

World Journal of *Gastroenterology*

World J Gastroenterol 2016 June 28; 22(24): 5459-5622



Editorial Board

2014-2017

The *World Journal of Gastroenterology* Editorial Board consists of 1376 members, representing a team of worldwide experts in gastroenterology and hepatology. They are from 68 countries, including Algeria (2), Argentina (7), Australia (31), Austria (9), Belgium (11), Brazil (20), Brunei Darussalam (1), Bulgaria (2), Cambodia (1), Canada (26), Chile (4), China (164), Croatia (2), Cuba (1), Czech (6), Denmark (2), Egypt (9), Estonia (2), Finland (6), France (20), Germany (58), Greece (31), Guatemala (1), Hungary (15), Iceland (1), India (33), Indonesia (2), Iran (10), Ireland (9), Israel (18), Italy (194), Japan (149), Jordan (1), Kuwait (1), Lebanon (7), Lithuania (1), Malaysia (1), Mexico (11), Morocco (1), Netherlands (5), New Zealand (4), Nigeria (3), Norway (6), Pakistan (6), Poland (12), Portugal (8), Puerto Rico (1), Qatar (1), Romania (10), Russia (3), Saudi Arabia (2), Singapore (7), Slovenia (2), South Africa (1), South Korea (69), Spain (51), Sri Lanka (1), Sudan (1), Sweden (12), Switzerland (5), Thailand (7), Trinidad and Tobago (1), Tunisia (2), Turkey (55), United Kingdom (49), United States (180), Venezuela (1), and Vietnam (1).

EDITORS-IN-CHIEF

Stephen C Strom, *Stockholm*
Andrzej S Tarnawski, *Long Beach*
Damian Garcia-Olmo, *Madrid*

ASSOCIATE EDITORS

Yung-Jue Bang, *Seoul*
Vincent Di Martino, *Besancon*
Daniel T Farkas, *Bronx*
Roberto J Firpi, *Gainesville*
Maria Gazouli, *Athens*
Chung-Feng Huang, *Kaohsiung*
Namir Katkhouda, *Los Angeles*
Anna Kramvis, *Johannesburg*
Wolfgang Kruis, *Cologne*
Peter L Lakatos, *Budapest*
Han Chu Lee, *Seoul*
Christine McDonald, *Cleveland*
Nahum Mendez-Sanchez, *Mexico City*
George K Michalopoulos, *Pittsburgh*
Suk Woo Nam, *Seoul*
Shu-You Peng, *Hangzhou*
Daniel von Renteln, *Montreal*
Angelo Sangiovanni, *Milan*
Hildegard M Schuller, *Knoxville*
Dong-Wan Seo, *Seoul*
Adrian John Stanley, *Glasgow*
Jurgen Stein, *Frankfurt*
Bei-Cheng Sun, *Nanjing*
Yoshio Yamaoka, *Yufu*

GUEST EDITORIAL BOARD MEMBERS

Jia-Ming Chang, *Taipei*
Jane CJ Chao, *Taipei*

Kuen-Feng Chen, *Taipei*
Tai-An Chiang, *Tainan*
Yi-You Chiou, *Taipei*
Seng-Kee Chuah, *Kaohsiung*
Wan-Long Chuang, *Kaohsiung*
How-Ran Guo, *Tainan*
Ming-Chih Hou, *Taipei*
Po-Shiuan Hsieh, *Taipei*
Ching-Chuan Hsieh, *Chiayi county*
Jun-Te Hsu, *Taoyuan*
Chung-Ping Hsu, *Taichung*
Chien-Ching Hsu, *Taipei*
Chao-Hung Hung, *Kaohsiung*
Chen-Guo Ker, *Kaohsiung*
Yung-Chih Lai, *Taipei*
Teng-Yu Lee, *Taichung City*
Wei-Jei Lee, *Taoyuan*
Jin-Ching Lee, *Kaohsiung*
Jen-Kou Lin, *Taipei*
Ya-Wen Lin, *Taipei*
Hui-kang Liu, *Taipei*
Min-Hsiung Pan, *Taipei*
Bor-Shyang Sheu, *Tainan*
Hon-Yi Shi, *Kaohsiung*
Fung-Chang Sung, *Taichung*
Dar-In Tai, *Taipei*
Jung-Fa Tsai, *Kaohsiung*
Yao-Chou Tsai, *New Taipei City*
Chih-Chi Wang, *Kaohsiung*
Liang-Shun Wang, *New Taipei City*
Hsiu-Po Wang, *Taipei*
Jaw-Yuan Wang, *Kaohsiung*
Yuan-Huang Wang, *Taipei*
Yuan-Chuen Wang, *Taichung*

Deng-Chyang Wu, *Kaohsiung*
Shun-Fa Yang, *Taichung*
Hsu-Heng Yen, *Changhua*

MEMBERS OF THE EDITORIAL BOARD



Algeria

Saadi Berkane, *Algiers*
Samir Rouabhia, *Batna*



Argentina

N Tolosa de Talamoni, *Córdoba*
Eduardo de Santibanes, *Buenos Aires*
Bernardo Frider, *Capital Federal*
Guillermo Mazzolini, *Pilar*
Carlos Jose Pirola, *Buenos Aires*
Bernabé Matías Quesada, *Buenos Aires*
María Fernanda Troncoso, *Buenos Aires*



Australia

Golo Ahlenstiel, *Westmead*
Minoti V Apte, *Sydney*
Jacqueline S Barrett, *Melbourne*
Michael Beard, *Adelaide*
Filip Braet, *Sydney*
Guy D Eslick, *Sydney*
Christine Feinle-Bisset, *Adelaide*
Mark D Gorrell, *Sydney*
Michael Horowitz, *Adelaide*

Gordon Stanley Howarth, *Roseworthy*
 Seungha Kang, *Brisbane*
 Alfred King Lam, *Gold Coast*
 Ian C Lawrence, *Perth/Fremantle*
 Barbara Anne Leggett, *Brisbane*
 Daniel A Lemberg, *Sydney*
 Rupert W Leong, *Sydney*
 Finlay A Macrae, *Victoria*
 Vance Matthews, *Melbourne*
 David L Morris, *Sydney*
 Reme Mountifield, *Bedford Park*
 Hans J Netter, *Melbourne*
 Nam Q Nguyen, *Adelaide*
 Liang Qiao, *Westmead*
 Rajvinder Singh, *Adelaide*
 Ross Cyril Smith, *St Leonards*
 Kevin J Spring, *Sydney*
 Debbie Trinder, *Fremantle*
 Daniel R van Langenberg, *Box Hill*
 David Ian Watson, *Adelaide*
 Desmond Yip, *Garran*
 Li Zhang, *Sydney*



Austria

Felix Aigner, *Innsbruck*
 Gabriela A Berlakovich, *Vienna*
 Herwig R Cerwenka, *Graz*
 Peter Ferenci, *Wien*
 Alfred Gangl, *Vienna*
 Kurt Lenz, *Linz*
 Markus Peck-Radosavljevic, *Vienna*
 Markus Raderer, *Vienna*
 Stefan Riss, *Vienna*



Belgium

Michael George Adler, *Brussels*
 Benedicte Y De Winter, *Antwerp*
 Mark De Ridder, *Jette*
 Olivier Detry, *Liege*
 Denis Dufrane Dufrane, *Brussels*
 Sven M Francque, *Edegem*
 Nikos Kotzampassakis, *Liège*
 Geert KMM Robaeyns, *Genk*
 Xavier Sagaert, *Leuven*
 Peter Starkel, *Brussels*
 Eddie Wisse, *Keerbergen*



Brazil

SMP Balzan, *Santa Cruz do Sul*
 JLF Caboclo, *Sao Jose do Rio Preto*
 Fábio Guilherme Campos, *Sao Paulo*
 Claudia RL Cardoso, *Rio de Janeiro*
 Roberto J Carvalho-Filho, *Sao Paulo*
 Carla Daltro, *Salvador*
 José Sebastiao dos Santos, *Ribeirao Preto*
 Eduardo LR Mello, *Rio de Janeiro*
 Sthela Maria Murad-Regadas, *Fortaleza*
 Claudia PMS Oliveira, *Sao Paulo*
 Júlio C Pereira-Lima, *Porto Alegre*
 Marcos V Perini, *Sao Paulo*
 Vietla Satyanarayana Rao, *Fortaleza*

Raquel Rocha, *Salvador*
 AC Simoes e Silva, *Belo Horizonte*
 Mauricio F Silva, *Porto Alegre*
 Aytan Miranda Sipahi, *Sao Paulo*
 Rosa Leonóra Salerno Soares, *Niterói*
 Cristiane Valle Tovo, *Porto Alegre*
 Eduardo Garcia Vilela, *Belo Horizonte*



Brunei Darussalam

Vui Heng Chong, *Bandar Seri Begawan*



Bulgaria

Tanya Kirilova Kadiyska, *Sofia*
 Mihaela Petrova, *Sofia*



Cambodia

Francois Rouet, *Phnom Penh*



Canada

Brian Bressler, *Vancouver*
 Frank J Burczynski, *Winnipeg*
 Wangxue Chen, *Ottawa*
 Francesco Crea, *Vancouver*
 Mirko Diksic, *Montreal*
 Jane A Foster, *Hamilton*
 Hugh J Freeman, *Vancouver*
 Shahrokh M Ghobadloo, *Ottawa*
 Yuewen Gong, *Winnipeg*
 Philip H Gordon, *Quebec*
 Rakesh Kumar, *Edmonton*
 Wolfgang A Kunze, *Hamilton*
 Patrick Labonte, *Laval*
 Zhikang Peng, *Winnipeg*
 Jayadev Raju, *Ottawa*
 Maitreyi Raman, *Calgary*
 Giada Sebastiani, *Montreal*
 Maida J Sewitch, *Montreal*
 Eldon A Shaffer, *Alberta*
 Christopher W Teshima, *Edmonton*
 Jean Sévigny, *Québec*
 Pingchang Yang, *Hamilton*
 Pingchang Yang, *Hamilton*
 Eric M Yoshida, *Vancouver*
 Bin Zheng, *Edmonton*



Chile

Marcelo A Beltran, *La Serena*
 Flavio Nervi, *Santiago*
 Adolfo Parra-Blanco, *Santiago*
 Alejandro Soza, *Santiago*



China

Zhao-Xiang Bian, *Hong Kong*
 San-Jun Cai, *Shanghai*
 Guang-Wen Cao, *Shanghai*
 Long Chen, *Nanjing*
 Ru-Fu Chen, *Guangzhou*

George G Chen, *Hong Kong*
 Li-Bo Chen, *Wuhan*
 Jia-Xu Chen, *Beijing*
 Hong-Song Chen, *Beijing*
 Lin Chen, *Beijing*
 Yang-Chao Chen, *Hong Kong*
 Zhen Chen, *Shanghai*
 Ying-Sheng Cheng, *Shanghai*
 Kent-Man Chu, *Hong Kong*
 Zhi-Jun Dai, *Xi'an*
 Jing-Yu Deng, *Tianjin*
 Yi-Qi Du, *Shanghai*
 Zhi Du, *Tianjin*
 Hani El-Nezami, *Hong Kong*
 Bao-Ying Fei, *Hangzhou*
 Chang-Ming Gao, *Nanjing*
 Jian-Ping Gong, *Chongqing*
 Zuo-Jiong Gong, *Wuhan*
 Jing-Shan Gong, *Shenzhen*
 Guo-Li Gu, *Beijing*
 Yong-Song Guan, *Chengdu*
 Mao-Lin Guo, *Luoyang*
 Jun-Ming Guo, *Ningbo*
 Yan-Mei Guo, *Shanghai*
 Xiao-Zhong Guo, *Shenyang*
 Guo-Hong Han, *Xi'an*
 Ming-Liang He, *Hong Kong*
 Peng Hou, *Xi'an*
 Zhao-Hui Huang, *Wuxi*
 Feng Ji, *Hangzhou*
 Simon Law, *Hong Kong*
 Yu-Yuan Li, *Guangzhou*
 Meng-Sen Li, *Haikou*
 Shu-De Li, *Shanghai*
 Zong-Fang Li, *Xi'an*
 Qing-Quan Li, *Shanghai*
 Kang Li, *Lasa*
 Han Liang, *Tianjin*
 Xing'e Liu, *Hangzhou*
 Zheng-Wen Liu, *Xi'an*
 Xiao-Fang Liu, *Yantai*
 Bin Liu, *Tianjin*
 Quan-Da Liu, *Beijing*
 Hai-Feng Liu, *Beijing*
 Fei Liu, *Shanghai*
 Ai-Guo Lu, *Shanghai*
 He-Sheng Luo, *Wuhan*
 Xiao-Peng Ma, *Shanghai*
 Yong Meng, *Shantou*
 Ke-Jun Nan, *Xi'an*
 Siew Chien Ng, *Hong Kong*
 Simon SM Ng, *Hong Kong*
 Zhao-Shan Niu, *Qingdao*
 Di Qu, *Shanghai*
 Ju-Wei Mu, *Beijing*
 Rui-Hua Shi, *Nanjing*
 Bao-Min Shi, *Shanghai*
 Xiao-Dong Sun, *Hangzhou*
 Si-Yu Sun, *Shenyang*
 Guang-Hong Tan, *Haikou*
 Wen-Fu Tang, *Chengdu*
 Anthony YB Teoh, *Hong Kong*
 Wei-Dong Tong, *Chongqing*
 Eric Tse, *Hong Kong*
 Hong Tu, *Shanghai*

Rong Tu, *Haikou*
 Jian-She Wang, *Shanghai*
 Kai Wang, *Jinan*
 Xiao-Ping Wang, *Xianyang*
 Xiu-Yan Wang, *Shanghai*
 Dao-Rong Wang, *Yangzhou*
 De-Sheng Wang, *Xi'an*
 Chun-You Wang, *Wuhan*
 Ge Wang, *Chongqing*
 Xi-Shan Wang, *Harbin*
 Wei-hong Wang, *Beijing*
 Zhen-Ning Wang, *Shenyang*
 Wai Man Raymond Wong, *Hong Kong*
 Chun-Ming Wong, *Hong Kong*
 Jian Wu, *Shanghai*
 Sheng-Li Wu, *Xi'an*
 Wu-Jun Wu, *Xi'an*
 Qing Xia, *Chengdu*
 Yan Xin, *Shenyang*
 Dong-Ping Xu, *Beijing*
 Jian-Min Xu, *Shanghai*
 Wei Xu, *Changchun*
 Ming Yan, *Jinan*
 Xin-Min Yan, *Kunming*
 Yi-Qun Yan, *Shanghai*
 Feng Yang, *Shanghai*
 Yong-Ping Yang, *Beijing*
 He-Rui Yao, *Guangzhou*
 Thomas Yau, *Hong Kong*
 Winnie Yeo, *Hong Kong*
 Jing You, *Kunming*
 Jian-Qing Yu, *Wuhan*
 Ying-Yan Yu, *Shanghai*
 Wei-Zheng Zeng, *Chengdu*
 Zong-Ming Zhang, *Beijing*
 Dian-Liang Zhang, *Qingdao*
 Ya-Ping Zhang, *Shijiazhuang*
 You-Cheng Zhang, *Lanzhou*
 Jian-Zhong Zhang, *Beijing*
 Ji-Yuan Zhang, *Beijing*
 Hai-Tao Zhao, *Beijing*
 Jian Zhao, *Shanghai*
 Jian-Hong Zhong, *Nanning*
 Ying-Qiang Zhong, *Guangzhou*
 Ping-Hong Zhou, *Shanghai*
 Yan-Ming Zhou, *Xiamen*
 Tong Zhou, *Nanchong*
 Li-Ming Zhou, *Chengdu*
 Guo-Xiong Zhou, *Nantong*
 Feng-Shang Zhu, *Shanghai*
 Jiang-Fan Zhu, *Shanghai*
 Zhao-Hui Zhu, *Beijing*



Croatia

Tajana Filipec Kanizaj, *Zagreb*
 Mario Tadic, *Zagreb*



Cuba

Damian Casadesus, *Havana*



Czech

Jan Bures, *Hradec Kralove*
 Marcela Kopacova, *Hradec Kralove*

Otto Kucera, *Hradec Kralove*
 Marek Minarik, *Prague*
 Pavel Soucek, *Prague*
 Miroslav Zavoral, *Prague*



Denmark

Vibeke Andersen, *Odense*
 E Michael Danielsen, *Copenhagen*



Egypt

Mohamed MM Abdel-Latif, *Assiut*
 Hussein Atta, *Cairo*
 Ashraf Elbahrawy, *Cairo*
 Mortada Hassan El-Shabrawi, *Cairo*
 Mona El Said El-Raziky, *Cairo*
 Elrashdy M Redwan, *New Borg Alrab*
 Zeinab Nabil Ahmed Said, *Cairo*
 Ragaa HM Salama, *Assiut*
 Maha Maher Shehata, *Mansoura*



Estonia

Margus Lember, *Tartu*
 Tamara Vorobjova, *Tartu*



Finland

Marko Kalliomäki, *Turku*
 Thomas Kietzmann, *Oulu*
 Kaija-Leena Kolho, *Helsinki*
 Eija Korkeila, *Turku*
 Heikki Makisalo, *Helsinki*
 Tanja Pessi, *Tampere*



France

Armando Abergel Clermont, *Ferrand*
 Elie K Chouillard, *Polssy*
 Pierre Cordelier, *Toulouse*
 Pascal P Crenn, *Garches*
 Catherine Daniel, *Lille*
 Fanny Daniel, *Paris*
 Cedric Dray, *Toulouse*
 Benoit Foligne, *Lille*
 Jean-Noel Freund, *Strasbourg*
 Hervé Guillou, *Toulouse*
 Nathalie Janel, *Paris*
 Majid Khatib, *Bordeaux*
 Jacques Marescaux, *Strasbourg*
 Jean-Claude Marie, *Paris*
 Driffa Moussata, *Pierre Benite*
 Hang Nguyen, *Clermont-Ferrand*
 Hugo Perazzo, *Paris*
 Alain L Servin, *Chatenay-Malabry*
 Chang Xian Zhang, *Lyon*



Germany

Stavros A Antoniou, *Monchengladbach*
 Erwin Biecker, *Siegburg*
 Hubert E Blum, *Freiburg*

Thomas Bock, *Berlin*
 Katja Breitkopf-Heinlein, *Mannheim*
 Elke Cario, *Essen*
 Güralp Onur Ceyhan, *Munich*
 Angel Cid-Arregui, *Heidelberg*
 Michael Clemens Roggendorf, *München*
 Christoph F Dietrich, *Bad Mergentheim*
 Valentin Fuhrmann, *Hamburg*
 Nikolaus Gassler, *Aachen*
 Andreas Geier, *Wuerzburg*
 Markus Gerhard, *Munich*
 Anton Gillessen, *Muenster*
 Thorsten Oliver Goetze, *Offenbach*
 Daniel Nils Gotthardt, *Heidelberg*
 Robert Grützmänn, *Dresden*
 Thilo Hackert, *Heidelberg*
 Claus Hellerbrand, *Regensburg*
 Harald Peter Hoensch, *Darmstadt*
 Jens Hoepfner, *Freiburg*
 Richard Hummel, *Muenster*
 Jakob Robert Izbicki, *Hamburg*
 Gernot Maximilian Kaiser, *Essen*
 Matthias Kapischke, *Hamburg*
 Michael Keese, *Frankfurt*
 Andrej Khandoga, *Munich*
 Jorg Kleeff, *Munich*
 Alfred Koenigsrainer, *Tuebingen*
 Peter Christopher Konturek, *Saalfeld*
 Michael Linnebacher, *Rostock*
 Stefan Maier, *Kaufbeuren*
 Oliver Mann, *Hamburg*
 Marc E Martignoni, *Munic*
 Thomas Minor, *Bonn*
 Oliver Moeschler, *Osnabrueck*
 Jonas Mudter, *Eutin*
 Sebastian Mueller, *Heidelberg*
 Matthias Ocker, *Berlin*
 Andreas Ommer, *Essen*
 Albrecht Piiper, *Frankfurt*
 Esther Raskopf, *Bonn*
 Christoph Reichel, *Bad Brückenau*
 Elke Roeb, *Giessen*
 Udo Rolle, *Frankfurt*
 Karl-Herbert Schafer, *Zweibrücken*
 Peter Schemmer, *Heidelberg*
 Andreas G Schreyer, *Regensburg*
 Manuel A Silva, *Penzberg*
 Georgios C Sotiropoulos, *Essen*
 Ulrike S Stein, *Berlin*
 Dirk Uhlmann, *Leipzig*
 Michael Weiss, *Halle*
 Hong-Lei Weng, *Mannheim*
 Karsten Wursthorn, *Hamburg*



Greece

Alexandra Alexopoulou, *Athens*
 Nikolaos Antonakopoulos, *Athens*
 Stelios F Assimakopoulos, *Patras*
 Grigoris Chatzimavroudis, *Thessaloniki*
 Evangelos Cholongitas, *Thessaloniki*
 Gregory Christodoulidis, *Larisa*
 George N Dalekos, *Larisa*
 Urania Georgopoulou, *Athens*
 Eleni Gigi, *Thessaloniki*

Stavros Gourgiotis, *Athens*
 Leontios J Hadjileontiadis, *Thessaloniki*
 Thomas Hyphantis, *Ioannina*
 Ioannis Kanellos, *Thessaloniki*
 Stylianos Karatapanis, *Rhodes*
 Michael Koutsilieris, *Athens*
 Spiros D Ladas, *Athens*
 Theodoros K Liakakos, *Athens*
 Emanuel K Manesis, *Athens*
 Spilios Manolakopoulos, *Athens*
 Gerassimos John Mantzaris, *Athens*
 Athanasios D Marinis, *Piraeus*
 Nikolaos Ioannis Nikiteas, *Athens*
 Konstantinos X Papamichael, *Athens*
 George Sgourakis, *Athens*
 Konstantinos C Thomopoulos, *Patras*
 Konstantinos Triantafyllou, *Athens*
 Christos Triantos, *Patras*
 Georgios Zacharakis, *Athens*
 Petros Zezos, *Alexandroupolis*
 Demosthenes E Ziogas, *Ioannina*



Guatemala

Carlos Maria Parellada, *Guatemala*



Hungary

Mihaly Boros, *Szeged*
 Tamás Decsi, *Pécs*
 Gyula Farkas, *Szeged*
 Andrea Furka, *Debrecen*
 Y vette Mandi, *Szeged*
 Peter L Lakatos, *Budapest*
 Pal Miheller, *Budapest*
 Tamás Molnar, *Szeged*
 Attila Olah, *Gyor*
 Maria Papp, *Debrecen*
 Zoltan Rakonczay, *Szeged*
 Ferenc Sipos, *Budapest*
 Miklós Tanyi, *Debrecen*
 Tibor Wittmann, *Szeged*



Iceland

Tryggvi Bjorn Stefánsson, *Reykjavík*



India

Brij B Agarwal, *New Delhi*
 Deepak N Amarapurkar, *Mumbai*
 Shams ul Bari, *Srinagar*
 Sriparna Basu, *Varanasi*
 Runu Chakravarty, *Kolkata*
 Devendra C Desai, *Mumbai*
 Nutan D Desai, *Mumbai*
 Suneela Sunil Dhaneshwar, *Pune*
 Radha K Dhiman, *Chandigarh*
 Pankaj Garg, *Mohali*
 Uday C Ghoshal, *Lucknow*
 Kalpesh Jani, *Vadodara*
 Premashis Kar, *New Delhi*
 Jyotdeep Kaur, *Chandigarh*
 Rakesh Kochhar, *Chandigarh*

Pradyumna K Mishra, *Mumbai*
 Asish K Mukhopadhyay, *Kolkata*
 Imtiyaz Murtaza, *Srinagar*
 P Nagarajan, *New Delhi*
 Samiran Nundy, *Delhi*
 Gopal Pande, *Hyderabad*
 Benjamin Perakath, *Vellore*
 Arun Prasad, *New Delhi*
 D Nageshwar Reddy, *Hyderabad*
 Lekha Saha, *Chandigarh*
 Sundeep Singh Saluja, *New Delhi*
 Mahesh Prakash Sharma, *New Delhi*
 Sadiq Saleem Sikora, *Bangalore*
 Sarman Singh, *New Delhi*
 Rajeew Sinha, *Jhansi*
 Rupjyoti Talukdar, *Hyderabad*
 Rakesh Kumar Tandon, *New Delhi*
 Narayanan Thirumoorthy, *Coimbatore*



Indonesia

David Handojo Muljono, *Jakarta*
 Andi Utama, *Jakarta*



Iran

Arezoo Aghakhani, *Tehran*
 Seyed Mohsen Dehghani, *Shiraz*
 Ahad Eshraghian, *Shiraz*
 Hossein Khedmat, *Tehran*
 Sadegh Massarrat, *Tehran*
 Marjan Mohammadi, *Tehran*
 Roja Rahimi, *Tehran*
 Farzaneh Sabahi, *Tehran*
 Majid Sadeghizadeh, *Tehran*
 Farideh Siavoshi, *Tehran*



Ireland

Gary Alan Bass, *Dublin*
 David J Brayden, *Dublin*
 Ronan A Cahill, *Dublin*
 Glen A Doherty, *Dublin*
 Liam J Fanning, *Cork*
 Barry Philip McMahon, *Dublin*
 RossMcManus, *Dublin*
 Dervla O'Malley, *Cork*
 Sinead M Smith, *Dublin*



Israel

Dan Carter, *Ramat Gan*
 Jorge-Shmuel Delgado, *Metar*
 Eli Magen, *Ashdod*
 Nitsan Maharshak, *Tel Aviv*
 Shaul Mordechai, *Beer Sheva*
 Menachem Moshkowitz, *Tel Aviv*
 William Bahij Nseir, *Nazareth*
 Shimon Reif, *Jerusalem*
 Ram Reifen, *Rehovot*
 Ariella Bar-Gil Shitrit, *Jerusalem*
 Noam Shussman, *Jerusalem*
 Igor Sukhotnik, *Haifa*
 Nir Wasserberg, *Petach Tikva*

Jacob Yahav, *Rehovot*
 Doron Levi Zamir, *Gedera*
 Shira Zelter-Sagi, *Haifa*
 Romy Zemel, *Petach-Tikva*



Italy

Ludovico Abenavoli, *Catanzaro*
 Luigi Elio Adinolfi, *Naples*
 Carlo Virginio Agostoni, *Milan*
 Anna Alisi, *Rome*
 Piero Luigi Almasio, *Palermo*
 Donato Francesco Altomare, *Bari*
 Amedeo Amedei, *Florence*
 Pietro Andreone, *Bologna*
 Imerio Angriman, *Padova*
 Vito Annese, *Florence*
 Paolo Aurello, *Rome*
 Salvatore Auricchio, *Naples*
 Gian Luca Baiocchi, *Brescia*
 Gianpaolo Balzano, *Milan*
 Antonio Basoli, *Rome*
 Gabrio Bassotti, *San Sisto*
 Mauro Bernardi, *Bologna*
 Alberto Biondi, *Rome*
 Ennio Biscaldi, *Genova*
 Massimo Bolognesi, *Padua*
 Luigi Bonavina, *Milano*
 Aldo Bove, *Chieti*
 Raffaele Bruno, *Pavia*
 Luigi Brusciano, *Napoli*
 Giuseppe Cabibbo, *Palermo*
 Carlo Calabrese, *Bologna*
 Daniele Calistri, *Meldola*
 Vincenza Calvaruso, *Palermo*
 Lorenzo Camellini, *Reggio Emilia*
 Marco Candela, *Bologna*
 Raffaele Capasso, *Naples*
 Lucia Carulli, *Modena*
 Renato David Caviglia, *Rome*
 Luigina Cellini, *Chieti*
 Giuseppe Chiarioni, *Verona*
 Claudio Chiesa, *Rome*
 Michele Cicala, *Roma*
 Rachele Ciccocioppo, *Pavia*
 Sandro Contini, *Parma*
 Gaetano Corso, *Foggia*
 Renato Costi, *Parma*
 Alessandro Cucchetti, *Bologna*
 Rosario Cuomo, *Napoli*
 Giuseppe Currò, *Messina*
 Paola De Nardi, *Milano*
 Giovanni D De Palma, *Naples*
 Raffaele De Palma, *Napoli*
 Giuseppina De Petro, *Brescia*
 Valli De Re, *Aviano*
 Paolo De Simone, *Pisa*
 Giuliana Decorti, *Trieste*
 Emanuele Miraglia del Giudice, *Napoli*
 Isidoro Di Carlo, *Catania*
 Matteo Nicola Dario Di Minno, *Naples*
 Massimo Donadelli, *Verona*
 Mirko D'Onofrio, *Verona*
 Maria Pina Dore, *Sassari*
 Luca Elli, *Milano*
 Massimiliano Fabozzi, *Aosta*

Massimo Falconi, *Ancona*
 Ezio Falletto, *Turin*
 Silvia Fargion, *Milan*
 Matteo Fassan, *Verona*
 Gianfranco Delle Fave, *Roma*
 Alessandro Federico, *Naples*
 Francesco Feo, *Sassari*
 Davide Festi, *Bologna*
 Natale Figura, *Siena*
 Vincenzo Formica, *Rome*
 Mirella Fraquelli, *Milan*
 Marzio Frazzoni, *Modena*
 Walter Fries, *Messina*
 Gennaro Galizia, *Naples*
 Andrea Galli, *Florence*
 Matteo Garcovich, *Rome*
 Eugenio Gaudio, *Rome*
 Paola Ghiorzo, *Genoa*
 Edoardo G Giannini, *Genova*
 Luca Gianotti, *Monza*
 Maria Cecilia Giron, *Padova*
 Alberto Grassi, *Rimini*
 Gabriele Grassi, *Trieste*
 Francesco Greco, *Bergamo*
 Luigi Greco, *Naples*
 Antonio Grieco, *Rome*
 Fabio Grizzi, *Rozzano*
 Laurino Grossi, *Pescara*
 Simone Guglielmetti, *Milan*
 Tiberiu Hershcovici, *Jerusalem*
 Calogero Iacono, *Verona*
 Enzo Ierardi, *Bari*
 Amedeo Indriolo, *Bergamo*
 Raffaele Iorio, *Naples*
 Paola Iovino, *Salerno*
 Angelo A Izzo, *Naples*
 Loreta Kondili, *Rome*
 Filippo La Torre, *Rome*
 Giuseppe La Torre, *Rome*
 Giovanni Latella, *L'Aquila*
 Salvatore Leonardi, *Catania*
 Massimo Libra, *Catania*
 Anna Licata, *Palermo*
 Carmela Loguercio, *Naples*
 Amedeo Lonardo, *Modena*
 Carmelo Luigiano, *Catania*
 Francesco Luzzza, *Catanzaro*
 Giovanni Maconi, *Milano*
 Antonio Macrì, *Messina*
 Mariano Malaguarnera, *Catania*
 Francesco Manguso, *Napoli*
 Tommaso Maria Manzia, *Rome*
 Daniele Marrelli, *Siena*
 Gabriele Masselli, *Rome*
 Sara Massironi, *Milan*
 Giuseppe Mazzarella, *Avellino*
 Michele Milella, *Rome*
 Giovanni Milito, *Rome*
 Antonella d'Arminio Monforte, *Milan*
 Fabrizio Montecucco, *Genoa*
 Giovanni Monteleone, *Rome*
 Mario Morino, *Torino*
 Vincenzo La Mura, *Milan*
 Gerardo Nardone, *Naples*
 Riccardo Nascimbeni, *Brescia*
 Gabriella Nesi, *Florence*
 Giuseppe Nigri, *Rome*

Erica Novo, *Turin*
 Veronica Ojetti, *Rome*
 Michele Orditura, *Naples*
 Fabio Pace, *Seriate*
 Lucia Pacifico, *Rome*
 Omero Alessandro Paoluzi, *Rome*
 Valerio Paziienza, *San Giovanni Rotondo*
 Rinaldo Pellicano, *Turin*
 Adriano M Pellicelli, *Rome*
 Nadia Peparini, *Ciampino*
 Mario Pescatori, *Rome*
 Antonio Picardi, *Rome*
 Alberto Pilotto, *Padova*
 Alberto Piperno, *Monza*
 Anna Chiara Piscaglia, *Rome*
 Maurizio Pompili, *Rome*
 Francesca Romana Ponziani, *Rome*
 Cosimo Prantero, *Rome*
 Girolamo Ranieri, *Bari*
 Carlo Ratto, *Tome*
 Barbara Renga, *Perugia*
 Alessandro Repici, *Rozzano*
 Maria Elena Riccioni, *Rome*
 Lucia Ricci-Vitiani, *Rome*
 Luciana Rigoli, *Messina*
 Mario Rizzetto, *Torino*
 Ballarin Roberto, *Modena*
 Roberto G Romanelli, *Florence*
 Claudio Romano, *Messina*
 Luca Roncucci, *Modena*
 Cesare Ruffolo, *Treviso*
 Lucia Sacchetti, *Napoli*
 Rodolfo Sacco, *Pisa*
 Lapo Sali, *Florence*
 Romina Salpini, *Rome*
 Giulio Aniello, *Santoro Treviso*
 Armando Santoro, *Rozzano*
 Edoardo Savarino, *Padua*
 Marco Senzolo, *Padua*
 Annalucia Serafino, *Rome*
 Giuseppe S Sica, *Rome*
 Pierpaolo Sileri, *Rome*
 Cosimo Sperti, *Padua*
 Vincenzo Stanghellini, *Bologna*
 Cristina Stasi, *Florence*
 Gabriele Stocco, *Trieste*
 Roberto Tarquini, *Florence*
 Mario Testini, *Bari*
 Guido Torzilli, *Milan*
 Guido Alberto Massimo, *Tiberio Brescia*
 Giuseppe Toffoli, *Aviano*
 Alberto Tommasini, *Trieste*
 Francesco Tonelli, *Florence*
 Cesare Tosetti Porretta, *Terme*
 Lucio Trevisani, *Cona*
 Guglielmo M Trovato, *Catania*
 Mariapia Vairetti, *Pavia*
 Luca Vittorio Valenti, *Milano*
 Mariateresa T Ventura, *Bari*
 Giuseppe Verlato, *Verona*
 Marco Vivarelli, *Ancona*
 Giovanni Li Volti, *Catania*
 Giuseppe Zanotti, *Padua*
 Vincenzo Zara, *Lecce*
 Gianguglielmo Zehender, *Milan*
 Anna Linda Zignego, *Florence*
 Rocco Antonio Zoccali, *Messina*

Angelo Zullo, *Rome*



Japan

Yasushi Adachi, *Sapporo*
 Takafumi Ando, *Nagoya*
 Masahiro Arai, *Tokyo*
 Makoto Arai, *Chiba*
 Takaaki Arigami, *Kagoshima*
 Itaru Endo, *Yokohama*
 Munechika Enjoji, *Fukuoka*
 Shunji Fujimori, *Tokyo*
 Yasuhiro Fujino, *Akashi*
 Toshiyoshi Fujiwara, *Okayama*
 Yosuke Fukunaga, *Tokyo*
 Toshio Fukusato, *Tokyo*
 Takahisa Furuta, *Hamamatsu*
 Osamu Handa, *Kyoto*
 Naoki Hashimoto, *Osaka*
 Yoichi Hiasa, *Toon*
 Masatsugu Hiraki, *Saga*
 Satoshi Hirano, *Sapporo*
 Keiji Hirata, *Fukuoka*
 Toru Hiyama, *Higashihiroshima*
 Akira Hokama, *Nishihara*
 Shu Hoteya, *Tokyo*
 Masao Ichinose, *Wakayama*
 Tatsuya Ide, *Kurume*
 Masahiro Iizuka, *Akita*
 Toshiro Iizuka, *Tokyo*
 Kenichi Ikejima, *Tokyo*
 Tetsuya Ikemoto, *Tokushima*
 Hiroyuki Imaeda, *Saitama*
 Atsushi Imagawa, *Kan-onji*
 Hiroo Imazu, *Tokyo*
 Shuji Isaji, *Tsu*
 Toru Ishikawa, *Niigata*
 Toshiyuki Ishiwata, *Tokyo*
 Soichi Itaba, *Kitakyushu*
 Yoshiaki Iwasaki, *Okayama*
 Tatehiro Kagawa, *Isehara*
 Satoru Kakizaki, *Maebashi*
 Naomi Kakushima, *Shizuoka*
 Terumi Kamisawa, *Tokyo*
 Akihide Kamiya, *Isehara*
 Osamu Kanauchi, *Tokyo*
 Tatsuo Kanda, *Chiba*
 Shin Kariya, *Okayama*
 Shigeyuki Kawa, *Matsumoto*
 Takumi Kawaguchi, *Kurume*
 Takashi Kawai, *Tokyo*
 Soo Ryang Kim, *Kobe*
 Shinsuke Kiriya, *Gunma*
 Tsuneo Kitamura, *Urayasu*
 Masayuki Kitano, *Osakasayama*
 Hirotohi Kobayashi, *Tokyo*
 Hironori Koga, *Kurume*
 Takashi Kojima, *Sapporo*
 Satoshi Kokura, *Kyoto*
 Shuhei Komatsu, *Kyoto*
 Tadashi Kondo, *Tokyo*
 Yasuteru Kondo, *Sendai*
 Yasuhiro Kuramitsu, *Yamaguchi*
 Yukinori Kurokawa, *Osaka*
 Shin Maeda, *Yokohama*
 Koutarou Maeda, *Toyoake*

Hitoshi Maruyama, *Chiba*
 Atsushi Masamune, *Sendai*
 Hiroyuki Matsubayashi, *Suntogun*
 Akihisa Matsuda, *Inzai*
 Hirofumi Matsui, *Tsukuba*
 Akira Matsumori, *Kyoto*
 Yoichi Matsuo, *Nagoya*
 Y Matsuzaki, *Ami*
 Toshihiro Mitaka, *Sapporo*
 Kouichi Miura, *Akita*
 Shinichi Miyagawa, *Matumoto*
 Eiji Miyoshi, *Suita*
 Toru Mizuguchi, *Sapporo*
 Nobumasa Mizuno, *Nagoya*
 Zenichi Morise, *Nagoya*
 Tomohiko Moriyama, *Fukuoka*
 Kunihiko Murase, *Tusima*
 Michihiro Mutoh, *Tsukiji*
 Akihito Nagahara, *Tokyo*
 Hikaru Nagahara, *Tokyo*
 Hidenari Nagai, *Tokyo*
 Koichi Nagata, *Shimotsuke-shi*
 Masaki Nagaya, *Kawasaki*
 Hisato Nakajima, *Nishi-Shinbashi*
 Toshifusa Nakajima, *Tokyo*
 Hiroshi Nakano, *Kawasaki*
 Hiroshi Nakase, *Kyoto*
 Toshiyuki Nakayama, *Nagasaki*
 Takahiro Nakazawa, *Nagoya*
 Shoji Natsugoe, *Kagoshima City*
 Tsutomu Nishida, *Suita*
 Shuji Nomoto, *Naogyu*
 Sachiyo Nomura, *Tokyo*
 Takeshi Ogura, *Takatsukishi*
 Nobuhiro Ohkohchi, *Tsukuba*
 Toshifumi Ohkusa, *Kashiwa*
 Hirohide Ohnishi, *Akita*
 Teruo Okano, *Tokyo*
 Satoshi Osawa, *Hamamatsu*
 Motoyuki Otsuka, *Tokyo*
 Michitaka Ozaki, *Sapporo*
 Satoru Saito, *Yokohama*
 Naoaki Sakata, *Sendai*
 Ken Sato, *Maebashi*
 Toshiro Sato, *Tokyo*
 Tomoyuki Shibata, *Toyoake*
 Tomohiko Shimatani, *Kure*
 Yukihiko Shimizu, *Nanto*
 Tadashi Shimoyama, *Hirosaki*
 Masayuki Sho, *Nara*
 Ikuo Shoji, *Kobe*
 Atsushi Sofuni, *Tokyo*
 Takeshi Suda, *Niigata*
 M Sugimoto, *Hamamatsu*
 Ken Sugimoto, *Hamamatsu*
 Haruhiko Sugimura, *Hamamatsu*
 Shoichiro Sumi, *Kyoto*
 Hidekazu Suzuki, *Tokyo*
 Masahiro Tajika, *Nagoya*
 Hitoshi Takagi, *Takasaki*
 Toru Takahashi, *Niigata*
 Yoshihisa Takahashi, *Tokyo*
 Shinsuke Takeno, *Fukuoka*
 Akihiro Tamori, *Osaka*
 Kyosuke Tanaka, *Tsu*
 Shinji Tanaka, *Hiroshima*

Atsushi Tanaka, *Tokyo*
 Yasuhito Tanaka, *Nagoya*
 Shinji Tanaka, *Tokyo*
 Minoru Tomizawa, *Yotsukaido City*
 Kyoko Tsukiyama-Kohara, *Kagoshima*
 Takuya Watanabe, *Niigata*
 Kazuhiro Watanabe, *Sendai*
 Satoshi Yamagiwa, *Niigata*
 Takayuki Yamamoto, *Yokkaichi*
 Hiroshi Yamamoto, *Otsu*
 Kosho Yamanouchi, *Nagasaki*
 Ichiro Yasuda, *Gifu*
 Yutaka Yata, *Maebashi-city*
 Shin-ichi Yokota, *Sapporo*
 Norimasa Yoshida, *Kyoto*
 Hiroshi Yoshida, *Tama-City*
 Hitoshi Yoshiji, *Kashihara*
 Kazuhiko Yoshimatsu, *Tokyo*
 Kentaro Yoshioka, *Toyoake*
 Nobuhiro Zaima, *Nara*



Jordan

Khaled Ali Jadallah, *Irbid*



Kuwait

Islam Khan, *Kuwait*



Lebanon

Bassam N Abboud, *Beirut*
 Kassem A Barada, *Beirut*
 Marwan Ghosn, *Beirut*
 Iyad A Issa, *Beirut*
 Fadi H Mourad, *Beirut*
 AIA Sharara, *Beirut*
 Rita Slim, *Beirut*



Lithuania

Antanas Mickevicius, *Kaunas*



Malaysia

Huck Joo Tan, *Petaling Jaya*



Mexico

Richard A Awad, *Mexico City*
 Carlos R Camara-Lemarroy, *Monterrey*
 Norberto C Chavez-Tapia, *Mexico City*
 Wolfgang Gaertner, *Mexico City*
 Diego Garcia-Compean, *Monterrey*
 Arturo Panduro, *Guadalajara*
 OT Teramoto-Matsubara, *Mexico City*
 Felix Tellez-Avila, *Mexico City*
 Omar Vergara-Fernandez, *Mexico City*
 Saúl Villa-Trevino, *Cuidad de México*



Morocco

Samir Ahboucha, *Khouribga*



Netherlands

Robert J de Knegt, *Rotterdam*
 Tom Johannes Gerardus Gevers, *Nijmegen*
 Menno Hoekstra, *Leiden*
 BW Marcel Spanier, *Arnhem*
 Karel van Erpecum, *Utrecht*



New Zealand

Leo K Cheng, *Auckland*
 Andrew Stewart Day, *Christchurch*
 Jonathan Barnes Koea, *Auckland*
 Max Petrov, *Auckland*



Nigeria

Olufunmilayo Adenike Lesi, *Lagos*
 Jesse Abiodun Otegbayo, *Ibadan*
 Stella Ifeanyi Smith, *Lagos*



Norway

Trond Berg, *Oslo*
 Trond Arnulf Buanes, *Krokkleiva*
 Thomas de Lange, *Rud*
 Magdy El-Salhy, *Stord*
 Rasmus Goll, *Tromso*
 Dag Arne Lihaug Hoff, *Aalesund*



Pakistan

Zaigham Abbas, *Karachi*
 Usman A Ashfaq, *Faisalabad*
 Muhammad Adnan Bawany, *Hyderabad*
 Muhammad Idrees, *Lahore*
 Saeed Sadiq Hamid, *Karachi*
 Yasir Waheed, *Islamabad*



Poland

Thomas Brzozowski, *Cracow*
 Magdalena Chmiela, *Lodz*
 Krzysztof Jonderko, *Sosnowiec*
 Anna Kasicka-Jonderko, *Sosnowiec*
 Michal Kukla, *Katowice*
 Tomasz Hubert Mach, *Krakow*
 Agata Mulak, *Wroclaw*
 Danuta Owczarek, *Krakow*
 Piotr Socha, *Warsaw*
 Piotr Stalke, *Gdansk*
 Julian Teodor Swierczynski, *Gdansk*
 Anna M Zawilak-Pawlik, *Wroclaw*



Portugal

Marie Isabelle Cremers, *Setubal*
 Ceu Figueiredo, *Porto*
 Ana Isabel Lopes, *Lisbon*
 M Paula Macedo, *Lisboa*
 Ricardo Marcos, *Porto*
 Rui T Marinho, *Lisboa*
 Guida Portela-Gomes, *Estoril*

Filipa F Vale, *Lisbon*



Puerto Rico

Caroline B Appleyard, *Ponce*



Qatar

Abdulbari Bener, *Doha*



Romania

Mihai Ciocirlan, *Bucharest*

Dan Lucian Dumitrascu, *Cluj-Napoca*

Carmen Fierbinteanu-Braticevici, *Bucharest*

Romeo G Mihaila, *Sibiu*

Lucian Negreanu, *Bucharest*

Adrian Saftoiu, *Craiova*

Andrada Seicean, *Cluj-Napoca*

Ioan Sporea, *Timisoara*

Letiția Adela Maria Streba, *Craiova*

Anca Trifan, *Iasi*



Russia

Victor Pasechnikov, *Stavropol*

Vasilii Ivanovich Reshetnyak, *Moscow*

Vitaly Skoropad, *Obninsk*



Saudi Arabia

Abdul-Wahed N Meshikhes, *Dammam*

M Ezzedien Rabie, *Khamis Mushait*



Singapore

Brian KP Goh, *Singapore*

Richie Soong, *Singapore*

Ker-Kan Tan, *Singapore*

Kok-Yang Tan, *Singapore*

Yee-Joo Tan, *Singapore*

Mark Wong, *Singapore*

Hong Ping Xia, *Singapore*



Slovenia

Matjaz Homan, *Ljubljana*

Martina Perse, *Ljubljana*



South Korea

Sang Hoon Ahn, *Seoul*

Seung Hyuk Baik, *Seoul*

Soon Koo Baik, *Wonju*

Soo-Cheon Chae, *Iksan*

Byung-Ho Choe, *Daegu*

Suck Chei Choi, *Iksan*

Hoon Jai Chun, *Seoul*

Yeun-Jun Chung, *Seoul*

Young-Hwa Chung, *Seoul*

Ki-Baik Hahm, *Seongnam*

Sang Young Han, *Busan*

Seok Joo Han, *Seoul*

Seung-Heon Hong, *Iksan*

Jin-Hyeok Hwang, *Seoungnam*

Jeong Won Jang, *Seoul*

Jin-Young Jang, *Seoul*

Dae-Won Jun, *Seoul*

Young Do Jung, *Kwangju*

Gyeong Hoon Kang, *Seoul*

Sung-Bum Kang, *Seoul*

Koo Jeong Kang, *Daegu*

Ki Mun Kang, *Jinju*

Chang Moo Kang, *Seodaemun-gu*

Gwang Ha Kim, *Busan*

Sang Soo Kim, *Goyang-si*

Jin Cheon Kim, *Seoul*

Tae Il Kim, *Seoul*

Jin Hong Kim, *Suwon*

Kyung Mo Kim, *Seoul*

Kyongmin Kim, *Suwon*

Hyung-Ho Kim, *Seongnam*

Seoung Hoon Kim, *Goyang*

Sang Il Kim, *Seoul*

Hyun-Soo Kim, *Wonju*

Jung Mogg Kim, *Seoul*

Dong Yi Kim, *Gwangju*

Kyun-Hwan Kim, *Seoul*

Jong-Han Kim, *Ansan*

Sang Wun Kim, *Seoul*

Ja-Lok Ku, *Seoul*

Kyu Taek Lee, *Seoul*

Hae-Wan Lee, *Chuncheon*

Inchul Lee, *Seoul*

Jung Eun Lee, *Seoul*

Sang Chul Lee, *Daejeon*

Song Woo Lee, *Ansan-si*

Hyuk-Joon Lee, *Seoul*

Seong-Wook Lee, *Yongin*

Kil Yeon Lee, *Seoul*

Jong-Inn Lee, *Seoul*

Kyung A Lee, *Seoul*

Jong-Baeck Lim, *Seoul*

Eun-Yi Moon, *Seoul*

SH Noh, *Seoul*

Seung Woon Paik, *Seoul*

Won Sang Park, *Seoul*

Sung-Joo Park, *Iksan*

Kyung Sik Park, *Daegu*

Se Hoon Park, *Seoul*

Yoonkyung Park, *Gwangju*

Seung-Wan Ryu, *Daegu*

Il Han Song, *Cheonan*

Myeong Jun Song, *Daejeon*

Yun Kyoung Yim, *Daejeon*

Dae-Yeul Yu, *Daejeon*



Spain

Mariam Aguas, *Valencia*

Raul J Andrade, *Málaga*

Antonio Arroyo, *Elche*

Josep M Bordas, *Barcelona*

Lisardo Boscá, *Madrid*

Ricardo Robles Campos, *Murcia*

Jordi Camps, *Reus*

Carlos Cervera, *Barcelona*

Alfonso Clemente, *Granada*

Pilar Codoner-Franch, *Valencia*

Fernando J Corrales, *Pamplona*

Fermin Sánchez de Medina, *Granada*

Alberto Herreros de Tejada, *Majadahonda*

Enrique de-Madaria, *Alicante*

JE Dominguez-Munoz, *Santiago de Compostela*

Vicente Felipe, *Valencia*

CM Fernandez-Rodriguez, *Madrid*

Carmen Frontela-Saseta, *Murcia*

Julio Galvez, *Granada*

Maria Teresa García, *Vigo*

MI Garcia-Fernandez, *Málaga*

Emilio Gonzalez-Reimers, *La Laguna*

Marcel Jimenez, *Bellaterra*

Angel Lanas, *Zaragoza*

Juan Ramón Larrubia, *Guadalajara*

Antonio Lopez-Sanroman, *Madrid*

Vicente Lorenzo-Zuniga, *Badalona*

Alfredo J Lucendo, *Tomelloso*

Vicenta Soledad Martinez-Zorzano, *Vigo*

José Manuel Martin-Villa, *Madrid*

Julio Mayol, *Madrid*

Manuel Morales-Ruiz, *Barcelona*

Alfredo Moreno-Egea, *Murcia*

Albert Pares, *Barcelona*

Maria Pellise, *Barcelona*

José Perea, *Madrid*

Miguel Angel Plaza, *Zaragoza*

María J Pozo, *Cáceres*

Enrique Quintero, *La Laguna*

Jose M Ramia, *Madrid*

Francisco Rodriguez-Frias, *Barcelona*

Silvia Ruiz-Gaspa, *Barcelona*

Xavier Serra-Aracil, *Barcelona*

Vincent Soriano, *Madrid*

Javier Suarez, *Pamplona*

Carlos Taxonera, *Madrid*

M Isabel Torres, *Jaén*

Manuel Vazquez-Carrera, *Barcelona*

Benito Velayos, *Valladolid*

Silvia Vidal, *Barcelona*



Sri Lanka

Arjuna Priyadarsin De Silva, *Colombo*



Sudan

Ishag Adam, *Khartoum*



Sweden

Roland G Andersson, *Lund*

Bergthor Björnsson, *Linköping*

Johan Christopher Bohr, *Örebro*

Mauro D'Amato, *Stockholm*

Thomas Franzen, *Norrköping*

Evangelos Kalaitzakis, *Lund*

Riadh Sadik, *Gothenburg*

Per Anders Sandstrom, *Linköping*

Ervin Toth, *Malmö*

Konstantinos Tsimogiannis, *Vasteras*

Apostolos V Tsolakis, *Uppsala*



Switzerland

Gieri Cathomas, *Liestal*
Jean Louis Frossard, *Geneve*
Christian Toso, *Geneva*
Stephan Robert Vavricka, *Zurich*
Dominique Velin, *Lausanne*



Thailand

Thawatchai Akaraviputh, *Bangkok*
P Yoysungnoen Chintana, *Pathumthani*
Veerapol Kukongviriyapan, *Muang*
Vijitra Leardkamolkarn, *Bangkok*
Varut Lohsiriwat, *Bangkok*
Somchai Pinlaor, *Khaon Kaen*
D Wattanasirichaigoon, *Bangkok*



Trinidad and Tobago

B Shivananda Nayak, *Mount Hope*



Tunisia

Ibtissem Ghedira, *Sousse*
Lilia Zouiten-Mekki, *Tunis*



Turkey

Inci Alican, *Istanbul*
Mustafa Altindis, *Sakarya*
Mutay Aslan, *Antalya*
Oktar Asoglu, *Istanbul*
Yasemin Hatice Balaban, *Istanbul*
Metin Basaranoglu, *Ankara*
Yusuf Bayraktar, *Ankara*
Süleyman Bayram, *Adiyaman*
Ahmet Bilici, *Istanbul*
Ahmet Sedat Boyacioglu, *Ankara*
Züleyha Akkan Cetinkaya, *Kocaeli*
Cavit Col, *Bolu*
Yasar Colak, *Istanbul*
Cagatay Erden Daphan, *Kirikkale*
Mehmet Demir, *Hatay*
Ahmet Merih Dobrucali, *Istanbul*
Gülüm Ozlem Elpek, *Antalya*
Ayse Basak Engin, *Ankara*
Eren Ersoy, *Ankara*
Osman Ersoy, *Ankara*
Yusuf Ziya Erzin, *Istanbul*
Mukaddes Esrefoglu, *Istanbul*
Levent Filik, *Ankara*
Ozgur Harmanci, *Ankara*
Koray Hekimoglu, *Ankara*
Abdurrahman Kadayifci, *Gaziantep*
Cem Kalayci, *Istanbul*
Selin Kapan, *Istanbul*
Huseyin Kayadibi, *Adana*
Sabahattin Kaymakoglu, *Istanbul*
Metin Kement, *Istanbul*
Mevlut Kurt, *Bolu*
Resat Ozaras, *Istanbul*
Elvan Ozbek, *Adapazari*

Cengiz Ozcan, *Mersin*
Hasan Ozen, *Ankara*
Halil Ozguc, *Bursa*
Mehmet Ozturk, *Izmir*
Orhan V Ozkan, *Sakarya*
Semra Paydas, *Adana*
Ozlem Durmaz Suoglu, *Istanbul*
Ilker Tasci, *Ankara*
Müge Tecder-ünal, *Ankara*
Mesut Tez, *Ankara*
Serdar Topaloglu, *Trabzon*
Murat Toruner, *Ankara*
Gokhan Tumgor, *Adana*
Oguz Uskudar, *Adana*
Mehmet Yalniz, *Elazig*
Mehmet Yaman, *Elazig*
Veli Yazisiz, *Antalya*
Yusuf Yilmaz, *Istanbul*
Ozlem Yilmaz, *Izmir*
Oya Yucel, *Istanbul*
Ilhami Yuksel, *Ankara*



United Kingdom

Nadeem Ahmad Afzal, *Southampton*
Navneet K Ahluwalia, *Stockport*
Yeng S Ang, *Lancashire*
Ramesh P Arasaradnam, *Coventry*
Ian Leonard Phillip Beales, *Norwich*
John Beynon, *Swansea*
Barbara Braden, *Oxford*
Simon Bramhall, *Birmingham*
Geoffrey Burnstock, *London*
Ian Chau, *Sutton*
Thean Soon Chew, *London*
Helen G Coleman, *Belfast*
Anil Dhawan, *London*
Sunil Dolwani, *Cardiff*
Piers Gatenby, *London*
Anil T George, *London*
Pasquale Giordano, *London*
Paul Henderson, *Edinburgh*
Georgina Louise Hold, *Aberdeen*
Stefan Hubscher, *Birmingham*
Robin D Hughes, *London*
Nusrat Husain, *Manchester*
Matt W Johnson, *Luton*
Konrad Koss, *Macclesfield*
Anastasios Koulaouzidis, *Edinburgh*
Simon Lal, *Salford*
John S Leeds, *Aberdeen*
JK K Limdi, *Manchester*
Hongxiang Liu, *Cambridge*
Michael Joseph McGarvey, *London*
Michael Anthony Mendall, *London*
Alexander H Mirnezami, *Southampton*
J Bernadette Moore, *Guildford*
Claudio Nicoletti, *Norwich*
Savvas Papagrigoriadis, *London*
Sylvia LF Pender, *Southampton*
David Mark Pritchard, *Liverpool*
James A Ross, *Edinburgh*
Kamran Rostami, *Worcester*
Xiong Z Ruan, *London*
Frank I Tovey, *London*
Dhiraj Tripathi, *Birmingham*

Vamsi R Velchuru, *Great Yarmouth*
Nicholas T Ventham, *Edinburgh*
Diego Vergani, *London*
Jack Westwood Winter, *Glasgow*
Terence Wong, *London*
Ling Yang, *Oxford*



United States

Daniel E Abbott, *Cincinnati*
Ghassan K Abou-Alfa, *New York*
Julian Abrams, *New York*
David William Adelson, *Los Angeles*
Jonathan Steven Alexander, *Shreveport*
Tauseef Ali, *Oklahoma City*
Mohamed R Ali, *Sacramento*
Rajagopal N Aravalli, *Minneapolis*
Hassan Ashktorab, *Washington*
Shashi Bala, *Worcester*
Charles F Barish, *Raleigh*
P Patrick Basu, *New York*
Robert L Bell, *Berkeley Heights*
David Bentrem, *Chicago*
Henry J Binder, *New Haven*
Joshua Bleier, *Philadelphia*
Wojciech Blonski, *Johnson City*
Kenneth Boorum, *Corvallis*
Brian Boulay, *Chicago*
Carla W Brady, *Durham*
Kyle E Brown, *Iowa City*
Adeel A Butt, *Pittsburgh*
Weibiao Cao, *Providence*
Andrea Castillo, *Cheney*
Fernando J Castro, *Weston*
Adam S Cheifetz, *Boston*
Xiaoxin Luke Chen, *Durham*
Ramsey Cheung, *Palo Alto*
Parimal Chowdhury, *Little Rock*
Edward John Ciaccio, *New York*
Dahn L Clemens, *Omaha*
Yingzi Cong, *Galveston*
Laura Iris Cosen-Binker, *Boston*
Joseph John Cullen, *Lowa*
Mark J Czaja, *Bronx*
Mariana D Dabeva, *Bronx*
Christopher James Damman, *Seattle*
Isabelle G De Plaen, *Chicago*
Punita Dhawan, *Nashville*
Hui Dong, *La Jolla*
Wael El-Rifai, *Nashville*
Sukru H Emre, *New Haven*
Paul Feuerstadt, *Hamden*
Josef E Fischer, *Boston*
Laurie N Fishman, *Boston*
Joseph Che Forbi, *Atlanta*
Temitope Foster, *Atlanta*
Amy E Foxx-Orenstein, *Scottsdale*
Daniel E Freedberg, *New York*
Shai Friedland, *Palo Alto*
Virgilio George, *Indianapolis*
Ajay Goel, *Dallas*
Oliver Grundmann, *Gainesville*
Stefano Guandalini, *Chicago*
Chakshu Gupta, *St. Joseph*
Grigoriy E Gurvits, *New York*

Xiaonan Han, *Cincinnati*
 Mohamed Hassan, *Jackson*
 Martin Hauer-Jensen, *Little Rock*
 Koichi Hayano, *Boston*
 Yingli Hee, *Atlanta*
 Samuel B Ho, *San Diego*
 Jason Ken Hou, *Houston*
 Lifang Hou, *Chicago*
 K-Qin Hu, *Orange*
 Jamal A Ibdah, *Columbia*
 Robert Thomas Jensen, *Bethesda*
 Huanguang "Charlie" Jia, *Gainesville*
 Rome Jutabha, *Los Angeles*
 Andreas M Kaiser, *Los Angeles*
 Avinash Kambadakone, *Boston*
 David Edward Kaplan, *Philadelphia*
 Randeep Kashyap, *Rochester*
 Rashmi Kaul, *Tulsa*
 Ali Keshavarzian, *Chicago*
 Amir Maqbul Khan, *Marshall*
 Nabeel Hasan Khan, *New Orleans*
 Sahil Khanna, *Rochester*
 Kusum K Kharbanda, *Omaha*
 Hyun Sik Kim, *Pittsburgh*
 Joseph Kim, *Duarte*
 Jae S Kim, *Gainesville*
 Miran Kim, *Providence*
 Timothy R Koch, *Washington*
 Burton I Korelitz, *New York*
 Betsy Kren, *Minneapolis*
 Shiu-Ming Kuo, *Buffalo*
 Michelle Lai, *Boston*
 Andreas Larentzakis, *Boston*
 Edward Wolfgang Lee, *Los Angeles*
 Daniel A Leffler, *Boston*
 Michael Leitman, *New York*
 Suthat Liangpunsakul, *Indianapolis*
 Joseph K Lim, *New Haven*
 Elaine Y Lin, *Bronx*
 Henry C Lin, *Albuquerque*
 Rohit Loomba, *La Jolla*
 James David Luketich, *Pittsburgh*

Li Ma, *Stanford*
 Mohammad F Madhoun, *Oklahoma City*
 Thomas C Mahl, *Buffalo*
 Ashish Malhotra, *Bettendorf*
 Pranoti Mandrekar, *Worcester*
 John Marks, *Wynnewood*
 Wendy M Mars, *Pittsburgh*
 Julien Vahe Matricon, *San Antonio*
 Craig J McClain, *Louisville*
 Tamir Miloh, *Phoenix*
 Ayse Leyla Mindikoglu, *Baltimore*
 Huanbiao Mo, *Denton*
 Klaus Monkemuller, *Birmingham*
 John Morton, *Stanford*
 Adnan Muhammad, *Tampa*
 Michael J Nowicki, *Jackson*
 Patrick I Okolo, *Baltimore*
 Giusepp Orlando, *Winston Salem*
 Natalia A Osna, *Omaha*
 Virendra N Pandey, *Newark*
 Mansour A Parsi, *Cleveland*
 Michael F Picco, *Jacksonville*
 Daniel S Pratt, *Boston*
 Xiaofa Qin, *Newark*
 Janardan K Reddy, *Chicago*
 Victor E Reyes, *Galveston*
 Jon Marc Rhoads, *Houston*
 Giulia Roda, *New York*
 Jean-Francois Armand Rossignol, *Tampa*
 Paul A Rufo, *Boston*
 Madhusudana Girija Sanal, *New York*
 Miguel Saps, *Chicago*
 Sushil Sarna, *Galveston*
 Ann O Scheimann, *Baltimore*
 Bernd Schnabl, *La Jolla*
 Matthew J Schuchert, *Pittsburgh*
 Ekihiro Seki, *La Jolla*
 Chanjuan Shi, *Nashville*
 David Quan Shih, *Los Angeles*
 Shadab A Siddiqi, *Orlando*
 William B Silverman, *Iowa City*
 Shashideep Singhal, *New York*

Bronislaw L Slomiany, *Newark*
 Steven F Solga, *Bethlehem*
 Byoung-Joon Song, *Bethesda*
 Dario Sorrentino, *Roanoke*
 Scott R Steele, *Fort Lewis*
 Branko Stefanovic, *Tallahassee*
 Arun Swaminath, *New York*
 Kazuaki Takabe, *Richmond*
 Naoki Tanaka, *Bethesda*
 Hans Ludger Tillmann, *Durham*
 George Triadafilopoulos, *Stanford*
 John Richardson Thompson, *Nashville*
 Andrew Ukleja, *Weston*
 Miranda AL van Tilburg, *Chapel Hill*
 Gilberto Vaughan, *Atlanta*
 Vijayakumar Velu, *Atlanta*
 Gebhard Wagener, *New York*
 Kasper Saonun Wang, *Los Angeles*
 Xiangbing Wang, *New Brunswick*
 Daoyan Wei, *Houston*
 Theodore H Welling, *Ann Arbor*
 C Mel Wilcox, *Birmingham*
 Jacqueline Lee Wolf, *Boston*
 Laura Ann Woollett, *Cincinnati*
 Harry Hua-Xiang Xia, *East Hanover*
 Wen Xie, *Pittsburgh*
 Guang Yu Yang, *Chicago*
 Michele T Yip-Schneider, *Indianapolis*
 Sam Zakhari, *Bethesda*
 Kezhong Zhang, *Detroit*
 Huiping Zhou, *Richmond*
 Xiao-Jian Zhou, *Cambridge*
 Richard Zubarik, *Burlington*



Venezuela

Miguel Angel Chiurillo, *Barquisimeto*



Vietnam

Van Bang Nguyen, *Hanoi*



TOPIC HIGHLIGHT

- 5459 Natural regression of fibrosis in chronic hepatitis B
Ohkoshi S, Hirono H, Watanabe K, Hasegawa K, Kamimura K, Yano M
- 5467 X region mutations of hepatitis B virus related to clinical severity
Kim H, Lee SA, Kim BJ

REVIEW

- 5479 Avoiding hepatic metastasis naturally: Lessons from the cotton top tamarin (*Saguinus oedipus*)
Tobi M, Thomas P, Ezekwudo D
- 5495 Current status of intragastric balloon for obesity treatment
Kim SH, Chun HJ, Choi HS, Kim ES, Keum B, Jeon YT

MINIREVIEWS

- 5505 Role and mechanisms of action of *Escherichia coli* Nissle 1917 in the maintenance of remission in ulcerative colitis patients: An update
Scaldaferri F, Gerardi V, Mangiola F, Lopetuso LR, Pizzoferrato M, Petito V, Papa A, Stojanovic J, Poscia A, Cammarota G, Gasbarrini A

ORIGINAL ARTICLE

Basic Study

- 5512 Transient receptor potential vanilloid 4-dependent calcium influx and ATP release in mouse and rat gastric epithelia
Mihara H, Suzuki N, Boudaka AA, Muhammad JS, Tominaga M, Tabuchi Y, Sugiyama T
- 5520 Intravoxel incoherent motion diffusion-weighted imaging for monitoring chemotherapeutic efficacy in gastric cancer
Song XL, Kang HK, Jeong GW, Ahn KY, Jeong YY, Kang YJ, Cho HJ, Moon CM
- 5532 MicroRNA-21 promotes phosphatase gene and protein kinase B/phosphatidylinositol 3-kinase expression in colorectal cancer
Sheng WZ, Chen YS, Tu CT, He J, Zhang B, Gao WD
- 5540 Effects of sphincter of Oddi motility on the formation of cholesterol gallstones
Rong ZH, Chen HY, Wang XX, Wang ZY, Xian GZ, Ma BZ, Qin CK, Zhang ZH

Case Control Study

- 5548** Comprehensive risk assessment for early neurologic complications after liver transplantation
Wu SY, Chen TW, Feng AC, Fan HL, Hsieh CB, Chung KP
- 5558** Relationships between cell cycle pathway gene polymorphisms and risk of hepatocellular carcinoma
Nan YL, Hu YL, Liu ZK, Duan FF, Xu Y, Li S, Li T, Chen DF, Zeng XY

Retrospective Cohort Study

- 5568** Hepatitis E in Israel: A nation-wide retrospective study
Erez-Granat O, Lachish T, Daudi N, Shouval D, Schwartz E
- 5578** Comprehensive mutation screening for 10 genes in Chinese patients suffering very early onset inflammatory bowel disease
Xiao Y, Wang XQ, Yu Y, Guo Y, Xu X, Gong L, Zhou T, Li XQ, Xu CD
- 5589** miR-422a is an independent prognostic factor and functions as a potential tumor suppressor in colorectal cancer
Zheng GX, Qu AL, Yang YM, Zhang X, Zhang SC, Wang CX

Retrospective Study

- 5598** Colostomy is a simple and effective procedure for severe chronic radiation proctitis
Yuan ZX, Ma TH, Wang HM, Zhong QH, Yu XH, Qin QY, Wang JP, Wang L

Observational Study

- 5609** Clinical study of anesthetization by dezocine combined with propofol for indolent colonoscopy
Xu BB, Zhao XL, Xu GP

CASE REPORT

- 5616** Successful treatment of ileal ulcers caused by immunosuppressants in two organ transplant recipients
Guo YW, Gu HY, Abassa KK, Lin XY, Wei XQ

ABOUT COVER

Editorial board member of *World Journal of Gastroenterology*, Manuel Morales-Ruiz, PhD, Doctor, Biochemistry and Molecular Genetics Department, Hospital Clinic/IDIBAPS/CIBERehd, Barcelona 08036, Spain

AIMS AND SCOPE

World Journal of Gastroenterology (*World J Gastroenterol*, *WJG*, print ISSN 1007-9327, online ISSN 2219-2840, DOI: 10.3748) is a peer-reviewed open access journal. *WJG* was established on October 1, 1995. It is published weekly on the 7th, 14th, 21st, and 28th each month. The *WJG* Editorial Board consists of 1376 experts in gastroenterology and hepatology from 68 countries.

The primary task of *WJG* is to rapidly publish high-quality original articles, reviews, and commentaries in the fields of gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, hepatobiliary surgery, gastrointestinal oncology, gastrointestinal radiation oncology, gastrointestinal imaging, gastrointestinal interventional therapy, gastrointestinal infectious diseases, gastrointestinal pharmacology, gastrointestinal pathophysiology, gastrointestinal pathology, evidence-based medicine in gastroenterology, pancreatology, gastrointestinal laboratory medicine, gastrointestinal molecular biology, gastrointestinal immunology, gastrointestinal microbiology, gastrointestinal genetics, gastrointestinal translational medicine, gastrointestinal diagnostics, and gastrointestinal therapeutics. *WJG* is dedicated to become an influential and prestigious journal in gastroenterology and hepatology, to promote the development of above disciplines, and to improve the diagnostic and therapeutic skill and expertise of clinicians.

INDEXING/ABSTRACTING

World Journal of Gastroenterology (*WJG*) is now indexed in Current Contents[®]/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch[®]), Journal Citation Reports[®], Index Medicus, MEDLINE, PubMed, PubMed Central, Digital Object Identifier, and Directory of Open Access Journals. The 2015 edition of Journal Citation Reports[®] released by Thomson Reuters (ISI) cites the 2015 impact factor for *WJG* as 2.787 (5-year impact factor: 2.848), ranking *WJG* as 38 among 78 journals in gastroenterology and hepatology (quartile in category Q2).

FLYLEAF

I-IX Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Shuai Ma*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Ze-Mao Gong*
Proofing Editorial Office Director: *Jin-Lei Wang*

NAME OF JOURNAL
World Journal of Gastroenterology

ISSN
ISSN 1007-9327 (print)
ISSN 2219-2840 (online)

LAUNCH DATE
October 1, 1995

FREQUENCY
Weekly

EDITORS-IN-CHIEF
Damian Garcia-Olmo, MD, PhD, Doctor, Professor, Surgeon, Department of Surgery, Universidad Autonoma de Madrid; Department of General Surgery, Fundacion Jimenez Diaz University Hospital, Madrid 28040, Spain

Stephen C Strom, PhD, Professor, Department of Laboratory Medicine, Division of Pathology, Karolinska Institutet, Stockholm 141-86, Sweden

Andrzej S Tarnawski, MD, PhD, DSc (Med), Professor of Medicine, Chief Gastroenterology, VA

Long Beach Health Care System, University of California, Irvine, CA, 5901 E. Seventh Str., Long Beach, CA 90822, United States

EDITORIAL OFFICE
Jin-Lei Wang, Director
Xiu-Xia Song, Vice Director
World Journal of Gastroenterology
Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China
Telephone: +86-10-59080039
Fax: +86-10-85381893
E-mail: editorialoffice@wjgnet.com
Help Desk: <http://www.wjgnet.com/esp/helpdesk.aspx>
<http://www.wjgnet.com>

PUBLISHER
Baishideng Publishing Group Inc
8226 Regency Drive,
Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.wjgnet.com/esp/helpdesk.aspx>
<http://www.wjgnet.com>

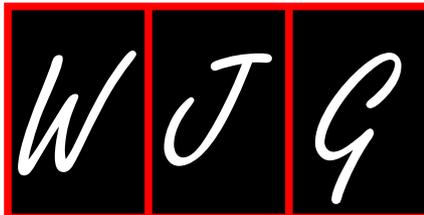
PUBLICATION DATE
June 28, 2016

COPYRIGHT
© 2016 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS
Full instructions are available online at http://www.wjgnet.com/bpg/g_info_20160116143427.htm

ONLINE SUBMISSION
<http://www.wjgnet.com/esp/>



2016 Hepatitis B virus: Global view

Natural regression of fibrosis in chronic hepatitis B

Shogo Ohkoshi, Haruka Hirono, Kazuhiko Watanabe, Katsuhiko Hasegawa, Kenya Kamimura, Masahiko Yano

Shogo Ohkoshi, Haruka Hirono, Kazuhiko Watanabe, Katsuhiko Hasegawa, Department of Internal Medicine, School of Life Dentistry at Niigata, The Nippon Dental University, Niigata 951-8580, Japan

Kenya Kamimura, Masahiko Yano, Division of Gastroenterology and Hepatology, Graduate School of Medical and Dental Sciences Niigata University, Niigata 951-8520, Japan

Author contributions: Ohkoshi S wrote the paper; Yano M and Kamimura K collected the data; Hirono H, Watanabe K and Hasegawa K had critical discussions regarding the study and manuscript with Ohkoshi S.

Supported by A Grant-in-Aid for Scientific Research (C) (25461012 to Ohkoshi S) from the Japan Society for the Promotion of Science.

Conflict-of-interest statement: The authors do not have any commercial affiliation or consultancy that could be construed as a conflict of interest.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Shogo Ohkoshi, MD, PhD, Department of Internal Medicine, School of Life Dentistry at Niigata, The Nippon Dental University, 1-8 Hamaura-Cho, Chuo-ku, Niigata 951-8580, Japan. okoshi@ngt.ndu.ac.jp
Telephone: +81-25-2118243
Fax: +81-25-2671582

Received: March 5, 2016
Peer-review started: March 7, 2016
First decision: April 14, 2016
Revised: April 20, 2016
Accepted: May 4, 2016

Article in press: May 4, 2016
Published online: June 28, 2016

Abstract

The fibrosis of liver cirrhosis was considered to be irreversible before the anti-viral drugs showed that it is reversible when they lead to continuous suppression of viral replication and inflammation. However, several reports previously showed that fibrosis of type B liver cirrhosis was almost completely absorbed after the natural remission of chronic inflammation. This phenomenon might not be limited to exceptional patients, but rather occur commonly, considering the dynamic clinical features of chronic hepatitis B (CHB), where inactive carrier stage normally follows aggravation of hepatitis and progression of fibrosis at the time of HBeAg seroconversion. Thus, fibrosis levels of CHB as a hepatocellular carcinoma (HCC)-surveillance marker, particularly those of the inactive stage, could be underestimated, because some of them might have been (pre)cirrhotic in the past and recovered with the natural regression of fibrosis. We argue that cirrhosis-induced HCC mechanisms, rather than direct action of viral genome, may be more common than generally considered in CHB patients. This may have some impact on reconsidering the surveillance rationale for HCC in CHB, from where advanced HCCs tended to be missed. In addition, a molecular marker to assess the cancer-prone characteristics of the liver will definitely be needed to resolve the issue.

Key words: Chronic hepatitis B; Cirrhosis; Spontaneous remission; Regression of fibrosis; Occult hepatitis B infection; Hepatocellular carcinoma surveillance of hepatitis B virus

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: The fibrosis of liver cirrhosis may be reversible. Regression of fibrosis in hepatitis B virus (HBV) patients with (pre)cirrhosis might be a more common phenomenon than generally considered. This might cause the underestimation of fibrosis levels in chronic hepatitis B, suggesting a difficulty with the surveillance system of HBV-hepatocellular carcinoma (HCC). That is, some HCC patients with non-cirrhotic liver might have been cirrhotic in the past, after which spontaneous regression of fibrosis occurred. Cirrhosis-HCC mechanisms, compared to the direct action of the viral genome, might be more prevalent than generally considered in HBV patients.

Ohkoshi S, Hirono H, Watanabe K, Hasegawa K, Kamimura K, Yano M. Natural regression of fibrosis in chronic hepatitis B. *World J Gastroenterol* 2016; 22(24): 5459-5466 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5459.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5459>

INTRODUCTION

Continuous inflammation of the liver induces deposition of extracellular matrix (ECM) and results in liver cirrhosis. Although several reports showed regression of fibrosis in animal models with liver cirrhosis^[1,2], and in human case reports^[3-5], established fibrosis of cirrhosis was generally considered to be irreversible^[6,7]. However, reversibility of fibrosis in the liver became commonly-known recently because elimination of viruses with anti-hepatitis B virus (HBV) or hepatitis C virus (HCV) drugs caused regression of fibrosis even in cirrhotic cases^[8,9].

The natural history of chronic HBV infection pursues a dynamic process caused by the conflict between virus and immune system, which results in the seroconversion of HBeAg to anti-HBe, followed by that of HBsAg to anti-HBs^[10]. Long-standing liver injury in the process of HBeAg-seroconversion induces particularly rapid progression of fibrosis that frequently results in cirrhosis. However, due to the dynamic course of the disease, stable clinical remission with low HBV DNA and transaminase levels follows, and the majority of patients become inactive carriers. Natural regression of fibrosis induced by this clinical remission has been reported^[3,11].

In general, hepatocellular carcinoma (HCC) with HBV arises more frequently in non-cirrhotic liver than in HCV infection^[12,13]. This is considered mainly due to the direct mutagenesis effect of the HBV genome that integrates into the host genome or the direct hepatocarcinogenic action of viral genes such as hepatitis B virus X (HBx)^[14]. However, if HCCs have arisen in the liver in which fibrosis naturally regressed from advanced liver fibrosis, they might be misinterpreted as having arisen from a non-cirrhotic liver.

Consequently, it should always be kept in mind that fibrosis levels of chronic hepatitis B (CHB) always have the possibility of being underestimated in terms of HCC-risk, because they might have regressed from a more advanced stage of fibrosis^[15]. We attempt to spotlight the natural regression of fibrosis in chronic HBV infection and re-evaluate the clinical background of HBV-induced HCC in this review.

CHRONIC HBV DISEASE AND PROGRESSION OF FIBROSIS

About 350 million people in the world have chronic HBV infection^[16]. Chronic HBV infection is a significant cause of liver cirrhosis, which bears a high risk of HCC. The annual rate of development of HCC has been reported to be 10% to 17% in HBV-induced liver cirrhosis^[17]. In general, fibrosis levels correlate well with the risk of HCC, as in the case with HCV infection.

The immunological attack against hepatocytes infected with HBV induces a dynamic clinical course which is represented by seroconversion of HBeAg to anti-HBe. Almost no or very mild liver injury is observed in the HBeAg-positive immunotolerant phase^[18]. The severity of liver injury and the progression of fibrosis depend on how long and severe the immunological attacks continue in the phase of HBeAg seroconversion. A rapid transition to anti-HBe normally results in a silent clinical course, referred to as the inactive carrier state, while a difficult transition that is accompanied by aggravation of chronic hepatitis result in liver cirrhosis or HBeAb-positive chronic active hepatitis, which frequently bears core-promoter mutations. Importantly, the majority of patients become inactive carriers afterwards, irrespective of their clinical course^[10,19].

MECHANISM OF HBV-INDUCED HCC AND CLINICAL BACKGROUND CHARACTERISTICS

Chronic inflammation induces a progression of fibrosis resulting in liver cirrhosis in which HCC frequently develops. This cirrhosis-HCC mechanism is common to both HBV and HCV infections and is considered to play the main role in hepatocarcinogenesis, to which a process of accumulation of genetic mutations and epigenetic events contributes. In addition to this common carcinogenetic mechanism, each virus infection has its own carcinogenic mechanism.

On the other hand, HBV, which is related to retroviruses, integrates into the host genome during the process of replication and acts as a mutagen. In addition, the HBx gene, which is a pleiotropic transactivator of many genes, has been known to be directly involved in hepatocarcinogenesis^[20]. Because "cirrhosis to HCC" (indirect) and these

Table 1 A comparison of clinic-pathological data between type-B and type-C hepatocellular carcinoma

	Type B (n = 97)	Type C (n = 81)	P value
Age (yr)	56.1 ± 11.9	66.7 ± 9.8	< 0.0001
Sex (M:F)	71.1%:28.9%	60.5%:39.5%	0.18
AST (IU/L)	82.2 ± 98.6	81.8 ± 44.1	0.18
ALT (IU/L)	67.7 ± 70.9	83.2 ± 56.9	0.11
T.Bil (mg/dL)	1.2 ± 1.1	1.0 ± 0.6	0.36
Alb (g/dL)	4.0 ± 0.6	3.9 ± 0.5	0.22
Plt (/mm ³)	13.9 ± 8.1	10.8 ± 5.5	< 0.005
Histology			
F1:2:3:4	2:20:3:32	1:6:8:21	< 0.005
A1:2:3	17:25:12	1:14:17	< 0.005
Clinical stage (I : II : III : IV)	12:29:25:31	15:32:29:5	< 0.0005

direct mechanisms are not mutually exclusive, it is unknown how much a role each mechanism plays in real hepatocarcinogenesis, especially in individual HCC patients with variable clinical backgrounds.

DIFFICULTY OF HCC SURVEILLANCE IN CHRONIC HBV PATIENTS

Because the carcinogenic processes of HBV and HCV are different, the rationale to perform effective cancer surveillance should be confirmed based on the difference in the specific clinical courses of these infections. Age, sex, fibrosis levels, the presence of HBeAg, HBV viral load, HBsAg levels accounts for the risks of HBV-HCC^[21]. In reports from Japan, HCC patients with HBV tended to be younger and have advanced tumor when diagnosed, as well as lower fibrosis levels with good liver function when compared to those with HCV^[22,23]. Our result also showed that patients with HBV were younger, with lower fibrosis levels and higher platelet counts than those with HCV (Table 1). Notably, the clinical stages of HBV-HCC were more advanced. These clinical features suggest that cancer surveillance of HBV is much more difficult than that of HCV infection, where continuous elevation of transaminase values induces gradually increasing fibrosis levels, and a low platelet count may be associated with advanced fibrosis levels^[24]. In general, a direct hepatocarcinogenic mechanism of HBV, such as the integration of the genome or a direct role of an HBV gene such as HBx, may be considered to be involved in the occurrence of HCC in these “unexpected” patients. However, our question is “Is this the full explanation?”

It seems very paradoxical that HBeAg-positive carriers in the immunotolerant phase with high viral loads and little inflammation, who are supposed have abundant viral gene expression and integration event, seldom suffer from HCC^[18]. On the other hand, most HCC patients who were younger than 20 years old were already seroconverted to anti-HBe, and the majority of them were cirrhotic^[25-27]. Thus, the cirrhosis to HCC mechanism might play a central role even in

these young HCC patients. These facts render the HCC surveillance system for HBV more complex.

REVERSIBILITY OF FIBROSIS WITH LIVER CIRRHOSIS WITH ANTI-HBV TREATMENT AND NATURAL COURSE

The generally accepted idea was that the fibrosis of liver cirrhosis is an irreversible process, and established fibrosis could not regress. However, the development of innovative anti-viral drugs for HBV and HCV changed this long-lasting concept^[8]. Fibrosis is reversed when inflammation is suppressed by effective anti-viral agents, and even cirrhosis can be cured. Dienstag *et al.*^[28] reported that the fibrosis of cirrhosis was reversed during 3.5 years lamivudine treatment. Several reports also confirmed the improvement of fibrosis levels with anti-HBV drugs^[29,30].

However, the regression of fibrosis occurs even in the natural course of CHB, where remission of inflammation usually follows the aggravation of hepatitis and progression of fibrosis that occurred during the HBeAg-positive stage^[31]. In the past, several reports showed the improvement of fibrosis during the natural course of CHB. Fong *et al.*^[3] showed the improvement of histology comparing before and after the seroclearance of HBsAg. Waneless *et al.*^[32] reported that most cirrhotic livers showed some findings indicating the regression of fibrosis, irrespective of etiology. Natural regression of fibrosis consists of three steps, that is, thinning of fibrous septa, regeneration of hepatocytes and recovery of acinal structure. The regression of fibrosis occurs at a particularly early stage of improvement^[6].

Recently, Jang *et al.*^[33] reported improvement of the long-term prognosis with anti-HBV drugs for patients with HBV-cirrhosis. However, suppression of HCC development was not observed in these cirrhotic patients, while it has been confirmed in patients with chronic hepatitis^[8]. These results strongly suggest that liver cirrhosis still provides a strong risk for HCC, even after the eradication of virus and suppression of

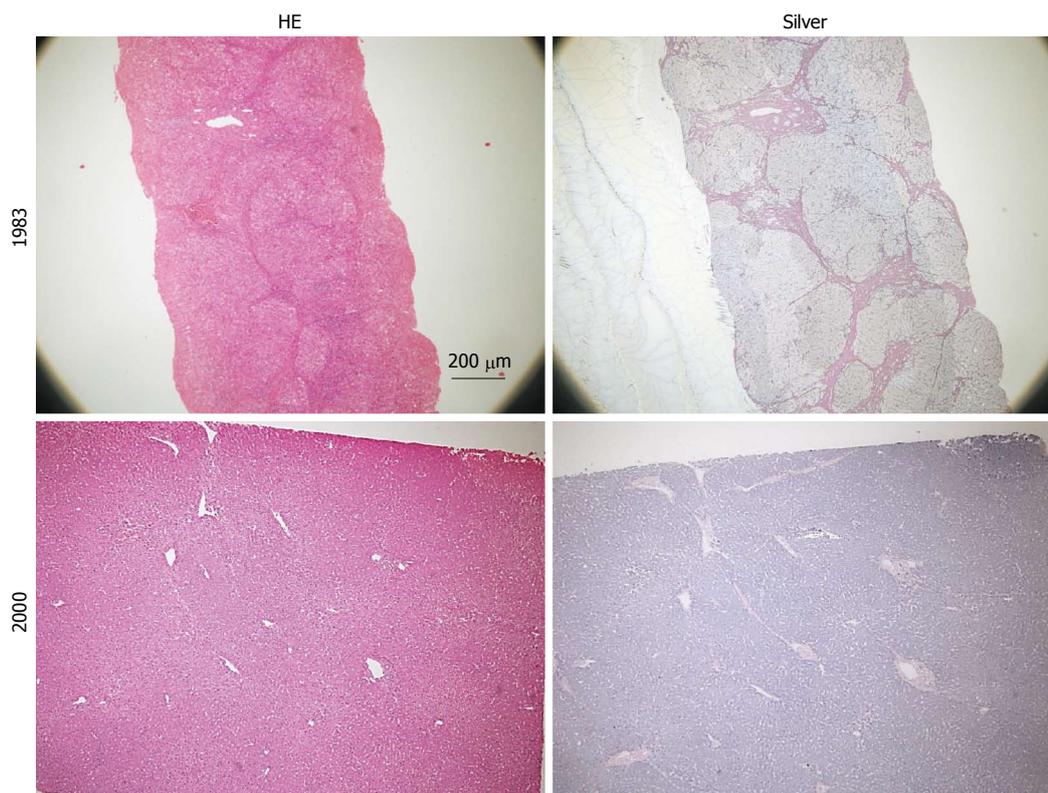


Figure 1 Liver histology of a patient obtained in 1983 (58 year old, needle biopsy) and 2000 (surgical resection of hepatocellular carcinoma). Slides stained with hematoxylin-eosin are shown on the left and those stained with silver are on the right. Marked reduction of fibrosis with improvement of inflammation is observed (This patient was patient 1 in our previous report^[4]).

inflammation.

THE LESSONS FROM THE HCC CASES THAT OCCURRED AFTER HBSAG SEROCLEARANCE

We previously reported several cases with HCC that developed after seroconversion from HBsAg to anti-HBs^[4,34]. They progressed to liver cirrhosis or precirrhosis when they had HBeAg-positive chronic hepatitis several decades earlier and became inactive carriers with low viral load and long-standing normal transaminase levels after the seroconversion to anti-HBe. HCC developed further after the seroconversion to anti-HBs. Hepatic fibrosis was dramatically reversed and found to become only thin septa. Changes in liver histology of one patient^[4] are shown in Figure 1. Similarly, Bortolotti *et al*^[5] reported 2 pediatric patients with HBV-induced liver cirrhosis, whose fibrosis was almost completely resolved after normalization of transaminase levels. Similar to our cases, these results suggested that fibrosis of cirrhosis could be dramatically resolved when viral replication and inflammation were suppressed^[31,35]. These cases, considering the dynamic natural course of HBV infection, give us some important suggestions about the clinical background characteristics of HBV-induced

hepatocarcinogenesis.

First, as with anti-HBV treatment, these cases are typical examples showing that advanced fibrosis could be absorbed even in the natural course of CHB. After HBeAg-positive chronic hepatitis progresses to liver cirrhosis, the majority of patients follow the clinical course of long-standing remission with a low viral load. Considering this typical clinical course of CHB, natural regression of fibrosis might not be an exceptional phenomenon limited to these case reports, but it might occur commonly. This also suggests that some population of HBV-HCC patients with non-cirrhotic liver might have regressed fibrosis that was once cirrhotic or pre-cirrhotic. Consequently, our hypothesis is that the incidence and role of cirrhosis-derived HCC might be underestimated in HBV-HCC, and the roles of direct carcinogenesis due to HBV integration or viral gene function with a slight involvement of necroinflammation in non-cirrhotic HCC are overestimated.

HBV-cirrhosis is "macronodular" and has thinner septa than HCV^[32]. Micronodular cirrhosis changes to macronodular with the suppression of inflammation^[36]. This morphological observation might reflect the clinical course of these viruses, where HCV takes a continuous progressive course, and HBV is characterized by the alternating increases and normalization of transaminase levels.

Another important suggestion relates to the assess-

ment of fibrosis in CHB patients for the establishment of an effective HCC surveillance system. Recently, not only liver biopsy which is a gold standard for the evaluation of fibrosis, but a handful of methods to assess fibrosis levels have been used clinically. However, these methods may not discriminate these cases of "regressed fibrosis from cirrhosis", which might have more risk of HCC than "ongoing fibrosis". Thus, it might be possible to underestimate the risk of HCC in these patients with regressed fibrosis levels using these current methods. Indeed, needle biopsy might give the diagnosis of an almost normal liver with completely regressed fibrosis^[32].

Cirrhosis-HCC mechanisms can be glimpsed in studies that examined the incidence of HCC after the seroclearance of HBsAg, which normally has a good prognosis^[10,37,38]. Yuen *et al.*^[39] reported that the risk factors of HCC at this stage were the presence of liver cirrhosis and old age. Liver cirrhosis was also counted as a significant factor for the seroclearance of HBsAg^[40]. This indicates that the strong immune reaction that caused liver cirrhosis eventually contributed to the seroclearance of HBsAg, providing some risk of HCC.

Similar suggestions could be applied to HCC cases with occult HBV infection (OHB). Some patients might have overt chronic HBV infection before entering the stage of OHB^[41]. These patients might have "regressed fibrosis from advanced fibrosis" during the process of becoming seronegative for HBsAg. They can easily be missed during clinical surveillance of HCC, unless they were recognized as HBV carriers, possibly in early life^[15]. Integration of HBV DNA or the direct action of an HBV gene such as HBx from long-standing viral replication is considered to play a role in the occurrence of HCC with OHB^[42]. However, cirrhosis-HCC mechanism might play a central role in more number of patients with OHB-HCC than generally considered.

Liver biopsy has been a gold standard of evaluation for liver fibrosis^[43]. Non-invasive methods for evaluation of fibrosis are widely used in clinical settings. They have been primarily used for the assessment of HCV-related fibrosis, and their application to HBV-induced fibrosis has always been under constant debate^[44]. Transient elastography (TE) is one of the most reliable recent modalities to assess the fibrosis level without biopsy^[45]. Several reports evaluated the efficacy of TE in both CHB and CHC patients and reported that diagnostic accuracy was almost equal between them^[46,47]. However, a lower liver stiffness (LS) cut-off in CHB than in CHC in the advanced fibrosis stage was reported^[48]. The interpretation of this was that fibrous septa were thinner in CHB than in CHC. Moreover, because CHB tended to have macronodular cirrhosis, TE waves go through liver parenchyma and produce a low stiffness value^[49]. Likewise, Castéra *et al.*^[50] reported that the cut off values of TE in cirrhosis were higher in HCV-LC than that of HBV-LC and this difference came from

the high prevalence of macronodular LC in HBV. Thus, although fibrosis markers are considered to be useful in the evaluation of fibrosis levels of CHB in general^[49], there have been no studies attempting to assess the fibrosis levels following CHB patients from aggravation of hepatitis to the inactive stage.

APPLICATION OF MOLECULAR ANALYSIS TO THE "HCC FROM REGRESSED FIBROSIS"

Innovative advances of molecular analysis recently allow comprehensive cancer genome analysis in a short time, resulting in the accumulation of an overwhelming body of information about the cancer genome^[51]. Schulze *et al.*^[52] analyzed the whole exon of 243 HCC tissues and reported that genomic pathways centered on three signaling abnormalities, β -catenin (CTNNB1), TP53, and AXIN1-related. They also suggested that they can distinguish HCC that arose from cirrhosis from those derived from non-cirrhosis using the difference in incidence of CTNNB1 TP53, a mutation of telomerase reverse transcriptase (TERT). In addition, mutations of TERT promoter have been noted as a characteristic genomic change of HBV-HCC, in which integration of the HBV genome frequently occurs^[53,54]. It is interesting to know whether HCC that arose from "liver with regressed fibrosis from cirrhosis" is close to a cirrhotic or a non-cirrhotic pattern.

Telomere shortening is a general genetic marker of liver cirrhosis causing senescence of hepatocytes that is associated with the development of HCC^[55-57]. Recently, TERT promoter mutations were observed already in premalignant nodules in cirrhotic liver^[58]. In addition, Hartmann *et al.*^[59] reported that TERT gene mutations were frequently detected in liver cirrhosis irrespective of etiology, suggesting that they may be used to evaluate cancer-prone characteristics of liver. This result might be applicable in the evaluation of whether the liver with regressed fibrosis has oncogenic potential close to that of real cirrhosis, helping the understandings of the carcinogenic potential of that state. Such molecular markers that can assess cancer-prone characteristics of cirrhotic or non-cirrhotic liver are much needed to prove that a cirrhosis-HCC mechanism might be involved in the development of HCC than currently considered, even in low-fibrosis cases.

CONCLUSION

The clinical course of chronic HBV infection is dynamic and complex, with changes of serological markers, which is distinct from HCV infection that takes a more continuous clinical course. Because of this clinical profile, long-standing remission after established cirrhosis, result in the regression of fibrosis, where only thin septa are recognized. On the other hand, the integration of

the HBV genome or a direct gene function, such as HBx, may play an important role in so-called direct hepatocarcinogenesis. HCCs from regressed fibrosis and those *via* direct hepatocarcinogenesis might be categorized together into "HCC from non-cirrhotic liver". Thus, these mechanisms render the assessment of fibrosis as a risk factor for HCC very complex. Therefore, there is an urgent need to establish effective cancer surveillance system that addresses these complex issues. In addition, molecular markers to evaluate the risk of HCC in non-cirrhotic as well as cirrhotic liver should be clarified in a future analysis.

REFERENCES

- Iredale JP, Thompson A, Henderson NC. Extracellular matrix degradation in liver fibrosis: Biochemistry and regulation. *Biochim Biophys Acta* 2013; **1832**: 876-883 [PMID: 23149387 DOI: 10.1016/j.bbdis.2012.11.002]
- Issa R, Zhou X, Constandinou CM, Fallowfield J, Millward-Sadler H, Gaca MD, Sands E, Suliman I, Trim N, Knorr A, Arthur MJ, Benyon RC, Iredale JP. Spontaneous recovery from micronodular cirrhosis: evidence for incomplete resolution associated with matrix cross-linking. *Gastroenterology* 2004; **126**: 1795-1808 [PMID: 15188175]
- Fong TL, Di Bisceglie AM, Gerber MA, Waggoner JG, Hoofnagle JH. Persistence of hepatitis B virus DNA in the liver after loss of HBsAg in chronic hepatitis B. *Hepatology* 1993; **18**: 1313-1318 [PMID: 8244254]
- Okoshi S, Igarashi M, Suda T, Iwamatsu H, Watanabe K, Ishihara K, Ogata N, Nomoto M, Takahashi T, Ichida T, Asakura H, Nihei K, Kurosaki I. Remote development of hepatocellular carcinoma in patients with liver cirrhosis type B serologically cured for HBs antigenemia with long-standing normalization of ALT values. *Dig Dis Sci* 2002; **47**: 2002-2006 [PMID: 12353845]
- Bortolotti F, Guido M, Cadrobbi P, Crivellaro C, Bartolacci S, Rugge M, Gatta A. Spontaneous regression of hepatitis B virus-associated cirrhosis developed in childhood. *Dig Liver Dis* 2005; **37**: 964-967 [PMID: 16249128 DOI: 10.1016/j.dld.2005.04.030]
- Bedossa P. Reversibility of hepatitis B virus cirrhosis after therapy: who and why? *Liver Int* 2015; **35** Suppl 1: 78-81 [PMID: 25529091 DOI: 10.1111/liv.12710]
- Calvaruso V, Craxi A. Regression of fibrosis after HBV antiviral therapy. Is cirrhosis reversible? *Liver Int* 2014; **34** Suppl 1: 85-90 [PMID: 24373083 DOI: 10.1111/liv.12395]
- Liaw YF. Impact of therapy on the outcome of chronic hepatitis B. *Liver Int* 2013; **33** Suppl 1: 111-115 [PMID: 23286854 DOI: 10.1111/liv.12057]
- Shiratori Y, Imazeki F, Moriyama M, Yano M, Arakawa Y, Yokosuka O, Kuroki T, Nishiguchi S, Sata M, Yamada G, Fujiyama S, Yoshida H, Omata M. Histologic improvement of fibrosis in patients with hepatitis C who have sustained response to interferon therapy. *Ann Intern Med* 2000; **132**: 517-524 [PMID: 10744587]
- McMahon BJ. The natural history of chronic hepatitis B virus infection. *Hepatology* 2009; **49**: S45-S55 [PMID: 19399792 DOI: 10.1002/hep.22898]
- Sharma SK, Saini N, Chwla Y. Hepatitis B virus: inactive carriers. *Viol J* 2005; **2**: 82 [PMID: 16191199 DOI: 10.1186/1743-422x-2-82]
- Lok AS. Hepatitis B: liver fibrosis and hepatocellular carcinoma. *Gastroenterol Clin Biol* 2009; **33**: 911-915 [PMID: 19577871 DOI: 10.1016/j.gcb.2009.06.001]
- Bralet MP, Régimbeau JM, Pineau P, Dubois S, Loas G, Degos F, Valla D, Belghiti J, Degott C, Terris B. Hepatocellular carcinoma occurring in nonfibrotic liver: epidemiologic and histopathologic analysis of 80 French cases. *Hepatology* 2000; **32**: 200-204 [PMID: 10915724 DOI: 10.1053/jhep.2000.9033]
- Neuveut C, Wei Y, Buendia MA. Mechanisms of HBV-related hepatocarcinogenesis. *J Hepatol* 2010; **52**: 594-604 [PMID: 20185200 DOI: 10.1016/j.jhep.2009.10.033]
- Bortolotti F, Guido M, Bartolacci S, Cadrobbi P, Crivellaro C, Noventa F, Morsica G, Moriondo M, Gatta A. Chronic hepatitis B in children after e antigen seroclearance: final report of a 29-year longitudinal study. *Hepatology* 2006; **43**: 556-562 [PMID: 16496323 DOI: 10.1002/hep.21077]
- Trépo C, Chan HL, Lok A. Hepatitis B virus infection. *Lancet* 2014; **384**: 2053-2063 [PMID: 24954675 DOI: 10.1016/s0140-6736(14)60220-8]
- Fattovich G, Bortolotti F, Donato F. Natural history of chronic hepatitis B: special emphasis on disease progression and prognostic factors. *J Hepatol* 2008; **48**: 335-352 [PMID: 18096267 DOI: 10.1016/j.jhep.2007.11.011]
- Hui CK, Leung N, Yuen ST, Zhang HY, Leung KW, Lu L, Cheung SK, Wong WM, Lau GK. Natural history and disease progression in Chinese chronic hepatitis B patients in immune-tolerant phase. *Hepatology* 2007; **46**: 395-401 [PMID: 17628874 DOI: 10.1002/hep.21724]
- Hsu YS, Chien RN, Yeh CT, Sheen IS, Chiou HY, Chu CM, Liaw YF. Long-term outcome after spontaneous HBeAg seroconversion in patients with chronic hepatitis B. *Hepatology* 2002; **35**: 1522-1527 [PMID: 12029639 DOI: 10.1053/jhep.2002.33638]
- Feitelson MA, Lee J. Hepatitis B virus integration, fragile sites, and hepatocarcinogenesis. *Cancer Lett* 2007; **252**: 157-170 [PMID: 17188425 DOI: 10.1016/j.canlet.2006.11.010]
- Wong VW, Janssen HL. Can we use HCC risk scores to individualize surveillance in chronic hepatitis B infection? *J Hepatol* 2015; **63**: 722-732 [PMID: 26026875 DOI: 10.1016/j.jhep.2015.05.019]
- Ishikawa T. Clinical features of hepatitis B virus-related hepatocellular carcinoma. *World J Gastroenterol* 2010; **16**: 2463-2467 [PMID: 20503445 DOI: 10.3748/wjg.v16.i20.2463]
- Tanizaki H, Ryu M, Kinoshita T, Kawano N, Konishi M, Cho A, Nakatsura T, Natsume T, Takahashi S, Sugita M, Izuishi K, Yoshino M, Furuse J, Iwasaki M, Tsubono Y. Comparison of clinical features and survival in patients with hepatitis B and C virus-related hepatocellular carcinoma. *Jpn J Clin Oncol* 1997; **27**: 67-70 [PMID: 9152792]
- Shiratori Y, Omata M. Predictors of the efficacy of interferon therapy for patients with chronic hepatitis C before and during therapy: how does this modify the treatment course? *J Gastroenterol Hepatol* 2000; **15** Suppl: E141-E151 [PMID: 10921398]
- Komatsu H, Inui A. Hepatitis B virus infection in children. *Expert Rev Anti Infect Ther* 2015; **13**: 427-450 [PMID: 25724218 DOI: 10.1586/14787210.2015.1019867]
- Tazawa Y, Nishinomiya F, Noguchi H, Tsuchiya S, Hayashi Y, Abukawa D, Watabe M, Nakagawa M, Imaizumi M, Ohi R. Hepatocellular carcinoma in children with hepatitis B surface antigen. *Tohoku J Exp Med* 1992; **167**: 47-55 [PMID: 1333650]
- Zhang XF, Liu XM, Wei T, Liu C, Li MX, Long ZD, Lv Y. Clinical characteristics and outcome of hepatocellular carcinoma in children and adolescents. *Pediatr Surg Int* 2013; **29**: 763-770 [PMID: 23794023 DOI: 10.1007/s00383-013-3334-4]
- Dienstag JL, Goldin RD, Heathcote EJ, Hann HW, Woessner M, Stephenson SL, Gardner S, Gray DF, Schiff ER. Histological outcome during long-term lamivudine therapy. *Gastroenterology* 2003; **124**: 105-117 [PMID: 12512035 DOI: 10.1053/gast.2003.50013]
- Chang TT, Liaw YF, Wu SS, Schiff E, Han KH, Lai CL, Safadi R, Lee SS, Halota W, Goodman Z, Chi YC, Zhang H, Hindes R, Iloeje U, Beebe S, Kreter B. Long-term entecavir therapy results in the reversal of fibrosis/cirrhosis and continued histological improvement in patients with chronic hepatitis B. *Hepatology* 2010; **52**: 886-893 [PMID: 20683932 DOI: 10.1002/hep.23785]
- Marcellin P, Gane E, Buti M, Afdhal N, Sievert W, Jacobson IM, Washington MK, Germanidis G, Flaherty JF, Aguilar Schall R, Bornstein JD, Kitrinis KM, Subramanian GM, McHutchison JG, Heathcote EJ. Regression of cirrhosis during treatment with

- tenofovir disoproxil fumarate for chronic hepatitis B: a 5-year open-label follow-up study. *Lancet* 2013; **381**: 468-475 [PMID: 23234725 DOI: 10.1016/s0140-6736(12)61425-1]
- 31 **Shah U**, Kelly D, Chang MH, Fujisawa T, Heller S, González-Peralta RP, Jara P, Mieli-Vergani G, Mohan N, Murray KF. Management of chronic hepatitis B in children. *J Pediatr Gastroenterol Nutr* 2009; **48**: 399-404 [PMID: 19322053 DOI: 10.1097/MPG.0b013e318197196e]
- 32 **Wanless IR**, Nakashima E, Sherman M. Regression of human cirrhosis. Morphologic features and the genesis of incomplete septal cirrhosis. *Arch Pathol Lab Med* 2000; **124**: 1599-1607 [PMID: 11079009 DOI: 10.1043/0003-9985(2000)124<1599:roh>2.0.co;2]
- 33 **Jang JW**, Choi JY, Kim YS, Woo HY, Choi SK, Lee CH, Kim TY, Sohn JH, Tak WY, Han KH. Long-term effect of antiviral therapy on disease course after decompensation in patients with hepatitis B virus-related cirrhosis. *Hepatology* 2015; **61**: 1809-1820 [PMID: 25627342 DOI: 10.1002/hep.27723]
- 34 **Tsuboi Y**, Ohkoshi S, Yano M, Suzuki K, Tsubata SS, Ishihara K, Ichida T, Sugitani S, Shibazaki K, Aoyagi Y. Common clinicopathological features of the patients with chronic hepatitis B virus infection who developed hepatocellular carcinoma after seroconversion to anti-HBs—a consideration of the pathogenesis of HBV-induced hepatocellular carcinoma and a strategy to inhibit it. *Hepatogastroenterology* 2006; **53**: 110-114 [PMID: 16506387]
- 35 **Bortolotti F**, Guido M. Reversal of liver cirrhosis: a desirable clinical outcome and its pathogenic background. *J Pediatr Gastroenterol Nutr* 2007; **44**: 401-406 [PMID: 17414134 DOI: 10.1097/MPG.0b013e318032069a]
- 36 **Fauerholdt L**, Schlichting P, Christensen E, Poulsen H, Tygstrup N, Juhl E. Conversion of micronodular cirrhosis into macronodular cirrhosis. *Hepatology* 1983; **3**: 928-931 [PMID: 6629323]
- 37 **Liaw YF**. Natural history of chronic hepatitis B virus infection and long-term outcome under treatment. *Liver Int* 2009; **29** Suppl 1: 100-107 [PMID: 19207972 DOI: 10.1111/j.1478-3231.2008.01941.x]
- 38 **Liu J**, Yang HI, Lee MH, Lu SN, Jen CL, Wang LY, You SL, Iloeje UH, Chen CJ. Incidence and determinants of spontaneous hepatitis B surface antigen seroclearance: a community-based follow-up study. *Gastroenterology* 2010; **139**: 474-482 [PMID: 20434450 DOI: 10.1053/j.gastro.2010.04.048]
- 39 **Yuen MF**, Wong DK, Sablon E, Tse E, Ng IO, Yuan HJ, Siu CW, Sander TJ, Bourne EJ, Hall JG, Condeary LD, Lai CL. HBsAg seroclearance in chronic hepatitis B in the Chinese: virological, histological, and clinical aspects. *Hepatology* 2004; **39**: 1694-1701 [PMID: 15185311 DOI: 10.1002/hep.20240]
- 40 **Liaw YF**, Sheen IS, Chen TJ, Chu CM, Pao CC. Incidence, determinants and significance of delayed clearance of serum HBsAg in chronic hepatitis B virus infection: a prospective study. *Hepatology* 1991; **13**: 627-631 [PMID: 2010157]
- 41 **Huang Y**, Wang W, Chen Y, Huang Y, Zhang J, He S, Tan Y, Qiang F, Li A, Røe OD, Wang S, Zhou Y, Zhou J. The opposite prognostic significance of nuclear and cytoplasmic p21 expression in resectable gastric cancer patients. *J Gastroenterol* 2014; **49**: 1441-1452 [PMID: 24127074 DOI: 10.1007/s00535-013-0900-4]
- 42 **Pollicino T**, Saitta C. Occult hepatitis B virus and hepatocellular carcinoma. *World J Gastroenterol* 2014; **20**: 5951-5961 [PMID: 24876718 DOI: 10.3748/wjg.v20.i20.5951]
- 43 **Bravo AA**, Sheth SG, Chopra S. Liver biopsy. *N Engl J Med* 2001; **344**: 495-500 [PMID: 11172192 DOI: 10.1056/nejm200102153440706]
- 44 **Xu XY**, Kong H, Song RX, Zhai YH, Wu XF, Ai WS, Liu HB. The effectiveness of noninvasive biomarkers to predict hepatitis B-related significant fibrosis and cirrhosis: a systematic review and meta-analysis of diagnostic test accuracy. *PLoS One* 2014; **9**: e100182 [PMID: 24964038 DOI: 10.1371/journal.pone.0100182]
- 45 **Fraquelli M**, Rigamonti C, Casazza G, Conte D, Donato MF, Ronchi G, Colombo M. Reproducibility of transient elastography in the evaluation of liver fibrosis in patients with chronic liver disease. *Gut* 2007; **56**: 968-973 [PMID: 17255218 DOI: 10.1136/gut.2006.111302]
- 46 **Afdhal NH**, Bacon BR, Patel K, Lawitz EJ, Gordon SC, Nelson DR, Challies TL, Nasser I, Garg J, Wei LJ, McHutchison JG. Accuracy of fibroscan, compared with histology, in analysis of liver fibrosis in patients with hepatitis B or C: a United States multicenter study. *Clin Gastroenterol Hepatol* 2015; **13**: 772-9. e1-772-9.e3 [PMID: 25528010 DOI: 10.1016/j.cgh.2014.12.014]
- 47 **Cardoso AC**, Carvalho-Filho RJ, Stern C, Dipumpo A, Giuly N, Ripault MP, Asselah T, Boyer N, Lada O, Castelnau C, Martinot-Peignoux M, Valla DC, Bedossa P, Marcellin P. Direct comparison of diagnostic performance of transient elastography in patients with chronic hepatitis B and chronic hepatitis C. *Liver Int* 2012; **32**: 612-621 [PMID: 22103765 DOI: 10.1111/j.1478-3231.2011.02660.x]
- 48 **Tsochatzis EA**, Gurusamy KS, Ntaoula S, Cholongitas E, Davidson BR, Burroughs AK. Elastography for the diagnosis of severity of fibrosis in chronic liver disease: a meta-analysis of diagnostic accuracy. *J Hepatol* 2011; **54**: 650-659 [PMID: 21146892 DOI: 10.1016/j.jhep.2010.07.033]
- 49 **Branchi F**, Conti CB, Baccarin A, Lampertico P, Conte D, Fraquelli M. Non-invasive assessment of liver fibrosis in chronic hepatitis B. *World J Gastroenterol* 2014; **20**: 14568-14580 [PMID: 25356021 DOI: 10.3748/wjg.v20.i40.14568]
- 50 **Castéra L**, Bernard PH, Le Bail B, Foucher J, Trimoulet P, Merrouche W, Couzigou P, de Ledinghen V. Transient elastography and biomarkers for liver fibrosis assessment and follow-up of inactive hepatitis B carriers. *Aliment Pharmacol Ther* 2011; **33**: 455-465 [PMID: 21235598 DOI: 10.1111/j.1365-2036.2010.04547.x]
- 51 **Zucman-Rossi J**, Villanueva A, Nault JC, Llovet JM. Genetic Landscape and Biomarkers of Hepatocellular Carcinoma. *Gastroenterology* 2015; **149**: 1226-1239.e4 [PMID: 26099527 DOI: 10.1053/j.gastro.2015.05.061]
- 52 **Schulze K**, Imbeaud S, Letouzé E, Alexandrov LB, Calderaro J, Rebouissou S, Couchy G, Meiller C, Shinde J, Soysouvanh F, Calatayud AL, Pinyol R, Pelletier L, Balabaud C, Laurent A, Blanc JF, Mazzaferro V, Calvo F, Villanueva A, Nault JC, Bioulac-Sage P, Stratton MR, Llovet JM, Zucman-Rossi J. Exome sequencing of hepatocellular carcinomas identifies new mutational signatures and potential therapeutic targets. *Nat Genet* 2015; **47**: 505-511 [PMID: 25822088 DOI: 10.1038/ng.3252]
- 53 **Buendia MA**, Neuvet C. Hepatocellular carcinoma. *Cold Spring Harb Perspect Med* 2015; **5**: a021444 [PMID: 25646384 DOI: 10.1101/cshperspect.a021444]
- 54 **Sung WK**, Zheng H, Li S, Chen R, Liu X, Li Y, Lee NP, Lee WH, Ariyaratne PN, Tennakoon C, Mulawadi FH, Wong KF, Liu AM, Poon RT, Fan ST, Chan KL, Gong Z, Hu Y, Lin Z, Wang G, Zhang Q, Barber TD, Chou WC, Aggarwal A, Hao K, Zhou W, Zhang C, Hardwick J, Buser C, Xu J, Kan Z, Dai H, Mao M, Reinhard C, Wang J, Luk JM. Genome-wide survey of recurrent HBV integration in hepatocellular carcinoma. *Nat Genet* 2012; **44**: 765-769 [PMID: 22634754 DOI: 10.1038/ng.2295]
- 55 **Kojima H**, Yokosuka O, Imazeki F, Saisho H, Omata M. Telomerase activity and telomere length in hepatocellular carcinoma and chronic liver disease. *Gastroenterology* 1997; **112**: 493-500 [PMID: 9024303]
- 56 **Suda T**, Isokawa O, Aoyagi Y, Nomoto M, Tsukada K, Shimizu T, Suzuki Y, Naito A, Igarashi H, Yanagi M, Takahashi T, Asakura H. Quantitation of telomerase activity in hepatocellular carcinoma: a possible aid for a prediction of recurrent diseases in the remnant liver. *Hepatology* 1998; **27**: 402-406 [PMID: 9462637 DOI: 10.1002/hep.510270213]
- 57 **Wiemann SU**, Satyanarayana A, Tshauridu M, Tillmann HL, Zender L, Klempnauer J, Flemming P, Franco S, Blasco MA, Manns MP, Rudolph KL. Hepatocyte telomere shortening and senescence are general markers of human liver cirrhosis. *FASEB J* 2002; **16**: 935-942 [PMID: 12087054 DOI: 10.1096/fj.01-0977com]
- 58 **Nault JC**, Calderaro J, Di Tommaso L, Balabaud C, Zafrani ES, Bioulac-Sage P, Roncalli M, Zucman-Rossi J. Telomerase

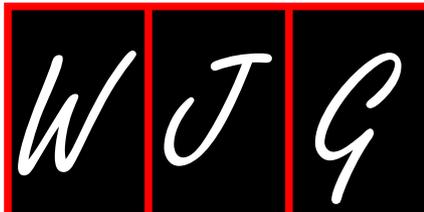
reverse transcriptase promoter mutation is an early somatic genetic alteration in the transformation of premalignant nodules in hepatocellular carcinoma on cirrhosis. *Hepatology* 2014; **60**: 1983-1992 [PMID: 25123086 DOI: 10.1002/hep.27372]

59 **Hartmann D**, Srivastava U, Thaler M, Kleinhans KN, N'kontchou G, Scheffold A, Bauer K, Kratzer RF, Kloos N, Katz SF, Song Z,

Begus-Nahrman Y, Kleger A, von Figura G, Strnad P, Lechel A, Günes C, Potthoff A, Deterding K, Wedemeyer H, Ju Z, Song G, Xiao F, Gillen S, Schrezenmeier H, Mertens T, Ziol M, Friess H, Jarek M, Manns MP, Beaugrand M, Rudolph KL. Telomerase gene mutations are associated with cirrhosis formation. *Hepatology* 2011; **53**: 1608-1617 [PMID: 21520174 DOI: 10.1002/hep.24217]

P- Reviewer: Sunbul M **S- Editor:** Yu J **L- Editor:** A
E- Editor: Zhang DN





2016 Hepatitis B virus: Global view

X region mutations of hepatitis B virus related to clinical severity

Hong Kim, Seoung-Ae Lee, Bum-Joon Kim

Hong Kim, Seoung-Ae Lee, Bum-Joon Kim, Department of Biomedical Sciences, Microbiology and Immunology, Liver Research Institute and Cancer Research Institute, College of Medicine, Seoul National University, Chongno-gu, Seoul 110-799, South Korea

Author contributions: Kim BJ conceived this research and participated in its design and coordination; Kim H and Lee SA analyzed and interpreted the data; Kim BJ and Kim H wrote this manuscript; Kim H and Lee SA reviewed the manuscript; All authors approved the final manuscript.

Supported by National Research Foundation grant of Ministry of Science, ICT and Future Planning, South Korea, No. NRF-2015R1C1A1A02037267; and Korea Health Technology R&D Project through the Korea Health Industry Development Institute, funded by the Ministry of Health and Welfare, South Korea, No. HI14C0955.

Conflict-of-interest statement: No conflict of interest.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Bum-Joon Kim, PhD, Department of Biomedical Sciences, Microbiology and Immunology, Liver Research Institute and Cancer Research Institute, College of Medicine, Seoul National University, 28 Yongon-dong, Chongno-gu, Seoul 110-799, South Korea. kbumjoon@snu.ac.kr
Telephone: + 82-2-7408316
Fax: + 82-2-7430881

Received: March 24, 2016
Peer-review started: March 25, 2016
First decision: May 12, 2016

Revised: May 17, 2016
Accepted: June 2, 2016
Article in press: June 2, 2016
Published online: June 28, 2016

Abstract

Chronic hepatitis B virus (HBV) infection remains a major health problem, with more than 240 million people chronically infected worldwide and potentially 650000 deaths per year due to advanced liver diseases including liver cirrhosis and hepatocellular carcinoma (HCC). HBV-X protein (HBx) contributes to the biology and pathogenesis of HBV *via* stimulating virus replication or altering host gene expression related to HCC. The HBV X region contains only 465 bp encoding the 16.5 kDa HBx protein, which also contains several critical cis-elements such as enhancer II, the core promoter and the microRNA-binding region. Thus, mutations in this region may affect not only the HBx open reading frame but also the overlapped cis-elements. Recently, several types of HBx mutations significantly associated with clinical severity have been described, although the functional mechanism in most of these cases remains unsolved. This review article will mainly focus on the HBx mutations proven to be significantly related to clinical severity *via* epidemiological studies.

Key words: Hepatitis B virus infection; Hepatitis B virus-X protein mutation; Hepatocellular carcinoma; Clinical severity

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Of hepatitis B virus (HBV)-X protein (HBx) mutations related to clinical severity, the A1762T/G1764A BCP mutation is one of the most frequently

encountered HBx mutations and plays a significant role in liver disease progression in chronic patients with HBV infections. It also further contributes to disease progression by inducing mutations of other HBx mutations related to clinical severity, such as G1386A/C (V5M/L), C1653T (H94Y), T1753V (I127V) and HBx C-terminal deletion or insertion. Moreover, T1753V (I127L,T,N,S) and C1653T (H94Y) mutations in the enhancer II/BCP region and A1383C, G1386A/C (V5M/L) and C1485T (P38S) in the negative regulation domain are significantly related to severe types of liver diseases, including hepatocellular carcinoma. Furthermore, deletions or insertions affecting the C-terminal region of HBx may also contribute to the severity of the clinical outcome in chronic patients. In addition, our recent study indicated that a novel mutation type (X8Del) composed of an 8-bp deletion in the C-terminal region of the HBx could contribute to occult HBV infection in vaccinated Korean individuals *via* a reduced secretion of HBsAg and virions. Therefore, several distinct types of HBx mutations may contribute to HBV pathogenesis by regulating HBV replication or host genes related to cell homeostasis.

Kim H, Lee SA, Kim BJ. X region mutations of hepatitis B virus related to clinical severity. *World J Gastroenterol* 2016; 22(24): 5467-5478 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5467.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5467>

INTRODUCTION

Chronic hepatitis B virus (HBV) infection remains a major health problem with more than 240 million people chronically infected worldwide, which potentially causes 650000 deaths per year due to advanced liver diseases including liver cirrhosis and hepatocellular carcinoma (HCC), particularly in endemic areas such as China and South Korea^[1,2]. It is generally accepted that HBV infection accounts for approximately 50% of the HCC cases worldwide and even 80%-90% in highly endemic areas^[1].

HBV is an enveloped Hepadnavirus belonging to the *Hepadnaviridae* family, with an incomplete double-stranded DNA genome of approximately 3.2 kb in length with four overlapping open reading frames (ORFs) encoding the polymerase (P), core (C), surface antigen (S), and X protein^[3]. The S gene encodes a family of surface antigen polypeptides embedded within the viral envelope, which is a major target for diagnosis and protective vaccines. The C gene encodes the core antigen, which forms the nucleocapsid, within which reverse transcription of pre-genomic RNA occurs. The P gene encodes the virus reverse transcriptase, which also has RNase H and DNA polymerase activities^[4-6]. Transcription of HBV proteins is controlled under four promoters (preS1, preS2, core and X) and

two enhancers (EnhI and EnhII) in the viral genome, which overlap with those ORFs. Because it contains a polymerase without proofreading activity and uses an RNA intermediate (pgRNA) during its replication, the HBV genome has a higher mutation ratio than other DNA viruses^[7-11]. Moreover, host immune pressures and interventions such as antiviral drugs and vaccines make the viral mutations more complicated^[12-18].

Based on an intergroup divergence of > 8% in its complete genome sequence, the HBV strains are classified into eight genotypes, which are designated A-H, with a distinct ethnic and geographical distribution^[1,19-21]. Different genotypes have distinct geographical distributions and usually induce various clinical outcomes. For instance, genotype C, the most prevalent genotype in Asia, is more prone to mutations and is associated with more severe liver diseases and lower antiviral responses compared with genotype B^[3,22,23]. In particular, genotype C2 is reportedly responsible for the most chronic infections in South Korea. Indeed, several types of HBV mutations that are never or rarely encountered in other areas have been found in South Korea and have been proven through molecular epidemiologic or functional studies to be related to disease progression in chronic patients^[24-44].

HBV X PROTEIN STRUCTURE AND FUNCTION

The HBV X protein (HBx) is a multifunctional non-structural protein that contributes to HBV biology and pathogenesis by stimulating virus replication or altering host gene expression related to HCC. HBx contains only 465 bp encoding the 16.5 kDa protein, which also contains several critical cis-elements such as EnhII, the core promoter and the microRNA-binding region^[45-47] (Figure 1).

HBx plays a significant role in sustained HBV replication, which is a major risk factor for HCC development *via* proteasome inhibition^[48,49], transactivation of HBV enhancer or promoters^[50], autophagy induction^[51,52], or polymerase activation by Ca²⁺-dependent signaling^[53-55]. HBx can also regulate HBV replication through epigenetic modifications, by being recruited onto the viral minichromosome in the nuclei of infected hepatocytes along with cellular histone acetyltransferases such as CREB-binding protein (CBP)/p300^[56,57] and histone deacetylases such as HDAC1 and hSirt1^[58]. HBx can help establish and maintain chronic infection by altering the patterns of host innate immunity, which causes the development and progression of chronic liver diseases in the absence of virus elimination^[59,60]. HBx blocks apoptotic signaling and activates signaling pathways (such as NF- κ B and PI3K) that override apoptotic signals from extrinsic ligands such as Fas or TNF- α ^[61,62]. HBx also plays an important role in hepatocarcinogenesis by inactivating the tumor suppressor p53^[63], promoting Rb inactivation by

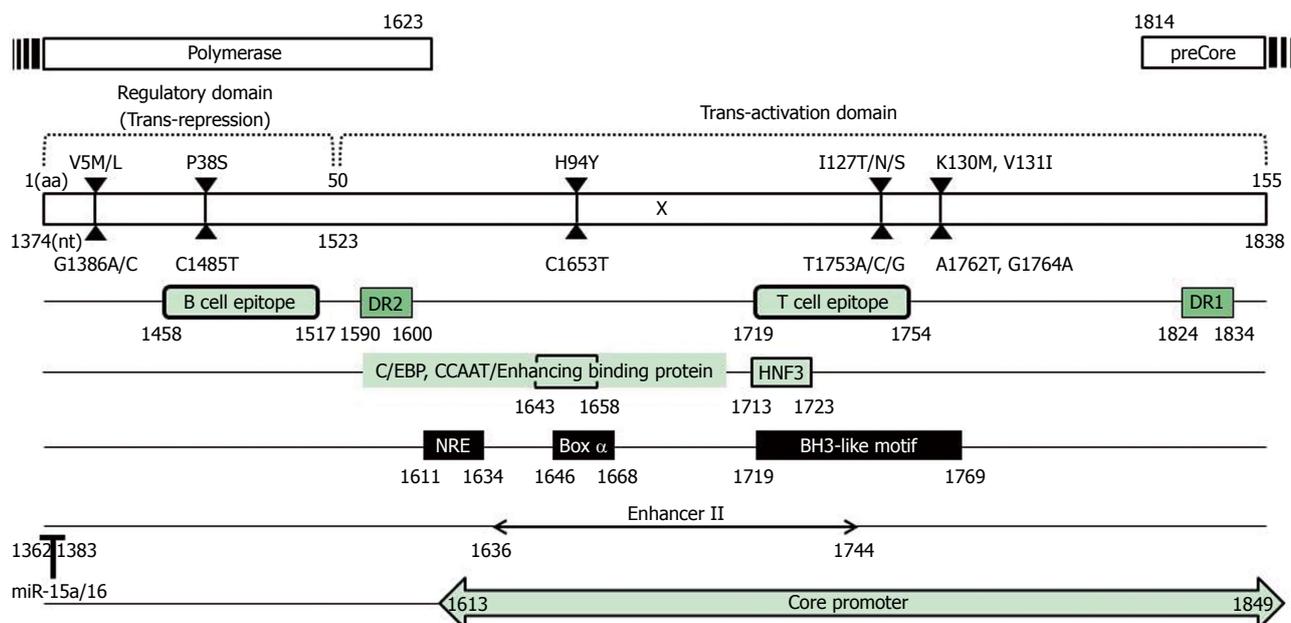


Figure 1 Hepatitis B virus-X protein genome structure. The HBV X region contains 465 bp (nt 1374 to 1838) encoding the 16.5 kDa HBx protein composed of 154 aa, which also contains several critical cis-elements such as EnhII (nt 1636 to 1744), the core promoter (nt 1613 to 1849) and the microRNA-binding region (nt 1362 to 1383). Thus, HBx mutations affected not only the HBx open reading frame but also the overlapped cis-elements. HBx: Hepatitis B virus-X protein.

phosphorylation^[64], and compromising DNA repair mechanisms^[65]. Consequently, mutations in the HBx ORF region may affect not only the HBx ORF and the overlapped cis-elements but also its binding capacity for host proteins. Recently, several types of HBx mutations significantly associated with clinical severity have been described mainly from chronic patients infected with genotype C^[28,31,39,66-83], although the functional mechanism in most of these cases remains unsolved. This review article will mainly focus on the HBx mutations that have been proven to be significantly related to clinical severity *via* epidemiological studies (Table 1).

HBx MUTATIONS RELATED TO CLINICAL SEVERITY

Mutations in EnhII and (or) the core promoter region (BCP mutation, T1753V, and C1653T)

In general, 3 types of mutations in the EnhII/BCP region [one mutation in EnhII (H94Y: C→T of nt 1653) and two mutations in BCP (I127L,T,N,S: T→V of nt 1753, K130M and V131I: A→T of nt 1762 and G→A of nt 1764)] are mutational "hot spots", namely, the most frequently encountered among naturally occurring HBx mutations related to clinical severance from chronic hepatitis B patients, irrespective of genotype or geographical distributions (Table 1). The A1762T/G1764A BCP mutation leading to two overlapped HBx amino acid changes, K130M and V131I, is the most frequent HBV DNA mutation identified in many studies as being associated with HCC risk and outcomes^[72,74,84-87]. The exact mechanism underlying

the role of this mutation in hepatocarcinogenesis is still unknown. However, some underlying mechanisms have been recently elucidated. The mutation can cause a substantial decrease in HBeAg expression and enhancement of viral genome replication, which contribute to the liver disease progression *via* increased inflammation and viral invasion^[88,89]. The mutation also leads to a truncated HBx protein, which not only promotes hepatocellular proliferation but also enhances HCC cell invasion and metastasis^[90,91]. In particular, in chronic patients infected with genotype C2, this mutation is reported to be related to HBV genome deletion^[26,31] or to be positively correlated with HBx M5V/L or H94Y mutations^[28,37]. In addition, it may also contribute to hepatocarcinogenesis *via* reduced p21 expression, leading to rapid and uncontrolled cell proliferation^[92]. A recent meta-analysis by Yang *et al.*^[82] revealed that the BCP mutation is present at significantly higher frequencies in HCC patients than in non-HCC controls, including patients with liver cirrhosis, chronic hepatitis and asymptomatic carriers. Our previous data using a Korean cohort with genotype C2-infected chronic patients also showed that the BCP mutation was the most frequently encountered mutation related to clinical severity (66.1%, 123/184 strains) and was significantly related to HCC [HCC (86.7%) vs chronic hepatitis (61%), $P = 0.017$; HCC (86.7%) vs asymptomatic carrier (24.4%), $P < 0.001$]^[28]. Our data also showed that during the natural course of HBV chronic infection, the most significant rise in the rate of the BCP mutation was found during the progression from asymptomatic carrier to chronic hepatitis (24.4%–61.0%), suggesting that the BCP mutation may play a major role in liver

Table 1 Mutations in the hepatitis B virus X region as related to clinical severity

Type of mutation	Mutations		Genotype	Clinical Significance		Region	Description	Ref.	
	Amino acid mutations			HCC (%)	Non-HCC (%)				P value
	aa mutations	nt mutations							
AS	4	NC	1383	A1383C	B/C	HCC (postoperative survival in patients with HBV-HCC) ($P = 0.028$)	Independent predictors of HCC survival	[65]	
AS	5	V5M/L	1386	G1386A/C	C	52.8 50.0 47.8 37.7	25.8 4.9 0.0 13.3	0.003 < 0.001 < 0.001 0.002	[66] [27] [38] [67]
AS	30	NC	1461	G1461A/T/C	B/C	HCC (postoperative survival in patients with HBV-HCC) ($P = 0.005$)	Independent predictors of HCC survival	[65]	
AS	36	T36P/S/A	1479	A/G1479C/T/G	A/C/D	49.3 15.3 80.0 21.7	22.7 7.8 17.1 4.9	0.034 < 0.001 < 0.010 0.023	[66] [66] [68] [27]
AS	38	P38S	1485	C1485T	B/C	HCC (postoperative survival in patients with HBV-HCC) ($P = 0.006$)	Independent predictors of HCC survival	[65]	
AS	44	A44V	1504, 1505	C1504G, C1505G	A/D	48.7 29.9 30.4 35.9	13.9 16.8 15.0 6.9	0.001 0.001 0.038 0.012	[69] [66] [67] [70]
AS	50	G50R	1521	G1521A/C	A/D	60.0	4.3	< 0.010	[68]
AS	57	NC	1544	T1544A/C	B/C	HCC (postoperative survival in patients with HBV-HCC) ($P = 0.039$)	Independent predictors of HCC survival	[65]	
AS	80	NC	1613	G1613A	B/C/C2	HCC (postoperative survival in patients with HBV-HCC) ($P = 0.006$)	Core promoter	Independent predictors of HCC survival	[65]
AS	81	NC	1613	G1613A	B/C	54.7 38.0	28.3 10.0	0.001 < 0.050	[71] [72]
AS	86	NC	1631	C1631T	C	50.0	8.6	0.001	[73]
AS	94	H94Y	1653	C1653T	B/C/C2	8.3 40.0	1.8 4.9	0.010 < 0.001	[66] [27]
						HCC (postoperative survival in patients with HBV-HCC) ($P = 0.015$)	Box α , C/EBP, CCAAT/enhancing binding protein, Core promoter, EnhII	Independent predictors of HCC survival	[65]
						61.3 56.0 45.0 31.6 35.4 41.2 55.5	25.3 30.0 19.0 19.1 18.6 13.3 2.9	< 0.001 0.0013 < 0.050 0.016 < 0.001 < 0.001 < 0.001	[71] [74] [72] [75] [66] [67] [73]
AS	101	S101Stop	1675	C1675A	B/C	8.9	2.2	0.017	[76]
AS	106	S106T	1689	T1689A	C2	35.3 19.3	5.3 4.4	0.001 < 0.001	[77] [76]

AS	116	L/V116V/ L	1719	T/G1719G/T	B/C	HCC (postoperative survival in patients with HBV-HCC) (<i>P</i> = 0.020)	BH3-like motif, Core promoter, EnhII, NRE	Independent predictors of HCC survival	[65]
						82.6		BH3-like motif, CP, EnhII, HNF3, T cell epitope	[66]
AS	117	NC	1724	T1724C	B/C	41.1	BH3-like motif, Core promoter, EnhII, NRE	EnhII	[77]
AS	118	NC	1727	A1727G	D1	35.0	BH3-like motif, Core promoter, EnhII, NRE		[78]
AS	123	I123S	1741	T1741C	D1	30.0	BH3-like motif, Core promoter, EnhII, NRE		[78]
AS, DM	127	I127L/T/ N/S	1753	T1753C/A	C	36.7	BH3-like motif, Core promoter, NRE		[78]
			1753	T1753C	B/C	12.2		Independent predictors of HCC survival	[65]
			1752, 1753	A1752C + T1753A/C/G	D	52.2			[79]
			1753	T1753A/C/G	C2	50.7		Significance of association with HCC	[71]
			1753	T1753A/C/G	C2	50.0			[74]
			1753	T1753C/G	A/D	43.6			[70]
			1753	T1753A/C/G	B/C	30.9		EnhII/BCP	[75]
AS	130	K130M	1762	T1753A/C/G A1762T	C	29.0			[67]
			1762	A1762T	C2	94.7	BH3-like motif, Core promoter	Significance of association with HCC	[71]
AS	131	V131I	1764	G1764A	B/C	80.0			[73]
			1773	C1773T	C2	98.7	BH3-like motif, Core promoter	Significance of association with HCC	[71]
AS, DM	134	NC	1773, 1775	C1773T + A1775G	B/C	95.0		Significance of association with HCC	[73]
			1800	T1800C	D1	95.0	Core promoter		[78]
AS	143	C143R	1800	T1800C	D1	17.5			[78]
AS, DM	100, 102	NC	1673, 1679	C1673T + A1679G	C	3.5	Core promoter	CP	[66]
AS, TM	128, 131	NC +	1757, 1764,	G1757A,	D1	17.5	Core promoter		[78]
			1766	G1764C + C1766G		37.5	Core promoter		[78]
AS, DM	130, 131	K130M + V131I	1762, 1764	A1762T + G1764A	C	86.7	Core promoter		[27]
					D	HCC (HBV-DNA ≥ 5 log copies/mL) vs CLD (HBV-DNA < 5 log copies/mL) (<i>P</i> < 0.05)			[79]
					C2	91.0			[74]
					A/D	73.0			[80]
					A/D	62.5			[70]
					B/C	64.1			[75]
					B/C	71.1		EnhII/BCP	[81]
					B/C	64.0			[82]
					A/D	44.9			[67]
					C	91.5			[76]
					C2	60.7			[6]
Del, Ins	129-154, 120-148, 115-149, 135-154, 137-151	Deletion	93-94 (4aa), 79-80 (2aa), 93-94 (4aa), 151-152 (3aa)	Insertion	C2	HCC + LC (7.6%) vs CH + C (1.5%) (<i>P</i> = 0.017)			[30]

AA: Amino acid; AS: Amino acid substitution; BCP: Basal core promoter; C: Carrier; CH: Chronic hepatitis; CLD: Chronic liver disease; CP: Core protein; Del: Deletion; DM: Double mutation; EnhII: Enhancer II; HCC: Hepatocellular carcinoma; HNF3: Hepatocyte nuclear factor 3; Ins: Insertion; LC: Liver cirrhosis; miRNA: MicroRNA; NC: No change; NRE: Negative regulatory element; TM: Triple mutation.

disease progression, especially in the progression from asymptomatic carrier to chronic hepatitis in chronic patients infected with genotype C2^[28]. This finding has also been confirmed by a recent meta-analysis^[82]. Yang *et al.*^[82] also demonstrated that HBV-infected patients with genotype C, including HCC patients, have a significantly higher risk of BCP mutation compared with those with genotype B, suggesting that the BCP mutation can increase the risk of HBV-related hepatocellular carcinoma, particularly in an HBV genotype C population.

An HBV genome transfection-based experiment indicated that the BCP mutation can reduce the synthesis of HBeAg and enhance viral replication. However, a meta-analysis and our previous report also showed that there is no significant difference in BCP mutation prevalence between HBeAg-positive and HBeAg-negative chronic HBV-infected patients^[28,82], suggesting that BCP mutation may occur in the HBeAg-positive immune tolerance phase earlier than in the HBeAg-negative immune clearance phase, at least in chronic patients infected with genotype C2.

The other HCC-associated T1753V mutation (I127L,T,N,S: T→V of nt 1753) was also shown to affect HCC survival^[93,94]. The mutations in the HBx protein, which include an I127L,T,N,S amino acid substitution, can change the HBx binding affinity to BCL2, thereby affecting HBx-induced apoptosis^[95]. Our previous data using Korean HBV-infected patients with genotype C2 showed that the prevalence of this mutation was also significantly higher in chronic patients with severe liver disease, HCC or liver cirrhosis than in patients who had milder types of diseases, were carriers or had chronic hepatitis [HCC and LC (34.3%) vs chronic hepatitis and carrier (13.4%), $P < 0.001$]^[28]. The other study using chronic patients from India who had genotype A or D revealed that this mutation is also usually associated with advanced forms of liver disease and had an increased risk of HCC^[69], suggesting that the T1753V mutation may play a significant role in liver disease progression. Our previous report showed that the T1753V mutation is significantly related to the BCP double mutation [patients with the BCP mutation (31.7%) vs patients without the BCP mutation (11.5%), $P = 0.003$], but not to HBeAg serostatus^[28]. A recent multivariate survival analysis by Xie *et al.*^[66] showed that the T1753V mutation is an independent predictor of HCC survival.

The C1653T mutation, leading to a simultaneous H94Y amino acid change in HBx, is located in box α , which is a strong activation element of the EnhII/core promoter, can enhance the box α binding affinity and EnhII/core promoter activity^[96,97]. Because many trans-regulated nuclear factors bind HBV at the 1653 site, this mutation can alter the binding affinity of these nuclear factors. The C1653T mutation has been recently reported to be a predictive factor for HCC in Japan^[75,98] and is associated with fulminant

hepatitis and the acute exacerbation of HCC^[99,100]. A recent multivariate survival analysis by Xie *et al.*^[66] showed that the C1653T mutation together with the T1753V mutation is also an independent predictor of HCC survival. Furthermore, our previous report showed that the C1653T mutation is significantly related to advanced liver diseases in Korean patients with genotype C2 infections [patients with HCC or LC (36.3%) vs patients who have chronic hepatitis or are carriers (12.2%), $P < 0.001$]. It has been reported that the C1653T mutation, together with 1762T/1764A mutations, can reduce the pre-C mRNA level (to approximately 55%) and HBeAg secretion in a transient transfection system using Huh7 cells^[101]. Our previous study also demonstrated that this mutation tended to be related to an HBeAg-negative serostatus ($P = 0.087$) and was significantly related to the BCP mutation [patients with the BCP mutation (35.0%) vs patients without the BCP mutation (6.6%), $P < 0.001$].

Mutation in the negative regulation domain of HBx (aa 1-50) (A1383C, G1386A/C-V5M/L, C1485T-P38S)

The A1383T synonymous mutation, which does not cause an amino acid change in the HBx protein, is located in the negative regulation domain of HBx (aa 1-50), and this mutation was first found to be associated with HCC in a Korean cohort^[28]. In one clinical study using Chinese cohort mostly infected with genotype B and C, this mutation was also associated with a worse prognosis in patients after liver transplantation^[66]. Recently, a comprehensive analysis study based on global data by Li *et al.*^[67] showed that A1383T is one of the HBx mutations identified as independent risk factors for genotype C HBV-related HCC. It has also been reported that tumor suppressor microRNA 15a/16 (miR-15a/16) can directly target the wildtype HBx RNA sequence (nt1362-1383), inducing Bcl-2 expression by acting as a sponge to bind and sequester endogenous miR-15a/16. Consequently, this mutation can lead to a reduced binding capacity of miR-15a/16 to the HBx protein^[47], which can prevent the infected cell from apoptosis by altering critical cell signal pathways and thereby contributing to hepatocarcinogenesis.

The G1386A/C mutation leading to a simultaneous V5M/L amino acid change at codon 5 of the HBx protein was first introduced by our previous study using a Korean cohort with genotype C2 infections^[28]. Our data showed that this mutation was significantly more frequently found in HCC patients than in patients in other disease groups. Notably, the prevalence of this mutation was abruptly increased in HCC patients rather than in liver cirrhosis patients during disease progression (HCC vs liver cirrhosis; 49.2% vs 25.6%, $P = 0.024$), strongly suggesting that this mutation is a genuine HCC-specific mutation that possibly plays a pivotal role in the progression from liver cirrhosis to HCC^[28]. Recently, the combination of both BCP double

mutations and both types of the V5M mutation, V5M and V5L, has also been reported to increase the HCC risk by 5.34 times compared with the wild type by inducing a higher NF- κ B activity in transformed cells^[86]. Our previous report showed that this mutation is significantly related to an HBeAg-negative serostatus [HBeAg-negative patients (40%) vs HBeAg-positive patients (19.1%), $P = 0.004$], suggesting that it may be generated from the immune clearance phase^[28]. This mutation was also significantly related to the BCP mutation [patients with the BCP mutation (36.6%) vs patients without the BCP mutation (9.2%), $P < 0.001$]. To date, its clinical relevance has not been introduced except for a Korean cohort with genotype C2 infections. It is tempting to speculate that this mutation may play a pivotal role in hepatocarcinogenesis during the HBeAg-negative immune clearance phase during the natural course of genotype C2 HBV infection.

The C1485T mutation, leading to simultaneous P38S in the HBx protein, were first introduced as an independent risk factor for HCC development in a study by Muroyama *et al.*^[70] using a Japanese cohort with genotype C infections. Both studies using Korean cohorts with genotype C2 infections^[28,68] and a recent investigation based on global data by Li *et al.*^[67] also revealed that this mutation is significantly related to HCC. A functional study supporting the relationship between the mutations with HCC still remains to be conducted. However, given that its mutation site is located at the B cell epitope region (Figure 1), this mutation may lead to persistent infection by providing a mechanism of evading the humoral immune response of the host.

Deletions or insertions in the C-terminal region of HBx

The C-terminal region of HBx plays a key role in controlling cell proliferation, viability, and transformation^[102-105]. Therefore, C-terminally deleted or inserted HBx has reduced transactivation activity and inhibitory effects on cell proliferation and thus may contribute to HCC generation^[106]. Moreover, its reduced transacting capacity might reduce HBV replication^[107]. The C-terminal deletion or insertion is one of the most frequently reported mutations of HBx and has been frequently detected in tissues and serum samples from HCC patients, irrespective of genotype or geographical distribution^[24,108,109]. Our previous report using a Korean cohort with genotype C2 infections showed that the prevalence of deletions or insertions was significantly higher in patients with severe liver disease, HCC, or cirrhosis of the liver (7.2%, 10/132), compared with patients who were carriers or had chronic hepatitis (1.5%, 2/135) ($P = 0.017$)^[31]. All deletions in six strains were concentrated at the C-terminal end of HBx, encompassing the 113th to 154th codons. Four types of insertions (PKLL, GM, FFN, and tt) were observed in six patients. Notably, all insertions were accompanied by a BCP double mutation^[31] (Figure

2). Furthermore, we first introduced a novel HBxAg vaccine escape mutation, X8Del with an 8-bp deletion in the C-terminal region of the HBx gene from 6 vaccinated Korean subjects^[38]. Our *in vitro* and *in vivo* studies showed that this mutation causes a reduced secretion of HBsAg and HBV virions compared with the wild type, suggesting that the X8Del mutation may contribute to occult HBV infection in vaccinated individuals *via* the reduced secretion of HBsAg and virions, possibly by compromising the transacting capacity of HBxAg^[38].

Other HBx mutations related to clinical severity

Recently, Xie *et al.*^[66] have reported 8 HBx mutational sites identified as significant independent risk predictors of HCC survival: 1383, 1461, 1485, 1544, 1613, 1653, 1719, and 1753 from a Chinese cohort mostly infected with genotype B and C. Despite the fact that the G1461V mutation is located at the B cell epitope, it (as a synonymous mutation) did not cause any simultaneous amino acid change in the HBx protein. Its regulatory modification in host cell or virion replication remains to be solved. The T1544V mutation also did not cause an amino acid change in the HBx protein. The G1613A mutation in the core promoter region is also a synonymous mutation, and its relationships with HCC have been reported in other previous studies.

Mutations in the BH-3-like motif of HBx can interfere with its interaction with two other Bcl-2 family members (Bcl-2 and Bcl-xL, which are critical for HBx to increase the intracellular calcium concentration), playing a significant role in viral replication and cell death^[110]. Previous studies have reported that several types of mutations in the BH-3-like motif, T/G1719G/T, T1724C, and T1741C, were also significantly related to HCC^[66,67,79].

The T1800C mutation leading to a simultaneous C143R amino acid change in the HBx protein is a novel genotype C HCC risk mutation identified by the Li *et al.*^[67] study, based on global data. To date, the function of this mutation in HCC remains unclear. However, of note, a recent study regarding HBV integration sites in 88 Chinese HCC patients showed that almost 40% of the integrated HBV genomes were cleaved at approximately nt1800, suggesting a potential role of this site in carcinogenesis, given that HBV genome integration has long been considered an important factor in HCC development.

CONCLUSION

In conclusion, HBx mutations may affect not only the HBx ORF but also the overlapped cis-elements. Considering all the HBx mutations related to clinical severity, the A1762T/G1764A BCP mutation is one of the most frequently encountered HBx mutations and plays a significant role in liver disease progression in

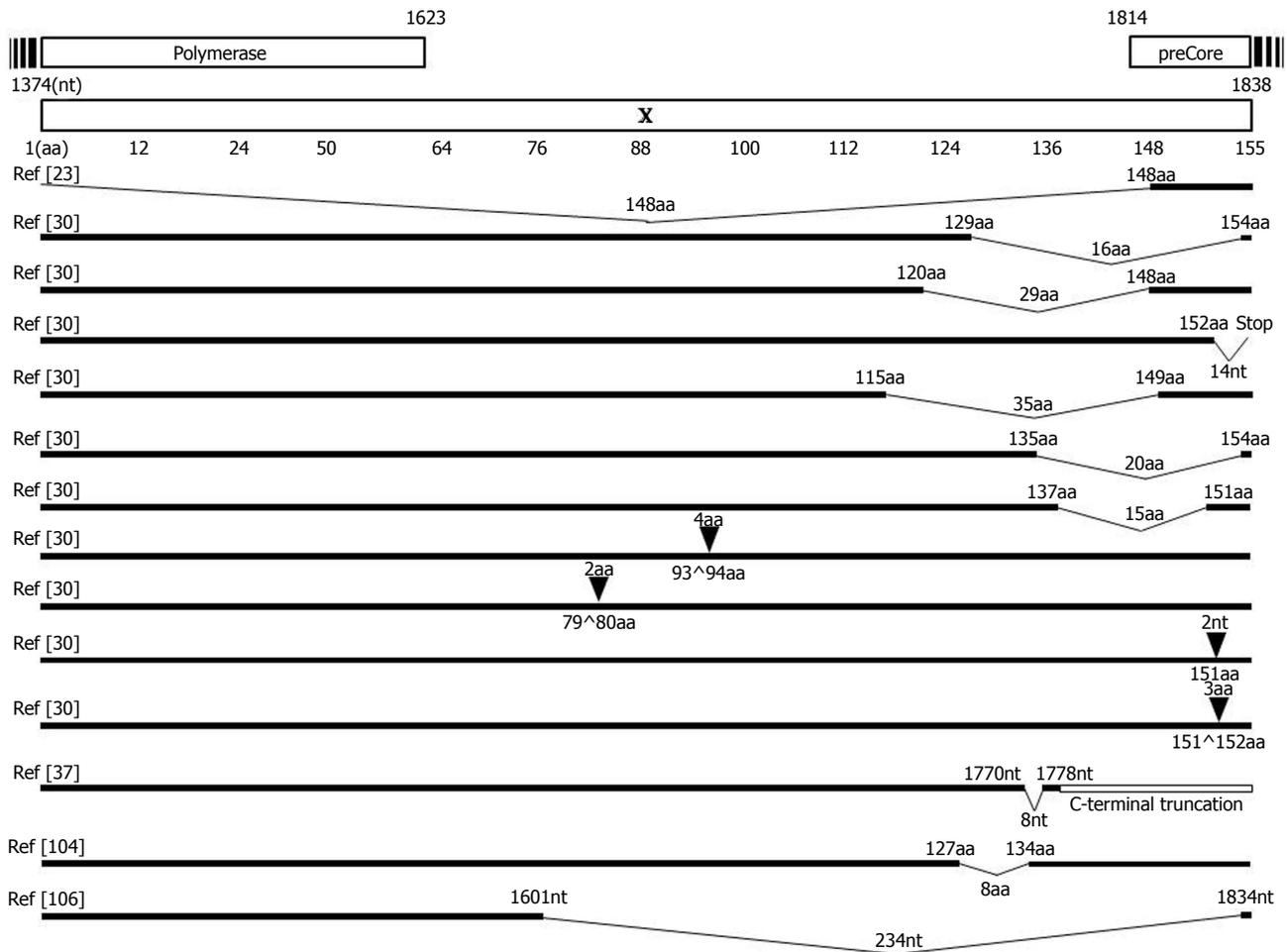


Figure 2 Mapping of deletions or insertions in the hepatitis B virus-X protein region. Deletions or insertions in the HBx region mainly occur in the C-terminal region of HBx, which may also contribute to the clinical outcome severity in chronic patients. HBx: Hepatitis B virus-X protein.

chronic patients with HBV infections. It also further contributes to disease progression by inducing mutations of other HBx mutations related to clinical severity, such as G1386A/C (V5M/L), C1653T (H94Y), T1753V (I127V) and HBx C terminal deletion or insertion. Moreover, T1753V (I127L,T,N,S) and C1653T (H94Y) mutations in the EnhII/BCP region and A1383C, G1386A/C (V5M/L) and C1485T (P38S) in the negative regulation domain were significantly related to severe types of liver diseases, including HCC. Furthermore, deletions or insertions affecting the C-terminal region of HBx can also contribute to the clinical outcome severity in chronic patients. In addition, our recent study indicated that a novel mutation type (X8Del) composed of an 8-bp deletion in the C-terminal region of the HBx contributes to occult HBV infection in vaccinated Korean individuals *via* a reduced secretion of HBsAg and virions. Thus, several distinct types of HBx mutations may contribute to HBV pathogenesis by regulating HBV replication or host genes related to cell homeostasis.

REFERENCES

1 Lee WM. Hepatitis B virus infection. *N Engl J Med* 1997; **337**:

1733-1745 [PMID: 9392700 DOI: 10.1056/NEJM199712113372406]
 2 Lozano R, Naghavi M, Foreman K, Lim S, Shibuya K, Aboyans V, Abraham J, Adair T, Aggarwal R, Ahn SY, Alvarado M, Anderson HR, Anderson LM, Andrews KG, Atkinson C, Baddour LM, Barker-Collo S, Bartels DH, Bell ML, Benjamin EJ, Bennett D, Bhalla K, Bikbov B, Bin Abdulhak A, Birbeck G, Blyth F, Bolliger I, Boufous S, Bucello C, Burch M, Burney P, Carapetis J, Chen H, Chou D, Chugh SS, Coffeng LE, Colan SD, Colquhoun S, Colson KE, Condon J, Connor MD, Cooper LT, Corriere M, Cortinovis M, de Vaccaro KC, Couser W, Cowie BC, Criqui MH, Cross M, Dabhadkar KC, Dahodwala N, De Leo D, Degenhardt L, Delossantos A, Denenberg J, Des Jarlais DC, Dharmaratne SD, Dorsey ER, Driscoll T, Duber H, Ebel B, Erwin PJ, Espindola P, Ezzati M, Feigin V, Flaxman AD, Forouzanfar MH, Fowkes FG, Franklin R, Fransen M, Freeman MK, Gabriel SE, Gakidou E, Gaspari F, Gillum RF, Gonzalez-Medina D, Halasa YA, Haring D, Harrison JE, Havmoeller R, Hay RJ, Hoen B, Hotez PJ, Hoy D, Jacobsen KH, James SL, Jasrasaria R, Jayaraman S, Johns N, Karthikeyan G, Kassebaum N, Keren A, Khoo JP, Knowlton LM, Kobusingye O, Koranteng A, Krishnamurthi R, Lipnick M, Lipshultz SE, Ohno SL, Mabweijano J, MacIntyre MF, Mallinger L, March L, Marks GB, Marks R, Matsumori A, Matzopoulos R, Mayosi BM, McAnulty JH, McDermott MM, McGrath J, Mensah GA, Merriman TR, Michaud C, Miller M, Miller TR, Mock C, Mocumbi AO, Mokdad AA, Moran A, Mulholland K, Nair MN, Naldi L, Narayan KM, Nasseri K, Norman P, O'Donnell M, Omer SB, Ortblad K, Osborne R, Ozgediz D, Pahari B, Pandian JD, Rivero AP, Padilla RP, Perez-Ruiz F, Perico N, Phillips D, Pierce K, Pope CA, Porrini E, Pourmalek F, Raju M, Ranganathan D, Rehm JT, Rein DB, Remuzzi G, Rivara FP, Roberts T, De León

- FR, Rosenfeld LC, Rushton L, Sacco RL, Salomon JA, Sampson U, Sanman E, Schwebel DC, Segui-Gomez M, Shepard DS, Singh D, Singleton J, Sliwa K, Smith E, Steer A, Taylor JA, Thomas B, Tleyjeh IM, Towbin JA, Truelsen T, Undurraga EA, Venketasubramanian N, Vijayakumar L, Vos T, Wagner GR, Wang M, Wang W, Watt K, Weinstock MA, Weintraub R, Wilkinson JD, Woolf AD, Wulf S, Yeh PH, Yip P, Zabetian A, Zheng ZJ, Lopez AD, Murray CJ, AlMazroa MA, Memish ZA. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* 2012; **380**: 2095-2128 [PMID: 23245604 DOI: 10.1016/S0140-6736]
- 3 **Guirgis BS**, Abbas RO, Azzazy HM. Hepatitis B virus genotyping: current methods and clinical implications. *Int J Infect Dis* 2010; **14**: e941-e953 [PMID: 20674432 DOI: 10.1016/j.ijid.2010.03.020]
 - 4 **Gao S**, Duan ZP, Coffin CS. Clinical relevance of hepatitis B virus variants. *World J Hepatol* 2015; **7**: 1086-1096 [PMID: 26052397 DOI: 10.4254/wjh.v7.i8.1086]
 - 5 **Jones SA**, Hu J. Hepatitis B virus reverse transcriptase: diverse functions as classical and emerging targets for antiviral intervention. *Emerg Microbes Infect* 2013; **2**: e56 [PMID: 26038488 DOI: 10.1038/emi.2013.56]
 - 6 **Nassal M**. Hepatitis B viruses: reverse transcription a different way. *Virus Res* 2008; **134**: 235-249 [PMID: 18339439 DOI: 10.1016/j.virusres.2007.12.024]
 - 7 **Datta S**, Chatterjee S, Veer V, Chakravarty R. Molecular biology of the hepatitis B virus for clinicians. *J Clin Exp Hepatol* 2012; **2**: 353-365 [PMID: 25755457 DOI: 10.1016/j.jceh.2012.10.003]
 - 8 **Moolla N**, Kew M, Arbutnot P. Regulatory elements of hepatitis B virus transcription. *J Viral Hepat* 2002; **9**: 323-331 [PMID: 12225325]
 - 9 **Park YM**. Clinical utility of complex mutations in the core promoter and proximal precore regions of the hepatitis B virus genome. *World J Hepatol* 2015; **7**: 113-120 [PMID: 25625002 DOI: 10.4254/wjh.v7.i1.113]
 - 10 **Quarleri J**. Core promoter: a critical region where the hepatitis B virus makes decisions. *World J Gastroenterol* 2014; **20**: 425-435 [PMID: 24574711 DOI: 10.3748/wjg.v20.i2.425]
 - 11 **Huan B**, Siddiqui A. Regulation of hepatitis B virus gene expression. *J Hepatol* 1993; **17** Suppl 3: S20-S23 [PMID: 8509635]
 - 12 **Gupta N**, Goyal M, Wu CH, Wu GY. The Molecular and Structural Basis of HBV-resistance to Nucleos(t)ide Analogs. *J Clin Transl Hepatol* 2014; **2**: 202-211 [PMID: 26357626 DOI: 10.14218/jcth.2014.00021]
 - 13 **Koumbi L**. Current and future antiviral drug therapies of hepatitis B chronic infection. *World J Hepatol* 2015; **7**: 1030-1040 [PMID: 26052392 DOI: 10.4254/wjh.v7.i8.1030]
 - 14 **Yano Y**, Azuma T, Hayashi Y. Variations and mutations in the hepatitis B virus genome and their associations with clinical characteristics. *World J Hepatol* 2015; **7**: 583-592 [PMID: 25848482 DOI: 10.4254/wjh.v7.i3.583]
 - 15 **Papatheodoridis GV**, Chan HL, Hansen BE, Janssen HL, Lampertico P. Risk of hepatocellular carcinoma in chronic hepatitis B: assessment and modification with current antiviral therapy. *J Hepatol* 2015; **62**: 956-967 [PMID: 25595883 DOI: 10.1016/j.jhep.2015.01.002]
 - 16 **Glebe D**, Geipel A. Selected phenotypic assays used to evaluate antiviral resistance and viral fitness of hepatitis B virus and its variants. *Intervirology* 2014; **57**: 225-231 [PMID: 25034492 DOI: 10.1159/000360950]
 - 17 **Luongo M**, Critelli R, Grottole A, Gitto S, Bernabucci V, Bevini M, Vecchi C, Montagnani G, Villa E. Acute hepatitis B caused by a vaccine-escape HBV strain in vaccinated subject: sequence analysis and therapeutic strategy. *J Clin Virol* 2015; **62**: 89-91 [PMID: 25542480 DOI: 10.1016/j.jcv.2014.11.029]
 - 18 **Locarnini SA**, Yuen L. Molecular genesis of drug-resistant and vaccine-escape HBV mutants. *Antivir Ther* 2010; **15**: 451-461 [PMID: 20516565 DOI: 10.3851/imp1499]
 - 19 **Delius H**, Gough NM, Cameron CH, Murray K. Structure of the hepatitis B virus genome. *J Virol* 1983; **47**: 337-343 [PMID: 6620456]
 - 20 **Liang TJ**. Hepatitis B: the virus and disease. *Hepatology* 2009; **49**: S13-S21 [PMID: 19399811 DOI: 10.1002/hep.22881]
 - 21 **Kidd-Ljunggren K**, Miyakawa Y, Kidd AH. Genetic variability in hepatitis B viruses. *J Gen Virol* 2002; **83**: 1267-1280 [PMID: 12029141 DOI: 10.1099/0022-1317-83-6-1267]
 - 22 **Miyakawa Y**, Mizokami M. Classifying hepatitis B virus genotypes. *Intervirology* 2003; **46**: 329-338 [PMID: 14688448]
 - 23 **Cho JH**, Yoon KH, Lee KE, Park DS, Lee YJ, Moon HB, Lee KR, Choi CS, Cho EY, Kim HC. [Distribution of hepatitis B virus genotypes in Korea]. *Korean J Hepatol* 2009; **15**: 140-147 [PMID: 19581766 DOI: 10.3350/kjhep.2009.15.2.140]
 - 24 **Kim H**, Jee Y, Mun HS, Park JH, Yoon JH, Kim YJ, Lee HS, Hyun JW, Hwang ES, Cha CY, Kook YH, Kim BJ. Characterization of two hepatitis B virus populations in a single Korean hepatocellular carcinoma patient with an HBeAg-negative serostatus: a novel X-Gene-deleted strain with inverted duplication sequences of upstream enhancer site II. *Intervirology* 2007; **50**: 273-280 [PMID: 17570929 DOI: 10.1159/000103915]
 - 25 **Kim H**, Jee Y, Mun HS, Song BC, Park JH, Hyun JW, Hwang ES, Cha CY, Kook YH, Kim BJ. Comparison of full genome sequences between two hepatitis B virus strains with or without preC mutation (A1896) from a single Korean hepatocellular carcinoma patient. *J Microbiol Biotechnol* 2007; **17**: 701-704 [PMID: 18051288]
 - 26 **Kim H**, Jee YM, Song BC, Hyun JW, Mun HS, Kim HJ, Oh EJ, Yoon JH, Kim YJ, Lee HS, Hwang ES, Cha CY, Kook YH, Kim BJ. Analysis of hepatitis B virus quasispecies distribution in a Korean chronic patient based on the full genome sequences. *J Med Virol* 2007; **79**: 212-219 [PMID: 17245716 DOI: 10.1002/jmv.20789]
 - 27 **Kim H**, Jee YM, Song BC, Shin JW, Yang SH, Mun HS, Kim HJ, Oh EJ, Yoon JH, Kim YJ, Lee HS, Hwang ES, Cha CY, Kook YH, Kim BJ. Molecular epidemiology of hepatitis B virus (HBV) genotypes and serotypes in patients with chronic HBV infection in Korea. *Intervirology* 2007; **50**: 52-57 [PMID: 17164558 DOI: 10.1159/000096313]
 - 28 **Kim HJ**, Park JH, Jee Y, Lee SA, Kim H, Song BC, Yang S, Lee M, Yoon JH, Kim YJ, Lee HS, Hwang ES, Kook YH, Kim BJ. Hepatitis B virus X mutations occurring naturally associated with clinical severity of liver disease among Korean patients with chronic genotype C infection. *J Med Virol* 2008; **80**: 1337-1343 [PMID: 18551606 DOI: 10.1002/jmv.21219]
 - 29 **Mun HS**, Lee SA, Jee Y, Kim H, Park JH, Song BC, Yoon JH, Kim YJ, Lee HS, Hyun JW, Hwang ES, Kook YH, Kim BJ. The prevalence of hepatitis B virus preS deletions occurring naturally in Korean patients infected chronically with genotype C. *J Med Virol* 2008; **80**: 1189-1194 [PMID: 18461612 DOI: 10.1002/jmv.21208]
 - 30 **Lee SA**, Cho YK, Lee KH, Hwang ES, Kook YH, Kim BJ. Gender disparity in distribution of the major hydrophilic region variants of hepatitis B virus genotype C according to hepatitis B e antigen serostatus. *J Med Virol* 2011; **83**: 405-411 [PMID: 21264860 DOI: 10.1002/jmv.21988]
 - 31 **Lee SA**, Mun HS, Kim H, Lee HK, Kim BJ, Hwang ES, Kook YH, Kim BJ. Naturally occurring hepatitis B virus X deletions and insertions among Korean chronic patients. *J Med Virol* 2011; **83**: 65-70 [PMID: 21108340 DOI: 10.1002/jmv.21938]
 - 32 **Mun HS**, Lee SA, Kim H, Hwang ES, Kook YH, Kim BJ. Novel F141L pre-S2 mutation in hepatitis B virus increases the risk of hepatocellular carcinoma in patients with chronic genotype C infections. *J Virol* 2011; **85**: 123-132 [PMID: 20962085 DOI: 10.1128/jvi.01524-10]
 - 33 **Kim DW**, Lee SA, Hwang ES, Kook YH, Kim BJ. Naturally occurring precore/core region mutations of hepatitis B virus genotype C related to hepatocellular carcinoma. *PLoS One* 2012; **7**: e47372 [PMID: 23071796 DOI: 10.1371/journal.pone.0047372]
 - 34 **Lee SA**, Kim K, Kim H, Kim BJ. Nucleotide change of codon 182 in the surface gene of hepatitis B virus genotype C leading to truncated surface protein is associated with progression of liver diseases. *J Hepatol* 2012; **56**: 63-69 [PMID: 21827734 DOI: 10.1016/j.jhep.2011.06.028]

- 35 **Kim H**, Lee SA, Kim DW, Lee SH, Kim BJ. Naturally occurring mutations in large surface genes related to occult infection of hepatitis B virus genotype C. *PLoS One* 2013; **8**: e54486 [PMID: 23349904 DOI: 10.1371/journal.pone.0054486]
- 36 **Lee SA**, Kim KJ, Kim DW, Kim BJ. Male-specific W4P/R mutation in the pre-S1 region of hepatitis B virus, increasing the risk of progression of liver diseases in chronic patients. *J Clin Microbiol* 2013; **51**: 3928-3936 [PMID: 24025913 DOI: 10.1128/jcm.01505-13]
- 37 **Kim BJ**. Hepatitis B virus mutations related to liver disease progression of Korean patients. *World J Gastroenterol* 2014; **20**: 460-467 [PMID: 24574714 DOI: 10.3748/wjg.v20.i2.460]
- 38 **Kim H**, Gong JR, Lee SA, Kim BJ. Discovery of a Novel Mutation (X8Del) Resulting in an 8-bp Deletion in the Hepatitis B Virus X Gene Associated with Occult Infection in Korean Vaccinated Individuals. *PLoS One* 2015; **10**: e0139551 [PMID: 26437447 DOI: 10.1371/journal.pone.0139551]
- 39 **Kim H**, Hong SH, Lee SA, Gong JR, Kim BJ. Development of Fok-I based nested polymerase chain reaction-restriction fragment length polymorphism analysis for detection of hepatitis B virus X region V5M mutation. *World J Gastroenterol* 2015; **21**: 13360-13367 [PMID: 26715821 DOI: 10.3748/wjg.v21.i47.13360]
- 40 **Kim H**, Kim BJ. Association of preS/S Mutations with Occult Hepatitis B Virus (HBV) Infection in South Korea: Transmission Potential of Distinct Occult HBV Variants. *Int J Mol Sci* 2015; **16**: 13595-13609 [PMID: 26084041 DOI: 10.3390/ijms160613595]
- 41 **Kim H**, Lee SA, Won YS, Lee H, Kim BJ. Occult infection related hepatitis B surface antigen variants showing lowered secretion capacity. *World J Gastroenterol* 2015; **21**: 1794-1803 [PMID: 25684944 DOI: 10.3748/wjg.v21.i6.1794]
- 42 **Lee H**, Kim H, Lee SA, Won YS, Kim HI, Inn KS, Kim BJ. Upregulation of endoplasmic reticulum stress and reactive oxygen species by naturally occurring mutations in hepatitis B virus core antigen. *J Gen Virol* 2015; **96**: 1850-1854 [PMID: 25828947 DOI: 10.1099/vir.0.000134]
- 43 **Lee SA**, Kim H, Won YS, Seok SH, Na Y, Shin HB, Inn KS, Kim BJ. Male-specific hepatitis B virus large surface protein variant W4P potentiates tumorigenicity and induces gender disparity. *Mol Cancer* 2015; **14**: 23 [PMID: 25645622 DOI: 10.1186/s12943-015-0303-7]
- 44 **Lee SA**, Kim KJ, Kim H, Choi WH, Won YS, Kim BJ. Hepatitis B virus preS1 deletion is related to viral replication increase and disease progression. *World J Gastroenterol* 2015; **21**: 5039-5048 [PMID: 25945020 DOI: 10.3748/wjg.v21.i16.5039]
- 45 **Liu WH**, Yeh SH, Chen PJ. Role of microRNAs in hepatitis B virus replication and pathogenesis. *Biochim Biophys Acta* 2011; **1809**: 678-685 [PMID: 21565290 DOI: 10.1016/j.bbagr.2011.04.008]
- 46 **Wang B**, Majumder S, Nuovo G, Kutay H, Volinia S, Patel T, Schmittgen TD, Croce C, Ghoshal K, Jacob ST. Role of microRNA-155 at early stages of hepatocarcinogenesis induced by choline-deficient and amino acid-defined diet in C57BL/6 mice. *Hepatology* 2009; **50**: 1152-1161 [PMID: 19711427 DOI: 10.1002/hep.23100]
- 47 **Liu N**, Zhang J, Jiao T, Li Z, Peng J, Cui Z, Ye X. Hepatitis B virus inhibits apoptosis of hepatoma cells by sponging the MicroRNA 15a/16 cluster. *J Virol* 2013; **87**: 13370-13378 [PMID: 24089558 DOI: 10.1128/jvi.02130-13]
- 48 **Hu Z**, Zhang Z, Doo E, Coux O, Goldberg AL, Liang TJ. Hepatitis B virus X protein is both a substrate and a potential inhibitor of the proteasome complex. *J Virol* 1999; **73**: 7231-7240 [PMID: 10438810]
- 49 **Zhang Z**, Protzer U, Hu Z, Jacob J, Liang TJ. Inhibition of cellular proteasome activities enhances hepatitis B virus replication in an HBX-dependent manner. *J Virol* 2004; **78**: 4566-4572 [PMID: 15078938]
- 50 **Colgrove R**, Simon G, Ganem D. Transcriptional activation of homologous and heterologous genes by the hepatitis B virus X gene product in cells permissive for viral replication. *J Virol* 1989; **63**: 4019-4026 [PMID: 2788226]
- 51 **Sir D**, Tian Y, Chen WL, Ann DK, Yen TS, Ou JH. The early autophagic pathway is activated by hepatitis B virus and required for viral DNA replication. *Proc Natl Acad Sci USA* 2010; **107**: 4383-4388 [PMID: 20142477 DOI: 10.1073/pnas.0911373107]
- 52 **Mao Y**, Da L, Tang H, Yang J, Lei Y, Tiollais P, Li T, Zhao M. Hepatitis B virus X protein reduces starvation-induced cell death through activation of autophagy and inhibition of mitochondrial apoptotic pathway. *Biochem Biophys Res Commun* 2011; **415**: 68-74 [PMID: 22020078 DOI: 10.1016/j.bbrc.2011.10.013]
- 53 **Li B**, Gao B, Ye L, Han X, Wang W, Kong L, Fang X, Zeng Y, Zheng H, Li S, Wu Z, Ye L. Hepatitis B virus X protein (HBx) activates ATF6 and IRE1-XBP1 pathways of unfolded protein response. *Virus Res* 2007; **124**: 44-49 [PMID: 17092596 DOI: 10.1016/j.virusres.2006.09.011]
- 54 **Bouchard MJ**, Wang LH, Schneider RJ. Calcium signaling by HBx protein in hepatitis B virus DNA replication. *Science* 2001; **294**: 2376-2378 [PMID: 11743208 DOI: 10.1126/science.294.5550.2376]
- 55 **McClain SL**, Clippinger AJ, Lizzano R, Bouchard MJ. Hepatitis B virus replication is associated with an HBx-dependent mitochondrion-regulated increase in cytosolic calcium levels. *J Virol* 2007; **81**: 12061-12065 [PMID: 17699583 DOI: 10.1128/jvi.00740-07]
- 56 **Belloni L**, Pollicino T, De Nicola F, Guerrieri F, Raffa G, Fanciulli M, Raimondo G, Levrero M. Nuclear HBx binds the HBV minichromosome and modifies the epigenetic regulation of cccDNA function. *Proc Natl Acad Sci USA* 2009; **106**: 19975-19979 [PMID: 19906987 DOI: 10.1073/pnas.0908365106]
- 57 **Cougot D**, Wu Y, Cairo S, Caramel J, Renard CA, Lévy L, Buendia MA, Neuveut C. The hepatitis B virus X protein functionally interacts with CREB-binding protein/p300 in the regulation of CREB-mediated transcription. *J Biol Chem* 2007; **282**: 4277-4287 [PMID: 17158882 DOI: 10.1074/jbc.M606774200]
- 58 **Ren JH**, Tao Y, Zhang ZZ, Chen WX, Cai XF, Chen K, Ko BC, Song CL, Ran LK, Li WY, Huang AL, Chen J. Sirtuin 1 regulates hepatitis B virus transcription and replication by targeting transcription factor AP-1. *J Virol* 2014; **88**: 2442-2451 [PMID: 24335313 DOI: 10.1128/jvi.02861-13]
- 59 **Wei C**, Ni C, Song T, Liu Y, Yang X, Zheng Z, Jia Y, Yuan Y, Guan K, Xu Y, Cheng X, Zhang Y, Yang X, Wang Y, Wen C, Wu Q, Shi W, Zhong H. The hepatitis B virus X protein disrupts innate immunity by downregulating mitochondrial antiviral signaling protein. *J Immunol* 2010; **185**: 1158-1168 [PMID: 20554965 DOI: 10.4049/jimmunol.0903874]
- 60 **Jiang J**, Tang H. Mechanism of inhibiting type I interferon induction by hepatitis B virus X protein. *Protein Cell* 2010; **1**: 1106-1117 [PMID: 21213104 DOI: 10.1007/s13238-010-0141-8]
- 61 **Pan J**, Duan LX, Sun BS, Feitelson MA. Hepatitis B virus X protein protects against anti-Fas-mediated apoptosis in human liver cells by inducing NF-kappa B. *J Gen Virol* 2001; **82**: 171-182 [PMID: 11125170 DOI: 10.1099/0022-1317-82-1-171]
- 62 **Kim JY**, Song EH, Lee HJ, Oh YK, Choi KH, Yu DY, Park SI, Seong JK, Kim WH. HBx-induced hepatic steatosis and apoptosis are regulated by TNFR1- and NF-kappaB-dependent pathways. *J Mol Biol* 2010; **397**: 917-931 [PMID: 20156456 DOI: 10.1016/j.jmb.2010.02.016]
- 63 **Chirillo P**, Pagano S, Natoli G, Puri PL, Burgio VL, Balsano C, Levrero M. The hepatitis B virus X gene induces p53-mediated programmed cell death. *Proc Natl Acad Sci USA* 1997; **94**: 8162-8167 [PMID: 9223332]
- 64 **Jung JK**, Park SH, Jang KL. Hepatitis B virus X protein overcomes the growth-inhibitory potential of retinoic acid by downregulating retinoic acid receptor-beta2 expression via DNA methylation. *J Gen Virol* 2010; **91**: 493-500 [PMID: 19828754 DOI: 10.1099/vir.0.015149-0]
- 65 **Arbuthnot P**, Capovilla A, Kew M. Putative role of hepatitis B virus X protein in hepatocarcinogenesis: effects on apoptosis, DNA repair, mitogen-activated protein kinase and JAK/STAT pathways. *J Gastroenterol Hepatol* 2000; **15**: 357-368 [PMID: 10824878]
- 66 **Xie Y**, Liu S, Zhao Y, Guo Z, Xu J. X protein mutations in hepatitis B virus DNA predict postoperative survival in hepatocellular carcinoma. *Tumour Biol* 2014; **35**: 10325-10331 [PMID: 25034530 DOI: 10.1007/s13277-014-2331-0]
- 67 **Li W**, Goto K, Matsubara Y, Ito S, Muroyama R, Li Q, Kato

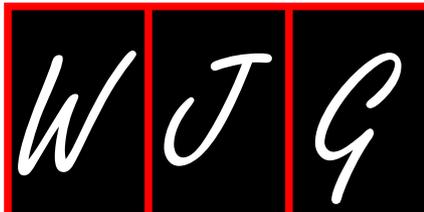
- N. The characteristic changes in hepatitis B virus x region for hepatocellular carcinoma: a comprehensive analysis based on global data. *PLoS One* 2015; **10**: e0125555 [PMID: 25942596 DOI: 10.1371/journal.pone.0125555]
- 68 **Cho EY**, Choi CS, Cho JH, Kim HC. Association between Hepatitis B Virus X Gene Mutations and Clinical Status in Patients with Chronic Hepatitis B Infection. *Gut Liver* 2011; **5**: 70-76 [PMID: 21461076 DOI: 10.5009/gnl.2011.5.1.70]
- 69 **Tuteja A**, Siddiqui AB, Madan K, Goyal R, Shalimar V, Kaur N, Panda SK, Narayanasamy K, Subodh S, Acharya SK. Mutation profiling of the hepatitis B virus strains circulating in North Indian population. *PLoS One* 2014; **9**: e91150 [PMID: 24637457 DOI: 10.1371/journal.pone.0091150]
- 70 **Muroyama R**, Kato N, Yoshida H, Otsuka M, Moriyama M, Wang Y, Shao RX, Dharel N, Tanaka Y, Ohta M, Tateishi R, Shiina S, Tatsukawa M, Fukai K, Imazeki F, Yokosuka O, Shiratori Y, Omata M. Nucleotide change of codon 38 in the X gene of hepatitis B virus genotype C is associated with an increased risk of hepatocellular carcinoma. *J Hepatol* 2006; **45**: 805-812 [PMID: 17050029 DOI: 10.1016/j.jhep.2006.07.025]
- 71 **Asim M**, Malik A, Sarma MP, Polipalli SK, Begum N, Ahmad I, Khan LA, Husain SA, Akhtar N, Husain S, Thayumanavan L, Singla R, Kar P. Hepatitis B virus BCP, Precore/core, X gene mutations/genotypes and the risk of hepatocellular carcinoma in India. *J Med Virol* 2010; **82**: 1115-1125 [PMID: 20513073 DOI: 10.1002/jmv.21774]
- 72 **Jang JW**, Chun JY, Park YM, Shin SK, Yoo W, Kim SO, Hong SP. Mutational complex genotype of the hepatitis B virus X/precure regions as a novel predictive marker for hepatocellular carcinoma. *Cancer Sci* 2012; **103**: 296-304 [PMID: 22136288 DOI: 10.1111/j.1349-7006.2011.02170.x]
- 73 **Tatsukawa M**, Takaki A, Shiraha H, Koike K, Iwasaki Y, Kobashi H, Fujioka S, Sakaguchi K, Yamamoto K. Hepatitis B virus core promoter mutations G1613A and C1653T are significantly associated with hepatocellular carcinoma in genotype C HBV-infected patients. *BMC Cancer* 2011; **11**: 458 [PMID: 22014121 DOI: 10.1186/1471-2407-11-458]
- 74 **Zhu Y**, Jin Y, Guo X, Bai X, Chen T, Wang J, Qian G, Groopman JD, Gu J, Li J, Tu H. Comparison study on the complete sequence of hepatitis B virus identifies new mutations in core gene associated with hepatocellular carcinoma. *Cancer Epidemiol Biomarkers Prev* 2010; **19**: 2623-2630 [PMID: 20699378 DOI: 10.1158/1055-9965.epi-10-0469]
- 75 **Shinkai N**, Tanaka Y, Ito K, Mukaide M, Hasegawa I, Asahina Y, Izumi N, Yatsuhashi H, Orito E, Joh T, Mizokami M. Influence of hepatitis B virus X and core promoter mutations on hepatocellular carcinoma among patients infected with subgenotype C2. *J Clin Microbiol* 2007; **45**: 3191-3197 [PMID: 17652471 DOI: 10.1128/jcm.00411-07]
- 76 **Qu LS**, Zhu J, Liu TT, Shen XZ, Chen TY, Ni ZP, Ni RZ, Lu CH. Effect of combined mutations in the enhancer II and basal core promoter of hepatitis B virus on development of hepatocellular carcinoma in Qidong, China. *Hepatol Res* 2014; **44**: 1186-1195 [PMID: 24341484 DOI: 10.1111/hepr.12291]
- 77 **Kim JK**, Chang HY, Lee JM, Baatarkhuu O, Yoon YJ, Park JY, Kim DY, Han KH, Chon CY, Ahn SH. Specific mutations in the enhancer II/core promoter/precure regions of hepatitis B virus subgenotype C2 in Korean patients with hepatocellular carcinoma. *J Med Virol* 2009; **81**: 1002-1008 [PMID: 19382267 DOI: 10.1002/jmv.21501]
- 78 **Fan W**, Shi B, Wei H, Du G, Song S. Comparison of hepatitis B X gene mutation between patients with hepatocellular carcinoma and patients with chronic hepatitis B. *Virus Genes* 2011; **42**: 162-170 [PMID: 21161360 DOI: 10.1007/s11262-010-0557-5]
- 79 **Khan A**, Al Balwi MA, Tanaka Y, Hajeer A, Sanai FM, Al Abdulkarim I, Al Ayyar L, Badri M, Saudi D, Tamimi W, Mizokami M, Al Knawy B. Novel point mutations and mutational complexes in the enhancer II, core promoter and precure regions of hepatitis B virus genotype D1 associated with hepatocellular carcinoma in Saudi Arabia. *Int J Cancer* 2013; **133**: 2864-2871 [PMID: 23740667 DOI: 10.1002/ijc.28307]
- 80 **Elkady A**, Tanaka Y, Kurbanov F, Oynsuren T, Mizokami M. Virological and clinical implication of core promoter C1752/V1753 and T1764/G1766 mutations in hepatitis B virus genotype D infection in Mongolia. *J Gastroenterol Hepatol* 2008; **23**: 474-481 [PMID: 18318825 DOI: 10.1111/j.1440-1746.2008.05321.x]
- 81 **Venard V**, Corsaro D, Kajzer C, Bronowicki JP, Le Faou A. Hepatitis B virus X gene variability in French-born patients with chronic hepatitis and hepatocellular carcinoma. *J Med Virol* 2000; **62**: 177-184 [PMID: 11002246]
- 82 **Yang Z**, Zhuang L, Lu Y, Xu Q, Tang B, Chen X. Naturally occurring basal core promoter A1762T/G1764A dual mutations increase the risk of HBV-related hepatocellular carcinoma: a meta-analysis. *Oncotarget* 2016; **7**: 12525-12536 [PMID: 26848866 DOI: 10.18632/oncotarget.7123]
- 83 **Malik A**, Singhal DK, Albanyan A, Husain SA, Kar P. Hepatitis B virus gene mutations in liver diseases: a report from New Delhi. *PLoS One* 2012; **7**: e39028 [PMID: 22720023 DOI: 10.1371/journal.pone.0039028]
- 84 **Chen QY**, Harrison TJ, Sabin CA, Li GJ, Huang GM, Yang JY, Wang XY, Li H, Liu MH, Fang ZL. The Effect of HBV Genotype C on the Development of HCC Differs Between Wild-Type Viruses and Those With BCP Double Mutations (T(1762)A(1764)). *Hepat Mon* 2014; **14**: e16214 [PMID: 24693312 DOI: 10.5812/hepatmon.16214]
- 85 **Muñoz A**, Chen JG, Egner PA, Marshall ML, Johnson JL, Schneider MF, Lu JH, Zhu YR, Wang JB, Chen TY, Kensler TW, Groopman JD. Predictive power of hepatitis B 1762T/1764A mutations in plasma for hepatocellular carcinoma risk in Qidong, China. *Carcinogenesis* 2011; **32**: 860-865 [PMID: 21474708 DOI: 10.1093/carcin/bgr055]
- 86 **Lee JH**, Han KH, Lee JM, Park JH, Kim HS. Impact of hepatitis B virus (HBV) x gene mutations on hepatocellular carcinoma development in chronic HBV infection. *Clin Vaccine Immunol* 2011; **18**: 914-921 [PMID: 21490166 DOI: 10.1128/cvi.00474-10]
- 87 **Datta S**, Banerjee A, Chandra PK, Biswas A, Panigrahi R, Mahapatra PK, Panda CK, Chakrabarti S, Bhattacharya SK, Chakravarty R. Analysis of hepatitis B virus X gene phylogeny, genetic variability and its impact on pathogenesis: implications in Eastern Indian HBV carriers. *Virology* 2008; **382**: 190-198 [PMID: 18952249 DOI: 10.1016/j.virol.2008.09.007]
- 88 **Buckwold VE**, Xu Z, Chen M, Yen TS, Ou JH. Effects of a naturally occurring mutation in the hepatitis B virus basal core promoter on precure gene expression and viral replication. *J Virol* 1996; **70**: 5845-5851 [PMID: 8709203]
- 89 **Hakami A**, Ali A, Hakami A. Effects of hepatitis B virus mutations on its replication and liver disease severity. *Open Virol J* 2013; **7**: 12-18 [PMID: 23400390 DOI: 10.2174/1874357901307010012]
- 90 **Yeh CT**, So M, Ng J, Yang HW, Chang ML, Lai MW, Chen TC, Lin CY, Yeh TS, Lee WC. Hepatitis B virus-DNA level and basal core promoter A1762T/G1764A mutation in liver tissue independently predict postoperative survival in hepatocellular carcinoma. *Hepatology* 2010; **52**: 1922-1933 [PMID: 20814897 DOI: 10.1002/hep.23898]
- 91 **Chen L**, Zhang Q, Chang W, Du Y, Zhang H, Cao G. Viral and host inflammation-related factors that can predict the prognosis of hepatocellular carcinoma. *Eur J Cancer* 2012; **48**: 1977-1987 [PMID: 22325840 DOI: 10.1016/j.ejca.2012.01.015]
- 92 **Kwon HJ**, Jang KL. Natural variants of hepatitis B virus X protein have differential effects on the expression of cyclin-dependent kinase inhibitor p21 gene. *Nucleic Acids Res* 2004; **32**: 2202-2213 [PMID: 15107488 DOI: 10.1093/nar/gkh553]
- 93 **Liu S**, Zhang H, Gu C, Yin J, He Y, Xie J, Cao G. Associations between hepatitis B virus mutations and the risk of hepatocellular carcinoma: a meta-analysis. *J Natl Cancer Inst* 2009; **101**: 1066-1082 [PMID: 19574418 DOI: 10.1093/jnci/djp180]
- 94 **Xu L**, Qian G, Tang L, Su J, Wang JS. Genetic variations of hepatitis B virus and serum aflatoxin-lysine adduct on high risk of hepatocellular carcinoma in Southern Guangxi, China. *J Hepatol* 2010; **53**: 671-676 [PMID: 20650537 DOI: 10.1016/

- j.jhep.2010.04.032]
- 95 **Geng X**, Harry BL, Zhou Q, Skeen-Gaar RR, Ge X, Lee ES, Mitani S, Xue D. Hepatitis B virus X protein targets the Bcl-2 protein CED-9 to induce intracellular Ca²⁺ increase and cell death in *Caenorhabditis elegans*. *Proc Natl Acad Sci USA* 2012; **109**: 18465-18470 [PMID: 23091037 DOI: 10.1073/pnas.1204652109]
 - 96 **López-Cabrera M**, Letovsky J, Hu KQ, Siddiqui A. Multiple liver-specific factors bind to the hepatitis B virus core/pregenomic promoter: trans-activation and repression by CCAAT/enhancer binding protein. *Proc Natl Acad Sci USA* 1990; **87**: 5069-5073 [PMID: 2367525]
 - 97 **Yuh CH**, Chang YL, Ting LP. Transcriptional regulation of precore and pregenomic RNAs of hepatitis B virus. *J Virol* 1992; **66**: 4073-4084 [PMID: 1602534]
 - 98 **Tanaka Y**, Mukaide M, Orito E, Yuen MF, Ito K, Kurbanov F, Sugauchi F, Asahina Y, Izumi N, Kato M, Lai CL, Ueda R, Mizokami M. Specific mutations in enhancer II/core promoter of hepatitis B virus subgenotypes C1/C2 increase the risk of hepatocellular carcinoma. *J Hepatol* 2006; **45**: 646-653 [PMID: 16935384 DOI: 10.1016/j.jhep.2006.06.018]
 - 99 **Kaneko M**, Uchida T, Moriyama M, Arakawa Y, Shikata T, Gotoh K, Mima S. Probable implication of mutations of the X open reading frame in the onset of fulminant hepatitis B. *J Med Virol* 1995; **47**: 204-208 [PMID: 8551270]
 - 100 **Uchida T**, Saitoh T, Shinzawa H. Mutations of the X region of hepatitis B virus and their clinical implications. *Pathol Int* 1997; **47**: 183-193 [PMID: 9103208]
 - 101 **Günther S**, Piwon N, Will H. Wild-type levels of pregenomic RNA and replication but reduced pre-C RNA and e-antigen synthesis of hepatitis B virus with C(1653) --> T, A(1762) --> T and G(1764) --> A mutations in the core promoter. *J Gen Virol* 1998; **79** (Pt 2): 375-380 [PMID: 9472623 DOI: 10.1099/0022-1317-79-2-375]
 - 102 **Bréchet C**. Pathogenesis of hepatitis B virus-related hepatocellular carcinoma: old and new paradigms. *Gastroenterology* 2004; **127**: S56-S61 [PMID: 15508104]
 - 103 **Kremsdorf D**, Soussan P, Paterlini-Brechot P, Brechot C. Hepatitis B virus-related hepatocellular carcinoma: paradigms for viral-related human carcinogenesis. *Oncogene* 2006; **25**: 3823-3833 [PMID: 16799624 DOI: 10.1038/sj.onc.1209559]
 - 104 **Feitelson MA**, Bonamassa B, Arzumanyan A. The roles of hepatitis B virus-encoded X protein in virus replication and the pathogenesis of chronic liver disease. *Expert Opin Ther Targets* 2014; **18**: 293-306 [PMID: 24387282 DOI: 10.1517/14728222.2014.867947]
 - 105 **Zhang H**, Shan CL, Li N, Zhang X, Zhang XZ, Xu FQ, Zhang S, Qiu LY, Ye LH, Zhang XD. Identification of a natural mutant of HBV X protein truncated 27 amino acids at the COOH terminal and its effect on liver cell proliferation. *Acta Pharmacol Sin* 2008; **29**: 473-480 [PMID: 18358094 DOI: 10.1111/j.1745-7254.2008.00764.x]
 - 106 **Ali A**, Abdel-Hafiz H, Suhail M, Al-Mars A, Zakaria MK, Fatima K, Ahmad S, Azhar E, Chaudhary A, Qadri I. Hepatitis B virus, HBx mutants and their role in hepatocellular carcinoma. *World J Gastroenterol* 2014; **20**: 10238-10248 [PMID: 25132741 DOI: 10.3748/wjg.v20.i30.10238]
 - 107 **Zhou F**, Xu H, Chen M, Xiao H, Zhang Z, Lu Y, Ren J, Dong J. X gene/core promoter deletion mutation: a novel mechanism leading to hepatitis B 'e' antigen-negative chronic hepatitis B. *Mol Med Rep* 2014; **10**: 799-803 [PMID: 24841504 DOI: 10.3892/mmr.2014.2248]
 - 108 **Wang WL**, London WT, Feitelson MA. Hepatitis B x antigen in hepatitis B virus carrier patients with liver cancer. *Cancer Res* 1991; **51**: 4971-4977 [PMID: 1654208]
 - 109 **Chen GG**, Li MY, Ho RL, Chak EC, Lau WY, Lai PB. Identification of hepatitis B virus X gene mutation in Hong Kong patients with hepatocellular carcinoma. *J Clin Virol* 2005; **34**: 7-12 [PMID: 16087118 DOI: 10.1016/j.jcv.2005.01.006]
 - 110 **Geng X**, Huang C, Qin Y, McCombs JE, Yuan Q, Harry BL, Palmer AE, Xia NS, Xue D. Hepatitis B virus X protein targets Bcl-2 proteins to increase intracellular calcium, required for virus replication and cell death induction. *Proc Natl Acad Sci USA* 2012; **109**: 18471-18476 [PMID: 23091012 DOI: 10.1073/pnas.1204668109]

P- Reviewer: Aghakhani A, Ma L, Wang K **S- Editor:** Ma YJ

L- Editor: A **E- Editor:** Ma S





Avoiding hepatic metastasis naturally: Lessons from the cotton top tamarin (*Saguinus oedipus*)

Martin Tobi, Peter Thomas, Daniel Ezekwudo

Martin Tobi, Department of Surgery, Aleda E. Lutz VAMC, Saginaw, MI 48601, United States

Martin Tobi, Daniel Ezekwudo, Internal Medicine Disciplines, Central Michigan University College of Medicine, Saginaw, MI 48601, United States

Peter Thomas, Department of Surgery, Creighton University School of Medicine, Omaha, NE 68178, United States

Author contributions: All authors participated equally in all aspects of this review.

Conflict-of-interest statement: The authors have no conflict of interest related to the manuscript.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Martin Tobi, MB, ChB, Department of Surgery, Aleda E. Lutz VAMC, 1500 Weiss Street, Saginaw, MI 48601, United States. martin.tobi@va.gov
Telephone: +1-989-4972500-13934
Fax: +1-989-32114946

Received: March 22, 2016

Peer-review started: March 22, 2016

First decision: April 14, 2016

Revised: April 26, 2016

Accepted: May 21, 2016

Article in press: May 23, 2016

Published online: June 28, 2016

Abstract

Much has been written about hepatic metastasis and animal models abound. In terms of the human experience, progress in treating this final common pathway, a terminal event of many human malignancies has been relatively slow. The current thinking is that primary prevention is best served by early detection of cancer and eradication of early stage cancers by screening. Some cancers spread early in their course and the role of screening may be limited. Until relatively recently there has not been a pathfinder model that makes the evasion of this unfortunate event a reality. This review discusses such an animal model and attempts to relate it to human disease in terms of intervention. Concrete proposals are also offered on how scientists may be able to intervene to prevent this deadly progression of the cancer process.

Key words: Cotton top tamarin; Hepatic metastasis; Carcinoembryonic antigen; Fibulin-5; Common marmoset

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Hepatic metastasis is a terminal event. Avoiding this complication would prolong life and current understanding of inflammatory mediators allows possible secondary intervention. The cotton top tamarin (CTT), like humans develops inflammatory bowel disease complicated by colorectal cancer but avoids liver metastasis. We suggest 5 mechanisms by which CTT avoid liver spread. They involve changes in ICAMs and their receptors, carcinoembryonic antigen (CEA) family mediators of angiogenesis, post translational modifications of molecules like CEA, and increased expression of anti-proliferative agents such as fibulins. This changes our perception from "monkey

see, monkey do” to “see what the monkeys do and do the same”. Possible avenues of intervention are suggested.

Tobi M, Thomas P, Ezekwudo D. Avoiding hepatic metastasis naturally: Lessons from the cotton top tamarin (*Saguinus oedipus*). *World J Gastroenterol* 2016; 22(24): 5479-5494 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5479.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5479>

INTRODUCTION

There are many animal models for human cancer and most of these do not occur naturally but are designed to mimic human disease by introduction of mutagen and may also employ promoters. Natural models lack this human “design” element but historically the differences may enhance understanding of disease causality and management for example, the Roux Sarcoma virus in the Plymouth Barred Rock fowl (chickens) and Bittner milk factor in murine mammary carcinoma among others. It is however unusual for a single animal to provide more than 1 natural model for disease^[1].

Enter the cotton top tamarin (CTT), a pint-sized lower order, New World primate of the order Callitrichidae^[2]. This remarkable monkey provides a model for human disease in inflammatory bowel disease and potential mechanisms^[3,4], cancer of the lymphatic system^[5], large bowel cancer^[6], and immune-altered states^[7], and recently liver metastases^[8]. While a review of these models apart from the latter is beyond the scope of this review, it is significant that the example of the liver metastasis model could best be described as a “negative” or “avoidance model” in that the CTT with colon cancer avoids liver metastasis with an estimated frequency of 97.2%.

The hypotheses of how the CTT achieves this are open for discussion and debate has been ongoing since the first symposium, reported in 1985^[9] where anatomical blood vessel variance was advanced as an explanation. This was refuted by veterinarians since the anatomy is similar to most mammals including humans (N. Clapp- personal communication). Our group has been exploring hypotheses based on adhesion molecules known to be important in human liver metastasis. While the article will focus on these molecular families in tamarins, it is important to first discuss our understanding of their importance in human “metastogenesis” as the findings in the CTT may enable intervention in the human where over 70% of GI cancer victims succumb to death caused by hepatic metastases.

Hypotheses of hepatic metastases in humans

Cancers of the colon and rectum comprise 14.5% of

all cancers diagnosed in the United States (149880) and 11.1% of cancer deaths (51710) each year^[10]. The primary treatment for large bowel cancer is surgery. Patients who recur do so largely in the liver, lung or peritoneum. While combination chemotherapy can increase survival even in advanced disease it is rarely long term. Metastatic colorectal cancer is therefore a major public health problem. Colorectal cancer is associated with chronic inflammation^[11-13]. Inflammatory cytokines can be induced by interaction of terminally differentiated macrophages with the cancer associated glycoprotein carcinoembryonic antigen (CEA)^[14]. CEA (CEACAM5) is a large heavily glycosylated glycoprotein of indeterminate function in the normal tissue in which it is expressed^[15,16]. Discovered in 1965 by Gold and Freedman^[17] it has become the most used tumor marker for colorectal cancers but failed to live up to the early expectations for use as an early detector of cancer^[15,18]. It is a glycoprotein that is a member of a large gene family that consists of 29 genes divided into three subgroups that include the CEA-like glycoproteins and the pregnancy-specific glycoproteins (PSGs)^[19]. These CEA like proteins are members of the much larger immunoglobulin supergene family^[15]. The nomenclature for the entire CEA and PSG families is published in Beauchemin *et al*^[20].

CEA

CEA is a GPI anchored glycoprotein with an average MW of 180 kDa this varies depending on the degree of glycosylation of the native protein. CEA contains 651 amino acids and these are found in 7 immunoglobulin like domains, an IgV domain at the N-terminus and six disulfide bridge linked IgC domains^[15,16]. There are 29 possible sites for N-linked glycosylation and they tend to be of the complex tetra antennary type^[15]. Recently a large number of functions in cancer cells have been attributed to CEA. These include a role in cell-cell adhesion^[21,22], apoptosis (anoikis)^[23], inflammatory responses leading to increased hepatic and possibly lung metastasis^[14] and angiogenesis^[24].

CEA AND ITS IMPACT ON LIVER METASTASIS: INDUCTION OF INFLAMMATORY RESPONSES IN THE LIVER

Liver metastasis is the major cause of death in patients with colorectal cancers. There is good evidence that cytokines play a major role in preparing the liver for implantation and subsequent growth of cancer cells^[14]. It appears that a relationship exists between the colon cancer derived glycoprotein CEA and macrophage related cytokine production particularly associated with CEA Kupffer cell interactions in metastasis of colorectal

cancers to the liver. The discovery of CEA in 1965^[17] marked a turning point in the study of cancer and led what might be called the age of tumor markers. CEA was the first commercially available tumor marker and may be considered the prototype^[25]. Serum elevations of CEA (above 3 ng/mL) are seen in about 60% of colorectal cancer patients at presentation^[26] and elevated levels are also seen in a number of other cancers including breast, gastric, lung and pancreas. Unfortunately, the early premise that CEA levels could be used as a screening test for colorectal cancer was not fulfilled. CEA was found not to be cancer specific and high false positive and false negative rates precluded its use in early detection of cancer. However, CEA is a useful marker in the determination of prognosis^[27] and for detection of recurrence in the post-surgical follow up of colorectal cancer patients^[28]. CEA has also been used for the detection of occult tumor by radioimmuno-detection^[29] and as a target for treatment using radioimmuno-therapy^[30]. In addition a number of studies have used CEA as the target antigen for immunotherapy^[31]. Clinically CEA is very well understood, but its biological role is not clear and its function in the normal colonocyte still evades us. However over the years a number of functions mostly associated with tumor cell behavior have been attributed to CEA. We focused on the molecular interactions between CEA and its receptor CEAR in colorectal cancer cells and how disruption of these interactions and inhibition of cytokine production will affect the spread and growth of colorectal cancers. Designing an effective mimetic-based therapy requires the identification of the responsible molecules, mechanisms of action and regulation.

The most important functions of CEA are the effects associated with tumor cell survival whether it is at the primary site or at a distant metastasis. High levels of CEA are also associated with malignant ascites from colorectal cancer (CRC)^[32]. CEA interacts with liver Kupffer cells through a cell surface receptor (CEAR) that we have identified as the heterogeneous RNA binding protein (hnRNP) M4^[33]. The hnRNP group of proteins are multifunctional and are involved in many cellular events including alternative splicing, translational regulation and packaging of transcripts^[34]. These proteins have also been implicated in the regulation of mRNA stability and translation in many cancers^[35]. CEAR is a highly conserved protein that can bind both RNA and DNA and can transport mature RNA to the cytoplasm as well as acting as a splicing factor^[36]. It can also be expressed both on the cell surface, in the cytoplasm and in the nucleus where it is most commonly found^[37]. There are 4 isoforms of hnRNP M known^[38]. Two of which we have shown bind CEA^[33]. One form (hnRNP M4) which we originally identified as the CEA receptor has a 38 amino acid deletion between the first and second RNA binding domains. The longer form also binds CEA^[33]. The two other forms have not been identified

other than as proteins on 2D gels. CEAR will also bind to CEA in HT-29 colon cancer cells although the functional significance of this is not known. It has been suggested that it may be involved with the resistance afforded to anoikis^[39,40]. Binding to CEAR occurs *via* a penta-peptide motif (PELPK) located at the hinge region between the CEA's N-terminal and first immunoglobulin loop domain. This interaction produces cytokines by activating a signaling cascade and these cytokines alter the liver microenvironment such that it becomes more hospitable to the implantation and growth of the cancer cells^[41,42]. Production of both IL-6 and IL-10 by CEA stimulated Kupffer cells improves the survival of highly metastatic human colorectal cancer cells in the nude mouse intra-splenic injection model for liver metastasis^[43] (see Figure 1).

CEA AS AN ADHESION MOLECULE

In 1989 Benchimol *et al*^[22] demonstrated the first potential function for CEA. By transfecting tumor cells with the CEA gene they showed increased cell to cell adhesion and demonstrated that this could be inhibited using anti CEA antibodies. Adhesion was due to interactions between the N-terminal and the 5th and 6th (A3B3) immunoglobulin domains^[44]. This suggested that these interactions may play a role in tumor cell behavior and in the development of metastases.

CEA AND ANOIKIS

Another important function that has been attributed to CEA expression is inhibition of apoptosis in particular the apoptosis caused by absence of attachment of cells to a substratum (anoikis)^[23,40]. This would play an important role in metastases formation to a distant organ as unattached tumor cells in the circulation would be more susceptible to anoikis.

CEA AND ANGIOGENESIS

Here, we suggest a relationship between CEA, its receptor CEAR and angiogenesis. Others have also suggested a relationship between CEA and angiogenesis though they did not establish it as causal or identify potential mechanisms^[45,46]. Recently, however they showed that secreted (soluble) CEA can directly activate endothelial cells *via* integrin $\beta 3$ signaling^[24,47]. We suggest that an alternate mechanism may also exist involving the relationship between CEA and its receptor CEAR. Interaction of CEAR with CEA in tumor associated stromal cells particularly macrophages is important. Disrupting that relationship may affect tumor growth and progression

It is significant that Low-Marchelli *et al*^[48] have shown that CCL2 recruits macrophages to promote angiogenesis. We therefore suggest that colon cancer cells recruit macrophages by secreting MCP-1 and these macrophages are activated by tumor derived

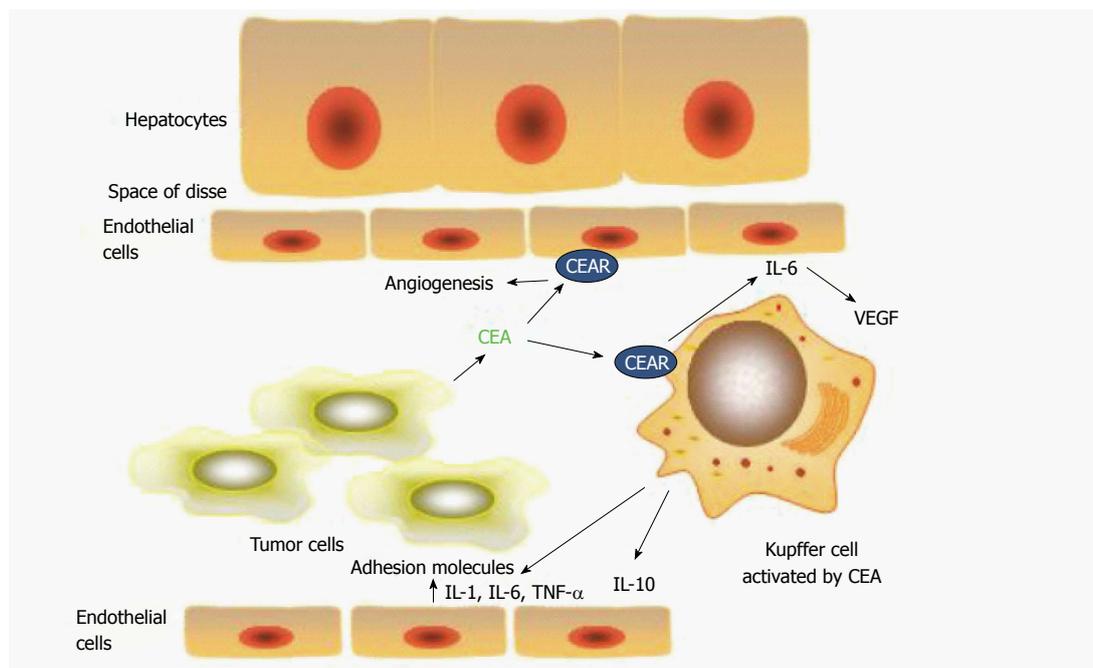


Figure 1 Schematic of the interactions of carcinoembryonic antigen in the hepatic sinusoid. Interactions of carcinoembryonic antigen (CEA) in the hepatic sinusoid. CEA released by the tumor cell binds with hnRNP M (CEAR) on the Kupffer cell surface resulting in release of the cytokines interleukin (IL)-1, IL-6, IL-10 and TNF- α . Effect of CEA induced cytokines on tumor cell interactions in the hepatic sinusoid. Cytokines IL-1, IL-6, IL-10 and TNF- α produced by Kupffer cells have a number of effects on the tumor cell microenvironment. These include up-regulation of adhesion molecules on hepatic sinusoidal endothelial cells. The most important of these seem to be E-selectin and ICAM-1. Cytokines such as IL-6 and IL-8 are pro-angiogenesis and they may also effect growth at the distant site^[14,42].

CEA to secrete IL-6 and IL-8 both known pro-angiogenic factors^[49]. Anti angiogenic factors are also produced (IL-10) and an imbalance between these two competing factions will tend towards pro-angiogenesis.

responses in PMA activated THP-1 macrophages. These types of study are important for the development of rational therapies that will enhance those currently available.

TARGETING CEA/CEAR INTERACTIONS AS A MEANS OF INHIBITING CYTOKINE PRODUCTION AS AN ANTI-METASTATIC THERAPY IN COLORECTAL CANCER

Because transformed cells home into tissues from the circulation in a highly selective way through complex molecular mechanisms, it provides a template for targeted therapy. Designing an effective mimetic-based therapy requires the identification of the responsible molecules and their mechanisms of action along with their regulation. The investigation of methods to block or modulate CEAR function *in vitro* and *in vivo* and relating this to tumor growth and development will increase our understanding of the molecular and biological mechanisms involved in the progression of gastrointestinal cancers. The investigation of methods to block or modulate CEAR function *in vitro* and *in vivo* and relating this to tumor growth and development has increased our understanding of the molecular and biological mechanisms involved in the progression of colorectal cancers. We have shown that the binding peptide YPELPK will induce cytokine responses in macrophages activated by CEA^[50]. We have shown that the binding peptide YPELPK will induce cytokine

OTHER POTENTIALLY PRO/ANTI-METASTATIC MOLECULES STUDIED

Fibulin-5

During the initial development of metastasis, the adhesion of tumor cells is mainly mediated through binding of extracellular matrix components to cell surface receptors^[51]. Interestingly, one study^[52] found an association of fibulin-5, the newest member of the fibulin family of extracellular matrix glycoproteins, with hepatic metastasis of colorectal carcinoma. Since fibulin-5 plays an important role in antagonizing angiogenesis^[51-53] in humans, we sought to confirm the expression of fibulin-5 expression level in the CTT.

Avoidance hypotheses in the CTT

Our understanding of cancer and metastases in primates began when colon cancer was described in old world monkeys as early as 1914 and then in the new world in inter alia, Goeldi's monkey in the early 1960s. The latter, *Callimico goeldii* shares the Colombian habitat with the CTT and is a small black-furred New World lower order primate. At that time tens of thousands of CTT were imported by the pharmaceutical industry utilizing the "tamarin alarm reaction" to test the efficacy of anxiolytics although literature to document

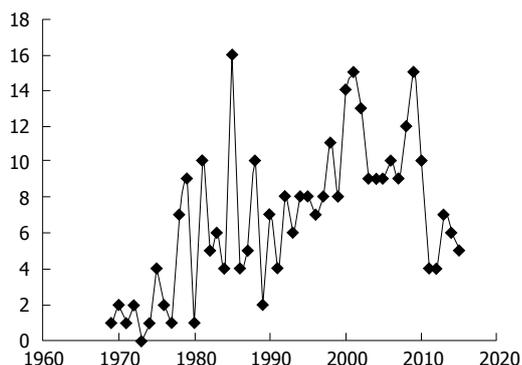


Figure 2 Graphic representation of published cotton top tamarin articles by year as estimated by a typical online search. The line chart is not necessarily inclusive of all cotton top tamarin (CTT) articles but merely confined to the search terms used. It is therefore used mainly to demonstrate the overall trends in annual publications devoted to this animal "CTT supermodel".

this is difficult to come by utilizing electronic search methods. Problems of CTT husbandry arose when the animals developed the "wasting marmoset syndrome"^[54] postulated to be caused by a coronavirus but later effectively countered by improving the nutritional intake. The major challenge facing the CTT is the reduction of its numbers in its shrinking natural habitat in northwestern Colombia and most CTT are today found in zoos. As a result the species was listed as endangered in 1973 by the Convention on International Trade in Endangered Species of Wild Flora and Fauna (CITES)^[55]. Since that time, the numbers of articles on the CTT had increased exponentially. This attests to the interest of the scientific community in the CTT specifically. Despite this interest, primate research in the United States is in decline with the closures of the ORAU (Oak Ridge Associated Universities) facility and that of the NEPC (New England Primate Center) which housed a significant proportion of CTT. The waning of public support and therefore funding opportunities combined with the expense of operations has largely been the cause of this decline. This is regrettable as it is generally not appreciated that monkey publication numbers are an extensive part of the scientific literature (678000 Medline™-listed publications) attesting to their usefulness and their command of the attention of the scientific community. Now, specifically CTT publications which were formerly about one tenth of non-human primate-themed publications (398 total CTT per National Library of Medicine ticket 28045-54653), are in decline in the example depicted graphically (Figure 2).

In view of the importance of this animal model applicable to many models of human disease, a CTT symposium was convened resulting in the publication of numerous articles in a supplement published in the journal *Digestive Diseases and Sciences* in 1985^[9]. Thereafter a comprehensive book on the CTT was edited by Dr. Neil K. Clapp and published in 1993 by CRC Press^[2]. This review seeks to build on that experience specifically focusing on the ability of the

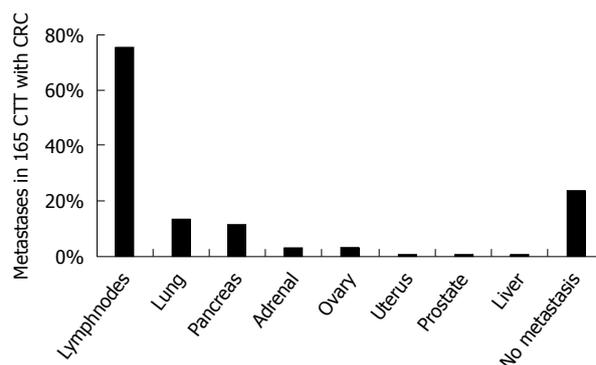


Figure 3 Schematic bar diagram depicting proportion of metastases in cotton top tamarin with colorectal cancer. The bar diagram shows the distribution of metastasis based on the ORAU colony cancer statistics. The paucity of liver metastases is remarkable. CTT: Cotton top tamarin; CRC: Colorectal cancer.

CTT to avoid hepatic metastases even though a large proportion of the animals develop advanced colorectal cancer. This is a unique feature of the CTT not typically found in other marmosets. It would appear that CTT peritoneal metastasis and ascites is also not common as it is with terminal human disease.

CTT MODEL FOR HEPATIC METASTASIS

The CTT develops colitis in the juvenile years and about one third (34.5%) go on to develop colon cancer and die of the disease. An observational study showed only 1.2% of animals had hepatic metastases^[56] (Figure 3).

The etiology of the cancer seems to be the de novo inflammation-dysplasia-cancer variety although animals with adenomas have been described^[56]. The risk of cancer deaths shows an increase from 3-8 years and then a gradual decline after age 12 similar to the recently described risk in humans with positive family history of CRC^[57]. The etiology is still unknown but stress and micro-organisms may play a role^[3,4]. Over half have multiple primaries diffusely distributed but the descending colon has the greatest proportion similar to the rectosigmoid colon predominance in ulcerative colitis cancer location.

CTT COLON CANCER BIOLOGY VIS A VIS HUMAN

Despite similarities in cancer biology between CTT and humans it can be seen from Table 1 that the anticipated mutations are either not described or characterization has not been attempted.

When considering changes associated with metastases^[63] no CXCR4, CXCL12 (fusins), CCR7 (chemokine receptor 7) chemokines thought to be important in metastases have been described as yet in the CTT. Interestingly CXCL12 has been described in the common marmoset^[64] and thus likely to be expressed in the CTT but is not necessarily a given.

Table 1 Comparison of cancer genetics in the cotton top tamarin and human

Stage of neoplasia	Sporadic human CRC	Human IBD-associated CRC	CTT colitis-CRC
Normal-appearing mucosa	APC initiator	Inflammation leading to mutations listed below	No cell line mutations in APC exon 4 and 15 ^[58]
Early adenoma/indefinite dysplasia	Aneuploidy, methylation, Sialyl Tn	Aneuploidy, methylation, Sialyl Tn, MSI	90% diploid; no methylation ^[59]
Early promotor	MSI; <i>kRAS</i> , COX-2	DCC	<i>kRAS</i> absent ^[60]
Intermediate adenoma/low-grade dysplasia	c-src	c-src	c-src ND
Intermediate promoter	DCC/DPC4	<i>kRAS</i>	DCC/DPC4 ND; <i>kRAS</i> absent ^[60]
Late promoter	p53	APC	p53 ND; as above for APC exon 4 and 15 ^[58]

Adapted from Itzkowitz and Harpaz 2004^[61]; and Mattar MC, Lough D, Pishvaian MJ, Charabaty A^[62]. APC: Adenomatous polyposis coli gene; MSI: Microsatellite instability; DCC: Deleted in colon cancer; *kRAS*: V-Ki-ras2 Kirsten rat sarcoma viral oncogene homolog; COX-2: Cyclooxygenase 2 gene; c-src: Rous sarcoma gene that encodes for usual C-terminal inhibitory phosphorylation site (tyrosine-527); DPC4: Deleted in pancreatic carcinoma, locus 4; p53: Protein 53 kilodaltons; ND: Not done.

The human kallikrein zymogens (hK2) closely related to prostate specific antigen (PSA) have been studied in the CTT^[65]. They concluded that the CTT has no functional hK2 or PSA. Since these moieties regulate cancer cell survival and growth the CTT may be a natural knock-out model for the study of this topic. We see in these findings yet another potential mechanism by which the CTT impedes cancer proliferation. Likely Toll-like receptors (TLR) also remain to be discovered in the CTT and may have likely similar functions to the human.

Since lectins^[66] and adhesion molecules have been described in the CTT we have focused our attention on these molecules, particularly CEA. We also explored extracellular matrix proteins as the interaction between these proteins and cells play an important role in tumorigenesis and metastases. One such protein, fibulin-5^[51-53] has calcium-binding EGF-like domains and associate with vascular and vascular structures. It is thought therefore to have anti-angiogenic actions and be an inhibitor of metastasis. It has not as yet been studied in the CTT.

CEA HOMOLOGUES IN THE CTT AND HUMAN

Cross reactive CEA species in monkeys were known when we published our seminal paper in 1994^[67]. Haagensen *et al*^[68] had reported CEA in higher order non-human primates, including the chimpanzee and gorilla. A mouse analogue had also been described by Abraham Fuks' group in 1989^[69]. Our initial experiments used a number of antibodies to CEA including the T84.66 antibody developed by Dr. J. Shively's laboratory group at the Beckman Institute at the City of Hope which was used in the CEA kit commercialized by RocheTM. This antibody when used in immunohistochemistry produced consistently negative results when using colon CTT fixed tissues and later native antigen in CTT colon tissue extracts in early experiments^[70]. We tested other antigens such

as organ specific neoantigen^[71], thought to be the mediator of the lymphocyte adherence inhibition test for cancer and found reactivity^[72]. In the early 1990's Karel Kithier of the Department of Pathology at Wayne State University was beta-testing a CEA kit produced by Tosoh Medics (CEA-AIA PACK kitTM) which seemed to have a greater proportion of positive results when compared to another CEA kit marketed by Abbot. These results were expanded and confirmed and found to yield similar levels in CTT and humans with IBD^[60,73] which opened a pathway to possible intervention^[33] in addition to other cancer related moieties such as telomerase^[60]. In order to show the CEA molecular moieties recognized by the kit we obtained both solid phase and tracer antibodies from Hybritech Inc., from Dr. Harry Rittenhouse. The resultant Western blotting^[67] showed a specific 50 kDa band shared by CTT and humans (Figure 4).

A control blot using T84.1EC monoclonal which reacts with both NCA and CEA did not recognize the 50 kDa shared antigen in the same samples. Based on the known epitopes on the antibodies concerned we were able to speculate that the shared antigen included the Ig-like loop domains however the exact structure of this moiety remains unknown. In this publication we did hypothesize that "the smaller CTT CEA molecule might lack the critical peptides necessary for uptake and hence explain the paucity of liver involvement in CTTs with CRC".

EXPRESSION OF THE CEA GENE FAMILY OF PROTEINS IN THE CTT AND HUMANS

The next question we needed to answer was if an entire repertoire of CEA-family adhesion molecules were expressed by the CTT and if this subspecies was unique in this respect in the *Callitrichidae* family. In a later paper in 2000^[73] we explored these questions by expanding the repertoire of antigens and extending our observations to include a close cousin of the CTT, the common marmoset (*Callithrix jacchus* - CM) which is

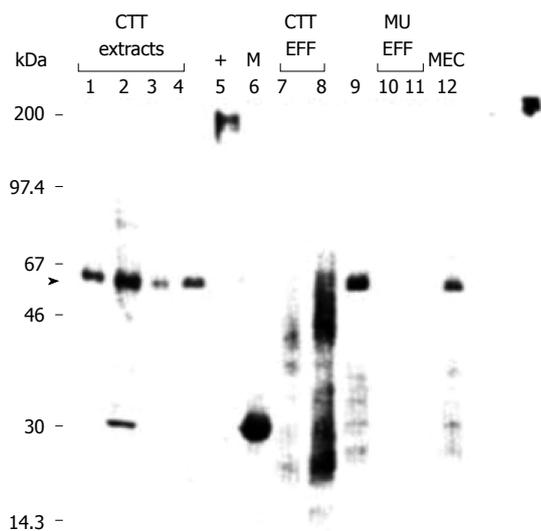


Figure 4 Specific carcinoembryonic antigen bands shared by cotton top tamarin and humans. Western blot using solid phase anti-CEA monoclonal antibody. Immunoblotting was performed using 5.6 pg protein/ml antibody after electrophoresis of a 12% SDS-polyacrylamide gel run under reducing conditions. Relative mobility (M) is shown on the left, and the type of samples loaded at 10 pg protein/lane are shown above the numbered lanes. Cotton-top tamarin extracts are in lanes 1 to 4 and effluent samples in 7 and 8. An extract from a patient with histologically proven rectal cancer and IBD is in lane 9, with human effluent samples in lanes 10 and 11 and human meconium in lane 12. Lane 5 contains the positive human CEA control, and the M, markers (M) are in lane 6. An arrow indicates the M, - 50000 band evident also in human extract (lane 5) likely a deglycosylated moiety. (Published with permission of Elsevier^[67] and modified). CEA: Carcinoembryonic antigen.

not endangered but does develop colitis uncomplicated by cancer. Aware that the changes likely to influence CEA uptake were at the N-terminal end of the CEA molecule we used antibodies directed at epitope in that region, including the aforementioned Tosoh Medics™ kit and T84.66 antibody to quantify the CEA in colonic washings.

We found that the concentration of CEA in washings was sevenfold times higher than in washings from humans with IBD. T84.66 antibody was able to demonstrate a faint high MW band in a specimen of colonic washing from a CTT by immunoblot. In the Western blot using T84.66 we could now also demonstrate specific bands (immunohistochemistry had been consistently negative) in CTT washings and tissue extract of Mr ≥ 90 kDa which is the size of the non-glycosylated protein core of the CEA molecule^[74]. CEA levels assayed by the Tosoh™ kit in CTT extracts were significantly higher than those from CM extracts ($P < 0.005$). Consistent with this finding was a high mean value in sera of CTT (134 ng/mL) compared to undetectable levels in the CM. NCA levels tended to be the lowest in the CTT as compared to humans and CM. Both animals' samples were low when reacted with a CEA-family antibody. In contrast binding with the anti-BGP was seen in both animals. We concluded that the similarity between human and tamarin native CEA is greater than previously believed and that it was likely that the N-terminal epitopes were conserved in the

tamarins.

OTHER ANTIGENS SHARED BY CTT AND HUMANS

By the end of 2000 we published an expanded group of antibody assays involving 7 additional common epitopes representing most colonic cell types and blood group/carbohydrate antigens, some of them accepted tumor markers^[60,70-76]. Mucin antigens, epidermal growth factor (EGFR) and telomerase were shared by CTT and humans but *k-ras* p21 and adenoma antigen (Adnab-9) were not. This supported the notion that carcinogenesis was likely *de novo* and did not progress through the adenoma-carcinoma sequence. Our finding of EGFR reactivity in the CTT provided a basis^[76] for fibulin-5 role in the anti-metastatic armamentarium and OSN (by reactivity of BAC 18 monoclonal) suggesting similarities in anti-cancer immune responsiveness *via* the lymphocyte migration reaction. We were able to demonstrate also significant levels of EGFR epitope in the urine of 5 CTT (0.152 ± 0.053 absorbance OD-background at 405 μm) by ELISA. CEA reactivity with various antibodies in these samples were low (mean < 0.05). Blood Group Substances (BGS) were detectable [Span 0.063; FBB 0.103; CaCo 0.054] as were Bac18.1 (anti-OSN antibody) at 0.06]. Wild-type (Table 1) Ras p21; src, common membrane antigen (CMA), were negative corresponding to the data derived from CTT tissue. The only pieces of the puzzle missing were the questions of CEA molecular homology and how to tie the available data together to explain how the CTT dodges liver metastases.

DEFINITIVE STUDIES ON THE HOMOLOGY OF THE CEA MOLECULE

A comparative study of the C-terminus of the CEA molecule by the Stanners' group looked at multiple primate and prosimian species. The membrane linkage in CTT and other monkeys was found to be GPI anchorage rather than transmembrane (TM) seen in mice and rats^[77]. This paper showed that the more versatile glycoposphatidyl inositol (GPI) linkage with a greater functionality constellation likely confers a tendency to tumorigenesis. Mutational packages evolved over time, one applicable to most primates including humans, and the second confined to the Cebidae radiation of New World monkeys leading to inhibition of cell differentiation. The rate of Ka (indicator of selective pressure acting on a protein-coding gene) nonsynonymous mutations for this radiation is 7X higher than the average Ka in primates. This led us to postulate that if such mutations occur at the C-terminus, it is quite feasible that similar mutations occur at the N-terminus where they may affect uptake of CEA, inhibiting hepatic metastases. Having

Table 2 Representative somatic sequences from the PELPK region of carcinoembryonic antigen in humans, cotton top tamarin and common marmoset

HCEA-A1	PELPKPSISSNNSKPVEDKDAVAFTCEPETQDA
CTT-WT	PELPKPSISSNNSKPVEDKDAVAFTCEPETQDA
CM-WT	PELPKPSISSNNSKPVEDKDAVAFTCEPETQDA
M1-A1 (CTT)	PEVPKPSISSNNSKPGGDKDAGAFWEPETQDA
M2-A1 (CTT)	PEVSKPSISSNNSKPGGDKDAGAFWEPETQDA
M3-A1 (CTT)	PEVSKPSISSNNSKPVGDKDAGAFWEPETQDA
M4-A1 (CTT)	PEVSKPSISSNNSKPVGDKDAGAFWEPETQDA
M5-A1 (CTT)	PEVSKPYISSNNSNPVENKDAVAFTWEPETQDA
M6-A1 (CTT)	PEVSKPFIFSNNSKPGGDKDAGAFWEPETQDA
M8-A1 (CTT)	PEVSKPFIFRNNSKPGGDKDAGAFWEPETQDA
M10-A1 (CTT)	PELPKPFIFSNNSKPVEDKDAVAFTCEPETQDT
M14-A1 (CTT)	PELPKPFIFSNNSKPVEDKDAVAFTCEPETQDA
M15-A1 (CTT)	PELPKPFIFSNNSKPVEDKDAVAFTCEPETQDA
M16-A1 (CTT)	PELPKPSILSNNSKPVEDKDAVAFTCEPETQDA
M17-A1 (CTT)	PELPKPSIPSNNNSNPVEDKDAVGLTCEPDTQNT
M19-A1 (CM)	PELPKPSISSYNSKPVEDKDAVAFTCEPETQDA
M20-A1 (CM)	PELPKPSISSYNSKPVEDKDAVAFTCEPETQDA
M21-A1 (CM)	PELPKPFIFSNNSKPVEDKDAVAFTCEPETQDA
M24-A1 (CM)	PELPKPSIFSNNSKPVEDKDAVAFTCEPETQDA
M27-A1 (CM)	PELPKPSISSNNSNPVEDKDAVAFTCEPETQDA
M31-A1 (CM)	PEVSKPFIFSNNSKPVGDKDAGAFWEPETQDA
M40-A1 (CM)	PGVVKPFIFRNNSKPVGDKDAGAFWEPETQDA

Sequences changes from those of wild type (germ line) are shown in bold. Published with permission of Springer^[8].

also found common CEA-family and fibulin-5 ligand antigens we resolved to also investigate for parallel pathways of metastasis inhibition.

SUMMARY: SPECIFIC METASTASIS-TARGETED STUDIES IN CTT AND HUMANS

Using DNA extracted from the CTT tissues a Hot start PCR was performed and (1) sequences around the hinge region between the N-terminal and the first loop domain of CEA obtained using a Big Dye Terminator™ sequencing kit. We also immunolabeled human and CTT liver tissue using: (2) BGP (CEACAM1) antibody kindly supplied by Christophe Wagener (University of Freiberg, Germany); (3) CEAR (CEA receptor) polyclonal antibody; and (4) an antibody directed to fibulin-5 supplied by W. P. Schiemann (Department of Pharmacology, University of Colorado, Aurora, CO, United States). Finally, (5) we deglycosylated CTT CEA to determine the degree of N-glycosylation, as glycosylation may correlate with hepatic metastases. We thus examined 5 potential ways in which the CTT may evade hepatic metastases in 10 CTT and 25 humans.

To briefly summarize the observed^[8] results (Table 2) we enumerate: (1) 63% of the CTT had non-synonymous PELPK pentapeptide mutations shown to be essential for hepatic uptake of numerous groups of protein including CEA and stromelysin, laminin, complement protein *etc.* CM had the PELPK mutations

Table 3 Biological attributes of the fibulin family^[51-53]

Attribute	Fibulin-1	Fibulin-2	Fibulin-3	Fibulin-4	Fibulin-5
Invasion of endothelium	Decrease	Increase	Decrease	Unknown	Decrease
Binding of tropoelastin	Moderate	High	low	Moderate	High
Effect on angiogenesis	Unknown	Unknown	Reduced	Unknown	Reduce
MMP/fibronectin change	Increase	Unknown	Reduced	Unknown	Reduced

to a lesser extent (73% vs 29%, *P* < 0.05). Sequence changes in the PELPK binding motif are found in the CTT that prevent binding to CEAR and do not allow activation of Kupffer cells and thus secretion of pro-inflammatory cytokines.

This could account for the high CEA serum levels in the CTT as compared to CM and humans and contribute to the lack of liver metastasis. Zimmer and Thomas in 2001 showed in a minority of patients with advanced colon cancer all of whom had high CEA levels, changes in the PELPK sequence. CEA from these patients could not bind to Kupffer cells. These patients did not have liver metastasis though peritoneal carcinomatosis was common^[78]. Serendipitously: (2) Normal sections of human livers bearing metastatic disease (our intended “negative” controls) showed translocation of the BGP to the cytosol of the hepatocyte whereas CTT livers did not have detectable BGP except in the gallbladder wall. BGP is usually a structural protein found in the bile canaliculus but conceivably, if translocated to the cytosol, may act as a downstream mediator of VEGF, facilitating the progression of liver metastasis. Human cirrhotic and hepatitis livers demonstrated orthotopic BGP compared to human livers bearing metastases (*P* < 0.02) where the BGP had translocated to the cytoplasm (Figure 5).

This may answer the age-old clinical question as to why patients with cirrhosis rarely get hepatic metastases from extrahepatic primary cancers. We also observed: (3) a diminution of CEAR staining (Figure 6) in the CTT liver sections compared to humans (*P* < 0.02), a further barrier to CEA hepatic uptake in the CTT as compared to the human liver. Surprisingly: (4) Extremely robust fibulin-5 hepatocyte labeling was detected in CTT livers (Figure 7) and finally: (5) Minimal glycosylation of CTT CEA was demonstrated as opposed to human CEA. 80% of CTT vs 60% of CM were positive for fibulin-5 binding with functions of the various fibulins contrasted in Table 3.

Hepatocytes were the predominant cell types expressing fibulin-5 in both types of animals but tended to be more intense (Figure 6) in the CTT (0.75 ± 0.289 and 0.4 ± 0.418 respectively) and also seen to be of the same distribution and intensity in the human liver controls. Kupffer cells did not stain (*P* = 0.033 CTT and *P* = 0.09 CM) but occasional bile duct

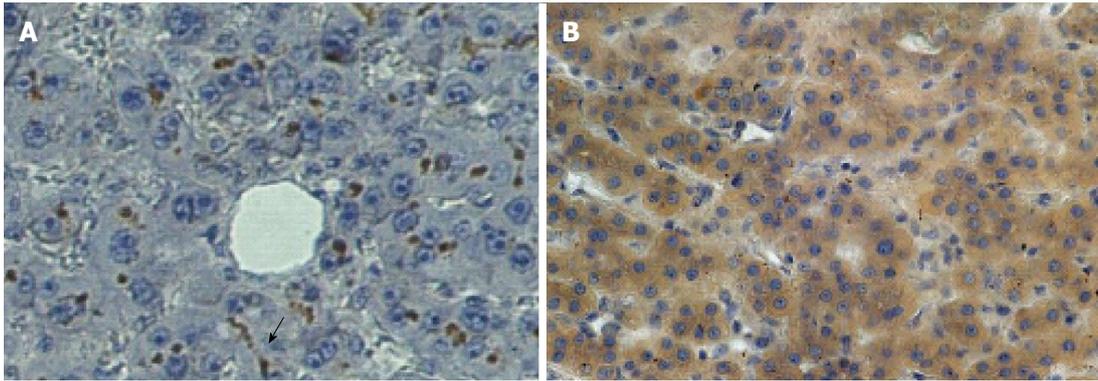


Figure 5 Distribution of CEACAM1 (BGP) in human liver. Photomicrographs of CEACAM1 staining with monoclonal antibody 4D1/C2 showing very intense brown staining mainly in the distribution of the biliary canaliculus in normal human liver (A). In contrast, in the normal portion of a liver from a patient with hepatic metastasis (B), dark staining is seen in the cytoplasm of the hepatocytes with no canalicular staining evident. Published with permission of Springer^[6]. The arrow points to the typical distribution of bound ant-BGP in the bile canaliculus in the normal liver (A) at center, bottom.

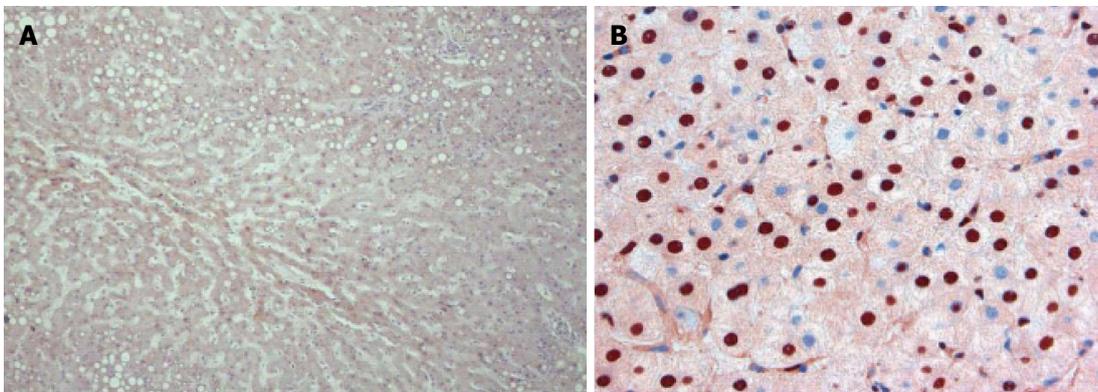


Figure 6 Distribution of carcinoembryonic antigen receptor in cotton top tamarin and human. The distribution of CEAR in the CTT (A) at low power can be seen in the cytoplasm of the hepatocyte by the light brown stain. In humans (B) at a higher power the increased intensity of staining in hepatocyte nuclei can be clearly seen. Published with permission of Springer^[6]. CEA: Carcinoembryonic antigen; CTT: Cotton top tamarin; CEAR: Carcinoembryonic antigen receptor.

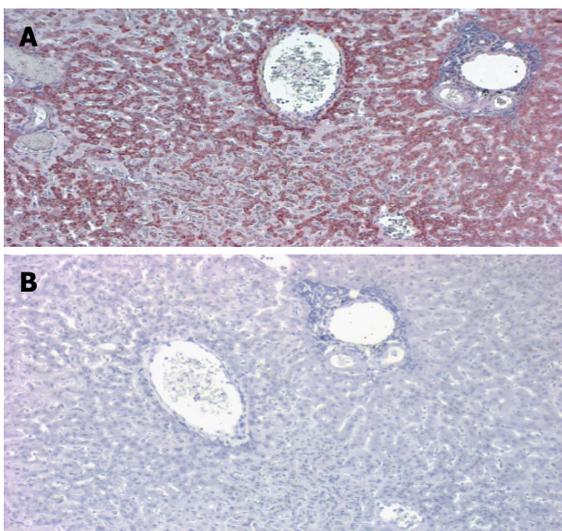


Figure 7 Immune labeling of CTT liver by anti fibulin-5 monoclonal antibody (A) and corresponding negative control (B). A: A central vein is seen in the upper center and a portal triad to the right. The dark red staining denotes the distribution of the antibody which is particularly intense surround the central vein. This is a low power magnification; B: In the negative control no staining is evident.

cells were positive but of lesser intensity in the CTT (0.125 ± 0.25 and 0.375 ± 0.479 respectively - Figure 8). Vascular staining was seen only in 2/5 CTT sections at a mean intensity of (0.3 ± 0.447). No appreciable uptake was noted in white blood cells in CTT and minimal in CM.

We therefore postulate 5 possible pathways of spontaneous hepatic metastasis evasion in the CTT, 4 of which we have previously reported^[8], summarized in Table 4.

OTHER POTENTIAL ANTI-METASTATIC AND METASTOGENIC MOIETIES IN THE CTT

Lastly using the Gelco Diagnostics antibody kit to POA we determined a low level of POA^[79] in extracts from CTT non-cancerous tissues $1.603 \pm 1.656 \mu\text{g/mL}$ (mean \pm SD). In contrast to serum in 13 humans with pancreatic cancer (9.622 ± 6.424) these levels were modest. Since normal serum levels in humans are $< 15 \text{ U/mL}$ even an order of magnitude increase in the

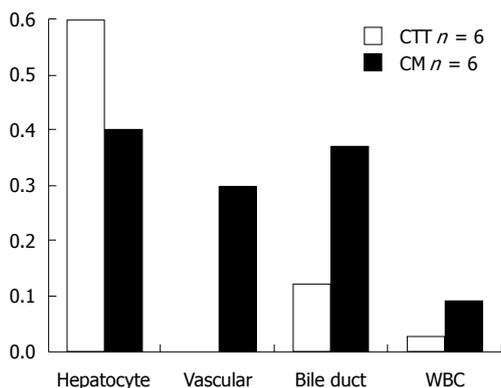


Figure 8 Summary of fibulin-5 immune labeling in cotton top tamarin, common marmoset and humans. Intensity of labeling: CTT and CM of Hepatocyte, Bile Ducts, Vascular Cells and White Blood Cells with Anti-fibulin-5. CTT: Cotton top tamarin; CM: Common marmoset.

CTT is unlikely to increase levels where they might influence the tumor invasiveness. In the study cited, a significant correlation was found between POA and CEA levels classified by stage of CRC ($P = 0.003$). Although further study is required it would appear that POA is not a major determinant in the CTT of the metastatic process to the extent that it is in humans making this another potential discouraging factor for the development of hepatic metastasis.

Ultimately, we would need to show that the cytokine environment milieu is similar in the CTT as described above for humans. Wilson *et al.*^[80] using 32 monoclonal antibodies has shown MHC Class I and II proteins on lymphocytic B cells (CD20-21, 23) and CD16-56 on NK cells; CD2 and CD3 and CD4/8 helper T cells. In addition IL-2 receptor CD25 receptor is present as well as B chain LFA-1 CD 18. There was however poor reactivity for CD11a; and ICAM-1 (CD54) was negative. 14 years later with more reagents available, Kap *et al.*^[81] confirmed much of the above findings with IL1, 2, 4, 6, 10, 12, and 17a found. In addition they found all 4 clones against CD11a to be positive in contrast to the above paper. CD11a and CD18 and CD29 are important factors for adhesion and are expressed in the CTT. In addition, CD40 of the TNF family is present thus demonstrating that the CTT has almost all the necessary repertoire of proteins as is present in humans to develop hepatic metastasis. Although ICAM-1 was not included in the repertoire, by virtue of CTT colitis to respond to an anti-integrin monoclonal^[82], we can conclude that selectins adhesion molecules are also likely expressed. This being the case, it would appear that the aforementioned inhibitory mechanisms are sufficient to abrogate this phenomenon almost completely.

We cannot rule out additional mechanisms and would encourage such research efforts. While ICAM1 may not have been confirmed in the above review (because it does not seem that it was included in the

Table 4 Summarization of our findings in this study

Parameter	CTT	CM	Human
PELPK Change	++++	++	-
CEAR expression	±	±	++++
CEA glycosylation	±	±	++++
CEACAM1 expression	-	-	++++
Fibulin-5 expression	+++	+++	++ ¹

¹Human data have limited staining data; Modified from and published with permission of Springer^[8]. CTT: Cotton top tamarin; CM: Common marmoset; CEA: Carcinoembryonic antigen.

battery of monoclonals described above), E-selectin and integrins certainly have been convincingly demonstrated to be actionable by therapeutic intervention in the CTT^[80-82].

Certainly, the above hypotheses are not established scientific facts but they are testable and applicable to a particularly vexing problem presented by hepatic metastases. The ability to intervene using novel approaches based on these ideas will constitute the final section of this review and may represent an encouraging future vision of hope for myriads of “terminal” cancer patients.

In quoting the final chapter of the late Neal K. Clapp’s book on this topic: “Our research opportunities using the cotton-top tamarins to study colonic diseases are truly only limited by our ingenuity”.

HOW MIGHT WE EXPLOIT THESE ADVANCES FOR THE BETTERMENT OF THE CANCER PATIENT?

Hepatic metastasis of CRC is a very common clinical situation in oncology with prevalence at the time of diagnosis of approximately 20%-25%. The liver has been shown to be the most common site of metastatic spread of colorectal cancer. The prognosis of colorectal hepatic metastases has improved in the last few years owing to surgical resection of liver metastases for patients with no extrahepatic disease; however, liver metastases are resectable in only 15% of the cases^[83,84]. Often, factors such as location, size and number of hepatic metastases make it difficult to resect these tumors^[83,85]. Thus early prevention of metastasis is crucial in improving the prognosis of patients diagnosed with primary colorectal cancer. As earlier documented by Smedsrød *et al.*^[86] colorectal hepatic metastasis involve four steps which are interrelated; the cancer cells must establish micrometastasis in the liver by passing through microvasculature which is guarded by Kupffer acting in concert with NK cells. To ensure nutritional supply and establish clinically evident metastatic disease, these cells engage growth factors and adhesion molecules such as VEGF, VCAMs *etc.* As can be appreciated from both clinical and experimental evidence, Kupffer cells are involved with all the rate

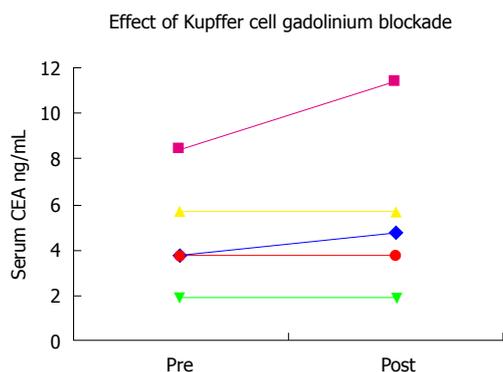


Figure 9 Carcinoembryonic antigen concentrations before and after gadolinium injection for magnetic resonance imaging. Five human patients had serial CEA levels measured before and after MRI with gadolinium contrast administration. No significant changes are seen after gadolinium for patients with normal or near-normal baseline levels. However the patient with a baseline CEA elevation showed a sizeable increase in the CEA level designated by the pink line. CEA: Carcinoembryonic antigen.

limiting step of this macrometastatic disease process which has been established in experimental systems but this role is not proven in human cancers^[86]. These phases represent target points of recent therapeutic interventional designs.

USE OF GADOLINIUM BLOCKADE TO DEplete KUPFFER CELLS

In our earlier study using CTT^[8], we showed that although they have CEA receptors similar to humans, they are able to evade CRC hepatic metastasis *via* two possible means; defective CEA receptors with much reduced receptor capacity or by sequence changes in the PELPK binding motif that is required for receptor activation (Table 2). We hypothesize exploiting methods to prevent CEA/CEAR binding in humans may prove effective in reducing the occurrence of hepatic and possibly lung metastasis. This hypothesis was tested earlier using gadolinium to block uptake of CEA by sinusoidal cells. Although the result was promising, only a transient blockage of CEA was achieved mostly due to the short wash out period for gadolinium (Figure 9). Therefore, using animal models this approach can be refined to optimize our earlier findings using gadolinium as blocking agent for Kupffer cells and the CEA/CEAR interaction.

As a novel approach to anti-metastatic therapy, gadolinium may be encapsulated in smart nano-devices that will prolong the residence time significantly. Using PEGylated-gellan-gum as the polymer of choice, gadolinium can be encapsulated in nanoparticles with average size of 250 nm^[87].

Gellan gum is an exocellular microbial polysaccharide with a natural propensity to absorb biological fluid *in vivo* thus regulating the rate of drug/agent release. It consists of repeating tetrasaccharide units of glucose, glucuronic acid and rhamnose in a molar ratio of 2:1:1^[88,89]. In addition to being used as a food additive,

gellan gum has a wide range of applications in the pharmaceutical industry. In this area, its use has been mainly concentrated in ophthalmic drug delivery and oral sustained-release preparations, in which its ability to undergo cation-induced gelation is utilized as the main fabrication platform^[87]. Gellan gum has a number of advantages as it can undergo temperature-dependent as well as cation-induced gelation, which may be of importance for *in vivo* slow release of encapsulated gadolinium. It also has proven non-toxicity in humans^[90]. The FDA has also approved gellan gum as a food additive for human consumption^[90], thus, its safety in humans is established. To improve the effectiveness of this targeting system, a three-dimensional matrix *via* covalent crosslinking with an end-reactive polyethylene glycol (PEG) through amide linkage can be used. Such modification provides a stronger gellan gum matrix that is nontoxic to normal cells and possesses the ability to deliver biological molecules such as gadolinium to target cells^[91] in a controlled manner as illustrated in Figure 10. There is some experimental animal model work that suggests that gadolinium in the form of metallofullerenol nanoparticles inhibit cancer metastases^[92] but the metastases observed were mainly pulmonary.

FIBULIN AS A TREATMENT FOR HEPATIC METASTASIS

The process of nanoprecipitation described by the schematics in Figure 10 can potentially be applied to protein encapsulation provided the formulation parameters are optimized as described earlier by our group^[91]. Fibulin-5 plays an important role in antagonizing angiogenesis in humans; furthermore its overexpression has been shown to inhibit HCC cell migration and invasion *in vitro*^[93], thus fibulin-5 could also be encapsulated in gellan gum nanoparticles to increase their *in vivo* residence time. Nanoparticles formulation should be optimized as described earlier^[91] prior to encapsulation of fibulin-5. We would expect fibulin-5 to be the most effective of the fibulins in reducing angiogenesis but fibulin-3 may also be considered (Table 3). While fibulin-5 has been shown to suppress metastasis to the liver and lung^[94], it has yet to be used directly in a nanotechnology formulation as we propose. A report that human oncogenic fibulin-5 may promote nasopharyngeal cancer metastasis^[95] would suggest that we proceed cautiously with this approach with both agents.

TARGETING CEA/CEAR INTERACTIONS AS A THERAPEUTIC APPROACH TO LIVER METASTASIS

CEA has been shown to be a mediator of metastasis for colorectal cancer by causing changes in the tumor microenvironment that increase the potential for tumor

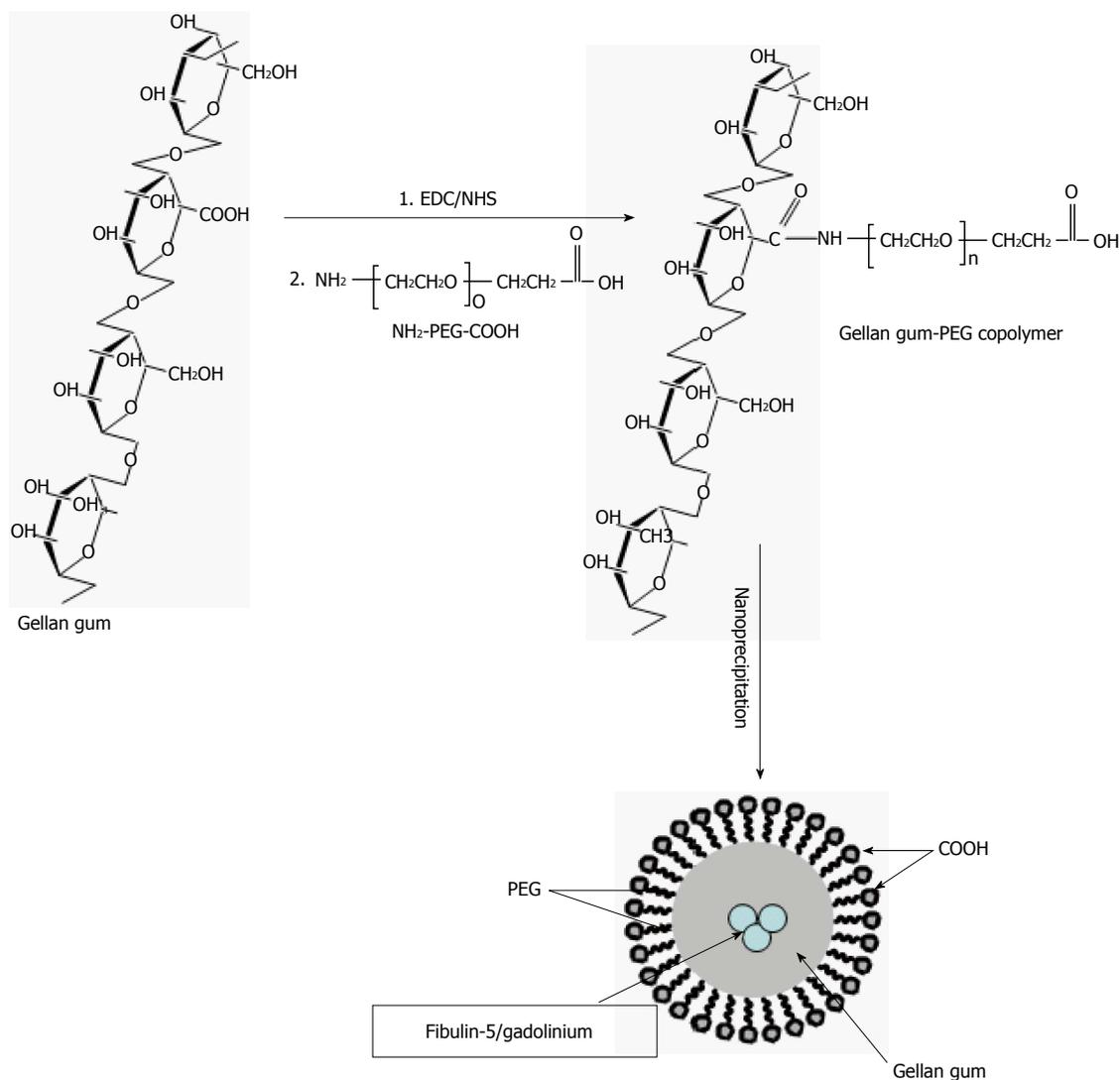


Figure 10 Schematic representation of the technique used for the synthesis of gadolinium encapsulated Gellan-gum-block-Polyethylene glycol copolymeric nanoparticles. The above figure is a representation of the technical steps to generate nanoparticles of gadolinium encapsulated by Gellan-gum-block-Polyethylene glycol copolymeric micelle nanoparticles. This should ensure delivery and advantageous dwell time in order to block Kupffer functions of uptake of CEA and reduce the pro-metastatic role of these cells. CEA: Carcinoembryonic antigen.

cell implantation and growth^[14]. CEA interaction with its receptor therefore suggests a targeted approach to therapy. A number of methods could be used to achieve disruption of CEA/CEAR including the use of antibodies specific for CEA or its receptor. In general this type of approach has worked well against various cancers and growth receptors *e.g.*, EGFR. Other approaches using small molecule inhibitors of ligand receptor interactions are also being used, In particular RNA aptamers hold great promise as therapeutic agents^[96]. These aptamers are now being used in studies involving CEA^[97]. Lee *et al*^[98] have described the use of a RNA aptamer directed against the PELPK motif of CEA to prevent liver metastasis from the mucinous colon cancer cell line LS174T. They suggest this occurred by disruption of CEA/CEAR interaction. LS174T produces large amounts of CEA and has a high metastatic potential to the liver^[99]. Recently, aptamers have also been used against the homotypic adhesion

sites of CEA to block tumor cell/cell interactions^[100] further emphasizing the feasibility of this approach^[101]. Aptamers are also being used for radiolocalization of CEA producing tumors and may even supplant the use of antibodies for this purpose^[102]. The use of CEA antagonists to reduce the metastatic potential has now become feasible especially with the advent of small molecule inhibitors. More pre-clinical trials are needed before these approaches can be used in cancer patients at high risk for metastases.

CONCLUSION

We thus have the means to potentially interfere with the uptake of CEA from the hepatic sinusoid Kupffer cell receptor and hopefully disrupt the inflammatory cascade and well as intervening in the fibulin pathways in delivery of fibulin-5 to shore up hepatic defenses against tumor spread to the liver. In time, additional

interventions will be designed to make this a reality based on the lessons learned from the cotton top tamarin- the artful liver metastasis dodger.

ACKNOWLEDGMENTS

Were we to mention every contributor to this substantial body of work this section would undoubtedly be the most extensive. We hope that the names listed in the references will be a curtain call for us to applaud their hard work and sincere efforts. At the same time there were a number of individuals without whom this work would not have been done. We recognize our mentors Norman Zamcheck and Neil Clapp of blessed memory. The guidance and generosity of Phil Gold, Abraham Fuks, Jack Shively, Young S Kim, Jim Fox and Sen-Itoh Hakamori, Clifford Stanners, Zvi Bentwich, Steven Itzkowitz and James Allard was peerless. Their mention here should not be construed to imply their approval of this work but we hope that they do. The tenacity and attention to detail by Karel Kithier and James Hatfield were legend. To the many students, resident and Fellows who appear in many meeting abstracts we are most grateful. Our work could not have progressed without various grants from National Institute of Health, Veterans Health Administration, Kaiser Permanente and Hybritech™ Inc. Thanks also to S. Jatczak and the library staff at the Saginaw VAMC. All previously published material that is used in this manuscript by the principal author emanates from work done as a US Government employee and is therefore in the public domain. However, this work does not necessarily reflect the views of the US Government. As a courtesy, permissions were sought from the relevant publishing houses and graciously granted. This paper is dedicated to the memory of Mendel Tobi whose love for the animal world was boundless.

REFERENCES

- 1 **Weiss RA**, Vogt PK. 100 years of Rous sarcoma virus. *J Exp Med* 2011; **208**: 2351-2355 [PMID: 22110182 DOI: 10.1084/jem.20112160]
- 2 **Clapp NK**, Nardi RV, Tobi M. Future Directions for Colon Disease Research Using Cotton-Top Tamarins. In: Clapp NK, editor. A Primate Model for the Study of Colitis and Colon Carcinoma The Cotton Top Tamarin *Saguinus oedipus*. Boca Raton: CRC Press, 1993: 319-324
- 3 **Wood JD**, Peck OC, Tefend KS, Stonerook MJ, Caniano DA, Mutabagani KH, Lhoták S, Sharma HM. Evidence that colitis is initiated by environmental stress and sustained by fecal factors in the cotton-top tamarin (*Saguinus oedipus*). *Dig Dis Sci* 2000; **45**: 385-393 [PMID: 10711456 DOI: 10.1023/A:1005485215128]
- 4 **Mansfield KG**, Lin KC, Xia D, Newman JV, Schauer DB, MacKey J, Lackner AA, Carville A. Enteropathogenic *Escherichia coli* and ulcerative colitis in cotton-top tamarins (*Saguinus oedipus*). *J Infect Dis* 2001; **184**: 803-807 [PMID: 11517446 DOI: 10.1086/322990]
- 5 **Hofmann P**, Kahnt K, Mätz-Rensing K, Brack M, Kaup FJ. Three spontaneous lymphomas in a colony of cotton-top tamarins (*Saguinus oedipus*). *J Med Primatol* 2001; **30**: 322-327 [PMID: 11990532 DOI: 10.1034/j.1600-0684.2001.300606.x]
- 6 **Lushbaugh CC**, Humason GL, Swartzendruber DC, Richter CB, Gengozian N. Spontaneous colonic adenocarcinoma in marmosets. *Primates Med* 1978; **10**: 119-134 [PMID: 417316 DOI: 10.1007/BF01296974]
- 7 **Watkins DL**, Letvin NL. Immunobiology of the Cotton Top Tamarin. In: Clapp NK, editor. A primate Model for the Study of Colitis and Colon Carcinoma The Cotton Top Tamarin *Saguinus oedipus*. Boca Raton: CRC Press, 1993: 299-307
- 8 **Tobi M**, Kim M, Zimmer R, Hatfield J, Kam M, Khoury N, Carville A, Lawson MJ, Schiemann WP, Thomas P. Colorectal cancer in the cotton top tamarin (*Saguinus oedipus*): how do they evade liver metastasis? *Dig Dis Sci* 2011; **56**: 397-405 [PMID: 20645001 DOI: 10.1007/s10620-010-1314-2]
- 9 **Lushbaugh C**, Humason G, Clapp N. Histology of colitis: *Saguinus oedipus* and other marmosets. *Dig Dis Sci* 1985; **30**: 45S-51S [PMID: 3933936]
- 10 **Siegel RL**, Miller KD, Jemal A. Cancer statistics, 2015. *CA Cancer J Clin* 2015; **65**: 5-29 [PMID: 25559415 DOI: 10.3322/caac.21254]
- 11 **Tarcic O**, Pateras IS, Cooks T, Shema E, Kanterman J, Ashkenazi H, Boocholez H, Hubert A, Rotkopf R, Baniyash M, Pikarsky E, Gorgoulis VG, Oren M. RNF20 Links Histone H2B Ubiquitylation with Inflammation and Inflammation-Associated Cancer. *Cell Rep* 2016; **14**: 1462-1476 [PMID: 26854224 DOI: 10.1016/j.celrep.2016.01.020]
- 12 **Bromberg J**, Wang TC. Inflammation and cancer: IL-6 and STAT3 complete the link. *Cancer Cell* 2009; **15**: 79-80 [PMID: 19185839 DOI: 10.1016/j.ccr.2009.01.009]
- 13 **Grivnennikov SI**, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell* 2010; **140**: 883-899 [PMID: 20303878 DOI: 10.1016/j.cell.2010.01.025]
- 14 **Thomas P**, Forse RA, Bajenova O. Carcinoembryonic antigen (CEA) and its receptor hnRNP M are mediators of metastasis and the inflammatory response in the liver. *Clin Exp Metastasis* 2011; **28**: 923-932 [PMID: 21901530 DOI: 10.1007/s10585-011-9419-3]
- 15 **Thomas P**, Toth CA, Saini KS, Jessup JM, Steele G. The structure, metabolism and function of the carcinoembryonic antigen gene family. *Biochim Biophys Acta* 1990; **1032**: 177-189 [PMID: 2261493]
- 16 **Hammarström S**. The carcinoembryonic antigen (CEA) family: structures, suggested functions and expression in normal and malignant tissues. *Semin Cancer Biol* 1999; **9**: 67-81 [PMID: 10202129 DOI: 10.1006/scbi.1998.0119]
- 17 **Gold P**, Freedman SO. Specific carcinoembryonic antigens of the human digestive system. *J Exp Med* 1965; **122**: 467-481 [PMID: 4953873 DOI: 10.1084/jem.122.3.467]
- 18 **Beauchemin N**, Arabzadeh A. Carcinoembryonic antigen-related cell adhesion molecules (CEACAMs) in cancer progression and metastasis. *Cancer Metastasis Rev* 2013; **32**: 643-671 [PMID: 23903773 DOI: 10.1007/s10555-013-9444-6]
- 19 **Paxton RJ**, Mooser G, Pande H, Lee TD, Shively JE. Sequence analysis of carcinoembryonic antigen: identification of glycosylation sites and homology with the immunoglobulin supergene family. *Proc Natl Acad Sci USA* 1987; **84**: 920-924 [PMID: 3469650]
- 20 **Beauchemin N**, Draber P, Dveksler G, Gold P, Gray-Owen S, Grunert F, Hammarström S, Holmes KV, Karlsson A, Kuroki M, Lin SH, Lucka L, Najjar SM, Neumaier M, Obrink B, Shively JE, Skubitz KM, Stanners CP, Thomas P, Thompson JA, Virji M, von Kleist S, Wagener C, Watt S, Zimmermann W. Redefined nomenclature for members of the carcinoembryonic antigen family. *Exp Cell Res* 1999; **252**: 243-249 [PMID: 11501563 DOI: 10.1006/excr.1999.4610]
- 21 **Aarons CB**, Bajenova O, Andrews C, Heydrick S, Bushell KN, Reed KL, Thomas P, Becker JM, Stucchi AF. Carcinoembryonic antigen-stimulated THP-1 macrophages activate endothelial cells and increase cell-cell adhesion of colorectal cancer cells. *Clin Exp Metastasis* 2007; **24**: 201-209 [PMID: 17487559 DOI: 10.1007/s10585-007-9069-7]
- 22 **Benchimol S**, Fuks A, Jothy S, Beauchemin N, Shirota K, Stanners CP. Carcinoembryonic antigen, a human tumor marker, functions as an intercellular adhesion molecule. *Cell* 1989; **57**: 327-334 [PMID: 2702691 DOI: 10.1016/0092-8674(89)90970-7]

- 23 **Ordoñez C**, Sreaton RA, Ilantzis C, Stanners CP. Human carcinoembryonic antigen functions as a general inhibitor of anoikis. *Cancer Res* 2000; **60**: 3419-3424 [PMID: 10910050]
- 24 **Bramswig KH**, Poettler M, Unseld M, Wrba F, Uhrin P, Zimmermann W, Zielinski CC, Prager GW. Soluble carcinoembryonic antigen activates endothelial cells and tumor angiogenesis. *Cancer Res* 2013; **73**: 6584-6596 [PMID: 24121495 DOI: 10.1158/0008-5472.CAN-13-0123]
- 25 **Zamcheck N**. The expanding field of colorectal cancer markers: CEA, the prototype. *Cancer Bull* 1981; **33**: 141-151
- 26 **Thomas P**, Zamcheck N. Role of the liver in clearance and excretion of circulating carcinoembryonic antigen (CEA). *Dig Dis Sci* 1983; **28**: 216-224 [PMID: 6337795 DOI: 10.1007/BF01295116]
- 27 **Zamcheck N**, Liu P, Thomas P, Steele G. Search for useful biomarkers of early malignant tumors. In: Steele G, Burt RW, Winawer SJ, Karr JP, editors. *Basic and Clinical Perspectives of Colorectal Polyps and Cancer (Progress in Clinical and Biological Research)*. New York: Clinical and Biological Research, 1988: 251-275
- 28 **Goldstein MJ**, Mitchell EP. Carcinoembryonic antigen in the staging and follow-up of patients with colorectal cancer. *Cancer Invest* 2005; **23**: 338-351 [PMID: 16100946 DOI: 10.1081/CNV-58878]
- 29 **Goldenberg DM**, Larson SM. Radioimmunodetection in cancer identification. *J Nucl Med* 1992; **33**: 803-814 [PMID: 1569493]
- 30 **Behr TM**, Sharkey RM, Juweid ME, Dunn RM, Vagg RC, Ying Z, Zhang CH, Swayne LC, Vardi Y, Siegel JA, Goldenberg DM. Phase I/II clinical radioimmunotherapy with an iodine-131-labeled anti-carcinoembryonic antigen murine monoclonal antibody IgG. *J Nucl Med* 1997; **38**: 858-870 [PMID: 9189130]
- 31 **Morse MA**, Nair SK, Mosca PJ, Hobeika AC, Clay TM, Deng Y, Boczkowski D, Proia A, Neidzwiecki D, Clavien PA, Hurwitz HI, Schlom J, Gilboa E, Lyerly HK. Immunotherapy with autologous, human dendritic cells transfected with carcinoembryonic antigen mRNA. *Cancer Invest* 2003; **21**: 341-349 [PMID: 12901279 DOI: 10.1081/CNV-120018224]
- 32 **O'Brien MJ**, Bronstein B, Zamcheck N, Saravis C, Burke B, Gottlieb LS. Cholestasis and hepatic metastases: a factor contributing to extreme elevations of carcinoembryonic antigen. *J Natl Cancer Inst* 1980; **64**: 1291-1294 [PMID: 6246298]
- 33 **Bajenova OV**, Zimmer R, Stolper E, Salisbury-Rowswell J, Nanji A, Thomas P. Heterogeneous RNA-binding protein M4 is a receptor for carcinoembryonic antigen in Kupffer cells. *J Biol Chem* 2001; **276**: 31067-31073 [PMID: 11406629 DOI: 10.1074/jbc.M104093200]
- 34 **Han SP**, Tang YH, Smith R. Functional diversity of the hnRNPs: past, present and perspectives. *Biochem J* 2010; **430**: 379-392 [PMID: 20795951 DOI: 10.1042/BJ20100396]
- 35 **Han N**, Li W, Zhang M. The function of the RNA-binding protein hnRNP in cancer metastasis. *J Cancer Res Ther* 2013; **9** Suppl: S129-S134 [PMID: 24516048 DOI: 10.4103/0973-1482.122506]
- 36 **Marko M**, Leichter M, Patrinoiu-Georgoula M, Guialis A. hnRNP M interacts with PSF and p54(nrb) and co-localizes within defined nuclear structures. *Exp Cell Res* 2010; **316**: 390-400 [PMID: 19874820 DOI: 10.1016/j.yexcr.2009.10.021]
- 37 **Bajenova O**, Stolper E, Gapon S, Sundina N, Zimmer R, Thomas P. Surface expression of heterogeneous nuclear RNA binding protein M4 on Kupffer cell relates to its function as a carcinoembryonic antigen receptor. *Exp Cell Res* 2003; **291**: 228-241 [PMID: 14597422 DOI: 10.1016/S0014-4827(03)00373-2]
- 38 **Datar KV**, Dreyfuss G, Swanson MS. The human hnRNP M proteins: identification of a methionine/arginine-rich repeat motif in ribonucleoproteins. *Nucleic Acids Res* 1993; **21**: 439-446 [PMID: 8441656 DOI: 10.1093/nar/21.3.439]
- 39 **Soeth E**, Wirth T, List HJ, Kumbhani S, Petersen A, Neumaier M, Czubayko F, Juhl H. Controlled ribozyme targeting demonstrates an antiapoptotic effect of carcinoembryonic antigen in HT29 colon cancer cells. *Clin Cancer Res* 2001; **7**: 2022-2030 [PMID: 11448920]
- 40 **Samara RN**, Laguigne LM, Jessup JM. Carcinoembryonic antigen inhibits anoikis in colorectal carcinoma cells by interfering with TRAIL-R2 (DR5) signaling. *Cancer Res* 2007; **67**: 4774-4782 [PMID: 17510406 DOI: 10.1158/0008-5472.CAN-06-4315]
- 41 **Gangopadhyay A**, Bajenova O, Kelly TM, Thomas P. Carcinoembryonic antigen induces cytokine expression in Kupffer cells: implications for hepatic metastasis from colorectal cancer. *Cancer Res* 1996; **56**: 4805-4810 [PMID: 8841002]
- 42 **Gangopadhyay A**, Lazure DA, Thomas P. Adhesion of colorectal carcinoma cells to the endothelium is mediated by cytokines from CEA stimulated Kupffer cells. *Clin Exp Metastasis* 1998; **16**: 703-712 [PMID: 10211983 DOI: 10.1023/A:1006576627429]
- 43 **Jessup JM**, Laguigne L, Lin S, Samara R, Aufman K, Battle P, Frantz M, Edmiston KH, Thomas P. Carcinoembryonic antigen induction of IL-10 and IL-6 inhibits hepatic ischemic/reperfusion injury to colorectal carcinoma cells. *Int J Cancer* 2004; **111**: 332-337 [PMID: 15221959 DOI: 10.1002/ijc.20264]
- 44 **Zhou H**, Fuks A, Alcaraz G, Bolling TJ, Stanners CP. Homophilic adhesion between Ig superfamily carcinoembryonic antigen molecules involves double reciprocal bonds. *J Cell Biol* 1993; **122**: 951-960 [PMID: 8349740 DOI: 10.1083/jcb.122.4.951]
- 45 **Bramswig KH**, Prager GW, Kallinowska W, Zielinski C, Prager G. Carcinoembryonic antigen (CEA) affects tumor angiogenesis in colon carcinomas. Denver, CO. Philadelphia (PA): AACR, 2009: Abstract 3183
- 46 **Bramswig KH**, Prager GW, Martel A, Heinze G, Binder BR, Brodowicz T, Kornek G, Scheithauer W, Zielinski C. Predictive role of carcinoembryonic antigen (CEA) for therapeutic efficacy of angiogenesis-targeting therapy in colorectal cancer: Novel clinical observation based on recently discovered CEA biology. *J Clin Oncol* 2010; **28**: 3574
- 47 **Prager GW**, Braemswig KH, Martel A, Unseld M, Heinze G, Brodowicz T, Scheithauer W, Kornek G, Zielinski CC. Baseline carcinoembryonic antigen (CEA) serum levels predict bevacizumab-based treatment response in metastatic colorectal cancer. *Cancer Sci* 2014; **105**: 996-1001 [PMID: 24850362 DOI: 10.1111/cas.12451]
- 48 **Low-Marchelli JM**, Ardi VC, Vizcarra EA, van Rooijen N, Quigley JP, Yang J. Twist1 induces CCL2 and recruits macrophages to promote angiogenesis. *Cancer Res* 2013; **73**: 662-671 [PMID: 23329645 DOI: 10.1158/0008-5472.CAN-12-0653]
- 49 **Cohen T**, Nahari D, Cerem LW, Neufeld G, Levi BZ. Interleukin 6 induces the expression of vascular endothelial growth factor. *J Biol Chem* 1996; **271**: 736-741 [PMID: 8557680 DOI: 10.1074/jbc.271.2.736]
- 50 **Palermo NY**, Thomas P, Murphy RF, Lovas S. Hexapeptide fragment of carcinoembryonic antigen which acts as an agonist of heterogeneous ribonucleoprotein M. *J Pept Sci* 2012; **18**: 252-260 [PMID: 22392880 DOI: 10.1002/psc.2393]
- 51 **Albig AR**, Schiemann WP. Fibulin-5 antagonizes vascular endothelial growth factor (VEGF) signaling and angiogenic sprouting by endothelial cells. *DNA Cell Biol* 2004; **23**: 367-379 [PMID: 15231070 DOI: 10.1089/104454904323145254]
- 52 **Albig AR**, Neil JR, Schiemann WP. Fibulins 3 and 5 antagonize tumor angiogenesis in vivo. *Cancer Res* 2006; **66**: 2621-2629 [PMID: 16510581 DOI: 10.1158/0008-5472.CAN-04-4096]
- 53 **Sullivan KM**, Bissonnette R, Yanagisawa H, Hussain SN, Davis EC. Fibulin-5 functions as an endogenous angiogenesis inhibitor. *Lab Invest* 2007; **87**: 818-827 [PMID: 17607303 DOI: 10.1038/labinvest.3700594]
- 54 **Barnard D**, Knapka JJ. Callitrichid Nutrition. In: Clapp NK, editor. *A Primate Model for the Study of Colitis and Colon Carcinoma The Cotton Top Tamarin Saguinus oedipus*. Boca Raton: CRC Press, 1993: 55-79
- 55 **Mast RB**, Rodriguez JV, Mittermeier RA. The Colombian Cotton Top Tamarin in the Wild. In: Clapp NK, editor. *A primate Model for the Study of Colitis and Colon Carcinoma The Cotton Top Tamarin Saguinus oedipus*. Boca Raton: CRC Press, 1993: 4-43
- 56 **Clapp NK**, Henke MA. Spontaneous colonic carcinoma observations in the Oak Ridge Associated Universities' 26-year-old Cotton-Top Tamarin (*Saguinus Oedipus*) Colony. In: Clapp NK, editor. *A primate Model for the Study of Colitis and Colon Carcinoma The Cotton Top Tamarin Saguinus oedipus*. Boca Raton: CRC Press, 1993: 171-185

- 57 **Murff HJ**. Cohort analysis finds that the proportion of people who meet high risk criteria for colorectal, breast or prostate cancer screening based on family history increases between age 30 and 50. *Evid Based Med* 2012; **17**: 50-51 [PMID: 21965625 DOI: 10.1136/ebm.2011]
- 58 **Mao X**, McGuire S, Hamoudi RA. Molecular and cytogenetic analysis of lymphoblastoid and colon cancer cell lines from cotton-top tamarin (*Saguinus oedipus*). *Cancer Genet Cytogenet* 2000; **120**: 6-10 [PMID: 10913670 DOI: 10.1016/S0165-4608(99)00237-X]
- 59 **Fuhr JE**, Van Meter S, Andrews RB, Clapp NK. Tamarin colon cancer and flow cytometry In: Clapp NK, editor. A primate Model for the Study of Colitis and Colon Carcinoma The Cotton Top Tamarin *Saguinus oedipus*. Boca Raton: CRC Press, 1993: 231-238
- 60 **Tobi M**, Chintalapani S, Kithier K, Clapp N. Gastrointestinal tract antigenic profile of cotton-top tamarin, *Saguinus oedipus*, is similar to that of humans with inflammatory bowel disease. *Dig Dis Sci* 2000; **45**: 2290-2297 [PMID: 11258547 DOI: 10.1023/A:1005622521294]
- 61 **Itzkowitz SH**, Harpaz N. Diagnosis and management of dysplasia in patients with inflammatory bowel diseases. *Gastroenterology* 2004; **126**: 1634-1648 [PMID: 15168373 DOI: 10.1053/j.gastro.2004.03.025]
- 62 **Mattar MC**, Lough D, Pishvaian MJ, Charabaty A. Current management of inflammatory bowel disease and colorectal cancer. *Gastrointest Cancer Res* 2011; **4**: 53-61 [PMID: 21673876]
- 63 **Murphy PM**. Chemokines and the molecular basis of cancer metastasis. *N Engl J Med* 2001; **345**: 833-835 [PMID: 11556308 DOI: 10.1056/NEJM200109133451113]
- 64 **Westernströer B**, Langenstroth D, Kliesch S, Troppmann B, Redmann K, Macdonald J, Mitchell R, Wistuba J, Schlatt S, Neuhaus N. Developmental expression patterns of chemokines CXCL11, CXCL12 and their receptor CXCR7 in testes of common marmoset and human. *Cell Tissue Res* 2015; **361**: 885-898 [PMID: 25810367 DOI: 10.1007/s00441-015-2164-1]
- 65 **Olsson AY**, Valtonen-André C, Lilja H, Lundwall A. The evolution of the glandular kallikrein locus: identification of orthologs and pseudogenes in the cotton-top tamarin. *Gene* 2004; **343**: 347-355 [PMID: 15588589 DOI: 10.1016/j.gene.2004.09.020]
- 66 **Boland CR**, Clapp NK. Glycoconjugates in the colons of New World monkeys with spontaneous colitis. Association between inflammation and neoplasia. *Gastroenterology* 1987; **92**: 625-634 [PMID: 3102306]
- 67 **Tobi M**, Memon M, Kithier K, Clapp N. A putative CEA moiety is shared by the cotton-top tamarin (*Saguinus oedipus*) and humans. *Cancer Lett* 1994; **77**: 7-13 [PMID: 8162564 DOI: 10.1016/0304-3835(94)90341-7]
- 68 **Haagensen DE**, Metzgar RS, Swenson B, Dilley WG, Cox CE, Davis S, Murdoch J, Zamcheck N, Wells SA. Carcinoembryonic antigen in nonhuman primates. *J Natl Cancer Inst* 1982; **69**: 1073-1076 [PMID: 6813550]
- 69 **Beauchemin N**, Turbide C, Afar D, Bell J, Raymond M, Stanners CP, Fuks A. A mouse analogue of the human carcinoembryonic antigen. *Cancer Res* 1989; **49**: 2017-2021 [PMID: 2702644]
- 70 **Tobi M**, Kaila V, Chintalapani S, Kithier K, Henke MA, Clapp NK. An antigenic profile in cotton-top tamarins (*Saguinus oedipus*): a model for human inflammatory bowel disease and colorectal cancer. In: Clapp NK, editor. A primate Model for the Study of Colitis and Colon Carcinoma The Cotton Top Tamarin *Saguinus oedipus*. Boca Raton: CRC Press, 1993: 113-125
- 71 **Tobi M**, Darmon CE, Rozen P, Harpaz N, Fink A, Maliakkal B, Halline A, Mobarhan S, Bentwich Z. Urinary organ specific neoantigen. A potentially diagnostic test for colorectal cancer. *Dig Dis Sci* 1995; **40**: 1531-1537 [PMID: 7628279 DOI: 10.1007/BF02285204]
- 72 **Tobi M**, Clapp N. Is the Cotton-Top Tamarin an appropriate model for human inflammatory bowel disease? *Agents and Actions* 1994; **41**: C246-C248 [DOI: 10.1007/BF01987655]
- 73 **Tobi M**, Chintalapani S, Kithier K, Clapp N. Carcinoembryonic antigen family of adhesion molecules in the cotton top tamarin (*Saguinus oedipus*). *Cancer Lett* 2000; **157**: 45-50 [PMID: 10893441 DOI: 10.1016/S0304-3835(00)00482-1]
- 74 **Tobi M**, Maliakkal B, Zitron I, Alousi M, Goo R, Nochomovitz L, Luk G. Adenoma-derived antibody, Adnab-9 recognizes a membrane-bound glycoprotein in colonic tissue and effluent material from patients with colorectal neoplasia. *Cancer Lett* 1992; **67**: 61-69 [PMID: 1423246 DOI: 10.1016/0304-3835(92)90009-k]
- 75 **Tobi M**, Yordanova V, Hatfield J, Hallman J, Khoury N, Bajenova O, Clapp NK, Lawson MJ, Sehgal PV, Carville A, Thomas P. Inhibition of Kupffer cell CEA-uptake averts liver metastases in a spontaneous colorectal cancer animal model: Chemoblockade in humans may provide a target for intervention. *Cancer Res* 2006; **66**: 820
- 76 **Hu Y**, Gao H, Vo C, Ke C, Pan F, Yu L, Siegel E, Hess KR, Linskey ME, Zhou YH. Anti-EGFR function of EFEMP1 in glioma cells and patient prognosis. *Oncoscience* 2014; **1**: 205-215 [PMID: 25594013 DOI: 10.18632/oncoscience.24]
- 77 **Naghibalhossaini F**, Yoder AD, Tobi M, Stanners CP. Evolution of a tumorigenic property conferred by glycoposphatidyl-inositol membrane anchors of carcinoembryonic antigen gene family members during the primate radiation. *Mol Biol Cell* 2007; **18**: 1366-1374 [PMID: 17287394 DOI: 10.1091/mbc.E06-10-0884]
- 78 **Zimmer R**, Thomas P. Mutations in the carcinoembryonic antigen gene in colorectal cancer patients: implications on liver metastasis. *Cancer Res* 2001; **61**: 2822-2826 [PMID: 11306451]
- 79 **Kithier K**, Samal B, Cejka J, Whitcomb MP, Mood DW. Pancreatic oncofetal antigen and carcinoembryonic antigen in breast and colon carcinoma. *Tumour Biol* 1988; **9**: 307-314 [PMID: 3206109 DOI: 10.1159/000217577]
- 80 **Wilson AD**, Shooshtari M, Finerty S, Watkins P, Morgan AJ. Selection of monoclonal antibodies for the identification of lymphocyte surface antigens in the New World primate *Saguinus oedipus oedipus* (cotton top tamarin). *J Immunol Methods* 1995; **178**: 195-200 [PMID: 7836781 DOI: 10.1016/0022-1757(94)00256-V]
- 81 **Kap VS**, van Meurs M, van Driel N, Koopman G, Melief MJ, Brok HP, Laman JD, Hart BA. A monoclonal antibody selection for immunohistochemical examination of lymphoid tissues from non-human primates. *J Histochem Cytochem* 2009; **57**: 1159-1167 [PMID: 19729671 DOI: 10.1369/jhc.2009.954123]
- 82 **Podolsky DK**, Lobb R, King N, Benjamin CD, Pepinsky B, Sehgal P, deBeaumont M. Attenuation of colitis in the cotton-top tamarin by anti-alpha 4 integrin monoclonal antibody. *J Clin Invest* 1993; **92**: 372-380 [PMID: 7686922 DOI: 10.1172/JCI116575]
- 83 **Adam R**. Chemotherapy and surgery: new perspectives on the treatment of unresectable liver metastases. *Ann Oncol* 2003; **14** Suppl 2: ii13-ii16 [PMID: 12810452 DOI: 10.1093/annonc/mdg731]
- 84 **Van Cutsem E**, Nordlinger B, Adam R, Köhne CH, Pozzo C, Poston G, Ychou M, Rougier P; European Colorectal Metastases Treatment Group. Towards a pan-European consensus on the treatment of patients with colorectal liver metastases. *Eur J Cancer* 2006; **42**: 2212-2221 [PMID: 16904315 DOI: 10.1016/j.ejca.2006.04.012]
- 85 **Nordlinger B**, Van Cutsem E, Rougier P, Köhne CH, Ychou M, Sobrero A, Adam R, Arvidsson D, Carrato A, Georgoulis V, Giuliante F, Glimelius B, Golling M, Gruenberger T, Tabernero J, Wasan H, Poston G; European Colorectal Metastases Treatment Group. Does chemotherapy prior to liver resection increase the potential for cure in patients with metastatic colorectal cancer? A report from the European Colorectal Metastases Treatment Group. *Eur J Cancer* 2007; **43**: 2037-2045 [PMID: 17766104 DOI: 10.1016/j.ejca.2007.07.017]
- 86 **Smedsrød B**, Le Couteur D, Ikejima K, Jaeschke H, Kawada N, Naito M, Knolle P, Nagy L, Senoo H, Vidal-Vanaclocha F, Yamaguchi N. Hepatic sinusoidal cells in health and disease: update from the 14th International Symposium. *Liver Int* 2009; **29**: 490-501 [PMID: 19210626 DOI: 10.1111/j.1478-3231.2009.01979.x]
- 87 **Roziar A**, Mazuel C, Grove J, Plazonnet B. Functionality testing of gellan gum, a polymeric excipient material for ophthalmic dosage forms. *Int J Pharm* 1997; **153**: 191-198 [DOI: 10.1016/S0378-3905173(97)00109]
- 88 **Kanari B**, Banik RR, Upadhyay SN. Effect of environmental factors and carbohydrate on gellan gum production. *Appl Biochem Biotechnol* 2002; **102-103**: 129-140 [PMID: 12396117 DOI: 10.1016/S0304-3835(00)00482-1]

- 10.1385/ABAB::102-103:1-6:129]
- 89 **Banik RM**, Santhiagu A. Improvement in production and quality of gellan gum by *Sphingomonas paucimobilis* under high dissolved oxygen tension levels. *Biotechnol Lett* 2006; **28**: 1347-1350 [PMID: 16820976 DOI: 10.1007/s10529-006-9098-3]
- 90 **Antony PJ**, Sanghavi NM. A New Binder for Pharmaceutic al Dosage Forms. *Drug Dev Ind Pharm* 1997; **23**: 417-418 [DOI: 10.3109/03639049709146147]
- 91 **Devineni D**, Ezekwudo D, Palaniappan R. Formulation of maltodextrin entrapped in polycaprolactone microparticles for protein and vaccine delivery: effect of size determining formulation process variables of microparticles on the hydrodynamic diameter of BSA. *J Microencapsul* 2007; **24**: 358-370 [PMID: 17497389 DOI: 10.1080/02652040701279104]
- 92 **Meng H**, Xing G, Blanco E, Song Y, Zhao L, Sun B, Li X, Wang PC, Korotcov A, Li W, Liang XJ, Chen C, Yuan H, Zhao F, Chen Z, Sun T, Chai Z, Ferrari M, Zhao Y. Gadolinium metallofullerenol nanoparticles inhibit cancer metastasis through matrix metalloproteinase inhibition: imprisoning instead of poisoning cancer cells. *Nanomedicine* 2012; **8**: 136-146 [PMID: 21930111 DOI: 10.1016/j.nano.2011.08.019]
- 93 **Tu K**, Dou C, Zheng X, Li C, Yang W, Yao Y, Liu Q. Fibulin-5 inhibits hepatocellular carcinoma cell migration and invasion by down-regulating matrix metalloproteinase-7 expression. *BMC Cancer* 2014; **14**: 938 [PMID: 25494879 DOI: 10.1186/1471-2407-14-938]
- 94 **Møller HD**, Ralfkjær U, Cremers N, Frankel M, Pedersen RT, Klingelhöfer J, Yanagisawa H, Grigorian M, Guldberg P, Sleeman J, Lukanidin E, Ambartsumian N. Role of fibulin-5 in metastatic organ colonization. *Mol Cancer Res* 2011; **9**: 553-563 [PMID: 21454378 DOI: 10.1158/1541-7786.MCR-11-0093]
- 95 **Hwang CF**, Shiu LY, Su LJ, Yu-Fang Yin WS, Huang SC, Chiu TJ, Huang CC, Zhen YY, Tsai HT, Fang FM, Huang TL, Chen CH. Oncogenic fibulin-5 promotes nasopharyngeal carcinoma cell metastasis through the FLJ10540/AKT pathway and correlates with poor prognosis. *PLoS One* 2013; **8**: e84218 [PMID: 24386352 DOI: 10.1371/journal.pone.0084218]
- 96 **Kaur G**, Roy I. Therapeutic applications of aptamers. *Expert Opin Investig Drugs* 2008; **17**: 43-60 [PMID: 18095918 DOI: 10.1517/13543784.17.1.43]
- 97 **Wang L**, Liu B, Yin H, Wei J, Qian X, Yu L. Selection of DNA aptamer that specific binding human carcinoembryonic antigen in vitro. *Nanjing Yike Daxue Zazhi* 2007; **21**: 277-278 [DOI: 10.1016/S1007-4376(07)60061-6]
- 98 **Lee YJ**, Han SR, Kim NY, Lee SH, Jeong JS, Lee SW. An RNA aptamer that binds carcinoembryonic antigen inhibits hepatic metastasis of colon cancer cells in mice. *Gastroenterology* 2012; **143**: 155-65.e8 [PMID: 22465431 DOI: 10.1053/j.gastro.2012.03.039]
- 99 **Wagner HE**, Toth CA, Steele GD, Thomas P. Metastatic potential of human colon cancer cell lines: relationship to cellular differentiation and carcinoembryonic antigen production. *Clin Exp Metastasis* 1992; **10**: 25-31 [PMID: 1733644 DOI: 10.1007/BF00163573]
- 100 **Orava EW**, Abdul-Wahid A, Huang EH, Mallick AI, Gariépy J. Blocking the attachment of cancer cells in vivo with DNA aptamers displaying anti-adhesive properties against the carcinoembryonic antigen. *Mol Oncol* 2013; **7**: 799-811 [PMID: 23656757 DOI: 10.1016/j.molonc.2013.03.005]
- 101 **Wang LF**, Wei J, Yin HT, Qian XP, Yu LX, Liu B. In vitro selection of specific binding DNA aptamer to human carcinoembryonic antigen. *Yixue Yanjiusheng Xuebao* 2007; **9**: 2004-2005 [DOI: 10.1016/s1007-4376(07)60061-6]
- 102 **Correa CR**, de Barros AL, Ferreira Cde A, de Goes AM, Cardoso VN, de Andrade AS. Aptamers directly radiolabeled with technetium-99m as a potential agent capable of identifying carcinoembryonic antigen (CEA) in tumor cells T84. *Bioorg Med Chem Lett* 2014; **24**: 1998-2001 [PMID: 24675379 DOI: 10.1016/j.bmcl.2014.02.048]

P- Reviewer: Berg T S- Editor: Ma YJ L- Editor: A
E- Editor: Wang CH



Current status of intragastric balloon for obesity treatment

Seung Han Kim, Hoon Jai Chun, Hyuk Soon Choi, Eun Sun Kim, Bora Keum, Yoon Tae Jeen

Seung Han Kim, Hoon Jai Chun, Hyuk Soon Choi, Eun Sun Kim, Bora Keum, Yoon Tae Jeen, Division of Gastroenterology and Hepatology, Department of Internal Medicine, Institute of Gastrointestinal Medical Instrument Research, Korea University College of Medicine, Seoul 02841, South Korea

Author contributions: Kim SH and Chun HJ designed the study; Kim ES and Jeon YT assisted in creating the tables and figures; Choi HS and Keum B analyzed the data; and Kim SH wrote the manuscript.

Supported by Korea Health Technology R and D Project through the Korea Health Industry Development Institute; and Ministry of Health and Welfare, South Korea, No. HI14C3477.

Conflict-of-interest statement: Seung Han Kim, Hoon Jai Chun, Hyuk Soon Choi, Eun Sun Kim, Bora Keum and Yoon Tae Jeen have no conflicts of interest or financial ties to disclose.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Hoon Jai Chun, MD, PhD, AGAF, Division of Gastroenterology and Hepatology, Department of Internal Medicine, Institute of Gastrointestinal Medical Instrument Research, Korea University College of Medicine, 126-1, Anam-dong 5 ga, Seongbuk-gu, Seoul 02841, South Korea. drchunhj@chol.com
Telephone: +82-2-9206555
Fax: +82-2-9531943

Received: March 27, 2016
Peer-review started: March 28, 2016
First decision: April 14, 2016
Revised: April 28, 2016
Accepted: May 21, 2016

Article in press: May 23, 2016
Published online: June 28, 2016

Abstract

Endoscopic bariatric therapy may be a useful alternative to pharmacological treatment for obesity, and it provides greater efficacy with lower risks than do conventional surgical procedures. Among the various endoscopic treatments for obesity, the intragastric balloon is associated with significant efficacy in body weight reduction and relief of comorbid disease symptoms. Anatomically, this treatment is based on gastric space-occupying effects that increase the feeling of satiety and may also affect gut neuroendocrine signaling. The simplicity of the intragastric balloon procedure may account for its widespread role in obesity treatment and its applicability to various degrees of obesity. However, advances in device properties and procedural techniques are still needed in order to improve its safety and cost-effectiveness. Furthermore, verification of the physiological outcomes of intragastric balloon treatment and the clinical predictive factors for treatment responses should be considered. In this article, we discuss the types, efficacy, safety, and future directions of intragastric balloon treatment.

Key words: Intragastric balloon; Obesity; Bariatric; Metabolic; Endoscopy

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Obesity is a complex metabolic illness that is associated with several comorbid diseases. There has been a constant demand for safe and more effective weight reduction interventions to fill the gap in the treatment of obesity. The intragastric balloon is a fascinating intermediate alternative solution between

medical obesity treatment and bariatric surgical procedures for obese patients that may provide better efficacy and have a more favorable risk profile.

Kim SH, Chun HJ, Choi HS, Kim ES, Keum B, Jeon YT. Current status of intra-gastric balloon for obesity treatment. *World J Gastroenterol* 2016; 22(24): 5495-5504 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5495.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5495>

INTRODUCTION

Obesity is a complex metabolic illness that results from excess accumulation of body fat and may lead to negative health consequences. Obesity increases the prevalence of various diseases, including diabetes mellitus, hypertension, coronary heart disease, sleep apnea, stroke, gastroesophageal reflux disease, gall bladder disease, certain types of malignancy, and non-alcoholic fatty liver disease^[1]. Moreover, it is also a major avoidable health detriment. Current therapeutic approaches to obesity are lifestyle changes, pharmacologic treatment, and bariatric surgery. Although intensive lifestyle modification was reportedly associated with only limited weight reduction^[2-4], when it is combined with weight-loss drugs approved for long-term use, an additional weight reduction of 3%-9% can occur within 1 year^[5]. Such drugs are said to improve several cardiometabolic risk factors, but they are also related to harmful adverse effects^[5]. Although new obesity medications have recently been approved and introduced^[6-8], they are associated with issues of safety and high costs. Weight-loss surgery provides the most sustained and effective therapeutic choice for obesity. Accessible methods include the adjustable gastric band, Roux-en-Y gastric bypass, or sleeve gastrectomy^[9,10]. Regardless of its proven effectiveness, only 1% of obese patients eligible for the surgical procedure choose to undergo it^[11]. The major issues with surgery are difficult accessibility, high costs, patient non-preference, and significant morbidity and mortality. Although its associated mortality has decreased considerably, the complication rate in the early and late stages of the bariatric procedure persist at 17% (95%CI: 11%-23%)^[10].

Therefore, minimally invasive and effective methods are needed for the treatment of obesity. As such, endoscopic bariatric treatment was recently introduced. It includes intra-gastric balloons, gastroplasty techniques, aspiration therapy, and gastrointestinal bypass sleeves. Among them, the intra-gastric balloon has been the most frequently used in practice and the most studied for obesity treatment.

INTRAGASTRIC BALLOON

In 1985, the Garren-Edwards gastric bubble (GEGB)

was the first intra-gastric balloon approved for obesity treatment and was introduced in the United States market. It was made of polyurethane, had a cylindrical design, and was filled with 200-220 mL of air^[12]. However, several adverse events were associated with its use, including small bowel obstruction associated with spontaneous deflation and gastric mucosal injury. Although the GEGB is no longer used, considerable advancements to its design have led to the development of a more effective and safer intra-gastric balloon. It is now being used in numerous countries. Additionally, the United States Food and Drug Administration (FDA) recently approved two new intra-gastric balloons.

The increased prevalence of obesity has motivated experts in bariatric medicine to advance minimally invasive endoscopic treatment for obesity management as well as innovative techniques that address important features of treatments, such as their efficiency and safety. A new meta-analysis showed that endoscopic obesity treatment could be effective and of substantial value if combined with a multidisciplinary and comprehensive treatment plan^[13].

The intra-gastric balloon technique has become an effective method of achieving significant weight reduction in obese people (Figure 1). One or more intra-gastric balloons can be placed in the stomach using endoscopic procedures under mild sedation in an outpatient setting. Intra-gastric balloons allow patients to sense fullness and ultimately reduce their food intake. It is hypothesized that the intra-gastric balloon facilitates satiety peripherally by being an obstacle to food consumption, decreasing intra-gastric volume, and delaying gastric emptying^[14]. Additionally, signals transmitted centrally through the vagal nerves by activated gastric stretching receptors could affect satiety^[14]. The intra-gastric balloon permits an early feeling of satiety, which is thought to be a consequence of gastric distention. The mechanical intra-gastric distention to a meaningful volume during mealtime significantly decreases the amount of food eaten^[15,16].

The intra-gastric balloon may also act *via* its relationship with various neurohumoral factors. It affects hunger control and gastric emptying by altering gut hormones and peptides such as ghrelin, leptin, cholecystokinin, and pancreatic polypeptide^[17,18] and may also be related to physiological adaptation to weight loss.

The intra-gastric balloon may play diverse roles in obesity treatment as a preemptive therapy, a metabolic therapy, or a primary therapy.

TYPES OF INTRAGASTRIC BALLOONS

Orbera intra-gastric balloon

The most frequently used balloon is the Orbera Intra-gastric Balloon (Apollo Endosurgery, Austin, TX, United States), which was known previously as the BioEnterics Intra-gastric Balloon (BIB). It is an elastic

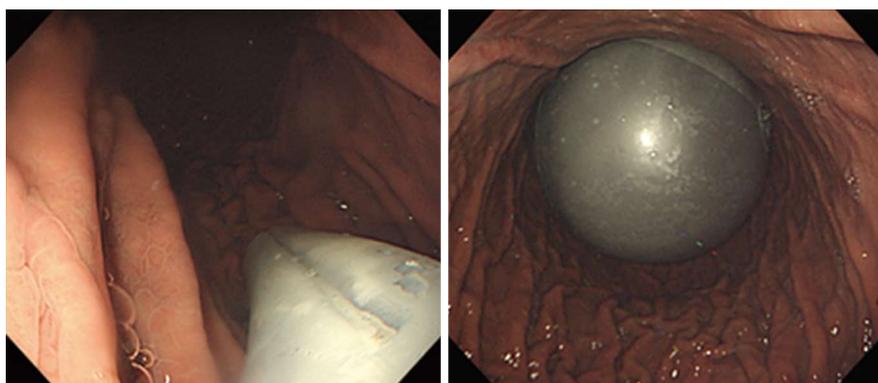


Figure 1 Intra-gastric balloon placement.

Table 1 Types of intra-gastric balloons

Balloon	Material	Volume	Weight loss ¹	Note
Fluid-supplied Orbera (Apollo Endosurgery)	Silicone	400-700 mL (saline)	16.9 ± 0.9 kg (at 6 mo) ^[58]	Most widely used and studied intra-gastric balloon Volume adjustment at the time of placement
Spatz Adjustable Balloon system (Spatz FGIA)	Silicone	400-600 mL (saline)	24 kg (at 12 mo) ^[64]	Totally adjustable balloon Approved for 12 mo of implantation
ReShape Duo [®] Integrated DualBalloon System (ReShape medical)	Silicone	900 mL (450 mL × 2) (saline)	25.1 ± 1.6 %EWL (at 6 mo) ^[62]	Two independent balloons connected to silicone shaft
The Elipse [™] (Allurion Technologies)	N/A	450-550 mL (filling fluid)	2.4 kg (at 6 wk) ^[19]	Naturally swallowed, self-emptying, and naturally excreted
Air-supplied Obalon [®] Gastric Balloon (Obalon Therapeutics)	N/A	250 mL (air, nitrogen)	5 kg (at 12 wk) ^[66]	Can be swallowed A second or a third balloon can be swallowed depending on patient's progress
Heliosphere BAG [®] (Helioscopie)	Polyurethane and silicone	950 mL (air)	16 ± 7 kg (at 6 mo) ^[23]	Less than 30g

¹Values extrapolated from representative reviews and clinical trials of each intervention. N/A: Not available; %EWL: Percentage of excess weight loss.

silicone balloon containing saline (450-700 mL) (Table 1). The positioning assembly, which comprises a balloon-filling tube and a catheter with the deflated balloon, is blindly advanced to the gastro-esophageal junction. An endoscopic device is inserted to ensure the precise deployment of the intra-gastric balloon, which is then filled with methylene-mixed saline under direct observation *via* the catheter. If an unexpected balloon rupture occurs, the methylene blue turns the urine green. The Orbera balloon is usually implanted for 6 mo, removed endoscopically by needle aspiration of the intra-gastric fluid, and retrieved with a snare or grasper. The FDA approved the use of the Orbera balloon on August 6, 2015. It is expected that the Orbera balloon could provide a valuable and less invasive therapeutic approach to bariatric treatment.

ReShape duo

The ReShape Duo[®] (ReShape Medical, San Clemente, CA, United States) aims to improve the space-occupying effects of intra-gastric balloons. This non-invasive device is delivered *via* the mouth through a

30-min endoscopic procedure. It is inflated with 900 mL of saline, which is equally distributed to each of two balloons. The balloon is inflated by a controller with methylene blue-mixed saline. This dual balloon system may reduce deflation-associated complications. If one of the balloons ruptures, the other balloon could sustain the location of the device in the stomach. It is recommended that the balloon be retrieved 6 mo after placement. The ReShape Duo was also approved by the FDA, on July 29, 2015.

Spatz

The Spatz Adjustable Balloon System (Spatz Medical, Great Neck, NY, United States) is a silicone balloon that is inflated with saline. It includes a filling catheter, which is extractable endoscopically, that permits an intra-gastric volume adjustment of 400-800 mL. The volume of the intra-gastric balloon can be modified to improve the patient's tolerance and increase weight reduction. The Spatz Adjustable Balloon System is allowed to be implanted in the stomach for 12 mo in locations outside the United States.

Elipse

The Elipse™ (Allurion Technologies, Wellesley, MA, United States) is a swallowable, self-draining, and naturally expelled intra-gastric balloon device for weight reduction. It is covered with a vegetarian shell and fixed to a flexible, slim tube. A resorbable substance inside the balloon degrades, allowing the balloon to empty naturally after a certain period. The deflated intra-gastric balloon is devised to be excreted *via* the digestive tract^[19]. It is inflated with 550 mL of fluid and can be placed in the stomach for 4 mo. Endoscopic procedures are not needed for Elipse™ placement or removal.

Obalon

The Obalon Gastric Balloon® (Obalon Therapeutics Inc, Carlsbad, CA, United States) is a balloon filled with 250 mL of air. It has a self-sealing valve linked to a catheter and is set inside a gelatin capsule. The balloon is packaged in the capsule and swallowed, and the thin catheter is extended alongside the esophagus into the stomach. Fluoroscopy is used to define the location of the intra-gastric balloon, and the gelatin capsule disintegrates and releases the balloon. The balloon is then inflated using a gas-contained canister. The catheter is separated from the balloon and removed after balloon inflation.

Heliosphere bag

The Heliosphere Bag® (Helioscopie, Vienne, France) comprises a double-bagged polymer covered with an external silicone pouch. It is slowly filled with 960 mL of air for a final inflation volume of 700 mL^[20,21]. The weight of the Heliosphere Bag is less than 30 g^[22,23].

Silimed balloon

The Silimed Gastric Balloon is a spherical silicone balloon set inside a thin silicone sheath^[24]. It is attached to the endoscope by a snare, advanced by an endoscope, and placed in the gastric fundus. It is filled with 650 mL of saline solution mixed with contrast dye and 10 mL of methylene blue.

Adjustable totally implantable intra-gastric prosthesis

The adjustable totally implantable intra-gastric prosthesis (ATIIP) is a polyurethane intra-gastric balloon that is rugby-shaped, 12 cm long, and has a volume of 300 mL when inflated with air^[25]. This balloon is placed with an endoscopic percutaneous gastrostomy technique followed by the deployment of a subcutaneous totally implantable system through a surgical procedure. This method may reduce the likelihood of device dislocation and allows for balloon volume adjustment. The proximal balloon position in the gastric fundus-corpora lesion could affect gastric accommodation, neurohormonal process, and electrical action, thus modifying various control processes related to satiety^[14].

INTRAGASTRIC BALLOON TREATMENT FOR OBESITY

Indications

The intra-gastric balloon may offer a minimally invasive and valuable method for managing obesity and related conditions. It is used to achieve weight loss in obese people, generally those with a body mass index (BMI) > 35 kg/m², or 30 kg/m² with certain comorbidities. Intra-gastric balloon treatment may play a different role in bariatric treatment based on the grade of obesity.

Preemptive therapy

Early intervention and preemptive therapy for weight reduction can be performed in obese patients (BMI ≥ 30 kg/m²) at risk for disease development, at high risk for all-cause mortality, and with a high cardiovascular risk profile^[26].

The objective of preemptive treatment is to achieve modest weight reduction, and, therefore, the overall risk/benefit ratio could validate the standards for procedures with this indication. Depending on the circumstances, indications for placement of intra-gastric balloons could also be extended to manage overweight people with a BMI < 30 kg/m² who desire weight reduction but who cannot achieve body weight loss with a controlled dietary program or with pharmacotherapy^[27].

Metabolic therapy

Body weight loss achieved with intra-gastric balloon placement is associated with improvements in obesity-related metabolic illness. The intra-gastric balloon for metabolic therapy may be performed in patients with mild obesity (BMI ≥ 30 kg/m²), where recovery from metabolic disease is the primary concern^[28]. Co-existing illnesses, such as hyperlipidemia, type II diabetes mellitus, and hypertension, could be particularly improved or resolved with even a modest reduction in body weight^[29,30]. Treatment for metabolic issues should be relatively low-risk and have superior stability.

Primary therapy

The goal of intra-gastric balloon treatment is to achieve weight reduction in severely obese people, generally those with a BMI > 35 kg/m² with or without comorbidities, and who could not achieve long-term weight loss with a weight-control regimen. In addition, intra-gastric balloon therapy could be performed in patients with a BMI ≥ 40 kg/m², primarily as a preparation for bariatric treatment or in patients with increased surgical risks. Obese patients who reject bariatric surgical procedures or who do not have an approach for surgery can also opt for it. Intra-gastric balloon therapy used as a primary therapy could induce weight reduction and improve obesity-related comorbidities with a level of safety and efficiency

comparable to that of bariatric surgery^[28]. However, lower efficiency is also acceptable because of the lower risk profile of intra-gastric balloon therapy.

Exclusion criteria

Exclusion criteria include any situation that could increase the risks related to intra-gastric balloon insertion, such as a large hiatal hernia (> 5 cm), active ulcer in the stomach or duodenum, previous surgical resection of the stomach, inflammatory bowel disease, gastrointestinal neoplasm, oropharyngeal abnormalities, active gastrointestinal bleeding, coagulative disorder, variceal disease, alcoholic disease or drug abuse, psychiatric disease, pregnancy, use of anti-coagulants or anti-inflammatory drugs, and cardiovascular, pulmonary or cerebrovascular diseases^[31].

EFFECTS OF INTRAGASTRIC BALLOONS

The purpose of intra-gastric balloon treatment is to stimulate weight loss and assist with recovery from associated comorbidities with adequate safety. Gastric capacity restriction is an essential factor in surgical bariatric management.

Surgical gastric restriction could induce early satiety and potentiate gastric mechanical and chemical stimulation through a relationship with various gastric or exogenic factors. It affects hunger control and gastric emptying through alterations in gut hormones and peptides^[32-34]. The intra-gastric balloon attempts to mimic surgical weight loss procedures by restricting the effective gastric volume.

Body weight loss

Results from previous studies indicated that the mean weight loss associated with intra-gastric balloon therapy ranged between 10.5 and 13.7 kg after 3 mo and between 12 and 26.3 kg after a 6-mo placement of the Orbera intra-gastric balloon^[22,23,35-49]. After balloon removal at 6 mo, the achieved weight reduction endured to some extent. The excess weight loss (EWL) at the 12-mo implantation mark (6 mo after removal) was 14% to 50.9%^[37,40,42,44,50-54]. The EWL at 12 mo after balloon extraction ranged from 14.2% to 27.2%^[51,55-57]. The Orbera intra-gastric balloon was most effective during the first 3 mo of therapy. During that time, average weight loss of obese patients was 12.9 kg, or 80% of the total achieved weight reduction^[58]. Additionally, the initial body weight loss (BWL) following intra-gastric balloon placement was associated with significant long-term weight maintenance. The percentage of BWL 1 mo after intra-gastric balloon placement was significantly associated with weight loss after 6, 12, and 18 mo (Pearson correlation coefficient = 0.77, 0.65, and 0.62, respectively, $P < 0.001$ for all)^[59].

A study reported that insertion of a second balloon is not difficult and achieved good results.

The mean %EWL was 31.5 ± 23.2 after the second balloon removal^[43]. Some studies showed long-term results after intra-gastric balloon placement. A multi-center European study presented results for weight loss 3 years after balloon removal, which accounted for 29.1% of mean EWL^[54]. A study that included 195 patients who were followed up for 5 years after intra-gastric balloon insertion demonstrated a %EWL of 12.97 ± 8.54 ^[44].

Additional limited studies have utilized other intra-gastric balloons. A Spanish study with 60 patients showed a total body weight reduction of 16.6 ± 9.33 kg 6 mo after implantation of the ReShape Duo double-balloon system^[60]. A prospective study with a double-balloon system showed a mean EWL of 18.3% in the control group compared to 31.8% in the treatment group 6 mo post-balloon implantation^[61]. Furthermore, 64% of the reduced body weight was maintained at 12 mo post-implantation (6 mo after removal). In the REDUCE Pivotal Trial, a prospective randomized trial with 326 patients, patients assigned to the double-balloon system plus exercise and diet showed significantly superior EWL compared to the sham endoscopy plus exercise and diet alone group at 6 mo post-implantation (25.1% vs 11.3%, $P < 0.05$) in an intent-to-treat analysis^[62].

A pilot trial showed 15.6 kg of mean weight loss at 24 wk and 24.4 kg at 52 wk after Spatz adjustable balloon placement^[63]. A small observational investigation in 73 obese patients showed 45.7% of EWL 12 mo after deployment of the Spatz adjustable balloon^[64].

Two preliminary studies with the Silimed Gastric Balloon reported results after the completion of 6 mo of treatment. The mean weight loss was 11.3 kg^[24] and 8.1 kg^[65]. A total of 57 morbidly obese patients underwent ATIIP placement. Mean EWL was 28.7% at 6 mo (38 patients) and 39.2% at 12 mo (20 patients)^[25]. The Obalon (orally ingestible intra-gastric balloon) showed median weight losses after 4 weeks, 8 weeks, and 12 wk as 2.2 kg, 4.0 kg, and 5 kg, respectively^[66]. A prospective study with 50 obese patients compared the effects of intra-gastric balloon therapy or pharmacotherapy on weight reduction. At 6 mo, patients in the intra-gastric balloon group had lost more weight than had patients in the pharmacotherapy group (percent of initial weight lost, %IWL = 14.5 ± 1.2 ; percent of excess BMI lost, %EBL = 37.7 ± 3.2 vs %IWL = 9.1 ± 1.5 , %EBL = 25.3 ± 4.1 , respectively, $P < 0.005$)^[67].

Although the intra-gastric balloon has been shown to be effective in causing a meaningful weight loss, several studies have reported that the results were short-lasting, with most patients regaining weight following intra-gastric balloon removal^[31,36,51,55,68-72].

Improvement in metabolic diseases

Obesity in patients is related to several comorbidities that are significant targets for obesity treatment. A

Table 2 Plasma ghrelin and leptin level after intra-gastric balloon treatment

Ref.	Balloon	Weight loss at 6 mo	Ghrelin (at 0 mo)	Ghrelin (after 3 mo)	Ghrelin (after 6 mo)	Ghrelin (after 12 mo)	Leptin (at 0 mo)	Leptin (after 3 mo)	Leptin (after 6 mo)	Leptin (after 12 mo)
Mathus-Vliegen <i>et al</i> ^[17]	Orbera	17.4 ± 7.8 kg	722.3 ± 151.5 pg/mL	791.5 ± 239.0 pg/mL	743.7 ± 115.2 pg/mL	N/A	N/A	N/A	N/A	N/A
Fuller <i>et al</i> ^[48]	N/A	14.2%	414.1 pmol/L	448 pmol/L	452.4 pmol/L	379.4 pmol/L	23.4 ng/mL	18.5 ng/mL	11.7 ng/mL	19.7 ng/mL
Bužga <i>et al</i> ^[85]	MedSil [®]	18.4 ± 8.2 kg	240.5 ± 101.5 μg/L	378.1 ± 155.8 μg/L	335.8 ± 149.2 μg/L	N/A	30.4 ± 17.2 μg/L	18.2 ± 15.8 μg/L	14.9 ± 15.5 μg/L	N/A
¹ Nikolic <i>et al</i> ^[86]	Orbera	N/A	958.3 pg/mL	1346.2 pg/mL	1050.1 pg/mL	922.6 pg/mL	25.1 ng/mL	14.3 ng/mL	10.5 ng/mL	17.5 ng/mL
Konopko-Zubrzycka <i>et al</i> ^[39]	Orbera	17.1 ± 8.0 kg	621.9 ± 182.4 pg/mL	903.9 ± 237 pg/mL (1 mo)	N/A	N/A	61.3 ± 36.7 ng/mL	39.9 ± 17.5 ng/mL (1 mo)	N/A	N/A
Martinez-Brocca <i>et al</i> ^[79]	Orbera	12.7 ± 5.6 kg (4 mo)	934.4 ± 199.2 pg/mL	947.1 ± 195.1 pg/mL (1 mo)	N/A	N/A	31.9 ± 16.4 ng/mL	22.4 ± 15.1 ng/mL (1 mo)	N/A	N/A
Mion <i>et al</i> ^[78]	Orbera	8.7 kg	3.2 ± 0.4 ng/mL	N/A	1.9 ± 0.1 ng/mL	N/A	27.8 ± 3.7 ng/mL	N/A	18.7 ± 2.7 ng/mL	N/A

¹Data from patients with body mass indexes < 40 kg/m². Data are presented as mean ± SD or median. N/A: Not available.

study with 143 obese patients described the effects of the Orbera intra-gastric balloon at the 12-mo follow-up^[51]. The incidence of metabolic syndrome declined from 34.8% (before balloon insertion) to 14.5%, 13%, and 11.6% at the time of removal, at the 6-mo follow-up, and at the 1-year follow-up, respectively. The incidence of type 2 diabetes mellitus decreased from 32.6% to 20.9%, 22.5%, and 21.3%, respectively. Likewise, the occurrence of hyperuricemia, hypertriglyceridemia, and hypercholesterolemia decreased from 26.1%, 37.7%, and 33.4% to 25.4%, 14.5%, and 16.7%, respectively, at the time of removal, 25.9%, 15.2%, and 16.7%, respectively, at the 6-mo follow-up, and 26.4%, 17.4%, and 18.9%, respectively, at the 1-year follow-up. A multi-center study presented data following treatment with the Orbera intra-gastric balloon in overweight populations^[54]. The percentage of patients with comorbidities at baseline and at the 3-year follow-up was 29% and 16% for hypertension, 15% and 10% for diabetes mellitus, 20% and 18% for dyslipidemia, 32% and 21% for hypercholesterolemia, and 25% and 13% for osteoarthropathy, respectively. A study with 119 obese patients assessed the effects of the Orbera intra-gastric balloon on obesity-associated diseases and quality of life^[42]. Six months after placement of the intra-gastric balloon, the rate of metabolic syndrome in the patients decreased from 42.9% to 15.1% ($P < 0.0005$). Cholesterol, triglycerides, fasting glucose, C-reactive protein levels, and blood pressure also improved after balloon treatment ($P < 0.005$). In patients with diabetes mellitus, the HbA1c level was decreased at 6 mo compared to that at baseline (7.4% to 5.8%; $P < 0.0005$). In addition, the quality of life of patients increased.

Obesity is one of the risk factors for nonalcoholic fatty liver disease (NAFLD). About 27% of patients with NAFLD can develop fibrosis, and 19% can develop cirrhosis^[73]. A randomized controlled study

showed that intra-gastric balloon therapy improved the histology of nonalcoholic steatohepatitis^[74]. Six months after implantation, a significant reduction in the median NAFLD activity scores was observed in the intra-gastric balloon-treated group (2 vs 4, $P = 0.030$) compared to that in the control group. Additionally, the median steatosis scores displayed a trend toward improvement in the balloon-treated group compared to those in the control group (1 vs 1, $P = 0.075$).

Alterations in gastrointestinal hormones

Body weight is controlled by multifaceted coordination of both central and peripheral factors. It is now obvious that a physiologic change in gut neurohumoral signaling is one important contributor to weight reduction and improvement in related diseases obtained by anatomic surgical manipulation of the gastrointestinal tract^[75]. Moreover, intra-gastric balloon treatment may affect weight changes through an interaction of gastric neurohumoral factors (Table 2). These factors include several gut peptides and hormones, such as ghrelin, leptin, cholecystokinin, peptide YY, pancreatic polypeptide, and glucagon-like peptide-1.

These factors are interrelated and participate in the peripheral mediation of satiety^[76,77]. Ghrelin is a hormone that has been known to influence energy balance. However, several studies presented varying results regarding ghrelin concentrations after balloon treatment. A study with 40 obese patients who underwent balloon placement indicated no effect on ghrelin levels when patients were fasting or meal-suppressed^[17]. In another study, 17 patients with non-morbid obesity underwent balloon placement, and fasting plasma ghrelin concentrations significantly decreased (3.2 to 1.9 ng/mL; $P = 0.021$) as a result^[78]. Martinez-Brocca *et al*^[79] reported that fasting and meal-suppressed plasma ghrelin levels did not



Figure 2 Gastric ulcer induced by intra-gastric balloon placement.

differ significantly between groups in morbidly obese patients. Konopko-Zubrzycka *et al.*^[39] reported that body weight reduction after balloon treatment is related to a transient elevation in plasma ghrelin levels and a decrease in plasma leptin levels.

Another study evaluated fasting and postprandial cholecystokinin and pancreatic polypeptide secretion after 13 wk of balloon treatment in obese patients. Baseline and meal-stimulated cholecystokinin levels were decreased^[18].

Maintenance of weight loss is controlled by the collaboration of several processes, including environmental, behavioral, and homeostatic factors^[80]. Physiological adaptation to weight reduction and weight regain show interplaying alterations in the stability of the levels of hunger hormones and energy homeostasis, in addition to changes in subjective appetite and nutrient metabolism. Appetite-related gastrointestinal hormones may play a crucial function in body weight regain after weight reduction^[81]. Bariatric procedures may prove to be effective methods to aid in altering obese patients' physiology and may provide a good opportunity for long-term maintenance. Procedures that affect the physiology of weight loss and regain may eventually fortify future approaches for obesity treatment.

COMPLICATIONS OF THE INTRAGASTRIC BALLOON

Adverse events rates following Orbera intra-gastric balloon placement showed pooled incidences of pain (33.7%) and nausea (29%) as common adverse side effects^[82]. The incidence of GERD, gastric ulcers (Figure 2), and balloon migration was 18.3%, 2%, and 1.4%, respectively. Serious adverse events with the Orbera balloon are uncommon, with prevalences of small bowel obstruction, perforation, and death as 0.3%, 0.1% and 0.08%, respectively^[82]. Similarly, the REDUCE pivotal trial ($n = 264$), which evaluated the efficacy and safety of the ReShape Duo intra-gastric balloon, showed that symptoms such as nausea, abdominal pain, and vomiting were common, but

decreased rapidly with medical management. Spontaneous intra-gastric balloon deflation occurred in 6% of patients, but balloon migrations did not occur concurrently. Early balloon removal occurred in 9.1% of study participants for non-ulcer-related intolerance. In 35% of the study participants, a gastric ulcer presented initially, but ulcer incidence and size decreased following subsequent device modification^[62].

FUTURE DIRECTIONS FOR THE INTRAGASTRIC BALLOON

New emerging technology trends in endoscopic treatment for obesity require an extensive and meticulous research plan to promote finding and recognize their optimal role in managing obese patients and their applications for clinical practice^[83]. Intra-gastric balloon placement can be performed through a simple endoscopic method and is easily reversible. This simplicity offers an expansive role in obesity treatment based on the degree of obesity. It is important to establish an appropriate method for intra-gastric balloon treatment by classifying the degree of obesity. Although serious complications with intra-gastric balloon treatment are rare, safety is an issue that cannot be overlooked. Various factors are associated with balloon safety, such as its durability and the simplicity of the procedure. Advances in device properties and procedural techniques could offer more cost-effectiveness with this outpatient procedure^[84]. There is also uncertainty about the ideal filling medium and content. A preference for fluid other than air has been mentioned^[14], but a debate still exists, with no established guidelines to date. There is a need to establish standards of intra-gastric balloon treatment practice, which could comprise pre- and post-procedural guidelines and long-term schedule management. In addition, collaboration with dietary counseling and an exercise program could offer quality assurance with intra-gastric balloon treatment.

Intra-gastric balloon treatment might produce only short-lasting effects in obesity treatment. Thus, it is important to maintain weight loss following intra-gastric balloon removal. Long-term management for weight reduction after intra-gastric balloon removal can also comprise intensive lifestyle modification, alone or with pharmacotherapy, and could be suggested to protect against weight regain.

Material and safety issues regarding the intra-gastric balloon should be further investigated. More research should be performed to investigate a pathophysiologic pattern of obesity, the uncertain role of gut hormones, potential predictive factors for the efficacy of the intra-gastric balloon in obesity treatment, and individualized treatment-induced changes.

CONCLUSION

Effective and safe weight reduction is very important

for the treatment of obesity, which is responsible for about 5% of all obesity-related deaths globally. Intra-gastric balloon treatment is more than just a transient curiosity. It shows promise in improving the quality of life and health status for obese patients. It offers a minimally invasive and effective method for managing obesity and associated conditions. Although there remains the possibility of improvement with research and development, gastroenterologists should maintain attention and interest in determining satisfactory outcomes and verifying standards of intra-gastric balloon treatment for the obesity epidemic.

REFERENCES

- 1 **Pi-Sunyer X.** The medical risks of obesity. *Postgrad Med* 2009; **121**: 21-33 [PMID: 19940414 DOI: 10.3810/pgm.2009.11.2074]
- 2 **Gregg EW,** Chen H, Wagenknecht LE, Clark JM, Delahanty LM, Bantle J, Pownall HJ, Johnson KC, Safford MM, Kitabchi AE, Pi-Sunyer FX, Wing RR, Bertoni AG. Association of an intensive lifestyle intervention with remission of type 2 diabetes. *JAMA* 2012; **308**: 2489-2496 [PMID: 23288372 DOI: 10.1001/jama.2012.67929]
- 3 **Knowler WC,** Fowler SE, Hamman RF, Christophi CA, Hoffman HJ, Brenneman AT, Brown-Friday JO, Goldberg R, Venditti E, Nathan DM. 10-year follow-up of diabetes incidence and weight loss in the Diabetes Prevention Program Outcomes Study. *Lancet* 2009; **374**: 1677-1686 [PMID: 19878986 DOI: 10.1016/S0140-6736(09)61457-4]
- 4 **Look AHEAD Research Group.** Eight-year weight losses with an intensive lifestyle intervention: the look AHEAD study. *Obesity* (Silver Spring) 2014; **22**: 5-13 [PMID: 24307184 DOI: 10.1002/oby.20662]
- 5 **Yanovski SZ,** Yanovski JA. Long-term drug treatment for obesity: a systematic and clinical review. *JAMA* 2014; **311**: 74-86 [PMID: 24231879 DOI: 10.1001/jama.2013.281361]
- 6 **Patel D.** Pharmacotherapy for the management of obesity. *Metabolism* 2015; **64**: 1376-1385 [PMID: 26342499 DOI: 10.1016/j.metabol.2015.08.001]
- 7 **Rankin W,** Wittert G. Anti-obesity drugs. *Curr Opin Lipidol* 2015; **26**: 536-543 [PMID: 26382553 DOI: 10.1097/MOL.0000000000000232]
- 8 **Fujioka K.** Current and emerging medications for overweight or obesity in people with comorbidities. *Diabetes Obes Metab* 2015; **17**: 1021-1032 [PMID: 26040215 DOI: 10.1111/dom.12502]
- 9 **Ochner CN,** Gibson C, Carnell S, Dambkowski C, Geliebter A. The neurohormonal regulation of energy intake in relation to bariatric surgery for obesity. *Physiol Behav* 2010; **100**: 549-559 [PMID: 20452367 DOI: 10.1016/j.physbeh.2010.04.032]
- 10 **Chang SH,** Stoll CR, Song J, Varela JE, Eagon CJ, Colditz GA. The effectiveness and risks of bariatric surgery: an updated systematic review and meta-analysis, 2003-2012. *JAMA Surg* 2014; **149**: 275-287 [PMID: 24352617 DOI: 10.1001/jamasurg.2013.3654]
- 11 **Buchwald H,** Oien DM. Metabolic/bariatric surgery worldwide 2011. *Obes Surg* 2013; **23**: 427-436 [PMID: 23338049 DOI: 10.1007/s11695-012-0864-0]
- 12 **Gleysteen JJ.** A history of intra-gastric balloons. *Surg Obes Relat Dis* 2016; **12**: 430-435 [PMID: 26775045 DOI: 10.1016/j.soard.2015.10.074]
- 13 **Abu Dayyeh BK,** Kumar N, Edmundowicz SA, Jonnalagadda S, Larsen M, Sullivan S, Thompson CC, Banerjee S. ASGE Bariatric Endoscopy Task Force systematic review and meta-analysis assessing the ASGE PIVI thresholds for adopting endoscopic bariatric therapies. *Gastrointest Endosc* 2015; **82**: 425-438.e5 [PMID: 26232362 DOI: 10.1016/j.gie.2015.03.1964]
- 14 **Mathus-Vliegen EM.** Endoscopic treatment: the past, the present and the future. *Best Pract Res Clin Gastroenterol* 2014; **28**: 685-702 [PMID: 25194184 DOI: 10.1016/j.bpg.2014.07.009]
- 15 **Geliebter A,** Westreich S, Gage D. Gastric distention by balloon and test-meal intake in obese and lean subjects. *Am J Clin Nutr* 1988; **48**: 592-594 [PMID: 3414573]
- 16 **Geliebter A,** Melton PM, McCray RS, Gage D, Heymsfield SB, Abiri M, Hashim SA. Clinical trial of silicone-rubber gastric balloon to treat obesity. *Int J Obes* 1991; **15**: 259-266 [PMID: 2071316]
- 17 **Mathus-Vliegen EM,** Eichenberger RI. Fasting and meal-suppressed ghrelin levels before and after intra-gastric balloons and balloon-induced weight loss. *Obes Surg* 2014; **24**: 85-94 [PMID: 23918282 DOI: 10.1007/s11695-013-1053-5]
- 18 **Mathus-Vliegen EM,** de Groot GH. Fasting and meal-induced CCK and PP secretion following intra-gastric balloon treatment for obesity. *Obes Surg* 2013; **23**: 622-633 [PMID: 23224567 DOI: 10.1007/s11695-012-0834-6]
- 19 **Machytka E,** Chuttani R, Bojkova M, Kupka T, Buzga M, Stecco K, Levy S, Gaur S. Elipse™, a Procedureless Gastric Balloon for Weight Loss: a Proof-of-Concept Pilot Study. *Obes Surg* 2016; **26**: 512-516 [PMID: 26253980 DOI: 10.1007/s11695-015-1783-7]
- 20 **Mion F,** Gincul R, Roman S, Beorchia S, Hedelius F, Claudel N, Bory RM, Malvoisin E, Trepo F, Napoleon B. Tolerance and efficacy of an air-filled balloon in non-morbidly obese patients: results of a prospective multicenter study. *Obes Surg* 2007; **17**: 764-769 [PMID: 17879576]
- 21 **Forestieri P,** De Palma GD, Formato A, Giuliano ME, Monda A, Pilone V, Romano A, Tramontano S. Heliosphere Bag in the treatment of severe obesity: preliminary experience. *Obes Surg* 2006; **16**: 635-637 [PMID: 16687034 DOI: 10.1381/096089206776945156]
- 22 **De Castro ML,** Morales MJ, Del Campo V, Pineda JR, Pena E, Sierra JM, Arbones MJ, Prada IR. Efficacy, safety, and tolerance of two types of intra-gastric balloons placed in obese subjects: a double-blind comparative study. *Obes Surg* 2010; **20**: 1642-1646 [PMID: 20390374 DOI: 10.1007/s11695-010-0128-9]
- 23 **Giardiello C,** Borrelli A, Silvestri E, Antognozzi V, Iodice G, Lorenzo M. Air-filled vs water-filled intra-gastric balloon: a prospective randomized study. *Obes Surg* 2012; **22**: 1916-1919 [PMID: 23054576 DOI: 10.1007/s11695-012-0786-x]
- 24 **Carvalho GL,** Barros CB, Okazaki M, Novaes ML, Albuquerque PC, Almeida NC, Albuquerque PP, Wakiyama C, Vilaça TG, Silva JS, Coelho RM. An improved intra-gastric balloon procedure using a new balloon: preliminary analysis of safety and efficiency. *Obes Surg* 2009; **19**: 237-242 [PMID: 18581191 DOI: 10.1007/s11695-008-9592-x]
- 25 **Gaggiotti G,** Tack J, Garrido AB, Palau M, Cappelluti G, Di Matteo F. Adjustable totally implantable intra-gastric prosthesis (ATIIP)-Endogast for treatment of morbid obesity: one-year follow-up of a multicenter prospective clinical survey. *Obes Surg* 2007; **17**: 949-956 [PMID: 17894156 DOI: 10.1007/s11695-007-9174-3]
- 26 **Calle EE,** Thun MJ, Petrelli JM, Rodriguez C, Heath CW. Body-mass index and mortality in a prospective cohort of U.S. adults. *N Engl J Med* 1999; **341**: 1097-1105 [PMID: 10511607 DOI: 10.1056/NEJM199910073411501]
- 27 **Mitura K,** Garnysz K. In search of the ideal patient for the intra-gastric balloon - short- and long-term results in 70 obese patients. *Wideochir Inne Tech Maloinwazyjne* 2016; **10**: 541-547 [PMID: 26865890 DOI: 10.5114/wiitm.2015.55748]
- 28 **Ginsberg GG,** Chand B, Cote GA, Dallal RM, Edmundowicz SA, Nguyen NT, Pryor A, Thompson CC. A pathway to endoscopic bariatric therapies. *Gastrointest Endosc* 2011; **74**: 943-953 [PMID: 22032311 DOI: 10.1016/j.gie.2011.08.053]
- 29 **Poobalan A,** Aucott L, Smith WC, Avenell A, Jung R, Broom J, Grant AM. Effects of weight loss in overweight/obese individuals and long-term lipid outcomes--a systematic review. *Obes Rev* 2004; **5**: 43-50 [PMID: 14969506 DOI: 10.1111/j.1467-789X.2004.00127.x]
- 30 **Poobalan AS,** Aucott LS, Smith WC, Avenell A, Jung R, Broom J. Long-term weight loss effects on all cause mortality in overweight/obese populations. *Obes Rev* 2007; **8**: 503-513 [PMID: 17949355]

- DOI: 10.1111/j.1467-789X.2007.00393.x]
- 31 **Dumonceau JM.** Evidence-based review of the Bioenterics intra-gastric balloon for weight loss. *Obes Surg* 2008; **18**: 1611-1617 [PMID: 18568377 DOI: 10.1007/s11695-008-9593-9]
 - 32 **Phillips RJ,** Powley TL. Gastric volume rather than nutrient content inhibits food intake. *Am J Physiol* 1996; **271**: R766-R769 [PMID: 8853402]
 - 33 **Kojima M,** Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature* 1999; **402**: 656-660 [PMID: 10604470]
 - 34 **Cummings DE,** Weigle DS, Frayo RS, Breen PA, Ma MK, Dellinger EP, Purnell JQ. Plasma ghrelin levels after diet-induced weight loss or gastric bypass surgery. *N Engl J Med* 2002; **346**: 1623-1630 [PMID: 12023994 DOI: 10.1056/Nejm0a012908]
 - 35 **Bonazzi P,** Petrelli MD, Lorenzini I, Peruzzi E, Nicolai A, Galeazzi R. Gastric emptying and intra-gastric balloon in obese patients. *Eur Rev Med Pharmacol Sci* 2005; **9**: 15-21 [PMID: 16457125]
 - 36 **Genco A,** Balducci S, Bacci V, Materia A, Cipriano M, Baglio G, Ribaud MC, Maselli R, Lorenzo M, Basso N. Intra-gastric balloon or diet alone? A retrospective evaluation. *Obes Surg* 2008; **18**: 989-992 [PMID: 18483834 DOI: 10.1007/s11695-007-9383-9]
 - 37 **Genco A,** Cipriano M, Materia A, Bacci V, Maselli R, Musmeci L, Lorenzo M, Basso N. Laparoscopic sleeve gastrectomy versus intra-gastric balloon: a case-control study. *Surg Endosc* 2009; **23**: 1849-1853 [PMID: 19169745 DOI: 10.1007/s00464-008-0285-2]
 - 38 **Göttig S,** Daskalakis M, Weiner S, Weiner RA. Analysis of safety and efficacy of intra-gastric balloon in extremely obese patients. *Obes Surg* 2009; **19**: 677-683 [PMID: 19291338 DOI: 10.1007/s11695-009-9820-z]
 - 39 **Konopko-Zubrzycka M,** Baniukiewicz A, Wróblewski E, Kowalska I, Zarzycki W, Górska M, Dabrowski A. The effect of intra-gastric balloon on plasma ghrelin, leptin, and adiponectin levels in patients with morbid obesity. *J Clin Endocrinol Metab* 2009; **94**: 1644-1649 [PMID: 19258408 DOI: 10.1210/jc.2008-1083]
 - 40 **Ohta M,** Kitano S, Kai S, Shiromizu A, Eguchi H, Endo Y, Masaki T, Kakuma T, Yoshimatsu H. Initial Japanese experience with intra-gastric balloon placement. *Obes Surg* 2009; **19**: 791-795 [PMID: 18592329 DOI: 10.1007/s11695-008-9612-x]
 - 41 **Coskun H,** Bostanci O. Assessment of the application of the intra-gastric balloon together with sibutramine: a prospective clinical study. *Obes Surg* 2010; **20**: 1117-1120 [PMID: 18712574 DOI: 10.1007/s11695-008-9662-0]
 - 42 **Mui WL,** Ng EK, Tsung BY, Lam CH, Yung MY. Impact on obesity-related illnesses and quality of life following intra-gastric balloon. *Obes Surg* 2010; **20**: 1128-1132 [PMID: 19015930 DOI: 10.1007/s11695-008-9766-6]
 - 43 **Lopez-Nava G,** Rubio MA, Prados S, Pastor G, Cruz MR, Companion E, Lopez A. BioEnterics® intra-gastric balloon (BIB®). Single ambulatory center Spanish experience with 714 consecutive patients treated with one or two consecutive balloons. *Obes Surg* 2011; **21**: 5-9 [PMID: 20306153 DOI: 10.1007/s11695-010-0093-3]
 - 44 **Kotzampassi K,** Grosomanidis V, Papakostas P, Penna S, Eleftheriadis E. 500 intra-gastric balloons: what happens 5 years thereafter? *Obes Surg* 2012; **22**: 896-903 [PMID: 22287051 DOI: 10.1007/s11695-012-0607-2]
 - 45 **Papavramidis TS,** Grosomanidis V, Papakostas P, Penna S, Kotzampassi K. Intra-gastric balloon fundal or antral position affects weight loss and tolerability. *Obes Surg* 2012; **22**: 904-909 [PMID: 22322378 DOI: 10.1007/s11695-012-0620-5]
 - 46 **Genco A,** Dellepiane D, Baglio G, Cappelletti F, Frangella F, Maselli R, Dante MC, Camoirano R, Lorenzo M, Basso N. Adjustable intra-gastric balloon vs non-adjustable intra-gastric balloon: case-control study on complications, tolerance, and efficacy. *Obes Surg* 2013; **23**: 953-958 [PMID: 23526067 DOI: 10.1007/s11695-013-0891-5]
 - 47 **Gümürdülü Y,** Doğan ÜB, Akın MS, Taşdoğan BE, Yalaki S. Long-term effectiveness of BioEnterics intra-gastric balloon in obese patients. *Turk J Gastroenterol* 2013; **24**: 387-391 [PMID: 24557961 DOI: 10.4318/tjg.2013.0696]
 - 48 **Fuller NR,** Lau NS, Denyer G, Caterson ID. An intra-gastric balloon produces large weight losses in the absence of a change in ghrelin or peptide YY. *Clin Obes* 2013; **3**: 172-179 [PMID: 25586733 DOI: 10.1111/cob.12030]
 - 49 **Totté E,** Hendrickx L, Pauwels M, Van Hee R. Weight reduction by means of intra-gastric device: experience with the bioenterics intra-gastric balloon. *Obes Surg* 2001; **11**: 519-523 [PMID: 11501367]
 - 50 **Sallet JA,** Marchesini JB, Paiva DS, Komoto K, Pizani CE, Ribeiro ML, Miguel P, Ferraz AM, Sallet PC. Brazilian multicenter study of the intra-gastric balloon. *Obes Surg* 2004; **14**: 991-998 [PMID: 15329191 DOI: 10.1381/0960892041719671]
 - 51 **Crea N,** Pata G, Della Casa D, Minelli L, Maifredi G, Di Betta E, Mitterpergher F. Improvement of metabolic syndrome following intra-gastric balloon: 1 year follow-up analysis. *Obes Surg* 2009; **19**: 1084-1088 [PMID: 19506981 DOI: 10.1007/s11695-009-9879-6]
 - 52 **Fuller NR,** Pearson S, Lau NS, Wlodarczyk J, Halstead MB, Tee HP, Chettiar R, Kaffes AJ. An intra-gastric balloon in the treatment of obese individuals with metabolic syndrome: a randomized controlled study. *Obesity (Silver Spring)* 2013; **21**: 1561-1570 [PMID: 23512773 DOI: 10.1002/oby.20414]
 - 53 **Al Kahtani K,** Khan MQ, Helmy A, Al Ashgar H, Rezeig M, Al Quaiz M, Kagevi I, Al Sofayan M, Al Fadda M. Bio-enteric intra-gastric balloon in obese patients: a retrospective analysis of King Faisal Specialist Hospital experience. *Obes Surg* 2010; **20**: 1219-1226 [PMID: 18752030 DOI: 10.1007/s11695-008-9654-0]
 - 54 **Genco A,** López-Nava G, Wahlen C, Maselli R, Cipriano M, Sanchez MM, Jacobs C, Lorenzo M. Multi-centre European experience with intra-gastric balloon in overweight populations: 13 years of experience. *Obes Surg* 2013; **23**: 515-521 [PMID: 23224509 DOI: 10.1007/s11695-012-0829-3]
 - 55 **Herve J,** Wahlen CH, Schaeken A, Dallemagne B, Dewandre JM, Markiewicz S, Monami B, Weerts J, Jehaes C. What becomes of patients one year after the intra-gastric balloon has been removed? *Obes Surg* 2005; **15**: 864-870 [PMID: 15978160 DOI: 10.1381/0960892054222894]
 - 56 **Tai CM,** Lin HY, Yen YC, Huang CK, Hsu WL, Huang YW, Chang CY, Wang HP, Mo LR. Effectiveness of intra-gastric balloon treatment for obese patients: one-year follow-up after balloon removal. *Obes Surg* 2013; **23**: 2068-2074 [PMID: 23832520 DOI: 10.1007/s11695-013-1027-7]
 - 57 **Dastis NS,** François E, Deviere J, Hittélet A, Ilah Mehdi A, Barea M, Dumonceau JM. Intra-gastric balloon for weight loss: results in 100 individuals followed for at least 2.5 years. *Endoscopy* 2009; **41**: 575-580 [PMID: 19588283 DOI: 10.1055/s-0029-1214826]
 - 58 **Gaur S,** Levy S, Mathus-Vliegen L, Chuttani R. Balancing risk and reward: a critical review of the intra-gastric balloon for weight loss. *Gastrointest Endosc* 2015; **81**: 1330-1336 [PMID: 25887720 DOI: 10.1016/j.gie.2015.01.054]
 - 59 **Dogan UB,** Gumurdulu Y, Akin MS, Yalaki S. Five percent weight lost in the first month of intra-gastric balloon treatment may be a predictor for long-term weight maintenance. *Obes Surg* 2013; **23**: 892-896 [PMID: 23404240 DOI: 10.1007/s11695-013-0876-4]
 - 60 **Lopez-Nava G,** Bautista-Castaño I, Jimenez-Baños A, Fernandez-Corbelle JP. Dual Intra-gastric Balloon: Single Ambulatory Center Spanish Experience with 60 Patients in Endoscopic Weight Loss Management. *Obes Surg* 2015; **25**: 2263-2267 [PMID: 25982804 DOI: 10.1007/s11695-015-1715-6]
 - 61 **Ponce J,** Quebbemann BB, Patterson EJ. Prospective, randomized, multicenter study evaluating safety and efficacy of intra-gastric dual-balloon in obesity. *Surg Obes Relat Dis* 2013; **9**: 290-295 [PMID: 22951075 DOI: 10.1016/j.soard.2012.07.007]
 - 62 **Ponce J,** Woodman G, Swain J, Wilson E, English W, Ikramuddin S, Bour E, Edmundowicz S, Snyder B, Soto F, Sullivan S, Holcomb R, Lehmann J. The REDUCE pivotal trial: a prospective, randomized controlled pivotal trial of a dual intra-gastric balloon for the treatment of obesity. *Surg Obes Relat Dis* 2015; **11**: 874-881 [PMID: 25868829 DOI: 10.1016/j.soard.2014.12.006]
 - 63 **Machytka E,** Klvana P, Kornbluth A, Peikin S, Mathus-Vliegen

- LE, Gostout C, Lopez-Nava G, Shikora S, Brooks J. Adjustable intra-gastric balloons: a 12-month pilot trial in endoscopic weight loss management. *Obes Surg* 2011; **21**: 1499-1507 [PMID: 21553304 DOI: 10.1007/s11695-011-0424-z]
- 64 **Brooks J**, Srivastava ED, Mathus-Vliegen EM. One-year adjustable intra-gastric balloons: results in 73 consecutive patients in the U.K. *Obes Surg* 2014; **24**: 813-819 [PMID: 24442419 DOI: 10.1007/s11695-014-1176-3]
- 65 **Carvalho GL**, Barros CB, Moraes CE, Okazaki M, Ferreira Mde N, Silva JS, de Albuquerque PP, Coelho Rde M. The use of an improved intra-gastric balloon technique to reduce weight in pre-obese patients--preliminary results. *Obes Surg* 2011; **21**: 924-927 [PMID: 19756895 DOI: 10.1007/s11695-009-9947-y]
- 66 **Mion F**, Ibrahim M, Marjoux S, Ponchon T, Dugardeyn S, Roman S, Deviere J. Swallowable Obalon® gastric balloons as an aid for weight loss: a pilot feasibility study. *Obes Surg* 2013; **23**: 730-733 [PMID: 23512445 DOI: 10.1007/s11695-013-0927-x]
- 67 **Farina MG**, Baratta R, Nigro A, Vinciguerra F, Puglisi C, Schembri R, Virgilio C, Vigneri R, Frittitta L. Intra-gastric balloon in association with lifestyle and/or pharmacotherapy in the long-term management of obesity. *Obes Surg* 2012; **22**: 565-571 [PMID: 21901285 DOI: 10.1007/s11695-011-0514-y]
- 68 **Imaz I**, Martínez-Cervell C, García-Alvarez EE, Sendra-Gutiérrez JM, González-Enríquez J. Safety and effectiveness of the intra-gastric balloon for obesity. A meta-analysis. *Obes Surg* 2008; **18**: 841-846 [PMID: 18459025 DOI: 10.1007/s11695-007-9331-8]
- 69 **Forlano R**, Ippolito AM, Iacobellis A, Merla A, Valvano MR, Niro G, Annese V, Andriulli A. Effect of the BioEnterics intra-gastric balloon on weight, insulin resistance, and liver steatosis in obese patients. *Gastrointest Endosc* 2010; **71**: 927-933 [PMID: 19863955 DOI: 10.1016/j.gie.2009.06.036]
- 70 **Angrisani L**, Lorenzo M, Borrelli V, Giuffrè M, Fonderico C, Capece G. Is bariatric surgery necessary after intra-gastric balloon treatment? *Obes Surg* 2006; **16**: 1135-1137 [PMID: 16989695 DOI: 10.1381/096089206778392365]
- 71 **Mathus-Vliegen EM**. Intra-gastric balloon treatment for obesity: what does it really offer? *Dig Dis* 2008; **26**: 40-44 [PMID: 18600014 DOI: 10.1159/000109385]
- 72 **Ganesh R**, Rao AD, Baladas HG, Leese T. The Bioenteric Intra-gastric Balloon (BIB) as a treatment for obesity: poor results in Asian patients. *Singapore Med J* 2007; **48**: 227-231 [PMID: 17342292]
- 73 **Younossi ZM**, Diehl AM, Ong JP. Nonalcoholic fatty liver disease: an agenda for clinical research. *Hepatology* 2002; **35**: 746-752 [PMID: 11915019 DOI: 10.1053/jhep.2002.32483]
- 74 **Lee YM**, Low HC, Lim LG, Dan YY, Aung MO, Cheng CL, Wee A, Lim SG, Ho KY. Intra-gastric balloon significantly improves nonalcoholic fatty liver disease activity score in obese patients with nonalcoholic steatohepatitis: a pilot study. *Gastrointest Endosc* 2012; **76**: 756-760 [PMID: 22840293 DOI: 10.1016/j.gie.2012.05.023]
- 75 **Ochner CN**, Kwok Y, Conceição E, Pantazatos SP, Puma LM, Carnell S, Teixeira J, Hirsch J, Geliebter A. Selective reduction in neural responses to high calorie foods following gastric bypass surgery. *Ann Surg* 2011; **253**: 502-507 [PMID: 21169809 DOI: 10.1097/SLA.0b013e318203a289]
- 76 **Wren AM**, Bloom SR. Gut hormones and appetite control. *Gastroenterology* 2007; **132**: 2116-2130 [PMID: 17498507 DOI: 10.1053/j.gastro.2007.03.048]
- 77 **Delzenne N**, Blundell J, Brouns F, Cunningham K, De Graaf K, Erkner A, Lluch A, Mars M, Peters HP, Westerterp-Plantenga M. Gastrointestinal targets of appetite regulation in humans. *Obes Rev* 2010; **11**: 234-250 [PMID: 20433660 DOI: 10.1111/j.1467-789X.2009.00707.x]
- 78 **Mion F**, Napoléon B, Roman S, Malvoisin E, Trepo F, Pujol B, Lefort C, Bory RM. Effects of intra-gastric balloon on gastric emptying and plasma ghrelin levels in non-morbid obese patients. *Obes Surg* 2005; **15**: 510-516 [PMID: 15946431 DOI: 10.1381/0960892053723411]
- 79 **Martinez-Brocca MA**, Belda O, Parejo J, Jimenez L, del Valle A, Pereira JL, Garcia-Pesquera F, Astorga R, Leal-Cerro A, Garcia-Luna PP. Intra-gastric balloon-induced satiety is not mediated by modification in fasting or postprandial plasma ghrelin levels in morbid obesity. *Obes Surg* 2007; **17**: 649-657 [PMID: 17658025]
- 80 **Greenway FL**. Physiological adaptations to weight loss and factors favouring weight regain. *Int J Obes (Lond)* 2015; **39**: 1188-1196 [PMID: 25896063 DOI: 10.1038/ijo.2015.59]
- 81 **Crujeiras AB**, Goyenechea E, Abete I, Lage M, Carreira MC, Martínez JA, Casanueva FF. Weight regain after a diet-induced loss is predicted by higher baseline leptin and lower ghrelin plasma levels. *J Clin Endocrinol Metab* 2010; **95**: 5037-5044 [PMID: 20719836 DOI: 10.1210/jc.2009-2566]
- 82 **Abu Dayyeh BK**, Edmundowicz SA, Jonnalagadda S, Kumar N, Larsen M, Sullivan S, Thompson CC, Banerjee S. Endoscopic bariatric therapies. *Gastrointest Endosc* 2015; **81**: 1073-1086 [PMID: 25828245 DOI: 10.1016/j.gie.2015.02.023]
- 83 **Kim SH**, Chun HJ. Endoscopic Treatment for Obesity: New Emerging Technology Trends. *Gut Liver* 2015; **9**: 431-432 [PMID: 26087857 DOI: 10.5009/gnl15125]
- 84 **Dai SC**, Paley M, Chandrasekhara V. Intra-gastric balloons: an introduction and removal technique for the endoscopist. *Gastrointest Endosc* 2015; **82**: 1122 [PMID: 26183825 DOI: 10.1016/j.gie.2015.06.008]
- 85 **Bužga M**, Evžen M, Pavel K, Tomáš K, Vladislava Z, Pavel Z, Svagera Z. Effects of the intra-gastric balloon MedSil on weight loss, fat tissue, lipid metabolism, and hormones involved in energy balance. *Obes Surg* 2014; **24**: 909-915 [PMID: 24488758 DOI: 10.1007/s11695-014-1191-4]
- 86 **Nikolic M**, Boban M, Ljubic N, Supanc V, Mirosevic G, Pezo Nikolic B, Krpan R, Posavec L, Zjajic-Rotkovic V, Bekavac-Beslin M, Gacina P. Morbidly obese are ghrelin and leptin hyporesponders with lesser intra-gastric balloon treatment efficiency: ghrelin and leptin changes in relation to obesity treatment. *Obes Surg* 2011; **21**: 1597-1604 [PMID: 21494811 DOI: 10.1007/s11695-011-0414-1]

P- Reviewer: Badiu C, Dogan UB, Farhat S, Shin JM
S- Editor: Ma YJ **L- Editor:** A **E- Editor:** Zhang DN



Role and mechanisms of action of *Escherichia coli* Nissle 1917 in the maintenance of remission in ulcerative colitis patients: An update

Franco Scaldaferrì, Viviana Gerardi, Francesca Mangiola, Loris Riccardo Lopetuso, Marco Pizzoferrato, Valentina Petito, Alfredo Papa, Jovana Stojanovic, Andrea Poscia, Giovanni Cammarota, Antonio Gasbarrini

Franco Scaldaferrì, Viviana Gerardi, Francesca Mangiola, Loris Riccardo Lopetuso, Marco Pizzoferrato, Valentina Petito, Alfredo Papa, Giovanni Cammarota, Antonio Gasbarrini, Polo Apparato digerente e sistema Endocrino Metabolico, Gastroenterology Division, Catholic University of Sacred Heart, 00168 Rome, Italy

Jovana Stojanovic, Andrea Poscia, Institute of Hygiene, Catholic University of Sacred Heart, 00168 Rome, Italy

Author contributions: Scaldaferrì F and Gerardi V equally contributed; Scaldaferrì F, Gerardi V and Gasbarrini A contributed to study design; Scaldaferrì F, Gerardi V, Mangiola F, Lopetuso LR, Pizzoferrato M, Petito V, Stojanovic J and Poscia A made literature search; Scaldaferrì F and Gerardi V made figures and wrote the paper; Stojanovic J and Poscia A performed the metanalysis; all authors contributed to data interpretation, editing.

Conflict-of-interest statement: Scaldaferrì F and Gasbarrini A were consultants for Ca.Digroup; however, each author has no financial interests or connections, direct or indirect, or other situations that might raise the question of bias in the work reported or the conclusions.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Franco Scaldaferrì, MD, PhD, Centre for the Research and Cure of Inflammatory Bowel Disease, UOC di Medicina Interna e Gastroenterologia, Institute of Medical Pathology, Catholic University of the Sacred Heart, Fondazione Policlinico "A. Gemelli" Hospital, Igo Gemelli 8, 00168 Roma, Italy. francoscaldaferrì@gmail.com
Telephone: +39-6-30155923
Fax: +39-6-30157249

Received: January 11, 2016
Peer-review started: January 13, 2016
First decision: February 18, 2016
Revised: March 12, 2016
Accepted: May 4, 2016
Article in press: May 4, 2016
Published online: June 28, 2016

Abstract

Ulcerative colitis (UC) is a chronic inflammatory disease, whose etiology is still unclear. Its pathogenesis involves an interaction between genetic factors, immune response and the "forgotten organ", Gut Microbiota. Several studies have been conducted to assess the role of antibiotics and probiotics as additional or alternative therapies for Ulcerative Colitis. *Escherichia coli* Nissle (EcN) is a nonpathogenic Gram-negative strain isolated in 1917 by Alfred Nissle and it is the active component of microbial drug Mutaflor® (Ardeypharm GmbH, Herdecke, Germany and EcN, Cadigroup, In Italy) used in many gastrointestinal disorder including diarrhea, uncomplicated diverticular disease and UC. It is the only probiotic recommended in ECCO guidelines as effective alternative to mesalazine in maintenance of remission in UC patients. In this review we propose an update on the role of EcN 1917 in maintenance of remission in UC patients, including data about efficacy and safety. Further studies may be helpful for this subject to further the full use of potential of EcN.

Key words: Ulcerative colitis; *Escherichia coli* Nissle; Metanalysis; Probiotic; Randomized trial; Inflammatory bowel disease

© The Author(s) 2016. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: *Escherichia coli* (*E. coli*) Nissle is a non-pathogenic Gram-negative strain used as a probiotic with very good quality paper assessing its bio-equivalence to mesalazine in maintaining remission in ulcerative colitis. Mechanisms of actions of this compound include immune-modulatory properties, reinforcement of intestinal barrier and inhibitory effect towards other pathogenic *E. coli*.

Scaldaferri F, Gerardi V, Mangiola F, Lopetuso LR, Pizzoferrato M, Petito V, Papa A, Stojanovic J, Poscia A, Cammarota G, Gasbarrini A. Role and mechanisms of action of *Escherichia coli* nissle 1917 in the maintenance of remission in ulcerative colitis patients: An update. *World J Gastroenterol* 2016; 22(24): 5505-5511 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5505.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5505>

INTRODUCTION

Ulcerative colitis (UC) is a chronic relapsing and relapsing disease, characterized by a continuous inflammation which can stretch from the rectum up to the entire colon, often resulting in mucosal ulceration, rectal bleeding, diarrhea, abdominal pain. Its etiology is still unclear, and it is multifactorial. Several factors have been identified as major determinants for induction or relapses and, among these, the imbalanced gut microbiota has become more crucial in recent years. The main hypothesis is that it is due to an excessive immune response to endogenous bacteria, in genetically predisposed individuals^[1].

The human microflora, known as "microbiota", includes more than thousand different species and higher than 15000 different bacterial strains, for an average total weight of 1 kg. In recent years several studies investigated the correlation between dysbiosis and intestinal and extra-intestinal diseases, including inflammatory bowel disease (IBD) and so UC^[2].

Probiotics are viable agents conferring benefits to the health of the human host^[3]. They can provide a beneficial effect on intestinal epithelial cells in numerous ways. Some strains can block pathogen entry into the epithelial cell by providing a physical barrier or by creating a mucus barrier; other probiotics maintain intestinal permeability acting on tight junctions. Some probiotic strains produce antimicrobial factors, other strains modulate the immune response^[4].

The role of microbiota in UC was supported by several evidences: inflammation is greatest in intestinal tracts with high concentration of bacteria, surgical reduction of the bacterial load is associated with improvement of inflammation and inflammation does not occur in germ free animals^[5].

The treatment goal of UC is the induction and the maintenance of remission. 5-aminosalicylic acid

(ASA) compounds, azathioprine/6 mercaptopurine, corticosteroids, cyclosporine, methotrexate, and anti-TNF α agents are conventional therapies used to control the disease. There are also other therapies, used as addition or alternative to conventional therapies in UC, in particular antibiotics and probiotics, which modulate gut microbiota.

Escherichia coli (*E. coli*) Nissle (EcN) 1917 is a non-pathogenic Gram-negative strain used in many gastrointestinal disorder including diarrhea^[6], uncomplicated diverticular disease^[7] and IBD, in particular UC^[8].

STRUCTURE AND MECHANISMS OF

ACTION OF EcN 1917

EcN 1917 (O6:K5:H1) was isolated by Prof. Alfred Nissle from Freiburg, Germany, in 1917 from the intestinal microflora of a young soldier. This soldier - unlike his comrades - did not develop infectious diarrhea, when stationed during World War I in Southeastern Europe (Dobrudja/Balkan peninsula), endemic for Shigella at that time. The strain was named *E. coli* strain Nissle 1917.

Using this *E. coli* strain Prof. Nissle developed the probiotic drug Mutaflor[®] and introduced it into medical practice in the same year. Since 1917, Mutaflor[®] is available in the German pharmaceutical market without interruption and recently also in Italy as EcN (cadigroup).

The lack of defined virulence factors (alpha-hemolysin, P-fimbrial adhesins, etc.) combined with the expression of fitness factors such as microcins, different iron uptake systems (enterobactin, yersiniabactin, aerobactin, salmochelin, ferric dicitrate transport system, and the *chu* heme transport locus), adhesins, and proteases may support its survival and efficacious colonization of the human gut, and contribute to the probiotic character of EcN 1917.

It exhibits a semi-rough lipopolysaccharide (LPS) phenotype and serum sensitivity and does not produce known toxins^[9]. EcN colonizes the intestine within few days and it remains as colonic flora for months after administration^[10].

EcN has an intestinal anti-inflammatory effect, but also systemic effects^[11], and there are many theories about its mechanism of action (Figure 1): (1) It has direct antimicrobial effects: it inhibits EHEC (*E. coli* EDL933) colonization in animal models^[12] and synthesis of Shiga-Toxins in co-cultivation experiment with STEC (Shiga-Toxin producing *E. coli*)^[13]. (2) It is involved in the bacterial-epithelial crosstalk ("Host cell signaling") by biofilm formation: it expresses F1C Fimbria. This is very important in the formation of biofilm, adherence to epithelial cells and persistence in infant mouse colonization^[14]. Its flagellum is the major "propulsor" *in vivo*, which allow this probiotic strain to efficiently compete with pathogens for binding sites on host tissue^[15]. It directly stimulates

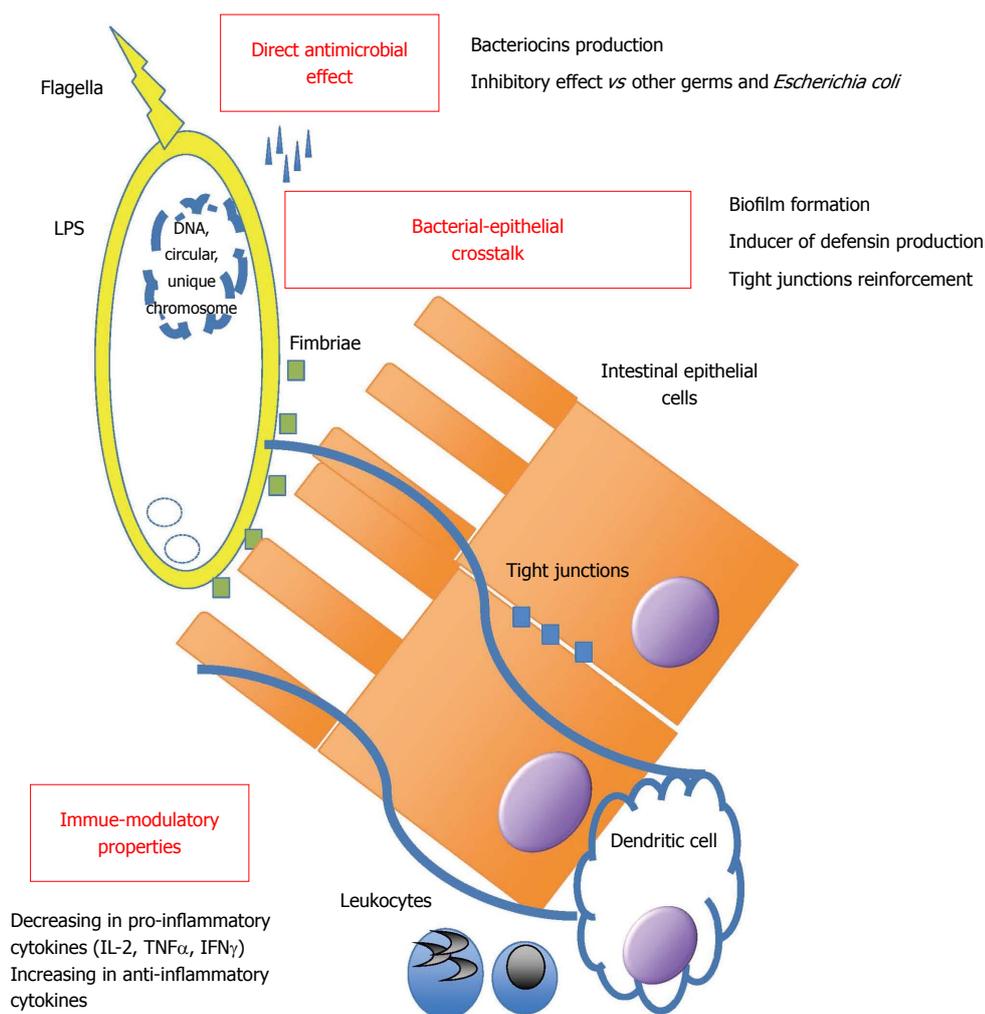


Figure 1 Structure and mechanisms of action of *Escherichia coli* Nissle 1917. LPS: Lipopolysaccharide; IL-2: Interleukin-2; TNF: Tumor necrosis factor; IFN: Interferon.

defensin production by intestinal epithelial cells, such as the human beta-defensin that inhibits adhesion and invasion of intestinal cells by pathogenic adherent invasive *E. coli*, which play a key role as trigger in immune response in IBD patients^[16-18]. It strengthens tight junctions of intestinal epithelial cells, by up-regulating the expression of the mRNA for the zonula occludens proteins ZO-1 and ZO-2, so it has an effect on the repair of the "leaky gut"^[19-21]. (3) It interacts with immune system by causing decreasing in pro-inflammatory cytokines (IL-2, TNF- α , IFN γ) and increasing in anti-inflammatory cytokines by peripheral blood mononuclear cells *in vitro*. It may reduce the expansion of newly recruited T cells into the intestinal mucosa and decrease intestinal inflammation, but it doesn't affect activated tissue-bound T cells, which may eliminate deleterious antigens in order to maintain immunological homeostasis^[22,23]. Furthermore EcN has a specific LPS that is responsible for its immunogenicity, without major immunotoxic properties at doses suggested and that provide, together with the other described features, a powerful effect on intestinal

immune function^[24].

Efficacy of EcN in animal models of colitis

Schultz *et al.*^[25] conducted a study on animal models of acute and chronic colitis. Acute colitis was induced by administration of dextran-sodium sulfate (DSS) in drinking water and chronic colitis was induced by transferin CD4⁺ CD62L⁺ T lymphocytes from BALB/c mice in SCID mice. These studies have shown that administration of EcN ameliorates intestinal inflammation (measured by histological scores) in chronic models but not in acute models, in accordance with clinical observations. Therefore, it was shown that EcN reduced secretion of pro-inflammatory cytokines, measured by enzyme-linked immune-sorbent assay^[25].

Grabig *et al.*^[5] demonstrated that EcN ameliorates experimental colitis induced by administration of 5% DSS in mice *via* TLR-2 and TLR-4 dependent pathway.

Decreasing of symptom scores and differences in body mass loss were shown in animal models of DSS colitis in BALB/c mice treated with EcN in the study conducted by Kokesová *et al.*^[26].

Table 1 Main results from trials on *Escherichia coli* Nissle on ulcerative colitis

Efficacy of EcN 1917 in maintenance of UC remission
Results from major randomized controlled clinical trials
EcN 200 mg/d is equivalent to Mesalazine 1000 mg/d in maintenance of UC remission ^[27]
EcN 400 mg/d is equivalent to Mesalazine 2400 mg/d in maintenance of UC remission following an acute flare ^[28]
EcN 200 mg/d is equivalent to Mesalazine 1500 mg/d in maintenance of UC remission ^[10]
Results from minor studies
Rectal administration of EcN 40 mL/d is effective in moderate distal active UC ^[30]
EcN 200 mg/d is equivalent to Mesalazine 1500 mg/d in maintenance of UC remission ^[6]

EcN: *Escherichia coli* Nissle; UC: Ulcerative colitis.

CLINICAL ROLE OF EcN 1917 IN MAINTANANCE OF UC

There are three major double-blind RCTs (Table 1), which compare EcN to mesalazine in prevention of relapse in UC patients, all of them designed to demonstrate equivalence of two treatment according to Schuirmann's two-one side test or "non inferiority trials".

The first trial was conducted by Kruis *et al.*^[27] in 1997. It was a randomized, double-blind, double-dummy study conducted on 120 out-patients in Germany, Czech Republic and Austria. Patients had a confirmed diagnosis of ulcerative colitis in remission. In particular patients had to be in remission for a maximum of 12 mo, with clinical activity index (CAI) < 4, no endoscopic or histological signs of acute inflammation. Each patient had to have had at least 2 relapses prior inclusion.

Patients received 500 mg mesalazine t.d.s. and a placebo form of EcN preparation or 200 mg/d of a preparation containing EcN in a single dose ("Mutaflor", Ardeypharm GmbH, Herdecke, Germany. 100 mg contains 25×10^9 viable *E. coli* bacteria) and a placebo form of mesalazine. The duration of the study was 12 wk. Study objectives included the assessment of the equivalence of the CAI under the two treatment modalities and the comparison of the relapse rates, relapse-free times and global assessment.

Study population was homogeneous into the two groups, with a prevalence in left sided colitis and small prevalence of active use of steroids (less than 25 % in both groups).

From the results of this study, no significant difference was observed between the two groups, although a low statistical power and a minor trend towards a slightly higher CAI in the EcN group. No serious adverse events reported for both groups.

The second trial was conducted by Rembacken *et al.*^[28] in 1999. This was a single-center, randomised, double-dummy study involving 116 patients, which

were treated with mesalazine 800 tds (Asacol formulation) or EcN 2 cp per 2 times daily ("Mutaflor", Ardeypharm GmbH, Herdecke, Germany. 100 mg contains 25×10^9 viable *E. coli* bacteria).

Inclusion criteria were: 18-80 years of age, clinical active (mild-moderate and severe) ulcerative colitis ("Leeds-Index") defined by number of 4 or more liquid stools/day for the last 7 d, with or without blood, erythema on sigmoidoscopy and histological confirmation of active ulcerative colitis.

The study populations were comparable. In particular: the median clinical activity index on study entry was 11 in the mesalazine group and nine in the EcN group (up to 30% of patients had a severe disease). The median sigmoidoscopy score was four in both groups. All patients received rectal or oral steroids at different doses together with a 1-wk course of oral gentamicin. At baseline both groups had an high active usage of steroids, around for 50% of the cases; furthermore, 2% in mesalazine group and 18% in EcN. In both groups the proportion between proctitis, left sided and pancolitis was similar (1/3 per each condition). No significant differences between the 2 groups were found.

Active treatment, started at the enrolment, was hydrocortisone enema twice daily, prednisolone 30 mg a day in moderate colitis and prednisolone 60 mg a day in severe colitis (according to Truelov Witts criteria). Only people in remission at 12 wk were enrolled in the follow up part of the trial assessing maintenance of remission.

Starting from remission, patients were maintained on either mesalazine or *E. coli* but the doses were reduced respectively at 1.2 g per day for the mesalazine group and 2 capsules per day for the EcN group. The follow up was at 12 mo. 59 were randomised to mesalazine and 57 to EcN. Of them 75% and 68% reached remission. Of those in remission, 73% of patients in the mesalazine group and 67% in the EcN group, relapsed by 12 mo. In the mesalazine group, the mean duration of remission was 206 d (median 175) compared with 221 (median 185) in the group given *E. coli*, ($P = 0.0174$). The treatment with EcN was proved safe, acceptable, and clinically equivalent to mesalazine in maintaining remission after an acute relapse of ulcerative colitis. Finally, this study is characterized by a very high rate of relapse: this is however not un-expected as the study population comprehends moderate and also severe patients.

The third trial was conducted by Kruis *et al.*^[10] in 2004. This was a double-blind, double dummy study in which 327 patients affected by ulcerative colitis in remission phase were recruited. 162 patients received Mutaflor 200 mg/d and 165 received Mesalazine 500 mg three times daily for 12 mo. Inclusion criteria were: age between 18-70 years, diagnosis of UC in remission [CAI ≤ 4 , endoscopic index (EI) ≤ 4 , and no signs of acute inflammation on histological examination]. Furthermore, within inclusion criteria there was at

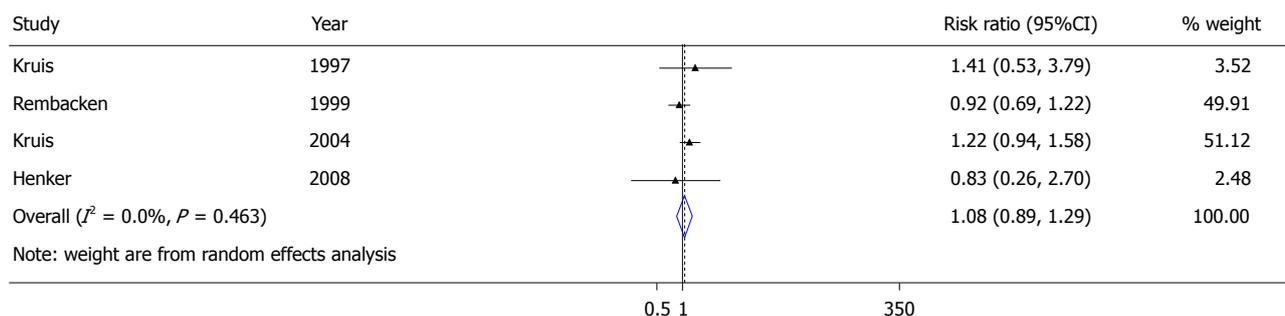


Figure 2 Metanalysis on randomized controlled trials assessing role of *Escherichia coli* Nissle on maintenance of remission in ulcerative colitis.

least two acute attacks of UC prior to the study and duration of the current remission of no longer than 12 mo. Primary objective of the study was meant to compare the number of patients experiencing a relapse during the 12 mo observation time between the two treatment groups. Secondary aims included efficacy variables like physician's and patient's assessment of general well being and calculation of a quality of life index. Additionally, time to relapse, CAI, EI, and histological findings were also evaluated. In the EcN group 36.4% of patients relapsed compared to 33.9% in the mesalazine group and statistical tests showed equivalence of the two treatments. A subgroup analyses showed no difference in terms of duration and localization of disease or pre-trial treatment. No difference of quality of life was shown in the two groups.

Overall same results on tolerance were found: it was very good or good in the EcN group in 80.0% and in the mesalazine group in 86.0%. According to the physician's assessment, the respective values were 85.1% and 90.3%. No unexpected drug reactions occurred during the study.

This is perhaps the best study based on the quality of data and also the large number of patients enrolled. Furthermore clinical outcomes were assessed by well-established endoscopic and histological activity indices, like in modern trials for more powerful drugs.

In addition to the above-described trials, there is a multicentric placebo-controlled study on 90 patients with moderate distal active UC conducted by Matthes *et al.*^[29] Patients in EcN groups received EcN 40, 20 or 10 mL (amount of bacteria 10E8/mL) enema once daily for at least 2 wk. A clinical DAI was recorded after 2, 4 and/or 8 wk.

The majority of patients also received concomitant medical treatment such as oral mesalazine. Remission rates and improvement of the histological score was showed particularly in the EcN 40 mL group, but further studies on largest population are required.

This study has shown that rectal administration of EcN is an effective treatment, with a dose dependent efficacy as shown in the Per Protocol analysis. Unfortunately the ITT analysis did not show significant results^[29].

Many meta-analysis present in the literature support the role of ECN in the therapy of ulcerative colitis^[6,10,27,28,30]. In particular, a very recent published meta-analysis, performed by Losurdo *et al.*^[31] (Figure 2), showed a non-significant inferiority of EcN in relapse prevention compared to mesalazine in preventing disease relapse, thus confirming current guideline recommendations^[31], despite a novel randomized double-blinded placebo controlled trial conducted in Denmark and published very recently^[32], with negative results. One hundred patients with active UC defined by CAI-score ≥ 6 and with calprotectin higher than 50 mg/kg, were enrolled and randomized into four groups of treatment: Ciprofloxacin (for 1 wk) followed by EcN (for 7 wk), Ciprofloxacin (for 7 wk) followed by placebo (for one week), placebo (for one week) followed by EcN (for 7 wk) and placebo (for one week) followed by placebo (for 7 wk). Aim of the study was the induction of the remission in ulcerative colitis and Kaplan-Meier curves were used to compare groups. In this study, the 54% of patients in the placebo/EcN group reached remission, compared to 89% of patients in the placebo/placebo group ($P < 0.05$), 78% of Ciprofloxacin/placebo group and 66% Ciprofloxacin/EcN group. Furthermore, the placebo/EcN group had the largest number of withdrawals. These impressive results, which would exclude a role of EcN in treatment of active ulcerative colitis, display several limitations, which make this study really weak. First of all this is a monocenter study, with a very not homogeneous population as showed in the table of patients characteristics. Mean CAI score at baseline was 10.5, 8.9, 9.3 and 8.9 in the Cipro/EcN, Cipro/placebo group, placebo/EcN group, placebo/placebo group, respectively. Furthermore patients clearly differed in concomitant medications use, in particular for use of active use of topical drugs and steroids as well as immunosuppressant. Taken together, these data suggest that this trial display major limitations regarding groups homogeneity. We confirmed the data from the published meta-analysis, which we performed independently before discovering that it was just published. In the present paper we report an extract of the recent published meta-analysis on the equivalence of the treatment between ECN and mesalazine^[31],

Table 2 Main potential clinical indications for *Escherichia coli* Nissle in gastroenterology

Maintenance of remission in ulcerative colitis ^[6,10,27,28]
Irritable bowel syndrome ^[34-37]
Constipation ^[38,39]
Acute diarrhea ^[6,40]
Collagenous colitis ^[41]
Uncomplicated diverticular disease ^[7]

starting from the major trials available. An equivalence between EcN and mesalamine on maintenance of remission in UC is still detectable (Figure 2).

Other studies assessing the use of EcN in ulcerative colitis

There is also an open-label multicenter pilot study that investigate the clinical benefit of EcN 1917 for maintenance therapy in young patients with UC. In this study 34 patients with UC in remission aged between 11 and 18 years were allocated either to EcN (2 capsules daily $n = 24$) or 5-ASA (median 1.5 g/d, $n = 10$), and observed over one year^[33]. Inclusion criteria were: 11-18 years of age, ulcerative colitis in remission for a maximum of 12 mo, at least 2 relapses prior inclusion, active therapy with mesalazine. Taking into account the low statistical power of the study, relapse rate was 25% (6/24) in the EcN group and 30% (3/10) in the 5-ASA group. Data on the patients' global health and development were favorable and no serious adverse events were reported^[33].

CONCLUSION

EcN is a well known probiotic, used in several countries for GI diseases (Table 2)^[6,10,27,28,34-41], registered as a drug in certain European countries, and it is the only one approved for maintenance of remission in UC patients by ECCO guidelines, based on data discussed also in the present paper. Trials designed to be non-inferiority/equivalence trials, comparing EcN to mesalazine, have reported equivalent rates of relapse between the two treatments, demonstrating that EcN is equivalent to mesalazine in the maintenance of remission in UC. Of the 3 major trials demonstrating these findings, the best and larger trial is the one conducted by Kruis *et al* and published on 2004. Finally, EcN showed a robust safe profile in UC patients. Further studies may be helpful to further dissect mechanisms of actions and perhaps optimize dose and newer indication of EcN.

REFERENCES

- 1 Sartor RB. Mechanisms of disease: pathogenesis of Crohn's disease and ulcerative colitis. *Nat Clin Pract Gastroenterol Hepatol* 2006; **3**: 390-407 [PMID: 16819502 DOI: 10.1038/ncpgasthep0528]
- 2 Scaldaferri F, Gerardi V, Lopetuso LR, Del Zompo F, Mangiola F, Bošković I, Bruno G, Petito V, Laterza L, Cammarota G, Gaetani

- E, Sgambato A, Gasbarrini A. Gut microbial flora, prebiotics, and probiotics in IBD: their current usage and utility. *Biomed Res Int* 2013; **2013**: 435268 [PMID: 23991417]
- 3 Reid G, Jass J, Sebulsky MT, McCormick JK. Potential uses of probiotics in clinical practice. *Clin Microbiol Rev* 2003; **16**: 658-672 [PMID: 14557292 DOI: 10.1128/CMR.16.4.658-672.2003]
- 4 Gareau MG, Sherman PM, Walker WA. Probiotics and the gut microbiota in intestinal health and disease. *Nat Rev Gastroenterol Hepatol* 2010; **7**: 503-514 [PMID: 20664519 DOI: 10.1038/nrgastro.2010.117]
- 5 Grabig A, Paclik D, Guzy C, Dankof A, Baumgart DC, Erckenbrecht J, Raupach B, Sonnenborn U, Eckert J, Schumann RR, Wiedenmann B, Dignass AU, Sturm A. *Escherichia coli* strain Nissle 1917 ameliorates experimental colitis via toll-like receptor 2- and toll-like receptor 4-dependent pathways. *Infect Immun* 2006; **74**: 4075-4082 [PMID: 16790781 DOI: 10.1128/IAI.01449-05]
- 6 Henker J, Laass MW, Blokhin BM, Maydannik VG, Bolbot YK, Elze M, Wolff C, Schreiner A, Schulze J. Probiotic *Escherichia coli* Nissle 1917 versus placebo for treating diarrhea of greater than 4 days duration in infants and toddlers. *Pediatr Infect Dis J* 2008; **27**: 494-499 [PMID: 18469732 DOI: 10.1097/INF.0b013e318169034c]
- 7 Fric P, Zavoral M. The effect of non-pathogenic *Escherichia coli* in symptomatic uncomplicated diverticular disease of the colon. *Eur J Gastroenterol Hepatol* 2003; **15**: 313-315 [PMID: 12610327 DOI: 10.1097/00042737-200303000-00015]
- 8 Schultz M. Clinical use of *E. coli* Nissle 1917 in inflammatory bowel disease. *Inflamm Bowel Dis* 2008; **14**: 1012-1018 [PMID: 18240278 DOI: 10.1002/ibd.20377]
- 9 Grozdanov L, Raasch C, Schulze J, Sonnenborn U, Gottschalk G, Hacker J, Dobrindt U. Analysis of the genome structure of the nonpathogenic probiotic *Escherichia coli* strain Nissle 1917. *J Bacteriol* 2004; **186**: 5432-5441 [PMID: 15292145 DOI: 10.1128/JB.186.16.5432-5441.2004]
- 10 Kruis W, Fric P, Pokrotnieks J, Lukás M, Fixa B, Kascák M, Kamm MA, Weismueller J, Beglinger C, Stolte M, Wolff C, Schulze J. Maintaining remission of ulcerative colitis with the probiotic *Escherichia coli* Nissle 1917 is as effective as with standard mesalazine. *Gut* 2004; **53**: 1617-1623 [PMID: 15479682 DOI: 10.1136/gut.2003.037747]
- 11 Arribas B, Rodríguez-Cabezas ME, Camuesco D, Comalada M, Bailón E, Utrilla P, Nieto A, Concha A, Zarzuelo A, Gálvez J. A probiotic strain of *Escherichia coli*, Nissle 1917, given orally exerts local and systemic anti-inflammatory effects in lipopolysaccharide-induced sepsis in mice. *Br J Pharmacol* 2009; **157**: 1024-1033 [PMID: 19486007 DOI: 10.1111/j.1476-5381.2009.00270.x]
- 12 Maltby R, Leatham-Jensen MP, Gibson T, Cohen PS, Conway T. Nutritional basis for colonization resistance by human commensal *Escherichia coli* strains HS and Nissle 1917 against *E. coli* O157: H7 in the mouse intestine. *PLoS One* 2013; **8**: e53957 [PMID: 23349773 DOI: 10.1371/journal.pone.0053957]
- 13 Reissbrodt R, Hammes WP, dal Bello F, Prager R, Fruth A, Hantke K, Rakin A, Starcic-Erjavec M, Williams PH. Inhibition of growth of Shiga toxin-producing *Escherichia coli* by nonpathogenic *Escherichia coli*. *FEMS Microbiol Lett* 2009; **290**: 62-69 [PMID: 19016876 DOI: 10.1111/j.1574-6968.2008.01405.x]
- 14 Lasaro MA, Salinger N, Zhang J, Wang Y, Zhong Z, Goulian M, Zhu J. F1C fimbriae play an important role in biofilm formation and intestinal colonization by the *Escherichia coli* commensal strain Nissle 1917. *Appl Environ Microbiol* 2009; **75**: 246-251 [PMID: 18997018 DOI: 10.1128/AEM.01144-08]
- 15 Troge A, Scheppach W, Schroeder BO, Rund SA, Heuner K, Wehkamp J, Stange EF, Oelschlaeger TA. More than a marine propeller--the flagellum of the probiotic *Escherichia coli* strain Nissle 1917 is the major adhesin mediating binding to human mucus. *Int J Med Microbiol* 2012; **302**: 304-314 [PMID: 23131416 DOI: 10.1016/j.ijmm.2012.09.004]
- 16 Boudeau J, Glasser AL, Julien S, Colombel JF, Darfeuille-Michaud A. Inhibitory effect of probiotic *Escherichia coli* strain Nissle 1917 on adhesion to and invasion of intestinal epithelial cells by adherent-invasive *E. coli* strains isolated from patients

- with Crohn's disease. *Aliment Pharmacol Ther* 2003; **18**: 45-56 [PMID: 12848625 DOI: 10.1046/j.1365-2036.2003.01638.x]
- 17 **Wehkamp J**, Harder J, Wehkamp K, Wehkamp-von Meissner B, Schlee M, Enders C, Sonnenborn U, Nuding S, Bengmark S, Fellermann K, Schröder JM, Stange EF. NF-kappaB- and AP-1-mediated induction of human beta defensin-2 in intestinal epithelial cells by Escherichia coli Nissle 1917: a novel effect of a probiotic bacterium. *Infect Immun* 2004; **72**: 5750-5758 [PMID: 15385474 DOI: 10.1128/IAI.72.10.5750-5758.2004]
 - 18 **Schlee M**, Wehkamp J, Altenhoefer A, Oelschlaeger TA, Stange EF, Fellermann K. Induction of human beta-defensin 2 by the probiotic Escherichia coli Nissle 1917 is mediated through flagellin. *Infect Immun* 2007; **75**: 2399-2407 [PMID: 17283097 DOI: 10.1128/IAI.01563-06]
 - 19 **Zyrek AA**, Cichon C, Helms S, Enders C, Sonnenborn U, Schmidt MA. Molecular mechanisms underlying the probiotic effects of Escherichia coli Nissle 1917 involve ZO-2 and PKCzeta redistribution resulting in tight junction and epithelial barrier repair. *Cell Microbiol* 2007; **9**: 804-816 [PMID: 17087734 DOI: 10.1111/j.1462-5822.2006.00836.x]
 - 20 **Otte JM**, Podolsky DK. Functional modulation of enterocytes by gram-positive and gram-negative microorganisms. *Am J Physiol Gastrointest Liver Physiol* 2004; **286**: G613-G626 [PMID: 15010363 DOI: 10.1152/ajpgi.00341.2003]
 - 21 **Ukena SN**, Singh A, Dringenberg U, Engelhardt R, Seidler U, Hansen W, Bleich A, Bruder D, Franzke A, Rogler G, Suerbaum S, Buer J, Gunzer F, Westendorf AM. Probiotic Escherichia coli Nissle 1917 inhibits leaky gut by enhancing mucosal integrity. *PLoS One* 2007; **2**: e1308 [PMID: 18074031 DOI: 10.1371/journal.pone.0001308]
 - 22 **Helwig U**, Lammers KM, Rizzello F, Brigidi P, Rohleder V, Caramelli E, Gionchetti P, Schrezenmeir J, Foelsch UR, Schreiber S, Campieri M. Lactobacilli, bifidobacteria and E. coli nissle induce pro- and anti-inflammatory cytokines in peripheral blood mononuclear cells. *World J Gastroenterol* 2006; **12**: 5978-5986 [PMID: 17009396 DOI: 10.3748/wjg.v12.i37.5978]
 - 23 **Sturm A**, Rilling K, Baumgart DC, Gargas K, Abou-Ghazalé T, Raupach B, Eckert J, Schumann RR, Enders C, Sonnenborn U, Wiedenmann B, Dignass AU. Escherichia coli Nissle 1917 distinctively modulates T-cell cycling and expansion via toll-like receptor 2 signaling. *Infect Immun* 2005; **73**: 1452-1465 [PMID: 15731043 DOI: 10.1128/IAI.73.3.1452-1465.2005]
 - 24 **Grozdánov L**, Zähringer U, Blum-Oehler G, Brade L, Henne A, Knirel YA, Schombel U, Schulze J, Sonnenborn U, Gottschalk G, Hacker J, Rietschel ET, Dobrindt U. A single nucleotide exchange in the wzy gene is responsible for the semirough O6 lipopolysaccharide phenotype and serum sensitivity of Escherichia coli strain Nissle 1917. *J Bacteriol* 2002; **184**: 5912-5925 [PMID: 12374825 DOI: 10.1128/JB.184.21.5912-5925.2002]
 - 25 **Schultz M**, Strauch UG, Linde HJ, Watzl S, Obermeier F, Göttl C, Dunger N, Grunwald N, Schölmerich J, Rath HC. Preventive effects of Escherichia coli strain Nissle 1917 on acute and chronic intestinal inflammation in two different murine models of colitis. *Clin Diagn Lab Immunol* 2004; **11**: 372-378 [PMID: 15013990 DOI: 10.1128/cdli.11.2.372-378.2004]
 - 26 **Kokesová A**, Frolová L, Kverka M, Sokol D, Rossmann P, Bártová J, Tlaskalová-Hogenová H. Oral administration of probiotic bacteria (E. coli Nissle, E. coli O83, Lactobacillus casei) influences the severity of dextran sodium sulfate-induced colitis in BALB/c mice. *Folia Microbiol (Praha)* 2006; **51**: 478-484 [PMID: 17176771 DOI: 10.1007/BF02931595]
 - 27 **Kruis W**, Schütz E, Fric P, Fixa B, Judmaier G, Stolte M. Double-blind comparison of an oral Escherichia coli preparation and mesalazine in maintaining remission of ulcerative colitis. *Aliment Pharmacol Ther* 1997; **11**: 853-858 [PMID: 9354192 DOI: 10.1046/j.1365-2036.1997.00225.x]
 - 28 **Rembacken BJ**, Snelling AM, Hawkey PM, Chalmers DM, Axon AT. Non-pathogenic Escherichia coli versus mesalazine for the treatment of ulcerative colitis: a randomised trial. *Lancet* 1999; **354**: 635-639 [PMID: 10466665 DOI: 10.1016/S0140-6736(98)06343-0]
 - 29 **Matthes H**, Krummnerl T, Giensch M, Wolff C, Schulze J. Clinical trial: probiotic treatment of acute distal ulcerative colitis with rectally administered Escherichia coli Nissle 1917 (EcN). *BMC Complement Altern Med* 2010; **10**: 13 [PMID: 20398311 DOI: 10.1186/1472-6882-10-13]
 - 30 **Jonkers D**, Penders J, Masclee A, Pierik M. Probiotics in the management of inflammatory bowel disease: a systematic review of intervention studies in adult patients. *Drugs* 2012; **72**: 803-823 [PMID: 22512365 DOI: 10.2165/11632710-000000000-00000]
 - 31 **Losurdo G**, Iannone A, Contaldo A, Ierardi E, Di Leo A, Principi M. Escherichia coli Nissle 1917 in Ulcerative Colitis Treatment: Systematic Review and Meta-analysis. *J Gastrointest Liver Dis* 2015; **24**: 499-505 [PMID: 26697577]
 - 32 **Petersen AM**, Mirsepasi H, Halkjær SI, Mortensen EM, Nordgaard-Lassen I, Krogfelt KA. Ciprofloxacin and probiotic Escherichia coli Nissle add-on treatment in active ulcerative colitis: a double-blind randomized placebo controlled clinical trial. *J Crohns Colitis* 2014; **8**: 1498-1505 [PMID: 24972748 DOI: 10.1016/j.crohns.2014.06.001]
 - 33 **Henker J**, Müller S, Laass MW, Schreiner A, Schulze J. Probiotic Escherichia coli Nissle 1917 (EcN) for successful remission maintenance of ulcerative colitis in children and adolescents: an open-label pilot study. *Z Gastroenterol* 2008; **46**: 874-875 [PMID: 18810672 DOI: 10.1055/s-2008-1027463]
 - 34 **Kruis W**, Chrubasik S, Boehm S, Stange C, Schulze J. A double-blind placebo-controlled trial to study therapeutic effects of probiotic Escherichia coli Nissle 1917 in subgroups of patients with irritable bowel syndrome. *Int J Colorectal Dis* 2012; **27**: 467-474 [PMID: 22130826 DOI: 10.1007/s00384-011-1363-9]
 - 35 **Róžańska D**, Regulska-Iłow B, Choroszy-Król I, Iłow R. [The role of Escherichia coli strain Nissle 1917 in the gastro-intestinal diseases]. *Postepy Hig Med Dosw (Online)* 2014; **68**: 1251-1256 [PMID: 25380207 DOI: 10.5604/17322693.1127882]
 - 36 **Plassmann D**, Schulte-Witte H. [Treatment of irritable bowel syndrome with Escherichia coli strain Nissle 1917 (EcN): a retrospective survey]. *Med Klin (Munich)* 2007; **102**: 888-892 [PMID: 17992479]
 - 37 **Krammer HJ**, Kämper H, von Büнау R, Zieseniss E, Stange C, Schlieger F, Clever I, Schulze J. [Probiotic drug therapy with E. coli strain Nissle 1917 (EcN): results of a prospective study of the records of 3,807 patients]. *Z Gastroenterol* 2006; **44**: 651-656 [PMID: 16902895 DOI: 10.1055/s-2006-926909]
 - 38 **Behnen J**, Deriu E, Sassone-Corsi M, Raffatelli M. Probiotics: properties, examples, and specific applications. *Cold Spring Harb Perspect Med* 2013; **3**: a010074 [PMID: 23457295 DOI: 10.1101/cshperspect.a010074]
 - 39 **Möllenbrink M**, Bruckschen E. [Treatment of chronic constipation with physiologic Escherichia coli bacteria. Results of a clinical study of the effectiveness and tolerance of microbiological therapy with the E. coli Nissle 1917 strain (Mutaflor)]. *Med Klin (Munich)* 1994; **89**: 587-593 [PMID: 7815986]
 - 40 **Henker J**, Laass M, Blokhin BM, Bolbot YK, Maydannik VG, Elze M, Wolff C, Schulze J. The probiotic Escherichia coli strain Nissle 1917 (EcN) stops acute diarrhoea in infants and toddlers. *Eur J Pediatr* 2007; **166**: 311-318 [PMID: 17287932 DOI: 10.1007/s00431-007-0419-x]
 - 41 **Tromm A**, Niewerth U, Khoury M, Baestlein E, Wilhelms G, Schulze J, Stolte M. The probiotic E. coli strain Nissle 1917 for the treatment of collagenous colitis: first results of an open-label trial. *Z Gastroenterol* 2004; **42**: 365-369 [PMID: 15136935 DOI: 10.1055/s-2004-812709]

P- Reviewer: Manguso F, Marie JC, Naito Y **S- Editor:** Gong ZM

L- Editor: A **E- Editor:** Ma S



Basic Study

Transient receptor potential vanilloid 4-dependent calcium influx and ATP release in mouse and rat gastric epithelia

Hiroshi Mihara, Nobuhiro Suzuki, Ammar Abdulkader Boudaka, Jibrán Sualeh Muhammad, Makoto Tominaga, Yoshiaki Tabuchi, Toshiro Sugiyama

Hiroshi Mihara, Nobuhiro Suzuki, Jibrán Sualeh Muhammad, Toshiro Sugiyama, Department of Gastroenterology, Graduate School of Medicine and Pharmaceutical Sciences, University of Toyama, Toyama 930-0194, Japan

Hiroshi Mihara, Ammar Abdulkader Boudaka, Makoto Tominaga, Division of Cell Signaling, Okazaki Institute for Integrative Bioscience (National Institute for Physiological Sciences), National Institutes of Natural Sciences, Okazaki 444-8787, Japan

Ammar Abdulkader Boudaka, Department of Physiology, College of Medicine and Health Sciences, Sultan Qaboos University, Muscat 123, Sultanate of Oman

Yoshiaki Tabuchi, Life Science Research Center, University of Toyama, Toyama 930-0194, Japan

Author contributions: Mihara H and Sugiyama T designed study concept; Mihara H, Suzuki N, Boudaka AA, Muhammad JS and Tabuchi Y acquired data; Mihara H and Suzuki N analyzed Data; Mihara H, Suzuki N, Tominaga M and Sugiyama T contributed to data interpretation; Mihara H drafted the manuscript; and all authors critically revised the manuscript.

Supported by Grants from the University of Toyama and JSPS KAKENHI to Mihara H, No. 26870214.

Institutional animal care and use committee statement: All procedures involving the care and use of animals were approved by The Institutional Animal Care and Use Committee of the National Institutes of Natural Sciences.

Conflict-of-interest statement: No conflict of interest exists.

Data sharing statement: There are no additional data available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on

different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Hiroshi Mihara, MD, PhD, Assistant Professor, Department of Gastroenterology, Graduate School of Medicine and Pharmaceutical Sciences, University of Toyama, 2630 Sugitani, Toyama 930-0194, Japan. m164-tym@umin.net
Telephone: +81-76-4347301
Fax: +81-76-4345072

Received: February 16, 2016
Peer-review started: February 17, 2016
First decision: March 31, 2016
Revised: April 11, 2016
Accepted: May 4, 2016
Article in press: May 4, 2016
Published online: June 28, 2016

Abstract

AIM: To explore the expression of transient receptor potential vanilloid 4 (TRPV4) and its physiological meaning in mouse and rat gastric epithelia.

METHODS: RT-PCR and immunochemistry were used to detect TRPV4 mRNA and protein expression in mouse stomach and a rat normal gastric epithelial cell line (RGE1-01), while Ca²⁺-imaging and electrophysiology were used to evaluate TRPV4 channel activity. ATP release was measured by a luciferin-luciferase assay. Gastric emptying was also compared between WT and TRPV4 knockout mice.

RESULTS: TRPV4 mRNA and protein were detected in mouse tissues and RGE1-01 cells. A TRPV4-specific agonist (GSK1016790A) increased intracellular Ca²⁺ concentrations and/or evoked TRPV4-like current activities in WT mouse gastric epithelial cells and

RGE1-01 cells, but not TRPV4KO cells. GSK1016790A or mechanical stimuli induced ATP release from RGE1-01 cells while TRPV4 knockout mice displayed delayed gastric emptying *in vivo*.

CONCLUSION: TRPV4 is expressed in mouse and rat gastric epithelium and contributes to ATP release and gastric emptying.

Key words: Transient receptor potential vanilloid 4; Stomach; Gastric emptying; ATP

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: A mechano-sensitive ion channel, transient receptor potential vanilloid 4 (TRPV4), is expressed in gastric epithelium and contributes to ATP release and gastric emptying. These findings suggest that gastric distension stimulates TRPV4 on gastric epithelium and released ATP stimulates sub-epithelial nerve fibers or acts on visceral smooth muscles. TRPV4 might be a promising novel diagnostic and therapeutic target for functional gastric disorders.

Mihara H, Suzuki N, Boudaka AA, Muhammad JS, Tominaga M, Tabuchi Y, Sugiyama T. Transient receptor potential vanilloid 4-dependent calcium influx and ATP release in mouse and rat gastric epithelia. *World J Gastroenterol* 2016; 22(24): 5512-5519 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5512.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5512>

INTRODUCTION

The transient receptor potential vanilloid 4 channel (TRPV4) is a non-selective cation channel that is involved in various cellular functions^[1] and is activated by several physical and chemical stimuli, including mechanical stimuli, endogenous arachidonic acid metabolites (epoxyeicosatrienoic acids)^[2], and heat. TRPV4 is also activated by the specific agonist GSK1016790A that elicits whole-cell currents in mouse and rat TRPV4-expressing cells with EC₅₀ values of 18.5 and 10 nmol/L, respectively^[3]. TRPV4 is widely expressed throughout the body, including gastrointestinal tract epithelium and the esophagus^[4]. Although the physiological function of TRPV4 expression in intestinal epithelial cells is unknown, TRPV4 activation in these cells causes increases in intracellular calcium concentrations, chemokine release, and incidence of colitis^[5], as well as increased paracellular permeability^[6]. Furthermore, TRPV4 antagonists are promising therapeutic options for colitis^[7,8]. However, TRPV4 expression in the gastric epithelium awaits evaluation.

In addition to its function as an intracellular energy donor, ATP is recognized as an important signaling molecule that mediates diverse biological effects *via*

cell surface receptors: the purinergic receptors^[9]. ATP is released by neurons of the central, peripheral, and enteric nervous system^[10,11], and acts as a non-adrenergic non-cholinergic (NANC) neurotransmitter that causes different responses or effects (either excitatory or inhibitory depending on the P2 receptor subtype upon which they act as well as the animal species under study). Several studies showed that purinergic neurotransmission (assuming that gut neurons are the sole source of released ATP) affects gastric motility^[12]. Recent reports showed that ATP is also released from non-neuronal tissues and has an effect on tissue function. Moreover, we found that ATP release in the esophagus and urothelium was mediated by TRPV4 stimulation^[4,13,14]. However, there are no data concerning whether TRPV4 is expressed in the stomach and, if so, whether TRPV4 stimulation plays a role in mediating ATP release. Therefore, this study explored the morphological (RT-PCR and immunostaining) and functional (Ca²⁺-imaging, patch clamp and gastric emptying) expression of TRPV4 in mouse and rat stomach with special focus on gastric epithelium.

MATERIALS AND METHODS

Animals

Eight week-old male C57BL/6Ncr (SLC) and TRPV4-knockout (TRPV4KO) mice^[15] weighing between 23 and 25 g were housed in a controlled environment (12-h light/12-h dark cycle; room temperature, 22-24 °C; 50%-60% relative humidity) with free access to food and water. All procedures involving the care and use of animals were approved by The Institutional Animal Care and Use Committee of the National Institutes of Natural Sciences.

Cell lines

RGE1-01 is an immortalized rat gastric mucosal cell line that shows distinct cell differentiation types and preserves some epithelial cell characteristics. RGE1-01 cells were maintained at 34 °C in Dulbecco's modified Eagle medium supplemented with 10% heat-inactivated fetal bovine serum, 100 µg/mL streptomycin and 100 U/mL penicillin with the addition of ITES (see reference^[16] for details).

Acute isolated mouse gastric epithelium

WT and TRPV4KO mice were sacrificed by cervical dislocation. The stomachs were washed in cold (4 °C) PBS (-) and then incubated in trypsin solution (Invitrogen) at 4 °C for 1 h. Gastric epithelial cells were harvested and plated on CELL-TAK (BD Biosciences)-coated glass cover slips and used for Ca²⁺-imaging and patch clamp experiments.

Reverse transcription PCR analysis

RT-PCR was performed as previously described^[4,17]. Total RNA (1 µg) was isolated using the RNeasy Mini

Table 1 Primer sequences for RT-PCR

Primer name	Sequence (5'→3')
mTRPV4-F	ACAACACCCGAGAGAACACC
mTRPV4-R	CCCAAACCTTACGCCACTTGT
mGAPDH-F	TGAAGGGTGGAGCCAAAAGG
mGAPDH-R	GGAAGAGTGGGAGTTGCTGTTC
rTRPV4-F	CCTGGCAGGGATCGAGGCCT
rTRPV4-R	GGATGGTGGTGGCCCACTGC
rGAPDH-F	GCCAAGGCTGTGGCAAGGT
rGAPDH-R	GAGCAATGCCAGCCCCAGCA

Kit (Qiagen, Courtaboeuf, France) and measured with a NanoDrop device (Thermo Fisher Scientific Inc., Wilmington, United States). Genomic DNA was eliminated in the process of reverse transcription (QuantiTect Reverse Transcription Kit, QIAGEN). PCR was performed using rTaq DNA polymerase (TaKaRa) in an iCycler (Bio-Rad) with specific primer sets (Table 1).

Immunocytochemistry

Immunocytochemistry was performed as previously described^[4] using the antibodies summarized in Table 2. For section preparation, mouse stomachs were fixed at 4 °C for 6 h. Tissues were placed in PBS-sucrose and embedded in OCT compound (Tissue Tek, Elkhart, IN, United States). Non-specific antibody binding was reduced by incubation in BlockAce (Yukijirushi, Sapporo, Japan) for 1 h at room temperature prior to antibody exposure. Preparations were analyzed using a confocal laser scanning microscope (LSM 700, Carl Zeiss). For immunocytochemistry, RGE1-01 cells were fixed at 4 °C for 20 min with the same fixative. Bovine serum albumin (3% BSA; Sigma) was used as a blocking solution.

Ca²⁺-imaging

Fura-2 fluorescence was measured in primary mouse gastric epithelial cells and RGE1-01 cells with a standard bath solution containing 140 mmol/L NaCl, 5 mmol/L KCl, 2 mmol/L MgCl₂, 2 mmol/L CaCl₂, 10 mmol/L HEPES, and 10 mmol/L glucose at pH 7.4 (adjusted with NaOH) at 25 °C. Results are presented as ratios of fluorescence intensities obtained with fura-2 emissions at 340 nm and 380 nm. GSK1016790A^[3] and ionomycin (both from Sigma) were used as a TRPV4 agonist and a positive control, respectively. F_{340}/F_{380} was calculated and acquired with an image processing system (IP-Lab, Scanalytics Inc., Rockville, MD or AQUA COSMOS, Hamamatsu Co., Japan) and ImageJ software (<http://rsb.info.nih.gov/ij/>). Changes in ratio (Δ) were calculated by subtracting the mean basal values from peak values. Since the degree of responses (strong, weak, or no response) to GSK varied from cell to cell in WT gastric epithelial cells, we expressed the observed changes in all ionomycin-responsive cells as a ratio between WT and TRPV4 knockout mice (Δ). We evaluated 53 and 41 ionomycin-responsive cells from six WT mice

Table 2 Primary and secondary antisera for immunochemistry

Tissue antigen/host	Dilution	Source
TRPV4/rabbit	1:500	B. Nilius, or Abcam
Goat anti-rabbit IgG-Alexa488	1:1500	Invitrogen, Inc.

and five TRPV4 knockout mice, respectively. Given the variations in response times, and that some WT cells responded 30 s after GSK application, we decided to incubate cells with GSK for 90 s.

Electrophysiology

The standard bath solution was the same as that used in the Ca²⁺-imaging experiments. Pipette solutions for whole-cell recordings contained 140 mmol/L KCl, 5 mmol/L EGTA and 10 mmol/L HEPES, pH 7.4. Whole-cell recording data on primary gastric epithelial cells three hours after insolation and RGE1-01 were sampled at 10 kHz and filtered at 5 kHz for analysis (Axon 200B amplifier with pCLAMP software, Molecular Devices, Foster City, CA, United States). Voltage ramp-pulses from -100 mV to +100 mV (500 ms) were applied every 5 s to generate an I-V curve.

ATP release measurement

ATP concentrations released from RGE1-01 rat gastric epithelial cells cultured in 12-well plates or stretch silicon chambers (STB-10-04 from STREX Inc., Osaka, Japan) were measured by a luciferin-luciferase assay (ATP Bioluminescence assay kit CLS II, Roche Diagnostics) and a luminometer (Lumat LB 9507, Berthold Technologies, Japan), using a previously described method that was slightly modified^[4]. For chemical stimuli, cells cultured to 70%-80% confluence and incubated in 500 μ L bath solution for 30 min at room temperature (25 °C) were used to measure basal ATP release. The superfusate was collected and replaced gently with another 500 μ L of bath solution with or without the TRPV4 agonists GSK1016790A or 5,6-EET. The superfusate was collected after 15 min and the ratio of released ATP (15 min stimulation/30 min basal condition) was calculated. An aliquot (200 μ L) of superfusate was then mixed with 200 μ L luciferin-luciferase reagent for luminometric ATP measurements. For mechanical stimuli, stretching was quantitatively applied with a STB-10 stretch machine (STREX Inc.) to RGE1-01 cells cultured on a silicon chamber. Three minutes after chamber placement in the stretch machine, the superfusate was washed away to exclude artificial ATP release and replaced with 500 μ L of bath solution for basal ATP measurement. After a further 3 min, the superfusate was collected and replaced, whereupon mechanical stimuli was applied for 3 min and the ratio of released ATP (3 min stimulation/3 min basal condition) was calculated. To block TRPV4 channels, cells were pre-treated with the specific TRPV4 antagonist HC 067047 (Sigma, 1 μ mol/L) for 3 min^[18].

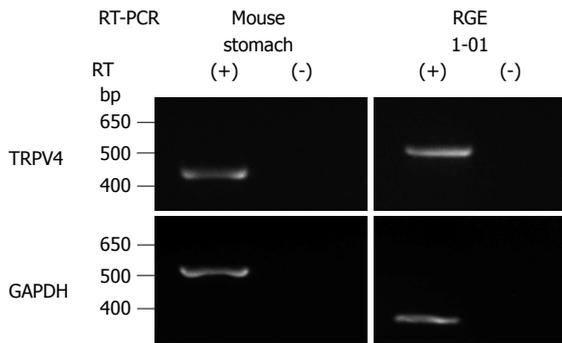


Figure 1 *TRPV4* mRNA expression in mouse stomach and RGE1-01 cells. *TRPV4* and *GAPDH* mRNA levels were examined with (+) and without (-) RT reaction. The expected sizes of the amplified fragments for *TRPV4* and *GAPDH* were 404 and 545 bp for mouse and 524 and 268 bp for rat, respectively. *TRPV4* mRNA was detected in mouse stomach and RGE1-01 cells (RGE1-01: normal rat gastric epithelial cell line). Band positions differed due to the use of different primers.

Gastric emptying

Eight-week-old WT and TRPV4KO male mice were used with a modified version of previously reported methods^[17]. Gastric emptying was determined by transit of a test meal containing phenol red. Mice were fasted for 14 h with *ad libitum* water before the experiment. Five mg/kg (200 μ L) of the test meal was administered into the stomach using a feeding needle. Fifteen minutes later, the mice were euthanized by cervical dislocation and the gastrointestinal tract was removed. The stomach was minced and the remaining phenol red concentration was measured. Gastric emptying was expressed as mean \pm SEM for each group.

Data analysis

Values for Ca^{2+} -imaging, patch-clamp experiments, ATP measurements, and gastric emptying are presented as mean \pm SEM from three or more independent experiments. A Student's *t*-test or non-parametric Bonferroni-type multiple comparison was used. Significance was accepted for $P < 0.05$.

RESULTS

TRPV4 expression in mouse and rat gastric epithelia

Given that *TRPV4* was shown to be expressed in the esophagus and intestinal epithelia^[4-6,19,20], we examined *TRPV4* mRNA expression in mouse stomach and a rat gastric epithelial cell line, RGE1-01. *TRPV4* mRNA was detected in mouse stomach and RGE1-01 cells (Figure 1). We next examined *TRPV4* protein expression in mouse and the RGE1-01 cells. A strong homogenous immunofluorescent signal was confined to the epithelial cell layer of the WT mouse gastric corpus and antrum but not in cells from TRPV4KO mice (Figure 2A). Meanwhile, Z-stack images obtained by confocal microscopy of RGE1-01 cells displayed apical

TRPV4 expression (Figure 2B).

TRPV4-mediated increase in cytosolic Ca^{2+} ($[Ca^{2+}]_i$) in mouse primary gastric epithelial cells

To confirm functional *TRPV4* expression in primary gastric epithelial cells and RGE1-01 cells, we examined the response to the reported specific *TRPV4* agonist, GSK1016790A (GSK)^[3], using a fluorescent Ca^{2+} -imaging system (10 μ mol/L, fura-2/AM). Response traces of $[Ca^{2+}]_i$ for WT and *TRPV4*KO gastric epithelial cells in the presence of GSK (100 nmol/L) showed that almost all cells isolated from WT stomach responded to GSK (Figure 3A) and the $[Ca^{2+}]_i$ increases were significantly larger in WT cells compared to *TRPV4*KO cells (Figure 3B). This finding suggests that the majority of gastric epithelial cells expressed *TRPV4* and $[Ca^{2+}]_i$ responses to GSK were *TRPV4* specific.

TRPV4-mediated current responses in mouse primary gastric epithelial cells and RGE1-01 cells

We next performed patch-clamp experiments with acute isolated mouse gastric epithelial cells in the presence of GSK (300 nmol/L) and observed inward current responses with an outwardly rectifying IV-relationship in WT but not *TRPV4*KO cells (Figure 4A)^[3]. Current responses were observed in all 5 trials with WT gastric epithelial cells but were completely absent with *TRPV4*KO cells, which indicates that the majority of gastric epithelial cells expressed *TRPV4*. Similar chemical stimulation with GSK (300 nmol/L) induced *TRPV4*-like current responses in the rat gastric epithelial cell line, RGE1-01 (Figure 4B). These data strongly indicated functional expression of *TRPV4* in mouse and rat gastric epithelial cells.

TRPV4 activators induced ATP release from RGE1-01 cells

Mechanical stimuli reportedly activate *TRPV4* expressed in esophageal keratinocytes that in turn leads to increased ATP release^[4]. To examine whether *TRPV4* stimulation has a similar effect in gastric epithelium, we measured ATP release in chemically- or mechanically-stimulated RGE1-01 cells using a luciferin-luciferase assay. *TRPV4* agonists GSK1016790A (GSK, 100 nmol/L) or 5,6-EET (500 nmol/L)^[2] significantly increased ATP release in RGE1-01 cells (Figure 5A, 2- to 3-fold higher vs control, $P < 0.05$). Additionally, 120% lateral stretch applied for 3 min to RGE1-01 cells cultured on a silicon chamber induced significantly higher amounts of ATP release (Figure 5B, about 2-fold vs without stretch (control), $P < 0.05$), and these responses were inhibited by the specific *TRPV4* inhibitor HC 067047 (1 μ mol/L) (stretched cells showed no detachment over the 3-min stretch period). These results suggested that chemical and mechanical stimuli-induced ATP release in RGE1-01 cells was mediated by *TRPV4* channel activation.

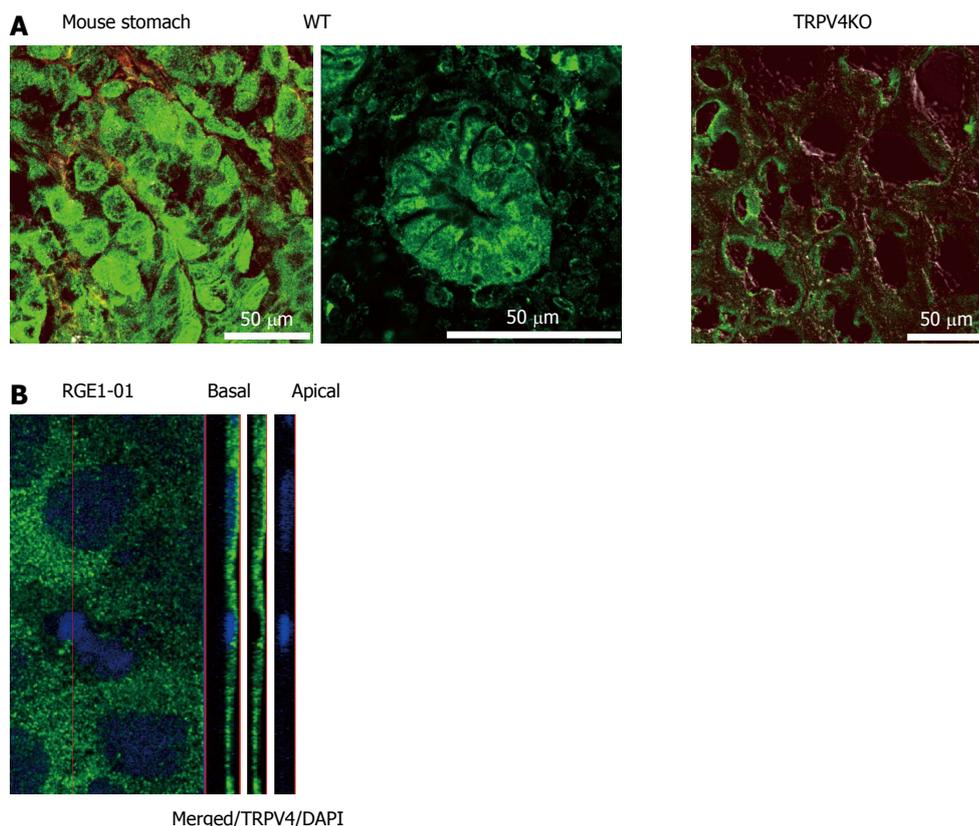


Figure 2 TRPV4 protein expression in mouse stomach and RGE1-01 cells. A: TRPV4 expression was homogeneously observed in WT but not TRPV4KO mouse gastric epithelium. Bars indicate 50 μm; B: Z-stack image of RGE1-01 cells demonstrated apical TRPV4 expression.

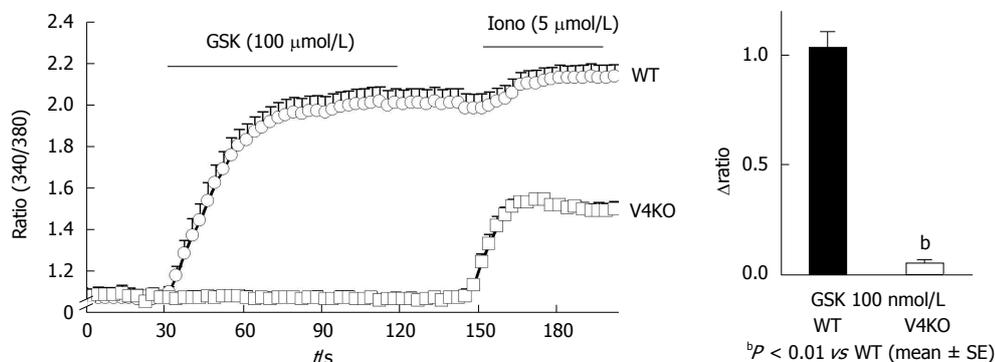


Figure 3 TRPV4-mediated increases in cytosolic Ca^{2+} ($[Ca^{2+}]_i$) in mouse primary gastric epithelial cells. A: $[Ca^{2+}]_i$ changes (340/380 ratio) in response to the TRPV4 specific agonist GSK1016790A (GSK, 100 nmol/L) in WT or TRPV4KO (V4KO) primary gastric epithelial cells (mean \pm SEM). Ionomycin (iono, 5 μmol/L) was used as a positive control. Bars indicate the period of chemical application; B: GSK significantly increased $[Ca^{2+}]_i$ in WT cells (means \pm SD; 1.03 \pm 0.07, $n = 20$) compared to TRPV4KO cells (0.05 \pm 0.01, $n = 20$) ($^bP < 0.01$ vs WT).

Delayed gastric emptying in TRPV4KO mice

Since TRPV4 has been shown to sense chemical and mechanical stimuli and contribute to ATP release from gastric epithelial cells, we hypothesized that TRPV4KO mice would exhibit altered gastric motility. To evaluate the physiological role of TRPV4 expressed in the gastric epithelium, we performed an *in vivo* experiment to compare gastric emptying rates of WT and TRPV4KO mice. Gastric emptying rates in TRPV4KO mice were about 2/3 of those in WT (Figure 6), suggesting that TRPV4 contributes to gastric motor function.

DISCUSSION

We identified morphological and functional TRPV4 expression in mouse gastric epithelial cells as well as the rat gastric epithelial cell line RGE1-01 (Figures 1-4). Furthermore, we demonstrated that chemical and mechanical stimuli can induce TRPV4-dependent ATP release from RGE1-01 cells (Figure 5), and that stimulation of gastric epithelial TRPV4 enhances gastric emptying *in vivo* (Figure 6). Using immunohistochemistry, Ca^{2+} imaging,

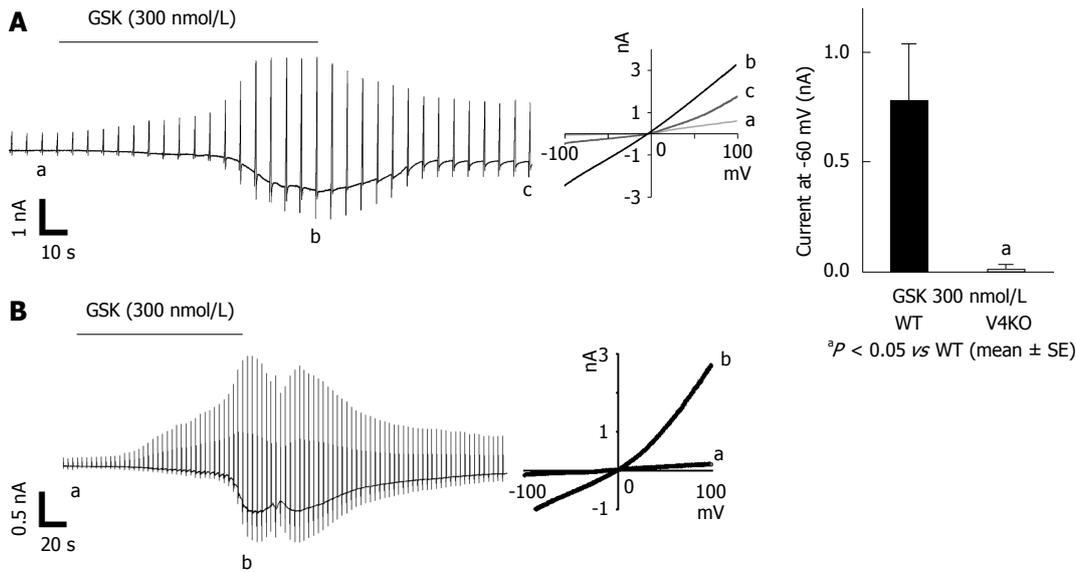


Figure 4 TRPV4-mediated current responses in mouse primary gastric epithelial cells and RGE1-01 cells. A: GSK (300 nmol/L) evoked inward current responses in WT primary gastric epithelial cells. Currents in response to ramp-pulses at points a, b and c (left in panel B) are shown (middle), with a strongly outwardly rectifying current-voltage relationship. Significantly larger inward currents at -60 mV were obtained from WT cells (means ± SEM; 0.76 ± 0.27 nA, $n = 5$) than in TRPV4KO cells (0.01 ± 0.00 nA, $n = 5$) (^a*P* < 0.05 vs WT). B: Similar current responses were also obtained in RGE1-01 cells.

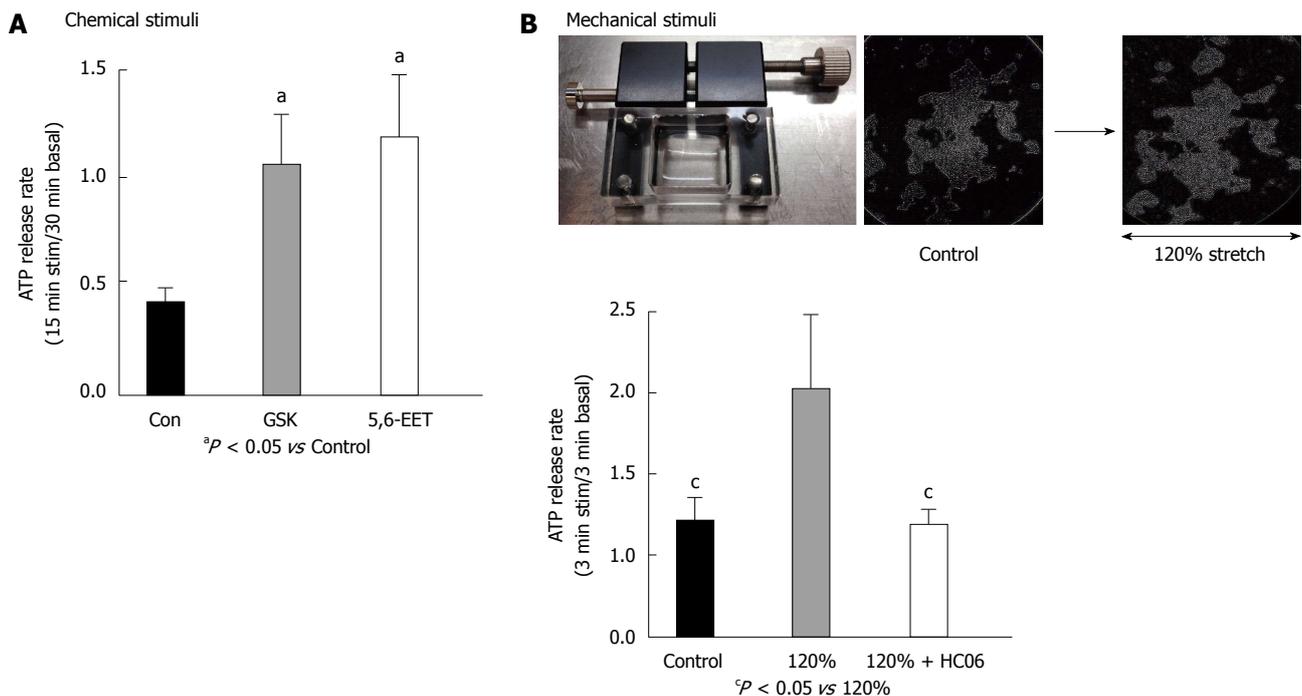


Figure 5 TRPV4 activator-induced ATP release in the RGE1-01 cells. A: GSK1016790A (GSK, 100 nmol/L) or 5,6-EET (500 nmol/L) induced significantly higher ATP release in RGE1-01 cells (^a*P* < 0.05 vs Control). B: Mechanical stimuli were quantitatively applied with a stretch apparatus. Microscopy images demonstrated that cells were stretched laterally without detachment. A 120% stretch induced significantly higher amounts of ATP release from RGE1-01 cells [^c*P* < 0.05 vs 0% stretch (control)] that could be inhibited by pre-treatment with specific TRPV4 antagonist HC 067047 (1 μ mol/L). TRPV4: Transient receptor potential vanilloid 4.

and electrophysiology, we found that the majority of mouse gastric epithelial cells exhibited abundant TRPV4 expression and responded to TRPV4 agonists, suggesting that TRPV4 is a candidate mechanoreceptor in gastric epithelial cells. In fact, ATP release was several hundred nmol/L in our *in vitro* study, suggesting that the corresponding ATP concentration *in vivo* might be estimated to be several μ mol/L, which

would be sufficient to activate the P2 receptor present in the wall of mouse stomach^[21,22]. These results suggested the hypothesis that luminal distension stimulates TRPV4 on gastric epithelial cells that in turn release ATP. The released ATP either stimulates sub-epithelial sensory nerve fibers that form the afferent limb of short or long gastrointestinal reflex arcs or acts directly on visceral smooth muscles expressing

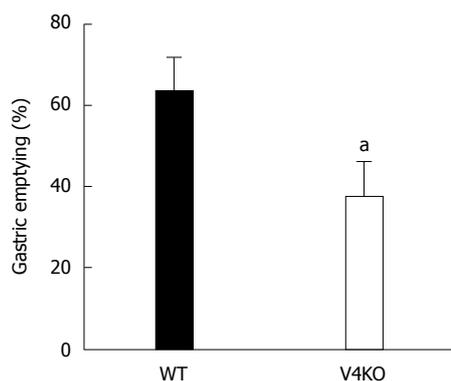


Figure 6 Delayed gastric emptying in TRPV4 knockout mice. Gastric emptying rates *in vivo* in TRPV4KO mice were significantly delayed relative to WT mice (^a $P < 0.05$, vs WT, $n = 7-9$).

purinergic receptors. The first possibility is supported by morphological evidence showing that purinergic receptors, mainly P2X2 and P2X3, were identified on putative gastric mechanosensing structures, including the vagal afferent intraganglionic laminar endings that are located in close proximity to the epithelium^[23,24]. These vagal afferents form the afferent limb of the central vago-vagal reflex (long reflex arc) and are known to increase gastric motility following stimulation^[25]. The hypothesis is also supported by findings from a previous study wherein P2X3-knockout mice show a blunted neural response to gastric distension and no differences in distension-evoked ATP release between knockout and control mice^[26]. The released ATP could also trigger a local reflex arc intrinsic to the stomach wall, which is supported by results from a previous study wherein ATP was shown to induce tetrodotoxin-sensitive contraction responses mediated by neuronal P2X receptors in an *in vitro* whole-stomach preparation^[27].

The possibility that ATP released from gastric epithelium could directly stimulate purinergic receptors expressed on gastric visceral smooth muscles is rather unlikely considering the short half-life of ATP and the distance that ATP must cross while diffusing from the gastric epithelium to visceral smooth muscles. Moreover, gastric smooth muscles are known to functionally express P2Y receptors that mediate relaxation in response to ATP^[27] and would be expected, upon stimulation, to delay gastric emptying, which is opposite to our current findings. This outcome further decreases the likelihood of a direct effect for ATP released from gastric epithelium on smooth muscles. However, the purinergic signaling pathway that mediates gastric distension-induced epithelial TRPV4 stimulation requires further future characterization.

In conclusion, TRPV4 is morphologically and functionally expressed in mouse and rat gastric epithelia and contributes to ATP release and gastric emptying. Our results suggest that TRPV4 could be a promising novel diagnostic and therapeutic target for functional gastrointestinal disorders.

ACKNOWLEDGMENTS

We thank Nilius B (Katholieke Universiteit, Belgium) for his generous gift of antibodies and Uchida K (NIPS) and Kozawa T (U. Toyama) for their technical assistance.

COMMENTS

Background

The transient receptor potential vanilloid 4 channel (TRPV4) is a non-selective cation channel that is activated by mechanical stimuli. ATP has been recognized as an important signaling molecule via cell surface ATP receptors. This study explored TRPV4 expression in mouse and rat stomach and whether the stimulation mediated ATP release.

Research frontiers

The authors have reported that ATP release in the esophagus and urothelium is mediated by TRPV4 stimulation.

Innovations and breakthroughs

This is the first study showing TRPV4 expression and ATP release by its stimulation in the mouse stomach and/or rat gastric epithelial cells.

Applications

These data suggested the hypothesis that luminal distension stimulates TRPV4 on gastric epithelial cells that in turn release ATP. However TRPV4 expression in human gastric epithelium and the purinergic signaling pathway requires further evaluation.

Terminology

TRPV4 is a non-selective cation channel, that is activated by several physical and chemical stimuli, including mechanical stimuli. The channel activation increases intracellular Ca^{2+} concentration and elicits whole-cell currents in TRPV4-expressing cells.

Peer-review

The manuscript describes an original research performed in mouse stomach and in rat epithelium cell line focusing on the expression and function of TRPV4 ion channels. The study concept is based on previous findings of the authors regarding TRPV4-mediated ATP release on the esophagus. The series of experiments in this manuscript demonstrated delayed gastric emptying in TRPV4 knockout mice compared to their WT littermates, as well as TRPV4 expression in mRNA and protein levels in both mouse and rat gastric epithelial cell line. In general, the idea is interesting, the various morphological and functional methods are sophisticated, the figures are demonstrative.

REFERENCES

- 1 **Everaerts W**, Nilius B, Owsianik G. The vanilloid transient receptor potential channel TRPV4: from structure to disease. *Prog Biophys Mol Biol* 2010; **103**: 2-17 [PMID: 19835908 DOI: 10.1016/j.pbiomolbio.2009.10.002]
- 2 **Watanabe H**, Vriens J, Prenen J, Droogmans G, Voets T, Nilius B. Anandamide and arachidonic acid use epoxyeicosatrienoic acids to activate TRPV4 channels. *Nature* 2003; **424**: 434-438 [PMID: 12879072 DOI: 10.1038/nature01807]
- 3 **Willette RN**, Bao W, Nerurkar S, Yue TL, Doe CP, Stankus G, Turner GH, Ju H, Thomas H, Fishman CE, Sulpizio A, Behm DJ, Hoffman S, Lin Z, Lozinskaya I, Casillas LN, Lin M, Trout RE, Votta BJ, Thorneloe K, Lashinger ES, Figueroa DJ, Marquis R, Xu X. Systemic activation of the transient receptor potential vanilloid subtype 4 channel causes endothelial failure and circulatory collapse: Part 2. *J Pharmacol Exp Ther* 2008; **326**: 443-452 [PMID: 18499744 DOI: 10.1124/jpet.107.134551]
- 4 **Mihara H**, Boudaka A, Sugiyama T, Moriyama Y, Tominaga M.

- Transient receptor potential vanilloid 4 (TRPV4)-dependent calcium influx and ATP release in mouse oesophageal keratinocytes. *J Physiol* 2011; **589**: 3471-3482 [PMID: 21540339 DOI: 10.1113/jphysiol.2011.207829]
- 5 **D'Aldebert E**, Cenac N, Rousset P, Martin L, Rolland C, Chapman K, Selves J, Alric L, Vinel JP, Vergnolle N. Transient receptor potential vanilloid 4 activated inflammatory signals by intestinal epithelial cells and colitis in mice. *Gastroenterology* 2011; **140**: 275-285 [PMID: 20888819 DOI: 10.1053/j.gastro.2010.09.045]
 - 6 **Yamawaki H**, Mihara H, Suzuki N, Nishizono H, Uchida K, Watanabe S, Tominaga M, Sugiyama T. Role of transient receptor potential vanilloid 4 activation in indomethacin-induced intestinal damage. *Am J Physiol Gastrointest Liver Physiol* 2014; **307**: G33-G40 [PMID: 24789205 DOI: 10.1152/ajpgi.00105.2013]
 - 7 **Fichna J**, Mokrowiecka A, Cygankiewicz AI, Zakrzewski PK, Malecka-Panas E, Janecka A, Krajewska WM, Storr MA. Transient receptor potential vanilloid 4 blockade protects against experimental colitis in mice: a new strategy for inflammatory bowel diseases treatment? *Neurogastroenterol Motil* 2012; **24**: e557-e560 [PMID: 22882778 DOI: 10.1111/j.1365-2982.2012.01999.x]
 - 8 **Vergnolle N**. TRPV4: new therapeutic target for inflammatory bowel diseases. *Biochem Pharmacol* 2014; **89**: 157-161 [PMID: 24440740 DOI: 10.1016/j.bcp.2014.01.005]
 - 9 **Ralevic V**, Burnstock G. Receptors for purines and pyrimidines. *Pharmacol Rev* 1998; **50**: 413-492 [PMID: 9755289]
 - 10 **Bertrand PP**. ATP and sensory transduction in the enteric nervous system. *Neuroscientist* 2003; **9**: 243-260 [PMID: 12934708 DOI: 10.1177/1073858403253768]
 - 11 **Galligan JJ**. Pharmacology of synaptic transmission in the enteric nervous system. *Curr Opin Pharmacol* 2002; **2**: 623-629 [PMID: 12482723 DOI: 10.1016/S1471-4892(02)00212-6]
 - 12 **Glasgow I**, Mattar K, Krantis A. Rat gastroduodenal motility in vivo: involvement of NO and ATP in spontaneous motor activity. *Am J Physiol* 1998; **275**: G889-G896 [PMID: 9815016]
 - 13 **Suzuki N**, Mihara H, Nishizono H, Tominaga M, Sugiyama T. Protease-Activated Receptor-2 Up-Regulates Transient Receptor Potential Vanilloid 4 Function in Mouse Esophageal Keratinocyte. *Dig Dis Sci* 2015; **60**: 3570-3578 [PMID: 26233549 DOI: 10.1007/s10620-015-3822-6]
 - 14 **Yoshiyama M**, Mochizuki T, Nakagomi H, Miyamoto T, Kira S, Mizumachi R, Sokabe T, Takayama Y, Tominaga M, Takeda M. Functional roles of TRPV1 and TRPV4 in control of lower urinary tract activity: dual analysis of behavior and reflex during the micturition cycle. *Am J Physiol Renal Physiol* 2015; **308**: F1128-F1134 [PMID: 25761879 DOI: 10.1152/ajprenal.00016.2015]
 - 15 **Mizuno A**, Matsumoto N, Imai M, Suzuki M. Impaired osmotic sensation in mice lacking TRPV4. *Am J Physiol Cell Physiol* 2003; **285**: C96-101 [PMID: 12777254 DOI: 10.1152/ajpcell.00559.2002]
 - 16 **Tabuchi Y**, Arai Y, Ohta S, Shioya H, Takahashi R, Ueda M, Takeguchi N, Asano S, Obinata M. Development and characterization of conditionally immortalized gastric epithelial cell lines from transgenic rats harboring temperature-sensitive simian virus 40 large T-antigen gene. *Cell Struct Funct* 2002; **27**: 71-79 [PMID: 12207048]
 - 17 **Mihara H**, Suzuki N, Yamawaki H, Tominaga M, Sugiyama T. TRPV2 ion channels expressed in inhibitory motor neurons of gastric myenteric plexus contribute to gastric adaptive relaxation and gastric emptying in mice. *Am J Physiol Gastrointest Liver Physiol* 2013; **304**: G235-G240 [PMID: 23203157 DOI: 10.1152/ajpgi.00256.2012]
 - 18 **Everaerts W**, Zhen X, Ghosh D, Vriens J, Gevaert T, Gilbert JP, Hayward NJ, McNamara CR, Xue F, Moran MM, Strassmaier T, Uykul E, Owsianik G, Vennekens R, De Ridder D, Nilius B, Fanger CM, Voets T. Inhibition of the cation channel TRPV4 improves bladder function in mice and rats with cyclophosphamide-induced cystitis. *Proc Natl Acad Sci USA* 2010; **107**: 19084-19089 [PMID: 20956320 DOI: 10.1073/pnas.1005333107]
 - 19 **Mizuno H**, Suzuki Y, Watanabe M, Sokabe T, Yamamoto T, Hattori R, Gotoh M, Tominaga M. Potential role of transient receptor potential (TRP) channels in bladder cancer cells. *J Physiol Sci* 2014; **64**: 305-314 [PMID: 24849279 DOI: 10.1007/s12576-014-0319-6]
 - 20 **Liedtke W**, Zhang JY, Hall RP, Steinhoff M. Keratinocyte growth regulation TRP-ed up over downregulated TRPV4? *J Invest Dermatol* 2014; **134**: 2310-2312 [PMID: 25120148 DOI: 10.1038/jid.2014.250]
 - 21 **North RA**. Molecular physiology of P2X receptors. *Physiol Rev* 2002; **82**: 1013-1067 [PMID: 12270951 DOI: 10.1152/physrev.00015.2002]
 - 22 **Coddou C**, Yan Z, Obsil T, Huidobro-Toro JP, Stojilkovic SS. Activation and regulation of purinergic P2X receptor channels. *Pharmacol Rev* 2011; **63**: 641-683 [PMID: 21737531 DOI: 10.1124/pr.110.003129]
 - 23 **Castelucci P**, Robbins HL, Furness JB. P2X(2) purine receptor immunoreactivity of intraganglionic laminar endings in the mouse gastrointestinal tract. *Cell Tissue Res* 2003; **312**: 167-174 [PMID: 12690440 DOI: 10.1007/s00441-003-0715-3]
 - 24 **Wang ZJ**, Neuhuber WL. Intraganglionic laminar endings in the rat esophagus contain purinergic P2X2 and P2X3 receptor immunoreactivity. *Anat Embryol (Berl)* 2003; **207**: 363-371 [PMID: 14624359 DOI: 10.1007/s00429-003-0351-4]
 - 25 **Travagli RA**, Hermann GE, Browning KN, Rogers RC. Musings on the wanderer: what's new in our understanding of vago-vagal reflexes? III. Activity-dependent plasticity in vago-vagal reflexes controlling the stomach. *Am J Physiol Gastrointest Liver Physiol* 2003; **284**: G180-G187 [PMID: 12529266 DOI: 10.1152/ajpgi.00413.2002]
 - 26 **McIlwrath SL**, Davis BM, Bielefeldt K. Deletion of P2X3 receptors blunts gastro-oesophageal sensation in mice. *Neurogastroenterol Motil* 2009; **21**: 890-e66 [PMID: 19368663 DOI: 10.1111/j.1365-2982.2009.01292.x]
 - 27 **Mulè F**, Naccari D, Serio R. Evidence for the presence of P2y and P2x receptors with different functions in mouse stomach. *Eur J Pharmacol* 2005; **513**: 135-140 [PMID: 15878718 DOI: 10.1016/j.ejphar.2005.01.052]

P- Reviewer: Cseko K S- Editor: Gong ZM L- Editor: A
E- Editor: Wang CH



Basic Study

Intravoxel incoherent motion diffusion-weighted imaging for monitoring chemotherapeutic efficacy in gastric cancer

Xiao-Li Song, Heoung Keun Kang, Gwang Woo Jeong, Kyu Youn Ahn, Yong Yeon Jeong, Yang Joon Kang, Hye Jung Cho, Chung Man Moon

Xiao-Li Song, Heoung Keun Kang, Yong Yeon Jeong, Yang Joon Kang, Department of Radiology, Chonnam National University Medical School, Chonnam National University Hwasun Hospital, Hwasun, Jeollanam-do 519-763, South Korea

Gwang Woo Jeong, Chung Man Moon, Department of Radiology, Chonnam National University Medical School, Chonnam National University Hospital, Gwangju 501-757, South Korea

Kyu Youn Ahn, Hye Jung Cho, Department of Anatomy, Chonnam National University Medical School, Gwangju 501-746, South Korea

Author contributions: Song XL and Jeong YY designed the studies; Song XL, Kang YJ and Moon CM performed the majority of experiments; Song XL, Ahn KY, Kang YJ and Cho HJ contributed to the analysis and interpretation of imaging data and histological examination; Song XL wrote the first draft of the manuscript; Jeong GW and Kang HK have approved the final manuscript and completed manuscript; Also, all authors agree with the content of the manuscript.

Supported by National Research Foundation of South Korea, No. NRF-2013R1A1A2013878 and No. 2015R1A2A2A01007827.

Institutional animal care and use committee statement: All procedures involving animals were reviewed and approved by the Institutional Animal Care and Use Committee of the Chonnam National University [CNU IACUC-H-2015-41].

Animal care and use statement: The animal protocol was designed to minimize pain or discomfort to the animals according to Institutional Animal Care and Use Committee Guidelines. All mice, fed with a standard diet with water ad libitum, were maintained at appropriate laboratory conditions (a photoperiod of 12 h light and darkness, 50% humidity, 23 °C). After MRI examination, mice were euthanized by overdose of isoflurane (over than 5%) for tissue collection.

Conflict-of-interest statement: The authors declared that they have no conflicts of interest to this work.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Heoung Keun Kang, MD, Professor of Medicine, Department of Radiology, Chonnam National University Medical School, Chonnam National University Hwasun Hospital, Hwasun, Jeollanam-do 519-763, South Korea. hkkang@jnu.ac.kr
Telephone: +82-61-3797101
Fax: +82-61-3797133

Received: February 24, 2016
Peer-review started: February 25, 2016
First decision: March 31, 2016
Revised: April 12, 2016
Accepted: April 20, 2016
Article in press: April 20, 2016
Published online: June 28, 2016

Abstract

AIM: To assess intravoxel incoherent motion diffusion-weighted imaging (IVIM-DWI) for monitoring early efficacy of chemotherapy in a human gastric cancer mouse model.

METHODS: IVIM-DWI was performed with 12 *b*-values (0-800 s/mm²) in 25 human gastric cancer-bearing nude mice at baseline (day 0), and then they were randomly divided into control and 1-, 3-, 5- and 7-d treatment groups (*n* = 5 per group). The control group underwent longitudinal MRI scans at days 1, 3, 5 and 7, and the treatment groups underwent subsequent MRI scans after a specified 5-fluorouracil/calcium

folinate treatment. Together with tumor volumes (TV), the apparent diffusion coefficient (ADC) and IVIM parameters [true water molecular diffusion coefficient (D), perfusion fraction (f) and pseudo-related diffusion coefficient (D^*)] were measured. The differences in those parameters from baseline to each measurement ($\Delta TV\%$, $\Delta ADC\%$, $\Delta D\%$, $\Delta f\%$ and $\Delta D^*\%$) were calculated. After image acquisition, tumor necrosis, microvessel density (MVD) and cellular apoptosis were evaluated by hematoxylin-eosin (HE), CD31 and terminal-deoxynucleotidyl transferase mediated nick end labeling (TUNEL) staining respectively, to confirm the imaging findings. Mann-Whitney test and Spearman's correlation coefficient analysis were performed.

RESULTS: The observed relative volume increase ($\Delta TV\%$) in the treatment group were significantly smaller than those in the control group at day 5 ($\Delta TV_{\text{treatment}}\% = 19.63\% \pm 3.01\%$ and $\Delta TV_{\text{control}}\% = 83.60\% \pm 14.87\%$, $P = 0.008$) and day 7 ($\Delta TV_{\text{treatment}}\% = 29.07\% \pm 10.01\%$ and $\Delta TV_{\text{control}}\% = 177.06\% \pm 63.00\%$, $P = 0.008$). The difference in $\Delta TV\%$ between the treatment and the control groups was not significant at days 1 and 3 after a short duration of treatment. Increases in ADC in the treatment group ($\Delta ADC_{\text{treatment}}$, median, $30.10\% \pm 18.32\%$, $36.11\% \pm 21.82\%$, $45.22\% \pm 24.36\%$) were significantly higher compared with the control group ($\Delta ADC_{\text{control}}$, median, $4.98\% \pm 3.39\%$, $6.26\% \pm 3.08\%$, $9.24\% \pm 6.33\%$) at days 3, 5 and 7 ($P = 0.008$, $P = 0.016$, $P = 0.008$, respectively). Increases in D in the treatment group ($\Delta D_{\text{treatment}}$, median $17.12\% \pm 8.20\%$, $24.16\% \pm 16.87\%$, $38.54\% \pm 19.36\%$) were higher than those in the control group ($\Delta D_{\text{control}}$, median $-0.13\% \pm 4.23\%$, $5.89\% \pm 4.56\%$, $5.54\% \pm 4.44\%$) at days 1, 3, and 5 ($P = 0.032$, $P = 0.008$, $P = 0.016$, respectively). Relative changes in f were significantly lower in the treatment group compared with the control group at days 1, 3, 5 and 7 follow-up (median, $-34.13\% \pm 16.61\%$ vs $1.68\% \pm 3.40\%$, $P = 0.016$; $-50.64\% \pm 6.82\%$ vs $3.01\% \pm 6.50\%$, $P = 0.008$; $-49.93\% \pm 6.05\%$ vs $0.97\% \pm 4.38\%$, $P = 0.008$, and $-46.22\% \pm 7.75\%$ vs $8.14\% \pm 6.75\%$, $P = 0.008$, respectively). D^* in the treatment group decreased significantly compared to those in the control group at all time points (median, $-32.10\% \pm 12.22\%$ vs $1.85\% \pm 5.54\%$, $P = 0.008$; $-44.14\% \pm 14.83\%$ vs $2.29\% \pm 10.38\%$, $P = 0.008$; $-59.06\% \pm 19.10\%$ vs $3.86\% \pm 5.10\%$, $P = 0.008$ and $-47.20\% \pm 20.48\%$ vs $7.13\% \pm 9.88\%$, $P = 0.016$, respectively). Furthermore, histopathologic findings showed positive correlations with ADC and D and tumor necrosis ($r_s = 0.720$, $P < 0.001$; $r_s = 0.522$, $P = 0.007$, respectively). The cellular apoptosis of the tumor also showed positive correlations with ADC and D ($r_s = 0.626$, $P = 0.001$; $r_s = 0.542$, $P = 0.005$, respectively). Perfusion-related parameters (f and D^*) were positively correlated to MVD ($r_s = 0.618$, $P = 0.001$; $r_s = 0.538$, $P = 0.006$, respectively), and negatively correlated to cellular apoptosis of the tumor ($r_s = -0.550$, $P = 0.004$; $r_s = -0.692$, $P < 0.001$, respectively).

CONCLUSION: IVIM-DWI is potentially useful for predicting the early efficacy of chemotherapy in a human gastric cancer mouse model.

Key words: Gastric cancer; Microvessel density; Nude mouse model; Intravoxel incoherent motion diffusion-weighted imaging; Terminal-deoxynucleotidyl transferase mediated nick end labeling

© The Author(s) 2016. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Intravoxel incoherent motion diffusion-weighted imaging (IVIM-DWI) is useful for monitoring changes of molecular diffusion and microcirculation in gastric cancer at the early stage of chemotherapy. The apparent diffusion coefficient (ADC) and IVIM parameters of true water molecular diffusion coefficient (D) could be reliable marker to detect the necrosis and cellular apoptosis, while perfusion-related IVIM parameters of perfusion fraction (f) and pseudo-related diffusion coefficient (D^*) are capable of noninvasive assessment of angiogenesis activity in gastric cancer undergoing chemotherapy.

Song XL, Kang HK, Jeong GW, Ahn KY, Jeong YY, Kang YJ, Cho HJ, Moon CM. Intravoxel incoherent motion diffusion-weighted imaging for monitoring chemotherapeutic efficacy in gastric cancer. *World J Gastroenterol* 2016; 22(24): 5520-5531 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5520.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5520>

INTRODUCTION

Gastric cancer (GC) remains the fourth most common malignancy and the second-leading cause of cancer deaths in the world^[1]. Despite many advances in cancer diagnosis and treatment, approximately two-thirds of patients are diagnosed with advanced gastric cancer (AGC) in many countries, excluding some trial results in Japanese and South Korean patients^[2,3]. For many years, 5-fluorouracil (5-FU)-based chemotherapy has been considered a standard chemotherapy regimen for AGC^[2-4]. Unfortunately, not all patients benefit from this regimen. If an ineffective therapy can be identified at an early stage of treatment, there will be an opportunity to change the therapy approach and clinical management in the individual patient quickly. The response evaluation criteria in solid tumors (RECIST) is widely adopted standard for evaluating therapy response based on the change in tumor size in clinical practice. However, this often takes several weeks to months to develop, and its evaluation period is too long to adjust patient management^[5]. Therefore, increasing demands are being placed on imaging modalities to identify early

and reliable surrogate markers for the evaluation of therapeutic effect in patients with GC^[6].

Diffusion-weighted magnetic resonance imaging (DWI) is capable of providing an apparent diffusion coefficient (ADC), which is a measure magnitude of diffusion (of water molecules) within tissue, and has become a favorite choice for oncologic studies^[7]. It is well known that a lower ADC is a characteristic of most tumors compare with native tissues because of their high cellularity. An increase in tissue diffusivity, which is induced by an enlarged extracellular space, cell swelling or a loss of membrane integrity under an effective therapy, makes it possible to use ADC to identify the treatment response^[8-11]. However, the ADC value derived from DWI based a mono-exponential model does not sufficiently demonstrate the characteristics of tissue behavior. The *b*-value, which represents the strength and timing of the gradients, determines DWI sensitivity to water motion. When a lower *b*-value is applied (≤ 100 s/mm²), microcirculation related protons have relative large diffusion distances, and are capable of changing diffusion signal intensities. In 1986, Le Bihan *et al.*^[12] first described the concept of intravoxel incoherent motion (IVIM), which can be used to estimate molecular diffusion and microcirculation in the capillaries separately through bi-exponential fitting of the DWI data using multiple *b*-values. In recent years, there has been increasing interests in IVIM-DWI, which allows acquisition of quantitative parameters that reflect tissue diffusivity and microcirculation perfusion simultaneously. The true water molecular diffusion coefficient (D), perfusion fractional (f) volume reflective of capillary blood volume and pseudo-related diffusion coefficient associated with capillary network blood flow (D*) can be measured using IVIM-DWI. The blood microcirculation within capillaries can be considered as a type of "pseudo-diffusion" because it has no specific orientation^[13].

It has been determined that IVIM-DWI can provide a new opportunity to gain an insight into the perfusion of a tumor without contrast agent administration for preclinical and clinical applications^[12,14,15]. Koh *et al.*^[16] reported that the calculated f values were lower in colorectal liver metastases, which are characterized by their hypovascular nature, than in the normal liver. In addition, DWI with 10 *b*-values between 0 and 700 s/mm² enables the measurement of diffusion and microcirculation contributions in renal allografts as early as 5-d after transplantation^[17]. Moreover, IVIM-DWI has been used to evaluate the treatment responses to radiofrequency ablation^[18] or a vascular disrupting agent of CKD-516^[19] in rabbit model with VX2 liver tumors, and to neoadjuvant chemotherapy in human locoregionally advanced nasopharyngeal carcinoma. Although DWI is increasingly being applied in the body, few studies have focused on gastric lesions due to the limitations of modality of the gastric^[20]. Recently, Cheng *et al.*^[21] applied IVIM with *b*-values of up to

1500 s/mm² to evaluate chemotherapeutic efficacy in a gastric cancer xenograft model. In that study, f increased after chemotherapy treatment, which refutes currents theories regarding angiogenesis activity within tumors after treatment^[22]. A previous IVIM study in prostate cancer also found that f in tumors significantly increased compared to that in normal prostate tissue when a *b*-value of less than 750 s/mm² was used, while f decrease or became indistinguishable from the normal prostate tissue when high *b*-values were employed^[23]. This phenomenon could be explained by the following theory: The departure of molecular diffusion at very high *b*-values may have an influence on perfusion-related parameters because both water diffusion and microcirculation contribute to the signal attenuation observed at lower *b*-values (≤ 100 s/mm²)^[13,15]. It should be noted that higher *b* values may affect the accuracy of the IVIM-derived parameters, which depend heavily on the *b*-value selection.

Here, we investigate IVIM-DWI with *b*-values below 800 s/mm² as a potential imaging marker for assessing the early chemotherapy response in term of tissue diffusion and microvascular perfusion, by comparing the tissue cellularity and microvascular density (MVD) properties revealed by histopathological analysis during the full course of treatment using a mouse human gastric cancer xenograft model.

MATERIALS AND METHODS

Animal model

This study included 25 male adult (6 weeks old) nude mice (BALB/c-nu/nu, Orient Bio, Gwangju, South Korea), each weighing 20-25 g. All mice, fed with a standard diet with water ad libitum, were maintained in appropriate laboratory conditions (a photoperiod of 12 h light and darkness, 50% humidity, 23 °C). After a two-week adaption period, 1×10^7 human gastric adenocarcinoma AGS cells (ATCC[®], CRL-1739[™]) suspended in 100 μ L cold PBS were subcutaneously injected into the lower right hind limbs of the nude mice with a 31 gauge syringe. To evaluate the therapeutic response, the tumor growth curves in each group were estimated based on the morphologic T2-weighted images by using the following formula: TV = $\pi/6 \times L \times W \times H$ (L, length, W, width, H, height of the tumor).

Study design

Following a baseline (day 0) MRI examination, human gastric cancer-bearing mice were randomly divided into control and 1-d, 3-d, 5-d, and 7-d treatment groups (*n* = 5 per group). 5-fluorouraci (5-FU) (15 mg/kg)/calcium folinate (5 mg/kg) was used for chemotherapy in the treatment groups, while same volume of saline was administered in the control group. 5-FU/calcium folinate was intraperitoneally injected on a bi-daily basis. Mice in the control group underwent longitudinal MRI at days 1, 3, 5 and 7. Each

treatment group underwent a second scan with same MRI protocol at days 1, 3, 5, or 7 after treatment. After the MRI examination, mice were euthanized by an overdose of isoflurane, and the tumor was stripped for further analysis. The short-term 7-d 5-FU treatment was performed, because this study was aimed to investigate the potential of IVIM-DWI for monitoring the early tumor diffusion and perfusion response to treatment.

MRI protocol

All MRI scans were performed using 3T MRI (GE Healthcare, Waukesha, WI, United States) with a wrist coil. Mice were placed on a heated pad and anesthetized with 2% isoflurane in oxygen (at a rate of 1.0 L/min) to void movement during imaging. After acquisition of the routine images for localization, a transverse T2-weighted image was obtained using fast spin echo (FSE) sequence [repetition time/echo time (TR/TE), 2000/99.6 ms; section thickness, 3 mm; matrix, 512 × 358; number of excitations (NEX), 4]. Subsequently, IVIM-DWI with 12 *b*-values (0, 10, 15, 20, 25, 30, 60, 75, 100, 200, 400 and 800 s/mm²) were acquired using a free-breathing single-shot echo-planar imaging (EPI) sequence with application of three diffusion-gradients directions (TR/TE, 2500/66.5; section thickness, 3 mm; number of sections, 8; field of view, 10 × 10 cm²; matrix, 128 × 128; NEX, 4) with an acquisition time of 7 min and 30 s for each study.

MR imaging analysis

The acquired datasets were transferred to a GE workstation (Advance Workstation 4.6) and analyzed using an in-house software. The ADC value was calculated by using a linear fit (least-squares fit) mono-exponential model based on the following equation: $S_b/S_0 = \exp(-b \times \text{ADC})$ ^[15], where S_0 is the signal intensity at a *b* value of 0 and S_b is the signal intensity at higher *b* values. IVIM parameters was calculated by a nonlinear fit (Levenberg-Marquardt fit) bi-exponential model, and the equation is shown as follow: $S_b/S_0 = (1-f) \times \exp(-bD) + f \times \exp[-b(D + D^*)]$ ^[15], where *D* represents true water molecular diffusion coefficient, *f* and *D** represent perfusion fraction and pseudo-related diffusion coefficient, respectively. A technician with 8 years of experience in MRI measured the tumor sizes and values of ADC, *D*, *f*, and *D**. The technician was blinded to the information regarding the treatment and control groups.

Regions of interest (ROIs) were drawn by outlining the tumor border on ADC maps, which showed the largest cross-section of the tumor. The ROIs on ADC map were copied and pasted on the corresponding *D*, *f*, and *D** maps. In each mouse, the change in tumor volume (TV) relative to the baseline was quantified to determine the treatment response as follows: $\Delta \text{TV}\% = [(TV_{\text{given time}} - TV_{\text{baseline}})/TV_{\text{baseline}}] \times 100$, where $TV_{\text{given time}}$ is the tumor volume on day 1, 3, 5, or 7

and TV_{baseline} is the tumor volume on day 0. For the ADC and IVIM parameters, the percentage changes in values compared to the baseline were recorded by using the following formula: $\Delta \text{Value}\% = [(Value_{\text{given time}} - Value_{\text{baseline}})/Value_{\text{baseline}}] \times 100$, where $value_{\text{given time}}$ is the value on day 1, 3, 5, or 7 and $Value_{\text{baseline}}$ is the value on day 0.

Histopathological analysis

The tumor tissue was paraffin-embedded, and sliced in the transverse plane at 0.6 μm intervals to match the corresponding MR image, was selected. The necrotic fraction (NF) of the tumor was evaluated through hematoxylin-eosin (HE) staining. CD31 (Dako, Carpinteria, CA, United States; mouse anti-human; used at 1:200) staining was carried out on pathological specimens by immunohistochemical methods, to analyze the angiogenesis activity of the tumor. The paraffin-embedded sections were incubated with the primary antibody at 4 °C overnight. Secondary antibody (Stem cell Technologies; rabbit anti-mouse; used at 1:300) was applied at room temperature for 1 h and then sections were rinsed with PBS. Positive reaction was visualized by DAB chromogen (Dako, Carpinteria, CA, United States) according to standard methods. Terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) (Roche Applied Science, Penzberg, Germany) was performed to evaluate cellular apoptosis of the tumor by following the manufacturer's instructions.

Histopathological analysis was performed by using Image J software (<http://rsb.info.nih.gov/ij>). Tumor necrosis was scored according to the following formula: $\text{NF} = \text{Area}_{\text{necrosis}}/\text{Area}_{\text{total tumor}}$. Cellular apoptosis of the tumor was defined as the percentage of positive TUNEL-stained cells among 200 nuclei from five randomly selected fields at a high magnification (× 200). The mean MVD of the tumor was defined by the CD31-stained vessels^[24], where any distinct area of positive for CD31 staining was defined as a single vessel, from five hot spots with higher vascular density compared to the remaining tissue in a high-power field (× 200; 0.578 mm²).

Statistical analysis

SPSS 21.0 (SPSS, Chicago, IL, United States) was used for statistical analysis. The differences in relative changes in the ADC and IVIM parameters, and tumor volumes between the control and treatment groups were determined by the Mann-Whitney test. Spearman's correlation coefficient was used to determine the correlations between histological features, including NF, MVD and TUNEL and the corresponding ADC and IVIM parameters. Spearman's coefficient was considered to be satisfactory with a critical value of $r_s = 0.415$ at *P* value < 0.05 (two-tailed test). A spearman's coefficient of 0.90-1.00 indicated almost perfect agreement; 0.70-0.90 indicated high

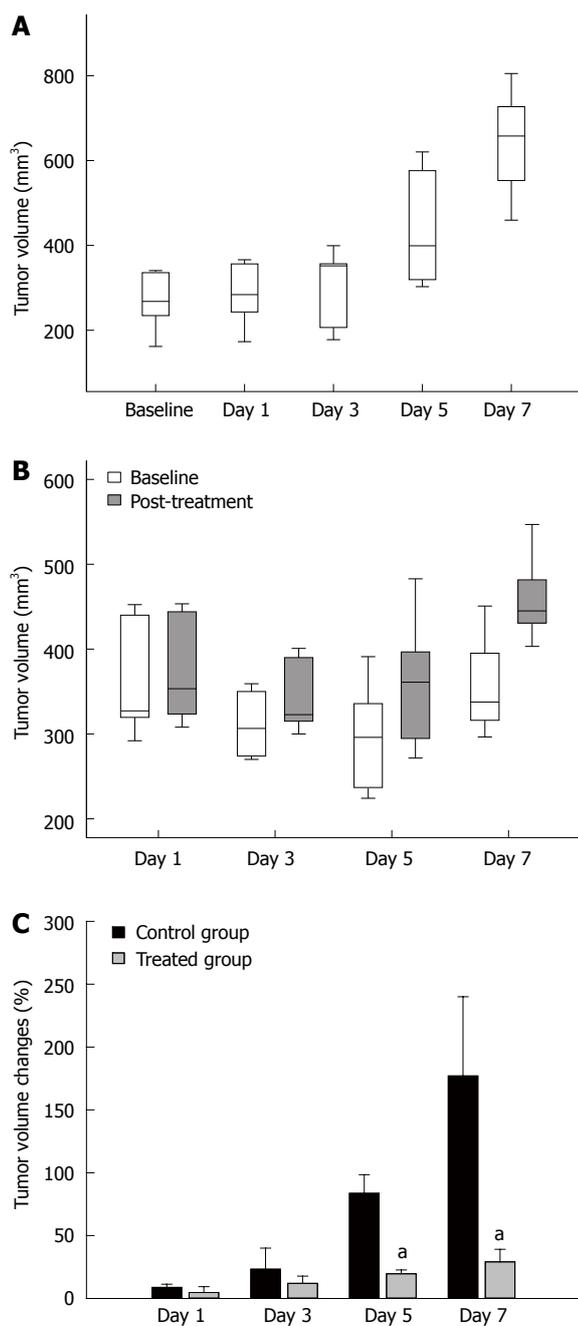


Figure 1 Anti-tumor effects of 5-fluorouracil therapy in mouse gastric cancer xenografts. A: Actual tumor volume changes in the control group; B: Actual tumor volume changes in the treatment groups; C: The comparison of the change in tumor volumes relative to baseline (day 0) between the control and treatment groups. The control group showed increases in tumor volume, while the treatment group showed as significant tumor growth delay from day 5. Center line = median; upper and lower margins of the box = 25th to the 75th percentile, respectively; whiskers = data from the minimum to the maximum. Error bars denote standard errors. ^a*P* < 0.05 vs control group, *n* = 5 in each group.

agreement; 0.50-0.70 indicate moderate agreement; 0.30-0.50 indicate low agreement; and 0.00-0.30 indicate negligible agreement^[25]. A two-tailed *P* value < 0.05 was considered as significant difference.

RESULTS

Effects of chemotherapy on tumor growth

Twenty-five tumors with a mean volume of 298.07 ± 103.44 mm³ (range: 194.32-432.34 mm³) before treatment were analyzed 20 to 25 d of implantation. The tumor volume in the control and treatment groups is shown in Figure 1A and B. 5-FU induced a significant growth delay, as assessed by the MRI-derived tumor volume measurements in comparison with that in the control group from day 5. As shown in Figure 1C, the observed relative volume increase ($\Delta TV\%$) in the therapy groups were significantly lower than in the control group at day 5 ($\Delta TV_{\text{treatment}}\% = 19.63\% \pm 3.01\%$ and $\Delta TV_{\text{control}}\% = 83.60\% \pm 14.87\%$, *P* = 0.008) and day 7 ($\Delta TV_{\text{treatment}}\% = 29.07\% \pm 10.01\%$ and $\Delta TV_{\text{control}}\% = 177.06\% \pm 63.00\%$, *P* = 0.008). The difference in $\Delta TV\%$ between the treatment and the control groups was not significant at day 1 ($\Delta TV_{\text{treatment}}\% = 4.97\% \pm 4.59\%$ and $\Delta TV_{\text{control}}\% = 8.08\% \pm 2.47\%$, *P* = 0.841) or day 3 ($\Delta TV_{\text{treatment}}\% = 15.36\% \pm 5.75\%$ and $\Delta TV_{\text{control}}\% = 23.28\% \pm 16.76\%$, *P* = 0.310) after a short treatment duration.

IVIM-DWI assessment of a human gastric cancer xenograft

Table 1 summarizes the ADC, D, f, and D* values of the tumors in the control and treatment groups, that were measured at baseline and at days 1, 3, 5 and 7. Figure 2 shows the mean percentage changes in the DWI parameters relative to baseline at each time point in each group. In the control group, all ADC values and IVIM parameters of the tumor remained relatively constant over the 7-d experiment. ADC increases in the treatment group ($\Delta ADC\%$ _{treatment}, median, 30.10% ± 18.32%, 36.11% ± 21.82%, 45.22% ± 24.36%) were significantly higher compared with the control group ($\Delta ADC\%$ _{control}, median, 4.98% ± 3.39%, 6.26% ± 3.08%, 9.24% ± 6.33%) at days 3, 5 and 7 (*P* = 0.008, *P* = 0.016, *P* = 0.008, respectively) (Figure 2A). Increases in D in the treatment group ($\Delta D\%$ _{treatment}, median 17.12% ± 8.20%, 24.16% ± 16.87%, 38.54% ± 19.36%) were higher than in the control group ($\Delta D\%$ _{control}, median -0.13% ± 4.23%, 5.89% ± 4.56%, 5.54% ± 4.44%) at days 1, 3, and 5 (*P* = 0.032, *P* = 0.008, *P* = 0.016, respectively) (Figure 2B). The relative changes in f were significantly lower in the treatment group than in the control group at days 1, 3, 5 and 7 follow-up (median, -34.13% ± 16.61% vs 1.68% ± 3.40%, *P* = 0.016; -50.64% ± 6.82% vs 3.01% ± 6.50%, *P* = 0.008; -49.93% ± 6.05% vs 0.97% ± 4.38%, *P* = 0.008, and -46.22% ± 7.75% vs 8.14% ± 6.75%, *P* = 0.008, respectively) (Figure 2C). D* in the treatment group decreased significantly compared with the control group at all time points (median, -32.10% ± 12.22% vs 1.85% ±

Table 1 Summary of the apparent diffusion coefficient and intravoxel incoherent motion parameters at baseline (day 0) and on days 1, 3, 5, and 7 in each groups (*n* = 5 per group)

	ADC (10^{-3} mm ² /s)	IVIM parameters		
		<i>D</i> (10^{-3} mm ² /s)	<i>f</i> (%)	<i>D</i> * (mm ² /s)
Control group				
Day 0	0.514 ± 0.050	0.480 ± 0.049	32.424 ± 6.647	0.112 ± 0.017
Day 1	0.520 ± 0.056	0.497 ± 0.052	33.010 ± 7.121	0.114 ± 0.016
Day 3	0.537 ± 0.029	0.506 ± 0.025	33.606 ± 8.113	0.115 ± 0.020
Day 5	0.545 ± 0.047	0.504 ± 0.021	32.602 ± 6.684	0.117 ± 0.018
Day 7	0.561 ± 0.053	0.524 ± 0.064	35.100 ± 7.858	0.120 ± 0.016
1-d treatment group				
Day 0	0.555 ± 0.028	0.524 ± 0.028	34.076 ± 7.247	0.171 ± 0.049
Day 1	0.597 ± 0.064	0.612 ± 0.065	21.912 ± 5.032	0.115 ± 0.040
3-d treatment group				
Day 0	0.528 ± 0.012	0.510 ± 0.019	37.066 ± 7.331	0.147 ± 0.025
Day 3	0.686 ± 0.107	0.636 ± 0.121	18.596 ± 5.766	0.083 ± 0.029
5-d treatment group				
Day 0	0.594 ± 0.069	0.567 ± 0.065	34.664 ± 8.337	0.173 ± 0.068
Day 5	0.804 ± 0.136	0.777 ± 0.107	17.344 ± 4.352	0.070 ± 0.045
7-d treatment group				
Day 0	0.560 ± 0.060	0.535 ± 0.081	37.598 ± 5.852	0.148 ± 0.082
Day 7	0.823 ± 0.221	0.710 ± 0.236	20.040 ± 3.042	0.065 ± 0.018

ADC: Apparent diffusion coefficient; IVIM: Intravoxel incoherent motion.

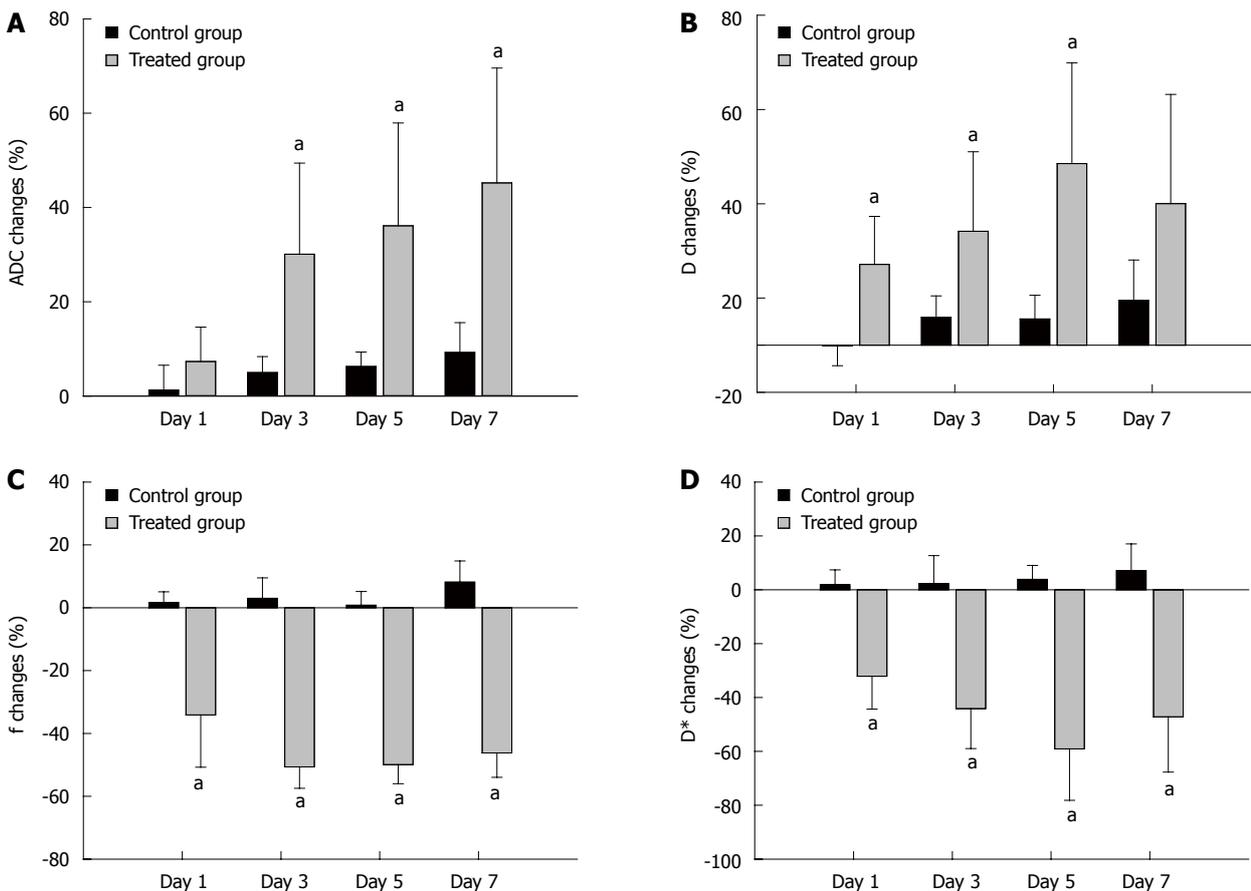


Figure 2 Comparison of the mean percentage changes from baseline in the intravoxel incoherent motion diffusion-weighted imaging derived values between the control (dark) and the 1-, 3-, 5- and 7-d treatment groups (grey). A: ADC value; B: D value; C: *f* value; D: *D** value. Standard deviations are represented by vertical bars. Relative changes were determined by comparing the values at baseline and those in follow-up. ^a*P* < 0.05 vs control. *n* = 5 in each group. ADC: Apparent diffusion coefficient.

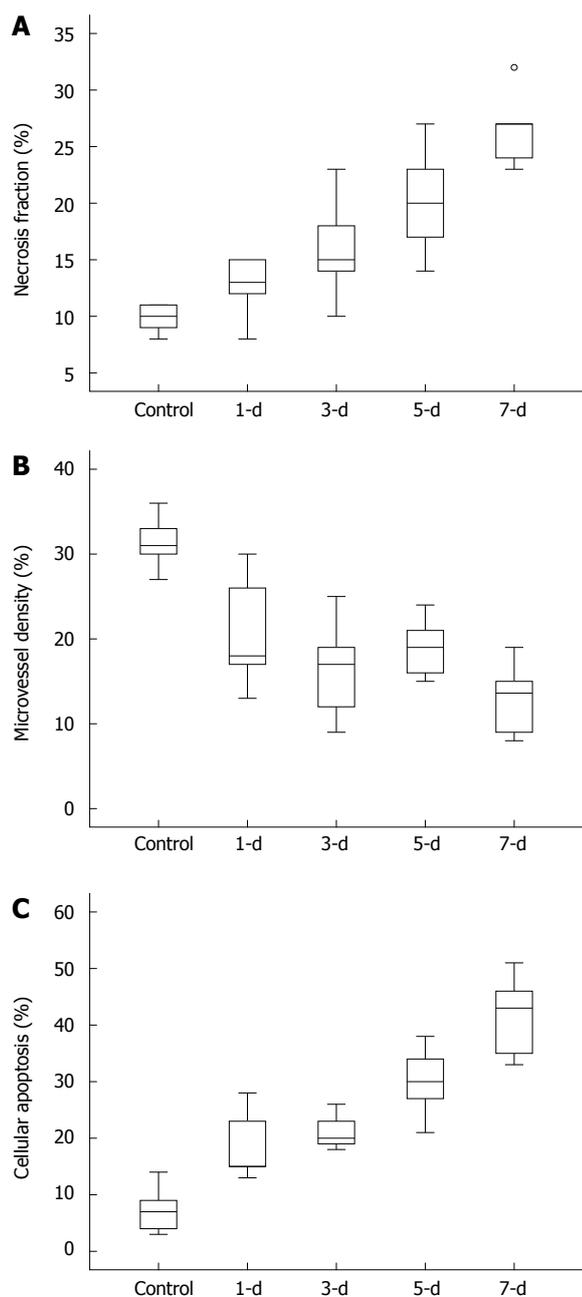


Figure 3 Box-and-Whisker plots show the results of histopathological analysis in the control and the 1-, 3-, 5- and 7-d treatment groups, respectively ($n = 5$ per group). A: Necrosis fraction of tumor; B: Microvessel density of tumor; C: Cellular apoptosis of tumor. Center line = median; upper and lower margins of box = 25th to the 75th percentile, respectively; whiskers = data from the minimum to the maximum; \circ = outlier.

5.54%, $P = 0.008$; $-44.14\% \pm 14.83\%$ vs $2.29\% \pm 10.38\%$, $P = 0.008$; $-59.06\% \pm 19.10\%$ vs $3.86\% \pm 5.10\%$, $P = 0.008$, and $-47.20\% \pm 20.48\%$ vs $7.13\% \pm 9.88\%$, $P = 0.016$, respectively) (Figure 2D).

Histopathological assessment of tumor response and its correlation with MR images

HE, CD31 and TUNEL staining were performed to confirm the tissue and vessel changes in the tumor. Figure 3 shows the quantification of the NF, MVD, and

cellular apoptosis in all animals of the control and the treatment groups ($n = 5$ per group). The MVD scores in 5-FU treatment tumors decreased significantly compared with the control group. The 5-FU treated tumors displayed a time-dependent increase in NF and cellular apoptosis compared to those in the control group, indicating an effective therapeutic response.

Table 2 summarizes the relationship among ADC, the IVIM parameters, MVD, cellular apoptosis and necrosis of tumors ($n = 25$) determined by Spearman's correlation coefficient analysis. ADC and D were positively correlated with NF ($r_s = 0.720$, $P < 0.001$; $r_s = 0.522$, $P = 0.007$, respectively) (Figure 4A, B) and the cellular apoptosis of the tumor ($r_s = 0.626$, $P = 0.001$; $r_s = 0.542$, $P = 0.005$, respectively) (Figure 4E, F). There is no significant correlation among ADC, D, and MVD. Perfusion-related parameters f and D^* shows positive correlations with MVD ($r_s = 0.618$, $P = 0.001$; $r_s = 0.538$, $P = 0.006$, respectively) (Figure 4C, D), and a negative correlation with the cellular apoptosis of the tumor ($r_s = -0.550$, $P = 0.004$; $r_s = -0.692$, $P < 0.001$, respectively) (Figure 4G, H). There is no significant correlation between the perfusion parameters and NF. Figure 5 shows the calculated ADC, D, f , and D^* maps and the correspondent histopathologic images from the control and 3 d treated mice.

DISCUSSION

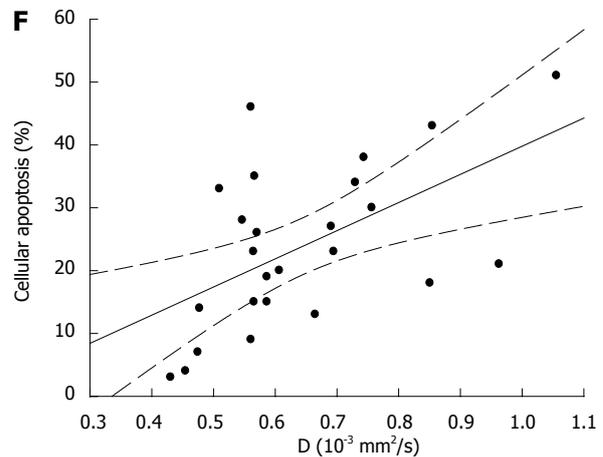
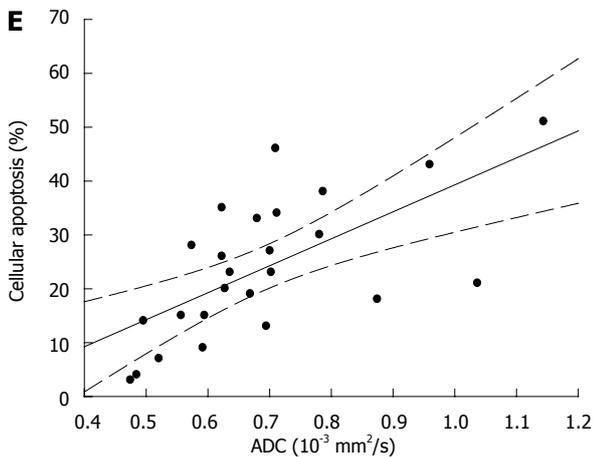
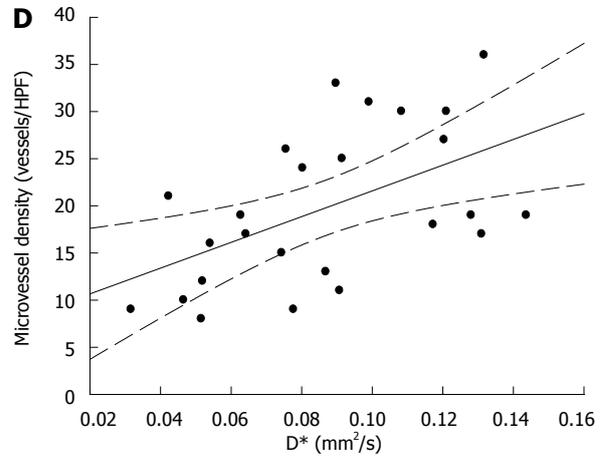
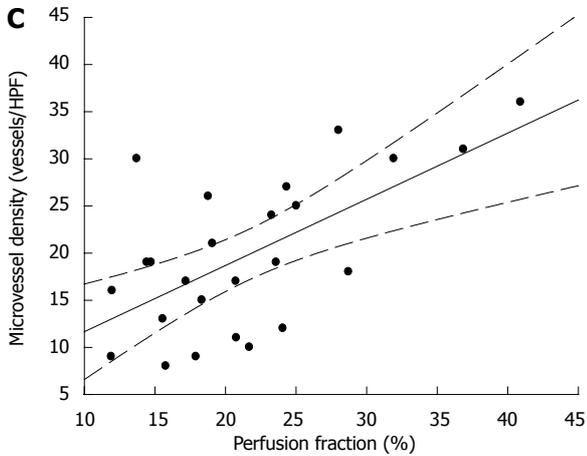
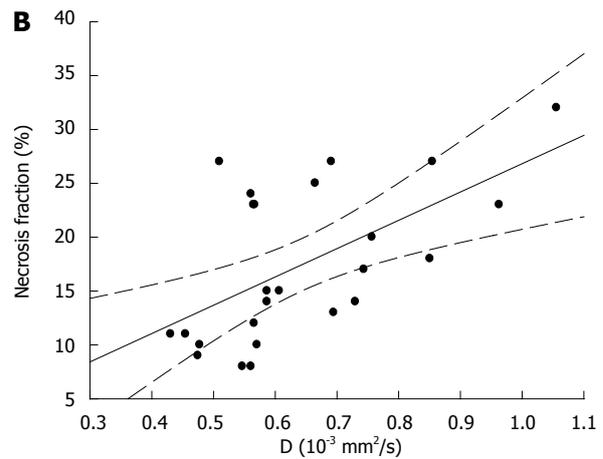
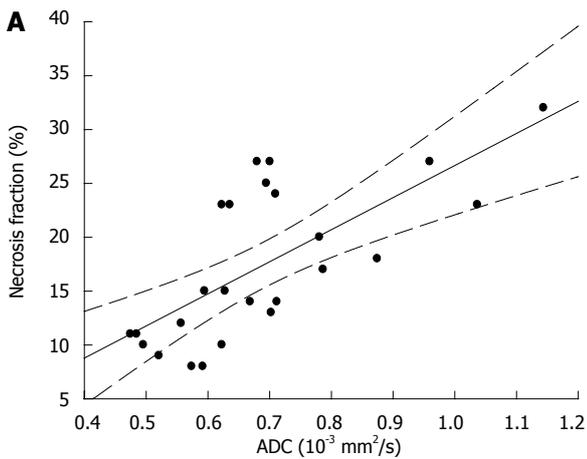
The ADC obtained from conventional DWI has been widely accepted as a marker to monitor the therapeutic efficacy of chemotherapy, radiotherapy or combined therapy with target medicine^[26,27]. In recent years, there has been a resurgent interest in IVIM studies, which allows to measure tissue diffusion and perfusion simultaneously. In this study, we performed IVIM-DWI using 12 b -values less than 800 s/mm^2 to monitor the efficacy of chemotherapy in a mouse model of human gastric cancer. A histopathological analysis was carried out to evaluate the tissue cellularity and MVD properties of the tumor.

Our results demonstrated that conventional ADC significantly increased after 3-d of treatment and it showed a positive correlation with the 5-FU induced intratumoral necrosis and cellular apoptosis of the tumor. Papaevangelou *et al.*^[28] found that ADC changes in the tumor were associated with the induction of a mixture of necrosis and apoptosis after irinotecan treatment. Therefore, ADC could be a reliable marker for detecting the necrotic and apoptotic cell death in gastric cancer patients during treatment. IVIM derived D values that showed a similar trend to ADC, were significantly increased as early as after one day treatment compared with the baseline values. Moreover, HE and TUNEL stain showed that D values were positively correlated with intratumoral necrosis and cellular apoptosis, indicating the possibility of

Table 2 Correlation between apparent diffusion coefficient and intravoxel incoherent motion parameters, microvessel density, cellular apoptosis and necrosis fraction of tumors ($n = 25$)

	NF (%)		MVD (vessels/HPF)		Apoptosis (%)	
	r_s	P value	r_s	P value	r_s	P value
ADC ($\times 10^{-3} \text{ mm}^2/\text{s}$)	0.720	< 0.001	-0.395	0.051	0.626	0.001
IVIM parameters						
D ($\times 10^{-3} \text{ mm}^2/\text{s}$)	0.522	0.007	-0.201	0.335	0.542	0.005
f (%)	-0.347	0.089	0.618	0.001	-0.550	0.004
D^* ($\times \text{ mm}^2/\text{s}$)	-0.378	0.062	0.538	0.006	-0.692	< 0.001

$P < 0.05$ is considered to be statistically significant. ADC: Apparent diffusion coefficient; IVIM: Intravoxel incoherent motion; MVD: Microvessel density; NF: Necrosis fraction.



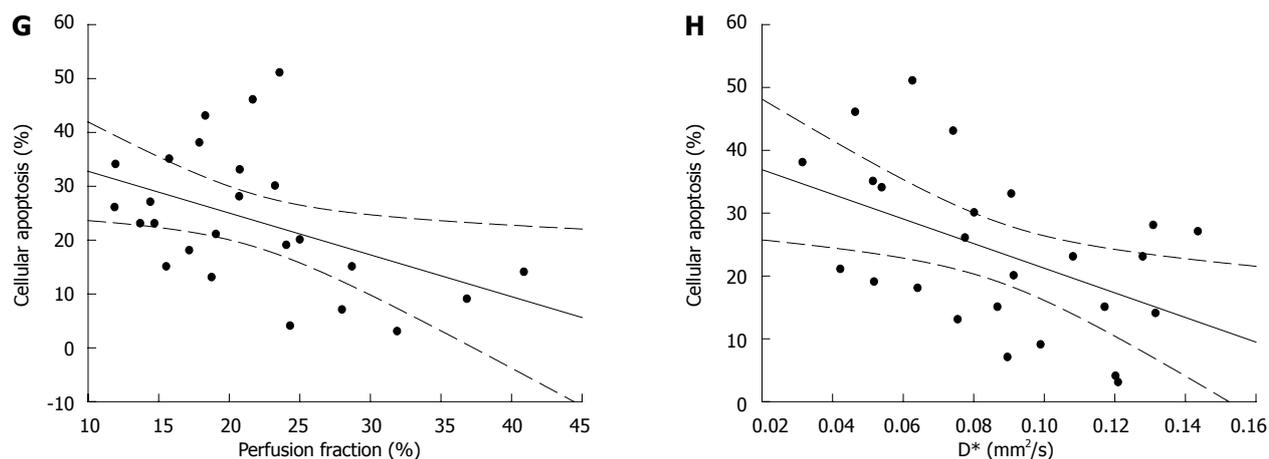


Figure 4 Representative scatter diagrams showing the relationships between intravoxel incoherent motion diffusion-weighted imaging derived parameters and the histological features ($n = 25$). A and B: ADC and D were positively correlated with tumor necrosis fraction; C and D: f and D^* were positively correlated with microvascular density of tumor; E and F: ADC, D were positively correlated with tumor cellular apoptosis; G and H: f and D^* were negatively correlated with tumor cellular apoptosis. P values are shown in Table 2. ADC: Apparent diffusion coefficient.

a noninvasive evaluation of necrosis or apoptotic. Our finding regarding the D value is consistent with previous studies which showed that an increase in D value can predict chemotherapeutic responsiveness in locoregionally advanced nasopharyngeal carcinoma^[20] and advanced cervical cancer^[29]. There is no denying that accurate quantification of the D value, which associates with the intra-to-extracellular spaces ratio and represents the true molecular diffusion, would eventually translate into a reliable marker to evaluate the tumor therapy response.

The f and D^* significantly decreased during the early phase of chemotherapy. In addition, the f and D^* were well correlated with the decrease in MVD revealed by the endothelial cell marker CD31 staining, which indicated that f and D^* have the potential to assess tumor angiogenesis activity noninvasively. There are experimental and clinical dates in body tissues with IVIM-DWI which support the claim that the signal attenuation is related to microcirculation in tissue when b -values less than 100 s/mm^2 were applied. Wang *et al*^[30] found an increase in the tumor blood flow induced by IV hydralazine injection was accompanied by an increase in the IVIM-derived D^* in a mammary adenocarcinoma rat model. In a human brain study, f and D^* were correlated separately with the relative blood volume and blood flow derived from dynamic susceptibility contrast enhancement imaging^[31]. Joo *et al*^[19] reported that IVIM-DWI has the potential to evaluate the early therapeutic effect induced by a vascular disrupting agent named CKD-516 in rabbit VX2 liver tumors. In addition, f and D^* also showed a weak negative correlation with the cellular apoptosis of the tumor, indicating that the decreased cellularity also contributed to the changes in f and D^* . This may reconfirm the finding that tissue water diffusion also contributes to the observed signal attenuation at low b -values. After effective chemotherapy, extracellular spaces would expand, resulting in less restriction

of migration of water molecules and weakening the process of pseudo-diffusion. This would then yield a higher D value and lower f and D^* than those obtained pre-treatment. Our results and the conclusions from previous studies^[13,32] indicated the usefulness of IVIM-DWI for depicting the therapeutic efficiency in gastric cancer before changes in tumor size are evident.

The option of a bi-exponential model for extracting molecular diffusion and microcirculation perfusion information from DWI remains controversial. A previous IVIM study with the b -values up to 1500 s/mm^2 found that f increased after treatment, which contradicts our finding^[21]. Pang *et al*^[23] found that f obtained from b -values below 750 s/mm^2 is more in keeping with the increased perfusion in prostate cancer tissue, while f decrease or became indistinguishable from the normal level of prostate tissue when high b -values employed. Both tissue microcirculation and water diffusion in the tissue contributed to signal attenuation at a low b -value; hence, departure of molecular diffusion at very high b -values may influence the perfusion-related parameters^[33,34]. Moreover, the IVIM method suffers significantly from the variations in the signal-to-noise ratio and is prone to generate measurement errors when b -values are lower than 100 s/mm^2 ^[13]. To date, no consensus has been reached on the magnitude and number of b -values that ought to be applied in preclinical and clinical studies. Therefore, more studies are required to optimize the process of image collection and image post-processing for deriving sufficiently accurate parameters based on the original IVIM model.

There are limitations to this study. First, the observation endpoint in this study is too short. Secondly, because of the small sample sizes, serial relative changes in ADC and the IVIM parameters of the tumor were not evaluated in the treatment groups. Therefore, we need further studies with larger sample size and long-term observations to clarify the limitation of this study.

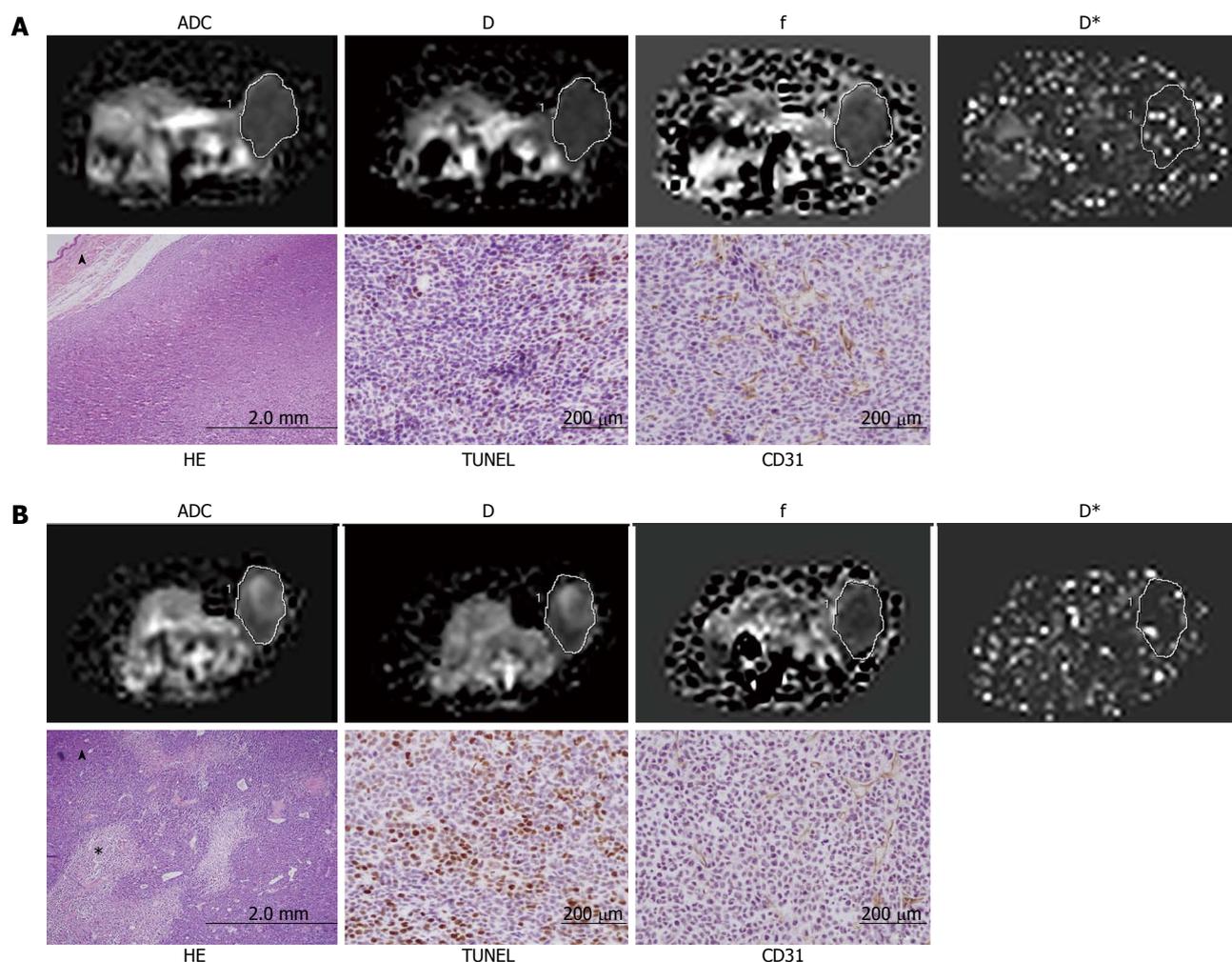


Figure 5 Calculated maps of intravoxel incoherent motion diffusion-weighted imaging parameters and the histopathological images. A: The lower ADC ($0.521 \times 10^{-3} \text{ mm}^2/\text{s}$) and D values ($0.475 \times 10^{-3} \text{ mm}^2/\text{s}$) and the higher f (40.92%) and D^* ($0.131 \text{ mm}^2/\text{s}$) values, which correspond to low necrosis (10%) and cellular apoptosis (7%) and high MVD (36) in the control group; B: Increased ADC ($0.875 \times 10^{-3} \text{ mm}^2/\text{s}$) and D ($0.851 \times 10^{-3} \text{ mm}^2/\text{s}$) values and reduced f (17.20%) and D^* ($0.098 \text{ mm}^2/\text{s}$) values, which correspond to the increased necrosis (18%) and cellular apoptosis (23%) and decreased MVD (17) in the 3-d treatment group. Note high signal intensity within tumor suggesting necrosis. asterisk means necrosis area; The triangle symbol means skin of the mouse. ADC: Apparent diffusion coefficient; MVD: Microvessel density.

In conclusion, IVIM-DWI raises the possibility of an effective, multi-parametric (D, f, D^*) imaging method without requirement of gadolinium enhancement. The IVIM method could potentially be used to assess tissue diffusivity changes in addition to evaluate the microcirculatory perfusion of gastric cancer in response to chemotherapy.

ACKNOWLEDGMENTS

The authors thank Mr. Kim Don Geun GE Healthcare for magnetic resonance imaging technical support and Pro of Moon Jai Dong Chonnam National University Medical School for advice on statistic analysis.

COMMENTS

Background

Chemotherapy is a standard treatment for advanced gastric cancer. Imaging modalities that help to identify early and reliable surrogate markers for the

evaluation of chemotherapy response are important in patients with gastric cancer. Intravoxel incoherent motion diffusion-weighted imaging (IVIM-DWI), which allows acquisition of quantitative parameters that reflect tissue diffusivity and tissue microcapillary perfusion, maybe a useful tool that can monitor the early chemotherapy response in terms of gastric cancer tissue diffusion and microvascular perfusion.

Research frontiers

IVIM-DWI has been used to monitor the treatment responses to radiofrequency ablation or a vascular disrupting agent of CKD-516 in rabbit model with VX2 liver tumors, and the response of human locoregionally advanced nasopharyngeal carcinoma to neoadjuvant chemotherapy. DWI is increasingly being applied in the human body, but few studies focused on the gastric lesions due to the limitation of the modality.

Innovations and breakthroughs

In this study, IVIM-DWI with 12 *b*-values less than $800 \text{ s}/\text{mm}^2$ was performed to avoid the potential impact of higher *b*-values on the accuracy of the IVIM-derived parameters.

Applications

IVIM-DWI raises the possibility of an effective, multi-parametric (D, f, D^*) imaging method that does not require gadolinium enhancement. The IVIM

method could potentially be used to assess tissue diffusivity changes in addition to measuring the microcirculatory perfusion of gastric cancer in response to chemotherapy.

Terminology

DWI is capable of providing a parameter of apparent diffusion coefficient (ADC). The measured ADC, which represents tissue diffusivity, has become a favorite choice in oncologic studies. IVIM-DWI allows acquisition of multi-quantitative parameters that reflect tissue diffusivity and microcirculation perfusion.

Peer-review

The authors have demonstrated the usefulness of IVIM-DWI for monitoring early chemotherapeutic efficacy in a human gastric cancer xenograft model with nude mouse. The manuscript presents interesting and novel findings. The data are well presented and important. However, noted by the authors, the limitation of this study is the short observation endpoint.

REFERENCES

- Carcas LP.** Gastric cancer review. *J Carcinog* 2014; **13**: 14 [PMID: 25589897 DOI: 10.4103/1477-3163.146506]
- Spampatti S, Rausei S, Galli F, Ruspi L, Peverelli C, Frattini F, Rovera F, Boni L, Dionigi G.** Neoadjuvant chemotherapy for locally advanced gastric cancer: the surgeon's role. *Transl Gastrointest Cancer* 2015; **4**: 141-147 [DOI: 10.3978/j.issn.2224-4778.2014.12.02]
- Digklia A, Wagner AD.** Advanced gastric cancer: Current treatment landscape and future perspectives. *World J Gastroenterol* 2016; **22**: 2403-2414 [PMID: 26937129 DOI: 10.3748/wjg.v22.i8.2403]
- Park SC, Chun HJ.** Chemotherapy for advanced gastric cancer: review and update of current practices. *Gut Liver* 2013; **7**: 385-393 [PMID: 23898376 DOI: 10.5009/gnl.2013.7.4.385]
- Therasse P, Arbuck SG, Eisenhauer EA, Wanders J, Kaplan RS, Rubinstein L, Verweij J, Van Glabbeke M, van Oosterom AT, Christian MC, Gwyther SG.** New guidelines to evaluate the response to treatment in solid tumors. European Organization for Research and Treatment of Cancer, National Cancer Institute of the United States, National Cancer Institute of Canada. *J Natl Cancer Inst* 2000; **92**: 205-216 [PMID: 10655437 DOI: 10.1093/jnci/92.3.205]
- Sadeghi-Naini A, Falou O, Hudson JM, Bailey C, Burns PN, Yaffe MJ, Stanisz GJ, Kolios MC, Czarnota GJ.** Imaging innovations for cancer therapy response monitoring. *Imaging Med* 2012; **4**: 311-327 [DOI: 10.2217/iim.12.23]
- Ross BD, Moffat BA, Lawrence TS, Mukherji SK, Gebarski SS, Quint DJ, Johnson TD, Junck L, Robertson PL, Muraszko KM, Dong Q, Meyer CR, Bland PH, McConville P, Geng H, Rehemtulla A, Chenevert TL.** Evaluation of cancer therapy using diffusion magnetic resonance imaging. *Mol Cancer Ther* 2003; **2**: 581-587 [PMID: 12813138]
- Nilsen L, Fangberget A, Geier O, Olsen DR, Seierstad T.** Diffusion-weighted magnetic resonance imaging for pretreatment prediction and monitoring of treatment response of patients with locally advanced breast cancer undergoing neoadjuvant chemotherapy. *Acta Oncol* 2010; **49**: 354-360 [PMID: 20397769 DOI: 10.3109/02841861003610184]
- Cui Y, Zhang XP, Sun YS, Tang L, Shen L.** Apparent diffusion coefficient: potential imaging biomarker for prediction and early detection of response to chemotherapy in hepatic metastases. *Radiology* 2008; **248**: 894-900 [PMID: 18710982 DOI: 10.1148/radiol.2483071407]
- Siegel MJ, Jokerst CE, Rajderkar D, Hildebolt CF, Goyal S, Dehdashti F, Wagner Johnston N, Siegel BA.** Diffusion-weighted MRI for staging and evaluation of response in diffuse large B-cell lymphoma: a pilot study. *NMR Biomed* 2014; **27**: 681-691 [PMID: 24700565 DOI: 10.1002/nbm.3105]
- Wu X, Kellokumpu-Lehtinen PL, Pertovaara H, Korkola P, Soimakallio S, Eskola H, Dastidar P.** Diffusion-weighted MRI in early chemotherapy response evaluation of patients with diffuse large B-cell lymphoma--a pilot study: comparison with 2-deoxy-2-fluoro- D-glucose-positron emission tomography/computed tomography. *NMR Biomed* 2011; **24**: 1181-1190 [PMID: 21387451 DOI: 10.1002/nbm.1689]
- Le Bihan D, Breton E, Lallemand D, Grenier P, Cabanis E, Laval-Jeantet M.** MR imaging of intravoxel incoherent motions: application to diffusion and perfusion in neurologic disorders. *Radiology* 1986; **161**: 401-407 [PMID: 3763909 DOI: 10.1148/radiology.161.2.3763909]
- Koh DM, Collins DJ, Orton MR.** Intravoxel incoherent motion in body diffusion-weighted MRI: reality and challenges. *AJR Am J Roentgenol* 2011; **196**: 1351-1361 [PMID: 21606299 DOI: 10.2214/AJR.10.5515]
- Kang KM, Lee JM, Yoon JH, Kiefer B, Han JK, Choi BI.** Intravoxel incoherent motion diffusion-weighted MR imaging for characterization of focal pancreatic lesions. *Radiology* 2014; **270**: 444-453 [PMID: 24126370 DOI: 10.1148/radiol.13122712]
- Le Bihan D, Breton E, Lallemand D, Aubin ML, Vignaud J, Laval-Jeantet M.** Separation of diffusion and perfusion in intravoxel incoherent motion MR imaging. *Radiology* 1988; **168**: 497-505 [PMID: 3393671 DOI: 10.1148/radiology.168.2.3393671]
- Koh DM, Scurr E, Collins DJ, Pirgon A, Kanber B, Karanjia N, Brown G, Leach MO, Husband JE.** Colorectal hepatic metastases: quantitative measurements using single-shot echo-planar diffusion-weighted MR imaging. *Eur Radiol* 2006; **16**: 1898-1905 [PMID: 16691378 DOI: 10.1007/s00330-006-0201-x]
- Eisenberger U, Thoeny HC, Binser T, Gugger M, Frey FJ, Boesch C, Vermathen P.** Evaluation of renal allograft function early after transplantation with diffusion-weighted MR imaging. *Eur Radiol* 2010; **20**: 1374-1383 [PMID: 20013274 DOI: 10.1007/s00330-009-1679-9]
- Guo Z, Zhang Q, Li X, Jing Z.** Intravoxel Incoherent Motion Diffusion Weighted MR Imaging for Monitoring the Instantly Therapeutic Efficacy of Radiofrequency Ablation in Rabbit VX2 Tumors without Evident Links between Conventional Perfusion Weighted Images. *PLoS One* 2015; **10**: e0127964 [PMID: 26020785 DOI: 10.1371/journal.pone.0127964]
- Joo I, Lee JM, Han JK, Choi BI.** Intravoxel incoherent motion diffusion-weighted MR imaging for monitoring the therapeutic efficacy of the vascular disrupting agent CKD-516 in rabbit VX2 liver tumors. *Radiology* 2014; **272**: 417-426 [PMID: 24697148 DOI: 10.1148/radiol.14131165]
- Xiao Y, Pan J, Chen Y, Chen Y, He Z, Zheng X.** Intravoxel Incoherent Motion-Magnetic Resonance Imaging as an Early Predictor of Treatment Response to Neoadjuvant Chemotherapy in Locoregionally Advanced Nasopharyngeal Carcinoma. *Medicine (Baltimore)* 2015; **94**: e973 [PMID: 26091468 DOI: 10.1097/MD.0000000000000973]
- Cheng J, Zhang C, Wang H, Wu W, Pan F, Hong N, Wang Y.** Evaluation of chemotherapy response of gastric cancer in a mouse model using intravoxel incoherent diffusion-weighted MRI correlated with histopathological characteristics. *ECR* 2015; C-0536 [DOI: 10.1594/ecr2015/C-0536]
- Ma J, Waxman DJ.** Combination of antiangiogenesis with chemotherapy for more effective cancer treatment. *Mol Cancer Ther* 2008; **7**: 3670-3684 [PMID: 19074844 DOI: 10.1158/1535-7163.MCT-08-0715]
- Pang Y, Turkbey B, Bernardo M, Kruecker J, Kadoury S, Merino MJ, Wood BJ, Pinto PA, Choyke PL.** Intravoxel incoherent motion MR imaging for prostate cancer: an evaluation of perfusion fraction and diffusion coefficient derived from different b-value combinations. *Magn Reson Med* 2013; **69**: 553-562 [PMID: 22488794 DOI: 10.1002/mrm.24277]
- Weidner N, Semple JP, Welch WR, Folkman J.** Tumor angiogenesis and metastasis--correlation in invasive breast carcinoma. *N Engl J Med* 1991; **324**: 1-8 [PMID: 1701519 DOI: 10.1056/NEJM199101033240101]
- Hinkle DE, Wiersma W, Jurs SG.** Applied statistics for the behavioral sciences. 5th ed. Boston: Houghton Mifflin, 2003: 89-90
- Woodhams R, Matsunaga K, Iwabuchi K, Kan S, Hata H,**

- Kuranami M, Watanabe M, Hayakawa K. Diffusion-weighted imaging of malignant breast tumors: the usefulness of apparent diffusion coefficient (ADC) value and ADC map for the detection of malignant breast tumors and evaluation of cancer extension. *J Comput Assist Tomogr* 2005; **29**: 644-649 [PMID: 16163035 DOI: 10.1097/01.rct.0000171913.74086.1b]
- 27 **Langer DL**, van der Kwast TH, Evans AJ, Trachtenberg J, Wilson BC, Haider MA. Prostate cancer detection with multi-parametric MRI: logistic regression analysis of quantitative T2, diffusion-weighted imaging, and dynamic contrast-enhanced MRI. *J Magn Reson Imaging* 2009; **30**: 327-334 [PMID: 19629981 DOI: 10.1002/jmri.21824]
- 28 **Papaevangelou E**, Almeida GS, Jamin Y, Robinson SP, deSouza NM. Diffusion-weighted MRI for imaging cell death after cytotoxic or apoptosis-inducing therapy. *Br J Cancer* 2015; **112**: 1471-1479 [PMID: 25880014 DOI: 10.1038/bjc.2015.134]
- 29 **Wang YC**, Hu DY, Hu XM, Shen YQ, Meng XY, Tang H, Li Z. Assessing the Early Response of Advanced Cervical Cancer to Neoadjuvant Chemotherapy Using Intravoxel Incoherent Motion Diffusion-weighted Magnetic Resonance Imaging: A Pilot Study. *Chin Med J (Engl)* 2016; **129**: 665-671 [PMID: 26960369 DOI: 10.4103/0366-6999.177995]
- 30 **Wang Z**, Su MY, Najafi A, Nalcioglu O. Effect of vasodilator hydralazine on tumor microvascular random flow and blood volume as measured by intravoxel incoherent motion (IVIM) weighted MRI in conjunction with Gd-DTPA-Albumin enhanced MRI. *Magn Reson Imaging* 2001; **19**: 1063-1072 [PMID: 11711230 DOI: 10.1016/S0730-725X(01)00431-3]
- 31 **Wirestam R**, Borg M, Brockstedt S, Lindgren A, Holtås S, Ståhlberg F. Perfusion-related parameters in intravoxel incoherent motion MR imaging compared with CBV and CBF measured by dynamic susceptibility-contrast MR technique. *Acta Radiol* 2001; **42**: 123-128 [PMID: 11281143 DOI: 10.1080/028418501127346459]
- 32 **Shinmoto H**, Tamura C, Soga S, Shiomi E, Yoshihara N, Kaji T, Mulkern RV. An intravoxel incoherent motion diffusion-weighted imaging study of prostate cancer. *AJR Am J Roentgenol* 2012; **199**: W496-W500 [PMID: 22997399 DOI: 10.2214/AJR.11.8347]
- 33 **Jensen JH**, Helpert JA. MRI quantification of non-Gaussian water diffusion by kurtosis analysis. *NMR Biomed* 2010; **23**: 698-710 [PMID: 20632416 DOI: 10.1002/nbm.1518]
- 34 **Shinmoto H**, Oshio K, Tanimoto A, Higuchi N, Okuda S, Kuribayashi S, Mulkern RV. Biexponential apparent diffusion coefficients in prostate cancer. *Magn Reson Imaging* 2009; **27**: 355-359 [PMID: 18768281 DOI: 10.1016/j.mri.2008.07.008]

P- Reviewer: Goetze TO, Martin-Villa JM, Zhang J **S- Editor:** Ma YJ
L- Editor: A **E- Editor:** Zhang DN



Basic Study

MicroRNA-21 promotes phosphatase gene and protein kinase B/phosphatidylinositol 3-kinase expression in colorectal cancer

Wei-Zhong Sheng, Yu-Sheng Chen, Chuan-Tao Tu, Juan He, Bo Zhang, Wei-Dong Gao

Wei-Zhong Sheng, Yu-Sheng Chen, Bo Zhang, Wei-Dong Gao, Department of General Surgery, Fudan University affiliated Zhongshan Hospital, Shanghai 200032, China

Chuan-Tao Tu, Juan He, Department of Digestion, Fudan University affiliated Zhongshan Hospital, Shanghai 200032, China

Author contributions: Sheng WZ and Chen YS contributed equally to this study, Sheng WZ, Chen YS and Gao WD designed research; Sheng WZ, Chen YS, Gao WD, Tu CT, He J and Zhang B performed research; Gao WD, Tu CT, He J and Zhang B contributed new reagents or analytic tools; Sheng WZ and Chen YS analyzed data; Sheng WZ and Gao WD wrote the paper.

Institutional review board statement: The study was reviewed and approved by the Fudan University affiliated Zhongshan Hospital Institutional review board.

Conflict-of-interest statement: We declare that there are no conflicts of interest to disclose.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Wei-Dong Gao, MD, Associate Professor, Department of General Surgery, Fudan University affiliated Zhongshan Hospital, NO. 180 Fenglin Road, Shanghai 200032, China. gao.weidong@zs-hospital.sh.cn
Telephone: +86-21-64041990
Fax: +86-21-64041990

Received: February 19, 2016

Peer-review started: February 20, 2016

First decision: March 21, 2016

Revised: April 11, 2016

Accepted: May 4, 2016

Article in press: May 4, 2016

Published online: June 28, 2016

Abstract

AIM: To explore the regulatory mechanism of the target gene of microRNA-21 (miR-21), phosphatase gene (PTEN), and its downstream proteins, protein kinase B (AKT) and phosphatidylinositol 3-kinase (PI3K), in colorectal cancer (CRC) cells.

METHODS: Quantitative real-time PCR (qRT-PCR) and Western blot were used to detect the expression levels of miR-21 and PTEN in HCT116, HT29, Colo32 and SW480 CRC cell lines. Also, the expression levels of PTEN mRNA and its downstream proteins AKT and PI3K in HCT116 cells after downregulating miR-21 were investigated.

RESULTS: Comparing the miR-21 expression in CRC cells, the expression levels of miR-21 were highest in HCT116 cells, and the expression levels of miR-21 were lowest in SW480 cells. In comparing miR-21 and PTEN expression in CRC cells, we found that the protein expression levels of miR-21 and PTEN were inversely correlated ($P < 0.05$); when miR-21 expression was reduced, mRNA expression levels of PTEN did not significantly change ($P > 0.05$), but the expression levels of its protein significantly increased ($P < 0.05$). In comparing the levels of PTEN protein and downstream AKT and PI3K in HCT116 cells after downregulation of miR-21 expression, the levels of AKT and PI3K protein expression significantly decreased ($P < 0.05$).

CONCLUSION: PTEN is one of the direct target genes

of miR-21. Thus, phosphatase gene and its downstream AKT and PI3K expression levels can be regulated by regulating the expression levels of miR-21, which in turn regulates the development of CRC.

Key words: MicroRNA-21; Protein kinase B; Colorectal cancer; Phosphatidylinositol 3-kinase; Phosphatase and tensin homolog

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: RT-PCR and Western blot were applied to detect the expression level of microRNA-21 (miR-21) and Phosphatase gene (PTEN), including its downstream proteins protein kinase B (AKT) and phosphatidylinositol 3-kinase (PI3K) in colorectal cancer (CRC) cell lines, and to explore the regulatory mechanism of the expression of miR-21 in inhibiting CRC, respectively. Their associations were investigated to clarify whether one of the direct target genes of miR-21 is PTEN. The expression levels of miR-21, PTEN and its downstream proteins AKT and PI3K are regulated and controlled to manage the occurrence and progression of CRC.

Sheng WZ, Chen YS, Tu CT, He J, Zhang B, Gao WD. MicroRNA-21 promotes phosphatase gene and protein kinase B/phosphatidylinositol 3-kinase expression in colorectal cancer. *World J Gastroenterol* 2016; 22(24): 5532-5539 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5532.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5532>

INTRODUCTION

MicroRNA (miRNA) is a kind of non-coding macromolecule RNA that includes nearly 22 nucleotides, and it is able to conjugate the 3'UTR of mRNA to facilitate target mRNA degradation and inhibition of the translation process by virtue of lowering gene expression^[1-4]. The research of Calin *et al* indicated that approximately 50% of miRNA was located in tumor-related genomic regions. This work also revealed aberrant expression levels in a number of tumors, which was probably on account of the oncogenic and tumor suppressing gene functions of miRNA molecules. In addition, miRNA is involved in the occurrence and progression of human tumors^[5-8]. Phosphatase gene (PTEN) is a phosphatase and tensin homologue gene derived from chromosome ten, which is associated with phosphohydrolase; its inactivation induces tumor occurrence in the human body^[9-11]. In recent years, the protein kinase B/phosphatase gene/phosphatidylinositol 3-kinase (AKT/PTEN/PI3K) signaling pathway has raised increasing concern, and a number of studies found that the atypical AKT/PTEN/PI3K signaling pathway was intimately linked with numerous tumor occurrences and progression,

immunity, drug resistance, metastasis, angiogenesis, *etc.*^[12-16]. In addition, the AKT/PTEN/PI3K signaling pathway has been reported in pulmonary, nasopharyngeal, gastric and renal tumors, as well as in neuroglioma^[17-21]. However, literature with regard to colorectal cancer (CRC) is rare. Moreover, there were reports that revealed that microRNA-21 (miR-21) could regulate the AKT/PTEN/PI3K signaling pathway to promote tumor occurrence and progression, and even tumor invasion^[22-25]. Hence, quantitative real-time PCR (qRT-PCR) and Western blot were applied to detect the expression level of miR-21 and PTEN, including its downstream proteins AKT and PI3K, in CRC cell lines. Furthermore, the mechanism of miR-21 expression in inhibiting CRC and their correlations were also explored. This study will provide a theoretical and experimental basis for the early diagnosis and therapy of CRC.

MATERIALS AND METHODS

Experimental materials

Primary reagents and equipment: PTEN antibody, PI3K mouse anti-human monoclonal antibody, immunohistochemistry (IHC) kit and AKT rabbit anti-human polyclonal antibody were purchased from Beijing Zhongshan Golden Bridge Biotechnology Company. Quantitative RT-PCR kit and miR-21 primer were purchased from Takara. Fluorescent quantitative PCR detection system was purchased from ABI (United States). The PCR instrument was purchased from Bio-Rad (United States). The inverted microscope was purchased from Olympus (Japan). The refrigerated centrifuge was purchased from Thermo Scientific (United States). HCT116, HT29, Colo32 and SW480 CRC cell strains were all purchased from the Cell Bank of the Chinese Academy of Sciences.

Preparation of primary reagents: Requisite reagents: phosphate buffer solution (PBS), bovine serum albumin (BSA) solution, Tris-buffered saline and Tween-20 (TBST) buffer, BSA blocking buffer, 0.25% trypsin solution, sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) electrophoresis buffer, transmembrane buffer, SDS-PAGE separation and stacking gel.

Experimental methods

CRC cell culture and transfection: A Dulbecco's modified Eagle's medium (DMEM) high glucose medium with 10% calf serum was used to culture HCT116, Ht29, Colo32 and SW480 CRC cell strains in 5% CO₂ at 37 °C. Cells in the logarithmic phase were used in experiments. All cells were sorted into three groups: miR-21 inhibition group (IG), negative control (NC) and blank control (BC). HCT116 cells were inoculated in 500 μL of medium (no antibiotics) to 30%-60% degree to spare. Then, 20 pmol/L of miR-21 inhibitor was

diluted with 50 mL of DMEM medium (no serum) and incubated for five minutes at room temperature after mixing. Next, 1 μ L of mixing Lipofectamine 2000 was diluted in 50 μ L of DMEM (no antibiotics and serum) and incubated at room temperature for 5 min. The diluted miR-21 inhibitor was mixed with Lipofectamine 2000 and incubated at room temperature for 20 min. Then, a 100- μ L transfection buffer was added, mixed and incubated for six hours in an incubator (5% CO₂ at 37 °C). Afterward, the medium was changed to normal medium and cells were incubated for another 48-72 hours for detection.

Transwell assay: Pre-cooling non-serum DMEM was used to dilute the Matrigel matrix gel. Then, it was paved on the chamber of the polycarbonate filtering membrane followed by inoculation of diluted HCT116 cells (100 μ L). Afterwards, 10% fetal calf serum was added to the lower chamber. After 24 h of culture in 5% CO₂ at 37 °C, the chamber was washed twice, stained with 0.1% crystal violet for five minutes, and washed again. Cells were randomly counted in five views at 100 \times objective and the average was calculated. This was performed in three replicates for each group.

Real-time quantitative RT-PCR to detect miR-21 and PTEN mRNA levels: Total RNA was extracted using Trizol RNA extraction methods, based on the Molecular Clone Technique Experimental Manual. One mL of Trizol reagent could be added to approximately 100-mg tissues. A mortar was used to grind the tissues to powder, which eventually reached complete decomposition. The lysate was drawn into a 1.5-mL tube and incubated for 10 min on ice. After centrifuging at 12000 rpm for 10 min at 4 °C, the supernatant was drawn into a new 1.5-mL tube. Isopropanol in equal volume was added and mixed, plated on ice for 10 min, and centrifuged at 12000 rpm for another 10 min at 4 °C. The supernatant was discarded, 1 mL of 75% ethanol was added, the precipitate was washed, and centrifuged for another five minutes. Afterwards, the supernatant was discarded, placed into a tube in room temperature for 10 min, waited until the residue entirely volatilized, then 100 μ L of diethyl pyrocarbonate H₂O was added to dissolve the precipitate, and it was stored in liquid nitrogen for use. Extracted RNA purity and concentration was detected using a spectrophotometer. The normal value of optical density (OD) OD₂₆₀/OD₂₈₀ was between 1.8 and 2.1, respectively. Then, cDNA was synthesized by reverse transcription reaction. RT-PCR was used to detect miR-21 reaction conditions: 95 °C for 10 s, one cycle, 95 °C for 5 s, 60 °C for 34 s, 45 cycles; computational formula: relative amount = $2^{-\Delta\Delta CT}$, where $-\Delta\Delta CT = (CT_{miR-21} - CT_{U6})_{tumor} - (CT_{miR-21} - CT_{U6})_{normal}$ tissue. RT-PCR was used to detect PTEN mRNA reaction conditions: 95 °C for 10 s, one cycle, 95 °C for 5 s, 60 °C for 20 s, 45 cycle; and PTEN mRNA

expression level was calculated on the basis of $\Delta Ct = Ct(PTEN) - Ct(\beta\text{-actin})$, where Folds = $2^{-\Delta\Delta CT}$, and the average result was used with three repeats.

Western blot: After 48-72 h of transfection, the medium was discarded and 100 μ L of RIPA lysate was added into each well. The lysate was mixed for 5 min, centrifuged at 12000 rpm for 15 min at 4 °C, and the supernatant was stored for use. Samples mixed with 5 \times loading were boiled for 10 min at 95 °C and loaded after centrifugation. Eight μ L of purified and desalted antibody was loaded, and 2-3 μ L of the marker was loaded. The antibody was confirmed according to the marker position. Electrophoresis: stacking gel in 80 V for 20-30 min, and separation gel in 120 V for 40 min. Transmembrane: 300 mA for 90 min. After transmembrane, the membrane was blocked with non-fat milk, washed with PBS for three times, and incubated with the primary antibody (PTEN antibody, PI3K antibody, AKT antibody and β -actin antibody; 1:500 dilution) overnight at 4 °C. On the next day, cells were washed with TBST for seven times and incubated with the secondary antibody (horseradish peroxidase-labeled goat-anti rabbit secondary antibody, 1:2500 dilution) for one hour. Then, cells were washed with TBST for another six times. Chemiluminescence reagent A and B solutions were mixed and added on the membrane based on a 1:1 proportion, and β -actin protein was used as an internal reference.

Statistical analysis

SPSS 19.0 was used to analyze all data; measurement data were presented as mean \pm SD. The *t*-test was used to compare miR-21 and PTEN expression levels, and the variance analysis method was used to compare the mean value of transfection in the IG, NC and BC groups. *P* < 0.05 was considered statistically significant.

RESULTS

miR-21 and PTEN expression in CRC cells

miR-21 had the highest protein expression in HCT116 cells, but had the lowest expression in SW480 cells (Figure 1). In contrast, PTEN protein expression was the lowest in HCT116 cells, but had the highest expression in SW480 cells (Figure 2). The protein expression level between miR-21 and PTEN was inversely related, and the difference was statistically significant (*P* < 0.01).

Invasive ability changes in cells after transfection

In counting the cells that crossed after transfection in the Transwell assay, we found that cells transfected with miR-21 in IG were reduced by 39.1% compared to BC and 36.9% compared to NC. However, the difference between BC and NC was not statistically significant (Figure 3).

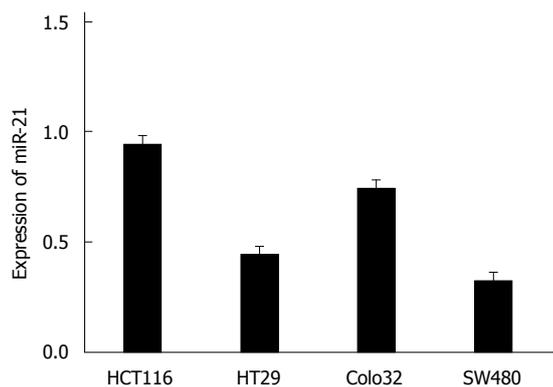


Figure 1 Expression of miR-21 in colorectal cancer cell lines ($n = 3$).

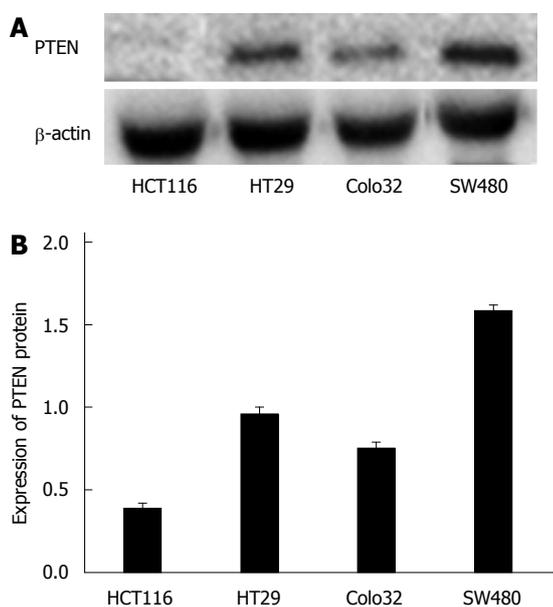


Figure 2 Expression of PTEN protein in colorectal cancer cell lines. A: Western blot assay; B: the expression of PTEN in colorectal cancer cell lines ($n = 3$).

RT-PCR detection of miR-21 expression after transfection

miR-21 expression levels after transfection in IG, NC and BC are shown in Figure 4. It can be observed that miR-21 expression levels in BC and NC were higher than in IG, and the difference was statistically significant ($P < 0.05$).

PTEN mRNA and protein expression in HCT116 cells after downregulation of miR-21

As shown in Figure 5A, after reducing the expression of miR-21, PTEN mRNA expression levels in HCT116 cells revealed no statistical significance after transfection in IG, NC and BC ($P > 0.05$). As illustrated in Figure 5B, when miR-21 was downregulated, PTEN protein expression levels were markedly elevated, and the difference was statistically significant ($P < 0.05$). The difference between BC and NC was not statistically significant ($P > 0.05$).

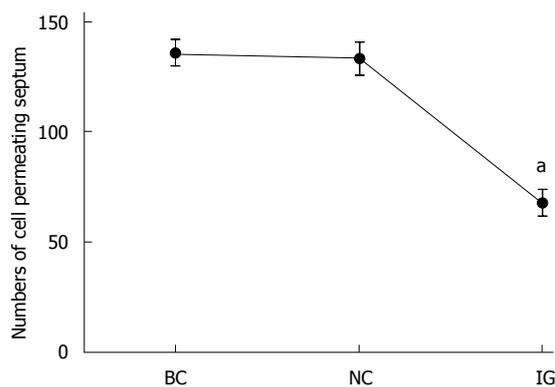


Figure 3 Changes in cell invasion ability after transfection ($^aP < 0.05$, $n = 3$). IG: Inhibition group; NC: Negative control; BC: Blank control.

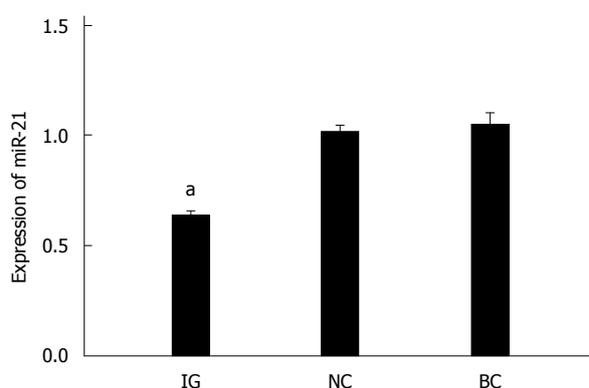


Figure 4 Effect of the miR-21 inhibitor on miR-21 expression in HCT116 cells ($^aP < 0.05$, $n = 3$). IG: Inhibition group; NC: Negative control; BC: Blank control.

PTEN and its downstream proteins AKT and PI3K expression in HCT116 cells after downregulating miR-21

As shown in Figure 6, PTEN protein expression levels in HCT116 cells were elevated in IG, and the difference was statistically significant ($P < 0.05$). However, AKT and PI3K expression levels in HCT116 cells in IG decreased; and the difference was statistically significant ($P < 0.05$).

DISCUSSION

Much research has indicated that CRC occurrence in patients is a complicated process, which consists of multiple stages and factors. One momentous feature is its oncogenic activation and tumor-suppressing gene expression dysregulation or absence^[26,27]. In recent years, it has been found miRNA possesses oncogenic and tumor suppressing gene functions^[22]. One representative miRNA, miR-21, possesses a crucial oncogenic function and exhibits high expression levels in a number of tumors; it is closely associated with tumor occurrence and chemotherapy sensitivity^[28,29]. PTEN is derived from chromosome ten, and acts as a tumor-suppressing gene. PTEN exerts vital effects on cell growth, proliferation, migration, signal transmission,

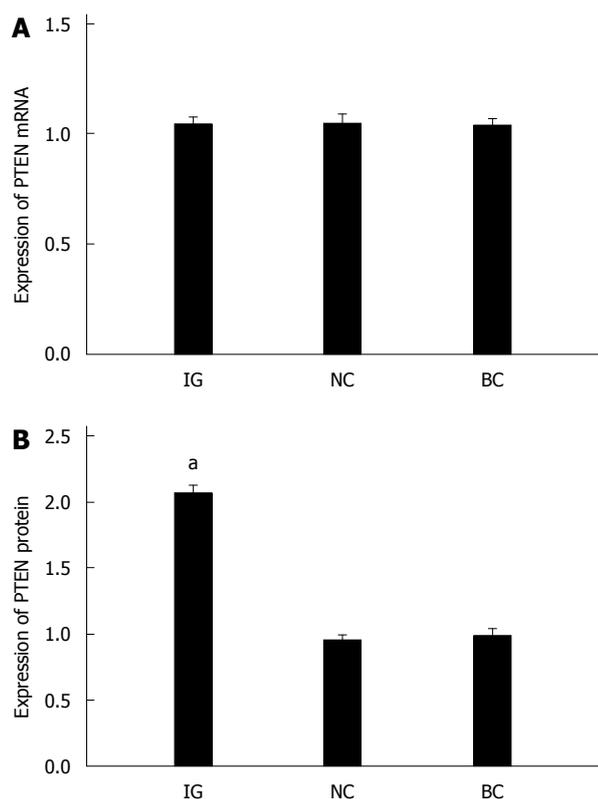


Figure 5 Effect of the miR-21 inhibitor on PTEN mRNA (A) and protein (B) expression in HCT116 cells ($^*P < 0.05$, $n = 3$). IG: Inhibition group; NC: Negative control; BC: Blank control.

invasion and apoptosis; it has an inhibitory role on tumor cell growth, invasion, proliferation, metastasis and apoptosis^[30]. The AKT/PTEN/PI3K signaling pathway plays a significant part in regulating cell growth, metabolism, differentiation and apoptosis; it is closely associated with tumor occurrence and progression^[31-36]. PTEN protein phosphatase activity (dephosphorylation) can inhibit AKT function, leading to cell apoptosis. Furthermore, the deletion or mutation of PTEN is able to enhance AKT activity to increase PI3K expression^[37]. AKT/PTEN/PI3K signaling pathway activation is capable of inhibiting cell apoptosis to facilitate cell differentiation and proliferation. This also exerts a crucial role in tumor occurrence and progression, and participates in tumor metastasis and invasion^[38-44].

miR-21 and PTEN expression in CRC cells

miR-21 and PTEN protein expression was detected in HCT116, HT29, Colo32 and SW480 CRC cell lines through RT-PCR and Western blot. Results revealed that miR-21 expression level was highest in HCT116 cells and lowest in SW480 cells. Furthermore, PTEN expression level was lowest in HCT116 cells and highest in SW480 cells. The protein expression level between miR-21 and PTEN was inversely correlated. These results indicate that miR-21 could be involved in the regulation of PTEN protein expression. In addition, due to the high expression level of miR-21 in HCT116

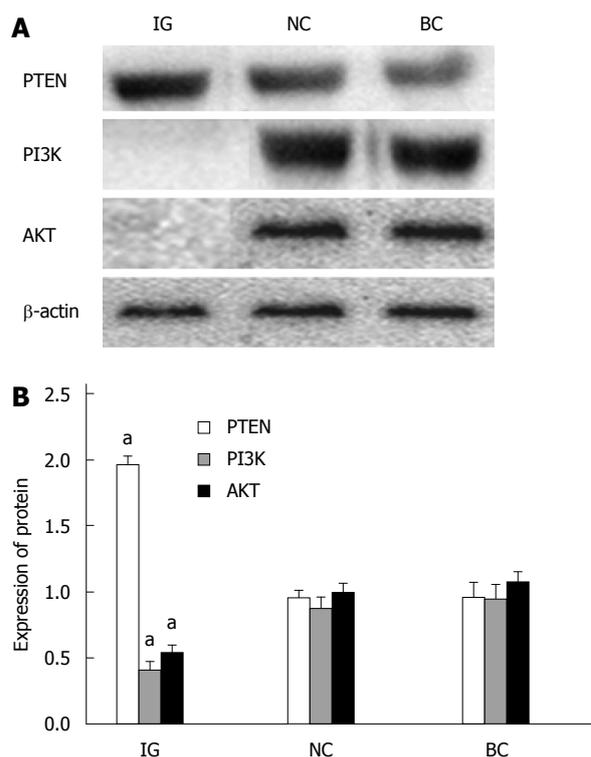


Figure 6 Protein expression of PTEN/AKT/PI3K in colorectal cancer HCT116 cell lines. A: Western blot assay; B: Protein expression in HCT116 cell lines ($^*P < 0.05$, $n = 3$). IG: Inhibition group; NC: Negative control; BC: Blank control.

cells, HCT116 cells were selected for this study. Transwell invasion assay revealed that the invasive capacity of tumor cells was obviously reduced after inhibiting CRC cell miR-21 expression, which in turn indicates that miR-21 probably plays a promoting role in cell malignancy transformation.

miR-21 and PTEN mRNA and protein expression in HCT116 cells after downregulating miR-21

After downregulating the expression of miR-21, the expression level of miR-21 in HCT116 cells in IG was obviously lower compared to NC and BC, which illustrates the successful inhibition of miR-21. As a known miR-21 target gene, PTEN has been verified to be a target gene of miR-21 by Western blot. When we inhibited the expression of miR-21, PTEN mRNA expression levels did not reveal apparent change, but protein levels were obviously elevated; which indicates that miR-21 could regulate PTEN expression at the post-transcriptional level and that PTEN was a target gene for miR-21. Meng *et al.*^[45] recently found that miR-21 targeted tumor-suppressing gene PTEN in liver and bile duct cancer cells to promote tumor growth, invasion and migration, manifesting that PTEN was a target gene of miR-21.

PTEN and its downstream AKT and PI3K expression in HCT116 cells after reducing miR-21 expression

After inhibiting the expression of miR-21, PTEN protein

level was significantly elevated, but its downstream AKT and PI3K expression obviously decreased, indicating that miR-21 could regulate and control PTEN and the expression level of downstream AKT and PI3K in terms of its effects on tumor cell invasion and migration. Certain studies have shown that change in miR-21 expression level is able to induce PTEN downstream molecule AKT phosphorylation, metalloproteinase 2 and focal adhesion kinase expression; these are altered to restrain tumor cell invasion and migration^[45-50].

In conclusion, our study revealed that PTEN is one of the direct target genes of miR-21. By altering miR-21 expression, downstream AKT and PI3K expression levels could be regulated and controlled, in order to control CRC occurrence and progression. Nevertheless, the mechanisms of miR-21 and PTEN function, as well as their effects in tumors, need to be further investigated. As more in-depth research on miRNA is being undertaken, especially regarding tumor cell regulatory mechanisms, more roles and targets of miR-21 will probably be unraveled.

COMMENTS

Background

Colorectal cancer (CRC) is complicated and combines multiple stages and factors. In recent years, we have found that miRNA possesses oncogenic and tumor-suppressing gene functions. One representative miRNA, microRNA-21 (miR-21), possesses a crucial oncogenic function that shows high expression levels in a number of tumors; this is closely associated with tumor occurrence and chemotherapy sensitivity. PTEN acts as a tumor-suppressing gene derived from chromosome ten. PTEN exerts vital effects in cell growth, proliferation, migration, signal transmission, invasion and apoptosis; it has an inhibitory role in tumor cell growth, invasion, proliferation, metastasis and apoptosis. The AKT/PTEN/PI3K signaling pathway plays a significant part in regulating cell growth, metabolism, differentiation, and apoptosis; it is closely associated with tumor occurrence and progression.

Research frontiers

In addition, the AKT/PTEN/PI3K signaling pathway has been reported in pulmonary, nasopharyngeal, gastric and renal tumors, as well as in neuroglioma; however, literature with regard to CRC is rare. Moreover, there have been reports that revealed that miR-21 could regulate the AKT/PTEN/PI3K signaling pathway, in order to promote tumor occurrence and progression, and even tumor invasion. Hence, RT-PCR and Western blot were applied to detect the expression levels of miR-21 and PTEN, including its downstream proteins AKT and PI3K in CRC cell lines, as well as the inhibition of miR-21 expression in CRC cell lines. Furthermore, their association was also explored. This study hopes to provide a theoretical and experimental basis for the early diagnosis and therapy of CRC.

Innovations and breakthroughs

RT-PCR and Western blot were applied to detect the expression level of miR-21 and PTEN including its downstream proteins AKT and PI3K, and the inhibition of miR-21 expression in CRC cell lines, respectively. This association was also investigated to demonstrate that PTEN was one of the direct target genes of miR-21. Through regulating and controlling the expression level of miR-21, PTEN and its downstream proteins AKT and PI3K expression level are regulated to control the occurrence and progression of CRC.

Applications

This study revealed that PTEN was one of the direct target genes of miR-21,

and that its downstream AKT and PI3K expression levels could be regulated and controlled through altering miR-21 expression, in order to control CRC occurrence and progression. Nevertheless, the mechanisms of miR-21 and PTEN function, as well as their effects in tumors, need to be further investigated. As in-depth research on mRNA is currently being implemented, particularly regarding the regulatory mechanism of tumor cells, more roles and targets of miR-21 will probably be unraveled.

Peer-review

This is an interesting manuscript about the miR-21 promotion of PTEN and its downstream proteins AKT and PI3K in CRC. Over all, this study is well designed and the manuscript is well written.

REFERENCES

- 1 **Lewis BP**, Burge CB, Bartel DP. Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell* 2005; **120**: 15-20 [PMID: 15652477 DOI: 10.1016/j.cell.2004.12.035]
- 2 **Janssen HL**, Reesink HW, Lawitz EJ, Zeuzem S, Rodriguez-Torres M, Patel K, van der Meer AJ, Patick AK, Chen A, Zhou Y, Persson R, King BD, Kauppinen S, Levin AA, Hodges MR. Treatment of HCV infection by targeting microRNA. *N Engl J Med* 2013; **368**: 1685-1694 [PMID: 23534542 DOI: 10.1056/NEJMoa1209026]
- 3 **Iorio MV**, Croce CM. MicroRNA dysregulation in cancer: diagnostics, monitoring and therapeutics. A comprehensive review. *EMBO Mol Med* 2012; **4**: 143-159 [PMID: 22351564 DOI: 10.1002/emmm.201100209]
- 4 **Pritchard CC**, Cheng HH, Tewari M. MicroRNA profiling: approaches and considerations. *Nat Rev Genet* 2012; **13**: 358-369 [PMID: 22510765 DOI: 10.1038/nrg3198]
- 5 **Wang B**, Zhang Q. The expression and clinical significance of circulating microRNA-21 in serum of five solid tumors. *J Cancer Res Clin Oncol* 2012; **138**: 1659-1666 [PMID: 22638884 DOI: 10.1007/s00432-012-1244-9]
- 6 **Liu ZL**, Wang H, Liu J, Wang ZX. MicroRNA-21 (miR-21) expression promotes growth, metastasis, and chemo- or radioresistance in non-small cell lung cancer cells by targeting PTEN. *Mol Cell Biochem* 2013; **372**: 35-45 [PMID: 22956424 DOI: 10.1007/s11010-012-1443-3]
- 7 **Kumarswamy R**, Volkmann I, Jazbutyte V, Dangwal S, Park DH, Thum T. Transforming growth factor- β -induced endothelial-to-mesenchymal transition is partly mediated by microRNA-21. *Arterioscler Thromb Vasc Biol* 2012; **32**: 361-369 [PMID: 22095988 DOI: 10.1161/ATVBAHA.111.234286]
- 8 **Parikh VN**, Jin RC, Rabello S, Gulbahce N, White K, Hale A, Cottrill KA, Shaik RS, Waxman AB, Zhang YY, Maron BA, Hartner JC, Fujiwara Y, Orkin SH, Haley KJ, Barabási AL, Loscalzo J, Chan SY. MicroRNA-21 integrates pathogenic signaling to control pulmonary hypertension: results of a network bioinformatics approach. *Circulation* 2012; **125**: 1520-1532 [PMID: 22371328 DOI: 10.1161/circulationaha.111.060269]
- 9 **Song MS**, Salmena L, Pandolfi PP. The functions and regulation of the PTEN tumour suppressor. *Nat Rev Mol Cell Biol* 2012; **13**: 283-296 [PMID: 22473468 DOI: 10.1038/nrm3330]
- 10 **Shrestha S**, Yang K, Guy C, Vogel P, Neale G, Chi H. Treg cells require the phosphatase PTEN to restrain TH1 and TFH cell responses. *Nat Immunol* 2015; **16**: 178-187 [PMID: 25559258 DOI: 10.1038/ni.3076]
- 11 **Carver BS**, Chapinski C, Wongvipat J, Hieronymus H, Chen Y, Chandralapaty S, Arora VK, Le C, Koutcher J, Scher H, Scardino PT, Rosen N, Sawyers CL. Reciprocal feedback regulation of PI3K and androgen receptor signaling in PTEN-deficient prostate cancer. *Cancer Cell* 2011; **19**: 575-586 [PMID: 21575859 DOI: 10.1016/j.ccr.2011.04.008]
- 12 **Zhang BG**, Li JF, Yu BQ, Zhu ZG, Liu BY, Yan M. microRNA-21 promotes tumor proliferation and invasion in gastric cancer by targeting PTEN. *Oncol Rep* 2012; **27**: 1019-1026 [PMID: 22510765 DOI: 10.1038/nrg3198]

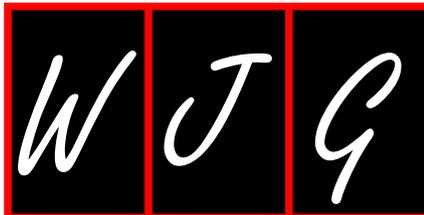
- 22267008 DOI: 10.3892/or.2012.1645]
- 13 **McCubrey JA**, Steelman LS, Chappell WH, Abrams SL, Franklin RA, Montalto G, Cervello M, Libra M, Candido S, Malaponte G, Mazzarino MC, Fagone P, Nicoletti F, Bäsecke J, Mijatovic S, Maksimovic-Ivanic D, Milella M, Tafuri A, Chiarini F, Evangelisti C, Cocco L, Martelli AM. Ras/Raf/MEK/ERK and PI3K/PTEN/Akt/mTOR cascade inhibitors: how mutations can result in therapy resistance and how to overcome resistance. *Oncotarget* 2012; **3**: 1068-1111 [PMID: 23085539 DOI: 10.18632/oncotarget.659]
 - 14 **Hafsi S**, Pezzino FM, Candido S, Ligresti G, Spandidos DA, Souza Z, McCubrey JA, Travali S, Libra M. Gene alterations in the PI3K/PTEN/AKT pathway as a mechanism of drug-resistance (review). *Int J Oncol* 2012; **40**: 639-644 [PMID: 22200790 DOI: 10.3892/ijo.2011.1312]
 - 15 **Hemmings BA**, Restuccia DF. The PI3K-PKB/Akt pathway. *Cold Spring Harb Perspect Biol* 2015; **7**: pii a026609 [PMID: 25833846 DOI: 10.1101/cshperspect.a026609]
 - 16 **Janku F**, Wheler JJ, Westin SN, Moulder SL, Naing A, Tsimberidou AM, Fu S, Falchook GS, Hong DS, Garrido-Laguna I, Luthra R, Lee JJ, Lu KH, Kurzrock R. PI3K/AKT/mTOR inhibitors in patients with breast and gynecologic malignancies harboring PIK3CA mutations. *J Clin Oncol* 2012; **30**: 777-782 [PMID: 22271473 DOI: 10.1200/jco.2011.36.1196]
 - 17 **Shih MC**, Chen JY, Wu YC, Jan YH, Yang BM, Lu PJ, Cheng HC, Huang MS, Yang CJ, Hsiao M, Lai JM. TOPK/PBK promotes cell migration via modulation of the PI3K/PTEN/AKT pathway and is associated with poor prognosis in lung cancer. *Oncogene* 2012; **31**: 2389-2400 [PMID: 21996732 DOI: 10.1038/ncr.2011.419]
 - 18 **Qu C**, Liang Z, Huang J, Zhao R, Su C, Wang S, Wang X, Zhang R, Lee MH, Yang H. MiR-205 determines the radioresistance of human nasopharyngeal carcinoma by directly targeting PTEN. *Cell Cycle* 2012; **11**: 785-796 [PMID: 22374676 DOI: 10.4161/cc.11.4.19228]
 - 19 **Wang F**, Li T, Zhang B, Li H, Wu Q, Yang L, Nie Y, Wu K, Shi Y, Fan D. MicroRNA-19a/b regulates multidrug resistance in human gastric cancer cells by targeting PTEN. *Biochem Biophys Res Commun* 2013; **434**: 688-694 [PMID: 23603256 DOI: 10.1016/j.bbrc.2013.04.010]
 - 20 **Wang KF**, Yang H, Jiang WQ, Li S, Cai YC. Puquintinib mesylate (XC-302) induces autophagy via inhibiting the PI3K/AKT/mTOR signaling pathway in nasopharyngeal cancer cells. *Int J Mol Med* 2015; **36**: 1556-1562 [PMID: 26499488 DOI: 10.3892/ijmm.2015.2378]
 - 21 **Mueller S**, Phillips J, Onar-Thomas A, Romero E, Zheng S, Wiencke JK, McBride SM, Cowdrey C, Prados MD, Weiss WA, Berger MS, Gupta N, Haas-Kogan DA. PTEN promoter methylation and activation of the PI3K/Akt/mTOR pathway in pediatric gliomas and influence on clinical outcome. *Neuro Oncol* 2012; **14**: 1146-1152 [PMID: 22753230 DOI: 10.1093/neuonc/nos140]
 - 22 **Xiong B**, Cheng Y, Ma L, Zhang C. MiR-21 regulates biological behavior through the PTEN/PI-3 K/Akt signaling pathway in human colorectal cancer cells. *Int J Oncol* 2013; **42**: 219-228 [PMID: 23174819 DOI: 10.3892/ijo.2012.1707]
 - 23 **Han M**, Liu M, Wang Y, Chen X, Xu J, Sun Y, Zhao L, Qu H, Fan Y, Wu C. Antagonism of miR-21 reverses epithelial-mesenchymal transition and cancer stem cell phenotype through AKT/ERK1/2 inactivation by targeting PTEN. *PLoS One* 2012; **7**: e39520 [PMID: 22761812 DOI: 10.1371/journal.pone.0039520]
 - 24 **Yang SM**, Huang C, Li XF, Yu MZ, He Y, Li J. miR-21 confers cisplatin resistance in gastric cancer cells by regulating PTEN. *Toxicology* 2013; **306**: 162-168 [PMID: 23466500 DOI: 10.1016/j.tox.2013.02.014]
 - 25 **Wei J**, Feng L, Li Z, Xu G, Fan X. MicroRNA-21 activates hepatic stellate cells via PTEN/Akt signaling. *Biomed Pharmacother* 2013; **67**: 387-392 [PMID: 23643356 DOI: 10.1016/j.biopha.2013.03.014]
 - 26 **Elez E**, Argilés G, Tabernero J. First-Line Treatment of Metastatic Colorectal Cancer: Interpreting FIRE-3, PEAK, and CALGB/SWOG 80405. *Curr Treat Options Oncol* 2015; **16**: 52 [PMID: 26374340 DOI: 10.1007/s11864-015-0369-x]
 - 27 **Vasen HF**, Watson P, Mecklin JP, Lynch HT. New clinical criteria for hereditary nonpolyposis colorectal cancer (HNPCC, Lynch syndrome) proposed by the International Collaborative group on HNPCC. *Gastroenterology* 1999; **116**: 1453-1456 [PMID: 10348829 DOI: 10.1016/s0016-5085(99)70510-x]
 - 28 **Witwer KW**. Circulating microRNA biomarker studies: pitfalls and potential solutions. *Clin Chem* 2015; **61**: 56-63 [PMID: 25391989 DOI: 10.1373/clinchem.2014.221341]
 - 29 **Yang JS**, Li BJ, Lu HW, Chen Y, Lu C, Zhu RX, Liu SH, Yi QT, Li J, Song CH. Serum miR-152, miR-148a, miR-148b, and miR-21 as novel biomarkers in non-small cell lung cancer screening. *Tumour Biol* 2015; **36**: 3035-3042 [PMID: 25501703 DOI: 10.1007/s13277-014-2938-1]
 - 30 **Fragoso R**, Barata JT. Kinases, tails and more: regulation of PTEN function by phosphorylation. *Methods* 2015; **77-78**: 75-81 [PMID: 25448482 DOI: 10.1016/j.ymeth.2014.10.015]
 - 31 **Wang G**, Shi Y, Jiang X, Leak RK, Hu X, Wu Y, Pu H, Li WW, Tang B, Wang Y, Gao Y, Zheng P, Bennett MV, Chen J. HDAC inhibition prevents white matter injury by modulating microglia/macrophage polarization through the GSK3 β /PTEN/Akt axis. *Proc Natl Acad Sci USA* 2015; **112**: 2853-2858 [PMID: 25691750 DOI: 10.1073/pnas.1501441112]
 - 32 **Yang X**, Cheng Y, Li P, Tao J, Deng X, Zhang X, Gu M, Lu Q, Yin C. A lentiviral sponge for miRNA-21 diminishes aerobic glycolysis in bladder cancer T24 cells via the PTEN/PI3K/AKT/mTOR axis. *Tumour Biol* 2015; **36**: 383-391 [PMID: 25266796 DOI: 10.1007/s13277-014-2617-2]
 - 33 **He C**, Dong X, Zhai B, Jiang X, Dong D, Li B, Jiang H, Xu S, Sun X. MiR-21 mediates sorafenib resistance of hepatocellular carcinoma cells by inhibiting autophagy via the PTEN/Akt pathway. *Oncotarget* 2015; **6**: 28867-28881 [PMID: 26311740 DOI: 10.18632/oncotarget.4814]
 - 34 **Paul-Samojedny M**, Pudelko A, Suchanek-Raif R, Kowalczyk M, Fila-Daniłow A, Borkowska P, Kowalski J. Knockdown of the AKT3 (PKB γ), PI3KCA, and VEGFR2 genes by RNA interference suppresses glioblastoma multiforme T98G cells invasiveness in vitro. *Tumour Biol* 2015; **36**: 3263-3277 [PMID: 25501707 DOI: 10.1007/s13277-014-2955-0]
 - 35 **Xu J**, Cai J, Jin X, Yang J, Shen Q, Ding X, Liang Y. PIG3 plays an oncogenic role in papillary thyroid cancer by activating the PI3K/AKT/PTEN pathway. *Oncol Rep* 2015; **34**: 1424-1430 [PMID: 26133772 DOI: 10.3892/or.2015.4096]
 - 36 **Malemud CJ**. The PI3K/Akt/PTEN/mTOR pathway: a fruitful target for inducing cell death in rheumatoid arthritis? *Future Med Chem* 2015; **7**: 1137-1147 [PMID: 26132523 DOI: 10.4155/fmc.15.55]
 - 37 **McCarroll JA**, Gan PP, Erlich RB, Liu M, Dwarte T, Sagnella SS, Akerfeldt MC, Yang L, Parker AL, Chang MH, Shum MS, Byrne FL, Kavallaris M. TUBB3/ β III-tubulin acts through the PTEN/AKT signaling axis to promote tumorigenesis and anoikis resistance in non-small cell lung cancer. *Cancer Res* 2015; **75**: 415-425 [PMID: 25414139 DOI: 10.1158/0008-5472.can-14-2740]
 - 38 **Hevner RF**. Brain overgrowth in disorders of RTK-PI3K-AKT signaling: a mosaic of malformations. *Semin Perinatol* 2015; **39**: 36-43 [PMID: 25432429 DOI: 10.1053/j.semperi.2014.10.006]
 - 39 **Sarbassov DD**, Guertin DA, Ali SM, Sabatini DM. Phosphorylation and regulation of Akt/PKB by the rictor-mTOR complex. *Science* 2005; **307**: 1098-1101 [PMID: 15718470 DOI: 10.1126/science.1106148]
 - 40 **Franke TF**, Yang SI, Chan TO, Datta K, Kazlauskas A, Morrison DK, Kaplan DR, Tsichlis PN. The protein kinase encoded by the Akt proto-oncogene is a target of the PDGF-activated phosphatidylinositol 3-kinase. *Cell* 1995; **81**: 727-736 [PMID: 7774014 DOI: 10.1016/0092-8674(95)90534-0]
 - 41 **Brunet A**, Bonni A, Zigmond MJ, Lin MZ, Juo P, Hu LS, Anderson MJ, Arden KC, Blenis J, Greenberg ME. Akt promotes cell survival by phosphorylating and inhibiting a Forkhead transcription factor. *Cell* 1999; **96**: 857-868 [PMID: 10102273 DOI: 10.1016/S0092-8674(00)80595-4]
 - 42 **Li J**, Yen C, Liaw D, Podsypanina K, Bose S, Wang SI, Puc J, Miliareis C, Rodgers L, McCombie R, Bigner SH, Giovannella BC,

- Ittmann M, Tycko B, Hibshoosh H, Wigler MH, Parsons R. PTEN, a putative protein tyrosine phosphatase gene mutated in human brain, breast, and prostate cancer. *Science* 1997; **275**: 1943-1947 [PMID: 9072974 DOI: 10.1126/science.275.5308.1943]
- 43 **Liaw D**, Marsh DJ, Li J, Dahia PL, Wang SI, Zheng Z, Bose S, Call KM, Tsou HC, Peacocke M, Eng C, Parsons R. Germline mutations of the PTEN gene in Cowden disease, an inherited breast and thyroid cancer syndrome. *Nat Genet* 1997; **16**: 64-67 [PMID: 9140396 DOI: 10.1038/ng0597-64]
- 44 **Maehama T**, Dixon JE. The tumor suppressor, PTEN/MMAC1, dephosphorylates the lipid second messenger, phosphatidylinositol 3,4,5-trisphosphate. *J Biol Chem* 1998; **273**: 13375-13378 [PMID: 9593664 DOI: 10.1074/jbc.273.22.13375]
- 45 **Meng F**, Henson R, Wehbe-Janek H, Ghoshal K, Jacob ST, Patel T. MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer. *Gastroenterology* 2007; **133**: 647-658 [PMID: 17681183 DOI: 10.1053/j.gastro.2007.05.022]
- 46 **Bao L**, Yan Y, Xu C, Ji W, Shen S, Xu G, Zeng Y, Sun B, Qian H, Chen L, Wu M, Su C, Chen J. MicroRNA-21 suppresses PTEN and hSulf-1 expression and promotes hepatocellular carcinoma progression through AKT/ERK pathways. *Cancer Lett* 2013; **337**: 226-236 [PMID: 23684551 DOI: 10.1016/j.canlet.2013.05.007]
- 47 **Vinciguerra M**, Sgroi A, Veyrat-Durebex C, Rubbia-Brandt L, Buhler LH, Foti M. Unsaturated fatty acids inhibit the expression of tumor suppressor phosphatase and tensin homolog (PTEN) via microRNA-21 up-regulation in hepatocytes. *Hepatology* 2009; **49**: 1176-1184 [PMID: 19072831 DOI: 10.1002/hep.22737]
- 48 **Ye JJ**, Cao J. MicroRNAs in colorectal cancer as markers and targets: Recent advances. *World J Gastroenterol* 2014; **20**: 4288-4299 [PMID: 24764666 DOI: 10.3748/wjg.v20.i15.4288]
- 49 **Gayral M**, Jo S, Hanoun N, Vignolle-Vidoni A, Lulka H, Delpu Y, Meulle A, Dufresne M, Humeau M, Chalret du Rieu M, Bournet B, Sélves J, Guimbaud R, Carrère N, Buscail L, Torrisani J, Cordelier P. MicroRNAs as emerging biomarkers and therapeutic targets for pancreatic cancer. *World J Gastroenterol* 2014; **20**: 11199-11209 [PMID: 25170204 DOI: 10.3748/wjg.v20.i32.11199]
- 50 **Iliopoulos D**, Jaeger SA, Hirsch HA, Bulyk ML, Struhl K. STAT3 activation of miR-21 and miR-181b-1 via PTEN and CYLD are part of the epigenetic switch linking inflammation to cancer. *Mol Cell* 2010; **39**: 493-506 [PMID: 20797623 DOI: 10.1016/j.molcel.2010.07.023]

P- Reviewer: Imai K, Mangiola F **S- Editor:** Gong ZM

L- Editor: Logan S **E- Editor:** Ma S





Basic Study

Effects of sphincter of Oddi motility on the formation of cholesterol gallstones

Zhong-Hou Rong, Hong-Yuan Chen, Xin-Xing Wang, Zhi-Yi Wang, Guo-Zhe Xian, Bang-Zhen Ma, Cheng-Kun Qin, Zhen-Hai Zhang

Zhong-Hou Rong, Hong-Yuan Chen, Xin-Xing Wang, Zhi-Yi Wang, Guo-Zhe Xian, Bang-Zhen Ma, Cheng-Kun Qin, Zhen-Hai Zhang, Department of Hepatobiliary Surgery, Provincial Hospital Affiliated to Shandong University, Jinan 250021, Shandong Province, China

Author contributions: Rong ZH and Zhang ZH contributed equally to this work; Zhang ZH, Rong ZH, Xian GZ, and Qin CK designed the research; Rong ZH, Chen HY, Wang XX, Wang ZY, and Ma BZ performed the research; Rong ZH and Zhang ZH analyzed the data; Rong ZH and Zhang ZH wrote the paper.

Supported by Natural Science Foundation of Shandong Province, China, No. ZR 2012 HM -079.

Institutional review board statement: The study was reviewed and approved by the Provincial Hospital Affiliated to Shandong University Institutional Review Board.

Institutional animal care and use committee statement: The animal protocol was designed to minimize pain or discomfort to the animals. The animals were acclimatized to laboratory conditions (room temperature 23 °C, 12-h light and dark cycle, 50% humidity, ad libitum access to food and water) for two weeks prior to experimentation. All animals were euthanized by barbiturate overdose (intravenous injection, 150 mg/kg pentobarbital sodium) for tissue collection.

Conflict-of-interest statement: The authors declare there is no conflict of interest related to this study.

Data sharing statement: Technical appendix, statistical code, and dataset available from the corresponding author at zhangzhenhai410@126.com. Participants gave informed consent for data sharing.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

[licenses/by-nc/4.0/](http://creativecommons.org/licenses/by-nc/4.0/)

Correspondence to: Zhen-Hai Zhang, PhD, Department of Hepatobiliary Surgery, Provincial Hospital Affiliated to Shandong University, No. 324 Jingwuweiqi Road, Jinan 250021, Shandong Province, China. zhangzhenhai410@126.com
Telephone: +86-531-68776363
Fax: +86-531-68776363

Received: January 31, 2016
Peer-review started: February 1, 2016
First decision: March 7, 2016
Revised: March 30, 2016
Accepted: April 20, 2016
Article in press: April 20, 2016
Published online: June 28, 2016

Abstract

AIM: To investigate the mechanisms and effects of sphincter of Oddi (SO) motility on cholesterol gallbladder stone formation in guinea pigs.

METHODS: Thirty-four adult male Hartley guinea pigs were divided randomly into two groups, the control group ($n = 10$) and the cholesterol gallstone group ($n = 24$), which was sequentially divided into four subgroups with six guinea pigs each according to time of sacrifice. The guinea pigs in the cholesterol gallstone group were fed a cholesterol lithogenic diet and sacrificed after 3, 6, 9, and 12 wk. SO manometry and recording of myoelectric activity were obtained by a multifunctional physiograph at each stage. Cholecystokinin-A receptor (CCKAR) expression levels in SO smooth muscle were detected by quantitative real-time PCR (qRT-PCR) and serum vasoactive intestinal peptide (VIP), gastrin, and cholecystokinin octapeptide (CCK-8) were detected by enzyme-linked immunosorbent assay at each stage in the process of cholesterol gallstone formation.

RESULTS: The gallstone formation rate was 0%, 0%, 16.7%, and 83.3% in the 3, 6, 9, and 12 wk groups, respectively. The frequency of myoelectric activity in the 9 wk group, the amplitude of myoelectric activity in the 9 and 12 wk groups, and the amplitude and the frequency of SO in the 9 wk group were all significantly decreased compared to the control group. The SO basal pressure and common bile duct pressure increased markedly in the 12 wk group, and the CCKAR expression levels increased in the 6 and 12 wk groups compared to the control group. Serum VIP was elevated significantly in the 9 and 12 wk groups and gastrin decreased significantly in the 3 and 9 wk groups. There was no difference in serum CCK-8 between the groups.

CONCLUSION: A cholesterol gallstone-causing diet can induce SO dysfunction. The increasing tension of the SO along with its decreasing activity may play an important role in cholesterol gallstone formation. Expression changes of CCKAR in SO smooth muscle and serum VIP and CCK-8 may be important causes of SO dysfunction.

Key words: Cholesterol gallstone; Sphincter of Oddi; Manometry; Myoelectric activity; Cholecystokinin-A receptor

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: This study investigated the role of sphincter of Oddi (SO) motility in cholesterol gallstone formation in a guinea pig model. The myoelectric activity and manometry of SO were measured at different stages of stone formation. As SO motility is controlled by neurological and hormonal factors, we detected the expression of serum vasoactive intestinal peptide (VIP), gastrin, cholecystokinin octapeptide (CCK-8), and CCK-A receptor (CCKAR) in the SO at different stages of stone formation. We found that a cholesterol gallstone-causing diet can induce SO dysfunction and expression changes of CCKAR in SO smooth muscle and serum VIP and CCK-8 may be important causes of SO dysfunction.

Rong ZH, Chen HY, Wang XX, Wang ZY, Xian GZ, Ma BZ, Qin CK, Zhang ZH. Effects of sphincter of Oddi motility on the formation of cholesterol gallstones. *World J Gastroenterol* 2016; 22(24): 5540-5547 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5540.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5540>

INTRODUCTION

Gallstone disease is one of the most common digestive disorders requiring hospital admission in Western countries, with a morbidity of 10%-15% in adults^[1]. In China, the incidence of cholesterol gallstones has been increasing in the past few decades due to

changing lifestyles^[2]. Cholesterol gallstone formation is a complicated process and is still not fully understood. Cholesterol supersaturation of bile, biliary stasis, and mucus hypersecretion are universally known to be important factors in the process of cholesterol gallstone formation^[3,4]. Among them, biliary stasis is thought to be a key factor because it allows time for cholesterol nucleation and then retention of the precipitated microcrystals^[4,5]. The sphincter of Oddi (SO) may play an important role in gallstone formation because it is the only way by which bile is discharged into the duodenum. There are very few reports investigating the relationship between SO motility and cholesterol gallstone formation, and the conclusions are controversial^[6-8].

It is well-established that cholecystokinin (CCK) is one of the major gastrointestinal hormones responsible for gallbladder contraction and SO relaxation^[9]. The biological actions of CCK in the alimentary tract are mediated by the CCK-A receptor (CCK-AR)^[10]. A series of studies focused on the expression level of CCK-AR in the gallbladder in the pathogenesis of cholesterol gallstone disease^[5,11]. However, no report on CCK-AR expression in the SO in animals with gallstones is available yet.

The aim of this study was to investigate the role of SO motility in cholesterol gallstone formation in a guinea pig model. The myoelectric activity and manometry of SO were measured at different stages of stone formation in this model. As SO motility is controlled by neurological and hormonal factors, we detected serum vasoactive intestinal peptide (VIP), gastrin, CCK-8, and CCK-AR expression in the SO at different stages of stone formation.

MATERIALS AND METHODS

Experimental animals

Thirty-four adult male Hartley guinea pigs, weighing between 230 g and 270 g, were purchased from Huishan Jiangnan Laboratory Animal Company (License SCXK SU: 2009-0005). The animals were housed in climate-controlled rooms under an alternating 12-h light and dark cycle and permitted continuous access to food and water. After a 2-wk equilibration, the animals were divided randomly into two groups (control and cholesterol stone groups): the control group ($n = 10$) was fed a normal diet, and the cholesterol stone group ($n = 24$) was sequentially divided into four subgroups with six guinea pigs each according to time of sacrifice, fed a cholesterol lithogenic diet^[6], and sacrificed after 3, 6, 9, and 12 wk. The lithogenic diet consisted of 1% cholesterol^[10] (purchased from Trophic Animal Feed High-Tech Co. Ltd., Nantong, China). Gallstones were defined as macroscopically visible sediment. The calculi were tested by infrared spectrometry to verify the sample as cholesterol gallstones. SO manometry and myoelectric activity of the guinea pigs were determined at 3, 6, 9, and 12 wk.

Table 1 Primer sequences of *CCKAR* and *GAPDH*

Genes	Forward primer (5'-3')	Reverse primer (5'-3')
<i>CCKAR</i>	ACGGAGGGTAGTGAACCTCCA	TCGCAGGCAGAAGTGATGTT
<i>GAPDH</i>	GCACCGTCAAGGCTGAGAAT	CATCACGAACATAGGGGCATC

CCKAR: Cholecystokinin-A receptor; GAPDH: Glyceraldehyde-3-phosphate dehydrogenase.

Measurement of myoelectric activity of the SO

At the end of the feeding period, the animals were anesthetized by injecting pentobarbital sodium (45 mg/kg) into the peritoneal cavity. The guinea pigs were fixed in the supine position, and the skin of the superior abdomen was prepared and sterilized. A longitudinal incision was made and the papilla determined. Details of the myoelectric system were described previously^[12]. In brief, two polar hook metal electrodes were inserted 0.2 mm into the subserosa of the SO by megaloscope ($\times 10$ magnification). The fan-out of the two signals was connected with the two polars of the physiological recorder (BL-420 F; Chengdu Taimeng Software, Chengdu, China), and a piece of metal needle was inserted into the legs of the animals to connect with the earth pole of the recorder. The myoelectric signal was collected by the electrode, imported into the computer, and stored after processing by a physiological recorder and the software system specialized in electromyographic signals. The setup parameters were as follows: scanning speed, 500 ms/div; sensitivity, 200 μ V; time parameter, 1 s; and frequency filtering, 10 Hz. Finally, the myoelectric figure was dealt with by digital filtering of 10-30 Hz.

SO manometry

Details of manometry were described previously^[12]. In brief, a manometry catheter was modified from a pedo bi-lumen central venous catheter (4F and 30 cm long). The catheter was inserted into the common bile duct (CBD) and SO through the duodenal ampulla. The pressure transducer was used for receiving the dynamic pressure change from the manometric lumen. The frequency of SO phasic contraction, SO amplitude, SO basal pressure, and CBD pressure were measured and recorded. A physiological recorder and relevant manometry program were used to record and analyze the tracings.

Detection of serum VIP, gastrin, and CCK-8

Four milliliters of venous blood were obtained from the guinea pigs in the early morning before they were sacrificed and placed in a test tube. The blood was centrifuged at 1500 r/min for 15 min, and serum was isolated, placed in Eppendorf tubes and stored at -70°C . Serum VIP, gastrin, and CCK-8 were measured by enzyme-linked immunosorbent assay (ELISA). The ELISA testing kit was supplied by USCN Life Science Inc. (Houston, TX, United States).

Quantification of *CCKAR* mRNA in SO tissues

The SO of each animal was quickly removed and transferred to normal saline after animals were euthanized. The smooth muscle was removed carefully using sharp dissection. Total RNA from SO tissue samples was extracted using TRIzol reagent (Invitrogen, Carlsbad, CA, United States). Every 50 mg of SO tissue sample was extracted with 1 mL of TRIzol reagent according to the manufacturer's protocol. One microgram of total RNA was used for the synthesis of cDNA using a FastQuant RT Kit and target genes were assayed using SYBR Green Real-time PCR Master Mix (*via* Roche Light Cycler, Roche, Basel, Switzerland) with their respective primers. The PCR conditions were as follows: 95°C for 30 s; 40 cycles of 95°C for 5 s, 57°C for 10 s, and 72°C for 15 s. Transcription levels of glyceraldehyde-3-phosphate dehydrogenase (GAPDH) were used as an internal control to calculate fold induction, and the fold changes in transcription levels were calculated using the $2^{-\Delta\Delta\text{Ct}}$ method. Primer sequences of *CCKAR* and *GAPDH* are shown in Table 1.

Statistical analysis

Statistical analysis was performed using Student's *t* test. Data were analyzed with software SPSS 17.0 (SPSS Inc. Chicago, IL, United States), and $P < 0.05$ was set as the level of significance. The results are expressed as mean \pm SE.

RESULTS

Gallstone formation rate

No gallstone was found in guinea pigs in the control group. In contrast, the gallstone formation rate in the cholesterol stone group fed with a cholesterol lithogenic diet was 0%, 0%, 16.7%, and 83.3% in the 3, 6, 9, and 12 wk groups, respectively.

SO myoelectric activity analysis

Compared with the control group, the frequency of myoelectric activity decreased markedly in the 9 wk group (Figure 1A and B, Table 2). The amplitude of myoelectric activity decreased significantly in the 9 and 12 wk groups (Figure 1A-C, Table 2).

SO manometry analysis

Compared with the control group, the amplitude and frequency of SO decreased significantly in the 9 wk group (Table 3), and SO basal pressure and common

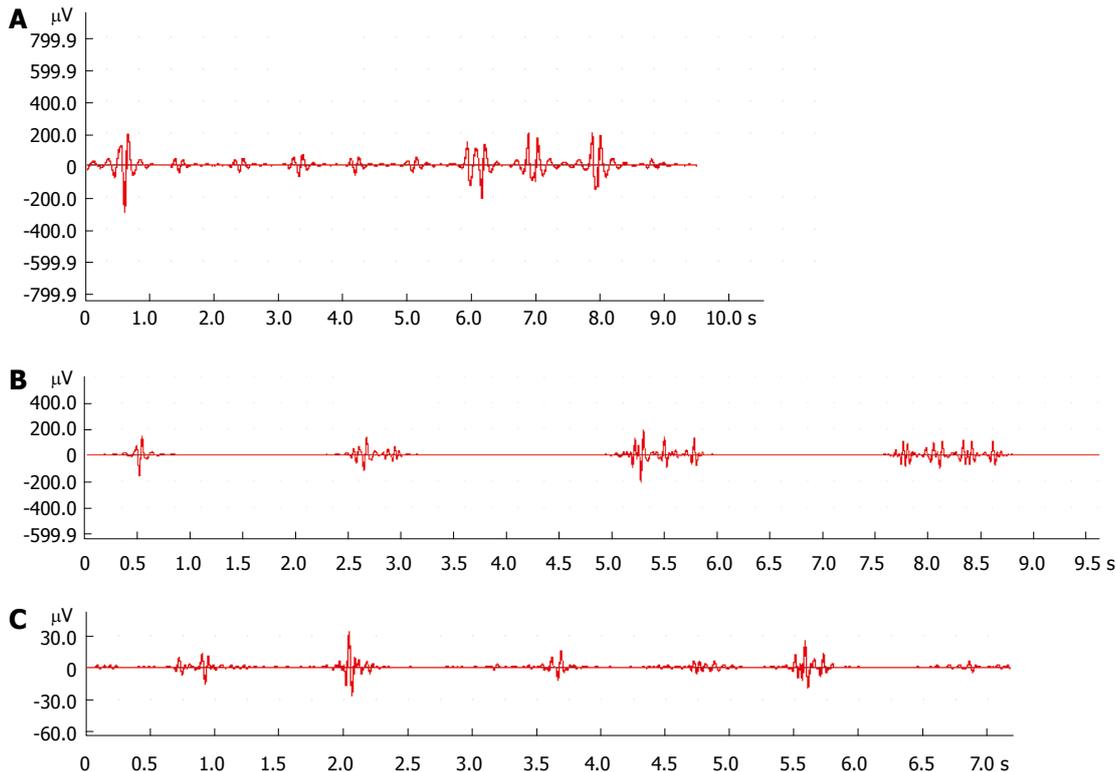


Figure 1 Spincter of Oddi myoelectric activity analysis. A: The frequency and amplitude of myoelectric activity in the control group; B: The frequency and amplitude of myoelectric activity in the 9 wk group; C: The frequency and amplitude of myoelectric activity in the 12 wk group.

Table 2 Changes of spincter of Oddi myoelectric activity in the process of gallstone formation

Group	Amplitude	Frequency
Control group	142.45 ± 71.25	16.56 ± 4.05
3-wk group	125.06 ± 59.76	13.70 ± 2.88
6-wk group	152.96 ± 81.05	15.10 ± 4.13
9-wk group	100.27 ± 50.42 ^a	8.68 ± 2.35 ^a
12-wk group	40.60 ± 15.03 ^c	13.80 ± 3.79

^a*P* < 0.05, ^c*P* < 0.01 vs the control group.

bile duct pressure increased significantly in the 12 wk group (Figure 2, Table 3).

Changes in serum VIP, gastrin and CCK-8

Compared with the control group, serum VIP was elevated significantly in the 9 and 12 wk groups (Table 4), and serum gastrin was decreased significantly in the 3 and 9 wk groups (Table 4). There was no difference in serum CCK-8 between the groups (Table 4).

Quantitative expression of CCK-AR mRNA

Compared with the control group, expression levels of the CCK-A receptor mRNA were increased significantly in the 6 and 12 wk groups (Figure 3).

DISCUSSION

Gallstone disease is one of the most common and most expensive digestive disorders requiring admission to

the hospital^[13-15], with a prevalence of 10%-15% in adults in Europe and the United States. In Western countries, cholesterol gallstones account for 80%-90% of gallstones at cholecystectomy^[15], but the mechanism underlying the pathogenesis of cholesterol gallstone disease is not completely understood.

Epidemiological evidence indicates that multiple environmental factors and genetic elements are involved in cholesterol gallstone formation. Among them, biliary stasis is thought to be an important factor. A series of studies in human and animal models have shown that formation of cholesterol gallstones is causatively related to decreased gallbladder contractility^[16]. Since SO is the only gate through which bile is discharged into the duodenum, bile filling and excretion of the gallbladder are closely related to the motility state of the SO^[17]. However, there is limited research on the role of SO motility in the process of cholesterol gallstone formation. SO manometry (SOM) is the only method that can assess directly the motor function of the SO and is considered the gold standard for assessing SO dysfunction (SOD)^[18]. However, manometry changes of the SO in cholelithiasis are controversial. Research by Pang *et al.*^[19] indicated that the base pressure of the SO was significantly increased in rabbits fed a cholesterol lithogenic diet. On the other hand, a study in prairie dogs showed that SO resistance remained normal throughout the period of gallstone formation. These completely different results may be species dependent. Meanwhile, the mechanical activity recorded may not represent the features of

Table 3 Sphincter of Oddi manometry in the process of gallstone formation

Group	SO basal pressure	CBD pressure	Amplitude of SO	Frequency of SO
Control group	26.59 ± 8.16	23.25 ± 8.35	8.72 ± 2.05	11.27 ± 3.74
3-wk group	21.25 ± 1.38	18.48 ± 1.94	8.33 ± 3.85	11.50 ± 1.64
6-wk group	27.57 ± 8.67	25.82 ± 8.26	8.03 ± 3.15	9.33 ± 3.27
9-wk group	34.11 ± 11.56	32.15 ± 11.64	5.89 ± 1.41 ^a	7.67 ± 3.44 ^a
12-wk group	52.38 ± 12.84 ^c	50.11 ± 12.59 ^e	6.82 ± 1.34	10.60 ± 3.51

^a $P < 0.05$, ^c $P < 0.01$, ^e $P < 0.001$ vs the control group. SO: Sphincter of Oddi; CBD: Common bile duct.

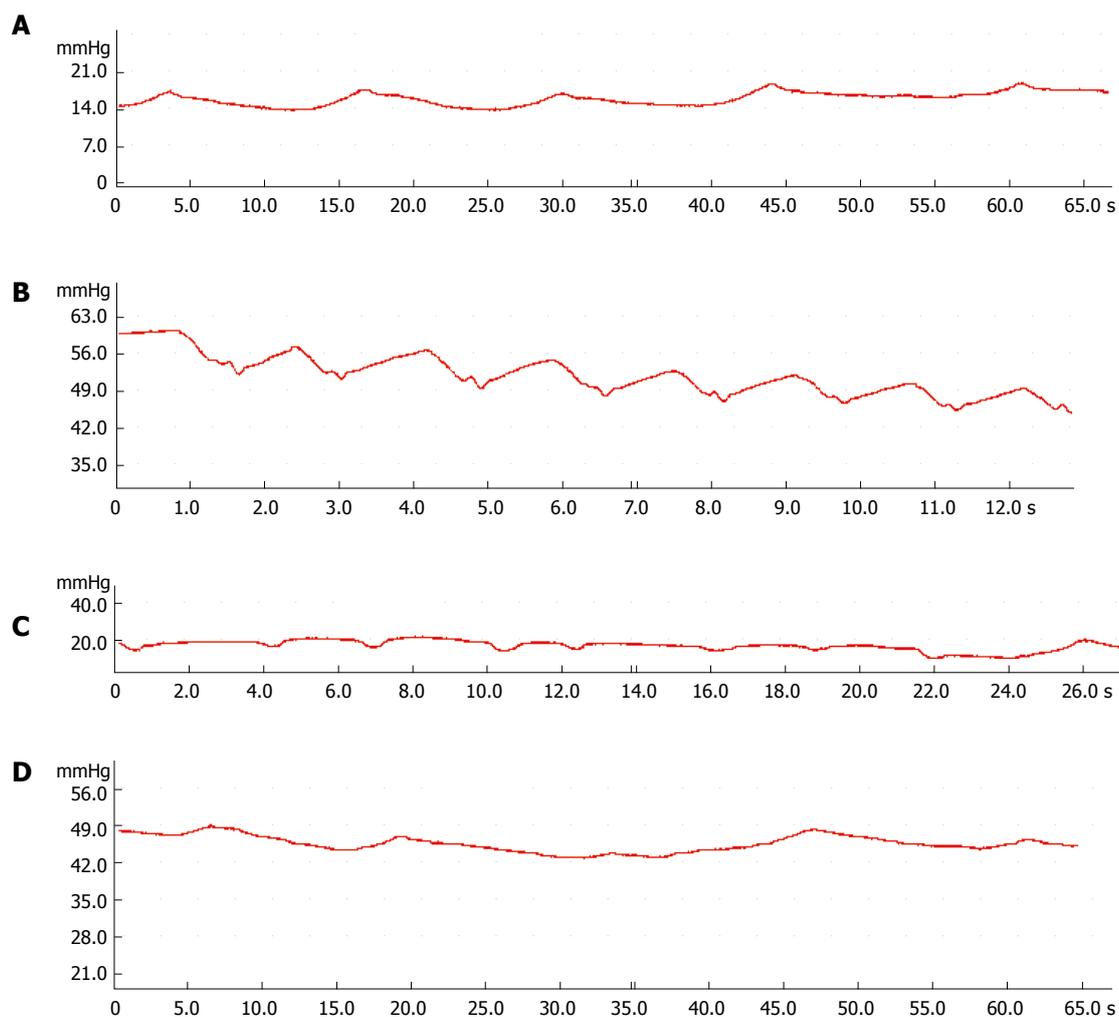


Figure 2 Sphincter of Oddi manometry analysis. A: Sphincter of Oddi (SO) basal pressure in the control group; B: SO basal pressure in the 12 wk group; C: Common bile duct pressure in the control group; D: Common bile duct pressure in the 12 wk group.

SO activity, especially in the relaxed SO. Another measurement reflecting SO function is recording the myoelectric activity of the SO^[20].

In this study, we investigated whether SOD was present and what role it played in the process of cholesterol gallstone formation in guinea pigs. SOM and myoelectric activity of SO were investigated simultaneously at different stages of stone formation. We found that gallstones did not occur until 9 wk, with an incidence rate of 16.7%, increasing to 83.3% after 12 wk. SO myoelectric activity analysis indicated that the frequency and amplitude of myoelectric activity

decreased significantly in the 9 wk group compared to control. Subsequent SO manometry analyses showed the same result: both SO amplitude and SO frequency decreased significantly in the 9 wk group, and the most important indications, the SO basal pressure and common bile duct pressure, increased significantly in the 12 wk group. All these findings are consistent with weakening of the myoelectric activity of SO and gradual increasing of SO tension in the process of gallstone formation. Disturbance of SO motor function impedes the flow of bile into the duodenum, and biliary stasis occurs.

Table 4 Changes in VIP, gastrin and CCK-8 in the process of cholesterol stone formation (pg/mL)

Group	3-wk	6-wk	9-wk	12-wk
VIP				
Control	9.36 ± 2.72	9.30 ± 8.91	6.91 ± 3.14	8.39 ± 0.99
Cholesterol stone	11.83 ± 3.57	7.08 ± 2.31	31.20 ± 7.78 ^a	22.15 ± 2.87 ^c
Gastrin				
Control	17.83 ± 2.35	18.56 ± 5.77	17.42 ± 6.39	19.41 ± 4.58
Cholesterol stone	0.08 ± 0.03 ^c	4.18 ± 2.87	2.16 ± 0.44 ^a	18.92 ± 8.37
CCK-8				
Control	1970.33 ± 439.82	2353.32 ± 13.18	2100.27 ± 29.70	2107.77 ± 171.70
Cholesterol stone	2214.61 ± 174.96	2044.18 ± 147.03	2034.75 ± 138.39	2042.61 ± 265.01

^a*P* < 0.05, ^c*P* < 0.01 vs the control group. VIP: Vasoactive intestinal peptide; CCK-8: Cholecystokinin octapeptide.

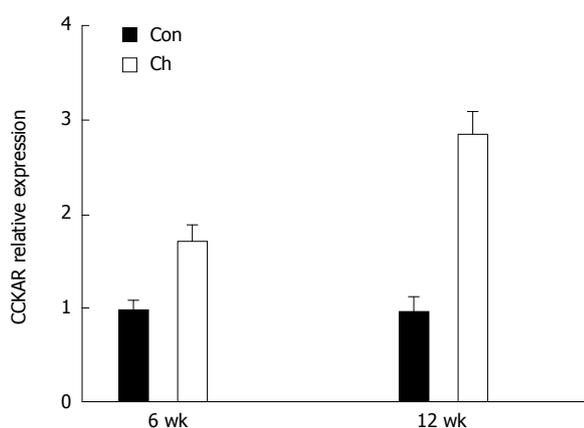


Figure 3 Cholecystokinin-A receptor mRNA expression levels in different groups. CCKAR: Cholecystokinin-A receptor.

The mechanism of cholesterol gallstone-causing diet-induced SOD has not been fully elucidated. Pang *et al.*^[19] considered that the potential mechanisms of hypercholesterolemia (HC)-induced SOD were the intracellular calcium overload and calcium oscillation abnormality. Szilvassy *et al.*^[21] suggested that the impairment of the SO relaxation function was related to the alteration of the nitric oxide signal caused by hypercholesterolemia and hypertriglyceridemia. SO motility was controlled by numerous neurotransmitters and gastrointestinal hormones^[22]. The most important hormone in the regulation of SO function is CCK^[23]. In normal conditions, CCK can contract the gallbladder smooth muscle and reduce tone in the SO to regulate bile flow from the liver through the bile duct into the duodenum. During fasting, hepatic bile enters the gallbladder for storage. Eating initiates gallbladder emptying by neural and hormonal (predominantly CCK) influences. We found serum CCK-8 level was not increased in the 9 and 12 wk groups, meaning that the gallbladder evacuation resistance increased during eating in these groups. Decreases in serum CCK-8 level can cause gallbladder bile stasis, increase the volume of the gallbladder, and induce SOD by increasing the tension of SO.

CCK modulates SO motility mainly *via* excitatory receptors on the smooth muscle and inhibitory

receptors in the neural endings^[24]. A study in cats^[24] showed that the SO relaxation by CCK was abolished by complete denervation induced by tetrodotoxin. Beagle dogs after cholecystectomy^[25] showed an increased CBD pressure, and SO relaxation response to CCK was weakened. These findings were thought to be due to destruction of neural pathways with the operation. In our study, we examined the expression of CCK-AR in the SO using quantitative real-time PCR for the first time. We found that the expression levels of CCK-AR mRNA increased in SO in the 6 and 12 wk groups. The increase in CCK-AR may result in changes in tension of SO, and, therefore, the excreting resistance of the bile may be enhanced significantly.

VIP, an alkaline intestinal peptide composed of 28 amino acids, can relax gallbladder smooth muscle and decrease gallbladder pressure. In the present study, serum VIP level in the 9 and 12 wk groups was higher than that of the control group. We restated the role of VIP in the formation of cholesterol gallstones that had been stressed in our previous study^[12]. We found that the frequency of myoelectric activity decreased markedly in the 9 wk group, the amplitude of myoelectric activity decreased in the 9 and 12 wk groups, and the SO basal pressure and common bile duct pressure increased significantly in the 12 wk group. Elevation of VIP may play an important role in the mechanism of SOD.

Gastrin can increase the SO basal pressure amplitude^[12]. Elevation of serum gastrin may be related to SOD with characteristics of high tension in patients with post-cholecystectomy pain^[26]. However, we found that serum gastrin was decreased significantly in the 3 and 9 wk groups. This effect may have resulted from changes in diet and may not necessarily be related to the formation of gallstones.

In conclusion, we have indicated that a cholesterol gallstone-causing diet may induce SOD, the increasing tension of SO along with its decreasing activity may play an important role in cholesterol gallstone formation, and expression changes in CCK-AR in SO smooth muscle and serum CCK-8 and VIP may be important causes of SOD. The exact mechanism of cholesterol gallstone-causing diet-induced SOD needs further study. Control

of a cholesterol diet and regulation of SO motility are important in the prevention of cholesterol gallstone formation.

COMMENTS

Background

The sphincter of Oddi (SO) may play an important role in gallstone formation because it is the only way by which bile is discharged into the duodenum. However, there is limited research on how the motor function of the SO works and what mechanism by which stones are formed in the process of cholesterol gallstone formation.

Research frontiers

SO manometry is the only method to directly assess the motor function of the SO and is considered the gold standard for assessing SO dysfunction (SOD). However, manometry changes of SO in cholelithiasis are quite controversial. Another measurement reflecting SO function is recording the myoelectric activity of the SO.

Innovations and breakthroughs

This study showed that a cholesterol gallstone-causing diet can induce SOD. The increasing tension of the SO along with its decreasing activity may play an important role in cholesterol gallstone formation. Expression changes of cholecystokinin-A receptor in SO smooth muscle, serum vasoactive intestinal peptide, and cholecystokinin-octapeptide may be important causes of SOD.

Applications

The study results suggest that a cholesterol gallstone-causing diet can induce SOD, and disturbance of SO motility may play a role in gallstone formation. Control of a cholesterol diet and regulation of SO motility are important in the prevention of cholesterol gallstone formation.

Peer-review

This is an interesting paper. The authors describe the results of a basic study in a guinea pig model to investigate the role of SO motility in cholesterol gallbladder stone formation. They concluded that a cholesterol gallstone causing diet may induce increasing tension and decreasing activity of SO, and these effects could play an important role in cholesterol gallstone formation. This work represents a well-conducted basic study that contributes useful information about the role of SO motility in the process of cholesterol gallstone formation and confirms previous limited data that indicated a significant increase in the base pressure of the SO in rabbits with a cholesterol lithogenic diet.

REFERENCES

- 1 **Di Ciaula A**, Wang DQ, Wang HH, Bonfrate L, Portincasa P. Targets for current pharmacologic therapy in cholesterol gallstone disease. *Gastroenterol Clin North Am* 2010; **39**: 245-64, viii-ix [PMID: 20478485 DOI: 10.1016/j.gtc.2010.02.005]
- 2 **Hou L**, Shu XO, Gao YT, Ji BT, Weiss JM, Yang G, Li HL, Blair A, Zheng W, Chow WH. Anthropometric measurements, physical activity, and the risk of symptomatic gallstone disease in Chinese women. *Ann Epidemiol* 2009; **19**: 344-350 [PMID: 19362277 DOI: 10.1016/j.annepidem.2008.12.002]
- 3 **Kong J**, Liu BB, Wu SD, Wang Y, Jiang QQ, Guo EL. Enhancement of interaction of BSEP and HAX-1 on the canalicular membrane of hepatocytes in a mouse model of cholesterol cholelithiasis. *Int J Clin Exp Pathol* 2014; **7**: 1644-1650 [PMID: 24817961]
- 4 **Matyja A**, Gil K, Pasternak A, Sztelfko K, Gajda M, Tomaszewski KA, Matyja M, Walocha JA, Kulig J, Thor P. Telocytes: new insight into the pathogenesis of gallstone disease. *J Cell Mol Med* 2013; **17**: 734-742 [PMID: 23551596 DOI: 10.1111/jcmm.12057]
- 5 **Wang HH**, Portincasa P, Liu M, Tso P, Samuelson LC, Wang DQ. Effect of gallbladder hypomotility on cholesterol crystallization and growth in CCK-deficient mice. *Biochim Biophys Acta* 2010; **1801**: 138-146 [PMID: 19836465]
- 6 **Wei JG**, Wang YC, Du F, Yu HJ. Dynamic and ultrastructural study of sphincter of Oddi in early-stage cholelithiasis in rabbits with hypercholesterolemia. *World J Gastroenterol* 2000; **6**: 102-106 [PMID: 11819533 DOI: 10.3748/wjg.v6.i1.102]
- 7 **Wang XJ**, Wei JG, Wang CM, Wang YC, Wu QZ, Xu JK, Yang XX. Effect of cholesterol liposomes on calcium mobilization in muscle cells from the rabbit sphincter of Oddi. *World J Gastroenterol* 2002; **8**: 144-149 [PMID: 11833091]
- 8 **De Masi E**, Corazziari E, Habib FI, Fontana B, Gatti V, Fegiz GF, Torsoli A. Manometric study of the sphincter of Oddi in patients with and without common bile duct stones. *Gut* 1984; **25**: 275-278 [PMID: 6698444]
- 9 **Chaudhri O**, Small C, Bloom S. Gastrointestinal hormones regulating appetite. *Philos Trans R Soc Lond B Biol Sci* 2006; **361**: 1187-1209 [PMID: 16815798 DOI: 10.1098/rstb.2006.1856]
- 10 **Varga G**, Bálint A, Burghardt B, D'Amato M. Involvement of endogenous CCK and CCK1 receptors in colonic motor function. *Br J Pharmacol* 2004; **141**: 1275-1284 [PMID: 15100163 DOI: 10.1038/sj.bjp.0705769]
- 11 **Xu GQ**, Xu CF, Chen HT, Liu S, Teng XD, Xu GY, Yu CH. Association of caveolin-3 and cholecystokinin A receptor with cholesterol gallstone disease in mice. *World J Gastroenterol* 2014; **20**: 9513-9518 [PMID: 25071346 DOI: 10.3748/wjg.v20.i28.9513]
- 12 **Zhang ZH**, Qin CK, Wu SD, Xu J, Cui XP, Wang ZY, Xian GZ. Roles of sphincter of Oddi motility and serum vasoactive intestinal peptide, gastrin and cholecystokinin octapeptide. *World J Gastroenterol* 2014; **20**: 4730-4736 [PMID: 24782626 DOI: 10.3748/wjg.v20.i16.4730]
- 13 **Portincasa P**, Ciaula AD, Bonfrate L, Wang DQ. Therapy of gallstone disease: What it was, what it is, what it will be. *World J Gastrointest Pharmacol Ther* 2012; **3**: 7-20 [PMID: 22577615 DOI: 10.4292/wjgpt.v3.i2.7]
- 14 **Ruhl CE**, Everhart JE. Gallstone disease is associated with increased mortality in the United States. *Gastroenterology* 2011; **140**: 508-516 [PMID: 21075109]
- 15 **Zhang D**, Xiang J, Wang L, Xu Z, Sun L, Zhou F, Zha X, Cai D. Comparative proteomic analysis of gallbladder bile proteins related to cholesterol gallstones. *PLoS One* 2013; **8**: e54489 [PMID: 23349907 DOI: 10.1371/journal.pone.0054489]
- 16 **Maurer KJ**, Carey MC, Fox JG. Roles of infection, inflammation, and the immune system in cholesterol gallstone formation. *Gastroenterology* 2009; **136**: 425-440 [PMID: 19109959 DOI: 10.1053/j.gastro.2008.12.031]
- 17 **Thune A**, Saccone GT, Scicchitano JP, Toouli J. Distension of the gall bladder inhibits sphincter of Oddi motility in humans. *Gut* 1991; **32**: 690-693 [PMID: 2060879 DOI: 10.1136/gut.32.6.690]
- 18 **Rice JP**, Spier BJ, Gopal DV, Soni A, Reichelderfer M, Pfau PR. Outcomes of sphincter of oddi manometry when performed in low volumes. *Diagn Ther Endosc* 2011; **2011**: 435806 [PMID: 21747651 DOI: 10.1155/2011/435806]
- 19 **Du P**, Cui GB, Wang YR, Zhang XY, Ma KJ, Wei JG. Down regulated expression of the beta1 subunit of the big-conductance Ca²⁺ sensitive K⁺ channel in sphincter of Oddi cells from rabbits fed with a high cholesterol diet. *Acta Biochim Biophys Sin (Shanghai)* 2006; **38**: 893-899 [PMID: 17151783 DOI: 10.1111/j.1745-7270.2006]
- 20 **Li ST**, Chen XW, Zhao HM, Li N, Yan J, Hu ZA. Effects of orexins on myoelectric activity of sphincter of Oddi in fasted rabbits. *Acta Pharmacol Sin* 2006; **27**: 212-216 [PMID: 16412271 DOI: 10.1111/j.1745-7254.2006.00266.x]
- 21 **Szilvássy Z**, Nagy I, Madácsy L, Hajnal F, Velösy B, Takács T, Lonovics J. Beneficial effect of lovastatin on sphincter of Oddi dyskinesia in hypercholesterolemia and hypertriglyceridemia. *Am J Gastroenterol* 1997; **92**: 900-902 [PMID: 9149215]
- 22 **Sonoda Y**, Takahata S, Jabar F, Schlothe AC, Grivell MA, Woods CM, Simula ME, Toouli J, Saccone GT. Electrical activation of common bile duct nerves modulates sphincter of Oddi motility in the Australian possum. *HPB (Oxford)* 2005; **7**: 303-312 [PMID:

- 18333212]
- 23 **Pálvölgyi A**, Sári R, Németh J, Szabolcs A, Nagy I, Hegyi P, Lonovics J, Szilvássy Z. Interplay between nitric oxide and VIP in CCK-8-induced phasic contractile activity in the rabbit sphincter of Oddi. *World J Gastroenterol* 2005; **11**: 3264-3266 [PMID: 15929179 DOI: 10.3748/wjg.v11.i21.3264]
- 24 **Behar J**, Biancani P. Effect of cholecystokinin and the octapeptide of cholecystokinin on the feline sphincter of Oddi and gallbladder. Mechanisms of action. *J Clin Invest* 1980; **66**: 1231-1239 [PMID: 7440712 DOI: 10.1172/JCI109974]
- 25 **Fan MM**, Li F, Zhang XW. Changes of the sphincter of Oddi motility in dog after cholecystectomy. *J Dig Dis* 2012; **13**: 40-46 [PMID: 22188915]
- 26 **Chen XX**, Mo JZ, Liu WZ. [A study on motility of sphincter of Oddi in postcholecystectomy syndrome]. *Zhonghua Nei Ke Zazhi* 1991; **30**: 337-339, 381 [PMID: 1914667]

P- Reviewer: Antonini F, Beltran MA, Tomkin GH **S- Editor:** Yu J
L- Editor: Filipodia **E- Editor:** Zhang DN



Case Control Study

Comprehensive risk assessment for early neurologic complications after liver transplantation

Si-Yuan Wu, Teng-Wei Chen, An-Chieh Feng, Hsiu-Lung Fan, Chung-Bao Hsieh, Kuo-Piao Chung

Si-Yuan Wu, Teng-Wei Chen, An-Chieh Feng, Hsiu-Lung Fan, Chung-Bao Hsieh, Division of Organ Transplantation Surgery, Department of Surgery, Tri-Service General Hospital, National Defense Medical Center, Taipei 11490, Taiwan

Kuo-Piao Chung, Chung-Bao Hsieh, Institute of Health Policy and Management, College of Public Health, National Taiwan University, Taipei 11490, Taiwan

Author contributions: Feng AC, Hsieh CB, Fan HL and Chen TW performed the research; Chung KP participated in the design of the study and performed the statistical analysis; Wu SY, Feng AC and Hsieh CB wrote the paper; all authors read and approved the final manuscript.

Supported by Tri-Service General Hospital, No. TSGH-C104-159.

Institutional review board statement: The study was approved by Institutional Review Board I of Tri-Service General Hospital, National Defense Medical Center (TSGHIRB No.100-05-220).

Informed consent statement: All transplantation procedures in the study were performed after approval of the Ethics Committee and informed consent from patients; Data were gathered retrospectively after approval of the IRB.

Conflict-of-interest statement: The authors declare no conflicts of interest.

Data sharing statement: Technical appendix, original data, and statistical code of manuscript are available from the corresponding author at albert0920@yahoo.com.tw.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Correspondence to: Chung-Bao Hsieh, MD, PhD, Division of Organ Transplantation Surgery, Department of Surgery, Tri-Service General Hospital, National Defense Medical Center, No. 325, Cheng-Kung Rd, Sec2, Neihu 11490, Taipei, Taiwan. albert0920@yahoo.com.tw
Telephone: +886-933980018
Fax: +886-2-87927372

Received: March 11, 2016
Peer-review started: March 11, 2016
First decision: April 14, 2016
Revised: April 26, 2016
Accepted: May 21, 2016
Article in press: May 23, 2016
Published online: June 28, 2016

Abstract

AIM: To determine risk factors for early neurologic complications (NCs) after liver transplantation from perspective of recipient, donor, and surgeon.

METHODS: In all, 295 adult recipients were enrolled consecutively between August 2001 and February 2014 from a single medical center in Taiwan. Any NC in the first 30 d post-liver transplantation, and perioperative variables from multiple perspectives were collected and analyzed. The main outcome was a 30-d NC. Generalized additive models were used to detect the non-linear effect of continuous variables on outcome, and to determine cut-off values for categorizing risk. Risk factors were identified using multiple logistic regression analysis.

RESULTS: In all, 288 recipients were included, of whom 142 (49.3%) experienced at least one NC, with encephalopathy being the most common 106 (73%). NCs prolonged hospital stay (35.15 ± 43.80 d vs 20.88 ± 13.58 d, $P < 0.001$). Liver recipients' age < 29 or ≥ 60 years, body mass index < 21.6 or $>$

27.6 kg/m², Child-Pugh class C, history of preoperative hepatoencephalopathy or mental disorders, day 7 tacrolimus level > 8.9 ng/mL, and postoperative intra-abdominal infection were more likely associated with NCs. Novel risk factors for NCs were donor age < 22 or ≥ 40 years, male-to-male gender matching, graft-recipient weight ratio 0.9%-1.9%, and sequence of transplantation between 31 and 174.

CONCLUSION: NCs post- liver transplantation occurs because of factors related to recipient, donor, and surgeon. Our results provide a basis of risk stratification for surgeon to minimize neurotoxic factors during transplantation.

Key words: Risk; Liver transplantation; Neurotoxicity syndromes; Donor; Learning curve

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: The study uses generalized additive models and logistic regression in statistics to control confounders in the case-control study. We identified 11 risk factors for early neurologic complication after transplantation. From liver recipients' perspective, age < 29 or ≥ 60 years, body mass index < 21.6 or > 27.6 kg/m², Child-Pugh class C, history of preoperative hepatoencephalopathy or mental disorders, day 7 tacrolimus level > 8.9 ng/mL, and postoperative intra-abdominal infection were at risk. From donors' perspective, age < 22 or ≥ 40 years, male-to-male gender matching, graft-recipient weight ratio 0.9%-1.9% was at risk. From surgeons' perspective, sequence of transplantation between 31 and 174 were at risk.

Wu SY, Chen TW, Feng AC, Fan HL, Hsieh CB, Chung KP. Comprehensive risk assessment for early neurologic complications after liver transplantation. *World J Gastroenterol* 2016; 22(24): 5548-5557 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5548.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5548>

INTRODUCTION

Neurologic complications (NCs) are common after solid organ transplantation, especially liver transplantation^[1]. While mortality rates decreased with advances in surgical technique and postoperative care in recent years, NCs continue to affect more than one-third of transplanted patients and cause significant morbidity, mortality, and prolonged hospital stay^[2,3]. The liver recipient is vulnerable to NCs since many patients have preoperative hepatic encephalopathy, which is a well-known risk factor^[3-6]. Moreover, the unfavorable condition of recipients including metabolic, nutritional, and electrolyte imbalances may predispose them to

NCs^[7,8]. Generally, NCs develop early, with more than 80% of patients developing NCs within 30 d after liver transplantation^[6,9]. Encephalopathy is the most frequent etiology^[3,5,9-12]. As it is multifactorial in nature, the transplant team should identify patients at risk and avoid predisposing variables during the perioperative period to minimize its incidence.

In the last decade, the risk factors and mechanism of NCs have been investigated in several studies, most of which were retrospective^[3,6,9-12]. The majority of these focused on recipients' factors depending on timing of transplant, such as presence of hepatic encephalopathy, etiology of cirrhosis before transplant, hyponatremia, cerebrovascular insult during transplant, immunosuppressant toxicity and central nervous system infection after transplant. However, the nature of liver graft, donor-recipient matching, and experience of surgeon and transplant team also impact outcome, and few studies have investigated NCs after liver transplantation from these perspectives. No study showed how the accumulated experience of the transplant team influences NC development. Besides, the non-linear effect of certain continuous variables, such as age, body mass index (BMI) and graft-recipient weight ratio (GRWR), was not considered in the statistical analysis of prior studies. Our study aimed to identify new risk factors of 30-d NCs after liver transplantation from multiple perspectives. In particular, we performed statistical analyses to explore how the non-linear effect of certain continuous variables increase NC risk.

MATERIALS AND METHODS

Patients

From August 2001 to February 2014, 295 consecutive adult liver transplantation surgeries were performed in the Tri-Service General Hospital (Taipei, Taiwan). All causes of mortality were included in the study except for surgical mortality unrelated to NCs. All procedures were performed after approval of the Ethics Committee. The transplant team included 2 qualified transplant surgeons, 2 fellows and 2 senior residents. The immunosuppressant induction protocol consisted of intravenous methylprednisolone, starting with a large bolus dose during the anhepatic phase, followed by daily tapering off until postoperative day 5, and triple oral immunosuppressants (tacrolimus, mycophenolate mofetil, steroids). Serum trough levels of tacrolimus were checked daily in the first postoperative week to maintain a level of 6-10 ng/mL. If basiliximab was included, the first 20 mg were given during anhepatic phase and the second dose on postoperative day 4, along with halving the steroid dose. Diagnoses of NCs were primarily made by the transplant team based on clinical examination, and ancillary examinations such as computed tomography, magnetic resonance imaging, and electroencephalography. Neurologists were consulted for major NCs. Only new onset of

postoperative neurologic disorders was regarded as NCs. The clinical data and patients' outcomes were retrospectively reviewed from medical charts. The final follow-up was conducted until March 31, 2014.

Data collection

The medical records and transplant database were reviewed after approval of the Institutional Review Board I of Tri-Service General Hospital, National Defense Medical Center (TSGHIRB No.100-05-220). Data were collected from 4 major perspectives: recipient, donor, donor-recipient match, and surgeon. The recipient-related variables were demographic details, comorbidities other than liver diseases, primary liver disease requiring liver transplantation, preoperative Child-Pugh and Model for End-Stage Liver Disease (MELD) scores, any complications of end-stage liver disease, such as ascites, hepatic encephalopathy, variceal bleeding and hepatorenal syndrome. In addition, any mental disorders related to alcoholism or encephalopathy were recorded. The perioperative variables included intraoperative blood loss, length of the procedure, complex vascular anatomy, and splenectomy. The postoperative variables included dose and serum level of tacrolimus, inclusion of basiliximab, and blood chemistry parameters (ammonia, sodium, and magnesium) on postoperative day 7. In addition, any postoperative complications as acute rejection, intra-abdominal infection, or kidney injury requiring dialysis were recorded. The donor and donor-recipient matching-related variables were graft's type and weight, gender match, GRWR, and ABO compatibility. The surgeon variables were surgeon in charge, and the chronological sequence of transplantations.

Outcomes' evaluation

The primary outcome was occurrence of any NCs within 30 d after liver transplantation. We adapted the NC classification from Dhar *et al.*^[3] as follows: (1) encephalopathy: delirium, psychosis, or alteration of conscious level diagnosed after excluding specific lesions of the central nervous system; (2) seizures; (3) drug neurotoxicity: symptoms subsiding after reduction or discontinuation of immunosuppressants, with severity varying from tremors to imaging-confirmed posterior reversible encephalopathy syndrome; (4) cerebrovascular insults; (5) central nervous system infection; and (6) central pontine myelinolysis: acute para- or quadriparesis, dysphagia, dysarthria, diplopia, loss of consciousness with evident change in serum sodium levels. Patients experiencing "locked-in syndrome", that is, awake but unable to move or communicate, were categorized into "central pontine myelinolysis" if rapid changes in serum sodium level coexisted. If the definite etiology could not be determined, they were recorded separately.

Other outcome data were hospital days and complications other than NCs during the first month. In addition, the complications were classified into

minor and major according to the clinical finding and its severity. Minor complications included those that improved spontaneously without sequela within 1 mo, while the others causing functional deficit, brain damage or death were considered as major complications.

Statistical analysis

Statistical analysis was performed using the R 3.1.0 software (R Foundation for Statistical Computing, Vienna, Austria). A *P*-value of < 0.05 was considered statistically significant.

Descriptive statistics were used to express data as mean \pm SD for continuous variables, and frequency and percentage for categorical variables. Differences between the two groups, patients developing NCs and those who did not, were analyzed using Wilcoxon's rank-sum test for continuous variables, and Fisher's exact test for categorical variables.

Some continuous variables had non-linear effect, such as age and BMI. The regression model may have described poor correlation between these variables and the outcome since linearity was usually assumed during the analysis. Generalized additive models (GAMs)^[13] for binary response (patients developing NCs and those who did not) were fitted to detect the potential nonlinear effects of continuous covariates; if nonlinearity existed, appropriate cut-off point(s) for discretizing the continuous covariate were selected in the GAM plots. This procedure was carried out using the *vgam* function (with the default values of smoothing parameters) of the VGAM package in R.

Multiple logistic regression analysis was conducted by fitting a generalized linear model to estimate the effects of predictors on the occurrence of NCs. First, all variables, donor-recipient gender match combinations, and new categorical variables obtained from previous cut-off points for discretizing continuous variables by GAM were selected. Next, a step-wise variable selection procedure went through iteration between the forward and backward steps with both the significant levels for entry and for stay set to 0.15 for being conservative. Then, the best candidate final logistic regression model was identified manually by dropping the covariates with *P* value > 0.05 one at a time until all regression coefficients were significantly different from 0. The final fitted logistic model was assessed with 3 goodness of fit measures: the estimated area under the receiver operating characteristic curve (≥ 0.7 acceptable), adjusted generalized R^2 (≥ 0.30 acceptable), and the Hosmer-Lemeshow test (*P* ≥ 0.05 , larger means better fit). In addition, the variance inflation factor was checked for multicollinearity (no more than 10 for continuous covariates and 2.5 for categorical covariates).

RESULTS

Among the 295 patients, 7 with surgical mortality

Table 1 Baseline characteristics of the 288 consecutive liver transplantation patients

Recipient variables	
Demographics	
Age (yr)	52.3 ± 9.81
Gender (M/F)	213/75
BMI (kg/m ²)	24.4 ± 3.72
Primary diagnosis, <i>n</i> (%)	
Hepatitis B	175 (60.8)
Hepatitis C	73 (25.3)
Alcoholic liver disease	77 (26.7)
HCC	138 (47.9)
Severity of liver disease	
Child-Pugh score	10.0 ± 2.49
MELD score	15.0 ± 9.01
Complication, <i>n</i> (%)	
Hepatic encephalopathy	132 (45.8)
Ascites	189 (65.6)
Variceal bleeding	114 (39.6)
Comorbidities, <i>n</i> (%)	
Diabetes mellitus	103 (35.7)
Hypertension	52 (18.1)
Uremia	14 (4.9)
Mental disorders	99 (34.4)
Blood test before transplant	
Glucose (mg/dL)	129 ± 76.07
Albumin (g/dL)	3.08 ± 0.66
Creatinine (mg/dL)	1.22 ± 1.53
INR	1.62 ± 0.99
T. bilirubin (mg/dL)	7.78 ± 11.19
Platelet (× 10 ³ /μL)	85.4 ± 53.85
Ammonia (μg/dL)	138.0 ± 97.05
Perioperative variables	
Blood loss (mL)	2896 ± 3409
Operation time (min)	575 ± 122.7
Splenectomy, <i>n</i> (%)	87 (30.2)
Complex vascularity, <i>n</i> (%)	75 (26.0)
Postoperative variables	
Tacrolimus dose (mg/d)	3.4 ± 1.67
Basiliximab, <i>n</i> (%)	51 (18)
Serum level - Day 7	
Tacrolimus level (ng/mL)	7.4 ± 6.24
Ammonia (μg/dL)	69.6 ± 130.8
Sodium (mmol/L)	136 ± 4.92
Magnesium (mEq/L)	1.73 ± 0.34
Donor variables	
Age (yr)	33 ± 11.21
Male gender, <i>n</i> (%)	171 (59.4)
Graft weight (g)	872 ± 357.88
Graft type	95/47/142/4
(whole/left/right/S67), <i>n</i> (%)	(33.0/16.3/49.3/1.4)
Donor-recipient matching	
GRWR (%)	1.34 ± 0.57
ABO incompatible, <i>n</i> (%)	9 (3.1)
Gender match	128/44/86/30
(MM/MF/FM/FF), <i>n</i> (%)	(44.4/15.3/29.9/10.4)
Surgeon variables	
Surgeon A/B/C, <i>n</i> (%)	234/51/3 (81/18/1)
Sequence of transplantation	1-288

BMI: Body mass index; HCC: Hepatocellular carcinoma; MELD: Model for end-stage liver disease; GRWR: Graft-recipient weight ratio.

unrelated to NCs were excluded: 2 failed to awake due to irreversible brain damage secondary to fulminant hepatitis, and 5 experienced primary graft failure. Finally, 288 patients were enrolled in this study, with 95

Table 2 Type of complication after liver transplantation *n* (%)

	Event	Minor/major
Neurologic complications		
Encephalopathy	145 (100)	97 (67)/48 (33)
Delirium	106 (73)	88 (83)/18 (17)
Psychosis	75 (52)	64 (85)/11 (15)
Change in consciousness	5 (4)	4 (80)/1 (20)
Seizures	26 (18)	20 (77)/6 (23)
Cerebrovascular events	10 (7)	0 (0)/10 (100)
Drug neurotoxicity	5 (4)	0 (0)/5 (100)
Locked-in syndrome	10 (7)	5 (50)/5 (50)
Central pontine myelinolysis	12 (8)	4 (33)/8 (67)
Other complications	2 (1)	0 (0)/2 (100)
Acute kidney injury	1	
Intra-abdominal infection	9	
Graft failure	2	
Reoperation	31	
Acute rejection	61	
Hepatitis B recurrence	10	
Tuberculosis infection	8	
Cytomegalovirus infection	7	
Total complications	85	

receiving deceased donor liver transplantation (DDLT) and 193 receiving living donor liver transplantation (LDLT). Baseline characteristics of the study population are shown in Table 1.

The outcome data are shown in Table 2. One hundred and forty two patients experienced 145 events of NCs, with an overall incidence of 49.3%. The most frequent events were encephalopathy (*n* = 106, 73%), including delirium (*n* = 75, 52%), conscious change (*n* = 26, 18%) and psychosis (*n* = 5, 4%). The average hospital stay was significantly longer in those with postoperative NCs (35.15 ± 43.80 d vs 20.88 ± 13.58 d, *P* < 0.001).

Variables associated with NCs

The differences between the patients with NC or not are shown in Table 3. Both groups were similar in demographics of recipient and donor, liver graft type, surgeon and sequence of transplantation. For primary diagnosis, alcoholic liver cirrhosis was more common in the NC group (*P* = 0.001), while hepatocellular carcinoma was prevalent in the control group (*P* < 0.001). The NC group included patients with more severe liver disease before transplant, more preoperative mental disorders and hepatic encephalopathy, more intraoperative complex vascular anatomy, higher day 7 tacrolimus level, more postoperative complications of acute rejection, intra-abdominal infection, and kidney injury requiring dialysis.

GAMs were fitted to continuous variables with potential non-linear effect on outcome. The selected GAM plots for continuous variables on the NC group are shown in Figure 1. According to the GAM plots, the following scales of variables were associated with higher probability of NCs: age < 29 or ≥ 60, recipient BMI < 21.6 or > 27.6 kg/m², Child-Pugh score >

Table 3 Comparison of variables from multiple perspectives between the neurologic complications and control groups

	NC (n = 142)	No NC (n = 146)	P value
Recipient variables			
Preoperative			
Age (yr)	51.75 ± 10.51	52.77 ± 9.09	0.567
Gender (M/F)	109/33	104/42	0.347
BMI (kg/m ²)	24.65 ± 4.25	24.26 ± 3.11	0.617
Hepatitis B	86	89	1.000
Hepatitis C	30	43	0.136
Alcoholic liver disease	50	27	0.001
HCC	51	87	< 0.0001
Child-Pugh score	10.26 ± 2.26	8.27 ± 2.32	< 0.0001
MELD score	20.3 ± 9.4	14.0 ± 7.4	< 0.0001
Hepatic encephalopathy	91	41	< 0.0001
Variceal bleeding	51	63	0.229
Ascites	106	83	0.002
Mental disorder	66	33	< 0.0001
Serum Albumin (g/dL)	2.95 ± 0.57	3.21 ± 0.72	0.01
Serum T. bilirubin (mg/dL)	10.91 ± 13.16	4.74 ± 7.79	< 0.0001
Perioperative			
Blood loss (mL)	3329 ± 3953	2474 ± 2728	0.081
Operation time (min)	558.98 ± 99.63	579.9 ± 127.0	0.160
Complex vascularity	45	30	0.033
Postoperative			
Day 7 tacrolimus level (ng/mL)	8.23 ± 7.42	6.54 ± 4.69	0.023
Acute rejection	38	23	0.030
Intra-abdominal infection	32	12	< 0.0001
Kidney injury requiring dialysis	18	7	0.021
Donor variables			
Donor age (yr)	32.5 ± 11.4	33.6 ± 11.0	0.234
Donor gender (M/F)	89/53	82/64	0.282
Graft type (whole/left/right/S67)	45/21/74/2	50/26/68/2	0.827
Donor-recipient matching			
GRWR	1.33 ± 0.56	1.35 ± 0.57	0.726
Surgeon variables			
Surgeon A/B/C	116/25/1	118/26/2	0.855
Sequence of transplantation	95.35 ± 69.66	108.12 ± 74.41	0.186

NCs: Neurologic complications; HCC: Hepatocellular carcinoma; MELD: Model for end-stage liver disease; GRWR: Graft-recipient weight ratio; BMI: Body mass index.

9, MELD score between 11 and 20, serum glucose > 8 mmol/L, serum albumin between 2.9 and 4.2 g/dL, serum creatinine between 0.6-1.6 mg/dL, the logarithm of INR > 0.3, platelet count < 70 × 10³/μL, serum ammonia > 268 μg/dL, day 7 tacrolimus level > 9 ng/mL, day 7 serum ammonia level > 98 μg/dL, day 7 serum sodium level > 143 mmol/L, day 7 serum magnesium level > 1.8 mEq/L, donor age < 22 or ≥ 40 years, GRWR between 0.9% and 1.9%, and sequence of transplantation between 31 and 174. New categorical variables for regression analysis were obtained after grouping these continuous variables.

After stepwise multiple logistic regression analysis, 12 independent variables were identified as significant in the final logistic regression model (Table 4). Eleven positive predictors of NCs were: recipient age < 29

or ≥ 60 years, BMI < 21.6 or > 27.6 kg/m², Child-Pugh score > 9, preoperative hepatic encephalopathy, history of mental disorder, day 7 tacrolimus level > 8.9, postoperative intra-abdominal infection, donor age < 22 or ≥ 40 years, male-to-male gender match, GRWR between 0.9% and 1.9%, and sequence of transplantation between 31 and 174. On the other hand, recipients with history of variceal bleeding were less likely to develop NCs (OR = 0.431; 95%CI: 0.221-0.821). The assessment of final logistic regression model showed fair goodness-of-fit (area under the receiver operating characteristic curve = 0.8553 > 0.7 with 95%CI: 0.8119-0.8988; adjusted R² = 0.471 > 0.3; Hosmer-Lemeshow goodness-of-fit test P = 0.1446 > 0.05). Furthermore, variance inflation factors for each covariate in the selected final logistic regression model were between 1.075 and 1.582, indicating no multicollinearity.

DISCUSSION

Our results confirmed the prevalence of NCs following liver transplantation, with three-quarters of patients developing encephalopathy. The risk factors were not only from recipient's perspective, but also from perspectives of donor, donor-recipient match and surgeon. Since randomization of subjects is hardly possible in critical conditions as those requiring liver transplantation, studies in this area almost have case-control design. The novelty we have in this study came from better control of confounding effects by flexible statistical tools. The GAM plots enabled more proper stratification of continuous variables on outcome, and the regression analysis controlled the confounding bias by considering multiple covariates all at a time.

The incidence of NCs in the study was relatively higher than in other reports (49.3% vs 20%-30%) because of more diagnoses of minor encephalopathy (88/145, 60.7% of overall NC events). In fact, the diagnostic criteria for encephalopathy are not universal among physicians, especially for minor degrees. In our institution we tend to broaden the diagnostic criteria to include any transient delirium, psychosis or consciousness' level change. This enables us to correct metabolic disorders and use immunosuppressants more properly. Our categorical system here is limited by not differentiating anoxic, septic or metabolic etiologies^[14] accounting for intra-abdominal infection as an independent risk factor (OR = 5.193, 95%CI: 2.114-13.67).

Preoperative hepatic encephalopathy and mental disorders significantly increased the risk of NCs, with an OR 2.432 and 2.517, respectively. Hepatic encephalopathy, both episodic and active before the operation, is a well-known risk factor^[3,4,6,12,14]. It is hypothesized that excess serum ammonia in end stage liver disease interferes with cerebral metabolism, and the condition is not immediately reversed after

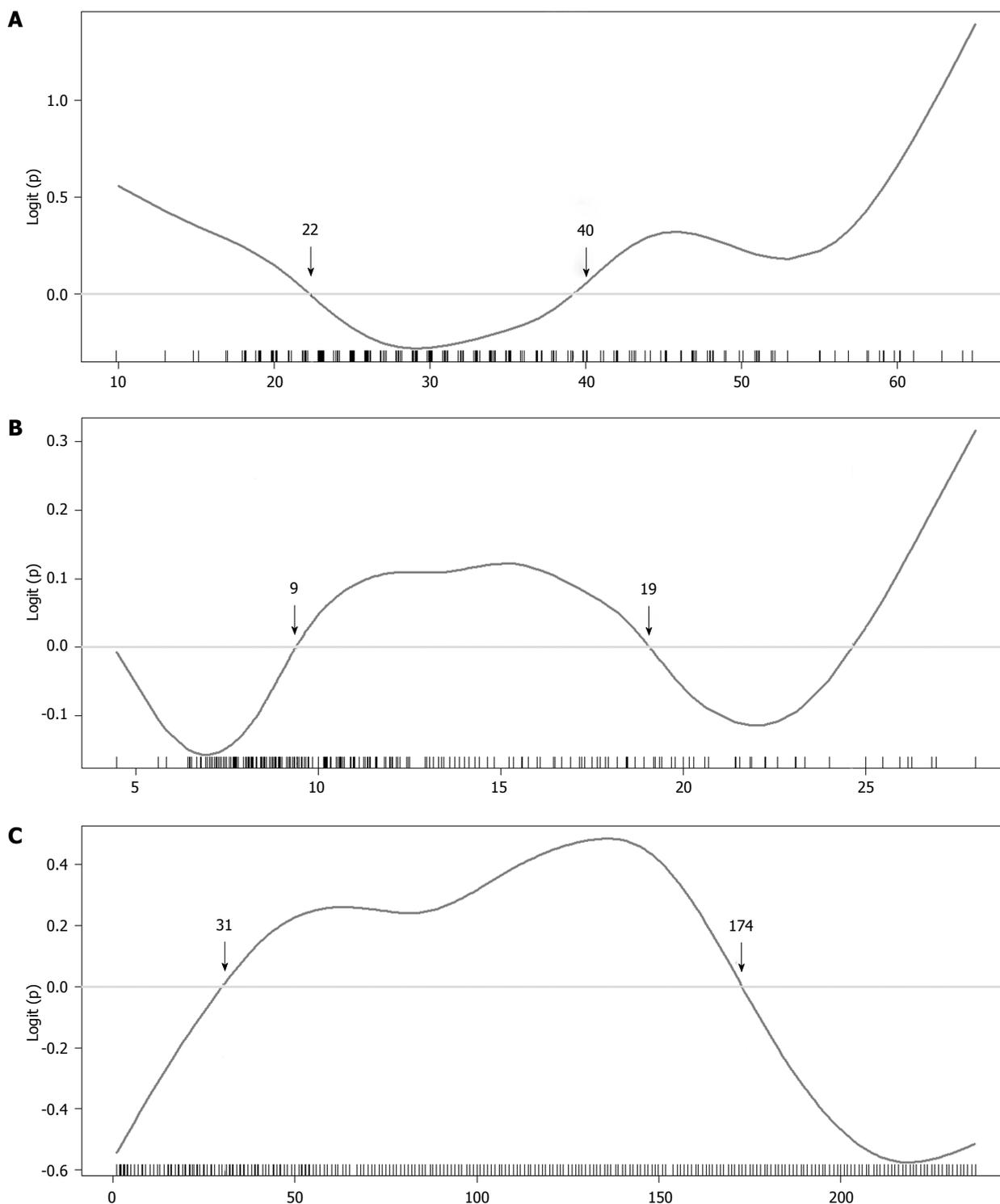


Figure 1 Influence of continuous covariates on neurologic complications resulting from fitting a generalized additive model to the data. A: Donor age; B: Graft-recipient weight ratio; C: Sequence of transplantation.

transplantation. Several investigators have emphasized the importance of comprehensive preoperative neurologic examinations including neuropsychiatric and neuromuscular assessments.

In addition to hepatic encephalopathy, chronic alcohol abuses also contribute to neurotoxicity, which impairs cognitive function especially memory. Alcoholism also renders patients at risk for thiamine

deficiency. A prospective study that performed neuropsychological assessments before and after liver transplantation found that recipients with alcoholic etiology had poorer cognitive indexes in memory; in addition, a multivariate analysis determined that alcohol etiology, diabetes mellitus, and hepatic encephalopathy were predictors of poor global cognitive function after transplantation^[15]. However,

Table 4 Multiple logistic regression model for neurologic complications

	OR	95%CI	P value
Recipient variables			
Preoperative			
Age < 29 or ≥ 60 yr	2.071	1.024-4.272	0.045
BMI < 21.6 or > 27.6 kg/m ²	1.877	1.007-3.552	0.049
Child-Pugh score > 9 (Class C)	1.509	1.288-1.790	< 0.001
Hepatic encephalopathy	2.432	1.232-4.860	0.011
Variceal bleeding	0.431	0.221-0.821	0.012
Mental disorder	2.517	1.279-5.064	0.008
Postoperative			
Day 7 tacrolimus level > 8.9 ng/mL	1.131	1.068-1.205	< 0.001
Intra-abdominal infection	5.193	2.114-13.67	< 0.001
Donor variable			
Donor age < 22 or ≥ 40 yr	2.245	1.207-4.271	0.012
Donor-recipient matching			
Male to male gender match	2.36	1.266-4.506	0.008
0.9% < GRWR < 1.9%	1.95	1.069-3.624	0.032
Surgeon variable			
31 ≤ Sequence of transplantation < 174	2.773	1.479-5.363	0.002

NCs: Neurologic complications; BMI: Body mass index; MELD: Model for end-stage liver disease; GRWR: Graft-recipient weight ratio.

in line with the result of recent studies on LDLT^[10,14,16], the current study, using regression analysis adjusted by other covariates, did not find alcoholic liver disease to be a predictor of NCs. The discrepancy may be related to the multifactorial nature of neurologic complications, and it is difficult to attribute to a specific cause of NC in patients with alcoholic liver disease. As a result, clinicians believe that hepatic encephalopathy, which is prevalent in cirrhotic patients, could be the most common cause of NC. A controlled study with the exact definition of alcohol abuse, complete neurologic examination before and after transplantation, standard postoperative care, and exclusion of postoperative complications confounding the result is needed to answer the question.

On the other hand, recipients with history of variceal bleeding were not likely to have NCs after controlling of potential confounders in the regression model. It was somewhat difficult to explain. One possible reason was that our recipients with variceal bleeding tended to receive living donor liver transplantation earlier than those without did, which enabled transplant proceeding under less severe liver disease. Another reason might be related to the shunt ligation procedure which we often done during transplantation blocking portosystemic encephalopathy.

Moreover, patients with NCs had more severe liver disease before transplant. Child-Pugh class C was an independent risk factor, with an odds ratio of 1.509 (95%CI: 1.288-1.790), while MELD score was not when adjusted by serum creatinine level and other factors in regression analysis. These results were consistent with data obtained in previous studies. In a retrospective study by Dhar *et al.*^[3] investigating factors associated with NCs, preoperative Child-Pugh Class

C was a significant variable in the univariate analysis, while only active preoperative hepatic encephalopathy was significant in the multivariate analysis. Another retrospective study by Kanwal *et al.*^[17] used age, sex and era-matched control group to identify risk factors of post-transplant mental status change. MELD score > 15 was an independent risk factor in both univariate and multivariate analyses and was one of the four factors included in the prediction model. These results are likely to be related to encephalopathy precipitated by higher serum level of endogenous neurotoxic substances (including ammonia) in patients with severe liver disease. Besides, the frequent changes in electrolyte levels, malnutrition and metabolic disorders in decompensated cirrhosis may result in an unfavorable postoperative environment, rendering liver recipients vulnerable to postoperative neurologic disorders.

Age < 29 or ≥ 60 years and BMI < 21.6 or ≥ 27.6 kg/m² showed increased probability of NCs in the GAM plot, and both were independent predictors in multiple logistic regression model, with odds ratios of 2.071 and 1.877, respectively (Table 4). Advanced age, preoperative cognitive impairment and multiple medical comorbidities were known risk factors for postoperative delirium after various procedures in several studies^[18-20]. We believe that age ≥ 60 years had greater impact on NCs since the mean recipients' age in our study was 52.3 ± 9.81 years, without pediatric recipients.

Underweight and overweight liver recipients had a significant risk for NCs. Extreme BMI values (< 18.5 and ≥ 40 kg/m²) are known risk factors for mortality after liver transplantation^[21]. Patients are also more likely to develop infectious complications owing to malnutrition, as well as prolonged treatment time, and NCs secondary to of vitamins' and trace elements' deficiencies^[22]. Early diagnosis and prompt treatment of nutritional deficits before transplantation may ameliorate encephalopathy and optimize transplant outcome^[22,23].

In the GAM plot for postoperative variables, we found that day 7 tacrolimus level > 8.9 ng/mL, serum ammonia level > 98 μg/dL, serum sodium level > 143 mmol/L, serum magnesium level > 1.8 mEq/L were associated with higher probabilities for NCs. In the final logistic regression model, only day 7 tacrolimus level was an independent variable. This is in line with prior study^[3].

Neurotoxicity during induction of immunosuppressants, mostly induced by calcineurin inhibitors (CNIs), is common in the early postoperative period. Clinical manifestations may vary from tremors, headache, and visual disturbances to altered mental status. In addition, seizures after transplantation are most often caused by drug neurotoxicity. The incidence is associated with high serum levels of CNIs while the occurrence is not excluded by normal serum CNI level. Coexisting hypomagnesemia and hypocholesterolemia

are risk factors. Rarely, patients using CNIs have posterior reversible encephalopathy syndrome, a radiographic diagnosis characterized by reversible vasogenic edema of the white matter involving the posterior circulation territory on serial magnetic resonance imaging. It is likely to occur in patients with concomitant alcoholic liver disease and infection/sepsis. Management is mainly dose reduction and shift to an immunosuppressant with another mechanism of action; however, these patients may be at risk of acute rejection from the lower maintenance dose^[24]. The process of optimizing immunosuppression, aiming to minimize graft rejection and avoid neurotoxicity, may need multidisciplinary teamwork and accumulated experience.

From the donor perspective, variables of graft type, donor age, donor gender, and GRWR were not significantly different between the NC and control groups. While age and GRWR had a biphasic effect on the GAM plots (Figure 1), we adjusted the cut-off values (donor age < 22 or > 40 years, and 0.9% < GRWR < 1.9%) and found that both variables were predictors in the regression analysis. In the literature, few studies investigated the influence of donor and donor-recipient match factors on NCs. Study from the SRTR database have shown an increased risk of graft failure with donor age > 40 years^[25]. Prior studies on NC risk factors including donor age and GRWR showed no significant difference of these variables^[5,11]. In contrast, our study indicated that risk of NC depends on either extreme donor age, or improper GRWR. In the GAM plots, we could see the probability of NCs decreasing when GRWR was above 1.9, which could suggest better detoxification capabilities of larger grafts. Besides, small size grafts are not likely to increase NC risk, suggesting non-inferiority of small graft when graft inflow is properly controlled^[26].

Male-to-male gender match was another independent risk factor for NCs in this study. The association between donor-gender match and neurologic complications has not been reported previously. The impact of gender mismatch on the outcome of liver transplantation is still controversial. Several studies demonstrated that female-to-male gender match is associated with negative outcomes in kidney, heart, lung, and liver transplantations, possibly due to the effect of estrogen and the relative small-for-size of female grafts^[27-30]. It is unclear whether other confounding variables specific to male gender play a role, such as prevalent alcoholic liver diseases. Further prospective, controlled studies are needed to confirm the gender match effect on NCs.

The sequence of transplantation was a notable finding. The risk of NCs decreased after 174 transplantations by the team, and the order between 31 and 174 was an independent risk factor for NC development. The lower risk in the first 30 cases could be related to strict patient selection and relatively conservative strategy of immunosuppression. Learning curve

effect in liver transplantation has been described, which is affected by the volume of the center or the year of transplant^[31-34]. For successful liver transplantation, multidisciplinary team work is needed. The improvement is important, not only in the surgical technique but also in donor selection, timely decision for surgery, standardized postoperative care and optimal immunosuppression. Compared to graft survival rate commonly used in studies with a learning curve, NCs are more likely to be related to non-operative factors^[35,36]. It is the environment influenced by perioperative risk factors from all perspectives that results in complications. To avoid NCs, the transplant team may need risk stratification during patient selection and donor matching, correcting risk factors before the operation as much as possible, and should remain watchful during the perioperative period. Ideal immunosuppression aims at minimizing acute rejection and avoids neurotoxicity. It is crucial for the transplant surgeon to recognize this process of development and to conduct interdisciplinary learning as well as continuously improving the patient care quality of the team.

This study has limitations owing to its retrospective design. First, the complications included were those that were identified and reported by the clinician. While serious complications are rarely excluded, clinicians may have missed minor complications that resolved spontaneously without treatment. Second, NCs often manifests as one or more signs, but it is possible that only the most serious ones were documented, leading to misclassification. Third, patients in the control group might have shared features and exposures with those in the NC group. Even if we have considered as many potential important variables as we could, residual confounders are still possible. Despite these limitations, the risk factors identified in the study could be subjective for validation in further prospective, randomized studies.

In conclusion, NCs after liver transplantation occur frequently and affect almost half of liver recipients, with encephalopathy being the most common (73%). In this study, we identified 11 risk factors for neurologic complications. From the recipient's perspective, age < 29 or \geq 60 years along with BMI < 21.6 or > 27.6 kg/m² significantly increased NC risk, and Child-Pugh score > 9, hepatic encephalopathy, mental disorder, 7-d tacrolimus level > 8.9 ng/mL, intraabdominal infection were complementary to previous studies. Patients with history of variceal bleeding were less likely to develop NCs. Novel risk factors from donor's and surgeon's perspective were donor age < 22 or \geq 40 years, male-to-male gender match, GRWR between 0.9 and 1.9 and sequence of transplantation between 31 and 174. Our results provide a basis for risk stratification, which would enable transplant surgeons to weigh the risk during patient selection, control unfavorable factors before operation, avoid neurotoxicity during perioperative period, and perform active surveillance

after operation. The manner in which the experience of transplant team influences NCs should also be taken into account by the team leader or health care manager who would allow the conduction of interdisciplinary education as well as strategies for continuous quality improvement.

ACKNOWLEDGMENTS

The authors are grateful for the statistical support provided by Dr. Fu-Chang Hu.

COMMENTS

Background

Neurologic complications (NCs) are common after liver transplantation and result in significant morbidity and mortality. Encephalopathy, the most frequent etiology, is multifactorial in nature. Surgeons should identify patients at risk and avoid predisposing variables during the perioperative period to minimize their incidence. However, known risk factors have been mainly investigated from the recipient's perspective.

Research frontiers

The authors have considered various factors from recipient, donor, donor-recipient matching, and surgeon perspectives that had potential impact on early NC after liver transplantation. They had better control of confounders from the case-control study design using more flexible statistical tools, including generalized additive models for variables, which had a nonlinear effect on outcome.

Innovations and breakthroughs

In addition to risk factors from the recipient perspective that were complementary to those in the literature, novel risk factors discovered in this study were donor age (< 22 or ≥ 40 years), male-to-male gender matching, graft-recipient weight ratio 0.9%-1.9%, and sequence of transplantation between 31 and 174.

Applications

The results provide a basis for risk stratification for surgeons to minimize neurotoxic factors during transplantation. In addition, the transplant team leader should be aware of the higher risk of neurologic complication during the team's earlier experiences to conduct interdisciplinary education and quality improvement program.

Peer-review

The manuscript is clearly written to describe the results with pretty good discussion. The method section is really well written as a clinical report and can be one of the greatest examples for physician-scientists.

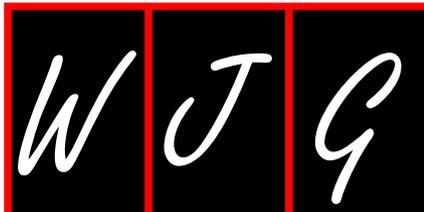
REFERENCES

- 1 **Senzolo M**, Ferronato C, Burra P. Neurologic complications after solid organ transplantation. *Transpl Int* 2009; **22**: 269-278 [PMID: 19076332 DOI: 10.1111/j.1432-2277.2008.00780.x]
- 2 **Zivković SA**. Neurologic complications after liver transplantation. *World J Hepatol* 2013; **5**: 409-416 [PMID: 24023979 DOI: 10.4254/wjh.v5.i8.409]
- 3 **Dhar R**, Young GB, Marotta P. Perioperative neurological complications after liver transplantation are best predicted by pre-transplant hepatic encephalopathy. *Neurocrit Care* 2008; **8**: 253-258 [PMID: 17928960 DOI: 10.1007/s12028-007-9020-4]
- 4 **Kim KM**, Kim GS, Ko JS, Gwak MS, Lee SK, Son MG. Factors associated with consciousness recovery time after liver transplantation in recipients with hepatic encephalopathy. *Transplant Proc* 2014; **46**: 712-715 [PMID: 24767331 DOI: 10.1016/j.transproceed.2013.12.031]
- 5 **Vizzini G**, Asaro M, Miraglia R, Gruttadauria S, Fili D, D'Antoni A, Petridis I, Marrone G, Pagano D, Gridelli B. Changing picture of central nervous system complications in liver transplant recipients. *Liver Transpl* 2011; **17**: 1279-1285 [PMID: 21770016 DOI: 10.1002/lt.22383]
- 6 **Saner FH**, Gensicke J, Olde Damink SW, Pavlaković G, Treckmann J, Dammann M, Kaiser GM, Sotiropoulos GC, Radtke A, Koeppe S, Beckebaum S, Cicinnati V, Nadalin S, Malagó M, Paul A, Broelsch CE. Neurologic complications in adult living donor liver transplant patients: an underestimated factor? *J Neurol* 2010; **257**: 253-258 [PMID: 19727899 DOI: 10.1007/s00415-009-5303-3]
- 7 **Amodio P**, Biancardi A, Montagnese S, Angeli P, Iannizzi P, Cillo U, D'Amico D, Gatta A. Neurological complications after orthotopic liver transplantation. *Dig Liver Dis* 2007; **39**: 740-747 [PMID: 17611177 DOI: 10.1016/j.dld.2007.05.004]
- 8 **Busuttil BW**, Klintmalm GK. *Transplantation of the Liver*. 2nd ed. Philadelphia, Pennsylvania: Saunders, 2005: 1029-1036
- 9 **Bronster DJ**, Emre S, Boccagni P, Sheiner PA, Schwartz ME, Miller CM. Central nervous system complications in liver transplant recipients--incidence, timing, and long-term follow-up. *Clin Transplant* 2000; **14**: 1-7 [PMID: 10693627 DOI: 10.1034/j.1399-0012.2000.140101.x]
- 10 **Lewis MB**, Howdle PD. Neurologic complications of liver transplantation in adults. *Neurology* 2003; **61**: 1174-1178 [PMID: 14610116 DOI: 10.1212/01.WNL.0000089487.42870.C6]
- 11 **Kim BS**, Lee SG, Hwang S, Park KM, Kim KH, Ahn CS, Moon DB, Ha TY, Song GW, Kim DS, Moon KM, Jung DH. Neurologic complications in adult living donor liver transplant recipients. *Clin Transplant* 2007; **21**: 544-547 [PMID: 17645717 DOI: 10.1111/j.1399-0012.2007.00687.x]
- 12 **Klintmalm GB**, Davis GL, Teperman L, Netto GJ, Washburn K, Rudich SM, Pomfret EA, Vargas HE, Brown R, Eckhoff D, Pruett TL, Roberts J, Mulligan DC, Charlton MR, Heffron TG, Ham JM, Douglas DD, Sher L, Baliga PK, Kinkhabwala M, Koneru B, Abecassis M, Millis M, Jennings LW, Fasola CG. A randomized, multicenter study comparing steroid-free immunosuppression and standard immunosuppression for liver transplant recipients with chronic hepatitis C. *Liver Transpl* 2011; **17**: 1394-1403 [PMID: 21850690 DOI: 10.1002/lt.22417]
- 13 **Yee TW**, Wild C. Vector generalized additive models. *J Roy Statist Soc Ser B* 1996; **58**: 481-493
- 14 **Pujol A**, Graus F, Rimola A, Beltrán J, Garcia-Valdecasas JC, Navasa M, Grande L, Galofré J, Visa J, Rodés J. Predictive factors of in-hospital CNS complications following liver transplantation. *Neurology* 1994; **44**: 1226-1230 [PMID: 8035920 DOI: 10.1212/WNL.44.7.1226]
- 15 **Garcia-Martinez R**, Rovira A, Alonso J, Jacas C, Simón-Talero M, Chavarria L, Vargas V, Córdoba J. Hepatic encephalopathy is associated with posttransplant cognitive function and brain volume. *Liver Transpl* 2011; **17**: 38-46 [PMID: 21254343 DOI: 10.1002/lt.22197]
- 16 **Buis CI**, Wiesner RH, Krom RA, Kremers WK, Wijdicks EF. Acute confusional state following liver transplantation for alcoholic liver disease. *Neurology* 2002; **59**: 601-605 [PMID: 12196657 DOI: 10.1212/WNL.59.4.601]
- 17 **Kanwal F**, Chen D, Ting L, Gornbein J, Saab S, Durazo F, Yersiz H, Farmer D, Ghobrial RM, Busuttil RW, Han SH. A model to predict the development of mental status changes of unclear cause after liver transplantation. *Liver Transpl* 2003; **9**: 1312-1319 [PMID: 14625832 DOI: 10.1016/j.lts.2003.09.023]
- 18 **Dasgupta M**, Dumbrell AC. Preoperative risk assessment for delirium after noncardiac surgery: a systematic review. *J Am Geriatr Soc* 2006; **54**: 1578-1589 [PMID: 17038078 DOI: 10.1111/j.1532-5415.2006.00893.x]
- 19 **Monk TG**, Weldon BC, Garvan CW, Dede DE, van der Aa MT, Heilman KM, Gravenstein JS. Predictors of cognitive dysfunction after major noncardiac surgery. *Anesthesiology* 2008; **108**: 18-30 [PMID: 18156878 DOI: 10.1097/01.anes.0000296071.19434.1e]

- 20 **Yoshimura Y**, Kubo S, Shirata K, Hirohashi K, Tanaka H, Shuto T, Takemura S, Kinoshita H. Risk factors for postoperative delirium after liver resection for hepatocellular carcinoma. *World J Surg* 2004; **28**: 982-986 [PMID: 15573252 DOI: 10.1007/s00268-004-7344-1]
- 21 **Dick AA**, Spitzer AL, Seifert CF, Deckert A, Carithers RL, Reyes JD, Perkins JD. Liver transplantation at the extremes of the body mass index. *Liver Transpl* 2009; **15**: 968-977 [PMID: 19642131 DOI: 10.1002/lt.21785]
- 22 **Bemeur C**. Neurological complications post-liver transplantation: impact of nutritional status. *Metab Brain Dis* 2013; **28**: 293-300 [PMID: 23129292 DOI: 10.1007/s11011-012-9352-4]
- 23 **Antar R**, Wong P, Ghali P. A meta-analysis of nutritional supplementation for management of hospitalized alcoholic hepatitis. *Can J Gastroenterol* 2012; **26**: 463-467 [PMID: 22803023 DOI: 10.1155/2012/945707]
- 24 **Cruz RJ**, DiMartini A, Akhavanheidari M, Iacovoni N, Boardman JF, Donaldson J, Humar A, Bartynski WS. Posterior reversible encephalopathy syndrome in liver transplant patients: clinical presentation, risk factors and initial management. *Am J Transplant* 2012; **12**: 2228-2236 [PMID: 22494636 DOI: 10.1111/j.1600-6143.2012.04048.x]
- 25 **Feng S**, Goodrich NP, Bragg-Gresham JL, Dykstra DM, Punch JD, DeRoy MA, Greenstein SM, Merion RM. Characteristics associated with liver graft failure: the concept of a donor risk index. *Am J Transplant* 2006; **6**: 783-790 [PMID: 16539636 DOI: 10.1111/j.1600-6143.2006.01242.x]
- 26 **Feng AC**, Fan HL, Chen TW, Hsieh CB. Hepatic hemodynamic changes during liver transplantation: a review. *World J Gastroenterol* 2014; **20**: 11131-11141 [PMID: 25170200 DOI: 10.3748/wjg.v20.i32.11131]
- 27 **Rodríguez-Castro KI**, De Martin E, Gambato M, Lazzaro S, Villa E, Burra P. Female gender in the setting of liver transplantation. *World J Transplant* 2014; **4**: 229-242 [PMID: 25540733 DOI: 10.5500/wjt.v4.i4.229]
- 28 **Burra P**, De Martin E, Gitto S, Villa E. Influence of age and gender before and after liver transplantation. *Liver Transpl* 2013; **19**: 122-134 [PMID: 23172830 DOI: 10.1002/lt.23574]
- 29 **Li C**, Wen TF, Yan LN, Li B, Yang JY, Wang WT, Xu MQ, Wei YG. Predictors of patient survival following living donor liver transplantation. *Hepatobiliary Pancreat Dis Int* 2011; **10**: 248-253 [PMID: 21669566 DOI: 10.1016/s1499-3872(11)60041-6]
- 30 **Marino IR**, Doyle HR, Aldrighetti L, Doria C, McMichael J, Gayowski T, Fung JJ, Tzakis AG, Starzl TE. Effect of donor age and sex on the outcome of liver transplantation. *Hepatology* 1995; **22**: 1754-1762 [PMID: 7489985 DOI: 10.1002/hep.1840220622]
- 31 **Adam R**, Cailliez V, Majno P, Karam V, McMaster P, Caine RY, O'Grady J, Pichlmayr R, Neuhaus P, Otte JB, Hoeckerstedt K, Bismuth H. Normalised intrinsic mortality risk in liver transplantation: European Liver Transplant Registry study. *Lancet* 2000; **356**: 621-627 [PMID: 10968434 DOI: 10.1016/S0140-6736(00)02603-9]
- 32 **Broering DC**, Kim JS, Mueller T, Fischer L, Ganschow R, Bickel T, Mueller L, Hillert C, Wilms C, Hinrichs B, Helmke K, Pothmann W, Burdelski M, Rogiers X. One hundred thirty-two consecutive pediatric liver transplants without hospital mortality: lessons learned and outlook for the future. *Ann Surg* 2004; **240**: 1002-1012; discussion 1012 [PMID: 15570206 DOI: 10.1097/01.sla.0000146148.01586.72]
- 33 **Morioka D**, Egawa H, Kasahara M, Ito T, Haga H, Takada Y, Shimada H, Tanaka K. Outcomes of adult-to-adult living donor liver transplantation: a single institution's experience with 335 consecutive cases. *Ann Surg* 2007; **245**: 315-325 [PMID: 17245187 DOI: 10.1097/01.sla.0000236600.24667.a4]
- 34 **Li C**, Mi K, Wen Tf, Yan Ln, Li B, Yang Jy, Xu Mq, Wang Wt, Wei Yg. A learning curve for living donor liver transplantation. *Dig Liver Dis* 2012; **44**: 597-602 [PMID: 22387283 DOI: 10.1016/j.dld.2012.01.016]
- 35 **Campagna F**, Biancardi A, Cillo U, Gatta A, Amodio P. Neurocognitive-neurological complications of liver transplantation: a review. *Metab Brain Dis* 2010; **25**: 115-124 [PMID: 20204483 DOI: 10.1007/s11011-010-9183-0]
- 36 **Teperman LW**. Impact of pretransplant hepatic encephalopathy on liver posttransplantation outcomes. *Int J Hepatol* 2013; **2013**: 952828 [PMID: 24324895 DOI: 10.1155/2013/952828]

P- Reviewer: Kita K, Rydzewski A **S- Editor:** Qi Y **L- Editor:** A
E- Editor: Ma S





Case Control Study

Relationships between cell cycle pathway gene polymorphisms and risk of hepatocellular carcinoma

Yue-Li Nan, Yan-Ling Hu, Zhi-Ke Liu, Fang-Fang Duan, Yang Xu, Shu Li, Ting Li, Da-Fang Chen, Xiao-Yun Zeng

Yue-Li Nan, Yang Xu, Shu Li, Ting Li, Xiao-Yun Zeng, Department of Epidemiology, School of Public Health, Guangxi Medical University, Nanning 530021, Guangxi Zhuang Autonomous Region, China

Yan-Ling Hu, Medical Scientific Research Centre, Guangxi Medical University, Nanning 530021, Guangxi Zhuang Autonomous Region, China

Zhi-Ke Liu, Fang-Fang Duan, Da-Fang Chen, Department of Epidemiology and Biostatistics, School of Public Health, Peking University Health Science Center, Beijing 100191, China

Xiao-Yun Zeng, Key Laboratory of High-Incidence-Tumor Prevention and Treatment (Guangxi Medical University), Ministry of Education, Nanning 530021, Guangxi Zhuang Autonomous Region, China

Author contributions: Zeng XY designed the research; Xu Y, Li S and Li T collected the materials and clinical data; Liu ZK and Duan FF performed the majority of experiments; Chen DF conceived the experimental assays; Nan YL performed the experiments, analyzed the data and wrote the manuscript; Hu YL made critical revisions of the manuscript.

Supported by National Natural Science Foundation of China, No. 81360448; Natural Science Foundation of Guangxi, No. 2014GXNSFAA118139; Fund of Key Laboratory of High-Incidence-Tumor Prevention and Treatment (Guangxi Medical University), Ministry of Education, No. GK2015-ZZ03 and No. GK2014-ZZ03; and Guangxi Outstanding Teacher Training Project for Colleges.

Institutional review board statement: The study was approved by the ethical review committee of Guangxi Medical University.

Informed consent statement: All study participants provided informed written consent prior to study enrollment.

Conflict-of-interest statement: The authors have declared that they have no competing interests.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Correspondence to: Xiao-Yun Zeng, MD, PhD, Department of Epidemiology, School of Public Health, Guangxi Medical University, No. 22 Shuangyong Road, Nanning 530021, Guangxi Zhuang Autonomous Region, China. zxyxjw@21cn.com
Telephone: +86-771-5358325
Fax: +86-771-5352523

Received: March 3, 2016

Peer-review started: March 7, 2016

First decision: April 14, 2016

Revised: April 29, 2016

Accepted: May 21, 2016

Article in press: May 23, 2016

Published online: June 28, 2016

Abstract

AIM: To investigate the associations between the polymorphisms of cell cycle pathway genes and the risk of hepatocellular carcinoma (HCC).

METHODS: We enrolled 1127 cases newly diagnosed with HCC from the Tumor Hospital of Guangxi Medical University and 1200 non-tumor patients from the First Affiliated Hospital of Guangxi Medical University. General demographic characteristics, behavioral information, and hematological indices were collected by unified questionnaires. Genomic DNA was isolated

from peripheral venous blood using Phenol-Chloroform. The genotyping was performed using the Sequenom MassARRAY iPLEX genotyping method. The association between genetic polymorphisms and risk of HCC was shown by *P*-value and the odd ratio (OR) with 95% confidence interval (CI) using the unconditional logistic regression after adjusting for age, sex, nationality, smoking, drinking, family history of HCC, and hepatitis B virus (HBV) infection. Moreover, stratified analysis was conducted on the basis of the status of HBV infection, smoking, and alcohol drinking.

RESULTS: The HCC risk was lower in patients with the *MCM4* rs2305952 CC (OR = 0.22, 95%CI: 0.08-0.63, *P* = 0.01) and with the *CHEK1* rs515255 TC, TT, TC/TT (OR = 0.73, 95%CI: 0.56-0.96, *P* = 0.02; OR = 0.67, 95%CI: 0.46-0.97, *P* = 0.04; OR = 0.72, 95%CI: 0.56-0.92, *P* = 0.01, respectively). Conversely, the HCC risk was higher in patients with the *KAT2B* rs17006625 GG (OR = 1.64, 95%CI: 1.01-2.64, *P* = 0.04). In addition, the risk was markedly lower for those who were carriers of *MCM4* rs2305952 CC and were also HBsAg-positive and non-drinking and non-smoking (*P* < 0.05, respectively) and for those who were carriers of *CHEK1* rs515255 TC, TT, TC/TT and were also HBsAg-negative and non-drinking (*P* < 0.05, respectively). Moreover, the risk was higher for those who were carriers of *KAT2B* rs17006625 GG and were also HBsAg-negative (*P* < 0.05).

CONCLUSION: Of 12 cell cycle pathway genes, *MCM4*, *CHEK1* and *KAT2B* polymorphisms may be associated with the risk of HCC.

Key words: Cell cycle pathway genes; Hepatocellular carcinoma; Single nucleotide polymorphism; Case-control study; Genetic susceptibility

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: We analyzed the effects of polymorphisms of 12 cell cycle pathway genes on the risk of hepatocellular carcinoma (HCC) in a large population of 1019 HCC cases and 1138 controls. The results suggest that *MCM4* rs2305952 CC and *CHEK1* rs515255 TC, TT, TC/TT may be significantly associated with a decreased risk of HCC. *KAT2B* rs17006625 GG may increase the risk of HCC.

Nan YL, Hu YL, Liu ZK, Duan FF, Xu Y, Li S, Li T, Chen DF, Zeng XY. Relationships between cell cycle pathway gene polymorphisms and risk of hepatocellular carcinoma. *World J Gastroenterol* 2016; 22(24): 5558-5567 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5558.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5558>

INTRODUCTION

Hepatocellular carcinoma (HCC) is a serious threat to human health worldwide. It is the fourth most common cancer and the second leading cause of cancer death, with nearly 746000 deaths per year^[1]. The incidence of this fatal disease continues to increase. HCC occurrence and development are related to environmental factors, such as infection with hepatitis B virus (HBV) or hepatitis C virus (HCV), cigarette smoking, and alcohol consumption, as well as genetic susceptibility^[2-4]. Many studies strongly support that single nucleotide polymorphisms (SNPs) of a variety of genes are associated with HCC^[5-7]. However, the genetic mechanism underlying the inherited component of HCC is still not fully understood.

The cell cycle comprises the events that result in the formation of two daughter cells through division of the parent cell. Cell cycle progression, including cell division, is influenced by three different types of molecules: cyclin, cyclin-dependent kinases, and cyclin kinase inhibitors^[8]. The associations between the genetic susceptibility of genes which regulate the cell cycle and the risk of cancer are well known. For instance, a polymorphism of the *p27* generates an increased risk of squamous cell carcinoma of the head and neck^[9], while polymorphisms of *p27* and *p21* are associated with a significantly increased risk of HCC^[10]. Other cell cycle pathway genes implicated in cancer include *cyclinD1*^[11], *p53*^[12], *CHEK2*^[13] and *P21*^[14].

During the last several decades, an increasing number of studies have shown an association between genetic variants, mainly in the form of SNPs, and the risk of cancer, including breast^[15], colorectal^[16], cervical, and vulvar cancers^[17], and HCC^[18]. Despite investigations into the association of polymorphisms in cell cycle pathway genes with cancer susceptibility^[19,20], in the case of HCC this association remains unclear. Therefore, in this hospital-based study we investigated the associations between the polymorphisms of SNPs in cell cycle pathway genes and the risk of HCC.

MATERIALS AND METHODS

Study population

For this case-control study, 2327 subjects were consecutively recruited from June 2007 to December 2013. The 1127 HCC patients were from the Tumor Hospital of Guangxi Medical University and were newly diagnosed with HCC based on biochemical (α -fetoprotein > 20 μ g/L) and histopathological examinations. None had undergone radiotherapy or chemotherapy before blood sampling. The 1200 controls from the First Affiliated Hospital of Guangxi Medical University consisted of non-tumor patients admitted within the same period of time. Informed

Table 1 Summarized information of selected single nucleotide polymorphisms in cell cycle pathway genes

Genes	SNPs	Chromosome (position)	Allele	MAF (hapmap-HCB)
MCM4	rs2305952	8 (47962049)	C/T	C = 0.18
YWHAB	rs2425675	20 (44906293)	A/G	A = 0.20
CDKN2A	rs3088440	9 (21968160)	A/G	A = 0.08
TGFB3	rs3917148	14 (75980178)	A/C	C = 0.10
RBL2	rs3929	16 (53490396)	C/G	C = 0.20
RAD21	rs6987652	8 (116870042)	A/G	A = 0.12
SMAD3	rs11556090	15 (67194045)	A/G	G = 0.09
	rs8025774	15 (67190938)	C/T	C = 0.45
KAT2B	rs17006625	3 (20119604)	A/G	G = 0.14
	rs4858770	3 (20152931)	C/T	T = 0.47
MCM7	rs2070215	7 (100099174)	A/G	G = 0.29
	rs2261360	7 (100095370)	A/C	A = 0.37
CDKN1A	rs3176320	6 (36679011)	A/G	G = 0.17
CDC25C	rs3734166	5 (138329634)	A/G	G = 0.38
CHEK1	rs515255	11 (125627250)	C/T	T = 0.44

MAF (minor allele frequency) was derived from HCB population in HapMap website (<http://hapmap.ncbi.nlm.nih.gov/>). SNPs: Single nucleotide polymorphisms.

consent was obtained from all participants, who also agreed to truthfully complete the questionnaires.

Information and sample collection

General demographic and behavioral information, hematological indices, and data on the patients' age, sex, nationality, drinking habit, smoking habit, HBV infection, and family history of HCC were obtained in face-to-face interviews by trained investigators. Peripheral venous blood was collected in a vacuum EDTA anticoagulant tube from each participant. Genomic DNA was extracted using a standard phenol-chloroform extraction method and stored at -80°C .

SNP selection

From the GEO database (<https://www.ncbi.nlm.nih.gov/geo/>), we found three sets of whole genome expression microarray data which were related to HCC (GSE14520, GSE25097, and GSE12941). A total of 3826 different genes were selected using SPSS 16.0 software (SPSS Inc., Chicago, IL, United States) ($P < 0.05$). Gene ontology classification and pathway enrichment analysis were performed by blast2GO and DAVID (<https://david.ncifcrf.gov/>) and 40 cell cycle pathway genes involved in the cellular process were chose. The genotype information was downloaded from Hapmap website (<http://hapmap.ncbi.nlm.nih.gov/>), and functional SNPs were selected using Haploview 4.2 software (Cambridge, MA o2141, United States) based on a function prediction website (<http://snpinfo.niehs.nih.gov/snpfunc.htm>). Referring to the existing literature on these SNPs with HCC, 15 SNPs in 12 genes (*MCM4* rs2305952, *YWHAB* rs2425675, *CDKN2A* rs3088440, *TGFB3* rs3917148, *RBL2* rs3929, *RAD21* rs6987652, *SMAD3* rs11556090, rs8025774,

KAT2B rs17006625, rs4858770, *MCM7* rs2070215, rs2261360, *CDKN1A* rs3176320, *CDC25C* rs3734166, and *CHEK1* rs515255) were selected in this study. Information of selected SNPs is shown in Table 1.

SNP genotyping

Before genotyping, each DNA sample was quantified using a UV-Vis spectrophotometer Q5000 (Quowell Technology, Inc., United States) and diluted to a final concentration of $50\text{ ng}/\mu\text{L}$. SNP genotyping was performed using a MassARRAY system (Sequenom, San Diego, CA, United States) and a matrix-assisted laser desorption ionization-time of flight mass spectrometry method according to the manufacturer's instructions. Primers for PCR and extension were designed using the Assay Designer software package (Sequenom). For quality control, 5% of the samples were randomly chosen and genotyped twice for each locus. Among the 1127 patient samples and 1200 control samples, genotyping was successful for all 15 SNPs in both groups, with a success rate of 92.7%. Thus, all 1019 HCC patients and 1138 controls were included in the final analysis.

Statistical analysis

Statistical analyses were performed using the SPSS 16.0 software (SPSS Inc., Chicago, IL, United States). Continuous variables were evaluated using the two-sample *t*-test. Categorical variables and genotype frequencies between the HCC patients and controls were compared using the Pearson's χ^2 and Fisher's exact test. Hardy-Weinberg equilibrium (HWE) was evaluated by a goodness-of-fit χ^2 test to compare the observed genotype frequencies with the expected ones. The association between SNP genotypes and HCC risk was estimated using unconditional logistic regression analysis and an odds ratio (OR) with 95% confidence interval (CI). All statistical tests were two-sided. A *P*-value < 0.05 was considered to indicate statistical significance.

RESULTS

Characteristics of the participants

The 2157 unrelated Chinese subjects enrolled in this study included 881 (86.5%) males and 138 (13.5%) females with HCC. The mean age of these patients was 48.54 ± 11.44 years. The control group consisted of 982 (86.3%) males and 156 (13.7%) females, with a mean age of 48.01 ± 11.5 years. The general demographic characteristics and behavior information on the patients and controls are provided in Table 2. There were no significant differences between the HCC patients and the controls in terms of age, sex, and nationality; however, HCC patients had a significantly higher rate of a positive history of HBV infection, a family history of HCC, smoking, and drinking.

Table 2 General demographic characteristics and behavioral information among hepatocellular carcinoma patients and controls

Variable	HCC patients <i>n</i> = 1019	Controls <i>n</i> = 1138	<i>t</i> / χ^2	<i>P</i> value
Age	48.54 ± 11.44	48.01 ± 11.50	-1.076	0.28
Gender				
Male	881	982	0.013	0.91
Female	138	156		
Nationality				
Han	673	708	3.591	0.17
Zhuang	332	410		
Others	14	20		
Drinking				
Yes	345	145	136.527	< 0.001
No	674	993		
Smoking				
Yes	355	158	130.222	< 0.001
No	664	980		
Chronic HBV infection				
Yes	794	109	1031.687	< 0.001
No	225	1029		
Family history of HCC				
Yes	80	2	86.597	< 0.001
No	939	1136		

HCC: Hepatocellular carcinoma.

Allele frequencies and genotype distribution

In the control group, the genotype frequencies of the 15 SNPs, all but *CDKN1A* rs3176320, were in line with the HWE (*P* > 0.05), which indicated that these study participants were from a homogeneous group. The allele frequencies and genotype distribution of SNPs among the HCC patients and controls from this study are listed in Table 3.

Association analysis of genetic polymorphisms and HCC

The association between SNPs and the risk of HCC was examined using unconditional logistic regression analysis. According to the crude ORs and their 95% CIs, *SMAD3* rs11556090 AG or AG/GG and *MCM7* rs2070215 GG carried an increased risk of HCC when compared with the wild genotype *SMAD3* rs11556090 AA and *MCM7* rs2070215 AA, respectively. Individuals with *CDC25C* rs3734166 GG or GA/GG and *KAT2B* rs4858770 TT had a lower risk of HCC than those with the wild genotype *CDC25C* rs3734166 AA and *KAT2B* rs4858770 CC, respectively. However, the association disappeared after adjusting for age, sex, nationality, smoking, drinking, family history of HCC, and HBV infection. Using individuals with the wild genotype AA as the reference, individuals carrying the GG variant of *KAT2B* rs17006625 had a higher risk of HCC (adjusted OR = 1.64, 95%CI: 1.01-2.64, *P* = 0.04) after adjusting for confounding factors. In addition, compared with the wild genotypes *MCM4* rs2305952 TT and *CHEK1* rs515255 CC, individuals carrying the CC variant of *MCM4* rs2305952 or the TC, TT, TC/TT

Table 3 Allele frequencies and genotype distribution of single nucleotide polymorphisms *n* (%)

SNP	Genotype	HCC patients <i>n</i> = 1019	Control <i>n</i> = 1138	χ^2	<i>P</i> value of HWE
rs2305952	TT	801 (78.61)	883 (77.59)	0.04	0.83
	TC	209 (20.51)	238 (20.91)		
	CC	9 (0.88)	17 (1.49)		
rs2425675	GG	632 (62.02)	724 (63.62)	0.96	0.33
	AG	348 (34.15)	374 (32.86)		
	AA	39 (3.83)	40 (3.51)		
rs3088440	GG	750 (73.60)	813 (71.44)	0.19	0.66
	GA	249 (24.44)	300 (26.36)		
	AA	20 (1.96)	25 (2.20)		
rs3917148	AA	773 (75.86)	882 (77.50)	1.32	0.25
	CA	233 (22.87)	235 (20.65)		
	CC	13 (1.28)	21 (1.85)		
rs3929	GG	619 (60.75)	688 (60.46)	0.03	0.86
	GC	349 (34.25)	395 (34.71)		
	CC	51 (5.00)	55 (4.83)		
rs6987652	GG	743 (72.91)	843 (74.08)	0.38	0.54
	AG	251 (24.63)	270 (23.73)		
	AA	25 (2.45)	25 (2.20)		
rs11556090	AA	622 (61.04)	749 (65.82)	0.15	0.70
	AG	352 (34.54)	346 (30.40)		
	GG	45 (4.42)	43 (3.78)		
rs17006625	AA	526 (51.62)	620 (54.48)	0.48	0.49
	AG	412 (40.43)	446 (39.19)		
	GG	81 (7.95)	72 (6.33)		
rs2070215	AA	465 (45.63)	554 (48.68)	< 0.01	1.00
	AG	424 (41.61)	480 (42.18)		
	GG	130 (12.76)	104 (9.14)		
rs2261360	CC	460 (45.14)	484 (42.53)	2.61	0.11
	CA	433 (42.49)	497 (43.67)		
	AA	126 (12.37)	157 (13.80)		
rs3176320	AA	579 (56.82)	687 (60.37)	5.05	0.02
	GA	383 (37.59)	377 (33.13)		
	GG	57 (5.59)	74 (6.50)		
rs3734166	AA	421 (41.32)	421 (36.99)	0.06	0.8
	GA	481 (47.20)	539 (47.36)		
	GG	117 (11.48)	178 (15.64)		
rs4858770	CC	445 (43.67)	465 (40.86)	0.65	0.42
	CT	461 (45.24)	515 (45.25)		
	TT	113 (11.09)	158 (13.88)		
rs515255	CC	408 (40.04)	411 (36.12)	0.29	0.59
	TC	469 (46.03)	553 (48.59)		
	TT	142 (13.94)	174 (15.29)		
rs8025774	CC	313 (30.72)	335 (29.44)	1.32	0.25
	CT	514 (50.44)	547 (48.07)		
	TT	192 (18.84)	256 (22.50)		

HCC: Hepatocellular carcinoma; SNP: Single nucleotide polymorphism; HWE: Hardy-Weinberg equilibrium

variants of *CHEK1* rs515255 had a significantly lower risk of HCC (adjusted OR = 0.22, 95%CI: 0.08-0.63, *P* = 0.01; adjusted OR = 0.73, 95%CI: 0.56-0.96, *P* = 0.02; adjusted OR = 0.67, 95%CI: 0.46-0.97, *P* = 0.04; adjusted OR = 0.72, 95%CI: 0.56-0.92, *P* = 0.01, respectively). The associations are shown in Table 4.

Association between SNPs and HCC risk stratified by behavioral factors

HBV infection, alcohol intake status, and smoking status are important behavioral factors that can

Table 4 Associations between single nucleotide polymorphisms with the risk of hepatocellular carcinoma

SNP	Genotype	OR (95%CI) ¹	P value ¹	OR (95%CI) ²	P value ²
rs2305952	TT	Reference		Reference	
	TC	0.97 (0.79-1.19)	0.76	0.97 (0.72-1.32)	0.85
	CC	0.58 (0.26-1.32)	0.19	0.22 (0.08-0.63)	0.01 ^a
rs2425675	TC/CC	0.94 (0.77-1.16)	0.57	0.89 (0.66-1.19)	0.43
	GG	Reference		Reference	
	AG	1.07 (0.89-1.28)	0.49	1.92 (0.71-1.20)	0.54
rs3088440	AA	1.12 (0.71-1.76)	0.63	0.97 (0.51-1.85)	0.93
	AG/AA	1.07 (0.90-1.28)	0.44	0.93 (0.72-1.20)	0.56
	GG	Reference		Reference	
rs3917148	GA	0.90 (0.74-1.09)	0.29	1.02 (0.76-1.35)	0.92
	AA	0.87 (0.48-1.58)	0.64	1.46 (0.62-3.44)	0.38
	GA/AA	0.90 (0.74-1.09)	0.26	1.04 (0.79-1.37)	0.77
rs3929	CA	1.13 (0.92-1.39)	0.24	1.18 (0.88-1.59)	0.28
	CC	0.71 (0.35-1.42)	0.33	1.05 (0.41-2.68)	0.92
	CA/CC	1.10 (0.90-1.34)	0.37	1.17 (0.88-1.56)	0.29
rs6987652	GG	Reference		Reference	
	GC	0.98 (0.82-1.18)	0.84	0.97 (0.75-1.26)	0.82
	CC	1.03 (0.69-1.53)	0.88	1.39 (0.80-2.42)	0.25
rs11556090	GC/CC	0.99 (0.83-1.18)	0.89	1.02 (0.79-1.30)	0.90
	GG	Reference		Reference	
	AG	1.06 (0.87-1.29)	0.60	0.92 (0.69-1.23)	0.59
rs17006625	AA	1.14 (0.65-1.99)	0.66	1.26 (0.55-2.88)	0.59
	AG/AA	1.06 (0.88-1.29)	0.54	0.95 (0.72-1.25)	0.71
	AA	Reference		Reference	
rs2070215	AG	1.23 (1.02-1.47)	0.03	1.11 (0.85-1.44)	0.44
	GG	1.26 (0.82-1.94)	0.29	1.02 (0.54-1.91)	0.96
	AG/GG	1.23 (1.03-1.47)	0.02	1.10 (0.85-1.42)	0.47
rs2261360	AA	Reference		Reference	
	AG	1.09 (0.91-1.30)	0.35	1.07 (0.83-1.38)	0.61
	GG	1.33 (0.95-1.86)	0.10	1.64 (1.01-2.64)	0.04 ^a
rs3734166	AG/GG	1.12 (0.95-1.33)	0.18	1.14 (0.89-1.46)	0.29
	AA	Reference		Reference	
	AG	1.05 (0.88-1.26)	0.58	0.95 (0.73-1.24)	0.71
rs4858770	GG	1.49 (1.12-1.98)	0.01	1.39 (0.93-2.08)	0.11
	AG/GG	1.13 (0.95-1.34)	0.16	1.03 (0.81-1.32)	0.81
	CC	Reference		Reference	
rs515255	CA	0.92 (0.77-1.10)	0.35	0.84 (0.64-1.09)	0.19
	AA	0.84 (0.65-1.10)	0.21	0.89 (0.60-1.31)	0.55
	CA/AA	0.90 (0.76-1.07)	0.22	0.85 (0.66-1.09)	0.19
rs8025774	AA	Reference		Reference	
	GA	0.89 (0.74-1.07)	0.22	0.92 (0.71-1.21)	0.56
	GG	0.66 (0.50-0.86)	0.002	0.86 (0.59-1.25)	0.43
rs2070215	GA/GG	0.83 (0.70-0.99)	0.04	0.91 (0.71-1.17)	0.45
	CC	Reference		Reference	
	CT	0.94 (0.78-1.12)	0.47	0.96 (0.74-1.24)	0.74
rs2261360	TT	0.75 (0.57-0.98)	0.04	0.80 (0.54-1.20)	0.28
	CT/TT	0.89 (0.75-1.06)	0.19	0.92 (0.72-1.18)	0.51
	CC	Reference		Reference	
rs3734166	TC	0.85 (0.71-1.03)	0.09	0.73 (0.56-0.96)	0.02 ^a
	TT	0.82 (0.63-1.07)	0.14	0.67 (0.46-0.97)	0.04 ^a
	TC/TT	0.85 (0.71-1.01)	0.06	0.72 (0.56-0.92)	0.01 ^a
rs4858770	CC	Reference		Reference	
	CT	1.01 (0.83-1.22)	0.95	0.95 (0.72-1.27)	0.74
	TT	0.80 (0.63-1.02)	0.08	0.94 (0.66-1.32)	0.71
rs515255	CT/TT	0.94 (0.78-1.13)	0.52	0.95 (0.73-1.24)	0.69

¹OR and 95%CI without adjusting for confounding factors; ²OR and 95%CI after adjusting for age, sex, nationality, smoking, drinking, family history of hepatocellular carcinoma, and HBV infection. ^a $P < 0.05$ was considered statistically significant. OR: Odds ratio; CI: Confidence interval; SNPs: Single nucleotide polymorphisms.

increase the risk of HCC. To account for the role of these factors, a stratified analysis was conducted. Thus, when the patients were stratified, we found that the variant genotype CC of *MCM4* rs2305952

was associated with a significantly lower risk of HCC among HBsAg-positive individuals, non-drinkers, and non-smokers (adjusted OR = 0.25, 95%CI: 0.08-0.80, $P = 0.02$; adjusted OR = 0.19, 95%CI: 0.06-0.60, P

Table 5 Stratified analysis on the association between single nucleotide polymorphism genotype and hepatocellular carcinoma risk according to hepatitis B virus infection status

SNP	HBsAg-positive				HBsAg-negative			
	Case	Control	OR (95%CI) ¹	P value ¹	Case	Control	OR (95%CI) ¹	P value ¹
rs2305952								
TT	624	80	Reference		177	803	Reference	
TC	161	24	0.86 (0.53-1.42)	0.56	48	214	1.05 (0.72-1.52)	0.80
CC	9	5	0.25 (0.08-0.80)	0.02 ^a	0	12	-	1.00
TC/CC	170	29	0.76 (0.48-1.21)	0.25	48	226	0.99 (0.68-1.43)	0.95
rs17006625								
AA	411	60	Reference		115	560	Reference	
AG	323	42	1.15 (0.75-1.76)	0.54	89	404	1.07 (0.77-1.48)	0.68
GG	60	7	1.36 (0.59-3.17)	0.47	21	65	1.79 (1.02-3.12)	0.04 ^a
AG/GG	383	49	1.18 (0.78-1.77)	0.44	110	469	1.17 (0.86-1.59)	0.32
rs515255								
CC	301	39	Reference		107	372	Reference	
TC	377	52	0.93 (0.59-1.46)	0.75	92	501	0.64 (0.46-0.89)	0.01 ^a
TT	116	18	0.81 (0.44-1.50)	0.51	26	156	0.69 (0.36-0.96)	0.03 ^a
TC/TT	493	70	0.90 (0.59-1.37)	0.62	118	657	0.63 (0.46-0.86)	0.003 ^a

¹OR and 95%CI after adjusting for age, sex, nationality, smoking, drinking and family history of hepatocellular carcinoma. ^aP < 0.05 was considered statistically significant. OR: Odds ratio; CI: Confidence interval; SNPs: Single nucleotide polymorphisms.

Table 6 Stratified analysis on the association between single nucleotide polymorphism genotype and hepatocellular carcinoma risk according to drinking status

SNP	Drinking				Non-drinking			
	Case	Control	OR (95%CI) ¹	P value ¹	Case	Control	OR (95%CI) ¹	P value ¹
rs2305952								
TT	273	111	Reference		528	772	Reference	
TC	69	33	0.82 (0.44-1.52)	0.53	140	205	1.02 (0.72-1.44)	0.93
CC	3	1	0.51 (0.03-9.74)	0.66	6	16	0.19 (0.06-0.60)	0.004 ^a
TC/CC	72	34	0.81 (0.44-1.49)	0.49	146	221	0.91 (0.65-1.27)	0.57
rs515255								
CC	145	56	Reference		263	355	Reference	
TC	154	71	0.69 (0.40-1.19)	0.18	315	482	0.73 (0.54-0.99)	0.05 ^a
TT	46	18	1.10 (0.50-2.43)	0.82	96	156	0.56 (0.36-0.86)	0.01 ^a
TC/TT	200	89	0.77 (0.46-1.29)	0.31	411	638	0.69 (0.52-0.92)	0.01 ^a

¹OR and 95%CI after adjusting for age, sex, nationality, smoking, family history of hepatocellular carcinoma, and hepatitis B virus infection. ^aP < 0.05 was considered statistically significant. OR: Odds ratio; CI: Confidence interval; SNPs: Single nucleotide polymorphisms.

= 0.004; adjusted OR = 0.17, 95%CI: 0.05-0.56, P = 0.004, respectively). The variant genotypes TC, TT, and TC/TT of *CHEK1* rs515255 were associated with a significantly lower risk of HCC in HBsAg-negative individuals (adjusted OR = 0.64, 95%CI: 0.46-0.89, P = 0.01; adjusted OR = 0.69, 95%CI: 0.36-0.96, P = 0.03; adjusted OR = 0.63, 95%CI: 0.46-0.86, P = 0.003) and in non-drinkers (adjusted OR = 0.73, 95%CI: 0.54-0.99, P = 0.05; adjusted OR = 0.56, 95%CI: 0.36-0.86, P = 0.01; adjusted OR = 0.69, 95%CI: 0.52-0.92, P = 0.01, respectively). Among smokers, those with the TC variant genotype of *CHEK1* rs515255 had a significantly lower risk of HCC (adjusted OR = 0.54, 95%CI: 0.32-0.93, P = 0.03), while among non-smokers the risk was significantly lower in those with the TT variant genotype (adjusted OR = 0.60, 95%CI: 0.39-0.94, P = 0.03). In addition, the variant genotype GG of *KAT2B* rs17006625 was shown to carry a significantly higher risk of HCC

among HBsAg-negative individuals (adjusted OR = 1.79, 95%CI: 1.02-3.12, P = 0.04). These findings are summarized in Tables 5-7 (only significant SNPs are shown).

DISCUSSION

We performed this case-control study to investigate the associations between the 15 SNPs in 12 cell cycle pathway genes and the risk of HCC. The *KAT2B* rs17006625 GG was associated with an increased risk of HCC. Furthermore, this harmful effect was more marked in HBsAg-negative carriers. Conversely, the *CHEK1* rs515255 TC, TT, TC/TT and the *MCM4* rs2305952 CC were associated with a decreased risk of HCC. In addition, the risk was markedly lower for those who were carriers of *MCM4* rs2305952 CC and were also HBsAg-positive and non-drinking and non-smoking and for those who were carriers of the TC,

Table 7 Stratified analysis on the association between single nucleotide polymorphism genotype and hepatocellular carcinoma risk according to smoking status

SNP	Smoking				Non-smoking			
	Case	Control	OR (95%CI) ¹	P value ¹	Case	Control	OR (95%CI) ¹	P value ¹
rs2305952								
TT	274	124	Reference		527	759	Reference	
TC	77	32	1.05 (0.58-1.91)	0.87	132	206	0.94 (0.66-1.34)	0.75
CC	4	2	0.54 (0.06-4.97)	0.59	5	15	0.17 (0.05-0.56)	0.004 ^a
TC/CC	81	34	1.01 (0.57-1.82)	0.96	137	221	0.84 (0.60-1.19)	0.33
rs515255								
CC	145	53	Reference		263	358	Reference	
TC	155	84	0.54 (0.32-0.93)	0.03 ^a	314	469	0.81 (0.59-1.10)	0.17
TT	55	21	0.87 (0.41-1.85)	0.72	87	153	0.60 (0.39-0.94)	0.03 ^a
TC/TT	210	105	0.61 (0.67-1.02)	0.06	401	622	0.75 (0.56-1.01)	0.06

¹OR and 95%CI after adjusting for age, sex, nationality, drinking, family history of hepatocellular carcinoma, and HBV infection. ^aP < 0.05 was considered statistically significant. OR: Odds ratio; CI: Confidence interval; SNPs: Single nucleotide polymorphisms.

TT, TC/TT genotype of *CHEK1* rs515255 and were also HBsAg-negative and non-drinking. No significant associations were observed between other 12 SNPs and HCC risk.

The cell cycle pathway is one of the most important cellular signaling pathways, as it regulates both cell division and apoptosis. DNA damage readily leads to dysregulation of the cell cycle, which is an essential step in the initiation and development of human malignancies^[21-23]. In the present study, we reported that three SNPs in cell cycle pathway genes (*MCM4*, *CHEK1*, and *KAT2B*) were significantly associated with the risk of HCC.

MCM4, a member of the mini-chromosome maintenance family of proteins, which interact with cell cycle checkpoints and recombinant proteins to stabilize the S phase, is essential for the initiation of eukaryotic genome replication^[24,25]. Several reports have shown that *MCM4* protein is overexpressed in esophageal carcinomas^[26], cervical cancer^[27], and cervical squamous cell carcinoma^[28]. In our study, we found that the polymorphism of *MCM4* rs2305952 was associated with a lower risk of HCC. However, the mechanism of *MCM4* polymorphisms in HCC development remains unclear. Ishimi *et al.*^[29] found that *MCM4* is one of the crucial targets of DNA replication checkpoint and the phosphorylation of *MCM4*, which is caused by the activation of ATR-CHK1 pathway and CDK2, results in the DNA replication through the inactivation of the *MCM4*/6/7 complex. It is also found that *MCM4* mutations may cause tumors by affecting the formation of the *MCM4*/6/7 complex^[30,31].

CHEK1 is a mediator of cell cycle arrest in response to DNA damage. In addition to controlling cell cycle progression^[32], it regulates DNA repair^[33] and coordinates cell survival and death^[34,35]. It is reported that *CHEK1* plays an important role in the checkpoint of DNA damage and DNA replication through the ATR-CHK1 pathway^[36-38]. Lin *et al.*^[39] performed a meta-analysis to explore the association of *CHEK1* SNPs with

breast cancer in patients registered in the database of the Utah Breast Cancer Study. They found that *CHEK1* polymorphisms are significantly associated with the risk of breast cancer. However, in that study common alleles of *CHEK1* are not implicated in breast cancer risk or in the survival of breast cancer patients after meta-analysis. Our results showed an association between the *CHEK1* rs515255 genetic variant and a decreased risk of HCC, after adjusting for age, sex, nationality, smoking, drinking, family history of HCC, and HBV infection. The conflicting results may reflect the different cancers evaluated and/or differences in the study population. This remains to be clarified in further investigations.

KAT2B, also known as *PCAF*, encodes the cofactor PCAF (P300/CBP associated factor) of activated nucleoprotein that is important in cell cycle regulation. *KAT2B* induces cell cycle arrest and/or apoptosis by regulating p53 and affects the acetylation and stability of E2F1 in the presence of DNA damage^[40,41]. Overexpression of PCAF was reported in samples of both central nervous system tumors and Wilm's tumors^[42]. In addition, an association between *KAT2B* gene polymorphisms and several human diseases and behaviors has been reported. For example, the *KAT2B* SNP rs9829896 is associated with drug abuse in African Americans^[43]. We also found that the risk of HCC was higher in individuals with the *KAT2B* rs17006625 GG genotype than with the AA genotype, after adjusting for age, sex, nationality, smoking, drinking, family history of HCC, and HBV infection.

HBV infection status, drinking status, and smoking status are well known to influence the occurrence and development of HCC^[44-47]. Moreover, some genotypes have no effect on HCC risk when considered within a population as a whole, but the subgroup analysis may show an effect on HCC risk among alcohol drinkers and/or smokers^[48,49]. Therefore, in our study, we evaluated the role of risk factors such as drinking status and smoking status in a stratified analysis and

found that these environmental factors may interact with the analyzed SNPs.

Our study had several limitations. First, the research population was drawn only from the Guangxi Zhuang Autonomous Region. Whether the results apply to the Chinese population as a whole or to other ethnic groups remains to be seen. Second, because our study used a case-control format, recall bias was difficult to avoid. However, we sought to minimize recall bias by choosing patients newly diagnosed with HCC. Finally, the functional influence of the examined SNPs and the potential mechanisms need to be determined in functional validation tests.

In conclusion, *MCM4* rs2305952 CC and *CHEK1* rs515255 TC, TT, TC/TT may decrease the risk of HCC and *KAT2B* rs17006625 GG may increase the risk of HCC. In addition, we observed an increased risk associated with *KAT2B* rs17006625 GG in HBsAg-negative patients. Furthermore, we also observed a decreased risk associated with *MCM4* rs2305952 CC in HBsAg-positive patients and in also non-drinking patients and non-smoking patients, and with *CHEK1* rs515255 TC, TT, TC/TT in HBsAg-negative patients and in also non-drinking patients. Our results suggest that the genetic variants in the cell cycle pathway genes affect the risk of HCC, however, further studies are needed to confirm the findings.

ACKNOWLEDGMENTS

We sincerely thank the staff of the First Affiliated Hospital of Guangxi Medical University and the Tumor Hospital of Guangxi Medical University for their support in recruiting the study participants. We also thank Da-Fang Chen and his students at the Peking University Health Science Center for technical help.

COMMENTS

Background

The uncontrollable proliferation of cancer cells is a crucial mechanism in cancer development and progression. Previous studies have shown that polymorphisms of cell cycle pathway genes are associated with cancer. However, their relationship with hepatocellular carcinoma (HCC) is unclear.

Research frontiers

Despite reports of an association between polymorphisms in cell cycle pathway genes and cancer risk, little is known about the relationship between these polymorphisms and HCC risk.

Innovations and breakthroughs

This study enrolled 1127 cases newly diagnosed with HCC and 1200 non-tumor patients. It comprehensively investigated the relationship between 15 SNPs in 12 cell cycle pathway genes and HCC risk.

Applications

Since individuals with the *KAT2B* rs17006625 GG genotype may have an increased risk of HCC, they should be carefully monitored to reduce the occurrence and development of HCC.

Terminology

A single nucleotide polymorphism (SNP) is a variation in the genomic DNA sequence. SNPs in some genes may cause an increased or decreased risk of HCC.

Peer-review

The manuscript is interesting and provides relevant information. The study is a descriptive paper analyzing the polymorphism in HCC in a wide number of patients. The analyses are consistent with the results and the conclusions asserted in the manuscript.

REFERENCES

- 1 **Torre LA**, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin* 2015; **65**: 87-108 [PMID: 25651787 DOI: 10.3322/caac.21262]
- 2 **Franceschi S**, Montella M, Polesel J, La Vecchia C, Crispo A, Dal Maso L, Casarin P, Izzo F, Tommasi LG, Chemin I, Trépo C, Crovatto M, Talamini R. Hepatitis viruses, alcohol, and tobacco in the etiology of hepatocellular carcinoma in Italy. *Cancer Epidemiol Biomarkers Prev* 2006; **15**: 683-689 [PMID: 16614109 DOI: 10.1158/1055-9965.EPI-05-0702]
- 3 **Dragani TA**. Risk of HCC: genetic heterogeneity and complex genetics. *J Hepatol* 2010; **52**: 252-257 [PMID: 20022654 DOI: 10.1016/j.jhep.2009.11.015]
- 4 **Kanda M**, Sugimoto H, Kodera Y. Genetic and epigenetic aspects of initiation and progression of hepatocellular carcinoma. *World J Gastroenterol* 2015; **21**: 10584-10597 [PMID: 26457018 DOI: 10.3748/wjg.v21.i37.10584]
- 5 **Labib HA**, Ahmed HS, Shalaby SM, Wahab EA, Hamed EF. Genetic polymorphism of IL-23R influences susceptibility to HCV-related hepatocellular carcinoma. *Cell Immunol* 2015; **294**: 21-24 [PMID: 25666505 DOI: 10.1016/j.cellimm.2015.01.012]
- 6 **Liu F**, Luo LM, Wei YG, Li B, Wang WT, Wen TF, Yang JY, Xu MQ, Yan LN. Polymorphisms of the CYP1B1 gene and hepatocellular carcinoma risk in a Chinese population. *Gene* 2015; **564**: 14-20 [PMID: 25796598 DOI: 10.1016/j.gene.2015.03.035]
- 7 **Son MS**, Jang MJ, Jeon YJ, Kim WH, Kwon CI, Ko KH, Park PW, Hong SP, Rim KS, Kwon SW, Hwang SG, Kim NK. Promoter polymorphisms of pri-miR-34b/c are associated with hepatocellular carcinoma. *Gene* 2013; **524**: 156-160 [PMID: 23632240 DOI: 10.1016/j.gene.2013.04.042]
- 8 **Bretones G**, Delgado MD, León J. Myc and cell cycle control. *Biochim Biophys Acta* 2015; **1849**: 506-516 [PMID: 24704206 DOI: 10.1016/j.bbagr.2014.03.013]
- 9 **Wang Z**, Sturgis EM, Zhang F, Lei D, Liu Z, Xu L, Song X, Wei Q, Li G. Genetic variants of p27 and p21 as predictors for risk of second primary malignancy in patients with index squamous cell carcinoma of head and neck. *Mol Cancer* 2012; **11**: 17 [PMID: 22449259 DOI: 10.1186/1476-4598-11-17]
- 10 **Liu F**, Wei YG, Luo LM, Wang WT, Yan LN, Wen TF, Xu MQ, Yang JY, Li B. Genetic variants of p21 and p27 and hepatocellular cancer risk in a Chinese Han population: a case-control study. *Int J Cancer* 2013; **132**: 2056-2064 [PMID: 23034899 DOI: 10.1002/ijc.27885]
- 11 **Liao D**, Wu Y, Pu X, Chen H, Luo S, Li B, Ding C, Huang GL, He Z. Cyclin D1 G870A polymorphism and risk of nasopharyngeal carcinoma: a case-control study and meta-analysis. *PLoS One* 2014; **9**: e113299 [PMID: 25409185 DOI: 10.1371/journal.pone.0113299]
- 12 **Xue L**, Han X, Liu R, Wang Z, Li H, Chen Q, Zhang P, Wang Z, Chong T. MDM2 and P53 polymorphisms contribute together to the risk and survival of prostate cancer. *Oncotarget* 2015; Epub ahead of print [PMID: 26025918 DOI: 10.18632/oncotarget.3923]
- 13 **Banaszkiewicz M**, Constantinou M, Pietrusiński M, Kepczyński L, Jędrzejczyk A, Roźniecki M, Marks P, Kałużewski B. Concomitance of oncogenic HPV types, CHEK2 gene mutations, and CYP1B1 gene polymorphism as an increased risk factor for malignancy. *Cent European J Urol* 2013; **66**: 23-29 [PMID: 23632240 DOI: 10.1016/j.gene.2013.04.042]

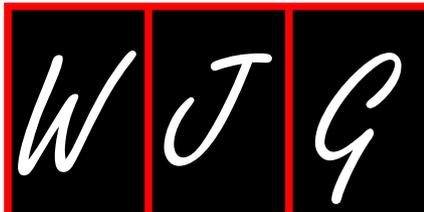
- 24578981 DOI: 10.5173/ceju.2013.01.art7]
- 14 **Wang N**, Wang S, Zhang Q, Lu Y, Wei H, Li W, Zhang S, Yin D, Ou Y. Association of p21 SNPs and risk of cervical cancer among Chinese women. *BMC Cancer* 2012; **12**: 589 [PMID: 23231583 DOI: 10.1186/1471-2407-12-589]
 - 15 **Ullah Shah A**, Mahjabeen I, Kayani MA. Genetic polymorphisms in cell cycle regulatory genes CCND1 and CDK4 are associated with susceptibility to breast cancer. *J BUON* 2015; **20**: 985-993 [PMID: 26416047]
 - 16 **Akbari Z**, Safari-Alighiarloo N, Taleghani MY, Mirfakhkar FS, Asadzadeh Aghdaei H, Vahedi M, Irani Shemirani A, Nazemalhosseini-Mojarad E, Zali MR. Polymorphism of SMAD7 gene (rs2337104) and risk of colorectal cancer in an Iranian population: a case-control study. *Gastroenterol Hepatol Bed Bench* 2014; **7**: 198-205 [PMID: 25289133]
 - 17 **Hardikar S**, Johnson LG, Malkki M, Petersdorf EW, Galloway DA, Schwartz SM, Madeleine MM. A population-based case-control study of genetic variation in cytokine genes associated with risk of cervical and vulvar cancers. *Gynecol Oncol* 2015; **139**: 90-96 [PMID: 26241630 DOI: 10.1016/j.ygyno.2015.07.110]
 - 18 **Qiu M**, Liu Y, Yu X, Qin L, Bei C, Zeng X, Qiu X, Tang B, He S, Yu H. Interaction between p53 codon 72 and MDM2 309T& gt; G polymorphisms and the risk of hepatocellular carcinoma. *Tumour Biol* 2016; **37**: 3863-3870 [PMID: 26476535 DOI: 10.1007/s13277-015-4222-4]
 - 19 **Murali A**, Nalinakumari KR, Thomas S, Kannan S. Association of single nucleotide polymorphisms in cell cycle regulatory genes with oral cancer susceptibility. *Br J Oral Maxillofac Surg* 2014; **52**: 652-658 [PMID: 24947332 DOI: 10.1016/j.bjoms.2014.05.010]
 - 20 **Wang W**, Spitz MR, Yang H, Lu C, Stewart DJ, Wu X. Genetic variants in cell cycle control pathway confer susceptibility to lung cancer. *Clin Cancer Res* 2007; **13**: 5974-5981 [PMID: 17908995 DOI: 10.1158/1078-0432.CCR-07-0113]
 - 21 **Fernández PL**, Jares P, Rey MJ, Campo E, Cardesa A. Cell cycle regulators and their abnormalities in breast cancer. *Mol Pathol* 1998; **51**: 305-309 [PMID: 10193510 DOI: 10.1136/mp.51.6.305]
 - 22 **Park MT**, Lee SJ. Cell cycle and cancer. *J Biochem Mol Biol* 2003; **36**: 60-65 [PMID: 12542976]
 - 23 **Todd R**, Hinds PW, Munger K, Rustgi AK, Opitz OG, Suliman Y, Wong DT. Cell cycle dysregulation in oral cancer. *Crit Rev Oral Biol Med* 2002; **13**: 51-61 [PMID: 12097237 DOI: 10.1177/154411130201300106]
 - 24 **Bailis JM**, Luche DD, Hunter T, Forsburg SL. Minichromosome maintenance proteins interact with checkpoint and recombination proteins to promote s-phase genome stability. *Mol Cell Biol* 2008; **28**: 1724-1738 [PMID: 18180284 DOI: 10.1128/MCB.01717-07]
 - 25 **Yu Z**, Feng D, Liang C. Pairwise interactions of the six human MCM protein subunits. *J Mol Biol* 2004; **340**: 1197-1206 [PMID: 15236977 DOI: 10.1016/j.jmb.2004.05.024]
 - 26 **Huang XP**, Zhang X, Su XD, Ma GW, Zhao JM, Rong TH. [Expression and significance of MCM4 in esophageal cancer]. *Ai Zheng* 2007; **26**: 96-99 [PMID: 17222376]
 - 27 **Das M**, Prasad SB, Yadav SS, Govardhan HB, Pandey LK, Singh S, Pradhan S, Narayan G. Over expression of minichromosome maintenance genes is clinically correlated to cervical carcinogenesis. *PLoS One* 2013; **8**: e69607 [PMID: 23874974 DOI: 10.1371/journal.pone.0069607]
 - 28 **Huber AR**, Tan D, Sun J, Dean D, Wu T, Zhou Z. High expression of carbonic anhydrase IX is significantly associated with glandular lesions in gastroesophageal junction and with tumorigenesis markers BMI1, MCM4 and MCM7. *BMC Gastroenterol* 2015; **15**: 80 [PMID: 26156831 DOI: 10.1186/s12876-015-0310-6]
 - 29 **Ishimi Y**, Komamura-Kohno Y, Kwon HJ, Yamada K, Nakanishi M. Identification of MCM4 as a target of the DNA replication block checkpoint system. *J Biol Chem* 2003; **278**: 24644-24650 [PMID: 12714602 DOI: 10.1074/jbc.M213252200]
 - 30 **Shima N**, Buske TR, Schimenti JC. Genetic screen for chromosome instability in mice: Mcm4 and breast cancer. *Cell Cycle* 2007; **6**: 1135-1140 [PMID: 17495541]
 - 31 **Watanabe E**, Ohara R, Ishimi Y. Effect of an MCM4 mutation that causes tumours in mouse on human MCM4/6/7 complex formation. *J Biochem* 2012; **152**: 191-198 [PMID: 22668557 DOI: 10.1093/jb/mvs060]
 - 32 **Maya-Mendoza A**, Petermann E, Gillespie DA, Caldecott KW, Jackson DA. Chk1 regulates the density of active replication origins during the vertebrate S phase. *EMBO J* 2007; **26**: 2719-2731 [PMID: 17491592 DOI: 10.1038/sj.emboj.7601714]
 - 33 **Sørensen CS**, Hansen LT, Dziegielewska J, Syljuåsen RG, Lundin C, Bartek J, Helleday T. The cell-cycle checkpoint kinase Chk1 is required for mammalian homologous recombination repair. *Nat Cell Biol* 2005; **7**: 195-201 [PMID: 15665856 DOI: 10.1038/ncb1212]
 - 34 **Sahu RP**, Batra S, Srivastava SK. Activation of ATM/Chk1 by curcumin causes cell cycle arrest and apoptosis in human pancreatic cancer cells. *Br J Cancer* 2009; **100**: 1425-1433 [PMID: 19401701 DOI: 10.1038/sj.bjc.6605039]
 - 35 **Smith J**, Tho LM, Xu N, Gillespie DA. The ATM-Chk2 and ATR-Chk1 pathways in DNA damage signaling and cancer. *Adv Cancer Res* 2010; **108**: 73-112 [PMID: 21034966 DOI: 10.1016/B978-0-12-380888-2.00003-0]
 - 36 **Sørensen CS**, Syljuåsen RG. Safeguarding genome integrity: the checkpoint kinases ATR, CHK1 and WEE1 restrain CDK activity during normal DNA replication. *Nucleic Acids Res* 2012; **40**: 477-486 [PMID: 21937510 DOI: 10.1093/nar/gkr697]
 - 37 **Reinhardt HC**, Yaffe MB. Kinases that control the cell cycle in response to DNA damage: Chk1, Chk2, and MK2. *Curr Opin Cell Biol* 2009; **21**: 245-255 [PMID: 19230643 DOI: 10.1016/j.jceb.2009.01.018]
 - 38 **Seiler JA**, Conti C, Syed A, Aladjem MI, Pommier Y. The intra-S-phase checkpoint affects both DNA replication initiation and elongation: single-cell and -DNA fiber analyses. *Mol Cell Biol* 2007; **27**: 5806-5818 [PMID: 17515603 DOI: 10.1128/MCB.02278-06]
 - 39 **Lin WY**, Brock IW, Connley D, Cramp H, Tucker R, Slate J, Reed MW, Balasubramanian SP, Cannon-Albright LA, Camp NJ, Cox A. Associations of ATR and CHEK1 single nucleotide polymorphisms with breast cancer. *PLoS One* 2013; **8**: e68578 [PMID: 23844225 DOI: 10.1371/journal.pone.0068578]
 - 40 **Liu L**, Scolnick DM, Trievel RC, Zhang HB, Marmorstein R, Halazonetis TD, Berger SL. p53 sites acetylated in vitro by PCAF and p300 are acetylated in vivo in response to DNA damage. *Mol Cell Biol* 1999; **19**: 1202-1209 [PMID: 9891054 DOI: 10.1128/MCB.19.2.1202]
 - 41 **Ianari A**, Gallo R, Palma M, Alesse E, Gulino A. Specific role for p300/CREB-binding protein-associated factor activity in E2F1 stabilization in response to DNA damage. *J Biol Chem* 2004; **279**: 30830-30835 [PMID: 15123636 DOI: 10.1074/jbc.M402403200]
 - 42 **Armas-Pineda C**, Arenas-Huertero F, Pérezpeña-Diazconti M, Chico-Ponce de León F, Sosa-Sáinz G, Lezama P, Recillas-Targa F. Expression of PCAF, p300 and Gen5 and more highly acetylated histone H4 in pediatric tumors. *J Exp Clin Cancer Res* 2007; **26**: 269-276 [PMID: 17725108]
 - 43 **Johnson EO**, Hancock DB, Levy JL, Gaddis NC, Page GP, Glasheen C, Saccone NL, Bierut LJ, Kral AH. KAT2B polymorphism identified for drug abuse in African Americans with regulatory links to drug abuse pathways in human prefrontal cortex. *Addict Biol* 2015; Epub ahead of print [PMID: 26202629 DOI: 10.1111/adb.12286]
 - 44 **Marrero JA**, Fontana RJ, Fu S, Conjeevaram HS, Su GL, Lok AS. Alcohol, tobacco and obesity are synergistic risk factors for hepatocellular carcinoma. *J Hepatol* 2005; **42**: 218-224 [PMID: 15664247 DOI: 10.1016/j.jhep.2004.10.005]
 - 45 **Tanaka M**, Katayama F, Kato H, Tanaka H, Wang J, Qiao YL, Inoue M. Hepatitis B and C virus infection and hepatocellular carcinoma in China: a review of epidemiology and control measures. *J Epidemiol* 2011; **21**: 401-416 [PMID: 22041528 DOI: 10.2188/jea.JE20100190]
 - 46 **Gambarin-Gelwan M**. Viral hepatitis, non-alcoholic fatty liver disease and alcohol as risk factors for hepatocellular carcinoma. *Chin Clin Oncol* 2013; **2**: 32 [PMID: 25841911 DOI: 10.3978/

j.issn.2304-3865.2013.09.02]

- 47 **Lin H**, Ha NB, Ahmed A, Ayoub W, Daugherty TJ, Lutchman GA, Garcia G, Nguyen MH. Both HCV and HBV are major causes of liver cancer in Southeast Asians. *J Immigr Minor Health* 2013; **15**: 1023-1029 [PMID: 23864445 DOI: 10.1007/s10903-013-9871-z]
- 48 **Zhang J**, Xu F, Ouyang C. Joint effect of polymorphism in the N-acetyltransferase 2 gene and smoking on hepatocellular carcinoma. *Tumour Biol* 2012; **33**: 1059-1063 [PMID: 22293947 DOI: 10.1007/s13277-012-0340-4]
- 49 **Hsieh YH**, Chang WS, Tsai CW, Tsai JP, Hsu CM, Jeng LB, Bau DT. DNA double-strand break repair gene XRCC7 genotypes were associated with hepatocellular carcinoma risk in Taiwanese males and alcohol drinkers. *Tumour Biol* 2015; **36**: 4101-4106 [PMID: 25944161 DOI: 10.1007/s13277-014-2934-5]

P- Reviewer: Alwahaibi NY, Patial V, Servillo G **S- Editor:** Yu J
L- Editor: Wang TQ **E- Editor:** Ma S





Retrospective Cohort Study

Hepatitis E in Israel: A nation-wide retrospective study

Ortal Erez-Granat, Tamar Lachish, Nili Daudi, Daniel Shouval, Eli Schwartz

Ortal Erez-Granat, Pediatrics Department, Sheba Medical Center, Tel Hashomer, Ramat Gan 5262110, Israel

Tamar Lachish, The Infectious Diseases Unit, Shaare-Zedek Medical Center, Jerusalem 9103102, Israel

Nili Daudi, The Liver Unit, the Hebrew University-Hadassah Medical School, Jerusalem 91120, Israel

Daniel Shouval, The Liver Unit, the Hebrew University-Hadassah Medical Center, Jerusalem 91120, Israel

Eli Schwartz, The Center for Geographic Medicine and Department of Medicine C, Sheba Medical Center, Tel Hashomer, Ramat Gan 5262110, Israel

Author contributions: Erez-Granat O, Schwartz E, Shouval D and Lachish T designed the research; Erez-Granat O performed the research and collected the data by telephone interviews and review of medical records; Daudi N was responsible for the laboratory work and analysis; all authors were responsible for interpretation of the data; Erez-Granat O wrote the paper; Schwartz E, Shouval D and Lachish T revised the paper; all authors were responsible for the final approval of the version to be published.

Supported by Puerto Rico and Gendal foundations (in part); and the Hadassah Liver Unit through an annual payment forwarded by the Hadassah Medical Organization, New-York, United States (to Portoricco-Gendal Endowment).

Institutional review board statement: The study was approved by the Sheba-Medical Centers' institutional review board.

Informed consent statement: All study participants received mail with the study protocol and were asked for consent to participate in this study. All study participants provided informed consent prior to study enrollment.

Conflict-of-interest statement: The authors have no conflict of interest to declare.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external

reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Eli Schwartz, MD, DTMH, Professor, The Center for Geographic Medicine and Department of Medicine C, Sheba Medical Center, Tel Hashomer 52621, Israel. elischwa@post.tau.ac.il
Telephone: +972-3-5308456
Fax: +972-3-5308456

Received: February 4, 2016
Peer-review started: February 9, 2016
First decision: April 14, 2016
Revised: May 11, 2016
Accepted: May 21, 2016
Article in press: May 23, 2016
Published online: June 28, 2016

Abstract

AIM: To investigate the epidemiology, risk factors and clinical course of acute hepatitis E virus (HEV) infection in Israel, an industrialized country.

METHODS: A retrospective analysis of acute HEV cases diagnosed in Israel from 1993 to 2013. Acute HEV was defined by ALT/AST elevation and a positive HEV PCR test or positive anti-HEV-IgM serology. HEV RNA was tested by quantitative reverse transcription PCR. Antibodies to HEV were tested retrospectively using an ELISA assay. HEV-RNA was sequenced using RT-PCR of ORF1 and ORF2 regions to diagnose genotype of the virus. Epidemiologic and clinical data were collected by reviewing the clinical files and through a telephone interview according to a structured questionnaire.

RESULTS: Acute HEV was diagnosed in 68 patients. Among the 59 patients who gave an informed consent and were interviewed, 41% of infections were autochthonous (acquired in Israel), 44% travel-related and 15% imported by foreign workers. Autochthonous patients were mainly females (62.5%), more than half of them pregnant, 26% recalled consuming food or water in areas with poor sanitation, 44% ate non-kosher meat. Fulminant hepatitis developed in 3 patients (5%), all of them were females, two of them with post-partum infection, all acquired the disease in Israel (autochthonous). Israeli travelers with imported infection were predominantly males (73%), acquired the disease in the Indian subcontinent (81%), with 100% reporting having consumed fresh vegetables and drinks with ice cubes abroad. Six patients' sera were tested for genotype and revealed HEV genotype 1 (all cases acquired in the Indian subcontinent).

CONCLUSION: This is the first report which highlights the existence of hepatitis E as an autochthonous infection in Israel. Imported HEV originates mostly from the Indian subcontinent.

Key words: Hepatitis E; Autochthonous; Travel; Foreign workers; Pregnancy; Post-partum; India; Nepal; Indian subcontinent; Israel

© The Author(s) 2016. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: This is the first epidemiologic report on hepatitis E virus (HEV) in Israel. This report demonstrates the significant presence of autochthonous acute HEV in Israel, serving as an example of occurrence in an industrialized country. Suspected risk factors in Israel include consumption of water and food in areas with poor sanitation, exposure to animals and eating a non-Kosher meat. The high risk group for fulminant hepatitis was pregnant women in their final trimester. Additionally, imported HEV, originating mainly from the Indian subcontinent, is also seen in Israel. Awareness of this disease is important both among physicians in Israel as well as those in other industrialized countries.

Erez-Granat O, Lachish T, Daudi N, Shouval D, Schwartz E. Hepatitis E in Israel: A nation-wide retrospective study. *World J Gastroenterol* 2016; 22(24): 5568-5577 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5568.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5568>

INTRODUCTION

Hepatitis E virus (HEV) infection is currently one of the leading causes of viral hepatitis worldwide^[1]. The first epidemic of HEV infection was recognized in India, by a retrospective epidemiologic and serologic survey

performed in India and the United States in the early 1980s. This led to the recognition of HEV as a water-borne associated hepatitis. Similar epidemics were subsequently identified in Central and Southeast Asia, the Middle East and North Africa^[2-4]. The viral genome was later cloned and sequenced using samples of bile obtained from experimentally infected macaques, and the virus was named Hepatitis E^[5-8].

HEV is a single strand RNA virus classified in the genus *Hepevirus*, family *Hepeviridae*^[9]. Identification of four HEV genotypes has subsequently been used to study the molecular epidemiology of HEV infection worldwide. Genotypes 1 and 2, restricted to humans, are mostly seen in developing countries causing large waterborne outbreaks of hepatitis. Genotype 1 is mostly associated with outbreaks in Asia and Africa, whereas genotype 2 has been detected in Mexico and some African countries^[10]. In recent years there has been increasing evidence of an autochthonous HEV infection in industrialized countries, contrasting with previous reports of HEV as only an imported infection from endemic, developing countries. In these cases, genotypes 3 and 4 have been identified, and found to be responsible for sporadic cases of autochthonous hepatitis E in industrialized countries^[11-15]. In contrast to genotypes 1 and 2, genotypes 3 and 4 infect not only humans but also other mammalian species such as pigs, boars, and deer^[11].

HEV is transmitted predominantly *via* the fecal-oral route, causing a self-limiting disease which resolves spontaneously within 4-6 wk^[6]. Occasionally, in immune-suppressed patients and in pregnant women, a fulminant form of hepatitis develops^[16]. Chronic infection has been identified almost exclusively among immunocompromised persons, including organ-transplant recipients, patients receiving cancer chemotherapy, and HIV-infected persons^[17].

Israel is an industrialized country located amid HEV endemic countries and home to immigrants and refugees from African countries (such as Egypt, Sudan and Ethiopia, all endemic for genotype 1 of the virus). Furthermore, since a portion of Israel's population eats only kosher food (*i.e.*, avoiding pork, game meat or seafood), the index of suspicion for autochthonous cases has been low. Data obtained through old and non-validated immune-assays regarding sero-epidemiology of hepatitis E in Israel revealed a seroprevalence of 2.81% and 1.81% in the Jewish and Arab population, respectively^[18]. However, data regarding acute HEV revealed only one case report of acute HEV infection acquired in Israel^[19] while the remaining published cases were travel-related^[20,21].

The aim of this study was to identify whether there is a change in the epidemiology of acute HEV in Israel, with cases acquired in Israel (autochthonous cases) and to characterize the epidemiology, risk factors and clinical presentation of all documented acute HEV infections in patients diagnosed in Israel.

MATERIALS AND METHODS

Study design

A descriptive, retrospective, nation-wide study.

Patient population

The study included all patients diagnosed with acute HEV infection in Israel from October 1993 to 2013 at the laboratory of the Liver Unit at the Hadassah Medical Center in Jerusalem. During the study period, this laboratory was the only reference laboratory in Israel for HEV detection. Epidemiologic and clinical data were collected by reviewing the clinical files and through telephone interviews in accordance with a structured questionnaire. The study was approved by the Sheba-Medical Centers' institutional review board.

Case definitions

"*Definite acute HEV*" was defined as acute hepatitis manifested by ALT/AST elevation and a positive HEV PCR (polymerase chain reaction) test or positive anti-HEV-IgM serology. "*Probable acute HEV*" was defined as acute non-A, non-B, non-C hepatitis with negative HEV PCR and negative anti-HEV-IgM serology but positive for anti-HEV-IgG serology, where serum samples were taken later in the course of the disease, and with a clinical course that fit HEV infection, and no other proven etiology. "*Fulminant hepatitis*" was defined as a rapid development of acute liver injury with evidence of coagulation abnormality, an international normalized ratio (INR) > 1.5, and any degree of hepatic encephalopathy in a patient without pre-existing liver disease; and after exclusion of the conventional etiologies for acute liver failure^[22].

Laboratory tests

Serologic detection of anti-HEV antibodies:

Antibodies to HEV (IgG and IgM) were tested using an ELISA micro-titer plates assay [DS-EIA-anti-HEV-G, DS-EIA-anti-HEV-M, DSI S.R.L. Serronno (VA), Milan, Italy] according to the manufacturer's instructions. Micro-titer plates were coated with HEV peptides able to detect all four genotypes of HEV. Ten μ L serum samples were diluted 1:10 with 90 μ L diluent and incubated for 30 min at 37 °C and rinsed three times, followed by a second incubation with 100 μ L conjugated antibodies for another 30 min at 37 °C. After rinsing three times, 100 μ L substrate was incubated for 30 min at room temperature. Finally, 100 μ L stop solution was added followed by reading the plates at a 450 nm wave length. Sample reading > 0.2 OD were considered as positive.

A pangenotypic evaluation by CDC of 6 serologic assays for IgM against HEV identified the assay manufactured by diagnostic systems, which was used in this study, as having the best performance characteristics. Its diagnostic sensitivity and specificity were 98% and 95.2%, respectively^[23].

We used an assay for the detection of IgG against HEV from the same manufacturer, with a sensitivity of 100% and specificity of 97.5%^[24].

Detection of HEV-RNA by Taqman real time

PCR: Detection of HEV RNA was performed by quantitative reverse transcription PCR (qRT-PCR)^[25]. RNA was extracted from 200 μ L serum with TRI reagent (Bio Lab), then diluted in 10 μ L DEPC water. A 10 μ L RNA aliquot was used for one step RT-PCR assay in a final volume of 20 μ L. The region used for the real time assay is a highly conserved region, junction of ORF's 2/3 of HEV: HEV Forward primer GGTGGTTTCTGGGGTGAC, HEV Reverse primer AGGGGTTGGTTGGATGAA, HEV Probe FAM-TGATTCTCAGCCCTTCGC-BHQ.

HEV sequence: To define HEV genotype in the sera samples, we ran reverse transcription polymerase chain reaction (RT-PCR) of two regions of the virus: ORF1 and ORF2. PCR products were sent for cleaning and sequencing in a service laboratory (HY-lab). We used the programs CHROMAS and CLUSTAL in order to analyze the sequences.

Statistical analysis

Quantitative variables are presented as mean \pm SD or as medians and range. Qualitative variables are presented as frequencies and ratios (percent). The χ^2 and the Fisher's exact tests were applied to assess associations between two qualitative variables. The comparison of quantitative variables between two independent groups was carried out using the two-sample *t*-test or the non-parametric Mann-Whitney test. All tests applied were two-tailed, and a *P*-value of 5% or less was considered statistically significant.

RESULTS

During the years 1993 to 2013, 651 patients with presumed acute non-A, non-B, non-C hepatitis were tested for HEV in Israel. Acute HEV was diagnosed in 68/651 patients (10.45%). Among them, 61 patients (90%) were classified as having "definite acute HEV" confirmed by a positive HEV-RNA PCR result (*n* = 50) or positive anti-HEV-IgM serology (*n* = 10). One patient had a positive PCR result from a stool sample taken abroad. Probable acute HEV was diagnosed in 7 patients.

Altogether the cohort of acute HEV infection included 68 patients, 58.8% male, with a mean age of 39.4 years. The greatest number of patients were between the ages of 17-40 years (63.5%). Comparing acute HEV positive patients with non-A-non-B-non-C-non-E acute hepatitis patients revealed no significant differences in gender or age distribution (Table 1).

Among this cohort of 68 patients with a history of acute HEV, 59 patients gave an informed consent and

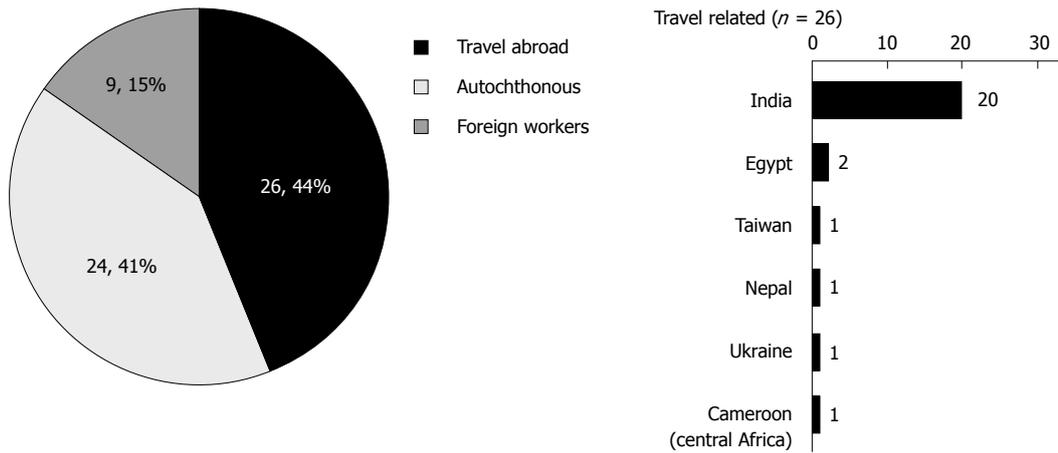


Figure 1 Hepatitis E infection in Israel according to place of acquisition (n = 59). Foreign workers origin: Nepal (n = 5), China (n = 1), 3 unknown.

Table 1 Demographic characteristics of patients with acute hepatitis E vs non A-B-C-E acute hepatitis

	Acute hepatitis	
	Non A-B-C-E hepatitis ¹ n = 583	Hepatitis E ¹ n = 68
Gender		
M	51.0%	58.8%
F	49.0%	41.1%
Age (yr)		
0-16	29 (7.0)	1 (1.6)
17-40	206 (49.8)	40 (63.5)
41-60	132 (31.9)	13 (20.6)
> 60	47 (11.4)	9 (14.3)
Mean	38.96	39.38
Range	0-86	16-87

¹Comparing the groups above in parameters of gender and age revealed non-significant P-value.

were further evaluated. Thirty five patients (35/59, 59%) had "imported" HEV; among them 26 cases (26/59, 44%) were travel-related HEV infections in Israeli patients and 9 cases (9/59, 15%) were diagnosed in foreign workers from HEV endemic countries (Figure 1). The majority (80%) of travel-related HEV cases were acquired in the Indian subcontinent. Finally, 41% of the patients (24/59) did not travel abroad and had no contact with people from endemic areas, and are therefore defined as "autochthonous HEV".

There was a trend of an increasing number of cases diagnosed with acute hepatitis E throughout the study years in both the travel-related and autochthonous groups (Figure 2).

Autochthonous HE

This group consisted of 24 patients, predominantly female (15/24, 62.5%), with a mean age of 42 years old (SD-15, range: 15-69 years old) and without any contact with a foreign worker in Israel. There were, however, 26% (5/19) who recalled consuming food or water from rural settlements and areas of

low sanitation (the West Bank, Bedouin villages) during the 6 wk before the onset of symptoms. Other probable risk factors for HEV infection are summarized in Table 2; 44%, (8/18) ate non-kosher meat (14% ate raw meat, 10% consumed sea food); 40% (8/20) reported contact with animals (cats, dogs, chicken, parrots, geese, fish, guinea pigs, horses or a monkey). Five out of the 24 with autochthonous infections (21%) had chronic liver disease before acquiring HEV (chronic HCV, HBsAg carrier, cystic fibrosis of liver or autoimmune hepatitis). Four of them were diagnosed by positive molecular test (PCR), and one by positive anti-IgM serology for HEV. Eight percent (2/24) received immune suppressing medications (Corticosteroids, Azathioprine, Mycophenolate Mofetil and Tacrolimus). Among the female patients, 53% (8/15) were pregnant or post-partum at the time of clinical presentation.

Fifteen of the patients with autochthonous HEV infection (15/24, 62.5%) were diagnosed by detection of HEV-RNA in their serum. Unfortunately, we were unable to retrospectively test for the genotype in this group of autochthonous patients due to a breakdown in refrigeration.

Travel related HEV

This group consisted 26 patients, predominantly male (19/26, 73%), with a mean age of 37.38 years, SD-16.7. Among the females, 1 out of 7 was pregnant (14.3%). Sixty four percent (16/25) developed symptoms after returning to Israel, with a mean time elapsing before symptoms of 16 d (range: 2-28 d). Duration of travel was on average 62.5 d (range: 3-240 d). Thirty six percent (9/25) were symptomatic before flying back to Israel, and among those patients, the average duration of travel was 6.5 mo (range: 1-24 mo).

Behavior during travel, possibly contributing to risk of infection is summarized in Table 3. As described, the vast majority had contact with suspected contaminated water and raw vegetables. The entire group consisted

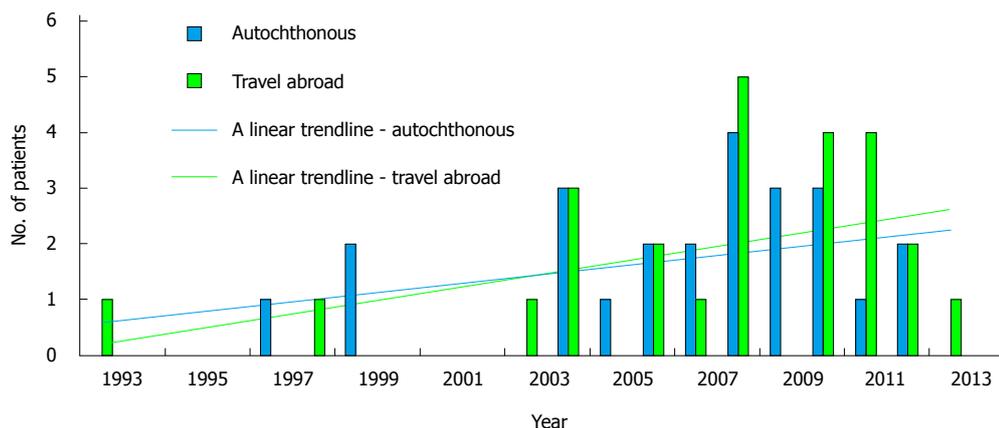


Figure 2 Number of cases of acute hepatitis E virus infection in travel related and autochthonous cases.

Table 2 Clinical characteristics and potential risk factors for hepatitis E virus in patients with autochthonous and travel-related hepatitis E virus infection¹

Character	Autochthonous infection (n = 24)	Travel related (n = 26)	P value ²	
Demography	Gender: M	9 (37.5)	19 (73.1)	0.011
	Age: mean	41.58	37.38	0.358
	Range	15-69	20-74	
Potential risk factors: food related	Eating non-kosher meat	8 (44.4)	22 (81.8)	0.014
	Eating raw meat	3 (14.3)	3 (13.0)	1.000
	Eating sea-food	2 (10.0)	6 (28.6)	0.238
	Consuming food/water from areas with poor sanitation	5 (26.3)	2 (12.5)	0.415
	Contact with animals ³	8 (40.0)	8 (34.8)	0.724
Potential risk factors: others	Pregnancy	8 (53.3)	1 (14.3)	0.165
	Immunosuppression ⁴	2 (8.3)	0 (0)	0.225
	Chronic liver disease	5 (20.8)	1 (3.8)	0.064
Clinical data	Time from onset of symptoms to diagnosis (d)	59.13 (n = 15)	25.21 (n = 19)	0.009
	Duration of symptoms (average weeks)	5.94 (n = 18)	4.08 (n = 20)	0.149
	Hospitalization (percent of patients)	68.2% (15/22)	20 (80.0)	0.345
	Duration of hospitalization (d)	22.15 (n = 13)	11.11 (n = 19)	0.195
Laboratory tests (average)	Bilirubin mg/dL (STD)	10.95 (10.84)	9.24 (5.93)	0.813
	GPT (ALT) U/L (STD)	1169.3 (1279.4)	2446.4 (1604.3)	0.043
	GOT U/L(STD)	1311.7 (2114.6)	1540.4 (1412.7)	0.436
	ALKP (STD)	566.5 (986.1)	205.6 (54.2)	0.673
	GGT U/L (STD)	470.0 (625.1)	232.2 (243.3)	0.730
	LDH U/L (STD)	2613 (6400.5)	1503 (1511.4)	0.440
	ALB g/dL (STD)	3.3 (0.94)	3.9 (0.42)	0.241
	INR (STD)	1.42 (0.8)	1.21 (0.2)	0.791
Outcome	Self-limited	20 (86.9)	26 (100)	0.085
	Fulminant hepatitis	3 (13)	0 (0)	
	Chronic hepatitis	0 (0)	0 (0)	

¹Excluding foreign workers with acute HEV; ²Mann-Whitney U Test, the χ^2 and the Fisher's exact tests were applied as detailed in the text (Methods);

³Contact with animals *i.e.*: cats, dogs, chicken, parrots, goose, fish, guinea pigs, horses, and monkeys; ⁴Immune-suppressed patients: (1) Forty five years old woman treated with immune-suppressed therapy (Imuran) for autoimmune hepatitis; and (2) 19 years old woman treated with immune suppressed therapy after liver and kidney transplantation due to cystic fibrosis.

of healthy travelers, none took immunosuppressive drugs or had a known systemic disease associated with immunosuppression, with the exception of one patient who had chronic hepatitis B, diagnosed in the past. Foreign workers from endemic countries (n = 9) were not included in the travel-related HEV group.

Twenty of the patients with travel-related HEV (20/26, 77%) were diagnosed by detection of HEV-RNA in their serum. Among them, we were able to sequence the HEV genome in six of the patients with

available sera. All cases revealed genotype 1.

Comparing patients with travel-related infection and autochthonous infection revealed that the autochthonous patients were predominantly female (62.5% vs 27%, P < 0.05), had lower levels of alanine aminotransferase on presentation (mean 1169 U/L vs 2446 U/L, P < 0.05) and the time from onset of symptoms to diagnosis was more than twice as long (59 d vs 25 d; P < 0.01) (Table 2). Although there was no significant statistical difference in outcome between the two

Table 3 Potential risk factors for travel-related hepatitis E virus infection

Risk factor	No. of patients (total: <i>n</i> = 26)	Incidence
Gender - M	19/26	73.07%
Pregnancy	1/7	14.28%
Chronic liver disease ¹	1/23	4.30%
Immunosuppression	0/23	0.00%
Eating non-kosher meat	18/22	81.81%
Eating raw meat	3/23	13.04%
Eating sea-food	6/21	28.57%
Drinking tap water abroad	8/23	34.78%
Consuming drinks with ice cubes	23/23	100.00%
Brushing teeth with tap water	20/23	86.95%
Eating fresh vegetables abroad	23/23	100.00%
Bath in fresh water	15/23	65.21%
Contact with animals ²	8/23	34.78%
Contact with travelers having similar symptoms	2/25	8.00%

¹Chronic hepatitis B virus; ²Contact with animals *i.e.*: cat, dog, chicken, parrot, goose, fish, guinea pig, horse and monkey.

groups (due to small sample size), the results may indicate that patients with autochthonous HEV are more prone to fulminant hepatitis than patients with travel-related HEV infection (13% vs 0% respectively). There were no significant differences in age, hospitalization rate and symptom duration between the groups.

Pregnant women

In this study, there were nine cases of HEV in pregnant women (9/28 women in the study, 32%). Only one of the women was a returning traveler from India, while the rest were diagnosed with autochthonous infection (8/9, 88.9%). Fulminant hepatitis occurred in two of the pregnant women (2/9, 22%), without fatality or vertical transmission. A detailed description of this cohort, together with other cases of acute HEV in pregnant women from Western countries, was published recently^[26].

In the entire cohort of the Israeli patients, the most common signs and symptoms were fatigue and non-obstructive-jaundice in 84% (26/31) and 78% (40/51) of patients respectively; 61% (11/18) of patients developed abdominal pain and 41% (19/46) had fever (> 37.5 °C). Other complaints were nausea (40%, 19/48), diarrhea (21%, 10/48) and headache (24%, 11/45). Nine percent (4/43), developed neurologic abnormalities including epilepsy, encephalopathy or loss of consciousness. Admission to hospital in Israel was reported by 78% of patients, for a mean duration of 15.6 d (range: 1-84 d, SD-19.4). One Israeli traveler was admitted to an Indian hospital while abroad where HEV-RNA was identified in a stool sample by PCR.

Results of conventional liver tests and INR at presentation or later were retrieved in only 22 patients (Table 2). The rate of bilirubin level varied between 0.39 to 27.9 mg/dL. No clinically significant bleeding disorders were reported although coagulopathy was

recorded in 41% of the patients (7/17), who presented with prolonged INR up to 3.49. Hepatocellular injury manifested in ALT elevation was significantly higher in patients with travel-related hepatitis compared to autochthonous infection (mean 2246.4 U/dL vs 1169.3 U/dL respectively, $P < 0.05$).

Ninety four percent (46/49) of the patients with HEV infection had self-limiting disease (Table 2). Among the travelers, none of the patients had a chronic or fulminant course of infection. This included a pregnant woman with HEV infection in her third trimester, who had a self-limiting infection, without any pregnancy or fetal related complications. In contrast, among the patients with autochthonous infection, three patients (3/23, 13%) developed fulminant hepatitis. All three were females in their reproductive years, two of them post-partum and the third had cystic fibrosis.

DISCUSSION

The current study was designed to review retrospectively the epidemiology and clinical outcome of acute HEV cases diagnosed in Israel in the past 20 years. During this period, 68 patients were diagnosed with acute HEV infection, and from those patients, 59 gave their consent for evaluation. In this cohort, 44% were travel related, 15% affected foreign workers and surprisingly 41% were autochthonous infection. This is the first study which recognizes the existence of a relatively large cohort of autochthonous HEV infections in Israel, while previous reports suggest that almost all cases of hepatitis E were travel related as summarized in Table 4.

Recently, reports on autochthonous HE in industrialized countries have become more frequent. The distribution of autochthonous vs travel-related cases varies widely between different countries^[13,14,27-30], as can be seen in Table 5. In Israel about half of the cases are autochthonous.

As shown in this report (Figure 2), the number of HEV cases increased during the study years in both travelers and the autochthonous groups. This may reflect an increase in infection rates, but may also be the consequence of improved diagnostic tools and growing awareness among physicians in Israel of HEV infection.

Previous studies in industrialized countries linked sporadic cases of HEV to consumption of undercooked pork^[1,12,31], raw game meat, shellfish^[32] and to blood transfusions^[33,34]. In autochthonous HEV in Israel, the most frequent risk factors included eating non-kosher meat and being exposed to animals. Furthermore, about a quarter of the patients reported that during the 6 wk that preceded the symptoms, they consumed food and/or water from the West Bank or other rural areas in Israel known to have relatively lower hygienic standards as compared to the rest of the country.

Table 4 Summary of the published case reports of acute hepatitis E virus in Israeli patients

Ref.	Year of follow-up	No. of patients	Travel related/autochthonous	Diagnosis
Schwartz <i>et al</i> ^[20]	1992-1998	5	Travel related (all cases acquired in the Indian subcontinent)	Serology tests (Abbott Laboratories, Abbott Park, IL, United States)
Lachish <i>et al</i> ^[21]	1997-2012	19	Travel related (84% acquired in the Indian subcontinent)	Molecular or Serology (IgM/IgG) tests (EIA, Abbott Laboratories, Abbott Park, IL, United States)
Mechnik <i>et al</i> ^[19]	2001	1	Autochthonous	Molecular test (HEV-RNA pos. in serum sample)

HEV: Hepatitis E virus.

Table 5 Hepatitis E virus in industrialized countries¹

Ref.	No. of patients	Travel related	Autochthonous
Norder <i>et al</i> ^[27]	248	88.3%	11.7%
Romanò <i>et al</i> ^[14]	134	81.3%	18.7%
Ramalingam <i>et al</i> ^[28]	16	62.5%	37.5%
Israel (current paper)	68	44.0% ²	41.0%
Drobeniuc <i>et al</i> ^[29]	26	42.0%	58.0%
Chalupa <i>et al</i> ^[13]	49	3.90%	96.1%
Mansuy <i>et al</i> ^[30]	62	3.20%	96.8%

¹A Literature review of the prevalence of HEV infection in industrialized countries^[13,14,27-30]; ²the rest 15% were foreign workers. HEV: Hepatitis E virus.

The other major risk group susceptible to HEV was the Israeli travelers (44% of acute HEV in Israel). In a prospective observational study from Israel, 1% of ill-returning Israeli travelers were diagnosed with acute hepatitis (1997-2012), while HEV has become the most common hepatitis, with a prevalence of 39%, of all hepatitis cases. In the aforementioned study, 84% of the travel-related HEV cases were "imported" from the Indian subcontinent^[21]. In this study, 80.7% of travel associated cases were acquired in the Indian subcontinent. In travel-related HEV, in contrast to autochthonous HEV, the majority of patients were young adult males (73% male, average age of 37 years). Only a minority of patients had chronic liver disease (3.8%). Regarding risk behaviors during travel in this population, although the majority of patients followed pre-travel instructions and did not drink tap water abroad, all patients ate raw vegetables, used ice cubes, brushed their teeth with tap water and bathed in fresh water (Table 3). A control group of travelers without HEV was not available for this study.

The comparison of characteristics of autochthonous and travel-related acute HEV infection, revealed significant differences between the two groups (Table 2); the majority of infected travelers were male, while in the autochthonous group, the majority were female. Time to diagnose was significantly longer in the autochthonous group (on average, more than twice longer), suggesting a delay in diagnosis. ALT levels were significantly higher in infected travelers than in patients with autochthonous infection.

HEV is generally a self-limiting disease, although on rare occasions it can be fulminant^[6,16]. In the present study, most of the patients with acute HEV infection

had a self-limited disease. Patients who had fulminant hepatitis reported one of the following risk factors: pregnancy (including post-partum women), immunosuppression or chronic liver disease.

There is scarce evidence about the mortality of pregnant women with acute HEV infection in industrialized countries^[35-38]. Our previous report indicated that in an industrialized country such as Israel, pregnant women had a high risk of fulminant hepatitis during their final trimester (2/9, 22.2%), though with no mortality or vertical transmission^[26].

Patients with underlying chronic liver disease who develop hepatitis E have a poor prognosis, as they frequently develop acute or sub-acute liver failure^[17]. In a study of a large cohort of patients in India with chronic liver disease, patients who developed HEV associated liver failure had a significantly worse prognosis than patients who decompensated due to other causes. In this cited cohort, the 12-mo mortality with HEV infection reached 70%^[39]. In industrialized countries, smaller studies have also shown a poor prognosis for HEV infected patients with underlying chronic liver disease^[40,41]. In our current study, evaluation of the entire group of 69 patients with acute HEV infection, revealed 7 patients with underlying chronic liver disease (chronic HCV, chronic HBV, cystic fibrotic liver, autoimmune hepatitis, idiopathic cirrhosis). Six of the patients survived and were interviewed and only one patient with chronic HBV infection and cirrhosis died 4 years after the diagnosis of chronic hepatitis B and 3 years after the diagnosis of acute HEV infection.

The differential diagnosis of autochthonous HEV infection includes drug induced liver injury (DILI). Two reports from the United Kingdom and the United States revealed that 21.4% and 3%, respectively, of the patients with an initial diagnosis of DILI, had autochthonous HEV. Such misdiagnosis is particularly common in elderly populations with autochthonous HEV taking various potentially hepato-toxic medications and herbs^[42,43]. In the present cohort, two patients treated with Isoniazide and Ketoconazole who presented with acute hepatitis, were initially misdiagnosed as DILI and subsequently were found positive for HEV RNA by PCR.

In the industrialized world, HEV genotypes 3 and 4 are responsible for sporadic cases of autochthonous HEV infection, where the disease is zoonotic. Imported,

travel related, cases are usually genotypes 1 and 2. In this study, genotyping of 6 travel-related cases with available serum revealed genotype 1 in all cases, a genotype which is prevalent in India where all cases were acquired. One case of hepatitis E in a foreign worker from Nepal who spent one month in Israel, revealed genotype 1 as well.

Israel is located amid countries endemic for HEV, with evidence for genotype 1 in most of the cases reported from those countries. For example, isolation of HEV circulating in Egypt, a country with HEV seroprevalence among the highest in the world, revealed HEV genotype 1 in acute HEV cases, both from rural and urban areas^[44-46]. Studies of North African countries revealed autochthonous infection with genotype 1 as well^[2]. Data on the HEV-genotype circulating in Lebanon, Syria, and Jordan is lacking.

Unfortunately, we were unable to retrospectively test for the genotype in the autochthonous group of patients. The cause included the retrospective design of this study which did not enable storage of large volumes of suspected sera and the physical state of some of the stored sera which were thawed and frozen several times prior to an attempt of sequencing.

However, a recent Israeli study confirmed the presence of HEV in 8.2% of the 169 sewage samples tested throughout the country during 2013-2015^[47]. Sequencing revealed genotype 3 in all sewage samples positive for HEV. These data along with the evidence for acute autochthonous HEV in Israel shown in the present study, suggest autochthonous infection with HEV genotype 3, as seen in Europe and in contrast to the circulating HEV genotype in Israel bordering countries.

The present study has several limitations. The information of exposure related to different risk factors, was based on patients' recollections of events that happened years before the survey, and consequently might be biased. And as mentioned, we were unable to retrospectively test for the HEV genotypes in the autochthonous group of patients. Thus phylogenetic analysis of HEV confirmed autochthonous cases in Israel remains an undertaking for the future.

This report, along with previous reports from Western countries mentioned above, provides evidence for the presence of sporadic autochthonous or travel-related acute HEV cases in the industrialized world. At present, treatment of active HEV infection with viremia with an effective anti-viral agent remains an elusive goal^[48].

To date, two types of recombinant HEV vaccine have been developed^[49-51], but neither of them is commercially available in the Western countries. Additionally both vaccines are genotype 1-based, and although they would be very useful in pregnant women and travelers to endemic regions, their efficacy in preventing HEV infection in non-endemic areas (where other genotypes predominate) needs to be investigated.

COMMENTS

Background

Hepatitis E virus (HEV), one of the leading causes of viral hepatitis worldwide, is transmitted predominantly *via* the fecal-oral route and responsible for epidemics of acute hepatitis in developing countries. However studies have shown that HEV is an emerging infection in industrialized countries as well, with increasing evidence of autochthonous cases (contracted locally) in addition to imported infection among immigrants and travelers from endemic countries. Israel is an industrialized country located amid endemic countries and absorbs immigrants from African countries, however a significant portion of Israel's population eats only kosher food (*i.e.*, do not eat pork, game meat or seafood), therefore the existence of autochthonous cases is not obvious.

Research frontiers

This study is a nation-wide epidemiological study of acute HEV cases diagnosed in Israel between the years 1993-2013. The aim of this manuscript was to identify whether there is any evidence of acute autochthonous HEV in Israel and to characterize the epidemiology, risk factors and clinical presentation of all documented acute HEV infection in Israel.

Innovations and breakthroughs

This report demonstrates, for the first time, the significant presence of autochthonous acute HEV in Israel (41% of all cases were autochthonous). Additionally, the imported HEV cases were from travelers and foreign workers acquiring the disease mainly in the Indian subcontinent.

Applications

The presence of autochthonous cases in Israel highlights the need for medical practitioners to be acquainted with the disease, even in non-endemic countries, and accurate diagnostic means should be easily accessible.

Terminology

"Autochthonous infection" is an infection contracted in the area where reported. Autochthonous HEV infection in Israel relates to patients with acute hepatitis E, with no history of travel during the last six months before disease onset, and who are not immigrants. In contrast, imported infection is one contracted in a country endemic for HEV, and occurs among travelers and immigrants.

Peer-review

The manuscript represents an interesting study of the etiology of acute hepatitis HEV in Israel reporting well documented data of interest to the scientific community.

REFERENCES

- 1 **Kamar N**, Bendall R, Legrand-Abravanel F, Xia NS, Ijaz S, Izopet J, Dalton HR. Hepatitis E. *Lancet* 2012; **379**: 2477-2488 [PMID: 22549046 DOI: 10.1016/S0140-6736(11)61849-7]
- 2 **Purcell RH**, Emerson SU. Hepatitis E: an emerging awareness of an old disease. *J Hepatol* 2008; **48**: 494-503 [PMID: 18192058 DOI: 10.1016/j.jhep.2007.12.008]
- 3 **Khuroo MS**. Study of an epidemic of non-A, non-B hepatitis. Possibility of another human hepatitis virus distinct from post-transfusion non-A, non-B type. *Am J Med* 1980; **68**: 818-824 [PMID: 6770682]
- 4 **Wong DC**, Purcell RH, Sreenivasan MA, Prasad SR, Pavri KM. Epidemic and endemic hepatitis in India: evidence for a non-A, non-B hepatitis virus aetiology. *Lancet* 1980; **2**: 876-879 [PMID: 6107544 DOI: 10.1016/S0140-6736(80)92045-0]
- 5 **Balayan MS**, Andjaparidze AG, Savinskaya SS, Ketiladze ES, Braginsky DM, Savinov AP, Poleschuk VF. Evidence for a virus in non-A, non-B hepatitis transmitted via the fecal-oral route. *Intervirology* 1983; **20**: 23-31 [PMID: 6409836]
- 6 **Hoofnagle JH**, Nelson KE, Purcell RH. Hepatitis E. *N Engl J Med* 2012; **367**: 1237-1244 [PMID: 23013075 DOI: 10.1056/NEJMra1204512]

- 7 **Reyes GR**, Purdy MA, Kim JP, Luk KC, Young LM, Fry KE, Bradley DW. Isolation of a cDNA from the virus responsible for enterically transmitted non-A, non-B hepatitis. *Science* 1990; **247**: 1335-1339 [PMID: 2107574 DOI: 10.1126/science.2107574]
- 8 **Tam AW**, Smith MM, Guerra ME, Huang CC, Bradley DW, Fry KE, Reyes GR. Hepatitis E virus (HEV): molecular cloning and sequencing of the full-length viral genome. *Virology* 1991; **185**: 120-131 [PMID: 1926770 DOI: 10.1016/0042-6822(91)90760-9]
- 9 **Sarin SK**, Kumar M. Hepatitis E. Boyer TD, Manns MP, Sanyal AJ. Zakim and Boyer's Hepatology: a Textbook of liver disease. 6th ed. Chapter 33. 2012: 605-628
- 10 **Mirazo S**, Ramos N, Mainardi V, Gerona S, Arbiza J. Transmission, diagnosis, and management of hepatitis E: an update. *Hepat Med* 2014; **6**: 45-59 [PMID: 24966702 DOI: 10.2147/HMER.S63417]
- 11 **Dalton HR**, Bendall R, Ijaz S, Banks M. Hepatitis E: an emerging infection in developed countries. *Lancet Infect Dis* 2008; **8**: 698-709 [PMID: 18992406 DOI: 10.1016/S1473-3099(08)70255-X]
- 12 **Wichmann O**, Schimanski S, Koch J, Kohler M, Rothe C, Plentz A, Jilg W, Stark K. Phylogenetic and case-control study on hepatitis E virus infection in Germany. *J Infect Dis* 2008; **198**: 1732-1741 [PMID: 18983248 DOI: 10.1086/593211]
- 13 **Chalupa P**, Vasickova P, Pavlik I, Holub M. Endemic hepatitis E in the Czech Republic. *Clin Infect Dis* 2014; **58**: 509-516 [PMID: 24280093 DOI: 10.1093/cid/cit782]
- 14 **Romanò L**, Paladini S, Tagliacarne C, Canuti M, Bianchi S, Zanetti AR. Hepatitis E in Italy: a long-term prospective study. *J Hepatol* 2011; **54**: 34-40 [PMID: 20888660 DOI: 10.1016/j.jhep.2010.06.017]
- 15 **Dalton HR**, Kamar N, Izopet J. Hepatitis E in developed countries: current status and future perspectives. *Future Microbiol* 2014; **9**: 1361-1372 [PMID: 25517900 DOI: 10.2217/fmb.14.89]
- 16 **Teo CG**. Fatal outbreaks of jaundice in pregnancy and the epidemic history of hepatitis E. *Epidemiol Infect* 2012; **140**: 767-787 [PMID: 22273541 DOI: 10.1017/S0950268811002925]
- 17 **Kamar N**, Dalton HR, Abravanel F, Izopet J. Hepatitis E virus infection. *Clin Microbiol Rev* 2014; **27**: 116-138 [PMID: 24396139 DOI: 10.1128/CMR.00057-13]
- 18 **Karetnyi YV**, Favorov MO, Khudyakova NS, Weiss P, Bar-Shani S, Handsher R, Aboudy Y, Varsano N, Schwartz E, Levin E. Serological evidence for hepatitis E virus infection in Israel. *J Med Virol* 1995; **45**: 316-320 [PMID: 7775954 DOI: 10.1002/jmv.1890450314]
- 19 **Mechnik L**, Bergman N, Attali M, Beergabel M, Mosenkis B, Sokolowski N, Malnick S. Acute hepatitis E virus infection presenting as a prolonged cholestatic jaundice. *J Clin Gastroenterol* 2011; **33**: 421-422 [PMID: 11606863]
- 20 **Schwartz E**, Jenks NP, Van Damme P, Galun E. Hepatitis E virus infection in travelers. *Clin Infect Dis* 1999; **29**: 1312-1314 [PMID: 10524982 DOI: 10.1086/313430]
- 21 **Lachish T**, Tandlich M, Schwartz E. Acute hepatitis in Israeli travelers. *J Travel Med* 2013; **20**: 232-236 [PMID: 23809073 DOI: 10.1111/jtm.12039]
- 22 **Lee WM**, Larson AM, Stravitz RT. AASLD Position Paper: The management of Acute Liver Failure: Update 2011. *Hepatology* 2011; **55**: 965-967
- 23 **Drobeniuc J**, Meng J, Reuter G, Greene-Montfort T, Khudyakova N, Dimitrova Z, Kamili S, Teo CG. Serologic assays specific to immunoglobulin M antibodies against hepatitis E virus: pangenotypic evaluation of performances. *Clin Infect Dis* 2010; **51**: e24-e27 [PMID: 20578874 DOI: 10.1086/654801]
- 24 **Ditah I**, Ditah F, Devaki P, Ditah C, Kamath PS, Charlton M. Current epidemiology of hepatitis E virus infection in the United States: low seroprevalence in the National Health and Nutrition Evaluation Survey. *Hepatology* 2014; **60**: 815-822 [PMID: 24824965 DOI: 10.1002/hep.27219]
- 25 **Jothikumar N**, Cromeans TL, Robertson BH, Meng XJ, Hill VR. A broadly reactive one-step real-time RT-PCR assay for rapid and sensitive detection of hepatitis E virus. *J Virol Methods* 2006; **131**: 65-71 [PMID: 16125257 DOI: 10.1016/j.jviromet.2005.07.004]
- 26 **Lachish T**, Erez O, Daudi N, Shouval D, Schwartz E. Acute hepatitis E virus in pregnant women in Israel and in other industrialized countries. *J Clin Virol* 2015; **73**: 20-24 [PMID: 26521225 DOI: 10.1016/j.jcv.2015.10.011]
- 27 **Norder H**, Sundqvist L, Magnusson L, Østergaard Breum S, Löfdahl M, Larsen LE, Hjulsgaard CK, Magnus L, Böttiger BE, Widén F. Endemic hepatitis E in two Nordic countries. *Euro Surveill* 2009; **14**: pii 19211 [PMID: 19442399]
- 28 **Ramalingam S**, Smith D, Wellington L, Vanek J, Simmonds P, MacGillchrist A, Bathgate A, Simpson K, Johannessen I. Autochthonous hepatitis E in Scotland. *J Clin Virol* 2013; **58**: 619-623 [PMID: 24200818 DOI: 10.1016/j.jcv.2013.10.002]
- 29 **Drobeniuc J**, Greene-Montfort T, Le NT, Mixson-Hayden TR, Ganova-Raeva L, Dong C, Novak RT, Sharapov UM, Tohme RA, Teshale E, Kamili S, Teo CG. Laboratory-based surveillance for hepatitis E virus infection, United States, 2005-2012. *Emerg Infect Dis* 2013; **19**: 218-222; quiz 353 [PMID: 23347695 DOI: 10.3201/eid1902.120961]
- 30 **Mansuy JM**, Abravanel F, Miedouge M, Mengelle C, Merviel C, Dubois M, Kamar N, Rostaing L, Alric L, Moreau J, Peron JM, Izopet J. Acute hepatitis E in south-west France over a 5-year period. *J Clin Virol* 2009; **44**: 74-77 [PMID: 18993112 DOI: 10.1016/j.jcv.2008.09.010]
- 31 **Colson P**, Borentain P, Queyriaux B, Kaba M, Moal V, Gallian P, Heyries L, Raoult D, Gerolami R. Pig liver sausage as a source of hepatitis E virus transmission to humans. *J Infect Dis* 2010; **202**: 825-834 [PMID: 20695796 DOI: 10.1086/655898]
- 32 **Said B**, Ijaz S, Kafatos G, Booth L, Thomas HL, Walsh A, Ramsay M, Morgan D. Hepatitis E outbreak on cruise ship. *Emerg Infect Dis* 2009; **15**: 1738-1744 [PMID: 19891860 DOI: 10.3201/eid1511.091094]
- 33 **Boxall E**, Herborn A, Kochethu G, Pratt G, Adams D, Ijaz S, Teo CG. Transfusion-transmitted hepatitis E in a 'nonhyperendemic' country. *Transfus Med* 2006; **16**: 79-83 [PMID: 16623913 DOI: 10.1111/j.1365-3148.2006.00652.x]
- 34 **Matsubayashi K**, Kang JH, Sakata H, Takahashi K, Shindo M, Kato M, Sato S, Kato T, Nishimori H, Tsuji K, Maguchi H, Yoshida J, Maekubo H, Mishiro S, Ikeda H. A case of transfusion-transmitted hepatitis E caused by blood from a donor infected with hepatitis E virus via zoonotic food-borne route. *Transfusion* 2008; **48**: 1368-1375 [PMID: 18651907 DOI: 10.1111/j.1537-2995.2008.01722.x]
- 35 **Khuroo MS**, Kamili S, Jameel S. Vertical transmission of hepatitis E virus. *Lancet* 1995; **345**: 1025-1026 [PMID: 7723501 DOI: 10.1016/S0140-6736(95)90761-0]
- 36 **Anty R**, Ollier L, Péron JM, Nicand E, Cannavo I, Bongain A, Giordanengo V, Tran A. First case report of an acute genotype 3 hepatitis E infected pregnant woman living in South-Eastern France. *J Clin Virol* 2012; **54**: 76-78 [PMID: 22336086 DOI: 10.1016/j.jcv.2012.01.016]
- 37 **Andersson MI**, Hughes J, Gordon FH, Ijaz S, Donati M. Of pigs and pregnancy. *Lancet* 2008; **372**: 1192 [PMID: 18926280 DOI: 10.1016/S0140-6736(08)61486-5]
- 38 **Thoden J**, Venhoff N, Miehle N, Klar M, Huzly D, Panther E, Jilg N, Kunze M, Warnatz K. Hepatitis E and jaundice in an HIV-positive pregnant woman. *AIDS* 2008; **22**: 909-910 [PMID: 18427215 DOI: 10.1097/QAD.0b013e3282f7cb9a]
- 39 **Kumar Acharya S**, Kumar Sharma P, Singh R, Kumar Mohanty S, Madan K, Kumar Jha J, Kumar Panda S. Hepatitis E virus (HEV) infection in patients with cirrhosis is associated with rapid decompensation and death. *J Hepatol* 2007; **46**: 387-394 [PMID: 17125878]
- 40 **Dalton HR**, Hazeldine S, Banks M, Ijaz S, Bendall R. Locally acquired hepatitis E in chronic liver disease. *Lancet* 2007; **369**: 1260 [PMID: 17434400 DOI: 10.1016/S0140-6736(07)60595-9]
- 41 **Péron JM**, Bureau C, Poirson H, Mansuy JM, Alric L, Selves J, Dupuis E, Izopet J, Vinel JP. Fulminant liver failure from acute autochthonous hepatitis E in France: description of seven patients with acute hepatitis E and encephalopathy. *J Viral Hepat* 2007; **14**: 298-303 [PMID: 17439518 DOI: 10.1111/j.1365-2893.2007.00858.x]
- 42 **Dalton HR**, Fellows HJ, Stableforth W, Joseph M, Thurairajah

- PH, Warshow U, Hazeldine S, Remnarace R, Ijaz S, Hussaini SH, Bendall RP. The role of hepatitis E virus testing in drug-induced liver injury. *Aliment Pharmacol Ther* 2007; **26**: 1429-1435 [PMID: 17850420 DOI: 10.1111/j.1365-2036.2007.03504.x]
- 43 **Davern TJ**, Chalasani N, Fontana RJ, Hayashi PH, Protiva P, Kleiner DE, Engle RE, Nguyen H, Emerson SU, Purcell RH, Tillmann HL, Gu J, Serrano J, Hoofnagle JH. Acute hepatitis E infection accounts for some cases of suspected drug-induced liver injury. *Gastroenterology* 2011; **141**: 1665-1672.e1-9 [PMID: 21855518 DOI: 10.1053/j.gastro.2011.07.051]
- 44 **Blackard JT**, Rouster SD, Nady S, Galal G, Marzuuk N, Rafaat MM, Daef E, El Din SS, Purcell RH, Emerson SU, Sherman KE, Shata MT. Genotypic characterization of symptomatic hepatitis E virus (HEV) infections in Egypt. *J Clin Virol* 2009; **46**: 140-144 [PMID: 19651539 DOI: 10.1016/j.jcv.2009.07.007]
- 45 **Tsarev SA**, Binn LN, Gomatos PJ, Arthur RR, Monier MK, van Cuyck-Gandre H, Longer CF, Innis BL. Phylogenetic analysis of hepatitis E virus isolates from Egypt. *J Med Virol* 1999; **57**: 68-74 [PMID: 9890424 DOI: 10.1002/(SICI)1096-9071(199901)57:1<68::AID-JMV10>3.0.CO;2-E]
- 46 **Delarocque-Astagneau E**, Abravanel F, Moshen A, Le Fouler L, Gad RR, El-Daly M, Ibrahim EM, El-Aidy S, Lashin T, El-Hoseiny M, Izopet J, Mohamed MK, Fontanet A, Abdel Hamid M. Epidemiological and virological characteristics of symptomatic acute hepatitis E in Greater Cairo, Egypt. *Clin Microbiol Infect* 2012; **18**: 982-988 [PMID: 22264267 DOI: 10.1111/j.1469-0691.2011.03727.x]
- 47 **Ram D**, Manor Y, Gozlan Y, Schwartz E, Ben-Ari Z, Mendelson E, Mor O. Hepatitis E virus genotype 3 in sewage and genotype 1 in sporadic acute hepatitis cases in Israel. *Am J Trop Med Hyg* 2016; **61**: 331-335
- 48 Hepatitis E vaccine: why wait? *Lancet* 2010; **376**: 845 [PMID: 20833284 DOI: 10.1016/S0140-6736(10)61393-1]
- 49 **Haffar S**, Bazerbachi F, Lake JR. Making the case for the development of a vaccination against hepatitis E virus. *Liver Int* 2015; **35**: 311-316 [PMID: 24836400 DOI: 10.1111/liv.12590]
- 50 **Shrestha MP**, Scott RM, Joshi DM, Mammen MP, Thapa GB, Thapa N, Myint KS, Fourneau M, Kuschner RA, Shrestha SK, David MP, Seriwatana J, Vaughn DW, Safary A, Endy TP, Innis BL. Safety and efficacy of a recombinant hepatitis E vaccine. *N Engl J Med* 2007; **356**: 895-903 [PMID: 17329696 DOI: 10.1056/NEJMoa061847]
- 51 **Zhu FC**, Zhang J, Zhang XF, Zhou C, Wang ZZ, Huang SJ, Wang H, Yang CL, Jiang HM, Cai JP, Wang YJ, Ai X, Hu YM, Tang Q, Yao X, Yan Q, Xian YL, Wu T, Li YM, Miao J, Ng MH, Shih JW, Xia NS. Efficacy and safety of a recombinant hepatitis E vaccine in healthy adults: a large-scale, randomised, double-blind placebo-controlled, phase 3 trial. *Lancet* 2010; **376**: 895-902 [PMID: 20728932 DOI: 10.1016/S0140-6736(10)61030-6]

P- Reviewer: Rodriguez-Frias F, Shenoy SM **S- Editor:** Ma YJ
L- Editor: O'Neill M **E- Editor:** Ma S



Retrospective Cohort Study

Comprehensive mutation screening for 10 genes in Chinese patients suffering very early onset inflammatory bowel disease

Yuan Xiao, Xin-Qiong Wang, Yi Yu, Yan Guo, Xu Xu, Ling Gong, Tong Zhou, Xiao-Qin Li, Chun-Di Xu

Yuan Xiao, Xin-Qiong Wang, Yi Yu, Yan Guo, Xu Xu, Ling Gong, Tong Zhou, Chun-Di Xu, Pediatric Department, Ruijin Hospital and Ruijin Hospital North, Shanghai Jiao Tong University, School of Medicine, Shanghai 200025, China

Xiao-Qin Li, Gastroenterology Department, Zhengzhou Children's Hospital, Zhengzhou 450053, Henan Province, China

Author contributions: Xiao Y and Wang XQ contributed equally to this work; Xiao Y and Xu CD designed this study; Xiao Y and Wang XQ performed the next generation sequencing; Yu Y, Guo Y, Xu X and Gong L collected the patients and recorded the data; Zhou T and Li XQ analyzed the data.

Supported by National Nature Science Foundation of China, No. 81400588.

Institutional review board statement: The study was reviewed and approved by the ethics committee of the Ruijin Hospital, Shanghai Jiao Tong University School of Medicine.

Informed consent statement: All the participants provided informed written consent prior to study enrollment.

Conflict-of-interest statement: All the authors listed declare no conflicts of interest.

Data sharing statement: Technical appendix, statistical code, and dataset is available from the corresponding author at chundixu55@163.com.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Chun-Di Xu, MD, PhD, Pediatric Department, Ruijin Hospital and Ruijin Hospital North, Shanghai

Jiao Tong University, School of Medicine, No. 197, Ruijin Er Road, Shanghai 200025, China. chundixu55@163.com
Telephone: +86-21-64370045
Fax: +86-21-64333414

Received: February 28, 2016
Peer-review started: February 29, 2016
First decision: March 31, 2016
Revised: April 13, 2016
Accepted: April 20, 2016
Article in press: April 20, 2016
Published online: June 28, 2016

Abstract

AIM: To perform sequencing analysis in patients with very early-onset inflammatory bowel disease (VEO-IBD) to determine the genetic basis for VEO-IBD in Chinese pediatric patients.

METHODS: A total of 13 Chinese pediatric patients with VEO-IBD were diagnosed from May 2012 and August 2014. The relevant clinical characteristics of these patients were analyzed. Then DNA in the peripheral blood from patients was extracted. Next generation sequencing (NGS) based on an Illumina-Miseq platform was used to analyze the exons in the coding regions of 10 candidate genes: *IL-10*, *IL-10RA*, *IL-10RB*, *NOD2*, *FUT2*, *IL23R*, *GPR35*, *GPR65*, *TNFSF15*, and *ADAM30*. The Sanger sequencing was used to verify the variations detected in NGS.

RESULTS: Out of the 13 pediatric patients, ten were diagnosed with Crohn's disease, and three diagnosed with ulcerative colitis. Mutations in *IL-10RA* and *IL-10RB* were detected in five patients. There were four patients who had single nucleotide polymorphisms associated with IBD. Two patients had *IL-10RA* and

FUT2 polymorphisms, and two patients had *IL-10RB* and *FUT2* polymorphisms. Gene variations were not found in the rest four patients. Children with mutations had lower percentile body weight (1.0% *vs* 27.5%, $P = 0.002$) and hemoglobin (87.4 g/L *vs* 108.5 g/L, $P = 0.040$) when compared with children without mutations. Although the age of onset was earlier, height was shorter, and the response to treatment was poorer in the mutation group, there was no significant difference in these factors between groups.

CONCLUSION: *IL-10RA* and *IL-10RB* mutations are common in Chinese children with VEO-IBD. Patients with mutations have an earlier disease onset, lower body weight and hemoglobin, and poorer prognosis.

Key words: Pediatric inflammatory bowel disease; Very early-onset inflammatory bowel disease; Interleukin 10 receptor; *NOD2* gene; *FUT2* gene

© The Author(s) 2016. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: In this small-sample size study, we performed next generation sequencing for 10 candidate genes in Chinese pediatric patients with very early onset inflammatory bowel disease. We found that *IL-10RA* and *IL-10RB* mutations were common. There were five patients harbouring mutations in these two genes and accounted for 38.5% of all samples. Besides, there were four patients who had single nucleotide polymorphisms associated with inflammatory bowel disease. Pediatric patients with mutations had an earlier disease onset, lower body weight, markedly lower hemoglobin, and poorer prognosis.

Xiao Y, Wang XQ, Yu Y, Guo Y, Xu X, Gong L, Zhou T, Li XQ, Xu CD. Comprehensive mutation screening for 10 genes in Chinese patients suffering very early onset inflammatory bowel disease. *World J Gastroenterol* 2016; 22(24): 5578-5588 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5578.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5578>

INTRODUCTION

Inflammatory bowel disease (IBD) is a chronic and recurrent gastrointestinal inflammatory disease in children. Based on clinical characteristics, laboratory tests, and endoscopic and pathological presentations, IBD can be subdivided into Crohn's disease (CD), ulcerative colitis (UC), and IBD-unclassified (IBD-U)^[1]. Our previous study showed that the annual incidence of IBD in the 0- to 14-year age group of Shanghai residents steadily increased from 2000 to 2010^[2]. Although pediatric IBD mainly occurs in adolescence^[2], approximately 15% of IBD pediatric patients have very early-onset IBD (VEO-IBD) that begins before 6

years of age, and 1% of children develop this disease before reaching 1 year of age^[3,4]. The majority of VEO-IBD cases have clinical characteristics that are distinct from those of classic IBD with adult and adolescent onset. VEO-IBD has more severe clinical symptoms, resistance to a variety of immunosuppressive therapies, and a poor prognosis after conventional treatments. Some scholars even consider VEO-IBD to be a completely different disease from classic IBD^[5].

Previous studies suggested that persistent intestinal immune dysfunction in a genetically susceptible individual exposed to adverse environmental factors is an important mechanism for IBD development. Genome-wide association studies (GWAS) have discovered a total of 163 loci associated with the risk for IBD development^[6]. However, disease onset at an early stage of life suggests a leading role for rare gene variations in VEO-IBD patients, especially in children with a disease onset before the age of 1 year. These low frequency mutations are difficult to detect using GWAS. Next generation sequencing technology allows for the high-throughput sequencing of exons in a series of genes concurrently; therefore, rare gene variations can be discovered^[7]. Since Glocker *et al*^[8] first discovered in 2009 that mutations in genes encoding the α subunit (*IL-10R1*, encoding gene *IL-10RA*) and the β subunit (*IL-10R2*, encoding gene *IL-10RB*) of the interleukin-10 (*IL-10*) receptor could induce VEO-IBD development, a few studies have continuously discovered mutations in genes encoding *IL-10R1*, *IL-10R2*, and *IL-10*^[5,9-12]. However, current reports are limited, and the majority of studies are small-size case studies. Reports on the Han Chinese population are scarcer^[13,14].

This study used the Illumina-Miseq platform to sequence candidate genes in Han Chinese children diagnosed with VEO-IBD. The candidate genes included genes involved in the *IL-10* signaling pathway, such as *IL-10*, *IL-10RA*, and *IL-10RB*, and genes highly associated with the development of CD in previous studies, including *NOD2*, *FUT2*, *IL-23R*, *GPR35*, *GPR65*, *TNFSF15*, and *ADAM30*. This study furthers our understanding of the genetic factors associated with VEO-IBD development in Han Chinese children.

MATERIALS AND METHODS

Patient consent and ethic committee approval

Verbal and written consent was obtained from the parents of all of children included this study. Ethic committee approval for the study was granted by Institutional Review Boards of Ruijin Hospital affiliated to Shanghai Jiao Tong University School of Medicine.

Study subjects

A total of 13 pediatric patients with repeated diarrhea, mucus and bloody stool, or abdominal pain who were diagnosed by laboratory tests and digestive endoscopy with VEO-IBD in the Pediatric Department of Ruijin

Hospital of Shanghai Jiao Tong University School of Medicine between May 2012 and August 2014 were included in this study. All of the patients were Han Chinese. VEO-IBD was defined as IBD onset before the age of 6 years, and a disease onset before 2 years of age was called infantile-onset IBD^[15,16]. The clinical characteristics of these pediatric patients, including gender, age of disease onset, body height, body weight, family history, clinical symptoms, complications, major laboratory examinations, endoscopic presentations, and therapeutic effects, were retrospectively analyzed.

Laboratory and digestive endoscopic examinations

Relevant laboratory examinations, including complete blood count (CBC), C-reactive protein (CRP), erythrocyte sedimentation rate (ESR), tumor necrosis factor α (TNF- α) level, immunoglobulins G, A, M, and E, vitamin D, human Immunodeficiency virus (HIV) and human cytomegalovirus (CMV) antibody detection in serum, T lymphocyte flow cytometry sorting, stool parasite tests, stool culture, and stool *Clostridium difficile* toxin detection, were performed when the patients were admitted to the hospital. Common infectious diseases and primary immunodeficiency diseases were excluded.

All patients received a colonoscopy under general anesthesia. A biopsy of the colonic mucosa under endoscopy was performed for pathological examination.

Illumina-Miseq platform sequencing

Genomic DNA extraction: After obtaining verbal and written informed consent from the patients' parents, genomic DNA in the peripheral blood from 13 pediatric patients was extracted using a FlexiGene DNA Kit (Qiagen Inc., Germany). Another 100 copies of DNA extracted from patients suffering from idiopathic short stature (ISS) in previous research were used to test frequency of mutant sites which were newly detected in our study.

Multiplex PCR primer design: Based on the stability of the Illumina-Miseq experiment and the operability of subsequent steps, the length requirement of target fragments for sequencing was < 400 bp. If the length of an exon was longer than 400 bp, an additional pair of primers was designed with overlapping bases of adjacent fragments. To avoid a high number of non-target fragment products, primers were grouped and suspended in a primer mix before the multiplex PCR was performed. The concentration of each primer in the primer mix was 10 mmol/L. The basic requirement for grouping was the lack of matching sequences between two of the amplified products. Oligo 7 software was used to design primers for exons of the encoding region of the 10 candidate genes: *IL-10*, *IL-10RA*, *IL-10RB*, *NOD2*, *FUT2*, *IL23R*, *GPR35*, *GPR65*, *TNFSF15*, and *ADAM30*. A total of 86 pairs of

primers were designed. The sequences are shown in supplementary Table 1.

Multiplex PCR amplification of candidate genes:

A Qiagen Multiple PCR Kit was used in this study. The PCR amplification reaction system had a total volume of 21 μ L, including 4 μ L of ddH₂O, 2 μ L of Q-solution (5 \times), 4 μ L of 10 mmol/L primer mix, 10 μ L of buffer mix, and 1 μ L of the DNA template (20 ng/ μ L). The reaction procedure consisted of pre-denaturation at 94 $^{\circ}$ C for 15 min, denaturation at 94 $^{\circ}$ C for 40 s, annealing at 63 $^{\circ}$ C for 1 min, and extension at 72 $^{\circ}$ C for 40 s. After each cycle, the annealing temperature was reduced by 0.5 $^{\circ}$ C for 10 cycles until the annealing temperature reached 58 $^{\circ}$ C. Next, the amplification was continued for 30 cycles with a constant annealing temperature of 58 $^{\circ}$ C. The final extension at 72 $^{\circ}$ C lasted for 10 min. The PCR products were stained with 100 \times GelRed and subjected to 1% agarose electrophoresis (120 V for 60 min).

The purified multiplex PCR products were sent to Shanghai South Gene Technology Co., Ltd. for sequencing analysis with the Illumina-Miseq platform. After sequencing, the nucleotide sequence information was compared with the standard gene sequences available in GenBank. The obtained gene mutation sites were compared with information in the dbSNP, HGMD, and OMIM databases to determine if the mutations had been previously reported.

To confirm the accuracy of the results, the corresponding gene sequences for the mutations discovered using the Illumina-Miseq platform were sequenced again using the Sanger sequencing method.

The newly discovered gene variation sites were analyzed to predict their influence on protein functions using two online databases: SIFT (<http://sift.jcvi.org/>) and PolyPhen 2 (<http://genetics.bwh.harvard.edu/pph2/>).

Statistical analysis

According to the sequencing results, the 13 pediatric patients were divided into two groups. The patients who harboured pathogenic mutations were in group 1. Those without pathogenic mutations (including presence of polymorphisms only or wild type) were in group 2. The differences in diagnosis, age of disease onset, growth indicators (percentiles of body weight and height were calculated according to WHO standards), complications (perianal diseases and recurrent infection), and therapeutic effects among all groups were compared. Because the sample size was small, quantitative and ranked ordinal data were subjected to nonparametric statistics. The Mann-Whitney Test was performed, and the difference was statistically analyzed using exact probability. SPSS13.0 for Windows software was used for the statistical analysis. $P < 0.05$ indicated a significant difference.

Table 1 Genotypes of 13 patients diagnosed with very early-onset inflammatory bowel disease

Patient	Gene	Variation	Homo/Heterozygote	Function defect
1	<i>IL-10RA</i>	p.R101W	Homozygote	Yes
2	<i>IL-10RA</i>	p.R101W	Compound	Yes
3	<i>IL-10RA</i>	p.V100G (novel mutation)	heterozygote	Pathogenic supporting by Polyphen 2 and SIFT
		p.R101W	Compound	Yes
4	<i>IL-10RA</i>	p.Y64C (novel mutation)	heterozygote	Pathogenic supporting by Polyphen 2 and SIFT
		p.R117H (rs199989396)	Heterozygote	Yes
5	<i>NOD2</i>	p.R703C (rs5743277)	Heterozygote	Susceptibility to CD recorded in HGMD
	<i>FUT2</i>	p.I140F (rs1047781)	Heterozygote	Susceptibility to CD in Chinese population reported by Hu <i>et al</i> ^[31]
	<i>IL-10RB</i>	p.K47E (rs2834167)	Homozygote	SNP in a VEO-UC child reported by Galatola <i>et al</i> ^[29]
6	<i>IL-10RA</i>	p.E141K (rs387907326)	Heterozygote	Pathogenic supporting by Polyphen 2 and SIFT
		p.P115P (rs22280554)	Homozygote	Susceptibility to VEO-IBD reported by Moran <i>et al</i> ^[30]
	<i>FUT2</i>	p.I224V (rs22280555)	Homozygote	
	<i>FUT2</i>	p.I140F (rs1047781)	Heterozygote	Susceptibility to CD in Chinese population reported by Hu <i>et al</i> ^[31]
7	<i>IL-10RA</i>	p.P115P (rs22280554)	Homozygote	Susceptibility to VEO-IBD reported by Moran <i>et al</i> ^[30]
		p.I224V (rs22280555)	Homozygote	
	<i>FUT2</i>	p.I140F (rs1047781)	Heterozygote	Susceptibility to CD in Chinese population reported by Hu <i>et al</i> ^[31]
8	<i>IL-10RB</i>	p.K47E (rs2834167)	Homozygote	SNP in a VEO-UC child reported by Galatola <i>et al</i> ^[29]
	<i>FUT2</i>	p.I140F (rs1047781)	Heterozygote	Susceptibility to CD in Chinese population reported by Hu <i>et al</i> ^[31]
9	<i>IL-10RB</i>	p.K47E (rs2834167)	Heterozygote	SNP in a VEO-UC child reported by Galatola <i>et al</i> ^[29]
	<i>FUT2</i>	p.I140F (rs1047781)	Heterozygote	Susceptibility to CD in Chinese population reported by Hu <i>et al</i> ^[31]

Patients 10, 11, 12 and 13 were wild types in all these genes. CD: Crohn's disease; UC: Ulcerative colitis; VEO-IBD: Very early-onset inflammatory bowel disease; VEO-UC: Very early-onset ulcerative colitis; SNP: Single nucleotide polymorphism; HGMD: The Human Gene Mutation Database (<http://www.hgmd.cf.ac.uk/ac/index.php>).

RESULTS

Genotyping in VEO-IBD patients

***IL-10RA*, *IL-10RB*, and *IL-10* mutations:** *IL-10RA* mutations were detected in four patients, an *IL-10RB* mutation was detected in one patient, and an *IL-10* mutation was not detected in any of the 13 patients.

The detected *IL-10RA* mutations were all in exon 3: c.A191G (p.Y64C), c.T299G (p.V100G), c.C301T (p.R101W), and c.G350A (rs199989396) (p.R117H). The p.R101W mutation was the most common and was detected in three patients (patients 1-3). The other mutations were detected in only one patient. Patient 1 had a homozygous mutation, patients 2 and 3 had compound mutations, and patient 4 had a heterozygous mutation (Table 1 and Figure 1).

Among detected *IL-10RA* mutations, p.Y64C and p.V100G were new mutations that were predicted to be deleterious by SIFT and Polyphen 2. These novel mutant sites were not found in 100 ISS children. The other two mutations had been confirmed to be deleterious in several studies^[5,12,14,17].

An *IL-10RB* heterozygous mutation was detected in one patient (patient 5) (Table 1 and Figure 1). This c.G421A (p.E141K) (rs387907326) mutation was located in exon 4 and was also predicted as a deleterious mutation by SIFT and Polyphen 2. A nonsense mutation in the same site was detected in previous studies^[11,18].

Candidate gene polymorphisms

After the sequence analysis of the coding regions of 10 candidate genes, we found that six patients (patient 4, 5, 6, 7, 8 and 9) had many IBD-associated single nucleotide polymorphisms (SNPs) in *IL-10RA*, *IL-10RB*, *NOD2*, and *FUT2*. The SNP loci in *IL-10RA* were rs22280554: c.G525A, p. P175P and rs22280555: c.A670G, p.I224V; the SNP locus in *IL-10RB* was rs2834167: c.A139G, p.K47E; the SNP locus in *NOD2* was rs5743277: c.C2107T, p.R703C; and the SNP locus in *FUT2* was rs1047781: c.A418T, p.I140F (Table 1).

In addition to the detected p.R117H heterozygous mutation in *IL-10RA*, patient 4 also had heterozygous SNPs in *NOD2* and *FUT2*.

Patient 5 had a heterozygous p.E141K mutation (rs387907326) in *IL-10RB* and SNPs in *IL-10RA*, *IL-10RB*, and *FUT2*. The SNP loci in *IL-10RA* were rs22280554 and rs22280555. The homozygous SNP loci for *IL-10RB* were rs2834167. The SNP in *FUT2* was heterozygous.

Patients 6 and 7 had SNPs in *IL-10RA* and *FUT2*. Patients 8 and 9 had SNPs in *IL-10RB* and *FUT2*.

Four patients did not show any IBD-associated variations in the coding regions of the 10 candidate genes.

There was no IBD-associated variation discovered in the coding regions of six genes: *IL-10*, *IL-23R*, *GPR35*, *GPR65*, *TNFSF15*, and *ADAM30*.

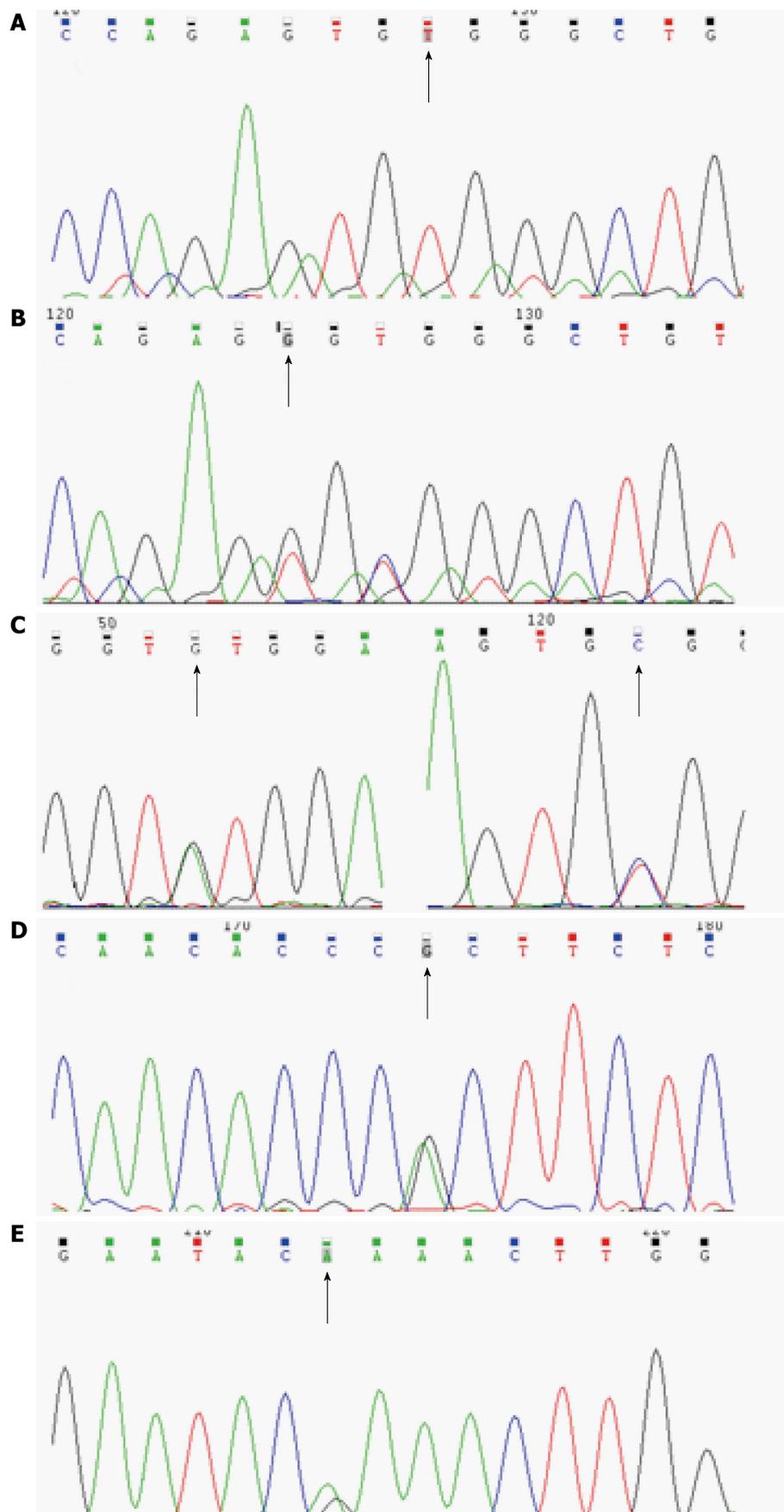


Figure 1 Causative mutations in *IL-10RA* (A-D) or *IL-10RB* (E). A: Patient 1, c.C301T, p.R101W, homozygote; B: Patient 2, c.T299G, p.V100G and c. C301T, p.R101W, compound heterozygote; C: Patient 3, c.A191G, p.Y64C and c. C301T, p.R101W, compound heterozygote; D: Patient 4, c.G35A, p.R117H (rs199989396), heterozygote; E: Patient 5, c.G421A, p.E141K (rs387907326), heterozygote.

Clinical characteristics of VEO-IBD pediatric patients

Out of the 13 VEO-IBD pediatric patients in this study, ten were diagnosed with CD (M:F = 9:1) and three had UC (M:F = 1:2). The mean age of disease onset was 5.8 ± 9.7 mo (range: birth to 3 years of age). None of the parents of the patients had a consanguineous marriage. Patient 8 had a brother that died as a neonate because of repeated diarrhea after birth. There was no clear diagnosis made at that time. The clinical symptoms of the pediatric patients included repeated abdominal pain (13/13), diarrhea (11/13), mucus and bloody stool (11/13), failure to thrive (8/13), recurrent infection (7/13), and perianal fistulas and abscesses (5/13). The colonoscopic presentation of patients with causative mutations showed pancolitis, cobblestone-like changes in mucosa, and deep and large ulcers (Figure 2). All patients received immunosuppressive treatment with glucocorticoids, 6-mercaptopurine and/or infliximab, and thalidomide; however, varying therapeutic effects were observed. Two patients died from severe sepsis or intestinal failure, 2 patients showed no change, 4 patients showed a partial alleviation of symptoms, and 5 patients showed complete clinical remission (Table 2).

Clinical characteristics of different genotypes

Based on the presence of causative mutations in *IL-10RA* and *IL-10RB*, 13 patients were divided into two groups for analysis (group 1: causative mutations in *IL-10RA* or *IL-10RB*; group 2: polymorphisms and no causative mutations). The five patients in group 1 were all diagnosed with CD (100%). Four of these patients had recurrent infections (80%), and three patients had perianal diseases (60%). In group 2 (eight patients), five patients were diagnosed with CD (62.5%), and the other three patients were diagnosed with UC (37.5%). There were only three (37.5%) and two (25%) patients that had recurrent infections and perianal diseases, respectively. Patients in group 1 had lower body weight percentile (1.0% vs 27.5%, $P = 0.002$) and hemoglobin concentrations (87.4 g/L vs 108.5 g/L, $P = 0.040$) when compared with group 2. Although patients in group 1 had a younger age of disease onset (2.7 mo), lower body height percentile (5.0%), and higher CRP (60.7 mg/L), there were no significant differences when compared with group 2 (Table 3).

DISCUSSION

The currently recognized pathogenetic mechanism of IBD is the involvement of many environmental triggers and genetic susceptibility that causes intestinal immune dysfunction. However, the influence of genes are likely more important than environmental factors for VEO-IBD patients with a disease onset prior to 6 years of age, especially for patients with an infantile onset prior to 1 year of age^[19]. GWAS

studies suggested that SNPs of *IL-10* and *STAT3* were associated with IBD^[20-23]. Previous studies confirmed that *IL-10* or *IL-10* receptor gene knockout mice had severe chronic inflammation of the intestinal tract^[24]. *IL-10* forms a complex with two molecules of *IL-10R1* and two molecules of *IL-10R2* to activate Janus kinase 1 (*Jak1*) and tyrosine kinase 2 (*Tyk2*). This activation results in the phosphorylation of signal transducer and activator of transcription 3 (*STAT3*), which regulates the transcription of specific genes. Studies suggested that *IL-10*-mediated signals effectively reduced the number of Th17 cells and relieved intestinal inflammation in CD^[25]. These data indicated that the anti-inflammatory *IL-10* signaling pathway plays a critical role in the regulation of intestinal immune homeostasis.

Since Glocker *et al.*^[8] first reported in 2009 that gene mutations in *IL-10RA* and *IL-10RB* caused infantile onset IBD^[8], studies have continuously reported mutations in *IL-10*, *IL-10RA*, and *IL-10RB* in patients with infantile onset IBD^[9-12,18,26]. In these limited data, the majority of patients were Arabian or Caucasian and the offspring of a consanguineous marriage. There are few reports on the Han Chinese population, which included only three pediatric patients to date^[13,14].

In this study, we used high-throughput next generation sequencing technology to sequence 10 IBD-associated genes, *IL-10*, *IL-10RA*, *IL-10RB*, *NOD2*, *FUT2*, *IL-23R*, *GPR35*, *GPR65*, *TNFSF15*, and *ADAM30*, in 13 Han Chinese children diagnosed with VEO-IBD. A total of four mutations were discovered in *IL-10RA*, including two novel mutations. There was one mutation in *IL-10RB*. These pathogenic mutations were found in five patients, which accounted for 38.5% of all VEO-IBD cases. Among these patients, one had an *IL-10RA* homozygous mutation, two had *IL-10RA* compound heterozygous mutations, one had an *IL-10RA* heterozygous mutation, and one had an *IL-10RB* heterozygous mutation. All *IL-10RA* mutations were in exon 3, and c.C301T (p.R101W) showed the highest frequency. The c.C301T (p.R101W) and c.G350A (p.R117H) mutations in *IL-10RA* were previously reported in similar pediatric patients. These mutations may disrupt signal transduction after activation of the *IL-10* receptor; therefore, *STAT3* is not phosphorylated and intractable inflammatory reactions in the intestinal tract of pediatric patients develop^[5,12]. The two novel mutations in *IL-10RA* discovered in this study were c.A191G (p.Y64C) and c.T299G (p.V100G). Because of condition limitations, we did not perform functional studies on these mutations. However, the SIFT prediction results for these two mutations were deleterious (scores of 0 and 0.002, respectively), and the Polyphen 2 prediction results were probably damaging (both scores were 1.000). These predictions suggest that these two mutations are pathogenic. According to the recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology^[27], these two

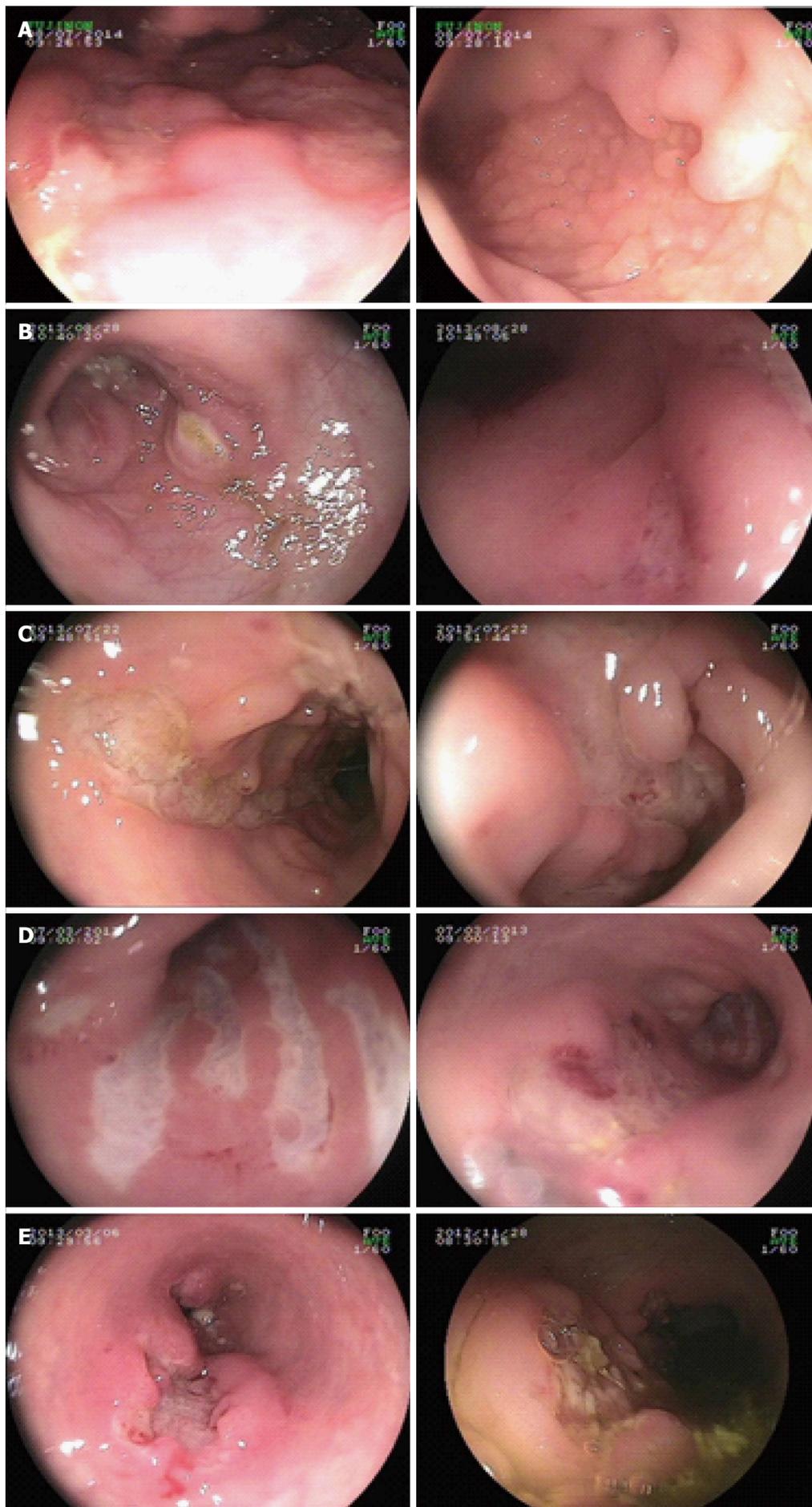


Figure 2 Colonoscopic presentation of patients with causative mutations showed pancolitis, cobblestone-like changes in mucosa, and deep and large ulcers. A to E presents patient 1 to patient 5, respectively.

Table 2 Clinical manifestations of very early-onset inflammatory bowel disease

	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5	Patient 6	Patient 7	Patient 8	Patient 9	Patient 10	Patient 11	Patient 12	Patient 13
Gender	F	M	M	M	M	M	M	M	M	M	M	F	F
Age of onset (mo)	8	1	0.3	0.3	4	0.2	9	2	0.5	3	0.7	10	36
Height percentile	19%	1%	3%	1%	1%	1%	52%	1%	15%	1%	19%	16%	20%
Weight percentile	1%	1%	1%	1%	20%	20%	55%	13%	8%	15%	16%	60%	33%
Diarrhea (times/ d)	>10	7-8	>10	10	5-10	5-6	7-8	2-4	7-8	No diarrhea	7-8	No diarrhea	2-3
Bloody stool	+	+	+	+	+	-	+	+	+	-	+	+	+
Infection	Sepsis	Pneumonia	No	Pneumonia, <i>Clostridium difficile</i> infection	Sepsis, oral candidiasis, fungemia, <i>Clostridium difficile</i> infection	Recurrent respiratory infection	No	No	No	Repeated fever of unknown origin	Oral candidiasis, gingivitis	No	No
Perianal lesion	Fistulae	No	No	Excrecence	Fistulae, abscess, excrecence	Fistulae, ulcer	No	No	No	No	Fistulae, abscess, excrecence	No	No
Clinical diagnosis	CD	CD	CD	CD	CD	CD	CD	CD	UC	CD	CD	UC	UC
Medication	GC, 6-MP	IFX, THD	GC, THD	GC, IFX ¹ , THD	GC, IFX, THD	GC, IFX ¹	GC, IFX, MIES	GC, IFX ¹ , THD, 6-MP	GC, MIES	GC, 6-MP, THD	GC, IFX, THD, 6-MP	GC, MIES	GC, MIES
Clinical status	NR	PR	Died at 2 yr because of severe sepsis	PR	Died at 3 yr because of intestinal failure	NR	CR	PR	CR	CR	PR	CR	CR

¹Allergic to IFX. CD: Crohn's disease; UC: Ulcerative colitis; GC: Glucocorticoid; 6-MP: 6-mercaptopurine; IFX: Infliximab; THD: Thalidomide; MIES: Mesalazine; NR: Non-remission; PR: Partial remission; CR: Complete remission.

mutations were defined as pathological supporting. Therefore, we speculate that these two novel mutations individually formed heterozygotes with the c.C301T (p.R101W) mutation to cause the disease symptoms observed in patients 2 and 3.

Previous analyses showed that the colitis caused by gene mutations in *IL-10* and its receptor exhibited an autosomal recessive inheritance pattern. In the current study, patients 4 and 5 were carriers of heterozygous mutations in *IL-10RA* and *IL-10RB*, respectively. The c.G350A (p.R117H) mutation in *IL-10RA* carried by patient 4 was a pathogenic mutation^[5,12,17]. The c.G421A (p.E141K) mutation in *IL-10RB* carried by patient 5 may affect protein function as predicted by SIFT (score = 0.026) and Polyphen 2 (score = 0.946). However, the clinical presentation of these two patients was similar to the symptoms of patients with other *IL-10* receptor mutations: disease onset within 1 year of age, the presence of perianal diseases and recurrent infection, and resistance to conventional medication treatment. Based on currently available knowledge, there are at least 50 single-gene genetic conditions that induce IBD-like diseases, and the majority of conditions are related to immunodeficiency^[4,6]. Therefore, the two patients that did not conform to a Mendelian genetic pattern might also carry abnormal sites on other genes that cause the disease symptoms. In addition to carrying a pathogenic mutation in *IL-10RA*, patient 4 also had a non-synonymous SNP (nsSNP): rs5743277 in *NOD2*. SIFT prediction results suggest that the nsSNP is deleterious (score = 0), and the Polyphen 2 prediction results suggest the nsSNP is probably damaging (score = 0.999). This polymorphism was already present in the HGMD database and has been considered to cause susceptibility to CD^[28]. Patient 5 had a similar condition. In addition to carrying an *IL-10RB* mutation, patient 5 also had multiple polymorphisms: rs22280554 (homozygous) and rs22280555 (homozygous) in *IL-10RA*,

Table 3 Comparison of features between patients with mutations and polymorphisms

	Group 1	Group 2
Size of sample	5	8
Age of onset (mo)	2.7	7.7
Height percentile	5.0%	15.6%
Weight percentile ^a	1.0%	27.5%
WBC ($\times 10^9$)	15.2	16.3
Hemoglobin (g/L) ^a	87.4	108.5
Platelets ($\times 10^9$)	538.4	424.0
C reactive protein (mg/L)	60.7	35.9
ESR (mm/H)	32.2	16.6
TNF α (pg/mL)	44.5	51.6
Diagnosis of CD	100.0%	62.5%
Recurrent infection	80.0%	25.0%
Perianal disease	60.0%	25.0%

^a $P < 0.05$. All measurement data are expressed as mean. Group 1: Mutations in *IL-10RA* or *IL-10RB*; Group 2: Polymorphisms. Height and weight percentile was calculated according to WHO charts. WBC: White blood cell; ESR: Erythrocyte sedimentation rate; TNF α : Tumor necrosis factor alpha; CD: Crohn's disease.

rs2834167 (homozygous) in *IL-10RB*, and rs1047781 (heterozygous) in *FUT2*. There are previous reports on the pathogenicity of these SNPs. For example, Galatola *et al.*^[29] reported that the heterozygous rs2834167 in *IL-10RB* and the heterozygous mutation in the promoter region of *IL-10RA* caused the development of UC in an 18-month-old patient. Although rs22280554 did not cause a change in the amino acid sequence of IL-10R1, a study by Moran *et al.*^[30] showed that rs22280554 and rs2228055 in *IL-10RA* may increase the risk for VEO-IBD, especially VEO-UC. Furthermore, in the Han Chinese population, the rs1047781 polymorphism in *FUT2* may increase the risk for CD development^[31]. The above SNPs were also detected in four patients in this study. Therefore, their disease development may be due to "trans-heterozygous": the collective effects of a variety of detected mutations. Another possible cause is that the pathogenic genes were not detected in this study.

When genotypes and phenotypes were combined for analyses, the results showed that the disease phenotype in patients with mutations were more severe. The age of disease onset was earlier, the patients were more likely to have combined recurrent infections and perianal diseases, their body weight and height were low, anemia was more severe, inflammatory indicators were high, and the prognosis was much poorer. These results are in accordance with previous studies^[5,8-14,18,32]. However, the sample size of this study was small, and significant differences were only found in body weight and hemoglobin parameters. Because of the influence of cultural ideas, family members find difficulty in accepting an ileostomy as a disease treatment. Past literature reported that pediatric patients with *IL-10RA* and *IL-10RB* mutations could be cured through hematopoietic stem cell transplantation^[4,6,8,17]; therefore, some

patients are waiting for a donor match.

In this study, we found that mutations in *IL-10RA* and *IL-10RB* were more common in Han Chinese VEO-IBD patients and accounted for 38.5% of all VEO-IBD cases. The high percentage is probably due to the small number of patients in the cohort as most of our patients who were referred by other clinical IBD centers were very ill. There was a selection bias. Because VEO-IBD is relatively rare, multi-center studies on the relationship between genotypes and phenotypes in VEO-IBD patients in China are necessary. The implementation of hematopoietic stem cell transplantation therapy is the focus in research agenda.

COMMENTS

Background

Very early-onset inflammatory bowel disease (VEO-IBD) may have stronger genetic contribution. Recently, a few studies on genetic defects in the IL-10 signaling pathway have provided new insights into IBD, especially in VEO-IBD. Furthermore, a lot of genes associated with IBD were identified, such as *NOD2*, *FUT2*, *IL-23R*, *GPR35*, *GPR65*, *TNFSF15*, and *ADAM30*. Because of different genetic background, this study was set to disclose whether mutations in these genes contributed to VEO-IBD in Chinese children.

Research frontiers

In addition to the polygenic variants associated with IBD, there are rare monogenic disorders, including many immunodeficiencies that can present with IBD-like intestinal inflammation, especially in early life.

Innovations and breakthroughs

To our knowledge, this is the first cohort study to apply NGS in 13 Chinese pediatric patients with VEO-IBD to discover gene variations in these children. The result revealed that *IL-10RA* and *IL-10RB* mutations were common in Chinese VEO-IBD, especially in infantile IBD. These monogenic IBD patients had more severe clinical features.

Applications

According to the results of this study and previous studies of VEO-IBD, the authors suggest that screening for gene mutations in IL-10 signaling pathway is necessary.

Peer-review

The clinical study is focused on gene mutation analysis in VEO-IBD by NGS. The authors conclude that mutations in the IL-10 pathway are common in VEO-IBD.

REFERENCES

- 1 **Levine A**, Koletzko S, Turner D, Escher JC, Cucchiara S, de Ridder L, Kolho KL, Veres G, Russell RK, Paerregaard A, Buderus S, Greer ML, Dias JA, Veereman-Wauters G, Lionetti P, Sladek M, Martin de Carpi J, Staiano A, Ruemmele FM, Wilson DC. ESPGHAN revised porto criteria for the diagnosis of inflammatory bowel disease in children and adolescents. *J Pediatr Gastroenterol Nutr* 2014; **58**: 795-806 [PMID: 24231644 DOI: 10.1097/mpg.0000000000000239]
- 2 **Wang XQ**, Zhang Y, Xu CD, Jiang LR, Huang Y, Du HM, Wang XJ. Inflammatory bowel disease in Chinese children: a multicenter analysis over a decade from Shanghai. *Inflamm Bowel Dis* 2013; **19**: 423-428 [PMID: 23340680 DOI: 10.1097/MIB.0b013e318286f9f2]
- 3 **Heyman MB**, Kirschner BS, Gold BD, Ferry G, Baldassano R, Cohen SA, Winter HS, Fain P, King C, Smith T, El-Serag HB.

- Children with early-onset inflammatory bowel disease (IBD): analysis of a pediatric IBD consortium registry. *J Pediatr* 2005; **146**: 35-40 [PMID: 15644819 DOI: 10.1016/j.jpeds.2004.08.043]
- 4 **Uhlig HH**, Schwerdt T, Koletzko S, Shah N, Kammermeier J, Elkadri A, Ouahed J, Wilson DC, Travis SP, Turner D, Klein C, Snapper SB, Muise AM. The diagnostic approach to monogenic very early onset inflammatory bowel disease. *Gastroenterology* 2014; **147**: 990-1007.e3 [PMID: 25058236 DOI: 10.1053/j.gastro.2014.07.023]
 - 5 **Shim JO**, Seo JK. Very early-onset inflammatory bowel disease (IBD) in infancy is a different disease entity from adult-onset IBD; one form of interleukin-10 receptor mutations. *J Hum Genet* 2014; **59**: 337-341 [PMID: 24785691 DOI: 10.1038/jhg.2014.32]
 - 6 **Uhlig HH**. Monogenic diseases associated with intestinal inflammation: implications for the understanding of inflammatory bowel disease. *Gut* 2013; **62**: 1795-1805 [PMID: 24203055 DOI: 10.1136/gutjnl-2012-303956]
 - 7 **Christodoulou K**, Wiskin AE, Gibson J, Tapper W, Willis C, Afzal NA, Upstill-Goddard R, Holloway JW, Simpson MA, Beattie RM, Collins A, Ennis S. Next generation exome sequencing of paediatric inflammatory bowel disease patients identifies rare and novel variants in candidate genes. *Gut* 2013; **62**: 977-984 [PMID: 22543157 DOI: 10.1136/gutjnl-2011-301833]
 - 8 **Glocker EO**, Kotlarz D, Boztug K, Gertz EM, Schäffer AA, Noyan F, Perro M, Diestelhorst J, Allroth A, Murugan D, Hätscher N, Pfeifer D, Sykora KW, Sauer M, Kreipe H, Lacher M, Nustede R, Woellner C, Baumann U, Salzer U, Koletzko S, Shah N, Segal AW, Sauerbrey A, Buderus S, Snapper SB, Grimbacher B, Klein C. Inflammatory bowel disease and mutations affecting the interleukin-10 receptor. *N Engl J Med* 2009; **361**: 2033-2045 [PMID: 19890111 DOI: 10.1056/NEJMoa0907206]
 - 9 **Beser OF**, Conde CD, Serwas NK, Cokugras FC, Kutlu T, Boztug K, Erkan T. Clinical features of interleukin 10 receptor gene mutations in children with very early-onset inflammatory bowel disease. *J Pediatr Gastroenterol Nutr* 2015; **60**: 332-338 [PMID: 25373860 DOI: 10.1097/mpg.0000000000000621]
 - 10 **Lee CH**, Hsu P, Nanan B, Nanan R, Wong M, Gaskin KJ, Leong RW, Murchie R, Muise AM, Stormon MO. Novel de novo mutations of the interleukin-10 receptor gene lead to infantile onset inflammatory bowel disease. *J Crohns Colitis* 2014; **8**: 1551-1556 [PMID: 24813381 DOI: 10.1016/j.crohns.2014.04.004]
 - 11 **Pigneur B**, Escher J, Elawad M, Lima R, Buderus S, Kierkus J, Guarais G, Canioni D, Lambot K, Talbotec C, Shah N, Begue B, Rieux-Laucat F, Goulet O, Cerf-Bensussan N, Neven B, Ruemmele FM. Phenotypic characterization of very early-onset IBD due to mutations in the IL10, IL10 receptor alpha or beta gene: a survey of the Genius Working Group. *Inflamm Bowel Dis* 2013; **19**: 2820-2828 [PMID: 24216686 DOI: 10.1097/01.MIB.0000435439.22484.d3]
 - 12 **Shim JO**, Hwang S, Yang HR, Moon JS, Chang JY, Ko JS, Park SS, Kang GH, Kim WS, Seo JK. Interleukin-10 receptor mutations in children with neonatal-onset Crohn's disease and intractable ulcerating enterocolitis. *Eur J Gastroenterol Hepatol* 2013; **25**: 1235-1240 [PMID: 23839161 DOI: 10.1097/MEG.0b013e328361a4f9]
 - 13 **Lu D**, Xu Y, Chen Y, Zeng P, Chen H, Zeng H. [Interleukin-10 receptor mutations in children with neonatal onset inflammatory bowel disease: genetic diagnosis and pathogenesis]. *Zhonghua Erke Zazhi* 2015; **53**: 348-354 [PMID: 26080664 DOI: 10.3760/cma.j.issn.0578-1310.2015.05.007]
 - 14 **Mao H**, Yang W, Lee PP, Ho MH, Yang J, Zeng S, Chong CY, Lee TL, Tu W, Lau YL. Exome sequencing identifies novel compound heterozygous mutations of IL-10 receptor 1 in neonatal-onset Crohn's disease. *Genes Immun* 2012; **13**: 437-442 [PMID: 22476154 DOI: 10.1038/gene.2012.8]
 - 15 **Hyams JS**. Standardized recording of parameters related to the natural history of inflammatory bowel disease: from Montreal to Paris. *Dig Dis* 2014; **32**: 337-344 [PMID: 24969277 DOI: 10.1159/000358133]
 - 16 **Levine A**, Griffiths A, Markowitz J, Wilson DC, Turner D, Russell RK, Fell J, Ruemmele FM, Walters T, Sherlock M, Dubinsky M, Hyams JS. Pediatric modification of the Montreal classification for inflammatory bowel disease: the Paris classification. *Inflamm Bowel Dis* 2011; **17**: 1314-1321 [PMID: 21560194 DOI: 10.1002/ibd.21493]
 - 17 **Engelhardt KR**, Shah N, Faizura-Yeop I, Kocacik Uygun DF, Frede N, Muise AM, Shteyer E, Filiz S, Chee R, Elawad M, Hartmann B, Arkwright PD, Dvorak C, Klein C, Puck JM, Grimbacher B, Glocker EO. Clinical outcome in IL-10- and IL-10 receptor-deficient patients with or without hematopoietic stem cell transplantation. *J Allergy Clin Immunol* 2013; **131**: 825-830 [PMID: 23158016 DOI: 10.1016/j.jaci.2012.09.025]
 - 18 **Begue B**, Verdier J, Rieux-Laucat F, Goulet O, Morali A, Canioni D, Hugot JP, Daussy C, Verkarre V, Pigneur B, Fischer A, Klein C, Cerf-Bensussan N, Ruemmele FM. Defective IL10 signaling defining a subgroup of patients with inflammatory bowel disease. *Am J Gastroenterol* 2011; **106**: 1544-1555 [PMID: 21519361 DOI: 10.1038/ajg.2011.112]
 - 19 **Imielinski M**, Baldassano RN, Griffiths A, Russell RK, Annese V, Dubinsky M, Kugathasan S, Bradfield JP, Walters TD, Sleiman P, Kim CE, Muise A, Wang K, Glessner JT, Saeed S, Zhang H, Frackelton EC, Hou C, Flory JH, Otieno G, Chiavacci RM, Grundmeier R, Castro M, Latiano A, Dallapiccola B, Stempak J, Abrams DJ, Taylor K, McGovern D, Silber G, Wrobel I, Quiros A, Barrett JC, Hansoul S, Nicolae DL, Cho JH, Duerr RH, Rioux JD, Brant SR, Silverberg MS, Taylor KD, Barmada MM, Bitton C, Dassopoulos T, Datta LW, Green T, Griffiths AM, Kistner EO, Murtha MT, Regueiro MD, Rotter JI, Schumm LP, Steinhart AH, Targan SR, Xavier RJ, Libioulle C, Sandor C, Lathrop M, Belaiche J, Dewit O, Gut I, Heath S, Laukens D, Mni M, Rutgeerts P, Van Gossom A, Zelenika D, Franchimont D, Hugot JP, de Vos M, Vermeire S, Louis E, Cardon LR, Anderson CA, Drummond H, Nimmo E, Ahmad T, Prescott NJ, Onnie CM, Fisher SA, Marchini J, Ghori J, Bumpstead S, Gwillam R, Tremelling M, Delukas P, Mansfield J, Jewell D, Satsangi J, Mathew CG, Parkes M, Georges M, Daly MJ, Heyman MB, Ferry GD, Kirschner B, Lee J, Essers J, Grand R, Stephens M, Levine A, Piccoli D, Van Limbergen J, Cucchiara S, Monos DS, Guthery SL, Denson L, Wilson DC, Grant SF, Daly M, Silverberg MS, Satsangi J, Hakonarson H. Common variants at five new loci associated with early-onset inflammatory bowel disease. *Nat Genet* 2009; **41**: 1335-1340 [PMID: 19915574 DOI: 10.1038/ng.489]
 - 20 **Franke A**, McGovern DP, Barrett JC, Wang K, Radford-Smith GL, Ahmad T, Lees CW, Balschun T, Lee J, Roberts R, Anderson CA, Bis JC, Bumpstead S, Ellinghaus D, Festen EM, Georges M, Green T, Haritunians T, Jostins L, Latiano A, Mathew CG, Montgomery GW, Prescott NJ, Raychaudhuri S, Rotter JI, Schumm P, Sharma Y, Simms LA, Taylor KD, Whiteman D, Wijmenga C, Baldassano RN, Barclay M, Bayless TM, Brand S, Büning C, Cohen A, Colombel JF, Cottone M, Stronati L, Denson T, De Vos M, D'Inca R, Dubinsky M, Edwards C, Florin T, Franchimont D, Gearry R, Glas J, Van Gossom A, Guthery SL, Halfvarson J, Verspaget HW, Hugot JP, Karban A, Laukens D, Lawrance I, Lemann M, Levine A, Libioulle C, Louis E, Mowat C, Newman W, Panés J, Phillips A, Proctor DD, Regueiro M, Russell R, Rutgeerts P, Sanderson J, Sans M, Seibold F, Steinhart AH, Stokkers PC, Torkvist L, Kullak-Ublick G, Wilson D, Walters T, Targan SR, Brant SR, Rioux JD, D'Amato M, Weersma RK, Kugathasan S, Griffiths AM, Mansfield JC, Vermeire S, Duerr RH, Silverberg MS, Satsangi J, Schreiber S, Cho JH, Annese V, Hakonarson H, Daly MJ, Parkes M. Genome-wide meta-analysis increases to 71 the number of confirmed Crohn's disease susceptibility loci. *Nat Genet* 2010; **42**: 1118-1125 [PMID: 21102463 DOI: 10.1038/ng.717]
 - 21 **Franke A**, Balschun T, Karlsen TH, Hedderich J, May S, Lu T, Schuldt D, Nikolaus S, Rosenstiel P, Krawczak M, Schreiber S. Replication of signals from recent studies of Crohn's disease identifies previously unknown disease loci for ulcerative colitis. *Nat Genet* 2008; **40**: 713-715 [PMID: 18438405 DOI: 10.1038/ng.148]
 - 22 **Barrett JC**, Hansoul S, Nicolae DL, Cho JH, Duerr RH, Rioux JD, Brant SR, Silverberg MS, Taylor KD, Barmada MM, Bitton

- A, Dassopoulos T, Datta LW, Green T, Griffiths AM, Kistner EO, Murtha MT, Regueiro MD, Rotter JI, Schumm LP, Steinhart AH, Targan SR, Xavier RJ, Libioulle C, Sandor C, Lathrop M, Belaiche J, Dewit O, Gut I, Heath S, Laukens D, Mni M, Rutgeerts P, Van Gossom A, Zelenika D, Franchimont D, Hugot JP, de Vos M, Vermeire S, Louis E, Cardon LR, Anderson CA, Drummond H, Nimmo E, Ahmad T, Prescott NJ, Onnie CM, Fisher SA, Marchini J, Ghori J, Bumpstead S, Gwilliam R, Tremelling M, Deloukas P, Mansfield J, Jewell D, Satsangi J, Mathew CG, Parkes M, Georges M, Daly MJ. Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. *Nat Genet* 2008; **40**: 955-962 [PMID: 18587394 DOI: 10.1038/ng.175]
- 23 **Franke A**, Balschun T, Karlsen TH, Sventoraityte J, Nikolaus S, Mayr G, Domingues FS, Albrecht M, Nothnagel M, Ellinghaus D, Sina C, Onnie CM, Weersma RK, Stokkers PC, Wijmenga C, Gazouli M, Strachan D, McArdle WL, Vermeire S, Rutgeerts P, Rosenstiel P, Krawczak M, Vatn MH, Mathew CG, Schreiber S. Sequence variants in IL10, ARPC2 and multiple other loci contribute to ulcerative colitis susceptibility. *Nat Genet* 2008; **40**: 1319-1323 [PMID: 18836448 DOI: 10.1038/ng.221]
- 24 **Kühn R**, Löhler J, Rennick D, Rajewsky K, Müller W. Interleukin-10-deficient mice develop chronic enterocolitis. *Cell* 1993; **75**: 263-274 [PMID: 8402911]
- 25 **Paul G**, Khare V, Gasche C. Inflamed gut mucosa: downstream of interleukin-10. *Eur J Clin Invest* 2012; **42**: 95-109 [PMID: 21631466 DOI: 10.1111/j.1365-2362.2011.02552.x]
- 26 **Glocker EO**, Frede N, Perro M, Sebire N, Elawad M, Shah N, Grimbacher B. Infant colitis--it's in the genes. *Lancet* 2010; **376**: 1272 [PMID: 20934598 DOI: 10.1016/S0140-6736]
- 27 **Richards S**, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, Grody WW, Hegde M, Lyon E, Spector E, Voelkerding K, Rehm HL. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med* 2015; **17**: 405-424 [PMID: 25741868 DOI: 10.1038/gim.2015.30]
- 28 **Lesage S**, Zouali H, Cézard JP, Colombel JF, Belaiche J, Almer S, Tysk C, O'Morain C, Gassull M, Binder V, Finkel Y, Modigliani R, Gower-Rousseau C, Macry J, Merlin F, Chamailard M, Jannot AS, Thomas G, Hugot JP. CARD15/NOD2 mutational analysis and genotype-phenotype correlation in 612 patients with inflammatory bowel disease. *Am J Hum Genet* 2002; **70**: 845-857 [PMID: 11875755 DOI: 10.1086/339432]
- 29 **Galatola M**, Miele E, Strisciuglio C, Paparo L, Rega D, Delrio P, Duraturo F, Martinelli M, Rossi GB, Staiano A, Izzo P, De Rosa M. Synergistic effect of interleukin-10-receptor variants in a case of early-onset ulcerative colitis. *World J Gastroenterol* 2013; **19**: 8659-8670 [PMID: 24379584 DOI: 10.3748/wjg.v19.i46.8659]
- 30 **Moran CJ**, Walters TD, Guo CH, Kugathasan S, Klein C, Turner D, Wolters VM, Bandsma RH, Mouzaki M, Zachos M, Langer JC, Cutz E, Benseler SM, Roifman CM, Silverberg MS, Griffiths AM, Snapper SB, Muise AM. IL-10R polymorphisms are associated with very-early-onset ulcerative colitis. *Inflamm Bowel Dis* 2013; **19**: 115-123 [PMID: 22550014 DOI: 10.1002/ibd.22974]
- 31 **Hu DY**, Shao XX, Xu CL, Xia SL, Yu LQ, Jiang LJ, Jin J, Lin XQ, Jiang Y. Associations of FUT2 and FUT3 gene polymorphisms with Crohn's disease in Chinese patients. *J Gastroenterol Hepatol* 2014; **29**: 1778-1785 [PMID: 24720527 DOI: 10.1111/jgh.12599]
- 32 **Shah N**, Kammermeier J, Elawad M, Glocker EO. Interleukin-10 and interleukin-10-receptor defects in inflammatory bowel disease. *Curr Allergy Asthma Rep* 2012; **12**: 373-379 [PMID: 22890722 DOI: 10.1007/s11882-012-0286-z]

P- Reviewer: Gassler N **S- Editor:** Ma YJ **L- Editor:** Wang TQ
E- Editor: Ma S



Retrospective Cohort Study

miR-422a is an independent prognostic factor and functions as a potential tumor suppressor in colorectal cancer

Gui-Xi Zheng, Ai-Lin Qu, Yong-Mei Yang, Xin Zhang, Shou-Cai Zhang, Chuan-Xin Wang

Gui-Xi Zheng, Ai-Lin Qu, Yong-Mei Yang, Xin Zhang, Shou-Cai Zhang, Chuan-Xin Wang, Department of Clinical Laboratory, Qilu Hospital of Shandong University, Jinan 250012, Shandong Province, China

Author contributions: Zheng GX and Wang CX conceived the study, participated in its design and coordination and helped draft the manuscript; Zheng GX and Qu AL performed the experiments and analyses; Yang YM, Zhang X and Zhang SC were responsible for collection of samples and acquiring of clinical data; all authors read and approved the final manuscript.

Supported by the National Natural Science Foundation of China, No. 81472025; Outstanding Young Scientist Research Award Fund of Shandong Province, No. BS2014YY023; Projects of Medical and Health Technology Development Program of Shandong Province, No. 2014WS0124; Science Foundation of Qilu Hospital of Shandong University, No. 2015QLQN37; Fundamental Research of Shandong University and the National Key Clinical Medical Specialties Foundation.

Institutional review board statement: The study was reviewed and approved by the Institution Review Board of Qilu Hospital of Shandong University.

Informed consent statement: All study participants provided informed written consent prior to study enrollment.

Conflict-of-interest statement: No potential conflicts of interest relevant to this article exist.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Correspondence to: Chuan-Xin Wang, MD, PhD, Department of Clinical Laboratory, Qilu Hospital of Shandong University, 107 Wenhua Xi Road, Jinan 250012, Shandong Province, China. cxwang@sdu.edu.cn
Telephone: +86-531-86927544
Fax: +86-531-86927544

Received: March 11, 2016
Peer-review started: March 11, 2016
First decision: March 31, 2016
Revised: April 25, 2016
Accepted: May 21, 2016
Article in press: May 23, 2016
Published online: June 28, 2016

Abstract

AIM: To determine the expression of miR-422a in colorectal cancer (CRC) tissues and to further explore the prognostic value and function of miR-422a in CRC carcinogenesis.

METHODS: miR-422a expression was analyzed in 102 CRC tissues and paired normal mucosa adjacent to carcinoma by quantitative real-time PCR. The relationship of miR-422a expression with clinicopathological parameters was also analyzed. Kaplan-Meier analysis and Cox multivariate analysis were performed to estimate the potential role of miR-422a. Cell proliferation, migration, and invasion were used for *in vitro* functional analysis of miR-422a.

RESULTS: The levels of miR-422a were dramatically reduced in CRC tissues compared with normal mucosa ($P < 0.05$), and significantly correlated with local invasion ($P = 0.004$) and lymph node metastasis ($P < 0.001$). Kaplan-Meier survival and Cox regression multivariate analyses revealed that miR-422a expression (HR = 0.568, $P = 0.015$) and clinical TNM stage (HR = 2.942, $P = 0.003$) were independent prognostic factors

for overall survival in CRC patients. Furthermore, *in vitro* experiments showed that overexpression of miR-422a inhibited the proliferation, migration, and invasion of SW480 and HT-29 cells.

CONCLUSION: Down-regulation of miR-422a may serve as an independent prognosis factor in CRC. MiR-422a functions as a tumor suppressor and regulates progression of CRC.

Key words: Colorectal cancer; MicroRNA; miR-422a; Prognosis; *In vitro* function

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: In the present study, we found that miR-422a was dramatically reduced in colorectal cancer (CRC) tissues, and significantly correlated with local invasion and lymph node metastasis. miR-422a expression and clinical TNM stage were independent prognostic factors for overall survival in CRC patients. Furthermore, *in vitro* experiments showed that overexpression of miR-422a inhibited the proliferation, migration, and invasion of SW480 and HT-29 cells. These results indicated that down-regulation of miR-422a might serve as an independent prognosis factor in CRC, and miR-422a functions as a tumor suppressor and regulates progression of CRC.

Zheng GX, Qu AL, Yang YM, Zhang X, Zhang SC, Wang CX. miR-422a is an independent prognostic factor and functions as a potential tumor suppressor in colorectal cancer. *World J Gastroenterol* 2016; 22(24): 5589-5597 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5589.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5589>

INTRODUCTION

Colorectal cancer (CRC) is one of the most common malignant tumors with increasing incidence and mortality over the past several decades^[1]. Despite the significant progress made in diagnostic methods and improved treatment strategies, the prognosis of CRC patients remains poor, especially in those with advanced CRC^[2,3]. CRC carcinogenesis is associated with multiple alterations in oncogenes and tumor suppressor genes. A growing number of studies have revealed that microRNAs (miRNAs) might regulate up to 30% of human genes and play a pivotal role in various cellular processes including proliferation, differentiation, apoptosis, migration, and invasion^[4-8]. While the functional mechanisms of miRNAs remain largely unknown in the pathogenesis of CRC, dysregulated expression of miRNAs can serve as potential biomarkers for the diagnosis and prognosis of cancer^[9-11].

Several studies have determined the importance of miR-422a in human diseases such as cancer, multiple sclerosis, and postmenopausal osteoporosis. Gougelet *et al.*^[12] reported that miR-422a could inhibit signaling pathways regulating tumor cell proliferation in osteosarcoma. A study by Mao *et al.*^[13] demonstrated that miR-422a targeted key mismatch repair protein (MutL α) by suppressing MLH1 expression, resulting in genome instability and tumorigenesis. Faltejškova *et al.*^[14] reported that dysregulation of miR-378, miR-375, miR-422a, miR-215 and miR-135b in CRC patients played an important role in CRC pathogenesis. However, further study is needed to confirm whether miR-422a is an independent predictive factor in patients with CRC. Moreover, the functional mechanism by which miR-422a regulates CRC progression and whether it can serve as a prognostic biomarker in CRC remain largely unknown.

Previously, we have identified a serum 4-miRNA panel that included miR-19a-3p, miR-92a-3p, miR-223-3p, and miR-422a. This miRNA panel served as biomarkers for early diagnosis of colorectal adenocarcinoma^[15,16]. In this study, we showed that the expression of miR-422a is dysregulated in CRC tissues. The correlation between miR-422a and certain clinical characteristics, as well as its potential as a prognostic marker for CRC, was also investigated. The effects of miR-422a on CRC cell proliferation, invasion, and migration were also assessed.

MATERIALS AND METHODS

Study population and sample collection

All written informed consent was obtained from every participant for use of tissue samples. This project was approved by the Clinical Research Ethics Committee of Qilu Hospital of Shandong University. All CRC patients were recruited from the Department of General Surgery, Qilu Hospital of Shandong University between November 2004 and December 2013. The diagnosis of CRC was confirmed by histopathology or histobiopsy. A total of 102 CRC tissues were collected and the adjacent normal mucosa tissues were used as controls since CRC is a malignant epithelial tumor and originates from glandular epithelium of the colorectal mucosa. All tissues were immediately frozen in liquid nitrogen and stored at -80 °C until RNA extraction. The median follow-up period for patients enrolled in this study was 63 mo (range, 14-78 mo).

Cell culture

SW480 and HT29 cells were purchased from the Type Culture Collection of the Chinese Academy of Sciences (Shanghai, China). The two cell lines were cultured in DMEM medium supplemented with 10% fetal bovine serum (Gibco, Carlsbad, CA, United States) at 37 °C in an incubator containing 5% CO₂.

Table 1 Relationship between miR-422a and clinical parameters in colorectal cancer tissues

Clinical parameter	Number of cases (<i>n</i> = 102)	miR-422a expression, median (min-max)	<i>P</i> value
Gender			0.704
Male	63	1.446 (0.074-130.774)	
Female	39	1.617 (0.167-33.543)	
Age (yr)			0.328
≤ median	49	1.637 (0.088-130.774)	
> median	53	1.715 (0.074-21.871)	
Tumor location			0.092
Colon	58	1.798 (0.088-130.774)	
Rectum	44	1.887 (0.074-33.543)	
Differentiation			0.150
Well	12	1.798 (0.074-130.744)	
Moderate	67	1.922 (0.166-21.871)	
Poor	23	2.088 (0.088-20.406)	
Tumor size			0.527
≤ 5 cm	64	2.127 (0.074-130.744)	
> 5 cm	38	2.353 (0.167-21.871)	
Local invasion			0.004
T1-T2	55	3.257 (0.206-130.744)	
T3-T4	47	1.617 (0.074-20.406)	
Lymph node metastasis			0.002
No	70	2.863 (0.359-130.744)	
Yes	32	1.445 (0.074-20.406)	
TNM stage			< 0.001
I	23	3.877 (0.265-15.999)	
II	42	3.253 (0.074-13.566)	
III	37	1.445 (0.329-130.744)	

Values are median (IQR: Inter-quartile range). TNM: Tumor node metastasis.

Cell transfection

CRC cells were seeded at 2×10^5 cells/well in 6-well plates until 30%-50% confluency and then were transfected with miR-422a mimics or negative control mimics (RiboBio, Guangzhou, China) using Lipofectamine 2000 (Invitrogen, Carlsbad, CA, United States) according to the manufacturer's introduction. The transfected cells were normalized and used in subsequent assays. The transfection efficiency was determined by RT-qPCR to verify the success of transfection.

RNA preparation, cDNA synthesis and RT-qPCR

Total RNA from cell lines, CRC tissues and normal mucosa was extracted using Trizol reagent (Invitrogen, Carlsbad, CA, United States). The cDNA was reverse-transcribed at 65 °C for 5 min in a 12 μL reaction system including 1 μg RNA, 1 μL reverse transcription primer (RiboBio, Guangzhou, China), 1 μL U6 reverse transcription primer and 1 μL dNTP. Then, 4 μL buffer, 2 μL DTT and 1 μL RNase inhibitor were added and incubated at 37 °C for 2 min followed by adding 1 μL MMLV and incubating at 37 °C for 50 min and 70 °C for 5 min. The PCR reaction was performed as follows: 95 °C for 1 min, and 45 cycles of 95 °C for

15 s, 60 °C for 30 s, and 72 °C for 45 s. U6 was used as a reference gene and each test was performed in triplicate in this study. RT-qPCR reactions were carried out on ABI Prism 7500 System (Applied Biosystems, Foster City, CA, United States).

Cell proliferation assay

miR-422a mimics and control mimics were transfected into SW480 and HT29 cells, then cultured for another 24, 48, 72, or 96 h. Cell proliferation rates was measured with CCK-8 reagent (Beyotime, Hangzhou, China) following the manufacturer's protocol. A microplate photometer (Multiskan FC, Thermo scientific, Shanghai, China) was used to determine the optical density at 450 nm.

Transwell migration and invasion assays

Transwell migration assay was performed using transwell chambers (Corning Costar, United States). After transfection for 24 h, cells were transferred into the upper chamber. RPMI 1640 medium containing 10% FBS functioning as a chemoattractant was added to matched lower chamber. SW480 and HT29 cells unable to migrate were removed from the upper surface of the transwell membrane after incubation for 48 h. Cells that were able to invade on the lower membrane surface were fixed in methanol, stained with 0.1% crystal violet, and counted using an inverted microscope (Olympus, Tokyo, Japan). For invasion assay, the inserts were pre-coated with matrix gel (BD Biosciences, Franklin Lakes, NJ, United States).

Statistical analysis

Data were analyzed using SPSS 17.0 software (IBM Corporation, Armonk, NY, United States). The expression of miR-422a between groups was compared using Mann-Whitney *U* test. Kaplan-Meier method was used for overall survival analysis. The Cox regression model was used for univariate and multivariate analyses to estimate the prognostic factors.

RESULTS

Expression of miR-422a in CRC tissues and correlative analysis with clinicopathological parameters

We detected the expression of miR-422a in 102 pairs of CRC tissues and normal mucosa adjacent to carcinoma using RT-qPCR. The results indicated that miR-422a levels were significantly lower in CRC tissues compared with normal mucosa ($P < 0.05$) (Figure 1A). Moreover, 65.7% (67 of 102) of CRC tissues had at least 2-fold down-regulated expression of miR-422a compared with normal mucosa (Figure 1B). The levels of miR-422a in stages I and II samples were significantly higher than those in stage III samples ($P < 0.05$). There was no significant difference in miR-422a expression between stages I and II samples (P

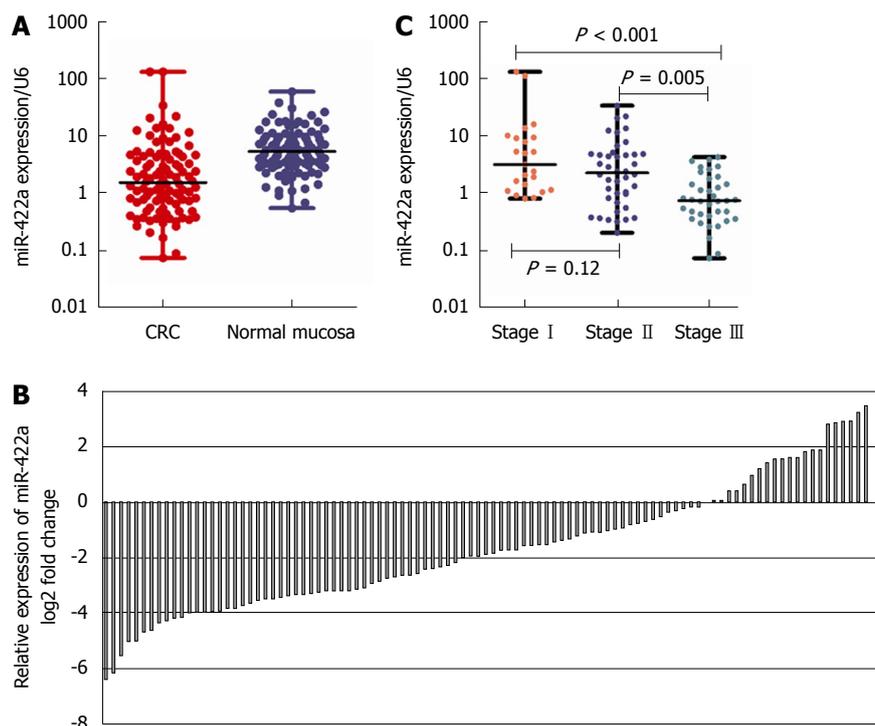


Figure 1 Expression of miR-422a in colorectal cancer and its association with TNM stage. A: Levels of miR-422a were determined in 102 colorectal cancer (CRC) tissues and their corresponding normal mucosa (NC); B: The fold changes of relative miR-422a level (CRC/NC) in each matched samples, with “< -1” defined as under-expression, “-1-1” as unchanged, and “> 1” as overexpression; C: Levels of miR-422a in stages I / II / III CRC.

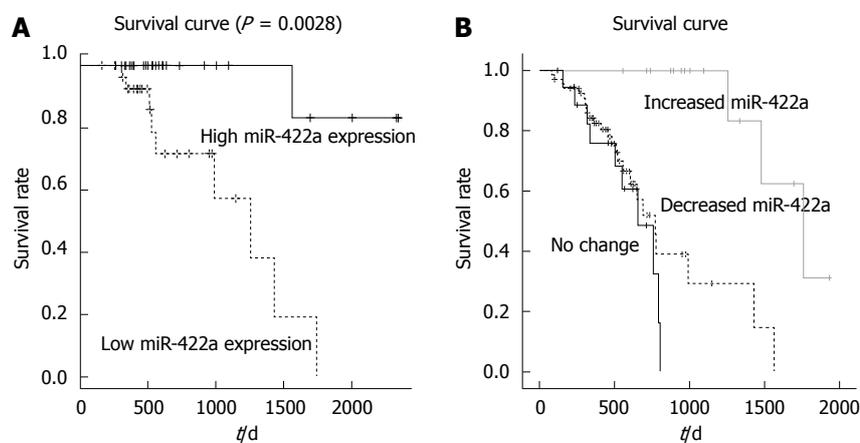


Figure 2 Kaplan-Meier curves for overall survival in 102 colorectal cancer patients based on miR-422a expression and the change of miR-422a expression compared to adjacent normal mucosa, respectively. Results showed that patients with low miR-422a expression had a significantly poorer prognosis than those with high miR-422a expression (A) ($P = 0.0028$), and patients with decreased miR-422a expression compared to adjacent normal mucosa had a significantly poorer prognosis than those with increased miR-422a expression (B) ($P < 0.001$).

> 0.05) (Figure 1C).

The association between miR-422a level and clinicopathological parameters of CRC patients is showed in Table 1. Our data showed that the level of miR-422a significantly correlated with local invasion ($P = 0.004$), lymph node metastasis ($P = 0.002$), and TNM stage ($P < 0.001$). However, there were no significant correlations between miR-422a level and gender, age, tumor location, differentiation or tumor size ($P > 0.05$).

miR-422a is an independent prognostic factor for overall survival in CRC patients

Our results demonstrated that 42 of 102 CRC patients died during the follow-up period. The 5-year overall survival rate was 58.8%. Patients were divided into high miR-422a expression and low miR-422a expression groups based on the median level. The prognosis was analyzed by Kaplan-Meier survival analysis, which revealed that patients with low miR-422a expression had a significantly poorer prognosis

Table 2 Univariate and multivariate analyses of overall survival in colorectal cancer patients

Variable	Category	Univariate analysis			Multivariate analysis		
		HR	95%CI	P value	HR	95%CI	P value
Gender	Male <i>vs</i> Female	0.943	0.493-1.872	0.810			
Age	≤ Median <i>vs</i> > Median	1.436	0.528-3.157	0.148			
Tumor location	Colon <i>vs</i> Rectum	0.862	0.484-1.706	0.603			
Tumor size	≤ 5 cm <i>vs</i> > 5 cm	1.093	0.568-2.194	0.873			
Differentiation	Well and moderate <i>vs</i> Poor	1.018	0.462-2.180	0.160	0.986	0.486-2.006	0.074
Local invasion	T1-T2 <i>vs</i> T3-T4	1.682	1.020-2.628	0.01 ¹	1.460	0.746-2.632	0.482
Lymph node metastasis	Yes <i>vs</i> No	1.182	0.634-2.530	< 0.001 ¹	1.262	0.584-2.680	0.306
TNM stage	I and II <i>vs</i> III	2.738	1.509-4.265	< 0.001 ¹	2.942	1.426-4.378	0.003 ¹
MiR-422a level	Low <i>vs</i> High	0.306	0.108-0.763	0.001 ¹	0.568	0.245-1.082	0.015 ¹

¹There was a significant difference between the two groups ($P < 0.05$).

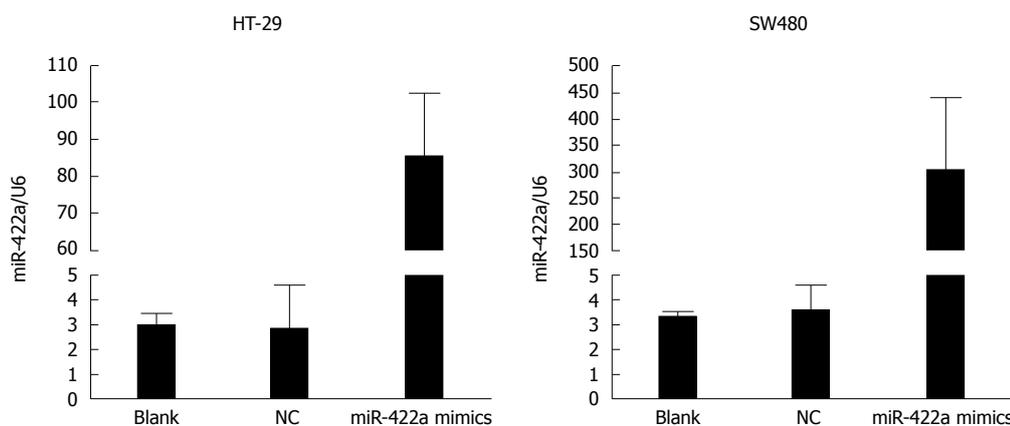


Figure 3 Transfection efficiency assays. The expression of miR-422a in HT-29 and SW480 cells was significantly increased after transfection with the mimics. All experiments were performed in triplicate.

than those with high miR-422a ($P = 0.0028$; Figure 2A). Furthermore, patients were divided into three groups (decreased, no change, and increased) based on the change of miR-422a expression compared to adjacent normal mucosa. Kaplan-Meier survival analysis showed that patients with decreased miR-422a expression had a significantly poorer prognosis than those with increased miR-422a expression ($P < 0.001$; Figure 2B). There was no significant difference between the decreased miR-422a group and no change group. In addition, Cox regression multivariate analysis was performed to determine whether miR-422a was an independent factor of overall survival in CRC patients. The analysis revealed that miR-422a expression (HR = 0.568, 95%CI: 0.245-1.082; $P = 0.015$) and clinical TNM stage (HR = 2.942, 95%CI: 1.426-4.378; $P = 0.003$) were independent prognostic factors for overall survival in CRC patients (Table 2).

Overexpression of miR-422a inhibits CRC cell proliferation

In our preliminary experiments, we detected the expression levels of miR-422a endogenously in several CRC cell lines, including HT-29, SW480, SW620 and HCT-116. The results showed that the levels of miR-422a were quite low in these CRC cells, and there

were no significant differences in the expression level of miR-422a among these cell lines (data not shown). To measure the biological properties of miR-422a in CRC cells, miR-422a mimics were transiently transfected into HT-29 and SW480 cells, respectively. Subsequently, real-time RT-qPCR was performed, which showed high transfection efficiencies in both cell lines (Figure 3). The results of MTT assay showed that transfection of miR-422a mimics could significantly decrease cell number in SW480 and HT29 CRC cells ($P < 0.05$; Figure 4). It means that overexpression of miR-422a can inhibit the proliferation potential of SW480 and HT29 CRC cells.

Effect of miR-422a overexpression on cell migration and invasion

The transwell assays with and without Matrigel were performed to determine the effects of miR-422a on the migration and invasiveness of CRC cells. Overexpression of miR-422a significantly inhibited the migration of SW480 ($P = 0.017$; Figure 5A and B) and HT-29 cells ($P = 0.007$; Figure 5C and D). Furthermore, transwell assay with Matrigel showed that the invasiveness of these cells was significantly suppressed in cells transfected with miR-422a compared with miR control (Figure 6A-D).

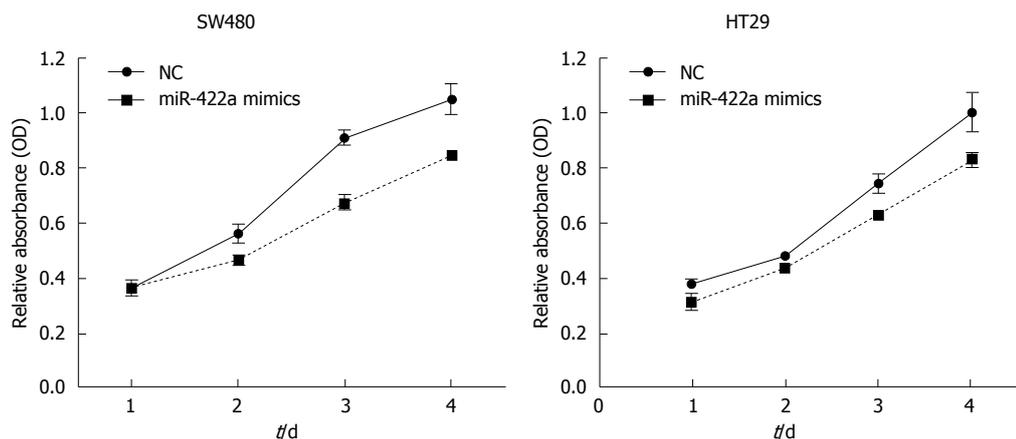


Figure 4 Overexpression of miR-422a inhibits colorectal cancer cell proliferation. Growth curves of miR-422a mimics and negative control mimics-transfected CRC cells were plotted after CCK-8 assay in SW480 and HT29 cells. All experiments were performed in triplicate. NC: Negative control.

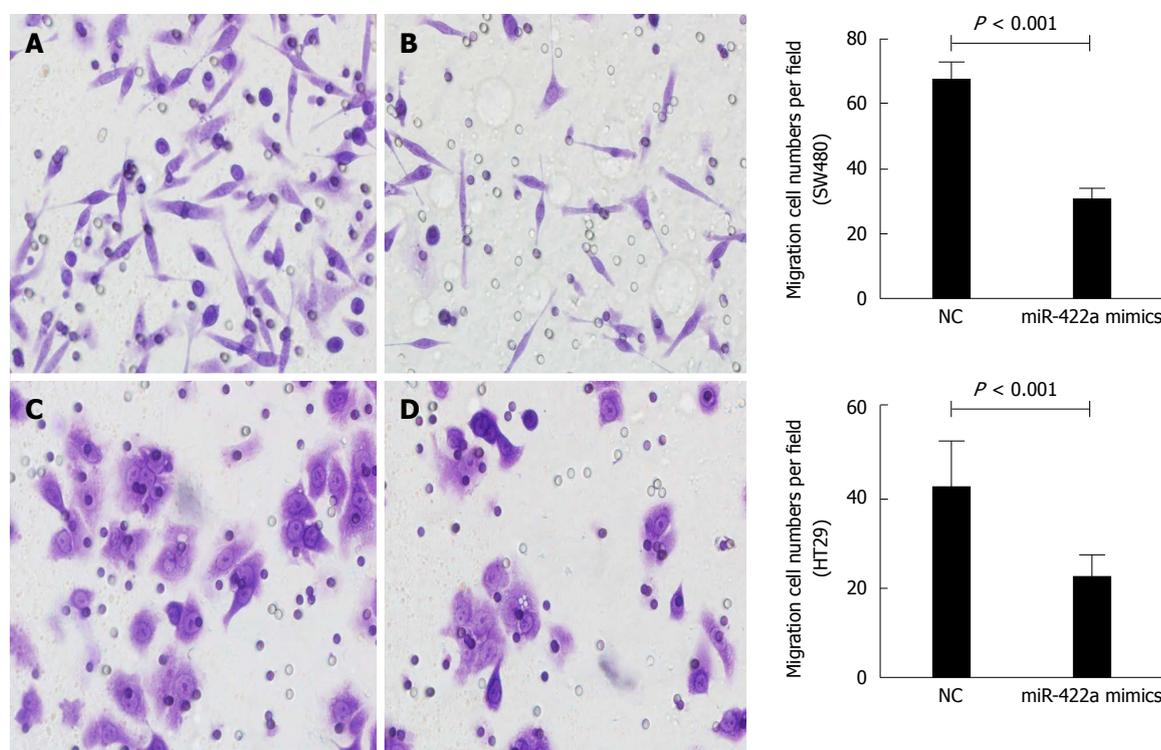


Figure 5 Effect of miR-422a overexpression on cell migration. Overexpression of miR-422a significantly inhibited the migration of SW480 (A and B) and HT29 (C and D) cells after transient transfection with miR-422a mimics. Stained cells were counted in five randomly selected fields under a light microscope. Representative photographs (left) and quantification (right) are shown. Magnification: $\times 400$. All assays were performed in triplicate to derive the confidence intervals.

DISCUSSION

Tumor progression in CRC is a multi-step process involving a large number of genetic and epigenetic alterations^[17-19]. Numerous studies have showed that miRNA target genes are involved in CRC tumorigenesis^[6-8,20]. Currently, there is an ongoing effort to elucidate new deregulated miRNAs and their roles in CRC progression.

miR-422a is encoded by gene MIR422A on 15q22.31 (64, 163, 129-64, 163, 218bp). Recently, several studies have studied the functional role of miR-422a in osteosarcoma, osteoporosis, HIV infection, and other

tumor types^[21-25]. Our previous study demonstrated that serum miR-422a was significantly downregulated in CRC patients and is a potential biomarker with a high diagnostic accuracy^[15]. However, the expression and role of miR-422a in CRC tissues, as well as its clinicopathological and prognostic value in CRC tumorigenesis remain undefined. In this study, our results demonstrated that miR-422a was down-regulated in CRC tissues compared with normal mucosa, which is consistent with a previous study^[14]. We also found that the expression of miR-422a was significantly associated with lymph node metastasis and clinical stage in CRC

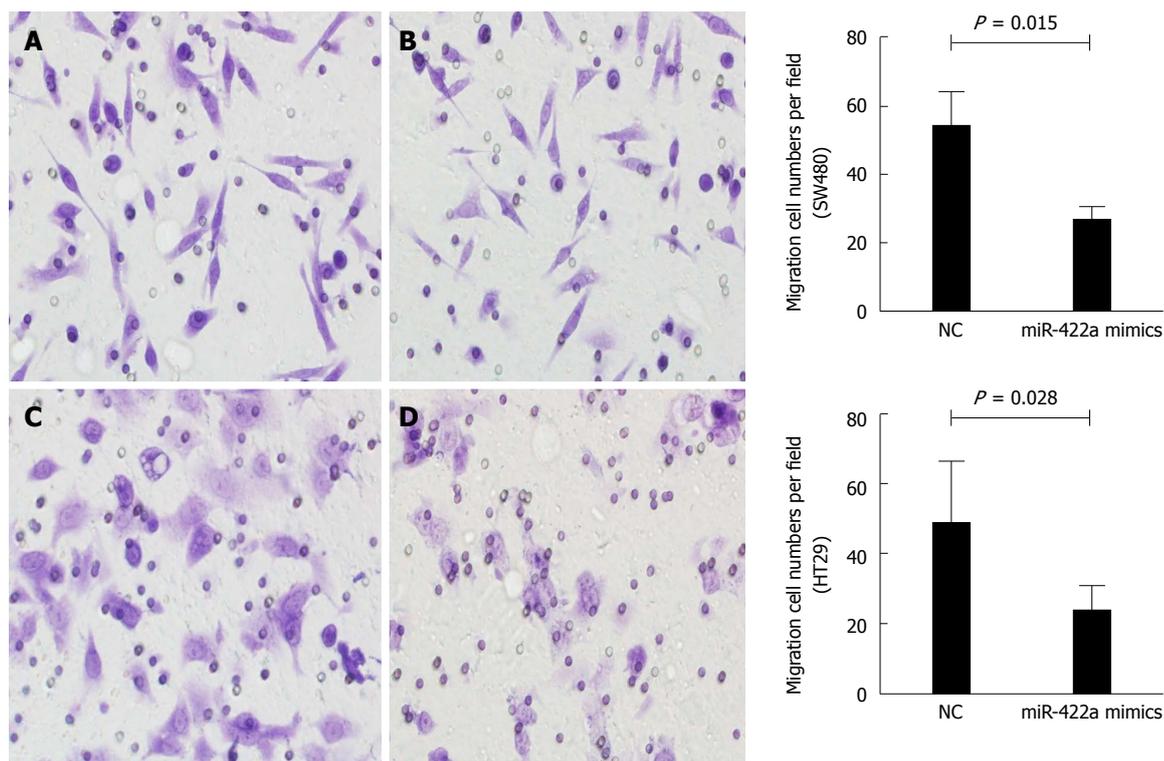


Figure 6 Effect of miR-422a overexpression on cell invasion. Overexpression of miR-422a significantly inhibited the invasiveness of in SW480 (A and B) and HT29 (C and D) cells after transient transfection with miR-422A mimics. Stained cells were counted in five randomly selected fields under a light microscope. Representative photographs (left) and quantification (right) are shown. Magnification: $\times 400$. All assays were performed in triplicate to derive the confidence intervals.

patients, suggesting that down-regulation of miR-422a might participate in CRC progression. To further explore the value of miR-422a as a prognostic factor, we investigated the correlation between miR-422a and overall survival in CRC patients. Our results showed that patients with low expression of miR-422a and decreased miR-422a expression compared with normal mucosa had poorer survival, suggesting that miR-422a could serve as an independent prognostic factor in CRC patients. Overall, these results suggest that miR-422a plays a protective role against CRC and could be used to evaluate prognosis in CRC patients.

Until now, few studies have described the function of miR-422a^[13,22,26]. A study has demonstrated that miR-422a and the mismatch repair protein MutL α (MLH1) are involved in a feedback loop, leading to genome instability and tumorigenesis^[13]. Another study showed that miR-422a significantly modulated the efficacy of IFN- α /RBV treatment *in vivo*. Exogenous IFN- α treatment led to decreased miR-422a and likely contributed to the IFN-mediated suppression of HIV-1, suggesting that restoring miR-422a treatment could be a potential therapeutic strategy for HIV-1 infection^[26]. Moreover, overexpression of human telomerase reverse transcriptase (hTERT) has been associated with the invasion and metastasis of CRC cells. miR-138-5p and miR-422a were found to be hTERT-targeting miRNAs, potentially inhibiting hTERT expression^[26]. Invasion and migration are required for

tumor cells to spread from the primary site to lymph or blood vessels. In the present study, we found that miR-422a expression was associated with lymph node metastasis and overexpression of miR-422a inhibited CRC cell proliferation, migration and invasion. These results suggest that multiple signaling pathways regulating different aspects of tumorigenesis may be regulated by miR-422a expression, suggesting that miR-422a may be a new therapeutic target to repress cancer progression.

COMMENTS

Background

A growing number of studies have shown that deregulated miRNAs play critical roles in tumorigenesis and progression of colorectal cancer (CRC). Nevertheless, an ongoing effort to elucidate new deregulated miRNAs and their roles in CRC is still urgently needed. Several studies have demonstrated the importance of miR-422a in different types of human diseases. In our previous study, miR-422a was found to be down-regulated in serum and could be used as a potential biomarker for CRC. Therefore, it is necessary to further determine the expression of miR-422a in CRC tissues and to explore its clinicopathological, prognostic value and role in CRC carcinogenesis.

Research frontiers

Several studies have determined the importance of miR-422a in human diseases such as cancer, multiple sclerosis, and postmenopausal osteoporosis. miR-422a could inhibit signaling pathways regulating tumor cell proliferation in osteosarcoma. It was also shown that miR-422a targeted key mismatch repair protein (MutL α) by suppressing MLH1 expression, resulting in genome instability and tumorigenesis. Previously, we identified a serum 4-miRNA panel that included miR-19a-3p, miR-92a-3p, miR-223-3p, and miR-

422a. This miRNA panel served as biomarkers for early diagnosis of colorectal adenocarcinoma.

Innovations and breakthroughs

The present study indicated that down-regulation of miR-422a might serve as an independent prognosis factor in CRC. miR-422a functions as a tumor suppressor and regulates progression of CRC.

Applications

By understanding the differential expression of miR-422a in CRC patients and its relationship with clinicopathological characteristics, prognosis and *in vitro* function, the present study may provide a new prognostic factor of CRC and further reveal the mechanism of miR-422a participating in CRC carcinogenesis.

Terminology

MicroRNAs are a class of short non-coding RNAs that regulate up to 30% of human genes and play a pivotal role in various cellular processes. A growing number of miRNAs have been implicated in the initiation and progression of tumors including miR-422a. miR-422a is encoded by gene *MIR422A* on 15q22.31 (64,163,129-64,163,218bp), which has been found to be important in human diseases such as cancer, multiple sclerosis, and postmenopausal osteoporosis.

Peer-review

The authors presented a study analyzing the prognostic value and function of miR-422a in CRC carcinogenesis. The results demonstrated that miR-422a was dramatically reduced in CRC tissues and significantly correlated with local invasion and lymph node metastasis. miR-422a expression was an independent prognostic factor for overall survival. *In vitro* experiments showed that overexpression of miR-422a inhibited the proliferation, migration, and invasion of SW480 and HT-29 cells. These data indicated that miR-422a might serve as an independent prognosis factor and regulate progression of CRC by functioning as a tumor suppressor.

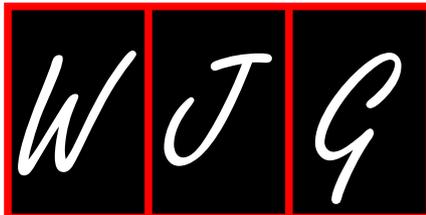
REFERENCES

- 1 Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin* 2015; **65**: 87-108 [PMID: 25651787 DOI: 10.3322/caac.21262]
- 2 Haggard FA, Boushey RP. Colorectal cancer epidemiology: incidence, mortality, survival, and risk factors. *Clin Colon Rectal Surg* 2009; **22**: 191-197 [PMID: 21037809 DOI: 10.1055/s-0029-1242458]
- 3 Lieberman DA. Clinical practice. Screening for colorectal cancer. *N Engl J Med* 2009; **361**: 1179-1187 [PMID: 19759380 DOI: 10.1056/NEJMc0902176]
- 4 Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell* 2009; **136**: 215-233 [PMID: 19167326 DOI: 10.1016/j.cell.2009.01.002]
- 5 Mirghasemi A, Taheriazam A, Karbasy SH, Torkaman A, Shakeri M, Yahaghi E, Mokarizadeh A. Down-regulation of miR-133a and miR-539 are associated with unfavorable prognosis in patients suffering from osteosarcoma. *Cancer Cell Int* 2015; **15**: 86 [PMID: 26388701 DOI: 10.1186/s12935-015-0237-6]
- 6 Xu W, Liu M, Peng X, Zhou P, Zhou J, Xu K, Xu H, Jiang S. miR-24-3p and miR-27a-3p promote cell proliferation in glioma cells via cooperative regulation of MXI1. *Int J Oncol* 2013; **42**: 757-766 [PMID: 23254855 DOI: 10.3892/ijo.2012.1742]
- 7 Lin C, Huang F, Li QZ, Zhang YJ. miR-101 suppresses tumor proliferation and migration, and induces apoptosis by targeting EZH2 in esophageal cancer cells. *Int J Clin Exp Pathol* 2014; **7**: 6543-6550 [PMID: 25400732]
- 8 Yunqiao L, Vanke H, Jun X, Tangmeng G. MicroRNA-206, down-regulated in hepatocellular carcinoma, suppresses cell proliferation and promotes apoptosis. *Hepatogastroenterology* 2014; **61**: 1302-1307 [PMID: 25513086]
- 9 Wang J, Zhang X, Wang L, Yang Y, Dong Z, Wang H, Du L, Wang C. MicroRNA-214 suppresses oncogenesis and exerts impact on prognosis by targeting PDRG1 in bladder cancer. *PLoS One* 2015; **10**: e0118086 [PMID: 25706919 DOI: 10.1371/journal.pone.0118086]
- 10 Kavitha N, Vijayarathna S, Jothy SL, Oon CE, Chen Y, Kanwar JR, Sasidharan S. MicroRNAs: biogenesis, roles for carcinogenesis and as potential biomarkers for cancer diagnosis and prognosis. *Asian Pac J Cancer Prev* 2014; **15**: 7489-7497 [PMID: 25292018]
- 11 Lyra-González I, Flores-Fong LE, González-García I, Medina-Preciado D, Armendáriz-Borunda J. MicroRNAs dysregulation in hepatocellular carcinoma: Insights in genomic medicine. *World J Hepatol* 2015; **7**: 1530-1540 [PMID: 26085912 DOI: 10.4254/wjh.v7.i11.1530]
- 12 Gougelet A, Pissaloux D, Besse A, Perez J, Duc A, Dutour A, Blay JY, Alberti L. Micro-RNA profiles in osteosarcoma as a predictive tool for ifosfamide response. *Int J Cancer* 2011; **129**: 680-690 [PMID: 20949564 DOI: 10.1002/ijc.25715]
- 13 Mao G, Lee S, Ortega J, Gu L, Li GM. Modulation of microRNA processing by mismatch repair protein MutLa. *Cell Res* 2012; **22**: 973-985 [PMID: 22290424 DOI: 10.1038/cr.2012.18]
- 14 Faltejiskova P, Svoboda M, Srutova K, Mlcochova J, Besse A, Nekvindova J, Radova L, Fabian P, Slaba K, Kiss I, Vyzula R, Slaby O. Identification and functional screening of microRNAs highly deregulated in colorectal cancer. *J Cell Mol Med* 2012; **16**: 2655-2666 [PMID: 22469014 DOI: 10.1111/j.1582-4934]
- 15 Zheng G, Du L, Yang X, Zhang X, Wang L, Yang Y, Li J, Wang C. Serum microRNA panel as biomarkers for early diagnosis of colorectal adenocarcinoma. *Br J Cancer* 2014; **111**: 1985-1992 [PMID: 25233400 DOI: 10.1038/bjc.2014.489]
- 16 Zheng G, Wang H, Zhang X, Yang Y, Wang L, Du L, Li W, Li J, Qu A, Liu Y, Wang C. Identification and validation of reference genes for qPCR detection of serum microRNAs in colorectal adenocarcinoma patients. *PLoS One* 2013; **8**: e83025 [PMID: 24349425 DOI: 10.1371/journal.pone.0083025]
- 17 Guda K, Veigl ML, Varadan V, Nosrati A, Ravi L, Lutterbaugh J, Beard L, Willson JK, Sedwick WD, Wang ZJ, Molyneaux N, Miron A, Adams MD, Elston RC, Markowitz SD, Willis JE. Novel recurrently mutated genes in African American colon cancers. *Proc Natl Acad Sci USA* 2015; **112**: 1149-1154 [PMID: 25583493 DOI: 10.1073/pnas.1417064112]
- 18 Vaiopoulos AG, Athanasoula KCh, Papavassiliou AG. Epigenetic modifications in colorectal cancer: molecular insights and therapeutic challenges. *Biochim Biophys Acta* 2014; **1842**: 971-980 [PMID: 24561654 DOI: 10.1016/j.bbdis.2014.02.006]
- 19 Bardhan K, Liu K. Epigenetics and colorectal cancer pathogenesis. *Cancers (Basel)* 2013; **5**: 676-713 [PMID: 24216997 DOI: 10.3390/cancers5020676]
- 20 Li J, Du L, Yang Y, Wang C, Liu H, Wang L, Zhang X, Li W, Zheng G, Dong Z. MiR-429 is an independent prognostic factor in colorectal cancer and exerts its anti-apoptotic function by targeting SOX2. *Cancer Lett* 2013; **329**: 84-90 [PMID: 23111103 DOI: 10.1016/j.canlet.2012.10.019]
- 21 Cao Z, Moore BT, Wang Y, Peng XH, Lappe JM, Recker RR, Xiao P. MiR-422a as a potential cellular microRNA biomarker for postmenopausal osteoporosis. *PLoS One* 2014; **9**: e97098 [PMID: 24820117 DOI: 10.1371/journal.pone.0097098]
- 22 Abdel-Mohsen M, Deng X, Danesh A, Liegler T, Jacobs ES, Rauch A, Ledergerber B, Norris PJ, Günthard HF, Wong JK, Pillai SK. Role of microRNA modulation in the interferon- α /ribavirin suppression of HIV-1 *in vivo*. *PLoS One* 2014; **9**: e109220 [PMID: 25275557 DOI: 10.1371/journal.pone.0109220]
- 23 Bidzhkeov K, Gan L, Denecke B, Rostalsky A, Hristov M, Koeppel TA, Zerneck A, Weber C. microRNA expression signatures and parallels between monocyte subsets and atherosclerotic plaque in humans. *Thromb Haemost* 2012; **107**: 619-625 [PMID: 22370758 DOI: 10.1160/TH11-09-0607]
- 24 Siegel SR, Mackenzie J, Chaplin G, Jablonski NG, Griffiths L. Circulating microRNAs involved in multiple sclerosis. *Mol Biol Rep* 2012; **39**: 6219-6225 [PMID: 22231906 DOI: 10.1007/s11033-011-1441-7]

- 25 **Song KH**, Li T, Owsley E, Chiang JY. A putative role of micro RNA in regulation of cholesterol 7alpha-hydroxylase expression in human hepatocytes. *J Lipid Res* 2010; **51**: 2223-2233 [PMID: 20351063 DOI: 10.1194/jlr.M004531]
- 26 **Qin YZ**, Xie XC, Liu HZ, Lai H, Qiu H, Ge LY. Screening and preliminary validation of miRNAs with the regulation of hTERT in colorectal cancer. *Oncol Rep* 2015; **33**: 2728-2736 [PMID: 25845814 DOI: 10.3892/or.2015.3892]

P- Reviewer: Voutsadakis IA **S- Editor:** Qi Y **L- Editor:** Wang TQ
E- Editor: Wang CH





Retrospective Study

Colostomy is a simple and effective procedure for severe chronic radiation proctitis

Zi-Xu Yuan, Teng-Hui Ma, Huai-Ming Wang, Qing-Hua Zhong, Xi-Hu Yu, Qi-Yuan Qin, Jian-Ping Wang, Lei Wang

Zi-Xu Yuan, Teng-Hui Ma, Huai-Ming Wang, Qing-Hua Zhong, Xi-Hu Yu, Qi-Yuan Qin, Jian-Ping Wang, Lei Wang, Department of Colorectal Surgery, The Sixth Affiliated Hospital of Sun Yat-Sen University, Guangzhou 510655, Guangdong Province, China

Zi-Xu Yuan, Clinical Research Division, Fred Hutchinson Cancer Research Center, University of Washington, Seattle, WA 98109, United States

Author contributions: Yuan ZX and Ma TH contributed equally to this work; Yuan ZX, Ma TH and Wang L conceived and designed the study; Yuan ZX, Zhong QH, Yu XH, and Qin QY and Wang L conducted the experiments; Yuan ZX, Ma TH, Wang HM and Wang JP analyzed and interpreted the data; Yuan ZX, Ma TH and Wang L wrote and revised the manuscript; all authors approved the final version to be published.

Supported by National Natural Science Foundation of China, No. 81201581, No. 81573078 and No. 81372566; Support Program from Ministry of Science and Technology of China, No. 2014BAI09B06; and Natural Science Foundation of Guangdong Province, China, No. 2016A030311021.

Institutional review board statement: The study was approved by the Ethical Committee of the Sixth Affiliated Hospital of Sun Yat-Sen University and fulfilled the guidelines of the local responsible governmental agency.

Informed consent statement: Informed consent was waived due to the retrospective nature of this study.

Conflict-of-interest statement: The authors declare no conflicts of interests related to the publication of this study.

Data sharing statement: No additional data was available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on

different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Lei Wang, MD, PhD, Professor, Department of Colorectal Surgery, The Sixth Affiliated Hospital of Sun Yat-Sen University, 26 Yuancunerheng Road, Guangzhou 510655, Guangdong Province, China. leiwangyinhu@163.com
Telephone: +86-20-38767131
Fax: +86-20-38254221

Received: March 13, 2016

Peer-review started: March 16, 2016

First decision: March 31, 2016

Revised: April 5, 2016

Accepted: May 4, 2016

Article in press: May 4, 2016

Published online: June 28, 2016

Abstract

AIM: To assess the efficacy and safety of diverting colostomy in treating severe hemorrhagic chronic radiation proctitis (CRP).

METHODS: Patients with severe hemorrhagic CRP who were admitted from 2008 to 2014 were enrolled into this study. All CRP patients were diagnosed by a combination of pelvic radiation history, clinical rectal bleeding, and endoscopic findings. Inclusion criteria were CRP patients with refractory bleeding with moderate to severe anemia with a hemoglobin level < 90 g/L. The study group included patients who were treated by diverting colostomy, while the control group included patients who received conservative treatment. The remission of bleeding was defined as complete cessation or only occasional bleeding that needed no further treatment. The primary outcome was bleeding remission at 6 mo after treatment. Quality of life before

treatment and at follow-up was evaluated according to EORTC QLQ C30. Severe CRP complications were recorded during follow-up.

RESULTS: Forty-seven consecutive patients were enrolled, including 22 in the colostomy group and 27 in the conservative treatment group. When compared to conservative treatment, colostomy obtained a higher rate of bleeding remission (94% *vs* 12%), especially in control of transfusion-dependent bleeding (100% *vs* 0%), and offered a better control of refractory perianal pain (100% *vs* 0%), and a lower score of bleeding ($P < 0.001$) at 6 mo after treatment. At 1 year after treatment, colostomy achieved better remission of both moderate bleeding (100% *vs* 21.5%, $P = 0.002$) and severe bleeding (100% *vs* 0%, $P < 0.001$), obtained a lower score of bleeding (0.8 *vs* 2.0, $P < 0.001$), and achieved obvious elevated hemoglobin levels ($P = 0.003$), when compared to the conservative treatment group. The quality of life dramatically improved after colostomy, which included global health, function, and symptoms, but it was not improved in the control group. Pathological evaluation after colostomy found diffused chronic inflammation cells, and massive fibrosis collagen depositions under the rectal wall, which revealed potential fibrosis formation.

CONCLUSION: Diverting colostomy is a simple, effective and safe procedure for severe hemorrhagic CRP. Colostomy can improve quality of life and reduce serious complications secondary to radiotherapy.

Key words: Chronic radiation proctitis; Rectal bleeding; Diverting colostomy; Quality of life; Serious complication

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: The study describes the efficacy and safety of diverting colostomy in treating severe hemorrhagic chronic radiation proctitis. The procedure focuses on improving the severe refractory bleeding and reducing severe complications. The advantages of diverting colostomy are as follows: it acts effectively and rapidly in controlling severe bleeding that does not respond to conservative treatment; it is a simple procedure that can be conducted in many medical centers; and it can improve quality of life dramatically and reduce serious complications that occur secondary to radiotherapy.

Yuan ZX, Ma TH, Wang HM, Zhong QH, Yu XH, Qin QY, Wang JP, Wang L. Colostomy is a simple and effective procedure for severe chronic radiation proctitis. *World J Gastroenterol* 2016; 22(24): 5598-5608 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5598.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5598>

INTRODUCTION

Chronic radiation proctitis (CRP) is a common complication after radiotherapy of pelvic malignancies, accounting for 5%-20% of cases^[1]. The onset of RP can be delayed for several months to years after radiotherapy. CRP develops as a result of ischemic lesions due to obliterative endarteritis and progressive fibrosis^[2,3]. Rectal bleeding is the most common symptom, which accounts for > 80% of CRP patients^[4]. Acute and mild CRP is usually self-limiting and easy to manage, but moderate to severe CRP is difficult to treat; especially those cases requiring blood transfusions and that are life threatening^[1,5].

Various treatment modalities have been tried. Medical agents include topical sucralfate, steroids^[6], sulfasalazine^[7], metronidazole^[8], rebamipide^[9] and short-chain fatty acid^[10]. Other treatment options include topical formalin^[11,12], endoscopic argon plasma coagulation (APC)^[13], laser therapy^[14], and hyperbaric oxygen therapy^[15]. However, most of these treatments are only useful for mild to moderate bleeding, and severe and refractory bleeding is still problematic^[16]. Furthermore, endoscopic treatments can bring severe side effects and multiple treatment sessions are needed for severe CRP^[17]. In addition, accompanying symptoms such as intractable perianal pain, urgency and tenesmus in CRP are usually hard to manage.

Diverting colostomy has been reported previously, mainly for severe CRP complications^[18,19]. Colostomy can reduce the irritation injury of fecal stream to the irradiated tissues and thus decrease rectal bleeding. However, unlike formalin or APC, colostomy is now not widely used. The issue of colostomy is not well studied to date. To the best of our knowledge, no study has compared diverting colostomy to conservative treatment. The aim of this study was to assess the efficacy and safety of diverting colostomy in severe CRP. The indications, quality of life, severe CRP complications, and stoma reversals after colostomy were also investigated, when compared to conservative treatment.

MATERIALS AND METHODS

Patients and ethical statements

Hemorrhagic CRP patients who were treated at the Sixth Affiliated Hospital of Sun Yat-Sen University (SYSU) from March 2008 to October 2014 were retrospectively enrolled in this study. Electronic files and medical records were both carefully collected to extract clinicopathological data. This study was approved by the Ethical Committee of the Sixth Affiliated Hospital of SYSU and the study was conducted in accordance with the provisions of the World Medical Association's Declaration of Helsinki in 1995 (revised in Tokyo, 2004). Due to the nature of

Table 1 Modified subjective objective management analysis system to assess the severity of bleeding in radiation proctitis

Grade	Bleeding	Severity	Anemia (Hb, g/L)
1	Mild bleeding	Occasionally or occult	Mild anemia (Hb: ≥ 90 g/L)
2	Moderate bleeding	Persistent	Moderate anemia (Hb: 70-90 g/L)
3	Severe bleeding	Gross	Severe anemia, transfusion needed (Hb: < 70 g/L)

Hb: Hemoglobin.

the retrospective study, informed consent was waived.

Inclusion and exclusion criteria

Inclusion criteria were CRP patients with refractory bleeding with moderate to severe anemia with a hemoglobin level < 90 g/L. Refractory bleeding was defined as no response to conservative treatment. Patients who were treated with diverting stomas were enrolled in the study group, while those who continued to non-surgical treatment were enrolled in the control group. Patients with tumor relapses, loss to follow-up, or who underwent rectal resection with preventive colostomy were excluded.

Diagnosis, scores and definitions

All patients were diagnosed by combination of pelvic radiation history, clinical rectal bleeding, and endoscopic findings of injured rectal mucosa. Flexible colonoscopy was performed in all patients to rule out other causes of bleeding, such as recurrent tumors, inflammatory bowel disease and anal benign hemorrhagic diseases. The Vienna Rectoscopy Score^[20] system was used to assess endoscopic severity.

Current scores to evaluate the severity of bleeding included Common Terminology Criteria for Adverse Events^[21], Radiation Therapy Oncology Group/European Organization for Research and Treatment of Cancer (EORTC) score^[22]. However, most of them are based on subjective complaints of patients, instead of accurate laboratory tests. Because the severity of bleeding was mainly reflected by a decrease in hemoglobin level, we designed a modified Subjective Objective Management Analysis (SOMA) system reported in a previous study^[23], to assess the severity of bleeding, included both subjective complaints of bleeding and objective hemoglobin level. All patients were scored by the system (Table 1). The remission of bleeding was defined as complete cessation of bleeding or only occasional bleeding that needed no further treatment. Failure in the conservative treatment group was defined as no improvement or even worse bleeding and decreased hemoglobin level 6 mo after treatment.

Indications of diverting colostomy

All patients, except those with fistulas, were initially

treated with medical agent enemas including almagate (one mucosa-protector like sucralfate), corticosteroids, and metronidazole. Topical formalin (details listed in our previous study)^[24] or endoscopic APC were suggested when they experienced recurrent bleeding. As for refractory and transfusion-dependent CRP after these conservative measures, physicians suggested a diverting stoma. If patients refused a colostomy and demanded to continue conservative treatment, they were enrolled in the control group. Other indications of diverting colostomy were as follow: (1) fistula, perforation or stricture; and (2) deep ulcer with refractory perianal pain.

Diverting loop colostomies were conducted under general anesthesia in the operating room. The transverse colon was pulled out through a small incision, then a soft catheter of a stent was inserted to prevent stoma retraction, and a double-cavity stoma of the transverse colon was then created. The catheter was removed postoperatively. Classical images of a double-cavity colostomy and a "gunsight" of stoma closure were shown (Figure 1). This technique of stoma closure can simplify wound care, decrease surgical site infection, and give a neat cosmetic result^[25,26].

Follow-up

Follow-up was scheduled through outpatient visits or telephone questionnaires at 6 mo and 1 year after treatment. The quality of life before treatment and at follow-up was evaluated according to EORTC QLQ C30^[27]. The primary outcome was the remission rate of bleeding at 6 mo after treatment. The secondary outcomes included hemoglobin level, remission rate of bleeding at 1 year after treatment, quality of life, stoma-related complications, severe CRP complications, and stoma reversal rate.

Statistical analysis

Comparisons of characteristics were made by Student's *t* test analysis for continuous variables. For categorical variables, the χ^2 test was used. Fisher's exact test was adopted when appropriate. For non-parameter variables, the Wilcoxon rank sum test was used. All statistical analyses were performed by SPSS version 17.0 (Chicago, IL, United States). $P < 0.05$ (two-tails) was considered to be statistically significant.

RESULTS

Demographics and characteristics

A total of 47 patients were analyzed. Twenty-two (46.8%) were treated by diverting colostomy, and 27 (53.2%) were managed by conservative treatment (Figure 2). Forty-three (91.5%) were women, and 40 (85.1%) primary malignancies were cervical cancer. Cumulative radiation dosage of one patient was about 80 Gy, which included the radiation for both sites

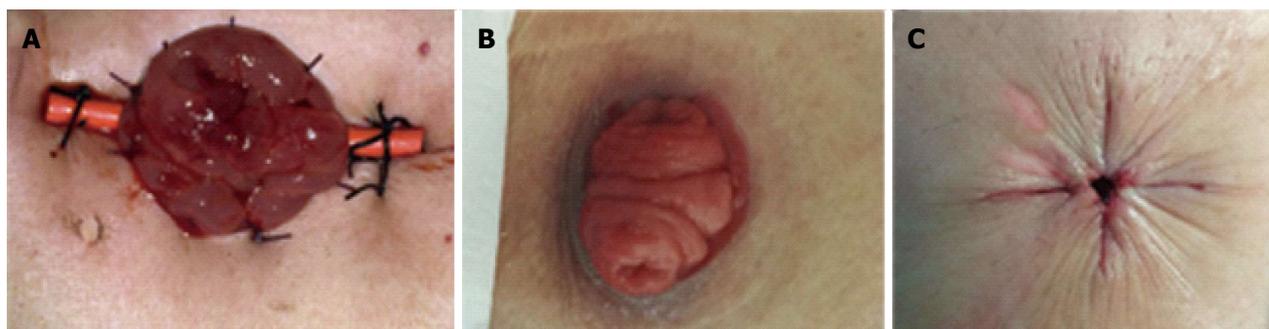


Figure 1 Classic representative images of a double-cavity colostomy. A: A double-cavity colostomy with a soft catheter as a stent; B: A colostomy after during follow-up; C: A "gunsight" skin incision and closure for stoma reversal.

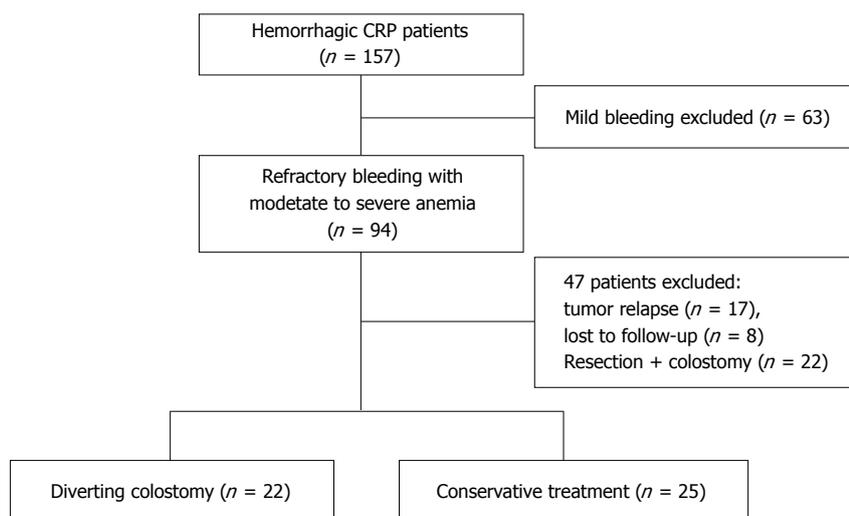


Figure 2 Flow chart of patient selection.

of primary malignancy and invasive lymph nodes. The detailed radiotherapy for those patients with gynecological cancers, especially cervical cancer, was 25 rounds (2 Gy/round) of external beam radiation and five or six episodes (6 Gy/episode) of intra-cavity brachytherapy. Patients with prostate or rectal cancers received only external beam radiation. When comparing demographics prior to treatment between the two groups, there were no significant differences in age, gender, type of primary malignancy, cumulative radiation dosage, latency period, duration from treatment to end of radiotherapy, duration of bleeding, albumin level, body mass index (BMI), concomitant radiation uropathy, radiation enteritis, and associated risk factors of CRP such as previous history of abdominal surgery, diabetes mellitus and hypertension (Table 2). Thus, these above characteristics were comparable between the two groups. However, the colostomy group had a higher score of bleeding (2.7 vs 2.0, $P < 0.001$) and a lower hemoglobin level (60.8 g/L vs 88.2 g/L, $P < 0.001$), when compared to the conservative treatment group, respectively. These results indicated that the colostomy group had more serious bleeding before treatment (Table 2).

Treatment

The indications for colostomy were as follows: (1) severe bleeding in eight (36.4%) cases; (2) fistulas in 11 (50%), including nine (40.9%) rectovaginal fistulas and two (9.1%) sigmoid-vesical-vaginal fistulas; (3) deep ulcer + refractory perianal pain in two (9.1%) cases; and (4) severe bleeding + deep ulcer + anal stricture in one (4.5%) case. Among these 11 patients with fistulas, five also had concomitant severe bleeding. Among these nine recto-vaginal fistulas, one had concomitant recto-urethral fistula, and another had concomitant recto-vesical fistula and small bowel fistula.

In the conservative treatment group, seven patients received topical formalin irrigation and one received APC treatment after enrollment. All eight (32%) cases transiently obtained bleeding remission, but only two (25%) obtained long-term remission of bleeding. The other six patients experienced recurrent bleeding and developed severe anemia. Repeat topical formalin achieved only limited efficacy in these patients with severe anemia (average 2 sessions of formalin at 2-4-wk intervals). The remaining 17 patients refused formalin treatment, and thus continued retention

Table 2 Comparisons of patient demographics between the colostomy group and conservative group before treatment *n* (%)

Characteristics	Diverting colostomy (<i>n</i> = 22)	Conservative treatment (<i>n</i> = 25)	<i>P</i> value
Age (mean ± SD)	60.1 ± 2.2	60.2 ± 2.4	0.964
Gender (female/male)	20/2	23/2	1.000 ¹
Primary malignancy			0.822 ²
Cervical cancer	19 (86.4)	21 (84)	
Endometrial cancer	2 (9.1)	2 (8)	
Rectal cancer	1 (4.5)	1 (4)	
Prostate cancer	0 (0)	1 (4)	
Cumulative radiation dosage (Gy), mean ± SD	80.5 ± 17.3	83.6 ± 20.5	0.842 ³
Latency period (mo), mean ± SD	8.3 ± 0.8	7.2 ± 1.1	0.252 ³
Duration from treatment to end of radiotherapy (mo), mean ± SD	16.3 ± 1.3	14.8 ± 1.6	0.277 ³
Duration of bleeding (month), mean ± SD	7.9 ± 1.0	8.0 ± 1.8	0.466 ¹
Score of bleeding, mean ± SD	2.7 ± 0.5	2.0 ± 0.5	< 0.001 ³
Mean hemoglobin (g/L), mean ± SD	60.8 ± 18.1	88.2 ± 19.3	< 0.001
Alb (g/L), (≤ 35/> 35)	6/16	3/22	0.339 ¹
BMI (kg/m ²), (≤ 17.5/> 17.5)	3/19	2/23	0.880 ¹
Concomitant radiation uropathy	8 (36.4)	6 (24)	0.355
Radiation enteritis	5 (22.7)	2 (8)	0.315 ¹
Previous abdominal surgery	6 (27.3)	11 (44)	0.234
Diabetes mellitus	2 (9.1)	3 (12)	1.000 ¹
Hypertension	6 (27.3)	7 (28)	0.956

¹χ² test; ²Fisher exact test; ³Wilcoxon rank-sum test. BMI: Body mass index; Alb: Albumin.

Table 3 Bleeding remissions in severe radiation proctitis after treatment

Variables	Diverting Colostomy (<i>n</i> = 22)	Conservative treatment (<i>n</i> = 25)	<i>P</i> value
6 mo after treatment			
Remission of bleeding	17/18 (94%)	3/25 (12%)	< 0.001 ¹
Remission of refractory perianal pain	8/8 (100%)	0/6 (0%)	< 0.001 ²
Score of bleeding, mean ± SD	1.1 ± 0.5	2.2 ± 0.7	< 0.001 ³
Elevated Hb, mean ± SD	34.1 ± 18.2	-12.3 ± 9.1 ⁴	< 0.001
Remission of moderate bleeding	8/8 (100%)	6/19 (21.5%)	0.002 ²
Remission of severe bleeding	11/11 (100%)	0/5 (0)	< 0.001 ²
Score of bleeding, mean ± SD	0.8 ± 0.5	2.0 ± 0.9	< 0.001 ³
Post-treatment recto-vaginal fistula	0/22	3/25 (12%)	0.237 ²
Elevated Hb, mean ± SD	40.3 ± 19.3	-1.9 ± 32.5 ⁴	0.003

¹χ² test; ²Fisher exact test; ³Wilcoxon rank-sum test; ⁴Represents decreased level.

enemas and transfusions when needed.

Outcomes

During a mean 22 (range: 6-77) mo of follow-up, eight (17%) patients died. The cause of death was recurrent malignancy in seven cases. The other one died of bladder perforation and sepsis that occurred secondary to radiation recto-vesical-vaginal perforation. At 6 mo after treatment, colostomy offered higher remission of bleeding (94% vs 12%, *P* < 0.001), higher remission of refractory perianal pain (100% vs 0%, *P* < 0.001), decreased scores of bleeding (1.1 vs 2.2, *P* < 0.001), and obvious increased hemoglobin levels (34.1 g/dL vs -12.3 g/dL, *P* < 0.001), compared to the conservative treatment group. At 1 year after treatment, colostomy achieved still higher remission of both moderate bleeding (100% vs 21.5%, *P* = 0.002) and severe bleeding (100% vs 0%, *P* < 0.001), acquired lower score of bleeding (0.8 vs 2.0, *P* <

0.001), and obviously elevated hemoglobin levels (40.3 g/dL vs -1.9 g/dL, *P* = 0.003), than those cases in the conservative treatment group. In addition, three recto-vaginal fistulas were found in the conservative treatment group during follow-up, but no new fistula occurred after the operation in the colostomy group. Patients who did not have bleeding remission continued conservative treatment at home (Table 3).

Stoma closure and complications

Of the eight patients who received colostomy to control severe bleeding, three (37.5%) underwent stoma closure (2 cases at 9 mo and 1 at 10 mo after colostomy). All three had no bleeding and remained well after stoma reversal. Of the remaining five, all obtained bleeding remission and improved hemoglobin levels. However, among these five, one had grade IV New York Heart Association heart failure and could not risk stoma closure. Four were unsuitable for closure

Table 4 Quality of life between diversion group and conservative group in CRP patients by EORTC QLQ-C30 scale

QLQ-C30 scale	Ref. (Normal German population)	Diverting colostomy group (<i>n</i> = 18), mean (SD)				Conservative treatment group (<i>n</i> = 23), mean (SD)			
		Pre-treatment	Follow-up	Δ(FU)-Pre ¹	Significance ²	Pre-treatment	Follow-up	Δ(FU)-Pre	Significance ²
Global health	63.2	23.1 (15.1)	64.8 (13.8)	41.7	< 0.001	47.1 (21.5)	62.3 (25.0)	15.2	0.033
Physical function	82.6	50.7 (17.8)	77.8 (16.6)	27.1	< 0.001	78.0 (22.7)	78.6 (26.1)	0.6	0.856
Role function	75.0	34.3 (23.9)	75.9 (27.3)	41.6	< 0.001	77.5 (24.9)	77.5 (29.1)	0.0	0.775
Emotional function	62.2	46.3 (27.4)	73.6 (27.0)	27.3	0.001	75.7 (17.6)	80.8 (23.8)	5.1	0.384
Cognition function	81.3	92.6 (12.7)	93.5 (9.8)	0.9	0.581	94.2 (15.6)	95.7 (9.0)	1.5	0.798
Social function	78.4	43.5 (28.9)	65.7 (31.7)	22.2	0.004	91.3 (20.6)	89.1 (21.1)	-2.2	0.916
Fatigue	34.1	72.8 (12.9)	36.4 (25.9)	-36.4	< 0.001	26.6 (24.1)	23.2 (27.2)	-3.4	0.695
Nausea/vomiting	5.7	4.6 (15.5)	1.9 (7.6)	-2.7	0.109	5.1 (15.4)	6.5 (16.5)	1.4	0.655
Pain	33.1	44.4 (28.3)	14.8 (22.8)	-29.6	0.001	14.5 (21.5)	11.6 (18.4)	-2.9	0.481
Dyspnea	18.8	42.6 (31.0)	18.5 (22.8)	-24.1	0.003	14.5 (19.7)	8.7 (18.0)	-5.8	0.210
Insomnia	38.5	48.1 (35.5)	29.6 (31.2)	-18.5	0.026	21.7 (21.6)	26.1 (31.7)	4.4	0.287
Appetite loss	9.4	22.2 (27.2)	13.0 (19.7)	-9.2	0.125	10.1 (25.5)	8.7 (18.0)	-1.4	0.785
Constipation	9.1	9.3 (14.9)	3.7 (10.5)	-5.6	0.066	8.7 (20.6)	7.2 (22.4)	-1.5	0.414
Diarrhea	9.2	33.3 (33.3)	22.2 (24.8)	-11.1	0.018	11.6 (23.8)	2.9 (9.6)	-8.7	0.078
Financial difficulties	17.1	59.3 (26.2)	50.0 (31.9)	-9.3	0.211	26.1 (31.7)	24.6 (30.5)	-1.5	0.595

¹Wilcoxon rank-sum test; ²Point (follow-up) - point (pre-treatment).

because of erythema and telangiectasia at 6 mo after colostomy, and two of these four had not yet reached 1 year follow-up to assess the lesion by colonoscopy.

Stoma complications were found in seven (31.8%) cases, which contained six stoma prolapses and one stoma stricture. Of these six stoma prolapses, four were managed with conservative measures by manual repositions (Grade II by Clavien-Dindo classification^[28]), and two required stoma rebuilding (Grade III). One stoma stricture occurred in a stoma of the descending colon, instead of the transverse colon, and stoma stricture was managed by finger dilatation (Grade II).

Quality of life

The quality of life was evaluated in 41 (87.2%) patients by the EORTC QLQ-C30 questionnaires. Because there were no similar reports in the Chinese population, the values were referred to the normal German population. Osoba *et al.*^[29] suggested that a difference of ≥ 20 points in global health was considered to be clinically relevant. In this study, when compared to pre-treatment, diverting colostomy improved quality of life, including improved global health (difference = 42, $P < 0.001$), improved functions like physical function ($P < 0.001$), role function ($P < 0.001$), emotional function ($P < 0.001$), social function ($P < 0.001$), and improved symptoms like fatigue ($P < 0.001$), pain ($P = 0.001$), dyspnea ($P = 0.003$), insomnia ($P = 0.026$), and diarrhea ($P = 0.018$). However, conservative treatment did not significantly improve quality of life at follow-up compared with before treatment (Table 4).

Endoscopic and pathological features

Classical endoscopic images prior to colostomy and at stoma closure from three patients with stoma closures were collected (Figure 3). After a mean 9.3 (range:

9-10) mo after colostomy, endoscopic lesions of active bleeding, multiple confluent telangiectasia, congested mucosa, or even ulcer were greatly improved and reached the criteria of stoma closure.

Pathological evaluation of endoscopic biopsy from rectal lesions was conducted at 31 mo after colostomy in Case 3. This patient obtained complete remission of bleeding and the biopsy sites were fully healed according to endoscopic observation at follow-up. Pathologically, diffuse chronic inflammatory cells were observed in the mucosa and sub-mucosa layers. In addition, massive fibrotic collagen depositions were found in the sub-mucosa layer, which revealed fibrosis formation (Figure 4).

DISCUSSION

Endoscopic treatments, such as topical formalin or APC, are extensively used for mild to moderate hemorrhagic CRP worldwide^[14,16]. However, these treatments show only limited long-term efficacy for severe CRP^[17]. It is also unclear how many patients have received these treatments for serious transfusion-dependent bleeding in previous studies^[17]. Meanwhile, serious complications can be caused by these endoscopic treatments^[5,17]. Our previous study also suggested that topical formalin should not be applied in CRP with ulcer because of the risk of fistula^[24]. In the present study, patients in the conservative treatment group received more topical formalin or APC treatment and developed more fistulas later, than patients in the colostomy group. Therefore, we suggest topical formalin or APC should be selected cautiously for patients with severe CRP. Recently, radiofrequency ablation (RFA) in treating CRP has been introduced, with improvement in hemoglobin level and decrease in clinical symptoms^[30,31]. Most RFA studies are based

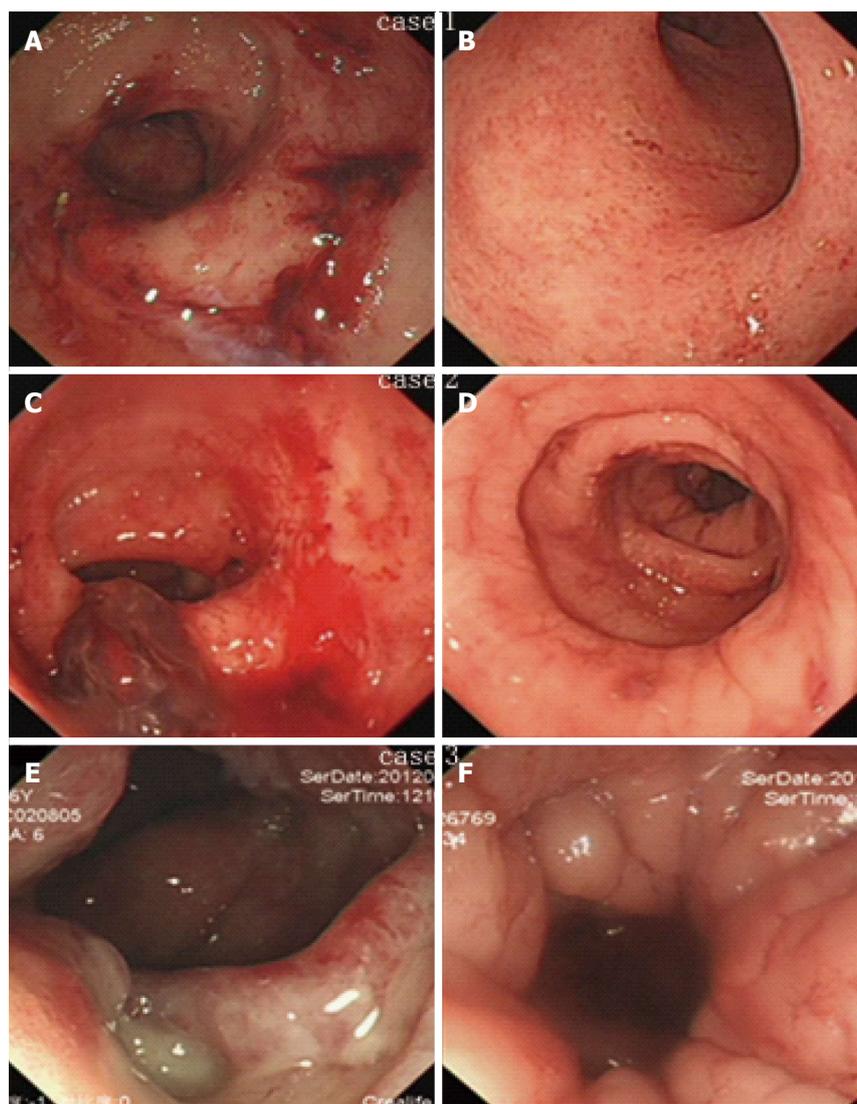


Figure 3 Classical endoscopic images before colostomy and at stoma reversal. Severe active bleeding (A and C), or confluent telangiectasia, edema and ulcer (E) were observed in cases 1-3 before colostomy, while these lesions were greatly improved at stoma reversal (B, D, F).

on retrospective case series without controls and current data are scarce, therefore, prospective trials of RFA should be conducted in the future to validate its efficacy and application in severe CRP.

Surgical intervention is often the last resort for severe CRP^[3,19]. Rectal resection is controversial, because it is difficult to perform a safe anastomosis in the radiation-injured tissue, and high risks of anastomotic leak and death from postoperative peritonitis are reported^[19]. Therefore, a simple and safe procedure to save life and relieve symptoms is mandatory. Theoretically, diverting colostomy can reduce bacterial contamination and decrease irritation injury by fecal stream, and colostomy can gain time to subside any radiation reaction to protect injured tissue^[32]. Thus, severe bleeding and refractory perianal pain can be controlled. Colostomy can also accelerate the course of fibrosis and relieve severe proctitis rapidly, which may prevent deep ulcers

progressing to fistulas. In our recent study^[33], we reported typical histopathological features of CRP: telangiectasia, abnormal hyaline-like wall vessels and sporadic radiation fibrocytes in the submucosal layer. In this study, consistently, massive fibrotic collagen depositions were observed in irradiated tissue after colostomy. Collectively, the nature of CRP is a progressive fibrosis course^[7]. The efficacy and safety of colostomy have been reported in previous studies^[18,19,34]. However, most previous studies did not contain controls, which could not discriminate the efficacy of interventions from the self-remission course. To the best of our knowledge, no previous study has reported quality of life after colostomy. It is also not clear whether colostomy can reduce serious CRP complications.

In this study, diverting colostomy resolved most cases of severe bleeding. No recurrent bleeding and no colostomy-related death were observed. Colostomy

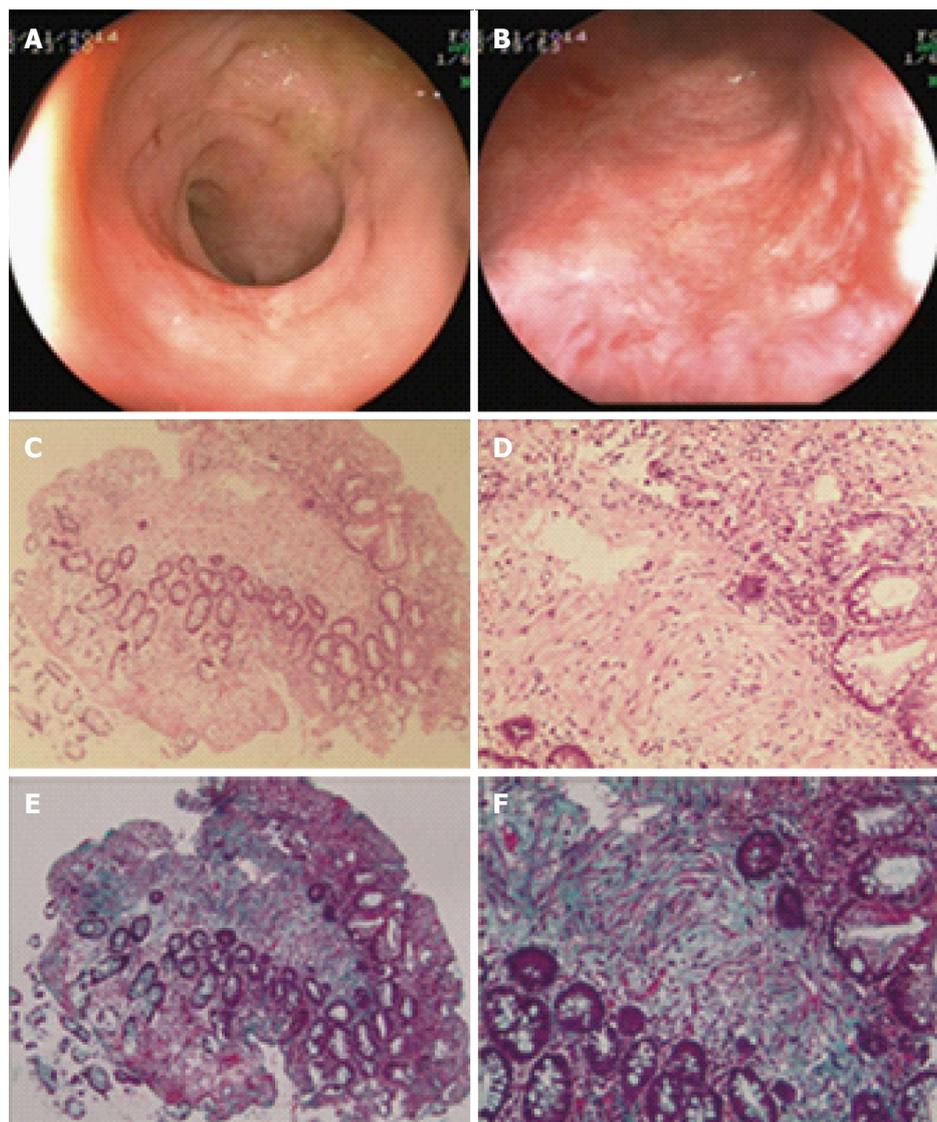


Figure 4 Histo-pathological features after colostomy in case 3. Complete remission of confluent telangiectasia, edema and ulcer were observed 31 mo after colostomy (A and B). Diffused chronic inflammatory cells and rebuilt of mucosal integrity were seen in hematoxylin and eosin-stained slides (C: $\times 100$; D: $\times 400$). Massive fibrotic collagen depositions were observed in the sub-mucosa layer, with green color by Masson staining (E: $\times 100$; F: $\times 400$).

can also decrease serious complications, including remission of long-term perianal pain, transfusion-dependent bleeding and fistula. Deep ulcer with severe bleeding is a contraindication to endoscopic treatment, because it is easy to progress to fistula, due to the poor-healing capacity and impaired blood supply of friable intestinal wall^[17,19]. We have previously tried topical revision and skin flap transplantation for some CRP fistulas. However, the efficacy was limited and new fistulas can occur rapidly, due to poor healing capacity of irradiated mucosa and bacterial infection from the fecal stream, which leads to treatment failure. In this study, two patients with deep ulcer and severe bleeding were successfully controlled by colostomy, and no fistula was found at follow-up.

Transverse colostomy is preferred because it provides a greater blood supply by preserving superior rectal and left marginal vessels, and provides more

options for later possible recto-sigmoid resection than sigmoid colostomy does, and transverse colostomy is easier to be closed and is more effective^[32,34]. In addition, the recto-sigmoid colon is expected to receive a higher radiation dose for pelvic malignancy, while the transverse colon receives the least radiation and damage, because it is located far from pelvic tumors, and thus causes the fewest complications^[19,32,35]. Loop ileostomy is not widely used because of the risk of high-volume fluid discharge. Colostomy-related complications were reported, ranging from 21.8% to 40%^[19,35]. Consistent with previous studies, colostomy-related complications in this study were 31.8%, including six (85%) cases of Grade II and one (15%) case of Grade III complications. Among them, stoma prolapse was a common complication.

Colostomy for recto-vaginal fistula and rectal stricture is permanent. However, colostomy for patients

unresponsive to medical treatment can be closed when severe proctitis improves sufficiently. Anseline *et al.*^[19] reported six (43%) colostomy reversals in 14 CRP patients, who were unresponsive to medical treatment. A similar result was observed in the present study. Three (38%) of eight patients with severe bleeding were closed successfully in a mean 9 mo after colostomy. Because the duration of follow-up after fecal diversion was short, many patients who obtained long-term remission of bleeding after colostomy had great potential to reverse the stoma.

In this study, we used a modified SOMA system, which coordinates subjective bleeding symptoms and objective accurate hemoglobin level. According to the system, we suggest that mild to moderate hemorrhagic CRP can be managed by medical or endoscopic treatment. However, for severe refractory bleeding colostomy should be considered to prevent development of serious complications. The scoring system will guide physicians in primary care to evaluate patient condition according to hemoglobin level, and then choose the appropriate treatment. Having a routine and easy protocol can reduce treatment-related delays and avoid unnecessary morbidity^[7].

In this study, 44 (94%) CRP patients enrolled had gynecological cancers, so most fistulas were documented in women. In western countries, patients with prostate cancer are the dominant population receiving pelvic radiation, and CRP is mainly reported in prostate cancer^[4,12]. However, prostate cancer receives only external beam radiation such as 3D conformal radiotherapy or intensity-modulated radiotherapy, and it does not receive intra-cavity brachytherapy, thus, fewer fistulas are observed. According to our clinical practice, intra-cavity radiation can result in more fistulas and other severe adverse radiation-related symptoms than external beam radiation can.

Although the colostomy group had more severe bleeding than the conservative treatment group, which could have resulted in selection bias, it achieved dramatically better control of bleeding, higher increased hemoglobin level, and improved quality of life compared with the conservative treatment group. These results have shown the advantages of diverting colostomy in treating severe CRP bleeding. Topical formalin or APC was not used in all the patients in the conservative treatment group, because some patients in China have not sufficient knowledge of CRP, poor compliance with physicians' advice, and poor economic status. Thus, they choose to continue self-enemas at home, when recurrent bleeding occurs. In addition, this study was limited by its non-randomized, retrospective design and small sample size. Additional randomized prospective studies of diverting colostomy are needed to confirm our findings.

ACKNOWLEDGMENTS

The authors thank Jie Zhao, Li-Li Chu for the contributions in the collection of stoma pictures and providing stoma cares. We also thank Yan-Qi Liu for the contribution in patient follow-up.

COMMENTS

Background

Chronic radiation proctitis (CRP) occurs in 5%-20% of patients receiving radiotherapy for pelvic malignant tumors. Mild to moderate CRP is usually self-limiting and easy to manage, but severe and refractory bleeding is still problematic, especially in cases requiring blood transfusions and that are life threatening. Furthermore, endoscopic treatment can cause severe side effects and only limited efficacy can be obtained for severe CRP. Thus, a simple and safe treatment with fewer complications to save life and relieve symptoms is mandatory.

Research frontiers

Diverting colostomy has been reported previously, mainly for severe CRP complications. However, unlike formalin or argon plasma coagulation, colostomy is now not widely used for severe bleeding in CRP patients. The issue of colostomy is not well studied to date. To the best of our knowledge, no study has compared diverting colostomy to conservative measures in treating severe hemorrhagic CRP.

Innovations and breakthroughs

In this series, the authors reported their experience that diverting colostomy was a simple, effective and safe procedure for severe hemorrhagic CRP. Furthermore, they found that colostomy improved quality of life and reduced serious complications secondary to radiotherapy, while conservative medical and endoscopic treatments did not show efficacy in severe CRP patients.

Applications

Diverting colostomy is a simple and safe procedure that can be performed in most medical centers. The authors also developed a modified Subjective Objective Management Analysis system, which coordinates subjective bleeding symptoms and objective accurate hemoglobin level, to guide physicians in primary care to evaluate patient condition according to hemoglobin level, and then choose the appropriate treatment. Having a routine and easy protocol can reduce treatment-related delays and avoid unnecessary morbidity.

Terminology

The underlying causes of CRP are endarteritis obliterans and progressive submucosal fibrosis due to radiotherapy. Diverting colostomy can reduce bacterial contamination and decrease irritation injury by the fecal stream, and can gain time to reduce any radiation reaction to protect injured tissue. Colostomy can also accelerate the course of fibrosis and relieve severe proctitis rapidly, which may prevent deep ulcers progressing to fistulas.

Peer-review

This is a single center, controlled, and retrospective case series of severe CRP patients who received diverting colostomy. Colostomy can relieve most of severe bleeding rapidly and unexpectedly, colostomy can also reduce serious CRP complications, including remission of long-term perianal pain, transfusion-dependent bleeding and fistula.

REFERENCES

- 1 Leiper K, Morris AI. Treatment of radiation proctitis. *Clin Oncol (R Coll Radiol)* 2007; 19: 724-729 [PMID: 17728120 DOI: 10.1016/

- j.clon.2007.07.008]
- 2 **Hasleton PS**, Carr N, Schofield PF. Vascular changes in radiation bowel disease. *Histopathology* 1985; **9**: 517-534 [PMID: 4007790 DOI: 10.1111/j.1365-2559.1985.tb02833.x]
 - 3 **Sahakitrungruang C**, Thum-Ummuaysuk S, Pati Wongpaisarn A, Atitthansakul P, Rojanasakul A. A novel treatment for haemorrhagic radiation proctitis using colonic irrigation and oral antibiotic administration. *Colorectal Dis* 2011; **13**: e79-e82 [PMID: 21114751 DOI: 10.1111/j.1463-1318.2010.02527.x]
 - 4 **Placer C**, Lizarazu A, Borda N, Elósegui JL, Enriquez Navascués JM. [Radiation proctitis and chronic and refractory bleeding. Experience with 4% formaldehyde]. *Cir Esp* 2013; **91**: 111-114 [PMID: 23036255 DOI: 10.1016/j.ciresp.2012.05.017]
 - 5 **Haas EM**, Bailey HR, Farragher I. Application of 10 percent formalin for the treatment of radiation-induced hemorrhagic proctitis. *Dis Colon Rectum* 2007; **50**: 213-217 [PMID: 17080283 DOI: 10.1007/s10350-006-0707-y]
 - 6 **Kochhar R**, Patel F, Dhar A, Sharma SC, Ayyagari S, Aggarwal R, Goenka MK, Gupta BD, Mehta SK. Radiation-induced proctosigmoiditis. Prospective, randomized, double-blind controlled trial of oral sulfasalazine plus rectal steroids versus rectal sucralfate. *Dig Dis Sci* 1991; **36**: 103-107 [PMID: 1670631]
 - 7 **Gul YA**, Prasannan S, Jabar FM, Shaker AR, Moissinac K. Pharmacotherapy for chronic hemorrhagic radiation proctitis. *World J Surg* 2002; **26**: 1499-1502 [PMID: 12297939 DOI: 10.1007/s00268-002-6529-8]
 - 8 **Cavčić J**, Turčić J, Martinac P, Jelincić Z, Zupancić B, Panijan-Pezerović R, Unusić J. Metronidazole in the treatment of chronic radiation proctitis: clinical trial. *Croat Med J* 2000; **41**: 314-318 [PMID: 10962052]
 - 9 **Kim TO**, Song GA, Lee SM, Kim GH, Heo J, Kang DH, Cho M. Rebampide enema therapy as a treatment for patients with chronic radiation proctitis: initial treatment or when other methods of conservative management have failed. *Int J Colorectal Dis* 2008; **23**: 629-633 [PMID: 18327596 DOI: 10.1007/s00384-008-0453-9]
 - 10 **Talley NA**, Chen F, King D, Jones M, Talley NJ. Short-chain fatty acids in the treatment of radiation proctitis: a randomized, double-blind, placebo-controlled, cross-over pilot trial. *Dis Colon Rectum* 1997; **40**: 1046-1050 [PMID: 9293933 DOI: 10.1007/BF02050927]
 - 11 **Nelamangala Ramakrishnaiah VP**, Javali TD, Dharanipragada K, Reddy KS, Krishnamachari S. Formalin dab, the effective way of treating haemorrhagic radiation proctitis: a randomized trial from a tertiary care hospital in South India. *Colorectal Dis* 2012; **14**: 876-882 [PMID: 22356304 DOI: 10.1111/j.1463-1318.2012.03008.x]
 - 12 **Patel P**, Subhas G, Gupta A, Chang YJ, Mittal VK, McKendrick A. Oral vitamin A enhances the effectiveness of formalin 8% in treating chronic hemorrhagic radiation proctopathy. *Dis Colon Rectum* 2009; **52**: 1605-1609 [PMID: 19690489 DOI: 10.1007/DCR.0b013e3181afbe3a]
 - 13 **Yeoh E**, Tam W, Schoeman M, Moore J, Thomas M, Botten R, Di Matteo A. Argon plasma coagulation therapy versus topical formalin for intractable rectal bleeding and anorectal dysfunction after radiation therapy for prostate carcinoma. *Int J Radiat Oncol Biol Phys* 2013; **87**: 954-959 [PMID: 24113059 DOI: 10.1016/j.ijrobp.2013.08.034]
 - 14 **Hanson B**, MacDonald R, Shaukat A. Endoscopic and medical therapy for chronic radiation proctopathy: a systematic review. *Dis Colon Rectum* 2012; **55**: 1081-1095 [PMID: 22965408 DOI: 10.1097/DCR.0b013e3182587aef]
 - 15 **Charneau J**, Bouachour G, Person B, Burtin P, Ronceray J, Boyer J. Severe hemorrhagic radiation proctitis advancing to gradual cessation with hyperbaric oxygen. *Dig Dis Sci* 1991; **36**: 373-375 [PMID: 1995275 DOI: 10.1007/BF01318212]
 - 16 **Karamanolis G**, Psatha P, Triantafyllou K. Endoscopic treatments for chronic radiation proctitis. *World J Gastrointest Endosc* 2013; **5**: 308-312 [PMID: 23858374 DOI: 10.4253/wjge.v5.i7.308]
 - 17 **Andreyev J**. Gastrointestinal symptoms after pelvic radiotherapy: a new understanding to improve management of symptomatic patients. *Lancet Oncol* 2007; **8**: 1007-1017 [PMID: 17976611 DOI: 10.1016/s1470-2045(07)70341-8]
 - 18 **Photopulos GJ**, Jones RW, Walton LA, Fowler WC. A simplified method of complete diversionary colostomy for patients with radiation-induced proctosigmoiditis. *Gynecol Oncol* 1977; **5**: 180-186 [PMID: 881129 DOI: 10.1016/0090-8258(77)90022-1]
 - 19 **Anselme PF**, Lavery IC, Fazio VW, Jagelman DG, Weakley FL. Radiation injury of the rectum: evaluation of surgical treatment. *Ann Surg* 1981; **194**: 716-724 [PMID: 7305485 DOI: 10.1097/0000658-198112000-00010]
 - 20 **Wachter S**, Gerstner N, Goldner G, Pötzi R, Wambersie A, Pötter R. Endoscopic scoring of late rectal mucosal damage after conformal radiotherapy for prostatic carcinoma. *Radiother Oncol* 2000; **54**: 11-19 [PMID: 10719695 DOI: 10.1016/S0167-8140(99)00173-5]
 - 21 Common Terminology Criteria for Adverse Events v4.0 (CTCAE) Available from: URL: http://evs.nci.nih.gov/ftp1/CTCAE/CTCAE_4.03_2010-06-14_QuickReference_8.5x11.pdf
 - 22 **Cox JD**, Stetz J, Pajak TF. Toxicity criteria of the Radiation Therapy Oncology Group (RTOG) and the European Organization for Research and Treatment of Cancer (EORTC) *Int J Radiat Oncol Biol Phys* 1995; **31**: 1341-1346 [PMID: 7713792 DOI: 10.1016/0360-3016(95)00060-c]
 - 23 **Coia LR**, Myerson RJ, Tepper JE. Late effects of radiation therapy on the gastrointestinal tract. *Int J Radiat Oncol Biol Phys* 1995; **31**: 1213-1236 [PMID: 7713784 DOI: 10.1016/0360-3016(94)00419-1]
 - 24 **Ma TH**, Yuan ZX, Zhong QH, Wang HM, Qin QY, Chen XX, Wang JP, Wang L. Formalin irrigation for hemorrhagic chronic radiation proctitis. *World J Gastroenterol* 2015; **21**: 3593-3598 [PMID: 25834325 DOI: 10.3748/wjg.v21.i12.3593]
 - 25 **Lim JT**, Shedda SM, Hayes IP. "Gunsight" skin incision and closure technique for stoma reversal. *Dis Colon Rectum* 2010; **53**: 1569-1575 [PMID: 20940608 DOI: 10.1007/DCR.0b013e3181f0535a]
 - 26 **Hsieh MC**, Kuo LT, Chi CC, Huang WS, Chin CC. Pursestring Closure versus Conventional Primary Closure Following Stoma Reversal to Reduce Surgical Site Infection Rate: A Meta-analysis of Randomized Controlled Trials. *Dis Colon Rectum* 2015; **58**: 808-815 [PMID: 26163961 DOI: 10.1097/dcr.0000000000000401]
 - 27 **Aaronson NK**, Ahmedzai S, Bergman B, Bullinger M, Cull A, Duez NJ, Filiberti A, Flechtner H, Fleishman SB, de Haes JC. The European Organization for Research and Treatment of Cancer QLQ-C30: a quality-of-life instrument for use in international clinical trials in oncology. *J Natl Cancer Inst* 1993; **85**: 365-376 [PMID: 8433390 DOI: 10.1093/jnci/85.5.365]
 - 28 **Clavien PA**, Barkun J, de Oliveira ML, Vauthey JN, Dindo D, Schulick RD, de Santibañes E, Pekolj J, Slankamenac K, Bassi C, Graf R, Vonlanthen R, Padbury R, Cameron JL, Makuuchi M. The Clavien-Dindo classification of surgical complications: five-year experience. *Ann Surg* 2009; **250**: 187-196 [PMID: 19638912 DOI: 10.1097/SLA.0b013e3181b13ca2]
 - 29 **Osoba D**, Rodrigues G, Myles J, Zee B, Pater J. Interpreting the significance of changes in health-related quality-of-life scores. *J Clin Oncol* 1998; **16**: 139-144 [PMID: 9440735]
 - 30 **Dray X**, Battaglia G, Wengrower D, Gonzalez P, Carlino A, Camus M, Adar T, Pérez-Roldán F, Marteau P, Repici A. Radiofrequency ablation for the treatment of radiation proctitis. *Endoscopy* 2014; **46**: 970-976 [PMID: 25290097 DOI: 10.1055/s-0034-1377756]
 - 31 **Rustagi T**, Corbett FS, Mashimo H. Treatment of chronic radiation proctopathy with radiofrequency ablation (with video). *Gastrointest Endosc* 2015; **81**: 428-436 [PMID: 24973172 DOI: 10.1016/j.gie.2014.04.038]
 - 32 **Pricolo VE**, Shellito PC. Surgery for radiation injury to the large intestine. Variables influencing outcome. *Dis Colon Rectum* 1994; **37**: 675-684 [PMID: 8026234 DOI: 10.1007/BF02054411]
 - 33 **Yuan ZX**, Ma TH, Zhong QH, Wang HM, Yu XH, Qin QY, Chu LL, Wang L, Wang JP. Novel and Effective Almagate Enema for Hemorrhagic Chronic Radiation Proctitis and Risk Factors for Fistula Development. *Asian Pac J Cancer Prev* 2016; **17**: 631-638

[PMID: 26925655 DOI: 10.7314/APJCP.2016.17.2.631]

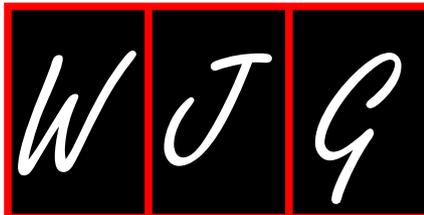
- 34 **Ayerdi J**, Moinuddeen K, Loving A, Wiseman J, Deshmukh N. Diverting loop colostomy for the treatment of refractory gastrointestinal bleeding secondary to radiation proctitis. *Mil Med*

2001; **166**: 1091-1093 [PMID: 11778411]

- 35 **Mäkelä J**, Nevasaari K, Kairaluoma MI. Surgical treatment of intestinal radiation injury. *J Surg Oncol* 1987; **36**: 93-97 [PMID: 3657181 DOI: 10.1002/jso.2930360204]

P- Reviewer: Francois A, Pigo F **S- Editor:** Qi Y **L- Editor:** Kerr C
E- Editor: Wang CH





Observational Study

Clinical study of anesthetization by dezocine combined with propofol for indolent colonoscopy

Bin-Bin Xu, Xiao-Liang Zhao, Gui-Ping Xu

Bin-Bin Xu, Xiao-Liang Zhao, Gui-Ping Xu, Department of Endoscopy Center, People's Hospital of Xinjiang Uygur Autonomous Region, Urumqi 830001, Xinjiang Uygur Autonomous Region, China

Author contributions: Xu BB designed the study; Zhao XL performed the study; Xu BB and Zhao XL contributed the analytical tools and analyzed the data; Xu BB and Xu GP wrote the paper.

Institutional review board statement: The study was reviewed and approved by the People's Hospital of Xinjiang Uygur Autonomous Region Review Board.

Conflict-of-interest statement: The authors declare that they have no conflicts of interest.

Data sharing statement: Technical appendix, statistical code and dataset available from the corresponding author at 56089778@qq.com. Participants have given informed consent for data sharing.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Correspondence to: Dr. Gui-Ping Xu, Department of Endoscopy Center, People's Hospital of Xinjiang Uygur Autonomous Region, No. 91 Tianchi Road, Urumqi 830001, Xinjiang Uygur Autonomous Region, China. xgpsyl@126.com
Telephone: +86-991-8562209
Fax: +86-991-8562209

Received: March 13, 2016

Peer-review started: March 13, 2016

First decision: March 31, 2016

Revised: April 12, 2016

Accepted: May 21, 2016

Article in press: May 23, 2016

Published online: June 28, 2016

Abstract

AIM: To assess the use of dezocine combined with propofol for the anesthetization of patients undergoing indolent colonoscopy.

METHODS: A cross-sectional survey of patients undergoing indolent colonoscopy in the Xinjiang People's Hospital was conducted from April 1 to April 30, 2015. The survey collected patient general information and anesthesia data, including overall medical experience and pain management. Thirty minutes after colonoscopy surgery, samples of venous blood were collected and the biochemical indicators of gastrointestinal function were analyzed.

RESULTS: There were 98 female and 62 male respondents. Indolent colonoscopy was found to be more suitable for mid to older-aged patients. The necessary conditions for the diagnosis of digestive diseases were required in 65 of the 73 inpatients. Adverse reactions to the intraoperative process included two cases of body movement and two cases of respiratory depression. Gastrin and vasoactive intestinal peptide levels were slightly increased. However, somatostatin and endothelin levels were slightly decreased.

CONCLUSION: This study revealed that dezocine combined with propofol can be successfully used for the anesthetization of indolent colonoscopy patients without pain and should be widely used.

Key words: Dezocine; Propofol; Colonoscopy; Patient

assessment; Anesthetization; Cross-sectional

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: A cross-sectional survey of patients undergoing indolent colonoscopy was carried out to study the clinical effect of anesthetization by dezocine combined with propofol. Thirty minutes after colonoscopy surgery, samples of venous blood were collected and the biochemical indicators of gastrointestinal function were analyzed. Indolent colonoscopy was found to be more suitable for mid to older-aged patients. Gastrin and vasoactive intestinal peptide levels were slightly increased. However, somatostatin and endothelin levels were slightly decreased. This study revealed that dezocine combined with propofol can be successfully used for the anesthetization of indolent colonoscopy patients without pain and should be widely used.

Xu BB, Zhao XL, Xu GP. Clinical study of anesthetization by dezocine combined with propofol for indolent colonoscopy. *World J Gastroenterol* 2016; 22(24): 5609-5615 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5609.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5609>

INTRODUCTION

Colonoscopy is an endoscopic examination of the large intestine and the distal part of the small intestine. It provides a diagnosis and therapeutic opportunity for colorectal lesions^[1-4]. At present, colonoscopy remains the gold standard investigation for inspecting colorectal lesions such as adenomas, cancer or inflammation. In many situations, it remains preferable to other imaging examinations such as computed tomography (CT) colonography or barium enema due to the capacity to intervene and sample or remove the pathology encountered.

Unlike sigmoidoscopy which can get to the distal portion (about 600 mm) of the colon, colonoscopy provides an investigation of the full colon (1200-1500 mm)^[5,6]. The pain is not caused by the scope of the insertion but rather by the inflation of the colon to allow the inspection. The scope itself is essentially a long, flexible tube about a centimeter in diameter, *i.e.*, around the diameter of the little finger, which is less than the diameter of an average stool^[7-9].

Increasingly, doctors like to operate on totally anesthetized patients because it allows for a calmer examination without perceived pain or discomfort. Compared with general anesthesia, twilight sedation is safer and it also allows the patient to follow simple commands and even to watch the operation on a monitor. Therefore, twilight sedation is generally best and the operator should not rush, even although the examination may cause discomfort. Tens of millions

of people need colonoscopies annually and yet many of them refuse because of concerns about the procedure^[9]. Propofol is a short-acting, intravenously administered hypnotic/amnestic agent. It is usually used in the induction and maintenance of general anesthesia, sedation for mechanically ventilated adults and procedural sedation. Propofol is also used in veterinary medicine^[9-11].

Recently, propofol has replaced sodium thiopental for the induction of anesthesia in many hospitals because anabiosis is faster and "clearer" using propofol. Propofol is not a pain medication so it may be combined with opioids such as fentanyl to alleviate pain. Whether this is always necessary is unclear.

Propofol has been known as "milk of amnesia" (a play on milk of magnesia) because the intravenous preparation has a milk-like appearance. It is on the World Health Organization Model List of Essential Medicines. More than 50 countries have approved it for use and generic versions are available^[11-15]. Some patients may develop propofol infusion syndrome (PRIS), a rare syndrome that affects patients who undergo long-term treatment with high doses of propofol (> 4 mg/kg per hour for more than 24 h). It can result in cardiac failure, kidney failure, metabolic acidosis and rhabdomyolysis and is often fatal. High blood potassium, high blood triglycerides and liver enlargement, possibly caused by either direct mitochondrial respiratory chain inhibition or impaired mitochondrial fatty acid metabolism, are also key features. Children are more likely to develop PRIS and critically ill patients who receive glucocorticoids and catecholamines are at high risk. The main treatment is supportive therapy^[9,16,17]. Early recognition of the syndrome and discontinuation of the propofol infusion reduces morbidity and mortality.

Dezocine is an opioid analgesic which was first synthesized in 1970. It is a mixed agonist/antagonist of opioid receptors and has the effects of analgesic action and euphoria. However, it is a silent antagonist of the κ -opioid receptor and thus has no side effects such as hallucinations or dysphoria^[18]. Dezocine was patented by the American Home Products Corp. in 1978. Clinical trials ran from 1979 to 1985 before its approval by the United States Food and Drug Administration in 1986. The use of dezocine was discontinued in the United States in 2011 but it is commonly used after surgery in China.

This study analyzed the clinical manifestations after the administration of dezocine combined with propofol in the anesthetization of indolent colonoscopy patients to assess its effects on gastrointestinal function. The incidence of adverse reactions and improvement of patient tolerance were also studied.

A cross-sectional survey was used to collect epidemiological evidence. The use of regularly collected data provides large cross-sectional studies at little or no expense^[19-21], which is an important advantage



Figure 1 Colonoscopy image.

over other styles of epidemiological studies. A natural progression has been suggested from cheap cross-sectional studies of regularly collected data that can put forward hypotheses. However case-control studies test more specific hypotheses, cohort studies and trials cost much more and take much longer but provide stronger evidence. A cross-sectional survey is conducted to examine a specific group to determine whether alcohol consumption is correlated with cirrhosis being investigated. If alcohol is related to cirrhosis, this would support the hypothesis that alcohol may induce cirrhosis.

MATERIALS AND METHODS

General information

After obtaining institutional review board approval and informed consent from the participants or their guardians, 160 consecutive eligible patients previously diagnosed with indolent colonoscopy were enrolled in the study. All the patients had an ASA physical status of I or II and were aged between 1 and 68 years old.

All the experiments were performed under the supervision of the Patients' Experimental Ethics Committee of the Beijing Army General Hospital (Beijing, China). The cross-sectional survey, conducted from April 1 to April 30, 2015, recruited patients undergoing indolent colonoscopy in the Xinjiang People's Hospital. Patients with severe organ damage and drug allergies were excluded. This department does not operate on patients aged over 70 or under 1 year, or with a body mass index over 28 or under 18.

General epidemiological data and a history of neurological anesthetic drugs, including intravenous injection or oral drugs, were recorded.

Anesthesia methods

After identification of the patients, intravenous access to the appropriate area was established. All patients received 5 mg of anisodamine, avoiding microvascular spasm. Baseline hemodynamic data were recorded on arrival in the operating room after placement of routine

monitors.

Anesthesia was induced with an intravenous injection of 5 mg of dezocine. Ten minutes later, 1.5-2.0 mg/kg of propofol was administered intravenously to induce anesthesia. The dose of propofol was adjusted to maintain the heart rate and blood pressure within 20% of the pre-induction values. No opioids were given intraoperatively. All patients received standardized colonoscopy by a practicing physician.

Evaluation index

The intraoperative index included the time the patient took to lose consciousness (from the induction of anesthesia until the eyelash conditioned reflex disappeared) and the total dosage of propofol.

The postoperative index included the length of time from the end of the colonoscopy surgery to the patient waking up and the patient's pain score in the observation room 30 min after surgery. Adverse reactions were investigated during two phases: the incidence of body movements and respiratory depression in the intraoperative phase and the incidence of vomiting and dizziness or headache in the postoperative phase.

Laboratory testing

Samples of venous blood were collected thirty minutes after the colonoscopy surgery. The biochemical indicators gastrin, vasoactive intestinal peptide, somatostatin and endothelin were tested using the radioimmunoassay (RIA) method. The RIA kits were purchased from the Chinese People's Liberation Army General Hospital, RIA Technology Development Center Institute (East Immunity Institute of Technology).

Statistical analysis

All data are represented as the means $x \pm SD$ ($x \pm s$) of three or more independent experiments. Data with a positively skewed distribution were logarithmically transformed into a normal distribution. If the data were homogenous, analysis of variance, the Student-Newman-Keuls test and Pearson's correlation were used. If the data were not homogenous, the Kruskal-Wallis test, Games-Howell test and Spearman's correlation analysis were used. All the analyses were carried out using SPSS17.0 software (SPSS Inc., Chicago, IL, United States). Values below 0.05 were considered to be statistically significant.

RESULTS

Population investigation

From April 1 to April 30, 2015, 160 patients who had undergone dezocine combined with propofol in the anesthetization of indolent colonoscopy took part in the cross-sectional survey. Table 1 summarizes the patients' characteristics. There were 98 females and 62 males; therefore, there were more than 1.6 times

Table 1 Sample characteristics of patients receiving dezocine combined with propofol

Items	Value
Gender	
Male	62
Female	98
Age (yr)	
Median age	43
Maximum age	68
Minimum age	1.3
Average age	48
Modal age	52
Source	
Outpatients	87
Inpatients	73
Feedback from all patients	
Good medical experience	92
Normal medical experience	58
Bad medical experience	9

as many females as males. They ranged from 1 to 68 years of age, with a median of 43, an average of 48 and a mode of 52 years. These figures indicate that indolent colonoscopy was more common among mid to older-aged patients.

There were 87 outpatients and 73 inpatients. For the purpose of health screening, more people chose indolent colonoscopy, especially the older patients. The necessary conditions for a diagnosis of digestive disease were required in 65 of the 73 inpatients (89.04%).

The patients' feedback about their overall medical experience indicated that 92 had a good medical experience, 58 had an average experience, but 9 had a bad experience and 1 had a very bad experience, which was concentrated in the younger age group. Only one child had no statistical significance.

Management survey for operation

According to the American Society of Anesthesiologists (ASA) grading, 89 patients were classified as P1, 62 as P2, and 9 as P3, but no patient was classified as P4, 5 or 6. All diagnoses met the standard clinical symptoms of the ASA.

The ASA physical status class risk stratification system is dependent on comorbid conditions that are a threat to life or limit activity, thus helping to predicting preoperative risks. They are as follows: (1) P1 - normal healthy patient; (2) P2 - patient with mild systemic disease; (3) P3 - patient with severe systemic disease; (4) P4 - patient with severe systemic disease that is a constant threat to life; (5) P5 - moribund patient who is not expected to survive without the operation; and (6) P6 - declared brain-dead patient whose organs are being removed for donor purposes. All the patients were assessed by the anesthetist before surgery and not by self-reporting.

Among the 160 anesthetized patients, two episodes of postoperative emesis were recorded which

Table 2 Management survey for operations conducted in this study

	Value
Anesthesiologists' (ASA) Grading	
P 1	89
P 2	62
P 3	9
P 4, 5 or 6	0
Adverse reactions	
Body movement	2
Respiratory depression	2
Vomiting	2
Dizziness	3
Exhaustion and fatigue	12
Pain management	
Level 0-3	132
Level 4-6	28
Above level 7-10	0

occurred after propofol anesthesia. No patient required additional medication for pain in the post-anesthesia care or normal unit. The anesthesia experience perceived by the parents of the children was more appropriate for dezocine combined with propofol. However, three patients reported dizziness 30 min after the operation.

Intraoperative adverse reactions included two cases of body movement and two cases of respiratory depression. The assessment and pain management methods were based on the World Health Organization classification. One hundred and thirty-two patients reported a pain level under 3. No patients reported a higher level of pain; however, 12 patients reported feeling exhausted and fatigued. These results are shown in Table 2.

Length of operation

The mean duration of surgery was 52 ± 24 min, the induction time was 10 ± 4 min and the duration of maintenance was 60 ± 23 min.

Laboratory results

Thirty minutes after the colonoscopy surgery, samples of venous blood were collected and four meaningful biochemical indicators of gastrointestinal function were assessed, as shown in Table 3. The results showed that gastrin and vasoactive intestinal peptide levels were slightly increased, whereas somatostatin and endothelin levels were slightly decreased.

Gastrin can increase gastric mucosal blood flow and protect the gastric mucosa. Vasoactive intestinal peptide can cause the relaxation of the pyloric sphincter. The decrease in somatostatin and endothelin levels helped to inhibit the relaxation of the smooth gastric muscles.

DISCUSSION

General anesthesia refers to a medically induced coma

Table 3 Biochemical indicators of gastrointestinal function

	Detection value	Reference value	Comparison
Gastrin (μg/mL)	88.94 ± 18.77	63.12 ± 28.71	Up
Vasoactive intestinal peptide (pg/mL)	58.33 ± 4.22	35.25 ± 3.12	Up
Somatostatin (pg/mL)	79.42 ± 4.26	108.25 ± 5.12	Down
Endothelin (pg/mL)	36.77 ± 9.12	48.91 ± 10.1	Down

with the lack of protective reflexes caused by one or more general anesthetic agents^[1,12,21]. Various kinds of medications may be administered, with the aim of amnesia, analgesia, ensuring unconsciousness, relaxing the skeletal muscles and loss of control of the autonomic nervous system reflexes^[9]. The optimal combination of these agents for any given patient and operation is usually chosen by the anesthesiologist or another provider, such as an anesthesiology assistant or nurse anesthetist, in consultation with the patient and the doctor conducting the operation.

Colonoscopy is the standard procedure for the diagnosis, screening, treatment and follow-up of many colorectal diseases^[22,23]. Although some patients can tolerate the colonoscopy procedure without sedation or analgesics, the use of drugs in some patients is associated with stress. There are difficulties in determining an optimal dose for sedation and monitoring patients adequately during the procedure^[24-27]. Many patients require intravenous benzodiazepines and opiates, which are associated with amnesic, anxiolytic and sedative properties. Combined administration of benzodiazepines and opioids has several undesirable effects, including a delay of several minutes from the time of injection before the drugs exert their effects, amnesia and the risk of respiratory depression^[7,28,29].

Hospitals increasingly prefer to use fospropofol for colonoscopy. Fospropofol (trade name Lusedra) is an intravenous sedative-hypnotic agent. It is currently used for the sedation of adult patients undergoing diagnostic or therapeutic operations such as endoscopy. Some water-soluble prodrugs of propofol have been developed, of which fospropofol is known as the most fit for clinical development to date. Fospropofol is often administered combined with an opioid such as fentanyl. As a prodrug of propofol, it is metabolized into propofol by alkaline phosphatases. Theoretically, one millimole (mmol) of propofol may be generated for each mmol of fospropofol sodium administered; 1.86 mg of fospropofol sodium is the molar equivalent of 1 mg propofol^[30-33].

Dezocine can be administered intravenously or intramuscularly. Dezocine is an effective painkiller compared with meperidine (pethidine) and a more effective analgesic than pentazocine but may cause greater respiratory depression. It is an effective drug for the treatment of pain but side effects such as

dizziness restrict its clinical use and it also causes opioid withdrawal syndrome in patients already using other opioids^[34].

A study conducted by the Department of Anesthesia, Xishan People's Hospital of Wuxi City in Jiangsu province explored the clinical effect of dezocine combined with propofol in the anesthetization of 60 patients undergoing indolent enteroscopy between July 2012 and June 2014. The patients were randomly divided into a research group that received dezocine combined with propofol and a control group that received fentanyl combined with propofol^[33-36]. The total dosage of propofol in the research group was less than that in the control group ($P < 0.05$), the awakening time in the research group was shorter than in the control group ($P < 0.05$) and the number of adverse effects during and after surgery was lower in the research group than in the control group ($P < 0.05$). Dezocine combined with propofol applied in the anesthetization of indolent enteroscopy can have a remarkable effect, improve operational safety and decrease the occurrence of adverse reactions. Our results were consistent with those of this study.

According to the results of a study by the Department of Anesthesiology at the First Hospital of Quanzhou, age was an important influence on the pharmacodynamics of propofol in patients receiving propofol combined with dezocine while undergoing colonoscopy^[37]. However, the results from the Department of Gastroenterology at the Second People's Hospital of Fujian province showed that endoscopic mucosal resection under painless colonoscopy shortened the recovery time, increased the success rate and improved the satisfaction rate of older patients who received propofol and fentanyl. Endoscopic mucosal resection under painless colonoscopy is an easy, safe and effective therapy for colorectal polyps.

Thirty minutes after colonoscopy surgery, samples of venous blood were collected and the biochemical indicators gastrin, vasoactive intestinal peptide, somatostatin and endothelin were tested. There was an increase in the plasma contents of gastrin and vasoactive intestinal peptide ($P < 0.05$) and a decrease in somatostatin and endothelin ($P < 0.05$) compared with the standard reference values^[38-40].

Anesthesia with dezocine combined with propofol evidently increases gastric mucosal blood flow, suggesting that it has a regulative effect on gastrointestinal function. The use of dezocine combined with propofol might change plasma brain-gut peptides levels and may be useful.

COMMENTS

Background

Increasingly, doctors like to operate on totally anesthetized patients because it allows for a calmer examination without perceived pain or discomfort. Twilight sedation is safer than general anesthesia and allows the patient to follow simple commands and even to watch the operation on a monitor. Therefore,

twilight sedation is generally best and the operator should not rush, even although the examination may cause discomfort. Tens of millions of adults need colonoscopies annually and yet many refuse because of concerns about the procedure.

Research frontiers

This study assessed the use of dezocine combined with propofol for the anesthetization of patients undergoing indolent colonoscopy for the assessment of gastrointestinal function. A cross-sectional survey of patients undergoing indolent colonoscopy in the Xinjiang People's Hospital was conducted from April 1 to April 30, 2015. The survey collected general information and anesthesia data, including overall medical experience and pain management.

Innovations and breakthroughs

Indolent colonoscopy was found to be more suitable for mid to older-aged patients. Somatostatin and endothelin levels were slightly decreased and patients had a good medical experience.

Applications

Gastrin and vasoactive intestinal peptide levels were slightly increased. However, somatostatin and endothelin levels were slightly decreased. This clinical study of dezocine combined with propofol indicates that it is a successful method for the anesthetization of indolent colonoscopy patients without pain and should therefore be widely used.

Peer-review

In this study, the authors assessed the use of dezocine combined with propofol for the anesthetization of patients undergoing indolent colonoscopy for the assessment of gastrointestinal function. The study collected general information and anesthesia data, including overall medical experience and pain management. This clinical study of dezocine combined with propofol indicates that it is a successful method for the anesthetization of indolent colonoscopy patients without pain and should therefore be widely used.

REFERENCES

- 1 **Triantafyllidis JK**, Merikas E, Nikolakis D, Papalois AE. Sedation in gastrointestinal endoscopy: current issues. *World J Gastroenterol* 2013; **19**: 463-481 [PMID: 23382625 DOI: 10.3748/wjg.v19.i4.463]
- 2 **Gasparović S**, Rustemović N, Opacić M, Premuzić M, Korusić A, Božikov J, Bates T. Clinical analysis of propofol deep sedation for 1,104 patients undergoing gastrointestinal endoscopic procedures: a three year prospective study. *World J Gastroenterol* 2006; **12**: 327-330 [PMID: 16482639 DOI: 10.3748/wjg.v12.i2.327]
- 3 **Ijspeert JE**, Bevan R, Senore C, Kaminski MF, Kuipers EJ, Mroz A, Bessa X, Cassoni P, Hassan C, Repici A, Balaguer F, Rees CJ, Dekker E. Detection rate of serrated polyps and serrated polyposis syndrome in colorectal cancer screening cohorts: a European overview. *Gut* 2016; Epub ahead of print [PMID: 26911398 DOI: 10.1136/gutjnl-2015-310784]
- 4 **Repici A**, Pagano N, Hassan C, Carlino A, Rando G, Strangio G, Romeo F, Zullo A, Ferrara E, Vitetta E, Ferreira Dde P, Danese S, Arosio M, Malesci A. Balanced propofol sedation administered by nonanesthesiologists: The first Italian experience. *World J Gastroenterol* 2011; **17**: 3818-3823 [PMID: 21987624 DOI: 10.3748/wjg.v17.i33.3818]
- 5 **Davila RE**, Rajan E, Adler DG, Egan J, Hirota WK, Leighton JA, Qureshi W, Zuckerman MJ, Fanelli R, Wheeler-Harbaugh J, Baron TH, Faigel DO. ASGE Guideline: the role of endoscopy in the patient with lower-GI bleeding. *Gastrointest Endosc* 2005; **62**: 656-660 [PMID: 16246674 DOI: 10.1016/j.gie.2005.07.032]
- 6 **Metwally M**, Agresti N, Hale WB, Ciofoaia V, O'Connor R, Wallace MB, Fine J, Wang Y, Gross SA. Conscious or unconscious: the impact of sedation choice on colon adenoma detection. *World J Gastroenterol* 2011; **17**: 3912-3915 [PMID: 22025879 DOI: 10.3748/wjg.v17.i34.3912]
- 7 **Zauber AG**, Winawer SJ, O'Brien MJ, Lansdorp-Vogelaar I, van

- 8 **Ballegooijen M**, Hankey BF, Shi W, Bond JH, Schapiro M, Panish JF, Stewart ET, Wayne JD. Colonoscopic polypectomy and long-term prevention of colorectal-cancer deaths. *N Engl J Med* 2012; **366**: 687-696 [PMID: 22356322 DOI: 10.1056/NEJMoa1100370]
- 9 **Hamdani U**, Naeem R, Haider F, Bansal P, Komar M, Diehl DL, Kirchner HL. Risk factors for colonoscopic perforation: a population-based study of 80118 cases. *World J Gastroenterol* 2013; **19**: 3596-3601 [PMID: 23801860 DOI: 10.3748/wjg.v19.i23.3596]
- 10 **Eisen GM**, Dominitz JA, Faigel DO, Goldstein JL, Kalloo AN, Petersen JL, Raddawi HM, Ryan ME, Vargo JJ, Young HS, Fanelli RD, Hyman NH, Wheeler-Harbaugh J. Endoscopic therapy of anorectal disorders. *Gastrointest Endosc* 2001; **53**: 867-870 [PMID: 11375620 DOI: 10.1016/S0016-5107(01)70308-2]
- 11 **Allen M**, Leslie K, Hebbard G, Jones I, Mettho T, Maruff P. A randomized controlled trial of light versus deep propofol sedation for elective outpatient colonoscopy: recall, procedural conditions, and recovery. *Can J Anaesth* 2015; **62**: 1169-1178 [PMID: 26335904 DOI: 10.1007/s12630-015-0463-3]
- 12 **Peytremann-Bridevaux I**, Arditi C, Froehlich F, O'Malley J, Fairclough P, Le Moine O, Dubois RW, Gonvers JJ, Schusselé Fillietaz S, Vader JP, Juillerat P, Pittet V, Burnand B. Appropriateness of colonoscopy in Europe (EPAGE II). Iron-deficiency anemia and hematochezia. *Endoscopy* 2009; **41**: 227-233 [PMID: 19280534 DOI: 10.1055/s-0028-1119644]
- 13 **Uezono S**, Goto T, Terui K, Ichinose F, Ishiguro Y, Nakata Y, Morita S. Emergence agitation after sevoflurane versus propofol in pediatric patients. *Anesth Analg* 2000; **91**: 563-566 [PMID: 10960377 DOI: 10.1213/00000539-200009000-00012]
- 14 **Savides TJ**, Jensen DM. Therapeutic endoscopy for nonvariceal gastrointestinal bleeding. *Gastroenterol Clin North Am* 2000; **29**: 465-487, vii [PMID: 10836190 DOI: 10.1016/S0889-8553(05)70123-0]
- 15 **Iwasaki J**, Sano Y, Fu KI, Machida A, Okuno T, Kuwamura H, Yoshino T, Mera K, Kato S, Ohtsu A, Yoshida S, Fujii T. Depressed-type (0-IIc) colorectal neoplasm in patients with family history of first-degree relatives with colorectal cancer: A cross-sectional study. *World J Gastroenterol* 2006; **12**: 3082-3087 [PMID: 16718792 DOI: 10.3748/wjg.v12.i19.3082]
- 16 **Lin YL**, Chiang JK, Lin SM, Tseng CE. Helicobacter pylori infection concomitant with metabolic syndrome further increase risk of colorectal adenomas. *World J Gastroenterol* 2010; **16**: 3841-3846 [PMID: 20698048 DOI: 10.3748/wjg.v16.i30.3841]
- 17 **Spinzi G**, Fante MD, Masci E, Buffoli F, Colombo E, Fiori G, Ravelli P, Ceretti E, Minoli G. Lack of colonic neoplastic lesions in patients under 50 yr of age with hematochezia: a multicenter prospective study. *Am J Gastroenterol* 2007; **102**: 2011-2015 [PMID: 17521401 DOI: 10.1111/j.1572-0241.2007.01332.x]
- 18 **Horiuchi A**, Nakayama Y, Kajiyama M, Kato N, Kamijima T, Ichise Y, Tanaka N. Safety and effectiveness of propofol sedation during and after outpatient colonoscopy. *World J Gastroenterol* 2012; **18**: 3420-3425 [PMID: 22807612 DOI: 10.3748/wjg.v18.i26.3420]
- 19 **Bae T**, Ha Y, Kim C, Lee J, Ha K, Shin S, Lee Y, Kang Y. Distribution of the colonoscopic adenoma detection rate according to age: is recommending colonoscopy screening for Koreans over the age of 50 safe? *Ann Coloproctol* 2015; **31**: 46-51 [PMID: 25960971 DOI: 10.3393/ac.2015.31.2.46]
- 20 **Ahn E**, Son KY, Shin DW, Han MK, Lee H, An AR, Kim EH, Cho B. Perceived risk as a barrier to appropriate diagnosis of irritable bowel syndrome. *World J Gastroenterol* 2014; **20**: 18360-18366 [PMID: 25561803 DOI: 10.3748/wjg.v20.i48.18360]
- 21 **Bjerregaard NC**, Tøttrup A, Sørensen HT, Laurberg S. Diagnostic value of self-reported symptoms in Danish outpatients referred with symptoms consistent with colorectal cancer. *Colorectal Dis* 2007; **9**: 443-451 [PMID: 17504342 DOI: 10.1111/j.1463-1318.2006.01170.x]
- 22 **Aydogmus MT**, Türk HS, Oba S, Gokalp O. A comparison of different proportions of a ketamine-propofol mixture administered in a single injection for patients undergoing colonoscopy. *Arch Med Sci* 2015; **11**: 570-576 [PMID: 26170850 DOI: 10.5114/

- aoms.2015.52360]
- 22 **Xie WQ**, Li YY, Zie WJ, Yan JJ, Jiang CC, Kang ZM. Effect of patient's age on propofol's pharmacodynamics while combining with low dose fentanyl in colonoscopy. *Linchuang Mazuixue Zazhi* 2014; **30**: 462-465
 - 23 **Pasha SF**, Shergill A, Acosta RD, Chandrasekhara V, Chathadi KV, Early D, Evans JA, Fisher D, Fonkalsrud L, Hwang JH, Khashab MA, Lightdale JR, Muthusamy VR, Saltzman JR, Cash BD. The role of endoscopy in the patient with lower GI bleeding. *Gastrointest Endosc* 2014; **79**: 875-885 [PMID: 24703084 DOI: 10.1016/j.gie.2013.10.039]
 - 24 **Heuss LT**, Sugandha SP, Beglinger C. Carbon dioxide accumulation during analgesedated colonoscopy: comparison of propofol and midazolam. *World J Gastroenterol* 2012; **18**: 5389-5396 [PMID: 23082055 DOI: 10.3748/wjg.v18.i38.5389]
 - 25 **Tong GX**, Chai J, Cheng J, Xia Y, Feng R, Zhang L, Wang DB. Diagnostic value of rectal bleeding in predicting colorectal cancer: a systematic review. *Asian Pac J Cancer Prev* 2014; **15**: 1015-1021 [PMID: 24568444 DOI: 10.7314/APJCP.2014.15.2.1015]
 - 26 **Sun XB**. Clinical study of dezocien combined with propofol in the anesthetization of indolent enteroscope. *Zhongguo Dangdai Yixue* 2015; **22**: 124-126
 - 27 **Choe EK**, Kim D, Kim HJ, Park KJ. Association of visceral obesity and early colorectal neoplasia. *World J Gastroenterol* 2013; **19**: 8349-8356 [PMID: 24363527 DOI: 10.3748/wjg.v19.i45.8349]
 - 28 **Lozano-Maya M**, Ponferrada-Díaz A, González-Asanza C, Nogales-Rincón O, Senent-Sánchez C, Pérez-de-Ayala V, Jiménez-Aleixandre P, Cos-Arregui E, Menchén-Fernández-Pacheco P. Usefulness of colonoscopy in ischemic colitis. *Rev Esp Enferm Dig* 2010; **102**: 478-483 [PMID: 20670068 DOI: 10.4321/S1130-01082010000800004]
 - 29 **Gan T**, Yang JL, Wu JC, Wang YP, Yang L. When and why a colonoscopist should discontinue colonoscopy by himself? *World J Gastroenterol* 2015; **21**: 7834-7841 [PMID: 26167083 DOI: 10.3748/wjg.v21.i25.7834]
 - 30 **Leggett B**, Whitehall V. Role of the serrated pathway in colorectal cancer pathogenesis. *Gastroenterology* 2010; **138**: 2088-2100 [PMID: 20420948 DOI: 10.1053/j.gastro.2009.12.066]
 - 31 **Fanti L**, Gemma M, Agostoni M, Rossi G, Ruggeri L, Azzolini ML, Dabizzi E, Beretta L, Testoni PA. Target Controlled Infusion for non-anaesthesiologist propofol sedation during gastrointestinal endoscopy: The first double blind randomized controlled trial. *Dig Liver Dis* 2015; **47**: 566-571 [PMID: 25840875 DOI: 10.1016/j.dld.2015.03.003]
 - 32 **Klare P**, Huth R, Haller B, Huth M, Weber A, Schlag C, Reindl W, Schmid RM, von Delius S. Patient position and hypoxemia during propofol sedation for colonoscopy: a randomized trial. *Endoscopy* 2015; **47**: 1159-1166 [PMID: 26126161 DOI: 10.1055/s-0034-1392329]
 - 33 **Nikpour S**, Ali Asgari A. Colonoscopic evaluation of minimal rectal bleeding in average-risk patients for colorectal cancer. *World J Gastroenterol* 2008; **14**: 6536-6540 [PMID: 19030208 DOI: 10.3748/wjg.14.6536]
 - 34 **Jass JR**. Classification of colorectal cancer based on correlation of clinical, morphological and molecular features. *Histopathology* 2007; **50**: 113-130 [PMID: 17204026 DOI: 10.1111/j.1365-2559.2006.02549.x]
 - 35 **Baykal Tural Z**, Gulec H, Dereli N, Babayigit M, Kurtay A, Inceoz H, Horasanlı E. Propofol-ketamine combination: a choice with less complications and better hemodynamic stability compared to propofol? On a prospective study in a group of colonoscopy patients. *Ir J Med Sci* 2015; Epub ahead of print [PMID: 26329313 DOI: 10.1007/s11845-015-1348-8]
 - 36 **Weng HF**, Hu JF, Yang JY, Wang QY, Wan LT. Analysis of complications of painless gastrointestinal endoscopy in the diagnosis and treatment. *Chendou Yixueyuan Xuebao* 2015; **10**: 324-326
 - 37 **Sun DY**, Huang YX, Gao W, Chu ZH, Wang QL. Effects of electroacupuncture on plasma levels of brain-gut peptides in dogs. *Jiefangjun Yixue Zazhi* 2002; **27**: 997-998
 - 38 **Goudra B**, Singh PM, Gouda G, Borle A, Carlin A, Yadwad A. Propofol and non-propofol based sedation for outpatient colonoscopy-prospective comparison of depth of sedation using an EEG based SEDLine monitor. *J Clin Monit Comput* 2015; Epub ahead of print [PMID: 26364193 DOI: 10.1007/s10877-015-9769-5]
 - 39 **Kim YW**, Choi H, Kim GJ, Ryu SJ, Park SM, Kim JS, Ji JS, Kim BW, Lee BI, Choi MG. [Role of Colonoscopy in Patients with Hematochezia]. *Korean J Gastroenterol* 2016; **67**: 87-91 [PMID: 26907484 DOI: 10.4166/kjg.2016.67.2.87]
 - 40 **Rees CJ**, Ngu WS. Temporal trends and variability of colonoscopy performance in gastroenterology practice. *Endoscopy* 2016; **48**: 213-214 [PMID: 26906313 DOI: 10.1055/s-0042-102054]

P- Reviewer: Arville B, Shimizu Y **S- Editor:** Gong ZM

L- Editor: Roemmele A **E- Editor:** Ma S



Successful treatment of ileal ulcers caused by immunosuppressants in two organ transplant recipients

Yun-Wei Guo, Hua-Ying Gu, Kodjo-Kunale Abassa, Xian-Yi Lin, Xiu-Qing Wei

Yun-Wei Guo, Hua-Ying Gu, Kodjo-Kunale Abassa, Xian-Yi Lin, Xiu-Qing Wei, Department of Gastroenterology, the Third Affiliated Hospital of Sun Yat-Sen University, Guangzhou 510630, Guangdong Province, China

Author contributions: Guo YW and Gu HY contributed equally to this work; Guo YW, Gu HY, Abassa KK, Lin XY, and Wei XQ analyzed the data and diagnosed and treated the patient; Guo YW, Gu HY, Abassa KK and Wei XQ wrote the paper.

Supported by Guangdong Science and Technology Program, No. 2012B061700072.

Institutional review board statement: The study was reviewed and approved by the Institutional Review Board of the Third Affiliated Hospital of Sun Yat-Sen University.

Informed consent statement: The patients reported in this article provided informed written consent prior to manuscript preparation.

Conflict-of-interest statement: There was no conflict of interest in this work.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Xiu-Qing Wei, MD, PhD, Professor, Department of Gastroenterology, the Third Affiliated Hospital of Sun Yat-Sen University, No. 600, Tianhe Road, Tianhe District, Guangzhou 510630, Guangdong Province, China. wei-xiuqing@163.com
Telephone: +86-20-85253095
Fax: +86-20-85253336

Received: March 3, 2016
Peer-review started: March 4, 2016
First decision: April 1, 2016

Revised: April 10, 2016

Accepted: April 20, 2016

Article in press: April 20, 2016

Published online: June 28, 2016

Abstract

Although gastroduodenal ulcers are common in solid organ transplant patients, there are few reports on multiple giant ulcers in the distal ileum and ileocecal valve caused by immunosuppressants. Herein, we report on a liver transplant recipient and a renal transplant recipient with multiple large ulcers in the distal ileum and ileocecal valve who rapidly achieved ulcer healing upon withdrawal of sirolimus or tacrolimus and administration of thalidomide. In case 1, a 56-year-old man with primary hepatocellular carcinoma had received a liver transplantation. Tacrolimus combined with sirolimus and prednisolone was used as the anti-rejection regimen. Colonoscopy was performed because of severe abdominal pain and diarrhea at post-operative month 10. Multiple giant ulcers were found at the ileocecal valve and distal ileum. The ulcers healed rapidly with withdrawal of sirolimus and treatment with thalidomide. There was no recurrence during 2 years of follow-up. In case 2, a 34-year-old man with end-stage kidney disease received kidney transplantation and was put on tacrolimus combined with mycophenolate mofetil and prednisolone as the anti-rejection regimen. Twelve weeks after the operation, the patient presented with hematochezia and severe anemia. Colonoscopy revealed multiple large ulcers in the ileocecal valve and distal ileum, with massive accumulation of fresh blood. The bleeding ceased after treatment with intravenous somatostatin and oral thalidomide. Tacrolimus was withdrawn at the same time. Colonoscopy at week 4 of follow-up revealed remarkable healing of the ulcers, and there was no recurrence of bleeding during 1 year of follow-up. No lymphoma, tuberculosis, or infection of cytomegalovirus, Epstein-Barr virus, or fungus was

found in either patient. In post-transplantation cases with ulcers in the distal ileum and ileocecal valve, sirolimus or tacrolimus should be considered a possible risk factor, and withdrawing them or switching to another immunosuppressant might be effective to treat these ulcers.

Key words: Ileal ulcers; Liver transplantation; Kidney transplantation; Sirolimus; Tacrolimus

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: There are few reports available on ileal ulcers caused by immunosuppressants. Herein, we report a liver transplant recipient and a renal transplant recipient who had multiple large ulcers in the distal ileum and ileocecal valve. Ulcers rapidly healed after withdrawal of sirolimus or tacrolimus and administration of thalidomide. No lymphoma, tuberculosis, or infection of cytomegalovirus, Epstein-Barr virus, or fungus was found in either patient. There was no recurrence of ulcers or organ rejection. In some post-transplantation cases with ileal ulcers, sirolimus or tacrolimus should be considered as a risk factor because of their inhibitory effects on wound healing. Withdrawing them or switching to other immunosuppressants might be effective.

Guo YW, Gu HY, Abassa KK, Lin XY, Wei XQ. Successful treatment of ileal ulcers caused by immunosuppressants in two organ transplant recipients. *World J Gastroenterol* 2016; 22(24): 5616-5622 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i24/5616.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i24.5616>

INTRODUCTION

Solid organ transplant recipients are susceptible to a variety of gastrointestinal (GI) complications, one of which is ulcer disease^[1-5]. Most of these ulcers are located at the anastomotic stoma or gastroduodenum; ulcers at the intestine or colon are rare. In addition to *Helicobacter pylori* infection and ischemia, infection with cytomegalovirus (CMV), Epstein-Barr virus (EBV), mycobacteria, and fungus can contribute to ulcer disease in transplant recipients^[1-5]. A few reports suggest that the use of immunosuppressants, especially high-dose immunosuppression after transplantation, might correlate with impairment of the gastrointestinal tract^[6,7]. Herein, we report a liver transplant recipient and a renal transplant recipient with multiple large ulcers in the distal ileum and ileocecal valve who rapidly achieved ulcers healing upon withdrawal of sirolimus or tacrolimus and administration of thalidomide.

CASE REPORT

Case 1

A 56-year-old man with primary hepatocellular carcinoma received orthotopic liver transplantation in our hospital. Prior to the transplantation, he was diagnosed with chronic hepatitis B virus (HBV), Child-C liver function, and elevated blood glucose. No history of renal or cardiac disease or mycobacterium tuberculosis (TB) infection was noted. He denied smoking and drinking. Tacrolimus combined with sirolimus and prednisolone was used as the anti-rejection regimen. The dose of tacrolimus and sirolimus was adjusted according to the drug serum concentration. The patient recovered and liver function improved to normal level. Entecavir was prescribed to prevent HBV reinfection.

After administration of the immunosuppressants, the patient began to develop mild peri-umbilical pain and diarrhea, which were tolerable at the time. Probiotics and antispasmodic treatment seemed not so effective. At 10 mo post-operation, he was admitted to our hospital because of severe diarrhea and abdominal pain. There were 7-10 bowel movements per day, with mucous blood stool. On physical examination, blood pressure, heart rate, and temperature were normal. There was slight tenderness on the peri-umbilicus area without rebound tenderness. Complete blood count revealed a leukocyte count of $11.50 \times 10^9/L$, erythrocyte count of $4.50 \times 10^{12}/L$, hemoglobin level of 121 g/L, platelet count of $295 \times 10^9/L$, neutrophils of 76.4%, and lymphocytes of 16.3%. Leukocytes and erythrocytes were found in the stool. The ratio of coccus to bacillus in stool was 1:9. Serum glutamic-oxaloacetic transaminase, glutamic-pyruvic transaminase, total bilirubin, BUN, creatine, and electrolytes were within normal range. Serum 1-3-β-D dextran was < 10 pg/mL. Blood and stool cultures for fungus and bacteria showed no growth. Chest computed tomography (CT) scan revealed no lesions and serum T-SPOT.TB was negative. Immunoglobulin (Ig)M and IgG of the EB virus were negative. Abdominal enhanced CT scan showed thickening of the distal ileum and ileocecal region, suggesting inflammatory lesions. The spleen was slightly enlarged, muddy stones were found in the common bile duct, and no enlarged lymph node was noted. Chronic erosive gastritis with negative *Helicobacter pylori* infection was confirmed through esophagogastroduodenoscopy. Colonoscopy revealed multiple giant and deep ulcers in the ileocecal valve and distal ileum, with polypoid hyperplasia. The length of the largest ulcer was up to 5.0 cm (Figure 1A-C). Histopathology of biopsy specimens revealed benign ulcers and chronic inflammation with non-caseous granulomas (Figure 2A and B), without signs of fungus and parasites infection. The immunohistochemical study was negative for CMV infection. EBV encoded

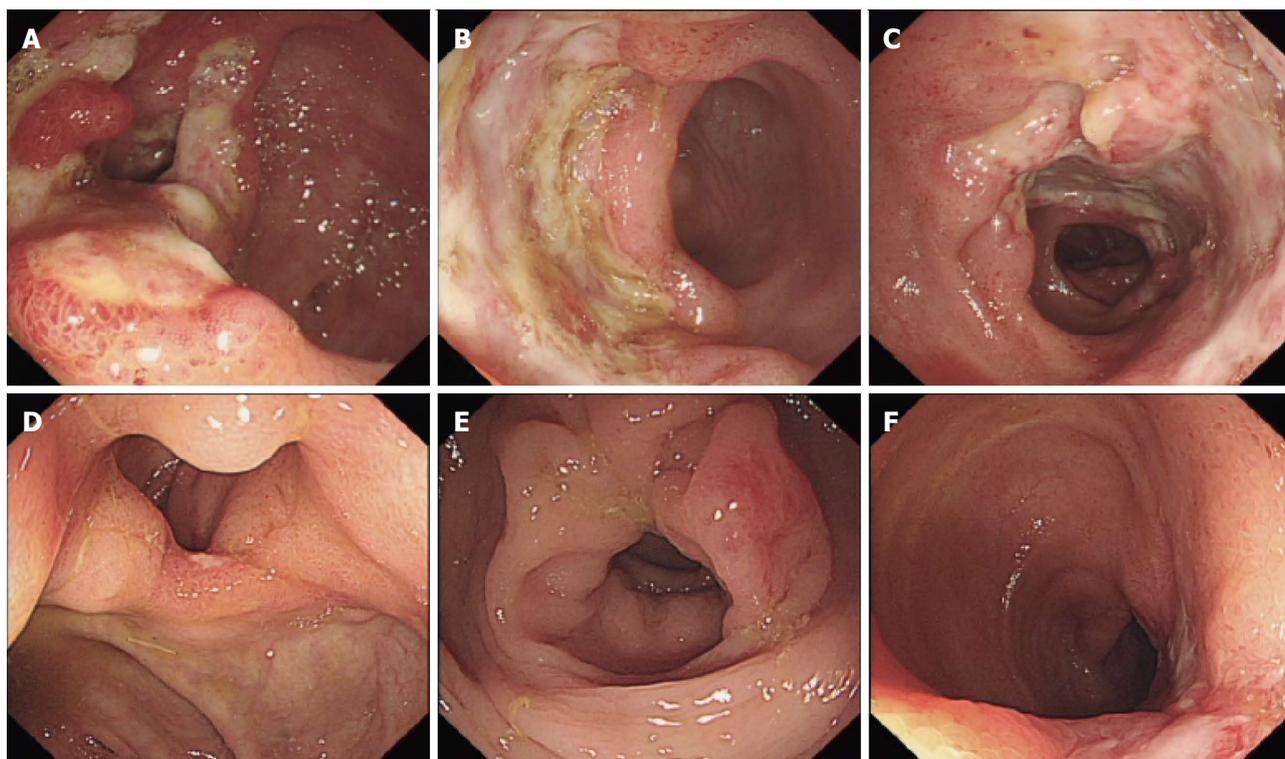


Figure 1 Colonoscopic images of case one. A-C: Multiple giant and deep ulcers in the ileocecal valve and distal ileum, with polypoid hyperplasia; D-F: Rapid healing of the ulcers in the ileocecal valve and distal ileum and only two healing 2 stage ulcers left.

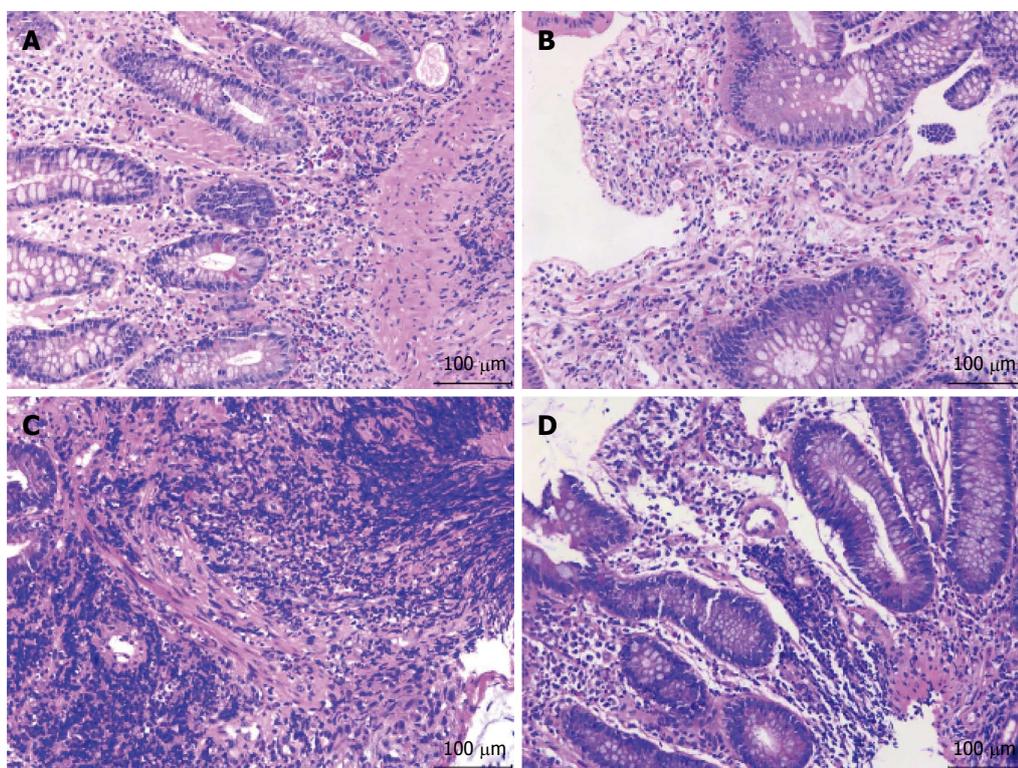


Figure 2 Photomicrograph of biopsy specimens. A and B: Biopsy specimens of ulcers from case one; C and D: Biopsy specimens of ulcers from case two. Hematoxylin-eosin staining, magnification $\times 200$.

early small RNA (EBER) was negative by *in situ* hybridization.

Considering sirolimus to have more gastrointestinal complications than tacrolimus in clinical application,

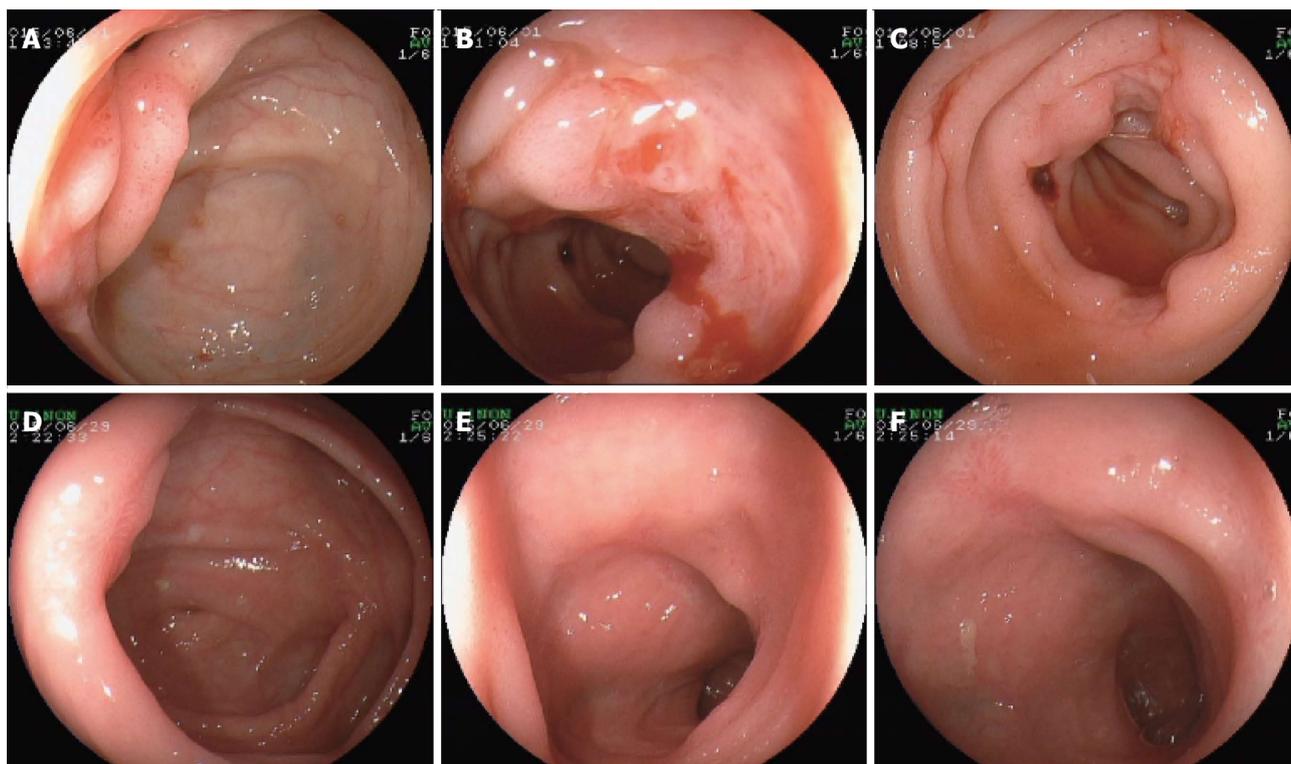


Figure 3 Multiple ulcers in ileocecal valve and distal ileum, with massive fresh blood accumulation (A-C); rapid healing of the ulcers with scar tissue in ileocecal valve and distal ileum (D-F).

sirolimus was withdrawn first. The patient was also put on oral thalidomide at a dose of 100 mg/d for 2 wk and intravenous antibiotics for one week. Diarrhea and abdominal pain were gradually relieved and subsided. Colonoscopy at week 6 of follow-up revealed remarkable healing of the ulcers in the ileocecal valve and distal ileum, and only two healing 2 stage ulcers were found (Figure 1D-F). No organ rejection was noted after withdrawing sirolimus. No recurrence of diarrhea and abdominal pain was noted during the 2 years of follow-up.

Case 2

A 34-year-old man with end-stage kidney disease was admitted to the department of renal transplantation for living-donor kidney transplantation. Except for kidney disease, he had no history of primary liver, heart, or head disease. He denied smoking and drinking. Tacrolimus combined with mycophenolate mofetil and prednisolone was applied as the anti-rejection regimen. The dose of tacrolimus was adjusted according to the drug serum concentration. At the same time, oral ganciclovir, voriconazole, and esomeprazole were used to prevent CMV and fungus infection and esophagogastroduodenal complications. The patient recovered smoothly. Serum creatinine level decreased to 152.0 $\mu\text{mol/L}$, and urine output was normal.

At postoperative week 12, the patient was admitted to our department because of repeated hematochezia for 2 wk, which was accompanied by

dizziness and weakness. No abdominal pain, nausea, or vomiting was observed. On physical examination, blood pressure, heart rate, and temperature were 95/55 mmHg, 102 beats/min, 37.5 $^{\circ}\text{C}$, respectively. There was mild tenderness at the peri-umbilicus area and the lower right abdomen, without rebound tenderness. Complete blood count revealed a leukocyte count of $7.45 \times 10^9/\text{L}$, erythrocyte count of $2.05 \times 10^{12}/\text{L}$, hemoglobin level of 57 g/L, and platelet count of $241 \times 10^9/\text{L}$. Mild elevated serum creatinine level of 168.0 $\mu\text{mol/L}$ was noted, with a normal BUN level of 6.8 mmol/L. Prothrombin time, apartprothrombin time, and thrombin time were within the normal range. Laboratory indices about hepatic, cardiac, and respiratory function were all normal. Blood and stool cultures for fungus and bacteria showed no growth. Chest CT scan revealed some pulmonary lesions of previous tuberculosis, and serum T-SPOT.TB was negative. There were no abnormal findings in the abdominal doppler ultrasound. Esophagogastroduodenoscopy was performed first, and only mild gastritis was observed, with negative rapid urease test for *Helicobacter pylori* infection. Colonoscopy revealed multiple ulcers in the ileocecal valve and distal ileum, with massive accumulation of fresh blood. These ulcers were oval and deep, covered with white fur or a blood scab, and the biggest had a diameter of 2.0 cm (Figure 3A-C). Histopathology revealed chronic inflammation with a large number of lymphocytes infiltration (Figure 2C

and D), without signs of fungus or parasite infection. Immunohistochemistry stain for CMV was negative. EBER was also negative by *in situ* hybridization.

The bleeding lessened and eventually ceased after treatment with intravenous somatostatin (1.2 mg/d) and oral thalidomide (100 mg/d) for 5 d. At the same time, because the multiple intestinal ulcers may be due to immunosuppressors, tacrolimus was withdrawn, and cyclosporine combined with mycophenolate mofetil and prednisolone were administered. Additionally, the patient was put on oral thalidomide at a dose of 100 mg/d for 4 wk. Colonoscopy at week 4 of follow-up revealed remarkable healing of the ulcers with scar tissue in the ileocecal valve and distal ileum (Figure 3D-F). There was no recurrent bleeding during the 1 year of follow-up. In addition, no organ rejection was found after withdrawing tacrolimus.

DISCUSSION

It is well known that solid organ transplantation patients are particularly at risk for GI complications. Severe GI complications such as GI bleeding and GI perforation may negatively influence long-term outcome and become deadly. It had been reported that GI bleeding occurred in 2.3%-6.4% patients after liver transplantation^[8,9] and GI perforation in 2.9% patients after renal transplantation^[7]. Ulcer diseases are an important cause of GI bleeding, and perforation can manifest with symptoms of abdominal pain or diarrhea. Most of these ulcers are located at the anastomotic stoma or gastroduodenum, whereas ulcers at the intestine or colon are rare. In the two cases described here, deep and large ulcers were located in the ileocecal valve and distal ileum with no lesions in the gastroduodenum. In case 1, there was a longer period of diarrhea and abdominal pain, while case 2 mainly presented with acute and massive GI bleeding.

Differentiation of ulcer diseases in post-transplantation patients is always difficult. Common and uncommon pathogenesis such as *Helicobacter pylori* infection, ischemia, infection of CMV, EBV, mycobacteria, and fungus, and post-transplant lymphoproliferative disorders (PTLDs) should be considered. In the present two cases, ulcers were in the ileocecal valve and distal ileum, and *Helicobacter pylori* infection was ruled out as the pathogen. Clinical manifestation and endoscopic characteristics of ulcers were not consistent with a pathogenesis of ischemia. Because of negative the findings on chest CT scan, serum T-SPOT.TB, and histopathology of ulcers, mycobacteria and fungus infection were also excluded. CMV infection is common in patients with solid organ transplantation, and attention should be paid in cases of gastrointestinal ulcers. In a study of renal transplant patients susceptible to a variety of GI complications, such as infections, ulcer disease, and malignancies, CMV infection occurred in 11% of all patients^[1]. Although oral ganci-

clovir preventive strategy was used in case 2, this treatment might be inefficient in some patients, and atypical symptoms might be present^[10,11]. Decreased leukocyte count and interstitial pneumonitis are always found in CMV infection, but in both of our cases, no positive signs about CMV infection were noted in leukocyte count or chest CT scan. More importantly, immunohistochemistry stain for CMV of ulcer biopsy tissue was negative, and both patients recovered without further anti-CMV therapy. Therefore, CMV infection was not considered the cause of ulcers in these two cases.

PTLD was most difficult to be excluded in both patients. This was especially true in case 1, where the ulcers in the ileocecal valve and distal ileum appeared deeper and larger, with surrounding proliferative tissue. PTLD is a severe complication after organ transplantation with a cumulative incidence of 1.1% at 18 mo and 4.7% at 15 years, and it is always associated with EBV infection^[12]. Lymph nodes, GI tract, and graft liver are the most common sites of involvement^[13]. The involvement of the GI tract could result in deadly perforation and hemorrhage. In both patients, no persistent fever, palpable superficial lymph nodes, enlarged liver, or lymph nodes were found on chest and abdominal CT scan or ultrasound; EBER was negative by *in situ* hybridization. Pathology was also not consistent with the characteristics of PTLD. Furthermore, PTLD usually deteriorates quickly and is difficult to treat. However, the present two patients were stable within 1-2 wk, and the ulcers healed rapidly in 4-6 wk. Taken together, the evidence did not support the diagnosis of PTLD.

Finally, we considered that ulcer development was related to the use of immunosuppressants. Current studies have shown that some immunosuppressants, such as mammalian target of rapamycin inhibitors, had inhibitory effects on wound healing. Sirolimus is the most common drug that can lead to impairment of wound healing, and the most common wound complication is skin or dermal eruption^[14-17]. Therefore, the immunosuppressant use was considered to be a possible cause of GI epithelium impairment. Fortunately, both patients recovered quickly after withdrawing sirolimus and tacrolimus, supporting our speculation. In Smith *et al.*^[6], three liver transplant patients taking sirolimus suffered from gastrointestinal hemorrhage due to complications of gastroduodenal ulcers. The ulcers in two patients healed only after discontinuation of sirolimus, and the third patient died of massive gastrointestinal bleeding^[6].

Thalidomide has anti-angiogenic properties and seems effective in some cases of GI bleeding, especially angiodysplasia-related bleeding^[18-20]. It is also used in some inflammatory and ulcerative diseases like inflammatory bowel disease and some skin and oral ulcers, because of its anti-inflammatory and immunomodulatory effects^[20-22]. In our clinical practice, thalidomide is effective in some unexplained

and refractory multiple ulcers of intestine and related GI bleeding. It also seemed to work in our present two patients. Thalidomide was administered at a dose of 100 mg/d for 2 wk and 4 wk respectively, and no severe side effects were found.

In summary, some types of immunosuppressants, such as sirolimus and tacrolimus, can lead to impairment of the GI track and sometimes to the development of severe ulcers. Withdrawing them or switching to other immunosuppressants might be effective to treat these ulcers.

COMMENTS

Case characteristics

A 56-year-old man presented with severe diarrhea and abdominal pain after orthotopic liver transplantation, and a 34-year-old man presented with hematochezia and severe anemia after living-donor kidney transplantation.

Clinical diagnosis

Multiple giant ulcers in the distal ileum and ileocecal valve were caused by immunosuppressants.

Differential diagnosis

Common and uncommon pathogenesis of gastrointestinal (GI) ulcers in solid organ transplant recipient, such as *Helicobacter pylori* infection, ischemia, infection of cytomegalovirus (CMV), Epstein-Barr virus (EBV), mycobacteria and fungus, and post-transplant lymphoproliferative disorders (PTLDs), should be considered.

Laboratory diagnosis

Blood and stool cultures for fungus and bacteria showed no growth, and serum T-SPOT.TB was negative.

Imaging diagnosis

Computed tomography (CT) scan revealed no current tuberculosis, and there were no abnormal findings on abdominal enhanced CT or doppler ultrasound.

Endoscopic diagnosis

Colonoscopy revealed multiple giant and deep ulcers in the ileocecal valve and distal ileum.

Pathological diagnosis

Histopathology revealed chronic inflammation without signs of fungus or parasite infection, negative immunohistochemistry stain for CMV, and negative stain for EBV encoded early small RNA by *in situ* hybridization.

Treatment

Sirolimus or tacrolimus was withdrawn, and thalidomide was administered.

Related reports

Most GI ulcers are located at the anastomotic stoma or gastroduodenum in post-transplant recipients, and ulcers at the intestine or colon are rare. Besides ischemia and infection of CMV, EBV, mycobacteria, and fungus, use of immunosuppressants might contribute to the impairment of gastrointestinal tract.

Term explanation

PTLDs are a severe complication of solid organ and hematopoietic stem cell transplantation, including lymphoproliferative entities varying from reactive hyperplasia to malignant lymphoma. EBV is the main pathogen of PTLD.

Experiences and lessons

In some post-transplantation cases with ileal ulcers, sirolimus or tacrolimus should be considered as a risk factor, and withdrawing them or switching to other immunosuppressants might be effective.

Peer-review

This article is very interesting for those in the field of liver and kidney transplantation. Since ulcers in the distal ileum in solid organ transplant recipients are very rare, experience about the management of these patients is very useful.

REFERENCES

- 1 **Ishaque M**, Rashid R, Mubarak M. Gastrointestinal complications in renal transplant recipients detected by endoscopic biopsies in a developing country. *Indian J Gastroenterol* 2015; **34**: 51-57 [PMID: 25757628 DOI: 10.1007/s12664-015-0537-8]
- 2 **Lahon B**, Mordant P, Thabut G, Georger JF, Dauriat G, Mal H, Lesèche G, Castier Y. Early severe digestive complications after lung transplantation. *Eur J Cardiothorac Surg* 2011; **40**: 1419-1424 [PMID: 21497510 DOI: 10.1016/j.ejcts.2011.02.069]
- 3 **Telkes G**, Peter A, Tulassay Z, Asderakis A. High frequency of ulcers, not associated with *Helicobacter pylori*, in the stomach in the first year after kidney transplantation. *Nephrol Dial Transplant* 2011; **26**: 727-732 [PMID: 20603242 DOI: 10.1093/ndt/gfq401]
- 4 **Grass F**, Schäfer M, Cristaudi A, Berutto C, Aubert JD, Gonzalez M, Demartines N, Ris HB, Soccia PM, Krueger T. Incidence and Risk Factors of Abdominal Complications After Lung Transplantation. *World J Surg* 2015; **39**: 2274-2281 [PMID: 26013207 DOI: 10.1007/s00268-015-3098-1]
- 5 **Gad EH**, Alsebaey A, Lotfy M, Eltabbakh M, Sherif AA. Complications and mortality after adult to adult living donor liver transplantation: A retrospective cohort study. *Ann Med Surg (Lond)* 2015; **4**: 162-171 [PMID: 26005570 DOI: 10.1016/j.amsu.2015.04.021]
- 6 **Smith AD**, Bai D, Marroquin CE, Tuttle-Newhall JE, Desai DM, Collins BH, Muir A, Kuo PC, McHutchison J, Rockey DC. Gastrointestinal hemorrhage due to complicated gastroduodenal ulcer disease in liver transplant patients taking sirolimus. *Clin Transplant* 2005; **19**: 250-254 [PMID: 15740563]
- 7 **Catena F**, Ansaloni L, Gazzotti F, Bertelli R, Severi S, Coccolini F, Fuga G, Nardo B, D'Alessandro L, Faenza A, Pinna AD. Gastrointestinal perforations following kidney transplantation. *Transplant Proc* 2008; **40**: 1895-1896 [PMID: 18675082 DOI: 10.1016/j.transproceed.2008.06.007]
- 8 **Kimura K**, Ikegami T, Bekki Y, Ninomiya M, Yamashita Y, Yoshizumi T, Yoshiya S, Soejima Y, Harada N, Shirabe K, Maehara Y. Clinical significance of gastrointestinal bleeding after living donor liver transplantation. *Transpl Int* 2014; **27**: 705-711 [PMID: 24673842 DOI: 10.1111/tri.12325]
- 9 **Ma Y**, He XS, Zhu XF, Wang GD, Wang DP, Hu AB, Ju WQ, Wu LW, Tai Q. [Etiology and management of postoperative gastrointestinal bleeding after orthotopic liver transplantation]. *Zhonghua Wei Chang Wai Ke Zazhi* 2010; **13**: 26-28 [PMID: 20099156]
- 10 **Bataille S**, Moal V, Gaudart J, Indreies M, Purgus R, Dussol B, Zandotti C, Berland Y, Vacher-Coponat H. Cytomegalovirus risk factors in renal transplantation with modern immunosuppression. *Transpl Infect Dis* 2010; **12**: 480-488 [PMID: 20629971 DOI: 10.1111/j.1399-3062.2010.00533.x]
- 11 **Pérez-Valentín MA**, Cofán F, Solé M, Llach J, Esforzado N, Campistol JM, Oppenheimer F. [Atypical cytomegalovirus in renal transplantation: a new form of presentation]. *Nefrologia* 2002; **22**: 381-385 [PMID: 12369131]
- 12 **Cornejo A**, Bohnenblust M, Harris C, Abrahamian GA. Intestinal perforation associated with rituximab therapy for post-transplant lymphoproliferative disorder after liver transplantation. *Cancer Chemother Pharmacol* 2009; **64**: 857-860 [PMID: 19588139 DOI: 10.1007/s00280-009-1062-1]

- 13 **Lo RC**, Chan SC, Chan KL, Chiang AK, Lo CM, Ng IO. Post-transplant lymphoproliferative disorders in liver transplant recipients: a clinicopathological study. *J Clin Pathol* 2013; **66**: 392-398 [PMID: 23423516 DOI: 10.1136/jclinpath-2012-201139]
- 14 **Nashan B**, Citterio F. Wound healing complications and the use of mammalian target of rapamycin inhibitors in kidney transplantation: a critical review of the literature. *Transplantation* 2012; **94**: 547-561 [PMID: 22941182]
- 15 **Tiong HY**, Flechner SM, Zhou L, Wee A, Mastroianni B, Savas K, Goldfarb D, Derweesh I, Modlin C. A systematic approach to minimizing wound problems for de novo sirolimus-treated kidney transplant recipients. *Transplantation* 2009; **87**: 296-302 [PMID: 19155988 DOI: 10.1097/TP.0b013e318192dd56]
- 16 **Pengel LH**, Liu LQ, Morris PJ. Do wound complications or lymphocele occur more often in solid organ transplant recipients on mTOR inhibitors? A systematic review of randomized controlled trials. *Transpl Int* 2011; **24**: 1216-1230 [PMID: 21955006 DOI: 10.1111/j.1432-2277.2011.01357.x]
- 17 **Knight RJ**, Villa M, Laskey R, Benavides C, Schoenberg L, Welsh M, Kerman RH, Podder H, Van Buren CT, Katz SM, Kahan BD. Risk factors for impaired wound healing in sirolimus-treated renal transplant recipients. *Clin Transplant* 2007; **21**: 460-465 [PMID: 17645704]
- 18 **Boey JP**, Hahn U, Sagheer S, McRae SJ. Thalidomide in angiodysplasia-related bleeding. *Intern Med J* 2015; **45**: 972-976 [PMID: 26332623 DOI: 10.1111/imj.12850]
- 19 **Moser S**, Tischer A, Karpi A, Schleicher M, Stavjanik S, Gschwantler M. Evidence that thalidomide is effective in recurrent bleeding from watermelon stomach associated with liver cirrhosis. *Endoscopy* 2014; **46** Suppl 1 UCTN: E384 [PMID: 25254589 DOI: 10.1055/s-0034-1377369]
- 20 **Amirshahrokhi K**, Khalili AR. The effect of thalidomide on ethanol-induced gastric mucosal damage in mice: involvement of inflammatory cytokines and nitric oxide. *Chem Biol Interact* 2015; **225**: 63-69 [PMID: 25478868 DOI: 10.1016/j.cbi.2014.11.019]
- 21 **Diamanti A**, Capriati T, Papadatou B, Knafelz D, Bracci F, Corsetti T, Elia D, Torre G. The clinical implications of thalidomide in inflammatory bowel diseases. *Expert Rev Clin Immunol* 2015; **11**: 699-708 [PMID: 25865355 DOI: 10.1586/1744666X.2015.1027687]
- 22 **Barrons RW**. Treatment strategies for recurrent oral aphthous ulcers. *Am J Health Syst Pharm* 2001; **58**: 41-50; quiz 51-53 [PMID: 11194135]

P- Reviewer: Markic D **S- Editor:** Yu J **L- Editor:** Filipodia
E- Editor: Ma S





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgooffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>



ISSN 1007-9327



9 771007 932045