

World Journal of *Hepatology*

World J Hepatol 2019 December 27; 11(12): 752-779



ORIGINAL ARTICLE**Retrospective Study**

- 752 Post-infantile giant cell hepatitis: A single center's experience over 25 years
Matta B, Cabello R, Rabinovitz M, Minervini M, Malik S

Observational Study

- 761 Lumacaftor/ivacaftor therapy is associated with reduced hepatic steatosis in cystic fibrosis patients
Kutney K, Donnola SB, Flask CA, Gubitosi-Klug R, O'Riordan M, McBennett K, Sferra TJ, Kaminski B

CASE REPORT

- 773 Intrahepatic and intra-abdominal splenosis: A case report and review of literature
Ananthan K, Yusuf GT, Kumar M

ABOUT COVER

Editorial Board Member of *World Journal of Hepatology*, Chiara Raggi, PhD, Research Scientist, Liver Unit and Center for Autoimmune Liver Diseases, Humanitas Clinical and Research Center, Rozzano (MI) 20089, Italy

AIMS AND SCOPE

The primary aim of *World Journal of Hepatology* (*WJH*, *World J Hepatol*) is to provide scholars and readers from various fields of hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJH mainly publishes articles reporting research results and findings obtained in the field of hepatology and covering a wide range of topics including chronic cholestatic liver diseases, cirrhosis and its complications, clinical alcoholic liver disease, drug induced liver disease autoimmune, fatty liver disease, genetic and pediatric liver diseases, hepatocellular carcinoma, hepatic stellate cells and fibrosis, liver immunology, liver regeneration, hepatic surgery, liver transplantation, biliary tract pathophysiology, non-invasive markers of liver fibrosis, viral hepatitis.

INDEXING/ABSTRACTING

The *WJH* is now abstracted and indexed in PubMed, PubMed Central, Emerging Sources Citation Index (Web of Science), Scopus, China National Knowledge Infrastructure (CNKI), China Science and Technology Journal Database (CSTJ), and Superstar Journals Database.

RESPONSIBLE EDITORS FOR THIS ISSUE

Responsible Electronic Editor: *Mei-Yi Liu*

Proofing Production Department Director: *Xiang Li*

NAME OF JOURNAL

World Journal of Hepatology

ISSN

ISSN 1948-5182 (online)

LAUNCH DATE

October 31, 2009

FREQUENCY

Monthly

EDITORS-IN-CHIEF

Ke-Qin Hu, Koo Jeong Kang, Nikolaos Pyrsopoulos

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/1948-5182/editorialboard.htm>

EDITORIAL OFFICE

Ruo-Yu Ma, Director

PUBLICATION DATE

December 27, 2019

COPYRIGHT

© 2019 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>

Retrospective Study

Post-infantile giant cell hepatitis: A single center's experience over 25 years

Bassem Matta, Ricardo Cabello, Mordechai Rabinovitz, Marta Minervini, Shahid Malik

ORCID number: Bassem Matta (0000-0003-2744-2928); Ricardo Cabello (0000-0002-2049-623X); Mordechai Rabinovitz (0000-0003-2340--8781); Marta Minervini (0000-0003-4404-6708); Shahid Malik (0000-0001-9646-0260).

Author contributions: Matta B and Malik SM designed the research study, obtained and analyzed data and wrote manuscript; Cabello R, Rabinovitz M and Minervini M contributed with data interpretation and critical review of manuscript for intellectual content; All authors have read and approve the final manuscript.

Institutional review board

statement: This study was reviewed and approved by the University of Pittsburgh Medical Center Institution Review Board PRO12030073.

Informed consent statement:

Patients were not required to give informed consent to the study because the analysis used anonymous data that were obtained after each patient agreed to treatment by written consent.

Conflict-of-interest statement:

There were no conflict of interests to disclose.

Data sharing statement: All data requests should be submitted to the corresponding author for consideration. Access to anonymized data may be granted following review.

Open-Access: This article is an open-access article which was selected by an in-house editor and

Bassem Matta, Mordechai Rabinovitz, Department of Medicine, Division of Gastroenterology, Hepatology and Nutrition, University of Pittsburgh, Pittsburgh, PA 15213, United States

Ricardo Cabello, Department of Internal Medicine, University of Pittsburgh Medical Center, Pittsburgh, PA 15213, United States

Marta Minervini, Department of Pathology, University of Pittsburgh, Pittsburgh, PA 15213, United States

Shahid Malik, Department of Medicine, University of Pittsburgh, Division of Gastroenterology Hepatology and Nutrition, Pittsburgh, PA 15213, United States

Corresponding author: Shahid Malik, MD, Department of Medicine, University of Pittsburgh, Division of Gastroenterology, Hepatology and Nutrition, 3471 5th Ave 916 Lillian Kaufman Building, Pittsburgh, PA 15213, United States. maliks@upmc.edu

Telephone: +1-412-6474932

Fax: +1-412-6479268

Abstract**BACKGROUND**

Giant cell hepatitis in the adult population remains very poorly defined with only 100 case reports published in the literature over the last three decades.

AIM

To present our center's experience in an attempt to learn about the predisposing factors, outcomes and efficacy of proposed therapeutic interventions for giant cell hepatitis.

METHODS

A retrospective chart review was conducted through the electronic records of the University of Pittsburgh Medical Center. We queried 36726 liver biopsy reports from January 1, 1991 to December 6, 2016. Our search yielded 50 patients who were identified as carrying a definite diagnosis of post-infantile giant cell hepatitis (PIGCH) by pathology. The data collected included demographic information, laboratory data (liver function tests, autoimmune markers) and transplant status. In order to better analyze patient characteristics and outcomes, subjects were separated into a non-transplant (native) liver group and a post-liver transplant (allograft) group.

RESULTS

The incidence of PIGCH was approximately 0.14% of all biopsies queried in the

fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Received: May 15, 2019

Peer-review started: May 17, 2019

First decision: July 4, 2019

Revised: September 9, 2019

Accepted: October 18, 2019

Article in press: October 18, 2019

Published online: December 27, 2019

P-Reviewer: Mrzljak A, Pop TL, Rubbini M

S-Editor: Ma YJ

L-Editor: Filipodia

E-Editor: Liu MY



25-year period. The mean age was 48 years with 66% females. Liver function tests were classified as 38.2% cholestatic, 35.3% hepatocellular and 26.5% mixed. Autoimmune hepatitis was found to be the most prevalent predisposing factor leading to PIGCH constituting 32% of cases. Management consisted mainly of immunosuppression, viral targeted therapy, supportive care and in six cases liver transplantations.

CONCLUSION

The diagnosis of PIGCH remains clinically challenging and requires a high index of suspicion as well as a thorough history, physical examination, serological workup and liver biopsy. Treatment of the underlying cause can result in clinical stability in a large number of cases.

Key words: Post-infantile giant cell hepatitis; Liver transplantation; Autoimmune hepatitis

©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Post-infantile giant cell hepatitis is a rare disorder and very poorly defined in the literature. Our study aimed to present our center's experience in an attempt to shed more light about the predisposing factors, outcomes and efficacy of proposed therapeutic interventions for post-infantile giant cell hepatitis.

Citation: Matta B, Cabello R, Rabinovitz M, Minervini M, Malik S. Post-infantile giant cell hepatitis: A single center's experience over 25 years. *World J Hepatol* 2019; 11(12): 752-760

URL: <https://www.wjgnet.com/1948-5182/full/v11/i12/752.htm>

DOI: <https://dx.doi.org/10.4254/wjh.v11.i12.752>

INTRODUCTION

Giant cell hepatitis (GCH) is a relatively common histologic finding in neonates. It is believed to occur secondary to insults to immature hepatocytes. In children, it typically presents with cholestasis, conjugated hyperbilirubinemia and variable degrees of inflammation^[1]. Idiopathic GCH refers to these histologic findings with a structurally intact biliary system as opposed to conditions where biliary abnormalities are present, such as biliary atresia^[1]. The most commonly proposed pathophysiological hypothesis to account for the presence of giant cells includes an ineffective cytoplasmic division in the setting of cellular fission (endomitosis) in contrast to cellular hepatocyte fusion secondary to hepatic injury^[2].

As common as GCH is in children, it is exceedingly rare in adults. GCH in the adult population remains very poorly defined with only 100 case reports published in the literature over the last three decades^[3]. In adults the entity is referred to as post-infantile giant cell hepatitis (PIGCH), also known as syncytial or adult onset GCH. PIGCH represents a histologic diagnosis that has been associated with a myriad of medical conditions including infectious, hematologic, autoimmune disorders and drug reactions (Table 1)^[3-13]. Pathological analysis is characterized by the presence of giant multinucleated syncytial hepatocytes. In particular, more than four to five nuclei in hepatocytes should be seen in a single lobule combined with other features of hepatitis such as lobular disarray, inflammation, Kupffer cell hypertrophy and spotty hepatocytes necrosis (Figure 1).

The clinical course of patients with giant cells on histology is widely variable, ranging from minimal symptoms without major clinical implications to acute liver failure that is often times fatal despite standard clinical care. In the current study, we aimed to present our center's experience with this very rare disease entity in an attempt to shed more light about its predisposing factors, outcomes and efficacy of proposed therapeutic interventions.

MATERIALS AND METHODS

After obtaining local institutional review board approval, we queried liver biopsy reports (36726) at the University of Pittsburgh Medical Center electronic records using

Table 1 Reported causes of post-infantile giant cell hepatitis

Infectious	Hepatitis A, B, C Epstein-Barr virus (EBV) Cytomegalovirus (CMV) Paramyxo-like virus Human immunodeficiency virus (HIV) Herpesvirus 6A Human papillomavirus (HPV)
Autoimmune	Autoimmune hepatitis (AIH) Ulcerative colitis (UC) Primary sclerosing cholangitis (PSC) Primary biliary cholangitis (PBC) Systemic lupus erythematosus (SLE) Rheumatoid arthritis (RA) Polyarteritis nodosa (PAN)
Drugs	Methotrexate 6 mercaptopurine Amytriptyline P-aminosalicylic acid Vinyl chloride Chropromazine Methotrexate
Hematologic	Chronic lymphocytic leukemia (CLL) Lymphoma Sickle cell disease (SCC) Hypereosinophilia Autoimmune hemolytic anemia
Endocrine	Hypoparathyroidism
Infiltrative	Sarcoidosis
Post-transplant	-
Idiopathic	-

the keywords “giant cell hepatitis” from January 1, 1991 to December 6, 2016. Our search yielded 127 individual patient records, of which 45 were diagnosed prior to 18 years of age. The remaining 82 records were evaluated by three physicians (BM, SM, MM) after which 50 patients were identified as carrying a definite diagnosis of PIGCH based on liver biopsy. In order to better analyze patient characteristics and outcomes, subjects were separated into a non-transplant (native) liver group and a post-liver transplant (allograft) group.

RESULTS

The incidence of PIGCH was approximately 0.14% of all biopsies queried in the 25-year period. The mean age of the studied patient sample was 48 years with 66% females. Liver function tests were classified as follows: 38.2% cholestatic, 35.3% hepatocellular and 26.5% mixed; 73.5% of patients had bilirubin values exceeding 1.5 mg/dL at the time of diagnosis and 42% of patients had bilirubin values exceeding 5 mg/dL. Mean follow up of the entire cohort was over six years (79 mo; SD = 76.1). Patient demographics and liver function tests for patients are outlined in [Table 2](#). Patients with GCH found in the native liver group were older, had higher aspartate aminotransferase, alanine aminotransferase and total bilirubin when compared to the allograft group.

Autoimmune hepatitis (AIH) was found to be the most prevalent predisposing factor leading to PIGCH constituting 32% of cases, while drugs accounted for 12% of cases. Other etiological associations included viral infections [hepatitis A, B, C (HCV), cytomegalovirus (CMV), Epstein-Barr virus], systemic autoimmune conditions (but not enough to give a diagnosis of AIH) and hematologic conditions. In nearly 1/3 of

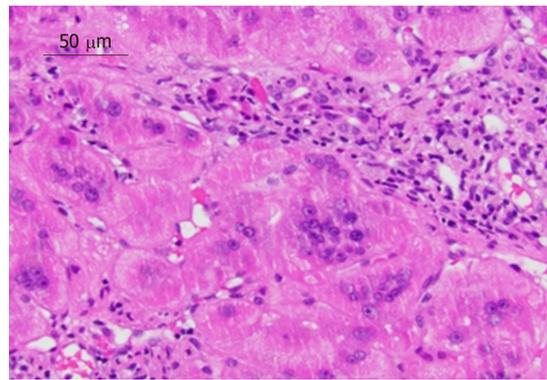


Figure 1 Liver biopsy of 44-year-old female with autoimmune hepatitis (hematoxylin and eosin stain 40 ×). Biopsy revealed chronic hepatitis with prominent giant multinucleated hepatocytes.

cases, no predisposing factor for PIGCH was found (idiopathic). In the post-transplant population, the most prevalent predisposing factor leading to PIGCH was AIH as well, accounting for 30% of cases.

Autoimmune markers related to liver disease were common: Anti-nuclear antibody in 34% of cases, elevated immunoglobulin G in 22% of cases, anti-smooth muscle antibody in 10% of cases, anti-mitochondrial antibody in 8% of cases and anti-liver kidney microsomal antibody in 2% of cases.

Drugs which were identified as the possible culprit for GCH development consisted of microdantin, ranitidine, omeprazole, moxifloxacin, ranitidine, plaquenil as well as chromium picolinate.

Notable pathological findings included diffuse necrosis in 24% of the patients, inflammation and acute hepatitis in 56% of patients and overt cirrhosis in 12% of patients. Of the ten patients with GCH post-liver transplant, five had concomitant features of acute cellular rejection.

Management of PIGCH consisted mainly of immunosuppression, viral targeted therapy, supportive care and in six cases liver transplantation. Management and outcomes are outlined in Table 3. Among the patients who were treated with immunosuppression, eight patients (53%) had improvement in their liver function tests. Of the patients treated with ganciclovir, two patients (100%) had improvement in their liver function tests.

Among the native group, five patients (13%) required liver transplantation, and one patient developed graft failure from post-transplant GCH and required a second transplant. Five (13%) patients died from liver-related complications in the native liver group compared to two (20%) in the allograft group. Among these seven patients, five died with acute liver failure. Patient #1 had received two liver transplants. The first transplant was for HCV cirrhosis and subsequently developed PIGCH in the allograft despite achieving a sustained virologic response after anti-viral therapy. This patient eventually developed allograft cirrhosis attributed to PIGCH and required a second transplant for this reason. The patient died of a spontaneous intracranial hemorrhage. Patient #2 had developed cirrhosis attributed to PIGCH and died of pneumonia and sepsis. The five remaining patients presented with acute liver failure. Patient #3 was urgently transplanted but developed infected necrotizing pancreatitis to which he succumbed. Patient #4 was found to develop a pneumothorax and died from hemothorax after placement of a thoracotomy tube. Patient #5 died after developing subcapsular hepatic bleeding following a liver biopsy. Patients #6 and #7 developed a massive variceal bleed and lower gastrointestinal bleeding (exact cause unknown), respectively, that led to their demise.

Of the 50 patients with GCH, 12 (6 native and 6 allograft) underwent a repeat liver biopsy of which 66% still had evidence of GCH despite treatment. Half of these patients had undergone liver transplantation (AIH, primary sclerosing cholangitis/AIH overlap, HCV, GCH, alcoholic and cryptogenic cirrhosis). These patients had persistent GCH on repeat biopsies despite immunosuppression. The patient with primary sclerosing cholangitis/AIH overlap had improvement of GCH findings on subsequent biopsy. One subject had evidence of acute cellular then chronic rejection on subsequent biopsies. Cirrhosis developed in a patient transplanted for alcoholic cirrhosis and GCH. Among the native liver group, six patients had recurrent GCH on biopsy. One had acute hepatitis B, while the rest did not have a specific predisposing factor.

Table 2 Patient characteristics and liver function tests

	GCH on native liver, n = 40	GCH on allograft, n = 10
Mean age in yr	50.4	43.4
Gender		
Male	14 (35%)	4 (40%)
Female	26 (65%)	6 (60%)
AST	433 ± 486	175 ± 158
ALT	488 ± 537	232 ± 206
Alkaline phosphatase	197 ± 151	296 ± 197
GGT	287 ± 582	246 ± 182
Bilirubin	10.9 ± 10.4	3.1 ± 3.8

GCH: Giant cell hepatitis; AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; GGT: γ -Glutamyl transpeptidase.

DISCUSSION

With only 100 cases reported in the adult literature, PIGCH remains poorly understood. The prevalence of this disease has been reported at 0.1% to 0.25%^[3], which is consistent with the incidence in our cohort (0.14%). Given the rarity of this entity, outcomes and management are largely based on anecdotal evidence. There are no approved therapies and no consensus on management strategies^[13].

The histological finding of giant cells in adults seems to be a manifestation of hepatic stress as opposed to a primary hepatic injury^[3,10]. The diagnosis is made based on the presence of multinucleated giant cells usually evident in zones 1 and 3 of the Rappaport acinus. More than four to five nuclei in hepatocytes should be seen in a single lobule combined with other features of hepatitis such as lobular disarray, acinar inflammation, Kupffer cell hypertrophy and spotty hepatocytes necrosis. Other common features may include non-suppurative cholangitis, ductopenia and different stages of periportal fibrosis leading to cirrhosis^[6,14]. Similar histological findings were observed among our patient cohort: The majority had notable inflammation on pathology, while a quarter of them exhibited evidence of hepatic necrosis (28% spotty necrosis, 48% bridging/confluent necrosis, 19% sub-massive necrosis and 5% massive necrosis), with 12% demonstrating overt cirrhosis, which is comparable to previous reported rates in the literature of about 13%^[3].

Out of the six liver transplant recipients for PICGH, two died with recurrent disease. The first patient died in the early post-transplant period, and the second patient died 11 years later. Two patients required two more liver transplants each for recurrent decompensated cirrhosis despite being on standard immunosuppression. One patient developed cirrhosis with features of chronic rejection, which was thought to be related to recurrent hepatitis C, and another was related to CMV hepatitis.

Scant data exists on PIGCH in the post-transplant setting with prior observations indicating the need for re-transplantation in the majority of recipients due to recurrent disease. Pappo *et al*^[15] examined the clinical and pathologic course of seven patients who developed GCH after liver transplantation. Five of these patients had GCH as their native liver disease. Two patients died. Two patients required re-transplantation because of recurrent GCH. One patient with recurrent GCH was still alive six years after transplantation. Similarly, in our study, ten patients developed GCH after liver transplantation. Two patients had GCH as their native liver disease. One patient died due to sepsis related to a second liver transplantation. Two patients developed recurrent GCH on the allograft; one of those patients had their immunosuppression increased and had survived at two years and the other patient required re-transplantation. Two patients developed *de novo* GCH that required an increase in immunosuppression; one patient eventually needed liver transplantation and the other one improved with medical management. One patient developed *de novo* GCH and CMV hepatitis and was treated with ganciclovir. The remaining four patients were lost to follow up.

Management strategies to treat recurrence mainly consisted of increasing immunosuppression and in rare cases the institution of ribavirin with variable success^[16,17].

Our results were consistent with prior reports indicating a potential autoimmune link to the findings of PIGCH. We concluded that an autoimmune type hepatitis was seen in 1/3 of our patients; 34% of the patients had a positive anti-nuclear antibody,

Table 3 Predisposing factors, n (%)

Predisposing factors	GCH on native liver	GCH on allograft
AIH	13 (32)	3 (30)
Drug induced	6 (15)	0
No factor identified	12 (30)	3 (30)
UC	2 (5)	3 (30)
PSC	3 (7)	1 (10)
HCV	2 (5)	1 (10)
CMV	1 (2)	1 (10)
SLE	2 (5)	0
Lymphoma	2 (5)	0
HAV	1 (2)	0
HBV	1 (2)	0
EBV	1 (2)	0
Sjogren	1 (2)	0
Autoimmune hemolytic anemia	1 (2)	0
CLL	1 (2)	0
Peripheral eosinophilia	1 (2)	0
SCC	1 (2)	0
Celiac disease	1 (2)	0

GCH: Giant cell hepatitis; AIH: Autoimmune hepatitis; UC: Ulcerative colitis; PSC: Primary sclerosing cholangitis; HCV: Hepatitis C virus; CMV: Cytomegalovirus; SLE: Systemic lupus erythematosus; HAV: Hepatitis A virus; HBV: Hepatitis B virus; EBV: Epstein-Barr virus; CLL: Chronic lymphocytic leukemia; SCC: Sickle cell disease.

22% had an elevated immunoglobulin G, while 12 patients would fulfill at least a probable diagnosis of AIH based on the AIH scoring system^[18] (Table 4).

The majority of our patients were female (66%), which is somewhat different to previous reports with approximately equal numbers between genders^[3]. Idiopathic PIGCH was present in 30% of our cohort, which is much higher than prior published studies. A higher incidence of idiopathic PIGCH in our cohort compared to the published literature is likely a manifestation of publication bias, *i.e.* cases of PIGCH where there is no clear link may be less apt to be reported^[3]. Drug induced liver injury was the culprit in 12% of cases with all of the reported drugs being novel associations with PIGCH (Table 4).

Viral causes amongst our cohort seem to have been less frequent than previously reported. Outcomes of those with a viral cause was variable, although the cases where CMV infection was felt to be the culprit did respond well to ganciclovir, similar to cases reported in the literature^[7,19].

The majority of deaths were in the group labeled idiopathic PIGCH, while only two out of sixteen patients with autoimmune like features died. Notably, all of the idiopathic patients were managed supportively while most of the autoimmune cases were managed with immunosuppression. One patient who died had chronic HCV in addition to AIH. HCV therapy (standard of therapy at the time was interferon-based treatment) was not offered given the patient's decompensated state. PIGCH has been described in both acute and chronically infected HCV patients (or co-infected with HIV) with a relatively good prognosis after treatment with interferon and ribavirin or immunosuppressive therapy when autoimmune features are present^[19-22]. No studies have been published to date using the highly potent direct acting antivirals that might potentially prove to have even better outcomes with higher rates of viral eradication^[23].

The presentations and outcomes of our patients coincide with previously reported observations in the literature of being highly variable. Some patients only manifested in mild elevations in liver function tests while others developed acute liver failure resulting in death or the need for liver transplantation (Table 2). Most patients responded well to immunosuppressive therapy that mainly consisted of intravenous hydrocortisone, prednisone, azathioprine and tacrolimus, especially with the presence of autoimmune features. One case (previously published) with PIGCH secondary to AIH complicating ulcerative colitis responded to prednisone with improved liver functions despite worsening ulcerative colitis (the patient ultimately required a

Table 4 Management and outcomes, n (%)

	GCH on native liver, n = 40	GCH on allograft, n = 10
Management		
Immunosuppression	11 (28)	4 (40)
Supportive care	10 (25)	0 (0)
Liver transplantation	5 (13)	1 (10)
Ganciclovir	1 (3)	1 (10)
Unknown	13 (33)	4 (40)
Outcomes		
Survived	25 (63)	4 (40)
Died	5 (13)	2 (20)
Unknown	10 (25)	4 (40)

GCH: Giant cell hepatitis.

colectomy)^[12]. Several cases of PIGCH associated with chronic lymphocytic leukemia have been reported with largely favorable outcomes after being managed with intravenous immunoglobulins (in the events where immunoglobulins are low), rituximab or steroids^[8,13]. This is similar to our patient with chronic lymphocytic leukemia who was managed successfully with prednisone but ultimately developed cirrhosis^[24].

Our study has several limitations. It is based on retrospective chart review and is mainly descriptive. That being said, it includes the largest number of unique cases of PIGCH from a single institution included in a single manuscript.

The exact etiology of PIGCH and mechanism of injury remains unknown, and the histological findings are likely related to an idiosyncratic or cytopathic response to various hepatocyte stimuli. Our series suggested an autoimmune cause as the most common association. The diagnosis of PIGCH remains clinically challenging and requires a high index of suspicion as well as a thorough history, physical examination and serological workup, which should include viral, hematologic and autoimmune causes. Ultimately a liver biopsy is required as PIGCH remains a purely histomorphological diagnosis. Treatment of the underlying cause (especially if it is autoimmune or viral) can result in clinical stability in a large number of cases. Treatment and monitoring should be done in close association with specialty centers including those capable of liver transplantation.

ARTICLE HIGHLIGHTS

Research background

Giant cell hepatitis in the adult population remains very poorly defined with only 100 case reports published in the literature over the last three decades. Pathological analysis is characterized by the presence of giant multinucleated syncytial hepatocytes. The clinical course of patients with giant cells on histology is widely variable, ranging from minimal symptoms without major clinical implications to acute liver failure that is often times fatal despite standard clinical care.

Research objectives

Our primary objective was to present our center's experience in an attempt to learn about the predisposing factors, outcomes and efficacy of proposed therapeutic interventions for giant cell hepatitis.

Research methods

A retrospective chart review was conducted through the electronic records of the University of Pittsburgh Medical Center. We queried 36726 liver biopsy reports from January 1, 1991 to December 6, 2016. Our search yielded 50 patients who were identified as carrying a definite diagnosis of post-infantile giant cell hepatitis (PIGCH) by pathology. The data collected included demographic information, laboratory data (liver function tests, autoimmune markers) and transplant status. In order to better analyze patient characteristics and outcomes, subjects were separated into a non-transplant (native) liver group and a post-liver transplant (allograft) group.

Research results

The incidence of PIGCH was approximately 0.14% of all biopsies queried in the 25-year period. The mean age was 48 years with 66% females. Liver function tests were classified as 38.2% cholestatic, 35.3% hepatocellular and 26.5% mixed. Autoimmune hepatitis was found to be the

most prevalent predisposing factor leading to PIGCH constituting 32% of cases. Management consisted mainly of immunosuppression, viral targeted therapy, supportive care and in six cases liver transplantation.

Research conclusions

The diagnosis of PIGCH remains clinically challenging and requires a high index of suspicion as well as a thorough history, physical examination, serological workup and liver biopsy. Treatment of the underlying cause can result in clinical stability in a large number of cases.

Research perspectives

This study reports our center's experience with PIGCH and the importance of thorough history, physical examination, serologic work up and liver biopsy in its diagnosis. Further research should aim at recognizing risk factors for progression from PIGCH to liver failure and further evaluation of therapeutic interventions (immunosuppression *vs* viral targeted therapy *vs* liver transplantation).

REFERENCES

- 1 **Torbenson M**, Hart J, Westerhoff M, Azzam RK, Elgendi A, Mziray-Andrew HC, Kim GE, Scheimann A. Neonatal giant cell hepatitis: histological and etiological findings. *Am J Surg Pathol* 2010; **34**: 1498-1503 [PMID: 20871223 DOI: 10.1097/PAS.0b013e3181f069ab]
- 2 **Correa KK**, Nanjundiah P, Wirtschafter DD, Alshak NS. Idiopathic neonatal giant cell hepatitis presenting with acute hepatic failure on postnatal day one. *J Perinatol* 2002; **22**: 249-251 [PMID: 11948391 DOI: 10.1038/sj.jp.7210670]
- 3 **Bihari C**, Rastogi A, Sarin SK. Postinfantile giant cell hepatitis: an etiological and prognostic perspective. *Hepat Res Treat* 2013; **2013**: 601290 [PMID: 23555054 DOI: 10.1155/2013/601290]
- 4 **Kinra P**, John BM. Hepatitis - A induced Non Infantile Giant Cell Hepatitis. *Med J Armed Forces India* 2007; **63**: 182-183 [PMID: 27407983 DOI: 10.1016/S0377-1237(07)80073-6]
- 5 **Johnson SJ**, Mathew J, MacSween RN, Bennett MK, Burt AD. Post-infantile giant cell hepatitis: histological and immunohistochemical study. *J Clin Pathol* 1994; **47**: 1022-1027 [PMID: 7829677 DOI: 10.1136/jcp.47.11.1022]
- 6 **Protzer U**, Dienes HP, Bianchi L, Lohse AW, Helmreich-Becker I, Gerken G, Meyer zum Büschenfelde KH. Post-infantile giant cell hepatitis in patients with primary sclerosing cholangitis and autoimmune hepatitis. *Liver* 1996; **16**: 274-282 [PMID: 8878001 DOI: 10.1111/j.1600-0676.1996.tb00743.x]
- 7 **Falasca L**, Nonno FD, Palmieri F, Licordari R, Iannicelli G, Antonucci G, Baiocchi A. Two cases of giant cell hepatitis in HIV-infected patients. *Int J STD AIDS* 2012; **23**: e3-e4 [PMID: 22844018 DOI: 10.1258/ijsa.2009.009407]
- 8 **Gupta E**, Yacoub M, Higgins M, Al-Katib AM. Syncytial giant cell hepatitis associated with chronic lymphocytic leukemia: a case report. *BMC Blood Disord* 2012; **12**: 8 [PMID: 22812631 DOI: 10.1186/1471-2326-12-8]
- 9 **Bouguila J**, Mabrouk S, Tilouche S, Bakir D, Trabelsi A, Hmila A, Boughammoura L. Giant cell hepatitis with autoimmune hemolytic anemia in a nine month old infant. *World J Hepatol* 2013; **5**: 226-229 [PMID: 23671728 DOI: 10.4254/wjh.v5.i4.226]
- 10 **Estradas J**, Pascual-Ramos V, Martínez B, Uribe M, Torre A. Autoimmune hepatitis with giant-cell transformation. *Ann Hepatol* 2009; **8**: 68-70 [PMID: 19221538 DOI: 10.1152/ajpgi.90298.2008]
- 11 **Moreno-Otero R**, Trapero-Marugán M, García-Buey L, García-Sánchez A. Drug-induced postinfantile giant cell hepatitis. *Hepatology* 2010; **52**: 2245-2246 [PMID: 20890941 DOI: 10.1002/hep.23830]
- 12 **Labowitz J**, Finklestein S, Rabinovitz M. Postinfantile giant cell hepatitis complicating ulcerative colitis: a case report and review of the literature. *Am J Gastroenterol* 2001; **96**: 1274-1277 [PMID: 11316183 DOI: 10.1111/j.1572-0241.2001.03711.x]
- 13 **Gupta N**, Njei B. Syncytial giant cell hepatitis in a patient with chronic lymphocytic leukemia. *J Dig Dis* 2015; **16**: 683-688 [PMID: 26147671 DOI: 10.1111/1751-2980.12273]
- 14 **Devaney K**, Goodman ZD, Ishak KG. Postinfantile giant-cell transformation in hepatitis. *Hepatology* 1992; **16**: 327-333 [PMID: 1639341 DOI: 10.1002/hep.1840160208]
- 15 **Pappo O**, Yunis E, Jordan JA, Jaffe R, Mateo R, Fung J, Demetris AJ. Recurrent and de novo giant cell hepatitis after orthotopic liver transplantation. *Am J Surg Pathol* 1994; **18**: 804-813 [PMID: 8037295 DOI: 10.1097/00000478-19940800-00007]
- 16 **Lerut JP**, Claeys N, Ciccarelli O, Pisa R, Galant C, Laterre PF, Palazzo U. Recurrent postinfantile syncytial giant cell hepatitis after orthotopic liver transplantation. *Transpl Int* 1998; **11**: 320-322 [PMID: 9704401 DOI: 10.1007/s001470050151]
- 17 **Durand F**, Degott C, Sauvanet A, Molas G, Sicot C, Marcellin P, Belghiti J, Erlinger S, Benhamou JP, Bernuau J. Subfulminant syncytial giant cell hepatitis: recurrence after liver transplantation treated with ribavirin. *J Hepatol* 1997; **26**: 722-726 [PMID: 9075682 DOI: 10.1016/s0168-8278(97)80440-0]
- 18 **Hennes EM**, Zeniya M, Czaja AJ, Parés A, Dalekos GN, Krawitt EL, Bittencourt PL, Porta G, Boberg KM, Hofer H, Bianchi FB, Shibata M, Schramm C, Eisenmann de Torres B, Galle PR, McFarlane I, Dienes HP, Lohse AW; International Autoimmune Hepatitis Group. Simplified criteria for the diagnosis of autoimmune hepatitis. *Hepatology* 2008; **48**: 169-176 [PMID: 18537184 DOI: 10.1002/hep.22322]
- 19 **Gábor L**, Pál K, Zsuzsa S. Giant cell hepatitis in adults. *Pathol Oncol Res* 1997; **3**: 215-218 [PMID: 18470733 DOI: 10.1007/BF02899924]
- 20 **Moreno A**, Moreno A, Pérez-Eliás MJ, Quereda C, Fernández-Muñoz R, Antela A, Moreno L, Bárcena R, López-San Román A, Celma ML, García-Martos M, Moreno S. Syncytial giant cell hepatitis in human immunodeficiency virus-infected patients with chronic hepatitis C: 2 cases and review of the literature. *Hum Pathol* 2006; **37**: 1344-1349 [PMID: 16949926 DOI: 10.1016/j.humpath.2006.05.003]
- 21 **Micchelli ST**, Thomas D, Boitnott JK, Torbenson M. Hepatic giant cells in hepatitis C virus (HCV) mono-infection and HCV/HIV co-infection. *J Clin Pathol* 2008; **61**: 1058-1061 [PMID: 18682418 DOI: 10.1136/jcp.2008.058560]
- 22 **Tajiri K**, Shimizu Y, Tokimitsu Y, Tsuneyama K, Sugiyama T. An elderly man with syncytial giant cell hepatitis successfully treated by immunosuppressants. *Intern Med* 2012; **51**: 2141-2144 [PMID: 22892492]

- DOI: [10.2169/internalmedicine.51.7870](https://doi.org/10.2169/internalmedicine.51.7870)]
- 23 **Zhang J**, Nguyen D, Hu KQ. Chronic Hepatitis C Virus Infection: A Review of Current Direct-Acting Antiviral Treatment Strategies. *N Am J Med Sci (Boston)* 2016; **9**: 47-54 [PMID: [27293521](https://pubmed.ncbi.nlm.nih.gov/27293521/)]
- 24 **Horsmans Y**, Galant C, Nicholas ML, Lamy M, Geubel AP. Failure of ribavirin or immunosuppressive therapy to alter the course of post-infantile giant-cell hepatitis. *J Hepatol* 1995; **22**: 382 [PMID: [7608496](https://pubmed.ncbi.nlm.nih.gov/7608496/) DOI: [10.1016/0168-8278\(95\)80298-3](https://doi.org/10.1016/0168-8278(95)80298-3)]

Observational Study

Lumacaftor/ivacaftor therapy is associated with reduced hepatic steatosis in cystic fibrosis patients

Katherine Kutney, Shannon B Donnola, Chris A Flask, Rose Gubitosi-Klug, MaryAnn O'Riordan, Kimberly McBennett, Thomas J Sferra, Beth Kaminski

ORCID number: Katherine Kutney (0000-0002-2573-2131); Shannon B Donnola (000-0002-7489-3011); Chris A Flask (0000-0002-1066-333X); Rose Gubitosi-Klug (0000-0002-7376-6870); MaryAnn O'Riordan (0000-0001-6713-8918); Kimberly McBennett (0000-0001-9642-3814); Thomas J Sferra (0000-0001-6893-9880); Beth Kaminski (0000-0003-1516-7455).

Author contributions: Kutney K designed the study, coordinated the research and wrote the manuscript; Donnola SB acquired and analyzed MRI images; Flask CA analyzed MRI images and edited the manuscript; Gubitosi-Klug R designed the study and edited the manuscript; O'Riordan M performed statistical analysis; McBennett K designed the study; Sferra TJ designed the study; Kaminski B designed the study and edited the manuscript.

Supported by a grant from the University Hospitals Fellowship Research Award Program (FRAP).

Institutional review board

statement: This study was approved by the University Hospitals Cleveland Medical Center Institutional Review Board.

Informed consent statement: All participants gave written informed consent prior to participating in this study.

Conflict-of-interest statement:

Flask CA and Kaminski B reports grants from Cystic Fibrosis Foundation, during the conduct of

Katherine Kutney, Rose Gubitosi-Klug, MaryAnn O'Riordan, Kimberly McBennett, Thomas J Sferra, Beth Kaminski, Department of Pediatric Endocrinology, University Hospitals Cleveland Medical Center, Cleveland, OH 44106, United States

Katherine Kutney, Chris A Flask, Rose Gubitosi-Klug, MaryAnn O'Riordan, Kimberly McBennett, Thomas J Sferra, Beth Kaminski, Department of Pediatrics, Case Western Reserve University, Cleveland, OH 44106, United States

Shannon B Donnola, Chris A Flask, Department of Radiology Case Western Reserve University, Cleveland, OH 44106, United States

Chris A Flask, Department of Biomedical Engineering, Case Western Reserve University, Cleveland, OH 44106, United States

Corresponding author: Katherine Kutney, MD, Assistant Professor, Department of Pediatric Endocrinology, University Hospitals Cleveland Medical Center, 11100 Euclid Ave., Suite 737, Cleveland, OH 44106, United States. katherine.kutney@uhhospitals.org

Telephone: +1-216-8443661

Fax: +1-216-8448900

Abstract**BACKGROUND**

Hepatic steatosis is a common form of cystic fibrosis associated liver disease (CFLD) seen in an estimated 15%-60% of patients with cystic fibrosis (CF). The pathophysiology and health implications of hepatic steatosis in cystic fibrosis remain largely unknown. In the general population, hepatic steatosis is strongly associated with insulin resistance and type 2 diabetes. Cystic fibrosis related diabetes (CFRD) impacts 40%-50% of CF adults and is characterized by both insulin insufficiency and insulin resistance. We hypothesized that patients with CFRD would have higher levels of hepatic steatosis than cystic fibrosis patients without diabetes.

AIM

To determine whether CFRD is associated with hepatic steatosis and to explore the impact of lumacaftor/ivacaftor therapy on hepatic steatosis in CF.

METHODS

Thirty patients with CF were recruited from a tertiary care medical center for this cross-sectional study. Only pancreatic insufficient patients with CFRD or normal glucose tolerance (NGT) were included. Patients with established CFLD, end

the study. The other authors declare that they have no conflict of interest.

STROBE statement: The authors have read the STROBE statement-checklist of items and the manuscript was prepared and revised according to the strobe guidelines.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Received: September 4, 2019

Peer-review started: September 4, 2019

First decision: October 14, 2019

Revised: October 28, 2019

Accepted: November 25, 2019

Article in press: November 25, 2019

Published online: December 27, 2019

P-Reviewer: Lucarelli M, Rowland M, Tjora E

S-Editor: Ma RY

L-Editor: A

E-Editor: Liu MY



stage lung disease, or persistently elevated liver enzymes were excluded. Mean magnetic resonance imaging (MRI) proton density fat fraction (PDFF) was obtained for all participants. Clinical characteristics [age, sex, body mass index, percent predicted forced expiratory volume at 1 s (FEV1), lumacaftor/ivacaftor use] and blood chemistries were assessed for possible association with hepatic steatosis. Hepatic steatosis was defined as a mean MRI PDFF > 5%. Patients were grouped by diabetes status (CFRD, NGT) and cystic fibrosis transmembrane conductance regulator (CFTR) modulator use (lumacaftor/ivacaftor, no lumacaftor/ivacaftor) to determine between group differences. Continuous variables were analyzed with a Wilcoxon rank sum test and discrete variables with a Chi square test or Fisher's exact test.

RESULTS

Twenty subjects were included in the final analysis. The median age was 22.3 years (11.3-39.0) and median FEV1 was 77% (33%-105%). Twelve subjects had CFRD and 8 had NGT. Nine subjects were receiving lumacaftor/ivacaftor. The median PDFF was 3.0% (0.0%-21.0%). Six subjects (30%) had hepatic steatosis defined as PDFF > 5%. Hepatic fat fraction was significantly lower in patients receiving lumacaftor/ivacaftor (median, range) (2.0%, 0.0%-6.4%) than in patients not receiving lumacaftor/ivacaftor (4.1%, 2.7-21.0%), $P = 0.002$. Though patients with CFRD had lower PDFF (2.2%, 0.0%-14.5%) than patients with NGT (4.9%, 2.4-21.0%) this did not reach statistical significance, $P = 0.06$. No other clinical characteristic was strongly associated with hepatic steatosis.

CONCLUSION

Use of the CFTR modulator lumacaftor/ivacaftor was associated with significantly lower hepatic steatosis. No association between CFRD and hepatic steatosis was found in this cohort.

Key words: Cystic fibrosis; Liver disease; Non-alcoholic fatty liver disease; Cystic fibrosis transmembrane conductance regulator; Lumacaftor/ivacaftor; Cystic fibrosis transmembrane conductance regulator modulator; Diabetes mellitus

©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Hepatic steatosis is a common manifestation of liver disease in cystic fibrosis (CF). It remains unknown whether hepatic steatosis contributes to the development of cirrhosis in patients with CF. Lumacaftor/ivacaftor is a cystic fibrosis transmembrane conductance regulator (CFTR) modulator drug targeting the defective chloride channel that causes CF. In this cross-sectional study, CF patients receiving lumacaftor/ivacaftor had significantly lower magnetic resonance imaging proton density fat fractions than CF patients not receiving the CFTR modulator. CFTR modulator use should be included in future studies of CF liver disease.

Citation: Kutney K, Donnola SB, Flask CA, Gubitosi-Klug R, O'Riordan M, McBennett K, Sferra TJ, Kaminski B. Lumacaftor/ivacaftor therapy is associated with reduced hepatic steatosis in cystic fibrosis patients. *World J Hepatol* 2019; 11(12): 761-772

URL: <https://www.wjgnet.com/1948-5182/full/v11/i12/761.htm>

DOI: <https://dx.doi.org/10.4254/wjh.v11.i12.761>

INTRODUCTION

The life expectancy for cystic fibrosis (CF) patients has improved dramatically over the past several decades, and continued improvement is expected with the widespread use of cystic fibrosis transmembrane conductance regulator (CFTR) modulator therapies^[1]. While pulmonary disease remains the leading cause of mortality in CF, extra pulmonary complications such as cystic fibrosis related diabetes (CFRD) and cystic fibrosis associated liver disease (CFLD) have emerged as important sources of morbidity in this population^[2-4]. As the life expectancy for CF patients improves, determining the impact of CFTR modulator therapy on extra-pulmonary

disease is of critical importance.

Hepatic manifestations of CF are broad, including: Neonatal cholestasis, transaminase elevation, hepatic steatosis, focal biliary cirrhosis, multilobular cirrhosis and portal hypertension^[3,5]. Cirrhosis with portal hypertension is the primary cause of morbidity and mortality from CFLD^[6]. Debate regarding the optimal diagnostic criteria for CFLD is ongoing^[6-9]. While older studies describe CFLD as a childhood-onset disease, recent data demonstrates that adult-onset CFLD is relatively common^[8,10,11]. Multilobular cirrhosis with portal hypertension is the end stage manifestation of CFLD and is the third leading cause of death in CF patients^[1,6].

Perhaps the most common manifestation of CFLD is hepatic steatosis, with prevalence estimates ranging from 15%-60%^[7,12,13]. Historically, hepatic steatosis in CF patients was attributed to malnutrition and considered a benign finding that did not increase risk for hepatic cirrhosis^[14]. Outside of CF, hepatic steatosis is strongly associated with obesity and type 2 diabetes^[15]. Hepatic steatosis can progress to non-alcoholic steatohepatitis and cirrhosis, which is now a common indication for liver transplantation among the general population in the United States^[16]. Little is known about the clinical implications of hepatic steatosis in CF and its relationship to other forms of CFLD^[12,17,18].

CFRD is another common extrapulmonary manifestation of CF, with a prevalence of approximately 20% in adolescents and 40%-50% in adults^[19]. CFRD is distinct from type 1 diabetes, which is characterized by absolute insulin deficiency, and type 2 diabetes in which peripheral insulin resistance predominates. CFRD is primarily a disease of insulin insufficiency, though insulin resistance occurs during illness and with increasing age^[20,21]. Patients with CFRD typically have lower body mass index (BMI), reduced pulmonary function and higher mortality rates. These effects are at least partially mitigated by insulin therapy^[22]. The prevalence of CFRD is also higher in CF patients with liver disease^[23].

Both CFRD and CFLD are almost exclusively seen in patients carrying two pathogenic CFTR variants that severely limit the chloride channel function^[24,25]. CFTR variants are generally categorized into five (or six) groups according to the underlying cause of channel malfunction. Class 1-3 variants result in little or no CFTR function while class 4-6 variants are characterized by residual CFTR function^[26]. Individualized CF therapy relies on understanding the functional defect causing CFTR malfunction^[27]. CFTR modulator therapies are a revolutionary class of small molecules targeting the underlying defect in CF^[26]. Ivacaftor is a CFTR potentiator that increases chloride conductance only if CFTR is present in the cell membrane^[28]. Lumacaftor and tezacaftor are correctors which redirect misfolded CFTR protein to the cell surface^[29,30]. Lumacaftor/ivacaftor combination therapy was approved in 2015 for patients carrying two copies of the F508del (p.Phe508del, c.1521_1523delCTT) pathogenic variant.

While pulmonary effects of CFTR modulators have been meticulously examined, the extrapulmonary effects are not well characterized^[28,30-33]. Two small studies demonstrated improved insulin secretion after modulator therapy; while two other studies failed to show improvement^[34-37]. No studies have systematically examined the impact of CFTR modulator therapy on hepatic steatosis or other liver disease in patients with CF. Thus, we sought to determine the impact of CFRD on hepatic steatosis in CF patients and explore other factors associated with elevated hepatic fat, including CFTR modulator use.

MATERIALS AND METHODS

Patient characteristics

All studies were conducted according to the approved Institutional Review Board protocols at University Hospitals Cleveland Medical Center between January 1 and December 31, 2017. Thirty subjects with CF were recruited from the LeRoy W. Matthews Cystic Fibrosis Center at University Hospitals Cleveland Medical Center/Rainbow Babies and Children's Hospital in Cleveland, OH, United States (Table 1 and Supplemental Table 1). Eligible subjects were identified using the local CF database and were approached for study involvement during routine clinic visits or during hospitalization for a CF pulmonary exacerbation. All subjects were diagnosed with cystic fibrosis based on sweat chloride and genetic testing according to established guidelines^[38]. Electronic medical records were reviewed to confirm eligibility. Inclusion criteria were age 10-40 years, pancreatic insufficiency and either normal glucose tolerance (NGT) or CFRD^[39]. Pancreatic insufficiency was defined by a clinical need for pancreatic enzyme replacement. No fecal elastase testing was performed as part of this study. Subjects with established CFLD, persistent transaminase elevation

for greater than one year, low baseline lung function (*i.e.*, FEV₁ < 30% predicted) or an ongoing pulmonary exacerbation were excluded. Only subjects with two copies of a class 1-3 pathogenic CFTR variant were included in the final analysis. CFTR variants were classified into classes 1-5 using the CFTR 2 database and existing guidelines for functional classification^[40,41]. Subjects with contraindications to MRI scanning (*i.e.*, metal implants, pregnancy) were also excluded. Informed consent was obtained in person prior to commencing study activities.

Clinical and laboratory evaluation

Clinical characteristics including BMI, percent predicted FEV₁, diabetes status, insulin use and CFTR modulator use were collected from the electronic medical record. Fasting blood chemistries including lipids, hepatic function tests and hemoglobin A1C were assessed during a period of baseline health in ambulatory subjects or after completing treatment for a pulmonary exacerbation in hospitalized subjects. An oral glucose tolerance test (OGTT) was performed in NGT subjects who had not had an OGTT in the past six months. Glucose tolerance testing was performed according to standard guidelines. After an eight hour fast, subjects ingested 1.75 g/kg (maximum 75 g) of glucose dissolved in water. Plasma glucose was evaluated at baseline and 2 h post glucose ingestion^[39]. Subjects whose study OGTT demonstrated impaired fasting glucose or impaired glucose tolerance as defined by standard criteria were excluded^[42].

All serum chemistries were collected from peripheral venous samples according to standard technique^[43]. Glucose, triglycerides, high density lipoprotein (HDL), low density lipoprotein (LDL), aspartate transaminase (AST), alanine transaminase (ALT), gamma-glutamyltransferase (GGT), alkaline phosphatase, and total bilirubin were analyzed using a Beckman AU 5800® analyzer. Hemoglobin A1C was analyzed using a BioRad-D-100® analyzer (University Hospitals Core Lab, Cleveland, OH, United States).

Hepatic fat fraction measurement

Proton density fat fraction (PDFF) was measured on a Siemens Skyra 3T magnetic resonance imaging (MRI) in the Imaging Research Core at Case Western Reserve University. Briefly, each subject was positioned supine within the MRI scanner. Spine and body array coils were used to obtain uniform images over the entire liver. A single-breathhold VIBE MRI acquisition was used to obtain axial liver PDFF maps for each subject (spatial resolution = 2 mm × 2 mm × 5 mm, 6 echoes). This MRI method also incorporates T2* correction to limit the effects of iron deposition and hepatic fibrosis^[44]. All images were exported for offline analysis in Matlab (The Mathworks, Natick, MA, United States). Mean liver PDFF was determined for the central 6-8 imaging slices in each subject using a region of interest (ROI) analysis. The mean liver PDFF in each slice was then averaged over all slices to calculate the overall mean liver PDFF for each subject.

Data and statistical considerations

For this study, we considered a PDFF > 5% to be consistent with clinical hepatic steatosis^[45]. We grouped subjects by the presence of hepatic steatosis > 5%, diabetes status, and CFTR modulator use to evaluate for significant associations. BMI percentiles were calculated for all subjects to account for age-related variation in BMI. Alkaline phosphatase measurements were standardized by subtracting the age and sex specific mean and dividing by the respective standard deviation. The resulting values were in units of standard deviation. Continuous variables were described with medians and ranges and nominal variables with frequencies and percent. Continuous variables were analyzed with a Wilcoxon rank sum test and nominal variables were analyzed using Chi square test or Fisher's exact test. Statistical analysis was performed using SAS software version 9.4 (SAS Institute, Cary, NC, United States). The level of significance was set at 0.05. The statistical analyses were performed by MaryAnn O'Riordan PhD, biomedical statistician.

RESULTS

Participant characteristics

We recruited 30 participants of whom 20 completed the study (Figure 1). Exclusions related to progression to impaired glucose tolerance on OGTT (*n* = 3), inability to perform breath hold for MRI (*n* = 2), failure to schedule or complete the MRI (*n* = 4) and presence of a class 4 CFTR pathogenic variant (*n* = 1). Participant characteristics are summarized in Table 1. The study population was primarily Caucasian, which is consistent with the demographic of the CF population overall. Eighty percent of the

Table 1 Demographics for all subjects and stratified by modulator (lumacaftor/ivacaftor) use

	All subjects (n = 20)	CFTR modulator (n = 9)	No CFTR modulator (n = 11)	P value
Age at MRI (yr)	22.3 (11.3-39.0)	26.4 (16.3-39.0)	21.9 (11.3-36.1)	0.29
Genotype				
F508del/F508del	10	9	1	< 0.01
F508del/other	9	0	9	< 0.01
Other/other	1	0	1	< 0.01
Male sex	16 (80%)	7 (78%)	9 (82%)	1.00
BMI percentile	39 (2-96)	51 (3-77)	23 (2-96)	0.21
% predicted FEV1	77 (33-105)	73 (33-89)	77 (48-105)	0.24
CFRD	12 (60%)	7 (78%)	5 (45%)	0.20
Insulin therapy	10 (50%)	6 (67%)	4 (36%)	0.37

Data presented as median (range) or frequency (percent) as appropriate. BMI (percentile): body mass index percentile adjusted for age; FEV1: forced expiratory volume at 1 s; CFRD: cystic fibrosis related diabetes.

study subjects were male. Median subject age at the time of MRI was 22.3 years with a range from 11.3 to 39.0 years. All participants had two severe class 1-3 CFTR pathogenic variants. Twelve subjects (60%) had CFRD and 8 subjects (40%) had NGT. Ten CFRD subjects (83%) were prescribed insulin therapy. Nine subjects (45%) had received the CFTR modulator lumacaftor/ivacaftor (Orkambi®) for more than 12 mo at the time of MRI. No subject received lumacaftor/ivacaftor for fewer than 12 mo.

Hepatic steatosis

The median hepatic fat fraction for all subjects was 3.0% with a range from 0.0%-21.0%. Six subjects (30%) had hepatic steatosis, defined as PDFFF > 5%. Subjects with hepatic steatosis showed a trend toward younger age that did not reach statistical significance. Alkaline phosphatase and age-adjusted alkaline phosphatase (z-score) were higher in subjects with hepatic steatosis, $P = 0.01$ and $P = 0.03$. LDL and HDL were both higher in patients with hepatic steatosis, $P = 0.05$ and $P = 0.02$. Total bilirubin, AST, ALT and GGT did not differ significantly between subjects with and without hepatic steatosis (Table 2). Missing data for alkaline phosphatase, AST, ALT, total bilirubin (1 missing), LDL, HDL, triglyceride (4 missing), and GGT (3 missing) were excluded from all analyses.

CFTR modulator

Hepatic fat fraction was significantly lower in the 9 subjects receiving CFTR modulator therapy (2.0%, 0.0%-6.4%) than in the 11 subjects not receiving CFTR modulators (4.1%, 2.7%-21.0%), $P = 0.002$ (Figure 2). Two CFRD subjects receiving CFTR modulators had exceptionally low hepatic fat fractions of 0.0%. Subjects receiving CFTR modulator therapy were not significantly different in terms of age, BMI percentile or diabetes status from subjects not receiving modulators. Absolute BMI was higher in the CFTR modulator group, which likely reflects expected age-related change in BMI, $P = 0.05$ (Table 3).

CFTR modulator use was associated with lower total bilirubin than no CFTR modulator use, $P = 0.003$. Alkaline phosphatase levels were also lower in the CFTR modulator group, but this difference may relate to younger age in the no CFTR modulator group, $P = 0.01$. Although age-adjusted alkaline phosphatase (z scores) were numerically lower in the CFTR modulator group than the no CFTR modulator group, this did not reach statistical significance, $P = 0.07$ (Table 3).

Diabetes

The median hepatic fat fraction was not statistically different between subjects with CFRD (median, range) (2.2%, 0.0-14.5%) and NGT (4.9%, 2.4-21.0%), $P = 0.06$. Subjects with CFRD were older (28.2 years, 17.0-39.0) than NGT subjects (18.0 years, 11.3-30.6), $P = 0.04$. As expected, the older CFRD cohort demonstrated a higher BMI, $P = 0.05$, but not BMI percentile, than NGT subjects. Patients with CFRD demonstrated lower percent predicted FEV1, which is known to be associated with CFRD, $P = 0.001$. Subjects with CFRD also demonstrated higher hemoglobin A1C levels, $P = 0.002$. Alkaline Phosphatase was lower in the CFRD group compared to the NGT group, $P = 0.04$; however, age adjusted alkaline phosphatase (z-score) was not different between groups (Table 2).

Importantly, CFTR modulator use was more common among patients with CFRD



Figure 1 Consort diagram. Consort diagram for the study. Modulator refers to lumacaftor/ivacaftor use. CF: Cystic fibrosis, MRI: Magnetic resonance imaging; CFTR: Cystic fibrosis transmembrane conductance regulator.

(7 of 12, 58%) than patients with NGT (2 of 8, 22%). Because we demonstrated that CFTR modulator use is associated with lower hepatic fat, we repeated the fat fraction analysis by diabetes status after excluding subjects receiving CFTR modulator therapy. The median hepatic fat fraction for the 5 CFRD subjects not receiving CFTR modulator was 4.1% (range 2.8%-14.5%) and for the 6 NGT subject not receiving CFTR modulators was 4.9% (range 2.7%-21.0%), which were not significantly different, $P = 0.92$.

DISCUSSION

In this cross-sectional study of 20 CF patients aged 11-39 years with either NGT or CFRD, we demonstrate a statistically significant association between use of the CFTR modulator lumacaftor/ivacaftor and reduced hepatic fat. CFTR modulator use was also associated with lower total bilirubin and a trend toward lower age-adjusted alkaline phosphatase levels (z-score). Interestingly, CFRD patients on lumacaftor/ivacaftor demonstrated particularly low hepatic fat fractions (0.0% in two cases), suggesting a particular sensitivity to modulator effects in patients with CFRD (Supplemental Table 1). CFRD was not found to be associated with increased hepatic steatosis as was originally hypothesized. In contrast, patients with CFRD showed a trend toward lower PDFE, which most likely reflects higher rates of CFTR modulator use in the CFRD group. Given the small number of subjects not receiving lumacaftor/ivacaftor, we cannot exclude a relationship between CFRD and hepatic steatosis in CF based on this study.

The multifactorial pathogenesis of hepatic steatosis has not been fully elucidated. While strongly associated with obesity and insulin resistance in the general population, hepatic steatosis has historically been attributed to nutritional deficiencies in CF patients. In 1999, Lindblad reported an association between hepatic steatosis and linoleic acid deficiency in a cohort of 41 CF patients^[14]. Others have proposed that

Table 2 Summary of data stratified by presence of steatosis, use of modulator (lumacaftor/ivacaftor), and diabetes status

	Normal Range	All subjects (n = 20)	Steatosis (n = 6)	No Steatosis (n = 14)	P value	CFTR modulator (n = 9)	No CFTR modulator (n = 11)	P value	CFRD (n = 12)	NGT (n = 8)	P value
Hepatic Fat Fraction	--	3.0 0.0-21.0	9.5 6.0-21.0	2.4 0-4.1	0.01	2.0 0.0-6.4	4.1 2.7-21	0.002	2.2 0.0-14.5	4.9 2.4-21	0.06
Age	--	22.3 11.3-39.0	16.7 13.4-36.1	26.0 11.3-39.0	0.08	26.4 16.3-39.0	21.9 11.3-36.1	0.29	28.2 17.0-39.0	18.0 11.3-30.6	0.04
Sex (male)	--	16 80%	5 83%	11 78%	1.00	7 78%	9 82%	1.00	8 67%	8 100%	0.12
BMI	--	21.0 16.9-32.4	20.5 16.9-32.4	21.0 18.8-25.7	0.6	22.4 18.8- 25.7	20.0 16.9- 32.4	0.05	21.0 18.8- 32.4	19.6 16.9- 22.4	0.05
BMI percentile	--	39 2-96	50 2-96	30 3-77	0.77	51 3-77	23 2-96	0.21	39 3-96	37.5 2-73	0.62
FEV1 %	--	77 33-105	86 62-105	74.5 33-97	0.22	73 33-89	77 48-105	0.24	63.5 33-105	88 65-97	0.001
CFTR modulator	--	9 45%	1 17%	8 57%	0.16	9 100%	11 100%	--	7 58%	2 25%	0.20
Hemoglobin A1C (%)	< 5.8	5.8 5.2-8.2	5.7 5.4-6.4	6.3 5.2-8.2	0.30	6.3 5.3-8.2	5.7 5.2-7.4	0.21	6.4 5.4-8.2	5.6 5.2-5.7	0.002
AST (U/L)	9-39	23 11-45	27 11-45	23 11-42	0.93	20 11-42	27 11-45	0.39	20 11-42	26 11-45	0.87
ALT (U/L)	10-52	22 10-58	24.5 12-58	22 10-45	0.57	18 10-45	29 12-58	0.19	18 10-58	26 14-47	0.46
GGT (U/L)	5-64	14 9-29	19 14-25	13.5 9-29	0.10	12 9-24	19 10-29	0.07	13.5 9-29	19 11-24	0.30
Alk Phos (U/L)	33-120	110 44-310	172 146-310	90 44-234	0.01	66 44-178	155 103-310	0.01	90 44-178	157 71-310	0.04
Alk Phos SD	-2-2	1.2 -1.5-4.8	3.1 -0.6-4.8	-0.3 -1.5-4.7	0.03	-0.3 -1.5-4.7	1.8 -0.6-4.8	0.07	0.6 -1.5-4.8	1.4 -0.6-3.2	0.65
Total Bili (µmol/L)	0-20.5	6.8 3.4-20.5	8.5 3.4-15.4	5.1 3.4-20.5	0.47	5.1 3.4-6.8	8.6 3.4-20.5	0.003	5.1 3.4-18.8	8.6 5.1-20.5	0.06
Triglyceride (mmol/L)	< 1.7	1.02 0.36-2.55	1.25 0.69-2.55	0.68 0.36-1.54	0.07	0.9 0.64-2.55	1.25 0.36-2.35	0.79	0.68 0.64-2.35	1.28 0.36-2.55	0.15
LDL (mmol/L)	< 3.4	1.7 0.8-2.4	1.4 0.9-1.6	1.7 0.8-2.4	0.05	1.6 0.8-2.2	1.7 0.9-2.4	0.53	1.7 0.9-2.4	1.6 0.8-2.0	0.71
HDL (mmol/L)	> 1.0	1.03 0.53-2.03	0.84 0.53-1.41	1.09 0.91-2.03	0.02	1.04 0.86-2.03	0.98 0.53-1.54	0.31	1.01 0.53-2.03	1.05 0.84-1.53	0.73

Data is stratified by steatosis (MRI proton density fat fraction >5%) or no steatosis (MRI proton density fat fraction <5%), use of CFTR modulator lumacaftor/ivacaftor, and diabetes status. Data are presented as median and range. To convert total bilirubin from µmol/L to mg/dL multiply by 0.0585. To convert triglycerides from mmol/L to mg/dL multiply by 88.5. To convert LDL from mmol/L to mg/dL multiply by 38.7. To convert HDL from mmol/L to mg/dL multiply by 38.7. CFRD: cystic fibrosis related diabetes. NGT: normal glucose tolerance; AST: Aspartate transaminase; ALT: Alanine transaminase; GGT: Gamma-glutamyltransferase; LDL: Low density lipoprotein; HDL: High density lipoprotein; BMI: Body mass index.

carnitine and choline deficiency cause hepatic steatosis in CF^[5,46]. In contrast, more recent data suggests that hepatic steatosis in CF is associated with higher BMI and better lung function^[45]. Importantly, one case report suggests that CFTR dysfunction may be responsible for hepatic steatosis in CF. Hayes *et al*^[47] reported rapid resolution of severe hepatic steatosis in a 17 year old female (F508del/G511D genotype) after initiation of ivacaftor therapy. Our results further support a role for CFTR dysfunction in the pathogenesis of hepatic steatosis in CF.

We have considered possible explanations for our findings. As CFTR is not expressed in hepatocytes, improvements in hepatic steatosis with CFTR modulators must be mediated by CFTR expression in other tissues^[48]. In CF, biliary stasis and impaired enterocyte function contribute to persistent fat malabsorption, even with

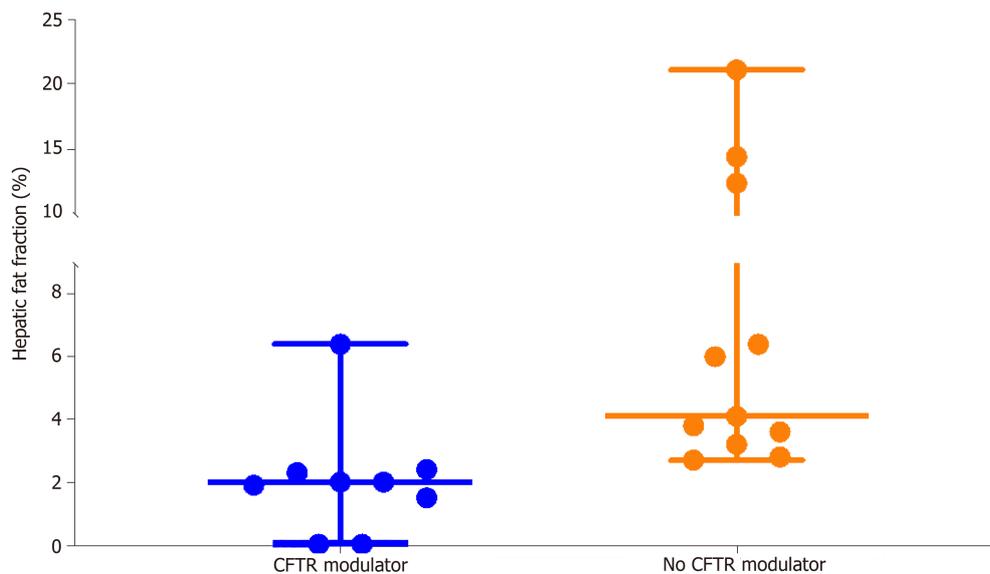


Figure 2 Hepatic steatosis (proton density fat fraction) stratified by lumacaftor/ivacaftor use. Subjects receiving cystic fibrosis transmembrane conductance regulator (CFTR) modulator (lumacaftor/ivacaftor) had a median proton density fat fraction of 2.1%. Subjects not receiving CFTR modulator had a median proton density fat fraction of 4.1%. Each dot represents one subject. Horizontal lines indicate the minimum, median and maximum for each group. $P = 0.002$. CFTR: Cystic fibrosis transmembrane conductance regulator.

adequate pancreatic enzyme replacement^[49,50]. Chronic fat malabsorption can lead to deficiencies in fat soluble nutrients including linoleic acid and choline-which have previously been associated with hepatic steatosis^[46]. Therefore, we theorize that CFTR modulator therapy may lead to resolution of hepatic steatosis by reversing nutritional deficiencies. Further mechanistic studies are needed to test this theory.

It is also possible that lumacaftor/ivacaftor therapy reduces hepatic steatosis through an off target, non-CFTR mediated, mechanism. Additionally, the extremely low hepatic fat seen in subjects receiving lumacaftor/ivacaftor may be secondary to the F508del/F508del genotype itself, rather than the modulator. Although the single F508del/F508del homozygous subject not on lumacaftor/ivacaftor had significant hepatic steatosis, a single observation cannot exclude a genotype effect. Ultimately, longitudinal study is needed to demonstrate that CFTR modulator therapy causes reduced hepatic steatosis and elucidate the mechanism behind this observation.

Prior studies of hepatic steatosis in CF patients have compared varied, qualitative measures of hepatic fat^[13,14]. Ours is the first study to utilize a single, precise, quantitative measure of hepatic fat, the MRI PDFF. Other strengths of our study include the collection of detailed biochemical and clinical information. We acknowledge important limitations. As this study utilized a cross sectional design, we can only demonstrate an association between lumacaftor/ivacaftor therapy and reduced hepatic fat. Moreover, we are unable to exclude the possibility that the F508del/F508del genotype, rather than CFTR modulator use, is associated with reduced hepatic steatosis. Prospective, longitudinal study of modulator therapy in patients expressing different pathogenic CFTR variants will help clarify this question. The relatively small sample size limited our power to detect differences in hepatic fat between CFRD and NGT subjects. This study does not eliminate a possible association between CFRD and hepatic steatosis in CF. Further longitudinal study is needed to understand how hepatic steatosis influences insulin sensitivity and risk for progression to CFRD.

In conclusion, we found no evidence that CFRD is associated with increased hepatic steatosis. We provide strong preliminary data suggesting that lumacaftor/ivacaftor is associated with reduced hepatic steatosis in CF patients. This finding raises many questions about the impact of CFTR modulator therapy on nutrient absorption and on the mechanisms of hepatic steatosis in CF patients. Our study raises the possibility that CFTR modulator therapy may impact other forms of CFLD and adds to the small but growing literature on the extrapulmonary impact of CFTR modulator therapy. CFTR modulator status should be included in future studies of hepatic steatosis or CFLD.

Table 3 Magnetic resonance imaging proton density fat fraction and biochemistry for all subjects and stratified by modulator (lumacaftor/ivacaftor) status

	Reference range	All subjects, n = 20	CFTR modulator, n = 9	No CFTR modulator, n = 11	P value
PDFF (%)	< 5%	3.0 (0.0-21.0)	2.0 (0.0-6.4)	4.1 (2.7-21.0)	0.002
HbA1C (%)	< 5.8%	5.8 (5.2-8.2)	6.3 (5.3-8.2)	5.7 (5.2-7.4)	0.21
Alk Phos (U/L)	¹	110 (44-310)	66 (44-178)	155 (103-310)	0.01
Alk Phos (SD)	-2.0-2.0	1.2 (-1.5-4.8)	-0.3 (-1.5-4.7)	1.8 (-0.6-4.8)	0.07
Total bilirubin (µmol/L) ²	0-20.5	6.8 (3.4-20.5)	5.1 (3.4-6.8)	8.6 (3.4-20.5)	0.003
AST (U/L)	9-39	23 (11-45)	20 (11-42)	27 (11-45)	0.39
ALT (U/L)	10-52	22 (10-58)	18 (10-45)	29 (12-58)	0.19
GGT (U/L)	5-64	14 (9-29)	12 (9-24)	19 (10-29)	0.07
Triglyceride (mmol/L) ³	< 1.7	1.02 (0.36-2.55)	0.9 (0.64-2.55)	1.25 (0.36-2.35)	0.79

¹Reference range is age dependent;

²To convert from µmol/L to mg/dL multiply by 0.0585;

³To convert to mg/dL multiply by 88.5. Results of magnetic resonance imaging proton density fat fraction and key laboratory parameters for all subjects and stratified by cystic fibrosis transmembrane conductance regulator modulator (lumacaftor/ivacaftor) use. Data presented as median (range) or number (percent) as appropriate. CFTR: Cystic fibrosis transmembrane conductance regulator; PDFF: Proton density fat fraction; AST: Aspartate transaminase; ALT: Alanine transaminase; GGT: Gamma-glutamyltransferase.

ARTICLE HIGHLIGHTS

Research background

Hepatic steatosis is a common form of cystic fibrosis associated liver disease (CFLD). The journal has published previous manuscripts regarding CFLD.

Research motivation

Cystic fibrosis (CF) transmembrane conductance regulator (CFTR) modulators are a revolutionary therapy which target the underlying cause of CF for the first time. Currently, very little is known about the impact of CFTR modulator therapy on hepatic disease in CF, despite liver failure being the third leading cause of death in CF patients.

Research objectives

The objectives of this study were therefore to determine whether CF related diabetes (CFRD) is associated with hepatic steatosis and to identify predictors of hepatic steatosis in CF.

Research methods

Patients with established CFLD, end stage lung disease, or persistently elevated liver enzymes were excluded. Mean magnetic resonance imaging (MRI) proton density fat fraction (PDFF) was obtained for all participants. Clinical characteristics and blood chemistries were assessed for possible association with hepatic steatosis. Hepatic steatosis was defined as a mean MRI PDFF > 5%. Patients were grouped by diabetes status and CFTR modulator use (lumacaftor/ivacaftor, no lumacaftor/ivacaftor) to determine between group differences. Continuous variables were analyzed with a Wilcoxon rank sum test and discrete variables with a Chi square test or Fisher's exact test.

Research results

Twelve subjects (60%) had CFRD and 8 subjects (40%) had normal glucose tolerance (NGT). The median hepatic fat fraction for all subjects was 3.0% with a range from 0.0%-21.0%. Six subjects (30%) had hepatic steatosis, defined as PDFF > 5%. Hepatic fat fraction was significantly lower in the 9 subjects receiving CFTR modulator therapy (2.0%, 0.0%-6.4%) than in the 11 subjects not receiving CFTR modulators (4.1%, 2.7%-21.0%), $P = 0.002$. The median hepatic fat fraction was not statistically different between subjects with CFRD (median, range) (2.2%, 0.0-14.5%) and NGT (4.9%, 2.4-21.0%), $P = 0.06$.

Research conclusions

In the enclosed manuscript, we demonstrate that lumacaftor/ivacaftor therapy is associated with reduced hepatic fat in CF patients. While hepatic steatosis has historically been considered a benign finding in CF, the spreading epidemic of liver failure from non-alcoholic steatohepatitis makes this doubtful.

Research perspectives

It suggests a previously unrecognized effect of CFTR modulators of CFLD. CFTR modulator status should be included in future studies of hepatic steatosis or CFLD.

REFERENCES

- 1 **Cystic Fibrosis Foundation.** 2016 Patient Registry Annual Data Report. Available from: <https://www.cff.org/Research/Researcher-Resources/Patient-Registry/2016-Patient-Registry-Annual-Data-Report.pdf>
- 2 **Brennan AL,** Beynon J. Clinical updates in cystic fibrosis-related diabetes. *Semin Respir Crit Care Med* 2015; **36**: 236-250 [PMID: 25826591 DOI: 10.1055/s-0035-1547319]
- 3 **Herrmann U,** Dockter G, Lammert F. Cystic fibrosis-associated liver disease. *Best Pract Res Clin Gastroenterol* 2010; **24**: 585-592 [PMID: 20955961 DOI: 10.1016/j.bpg.2010.08.003]
- 4 **Boëlle PY,** Debray D, Guillot L, Clement A, Corvol H; French CF Modifier Gene Study Investigators. Cystic Fibrosis Liver Disease: Outcomes and Risk Factors in a Large Cohort of French Patients. *Hepatology* 2019; **69**: 1648-1656 [PMID: 30058245 DOI: 10.1002/hep.30148]
- 5 **Colombo C.** Liver disease in cystic fibrosis. *Curr Opin Pulm Med* 2007; **13**: 529-536 [PMID: 17901760 DOI: 10.1097/MCP.0b013e3282f10a16]
- 6 **Flass T,** Narkewicz MR. Cirrhosis and other liver disease in cystic fibrosis. *J Cyst Fibros* 2013; **12**: 116-124 [PMID: 23266093 DOI: 10.1016/j.jcf.2012.11.010]
- 7 **Debray D,** Kelly D, Houwen R, Strandvik B, Colombo C. Best practice guidance for the diagnosis and management of cystic fibrosis-associated liver disease. *J Cyst Fibros* 2011; **10** Suppl 2: S29-S36 [PMID: 21658639 DOI: 10.1016/S1569-1993(11)60006-4]
- 8 **Koh C,** Sakiani S, Surana P, Zhao X, Eccleston J, Kleiner DE, Herion D, Liang TJ, Hoofnagle JH, Chernick M, Heller T. Adult-onset cystic fibrosis liver disease: Diagnosis and characterization of an underappreciated entity. *Hepatology* 2017; **66**: 591-601 [PMID: 28422310 DOI: 10.1002/hep.29217]
- 9 **Debray D,** Narkewicz MR, Bodewes FAJA, Colombo C, Housset C, de Jonge HR, Jonker JW, Kelly DA, Ling SC, Poynard T, Sogni P, Trauner M, Witters P, Baumann U, Wilschanski M, Verkade HJ. Cystic Fibrosis-related Liver Disease: Research Challenges and Future Perspectives. *J Pediatr Gastroenterol Nutr* 2017; **65**: 443-448 [PMID: 28753176 DOI: 10.1097/MPG.0000000000001676]
- 10 **Wilschanski M,** Rivlin J, Cohen S, Augarten A, Blau H, Aviram M, Bentur L, Springer C, Vila Y, Branski D, Kerem B, Kerem E. Clinical and genetic risk factors for cystic fibrosis-related liver disease. *Pediatrics* 1999; **103**: 52-57 [PMID: 9917439 DOI: 10.1542/peds.103.1.52]
- 11 **Terlizzi V,** Lucarelli M, Salvatore D, Angioni A, Bisogno A, Braggion C, Buzzetti R, Carnovale V, Casciaro R, Castaldo G, Cirilli N, Collura M, Colombo C, Di Lullo AM, Elce A, Lucidi V, Madarena E, Padoan R, Quattrucci S, Raia V, Seia M, Termini L, Zarrilli F. Clinical expression of cystic fibrosis in a large cohort of Italian siblings. *BMC Pulm Med* 2018; **18**: 196 [PMID: 30577776 DOI: 10.1186/s12890-018-0766-6]
- 12 **Bader RM,** Jonas MM, Mitchell PD, Wiggins S, Lee CK. Controlled attenuation parameter: A measure of hepatic steatosis in patients with cystic fibrosis. *J Cyst Fibros* 2019; **18**: 280-285 [PMID: 30509601 DOI: 10.1016/j.jcf.2018.11.004]
- 13 **Ayoub F,** Trillo-Alvarez C, Morelli G, Lascano J. Risk factors for hepatic steatosis in adults with cystic fibrosis: Similarities to non-alcoholic fatty liver disease. *World J Hepatol* 2018; **10**: 34-40 [PMID: 29399276 DOI: 10.4254/wjh.v10.i1.34]
- 14 **Lindblad A,** Glaumann H, Strandvik B. Natural history of liver disease in cystic fibrosis. *Hepatology* 1999; **30**: 1151-1158 [PMID: 10534335 DOI: 10.1002/hep.510300527]
- 15 **Saponaro C,** Gaggini M, Gastaldelli A. Nonalcoholic fatty liver disease and type 2 diabetes: common pathophysiologic mechanisms. *Curr Diab Rep* 2015; **15**: 607 [PMID: 25894944 DOI: 10.1007/s11892-015-0607-4]
- 16 **Agopian VG,** Kaldas FM, Hong JC, Whittaker M, Holt C, Rana A, Zarrinpar A, Petrowsky H, Farmer D, Yersiz H, Xia V, Hiatt JR, Busuttill RW. Liver transplantation for nonalcoholic steatohepatitis: the new epidemic. *Ann Surg* 2012; **256**: 624-633 [PMID: 22964732 DOI: 10.1097/SLA.0b013e31826b4b7e]
- 17 **Cortes-Santiago N,** Leung DH, Castro E, Finegold M, Wu H, Patel KR. Hepatic Steatosis Is Prevalent Following Orthotopic Liver Transplantation in Children With Cystic Fibrosis. *J Pediatr Gastroenterol Nutr* 2019; **68**: 96-103 [PMID: 30234762 DOI: 10.1097/MPG.0000000000002154]
- 18 **Haber HP.** Cystic fibrosis in children and young adults: findings on routine abdominal sonography. *AJR Am J Roentgenol* 2007; **189**: 89-99 [PMID: 17579157 DOI: 10.2214/AJR.06.1046]
- 19 **Moran A,** Dunitz J, Nathan B, Saeed A, Holme B, Thomas W. Cystic fibrosis-related diabetes: current trends in prevalence, incidence, and mortality. *Diabetes Care* 2009; **32**: 1626-1631 [PMID: 19542209 DOI: 10.2337/dc09-0586]
- 20 **Battezzati A,** Bedogni G, Zazzeron L, Mari A, Battezzati PM, Alicandro G, Bertoli S, Colombo C. Age- and Sex-Dependent Distribution of OGTT-Related Variables in a Population of Cystic Fibrosis Patients. *J Clin Endocrinol Metab* 2015; **100**: 2963-2971 [PMID: 26057180 DOI: 10.1210/jc.2015-1512]
- 21 **Colomba J,** Boudreau V, Lehoux-Dubois C, Desjardins K, Coriati A, Tremblay F, Rabasa-Lhoret R. The main mechanism associated with progression of glucose intolerance in older patients with cystic fibrosis is insulin resistance and not reduced insulin secretion capacity. *J Cyst Fibros* 2019; **18**: 551-556 [PMID: 30711385 DOI: 10.1016/j.jcf.2019.01.009]
- 22 **Moran A,** Pekow P, Grover P, Zorn M, Slovis B, Pilewski J, Tullis E, Liou TG, Allen H; Cystic Fibrosis Related Diabetes Therapy Study Group. Insulin therapy to improve BMI in cystic fibrosis-related diabetes without fasting hyperglycemia: results of the cystic fibrosis related diabetes therapy trial. *Diabetes Care* 2009; **32**: 1783-1788 [PMID: 19592632 DOI: 10.2337/dc09-0585]
- 23 **Minicucci L,** Lorini R, Giannattasio A, Colombo C, Iapichino L, Reali MF, Padoan R, Calevo MG, Casciaro R, De Alessandri A, Haupt R. Liver disease as risk factor for cystic fibrosis-related diabetes development. *Acta Paediatr* 2007; **96**: 736-739 [PMID: 17381470 DOI: 10.1111/j.1651-2227.2007.00250.x]
- 24 **Adler AI,** Shine BS, Chamnan P, Haworth CS, Bilton D. Genetic determinants and epidemiology of cystic fibrosis-related diabetes: results from a British cohort of children and adults. *Diabetes Care* 2008; **31**: 1789-1794 [PMID: 18535191 DOI: 10.2337/dc08-0466]
- 25 **Ivanov M,** Matsvay A, Glazova O, Krasovskiy S, Usacheva M, Amelina E, Chernyak A, Ivanov M, Musienko S, Prodanov T, Kovalenko S, Baranova A, Khafizov K. Targeted sequencing reveals complex, phenotype-correlated genotypes in cystic fibrosis. *BMC Med Genomics* 2018; **11**: 13 [PMID: 29504914 DOI: 10.1186/s12920-018-0328-z]
- 26 **Elborn JS.** Cystic fibrosis. *Lancet* 2016; **388**: 2519-2531 [PMID: 27140670 DOI: 10.1016/S0140-6736(16)00576-6]
- 27 **Cutting GR.** Cystic fibrosis genetics: from molecular understanding to clinical application. *Nat Rev Genet*

- 2015; **16**: 45-56 [PMID: 25404111 DOI: 10.1038/nrg3849]
- 28 **McKone EF**, Borowitz D, Drevinek P, Griese M, Konstan MW, Wainwright C, Ratjen F, Sermet-Gaudelus I, Plant B, Munck A, Jiang Y, Gilmartin G, Davies JC; VX08-770-105 (PERSIST) Study Group. Long-term safety and efficacy of ivacaftor in patients with cystic fibrosis who have the Gly551Asp-CFTR mutation: a phase 3, open-label extension study (PERSIST). *Lancet Respir Med* 2014; **2**: 902-910 [PMID: 25311995 DOI: 10.1016/S2213-2600(14)70218-8]
- 29 **Wainwright CE**, Elborn JS, Ramsey BW, Marigowda G, Huang X, Cipolli M, Colombo C, Davies JC, De Boeck K, Flume PA, Konstan MW, McColley SA, McCoy K, McKone EF, Munck A, Ratjen F, Rowe SM, Waltz D, Boyle MP; TRAFFIC Study Group; TRANSPORT Study Group. Lumacaftor-Ivacaftor in Patients with Cystic Fibrosis Homozygous for Phe508del CFTR. *N Engl J Med* 2015; **373**: 220-231 [PMID: 25981758 DOI: 10.1056/NEJMoa1409547]
- 30 **Taylor-Cousar JL**, Munck A, McKone EF, van der Ent CK, Moeller A, Simard C, Wang LT, Ingenito EP, McKee C, Lu Y, Lekstrom-Himes J, Elborn JS. Tezacaftor-Ivacaftor in Patients with Cystic Fibrosis Homozygous for Phe508del. *N Engl J Med* 2017; **377**: 2013-2023 [PMID: 29099344 DOI: 10.1056/NEJMoa1709846]
- 31 **Davies JC**, Moskowitz SM, Brown C, Horsley A, Mall MA, McKone EF, Plant BJ, Prais D, Ramsey BW, Taylor-Cousar JL, Tullis E, Uluer A, McKee CM, Robertson S, Shilling RA, Simard C, Van Goor F, Waltz D, Xuan F, Young T, Rowe SM; VX16-659-101 Study Group. VX-659-Tezacaftor-Ivacaftor in Patients with Cystic Fibrosis and One or Two Phe508del Alleles. *N Engl J Med* 2018; **379**: 1599-1611 [PMID: 30334693 DOI: 10.1056/NEJMoa1807119]
- 32 **Keating D**, Marigowda G, Burr L, Daines C, Mall MA, McKone EF, Ramsey BW, Rowe SM, Sass LA, Tullis E, McKee CM, Moskowitz SM, Robertson S, Savage J, Simard C, Van Goor F, Waltz D, Xuan F, Young T, Taylor-Cousar JL; VX16-445-001 Study Group. VX-445-Tezacaftor-Ivacaftor in Patients with Cystic Fibrosis and One or Two Phe508del Alleles. *N Engl J Med* 2018; **379**: 1612-1620 [PMID: 30334692 DOI: 10.1056/NEJMoa1807120]
- 33 **Konstan MW**, McKone EF, Moss RB, Marigowda G, Tian S, Waltz D, Huang X, Lubarsky B, Rubin J, Millar SJ, Pasta DJ, Mayer-Hamblett N, Goss CH, Morgan W, Sawicki GS. Assessment of safety and efficacy of long-term treatment with combination lumacaftor and ivacaftor therapy in patients with cystic fibrosis homozygous for the F508del-CFTR mutation (PROGRESS): a phase 3, extension study. *Lancet Respir Med* 2017; **5**: 107-118 [PMID: 28011037 DOI: 10.1016/S2213-2600(16)30427-1]
- 34 **Bellin MD**, Laguna T, Leschyshyn J, Regelman W, Dunitz J, Billings J, Moran A. Insulin secretion improves in cystic fibrosis following ivacaftor correction of CFTR: a small pilot study. *Pediatr Diabetes* 2013; **14**: 417-421 [PMID: 23952705 DOI: 10.1111/medi.12026]
- 35 **Kelly A**, De Leon DD, Sheikh S, Camburn D, Kubrak C, Peleckis AJ, Stefanovski D, Hadjiliadis D, Rickels MR, Rubenstein RC. Islet Hormone and Incretin Secretion in Cystic Fibrosis after Four Months of Ivacaftor Therapy. *Am J Respir Crit Care Med* 2019; **199**: 342-351 [PMID: 30130412 DOI: 10.1164/rccm.201806-1018OC]
- 36 **Thomassen JC**, Mueller MI, Alejandro Alcazar MA, Rietschel E, van Koningsbruggen-Rietschel S. Effect of Lumacaftor/Ivacaftor on glucose metabolism and insulin secretion in Phe508del homozygous cystic fibrosis patients. *J Cyst Fibros* 2018; **17**: 271-275 [PMID: 29249670 DOI: 10.1016/j.jcf.2017.11.016]
- 37 **Li A**, Vigers T, Pyle L, Zemanick E, Nadeau K, Sagel SD, Chan CL. Continuous glucose monitoring in youth with cystic fibrosis treated with lumacaftor-ivacaftor. *J Cyst Fibros* 2019; **18**: 144-149 [PMID: 30104123 DOI: 10.1016/j.jcf.2018.07.010]
- 38 **Farrell PM**, White TB, Ren CL, Hempstead SE, Accurso F, Derichs N, Howenstine M, McColley SA, Rock M, Rosenfeld M, Sermet-Gaudelus I, Southern KW, Marshall BC, Sosnay PR. Diagnosis of Cystic Fibrosis: Consensus Guidelines from the Cystic Fibrosis Foundation. *J Pediatr* 2017; **181S**: S4-S15.e1 [PMID: 28129811 DOI: 10.1016/j.jpeds.2016.09.064]
- 39 **Moran A**, Brunzell C, Cohen RC, Katz M, Marshall BC, Onady G, Robinson KA, Sabadosa KA, Stecenko A, Slovis B; CFRD Guidelines Committee. Clinical care guidelines for cystic fibrosis-related diabetes: a position statement of the American Diabetes Association and a clinical practice guideline of the Cystic Fibrosis Foundation, endorsed by the Pediatric Endocrine Society. *Diabetes Care* 2010; **33**: 2697-2708 [PMID: 21115772 DOI: 10.2337/dc10-1768]
- 40 **Castellani C**, Cuppens H, Macek M, Cassiman JJ, Kerem E, Durie P, Tullis E, Assael BM, Bombieri C, Brown A, Casals T, Claustres M, Cutting GR, Dequeker E, Dodge J, Doull I, Farrell P, Ferec C, Girodon E, Johannesson M, Kerem B, Knowles M, Munck A, Pignatti PF, Radojkovic D, Rizzotti P, Schwarz M, Stuhmann M, Tzetis M, Zielenski J, Elborn JS. Consensus on the use and interpretation of cystic fibrosis mutation analysis in clinical practice. *J Cyst Fibros* 2008; **7**: 179-196 [PMID: 18456578 DOI: 10.1016/j.jcf.2008.03.009]
- 41 **Castellani C**; CFTR2 team. CFTR2: How will it help care? *Paediatr Respir Rev* 2013; **14** Suppl 1: 2-5 [PMID: 23466340 DOI: 10.1016/j.prrv.2013.01.006]
- 42 **Moran A**, Pillay K, Becker DJ, Acerini CL; International Society for Pediatric and Adolescent Diabetes. ISPAD Clinical Practice Consensus Guidelines 2014. Management of cystic fibrosis-related diabetes in children and adolescents. *Pediatr Diabetes* 2014; **15** Suppl 20: 65-76 [PMID: 25182308 DOI: 10.1111/medi.12178]
- 43 **Clinical and Laboratory Standards Institute/NCCLS**. Procedures for the Collection of Diagnostic Blood Specimens by Venipuncture, 5th Edition 2003. Available from: <https://clsi.org/standards/>
- 44 **Yokoo T**, Shieh-morteza M, Hamilton G, Wolfson T, Schroeder ME, Middleton MS, Bydder M, Gamst AC, Kono Y, Kuo A, Patton HM, Horgan S, Lavine JE, Schwimmer JB, Sirlin CB. Estimation of hepatic proton-density fat fraction by using MR imaging at 3.0 T. *Radiology* 2011; **258**: 749-759 [PMID: 21212366 DOI: 10.1148/radiol.10100659]
- 45 **Chalasani N**, Younossi Z, Lavine JE, Charlton M, Cusi K, Rinella M, Harrison SA, Brunt EM, Sanyal AJ. The diagnosis and management of nonalcoholic fatty liver disease: Practice guidance from the American Association for the Study of Liver Diseases. *Hepatology* 2018; **67**: 328-357 [PMID: 28714183 DOI: 10.1002/hep.29367]
- 46 **Schall JI**, Mascarenhas MR, Maqbool A, Dougherty KA, Elci O, Wang DJ, Altes TA, Hommel KA, Shaw W, Moore J, Stallings VA. Choline Supplementation With a Structured Lipid in Children With Cystic Fibrosis: A Randomized Placebo-Controlled Trial. *J Pediatr Gastroenterol Nutr* 2016; **62**: 618-626 [PMID: 26465792 DOI: 10.1097/MPG.0000000000001004]
- 47 **Hayes D**, Warren PS, McCoy KS, Sheikh SI. Improvement of hepatic steatosis in cystic fibrosis with ivacaftor therapy. *J Pediatr Gastroenterol Nutr* 2015; **60**: 578-579 [PMID: 25688481 DOI: 10.1097/MPG.0000000000000765]

- 48 **Olivier AK**, Gibson-Corley KN, Meyerholz DK. Animal models of gastrointestinal and liver diseases. Animal models of cystic fibrosis: gastrointestinal, pancreatic, and hepatobiliary disease and pathophysiology. *Am J Physiol Gastrointest Liver Physiol* 2015; **308**: G459-G471 [PMID: [25591863](#) DOI: [10.1152/ajpgi.00146.2014](#)]
- 49 **Mailhot G**, Rabasa-Lhoret R, Moreau A, Berthiaume Y, Levy E. CFTR depletion results in changes in fatty acid composition and promotes lipogenesis in intestinal Caco 2/15 cells. *PLoS One* 2010; **5**: e10446 [PMID: [20463919](#) DOI: [10.1371/journal.pone.0010446](#)]
- 50 **Peretti N**, Marcil V, Drouin E, Levy E. Mechanisms of lipid malabsorption in Cystic Fibrosis: the impact of essential fatty acids deficiency. *Nutr Metab (Lond)* 2005; **2**: 11 [PMID: [15869703](#) DOI: [10.1186/1743-7075-2-11](#)]

Intrahepatic and intra-abdominal splenosis: A case report and review of literature

Kiruthika Ananthan, Gibran Timothy Yusuf, Mayur Kumar

ORCID number: Kiruthika Ananthan (0000-0003-0332-904X); Gibran Timothy Yusuf (0000-0002-5621-4249); Mayur Kumar (0000-0003-4920-6918).

Author contributions: Ananthan K, Yusuf GT and Kumar M wrote and reviewed the paper; Yusuf GT additionally selected the images for the case report.

Informed consent statement: Informed written consent was obtained from the patient for publication of this report and accompanying images.

Conflict-of-interest statement: The authors declare that they have no conflict of interest.

CARE Checklist (2016) statement: The authors have read the CARE Checklist (2016) and the manuscript was prepared and revised according to the CARE Checklist (2016).

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited

Kiruthika Ananthan, GKT School of Medical Education, King's College London, London WC2R 2LS, United Kingdom

Gibran Timothy Yusuf, Department of Radiology, Princess Royal University Hospital, King's College Hospital NHS Foundation Trust, Kent BR6 8ND, United Kingdom

Mayur Kumar, Department of Gastroenterology, Princess Royal University Hospital, King's College Hospital NHS Foundation Trust, Kent BR6 8ND, United Kingdom

Corresponding author: Mayur Kumar, FRCP (Hon), MBBS, MRCP, Doctor, Department of Gastroenterology, Princess Royal University Hospital, King's College Hospital NHS Foundation Trust, Farnborough Common, Orpington, Kent BR6 8ND, United Kingdom. mayurkumar@nhs.net

Telephone: +44-203-2995844

Fax: +44-203-2996474

Abstract

BACKGROUND

Splenosis is defined as the process by which tissue from the spleen disseminates through the body and grows in an ectopic location following trauma or a splenectomy. Visceral sites of splenosis are rare.

CASE SUMMARY

We report a case of intrahepatic splenosis in a 57-year-old man with a history of trauma over 40 years ago who initially presented with chest pain. Findings initially mimicked malignancy but a diagnosis of intrahepatic splenosis was confirmed using computed tomography and scintigraphy with technetium-99m heat-denatured red blood cells (Tc-99 DRBC).

CONCLUSION

Scintigraphy with Tc-99 DRBC is a reliable technique to diagnose splenosis and should be performed before using more invasive procedures are carried out. Splenosis should be considered as a possible differential diagnosis for a hepatic nodule in any patient with a history of abdominal trauma, previous splenectomy or atypical radiological features on imaging.

Key words: Intrahepatic splenosis; Abdominal splenosis; Computed tomography; Scintigraphy; Hepatocellular carcinoma; Case report

©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

manuscript

Received: August 22, 2019

Peer-review started: August 22, 2019

First decision: November 2, 2019

Revised: November 11, 2019

Accepted: November 25, 2019

Article in press: November 25, 2019

Published online: December 27, 2019

P-Reviewer: Aoki H, Goral V, Surucu E, Tsoulfas G

S-Editor: Ma RY

L-Editor: A

E-Editor: Liu MY



Core tip: Intrahepatic splenosis is rare. On imaging it is difficult to distinguish splenosis from hepatic malignancy, particularly hepatocellular carcinoma. We report a case of a patient with intrahepatic and intra-abdominal splenosis diagnosed using scintigraphy with technetium-99m heat-denatured red blood cells. To the author's knowledge, this is the first case where hepatic splenosis was confirmed without using invasive procedures such as biopsy or surgery. Splenosis should be considered as an important differential for a hepatic lesion in a patient with a history of trauma or splenectomy, particularly if the lesion is located near the capsule and associated with multiple abdominal deposits.

Citation: Ananthan K, Yusuf GT, Kumar M. Intrahepatic and intra-abdominal splenosis: A case report and review of literature. *World J Hepatol* 2019; 11(12): 773-779

URL: <https://www.wjgnet.com/1948-5182/full/v11/i12/773.htm>

DOI: <https://dx.doi.org/10.4254/wjh.v11.i12.773>

INTRODUCTION

Splenosis is a benign acquired condition. Following trauma or a splenectomy, splenic tissue may autotransplant in an ectopic location. Common sites include the serosal surface of the small or large intestine, greater omentum or the peritoneum^[1]. Less frequently, splenic nodules may be found in the liver^[2], stomach^[3], pancreas^[4] and following rupture of the diaphragm in the thorax^[5]. The kidneys^[6], ovaries^[7] and subcutaneous tissue^[8] are even rarer sites of splenosis. Splenosis is usually asymptomatic and when incidentally discovered can be difficult to distinguish from malignancy using computed tomography (CT) or magnetic resonance imaging (MRI).

CASE PRESENTATION

Chief complaints

A 57-year-old male presented to the Emergency Department with severe right-sided pleuritic chest pain radiating to his back. There was no associated breathlessness, fever, cough, haemoptysis, dizziness, syncope, numbness, paraesthesia or weakness.

History of present illness

The patient reported that the symptoms began abruptly two days ago and progressively worsened, without any triggers. The pain settled following the administration of morphine after admission to hospital.

History of past illness

In 2015 he was diagnosed with benign prostatic hypertrophy and had suffered traumatic injury following a road traffic accident over 40 years ago. Of note, he had no history of hepatic disease.

Physical examination

There was marked tenderness on inspiration on the right side of the chest, but otherwise physical examination was unremarkable. The patient's vital signs were normal with a temperature of 37.0 °C, heart rate of 74 bpm, blood pressure of 125/75 mmHg, respiratory rate of 15 breaths/min and oxygen saturations of 98% in room air.

Laboratory testing

Routine blood tests were within normal ranges including liver function tests, alpha-fetoprotein, prothrombin time and a normal D-Dimer.

Imaging examinations

A CT angiogram was performed to rule out aortic dissection due to his acute presentation. No evidence of the latter was seen but there was a 3-cm large arterially enhancing lesion in segment IV of the liver (Figure 1A). The lesion was arterialised with faint hypoenhancement in the portal venous phase (Figure 1B and C). Multiple arterially enhancing peritoneal, lesser sac and retroperitoneal nodules were additionally seen following the same enhancement pattern. The patient's spleen was observed to be lobulated and the right inferior ribs and right iliac crest had an abnormal appearance suggestive of previous trauma.

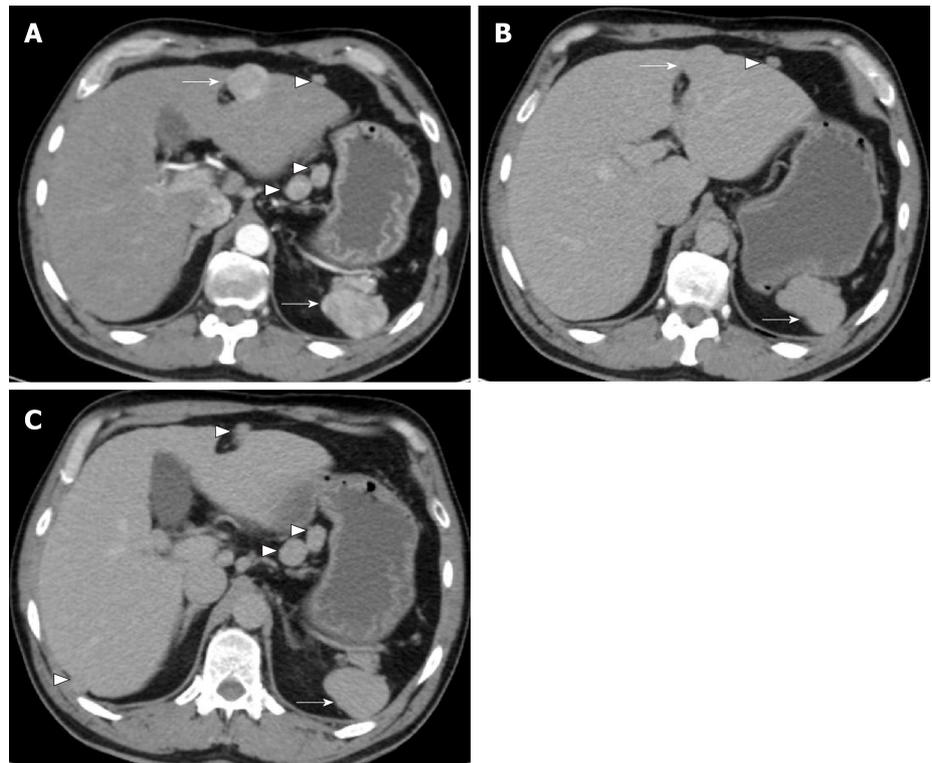


Figure 1 Computed tomography axial images from a patient with intrahepatic splenosis. A: arterial phase; a heterogeneously enhancing left lobe liver lesion is present 3 cm in diameter (thick arrow). Similar heterogeneously enhancing peritoneal and lesser sac nodules are seen (arrowhead). The deformed spleen shows typical heterogenous "zebra stripe" arterial enhancement; B and C: portal venous phase; the liver lesion (thick arrow) and the peritoneal and retroperitoneal nodules (arrowhead) are isodense and the same density as the spleen (thin arrow).

FINAL DIAGNOSIS

Following discussion of these imaging findings and the patient's history in a specialist multidisciplinary team meeting, the possibility of intrahepatic splenosis with additional intra-abdominal splenosis was considered. This diagnosis was confirmed using scintigraphy with technetium-99m heat-denatured red blood cells (Tc-99 DRBC), which demonstrated uptake of the radiolabelled red blood cells by the multiple peritoneal nodules, as well as the lesion within the liver and the anterior abdominal wall (Figure 2).

OUTCOME AND FOLLOW UP

Due to the extensively reported benign nature of this condition, treatment was not required. The patient was informed of the incidental imaging finding and reassured. He was also informed that his chest pain was likely to be musculoskeletal in nature.

DISCUSSION

We describe a case of intrahepatic splenosis diagnosed radiologically, in a patient with a history of trauma. To the authors' knowledge, this is the first case of intrahepatic splenosis diagnosed without the need for histological analysis, thereby avoiding the potential risk of complications secondary to invasive investigations such as a liver biopsy or laparoscopic surgery.

To date, 21 case reports of intrahepatic splenosis have been described in the literature. We specifically review 13 cases which include CT and MRI (Table 1). Nine cases describe solitary lesions^[9-17], whilst four cases involved multiple lesions^[18-21]. The nodules ranged in size from 1.5-5.0 cm and were primarily found in the left lobe of the liver. Additional abdominal splenic nodules were reported in four cases to be located in close proximity to the upper pole of the left kidney^[19], pancreatic tail^[19], mesentery of the colon^[20], paravesical space^[21], caecum^[21] and abdominal wall^[15].

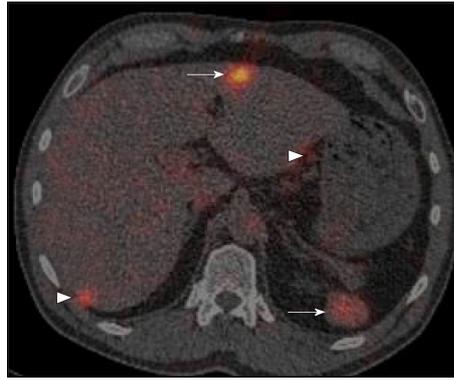


Figure 2 Denatured red cell scan with fused computed tomography images. The liver lesion (thick arrow), peritoneal and retroperitoneal nodules (arrowheads) and spleen (thin arrow) show uptake in keeping with multiple areas of splenic tissue.

All patients except one^[14] had undergone a splenectomy in the past. Notably, a wrong diagnosis was made in all but one case^[21], primarily of hepatocellular carcinoma (HCC)^[9,13-18,20], leading to unnecessary surgery with the correct diagnosis only being made following post-operative histological analysis. This is partly due to the fact that splenosis is rare and hence is often not considered amongst the differential diagnosis. Additionally, six of the patients included in the literature review had chronic liver disease^[9,13-16,18] including hepatitis B; a major risk factor for the development of HCC and four patients had raised tumour markers^[9,14,15,18]. In such cases, HCC presents a more likely diagnosis rather than hepatic splenosis. In the isolated case where splenosis was correctly suspected, percutaneous biopsy was still carried out for confirmation^[21].

Imaging is a useful diagnostic tool to distinguish splenosis from other lesions such as HCC, hepatic metastasis, haemangioma and focal nodular hyperplasia (FNH). CT and MRI provide panoramic imaging of the abdomen and can identify the size, location and enhancement characteristics of all lesions. Critically, all the splenic deposits exhibit an enhancement pattern identical to the native spleen on all imaging, with a heterogenous classical striped arterial hyperenhancement. However, this may be difficult to characterise if the native spleen is small or has been removed.

Classically, on unenhanced CT splenic tissue appears hypointense relative to the liver, whilst on MRI it appears hypodense on T1 and hyperintense on T2-weighted images. Five and six cases in the literature review exhibited these CT^[14-17,19] and MRI^[10,11,16,18,19,21] findings respectively (Table 1). On administration of contrast, splenic nodules are hyperintense in the arterial phase^[9,10,13-20] often with a striated appearance as seen in our patient. They vary in appearance in the portal venous phase and may be hypointense^[10,14,15,17,20], isointense^[19] or hyperintense^[9,13,16,18].

HCC has a variable appearance on both CT and MRI depending on biological characteristics including their degree of differentiation^[22]. Since their blood supply is derived from the neoangiogenesis of non-triadial arteries, HCC, like splenosis typically appear hyperenhancing in the arterial phase with portal venous washout^[22] (Table 2).

Hepatic metastasis also varies widely in appearance depending on the location of the primary tumour. They may appear hypo or hypervascular but typically show portal venous washout^[23]. Haemangioma, FNH and adenoma are benign lesions which typically show arterial hyperenhancement^[24-26] and hence may be mimicked by splenosis.

Of note, MRI using superparamagnetic iron oxide contrast instead of gadopentetate dimeglumine has been used to distinguish hepatic splenosis from malignancy^[27]. Following intravenous administration, these particles are removed from the circulation specifically by the reticuloendothelial cells of the liver and spleen, leading to a reduction in signal intensity of the hepatic and splenic parenchyma on T2-weighted MRI. Such a reduction in signal intensity is however not seen in malignant lesions except some well differentiated HCCs^[18,28]. Splenic nodules still have a higher intensity than the hypointense liver as they take up more contrast. Nonetheless, uptake of contrast still occurs in FNH and so the specificity of this technique is limited in isolation^[29].

Scintigraphy using Tc-99 DRBC is the current diagnostic tool of choice. This is due to its high specificity in identifying splenic tissue. It involves intravenous injection of heat denatured erythrocytes labelled with Tc-99. The majority, as many as 90% of the

Table 1 Characteristics of previous cases of intrahepatic splenosis identified in our literature review

Ref.	Age/sex	Blood results	Liver disease	Existing malignancy	Initial diagnosis	CT	MRI-T1	MRI-T2	Enhancement	Technique for diagnosis
[9]	60/M	Abnormal LFTs, ↑AFP	Chronic HepC	None	HCC	NA	NA	NA	A: Hyper V: Hyper	Laparoscopic surgery
[10]	54/M	Normal	None	None	NA	NA	Hypo	Slightly hyper	A: Hyper V: Hypo	Laparotomy
[11]	54/M	Normal	None	Gastric cancer	Liver metastasis	ND	Hypo	Slightly hyper	NA	Laparotomy
[12]	52/M	Normal	None	None	Neuroendocrine tumour	NA	NA	NA	Hypervascular	Surgery
[13]	53/M	↑γGT	NASH	None	HCC/hepatic adenoma	NA	NA	NA	A: Hyper V: Hyper	Laparoscopic surgery
[14]	58/M	↑AST, ↑ALT, ↑AFP, ↓PT, +HepC	Chronic HepC	None	HCC	Hypo	NA	Hyper	A: Hyper V: Hypo	Surgery
[15]	67/F	Slightly abnormal LFTs, ↑AFP	Chronic HepC	None	HCC	Hypo	Slightly hyper	Slightly hyper	A: Hyper V: Hypo	Surgery
[16]	42/M	+HepB +HepC	NASH	None	HCC	Hypo	Hypo	Hyper	A: Hyper V: Hyper	Laparotomy
[17]	54/M	Normal	None	None	HCC	Slightly hypo	Slightly hypo	Slightly hypo	A: Hyper V: Hypo	Surgery
[18]	32/M	↑AFP, ↑AST, +HepB	Chronic HepB	None	HCC	NA	Hypo	Hyper	A: Hyper V: Slightly hyper	Laparotomy
[19]	39/M	Normal	NA	None	Renal malignancy	Hypo	Hypo	Slightly hyper	A: Hyper V: Iso	Surgery
[20]	49/F	Normal	None	None	Hepatic malignancy	NA	NA	NA	A: Hyper V: Hypo	Laparotomy
[21]	69/M	Normal	None	None	Intrahepatic, abdominal splenosis	NA	Hypo	Slightly hyper	Hypovascular	Percutaneous biopsy

M: Male; F: Female; AFP: Alpha-fetoprotein; AST: Aspartate transaminase; ALT: Alanine transaminase; γGT: Gamma-glutamyltransferase; PT: Prothrombin time; LFTs: Liver function tests; Hep B+: Hepatitis B positive; Hep C+: Hepatitis C positive; NASH: Non-alcoholic steatohepatitis; HCC: Hepatocellular carcinoma; A: Arterial phase; V: Portal venous phase; Iso: Isointense; Hypo: Hypointense; Hyper: Hyperintense (relative to the liver parenchyma); ND: Lesion not detected; NA: Not available.

erythrocytes are sequestered in splenic tissue, whilst normal liver tissue or malignant lesions have relatively modest uptake of the radioactive isotope^[30]. This technique is therefore a reliable means of distinguishing splenic tissue from other hepatic lesions and avoids subjecting a patient to invasive procedures such as biopsy or surgery which are associated with their own risks.

Patients with splenosis are typically asymptomatic. Hence, surgery is only indicated if rare complications such as infarction^[31], bleeding^[32] or adhesions resulting in bowel obstruction occur^[33]. It is suggested that splenosis may even be beneficial, providing some degree of immunological protection^[13]. As it is a benign condition, it is often only diagnosed incidentally decades after the initial trauma following imaging for an unrelated condition. It is therefore difficult to ascertain the time taken for splenosis to occur. However, the process of splenic cells seeding in and growing on the serosal surface of the liver after recruiting nearby hepatic vasculature is likely to take several years. In the literature cases of intra-hepatic splenosis were diagnosed from a range of 5 to 46 years after trauma or splenectomy^[17].

CONCLUSION

In conclusion, splenosis should be considered as a possible differential diagnosis for a hepatic nodule in any patients with a history of abdominal trauma or previous splenectomy, especially when the nodules are located near the capsule of the liver and are associated with multiple intra-abdominal deposits. Scintigraphy using Tc-99

Table 2 Typical enhancement characteristics of ectopic splenic tissue and other hepatic lesions

Enhancement pattern	
Splenosis	Arterially hyperenhancing; variable venous enhancement
HCC	Arterially hyperenhancing; venous hypoenhancement
Hepatic metastasis	Arterially variable enhancement; venous hypoenhancement
Haemangioma	Arterially peripheral nodular enhancement; venous infilling
FNH	Arterially hyperenhancing; venous iso/hyperenhancing with late enhancement of scar on MRI
Hepatic adenoma	Arterially hyperenhancing; variable venous enhancement

HCC: Hepatocellular carcinoma; FNH: Focal nodular hyperplasia; MRI: Magnetic resonance imaging.

DRBC is a reliable technique to diagnose splenosis and should be carried out in all patients suspected of the condition before more invasive diagnostic procedures are considered. Greater awareness of this condition could reduce the high incidence of unnecessary invasive interventions in these patients.

REFERENCES

- Arena R**, Gasperoni S, Lisotti A, Petrini CAA, Brancaccio ML, Triossi O, Mussetto A. An unusual cause of gastrointestinal bleeding and intestinal obstruction. *Turk J Gastroenterol* 2018; **29**: 365-366 [PMID: 29755023 DOI: 10.5152/tjg.2018.17647]
- Guedes TP**, Fernandes B, Pedroto I. Hepatobiliary and Pancreatic: Symptomatic hepatic splenosis. *J Gastroenterol Hepatol* 2019 [PMID: 31317571 DOI: 10.1111/jgh.14743]
- Guan B**, Li XH, Wang L, Zhou M, Dong ZW, Luo GJ, Meng LP, Hu J, Jin WY. Gastric fundus splenosis with hemangioma masquerading as a gastrointestinal stromal tumor in a patient with schistosomiasis and cirrhosis who underwent splenectomy: A case report and literature review. *Medicine (Baltimore)* 2018; **97**: e11461 [PMID: 29979450 DOI: 10.1097/MD.00000000000011461]
- Bastidas AB**, Holloman D, Lankarani A, Nieto JM. Endoscopic Ultrasound-Guided Needle-Based Probe Confocal Laser Endomicroscopy (nCLE) of Intrapancreatic Ectopic Spleen. *ACG Case Rep J* 2016; **3**: 196-198 [PMID: 27144203 DOI: 10.14309/crj.2016.48]
- Ha YJ**, Hong TH, Choi YS. Thoracic Splenosis after Splenic and Diaphragmatic Injury. *Korean J Thorac Cardiovasc Surg* 2019; **52**: 47-50 [PMID: 30834219 DOI: 10.5090/kjtcs.2019.52.1.47]
- Page JB**, Lenz DL, Wong C. Right-sided intrarenal splenosis mimicking a renal carcinoma. *ScientificWorldJournal* 2006; **6**: 2442-2444 [PMID: 17619715 DOI: 10.1100/tsw.2006.380]
- Disanto MG**, Mercalli F, Palicelli A, Arnulfo A, Boldorini R. A unique case of bilateral ovarian splenosis and review of the literature. *APMIS* 2017; **125**: 844-848 [PMID: 28543860 DOI: 10.1111/apm.12714]
- Kim JM**, Lee SM, Choi J, Lyu J, Lee MS. Two Rare Cases of Intrathoracic Splenosis and Subcutaneous Splenosis: Spleen Scintigraphy Avoided the Need for Invasive Procedures. *Nucl Med Mol Imaging* 2016; **50**: 76-79 [PMID: 26941863 DOI: 10.1007/s13139-015-0370-0]
- Abu Hilal M**, Harb A, Zeidan B, Steadman B, Primrose JN, Pearce NW. Hepatic splenosis mimicking HCC in a patient with hepatitis C liver cirrhosis and mildly raised alpha fetoprotein; the important role of explorative laparoscopy. *World J Surg Oncol* 2009; **7**: 1 [PMID: 19123935 DOI: 10.1186/1477-7819-7-1]
- Yu H**, Xia L, Li T, Ju M, Liu L, Wu Z, Tang Z. Intrahepatic splenosis mimicking hepatoma. *BMJ Case Rep* 2009; 2009 [PMID: 21691391 DOI: 10.1136/bcr.06.2008.0230]
- Kang KC**, Cho GS, Chung GA, Kang GH, Kim YJ, Lee MS, Kim HK, Park SJ. Intrahepatic splenosis mimicking liver metastasis in a patient with gastric cancer. *J Gastric Cancer* 2011; **11**: 64-68 [PMID: 22076204 DOI: 10.5230/jgc.2011.11.1.64]
- Leong CW**, Menon T, Rao S. Post-traumatic intrahepatic splenosis mimicking a neuroendocrine tumour. *BMJ Case Rep* 2013; 2013 [PMID: 23470668 DOI: 10.1136/bcr-2012-007885]
- Inchingolo R**, Peddu P, Karani J. Hepatic splenosis presenting as arterialisied liver lesion in a patient with NASH. *Eur Rev Med Pharmacol Sci* 2013; **17**: 2853-2856 [PMID: 24254551]
- Sato N**, Abe T, Suzuki N, Waragai M, Teranishi Y, Takano Y, Sato A, Azami A, Gotoh M. Intrahepatic splenosis in a chronic hepatitis C patient with no history of splenic trauma mimicking hepatocellular carcinoma. *Am J Case Rep* 2014; **15**: 416-420 [PMID: 25261602 DOI: 10.12659/AJCR.890999]
- Li T**, Yang XY, Tang ZY. Intrahepatic and intraperitoneal splenosis mimicking hepatocellular carcinoma with abdominal wall metastasis in a patient with hepatitis C cirrhotic liver. *Surgery* 2015; **157**: 954-956 [PMID: 25934025 DOI: 10.1016/j.surg.2013.12.018]
- Wang MY**, Li B, Chen D, Liu AL, Qamar S, Sun MY. Spleen implanting in the fatty liver mimicking hepatocarcinoma in a patient with hepatitis B&C: A case report and literature review. *Medicine (Baltimore)* 2017; **96**: e7217 [PMID: 28640113 DOI: 10.1097/MD.00000000000007217]
- Xuan Z**, Chen J, Song P, Du Y, Wang L, Wan D, Zheng S. Management of intrahepatic splenosis: a case report and review of the literature. *World J Surg Oncol* 2018; **16**: 119 [PMID: 29954390 DOI: 10.1186/s12957-018-1419-1]
- Choi GH**, Ju MK, Kim JY, Kang CM, Kim KS, Choi JS, Han KH, Park MS, Park YN, Lee WJ, Kim BR. Hepatic splenosis preoperatively diagnosed as hepatocellular carcinoma in a patient with chronic hepatitis B: a case report. *J Korean Med Sci* 2008; **23**: 336-341 [PMID: 18437023 DOI: 10.3346/jkms.2008.23.2.336]
- Imbriaco M**, Camera L, Mancurina A, Salvatore M. A case of multiple intra-abdominal splenosis with computed tomography and magnetic resonance imaging correlative findings. *World J Gastroenterol* 2008; **14**: 1453-1455 [PMID: 18322965 DOI: 10.3748/wjg.14.1453]
- Liu Y**, Ji B, Wang G, Wang Y. Abdominal multiple splenosis mimicking liver and colon tumors: a case

- report and review of the literature. *Int J Med Sci* 2012; **9**: 174-177 [PMID: 22359484 DOI: 10.7150/ijms.3983]
- 21 **Vergara D**, Ginolfi F, Moscatti S, Giordano B, Ferrara N, Panico C, Imbriaco M. Multiple intra-hepatic and abdominal splenosis: an easy call if you know about it. *Acta Radiol Open* 2018; **7**: 2058460118772324 [PMID: 29780616 DOI: 10.1177/2058460118772324]
 - 22 **Choi JY**, Lee JM, Sirlin CB. CT and MR imaging diagnosis and staging of hepatocellular carcinoma: part I. Development, growth, and spread: key pathologic and imaging aspects. *Radiology* 2014; **272**: 635-654 [PMID: 25153274 DOI: 10.1148/radiol.14132361]
 - 23 **Sica GT**, Ji H, Ros PR. CT and MR imaging of hepatic metastases. *AJR Am J Roentgenol* 2000; **174**: 691-698 [PMID: 10701611 DOI: 10.2214/ajr.174.3.1740691]
 - 24 **Klotz T**, Montoriol PF, Da Ines D, Petitcolin V, Joubert-Zakeyh J, Garcier JM. Hepatic haemangioma: common and uncommon imaging features. *Diagn Interv Imaging* 2013; **94**: 849-859 [PMID: 23796395 DOI: 10.1016/j.diii.2013.04.008]
 - 25 **Mortelé KJ**, Praet M, Van Vlierberghe H, Kunnen M, Ros PR. CT and MR imaging findings in focal nodular hyperplasia of the liver: radiologic-pathologic correlation. *AJR Am J Roentgenol* 2000; **175**: 687-692 [PMID: 10954451 DOI: 10.2214/ajr.175.3.1750687]
 - 26 **Faria SC**, Iyer RB, Rashid A, Whitman GJ. Hepatic adenoma. *AJR Am J Roentgenol* 2004; **182**: 1520 [PMID: 15149999 DOI: 10.2214/ajr.182.6.1821520]
 - 27 **Kondo M**, Okazaki H, Takai K, Nishikawa J, Ohta H, Uekusa T, Yoshida H, Tanaka K. Intrahepatic splenosis in a patient with chronic hepatitis C. *J Gastroenterol* 2004; **39**: 1013-1015 [PMID: 15549458 DOI: 10.1007/s00535-004-1439-1]
 - 28 **De Vuysere S**, Van Steenberghe W, Aerts R, Van Hauwaert H, Van Beckevoort D, Van Hoe L. Intrahepatic splenosis: imaging features. *Abdom Imaging* 2000; **25**: 187-189 [PMID: 10675464 DOI: 10.1007/s002619910042]
 - 29 **Zheng WW**, Zhou KR, Chen ZW, Shen JZ, Chen CZ, Zhang SJ. Characterization of focal hepatic lesions with SPIO-enhanced MRI. *World J Gastroenterol* 2002; **8**: 82-86 [PMID: 11833077 DOI: 10.3748/wjg.v8.i1.82]
 - 30 **Massey MD**, Stevens JS. Residual spleen found on denatured red blood cell scan following negative colloid scans. *J Nucl Med* 1991; **32**: 2286-2287 [PMID: 1744718]
 - 31 **Książczyńska D**. A case report of abdominal splenosis - a practical mini-review for a gastroenterologist. *J Gastrointest Liver Dis* 2011; **20**: 321-324 [PMID: 21961104]
 - 32 **Basile RM**, Morales JM, Zupanec R. Splenosis. A cause of massive gastrointestinal hemorrhage. *Arch Surg* 1989; **124**: 1087-1089 [PMID: 2789029 DOI: 10.1001/archsurg.1989.01410090097022]
 - 33 **El-Kheir A**, Abdelnour M, Boutros JG. Simultaneous small bowel and colon obstruction due to splenosis. A case report and review of literature. *Int J Surg Case Rep* 2019; **58**: 63-66 [PMID: 31009896 DOI: 10.1016/j.ijscr.2019.03.040]



Published By Baishideng Publishing Group Inc
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA
Telephone: +1-925-2238242
E-mail: bpgoffice@wjgnet.com
Help Desk: <https://www.f6publishing.com/helpdesk>
<https://www.wjgnet.com>

