

# World Journal of *Hepatology*

*World J Hepatol* 2015 December 8; 7(28): 2781-2858



## Editorial Board

2014-2017

The *World Journal of Hepatology* Editorial Board consists of 469 members, representing a team of worldwide experts in hepatology. They are from 53 countries, including Algeria (1), Argentina (6), Armenia (1), Australia (1), Austria (4), Bangladesh (2), Belgium (3), Botswana (2), Brazil (13), Bulgaria (2), Canada (3), Chile (1), China (98), Czech Republic (1), Denmark (2), Egypt (12), France (6), Germany (19), Greece (11), Hungary (5), India (15), Indonesia (2), Iran (4), Israel (1), Italy (52), Japan (35), Jordan (1), Malaysia (2), Mexico (3), Moldova (1), Netherlands (3), Nigeria (1), Pakistan (1), Philippines (2), Poland (1), Portugal (2), Qatar (1), Romania (6), Russia (2), Saudi Arabia (4), Singapore (1), South Korea (11), Spain (20), Sri Lanka (1), Sudan (1), Sweden (1), Switzerland (1), Thailand (4), Turkey (21), Ukraine (3), United Kingdom (17), and United States (56).

### EDITORS-IN-CHIEF

Clara Balsano, *Rome*  
Wan-Long Chuang, *Kaohsiung*

### GUEST EDITORIAL BOARD MEMBERS

King-Wah Chiu, *Kaohsiung*  
Tai-An Chiang, *Tainan*  
Chi-Tan Hu, *Hualien*  
Sen-Yung Hsieh, *Taoyuan*  
Wenya Huang, *Tainan*  
Liang-Yi Hung, *Tainan*  
Jih RU Hwu, *Hsinchu*  
Jing-Yi Lee, *Taipei*  
Mei-Hsuan Lee, *Taipei*  
Chih-Wen Lin, *Kaohsiung*  
Chun-Che Lin, *Taichung*  
Wan-Yu Lin, *Taichung*  
Tai-Long Pan, *Tao-Yuan*  
Suh-Ching Yang, *Taipei*  
Chun-Yan Yeung, *Taipei*

### MEMBERS OF THE EDITORIAL BOARD



**Algeria**

Samir Rouabhia, *Batna*



**Argentina**

Fernando O Bessone, *Rosario*  
Maria C Carrillo, *Rosario*  
Melisa M Dirchwolf, *Buenos Aires*  
Bernardo Frider, *Buenos Aires*

Jorge Quarleri, *Buenos Aires*  
Adriana M Torres, *Rosario*



**Armenia**

Narina Sargsyants, *Yerevan*



**Australia**

Mark D Gorrell, *Sydney*



**Austria**

Harald Hofer, *Vienna*  
Gustav Paumgartner, *Vienna*  
Matthias Pinter, *Vienna*  
Thomas Reiberger, *Vienna*



**Bangladesh**

Shahinul Alam, *Dhaka*  
Mamun Al Mahtab, *Dhaka*



**Belgium**

Nicolas Lanthier, *Brussels*  
Philip Meuleman, *Ghent*  
Luisa Vonghia, *Antwerp*



**Botswana**

Francesca Cainelli, *Gaborone*

Sandro Vento, *Gaborone*



**Brazil**

Edson Abdala, *Sao Paulo*  
Ilka FSF Boin, *Campinas*  
Niels OS Camara, *Sao Paulo*  
Ana Carolina FN Cardoso, *Rio de Janeiro*  
Roberto J Carvalho-Filho, *Sao Paulo*  
Julio CU Coelho, *Curitiba*  
Flavio Henrique Ferreira Galvao, *São Paulo*  
Janaina L Narciso-Schiavon, *Florianopolis*  
Sílvia HC Sales-Peres, *Bauru*  
Leonardo L Schiavon, *Florianópolis*  
Luciana D Silva, *Belo Horizonte*  
Vanessa Souza-Mello, *Rio de Janeiro*  
Jaques Waisberg, *Santo André*



**Bulgaria**

Mariana P Penkova-Radicheva, *Stara Zagora*  
Marieta Simonova, *Sofia*



**Canada**

Runjan Chetty, *Toronto*  
Michele Molinari, *Halifax*  
Giada Sebastiani, *Montreal*



**Chile**

Luis A Videla, *Santiago*



## China

Guang-Wen Cao, *Shanghai*  
 En-Qiang Chen, *Chengdu*  
 Gong-Ying Chen, *Hangzhou*  
 Jin-lian Chen, *Shanghai*  
 Jun Chen, *Changsha*  
 Alfred Cheng, *Hong Kong*  
 Chun-Ping Cui, *Beijing*  
 Shuang-Suo Dang, *Xi'an*  
 Ming-Xing Ding, *Jinhua*  
 Zhi-Jun Duang, *Dalian*  
 He-Bin Fan, *Wuhan*  
 Xiao-Ming Fan, *Shanghai*  
 James Yan Yue Fung, *Hong Kong*  
 Yi Gao, *Guangzhou*  
 Zuo-Jiong Gong, *Wuhan*  
 Zhi-Yong Guo, *Guangzhou*  
 Shao-Liang Han, *Wenzhou*  
 Tao Han, *Tianjin*  
 Jin-Yang He, *Guangzhou*  
 Ming-Liang He, *Hong Kong*  
 Can-Hua Huang, *Chengdu*  
 Bo Jin, *Beijing*  
 Shan Jin, *Hohhot*  
 Hui-Qing Jiang, *Shijiazhuang*  
 Wan-Yee Joseph Lau, *Hong Kong*  
 Guo-Lin Li, *Changsha*  
 Jin-Jun Li, *Shanghai*  
 Qiang Li, *Jinan*  
 Sheng Li, *Jinan*  
 Zong-Fang Li, *Xi'an*  
 Xu Li, *Guangzhou*  
 Xue-Song Liang, *Shanghai*  
 En-Qi Liu, *Xi'an*  
 Pei Liu, *Shenyang*  
 Zhong-Hui Liu, *Changchun*  
 Guang-Hua Luo, *Changzhou*  
 Yi Lv, *Xi'an*  
 Guang-Dong Pan, *Liuzhou*  
 Wen-Sheng Pan, *Hangzhou*  
 Jian-Min Qin, *Shanghai*  
 Wai-Kay Seto, *Hong Kong*  
 Hong Shen, *Changsha*  
 Xiao Su, *Shanghai*  
 Li-Ping Sun, *Beijing*  
 Wei-Hao Sun, *Nanjing*  
 Xue-Ying Sun, *Harbin*  
 Hua Tang, *Tianjin*  
 Ling Tian, *Shanghai*  
 Eric Tse, *Hong Kong*  
 Guo-Ying Wang, *Changzhou*  
 Yue Wang, *Beijing*  
 Shu-Qiang Wang, *Chengdu*  
 Mary MY Wayne, *Hong Kong*  
 Hong-Shan Wei, *Beijing*  
 Danny Ka-Ho Wong, *Hong Kong*  
 Grace Lai-Hung Wong, *Hong Kong*  
 Bang-Fu Wu, *Dongguan*  
 Feng Wu, *Chongqing*  
 Xiong-Zhi Wu, *Tianjin*  
 Chun-Fang Xu, *Suzhou*  
 Rui-An Xu, *Quanzhou*  
 Rui-Yun Xu, *Guangzhou*  
 Wei-Li Xu, *Shijiazhuang*  
 Shi-Ying Xuan, *Qingdao*  
 Ming-Xian Yan, *Jinan*  
 Lv-Nan Yan, *Chengdu*  
 Jin Yang, *Hangzhou*  
 Ji-Hong Yao, *Dalian*  
 Winnie Yeo, *Hong Kong*

Zheng Zeng, *Beijing*  
 Qi Zhang, *Hangzhou*  
 Shi-Jun Zhang, *Guangzhou*  
 Xiao-Lan Zhang, *Shijiazhuang*  
 Xiao-Yong Zhang, *Guangzhou*  
 Xin-Chen Zhang, *Harbin*  
 Yong Zhang, *Xi'an*  
 Hong-Chuan Zhao, *Hefei*  
 Ming-Hua Zheng, *Wenzhou*  
 Yu-Bao Zheng, *Guangzhou*  
 Ren-Qian Zhong, *Shanghai*  
 Fan Zhu, *Wuhan*  
 Xiao Zhu, *Dongguan*



## Czech Republic

Kamil Vyslouzil, *Olomouc*



## Denmark

Henning Gronbaek, *Aarhus*  
 Christian Mortensen, *Hvidovre*



## Egypt

Ihab T Abdel-Raheem, *Damanhour*  
 NGB G Bader EL Din, *Cairo*  
 Hatem Elalfy, *Mansoura*  
 Mahmoud M El-Bendary, *Mansoura*  
 Mona El SH El-Raziky, *Cairo*  
 Mohammad El-Sayed, *Cairo*  
 Yasser M Fouad, *Minia*  
 Mohamed AA Metwally, *Benha*  
 Hany Shehab, *Cairo*  
 Mostafa M Sira, *Shebin El-koom*  
 Ashraf Taye, *Minia*  
 MA Ali Wahab, *Mansoura*



## France

Laurent Alric, *Toulouse*  
 Sophie Conchon, *Nantes*  
 Daniel J Felmlee, *Strasbourg*  
 Herve Lerat, *Creteil*  
 Dominique Salmon, *Paris*  
 Jean-Pierre Vartanian, *Paris*



## Germany

Laura E Buitrago-Molina, *Hannover*  
 Enrico N De Toni, *Munich*  
 Oliver Ebert, *Muenchen*  
 Rolf Gebhardt, *Leipzig*  
 Janine V Hartl, *Regensburg*  
 Sebastian Hinz, *Kiel*  
 Benjamin Juntermanns, *Essen*  
 Roland Kaufmann, *Jena*  
 Viola Knop, *Frankfurt*  
 Veronika Lukacs-Kornek, *Homburg*  
 Benjamin Maasoumy, *Hannover*  
 Jochen Mattner, *Erlangen*  
 Nadja M Meindl-Beinker, *Mannheim*  
 Ulf P Neumann, *Aachen*  
 Margarete Odenthal, *Cologne*  
 Yoshiaki Sunami, *Munich*

Christoph Roderburg, *Aachen*  
 Frank Tacke, *Aachen*  
 Yuchen Xia, *Munich*



## Greece

Alex P Betrosian, *Athens*  
 George N Dalekos, *Larissa*  
 Ioanna K Delladetsima, *Athens*  
 Nikolaos K Gatselis, *Larissa*  
 Stavros Gourgiotis, *Athens*  
 Christos G Savopoulos, *Thessaloniki*  
 Tania Siahaniidou, *Athens*  
 Emmanouil Sinakos, *Thessaloniki*  
 Nikolaos G Symeonidi, *Thessaloniki*  
 Konstantinos C Thomopoulos, *Larissa*  
 Konstantinos Tziomalos, *Thessaloniki*



## Hungary

Gabor Banhegyi, *Budapest*  
 Peter L Lakatos, *Budapest*  
 Maria Papp, *Debrecen*  
 Ferenc Sipos, *Budapest*  
 Zsolt J Tulassay, *Budapest*



## India

Deepak N Amarapurkar, *Mumbai*  
 Girish M Bhopale, *Pune*  
 Sibnarayan Datta, *Tezpur*  
 Nutan D Desai, *Mumbai*  
 Sorabh Kapoor, *Mumbai*  
 Jaswinder S Maras, *New Delhi*  
 Nabeen C Nayak, *New Delhi*  
 C Ganesh Pai, *Manipal*  
 Amit Pal, *Chandigarh*  
 K Rajeshwari, *New Delhi*  
 Anup Ramachandran, *Vellore*  
 D Nageshwar Reddy, *Hyderabad*  
 Shivaram P Singh, *Cuttack*  
 Ajith TA, *Thrissur*  
 Balasubramaniyan Vairappan, *Pondicherry*



## Indonesia

Cosmas RA Lesmana, *Jakarta*  
 Neneng Ratnasari, *Yogyakarta*



## Iran

Seyed M Jazayeri, *Tehran*  
 Sedigheh Kafi-Abad, *Tehran*  
 Iradj Maleki, *Sari*  
 Fakhreddin Naghibalhossaini, *Shiraz*



## Israel

Stephen DH Malnick, *Rehovot*



## Italy

Francesco Angelico, *Rome*

Alfonso W Avolio, *Rome*  
 Francesco Bellanti, *Foggia*  
 Marcello Bianchini, *Modena*  
 Guglielmo Borgia, *Naples*  
 Mauro Borzio, *Milano*  
 Enrico Brunetti, *Pavia*  
 Valeria Cento, *Roma*  
 Beatrice Conti, *Rome*  
 Francesco D'Amico, *Padova*  
 Samuele De Minicis, *Fermo*  
 Fabrizio De Ponti, *Bologna*  
 Giovan Giuseppe Di Costanzo, *Napoli*  
 Luca Fabris, *Padova*  
 Giovanna Ferraioli, *Pavia*  
 Andrea Galli, *Florence*  
 Matteo Garcovich, *Rome*  
 Edoardo G Giannini, *Genova*  
 Rossano Girometti, *Udine*  
 Alessandro Granito, *Bologna*  
 Alberto Grassi, *Rimini*  
 Alessandro Grasso, *Savona*  
 Salvatore Gruttadauria, *Palermo*  
 Francesca Guerrieri, *Rome*  
 Quirino Lai, *Aquila*  
 Andrea Lisotti, *Bologna*  
 Marcello F Maida, *Palermo*  
 Lucia Malaguarnera, *Catania*  
 Andrea Mancuso, *Palermo*  
 Luca Maroni, *Ancona*  
 Francesco Marotta, *Milano*  
 Pierluigi Marzuillo, *Naples*  
 Sara Montagnese, *Padova*  
 Giuseppe Nigri, *Rome*  
 Claudia Piccoli, *Foggia*  
 Camillo Porta, *Pavia*  
 Chiara Raggi, *Rozzano (MI)*  
 Maria Rendina, *Bar*  
 Maria Ripoli, *San Giovanni Rotondo*  
 Kryssia I Rodriguez-Castro, *Padua*  
 Raffaella Romeo, *Milan*  
 Amedeo Sciarra, *Milano*  
 Antonio Solinas, *Sassari*  
 Aurelio Sonzogni, *Bergamo*  
 Giovanni Squadrito, *Messina*  
 Salvatore Sutti, *Novara*  
 Valentina Svicher, *Rome*  
 Luca Toti, *Rome*  
 Elvira Verduci, *Milan*  
 Umberto Vespasiani-Gentilucci, *Rome*  
 Maria A Zocco, *Rome*



#### Japan

Yasuhiro Asahina, *Tokyo*  
 Nabil AS Eid, *Takatsuki*  
 Kenichi Ikejima, *Tokyo*  
 Shoji Ikuo, *Kobe*  
 Yoshihiro Ikura, *Takatsuki*  
 Shinichi Ikuta, *Nishinomiya*  
 Kazuaki Inoue, *Yokohama*  
 Toshiya Kamiyama, *Sapporo*  
 Takanobu Kato, *Tokyo*  
 Saiho Ko, *Nara*  
 Haruki Komatsu, *Sakura*  
 Masanori Matsuda, *Chuo-city*  
 Yasunobu Matsuda, *Niigata*  
 Yoshifumi Nakayama, *Kitakyushu*  
 Taichiro Nishikawa, *Kyoto*

Satoshi Oeda, *Saga*  
 Kenji Okumura, *Urayasu*  
 Michitaka Ozaki, *Sapporo*  
 Takahiro Sato, *Sapporo*  
 Junichi Shindoh, *Tokyo*  
 Ryo Sudo, *Yokohama*  
 Atsushi Suetsugu, *Gifu*  
 Haruhiko Sugimura, *Hamamatsu*  
 Reiji Sugita, *Sendai*  
 Koichi Takaguchi, *Takamatsu*  
 Shinji Takai, *Takatsuki*  
 Akinobu Takaki, *Okayama*  
 Yasuhito Tanaka, *Nagoya*  
 Takuji Tanaka, *Gifu City*  
 Atsunori Tsuchiya, *Niigata*  
 Koichi Watashi, *Tokyo*  
 Hiroshi Yagi, *Tokyo*  
 Taro Yamashita, *Kanazawa*  
 Shuhei Yoshida, *Chiba*  
 Hitoshi Yoshiji, *Kashiwara*



#### Jordan

Kamal E Bani-Hani, *Zarqa*



#### Malaysia

Peng Soon Koh, *Kuala Lumpur*  
 Yeong Yeh Lee, *Kota Bahru*



#### Mexico

Francisco J Bosques-Padilla, *Monterrey*  
 María de F Higuera-de la Tijera, *Mexico City*  
 José A Morales-Gonzalez, *México City*



#### Moldova

Angela Peltec, *Chishinev*



#### Netherlands

Wybrich R Cnossen, *Nijmegen*  
 Frank G Schaap, *Maastricht*  
 Fareeba Sheedfar, *Groningen*



#### Nigeria

CA Asabamaka Onyekwere, *Lagos*



#### Pakistan

Bikha Ram Devrajani, *Jamshoro*



#### Philippines

Janus P Ong, *Pasig*  
 JD Decena Sollano, *Manila*



#### Poland

Jacek Zielinski, *Gdansk*



#### Portugal

Rui T Marinho, *Lisboa*  
 Joao B Soares, *Braga*



#### Qatar

Reem Al Olaby, *Doha*



#### Romania

Bogdan Dorobantu, *Bucharest*  
 Liana Gheorghe, *Bucharest*  
 George S Gherlan, *Bucharest*  
 Romeo G Mihaila, *Sibiu*  
 Bogdan Procopet, *Cluj-Napoca*  
 Streba T Streba, *Craiova*



#### Russia

Anisa Gumerova, *Kazan*  
 Pavel G Tarazov, *St.Petersburg*



#### Saudi Arabia

Abdulrahman A Aljumah, *Riyadh*  
 Ihab MH Mahmoud, *Riyadh*  
 Ibrahim Masoodi, *Riyadh*  
 Mhoammad K Parvez, *Riyadh*



#### Singapore

Ser Yee Lee, *Singapore*



#### South Korea

Young-Hwa Chung, *Seoul*  
 Dae-Won Jun, *Seoul*  
 Bum-Joon Kim, *Seoul*  
 Do Young Kim, *Seoul*  
 Ji Won Kim, *Seoul*  
 Moon Young Kim, *Wonu*  
 Mi-Kyung Lee, *Suncheon*  
 Kwan-Kyu Park, *Daegu*  
 Young Nyun Park, *Seoul*  
 Jae-Hong Ryoo, *Seoul*  
 Jong Won Yun, *Kyungsan*



#### Spain

Ivan G Marina, *Madrid*  
 Juan G Acevedo, *Barcelona*  
 Javier Ampuero, *Sevilla*  
 Jaime Arias, *Madrid*  
 Andres Cardenas, *Barcelona*  
 Agustin Castiella, *Mendaro*  
 Israel Fernandez-Pineda, *Sevilla*  
 Rocio Gallego-Duran, *Sevilla*  
 Rita Garcia-Martinez, *Barcelona*

José M González-Navajas, *Alicante*  
Juan C Laguna, *Barcelona*  
Elba Llop, *Madrid*  
Laura Ochoa-Callejero, *La Rioja*  
Albert Pares, *Barcelona*  
Sonia Ramos, *Madrid*  
Francisco Rodríguez-Frias, *Córdoba*  
Manuel L Rodríguez-Peralvarez, *Córdoba*  
Marta R Romero, *Salamanca*  
Carlos J Romero, *Madrid*  
Maria Traperó-Marugan, *Madrid*



#### **Sri Lanka**

Niranga M Devanarayana, *Ragama*



#### **Sudan**

Hatim MY Mudawi, *Khartoum*



#### **Sweden**

Evangelos Kalaitzakis, *Lund*



#### **Switzerland**

Christoph A Maurer, *Liestal*



#### **Thailand**

Taned Chitapanarux, *Chiang mai*  
Temduang Limpaboon, *Khon Kaen*  
Sith Phongkitkarun, *Bangkok*  
Yong Poovorawan, *Bangkok*



#### **Turkey**

Osman Abbasoglu, *Ankara*  
Mesut Akarsu, *Izmir*  
Umit Akyuz, *Istanbul*  
Hakan Alagozlu, *Sivas*  
Yasemin H Balaban, *Istanbul*  
Bulent Baran, *Van*  
Mehmet Celikbilek, *Yozgat*

Levent Doganay, *Istanbul*  
Fatih Eren, *Istanbul*  
Abdurrahman Kadayifci, *Gaziantep*  
Ahmet Karaman, *Kayseri*  
Muhsin Kaya, *Diyarbakir*  
Ozgur Kemik, *Van*  
Serdar Moralioglu, *Uskudar*  
A Melih Ozel, *Gebze - Kocaeli*  
Seren Ozenirler, *Ankara*  
Ali Sazci, *Kocaeli*  
Goktug Sirin, *Kocaeli*  
Mustafa Sunbul, *Samsun*  
Nazan Tuna, *Sakarya*  
Ozlem Yonem, *Sivas*



#### **Ukraine**

Rostyslav V Bubnov, *Kyiv*  
Nazarii K Kobyljak, *Kyiv*  
Igor N Skrypnyk, *Poltava*



#### **United Kingdom**

Safa Al-Shamma, *Bournemouth*  
Jayantha Arnold, *Southall*  
Marco Carbone, *Cambridge*  
Rajeev Desai, *Birmingham*  
Ashwin Dhanda, *Bristol*  
Matthew Hoare, *Cambridge*  
Stefan G Hubscher, *Birmingham*  
Nikolaos Karidis, *London*  
Lemonica J Koumbi, *London*  
Patricia Lalor, *Birmingham*  
Ji-Liang Li, *Oxford*  
Evaggelia Liaskou, *Birmingham*  
Rodrigo Liberal, *London*  
Wei-Yu Lu, *Edinburgh*  
Richie G Madden, *Truro*  
Christian P Selinger, *Leeds*  
Esther Una Cidon, *Bournemouth*



#### **United States**

Naim Alkhouri, *Cleveland*  
Robert A Anders, *Baltimore*  
Mohammed Sawkat Anwer, *North Grafton*  
Kalyan Ram Bhamidimarri, *Miami*

Brian B Borg, *Jackson*  
Ronald W Busuttill, *Los Angeles*  
Andres F Carrion, *Miami*  
Saurabh Chatterjee, *Columbia*  
Disaya Chavalitdhamrong, *Gainesville*  
Mark J Czaja, *Bronx*  
Jonathan M Fenkel, *Philadelphia*  
Catherine Frenette, *La Jolla*  
Lorenzo Gallon, *Chicago*  
Kalpana Ghoshal, *Columbus*  
Grigoriy E Gurvits, *New York*  
Hie-Won L Hann, *Philadelphia*  
Shuang-Teng He, *Kansas City*  
Wendong Huang, *Duarte*  
Rachel Hudacko, *Suffern*  
Lu-Yu Hwang, *Houston*  
Ijaz S Jamall, *Sacramento*  
Neil L Julie, *Bethesda*  
Hetal Karsan, *Atlanta*  
Ahmed O Kaseb, *Houston*  
Zeid Kayali, *Pasadena*  
Kusum K Kharbanda, *Omaha*  
Timothy R Koch, *Washington*  
Gursimran S Kochhar, *Cleveland*  
Steven J Kovacs, *East Hanover*  
Mary C Kuhns, *Abbott Park*  
Jiang Liu, *Silver Spring*  
Li Ma, *Stanford*  
Francisco Igor Macedo, *Southfield*  
Sandeep Mukherjee, *Omaha*  
Natalia A Osna, *Omaha*  
Jen-Jung Pan, *Houston*  
Christine Pocha, *Minneapolis*  
Yury Popov, *Boston*  
Davide Povero, *La Jolla*  
Phillip Ruiz, *Miami*  
Takao Sakai, *Cleveland*  
Nicola Santoro, *New Haven*  
Eva Schmelzer, *Pittsburgh*  
Zhongjie Shi, *Philadelphia*  
Nathan J Shores, *New Orleans*  
Siddharth Singh, *Rochester*  
Veysel Tahan, *Iowa City*  
Mehlika Toy, *Boston*  
Hani M Wadei, *Jacksonville*  
Gulam Waris, *North Chicago*  
Ruliang Xu, *New York*  
Jun Xu, *Los Angeles*  
Matthew M Yeh, *Seattle*  
Xuchen Zhang, *West Haven*  
Lixin Zhu, *Buffalo*  
Sasa Zivkovic, *Pittsburgh*

**TOPIC HIGHLIGHT**

- 2781 Long noncoding RNAs in hepatocellular carcinoma: Novel insights into their mechanism

*Liu YR, Tang RX, Huang WT, Ren FH, He RQ, Yang LH, Luo DZ, Dang YW, Chen G*

- 2792 Hepatitis C genotype 4: The past, present, and future

*Abdel-Ghaffar TY, Sira MM, El Naghi S*

- 2811 Bile acid receptors and nonalcoholic fatty liver disease

*Yuan L, Bambha K*

**REVIEW**

- 2819 Treating morbid obesity in cirrhosis: A quest of holy grail

*Kumar N, Choudhary NS*

**MINIREVIEWS**

- 2829 Update on hepatitis C: Direct-acting antivirals

*Seifert LL, Perumpail RB, Ahmed A*

- 2834 Contributions of transgenic mouse studies on the research of hepatitis B virus and hepatitis C virus-induced hepatocarcinogenesis

*Ohkoshi S, Hirono H, Watanabe K, Hasegawa K, Yano M*

**ORIGINAL ARTICLE**
**Retrospective Cohort Study**

- 2841 Comparison of peg-interferon, ribavirin plus telaprevir vs simeprevir by propensity score matching

*Fujii H, Nishimura T, Umemura A, Nishikawa T, Yamaguchi K, Moriguchi M, Sumida Y, Mitsuyoshi H, Yokomizo C, Tanaka S, Ishikawa H, Nishioji K, Kimura H, Takami S, Nagao Y, Takeuchi T, Shima T, Sawa Y, Minami M, Yasui K, Itoh Y*

**SYSTEMATIC REVIEWS**

- 2849 Epidemiology of hepatitis C virus exposure in Egypt: Opportunities for prevention and evaluation

*Miller FD, Elzalabany MS, Hassani S, Cuadros DF*

**ABOUT COVER**

Editorial Board Member of *World Journal of Hepatology*, Ji Won Kim, MD, PhD, Associate Professor, Department of Internal Medicine, Seoul National University College of Medicine, SMG-SNU Medical Center, Seoul 156-707, South Korea

**AIM AND SCOPE**

*World Journal of Hepatology* (*World J Hepatol*, *WJH*, online ISSN 1948-5182, DOI: 10.4254), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

*WJH* covers topics concerning liver biology/pathology, cirrhosis and its complications, liver fibrosis, liver failure, portal hypertension, hepatitis B and C and inflammatory disorders, steatohepatitis and metabolic liver disease, hepatocellular carcinoma, biliary tract disease, autoimmune disease, cholestatic and biliary disease, transplantation, genetics, epidemiology, microbiology, molecular and cell biology, nutrition, geriatric and pediatric hepatology, diagnosis and screening, endoscopy, imaging, and advanced technology. Priority publication will be given to articles concerning diagnosis and treatment of hepatology diseases. The following aspects are covered: Clinical diagnosis, laboratory diagnosis, differential diagnosis, imaging tests, pathological diagnosis, molecular biological diagnosis, immunological diagnosis, genetic diagnosis, functional diagnostics, and physical diagnosis; and comprehensive therapy, drug therapy, surgical therapy, interventional treatment, minimally invasive therapy, and robot-assisted therapy.

We encourage authors to submit their manuscripts to *WJH*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

**INDEXING/ ABSTRACTING**

*World Journal of Hepatology* is now indexed in PubMed Central, PubMed, Digital Object Identifier, Directory of Open Access Journals, and Scopus.

**FLYLEAF**

I-IV Editorial Board

**EDITORS FOR THIS ISSUE**

**Responsible Assistant Editor:** *Xiang Li*  
**Responsible Electronic Editor:** *Su-Qing Liu*  
**Proofing Editor-in-Chief:** *Lian-Sheng Ma*

**Responsible Science Editor:** *Fang-Fang Ji*  
**Proofing Editorial Office Director:** *Xiu-Xia Song*

**NAME OF JOURNAL**  
*World Journal of Hepatology*

**ISSN**  
 ISSN 1948-5182 (online)

**LAUNCH DATE**  
 October 31, 2009

**FREQUENCY**  
 36 Issues/Year (8<sup>th</sup>, 18<sup>th</sup>, and 28<sup>th</sup> of each month)

**EDITORS-IN-CHIEF**  
**Clara Balsano, PhD, Professor**, Departement of Biomedicine, Institute of Molecular Biology and Pathology, Rome 00161, Italy

**Wan-Long Chuang, MD, PhD, Doctor, Professor**, Hepatobiliary Division, Department of Internal Medicine, Kaohsiung Medical University Hospital, Kaohsiung Medical University, Kaohsiung 807, Taiwan

**EDITORIAL OFFICE**  
 Jin-Lei Wang, Director

Xiu-Xia Song, Vice Director  
*World Journal of Hepatology*  
 Room 903, Building D, Ocean International Center,  
 No. 62 Dongsihuan Zhonglu, Chaoyang District,  
 Beijing 100025, China  
 Telephone: +86-10-59080039  
 Fax: +86-10-85381893  
 E-mail: editorialoffice@wjnet.com  
 Help Desk: <http://www.wjnet.com/esps/helpdesk.aspx>  
<http://www.wjnet.com>

**PUBLISHER**  
 Baishideng Publishing Group Inc  
 8226 Regency Drive,  
 Pleasanton, CA 94588, USA  
 Telephone: +1-925-223-8242  
 Fax: +1-925-223-8243  
 E-mail: [bpgoffice@wjnet.com](mailto:bpgoffice@wjnet.com)  
 Help Desk: <http://www.wjnet.com/esps/helpdesk.aspx>  
<http://www.wjnet.com>

**PUBLICATION DATE**  
 December 8, 2015

**COPYRIGHT**  
 © 2015 Baishideng Publishing Group Inc. Articles published by this Open Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

**SPECIAL STATEMENT**  
 All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

**INSTRUCTIONS TO AUTHORS**  
 Full instructions are available online at [http://www.wjnet.com/1948-5182/g\\_info\\_20100316080002.htm](http://www.wjnet.com/1948-5182/g_info_20100316080002.htm)

**ONLINE SUBMISSION**  
<http://www.wjnet.com/esps/>

## 2015 Advances in Hepatocellular Carcinoma

**Long noncoding RNAs in hepatocellular carcinoma: Novel insights into their mechanism**

Yong-Ru Liu, Rui-Xue Tang, Wen-Ting Huang, Fang-Hui Ren, Rong-Quan He, Li-Hua Yang, Dian-Zhong Luo, Yi-Wu Dang, Gang Chen

Yong-Ru Liu, Rui-Xue Tang, Wen-Ting Huang, Fang-Hui Ren, Dian-Zhong Luo, Yi-Wu Dang, Gang Chen, Department of Pathology, First Affiliated Hospital of Guangxi Medical University, Nanning 530021, Guangxi Zhuang Autonomous Region, China

Rong-Quan He, Li-Hua Yang, Department of Medical Oncology, First Affiliated Hospital of Guangxi Medical University, Nanning 530021, Guangxi Zhuang Autonomous Region, China

**Author contributions:** Liu YR, Tang RX, Huang WT, Ren FH, He RQ and Yang LH performed the literature search, wrote the first draft of the manuscript and approved the final version; Luo DZ, Dang YW and Chen G edited the final draft of the manuscript and approved the final version.

**Supported by** Partly Fund of Guangxi Provincial Health Bureau Scientific Research Project, No. Z2014054; Youth Science Foundation of Guangxi Medical University, No. GXMUYSF201311; Guangxi University Science and Technology Research Projects, No. LX2014075; Natural Science Foundation of Guangxi, Nos. 2015GXNSFBA139157, 2015GXNSFCA139009; and National Natural Science Foundation of China, Nos. NSFC81360327, NSFC81560489, NSFC81560469.

**Conflict-of-interest statement:** The authors have not declared any conflicts of interest.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to:** Dr. Gang Chen, MD, PhD, Department of Pathology, First Affiliated Hospital of Guangxi Medical University, 6 Shuangyong Road, Nanning 530021, Guangxi Zhuang Autonomous Region, China. [chen\\_gang\\_triones@163.com](mailto:chen_gang_triones@163.com)

Telephone: +86-771-5356534  
Fax: +86-771-5358943

Received: April 30, 2015  
Peer-review started: May 8, 2015  
First decision: September 8, 2015  
Revised: November 3, 2015  
Accepted: November 24, 2015  
Article in press: November 25, 2015  
Published online: December 8, 2015

**Abstract**

Hepatocellular carcinoma (HCC) is the predominant subject of liver malignancies which arouse global concern. Advanced studies have found that long non-coding RNAs (lncRNAs) are differentially expressed in HCC and implicate they may play distinct roles in the pathogenesis and metastasis of HCC. However, the underlying mechanisms remain largely unclear. In this review, we summarized the functions and mechanisms of those known aberrantly expressed lncRNAs identified in human HCC tissues. We hope to enlighten more comprehensive researches on the detailed mechanisms of lncRNAs and their application in clinic, such as being used as diagnostic and prognostic biomarkers and the targets for potential therapy. Although studies on lncRNAs in HCC are still deficient, an improved understanding of the roles played by lncRNAs in HCC will lead to a much more effective utilization of those lncRNAs as novel candidates in early detection, diagnosis, prevention and treatment of HCC.

**Key words:** Hepatocellular carcinoma; Long noncoding RNA; Dysregulation; Mechanism; Pathway

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Hepatocellular carcinoma (HCC) is a global concern. Long noncoding RNAs (lncRNAs) are likely to play crucial roles in various pathogenesis of HCC, including tumor growth, proliferation, invasion, metastasis and recurrence. Here, we focus on recent studies of human HCC associated lncRNAs and highlight their functions, mechanisms, as well as their potential to act as novel candidates for early detection, diagnosis, prevention and treatment of HCC.

Liu YR, Tang RX, Huang WT, Ren FH, He RQ, Yang LH, Luo DZ, Dang YW, Chen G. Long noncoding RNAs in hepatocellular carcinoma: Novel insights into their mechanism. *World J Hepatol* 2015; 7(28): 2781-2791 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i28/2781.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i28.2781>

## INTRODUCTION

Hepatocellular carcinoma (HCC), one of the most common histologic subtype of primary liver cancers, accounting for 70%-85% of liver cancer cases in most countries, is the third-leading cause of worldwide mortality for various cancers<sup>[1-3]</sup>. It causes nearly 695900 deaths per year, half of which occur in China, as a result of the high chronic hepatitis B virus (HBV) infection incidence<sup>[4]</sup>. Besides, infection with hepatitis C virus, hepatosteatosis and chronic exposure to toxic chemical substances are the major HCC risk factors<sup>[5,6]</sup>. Especially, exposure to aflatoxin plays a key role in inducing HCC in our Guangxi Zhuang Autonomous Region<sup>[7]</sup>. In addition, many key signal transduction pathways have been verified to be involved in the pathogenesis of HCC, including PI3K/Akt/mTOR pathway, Raf/MAPK/ERK pathway, Jak/Stat pathway, WNT-b-catenin pathway, so on and so forth<sup>[8-12]</sup>. Usually, the treatments for HCC are finite and are only available in the early stage, while most HCC are only detected in their advanced stage when traditional chemotherapy has marginal effects and leads to poor prognosis. Because of its dismal outcome, etiology and carcinogenesis investigation are in urgent need. Recently, human genome analysis in non-protein coding has made new progress. It discovers massive transcription of large RNA transcripts which lack coding protein function, termed as long noncoding RNAs (lncRNAs).

With recent application of next generation sequencing techniques, significant numbers of non-coding RNAs (ncRNAs) and lncRNAs have been discovered to be associated with HCC. And HCC is characterized by dysregulation of numerous gene networks while both protein-coding genes and ncRNA genes are involved, just like many other cancers. It is estimated that protein-coding genes only account for less than 2% of the human genome while nearly 70% of the human genome is transcribed pervasively<sup>[13]</sup>. Accordingly, ample ncRNAs are transcribed from human genome, for instance lncRNAs, microRNAs (miRNAs), small interfering RNAs

and PIWI-interacting RNAs<sup>[14-16]</sup>.

ncRNAs were once thought to be body "garbage" or transcriptional "noise". However, accumulating reports have demonstrated that miRNAs and lncRNAs play valid regulatory roles in cancer<sup>[17-22]</sup>. Recent studies<sup>[23]</sup> have also elucidated that lncRNAs possess a significant role in epigenetic regulation. *Via* regulating gene expression by miscellaneous mechanisms, including genomic imprinting, chromatin modification, regulation of protein function, transcription and post-transcriptional processing<sup>[24-26]</sup>, lncRNAs are involved in multitudinous physiological functions and pathological processes.

lncRNAs, larger than 200 nucleotides (nt) in length, are commonly defined as endogenous cellular RNA molecules, which are poorly conserved and not capable of being translated into proteins<sup>[21,23,27]</sup>. They can be monitored by a high-throughput analysis such as transcriptome analysis and microarrays, or through bioinformatics prediction<sup>[28]</sup>. lncRNAs can be transcribed by RNA polymerase II, and then undergo cotranscriptional modifications including polyadenylation and pre-RNA splicing<sup>[29]</sup>. Many studies have pointed out that lncRNA transcripts play vital roles in various biological processes as they function in gene imprinting and splicing, chromatin modification, immunosurveillance, cell fate specification, cell cycle control and cell apoptosis, or act as nuclear architecture, subnuclear compartments, RNA processing enhancer and promoter<sup>[30,31]</sup>.

lncRNAs have assorted mechanisms in biological processes. Generally speaking, the role of lncRNA as a gene expressing regulator could be found in transcriptional level and posttranscriptional level. Cis-regulation and trans-regulation are two main transcriptional regulation means, by which lncRNAs can target local and distant genes, respectively. The posttranscriptional regulating mechanism is involved in posttranscriptional process of mRNAs which includes splicing, editing, trafficking, translation and degradation. lncRNAs can also function as competing endogenous RNAs for shared miRNA<sup>[32,33]</sup>. In brief, there are four known molecular functions of lncRNAs: Signal, decoy, guide, and scaffold<sup>[21,34]</sup>.

There are five species of lncRNAs, listed as follow<sup>[31,35,36]</sup>: Sense or antisense (when overlapping at least one exon of another transcript on the opposite or same strand), bidirectional (when a neighboring coding transcript or its expression on the opposite strand is initiated in close genomic proximity), intronic (when derived from an intron of a second transcript), and intergenic (when it lies as an independent unit within the genomic interval between two genes). An updated definition was given to lncRNAs by other researchers regardless of their length and non-protein coding capability<sup>[37]</sup>. It described lncRNAs as RNA molecules who may have the function as primary or spliced transcripts which do not confirm to the known varieties of small RNAs or structural RNAs<sup>[38]</sup>. In recent years, the number of articles focused on lncRNAs has increased greatly. Recent studies have demonstrated that certain

**Table 1 Upregulated long noncoding RNAs in hepatocellular carcinoma**

Name	Gene locus	Size (bp)	Dysregulation	Potential role in HCC	Ref.
HULC	6p24.3	1638	Upregulated	Associate with HBV infection or histological grade. Associate with tumor growth	[22,29,44-50]
H19	11p15.5	2660	Upregulated	Suppress progression, metastasis. Promote cell proliferation	[22,29,50-57]
TUC338	12q13.13	590	Upregulated	Increased in liver cirrhosis. Modulate cell growth	[58]
MALAT1	11q13.1	8708	Upregulated	Associate with tumor metastasis, recurrence	[21,22,29,59,60]
HOTAIR	12q13.13	12649	Upregulated	Associate with invasion and metastasis. Increases chemosensitivity	[21,22,29,61-65]
HOTTIP	7p15.2	6839	Upregulated	Associate with tumor progression and disease outcome	[28,66-68]
HEIH	5q35.3	1665	Upregulated	Associated with HBV-HCC. Associate with prognosis	[21,29,69]
MDIG	3q11.2	30635	Upregulated	Associate with DNA repair and prognosis	[70]
PVT1	8q24.21	210626	Upregulated	Associate with HCC progression and predict recurrence	[71,72]
Linc00974	17q21.31	4890	Upregulated	Predict tumor growth and metastasis	[73]
UFC1	1q23.3	5113	Upregulated	Promote HCC cell proliferation, inhibit cell apoptosis and induce cell cycle progression	[10,12,74]
PCNA-AS1	20p12.3	384	Upregulated	Promote tumor growth	[75]
UCA1	19p13.12	7375	Upregulated	Involved in chemotherapeutic resistance. Associate with TNM stage, metastasis and postoperative survival	[50,76]
CCAT1	8q24.21	11887	Upregulated	Promotes HCC progression	[77-79]
ATB	19q13.3	2895	Upregulated	Associate with poor prognosis	[80,81]
URHC	2q24.2	192173	Upregulated	Promote cell proliferation and inhibit apoptosis	[82]

HCC: Hepatocellular carcinoma; HULC: Highly upregulated in liver cancer; MALAT1: Metastasis-associated lung adenocarcinoma transcript 1; HOTAIR: HOX transcript antisense RNA; HEIH: Long noncoding RNA high expression in HCC; CCAT1: Colon cancer associated transcript-1; ATB: Long noncoding RNA-activated by transforming growth factor- $\beta$ ; URHC: Up-regulated in hepatocellular carcinoma; HBV: Hepatitis B virus; TNM: Tumor node metastasis.

**Table 2 Downregulated long noncoding RNAs in hepatocellular carcinoma**

Name	Gene locus	Size (bp)	Dysregulation	Potential role in HCC	Ref.
MEG3 (GTL2)	14q32.3	34919	Downregulated	Associate with methylation. Predictive biomarker for monitoring epigenetic therapy	[21,83-87]
LncRNA-LET	15q24.1	2606	Downregulated	Reduces hepatic invasion and abdominal metastases	[88]
PTENP1	9p13.3	3917	Downregulated	Repress the tumorigenic properties of HCC cells	[22,89]
SRHC	5p15.31	6365	Downregulated	Inhibit cancer proliferation	[90]
MT1DP	16q13	1255	Downregulated	Act as a tumor suppressor. Overexpression of MT1DP decreases cell proliferation but increases apoptosis	[91]

HCC: Hepatocellular carcinoma; MT1DP: Metallothionein 1D, pseudogene; MEG3: Maternally expressed gene 3; LncRNA: Long noncoding RNA.

lncRNAs are specifically correlated with certain classes of cancer and the different expression level of lncRNAs may function as an indicator for metastasis and prognosis<sup>[39-41]</sup>. Genome-wide transcriptomic analyses have found that a number of lncRNAs are dysregulated in HCC cell lines or cancer tissues<sup>[21,28,29,42,43]</sup>. Given the fact that such large scales of lncRNAs are aberrantly regulated in HCC, it is of highly possibility that lncRNAs are directly associated with carcinogenesis of HCC. In this review, we concentrated on cancer-related lncRNAs which have been validated in human HCC. Furthermore, we summarized their mechanism and signaling pathways in HCC.

## REPRESENTATIVE LNCRNAs DYSREGULATED IN HCC

Abnormal expressed lncRNAs have been found to be associated with hepatocarcinogenesis and play a key role in metastasis and prognosis<sup>[29,34,42]</sup>. Accumulating studies recently have focused on the contributions of lncRNAs in HCC development. Here, we summarized differential expressions of lncRNAs and their potential roles in HCC (Tables 1 and 2).

## MECHANISMS AND SIGNALING PATHWAY IN HCC

After attentive study, we divided the researched lncRNAs into 3 groups and made the above conclusive table. And with the purpose of identifying the roles of different lncRNAs who might play in the early detection and diagnosis or prevention and treatment of HCC, we further summarized their function and mechanism as follow.

### LNCRNAs ASSOCIATED WITH TUMOR GROWTH AND PROLIFERATION IN HCC

#### Highly upregulated in liver cancer

Highly upregulated in liver cancer (HULC), a well-researched lncRNA who associates with HBV infection and HCC tumor growth is upregulated in HCC and associated with its grades<sup>[45]</sup>. By viewing dozens of research articles, we found that HULC can accelerate the growth of HCC *via* downregulating its neighbor gene p18 (known as eukaryotic translation elongation factor 1, EEF1E1 or AIMP3) and can disturb the circadian rhythm of HCC *via* upregulating oscillator CLOCK<sup>[47]</sup>.

The complementary base which pairs between 5'UTR of CLOCK mRNA and HULC takes responsibility for the modulation of CLOCK mediated by HULC<sup>[91]</sup>. With regard to the mechanism of HULC upregulation, HBx regulates the transcription of CREB-dependent promoters by interacting with CREB, and then, the transcription factor CREB contributes to the activation of HULC promoter<sup>[47,48]</sup>. The variant genotypes of rs7763881 in HULC contribute to decreasing HCC susceptibility in persistent HBV carriers. And single nucleotide polymorphisms (SNPs) in HULC contribute to the risk of HBV chronic infection and HCC<sup>[92]</sup>. Besides, HULC acts as an endogenous "sponge", who downregulates a series of miRNAs activities, including miR-372. Studies indicate that inhibition of miR-372 results in decreased translational repression of its target gene, PRKACB, and inducing phosphorylation of CREB in turn<sup>[48]</sup>.

Based on the above mechanisms, HULC may have the potential of predicting prognosis in clinical practice. However, only cell lines research were done by researchers, other confirmatory experiments are requisite.

#### **LncRNA-hPVT1**

Two studies investigated closely into lncRNA-hPVT1 and concluded that it has a function of promoting cell proliferation, cell cycling and it also functions as an acquisition of stem-cell like contents in HCC cells. LncRNA-hPVT1 upregulates nucleolar protein p120 (NOP2) *via* enhancing the stability of NOP2 proteins and its above functions depend on the presence of NOP2. Studies show that the transforming growth factor (TGF)- $\beta$ 1/lncRNA-hPVT1/NOP2 pathway is compromised in the progression of HCC. Hence, lncRNA-hPVT1 influences the stem-cell like potential of HCC cells and promotes the growth of HCC. Regulation of the lncRNA-hPVT1/NOP2 pathway has a beneficial effect in the treatment of HCC<sup>[71]</sup>. More researches are needed to be done in order to find effective therapy targeted at lncRNA-hPVT1.

#### **UFC1**

Other than the above three lncRNAs, another well-studied lncRNA associated with HCC proliferation is lncRNA-UFC1 (GenBank Accession No. KJ809564), who promotes HCC cell proliferation, induces cell cycle progression and inhibits cell apoptosis<sup>[73]</sup>. It induces HuR translocation and by silencing HuR expression can abrogate the function of lncRNA-UFC1 function in HCC. Moreover, lncRNA-UFC1 is targeted by miR-34a and the overexpression of miR-34a significantly suppresses the expression levels of cell cycle related proteins, cellular proliferation and HuR expression in lncRNA-UFC1-overexpressing cells<sup>[10]</sup>. As molecularly targeted therapies are heated studied, UFC1 offers us a new aspect, clinical research are in the urgent need.

#### **ZNRD1 antisense RNA 1**

A large case-control study including 1344 HBV natural-clearance subjects, 1344 HBV persistent carriers and

1300 HBV-positive HCC patients was done<sup>[93]</sup>. The study found out that ZNRD1 antisense RNA 1 (ZNRD1-AS1) is a crucial regulator of ZNRD1 (human zinc ribbon domain containing 1). In ZNRD1-AS1, several SNPs (nucleotide polymorphisms) is identified as expression quantitative trait loci (eQTLs) SNPs, which are connected with the expression of ZNRD1<sup>[94,95]</sup>. ZNRD1 is involved in DNA damage and repair *via* regulating the expression of excision repair cross-complementing 1 (ERCC1)<sup>[96]</sup>, to restrain cell proliferation and to regulate the expression of miRNAs in cancers<sup>[97,98]</sup>. Furthermore, ZNRD1 eQTLs SNPs in lncRNA ZNRD1-AS1 have an increased risk for persistent HBV-carriers HCC but a protective influence against chronic HBV infection<sup>[93]</sup>. As a result, the different roles which ZNRD1-AS1 play make a difference in the treatment of HBV-positive HCC patients and HBV-negative HCC patients.

#### **Colon cancer associated transcript-1**

Dysregulation of colon cancer associated transcript-1 (CCAT1) is in association with tumor size, microvascular invasion, AFP and prognosis in patients with HCC. Besides, it is demonstrated that *in vitro* CCAT1 could promote proliferation and migration in HCC by binding to let-7, which contributes to the up-regulation of HMGA2 and c-Myc<sup>[76]</sup>. Herein, the complex of CCAT1 and let-7 may have the diagnostic function in early detection of HCC and its migration.

#### **Maternally expressed gene 3**

Maternally expressed gene 3 (*MEG3*) regulates tumor cell proliferation and apoptosis in HCC partially through the accumulation of p53<sup>[83]</sup>. UHRF1, as a new identified oncogene, contributes to the upregulation of *MEG3* in HCC by regulating DNMT1, while upregulation of *MEG3* in HCC cells can partially diminish the promotion of proliferation produced by UHRF1. In addition, UHRF1/DNMT1/*MEG3*/p53 axis signaling pathway is involved in HCC progression<sup>[99]</sup>. Furthermore, loss of *MEG3* gene expression is related to hypermethylation of the promoter region in HCC<sup>[82]</sup>. Impressively, enforced expression of *MEG3* in HCC remarkably decreases both anchorage-independent and anchorage-dependent cell growth, and induces cell apoptosis<sup>[83]</sup>. Associated with anti-oncogene p53, *MEG3* shows a promising future in being one of the therapeutic targets for HCC treatment.

#### **PTENP1**

The over-expression of PTENP1 (a pseudogene of PTEN) represses the oncogenic PI3K/AKT pathway and elicits pro-death autophagy by sequestering miR-20a, miR-19b and miR-17 *in vitro*. It also inhibits tumor growth *in vivo*. These are accompanied by dampened angiogenesis or neovasculature maturation, enhanced apoptosis and autophagy<sup>[88]</sup>. It's necessary to do further *in vivo* experiments to confirm its detailed mechanism.

The above 7 lncRNAs have been widely studied, and they are considered to be associated with tumor growth and proliferation in HCC. With further clinical trials, their

application in the prediction and diagnosis of HCC would be possible.

## LNCRNAs ASSOCIATED WITH METASTASIS AND PROGNOSIS IN HCC

Invasion and metastasis often adumbrate an advanced stage while recurrence often indicates a poor prognosis. It's the same in HCC development. The following lncRNAs were found associated with metastases and recurrence which may predict a dismal outcome.

### H19

A dozen of studies have elucidated that H19 serves as a potential prognostic marker as well as potential target for HCC therapy. By decreasing the expression of markers for epithelial-to-mesenchymal transition, such as claudin 1, cytokeratin-8 (KRT-8), KRT-19 and CDH1 (E-cadherin), H19 suppresses the progression of HCC. By mediating hnRNP/PCAF/RNAPol II, H19 suppresses the migration of HCC. By increasing histone acetylation, H19 can epigenetically activates miR-200 family, and thus, it suppresses HCC metastasis<sup>[50]</sup>. Moreover, the identification of AKT/GSK-3 $\beta$ /Cdc25A signaling pathway as the downstream signaling pathway of H19 explains the molecular mechanism of metastasis and invasion in HCC<sup>[100]</sup>. And it has also been demonstrated that the deletion of H19 endodermal enhancer can regulate expression of insulin-like growth factor 2 (IGF2) and H19 in the early stage of liver carcinogenesis as well as that paternal inheritance of the deletion of H19 endodermal enhancer can delay tumor formation by increasing apoptosis of the hepatocytes and reducing IGF2 expression<sup>[101]</sup>. In the main, by various signaling pathway, H19 can be a promising indicator for prognosis and can be targeted in the treatment of HCC.

### Metastasis-associated lung adenocarcinoma transcript 1

Metastasis-associated lung adenocarcinoma transcript 1 (MALAT1) is a critical regulator of maintaining the transformative phenotype in HCC, it associates with tumor metastasis and recurrence<sup>[102]</sup>. MALAT1 is a genuine target gene of the Wnt/TCF/ $\beta$ -catenin and Hippo/yes associated protein (YAP) signaling pathway. It is negatively regulated by SRSF1 *via* the following two pathways. The first pathway is by accelerating the degradation of MALAT1 which is blocked by YAP at the stage of post-transcriptional. When overexpressed, YAP stimulates the translocation of SRSF1 from the nucleus to the cytoplasm, thus the nuclear-retained MALAT1 avoids degradation<sup>[103]</sup>. The second pathway is by binding to YAP. SRSF1 inhibits the transcriptional activity of YAP and prevents the recruitment of YAP on the MALAT1 promoter at a transcriptional stage<sup>[104]</sup>.

MALAT1 can interact with the arginine/serine SR proteins and modulate their distribution to the nuclear speckles. Furthermore, MALAT1 can regulate alternative splicing of pre-mRNAs *via* controlling active SR proteins'

levels<sup>[103]</sup>.

To sum up, by participating in different pathway, MALAT1 shows a crucial role in the carcinogenesis of HCC which lays the foundation of early detection of metastases and assessment of recurrence.

### HOX transcript antisense RNA

High expression of HOX transcript antisense RNA (HOTAIR) indicates a notably poorer prognosis with respect to overall survival (OS) and a remarkably larger tumor size in HCC patients<sup>[62]</sup>. HOTAIR is selectively required to target polycomb repressive complex 2 (PRC2) occupancy, thus, induces histone H3 tri-methylated at lysine 27 (H3K27) trimethylation and silenced transcription of the HOXD locus<sup>[105]</sup>. The 5' domain of HOTAIR can bind to PRC2, while the 3' domain of HOTAIR can bind to the LSD1 (lysine specific demethylase 1)/CoREST (Co-repressor of RE1-silencing transcription factor)/REST complex; after that, the complexes are targeted and assembled to the HOXD locus; and co-ordinately regulate histone H3K27 methylation and histone histone 3 methylated at lysine 4 demethylation; consequently, the transcription across 40 kb of the HOXD locus is silenced in trans by DNA methylation<sup>[106]</sup>. These indicate that the HOTAIR-induced assembly and targeting of LSD1 and PRC2 complexes is a general mechanism for gene silencing across the genome, which plays a vital role in the association with invasion and metastasis in HCC<sup>[64]</sup>. Expression of HOTAIR is also associated with tumor size and lymph node metastasis in HCC patients<sup>[62]</sup>. Knockdown of HOTAIR reduces the levels of matrix metalloproteinase 9 and vascular endothelial growth factor, which play an important role in metastasis and cell motility<sup>[60]</sup>.

In addition, HOTAIR is found to promote invasion and migration in HCC by inhibiting RBM38. Silencing of RBM38 restores cell motility, while knockdown of HOTAIR conspicuously reduces cell motility as the downregulation of HOTAIR increases the expression of RBM38 both on mRNA levels and protein levels<sup>[63]</sup>.

With its association with tumor size, tumor metastasis and OS, HOTAIR is regarded as one of the prognosis indicator. It's of great potential value to use it in the clinic in the future.

### HOTTIP

HOTTIP (the transcript of HOXA at the distal tip with 3.8 kb), transcribed from the 5' tip of the HOXA locus, coordinates the activation of some 5' HOXA genes and is negatively regulated by miR-125b. High HOTTIP expression indicates an increased metastasis formation and a decreased overall survival<sup>[65]</sup>. HOTTIP binds the adaptor protein WDR5 directly and targets WDR5/MLL complexes across HOXA, thereby driving histone H3 lysine 4 trimethylation and the gene transcription and influences HCC cell proliferation rates as a result<sup>[27]</sup>. Its overexpression contributes to hepatocarcinogenesis through regulating the expression of its neighboring protein-coding genes<sup>[67]</sup>. With these function, HOTTIP

can be potentially used in clinic as a prognosis predictor.

### **LncRNA high expression in HCC**

LncRNA high expression in HCC (HEIH), associated with recurrence and overall survival in HBV-related HCC, is overexpressed in HCC tissues<sup>[68]</sup>. HEIH influences the expression of enhancer of zeste homolog 2 (EZH2) target genes by increasing the binding of EZH2 levels. Downregulation of HEIH induces G(0)/G(1) arrest which is caused by the interaction of EZH2 with HEIH. Thus, level of HEIH is significantly associated with recurrence and is an independent prognostic factor for survival in HCC<sup>[68]</sup>. In summary, HEIH connects with HBV-related HCC and plays a role as the risk factor for HCC recurrence. Therefore, predicting the possibility of HCC recurrence by detecting the relative change of the HBV virus and HEIH level in serum is worth further studying.

### **ATB**

Overexpression of lncRNA-ATB (lncRNA-activated by TGF- $\beta$ ) significantly correlates with EMT gene signature expression, macrovascular invasion, microvascular invasion, portal vein tumor thrombus and encapsulation. Moreover, higher expression of lncRNA-ATB is significantly correlated with shorter recurrence-free survival and overall survival, suggesting that lncRNA-ATB contributes to HCC progression<sup>[27]</sup>.

LncRNA-ATB upregulates ZEB1 and ZEB2 *via* competitively binding with the miR-200 family, then it induces EMT and invasion. Besides, lncRNA-ATB promotes organ colonization of disseminated HCC tumor cells through binding with autocrine induction of interleukin 11 (IL-11), IL-11 mRNA and triggering signal transducer and activator of transcription 3 signaling<sup>[79,80]</sup>. On the whole, lncRNA-ATB facilitates the invasion-metastasis cascade, which makes it a predictor for HCC prognosis.

### **LncRNA-p21**

LncRNA-p21, located upstream of *CDKN1A* gene, who triggers apoptosis in HCC, is a transcriptional target of p53 and *p53* gene is an important tumor suppressor gene<sup>[107,108]</sup>. Importantly, lncRNA-p21 can bind to heterogeneous nuclear ribonucleoprotein K (hnRNP-K), therefore it contributes to the localization of hnRNP-K and transcriptional repression of p53-regulated genes<sup>[109]</sup> and as a result, triggers apoptosis. Theoretically, lncRNA-p21 should be down-regulated in HCC, however, corroborating experiments are needed to delineate the exact underlying mechanism. As the transcriptional target of anti-oncogene p53, p21 shows a promising future in being one of the therapeutic targets for inducing cellular apoptosis.

The above 7 lncRNAs are considered to be associated with metastasis and prognosis in HCC. Although many researches have been complicated, studies with more cases and further ward clinical trials would be needed in order to develop novel therapeutics and treatment for

HCC patients.

## **LATEST FIND OF LNCRNAs RELATED TO HCC**

### **Linc00974**

Dysregulation of Linc00974 increases KRT19 levels, which results in the activation of both TGF- $\beta$  and Notch signaling pathways, which causes the invasion and proliferation of HCC both *in vivo* and *in vitro*. Linc00974 influenced KRT19 expression by interacting with miR-642<sup>[72]</sup>. Being significantly correlated with tumor differentiation grade, size and metastasis makes Linc00974 a feasible predictor in the carcinogenesis of HCC.

### **Up-regulated in hepatocellular carcinoma**

High level of up-regulated in hepatocellular carcinoma (URHC) expression is significantly associated with tumor size, tumor number and shorter overall survival after surgery<sup>[81]</sup>. URHC can regulate cell proliferation and apoptosis by repressing ZAK expression. ZAK, also known as MLK-like MAP triple kinase- $\alpha$  or ZAK- $\alpha$ , belongs to the mixed lineage kinase family and functions as a tumor-suppressor gene in HCC<sup>[110]</sup>. Inactivation of the ERK/MAPK pathway is required for the increase in HCC growth, which is induced by URHC-ZAK regulation<sup>[81]</sup>. However, only microarray analysis was done, which limited its conviction. Experiments *in vivo* and *in vitro* are desperately needed.

### **SRHC**

SRHC (NCBI No: uc003jdr) is an important downstream target gene of HNF-4A and it is correlated with  $\alpha$ -feto-protein (AFP) levels and the degree of differentiated tumors<sup>[89]</sup>. SRHC is combined with HNF-4A to promote its transcription, thus inhibiting the proliferation of tumor cells and promoting cell differentiation in HCC<sup>[89]</sup>. Serum AFP level monitoring is well developed and have been used as a standardized index of HCC diagnosis, therefore, detecting SRHC in predicting HCC may have great value in clinical practice as well.

### **Metallothionein 1D, pseudogene**

Metallothionein 1D, pseudogene (MT1DP) inhibits tumor cell growth could be rescued by a combination of overexpression of Runt related transcription factor 2, FoxA1 and YAP. In addition, MT1DP inhibited trans-formative phenotype of liver cancer cells and cell proliferation by reducing protein synthesis of FoxA1<sup>[90]</sup>. With this inhibiting function, it's of great research value whether MT1DP can be a potential therapeutic target in the treatment of HCC or not.

### **LncRNA low expression in tumor**

LncRNA low expression in tumor (LncRNA-LET) plays a decisive role in hypoxia-induced metastasis in HCC through a hypoxia-inducible factor 1, alpha subunit

(HIF-1a)/histone deacetylase 3 (HDAC3)/LET/NF90 pathway<sup>[110]</sup>. Precisely, HDAC3 represses LET by decreasing the LET promoter region's histone acetylation-mediated modulation under hypoxic conditions. And then, down-regulation of LET recedes the direct interactions between LET and NF90, then enhances the stabilization of NF90 and increases the expression of HIF-1a (a target mRNA of NF90 involved in hypoxia-induced metastasis). As a result, LET inhibits the metastasis of HCC *via* this positive feedback loop<sup>[87]</sup>. Therefore, when LET is downregulated, patients usually face a poor prognosis.

For these 5 newly found lncRNAs, investigations with more preclinical models of HCC would be desired in order to further strengthen the conclusions and provide a more rational support for ward clinical application.

## CONCLUSION

In conclusion, lncRNAs play a crucial role in various biological processes in HCC, such as initiation, progression, metastasis, treatment and prognosis.

Two latest published articles also reached the same conclusion. On the basis of dozens of studies, Yang *et al.*<sup>[111]</sup> discussed the probable molecular mechanisms depended on lncRNA level change, and drew the conclusion that lncRNAs can be applied in HCC diagnosis and treatment. However, their summary was omissive as they only analyzed the upregulated lncRNAs, and even some upregulated lncRNAs were left out, such as HOTTIP, MVIH, UFC1, UCA1, CCAT1, *etc.* In our article, we not only analyzed the differential expression of lncRNAs in human HCC (upregulated as well as downregulated), but also summarized their specific mechanisms and pathways and gave an outlook in their potential as candidates in diagnosis and treatment of HCC. Another review<sup>[111]</sup> provided different lncRNAs compared to us, such as RP11-160H22.5, XLOC\_014172 and LOC149086. However, these lncRNAs were not embodied in NCBI database, and their mechanisms were unavailable, which brought us new research direction.

Although plenty of studies<sup>[28,112-115]</sup> cast light on the characteristics and mechanisms of different lncRNAs involved in HCC as we summarized above, till now, research on lncRNA still remains in its infancy and a large portion of lncRNAs surely remains to be further discovered. As systematic identification of lncRNAs and their well-understanding of mechanisms can pave the way for early diagnosis and therapeutics designing for HCC, there is still a long way to go in the research field of HCC-related lncRNAs.

Continuous researches are needed to verify the detailed function and mechanisms of revealed lncRNAs and to find innovative lncRNAs and sequentially, the predictive and diagnostic roles of lncRNAs in HCC can be validated. Thereby, diagnosis of HCC in an early stage and controlling its development and progression will become possible. Further clinical and ward trials are also in the urge, so that we can develop therapeutic roles of lncRNAs in HCC. In a word, future studies should aim

at investigating how can a discovered lncRNA be used to identify HCC patients and used as a guidance being applied in treatment to have optimal responses and to reduce the likelihood of relapse.

## REFERENCES

- 1 **Schmieder R**, Puehler F, Neuhaus R, Kissel M, Adjei AA, Miner JN, Mumberg D, Ziegelbauer K, Scholz A. Allosteric MEK1/2 inhibitor refametinib (BAY 86-9766) in combination with sorafenib exhibits antitumor activity in preclinical murine and rat models of hepatocellular carcinoma. *Neoplasia* 2013; **15**: 1161-1171 [PMID: 24204195 DOI: 10.1593/neo.13812]
- 2 **Su JC**, Tseng PH, Wu SH, Hsu CY, Tai WT, Li YS, Chen IT, Liu CY, Chen KF, Shiau CW. SC-2001 overcomes STAT3-mediated sorafenib resistance through RFX-1/SHP-1 activation in hepatocellular carcinoma. *Neoplasia* 2014; **16**: 595-605 [PMID: 25047655 DOI: 10.1016/j.neo.2014.06.005]
- 3 **Xia H**, Ooi LL, Hui KM. MicroRNA-216a/217-induced epithelial-mesenchymal transition targets PTEN and SMAD7 to promote drug resistance and recurrence of liver cancer. *Hepatology* 2013; **58**: 629-641 [PMID: 23471579 DOI: 10.1002/hep.26369]
- 4 **El-Serag HB**, Rudolph KL. Hepatocellular carcinoma: epidemiology and molecular carcinogenesis. *Gastroenterology* 2007; **132**: 2557-2576 [PMID: 17570226 DOI: 10.1053/j.gastro.2007.04.061]
- 5 **Sakurai T**, Kudo M, Umemura A, He G, Elsharkawy AM, Seki E, Karin M. p38 $\alpha$  inhibits liver fibrogenesis and consequent hepatocarcinogenesis by curtailing accumulation of reactive oxygen species. *Cancer Res* 2013; **73**: 215-224 [PMID: 23271722 DOI: 10.1158/0008-5472.can-12-1602]
- 6 **Zhang T**, Zhang J, Cui M, Liu F, You X, Du Y, Gao Y, Zhang S, Lu Z, Ye L, Zhang X. Hepatitis B virus X protein inhibits tumor suppressor miR-205 through inducing hypermethylation of miR-205 promoter to enhance carcinogenesis. *Neoplasia* 2013; **15**: 1282-1291 [PMID: 24339740 DOI: 10.1593/neo.131362]
- 7 **Qi LN**, Bai T, Chen ZS, Wu FX, Chen YY, De Xiang B, Peng T, Han ZG, Li LQ. The p53 mutation spectrum in hepatocellular carcinoma from Guangxi, China : role of chronic hepatitis B virus infection and aflatoxin B1 exposure. *Liver Int* 2015; **35**: 999-1009 [PMID: 24461059 DOI: 10.1111/liv.12460]
- 8 **Aravalli RN**, Cressman EN, Steer CJ. Cellular and molecular mechanisms of hepatocellular carcinoma: an update. *Arch Toxicol* 2013; **87**: 227-247 [PMID: 23007558 DOI: 10.1007/s00204-012-0931-2]
- 9 **Shin JW**, Chung YH. Molecular targeted therapy for hepatocellular carcinoma: current and future. *World J Gastroenterol* 2013; **19**: 6144-6155 [PMID: 24115810 DOI: 10.3748/wjg.v19.i37.6144]
- 10 **Cao C**, Sun J, Zhang D, Guo X, Xie L, Li X, Wu D, Liu L. The long intergenic noncoding RNA UFC1, a target of MicroRNA 34a, interacts with the mRNA stabilizing protein HuR to increase levels of  $\beta$ -catenin in HCC cells. *Gastroenterology* 2015; **148**: 415-426. e18 [PMID: 25449213 DOI: 10.1053/j.gastro.2014.10.012]
- 11 **Sherwood V**. WNT signaling: an emerging mediator of cancer cell metabolism? *Mol Cell Biol* 2015; **35**: 2-10 [PMID: 25348713 DOI: 10.1128/mcb.00992-14]
- 12 **Collins JF**. Long noncoding RNAs and hepatocellular carcinoma. *Gastroenterology* 2015; **148**: 291-294 [PMID: 25529811 DOI: 10.1053/j.gastro.2014.12.011]
- 13 **Huang B**, Zhang R. Regulatory non-coding RNAs: revolutionizing the RNA world. *Mol Biol Rep* 2014; **41**: 3915-3923 [PMID: 24549720 DOI: 10.1007/s11033-014-3259-6]
- 14 **Gehrau RC**, Mas VR, Villamil FG, Dumur CI, Mehta NK, Suh JL, Maluf DG. MicroRNA signature at the time of clinical HCV recurrence associates with aggressive fibrosis progression post-liver transplantation. *Am J Transplant* 2013; **13**: 729-737 [PMID: 23312020 DOI: 10.1111/ajt.12047]
- 15 **Dong P**, Yu F, Fan X, Lin Z, Chen Y, Li J. Inhibition of ATIR by shRNA prevents collagen synthesis in hepatic stellate cells. *Mol Cell Biochem* 2010; **344**: 195-202 [PMID: 20703514 DOI:

- 10.1007/s11010-010-0542-2]
- 16 **Gomes AQ**, Nolasco S, Soares H. Non-coding RNAs: multi-tasking molecules in the cell. *Int J Mol Sci* 2013; **14**: 16010-16039 [PMID: 23912238 DOI: 10.3390/ijms140816010]
  - 17 **Li X**, Wu J, Zheng J, Li Y, Yang T, Hu G, Dai J, Yang Q, Dai L, Jiang Y. Altered miRNA expression profiles and miR-1a associated with urethane-induced pulmonary carcinogenesis. *Toxicol Sci* 2013; **135**: 63-71 [PMID: 23761296 DOI: 10.1093/toxsci/kft131]
  - 18 **Zheng L**, Pu J, Qi T, Qi M, Li D, Xiang X, Huang K, Tong Q. miRNA-145 targets v-ets erythroblastosis virus E26 oncogene homolog 1 to suppress the invasion, metastasis, and angiogenesis of gastric cancer cells. *Mol Cancer Res* 2013; **11**: 182-193 [PMID: 23233482 DOI: 10.1158/1541-7786.mcr-12-0534]
  - 19 **Wang Y**, Zhang X, Li H, Yu J, Ren X. The role of miRNA-29 family in cancer. *Eur J Cell Biol* 2013; **92**: 123-128 [PMID: 23357522 DOI: 10.1016/j.ejcb.2012.11.004]
  - 20 **Li J**, Kong X, Zhang J, Luo Q, Li X, Fang L. MiRNA-26b inhibits proliferation by targeting PTGS2 in breast cancer. *Cancer Cell Int* 2013; **13**: 7 [PMID: 23374284 DOI: 10.1186/1475-2867-13-7]
  - 21 **He Y**, Meng XM, Huang C, Wu BM, Zhang L, Lv XW, Li J. Long noncoding RNAs: Novel insights into hepatocellular carcinoma. *Cancer Lett* 2014; **344**: 20-27 [PMID: 24183851 DOI: 10.1016/j.canlet.2013.10.021]
  - 22 **Qiu MT**, Hu JW, Yin R, Xu L. Long noncoding RNA: an emerging paradigm of cancer research. *Tumour Biol* 2013; **34**: 613-620 [PMID: 23359273 DOI: 10.1007/s13277-013-0658-6]
  - 23 **Cheetham SW**, Gruhl F, Mattick JS, Dinger ME. Long noncoding RNAs and the genetics of cancer. *Br J Cancer* 2013; **108**: 2419-2425 [PMID: 23660942 DOI: 10.1038/bjc.2013.233]
  - 24 **Huang JF**, Guo YJ, Zhao CX, Yuan SX, Wang Y, Tang GN, Zhou WP, Sun SH. Hepatitis B virus X protein (HBx)-related long noncoding RNA (lncRNA) down-regulated expression by HBx (Dreh) inhibits hepatocellular carcinoma metastasis by targeting the intermediate filament protein vimentin. *Hepatology* 2013; **57**: 1882-1892 [PMID: 23239537 DOI: 10.1002/hep.26195]
  - 25 **Pandey RR**, Mondal T, Mohammad F, Enroth S, Redrup L, Komorowski J, Nagano T, Mancini-Dinardo D, Kanduri C. Kcnq1o1 antisense noncoding RNA mediates lineage-specific transcriptional silencing through chromatin-level regulation. *Mol Cell* 2008; **32**: 232-246 [PMID: 18951091 DOI: 10.1016/j.molcel.2008.08.022]
  - 26 **Wang X**, Arai S, Song X, Reichart D, Du K, Pascual G, Tempst P, Rosenfeld MG, Glass CK, Kurokawa R. Induced ncRNAs allosterically modify RNA-binding proteins in cis to inhibit transcription. *Nature* 2008; **454**: 126-130 [PMID: 18509338 DOI: 10.1038/nature06992]
  - 27 **Wang KC**, Yang YW, Liu B, Sanyal A, Corces-Zimmerman R, Chen Y, Lajoie BR, Protacio A, Flynn RA, Gupta RA, Wysocka J, Lei M, Dekker J, Helms JA, Chang HY. A long noncoding RNA maintains active chromatin to coordinate homeotic gene expression. *Nature* 2011; **472**: 120-124 [PMID: 21423168 DOI: 10.1038/nature09819]
  - 28 **Wang Z**, Li X. The role of noncoding RNA in hepatocellular carcinoma. *Gland Surg* 2013; **2**: 25-29 [PMID: 25083452 DOI: 10.3978/j.issn.2227-684X.2013.02.07]
  - 29 **Shibata C**, Otsuka M, Kishikawa T, Ohno M, Yoshikawa T, Takata A, Koike K. Diagnostic and therapeutic application of noncoding RNAs for hepatocellular carcinoma. *World J Hepatol* 2015; **7**: 1-6 [PMID: 25624991 DOI: 10.4254/wjh.v7.i1.1]
  - 30 **Yan B**, Wang Z. Long noncoding RNA: its physiological and pathological roles. *DNA Cell Biol* 2012; **31** Suppl 1: S34-S41 [PMID: 22612272 DOI: 10.1089/dna.2011.1544]
  - 31 **Isin M**, Dalay N. LncRNAs and neoplasia. *Clin Chim Acta* 2015; **444**: 280-288 [PMID: 25748036 DOI: 10.1016/j.cca.2015.02.046]
  - 32 **Salmena L**, Poliseno L, Tay Y, Kats L, Pandolfi PP. A ceRNA hypothesis: the Rosetta Stone of a hidden RNA language? *Cell* 2011; **146**: 353-358 [PMID: 21802130 DOI: 10.1016/j.cell.2011.07.014]
  - 33 **Li X**, Wang Z. The role of noncoding RNA in thyroid cancer. *Gland Surg* 2012; **1**: 146-150 [PMID: 25083438 DOI: 10.3978/j.issn.2227-684X.2012.10.07]
  - 34 **Wang KC**, Chang HY. Molecular mechanisms of long noncoding RNAs. *Mol Cell* 2011; **43**: 904-914 [PMID: 21925379 DOI: 10.1016/j.molcel.2011.08.018]
  - 35 **Ponting CP**, Oliver PL, Reik W. Evolution and functions of long noncoding RNAs. *Cell* 2009; **136**: 629-641 [PMID: 19239885 DOI: 10.1016/j.cell.2009.02.006]
  - 36 **Ma L**, Bajic VB, Zhang Z. On the classification of long non-coding RNAs. *RNA Biol* 2013; **10**: 925-933 [PMID: 23696037 DOI: 10.4161/rna.24604]
  - 37 **Spizzo R**, Almeida MI, Colombatti A, Calin GA. Long non-coding RNAs and cancer: a new frontier of translational research? *Oncogene* 2012; **31**: 4577-4587 [PMID: 22266873 DOI: 10.1038/onc.2011.621]
  - 38 **Mercer TR**, Dinger ME, Mattick JS. Long non-coding RNAs: insights into functions. *Nat Rev Genet* 2009; **10**: 155-159 [PMID: 19188922 DOI: 10.1038/nrg2521]
  - 39 **Gupta RA**, Shah N, Wang KC, Kim J, Horlings HM, Wong DJ, Tsai MC, Hung T, Argani P, Rinn JL, Wang Y, Brzoska P, Kong B, Li R, West RB, van de Vijver MJ, Sukumar S, Chang HY. Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. *Nature* 2010; **464**: 1071-1076 [PMID: 20393566 DOI: 10.1038/nature08975]
  - 40 **Matouk IJ**, Abbasi I, Hochberg A, Galun E, Dweik H, Akkawi M. Highly upregulated in liver cancer noncoding RNA is overexpressed in hepatic colorectal metastasis. *Eur J Gastroenterol Hepatol* 2009; **21**: 688-692 [PMID: 19445043 DOI: 10.1097/MEG.0b013e328306a3a2]
  - 41 **Ji P**, Diederichs S, Wang W, Böing S, Metzger R, Schneider PM, Tidow N, Brandt B, Buerger H, Bulk E, Thomas M, Berdel WE, Serve H, Müller-Tidow C. MALAT-1, a novel noncoding RNA, and thymosin beta4 predict metastasis and survival in early-stage non-small cell lung cancer. *Oncogene* 2003; **22**: 8031-8041 [PMID: 12970751 DOI: 10.1038/sj.onc.1206928]
  - 42 **Huang JL**, Zheng L, Hu YW, Wang Q. Characteristics of long non-coding RNA and its relation to hepatocellular carcinoma. *Carcinogenesis* 2014; **35**: 507-514 [PMID: 24296588 DOI: 10.1093/carcin/bgt405]
  - 43 **Ma L**, Chua MS, Andrisani O, So S. Epigenetics in hepatocellular carcinoma: an update and future therapy perspectives. *World J Gastroenterol* 2014; **20**: 333-345 [PMID: 24574704 DOI: 10.3748/wjg.v20.i2.333]
  - 44 **Zhao Y**, Guo Q, Chen J, Hu J, Wang S, Sun Y. Role of long non-coding RNA HULC in cell proliferation, apoptosis and tumor metastasis of gastric cancer: a clinical and in vitro investigation. *Oncol Rep* 2014; **31**: 358-364 [PMID: 24247585 DOI: 10.3892/or.2013.2850]
  - 45 **Xie H**, Ma H, Zhou D. Plasma HULC as a promising novel biomarker for the detection of hepatocellular carcinoma. *Biomed Res Int* 2013; **2013**: 1361106 [PMID: 23762823 DOI: 10.1155/2013/1361106]
  - 46 **Hämmerle M**, Gutschner T, Uckelmann H, Ozgur S, Fiskin E, Gross M, Skawran B, Geffers R, Longerich T, Breuhahn K, Schirmacher P, Stoecklin G, Diederichs S. Posttranscriptional destabilization of the liver-specific long noncoding RNA HULC by the IGF2 mRNA-binding protein 1 (IGF2BP1). *Hepatology* 2013; **58**: 1703-1712 [PMID: 23728852 DOI: 10.1002/hep.26537]
  - 47 **Du Y**, Kong G, You X, Zhang S, Zhang T, Gao Y, Ye L, Zhang X. Elevation of highly up-regulated in liver cancer (HULC) by hepatitis B virus X protein promotes hepatoma cell proliferation via down-regulating p18. *J Biol Chem* 2012; **287**: 26302-26311 [PMID: 22685290 DOI: 10.1074/jbc.M112.342113]
  - 48 **Wang J**, Liu X, Wu H, Ni P, Gu Z, Qiao Y, Chen N, Sun F, Fan Q. CREB up-regulates long non-coding RNA, HULC expression through interaction with microRNA-372 in liver cancer. *Nucleic Acids Res* 2010; **38**: 5366-5383 [PMID: 20423907 DOI: 10.1093/nar/gkq285]
  - 49 **Yang Z**, Lu Y, Xu Q, Tang B, Park CK, Chen X. HULC and H19 Played Different Roles in Overall and Disease-Free Survival from Hepatocellular Carcinoma after Curative Hepatectomy: A Preliminary Analysis from Gene Expression Omnibus. *Dis Markers* 2015; **2015**: 191029 [PMID: 26136615 DOI: 10.1155/2015/191029]

- 50 **Zhang L**, Yang F, Yuan JH, Yuan SX, Zhou WP, Huo XS, Xu D, Bi HS, Wang F, Sun SH. Epigenetic activation of the MiR-200 family contributes to H19-mediated metastasis suppression in hepatocellular carcinoma. *Carcinogenesis* 2013; **34**: 577-586 [PMID: 23222811 DOI: 10.1093/carcin/bgs381]
- 51 **Shi X**, Sun M, Liu H, Yao Y, Song Y. Long non-coding RNAs: a new frontier in the study of human diseases. *Cancer Lett* 2013; **339**: 159-166 [PMID: 23791884 DOI: 10.1016/j.canlet.2013.06.013]
- 52 **Lv J**, Yu YQ, Li SQ, Luo L, Wang Q. Aflatoxin B1 promotes cell growth and invasion in hepatocellular carcinoma HepG2 cells through H19 and E2F1. *Asian Pac J Cancer Prev* 2014; **15**: 2565-2570 [PMID: 24761865 DOI: 10.7314/APJCP.2014.15.6.2565]
- 53 **Tsang WP**, Kwok TT. Riboregulator H19 induction of MDR1-associated drug resistance in human hepatocellular carcinoma cells. *Oncogene* 2007; **26**: 4877-4881 [PMID: 17297456 DOI: 10.1038/sj.onc.1210266]
- 54 **Iizuka N**, Oka M, Yamada-Okabe H, Mori N, Tamesa T, Okada T, Takemoto N, Tangoku A, Hamada K, Nakayama H, Miyamoto T, Uchimura S, Hamamoto Y. Comparison of gene expression profiles between hepatitis B virus- and hepatitis C virus-infected hepatocellular carcinoma by oligonucleotide microarray data on the basis of a supervised learning method. *Cancer Res* 2002; **62**: 3939-3944 [PMID: 12124323]
- 55 **Ariel I**, Miao HQ, Ji XR, Schneider T, Roll D, de Groot N, Hochberg A, Ayes S. Imprinted H19 oncofetal RNA is a candidate tumour marker for hepatocellular carcinoma. *Mol Pathol* 1998; **51**: 21-25 [PMID: 9624415 DOI: 10.1136/mp.51.1.21]
- 56 **Gabory A**, Jammes H, Dandolo L. The H19 locus: role of an imprinted non-coding RNA in growth and development. *Bioessays* 2010; **32**: 473-480 [PMID: 20486133 DOI: 10.1002/bies.200900170]
- 57 **Braconi C**, Valeri N, Kogure T, Gasparini P, Huang N, Nuovo GJ, Terracciano L, Croce CM, Patel T. Expression and functional role of a transcribed noncoding RNA with an ultraconserved element in hepatocellular carcinoma. *Proc Natl Acad Sci USA* 2011; **108**: 786-791 [PMID: 21187392 DOI: 10.1073/pnas.1011098108]
- 58 **Lai MC**, Yang Z, Zhou L, Zhu QQ, Xie HY, Zhang F, Wu LM, Chen LM, Zheng SS. Long non-coding RNA MALAT-1 overexpression predicts tumor recurrence of hepatocellular carcinoma after liver transplantation. *Med Oncol* 2012; **29**: 1810-1816 [PMID: 21678027 DOI: 10.1007/s12032-011-0004-z]
- 59 **Lin R**, Maeda S, Liu C, Karin M, Edgington TS. A large noncoding RNA is a marker for murine hepatocellular carcinomas and a spectrum of human carcinomas. *Oncogene* 2007; **26**: 851-858 [PMID: 16878148 DOI: 10.1038/sj.onc.1209846]
- 60 **Geng YJ**, Xie SL, Li Q, Ma J, Wang GY. Large intervening non-coding RNA HOTAIR is associated with hepatocellular carcinoma progression. *J Int Med Res* 2011; **39**: 2119-2128 [PMID: 22289527 DOI: 10.1177/147323001103900608]
- 61 **Yang Z**, Zhou L, Wu LM, Lai MC, Xie HY, Zhang F, Zheng SS. Overexpression of long non-coding RNA HOTAIR predicts tumor recurrence in hepatocellular carcinoma patients following liver transplantation. *Ann Surg Oncol* 2011; **18**: 1243-1250 [PMID: 21327457 DOI: 10.1245/s10434-011-1581-y]
- 62 **Ishibashi M**, Kogo R, Shibata K, Sawada G, Takahashi Y, Kurashige J, Akiyoshi S, Sasaki S, Iwaya T, Sudo T, Sugimachi K, Mimori K, Wakabayashi G, Mori M. Clinical significance of the expression of long non-coding RNA HOTAIR in primary hepatocellular carcinoma. *Oncol Rep* 2013; **29**: 946-950 [PMID: 23292722 DOI: 10.3892/or.2012.2219]
- 63 **Ding C**, Cheng S, Yang Z, Lv Z, Xiao H, Du C, Peng C, Xie H, Zhou L, Wu J, Zheng S. Long non-coding RNA HOTAIR promotes cell migration and invasion via down-regulation of RNA binding motif protein 38 in hepatocellular carcinoma cells. *Int J Mol Sci* 2014; **15**: 4060-4076 [PMID: 24663081 DOI: 10.3390/ijms15034060]
- 64 **Cai B**, Song XQ, Cai JP, Zhang S. HOTAIR: a cancer-related long non-coding RNA. *Neoplasma* 2014; **61**: 379-391 [PMID: 25027739 DOI: 10.4149/neo\_2014\_075]
- 65 **Quagliata L**, Matter MS, Piscuoglio S, Arabi L, Ruiz C, Procino A, Kovac M, Moretti F, Makowska Z, Boldanova T, Andersen JB, Hämmerle M, Tornillo L, Heim MH, Diederichs S, Cillo C, Terracciano LM. Long noncoding RNA HOTTIP/HOXA13 expression is associated with disease progression and predicts outcome in hepatocellular carcinoma patients. *Hepatology* 2014; **59**: 911-923 [PMID: 24114970 DOI: 10.1002/hep.26740]
- 66 **Cillo C**, Schiavo G, Cantile M, Bihl MP, Sorrentino P, Carafa V, D'Armiento M, Roncalli M, Sansano S, Vecchione R, Tornillo L, Mori L, De Libero G, Zucman-Rossi J, Terracciano L. The HOX gene network in hepatocellular carcinoma. *Int J Cancer* 2011; **129**: 2577-2587 [PMID: 21626505 DOI: 10.1002/ijc.25941]
- 67 **Tsang FH**, Au SL, Wei L, Fan DN, Lee JM, Wong CC, Ng IO, Wong CM. Long non-coding RNA HOTTIP is frequently up-regulated in hepatocellular carcinoma and is targeted by tumour suppressive miR-125b. *Liver Int* 2015; **35**: 1597-1606 [PMID: 25424744 DOI: 10.1111/liv.12746]
- 68 **Yang F**, Zhang L, Huo XS, Yuan JH, Xu D, Yuan SX, Zhu N, Zhou WP, Yang GS, Wang YZ, Shang JL, Gao CF, Zhang FR, Wang F, Sun SH. Long noncoding RNA high expression in hepatocellular carcinoma facilitates tumor growth through enhancer of zeste homolog 2 in humans. *Hepatology* 2011; **54**: 1679-1689 [PMID: 21769904 DOI: 10.1002/hep.24563]
- 69 **Chen B**, Yu M, Chang Q, Lu Y, Thakur C, Ma D, Yi Z, Chen F. Mdig de-represses H19 large intergenic non-coding RNA (lincRNA) by down-regulating H3K9me3 and heterochromatin. *Oncotarget* 2013; **4**: 1427-1437 [PMID: 23965803]
- 70 **Ding C**, Yang Z, Lv Z, DU C, Xiao H, Peng C, Cheng S, Xie H, Zhou L, Wu J, Zheng S. Long non-coding RNA PVT1 is associated with tumor progression and predicts recurrence in hepatocellular carcinoma patients. *Oncol Lett* 2015; **9**: 955-963 [PMID: 25624916 DOI: 10.3892/ol.2014.2730]
- 71 **Wang F**, Yuan JH, Wang SB, Yang F, Yuan SX, Ye C, Yang N, Zhou WP, Li WL, Li W, Sun SH. Oncofetal long noncoding RNA PVT1 promotes proliferation and stem cell-like property of hepatocellular carcinoma cells by stabilizing NOP2. *Hepatology* 2014; **60**: 1278-1290 [PMID: 25043274 DOI: 10.1002/hep.27239]
- 72 **Tang J**, Zhuo H, Zhang X, Jiang R, Ji J, Deng L, Qian X, Zhang F, Sun B. A novel biomarker Linc00974 interacting with KRT19 promotes proliferation and metastasis in hepatocellular carcinoma. *Cell Death Dis* 2014; **5**: e1549 [PMID: 25476897 DOI: 10.1038/cddis.2014.518]
- 73 **Liu H**, Gong M, French BA, Li J, Tillman B, French SW. Mallory-Denk Body (MDB) formation modulates Ufmylation expression epigenetically in alcoholic hepatitis (AH) and non-alcoholic steatohepatitis (NASH). *Exp Mol Pathol* 2014; **97**: 477-483 [PMID: 25290169 DOI: 10.1016/j.yexmp.2014.10.001]
- 74 **Yuan SX**, Tao QF, Wang J, Yang F, Liu L, Wang LL, Zhang J, Yang Y, Liu H, Wang F, Sun SH, Zhou WP. Antisense long non-coding RNA PCNA-AS1 promotes tumor growth by regulating proliferating cell nuclear antigen in hepatocellular carcinoma. *Cancer Lett* 2014; **349**: 87-94 [PMID: 24704293 DOI: 10.1016/j.canlet.2014.03.029]
- 75 **Wang F**, Ying HQ, He BS, Pan YQ, Deng QW, Sun HL, Chen J, Liu X, Wang SK. Upregulated lincRNA-UCA1 contributes to progression of hepatocellular carcinoma through inhibition of miR-216b and activation of FGFR1/ERK signaling pathway. *Oncotarget* 2015; **6**: 7899-7917 [PMID: 25760077 DOI: 10.18632/oncotarget.3219]
- 76 **Deng L**, Yang SB, Xu FF, Zhang JH. Long noncoding RNA CCAT1 promotes hepatocellular carcinoma progression by functioning as let-7 sponge. *J Exp Clin Cancer Res* 2015; **34**: 18 [PMID: 25884472 DOI: 10.1186/s13046-015-0136-7]
- 77 **Zhu H**, Zhou X, Chang H, Li H, Liu F, Ma C, Lu J. CCAT1 promotes hepatocellular carcinoma cell proliferation and invasion. *Int J Clin Exp Pathol* 2015; **8**: 5427-5434 [PMID: 26191246]
- 78 **Zhu HQ**, Zhou X, Chang H, Li HG, Liu FF, Ma CQ, Lu J. Aberrant Expression of CCAT1 Regulated by c-Myc Predicts the Prognosis of Hepatocellular Carcinoma. *Asian Pac J Cancer Prev* 2015; **16**: 5181-5185 [PMID: 26225650]

- 79 **Sun T**, Wong N. Transforming growth factor- $\beta$ -induced long noncoding RNA promotes liver cancer metastasis via RNA-RNA crosstalk. *Hepatology* 2015; **61**: 722-724 [PMID: 25380484 DOI: 10.1002/hep.27599]
- 80 **Yuan JH**, Yang F, Wang F, Ma JZ, Guo YJ, Tao QF, Liu F, Pan W, Wang TT, Zhou CC, Wang SB, Wang YZ, Yang Y, Yang N, Zhou WP, Yang GS, Sun SH. A long noncoding RNA activated by TGF- $\beta$  promotes the invasion-metastasis cascade in hepatocellular carcinoma. *Cancer Cell* 2014; **25**: 666-681 [PMID: 24768205 DOI: 10.1016/j.ccr.2014.03.010]
- 81 **Xu WH**, Zhang JB, Dang Z, Li X, Zhou T, Liu J, Wang DS, Song WJ, Dou KF. Long non-coding RNA URHC regulates cell proliferation and apoptosis via ZAK through the ERK/MAPK signaling pathway in hepatocellular carcinoma. *Int J Biol Sci* 2014; **10**: 664-676 [PMID: 25013376 DOI: 10.7150/ijbs.8232]
- 82 **Anwar SL**, Krech T, Hasemeier B, Schipper E, Schweitzer N, Vogel A, Kreipe H, Lehmann U. Loss of imprinting and allelic switching at the DLK1-MEG3 locus in human hepatocellular carcinoma. *PLoS One* 2012; **7**: e49462 [PMID: 23145177 DOI: 10.1371/journal.pone.0049462]
- 83 **Braconi C**, Kogure T, Valeri N, Huang N, Nuovo G, Costinean S, Negrini M, Miotto E, Croce CM, Patel T. microRNA-29 can regulate expression of the long non-coding RNA gene MEG3 in hepatocellular cancer. *Oncogene* 2011; **30**: 4750-4756 [PMID: 21625215 DOI: 10.1038/onc.2011.193]
- 84 **Huang J**, Zhang X, Zhang M, Zhu JD, Zhang YL, Lin Y, Wang KS, Qi XF, Zhang Q, Liu GZ, Yu J, Cui Y, Yang PY, Wang ZQ, Han ZG. Up-regulation of DLK1 as an imprinted gene could contribute to human hepatocellular carcinoma. *Carcinogenesis* 2007; **28**: 1094-1103 [PMID: 17114643 DOI: 10.1093/carcin/bgl215]
- 85 **Zhang X**, Zhou Y, Mehta KR, Danila DC, Scolavino S, Johnson SR, Klibanski A. A pituitary-derived MEG3 isoform functions as a growth suppressor in tumor cells. *J Clin Endocrinol Metab* 2003; **88**: 5119-5126 [PMID: 14602737 DOI: 10.1210/jc.2003-030222]
- 86 **Zhou Y**, Zhong Y, Wang Y, Zhang X, Batista DL, Gejman R, Ansell PJ, Zhao J, Weng C, Klibanski A. Activation of p53 by MEG3 non-coding RNA. *J Biol Chem* 2007; **282**: 24731-24742 [PMID: 17569660 DOI: 10.1074/jbc.M702029200]
- 87 **Yang F**, Huo XS, Yuan SX, Zhang L, Zhou WP, Wang F, Sun SH. Repression of the long noncoding RNA-LET by histone deacetylase 3 contributes to hypoxia-mediated metastasis. *Mol Cell* 2013; **49**: 1083-1096 [PMID: 23395002 DOI: 10.1016/j.molcel.2013.01.010]
- 88 **Chen CL**, Tseng YW, Wu JC, Chen GY, Lin KC, Hwang SM, Hu YC. Suppression of hepatocellular carcinoma by baculovirus-mediated expression of long non-coding RNA PTENP1 and MicroRNA regulation. *Biomaterials* 2015; **44**: 71-81 [PMID: 25617127 DOI: 10.1016/j.biomaterials.2014.12.023]
- 89 **Zheng H**, Yang S, Yang Y, Yuan SX, Wu FQ, Wang LL, Yan HL, Sun SH, Zhou WP. Epigenetically silenced long noncoding-SRHC promotes proliferation of hepatocellular carcinoma. *J Cancer Res Clin Oncol* 2015; **141**: 1195-1203 [PMID: 25512078 DOI: 10.1007/s00432-014-1871-4]
- 90 **Yu W**, Qiao Y, Tang X, Ma L, Wang Y, Zhang X, Weng W, Pan Q, Yu Y, Sun F, Wang J. Tumor suppressor long non-coding RNA, MT1DP is negatively regulated by YAP and Runx2 to inhibit FoxA1 in liver cancer cells. *Cell Signal* 2014; **26**: 2961-2968 [PMID: 25261601 DOI: 10.1016/j.cellsig.2014.09.011]
- 91 **Cui M**, Zheng M, Sun B, Wang Y, Ye L, Zhang X. A long noncoding RNA perturbs the circadian rhythm of hepatoma cells to facilitate hepatocarcinogenesis. *Neoplasia* 2015; **17**: 79-88 [PMID: 25622901 DOI: 10.1016/j.neo.2014.11.004]
- 92 **Liu Y**, Pan S, Liu L, Zhai X, Liu J, Wen J, Zhang Y, Chen J, Shen H, Hu Z. A genetic variant in long non-coding RNA HULC contributes to risk of HBV-related hepatocellular carcinoma in a Chinese population. *PLoS One* 2012; **7**: e35145 [PMID: 22493738 DOI: 10.1371/journal.pone.0035145]
- 93 **Wen J**, Liu Y, Liu J, Liu L, Song C, Han J, Zhu L, Wang C, Chen J, Zhai X, Shen H, Hu Z. Expression quantitative trait loci in long non-coding RNA ZNRD1-AS1 influence both HBV infection and hepatocellular carcinoma development. *Mol Carcinog* 2015; **54**: 1275-1282 [PMID: 25110835 DOI: 10.1002/mc.22200]
- 94 **Stranger BE**, Nica AC, Forrest MS, Dimas A, Bird CP, Beazley C, Ingle CE, Dunning M, Flicek P, Koller D, Montgomery S, Tavaré S, Deloukas P, Dermitzakis ET. Population genomics of human gene expression. *Nat Genet* 2007; **39**: 1217-1224 [PMID: 17873874 DOI: 10.1038/ng2142]
- 95 **Veyrieras JB**, Kudaravalli S, Kim SY, Dermitzakis ET, Gilad Y, Stephens M, Pritchard JK. High-resolution mapping of expression-QTLs yields insight into human gene regulation. *PLoS Genet* 2008; **4**: e1000214 [PMID: 18846210 DOI: 10.1371/journal.pgen.1000214]
- 96 **Guo W**, Zhao YP, Jiang YG, Wang RW, Hong L, Fan DM. ZNRD1 might mediate UV irradiation related DNA damage and repair in human esophageal cancer cells by regulation of ERCC1. *Dis Esophagus* 2008; **21**: 730-736 [PMID: 18564169 DOI: 10.1111/j.1442-2050.2008.00846.x]
- 97 **Hong L**, Han Y, Li S, Yang J, Gong T, Li J, Zheng J, Zhang H, Zhao Q, Wu K, Fan D. Role of ZNRD1 (zinc ribbon domain-containing 1) in angiogenesis of leukaemia cells. *Cell Biol Int* 2011; **35**: 321-324 [PMID: 21080911 DOI: 10.1042/cbi20100506]
- 98 **Hong L**, Han Y, Shi R, Shao X, Sun L, Zhang Y, Huang D, Chen Z, Zhang G, Liang J, Hu S, Fan D. ZNRD1 gene suppresses cell proliferation through cell cycle arrest in G1 phase. *Cancer Biol Ther* 2005; **4**: 60-64 [PMID: 15662122]
- 99 **Zhuo H**, Tang J, Lin Z, Jiang R, Zhang X, Ji J, Wang P, Sun B. The aberrant expression of MEG3 regulated by UHRF1 predicts the prognosis of hepatocellular carcinoma. *Mol Carcinog* 2015; Epub ahead of print [PMID: 25641194 DOI: 10.1002/mc.22270]
- 100 **Lv J**, Ma L, Chen XL, Huang XH, Wang Q. Downregulation of LncRNAH19 and MiR-675 promotes migration and invasion of human hepatocellular carcinoma cells through AKT/GSK-3 $\beta$ /Cdc25A signaling pathway. *J Huazhong Univ Sci Technolog Med Sci* 2014; **34**: 363-369 [PMID: 24939300 DOI: 10.1007/s11596-014-1284-2]
- 101 **Vernucci M**, Cerrato F, Besnard N, Casola S, Pedone PV, Brunni CB, Riccio A. The H19 endodermal enhancer is required for Igf2 activation and tumor formation in experimental liver carcinogenesis. *Oncogene* 2000; **19**: 6376-6385 [PMID: 11175353 DOI: 10.1038/sj.onc.1204024]
- 102 **Wang J**, Wang H, Zhang Y, Zhen N, Zhang L, Qiao Y, Weng W, Liu X, Ma L, Xiao W, Yu W, Chu Q, Pan Q, Sun F. Mutual inhibition between YAP and SRSF1 maintains long non-coding RNA, Malat1-induced tumorigenesis in liver cancer. *Cell Signal* 2014; **26**: 1048-1059 [PMID: 24468535]
- 103 **Tripathi V**, Ellis JD, Shen Z, Song DY, Pan Q, Watt AT, Freier SM, Bennett CF, Sharma A, Bubulya PA, Blencowe BJ, Prasad SM, Prasad KV. The nuclear-retained noncoding RNA MALAT1 regulates alternative splicing by modulating SR splicing factor phosphorylation. *Mol Cell* 2010; **39**: 925-938 [PMID: 20797886 DOI: 10.1016/j.molcel.2010.08.011]
- 104 **Bernard D**, Prasad KV, Tripathi V, Colasse S, Nakamura T, Xuan Z, Zhang MQ, Sedel F, Jourden L, Couplier F, Triller A, Spector DL, Bessis A. A long nuclear-retained non-coding RNA regulates synaptogenesis by modulating gene expression. *EMBO J* 2010; **29**: 3082-3093 [PMID: 20729808 DOI: 10.1038/emboj.2010.199]
- 105 **Rinn JL**, Kertesz M, Wang JK, Squazzo SL, Xu X, Bruggmann SA, Goodnough LH, Helms JA, Farnham PJ, Segal E, Chang HY. Functional demarcation of active and silent chromatin domains in human HOX loci by noncoding RNAs. *Cell* 2007; **129**: 1311-1323 [PMID: 17604720 DOI: 10.1016/j.cell.2007.05.022]
- 106 **Tsai MC**, Manor O, Wan Y, Mosammamaparast N, Wang JK, Lan F, Shi Y, Segal E, Chang HY. Long noncoding RNA as modular scaffold of histone modification complexes. *Science* 2010; **329**: 689-693 [PMID: 20616235 DOI: 10.1126/science.1192002]
- 107 **Lujambio A**, Akkari L, Simon J, Grace D, Tschaharganeh DF, Bolden JE, Zhao Z, Thapar V, Joyce JA, Krizhanovsky V, Lowe SW. Non-cell-autonomous tumor suppression by p53. *Cell* 2013; **153**: 449-460 [PMID: 23562644 DOI: 10.1016/j.cell.2013.03.020]

- 108 **Mellert H**, Espinosa JM. Tumor suppression by p53: is apoptosis important or not? *Cell Rep* 2013; **3**: 1335-1336 [PMID: 23726020 DOI: 10.1016/j.celrep.2013.05.011]
- 109 **Huarte M**, Guttman M, Feldser D, Garber M, Koziol MJ, Kenzelmann-Broz D, Khalil AM, Zuk O, Amit I, Rabani M, Attardi LD, Regev A, Lander ES, Jacks T, Rinn JL. A large intergenic noncoding RNA induced by p53 mediates global gene repression in the p53 response. *Cell* 2010; **142**: 409-419 [PMID: 20673990 DOI: 10.1016/j.cell.2010.06.040]
- 110 **Yang JJ**, Lee YJ, Hung HH, Tseng WP, Tu CC, Lee H, Wu WJ. ZAK inhibits human lung cancer cell growth via ERK and JNK activation in an AP-1-dependent manner. *Cancer Sci* 2010; **101**: 1374-1381 [PMID: 20331627 DOI: 10.1111/j.1349-7006.2010.01537.x]
- 111 **Yang X**, Xie X, Xiao YF, Xie R, Hu CJ, Tang B, Li BS, Yang SM. The emergence of long non-coding RNAs in the tumorigenesis of hepatocellular carcinoma. *Cancer Lett* 2015; **360**: 119-124 [PMID: 25721084 DOI: 10.1016/j.canlet.2015.02.035]
- 112 **Amicone L**, Citarella F, Cicchini C. Epigenetic regulation in hepatocellular carcinoma requires long noncoding RNAs. *Biomed Res Int* 2015; **2015**: 473942 [PMID: 25861629 DOI: 10.1155/2015/473942]
- 113 **Sun J**, Bie B, Zhang S, Yang J, Li Z. Long non-coding RNAs: critical players in hepatocellular carcinoma. *Int J Mol Sci* 2014; **15**: 20434-20448 [PMID: 25387074 DOI: 10.3390/ijms151120434]
- 114 **Tang J**, Jiang R, Deng L, Zhang X, Wang K, Sun B. Circulation long non-coding RNAs act as biomarkers for predicting tumorigenesis and metastasis in hepatocellular carcinoma. *Oncotarget* 2015; **6**: 4505-4515 [PMID: 25714016 DOI: 10.18632/oncotarget.2934]
- 115 **Li CH**, Chen Y. Targeting long non-coding RNAs in cancers: progress and prospects. *Int J Biochem Cell Biol* 2013; **45**: 1895-1910 [PMID: 23748105 DOI: 10.1016/j.biocel.2013.05.030]

**P- Reviewer:** Dalay N, Slomiany BL, Tomizawa M  
**S- Editor:** Ji FF **L- Editor:** A **E- Editor:** Liu SQ



## 2015 Advances in Hepatitis C Virus

**Hepatitis C genotype 4: The past, present, and future**

Tawhida Y Abdel-Ghaffar, Mostafa M Sira, Suzan El Naghi

Tawhida Y Abdel-Ghaffar, Pediatric Department, Ain Shams University, Cairo 11566, Egypt

Tawhida Y Abdel-Ghaffar, Suzan El Naghi, Yassin Abdel Ghaffar Charity Center for Liver Disease and Research, Cairo 11566, Egypt

Mostafa M Sira, Pediatric Hepatology Department, National Liver Institute, Menofiya University, Shebin El-koom, Menofiya 32511, Egypt

Suzan El Naghi, Pediatric Department, National Hepatology and Tropical Medicine Research Institute, Cairo 11441, Egypt

**Author contributions:** Abdel-Ghaffar TY, Sira MM and El Naghi S were involved in the study concept and design and shared in preparation; El Naghi S wrote the article; Abdel-Ghaffar TY and Sira MM reviewed the article.

**Conflict-of-interest statement:** The authors declare no conflict of interest for this article.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to:** Suzan El Naghi, MD, Pediatric Department, National Hepatology and Tropical Medicine Research Institute, 10 - Kasr El Aini Street, Cairo 11441, Egypt. [suzan\\_elnaghi@yahoo.com](mailto:suzan_elnaghi@yahoo.com)  
Telephone: +20-2-24197255  
Fax: +20-2-23504240

Received: April 22, 2015

Peer-review started: April 24, 2015

First decision: July 21, 2015

Revised: September 24, 2015

Accepted: November 24, 2015

Article in press: November 25, 2015

Published online: December 8, 2015

**Abstract**

Hepatitis C virus (HCV) genotype (GT) 4 represents 12%-15% (15-18 million) of total global HCV infection. It is prevalent in Northern and Equatorial Africa and the Middle East, and is also present in some countries in Europe. GT-4 (and subtype 4a in particular) dominates the HCV epidemic in Egypt. In underdeveloped countries, risk factors associated with HCV infection may be due to unsafe medical practices or other factors such as familial transmission, mother's HCV status, or illiteracy. HCV prevention and control programs should include health education, increased community awareness towards the disease, controlling infection distribution in health-care centers, proper sterilization of medical and dental instruments, and ensuring safe supply of blood and blood-products. Response rates to a 48-wk combined pegylated-interferon (PEG-IFN) and ribavirin (RBV) treatment range from 40%-69%, and HCV-GT-4 has been considered better than GT-1 but worse than GT-2 and GT-3 in treatment with PEG-IFN/RBV. However, with the introduction of the HCV-GT-1 effective protease inhibitors boceprevir and telaprevir in 2011, HCV-GT-4 became the "most difficult (GT) to treat". Recently, the direct-acting antivirals (DAAs) with pan-genotypic activities simeprevir, sofosbuvir, and daclatasvir have been recommended in triple regimens with PEG-IFN/RBV for the treatment of HCV-GT-4. An IFN-free regimen will be available for treatment of all genotypes of HCV in the near future. To date, several DAAs have been developed and are currently being evaluated in various combinations in clinical trials. As new regimens and new agents are being approved by the Food and Drug Administration, we can expect the guidelines for HCV treatment to be changed. The availability of shorter, simpler, and more tolerable treatment regimens can reduce the morbidity and mortality associated with HCV infection. With such a large number of therapeutic agents available, we can end up with a range of choices that we can select from to treat patients.

**Key words:** Hepatitis C virus; Genotypes; Transmission; Pegylated-interferon; Ribavirin; Direct acting antivirals;

## Hepatitis C virus vaccine

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Hepatitis C virus (HCV) genotype (GT) 4 represents 12%-15% of total global HCV infection. It is higher in limited resource countries. Response rates to a 48-wk peg-interferon/ribavirin combination ranges from 40%-69% for HCV-GT-4. Direct-acting antivirals may significantly improve treatment outcomes in HCV-GT-4, but use of these agents in countries endemic for HCV-GT-4 is currently precluded by the very high costs. A new hepatitis C vaccine from GlaxoSmithKline has shown promise in early clinical tests, prompting strong and broad immune responses. Another Egyptian clinical trial in the field of HCV vaccination: Clinical Trials phases I and II, started on March 2011. ClinicalTrials.gov Identifier NCT01718834.

Abdel-Ghaffar TY, Sira MM, El Naghi S. Hepatitis C genotype 4: The past, present, and future. *World J Hepatol* 2015; 7(28): 2792-2810 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i28/2792.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i28.2792>

## INTRODUCTION

Harvey J Alter is an American Virologist, Medical Researcher, and Physician. In mid-1970s, he was the first person to prove a new type of hepatitis virus, initially called non-A, non-B hepatitis (NANBH). During the following years, medical researchers Michael Houghton, George Kuo, and Qui-Lim Choo from Chiron Corporation (an American multinational biotechnology firm) and Dr. Daniel W Bradley from the Centers for Disease Control and Prevention of America (CDC) collaborated in identifying the virus: Using the molecular cloning process to identify an unknown organism, and confirming that the organism is a virus after finding an NANBH specimen in the organism in 1988. Two articles were published on it, and the name was changed from NANBH to hepatitis C virus (HCV) in April 1989. Later on, HCV was shown to be the principal cause of parenterally transmitted NANBH worldwide<sup>[1]</sup>. After the discovery of this virus, a flurry of international studies was conducted to document its distribution and prevalence in humans<sup>[2]</sup>. It is now well established that HCV is a global health challenge, with an estimated 2%-3% of the global population having chronic HCV infection<sup>[3]</sup>. Estimates over the last 15 years show HCV affection to have increased to 2.8%, which means > 185 million infections worldwide<sup>[4]</sup>.

## EPIDEMIOLOGY

HCV is a small single-stranded ribonucleic acid (RNA) of positive polarity, and is an enveloped virus belonging to

the *Hepacivirus* genus within the *Flaviviridae* family<sup>[5]</sup>. It consists of approximately 9600 nucleotides in length, which encode three structural proteins (core, E1, and E2), the ion channel protein p7, and six nonstructural (NS) proteins (NS2, NS3, NS4A, NS4B, NS5A, and NS5B)<sup>[6]</sup>. Because each protein is involved in HCV entry, infection, replication, or maturation, they are potential antiviral targets. Hepatitis C virus replication takes place entirely within the cytoplasm, therefore it does not establish latency making it easier to cure<sup>[7]</sup>.

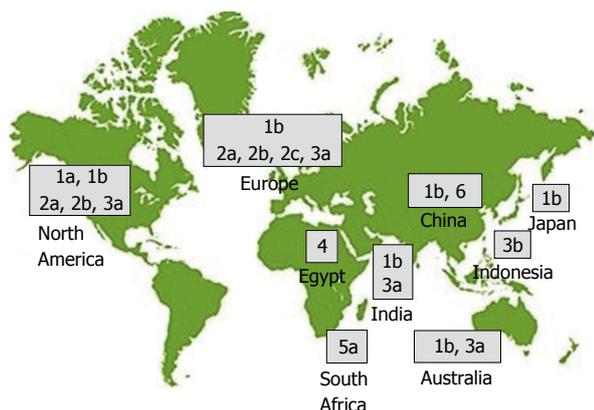
All RNA viruses show a high degree of genome-sequence heterogeneity. HCV RNA is characterized by three tiers of variability: Genotype (GT), subtype, and quasispecies, *i.e.*, the sequence variation within a single patient. The most recent classification includes seven GTs (numbered 1-7), 67 confirmed and 20 provisional subtypes, which differ in sequence by approximately 30% to 35%, and approximately 20% to 25%, respectively. All GTs except 5 and 7 are subdivided into numerous subtypes (1a, 1b, 1c, 2a, 2b, *etc.*)<sup>[8]</sup>. There is a clear geographic pattern in the distribution of HCV genetic diversity (Figure 1). Highly divergent "endemic" strains that belong to the same GT are typically found in a restricted geographic area, indicating the presence of the GT in that location for hundreds or even thousands of years<sup>[9]</sup>. GT-1 dominates worldwide (83.4 million cases, 46.2% of all HCV cases), approximately one-third of which are in East Asia. GT-3 comes second (54.3 million, 30.1%). GT-2, GT-4, and GT-6 compromise a total 22.8% of all cases, and GT-5 comprises the remaining < 1%. Subtypes, specifically 1a, 1b, 2a, and 3a, are distributed worldwide, and are especially dominant in high income countries<sup>[10]</sup>. Within the United States, 75% of isolates are HCV GT-1a or GT-1b and the remainder are generally GT-2 or GT-3<sup>[11]</sup>. In Europe, HCV-GT-1a and GT-1b are the commonest subtypes. GT-2 dominates in West Africa, and GT-3 dominates in South Asia and parts of Scandinavia. While GT-1 and GT-3 dominate in most countries, GT-4 and GT-5 are mostly found in lower-income countries<sup>[10]</sup>.

HCV GT-4 is prevalent in Northern and Equatorial Africa and the Middle East, while GT-5 and GT-6 have been identified in South Africa and Hong Kong, respectively<sup>[12]</sup>. GT-4 represents 12%-15% (15-18 million) of total global HCV infection (Table 1)<sup>[13]</sup>, and its distribution is restricted to Egypt, Central Africa, and the Middle East regions<sup>[10]</sup>, although it has recently been reported in the Caribbean region and in India<sup>[14-16]</sup>. GT-4 (and subtype 4a in particular) dominates the HCV epidemic in Egypt<sup>[10,17]</sup>.

## EPIDEMIOLOGY OF HCV INFECTION IN EGYPT

### Prevalence

An anomaly in the distribution of HCV infection, however, was discovered in Egypt, where the prevalence was approximately 10-fold higher than in other countries<sup>[18]</sup>.



**Figure 1 Global hepatitis C virus genotype distribution.** Johns Hopkins Hospital Division of Gastroenterology and Hepatology. Available from: URL: [https://gi.jhsp.org/Upload/200710291122\\_15908\\_000.jpg](https://gi.jhsp.org/Upload/200710291122_15908_000.jpg).

The prevalence is higher in Egypt than in industrialized countries (ranging from 0.5% to 2.3%)<sup>[2]</sup>, as well as in limited-resource countries, even Pakistan (high prevalence rate of 6.5%)<sup>[19]</sup>.

In 2008, an Egyptian Demographic Health Survey (EDHS) was carried out in Egypt on 11126 women and men aged 15-59 years, and interviews were obtained for each individual. It was the first nationwide representative sample for HCV antibody testing done in Egypt. The blood samples were tested by a third generation enzyme-linked immunosorbent assay to detect the HCV antibody (Adaltis EIAgen HCV Ab, Adaltis Italia, Casalecchio di Reno, Italy) at the Central Laboratory in Cairo. HCV antibodies that were Sera positive were tested for HCV RNA<sup>[20]</sup>. Results showed HCV antibody prevalence was 14.7% (95%CI: 13.9%-15.5%). Most (> 90%) HCV isolates were found to belong to GT-4 with the remaining belonging to GT-1<sup>[21]</sup>.

Prevalence was highest in Lower Egypt (Nile Delta), followed by Upper Egypt, then by Urban governorates (Cairo, Alexandria, Port-Said, and Suez), and then by frontier governorates (17.5%, 14.7%, 9.5% and 3.8% respectively). Increased HCV antibody prevalence was shown with increasing age, in males ( $P < 0.001$ ), and in rural areas ( $P < 0.001$ ). Two thirds of the participants positive for HCV antibody were viremic, and viremia was more prevalent in males, but there was no difference in prevalence according to age or type of exposure. Decreased HCV antibody prevalence was shown with increasing educational level and wealth, but the prevalence was increased with increasing number of people in the same household. Previous history of blood transfusion, parenteral anti-schistosomiasis treatment (PAT), contaminated syringes, and female circumcision were all associated with HCV infection in univariate analysis<sup>[20]</sup>.

The prevalence of HCV antibody positivity in 2008 (after adjusting for the younger than 15 years and the older than 59 years individuals) was estimated at 12%. However, only two thirds of the infected population were viremic in the EDHS, resulting in an all age group viremic prevalence of 8.5% in 2008<sup>[22]</sup>. Mass campaigns

**Table 1 Hepatitis C virus genotype-4 % among infected patients in a selection of countries in Europe, the Middle East, Africa, and India<sup>[13]</sup>**

Country or region	GT-4 % of the total HCV infected patients in the country
Southwestern France	7.4%
Germany	3.6%
Southern Italy	1.4%
Northern Italy	3.1%
Southern Spain	14%
Saudi Arabia	60%
Lebanon	30%
Syria	30%
Cameroon	76%
Nigeria	60%
Egypt	91%
Gabon	71%
Southern India	6.2%

HCV: Hepatitis C virus; GT: Genotype.

of PAT were blamed for the HCV epidemic in Egypt<sup>[23,24]</sup>. Between 1964 and 1982, 2 million Egyptians, most of whom were children above five years of age and young adults who lived in areas where schistosomiasis was prevalent, received intravenous weekly injections of antimony salts for 12-16 wk. Insufficient sterilization of the syringes was considered the cause of the HCV transmission at that time<sup>[23]</sup>. EDHS, performed 30 years after the treatment campaigns, showed PAT to be related to 7.8% and 11.0% of infected people in rural and urban areas, respectively, whereas other means of transmission attributed to the rest of the patients. The introduction of Praziquantel in 1982, an oral drug to treat schistosomiasis<sup>[24]</sup>, did not stop the transmission of HCV due to multifactorial mechanisms including blood transfusion<sup>[25-27]</sup>, contaminated syringes<sup>[26,28]</sup>, dental intervention<sup>[26-28]</sup>, surgical and invasive medical procedures<sup>[26-30]</sup>.

An attempt to determine HCV prevalence in special populations estimated a vertical transmission rate among 1224 pregnant women. Presence of maternal positive HCV antibodies was in 105 of the women (8.6%, 95%CI: 7.05-10.17) with only 83 (6.8%) positive for HCV-RNA. Tests on infants during their first month showed that 43 out of 53 infants (81%) were positive for HCV antibodies and 7 of them (13%) were HCV-RNA positive. Six months later, only two infants (3.8%) remained HCV-RNA positive<sup>[31]</sup>.

In another cohort study<sup>[32]</sup> to detect mother-to-infant infection, 1863 pregnant women were included and tested for HCV infection. Among those pregnant women, 15.7% and 10.9% tested positive for HCV antibodies and HCV-RNA, respectively. Out of 33 (10%) infants positive for both antibodies and RNA, 29 had RNA-positive mothers and four had only antibody positive mothers who underwent clearing of infection. Fifteen (4.6%) had detectable HCV-RNA at 12 mo and 14 had cleared infection early. At 2-3 years of age, only eight (2.4%) had persistent HCV-RNA while

seven had late clearance of infection. The rate of HCV vertical transmission can be increased in the presence of human immunodeficiency virus (HIV) coinfection or elevated maternal HCV viral load, but it is not affected by breastfeeding or HCV GTs<sup>[33]</sup>.

Prevalence ranged among blood donors between 5%-25%, among blood-transfusion dependent patients between 10%-55%, and among dialyzed patients between 50%-90%<sup>[34]</sup>. Many studies were conducted in the pediatric population. Among hemophilic children, HCV-antibodies were positive in 40% of patients<sup>[35]</sup>, 47.5% of them were positive for HCV-RNA. In 2011, Barakat *et al*<sup>[36]</sup> screened 500 school children from 10 schools and found 5.8% HCV seroprevalence with viremia in 75% of them. In 2010, El-Karakasy *et al*<sup>[37]</sup> reported that the prevalence of HCV in diabetic and non-diabetic children aged below nine years was 2.5% vs 1.4% ( $P = 0.25$ ).

### Incidence

In the last estimate of HCV incidence in Egypt it was found that the number of new infections annually is < 150000<sup>[38]</sup>. In 2008, the EDHS estimated the incidence in Egypt at a rate of 6.9/1000 (95%CI: 5.5-7.4 per person per year), indicative of possibly ongoing hyper epidemic transmission<sup>[18]</sup>.

Another study was conducted on 2852 uninfected infants who were followed from birth up to 5.5 years, to detect incidence and risk factors for acquired HCV in rural Egyptian children. Fifteen infants (0.53%) seroconverted to either HCV antibodies and/or HCV-RNA: 10 of them had both antibody and HCV-RNA positive, 4 of them had only antibody positive, whereas the last one had only HCV-RNA positive. The incidence rate at all ages was 2.7/1000 person-years (PY): Higher during infancy than between 1-5 years of age (3.8/1000 PY and 2.0/1000 PY respectively). It was shown that prolonged hospitalization and low birth weight increased the risk of infection, whereas maternal HCV was the source of infection in only two older children. By the end of the follow-up period, six children (40%) had natural clearance<sup>[39]</sup>. A four-year population-based cohort study was conducted on seronegative villagers to calculate the incidence of new HCV cases. Out of 10578 participants, 25 (11 females and 14 males) caught the infection (incidence rate of 2.4/1000 PY; 95%CI: 1.6-3.5)<sup>[40]</sup>.

### Risk factors associated with HCV infection in Egypt

Using systematic literature review methods, Reker *et al*<sup>[41]</sup> selected 11 articles published between 2008 and 2013 which met the study selection criteria aiming to determine risk factors responsible for the high incidence and prevalence of HCV in Egypt. They categorized the risk factors into two major groups: "Unsafe medical practices and other risk factors". Unsafe medical practices included surgery, intravenous injections, PAT, dental intervention, stitches, and catheterization. The other risk factors included illiteracy, maternal HCV, and familial transmission<sup>[41]</sup>.

## PREVENTION

Primary prevention is the best strategy. Health education and increased community awareness towards the disease is needed. Development of new approaches to learn more about HCV transmission is required. Effort should be invested towards making HCV vaccine and direct-acting antivirals (DAAs) accessible for patients during infancy and early childhood<sup>[33]</sup>.

### Prevention inside Egypt

The Egyptian Ministry of Health and Population implemented a program in 2001 to decrease HCV transmission through medical practices, and launched a comprehensive national viral hepatitis control program in 2008 for treatment. This led to a decrease in the incidence of infection among dialyzed patients from 28% to 6%<sup>[42]</sup>. But Egypt still continues to suffer from an ongoing HCV epidemic. Therefore, a comprehensive plan is required to control the infection, which includes increase in community awareness and health education, proper sterilization of medical and dental instruments, a safe blood supply, monitoring the effect of programs, and providing proper treatment<sup>[41-48]</sup>.

## CLINICAL PRESENTATIONS

### Acute HCV

Primary infection is mostly asymptomatic which makes early detection of the disease difficult, and this leads to underestimation of its true incidence rate. Its diagnosis may be confirmed *via* a documented seroconversion to anti-HCV in a person who was previously negative. Primary HCV infection has no specific markers, and it may be diagnosed with or without an increase in alanine aminotransferase (ALT) levels<sup>[49,50]</sup>.

In primary exposure, serum HCV RNA cannot be detected before a window of 1-3 wk. Symptoms are mild and non-specific, so patients often do not seek medical assistance. Elevated ALT levels indicating the first signs of liver injury can be detected 4-12 wk after infection, and wide fluctuations are common. Severe liver inflammation is uncommon, and fulminant hepatitis is rare. Seroconversion may occur between 4 and 10 wk after exposure<sup>[51]</sup>.

When HCV viremia persists more than 6 mo, it is defined as chronic. The risk of chronic HCV developing from acute infection is high, ranging between 55% and 85%<sup>[52,53]</sup>. Natural clearance is more likely to occur in symptomatic cases<sup>[54]</sup>. On the other hand, asymptomatic cases are more likely to progress to chronicity. Spontaneous clearance in adults with chronic HCV is rarely seen<sup>[55]</sup>, but is more observed in children (8% to 45%). Spontaneous viral clearance is unlikely beyond four years of age, and viral infection that does not get cleared in the first years will progress to chronicity<sup>[56-59]</sup>. We can expect natural clearance mostly to occur within the first three months after exposure<sup>[60]</sup>. Other factors

may also increase the chance of viral clearance include interferon *L3* gene, formerly known as interleukin 28B (IL28B polymorphisms)<sup>[61]</sup> and the intensity of the cellular immune response<sup>[51]</sup>. El-Awady *et al.*<sup>[62]</sup> conducted a study in 2012 on a total of 404 subjects divided into patients infected with HCV-GT-4a ( $n = 304$ ) and a healthy group ( $n = 100$ ). They found a significant increase ( $P < 0.0005$ ) in frequencies of IL28B rs12979860 C/C GTs in the healthy population than in the other group. On the other hand, the C/C GT was significantly higher ( $P < 0.0005$ ) in spontaneous resolvers cases ( $n = 84$ ) than in healthy subjects, so they reported that GT C/C was associated with viral clearance during acute infection and suggested a central role of this GT against HCV disease progression<sup>[62]</sup>.

Higher rates of spontaneous resolution have been found in infants with the rs12979860 CC GT for the IL28B polymorphism. Infants, in particular, may have defense mechanisms that explain the inefficiency of HCV perinatal transmission, including the placenta, which has an immunoprotective role, and human leukocyte antigen DR13, where they are less likely to experience chronic HCV from vertical transmission<sup>[63]</sup>. Co-infection with HIV seems to hinder resolution. The role of type of exposure, viral load, age, sex, and previous recovery following HCV exposure is debatable. Patients, especially those with higher risk of transmission, should be educated that HCV re-infection is possible even after viral eradication<sup>[64,65]</sup>.

### Chronic HCV - disease progression

HCV can progress to cirrhosis after decades. It can also lead to liver cell failure or hepatocellular carcinoma (HCC) (approximately 2% to 4% yearly). Several factors influence the progression of the disease, the most important of which is the extent of intrahepatic inflammation elicited by HCV<sup>[66]</sup>. Persistent normal ALT level indicates slow progression of the disease<sup>[67]</sup>. Unlike the case with other viral infections, serum HCV RNA is not an indicator for disease progression<sup>[68]</sup>. GT-3 HCV is associated with accelerated fibrogenesis and an increased risk of developing HCC relative to other GTs<sup>[69-71]</sup>. Host factors are important in influencing disease progression, and those include gender, race and age. Liver fibrogenesis and HCC incidence are more seen in males<sup>[68,72]</sup>, whereas mild liver fibrosis and normal liver enzymes are more observed in females. Spontaneous eradication of the virus is also more seen in females<sup>[73]</sup>. A study on black and white Americans comparing disease progression showed no significant difference, making the impact of race unclear<sup>[74]</sup>. Egyptians infected with HCV GT-4 have high rates of advanced fibrosis. An Egyptian study reported strong correlation between subtypes 4a and 4o with HCC<sup>[75]</sup>.

Age at infection affects prognosis<sup>[76,77]</sup>. Slower progression of the disease, at least during the first 1-2 decades, is observed in children<sup>[78]</sup> and females when infected at a young age<sup>[79]</sup>. A study by Yosry *et al.*<sup>[80]</sup> was conducted on Egyptian children aged 3-17 years with chronic HCV on the relationship between HLA class

II with clinical chemical and histopathological state in that special population. The most frequent alleles were DRB1\*03, DRB1\*04, and DRB1\*13 (45.6%, 39.1% and 26.1%), respectively. Nearly half of the patients had DRB1\*03, and it was associated with a minimal amount of liver affection. Low serum albumin ( $P = 0.04$ ) was shown in patients with DRB1\*04, while high aspartate aminotransferase (AST) level ( $P = 0.05$ ) was shown in patients with DRB1\*13. In comparison to controls, DRB1\*15 was significantly reduced among cases.

Environmental factors are important in influencing disease progression, and those include alcohol consumption, tobacco inhalation and coffee consumption. Excessive alcohol intake affects disease progression and HCC risk<sup>[77,81]</sup>. Tobacco inhalation is associated with liver affection, and consequently with increased fibrosis score<sup>[82]</sup>, whereas coffee has a protective role against fibrosis<sup>[83]</sup> and HCC<sup>[84]</sup>. Steatosis<sup>[66,85]</sup>, insulin resistance<sup>[86,87]</sup>, and type 2 diabetes<sup>[88,89]</sup> are associated with disease progression and the possibility of HCC. Iron overload is related to severe fibrosis<sup>[90]</sup>. HCV and HBV coinfection increases the incidence of HCC<sup>[81]</sup>. Large meta-analyses on patients co-infected with HIV have also shown accelerated disease progression<sup>[91,92]</sup>. Liver transplantation almost always leads to HCV re-infection, as well as accelerated disease progression (compared to non-transplant patients) which may be related to a variety of factors: Chronic HCV infection is observed in liver biopsy after 1 year post-transplantation in 50%-90% of patients, whereas cirrhosis is seen in around 20% within 5 years<sup>[93,94]</sup>.

### Extrahepatic manifestations

Several extrahepatic manifestations have been associated with HCV infection that may significantly affect its morbidity and mortality. Between 38% and 76% of chronic HCV patients develop at least one extrahepatic manifestation, the presence of which when clinically significant may sometimes represent a sufficient indication for treatment, even in the presence of mild liver disease<sup>[95]</sup>. HCV infection can lead to various extrahepatic manifestations, including diseases that affect the small vessels, skin, kidneys, salivary gland, eyes, thyroid, and immune system. The majority of these manifestations are immune mediated<sup>[96]</sup>.

The most common extrahepatic manifestation associated with HCV infection is mixed cryoglobulinemia, or type II or III cryoglobulinemia, a lymphoproliferative disorder characterized by the production and tissue deposition of immune complexes formed by monoclonal and polyclonal immunoglobulins<sup>[97,98]</sup>. Anti-HCV antibodies and HCV RNA tend to concentrate in the cryoprecipitate<sup>[97]</sup>. The vast majority (up to approximately 95% in some studies) of patients with essential mixed cryoglobulinemia have HCV infection<sup>[98]</sup>. Approximately one half of HCV-infected patients have circulating cryoglobulins. Complexes tend to accumulate in small- to medium-sized blood vessels: Leukocytoclastic vasculitis is

the typical histopathologic finding and can be found in the skin as well as in various organs and tissues, including the brain, gut, and peripheral nerves<sup>[99]</sup>. Deposition of immune complexes may affect the kidneys, resulting in glomerulonephritis, mostly of the membranoproliferative type, that may lead to renal insufficiency. A long-term consequence of the syndrome is the establishment of B-cell non-Hodgkin's lymphoma<sup>[100]</sup>. Successful treatment of HCV infection with antivirals leads to a decrease of cryoglobulin levels in serum and to the remission of cryoglobulin - related symptoms and pathologic lesions<sup>[101]</sup>.

Type 2 diabetes is more frequent in HCV infection than in HBV<sup>[102]</sup>, affecting patients who are already at risk for glucose metabolism disturbances. In such patients, overt diabetes may develop earlier than in HCV-negative persons<sup>[103]</sup>. Glucose metabolism is altered by HCV at early stages of the infection, leading to insulin resistance<sup>[86]</sup> which, when combined with type 2 diabetes, accelerates liver disease progression<sup>[86,88]</sup> and reduces the response to combined pegylated interferon (PEG-IFN) with Ribavirin (RBV)<sup>[104,105]</sup>, although the response to regimens containing direct-acting antiviral (DAA) does not seem to be affected by glucose metabolism alterations<sup>[106]</sup>. Other extrahepatic-associated diseases include porphyria cutanea tarda, lichen planus, necrolytic acral erythema, sialadenitis, sicca syndrome, and autoimmune thyroiditis<sup>[96]</sup>.

## SCREENING - DIAGNOSIS

HCV screening is necessary for all individuals at high risk of HCV infection due to a history of illicit injection drug use, history of hemodialysis, history of tattooing, healthcare workers upon accidental exposure, infants born to HCV-positive mothers, history of transfusion with blood or organ transplantation, HIV infection, or chronic liver disease/hepatitis with unknown cause including elevated liver enzymes<sup>[107,108]</sup>.

### Serologic assays

Two types of assays for detecting the presence of HCV infection include serologic assays to test for antibody to HCV and molecular assays to test for HCV RNA. Identifying patient GT is important to establish prognostic risk and to guide management. HCV GT helps predict the degree of response to treatment: Patients with GT-1 or GT-4 are less likely to be cured than those with GT-2 or GT-3 with combined therapy<sup>[109]</sup>.

The American Association for the Study of Liver Diseases (AASLD) and the Infectious Diseases Society of America (IDSA) recommend that persons for whom HCV testing is advised should initially be tested for HCV antibodies (Management Guidelines)<sup>[108]</sup>. HCV antibody testing is sensitive and inexpensive, and it detects antibodies against the core and the NS3, NS4, and NS5 proteins. Current serologic assays are highly specific (> 99%). In children born to HCV-positive mothers, testing for anti-HCV antibodies is unreliable before 18

mo of age because their detection may be related to the passive transfer of maternal antibodies and not active infection<sup>[110]</sup>.

In 2011, the Food and Drug Administration (FDA) granted approval to the OraQuick HCV Rapid Antibody test (OraSure Technologies, Bethlehem, PA), for detection of HCV antibody in finger stick capillary blood and venipuncture whole blood. Its sensitivity and specificity are similar to those of FDA-approved, laboratory-conducted HCV antibody assays<sup>[111]</sup>. Because the test is rapid, it can be performed for a larger population at risk<sup>[110]</sup>.

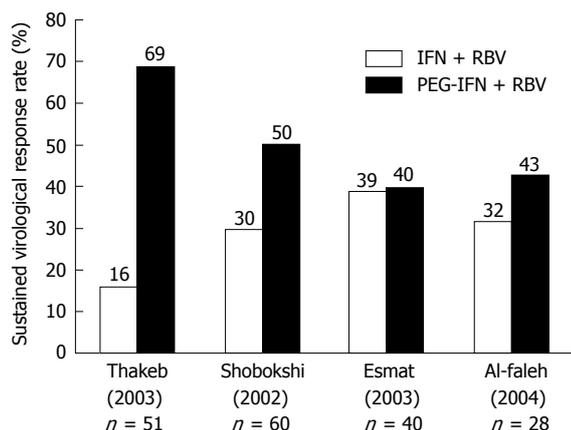
### Molecular tests (HCV-RNA and genotyping)

The CDC has recommended that positive HCV antibody results can be confirmed with a HCV RNA test that detects HCV viremia<sup>[112]</sup>. It is recommended to perform quantitative HCV RNA testing on all patients candidate for HCV treatment. HCV RNA testing is also recommended for patients with negative HCV antibody test results if they are immunocompromised. In special situations in which acute HCV infection is suspected, it is important to remember that HCV antibodies may not be present. Although the seronegative window of acute infection has diminished with improved sensitivity of HCV antibody testing, HCV RNA testing is recommended as early as 1-2 wk after the initial exposure in individuals in whom early detection is desired<sup>[108,112,113]</sup>.

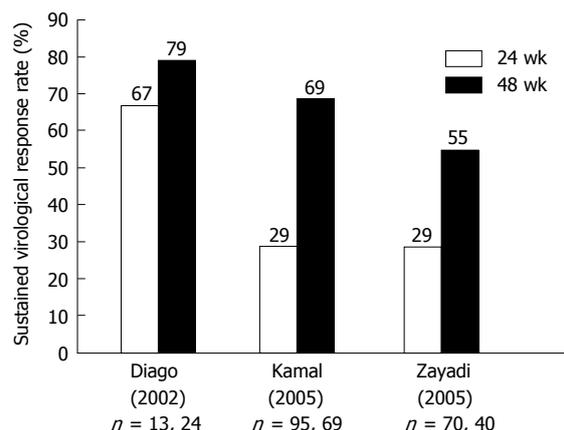
### Liver histology

The AASLD, IDSA, and European Association for the Study of the Liver (EASL) guidelines recommend that the fibrosis stage be initially determined either by liver biopsy (LB) or validated noninvasive techniques in all patients with HCV infection<sup>[108,113]</sup>. A LB provides the grade and stage of liver disease and may reveal unsuspected cirrhosis, necessitating surveillance for HCC. In children with HCV infection there is great variability in the degree of inflammation and fibrosis reported. Badizadegan *et al.*<sup>[114]</sup> found periportal fibrosis in 78% and cirrhosis in 8% of 40 children younger than 18 years of age treated at Boston Children's Hospital. A study in Cairo, Egypt, found fibrosis in 72.1% of children with HCV GT-4 (46.5% with mild and 25.6% with moderate to severe fibrosis stages), with a median age for progression of fibrosis at 5.5 years<sup>[115]</sup>.

A major effort has been devoted to identifying alternative noninvasive or minimally invasive procedures for assessing liver disease by detecting the liver fibrosis stage and establishing antiviral therapy. Several algorithms have been created using clinical information or serologic markers commonly gathered at the time of a routine diagnostic workup, such as AST, ALT, and platelet count<sup>[116]</sup> or more specific substances, such as  $\alpha$ -2-macroglobulin or hyaluronate. Combining more than one noninvasive assay appears to increase the diagnostic accuracy and may eliminate the need for LB. Transient elastography is another noninvasive approach that defines liver fibrosis using ultrasound and low-frequency



**Figure 2** Sustained virologic response to 48 wk of combination therapy in patients with hepatitis C virus genotype-4 using standard-dose pegylated-interferon and ribavirin (pegylated-interferon- $\alpha$ 2a 180  $\mu$ g or pegylated-interferon- $\alpha$ 2b 1.5 mg/kg and RBV 1-1.2 g/d). All studies were randomized control trials with intention-to-treat analysis: ( $P < 0.001$ ) ( $P < 0.01$ ) ( $P =$  not supplied) ( $P = 0.43$ )<sup>[128-131]</sup>. PEG-IFN: Pegylated-interferon; RBV: Ribavirin.



**Figure 3** Results of hepatitis C virus genotype-4 treatments with pegylated-interferon and ribavirin: 24 wk vs 48 wk. ( $P =$  not reported) ( $P = 0.001$ ) ( $P = 0.006$ ) showing far less sustained virologic response rates with 24 wk of therapy<sup>[132-134]</sup>.

elastic waves<sup>[117]</sup>. A study was conducted on chronic HCV Egyptian patients to evaluate different new noninvasive methods for assessment of liver fibrosis. The aim of that study was to evaluate whether GT-4, increased body mass index, and co-infection with schistosomiasis can interfere with liver fibrosis assessment. Egyptian HCV chronic patients ( $n = 312$ ) with GT-4 underwent a LB, an elastometry measurement (Fibroscan $\text{\textcircled{C}}$ ), and serum markers: AST-to-platelet ratio index, fibrosis-4 score (Fib4), and Fibrotest $\text{\textcircled{C}}$ . The researchers found that the algorithm using the Fib4 for identifying patients with F2 stage or more reduced by nearly 90% the number of LBs, and reported that noninvasive techniques were feasible in Egypt for HCV GT-4-infected patients and that Fib4 may be used to assess the F2 threshold, which decides whether treatment should be proposed or delayed<sup>[118]</sup>.

## BACKGROUND IN HCV TREATMENT

At the start of the millennium, two major advances in the management of HCV took place: one, the approval by the FDA of PEG-IFN for the treatment of HCV infection, which allowed weekly subcutaneous injections instead of the previous daily or thrice weekly injections with standard IFN; and the use of weight-based RBV. By the mid-2000s, it was established that PEG-IFN- $\alpha$ 2a or PEG-IFN- $\alpha$ 2b could be combined with weight-based RBV for GT-1 infected patients, or with flat-dosed RBV for GT-2 or GT-3 infected patients, and that combination was better than treatment with standard IFN and RBV<sup>[119]</sup>. HCV-GT-4 response to treatment with PEG-IFN/RBV has been considered better than GT-1, and worse than GT-2 and GT-3<sup>[120,121]</sup>.

With the introduction of the HCV-GT-1 effective protease inhibitors boceprevir (BOC)<sup>[122,123]</sup> and telaprevir<sup>[122,124-126]</sup> in 2011, HCV-GT-4 became the "most difficult (GT) to treat", as both protease inhibitors are not

indicated for treatment of GT-4<sup>[127]</sup>. Many studies used 48 wk duration of combined therapy to treat HCV-GT-4, and a few of them compared responses between 24-wk treatment and 48-wk treatment response rates. Figures 2 and 3 summarize the results of those studies after using standard-dose PEG-IFN and RBV. The results show sustained virologic response (SVR) rates with 24 wk of therapy to be far less than rates with 48-wk, making the longer duration the standard of care<sup>[13]</sup>.

Drawbacks for the IFN/RBV treatment include its long course duration, severe side effects, and cost, as it is not as affordable for patients in limited-resource countries<sup>[135-137]</sup>.

A major indicator of good response to PEG-IFN/RBV therapy in patients with HCV GT-4 is the IL28B GT<sup>[138-140]</sup>. The favorable CC phenotype is found in 20%-30% of Egyptian patients with chronic HCV<sup>[141,142]</sup>. In Europe, SVR rates for HCV GT-4 infected patients who are IL28B CC is  $> 80\%$ <sup>[139,140,143]</sup>. In Egypt, CC patients had response rates between 67% and 87%<sup>[141,142]</sup>. Mutations in the NS5A region, particularly in patients with more than 6 aa mutations in the Interferon RBV resistance - determining region (IRRDR) are highly associated with good response to PEG-IFN and RBV combination therapy, while a less diverse ( $\leq 5$ ) IRRDR sequence is associated with non-response<sup>[144-146]</sup>. Another predictor of response to PEG-IFN/RBV therapy was found to be insulin resistance, which impairs response rates to PEG-IFN/RBV therapy in HCV GT-4 patients, and patients with Homeostasis Model Assessment for Insulin Resistance scores  $> 2$  had lower SVR rates than those with scores  $< 2$  (36% vs 72%)<sup>[147]</sup>.

Abu-Mouch *et al.*<sup>[148]</sup> found that adding vitamin-D to the standard of care with PEG-IFN/RBV therapy for HCV-GT-1 infected patients increases SVR rates from 42% to 82%. Esmat *et al.*<sup>[149]</sup> reported that vitamin D supplementation, despite its role in other GTs, has no positive impact on treatment outcome in HCV-GT-4 patients where SVR was achieved in 51.2% in group 2, [who received the standard of care therapy (SOC

therapy) plus vitamin D3 (Cholecalciferol) in a dose of 15000 IU/wk during the treatment course] and 71.4% in group 1 (who received the SOC therapy consisting of PEG-IFN- $\alpha$ 2b plus RBV) by per-protocol analysis and in 44% in group 2 and in 68.6% in group 1 by intention to treat analysis (*P* value 0.22 and 0.220, respectively)<sup>[149]</sup>.

The Ministry of Health in Egypt has embarked on a national treatment program since 2006, where all eligible patients are treated with PEG-IFN/RBV for 48 wk. Annually, 40000-50000 patients have been treated, and 350000 patients had received therapy in this program by 2013<sup>[150]</sup>. SVR rates for patients treated with the original PEG-IFN- $\alpha$ 2a and alfa-2b were 54%-59%<sup>[151,152]</sup> and response rates to a locally produced biosimilar PEG-IFN were reported at 52%<sup>[153]</sup>.

## TREATMENT CONSIDERATIONS IN CHILDREN

Infected children are generally asymptomatic and often have normal ALT levels. However, children are likely to clear the virus. Given the typically slow histologic progression of liver disease in children, some argue that treatment should be postponed until adulthood. In addition, response rates with currently available antiviral therapies approved by the FDA for the pediatric population remain suboptimal, and are associated with high costs and potential toxicities<sup>[154]</sup>.

On the other hand, early eradication of HCV is likely to reduce the social stigma associated with viral infection, a source of significant caregiver stress<sup>[155]</sup>, and improve the psychosocial status of patients and their families. Children also possess multiple characteristics that make them ideal candidates for treatment. Shorter duration of infection and a lesser degree of hepatic fibrosis are associated with improved response to antiviral therapy for HCV. In addition, children have fewer co-morbidities than adults and parental motivation enhances adherence to treatment. Children also tolerate currently available therapies better than adults, with mild adverse effects<sup>[156]</sup>. Economically, the cost of treatment is less than that in adults (fewer drugs used). It is also expected that eradicating the infection sooner will decrease the risk of transmission to the population at large<sup>[157-164]</sup>.

The decision to initiate treatment should be individualized to each patient. However, for children with GT-1 and GT-4 who have mild disease at the time of biopsy, a watch and wait approach is acceptable, anticipating the availability of more efficacious drugs. According to the North American Society for Pediatric Gastroenterology, Hepatology and Nutrition and the AASLD guidelines for management of HCV infection, considerations for treating children aged 3-17 years follow the same guidelines as adults<sup>[109]</sup>.

The results of the first randomized, placebo-controlled trial of chronic pediatric HCV treatment in the United States have been published. They show that 21% of children treated with PEG-IFN alone in contrast with

53% of those treated with PEG-IFN in combination with RBV achieved a SVR, demonstrating the superiority of the combination over single therapy<sup>[165]</sup>. An international multicenter study of 107 children published by Wirth *et al*<sup>[166]</sup> demonstrated a SVR in 53% of patients with GT-1, 93% of those with GT-2 and GT-3, and 80% of those with GT-4.

In 2008, the results of these two trials led to FDA approval of combination therapy with PEG-IFN- $\alpha$ 2b and RBV for use in children three years of age or older infected with HCV. The established duration of treatment at present is 48 wk for GT-1 and GT-4<sup>[154]</sup>. A study was conducted on 118 Egyptian children with chronic HCV, including both naïve patients and previous non-responders. All received PEG-IFN- $\alpha$ 2b plus RBV. Early virological response was achieved in 69.5%, end of treatment response (ETR) was 51.7%, and SVR was achieved in 50% of the patients. Children with previously failed treatment achieved a SVR rate of 28.6%<sup>[167]</sup>. A pilot study was conducted on a small sample of Egyptian chronic HCV children (seven cases). ETR was 42.9% and SVR was 28.6%. Children with SVR were the youngest; their mean duration of infection was 4.5 years vs 12.7 years in the others. Side effects of both IFN and RBV were mild, and required no reduction in doses<sup>[168]</sup>.

In another study on chronic HCV children, the novel *Hansenula*-derived PEG-IFN: IFN- $\alpha$ 2a (Reiferon Retard) was used in the infected patients, whether naïve or previously treated. At week 12, patients were divided according to polymerase chain reaction (PCR) results into two groups: The first group included patients who continued treatment on a weekly basis (7-d schedule) and the second group included patients who continued treatment on a 5-d schedule. Patients from either group who were PCR-negative at week 48, but had at least one PCR positive test during therapy, were assigned to have an extended treatment course of up to 72 wk. The study was registered at [www.ClinicalTrials.gov](http://www.ClinicalTrials.gov) (NCT02027493). The 5-d schedule did not affect the response rate. Treatment duration (whether 48 wk or extended 72 wk) gave similar response rates (*P* = 0.49). Type of previous treatment (short acting IFN vs PEG-IFN) did not affect the response to retreatment. On the other hand, SVR was significantly higher in previous relapsers than in previous non-responders (*P* = 0.039). It was safe, but the customized regimen did not improve response rate as SVR was detected in 23.9% of children, breakthrough was seen in 39.1% of patients, and 30.4% were non-responders. Despite recent success after the introduction of combination therapy with IFN-alpha and RBV, GT-4 is considered difficult to treat. Approximately 60% of patients fail to respond. Resistance to antiviral therapy remains a serious problem in the management of chronic HCV<sup>[169]</sup>.

## EVOLUTION OF TREATMENT FOR HCV

### *Development of future treatments: DAAs*

Currently, research is orientated to the development

and approval of DAA agents regulated within the Center for Drug Evaluation and Research at the FDA for the treatment of chronic HCV. DAA agents inhibit specific stages in the HCV replication cycle through targeting the HCV polyprotein and its cleavage products<sup>[170]</sup>.

**The HCV life cycle:** The HCV genome is composed of approximately 9600 nucleotides and generates structural and non structural (NS) proteins. The structural proteins are used to assemble new viral particles and the NS proteins support viral RNA replication. The NS3/4A, a serine protease (NS3) and cofactor (NS4A) catalyze the post-translational processing of NS proteins from the polyprotein. The products released go on to form a replicative complex (NS5A) responsible for producing viral RNA using the RNA dependent/RNA polymerase (NS5B). Finally, virions are assembled, packaged, and released<sup>[118]</sup>.

#### **HCV NS3/4A serine protease inhibitors**

Targeting this protease may inhibit viral replication. Telaprevir (TPV), BOC, and simeprevir (SMV) are all examples of HCV protease inhibitors<sup>[120,122,171]</sup>.

#### **HCV NS5B inhibitors**

The HCV polymerase inhibitors are another promising DAA class. These molecules are divided into nucleoside/nucleotide competitive polymerase [nucleoside inhibitors (NIs)] and allosteric inhibitors of RNA polymerase [non-NIs (NNIs)]. NS5B NIs as sofosbuvir (SOF) are structural analogues to the natural substrates of the polymerase and are incorporated into the RNA chain. This causes direct chain termination<sup>[172]</sup>. Since the active site of NS5B is highly conserved, NIs are effective against all GTs, and resistance to NIs is usually very low. Allosteric polymerase inhibitors inhibit the NS5B by binding to one of several discrete sites on the HCV polymerase, resulting in a conformational protein change. They are less potent than NIs, resistance occurs more frequently<sup>[173]</sup> and they appear to be GT specific. SOF is a nucleotide analogue inhibitor of the HCV NS5B RNA-dependent RNA polymerase<sup>[174]</sup>.

#### **HCV NS5A inhibitors**

Inhibition of NS5A is associated with significant reductions in HCV RNA levels, which makes these agents among the most potent antiviral molecules yet developed. NS5A inhibitors have pan-genotypic activity, *i.e.*, they suppress replication of all HCV GTs. Daclatasvir (DCV) and ledipasvir (LDV) are examples of NS5A inhibitors<sup>[171]</sup>.

With the advent and improvement of DAAs, drugs now can target almost all steps of the HCV life cycle, including entry, translation, RNA replication, assembly, and export of progeny viruses<sup>[175]</sup>. This has facilitated the designing of highly potent oral drugs characterized by shorter treatment duration, simplified dosing, a high genetic barrier to resistance, and improved safety profiles<sup>[176]</sup>.

## **COMBINATION THERAPY WITH PEG-IFN, RBV AND A DAA**

### **Combination therapy with PEG-IFN, RBV and a HCV NS3/4A serine protease inhibitor**

Selective inhibitors of HCV NS3/4 serine protease, BOC and TPV were developed and found to be effective in treating HCV-infected GT-1 patients in combination with PEG-IFN and RBV. Both agents are not indicated for use in GT-2 and GT-3 patients. Combination therapy using TPV or BOC with PEG-IFN and RBV resulted in higher rates of SVR for the treatment of naïve HCV GT-1 patients (range 61%-75%) compared with treatment with PEG-IFN and RBV (range 38%-49%). However, the addition of these drugs resulted in increased adverse effects such as anemia, neutropenia, fatigue, pyrexia, and insomnia<sup>[120,122]</sup>. Telaprevir may cause skin rash, in up to 5% of cases it can be severe, and it may cause Stevens-Johnson Syndrome (SJS)<sup>[177]</sup>. On December 2012; FDA Drug Safety Communication reported severe skin affection after combination treatment of PEG-IFN and RBV with Incivek (Telaprevir). These types of skin reactions (toxic epidermal necrolysis, drug rash with eosinophilia and systemic symptoms, and SJS) lie along a continuum of serious skin reactions that can be difficult to distinguish from each other. FDA added instructions on the drug label to stop Incivek combination treatment whenever symptoms of severe skin rash appear<sup>[178]</sup>.

The rates of stoppage of treatment containing TPV or BOC were higher than treatment of PEG-IFN and RBV alone. This affected compliance and the ability to complete treatment duration<sup>[179]</sup>.

### **Combination therapy with PEG-IFN, RBV and a second-generation protease inhibitor, SMV**

SMV, a HCV NS3/4 serine protease inhibitor, was recently developed, approved by the FDA in November 2013, and is now used to treat HCV GT-1 patients in combination with PEG-IFN and RBV<sup>[171,180]</sup>. The response rate is much higher than what was obtained with the use of first-generation protease inhibitors, BOC and TPV. Recently, DAAs with pan-genotypic activities SMV, SOF and DCV have been recommended in triple regimens with PEG-IFN/RBV for the treatment of HCV genotypes 4<sup>[181]</sup>. SMV is active against genotypes 1, 2, 4, 5 and 6. It is administered as a once-daily tablet orally and has demonstrated a favorable safety profile and limited drug-drug interactions<sup>[182]</sup>. RESTORE, a phase III, multicenter, single-arm, open-label study, conducted in France and Belgium, evaluated SMV (150 mg once-daily for 12 wk in combination with PegIFN/RBV, followed by 12-36 wk of Peg-IFN/RBV only) in 107 patients with HCV 4, either naïve or treatment-experienced<sup>[183]</sup>. Recent European guidelines have included a 24-48 wk SMV plus PEG-IFN/RBV combination as an option for HCV 4-related compensated liver disease (including cirrhotics), suggesting interruption of treatment if HCV-RNA levels are  $\geq$  25 IU/mL at week 4, 12 or 24<sup>[123]</sup>.

### **Combination therapy with PEG-IFN, RBV and HCV NS5B polymerase inhibitor, SOF**

Recently (in December 2013), the FDA approved SOF as a new component of combined HCV antiviral treatment. Phase III clinical trial using combination therapy with PEG-IFN, weight-based RBV, and SOF for 12 wk resulted in high SVR rates in GT-1, GT-4, GT-5, and GT-6 patients (89%-90%)<sup>[174,184,185]</sup>. The addition of SOF to PEG-IFN and RBV reduces the total duration of treatment.

The recent approval of SOF for treatment of HCV-GT-4 promises significant improvement in the outcome of therapy. When given as part of triple therapy with PEG-IFN/RBV for 12 wk in the phase III Neutrino trial, SOF resulted in a SVR rate of 96% in HCV-GT-4 patients<sup>[185]</sup>. When it was given for 12 wk without IFN as a dual therapy with RBV to HCV-GT-4 infected patients of Egyptian origin living in the United States, SVR rates of 79% and 59% were found in naïve and treatment experienced patients, respectively. Extending the treatment for 24 wk yielded higher SVR rates: 100% and 93%<sup>[186]</sup>.

### **Combination therapy with PEG-IFN, RBV and HCV NS5A inhibitors, DCV**

Another compound with promising efficacy against HCV-GT-4 is DCV (a NS5A inhibitor) in combination with PEG-IFN- $\alpha$ 2a and RBV. Hézode *et al.*<sup>[187]</sup> reported that this was generally well tolerated and achieved higher SVR rates at week 24 compared with placebo/PEG-IFN alfa/RBV among patients infected with HCV-GT-1 or GT-4.

## **IFN-FREE DAA THERAPY (IFN-SPARING REGIMEN)**

Over the past year, IFN-sparing regimens for treating chronic HCV have become available and more superior, as those are characterized by high rates of SVR, short duration of treatment, increased tolerability, as well as room to tailor treatment according to the patients' individual needs due to the presence of multiple agents that can interrupt several stages of the HCV lifecycle. IFN-sparing treatment is currently the new approach for both treatment-naïve and treatment-experienced patients, including even cirrhotic patients. Most treatment-experienced patients can now achieve high SVR rates (above 90%). Cirrhotic patients can reach high SVR with longer duration of treatment and/or addition of RBV therapy<sup>[188]</sup>.

### **Single DAA therapy (SOF and RBV)**

The first attempt to use an IFN-free regimen for HCV involved a combination of SOF and RBV. SOF is active against all GTs<sup>[185]</sup> and has an excellent safety profile and high barrier to resistance. All oral combination therapies with SOF and RBV for 12-24 wk in HCV GT-1 were evaluated<sup>[189,190]</sup>. SVR rates using weight-based

dosing of RBV with SOF were higher in GT-1 treatment-naïve patients (SVR 68%-84%)<sup>[184,190]</sup>. In patients with HCV GT-2 or GT-3, a study showed higher rates of SVR after SOF plus RBV for 12 wk in comparison with PEG plus RBV for 24 wk. For HCV GT-2-infected patients, SVR rates were 97% vs 78% for each treatment group, respectively. However, for HCV GT-3-infected patients, the improvement in SVR rates in the SOF group was not observed (SVR 56% vs 63%, respectively)<sup>[185]</sup>. Another study confirmed the high SVR rate for SOF plus RBV in treatment-naïve and treatment-experienced GT-2 patients (SVR 93%) and showed an improved SVR rate when this combination is used for 24 wk in patients with HCV GT-3 (SVR 80%)<sup>[191]</sup>.

A randomized, open-label study was conducted at three centers in Egypt (ClinicalTrials, NCT01838590) using SOF plus RBV for 12 wk (52 patients) or SOF plus RBV for 24 wk (51 patients). Treatment-naïve (TN) and treatment-experienced (TE) patients with chronic HCV-GT-4 (up to 20% with compensated cirrhosis) were included, 74 were GT-4a while 11 were GT- 4l, 4n, 4o, 4p, and 4u. SOF plus RBV for 24 wk resulted in a 90% SVR12 rate in patients regardless of prior treatment experience. SVR12 rates with SOF plus RBV for 12 wk were higher in TN vs TE patients. No SOF-resistance mutation S282T was found in any patient with virologic failure. Combined SOF plus RBV for 12 or 24 wk were well tolerated. The authors concluded that SOF plus RBV for 24 wk provides a simple, effective, IFN-free regimen for patients with HCV-GT-4<sup>[192]</sup>.

Phase 2 trials with SOF and RBV in adolescents and children (3-17 years) with GT-2 and GT-3 began in 2014 with an estimated study completion date in May 2018 (ClinicalTrials, Identifier: NCT02175758). HCV infected children may soon realize the benefits from the tremendous research in anti-HCV therapy in the last five years<sup>[185,190,193]</sup>.

### **Dual DDA combination therapy**

Dual therapy tends to be more effective than monotherapy in regards to viral eradication and decreasing the risk of viral resistance<sup>[194]</sup>. These targets include NS3/4A protease inhibitors (PI), NS5A inhibitors, and NS5B polymerase inhibitors [both NI/tide inhibitors and NNI]. More recently, combinations of these DAAs have been effectively used without the use of IFN and RBV to achieve high rates of SVR. In an open-label study, oral SOF and DCV taken once daily were associated with high SVR in patients infected with HCV GT-1, GT-2, and GT-3, including patients with no response to prior therapy with TPV or BOC<sup>[195]</sup>.

Phase 3 is a multicenter, open-label, single-arm study conducted at multiple sites in Spain, to investigate the efficacy and safety of a 12-wk regimen of SMV in combination with SOF in TN or TE subjects (age: 18-70 years) with chronic GT-4 HCV infection (ClinicalTrials, Identifier: NCT02250807, started January 2015 with estimated study completion date January 2016). The FDA approved SMV to be used with SOF as a combination

therapy in GT-1 in November 2014. Another Egyptian study (NCT 02278419) to investigate the efficacy and safety of an 8- or 12-wk treatment regimen of SMV in combination with SOF in TN and TE adult participants with chronic HCV-GT-4 is ongoing.

For children and adolescents with chronic HCV infection, a Phase 2 study will be conducted to investigate the safety and efficacy of LDV/SOF fixed dose combination in this particular age group (ClinicalTrials, Identifier: NCT02249182). The FDA approved the first combination pill-LDV/SOF (Harvoni) Gilead, for treatment of HCV GT-1 in October 2014.

### **Triple DDA combination therapy**

The FDA approved a three drug regimen called the AbbVie Viekira Pak (Ombitasvir, Paritaprevir, and Ritonavir tablets co-packed with Dasabuvir tablets) to treat patients with chronic HCV-GT-1 infection including cirrhotic cases in December 2014. In a randomized, open-label trial of Faldaprevir (a NS3/4A protease inhibitor) and Deleobuvir (a nonnucleoside NS5B polymerase inhibitor) with or without RBV (phase II B), used by 362 TN patients infected with HCV GT-1, the SVR at 12 wk was 52%-59% among patients who received IFN-free treatment with Faldaprevir plus Deleobuvir plus RBV<sup>[196]</sup>.

The AASLD announced detailed results from The PEARL- I study [ABT-450/R (Protease inhibitor and Ritonavir) + ABT-267 (NS5A inhibitor) ± RBV] (open-label Phase 2b), which demonstrated that 100% of GT-4 patients who were new to therapy ( $n = 42/42$ ) or who had failed previous treatment with PEG-IFN and RBV ( $n = 49/49$ ) achieved SVR rate at 12 wk post-treatment after taking the AbbVie investigational treatment with RBV. Additionally, 90.9% of patients who were new to therapy achieved SVR 12 ( $n = 40/44$ ) after taking the treatment without RBV<sup>[197]</sup>.

Another open-label Egyptian study (phase 3) began in November 2014 to evaluate the safety and efficacy of the co-administration of ABT-450/Ritonavir/ABT-267 (ABT-450/r/ABT-267) with RBV in adults with chronic HCV-GT-4 in Egypt. It includes 160 patients and has an estimated study completion date of August 2016 (ClinicalTrials, Identifier: NCT02247401). Currently, Egypt is conducting a national mass treatment program for HCV GT-4 patients based on the EASL's 2014 practice guidelines for HCV-GT-4, which (based on two studies: Neutrino study for IFN-based therapy and Ruane study for IFN-free regimens) involves triple therapy using PEG-IFN/weight-based RBV/SOF 400 mg for 12 wk for IFN-eligible patients or RBV/SOF 400 mg for 24 wk for patients who are unable to tolerate IFN. Advanced liver fibrosis patients are included in the IFN-free regimen.

Due to advancements in HCV therapy, the AASLD and IDSA designed a new protocol for the use of new agents<sup>[108]</sup>. The EASL also announced new protocols on HCV therapy in 2014, including the use of all-oral combinations of SOF/SMV and SOF/DCV<sup>[113]</sup>. The AASLD, IDSA, and EASL will release updated guidelines as new

therapeutics and regimens become approved by the FDA and European Medicines Agency.

The field of IFN-free HCV therapy is under constant development. Currently, DAA's provide patients with high SVR rates (above 90%) with short duration treatment and increased tolerable adverse effect profiles. Various oral therapy combinations can be used to improve SVR outcomes in the TE patient, according to HCV GT/subtype, type of prior regimen, and presence of cirrhosis<sup>[188]</sup>.

The next steps in the clinical development of anti-HCV therapy are expected in late 2015 and early 2016 with the availability of pangenotypic ultrarapid (4-8 wk) single pill regimens such as Grazoprevir/MK8742, SOF/GS5816, and BMS791325/DAC/Asunaprevir<sup>[198]</sup>.

The most common and tolerable adverse effects of DAA combination therapy are nasopharyngitis, headache and malaise<sup>[199]</sup>. However The FDA warned on March 2015 that serious slowing of the heart rate can occur when the antiarrhythmic drug amiodarone is taken together with either Harvoni (Ledipasvir/SOF) or with SOF taken in combination with another direct acting antiviral for the treatment of hepatitis C infection. They recommended that health care professionals should not prescribe either Harvoni or SOF combined with another direct acting antiviral, such as DCV or Olysio (SMV), with amiodarone<sup>[200]</sup>.

---

## **HCV VACCINE SHOWS PROMISE**

A new HCV vaccine from GlaxoSmithKline has shown promise in early clinical tests, prompting strong and broad immune responses. Researchers have evaluated the vaccine in humans, and it is now ready for phase 2 efficacy studies<sup>[201]</sup>.

Another Egyptian clinical trial in the field of HCV vaccination is promising: Safety and efficacy of a novel candidate peptide vaccine against HCV infection in healthy volunteers and in treated (Non-responders/responders) chronic HCV patients. Clinical Trials (phases I and II) started in March 2011 (Clinical Trial, Identifier: NCT01718834; National Liver Institute, Menofya University, Egypt).

---

## **CONCLUSION**

HCV therapy is steadily moving to an all oral, well-tolerated, DAA, short-term, and more efficacious regimen. An IFN-free regimen will be available for treatment of all GTs of HCV in the near future. To date, several DAAs have been developed and are currently being evaluated in various combinations in clinical trials (for current management strategies of chronic HCV see <http://www.aasld.org>). We can expect changes in treatment recommendations of HCV as new regimens are developed and new agents are approved by the FDA. The availability of shorter, simpler, well-tolerated treatment regimens can have a major impact in reducing the morbidity and mortality associated with HCV infec-

tion. With such a large number of therapeutic agents we can end up with a world of choices that we can select from to treat patients. We hope not just to treat some patients with HCV infection but also treat all patients to achieve a cure regardless of their fibrosis state.

## REFERENCES

- Houghton M. Discovery of the hepatitis C virus. *Liver Int* 2009; **29** Suppl 1: 82-88 [PMID: 19207970 DOI: 10.1111/j.1478-3231.2008.01925.x]
- Shepard CW, Finelli L, Alter MJ. Global epidemiology of hepatitis C virus infection. *Lancet Infect Dis* 2005; **5**: 558-567 [PMID: 16122679 DOI: 10.1016/S1473-3099(05)70216-4]
- WHO. Hepatitis C Fact sheet No. 164, 2014. [Accessed 2015 May 1]. Available from: URL: <http://www.who.int/mediacentre/factsheets/fs164/en/>
- Mohd Hanafiah K, Groeger J, Flaxman AD, Wiersma ST. Global epidemiology of hepatitis C virus infection: new estimates of age-specific antibody to HCV seroprevalence. *Hepatology* 2013; **57**: 1333-1342 [PMID: 23172780 DOI: 10.1002/hep.26141]
- Preciado MV, Valva P, Escobar-Gutierrez A, Rahal P, Ruiz-Tovar K, Yamasaki L, Vazquez-Chacon C, Martinez-Guarneros A, Carpio-Pedroza JC, Fonseca-Coronado S, Cruz-Rivera M. Hepatitis C virus molecular evolution: transmission, disease progression and antiviral therapy. *World J Gastroenterol* 2014; **20**: 15992-16013 [PMID: 25473152 DOI: 10.3748/wjg.v20.i43.15992]
- Tang H, Grisé H. Cellular and molecular biology of HCV infection and hepatitis. *Clin Sci (Lond)* 2009; **117**: 49-65 [PMID: 19515018 DOI: 10.1042/CS20080631]
- Lambers FA, Prins M, Thomas X, Molenkamp R, Kwa D, Brinkman K, van der Meer JT, Schinkel J. Alarming incidence of hepatitis C virus re-infection after treatment of sexually acquired acute hepatitis C virus infection in HIV-infected MSM. *AIDS* 2011; **25**: F21-F27 [PMID: 21857492 DOI: 10.1097/QAD.0b013e32834bac44]
- Smith DB, Bukh J, Kuiken C, Muerhoff AS, Rice CM, Stapleton JT, Simmonds P. Expanded classification of hepatitis C virus into 7 genotypes and 67 subtypes: updated criteria and genotype assignment web resource. *Hepatology* 2014; **59**: 318-327 [PMID: 24115039 DOI: 10.1002/hep.26744]
- Simmonds P. Genetic diversity and evolution of hepatitis C virus--15 years on. *J Gen Virol* 2004; **85**: 3173-3188 [PMID: 15483230 DOI: 10.1099/vir.0.80401-0]
- Messina JP, Humphreys I, Flaxman A, Brown A, Cooke GS, Pybus OG, Barnes E. Global distribution and prevalence of hepatitis C virus genotypes. *Hepatology* 2015; **61**: 77-87 [PMID: 25069599 DOI: 10.1002/hep.27259]
- Zein NN, Rakela J, Krawitt EL, Reddy KR, Tominaga T, Persing DH. Hepatitis C virus genotypes in the United States: epidemiology, pathogenicity, and response to interferon therapy. Collaborative Study Group. *Ann Intern Med* 1996; **125**: 634-639 [PMID: 8849147]
- Simmonds P, Holmes EC, Cha TA, Chan SW, McOmish F, Irvine B, Beall E, Yap PL, Kolberg J, Urdea MS. Classification of hepatitis C virus into six major genotypes and a series of subtypes by phylogenetic analysis of the NS-5 region. *J Gen Virol* 1993; **74** (Pt 11): 2391-2399 [PMID: 8245854]
- Wantuck JM, Ahmed A, Nguyen MH. Review article: the epidemiology and therapy of chronic hepatitis C genotypes 4, 5 and 6. *Aliment Pharmacol Ther* 2014; **39**: 137-147 [PMID: 24251930 DOI: 10.1111/apt.12551]
- Martial J, Morice Y, Abel S, Cabié A, Rat C, Lombard F, Edouard A, Pierre-Louis S, Garsaud P, Béra O, Chout R, Gordien E, Deny P, Césaire R. Hepatitis C virus (HCV) genotypes in the Caribbean island of Martinique: evidence for a large radiation of HCV-2 and for a recent introduction from Europe of HCV-4. *J Clin Microbiol* 2004; **42**: 784-791 [PMID: 14766854]
- Raghuraman S, Abraham P, Sridharan G, Daniel HD, Rama-krishna BS, Shaji RV. HCV genotype 4--an emerging threat as a cause of chronic liver disease in Indian (south) patients. *J Clin Virol* 2004; **31**: 253-258 [PMID: 15494265 DOI: 10.1016/j.jcv.2004.03.019]
- Singh S, Malhotra V, Sarin SK. Distribution of hepatitis C virus genotypes in patients with chronic hepatitis C infection in India. *Indian J Med Res* 2004; **119**: 145-148 [PMID: 15147119]
- Gower E, Estes C, Blach S, Razavi-Shearer K, Razavi H. Global epidemiology and genotype distribution of the hepatitis C virus infection. *J Hepatol* 2014; **61**: S45-S57 [PMID: 25086286 DOI: 10.1016/j.jhep.2014.07.027]
- El-Zanaty F, Way A. Demographic and Health Survey. Egypt Demographic and Health Survey 2008. Cairo, Egypt: Ministry of Health, El-Zanaty and Associates, and Macro International, 2009. Available from: URL: <http://dhsprogram.com/pubs/pdf/FR220/FR220.pdf>
- Baatarhuu O, Kim do Y, Ahn SH, Nymadawa P, Dahgwahdorj Y, Shagdarsuren M, Park JY, Choi JW, Oyunbileg J, Oyunsuren T, Han KH. Prevalence and genotype distribution of hepatitis C virus among apparently healthy individuals in Mongolia: a population-based nationwide study. *Liver Int* 2008; **28**: 1389-1395 [PMID: 18647237 DOI: 10.1111/j.1478-3231.2008.01820.x]
- Guerra J, Garenne M, Mohamed MK, Fontanet A. HCV burden of infection in Egypt: results from a nationwide survey. *J Viral Hepat* 2012; **19**: 560-567 [PMID: 22762140 DOI: 10.1111/j.1365-2893.2011.01576.x]
- Ray SC, Arthur RR, Carella A, Bukh J, Thomas DL. Genetic epidemiology of hepatitis C virus throughout egypt. *J Infect Dis* 2000; **182**: 698-707 [PMID: 10950762 DOI: 10.1086/315786]
- Razavi H, Waked I, Sarrazin C, Myers RP, Idilman R, Calinas F, Vogel W, Mendes Correa MC, Hézode C, Lázaro P, Akarca U, Aleman S, Balik I, Berg T, Bihl F, Bilodeau M, Blasco AJ, Brandão Mello CE, Bruggmann P, Buti M, Calleja JL, Cheinquer H, Christensen PB, Clausen M, Coelho HS, Cramp ME, Dore GJ, Doss W, Duberg AS, El-Sayed MH, Ergör G, Esmat G, Falconer K, Félix J, Ferraz ML, Ferreira PR, Frankova S, García-Samaniego J, Gerstoft J, Giria JA, Gonçalves FL, Gower E, Gschwantler M, Guimarães Pessôa M, Hindman SJ, Hofer H, Husa P, Kåberg M, Kaita KD, Kautz A, Kaymakoglu S, Krajden M, Krarup H, Laleman W, Lavanchy D, Marinho RT, Marotta P, Mauss S, Moreno C, Murphy K, Negro F, Nemecek V, Örmeci N, Øvrehus AL, Parkes J, Pasini K, Peltekian KM, Ramji A, Reis N, Roberts SK, Rosenberg WM, Roudot-Thoraval F, Ryder SD, Sarmento-Castro R, Semela D, Sherman M, Shiha GE, Sievert W, Sperl J, Stärkel P, Stauber RE, Thompson AJ, Urbanek P, Van Damme P, van Thiel I, Van Vlierberghe H, Vandijck D, Wedemeyer H, Weis N, Wiegand J, Yosry A, Zekry A, Cornberg M, Müllhaupt B, Estes C. The present and future disease burden of hepatitis C virus (HCV) infection with today's treatment paradigm. *J Viral Hepat* 2014; **21** Suppl 1: 34-59 [PMID: 24713005 DOI: 10.1111/jvh.12248]
- Frank C, Mohamed MK, Strickland GT, Lavanchy D, Arthur RR, Magder LS, El Khoby T, Abdel-Wahab Y, Aly Ohn ES, Anwar W, Sallam I. The role of parenteral antischistosomal therapy in the spread of hepatitis C virus in Egypt. *Lancet* 2000; **355**: 887-891 [PMID: 10752705]
- Rao MR, Naficy AB, Darwish MA, Darwish NM, Schisterman E, Clemens JD, Edelman R. Further evidence for association of hepatitis C infection with parenteral schistosomiasis treatment in Egypt. *BMC Infect Dis* 2002; **2**: 29 [PMID: 12464161]
- El Sherbini A, Mohsen SA, Hasan W, Mostafa S, El Gohary K, Moneib A, Abaza AH. The peak impact of an Egyptian outbreak of hepatitis C virus: has it passed or has not yet occurred? *Liver Int* 2007; **27**: 876-877 [PMID: 17617132 DOI: 10.1111/j.1478-3231.2007.01501.x]
- Habib M, Mohamed MK, Abdel-Aziz F, Magder LS, Abdel-Hamid M, Gamil F, Madkour S, Mikhail NN, Anwar W, Strickland GT, Fix AD, Sallam I. Hepatitis C virus infection in a community in the Nile Delta: risk factors for seropositivity. *Hepatology* 2001; **33**: 248-253 [PMID: 11124843 DOI: 10.1053/jhep.2001.20797]
- Medhat A, Shehata M, Magder LS, Mikhail N, Abdel-Baki L,

- Nafeh M, Abdel-Hamid M, Strickland GT, Fix AD. Hepatitis c in a community in Upper Egypt: risk factors for infection. *Am J Trop Med Hyg* 2002; **66**: 633-638 [PMID: 12201604]
- 28 **Arafa N**, El Hoseiny M, Rekecewicz C, Bakr I, El-Kafrawy S, El Daly M, Aoun S, Marzouk D, Mohamed MK, Fontanet A. Changing pattern of hepatitis C virus spread in rural areas of Egypt. *J Hepatol* 2005; **43**: 418-424 [PMID: 16019104 DOI: 10.1016/j.jhep.2005.03.021]
- 29 **Darwish MA**, Faris R, Darwish N, Shouman A, Gadallah M, El-Sharkawy MS, Edelman R, Grumbach K, Rao MR, Clemens JD. Hepatitis c and cirrhotic liver disease in the Nile delta of Egypt: a community-based study. *Am J Trop Med Hyg* 2001; **64**: 147-153 [PMID: 11442209]
- 30 **Mohamed MK**, Magder LS, Abdel-Hamid M, El-Daly M, Mikhail NN, Abdel-Aziz F, Medhat A, Thiers V, Strickland GT. Transmission of hepatitis C virus between parents and children. *Am J Trop Med Hyg* 2006; **75**: 16-20 [PMID: 16837701]
- 31 **AbdulQawi K**, Youssef A, Metwally MA, Ragih I, AbdulHamid M, Shaheen A. Prospective study of prevalence and risk factors for hepatitis C in pregnant Egyptian women and its transmission to their infants. *Croat Med J* 2010; **51**: 219-228 [PMID: 20564765]
- 32 **Shebl FM**, El-Kamary SS, Saleh DA, Abdel-Hamid M, Mikhail N, Allam A, El-Arabi H, Elhenawy I, El-Kafrawy S, El-Daly M, Selim S, El-Wahab AA, Mostafa M, Sharaf S, Hashem M, Heyward S, Stine OC, Magder LS, Stoszek S, Strickland GT. Prospective cohort study of mother-to-infant infection and clearance of hepatitis C in rural Egyptian villages. *J Med Virol* 2009; **81**: 1024-1031 [PMID: 19382251 DOI: 10.1002/jmv.21480]
- 33 **Yeung CY**, Lee HC, Chan WT, Jiang CB, Chang SW, Chuang CK. Vertical transmission of hepatitis C virus: Current knowledge and perspectives. *World J Hepatol* 2014; **6**: 643-651 [PMID: 25276280 DOI: 10.4254/wjh.v6.i9.643]
- 34 **Mohamoud YA**, Mumtaz GR, Riome S, Miller D, Abu-Raddad LJ. The epidemiology of hepatitis C virus in Egypt: a systematic review and data synthesis. *BMC Infect Dis* 2013; **13**: 288 [PMID: 23799878 DOI: 10.1186/1471-2334-13-288]
- 35 **Abdelwahab MS**, El-Raziky MS, Kaddah NA, Abou-Elewh HH. Prevalence of hepatitis C virus infection and human immunodeficiency virus in a cohort of Egyptian hemophiliac children. *Ann Saudi Med* 2012; **32**: 200-202 [PMID: 22366833]
- 36 **Barakat SH**, El-Bashir N. Hepatitis C virus infection among healthy Egyptian children: prevalence and risk factors. *J Viral Hepat* 2011; **18**: 779-784 [PMID: 21992795 DOI: 10.1111/j.1365-2893.2010.01381.x]
- 37 **El-Karakasy H**, Anwar GH, El-Raziky MS, El-Hawary M, Hashem M, El-Sayed R, El-Shabrawi M, Mohsen N, Fouad H, Esmat G. Anti-HCV prevalence among diabetic and non-diabetic Egyptian children. *Curr Diabetes Rev* 2010; **6**: 388-392 [PMID: 20879976]
- 38 **Breban R**, Doss W, Esmat G, Elsayed M, Hellard M, Ayscuse P, Albert M, Fontanet A, Mohamed MK. Towards realistic estimates of HCV incidence in Egypt. *J Viral Hepat* 2013; **20**: 294-296 [PMID: 23490375 DOI: 10.1111/j.1365-2893.2012.01650.x]
- 39 **Saleh DA**, Shebl FM, El-Kamary SS, Magder LS, Allam A, Abdel-Hamid M, Mikhail N, Hashem M, Sharaf S, Stoszek SK, Strickland GT. Incidence and risk factors for community-acquired hepatitis C infection from birth to 5 years of age in rural Egyptian children. *Trans R Soc Trop Med Hyg* 2010; **104**: 357-363 [PMID: 20153495 DOI: 10.1016/j.trstmh.2010.01.009]
- 40 **Mostafa A**, Taylor SM, el-Daly M, el-Hoseiny M, Bakr I, Arafa N, Thiers V, Rimlinger F, Abdel-Hamid M, Fontanet A, Mohamed MK. Is the hepatitis C virus epidemic over in Egypt? Incidence and risk factors of new hepatitis C virus infections. *Liver Int* 2010; **30**: 560-566 [PMID: 20141592 DOI: 10.1111/j.1478-3231.2009.02204.x]
- 41 **Reker C**, Islam KM. Risk factors associated with high prevalence rates of hepatitis C infection in Egypt. *Int J Infect Dis* 2014; **25**: 104-106 [PMID: 24865321 DOI: 10.1016/j.ijid.2014.02.003]
- 42 **Centers for Disease Control and Prevention (CDC)**. Progress toward prevention and control of hepatitis C virus infection--Egypt, 2001-2012. *MMWR Morb Mortal Wkly Rep* 2012; **61**: 545-549 [PMID: 22832935]
- 43 **Esmat G**, Hashem M, El-Raziky M, El-Akel W, El-Naghy S, El-Koofy N, El-Sayed R, Ahmed R, Atta-Allah M, Hamid MA, El-Kamary SS, El-Karakasy H. Risk factors for hepatitis C virus acquisition and predictors of persistence among Egyptian children. *Liver Int* 2012; **32**: 449-456 [PMID: 22098096 DOI: 10.1111/j.1478-3231.2011.02643.x]
- 44 **Kandeel AM**, Talaat M, Afifi SA, El-Sayed NM, Abdel Fadeel MA, Hajjeh RA, Mahoney FJ. Case control study to identify risk factors for acute hepatitis C virus infection in Egypt. *BMC Infect Dis* 2012; **12**: 294 [PMID: 23145873 DOI: 10.1186/1471-2334-12-294]
- 45 **Paez Jimenez A**, Sharaf Eldin N, Rimlinger F, El-Daly M, El-Hariri H, El-Hoseiny M, Mohsen A, Mostafa A, Delarocque-Astagneau E, Abdel-Hamid M, Fontanet A, Mohamed MK, Thiers V. HCV iatrogenic and intrafamilial transmission in Greater Cairo, Egypt. *Gut* 2010; **59**: 1554-1560 [PMID: 20947889 DOI: 10.1136/gut.2009.194266]
- 46 **Plancoulaine S**, Mohamed MK, Arafa N, Bakr I, Rekecewicz C, Trégouët DA, Obach D, El Daly M, Thiers V, Féray C, Abdel-Hamid M, Abel L, Fontanet A. Dissection of familial correlations in hepatitis C virus (HCV) seroprevalence suggests intrafamilial viral transmission and genetic predisposition to infection. *Gut* 2008; **57**: 1268-1274 [PMID: 18480169 DOI: 10.1136/gut.2007.140681]
- 47 **Saleh DA**, Shebl F, Abdel-Hamid M, Naroos S, Mikhail N, El-Batanony M, El-Kafrawy S, El-Daly M, Sharaf S, Hashem M, El-Kamary S, Magder LS, Stoszek SK, Strickland GT. Incidence and risk factors for hepatitis C infection in a cohort of women in rural Egypt. *Trans R Soc Trop Med Hyg* 2008; **102**: 921-928 [PMID: 18514243 DOI: 10.1016/j.trstmh.2008.04.011]
- 48 **Zahrán KM**, Badary MS, Agban MN, Abdel Aziz NH. Pattern of hepatitis virus infection among pregnant women and their newborns at the Women's Health Center of Assiut University, Upper Egypt. *Int J Gynaecol Obstet* 2010; **111**: 171-174 [PMID: 20708181 DOI: 10.1016/j.ijgo.2010.06.013]
- 49 **Armstrong GL**, Alter MJ, McQuillan GM, Margolis HS. The past incidence of hepatitis C virus infection: implications for the future burden of chronic liver disease in the United States. *Hepatology* 2000; **31**: 777-782 [PMID: 10706572 DOI: 10.1002/hep.510310332]
- 50 **Centers for Disease Control and Prevention**. Disease Burden from Viral Hepatitis A, B, and C in the United States. 2015. Available from: URL: <http://www.cdc.gov/hepatitis/Statistics/index.htm>
- 51 **Santantonio T**, Wiegand J, Gerlach JT. Acute hepatitis C: current status and remaining challenges. *J Hepatol* 2008; **49**: 625-633 [PMID: 18706735 DOI: 10.1016/j.jhep.2008.07.005]
- 52 **McHutchison JG**. Understanding hepatitis C. *Am J Manag Care* 2004; **10**: S21-S29 [PMID: 15084064]
- 53 **Seeff LB**. Natural history of chronic hepatitis C. *Hepatology* 2002; **36**: S35-S46 [PMID: 12407575 DOI: 10.1053/jhep.2002.36806]
- 54 **Gerlach JT**, Diepolder HM, Zachoval R, Gruener NH, Jung MC, Ulsenheimer A, Schraut WW, Schirren CA, Waehtler M, Backmund M, Pape GR. Acute hepatitis C: high rate of both spontaneous and treatment-induced viral clearance. *Gastroenterology* 2003; **125**: 80-88 [PMID: 12851873]
- 55 **Yoshikawa M**, Morimoto Y, Shiroi A, Yoshiji H, Kuriyama S, Fukui H. Spontaneous elimination of serum HCV-RNA after total gastrectomy for early gastric cancer in a patient with chronic hepatitis C. *Am J Gastroenterol* 2001; **96**: 922-923 [PMID: 11280585 DOI: 10.1111/j.1572-0241.2001.03650.x]
- 56 **Fujisawa T**, Komatsu H, Inui A, Miyagawa Y, Onoue M, Sekine I, Yokota S, Hanada R, Yamamoto K, Inui M. Spontaneous remission of chronic hepatitis C in children. *Eur J Pediatr* 1997; **156**: 773-776 [PMID: 9365066]
- 57 **Iorio R**, Giannattasio A, Sepe A, Terracciano LM, Vecchione R, Vegnente A. Chronic hepatitis C in childhood: an 18-year experience. *Clin Infect Dis* 2005; **41**: 1431-1437 [PMID: 16231253 DOI: 10.1086/497141]
- 58 **Vogt M**, Lang T, Frösner G, Klingler C, Sendl AF, Zeller A,

- Wiebecke B, Langer B, Meisner H, Hess J. Prevalence and clinical outcome of hepatitis C infection in children who underwent cardiac surgery before the implementation of blood-donor screening. *N Engl J Med* 1999; **341**: 866-870 [PMID: 10498458 DOI: 10.1056/NEJM199909163411202]
- 59 **Wirth S.** Current treatment options and response rates in children with chronic hepatitis C. *World J Gastroenterol* 2012; **18**: 99-104 [PMID: 22253515 DOI: 10.3748/wjg.v18.i2.99]
- 60 **Santantonio T,** Medda E, Ferrari C, Fabris P, Cariti G, Massari M, Babudieri S, Toti M, Francavilla R, Ancarani F, Antonucci G, Scotto G, Di Marco V, Pastore G, Stroffolini T. Risk factors and outcome among a large patient cohort with community-acquired acute hepatitis C in Italy. *Clin Infect Dis* 2006; **43**: 1154-1159 [PMID: 17029134 DOI: 10.1086/507640]
- 61 **Thomas DL,** Thio CL, Martin MP, Qi Y, Ge D, O'Huigin C, Kidd J, Kidd K, Khakoo SI, Alexander G, Goedert JJ, Kirk GD, Donfield SM, Rosen HR, Tobler LH, Busch MP, McHutchison JG, Goldstein DB, Carrington M. Genetic variation in IL28B and spontaneous clearance of hepatitis C virus. *Nature* 2009; **461**: 798-801 [PMID: 19759533 DOI: 10.1038/nature08463]
- 62 **El-Awady MK,** Mostafa L, Tabll AA, Abdelhafez TH, Bader El Din NG, Zayed N, Shenawy RE, El Abd Y, Hasan RM, Zaghlol H, El Khayat H, Abdel Aziz AO. Association of IL28B SNP With Progression of Egyptian HCV Genotype 4 Patients to End Stage Liver Disease. *Hepat Mon* 2012; **12**: 271-277 [PMID: 22690235 DOI: 10.5812/hepatmon.835]
- 63 **Ruiz-Extremera A,** Muñoz-Gámez JA, Salmerón-Ruiz MA, de Rueda PM, Quiles-Pérez R, Gila-Medina A, Casado J, Belén Martín A, Sanjuan-Núñez L, Carazo A, Pavón EJ, Ocete-Hita E, León J, Salmerón J. Genetic variation in interleukin 28B with respect to vertical transmission of hepatitis C virus and spontaneous clearance in HCV-infected children. *Hepatology* 2011; **53**: 1830-1838 [PMID: 21413051 DOI: 10.1002/hep.24298]
- 64 **Danta M,** Semmo N, Fabris P, Brown D, Pybus OG, Sabin CA, Bhagani S, Emery VC, Dusheiko GM, Klenerman P. Impact of HIV on host-virus interactions during early hepatitis C virus infection. *J Infect Dis* 2008; **197**: 1558-1566 [PMID: 18419344 DOI: 10.1086/587843]
- 65 **Schnuriger A,** Dominguez S, Guiguet M, Harfouch S, Samri A, Ouazene Z, Slama L, Simon A, Valantin MA, Thibault V, Autran B. Acute hepatitis C in HIV-infected patients: rare spontaneous clearance correlates with weak memory CD4 T-cell responses to hepatitis C virus. *AIDS* 2009; **23**: 2079-2089 [PMID: 19710595 DOI: 10.1097/QAD.0b013e328330ed24]
- 66 **Leandro G,** Mangia A, Hui J, Fabris P, Rubbia-Brandt L, Colloredo G, Adinolfi LE, Asselah T, Jonsson JR, Smedile A, Terrault N, Pazienza V, Giordani MT, Giostra E, Sonzogni A, Ruggiero G, Marcellin P, Powell EE, George J, Negro F. Relationship between steatosis, inflammation, and fibrosis in chronic hepatitis C: a meta-analysis of individual patient data. *Gastroenterology* 2006; **130**: 1636-1642 [PMID: 16697727 DOI: 10.1053/j.gastro.2006.03.014]
- 67 **Mathurin P,** Moussalli J, Cadranet JF, Thibault V, Charlotte F, Dumouchel P, Cazier A, Huriaux JM, Devergie B, Vidaud M, Opolon P, Poynard T. Slow progression rate of fibrosis in hepatitis C virus patients with persistently normal alanine transaminase activity. *Hepatology* 1998; **27**: 868-872 [PMID: 9500720 DOI: 10.1002/hep.510270333]
- 68 **Bochud PY,** Cai T, Overbeck K, Bochud M, Dufour JF, Müllhaupt B, Borovicka J, Heim M, Moradpour D, Cerny A, Malinverni R, Francioli P, Negro F. Genotype 3 is associated with accelerated fibrosis progression in chronic hepatitis C. *J Hepatol* 2009; **51**: 655-666 [PMID: 19665246 DOI: 10.1016/j.jhep.2009.05.016]
- 69 **Kanwal F,** Kramer JR, Ilyas J, Duan Z, El-Serag HB. HCV genotype 3 is associated with an increased risk of cirrhosis and hepatocellular cancer in a national sample of U.S. Veterans with HCV. *Hepatology* 2014; **60**: 98-105 [PMID: 24615981 DOI: 10.1002/hep.27095]
- 70 **Nkontchou G,** Zioli M, Aout M, Lhabadie M, Baazia Y, Mahmoudi A, Roulot D, Ganne-Carrie N, Grando-Lemaire V, Trinchet JC, Gordien E, Vicaute E, Baghdad I, Beaugrand M. HCV genotype 3 is associated with a higher hepatocellular carcinoma incidence in patients with ongoing viral C cirrhosis. *J Viral Hepat* 2011; **18**: e516-e522 [PMID: 21914071 DOI: 10.1111/j.1365-2893.2011.01441.x]
- 71 **van der Meer AJ,** Veldt BJ, Feld JJ, Wedemeyer H, Dufour JF, Lammert F, Duarte-Rojo A, Heathcote EJ, Manns MP, Kuske L, Zeuzem S, Hofmann WP, de Knecht RJ, Hansen BE, Janssen HL. Association between sustained virological response and all-cause mortality among patients with chronic hepatitis C and advanced hepatic fibrosis. *JAMA* 2012; **308**: 2584-2593 [PMID: 23268517 DOI: 10.1001/jama.2012.144878]
- 72 **Chiba T,** Matsuzaki Y, Abei M, Shoda J, Aikawa T, Tanaka N, Osuga T. Multivariate analysis of risk factors for hepatocellular carcinoma in patients with hepatitis C virus-related liver cirrhosis. *J Gastroenterol* 1996; **31**: 552-558 [PMID: 8844477]
- 73 **Narciso-Schiavon JL,** Schiavon LL, Carvalho-Filho RJ, Freire FC, Cardoso JR, Bordin JO, Silva AE, Ferraz ML. Anti-hepatitis C virus-positive blood donors: are women any different? *Transfus Med* 2008; **18**: 175-183 [PMID: 18598280 DOI: 10.1111/j.1365-3148.2008.00859.x]
- 74 **Terrault NA,** Im K, Boylan R, Bacchetti P, Kleiner DE, Fontana RJ, Hoofnagle JH, Belle SH. Fibrosis progression in African Americans and Caucasian Americans with chronic hepatitis C. *Clin Gastroenterol Hepatol* 2008; **6**: 1403-1411 [PMID: 19081528 DOI: 10.1016/j.cgh.2008.08.006]
- 75 **Abdel-Hamid M,** El-Daly M, Molnegren V, El-Kafrawy S, Abdel-Latif S, Esmat G, Strickland GT, Loffredo C, Albert J, Widell A. Genetic diversity in hepatitis C virus in Egypt and possible association with hepatocellular carcinoma. *J Gen Virol* 2007; **88**: 1526-1531 [PMID: 17412982 DOI: 10.1099/vir.0.82626-0]
- 76 **Minola E,** Prati D, Suter F, Maggiolo F, Caprioli F, Sonzogni A, Fraquelli M, Paggi S, Conte D. Age at infection affects the long-term outcome of transfusion-associated chronic hepatitis C. *Blood* 2002; **99**: 4588-4591 [PMID: 12036892 DOI: 10.1182/blood-2001-12-0192]
- 77 **Poynard T,** Bedossa P, Opolon P. Natural history of liver fibrosis progression in patients with chronic hepatitis C. The OBSVIRC, METAVIR, CLINIVIR, and DOSVIRC groups. *Lancet* 1997; **349**: 825-832 [PMID: 9121257]
- 78 **Rerksuppaphol S,** Hardikar W, Dore GJ. Long-term outcome of vertically acquired and post-transfusion hepatitis C infection in children. *J Gastroenterol Hepatol* 2004; **19**: 1357-1362 [PMID: 15610308 DOI: 10.1111/j.1440-1746.2004.03463.x]
- 79 **Wiese M,** Grüngreiff K, Güthoff W, Lafrenz M, Oesen U, Porst H. Outcome in a hepatitis C (genotype 1b) single source outbreak in Germany—a 25-year multicenter study. *J Hepatol* 2005; **43**: 590-598 [PMID: 16237783]
- 80 **Yosry A,** Fouad R, Mahmoud S, El-Raziky MS, El-Hennawy A, Ghoneim MA. The association of HLA class II DR B1 alleles with HCV infection in Egyptian children. *Arab J Gastroenterol* 2011; **12**: 25-28 [PMID: 21429451 DOI: 10.1016/j.ajg.2011.01.007]
- 81 **Tagger A,** Donato F, Ribero ML, Chiesa R, Portera G, Gelatti U, Albertini A, Fasola M, Boffetta P, Nardi G. Case-control study on hepatitis C virus (HCV) as a risk factor for hepatocellular carcinoma: the role of HCV genotypes and the synergism with hepatitis B virus and alcohol. Brescia HCC Study. *Int J Cancer* 1999; **81**: 695-699 [PMID: 10328218]
- 82 **Hézode C,** Lonjon I, Roudot-Thoraval F, Mavier JP, Pawlotsky JM, Zafrani ES, Dhumeaux D. Impact of smoking on histological liver lesions in chronic hepatitis C. *Gut* 2003; **52**: 126-129 [PMID: 12477773]
- 83 **Freedman ND,** Everhart JE, Lindsay KL, Ghany MG, Curto TM, Shiffman ML, Lee WM, Lok AS, Di Bisceglie AM, Bonkovsky HL, Hoefs JC, Dienstag JL, Morishima C, Abnet CC, Sinha R. Coffee intake is associated with lower rates of liver disease progression in chronic hepatitis C. *Hepatology* 2009; **50**: 1360-1369 [PMID: 19676128 DOI: 10.1002/hep.23162]
- 84 **Ohishi W,** Fujiwara S, Cologne JB, Suzuki G, Akahoshi M, Nishi N, Takahashi I, Chayama K. Risk factors for hepatocellular carcinoma in a Japanese population: a nested case-control study.

- Cancer Epidemiol Biomarkers Prev* 2008; **17**: 846-854 [PMID: 18398026 DOI: 10.1158/1055-9965.EPI-07-2806]
- 85 **Pekow JR**, Bhan AK, Zheng H, Chung RT. Hepatic steatosis is associated with increased frequency of hepatocellular carcinoma in patients with hepatitis C-related cirrhosis. *Cancer* 2007; **109**: 2490-2496 [PMID: 17487861 DOI: 10.1002/cncr.22701]
- 86 **Hui JM**, Sud A, Farrell GC, Bandara P, Byth K, Kench JG, McCaughan GW, George J. Insulin resistance is associated with chronic hepatitis C virus infection and fibrosis progression [corrected]. *Gastroenterology* 2003; **125**: 1695-1704 [PMID: 14724822]
- 87 **Hung CH**, Wang JH, Hu TH, Chen CH, Chang KC, Yen YH, Kuo YH, Tsai MC, Lu SN, Lee CM. Insulin resistance is associated with hepatocellular carcinoma in chronic hepatitis C infection. *World J Gastroenterol* 2010; **16**: 2265-2271 [PMID: 20458764]
- 88 **Hu SX**, Kyulo NL, Xia VW, Hillebrand DJ, Hu KQ. Factors associated with hepatic fibrosis in patients with chronic hepatitis C: a retrospective study of a large cohort of U.S. patients. *J Clin Gastroenterol* 2009; **43**: 758-764 [PMID: 19238091 DOI: 10.1097/MCG.0b013e31818be17c]
- 89 **Veldt BJ**, Chen W, Heathcote EJ, Wedemeyer H, Reichen J, Hofmann WP, de Knecht RJ, Zeuzem S, Manns MP, Hansen BE, Schalm SW, Janssen HL. Increased risk of hepatocellular carcinoma among patients with hepatitis C cirrhosis and diabetes mellitus. *Hepatology* 2008; **47**: 1856-1862 [PMID: 18506898 DOI: 10.1002/hep.22251]
- 90 **Bonkovsky HL**, Lambrecht RW, Shan Y. Iron as a co-morbid factor in nonhemochromatotic liver disease. *Alcohol* 2003; **30**: 137-144 [PMID: 12957298]
- 91 **Deng LP**, Gui XE, Zhang YX, Gao SC, Yang RR. Impact of human immunodeficiency virus infection on the course of hepatitis C virus infection: a meta-analysis. *World J Gastroenterol* 2009; **15**: 996-1003 [PMID: 19248201]
- 92 **Thein HH**, Yi Q, Dore GJ, Krahn MD. Natural history of hepatitis C virus infection in HIV-infected individuals and the impact of HIV in the era of highly active antiretroviral therapy: a meta-analysis. *AIDS* 2008; **22**: 1979-1991 [PMID: 18784461 DOI: 10.1097/QAD.0b013e32830e6d51]
- 93 **Germani G**, Tsochatzis E, Papastergiou V, Burroughs AK. HCV in liver transplantation. *Semin Immunopathol* 2013; **35**: 101-110 [PMID: 22829333 DOI: 10.1007/s00281-012-0329-5]
- 94 **Meriden Z**, Forde KA, Pasha TL, Hui JJ, Reddy KR, Furth EE, Wells RG. Histologic predictors of fibrosis progression in liver allografts in patients with hepatitis C virus infection. *Clin Gastroenterol Hepatol* 2010; **8**: 289-296, 296.e1-8 [PMID: 19913638 DOI: 10.1016/j.cgh.2009.10.034]
- 95 **Himoto T**, Masaki T. Extrahepatic manifestations and autoantibodies in patients with hepatitis C virus infection. *Clin Dev Immunol* 2012; **2012**: 871401 [PMID: 22988469 DOI: 10.1155/2012/871401]
- 96 **Ko HM**, Hernandez-Prera JC, Zhu H, Dikman SH, Sidhu HK, Ward SC, Thung SN. Morphologic features of extrahepatic manifestations of hepatitis C virus infection. *Clin Dev Immunol* 2012; **2012**: 740138 [PMID: 22919404 DOI: 10.1155/2012/740138]
- 97 **Agnello V**, Chung RT, Kaplan LM. A role for hepatitis C virus infection in type II cryoglobulinemia. *N Engl J Med* 1992; **327**: 1490-1495 [PMID: 1383822 DOI: 10.1056/NEJM199211193272104]
- 98 **Sène D**, Limal N, Cacoub P. Hepatitis C virus-associated extrahepatic manifestations: a review. *Metab Brain Dis* 2004; **19**: 357-381 [PMID: 15554428]
- 99 **Ferri C**. Mixed cryoglobulinemia. *Orphanet J Rare Dis* 2008; **3**: 25 [PMID: 18796155 DOI: 10.1186/1750-1172-3-25]
- 100 **Pozzato G**, Mazzaro C, Crovatto M, Modolo ML, Ceselli S, Mazzi G, Sulfaro S, Franzin F, Tulissi P, Moretti M. Low-grade malignant lymphoma, hepatitis C virus infection, and mixed cryoglobulinemia. *Blood* 1994; **84**: 3047-3053 [PMID: 7949176]
- 101 **Saadoun D**, Delluc A, Piette JC, Cacoub P. Treatment of hepatitis C-associated mixed cryoglobulinemia vasculitis. *Curr Opin Rheumatol* 2008; **20**: 23-28 [PMID: 18281853 DOI: 10.1097/BOR.0b013e3282f1330c]
- 102 **White DL**, Ratziu V, El-Serag HB. Hepatitis C infection and risk of diabetes: a systematic review and meta-analysis. *J Hepatol* 2008; **49**: 831-844 [PMID: 18814931 DOI: 10.1016/j.jhep.2008.08.006]
- 103 **Mehta SH**, Brancati FL, Strathdee SA, Pankow JS, Netski D, Coresh J, Szklo M, Thomas DL. Hepatitis C virus infection and incident type 2 diabetes. *Hepatology* 2003; **38**: 50-56 [PMID: 12829986 DOI: 10.1053/jhep.2003.50291]
- 104 **Deltenre P**, Louvet A, Lemoine M, Mourad A, Fartoux L, Moreno C, Henrion J, Mathurin P, Serfaty L. Impact of insulin resistance on sustained response in HCV patients treated with pegylated interferon and ribavirin: a meta-analysis. *J Hepatol* 2011; **55**: 1187-1194 [PMID: 21703195 DOI: 10.1016/j.jhep.2011.03.010]
- 105 **Elgouhari HM**, Zein CO, Hanouneh I, Feldstein AE, Zein NN. Diabetes mellitus is associated with impaired response to antiviral therapy in chronic hepatitis C infection. *Dig Dis Sci* 2009; **54**: 2699-2705 [PMID: 19148751 DOI: 10.1007/s10620-008-0683-2]
- 106 **Younossi Z**, Negro F, Serfaty L, Pol S, Diago M, Zeuzem S, Andreone P, Lawitz EJ, Roberts S, Focaccia R, Foster GR, Horban A, Lonjon-Domanec I, Coate B, DeMasi R, Picchio G, Witek J. Homeostasis model assessment of insulin resistance does not seem to predict response to telaprevir in chronic hepatitis C in the REALIZE trial. *Hepatology* 2013; **58**: 1897-1906 [PMID: 24382638 DOI: 10.1002/hep.26437]
- 107 Recommendations for prevention and control of hepatitis C virus (HCV) infection and HCV-related chronic disease. Centers for Disease Control and Prevention. *MMWR Recomm Rep* 1998; **47**: 1-39 [PMID: 9790221]
- 108 **AASLD/IDSA/IAS-USA**. Recommendations for testing, managing, and treating hepatitis C. [Accessed 2015 Apr 29]. Available from: URL: <http://hcvguidelines.org/full-report-view>
- 109 **Rose R**, Markov PV, Lam TT, Pybus OG. Viral evolution explains the associations among hepatitis C virus genotype, clinical outcomes, and human genetic variation. *Infect Genet Evol* 2013; **20**: 418-421 [PMID: 24140473 DOI: 10.1016/j.meegid.2013.09.029]
- 110 **Ghany MG**, Strader DB, Thomas DL, Seeff LB. Diagnosis, management, and treatment of hepatitis C: an update. *Hepatology* 2009; **49**: 1335-1374 [PMID: 19330875 DOI: 10.1002/hep.22759]
- 111 **Shivkumar S**, Peeling R, Jafari Y, Joseph L, Pant Pai N. Accuracy of rapid and point-of-care screening tests for hepatitis C: a systematic review and meta-analysis. *Ann Intern Med* 2012; **157**: 558-566 [PMID: 23070489 DOI: 10.7326/0003-4819-157-8-201210160-00006]
- 112 **Centers for Disease Control and Prevention (CDC)**. Testing for HCV infection: an update of guidance for clinicians and laboratorians. *MMWR Morb Mortal Wkly Rep* 2013; **62**: 362-365 [PMID: 23657112]
- 113 **European Association for the Study of the Liver**. EASL recommendations on treatment of hepatitis C 2014. *J Hepatol* 2014; **61**: 373-395 [PMID: 24818984 DOI: 10.1016/j.jhep.2014.05.001]
- 114 **Badizadegan K**, Jonas MM, Ott MJ, Nelson SP, Perez-Atayde AR. Histopathology of the liver in children with chronic hepatitis C viral infection. *Hepatology* 1998; **28**: 1416-1423 [PMID: 9794930 DOI: 10.1002/hep.510280534]
- 115 **El-Hawary MA**, El-Raziky MS, Esmat G, Soliman H, Abouzied A, El-Raziky M, El-Akel W, El-Sayed R, Shebl F, Shaheen AA, El-Karakasy H. Assessment of hepatic fibrosis in pediatric cases with hepatitis C virus in Egypt. *World J Gastroenterol* 2007; **13**: 2846-2851 [PMID: 17569121]
- 116 **Castera L**. Transient elastography and other noninvasive tests to assess hepatic fibrosis in patients with viral hepatitis. *J Viral Hepat* 2009; **16**: 300-314 [PMID: 19254351 DOI: 10.1111/j.1365-2893.2009.01087.x]
- 117 **Sandrin L**, Fourquet B, Hasquenoph JM, Yon S, Fournier C, Mal F, Christidis C, Ziol M, Poulet B, Kazemi F, Beaugrand M, Palau R. Transient elastography: a new noninvasive method for assessment of hepatic fibrosis. *Ultrasound Med Biol* 2003; **29**: 1705-1713 [PMID: 14698338]
- 118 **Bonnard P**, Elsharkawy A, Zalata K, Delarocque-Astagneau E, Biard L, Le Foulher L, Hassan AB, Abdel-Hamid M, El-Daly M,

- Gamal ME, El Kassas M, Bedossa P, Carrat F, Fontanet A, Esmat G. Comparison of liver biopsy and noninvasive techniques for liver fibrosis assessment in patients infected with HCV-genotype 4 in Egypt. *J Viral Hepat* 2015; **22**: 245-253 [PMID: 25073725 DOI: 10.1111/jvh.12285]
- 119 **Cortez KJ**, Kottitil S. Beyond interferon: rationale and prospects for newer treatment paradigms for chronic hepatitis C. *Ther Adv Chronic Dis* 2015; **6**: 4-14 [PMID: 25553238 DOI: 10.1177/2040622314551934]
- 120 **Irshad M**, Ansari MA, Singh A, Nag P, Raghvendra L, Singh S, Badhal SS. HCV-genotypes: a review on their origin, global status, assay system, pathogenecity and response to treatment. *Hepatogastroenterology* 2010; **57**: 1529-1538 [PMID: 21443116]
- 121 **Varghese R**, Al-Khaldi J, Asker H, Fadili AA, Al Ali J, Hassan FA. Treatment of chronic hepatitis C genotype 4 with peginterferon alpha-2a plus ribavirin. *Hepatogastroenterology* 2009; **56**: 218-222 [PMID: 19453061]
- 122 **Bacon BR**, Gordon SC, Lawitz E, Marcellin P, Vierling JM, Zeuzem S, Poordad F, Goodman ZD, Sings HL, Boparai N, Burroughs M, Brass CA, Albrecht JK, Esteban R. Boceprevir for previously treated chronic HCV genotype 1 infection. *N Engl J Med* 2011; **364**: 1207-1217 [PMID: 21449784 DOI: 10.1056/NEJMoa1009482]
- 123 **Poordad F**, McCone J, Bacon BR, Bruno S, Manns MP, Sulkowski MS, Jacobson IM, Reddy KR, Goodman ZD, Boparai N, DiNubile MJ, Snickiene V, Brass CA, Albrecht JK, Bronowicki JP. Boceprevir for untreated chronic HCV genotype 1 infection. *N Engl J Med* 2011; **364**: 1195-1206 [PMID: 21449783 DOI: 10.1056/NEJMoa1010494]
- 124 **Jacobson IM**, McHutchison JG, Dusheiko G, Di Bisceglie AM, Reddy KR, Bzowej NH, Marcellin P, Muir AJ, Ferenci P, Flisiak R, George J, Rizzetto M, Shouval D, Sola R, Terg RA, Yoshida EM, Adda N, Bengtsson L, Sankoh AJ, Kieffer TL, George S, Kauffman RS, Zeuzem S. Telaprevir for previously untreated chronic hepatitis C virus infection. *N Engl J Med* 2011; **364**: 2405-2416 [PMID: 21696307 DOI: 10.1056/NEJMoa1012912]
- 125 **Sherman KE**, Flamm SL, Afdhal NH, Nelson DR, Sulkowski MS, Everson GT, Fried MW, Adler M, Reesink HW, Martin M, Sankoh AJ, Adda N, Kauffman RS, George S, Wright CI, Poordad F. Response-guided telaprevir combination treatment for hepatitis C virus infection. *N Engl J Med* 2011; **365**: 1014-1024 [PMID: 21916639 DOI: 10.1056/NEJMoa1014463]
- 126 **Zeuzem S**, Andreone P, Pol S, Lawitz E, Diago M, Roberts S, Focaccia R, Younossi Z, Foster GR, Horban A, Ferenci P, Nevens F, Müllhaupt B, Pockros P, Terg R, Shouval D, van Hoek B, Weiland O, Van Heeswijk R, De Meyer S, Luo D, Boogaerts G, Polo R, Picchio G, Beumont M. Telaprevir for retreatment of HCV infection. *N Engl J Med* 2011; **364**: 2417-2428 [PMID: 21696308 DOI: 10.1056/NEJMoa1013086]
- 127 **Benhamou Y**, Moussalli J, Ratziu V, Lebray P, De Backer K, De Meyer S, Ghys A, Luo D, Picchio GR, Beumont M. Telaprevir activity in treatment-naïve patients infected hepatitis C virus genotype 4: a randomized trial. *J Infect Dis* 2013; **208**: 1000-1007 [PMID: 23801602 DOI: 10.1093/infdis/jit274]
- 128 **Alfaleh FZ**, Hadad Q, Khuroo MS, Aljumah A, Algamedi A, Alashgar H, Al-Ahdal MN, Mayet I, Khan MQ, Kessie G. Peginterferon alpha-2b plus ribavirin compared with interferon alpha-2b plus ribavirin for initial treatment of chronic hepatitis C in Saudi patients commonly infected with genotype 4. *Liver Int* 2004; **24**: 568-574 [PMID: 15566506 DOI: 10.1111/j.1478-3231.2004.0976.x]
- 129 **Esmat G**, Mohamed MK, Abdel Hamid M, Zalata K, Khatab H, El Batanony M, Abouzied AM, El Raziky M, Shaheen AM, Ismail A, Strickland GT, Fix A, Sjogren M. The impact of steatosis on baseline characteristic and end of treatment response for chronic hepatitis (C) genotype 4 patients treated with interferon. *J Hepato* 2003; **38**: 139 [DOI: 10.1016/s0168-8278(03)80743-2]
- 130 **Shobokshi O**, Serebour F, Skakni L, Tantawi A, Dinish T, Al Quaiz M, Sandokji A, Al-Kayyal B, Al Momen S, Akbar H, Ayoola A, Amer H, Hussein E, Khawaja F, Al-Jasser N. Efficacy of pegylated (40 KDA) IFN alfa-2a (PEGASYS) plus ribavirin in the treatment of hepatitis C genotype 4 chronic active patients in Saudi Arabia. *J Hepato* 2002; **36** Supplement 1: 129 [DOI: 10.1016/S0168-8278(02)80462-7]
- 131 **Thakeb F**, Omar M, Bilharz T, Awady M, Isshak S. Randomized controlled trial of peginterferon alfa-2a plus ribavirin for chronic hepatitis C virus-genotype 4 among Egyptian patients. *Hepatology* 2003; **38**: 278A [DOI: 10.1002/hep.1840380504]
- 132 **Diago M**, Hadziyannous S, Bodenheimer H, Hassanein T, Uchman S, Marcellin P, Ramadori G, Delwaide J, Sedarati F. Optimized virological response in genotype 4 chronic hepatitis C patients treated with peginterferon alfa-2a (PEGASYS) in combination with ribavirin (RBV). *Hepatology* 2002; **36**: 364A
- 133 **El-Zayadi AR**, Attia M, Barakat EM, Badran HM, Hamdy H, El-Tawil A, El-Nakeeb A, Selim O, Saied A. Response of hepatitis C genotype-4 naïve patients to 24 weeks of Peg-interferon-alpha2b/ribavirin or induction-dose interferon-alpha2b/ribavirin/amantadine: a non-randomized controlled study. *Am J Gastroenterol* 2005; **100**: 2447-2452 [PMID: 16279899 DOI: 10.1111/j.1572-0241.2005.00253.x]
- 134 **Kamal SM**, El Tawil AA, Nakano T, He Q, Rasenack J, Hakam SA, Saleh WA, Ismail A, Aziz AA, Madwar MA. Peginterferon {alpha}-2b and ribavirin therapy in chronic hepatitis C genotype 4: impact of treatment duration and viral kinetics on sustained virological response. *Gut* 2005; **54**: 858-866 [PMID: 15888797 DOI: 10.1136/gut.2004.057182]
- 135 **Davies A**, Singh KP, Shubber Z, Ducros P, Mills EJ, Cooke G, Ford N. Treatment outcomes of treatment-naïve Hepatitis C patients co-infected with HIV: a systematic review and meta-analysis of observational cohorts. *PLoS One* 2013; **8**: e55373 [PMID: 23393570 DOI: 10.1371/journal.pone.0055373]
- 136 **Ford N**, Kirby C, Singh K, Mills EJ, Cooke G, Kamarulzaman A, duCros P. Chronic hepatitis C treatment outcomes in low- and middle-income countries: a systematic review and meta-analysis. *Bull World Health Organ* 2012; **90**: 540-550 [PMID: 22807600 DOI: 10.2471/BLT.11.097147]
- 137 **Ford N**, Singh K, Cooke GS, Mills EJ, von Schoen-Angerer T, Kamarulzaman A, du Cros P. Expanding access to treatment for hepatitis C in resource-limited settings: lessons from HIV/AIDS. *Clin Infect Dis* 2012; **54**: 1465-1472 [PMID: 22431808 DOI: 10.1093/cid/cis227]
- 138 **Antaki N**, Bibert S, Kebbewar K, Asaad F, Baroudi O, Alideeb S, Hadad M, Abboud D, Sabah H, Bochud PY, Negro F. IL28B polymorphisms predict response to therapy among chronic hepatitis C patients with HCV genotype 4. *J Viral Hepat* 2013; **20**: 59-64 [PMID: 23231085 DOI: 10.1111/j.1365-2893.2012.01621.x]
- 139 **De Nicola S**, Aghemo A, Rumi MG, Galmozzi E, Valenti L, Soffredini R, De Francesco R, Prati GM, D'Ambrosio R, Cheroni C, Donato MF, Colombo M. Interleukin 28B polymorphism predicts pegylated interferon plus ribavirin treatment outcome in chronic hepatitis C genotype 4. *Hepatology* 2012; **55**: 336-342 [PMID: 21932415 DOI: 10.1002/hep.24683]
- 140 **Stättermayer AF**, Strassl R, Maieron A, Rutter K, Stauber R, Strasser M, Beinhardt S, Datz C, Scherzer TM, Steindl-Munda P, Gschwantler M, Trauner M, Hofer H, Ferenci P. Polymorphisms of interferon- $\lambda$ 4 and IL28B - effects on treatment response to interferon/ribavirin in patients with chronic hepatitis C. *Aliment Pharmacol Ther* 2014; **39**: 104-111 [PMID: 24205831 DOI: 10.1111/apt.12547]
- 141 **El Awady MK**, Bader El Din NG, Tabll A, El Hosary Y, Abdel Aziz AO, El Khayat H, Salama M, Abdelhafez TH. IL28B polymorphism and cytomegalovirus predict response to treatment in Egyptian HCV type 4 patients. *World J Gastroenterol* 2013; **19**: 290-298 [PMID: 23345953 DOI: 10.3748/wjg.v19.i2.290]
- 142 **Ragheb MM**, Nemr NA, Kishk RM, Mandour MF, Abdou MM, Matsuura K, Watanabe T, Tanaka Y. Strong prediction of virological response to combination therapy by IL28B gene variants rs12979860 and rs8099917 in chronic hepatitis C genotype 4. *Liver Int* 2014; **34**: 890-895 [PMID: 24102823 DOI: 10.1111/liv.12321]
- 143 **Asselah T**, De Muynck S, Broët P, Masliah-Planchon J, Blanluet

- M, Bièche I, Lapalus M, Martinot-Peignoux M, Lada O, Estrabaud E, Zhang Q, El Ray A, Vidaud D, Ripault MP, Boyer N, Bedossa P, Valla D, Vidaud M, Marcellin P. IL28B polymorphism is associated with treatment response in patients with genotype 4 chronic hepatitis C. *J Hepatol* 2012; **56**: 527-532 [PMID: 21951981 DOI: 10.1016/j.jhep.2011.09.008]
- 144 **Kim SR**, El-Shamy A, Imoto S, Kim KI, Ide YH, Deng L, Shoji I, Tanaka Y, Hasegawa Y, Ota M, Hotta H. Prediction of response to pegylated interferon/ribavirin combination therapy for chronic hepatitis C genotype 1b and high viral load. *J Gastroenterol* 2012; **47**: 1143-1151 [PMID: 22441534 DOI: 10.1007/s00535-012-0578-z]
- 145 **El-Shamy A**, Nagano-Fujii M, Sasase N, Imoto S, Kim SR, Hotta H. Sequence variation in hepatitis C virus nonstructural protein 5A predicts clinical outcome of pegylated interferon/ribavirin combination therapy. *Hepatology* 2008; **48**: 38-47 [PMID: 18537193 DOI: 10.1002/hep.22339]
- 146 **Yano Y**, Seo Y, Miki A, Saito M, Kato H, Hamano K, Oya M, Ouchi S, Fujisawa T, Yamada H, Yamashita Y, Tani S, Hirohata S, Yoon S, Kitajima N, Kitagaki K, Kawara A, Nakashima T, Yu H, Maeda T, Azuma T, El-Shamy A, Hotta H, Hayashi Y. Mutations in non-structural 5A and rapid viral response to pegylated interferon- $\alpha$ -2b plus ribavirin therapy are associated with therapeutic efficacy in patients with genotype 1b chronic hepatitis C. *Int J Mol Med* 2012; **30**: 1048-1052 [PMID: 22899224 DOI: 10.3892/ijmm.2012.1093]
- 147 **Moucari R**, Ripault MP, Martinot-Peignoux M, Voitot H, Cardoso AC, Stern C, Boyer N, Maylin S, Nicolas-Chanoine MH, Vidaud M, Valla D, Bedossa P, Marcellin P. Insulin resistance and geographical origin: major predictors of liver fibrosis and response to peginterferon and ribavirin in HCV-4. *Gut* 2009; **58**: 1662-1669 [PMID: 19671541 DOI: 10.1136/gut.2009.185074]
- 148 **Abu-Mouch S**, Fireman Z, Jarchovsky J, Zeina AR, Assy N. Vitamin D supplementation improves sustained virologic response in chronic hepatitis C (genotype 1)-naïve patients. *World J Gastroenterol* 2011; **17**: 5184-5190 [PMID: 22215943 DOI: 10.3748/wjg.v17.i47.5184]
- 149 **Esmat G**, El Raziky M, Elsharkawy A, Sabry D, Hassany M, Ahmed A, Assem N, El Kassas M, Doss W. Impact of vitamin D supplementation on sustained virological response in chronic hepatitis C genotype 4 patients treated by pegylated interferon/ribavirin. *J Interferon Cytokine Res* 2015; **35**: 49-54 [PMID: 25061714 DOI: 10.1089/jir.2014.0060]
- 150 **Doss W**, Mohamed MK, Esmat G, El Sayed M, Fontanet A, Cooper S, El Sayed N. Arab Republic of Egypt, Ministry of Health and Population National Committee for the Control of Viral Hepatitis 2008. Available from: URL: [http://www.hepnile.org/images/stories/doc/NSP\\_10\\_April\\_2008\\_final2.pdf](http://www.hepnile.org/images/stories/doc/NSP_10_April_2008_final2.pdf)
- 151 **El Raziky M**, Fathalah WF, El-Akel WA, Salama A, Esmat G, Mabrouk M, Salama RM, Khatib HM. The Effect of Peginterferon Alpha-2a vs. Peginterferon Alpha-2b in Treatment of Naive Chronic HCV Genotype-4 Patients: A Single Centre Egyptian Study. *Hepat Mon* 2013; **13**: e10069 [PMID: 23922556 DOI: 10.5812/hepatmon.10069]
- 152 **Esmat G**, El Kassas M, Hassany M, Gamil M, El Raziky M. Optimizing treatment for HCV genotype 4: PEG-IFN  $\alpha$  2a vs. PEG-IFN  $\alpha$  2b; the debate continues. *Liver Int* 2014; **34** Suppl 1: 24-28 [PMID: 24373075 DOI: 10.1111/liv.12397]
- 153 **Taha AA**, El-Ray A, El-Ghannam M, Mounir B. Efficacy and safety of a novel pegylated interferon alpha-2a in Egyptian patients with genotype 4 chronic hepatitis C. *Can J Gastroenterol* 2010; **24**: 597-602 [PMID: 21037988]
- 154 **Fifi A**, Barreto A, Delgado-Borrego A. Optimal management of pediatric hepatitis C infection: a review. *Pediatric Health Med Ther* 2014; **5**: 173-84 [DOI: 10.2147/PHMT.S45256]
- 155 **Rodrigue JR**, Balistreri W, Haber B, Jonas MM, Mohan P, Molleston JP, Murray KF, Narkewicz MR, Rosenthal P, Smith LJ, Schwarz KB, Robuck P, Barton B, González-Peralta RP. Impact of hepatitis C virus infection on children and their caregivers: quality of life, cognitive, and emotional outcomes. *J Pediatr Gastroenterol Nutr* 2009; **48**: 341-347 [PMID: 19242286 DOI: 10.1097/MPG.0b013e318185998f]
- 156 **Wirth S**, Pieper-Boustani H, Lang T, Ballauff A, Kullmer U, Gerner P, Wintermeyer P, Jenke A. Peginterferon alfa-2b plus ribavirin treatment in children and adolescents with chronic hepatitis C. *Hepatology* 2005; **41**: 1013-1018 [PMID: 15793840 DOI: 10.1002/hep.20661]
- 157 **Ceci O**, Margiotta M, Marelllo F, Francavilla R, Loizzi P, Francavilla A, Mautone A, Impedovo L, Ierardi E, Mastroianni M, Bettocchi S, Selvaggi L. Vertical transmission of hepatitis C virus in a cohort of 2,447 HIV-seronegative pregnant women: a 24-month prospective study. *J Pediatr Gastroenterol Nutr* 2001; **33**: 570-575 [PMID: 11740231]
- 158 **Conte D**, Fraquelli M, Prati D, Colucci A, Minola E. Prevalence and clinical course of chronic hepatitis C virus (HCV) infection and rate of HCV vertical transmission in a cohort of 15,250 pregnant women. *Hepatology* 2000; **31**: 751-755 [PMID: 10706568 DOI: 10.1002/hep.510310328]
- 159 **Delgado-Borrego A**, Smith L, Jonas MM, Hall CA, Negre B, Jordan SH, Ogrodowicz M, Raza R, Ludwig DA, Miller T, Lipshultz SE, Gonzalez-Peralta R, Chung RT. Expected and actual case ascertainment and treatment rates for children infected with hepatitis C in Florida and the United States: epidemiologic evidence from statewide and nationwide surveys. *J Pediatr* 2012; **161**: 915-921 [PMID: 22765955 DOI: 10.1016/j.jpeds.2012.05.002]
- 160 **González-Peralta RP**, Langham MR, Andres JM, Mohan P, Colombani PM, Alford MK, Schwarz KB. Hepatocellular carcinoma in 2 young adolescents with chronic hepatitis C. *J Pediatr Gastroenterol Nutr* 2009; **48**: 630-635 [PMID: 19412012 DOI: 10.1097/MPG.0b013e318170af04]
- 161 **Karnsakul W**, Alford MK, Schwarz KB. Managing pediatric hepatitis C: current and emerging treatment options. *Ther Clin Risk Manag* 2009; **5**: 651-660 [PMID: 19707281]
- 162 **Mohan N**, González-Peralta RP, Fujisawa T, Chang MH, Heller S, Jara P, Kelly D, Mieli-Vergani G, Shah U, Murray KF. Chronic hepatitis C virus infection in children. *J Pediatr Gastroenterol Nutr* 2010; **50**: 123-131 [PMID: 20038846 DOI: 10.1097/MPG.0b013e3181c61995]
- 163 **Shiraki K**, Ohto H, Inaba N, Fujisawa T, Tajiri H, Kanzaki S, Matsui A, Morishima T, Goto K, Kimura A, Hino S. Guidelines for care of pregnant women carrying hepatitis C virus and their infants. *Pediatr Int* 2008; **50**: 138-140 [PMID: 18279227 DOI: 10.1111/j.1442-200X.2007.02518.x]
- 164 **Tovo PA**, Palomba E, Ferraris G, Principi N, Ruga E, Dallacasa P, Maccabruni A. Increased risk of maternal-infant hepatitis C virus transmission for women coinfecting with human immunodeficiency virus type 1. Italian Study Group for HCV Infection in Children. *Clin Infect Dis* 1997; **25**: 1121-1124 [PMID: 9402369]
- 165 **Schwarz KB**, Gonzalez-Peralta RP, Murray KF, Molleston JP, Haber B, Jonas MM, Mohan P, Balistreri WF, Rosenthal P, Narkewicz MR, Smith LJ, Robuck PR, Barton B. Peginterferon with or without ribavirin for chronic hepatitis C in children and adolescents: final results of the Peds-C trial. *Hepatology* 2008; **48**: 418A [DOI: 10.1002/hep.22641]
- 166 **Wirth S**, Ribes-Koninckx C, Calzado MA, Bortolotti F, Zancan L, Jara P, Shelton M, Kerkar N, Galoppo M, Pedreira A, Rodriguez-Baez N, Ciocca M, Lachaux A, Lacaille F, Lang T, Kullmer U, Huber WD, Gonzalez T, Pollack H, Alonso E, Broue P, Ramakrishna J, Neigut D, Valle-Segarra AD, Hunter B, Goodman Z, Xu CR, Zheng H, Noviello S, Sniukiene V, Brass C, Albrecht JK. High sustained virologic response rates in children with chronic hepatitis C receiving peginterferon alfa-2b plus ribavirin. *J Hepatol* 2010; **52**: 501-507 [PMID: 20189674 DOI: 10.1016/j.jhep.2010.01.016]
- 167 **El Naghi S**, Mahmoud F. Study of outcome of naïve and previous nonresponder hepatitis C virus Egyptian children treated with combined therapy of pegylated interferon plus ribavirin. *Egyptian Liver Journal* 2014; **4**: 87-93
- 168 **Ghaffar TY**, El Naghy S, El Sebaie H, El Monaiery M, Ghaffar AY. Pegylated alpha interferon 2B plus ribavirin in the treatment of HCV genotype 4 infection. *Indian J Pediatr* 2009; **76**: 895-898

- [PMID: 19904504 DOI: 10.1007/s12098-009-0187-x]
- 169 **El Naghi S**, Abdel-Ghaffar TY, El-Karaksy H, Abdel-Aty EF, El-Raziky MS, Allam AA, Helmy H, El-Araby HA, Behairy BE, El-Guindi MA, El-Sebaie H, Abdel-Ghaffar AY, Ehsan NA, El-Hennawy AM, Sira MM. Safety and efficacy of Hansenula-derived PEGylated-interferon alpha-2a and ribavirin combination in chronic hepatitis C Egyptian children. *World J Gastroenterol* 2014; **20**: 4681-4691 [PMID: 24782620 DOI: 10.3748/wjg.v20.i16.4681]
- 170 **(CDER) USDoHaHSFaDACfDEaR**. Guidance for Industry Chronic Hepatitis C Virus Infection: Developing Direct-Acting Antiviral Drugs for Treatment 2013. Available from: URL: <http://www.fda.gov/downloads/drugs/guidancecomplianceregulatoryinformation/guidances/ucm225333.pdf>
- 171 **Hayashi N**, Seto C, Kato M, Komada Y, Goto S. Once-daily simeprevir (TMC435) with peginterferon/ribavirin for treatment-naïve hepatitis C genotype 1-infected patients in Japan: the DRAGON study. *J Gastroenterol* 2014; **49**: 138-147 [PMID: 24005956 DOI: 10.1007/s00535-013-0875-1]
- 172 **Asselah T**, Marcellin P. Direct acting antivirals for the treatment of chronic hepatitis C: one pill a day for tomorrow. *Liver Int* 2012; **32** Suppl 1: 88-102 [PMID: 22212578 DOI: 10.1111/j.1478-3231.2011.02699.x]
- 173 **Schneider MD**, Sarrazin C. Antiviral therapy of hepatitis C in 2014: do we need resistance testing? *Antiviral Res* 2014; **105**: 64-71 [PMID: 24583028 DOI: 10.1016/j.antiviral.2014.02.011]
- 174 **Kowdley KV**, Lawitz E, Crespo I, Hassanein T, Davis MN, DeMicco M, Bernstein DE, Afdhal N, Vierling JM, Gordon SC, Anderson JK, Hyland RH, Dvory-Sobol H, An D, Hinds RG, Albanis E, Symonds WT, Berrey MM, Nelson DR, Jacobson IM. Sofosbuvir with pegylated interferon alfa-2a and ribavirin for treatment-naïve patients with hepatitis C genotype-1 infection (ATOMIC): an open-label, randomised, multicentre phase 2 trial. *Lancet* 2013; **381**: 2100-2107 [PMID: 23499440 DOI: 10.1016/S0140-6736(13)60247-0]
- 175 **Schmidt WN**, Nelson DR, Pawlotsky JM, Sherman KE, Thomas DL, Chung RT. Direct-acting antiviral agents and the path to interferon independence. *Clin Gastroenterol Hepatol* 2014; **12**: 728-737 [PMID: 23872239 DOI: 10.1016/j.cgh.2013.06.024]
- 176 **Schinazi R**, Halfon P, Marcellin P, Asselah T. HCV direct-acting antiviral agents: the best interferon-free combinations. *Liver Int* 2014; **34** Suppl 1: 69-78 [PMID: 24373081 DOI: 10.1111/liv.12423]
- 177 **Pellicer Corbí M**, Matoses Asensio S, Garcia Muñoz C, Ortiz Campos M, Herranz Muñoz C, Fernandez-Pacheco M, Garcia-Valdecasas, Luque Infantes M. PS-071 Telaprevir-induced Stevens-Johnson Syndrome. A case report. *Eur J Hosp Pharm-S P* 2014; **21** (Suppl 1): A172-A173 [DOI: 10.1136/ejpharm-2013-000436.422]
- 178 **FDA Drug Safety Communication**. Serious skin reactions after combination treatment with the Hepatitis C drugs Icodecik (telaprevir), peginterferon alfa, and ribavirin. 2012. Available from: URL: <http://www.fda.gov/Drugs/DrugSafety/ucm332731.htm>
- 179 **Hézode C**, Fontaine H, Dorival C, Zoulim F, Larrey D, Canva V, De Ledinghen V, Poynard T, Samuel D, Bourliere M, Alric L, Raabe JJ, Zarski JP, Marcellin P, Riachi G, Bernard PH, Loustaud-Ratti V, Chazouilleres O, Abergel A, Guyader D, Metivier S, Tran A, Di Martino V, Causse X, Dao T, Lucidarme D, Portal I, Cacoub P, Gournay J, Grando-Lemaire V, Hillon P, Attali P, Fontanges T, Rosa I, Petrov-Sanchez V, Barthe Y, Pawlotsky JM, Pol S, Carrat F, Bronowicki JP. Effectiveness of telaprevir or boceprevir in treatment-experienced patients with HCV genotype 1 infection and cirrhosis. *Gastroenterology* 2014; **147**: 132-142.e4 [PMID: 24704719 DOI: 10.1053/j.gastro.2014.03.051]
- 180 **Fried MW**, Buti M, Dore GJ, Flisiak R, Ferenci P, Jacobson I, Marcellin P, Manns M, Nikitin I, Poordad F, Sherman M, Zeuzem S, Scott J, Gilles L, Lenz O, Peeters M, Sekar V, De Smedt G, Beumont-Mauviel M. Once-daily simeprevir (TMC435) with pegylated interferon and ribavirin in treatment-naïve genotype 1 hepatitis C: the randomized PILLAR study. *Hepatology* 2013; **58**: 1918-1929 [PMID: 23907700 DOI: 10.1002/hep.26641]
- 181 **Papastergiou V**, Karatapanis S. Current status and emerging challenges in the treatment of hepatitis C virus genotypes 4 to 6. *World J Clin Cases* 2015; **3**: 210-220 [PMID: 25789294 DOI: 10.12998/wjcc.v3.i3.210]
- 182 **Gaetano JN**. Benefit-risk assessment of new and emerging treatments for hepatitis C: focus on simeprevir and sofosbuvir. *Drug Healthc Patient Saf* 2014; **6**: 37-45 [PMID: 24729731 DOI: 10.2147/DHPS.S43304]
- 183 **Moreno C**, Hezode C, Marcellin P, Bourgeois S, Francque S, Samuel D, Zoulim F, Grange JD, Lenz O, Ouwerkerk-Mahadevan S, Peeters M, Beumont-Mauviel M, Jessner W. P1319 once-daily simeprevir (tmc435) with peginterferon/ribavirin in treatment-naïve or treatment-experienced chronic hcv genotype 4-infected patients: final results of a phase iii trial. *J Hepatol* 2014; **60**: S535 [DOI: 10.1016/s0168-8278(14)61486-0]
- 184 **Lawitz E**, Lalezari JP, Hassanein T, Kowdley KV, Poordad FF, Sheikh AM, Afdhal NH, Bernstein DE, Dejesus E, Freilich B, Nelson DR, Dieterich DT, Jacobson IM, Jensen D, Abrams GA, Darling JM, Rodriguez-Torres M, Reddy KR, Sulkowski MS, Bzowej NH, Hyland RH, Mo H, Lin M, Mader M, Hinds R, Albanis E, Symonds WT, Berrey MM, Muir A. Sofosbuvir in combination with peginterferon alfa-2a and ribavirin for non-cirrhotic, treatment-naïve patients with genotypes 1, 2, and 3 hepatitis C infection: a randomised, double-blind, phase 2 trial. *Lancet Infect Dis* 2013; **13**: 401-408 [PMID: 23499158 DOI: 10.1016/S1473-3099(13)70033-1]
- 185 **Lawitz E**, Mangia A, Wyles D, Rodriguez-Torres M, Hassanein T, Gordon SC, Schultz M, Davis MN, Kayali Z, Reddy KR, Jacobson IM, Kowdley KV, Nyberg L, Subramanian GM, Hyland RH, Arterburn S, Jiang D, McNally J, Brainard D, Symonds WT, McHutchison JG, Sheikh AM, Younossi Z, Gane EJ. Sofosbuvir for previously untreated chronic hepatitis C infection. *N Engl J Med* 2013; **368**: 1878-1887 [PMID: 23607594 DOI: 10.1056/NEJMoa1214853]
- 186 **Ruane PJ**, Ain D, Stryker R, Meshrekey R, Soliman M, Wolfe PR, Riad J, Mikhail S, Kersey K, Jiang D, Massetto B, Doehle B, Kirby BJ, Knox SJ, McHutchison JG, Symonds WT. Sofosbuvir plus ribavirin for the treatment of chronic genotype 4 hepatitis C virus infection in patients of Egyptian ancestry. *J Hepatol* 2015; **62**: 1040-1046 [PMID: 25450208 DOI: 10.1016/j.jhep.2014.10.044]
- 187 **Hézode C**, Hirschfield GM, Ghesquiere W, Sievert W, Rodriguez-Torres M, Shafran SD, Thuluvath PJ, Tatum HA, Waked I, Esmat G, Lawitz EJ, Rustgi VK, Pol S, Weis N, Pockros PJ, Bourliere M, Serfaty L, Vierling JM, Fried MW, Weiland O, Brunetto MR, Everson GT, Zeuzem S, Kwo PY, Sulkowski M, Bräu N, Hernandez D, McPhee F, Wind-Rotolo M, Liu Z, Noviello S, Hughes EA, Yin PD, Schnittman S. Daclatasvir plus peginterferon alfa and ribavirin for treatment-naïve chronic hepatitis C genotype 1 or 4 infection: a randomised study. *Gut* 2015; **64**: 948-956 [PMID: 25080450 DOI: 10.1136/gutjnl-2014-307498]
- 188 **Peter J**, Nelson DR. Optimal interferon-free therapy in treatment-experienced chronic hepatitis C patients. *Liver Int* 2015; **35** Suppl 1: 65-70 [PMID: 25529089 DOI: 10.1111/liv.12718]
- 189 **Gane EJ**, Stedman CA, Hyland RH, Ding X, Svarovskaia E, Symonds WT, Hinds RG, Berrey MM. Nucleotide polymerase inhibitor sofosbuvir plus ribavirin for hepatitis C. *N Engl J Med* 2013; **368**: 34-44 [PMID: 23281974 DOI: 10.1056/NEJMoa1208953]
- 190 **Osinusi A**, Meissner EG, Lee YJ, Bon D, Heytens L, Nelson A, Sneller M, Kohli A, Barrett L, Proschan M, Herrmann E, Shivakumar B, Gu W, Kwan R, Teferi G, Talwani R, Silk R, Kotb C, Wroblewski S, Fishbein D, Dewar R, Highbarger H, Zhang X, Kleiner D, Wood BJ, Chavez J, Symonds WT, Subramanian M, McHutchison J, Polis MA, Fauci AS, Masur H, Kottlilil S. Sofosbuvir and ribavirin for hepatitis C genotype 1 in patients with unfavorable treatment characteristics: a randomized clinical trial. *JAMA* 2013; **310**: 804-811 [PMID: 23982366 DOI: 10.1001/jama.2013.109309]
- 191 **SOVALDI US full Prescribing Information**. CA: Gilead Sciences, Inc. Foster City. [Accessed 2015 May 1]. Available from: URL: <http://hcp.sovaldi.com/>
- 192 **Esmat GE**, Shiha G, Omar RF, Hassany M, Hammad R, Khairy M,

- Samir W, Soliman R, Brainard DM, Jiang D, Kersey K, Knox SJ, Masetto B, McHutchison JG, Doss WH. Sofosbuvir plus Ribavirin in the treatment of Egyptian patients with chronic genotype 4 HCV infection. The Liver Meeting®, November 7-11, Boston, MA (Poster presentation) no.: 959. 2014
- 193 **Jacobson IM**, Gordon SC, Kowdley KV, Yoshida EM, Rodriguez-Torres M, Sulkowski MS, Shiffman ML, Lawitz E, Everson G, Bennett M, Schiff E, Al-Assi MT, Subramanian GM, An D, Lin M, McNally J, Brainard D, Symonds WT, McHutchison JG, Patel K, Feld J, Pianko S, Nelson DR. Sofosbuvir for hepatitis C genotype 2 or 3 in patients without treatment options. *N Engl J Med* 2013; **368**: 1867-1877 [PMID: 23607593 DOI: 10.1056/NEJMoa1214854]
- 194 **Rong L**, Dahari H, Ribeiro RM, Perelson AS. Rapid emergence of protease inhibitor resistance in hepatitis C virus. *Sci Transl Med* 2010; **2**: 30ra32 [PMID: 20445200 DOI: 10.1126/scitranslmed.3000544]
- 195 **Sulkowski MS**, Gardiner DF, Rodriguez-Torres M, Reddy KR, Hassanein T, Jacobson I, Lawitz E, Lok AS, Hinestrosa F, Thuluvath PJ, Schwartz H, Nelson DR, Everson GT, Eley T, Wind-Rotolo M, Huang SP, Gao M, Hernandez D, McPhee F, Sherman D, Hindes R, Symonds W, Pasquinelli C, Grasela DM. Daclatasvir plus sofosbuvir for previously treated or untreated chronic HCV infection. *N Engl J Med* 2014; **370**: 211-221 [PMID: 24428467 DOI: 10.1056/NEJMoa1306218]
- 196 **Zeuzem S**, Soriano V, Asselah T, Bronowicki JP, Lohse AW, Müllhaupt B, Schuchmann M, Bourlière M, Buti M, Roberts SK, Gane EJ, Stern JO, Vinisko R, Kukulj G, Gallivan JP, Böcher WO, Mensa FJ. Faldaprevir and deleobuvir for HCV genotype 1 infection. *N Engl J Med* 2013; **369**: 630-639 [PMID: 23944300 DOI: 10.1056/NEJMoa1213557]
- 197 **Hezode C**, Marcellin P, Pol S, Hassanein T, Fleischer-Stepniewska K, Baykal T, Wang T, Lovell SS, Pilot-Matias T, Vilchez RA. O58 results from the phase 2 pearl-i study: interferon-free regimens of abt-450/r abt-267 with or without ribavirin in patients with hcv genotype 4 infection. *J Hepatol* 2014; **60**: S24 [DOI: 10.1016/s0168-8278(14)60060-x]
- 198 **Petta S**, Craxi A. Current and future HCV therapy: do we still need other anti-HCV drugs? *Liver Int* 2015; **35** Suppl 1: 4-10 [PMID: 25529081 DOI: 10.1111/liv.12714]
- 199 **Mizokami M**, Yokosuka O, Takehara T, Sakamoto N, Korenaga M, Mochizuki H, Nakane K, Enomoto H, Ikeda F, Yanase M, Toyoda H, Genda T, Umemura T, Yatsuhashi H, Ide T, Toda N, Nirei K, Ueno Y, Nishigaki Y, Betular J, Gao B, Ishizaki A, Omote M, Mo H, Garrison K, Pang PS, Knox SJ, Symonds WT, McHutchison JG, Izumi N, Omata M. Ledipasvir and sofosbuvir fixed-dose combination with and without ribavirin for 12 weeks in treatment-naive and previously treated Japanese patients with genotype 1 hepatitis C: an open-label, randomised, phase 3 trial. *Lancet Infect Dis* 2015; **15**: 645-653 [PMID: 25863559 DOI: 10.1016/S1473-3099(15)70099-X]
- 200 Hepatitis C Treatments Containing Sofosbuvir in Combination With Another Direct Acting Antiviral Drug: Drug Safety Communication - Serious Slowing of Heart Rate When Used With Antiarrhythmic Drug Amiodarone. 2015. Available from: URL: <http://www.fda.gov/Safety/MedWatch/SafetyInformation/SafetyAlertsforHumanMedicalProducts/ucm439662.htm>
- 201 **Franciscus A**. HCV Advocate Eblast: November 15, 2015. Feds to Medicaid: Stop HCV Treatment Restrictions. The latest news on hepatitis C and coinfection, research, drugs in development, treatment side effects and interferon-free direct antiviral combinations. [Accessed 2015 May 1]. Available from: URL: <http://hcvadvocate.org/hcv-advocate-eblast-october-15-2015-2/>

**P- Reviewer:** Basu PP, Kim SR **S- Editor:** Ji FF  
**L- Editor:** A **E- Editor:** Liu SQ



## 2015 Advances in Nonalcoholic Fatty Liver Disease

**Bile acid receptors and nonalcoholic fatty liver disease**

Liyun Yuan, Kiran Bambha

Liyun Yuan, Department of Gastroenterology and Liver Diseases, Keck USC School of Medicine, University of Southern California, Los Angeles, CA 90033, United States

Kiran Bambha, Division of Gastroenterology and Hepatology, University of Colorado, Aurora, CO 80045, United States

**Author contributions:** Yuan L performed the literature search and initial publication review, drafted the manuscript, and gave final approval of the version of the article to be submitted; Bambha K developed the concept for the manuscript, performed a literature search and publication review, critically revised the manuscript, and gave final approval of the version of the article to be submitted.

**Conflict-of-interest statement:** The authors have no conflicts of interest regarding this manuscript submission to report.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to:** Kiran Bambha, MD, MSc, Assistant Professor of Medicine, Division of Gastroenterology and Hepatology, University of Colorado, Anschutz Medical Campus, 12631 E. 17<sup>th</sup> Avenue, MS B-158, Aurora, CO 80045, United States. [kiran.bambha@ucdenver.edu](mailto:kiran.bambha@ucdenver.edu)  
Telephone: +1-303-7241857  
Fax: +1-303-7241891

Received: July 14, 2015

Peer-review started: July 17, 2015

First decision: September 2, 2015

Revised: November 5, 2015

Accepted: November 17, 2015

Article in press: November 25, 2015

Published online: December 8, 2015

**Abstract**

With the high prevalence of obesity, diabetes, and other

features of the metabolic syndrome in United States, nonalcoholic fatty liver disease (NAFLD) has inevitably become a very prevalent chronic liver disease and is now emerging as one of the leading indications for liver transplantation. Insulin resistance and derangement of lipid metabolism, accompanied by activation of the pro-inflammatory response and fibrogenesis, are essential pathways in the development of the more clinically significant form of NAFLD, known as non-alcoholic steatohepatitis (NASH). Recent advances in the functional characterization of bile acid receptors, such as farnesoid X receptor (FXR) and transmembrane G protein-coupled receptor (TGR) 5, have provided further insight in the pathophysiology of NASH and have led to the development of potential therapeutic targets for NAFLD and NASH. Beyond maintaining bile acid metabolism, FXR and TGR5 also regulate lipid metabolism, maintain glucose homeostasis, increase energy expenditure, and ameliorate hepatic inflammation. These intriguing features have been exploited to develop bile acid analogues to target pathways in NAFLD and NASH pathogenesis. This review provides a brief overview of the pathogenesis of NAFLD and NASH, and then delves into the biological functions of bile acid receptors, particularly with respect to NASH pathogenesis, with a description of the associated experimental data, and, finally, we discuss the prospects of bile acid analogues in the treatment of NAFLD and NASH.

**Key words:** Bile acids; Bile acid receptors; Nonalcoholic steatohepatitis; Farnesoid X receptor; Transmembrane G protein-coupled receptor 5; Nonalcoholic fatty liver disease; Hepatic steatosis

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Bile acids and bile acid receptors play important roles in modulation of feature of the metabolic syndrome, hepatic steatosis, and hepatic inflammation. Development of bile acid analogues specifically targeting farnesoid X receptor and transmembrane G protein-

coupled receptor 5 provide potential novel classes of drugs for the treatment of nonalcoholic steatohepatitis.

Yuan L, Bambha K. Bile acid receptors and nonalcoholic fatty liver disease. *World J Hepatol* 2015; 7(28): 2811-2818 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i28/2811.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i28.2811>

## INTRODUCTION

Nonalcoholic fatty liver disease (NAFLD) is now a very prevalent liver disease in the United States. It affects up to 20% of the United States population<sup>[1]</sup>, with an estimated prevalence of 43%-60% in patients with diabetes<sup>[2]</sup>, and 90% in patients with hyperlipidemia<sup>[3]</sup>. NAFLD, by definition, is macrovesicular fat accumulation in more than 5% of hepatocytes in patients who drink less than 20 g/d. NAFLD represent a spectrum of diseases ranging from simple hepatic steatosis to steatohepatitis. Simple steatosis rarely progresses to advanced fibrosis and thus does not carry an increased liver-related mortality. Nonalcoholic steatohepatitis (NASH) instead describes hepatic inflammation and hepatocyte damage within liver including lobular inflammation and hepatic ballooning in addition to macrovesicular fat. Fifteen percent to 30% of patients with NASH progresses to fibrosis, cirrhosis and cancer<sup>[4,5]</sup>, leading to the need for a liver transplant. Based on the data from the United Network for Organ Sharing and Organ Procurement and Transplantation Network Registry, the percentage of patients who underwent a liver transplant for NASH has increased to 9.7% in 2009 compared to 1.2% in 2001<sup>[6]</sup>. The number of adults with NASH awaiting liver transplant has almost tripled in 2013, compared to the year 2004<sup>[7]</sup>. NASH is projected to become the leading etiology for liver transplant in the United States.

The "two hit hypothesis" and the "multiple hits hypothesis" have been proposed to explain the underlying pathogenesis of NAFLD<sup>[8,9]</sup> (Figure 1). Simple hepatic steatosis reflects the accumulation of triglyceride in the liver, as result of influx of lipids and *de novo* lipogenesis exceeding the export of lipids in the forms of lipoproteins. Insulin resistance has been considered a primary driving force for lipid influx by promoting the lipolysis of peripheral adipose tissue, and increasing the liver uptake of free fatty acids for *de novo* lipogenesis<sup>[10]</sup>. Hyperinsulinemia and hyperglycemia also inhibit fatty acid oxidation and accelerate lipogenesis<sup>[11]</sup>. Triglyceride is exported out of liver to peripheral tissues by incorporation into very-low-density lipoprotein (VLDL) carriers, and impairment of VLDL synthesis/export has been implicated in NAFLD pathogenesis<sup>[12]</sup>. Accumulation of lipids, including triglycerides and free fatty acids, as a first hit, primes the liver - making it susceptible to additional hepatotoxic insults (second or multiple hits), which then lead to hepatocyte injury, inflammation, and fibrosis. The second hit or multiple hits involve pro-

inflammatory processes mediated by the gut-liver-axis with microbiota imbalance, mitochondrial dysfunction, oxidative stress pathway activation, and activation of intracellular signal such as nuclear factor  $\kappa$ B and c-Jun N-terminal kinase (JNK) pathways<sup>[13-17]</sup>.

Unraveling the pathogenesis of NAFLD has established several important drug targets in recent years. Among them are bile acid receptors including farnesoid X receptor (FXR) and transmembrane G protein-coupled receptor (TGR) 5, which play pivotal roles in regulation of metabolism, inflammation and cell proliferation. These receptors have emerged as attractive targets for drug development for the treatment of NAFLD, and are the focus of this review.

## BILE ACID METABOLISM

### *Bile acid synthesis*

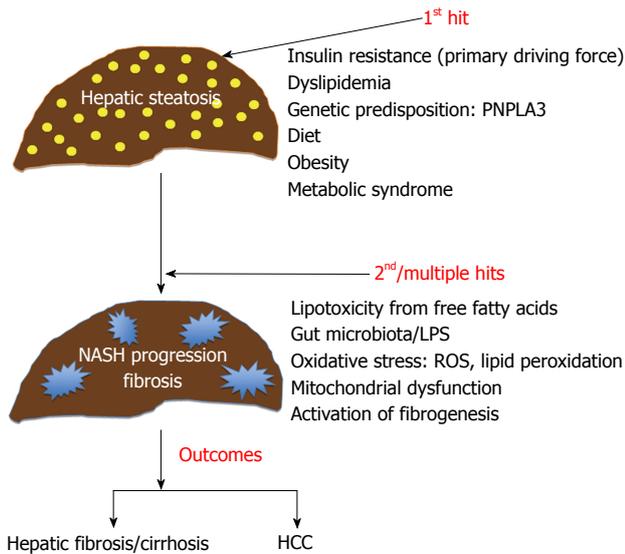
Bile acids are generated from cholesterol oxidation in the liver through two major pathways: "classic pathway" also called the neutral pathway, and the "alternative pathway" also called the acidic pathway. Cholesterol 7 $\alpha$ -hydroxylase (CYP7A1) is a rate-limiting enzyme in the classic pathway. Both of the primary bile acids, cholic acid (CA) and chenodeoxycholic acids (CDCA), are end products of the classic pathway. The alternative pathway of bile acid synthesis is initiated by sterol 27-hydroxylase (CYP27A1), an enzyme located on the inner membrane of mitochondria and widely expressed in various tissues. The alternative pathway produces oxysterols, notably 25-hydroxycholesterol and 27-hydroxycholesterol, which are important ligands in regulating inflammation, lipid metabolism, and cell proliferation.

### *Bile acid recycling*

Once synthesized in the liver, bile acids are conjugated to glycine or taurine, excreted out of liver, and stored in the gallbladder. In response to a meal, the contraction of the gallbladder delivers bile salts to the small intestine, facilitating the digestion of dietary fat. In the gastrointestinal tract, CA and CDCA are further metabolized by intestinal microbiota to secondary bile acids: Lithocholic acid (LCA) and deoxycholic acid (DCA) by de-conjugation and dehydroxylation. DCA is unable to convert back to CA in the liver, and thus the proportion of DCA in the bile acid pool varies from 1% to 50%, depending on the level and activity of bile acid 7 $\alpha$ -dehydroxylating gut bacteria and intestinal transient time. LCA is reabsorbed and reduced to CA in the liver. Overall, approximately 95% of bile acids are reabsorbed in the ileum and transported back to the liver *via* the enterohepatic circulation. Approximately 5% of bile acids are lost in the feces daily. But bile acids do not just aid in digestion and participate in the enterohepatic circulation, they also function as signaling molecules both within and outside of the liver.

## FXR AND NAFLD

FXR belongs to the family of nuclear hormone receptors



**Figure 1 Pathogenesis of nonalcoholic steatohepatitis.** Insulin resistance is considered a primary driving force for hepatic steatosis by promoting lipolysis of peripheral adipose tissue, and increasing hepatic uptake of free fatty acids for *de novo* lipogenesis. The second hit, or multiple hits, involve genetic predisposition such as PNPLA3, pro-inflammatory processes mediated by the gut-liver-axis with microbiota imbalance, mitochondrial dysfunction, activation of oxidative stress pathways, and induction of lipotoxicity from free fatty acids. HCC: Hepatocellular carcinoma; NASH: Nonalcoholic steatohepatitis; PNPLA3: Patatin-like phospholipase domain containing 3; LPS: Lipopolysaccharides; ROS: Reactive oxygen species.

that regulate expression of genes involved in a wide array of biologic processes including, development, reproduction, and metabolism, and was first described in 1995<sup>[18-20]</sup>. Bile acids were subsequently identified as unique endogenous ligands for FXR at physiologic levels<sup>[21,22]</sup> in 1999. FXR is richly expressed at the ileum, and in liver parenchymal cells. It is also expressed in liver non-parenchymal cells such as endothelial cells, Kupffer cells and stellate cells at very low level. Various bile acids activate FXR in the following order of activity: CDCA > DCA > CA > LCA. The targets and effects of FXR are outlined in detail below and summarized in Figure 2.

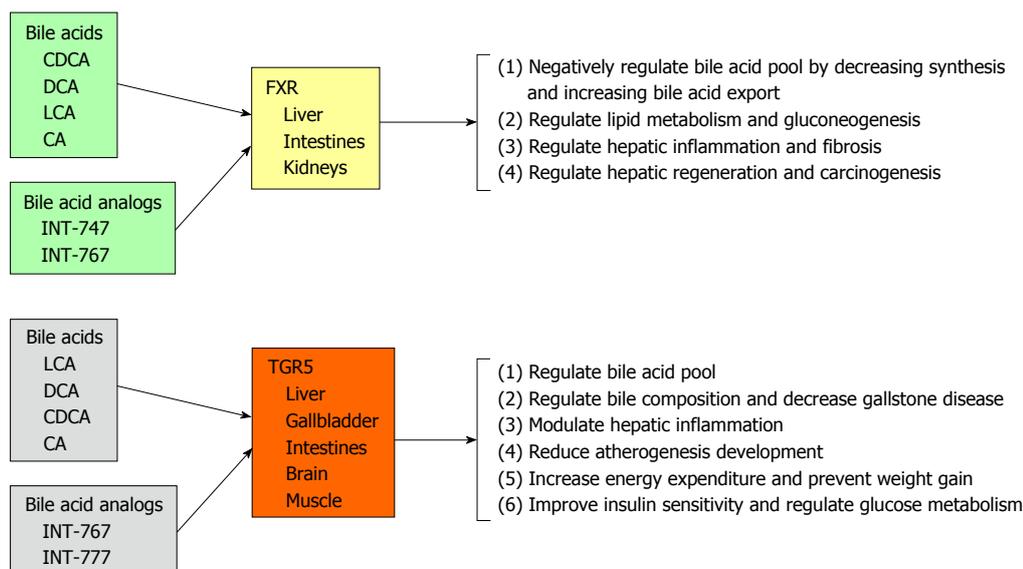
### FXR biologic functions

**Bile acid synthesis:** FXR plays an essential role in the feedback regulation of bile acid biosynthesis by repression of CYP7A1, and CYP8B1, two key enzymes in bile acids synthesis. Repression of CYP7A1 is mediated by activation of the orphan nuclear receptor small heterodimer partner (SHP)<sup>[22-24]</sup>, which in turn interact with liver receptor homolog (LRH-1). SHP protein blocks activities of LRH-1 that is known to positively regulate CYP7A1 expression. Mice lacking SHP (SHP<sup>-/-</sup>) failed to repress CYP7A1 in response to a specific agonist for FXR. Yet, Bile acid feeding can restore expression of CYP7A1 in SHP-null mice, indicating the existence of SHP-independent regulation pathways, as well. One of these pathways involves JNK mitogen-activated protein kinase activation<sup>[24,25]</sup> and fibroblast growth factor 19 (human FGF19, mouse FGF15)<sup>[26]</sup>. Additionally, FXR induces the

expression of ATP-binding cassette transporters such as bile salt export pump, multidrug resistance protein 3 (MDR3) and multidrug resistance-associated protein 2. These transporters export bile acids from hepatocytes into bile canaliculi. Activation of FXR was also found to stimulate the expression of intestinal bile acid-binding protein at ileum, which facilitates enterohepatic recycling of bile acids<sup>[21,27]</sup>.

**Lipid and glucose metabolism:** FXR also regulates lipid metabolism and gluconeogenesis<sup>[28,29]</sup>. FXR-null mice develop severe fatty liver with elevated circulating plasma cholesterol, elevated triglycerides and free fatty acids. Several lipoproteins such as phospholipid transfer protein, apoC-II, apoC-III, and apoA-1 are FXR targets<sup>[30-32]</sup> and their decreased expression likely accounts for lipid derangements in FXR-null mice. FXR also regulates lipid synthesis by involving acetyl-CoA carboxylase 1 (Acc1), Acc2, Cd36, and sterol regulatory element-binding protein 1C, with the latter being a major regulator of lipogenesis *via* stimulation of *de novo* lipogenesis, and these FXR effects likely occur through activation of SHP<sup>[33]</sup> and FGF19<sup>[34]</sup>. Beyond regulation of lipid metabolism, FXR also plays an important role in glucose homeostasis. Loss of FXR in mice lead to development of impaired glucose tolerance and insulin resistance both in liver and skeletal muscles which is associated hepatic steatosis and elevated circulating free fatty acids<sup>[35]</sup>. Bile acids alter the expression of genes involved in gluconeogenesis, including phosphoenolpyruvate carboxykinase (PEPCK), glucose-6-phosphatase (G-6-Pase), and fructose-1,6-biphosphatase<sup>[36-38]</sup>. SHP activation may modulate gluconeogenesis through repression of PEPCK and G-6-Pase<sup>[38,39]</sup>. FGF15/19 appear also to be critical in glucose regulation. In the postprandial state, FGF15/19 are released from the small intestine and inhibit hepatic gluconeogenesis, like insulin, through dephosphorylation and inactivation of cAMP regulatory element-binding protein<sup>[40]</sup>.

**Hepatic inflammation and fibrosis:** FXR also regulates hepatic inflammation and fibrosis<sup>[29]</sup>. FXR is expressed at very low levels on hepatic Kupffer, stellate, and endothelial cells. Porcine serum treatment or bile duct ligation (BDL) are commonly used experimental methods to induce cirrhosis in rats. Treatment of these rats with 6-ethyl chenodeoxycholic acid (6-ECDCA), an FXR ligand, prevents liver fibrosis in porcine serum-treated rats or BDL-treated rats, and decreases expression of matrix proteins including,  $\alpha$ 1-collagen, transforming growth factor  $\beta$ -1,  $\alpha$ SMA, and tissue inhibitors of metalloproteinase 1 and 2<sup>[41]</sup>. Interestingly, Fickert *et al.*<sup>[42]</sup> showed that FXR loss reduced fibrosis of the hepatic biliary tree. In the study, hepatic fibrosis was induced in wild type and FXR knock-out mice (FXR<sup>-/-</sup>) by a variety of methods, including carbon tetrachloride (CCl<sub>4</sub>) intoxication, 3,5-diethoxycarbonyl-1,4-dihydrocollidine feeding, BDL, or *Schistosoma mansoni* (S.m.)-infection.



**Figure 2** Characteristics of farnesoid X receptor and transmembrane G protein-coupled receptor 5, and their functions. CA: Cholic acid; LCA: Lithocholic acid; DCA: Deoxycholic acid; FXR: Farnesoid X receptor; TGR5: Transmembrane G protein-coupled receptor 5; CDCA: Chenodeoxycholic acids.

Only biliary-type hepatic fibrosis was reduced in FXR (-/-) mice with BDL and 3,5-diethoxycarbonyl-1,4-dihydrocollidine. FXR loss had no effect on the prevention of non-cholestatic liver fibrosis in the study.

**Hepatic regeneration and carcinogenesis:** FXR appears to regulate liver regeneration and carcinogenesis. CA feeding has been shown to induce liver growth and decrease mortality in mice that have undergone partial hepatectomy. This effect may involve activation of FGF15/19. Studies have shown that the protective effects with CA feeding after partial hepatectomy were significantly abolished in FGF15 (-/-) mice<sup>[43]</sup>, and proliferation of hepatocytes and cholangiocytes was also noticeably reduced in CA-fed FGF15 (-/-) mice<sup>[43]</sup>. FXR (-/-) mice developed spontaneous hepatocellular carcinoma (HCC) at age > 12 mo<sup>[44,45]</sup>. And FXR had a direct effect in down-regulating a number of tumor suppressor genes such as N-myc downstream-regulated gene 2<sup>[46]</sup> and gankyrin, a proteasomal subunit that assists in degradation of a number of tumor suppressor proteins<sup>[46,47]</sup>. Interestingly, selective reactivation of intestinal FXR can restore bile acid enterohepatic circulation and protect FXR (-/-) mice from spontaneous HCC development<sup>[48]</sup>.

### FXR agonists in the treatment of NAFLD

6-ECDCA, also known as INT-747 or obeticholic acid (OCA), is a lipophilic bile acid derivative and a potent selective FXR activator<sup>[49]</sup>. In animal studies, it improves hepatic steatosis<sup>[50]</sup>, fibrosis<sup>[42]</sup>, and portal hypertension<sup>[51]</sup>. The FLINT trial<sup>[52]</sup>, a phase II B randomized, placebo-controlled trial of OCA in human NASH, demonstrated that OCA significantly improved the NAFLD activity score in all components, including steatosis, lobular inflammation, and hepatocellular ballooning, compared to placebo, establishing a clear benefit of in

alleviating liver injury and inflammation in NAFLD. There was also some improvement in fibrosis score in the OCA group in the FLINT trial, as well, but the trial was not powered to detect the statistical significance in fibrosis changes. It remains to be determined whether or not OCA will resolve NASH and ameliorate advanced fibrosis, but further trials are ongoing.

### TGR5 and NAFLD

TGR5 is a classic G-protein coupled cell surface receptor<sup>[53,54]</sup> that is activated by bile acids in the order of LCA > DCA > CDCA > CA<sup>[53]</sup>. In the absence of bile acid binding, TGR5 is tightly associated with a G-protein complex consisting of  $\alpha$ ,  $\beta$  and  $\gamma$  subunits. Upon binding to bile acids, TGR5 allows the release of  $\alpha$  subunit, which in turn activates adenylyl cyclase, leading to the accumulation of cAMP and activation of protein kinase A. TGR5 is widely expressed in various tissues including the liver, gallbladder, bile ducts, adipose tissue, spleen, intestines, and kidneys. Within the liver, TGR5 is abundantly expressed in Kupffer cells and endothelial cells, but not in hepatocytes. The targets and effects of TGR5 are outlined in detail below and summarized in Figure 2.

### TGR5 functions and NAFLD

**Regulation of the bile acid pool:** TGR5 regulates the bile acid pool. The bile acid pool is significantly reduced in TGR (-/-) mice compared to wild-type mice. TGR5 also regulates bile composition as demonstrated by an experiment showing that, when fed with lithogenic diet, TGR (-/-) mice were protected from gallstone diseases. The expression of TGR5 was shown to be present in gallbladder epithelial cells and cholangiocytes, and TGR5 activation induced bicarbonate and chloride secretion from cholangiocytes, which may account for the alteration of bile composition.

**Modulation of the immune response:** TGR5 also modulates immune responses of immune cells *via* increasing intracellular cAMP<sup>[53]</sup>, and this function appears relevant to the regulation of hepatic inflammation and atherosclerosis development. TGR5 is highly expressed in monocytes and macrophages. Activation of TGR5 increases cAMP in rat alveolar macrophages and, as a result, it reduces the phagocytic activity of the macrophage and inhibits lipopolysaccharide (LPS)-induced production of pro-inflammatory cytokines such as tumor necrosis factor (TNF)- $\alpha$ , interleukin-1 (IL-1), IL-6 and IL-8. This finding was also demonstrated in resident hepatic macrophage Kupffer cells. Activation of TGR5 in isolated Kupffer cells causes an increase in cAMP and reduced expression of pro-inflammatory cytokines, including: TNF- $\alpha$ , IL-1, IL-6 and IL-8, following LPS treatment. Notably, TGR5-deficient mice are more susceptible to LPS-induced liver injury<sup>[55]</sup>. TGR5 activation also attenuates the formation of atheromatous plaque in low-density lipoprotein receptor knockout mice (LDL<sup>-/-</sup>), a commonly used murine model for atherosclerosis studies that have functioning TGR5 (LDL<sup>-/-</sup>; TGR5<sup>+/+</sup>). This attenuation of plaque formation was achieved through decreases in intra-plaque inflammation and macrophage activation<sup>[56]</sup>. Recent study has also showed that a TGR5 agonist increased the production of nitric oxide (NO) in endothelial cells, a key anti-atherogenic molecule. This suggests that NO might be one of downstream effectors of TGR5 signaling.

**Energy expenditure and metabolism:** TGR5 enhances energy expenditure and mitigates obesity and insulin resistance in obese mice. TGR5 is expressed in human brown adipocytes and skeletal myocytes. In brown adipose and skeletal muscle, interactions between bile acids and TGR5 promote the expression of cAMP-dependent 2-iodothyronine de-iodinase (D2), which converts inactive thyroxine (T4) to active 3,5,3-triiodothyronine (T3), a major hormone in increasing basal metabolism and thus, inducing energy expenditure<sup>[57]</sup>. TGR5 signaling also regulates glucose homeostasis<sup>[58]</sup>. TGR5 is expressed on enteroendocrine L-cells, and activation of TGR5 on L-cells modulates mitochondrial oxidative phosphorylation and alters ATP/ADP ratio. This leads to the release of glucagon like peptide-1 (GLP-1) from L-cells. GLP-1 further stimulates insulin secretion from pancreas and maintains glucose homeostasis.

### **TGR5 agonists in the treatment of NALFD**

Given the aforementioned biological effects of TGR5, TGR5 becomes an enticing potential target for NASH therapeutics. A specific CA derivative, 6 $\alpha$ -ethyl-23(S)-methylcholic acid (INT-777) has been developed as a selective TGR5 agonist<sup>[59]</sup>. Treatment of high-fat fed mice with INT-777 increased energy expenditure and attenuated both weight gain and expansion of fat pad mass<sup>[58]</sup>. In addition, INT-777 treatment reduced hepatic steatosis and improved liver enzyme levels without evidence of hepatic fibrosis<sup>[58]</sup>. INT-777 treatment has

also been shown to improve insulin sensitivity, likely through the release of GLP-1 in the intestines, in both diet-induced obese mice and in genetically obese mice that have a leptin receptor gene mutation (*db/db*), which is a well-established model of obesity and diabetes<sup>[58]</sup>. This effect was blunted in TGR5<sup>-/-</sup> mice, indicating the specificity of INT-777 treatment in targeting TGR5. With all these features, TGR5 agonists such as INT-777 are very attractive treatment candidates for NASH and other features of the metabolic syndrome<sup>[60]</sup>.

## **TARGETING BOTH FXR AND TGR5 IN THE TREATMENT OF NAFLD**

INT-767, the 23-sulphate derivative of OCA, is a dual FXR/TGR5 agonist<sup>[61]</sup>. INT-767 has been demonstrated to induce FXR-dependent lipid uptake by adipocytes, mobilizing lipid from the circulation and the liver to peripheral adipose tissue. INT-767 also promotes TGR5-dependant GLP-1 release. Treatment of obese mice with INT-767 significantly decreased total plasma cholesterol and triglyceride levels<sup>[61]</sup>, and improved the histological features of NASH in these mice<sup>[62]</sup>. These effects have been postulated to be due to INT-767-mediated alterations in the phenotypes of intrahepatic macrophage populations and modulation of cytokine production<sup>[62]</sup>. Uniquely, INT-767, but not INT-777 or INT-747, ameliorates hepatic injury in MDR2 (-/-) mice, a model for chronic cholangiopathy. This hepatoprotective effect is manifested by a reduction in bile acid synthesis and an increase in bile flow and biliary HCO<sub>3</sub><sup>-</sup> output<sup>[63]</sup>.

## **CONCLUSION**

Bile acids and bile acid receptors have pluripotent functions in energy expenditure, regulation of lipids and glucose metabolism, modulation of hepatic inflammation, fibrosis, regeneration, and carcinogenesis. These effects translate into attractive therapeutic targets for NASH and to improve metabolic profiles, ameliorate hepatic injury, and halt hepatic fibrosis. One currently very promising drug is OCA, but additional new drugs are expected in the not-too-distant future that will target the pathways in NASH pathogenesis.

## **REFERENCES**

- 1 **Wanless IR**, Lentz JS. Fatty liver hepatitis (steatohepatitis) and obesity: an autopsy study with analysis of risk factors. *Hepatology* 1990; **12**: 1106-1110 [PMID: 2227807 DOI: 10.1002/hep.1840120505]
- 2 **Williamson RM**, Price JF, Glancy S, Perry E, Nee LD, Hayes PC, Frier BM, Van Look LA, Johnston GI, Reynolds RM, Strachan MW. Prevalence of and risk factors for hepatic steatosis and nonalcoholic fatty liver disease in people with type 2 diabetes: the Edinburgh Type 2 Diabetes Study. *Diabetes Care* 2011; **34**: 1139-1144 [PMID: 21478462 DOI: 10.2337/dc10-2229]
- 3 **Gaggini M**, Morelli M, Buzzigoli E, DeFronzo RA, Bugianesi E, Gastaldelli A. Non-alcoholic fatty liver disease (NAFLD) and its connection with insulin resistance, dyslipidemia, atherosclerosis

- and coronary heart disease. *Nutrients* 2013; **5**: 1544-1560 [PMID: 23666091 DOI: 10.3390/nu5051544]
- 4 **Angulo P.** Long-term mortality in nonalcoholic fatty liver disease: is liver histology of any prognostic significance? *Hepatology* 2010; **51**: 373-375 [PMID: 20101746 DOI: 10.1002/hep.23521]
  - 5 **Vernon G,** Baranova A, Younossi ZM. Systematic review: the epidemiology and natural history of non-alcoholic fatty liver disease and non-alcoholic steatohepatitis in adults. *Aliment Pharmacol Ther* 2011; **34**: 274-285 [PMID: 21623852 DOI: 10.1111/j.1365-2036.2011.04724.x]
  - 6 **Charlton MR,** Burns JM, Pedersen RA, Watt KD, Heimbach JK, Dierkhising RA. Frequency and outcomes of liver transplantation for nonalcoholic steatohepatitis in the United States. *Gastroenterology* 2011; **141**: 1249-1253 [PMID: 21726509 DOI: 10.1053/j.gastro.2011.06.061]
  - 7 **Wong RJ,** Aguilar M, Cheung R, Perumpail RB, Harrison SA, Younossi ZM, Ahmed A. Nonalcoholic steatohepatitis is the second leading etiology of liver disease among adults awaiting liver transplantation in the United States. *Gastroenterology* 2015; **148**: 547-555 [PMID: 25461851 DOI: 10.1053/j.gastro.2014.11.039]
  - 8 **Day CP,** James OF. Steatohepatitis: a tale of two "hits"? *Gastroenterology* 1998; **114**: 842-845 [PMID: 9547102 DOI: 10.1016/S0016-5085(98)70599-2]
  - 9 **Tilg H,** Moschen AR. Evolution of inflammation in nonalcoholic fatty liver disease: the multiple parallel hits hypothesis. *Hepatology* 2010; **52**: 1836-1846 [PMID: 21038418 DOI: 10.1002/hep.24001]
  - 10 **Browning JD,** Horton JD. Molecular mediators of hepatic steatosis and liver injury. *J Clin Invest* 2004; **114**: 147-152 [PMID: 15254578 DOI: 10.1172/JCI22422]
  - 11 **Azzout-Marniche D,** Bécard D, Guichard C, Foretz M, Ferré P, Foufelle F. Insulin effects on sterol regulatory-element-binding protein-1c (SREBP-1c) transcriptional activity in rat hepatocytes. *Biochem J* 2000; **350** Pt 2: 389-393 [PMID: 10947952 DOI: 10.1042/bj3500389]
  - 12 **Fon Tacer K,** Rozman D. Nonalcoholic Fatty liver disease: focus on lipoprotein and lipid deregulation. *J Lipids* 2011; **2011**: 783976 [PMID: 21773052 DOI: 10.1155/2011/783976]
  - 13 **Farhadi A,** Gundlapalli S, Shaikh M, Frantzides C, Harrell L, Kwasny MM, Keshavarzian A. Susceptibility to gut leakiness: a possible mechanism for endotoxaemia in non-alcoholic steatohepatitis. *Liver Int* 2008; **28**: 1026-1033 [PMID: 18397235 DOI: 10.1111/j.1478-3231.2008.01723.x]
  - 14 **Yang SQ,** Lin HZ, Lane MD, Clemens M, Diehl AM. Obesity increases sensitivity to endotoxin liver injury: implications for the pathogenesis of steatohepatitis. *Proc Natl Acad Sci USA* 1997; **94**: 2557-2562 [PMID: 9122234 DOI: 10.1073/pnas.94.6.2557]
  - 15 **Saberi M,** Woods NB, de Luca C, Schenk S, Lu JC, Bandyopadhyay G, Verma IM, Olefsky JM. Hematopoietic cell-specific deletion of toll-like receptor 4 ameliorates hepatic and adipose tissue insulin resistance in high-fat-fed mice. *Cell Metab* 2009; **10**: 419-429 [PMID: 19883619 DOI: 10.1016/j.cmet.2009.09.006]
  - 16 **Rivera CA,** Adegboyega P, van Rooijen N, Tagalicud A, Allman M, Wallace M. Toll-like receptor-4 signaling and Kupffer cells play pivotal roles in the pathogenesis of non-alcoholic steatohepatitis. *J Hepatol* 2007; **47**: 571-579 [PMID: 17644211 DOI: 10.1016/j.jhep.2007.04.019]
  - 17 **Miura K,** Kodama Y, Inokuchi S, Schnabl B, Aoyama T, Ohnishi H, Olefsky JM, Brenner DA, Seki E. Toll-like receptor 9 promotes steatohepatitis by induction of interleukin-1beta in mice. *Gastroenterology* 2010; **139**: 323-34.e7 [PMID: 20347818 DOI: 10.1053/j.gastro.2010.03.052]
  - 18 **Evans RM.** The steroid and thyroid hormone receptor superfamily. *Science* 1988; **240**: 889-895 [PMID: 3283939 DOI: 10.1126/science.3283939]
  - 19 **Giguère V,** Yang N, Segui P, Evans RM. Identification of a new class of steroid hormone receptors. *Nature* 1988; **331**: 91-94 [PMID: 3267207 DOI: 10.1038/331091a0]
  - 20 **Forman BM,** Goode E, Chen J, Oro AE, Bradley DJ, Perlmann T, Noonan DJ, Burka LT, McMorris T, Lamph WW, Evans RM, Weinberger C. Identification of a nuclear receptor that is activated by farnesol metabolites. *Cell* 1995; **81**: 687-693 [PMID: 7774010 DOI: 10.1016/0092-8674(95)90530-8]
  - 21 **Makishima M,** Okamoto AY, Repa JJ, Tu H, Learned RM, Luk A, Hull MV, Lustig KD, Mangelsdorf DJ, Shan B. Identification of a nuclear receptor for bile acids. *Science* 1999; **284**: 1362-1365 [PMID: 10334992 DOI: 10.1126/science.284.5418.1362]
  - 22 **Wang H,** Chen J, Hollister K, Sowers LC, Forman BM. Endogenous bile acids are ligands for the nuclear receptor FXR/BAR. *Mol Cell* 1999; **3**: 543-553 [PMID: 10360171 DOI: 10.1016/S1097-2765(00)80348-2]
  - 23 **Goodwin B,** Jones SA, Price RR, Watson MA, McKee DD, Moore LB, Galardi C, Wilson JG, Lewis MC, Roth ME, Maloney PR, Willson TM, Kliewer SA. A regulatory cascade of the nuclear receptors FXR, SHP-1, and LXR-1 represses bile acid biosynthesis. *Mol Cell* 2000; **6**: 517-526 [PMID: 11030332 DOI: 10.1016/S1097-2765(00)00051-4]
  - 24 **Wang L,** Lee YK, Bundman D, Han Y, Thevananther S, Kim CS, Chua SS, Wei P, Heyman RA, Karin M, Moore DD. Redundant pathways for negative feedback regulation of bile acid production. *Dev Cell* 2002; **2**: 721-731 [PMID: 12062085 DOI: 10.1016/S1534-5807(02)00187-9]
  - 25 **Kerr TA,** Saeki S, Schneider M, Schaefer K, Berdy S, Redder T, Shan B, Russell DW, Schwarz M. Loss of nuclear receptor SHP impairs but does not eliminate negative feedback regulation of bile acid synthesis. *Dev Cell* 2002; **2**: 713-720 [PMID: 12062084]
  - 26 **Holt JA,** Luo G, Billin AN, Bisi J, McNeill YY, Kozarsky KF, Donahee M, Wang DY, Mansfield TA, Kliewer SA, Goodwin B, Jones SA. Definition of a novel growth factor-dependent signal cascade for the suppression of bile acid biosynthesis. *Genes Dev* 2003; **17**: 1581-1591 [PMID: 12815072 DOI: 10.1101/gad.1083503]
  - 27 **Hwang ST,** Urizar NL, Moore DD, Henning SJ. Bile acids regulate the ontogenic expression of ileal bile acid binding protein in the rat via the farnesoid X receptor. *Gastroenterology* 2002; **122**: 1483-1492 [PMID: 11984532]
  - 28 **Claudel T,** Staels B, Kuipers F. The Farnesoid X receptor: a molecular link between bile acid and lipid and glucose metabolism. *Arterioscler Thromb Vasc Biol* 2005; **25**: 2020-2030 [PMID: 16037564 DOI: 10.1161/01.ATV.0000178994.21828.a7]
  - 29 **Schreuder TC,** Marsman HA, Lenicek M, van Werven JR, Nederveen AJ, Jansen PL, Schaap FG. The hepatic response to FGF19 is impaired in patients with nonalcoholic fatty liver disease and insulin resistance. *Am J Physiol Gastrointest Liver Physiol* 2010; **298**: G440-G445 [PMID: 20093562 DOI: 10.1152/ajpgi.00322.2009]
  - 30 **Urizar NL,** Dowhan DH, Moore DD. The farnesoid X-activated receptor mediates bile acid activation of phospholipid transfer protein gene expression. *J Biol Chem* 2000; **275**: 39313-39317 [PMID: 10998425 DOI: 10.1074/jbc.M007998200]
  - 31 **Claudel T,** Inoue Y, Barbier O, Duran-Sandoval D, Kosykh V, Fruchart J, Fruchart JC, Gonzalez FJ, Staels B. Farnesoid X receptor agonists suppress hepatic apolipoprotein CIII expression. *Gastroenterology* 2003; **125**: 544-555 [PMID: 12891557]
  - 32 **Claudel T,** Sturm E, Duez H, Torra IP, Sirvent A, Kosykh V, Fruchart JC, Dallongeville J, Hum DW, Kuipers F, Staels B. Bile acid-activated nuclear receptor FXR suppresses apolipoprotein A-I transcription via a negative FXR response element. *J Clin Invest* 2002; **109**: 961-971 [PMID: 11927623 DOI: 10.1172/JCI14505]
  - 33 **Watanabe M,** Houten SM, Wang L, Moschetta A, Mangelsdorf DJ, Heyman RA, Moore DD, Auwerx J. Bile acids lower triglyceride levels via a pathway involving FXR, SHP, and SREBP-1c. *J Clin Invest* 2004; **113**: 1408-1418 [PMID: 15146238 DOI: 10.1172/JCI21025]
  - 34 **Miyata M,** Sakaida Y, Matsuzawa H, Yoshinari K, Yamazoe Y. Fibroblast growth factor 19 treatment ameliorates disruption of hepatic lipid metabolism in farnesoid X receptor (Fxr)-null mice. *Biol Pharm Bull* 2011; **34**: 1885-1889 [PMID: 22130247]
  - 35 **Ma K,** Saha PK, Chan L, Moore DD. Farnesoid X receptor is essential for normal glucose homeostasis. *J Clin Invest* 2006; **116**: 1102-1109 [PMID: 16557297 DOI: 10.1172/JCI25604]
  - 36 **De Fabiani E,** Mitro N, Gilardi F, Caruso D, Galli G, Crestani

- M. Coordinated control of cholesterol catabolism to bile acids and of gluconeogenesis via a novel mechanism of transcription regulation linked to the fasted-to-fed cycle. *J Biol Chem* 2003; **278**: 39124-39132 [PMID: 12865425 DOI: 10.1074/jbc.M305079200]
- 37 **Duran-Sandoval D**, Mautino G, Martin G, Percevault F, Barbier O, Fruchart JC, Kuipers F, Staels B. Glucose regulates the expression of the farnesoid X receptor in liver. *Diabetes* 2004; **53**: 890-898 [PMID: 15047603 DOI: 10.2337/diabetes.53.4.890]
- 38 **Yamagata K**, Daitoku H, Shimamoto Y, Matsuzaki H, Hirota K, Ishida J, Fukamizu A. Bile acids regulate gluconeogenic gene expression via small heterodimer partner-mediated repression of hepatocyte nuclear factor 4 and Foxo1. *J Biol Chem* 2004; **279**: 23158-23165 [PMID: 15047713 DOI: 10.1074/jbc.M314322200]
- 39 **Kim JY**, Kim HJ, Kim KT, Park YY, Seong HA, Park KC, Lee IK, Ha H, Shong M, Park SC, Choi HS. Orphan nuclear receptor small heterodimer partner represses hepatocyte nuclear factor 3/Foxa transactivation via inhibition of its DNA binding. *Mol Endocrinol* 2004; **18**: 2880-2894 [PMID: 15358835 DOI: 10.1210/me.2004-0211]
- 40 **Potthoff MJ**, Boney-Montoya J, Choi M, He T, Sunny NE, Satapati S, Suino-Powell K, Xu HE, Gerard RD, Finck BN, Burgess SC, Mangelsdorf DJ, Kliewer SA. FGF15/19 regulates hepatic glucose metabolism by inhibiting the CREB-PGC-1 $\alpha$  pathway. *Cell Metab* 2011; **13**: 729-738 [PMID: 21641554 DOI: 10.1016/j.cmet.2011.03.019]
- 41 **Fiorucci S**, Antonelli E, Rizzo G, Renga B, Mencarelli A, Riccardi L, Orlandi S, Pellicciari R, Morelli A. The nuclear receptor SHP mediates inhibition of hepatic stellate cells by FXR and protects against liver fibrosis. *Gastroenterology* 2004; **127**: 1497-1512 [PMID: 15521018 DOI: 10.1053/j.gastro.2004.08.001]
- 42 **Fickert P**, Fuchsichler A, Moustafa T, Wagner M, Zollner G, Halilbasic E, Stöger U, Arrese M, Pizarro M, Solis N, Carrasco G, Caligiuri A, Sombetzki M, Reisinger E, Tsybrovskyy O, Zatloukal K, Denk H, Jaeschke H, Pinzani M, Trauner M. Farnesoid X receptor critically determines the fibrotic response in mice but is expressed to a low extent in human hepatic stellate cells and periductal myofibroblasts. *Am J Pathol* 2009; **175**: 2392-2405 [PMID: 19910507 DOI: 10.2353/ajpath.2009.090114]
- 43 **Uriarte I**, Fernandez-Barrena MG, Monte MJ, Latasa MU, Chang HC, Carotti S, Vespasiani-Gentilucci U, Morini S, Vicente E, Concepcion AR, Medina JF, Marin JJ, Berasain C, Prieto J, Avila MA. Identification of fibroblast growth factor 15 as a novel mediator of liver regeneration and its application in the prevention of post-resection liver failure in mice. *Gut* 2013; **62**: 899-910 [PMID: 23292666 DOI: 10.1136/gutjnl-2012-302945]
- 44 **Zhang Y**, Ge X, Heemstra LA, Chen WD, Xu J, Smith JL, Ma H, Kasim N, Edwards PA, Novak CM. Loss of FXR protects against diet-induced obesity and accelerates liver carcinogenesis in ob/ob mice. *Mol Endocrinol* 2012; **26**: 272-280 [PMID: 22261820 DOI: 10.1210/me.2011-1157]
- 45 **Kim I**, Morimura K, Shah Y, Yang Q, Ward JM, Gonzalez FJ. Spontaneous hepatocarcinogenesis in farnesoid X receptor-null mice. *Carcinogenesis* 2007; **28**: 940-946 [PMID: 17183066 DOI: 10.1093/carcin/bgl249]
- 46 **Deuschle U**, Schüler J, Schulz A, Schlüter T, Kinzel O, Abel U, Kremoser C. FXR controls the tumor suppressor NDRG2 and FXR agonists reduce liver tumor growth and metastasis in an orthotopic mouse xenograft model. *PLoS One* 2012; **7**: e43044 [PMID: 23056173 DOI: 10.1371/journal.pone.0043044]
- 47 **Jiang Y**, Iakova P, Jin J, Sullivan E, Sharin V, Hong IH, Anakk S, Mayor A, Darlington G, Finegold M, Moore D, Timchenko NA. Farnesoid X receptor inhibits gankyrin in mouse livers and prevents development of liver cancer. *Hepatology* 2013; **57**: 1098-1106 [PMID: 23172628 DOI: 10.1002/hep.26146]
- 48 **Degrolamo C**, Modica S, Vacca M, Di Tullio G, Morgano A, D'Orazio A, Kannisto K, Parini P, Moschetta A. Prevention of spontaneous hepatocarcinogenesis in farnesoid X receptor-null mice by intestinal-specific farnesoid X receptor reactivation. *Hepatology* 2015; **61**: 161-170 [PMID: 24954587 DOI: 10.1002/hep.27274]
- 49 **Pellicciari R**, Fiorucci S, Camaioni E, Clerici C, Costantino G, Maloney PR, Morelli A, Parks DJ, Willson TM. 6 $\alpha$ -ethyl-chenodeoxycholic acid (6-ECDCA), a potent and selective FXR agonist endowed with anticholestatic activity. *J Med Chem* 2002; **45**: 3569-3572 [PMID: 12166927 DOI: 10.1021/jm025529g]
- 50 **Cipriani S**, Mencarelli A, Palladino G, Fiorucci S. FXR activation reverses insulin resistance and lipid abnormalities and protects against liver steatosis in Zucker (fa/fa) obese rats. *J Lipid Res* 2010; **51**: 771-784 [PMID: 19783811 DOI: 10.1194/jlr.M001602]
- 51 **Verbeke L**, Farre R, Trebicka J, Komuta M, Roskams T, Klein S, Elst IV, Windmolders P, Vanuysel T, Nevens F, Laleman W. Obeticholic acid, a farnesoid X receptor agonist, improves portal hypertension by two distinct pathways in cirrhotic rats. *Hepatology* 2014; **59**: 2286-2298 [PMID: 24259407 DOI: 10.1002/hep.26939]
- 52 **Neuschwander-Tetri BA**, Loomba R, Sanyal AJ, Lavine JE, Van Natta ML, Abdelmalek MF, Chalasani N, Dasarathy S, Diehl AM, Hameed B, Kowdley KV, McCullough A, Terrault N, Clark JM, Tonascia J, Brunt EM, Kleiner DE, Doo E. Farnesoid X nuclear receptor ligand obeticholic acid for non-cirrhotic, non-alcoholic steatohepatitis (FLINT): a multicentre, randomised, placebo-controlled trial. *Lancet* 2015; **385**: 956-965 [PMID: 25468160 DOI: 10.1016/S0140-6736(14)61933-4]
- 53 **Kawamata Y**, Fujii R, Hosoya M, Harada M, Yoshida H, Miwa M, Fukusumi S, Habata Y, Itoh T, Shintani Y, Hinuma S, Fujisawa Y, Fujino M. A G protein-coupled receptor responsive to bile acids. *J Biol Chem* 2003; **278**: 9435-9440 [PMID: 12524422 DOI: 10.1074/jbc.M209706200]
- 54 **Takeda S**, Kadowaki S, Haga T, Takaesu H, Mitaku S. Identification of G protein-coupled receptor genes from the human genome sequence. *FEBS Lett* 2002; **520**: 97-101 [PMID: 12044878 DOI: 10.1016/S0014-5793(02)02775-8]
- 55 **Wang YD**, Chen WD, Yu D, Forman BM, Huang W. The G-protein-coupled bile acid receptor, Gpbar1 (TGR5), negatively regulates hepatic inflammatory response through antagonizing nuclear factor  $\kappa$  light-chain enhancer of activated B cells (NF- $\kappa$ B) in mice. *Hepatology* 2011; **54**: 1421-1432 [PMID: 21735468]
- 56 **Pols TW**, Nomura M, Harach T, Lo Sasso G, Oosterveer MH, Thomas C, Rizzo G, Gioiello A, Adorini L, Pellicciari R, Auwerx J, Schoonjans K. TGR5 activation inhibits atherosclerosis by reducing macrophage inflammation and lipid loading. *Cell Metab* 2011; **14**: 747-757 [PMID: 22152303 DOI: 10.1016/j.cmet.2011.11.006]
- 57 **Watanabe M**, Houten SM, Matakai C, Christoffolete MA, Kim BW, Sato H, Messaddeq N, Harney JW, Ezaki O, Kodama T, Schoonjans K, Bianco AC, Auwerx J. Bile acids induce energy expenditure by promoting intracellular thyroid hormone activation. *Nature* 2006; **439**: 484-489 [PMID: 16400329 DOI: 10.1038/nature04330]
- 58 **Thomas C**, Gioiello A, Noriega L, Strehle A, Oury J, Rizzo G, Macchiarulo A, Yamamoto H, Matakai C, Pruzanski M, Pellicciari R, Auwerx J, Schoonjans K. TGR5-mediated bile acid sensing controls glucose homeostasis. *Cell Metab* 2009; **10**: 167-177 [PMID: 19723493 DOI: 10.1016/j.cmet.2009.08.001]
- 59 **Pellicciari R**, Gioiello A, Macchiarulo A, Thomas C, Rosatelli E, Natalini B, Sardella R, Pruzanski M, Roda A, Pastorini E, Schoonjans K, Auwerx J. Discovery of 6 $\alpha$ -ethyl-23(S)-methylcholic acid (S-EMCA, INT-777) as a potent and selective agonist for the TGR5 receptor, a novel target for diabetes. *J Med Chem* 2009; **52**: 7958-7961 [PMID: 20014870 DOI: 10.1021/jm901390p]
- 60 **Tiwari A**, Maiti P. TGR5: an emerging bile acid G-protein-coupled receptor target for the potential treatment of metabolic disorders. *Drug Discov Today* 2009; **14**: 523-530 [PMID: 19429513 DOI: 10.1016/j.drudis.2009.02.005]
- 61 **Rizzo G**, Passeri D, De Franco F, Ciaccioli G, Donadio L, Rizzo G, Orlandi S, Sadeghpour B, Wang XX, Jiang T, Levi M, Pruzanski M, Adorini L. Functional characterization of the semisynthetic bile acid derivative INT-767, a dual farnesoid X receptor and TGR5 agonist. *Mol Pharmacol* 2010; **78**: 617-630 [PMID: 20631053 DOI: 10.1124/mol.110.064501]
- 62 **McMahan RH**, Wang XX, Cheng LL, Krisko T, Smith M, El Kasmi K, Pruzanski M, Adorini L, Golden-Mason L, Levi M, Rosen HR. Bile acid receptor activation modulates hepatic

monocyte activity and improves nonalcoholic fatty liver disease.  
*J Biol Chem* 2013; **288**: 11761-11770 [PMID: 23460643 DOI:  
10.1074/jbc.M112.446575]

- 63 **Baghdasaryan A**, Claudel T, Gumhold J, Silbert D, Adorini L,  
Roda A, Vecchiotti S, Gonzalez FJ, Schoonjans K, Strazzabosco

M, Fickert P, Trauner M. Dual farnesoid X receptor/TGR5  
agonist INT-767 reduces liver injury in the Mdr2<sup>-/-</sup> (Abcb4<sup>-/-</sup>)  
mouse cholangiopathy model by promoting biliary HCO<sub>3</sub><sup>-</sup> output.  
*Hepatology* 2011; **54**: 1303-1312 [PMID: 22006858 DOI: 10.1002/  
hep.24537]

**P- Reviewer:** Garcia-Ruiz I, Kayadibi H, Tomizawa M  
**S- Editor:** Kong JX **L- Editor:** A **E- Editor:** Liu SQ



## Treating morbid obesity in cirrhosis: A quest of holy grail

Naveen Kumar, Narendra Singh Choudhary

Naveen Kumar, Narendra Singh Choudhary, Department of Transplant Hepatology, Medanta Liver Institute, Medanta the Medicity, Gurgaon 122001, Haryana, India

**Author contributions:** Kumar N and Choudhary NS both wrote the paper.

**Conflict-of-interest statement:** The authors declare no conflict of interest for this article.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to:** Dr. Narendra Singh Choudhary, Consultant, Department of Transplant Hepatology, Medanta Liver Institute, Medanta the Medicity, CH Bakhtawar Singh Road, Gurgaon 122001, Haryana, India. [docnarendra@gmail.com](mailto:docnarendra@gmail.com)  
Telephone: +91-124-4141414  
Fax: +91-124-4834111

Received: June 28, 2015  
Peer-review started: July 5, 2015  
First decision: July 28, 2015  
Revised: October 13, 2015  
Accepted: November 17, 2015  
Article in press: November 25, 2015  
Published online: December 8, 2015

### Abstract

The problem of obesity is increasing worldwide in epidemic proportions; the situation is similarly becoming more common in patients with cirrhosis which negatively affect the prognosis of disease and also makes liver transplantation difficult especially in the living donor liver transplantation setting where low graft to recipient

weight ratio negatively affects survival. Treatment of obesity is difficult in cirrhosis due to difficulty in implementation of lifestyle measures, limited data on safety of anti-obesity drugs and high risk of surgery. Currently approved anti-obesity drugs have limited data in patients with cirrhosis. Bariatric surgery remains an option in selected compensated cirrhotic patients. Endoscopic interventions for obesity are emerging and are quite promising in patients with cirrhosis as these are minimally invasive. In present review, we briefly discuss various modalities of weight reduction in obese patients and their applicability in patients with cirrhosis.

**Key words:** Obesity; Intra-gastric balloon; Antiobesity drugs; Bariatric surgery; Cirrhosis

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** The rising obesity problem is also associated with increased incidence of simultaneous obesity and cirrhosis. This is a particularly difficult subset of obese patients to treat as there is difficulty in implementation of lifestyle measures, limited data on safety of anti-obesity drugs and high risk of surgery. In present review, we briefly discuss various modalities of weight reduction in obese patients and their applicability in patients with cirrhosis.

Kumar N, Choudhary NS. Treating morbid obesity in cirrhosis: A quest of holy grail. *World J Hepatol* 2015; 7(28): 2819-2828 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i28/2819.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i28.2819>

### INTRODUCTION

Obesity has been defined as abnormal or excessive fat accumulation that can lead to impairment of health. It's one of the most significant public health problems faced by people of industrialized countries and is rapidly

catching up in developing countries also. Worldwide obesity prevalence has almost doubled since 1980<sup>[1]</sup>. Obesity has reached epidemic proportions over the world and it is simultaneously associated with various comorbidities, namely diabetes mellitus, hypertension, and cardiac diseases<sup>[2]</sup>. In 2014, according to estimate more than 1.9 billion adults were overweight with 600 million likely obese. Approximately 39% of adults aged 18 years and above were overweight in 2014, and 13% were obese<sup>[3]</sup>. Due to various co-morbidities, obesity represents a very serious health problem worldwide. Obesity management is a unique challenge due to the rapid evolution of unfavourable lifestyles<sup>[4]</sup>.

Obesity can be associated with cirrhosis as a virtue of non-alcoholic steatohepatitis (NASH), an important cause of cirrhosis, being a component of the metabolic syndrome and it can also exacerbate co-existing liver injury due to other causes and is associated with more risk of decompensation of cirrhosis<sup>[5]</sup>. The pathophysiology of NASH has been considered a "two hit" process<sup>[6]</sup>. The "first hit", hepatic steatosis, makes the liver susceptible to injury mediated by "second hits", like inflammatory cytokines/adipokines, oxidative stress, and mitochondrial dysfunction, leading to steatohepatitis and/or fibrosis<sup>[7]</sup>. Impaired hepatocyte proliferation progenitors due to cell death has been proposed as "third hit" in pathogenesis of non-alcoholic fatty liver disease<sup>[8]</sup>. Various cytokines/adipokines involved in NASH pathogenesis includes tumor necrosis factor- $\alpha$ , leptin, adiponectin, interleukin-6 (IL-6), *etc.*<sup>[9]</sup>. Fibrosis/cirrhosis represents the final common endpoint of pathway of almost all chronic liver diseases including NASH. Mechanisms for fibrosis include the secretion of profibrogenic cytokines (tumor growth factor- $\beta$ , IL-6, IL-8, *etc.*) by the ductular reaction, as well as epithelial to mesenchymal transition of stellate cells to myofibroblasts<sup>[10,11]</sup>. Steatosis is very commonly associated with hepatitis C, particularly with genotype 3. In chronic hepatitis C, obesity is associated with inflammation, steatosis, insulin resistance, faster progression of fibrosis, and nonresponse to treatment with interferon<sup>[12]</sup>.

## OBESITY AND CIRRHOSIS: THE CHALLENGES AND RATIONALE FOR MANAGEMENT

Obesity with cirrhosis is a complex problem. Once cirrhosis is decompensated, lifestyle measures are very difficult to implement and bariatric surgery becomes risky due to increased morbidity and mortality<sup>[13,14]</sup>. Pharmacological measures (drugs) have a very limited role in management of obesity, are not as effective as surgery and there is rebound weight gain once stopped. The limited drug arsenal available to treat obesity is not well studied in patients with liver disease. No safe anti-obesity drug in cirrhosis is available at the moment. Proportion of patients with NASH associated end stage liver disease as an indication for liver transplantation is increasing

gradually<sup>[15,16]</sup> and these patients are more prone for comorbidities associated with NASH like coronary artery disease, diabetes, hypertension, dyslipidemia, metabolic syndrome and chronic kidney disease<sup>[17]</sup>. Obesity in cirrhosis becomes a multi-headed monster leading to more rapid worsening of liver disease and also makes liver transplantation difficult. There is difficulty in finding a suitable donor for morbidly obese patients due to risk of low graft to recipient ratio and subsequent risk of poor graft function and higher mortality in living donor liver transplantation (LDLT) programs which predominant from of liver transplantation in Asia<sup>[18]</sup>. Increased rates of complications and mortality, as well as decreased graft survival, have been reported in morbidly obese patients often discouraging transplantation in this population and have resulted in the exclusion of morbidly obese patients from liver transplantation at some centres<sup>[19]</sup>. With increasing number of non-alcoholic steatohepatitis associated end stage liver disease as an indication for liver transplantation, problem of morbid obesity before liver transplantation is going to rise<sup>[20]</sup>.

There are multiple benefits of treating obesity in patients with cirrhosis. Firstly there is a reduction in risk of decompensation as studies have shown higher decompensation over time in overweight and obese cirrhotic<sup>[5]</sup>. Some patients may improve from compensated stage to lesser degree of fibrosis as shown by bariatric surgery studies<sup>[21]</sup>. This may avoid need for liver transplantation in many patients. Secondly reduction in weight may improve their candidacy for liver transplantation by improving co-morbidities (like diabetes control), decreasing risk in surgery and improving their graft recipient weight ratio especially in LDLT settings. Thirdly there can be reduced incidence of hepatocellular carcinoma (HCC) as obesity is considered to be an independent risk factor for development of HCC<sup>[22]</sup>.

In current review article we will discuss the various weight reductions strategies in brief and their applicability in patients with cirrhosis.

## DIET, LIFE STYLE MODIFICATION AND EXERCISE IN THE MANAGEMENT OF OBESITY IN CIRRHOSIS

The recent worldwide increase in the population of obese individual is also seen in liver cirrhosis patients<sup>[23]</sup>. At present, in liver cirrhosis due to alcohol and chronic hepatitis C infection, nutritional intake is a spectrum ranging from being either sufficient or excessive<sup>[24,25]</sup>. Excessive nutrients has to be assessed in every patient, and the various nutritional parameters, like serum albumin and lean body mass, should be evaluated for the appropriate nutritional therapy. However, the amount of body weight reduction has not been evaluated properly in the obese liver cirrhosis patients. The addition of oral Branched chain amino acids (BCAA) granules to diet has been shown to reduce the incidence of HCC<sup>[23]</sup>. It has also been shown that oral BCAA supplementation

increases serum albumin levels<sup>[26]</sup>. The mechanism involved may be improved insulin sensitivity in muscle, increase in and reduced oxidative stress<sup>[27]</sup>. Thus, in obese liver cirrhosis patients, oral BCAA treatment is recommended in addition to correction of nutritional intake. The current epidemic of global obesity has created a new entity: The unique combination of sarcopenia and obesity, now commonly described as sarcopenic obesity<sup>[28]</sup>. A recent study has shown that sarcopenic obesity is more closely linked with insulin resistance than either sarcopenia or obesity alone<sup>[29]</sup>.

Physical activity levels and also exercise capacity are generally lower in liver cirrhosis patients than in healthy controls<sup>[30,31]</sup>. Exercise is a key component of management of liver cirrhosis patients because it leads to increased calorie burning, increased skeletal muscle mass, along with exercise capacity, leading to improved quality of life. The advice regarding exercise is made complex in patients with cirrhosis as portal pressure has been shown to increase with moderate exercise (up to 30% of the maximum), which poses a risk for variceal bleeding<sup>[32]</sup>. The optimal exercise regimen in liver cirrhosis patients remains uncertain. Researcher's recommendation is walking 5000 or more steps every day with a caloric intake of 30 kcal/kg based on a survey done on compensated cirrhosis patients<sup>[31]</sup>. A randomized pilot study involving liver cirrhosis patients, mostly Child-Pugh A cirrhosis, examined the effect of exercise combined with leucine supplementation (10 g/d). The program included three sessions every week of one hour treadmill along-with cycle ergometry at 60%-70% of the maximum heart rate, over a total period of 12 wk. The intervention group had improved exercise capacity, shown by the 6-min walk test and the 2-min step test with associated improvement in quality of life parameters with no adverse events<sup>[33]</sup>. Aerobic exercise is expected to improve insulin resistance in patients with cirrhosis which is particularly important for obese patients<sup>[34,35]</sup>. Future studies will establish efficacious along with safe exercise regimen needed for liver cirrhosis patients.

## DRUG THERAPY

### **Orlistat**

Orlistat at a dose of 120 mg was approved by the Food and Drug Administration (FDA) in 1999 for the management of obesity in association with reduced calorie diet, and also to reduce the risk of regaining weight after previous weight loss. Orlistat was the first treatment for obesity that was not an appetite suppressant, but acted by interfering with the action of hormone lipase involved in fat digestion<sup>[36]</sup>. In one of the longest trials comprising of 3304 patients, 21% also having impaired glucose tolerance, were randomized to receive either placebo or orlistat. During the first year, weight loss was greater in the orlistat-treated group (11% compared with 6% in the placebo group)<sup>[37]</sup>. Despite being FDA-approved fewer than 10% patients take it for 1 year and less than 2% of patients for 2 years due

to poor compliance secondarily to side effects<sup>[38,39]</sup>. It is advisable to give vitamin supplements to patients treated with this drug. Severe liver injury has been reported rarely with a United States FDA review identifying 13 reports of severe liver damage<sup>[40]</sup>. Given the side effect profile it is unlikely to become a commonly prescribed drug in cirrhosis patients who may have malnutrition despite obesity.

### **Lorcaserin**

Lorcaserin is a selective agonist of 5-hydroxytryptamine receptor 2C (5-HT<sub>2C</sub>), which is expressed in hypothalamic pro-opiomelanocortin (POMC)-producing neurons of central nervous system, the centre controlling appetite and satiety<sup>[41]</sup>. Lorcaserin causes activation of the 5-HT<sub>2C</sub> receptors which stimulates release of melanotropin- $\alpha$ , subsequently decreasing appetite through stimulation of melanocortin receptor 4<sup>[41]</sup>. Of significance is the low affinity lorcaserin has for other 5-hydroxytryptamine receptor subtypes, especially 5-HT<sub>2B</sub>, which has previously been associated with the development of valvular heart disease. Lorcaserin approval by FDA was largely based on two placebo-controlled trials in nondiabetic patients (BLOOM and BLOSSOM) along with a third smaller trial in adults with diabetes (BLOOM-DM)<sup>[42-44]</sup>. Lorcaserin caused a modest weight reduction of approximately 3.2 kg more than placebo. Adverse effects include headache, nausea, fatigue, and dizziness<sup>[45]</sup>. Lorcaserin should be discontinued if there is less than 5% weight reduction in 12 wk. No dose adjustment is required in patients with mild to moderate hepatic impairment. It has not been studied in patients with severe hepatic impairment and is not recommended in these groups of patients.

### **Phentermine/topiramate-extended-release**

In 2012, the United States FDA approved a preparation of phentermine and extended-release topiramate for use in adults with a body mass index (BMI)  $\geq 30$  kg/m<sup>2</sup> or with BMI  $\geq 27$  kg/m<sup>2</sup> with associated comorbidity (hypertension, diabetes, dyslipidemia). Phentermine plus topiramate-extended-release (ER) was recommended for approval based largely on two phase 3 clinical trials (EQUIP and CONQUER)<sup>[46,47]</sup>. In the EQUIP trial ( $n = 1267$ ) participants given the top dose vs placebo, the mean 1-year weight loss was 10.9% vs 1.6%<sup>[46]</sup>. In CONQUER trial ( $n = 2487$ ) one-year mean weight loss was 8.1 kg (7.8%) with the recommended dose and 10.2 kg (9.8%) with the top dose vs 1.4 kg (1.2%) with placebo<sup>[47]</sup>.

The labelling recommends against prescription in patients with recent or unstable cardiac or cerebrovascular disease, and suggests regular monitoring of resting heart rate. No dose adjustment is needed in patients with mild hepatic impairment. In patients with moderate hepatic impairment, the maximum dose is Phentermine/topiramate-ER 7.5 mg/46 mg once daily. Phentermine/topiramate-ER is not studied in patients with severe hepatic impairment where it has to be

**Table 1 Newer weight reduction drugs in pipeline**

Drug	Mechanism of action
Cetilistat	Gastrointestinal and pancreatic lipase inhibitor
Velneperit	Neuropeptide Y5 receptor inhibitor, appetite suppression
Tesofensine	Inhibition of serotonin, dopamine, and noradrenaline reuptake
Metreleptin	Leptin receptor agonist
Obinipitide	Dual neuropeptide Y2/Y4 receptor agonist
Beloranib	MetAP2 inhibition

MetAP2: Methionine aminopeptidase 2.

avoided<sup>[48]</sup>.

### **Bupropion-naltrexone**

In September 2014, a sustained release formulation of bupropion-naltrexone was approved by FDA<sup>[49]</sup>. Bupropion activates POMC neurons in the hypothalamus which gives downstream effects of appetite reduction and increased energy output. The POMC is regulated by endogenous opioids *via* opioid-mediated negative feedback. Naltrexone is a pure opioid antagonist, which further augments bupropion's activation of the POMC.

In a randomized trial of bupropion and naltrexone (varying doses) vs double placebo, weight loss was greater in those assigned to active treatment (mean change in body weight in low dose naltrexone and high dose naltrexone was -5% and -6.1% vs -1.3% in placebo arm)<sup>[50]</sup>. Compared with placebo, the combination of bupropion-naltrexone has been shown to reduce weight by approximately 4% to 5%<sup>[50-53]</sup>. Contraindications include uncontrolled hypertension, seizure history, eating disorders, simultaneously using other bupropion-containing products, chronic opioid use, and monoamine oxidase inhibitors use within last 14 d. Cases of hepatitis and clinically significant liver dysfunction have been seen in association with naltrexone use during naltrexone clinical trials and in post marketing reports of naltrexone use<sup>[54]</sup>. Thus the combination of bupropion-naltrexone doesn't look too exciting for the patient of cirrhosis and in the absence of strong data for liver disease patients, shouldn't be prescribed.

### **Liraglutide**

Liraglutide, is a long-acting glucagon-like peptide-1 analog, and a promising option for obese patients with type 2 diabetes. It is the most recent drug to be approved for obesity by FDA in December 2014. In diabetes trials, liraglutide (1.8 mg daily) was associated with a greater reduction in weight (2.0 to 2.5 kg) when compared with placebo or glimepiride<sup>[55]</sup>. In a randomized trial comparing liraglutide (1.2 to 3 mg), placebo, and open-label orlistat (120 mg orally three times daily) in 564 patients with a mean BMI of 35, weight loss increased with increasing doses of liraglutide, with the mean weight loss ranging from 4.8 to 7.2 kg<sup>[56]</sup>. Patients who were randomly assigned to receive any dose of liraglutide were found to lose significantly more

weight compared to placebo (mean weight loss 2.8 kg). Patients taking the two highest doses of liraglutide (2.4 and 3.0 mg) lost significantly more weight than those assigned to orlistat (6.3, 7.2 and 4.1 kg, respectively)<sup>[56]</sup>. In a 56-wk SCALE Maintenance randomized study trial comparing liraglutide 3 mg once daily with placebo injection in 422 patients a greater proportion of patients maintained weight loss in the liraglutide group (81.4%) vs 48.9% in placebo group<sup>[57]</sup>. Common side effects included nausea (37%-47%), vomiting (12%-14%), diarrhoea, reduction of blood sugar levels, and loss of appetite. Less common side effects included pancreatitis, renal impairment, and suicidal tendencies. In rodent studies, liraglutide has been associated with benign and malignant thyroid C-cell tumors. Liraglutide is not recommended in patients with a personal or family history of medullary thyroid cancer or multiple endocrine neoplasia 2A or 2B. Because of limited experience in patients with hepatic impairment, it must be used cautiously in patients with liver impairment. If data regarding safety becomes available it would be an interesting drug given its potential to improve various cardio metabolic factors<sup>[58]</sup>.

### **Antiobesity drugs: In the pipeline**

Apart from the current approved drugs for treatment of obesity many others are in various stages of development. Endogenous cannabinoids which are ubiquitous lipid signalling molecules having both central and peripheral effects mediated by the specific receptors CB1 and CB2<sup>[59]</sup>. Compounds targeting the peripheral CB1 receptors selectively are under evaluation<sup>[60,61]</sup>. Various drugs which may hold promise in near future are listed in the table. Other drugs in various stages of development are summarized in the Table 1<sup>[62-67]</sup>.

Overall there is hope of some drugs being available in near future. The role of combination polytherapy needs further evaluation due to paucity of efficacy and safety data in cirrhosis patients currently.

## **SURGERY**

The number of obese patients awaiting organ transplantation is increasing in parallel with the increasing prevalence of obesity. For patients with advanced fibrosis and cirrhosis, historically, bariatric surgery was not advised or offered. Complications of bariatric surgery, including bleeding, gastrointestinal symptoms, nutritional or electrolyte abnormalities, and stomal stenosis, can be seen in 10% to 17% of patients without cirrhosis<sup>[68]</sup>. Bariatric surgery may be useful in cirrhosis patients needing LT who were denied evaluation primarily because of weight. Furthermore reduction in weight can lead to improvement in liver parameters reducing chances of decompensation over time.

Patients with cirrhosis undergoing surgery of any kind are placed at an increased risk of mortality from liver failure, renal failure, or even postoperative bleeding due to impaired coagulation. This risk depends on the

degree of liver dysfunction or model for end-stage liver disease scores<sup>[69]</sup>. The mortality rates from bariatric operations have been reported to be in the range of 0.28%-0.35%<sup>[14,70]</sup>. A large population-based study ( $n = 674900$ ) has reported that patients with cirrhosis had higher in-hospital mortality rates than those without cirrhosis after bariatric surgery (1.2% vs 0.3%)<sup>[14]</sup>.

Takata *et al*<sup>[71]</sup> reviewed 15 patients of end-stage organ failure, of which 6 with cirrhosis underwent laparoscopic sleeve gastrectomy (LSG). Complications were noted in 2 patients with cirrhosis but there was no mortality. The mean follow-up was 12.4 mo, and the mean excess weight loss noted was 33% for cirrhosis patients at 9 mo<sup>[71]</sup>. The LSG was selected instead of Roux-en-Y gastric bypass (LRYGB) in this study for the following reasons: (1) some evidence has shown previously that the operative time and overall morbidity are reduced compared with those with LRYGB<sup>[13]</sup>; (2) the remaining gastric tube remains endoscopically accessible in the case of variceal bleeding; (3) endoscopic access to the biliary system after liver transplantation is preserved; and (4) it is expected that intake and absorption of critical medications will not be significantly altered.

Lin *et al*<sup>[72]</sup> studied 26 pretransplant patients who underwent LSG. The mean age of patients was 57 years with 17 (65%) of patients being female. Six patients had end-stage renal disease, and 20 patients had end-stage liver disease. There were 6 postoperative complications but no death, the complications being two superficial wound infections, one staple line leak, one postoperative bleed, one transient encephalopathy, and one renal insufficiency that resolved. The mean excess weight loss at 1, 3 and 12 mo was 17%, 26% and 50% respectively<sup>[72]</sup>. This lead to liver transplantation in seven patients showing LSG is well tolerated, is technically feasible, and improves candidacy for transplantation.

Shimizu *et al*<sup>[73]</sup> prospectively reviewed 23 patients (12 with known cirrhosis and 11 with unknown cirrhosis). There were 14 females and 9 males with a mean age of  $51.5 \pm 8.3$  and a mean body mass index of  $48.2 \pm 8.6$  kg/m<sup>2</sup>. Child-Pugh classes were A ( $n = 22$ ) and B ( $n = 1$ ). Procedures performed were LRYGB ( $n = 14$ ), LSG ( $n = 8$ ), and laparoscopic adjustable gastric banding (LAGB) ( $n = 1$ ). No patients had liver decompensation after surgery. The patients lost  $67.4\% \pm 30.9\%$  of their excess weight at 12 mo follow-up and  $67.7\% \pm 24.8\%$  at 37 mo follow-up<sup>[73]</sup>.

Most recently Pestana *et al*<sup>[74]</sup> reviewed 14 patients [11 patients underwent sleeve gastrectomy (78.6%) and 3 gastric bypass (21.4%)] with Child's A cirrhosis with or without portal hypertension. The mean patient age was 55.5 years, and 10 of 14 patients were women. At 1-year post surgery, only 1 of 8 patients who underwent follow-up ultrasound imaging showed steatosis. The bilirubin level above 2 mg/dL was seen in a patient one year post surgery. One patient developed encephalopathy at 2-year post-surgery. Bariatric surgery in patients with compensated cirrhosis even with mild portal hypertension seems well tolerated<sup>[74]</sup>.

Woodford *et al*<sup>[75]</sup> studied 14 patients intraoperatively detected with cirrhosis undergoing LAGB. No patients had preoperative clinical evidence of decompensated liver disease. There was no operative mortality.

Table 2 reviews the various studies done on bariatric surgery in cirrhosis patients<sup>[71,73-78]</sup>. Overall the literature suggests that bariatric surgery is tolerated in compensated cirrhosis although with slightly higher but acceptable complication rate and should be offered to obese cirrhosis patients. This will delay progression of liver disease to decompensation and also increase the candidacy for transplantation in both living donor liver transplantation and dead donor liver transplantation setting.

## ENDOSCOPIC INTERVENTIONS FOR MORBID OBESITY IN CIRRHOSIS

Endoluminal interventions performed through the gastrointestinal (GI) tract using endoscope offers potential for a weight loss procedure which is safer and more cost-effective than the current laparoscopic approaches<sup>[79]</sup>. Endoscopic techniques try to mimic the anatomical features produced by bariatric surgery. There are mainly two types of endoscopic weight loss modalities - restrictive and malabsorptive. Restrictive procedures causes reduction of gastric volume through use of space-occupying prosthesis or through suturing/stapling devices, while malabsorptive procedures causes reduced absorption by preventing contact of food with the duodenum and proximal jejunum. Restrictive procedures include intragastric balloon insertion, endoluminal vertical gastropasty, transoral gastropasty and transoral endoscopic restrictive implant system, while malabsorptive procedure include duodenojejunal bypass sleeve. Gastroduodenojejunal bypass sleeve is combines both restrictive and malabsorptive features. Except for intragastric balloon, all the mentioned procedures are comparatively new, with no data on cirrhosis patients.

Intra-gastric balloon placement is minimally invasive modality for weight loss. While this procedure has a well-established role in patients without liver disease, data on cirrhosis is not there. A meta-analysis of intra-gastric balloon placement in general patients including 15 articles (3608 patients) showed weight loss of 14.7 kg, 12.2% of initial weight, 5.7 kg/m<sup>2</sup>, and 32.1% of excess weight at 6 mo. Complications of intra-gastric balloon placement are uncommon and most common side effect is nausea and vomiting (8.6%)<sup>[80]</sup>. Other side effects included intolerance to the balloon which resulted in early removal, gastric ulcers and erosions, esophagitis, spontaneous deflation, persistent vomiting, gastroesophageal reflux and abdominal pain. However, severe complications are rare with a large Italian series of 2525 cases showing the following complications; 0.08% acute gastric dilatation, gastric perforation in 5 (0.19%, 4 of these had gastric surgery earlier), gastric obstruction in 0.76%, balloon rupture in 0.36%, esopha-

**Table 2 Studies of bariatric surgery in cirrhotic patients**

Ref.	Study characteristics	Cirrhosis diagnosis	Child pugh	Procedures	Complications	Liver decompensation	Mortality
Pestana <i>et al</i> <sup>[74]</sup> n = 14 F:M = 10:4	Mean age = 55.5 yr	Known cirrhosis	A = 14	SG = 11 RYGB = 3	0	1 (late HE)	0
Shimizu <i>et al</i> <sup>[73]</sup> n = 23 F:M = 14:9	Mean age = 51.5 yr Mean BMI = 48.2 kg/m <sup>2</sup> Mean stay = 4.3 d	12 preoperatively 11 intraoperatively	A = 22 B = 1	RYGB = 14 SG = 8 AGB = 1	8	0	0
Rebibo <i>et al</i> <sup>[78]</sup> n = 13 F:M = 7:6	Median age = 52 yr Median BMI = 46.3 kg/m <sup>2</sup>	All intraoperatively	A = 13	SG = 13	2	1 (ascites)	0
Takata <i>et al</i> <sup>[71]</sup> n = 6 F:M = 4:2	Mean age = 52 yr Mean BMI = 49 kg/m <sup>2</sup>	All preoperatively	A = 4 B = 2	SG = 6	2	1 (ascites) 1 (HE)	0
Dallal <i>et al</i> <sup>[76]</sup> n = 30 F:M = 20:20	Mean age = 50 yr Mean BMI = 52.6 kg/m <sup>2</sup> Mean hospital stay = 4 d	Diagnosed intraoperatively in 27 (90%)	A = 30	RYGB = 27 SG = 3	9	0	0
Kral <i>et al</i> <sup>[77]</sup> n = 14 F:M = 10:4	Mean age = 40 yr Mean BMI = 54 kg/m <sup>2</sup>	All intraoperatively	NA	BPD = 14	2	2	2 (one late hepatic failure)
Woodford <i>et al</i> <sup>[75]</sup> n = 14 F:M = 10:4	Mean age = 52.5 yr Mean BMI = 38.9 kg/m <sup>2</sup>	All intraoperatively	A or B	AGB = 14	2	0	0

RYGB: Roux-en-Y gastric bypass; SG: Sleeve gastrectomy; BPD: Bilio-pancreatic diversion; AGB: Adjustable gastric banding; HE: Hepatic encephalopathy; M: Male; F: Female; BMI: Body mass index.

gitis in 1.27% and gastric ulcer in 0.2%. They noted significant improvement of co-morbidities<sup>[81]</sup>. It should be noted that above meta-analysis and Italian study used BioEnterics intra-gastric balloon, the newer Spatz balloon provides option of gradual increase (or decrease) in balloon volume, thus should be associated with less complications and it can be kept for 1 year as compared to 6 mo duration for earlier. If dietary and lifestyle measures continued after balloon removal, these patients sustain initial weight loss. A Brazilian multicenter study of 483 patients showed that significant number of patients maintained their weight loss after balloon removal with a multidisciplinary program which involved clinical, psychiatric, exercise, and dietary therapy<sup>[82]</sup>.

We published use of intragastric balloon in decompensated cirrhosis for the first time in 2012 as letter to editor. The 61-year-old patient had decompensated alcoholic liver disease (CTP score 9). His BMI decreased from 48.3 kg/m<sup>2</sup> to 39.2 kg/m<sup>2</sup> (resulting in a total of 24 kg weight loss) at 6 mo after intragastric balloon placement. His diabetic control also improved, HbA1c level decreasing from 9.2 to 5.4<sup>[83]</sup>.

We have placed a total of 8 intragastric balloons (7 had decompensated cirrhosis) and five of them had successful liver transplantation (3 deceased donor liver transplantation and 2 LDLT), this data is submitted for publication elsewhere. None of these patients had any severe complication other than vomiting in initial few days. One patient didn't lose weight out of these 8 patients and in one patient we had to decrease initial volume of Spatz balloon due to persistent vomiting at day 7. Although intra-gastric balloon appears to be a promising modality for weight loss in decompensated cirrhosis,

it cannot be placed in all patients. Contraindications of intra-gastric balloon include severe coagulopathy, upper gastro-intestinal tract conditions with potential bleeding risks (large or high risk esophageal varices, gastric varices, ulceration), presence of eating disorders, history of prior gastroesophageal surgery, presence of autoimmune connective tissue disorder affecting GI tract, significant hiatal hernia, esophageal stenosis, GI motility disorders, unwillingness for supervised diet and behaviour modification program and allergy to Silicon (product information). In conclusion, there is plenty of data about use of intra-gastric balloon for weight loss in morbidly obese patients and it has proven to be a safe modality. However, its use in morbidly obese patients with cirrhosis who are awaiting liver transplantation has not been studied.

The endoscopic administration of botulinum toxin type A in gastric wall is thought to aid in weight reduction by inhibiting antral motility and slowing gastric emptying by inhibiting acetylcholine release at the neuromuscular junction causing local paralysis of muscle. In published randomized placebo controlled trials no statistically significant weight loss has been shown<sup>[84,85]</sup>. When fundal injections were also applied, significantly greater short-term weight loss, reduction in BMI and prolongation of gastric emptying was achieved compared with controls<sup>[86]</sup>. A recent meta-analysis by Bang *et al*<sup>[87]</sup> analysed a total of 115 patients in 8 studies. Wide area injection including the fundus or body rather than the antrum only and multiple injections (> 10) were associated with weight loss. The safety and efficacy of this approach needs to be studied in cirrhosis.

There has been a lot of enthusiasm in gastric elec-

trical stimulation (GES) and devices innervating the stomach for bariatric applications. The exact mechanisms of GES are largely unknown, but causes delayed gastric emptying and increased satiety<sup>[88]</sup>. Recently in January 2015 the Maestro Rechargeable System, was approved<sup>[89]</sup>. These devices are generally implanted through open or laparoscopic means, but electrical stimulation systems deployed endoluminally has shown to be feasible and safe<sup>[90,91]</sup>.

## CONCLUSION

There has been a worldwide rise in patients having obesity associated with cirrhosis. Obesity with cirrhosis is a double trouble leading to early decompensation and also making liver transplantation difficult. Weight reduction is generally more difficult in this group of patients. Lifestyle changes should include a diet of around 30 kcal/kg and walk of greater than 5000 steps/d but optimal safe exercise regimen is unknown.

Among the current FDA approved anti-obesity drugs (orlistat, phenteramine/topiramate-ER, lorcaserin, naltrexone-bupropion ER and liraglutide) none are well studied in patients with cirrhosis but lorcaserin and liraglutide have similar pharmacokinetics in patients with mild hepatic impairment and are not contraindicated.

Bariatric surgery can be relatively safely performed in compensated cirrhosis patients with a slightly higher but acceptable complication rate.

Role of endoscopic intervention for management of obesity in cirrhosis especially intragastric balloon placement is evolving but promising and seems feasible even in those with decompensated cirrhosis.

## REFERENCES

- Hainer V, Aldhoon-Hainerová I. Tolerability and safety of the new anti-obesity medications. *Drug Saf* 2014; **37**: 693-702 [PMID: 25096956 DOI: 10.1007/s40264-014-0206-3]
- Middleton KM, Patidar SM, Perri MG. The impact of extended care on the long-term maintenance of weight loss: a systematic review and meta-analysis. *Obes Rev* 2012; **13**: 509-517 [PMID: 22212682 DOI: 10.1111/j.1467-789X.2011.00972.x]
- World Health Organization. Obesity and overweight. [Accessed 2015 Jun 6]. Available from: URL: <http://www.who.int/mediacentre/factsheets/fs311/en/>
- Casazza K, Fontaine KR, Astrup A, Birch LL, Brown AW, Bohan Brown MM, Durant N, Dutton G, Foster EM, Heymsfield SB, McIver K, Mehta T, Menachemi N, Newby PK, Pate R, Rolls BJ, Sen B, Smith DL, Thomas DM, Allison DB. Myths, presumptions, and facts about obesity. *N Engl J Med* 2013; **368**: 446-454 [PMID: 23363498 DOI: 10.1056/NEJMs1208051]
- Berzigotti A, Garcia-Tsao G, Bosch J, Grace ND, Burroughs AK, Morillas R, Escorsell A, Garcia-Pagan JC, Patch D, Matloff DS, Groszmann RJ. Obesity is an independent risk factor for clinical decompensation in patients with cirrhosis. *Hepatology* 2011; **54**: 555-561 [PMID: 21567436 DOI: 10.1002/hep.24418]
- Gentile CL, Pagliassotti MJ. The role of fatty acids in the development and progression of nonalcoholic fatty liver disease. *J Nutr Biochem* 2008; **19**: 567-576 [PMID: 18430557 DOI: 10.1016/j.jnutbio.2007.10.001]
- Day CP. From fat to inflammation. *Gastroenterology* 2006; **130**: 207-210 [PMID: 16401483 DOI: 10.1053/j.gastro.2005.11.017]
- Jou J, Choi SS, Diehl AM. Mechanisms of disease progression in nonalcoholic fatty liver disease. *Semin Liver Dis* 2008; **28**: 370-379 [PMID: 18956293 DOI: 10.1055/s-0028-1091981]
- Dowman JK, Tomlinson JW, Newsome PN. Pathogenesis of non-alcoholic fatty liver disease. *QJM* 2010; **103**: 71-83 [PMID: 19914930 DOI: 10.1093/qjmed/hcp158]
- Svegliati-Baroni G, De Minicis S, Marziani M. Hepatic fibrogenesis in response to chronic liver injury: novel insights on the role of cell-to-cell interaction and transition. *Liver Int* 2008; **28**: 1052-1064 [PMID: 18783548 DOI: 10.1111/j.1478-3231.2008.01825.x]
- Xia JL, Dai C, Michalopoulos GK, Liu Y. Hepatocyte growth factor attenuates liver fibrosis induced by bile duct ligation. *Am J Pathol* 2006; **168**: 1500-1512 [PMID: 16651617 DOI: 10.2353/ajpath.2006.050747]
- Charlton MR, Pockros PJ, Harrison SA. Impact of obesity on treatment of chronic hepatitis C. *Hepatology* 2006; **43**: 1177-1186 [PMID: 16729327 DOI: 10.1002/hep.21239]
- Lee CM, Cirangle PT, Jossart GH. Vertical gastrectomy for morbid obesity in 216 patients: report of two-year results. *Surg Endosc* 2007; **21**: 1810-1816 [PMID: 17356932 DOI: 10.1007/s00464-007-9276-y]
- Mosko JD, Nguyen GC. Increased perioperative mortality following bariatric surgery among patients with cirrhosis. *Clin Gastroenterol Hepatol* 2011; **9**: 897-901 [PMID: 21782772 DOI: 10.1016/j.cgh.2011.07.007]
- Angulo P. Nonalcoholic fatty liver disease and liver transplantation. *Liver Transpl* 2006; **12**: 523-534 [PMID: 16555318 DOI: 10.1002/lt.20738]
- Afzali A, Berry K, Ioannou GN. Excellent posttransplant survival for patients with nonalcoholic steatohepatitis in the United States. *Liver Transpl* 2012; **18**: 29-37 [PMID: 21932374 DOI: 10.1002/lt.22435]
- Armstrong MJ, Adams LA, Canbay A, Syn WK. Extrahepatic complications of nonalcoholic fatty liver disease. *Hepatology* 2014; **59**: 1174-1197 [PMID: 24002776 DOI: 10.1002/hep.26717]
- Gunay Y, Guler N, Dayangac M, Taskesen F, Yaprak O, Emek E, Akyildiz M, Altaca G, Yuzer Y, Tokat Y. Living donor liver transplantation for obese patients: challenges and outcomes. *Liver Transpl* 2014; **20**: 311-322 [PMID: 24243642 DOI: 10.1002/lt.23794]
- Leonard J, Heimbach JK, Malinchoc M, Watt K, Charlton M. The impact of obesity on long-term outcomes in liver transplant recipients-results of the NIDDK liver transplant database. *Am J Transplant* 2008; **8**: 667-672 [PMID: 18294163 DOI: 10.1111/j.1600-6143.2007.02100.x]
- Agopian VG, Kaldas FM, Hong JC, Whittaker M, Holt C, Rana A, Zarrinpar A, Petrowsky H, Farmer D, Yersiz H, Xia V, Hiatt JR, Busuttill RW. Liver transplantation for nonalcoholic steatohepatitis: the new epidemic. *Ann Surg* 2012; **256**: 624-633 [PMID: 22964732 DOI: 10.1097/SLA.0b013e31826b4b7e]
- Jan A, Narwaria M, Mahawar KK. A Systematic Review of Bariatric Surgery in Patients with Liver Cirrhosis. *Obes Surg* 2015; **25**: 1518-1526 [PMID: 25982807 DOI: 10.1007/s11695-015-1727-2]
- Chen Y, Wang X, Wang J, Yan Z, Luo J. Excess body weight and the risk of primary liver cancer: an updated meta-analysis of prospective studies. *Eur J Cancer* 2012; **48**: 2137-2145 [PMID: 22446023 DOI: 10.1016/j.ejca.2012.02.063]
- Muto Y, Sato S, Watanabe A, Moriwaki H, Suzuki K, Kato A, Kato M, Nakamura T, Higuchi K, Nishiguchi S, Kumada H, Ohashi Y. Overweight and obesity increase the risk for liver cancer in patients with liver cirrhosis and long-term oral supplementation with branched-chain amino acid granules inhibits liver carcinogenesis in heavier patients with liver cirrhosis. *Hepatol Res* 2006; **35**: 204-214 [PMID: 16737844 DOI: 10.1016/j.hepres.2006.04.007]
- Campillo B, Bories PN, Leluan M, Pornin B, Devanlay M, Fouet P. Short-term changes in energy metabolism after 1 month of a regular oral diet in severely malnourished cirrhotic patients. *Metabolism* 1995; **44**: 765-770 [PMID: 7783661 DOI: 10.1016/0026-0495(95)90190-6]
- Yasutake K, Bekki M, Ichinose M, Ikemoto M, Fujino T,

- Ryu T, Wada Y, Takami Y, Saitsu H, Kohjima M, Fukuizumi K, Nakashima M, Nakamuta M, Enjoji M. Assessing current nutritional status of patients with HCV-related liver cirrhosis in the compensated stage. *Asia Pac J Clin Nutr* 2012; **21**: 400-405 [PMID: 22705430]
- 26 **Yatsuhashi H**, Ohnishi Y, Nakayama S, Iwase H, Nakamura T, Imawari M. Anti-hypoalbuminemic effect of branched-chain amino acid granules in patients with liver cirrhosis is independent of dietary energy and protein intake. *Hepatol Res* 2011; **41**: 1027-1035 [PMID: 21951974 DOI: 10.1111/j.1872-034X.2011.00864.x]
- 27 **Ohno T**, Tanaka Y, Sugauchi F, Orito E, Hasegawa I, Nukaya H, Kato A, Matunaga S, Endo M, Tanaka Y, Sakakibara K, Mizokami M. Suppressive effect of oral administration of branched-chain amino acid granules on oxidative stress and inflammation in HCV-positive patients with liver cirrhosis. *Hepatol Res* 2008; **38**: 683-688 [PMID: 18328070 DOI: 10.1111/j.1872-034X.2008.00319.x]
- 28 **Zamboni M**, Mazzali G, Fantin F, Rossi A, Di Francesco V. Sarcopenic obesity: a new category of obesity in the elderly. *Nutr Metab Cardiovasc Dis* 2008; **18**: 388-395 [PMID: 18395429 DOI: 10.1016/j.numecd.2007.10.002]
- 29 **Lim S**, Kim JH, Yoon JW, Kang SM, Choi SH, Park YJ, Kim KW, Lim JY, Park KS, Jang HC. Sarcopenic obesity: prevalence and association with metabolic syndrome in the Korean Longitudinal Study on Health and Aging (KLoSHA). *Diabetes Care* 2010; **33**: 1652-1654 [PMID: 20460442 DOI: 10.2337/dc10-0107]
- 30 **Hayashi F**, Momoki C, Yuikawa M, Simotani Y, Kawamura E, Hagihara A, Fujii H, Kobayashi S, Iwai S, Morikawa H, Enomoto M, Tamori A, Kawada N, Ohfuji S, Fukusima W, Habu D. Nutritional status in relation to lifestyle in patients with compensated viral cirrhosis. *World J Gastroenterol* 2012; **18**: 5759-5770 [PMID: 23155318 DOI: 10.3748/wjg.v18.i40.5759]
- 31 **Hayashi F**, Matsumoto Y, Momoki C, Yuikawa M, Okada G, Hamakawa E, Kawamura E, Hagihara A, Toyama M, Fujii H, Kobayashi S, Iwai S, Morikawa H, Enomoto M, Tamori A, Kawada N, Habu D. Physical inactivity and insufficient dietary intake are associated with the frequency of sarcopenia in patients with compensated viral liver cirrhosis. *Hepatol Res* 2013; **43**: 1264-1275 [PMID: 23489325 DOI: 10.1111/hepr.12085]
- 32 **García-Pagán JC**, Santos C, Barberá JA, Luca A, Roca J, Rodríguez-Roisin R, Bosch J, Rodés J. Physical exercise increases portal pressure in patients with cirrhosis and portal hypertension. *Gastroenterology* 1996; **111**: 1300-1306 [PMID: 8898644 DOI: 10.1053/gast.1996.v111.pm8898644]
- 33 **Román E**, Torrades MT, Nadal MJ, Cárdenas G, Nieto JC, Vidal S, Bascuñana H, Juárez C, Guarner C, Córdoba J, Soriano G. Randomized pilot study: effects of an exercise programme and leucine supplementation in patients with cirrhosis. *Dig Dis Sci* 2014; **59**: 1966-1975 [PMID: 24599772 DOI: 10.1007/s10620-014-3086-6]
- 34 **Hickman IJ**, Jonsson JR, Prins JB, Ash S, Purdie DM, Clouston AD, Powell EE. Modest weight loss and physical activity in overweight patients with chronic liver disease results in sustained improvements in alanine aminotransferase, fasting insulin, and quality of life. *Gut* 2004; **53**: 413-419 [PMID: 14960526 DOI: 10.1136/gut.2003.027581]
- 35 **Konishi I**, Hiasa Y, Tokumoto Y, Abe M, Furukawa S, Toshimitsu K, Matsuura B, Onji M. Aerobic exercise improves insulin resistance and decreases body fat and serum levels of leptin in patients with hepatitis C virus. *Hepatol Res* 2011; **41**: 928-935 [PMID: 21707884 DOI: 10.1111/j.1872-034X.2011.00833.x]
- 36 **Leung WY**, Thomas GN, Chan JC, Tomlinson B. Weight management and current options in pharmacotherapy: orlistat and sibutramine. *Clin Ther* 2003; **25**: 58-80 [PMID: 12637112 DOI: 10.1016/S0149-2918(03)90009-9]
- 37 **Torgerson JS**, Hauptman J, Boldrin MN, Sjörström L. XENical in the prevention of diabetes in obese subjects (XENDOS) study: a randomized study of orlistat as an adjunct to lifestyle changes for the prevention of type 2 diabetes in obese patients. *Diabetes Care* 2004; **27**: 155-161 [PMID: 14693982 DOI: 10.2337/diacare.27.1.155]
- 38 **Hampp C**, Kang EM, Borders-Hemphill V. Use of prescription antiobesity drugs in the United States. *Pharmacotherapy* 2013; **33**: 1299-1307 [PMID: 24019195 DOI: 10.1002/phar.1342]
- 39 **Padwal R**, Kezouh A, Levine M, Etmnan M. Long-term persistence with orlistat and sibutramine in a population-based cohort. *Int J Obes (Lond)* 2007; **31**: 1567-1570 [PMID: 17420781 DOI: 10.1038/sj.ijo.0803631]
- 40 **United States Food and Drug Administration**. FDA Drug Safety Communication: Completed safety review of Xenical/ Alli (orlistat) and severe liver injury. [Accessed 2011 Oct 20]. Available from: URL: <http://www.fda.gov/Drugs/DrugSafety/PostmarketDrugSafetyInformationforPatientsandProviders/ucm213038.htm>
- 41 **Smith SR**, Prosser WA, Donahue DJ, Morgan ME, Anderson CM, Shanahan WR. Lorcaserin (APD356), a selective 5-HT(2C) agonist, reduces body weight in obese men and women. *Obesity (Silver Spring)* 2009; **17**: 494-503 [PMID: 19057523 DOI: 10.1038/oby.2008.537]
- 42 **Smith SR**, Weissman NJ, Anderson CM, Sanchez M, Chuang E, Stubbe S, Bays H, Shanahan WR; Behavioral Modification and Lorcaserin for Overweight and Obesity Management (BLOOM) Study Group. Multicenter, placebo-controlled trial of lorcaserin for weight management. *N Engl J Med* 2010; **363**: 245-256 [PMID: 20647200 DOI: 10.1056/NEJMoa0909809]
- 43 **Fidler MC**, Sanchez M, Raether B, Weissman NJ, Smith SR, Shanahan WR, Anderson CM; BLOSSOM Clinical Trial Group. A one-year randomized trial of lorcaserin for weight loss in obese and overweight adults: the BLOSSOM trial. *J Clin Endocrinol Metab* 2011; **96**: 3067-3077 [PMID: 21795446 DOI: 10.1210/jc.2011-1256]
- 44 **O'Neil PM**, Smith SR, Weissman NJ, Fidler MC, Sanchez M, Zhang J, Raether B, Anderson CM, Shanahan WR. Randomized placebo-controlled clinical trial of lorcaserin for weight loss in type 2 diabetes mellitus: the BLOOM-DM study. *Obesity (Silver Spring)* 2012; **20**: 1426-1436 [PMID: 22421927 DOI: 10.1038/oby.2012.66]
- 45 **Eisai Inc**. BELVIQ (lorcaserin hydrochloride) tablets, for oral use. Patient package insert January 4, 2013. [Accessed 2013 Jun 27]. Available from: URL: [http://www.accessdata.fda.gov/drugsatfda\\_docs/label/2012/0225291bl.pdf](http://www.accessdata.fda.gov/drugsatfda_docs/label/2012/0225291bl.pdf)
- 46 **Allison DB**, Gadde KM, Garvey WT, Peterson CA, Schwiens ML, Najarian T, Tam PY, Troupin B, Day WW. Controlled-release phentermine/topiramate in severely obese adults: a randomized controlled trial (EQUIP). *Obesity (Silver Spring)* 2012; **20**: 330-342 [PMID: 22051941 DOI: 10.1038/oby.2011.330]
- 47 **Gadde KM**, Allison DB, Ryan DH, Peterson CA, Troupin B, Schwiens ML, Day WW. Effects of low-dose, controlled-release, phentermine plus topiramate combination on weight and associated comorbidities in overweight and obese adults (CONQUER): a randomized, placebo-controlled, phase 3 trial. *Lancet* 2011; **377**: 1341-1352 [PMID: 21481449 DOI: 10.1016/S0140-6736(11)60205-5]
- 48 **Highlights of prescribing information**. QSYMIA (phentermine and topiramate extended-release). [Accessed 2015 Jun 6]. Available from: URL: [http://www.accessdata.fda.gov/drugsatfda\\_docs/label/2012/022580s0001bl.pdf](http://www.accessdata.fda.gov/drugsatfda_docs/label/2012/022580s0001bl.pdf)
- 49 **United States Food and Drug Administration**. FDA approves weight-management drug Contrave. Available from: URL: <http://www.fda.gov/newsevents/newsroom/pressannouncements/ucm413896.htm>
- 50 **Greenway FL**, Fujioka K, Plodkowski RA, Mudaliar S, Guttadauria M, Erickson J, Kim DD, Dunayevich E; COR-I Study Group. Effect of naltrexone plus bupropion on weight loss in overweight and obese adults (COR-I): a multicentre, randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet* 2010; **376**: 595-605 [PMID: 20673995 DOI: 10.1016/S0140-6736(10)60888-4]
- 51 **Apovian CM**, Aronne L, Rubino D, Still C, Wyatt H, Burns C, Kim D, Dunayevich E; COR-II Study Group. A randomized, phase 3 trial of naltrexone SR/bupropion SR on weight and obesity-related risk factors (COR-II). *Obesity (Silver Spring)* 2013; **21**: 935-943 [PMID: 23408728 DOI: 10.1002/oby.20309]

- 52 **Caixàs A**, Albert L, Capel I, Rigla M. Naltrexone sustained-release/bupropion sustained-release for the management of obesity: review of the data to date. *Drug Des Devel Ther* 2014; **8**: 1419-1427 [PMID: 25258511 DOI: 10.2147/DDDT.S55587]
- 53 **Greenway FL**, Dunayevich E, Tollefson G, Erickson J, Guttadauria M, Fujioka K, Cowley MA; NB-201 Study Group. Comparison of combined bupropion and naltrexone therapy for obesity with monotherapy and placebo. *J Clin Endocrinol Metab* 2009; **94**: 4898-4906 [PMID: 19846734 DOI: 10.1210/jc.2009-1350]
- 54 **Highlights of prescribing information.** CONTRAVE (naltrexone HCl and bupropion HCl) Extended-Release Tablets. [Accessed 2015 Jun 6]. Available from: URL: <http://general.takedapharm.com/content/file.aspx?filetypecode=CONTRAWEPI&Country-Code=US&LanguageCode=EN&cacheRandomizer=30e83f2b-9c2f-4df1-8fb8-7d2bf6cf7de1>
- 55 **Scott LJ.** Liraglutide: a review of its use in the management of obesity. *Drugs* 2015; **75**: 899-910 [PMID: 25985864 DOI: 10.1007/s40265-015-0408-8]
- 56 **Astrup A**, Rössner S, Van Gaal L, Rissanen A, Niskanen L, Al Hakim M, Madsen J, Rasmussen MF, Lean ME; NN8022-1807 Study Group. Effects of liraglutide in the treatment of obesity: a randomised, double-blind, placebo-controlled study. *Lancet* 2009; **374**: 1606-1616 [PMID: 19853906 DOI: 10.1016/S0140-6736(09)61375-1]
- 57 **Wadden TA**, Hollander P, Klein S, Niswender K, Woo V, Hale PM, Aronne L; NN8022-1923 Investigators. Weight maintenance and additional weight loss with liraglutide after low-calorie-diet-induced weight loss: the SCALE Maintenance randomized study. *Int J Obes (Lond)* 2013; **37**: 1443-1451 [PMID: 23812094 DOI: 10.1038/ijo.2013.120]
- 58 **United States Food and Drug Administration.** FDA approves weight-management drug Saxenda. [Accessed 2015 Jun 6]. Available from: URL: <http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm427913.htm>
- 59 **Mackie K.** Cannabinoid receptors: where they are and what they do. *J Neuroendocrinol* 2008; **20** Suppl 1: 10-14 [PMID: 18426493 DOI: 10.1111/j.1365-2826.2008.01671.x]
- 60 **Tam J**, Cinar R, Liu J, Godlewski G, Wesley D, Jourdan T, Szanda G, Mukhopadhyay B, Chedester L, Liow JS, Innis RB, Cheng K, Rice KC, Deschamps JR, Chorvat RJ, McElroy JF, Kunos G. Peripheral cannabinoid-1 receptor inverse agonism reduces obesity by reversing leptin resistance. *Cell Metab* 2012; **16**: 167-179 [PMID: 22841573 DOI: 10.1016/j.cmet.2012.07.002]
- 61 **Tam J**, Vemuri VK, Liu J, Bátkai S, Mukhopadhyay B, Godlewski G, Osei-Hyiaman D, Ohnuma S, Ambudkar SV, Pickel J, Makriyannis A, Kunos G. Peripheral CB1 cannabinoid receptor blockade improves cardiometabolic risk in mouse models of obesity. *J Clin Invest* 2010; **120**: 2953-2966 [PMID: 20664173 DOI: 10.1172/JCI42551]
- 62 **Gras J.** Cetilistat for the treatment of obesity. *Drugs Today (Barc)* 2013; **49**: 755-759 [PMID: 24524093 DOI: 10.1358/dot.2013.49.1.2.2099318]
- 63 **Moon HS**, Matarese G, Brennan AM, Chamberland JP, Liu X, Fiorenza CG, Mylvaganam GH, Abanni L, Carbone F, Williams CJ, De Paoli AM, Schneider BE, Mantzoros CS. Efficacy of metreleptin in obese patients with type 2 diabetes: cellular and molecular pathways underlying leptin tolerance. *Diabetes* 2011; **60**: 1647-1656 [PMID: 21617185 DOI: 10.2337/db10-1791]
- 64 **Nielsen AL**, Larsen TM, Madsbad S, Breum L, Jensen TJ, Kroustrup JP, Astrup A. [The effect of tesofensin on body weight and body composition in obese subjects--secondary publication]. *Ugeskr Laeger* 2009; **171**: 2974-2977 [PMID: 19824222]
- 65 **Powell AG**, Apovian CM, Aronne LJ. New drug targets for the treatment of obesity. *Clin Pharmacol Ther* 2011; **90**: 40-51 [PMID: 21654742 DOI: 10.1038/clpt.2011.82]
- 66 **George M**, Rajaram M, Shanmugam E. New and emerging drug molecules against obesity. *J Cardiovasc Pharmacol Ther* 2014; **19**: 65-76 [PMID: 24064009 DOI: 10.1177/1074248413501017]
- 67 **Joharapurkar AA**, Dhanesha NA, Jain MR. Inhibition of the methionine aminopeptidase 2 enzyme for the treatment of obesity. *Diabetes Metab Syndr Obes* 2014; **7**: 73-84 [PMID: 24611021 DOI: 10.2147/DMSO.S56924]
- 68 **Chang SH**, Stoll CR, Song J, Varela JE, Eagon CJ, Colditz GA. The effectiveness and risks of bariatric surgery: an updated systematic review and meta-analysis, 2003-2012. *JAMA Surg* 2014; **149**: 275-287 [PMID: 24352617 DOI: 10.1001/jamasurg.2013.3654]
- 69 **Teh SH**, Nagorney DM, Stevens SR, Offord KP, Therneau TM, Plevak DJ, Talwalkar JA, Kim WR, Kamath PS. Risk factors for mortality after surgery in patients with cirrhosis. *Gastroenterology* 2007; **132**: 1261-1269 [PMID: 17408652 DOI: 10.1053/j.gastro.2007.01.040]
- 70 **Buchwald H**, Estok R, Fahrenbach K, Banel D, Sledge I. Trends in mortality in bariatric surgery: a systematic review and meta-analysis. *Surgery* 2007; **142**: 621-632; discussion 632-635 [PMID: 17950357 DOI: 10.1016/j.surg.2007.07.018]
- 71 **Takata MC**, Campos GM, Ciofica R, Rabl C, Rogers SJ, Cello JP, Ascher NL, Posselt AM. Laparoscopic bariatric surgery improves candidacy in morbidly obese patients awaiting transplantation. *Surg Obes Relat Dis* 2008; **4**: 159-164; discussion 164-165 [PMID: 18294923 DOI: 10.1016/j.soard.2007.12.009]
- 72 **Lin MY**, Tavakol MM, Sarin A, Amirikiai SM, Rogers SJ, Carter JT, Posselt AM. Laparoscopic sleeve gastrectomy is safe and efficacious for pretransplant candidates. *Surg Obes Relat Dis* 2013; **9**: 653-658 [PMID: 23701857 DOI: 10.1016/j.soard.2013.02.013]
- 73 **Shimizu H**, Phuong V, Maia M, Kroh M, Chand B, Schauer PR, Brethauer SA. Bariatric surgery in patients with liver cirrhosis. *Surg Obes Relat Dis* 2013; **9**: 1-6 [PMID: 23201210 DOI: 10.1016/j.soard.2012.07.021]
- 74 **Pestana L**, Swain J, Dierkhising R, Kendrick ML, Kamath PS, Watt KD. Bariatric surgery in patients with cirrhosis with and without portal hypertension: a single-center experience. *Mayo Clin Proc* 2015; **90**: 209-215 [PMID: 25659239 DOI: 10.1016/j.mayocp.2014.11.012]
- 75 **Woodford RM**, Burton PR, O'Brien PE, Laurie C, Brown WA. Laparoscopic Adjustable Gastric Banding In Patients with Unexpected Cirrhosis: Safety and Outcomes. *Obes Surg* 2015; **25**: 1858-1862 [PMID: 25708241 DOI: 10.1007/s11695-015-1623-9]
- 76 **Dallal RM**, Mattar SG, Lord JL, Watson AR, Cottam DR, Eid GM, Hamad G, Rabinovitz M, Schauer PR. Results of laparoscopic gastric bypass in patients with cirrhosis. *Obes Surg* 2004; **14**: 47-53 [PMID: 14980033 DOI: 10.1381/096089204772787284]
- 77 **Kral JG**, Thung SN, Biron S, Hould FS, Lebel S, Marceau S, Simard S, Marceau P. Effects of surgical treatment of the metabolic syndrome on liver fibrosis and cirrhosis. *Surgery* 2004; **135**: 48-58 [PMID: 14694300 DOI: 10.1016/j.surg.2003.10.003]
- 78 **Rebibo L**, Gerin O, Verhaeghe P, Dhahri A, Cosse C, Regimbeau JM. Laparoscopic sleeve gastrectomy in patients with NASH-related cirrhosis: a case-matched study. *Surg Obes Relat Dis* 2014; **10**: 405-410; quiz 565 [PMID: 24355322 DOI: 10.1016/j.soard.2013.09.015]
- 79 **Coté GA**, Edmundowicz SA. Emerging technology: endoluminal treatment of obesity. *Gastrointest Endosc* 2009; **70**: 991-999 [PMID: 19879407 DOI: 10.1016/j.gie.2009.09.016]
- 80 **Imaz I**, Martínez-Cervell C, García-Alvarez EE, Sendra-Gutiérrez JM, González-Enríquez J. Safety and effectiveness of the intra-gastric balloon for obesity. A meta-analysis. *Obes Surg* 2008; **18**: 841-846 [PMID: 18459025 DOI: 10.1007/s11695-007-9331-8]
- 81 **Genco A**, Bruni T, Doldi SB, Forestieri P, Marino M, Busetto L, Giardiello C, Angrisani L, Pecchioli L, Stornelli P, Puglisi F, Alkilani M, Nigri A, Di Lorenzo N, Furbetta F, Cascardo A, Cipriano M, Lorenzo M, Basso N. BioEnterics Intra-gastric Balloon: The Italian Experience with 2,515 Patients. *Obes Surg* 2005; **15**: 1161-1164 [PMID: 16197790 DOI: 10.1381/0960892055002202]
- 82 **Sallet JA**, Marchesini JB, Paiva DS, Komoto K, Pizani CE, Ribeiro ML, Miguel P, Ferraz AM, Sallet PC. Brazilian multicenter study of the intra-gastric balloon. *Obes Surg* 2004; **14**: 991-998 [PMID: 15329191 DOI: 10.1381/0960892041719671]
- 83 **Choudhary NS**, Saigal S, Saraf N, Puri R, Soim A. Innovative approach using an intra-gastric balloon for weight loss in a morbidly

- obese patient undergoing liver transplantation. *Liver Transpl* 2013; **19**: 235 [PMID: 23161847 DOI: 10.1002/lt.23567]
- 84 **Gui D**, Mingrone G, Valenza V, Spada PL, Mutignani M, Runfola M, Scarfone A, Di Mugno M, Panunzi S. Effect of botulinum toxin antral injection on gastric emptying and weight reduction in obese patients: a pilot study. *Aliment Pharmacol Ther* 2006; **23**: 675-680 [PMID: 16480407 DOI: 10.1111/j.1365-2036.2006.02773.x]
- 85 **Mittermair R**, Keller C, Geibel J. Intra-gastric injection of botulinum toxin A for the treatment of obesity. *Obes Surg* 2007; **17**: 732-736 [PMID: 17879570]
- 86 **Foschi D**, Corsi F, Lazzaroni M, Sangaletti O, Riva P, La Tartara G, Bevilacqua M, Osio M, Alciati A, Bianchi Porro G, Trabucchi E. Treatment of morbid obesity by intraparietogastric administration of botulinum toxin: a randomized, double-blind, controlled study. *Int J Obes (Lond)* 2007; **31**: 707-712 [PMID: 17006442 DOI: 10.1038/sj.ijo.0803451]
- 87 **Bang CS**, Baik GH, Shin IS, Kim JB, Suk KT, Yoon JH, Kim YS, Kim DJ. Effect of intra-gastric injection of botulinum toxin A for the treatment of obesity: a meta-analysis and meta-regression. *Gastrointest Endosc* 2015; **81**: 1141-1149.e1-7 [PMID: 25765772 DOI: 10.1016/j.gie.2014.12.025]
- 88 **Chen J**. Mechanisms of action of the implantable gastric stimulator for obesity. *Obes Surg* 2004; **14** Suppl 1: S28-S32 [PMID: 15479587 DOI: 10.1381/0960892041978962]
- 89 **United States Food and Drug Administration**. FDA approves first-of-kind device to treat obesity. Available from: URL: <http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm430223.htm>
- 90 **Sallam HS**, Chen JD, Pasricha PJ. Feasibility of gastric electrical stimulation by percutaneous endoscopic transgastric electrodes. *Gastrointest Endosc* 2008; **68**: 754-759 [PMID: 18718585 DOI: 10.1016/j.gie.2008.04.060]
- 91 **Xu X**, Pasricha PJ, Chen JD. Feasibility of gastric electrical stimulation by use of endoscopically placed electrodes. *Gastrointest Endosc* 2007; **66**: 981-986 [PMID: 17963885 DOI: 10.1016/j.gie.2007.05.020]

**P- Reviewer:** Naqvi IH, Su ZJ **S- Editor:** Gong XM  
**L- Editor:** A **E- Editor:** Liu SQ



## Update on hepatitis C: Direct-acting antivirals

Leon L Seifert, Ryan B Perumpail, Aijaz Ahmed

Leon L Seifert, Department of Transplantation Medicine, University Hospital Münster, 48149 Münster, Germany

Ryan B Perumpail, Aijaz Ahmed, Division of Gastroenterology and Hepatology, Stanford University School of Medicine, Stanford, CA 94305, United States

Author contributions: Seifert LL, Perumpail RB and Ahmed A designed research and analyzed data; Seifert LL performed research and wrote the paper.

Conflict-of-interest statement: We declare that we have no conflicts of interest.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Aijaz Ahmed, MD, Associate Professor of Medicine, Medical Director Liver Transplant Program, Division of Gastroenterology and Hepatology, Stanford University School of Medicine, 750 Welch Road, Suite 210, Stanford, CA 94305, United States. [aijazahmed@stanford.edu](mailto:aijazahmed@stanford.edu)  
 Telephone: +1-650-4986091  
 Fax: +1-650-4985692

Received: July 20, 2015  
 Peer-review started: July 24, 2015  
 First decision: October 22, 2015  
 Revised: October 24, 2015  
 Accepted: November 23, 2015  
 Article in press: November 25, 2015  
 Published online: December 8, 2015

### Abstract

Hepatitis C virus (HCV) was discovered 26 years ago. For decades, interferon-based therapy has been the mainstay of treatment for HCV. Recently, several direct-

acting antivirals (DAAs) have been approved for treatment of HCV-infected patients and to help combat the virus. These drugs have revolutionized the management of HCV as all-oral regimens with favorable side effect profiles and superior rates of sustained virological response. Emerging real-world data are demonstrating results comparable to registration trials for DAA agents. Suddenly, the potential for eradicating HCV is on the horizon.

**Key words:** Hepatitis C virus; Direct-acting antivirals; Sustained virologic response; Management; Treatment

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Recently, several direct-acting antivirals (DAAs) have been approved for treatment of hepatitis C virus (HCV)-infected patients and to help combat the virus. These drugs have revolutionized the management of HCV as all-oral regimens with favorable side effect profiles and superior rates of sustained virological response. Emerging real-world data are demonstrating results comparable to registration trials for DAA agents. Suddenly, the potential for eradicating HCV is on the horizon.

Seifert LL, Perumpail RB, Ahmed A. Update on hepatitis C: Direct-acting antivirals. *World J Hepatol* 2015; 7(28): 2829-2833 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i28/2829.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i28.2829>

### INTRODUCTION

Hepatitis C virus (HCV) was discovered 26 years ago in 1989, previously the HCV-related clinical entity was referred to as non-A, non-B hepatitis<sup>[1]</sup>. Currently, HCV has created a major health burden, with approximately 180 million infected people worldwide, representing about 2%-3% of the world's population<sup>[2]</sup>. This single-stranded, positive-sense 9.6 kb RNA-virus is globally

prevalent, showing geographic variation in its genotypic distribution and represents a major cause of end-stage liver disease<sup>[3,4]</sup>. About 4 out of 5 patients acutely infected with HCV develop a chronic hepatitis while only 20% of patients demonstrate spontaneous recovery with eradication of HCV<sup>[5]</sup>. Chronic hepatitis C (CHC) is a leading cause of cirrhosis and is complicated by development of hepatocellular carcinoma in 1%-4% of cirrhotic patients<sup>[6,7]</sup>.

Until recently, interferon (IFN)-based therapies represented the mainstay of treatment for HCV infection. Modifications of the treatment-regimens including pegylation of IFN and the addition of ribavirin (RBV) resulted in suboptimal improvement sustained virologic response (SVR) and an unfavourable adverse effects profile. Based on the HCV genotype (GT) and the treatment-experience, only 40% to 70% of patients achieved SVR, with poorer outcomes among people infected with the more prevalent GT1<sup>[8]</sup>. The approval of the first-generation direct acting antiviral (DAA) agents, telaprevir (TLV) and boceprevir (BCV), in 2011 provided improvement in SVR for the targeted HCV GT1<sup>[9]</sup>. Unfortunately, TLV and BCV therapy was complicated by cumbersome schema of drug intake and the broad range of adverse events.

With the release of sofosbuvir in 2013 and 2014 in most Western countries, a new era in the treatment of CHC began. An all-oral, IFN-free antiviral treatment for CHC with DAA agents became available for the first time. In addition to sofosbuvir, approvals of other second-generation DAA agents, which target different proteins of HCV have improved the efficacy of antiviral therapy with better tolerance. The superior SVR rates from several phase III trials have recently been confirmed by a number of real-life experience reports. We review various DAA-based antiviral regimens for HCV-infected patients.

## MOLECULAR STRUCTURE OF HCV - TARGET SITES FOR DAA AGENTS

HCV is a member of the Flaviviridae virus family<sup>[10-12]</sup>. Its RNA is single-stranded and positive-sensed with a size of approximately 9.6 kb. The precursor-polyprotein is post-translationally processed and modified by a cooperation of cellular and viral proteases<sup>[13,14]</sup>. Bench molecular biology research on HCV has led to a better understanding of its replication cycle and has been instrumental in the discovery and development of molecules blocking viral proteins, specifically the DAAs<sup>[15-17]</sup>. The HCV-genome encodes for 9 proteins - 2 are structural (E1 and E2) and 7 non-structural (p7, NS2, NS3, NS4A, NS4B, NS5A, NS5B)<sup>[10,11,13,14]</sup>. These proteins provide targets for the DAAs, being mostly essential in the virus cycle of replication<sup>[10-14]</sup>. NS5B is a polymerase and a prime target for antiviral agents<sup>[18,19]</sup>. Antiviral agents are classified as inhibitors of nucleoside-type and non-nucleoside type. The active site of NS5B is highly conserved compared to other parts of the HCV-genome<sup>[18,19]</sup>. Currently, sofosbuvir is the only clinically available NS5B-inhibitor

(nucleoside-type) with pan-genotypic antiviral activity and higher barrier to resistance compared to other DAAs<sup>[18,19]</sup>. It is a pro-drug and currently represents the backbone of most treatment-regimens<sup>[20,21]</sup>. Inhibition of the NS3/4A protease-complex is another potential target for DAAs. The first-generation DAAs, TLV and BCV were inhibitors of NS3/4A, and referred to as protease-inhibitors. Currently, two NS3/4A-inhibitors are approved in the United States and the European Union - paritaprevir, which is approved for the treatment of HCV GT1 in combination with ombitasvir and dasabuvir; and simeprevir, which is approved in combination with sofosbuvir for GT1 patients. DAAs targeting NS5A have also been approved<sup>[22-25]</sup>. Currently, three different NS5A-Inhibitors are approved in the United States and/or the European Union - daclatasvir, which is given in combination with sofosbuvir  $\pm$  RBV for the treatment of the GTs 1-4; ombitasvir (ABT-267), which is approved for the treatment of GT1 in combination with paritaprevir and dasabuvir; and ledipasvir, which is approved for GT1, 3 and 4 in combination with sofosbuvir  $\pm$  RBV.

## DAA AGENTS - REGIMEN BASED ON HCV GT

With the approval of sofosbuvir in December of 2013 in North America (United States and Canada) and in January 2014 in Europe, an all-oral antiviral treatment for CHC with DAAs was available for the first time. In 2014, several studies analyzing the efficacy and the impact of the DAA-based therapies have been published. The response rates have been reproduced in real-life experiences (TRIO and TARGET 2.0) as well<sup>[26-29]</sup>.

HCV GT1 is the most common GT with an overall prevalence of 46.2%. In particular, GT1 is more prevalent in the Western countries of North America and Western Europe (75.8% and 59.0% respectively). Accordingly, most studies have focused on the treatment of GT1. Patients with GT2 and GT3 are less prevalent worldwide (GT2 9.1% and GT3 30.1%) with a noticeable variation in distribution within Western countries - North America (GT2 12.0% and GT3 10.4%) and Western Europe (GT2 10.8% and GT3 24.8%). Patient with GT 4, 5 and 6 demonstrate the lowest prevalence (GT4 8.3%, GT5 0.8%, and GT6 5.4%) worldwide, with highest prevalence in low-income countries, and limited data on experience with second-generation DAA agents<sup>[4]</sup>.

In phase-3 SAPPPIRE- I clinical trial the combination of ritonavir-boosted ABT-450/r (protease inhibitor)-ombitasvir (NS5A inhibitor), and dasabuvir (non-nucleoside NS5B) with RBV were studied in treatment-naïve, non-cirrhotic HCV-infected non-cirrhotic patients with GT1. RBV was added according to body weight ( $\geq$  75 kg 1200 mg/d or  $<$  75 kg 1000 mg/d). Overall, 96.2% of patients achieved SVR (GT1b 98.0% and GT1a 95.3%). A higher stage of fibrosis and obesity were the negative predictive factors with SVR-12 rates still  $>$  90% and thus satisfactory<sup>[30]</sup>. Treatment-experienced patients were studied in the SAPPPIRE- II clinical trial.

**Table 1** Direct-acting antiviral-based regimens for treatment-naïve hepatitis C virus-infected patients

Genotype	Recommended regimens options
GT1a	SOF/LDV × 12 wk
	PrOD + RBV × 12 wk (no cirrhosis) - 24 wk (cirrhosis)
	SOF + SMV ± RBV × 12 wk (no cirrhosis) - 24 wk (cirrhosis without Q80K variant)
GT1b	SOF + DCV × 12 wk (no cirrhosis) - 24 wk (cirrhosis ± RBV)
	SOF/LDV × 12 wk
GT2	PrOD + RBV × 12 wk
	SOF + SMV ± RBV × 12 wk (no cirrhosis) - 24 wk (cirrhosis)
	SOF + DCV × 12 wk (no cirrhosis) - 24 wk (cirrhosis ± RBV)
GT3	SOF + RBV × 12 wk (no cirrhosis) - 16 wk (cirrhosis)
	SOF + DCV × 12 wk (RBV intolerant)
GT4	SOF + PegIFN + RBV × 12 wk (PegIFN eligible)
	SOF + DCV × 12 wk (no cirrhosis) - 24 wk (cirrhosis ± RBV)
GT5	SOF + RBV × 24 wk (PegIFN ineligible)
	SOF/LDV × 12 wk
	PrO + RBV × 12 wk
GT6	SOF + RBV × 24 wk
	SOF + PegIFN + RBV × 12 wk
GT7	SOF + SMV ± RBV × 12 wk
	SOF/LDV × 12 wk
GT8	SOF + PegIFN + RBV × 12 wk
	SOF/LDV × 12 wk
GT9	SOF + PegIFN + RBV × 12 wk
	SOF/LDV × 12 wk

GT: Genotype; SOF: Sofosbuvir; LDV: Ledipasvir; PrOD: Paritaprevir + ritonavir + ombitasvir + dasabuvir; RBV: Ribavirin; SMV: Simeprevir; DCV: Daclatasvir; PegIFN: Pegylated interferon.

Again, a high grade of fibrosis and obesity were negative predictive factors, with an overall SVR-12 of 96.3% (GT1a 96% and GT1b 96.7%)<sup>[31]</sup>.

The ION clinical trials (ION- I , ION- II and ION-III) examined the efficacy of sofosbuvir and ledipasvir co-formulation with and without RBV for 12 to 24 wk in treatment-naïve (16% with cirrhosis) HCV-infected GT1 patients<sup>[31]</sup>. SVR-12 was 97%-99% in ION- I clinical trial. There was no statistically significant difference between the duration of the treatment (12 wk vs 24 wk), HCV sub-GT (GT1a vs GT1b) or RBV use. Even the presence of cirrhosis did not impact the SVR<sup>[32,33]</sup>. Treatment-experienced HCV-infected GT1. Patients were treated with sofosbuvir and ledipasvir co-formulation ± RBV for 12 or 24 wk in ION- II clinical trial. In these patients, addition of RBV did not impact the SVR. Previously treated patients with cirrhosis were the only sub-group that demonstrated a higher SVR with 24 wk of therapy. Therefore, 24 wk of treatment was recommended for previously treated patients with cirrhosis<sup>[34]</sup>. In The ION- III clinical trial, the possibility of shortening the treatment to 8 wk in previously untreated patients without cirrhosis was evaluated. A high number of patients reached SVR in all groups (93% to 95%) without a significant impact of the duration of the treatment or the addition of RBV in the 8-wk treatment<sup>[35]</sup>. Based on secondary analysis, patients with baseline HCV RNA level greater than 6 million international units per milliliter demonstrated a higher risk of relapse with 8 wk of therapy. Therefore, 8 wk of therapy is recommended for treatment-naïve, non-cirrhotic HCV-infected patients with pre-treatment

**Table 2** Direct-acting antiviral-based regimens for treatment-experienced hepatitis C virus-infected patients

Genotype	Recommended regimens options
GT1a	SOF/LDV <sup>1</sup> × 12 wk (no cirrhosis) - 24 wk (cirrhosis) <sup>2</sup>
	PrOD + RBV × 12 wk (no cirrhosis) - 24 wk (cirrhosis)
	SOF + SMV ± RBV × 12 wk (no cirrhosis) - 24 wk (cirrhosis without Q80K variant)
GT1b	SOF + DCV × 12 wk (no cirrhosis) - 24 wk (cirrhosis ± RBV)
	SOF/LDV <sup>1</sup> × 12 wk (no cirrhosis) - 24 wk (cirrhosis)
GT2	PrOD + RBV × 12 wk
	SOF + SMV ± RBV × 12 wk (no cirrhosis) - 24 wk (cirrhosis)
	SOF + DCV × 12 wk (no cirrhosis) - 24 wk (cirrhosis and ± RBV)
GT3	SOF + RBV × 16-24 wk
	SOF + PegIFN + RBV × 12 wk
GT4	SOF + PegIFN + RBV × 12 wk (PegIFN eligible)
	SOF + DCV × 12 wk (no cirrhosis) - 24 wk (cirrhosis ± RBV)
GT5	SOF/LDV × 12 wk
	PrO + RBV × 12 wk
	SOF + RBV × 24 wk
GT6	SOF + PegIFN + RBV × 12 wk
	SOF + SMV ± RBV × 12 wk
GT7	SOF/LDV × 12 wk
	SOF + PegIFN + RBV × 12 wk
GT8	SOF/LDV × 12 wk
	SOF + PegIFN + RBV × 12 wk
GT9	SOF/LDV × 12 wk
	SOF + PegIFN + RBV × 12 wk

<sup>1</sup>Add RBV if previously treated with SOF + RBV or SOF + PegIFN + RBV; <sup>2</sup>Alternative option SOF/LDV + RBV × 12 wk. GT: Genotype; SOF: Sofosbuvir; LDV: Ledipasvir; PrOD: Paritaprevir + ritonavir + ombitasvir + dasabuvir; RBV: Ribavirin; SMV: Simeprevir; DCV: Daclatasvir; PegIFN: Pegylated interferon.

HCV RNA viral load of less than 6 million international units per milliliter<sup>[35]</sup>.

In the COSMOS trial, the SVR to sofosbuvir and simeprevir combination in previous non-responders with METAVIR scores between F0 and F2 was compared to previous non-responders and treatment-naïve patients with METAVIR scores between F3 and F4. The SVR-12 rates were similar in both groups, showing 90% SVR-12 in patients with METAVIR scores F0-F2 and 94% SVR-12 in patients with METAVIR score F3-F4. Neither the duration of the treatment (12 wk vs 24 wk) nor the addition of RBV seemed to influence the SVR<sup>[36]</sup>.

The combination of sofosbuvir and daclatasvir DCV has been safe and effective, both, in previously treated and untreated HCV-patients with GT1<sup>[37,38]</sup>. In previously untreated HCV-infected GT1 patients, a SVR-12 of 98% was achieved with no significant impact of the duration of the treatment (12 wk vs 24 wk) or the addition of RBV<sup>[37]</sup>. In previously treated patients, 24 wk of treatment with sofosbuvir and daclatasvir demonstrated a SVR-12 of 97.5% with no influence from RBV addition<sup>[37]</sup>.

Please refer to Tables 1 (treatment-naïve) and 2 (treatment-experienced) for treatment recommendation by HCV GT with DAA agents<sup>[38-44]</sup>.

## CONCLUSION

The current developments in the treatment of CHC are extraordinary. A significant improvement in efficacy

provided by the DAA agents has been long awaited. In addition to higher efficacy, DAA agents are tolerable with favorable adverse effects profile. Improved efficacy combined with easy tolerability is welcome news for a wide spectrum of patients who were not able to pursue interferon-based antiviral therapy for CHC. Impediments to DAA-based therapy include the high cost of therapy. Efforts are underway to make DAA agents affordable in Asia and Africa. Other issues include a cumbersome insurance authorization process in the United States. Importance of screening patients with risk factors for CHC and linkage to care remains a global issue. It is important to educate the patients that HCV treatment with DAA agents does not confer immunity and exposure to risk factors can lead to re-infection.

## REFERENCES

- 1 **Choo QL**, Kuo G, Weiner AJ, Overby LR, Bradley DW, Houghton M. Isolation of a cDNA clone derived from a blood-borne non-A, non-B viral hepatitis genome. *Science* 1989; **244**: 359-362 [PMID: 2523562 DOI: 10.1126/science.2523562]
- 2 **Mohd Hanafiah K**, Groeger J, Flaxman AD, Wiersma ST. Global epidemiology of hepatitis C virus infection: new estimates of age-specific antibody to HCV seroprevalence. *Hepatology* 2013; **57**: 1333-1342 [PMID: 23172780 DOI: 10.1002/hep.26141]
- 3 **Halliday J**, Klenerman P, Barnes E. Vaccination for hepatitis C virus: closing in on an evasive target. *Expert Rev Vaccines* 2011; **10**: 659-672 [PMID: 21604986 DOI: 10.1586/erv.11.55]
- 4 **Messina JP**, Humphreys I, Flaxman A, Brown A, Cooke GS, Pybus OG, Barnes E. Global distribution and prevalence of hepatitis C virus genotypes. *Hepatology* 2015; **61**: 77-87 [PMID: 25069599 DOI: 10.1002/hep.27259]
- 5 **Marcellin P**. Hepatitis C: the clinical spectrum of the disease. *J Hepatol* 1999; **31** Suppl 1: 9-16 [PMID: 10622554 DOI: 10.1016/S0168-8278(99)80368-7]
- 6 **Lauer GM**, Walker BD. Hepatitis C virus infection. *N Engl J Med* 2001; **345**: 41-52 [PMID: 11439948 DOI: 10.1056/NEJM200107053450107]
- 7 **Zoulim F**, Chevallerier M, Maynard M, Trepo C. Clinical consequences of hepatitis C virus infection. *Rev Med Virol* 2003; **13**: 57-68 [PMID: 12516062 DOI: 10.1002/rmv.371]
- 8 **Manns M**, Pol S, Jacobson IM, Marcellin P, Gordon SC, Peng CY, Chang TT, Everson GT, Heo J, Gerken G, Yoffe B, Towner WJ, Bourliere M, Metivier S, Chu CJ, Sievert W, Bronowicki JP, Thabut D, Lee YJ, Kao JH, McPhee F, Kopit J, Mendez P, Linaberry M, Hughes E, Noviello S. All-oral daclatasvir plus asunaprevir for hepatitis C virus genotype 1b: a multinational, phase 3, multicohort study. *Lancet* 2014; **384**: 1597-1605 [PMID: 25078304 DOI: 10.1016/S0140-6736(14)61059-X]
- 9 **Hézode C**, Fontaine H, Dorival C, Zoulim F, Larrey D, Canva V, De Ledinghen V, Poynard T, Samuel D, Bourliere M, Alric L, Raabe JJ, Zarski JP, Marcellin P, Riachi G, Bernard PH, Loustaud-Ratti V, Chazouilleres O, Abergel A, Guyader D, Metivier S, Tran A, Di Martino V, Causse X, Dao T, Lucidarme D, Portal I, Cacoub P, Gournay J, Grando-Lemaire V, Hillon P, Attali P, Fontanges T, Rosa I, Petrov-Sanchez V, Barthe Y, Pawlowsky JM, Pol S, Carrat F, Bronowicki JP. Effectiveness of telaprevir or boceprevir in treatment-experienced patients with HCV genotype 1 infection and cirrhosis. *Gastroenterology* 2014; **147**: 132-142.e4 [PMID: 24704719 DOI: 10.1053/j.gastro.2014.03.051]
- 10 **Lindenbach BD**, Murray CL, Thiel HJ, Rice CM. Flaviviridae: the viruses and their replication. In DM Knipe and PM Howley (editors). *Fields virology*, 6th ed. Lippincott Williams & Wilkins: New York, NY, 2013
- 11 **Moradpour D**, Penin F, Rice CM. Replication of hepatitis C virus. *Nat Rev Microbiol* 2007; **5**: 453-463 [PMID: 17487147 DOI: 10.1038/nrmicro1645]
- 12 **Tellinghuisen TL**, Evans MJ, von Hahn T, You S, Rice CM. Studying hepatitis C virus: making the best of a bad virus. *J Virol* 2007; **81**: 8853-8867 [PMID: 17522203 DOI: 10.1128/JVI.00753-07]
- 13 **Blight KJ**, Kolykhalov AA, Rice CM. Efficient initiation of HCV RNA replication in cell culture. *Science* 2000; **290**: 1972-1974 [PMID: 11110665 DOI: 10.1126/science.290.5498.1972]
- 14 **Lohmann V**, Körner F, Koch J, Herian U, Theilmann L, Bartenschlager R. Replication of subgenomic hepatitis C virus RNAs in a hepatoma cell line. *Science* 1999; **285**: 110-113 [PMID: 10390360 DOI: 10.1126/science.285.5424.110]
- 15 **Wölk B**, Büchele B, Moradpour D, Rice CM. A dynamic view of hepatitis C virus replication complexes. *J Virol* 2008; **82**: 10519-10531 [PMID: 18715913 DOI: 10.1128/JVI.00640-08]
- 16 **Pawlowsky JM**, Chevaliez S, McHutchison JG. The hepatitis C virus life cycle as a target for new antiviral therapies. *Gastroenterology* 2007; **132**: 1979-1998 [PMID: 17484890 DOI: 10.1053/j.gastro.2007.03.116]
- 17 **Soriano V**, Vispo E, Poveda E, Labarga P, Martin-Carbonero L, Fernandez-Montero JV, Barreiro P. Directly acting antivirals against hepatitis C virus. *J Antimicrob Chemother* 2011; **66**: 1673-1686 [PMID: 21652618 DOI: 10.1093/jac/ckr215]
- 18 **Sofia MJ**. Nucleotide prodrugs for the treatment of HCV infection. *Adv Pharmacol* 2013; **67**: 39-73 [PMID: 23885998 DOI: 10.1016/B978-0-12-405880-4.00002-0]
- 19 **Le Pogam S**, Seshadri A, Kosaka A, Chiu S, Kang H, Hu S, Rajyaguru S, Symons J, Cammack N, Nájera I. Existence of hepatitis C virus NS5B variants naturally resistant to non-nucleoside, but not to nucleoside, polymerase inhibitors among untreated patients. *J Antimicrob Chemother* 2008; **61**: 1205-1216 [PMID: 18343801 DOI: 10.1093/jac/dkn085]
- 20 **Gane EJ**, Hyland RH, An D, Svarovskaia ES, Pang PS, Symonds WT, McHutchinson JG, Stedman CA. High efficacy of LDV/SOF regimens for 12 weeks for patients with HCV genotype 3 or 6 infection. AASLD, Boston, MA, USA, 2014: Abstract LB-11
- 21 **Koff RS**. Review article: the efficacy and safety of sofosbuvir, a novel, oral nucleotide NS5B polymerase inhibitor, in the treatment of chronic hepatitis C virus infection. *Aliment Pharmacol Ther* 2014; **39**: 478-487 [PMID: 24387618 DOI: 10.1111/apt.12601]
- 22 **Appel N**, Schaller T, Penin F, Bartenschlager R. From structure to function: new insights into hepatitis C virus RNA replication. *J Biol Chem* 2006; **281**: 9833-9836 [PMID: 16407182 DOI: 10.1074/jbc.R500026200]
- 23 **He Y**, Staschke KA, Tan SL. HCV NS5A: a multifunctional regulator of cellular pathway and virus replication. In *Hepatitis C Viruses*. In: Tan SL, editor. *Source Hepatitis C Viruses: Genomes and Molecular Biology*. Norfolk (UK): Horizon Bioscience, 2006: Chapter 9 [PMID: 21250384]
- 24 **Szabo G**. Hepatitis C virus NS5A protein--a master regulator? *Gastroenterology* 2006; **130**: 995-999 [PMID: 16530536 DOI: 10.1053/j.gastro.2006.01.072]
- 25 **Huang Y**, Staschke K, De Francesco R, Tan SL. Phosphorylation of hepatitis C virus NS5A nonstructural protein: a new paradigm for phosphorylation-dependent viral RNA replication? *Virology* 2007; **364**: 1-9 [PMID: 17400273 DOI: 10.1016/j.virol.2007.01.042]
- 26 **Backus LI**, Belperio PS, Shahoumian TA, Loomis TP, Mole LA. Effectiveness of sofosbuvir-based regimens in genotype 1 and 2 hepatitis C virus infection in 4026 U.S. Veterans. *Aliment Pharmacol Ther* 2015; **42**: 559-573 [PMID: 26113432 DOI: 10.1111/apt.13300]
- 27 **Höner Zu Siederdisen C**, Maasoumy B, Deterding K, Port K, Sollik L, Mix C, Kirschner J, Cornberg J, Manns MP, Wedemeyer H, Cornberg M. Eligibility and safety of the first interferon-free therapy against hepatitis C in a real-world setting. *Liver Int* 2015; **35**: 1845-1852 [PMID: 25556625 DOI: 10.1111/liv.12774]
- 28 **Dieterich D**, Bacon BR, Flamm SL, Kowdley KV, Milligan S, Tsai N, Younossi ZM, Lawitz E. Evaluation of sofosbuvir and simeprevir-based regimens in the TRIO network: academic and community treatment of a real-world, heterogeneous population. 65th Annual Meeting of the American Association for the Study of

- Liver diseases. Boston, USA, 2014: Abstract 46
- 29 **Jensen DM**, O'Leary JG, Pockros PJ, Sherman KE, Kwo PY, Mailliard ME, Kowdley KV, Muir AJ, Dickson RC, Ramani A, Manns MP, Lok AS, Akuskevich L, Nelson DR, Fried MW. Safety and efficacy of sofosbuvir-containing regimens for hepatitis C: real-world experience in a diverse, longitudinal observational cohort. 65th Annual Meeting of the American Association for the Study of Liver diseases. Boston, USA, 2015: Abstract 45
  - 30 **Feld JJ**, Kowdley KV, Coakley E, Sigal S, Nelson DR, Crawford D, Weiland O, Aguilar H, Xiong J, Pilot-Matias T, DaSilva-Tillmann B, Larsen L, Podsadecki T, Bernstein B. Treatment of HCV with ABT-450/r-ombitasvir and dasabuvir with ribavirin. *N Engl J Med* 2014; **370**: 1594-1603 [PMID: 24720703 DOI: 10.1056/NEJMoa1315722]
  - 31 **Zeuzem S**, Jacobson IM, Baykal T, Marinho RT, Poordad F, Bourlière M, Sulkowski MS, Wedemeyer H, Tam E, Desmond P, Jensen DM, Di Bisceglie AM, Varunok P, Hassanein T, Xiong J, Pilot-Matias T, DaSilva-Tillmann B, Larsen L, Podsadecki T, Bernstein B. Retreatment of HCV with ABT-450/r-ombitasvir and dasabuvir with ribavirin. *N Engl J Med* 2014; **370**: 1604-1614 [PMID: 24720679 DOI: 10.1056/NEJMoa1401561]
  - 32 **Younossi ZM**, Stepanova M, Marcellin P, Afdhal N, Kowdley KV, Zeuzem S, Hunt SL. Treatment with ledipasvir and sofosbuvir improves patient-reported outcomes: Results from the ION-1, -2, and -3 clinical trials. *Hepatology* 2015; **61**: 1798-1808 [PMID: 25627448 DOI: 10.1002/hep.27724]
  - 33 **Afdhal N**, Reddy KR, Nelson DR, Lawitz E, Gordon SC, Schiff E, Nahass R, Ghalib R, Gitlin N, Herring R, Lalezari J, Younes ZH, Pockros PJ, Di Bisceglie AM, Arora S, Subramanian GM, Zhu Y, Dvory-Sobol H, Yang JC, Pang PS, Symonds WT, McHutchison JG, Muir AJ, Sulkowski M, Kwo P. Ledipasvir and sofosbuvir for previously treated HCV genotype 1 infection. *N Engl J Med* 2014; **370**: 1483-1493 [PMID: 24725238]
  - 34 **Afdhal N**, Zeuzem S, Kwo P, Chojkier M, Gitlin N, Puoti M, Romero-Gomez M, Zarski JP, Agarwal K, Buggisch P, Foster GR, Bräu N, Buti M, Jacobson IM, Subramanian GM, Ding X, Mo H, Yang JC, Pang PS, Symonds WT, McHutchison JG, Muir AJ, Mangia A, Marcellin P. Ledipasvir and sofosbuvir for untreated HCV genotype 1 infection. *N Engl J Med* 2014; **370**: 1889-1898 [PMID: 24725239]
  - 35 **Kowdley KV**, Gordon SC, Reddy KR, Rossaro L, Bernstein DE, Lawitz E, Shiffman ML, Schiff E, Ghalib R, Ryan M, Rustgi V, Chojkier M, Herring R, Di Bisceglie AM, Pockros PJ, Subramanian GM, An D, Svarovskaia E, Hyland RH, Pang PS, Symonds WT, McHutchison JG, Muir AJ, Pound D, Fried MW. Ledipasvir and sofosbuvir for 8 or 12 weeks for chronic HCV without cirrhosis. *N Engl J Med* 2014; **370**: 1879-1888 [PMID: 24720702 DOI: 10.1056/NEJMoa1402355]
  - 36 **Lawitz E**, Sulkowski MS, Ghalib R, Rodriguez-Torres M, Younossi ZM, Corregidor A, DeJesus E, Pearlman B, Rabinovitz M, Gitlin N, Lim JK, Pockros PJ, Scott JD, Fevery B, Lambrecht T, Ouwerkerk-Mahadevan S, Callewaert K, Symonds WT, Picchio G, Lindsay KL, Beumont M, Jacobson IM. Simeprevir plus sofosbuvir, with or without ribavirin, to treat chronic infection with hepatitis C virus genotype 1 in non-responders to pegylated interferon and ribavirin and treatment-naïve patients: the COSMOS randomised study. *Lancet* 2014; **384**: 1756-1765 [PMID: 25078309]
  - 37 **Sulkowski MS**, Gardiner DF, Rodriguez-Torres M, Reddy KR, Hassanein T, Jacobson I, Lawitz E, Lok AS, Hineostroza F, Thuluvath PJ, Schwartz H, Nelson DR, Everson GT, Eley T, Wind-Rotolo M, Huang SP, Gao M, Hernandez D, McPhee F, Sherman D, Hindes R, Symonds W, Pasquinelli C, Grasele DM. Daclatasvir plus sofosbuvir for previously treated or untreated chronic HCV infection. *N Engl J Med* 2014; **370**: 211-221 [PMID: 24428467 DOI: 10.1056/NEJMoa1306218]
  - 38 **Kumada H**, Suzuki Y, Ikeda K, Toyota J, Karino Y, Chayama K, Kawakami Y, Ido A, Yamamoto K, Takaguchi K, Izumi N, Koike K, Takehara T, Kawada N, Sata M, Miyagoshi H, Eley T, McPhee F, Damokosh A, Ishikawa H, Hughes E. Daclatasvir plus asunaprevir for chronic HCV genotype 1b infection. *Hepatology* 2014; **59**: 2083-2091 [PMID: 24604476 DOI: 10.1002/hep.27113]
  - 39 **Zeuzem S**, Dusheiko GM, Salupere R, Mangia A, Flisiak R, Hyland RH, Illeperuma A, Svarovskaia E, Brainard DM, Symonds WT, Subramanian GM, McHutchison JG, Weiland O, Reesink HW, Ferenci P, Hézode C, Esteban R. Sofosbuvir and ribavirin in HCV genotypes 2 and 3. *N Engl J Med* 2014; **370**: 1993-2001 [PMID: 24795201 DOI: 10.1056/NEJMoa1316145]
  - 40 **Omata M**, Nishiguchi S, Ueno Y, Mochizuki H, Izumi N, Ikeda F, Toyoda H, Yokosuka O, Nirei K, Genda T, Umemura T, Takehara T, Sakamoto N, Nishigaki Y, Nakane K, Toda N, Ide T, Yanase M, Hino K, Gao B, Garrison KL, Dvory-Sobol H, Ishizaki A, Omote M, Brainard D, Knox S, Symonds WT, McHutchison JG, Yatsushashi H, Mizokami M. Sofosbuvir plus ribavirin in Japanese patients with chronic genotype 2 HCV infection: an open-label, phase 3 trial. *J Viral Hepat* 2014; **21**: 762-768 [PMID: 25196837 DOI: 10.1111/jvh.12312]
  - 41 **Gane EJ**, Hyland RH, An D, Pang PS, Symonds WT, McHutchison JG, Stedman CA. Sofosbuvir/ledipasvir fixed dose combination is safe and effective in difficult-to-treat populations including genotype-3 patients, decompensated genotype-1 patients, and genotype-1 patients with prior sofosbuvir treatment experience. *J Hepatol* 2014; **60**: S1-S22 [DOI: 10.1016/S0168-8278(14)60008-8]
  - 42 **Ruane PJ**, Ain D, Meshrekey R, Riad J, Stryker R, Soliman M, Mikhail S, Wolfe PR, Kersey K, Doehle B, Deyuan J, Symonds WT. Sofosbuvir plus ribavirin, an interferon-free regimen, in the treatment of treatment-naïve and treatment-experienced patients with chronic genotype 4 HCV infection. *J Hepatol* 2014; **60**: S503-S504 [DOI: 10.1016/S0168-8278(14)61403-3]
  - 43 **AASLD/IDSA HCV Guidance Panel**. Hepatitis C guidance: AASLD-IDSA recommendations for testing, managing, and treating adults infected with hepatitis C virus. *Hepatology* 2015; **62**: 932-954 [PMID: 26111063 DOI: 10.1002/hep.27950]
  - 44 **European Association for Study of Liver**. EASL Recommendations on Treatment of Hepatitis C 2015. *J Hepatol* 2015; **63**: 199-236 [PMID: 25911336 DOI: 10.1016/j.jhep.2015.03.025]

**P- Reviewer:** Chiu KW, Kanda T, Malnick SDH, Panduro A

**S- Editor:** Ji FF **L- Editor:** A **E- Editor:** Liu SQ



## Contributions of transgenic mouse studies on the research of hepatitis B virus and hepatitis C virus-induced hepatocarcinogenesis

Shogo Ohkoshi, Haruka Hirono, Kazuhiko Watanabe, Katsuhiko Hasegawa, Masahiko Yano

Shogo Ohkoshi, Haruka Hirono, Kazuhiko Watanabe, Katsuhiko Hasegawa, Department of Internal Medicine, School of Life Dentistry at Niigata, the Nippon Dental University, Niigata-city 951-8580, Japan

Masahiko Yano, Division of Gastroenterology and Hepatology, Graduate School of Medical and Dental Sciences Niigata University, Niigata-city 951-8520, Japan

**Author contributions:** All authors contributed to this manuscript.

**Supported by** A Grant-in-Aid for Scientific Research (C) (25461012 to Shogo Ohkoshi) from the Japan Society for the Promotion of Science (JSPS).

**Conflict-of-interest statement:** The authors do not have any commercial affiliation or consultancy that could be construed as a conflict of interest.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to:** Shogo Ohkoshi, MD, PhD, Department of Internal Medicine, School of Life Dentistry at Niigata, the Nippon Dental University, 1-8 Hamaura-Cho, Chuo-ku, Niigata-city 951-8580, Japan. [okoshi@ngt.ndu.ac.jp](mailto:okoshi@ngt.ndu.ac.jp)  
Telephone: +81-25-2118243  
Fax: +81-25-2671582

Received: July 22, 2015  
Peer-review started: July 27, 2015  
First decision: September 22, 2015  
Revised: September 28, 2015  
Accepted: November 24, 2015  
Article in press: November 25, 2015  
Published online: December 8, 2015

### Abstract

Transgenic mouse technology has enabled the investigation of the pathogenic effects, including those on development, immunological reactions and carcinogenesis, of viral genes directly in living organism in a real-time manner. Although viral hepatocarcinogenesis comprises multiple sequences of pathological events, that is, chronic necroinflammation and the subsequent regeneration of hepatocytes that induces the accumulation of genetic alterations and hepatocellular carcinoma (HCC), the direct action of viral proteins also play significant roles. The pathogenesis of hepatitis B virus X and hepatitis C virus (HCV) core genes has been extensively studied by virtue of their functions as a transactivator and a steatosis inducer, respectively. In particular, the mechanism of steatosis in HCV infection and its possible association with HCC has been well studied using HCV core gene transgenic mouse models. Although transgenic mouse models have remarkable advantages, they are intrinsically accompanied by some drawbacks when used to study human diseases. Therefore, the results obtained from transgenic mouse studies should be carefully interpreted in the context of whether or not they are well associated with human pathogenesis.

**Key words:** Transgenic mouse; Hepatocarcinogenesis; Hepatitis C virus; Hepatitis B virus X; Hepatitis B virus; Hepatitis C virus core protein; Steatosis

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Transgenic technology offers researchers several advantages over *in vitro* experiments, including the ability to trace the pathogenic effects of viral genes in living organisms. Transgenic mouse studies have provided evidence that the direct action of viral genes, especially the genes encoding hepatitis B virus

X and hepatitis C virus core proteins, is involved in hepatocarcinogenesis. However, such results should be considered carefully as transgenic mouse experiments have intrinsic advantages and drawbacks. As such, the results including phenotypes and molecular mechanisms from transgenic mouse studies must always be verified by comparing them to those of human studies for evidence of an association.

Ohkoshi S, Hirono H, Watanabe K, Hasegawa K, Yano M. Contributions of transgenic mouse studies on the research of hepatitis B virus and hepatitis C virus-induced hepatocarcinogenesis. *World J Hepatol* 2015; 7(28): 2834-2840 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i28/2834.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i28.2834>

## INTRODUCTION

Primary liver cancer has a high mortality rate worldwide and ranks 5<sup>th</sup> as the most common cancer among men and 7<sup>th</sup> among women; as such, therapeutic options to cure this disease are urgently needed<sup>[1,2]</sup>. Hepatocellular carcinoma (HCC) accounts for the majority of primary liver cancers. The most prevalent etiological agents of HCC are hepatitis B virus (HBV) and hepatitis C virus (HCV) infections. Once HBV- or HCV-related cirrhosis is established, HCC develops with an annual rate of approximately 4.3% or 7.1%, respectively in Japan, and 2.2% or 3.7%, respectively, in Western countries<sup>[3,4]</sup>.

Individual viral proteins may play significant roles in and confer characteristics that are peculiar to the viral pathogenesis of HBV or HCV. Transgenic mouse strategies are applied to explore the functions of viral genes *in vivo* and can provide significant information on the mechanisms of viral pathogenesis by allowing the elucidation of the effects of individual viral proteins<sup>[5-7]</sup>. In this review, we summarize the past contributions of transgenic mouse studies to HBV- and HCV-induced viral pathogenesis with a particular focus on the evaluation of how well the results of these transgenic mouse studies correlate with human pathogenesis and how useful they are in the development of therapeutic strategies for viral disease, especially hepatocarcinogenesis.

## MECHANISMS OF HEPATOCARCINOGENESIS DUE TO HBV AND HCV

When considering the contribution of transgenic mouse studies to the research on hepatocarcinogenesis caused by HBV or HCV, it is important to outline the mechanisms of the disease and consider for which mechanisms transgenic mouse studies can be applied to provide pathogenic and therapeutic contributions.

First, the common mechanism between HBV and HCV is as follows: Once these viruses infect liver, they

skillfully evade host immune surveillance and induce chronic necroinflammation. These injuries cause fibrosis and result in liver cirrhosis. Hepatocytes, using their intrinsic regenerative capability, continue to proliferate in order to compensate for the necrotic tissues. Genetic alterations continuously accumulate during these processes, resulting in the formation of a pathogenic state such as cirrhosis from which HCC frequently arises.

Second, viral genes may be involved in hepatocarcinogenesis by directly affecting cellular machineries. The most representative genes of this type that have drawn clinical attention are the genes for HBV X (HBx) and HCV core protein. HBx is multifunctional and may induce the transactivation of many cellular genes<sup>[8]</sup>. On the other hand, the HCV core protein causes steatosis in the liver and subsequent HCC<sup>[9]</sup>. Transgenic mouse studies can shed light on the mechanisms of HBx and HCV core protein by enabling assays on the direct actions of these viral genes *in vivo*.

Third, a mechanism specific to HBV is its integration into the cellular DNA of the host; this may increase the genomic instability and cis-activation of the adjacent cellular genome that may possibly be involved in the regulation of the cell cycle<sup>[10]</sup>. Importantly, most integrated viral DNA retain the sequences encoding HBxAg, and the HBxAg expressed from the integrated HBV DNA further promotes genetic instability of the host by a variety of mechanisms<sup>[10]</sup>.

## ADVANTAGES AND LIMITATIONS OF RESEARCH USING TRANSGENIC MICE

### Advantages

The mechanism of viral hepatocarcinogenesis comprises a number of complex factors<sup>[11]</sup>. Although the majority of research in this field has been performed using human samples, human resources have limitations in both ethical and quantitative terms, and it is sometimes difficult to extract significant conclusions from the final results that are dependent of a combination of vastly complex molecular events. The cell lines used for *in vitro* experiments are mainly derived from HCC cells in which carcinogenic events have already finished and little information on the real-time carcinogenic process can be obtained. Transgenic mouse research can compensate for these drawbacks.

### Limitations

Transgenic technology offers the researchers several advantages over *in vitro* experiments<sup>[12,13]</sup>: It enables the investigation of the pathogenic effects, including those on development, immunological reactions, and carcinogenesis, of viral genes directly in living organisms in a real-time manner. Viral genes can be designed to be placed under their own or another appropriate promoter for specific expression in permissive cell types or tissues<sup>[14,15]</sup>. Thus, it is possible to identify particular cell types or organs that allow the expression of viral

proteins. In addition, it allows a sufficient number or amount of experimental material to be obtained at any specific condition, enabling the analysis of comprehensive and objective data.

Unfortunately, there are also several drawbacks to transgenic mouse research. First, because viral proteins of HBV or HCV, which normally only infect human or chimpanzee, are forced to be expressed in mouse tissue, the resulting protein-protein interactions may not be the same as those that would occur in the natural hosts and unexpected molecular reactions may result. Second, expressed viral proteins may become immune-tolerant in mice and cannot induce immunological response<sup>[16]</sup>, and as eliciting an immune response is the main mechanism of viral liver pathogenesis, it becomes difficult to completely simulate this human disease; however, this may actually be an advantage as it may allow the evaluation of the direct role of viral proteins on liver pathogenesis without local inflammation<sup>[6]</sup>. Third, because transgenes are randomly integrated into the genome, the expression of transgene can be affected by the adjacent genomic structures. Phenotypes should be confirmed by the results obtained in mouse lines established using several founder mice independently. It is extremely difficult to establish viral replication in mice and cell culture systems are more suitable for analyzing viral replication, including the identification of viral receptors. Thus, because transgenic mouse models for viral diseases are not multipotent and can produce results that are easily hampered by experimental artifact, the obtained results should always be strictly scrutinized and evaluated in both scientific and clinical aspects. Namely, confirmation as to whether the phenotypes really correlate with the pathogenesis of human liver diseases is needed.

## HBV AND TRANSGENIC MOUSE

HBV is a DNA virus with a length of 3.2 kb that replicates *via* an RNA intermediate for viral replication. Similar to retroviruses, which undergo reverse transcription for replication, it is integrated into the host genome.

### **Hepatitis B surface antigen transgenic mouse: Immunopathology of HBV**

Chisari's group has made vast contributions to the clinical field by performing transgenic mouse studies to investigate immune-mediated hepatitis and hepatocarcinogenesis<sup>[17,18]</sup>. In the 1980s, they produced a transgenic mouse model that overexpressed the large protein of the hepatitis B surface antigen (HBsAg). It induced severe, prolonged hepatocellular injury that was characterized by inflammation and regenerative hyperplasia, resulting in the development of HCC<sup>[19]</sup>. This was the first transgenic mouse model in which the development of HCC was observed from the function of a single viral protein. However, these results were considered to be the outcome of the storage effect of the large HBsAg, since overexpression of the HBV

core, precore, X, small or middle envelope protein was not associated with any evidence of liver disease in the transgenic mouse model due to immunological tolerance for the inherently expressed viral proteins<sup>[16]</sup>. In order to better mimic the HBV pathogenesis seen in human where immunological reactions to viral proteins are essential, HBsAg-specific cytotoxic T cells were transferred to the mice to induce viral antigen-mediated acute hepatitis; this provided direct evidence that hepatocellular injury in HBV infection may be immunologically mediated<sup>[20,21]</sup>. Moreover, further induction of cytokines such as interferon (IFN)- $\gamma$  after acute liver injury may magnify the degree of inflammation, resulting in fulminant hepatitis<sup>[22]</sup>. These transgenic mouse studies clarified the immunological aspects of HBV infection, showing that the balance between viral load and the strength of the immunological reactivity towards HBV antigen determines the fate of the disease<sup>[18]</sup>. As for the immune pathogenesis of HCC, the adoptive transfer of CD8 lymphocytes to HBsAg transgenic mice generated a pathogenesis that closely resembled that of human chronic viral hepatitis and finally resulted in the development of HCC<sup>[23]</sup>. This model strengthened the notion that only immune-mediated hepatocellular injury, and not insertional mutagenesis from the integration of the HBV genome or the expression of the *HBx* gene, could cause hepatocarcinogenesis.

In the early era of transgenic mouse studies, several reports showed a high level of HBV replication *in vivo*, injecting a duplicated HBV plasmid<sup>[24-26]</sup>. These results demonstrated that the HBV genome integrated into the mouse chromosome acted as a template for viral gene expression, allowing viral replication. Although these mice did not show the HCC phenotype, they contributed to the detailed studies of the replication and expression of HBV and to pathological studies of hepatitis. In addition, HBs large envelope protein expression in transgenic mouse showed the inhibition of HBsAg secretion, suggesting an inhibitory effect of the pre-S-containing domain of the large envelope peptide<sup>[27]</sup>.

### **HBxAg transgenic mouse: Direct hepatocarcinogenic role of HBV**

The *HBx* gene is a known oncogene with pleiotropic functions that transactivate multiple cellular genes; it has attracted extensive clinical attention and has been regarded as a suitable target for transgenic mouse research<sup>[8,28,29]</sup>. Kim *et al.*<sup>[30]</sup> first observed the occurrence of HCC in HBx transgenic mouse. Koike *et al.*<sup>[31,32]</sup> further showed that HBx induced hepatocyte proliferation and contributed to hepatocarcinogenesis. Inspired by these studies, many researchers have attempted to produce the HCC phenotype in HBx transgenic mice, but most of them failed<sup>[33-36]</sup>. These inconsistent observations in terms of HCC development among transgenic mice might be partly due to differences in the sequences<sup>[37]</sup> or subtypes<sup>[38]</sup> of HBV or the genetic background of the mice used. However, strong and continuous expression of HBx might be a requirement for observing the HCC

phenotype<sup>[38,39]</sup>. Nonetheless, it remains unclear whether this high level of HBx expression is physiologically relevant to human pathogenesis<sup>[8]</sup>. Moreover, there is still a lack of clinical and molecular evaluation methods that can be used to measure how much the direct carcinogenetic function of HBx contributes to human HBV hepatocarcinogenesis where necrosis-regeneration sequence of hepatocytes by immunological reactions to HBV is generally considered to be an essential mechanism. How a single HBx gene is involved in this huge complex pathogenesis process remains to be clarified.

It has been reported that fibrosis levels of liver complicated with HBV-HCC is milder than that of HCV-HCC<sup>[40-42]</sup>. In our cohort of 57 patients with HBV-HCC who were treated surgically, 25 (44%) did not have cirrhosis, and 22 (39%) had a mild level of fibrosis (F1 or 2) (Unpublished results). Therefore, the direct carcinogenic role of HBx or the dysregulation of the cell cycle due to insertional mutagenesis by the HBV genome might play a large role, especially in those who have a mild level of fibrosis, than cirrhosis, which is the final state of necrosis-regeneration sequence. In addition, our past studies have shown that the HBx transgenic mouse is a good model for testing the anti-hepatocarcinogenic function of IFN- $\beta$ <sup>[29,43]</sup>.

## HCV AND TRANSGENIC MOUSE

### **HCV and steatosis: Close association with human pathogenesis**

While HBx transgenic mouse research has been confronted by difficulties in correlating the research results with specific clinical or molecular landmarks, HCV transgenic mice have provided fruitful experimental observations in terms of steatosis<sup>[6,44]</sup> which is commonly observed in both HCV-infected humans and HCV-transgenic mice.

Steatosis has been reported to be a characteristic finding of chronic HCV infection<sup>[45-47]</sup>. Moriya *et al*<sup>[9]</sup> observed steatosis in HCV core gene transgenic mice at as early as 3 mo of age and found that about a quarter of the mice developed HCC in their late life, demonstrating that the HCV core protein itself has a direct role in hepatocarcinogenesis by virtue of steatosis<sup>[9]</sup>. Lerat *et al*<sup>[48]</sup> also observed hepatic steatosis and HCC in transgenic mouse models that express complete viral and structural proteins without immunological reactions.

It is well known that HCV genotype 3 directly induce steatosis in liver<sup>[44,49]</sup>, supporting the observations obtained in transgenic mouse. Importantly, an association between steatosis and fibrosis has also been demonstrated in a meta-analysis of chronically infected patients with HCV<sup>[50]</sup>, and hepatic steatosis is a risk factor for HCC in chronic hepatitis C patients<sup>[51,52]</sup>. These results indicate that the findings obtained from transgenic research are well associated with findings from human research.

### **HCV core protein and PRAR $\alpha$**

In addition, HCV core gene transgenic mice became the base for further studies which explored the mechanisms of steatosis and its relationship with HCC development. Tanaka *et al*<sup>[53]</sup> generated peroxisome proliferator-activated receptor alpha (PRAR $\alpha$ -homozygous, -heterozygous, and -null mice with HCV core protein expression and showed that severe steatosis developed in mice that had both PRAR $\alpha$  alleles, revealing that the expression of PRAR $\alpha$ , which is important in maintaining triglyceride homeostasis, was essential for the development of HCV core protein-induced steatosis and HCC<sup>[53]</sup>. Moriishi *et al*<sup>[54]</sup> showed that a knockout of the proteasome activator 28 gamma (PA28 $\gamma$ ) gene induces the accumulation of HCV core protein in the nucleus of hepatocytes of HCV core gene transgenic mice and disrupts the development of both hepatic steatosis and HCC, thus revealing that PA28 $\gamma$  plays a crucial role in the development of HCV-induced liver pathogenesis<sup>[54]</sup>.

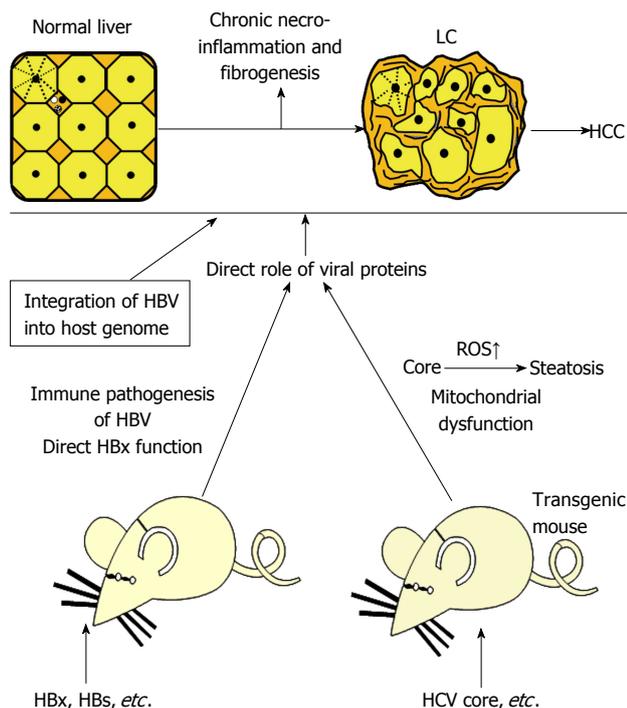
### **HCV core protein and reactive oxygen species**

Transgenic mouse studies have further provided significant findings on the mechanisms of the progression from steatosis to hepatocarcinogenesis. Okuda *et al*<sup>[55]</sup>, reported that HCV core protein increases the production of reactive oxygen species (ROS) *via* a direct effect on the mitochondrial electron transport system. Korenaga *et al*<sup>[56]</sup> reported that reduced activity of electron transport complex 1 enhances the production of ROS in HCV core gene transgenic mouse. Consistent with these observations, both mitochondrial dysfunction<sup>[57]</sup> and high levels of oxidative stress have been demonstrated in HCV-infected patients<sup>[58,59]</sup>. 8-Hydroxy-2'-deoxy-guanosine which is generated by ROS and leads to an increased frequency of mutations, accumulates in HCV core gene transgenic mouse<sup>[60]</sup> and causes mutations in cellular genes<sup>[61]</sup>. In addition to its direct effect on mitochondria, HCV core protein has been shown to cause endoplasmic reticulum stress that results in an oxidized redox state in hepatocytes, interfering with immune responses and potentiating fibrosis and carcinogenesis<sup>[62]</sup>. Moreover, Klopstock *et al*<sup>[63]</sup> used HCV transgenic mice that were crossed with Mdr2-knockout mice to demonstrate that the HCV transgene accelerates inflammation-associated hepatocarcinogenesis, which has a pathogenesis similar to that of human HCV-induced carcinogenesis<sup>[63]</sup>.

It was also reported in a transgenic mouse study that the production of ROS induces high levels of iron deposition in liver, resulting in an increased risk of HCC<sup>[64]</sup>. A strong correlation between hepatic DNA damage and iron overload has been confirmed in a human study<sup>[65]</sup>. Mitochondrial ROS may be linked to metabolic disorders such as insulin resistance, hepatic steatosis, and hepatic iron accumulation, all of which are characteristic features of chronic HCV infection<sup>[66]</sup>.

### **Direct role of HCV core protein in hepatocarcinogenesis**

The mainstream mechanism of HCV-induced hepato-



**Figure 1** Outline of the overall aspects of this review. The mainstream mechanism of HBV- or HCV-induced hepatocarcinogenesis is necroinflammation and hepatocyte-regeneration sequences that eventually result in cirrhosis where HCC frequently develops. However, transgenic mouse studies have clarified the significant contribution of viral proteins to hepatocarcinogenesis by directly affecting cellular machinery. A close association of HCV core protein and hepatic steatosis has been established by transgenic mouse studies. In addition to the inherent insertional mutagenesis mechanism of HBV, results from transgenic mouse studies have also suggested the direct involvement of HBV proteins in hepatocarcinogenesis. HBV: Hepatitis B virus; HCV: Hepatitis C virus; HCC: Hepatocellular carcinoma; ROS: Reactive oxygen species; HBs: Hepatitis B surface; HBx: Hepatitis B virus X.

carcinogenesis is persistent necroinflammation that induces the irregular regeneration of hepatocytes, allowing the accumulation of genetic or epigenetic alterations. In addition to this, transgenic mouse studies have shown that the HCV core protein plays a significant role in hepatocarcinogenesis by inducing steatosis *via* the production of ROS from by the dysregulation of lipid metabolism or functional abnormalities of the mitochondria<sup>[5]</sup>. Thus, transgenic mouse studies have shown that viral proteins, especially the HCV core protein, directly interact with lipid-metabolizing pathways and contribute to HCC development; these are the major achievements of transgenic mouse studies on HCV-induced carcinogenesis.

## CONCLUSION

The mechanisms of hepatocarcinogenesis common between HBV and HCV include persistent necroinflammation and the regeneration of hepatocytes that allows the accumulation of genetic changes. However, there are yet no reports on common genetic changes that can fully explain these complex pathways; rather, multiple dysfunctions resulting from abnormalities in a number

of signal transduction pathways appear to converge to produce the common HCC phenotype. However, the use of transgenic mouse technology has clarified that even a single viral gene, such as the gene for HBx or HCV core protein can directly affect the cellular machinery and impact the mainstream mechanism of persistent necroinflammation-induced hepatocarcinogenesis (Figure 1). Especially for HCV, a good correlation has been found between the experimental findings from transgenic mouse studies and clinical observations. Thus, transgenic mouse models may provide an efficient method for evaluating the effectiveness of anti-hepatocarcinogenesis agents in the future.

## REFERENCES

- 1 **El-Serag HB.** Epidemiology of viral hepatitis and hepatocellular carcinoma. *Gastroenterology* 2012; **142**: 1264-1273.e1 [PMID: 22537432 DOI: 10.1053/j.gastro.2011.12.061]
- 2 **Forner A, Llovet JM, Bruix J.** Hepatocellular carcinoma. *Lancet* 2012; **379**: 1245-1255 [PMID: 22353262 DOI: 10.1016/s0140-6736(11)61347-0]
- 3 **Fattovich G, Stroffolini T, Zagni I, Donato F.** Hepatocellular carcinoma in cirrhosis: incidence and risk factors. *Gastroenterology* 2004; **127**: S35-S50 [PMID: 15508101]
- 4 **Fattovich G, Pantalena M, Zagni I, Realdi G, Schalm SW, Christensen E.** Effect of hepatitis B and C virus infections on the natural history of compensated cirrhosis: a cohort study of 297 patients. *Am J Gastroenterol* 2002; **97**: 2886-2895 [PMID: 12425564 DOI: 10.1111/j.1572-0241.2002.07057.x]
- 5 **Koike K.** Hepatitis C virus contributes to hepatocarcinogenesis by modulating metabolic and intracellular signaling pathways. *J Gastroenterol Hepatol* 2007; **22** Suppl 1: S108-S111 [PMID: 17567457 DOI: 10.1111/j.1440-1746.2006.04669.x]
- 6 **Lerat H, Higgs M, Pawlotsky JM.** Animal models in the study of hepatitis C virus-associated liver pathologies. *Expert Rev Gastroenterol Hepatol* 2011; **5**: 341-352 [PMID: 21651352 DOI: 10.1586/egh.11.14]
- 7 **Li Y, Tang ZY, Hou JX.** Hepatocellular carcinoma: insight from animal models. *Nat Rev Gastroenterol Hepatol* 2012; **9**: 32-43 [PMID: 22025031 DOI: 10.1038/nrgastro.2011.196]
- 8 **Slagle BL, Andrisani OM, Bouchard MJ, Lee CG, Ou JH, Siddiqui A.** Technical standards for hepatitis B virus X protein (HBx) research. *Hepatology* 2015; **61**: 1416-1424 [PMID: 25099228 DOI: 10.1002/hep.27360]
- 9 **Moriya K, Fujie H, Shintani Y, Yotsuyanagi H, Tsutsumi T, Ishibashi K, Matsuura Y, Kimura S, Miyamura T, Koike K.** The core protein of hepatitis C virus induces hepatocellular carcinoma in transgenic mice. *Nat Med* 1998; **4**: 1065-1067 [PMID: 9734402 DOI: 10.1038/2053]
- 10 **Feitelson MA, Lee J.** Hepatitis B virus integration, fragile sites, and hepatocarcinogenesis. *Cancer Lett* 2007; **252**: 157-170 [PMID: 17188425 DOI: 10.1016/j.canlet.2006.11.010]
- 11 **Farazi PA, DePinho RA.** Hepatocellular carcinoma pathogenesis: from genes to environment. *Nat Rev Cancer* 2006; **6**: 674-687 [PMID: 16929323 DOI: 10.1038/nrc1934]
- 12 **Hinrichs SH, Vogel J, Rhim MJ, Jay G.** Use of transgenic animals to study human retroviruses. *Cancer Metastasis Rev* 1988; **7**: 311-320 [PMID: 3145157]
- 13 **Jaenisch R.** Transgenic animals. *Science* 1988; **240**: 1468-1474 [PMID: 3287623]
- 14 **Okoshi S, Takeda Y, Takimoto M, Shimotohno K, Asakura H, Jay G.** Molecular dissection of hepatitis C virus expression. *Intervirology* 2001; **44**: 21-28 [PMID: 11223716]
- 15 **Takeda Y, Okoshi S, Suzuki K, Yano M, Gangemi JD, Jay G, Asakura H, Aoyagi Y.** Effect of interferon alpha and cell cycle progression on translation mediated by the hepatitis C virus 5'

- untranslated region: a study using a transgenic mouse model. *J Viral Hepat* 2004; **11**: 33-44 [PMID: 14738556]
- 16 **Chisari FV**, Pinkert CA, Milich DR, Filippi P, McLachlan A, Palmiter RD, Brinster RL. A transgenic mouse model of the chronic hepatitis B surface antigen carrier state. *Science* 1985; **230**: 1157-1160 [PMID: 3865369]
  - 17 **Chisari FV**. Hepatitis B virus transgenic mice: insights into the virus and the disease. *Hepatology* 1995; **22**: 1316-1325 [PMID: 7557887]
  - 18 **Chisari FV**. Rous-Whipple Award Lecture. Viruses, immunity, and cancer: lessons from hepatitis B. *Am J Pathol* 2000; **156**: 1117-1132 [PMID: 10751335]
  - 19 **Chisari FV**, Klopchin K, Moriyama T, Pasquinelli C, Dunsford HA, Sell S, Pinkert CA, Brinster RL, Palmiter RD. Molecular pathogenesis of hepatocellular carcinoma in hepatitis B virus transgenic mice. *Cell* 1989; **59**: 1145-1156 [PMID: 2598264]
  - 20 **Ando K**, Guidotti LG, Wirth S, Ishikawa T, Missale G, Moriyama T, Schreiber RD, Schlicht HJ, Huang SN, Chisari FV. Class I-restricted cytotoxic T lymphocytes are directly cytopathic for their target cells in vivo. *J Immunol* 1994; **152**: 3245-3253 [PMID: 8144915]
  - 21 **Moriyama T**, Guillhot S, Klopchin K, Moss B, Pinkert CA, Palmiter RD, Brinster RL, Kanagawa O, Chisari FV. Immunobiology and pathogenesis of hepatocellular injury in hepatitis B virus transgenic mice. *Science* 1990; **248**: 361-364 [PMID: 1691527]
  - 22 **Ando K**, Moriyama T, Guidotti LG, Wirth S, Schreiber RD, Schlicht HJ, Huang SN, Chisari FV. Mechanisms of class I restricted immunopathology. A transgenic mouse model of fulminant hepatitis. *J Exp Med* 1993; **178**: 1541-1554 [PMID: 8228807]
  - 23 **Nakamoto Y**, Guidotti LG, Kuhlen CV, Fowler P, Chisari FV. Immune pathogenesis of hepatocellular carcinoma. *J Exp Med* 1998; **188**: 341-350 [PMID: 9670046]
  - 24 **Araki K**, Miyazaki J, Hino O, Tomita N, Chisaka O, Matsubara K, Yamamura K. Expression and replication of hepatitis B virus genome in transgenic mice. *Proc Natl Acad Sci USA* 1989; **86**: 207-211 [PMID: 2911569]
  - 25 **Guidotti LG**, Matzke B, Schaller H, Chisari FV. High-level hepatitis B virus replication in transgenic mice. *J Virol* 1995; **69**: 6158-6169 [PMID: 7666518]
  - 26 **Wirth S**, Guidotti LG, Ando K, Schlicht HJ, Chisari FV. Breaking tolerance leads to autoantibody production but not autoimmune liver disease in hepatitis B virus envelope transgenic mice. *J Immunol* 1995; **154**: 2504-2515 [PMID: 7868916]
  - 27 **Chisari FV**, Filippi P, McLachlan A, Milich DR, Riggs M, Lee S, Palmiter RD, Pinkert CA, Brinster RL. Expression of hepatitis B virus large envelope polypeptide inhibits hepatitis B surface antigen secretion in transgenic mice. *J Virol* 1986; **60**: 880-887 [PMID: 3783819]
  - 28 **Ng SA**, Lee C. Hepatitis B virus X gene and hepatocarcinogenesis. *J Gastroenterol* 2011; **46**: 974-990 [PMID: 21647825 DOI: 10.1007/s00535-011-0415-9]
  - 29 **Yamazaki K**, Suzuki K, Ohkoshi S, Yano M, Kurita S, Aoki YH, Toba K, Takamura MA, Yamagiwa S, Matsuda Y, Aoyagi Y. Temporal treatment with interferon-beta prevents hepatocellular carcinoma in hepatitis B virus X gene transgenic mice. *J Hepatol* 2008; **48**: 255-265 [PMID: 18083266 DOI: 10.1016/j.jhep.2007.09.012]
  - 30 **Kim CM**, Koike K, Saito I, Miyamura T, Jay G. HBx gene of hepatitis B virus induces liver cancer in transgenic mice. *Nature* 1991; **351**: 317-320 [PMID: 2034275 DOI: 10.1038/351317a0]
  - 31 **Koike K**, Moriya K, Iino S, Yotsuyanagi H, Endo Y, Miyamura T, Kurokawa K. High-level expression of hepatitis B virus HBx gene and hepatocarcinogenesis in transgenic mice. *Hepatology* 1994; **19**: 810-819 [PMID: 8138251]
  - 32 **Koike K**, Moriya K, Yotsuyanagi H, Shintani Y, Fujie H, Tsutsumi T, Kimura S. Compensatory apoptosis in preneoplastic liver of a transgenic mouse model for viral hepatocarcinogenesis. *Cancer Lett* 1998; **134**: 181-186 [PMID: 10025879]
  - 33 **Balsano C**, Billet O, Bennoun M, Cavard C, Zider A, Grimber G, Natoli G, Briand P, Levrero M. Hepatitis B virus X gene product acts as a transactivator in vivo. *J Hepatol* 1994; **21**: 103-109 [PMID: 7963409]
  - 34 **Lee TH**, Finegold MJ, Shen RF, DeMayo JL, Woo SL, Butel JS. Hepatitis B virus transactivator X protein is not tumorigenic in transgenic mice. *J Virol* 1990; **64**: 5939-5947 [PMID: 2243380]
  - 35 **Perfumo S**, Amicone L, Colloca S, Giorgio M, Pozzi L, Tripodi M. Recognition efficiency of the hepatitis B virus polyadenylation signals is tissue specific in transgenic mice. *J Virol* 1992; **66**: 6819-6823 [PMID: 1357192]
  - 36 **Reifenberg K**, Löhler J, Pudollek HP, Schmitteckert E, Spindler G, Köck J, Schlicht HJ. Long-term expression of the hepatitis B virus core-e- and X-proteins does not cause pathologic changes in transgenic mice. *J Hepatol* 1997; **26**: 119-130 [PMID: 9148002]
  - 37 **Kwun HJ**, Jang KL. Natural variants of hepatitis B virus X protein have differential effects on the expression of cyclin-dependent kinase inhibitor p21 gene. *Nucleic Acids Res* 2004; **32**: 2202-2213 [PMID: 15107488 DOI: 10.1093/nar/gkh553]
  - 38 **Koike K**. Hepatocarcinogenesis in hepatitis viral infection: lessons from transgenic mouse studies. *J Gastroenterol* 2002; **37** Suppl 13: 55-64 [PMID: 12109667]
  - 39 **Yu DY**, Moon HB, Son JK, Jeong S, Yu SL, Yoon H, Han YM, Lee CS, Park JS, Lee CH, Hyun BH, Murakami S, Lee KK. Incidence of hepatocellular carcinoma in transgenic mice expressing the hepatitis B virus X-protein. *J Hepatol* 1999; **31**: 123-132 [PMID: 10424292]
  - 40 **Shiratori Y**, Shiina S, Imamura M, Kato N, Kanai F, Okudaira T, Teratani T, Tohgo G, Toda N, Ohashi M. Characteristic difference of hepatocellular carcinoma between hepatitis B- and C- viral infection in Japan. *Hepatology* 1995; **22**: 1027-1033 [PMID: 7557847]
  - 41 **Takano S**, Yokosuka O, Imazeki F, Tagawa M, Omata M. Incidence of hepatocellular carcinoma in chronic hepatitis B and C: a prospective study of 251 patients. *Hepatology* 1995; **21**: 650-655 [PMID: 7875662]
  - 42 **Lok AS**. Hepatitis B: liver fibrosis and hepatocellular carcinoma. *Gastroenterol Clin Biol* 2009; **33**: 911-915 [PMID: 19577871 DOI: 10.1016/j.gcb.2009.06.001]
  - 43 **Yano M**, Ohkoshi S, Aoki YH, Takahashi H, Kurita S, Yamazaki K, Suzuki K, Yamagiwa S, Sanpei A, Fujimaki S, Wakai T, Kudo SE, Matsuda Y, Aoyagi Y. Hepatitis B virus X induces cell proliferation in the hepatocarcinogenesis via up-regulation of cytoplasmic p21 expression. *Liver Int* 2013; **33**: 1218-1229 [PMID: 23590292 DOI: 10.1111/liv.12176]
  - 44 **Negro F**, Sanyal AJ. Hepatitis C virus, steatosis and lipid abnormalities: clinical and pathogenic data. *Liver Int* 2009; **29** Suppl 2: 26-37 [PMID: 19187070 DOI: 10.1111/j.1478-3231.2008.01950.x]
  - 45 **Bach N**, Thung SN, Schaffner F. The histological features of chronic hepatitis C and autoimmune chronic hepatitis: a comparative analysis. *Hepatology* 1992; **15**: 572-577 [PMID: 1551632]
  - 46 **Czaja AJ**, Carpenter HA. Sensitivity, specificity, and predictability of biopsy interpretations in chronic hepatitis. *Gastroenterology* 1993; **105**: 1824-1832 [PMID: 8253358]
  - 47 **Wong VS**, Wight DG, Palmer CR, Alexander GJ. Fibrosis and other histological features in chronic hepatitis C virus infection: a statistical model. *J Clin Pathol* 1996; **49**: 465-469 [PMID: 8763259]
  - 48 **Lerat H**, Honda M, Beard MR, Loesch K, Sun J, Yang Y, Okuda M, Gosert R, Xiao SY, Weinman SA, Lemon SM. Steatosis and liver cancer in transgenic mice expressing the structural and nonstructural proteins of hepatitis C virus. *Gastroenterology* 2002; **122**: 352-365 [PMID: 11832450]
  - 49 **Rubbia-Brandt L**, Quadri R, Abid K, Giostra E, Malé PJ, Mentha G, Spahr L, Zarski JP, Borisch B, Hadengue A, Negro F. Hepatocyte steatosis is a cytopathic effect of hepatitis C virus genotype 3. *J Hepatol* 2000; **33**: 106-115 [PMID: 10905593]
  - 50 **Leandro G**, Mangia A, Hui J, Fabris P, Rubbia-Brandt L, Colloredo G, Adinolfi LE, Asselah T, Jonsson JR, Smedile A, Terrault N, Paziienza V, Giordani MT, Giostra E, Sonzogni A, Ruggiero G, Marcellin P, Powell EE, George J, Negro F. Relationship between steatosis, inflammation, and fibrosis in chronic hepatitis C: a meta-analysis of individual patient data. *Gastroenterology* 2006; **130**:

- 1636-1642 [PMID: 16697727 DOI: 10.1053/j.gastro.2006.03.014]
- 51 **Ohata K**, Hamasaki K, Toriyama K, Matsumoto K, Saeki A, Yanagi K, Abiru S, Nakagawa Y, Shigeno M, Miyazoe S, Ichikawa T, Ishikawa H, Nakao K, Eguchi K. Hepatic steatosis is a risk factor for hepatocellular carcinoma in patients with chronic hepatitis C virus infection. *Cancer* 2003; **97**: 3036-3043 [PMID: 12784339 DOI: 10.1002/cncr.11427]
- 52 **Pekow JR**, Bhan AK, Zheng H, Chung RT. Hepatic steatosis is associated with increased frequency of hepatocellular carcinoma in patients with hepatitis C-related cirrhosis. *Cancer* 2007; **109**: 2490-2496 [PMID: 17487861 DOI: 10.1002/cncr.22701]
- 53 **Tanaka N**, Moriya K, Kiyosawa K, Koike K, Gonzalez FJ, Aoyama T. PPARalpha activation is essential for HCV core protein-induced hepatic steatosis and hepatocellular carcinoma in mice. *J Clin Invest* 2008; **118**: 683-694 [PMID: 18188449 DOI: 10.1172/jci33594]
- 54 **Moriishi K**, Mochizuki R, Moriya K, Miyamoto H, Mori Y, Abe T, Murata S, Tanaka K, Miyamura T, Suzuki T, Koike K, Matsuura Y. Critical role of PA28gamma in hepatitis C virus-associated steatogenesis and hepatocarcinogenesis. *Proc Natl Acad Sci USA* 2007; **104**: 1661-1666 [PMID: 17234812 DOI: 10.1073/pnas.0607312104]
- 55 **Okuda M**, Li K, Beard MR, Showalter LA, Scholle F, Lemon SM, Weinman SA. Mitochondrial injury, oxidative stress, and antioxidant gene expression are induced by hepatitis C virus core protein. *Gastroenterology* 2002; **122**: 366-375 [PMID: 11832451]
- 56 **Korenaga M**, Wang T, Li Y, Showalter LA, Chan T, Sun J, Weinman SA. Hepatitis C virus core protein inhibits mitochondrial electron transport and increases reactive oxygen species (ROS) production. *J Biol Chem* 2005; **280**: 37481-37488 [PMID: 16150732 DOI: 10.1074/jbc.M506412200]
- 57 **Barbaro G**, Di Lorenzo G, Asti A, Ribersani M, Belloni G, Grisorio B, Filice G, Barbarini G. Hepatocellular mitochondrial alterations in patients with chronic hepatitis C: ultrastructural and biochemical findings. *Am J Gastroenterol* 1999; **94**: 2198-2205 [PMID: 10445550 DOI: 10.1111/j.1572-0241.1999.01294.x]
- 58 **Horiike S**, Kawanishi S, Kaito M, Ma N, Tanaka H, Fujita N, Iwasa M, Kobayashi Y, Hiraku Y, Oikawa S, Murata M, Wang J, Semba R, Watanabe S, Adachi Y. Accumulation of 8-nitroguanine in the liver of patients with chronic hepatitis C. *J Hepatol* 2005; **43**: 403-410 [PMID: 16023246 DOI: 10.1016/j.jhep.2005.03.026]
- 59 **Chuma M**, Hige S, Nakanishi M, Ogawa K, Natsuzaka M, Yamamoto Y, Asaka M. 8-Hydroxy-2'-deoxy-guanosine is a risk factor for development of hepatocellular carcinoma in patients with chronic hepatitis C virus infection. *J Gastroenterol Hepatol* 2008; **23**: 1431-1436 [PMID: 18854000 DOI: 10.1111/j.1440-1746.2008.05502.x]
- 60 **Machida K**, Cheng KT, Lai CK, Jeng KS, Sung VM, Lai MM. Hepatitis C virus triggers mitochondrial permeability transition with production of reactive oxygen species, leading to DNA damage and STAT3 activation. *J Virol* 2006; **80**: 7199-7207 [PMID: 16809325 DOI: 10.1128/jvi.00321-06]
- 61 **Machida K**, Cheng KT, Sung VM, Lee KJ, Levine AM, Lai MM. Hepatitis C virus infection activates the immunologic (type II) isoform of nitric oxide synthase and thereby enhances DNA damage and mutations of cellular genes. *J Virol* 2004; **78**: 8835-8843 [PMID: 15280491 DOI: 10.1128/jvi.78.16.8835-8843.2004]
- 62 **Wang T**, Weinman SA. Causes and consequences of mitochondrial reactive oxygen species generation in hepatitis C. *J Gastroenterol Hepatol* 2006; **21** Suppl 3: S34-S37 [PMID: 16958669 DOI: 10.1111/j.1440-1746.2006.04591.x]
- 63 **Klopstock N**, Katzenellenbogen M, Pappo O, Sklair-Levy M, Olam D, Mizrahi L, Potikha T, Galun E, Goldenberg D. HCV tumor promoting effect is dependent on host genetic background. *PLoS One* 2009; **4**: e5025 [PMID: 19340302 DOI: 10.1371/journal.pone.0005025]
- 64 **Nishina S**, Hino K, Korenaga M, Vecchi C, Pietrangelo A, Mizukami Y, Furutani T, Sakai A, Okuda M, Hidaka I, Okita K, Sakaida I. Hepatitis C virus-induced reactive oxygen species raise hepatic iron level in mice by reducing hepcidin transcription. *Gastroenterology* 2008; **134**: 226-238 [PMID: 18166355 DOI: 10.1053/j.gastro.2007.10.011]
- 65 **Tanaka H**, Fujita N, Sugimoto R, Urawa N, Horiike S, Kobayashi Y, Iwasa M, Ma N, Kawanishi S, Watanabe S, Kaito M, Takei Y. Hepatic oxidative DNA damage is associated with increased risk for hepatocellular carcinoma in chronic hepatitis C. *Br J Cancer* 2008; **98**: 580-586 [PMID: 18231107 DOI: 10.1038/sj.bjc.6604204]
- 66 **Hino K**, Hara Y, Nishina S. Mitochondrial reactive oxygen species as a mystery voice in hepatitis C. *Hepatol Res* 2014; **44**: 123-132 [PMID: 24112394 DOI: 10.1111/hepr.12247]

**P- Reviewer:** He JY, Kim K, Zhang WZ **S- Editor:** Ji FF

**L- Editor:** A **E- Editor:** Liu SQ



## Retrospective Cohort Study

**Comparison of peg-interferon, ribavirin plus telaprevir vs simeprevir by propensity score matching**

Hideki Fujii, Takeshi Nishimura, Atsushi Umemura, Taichiro Nishikawa, Kanji Yamaguchi, Michihisa Moriguchi, Yoshio Sumida, Hironori Mitsuyoshi, Chihiro Yokomizo, Saiyu Tanaka, Hiroki Ishikawa, Kenichi Nishioji, Hiroyuki Kimura, Shiro Takami, Yasuyuki Nagao, Takayuki Takeuchi, Toshihide Shima, Yoshihiko Sawa, Masahito Minami, Kohichiroh Yasui, Yoshito Itoh

Hideki Fujii, Takeshi Nishimura, Atsushi Umemura, Taichiro Nishikawa, Kanji Yamaguchi, Michihisa Moriguchi, Yoshio Sumida, Hironori Mitsuyoshi, Masahito Minami, Kohichiroh Yasui, Yoshito Itoh, Department of Molecular Gastroenterology and Hepatology, Kyoto Prefectural University of Medicine, Kyoto 602-8566, Japan

Hideki Fujii, Hiroyuki Kimura, Department of Gastroenterology, Japanese Red Cross Kyoto Daiichi Hospital, Kyoto 605-0981, Japan

Takeshi Nishimura, Department of Gastroenterology and Hepatology, North Medical Center, Kyoto Prefectural University of Medicine, Kyoto 629-2261, Japan

Chihiro Yokomizo, Department of Gastroenterology, Osaka General Hospital of West Japan Railway Company, Osaka 545-0053, Japan

Saiyu Tanaka, Center for Digestive and Liver Diseases, Nara City Hospital, Nara 630-8305, Japan

Hiroki Ishikawa, Department of Gastroenterology and Hepatology, Omihachiman Community Medical Center, Shiga 523-0082, Japan

Kenichi Nishioji, Health Care Division, Kyoto Second Red Cross Hospital, Kyoto 602-8026, Japan

Shiro Takami, Department of Gastroenterology, Otsu Municipal Hospital, Otsu 520-0804, Japan

Yasuyuki Nagao, Department of Gastroenterology, Matsushita Memorial Hospital, Moriguchi 570-8540, Japan

Takayuki Takeuchi, Higashiomi City Notogawa Hospital, Higashiomi 521-1223, Japan

Toshihide Shima, Center of Gastroenterology and Hepatology, Saiseikai Suita Hospital, Suita 564-0013, Japan

Yoshihiko Sawa, Masahito Minami, Department of Internal Medicine, Aiseikai Yamashina Hospital, Kyoto 607-8086, Japan

**Author contributions:** Fujii H, Nishimura T and Itoh Y designed the research; Umemura A, Nishikawa T, Yamaguchi K, Moriguchi M, Sumida Y, Mitsuyoshi H, Yokomizo C, Tanaka S, Ishikawa H, Nishioji K, Kimura H, Takami S, Nagao Y, Takeuchi T, Shima T, Sawa Y, Minami M and Yasui K performed the research; Fujii H analyzed the data; Fujii H and Itoh Y wrote the paper.

**Institutional review board statement:** The study was reviewed and approved by the Kyoto Prefectural University of Medicine Institutional Review Board.

**Informed consent statement:** All study participants provided informed written consent prior to study enrollment.

**Conflict-of-interest statement:** Yoshito Itoh receives honoraria from Bristol-Myers K.K., MSD K.K.. Yoshito Itoh receives research grant from MSD K.K., Chugai Phram, Bristol-Myers K.K., Gilead Sciences, Inc., Mitsubishi Tanabe Pharma Corporation, Dainippon Sumitomo Pharma Co., Ltd.

**Data sharing statement:** No additional data are available. Responses to the request for the raw data will be judged by the IRB.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to:** Hideki Fujii, MD, PhD, Department of Molecular Gastroenterology and Hepatology, Kyoto Prefectural

University of Medicine, 465 Kajii-cho, Hirokoji Agaru, Kawaramachi-dori, Kamigyo-ku, Kyoto 602-8566, Japan. hidekifujii710810@yahoo.co.jp  
Telephone: +81-75-2515519  
Fax: +81-75-2510710

Received: August 16, 2015  
Peer-review started: August 18, 2015  
First decision: October 14, 2015  
Revised: October 22, 2015  
Accepted: November 10, 2015  
Article in press: November 11, 2015  
Published online: December 8, 2015

## Abstract

**AIM:** To compare efficacy of telaprevir (TVR) and simeprevir (SMV) combined with pegylated interferon (PEG-IFN) and ribavirin (RBV) while treating chronic hepatitis C (CHC).

**METHODS:** In all, 306 CHC patients were included in this study. There were 159 patients in the TVR combination therapy group and 147 patients in the SMV combination therapy group. To evaluate pretreatment factors contributing to sustained virological response at 12 wk (SVR12), univariate and multivariate analyses were performed in TVR and SMV groups. To adjust for patient background between TVR and SMV groups, propensity score matching was performed. Virological response during treatment and SVR12 were evaluated.

**RESULTS:** Overall rates of SVR12 [undetectable serum hepatitis C virus (HCV) RNA levels] were 79.2% and 69.4% in TVR and SMV groups, respectively. Patients in the SMV group were older, had higher serum HCV RNA levels, lower hemoglobin, higher prevalence of unfavorable interleukin-28B (*IL28B*) genotype (rs8099917), and poorer response to previous PEG-IFN and RBV treatment. Propensity score matching was performed to adjust for backgrounds ( $n = 104$ ) and demonstrated SVR12 rates of 74.0% and 73.1% in the TVR and SMV groups, respectively. In the TVR group, discontinuation rates were higher because of adverse events; however, breakthrough and nonresponse was more frequent in the SMV group. Multivariate analysis revealed *IL28B* genotype (rs8099917) as the only independent predictive factor of SVR12 in both groups.

**CONCLUSION:** SVR12 rates were almost identical following propensity score matching.

**Key words:** Chronic hepatitis C; Combination therapy; Pegylated interferon; Simeprevir; Telaprevir; Propensity score matching; Protease inhibitor

© The Author(s) 2015. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** We evaluated and compared the efficacy of telaprevir (TVR) and simeprevir (SMV) in combination

with pegylated interferon and ribavirin in the treatment of chronic hepatitis C. patients in real-world clinical settings. In the TVR group, the proportion of patients achieving a virological response was higher than that in the SMV group according to the original data. After propensity score matching, the proportion of patients achieving a virological response during treatment and after 12 wk was almost identical between the two groups with no significant difference observed.

Fujii H, Nishimura T, Umemura A, Nishikawa T, Yamaguchi K, Moriguchi M, Sumida Y, Mitsuyoshi H, Yokomizo C, Tanaka S, Ishikawa H, Nishioji K, Kimura H, Takami S, Nagao Y, Takeuchi T, Shima T, Sawa Y, Minami M, Yasui K, Itoh Y. Comparison of peg-interferon, ribavirin plus telaprevir vs simeprevir by propensity score matching. *World J Hepatol* 2015; 7(28): 2841-2848 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i28/2841.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i28.2841>

## INTRODUCTION

Chronic hepatitis C (CHC) infection is associated with a greatly increased risk of liver cirrhosis and hepatocellular carcinoma. There are an estimated 130-170 million people infected with hepatitis C virus (HCV) worldwide<sup>[1]</sup> and approximately 1.5-2 million in Japan<sup>[2]</sup>. The combination of pegylated interferon (PEG-IFN) and ribavirin (RBV) dual therapy was previously the standard care for CHC, and was administered for 48-72 wk in patients with genotype 1 and for 24 wk in genotype 2. Sustained virological response (SVR) rates are approximately 40%-50% in former group treated for 48 wk and approximately 80% in the latter treated for 24 wk<sup>[3-5]</sup>.

Novel drug classes, including inhibitors of the NS3/NS4 protease of HCV polyprotein (protease inhibitors), have recently become available<sup>[6-8]</sup>. Of these, telaprevir (TVR) was the first to be approved in Japan for the treatment of CHC. In a clinical trial of TVR triple combination therapy (TVR, PEG-IFN, and RBV) for 24 wk in Japan, rapid reductions in serum HCV RNA levels were observed with a SVR rate of approximately 70%<sup>[9,10]</sup>. However, treatment discontinuation because of adverse events, including skin rash, anemia, and thrombocytopenia, occurred in up to 21% patients<sup>[11]</sup>. Thus, the TVR triple combination therapy is no longer recommended<sup>[12]</sup>.

Simeprevir (SMV) is a second generation NS3/NS4 protease inhibitor<sup>[13]</sup>. The QUEST 1 and QUEST 2 phase 3 clinical trials demonstrated SVRs of 80% and 81% in patients treated with SMV triple combination therapy (SMV, PEG-IFN, and RBV), respectively. Similar results have been reported in phase 3 clinical trials conducted in Japan<sup>[14-16]</sup>. TVR and SMV were approved for use in clinical practice in Japan in December 2011 and December 2013, respectively. We previously treated patients with CHC using TVR or SMV as PEG-IFN and

RBV-based triple combination therapy with an NS3/NS4 protease inhibitor; however, “drug lag” between TVR and SMV, causing a difference in clinical backgrounds between the two regimens prior to treatment initiation, prevented fair comparison of the efficacy of TVR and SMV in real-world clinical practice. The aim of this study was to evaluate and compare the efficacy of TVR or SMV for the treatment of CHC patients in Japan.

## MATERIALS AND METHODS

### Patients

Patients were enrolled at Kyoto Prefectural University of Medicine and 8 affiliated hospitals in Kinki area of Japan (Kyoto, Osaka, Nara, Shiga Prefecture) from 2012 to 2014. Study protocols were approved by the ethics committee of each institution and conformed to the provisions of the Declaration of Helsinki. Patients enrolled in this study were diagnosed with CHC by board-certified hepatologists. Eligible patients were 20–80 years of age and had chronic HCV genotype 1 infection with HCV RNA levels of 5.0 log<sub>10</sub> IU/mL or higher at screening.

Patients with decompensated liver disease, chronic hepatitis B, co-infection with human immunodeficiency virus, autoimmune hepatitis, primary biliary cirrhosis, hemochromatosis, or Wilson’s disease were excluded. Patients with uncontrollable hypertension or diabetes mellitus, and those with a history of alcohol abuse, were also excluded. Patients were followed-up monthly for the assessment of liver function and virological markers during treatment and until 12 wk after the completion of triple therapy. All patients gave informed consent to participate in this study.

In the TVR group, three patients were lost to follow-up and extreme protocol deviation (*e.g.*, extended PEG-IFN and RBV therapy for up to 48 wk) occurred in seven patients. In the SMV group, two patients were lost to follow-up and extreme protocol deviation (*e.g.*, extended PEG-IFN and RBV therapy for up to 48 wk) occurred in nine patients. Patients lost to follow-up or with extreme protocol deviation were excluded from the analysis.

The therapeutic outcomes of previous PEG-IFN and RBV therapy were classified into the following two groups: Undetectable serum HCV RNA levels at the end of the treatment period with quantifiable HCV RNA levels during follow-up (relapse group); and detectable HCV RNA levels at the end of the treatment period (other group).

### Study design

Patients received per os telaprevir (Telavic; Mitsubishi Tanabe Pharma, Osaka, Japan) 2250 mg/d or simeprevir (Sovriad; Janssen Pharmaceutical K.K., Tokyo, Japan) 100 mg/d, combined with weekly subcutaneous injections of PEG-IFN alpha 2b (Peg-Intron; MSD, Tokyo, Japan) of 1.5 µg/kg and per os administration of RBV (Rebetol; MSD, Tokyo, Japan) of 600–1000 mg/d in accordance with prescribing information for 12 wk followed by PEG-IFN alpha 2b and RBV between weeks

12 and 24. In the TVR group, patients with lower serum hemoglobin levels began therapy at a reduced dose of TVR 1500 mg/d according to the judgment of treating physicians (2250 mg/d, 66 patients; 1500 mg/d, 93 patients). In the SMV group, patients began therapy at a dose of 100 mg/d. Dose reductions or discontinuation of TVR, SMV, PEG-IFN, and RBV were according to the judgment of treating physicians. Patients were followed-up for at least 12 wk after final treatment administration to assess SVR.

HCV RNA responses during therapy were classified into the following groups: Detectable HCV RNA levels at the end of the treatment period (nonresponse group); reappearance of HCV RNA during treatment (break-through group); and undetectable serum HCV RNA levels at the end of the treatment period with quantifiable HCV RNA levels during follow-up (relapse group). SVR<sub>12</sub> was defined as undetectable serum HCV RNA levels at 12 wk after the end of treatment. Therapeutic effects were evaluated using intention-to-treat analysis.

### Laboratory assessments

Blood samples were obtained for routine biochemical and hematological assessments at treatment initiation, on treatment weeks 2, 4, 8, 12, 16, 20, 24, at the end of treatment (EOT), and at 12 wk after EOT. The antiviral effects were assessed by measuring serum HCV RNA levels using the COBAS TaqMan HCV test (Roche Molecular Diagnostics, Tokyo, Japan) with a lower limit of quantitation of 15 IU/mL. Interleukin 28B (*IL28B*; rs8099917) genotyping was accordingly performed in the majority of patients. In brief, DNA was extracted from peripheral whole blood (100 µL) with DNeasy Blood and Tissue Kits (QIAGEN, Valencia, CA) according to the manufacturer’s instructions. Genotypes were determined using a Light Cycler (Roche, Osaka, Japan). Subsequent gene sequencing was performed to validate amplified polymerase chain reaction (PCR) products. Primers and probes used for PCR were as follows: Forward primer, 5'-CAACATGGAGAGTTAAAGTAAGTCTTG-3'; reverse primer, 5'-TGCTGGGCCCTAACTGAT-3'; probe 1, LC Red 640-TTGGGTGACATTGCTCACAGAAAGG-Phosphate; and probe 2, CCAGCTACCAAAGTGTATACAGCATGGTTCCA-Fluorescein.

### Statistical analysis

Baseline continuous data were expressed as median with interquartile ranges in parentheses, and categorical variables were expressed as numbers. Univariate analyses were performed using chi-squared or Mann-Whitney *U*-tests as appropriate. All *P*-values of < 0.05 of two-tailed tests were considered significant. Multivariate logistic regression was used to identify significant independent predictive factors of SVR<sub>12</sub>. Results were expressed as Odds ratios and 95%CI. All statistical analyses were performed using the SPSS 22.0 statistical package (SPSS Incorporated, Chicago, Illinois, United States).

To adjust for patient background between TVR and

**Table 1** Baseline characteristics of patients who received triple therapy with pegylated interferon, ribavirin, and telaprevir or simeprevir

	Unmatched patients		<i>P</i> value	Standardized difference	Propensity score matched patients		<i>P</i> value	Standardized difference
	Telaprevir	Simeprevir			Telaprevir	Simeprevir		
No. of patients	<i>n</i> = 159	<i>n</i> = 147			<i>n</i> = 104	<i>n</i> = 104		
Age (yr)	60 (51.0-65.0)	63 (54.5-70.0)	0.002	0.348	61.5 (53.0-65.8)	60.5 (52.0-67.0)	NS	0.0154
Gender (male/female)	77/82	67/80	NS	0.057	45/59	49/55	NS	0.0773
Body mass index (kg/m <sup>2</sup> )	23.9 (21.7-25.7)	23.2 (21.1-25.0)	NS	0.202	23.6 (21.1-25.3)	23.4 (21.2-25.2)	NS	0.0747
Laboratory data								
Level of viremia (log IU/mL)	6.7 (6.3-7.0)	6.8 (6.3-7.2)	NS	0.210	6.7 (6.3-7.0)	6.6 (6.2-7.1)	NS	0.0158
Leukocyte count (/mm <sup>3</sup> )	5060 (4200-5800)	4920 (4100-5800)	NS	0.094	5000 (4200-5700)	5020 (4150-5800)	NS	0.0272
Hemoglobin (g/dL)	14.1 (13.1-15.0)	13.8 (12.9-14.7)	NS	0.175	14 (13.0-14.8)	13.9 (12.9-15.0)	NS	0.0264
Platelet count (× 10 <sup>4</sup> /mm <sup>3</sup> )	15 (12.5-19.8)	15.1 (11.7-20.1)	NS	0.053	15 (12.9-20.0)	15.1 (11.8-20.1)	NS	0.0032
SNP of <i>IL28B</i> (TT/non-TT/unknown)	99/34/26	89/43/15	NS	0.155	74/30	73/31	NS	0.0211
Other data								
Prior treatment response relapse/other	43/30	31/32	NS	0.196	31/22	23/20	NS	0.100

“Unmatched patients” refer to values prior to propensity score matching and “Propensity score matched patients” refer to values after adjustment by propensity score matching. Data are presented as numbers or medians with interquartile ranges in parentheses. *P*-values were calculated using the  $\chi^2$  or Mann-Whitney *U*-test for continuous variables. SNP: Single-nucleotide polymorphism; *IL28B*: Interleukin 28B; NS: Not significant.

SMV groups, propensity score matching was performed. Propensity score models were estimated using a logistic regression model that adjusts for patient characteristics (age, gender, body mass index, HCV RNA level, leukocyte count, hemoglobin, platelet count, and *IL28B* SNPs) listed in Table 1. Confounders were selected according to their potential association with the outcome on the basis of clinical knowledge and previous studies<sup>[17]</sup>. The propensity score matching model was validated by the Hosmer and Lemeshow goodness-of-fit test (*P* = 0.638) and by the value of the area under the curve (0.66, 95%CI: 0.594-0.724). One SMV patient was matched to one TVR patient using nearest neighbor matching without replacement. Propensity scores were matched using a caliper width 0.25 logit of the SD. The standardized difference was used to assess the covariate balance. McNemarr's tests were performed after matching.

## RESULTS

### Baseline characteristics

The baseline patient characteristics in the TVR group (*n* = 159) and SMV group (*n* = 147) are shown in Table 1 as “unmatched patients”. Patients in the SMV group were significantly older than patients in the TVR group. High viral load, low hemoglobin levels, the non-TT *IL28B* genotype, and relapse following previous PEG-IFN and RBV treatment were more commonly observed in the SMV group compared with the TVR group.

### Virological response to therapy and loss of HCV RNA during treatment

In the TVR group, the overall SVR12 was 79.2% (126 of 159 patients). Undetectable HCV RNA levels were achieved during treatment in 33.3% (41 of 123), 80.8% (118 of 146), 92.4% (146 of 158), and 91.2% (145 of 159) of patients at 2, 4, 8 wk, and EOT or 24 wk, respectively. In the SMV group, the overall SVR12 rate was 69.4% (102 of 147 patients). Undetectable HCV

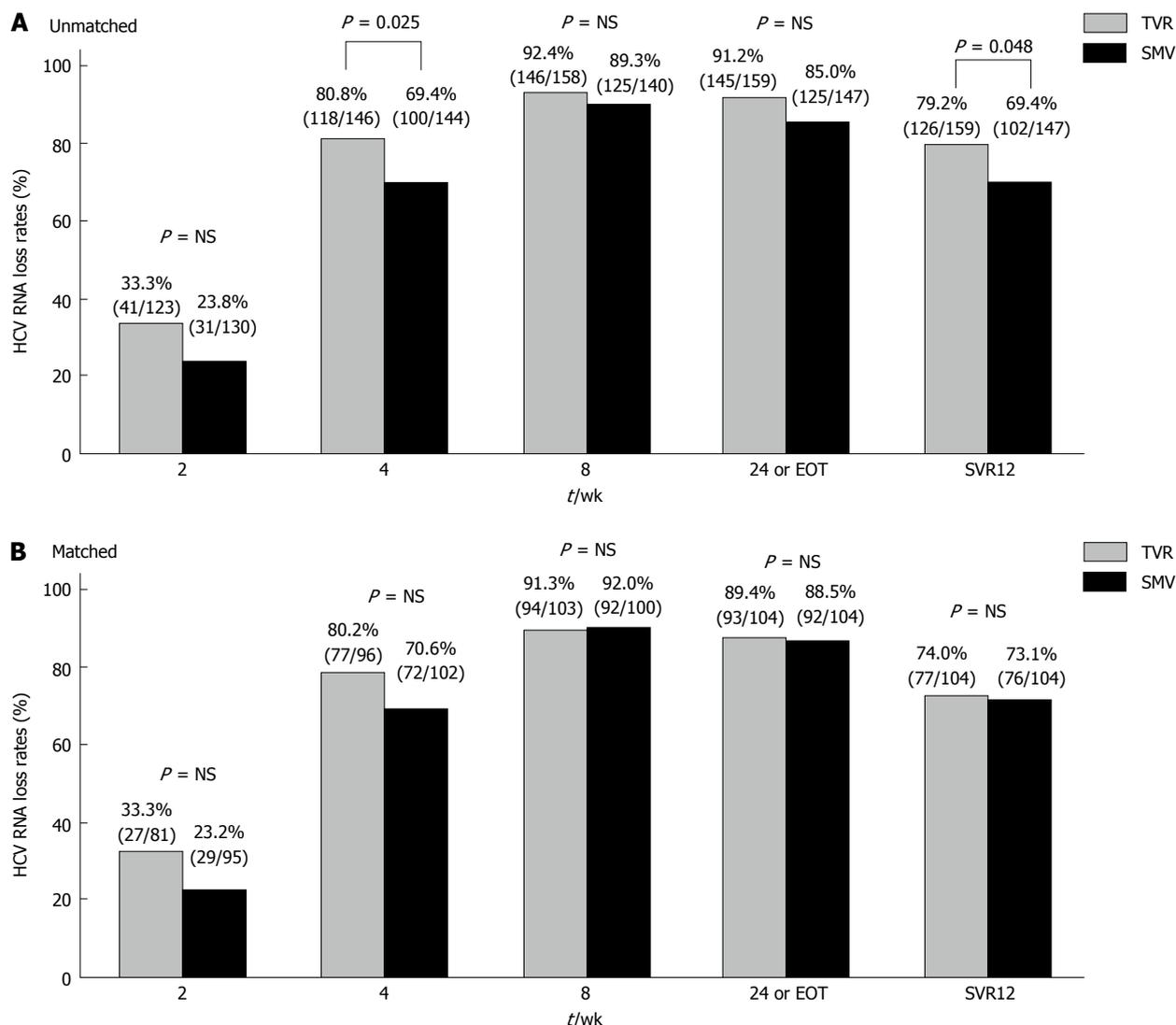
RNA levels were achieved during treatment in 23.8% (31 of 130), 69.4% (100 of 144), 89.3% (125 of 140), and 85.0% (125 of 147) of patients at 2, 4, 8 and EOT or 24 wk, respectively (Figure 1A).

### Safety and tolerability

In the TVR group, 10 patients demonstrated non-response, and breakthrough occurred in 4 patients. Relapse occurred in 19 patients. In patients with nonresponse, 8 patients discontinued TVR because of adverse events within the first 4 wk of treatment (four skin rash, one renal dysfunction, two appetite loss, one unknown). In the SMV group, 15 patients demonstrated non-response, and breakthrough occurred in eight patients. Relapse occurred in 22 patients. In patients with non-response, one patient discontinued within the first 4 wk of treatment (transient visual field defect). There was a trend toward greater rates of treatment discontinuation because of adverse events in the TVR group and nonresponse and breakthrough in the SMV group.

### Pretreatment factors contributing to SVR12 in TVR and SMV groups

To evaluate pretreatment factors contributing to SVR12, univariate and multivariate analyses were performed in TVR and SMV groups including the following variables: Age, gender, body mass index, *IL28B* (rs8099917) genotype, viral load, leukocyte count, hemoglobin, and platelet counts (Table 2). In the TVR group, *IL28B* genotypes significantly correlated with SVR12 according to univariate analysis. In multivariable logistic regression analysis, *IL28B* genotype was found to be a significant independent predictor of SVR12 (OR = 4.316; 95%CI: 1.804-10.327, *P* = 0.001). In the SMV group, age and *IL28B* genotype significantly correlated with SVR12 according to univariate analysis. In multivariable logistic regression analysis, significant independent predictors of SVR were *IL28B* genotype (OR = 8.598; 95%CI: 3.388-21.817; *P* < 0.001), age (OR = 0.933; 95%CI:



**Figure 1** Rates of virological response to telaprevir and simeprevir according to serum hepatitis C virus RNA levels before and after adjustment by propensity score matching. Percentages represent the proportion of patients with undetectable serum hepatitis C virus (HCV) RNA levels. Patient numbers are shown in parentheses. *P*-values were calculated using the  $\chi^2$  test prior to matching and McNemarr's test after matching. A: Before adjustment. Rates of virological response at 4 and 12 wk after treatment were significantly different between the telaprevir (TVR) group and simeprevir (SMV) group; B: After adjustment. No significant difference in the virological response was observed between the two groups. NS: Not significant.

0.889–0.980;  $P = 0.006$ ), and viral load (OR = 0.335; 95%CI: 0.157–0.715,  $P = 0.005$ ). Propensity score matching analysis was subsequently performed to reduce bias caused by differing baseline patient characteristics between TVR and SMV groups (Tables 1 and 2, propensity score matched patients). Following one-to-one matching of the two groups according to propensity score, 104 patients from the TVR group and 104 patients from the SMV group were matched according to baseline characteristics (Tables 1 and 2). Majority of covariates were statistically similar between the two groups (Table 1, Propensity score matched patients). Multivariable logistic regression analysis demonstrated that *IL28B* genotype significantly associated with SVR12 in both groups (TVR: OR = 7.739; 95%CI: 2.111–28.375,  $P = 0.002$ ; and SMV: OR = 8.594; 95%CI: 2.777–26.598,  $P < 0.001$ ; Table 2).

### Virological response during treatment and SVR12 after propensity score matching

Before adjustment, the proportion of patients achieving virological responses at 4 wk and after 12 wk treatment significantly differed between the TVR group and SMV group. In general, a greater proportion of patients in the TVR group had a virological response than that in the SMV group (Figure 1A). After one-to-one propensity score matching, the proportions of patients achieving a virological response during treatment and after 12 wk treatment were similar between the two groups (SVR12: TVR, 74.0%; SMV, 73.1%; Figure 1B).

## DISCUSSION

In the present study, we evaluated and compared the efficacy of TVR and SMV in combination with PEG-IFN

**Table 2** Univariate and multivariate analysis of factors associated with sustained virological response following pegylated interferon, ribavirin, and telaprevir or simeprevir triple therapy

Parameters	Telaprevir			Simeprevir		
	SVR	Non-SVR	P value	SVR	Non-SVR	P value
Unmatched patients						
Univariate analysis	<i>n</i> = 126	<i>n</i> = 33		<i>n</i> = 102	<i>n</i> = 45	
Age (yr)	58.0 (51.0-65.0)	62.0 (59.6-65.5)	NS	61.0 (52.8-67.3)	66.0 (56.5-71.0)	0.016
Gender (male/female)	62/64	15/18	NS	48/54	19/26	NS
Body mass index (kg/m <sup>2</sup> )	23.8 (21.7-25.7)	24.6 (21.7-26.0)	NS	23.2 (21.0-25.1)	23.5 (21.3-26.0)	NS
Level of viremia (log IU/mL)	6.7 (6.3-7.0)	6.6 (6.3-7.0)	NS	6.8 (6.2-7.1)	6.9 (6.4-7.3)	NS
Leukocyte count (/mm <sup>3</sup> )	5100 (4200-5700)	5100 (4400-6600)	NS	5000 (4300-5800)	4800 (3800-5800)	NS
Hemoglobin (g/dL)	14.1 (13.2-15.1)	14.1 (12.8-14.8)	NS	13.9 (13.1-14.7)	13.7 (12.7-14.8)	NS
Platelet count (× 10 <sup>4</sup> /mm <sup>3</sup> )	15.0 (12.8-19.8)	15.0 (12.5-20.1)	NS	15.3 (11.9-20.5)	15.0 (11.1-18.3)	NS
SNP of IL28B (TT/non-TT)	84/19	15/15	< 0.001	72/19	17/24	< 0.001
Multivariate analysis	Odds ratio (95%CI)			Odds ratio (95%CI)		
SNP of IL28B (TT/non-TT)	4.316 (1.804-10.327)		0.001	8.598 (3.388-21.817)		< 0.001
Age (yr)				0.933 (0.889-0.980)		0.006
Level of viremia (log IU/mL)				0.335 (0.157-0.715)		0.005
Propensity score matched patients						
Univariate analysis	<i>n</i> = 77	<i>n</i> = 27		<i>n</i> = 76	<i>n</i> = 28	
Age (yr)	60.0 (52.5-66.5)	64.0 (60.0-65.0)	NS	59.0 (51.0-66.0)	65.0 (56.0-71.0)	0.021
Gender (male/female)	33/44	12/15	NS	37/39	12/16	NS
Body mass index (kg/m <sup>2</sup> )	23.1 (20.6-25.0)	24.0 (21.4-26.6)	NS	23.2 (21.3-25.1)	23.6 (21.1-26.1)	NS
Level of viremia (log IU/mL)	6.7 (6.3-7.0)	6.7 (6.4-6.9)	NS	6.7 (6.1-7.0)	6.6 (6.3-7.2)	NS
Leukocyte count (/mm <sup>3</sup> )	5000 (4100-5700)	5100 (4400-6700)	NS	5100 (4400-5900)	4600 (3500-5700)	NS
Hemoglobin (g/dL)	14.0 (13.1-14.8)	13.9 (13.0-14.9)	NS	14.0 (12.9-15.0)	13.8 (12.9-14.7)	NS
Platelet count (× 10 <sup>4</sup> /mm <sup>3</sup> )	15.0 (12.3-19.7)	15.4 (12.9-20.3)	NS	15.1 (11.9-20.4)	15.0 (11.0-17.2)	NS
SNP of IL28B (TT/non-TT)	61/16	13/14	0.002	61/15	12/16	< 0.001
Multivariate analysis	Odds ratio (95%CI)			Odds ratio (95%CI)		
SNP of IL28B (TT/non-TT)	7.739 (2.111-28.375)		0.002	8.594 (2.777-26.598)		< 0.001

Values are presented as numbers or medians with interquartile ranges in parentheses. *P*-values were calculated using the  $\chi^2$  test or Mann-Whitney *U*-test for continuous variables. SNP: Single-nucleotide polymorphism; IL28: Interleukin 28B; SVR: Sustained virological response; NS: Not significant.

and RBV in the treatment of CHC patients in real-world clinical settings in Japan. Both regimens achieved higher SVR rates compared with that using the dual combination therapy with PEG-IFN and RBV<sup>[6-10,14-16]</sup>. In the TVR group, the proportion of patients achieving a virological response was higher than in the SMV group according to the original data. A number of patients discontinued TVR therapy because of adverse events at the beginning of treatment. After propensity score matching, the proportion of patients achieving a virological response during treatment and after 12 wk was almost identical between the two groups with no significantly difference observed (Figure 1B).

Patients in the SMV group appeared to have a greater prevalence of unfavorable baseline characteristics. Patients in SMV group were statistically older, had higher viral loads, lower hemoglobin levels, and a higher prevalence of unfavorable *IL28* genotypes (rs809997) compared with that in the TVR group. These pretreatment factors are known to influence the efficacy of IFN-based therapies<sup>[17]</sup>. As previously reported, Japanese patients infected with HCV genotype 1b are substantially older than Western patients<sup>[18]</sup>. A large proportion of patients able to tolerate IFN-based therapies were cured with previous therapies. Patients with unfavorable baseline characteristics remain untreated. In addition, according to academic guidelines<sup>[19]</sup>, TVR therapy should be avoided in older patients with low hemoglobin levels in anticipation of future

therapeutic options. As a result, a greater prevalence of unfavorable baseline characteristics were observed in patients in the SMV group.

In the present study, a greater proportion of patients in the TVR group discontinued treatment because of adverse events. Previously reported adverse events associated with TVR treatment include anemia, skin rash, and severe fatigue<sup>[11]</sup>. Cutaneous adverse effects caused by TVR have been frequently reported and are rare but are characterized by rapid development of lethal severe skin complications, such as Stevens-Johnson syndrome and drug-induced hypersensitivity syndrome<sup>[20,21]</sup>. Patients with these skin complications may have stopped the TVR treatment earlier. We administered an initial dose of TVR 1500 mg/d in majority of patients to prevent treatment-induced anemia<sup>[22]</sup>. In contrast, the incidence of severe adverse events was low in the SMV group. Therefore, a smaller number of patients discontinued therapy in the SMV group.

Viral dynamics during treatment were similar to previous reports in both groups<sup>[16,23]</sup>. However, breakthrough and nonresponse was more frequent in the SMV group. Before matching, the TVR group had a higher SVR12 rate than that of the SMV group. After propensity score matching, this difference diminished and SVR12 rates were similar between the two groups. Reddy *et al.*<sup>[24]</sup> reported a randomized control study between SMV and TVR for previous null or partial responders. Although the differences were observed in dosage, race, approved

combined interferon, and treatment duration in their report, viral breakthrough was more frequent with SMV therapy than with TVR therapy similar to the present report.

The SVR rate in the SMV group in the present study was lower than in the CONCERTO-4 study<sup>[16]</sup>. As our study was in a real-world clinical setting, patients were generally older (proportion of patients aged > 65 years, 42.3% vs CONCERTO-4, 22.8%) and had lower platelet counts (platelet counts < 15 × 10<sup>4</sup>/mm<sup>3</sup>: 47.7% vs CONCERTO-4, 31.6%) in our study. Baseline patient characteristics in our study may have resulted in a lower SVR12 rate.

The major limitation of the present study was the inability to evaluate several factors known to influence treatment efficacy. We did not examine amino acid substitutions of the HCV core region 70 and 91<sup>[23]</sup>, NS5A interferon sensitivity determining region<sup>[25]</sup>, interferon/ribavirin resistance determining region<sup>[26]</sup>, or resistance-associated mutations of HCV NS3/NS4 proteases<sup>[27-29]</sup>.

Treatment approaches to CHC are rapidly changing worldwide<sup>[30,31]</sup>. At present, direct-acting antiviral agent (DAA) combination therapy (daclatasvir and asunaprevir) is available for patients with HCV genotype 1 in Japan. Interferon-free DAA combination therapy has demonstrated an overall SVR12 rate of 85%<sup>[32]</sup>. Although the majority of patients with HCV infection may be treated with DAAs combination regimens, PEG-IFN and RBV-based treatment may still have utility in a small number of patients that do not respond to DAAs therapies.

In conclusion, both TVR and SMV regimens achieved high SVR12 rates. In the original analysis, TVR appeared to demonstrate an increased anti-viral efficacy compared with that of SMV. After propensity score matching, the proportion of patients achieving a virological response during treatment and after 12 wk treatment was almost identical between the two groups. Treatment discontinuation was more frequent in the TVR group because of adverse events at the beginning of treatment; however, nonresponse and breakthrough were more frequently observed in the SMV group.

## COMMENTS

### Background

The combination of pegylated interferon (PEG-IFN) and ribavirin (RBV) dual therapy was previously the standard care for chronic hepatitis C (CHC). Novel drug classes, including inhibitors of the NS3/NS4 protease of hepatitis C virus (HCV) polyprotein (protease inhibitors), have recently become available. Of these, telaprevir (TVR) triple combination therapy (TVR, PEG-IFN, and RBV) was the first to be approved in Japan in December 2011. Simeprevir (SMV), second generation protease inhibitor, was approved in Japan in December 2013. "Drug lag" between TVR and SMV, causing a difference in clinical backgrounds between the two regimens prior to treatment initiation, prevented fair comparison of the efficacy of TVR and SMV in real-world clinical practice.

### Research frontiers

The authors' group evaluated and compared the efficacy of TVR or SMV for the treatment of CHC patients in Japan with propensity score matching to adjust for patient background between two groups.

## Innovations and breakthroughs

Before adjustment, the proportion of patients achieving virological responses significantly differed between the TVR group and SMV group. In general, a greater proportion of patients in the TVR group had a virological response than that in the SMV group. After one-to-one propensity score matching, the proportions of patients achieving a virological response during treatment and after 12 wk treatment were similar between the two groups.

## Applications

In the TVR group, the proportion of patients achieving a virological response was higher than in the SMV group according to the original data. A number of patients discontinued TVR therapy because of adverse events at the beginning of treatment. Breakthrough and nonresponse was more frequent in the SMV group. After propensity score matching, this difference diminished and sustained virological response 12 rates were similar between the two groups.

## Terminology

TVR is the first inhibitor of the NS3/NS4 protease of HCV polyprotein (protease inhibitors) in Japan. SMV is a second generation NS3/NS4 protease inhibitor. Propensity score matching attempt is used to reduce the background difference between TVR and SMV groups.

## Peer-review

The article is well written. It's clear and can help the authors' to understand the new drug treatment efficiency in a big cohort of HCV patients.

## REFERENCES

- 1 **Mohd Hanafiah K**, Groeger J, Flaxman AD, Wiersma ST. Global epidemiology of hepatitis C virus infection: new estimates of age-specific antibody to HCV seroprevalence. *Hepatology* 2013; **57**: 1333-1342 [PMID: 23172780 DOI: 10.1002/hep.26141]
- 2 **Namiki I**, Nishiguchi S, Hino K, Suzuki F, Kumada H, Itoh Y, Asahina Y, Tamori A, Hiramatsu N, Hayashi N, Kudo M. Management of hepatitis C; Report of the Consensus Meeting at the 45th Annual Meeting of the Japan Society of Hepatology (2009). *Hepatol Res* 2010; **40**: 347-368 [PMID: 20394674]
- 3 **Fried MW**, Shiffman ML, Reddy KR, Smith C, Marinos G, Gonçales FL, Häussinger D, Diago M, Carosi G, Dhumeaux D, Craxi A, Lin A, Hoffman J, Yu J. Peginterferon alfa-2a plus ribavirin for chronic hepatitis C virus infection. *N Engl J Med* 2002; **347**: 975-982 [PMID: 12324553 DOI: 10.1056/NEJMoa02047347/13/975]
- 4 **Manns MP**, McHutchison JG, Gordon SC, Rustgi VK, Shiffman M, Reindollar R, Goodman ZD, Koury K, Ling M, Albrecht JK. Peginterferon alfa-2b plus ribavirin compared with interferon alfa-2b plus ribavirin for initial treatment of chronic hepatitis C: a randomised trial. *Lancet* 2001; **358**: 958-965 [PMID: 11583749]
- 5 **Manns MP**, Wedemeyer H, Cornberg M. Treating viral hepatitis C: efficacy, side effects, and complications. *Gut* 2006; **55**: 1350-1359 [PMID: 16905701]
- 6 **Jacobson IM**, McHutchison JG, Dusheiko G, Di Bisceglie AM, Reddy KR, Bzowej NH, Marcellin P, Muir AJ, Ferenci P, Flisiak R, George J, Rizzetto M, Shouval D, Sola R, Terg RA, Yoshida EM, Adda N, Bengtsson L, Sankoh AJ, Kieffer TL, George S, Kauffman RS, Zeuzem S. Telaprevir for previously untreated chronic hepatitis C virus infection. *N Engl J Med* 2011; **364**: 2405-2416 [PMID: 21696307 DOI: 10.1056/NEJMoa1012912]
- 7 **Poordad F**, McCone J, Bacon BR, Bruno S, Manns MP, Sulkowski MS, Jacobson IM, Reddy KR, Goodman ZD, Boparai N, DiNubile MJ, Sniukiene V, Brass CA, Albrecht JK, Bronowicki JP. Boceprevir for untreated chronic HCV genotype 1 infection. *N Engl J Med* 2011; **364**: 1195-1206 [PMID: 21449783 DOI: 10.1056/NEJMoa1010494]
- 8 **Zeuzem S**, Andreone P, Pol S, Lawitz E, Diago M, Roberts S, Focaccia R, Younossi Z, Foster GR, Horban A, Ferenci P, Nevens F, Müllhaupt B, Pockros P, Terg R, Shouval D, van Hoek B, Weiland O, Van Heeswijk R, De Meyer S, Luo D, Boogaerts G, Polo R, Picchio G, Beumont M. Telaprevir for retreatment of HCV

- infection. *N Engl J Med* 2011; **364**: 2417-2428 [PMID: 21696308 DOI: 10.1056/NEJMoa1013086]
- 9 **Akuta N**, Suzuki F, Hirakawa M, Kawamura Y, Yatsuji H, Sezaki H, Suzuki Y, Hosaka T, Kobayashi M, Kobayashi M, Saitoh S, Arase Y, Ikeda K, Chayama K, Nakamura Y, Kumada H. Amino acid substitution in hepatitis C virus core region and genetic variation near the interleukin 28B gene predict viral response to telaprevir with peginterferon and ribavirin. *Hepatology* 2010; **52**: 421-429 [PMID: 20648473 DOI: 10.1002/hep.23690]
  - 10 **Kumada H**, Toyota J, Okanoue T, Chayama K, Tsubouchi H, Hayashi N. Telaprevir with peginterferon and ribavirin for treatment-naïve patients chronically infected with HCV of genotype 1 in Japan. *J Hepatol* 2012; **56**: 78-84 [PMID: 21827730]
  - 11 **Sherman KE**. Managing adverse effects and complications in completing treatment for hepatitis C virus infection. *Top Antivir Med* 2012; **20**: 125-128 [PMID: 23154251]
  - 12 **Drafting Committee for Hepatitis Management Guidelines, the Japan Society of Hepatology**. JSH Guidelines for the Management of Hepatitis C Virus Infection: A 2014 Update for Genotype 1. *Hepatol Res* 2014; **44** Suppl S1: 59-70 [PMID: 24397840 DOI: 10.1111/hepr.12272]
  - 13 **Welch NM**, Jensen DM. Pegylated interferon based therapy with second-wave direct-acting antivirals in genotype 1 chronic hepatitis C. *Liver Int* 2015; **35** Suppl 1: 11-17 [PMID: 25529082 DOI: 10.1111/liv.12715]
  - 14 **Hayashi N**, Izumi N, Kumada H, Okanoue T, Tsubouchi H, Yatsuhashi H, Kato M, Ki R, Komada Y, Seto C, Goto S. Simeprevir with peginterferon/ribavirin for treatment-naïve hepatitis C genotype 1 patients in Japan: CONCERTO-1, a phase III trial. *J Hepatol* 2014; **61**: 219-227 [PMID: 24727123]
  - 15 **Izumi N**, Hayashi N, Kumada H, Okanoue T, Tsubouchi H, Yatsuhashi H, Kato M, Ki R, Komada Y, Seto C, Goto S. Once-daily simeprevir with peginterferon and ribavirin for treatment-experienced HCV genotype 1-infected patients in Japan: the CONCERTO-2 and CONCERTO-3 studies. *J Gastroenterol* 2014; **49**: 941-953 [PMID: 24626851 DOI: 10.1007/s00535-014-0949-8]
  - 16 **Kumada H**, Hayashi N, Izumi N, Okanoue T, Tsubouchi H, Yatsuhashi H, Kato M, Rito K, Komada Y, Seto C, Goto S. Simeprevir (TMC435) once daily with peginterferon- $\alpha$ -2b and ribavirin in patients with genotype 1 hepatitis C virus infection: The CONCERTO-4 study. *Hepatol Res* 2015; **45**: 501-513 [PMID: 24961662 DOI: 10.1111/hepr.12375]
  - 17 **Kau A**, Vermehren J, Sarrazin C. Treatment predictors of a sustained virologic response in hepatitis B and C. *J Hepatol* 2008; **49**: 634-651 [PMID: 18715665]
  - 18 **Yoshizawa H**, Tanaka J, Miyakawa Y. National prevention of hepatocellular carcinoma in Japan based on epidemiology of hepatitis C virus infection in the general population. *Intervirology* 2006; **49**: 7-17 [PMID: 16166783]
  - 19 **Editors of the Drafting Committee for Hepatitis Management Guidelines: The Japan Society of Hepatology**. Guidelines for the Management of Hepatitis C Virus Infection: First edition, May 2012, The Japan Society of Hepatology. *Hepatol Res* 2013; **43**: 1-34 [PMID: 23332085 DOI: 10.1111/hepr.12020]
  - 20 **Shuster M**, Do D, Nambudiri V. Severe cutaneous adverse reaction to telaprevir. *Dermatol Online J* 2015; **21**: pii: 13030/qt2zq8z9zt [PMID: 25612120]
  - 21 **Federico A**, Sgambato D, Cotticelli G, Gravina AG, Dallio M, Beneduce F, Ruocco E, Romano M, Loguercio C. Skin Adverse Events During Dual and Triple Therapy for HCV-Related Cirrhosis. *Hepat Mon* 2014; **14**: e16632 [PMID: 24734094 DOI: 10.5812/hepatmon.16632]
  - 22 **Sezaki H**, Suzuki F, Hosaka T, Akuta N, Fukushima T, Hara T, Kawamura Y, Kobayashi M, Suzuki Y, Saitoh S, Arase Y, Ikeda K, Kumada H. Effectiveness and safety of reduced-dose telaprevir-based triple therapy in chronic hepatitis C patients. *Hepatol Res* 2014; **44**: E163-E171 [PMID: 24397402 DOI: 10.1111/hepr.12268]
  - 23 **Akuta N**, Suzuki F, Hirakawa M, Kawamura Y, Yatsuji H, Sezaki H, Suzuki Y, Hosaka T, Kobayashi M, Kobayashi M, Saitoh S, Arase Y, Ikeda K, Chayama K, Nakamura Y, Kumada H. Amino acid substitution in HCV core region and genetic variation near the IL28B gene affect viral dynamics during telaprevir, peginterferon and ribavirin treatment. *Intervirology* 2012; **55**: 417-425 [PMID: 21325786]
  - 24 **Reddy KR**, Zeuzem S, Zoulim F, Weiland O, Horban A, Stanciu C, Villamil FG, Andreone P, George J, Dammers E, Fu M, Kurland D, Lenz O, Ouwerkerk-Mahadevan S, Verbinen T, Scott J, Jessner W. Simeprevir versus telaprevir with peginterferon and ribavirin in previous null or partial responders with chronic hepatitis C virus genotype 1 infection (ATTAIN): a randomised, double-blind, non-inferiority phase 3 trial. *Lancet Infect Dis* 2015; **15**: 27-35 [PMID: 25482330]
  - 25 **Enomoto N**, Sakuma I, Asahina Y, Kurosaki M, Murakami T, Yamamoto C, Ogura Y, Izumi N, Marumo F, Sato C. Mutations in the nonstructural protein 5A gene and response to interferon in patients with chronic hepatitis C virus 1b infection. *N Engl J Med* 1996; **334**: 77-81 [PMID: 8531962 DOI: 10.1056/NEJM199601113340203]
  - 26 **El-Shamy A**, Nagano-Fujii M, Sasase N, Imoto S, Kim SR, Hotta H. Sequence variation in hepatitis C virus nonstructural protein 5A predicts clinical outcome of pegylated interferon/ribavirin combination therapy. *Hepatology* 2008; **48**: 38-47 [PMID: 18537193 DOI: 10.1002/hep.22339]
  - 27 **Lin C**, Gates CA, Rao BG, Brennan DL, Fulghum JR, Luong YP, Frantz JD, Lin K, Ma S, Wei YY, Perni RB, Kwong AD. In vitro studies of cross-resistance mutations against two hepatitis C virus serine protease inhibitors, VX-950 and BILN 2061. *J Biol Chem* 2005; **280**: 36784-36791 [PMID: 16087668]
  - 28 **Halfon P**, Locarnini S. Hepatitis C virus resistance to protease inhibitors. *J Hepatol* 2011; **55**: 192-206 [PMID: 21284949]
  - 29 **Sarrazin C**, Zeuzem S. Resistance to direct antiviral agents in patients with hepatitis C virus infection. *Gastroenterology* 2010; **138**: 447-462 [PMID: 20006612]
  - 30 **Scheel TK**, Rice CM. Understanding the hepatitis C virus life cycle paves the way for highly effective therapies. *Nat Med* 2013; **19**: 837-849 [PMID: 23836234]
  - 31 **Webster DP**, Klennerman P, Dusheiko GM. Hepatitis C. *Lancet* 2015; **385**: 1124-1135 [PMID: 25687730]
  - 32 **Kumada H**, Suzuki Y, Ikeda K, Toyota J, Karino Y, Chayama K, Kawakami Y, Ido A, Yamamoto K, Takaguchi K, Izumi N, Koike K, Takehara T, Kawada N, Sata M, Miyagoshi H, Eley T, McPhee F, Damokosh A, Ishikawa H, Hughes E. Daclatasvir plus asunaprevir for chronic HCV genotype 1b infection. *Hepatology* 2014; **59**: 2083-2091 [PMID: 24604476 DOI: 10.1002/hep.27113]

**P- Reviewer:** Conti B **S- Editor:** Qi Y  
**L- Editor:** A **E- Editor:** Liu SQ



## Epidemiology of hepatitis C virus exposure in Egypt: Opportunities for prevention and evaluation

F DeWolfe Miller, Mahmoud S Elzalabany, Sara Hassani, Diego F Cuadros

F DeWolfe Miller, Department of Tropical Medicine, Medical Microbiology and Pharmacology, John A. Burns School of Medicine, University of Hawaii, Honolulu, HI 96813, United States

Mahmoud S Elzalabany, Department of Internal Medicine, Ahmed Maher Teaching Hospital, Cairo 11638, Egypt

Sara Hassani, School of Medicine, Boston University, Boston, MA 02118, United States

Diego F Cuadros, Weill Cornell Medical College - Qatar, Qatar Foundation - Education City, PO Box 24144, Doha, Qatar

**Author contributions:** All authors contributed to the content of the review and editing manuscript drafts; Miller FD and Cuadros DF analyzed the data on injections; Miller FD wrote the manuscript.

**Conflict-of-interest statement:** Authors declare no conflicts of interests for this article.

**Data sharing statement:** The review is based on published work cited in the References. No additional data are available.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to:** F DeWolfe Miller, MPH, PhD, FACE, Professor of Epidemiology, Department of Tropical Medicine, Medical Microbiology and Pharmacology, John A. Burns School of Medicine, University of Hawaii, 651 Ilalo Street, BSB 320C, Honolulu, HI 96813, United States. [dewolfe@hawaii.edu](mailto:dewolfe@hawaii.edu)  
Telephone: +1-808-6921605  
Fax: +1-808-6921979

Received: July 1, 2015

Peer-review started: July 6, 2015

First decision: September 22, 2015

Revised: October 24, 2015

Accepted: November 23, 2015

Article in press: November 25, 2015

Published online: December 8, 2015

### Abstract

**AIM:** To critically evaluate the current epidemiology data on exposures, rather than infection, to hepatitis C virus (HCV) transmission and recommend epidemiologic strategies to fill gaps.

**METHODS:** Standard methods for identifying and evaluating relevant epidemiologic literature and available data were used.

**RESULTS:** There is a large body of literature on the epidemiology of HCV transmission in Egypt that collectively identifies ongoing iatrogenic exposures as the major driver for HCV transmission due to shortcomings in infection control and standard procedures. Additional epidemiologic studies on HCV transmission that requires the participation of human subject is unwarranted. Alternatively, very little literature was found on the epidemiology of exposure to HCV, infection control, and safe injection practices. The information that is available on patterns of HCV exposure shows high frequencies of inadequate infection control, problems in sterilization in health care facilities, low rates of hand washing, untrained personnel, lack of stated policies in facilities, HCV contamination of instruments and very large injection frequencies with low but very significant syringe and needle reuse. There is an important need to increase the number, size, and diversity of epidemiologic studies on HCV exposures, patterns of risk factors for infection, infection control, and safe injection practices. In addition to health care facilities evaluation, relevant knowledge attitude and practice studies are recommended.

**CONCLUSION:** Epidemiologic methods on HCV ex-

posure can be used to characterize the magnitude of exposures to HCV infection, target interventions to reduce exposures, and provide the best method for evaluating interventions by demonstrating the reduction of exposure to HCV infection.

**Key words:** Epidemiology; Hepatitis C virus; Egypt; Exposure; Prevention; Epidemic

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Much has been published on the epidemiology of hepatitis C virus epidemic in Egypt. The exposures that drive this epidemic are iatrogenic. This review focuses on what has been published on the epidemiology (patterns, distributions, and related factors) of the iatrogenic exposures. The review found that very little has been published on epidemiology of the exposures driving the epidemic. This is essential for developing effective interventions and evaluating prevention programs. Recommendations are given.

Miller FD, Elzabalany MS, Hassani S, Cuadros DF. Epidemiology of hepatitis C virus exposure in Egypt: Opportunities for prevention and evaluation. *World J Hepatol* 2015; 7(28): 2849-2858 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i28/2849.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i28.2849>

## INTRODUCTION

Kamel *et al.*<sup>[1]</sup> reported in 1992 an unusually high prevalence of anti-hepatitis C virus (HCV) antibodies in over 2000 first time healthy Egyptian blood donors in Cairo, Egypt. The prevalence was 10.1%, five to ten times higher than what had been reported elsewhere in the world<sup>[2]</sup>. This was a population based study of first time blood donors in urban Egypt and likely underestimated anti-HCV antibodies prevalence in the general population.

A follow up to the blood donor study, Kamel *et al.*<sup>[3]</sup> completed a rural population based community study of anti-HCV antibodies in 1994. The study included the entire population of a remote village in the northern Nile Delta. The overall anti-HCV antibodies prevalence in the village was 17.6%. In both of these population based epidemiologic studies, prevalence of anti-HCV antibodies increased strongly with age as shown in Figure 1. In the blood donors and the village study, the prevalence of anti-HCV antibodies was similar in both sexes.

These were the first two population based studies reported in Egypt providing the initial evidence that there was an extraordinary HCV epidemic unlike anywhere else in the world. These and many similar studies that followed<sup>[4-9]</sup>, including two national studies<sup>[10,11]</sup>, reinforced these observations. The 2008 national estimate of anti-HCV antibodies prevalence was 14.7% and the estimate for HCV RNA prevalence was 9.7%<sup>[10]</sup>. Given

a national population of about 80 million persons, 7.8 million were estimated to be asymptotically infected with HCV comprising a large reservoir of HCV in the population. It is now well established that Egypt has a HCV epidemic, which is the largest HCV epidemic in the world, and the epidemic is ongoing<sup>[2,9,12-14]</sup>.

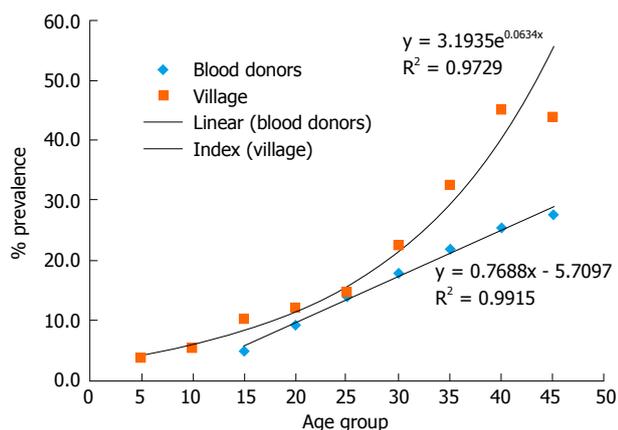
Epidemiologic tools are needed in Egypt to evaluate the magnitude and patterns of exposure to HCV transmission. Exposure to HCV is similar in concept to risk factors or independent variables associated with HCV transmission. The distribution and determinates of HCV exposures and related factors are fundamental for rationale allocations of resources for intervention by reducing exposure to HCV infection. Secondly, an extension of these epidemiologic tools is needed to evaluate intervention programs by demonstrating a reduction in the magnitude and patterns of HCV exposure.

The aim of our study was to first briefly summarize the epidemiology of HCV transmission (HCV T), identify all epidemiologic studies completed in Egypt to date that were designed to describe the magnitude, patterns and determinates of exposures (predominately iatrogenic exposures) to HCV transmission (HCV E), characterize gaps in HCV E and finally demonstrate and provide examples for the application of HCV E for the evaluation of public health interventions to reduce exposure to HCV.

## MATERIALS AND METHODS

A literature review was conducted to identify publications on the epidemiology of HCV transmission and on the epidemiology of iatrogenic exposures in Egypt using methods previously published by us<sup>[14]</sup>. Briefly, a search of all published peer-reviewed literature (English language) from 1992 to 2015 on HCV and Egypt was made using the National Library of Medicine, PubMed, Google Scholar, Web of Science, Biological Abstracts, manual review of citations in search-identified publications and in Egypt for reports available only locally. Studies that: (1) reported HCV prevalence or incidence; (2) described the serologic methods; (3) were of cross-sectional or prospective epidemiologic design; and (4) could be abstracted for the purposes of the study were included. Studies on infection control were included as proxy for iatrogenic exposures.

One of our objectives was to build a complete bibliographic database on iatrogenic exposures and infection control practices in Egypt. From this, an assessment of epidemiologic methods for investigations on iatrogenic exposures or HCV E could be evaluated. Methodologically sound studies were sought as examples for investigation as well as methods for evaluation of intervention programs to reduce and prevent iatrogenic transmission of HCV in Egypt. Additional data were based on personal onsite visits in Egypt and anecdotal observation as some potentially iatrogenic practices have not been formally published and appear to be unique to Egypt. An example is the widely practiced re-use of latex gloves or not using



**Figure 1** Prevalence of anti-hepatitis C virus antibody in urban blood donors and rural villagers.

latex gloves when indicated.

## RESULTS

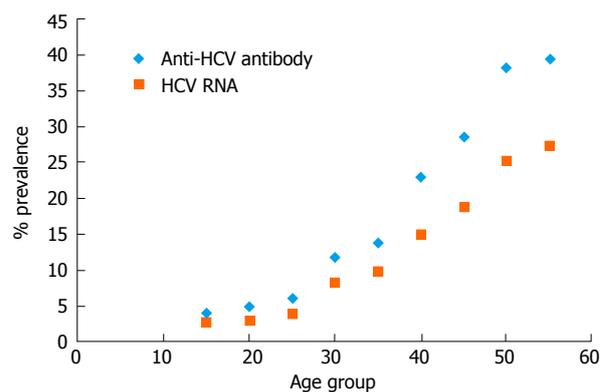
### *History of HCV epidemic in Egypt*

Following the pioneering work of Kamel *et al.*<sup>[1,3]</sup>, many similar studies in rural communities and selected health clinics followed without adding any significant new findings. Beyond the national blood donor screening program, no national program for the prevention of HCV emerged in the first decade after discovery. In part, this may have been due to the natural history of HCV infection. In a primary HCV infection, there is rarely an acute phase. Manifestations of liver dysfunction and disease usually do not occur until one or more decades later. HCV became a silent epidemic.

The origins of the HCV epidemic in Egypt are not clear but thought to be due to past and ongoing iatrogenic exposures<sup>[15,16]</sup>. Iatrogenic exposures and failure in infection control could be frequently seen on visits to health care facilities throughout the large Egyptian health care system.

A report in 1997 showed an association between anti-HCV antibodies and a history of parenteral therapy for schistosomiasis and surgery<sup>[17]</sup>. In 2000, a report in the *Lancet* suggested that the epidemic was due in part to the previous wide spread rural campaigns of parenteral anti-schistosomiasis therapy (PAT)<sup>[15]</sup>. The Egyptian medical care system embraced this report as the cause of the epidemic. More importantly, Egypt's physicians concluded that since these campaigns had ended more than three decades ago, the cause of the epidemic, PAT, had ended and therefore transmission had ended as well. If this was true, then epidemiologically the prevalence of anti-HCV antibodies in Egyptians 30 years old and younger should be similar to other countries. That is from 1% to 3% or less. As shown in Figures 1 and 2, anti-HCV antibodies prevalence increase from the earliest age. There is now abundant evidence that there is an ongoing HCV epidemic in Egypt.

The first formal epidemiologic study on infection



**Figure 2** Prevalence by age of anti-hepatitis C virus antibody and hepatitis C virus RNA in 2008 Demographic Health Survey survey. HCV: Hepatitis C virus.

control and iatrogenic exposures (HCV E) to HCV transmission in Egypt was in 2006<sup>[18]</sup>. This was a survey of large and small health care facilities. The study showed a complete lack of infection control practices in all facilities. In this study the presence or absence of infection control programs was an index of iatrogenic exposure to HCV. The American Centers for Disease Control and Prevention reported in 2012 that Egypt continued to face an ongoing HCV epidemic and that a comprehensive prevention plan was needed<sup>[13]</sup>.

### *HCV transmission in Egypt*

HCV is a blood borne pathogen and the transmission and epidemiology of HCV is well established<sup>[16]</sup>. The virus is inherently liable with exponential die off in 24 h under laboratory conditions<sup>[19,20]</sup>, is less infectious than hepatitis B virus (HBV) and slightly more infectious than human immunodeficiency virus (HIV)<sup>[21]</sup>.

The probability of HCV transmission varies by different routes of exposure as shown in Table 1. Note that the significance relative to the ongoing epidemic in Egypt of a given exposure route of transmission has been included in this table. For example, the probability of HCV transmission by contaminated blood or organ donation results in all recipients becoming infected. In this case the probability of transmission and infection is one (100%). Contaminated blood and organ donation have the highest HCV transmission. Nationally mandated HCV blood donor screening program in 1994 reduced exposure frequency to the point that blood transfusion, once a significant route of transmission in Egypt and elsewhere, no longer plays a significant role in regard to the general population<sup>[22]</sup>.

Injection drug users (IDU) were found to have very high prevalence of HCV infection presumably due to the reuse and sharing of drug injection equipment<sup>[23,24]</sup>. Contaminated drug injection equipment has a high probability of transmission although there is no exact probability estimate available. The exact population of IDU in Egypt is not well defined although considered small. Moreover, HCV transmission with IDU groups, unlike HIV, poses a very low probability of exposure to

**Table 1** The probability of hepatitis C virus transmission by different routes of exposure

Exposure route	Exposure frequency	Transmission probability	Population exposed	Significance <sup>1</sup>	Ref.
Blood transfusion	Very low	1 (100%)	<sup>2</sup> 300 k/yr	Zero	[2,16]
Organ donation	Very low	1	< 100/yr	Zero	[2]
Injection drug users	High	≥ 0.8	Very small	Very low	[23,24]
Hemodialysis	High	≥ 0.75	Small	Very low	[26-28]
Sexual	High	Unk <sup>3</sup>	Adults	Zero	[29]
Intrafamilial	Unknown	Unk	General	Very low	[15,16]
Needle stick	High	≤ 0.02	Occupational	Low	[31]
Injections	<sup>4</sup> 4.1/p per year	≤ 0.02	General	High	[17,30,53,60]
Maternal	High	0.02	New born	Low	[33,43]
Dental	High	≤ 0.02	General	High	[62-65]
Iatrogenic	High	≤ 0.02	General	High	[14,42,43,45,49,50,54]

<sup>1</sup>Significance refers to the extent that the specific exposure route contributes to overall national HCV transmission in Egypt; <sup>2</sup>1000 persons per year;

<sup>3</sup>Unknown; <sup>4</sup>Persons per year. HCV: Hepatitis C virus.

the general population<sup>[25]</sup>.

It has been long recognized that HCV infection, like HIV, was a risk for hemodialysis patients<sup>[26-28]</sup>. In Egypt, this became a national scandal. From 46.1% to 100% of HCV negative dialysis patients would convert to HCV positive within a year in dialysis centers throughout the country<sup>[16,26,28]</sup>. Considerable efforts both in the public and private sector have reduced HCV transmission in local dialysis centers. However, the general population exposed to this risk is small and the significance of HCV positive dialysis patients to the overall epidemic is very low.

Sexual and intra-familial transmission of HCV remains to be unequivocally established in Egypt or elsewhere. Sexual transmission of HCV is controversial. HCV discordant monogamous couples showed almost no transmission for long periods and recovery of HCV from semen or other genital fluids has proved to be difficult<sup>[29]</sup>. Sexual transmission does not play a significant role in Egypt. The studies of intra-familial transmission of HCV in Egypt<sup>[30-32]</sup> have validity issues (small numbers, confounding, selection biases) and have not been replicated. No specific intra-familial exposure to HCV transmission has been identified. Familial sharing of any medical equipment such as syringe and needles or diabetic testing equipment could result in exposure to HCV transmission, but this remains to be established.

Confirmed occupationally related accidental needle sticks from HCV positive patients have a probability of infection slightly greater than HIV but much lower relative to the probability of HBV infection. The probability of transmission has been estimated to be approximately 0.01 to 0.02<sup>[21]</sup>. Accidental occupational needle sticks in Egypt is a significant exposure<sup>[33-36]</sup>. However, the extent that this exposure contributes to transmission in the general population is not known but not likely to be significant.

Transmission of HCV infection from mother to new born however is well established. In Egypt, we have estimated that there are 5000 newborns infected with HCV every year<sup>[37]</sup>.

Iatrogenic transmission of HCV has been documented

globally<sup>[2,24]</sup>. Iatrogenic transmission can be complex due to the many possible routes of exposure from contaminated medical and dental instruments, sharps, needles, invasive procedures, contaminated multi-dose vials, blood, or blood product transfusion, organ transplantation or any of many kinds of medical/dental percutaneous exposures.

A key element of iatrogenic transmission is patient to patient exposure where the first patient is knowingly or more likely unknowingly asymptotically infected. This patient is a key to the exposure and contamination of medical or dental instruments, sharps, or needles. Failure in infection control or standard procedures to prevent a second patient to be percutaneously or parenterally exposed to a contaminated instrument or sharp has a relatively low probability of HCV transmission and infection<sup>[21]</sup>.

Iatrogenic transmission of HCV in Egypt is well documented<sup>[7,17,38-54]</sup>. These reports identify iatrogenic transmission as the principal driver of the HCV epidemic in Egypt. Accordingly, in Egypt, if the probability of iatrogenic exposure is the same across a health care system, then the probability of transmission will be greater in patient populations who have a higher prevalence of chronic HCV RNA infection, symptomatic or not. This reservoir of chronic HCV infected patients is known to be high in Egypt. Overall, 10% of the Egyptian population is HCV RNA positive<sup>[10]</sup>. This varies with rural populations having a higher prevalence of HCV RNA relative to urban populations. In Egypt, like anti-HCV antibodies, HCV RNA positivity increases directly with age as shown in Figure 2. The reservoir of HCV infection is therefore higher in older patients relative to younger patients.

### **Epidemiology of HCV transmission and epidemiology of HCV exposure in Egypt**

The epidemiology of HCV T in Egypt is defined as reports which estimate patterns of HCV infection in people and associations with a possible exposure to HCV infection. For example the report by el-Sayed *et al*<sup>[17]</sup> showed an odds ratio (OR = 7.9) with a history of

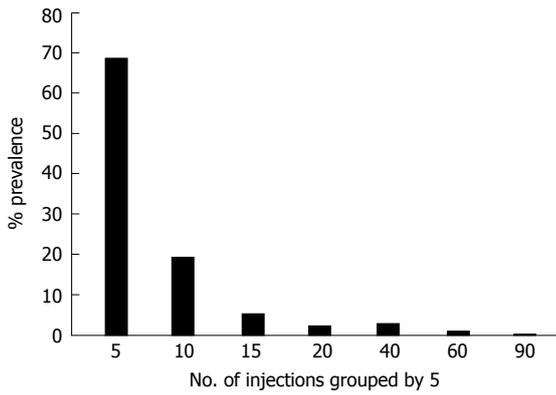


Figure 3 Percentage and number of injections reported in the past 6 mo.

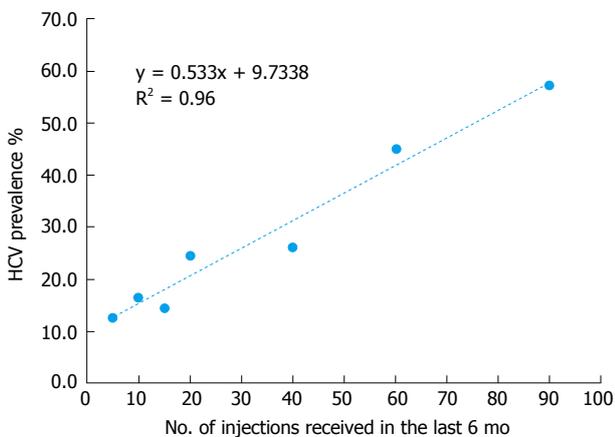


Figure 4 Injections received in last 6 mo and hepatitis C virus antibody percent prevalence. HCV: Hepatitis C virus.

PAT exposure. It is important to note that, as shown in this study, there is an abundance of similar HCV T literature on the prevalence, incidence, and risk factors for infection. Additional HCV T studies are unwarranted. There is no continued justification to test individuals for HCV antibodies or HCV RNA for the purposes of HCV T studies. Conversely, the epidemiology of exposure to HCV transmission (HCV E) would provide data on the patterns and frequency of exposures. Examples of HCV E are given below.

**Injections:** Reuse of inadequately sterilized needles and syringes during the PAT campaigns in rural Egypt over 30 years ago is cited as a major factor in the origin of the Egyptian HCV epidemic<sup>[15,21,33,34,55-60]</sup>. The PAT hypothesis has been challenged on the bases that there was a greater concurrent abundance of iatrogenic exposures throughout Egypt in addition to PAT exposures<sup>[49]</sup>. Accordingly, HCV E studies would characterize the patterns and frequency of injections and identify the magnitude and determinates of safe and unsafe injection practices.

Talaat *et al.*<sup>[60]</sup> has estimated a high rate of injections in Egypt at 281 million per year. These injections were administered by public and private sector physicians, pharmacists, barbers, doctor assistants, housekeepers, relatives and friends and 8.4% reported that the syringe

Table 2 The number of persons who received one or more injections in the last 6 mo by gender

Gender	Yes	No	Total	Prevalence %	95%CI (lower-upper)
Female	1493	4661	6154	24.3	23.2-25.4
Male	625	4403	5028	12.4	11.5-13.3
Total	2118	9064	11182	18.9	18.2-19.6
OR = 2.3					2.0-2.5

Demographic Health Survey 2008<sup>[10]</sup>. OR: Odds ratio.

was not taken from a closed packet<sup>[60]</sup>. Many in rural areas resort to informal nonprofessionals for injections<sup>[56]</sup>.

Safe injection practices were directly observed in a cross sectional study among 1100 healthcare workers located in 25 healthcare facilities in the Gharbiya governorate in the Nile Delta<sup>[33,34]</sup>. Noted was a lack of supplies needed for safe injections and safe practice was infrequent. Important policies were lacking including an infection control committee and dedicated infection control personal. Most importantly, there were an estimated 13.2% of syringes and needles reused. The important data on exposure in this study included lack of supplies, safe injection practices, essential policies, lack of infection control committee and personal and syringe and needle reuse.

Shown here for the first time is an analysis of injection data collected by the 2008 Egyptian Demographic Health Survey<sup>[10]</sup>. Shown in Table 2 is the frequency of injections reported by participants in the 2008 nationally representative study. Almost 19% (18.5%) received one or more injections in the last six months. Women reported receiving twice as many injections as men (OR = 2.3; 95%CI: 2.0-2.5). Figure 3 shows the pattern of participants reporting multiple injections in groups by 5. By far, the largest group received 1 to 5 injections in the last 6 mo followed by a sharp decline. Figure 4 shows the relationship between HCV antibodies prevalence and the pattern of multiple injections. A strong ( $R^2 = 0.96$ ) and direct relationship of increasing HCV antibody prevalence and the number of injections is shown. These results suggest that increased frequency of injections has a direct relation with increased HCV antibodies positivity. An alternate interpretation is that individuals who are receiving multiple frequent injections have serious medical conditions and more likely have additional iatrogenic exposures.

The measure of exposure to HCV was the following question: "The last time you had an injection from a health worker, did the person who gave you that injection take the syringe and needle from a new, unopened package?". Of those that received injections, 14.8% answered "No". Data are shown in Table 3. There may be concerns about how this question was interpreted by the participant. The injection may have been prepared in a separate area from the patient. The patient may not know or be able to confirm that the syringe and needle came from a new unopened package. However, this estimate is similar in magnitude

**Table 3** To answer the question: "The last time you had an injection from a health worker did the person who gave you that injection take the syringe and needle from a new, unopened package?" Demographic Health Survey 2008<sup>[10]</sup>

Question <sup>1</sup>	Yes	No	Total	Prevalence %	95%CI (lower-upper)
New Unopened Package	1273	221	1494	14.80	13.0-16.6

<sup>1</sup>Among those who received an injection and replied to the question.

to that from the study conducted in Gharbiya<sup>[33,34]</sup>, in which Talaat *et al.*<sup>[60]</sup> estimated about 8.4% in an answer to a similar question.

**Dental health care:** Dental health care has a significant potential for iatrogenic HCV transmission<sup>[61-63]</sup>. Only a single report of HCV iatrogenic exposure in Egyptian dental clinics was found published by Hashish *et al.*<sup>[64]</sup> in 2012. The study measured the presence of HCV RNA by reverse transcription polymerase chain reaction (RT-PCR) on various dental instruments in selected dental clinics in Alexandria. The study found that 18% of the dental instruments in the dental clinics visited were positive for HCV RNA indicating the presence of the virus. This study demonstrates a method for measuring iatrogenic exposure to HCV infection from dental health care by showing the presence of HCV contaminated dental instruments.

The exact reason for the contamination of these instruments was not given. The authors did suggest that the lack of sterilization equipment for instruments was a short coming. What was not provided was the number of sets of instruments, patient load, hand washing practices, use of latex gloves, how instruments were cleaned and disinfected, the presence of operating sterilization equipment, and if there had been specific efforts and policies present for training and preventing HCV transmission.

The prevalence of HCV is lowest in Alexandria most likely due to better infection control than most other areas of Egypt, especially rural areas. The observed results are therefore most likely an under estimate.

The correct method to expand this epidemiologic approach to HCV exposure in dental clinics would be to include other measures of infection control mentioned above in recording data. Secondly, this study design could be used at the community level to describe the epidemiology of HCV exposure in dental clinics by obtaining a list of all dental care facilities in the community and decide on drawing a representative sample of dental care facilities or to include all facilities in the study. Results of a sample could be used to describe the magnitude of HCV contamination and provide justification for professional improvement or training programs. Results based on facility specific level would be used for compliance and licensing.

Detection of HCV RNA by RT-PCR could be resource

challenging. A lower resource approach would have the same primary step of community and facility identification and sampling. Each facility would provide details on cleaning and sterilization methods and an inventory of dental equipment sets and daily patient loads. Large patient to equipment ratios would suggest a trigger for compliance or licensing issues.

**Health care facilities:** Egypt has a national health care system dating back to the 19<sup>th</sup> century. Health care facilities in Egypt are a complex organization of public and private sector facilities that include medical care, dental care, clinical laboratories, and pharmacies.

Pharmacies in Egypt give injections, intravenous fluids, and provide testing for glucose levels. Glucometers are an overlooked exposure to HCV transmission. Testing is carried out by pharmacy technicians who may have only secondary school education and do unsupervised home visits. No studies have been done on exposure to HCV infections related to Egyptian pharmacies.

El-Zanaty *et al.*<sup>[65]</sup> carried out a national service provision assessment survey in 2004. A portion of this study was on "Systems for Infection Control". Medical care facilities rather than individuals were surveyed in a nationally representative sample using very specific standardized data forms. This report has considerable details representing all regions of Egypt and covering all types of medical care facilities. Private clinics, private pharmacies, and dental services were not included.

A statement in a summary of the findings reported a significant decrease from 2002 to 2004 in almost all indicators of infection control. It was concluded that infection control practices were extremely weak. Only 4% of all facilities were adherent to all infection control measures. New disposable syringes and needles were however universal. The study could uniquely compare changes over time to their previous publication in 2002<sup>[66]</sup>.

**Informal health care providers:** Egypt has a large undocumented sector of informal health care provides. These providers do not have formal education or training and provide services for injections, dentistry, wound treatment, and male circumcision. Traditional birth attendants were reported to oversee > 50% of all births. "Injectionists" included barbers and staff at pharmacies. A study of Informal health providers in two Egyptian villages found that these providers, "knew little about HCV" and its transmission<sup>[56]</sup>.

A number of studies in Egypt have included barbers as a possible exposure to HCV<sup>[41,43,44,56,67,68]</sup>. This is based on the assumption of percutaneous exposure by shaving. Most studies found that there is no evidence for transmission. This is consistent with viral fragility and the unlikelihood of the virus remaining viable in the soap used in shaving. In a study of barbers in the Gharbia governorate<sup>[68]</sup>, anti-HCV antibodies were detected in 12.3% of barbers and 12.7% of clients. Knowledge of HCV prevention was reported to be high among the majority of participating barbers and good practices

during shaving and hair-cutting were observed for the majority of barbers.

### **HCV prevention in Egypt**

The transmission of HCV is entirely preventable. Throughout the world, HCV transmission is prevented by good medical and dental care practices which reduce and eliminate iatrogenic exposure to infection. This includes following well established aseptic techniques, standard procedures and universal precautions<sup>[13]</sup>. Measures should and can be taken to reduce and eliminate all HCV transmission routes listed in Table 1.

### **HCV prevention in the Egyptian health care educational system:**

Aseptic techniques, standard procedures, and universal precautions are or should be introduced in all health care curricular in the Egyptian health care educational system and reinforced at every level of health care service. This educational intervention is not intellectually complex or challenging to teach. In fact, teaching and training could be done entirely using modern computer based social media. Moreover, the interventions to reduce HCV exposure do not require new or costly technology. There is no publicly available information on the evaluation of the Egyptian health care educational system in regard to HCV prevention curricular. Technically, evaluation of the Egyptian health care educational system is straight forward.

**Recommendation:** Conduct an examination on a sample of recent and past graduates from medicine, nursing, dentistry, and pharmacy on HCV prevention. This is an essential data for status on current knowledge and a benchmark to which future evaluation can be based.

Evaluation of knowledge, attitudes, and practices (KAP) of health care providers on standard procedures and infection control is necessary for the reduction of exposure to HCV. Well-designed repeated KAP studies based on representative samples stratified by health care provider categories can provide direct evidence of HCV exposures defined as incomplete knowledge and practice errors. The large Egyptian health care syndicates can provide needed sampling frames. These syndicates can also provide opportunities to spread important messages about HCV exposure prevention.

**Recommendation:** KAP study on health care providers on HCV prevention is needed.

In addition to curricular interventions, professional development or continued health care education should be developed for online certification or re-certification for preventing exposure to HCV infection.

**Screening of blood and blood products:** Globally, blood donations are screened for HCV. Egypt mandated a national blood donation HCV screening program by 1994<sup>[22]</sup>. Before this national program, approximately 9% of blood and blood products recipients would have

become infected. Assuming the probability of HCV transmission at 100% among exposed recipients and 300000 recipients per year, this program has prevented to date at least half a million people from becoming infected. Maintaining and enhancing Egypt's national blood donation HCV screening program is essential.

## **DISCUSSION**

The number and extent of studies which test for the presence of HCV in individuals (HCV T) have been considerable dating from the first reports in 1992. Given the abundance of literature on HCV T, additional HCV T studies are unwarranted. There is no continued justification to test individuals for HCV antibodies or RNA for the purposes of HCV T studies. Aside from cost, there are additional methodological limitations for both cross sectional and prospective HCV T studies in Egypt that further support discontinuation of these investigations. Due to the natural history of HCV, there are serious validity issues for cross sectional, prospective, and case control study designs undermining hypothesis testing related inferences. Population prevalence estimates are not useful for evaluating interventions and prevention measures<sup>[69]</sup>. Generating national incidence rates requires very large samples, is very costly, requires long term follow ups, and has limited validity determining past exposures. Prospective incidence studies are strongly discouraged especially for intervention assessment and project evaluation. The difficulty of using prospective incidence studies for intervention assessment is not unique to this public health problem but is widely recognized by public health practitioners.

Successful interventions will reduce exposure to HCV infection which will decrease the incidence. It is strongly recommended to use HCV E studies, rather than prospective incidence studies to evaluate the control of the ongoing epidemic. For that, benchmark HCV E data are needed which can be obtained from cross sectional studies.

Few studies on HCV E were found. However, these studies illustrate the basic low cost low technology methodology needed to document exposures to HCV infection. The only national level study using facility surveys completed by El-Zanaty *et al*<sup>[65,66]</sup> in 2002 and 2004 is a good example. These facility studies could be modified to focus more on infection control evaluation. This would provide direct data on the frequency and patterns of HCV exposures. These observational based studies can be complemented by interview based KAP studies adapted to the unique Egyptian epidemic directed at infection control documentation and evaluation.

It is essential to conceptualize the methodological approach epidemiologically for HCV E. The objective is to epidemiologically describe the prevalence of a specifically defined set of health care practices and procedures which can be evaluated as being correctly or incorrectly done with regard to standard procedures<sup>[64,65]</sup> in a given health care setting. As mentioned above this

should include both observational and interview based data collection. Specific modifications are needed for the different types of Egyptian health care facilities.

Methods for standardizing data collection on infection control in general and injection practices in specific for Egypt are needed. Standardized methods are also needed for direct and indirect observational data collection and data collected by interview. Standards for collecting photographic documentation would be useful. Pilot testing with independent verification is advisable. This capacity exist in Egypt as demonstrated by the studies carried out by El-Zanaty *et al.*<sup>[10,65]</sup>.

There are many unique infection control violations in Egypt which have the potential for exposure to HCV infection. These exposures unique to Egypt are poorly documented and should be thoroughly investigated and included in any comprehensive prevention program to reduce exposure to HCV transmission. There is a large unregulated informal health care system in Egypt that contributes significantly to injections and other poorly regulated procedures<sup>[18,60]</sup>.

It is recommended that efforts be made to develop strong HCV E studies that generate a comprehensive inventory of all typical and unique iatrogenic exposures, where these exposures are occurring, the magnitude of these exposures (number of individuals potentially exposed), probability of transmission and create an index of iatrogenic transmission. The HCV index of transmission (HCV IT) would incorporate the magnitude of population exposed and probability of transmission. For example, HCV contaminated blood transfusion has 100% probability of transmission<sup>[2,16]</sup>, a restricted population exposed (blood donor recipients) and with the current level of blood donor screening an overall low index. Given the magnitude of injections received in Egypt with much lower probability of transmission *via* contaminated drug vials or syringe and or needle reuse, the overall HCV IT is likely to be very significant<sup>[53]</sup>.

Strong well designed HCV E studies have a dual function. The first objective is to better and more precisely document the distribution and determinates of specific iatrogenic exposures. This is essential to provide a baseline for evaluation. The second objective is evaluation of intervention programs. That is to quantitatively demonstrate a reduction in exposure to HCV transmission by an improvement in infection control measures and safe injection practices by direct and indirect measures<sup>[10,60,64,65]</sup> with follow up HCV E studies.

## COMMENTS

### Background

Egypt has the largest epidemic of hepatitis C virus (HCV) in the world. A review of the epidemiologic literature on HCV was completed.

### Research frontiers

HCV is entirely preventable. The exposure factors to HCV infection driving the epidemic in Egypt have been thoroughly identified as iatrogenic. The authors' aim was to assess the magnitude of the epidemiologic literature on iatrogenic exposures in Egypt.

### Innovations and breakthroughs

The authors found that the amount of epidemiologic information on iatrogenic exposures needed for designing the prevention of HCV transmission was very limited, especially in contrast to the epidemiology on HCV infection or transmission.

### Applications

The epidemiology of exposure to HCV transmission, that is predominately iatrogenic exposures, is essential information and knowledge needed to design, guide, and evaluate interventions to reduce iatrogenic exposures and prevent HCV transmission. The application of epidemiologic investigation on exposures does not include human subjects which vastly reduces the cost and complexity of data collection. Recommendations and suggested epidemiologic approaches and designs were given.

### Terminology

The authors refer to classical epidemiology of HCV infection, transmission, and prevalence as HCV transmission. Individuals participating in these types of studies have to provide a specimen for HCV testing, understand the consequences of being found HCV positive, and if viremic, referred for treatment. The authors refer to the epidemiologic investigation of exposures to HCV infection as HCV E. Knowledge, attitude, practice (KAP) studies are an example of methodology that could be adapted to exposure epidemiology. The KAP of injection preparation and administration is an example.

### Peer-review

The review is very interesting and gives a fairly comprehensive overview of the situation in Egypt.

## REFERENCES

- 1 **Kamel MA**, Ghaffar YA, Wasef MA, Wright M, Clark LC, Miller FD. High HCV prevalence in Egyptian blood donors. *Lancet* 1992; **340**: 427 [PMID: 1353577 DOI: 10.1016/0140-6736(92)91508-6]
- 2 **Alter MJ**. Epidemiology of hepatitis C virus infection. *World J Gastroenterol* 2007; **13**: 2436-2441 [PMID: 17552026 DOI: 10.3748/wjg.v13.i17.2436]
- 3 **Kamel MA**, Miller FD, el Masry AG, Zakaria S, Khattab M, Essmat G, Ghaffar YA. The epidemiology of Schistosoma mansoni, hepatitis B and hepatitis C infection in Egypt. *Ann Trop Med Parasitol* 1994; **88**: 501-509 [PMID: 7979640]
- 4 **el Gohary A**, Hassan A, Nooman Z, Lavanchy D, Mayerat C, el Ayat A, Fawaz N, Gobran F, Ahmed M, Kawano F. High prevalence of hepatitis C virus among urban and rural population groups in Egypt. *Acta Trop* 1995; **59**: 155-161 [PMID: 7545863 DOI: 10.1016/0001-706X(95)00075-P]
- 5 **Quinti I**, Renganathan E, El Ghazzawi E, Divizia M, Sawaf G, Awad S, Pana A, Rocchi G. Seroprevalence of HIV and HCV infections in Alexandria, Egypt. *Zentralbl Bakteriol* 1995; **283**: 239-244 [PMID: 8825115 DOI: 10.1016/S0934-8840(11)80205-7]
- 6 **Darwish MA**, Faris R, Clemens JD, Rao MR, Edelman R. High seroprevalence of hepatitis A, B, C, and E viruses in residents in an Egyptian village in The Nile Delta: a pilot study. *Am J Trop Med Hyg* 1996; **54**: 554-558 [PMID: 8686770]
- 7 **el-Sayed NM**, Gomatos PJ, Rodier GR, Wierzbza TF, Darwish A, Khashaba S, Arthur RR. Seroprevalence survey of Egyptian tourism workers for hepatitis B virus, hepatitis C virus, human immunodeficiency virus, and Treponema pallidum infections: association of hepatitis C virus infections with specific regions of Egypt. *Am J Trop Med Hyg* 1996; **55**: 179-184 [PMID: 8780457]
- 8 **Mohamed MK**, Rakhaa M, Shoeir S, Saber M. Viral hepatitis C infection among Egyptians the magnitude of the problem: epidemiological and laboratory approach. *J Egypt Public Health Assoc* 1996; **71**: 79-111 [PMID: 17217003]
- 9 **Lehman EM**, Wilson ML. Epidemic hepatitis C virus infection in Egypt: estimates of past incidence and future morbidity and mortality. *J Viral Hepat* 2009; **16**: 650-658 [PMID: 19413698 DOI: 10.1111/j.1365-2893.2009.01115.x]
- 10 **El-Zanaty F**, Way A. Egypt Demographic and Health Survey

2008. Cairo: El-Zanaty and Associates, and Macro International, 2009
- 11 **Mohamed MK.** Epidemiology of HCV in Egypt. *Afro-Arab Liver J* 2004; **3**: 41-52
  - 12 **Lavanchy D.** Evolving epidemiology of hepatitis C virus. *Clin Microbiol Infect* 2011; **17**: 107-115 [PMID: 21091831 DOI: 10.1111/j.1469-0691.2010.03432.x]
  - 13 **Centers for Disease Control and Prevention (CDC).** Progress toward prevention and control of hepatitis C virus infection--Egypt, 2001-2012. *MMWR Morb Mortal Wkly Rep* 2012; **61**: 545-549 [PMID: 22832935]
  - 14 **Miller FD, Abu-Raddad LJ.** Evidence of intense ongoing endemic transmission of hepatitis C virus in Egypt. *Proc Natl Acad Sci USA* 2010; **107**: 14757-14762 [PMID: 20696911 DOI: 10.1073/pnas.1008877107]
  - 15 **Frank C, Mohamed MK, Strickland GT, Lavanchy D, Arthur RR, Magder LS, El Khoby T, Abdel-Wahab Y, Aly Ohn ES, Anwar W, Sallam I.** The role of parenteral antischistosomal therapy in the spread of hepatitis C virus in Egypt. *Lancet* 2000; **355**: 887-891 [PMID: 10752705 DOI: 10.1016/S0140-6736(99)06527-7]
  - 16 **Mohamoud YA, Mumtaz GR, Riome S, Miller D, Abu-Raddad LJ.** The epidemiology of hepatitis C virus in Egypt: a systematic review and data synthesis. *BMC Infect Dis* 2013; **13**: 288 [PMID: 23799878 DOI: 10.1186/1471-2334-13-288]
  - 17 **el-Sayed HF, Abaza SM, Mehanna S, Winch PJ.** The prevalence of hepatitis B and C infections among immigrants to a newly reclaimed area endemic for *Schistosoma mansoni* in Sinai, Egypt. *Acta Trop* 1997; **68**: 229-237 [PMID: 9386797 DOI: 10.1016/S0001-706X(97)00097-1]
  - 18 **Talaat M, Kandeel A, Rasslan O, Hajjeh R, Hallaj Z, El-Sayed N, Mahoney FJ.** Evolution of infection control in Egypt: achievements and challenges. *Am J Infect Control* 2006; **34**: 193-200 [PMID: 16679176 DOI: 10.1016/j.ajic.2005.05.028]
  - 19 **Kamili S, Krawczynski K, McCaustland K, Li X, Alter MJ.** Infectivity of hepatitis C virus in plasma after drying and storing at room temperature. *Infect Control Hosp Epidemiol* 2007; **28**: 519-524 [PMID: 17464909 DOI: 10.1086/513727]
  - 20 **Song H, Li J, Shi S, Yan L, Zhuang H, Li K.** Thermal stability and inactivation of hepatitis C virus grown in cell culture. *Virology* 2010; **7**: 40 [PMID: 20167059 DOI: 10.1186/1743-422X-7-40]
  - 21 **Simonsen L, Kane A, Lloyd J, Zaffran M, Kane M.** Unsafe injections in the developing world and transmission of bloodborne pathogens: a review. *Bull World Health Organ* 1999; **77**: 789-800 [PMID: 10593026]
  - 22 **Moftah FM.** Regionalization of the blood transfusion service in Egypt. *Vox Sang* 2002; **83** Suppl 1: 197-199 [PMID: 12617136 DOI: 10.1111/j.1423-0410.2002.tb05300.x]
  - 23 **el-Ghazzawi E, Drew L, Hamdy L, El-Sherbini E, Sadek Sel-D, Saleh E.** Intravenous drug addicts: a high risk group for infection with human immunodeficiency virus, hepatitis viruses, cytomegalovirus and bacterial infections in Alexandria Egypt. *J Egypt Public Health Assoc* 1995; **70**: 127-150 [PMID: 17214204]
  - 24 **Shepard CW, Finelli L, Alter MJ.** Global epidemiology of hepatitis C virus infection. *Lancet Infect Dis* 2005; **5**: 558-567 [PMID: 16122679 DOI: 10.1016/S1473-3099(05)70216-4]
  - 25 **Bruggmann P, Berg T, Øvrehus AL, Moreno C, Brandão Mello CE, Roudot-Thoraval F, Marinho RT, Sherman M, Ryder SD, Sperl J, Akarca U, Balik I, Bihl F, Bilodeau M, Blasco AJ, Buti M, Calinas F, Calleja JL, Cheinquer H, Christensen PB, Clausen M, Coelho HS, Cornberg M, Cramp ME, Dore GJ, Doss W, Duberg AS, El-Sayed MH, Ergör G, Esmat G, Estes C, Falconer K, Félix J, Ferraz ML, Ferreira PR, Frankova S, García-Samaniego J, Gerstoft J, Gira JA, Gonçalves FL, Gower E, Gschwantler M, Guimarães Pessôa M, Hézode C, Hofer H, Husa P, Idilman R, Kåberg M, Kaita KD, Kautz A, Kaymakoglu S, Krajdin M, Krarup H, Laleman W, Lavanchy D, Lázaro P, Marotta P, Mauss S, Mendes Correa MC, Müllhaupt B, Myers RP, Negro F, Nemecek V, Örmeci N, Parkes J, Peltekian KM, Ramji A, Razavi H, Reis N, Roberts SK, Rosenberg WM, Sarmiento-Castro R, Sarrazin C, Semela D, Shiha GE, Sievert W, Stärkel P, Stauber RE, Thompson AJ, Urbanek P, van Thiel I, Van Vlierberghe H, Vandijck D, Vogel W, Waked I, Wedemeyer H, Weis N, Wiegand J, Yosry A, Zekry A, Van Damme P, Aleman S, Hindman SJ.** Historical epidemiology of hepatitis C virus (HCV) in selected countries. *J Viral Hepat* 2014; **21** Suppl 1: 5-33 [PMID: 24713004 DOI: 10.1111/jvh.12247]
  - 26 **Hassan AA, Khalil R.** Hepatitis C in dialysis patients in Egypt: relationship to dialysis duration, blood transfusion, and liver disease. *Saudi J Kidney Dis Transpl* 2000; **11**: 72-73 [PMID: 18209303]
  - 27 **El Sayed NM, Gomatos PJ, Beck-Sagué CM, Dietrich U, von Briesen H, Osmanov S, Esparza J, Arthur RR, Wahdan MH, Jarvis WR.** Epidemic transmission of human immunodeficiency virus in renal dialysis centers in Egypt. *J Infect Dis* 2000; **181**: 91-97 [PMID: 10608755]
  - 28 **Hassan NF, el Ghorab NM, Abdel Rehim MS, el Sharkawy MS, el Sayed NM, Emara K, Soltant Y, Sanad M, Hibbs RG, Arthur RR.** HIV infection in renal dialysis patients in Egypt. *AIDS* 1994; **8**: 853 [PMID: 8086148 DOI: 10.1097/00002030-199406000-00023]
  - 29 **Vandelli C, Renzo F, Romano L, Tisminetzky S, De Palma M, Stroffolini T, Ventura E, Zanetti A.** Lack of evidence of sexual transmission of hepatitis C among monogamous couples: results of a 10-year prospective follow-up study. *Am J Gastroenterol* 2004; **99**: 855-859 [PMID: 15128350 DOI: 10.1111/j.1572-0241.2004.04150.x]
  - 30 **Magder LS, Fix AD, Mikhail NN, Mohamed MK, Abdel-Hamid M, Abdel-Aziz F, Medhat A, Strickland GT.** Estimation of the risk of transmission of hepatitis C between spouses in Egypt based on seroprevalence data. *Int J Epidemiol* 2005; **34**: 160-165 [PMID: 15647312 DOI: 10.1093/ije/dyh370]
  - 31 **Mohamed MK, Abdel-Hamid M, Mikhail NN, Abdel-Aziz F, Medhat A, Magder LS, Fix AD, Strickland GT.** Intrafamilial transmission of hepatitis C in Egypt. *Hepatology* 2005; **42**: 683-687 [PMID: 16032698 DOI: 10.1002/hep.20811]
  - 32 **Mohamed MK, Magder LS, Abdel-Hamid M, El-Daly M, Mikhail NN, Abdel-Aziz F, Medhat A, Thiers V, Strickland GT.** Transmission of hepatitis C virus between parents and children. *Am J Trop Med Hyg* 2006; **75**: 16-20 [PMID: 16837701]
  - 33 **Ismail NA, Aboul Ftouh AM, El Shoubary WH.** Safe injection practice among health care workers, Gharbiya, Egypt. *J Egypt Public Health Assoc* 2005; **80**: 563-583 [PMID: 17187743]
  - 34 **Ismail NA, Aboul Ftouh AM, El-Shoubary WH, Mahaba H.** Safe injection practice among health-care workers in Gharbiya Governorate, Egypt. *East Mediterr Health J* 2007; **13**: 893-906 [PMID: 17955773]
  - 35 **Talaat M, Kandeel A, El-Shoubary W, Bodenschatz C, Khairy I, Oun S, Mahoney FJ.** Occupational exposure to needlestick injuries and hepatitis B vaccination coverage among health care workers in Egypt. *Am J Infect Control* 2003; **31**: 469-474 [PMID: 14647109 DOI: 10.1016/j.ajic.2003.03.003]
  - 36 **Washer P.** Hepatitis C: transmission, treatment and occupational risk. *Nurs Stand* 2001; **15**: 43-46 [PMID: 12206075 DOI: 10.7748/ns2001.06.15.40.43.c3046]
  - 37 **Benova L, Awad SF, Miller FD, Abu-Raddad LJ.** Estimation of hepatitis C virus infections resulting from vertical transmission in Egypt. *Hepatology* 2015; **61**: 834-842 [PMID: 25366418 DOI: 10.1002/hep.27596]
  - 38 **Arafa N, El Hoseiny M, Rekecewicz C, Bakr I, El-Kafrawy S, El Daly M, Aoun S, Marzouk D, Mohamed MK, Fontanet A.** Changing pattern of hepatitis C virus spread in rural areas of Egypt. *J Hepatol* 2005; **43**: 418-424 [PMID: 16019104 DOI: 10.1016/j.jhep.2005.03.021]
  - 39 **El Gaafary MM, Rekecewicz C, Abdel-Rahman AG, Allam MF, El Hosseiny M, Hamid MA, Colombani F, Sultan Y, El-Aidy S, Fontanet A, Mohamed MK.** Surveillance of acute hepatitis C in Cairo, Egypt. *J Med Virol* 2005; **76**: 520-525 [PMID: 15977225 DOI: 10.1002/jmv.20392]
  - 40 **El-Raziky MS, El-Hawary M, Esmat G, Abouzied AM, El-Koofy N, Mohsen N, Mansour S, Shaheen A, Abdel Hamid M, El-Karakasy H.** Prevalence and risk factors of asymptomatic hepatitis C virus infection in Egyptian children. *World J Gastroenterol* 2007; **13**: 1828-1832 [PMID: 17465475 DOI: 10.3748/wjg.v13.i12.1828]
  - 41 **el-Sadawy M, Ragab H, el-Touky H, el-Mor Ael-L, Mangoud AM, Eissa MH, Afefy AF, el-Shorbagy E, Ibrahim IA, Mahrous S, Abdel-Monem A, Sabee EI, Ismail A, Morsy TA, Etewa S, Nor**

- Edin E, Mostafa Y, Abouel-Magd Y, Hassan MI, Lakouz K, Abdel-Aziz K, el-Hady G, Saber M. Hepatitis C virus infection at Sharkia Governorate, Egypt: seroprevalence and associated risk factors. *J Egypt Soc Parasitol* 2004; **34**: 367-384 [PMID: 15124747]
- 42 **El-Zanaty F**, Way A. Egypt Demographic and Health Survey 2008. Egyptian: Ministry of Health. Cairo: El-Zanaty and Associates, and Macro International, 2009: 431
- 43 **Habib M**, Mohamed MK, Abdel-Aziz F, Magder LS, Abdel-Hamid M, Gamil F, Madkour S, Mikhail NN, Anwar W, Strickland GT, Fix AD, Sallam I. Hepatitis C virus infection in a community in the Nile Delta: risk factors for seropositivity. *Hepatology* 2001; **33**: 248-253 [PMID: 11124843 DOI: 10.1053/jhep.2001.20797]
- 44 **Medhat A**, Shehata M, Magder LS, Mikhail N, Abdel-Baki L, Nafeh M, Abdel-Hamid M, Strickland GT, Fix AD. Hepatitis c in a community in Upper Egypt: risk factors for infection. *Am J Trop Med Hyg* 2002; **66**: 633-638 [PMID: 12201604]
- 45 **Saleh DA**, Shebl F, Abdel-Hamid M, Narooz S, Mikhail N, El-Batanony M, El-Kafrawy S, El-Daly M, Sharaf S, Hashem M, El-Kamary S, Magder LS, Stoszek SK, Strickland GT. Incidence and risk factors for hepatitis C infection in a cohort of women in rural Egypt. *Trans R Soc Trop Med Hyg* 2008; **102**: 921-928 [PMID: 18514243 DOI: 10.1016/j.trstmh.2008.04.011]
- 46 **Saleh DA**, Shebl FM, El-Kamary SS, Magder LS, Allam A, Abdel-Hamid M, Mikhail N, Hashem M, Sharaf S, Stoszek SK, Strickland GT. Incidence and risk factors for community-acquired hepatitis C infection from birth to 5 years of age in rural Egyptian children. *Trans R Soc Trop Med Hyg* 2010; **104**: 357-363 [PMID: 20153495 DOI: 10.1016/j.trstmh.2010.01.009]
- 47 **Farghaly AG**, Barakat RM. Prevalence, impact and risk factors of hepatitis C infection. *J Egypt Public Health Assoc* 1993; **68**: 63-79 [PMID: 7504049]
- 48 **Kandeel AM**, Talaat M, Afifi SA, El-Sayed NM, Abdel Fadeel MA, Hajjeh RA, Mahoney FJ. Case control study to identify risk factors for acute hepatitis C virus infection in Egypt. *BMC Infect Dis* 2012; **12**: 294 [PMID: 23145873 DOI: 10.1186/1471-2334-12-294]
- 49 **Cuadros DF**, Branscum AJ, Miller FD, Abu-Raddad LJ. Spatial epidemiology of hepatitis C virus infection in Egypt: analyses and implications. *Hepatology* 2014; **60**: 1150-1159 [PMID: 24913187 DOI: 10.1002/hep.27248]
- 50 **Chemaitelly H**, Abu-Raddad LJ, Miller FD. An apparent lack of epidemiologic association between hepatitis C virus knowledge and the prevalence of hepatitis C infection in a national survey in Egypt. *PLoS One* 2013; **8**: e69803 [PMID: 23922806 DOI: 10.1371/journal.pone.0069803]
- 51 **Mostafa A**, Taylor SM, el-Daly M, el-Hoseiny M, Bakr I, Arafat N, Thiers V, Rimlinger F, Abdel-Hamid M, Fontanet A, Mohamed MK. Is the hepatitis C virus epidemic over in Egypt? Incidence and risk factors of new hepatitis C virus infections. *Liver Int* 2010; **30**: 560-566 [PMID: 20141592 DOI: 10.1111/j.1478-3231.2009.02204.x]
- 52 **Paez Jimenez A**, Sharaf Eldin N, Rimlinger F, El-Daly M, El-Hariri H, El-Hoseiny M, Mohsen A, Mostafa A, Delarocque-Astagneau E, Abdel-Hamid M, Fontanet A, Mohamed MK, Thiers V. HCV iatrogenic and intrafamilial transmission in Greater Cairo, Egypt. *Gut* 2010; **59**: 1554-1560 [PMID: 20947889 DOI: 10.1136/gut.2009.194266]
- 53 **Breban R**, Arafat N, Leroy S, Mostafa A, Bakr I, Tondeur L, Abdel-Hamid M, Doss W, Esmat G, Mohamed MK, Fontanet A. Effect of preventive and curative interventions on hepatitis C virus transmission in Egypt (ANRS 1211): a modelling study. *Lancet Glob Health* 2014; **2**: e541-e549 [PMID: 25304421 DOI: 10.1016/S2214-109X(14)70188-3]
- 54 **Breban R**, Fontanet A. Feasible HCV targets in Egypt - authors' reply. *Lancet Glob Health* 2014; **2**: e688 [PMID: 25433621 DOI: 10.1016/S2214-109X(14)70327-4]
- 55 **Drain PK**, Nelson CM, Lloyd JS. Single-dose versus multi-dose vaccine vials for immunization programmes in developing countries. *Bull World Health Organ* 2003; **81**: 726-731 [PMID: 14758432]
- 56 **El Katsha S**, Labeeb S, Watts S, Younis A. Informal health providers and the transmission of hepatitis C virus: pilot study in two Egyptian villages. *East Mediterr Health J* 2006; **12**: 758-767 [PMID: 17333820]
- 57 **Hutin YJ**, Hauri AM, Armstrong GL. Use of injections in healthcare settings worldwide, 2000: literature review and regional estimates. *BMJ* 2003; **327**: 1075 [PMID: 14604927 DOI: 10.1136/bmj.327.7423.1075]
- 58 **Kane A**, Lloyd J, Zaffran M, Simonsen L, Kane M. Transmission of hepatitis B, hepatitis C and human immunodeficiency viruses through unsafe injections in the developing world: model-based regional estimates. *Bull World Health Organ* 1999; **77**: 801-807 [PMID: 10593027]
- 59 **Mast EE**, Alter MJ, Margolis HS. Strategies to prevent and control hepatitis B and C virus infections: a global perspective. *Vaccine* 1999; **17**: 1730-1733 [PMID: 10194830 DOI: 10.1016/S0264-410X(98)00415-0]
- 60 **Talaat M**, el-Oun S, Kandeel A, Abu-Rabeii W, Bodenschatz C, Lohiniva AL, Hallaj Z, Mahoney FJ. Overview of injection practices in two governorates in Egypt. *Trop Med Int Health* 2003; **8**: 234-241 [PMID: 12631314 DOI: 10.1046/j.1365-3156.2003.01015.x]
- 61 **Garbin CA**, de Souza NP, de Vasconcelos RR, Garbin AJ, Villar LM. Hepatitis C virus and dental health workers: an update. *Oral Health Prev Dent* 2014; **12**: 313-321 [PMID: 24914431 DOI: 10.3290/j.ohpd.a32134]
- 62 **Klevens RM**, Moorman AC. Hepatitis C virus: an overview for dental health care providers. *J Am Dent Assoc* 2013; **144**: 1340-1347 [PMID: 24282263 DOI: 10.14219/jada.archive.2013.0069]
- 63 **Mahboobi N**, Porter SR, Karayiannis P, Alavian SM. Dental treatment as a risk factor for hepatitis B and C viral infection. A review of the recent literature. *J Gastrointest Liver Dis* 2013; **22**: 79-86 [PMID: 23539395]
- 64 **Hashish MH**, Selim HS, Elshazly SA, Diab HH, Elsayed NM. Screening for the hepatitis C virus in some dental clinics in Alexandria, Egypt. *J Egypt Public Health Assoc* 2012; **87**: 109-115 [PMID: 23196884 DOI: 10.1097/01.EPX.0000421670.02166.ec]
- 65 **El-Zanaty F**, Population MoHa, Macro O. Egypt Service Provision Assessment Survey 2004. Calverton, Maryland, USA: Ministry of Health and Population and ORC Macro, 2005
- 66 **El-Zanaty F**, Macro O; Ministry of Health and Population (MOHP) (Egypt). Egypt Service Provision Assessment Survey 2002: Calverton, Maryland: Ministry of Health and Population, El-Zanaty Associates, and ORC Macro. 2003
- 67 **Eassa S**, Eissa M, Sharaf SM, Ibrahim MH, Hassanein OM. Prevalence of hepatitis C virus infection and evaluation of a health education program in el-ghar village in zagazig, egypt. *J Egypt Public Health Assoc* 2007; **82**: 379-404 [PMID: 18706295]
- 68 **Shalaby S**, Kabbash IA, El Saleet G, Mansour N, Omar A, El Nawawy A. Hepatitis B and C viral infection: prevalence, knowledge, attitude and practice among barbers and clients in Gharbia governorate, Egypt. *East Mediterr Health J* 2010; **16**: 10-17 [PMID: 20214151]
- 69 **Kleinbaum D**, Kupper L, Morgenstern H. Epidemiologic Research: Principles and Quantitative Methods. New York, NY: John Wiley and Sons, 1982

**P- Reviewer:** De Paschale M, McQuillan GM, Yokota S  
**S- Editor:** Ji FF **L- Editor:** A **E- Editor:** Liu SQ





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

