

World Journal of Gastroenterology®

Volume 18 Number 5
February 7, 2012



Published by Baishideng Publishing Group Co., Limited
Room 1701, 17/F, Henan Building,
No. 90 Jaffe Road, Wanchai, Hong Kong, China
Fax: +852-31158812
Telephone: +852-58042046
E-mail: bpg@baishideng.com
<http://www.wjgnet.com>

ISSN 1007-9327



World Journal of *Gastroenterology*

World J Gastroenterol 2012 February 7; 18(5): 393-490



Editorial Board

2010-2013

The *World Journal of Gastroenterology* Editorial Board consists of 1361 members, representing a team of worldwide experts in gastroenterology and hepatology. They are from 64 countries, including Albania (1), Argentina (8), Australia (34), Austria (14), Belgium (14), Brazil (13), Brunei Darussalam (1), Bulgaria (2), Canada (23), Chile (3), China (82), Colombia (1), Croatia (2), Cuba (1), Czech (6), Denmark (9), Ecuador (1), Egypt (4), Estonia (2), Finland (8), France (31), Germany (86), Greece (22), Hungary (11), India (32), Indonesia (2), Iran (10), Ireland (6), Israel (13), Italy (126), Japan (141), Jordan (2), Kuwait (1), Lebanon (4), Lithuania (2), Malaysia (1), Mexico (11), Morocco (1), Moldova (1), Netherlands (32), New Zealand (2), Norway (13), Pakistan (2), Poland (11), Portugal (6), Romania (4), Russia (1), Saudi Arabia (3), Serbia (3), Singapore (12), Slovenia (1), South Africa (3), South Korea (46), Spain (43), Sri Lanka (1), Sweden (17), Switzerland (12), Thailand (1), Trinidad and Tobago (1), Turkey (30), United Arab Emirates (2), United Kingdom (95), United States (287), and Uruguay (1).

HONORARY EDITORS-IN-CHIEF

James L Boyer, *New Haven*
Ke-Ji Chen, *Beijing*
Martin H Floch, *New Haven*
Bo-Rong Pan, *Xi'an*
Emmet B Keeffe, *Palo Alto*
Eamonn M Quigley, *Cork*
Rafiq A Sheikh, *Sacramento*
Nicholas J Talley, *Rochester*

EDITOR-IN-CHIEF

Ferruccio Bonino, *Pisa*
Myung-Hwan Kim, *Seoul*
Kjell Öberg, *Uppsala*
Matt Rutter, *Stockton-on-Tees*
Andrzej S Tarnawski, *Long Beach*

STRATEGY ASSOCIATE

EDITORS-IN-CHIEF

You-Yong Lu, *Beijing*
Peter Draganov, *Florida*
Hugh J Freeman, *Vancouver*
María Concepción Gutiérrez-Ruiz, *México*
Kazuhiro Hanazaki, *Kochi*
Akio Inui, *Kagoshima*
Kalpesh Jani, *Baroda*
Javier S Martin, *Punta del Este*
Natalia A Osna, *Omaha*
Wei Tang, *Tokyo*
Alan BR Thomson, *Edmonton*
Harry HX Xia, *East Hanover*
John M Luk, *Hong Kong*
Hiroschi Shimada, *Yokohama*

GUEST EDITORIAL BOARD

MEMBERS

Jiunn-Jong Wu, *Tainan*

Cheng-Shyong Wu, *Chia-Yi*
Ta-Sen Yeh, *Taoyuan*
Tsung-Hui Hu, *Kaohsiung*
Chuah Seng-Kee, *Kaohsiung*
I-Rue Lai, *Taipei*
Jin-Town Wang, *Taipei*
Ming-Shiang Wu, *Taipei*
Teng-Yu Lee, *Taichung*
Yang-Yuan Chen, *Changhua*
Po-Shiuan Hsieh, *Taipei*
Chao-Hung Hung, *Kaohsiung*
Hon-Yi Shi, *Kaohsiung*
Hui-kang Liu, *Taipei*
Jen-Hwey Chiu, *Taipei*
Chih-Chi Wang, *Kaohsiung*
Wan-Long Chuang, *Kaohsiung*
Wen-Hsin Huang, *Taichung*
Hsu-Heng Yen, *Changhua*
Ching Chung Lin, *Taipei*
Chien-Jen Chen, *Taipei*
Jaw-Ching Wu, *Taipei*
Ming-Chih Hou, *Taipei*
Kevin Cheng-Wen Hsiao, *Taipei*
Chiun Hsu, *Taipei*
Yu-Jen Chen, *Taipei*
Chen Hsiu-Hsi Chen, *Taipei*
Liang-Shun Wang, *Taipei*
hun-Fa Yang, *Taichung*
Min-Hsiung Pan, *Kaohsiung*
Chun-Hung Lin, *Taipei*
Ming-Whei Yu, *Taipei*
Chuen Hsueh, *Taoyuan*
Hsiu-Po Wang, *Taipei*
Lein-Ray Mo, *Tainan*
Ming-Lung Yu, *Kaohsiung*

MEMBERS OF THE EDITORIAL BOARD



Albania

Bashkim Resuli, *Tirana*



Argentina

Julio H Carri, *Córdoba*
Bernabe Matias Quesada, *Buenos Aires*
Bernardo Frider, *Buenos Aires*
Maria Ines Vaccaro, *Buenos Aires*
Eduardo de Santibañes, *Buenos Aires*
Adriana M Torres, *Rosario*
Carlos J Pirola, *Buenos Aires*
Silvia Sookoian, *Buenos Aires*



Australia

Finlay A Macrae, *Victoria*
David Ian Watson, *Bedford Park*
Jacob George, *Westmead*
Leon Anton Adams, *Nedlands*
Minoti V Apte, *Liverpool*
Andrew V Biankin, *Sydney*
Filip Braet, *Sydney*
Guy D Eslick, *Sydney*
Michael A Fink, *Melbourne*
Mark D Gorrell, *Sydney*
Michael Horowitz, *Adelaide*
John E Kellow, *Sydney*
Daniel Markovich, *Brisbane*

Phillip S Oates, *Perth*
 Ross C Smith, *Sydney*
 Kevin J Spring, *Brisbane*
 Philip G Dinning, *Koagarah*
 Christopher Christophi, *Melbourne*
 Cuong D Tran, *North Adelaide*
 Shan Rajendra, *Tasmania*
 Rajvinder Singh, *Elizabeth Vale*
 William Kemp, *Melbourne*
 Phil Sutton, *Melbourne*
 Richard Anderson, *Victoria*
 Vance Matthews, *Melbourne*
 Alexander G Heriot, *Melbourne*
 Debbie Trinder, *Fremantle*
 Ian C Lawrance, *Perth*
 Adrian G Cummins, *Adelaide*
 John K Olynyk, *Fremantle*
 Alex Boussioutas, *Melbourne*
 Emilia Prakoso, *Sydney*
 Robert JL Fraser, *Daw Park*
 Stefan Riss, *Vienna*



Austria

Wolfgang Mikulits, *Vienna*
 Alfred Gangl, *Vienna*
 Dietmar Öfner, *Salzburg*
 Georg Roth, *Vienna*
 Herwig R Cerwenka, *Graz*
 Ashraf Dahaba, *Graz*
 Markus Raderer, *Vienna*
 Alexander M Hirschl, *Wien*
 Thomas Wild, *Kapellerfeld*
 Peter Ferenci, *Vienna*
 Valentin Fuhrmann, *Vienna*
 Kurt Lenz, *Linz*
 Markus Peck-Radosavljevic, *Vienna*
 Michael Trauner, *Graz*



Belgium

Rudi Beyaert, *Gent*
 Inge I Depoortere, *Leuven*
 Olivier Detry, *Liège*
 Benedicte Y De Winter, *Antwerp*
 Etienne M Sokal, *Brussels*
 Marc Peeters, *De Pintelaan*
 Eddie Wisse, *Keerbergen*
 Jean-Yves L Reginster, *Liège*
 Mark De Ridder, *Brussel*
 Freddy Penninckx, *Leuven*
 Kristin Verbeke, *Leuven*
 Lukas Van Oudenhove, *Leuven*
 Leo van Grunsven, *Brussels*
 Philip Meuleman, *Ghent*



Brazil

Heitor Rosa, *Goiania*
 Roberto J Carvalho-Filho, *Sao Paulo*
 Damiao Carlos Moraes Santos, *Rio de Janeiro*
 Marcelo Lima Ribeiro, *Braganca Paulista*
 Eduardo Garcia Vilela, *Belo Horizonte*
 Jaime Natan Eisig, *São Paulo*
 Andre Castro Lyra, *Salvador*
 José Liberato Ferreira Caboclo, *Brazil*
 Yukié Sato-Kuwabara, *São Paulo*
 Raquel Rocha, *Salvador*

Paolo R Salvalaggio, *Sao Paulo*
 Ana Cristina Simões e Silva, *Belo Horizonte*
 Joao Batista Teixeira Rocha, *Santa Maria*



Brunei Darussalam

Vui Heng Chong, *Bandar Seri Begawan*



Bulgaria

Zahariy Krastev, *Sofia*
 Mihaela Petrova, *Sofia*



Canada

Eldon Shaffer, *Calgary*
 Nathalie Perreault, *Sherbrooke*
 Philip H Gordon, *Montreal*
 Ram Prakash Galwa, *Ottawa*
 Baljinder Singh Salh, *Vancouver*
 Claudia Zwingmann, *Montreal*
 Kris Chadee, *Calgary*
 Alain Bitton, *Montreal*
 Pingchang Yang, *Hamilton*
 Michael F Byrne, *Vancouver*
 Andrew L Mason, *Alberta*
 John K Marshall, *Ontario*
 Kostas Pantopoulos, *Quebec*
 Martin Storr, *Calgary*
 Waliul Khan, *Ontario*
 Eric M Yoshida, *Vancouver*
 Geoffrey C Nguyen, *Toronto*
 Devendra K Amre, *Montreal*
 Tedros Bezabeh, *Winnipeg*
 Wangxue Chen, *Ottawa*
 Qiang Liu, *Saskatoon*



Chile

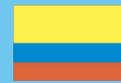
De Aretxabala Xabier, *Santiago*
 Marcelo A Beltran, *La Serena*
 Silvana Zanlungo, *Santiago*



China

Chi-Hin Cho, *Hong Kong*
 Chun-Qing Zhang, *Jinan*
 Ren Xiang Tan, *Nanjing*
 Fei Li, *Beijing*
 Hui-Jie Bian, *Xi'an*
 Xiao-Peng Zhang, *Beijing*
 Xing-Hua Lu, *Beijing*
 Fu-Sheng Wang, *Beijing*
 An-Gang Yang, *Xi'an*
 Xiao-Ping Chen, *Wuhan*
 Zong-Jie Cui, *Beijing*
 Ming-Liang He, *Hong Kong*
 Yuk-Tong Lee, *Hong Kong*
 Qin Su, *Beijing*
 Jian-Zhong Zhang, *Beijing*
 Paul Kwong-Hang Tam, *Hong Kong*
 Wen-Rong Xu, *Zhenjiang*
 Chun-Yi Hao, *Beijing*
 San-Jun Cai, *Shanghai*
 Simon Law, *Hong Kong*
 Yuk Him Tam, *Hong Kong*

De-Liang Fu, *Shanghai*
 Eric WC Tse, *Hong Kong*
 Justin CY Wu, *Hong Kong*
 Nathalie Wong, *Hong Kong*
 Jing Yuan Fang, *Shanghai*
 Yi-Min Mao, *Shanghai*
 Wei-Cheng You, *Beijing*
 Xiang-Dong Wang, *Shanghai*
 Xuan Zhang, *Beijing*
 Zhao-Shen Li, *Shanghai*
 Guang-Wen Cao, *Shanghai*
 En-min Li, *Shantou*
 Yu-Yuan Li, *Guangzhou*
 Fook Hong Ng, *Hong Kong*
 Hsiang-Fu Kung, *Hong Kong*
 Wai Lun Law, *Hong Kong*
 Eric CH Lai, *Hong Kong*
 Jun Yu, *Hong Kong*
 Ze-Guang Han, *Shanghai*
 Bian zhao-xiang, *Hong Kong*
 Wei-Dong Tong, *Chongqing*



Colombia

Germán Campuzano-Maya, *Medellín*



Croatia

Tamara Cacev, *Zagreb*
 Marko Duvnjak, *Zagreb*



Cuba

Damian C Rodriguez, *Havana*



Czech

Milan Jirsa, *Praha*
 Pavel Trunečka, *Prague*
 Jan Bures, *Hradec Kralove*
 Marcela Kopacova, *Hradec Kralove*
 Ondrej Slaby, *Brno*
 Radan Bruha, *Prague*



Denmark

Asbjørn M Drewes, *Aalborg*
 Leif Percival Andersen, *Copenhagen*
 Jan Mollenhauer, *Odense C*
 Morten Frisch, *Copenhagen S*
 Jorgen Rask-Madsen, *Skodsborg*
 Morten Hylander Møller, *Holte*
 Søren Rafaelsen, *Vejle*
 Vibeke Andersen, *Viborg*
 Ole Haagen Nielsen, *Herlev*



Ecuador

Fernando E Sempértegui, *Quito*



Egypt

Zeinab Nabil Ahmed, *Cairo*

Hussein M Atta, *El-Minia*
Asmaa Gaber Abdou, *Shebein Elkom*
Maha Maher Shehata, *Mansoura*



Estonia

Riina Salupere, *Tartu*
Tamara Vorobjova, *Tartu*



Finland

Saila Kauhanen, *Turku*
Pauli Antero Puolakkainen, *Turku*
Minna Nyström, *Helsinki*
Juhani Sand, *Tampere*
Jukka-Pekka Mecklin, *Jyväskylä*
Lea Veijola, *Helsinki*
Kaija-Leena Kolho, *Helsinki*
Thomas Kietzmann, *Oulu*



France

Boris Guiu, *Dijon*
Baumert F Thomas, *Strasbourg*
Alain L Servin, *Châtenay-Malabry*
Patrick Marcellin, *Paris*
Jean-Jacques Tuech, *Rouen*
Francoise L Fabiani, *Angers*
Jean-Luc Faucheron, *Grenoble*
Philippe Lehours, *Bordeaux*
Stephane Supiot, *Nantes*
Lionel Bueno, *Toulouse*
Claire Bonithon-Kopp, *Dijon*
Flavio Maina, *Marseille*
Paul Hofman, *Nice*
Abdel-Majid Khatib, *Paris*
Annie Schmid-Alliana, *Nice cedex 3*
Frank Zerbib, *Bordeaux Cedex*
Jean Paul Galmiche, *Nantes cedex*
Rene Gerolami Santandera, *Marseille*
Sabine Colnot, *Paris*
Catherine Daniel, *Lille Cedex*
Thabut Dominique, *Paris*
Laurent Huwart, *Paris*
Alain Braillon, *Amiens*
Bruno Bonaz, *Grenoble*
Evelyne Schvoerer, *Strasbourg*
M Coeffier, *Rouen*
Mathias Chamailard, *Lille*
Hang Nguyen, *Clermont-Ferrand*
Veronique Vitton, *Marseille*
Alexis Desmoulière, *Limoges*
Juan Iovanna, *Marseille*



Germany

Hans L Tillmann, *Leipzig*
Stefan Kubicka, *Hannover*
Elke Cario, *Essen*
Hans Scherubl, *Berlin*
Harald F Teutsch, *Ulm*
Peter Konturek, *Erlangen*
Thilo Hackert, *Heidelberg*
Jurgen M Stein, *Frankfurt*
Andrej Khandoga, *Munich*
Karsten Schulmann, *Bochum*
Jutta Elisabeth Lüttges, *Riegelsberg*

Wolfgang Haggmann, *Heidelberg*
Hubert Blum, *Freiburg*
Thomas Bock, *Tuebingen*
Christa Buechler, *Regensburg*
Christoph F Dietrich, *Bad Mergentheim*
Ulrich R Fölsch, *Kiel*
Nikolaus Gassler, *Aachen*
Markus Gerhard, *Munich*
Dieter Glebe, *Giessen*
Klaus R Herrlinger, *Stuttgart*
Eberhard Hildt, *Berlin*
Joerg C Hoffmann, *Berlin*
Joachim Labenz, *Siegen*
Peter Malferttheiner, *Magdeburg*
Sabine Mihm, *Göttingen*
Markus Reiser, *Bochum*
Steffen Rickes, *Magdeburg*
Andreas G Schreyer, *Regensburg*
Henning Schulze-Bergkamen, *Mainz*
Ulrike S Stein, *Berlin*
Wolfgang R Stremmel, *Heidelberg*
Fritz von Weizsäcker, *Berlin*
Stefan Wirth, *Wuppertal*
Dean Bogoevski, *Hamburg*
Bruno Christ, *Halle/Saale*
Peter N Meier, *Hannover*
Stephan Johannes Ott, *Kiel*
Arndt Vogel, *Hannover*
Dirk Haller, *Freising*
Jens Standop, *Bonn*
Jonas Mudter, *Erlangen*
Jürgen Büning, *Lübeck*
Matthias Ocker, *Erlangen*
Joerg Trojan, *Frankfurt*
Christian Trautwein, *Aachen*
Jorg Kleeff, *Munich*
Christian Rust, *Munich*
Claus Hellerbrand, *Regensburg*
Elke Roeb, *Giessen*
Erwin Biecker, *Siegburg*
Ingmar Königsrainer, *Tübingen*
Jürgen Borlak, *Hannover*
Axel M Gressner, *Aachen*
Oliver Mann, *Hamburg*
Marty Zdichavsky, *Tübingen*
Christoph Reichel, *Bad Brückenau*
Nils Habbe, *Marburg*
Thomas Wex, *Magdeburg*
Frank Ulrich Weiss, *Greifswald*
Manfred V Singer, *Mannheim*
Martin K Schilling, *Homburg*
Philip D Hard, *Giessen*
Michael Linnebacher, *Rostock*
Ralph Graeser, *Freiburg*
Rene Schmidt, *Freiburg*
Robert Obermaier, *Freiburg*
Sebastian Mueller, *Heidelberg*
Andrea Hille, *Goettingen*
Klaus Mönkemüller, *Bottrop*
Elfriede Bollschweiler, *Köln*
Siegfried Wagner, *Deggendorf*
Dieter Schilling, *Mannheim*
Joerg F Schlaak, *Essen*
Michael Keese, *Frankfurt*
Robert Grützmänn, *Dresden*
Ali Canbay, *Essen*
Dirk Domagk, *Muenster*
Jens Hoepfner, *Freiburg*
Frank Tacke, *Aachen*
Patrick Michl, *Marburg*
Alfred A Königsrainer, *Tübingen*

Kilian Weigand, *Heidelberg*
Mohamed Hassan, *Duesseldorf*
Gustav Paumgartner, *Munich*
Philippe N Khalil, *Munich*



Greece

Andreas Larentzakis, *Athens*
Tsianos Epameinondas, *Ioannina*
Elias A Kouroumalis, *Heraklion*
Helen Christopoulou-Aletra, *Thessaloniki*
George Papatheodoridis, *Athens*
Ioannis Kanellos, *Thessaloniki*
Michael Koutsilieris, *Athens*
T Choli-Papadopoulou, *Thessaloniki*
Emanuel K Manesis, *Athens*
Evangelos Tsiambas, *Ag Paraskevi Attiki*
Konstantinos Mimidis, *Alexandroupolis*
Spilios Manolakopoulos, *Athens*
Spiros Sgouros, *Athens*
Ioannis E Koutroubakis, *Heraklion*
Stefanos Karagiannis, *Athens*
Spiros Ladas, *Athens*
Vezali Elena, *Athens*
Dina G Tiniakos, *Athens*
Ekaterini Chatzaki, *Alexandroupolis*
Dimitrios Roukos, *Ioannina*
George Sgourakis, *Athens*
Maroulio Talieri, *Athens*



Hungary

Peter L Lakatos, *Budapest*
Yvette Mándi, *Szeged*
Ferenc Sipos, *Budapest*
György M Buzás, *Budapest*
László Czakó, *Szeged*
Peter Hegyi, *Szeged*
Zoltan Rakonczay, *Szeged*
Gyula Farkas, *Szeged*
Zsuzsa Szondy, *Debrecen*
Gabor Veres, *Budapest*
Zsuzsa Schaff, *Budapest*



India

Philip Abraham, *Mumbai*
Sri P Misra, *Allahabad*
Ramesh Roop Rai, *Jaipur*
Nageshwar D Reddy, *Hyderabad*
Rakesh Kumar Tandon, *New Delhi*
Jai Dev Wig, *Chandigarh*
Uday C Ghoshal, *Lucknow*
Pramod Kumar Garg, *New Delhi*
Barjesh Chander Sharma, *New Delhi*
Gopal Nath, *Varanasi*
Bhupendra Kumar Jain, *Delhi*
Devinder Kumar Dhawan, *Chandigarh*
Jagannath Palepu, *Mumbai*
Ashok Kumar, *Lucknow*
Benjamin Perakath, *Tamil Nadu*
Debidas Ghosh, *Midnapore*
Pankaj Garg, *Panchkula*
Samiran Nundy, *New Delhi*
Virendra Singh, *Chandigarh*
Bikash Medhi, *Chandigarh*
Radha K Dhiman, *Chandigarh*
Vandana Panda, *Mumbai*
Vineet Ahuja, *New Delhi*

SV Rana, *Chandigarh*
Deepak N Amarapurkar, *Mumbai*
Abhijit Chowdhury, *Kolkata*
Jasbir Singh, *Kurukshehra*
B Mittal, *Lucknow*
Sundeep Singh Saluja, *New Delhi*
Pradyumna Kumar Mishra, *Mumbai*
Runu Chakravarty, *Kolkata*



Indonesia

David handoyo Muljono, *Jakarta*
Andi Utama, *Tangerang*



Iran

Seyed-Moayed Alavian, *Tehran*
Reza Malekzadeh, *Tehran*
Peyman Adibi, *Isfahan*
Alireza Mani, *Tehran*
Seyed Mohsen Dehghani, *Shiraz*
Mohammad Abdollahi, *Tehran*
Majid Assadi, *Bushehr*
Arezoo Aghakhani, *Tehran*
Marjan Mohammadi, *Tehran*
Fariborz Mansour-Ghanaei, *Rasht*



Ireland

Ross McManus, *Dublin*
Billy Bourke, *Dublin*
Catherine Greene, *Dublin*
Ted Dinan, *Cork*
Marion Rowland, *Dublin*



Israel

Abraham R Eliakim, *Haifa*
Simon Bar-Meir, *Hashomer*
Ami D Sperber, *Beer-Sheva*
Boris Kirshtein, *Beer Sheva*
Mark Pines, *Bet Dagan*
Menachem Moshkowitz, *Tel-Aviv*
Ron Shaoul, *Haifa*
Shmuel Odes, *Beer Sheva*
Sigal Fishman, *Tel Aviv*
Alexander Becker, *Afula*
Assy Nimer, *Safed*
Eli Magen, *Ashdod*
Amir Shlomai, *Tel-Aviv*



Italy

Mauro Bortolotti, *Bologna*
Gianlorenzo Dionigi, *Varese*
Fiorucci Stefano, *Perugia*
Roberto Berni Canani, *Naples*
Ballarin Roberto, *Modena*
Bruno Annibale, *Roma*
Vincenzo Stanghellini, *Bologna*
Giovanni B Gaeta, *Napoli*
Claudio Bassi, *Verona*
Mauro Bernardi, *Bologna*
Giuseppe Chiarioni, *Valeggio*
Michele Cicala, *Rome*

Dario Conte, *Milano*
Francesco Costa, *Pisa*
Giovanni D De Palma, *Naples*
Giammarco Fava, *Ancona*
Francesco Feo, *Sassari*
Edoardo G Giannini, *Genoa*
Fabio Grizzi, *Milan*
Salvatore Gruttadauria, *Palermo*
Pietro Invernizzi, *Milan*
Ezio Laconi, *Cagliari*
Giuseppe Montalto, *Palermo*
Giovanni Musso, *Torino*
Gerardo Nardone, *Napoli*
Valerio Nobili, *Rome*
Raffaele Pezzilli, *Bologna*
Alberto Piperno, *Monza*
Anna C Piscaglia, *Roma*
Piero Portincasa, *Bari*
Giovanni Tarantino, *Naples*
Cesare Tosetti, *Porretta Terme*
Alessandra Ferlini, *Ferrara*
Alessandro Ferrero, *Torino*
Donato F Altomare, *Bari*
Giovanni Milito, *Rome*
Giuseppe Sica, *Rome*
Guglielmo Borgia, *Naples*
Giovanni Latella, *L'Aquila*
Salvatore Auricchio, *Naples*
Alberto Biondi, *Rome*
Alberto Tommasini, *Trieste*
Antonio Basoli, *Roma*
Giuliana Decorti, *Trieste*
Marco Silano, *Roma*
Michele Reni, *Milan*
Pierpaolo Sileri, *Rome*
Achille Iolascon, *Naples*
Alessandro Granito, *Bologna*
Angelo A Izzo, *Naples*
Giuseppe Currò, *Messina*
Pier Mannuccio Mannucci, *Milano*
Marco Vivarelli, *Bologna*
Massimo Levrero, *Rome*
Massimo Rugge, *Padova*
Paolo Angeli, *Padova*
Silvio Danese, *Milano*
Antonello Trecca, *Rome*
Antonio Gasbarrini, *Rome*
Cesare Ruffolo, *Treviso*
Massimo Falconi, *Verona*
Fausto Catena, *Bologna*
Francesco Manguso, *Napoli*
Giancarlo Mansueti, *Verona*
Luca Morelli, *Trento*
Marco Scarpa, *Padova*
Mario M D'Elios, *Florence*
Francesco Luzzza, *Catanzaro*
Franco Roviello, *Siena*
Guido Torzilli, *Rozzano Milano*
Luca Frulloni, *Verona*
Lucia Malaguarnera, *Catania*
Lucia Ricci Vitiani, *Rome*
Mara Massimi, *Coppito*
Mario Nano, *Torino*
Mario Pescatori, *Rome*
Mario Rizzetto, *Torino*
Mirko D'Onofrio, *Verona*
Nadia Peparini, *Rome*
Paola De Nardi, *Milan*
Paolo Aurello, *Rome*
Piero Amodio, *Padova*
Riccardo Nascimbeni, *Brescia*

Vincenzo Villanacci, *Brescia*
Vittorio Ricci, *Pavia*
Silvia Fargion, *Milan*
Luigi Bonavina, *Milano*
Oliviero Riggio, *Rome*
Fabio Pace, *Milano*
Gabrio Bassotti, *Perugia*
Giulio Marchesini, *Bologna*
Roberto de Franchis, *Milano*
Giovanni Monteleone, *Rome*
Carmelo Scarpignato, *Parma*
Luca VC Valenti, *Milan*
Urgesi Riccardo, *Rome*
Marcello Persico, *Naples*
Pietro Valdastrì, *Pontedera*
Antonio Moschetta, *Bari*
Luigi Muratori, *Bologna*
Angelo Zullo, *Roma*
Vito Annese, *Florence*
Simone Lanini, *Rome*
Alessandro Grasso, *Savona*
Giovanni Targher, *Verona*
Domenico Girelli, *Verona*
Alessandro Cucchetti, *Bologna*
Fabio Marra, *Florence*
Michele Milella, *Rome*
Francesco Franceschi, *Rome*
Giuseppina De Petro, *Brescia*
Salvatore Leonardi, *Catania*
Cristiano Simone, *Santa Maria Imbaro*
Bernardino Rampone, *Salerno*
Francesco Crea, *Pisa*
Walter Fries, *Messina*
Antonio Craxi, *Palermo*
Gerardo Rosati, *Potenza*
Mario Guslandi, *Milano*
Gianluigi Giannelli, *Bari*
Paola Loria, *Modena*
Paolo Sorrentino, *Avellino*
Armando Santoro, *Rozzano*
Gabriele Grassi, *Trieste*
Antonio Orlacchio, *Rome*



Japan

Tsuneo Kitamura, *Chiba*
Katsutoshi Yoshizato, *Higashihiroshima*
Masahiro Arai, *Tokyo*
Shinji Tanaka, *Hiroshima*
Keiji Hirata, *Kitakyushu*
Yoshio Shirai, *Niigata*
Susumu Ohmada, *Maebashi*
Kenichi Ikejima, *Bunkyo-ku*
Masatoshi Kudo, *Osaka*
Yoshiaki Murakami, *Hiroshima*
Masahiro Tajika, *Nagoya*
Kentaro Yoshika, *Toyoake*
Kyoichi Adachi, *Izumo*
Yasushi Adachi, *Sapporo*
Takafumi Ando, *Nagoya*
Akira Andoh, *Otsu*
Hitoshi Asakura, *Tokyo*
Mitsuhiro Fujishiro, *Tokyo*
Toru Hiyama, *Higashihiroshima*
Yutaka Inagaki, *Kanagawa*
Hiromi Ishibashi, *Nagasaki*
Shunji Ishihara, *Izumo*
Toru Ishikawa, *Niigata*
Yoshiaki Iwasaki, *Okayama*
Terumi Kamisawa, *Tokyo*

Norihiro Kokudo, *Tokyo*
 Shin Maeda, *Tokyo*
 Yasushi Matsuzaki, *Tsukuba*
 Kenji Miki, *Tokyo*
 Hiroto Miwa, *Hyogo*
 Yoshiharu Motoo, *Kanazawa*
 Kunihiko Murase, *Tusima*
 Atsushi Nakajima, *Yokohama*
 Yuji Naito, *Kyoto*
 Hisato Nakajima, *Tokyo*
 Hiroki Nakamura, *Yamaguchi*
 Shotaro Nakamura, *Fukuoka*
 Mikio Nishioka, *Niihama*
 Hirohide Ohnishi, *Akita*
 Kazuichi Okazaki, *Osaka*
 Morikazu Onji, *Ehime*
 Satoshi Osawa, *Hamamatsu*
 Hidetsugu Saito, *Tokyo*
 Yutaka Saito, *Tokyo*
 Yasushi Sano, *Chiba*
 Tomohiko Shimatan, *Hiroshima*
 Yukihiko Shimizu, *Toiyama*
 Shinji Shimoda, *Fukuoka*
 Masayuki Sho, *Nara*
 Hidekazu Suzuki, *Tokyo*
 Shinji Togo, *Yokohama*
 Satoshi Yamagiwa, *Niigata*
 Takayuki Yamamoto, *Yokkaichi*
 Hiroshi Yoshida, *Tokyo*
 Norimasa Yoshida, *Kyoto*
 Akihito Nagahara, *Tokyo*
 Hiroaki Takeuchi, *Kochi*
 Keiji Ogura, *Tokyo*
 Kotaro Miyake, *Tokushima*
 Mitsunori Yamakawa, *Yamagata*
 Naoaki Sakata, *Sendai*
 Naoya Kato, *Tokyo*
 Satoshi Mamori, *Hyogo*
 Shogo Kikuchi, *Aichi*
 Shoichi Sumi, *Kyoto*
 Susumu Ikehara, *Osaka*
 Taketo Yamaguchi, *Chiba*
 Tokihiko Sawada, *Tochigi*
 Tomoharu Yoshizumi, *Fukuoka*
 Toshiyuki Ishiwata, *Tokyo*
 Yasuhiro Fujino, *Akashi*
 Yasuhiro Koga, *Isehara city*
 Yoshihisa Takahashi, *Tokyo*
 Yoshitaka Takuma, *Okayama*
 Yutaka Yata, *Maebashi-city*
 Itaru Endo, *Yokohama*
 Kazuo Chijiwa, *Miyazaki*
 Kouhei Fukushima, *Sendai*
 Masahiro Iizuka, *Akita*
 Mitsuyoshi Urashima, *Tokyo*
 Munechika Enjoji, *Fukuoka*
 Takashi Kojima, *Sapporo*
 Takumi Kawaguchi, *Kurume*
 Yoshiyuki Ueno, *Sendai*
 Yuichiro Eguchi, *Saga*
 Akihiro Tamori, *Osaka*
 Atsushi Masamune, *Sendai*
 Atsushi Tanaka, *Tokyo*
 Hitoshi Tsuda, *Tokyo*
 Takashi Kobayashi, *Tokyo*
 Akimasa Nakao, *Nagoya*
 Hiroyuki Uehara, *Osaka*
 Masahito Uemura, *Kashihara*
 Satoshi Tanno, *Hokkaido*
 Toshinari Takamura, *Kanazawa*
 Yohei Kida, *Kainan*

Kenji Okajima, *Nagoya*
 Masanori Hatakeyama, *Tokyo*
 Satoru Kakizaki, *Gunma*
 Shuhei Nishiguchi, *Hyogo*
 Yuichi Yoshida, *Osaka*
 Manabu Morimoto, *Japan*
 Mototsugu Kato, *Sapporo*
 Naoki Ishii, *Tokyo*
 Noriko Nakajima, *Tokyo*
 Nobuhiro Ohkohchi, *Tsukuba*
 Takanori Kanai, *Tokyo*
 Kenichi Goda, *Tokyo*
 Mitsugi Shimoda, *Mibu*
 Zenichi Morise, *Nagoya*
 Hitoshi Yoshiji, *Kashihara*
 Takahiro Nakazawa, *Nagoya*
 Utaroh Motosugi, *Yamanashi*
 Nobuyuki Matsuhashi, *Tokyo*
 Yasuhiro Kodera, *Nagoya*
 Takayoshi Ito, *Tokyo*
 Yasuhiro Tanaka, *Nagoya*
 Haruhiko Sugimura, *Hamamatsu*
 Hiroki Yamaue, *Wakayama*
 Masao Ichinose, *Wakayama*
 Takaaki Arigami, *Kagoshima*
 Nobuhiro Zaima, *Nara*
 Naoki Tanaka, *Matsumoto*
 Satoru Motoyama, *Akita*
 Tomoyuki Shibata, *Toyoake*
 Tatsuya Ide, *Kurume*
 Tsutomu Fujii, *Nagoya*
 Osamu Kanauchi, *Tokyo*
 Atsushi Irisawa, *Aizuwakamatsu*
 Hikaru Nagahara, *Tokyo*
 Keiji Hanada, *Onomichi*
 Keiichi Mitsuyama, *Fukuoka*
 Shin Maeda, *Yokohama*
 Takuya Watanabe, *Niigata*
 Toshihiro Mitaka, *Sapporo*
 Yoshiki Murakami, *Kyoto*
 Tadashi Shimoyama, *Hirosaki*



Jordan

Ismail Matalka, *Irbid*
 Khaled Jadallah, *Irbid*



Kuwait

Islam Khan, *Safat*



Lebanon

Bassam N Abboud, *Beirut*
 Rami Moucari, *Beirut*
 Ala I Sharara, *Beirut*
 Rita Slim, *Beirut*



Lithuania

Giedrius Barauskas, *Kaunas*
 Limas Kupcinskas, *Kaunas*



Malaysia

Andrew Seng Boon Chua, *Ipoh*



Mexico

Saúl Villa-Trevio, *México*
 Omar Vergara-Fernandez, *Tlalpan*
 Diego Garcia-Compean, *Monterrey*
 Arturo Panduro, *Jalisco*
 Miguel Angel Mercado, *Distrito Federal*
 Richard A Awad, *Mexico*
 Aldo Torre Delgadillo, *México*
 Paulino Martínez Hernández Magro, *Celaya*
 Carlos A Aguilar-Salinas, *Mexico*
 Jesus K Yamamoto-Furusho, *Mexico*



Morocco

Samir Ahboucha, *Marrakech*



Moldova

Igor Mishin, *Kishinev*



Netherlands

Ulrich Beuers, *Amsterdam*
 Albert Frederik Pull ter Gunne, *Tilburg*
 Jantine van Baal, *Heidelberglaan*
 Wendy Wilhelmina Johanna de Leng, *Utrecht*
 Gerrit A Meijer, *Amsterdam*
 Lee Bouwman, *Leiden*
 J Bart A Crusius, *Amsterdam*
 Frank Hoentjen, *Haarlem*
 Servaas Morré, *Amsterdam*
 Chris JJ Mulder, *Amsterdam*
 Paul E Sijens, *Groningen*
 Karel van Erpecum, *Utrecht*
 BW Marcel Spanier, *Arnhem*
 Misha Luyer, *Sittard*
 Pieter JF de Jonge, *Rotterdam*
 Robert Christiaan Verdonk, *Groningen*
 John Plukker, *Groningen*
 Maarten Tushuizen, *Amsterdam*
 Wouter de Herder, *Rotterdam*
 Erwin G Zoetendal, *Wageningen*
 Robert J de Knegt, *Rotterdam*
 Albert J Bredenoord, *Nieuwegein*
 Annemarie de Vries, *Rotterdam*
 Astrid van der Velde, *The Hague*
 Lodewijk AA Brosens, *Utrecht*
 James CH Hardwick, *Leiden*
 Loes van Keimpema, *Nijmegen*
 WJ de Jonge, *Amsterdam*
 Zuzana Zelinkova, *Rotterdam*
 LN van Steenberg, *Eindhoven*
 Frank G Schaap, *Amsterdam*
 Jeroen Maljaars, *Maastricht*



New Zealand

Andrew S Day, *Christchurch*
 Max S Petrov, *Auckland*



Norway

Espen Melum, *Oslo*
 Trine Olsen, *Tromsø*
 Eyvind J Paulssen, *Tromsø*

Rasmus Goll, *Tromsø*
Asle W Medhus, *Oslo*
Jon Arne Søreide, *Stavanger*
Kjetil Søreide, *Stavanger*
Reidar Fossmark, *Trondheim*
Trond Peder Flaten, *Trondheim*
Olav Dalgard, *Oslo*
Ole Høie, *Arendal*
Magdy El-Salhy, *Stord*
Jørgen Valeur, *Oslo*



Pakistan

Shahab Abid, *Karachi*
Syed MW Jafri, *Karachi*



Poland

Beata Jolanta Jabłońska, *Katowice*
Halina Cichoż-Lach, *Lublin*
Tomasz Brzozowski, *Cracow*
Hanna Gregorek, *Warsaw*
Marek Hartleb, *Katowice*
Stanislaw J Konturek, *Krakow*
Andrzej Dabrowski, *Bialystok*
Jan Kulig, *Kraków*
Julian Swierczynski, *Gdansk*
Marek Bebenek, *Wroclaw*
Dariusz M Lebensztejn, *Bialystok*



Portugal

Ricardo Marcos, *Porto*
Guida Portela-Gomes, *Estoril*
Ana Isabel Lopes, *Lisboa Codex*
Raquel Almeida, *Porto*
Rui Tato Marinho, *Lisbon*
Ceú Figueiredo, *Porto*



Romania

Dan L Dumitrascu, *Cluj*
Adrian Saftoiu, *Craiova*
Andrada Seicean, *Cluj-Napoca*
Anca Trifan, *Iasi*



Russia

Vasilii I Reshetnyak, *Moscow*



Saudi Arabia

Ibrahim A Al Mofleh, *Riyadh*
Abdul-Wahed Meshikhes, *Qatif*
Faisal Sanai, *Riyadh*



Serbia

Tamara M Alempijevic, *Belgrade*
Dusan M Jovanovic, *Sremska Kamenica*
Zoran Krivokapic, *Belgrade*



Singapore

Brian Kim Poh Goh, *Singapore*

Khek-Yu Ho, *Singapore*
Fock Kwong Ming, *Singapore*
Francis Seow-Choen, *Singapore*
Kok Sun Ho, *Singapore*
Nagarajan Perumal, *Singapore*
Kong Weng Eu, *Singapore*
Madhav Bhatia, *Singapore*
London Lucien Ooi, *Singapore*
Wei Ning Chen, *Singapore*
Richie Soong, *Singapore*
Kok Ann Gwee, *Singapore*



Slovenia

Matjaz Homan, *Ljubljana*



South Africa

Rosemary Joyce Burnett, *Pretoria*
Michael Kew, *Cape Town*
Roland Ndip, *Alice*



South Korea

Byung Chul Yoo, *Seoul*
Jae J Kim, *Seoul*
Jin-Hong Kim, *Suwon*
Marie Yeo, *Suwon*
Jeong Min Lee, *Seoul*
Eun-Yi Moon, *Seoul*
Joong-Won Park, *Goyang*
Hoon Jai Chun, *Seoul*
Myung-Gyu Choi, *Seoul*
Sang Kil Lee, *Seoul*
Sang Yeoup Lee, *Gyeongsangnam-do*
Won Ho Kim, *Seoul*
Dae-Yeul Yu, *Daejeon*
Kim Donghee, *Seoul*
Sang Geon Kim, *Seoul*
Sun Pyo Hong, *Geonggi-do*
Sung-Gil Chi, *Seoul*
Yeun-Jun Chung, *Seoul*
Ki-Baik Hahm, *Incheon*
Ji Kon Ryu, *Seoul*
Kyu Taek Lee, *Seoul*
Yong Chan Lee, *Seoul*
Seong Gyu Hwang, *Seongnam*
Seung Woon Paik, *Seoul*
Sung Kim, *Seoul*
Hong Joo Kim, *Seoul*
Hyoung-Chul Oh, *Seoul*
Nayoung Kim, *Seongnam-si*
Sang Hoon Ahn, *Seoul*
Seon Hahn Kim, *Seoul*
Si Young Song, *Seoul*
Young-Hwa Chung, *Seoul*
Hyo-Cheol Kim, *Seoul*
Kwang Jae Lee, *Swon*
Sang Min Park, *Seoul*
Young Chul Kim, *Seoul*
Do Hyun Park, *Seoul*
Dae Won Jun, *Seoul*
Dong Wan Seo, *Seoul*
Soon-Sun Hong, *Incheon*
Hoguen Kim, *Seoul*
Ho-Young Song, *Seoul*
Joo-Ho Lee, *Seoul*
Jung Eun Lee, *Seoul*
Jong H Moon, *Bucheon*



Spain

Eva Vaquero, *Barcelona*
Andres Cardenas, *Barcelona*
Laureano Fernández-Cruz, *Barcelona*
Antoni Farré, *Spain*
Maria-Angeles Aller, *Madrid*
Raul J Andrade, *Málaga*
Fernando Azpiroz, *Barcelona*
Josep M Bordas, *Barcelona*
Antoni Castells, *Barcelona*
Vicente Felipo, *Valencia*
Isabel Fabregat, *Barcelona*
Angel Lanas, *Zaragoza*
Juan-Ramón Larrubia, *Guadalajara*
María IT López, *Jaén*
Jesús M Prieto, *Pamplona*
Mireia Miquel, *Sabadell*
Ramon Bataller, *Barcelona*
Fernando J Corrales, *Pamplona*
Julio Mayol, *Madrid*
Matias A Avila, *Pamplona*
Juan Macías, *Seville*
Juan Carlos Laguna Egea, *Barcelona*
Juli Busquets, *Barcelona*
Belén Beltrán, *Valencia*
José Manuel Martín-Villa, *Madrid*
Lisardo Boscá, *Madrid*
Luis Grande, *Barcelona*
Pedro Lorenzo Majano Rodriguez, *Madrid*
Adolfo Benages, *Valencia*
Domínguez-Muñoz JE, *Santiago de Compostela*
Gloria González Aseguinolaza, *Navarra*
Javier Martin, *Granada*
Luis Bujanda, *San Sebastián*
Matilde Bustos, *Pamplona*
Luis Aparisi, *Valencia*
José Julián calvo Andrés, *Salamanca*
Benito Velayos, *Valladolid*
Javier Gonzalez-Gallego, *León*
Ruben Ciria, *Córdoba*
Francisco Rodriguez-Frias, *Barcelona*
Manuel Romero-Gómez, *Sevilla*
Albert Parés, *Barcelona*
Joan Roselló-Catafau, *Barcelona*



Sri Lanka

Arjuna De Silva, *Colombo*



Sweden

Stefan G Pierzynowski, *Lund*
Hanns-Ulrich Marschall, *Stockholm*
Lars A Pahlman, *Uppsala*
Helena Nordenstedt, *Stockholm*
Bobby Tingstedt, *Lund*
Evangelos Kalaitzakis, *Gothenburg*
Lars Erik Agréus, *Huddinge*
Annika Lindblom, *Stockholm*
Roland Andersson, *Lund*
Zongli Zheng, *Stockholm*
Mauro D'Amato, *Huddinge*
Greger Lindberg, *Stockholm*
Pär Erik Myrelid, *Linköping*
Sara Lindén, *Göteborg*

Sara Regné, *Malmö*
Åke Nilsson, *Lund*



Switzerland

Jean L Frossard, *Geneva*
Andreas Geier, *Zürich*
Bruno Stieger, *Zürich*
Pascal Gervaz, *Geneva*
Paul M Schneider, *Zurich*
Felix Stickel, *Berne*
Fabrizio Montecucco, *Geneva*
Inti Zlobec, *Basel*
Michelangelo Foti, *Geneva*
Pascal Bucher, *Geneva*
Andrea De Gottardi, *Berne*
Christian Toso, *Geneva*



Thailand

Weekitt Kittisupamongkol, *Bangkok*



Trinidad and Tobago

Shivananda Nayak, *Mount Hope*



Turkey

Tarkan Karakan, *Ankara*
Yusuf Bayraktar, *Ankara*
Ahmet Tekin, *Mersin*
Aydin Karabacakoglu, *Konya*
Osman C Ozdogan, *Istanbul*
Özlem Yilmaz, *Izmir*
Bülent Salman, *Ankara*
Can GONEN, *Kutahya*
Cuneyt Kayaalp, *Malatya*
Ekmel Tezel, *Ankara*
Eren Ersoy, *Ankara*
Hayrullah Derici, *Balikesir*
Mehmet Refik Mas, *Etlik-Ankara*
Sinan Akay, *Tekirdag*
A Mithat Bozdayi, *Ankara*
Metin Basaranoglu, *Istanbul*
Mesut Tez, *Ankara*
Orhan Sezgin, *Mersin*
Mukaddes Esrefoglu, *Malatya*
Ilker Tasci, *Ankara*
Kemal Kismet, *Ankara*
Selin Kapan, *Istanbul*
Seyfettin Köklü, *Ankara*
Murat Sayan, *Kocaeli*
Sabahattin Kaymakoglu, *Istanbul*
Yucel Ustundag, *Zonguldak*
Can Gonen, *Istanbul*
Yusuf Yilmaz, *Istanbul*
Müge Tecder-Ünal, *Ankara*
İlhami Yüksel, *Ankara*



United Arab Emirates

Fikri M Abu-Zidan, *Al-Ain*
Sherif M Karam, *Al-Ain*



United Kingdom

Anastasios Koulaouzidis, *Edinburgh*
Sylvia LF Pender, *Southampton*
Hong-Xiang Liu, *Cambridge*
William Dickey, *London*
Simon D Taylor-Robinson, *London*
James Neuberger, *Birmingham*
Frank I Tovey, *London*
Kevin Robertson, *Glasgow*
Chew Thean Soon, *Manchester*
Geoffrey Burnstock, *London*
Vamsi R Velchuru, *United Kingdom*
Simon Afford, *Birmingham*
Navneet K Ahluwalia, *Stockport*
Lesley A Anderson, *Belfast*
Anthony TR Axon, *Leeds*
Jim D Bell, *London*
Alastair D Burt, *Newcastle*
Tatjana Crnogorac-Jurcevic, *London*
Daniel R Gaya, *Edinburgh*
William Greenhalf, *Liverpool*
Indra N Guha, *Southampton*
Stefan G Hübscher, *Birmingham*
Robin Hughes, *London*
Pali Hungin, *Stockton*
Janusz AZ Jankowski, *Oxford*
Peter Karayiannis, *London*
Patricia F Lalor, *Birmingham*
Giorgina Mieli-Vergani, *London*
D Mark Pritchard, *Liverpool*
Marco Senzolo, *Padova*
Roger Williams, *London*
M H Ahmed, *Southampton*
Christos Paraskeva, *Bristol*
Emad M El-Omar, *Aberdeen*
A M El-Tawil, *Birmingham*
Anne McCune, *Bristol*
Charles B Ferguson, *Belfast*
Chin Wee Ang, *Liverpool*
Clement W Imrie, *Glasgow*
Dileep N Lobo, *Nottingham*
Graham MacKay, *Glasgow*
Guy Fairbairn Nash, *Poole*
Ian Lindsey, *Oxford*
Jason CB Goh, *Birmingham*
Jeremy FL Cobbold, *London*
Julian RF Walters, *London*
Jamie Murphy, *London*
John Beynon, *Swansea*
John B Schofield, *Kent*
Anil George, *London*
Aravind Suppiah, *East Yorkshire*
Basil Ammori, *Salford*
Catherine Walter, *Cheltenham*
Chris Briggs, *Sheffield*
Jeff Butterworth, *Shrewsbury*
Nawfal Hussein, *Nottingham*
Patrick O'Dwyer, *Glasgow*
Rob Glynne-Jones, *Northwood*
Sharad Karandikar, *Birmingham*
Venkatesh Shanmugam, *Derby*
Yeng S Ang, *Wigan*
Alberto Quaglia, *London*
Andrew Fowell, *Southampton*
Gianpiero Gravante, *Leicester*
Piers Gatenby, *London*
Kondragunta Rajendra Prasad, *Leeds*

Sunil Dolwani, *Cardiff*
Andrew McCulloch Veitch, *Wolverhampton*
Brian Green, *Belfast*
Noriko Suzuki, *Middlesex*
Richard Parker, *North Staffordshire*
Shahid A Khan, *London*
Akhilesh B Reddy, *Cambridge*
Jean E Crabtree, *Leeds*
John S Leeds, *Sheffield*
Paul Sharp, *London*
Sumita Verma, *Brighton*
Thamara Perera, *Birmingham*
Donald Campbell McMillan, *Glasgow*
Kathleen B Bamford, *London*
Helen Coleman, *Belfast*
Eyad Elkord, *Manchester*
Mohammad Ilyas, *Nottingham*
Simon R Carding, *Norwich*
Ian Chau, *Sutton*
Claudio Nicoletti, *Norwich*
Hendrik-Tobias Arkenau, *London*
Muhammad Imran Aslam, *Leicester*
Giuseppe Orlando, *Oxford*
John S Leeds, *Aberdeen*
S Madhusudan, *Nottingham*
Amin Ibrahim Amin, *Dunfermline*
David C Hay, *Edinburgh*
Alan Burns, *London*



United States

Tauseef Ali, *Oklahoma City*
George Y Wu, *Farmington*
Josef E Fischer, *Boston*
Thomas Clancy, *Boston*
John Morton, *Stanford*
Luca Stocchi, *Cleveland*
Kevin Michael Reavis, *Orange*
Shiu-Ming Kuo, *Buffalo*
Gary R Lichtenstein, *Philadelphia*
Natalie J Torok, *Sacramento*
Scott A Waldman, *Philadelphia*
Georgios Papachristou, *Pittsburgh*
Carla W Brady, *Durham*
Robert CG Martin, *Louisville*
Eugene P Ceppia, *Durham*
Shashi Bala, *Worcester*
Imran Hassan, *Springfield*
Klaus Thaler, *Columbia*
Andreas M Kaiser, *Los Angeles*
Shawn D Safford, *Norfolk*
Massimo Raimondo, *Jacksonville*
Kazuaki Takabe, *Richmond VA*
Stephen M Kavic, *Baltimore*
T Clark Gamblin, *Pittsburgh*
BS Anand, *Houston*
Ananthanarayanan M, *New York*
Anthony J Bauer, *Pittsburgh*
Edmund J Bini, *New York*
Xian-Ming Chen, *Omaha*
Ramsey Chi-man Cheung, *Palo Alto*
Parimal Chowdhury, *Arkansas*
Mark J Czaja, *New York*
Conor P Delaney, *Cleveland*
Sharon DeMorrow, *Temple*
Bijan Eghtesad, *Cleveland*
Alessandro Fichera, *Chicago*
Glenn T Furuta, *Aurora*
Jean-Francois Geschwind, *Baltimore*

Shannon S Glaser, *Temple*
Ajay Goel, *Dallas*
James H Grendell, *New York*
Anna S Gukovskaya, *Los Angeles*
Jamal A Ibdah, *Columbia*
Atif Iqbal, *Omaha*
Hajime Isomoto, *Rochester*
Hartmut Jaeschke, *Kansas*
Leonard R Johnson, *Memphis*
Rashmi Kaul, *Tulsa*
Ali Keshavarzian, *Chicago*
Miran Kim, *Providence*
Burton I Korelitz, *New York*
Richard A Kozarek, *Seattle*
Alyssa M Krasinskas, *Pittsburgh*
Ming Li, *New Orleans*
Zhiping Li, *Baltimore*
Chen Liu, *Gainesville*
Michael R Lucey, *Madison*
James D Luketich, *Pittsburgh*
Patrick M Lynch, *Houston*
Willis C Maddrey, *Dallas*
Mercedes Susan Mandell, *Aurora*
Wendy M Mars, *Pittsburgh*
Laura E Matarese, *Pittsburgh*
Lynne V McFarland, *Washington*
Stephan Menne, *New York*
Didier Merlin, *Atlanta*
George Michalopoulos, *Pittsburgh*
James M Millis, *Chicago*
Pramod K Mistry, *New Haven*
Emiko Mizoguchi, *Boston*
Peter L Moses, *Burlington*
Masaki Nagaya, *Boston*
Robert D Odze, *Boston*
Stephen JD O'Keefe, *Pittsburgh*
Zhiheng Pei, *New York*
Raymund R Razonable, *Minnesota*
Basil Rigas, *New York*
Richard A Rippe, *Chapel Hill*
Philip Rosenthal, *San Francisco*
Stuart Sherman, *Indianapolis*
Christina Surawicz, *Seattle*
Wing-Kin Syn, *Durham*
Yvette Taché, *Los Angeles*
K-M Tchou-Wong, *New York*
George Triadafilopoulos, *Stanford*
Chung-Jyi Tsai, *Lexington*
Andrew Ukleja, *Florida*
Arnold Wald, *Wisconsin*
Irving Waxman, *Chicago*
Steven D Wexner, *Weston*
Jackie Wood, *Ohio*
Jian Wu, *Sacramento*
Zobair M Younossi, *Virginia*
Liqing Yu, *Winston-Salem*
Ruben Zamora, *Pittsburgh*
Michael E Zenilman, *New York*
Michael A Zimmerman, *Colorado*
Beat Schnüriger, *California*
Clifford S Cho, *Madison*
R Mark Ghobrial, *Texas*
Anthony T Yeung, *Philadelphia*
Chang Kim, *West Lafayette*
Balamurugan N Appakalai, *Minneapolis*
Aejaz Nasir, *Tampa*
Ashkan Farhadi, *Irvine*
Kevin E Behrns, *Gainesville*

Joseph J Cullen, *Iowa City*
David J McGee, *Shreveport*
Anthony J Demetris, *Pittsburgh*
Dimitrios V Avgerinos, *New York*
Dong-Hui Li, *Houston*
Eric S Hungness, *Chicago*
Giuseppe Orlando, *Winston Salem*
Hai-Yong Han, *Phoenix*
Huanbiao Mo, *Denton*
Jong Park, *Tampa*
Justin MM Cates, *Nashville*
Charles P Heise, *Madison*
Craig D Logsdon, *Houston*
Ece A Mutlu, *Chicago*
Jessica A Davila, *Houston*
Rabih M Salloum, *Rochester*
Amir Maqbul Khan, *Marshall*
Bruce E Sands, *Boston*
Chakshu Gupta, *Saint Joseph*
Ricardo Alberto Cruciani, *New York*
Mariana D Dabeva, *Bronx*
Edward L Bradley III, *Sarasota*
Martín E Fernández-Zapico, *Rochester*
Henry J Binder, *New Haven*
John R Grider, *Richmond*
Ronnie Fass, *Tucson*
Dinesh Vyas, *Washington*
Wael El-Rifai, *Nashville*
Craig J McClain, *Louisville*
Christopher Mantyh, *Durham*
Daniel S Straus, *Riverside*
David A Brenner, *San Diego*
Eileen F Grady, *San Francisco*
Ekihiro Seki, *La Jolla*
Fang Yan, *Nashville*
Fritz Francois, *New York*
Giamila Fantuzzi, *Chicago*
Guang-Yin Xu, *Galveston*
Jianyuan Chai, *Long Beach*
JingXuan Kang, *Charlestown*
Le Shen, *Chicago*
Lin Zhang, *Pittsburgh*
Mitchell L Shiffman, *Richmond*
Douglas K Rex, *Indianapolis*
Bo Shen, *Cleveland*
Edward J Ciccio, *New York*
Jean S Wang, *Saint Louis*
Bao-Ting Zhu, *Kansas*
Tamir Miloh, *Phoenix*
Eric R Kallwitz, *Chicago*
Yujin Hoshida, *Cambridge*
C Chris Yun, *Atlanta*
Alan C Moss, *Boston*
Oliver Grundmann, *Gainesville*
Linda A Feagins, *Dallas*
Chanjuan Shi, *Nashville*
Xiaonan Han, *Cincinnati*
William R Brugge, *Boston*
Richard W McCallum, *El Paso*
Lisa Ganley-Leal, *Boston*
Lin-Feng Chen, *Urbana*
Elaine Y Lin, *New York*
Julian Abrams, *New York*
Arun Swaminath, *New York*
Huiping Zhou, *Richmond*
Korkut Uygun, *Boston*
Anupam Bishayee, *Signal Hill*
C Bart Rountree, *Hershey*

Avinash Kambadakone, *Boston*
Courtney W Houchen, *Oklahoma*
Joshua R Friedman, *Philadelphia*
Justin H Nguyen, *Jacksonville*
Sophoclis Alexopoulos, *Los Angeles*
Suryakanth R Gurudu, *Scottsdale*
Wei Jia, *Kannapolis*
Yoon-Young Jang, *Baltimore*
Ourania M Andrisani, *West Lafayette*
Roderick M Quiros, *Bethlehem*
Timothy R Koch, *Washington*
Adam S Cheifetz, *Boston*
Lifang Hou, *Chicago*
Thiru vengadam Muniraj, *Pittsburgh*
Dhiraj Yadav, *Pittsburgh*
Ying Gao, *Rockville*
John F Gibbs, *Buffalo*
Aaron Vinik, *Norfolk*
Charles Thomas, *Oregon*
Robert Jensen, *Bethesda*
John W Wiley, *Ann Arbor*
Jonathan Strosberg, *Tampa*
Randeep Singh Kashyap, *New York*
Kaye M Reid Lombardo, *Rochester*
Lygia Stewart, *San Francisco*
Martin D Zielinski, *Rochester*
Matthew James Schuchert, *Pittsburgh*
Michelle Lai, *Boston*
Million Mulugeta, *Los Angeles*
Patricia Sylla, *Boston*
Pete Muscarella, *Columbus*
Raul J Rosenthal, *Weston*
Robert V Rega, *Dallas*
Roberto Bergamaschi, *New York*
Ronald S Chamberlain, *Livingston*
Alexander S Rosemurgy, *Tampa*
Run Yu, *Los Angeles*
Samuel B Ho, *San Diego*
Sami R Achem, *Florida*
Sandeep Mukherjee, *Omaha*
Santhi Swaroop Vege, *Rochester*
Scott Steele, *Fort Lewis*
Steven Hochwald, *Gainesville*
Udayakumar Navaneethan, *Cincinnati*
Radha Krishna Yellapu, *New York*
Rupjyoti Talukdar, *Rochester*
Shi-Ying Cai, *New Haven*
Thérèse Tuohy, *Salt Lake City*
Tor C Savidge, *Galveston*
William R Parker, *Durham*
Xiaofa Qin, *Newark*
Zhang-Xu Liu, *Los Angeles*
Adeel A Butt, *Pittsburgh*
Dean Y Kim, *Detroit*
Denesh Chitkara, *East Brunswick*
Mohamad A Eloubeidi, *Alabama*
JiPing Wang, *Boston*
Oscar Joe Hines, *Los Angeles*
John Y Kao, *Michigan*
Jon C Gould, *Madison*
Kirk Ludwig, *Wisconsin*
Mansour A Parsi, *Cleveland*
Perry Shen, *Winston-Salem*
Piero Marco Fisichella, *Maywood*
Marco Giuseppe Patti, *Chicago*
Michael Leitman, *New York*
Parviz M Pour, *Omaha*
Florencia Georgina Que, *Rochester*

Richard Hu, *Los Angeles*
Robert E Schoen, *Pittsburgh*
Valentina Medici, *Sacramento*
Wojciech Blonski, *Philadelphia*
Yoram Elitsur, *Huntington*
Yuan-Ping Han, *Los Angeles*
Grigoriy E Gurvits, *New York*
Robert C Moesinger, *Ogden*
Mark Bloomston, *Columbus*
Bronislaw L Slomiany, *Newark*
Laurie DeLeve, *Los Angeles*
Michel M Murr, *Tampa*

John Marshall, *Columbia*
Wilfred M Weinstein, *Los Angeles*
Jonathan D Kaunitz, *Los Angeles*
Josh Korzenik, *Boston*
Kareem M Abu-Elmagd, *Pittsburgh*
Michael L Schilsky, *New Haven*
John David Christein, *Birmingham*
Mark A Zern, *Sacramento*
Ana J Coito, *Los Angeles*
Golo Ahlenstiel, *Bethesda*
Smruti R Mohanty, *Chicago*
Victor E Reyes, *Galveston*

CS Pitchumoni, *New Brunswick*
Yoshio Yamaoka, *Houston*
Sukru H Emre, *New Haven*
Branko Stefanovic, *Tallahassee*
Jack R Wands, *Providence*
Wen Xie, *Pittsburgh*
Robert Todd Striker, *Madison*
Shivendra Shukla, *Columbia*
Laura E Nagy, *Cleveland*
Fei Chen, *Morgantown*
Kusum K Kharbanda, *Omaha*
Pal Pacher, *Rockville*

Contents

Weekly Volume 18 Number 5 February 7, 2012

- | | | |
|-------------------------|-----|--|
| EDITORIAL | 393 | Estrogen, male dominance and esophageal adenocarcinoma: Is there a link?
<i>Yang H, Sukocheva OA, Hussey DJ, Watson DI</i> |
| TOPIC HIGHLIGHT | 401 | Update of endoscopy in liver disease: More than just treating varices
<i>Krystallis C, Masterton GS, Hayes PC, Plevris JN</i> |
| REVIEW | 412 | Role of <i>ATG16L</i> , <i>NOD2</i> and <i>IL23R</i> in Crohn's disease pathogenesis
<i>Naser SA, Arce M, Khaja A, Fernandez M, Naser N, Elwasila S, Thanigachalam S</i> |
| ORIGINAL ARTICLE | 425 | Effect of <i>Helicobacter pylori cdrA</i> on interleukin-8 secretions and nuclear factor kappa B activation
<i>Takeuchi H, Zhang YN, Israel DA, Peek Jr RM, Kamioka M, Yanai H, Morimoto N, Sugiura T</i> |
| | 435 | <i>A Macaca mulatta</i> model of fulminant hepatic failure
<i>Zhou P, Xia J, Guo G, Huang ZX, Lu Q, Li L, Li HX, Shi YJ, Bu H</i> |
| BRIEF ARTICLE | 445 | Chronic fatigue is associated with increased disease-related worries and concerns in inflammatory bowel disease
<i>Jelsness-Jørgensen LP, Bernklev T, Henriksen M, Torp R, Moum B</i> |
| | 453 | Pneumatosis cystoides intestinalis: A single center experience
<i>Arikanoglu Z, Aygen E, Camci C, Akbulut S, Basbug M, Dogru O, Cetinkaya Z, Kirkil C</i> |
| | 458 | Survival analysis of cholangiocarcinoma: A 10-year experience in Malaysia
<i>Yusoff AR, Abdul Razak MM, Yoong BK, Vijeyasingam R, Siti ZM</i> |
| | 466 | Thalidomide-based multidisciplinary treatment for patients with advanced hepatocellular carcinoma: A retrospective analysis
<i>Chen YY, Yen HH, Chou KC, Wu SS</i> |
| | 472 | A pharmacodynamic model of portal hypertension in isolated perfused rat liver
<i>Zhang T, Xu XY, Zhou H, Zhao X, Song M, Zhang TT, Yin H, Li T, Li PT, Cai DY</i> |

- 479 Safety assessment of *Bifidobacterium longum* JDM301 based on complete genome sequences

Wei YX, Zhang ZY, Liu C, Malakar PK, Guo XK

LETTERS TO THE EDITOR 489

- Potential implications of *Helicobacter pylori*-related neutrophil-activating protein
Kountouras J, Zavos C, Deretzi G, Gavalas E, Chatzopoulos D, Katsinelos P, Tsiaousi E, Gagalis S, Polyzos SA, Venizelos I

Contents

World Journal of Gastroenterology
Volume 18 Number 5 February 7, 2012

ACKNOWLEDGMENTS I Acknowledgments to reviewers of *World Journal of Gastroenterology*

APPENDIX I Meetings
I-VI Instructions to authors

ABOUT COVER Editor-in-Chief of *World Journal of Gastroenterology*, Matt D Rutter, MBBS, MD, FRCP, Consultant Gastroenterologist, Senior Lecturer, Director, Tees Bowel Cancer Screening Centre, University Hospital of North Tees, Durham University, Stockton-on-Tees, Cleveland TS19 8PE, United Kingdom

AIM AND SCOPE *World Journal of Gastroenterology* (*World J Gastroenterol*, *WJG*, print ISSN 1007-9327, DOI: 10.3748) is a weekly, open-access, peer-reviewed journal supported by an editorial board of 1361 experts in gastroenterology and hepatology from 64 countries.
The major task of *WJG* is to report rapidly the most recent results in basic and clinical research on esophageal, gastrointestinal, liver, pancreas and biliary tract diseases, *Helicobacter pylori*, endoscopy and gastrointestinal surgery, including: gastroesophageal reflux disease, gastrointestinal bleeding, infection and tumors; gastric and duodenal disorders; intestinal inflammation, microflora and immunity; celiac disease, dyspepsia and nutrition; viral hepatitis, portal hypertension, liver fibrosis, liver cirrhosis, liver transplantation, and metabolic liver disease; molecular and cell biology; geriatric and pediatric gastroenterology; diagnosis and screening, imaging and advanced technology.

FLYLEAF I-IX Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: Yuan Zhou
Responsible Electronic Editor: Jun-Yao Li
Proofing Editor-in-Chief: Lian-Sheng Ma

Responsible Science Editor: Xing Wu
Proofing Editorial Office Director: Jian-Xia Cheng

NAME OF JOURNAL
World Journal of Gastroenterology

ISSN AND EISSN
ISSN 1007-9327 (print)
ISSN 2219-2840 (online)

LAUNCH DATE
October 1, 1995

FREQUENCY
Weekly

RESPONSIBLE INSTITUTION
Department of Science and Technology of Shanxi Province

SPONSOR
Taiyuan Research and Treatment Center for Digestive Diseases, 77 Shuangta Xijie, Taiyuan 030001, Shanxi Province, China

EDITING
Editorial Board of *World Journal of Gastroenterology*
Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China
Telephone: +86-10-59080039
Fax: +86-10-85381893
E-mail: wjg@wjgnet.com
http://www.wjgnet.com

EDITOR-IN-CHIEF
Ferruccio Bonino, MD, PhD, Professor of Gastroenterology, Director of Liver and Digestive Disease Division, Department of Internal Medicine, Uni-

versity of Pisa, Director of General Medicine 2 Unit University Hospital of Pisa, Via Roma 67, 56124 Pisa, Italy

Myung-Hwan Kim, MD, PhD, Professor, Head, Department of Gastroenterology, Director, Center for Biliary Diseases, University of Ulsan College of Medicine, Asan Medical Center, 388-1 Pungnap-2dong, Songpa-gu, Seoul 138-736, South Korea

Kjell Öberg, MD, PhD, Professor, Department of Endocrine Oncology, Uppsala University Hospital, SE-751 85 Uppsala, Sweden

Matt D Rutter, MBBS, MD, FRCP, Consultant Gastroenterologist, Senior Lecturer, Director, Tees Bowel Cancer Screening Centre, University Hospital of North Tees, Durham University, Stockton-on-Tees, Cleveland TS19 8PE, United Kingdom

Andrzej S Tarnawski, MD, PhD, DSc (Med), Professor of Medicine, Chief of Gastroenterology, VA Long Beach Health Care System, University of California, Irvine, CA, 5901 E. Seventh Str., Long Beach, CA 90822, United States

EDITORIAL OFFICE
Jian-Xia Cheng, Director
World Journal of Gastroenterology
Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China
Telephone: +86-10-59080039
Fax: +86-10-85381893
E-mail: wjg@wjgnet.com
http://www.wjgnet.com

PUBLISHER
Baishideng Publishing Group Co., Limited
Room 1701, 17/F, Henan Building,
No.90 Jaffe Road, Wanchai, Hong Kong, China
Fax: +852-31158812
Telephone: +852-58042046
E-mail: bpg@baishideng.com
http://www.wjgnet.com

PRINT SUBSCRIPTION
RMB 300 Yuan for each issue, RMB 14400 Yuan for one year.

PUBLICATION DATE
February 7, 2012

COPYRIGHT
© 2012 Baishideng. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

INSTRUCTIONS TO AUTHORS
Full instructions are available online at http://www.wjgnet.com/1007-9327/g_info_20100315215714.htm

ONLINE SUBMISSION
<http://www.wjgnet.com/1007-9327office/>

Estrogen, male dominance and esophageal adenocarcinoma: Is there a link?

Huiqi Yang, Olga A Sukocheva, Damian J Hussey, David I Watson

Huiqi Yang, Olga A Sukocheva, Damian J Hussey, David I Watson, Department of Surgery, Flinders Medical Centre, Flinders University, Bedford Park, South Australia 5042, Australia
Olga A Sukocheva, Damian J Hussey, David I Watson, Flinders Centre for Cancer Prevention and Control, Flinders Medical Centre, Bedford Park, South Australia 5042, Australia
Author contributions: Yang H reviewed the related research papers and wrote this paper; Hussey DJ and Sukocheva OA critically reviewed the scientific sections and co-wrote the paper; Watson DI designed the outline of this review, edited the manuscript and critically reviewed its content.

Correspondence to: David I Watson, Professor, Department of Surgery, Flinders Medical Centre, Flinders University, Bedford Park, South Australia 5042,

Australia. david.watson@flinders.edu.au

Telephone: +61-8-82046086 Fax: +61-8-82046130

Received: May 28, 2011 Revised: August 11, 2011

Accepted: August 15, 2011

Published online: February 7, 2012

in the male gender bias in esophageal adenocarcinoma, but further studies are required.

© 2012 Baishideng. All rights reserved.

Key words: Estrogen; Estrogen receptors; Male dominance; Esophageal adenocarcinoma

Peer reviewer: Guy D Eslick, PhD, MmedSc (Clin Epi), MmedStat, Department of Medicine, The University of Sydney Nepean Hospital, Level 5, South Block, PO Box 63, Penrith, Sydney NSW 2751, Australia

Yang H, Sukocheva OA, Hussey DJ, Watson DI. Estrogen, male dominance and esophageal adenocarcinoma: Is there a link? *World J Gastroenterol* 2012; 18(5): 393-400 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/393.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.393>

Abstract

Esophageal adenocarcinoma is a cancer with poor prognosis, and its incidence has risen sharply over recent decades. Obesity is a major risk factor for developing this cancer and there is a clear male gender bias in the incidence that cannot be fully explained by known risk factors. It is possible that a difference in the expression of estrogen, or its signaling axes, may contribute to this gender bias. We undertook a comprehensive literature search and analyzed the available data regarding estrogen and estrogen receptor expression, and the possible sex-specific links with esophageal adenocarcinoma development. Potentially relevant associations between visceral *vs* subcutaneous fat deposition and estrogen expression, and the effect of crosstalk between estrogen and leptin signaling were identified. We also found limited studies suggesting a role for estrogen receptor β expression in esophageal adenocarcinoma development. The current literature supports speculation on an etiological role for estrogen

INTRODUCTION

Esophageal carcinoma is the eighth most common cancer worldwide, and over the last three decades its incidence has risen significantly in all Western countries^[1-4]. This change is entirely due to an increase in the adenocarcinoma subtype, and this has predominantly occurred in males^[2,5-9]. Recent Western experiences report that the male:female ratio for patients undergoing esophagectomy for adenocarcinoma now exceeds 8:1. However, the identified risk factors for esophageal adenocarcinoma, including gastroesophageal reflux disease, Barrett's esophagus, obesity, alcohol, and tobacco consumption cannot adequately explain this profound gender difference. The dramatic gender difference for esophageal adenocarcinoma suggests there should be a gender-related mechanism underpinning this phenomenon. Estrogens, the primary female sex hormones, are mechanistically linked to aspects of cancer risk and cancer development. Therefore it seems reasonable to consider that estrogens might

contribute towards the gender difference for esophageal adenocarcinoma.

A link between estrogen-activated signaling and carcinogenesis in many organs, including mammary glands^[10], ovaries and colon^[11] has been clearly defined, although it is unclear whether a similar connection exists for the esophagus, and esophageal adenocarcinoma in particular. Additionally, estrogen is actively involved in the regulation of metabolism in adipose tissues^[12], and it can be synthesized locally by activated aromatase in adipocytes in both men and women^[13-15]. Involvement of estrogen signaling in regulation of adipose tissue metabolism indicates a possible connection between the effects of estrogen and male obesity - one of the main risk factors for esophageal adenocarcinoma. Given the established regulatory role for estrogen in carcinogenesis and metabolic homeostasis for other cancers, and the strong gender differences for the incidence of esophageal adenocarcinoma, it is plausible to suggest that the estrogen signaling network is involved in the progression of this cancer, and an understanding of estrogen and estrogen receptor (ER) roles in the regulation of carcinogenesis, and how this might be relevant in the esophagus, could provide a basis for developing either preventive measures or new treatments.

In this paper we review potential links between estrogen signaling and esophageal adenocarcinoma, to determine whether this might contribute to the dominance of esophageal adenocarcinoma in males. Literature pertinent to gender specific differences in estrogen synthesis, estrogen-regulated carcinogenesis, specific differences between ERs and signaling in cancer cells, and available information about estrogen signaling in esophageal adenocarcinoma is reviewed.

ESTROGEN IN WOMEN AND MEN: AGE RELATED CHANGES

In premenopausal women the ovaries are the principal source of estrogen^[16]. Serum estradiol concentration is much higher in premenopausal women, compared to men, but decreases substantially after the menopause, and ultimately becomes lower than in elderly men^[17]. Mass spectrometry has shown that average levels of serum estradiol in elderly men are approximately 73 pmol/L, whereas levels in postmenopausal women are markedly lower (about 15 pmol/L)^[17].

When the ovaries cease to produce estrogens in postmenopausal women the main characteristics of estradiol function change, and it is produced in extragonadal sites, and acts locally at these sites as a paracrine or intracrine factor^[16-20]. These sites are similar in men and postmenopausal women and include the mesenchymal cells of adipose tissue^[13], osteoblasts and chondrocytes of bone^[21], the vascular endothelium and aortic smooth muscle cells^[22], and numerous sites in the brain^[16,17,23].

Importantly, in men and postmenopausal women, circulating estrogens are not the main drivers of estro-

gen action, but locally produced estrogens originating in extragonadal sites are responsible for the majority of paracrine and intracrine effects of these hormones^[20]. The total amount of estrogen synthesized by these extragonadal sites may be small, but the local tissue concentrations achieved are probably high and exert biological influence locally. This might impact on tumor biology. For example, it has been determined that the concentration of estradiol present in breast tumors in postmenopausal women is at least 20-fold higher than in the plasma. Aromatase inhibitor therapy is associated with a major decrease in intratumoral concentrations of estradiol and estrone and loss of intratumoral aromatase activity, which is followed by downregulation of cancer cell growth^[10,24]. Local estrogen biosynthesis has also been demonstrated in men, where aromatase expression in adipose tissue is greatly increased by this process^[25,26]. However, with respect to esophageal adenocarcinoma, no studies have evaluated whether the amount of estrogen synthesized in abdominal adipose tissue is sufficient to exert any paracrine effect in the esophagus.

ESTROGEN RECEPTORS: STRUCTURE AND FUNCTIONS

The effects of estrogens are mediated by their ligation to ER α and ER β . ER α and ER β both belong to the nuclear steroid/thyroid hormone receptor family and they are encoded by two distinct genes [encoding estrogen receptor 1 (*ESR1*) and *ESR2*] which are located on two different chromosomes 6q25.1 and 14q22-24^[27,28]. ER α and ER β have distinct cellular distributions and regulate separate sets of genes. ER α is predominantly expressed in female sex organs such as the breast, uterus and ovaries especially during the reproductive years. ER β is widely expressed in many other tissues in both genders, but to a lesser degree in males compared to females^[15,29]. Although the role of ERs in male physiology has long been neglected, there is growing evidence for estrogen involvement in multiple areas of male physiology^[15-17].

The mechanism for ER signaling has been widely investigated. ER α and ER β share common functional domains, with a conserved central DNA-binding domain which is often involved in receptor dimerization^[30,31]. ERs possess two activation function domains; activation function-1 and activation function-2, with the former interacting with non-ER transcription factors, and the latter containing the ligand binding domain^[31,32]. One of the most important differences between ER α and ER β is that activation function-1 in ER β lacks functional activity^[30]. Also, it has been suggested that the main function of ER β is to bind ER α and suppress its activation, so that ER α and ER β as a dimer might exert inverse biological effects. Another difference between ER α and ER β signaling is their interaction with the activator protein-1^[32-35]. The activator protein-1 complex of Jun/Fos hetero- or homo-dimers is a key regulator of cell proliferation, with one of its target genes identified as cyclin

D1^[33]. Depending on whether ER α or ER β is activated, the activator protein-1 complex acts in a reciprocal fashion to stimulate or inhibit cell proliferation^[35].

After binding estrogen, the receptor ligand-binding domain undergoes a conformational and surface-charge change that results in receptor dimerization. Ligand-binding is accompanied by the dissociation of intracellular ER from chaperone proteins, subsequently releasing the hormone/ER complex for attachment to estrogen response elements in the promoter region of target genes. The dimer then binds DNA to regulate gene expression at specific regions of the DNA named hormone response elements^[31,35]. As a consequence, transcription of 17 β -estradiol-responsive genes increases, and proliferation or differentiation of steroid-sensitive tissue is augmented. Although most steroid hormone receptors primarily localize to the nuclei, additional ERs have been identified in the cytoplasm and on the plasma membrane. Activation of cytoplasm signaling cascades has been detected after estrogen binding to its plasma membrane receptors^[36].

Several isoforms of ER β able to mediate estrogen signaling have also been found. The isoforms can exert diverse functions, and significantly complicate understanding of cellular responses to estrogens. ER β isoforms can inhibit ER α transcriptional activity at the estrogen response elements and potentially reverse estrogen signaling^[34]. A splice variant of ER β , termed ER β cx, has been characterized^[37]. ER β cx is expressed in the breast^[38], the prostate and testis^[37], the esophagus^[39], and in gastric tissue^[40]. Interestingly, ER β cx does not bind estrogen^[41]. Instead it inhibits ER α from binding DNA, whilst it does not influence ER β . The role and mechanism of ER β cx downstream signaling in esophageal tissue is largely unclear and needs to be further investigated.

ROLE OF ESTROGEN AND ESTROGEN RECEPTORS IN VARIOUS MALIGNANCIES: HARMFUL OR HELPFUL?

The biological significance of ERs in breast tumorigenesis has been studied extensively. In breast tumors, ER signaling promotes malignancy due to oncogenic mutations, sustained exposure of ER α with endogenous or exogenous estrogen, and abnormal coupling of estrogen-activated cytoplasmic machinery to growth and anti-apoptosis, all well established causative triggers of cancer in postmenopausal women^[41]. Several large prospective studies have confirmed the role of estrogen in stimulation of breast tumor growth, and have demonstrated that the risk of breast cancer is increased in women taking estradiol after the menopause^[42-44].

In females with breast cancer, ER α is instrumental in promoting cell proliferation and cancer progression, whereas ER β exerts anti-proliferative effects by induction of cell cycle and growth arrest^[34]. For instance, the

downregulation of the cyclin D1 gene by ER β prevents cellular progression from the G1 to S-phase of the cell cycle^[45]. Loss of ER β expression is considered to be a common feature in estrogen-dependent breast tumor progression^[34,35] supporting the hypothesis that ER β acts as a protector against the mitogenic activity of estrogen in breast pre-malignant tissues.

Estrogen is also critical for the progression of ovarian cancer^[46-48]. A strong association between long-term estrogen replacement therapy and increased risk of ovarian cancer has been detected in several studies^[45-47]. Similar to breast cancer, the imbalance between ER α and ER β , along with decreasing expression of ER β in the ovaries can also lead to uncontrolled cellular proliferation, subsequent malignancy and metastasis^[49,50]. Thus, ER β appears to be pro-apoptotic, facilitating the destruction of malignant cells, whereas ER α has anti-apoptotic activity, indicating its growth stimulatory role^[34,45,49]. Confirming the role of ER β as a tumor-suppressor, deletion of chromosome 14q, where ER β co-localizes with some other tumor suppressors, is often detected in breast, colon, ovarian and prostate malignant tissue^[51-54].

In contrast to the cancer-promoting role of estrogen in breast and ovarian cancers, it has been shown that estrogen works as a cancer suppressor for several gastrointestinal malignancies^[41,42,44-56]. The Women's Health Initiative study, which included a cohort of 16 608 women randomized to hormone replacement therapy (HRT) *vs* no HRT, showed that the risk of colorectal cancer was almost halved in women using HRT^[55]. A similar study in the United Kingdom of patients with esophageal and gastric cancer concluded that HRT was associated with a 50% reduction in the risk of gastric and colon adenocarcinoma, but had no significant benefit for esophageal adenocarcinoma^[56]. However, due to the relatively small number of females with esophageal adenocarcinoma in this study ($n = 299$), the power of the study was limited and the question remains, thus, unresolved^[41,42].

The male predominance of approximately 2:1 in gastric cancer incidence across the world cannot be explained on the basis of gender differences for the prevalence of known risk factors^[57]. It has been hypothesized that estrogens play a protective role against gastric cancer. This statement has gained further support from a clinical study of a male cohort of patients with prostate cancer. In this study the risk of developing gastric cancer was lower amongst those who had been treated with estrogen than in those without such treatment (standardized incidence ratio, 0.87; 95% confidence interval, 0.78-0.98)^[58]. Further supporting this argument are studies which have shown decreased ER β expression in other gastrointestinal cancers, such as colon cancer, compared to benign tumors and normal tissues^[59]. Tamoxifen exposure has also been shown to be a risk factor for gastric cancer^[60,61], adding support to the idea that estrogen signaling has a protective role against gastrointestinal cancer.

FAT DISTRIBUTION, LEPTIN AND ESTROGEN: IS THERE A LINK?

There is a growing appreciation that estrogens are not only directly involved in the reproductive process and in regulation of carcinogenesis, but also have general metabolic roles in both sexes^[15-17]. Estrogen signaling has a complex relationship with obesity that differs for premenopausal and postmenopausal women^[12]. Importantly, obesity is a risk factor for esophageal adenocarcinoma in both women^[62] and men^[63]. In a recent study of 23 women with esophageal adenocarcinoma^[63], 21 (91.3%) were in the top half of the distribution of the studied cohort with regard to waist-to-hip ratio, waist circumference, and body mass index. Multiple studies of male cohorts have demonstrated a strong association between increased abdominal diameter and esophageal adenocarcinoma, after controlling for body mass index and gastroesophageal reflux^[63-68]. It is possible that associations between obesity and esophageal cancer are similar for both sexes, even though the regulation of adiposity in men and women differs significantly. For instance, distribution of body fat in men is characterized by the accumulation of visceral fat, but in women by subcutaneous fat.

Subcutaneous and visceral fat tissues express variable levels of both types of ER^[69-71]. However, only ER α has a significant influence on energy homeostasis. The role of ER α in estradiol regulation of body weight and obesity is supported by the following observations: (1) both male and female mice that have been genetically altered to reduce the ability to produce estrogen by knocking out aromatase (an enzyme that catalyzes the conversion of androgen to estrogen) became obese when fed the same amounts as normal mice^[72]; and (2) increased white adipose tissue and body fat were seen in both sexually mature male and female ER α -knockout mice^[73,74]. Further supporting a role for estrogen signaling through ER α in the regulation of body weight are the findings that abnormal adiposity has been associated with the XbaI polymorphism of the human ER α gene^[75,76].

The role of ER β in estradiol regulation of body weight and obesity is less clear and somewhat controversial suggesting that ER β functions more as a modulator of estrogen actions^[71].

Estrogen has also been shown to contribute to the regulation of body adiposity and fat distribution through ERs in the brain^[77], and by interacting with leptin signaling pathways^[78]. 17 β -estradiol increases leptin mRNA levels in adipose tissue^[79]. Consistently, estrogen deficiency impairs central leptin sensitivity^[77,78]. In women, leptin fluctuations during the menstrual cycle correlate directly with secretion of estrogen^[79,80]. Estrogen has also been found to influence leptin receptor expression and hypothalamic sensitivity to leptin driving subcutaneous body fat accrual over visceral fat during the estrous cycle in rats^[81]. Hence, visceral fat varies inversely with estrogen levels. Visceral fat accumulates in females when circulating estrogen levels become sufficiently low, as

in postmenopausal women^[76,78,82]. The accumulation of visceral fat is associated with an increased risk of various gastrointestinal malignancies, including esophageal adenocarcinoma^[83]. Thus, estrogen regulation of leptin levels in women may play a protective role, directing accumulation of subcutaneous in preference to visceral fat.

The situation for men, however, is less clear, although a high level of leptin is considered to be a risk factor for males to develop esophageal adenocarcinoma^[63,83]. Speculatively, the production of, and sensitivity to, leptin in men may be increased in visceral fat, and locally in tissues located in close proximity to adipose tissue where estrogen synthesis may be increased. However, mechanisms of ER and leptin signaling in males remain obscure, mostly because the majority of laboratory findings and clinical investigations of leptin and estrogen signaling have used tissues from females. To address this issue, studies are needed that specifically address the role of estrogen signaling in male adipose tissue.

IMPLICATIONS OF ESTROGEN RECEPTOR EXPRESSION IN ESOPHAGEAL ADENOCARCINOMA

In 1998, Lagergren *et al*^[83] hypothesized that high estrogen and/or progesterone levels, low testosterone, or a combination of both, might contribute to the lower incidence of esophageal carcinoma in women. Epidemiological data for esophageal adenocarcinoma demonstrates a profound gender difference, with the male:female ratio exceeding 8:1, strongly supporting this hypothesis^[1-4]. There are no detailed studies that compare the expression of ERs in esophageal tissues between males and females, but a limited number of studies have provided some preliminary data comparing ER expression in esophageal adenocarcinoma and its precursor lesion, Barrett's esophagus. These studies are summarized in Table 1.

In contrast to the anti-tumor role of ER β in other cancers, some studies have identified a positive association between ER β expression and esophageal adenocarcinoma development. Akgun *et al*^[84] determined ER β expression in the esophageal mucosa from patients with Barrett's metaplasia negative for dysplasia, Barrett's metaplasia with low grade dysplasia and Barrett's metaplasia with high grade dysplasia. The results of this study showed significant expression of ER β (more than 50% of cells positive) in all patients with esophageal adenocarcinoma, and there was a trend towards increased expression of ER β as the esophageal lesions progressed^[85]. These results raise the possibility of ER β as a target of therapy for esophageal adenocarcinoma. Similarly, another investigation showed a moderate increase in ER expression in tissue samples from men and women with Barrett's esophagus and esophageal adenocarcinoma. However, the subtype of ER was not determined in this study^[59].

As ER β has several isoforms, and these isoforms

Table 1 Estrogen receptor expression in patients with esophageal adenocarcinoma				
	No. of patients with EAC	ER α	ER β	Conclusion
Akgun <i>et al</i> ^[84]	31	Not expressed	Increased expression as esophageal lesions progressed	ER β is suggested as a EAC therapy target
Tiffin <i>et al</i> ^[85]	20 (8)	Type of ER was not specified; ER were detected in EAC patients		ER may be important for further investigation
Liu <i>et al</i> ^[39]	33	Not expressed	Expressed in EAC, but not in Barrett's esophagus	Anti-estrogen treatment could be a promising therapeutic target for EAC
Kalayaransan <i>et al</i> ^[86]	45 (15)	Not expressed	Detected in all 45 patients; Expressed higher in EAC, compared to normal esophageal mucosa	ER β suggested as marker and/or prognostic factor

EAC: Esophageal adenocarcinoma; ER: Estrogen receptor.

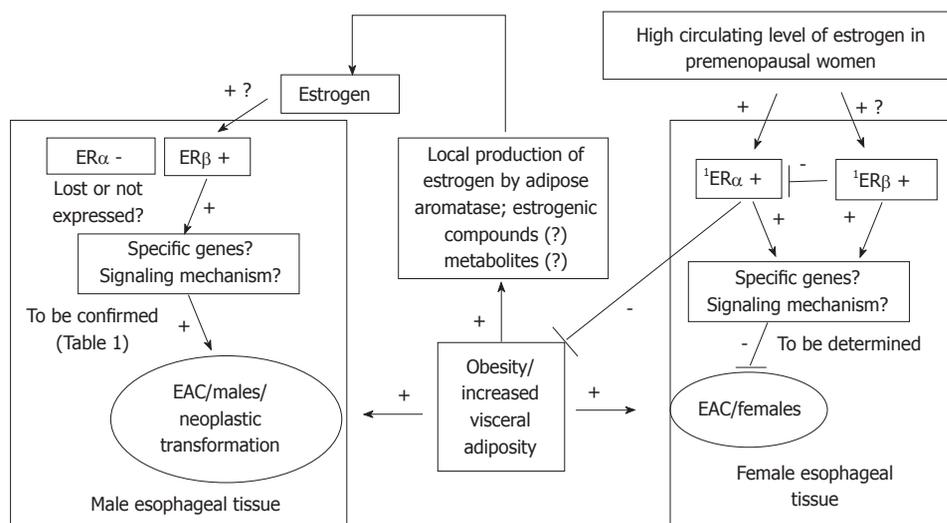


Figure 1 Role of estrogen-activated signaling in the development of esophageal adenocarcinoma: hypothetical pathway. ¹Expression level of estrogen receptor (ER) needs to be determined in esophageal tissue from females and compared to expression in males. EAC: Esophageal adenocarcinoma.

have different functions, Liu *et al*^[39] identified which isoforms of ER β were expressed in esophageal adenocarcinoma but not in Barrett's esophagus. All isoforms of ER β showed much higher expression in esophageal adenocarcinoma, than in its precursor lesion, Barrett's esophagus. Thus, a possible role for ER β isoforms in the maintenance and evolution of esophageal adenocarcinoma was suggested. Although the study did not find a correlation between immunoreactivity and cancer proliferative activity, it showed that ER β 1 tended to have higher expression in invasive tumors which had penetrated the full thickness of the esophageal wall, compared to tumors limited to the esophageal wall ($P = 0.05$), and ER β 1 immunostaining tended to be most prominent in invasive esophageal adenocarcinoma. Conclusively, the study detected the presence of ER β isoforms in esophageal adenocarcinoma and suggested the potential use of anti-estrogen treatment as a therapeutic target for esophageal adenocarcinoma. Manipulation of ER β signaling may be considered as a potential prevention strategy to delay or block progression from dysplasia to esophageal adenocarcinoma. Figure 1 summarizes a potential mechanism for interaction between estrogen, ERs

and esophageal adenocarcinoma.

Another study by Kalayaransan *et al*^[86] determined the expression of ER α and ER β in esophageal adenocarcinoma across various classifications of tumor stage, and compared expression with adjacent normal esophageal mucosa. No significant expression levels of ER α were found in esophageal adenocarcinoma, suggesting ER α is unlikely to mediate the growth of esophageal adenocarcinoma. However, immunostaining with ER β antibodies yielded significantly higher results in esophageal adenocarcinoma, compared to normal esophageal mucosa^[87]. In each group with the same degree of tumor differentiation, tumor samples had significantly higher staining scores compared to normal esophageal mucosa. Tumors with good or moderate differentiation had lower staining scores than those which were poorly differentiated, indicating that the potential effect of estrogen on esophageal adenocarcinoma could be mediated by ER β ^[84,86,87]. Overall, most studies that have evaluated esophageal adenocarcinoma are consistent in suggesting a detrimental effect and prognostic value for ER β .

Unfortunately, these clinical findings have not yet been supported by *in vitro* experiments using esopha-

geal adenocarcinoma cells. The few *in vitro* studies that have addressed the role of estrogen in the regulation of esophageal cell growth were conducted using squamous cancer cells^[87,88]. It has been shown that the growth of an ER-positive esophageal squamous carcinoma cell line (ES-25C) is significantly inhibited by 17 β -estradiol, whereas this effect is not observed in an ER-negative squamous carcinoma cell line (ES-8C)^[87]. A similar finding was seen in another study, in which the proliferation of the ER-positive KSE-1 esophageal squamous carcinoma cell line was inhibited by 17 β -estradiol^[88]. In addition, *in vivo* growth of this cell line in both female and male mice was suppressed by the administration of 17 β -estradiol, raising the possibility of manipulating the growth of esophageal carcinoma by manipulating the estrogen-ER system^[88]. However, esophageal squamous cell carcinoma and esophageal adenocarcinoma are two biologically distinct diseases, so estrogen responsiveness in squamous cell carcinoma lines does not automatically mean that esophageal adenocarcinoma cell lines will also respond. Similar experiments need to be performed on esophageal adenocarcinoma cell lines in order to explore this possibility further.

FUTURE PERSPECTIVES

Current literature provides only limited evidence for a link between estrogen and the development of esophageal adenocarcinoma. Hence, a series of questions can be proposed, and further studies will be needed to determine whether there is any link. It is unclear whether there is a gender difference for the expression of ER β , or correlation between tumor stage and the expression of ER β . Most previous studies have not compared estrogen effects in both genders, and have only addressed men and women separately. Detailed comparisons have not been done for various esophageal pathologies *vs* normal esophageal mucosa within both gender groups. Another limitation of previous studies is the small number of patients studied, and for this reason reported data is yet to be verified. A systematic study which includes a sufficiently large number of men and women is needed to determine whether, within each gender group, ER β expression is associated with the development and progression of esophageal adenocarcinoma. A confirmed link might provide support for ER β to be used as a target for therapy, or as a prognostic marker.

REFERENCES

- 1 **Armstrong RW**, Borman B. Trends in incidence rates of adenocarcinoma of the oesophagus and gastric cardia in New Zealand, 1978-1992. *Int J Epidemiol* 1996; **25**: 941-947
- 2 **Hansson LE**, Sparén P, Nyrén O. Increasing incidence of both major histological types of esophageal carcinomas among men in Sweden. *Int J Cancer* 1993; **54**: 402-407
- 3 **Lepage C**, Rachet B, Jooste V, Faivre J, Coleman MP. Continuing rapid increase in esophageal adenocarcinoma in England and Wales. *Am J Gastroenterol* 2008; **103**: 2694-2699
- 4 **Brown LM**, Devesa SS, Chow WH. Incidence of adenocar-

- cinoma of the esophagus among white Americans by sex, stage, and age. *J Natl Cancer Inst* 2008; **100**: 1184-1187
- 5 **Powell J**, McConkey CC, Gillison EW, Spychal RT. Continuing rising trend in oesophageal adenocarcinoma. *Int J Cancer* 2002; **102**: 422-427
- 6 **Hansen S**, Wiig JN, Giercksky KE, Tretli S. Esophageal and gastric carcinoma in Norway 1958-1992: incidence time trend variability according to morphological subtypes and organ subsites. *Int J Cancer* 1997; **71**: 340-344
- 7 **McKinney A**, Sharp L, Macfarlane GJ, Muir CS. Oesophageal and gastric cancer in Scotland 1960-90. *Br J Cancer* 1995; **71**: 411-415
- 8 **Parkin DM**, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. *CA Cancer J Clin* 2005; **55**: 74-108
- 9 **Nordenstedt H**, El-Serag H. The influence of age, sex, and race on the incidence of esophageal cancer in the United States (1992-2006). *Scand J Gastroenterol* 2011; **46**: 597-602
- 10 **Russo J**, Russo IH. Breast development, hormones and cancer. *Adv Exp Med Biol* 2008; **630**: 52-56
- 11 **Chen JQ**, Brown TR, Yager JD. Mechanisms of hormone carcinogenesis: evolution of views, role of mitochondria. *Adv Exp Med Biol* 2008; **630**: 1-18
- 12 **Rose DP**, Vona-Davis L. Interaction between menopausal status and obesity in affecting breast cancer risk. *Maturitas* 2010; **66**: 33-38
- 13 **Zhao Y**, Nichols JE, Bulun SE, Mendelson CR, Simpson ER. Aromatase P450 gene expression in human adipose tissue. Role of a Jak/STAT pathway in regulation of the adipose-specific promoter. *J Biol Chem* 1995; **270**: 16449-16457
- 14 **Zhou C**, Zhou D, Esteban J, Murai J, Siiteri PK, Wilczynski S, Chen S. Aromatase gene expression and its exon 1 usage in human breast tumors. Detection of aromatase messenger RNA by reverse transcription-polymerase chain reaction. *J Steroid Biochem Mol Biol* 1996; **59**: 163-171
- 15 **Sharpe RM**. The roles of oestrogen in the male. *Trends Endocrinol Metab* 1998; **9**: 371-377
- 16 **Simpson E**, Rubin G, Clyne C, Robertson K, O'Donnell L, Jones M, Davis S. The role of local estrogen biosynthesis in males and females. *Trends Endocrinol Metab* 2000; **11**: 184-188
- 17 **Simpson ER**, Clyne C, Speed C, Rubin G, Bulun S. Tissue-specific estrogen biosynthesis and metabolism. *Ann N Y Acad Sci* 2001; **949**: 58-67
- 18 **Labrie F**, Cusan L, Gomez JL, Martel C, Bérubé R, Bélanger P, Bélanger A, Vandenput L, Mellström D, Ohlsson C. Comparable amounts of sex steroids are made outside the gonads in men and women: strong lesson for hormone therapy of prostate and breast cancer. *J Steroid Biochem Mol Biol* 2009; **113**: 52-56
- 19 **Labrie F**, Bélanger A, Cusan L, Candas B. Physiological changes in dehydroepiandrosterone are not reflected by serum levels of active androgens and estrogens but of their metabolites: intracrinology. *J Clin Endocrinol Metab* 1997; **82**: 2403-2409
- 20 **Labrie F**, Bélanger A, Cusan L, Gomez JL, Candas B. Marked decline in serum concentrations of adrenal C19 sex steroid precursors and conjugated androgen metabolites during aging. *J Clin Endocrinol Metab* 1997; **82**: 2396-2402
- 21 **Vandenput L**, Ohlsson C. Estrogens as regulators of bone health in men. *Nat Rev Endocrinol* 2009; **5**: 437-443
- 22 **Cho JJ**, Cadet P, Salamon E, Mantione K, Stefano GB. The nongenomic protective effects of estrogen on the male cardiovascular system: clinical and therapeutic implications in aging men. *Med Sci Monit* 2003; **9**: RA63-RA68
- 23 **Cornil CA**, Ball GF, Balthazart J. Functional significance of the rapid regulation of brain estrogen action: where do the estrogens come from? *Brain Res* 2006; **1126**: 2-26
- 24 **Huiart L**, Dell'Aniello S, Suissa S. Use of tamoxifen and aromatase inhibitors in a large population-based cohort of women with breast cancer. *Br J Cancer* 2011; **104**: 1558-1563
- 25 **Dieudonné MN**, Sammari A, Dos Santos E, Leneuve MC,

- Giudicelli Y, Pecquery R. Sex steroids and leptin regulate 11beta-hydroxysteroid dehydrogenase I and P450 aromatase expressions in human preadipocytes: Sex specificities. *J Steroid Biochem Mol Biol* 2006; **99**: 189-196
- 26 **Wilson JD**, Aiman J, MacDonald PC. The pathogenesis of gynecomastia. *Adv Intern Med* 1980; **25**: 1-32
- 27 **McInerney EM**, Tsai MJ, O'Malley BW, Katzenellenbogen BS. Analysis of estrogen receptor transcriptional enhancement by a nuclear hormone receptor coactivator. *Proc Natl Acad Sci USA* 1996; **93**: 10069-10073
- 28 **Enmark E**, Pelto-Huikko M, Grandien K, Lagercrantz S, Lagercrantz J, Fried G, Nordenskjöld M, Gustafsson JA. Human estrogen receptor beta-gene structure, chromosomal localization, and expression pattern. *J Clin Endocrinol Metab* 1997; **82**: 4258-4265
- 29 **Rochira V**, Granata AR, Madeo B, Zirilli L, Rossi G, Carani C. Estrogens in males: what have we learned in the last 10 years? *Asian J Androl* 2005; **7**: 3-20
- 30 **DeLaunay F**, Pettersson K, Tujague M, Gustafsson JA. Functional differences between the amino-terminal domains of estrogen receptors alpha and beta. *Mol Pharmacol* 2000; **58**: 584-590
- 31 **Nilsson S**, Mäkelä S, Treuter E, Tujague M, Thomsen J, Andersson G, Enmark E, Pettersson K, Warner M, Gustafsson JA. Mechanisms of estrogen action. *Physiol Rev* 2001; **81**: 1535-1565
- 32 **Hayashi SI**, Eguchi H, Tanimoto K, Yoshida T, Omoto Y, Inoue A, Yoshida N, Yamaguchi Y. The expression and function of estrogen receptor alpha and beta in human breast cancer and its clinical application. *Endocr Relat Cancer* 2003; **10**: 193-202
- 33 **Liu MM**, Albanese C, Anderson CM, Hilty K, Webb P, Uht RM, Price RH, Pestell RG, Kushner PJ. Opposing action of estrogen receptors alpha and beta on cyclin D1 gene expression. *J Biol Chem* 2002; **277**: 24353-24360
- 34 **Bardin A**, Bouille N, Lazennec G, Vignon F, Pujol P. Loss of ERbeta expression as a common step in estrogen-dependent tumor progression. *Endocr Relat Cancer* 2004; **11**: 537-551
- 35 **Chen GG**, Zeng Q, Tse GM. Estrogen and its receptors in cancer. *Med Res Rev* 2008; **28**: 954-974
- 36 **Levin ER**, Pietras RJ. Estrogen receptors outside the nucleus in breast cancer. *Breast Cancer Res Treat* 2008; **108**: 351-361
- 37 **Ogawa S**, Inoue S, Watanabe T, Orimo A, Hosoi T, Ouchi Y, Muramatsu M. Molecular cloning and characterization of human estrogen receptor betacx: a potential inhibitor of estrogen action in human. *Nucleic Acids Res* 1998; **26**: 3505-3512
- 38 **Palmieri C**, Lam EW, Mansi J, MacDonald C, Shousha S, Madden P, Omoto Y, Sunter A, Warner M, Gustafsson JA, Coombes RC. The expression of ER beta cx in human breast cancer and the relationship to endocrine therapy and survival. *Clin Cancer Res* 2004; **10**: 2421-2428
- 39 **Liu L**, Chirala M, Younes M. Expression of estrogen receptor-beta isoforms in Barrett's metaplasia, dysplasia and esophageal adenocarcinoma. *Anticancer Res* 2004; **24**: 2919-2924
- 40 **Hankinson SE**, Colditz GA, Willett WC. Towards an integrated model for breast cancer etiology: the lifelong interplay of genes, lifestyle, and hormones. *Breast Cancer Res* 2004; **6**: 213-218
- 41 **Nelson HD**, Humphrey LL, Nygren P, Teutsch SM, Allan JD. Postmenopausal hormone replacement therapy: scientific review. *JAMA* 2002; **288**: 872-881
- 42 **Rossouw JE**, Anderson GL, Prentice RL, LaCroix AZ, Kooperberg C, Stefanick ML, Jackson RD, Beresford SA, Howard BV, Johnson KC, Kotchen JM, Ockene J. Risks and benefits of estrogen plus progestin in healthy postmenopausal women: principal results From the Women's Health Initiative randomized controlled trial. *JAMA* 2002; **288**: 321-333
- 43 **Beral V**. Breast cancer and hormone-replacement therapy in the Million Women Study. *Lancet* 2003; **362**: 419-427
- 44 **Paruthiyil S**, Parmar H, Kerekatte V, Cunha GR, Firestone GL, Leitman DC. Estrogen receptor beta inhibits human breast cancer cell proliferation and tumor formation by causing a G2 cell cycle arrest. *Cancer Res* 2004; **64**: 423-428
- 45 **Lacey JV**, Mink PJ, Lubin JH, Sherman ME, Troisi R, Hartge P, Schatzkin A, Schairer C. Menopausal hormone replacement therapy and risk of ovarian cancer. *JAMA* 2002; **288**: 334-341
- 46 **Anderson GL**, Judd HL, Kaunitz AM, Barad DH, Beresford SA, Pettinger M, Liu J, McNeeley SG, Lopez AM. Effects of estrogen plus progestin on gynecologic cancers and associated diagnostic procedures: the Women's Health Initiative randomized trial. *JAMA* 2003; **290**: 1739-1748
- 47 **Folsom AR**, Anderson JP, Ross JA. Estrogen replacement therapy and ovarian cancer. *Epidemiology* 2004; **15**: 100-104
- 48 **Pujol P**, Rey JM, Nirde P, Roger P, Gastaldi M, Laffargue F, Rochefort H, Maudelonde T. Differential expression of estrogen receptor-alpha and -beta messenger RNAs as a potential marker of ovarian carcinogenesis. *Cancer Res* 1998; **58**: 5367-5373
- 49 **Rutherford T**, Brown WD, Sapi E, Aschkenazi S, Muñoz A, Mor G. Absence of estrogen receptor-beta expression in metastatic ovarian cancer. *Obstet Gynecol* 2000; **96**: 417-421
- 50 **Bandera CA**, Takahashi H, Behbakht K, Liu PC, LiVolsi VA, Benjamin I, Morgan MA, King SA, Rubin SC, Boyd J. Deletion mapping of two potential chromosome 14 tumor suppressor gene loci in ovarian carcinoma. *Cancer Res* 1997; **57**: 513-515
- 51 **Young J**, Leggett B, Gustafson C, Ward M, Searle J, Thomas L, Buttenshaw R, Chenevix-Trench G. Genomic instability occurs in colorectal carcinomas but not in adenomas. *Hum Mutat* 1993; **2**: 351-354
- 52 **Loveday RL**, Greenman J, Simcox DL, Speirs V, Drew PJ, Monson JR, Kerin MJ. Genetic changes in breast cancer detected by comparative genomic hybridisation. *Int J Cancer* 2000; **86**: 494-500
- 53 **Kasahara K**, Taguchi T, Yamasaki I, Kamada M, Yuri K, Shuin T. Detection of genetic alterations in advanced prostate cancer by comparative genomic hybridization. *Cancer Genet Cytogenet* 2002; **137**: 59-63
- 54 **Chlebowski RT**, Wactawski-Wende J, Ritenbaugh C, Hubbell FA, Ascensao J, Rodabough RJ, Rosenberg CA, Taylor VM, Harris R, Chen C, Adams-Campbell LL, White E. Estrogen plus progestin and colorectal cancer in postmenopausal women. *N Engl J Med* 2004; **350**: 991-1004
- 55 **Lindblad M**, Garcia Rodríguez LA, Chandanos E, Lagergren J. Hormone replacement therapy and risks of oesophageal and gastric adenocarcinomas. *Br J Cancer* 2006; **94**: 136-141
- 56 **Sipponen P**, Correa P. Delayed rise in incidence of gastric cancer in females results in unique sex ratio (M/F) pattern: etiologic hypothesis. *Gastric Cancer* 2002; **5**: 213-219
- 57 **Lindblad M**, Ye W, Rubio C, Lagergren J. Estrogen and risk of gastric cancer: a protective effect in a nationwide cohort study of patients with prostate cancer in Sweden. *Cancer Epidemiol Biomarkers Prev* 2004; **13**: 2203-2207
- 58 **Konstantinopoulos PA**, Kominea A, Vandoros G, Sykiotis GP, Andricopoulos P, Varakis I, Sotiropoulou-Bonikou G, Papavassiliou AG. Oestrogen receptor beta (ERbeta) is abundantly expressed in normal colonic mucosa, but declines in colon adenocarcinoma paralleling the tumour's dedifferentiation. *Eur J Cancer* 2003; **39**: 1251-1258
- 59 **Rutqvist LE**, Johansson H, Signomklao T, Johansson U, Fornander T, Wilking N. Adjuvant tamoxifen therapy for early stage breast cancer and second primary malignancies. Stockholm Breast Cancer Study Group. *J Natl Cancer Inst* 1995; **87**: 645-651
- 60 **Curtis RE**, Boice JD, Shriner DA, Hankey BF, Fraumeni JF. Second cancers after adjuvant tamoxifen therapy for breast

- cancer. *J Natl Cancer Inst* 1996; **88**: 832-834
- 61 **Bodelon C**, Anderson GL, Rossing MA, Chlebowski RT, Ochs-Balcom HM, Vaughan TL. Hormonal factors and risks of esophageal squamous cell carcinoma and adenocarcinoma in postmenopausal women. *Cancer Prev Res (Phila)* 2011; **4**: 840-850
- 62 **Whiteman DC**, Sadeghi S, Pandeya N, Smithers BM, Gotley DC, Bain CJ, Webb PM, Green AC. Combined effects of obesity, acid reflux and smoking on the risk of adenocarcinomas of the oesophagus. *Gut* 2008; **57**: 173-180
- 63 **Edelstein ZR**, Farrow DC, Bronner MP, Rosen SN, Vaughan TL. Central adiposity and risk of Barrett's esophagus. *Gastroenterology* 2007; **133**: 403-411
- 64 **MacInnis RJ**, English DR, Hopper JL, Giles GG. Body size and composition and the risk of gastric and oesophageal adenocarcinoma. *Int J Cancer* 2006; **118**: 2628-2631
- 65 **Corley DA**, Kubo A, Levin TR, Block G, Habel L, Zhao W, Leighton P, Quesenberry C, Rumore GJ, Buffler PA. Abdominal obesity and body mass index as risk factors for Barrett's esophagus. *Gastroenterology* 2007; **133**: 34-41; quiz 311
- 66 **Corley DA**, Kubo A, Zhao W. Abdominal obesity and the risk of esophageal and gastric cardia carcinomas. *Cancer Epidemiol Biomarkers Prev* 2008; **17**: 352-358
- 67 **Renehan AG**, Tyson M, Egger M, Heller RF, Zwahlen M. Body-mass index and incidence of cancer: a systematic review and meta-analysis of prospective observational studies. *Lancet* 2008; **371**: 569-578
- 68 **Mizutani T**, Nishikawa Y, Adachi H, Enomoto T, Ikegami H, Kurachi H, Nomura T, Miyake A. Identification of estrogen receptor in human adipose tissue and adipocytes. *J Clin Endocrinol Metab* 1994; **78**: 950-954
- 69 **Price TM**, O'Brien SN. Determination of estrogen receptor messenger ribonucleic acid (mRNA) and cytochrome P450 aromatase mRNA levels in adipocytes and adipose stromal cells by competitive polymerase chain reaction amplification. *J Clin Endocrinol Metab* 1993; **77**: 1041-1045
- 70 **Schomberg DW**, Couse JF, Mukherjee A, Lubahn DB, Sar M, Mayo KE, Korach KS. Targeted disruption of the estrogen receptor-alpha gene in female mice: characterization of ovarian responses and phenotype in the adult. *Endocrinology* 1999; **140**: 2733-2744
- 71 **Jones ME**, Thorburn AW, Britt KL, Hewitt KN, Wreford NG, Proietto J, Oz OK, Leury BJ, Robertson KM, Yao S, Simpson ER. Aromatase-deficient (ArKO) mice have a phenotype of increased adiposity. *Proc Natl Acad Sci USA* 2000; **97**: 12735-12740
- 72 **Heine PA**, Taylor JA, Iwamoto GA, Lubahn DB, Cooke PS. Increased adipose tissue in male and female estrogen receptor-alpha knockout mice. *Proc Natl Acad Sci USA* 2000; **97**: 12729-12734
- 73 **Cooke PS**, Heine PA, Taylor JA, Lubahn DB. The role of estrogen and estrogen receptor-alpha in male adipose tissue. *Mol Cell Endocrinol* 2001; **178**: 147-154
- 74 **Speer G**, Cseh K, Winkler G, Vargha P, Braun E, Takács I, Lakatos P. Vitamin D and estrogen receptor gene polymorphisms in type 2 diabetes mellitus and in android type obesity. *Eur J Endocrinol* 2001; **144**: 385-389
- 75 **Bjørntorp P**. Hormonal effects on fat distribution and its relationship to health risk factors. *Acta Paediatr Suppl* 1992; **383**: 59-60; discussion 61
- 76 **Ainslie DA**, Morris MJ, Wittert G, Turnbull H, Proietto J, Thorburn AW. Estrogen deficiency causes central leptin insensitivity and increased hypothalamic neuropeptide Y. *Int J Obes Relat Metab Disord* 2001; **25**: 1680-1688
- 77 **Clegg DJ**, Brown LM, Woods SC, Benoit SC. Gonadal hormones determine sensitivity to central leptin and insulin. *Diabetes* 2006; **55**: 978-987
- 78 **Quinton ND**, Laird SM, Okon MA, Li TC, Smith RF, Ross RJ, Blakemore AI. Serum leptin levels during the menstrual cycle of healthy fertile women. *Br J Biomed Sci* 1999; **56**: 16-19
- 79 **Quinton ND**, Smith RF, Clayton PE, Gill MS, Shalet S, Justice SK, Simon SA, Walters S, Postel-Vinay MC, Blakemore AI, Ross RJ. Leptin binding activity changes with age: the link between leptin and puberty. *J Clin Endocrinol Metab* 1999; **84**: 2336-2341
- 80 **Asarian L**, Geary N. Cyclic estradiol treatment normalizes body weight and restores physiological patterns of spontaneous feeding and sexual receptivity in ovariectomized rats. *Horm Behav* 2002; **42**: 461-471
- 81 **Morita Y**, Iwamoto I, Mizuma N, Kuwahata T, Matsuo T, Yoshinaga M, Douchi T. Precedence of the shift of body-fat distribution over the change in body composition after menopause. *J Obstet Gynaecol Res* 2006; **32**: 513-516
- 82 **Howard JM**, Beddy P, Ennis D, Keogan M, Pidgeon GP, Reynolds JV. Associations between leptin and adiponectin receptor upregulation, visceral obesity and tumour stage in oesophageal and junctional adenocarcinoma. *Br J Surg* 2010; **97**: 1020-1027
- 83 **Lagergren J**, Nyrén O. Do sex hormones play a role in the etiology of esophageal adenocarcinoma? A new hypothesis tested in a population-based cohort of prostate cancer patients. *Cancer Epidemiol Biomarkers Prev* 1998; **7**: 913-915
- 84 **Akgun H**, Lechago J, Younes M. Estrogen receptor-beta is expressed in Barrett's metaplasia and associated adenocarcinoma of the esophagus. *Anticancer Res* 2002; **22**: 1459-1461
- 85 **Tiffin N**, Suvarna SK, Trudgill NJ, Riley SA. Sex hormone receptor immunohistochemistry staining in Barrett's oesophagus and adenocarcinoma. *Histopathology* 2003; **42**: 95-96
- 86 **Kalayarasan R**, Ananthkrishnan N, Kate V, Basu D. Estrogen and progesterone receptors in esophageal carcinoma. *Dis Esophagus* 2008; **21**: 298-303
- 87 **Utsumi Y**, Nakamura T, Nagasue N, Kubota H, Harada T, Morikawa S. Effect of 17 beta-estradiol on the growth of an estrogen receptor-positive human esophageal carcinoma cell line. *Cancer* 1991; **67**: 2284-2289
- 88 **Ueo H**, Matsuoka H, Sugimachi K, Kuwano H, Mori M, Akiyoshi T. Inhibitory effects of estrogen on the growth of a human esophageal carcinoma cell line. *Cancer Res* 1990; **50**: 7212-7215

S- Editor Tian L L- Editor Cant MR E- Editor Li JY

Anastasios Koulaouzidis, MD, MRCP, *Series Editor*

Update of endoscopy in liver disease: More than just treating varices

Christoforos Krystallis, Gail S Masterton, Peter C Hayes, John N Plevris

Christoforos Krystallis, Gail S Masterton, Peter C Hayes, John N Plevris, Centre for Liver and Digestive Disorders, The Royal Infirmary, University of Edinburgh, Edinburgh EH16 4SA, United Kingdom

Author contributions: Krystallis C performed the literature review; Krystallis C, Masterton GS, Hayes PC and Plevris JN wrote the paper.

Correspondence to: John N Plevris, MD, PhD, FRCPE, FEBGH, Centre for Liver and Digestive Disorders, The Royal Infirmary, University of Edinburgh, 51, Little France Crescent, Edinburgh EH16 4SA, United Kingdom. j.plevris@ed.ac.uk
 Telephone: +44-131-242163 Fax: +44-131-2421638

Received: April 11, 2011 Revised: June 2, 2011

Accepted: June 9, 2011

Published online: February 7, 2012

Key words: Cirrhosis; Portal hypertension; Varices; Endoscopic ultrasound; Esophageal capsule; Endoscopic retrograde cholangiopancreatography; Endomicroscopy; Spyglass

Peer reviewers: Erwin Biecker, MD, PhD, Department of Gastroenterology and Hepatology, Helios Klinikum Siegburg, Siegburg 53343, Germany; Yu-Yuan Li, Professor, Department of Gastroenterology, First Municipal People's Hospital of Guangzhou, 1 Panfu Road, Guangzhou 510180, Guangdong Province, China

Krystallis C, Masterton GS, Hayes PC, Plevris JN. Update of endoscopy in liver disease: More than just treating varices. *World J Gastroenterol* 2012; 18(5): 401-411 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/401.htm>
 DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.401>

Abstract

The management of complications in liver disease is often complex and challenging. Endoscopy has undergone a period of rapid expansion with numerous novel and specialized endoscopic modalities that are of increasing value in the investigation and management of the patient with liver disease. In this review, relevant literature search and expert opinions have been used to provide a brief overview and update of the current endoscopic management of patients with liver disease and portal hypertension. The main areas covered are safety of endoscopy in patients with liver disease, the use of standard endoscopy for the treatment of varices and the role of new endoscopic modalities such as endoscopic ultrasound, esophageal capsule, argon plasma coagulation, spyglass and endomicroscopy in the investigation and treatment of liver-related gastrointestinal and biliary pathology. It is clear that the role of the endoscopy in liver disease is well beyond that of just treating varices. As the technology in endoscopy expands, so does the role of the endoscopist in liver disease.

© 2012 Baishideng. All rights reserved.

INTRODUCTION

Liver disease and cirrhosis are common causes of mortality worldwide^[1]. The role of endoscopy in liver disease is both diagnostic and interventional: endoscopy should be offered to patients with relevant symptoms (unsuspected liver disease may be diagnosed in this manner) and for variceal screening and treatment. Patients with liver disease can be challenging to sedate, and the complexity of endoscopy in liver disease continues to increase with rising numbers of patients with a liver transplant, and the advent of new endoscopic modalities such as capsule endoscopy and endoscopic ultrasound (EUS).

SEDATION AND ANALGESIA IN LIVER DISEASE

Pharmacodynamics are altered in advanced liver disease as a result of changes in hepatic conjugation and oxidation, shunting, decreased protein binding and an increased vol-

ume of distribution^[2]. The common agents used for sedation in endoscopy are discussed, however, specific doses cannot be recommended because these are dependent on patient factors. We would recommend that an endoscopist or anesthetist who has experience with this patient group undertakes the sedation of liver patients.

Benzodiazepines

Midazolam is the benzodiazepine of choice in most endoscopy units. It is protein bound and metabolized in the liver by cytochrome P3A4. In cirrhosis, clearance of midazolam is impaired and elimination half-life is doubled. As a result, midazolam should be used with caution in patients with cirrhosis^[3].

Opiates

Pethidine and fentanyl are the most commonly used analgesics for endoscopic procedures. The liver is the major site of biotransformation for most opiates. The oxidation of pethidine is reduced in patients with cirrhosis and its clearance is diminished. Therefore, there is increased bioavailability, and pethidine should be avoided in patients with liver disease^[4]. The half-life of fentanyl is shorter and does not seem to be influenced by cirrhosis. Its use is preferred to pethidine^[5].

Anesthetic agents

The pharmacokinetics of propofol, an anesthetic agent that is widely used in endoscopy, appears to be unaffected by cirrhosis; again, perhaps secondary to its short half-life. One study has suggested that the use of propofol rather than midazolam in patients with compensated cirrhosis facilitates a faster recovery time with less exacerbation of subclinical encephalopathy^[6].

Endotracheal intubation

Gastrointestinal (GI) bleeding in patients with liver disease may be life threatening. If bleeding varices are suspected or patients are hemodynamically unstable, there is often a low threshold for endotracheal intubation to protect the airway. There is little literature on prophylactic intubation for airway protection in such patients, and two retrospective studies^[7,8] have concluded that it does not prevent cardiopulmonary complications, or pneumonia. We are of the opinion that airway protection at emergency endoscopy is extremely important in patients with suspected variceal bleeding, who present with hematemesis, and particularly in hemodynamically unstable patients and those with hepatic encephalopathy or alcohol withdrawal symptoms. In such patients, endoscopy is best undertaken in a critical care environment with immediate access to anesthetic support and endotracheal intubation^[9].

ENDOSCOPY IN THE CIRRHOTIC PATIENTS WITH COAGULATION ABNORMALITIES

Coagulopathy and thrombocytopenia are common in pa-

tients with chronic liver disease. The mechanisms behind coagulation abnormalities are often complex, and it is now thought that prolongation of prothrombin time may not directly relate to the risk of bleeding, and rather, it is the balance of pro- and antithrombotic factors that is important. In practical terms, there is currently no reliable way of quantifying this. Routine correction of coagulopathy at endoscopy is not recommended, although patients with chronic liver disease should receive vitamin K to correct any dietary deficiency that may result in coagulopathy.

It is recognized that diagnostic endoscopy is a low-risk procedure and safe in patients with altered coagulation. However, high-risk endoscopic therapeutic procedures have a significantly increased risk of hemorrhage and, as such, coagulopathy should be treated^[10]. It is therefore common practice in cirrhotic patients to correct significant thrombocytopenia ($< 50 \times 10^6/\text{mL}$) with platelet transfusions and to correct coagulopathy with fresh frozen plasma (FFP) if prothrombin time is > 20 s to an international normalized ratio < 1.5 , before high-risk procedures. Platelet and FFP transfusions are particularly helpful during an acute bleeding episode if the prothrombin time is prolonged or platelets are low, similar to the previously mentioned values^[11].

Novel treatments include recombinant factor VIIa. This has been used as a hemostatic agent in acute variceal bleeding, but failed to show efficacy in a large randomized study^[12]. Another trial has demonstrated that addition of desmopressin does not improve and may worsen the efficacy of terlipressin in controlling acute variceal bleeding in cirrhotic patients^[13].

Certain endoscopic investigations have been shown to be safe in coagulopathic patients with cirrhosis, despite being relatively invasive; EUS-fine needle aspiration of the liver has been shown to be a safe alternative to percutaneous liver biopsy, particularly in patients with advanced liver disease, coagulopathy and high risk of bleeding^[14]. At endoscopic retrograde cholangiopancreatography (ERCP), endoscopic papillary balloon dilation is safer than endoscopic biliary sphincterotomy for the treatment of choledocholithiasis in patients with advanced cirrhosis and coagulopathy, because it has a reduced risk of bleeding^[15].

COMMON ENDOSCOPIC DIAGNOSES AND MANAGEMENT IN PATIENTS WITH LIVER DISEASE

Peptic ulcer disease

The correlation between peptic ulcer disease and cirrhosis is well described. Both duodenal and gastric ulcers are more common in cirrhosis: the reported prevalence is 24.1%^[16]. It is recognized that the prevalence of gastric ulceration increases with the severity of liver disease and is related to changes in the hepatic venous pressure gradient^[16,17].

A high prevalence of *Helicobacter pylori* (*H. pylori*), up

Table 1 Summary of primary prophylaxis of esophageal varices

Grade	Appearance	High-risk stigmata	Treatment
Grade 1: Small varices	Barely noticeable varices; disappear easily with insufflation	No red signs	No treatment
Grade 2: Small/medium varices	Small or medium varices; do not easily disappear with insufflation	± Red signs	NSBB or VBL
Grade 3: Medium/large varices	Medium or large varices; do not disappear with insufflation	± Red signs	NSBB or VBL

NSBB: Non-selective β blockers; VBL: Variceal band ligation.

to 89%, in patients with cirrhosis has been reported^[18]. ¹³C urea breath testing and gastric body histology remain highly accurate in detecting *H. pylori* in cirrhosis, whereas rapid urease tests and serology are less reliable than in non-cirrhotic patients^[19]. A meta-analysis of seven studies with almost 1000 patients has strongly suggested that, as with non-cirrhotic patients, *H. pylori* infection increases the risk for peptic ulcer disease in cirrhosis^[20]. *H. pylori* eradication therapy is effective in chronic liver disease^[21]. However, two recent studies have suggested that *H. pylori* eradication in cirrhotic patients with duodenal ulcers is not as effective at reducing ulcer recurrence as it is in the general population. These patients require maintenance acid suppression therapy^[22,23].

Portal hypertension

The development of portal hypertension and formation of portosystemic shunts is a major event in the natural history of liver disease. Measurement of the portal pressure gradient is invasive and not widely available for clinical use; instead the hepatic venous pressure gradient (HVPG) is commonly used in clinical practice and it is of prognostic value: HVPG ≥ 10 mmHg strongly predicts the development of esophageal varices^[24]. Similarly, the most significant risk factor associated with failure to control bleeding or early rebleeding of esophageal varices is HVPG > 20 mmHg. This is also associated with increased mortality^[25].

Gastroesophageal varices are present in $> 50\%$ of patients with portal hypertension and are more likely as liver disease progresses^[26]. Ectopic varices are located in sites other than the gastroesophageal region and are more common than previously thought: duodenal or colonic varices are seen at angiography or colonoscopy in up to 40% of patients with intrahepatic portal hypertension^[27].

Esophageal varices

It is recommended that all patients undergo endoscopy to assess the presence and the size of varices at the time of the diagnosis of cirrhosis. Thereafter, guidelines for the interval of endoscopic screening vary. Currently, the American Association for the Study of the Liver (AASLD) recommends that, if no varices are present at index endoscopy, this should be repeated at 2-3 years in compensated cirrhosis and annually in decompensated cirrhosis^[11]. The British Society of Gastroenterology recommends annual screening if grade 1 varices are present at initial screening (Table 1, grading and treatment of esophageal varices), and an interval of 3 years if there is

no evidence of varices at index endoscopy^[28].

Esophageal variceal bleeding occurs at a rate of 5%-15% per year in untreated patients. The main risk factors for bleeding are variceal size (grade 2 or 3), decompensated cirrhosis, and the presence of high-risk stigmata at endoscopy^[29]. Variceal bleeding is a significant clinical event with a mortality rate of approximately 20% at 6 wk, and a recurrence rate of up to 60% at 2 years if secondary prophylaxis is not commenced^[30].

The management of esophageal varices may be divided into pre-primary, primary and secondary prophylaxis and control of active bleeding. At present, there is no evidence to support treatment to prevent the development of varices in patients with liver disease (pre-primary prophylaxis)^[31,24].

For primary prophylaxis of esophageal varices, there is no evidence that variceal band ligation (VBL) is superior to β -blockade. Due to issues with access to endoscopy and patient preference, non-selective β -blockade, typically with propranolol, is often first line when treatment is indicated^[32]. Carvedilol is a potent non-selective β -blocker, with weak vasodilating properties. A reduction in the HVPG in the range of 10%-43% has been reported with a 12.5 mg/d dose. Carvedilol has therefore been adopted as the β -blocker of choice for primary prophylaxis of variceal bleeding in some centers^[33-35]. Primary prophylaxis with VBL is recommended if there are contraindications to β -blockers, or concerns about patient compliance^[11].

Secondary prophylaxis is indicated for patients who have had an episode of variceal hemorrhage. β -blocker monotherapy is not used as secondary prophylaxis, and patients should either be entered into a variceal banding program or receive a combination of a β -blocker and nitrate^[36]. AASLD recommends a combination of β blockade and VBL^[11]. However, there is no strong evidence to suggest that this strategy is associated with improved mortality^[37] and our local practice is to use VBL alone. VBL should be repeated every 2 wk until obliteration of varices is achieved. Following this, a surveillance endoscopy at 1-3 mo to confirm eradication is required, and this should be repeated every 6-12 mo^[11]. VBL is a safe technique: although asymptomatic banding ulcers are common after VBL, the rate of bleeding from these and requiring hospitalization does not exceed 5%^[38].

The combination of terlipressin and VBL is the preferred treatment for acute variceal bleeding in many centers, and using terlipressin before endoscopy is not unreasonable if there is a delay to the endoscopy. Endo-

scopic hemostasis is usually achieved in the majority of cases^[11]. Transjugular intrahepatic portal systemic shunt (TIPSS) may be considered if VBL has been unsuccessful or there is an early re-bleed (defined at Baveno V as a repeat bleed within 5 d of the index bleed). A reduction in HVPG below 12 mmHg or a 20% reduction from the baseline value, even without reaching < 12 mmHg, protects against rebleeding^[39].

The use of sclerosing agents (variceal sclerotherapy) is no longer recommended as first-line treatment, because of increased mortality rates^[40], nor for secondary prophylaxis because VBL treatment has been shown to be safer and more effective^[41].

Recently, endoscopic placement of a specifically designed self-expanding covered metal stent has proved effective in the treatment of esophageal varices in patients in whom initial endoscopic methods have failed to achieve hemostasis^[42]. This method appears to be a safe and effective means of controlling ongoing bleeding. The stent is usually removed 1 wk after the acute bleed. Currently, this technique is limited by its relative complexity of stent insertion in acute bleeding, but stenting with covered biodegradable stents when available, which do not require removal, may play an important role in the management of acute esophageal variceal bleeding^[43].

Gastric and ectopic varices

Gastric varices are less prevalent than esophageal varices and less prone to bleeding; around 25% over a 2-year period^[44]. There is no evidence to support the primary prophylaxis of gastric varices. The tissue adhesive cyanoacrylate (“glue”) is used widely in the management of acutely bleeding gastric varices. Cyanoacrylate is a liquid with consistency similar to water, which when added to a physiological fluid like blood, polymerizes to form a solid substance^[45]. Two randomized controlled studies have compared cyanoacrylate with VBL for management of bleeding gastric varices^[46,47]. In one study, cyanoacrylate was more effective than VBL in achieving homeostasis, and in the second, no difference was reported, although both reported less recurrence of bleeding. Current evidence suggests that cyanoacrylate achieves control of bleeding in 87%-93% of cases, and that bleeding-related mortality is between 6.5% and 10%^[46,47].

The use of bovine or more recently, human thrombin has been described as an alternative treatment for active gastric variceal bleeding. Thrombin [activated factor II (IIa)] is a serine protease that converts soluble fibrinogen into insoluble strands of fibrin clot. It has additional effects including promotion of platelet aggregation. Initial hemostasis rates have been reported at 94%-100%, and rebleeding rates of between 23% and 25%^[48,49]. A single center experience of 13 patients treated with thrombin for bleeding gastric varices has reported efficient hemostasis and an overall mortality of 38% in a median follow-up of 22 mo^[50].

Technical difficulties that include the risk of equipment damage and reports of severe thromboembolic

complications may limit the use of cyanoacrylate, and thrombin (Figure 1) may become more widespread in the future^[51,52].

The role of TIPSS as primary treatment in actively bleeding gastric varices has also been explored. Although TIPSS has a comparable mortality and rebleeding rate to cyanoacrylate, it is associated with significantly higher morbidity and is not used as first-line treatment^[53,54], but remains an effective treatment when endoscopy fails to control bleeding.

Nonvariceal manifestations of portal hypertension

Portal hypertensive gastropathy (PHG), with its typical “snake skin” appearance, is present in approximately 80% of patients with cirrhosis^[55]. PHG accounts for 8% of nonvariceal bleeds in patients with liver disease, although this condition more commonly presents with anemia^[56]. Patients with cirrhosis and severe PHG-related bleeding may respond to β -blockade. Endoscopic measures such as argon plasma coagulation (APC) therapy can reduce bleeding, thus controlling anemia. TIPSS should be reserved for those patients with pharmacological treatment failure^[57].

The prevalence of portal hypertensive enteropathy (PHE), determined by capsule endoscopy, is as high as 63% in patients with end-stage liver disease who also have esophageal or gastric varices^[58]. Portal hypertensive duodenopathy is present in around half of patients with cirrhosis, and it is more common in patients with severe PHG^[59].

Gastric antral vascular ectasia

Gastric antral vascular ectasia (GAVE) is related to portal hypertension in about 30% of patients, and accounts for 4% of nonvariceal upper GI bleeds^[60]. Unlike PHG, GAVE does not respond well to reduction in portal pressure^[61]. The Nd:YAG laser has been widely used in the treatment of GAVE and is the most commonly reported endoscopic modality in cirrhotic and non-cirrhotic patients, with a reported overall success rate of almost 90%, although the authors did not distinguish the etiology of GAVE when reporting the outcomes^[62].

APC has also proved to be effective for the treatment of GAVE-related bleeding, and it is extensively used in our unit, with a success rate of > 85%; success is defined as control of bleeding, stabilization of hemoglobin at > 100 g/dL, or hemoglobin increase > 10% from pretreatment level, and reduction of transfusion requirements by > 50% in transfusion-dependent patients. An average of four sessions of APC is usually required (Figures 2 and 3). In two published studies, with a total of 37 patients with cirrhosis, success rates in controlling bleeding were very high, and the reported rebleeding rates were between 12% and 20% after 2 years follow-up^[62,63].

Although the success rates of the two aforementioned modalities are comparable, APC treatment is probably the therapy of choice due to the technical ease, safety and low cost^[61]. Other endoscopic techniques have been positively reported in small series, such as VBL^[64],

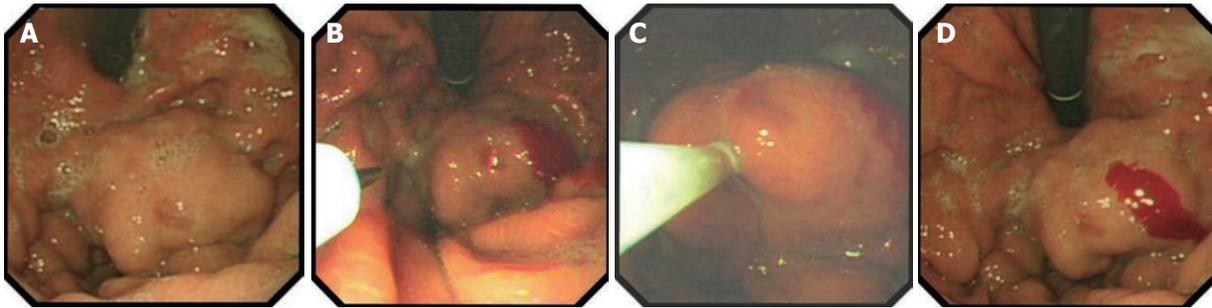


Figure 1 Endoscopic images of fundic gastric varices before (A), during (B, C) and after (D) thrombin injection.

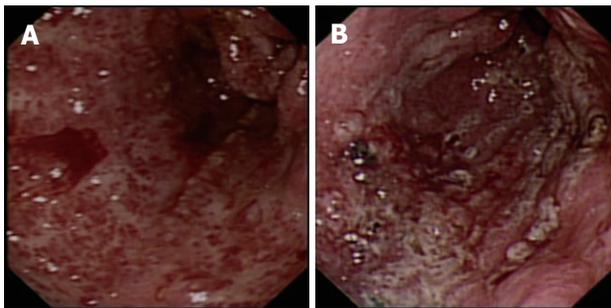


Figure 2 Endoscopic image. A: Gastric antral vascular ectasia (GAVE) (diffuse type) with active bleeding prior to argon plasma coagulation (APC) treatment; B: GAVE (diffuse type) immediately after APC treatment.

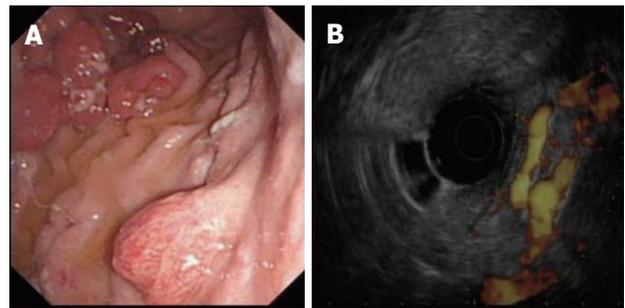


Figure 4 Endoscopic image of gastric irregular submucosal lesion. A: Gastric irregular submucosal lesion in a patient with portal hypertension; B: The same lesion examined under color Doppler endoscopic ultrasound. The submucosal lesion was hypervascular and represented a gastric varix.

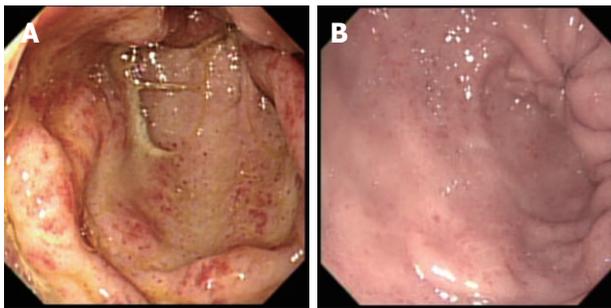


Figure 3 Endoscopic image. A: Gastric antral vascular ectasia-induced symptomatic anemia; B: Endoscopic image of the same patient 2 years later, after several argon plasma coagulation sessions. The number of angioectatic lesions in the gastric outlet had dramatically decreased.

endoscopic mucosal ablation^[65] and cryotherapy^[66].

ADVANCED ENDOSCOPIC PROCEDURES AND THEIR VALUE IN LIVER DISEASE

EUS

Recently, EUS has been used to assist in the management of portal hypertension. Doppler EUS is of significant value in differentiating ectopic varices from other submucosal lesions^[67] (Figure 4), and several EUS-assisted techniques have been used to identify the precise site for the intravariceal injection of sclerosant agents. Linear color EUS-guided sclerotherapy has proved to be effective in the eradication of esophageal varices, in two small studies with recurrence reported at 0% and 8.3%, respective-

ly^[68,69]. EUS catheter probes and high-frequency (20 MHz) miniprobes have both been used successfully before and after esophageal variceal sclerosant injection in two different studies to assess eradication and variceal recurrence. After a mean follow-up of 24 mo, variceal recurrence was reported at 16.6% and 26.3%, respectively^[70,71].

Although the overall patient numbers are small, linear EUS seems to be the superior modality in assisting treatment of esophageal varices, because it permits the targeting of the feeding vessels.

EUS-assisted injection of cyanoacrylate for the treatment of gastric varices has been described in 54 patients with a mean follow-up of 24 mo. Varices recurred in 35% of patients^[72]. Furthermore, a series of 15 patients with gastric or ectopic varices treated with thrombin injection in conjunction with a variety of EUS techniques (Figure 5) has recently been reported in our unit, and this proved to be effective in controlling active bleeding and achieving variceal eradication^[73].

Capsule endoscopy

Esophageal capsule endoscopy (OCE) is an alternative to conventional upper GI endoscopy for the diagnosis of varices in complex patients with portal hypertension. In a recent meta-analysis of seven studies involving 446 patients, OCE had a sensitivity of 85.8% and specificity 80.5% in detecting esophageal varices^[74]. However, a multicenter trial evaluating the efficacy of OCE in esophageal varices screening was less encouraging, because the standard of < 10% difference between capsule



Figure 5 Color Doppler endoscopic ultrasound image of duodenal varices after thrombin injection. The absence of blood flow and the speckled appearances were suggestive of thrombus formation.

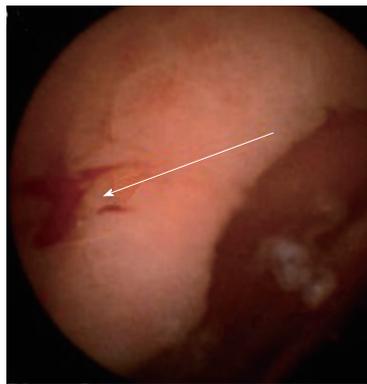


Figure 7 Small bowel capsule image of portal hypertensive enteropathy with snake-skin-like appearance of the mucosa and red spots as stigmata of recent bleeding.



Figure 6 Small bowel capsule image of portal hypertensive enteropathy and stigmata of recent bleeding. Engorged small bowel villi and micro-hemorrhagic spots were visible.

and conventional endoscopy was not met^[75]. This is not surprising, because the two techniques differ in that during conventional endoscopy, the esophagus is inevitably insufflated with air and varices can appear more flattened than during OCE examination. Further studies to take this into account are necessary. Nevertheless, OCE remains a useful tool for screening of varices in certain patient groups; patients who poorly tolerate endoscopy or who have significant comorbidity, thus increasing the risks of repeated endoscopy, and patients with high risk of variant Creutzfeldt-Jakob disease. Although this technique is limited by availability and high costs, OCE can be cost-effective for variceal screening of patients with coagulation abnormalities (e.g., hemophilia) with coexisting liver disease, because it does not require prophylactic clotting factor administration, unlike conventional endoscopy. Serial capsule examinations in the same patient may provide significant diagnostic information regarding progression of varices.

Small bowel capsule endoscopy (SBCE) has been used to characterize PHE (Figure 6), and is of value in the diagnosis of this condition in patients with advanced liver disease who continue to bleed despite treatment of esophageal/gastric varices or portal gastropathy (Figure 7).

The role of SBCE in portal hypertension has yet to be defined, but it is likely that it will remain a valuable tool in certain groups of patients with liver disease.

Endoscopic retrograde cholangiography

Primary sclerosing cholangitis (PSC) is characterized by fibrosis of both intrahepatic and/or extrahepatic biliary ducts. These patients are at risk of developing infectious cholangitis and up to 20% develop cholangiocarcinoma. A strategy of initial magnetic resonance cholangiopancreatography (MRCP) followed, if necessary, by ERCP is currently the most cost-effective approach to the work-up of patients with suspected sclerosing cholangitis^[76]. ERCP plus diagnostic brushing have a sensitivity of 60%-100%, and specificity of 85%-89% in differentiating between a benign dominant stricture and cholangiocarcinoma^[77,78]. Recently, two advanced cytological techniques (digital image analysis and fluorescence *in situ* hybridization) have been used for the detection of malignancy in PSC-related strictures and have proved to be more sensitive and equally specific to conventional cytology^[79].

In addition, ERCP permits therapeutic interventions with balloon dilation or stent placement as appropriate.

Novel techniques and the biliary tree

Novel endoscopic modalities have been compared with conventional ERCP and brush cytology. Transpapillary cholangioscopy with tissue sampling has proved to be more sensitive (92% *vs* 66%) and specific (93% *vs* 51%) than ERCP to detect cholangiocarcinoma in PSC^[80]. In a small study, narrow band imaging has demonstrated superior visualization of biliary lesions compared with conventional white light imaging^[81]. In another study, transpapillary intraductal ultrasound was superior to ERCP for the detection of cholangiocarcinoma in PSC in terms of sensitivity (87.5% *vs* 62.5%) and specificity (90.6% *vs* 53.1%)^[82].

SpyglassTM is a new single-operator system used for the diagnosis of a variety of pancreatobiliary disorders, such as the definition of indeterminate strictures and filling defects prior to stone extraction^[83]. Although the

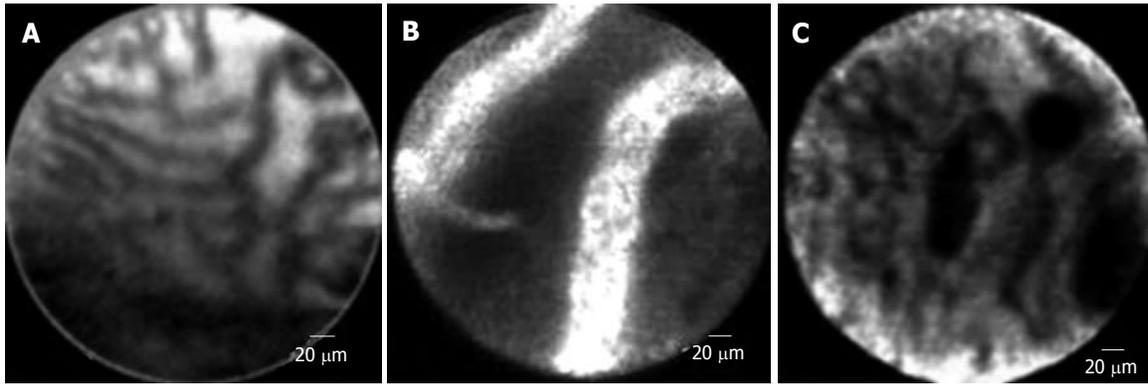


Figure 8 Endomicroscopy image. A: Image from Cellvizio® bile duct endomicroscopy. The regular reticular pattern of thin dark structures with low signal (dark) characterized the normal bile duct (Image courtesy of www.cellvizio.net); B: Abnormal bile duct appearances in Cellvizio® endomicroscopy; isolated blood vessels with very strong signal (with strands) suggestive of tumor neovascularization of cholangiocarcinoma (Image courtesy of www.cellvizio.net); C: Reticular pattern of dark bands and dark clumps or glands suggestive of cholangiocarcinoma (Image courtesy of www.cellvizio.net).

initial experience is promising, the modality has been only been tested in a limited number of PSC-related strictures, and unlike ERCP, this is a purely diagnostic technique.

Confocal laser endomicroscopy (“miniprobe”) is a new field of endoluminal imaging that offers extremely high magnification and resolution. This technique allows visualization of pancreatic and biliary ducts. In a pilot study of 14 patients with biliary strictures, miniprobe-based microscopy after fluorescein administration proved to be more accurate than brushings and biopsy in distinguishing benign from malignant strictures (Figure 8)^[84].

A novel alternative is direct cholangioscopy using ultra-slim endoscopes (4.9-5.9 mm). These endoscopes, initially developed for transnasal endoscopy, can be safely inserted into the bile duct following sphincterotomy, and not only permit high-resolution images, but also biopsy and other interventional procedures in the bile ducts, such as hydraulic lithotripsy and division of strictures in benign biliary disease. This technique is currently under development and after a full range of endoscopic accessories are available for endobiliary interventions, it could be an effective and safe approach for patients with difficult to manage biliary disease^[85].

Endoscopic modalities in the lower GI tract

In comparison to the upper GI tract, colonic manifestations of portal hypertension much less often present with acute bleeding, and are more often found incidentally or during investigation of anemia. As such, data are sparse and less consistent. The reported prevalence of portal hypertensive colonopathy is 24%^[86].

The most significant feature of portal hypertension in the colon is arguably the presence of rectal varices. These can be present in up to 44% of patients with cirrhosis at colonoscopy, although the reported prevalence varies widely. They are more frequent in patients with advanced liver disease^[87].

Although bleeding from rectal varices is uncommon, it can be life threatening. Due to their rarity, no firm

guidelines have been established for the management of bleeding colonic varices, and there is a limited evidence base. The most commonly used treatment modalities are sclerotherapy and band ligation. In a small retrospective comparative study of 15 patients, endoscopic injection sclerotherapy proved to be superior to endoscopic band ligation and achieved lower recurrence rates^[88]. In our unit, thrombin has been successfully used to manage rectal variceal bleeding. Patients may require TIPSS if bleeding cannot be controlled endoscopically.

ENDOSCOPY AND THE LIVER TRANSPLANT PATIENT

Luminal diseases in liver transplant patients

Peptic ulcer disease is the most common cause of GI bleeding in post-orthotopic liver transplant (OLT) recipients, accounting for 27% of all bleeding^[89]. Varices rarely recur post-transplant, and if present, require investigation to exclude portal vein thrombosis or disease recurrence^[90].

Liver transplant patients are at increased risk of opportunistic infections, particularly candidiasis and cytomegalovirus^[91]. These often present with GI symptoms and require endoscopic evaluation with biopsies or brushings to confirm the diagnosis.

The association of PSC with ulcerative colitis is well recognized and is an additional risk factor for the development of colorectal cancer in immunosuppressed transplant patients. It is currently recommended that these patients have an annual surveillance colonoscopy commencing 10 years after the onset of bowel symptoms^[92]. Colectomy is safe in patients who have undergone OLT, and in some high-risk cases, such as when high-grade dysplasia has already been identified, a prophylactic colectomy may be performed at the time of transplantation^[93].

Transplant-related biliary disease

Biliary complications (biliary strictures and leaks) following liver transplantation are a challenging and common

issue that affects 10%-30% of OLT patients. Biliary strictures are classified as anastomotic or non-anastomotic^[94]. The initial approach in suspected post-transplant biliary strictures is usually MRCP, restricting the use of ERCP to patients who require intervention, or where MRCP results are equivocal^[95]. Further imaging techniques include SpyGlassTM, which has been successfully used for the investigation of post-transplant biliary strictures^[96], and contrast-enhanced ultrasound. This is a non-invasive technique for the detection of strictures relating to hepatic artery stenosis in liver transplant patients. It provides details on the presence, location, degree, and type of stricture^[97]. Treatment comprises a combination of balloon dilation and stent placement, repeated if necessary until stricture resolution.

Biliary leaks can occur in up to 22% of patients. Evidence suggests that sphincterotomy with stent placement is the best treatment option for biliary leaks following OLT^[94,98]. Surgical revision and biliary reconstruction with the formation of hepaticojejunostomy is indicated when endoscopic or percutaneous treatment fails^[94,98].

In patients who have a roux-en-Y anastomosis, the technique of double balloon ERCP has been devised with promising results. The technique uses a double balloon colonoscope to approach the ampulla, although there are some limitations regarding endoscopic accessories. With further development of this technique, the endoscopic treatment of biliary complications may become easier and will play a larger role in the management of such patients^[99].

CONCLUSION

Endoscopy has undergone rapid expansion with numerous novel endoscopic modalities and techniques directly applicable to the diagnosis and management of complications of liver disease. Although conventional upper GI endoscopy is still the modality of choice for esophageal variceal surveillance and treatment, further options are now available with the use of capsule endoscopy and EUS. Therapeutic options for the management of upper GI bleeding in portal hypertension have also been developed. Band ligation remains the treatment of choice for esophageal variceal bleeding, whereas for gastric and ectopic varices, the use of sclerosants, particularly "glue" and thrombin are increasingly being used. APC is the preferred modality for GAVE and PHG. Biliary strictures and the risk of cholangiocarcinoma are major issues in patients with PSC. ERCP is both diagnostic and therapeutic in this setting and can differentiate benign from malignant lesions in the majority of cases. Novel endoscopic techniques such as transpapillary cholangioscopy, Spyglass Direct Visualization System, confocal laser endomicroscopy ("miniprobe") and ultra-thin cholangioscopy are increasingly being used to assist diagnosis in selected patients. Finally, in the post-liver transplant patient, upper and lower endoscopies are used to detect gastrointestinal opportunistic infections, as well as to screen for colorec-

tal cancer in high-risk patients. Biliary complications are common after transplantation and ERCP is the modality of choice for treating such patients.

ACKNOWLEDGMENTS

We would like to thank the Hellenic Foundation of Gastroenterology and Nutrition for providing a research scholarship to Dr. C Krystallis.

REFERENCES

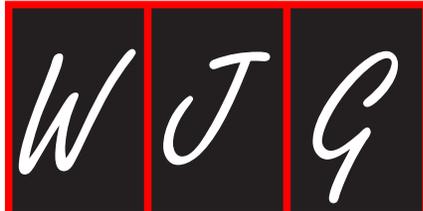
- 1 **World Health Organization.** WHO European Health for All Database. Geneva: Author, 2009
- 2 **Verbeeck RK.** Pharmacokinetics and dosage adjustment in patients with hepatic dysfunction. *Eur J Clin Pharmacol* 2008; **64**: 1147-1161
- 3 **MacGilchrist AJ,** Birnie GG, Cook A, Scobie G, Murray T, Watkinson G, Brodie MJ. Pharmacokinetics and pharmacodynamics of intravenous midazolam in patients with severe alcoholic cirrhosis. *Gut* 1986; **27**: 190-195
- 4 **Neal EA,** Meffin PJ, Gregory PB, Blaschke TF. Enhanced bioavailability and decreased clearance of analgesics in patients with cirrhosis. *Gastroenterology* 1979; **77**: 96-102
- 5 **Tegeder I,** Lötsch J, Geisslinger G. Pharmacokinetics of opioids in liver disease. *Clin Pharmacokinet* 1999; **37**: 17-40
- 6 **Khamaysi I,** William N, Olga A, Alex I, Vladimir M, Kamal D, Nimer A. Sub-clinical hepatic encephalopathy in cirrhotic patients is not aggravated by sedation with propofol compared to midazolam: a randomized controlled study. *J Hepatol* 2011; **54**: 72-77
- 7 **Rudolph SJ,** Landsverk BK, Freeman ML. Endotracheal intubation for airway protection during endoscopy for severe upper GI hemorrhage. *Gastrointest Endosc* 2003; **57**: 58-61
- 8 **Rehman A,** Iscimen R, Yilmaz M, Khan H, Belsher J, Gomez JF, Hanson AC, Afessa B, Baron TH, Gajic O. Prophylactic endotracheal intubation in critically ill patients undergoing endoscopy for upper GI hemorrhage. *Gastrointest Endosc* 2009; **69**: e55-e59
- 9 **Liebler JM,** Benner K, Putnam T, Vollmer WM. Respiratory complications in critically ill medical patients with acute upper gastrointestinal bleeding. *Crit Care Med* 1991; **19**: 1152-1157
- 10 **Veitch AM,** Baglin TP, Gershlick AH, Harnden SM, Tighe R, Cairns S. Guidelines for the management of anticoagulant and antiplatelet therapy in patients undergoing endoscopic procedures. *Gut* 2008; **57**: 1322-1329
- 11 **Garcia-Tsao G,** Sanyal AJ, Grace ND, Carey W. Prevention and management of gastroesophageal varices and variceal hemorrhage in cirrhosis. *Hepatology* 2007; **46**: 922-938
- 12 **Bosch J,** Thabut D, Albillos A, Carbonell N, Spicak J, Massard J, D'Amico G, Lebrech D, de Franchis R, Fabricius S, Cai Y, Bendtsen F. Recombinant factor VIIa for variceal bleeding in patients with advanced cirrhosis: A randomized, controlled trial. *Hepatology* 2008; **47**: 1604-1614
- 13 **de Franchis R,** Arcidiacono PG, Carpinelli L, Andreoni B, Cestari L, Brunati S, Zambelli A, Battaglia G, Mannucci PM. Randomized controlled trial of desmopressin plus terlipressin vs. terlipressin alone for the treatment of acute variceal hemorrhage in cirrhotic patients: a multicenter, double-blind study. *New Italian Endoscopic Club. Hepatology* 1993; **18**: 1102-1107
- 14 **Hollerbach S,** Willert J, Topalidis T, Reiser M, Schmiegel W. Endoscopic ultrasound-guided fine-needle aspiration biopsy of liver lesions: histological and cytological assessment. *Endoscopy* 2003; **35**: 743-749
- 15 **Park DH,** Kim MH, Lee SK, Lee SS, Choi JS, Song MH, Seo DW, Min YI. Endoscopic sphincterotomy vs. endoscopic

- papillary balloon dilation for choledocholithiasis in patients with liver cirrhosis and coagulopathy. *Gastrointest Endosc* 2004; **60**: 180-185
- 16 **Wu CS**, Lin CY, Liaw YF. Helicobacter pylori in cirrhotic patients with peptic ulcer disease: a prospective, case controlled study. *Gastrointest Endosc* 1995; **42**: 424-427
 - 17 **Chen LS**, Lin HC, Hwang SJ, Lee FY, Hou MC, Lee SD. Prevalence of gastric ulcer in cirrhotic patients and its relation to portal hypertension. *J Gastroenterol Hepatol* 1996; **11**: 59-64
 - 18 **Pellicano R**, Leone N, Berrutti M, Cutufia MA, Fiorentino M, Rizzetto M, Ponzetto A. Helicobacter pylori seroprevalence in hepatitis C virus positive patients with cirrhosis. *J Hepatol* 2000; **33**: 648-650
 - 19 **Calvet X**, Sanfeliu I, Musulen E, Mas P, Dalmau B, Gil M, Bella MR, Campo R, Brullet E, Valero C, Puig J. Evaluation of Helicobacter pylori diagnostic methods in patients with liver cirrhosis. *Aliment Pharmacol Ther* 2002; **16**: 1283-1289
 - 20 **Vergara M**, Calvet X, Roqué M. Helicobacter pylori is a risk factor for peptic ulcer disease in cirrhotic patients. A meta-analysis. *Eur J Gastroenterol Hepatol* 2002; **14**: 717-722
 - 21 **Jung SW**, Lee SW, Hyun JJ, Kim DI, Koo JS, Yim HJ, Park JJ, Lee HS, Chun HJ, Um SH, Choi JH, Kim CD, Ryu HS. Efficacy of Helicobacter pylori eradication therapy in chronic liver disease. *Dig Liver Dis* 2009; **41**: 134-140
 - 22 **Lo GH**, Yu HC, Chan YC, Chen WC, Hsu PI, Lin CK, Lai KH. The effects of eradication of Helicobacter pylori on the recurrence of duodenal ulcers in patients with cirrhosis. *Gastrointest Endosc* 2005; **62**: 350-356
 - 23 **Tzathas C**, Triantafyllou K, Mallas E, Triantafyllou G, Ladas SD. Effect of Helicobacter pylori eradication and antisecretory maintenance therapy on peptic ulcer recurrence in cirrhotic patients: a prospective, cohort 2-year follow-up study. *J Clin Gastroenterol* 2008; **42**: 744-749
 - 24 **Groszmann RJ**, Garcia-Tsao G, Bosch J, Grace ND, Burroughs AK, Planas R, Escorsell A, Garcia-Pagan JC, Patch D, Matloff DS, Gao H, Makuch R. Beta-blockers to prevent gastroesophageal varices in patients with cirrhosis. *N Engl J Med* 2005; **353**: 2254-2261
 - 25 **Moitinho E**, Escorsell A, Bandi JC, Salmerón JM, García-Pagán JC, Rodés J, Bosch J. Prognostic value of early measurements of portal pressure in acute variceal bleeding. *Gastroenterology* 1999; **117**: 626-631
 - 26 **Kovalak M**, Lake J, Mattek N, Eisen G, Lieberman D, Zaman A. Endoscopic screening for varices in cirrhotic patients: data from a national endoscopic database. *Gastrointest Endosc* 2007; **65**: 82-88
 - 27 **Norton ID**, Andrews JC, Kamath PS. Management of ectopic varices. *Hepatology* 1998; **28**: 1154-1158
 - 28 **Jalan R**, Hayes PC. UK guidelines on the management of variceal haemorrhage in cirrhotic patients. British Society of Gastroenterology. *Gut* 2000; **46** Suppl 3-4: III1-III15
 - 29 Prediction of the first variceal hemorrhage in patients with cirrhosis of the liver and esophageal varices. A prospective multicenter study. *N Engl J Med* 1988; **319**: 983-989
 - 30 **Garcia-Tsao G**, Bosch J. Management of varices and variceal hemorrhage in cirrhosis. *N Engl J Med* 2010; **362**: 823-832
 - 31 **Calés P**, Oberti F, Payen JL, Naveau S, Guyader D, Blanc P, Abergel A, Bichard P, Raymond JM, Canva-Delcambre V, Vetter D, Valla D, Beauchant M, Hadengue A, Champigneulle B, Pascal JP, Poynard T, Lebrech D. Lack of effect of propranolol in the prevention of large oesophageal varices in patients with cirrhosis: a randomized trial. French-Speaking Club for the Study of Portal Hypertension. *Eur J Gastroenterol Hepatol* 1999; **11**: 741-745
 - 32 **Schepke M**, Kleber G, Nürnberg D, Willert J, Koch L, Veltzke-Schlieker W, Hellerbrand C, Kuth J, Schanz S, Kahl S, Fleig WE, Sauerbruch T. Ligation versus propranolol for the primary prophylaxis of variceal bleeding in cirrhosis. *Hepatology* 2004; **40**: 65-72
 - 33 **de Franchis R**. Revising consensus in portal hypertension: report of the Baveno V consensus workshop on methodology of diagnosis and therapy in portal hypertension. *J Hepatol* 2010; **53**: 762-768
 - 34 **Tripathi D**, Hayes PC. The role of carvedilol in the management of portal hypertension. *Eur J Gastroenterol Hepatol* 2010; **22**: 905-911
 - 35 **Tripathi D**, Ferguson JW, Kochar N, Leithead JA, Therapondos G, McAvoy NC, Stanley AJ, Forrest EH, Hislop WS, Mills PR, Hayes PC. Randomized controlled trial of carvedilol versus variceal band ligation for the prevention of the first variceal bleed. *Hepatology* 2009; **50**: 825-833
 - 36 **de Franchis R**. Evolving consensus in portal hypertension. Report of the Baveno IV consensus workshop on methodology of diagnosis and therapy in portal hypertension. *J Hepatol* 2005; **43**: 167-176
 - 37 **Kumar A**, Jha SK, Sharma P, Dubey S, Tyagi P, Sharma BC, Sarin SK. Addition of propranolol and isosorbide mononitrate to endoscopic variceal ligation does not reduce variceal rebleeding incidence. *Gastroenterology* 2009; **137**: 892-901, 901.e1
 - 38 **García-Pagán JC**, Villanueva C, Albillos A, Bañares R, Morillas R, Abraldes JG, Bosch J. Nadolol plus isosorbide mononitrate alone or associated with band ligation in the prevention of recurrent bleeding: a multicentre randomised controlled trial. *Gut* 2009; **58**: 1144-1150
 - 39 **Bosch J**, García-Pagán JC. Prevention of variceal rebleeding. *Lancet* 2003; **361**: 952-954
 - 40 Prophylactic sclerotherapy for esophageal varices in men with alcoholic liver disease. A randomized, single-blind, multicenter clinical trial. The Veterans Affairs Cooperative Variceal Sclerotherapy Group. *N Engl J Med* 1991; **324**: 1779-1784
 - 41 **de Franchis R**, Primignani M. Endoscopic treatments for portal hypertension. *Semin Liver Dis* 1999; **19**: 439-455
 - 42 **Wright G**, Lewis H, Hogan B, Burroughs A, Patch D, O'Beirne J. A self-expanding metal stent for complicated variceal hemorrhage: experience at a single center. *Gastrointest Endosc* 2010; **71**: 71-78
 - 43 **Zehetner J**, Shamiyeh A, Wayand W, Hubmann R. Results of a new method to stop acute bleeding from esophageal varices: implantation of a self-expanding stent. *Surg Endosc* 2008; **22**: 2149-2152
 - 44 **Sarin SK**, Lahoti D, Saxena SP, Murthy NS, Makwana UK. Prevalence, classification and natural history of gastric varices: a long-term follow-up study in 568 portal hypertension patients. *Hepatology* 1992; **16**: 1343-1349
 - 45 **Binmoeller KF**. Glue for gastric varices: some sticky issues. *Gastrointest Endosc* 2000; **52**: 298-301
 - 46 **Tan PC**, Hou MC, Lin HC, Liu TT, Lee FY, Chang FY, Lee SD. A randomized trial of endoscopic treatment of acute gastric variceal hemorrhage: N-butyl-2-cyanoacrylate injection versus band ligation. *Hepatology* 2006; **43**: 690-697
 - 47 **Lo GH**, Lai KH, Cheng JS, Chen MH, Chiang HT. A prospective, randomized trial of butyl cyanoacrylate injection versus band ligation in the management of bleeding gastric varices. *Hepatology* 2001; **33**: 1060-1064
 - 48 **Przemioslo RT**, McNair A, Williams R. Thrombin is effective in arresting bleeding from gastric variceal hemorrhage. *Dig Dis Sci* 1999; **44**: 778-781
 - 49 **Yang WL**, Tripathi D, Therapondos G, Todd A, Hayes PC. Endoscopic use of human thrombin in bleeding gastric varices. *Am J Gastroenterol* 2002; **97**: 1381-1385
 - 50 **Ramesh J**, Limdi JK, Sharma V, Makin AJ. The use of thrombin injections in the management of bleeding gastric varices: a single-center experience. *Gastrointest Endosc* 2008; **68**: 877-882
 - 51 **Roesch W**, Rexroth G. Pulmonary, cerebral and coronary emboli during bucrylate injection of bleeding fundic varices. *Endoscopy* 1998; **30**: S89-S90

- 52 **Palejwala AA**, Smart HL, Hughes M. Multiple pulmonary glue emboli following gastric variceal obliteration. *Endoscopy* 2000; **32**: S1-S2
- 53 **Procaccini NJ**, Al-Osaimi AM, Northup P, Argo C, Caldwell SH. Endoscopic cyanoacrylate versus transjugular intrahepatic portosystemic shunt for gastric variceal bleeding: a single-center U.S. analysis. *Gastrointest Endosc* 2009; **70**: 881-887
- 54 **Lo GH**, Liang HL, Chen WC, Chen MH, Lai KH, Hsu PI, Lin CK, Chan HH, Pan HB. A prospective, randomized controlled trial of transjugular intrahepatic portosystemic shunt versus cyanoacrylate injection in the prevention of gastric variceal rebleeding. *Endoscopy* 2007; **39**: 679-685
- 55 **Primignani M**, Carpinelli L, Preatoni P, Battaglia G, Carta A, Prada A, Cestari R, Angeli P, Gatta A, Rossi A, Spinzi G, De Franchis R. Natural history of portal hypertensive gastropathy in patients with liver cirrhosis. The New Italian Endoscopic Club for the study and treatment of esophageal varices (NIEC). *Gastroenterology* 2000; **119**: 181-187
- 56 **Burak KW**, Lee SS, Beck PL. Portal hypertensive gastropathy and gastric antral vascular ectasia (GAVE) syndrome. *Gut* 2001; **49**: 866-872
- 57 **Kamath PS**, Lacerda M, Ahlquist DA, McKusick MA, Andrews JC, Nagorney DA. Gastric mucosal responses to intrahepatic portosystemic shunting in patients with cirrhosis. *Gastroenterology* 2000; **118**: 905-911
- 58 **Canlas KR**, Dobozi BM, Lin S, Smith AD, Rockey DC, Muir AJ, Agrawal NM, Poleski MH, Patel K, McHutchison JG. Using capsule endoscopy to identify GI tract lesions in cirrhotic patients with portal hypertension and chronic anemia. *J Clin Gastroenterol* 2008; **42**: 844-848
- 59 **Barakat M**, Mostafa M, Mahran Z, Soliman AG. Portal hypertensive duodenopathy: clinical, endoscopic, and histopathologic profiles. *Am J Gastroenterol* 2007; **102**: 2793-2802
- 60 **Selinger CP**, Ang YS. Gastric antral vascular ectasia (GAVE): an update on clinical presentation, pathophysiology and treatment. *Digestion* 2008; **77**: 131-137
- 61 **Sebastian S**, O'Morain CA, Buckley MJ. Review article: current therapeutic options for gastric antral vascular ectasia. *Aliment Pharmacol Ther* 2003; **18**: 157-165
- 62 **Lecleire S**, Ben-Soussan E, Antonietti M, Gorla O, Riachi G, Lerebours E, Ducrotte P. Bleeding gastric vascular ectasia treated by argon plasma coagulation: a comparison between patients with and without cirrhosis. *Gastrointest Endosc* 2008; **67**: 219-225
- 63 **Fuccio L**, Zagari RM, Serrani M, Eusebi LH, Grilli D, Cennamo V, Laterza L, Asioli S, Ceroni L, Bazzoli F. Endoscopic argon plasma coagulation for the treatment of gastric antral vascular ectasia-related bleeding in patients with liver cirrhosis. *Digestion* 2009; **79**: 143-150
- 64 **Wells CD**, Harrison ME, Gurudu SR, Crowell MD, Byrne TJ, Depetris G, Sharma VK. Treatment of gastric antral vascular ectasia (watermelon stomach) with endoscopic band ligation. *Gastrointest Endosc* 2008; **68**: 231-236
- 65 **Gross SA**, Al-Haddad M, Gill KR, Schore AN, Wallace MB. Endoscopic mucosal ablation for the treatment of gastric antral vascular ectasia with the HALO90 system: a pilot study. *Gastrointest Endosc* 2008; **67**: 324-327
- 66 **Cho S**, Zanati S, Yong E, Cirocco M, Kandel G, Kortan P, May G, Marcon N. Endoscopic cryotherapy for the management of gastric antral vascular ectasia. *Gastrointest Endosc* 2008; **68**: 895-902
- 67 **Shen EF**, Arnott ID, Plevris J, Penman ID. Endoscopic ultrasonography in the diagnosis and management of suspected upper gastrointestinal submucosal tumours. *Br J Surg* 2002; **89**: 231-235
- 68 **Lahoti S**, Catalano MF, Alcocer E, Hogan WJ, Geenen JE. Obliteration of esophageal varices using EUS-guided sclerotherapy with color Doppler. *Gastrointest Endosc* 2000; **51**: 331-333
- 69 **de Paulo GA**, Ardengh JC, Nakao FS, Ferrari AP. Treatment of esophageal varices: a randomized controlled trial comparing endoscopic sclerotherapy and EUS-guided sclerotherapy of esophageal collateral veins. *Gastrointest Endosc* 2006; **63**: 396-402; quiz 463
- 70 **Suzuki T**, Matsutani S, Umebara K, Sato G, Maruyama H, Mitsuhashi O, Nakano Y, Fukamachi T, Saisho H. EUS changes predictive for recurrence of esophageal varices in patients treated by combined endoscopic ligation and sclerotherapy. *Gastrointest Endosc* 2000; **52**: 611-617
- 71 **Irisawa A**, Saito A, Obara K, Shibukawa G, Takagi T, Shishido H, Sakamoto H, Sato Y, Kasukawa R. Endoscopic recurrence of esophageal varices is associated with the specific EUS abnormalities: severe periesophageal collateral veins and large perforating veins. *Gastrointest Endosc* 2001; **53**: 77-84
- 72 **Lee YT**, Chan FK, Ng EK, Leung VK, Law KB, Yung MY, Chung SC, Sung JJ. EUS-guided injection of cyanoacrylate for bleeding gastric varices. *Gastrointest Endosc* 2000; **52**: 168-174
- 73 Abstracts of the British Society of Gastroenterology Annual General Meeting. March 23-26, 2009. Glasgow, Scotland. *Gut* 2009; **58** Suppl 1: A1-156
- 74 **Lu Y**, Gao R, Liao Z, Hu LH, Li ZS. Meta-analysis of capsule endoscopy in patients diagnosed or suspected with esophageal varices. *World J Gastroenterol* 2009; **15**: 1254-1258
- 75 **de Franchis R**, Eisen GM, Laine L, Fernandez-Urien I, Herrierias JM, Brown RD, Fisher L, Vargas HE, Vargo J, Thompson J, Eliakim R. Esophageal capsule endoscopy for screening and surveillance of esophageal varices in patients with portal hypertension. *Hepatology* 2008; **47**: 1595-1603
- 76 **Meagher S**, Yusoff I, Kennedy W, Martel M, Adam V, Barkun A. The roles of magnetic resonance and endoscopic retrograde cholangiopancreatography (MRCP and ERCP) in the diagnosis of patients with suspected sclerosing cholangitis: a cost-effectiveness analysis. *Endoscopy* 2007; **39**: 222-228
- 77 **Ponsioen CY**, Vrouenraets SM, van Milligen de Wit AW, Sturm P, Tascilar M, Offerhaus GJ, Prins M, Huijbregtse K, Tytgat GN. Value of brush cytology for dominant strictures in primary sclerosing cholangitis. *Endoscopy* 1999; **31**: 305-309
- 78 **Lindberg B**, Arnelo U, Bergquist A, Thörne A, Hjerpe A, Granqvist S, Hansson LO, Tribukait B, Persson B, Broomé U. Diagnosis of biliary strictures in conjunction with endoscopic retrograde cholangiopancreatography, with special reference to patients with primary sclerosing cholangitis. *Endoscopy* 2002; **34**: 909-916
- 79 **Moreno Luna LE**, Kipp B, Halling KC, Sebo TJ, Kremers WK, Roberts LR, Barr Fritcher EG, Levy MJ, Gores GJ. Advanced cytologic techniques for the detection of malignant pancreaticobiliary strictures. *Gastroenterology* 2006; **131**: 1064-1072
- 80 **Tischendorf JJ**, Krüger M, Trautwein C, Duckstein N, Schneider A, Manns MP, Meier PN. Cholangioscopic characterization of dominant bile duct stenoses in patients with primary sclerosing cholangitis. *Endoscopy* 2006; **38**: 665-669
- 81 **Itoi T**, Sofuni A, Itokawa F, Tsuchiya T, Kurihara T, Ishii K, Tsuji S, Moriyasu F, Gotoda T. Peroral cholangioscopic diagnosis of biliary-tract diseases by using narrow-band imaging (with videos). *Gastrointest Endosc* 2007; **66**: 730-736
- 82 **Tischendorf JJ**, Meier PN, Schneider A, Manns MP, Krüger M. Transpapillary intraductal ultrasound in the evaluation of dominant bile duct stenoses in patients with primary sclerosing cholangitis. *Scand J Gastroenterol* 2007; **42**: 1011-1017
- 83 **Fishman DS**, Tarnasky PR, Patel SN, Rajiman I. Management of pancreaticobiliary disease using a new intra-ductal endoscope: the Texas experience. *World J Gastroenterol* 2009; **15**: 1353-1358
- 84 **Meining A**, Frimberger E, Becker V, Von Delius S, Von

- Weyhern CH, Schmid RM, Prinz C. Detection of cholangiocarcinoma in vivo using miniprobe-based confocal fluorescence microscopy. *Clin Gastroenterol Hepatol* 2008; **6**: 1057-1060
- 85 **Moon JH**, Ko BM, Choi HJ, Koo HC, Hong SJ, Cheon YK, Cho YD, Lee MS, Shim CS. Direct peroral cholangioscopy using an ultra-slim upper endoscope for the treatment of retained bile duct stones. *Am J Gastroenterol* 2009; **104**: 2729-2733
- 86 **Diaz-Sanchez A**, Nuñez-Martinez O, Gonzalez-Asanza C, Matilla A, Merino B, Rincon D, Beceiro I, Catalina MV, Salcedo M, Bañares R, Clemente G. Portal hypertensive colopathy is associated with portal hypertension severity in cirrhotic patients. *World J Gastroenterol* 2009; **15**: 4781-4787
- 87 **Hosking SW**, Smart HL, Johnson AG, Triger DR. Anorectal varices, haemorrhoids, and portal hypertension. *Lancet* 1989; **1**: 349-352
- 88 **Sato T**, Yamazaki K, Toyota J, Karino Y, Ohmura T, Suga T. The value of the endoscopic therapies in the treatment of rectal varices: a retrospective comparison between injection sclerotherapy and band ligation. *Hepatol Res* 2006; **34**: 250-255
- 89 **Tabasco-Minguillán J**, Jain A, Naik M, Weber KM, Irish W, Fung JJ, Rakela J, Starzl TE. Gastrointestinal bleeding after liver transplantation. *Transplantation* 1997; **63**: 60-67
- 90 **Hirata M**, Kita Y, Harihara Y, Hisatomi S, Sano K, Mizuta K, Yoshino H, Sugawara Y, Takayama T, Kawarasaki H, Hashizume K, Makuuchi M. Gastrointestinal bleeding after living-related liver transplantation. *Dig Dis Sci* 2002; **47**: 2386-2388
- 91 **Sharma S**, Gurakar A, Camci C, Jabbour N. Avoiding pitfalls: what an endoscopist should know in liver transplantation--part II. *Dig Dis Sci* 2009; **54**: 1386-1402
- 92 **Cairns SR**, Scholefield JH, Steele RJ, Dunlop MG, Thomas HJ, Evans GD, Eaden JA, Rutter MD, Atkin WP, Saunders BP, Lucassen A, Jenkins P, Fairclough PD, Woodhouse CR. Guidelines for colorectal cancer screening and surveillance in moderate and high risk groups (update from 2002). *Gut* 2010; **59**: 666-689
- 93 **Vera A**, Gunson BK, Ussatoff V, Nightingale P, Candinas D, Radley S, Mayer AD, Buckels JA, McMaster P, Neuberger J, Mirza DF. Colorectal cancer in patients with inflammatory bowel disease after liver transplantation for primary sclerosing cholangitis. *Transplantation* 2003; **75**: 1983-1988
- 94 **Jue TL**, Imperial JC. Management of post-liver-transplant biliary strictures: a work in progress. *Gastrointest Endosc* 2008; **67**: 886-889
- 95 **Boraschi P**, Braccini G, Gigoni R, Sartoni G, Neri E, Filipponi F, Mosca F, Bartolozzi C. Detection of biliary complications after orthotopic liver transplantation with MR cholangiography. *Magn Reson Imaging* 2001; **19**: 1097-1105
- 96 **Wright H**, Sharma S, Gurakar A, Sebastian A, Kohli V, Jabbour N. Management of biliary stricture guided by the Spyglass Direct Visualization System in a liver transplant recipient: an innovative approach. *Gastrointest Endosc* 2008; **67**: 1201-1203
- 97 **Zheng RQ**, Mao R, Ren J, Xu EJ, Liao M, Wang P, Lu MQ, Yang Y, Cai CJ, Chen GH. Contrast-enhanced ultrasound for the evaluation of hepatic artery stenosis after liver transplantation: potential role in changing the clinical algorithm. *Liver Transpl* 2010; **16**: 729-735
- 98 **Wojcicki M**, Milkiewicz P, Silva M. Biliary tract complications after liver transplantation: a review. *Dig Surg* 2008; **25**: 245-257
- 99 **Aabakken L**, Bretthauer M, Line PD. Double-balloon enteroscopy for endoscopic retrograde cholangiography in patients with a Roux-en-Y anastomosis. *Endoscopy* 2007; **39**: 1068-1071

S- Editor Tian L L- Editor Kerr C E- Editor Li JY



Role of *ATG16L*, *NOD2* and *IL23R* in Crohn's disease pathogenesis

Saleh A Naser, Melissa Arce, Anam Khaja, Marlene Fernandez, Najih Naser, Sammer Elwasila, Saisathya Thanigachalam

Saleh A Naser, Melissa Arce, Anam Khaja, Marlene Fernandez, Sammer Elwasila, Saisathya Thanigachalam, Department of Molecular Biology and Microbiology, Burnett School of Biomedical Sciences, College of Medicine, University of Central Florida, Orlando, FL 32816, United States

Najih Naser, Division of Research and Development, BioSynse, Cary, NC 27513, United States

Author contributions: All authors made substantial contributions to conception and design, drafting the article and revising it critically for important intellectual content; and all authors approved the version to be published.

Supported by The Broad Foundation grant, No. IBD-0207R

Correspondence to: Saleh A Naser, PhD, Professor, Graduate, Coordinator, Department of Molecular Biology and Microbiology, Burnett School of Biomedical Sciences, College of Medicine, University of Central Florida, Orlando, FL 32816, United States. nasers@mail.ucf.edu

Telephone: +1-407-8230955 Fax: +1-407-8230956

Received: April 1, 2011 Revised: August 8, 2011

Accepted: August 27, 2011

Published online: February 7, 2012

key in acquiring CD. Many studies have proven the link between mutations in the *ATG16L*, *NOD2/CARD15*, *IBD5*, *CTLA4*, *TNFSF15* and *IL23R* genes, and CD. The purpose of this review is to examine all genetic aspects and theories of CD, including up to date multiple population studies performed worldwide.

© 2012 Baishideng. All rights reserved.

Key words: Crohn's disease; *ATG16L*; *NOD2/CARD15*; *IBD5*; *CTLA4*; *TNFSF15*; *IL23R*

Peer reviewer: José Manuel Martín-Villa, PhD, Professor, Department of Immunología, Facultad de Medicina, Universidad Complutense de Madrid, Pabellón V. Planta 4^a, Madrid 28040, Spain

Naser SA, Arce M, Khaja A, Fernandez M, Naser N, Elwasila S, Thanigachalam S. Role of *ATG16L*, *NOD2* and *IL23R* in Crohn's disease pathogenesis. *World J Gastroenterol* 2012; 18(5): 412-424 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/412.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.412>

Abstract

Inflammatory bowel disease is a group of diseases that includes Crohn's disease (CD) and ulcerative colitis. CD is characterized as a chronic inflammatory disease of the gastrointestinal tract, ranging from the mouth to the anus. Although there are gross pathological and histological similarities between CD and Johne's disease of cattle, the cause of CD remains controversial. It is vital to understand fully the cause of this disease because it affects approximately 500 000 people in North America and Europe. It ranges from 27 to 48 cases per 100 000 people. There are many theories on the cause of CD ranging from possible association with environmental factors including microorganisms to imbalance in the intestinal normal flora of the patients. Regardless of the environmental trigger, there is strong evidence that a genetic disposition is a major

INTRODUCTION

The first description of Crohn's disease (CD) was made in 1769 by an Italian physician, Giovanni Battista Morgagni, when he diagnosed a man with chronic diarrhea. In 1898, consecutive cases were reported by John Berg, and then in 1904 by Antoni Lesniowski. Throughout the 1920s and 1930s, young adults were thought to have this same condition as they suffered from symptoms, such as abdominal cramps, diarrhea, fever and significant weight loss. In 1923, surgeons at Mt. Sinai Hospital in New York had also identified patients with comparable symptoms. In addition, in 1930, Dr. Burrill B Crohn saw a connection between this unknown debilitating condition and two of his patients. Consequently, Dr. Crohn and his

colleagues presented a paper in 1932, "Regional ileitis: a pathologic and chronic entity", describing the features of this disease to the American Medical Association. CD was named after Burrill Crohn and it became an official medical entity in 1932.

CD is a chronic inflammatory disease that can affect any portion of the digestive tract including the mouth, esophagus, and small and large intestines, but is most common in the ileum. It is also characterized as an autoimmune disease in which the body attacks itself and causes inflammation. In its mild form, it causes erosions called aphthous ulcers in the inner surface of the bowel. In severe cases, deeper and larger ulcers develop that can lead to bowel obstruction and holes in the bowel wall. If a hole in the bowel wall arises, infection to neighboring organs can occur. CD branches off to many forms depending on the location of erosion. If these symptoms take place within the large intestine, it is called Crohn's or granulomatous colitis. If it occurs in the small intestine, it is known as Crohn's enteritis, and more specifically if it occurs in the ileum, it is called Crohn's ileitis. During severe cases in which the small and large intestine are both involved, it is known as Crohn's enterocolitis or ileocolitis. CD is separated into three phenotypes: non-stricturing and non-penetrating, stricturing and penetrating CD^[1,2].

CD is most prevalent in North America and Europe, and least prevalent among African Americans and Asians^[3]. It affects approximately 500 000 people in North America and Europe. It ranges from 27 to 48 cases per 100 000 people. There is no difference in prevalence among males or females. Individuals with siblings affected by CD have a higher risk of acquiring the disease. There is evidence of a cause from environmental factors, thus, there are a higher number of cases in western industrialized countries. Symptoms of CD typically begin in the teens and twenties and then go into remission on and off with appropriate therapies. There is a peak incidence between 50 years and 70 years of age, which often leads to major complications due to age and the necessity of surgery.

Although the etiology is still unknown, there are many theories about what causes CD and ulcerative colitis (UC). Many believe it is caused by environmental factors, such as certain foods, bacteria, viruses, or cigarette smoke, which all can trigger an immune system response. Scientists have linked inflammatory bowel diseases (IBDs) as an autoimmune problem. In a healthy person, the immune system defends the body against harmful microbes that have entered it. Upon triggering the immune system, an inflammatory response occurs in which immune cells aggregate at the site of infection and overcome the threat. There are microbes native to our bodies that are useful rather than harmful to which the immune system does not trigger a response. In patients with CD, the immune system will attack these native luminal bacteria, disrupting the normal flora, thus characterizing this condition as an autoimmune disease.

Recent research has indicated specific genetic variations as a direct cause of CD and UC. The genetic aspects of CD have been linked by observing familial clustering of IBD cases^[4]. Genetic variations in the *ATG16L*, *NOD2/CARD15* and *IL23R* genes have strongly been linked to the onset of CD^[5]. Not only are individual gene mutations listed as a cause of CD, but a combination of them has also been shown in CD patients by conducting many population studies. There have also been many studies that have predicted surgical outcomes in both adults and children with specific genetic variations.

Diagnosis of CD can be tricky and requires a number of tests to be certain. Colonoscopy is the most effective way to diagnose CD but not in all cases because it only allows the physician to visualize the colon, ileum, and lower portion of the small intestine^[6]. If the ulcers are located within the upper portion of the small intestine, this test will not be effective. In this case, a barium follow-through X-ray is useful because barium sulfate gives fluoroscopic images of the bowel and the physician can see areas of inflammation and narrowing^[6]. Another effective method of diagnosis is the use of white blood cell scans. During this procedure, white blood cells are tagged with a radioisotope and then injected back into the patient. At specific intervals of time, the scan can locate accumulations of white blood cells in the intestine at the site of CD. This method is also useful to monitor the disease and show effectiveness in other therapies. Furthermore, a simple blood test can also diagnose CD because it can determine whether the patient is anemic or has a vitamin B12 deficiency because vitamin B12 is absorbed in the ileum and a deficiency can be due to ileitis.

CD causes a wide variety of symptoms and can be confused with UC, which is a similar disease, both under the group of IBDs. UC is only found in the colon and affects the mucosal membrane, whereas CD can occur anywhere throughout the gastrointestinal (GI) tract and affects the thickness of the GI wall. Individuals with CD can experience flare-ups followed by remissions. It can vary from one flare-up in a lifetime to multiple flare-ups that need surgical treatment. Symptoms include persistent diarrhea, abdominal pain in the affected area, fever, and weight loss^[6]. There are also signs and symptoms that may occur unrelated to the GI tract, such as reddening and inflammation of the eye, joint pain, skin lesions, and sores inside the mouth.

Currently, there is no cure for CD. Treatment is focused on relieving the symptoms and putting it into remission. Since CD is characterized as an autoimmune disease, medications to suppress the immune system include 5-aminosalicylic acid and steroids, such as prednisone^[6]. Antibiotics such as clarithromycin, ampicillin and metronidazole, can also be used. More than 50% of patients with CD will have to undergo surgical treatment to correct a fistula, drain an abscess, open a narrow or obstructed bowel, or remove a segment of infected intestine.

AUTOPHAGY-RELATED 16-LIKE 1 PROTEIN COMPLEX

Autophagy is a catabolic process of intracellular degradation in which cytoplasmic components are sequestered within vesicles and delivered to the lysosomes. Cells use this pathway during nutrient starvation because they can break down non-vital components and use them as nutrients. Autophagy plays a role during infection by helping rid the cell of foreign antigens by breakdown of the pathogen. This pathway can also be implemented as a repair mechanism to degrade damaged organelles and proteins. Autophagosomes, formed by the fusion of lysosomes and vesicles, are also implicated in the processing of intracellular bacteria. If the gene responsible for autophagy is mutated, it can cause a shift in normal flora as previously mentioned, and lead to many GI problems, which has been pointed out as a possible cause of CD. If the cell cannot regain nutrients or fight off foreign antigens within the GI tract, these cells will undergo programmed cell death and cause tissue damage. This damage can be seen as lesions and ulcers within the intestines, creating dead infected patches of tissue along the GI tract. The only option of treatment is surgery to remove the sections of the intestines with the diseased tissue so that the necrosis will not spread among neighboring cells.

An autophagosome, a double-membraned vesicle formed by autophagy, envelops part of the cytoplasm and delivers it to the lysosomes where it is degraded and recycled. There have been approximately 30 autophagy-related (Atg) genes identified, with two proteins having ubiquitin-like characteristics, Atg12 and Atg8^[7]. These proteins covalently modify their target protein with molecules such as ubiquitin-like proteins to tag them for degradation. Both proteins also contain a conserved ubiquitin-fold region^[7]. Autophagosomes use two conjugation systems, the Atg12 and LC3-II systems^[8]. These systems were first discovered during yeast genetic studies revealing a set of 17 ATG genes involved in the autophagy pathway^[9]. In the Atg12 conjugation system, an Atg12-Atg5-Atg16L complex forms, and dissociates from the membrane just before or after completion of the autophagosome^[8]. The ATG16L1 protein is expressed in the colon, small intestine, intestinal epithelial cells, leukocytes, and spleen^[8]. Recent independent studies have shown that an ATG16L mutation, located on chromosome 2, is associated with the onset of ileal CD, and is therefore a key molecule in elucidating the genetic aspects of this disease^[10].

Multiple studies have been performed with each resulting in the same conclusion that ATG16L is implicated in CD. During a genome-wide survey of 19 779 non-synonymous single nucleotide polymorphisms (SNPs), Thr300Ala within the N terminus of ATG16L was found to be highly associated with CD by using a haplotype and regression analysis^[4]. This study used a total of 735 CD patients, 368 controls and 72 SNPs. A sec-

ond report from the North American CD genome-wide study submitted by Rioux and colleagues also shows an association with ATG16L using a case-control analysis in 988 CD patients and 1007 controls^[11]. Another group of German and British collaborators demonstrated that rs2241880, another non-synonymous variant of the *ATG16L* gene on chromosome 2q37.1, is implicated in the autophagy pathway^[8].

In a study performed on an Italian cohort, the same polymorphism, rs2241880, was observed in 667 CD and 668 UC patients^[12]. Both the frequency of the G allele and number of carriers of the G allele were increased in CD patients when compared to the controls^[12]. These differences were only significant in the adult subgroup, which could be due to the small sample size of the pediatric subgroup. In comparison, there were no significant allele or genotype frequencies found between UC patients and controls for the groups as a whole^[12]. During analysis of genotype/phenotype correlation of the rs2241880 SNP, there were no associations with disease location, behavior, and age at diagnosis based on the Montreal Classification of CD^[12]. There were also no associations found in sex, smoking, and perianal fistulae^[12]. For the rs2241880 variant, recent studies, specifically by Prescott^[13], demonstrate an association with the ileal form of CD with or without colonic involvement, but not with isolated colonic disease^[12].

In addition, a study from Oxford compared 645 CD patients with 1190 controls and showed an association of ATG16L1 with CD^[14]. To understand fully the function of this gene, a study utilized oligo-based silencing RNA directed against ATG16L1 isoforms, where autophagy was induced by *Salmonella typhimurium* in ATG16L1 knockdown HEK293 cells. There was a significant difference between the knockdown cells compared to the control cells during the autophagy pathway^[11]. It is clear to say that variants of this gene have been proven without a doubt to be directly associated with CD because autophagy plays a critical role in disease pathogenesis. Further research needs to focus on understanding how ATG16L1 variants contribute to disease susceptibility in IBD patients, and their possible therapeutic implications.

TUMOR NECROSIS FACTOR SUPER FAMILY 15

Tumor necrosis factor super family 15 (TNFSF15) is a Th-1 polarizing cytokine involved in systemic inflammation. TNF functions to regulate immune cells, induce apoptosis, induce inflammation, and inhibit tumorigenesis. TNFs are produced by macrophages, lymphoid cells, mast cells and endothelial cells. During immunological studies, it has been found that CD patients have an increased expression of TNFSF by multiple cells in the intestinal tissues when compared to controls^[7]. The *TNFSF15* gene is a candidate for increased susceptibility in IBD. It binds to a specific T-cell receptor to enhance

cytokine-induced interferon expression in mucosal CD4⁺ T cells^[7].

In 2005, the first genome study involving IBD tested nearly 80 000 SNPs in Japanese CD patients. This study identified haplotypes within the *TNFSF15* gene, which included seven SNPs within a 280-kb region on chromosome 9q32^[15]. By resequencing *TNFSF15* from the same CD cases but with a new control group, *TNFSF15* was found to be strongly associated with CD with an odds ratio (OR) of 2.17 (95% CI: 1.78-2.66), $P = 1.71 \times 10^{-14}$ ^[15]. The Japanese wanted to see if the same patterns were seen in other population groups so they replicated their associate in two panels from Oxford, United Kingdom. Although the risk haplotype was identified in both cohorts, there was a weaker effect size ($P = 0.02$ in both family-based and case-control association panels). In addition, another study involving a Jewish cohort also showed an association of *TNFSF15* with CD, and also suggested that in response to FC-gamma receptor stimulation, *TNFSF15* gene variation aggravates induction of *TNFSF15*^[16]. However, in a separate study using a Belgian CD cohort, no significant association was observed between CD and *TNFSF15*^[4]. This may have been due to different marker genotypes or differences in susceptibility genes between Asian and European cohorts^[4]. Not enough studies have been performed regarding *TNFSF15* and its possible implications in CD. Future studies have to focus on different populations to provide efficient insight.

NOD2/CARD15 GENE

Nucleotide-binding oligomerization domain containing 2 (NOD2), located on chromosome 16q12, is a protein that plays an essential role in the immune system by controlling commensal bacterial flora in the intestine^[17]. NOD2 belongs to a nucleotide-binding domain, leucine-rich repeat family of cytoplasmic proteins that may detect a variety of bacteria by acting as an intracellular sensor for bacterial peptidoglycan^[18]. It has the ability to respond to N-acetylmuramyl-L-alanyl-D-isoglutamine (MDP) *via* the leucine-rich repeat domain. The MDP is conserved in both Gram-negative and positive bacteria^[10,19]. This leucine-rich repeat domain plays a role in protein-protein interactions and the middle portion of the protein is responsible for self-oligomerization. The N-terminal portion of NOD2 contains two caspase recruitment domains (CARDs) which play a role in apoptosis. The *CARD15* gene, which encodes for the CARD domain within NOD2, has been specifically identified as a genetic factor for CD. Three SNPs were found to be independently associated with CD: rs2066844, rs2066845, and an insertion mutation 3020insC^[20]. Each variant may result in distinct phenotypic expression of CD^[4]. To activate NOD2, Rip2 kinase is required because it is necessary for downstream signaling of NOD2 and signaling cascades such as nuclear factor (NF)- κ B and mitogen-activated protein kinase cascades^[21].

The intestinal mucosa is constantly exposed to a large number of commensal microorganisms; the majority of which inhabit the large intestine. In a healthy individual, there is a basal immune response elicited from the interaction between the intestinal immune system and commensal bacteria. This immune response is constantly present to protect the host from pathogenic and non-pathogenic bacteria. When there are changes within this balance, the intestines are susceptible to chronic intestinal inflammatory conditions, such as CD^[21]. Multiple genetic studies have linked NOD2 with susceptibility to CD. However, NOD2-deficient mice do not develop colitis, suggesting that dysregulation of the NOD2 pathway is not sufficient to provoke CD^[22]. This is not a surprise because the pathogenesis of CD is caused by several factors including environmental, dysfunctional immune system, and a shift in normal bacterial flora^[21].

NOD2 is expressed in Paneth cells, which are found in the intestines. The exact functions of Paneth cells are still unknown, but they are likely to contribute to the host defense by secreting antibacterial compounds due to the presence of lysozyme. A NOD2 mutation can alter the function of Paneth cells, which alters their antimicrobial activity and leads to the development of ileal lesions, which correspond to the location of these cells. Of the three NOD2 mutations associated with CD, both rs2066844 and rs2066845 are the result of a two-amino-acid substitution in which rs2066844 is encoded by exon 4 and rs2066845 is encoded by exon 8^[23]. The variant 1007fs is created by a frameshift mutation in exon 11^[23]. Each of the three mutations occurs within or near the leucine-rich repeat and decrease the cells ability to activate NF- κ B in response to peptidoglycan^[18]. Interestingly, these mutations are observed in Caucasian patients but not in Japanese, Chinese and Korean patients with IBD, and they are very rare in African Americans with IBD^[24,25]. An individual heterozygous for at least one NOD2 mutation is at a 2-4-fold increased risk of developing CD, whereas a homozygous individual for at least one NOD2 mutation is at a 20-40-fold increased risk when compared to healthy individuals^[14].

Studies of each individual mutation in NOD2 have shown that 1007fs causes a decrease in defensin expression^[26]. Defensins are cells of the immune system that assist in killing phagocytized bacteria. They function by binding to the microbial cell membrane and forming a pore in the membrane that allows outward flow of nutrients and essential ions. CD patients who are homozygous and/or heterozygous for NOD2 mutations typically have lower defensin levels in their ileostomy fluid^[17]. As a result of a mutation in NOD2, the function of this protein is diminished, therefore allowing subsequent entry of bacteria into epithelial cells because they are no longer able to recognize them. This in turn alters the bacterial population in the intestines and thus, defensins are not able to function correctly because they are working in an impaired bactericidal capacity^[22].

In a recent study by Van Limbergen *et al*^[17], it was

proven that NOD2 is required for the regulation of commensal microbiota in the intestine. The regulation of NOD2 depends on the downstream kinase Rip2 because Rip2-deficient mice fail to establish and regulate commensal bacteria in their terminal ileum. NOD2-deficient mice do not develop spontaneous intestinal inflammation. In conclusion, the NOD2–Rip2 pathway is critical for the regulation of homeostasis between the body's normal bacterial flora and innate immunity^[17]. As a controlled balance, it has been found that the expression of NOD2 and Rip2 is dependent upon and regulated by the presence of commensal bacteria^[17]. This creates a negative feedback by which the commensal bacteria positively regulate NOD2, and in turn negatively regulate the normal flora. NOD2 mutations directly affect the ileum in CD patients, thus, it is possible that they are responsible for the composition of the bacterial flora in the terminal ileum^[17]. This may facilitate both disease pathology and progression.

Although the exact mechanism by which NOD2 contributes to the control of commensals in the intestines is still not known, there are many possible theories proposed by this study. The first theory entertains the possibility that NOD2 regulates the commensal flora through the bactericidal activity of ileal crypt secretions^[17]. The second theory is that NOD2 regulates the adaptive immune system by inducing lymphoid tissue genesis^[17]. The third theory describes how NOD2 expression in myeloid lineage cells contributes indirectly to maintain the normal microbiota flora^[17]. The fourth theory states that there may be other cells in the intestines, in addition to Paneth cells, that may play a role in the regulation of commensal bacteria in the intestines^[17]. Further research is needed to elucidate this mechanism.

It has been reported that both adults and pediatric cohorts share a strong association between the NOD2 variants and ileal disease location^[23]. Some studies have suggested that adult patients show a correlation in fibrostenotic behavior and NOD2 mutation status, whereas other studies have failed to replicate the same phenotypic effect^[12]. It is necessary to study different CD cohorts from different countries to obtain an accurate effect of each mutation. Exploring the genotype-phenotype interactions in children is beneficial because they show a higher gene dosage and have less environmental influences.

A study by Lacher and colleagues has explored the association of the NOD2 mutation in German pediatric CD patients and the risk of surgery. The risk of surgery in children is important to predict because surgery is sometimes the only method of treatment. Out of 171 young CD patients, 78 (45.6%) carried at least one NOD2 mutation, with 11 being compound heterozygous, and 14 being homozygous for two NOD2 mutations^[14]. The presence of rs2066844 was found in 29 (17%) children, rs2066845 was found in 18 (10.5%), and 1007fs was found in 42 (24.6%)^[14]. Overall, one out of three German children with CD had at least one NOD2 mutation. In comparison, 36% of adult CD patients

were heterozygous for at least one NOD2 mutation: 17.2% for rs2066844, 9.6% for rs2066845, and 11.7% for 1007fs^[14]. In conclusion, the genetic alterations were observed more predominantly in German pediatric CD patients than in adults. The study also looked into each mutation and its association with localization and symptoms of the disease. A 4.73-fold increased risk of isolated ileal localization was observed in patients that were identified as 1007fs carriers when compared to children with none of the NOD2 mutations^[14]. Although the 1007fs carriers were predisposed to ileal disease, there was no involvement of the ileocolonic or upper GI tract^[14]. The next characterization the study explored was the association of NOD2 mutations and stricturing disease or perianal fistulae in their German pediatric CD cohort. Only 17% of patients showed a stricturing phenotype and 18.1% had a perianal fistula^[14]. Among the children that showed a stricturing phenotype, 79.3% required surgery and those with the 1007fs mutation had surgical complications^[14]. The outcome showed a 9.8-fold increase in surgical complications when the child carried at least one allele for the 1007fs mutation^[14]. Not only is the 1007fs mutation strongly associated with isolated ileal disease, but children are at a high risk for surgical intervention. Therefore, this mutation can act as a prognostic tool in Caucasian children with CD^[14].

Another study by Jurgens and colleagues has investigated the presence of fistulas and their association with NOD2 homozygosity and how they predict intestinal stenosis in CD patients. It was observed that patients with fistulas had simultaneous intestinal stenosis^[15]. In another study using the same research, it was found that NOD2/CARD15 variants, especially with 1007fs homozygosity, could predict the occurrence of intestinal stenosis^[27]. By using a strict screening process based on phenotypes including stenoses (stricturing CD) and fistulae (penetrating CD), and genotype, the study isolated a total of 145 patients. One hundred and twenty-five of the patients had penetrating CD with simultaneous stenosis within a 6-mo interval^[28]. It was also found that all 14 CD patients homozygous for NOD2 variants suffered from stenosis^[28]. To no one's surprise, 11 out of the 14 patients with stenosis and fistulas carried the NOD2 1007fs mutation^[28]. These data confirm the strong risk factor of the 1007fs NOD2 mutation in the prevalence of intestinal stenosis, which may be related to the decreased intestinal barrier function found in CD patients with NOD2/CARD15 variant mutations^[28]. This study has isolated the 1007fs variant as the cause in disrupting the intestinal barrier and causing the many problems often observed in CD patients.

To investigate whether stenosis occurs in a specific anatomical region, 223 patients with 248 stenoses located at different intestinal regions were examined^[28]. The most common anatomical region was the terminal ileum with 68.8% of stenoses, followed by 11.7% found in the rectosigmoidal segment, and 9.3% in the jejunum or proximal ileum^[28]. In conclusion, homozygosity in the NOD2/

CARD15 mutation is a strong risk factor for intestinal stenosis^[28]. When the study looked into the possible association between stenosis and interleukin-23 receptor (IL-23R) mutation variants, there was a weak connection with no influence on stenosis^[28]. This study suggests classifying CD into four disease phenotypes instead of the current three in the Montreal Classification System. The four new classifications would be: non-stricturing^[12], non-fistulizing CD^[12]; stricturing, non-fistulizing CD^[18]; non-stricturing, fistulizing CD^[29]; and stricturing, fistulizing CD^[28]. It is important to link any possible association between fistulas and stenoses because there is a strong risk factor for recurrence of CD after surgery^[28].

Many studies have investigated North American, European and Asian countries and their CD patients with specific genotypes. Interestingly, the NOD2/CARD15 mutations are very rare and even absent in Asians (Japanese, Chinese and Korean), Arabs, Africans and African Americans^[28]. A study by Baptista and colleagues has concentrated on a South American population for the first time. There were a total of 187 CD patients used for the study with a median age of 33 years and a median age of onset of 23 years^[24]. Their patients were ethnically classified into the following groups: 58.8% were in the Brazilian subgroup, 36.9% shared a common European ancestry and were in the European-Brazilian subgroup, three were Amerindian-Brazilian, and two were Afro-Brazilian^[24]. The alleles related to CD within the CARD15, rs2066844 and 3020insC variants were significant for CD susceptibility^[24]. Both the rs2066845 and 1007fs variants failed to show any significant association^[24]. Among their patients, 30% had at least one NOD2/CARD15 variant allele^[24]. The frequency of rs2066844 (9.63%) in the Brazilian CD cohort was consistent with the reports for European populations^[24]. In conclusion, this study confirmed that CARD15 variants lead to greater susceptibility to CD in the Brazilian population^[24].

INTERLEUKIN-23 RECEPTOR

IL-23R is a protein consisting of an IL-12 β 1 and an IL-23R chain^[27]. The molecular location of the *IL23R* gene is on chromosome 1 and is formed by the binding of IL-12p40 and a p19 protein^[30]. It is highly expressed on the cell membrane of memory T cells and other immune cells, such as natural killer cells, monocytes, and dendritic cells, which identify foreign substances to defend the body against infection. It is highly involved in the mediation of proinflammatory activities by the production of IL-17 *via* the activation of Th17 lymphocytes^[20]. IL-23R interacts with IL-23, which is a cytokine that regulates the activity of immune cells and plays an important role in the inflammatory response against infection by bacteria and viruses. It also has been suggested that the functional IL23R pathway polymorphisms play a role in modulating neonatal development of intestinal tolerance and bacterial colonization^[4].

Th-17 lymphocytes are a distinct subset of T-helper cells, which mainly produce IL-17 and to a lesser extent IL-6 and TNF- α ^[26]. IL-17 *in vitro* and *in vivo* acts as a potent inflammatory cytokine and is involved in the destruction of cartilage and bone, as seen in rheumatoid arthritis^[12]. It has been reported that IL-23 could be a key regulator in the differentiation of Th-17 lymphocytes from memory T cells. It also has been suggested that IL-23 plays a role in providing a survival advantage to already differentiated Th-17 cells^[21]. The expression of the heterodimeric receptor complex, IL-23R and IL-12R β 1, regulates activities of IL-23^[12]. Therefore, the IL-23-IL17 cytokine axis is a key pathogenic mechanism that mediates the development and progress of inflammation by Th-17 cells. The role for the IL23-IL17 axis in CD patients was supported in human patients and animal models of colitis^[51]. Both cytokines are increased in knockout mouse models of IBD. More specifically, IL-17 levels are increased in both intestinal mucosa and in the serum of CD patients^[12]. In a study using IL-17R knockout mouse models, an association was found with colitis and disease severity^[27]. Therefore, the use of anti-IL-12p40 antibody to treat CD patients is a therapeutic option because it is known to reduce production of IL-23 and IL-17 in the lamina propria cells^[12]. The mechanism of IL-23R is clearly important to understand because it is directly associated with CD.

During a genome-wide association study, 2877 DNA samples from IBD patients (two-thirds CD and one-third UC), identified rs11209026 as a possible protective variant, in the *IL23* gene on chromosome 1p31^[4]. There are many other variants within IL23R that are associated with IBD, but rs11209026 has the strongest association with conferring protection against CD^[4]. Although the effect of IL23R variants is greatest in CD, it may have an overall effect on susceptibility to chronic intestinal inflammation.

Additional studies have confirmed the susceptibility of the *IL23R* gene to CD in North American and European populations. They include cohorts ranging from Scottish pediatric IBD, Belgian CD and an independent cohort of 883 families^[4]. Due to the role of IL-23 in activation of inflammatory responses, targeting this pathway may be a good therapeutic approach. Some promising research is underway using anti-p40, which blocks IL-23 and IL-12 activities. The variant allele, rs11209026, could be exploited to define clinical outcomes, such as a pharmacological approach to mimic the rs11209026 polymorphism^[4].

A study by Schmechel and colleagues has shown a link in susceptibility to CD with Th17 cell function, IL-22 serum levels, and IL23R genotype. IL-22 is a strong activator of proinflammatory gene expression and upregulates SOC3 mRNA in intestinal epithelial cells^[29]. Recent evidence has shown that Th17 cells expressing IL23R play a key role in the mechanism by which IL23R modulates IBD susceptibility^[32]. Previous studies have shown that Th17 plays a role in autoimmune diseases,

such as rheumatoid arthritis and CD. However, Th17 is responsible for the important function of antimicrobial immunity at epithelial barriers where it produces cytokines such as IL-22. Because it produces IL-22 in epithelial barriers such as in the intestines, theoretically by testing for increased IL-22 serum levels, a physician can determine CD and disease activity.

It has been confirmed that IL-22 serum level is increased in CD and correlates with disease activity^[33]. IL-22 serum levels are also independent of CD phenotype and CARD15 genotype, but are modulated by IL23R polymorphisms^[33]. The study investigated IL-17 serum because Th17 cells also produce IL-17. There was no correlation between IL-22 and IL-17 serum levels^[33]. Currently, serum levels of TNF- α and IL-6 are used as inflammation markers in determining CD. There was no difference in TNF- α and IL-6 levels whether CD was active or in remission, whereas IL-22 levels were significantly higher in active CD compared to CD in remission^[33]. Therefore, measuring IL-22 levels are clinically relevant in determining the disease activity in CD patients^[33].

There are also strong associations between IL-22 serum levels and *IL23R* gene variants. Previous studies have shown both protective and inducing variants of IL23R in susceptibility to CD. Although rs1004819 is the CD increasing variant of IL23R, rs11209026 has shown to be a protective IL23R variant against CD. As predicted, when the IL-22 mean serum levels were tested against each variant, the serum levels were highest among SNPs that increased CD risk as opposed to the identified protective SNPs of IL23R, which had low serum levels^[33]. In contrast, the three main variants within NOD2/CARD15 did not show any differences in IL-22 serum levels^[33]. Although, the exact function of IL-22 in human IBD is still unknown, recent studies have observed increased β -defensin-2 expression and intestinal epithelial cell migration and proliferation upon stimulation with IL-22^[29]. These data ultimately suggests a clinically useful marker to assess disease severity and Th17 cell activity in CD patients^[33].

As mentioned above, a rare glutamine allele, rs11209026, in IL23R conferred protection against CD during a recent genome-wide association study. Recent studies have proven that this variant protects against CD in both Jewish and non-Jewish populations. In addition, in a study involving IBD (both UC and CD) patients of Spanish Caucasian origin, the rs11209026 variant was seen to be most significantly associated with IBD protection (OR: 0.4; 95% CI: 0.3-0.7)^[34]. A study by Dubinsky and colleagues has investigated this rare allele further and how it protects pediatric CD patients. They used the transmission disequilibrium test (TDT) analysis and genotyping of whole blood samples from children with IBD and their two parents. This rare rs11209026 SNP was present in 2.67% non-Jewish CD patients and 2.94% of non-Jewish UC patients^[35]. The TDT demonstrated that the allele was under-transmitted in all CD offspring and confirmed

the negative association between rs11209026 and CD^[35].

Another way to confirm the association of alleles with CD and UC is to use a population that is genetically isolated, such as the Finnish population. This will provide an advantage in molecular genetic studies in complex disorders^[36]. A study by Lappalainen has used such a strategy to confirm the association of IL23R, TNFRSF1A, and the HLA-DRB1*0103 allele variants. The strongest association of IL23R with the marker rs2201841 (IL-23R risk variant) showed a frequency of 37.2% in CD. In most studies, the non-synonymous SNP, rs11209026 (the protective IL23R allele), has had the highest association, but is only marginally associated with Finnish CD^[36]. No association has been observed between the IL23R markers and UC patients^[36].

Previous studies have shown that the HLA-DRB1*0103 allele is associated with both UC and CD. In the Finnish population, this allele only has a frequency of 0.6%, which is not statistically significant, whereas it is significant for UC and IBD^[36]. When looking into the genotype-phenotype association, patients carrying the rare HLA-DRB1*0103 allele have colonic involvement in CD^[36]. During the TNFRSF1A analysis, the investigators genotyped a rare A36G variant and an IVS6+10A (rs1800693) variant. CD patients with both variants often show ileocolonic disease in comparison with patients without these two variants^[36]. The weak association is probably due to the small sample size. Interestingly, when the protective haplotype of IL-23R (described in the above paragraph) was sequenced and compared to North American Caucasian CD patients, there was a one-nucleotide difference between Finnish (CCTGATCG) and North American (CGTGATCG) CD patients^[36]. In conclusion, the HLA-DRB1*0103 allele has been confirmed in CD patients, which shows an inherited susceptibility of colonic inflammation. The *TNFRSF1A* gene variants are markers of ileocolonic involvement in CD^[36]. Although this study shows a weaker association of the *IL23R* gene, it confirms the genetic involvement within a Finnish population.

In another population study among French-Canadian and English-Canadian children, the association between genetic variants of the *IL23R* gene and early-onset of CD has been investigated. To study the associations accurately, they have carried out both a case-control and a family-based study. They have targeted the 10 SNPs in IL23R achieved by the genome-wide study and the three CARD15 SNPs. In total, 259 CD patients and 139 controls were recruited with a mean age at diagnosis of 13.3 years (range: 2.6-20 years)^[12]. The IL23R protective allele, rs11209026, was only present in 2% of CD case chromosomes and 6% of the control chromosomes^[12]. All CARD15 variant allele frequencies were higher among CD patients when compared to the control group^[12]. In the *IL23R* gene, they observed a significant association among four SNPs that did not possess any CARD15 variants^[12]. Therefore, variants in the *IL23R* gene were associated with early-onset CD among Canadian children.

This study confirms previously reported findings in CD patients among North Americans.

In continuation of a study that was described in the ATG16L section of this paper, Latiano and colleagues have shown that the replication of IL23R is associated in adult and pediatric onset of IBD in Italy. Approximately 730 CD patients were genotyped for the rs7517847 and rs11209026 variants. For the rs7517847 polymorphism, significant reductions were found in minor allele (G) frequency in CD patients when compared with controls and in the number of carriers^[12]. No differences were found in UC patients. When the rs11209026 polymorphism was examined, a significant increase in the frequency of the risk genotype was observed in CD patients. In either SNP, there were no correlations between phenotypes of CD and risk alleles or genotypes of the *IL23R* gene.

To specify which IL23R variant is the main disease associated variant, a study using German CD patients was conducted. Among all 10 IL23R SNPs, they chose to focus their study on the rs1004819 variant because it had the strongest association to CD when compared to controls. This variant showed high prevalence of ileal involvement when carriers had the TT genotype versus the CC wild-type genotype^[28]. This identification is different from recent data published by Roberts *et al.*^[37], which have identified the rs7517847 variant as having the strongest association with CD, along with other overlapping North American study populations. They have only analyzed ileal cases of CD, therefore, it is assumed that IL23R variants are predisposed to an ileal disease phenotype^[37]. In a recent British study, they could not identify any association between disease phenotype and IL23R variants^[9]. Interestingly, they found that the rs1004819 variant was 1000-fold weaker than that reported in the German study.

Not only are the IL23R variants highly associated with CD, they are also associated with other chronic inflammatory diseases, such as the rs10489629 variant with chronic periodontitis. In psoriasis, the variant rs11209026 has been described as a predisposing haplotype^[17]. In both CD and psoriasis, treatment with an anti-p40 IL-12/23 antibody has shown promising results. Antibodies to the p40 subunit block both IL-12 and IL-23, although in knockout mice studies, it has been proven that IL-23 drives chronic intestinal inflammation^[28]. Therefore, IL23R can serve as a therapeutic target in many different chronic inflammatory diseases, although the variants may differ among the different diseases. It is hypothesized that this is due to alternative mRNA splicing, which results in corresponding IL23R isoforms with different tissue distribution^[28].

IBD5 GENE

The *IBD5* gene is about 250 kb and is located at position 5q31. During a genome-wide linkage analysis, mapping studies have identified a risk haplotype within the *IBD5* locus. Many studies have linked this gene and its two vari-

ants, *SLC22A4* (*OCTN1*) and *SLC22A5* (*OCTN2*), to CD. IBD is believed to originate from an uncontrolled mucosal immunity of the GI tract^[17]. Although these variants are associated with CD, they act independently because there is no statistical evidence for interaction between *IBD5* and the *IL23R*, *ATG16L1* or *CARD15* genes^[12].

CTLA4 VARIANTS

The *CTLA4* gene is a member of the immunoglobulin superfamily and is expressed on the surface of helper T cells. It is located within the 2q33 region, translates into a protein that plays a role in the immune system, and may have a genetic association with IBD. It is a T-cell suppressor, which is essential in the function of the CD25⁺ CD4⁺ regulatory cells^[7]. These regulator cells control the process of intestinal inflammation. Many SNPs have been studied within this gene and it has been found that the rs3087243 variant shows the most association with IBD followed by the rs11571302, rs7565213 and rs11571297 variants^[53]. Although studies have shown that the three variants in the *CTLA4* gene, g.49A > G (rs231775), g.-318C > T (rs5742909), and rs3087243, have no association with CD, other work has suggested that these variants may control the phenotype of CD^[14].

A study performed by Hradsky has shown no crude association between CD and SNPs within the *CTLA4* gene^[12]. The study explored the possibility of interactions in *CTLA4* SNPs with variants in *IL23R* and *NOD2*. The R-project package SNPAssoc was used and significant interactions between the three *CTLA4* variants with *NOD2* p.Leu1007fsX1008 and *IL23R* rs11209026 were observed^[12]. This may be due to complex gene-gene interactions. To characterize further the different variants and whether they determine phenotype in CD patients, the study used a case-only design. They observed a difference of minor allele frequency at the rs3087243 gene between pediatric-onset and adult-onset of CD^[12]. It seems that a genetic factor has a greater impact in early-onset patients when compared to the adult-onset patients^[38]. Within this study, the age of diagnosis and localization of the disease was strongly associated with the rs3087243, rs11571302 and rs11571297 variants^[12].

NOD2, IL23R, OCTN1/2 AND ATG16L1 POLYMORPHISMS

Gene-gene interactions

Interactions of the major IBD alleles show a high susceptibility in CD patients. Gene-gene interaction can either enhance or weaken the effects of an individual gene, therefore making it more important than independent studies looking into the effects of single susceptibility genes. Csongei and colleagues have performed a gene-gene interaction analysis in the Hungarian CD population^[5]. They concentrated on the two *IL23R* gene risk variants (rs2201841 and rs1004819), the *ATG16L1* gene variant (Thr300Ala), and the three *NOD2/CARD15*

variants (rs2066844, L1007fs and rs2066845). Logistic regression analysis showed that the IL23R variants, both rs1004819 and rs2201841 and the NOD2/CARD15 variants (rs2066844 and L1007fs) conferred significant risk for CD. When the patients were homozygous for IL23R (rs1004819 and rs2201841) variants or ATG16L1, there was a highly increased risk for CD. When they analyzed possible statistical interactions between pairs of ATG16L1, IL23R and CARD15 variants, no evidence of interactions was found. Therefore, all examined loci contribute independently to CD risk. Although significant statistical interactions were not detected, these susceptibility factors may have a cumulative effect in the Hungarian population.

Further gene-gene interaction studies have been performed in another population study, as well as assessment of CD genetic risk factors. The study included five SNP variants for IL23R, the functional variants SLC22A4 and SLC22A5, and the ATG16L1 mis-sense risk polymorphism Thr300Ala. All the SNPs, except for rs1495965 of IL23R, showed significant association when carriers were either homozygous or heterozygous for the alleles^[2]. Among them all, rs10889677 of IL23R, showed the strongest association with CD risk^[2]. Unsurprisingly, ATG16L1 also showed a strong association with CD^[2]. None of the SNPs showed an association with UC. When they evaluated correlations between genotype and phenotype for intestinal complications, carriers of the rs7517848 allele of IL23R, particularly those who were homozygous for the allele, were found to be at risk for ileal disease. No other SNPs showed differences in genotype or carrier frequencies when compared to CD patients and their history of complications.

Homozygote variants from the IBD5 and ATG16L1 genes had a greater risk than heterozygotes, suggesting a gene dosage effect^[2]. When SNPs were considered two at a time, the best interactions were shown between IL23R_rs10889677 and IBD5_rs17622208, which were not statistically significant^[2]. However, when using a logistic regression approach between these two markers, the IBD5_rs17622208 risk was only significant in the presence of the IL23R_rs10889677 risk allele^[2]. When SNPs were considered three at a time, the model suggested statistically significant interactions between IL23R_rs10889677, IBD5_rs11739135, and ATG16L1_rs2241880^[2]. When SNPs were considered four at a time, the model suggested interaction between IL23R_rs2201841, IL23R_rs7517847, and IBD5_rs11739135^[2]. A study in Oxford, UK has shown that certain IL23R polymorphisms have an association with CD only when the person is positive for IBD5^[2]. In conclusion, the logistic regression model did not show any significant evidence of gene-gene interaction, due to the small size of the study, even though there was a consistently small association between IBD5 and IL23R.

Childhood- vs adult-onset CD

The pathogenesis of pediatric and adult IBD differ.

The age of childhood and adult onset varies between cultures to culture because different cultures base it on different aspects such as physical, mental, or puberty. North American studies choose an age cut-off of 18 years, and < 18 years is considered a child, even though an individual at this age is physically mature. This is why Canadian studies have chosen to describe a child as < 16 years old. The belief is that the lower the age cut-off, the better the results are when comparing pediatric and adult onset of disease. Recent studies have shown that a subgroup of patients with early-onset IBD may have specific phenotypes that differ from adult-onset IBD^[39]. Many believe that pediatric-onset IBD is influenced by genetics compared to adult onset because there is less time for exposure to environmental modifiers to influence the onset of disease. Adult-onset is probably due to a mixture of genetics and abundant environmental exposure^[25]. For example, smoking is a major variable in adult IBD patients, but has little influence on pediatric IBD cohorts. Currently, there are conflicting studies on whether NOD2/CARD15 polymorphisms are associated with the age of onset of IBD because some show an association towards a younger age, while others show no effect^[40].

A study by Gazouli *et al.*^[25] has investigated the main polymorphisms and their association with childhood-onset of CD in a Greek cohort. While investigating the genotype and allele frequencies of the NOD2/CARD15 polymorphisms, rs2066844, rs2066845 and 3020insC, a statistically significant association between rs2066844 and adult-onset CD was observed. In both pediatric- and adult-onset CD, individuals with at least one NOD2/CARD15 polymorphism showed a genotype-phenotype correlation with ileal involvement^[25]. The study also confirmed the recently described association between IL23R variants in both child- and adult-onset CD^[35]. There has been conflicting evidence in studies regarding the ATG16L1 SNPs. One study has shown no association with early-onset or adult-onset CD^[25]. Recent research has indicated that the ATG16L1 rs2144880 variant is associated with adult-pediatric-onset CD, whereas other studies have demonstrated an association with diagnosis at an earlier age^[25]. In conclusion, the 3020insC variant in the NOD2/CARD15 gene is associated with CD and occurs considerably more often in childhood- than in adult-onset patients with CD^[25].

There is growing evidence that pediatric-onset IBD shows distinct differences when compared to its adult counterpart. Familial aggregation studies have shown an age-adjusted risk of developing IBD in first-degree relatives of affected individuals compared to the general population^[23]. The risk increases to > 30% for children when both parents are affected with IBD, suggesting that family history is the strongest risk factor^[5]. This is especially true among CD patients. Familial cases of CD occur at a younger age with greater severity than random sporadic cases^[36]. One main difference observed between pediatric- and adult-onset is that early onset shows a distinct and

more aggressive phenotype, such as the need for surgery, than similar IBD in individuals > 20 years old^[41].

Although NOD2/CARD15 mutations are neither sufficient nor necessary for the development of IBD, they are associated with a younger age of onset, presence of ileal involvement, and the development of strictures^[23]. There is also a gene dosage effect for CD location and complications. For example, stricture complications occur more frequently in CD children with the 1007fs mutation in the NOD2/CARD15 gene compared to children without this variant due to early surgery^[42]. Therefore, children with this mutation have a sixfold increased risk for developing a stricture complication^[23].

There has been conflicting evidence among studies that have attempted to show a link between the IBD5 locus and early-onset CD. A study by Rioux has found that the IBD5 locus is associated with early-onset CD where children were defined with an age of onset of < 16 years old^[23]. However, studies from pediatric-onset CD cohorts have demonstrated that the risk of IBD5 is lower compared to that in adult-onset CD, while others have demonstrated enhanced risk of developing CD when an individual has both SLC22A4-A5 and NOD2/CARD15 mutations^[43]. This connection may be due to a common pathophysiological mechanism^[23].

Although there is no sex difference among patients with adult-onset IBD, there is a clear and distinct difference among children with CD. Many studies from pediatric CD cohorts in the United States, Canada and the United Kingdom have shown an increase in male incidence. The higher male to female ratio continues to be unexplained. These differences are not observed among UC pediatric patients^[23]. It seems that sex is an age-dependent variable that has more influence on children than adults with IBD.

There are also phenotypic differences, such as disease location, among children and adult CD. For example, increased rates of upper GI tract disease and pure colonic disease in pediatric-onset CD have been identified. This difference may be due to the amount of examination during the onset of disease. Children undergo extensive GI endoscopy, whereas adults do not^[23]. These findings may be artificial or represent a true disease distinction among children and adults. Another distinction is the occurrence of colon-predominant disease during childhood-onset IBD under the age of 10 years old. With children < 5 years old, all have colon-only disease^[44]. During a study of approximately 1400 North American early-onset patients, data showed a colon-predominant phenotype in children < 8 years old. In another study in Europe, the acquisition of ileal CD became increasingly common as an individual approached 16 years old^[23]. These data confirm an association of colon-predominant phenotype in early diagnosed children that changes as they grow older.

Population studies with NOD2, IL23R and ATG16L1 polymorphisms

NOD2, IL23R and ATG16L1 polymorphisms were stud-

ied in a Lithuanian cohort with IBD. The study included 57 unrelated patients with CD, 123 with UC and 186 healthy individuals as controls. The three NOD2 variants, the IL23R variant rs11209026, and the ATG16L1 variant Thr300Ala, were genotyped among the population sample. No individuals were carriers of all three NOD2 risk alleles, whereas two CD patients were compound heterozygotes^[10]. Carriers of at least one NOD2 variant were highest among CD patients^[10]. There were no significant differences observed between UC patients and the controls. The NOD2 variant, Leu1007insC, was significantly associated with increased susceptibility in the Lithuanian CD population^[10]. In comparison to the other two NOD2 variants, the frequencies were very low and not significant in controls and among the IBD patients^[10]. In contrast to other European studies, a positive association between rs2066844, rs2066845 and CD was not found^[10]. When they further tried to analyze IL23R and ATG16L1, they were unable to replicate previous findings of increased susceptibility to IBD within the Lithuanian population^[10]. They were was a trend for a possible association with the ATG16L1 risk allele^[10]. It is of particular interest to study this population because Baltic countries have low IBD incidence rates, especially for CD. To confirm distinct IBD subtypes, a study using a larger North-Eastern European IBD sample needs to be investigated.

Another population study was performed among New Zealand Caucasians with IBD. Their cohort included 466 UC patients, 496 CD patients and 591 controls. All individuals were genotyped for the IL23R rs11209026 SNP, the ATG16L1 rs2241880 SNP, and the CARD15 variants. Significant interaction was detected between variants in ATG16L1, IL23R and CARD15 and CD susceptibility, whereas no significant association was observed between IL23R or ATG16L1 genotypes and IBD sub-phenotypes^[37]. The strongest association occurred between the ATG16L1 rs2241880 variant and CD, with no association detected with UC^[37]. ATG16L1 is suggested to have a CD-specific susceptibility locus. Unlike ATG16L1, the IL23R rs11209026 variant was strongly associated with both CD and UC. In their patient cohort, there was no evidence that IL23R or ATG16L1 genotypes influenced disease behavior, age of onset, location, or the need for surgical bowel resection^[37]. On the other hand, CARD15 was consistently a susceptibility factor and predictor of CD phenotype. Similar to many other studies, all three CARD15 SNPs are significantly overrepresented in patients with IBD family history, early onset of disease, ileal disease involvement, and development of complications^[33].

Interactions between NOD2 and IL23R variants with toll-like receptor-9 polymorphisms

Toll-like receptors (TLRs) are single, membrane-spanning proteins that play a key role in the innate immune system. These receptors recognize microbes that have structurally conserved molecules that breach the physical

Table 1 Key gene polymorphisms and their significance in Crohn's disease

Gene	Polymorphism	Relationship significance	Ref.
ATG16L	rs2241880 Thr300Ala	Associated with ileal form of CD with or without colonic involvement Highly associated with CD	[4,12]
NOD2/CARD15	rs2066844, rs2066845, 3020insC, 1007fs	Independently associated with CD	[20]
IBD5	SLC22A4 (OCTN1), SLC22A5 (OCTN2)	Independently associated with CD	[12]
CTLA4	rs3087243, rs11571302, rs11571297, rs7565213	Associated with IBD; no crude association to CD	[35]
TNFSF15	80 000 SNPs tested, including 7 SNPs within a 280 kb region on chromosome 9q32	Strongly associated with CD for Japanese and Jewish cohorts, but not for Europeans	[4,15,16]
IL23R	rs11209026 rs1004819	Strongly associated with conferring protection against CD Highly associated with CD	[4,33]

SNPs: Single-nucleotide polymorphism; CD: Crohn's disease.

barriers. Once bound, TLRs activate the immune system. The responsiveness of the GI tract to luminal bacteria is dependent on the interaction of transmembrane TLRs and the intracellular NOD2 receptor^[18]. Specifically, TLR9 plays a role in the maintenance of intestinal inflammation in IBD. TLR9 is also responsible for stimulating NOD2, which in turn enhances innate immune responses. Patients with two NOD2 mutations lose the synergistic effect between NOD2 and TLR9 stimulation. Therefore, interactions of both receptors have implications for intestinal homeostasis and inflammation^[18]. The TLR9 gene is located on chromosome 3p21.3, which is close to other CD susceptible loci^[9]. There are four SNPs in TLR9, but two of them are sufficient to distinguish between the haplotypes, which are rs5743836 and rs352140.

There might be a synergistic effect of NOD2 and TLR9 stimulation, therefore, Torok and colleagues have tested for gene interactions between TLR9 and CD-associated variants of NOD2^[18]. Significant associations between the two were observed that were specific to CD^[18]. The controls and UC showed no difference in distribution of TLR9 polymorphisms and NOD2 variants^[18]. Other CD variants in IL23R, ATG16L1 and IBD5 were analyzed for epistatic interactions. Aside from NOD2, the most significant association was found in the IL23R variant rs1004819, with ATG16L1 showing weaker associations with CD. There was no significant association between TLR9 polymorphisms with CD or UC phenotypes^[18]. Along with previous studies, they also showed that NOD2 mutations were associated with younger age of diagnosis of CD, ileal disease, and need for surgery^[18]. When there were two NOD2 mutations, there was a higher frequency of penetrating disease^[18].

In conclusion, a new association between CD and a TLR9 polymorphism has been found. There is evidence that when CD patients carry CD-associated NOD2 variants, they have an increased incidence of TLR9 polymorphisms. This is not surprising because there is a synergistic effect of NOD2 and TLR9 stimulation and it is important for the maintenance of intestinal homeostasis and inflammation. TLR9 also demonstrates significant epistatic interactions with IL23R variants, but unlike NOD2, there is no association with the frequency

in TLR9 polymorphisms present. This study shows the first evidence for interaction between polymorphisms in TLR9 and variants between NOD2 and IL23R^[18].

CONCLUSION

CD is an autoimmune disease characteristic of chronic intestinal inflammation and lesions. It can affect people of all ages and ethnicities worldwide. Recent genome-wide studies have shown significant genetic associations of several variants and susceptibility to CD. This is true for North American, South American and European populations. Certain variants have been linked only to Asian and African cohorts. It is vital to understand the pathology of CD and the underlying genetic interactions to increase efficiency of diagnosis and develop drugs that target specific immune system pathways. There have been several genetic variants highly associated with CD, which include ATG16L1, TNFSF15, NOD2/CARD15, IL23R and IBD5 (Table 1).

The process of autophagy is an important aspect of our immune system. It is a way to destroy foreign pathogens that enter the body. The Thr300Ala variant within the N terminus of ATG16L, which is part of a complex that forms autophagosomes, has significant associations with CD. Different ethnicities show different genotype markers. This is seen in TNFSF15, which is involved in systemic inflammation and regulation of various immune cells. Seven haplotypes have been identified within a Japanese cohort, but not in other ethnic cohorts. More research needs to be conducted to characterize fully this possible genetic link to CD. The most significant association with CD is observed among NOD2/CARD15 and IL23R variants. NOD2/CARD15 plays an essential role in maintaining the intestinal normal flora. There are many theories about the cause of CD, and one of them includes a shift in the intestinal bacterial flora. Therefore, it is no surprise that NOD2 variants are highly associated with CD. There are three independent variants associated with CD: rs2066844, rs2066845 and 1007fs. The other highly associated genetic link is observed within the IL23R gene. One variant that is highly associated with CD is rs1004819, whereas rs11209026 confers protection against CD. IL23R plays an essential role in

mediating proinflammatory activities. Another highly associated genetic link with CD is observed in two variants within IBD5. The two variants, SLC22A4 (OCTN1) and SLC22A5 (OCTN2), show significant association with CD, but no interactions between these variants and other CD-associated genetic variants have been observed. The last highly associated genetic link with CD is observed within the *CTLA4* gene. This gene plays a role in the immune system. The four polymorphisms associated with IBD are rs3087243, rs11571302, rs7565213 and rs11571297.

Many CD variants are associated with a specific CD phenotype, age of diagnosis, severity and location of CD, and surgical outcome. Gene-gene interactions have also been characterized among the different variants and other immune system receptors. We need to understand fully the pathogenesis of CD to target pathways for effective treatment. Further research is needed to explore all possible gene-gene interactions due to a gene dose affect associated with CD. It seems that new genetic associations are constantly being uncovered within IBD and then associated with either CD or UC. Overall, CD shows a greater genetic link than UC. Recent discoveries have led to therapies and treatments that show much promise. Researchers need to continue the search for genetic links and diseases because this may be the only way to understand the pathology of CD and develop effective treatments.

REFERENCES

- Brand S, Staudinger T, Schnitzler F, Pfennig S, Hofbauer K, Dambacher J, Seiderer J, Tillack C, Konrad A, Crispin A, Göke B, Lohse P, Ochsenkühn T. The role of Toll-like receptor 4 Asp299Gly and Thr399Ile polymorphisms and CARD15/NOD2 mutations in the susceptibility and phenotype of Crohn's disease. *Inflamm Bowel Dis* 2005; **11**: 645-652
- Cummings JR, Ahmad T, Geremia A, Beckly J, Cooney R, Hancock L, Pathan S, Guo C, Cardon LR, Jewell DP. Contribution of the novel inflammatory bowel disease gene IL23R to disease susceptibility and phenotype. *Inflamm Bowel Dis* 2007; **13**: 1063-1068
- Cho JH. Inflammatory bowel disease: genetic and epidemiologic considerations. *World J Gastroenterol* 2008; **14**: 338-347
- Libioule C, Louis E, Hansoul S, Sandor C, Farnir F, Franchimont D, Vermeire S, Dewit O, de Vos M, Dixon A, Demarche B, Gut I, Heath S, Foglio M, Liang L, Laukens D, Mni M, Zelenika D, Van Gossum A, Rutgeerts P, Belaiche J, Lathrop M, Georges M. Novel Crohn disease locus identified by genome-wide association maps to a gene desert on 5p13.1 and modulates expression of PTGER4. *PLoS Genet* 2007; **3**: e58
- Csöngéi V, Járomi L, Sáfrány E, Sipeky C, Magyari L, Faragó B, Bene J, Polgár N, Lakner L, Sarlós P, Varga M, Melegh B. Interaction of the major inflammatory bowel disease susceptibility alleles in Crohn's disease patients. *World J Gastroenterol* 2010; **16**: 176-183
- Beattie RM, Croft NM, Fell JM, Afzal NA, Heuschkel RB. Inflammatory bowel disease. *Arch Dis Child* 2006; **91**: 426-432
- Read S, Malmström V, Powrie F. Cytotoxic T lymphocyte-associated antigen 4 plays an essential role in the function of CD25(+)CD4(+) regulatory cells that control intestinal inflammation. *J Exp Med* 2000; **192**: 295-302
- Fujita N, Itoh T, Omori H, Fukuda M, Noda T, Yoshimori T. The Atg16L complex specifies the site of LC3 lipidation for membrane biogenesis in autophagy. *Mol Biol Cell* 2008; **19**: 2092-2100
- Tremelling M, Cummings F, Fisher SA, Mansfield J, Gwilliam R, Keniry A, Nimmo ER, Drummond H, Onnie CM, Prescott NJ, Sanderson J, Bredin F, Berzuini C, Forbes A, Lewis CM, Cardon L, Deloukas P, Jewell D, Mathew CG, Parkes M, Satsangi J. IL23R variation determines susceptibility but not disease phenotype in inflammatory bowel disease. *Gastroenterology* 2007; **132**: 1657-1664
- Sventoraityte J, Zvirbliene A, Franke A, Kwiatkowski R, Kiudelis G, Kupcinskas L, Schreiber S. NOD2, IL23R and ATG16L1 polymorphisms in Lithuanian patients with inflammatory bowel disease. *World J Gastroenterol* 2010; **16**: 359-364
- Rioux JD, Xavier RJ, Taylor KD, Silverberg MS, Goyette P, Huett A, Green T, Kuballa P, Barmada MM, Datta LW, Shugart YY, Griffiths AM, Targan SR, Ippoliti AF, Bernard EJ, Mei L, Nicolae DL, Regueiro M, Schumm LP, Steinhart AH, Rotter JI, Duerr RH, Cho JH, Daly MJ, Brant SR. Genome-wide association study identifies new susceptibility loci for Crohn disease and implicates autophagy in disease pathogenesis. *Nat Genet* 2007; **39**: 596-604
- Hradsky O, Dusatkova P, Lenicek M, Bronsky J, Nevala J, Vitek L, Lukas M, Zeniskova I, Cinek O. The CTLA4 variants may interact with the IL23R- and NOD2-conferred risk in development of Crohn's disease. *BMC Med Genet* 2010; **11**: 91
- Prescott NJ, Fisher SA, Franke A, Hampe J, Onnie CM, Soars D, Bagnall R, Mirza MM, Sanderson J, Forbes A, Mansfield JC, Lewis CM, Schreiber S, Mathew CG. A non-synonymous SNP in ATG16L1 predisposes to ileal Crohn's disease and is independent of CARD15 and IBD5. *Gastroenterology* 2007; **132**: 1665-1671
- Xia B, Crusius JB, Wu J, Zwiers A, van Bodegraven AA, Peña AS. CTLA4 gene polymorphisms in Dutch and Chinese patients with inflammatory bowel disease. *Scand J Gastroenterol* 2002; **37**: 1296-1300
- Yamazaki K, McGovern D, Ragoussis J, Paolucci M, Butler H, Jewell D, Cardon L, Takazoe M, Tanaka T, Ichimori T, Saito S, Sekine A, Iida A, Takahashi A, Tsunoda T, Lathrop M, Nakamura Y. Single nucleotide polymorphisms in TNFSF15 confer susceptibility to Crohn's disease. *Hum Mol Genet* 2005; **14**: 3499-3506
- Michelsen KS, Thomas LS, Taylor KD, Yu QT, Mei L, Landers CJ, Derkowski C, McGovern DP, Rotter JI, Targan SR. IBD-associated TL1A gene (TNFSF15) haplotypes determine increased expression of TL1A protein. *PLoS One* 2009; **4**: e4719
- Van Limbergen J, Russell RK, Nimmo ER, Ho GT, Arnott ID, Wilson DC, Satsangi J. Genetics of the innate immune response in inflammatory bowel disease. *Inflamm Bowel Dis* 2007; **13**: 338-355
- Török HP, Glas J, Endres I, Tonenchi L, Teshome MY, Wetzke M, Klein W, Lohse P, Ochsenkühn T, Folwaczny M, Göke B, Folwaczny C, Müller-Myhsok B, Brand S. Epistasis between Toll-like receptor-9 polymorphisms and variants in NOD2 and IL23R modulates susceptibility to Crohn's disease. *Am J Gastroenterol* 2009; **104**: 1723-1733
- Kobayashi KS, Chamaillard M, Ogura Y, Henegariu O, Inohara N, Nuñez G, Flavell RA. Nod2-dependent regulation of innate and adaptive immunity in the intestinal tract. *Science* 2005; **307**: 731-734
- Vermeire S, Wild G, Kocher K, Cousineau J, Dufresne L, Bitton A, Langelier D, Pare P, Lapointe G, Cohen A, Daly MJ, Rioux JD. CARD15 genetic variation in a Quebec population: prevalence, genotype-phenotype relationship, and haplotype structure. *Am J Hum Genet* 2002; **71**: 74-83
- Veldhoen M, Hocking RJ, Atkins CJ, Locksley RM, Stock-

- inger B. TGFbeta in the context of an inflammatory cytokine milieu supports de novo differentiation of IL-17-producing T cells. *Immunity* 2006; **24**: 179-189
- 22 **Traub S**, von Aulock S, Hartung T, Hermann C. MDP and other muropeptides--direct and synergistic effects on the immune system. *J Endotoxin Res* 2006; **12**: 69-85
 - 23 **Biank V**, Broeckel U, Kugathasan S. Pediatric inflammatory bowel disease: clinical and molecular genetics. *Inflamm Bowel Dis* 2007; **13**: 1430-1438
 - 24 **Baptista ML**, Amarante H, Picheth G, Sdepanian VL, Peterson N, Babasukumar U, Lima HC, Kugathasan S. CARD15 and IL23R influences Crohn's disease susceptibility but not disease phenotype in a Brazilian population. *Inflamm Bowel Dis* 2008; **14**: 674-679
 - 25 **Gazouli M**, Pachoula I, Panayotou I, Mantzaris G, Chrousos G, Anagnou NP, Roma-Giannikou E. NOD2/CARD15, ATG16L1 and IL23R gene polymorphisms and childhood-onset of Crohn's disease. *World J Gastroenterol* 2010; **16**: 1753-1758
 - 26 **Wehkamp J**, Salzman NH, Porter E, Nuding S, Weichenthal M, Petras RE, Shen B, Schaeffeler E, Schwab M, Linzmeier R, Feathers RW, Chu H, Lima H, Fellermann K, Ganz T, Stange EF, Bevins CL. Reduced Paneth cell alpha-defensins in ileal Crohn's disease. *Proc Natl Acad Sci USA* 2005; **102**: 18129-18134
 - 27 **Zhang Z**, Zheng M, Bindas J, Schwarzenberger P, Kolls JK. Critical role of IL-17 receptor signaling in acute TNBS-induced colitis. *Inflamm Bowel Dis* 2006; **12**: 382-388
 - 28 **Glas J**, Seiderer J, Wetzke M, Konrad A, Török HP, Schmechel S, Tonenchi L, Grassl C, Dambacher J, Pfennig S, Maier K, Griga T, Klein W, Eppel JT, Schiemann U, Folwaczny C, Lohse P, Göke B, Ochsenkühn T, Müller-Myhsok B, Folwaczny M, Mussack T, Brand S. rs1004819 is the main disease-associated IL23R variant in German Crohn's disease patients: combined analysis of IL23R, CARD15, and OCTN1/2 variants. *PLoS One* 2007; **2**: e819
 - 29 **Tack J**, Carethers JM. This Month in Gastroenterology. *Gastroenterology* 2007; **132**: 1641-1643
 - 30 **Oppmann B**, Lesley R, Blom B, Timans JC, Xu Y, Hunte B, Vega F, Yu N, Wang J, Singh K, Zonin F, Vaisberg E, Churakova T, Liu M, Gorman D, Wagner J, Zurawski S, Liu Y, Abrams JS, Moore KW, Rennick D, de Waal-Malefyt R, Hannum C, Bazan JF, Kastelein RA. Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. *Immunity* 2000; **13**: 715-725
 - 31 **Schmidt C**, Giese T, Ludwig B, Mueller-Molaian I, Marth T, Zeuzem S, Meuer SC, Stallmach A. Expression of interleukin-12-related cytokine transcripts in inflammatory bowel disease: elevated interleukin-23p19 and interleukin-27p28 in Crohn's disease but not in ulcerative colitis. *Inflamm Bowel Dis* 2005; **11**: 16-23
 - 32 **Neurath MF**. IL-23: a master regulator in Crohn disease. *Nat Med* 2007; **13**: 26-28
 - 33 **Schmechel S**, Konrad A, Diegelmann J, Glas J, Wetzke M, Paschos E, Lohse P, Göke B, Brand S. Linking genetic susceptibility to Crohn's disease with Th17 cell function: IL-22 serum levels are increased in Crohn's disease and correlate with disease activity and IL23R genotype status. *Inflamm Bowel Dis* 2008; **14**: 204-212
 - 34 **Oliver J**, Rueda B, López-Nevot MA, Gómez-García M, Martín J. Replication of an association between IL23R gene polymorphism with inflammatory bowel disease. *Clin Gastroenterol Hepatol* 2007; **5**: 977-981, 981.e1-2
 - 35 **Van Limbergen J**, Russell RK, Nimmo ER, Drummond HE, Smith L, Anderson NH, Davies G, Gillett PM, McGrogan P, Weaver LT, Bisset WM, Mahdi G, Arnott ID, Wilson DC, Satsangi J. Autophagy gene ATG16L1 influences susceptibility and disease location but not childhood-onset in Crohn's disease in Northern Europe. *Inflamm Bowel Dis* 2008; **14**: 338-346
 - 36 **Lappalainen M**, Halme L, Turunen U, Saavalainen P, Einarsdottir E, Färkkilä M, Kontula K, Paavola-Sakki P. Association of IL23R, TNFRSF1A, and HLA-DRB1*0103 allele variants with inflammatory bowel disease phenotypes in the Finnish population. *Inflamm Bowel Dis* 2008; **14**: 1118-1124
 - 37 **Roberts RL**, Geary RB, Hollis-Moffatt JE, Miller AL, Reid J, Abkevich V, Timms KM, Gutin A, Lanchbury JS, Merriman TR, Barclay ML, Kennedy MA. IL23R R381Q and ATG16L1 T300A are strongly associated with Crohn's disease in a study of New Zealand Caucasians with inflammatory bowel disease. *Am J Gastroenterol* 2007; **102**: 2754-2761
 - 38 **Cummings JR**, Cooney R, Pathan S, Anderson CA, Barrett JC, Beckly J, Geremia A, Hancock L, Guo C, Ahmad T, Cardon LR, Jewell DP. Confirmation of the role of ATG16L1 as a Crohn's disease susceptibility gene. *Inflamm Bowel Dis* 2007; **13**: 941-946
 - 39 **Mamula P**, Telega GW, Markowitz JE, Brown KA, Russo PA, Piccoli DA, Baldassano RN. Inflammatory bowel disease in children 5 years of age and younger. *Am J Gastroenterol* 2002; **97**: 2005-2010
 - 40 **Russell RK**, Drummond HE, Nimmo EE, Anderson N, Smith L, Wilson DC, Gillett PM, McGrogan P, Hassan K, Weaver LT, Bisset M, Mahdi G, Satsangi J. Genotype-phenotype analysis in childhood-onset Crohn's disease: NOD2/CARD15 variants consistently predict phenotypic characteristics of severe disease. *Inflamm Bowel Dis* 2005; **11**: 955-964
 - 41 **Polito JM**, Childs B, Mellits ED, Tokayer AZ, Harris ML, Bayless TM. Crohn's disease: influence of age at diagnosis on site and clinical type of disease. *Gastroenterology* 1996; **111**: 580-586
 - 42 **Kugathasan S**, Collins N, Maresso K, Hoffmann RG, Stephens M, Werlin SL, Rudolph C, Broeckel U. CARD15 gene mutations and risk for early surgery in pediatric-onset Crohn's disease. *Clin Gastroenterol Hepatol* 2004; **2**: 1003-1009
 - 43 **Newman B**, Gu X, Wintle R, Cescon D, Yazdanpanah M, Liu X, Peltekova V, Van Oene M, Amos CI, Siminovitch KA. A risk haplotype in the Solute Carrier Family 22A4/22A5 gene cluster influences phenotypic expression of Crohn's disease. *Gastroenterology* 2005; **128**: 260-269
 - 44 **Heyman MB**, Kirschner BS, Gold BD, Ferry G, Baldassano R, Cohen SA, Winter HS, Fain P, King C, Smith T, El-Serag HB. Children with early-onset inflammatory bowel disease (IBD): analysis of a pediatric IBD consortium registry. *J Pediatr* 2005; **146**: 35-40

S- Editor Tian L L- Editor Kerr C E- Editor Li JY

Effect of *Helicobacter pylori* *cdrA* on interleukin-8 secretions and nuclear factor kappa B activation

Hiroaki Takeuchi, Ya-Nan Zhang, Dawn A Israel, Richard M Peek Jr, Mikio Kamioka, Hideo Yanai, Norihito Morimoto, Tetsuro Sugiura

Hiroaki Takeuchi, Mikio Kamioka, Norihito Morimoto, Tetsuro Sugiura, Department of Clinical Laboratory Medicine, Kochi Medical School, Kohasu, Oko-cho, Nankoku-city, Kochi 783-8505, Japan

Ya-Nan Zhang, Laboratory Department, Beijing Tiantan Hospital Affiliated of Capital Medical University, Beijing 100050, China

Dawn A Israel, Richard M Peek Jr, Division of Gastroenterology and Cancer Prevention, Department of Medicine, Vanderbilt University Medical Center, Nashville, TN 37232, United States
Hideo Yanai, Department of Internal Medicine, Kanmon Medical Center, 1-1-1 Ushiroda-cho, Shimonoseki, Yamaguchi 751-8501, Japan

Author contributions: Takeuchi H designed and performed the research, and wrote the paper; Zhang YN and Kamioka M performed the research; Peek Jr RM, Yanai H, Morimoto N and Sugiura T contributed new reagents/analytic tools; Israel DA revised the paper.

Supported by The Project Research Fund from Kochi University, to Takeuchi H; and a Grant-in-Aid for Scientific Research from the Ministry of Education, Science and Culture of Japan, No. 21590631 and 21590629, in part

Correspondence to: Dr. Hiroaki Takeuchi, Department of Clinical Laboratory Medicine, Kochi Medical School, Kohasu, Oko-cho, Nankoku-city, Kochi 783-8505,

Japan. htake@kochi-u.ac.jp

Telephone: +81-88-8802427 Fax: +81-88-8802428

Received: April 15, 2011 Revised: May 19, 2011

Accepted: May 26, 2011

Published online: February 7, 2012

Abstract

AIM: To investigate genetic diversity of *Helicobacter pylori* (*H. pylori*) cell division-related gene A (*cdrA*) and its effect on the host response.

METHODS: Inactivation of *H. pylori* *cdrA*, which is involved in cell division and morphological elongation, has a role in chronic persistent infections. Genetic property of *H. pylori* *cdrA* was evaluated using

polymerase chain reaction and sequencing in 128 (77 American and 51 Japanese) clinical isolates obtained from 48 and 51 patients, respectively. Enzyme-linked immunosorbent assay was performed to measure interleukin-8 (IL-8) secretion with gastric biopsy specimens obtained from American patients colonized with *cdrA*-positive or -negative strains and AGS cells co-cultured with wild-type HPK5 (*cdrA*-positive) or its derivative HPKT510 (*cdrA*-disruptant). Furthermore, the cytotoxin-associated gene A (*cagA*) status (translocation and phosphorylation) and kinetics of transcription factors [nuclear factor-kappa B (NF- κ B) and inhibition kappa B] were investigated in AGS cells co-cultured with HPK5, HPKT510 and its derivative HPK5CA (*cagA*-disruptant) by western blotting analysis with immunoprecipitation.

RESULTS: Genetic diversity of the *H. pylori* *cdrA* gene demonstrated that the *cdrA* status segregated into two categories including four allele types, *cdrA*-positive (allele types; I and II) and *cdrA*-negative (allele types; III and IV) categories, respectively. Almost all Japanese isolates were *cdrA*-positive (I: 7.8% and II: 90.2%), whereas 16.9% of American isolates were *cdrA*-positive (II) and 83.1% were *cdrA*-negative (III: 37.7% and IV: 45.5%), indicating extended diversity of *cdrA* in individual American isolates. Comparison of each isolate from different regions (antrum and corpus) in the stomach of 29 Americans revealed that *cdrA* status was identical in both isolates from different regions in 17 cases. However, 12 cases had a different *cdrA* allele and 6 of them exhibited a different *cdrA* category between two regions in the stomach. Furthermore, in 5 of the 6 cases possessing a different *cdrA* category, *cdrA*-negative isolate existed in the corpus, suggesting that *cdrA*-negative strain is more adaptable to colonization in the corpus. IL-8 secretions from AGS revealed that IL-8 levels induced by a *cdrA*-disrupted HPKT510 was significantly lower ($P < 0.01$) compared to wild-type HPK5: corresponding to 50%-60% of those of

wild-type HPK5. These data coincided with *in vivo* data that an average value of IL-8 in biopsy specimens from *cdrA*-positive and *cdrA*-negative groups was 215.6 and 135.9 pg/mL, respectively. Western blotting analysis documented that HPKT510 had no effect on CagA translocation and phosphorylation, however, nuclear accumulation of NF- κ B was lower by HPKT510 compared to HPK5.

CONCLUSION: Colonization by a *cdrA*-negative or *cdrA*-dysfunctional strain resulted in decreased IL-8 production and repression of NF- κ B, and hence, attenuate the host immunity leading to persistent infection.

© 2012 Baishideng. All rights reserved.

Key words: *Helicobacter pylori* cell division-related gene A; Genetic diversity; Host immune response; Interleukin-8 secretion; Nuclear factor kappa B; Persistent infection

Peer reviewers: Marcelo Lima Ribeiro, PhD, Clinical Pharmacology and Gastroenterology Unit, Laboratory of Microbiology and Molecular Biology, Sao Francisco University Medical School, Av Sao Francisco de Assis, 218, Braganca Paulista-SP 12916-900, Brazil; Noriko Nakajima, MD, PhD, Associate Professor, Division of Gastroenterology and Hepatology, Department of Internal Medicine, Nihon University School of Medicine, 1-8-13 Kandasurugadai Chiyoda-ku, Tokyo 101-8309, Japan; Tamara Vorobjova, MD, PhD, Senior Researcher in Immunology, Department of Immunology, Institute of General and Molecular Pathology, University of Tartu, Ravila, 19, Tartu 50411, Estonia

Takeuchi H, Zhang YN, Israel DA, Peek Jr RM, Kamioka M, Yanai H, Morimoto N, Sugiura T. Effect of *Helicobacter pylori cdrA* on interleukin-8 secretions and nuclear factor kappa B activation. *World J Gastroenterol* 2012; 18(5): 425-434 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/425.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.425>

INTRODUCTION

Helicobacter pylori (*H. pylori*), a Gram-negative spiral bacterium, colonizes the human stomach and causes chronic inflammation, which may progress to peptic ulceration, atrophic gastritis and gastric cancer^[1]. Although the *H. pylori* population structure appears to be clonal over short periods of time, isolates obtained from different individuals exhibit substantial genetic diversity, consistent with extensive recombination and a panmictic population structure^[2-7]. Putative mechanisms for the generation of diversity within *H. pylori* include frequent horizontal genetic exchange among strains and a high level of spontaneous mutation occurring over a long evolutionary time period within a highly restricted niche^[6,8].

Infection by cytotoxin-associated gene A (*cagA*)-positive *H. pylori* is a known risk factor for the development of gastroduodenal disease due to major changes in cellular morphology and the release of molecules, including

cytokines, from the gastric epithelium. *H. pylori* infection up-regulates secretion of various inflammatory cytokines, including interleukin (IL)-1, IL-6, IL-8 and tumor necrosis factor α , and contributes to the pronounced inflammatory response^[9]. IL-8 production induced by *H. pylori in vitro* and *in vivo* is recognized as a host response to microbes^[10,11], but the role of CagA on IL-8 production is less clear^[12]. IL-8 is a potent pro-inflammatory cytokine which regulates neutrophil infiltration of gastric mucosa in *H. pylori* gastritis and is released through the signaling pathway concerning nuclear factor-kappa B (NF- κ B), which includes extracellular signal-regulated kinase (ERK) activity^[12,13] and mitogen-activated protein kinase^[14]. Among the strongest transcriptional regulators of *H. pylori*-induced cytokine expression is the NF- κ B family of transcription factors^[15]. Activation of NF- κ B can affect the expression of several hundred genes, and activation of its signal transduction pathway occurs in response to a wide range of stimuli and results in nuclear accumulation of NF- κ B transcription factors, causing changes in the expression of target genes involved in innate and adaptive immunity, and inflammation.

We described previously that the *H. pylori* cell division-related gene A (*cdrA*) not only has a repressive role on cell division but is also involved in cell elongation and cell death *via* cell wall synthesis at the division site^[16,17]. The *cdrA*-disrupted mutant HPKT510 was able to survive for the long-term in liquid medium, even under serum-free and aerobic conditions, and was more resistant to bactericidal of beta-lactam antibiotics than the wild-type HPK5^[16]. Inactivation of *cdrA* during infection in the stomach may have contributed to ensuring persistent infection by altering its ability to adapt to the microenvironment. In fact, the *cdrA* gene was found to be absent in 3 out of 4 colonies recovered from a mouse infected with *H. pylori* strain B128^[18]. Furthermore, additional isolates of the sequenced *H. pylori* strain J99 from a patient after a 6-year interval were subjected to microarray analysis, which indicated that the *cdrA* gene was missing in additional isolates^[19]. Thus, we hypothesize that a loss of *cdrA* in *H. pylori* during infection might be an evolutionary event to alter its biological characteristics, which affects the host immune response to the microbes and promotes persistent infection.

In this study, the level of IL-8 secretion induced by a *cdrA*-negative strain was approximately 50% lower than those induced by a *cdrA*-positive strain *in vitro* and *in vivo*. Genetic diversity of *cdrA* was extended in American isolates and *cdrA*-negative isolates might be more adaptable to colonize in the corpus. Western blotting analysis documented that the CagA status at the point of its translocation and phosphorylation was not different between HPK5 and HPKT510 strains. However, expression of NF- κ B was lower in HPKT510 than that in HPK5, indicating that *H. pylori* with inactivation of *cdrA* might escape from rigorous immune clearance and facilitate chronic persistent infection caused by decreased levels of IL-8 and nuclear accumulation of NF- κ B.

MATERIALS AND METHODS

Bacterial strains and growth conditions

Wild-type *H. pylori* HPK5 and its derivatives such as HPKT510, a *cdrA*-disrupted mutant carrying *xyIE-Km* cassette^[17], and HPK5CA, a *cagA*-disrupted mutant carrying *xyIE-Km* cassette^[20], were cultivated in Brucella broth (Becton Dickinson, United States) supplemented with 10% horse serum at 37 °C in an atmosphere containing 10% CO₂ as previously described^[21]. Bacteria grown in 10 mL of Brucella-serum medium in 100-mL conical flasks following sub-culture were subjected to co-culture with AGS cells. Bacterial growth was measured by determining absorbance at 600 nm (*A*₆₀₀) with a spectrophotometer (GENEQUANT pro, Amersham Pharmacia Biotech), and colony forming units were determined for bacterial viability. Additional clinical isolates, 77 American and 51 Japanese isolates endoscopically obtained from 48 and 51 patients, respectively, were cultured on Brucella-serum agar at 37 °C in an atmosphere containing 10% CO₂. Isolates from different regions in the stomach, such as the antrum and corpus, were also included in 29 of 77 American isolates. The bacterial genomic DNA extracted by a DNA kit (Qiagen, Tokyo, Japan) according to the manufacturer's instructions were subjected to polymerase chain reaction (PCR) with specific primer pairs to examine *cdrA* gene diversity.

AGS cell culture conditions

AGS cells (human gastric adenocarcinoma epithelial cell line, CRL1739c) were grown in RPMI 1640 + 7 medium including streptomycin (20 µg/mL) and kanamycin (60 µg/mL) (Nikken bio medical Laboratory, Japan) supplemented with 10% (v/v) heat-inactivated fetal bovine serum (Biowest, France) at 37 °C in an atmosphere containing 5% CO₂. AGS cells were seeded at a density of 0.5 × 10⁶ cells in six-well plates (SUMITOMO BAKELIFE, Japan) and grown to about 80% confluence prior to co-culture experiments.

Co-culture of AGS cells with *H. pylori* strains

Subconfluent AGS cells (1 × 10⁶) cultured in RPMI 1640 + 7 medium supplemented with 10% serum were washed twice by phosphate-buffered saline (PBS) and subsequently cultured with or without each *H. pylori* strain (HPK5, HPKT510 and HPK5CA) at a multiplicity of infection (MOI) of 150 or 300 in 1.5 mL of RPMI 1640 medium (Gibco BRL, Eggenstein, Germany) alone for 48 h at 37 °C in an atmosphere containing 10% CO₂. The supernatant was collected at various times (3–48 h), centrifuged at 7000 r/min for 5 min to pellet bacteria and AGS cells, and subjected to sandwich enzyme-linked immunosorbent assay (ELISA) to measure IL-8 productions. AGS cells after co-culture were collected at the appropriate time and used in the following analyses.

Preparation of whole-cell and nuclear extracts

AGS cells were washed five times in ice-cold PBS, incu-

bated with lysis buffer containing 50 mmol/L NaCl, 1 mmol/L EDTA, 1 mmol/L NaF, 100 µmol/L sodium orthovanadate, 10 µmol/L phenylmethylsulfonyl fluoride (PMSF) and a commercially available protease inhibitor mixture tablet (Complete, Roche Molecular biochemical, Indianapolis, IN), harvested by scraping and transferred to a microcentrifuge tube. Debris was removed by centrifugation at 10 000 *g* for 10 min (TOMY MRX-150) to collect the total proteins for whole-cell extracts. Nuclear proteins were prepared by Cellytic Nu-CLEAR Extraction Kit (SIGMA, Japan) according to a previous report^[22]. Briefly, AGS cells were incubated with lysis buffer (10 mmol/L HEPES, pH 7.9, 1.5 mmol/L MgCl₂, 10 mmol/L KCl, 1 mmol/L DTT and 0.25 mmol/L PMSF) on ice for 15 min. The pellet precipitated containing nuclei following centrifugation for 30 s at 10 000 *g* was resuspended in 100 µL extraction buffer (20 mmol/L HEPES, pH 7.9, 1.5 mmol/L MgCl₂, 0.42 mol/L NaCl, 0.2 mmol/L EDTA, 25% glycerol, 1 mmol/L DTT and 0.25 mmol/L PMSF) for 30 min at 4 °C with agitation. After centrifuging for 5 min at 20 000 *g*, the supernatant (nuclear fraction) was collected for nuclear extracts.

Immunoprecipitation

One milligram of the lysate proteins was incubated with polyclonal rabbit anti-*H. pylori* CagA antibody (Austral Biologicals, CA, United States) for 1 h at 4 °C, followed by an overnight incubation with a 20 µL aliquot of Protein G Plus-agarose beads (Santa Cruz, United States) at 4 °C, as previously described^[20]. Briefly, the beads were washed with lysis buffer and boiled for 10 min in 2 × electrophoresis sample buffer (50 mmol/L Tris (pH 6.8), 10% sodium dodecyl sulfate, 12% 2-mercaptoethanol, 20% (wt/vol) glycine and 1% (v/v) bromophenol blue) to elute the immunoprecipitated proteins.

Western blotting analysis

For detection of CagA and its phosphorylation, equivalent amounts of the immunoprecipitated proteins (100 µg) were resolved by 5% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). Proteins were electrotransferred onto nitrocellulose membranes (Millipore Corporation, United States) by a semidry blotting apparatus KS-8460 (System Instruments Co., Ltd. Tokyo). The blots were blocked overnight with 4% (wt/vol) dried skim milk in Tris-buffered saline with Tween 20 at room temperature and then incubated with monoclonal antibody PY99 (diluted 1/1000; Santa Cruz, United States) for 1 h and anti-mouse IgG peroxidase-linked species-specific whole secondary antibody (diluted 1/1000; GE Healthcare Biosciences, Co. Ltd., United Kingdom) for 1 h to detect phosphorylated tyrosine protein. Immunodetection was performed by enhanced chemiluminescence Plus Western Blotting Detection Reagents (GE Healthcare Biosciences). Next, blots were stripped, reprobbed with a specific polyclonal rabbit anti-*H. pylori* CagA antibody (diluted 1/1000; Austral Biologicals, CA, United States) and incubated with a horseradish perox-

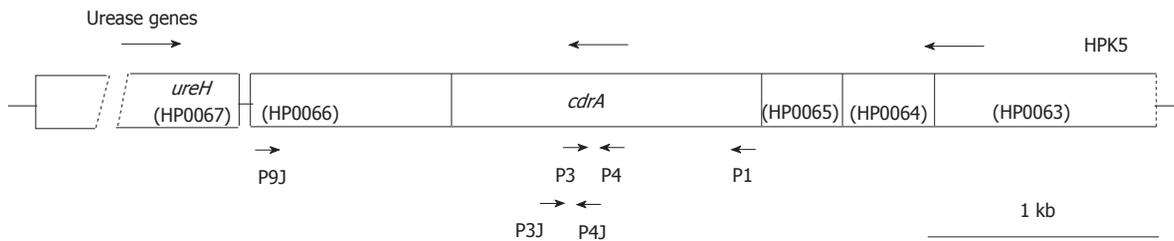


Figure 1 Location of primers used for cell division-related gene A in the region downstream of the urease gene cluster of *Helicobacter pylori* HPK5. Arrows above and below the map depict the direction of transcription and primers, respectively. The open reading frames (HP0063 to HP0067) are shown based on strain 26695.

idase-conjugated goat anti-rabbit secondary antibody (diluted 1/10 000; Jackson ImmunoResearch, PA, United States)^[20]. Each 10 µg of nuclear and total proteins was subjected to 12% SDS-PAGE for detection of NF-κB and inhibition kappa B (IκB), respectively. Procedures of western blot analysis were followed as described above. Rabbit anti-human-NF-κB (p65) (diluted 1/200; Cell Signaling Technology, United States) and rabbit anti-human-IκB antibodies (diluted 1/200; Santa Cruz Biotechnology Inc, United States), and an anti-rabbit IgG antibody-HRP (diluted 1/2000; Santa Cruz Biotechnology Inc, United States) were used as primary and secondary antibodies, respectively. For a standard control, a rabbit anti-human-ERK2 antibody (diluted 1/200; Santa Cruz Biotechnology Inc, United States) was used to detect unphosphorylated ERK2 to confirm equal protein load^[23]. The analyses were performed for least three independent experiments.

Measurement of IL-8 secretion from AGS cells

The amount of IL-8 secreted into culture medium after co-culture with *H. pylori* strains was determined by ELISA using the CytoSets system (BioSource International) according to the manufacturer’s instructions. Fifty µL aliquots of the supernatant was briefly centrifuged at 7000 r/min for 10 min to remove bacteria and AGS cells, after which supernatants were added to an equal volume of reagent on the 96-well ELISA plate. The absorbance of samples was measured at 550 nm using a 96-well microplate reader (Multiskan JX ver1.1, Thermo Labsystems, Finland), and the data was expressed as pg/mL. All samples were measured in triplicate in at least three independent experiments.

Measurement of mucosal IL-8 secretion

Frozen gastric antral biopsy specimens from 25 clinical patients infected with *H. pylori* were homogenized in 1 mL of PBS, and supernatants obtained by centrifugation were used for determination of IL-8 proteins by ELISA, as described previously^[24]. Cytokine concentrations in homogenates were normalized in terms of total protein concentration of the biopsy specimen and were expressed as pg/mL.

PCR for cdrA gene diversity

To compare the *in vivo* IL-8 level in the biopsy speci-

mens between the patients infected with *cdrA*-positive or -negative strains, PCR was utilized to examine the *cdrA* status in clinical isolates. The genomic DNA of clinical isolates, including 77 American and 51 Japanese strains, were subjected to PCR with specific primers for the *cdrA* gene of the HPK5 strain such as P1 (forward), P3 (reverse) and P4 (forward), previously described^[17]. Additional new primers, P9J (reverse), P3J (reverse) and P4J (forward), based on the sequence of the *cdrA* gene of J99 strains, were also utilized in this study. The sequence of primer P1 was identical in strains, HPK5 and J99. The conditions used for PCR to amplify the 194-bp or 336-bp products in the central region of *cdrA*, which is highly conserved among strains using P3-P4 or P3J-P4J primer pairs, respectively, were as follows: pre-heat for 2 min at 96 °C, followed by 40 cycles of 96 °C for 30 s, 55 °C for 30 s, and 72 °C for 30 s. To amplify the N-terminal region of *cdrA* using P1-P3 or P1-P3J primer pairs, PCR was performed with the following conditions: pre-heat for 2 min at 96 °C, followed by 40 cycles of 96 °C for 30 s, 52 °C for 30 s, and 72 °C for 2.5 min. PCR was carried out at least two times with all sets of primer pairs, P1-P3, P1-P3J, P3-P4 and P3J-P4J, for determining the standing of the *cdrA* gene. In particular, for the strains representing no PCR product using various primer pairs mentioned above, P9J-P4 or P9J-P4J primer pairs were used to confirm the existence of the region from central to C-terminal on *cdrA* corresponding to flanking the downstream region of urease gene cluster. The PCR conditions were as follows: pre-heat for 2 min at 96 °C, followed by 40 cycles of 96 °C for 30 s, 55 °C for 30 s, and 72 °C for 1 min. The location of these primers used for *cdrA* amplification (Figure 1) and primer sequences are shown (Table 1).

Statistical analysis

Fischer’s exact test and the χ^2 test with Yates’ continuity correction were applied using SPSS version 10.0 for Windows to compare differences on the level of IL-8 productions induced by *cdrA*-positive and -negative *H. pylori* strains. *P* value of < 0.05 were considered significant.

RESULTS

IL-8 secretion from AGS cells

To determine the effect of host response to *H. pylori*

Table 1 The sequence of primers used and the target region of cell division-related gene A amplified with combination of primers

Primers	Sequence (5'-3')	Reference	Combination of primers	Target region of cell division-related gene A
P1 (forward)	TGAAGCACACGAAAGGA	17	P1-P3	N-terminal to central
P3 (reverse)	CATGCTCTAAAATCGTGC	17	P1-P3J	N-terminal to central
P3J (reverse)	ATCACACGATCAGTCTG	This study	P3-P4	Central
P4 (forward)	TTTTGAGATCGGTGAAGC	17	P3J-P4J	Central
P4J (forward)	AACCACACGCTCATTTCG	This study	P4-P9J	Central to C-terminal
P9J (reverse)	GCTGAAAGGCTGAATTCAG	This study	P4J-P9J	Central to C-terminal

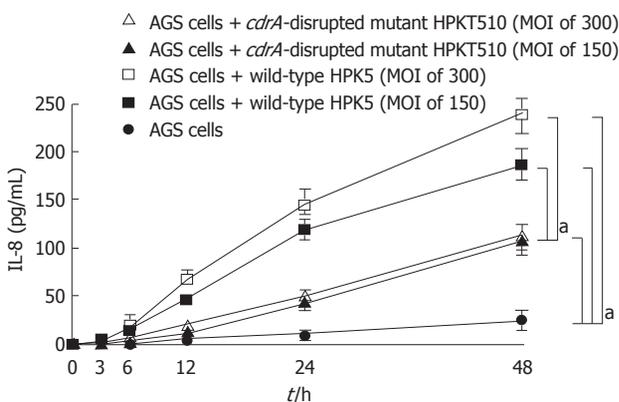


Figure 2 Interleukin-8 production from AGS cells induced by either wild-type or cell division-related gene A-disrupted mutant strains. ^a*P* < 0.01.

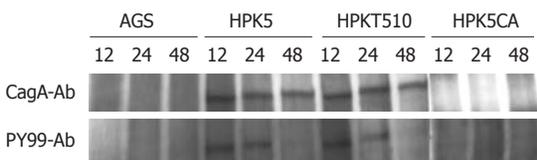


Figure 3 Cytotoxin-associated gene A status in AGS cells co-cultured with or without *Helicobacter pylori* strains. The immunoprecipitated proteins with anti-*Helicobacter pylori* (*H. pylori*) cytotoxin-associated gene A (CagA) antibody (CagA-Ab) were subjected to Western blotting with CagA-Ab (upper) and PY-99 antibody (PY99-Ab) (bottom), respectively. The assay was carried out with strains at MOI of 150. HPK5: Wild-type; HPKT510: Cell division-related gene A-disrupted mutant; HPK5CA: *cagA*-disrupted mutant.

during persistent infection in the human stomach, *in vitro* levels of IL-8 production from AGS cells co-cultured with HPK5 or HPKT510 strains were examined relative to time (Figure 2). Both strains induced the secretion of IL-8 from AGS cells and the levels increased time-dependently, but HPKT510, the *cdrA*-disrupted mutant, showed significantly lower levels, corresponding to 50%-60% of levels induced by wild-type HPK5 at MOI of 150 (*P* < 0.01). Furthermore, the levels of IL-8 were increased dose (MOI)-dependently by HPK5, but not HPKT510, which showed identical levels between MOI of 150 and 300.

Western blotting for CagA

Ultrastructural analyses revealed that the HPKT510 strain had a slightly wider periplasmic space between the inner and the outer membrane than that of HPK5^[16]. To determine whether such morphological difference af-

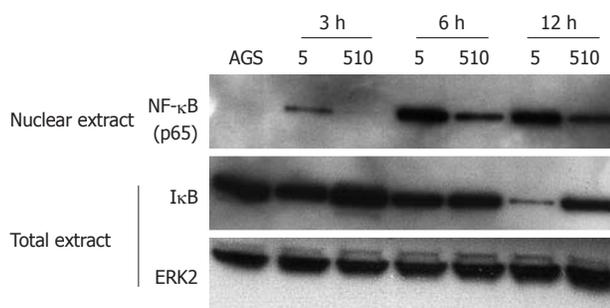


Figure 4 Detection of nuclear factor kappa B (p65) (upper), inhibition kappa B (middle) and extracellular signal-regulated kinase 2 (ERK2) in nuclear and total extracts, respectively at 3, 6 and 12 h after being co-cultured with *Helicobacter pylori*. Unphosphorylated extracellular signal-regulated kinase 2 (ERK2) was detected as the control in this study^[23]. Molecular weights of nuclear factor kappa B (NF-κB), inhibition kappa B (IκB) and ERK2 are 65, 35-37 and 42 kDa, respectively. The assay was carried out with strains at a MOI of 150. AGS: AGS cells co-cultured without *H. pylori*; HPK5: AGS cells co-cultured with wild-type HPK5; HPKT510: AGS cells co-cultured with *cdrA*-disrupted mutant HPKT510.

fects translocation of CagA into AGS *via* type IV secretory component system (T4SS) and its phosphorylation, which are considered to be risk factor for development of gastroduodenal disease, western blotting following immunoprecipitation with anti-*H. pylori* CagA antibody was carried out. CagA was detected in AGS cells co-cultured with HPK5 or HPKT510 strains, but no CagA was observed in AGS alone or when co-cultured with a *cagA*-disrupted mutant, HPK5CA as a negative control (Figure 3). The intensities of each CagA band are similar between both strains. Phosphorylation of CagA was detected up to 24 h after co-culture with HPK5 or HPKT510 strains and had similar intensity, indicating that there were no differences in the status of CagA between both strains.

Western blotting for NF-κB (p65) and IκB

To determine how the expression level of NF-κB transcription factor affects the expression of many genes involved in inflammation and immunities, western blotting was carried out utilizing AGS cells co-cultured with HPK5 or HPKT510 strains for 3, 6 and 12 h. Overall, NF-κB levels in the nuclear extracts from AGS cells with HPKT510 were definitely lower compared to those with HPK5, whereas IκB levels in total extracts were found to be higher in HPKT510 (Figure 4), indicating that activation of NF-κB was relatively repressed in AGS cells stimulated by *H. pylori* with disruption of *cdrA*.

Table 2 *Helicobacter pylori* cell division-related gene A status (positive or negative) and genotype (allele type) in clinical isolates *n* (%)

<i>H. pylori cdrA</i>		United States (48 patients) ¹	Japan (51 patients)
Status	Genotype		
Positive	Type I	0 (0)	4 (7.8)
	Type II	13 (16.9)	46 (90.2)
Negative	Type III	29 (37.7)	1 (2.0)
	Type IV	35 (45.4)	0 (0)
Total		77 ²	51

H. pylori cdrA: *Helicobacter pylori* cell division-related gene A. ¹In 29 out of 48 patients, each isolate was obtained from both antrum and corpus in same stomach; ²The total 77 isolates include 44 and 33 isolates obtained from antrum and corpus, respectively.

Genetic diversity of *H. pylori cdrA*

To examine the *cdrA* status [category such as *cdrA*-positive and -negative, including 4 allele types (I to IV)] in 128 clinical isolates, PCR was performed with appropriate specific primer pairs. In a previous report, PCR with P1-P3 yielded the 731 bp (short fragment) and approximately 2 kb (long fragment) in HPK5 and other three strains examined respectively, demonstrating that the shorter product of HPK5 was due to partial deletion in the N-terminal region of *cdrA*. However, *cdrA* was functionally involved in cell division, morphology, long-term survival and susceptibilities to beta-lactam and its transcript could be detected^{116,171}. On the other hand, the 194-bp amplified in the central region of *cdrA* was detected in all strains, indicating that size diversity exists within the N-terminal region of *cdrA* among individual strains¹¹⁷. In this study, 128 clinical isolates (77 American and 51 Japanese) were subjected to PCR with N-terminal primer pairs (P1-P3 and P1-P3J) which demonstrated that the short fragment (allele type I) was obtained in only 4 Japanese isolates (7.8%) including HPK5 and the long fragment (allele type II) was detected in 13 American (16.9%) and 46 Japanese (90.2%) isolates. In addition, no product was observed in the 64 American (83.1%) and 1 Japanese (2.0%) isolates by PCR with central region primer pairs (P3-P4 and P3J-P4J). Of the 64 American isolates showing no amplification on the central region, 35 isolates have no product amplified on the C-terminal region with P9J-P4J and P9J-P4 pairs. The results revealed that the downstream region of the urease gene cluster, including *cdrA*, was absent in the 35 isolates (allele type IV). However, PCR product on the C-terminal region with P9J-P4J or P9J-P4 pairs was detected in remaining 29 isolates, demonstrating that they have partial *cdrA* sequence loss between the N-terminal and central regions (allele type III). Taken together, the status of the *cdrA* gene among individual isolates examined was divided into two categories; *cdrA*-positive including allele types I (HPK5) and II (J99 and 26695), and *cdrA*-negative including allele types III and IV (Table 2).

Comparison of *cdrA* status between antrum and corpus

The *cdrA* allele type of each isolate from the antrum and

Table 3 Comparison of cell division-related gene A status and genotype (allele type) between antrum and corpus isolates in 29 American patients

	<i>cdrA</i> status ¹	<i>cdrA</i> genotype (allele type)
<i>cdrA</i> positive (6 cases)	Same ² : 1	Antrum (+) ³ Corpus (+)
	Difference: 5	Antrum (+) Corpus (-)
		Antrum (-) Corpus (-)
<i>cdrA</i> negative (23 cases)	Same: 22	Antrum (-) Corpus (-)
	Difference: 1	Antrum (-) Corpus (+)
		Antrum (+) Corpus (+)
Total (%)	Same: 23 (79) Difference: 6 (21)	Identical ⁴ : 1 Difference: 5 Identical: 16 Difference: 7 Same: 17 (59) Difference: 12 (41)

¹The status of cell division-related gene A (*cdrA*) determined with isolate from antrum was divided to two category groups (*cdrA* positive and negative); ²Same or difference: *cdrA* status is same or difference between isolate (antrum and corpus); ³+: *cdrA* positive isolate; -: *cdrA* negative isolate; ⁴Identical or difference: the genotypes of *cdrA* is identical or difference between isolate (antrum and corpus). Bold denotes the 6 cases representing differences at *cdrA* status and genotype between isolates (antrum and corpus).

corpus in the stomach of 29 American patients demonstrated that *cdrA* allele between the regions was identical in 17 patients (59%); however, 9 of the remaining 12 patients possessing a different allele had a isolate with more loss of the *cdrA* sequence in the corpus compared to the antrum (Table 3). Regarding the status of *cdrA*, 6 out of 29 patients had different *cdrA* status between the regions, and of which, 5 patients harbored *cdrA*-positive and -negative isolates in the antrum and corpus, respectively (Table 3).

Measurement of mucosal IL-8 secretion

The *cdrA*-disrupted mutant HPKT510 showed significantly lower levels of IL-8 secretion *in vitro* compared to wild-type HPK5. To investigate whether *cdrA* is associated with IL-8 production *in vivo*, mucosal IL-8 secretion of biopsy specimens was measured using samples obtained from 20 American patients infected with *cdrA*-negative and 5 American patients with *cdrA*-positive *H. pylori*. Measurement of IL-8 in these specimens demonstrated that the *cdrA*-negative group exhibited lower IL-8 levels than those of *cdrA*-positive group. The average value of IL-8 in the *cdrA*-negative group was 111 pg/mL, corresponding to the average of 60% in the *cdrA*-positive group (156 pg/mL). This tendency was consistent with *in vitro* data, revealing that *cdrA*-negative *H. pylori* does not induce IL-8 secretion as strongly in the human stomach as *cdrA*-positive strains.

DISCUSSION

H. pylori colonizes more than half of the world's population. While it is clear that this organism induces a strong innate and adaptive immune response leading to active inflammation in the gastric mucosa, the ability of *H. pylori* to establish persistent infection so efficiently has not been fully elucidated. *H. pylori* possesses a number

of virulence factors, and some, such as urease, flagella and lipopolysaccharide (LPS), contribute to its persistence^[25-27], whereas others, such as CagA and vacuolating cytotoxin, appear to confer increased virulence^[28,29]. Furthermore, the strain-specific genes found outside of the *cag* pathogenicity island, especially genes in the plasticity regions^[30] and genes with variable structures/genotypes, have considerable interest in their contribution to pathogenesis and the host immune response. Nearly half of the strain-specific genes of *H. pylori* are located in the plasticity regions in strains 26695^[31] and J99^[30,32]. There is evidence that genetic recombination among *H. pylori* may occur during the course of infection in the stomach. The diversity or phase variation in *H. pylori* genes such as *babA*^[33] and *fucT1*^[26] likely contributes to the evasion of the host immune response and thereby has a role in the establishment of persistent infection in the stomach. Based on the genomic information of the sequenced *H. pylori* strains 26695^[31] and J99^[32], more than a third of *H. pylori* genes have unknown function, suggesting that there may be strain-specific genes such as *atkA*^[34] involved in the mechanisms of persistence as well as the host immune response.

One of unique genes found in the HPK5 strain, *cdrA* located in the downstream region of urease gene cluster, has not only a inhibitory role for cell division, but is also involved in elongation and death of cells *via* cell wall synthesis at the site of division^[16,17]. Furthermore, the *cdrA*-disrupted mutant HPKT510 had a longer survival time compared to the wild-type HPK5 in both liquid and solid media, as well as in serum-free medium and aerobic conditions^[16]. Loss of the *cdrA* gene during infection is frequently found in a mouse infected with *H. pylori* strain B128^[18] and in the human stomach infected with J99^[19]. The present study found that *cdrA*-negative strains resulted in lower levels of IL-8 production *in vitro* and *in vivo* compared to *cdrA*-positive strains. *In vitro* data showed that increased IL-8 production was dose-dependently observed by *cdrA*-positive HPK5, but not *cdrA*-disrupted HPKT510. In addition, nuclear accumulation of NF- κ B in AGS co-cultured with HPKT510 was lower compared to HPK5, suggesting that activation of the NF- κ B signaling pathway was relatively repressed by stimulation of the *cdrA*-negative strain, which coincided with decreased IL-8 production. These indicated that *cdrA*-defective *H. pylori* may evade immune clearance due to limiting bactericidal effects of pro-inflammatory molecules leading to promotion for persistent infection, likely *via* repression of the NF- κ B signaling pathway. Persistent infection was not observed in IL-10-deficient mice with strong inflammation, implying that attenuate host immunity is necessary for *H. pylori* colonization^[35]. Accordingly, it is acceptable that *H. pylori* infection occurred in the stomach with immature immunity during infantile generation. The proper NF- κ B transcriptional response is primarily regulated by post-translational modification of NF- κ B signaling components. On the

other hand, alternative splicing of NF- κ B signaling components is another way to control the NF- κ B signaling pathway^[36,37]. Alternative splicing of some NF- κ B signaling components can be induced by prolonged exposure to a NF- κ B-activating signal, such as LPS, suggesting a mechanism for negative feedback to dampen excessive NF- κ B signaling. In particular, alternative splicing events in Toll/interleukin-1 NF- κ B signaling pathways *via* Toll like receptors can inhibit the NF- κ B response^[38]. This raises the possibility that changes in bacterial constituents concerned with disruption of *cdrA*, such as cell envelope components including LPS, may exert on the complicated NF- κ B signaling pathway.

As HPKT510 had a slightly wider periplasmic space^[16], the effector protein, CagA, was investigated to determine whether this morphological change altered the function of T4SS. The CagA production, its translocation and phosphorylation were indistinguishable between HPK5 and HPKT510 strains, indicating that such morphological differences related with inactivation of *cdrA* had no effect on CagA-associated pathogenicity, including CagA-related signaling pathway and T4SS. Cytoplasmic nucleotide-binding oligomerization domain (Nod) molecules, Nod1 and Nod2, have been shown to be specific ligands of diaminopimelic acid-containing muropeptides and muramyl dipeptide, respectively^[39,40], and were important components of the innate immune response. A peptidoglycan-derived muropeptide possessing a Nod1 motif was translocated through T4SS and affected cell signaling *via* activation of NF- κ B in the host cell, leading to the stimulation of an intracellular pro-inflammatory response^[41]. In terms of conserved domains, part of HPK5 *cdrA* belongs to the SM1-NRK4 family, beta glucan and cell wall synthesis enzyme family, and another belongs to the SpoIIE-FtsK-ATPase family. The properties of penicillin-binding proteins such as PBP1, PBP2, PBP3 and PBP4 varied between HPK5 and HPKT510 strains^[16], suggesting that an alteration of such peptidoglycans influences the level of IL-8 production *via* the Nod1-NF- κ B signaling pathway. In fact, the measurement of *nod1* transcript level in AGS by real-time RT-PCR documented that HPKT510 induced lower level of *nod1* transcript compared to HPK5 (data not shown), which supports the Nod1-NF- κ B pathway involved in the difference of IL-8 production.

The *cdrA* status in almost all Japanese isolates were *cdrA*-positive (98%), but not in isolates obtained from Americans, demonstrating that American isolates had a greater diversity and higher prevalence of *cdrA*-negative strains (83.1%). Evaluation of *cdrA* status in isolates obtained from different regions of the stomach revealed that in 5 of the 6 cases possessing different status between the antrum and corpus of the stomach, the *cdrA*-negative isolate was from the corpus, suggesting that a *cdrA*-negative strain might be more adaptable to colonize in the corpus over a longer time period than the *cdrA*-positive strain. A *cdrA*-disrupted mutant could survive

longer under the stresses, such as presence of beta-lactam antibiotics, serum-free and aerobic conditions, than the wild-type strain^[16]. It is possible that changes in biological behavior accompanied by inactivation of *cdrA* are necessary to stay balanced for the establishment of long-term infection in the human stomach. When the HPK5 *cdrA* sequence was compared to reference strain 26695^[31], the *cdrA* sequence was a fusion of the ~130 codon hp0064 gene to the middle ~one third of the hp0066 gene, with deletion of the interstitial hp0065 and first part of hp0066. A similar fusion is evident with respect to the reference strain J99 (jhp0059 and jhp0061) or HPAG1 (hpag1_0064 and hpag1_0067)^[42]. These two reference strains might represent the majority class in the Japanese population, which gave 2 kb instead of 730 bp PCR products. In contrast, two other sequenced strains, Shi470^[43] and G27^[44], seem to contain only the ~130 codon ORF, not the longer one, and thus might possibly represent one of the major classes in the United States population. The HPK5 type *cdrA* stems from gene fusion and synthesizes a partially defective protein that impacts cell wall synthesis or structure and a greater release of peptidoglycan fragments, which can be proinflammatory through the Nod1-NF- κ B pathway. More studies are required in the future to dissect the mechanistic role of *cdrA* and the alteration of comprehensive genomics/proteomics affected by inactivation of *cdrA*. Analysis for putative deletion alleles might actually provide us with insights to elucidate whether the phenomenon resulted from such sources as large insertions or genome rearrangements.

In histological observation, as far the confined tissue sections of small biopsy specimens were examined, no significant finding was observed in the gastric mucosa colonized with either *cdrA*-positive or *cdrA*-negative *H. pylori* strains. We may need more examinations in detail with extended sections and specimens to confirm the differences in histological findings, including inflammation with neutrophil infiltration.

We concluded that the presence of *H. pylori cdrA* was associated with effective production of pro-inflammatory cytokine, IL-8, both *in vitro* and *in vivo*. Therefore, *cdrA*-inactivated *H. pylori* strains may result in attenuated host immunity and evade immune clearance due to repression of the NF- κ B signaling pathway in the host cell, leading to persistent infection. Finally, our studies suggest that *cdrA*-negative *H. pylori* strains are more likely to colonize in the corpus over long time periods compared with *cdrA*-positive strains.

COMMENTS

Background

Helicobacter pylori (*H. pylori*) infection up-regulates secretion of various inflammatory cytokines including interleukin-8 (IL-8), whose production *in vitro* and *in vivo* is recognized as a host response to microbes. The levels of IL-8 production differ among individual strains, suggesting the existence of unique genes involved in host response and genetic diversity during infection in the stomach. Furthermore, the effect of fundamental constituents belonging to cell division of

H. pylori on host response is unclear.

Research frontiers

H. pylori infection and its persistence in the stomach is the most important event to actually lead a variety of diseases. However, the functions and practices of the cell division of strains colonized in an individual stomach remain unclear. Furthermore, how the bacterial behaviors concerned with the host response is still unknown. In this study, the authors demonstrate that *H. pylori* cell division-related gene A (*cdrA*) required for cell division and morphological shape could be a potential role for mediating IL-8 secretion.

Innovations and breakthroughs

Reports have highlighted the importance of *H. pylori*-host interaction to understand host immunity and pathogenesis. Certain molecules, such as cytotoxin-associated gene A (*CagA*) and outer membrane proteins, were shown to be involved in these phenomena; however, the relationship between cell division- and morphological shape-related molecules and host response associated with persistent infection in the stomach is unknown. This is the first study to report that *cdrA* influenced IL-8 secretion (*vivo* and *vitro*) and loss of *cdrA* may attenuate the host immunity due to repression of NF- κ B, leading to persistence.

Applications

In this study, how *H. pylori cdrA* functions and influences bacterium-host interaction was investigated, which provides insights into understanding bacterial fundamental components and their effect on the stomach, including host response, and a future strategy for therapeutic intervention in the treatment of patients with *H. pylori* infection.

Terminology

H. pylori cdrA, one of the unique genes discovered in strain HPK5, has not only a repressive role on cell division but is also involved in cell elongation and cell death *via* cell wall synthesis (mainly penicillin binding proteins) at the division site. The *cdrA*-disrupted mutant is able to survive for long time periods and is more resistant to the bactericidal of beta-lactam antibiotics than the wild-type. The *cdrA* gene tends to be lost during infection to facilitate adaption in the stomach, resulting in a persistent infection.

Peer review

This interesting manuscript showed that the presence of *H. pylori cdrA* was associated with the effective production of IL-8 compared to the inactivation of *cdrA in vitro* and *in vivo*. Additionally, they observed that *cdrA*-inactivated *H. pylori* strains may result in attenuated host immunity and evade immune clearance due to repression of the NF- κ B pathway, leading to persistent infection.

REFERENCES

- 1 Peek RM, Blaser MJ. Helicobacter pylori and gastrointestinal tract adenocarcinomas. *Nat Rev Cancer* 2002; **2**: 28-37
- 2 Achtman M, Azuma T, Berg DE, Ito Y, Morelli G, Pan ZJ, Suerbaum S, Thompson SA, van der Ende A, van Doorn LJ. Recombination and clonal groupings within Helicobacter pylori from different geographical regions. *Mol Microbiol* 1999; **32**: 459-470
- 3 Akopyanz N, Bukanov NO, Westblom TU, Kresovich S, Berg DE. DNA diversity among clinical isolates of Helicobacter pylori detected by PCR-based RAPD fingerprinting. *Nucleic Acids Res* 1992; **20**: 5137-5142
- 4 Go MF, Kapur V, Graham DY, Musser JM. Population genetic analysis of Helicobacter pylori by multilocus enzyme electrophoresis: extensive allelic diversity and recombinational population structure. *J Bacteriol* 1996; **178**: 3934-3938
- 5 Salaün L, Audibert C, Le Lay G, Burucoa C, Fauchère JL, Picard B. Panmictic structure of Helicobacter pylori demonstrated by the comparative study of six genetic markers. *FEMS Microbiol Lett* 1998; **161**: 231-239
- 6 Suerbaum S, Smith JM, Bapumia K, Morelli G, Smith NH, Kunstmann E, Dyrek I, Achtman M. Free recombination within Helicobacter pylori. *Proc Natl Acad Sci USA* 1998; **95**: 12619-12624
- 7 van Doorn NEM, Namavar F, Kusters JG, van Rees EP, Kuipers EJ, de Graaff J. Genomic DNA fingerprinting of clinical isolates of Helicobacter pylori by REP-PCR and restriction fragment end-labelling. *FEMS Microbiol Lett* 1998; **160**: 145-150

- 8 **Blaser MJ**. In *Helicobacter pylori*. In: Hunt RH, Tytgat GNH. Basic Mechanisms to Clinical Cure. Dordrecht: Kluwer, 1996: 33-39
- 9 **Yamaoka Y**, Kita M, Kodama T, Sawai N, Kashima K, Imanishi J. Induction of various cytokines and development of severe mucosal inflammation by *cagA* gene positive *Helicobacter pylori* strains. *Gut* 1997; **41**: 442-451
- 10 **Crabtree JE**, Farmery SM, Lindley IJ, Figura N, Peichl P, Tompkins DS. *CagA*/cytotoxic strains of *Helicobacter pylori* and interleukin-8 in gastric epithelial cell lines. *J Clin Pathol* 1994; **47**: 945-950
- 11 **Sharma SA**, Tummuru MK, Miller GG, Blaser MJ. Interleukin-8 response of gastric epithelial cell lines to *Helicobacter pylori* stimulation in vitro. *Infect Immun* 1995; **63**: 1681-1687
- 12 **Nozawa Y**, Nishihara K, Peek RM, Nakano M, Uji T, Ajioka H, Matsuura N, Miyake H. Identification of a signaling cascade for interleukin-8 production by *Helicobacter pylori* in human gastric epithelial cells. *Biochem Pharmacol* 2002; **64**: 21-30
- 13 **Sharma SA**, Tummuru MK, Blaser MJ, Kerr LD. Activation of IL-8 gene expression by *Helicobacter pylori* is regulated by transcription factor nuclear factor-kappa B in gastric epithelial cells. *J Immunol* 1998; **160**: 2401-2407
- 14 **Keates S**, Keates AC, Warny M, Peek RM, Murray PG, Kelly CP. Differential activation of mitogen-activated protein kinases in AGS gastric epithelial cells by *cag+* and *cag-* *Helicobacter pylori*. *J Immunol* 1999; **163**: 5552-5559
- 15 **Ghosh S**, Karin M. Missing pieces in the NF-kappaB puzzle. *Cell* 2002; **109** Suppl: S81-S96
- 16 **Takeuchi H**, Nakazawa T, Okamoto T, Shirai M, Kimoto M, Nishioka M, Con SA, Morimoto N, Sugiura T. Cell elongation and cell death of *Helicobacter pylori* is modulated by the disruption of *cdrA* (cell division-related gene A). *Microbiol Immunol* 2006; **50**: 487-497
- 17 **Takeuchi H**, Shirai M, Akada JK, Tsuda M, Nakazawa T. Nucleotide sequence and characterization of *cdrA*, a cell division-related gene of *Helicobacter pylori*. *J Bacteriol* 1998; **180**: 5263-5268
- 18 **Israel DA**, Salama N, Arnold CN, Moss SF, Ando T, Wirth HP, Tham KT, Camorlinga M, Blaser MJ, Falkow S, Peek RM. *Helicobacter pylori* strain-specific differences in genetic content, identified by microarray, influence host inflammatory responses. *J Clin Invest* 2001; **107**: 611-620
- 19 **Israel DA**, Salama N, Krishna U, Rieger UM, Atherton JC, Falkow S, Peek RM. *Helicobacter pylori* genetic diversity within the gastric niche of a single human host. *Proc Natl Acad Sci USA* 2001; **98**: 14625-14630
- 20 **Zhang Y**, Takeuchi H, Nishioka M, Morimoto N, Kamioka M, Kumon Y, Sugiura T. Relationship of IL-8 production and the *CagA* status in AGS cells infected with *Helicobacter pylori* exposed to low pH and activating transcription factor 3 (ATF3). *Microbiol Res* 2009; **164**: 180-190
- 21 **Trang VT**, Takeuchi H, Kudo H, Aoki A, Katsuno S, Shimamura T, Sugiura T, Ukeda H. Antimicrobial activity of aminoreductone against *Helicobacter pylori*. *J Agric Food Chem* 2009; **57**: 11343-11348
- 22 **Mi J**, Zhang X, Liu Y, Reddy SK, Rabbani ZN, Sullenger BA, Clary BM. NF-kB inhibition by an adenovirus expressed aptamer sensitizes TNF α -induced apoptosis. *Biochem Biophys Res Commun* 2007; **359**: 475-480
- 23 **Schmeck B**, N'Guessan PD, Ollomang M, Lorenz J, Zahlten J, Opitz B, Flieger A, Suttrop N, Hippenstiel S. Legionella pneumophila-induced NF-kappaB- and MAPK-dependent cytokine release by lung epithelial cells. *Eur Respir J* 2007; **29**: 25-33
- 24 **Peek RM**, Thompson SA, Donahue JP, Tham KT, Atherton JC, Blaser MJ, Miller GG. Adherence to gastric epithelial cells induces expression of a *Helicobacter pylori* gene, *iceA*, that is associated with clinical outcome. *Proc Assoc Am Physicians* 1998; **110**: 531-544
- 25 **Eaton KA**, Morgan DR, Krakowka S. Motility as a factor in the colonisation of gnotobiotic piglets by *Helicobacter pylori*. *J Med Microbiol* 1992; **37**: 123-127
- 26 **Khamri W**, Moran AP, Worku ML, Karim QN, Walker MM, Annuk H, Ferris JA, Appelmelk BJ, Eggleton P, Reid KB, Thursz MR. Variations in *Helicobacter pylori* lipopolysaccharide to evade the innate immune component surfactant protein D. *Infect Immun* 2005; **73**: 7677-7686
- 27 **Meyer-Rosberg K**, Scott DR, Rex D, Melchers K, Sachs G. The effect of environmental pH on the proton motive force of *Helicobacter pylori*. *Gastroenterology* 1996; **111**: 886-900
- 28 **Covacci A**, Censini S, Bugnoli M, Petracca R, Burrioni D, Macchia G, Massone A, Papini E, Xiang Z, Figura N, Rappuoli R. Molecular characterization of the 128-kDa immunodominant antigen of *Helicobacter pylori* associated with cytotoxicity and duodenal ulcer. *Proc Natl Acad Sci USA* 1993; **90**: 5791-5795
- 29 **Leunk RD**, Johnson PT, David BC, Kraft WG, Morgan DR. Cytotoxic activity in broth-culture filtrates of *Campylobacter pylori*. *J Med Microbiol* 1988; **26**: 93-99
- 30 **Yamaoka Y**. Roles of the plasticity regions of *Helicobacter pylori* in gastroduodenal pathogenesis. *J Med Microbiol* 2008; **57**: 545-553
- 31 **Tomb JF**, White O, Kerlavage AR, Clayton RA, Sutton GG, Fleischmann RD, Ketchum KA, Klenk HP, Gill S, Dougherty BA, Nelson K, Quackenbush J, Zhou L, Kirkness EF, Peterson S, Loftus B, Richardson D, Dodson R, Khalak HG, Glodek A, McKenney K, Fitzgerald LM, Lee N, Adams MD, Hickey EK, Berg DE, Gocayne JD, Utterback TR, Peterson JD, Kelley JM, Cotton MD, Weidman JM, Fujii C, Bowman C, Watthey L, Wallin E, Hayes WS, Borodovsky M, Karp PD, Smith HO, Fraser CM, Venter JC. The complete genome sequence of the gastric pathogen *Helicobacter pylori*. *Nature* 1997; **388**: 539-547
- 32 **Alm RA**, Ling LS, Moir DT, King BL, Brown ED, Doig PC, Smith DR, Noonan B, Guild BC, deJonge BL, Carmel G, Tummino PJ, Caruso A, Uria-Nickelsen M, Mills DM, Ives C, Gibson R, Merberg D, Mills SD, Jiang Q, Taylor DE, Vovis GF, Trust TJ. Genomic-sequence comparison of two unrelated isolates of the human gastric pathogen *Helicobacter pylori*. *Nature* 1999; **397**: 176-180
- 33 **Aspholm-Hurtig M**, Dailide G, Lahmann M, Kalia A, Ilver D, Roche N, Vikström S, Sjöström R, Lindén S, Bäckström A, Lundberg C, Arnqvist A, Mahdavi J, Nilsson UJ, Velapataño B, Gilman RH, Gerhard M, Alarcon T, López-Brea M, Nakazawa T, Fox JG, Correa P, Dominguez-Bello MG, Perez-Perez GI, Blaser MJ, Normark S, Carlstedt I, Oscarson S, Teneberg S, Berg DE, Borén T. Functional adaptation of *BabA*, the *H. pylori* ABO blood group antigen binding adhesin. *Science* 2004; **305**: 519-522
- 34 **Kim do J**, Park KS, Kim JH, Yang SH, Yoon JY, Han BG, Kim HS, Lee SJ, Jang JY, Kim KH, Kim MJ, Song JS, Kim HJ, Park CM, Lee SK, Lee BI, Suh SW. *Helicobacter pylori* proinflammatory protein up-regulates NF- κ B as a cell-translocating Ser/Thr kinase. *Proc Natl Acad Sci USA* 2010; **107**: 21418-21423
- 35 **Matsumoto Y**, Blanchard TG, Drakes ML, Basu M, Redline RW, Levine AD, Czinn SJ. Eradication of *Helicobacter pylori* and resolution of gastritis in the gastric mucosa of IL-10-deficient mice. *Helicobacter* 2005; **10**: 407-415
- 36 **Courtois G**, Gilmore TD. Mutations in the NF-kappaB signaling pathway: implications for human disease. *Oncogene* 2006; **25**: 6831-6843
- 37 **Gerondakis S**, Grumont R, Gugasyan R, Wong L, Isomura I, Ho W, Banerjee A. Unravelling the complexities of the NF-kappaB signalling pathway using mouse knockout and transgenic models. *Oncogene* 2006; **25**: 6781-6799
- 38 **Leeman JR**, Gilmore TD. Alternative splicing in the NF-kappaB signaling pathway. *Gene* 2008; **423**: 97-107

- 39 **Chamaillard M**, Hashimoto M, Horie Y, Masumoto J, Qiu S, Saab L, Ogura Y, Kawasaki A, Fukase K, Kusumoto S, Valvano MA, Foster SJ, Mak TW, Nunez G, Inohara N. An essential role for NOD1 in host recognition of bacterial peptidoglycan containing diaminopimelic acid. *Nat Immunol* 2003; **4**: 702-707
- 40 **Inohara N**, Ogura Y, Fontalba A, Gutierrez O, Pons F, Crespo J, Fukase K, Inamura S, Kusumoto S, Hashimoto M, Foster SJ, Moran AP, Fernandez-Luna JL, Nuñez G. Host recognition of bacterial muramyl dipeptide mediated through NOD2. Implications for Crohn's disease. *J Biol Chem* 2003; **278**: 5509-5512
- 41 **Viala J**, Chaput C, Boneca IG, Cardona A, Girardin SE, Moran AP, Athman R, Mémet S, Huerre MR, Coyle AJ, DiStefano PS, Sansonetti PJ, Labigne A, Bertin J, Philpott DJ, Ferrero RL. Nod1 responds to peptidoglycan delivered by the *Helicobacter pylori* cag pathogenicity island. *Nat Immunol* 2004; **5**: 1166-1174
- 42 **Oh JD**, Kling-Bäckhed H, Giannakis M, Xu J, Fulton RS, Fulton LA, Cordum HS, Wang C, Elliott G, Edwards J, Mardis ER, Engstrand LG, Gordon JI. The complete genome sequence of a chronic atrophic gastritis *Helicobacter pylori* strain: evolution during disease progression. *Proc Natl Acad Sci USA* 2006; **103**: 9999-10004
- 43 **Thiberge JM**, Boursaux-Eude C, Lehours P, Dillies MA, Creno S, Coppée JY, Rouy Z, Lajus A, Ma L, Burucoa C, Ruskoné-Foumestreaux A, Courillon-Mallet A, De Reuse H, Boneca IG, Lamarque D, Mégraud F, Delchier JC, Médigue C, Bouchier C, Labigne A, Raymond J. From array-based hybridization of *Helicobacter pylori* isolates to the complete genome sequence of an isolate associated with MALT lymphoma. *BMC Genomics* 2010; **11**: 368
- 44 **Baltrus DA**, Amieva MR, Covacci A, Lowe TM, Merrell DS, Ottemann KM, Stein M, Salama NR, Guillemin K. The complete genome sequence of *Helicobacter pylori* strain G27. *J Bacteriol* 2009; **191**: 447-448

S- Editor Tian L L- Editor Rutherford A E- Editor Li JY

A *Macaca mulatta* model of fulminant hepatic failure

Ping Zhou, Jie Xia, Gang Guo, Zi-Xing Huang, Qiang Lu, Li Li, Hong-Xia Li, Yu-Jun Shi, Hong Bu

Ping Zhou, Hong Bu, Department of Pathology, West China Hospital, Sichuan University, Chengdu 610041, Sichuan Province, China

Jie Xia, Gang Guo, Li Li, Yu-Jun Shi, Hong Bu, Key Laboratory of Transplant Engineering and Immunology, Ministry of Health, West China Hospital, Sichuan University, Chengdu 610041, Sichuan Province, China

Zi-Xing Huang, Department of Radiology, West China Hospital, Sichuan University, Chengdu 610041, Sichuan Province, China

Qiang Lu, Department of Ultrasound, West China Hospital, Sichuan University, Chengdu 610041, Sichuan Province, China

Hong-Xia Li, The National Chengdu Center for Safety Evaluation of Drugs, West China Hospital, Sichuan University, Chengdu 610041, Sichuan Province, China

Author contributions: Zhou P, Shi YJ and Bu H designed the project; Zhou P, Xia J, Guo G and Shi YJ performed the experiments; Zhou P and Shi YJ contributed to the discussion of the data and wrote the manuscript; Huang ZX contributed to the MRI examination; Lu Q contributed to the ultrasound examination; Li L contributed to the production and staining of section; Li HX provided the experimental animals and contributed to the biochemical examination.

Supported by National Basic Research Program of China, No. 2009CB522401; and grand from Natural Science Foundation of China, No. 30870983 and 30971118

Correspondence to: Yu-Jun Shi, MD, PhD, Key Laboratory of Transplant Engineering and Immunology, Ministry of Health, West China Hospital, Sichuan University, No. 1 Keyuan 4th Road, Chengdu 610041, Sichuan Province,

China. shiyujun@scu.edu.cn

Telephone: +86-28-85164030 Fax: +86-28-85164034

Received: May 19, 2011 Revised: June 23, 2011

Accepted: June 30, 2011

Published online: February 7, 2012

Abstract

AIM: To establish an appropriate primate model of fulminant hepatic failure (FHF).

METHODS: We have, for the first time, established a large animal model of FHF in *Macaca mulatta* by intra-

peritoneal infusion of amatoxin and endotoxin. Clinical features, biochemical indexes, histopathology and iconography were examined to dynamically investigate the progress and outcome of the animal model.

RESULTS: Our results showed that the enzymes and serum bilirubin were markedly increased and the enzyme-bilirubin segregation emerged 36 h after toxin administration. Coagulation activity was significantly decreased. Gradually deteriorated parenchymal abnormality was detected by magnetic resonance imaging (MRI) and ultrasonography at 48 h. The liver biopsy showed marked hepatocyte steatosis and massive parenchymal necrosis at 36 h and 49 h, respectively. The autopsy showed typical yellow atrophy of the liver. Hepatic encephalopathy of the models was also confirmed by hepatic coma, MRI and pathological changes of cerebral edema. The lethal effects of the extrahepatic organ dysfunction were ruled out by their biochemical indices, imaging and histopathology.

CONCLUSION: We have established an appropriate large primate model of FHF, which is closely similar to clinic cases, and can be used for investigation of the mechanism of FHF and for evaluation of potential medical therapies.

© 2012 Baishideng. All rights reserved.

Key words: Fulminant hepatic failure; *Macaca mulatta*; Biochemistry; Imaging; Pathology

Peer reviewer: Dr. Yasuji Arase, Department of Gastroenterology, Toranomon Hospital, 2-2-2 Toranomon Minato-ku, Tokyo 105-8470, Japan

Zhou P, Xia J, Guo G, Huang ZX, Lu Q, Li L, Li HX, Shi YJ, Bu H. A *Macaca mulatta* model of fulminant hepatic failure. *World J Gastroenterol* 2012; 18(5): 435-444 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/435.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.435>

INTRODUCTION

Fulminant hepatic failure (FHF) is an uncommon and challenging clinical disease characterized by sudden and severe hepatic injury and dysfunction, with many different symptoms and complications, and high mortality. The term was first used in 1970 to describe a potentially reversible disorder that was the result of severe liver injury in the absence of pre-existing liver disease, with an onset of encephalopathy within 8 wk of symptom appearance^[1]. The incidence and etiology of FHF vary according to different geographic regions^[1,2], and viral hepatitis and drug or toxin ingestion are the most common causes. As the pathophysiological changes are complicated and still need to be investigated, a clinically relevant large-animal model would be an indispensable tool. Orthotopic liver transplantation has been proved to be an effective treatment^[3-6], but the shortage of donor organs has restricted its wide application^[7-9]. The development of new therapies, including hepatocyte transplantation, stem cell transplantation, tissue engineered liver and bioartificial liver support systems, is still under investigation^[3-6]. Before these treatments can be used clinically, their effects should be determined in large animal models. For a satisfactory animal model of FHF, seven criteria were suggested by Terblanche *et al.*^[10] and Fourneau *et al.*^[11]: (1) reversibility; (2) reproducibility; (3) death from liver failure; (4) presence of a therapeutic window; (5) a large animal model; (6) minimal hazard to personnel; and (7) a conscious animal model. Previous reports have described different animal models of FHF induced by various methods^[7,12,13], but all the established models did not entirely satisfy all these criteria. Moreover, selection of a species with similar metabolic and physiological properties to humans is of importance. In this article, we describe, for the first time, the establishment of a *Macaca mulatta* model of FHF and the dynamic biochemical, pathological and imaging changes are also described.

MATERIALS AND METHODS

Chemicals

Lipopolysaccharide (LPS) from *Escherichia coli* was purchased from Sigma-Aldrich, Inc., St. Louis, MO. α -amanitin was from Alexis Biochemicals Co., Lausen, Switzerland. Gd-BOPTA (multihance) and SonoVue, contrast media for magnetic resonance imaging (MRI) and ultrasonography, respectively, were from Bracco Sine Pharmaceutical Co., Shanghai, China.

Animals

Two healthy female *Macaca mulattas*, 3 and 4 years old and with body weights of 5.5 kg and 4.6 kg, respectively, were purchased from the Safe and Secure Experimental Animal Breeding Base in Chengdu, China. They were housed in a large animal care facility with a constant temperature of 20 °C \pm 1 °C and a 6 am to 6 pm light cycle. Two weeks were allowed for the animals to acclimatize to

the animal facility before the study. They were fed with standard dry monkey food, washed apples and water *ad libitum*. Animal procedures and care were conducted in accordance with the institutional guidelines and in compliance with national and international laws and policies.

Study design

After premedication with ketamine (15 mg/kg im), skin preparation was performed on the posterior legs for blood collection from the saphenous vein and on the abdomen for intraperitoneal infusion and hepatic needle biopsy. LPS (1 μ g/kg) and α -amanitin (0.1 mg/kg) were mixed in 50 mL physiological saline and slowly infused into the peritoneal cavity. Before their administration, blood biochemical parameters of the liver, kidney, heart and pancreas were measured, and total body MRI and ultrasonography were performed to acquire the physiological data as baseline values (expressed as "0" in the figures). After administration, blood biochemical parameters were measured every 12 h. An imaging examination was performed every 24 h.

Core needle biopsy of the liver guided by ultrasonography scan was performed 12 h after administration and repeated every 24 h. Part of the needle biopsy specimens was subjected to frozen section and oil red O staining.

The state of consciousness and behavior of the animals was observed and the hepatic encephalopathy was defined according to the West-Haven criteria in humans^[14].

The two *Macaca mulattas* underwent necropsies after death, which included a full macroscopy and a histological evaluation of the liver, kidneys, lungs, heart, brain, spleen, pancreas and lymph nodes. Tissues were fixed in 10% neutral buffered formalin, sectioned at 5 μ m and stained with hematoxylin and eosin (H and E).

RESULTS

Clinical features and animal survival

The animals showed a gradual increase in listlessness and loss of appetite after toxin administration. Anorexia and vomiting occurred 24 h after administration, followed by grasping disability, mental indifference, drowsiness and coma. Hepatic coma appeared at 42 h and 56 h, and death occurred at 49 h and 70 h, respectively.

Biochemical results

We measured the liver enzymes, bilirubin and coagulation indices to determine the extent of liver injury. The data below were the average values from the two *Macaca mulatta* models when both were alive, or from one monkey when one died. After administration, the values of alanine aminotransferase and aspartate aminotransferase began to increase significantly and reached a peak at 36 h with over 300-fold and 200-fold increases from the baseline, respectively (Figure 1A), then the values sharply decreased. The total bilirubin, direct bilirubin and indirect bilirubin all increased gradually, and the final measured values exceeded a 30-fold rise from the corresponding

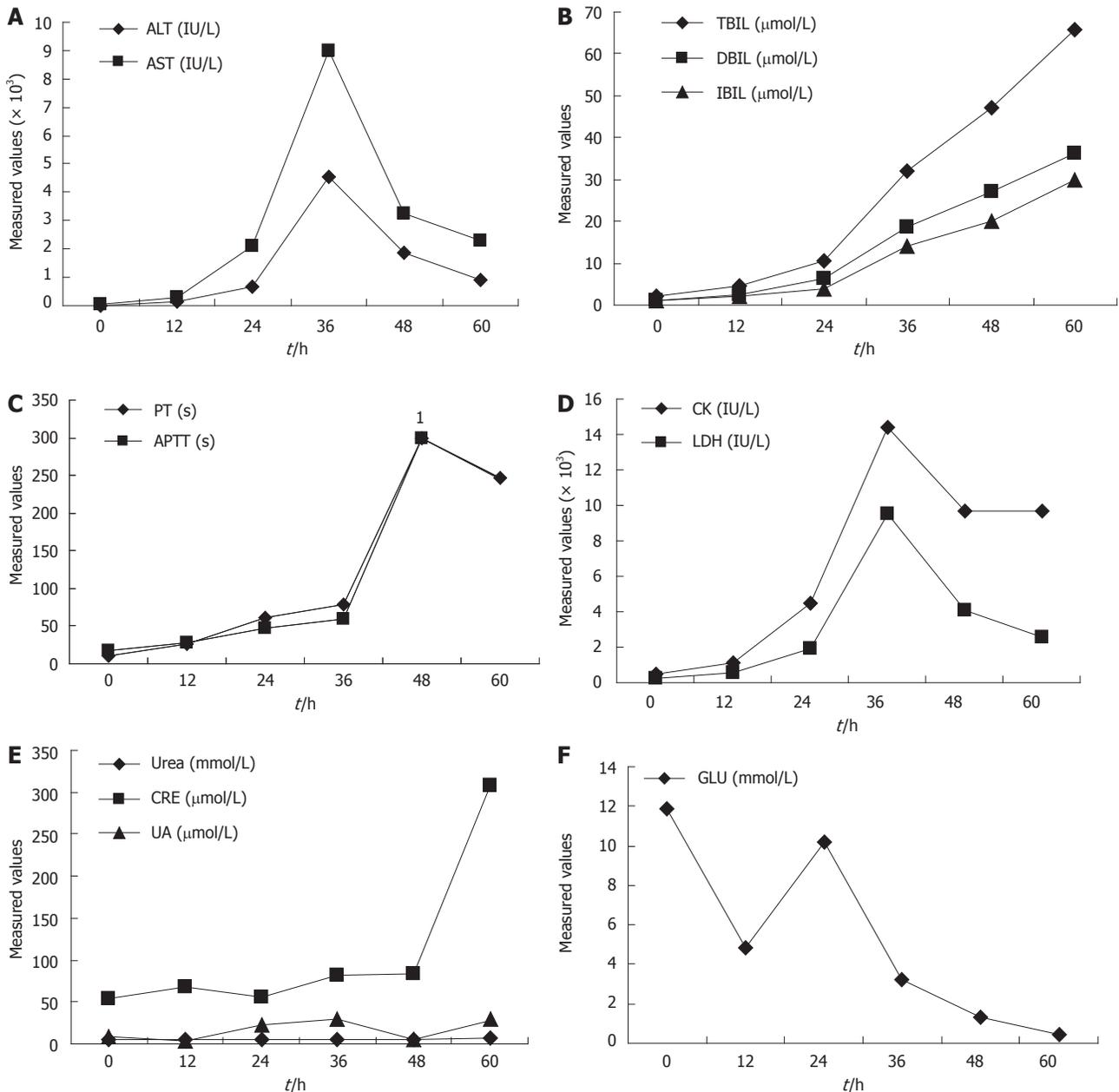


Figure 1 Blood biochemical parameters of the *Macaca mulatta* model of fulminant hepatic failure. The abscissa and ordinate represent measured time and value, respectively. ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; TBIL: Total bilirubin; DBIL: Direct bilirubin; IBIL: Indirect bilirubin; PT: Prothrombin time; APTT: Activated partial thromboplastin time; CK: Creatine kinase; LDH: Lactate dehydrogenase; CRE: Creatinine; UA: Uric acid; GLU: Glucose; FHF: Fulminant hepatic failure. *Represents values > 300 s, which exceeded the range of the equipment, so we did not obtain an exact measurement.

baseline values (Figure 1B). The consistent increase in bilirubin and the notable decrease in enzymes after 36 h suggest the emergence of enzyme-bilirubin segregation. The prothrombin time (PT) and activated partial thromboplastin time (APTT) were also prolonged after administration. At 48 h, both the levels exceeded 300 s so that we could not measure the exact number using the machine. Even the last two measured levels of PT and APTT showed little diminution, and at 250 s were still 14- to 22-fold of the baseline (Figure 1C). The ratios of prothrombin activity were 1.18% and 5.53%.

To determine whether the toxins damaged other organs, the biochemical parameters of the heart, kidneys

and pancreas were measured. Creatine kinase (CK) and lactate dehydrogenase (LDH), markers of myocardial injury, also displayed peaks at 36 h with values 27-fold and 38-fold that of baseline, respectively (Figure 1D). Of the three kidney-associated parameters, i.e., urea, creatinine (CRE) and uric acid, only the level of CRE obviously increased after 48 h (Figure 1E). The blood glucose concentration decreased (Figure 1F), and the serum amylase of the pancreas did not show any significant changes.

Imaging

MRI and ultrasonography were performed every 24 h, and the final scans were performed when the animal

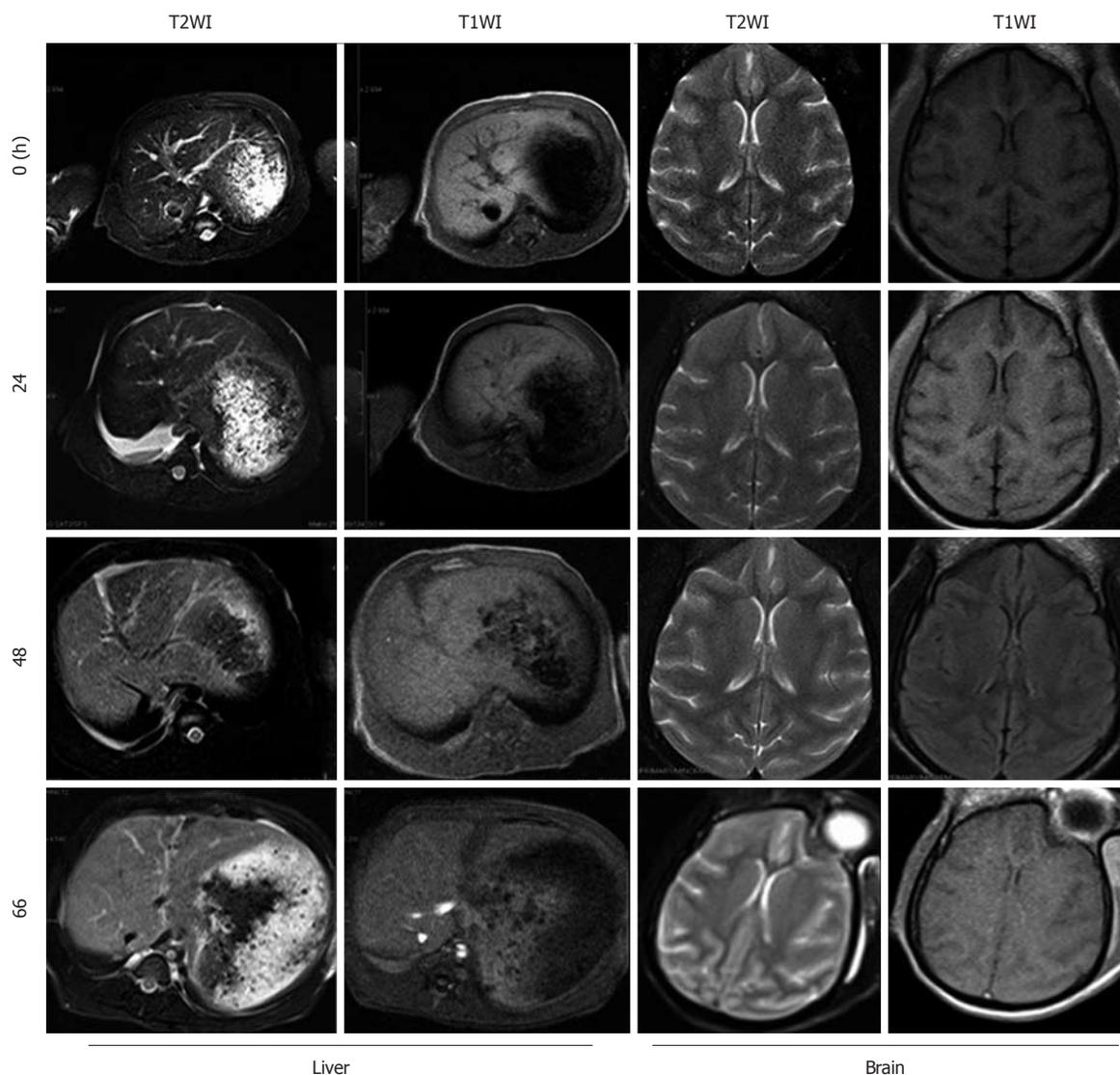


Figure 2 Magnetic resonance imaging of the liver and brain of *Macaca mulatta* model at different times. T2WI: T2-weighted imaging; T1WI: T1-weighted imaging.

died. At 24 h, MRI showed slight signals emerging in the left lobe of the liver, and Gd-enhanced T1-weighted imaging demonstrated homogeneous intensity. The signal began to increase heterogeneously in the parenchyma at 48 h, and became more heterogeneous and diffuse at 66 h, and the intensity decreased on a widespread basis in the hepatobiliary period (Figure 2). In ultrasonography, the echo of the liver parenchyma was slightly enhanced at 48 h, especially in the left lobe, and the contrast sonogram with SonoVue showed low perfusion. The hepatic artery resistance index increased. At 60 h, the echo was still enhanced slightly, but the portal vein velocity decreased and the hepatic artery resistance index increased further (Figure 3). Before 48 h, no predominant changes were found in brain MRI, but at 66 h, abnormal rhythmic signals emerged in the frontal lobe and both temporal lobes, which demonstrated hyperacute ischemia (Figure 2). We also scanned the thoracic and abdominal organs and no marked abnormality was found (data not shown).

Pathological changes

Core needle biopsies were performed at 12 h and 36 h af-

ter administration and the animals underwent necropsies after death. The main organs were evaluated, including the liver, kidneys, lungs, heart, brain, spleen, pancreas and lymph nodes, for macroscopic and histological changes.

H and E and oil red O staining showed that the liver developed severe steatosis at 36 h after toxin infusion, and in the necropsy liver, extensive parenchymal necrosis was found. The hepatic cord was dissociated, with disordered hepatocytes, and the hepatic sinusoid was extended with hyperemia. Vacuoles appeared in the cytoplasm, and patchy necrosis was found in the portal areas. In the non-necrotic areas, most hepatocytes were stained reddish-orange by oil red O. In the necropsy section, massive necrosis caused loss of almost all of the hepatocytes, and only the reticular structure remained, which is similar to the case in humans suffering from acute severe viral hepatitis (Figure 4).

With the naked eye, the livers appeared softer and smaller, with sharper and thinner edges, than the normal liver. The surface color was red and yellow, and it was a uniform yellow at the cut surface (Figure 4). Vascular dilatation and hyperemia were found on the surface of

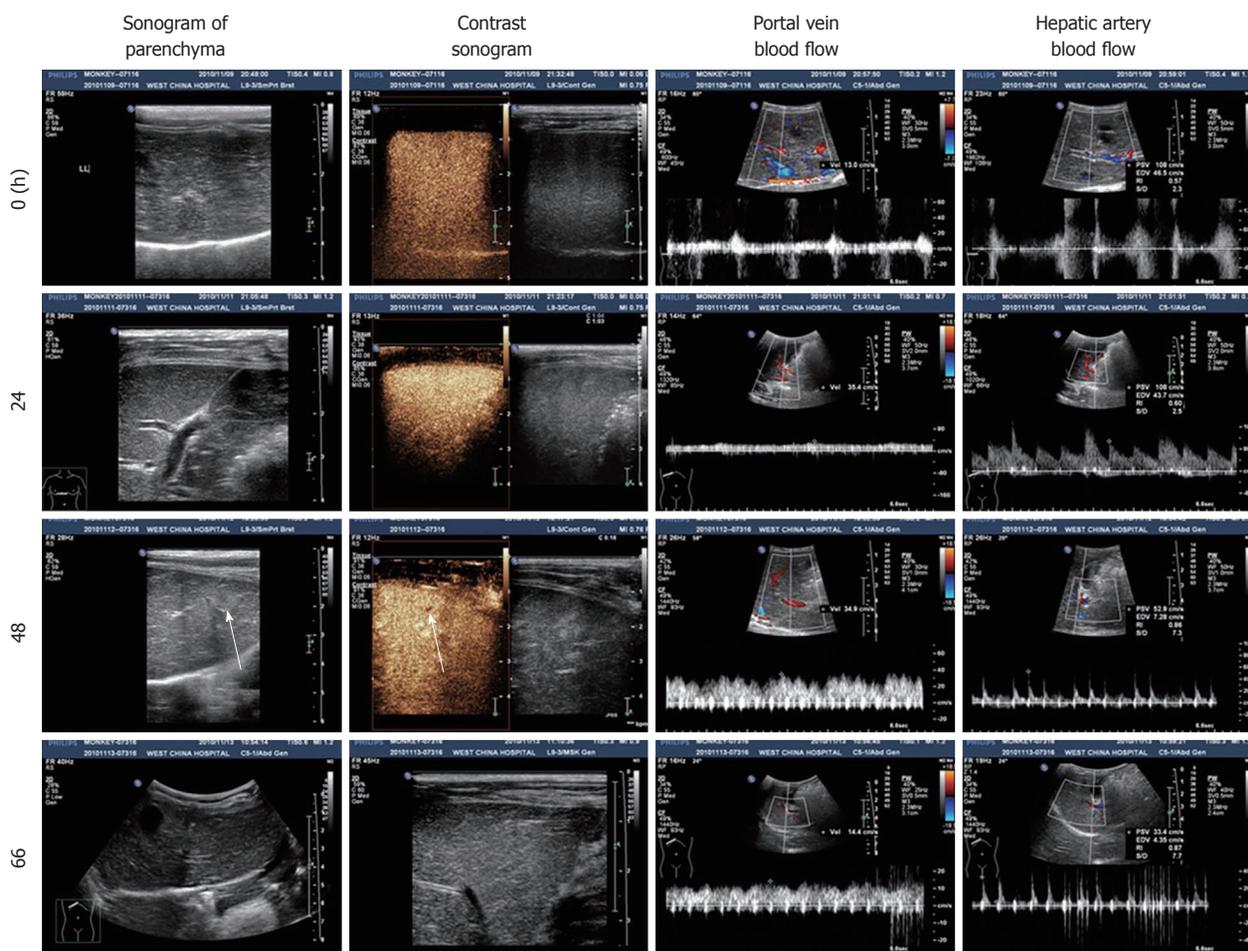


Figure 3 Liver sonogram of the *Macaca mulatta* model at different times. Arrows show a slightly enhanced echo and lower perfusion of SonoVue at 48 h.

the brain and cerebral edema was proved by widened gyri and shallowed sulci (Figure 5). Under microscopy, the superficial layer of the brain was loose, edematous and weakly stained (Figure 6).

Both kidneys showed hyperemia. The heart, spleen and pancreas did not display obvious gross changes (Figure 5). No peritoneal hyperemia, adhesions or edema in the peritoneal cavity was found. The lymph nodes located in the mesentery were enlarged and darkened. Under microscopy, they showed diffuse bleeding and lymphocytes were extensively reduced with much histiocyte proliferation. Arterioles in the lymphoid follicles were found to have hyaline degeneration. Both kidneys were also injured and demonstrated ectasia and hyperemia of the capillaries in the renal glomerulus and the stroma of the kidney tubules, and there was cellular swelling in the tubule epithelial cells. Only a wave-like change of the myocardium was found in the heart (Figure 6).

DISCUSSION

In this study, we have established, for the first time, a *Macaca mulatta* model of FHF, which satisfies all of the criteria for a large animal model of FHF proposed by Terblanche *et al*^[10] and Fourneau *et al*^[11], especially a con-

scious animal model for observation of the development of hepatic encephalopathy. All the clinical features and biochemical parameters, including liver enzymes, bilirubin as well as coagulation activity, confirmed that this model matched all the diagnostic criteria of human FHF. In addition, we dynamically examined and recorded the pathological and imaging changes in the liver and extrahepatic organs during disease progression.

Animal models of FHF are urgently needed to fully investigate the pathogenesis, progression, diagnosis and treatment of this serious disease clinically. Novel therapeutic strategies including hepatocyte transplantation, stem cell transplantation, tissue engineered liver and bioartificial liver support systems are under investigation. However, all the therapies need to be tested in an animal model before clinic use. Over the past 30 years, various animal models of FHF have been created with different methods^[7,12,13], including models using rodents^[15-18], dogs^[19-23] and pigs^[24,25]. However, differences in the metabolic and physiological properties in these distantly relative species restricted the application of the results to humans. As the closest relative to the human, the primate possesses much more similar metabolic and physiological properties, and a primate with FHF is considered to be the best large animal model for use in basic and pre-clin-

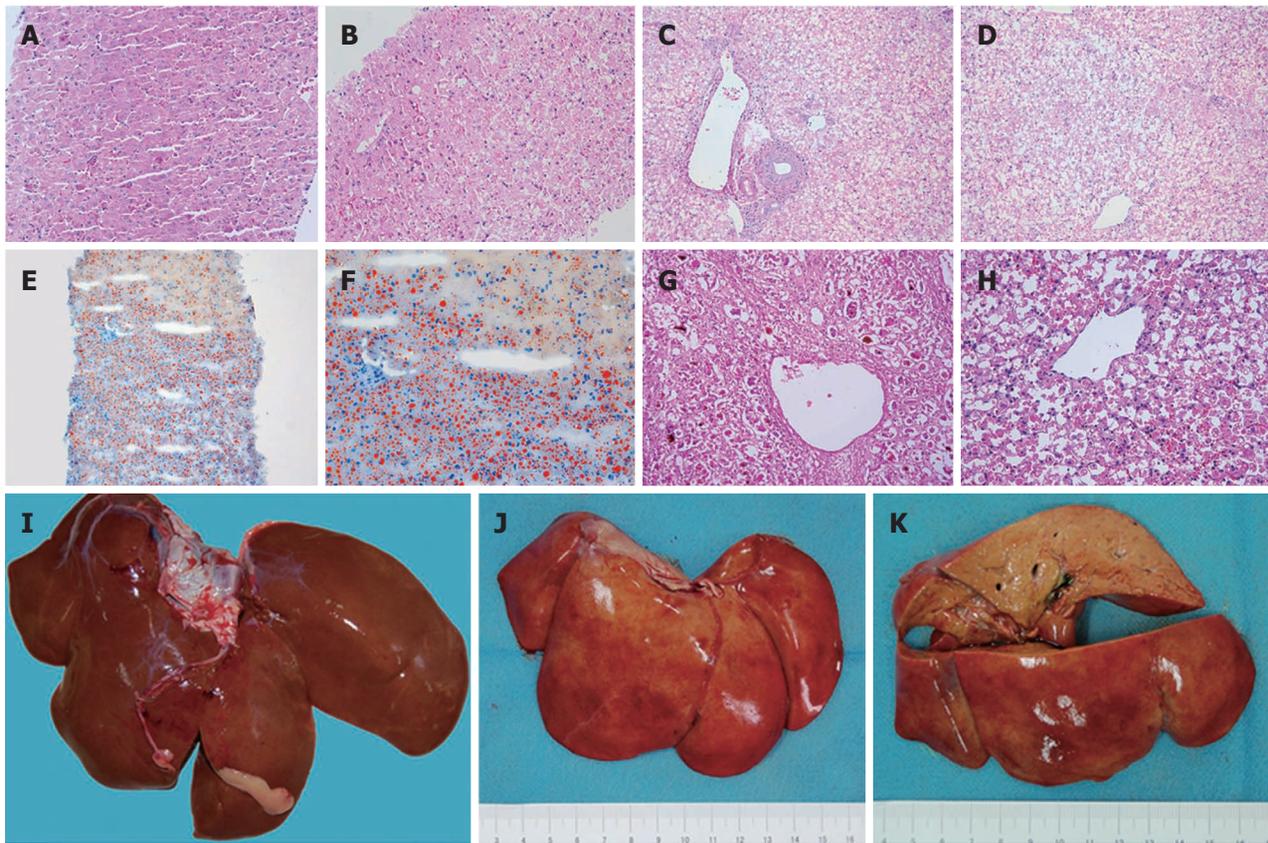


Figure 4 Histopathological changes of the liver. A: Tissue from needle biopsy at 12 h. Changes in organizational structure and cell morphology were not obvious [hematoxylin and eosin (H and E) stain, $\times 200$]; B: Tissue from needle biopsy at 36 h. Vacuoles appeared in the hepatocellular cytoplasm (H and E stain, $\times 200$); C, D: Tissue from necropsy after death at 70 h. Hepatocellular necrosis was distributed in the portal area (C) and central area (D) (H and E stain, $\times 200$). Massive necrosis caused almost all of the hepatocytes to be lost, and only the support structure remained; E: Frozen tissue from needle biopsy at 36 h. The extensive reddish-orange color suggested serious fatty degeneration (oil red O stain, $\times 100$); F: Frozen tissue from needle biopsy at 36 h. In the borderline between necrosis and steatosis, the reddish-orange color was obvious (oil red O stain, $\times 200$); G, H: Comparison of pathological changes of fulminant hepatic failure in humans induced by viral hepatitis (G) and a *Macaca mulatta* model induced by α -amanitin and lipopolysaccharide (H) (H and E stain, $\times 200$); I: Normal liver of *Macaca mulatta* for comparison; J: The surface view of the experimental liver on necropsy appeared red and yellow; K: The cut surface view of the experimental liver was uniformly yellow.

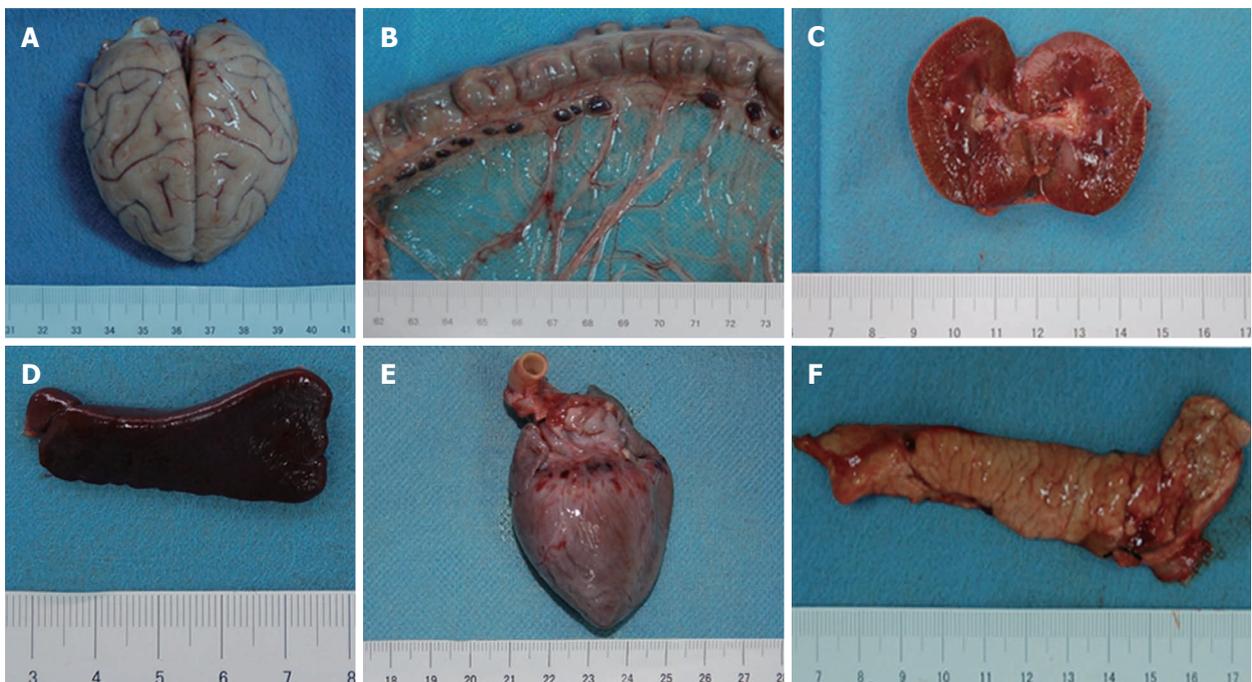


Figure 5 Other main organs on necropsy. A: The brain with cerebral edema; B: Mesenteric lymph nodes enlarged and beaded; Hyperemia of the kidney (C), spleen (D), heart (E), and pancreas (F) without obvious gross lesions.

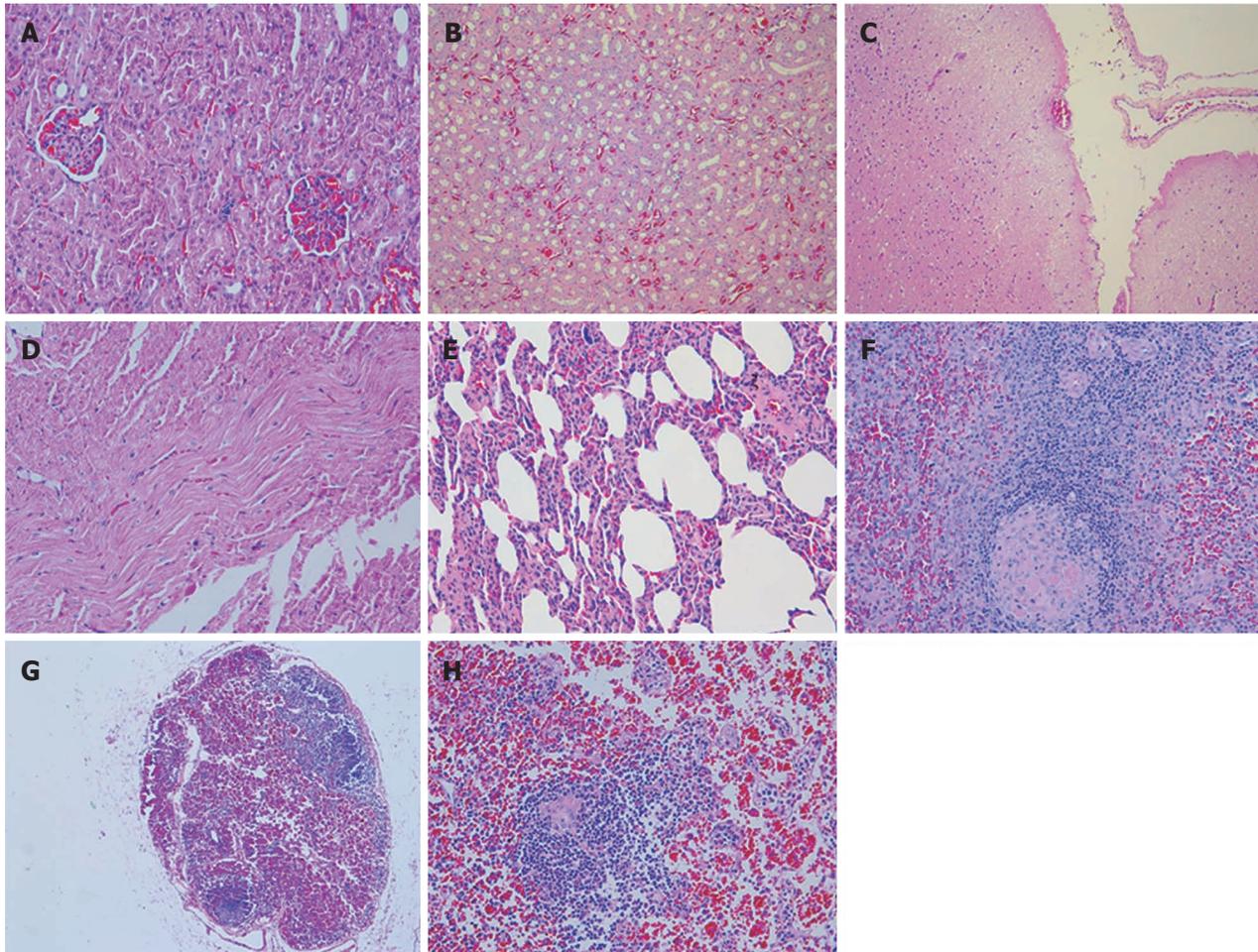


Figure 6 Histopathological changes of other organs. A: Ectasia and hyperemia of capillaries in the renal glomerulus [hematoxylin and eosin (H and E) stain, $\times 200$]; B: Widespread hyperemia of capillaries in the renal stroma and cellular swelling in the tubular epithelial cells (H and E stain, $\times 100$); C: Weak staining in the superficial layer of the brain suggest cerebral edema (H and E stain, $\times 100$); D: Wave-like changes in the myocardium (H and E stain, $\times 200$); E: Hyperemia of capillaries in the stroma and hyaline changes in the arterioles of the lung (H and E stain, $\times 200$); F: Hyaline changes in the central artery of the spleen (H and E stain, $\times 200$); G: Widespread reduction in lymphocytes and hemorrhage in the mesenteric lymph nodes (H and E stain, $\times 50$); H: Hemorrhage and histiocyte proliferation in the mesenteric lymph nodes. Hyaline change appeared in the vessel wall of lymphoid follicles (H and E stain, $\times 200$).

ical research. To the best of our knowledge, our *Macaca mulatta* model of FHF is the first large primate model in this field.

Methods used to induce the FHF models included surgical intervention and hepatotoxin administration. Total or partial hepatectomy and hepatic ischemic injury were widely used, but these relied on extensive surgical expertise and were influenced by individual variations in the animals. Hepatotoxins, which induce massive hepatic necrosis, are widely used in the establishment of a FHF model. Acetaminophen and galactosamine were the two most widely used hepatotoxins^[17,19,22,26-31], and in some studies, carbon tetrachloride^[32], thioacetamide^[33,34], azoxymethane^[35,36], concanavalin A^[37] and amanita phalloides^[38,39] were also employed. However, a lack of standardization, especially a lack of reproducibility, is the major disadvantage of models induced by acetaminophen^[7,10]. The high costs restrict the application of galactosamine in large-scale models^[40]. Amatoxin, a polypeptide extracted from a hypertoxic mushroom, has been used as a hepatotoxin in pigs with an endotoxin (LPS), and this

model was reported to satisfy the criteria of near-universal mortality from highly reproducible hepatic failure, of mortality occurring in a defined time range, of damage specific to the liver, and of potential for recovery of the damaged liver^[38,39]. In our preliminary experiment, LPS (2.25 $\mu\text{g}/\text{kg}$) and α -amanitin (0.225 mg/kg) were injected in a *Macaca mulatta* through a branch of the mesenteric vein by laparotomy at the dose converted from that used in pigs, which received LPS and α -amanitin at a dose of 1 $\mu\text{g}/\text{kg}$ and 0.1 mg/kg , respectively^[38,39]. However, the animal died within 8 h. We supposed that the direct and undiluted toxic effects to the liver might be the main cause of death and the surgery might promote the death of the *Macaca mulatta*. Thus, we reduced the dose of the two hepatotoxins to the level used in pigs, which was nearly half of our preliminary dose. We then improved the administration pathway by slow intraperitoneal infusion of the toxins diluted in 50 mL physiological saline instead of direct portal injection. Our results showed that the animals survived for at least 49 h and all the features of FHF were clearly presented. Most importantly, the

prolonged disease course ensures a wide therapeutic window. It is possible that intraperitoneal infusion of toxins could lead to severe peritonitis. Fortunately, we did not find any symptoms and signs of peritonitis, either in imaging examination or in necropsy.

Induced by α -amanitin and LPS, the hepatocytes presented severe steatosis first and massive necrosis at the end stage of FHF, which is similar to the case in humans suffering from acute severe viral hepatitis. Serum parameters, including hepatic enzymes, bilirubin, and coagulation activities, as well as serum ammonia, are often considered as the markers representing the level of damage of hepatocytes, since a biopsy is seldom carried out because of the restriction of invasive operation and bleeding. In fact, the hepatic injury during the course of FHF still needs direct confirmation. Our model offers an appropriate model to obtain real-time pathological changes in the hepatocytes, which makes it valuable for study of clinical features based on the pathological changes.

We also monitored the continuous alteration of the liver by imaging, and this is the first report on dynamic MRI of the liver during the course of FHF, either in patients or in an animal model. Combining the biopsy and the MRI, it is feasible to establish the diagnostic criteria of the damage level based on the MRI values, thus offering a non-invasive strategy to reveal the level of hepatic injury.

Hepatic encephalopathy is one of the most important clinical features and causes of death of FHF. Being consanguineous with humans, the *Macaca mulatta* presents similar behavior and consciousness to humans suffering from FHF. In addition, the brain volume and the similar gyrus outline make it suitable for MRI examination of the cerebral edema, ischemia or infarction. In our model, the progressing symptoms of listlessness, anorexia, grasping disability, mental indifference, drowsiness and coma are fully developed and thoroughly recorded. The behavior and conscious alterations, combined with the MRI and pathological changes, make it a valuable model for further investigation of the pathogenesis and pathophysiology of hepatic encephalopathy, which could not be reproduced in humans or in any other animals. As a noninvasive means of detection, MRI may be an appropriate strategy to monitor the progression of the hepatic encephalopathy. The first objective assessment of MR brain changes specific to hepatic encephalopathy was made in the early 1990s. It was noted that in patients with cirrhosis, the basal ganglia appeared hyperintense on T1-weighted MRI^[41]. However, continuous data are still absent to reveal the correlation between the degree of encephalopathy and the MRI parameters during the course of FHF. With our model, it is feasible to obtain the dynamic imaging alterations during the whole course of hepatic encephalopathy.

It is necessary to rule out that animal death was induced by extrahepatic organ failure. In our experiment, the main extrahepatic organs were also evaluated. The in-

creasing of CRE happened late, suggesting that the damage to the kidneys was a result of hepatorenal syndrome rather than direct toxin damage. The values of CK and LDH increased immediately after toxin administration, but no marked abnormality was found by echocardiogram and no myocardial necrosis was observed in histopathological examination. The cause of the increased CK and LDH still needs clarification. The lesion of the mesenteric lymph nodes might be associated with the toxins, which needs further study. We did not find any pathological evidences indicating cell injury of the lung and pancreas. Above all, we can conclude that the toxicity of intraperitoneal infusion of amatoxin and endotoxin is liver-specific and animal death is directly correlated with FHF and not extrahepatic organ failure.

We describe, for the first time, a *Macaca mulatta* model of fulminant hepatic failure, which satisfies all of the criteria for a large animal model of FHF^[10,11]. With similar metabolic and physiological properties to humans, our primate animal model of FHF offers a valuable model to be used for investigation of the pathophysiology of FHF and for evaluation of potential medical therapies. Compared with other animals, primate models are more expensive, and the ethics of using such animals should be taken into consideration.

COMMENTS

Background

Fulminant hepatic failure (FHF) is an uncommon and challenging clinical disease characterized by sudden and severe hepatic injury and dysfunction, and it results in many different symptoms and complications with a high mortality. Previous reports have described different animal models of FHF induced by various methods, but all the established models did not entirely satisfy all the criteria.

Research frontiers

Animal models of FHF are urgently needed to fully investigate the pathogenesis, progression, diagnosis and treatment of this serious disease in the clinic. Recently, more studies have focused on large animal models and therapy for FHF. Novel therapeutic strategies including hepatocyte transplantation, stem cell transplantation, tissue engineered liver and BAL support systems are under investigation. Also, various animal models of FHF have been created with different methods, including models using rodents, dogs and pigs. However, differences in metabolic and physiological properties in these distantly relative species restricted the application of the results to humans. No primate animal model with FHF has been created before.

Innovations and breakthroughs

In this article, the authors describe, for the first time, a *Macaca mulatta* model of FHF, which satisfies all of the criteria for the large animal model of FHF. It is important that with the model, it is feasible to obtain dynamic imaging and pathologic alterations, and this is the first report about dynamic MRI of the liver during the course of FHF, either in patients or in animal models. Though lipopolysaccharide and α -amanitin were reported to induce a pig model of FHF, the main extrahepatic organs were not evaluated. In this article, the authors improved the administration pathway and evaluated the main extrahepatic organs.

Applications

The primate animal model of FHF offers a valuable model to be used for investigation of the mechanism of FHF and for evaluation of potential medical therapies.

Terminology

Macaca Mulatta: A species of the genus *MACACA* inhabiting India, China, and other parts of Asia. The species is used extensively in biomedical research and adapts very well to living with humans.

Peer review

The authors show an appropriate large primate model of fulminant hepatic failure. They described the rigid data of hepatic failure by biochemical results, imaging, and pathological changes. The primate animal model of fulminant hepatic failure has clinical benefit because of similar metabolic and physiological properties to human. This paper is an interesting and instructive manuscript. It is well written.

REFERENCES

- Bernal W**, Auzinger G, Dhawan A, Wendon J. Acute liver failure. *Lancet* 2010; **376**: 190-201
- Fontana RJ**. Acute liver failure including acetaminophen overdose. *Med Clin North Am* 2008; **92**: 761-794
- Adam R**, Cailliez V, Majno P, Karam V, McMaster P, Caine RY, O'Grady J, Pichlmayr R, Neuhaus P, Otte JB, Hoeckerstedt K, Bismuth H. Normalised intrinsic mortality risk in liver transplantation: European Liver Transplant Registry study. *Lancet* 2000; **356**: 621-627
- Lidofsky SD**, Bass NM, Prager MC, Washington DE, Read AE, Wright TL, Ascher NL, Roberts JP, Scharschmidt BF, Lake JR. Intracranial pressure monitoring and liver transplantation for fulminant hepatic failure. *Hepatology* 1992; **16**: 1-7
- de Rave S**, Tilanus HW, van der Linden J, de Man RA, van der Berg B, Hop WC, Ijzermans JN, Zondervan PE, Metseelaar HJ. The importance of orthotopic liver transplantation in acute hepatic failure. *Transpl Int* 2002; **15**: 29-33
- Wall WJ**, Adams PC. Liver transplantation for fulminant hepatic failure: North American experience. *Liver Transpl Surg* 1995; **1**: 178-182
- Newsome PN**, Plevris JN, Nelson LJ, Hayes PC. Animal models of fulminant hepatic failure: a critical evaluation. *Liver Transpl* 2000; **6**: 21-31
- Nussler A**, Konig S, Ott M, Sokal E, Christ B, Thasler W, Brulport M, Gabelein G, Schormann W, Schulze M, Ellis E, Kraemer M, Nocken F, Fleig W, Manns M, Strom SC, Hengstler JG. Present status and perspectives of cell-based therapies for liver diseases. *J Hepatol* 2006; **45**: 144-159
- Stravitz RT**. Critical management decisions in patients with acute liver failure. *Chest* 2008; **134**: 1092-1102
- Terblanche J**, Hickman R. Animal models of fulminant hepatic failure. *Dig Dis Sci* 1991; **36**: 770-774
- Fourneau I**. A model to test the efficiency of a bioartificial liver. In: A potential reversible model of acute liver failure in the pig. Belgium: Leuven University Press, 2001; 41-56
- van de Kerckhove MP**, Hoekstra R, van Gulik TM, Chamuleau RA. Large animal models of fulminant hepatic failure in artificial and bioartificial liver support research. *Biomaterials* 2004; **25**: 1613-1625
- Tuñón MJ**, Alvarez M, Culebras JM, González-Gallego J. An overview of animal models for investigating the pathogenesis and therapeutic strategies in acute hepatic failure. *World J Gastroenterol* 2009; **15**: 3086-3098
- Ferenci P**, Lockwood A, Mullen K, Tarter R, Weissenborn K, Blei AT. Hepatic encephalopathy—definition, nomenclature, diagnosis, and quantification: final report of the working party at the 11th World Congresses of Gastroenterology, Vienna, 1998. *Hepatology* 2002; **35**: 716-721
- Walker RM**, Racz WJ, McElligott TF. Acetaminophen-induced hepatotoxicity in mice. *Lab Invest* 1980; **42**: 181-189
- Olafsson S**, Gottstein J, Blei AT. Brain edema and intracranial hypertension in rats after total hepatectomy. *Gastroenterology* 1995; **108**: 1097-1103
- Blitzer BL**, Waggoner JG, Jones EA, Gralnick HR, Towne D, Butler J, Weise V, Kopin IJ, Walters I, Teychenne PF, Goodman DG, Berk PD. A model of fulminant hepatic failure in the rabbit. *Gastroenterology* 1978; **74**: 664-671
- Horowitz ME**, Schafer DF, Molnar P, Jones EA, Blasberg RG, Patlak CS, Waggoner J, Fenstermacher JD. Increased blood-brain transfer in a rabbit model of acute liver failure. *Gastroenterology* 1983; **84**: 1003-1011
- Francavilla A**, Makowka L, Polimeno L, Barone M, Demetris J, Prelich J, Van Thiel DH, Starzl TE. A dog model for acetaminophen-induced fulminant hepatic failure. *Gastroenterology* 1989; **96**: 470-478
- Kelly JH**, Koussayer T, He DE, Chong MG, Shang TA, Whisennand HH, Sussman NL. An improved model of acetaminophen-induced fulminant hepatic failure in dogs. *Hepatology* 1992; **15**: 329-335
- Rozga J**, Williams F, Ro MS, Neuzil DF, Giorgio TD, Backfisch G, Moscioni AD, Hakim R, Demetriou AA. Development of a bioartificial liver: properties and function of a hollow-fiber module inoculated with liver cells. *Hepatology* 1993; **17**: 258-265
- Sielaff TD**, Hu MY, Rollins MD, Bloomer JR, Amiot B, Hu WS, Cerra FB. An anesthetized model of lethal canine galactosamine fulminant hepatic failure. *Hepatology* 1995; **21**: 796-804
- Diaz-Buxo JA**, Blumenthal S, Hayes D, Gores P, Gordon B. Galactosamine-induced fulminant hepatic necrosis in unanesthetized canines. *Hepatology* 1997; **25**: 950-957
- Miller DJ**, Hickman R, Fratter R, Terblanche J, Saunders SJ. An animal model of fulminant hepatic failure: a feasibility study. *Gastroenterology* 1976; **71**: 109-113
- Hanid MA**, Mackenzie RL, Jenner RE, Chase RA, Mellon PJ, Trewby PN, Janota I, Davis M, Silk DB, Williams R. Intracranial pressure in pigs with surgically induced acute liver failure. *Gastroenterology* 1979; **76**: 123-131
- Sussman NL**, Chong MG, Koussayer T, He DE, Shang TA, Whisennand HH, Kelly JH. Reversal of fulminant hepatic failure using an extracorporeal liver assist device. *Hepatology* 1992; **16**: 60-65
- Gardner CR**, Heck DE, Yang CS, Thomas PE, Zhang XJ, DeGeorge GL, Laskin JD, Laskin DL. Role of nitric oxide in acetaminophen-induced hepatotoxicity in the rat. *Hepatology* 1998; **27**: 748-754
- Gunawan BK**, Liu ZX, Han D, Hanawa N, Gaarde WA, Kaplowitz N. c-Jun N-terminal kinase plays a major role in murine acetaminophen hepatotoxicity. *Gastroenterology* 2006; **131**: 165-178
- Nakagawa H**, Maeda S, Hikiba Y, Ohmae T, Shibata W, Yanai A, Sakamoto K, Ogura K, Noguchi T, Karin M, Ichijo H, Omata M. Deletion of apoptosis signal-regulating kinase 1 attenuates acetaminophen-induced liver injury by inhibiting c-Jun N-terminal kinase activation. *Gastroenterology* 2008; **135**: 1311-1321
- Namisaki T**, Yoshiji H, Kojima H, Yoshiji J, Ikenaka Y, Noguchi R, Sakurai S, Yanase K, Kitade M, Yamazaki M, Asada K, Uemura M, Nakamura M, Fukui H. Salvage effect of the vascular endothelial growth factor on chemically induced acute severe liver injury in rats. *J Hepatol* 2006; **44**: 568-575
- Ma KF**, Yang HY, Chen Z, Qi LY, Zhu DY, Lou YJ. Enhanced expressions and activations of leukotriene C4 synthesis enzymes in D-galactosamine/lipopolysaccharide-induced rat fulminant hepatic failure model. *World J Gastroenterol* 2008; **14**: 2748-2756
- Brattin WJ**, Glende EA, Recknagel RO. Pathological mechanisms in carbon tetrachloride hepatotoxicity. *J Free Radic Biol Med* 1985; **1**: 27-38
- Chieli E**, Malvaldi G. Role of the microsomal FAD-containing monooxygenase in the liver toxicity of thioacetamide S-oxide. *Toxicology* 1984; **31**: 41-52
- Pallottini V**, Martini C, Bassi AM, Romano P, Nanni G, Trentalancia A. Rat HMGCoA reductase activation in thioacetamide-induced liver injury is related to an increased reactive oxygen species content. *J Hepatol* 2006; **44**: 368-374

- 35 **Matkowskyj KA**, Marrero JA, Carroll RE, Danilkovich AV, Green RM, Benya RV. Azoxymethane-induced fulminant hepatic failure in C57BL/6J mice: characterization of a new animal model. *Am J Physiol* 1999; **277**: G455-G462
- 36 **Bélanger M**, Côté J, Butterworth RF. Neurobiological characterization of an azoxymethane mouse model of acute liver failure. *Neurochem Int* 2006; **48**: 434-440
- 37 **Toritsu T**, Nakaya M, Watanabe S, Hashimoto M, Yoshida H, Chinen T, Yoshida R, Okamoto F, Hanada T, Torisu K, Takaesu G, Kobayashi T, Yasukawa H, Yoshimura A. Suppressor of cytokine signaling 1 protects mice against concanavalin A-induced hepatitis by inhibiting apoptosis. *Hepatology* 2008; **47**: 1644-1654
- 38 **Takada Y**, Ishiguro S, Fukunaga K, Gu M, Taniguchi H, Seino KI, Yuzawa K, Otsuka M, Todoroki T, Fukao K. Increased intracranial pressure in a porcine model of fulminant hepatic failure using amatoxin and endotoxin. *J Hepatol* 2001; **34**: 825-831
- 39 **Takada Y**, Ishiguro S, Fukunaga K. Large-animal models of fulminant hepatic failure. *J Artif Organs* 2003; **6**: 9-13
- 40 **Rahman TM**, Hodgson HJ. Animal models of acute hepatic failure. *Int J Exp Pathol* 2000; **81**: 145-157
- 41 **Zeneroli ML**, Cioni G, Crisi G, Vezzelli C, Ventura E. Globus pallidus alterations and brain atrophy in liver cirrhosis patients with encephalopathy: an MR imaging study. *Magn Reson Imaging* 1991; **9**: 295-302

S- Editor Sun H L- Editor Cant MR E- Editor Zhang DN

Chronic fatigue is associated with increased disease-related worries and concerns in inflammatory bowel disease

Lars-Petter Jelsness-Jørgensen, Tømm Bernklev, Magne Henriksen, Roald Torp, Bjørn Moum

Lars-Petter Jelsness-Jørgensen, Department of Gastroenterology, Oslo University Hospital Aker and Østfold Hospital Trust, 1603 Fredrikstad, Norway

Tømm Bernklev, Department of Research, Telemark Hospital Trust, 3710 Skien, Norway

Magne Henriksen, Department of Gastroenterology, Østfold Hospital Trust, 1603 Fredrikstad, Norway

Roald Torp, Department of Gastroenterology, Innlandet Hospital Trust, 2380 Brumundal, Norway

Bjørn Moum, Department of Gastroenterology, Oslo University Hospital and Institute of Clinical Medicine, University of Oslo, 0586 Oslo, Norway

Author contributions: Jelsness-Jørgensen LP gathered the data, analysed it and wrote the manuscript; Henriksen M and Torp R contributed to the data gathering and analysis, and checked the final version of the manuscript for intellectual content; Bernklev T contributed to the design of the study, the analysis and the manuscript preparation; Moum B contributed to the design of the study, gathering data, the analysis and the manuscript preparation.

Supported by Oslo University Hospital Aker; Østfold Hospital Trust; and Ferring Pharmaceuticals

Correspondence to: Dr. Lars-Petter Jelsness-Jørgensen, Department of Gastroenterology, Østfold Hospital Trust, Cicignongt 19, N-1603, 1603 Fredrikstad,

Norway. l.p.jelsness-jorgensen@medisin.uio.no

Telephone: +47-69-303024 Fax: +47-69-861173

Received: February 5, 2011 Revised: June 16, 2011

Accepted: June 23, 2011

Published online: February 7, 2012

Abstract

AIM: To investigate the impact of chronic fatigue on disease-related worries in inflammatory bowel disease (IBD) and the potential multicollinearity between subjective questionnaires.

METHODS: Patients in remission or with mild-to-moderate disease activity completed the fatigue questionnaire (FQ), the rating form of IBD patient concerns (RFIPC), the Short-Form 36 (SF-36), and IBD question-

naire (N-IBDQ). In addition, clinical and epidemiological data were obtained.

RESULTS: In total, 140 patients were included; of which 92 were diagnosed with ulcerative colitis and 48 with Crohn's disease. The mean age of patients with chronic fatigue was 44.2 years (SD = 15.8) and for non-fatigued patients was 44.7 years (SD = 16.0). Chronic fatigued patients had clinically significantly increased levels of disease-related worries, as measured by Cohen's *d* effect size. Worries about having an ostomy bag, loss of bowel control, and energy levels were most prominent in both chronic fatigued and non-chronic fatigued IBD patients. Variance inflation factor (VIF) and tolerance indicated that there were no problematic multicollinearity among the FQ, RFIPC, SF-36 and N-IBDQ responses (VIF < 5 and tolerance > 2).

CONCLUSION: Chronic fatigue is associated with increased levels of disease-related worries and concerns in IBD. Increased levels of worries were also associated with impaired health-related quality of life.

© 2012 Baishideng. All rights reserved.

Key words: Fatigue; Worries; Health-related quality of life; Patient reported outcome; Inflammatory bowel disease

Peer reviewer: Wojciech Blonski, MD, PhD, University of Pennsylvania, GI Research-Ground Centrex, 3400 Spruce St, Philadelphia, PA 19104, United States

Jelsness-Jørgensen LP, Bernklev T, Henriksen M, Torp R, Moum B. Chronic fatigue is associated with increased disease-related worries and concerns in inflammatory bowel disease. *World J Gastroenterol* 2012; 18(5): 445-452 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/445.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.445>

INTRODUCTION

Inflammatory bowel disease (IBD) includes ulcerative colitis (UC) and Crohn's disease (CD). Both UC and CD are chronic, recurrent diseases of the gastrointestinal tract, with symptoms that include abdominal pain, frequent bowel movements, and rectal bleeding^[1,2]. Traditionally, these diseases have been defined as abnormalities in the structure and function of both organs and tissues^[3]. However, given that IBD is associated with impaired health-related quality of life (HRQoL), increased fatigue, and increased worrying regarding the potential consequences of the disease^[4-7], this definition seems too restrictive. The concept of illness might therefore be broader, embodying the subjective experiences of reduced health and bodily function^[3].

Living with chronic illness can be challenging, and individual responses to these challenges vary widely^[8]. Several HRQoL studies have found that many patients with IBD tend to do well^[9]. However, there are subgroups of patients - such as patients with more severe disease or anxiety - who seem to be particularly affected^[4,5,9]. It was recently reported that IBD patients with chronic fatigue (CF) (defined as an elevated fatigue level of a duration longer than six months) had clinically significant reductions in HRQoL, compared to patients without CF^[10]. Thus, an important aspect of measuring subjective health is to identify patients who are most severely affected.

Both generic (i.e., irrespective of illness/condition) and disease-specific HRQoL questionnaires have been developed^[11-13]. In addition, questionnaires such as the rating form of IBD patient concerns (RFIPC) and the fatigue questionnaire (FQ) are used to quantify other aspects of living with chronic illness^[7,14]. We hypothesised that measuring aspects such as worries/concerns (RFIPC) and fatigue (FQ) might help us to identify subgroups of patients that tend to have a poorer outcome. Potentially, however, there might be areas of overlap between the various questionnaires, which is often referred to as multicollinearity. Multicollinearity refers to a situation wherein predictor variables in a regression model are strongly correlated and, consequently, that variables included in the model are closely related.

The primary aim of this study was to examine the impact of chronic fatigue on disease-related worries in IBD. In addition, we wanted to investigate whether the potential association between worries, fatigue and HRQoL in IBD exhibit multicollinearity.

MATERIALS AND METHODS

Subjects

Patients, who were over the age of 18 years, had IBD that was previously verified clinically, endoscopically, or histologically, and who were either in remission or with mild-to-moderate disease activity [defined as Simple Clinical Colitis Activity Index (SCCAI)^[15] or Simple Crohn's Disease Activity Index (SCDAI)^[16] score of less than 10], were eligible for inclusion in this study. Patients

were excluded if they had cognitive impairment, were deemed unlikely to comply with the study procedures, or if they participated in another study. Participants were consecutively recruited from three outpatient clinics in South-Eastern Norway (the counties of Østfold and Hedmark) during routine follow-up visits. At each of the centre, a senior gastroenterologist was in charge of the study protocol. The inclusion period was from August 23, 2005 to January 29, 2007.

Clinical and sociodemographic data

Sociodemographic variables were gathered by interview, and data regarding clinical status and symptoms were obtained from laboratory tests, medical records and disease activity indices (SCCAI/SCDAI)^[15,16]. In addition, we asked the patients to complete a symptom-based questionnaire that graded their self-perceived IBD symptoms during the previous 14 d, using the following categories; no symptoms, mild symptoms (did not interfere with everyday activities), moderate symptoms (interfered with everyday activities and may have resulted in sick leave), and severe symptoms (unable to perform everyday activities, on sick leave, or hospitalized)^[5].

Each patient's phenotype was classified according to the Vienna Classification for CD patients, as the Montreal Classification did not exist when the study protocol was designed. The UC patients were classified into three subgroups: proctitis, left-sided colitis (with inflammation up to the splenic flexure), and extensive colitis (with inflammation beyond the splenic flexure).

The information regarding fatigue was collected with the FQ^[14], the generic HRQoL with the Short-Form 36 (SF-36)^[11], the disease specific HRQoL with the Norwegian version of the IBD questionnaire (N-IBDQ)^[17], and disease-related worries and concerns with the RFIPC^[7] (Table 1). The questionnaires were self-administered by the patients at the various centres, following a standardized procedure, which allowed the patients to fill out the questionnaires in the peace and quiet of the hospital's outpatient clinic.

Questionnaires

The RFIPC: The RFIPC is a disease-specific questionnaire that was developed by Drossman *et al.*^[7]. This questionnaire rates various important worries and concerns that are raised by IBD patients. The questionnaire consists of the 25 most frequently reported concerns reported by IBD patients, with every item framed in the same style: "Because of your condition, how concerned are you with...?" The responses were scored on a 100-mm horizontal visual analog scale. A score of 0-mm represents no worries/concerns, and a score of 100-mm represents the highest possible worries and concerns. The mean scores of all 25 items yields the "sum score". The RFIPC has been translated into Norwegian and validated (Jelsness-Jørgensen LP, Moum B, Bernklev T. Worries and concerns among inflammatory bowel disease patients followed prospectively over one year. Submitted:

Table 1 Main characteristics of the questionnaires used in this study

Questionnaire	Number of items and dimensions	Main characteristics
SF-36	36 items, divided into 8 dimensions	Measure generic HRQoL
Ware <i>et al</i> ^[11]		Scale scores from 0-100, with higher scores indicating better HRQoL
N-IBDQ	32 items, divided into 5 dimensions	Measure disease-specific HRQoL
Bernlev <i>et al</i> ^[17]		Scale scores from 32-224, with higher scores indicating better HRQoL
FQ	11 items, divided into 2 dimensions	Measure both physical and mental fatigue
Chalder <i>et al</i> ^[14]	The FQ contains 2 items asking about duration and extent of fatigue symptoms	Scale scores from 0-33 with higher scores indicating higher levels of fatigue
Jelsness-Jørgensen <i>et al</i> ^[10]		In addition scored as a dichotomized scale where original scores 0 + 1 = 0 and 2 + 3 = 1
		Chronic fatigue defined as a score of ≥ 4 on the dichotomized scale and duration of fatigue symptoms ≥ 6 mo
RFIPC	25 items, divided into 6 dimensions	Measure the 25 most frequently reported disease-related worries/concerns by IBD patients
Drossman <i>et al</i> ^[7]		

IBD: Inflammatory bowel disease; SF-36: Short-Form 36; N-IBDQ: IBD questionnaire (Norwegian version); FQ: Fatigue questionnaire; RFIPC: Rating form of IBD patient concerns; HRQoL: Health-related quality of life.

Gastroenterol Res Pract).

Factor analysis of the Norwegian version of the RFIPC yielded six factors. F1: Impact of disease (e.g., financial difficulties/energy/loss of bowel control); F2: Expectancy (e.g., developing cancer/dying early); F3: Treatment (e.g., effects of medication); F4: Intimacy (e.g., ability to perform sexually); F5: Stigma (e.g., feeling dirty/smelly); and F6: Complications (e.g., undergoing surgery/ostomy bag placement).

FQ: The FQ was developed by Chalder *et al*^[14] and consists of 11 items that are divided into two main dimensions: physical fatigue (PF), which contains seven items, and mental fatigue (MF), which contains four items. The available responses included four options: 0 = better than usual, 1 = no more than usual, 2 = worse than usual, and 3 = much worse than usual. A higher score reflects a higher level of fatigue. Combining the scores of PF and MF yields the total fatigue score, with a maximum possible scale score of 33. The scale scores of the FQ were also scored on a dichotomized scale (0 = better than usual and no more than usual; 1 = worse than usual and much worse than usual). In addition to measuring episodic fatigue (irrespective of the duration of symptoms), the FQ contains two questions regarding the duration and extent of fatigue symptoms. Based on the results of the original validation study, the Norwegian validation study, and general consensus^[14,18,19], CF was defined as dichotomized scores ≥ 4 and duration > 6 mo. The FQ was recently validated for use in IBD^[6].

SF-36: The SF-36 is a generic, self-administered questionnaire containing 36 questions^[11] that are divided into eight multi-item scales consisting of: physical functioning, role limitations due to physical problems, bodily pain, general health, vitality, social functioning, role limitations due to emotional problems, and mental health (MH). For each question, the raw score was coded and transformed into a scale from 0 to 100, with 0 and 100 representing the lowest level and highest level of function, respectively. The SF-36 has been validated by others^[20]. Missing

data were treated following published recommendations: if data for half of the items within a scale or fewer were missing, they were replaced by the mean value of the respondent's completed items in the same scale in accordance with the SF-36 scoring algorithms^[21].

N-IBDQ: The IBDQ is a disease-specific questionnaire that was developed by Irvine^[12,13]. The original version consists of 32 items divided into four dimensions: bowel symptoms (e.g., loose stools or abdominal pain), systemic symptoms (e.g., fatigue or altered sleep patterns), social function (e.g., work attendance) and emotional function (e.g., anger or depression). The Norwegian validation study (N-IBDQ) revealed a five-dimensional structure: emotional function-1 (fatigue, energy), bowel function-1 (stool consistency and pattern), bowel function-2 (bowel pain and discomfort), social function (work attendance, cancelling social events) and emotional function-2 (worries)^[17]. All of the responses were scored on a 7-point Likert Scale, with a score of 7 representing no problems and a score of 1 representing severe problems. This gives a possible score range of 32-224, with a higher score reflecting improved HRQoL^[12,13,17].

Statistical analysis

To assess the characteristics of the patients, we used descriptive analyses and frequencies. A student's *t* test was used to evaluate the differences in the distribution of epidemiological and clinical variables between the diagnostic groups.

To test potential associations between the RFIPC, FQ, N-IBDQ and SF-36 questionnaires, both bivariate correlation analysis with spearman's ρ and linear regression analysis were used. In addition, the latter analysis was used to explore potential multicollinearity. A variance inflation factor (VIF) greater than 5 and a reciprocal tolerance value of less than 0.20 were defined as indicative of colinearity in accordance with published recommendations^[22,23]. We chose to analyse the RFIPC sum score, the FQ and N-IBDQ total score as dependents only, to reduce the number of dimensions tested.

Table 2 Main clinical/sociodemographic characteristics *n* (%)

	UC (<i>n</i> = 92)	CD (<i>n</i> = 48)
Age, (yr), mean ± SD	46.9 ± 5.8	40.0 ± 15.0
Age range, (yr)	20-82	19-69
Gender		
Female	43 (47)	36 (75)
Male	49 (53)	12 (25)
Disease duration, (yr), mean ± SD	8.5 ± 9.5	9.2 ± 9.6
Educational level		
Second level, first stage (lower)	16 (17.4)	5 (10.4)
Second level, second stage (medium)	42 (45.7)	22 (45.8)
Third level (university)	34 (37.0)	21 (43.8)
UC extension		
Proctitis	27 (29.3)	
Left-sided colitis	23 (25.0)	
Extensive colitis	42 (45.7)	
CD extension		
L1 terminal ileum		11 (22.9)
L2 colon		17 (35.4)
L3 ileocolon		18 (37.6)
L4 upper GI		2 (4.1)
Perceived IBD symptom score		
No symptoms	23 (25.0)	9 (18.8)
Mild symptoms	41 (44.6)	22 (45.8)
Moderate symptoms	25 (27.2)	13 (27.1)
Severe symptoms	3 (3.2)	4 (8.3)

UC: Ulcerative colitis; CD: Crohn's disease; IBD: Inflammatory bowel disease; GI: Gastrointestinal.

To control for and eliminate the effect of potential confounding factors that are known to influence RFIPC scores, age, gender, and educational level were entered as covariates through univariate analysis of variance. Perceived IBD symptoms were hypothesized to affect the RFIPC scores and we therefore additionally chose to correct for this factor as well. The effect size was calculated with Cohen's d^{24} . Operational definitions of 0.2, 0.5 and 0.8 were categorized as small, medium and large effect sizes, respectively^[24].

All tests were 2-sided with a 5% significance level. All statistics were performed using the Predictive Analytics Software, PASW, version 18.0 (IBM Corporation, Route 100 Somers, NY 10589).

Ethical considerations

This study was performed in accordance with the principles of the Helsinki declaration and approval was obtained from the Regional Ethics committee and the Norwegian Data Inspectorate.

RESULTS

One hundred and forty-four patients who were diagnosed with either UC or CD gave their written informed consent for participation in the study. One patient was excluded due to severe disease activity at inclusion (SC-DAI > 10), one patient withdrew from the study after a few weeks, and two patients were excluded from analysis due to incomplete responses to the questionnaires. A to-

tal of 140 patients provided complete data sets and were suitable for statistical analysis.

Epidemiological and clinical characteristics

The CD patients were significantly younger than the UC patients ($P = 0.014$). There were no significant differences in the perceived IBD symptoms or the duration of disease between the two diagnostic groups. Twenty of the 92 (22%) UC patients and 14 of the 48 (29%) CD patients were judged to have CF. There were no significant differences in age, gender, disease duration or educational level between CF and non-CF patients. However, the perceived IBD symptoms were more severe in CF than in non-CF patients ($P < 0.01$). The primary characteristics of the participants are presented in Table 2.

CF and RFIPC scores

CF was associated with significantly higher RFIPC scores in one of the six dimensions in CD and in five of the six dimensions in UC. In addition, the RFIPC sum score was significantly higher in UC patients with CF compared to those without CF (Table 2). Among the individual RFIPC items, the differences between UC patients with and without CF were most pronounced in worries regarding loss of bowel control, developing cancer, dying early, energy level, being a burden to others, having surgery, or an ostomy bag ($P < 0.01$ for all items). In addition, the scores for the following RFIPC items were significantly higher in CD patients with CF compared to those without CF: ability to achieve full potential, financial difficulties, and energy level ($P < 0.05$ for all items). When comparing RFIPC dimensional scores for UC patients with CF to CD patients with CF, the analysis revealed no statistical difference. Only one of the twenty-five individual RFIPC items (worries about pain and suffering) was significantly different between UC and CD patients with CF, with CD patients reporting more worries in this item ($P < 0.01$).

Effect size (Cohen's d) is a measure of the estimated magnitude of a relationship between two variables and is calculated by subtracting the mean dimensional RFIPC scores of CF patients from non-CF patients, then dividing by the common σ of both groups. The analysis revealed that the RFIPC factors that were statistically significant in CF *vs* non-CF patients - regardless of their IBD diagnosis - also produced a large Cohen's d ($d > 0.80$) (Table 3). The analysis further revealed that although they did not reach statistical significance, the numerical differences produced small-to- medium effect sizes. Among the CD group, the differences between CF and non-CF patients resulted in a medium effect size ($d > 0.50$) in the RFIPC sum score, whereas the effect size in the remaining five out of six dimensions were small ($d > 0.20$). Among the UC group, one out of the six RFIPC factors did not differ significantly between CF and non-CF patients, producing a small Cohen's d ($d > 0.20$).

After the mean dimensional RFIPC scores (Table 4)

Table 3 Cohen's *d* effect size calculated from crude dimensional rating form of inflammatory bowel disease patient concerns scores and presence of chronic fatigue

RFIPC factors	CD no CF (n = 34)	CD CF (n = 14)	P value	Cohen's <i>d</i>	UC no CF (n = 72)	UC CF (n = 20)	P value	Cohen's <i>d</i>
Impact of disease	31.1 (20.6)	46.7 (17.7)	0.017	-0.81	25.6 (19.2)	45.7 (18.3)	< 0.001	-1.07
Expectancy	38.1 (30.5)	51.1 (23.1)	0.160	-0.48	32.6 (23.7)	55.3 (20.8)	< 0.001	-1.02
Treatment	27.5 (21.2)	38.0 (25.4)	0.146	-0.44	27.5 (22.0)	49.1 (22.5)	< 0.001	-0.97
Intimacy	20.3 (21.6)	28.3 (28.5)	0.299	-0.31	18.5 (18.9)	27.2 (20.1)	0.076	-0.44
Stigma	21.2 (24.3)	26.6 (20.2)	0.467	-0.24	17.3 (19.7)	35.0 (22.0)	0.001	-0.84
Complications	26.3 (23.6)	30.6 (18.6)	0.549	-0.20	21.2 (17.9)	35.7 (17.3)	0.002	-0.82
Sum score	29.2 (19.1)	39.6 (16.3)	0.082	-0.58	25.0 (15.9)	43.5 (14.5)	< 0.001	-1.21

Data are presented as mean (SD), calculated with independent samples *t* tests. RFIPC: Rating form of inflammatory bowel disease patient concerns; CD: Crohn's disease; UC: Ulcerative colitis; CF: Chronic fatigue. Cohen's *d* effect size: small, *d* = 0.2; medium, *d* = 0.5; large, *d* = 0.8.

Table 4 Univariate analysis of mean rating form of inflammatory bowel disease patient concerns scores in non-chronic fatigue and chronic fatigue patients

	Mean RFIPC adjusted for age, gender and education		Mean RFIPC adjusted for age, gender, education and IBD symptoms	
	No CF mean	CF mean	No CF mean	CF mean
CD (n = 48)	(n = 34)	(n = 14)	(n = 34)	(n = 14)
Impact of disease	30.7	47.6 ^a	32.2	44.0 ^a
Expectancy	37.3	53.0	37.8	52.0
Treatment	27.5	38.0	27.2	39.0
Intimacy	19.7	30.0	20.6	27.8
Stigma	20.9	27.3	20.8	27.6
Complications	26.4	30.3	25.9	31.8
Sum score	28.8	40.3	29.4	39.0
UC (n = 92)	(n = 72)	(n = 20)	(n = 72)	(n = 20)
Impact of disease	25.7	45.3 ^c	26.3	43.1 ^c
Expectancy	33.0	53.8 ^b	33.4	52.4 ^c
Treatment	27.9	47.8 ^c	28.5	45.6 ^c
Intimacy	18.7	26.7	19.0	25.6 ^a
Stigma	17.5	34.3 ^b	17.9	33.1 ^c
Complications	21.4	35.0 ^b	21.7	34.0 ^b
Sum score	25.2	42.8 ^c	25.6	41.2 ^c

RFIPC: Rating form of inflammatory bowel disease patient concerns; CD: Crohn's disease; UC: Ulcerative colitis; CF: Chronic fatigue; IBD: Inflammatory bowel disease. Significance levels are between patients reporting CF or not: ^a*P* ≤ 0.05, ^b*P* ≤ 0.01, ^c*P* ≤ 0.001.

were adjusted for the covariates age, gender and education, statistically significant differences were reproduced in the same RFIPC factors as in the raw, unadjusted analysis for both the UC and CD patients. Furthermore, when controlling for the perceived IBD symptom score, the significance level increased in two of the six factors (expectancy/stigma) for the UC group. One RFIPC factor (intimacy) in the UC group changed from non-significant to significant. Among the CD patients, controlling for perceived IBD symptoms had only a minor effect.

The correlation between RFIPC sum score, total fatigue, N-IBDQ total score and subdimensions of the SF-36 questionnaire

In both the UC and CD patient groups, increased RFIPC sum scores were associated with higher fatigue levels (measured as total fatigue) and reduced HRQoL (Table 5).

Table 5 Correlation (Spearman's ρ) between rating form of inflammatory bowel disease patient concerns sum, fatigue questionnaire sum, Norwegian inflammatory bowel disease questionnaire total and Short Form 36 dimensions

	CD (n = 48) RFIPC sum score	UC (n = 92) RFIPC sum score
FQ		
TF	0.36	0.49
N-IBDQ		
IBDQ Total	-0.53	-0.51
SF-36		
PF	-0.34	-0.40
RP	-0.28	-0.48
BP	-0.22	-0.44
GH	-0.49	-0.43
VT	-0.32	-0.48
SF	-0.40	-0.57
RE	-0.12	-0.49
MH	-0.31	-0.50

RFIPC: Rating form of inflammatory bowel disease patient concerns; CD: Crohn's disease; UC: Ulcerative colitis; FQ: Fatigue questionnaire; TF: Total fatigue; N-IBDQ: Norwegian inflammatory bowel disease questionnaire; SF-36: Short Form 36; PF: Physical functioning; RP: Role physical; BP: Bodily pain; GH: General health; VT: Vitality; SF: Social functioning; RE: Role emotional; MH: Mental health.

The latter showed a negative Spearman's ρ both for N-IBDQ and the SF-36 subdimensions. In general, these associations were stronger for the UC group than for the CD group.

Multicollinearity

Testing for multicollinearity revealed satisfactory values of both the VIF and tolerance (Table 6). The VIF was below the limit of 5 and the tolerance above the crucial threshold of 0.2 for both the UC and CD group. In the CD group, the ranges for the VIF and tolerance were 1.6-4.9 and 0.21-0.61, respectively. In the UC group, the ranges of the VIF and tolerance were 1.9-4.1 and 0.24-0.54, respectively.

DISCUSSION

The ramifications of IBD on patients subjective health has been thoroughly studied, identifying both a sub-

Table 6 Linear regression analysis of the rating form of inflammatory bowel disease patient concerns sum score, Norwegian inflammatory bowel disease questionnaire total score, total fatigue, chronic fatigue, Short Form 36 subdimensions and calculation of multicollinearity

	Dependent	Independent	β	P value	Tolerance	VIF
UC	RFIPC sum	N-IBDQ total	-0.70	< 0.001	0.24	4.1
	N-IBDQ total	RFIPC sum	0.29	< 0.001	0.58	1.7
		SF-36 BP	0.14	0.032	0.57	1.7
		SF-36 VT	0.17	0.019	0.50	2.0
		SF-36 SF	0.29	< 0.001	0.39	2.6
TF	SF-36 VT	-0.25	0.031	0.49	2.0	
	CF	RFIPC sum	0.34	0.018	0.46	2.2
CD	RFIPC sum	N-IBDQ total	-0.60	0.047	0.21	4.9
	N-IBDQ total	RFIPC sum	-0.17	0.047	0.71	1.4
		SF-36 SF	0.35	0.004	0.39	2.6
		SF-36 PF	0.22	0.033	0.49	2.0
		TF	-0.31	0.015	0.34	2.9

RFIPC: Rating form of inflammatory bowel disease patient concerns; CD: Crohn's disease; UC: Ulcerative colitis; TF: Total fatigue; CF: Chronic fatigue; N-IBDQ: Norwegian inflammatory bowel disease questionnaire; SF-36: Short Form 36; PF: Physical functioning; BP: Bodily pain; VT: Vitality; SF: Social functioning; VIF: Variance inflation factor.

group of patients with a worsening in HRQoL scores and other subgroups of patients with HRQoL scores that are comparable to the general population^[4,5,9,13]. This division of patient responses to IBD can partly be explained by demographic differences and differences in clinical variables^[4,5,9,13]. Recently, CF was reported to be at least twice as prevalent in IBD as in the general population, and CF has been reported to lead to clinically significant reductions in HRQoL^[6,10].

In the present study, we hypothesized that chronic fatigue might influence the level of disease-related concerns in IBD patients, and we found significant differences in the RFIPC scores between UC patients with CF and UC patients without CF. Although only one factor was significantly higher for CD patients with CF compared to CD patients without CF, there was a tendency of elevated scores in all RFIPC factors. In addition, we calculated effect sizes according to Cohen^[24], because statistical significant differences need not be of clinical importance and insignificant differences might be. As expected, the effect sizes were more pronounced in UC patients than in CD patients. The five RFIPC factors that were not significant in CD were, however, within Cohen's limits of small-to-medium effect sizes^[24]. This finding indicates that there are clinically important differences between IBD patients with CF and IBD patients without CF among all of the RFIPC scales.

IBD is often characterized by periods of remission and exacerbation; therefore, potentially negative consequences of the disease may become the primary focus in daily living^[1,2,5]. This has also been reported by Drossman^[7]. Worries and concerns are aspects of subjective health that can lead to decreased well-being^[7]. These worries may potentially be linked to situations in which the patient's expectations regarding physical and mental

functioning in "normal life" do not match their experienced reality^[8].

Within the published literature, there appears to be a pattern regarding which of the individual RFIPC items are rated by IBD patients as being most important^[7,25-27]. Our results concur with previous studies in this regard. However, the presence of chronic fatigue seems to further increase worries in IBD patients. To our knowledge, this association has not been previously reported for IBD. Among CD patients, we found that CF was associated with increased worries regarding pain and suffering. Research has highlighted the negative association between body pain and CD^[5] and recently it was reported that chronic fatigue further decreased body pain scores^[10]. In addition, patients with CD and CF seem to negatively relate their perceived energy capacity with their ability to achieve full potential and experience financial difficulties. The latter may be related to their ability to work and provide a household income. Indeed, studies have found that patients who are on sick-leave because of their IBD have a significant deterioration in HRQoL^[28].

The nature of worry is not clear; however, it is most often prospective and predominated by negative thinking^[29]. In IBD patients, these processes are linked to potentially negative events that may or may not occur in the future, including requiring surgery or an ostomy bag^[3,7,25-27]. IBD patients are reported to believe that stress influences the course of the disease^[30]. However, this potential link between psychological stress and inflammatory exacerbations is the subject of debate^[31-33]. Addressing the worries and concerns of patients might therefore have a potentially positive influence on both fatigue and HRQoL; on the other hand, Borgaonkar *et al*^[34] found that providing disease-related information to IBD patients worsens their short-term HRQoL.

In the present study, we found that increased levels of worrying were associated with both increased fatigue levels and reduced HRQoL. When investigating the potential association between worries, fatigue, and HRQoL in IBD patients, it is natural to wonder which is the cause and which is the effect. Is increased worrying a result of chronic fatigue or vice versa? Does impaired HRQoL increase worries and fatigue, or is HRQoL exaggerated by worries? The cross-sectional design of our study makes it difficult to reach any final conclusions. Our hypothesis, which we regard as plausible, is that both reduced HRQoL and increased fatigue are secondary effects of increased disease-related worries and concerns. This theory has also been proposed by others^[30]. Potentially, elevated levels of disease-related concerns may increase energy loss and thus manifest as increased fatigue and reduced HRQoL^[35].

The RFIPC was developed just a few years after the IBDQ, which may explain why the questionnaire has received much less attention in the literature^[7,12,36,37]. As shown by our study, employing a set of various questionnaires that measure subjective health increases the

likelihood of detecting subgroups of IBD patients who are at risk of coping less successfully^[8]. Naturally, there is the potential risk that subdimensions of different questionnaires are in fact operationalizations of the same phenomena^[22,23]. However, both the VIF and tolerance were within acceptable limits for both UC and CD patients, indicating that the questionnaires seem to measure different aspects of health perception. Given these results, we would argue that patient-reported outcome, rather than HRQoL, seems to be a more adequate definition in cases where one is interested in a wide variety of outcomes (e.g., worries, concerns, fatigue, HRQoL).

A limitation of the present study is that we did not include a specific questionnaire for measuring depression. This precludes the possibility of adjusting for depression as a potentially confounding variable for fatigue. Several reports have indicated a connection and overlap between fatigue and depression^[7,19,38]. However, the MH dimension of the SF-36 questionnaire indicated that the patients in our study did not differ from the general population in this respect. Consequently, depression does not appear to be a particular problem in this patient population. Moreover, in the CD patient group there may be type 2 statistical errors, as we were not able to obtain the strong associations between levels of worries and fatigue, as was shown for UC patients.

In conclusion, we report that chronic fatigue is associated with clinically significantly increased levels of disease-related worries in both UC and CD patients. This study provides additional information to the complex nature of understanding how IBD patients perceive their own subjective health.

ACKNOWLEDGMENTS

The authors wish to thank Professor Trudie Chalder, Kings College London, for the permission to use the fatigue questionnaire (FQ) in this study; Professor Jon Håvard Loge, University of Oslo and Professor Stein Kaasa, University Hospital of Trondheim, for the permission to use the validated Norwegian version of FQ; and Professor Leiv Sandvik, University of Oslo, for the statistics. Hege Hølmo Johannessen is acknowledged for correcting the manuscript for grammatical errors. Furthermore, we wish to thank Marita Nilsen (Innlandet Hospital), Hilde Næs, Brita Olafsen and Margrethe Solheim (Østfold Hospital Trust) for their contribution in gathering the data for this study.

COMMENTS

Background

Fatigue is reported to be a prevalent symptom in chronic diseases. In inflammatory bowel disease (IBD), however, there has been a lack of studies with fatigue as the primary endpoint. Recent publications have revealed that the amount of patients reporting long-lasting, chronic fatigue is two to three times elevated in patients with IBD compared to the background population.

Research frontiers

Chronic fatigue has been reported to significantly reduce IBD patients' health-re-

lated quality of life. In this study, the authors demonstrate that chronic fatigue is a potential important contributor to increased disease-related worries and concerns.

Innovations and breakthroughs

Recent studies have highlighted the importance of fatigue as a subjective symptom in IBD. This is the first study to report that chronic, long-lasting fatigue symptoms is associated with increased levels of disease-related worries.

Applications

By adding further complexity to the understanding of subjective health experiences in IBD, this study might help clinicians to detect patients at risk of less successful coping, potentially enhancing the patient's quality of life, and also their course of disease.

Terminology

Chronic fatigue refers to fatigue symptoms of some intensity, which have a long-lasting duration of six months or more.

Peer review

This is a very interesting and novel original contribution analyzing the problem of chronic fatigue and its relationship with worries associated with IBD. The study is interesting, seems well done and well reported. It adds a valuable bit of information to the IBD literature.

REFERENCES

- 1 **Cantor M**, Bernstein CN. Clinical course and natural history of ulcerative colitis. In: Sartor RB, Sanborn WJ (editor). *Kirsner's Inflammatory Bowel Diseases*. Edinburgh: Saunders, 2004: 280-288
- 2 **Munkholm P**, Binder V. Clinical features and natural history of Crohn's disease. In: Sartor RB, Sanborn WJ (editor). *Kirsner's Inflammatory Bowel Diseases*. Edinburgh: Saunders, 2004: 289-300
- 3 **Drossman DA**, Ringel Y. Psychosocial factors in ulcerative colitis and Crohn's disease. In: Sartor RB, Sanborn WJ (editor). *Kirsner's Inflammatory Bowel Diseases*. Edinburgh: Saunders, 2004: 340-356
- 4 **Mitchell A**, Guyatt G, Singer J, Irvine EJ, Goodacre R, Tompkins C, Williams N, Wagner F. Quality of life in patients with inflammatory bowel disease. *J Clin Gastroenterol* 1988; **10**: 306-310
- 5 **Bernklev T**, Jahnsen J, Lygren I, Henriksen M, Vatn M, Moum B. Health-related quality of life in patients with inflammatory bowel disease measured with the short form-36: psychometric assessments and a comparison with general population norms. *Inflamm Bowel Dis* 2005; **11**: 909-918
- 6 **Jelsness-Jørgensen LP**, Bernklev T, Henriksen M, Torp R, Moum BA. Chronic fatigue is more prevalent in patients with inflammatory bowel disease than in healthy controls. *Inflamm Bowel Dis* 2011; **17**: 1564-1572
- 7 **Drossman DA**, Leserman J, Li ZM, Mitchell CM, Zagami EA, Patrick DL. The rating form of IBD patient concerns: a new measure of health status. *Psychosom Med* 1991; **53**: 701-712
- 8 **Martz E**, Livneh H. *Coping with Chronic Illness and Disability: Theoretical, Empirical and Clinical Aspects*. New York: Springer, 2007
- 9 **Irvine EJ**. Measuring quality of life in inflammatory bowel disease. In: Targan SR, Shanahan F, Karp LC (editor). *Inflammatory Bowel Disease: From Bench to Bedside*. New York: Springer, 2003: 481-494
- 10 **Jelsness-Jørgensen LP**, Bernklev T, Henriksen M, Torp R, Moum BA. Chronic fatigue is associated with impaired health-related quality of life in inflammatory bowel disease. *Aliment Pharmacol Ther* 2011; **33**: 106-114
- 11 **Ware JE**, Sherbourne CD. The MOS 36-item short-form health survey (SF-36). I. Conceptual framework and item selection. *Med Care* 1992; **30**: 473-483
- 12 **Guyatt G**, Mitchell A, Irvine EJ, Singer J, Williams N, Goodacre R, Tompkins C. A new measure of health status for clinical trials in inflammatory bowel disease. *Gastroenterol-*

- ogy 1989; **96**: 804-810
- 13 **Irvine EJ**, Feagan B, Rochon J, Archambault A, Fedorak RN, Groll A, Kinnear D, Saibil F, McDonald JW. Quality of life: a valid and reliable measure of therapeutic efficacy in the treatment of inflammatory bowel disease. Canadian Crohn's Relapse Prevention Trial Study Group. *Gastroenterology* 1994; **106**: 287-296
 - 14 **Chalder T**, Berelowitz G, Pawlikowska T, Watts L, Wessely S, Wright D, Wallace EP. Development of a fatigue scale. *J Psychosom Res* 1993; **37**: 147-153
 - 15 **Walmsley RS**, Ayres RC, Pounder RE, Allan RN. A simple clinical colitis activity index. *Gut* 1998; **43**: 29-32
 - 16 **Harvey RF**, Bradshaw JM. A simple index of Crohn's-disease activity. *Lancet* 1980; **1**: 514
 - 17 **Bernklev T**, Moum B, Moum T. Quality of life in patients with inflammatory bowel disease: translation, data quality, scaling assumptions, validity, reliability and sensitivity to change of the Norwegian version of IBDQ. *Scand J Gastroenterol* 2002; **37**: 1164-1174
 - 18 **Loge JH**, Ekeberg O, Kaasa S. Fatigue in the general Norwegian population: normative data and associations. *J Psychosom Res* 1998; **45**: 53-65
 - 19 **Swain MG**. Fatigue in chronic disease. *Clin Sci (Lond)* 2000; **99**: 1-8
 - 20 **Loge JH**, Kaasa S. Short form 36 (SF-36) health survey: normative data from the general Norwegian population. *Scand J Soc Med* 1998; **26**: 250-258
 - 21 **Medical Outcomes Trust**. How to score the SF-36 health survey. Boston: Medical Outcomes Trust, 1994
 - 22 **Hocking RR**. Methods and Applications of Linear Models; regression and the analysis of variance. 2nd ed. New Jersey: John Wiley and Sons, 2003
 - 23 **O'Brian RM**. A Caution Regarding Rules of Thumb of Variance Inflation Factors. *Quality and Quantity* 2007; **41**: 673-690
 - 24 **Cohen J**. Statistical Power Analysis for the Behavioural Sciences. Mahwah: Lawrence Erlbaum Associates, 1988
 - 25 **Stjernman H**, Tysk C, Almer S, Ström M, Hjortswang H. Worries and concerns in a large unselected cohort of patients with Crohn's disease. *Scand J Gastroenterol* 2010; **45**: 696-706
 - 26 **Moser G**, Tillinger W, Sachs G, Genser D, Maier-Dobersberger T, Spiess K, Wyatt J, Vogelsang H, Lochs H, Gangl A. Disease-related worries and concerns: a study on out-patients with inflammatory bowel disease. *Eur J Gastroenterol Hepatol* 1995; **7**: 853-858
 - 27 **de Rooy EC**, Toner BB, Maunder RG, Greenberg GR, Baron D, Steinhart AH, McLeod R, Cohen Z. Concerns of patients with inflammatory bowel disease: results from a clinical population. *Am J Gastroenterol* 2001; **96**: 1816-1821
 - 28 **Bernklev T**, Jahnsen J, Henriksen M, Lygren I, Aadland E, Sauar J, Schulz T, Stray N, Vatn M, Moum B. Relationship between sick leave, unemployment, disability, and health-related quality of life in patients with inflammatory bowel disease. *Inflamm Bowel Dis* 2006; **12**: 402-412
 - 29 **Borkovec TD**, Ray WJ, Stober J. Worry: A Cognitive Phenomenon Intimately Linked to Affective, Physiological, and Interpersonal Behavioral Processes. *Cogn Ther Res* 1998; **22**: 561-576
 - 30 **Moser G**, Maeir-Dobersberger T, Vogelsang H, Lochs H. Inflammatory bowel disease: Patients' beliefs about the etiology of their disease - A controlled study. *Psychosom Med* 1993; **55**: 131
 - 31 **Cámara RJ**, Ziegler R, Begré S, Schoepfer AM, von Känel R. The role of psychological stress in inflammatory bowel disease: quality assessment of methods of 18 prospective studies and suggestions for future research. *Digestion* 2009; **80**: 129-139
 - 32 **Mawdsley JE**, Rampton DS. Psychological stress in IBD: new insights into pathogenic and therapeutic implications. *Gut* 2005; **54**: 1481-1491
 - 33 **Bernstein CN**, Walker JR, Graff LA. On studying the connection between stress and IBD. *Am J Gastroenterol* 2006; **101**: 782-785
 - 34 **Borgaonkar MR**, Townson G, Donnelly M, Irvine EJ. Providing disease-related information worsens health-related quality of life in inflammatory bowel disease. *Inflamm Bowel Dis* 2002; **8**: 264-269
 - 35 **Lerdal A**. A theoretical extension of the concept of energy through an empirical study. *Scand J Caring Sci* 2002; **16**: 197-206
 - 36 **Fayers PM**, Machin D. Quality of Life: The assessment, analysis and interpretation of patient-reported outcomes. Chichester: John Wiley and Sons, 2007
 - 37 **Feagan B**. Quality of life and pharmacoeconomics. In: Sartor RB, Sanborn WJ (editor). *Kirsner's Inflammatory Bowel Diseases*. Edinburgh: Saunders, 2004: 469-483
 - 38 **Wessely S**. Chronic fatigue: symptom and syndrome. *Ann Intern Med* 2001; **134**: 838-843

S- Editor Yang XC L- Editor Stewart GJ E- Editor Li JY

Pneumatosis cystoides intestinalis: A single center experience

Zulfu Arikanoglu, Erhan Aygen, Cemalettin Camci, Sami Akbulut, Murat Basbug, Osman Dogru, Ziya Cetinkaya, Cuneyt Kirkil

Zulfu Arikanoglu, Sami Akbulut, Murat Basbug, Department of Surgery, Diyarbakir Education and Research Hospital, 21400 Diyarbakir, Turkey

Erhan Aygen, Cemalettin Camci, Osman Dogru, Ziya Cetinkaya, Cuneyt Kirkil, Department of Surgery, Faculty of Medicine, Firat University, 23119 Elazig, Turkey

Author contributions: Arikanoglu Z, Basbug M, Aygen E and Kirkil C performed the surgical procedure; Akbulut S, Dogru O, Camci C, Arikanoglu Z and Cetinkaya Z contributed to writing the article and reviewing the literature, as well as undertaking a comprehensive literature search.

Correspondence to: Sami Akbulut, MD, Department of Surgery, Diyarbakir Education and Research Hospital, 21400 Diyarbakir, Turkey. akbulutsami@gmail.com

Telephone: +90-412-2580052 Fax: +90-412-2580050

Received: April 25, 2011 Revised: August 25, 2011

Accepted: August 31, 2011

Published online: February 7, 2012

Abstract

AIM: To share our experience of the management and outcomes of patients with pneumatosis cystoides intestinalis (PCI).

METHODS: The charts of seven patients who underwent surgery for PCI between 2001 and 2009 were reviewed retrospectively. Clinical features, diagnoses and surgical interventions of patients with PCI are discussed.

RESULTS: Seven patients with PCI (3 males, 4 females; mean age, 50 ± 16.1 years; range, 29-74 years) were analyzed. In three of the patients, abdominal pain was the only complaint, whereas additional vomiting and/or constipation occurred in four. Leukocytosis was detected in four patients, whereas it was within normal limits in three. Subdiaphragmatic free air was observed radiologically in four patients but not in three. Six of the patients underwent an applied laparotomy, whereas one underwent an applied explorative laparoscopy. PCI localized to the small intestine only was detected in four patients, whereas it was localized

to the small intestine and the colon in three. Three patients underwent a partial small intestine resection and four did not after PCI was diagnosed. Five patients were diagnosed with secondary PCI and two with primary PCI when the surgical findings and medical history were assessed together. Gastric atony developed in one case only, as a complication during a postoperative follow-up of 5-14 d.

CONCLUSION: Although rare, PCI should be considered in the differential diagnosis of acute abdomen. Diagnostic laparoscopy and preoperative radiological tests, including computed tomography, play an important role in confirming the diagnosis.

© 2012 Baishideng. All rights reserved.

Key words: Pneumatosis cystoides intestinalis; Peritoneal free air; Radiological tools; Diagnosis

Peer reviewer: Kjetil Soreide, MD, PhD, Associate Professor, Department of Surgery, Stavanger University Hospital, Armauer Hansensvei 20, PO Box 8100, N-4068 Stavanger, Norway

Arikanoglu Z, Aygen E, Camci C, Akbulut S, Basbug M, Dogru O, Cetinkaya Z, Kirkil C. Pneumatosis cystoides intestinalis: A single center experience. *World J Gastroenterol* 2012; 18(5): 453-457 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/453.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.453>

INTRODUCTION

Pneumatosis cystoides intestinalis (PCI) is a relatively uncommon condition, characterized by the presence of multiple gas-filled cysts within the wall of the gastrointestinal tract^[1-12]. The term "pneumatosis intestinalis" was first used by Duo Vernoi while observing autopsy specimens in 1730. The entity defined by Duo Vernoi is what we now know as primary PCI. The term "second-

ary PCI” was termed by Koss in 1952, who analyzed 213 pathological specimens and attributed 85% of the cases to a secondary disease^[1,2].

One of the pathognomonic features of PCI is pneumoperitoneum without peritoneal irritation as a result of a cyst rupture. In contrast, air retention leading to acute abdominal findings may be seen in some cases^[3].

PCI is a radiological or exploratory entity, not a disease, and the underlying causes are numerous. PCI may develop either after a benign procedure, such as endoscopy, or from an unknown cause (primary or idiopathic). In some cases, a more serious disease, such as secondary necrotizing enterocolitis, may be the cause. No clear consensus has yet been established, although many mechanical, bacterial, and pulmonary hypotheses have been proposed regarding the etiopathogenesis of PCI^[4]. PCI usually does not lead to clinical findings and may disappear spontaneously in cases in which the primary disease is treated. Steroids, an elemental diet, hyperbaric oxygen, antibiotics, and surgery have been used as treatments. In this study, we describe seven PCI cases, which were diagnosed and treated at our clinic.

MATERIALS AND METHODS

Seven patients were admitted to Firat University Faculty of Medicine, Department of Surgery, Emergency Unit, between January 2001 and August 2009. Their medical records were evaluated retrospectively to obtain follow-up and clinical data, including age, sex, initial complaints, medical histories, white blood cell, abdominal and thoracic radiography, intraoperative findings, surgical intervention, duration of hospital stay, complications and follow-up time (Table 1). Preoperative tests were performed, including routine biochemistry and thoracic and abdominal radiography. Five of the patients had findings consistent with an acute abdomen and were operated on. Contrast-enhanced abdominal computed tomography (CT) was performed in one case due to vague abdominal findings. The CT findings were consistent with a rectal perforation. Abdominal ultrasonography (USG) was also used in one patient who had marked tenderness in the right upper quadrant. All patients operated on underwent emergent surgery, and all patients were diagnosed with PCI intraoperatively. Surgical team consensus was used to determine which patients would be resected after a laparotomy and/or laparoscopic exploration. The affected segment was resected in cases with suspicion of bowel perforation and ischemia, whereas no additional surgical intervention was performed in cases in which only PCI was detected. The primary or secondary nature of the PCI was determined by considering the medical history and preoperative findings. Cases with no underlying predisposing disease were considered primary or idiopathic PCI, whereas those accompanying some disease, such as appendicitis, Crohn’s disease, pyloric stenosis, necrotizing enterocolitis, peptic ulcers, cystic fibrosis, or chronic obstructive lung disease, were regarded as sec-

ondary PCI. Follow-up time was determined from the time of surgery to the last visit to our outpatient clinic.

RESULTS

Data for seven patients with PCI (3 males, 4 females; age, 50 ± 16.1 years (mean \pm SD); range, 29-74 years) were analyzed retrospectively. Of the patients who presented at the Emergency Unit, three had severe abdominal pain, two had abdominal pain and vomiting, and two had abdominal pain, vomiting, and constipation. Liver and renal function tests as well as electrolyte values were normal in all patients, while marked leukocytosis was detected in four. Exam findings in six of the patients were consistent with acute peritonitis, whereas no findings other than minimal tenderness were noted in one patient (female, aged 29 years). Subdiaphragmatic free air was detected on plain thoracic and abdominal radiographs in four patients. Abdominal USG used in one patient (female, aged 74 years), who had marked tenderness in the right upper quadrant, revealed cholecystitis together with an image consistent with a stone in the lower tip of the choledoch. An abdominal CT of a 29-year-old female patient with vague findings revealed free air extending into the retroperitoneum, indicating a rectal perforation. This patient was diagnosed with acute abdomen and was scheduled for urgent surgery. A PCI diagnosis was established in seven patients after a laparotomy in six and a laparoscopic exploration in one. An appearance consistent with PCI was observed in the small intestine of four patients and in the small intestine and colon of three (Figure 1A). A small intestinal perforation was observed in only one (female, aged 34 years) of these cases (Figure 1B). PCI was detected incidentally in a 74-year-old female patient who was scheduled for bile duct surgery (cholecystectomy and choledoch exploration only). Three patients underwent a partial small intestinal resection and anastomosis, while four had no additional surgical procedures after PCI was diagnosed. All patients were given 3 L/min oxygen during the first 3 postoperative days. During the 5-14 d clinical follow-up, a 48-year-old male developed gastric atony, whereas the remaining six patients were discharged with no complications. No additional complications were observed in any of the patients during the 15 ± 5.4 mo (range, 8-23 mo) follow-up. After considering the surgical findings and medical and surgical histories, five of these patients had secondary PCI and two had primary PCI.

DISCUSSION

PCI is a rare condition characterized by multilocular gas-filled cysts localized in the submucosa and subserosa of the gastrointestinal tract^[5,13-17]. The term “pneumatosis intestinalis” was first used by Duo Vernoi during postmortem observations. A PCI diagnosis in surviving patients was first established by Hahn in 1899. A PCI diagnosis *via* preoperative radiological findings was first

Table 1 Demographic and clinical characteristics of the seven patients with pneumatosis cystoides intestinalis

No.	Age	Sex	Complaint	Medical history	WBC	Radiologic tools	Loc.	Etiology	Surgical intervention	Length of hospital stay (d)	Postoperative complication	Follow-up (mo)
1	29	F	AP + V	Endoscopy	11.1	X-ray, CT ⁴	SB	Secondary	Ileal resection + anastomosis	7	No	14
2	48	M	AP + V + C ²	Peptic ulcer perforation	NR	X-ray	SB	Secondary	Ileal resection + anastomosis	14	Gastric atony	21
3	71	M	AP + V + C ²	CLL (CT ³)	35	X-ray	SB	Secondary	Laparotomy	5	No	8
4	74	F	AP	Normal	NR	X-ray, US	SB	Secondary	Cholecystectomy + choledocotomy + drainage	11	No	23
5	53	F	AP	Colonoscopy	NR	X-ray	SB + C ¹	Secondary	Laparotomy	8	No	18
6	34	F	AP + V	Normal	23	X-ray	SB + C ¹	Primary	Ileal resection + anastomosis	10	No	12
7	41	M	AP	Normal	16	X-ray	SB + C ¹	Primary	Laparoscopic exploration	7	No	9

WBC: White blood cell; SB: Small bowel; ¹C: Colon; AP: Abdominal pain; V: Vomiting; ²C: Constipation; CLL: Chronic lymphocytic leukemia; ³CT: Chemotherapy; NR: Normal range; ⁴CT: Computed tomography; US: Ultrasonography.

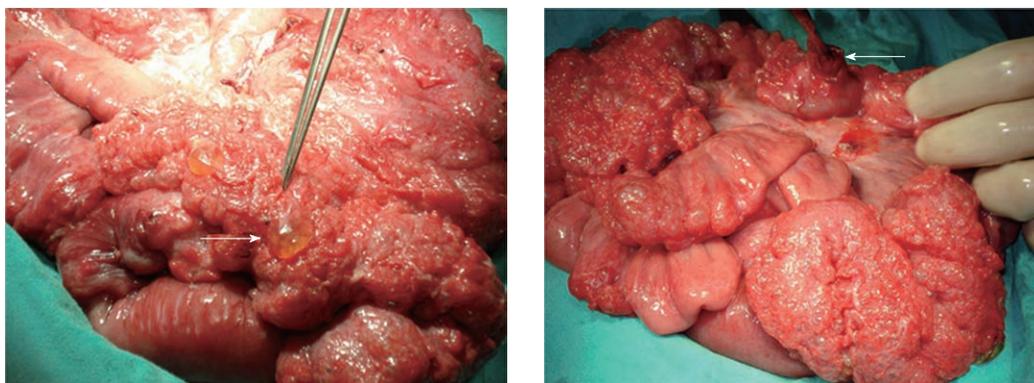


Figure 1 Intraoperative appearance of multiple air sacs in the small intestine. A: Multiple air sacs (white arrow); B: A perforation and multiple air sacs (white arrow).

described by Baumann-Schender in 1939. The condition originally described by Duo Vernoi is what we now consider primary PCI. The term “secondary PCI” was coined by Koss in 1952, who analyzed 213 pathological specimens and attributed 85% of the cases to a secondary disease^[1-3].

Several hypotheses have been proposed regarding the development of PCI, although its pathogenesis is still controversial. Two main hypotheses regarding the fundamental pathogenesis of PCI are mechanical and bacterial^[17]. The mechanical hypothesis postulates that PCI develops when defects in the mucosa, in combination with increased intraluminal pressure, allow gas to infiltrate the gastrointestinal (GI) tract wall. A subgroup of patients with severe pulmonary conditions may present with PCI arising from pulmonary causes, such as cough and rapid changes in intra-abdominal pressure. The bacterial hypothesis proposes that PCI develops when gas-producing bacteria gain entry into the GI tract wall and produce gas pockets. Much of the supporting evidence for these two hypotheses is derived from observational studies, and mechanical and bacterial mechanisms may occur simultaneously^[4,6,7].

Although PCI may occur anywhere in the gastrointestinal tract, from the esophagus to the rectum, it is usually seen in the intestine. A previous study reported that 20%-51.6% of all PCI cases involve the small intestine, 36%-78% involve the colon, and 2%-22% include both the small intestine and colon. The small intestine was involved in 57.1% of the cases we presented here and 42.9% involved the small intestine and the colon^[1,2,6,8].

PCI is not a disease but a clinical entity. The etiology can be classified by considering factors thought to play a role in its development. Based on this notion, PCI can be divided into primary and idiopathic (15%) or secondary (85%) type^[9]. No identifiable underlying or predisposing factor is present in the primary or idiopathic type. However, numerous gastrointestinal diseases, including appendicitis, necrotizing enterocolitis, Crohn's disease, pyloric stenosis, ulcerative colitis, diverticular disease, necrotizing enterocolitis, gastroduodenal ulcer, and sigmoid volvulus, may accompany PCI as a secondary cause. PCI has also been reported as accompanying some non-gastrointestinal diseases, such as chronic obstructive pulmonary disease, collagen tissue diseases, acquired immune deficiency syndrome, and glucocorticoid use. PCI

cases secondary to surgical or endoscopic trauma have also been reported^[10,11].

Lesions are usually localized to the left hemicolon or its mesentery or to the submucosal layer and are frequently characterized by segmentary involvement in the primary form of the disease. However, involvement is usually subserosal in the secondary form, and occurs in the stomach, small intestine, and right colon, usually in a generalized or segmented pattern^[7].

The incidence of PCI is unknown, because it is usually asymptomatic. Symptoms, if any, are usually secondary to an underlying disease. Together with non-specific symptoms, such as abdominal discomfort, diarrhea, constipation, rectal bleeding, tenesmus, or loss of weight, severe complications, including volvulus, intestinal obstruction, tension pneumoperitoneum, bleeding, intussusception, and intestinal perforation may be seen in 3% of patients^[18-23].

Radiological tools are important for diagnosing PCI. These include plain radiographs, USG, barium series, CT, CT-colonoscopy, magnetic resonance imaging and MRI-colonography, endoscopy, and colonoscopy^[19,20]. X-ray is of great importance, because it is readily available in every emergency room. Cysts usually appear as radiolucent shadows, similar to a bunch of grapes, close to the intestinal lumen on radiographs. Free air underneath the diaphragm may be seen if these cysts perforate. An appearance of bulging into the lumen as a filling defect is seen on barium-colon radiographs^[15,23]. Linear or spot-like hyperechoic images may be seen in the intestinal wall on USG. CT is the most useful method for diagnosing PCI and is important because it provides data on other abdominal pathologies. However, CT may not provide data on intestinal ischemia and necrosis^[1,4,7,12]. The colonoscopic findings may be similar to multiple polyposis or collections of submucosal tumors, but subserous pneumatosis may go undetected^[20]. A laparoscopic exploration is quite useful to confirm a PCI diagnosis, if the physical examination findings are suspicious, and particularly in cases that are not preoperatively diagnosed clearly using the above-mentioned radiological methods. Diagnostic laparoscopy provides the convenience of converting to open surgery as well as confirming the diagnosis.

When presence of such an entity is confirmed radiologically, gastroenterologic surgeons begin to feel annoyance. The answer to the question, "What should we do to these patients?" is correlated with the experience of each surgeon on that entity. The approach to a patient with PCI should be determined by evaluating the underlying causes and exam findings together. A specific treatment is not recommended in asymptomatic patients who are detected as having PCI radiologically and whose examination findings are negative. Conservative approaches, including nasogastric decompression, intestinal rest, antibiotic therapy and oxygen, are recommended for patients with positive examination findings and normal biochemical parameters who are confirmed radiologically to have no intestinal ischemia or perforation^[24]. Applying 250 mmHg PO₂

pressure or 70% oxygen inhalation for 5 d or 2.5 atmospheres of hyperbaric oxygen pressure for 150 min/d for 3 consecutive days can lead to resolution of gas collection within a cyst^[10,13,24,25]. An urgent laparotomy is necessary in cases of intestinal ischemia, obstruction, intestinal bleeding, or peritonitis^[14,16]. Definitive surgery should be performed during a laparotomy if necrosis, perforation, or marked ischemia is observed in the intestine. Furthermore, no additional surgical procedures should be conducted unless other pathology is detected in addition to serosal or subserosal air cysts.

Consequently, clinical suspicion, physician experience, radiological tools, and team spirit are important in terms of the approach to PCI. When and how to treat these patients is the main issue to lower mortality and morbidity.

COMMENTS

Background

Pneumatosis cystoides intestinalis (PCI) is a pathologic condition defined as infiltration of gas into the wall of the gastrointestinal tract.

Research frontiers

The authors retrospectively reviewed the diagnosis and management of seven patients with pneumatosis cystoides intestinalis.

Innovations and breakthroughs

Clinical suspicion, physician experience, radiological tools and team spirit are important in terms of the approach to PCI. When and how to treat these patients is the main issue to lower mortality and morbidity.

Applications

According to authors' opinion, specific treatment is not recommended in asymptomatic patients who are detected to have PCI radiologically and whose examination findings are negative. However, laparotomy is necessary in cases of intestinal ischemia, obstruction, intestinal bleeding or peritonitis

Terminology

The primary and idiopathic or secondary nature of the PCI is determined by considering the medical history and by preoperative examination. Cases with no underlying predisposing disease are considered primary PCI, whereas those accompanying some disease, such as appendicitis, Crohn's disease, pyloric stenosis, necrotizing enterocolitis, peptic ulcers, cystic fibrosis or chronic obstructive lung disease, are regarded as secondary PCI.

Peer review

This is a well written report on a small series of a rare entity. It has some educational value in the presentation and the figures.

REFERENCES

- 1 **Morris MS**, Gee AC, Cho SD, Limbaugh K, Underwood S, Ham B, Schreiber MA. Management and outcome of pneumatosis intestinalis. *Am J Surg* 2008; **195**: 679-682; discussion 682-683
- 2 **Greenstein AJ**, Nguyen SQ, Berlin A, Corona J, Lee J, Wong E, Factor SH, Divino CM. Pneumatosis intestinalis in adults: management, surgical indications, and risk factors for mortality. *J Gastrointest Surg* 2007; **11**: 1268-1274
- 3 **Voboril R**. Pneumatosis cystoides intestinalis--a review. *Acta Medica (Hradec Kralove)* 2001; **44**: 89-92
- 4 **St Peter SD**, Abbas MA, Kelly KA. The spectrum of pneumatosis intestinalis. *Arch Surg* 2003; **138**: 68-75
- 5 **Bilici A**, Karadag B, Doventas A, Seker M. Gastric pneumatosis intestinalis associated with malignancy: an unusual case report. *World J Gastroenterol* 2009; **15**: 758-760
- 6 **Wayne E**, Ough M, Wu A, Liao J, Andresen KJ, Kuehn D,

- Wilkinson N. Management algorithm for pneumatosis intestinalis and portal venous gas: treatment and outcome of 88 consecutive cases. *J Gastrointest Surg* 2010; **14**: 437-448
- 7 **Bolukbas FF**, Bolukbas C. Pnomatozis sistoides intestinalis. *Guncel Gastroenteroloji* 2004; **8**: 182-185
 - 8 **Jamart J**. Pneumatosis cystoides intestinalis. A statistical study of 919 cases. *Acta Hepatogastroenterol (Stuttg)* 1979; **26**: 419-422
 - 9 **Kim KM**, Lee CH, Kim KA, Park CM. CT Colonography of pneumatosis cystoides intestinalis. *Abdom Imaging* 2007; **32**: 602-605
 - 10 **Sakurai Y**, Hikichi M, Isogaki J, Furuta S, Sunagawa R, Inaba K, Komori Y, Uyama I. Pneumatosis cystoides intestinalis associated with massive free air mimicking perforated diffuse peritonitis. *World J Gastroenterol* 2008; **14**: 6753-6756
 - 11 **Türk E**, Karagülle E, Ocak I, Akkaya D, Moray G. [Pneumatosis intestinalis mimicking free intraabdominal air: a case report]. *Ulus Trauma Acil Cerrahi Derg* 2006; **12**: 315-317
 - 12 **Ho LM**, Paulson EK, Thompson WM. Pneumatosis intestinalis in the adult: benign to life-threatening causes. *AJR Am J Roentgenol* 2007; **188**: 1604-1613
 - 13 **Goel A**, Tiwari B, Kujur S, Ganguly PK. Pneumatosis cystoides intestinalis. *Surgery* 2005; **137**: 659-660
 - 14 **Mclaughlin SA**, Nguyen JH. Conservative management of nongangrenous esophageal and gastric pneumatosis. *Am Surg* 2007; **73**: 862-864
 - 15 **Di Giorgio A**, Sofo L, Ridolfini MP, Alfieri S, Doglietto GB. Pneumatosis cystoides intestinalis. *Lancet* 2007; **369**: 766
 - 16 **Kala Z**, Hermanova M, Kysela P. Laparoscopically assisted subtotal colectomy for idiopathic pneumatosis cystoides intestinalis. *Acta Chir Belg* 2006; **106**: 346-347
 - 17 **Yellapu RK**, Rajekar H, Martin JD, Schiano TD. Pneumatosis intestinalis and mesenteric venous gas - a manifestation of bacterascites in a patient with cirrhosis. *J Postgrad Med* 2011; **57**: 42-43
 - 18 **Frossard JL**, Braude P, Berney JY. Computed tomography colonography imaging of pneumatosis intestinalis after hyperbaric oxygen therapy: a case report. *J Med Case Reports* 2011; **5**: 375
 - 19 **Donati F**, Boraschi P, Giusti S, Spallanzani S. Pneumatosis cystoides intestinalis: imaging findings with colonoscopy correlation. *Dig Liver Dis* 2007; **39**: 87-90
 - 20 **Tsujimoto T**, Shioyama E, Moriya K, Kawaratani H, Shirai Y, Toyohara M, Mitoro A, Yamao J, Fujii H, Fukui H. Pneumatosis cystoides intestinalis following alpha-glucosidase inhibitor treatment: a case report and review of the literature. *World J Gastroenterol* 2008; **14**: 6087-6092
 - 21 **Koreishi A**, Lauwers GY, Misdraji J. Pneumatosis intestinalis: a challenging biopsy diagnosis. *Am J Surg Pathol* 2007; **31**: 1469-1475
 - 22 **Doumit M**, Saloojee N, Seppala R. Pneumatosis intestinalis in a patient with chronic bronchiectasis. *Can J Gastroenterol* 2008; **22**: 847-850
 - 23 **Nagata S**, Ueda N, Yoshida Y, Matsuda H. Pneumatosis coli complicated with intussusception in an adult: report of a case. *Surg Today* 2010; **40**: 460-464
 - 24 **Mizoguchi F**, Nanki T, Miyasaka N. Pneumatosis cystoides intestinalis following lupus enteritis and peritonitis. *Intern Med* 2008; **47**: 1267-1271
 - 25 **Togawa S**, Yamami N, Nakayama H, Shibayama M, Mano Y. Evaluation of HBO2 therapy in pneumatosis cystoides intestinalis. *Undersea Hyperb Med* 2004; **31**: 387-393

S- Editor Tian L L- Editor Logan S E- Editor Li JY

Survival analysis of cholangiocarcinoma: A 10-year experience in Malaysia

Ahmad Ramzi Yusoff, Mohd Muzammil Abdul Razak, Yoong Boon Koon, R Vijeyasingam, Siti Zuraidah Mahmud

Ahmad Ramzi Yusoff, Faculty of Medicine, Universiti Teknologi MARA, Sg. Buloh Campus, 47000 Sg. Buloh, Selangor, Malaysia

Mohd Muzammil Abdul Razak, Yoong Boon Koon, R Vijeyasingam, Department of Surgery, University Malaya Medical Centre, 59100 Kuala Lumpur, Malaysia

Siti Zuraidah Mahmud, Institute for Medical Research, Jalan Pahang, 50588 Kuala Lumpur, Malaysia

Author contributions: Yusoff AR designed the study, collected and analyzed data, and drafted the manuscript; Abdul Razak MM and Yoong BK supervised the research; Vijeyasingam R operated on patients and provided the data; and Siti ZM edited the article and conducted the literature review.

Correspondence to: Dr. Ahmad Ramzi Yusoff, Faculty of Medicine, Universiti Teknologi MARA, Sg. Buloh Campus, 47000 Sg. Buloh, Selangor, Malaysia. dr_aramzy@yahoo.com
Telephone: +60-3-61564182 Fax: +60-3-61564292

Received: February 2, 2011 Revised: April 26, 2011

Accepted: May 3, 2011

Published online: February 7, 2012

Abstract

AIM: To investigate the clinical features and survival of patients treated for cholangiocarcinoma in our institution and to analyze the factors affecting their survival.

METHODS: This retrospective cohort study assessed patients diagnosed with cholangiocarcinoma between January 1997 and December 2007 at the University Malaya Medical Centre in Malaysia. The clinical data and associated outcomes were collected using a structured proforma.

RESULTS: Of the 69 patients diagnosed with cholangiocarcinoma, 38 (55%) were male; mean patient age was 61 years. Twelve patients (17%) had intrahepatic, 38 (55%) had perihilar and 19 (28%) had distal tumors. Only 12 patients underwent curative surgery, including seven R0 resections. Only one patient died within 30 d after surgery. The overall median survival

was 4 mo, whereas the median survival of R0 resected patients was 16 mo. The overall 1-, 2- and 3-year cumulative survival rates were 67%, 17% and 17%, respectively. Survival rates were significantly associated with curative resection ($P = 0.002$), intrahepatic tumor ($P = 0.003$), negative margin status ($P = 0.013$), early tumor stage ($P = 0.016$), higher tumor differentiation ($P = 0.032$) and absence of jaundice ($P = 0.038$). Multivariate analysis showed that tumor location was a significant independent predictor of patient survival.

CONCLUSION: Curative, margin-negative resection of early stage, well-differentiated intrahepatic tumors is associated with improved patient survival.

© 2012 Baishideng. All rights reserved.

Key words: Cholangiocarcinoma; Bile duct tumor; Surgery; Malaysia

Peer reviewer: Richard A Kozarek, MD, Executive Director, Digestive Disease Institute, Virginia Mason Medical Center 1100 Ninth Avenue, PO Box 900, Seattle, WA 98111-0900, United States

Yusoff AR, Abdul Razak MM, Yoong BK, Vijeyasingam R, Siti ZM. Survival analysis of cholangiocarcinoma: A 10-year experience in Malaysia. *World J Gastroenterol* 2012; 18(5): 458-465 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/458.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.458>

INTRODUCTION

Cholangiocarcinoma is a rare malignant neoplasm of biliary tract epithelium, accounting for less than 2% of all human malignancies^[1]. Cholangiocarcinoma is the second most common primary hepatic malignancy after hepatocellular carcinoma (HCC)^[2] and is associated with

poor patient outcomes. Over the past three decades, however, the worldwide incidence of and mortality from cholangiocarcinoma have steadily increased^[3].

Cholangiocarcinoma is difficult to diagnose and is usually fatal, due to its late clinical presentation and the absence of effective non-surgical therapeutic modalities^[4]. The incidence of cholangiocarcinoma peaks in patients aged 50-70 years, and there is a slight male predominance^[5]. Most patients have unresectable disease at the time of diagnosis and usually die within 6-12 mo from the cancer cachexia, liver failure and biliary sepsis^[6]. The 5-year survival rate is low (about 5%), and has remained virtually unchanged over the past 20 years^[5,7]. Surgical resection remains the only hope for cure, and radical resection has improved outcomes, although it is also associated with high perioperative morbidity and mortality rates^[8]. Since most patients with cholangiocarcinoma present with advanced disease, palliative stenting and chemoradiation are reserved for non-resectable patients, those with recurrence, and those who refuse surgical treatment^[3].

Little is known about the survival of cholangiocarcinoma patients in Malaysia. We therefore analyzed the factors affecting survival of patients with cholangiocarcinoma treated at the University Malaya Medical Centre, Malaysia.

MATERIALS AND METHODS

This retrospective cohort study assessed patients diagnosed with cholangiocarcinoma and treated between January 1997 and December 2007 at our center, a tertiary referral center in Malaysia with a specialized hepatobiliary surgery unit and a gastroenterology unit. Patients with a diagnosis of cholangiocarcinoma confirmed histologically by tissue biopsy, and patients without a histologically confirmed diagnosis but with a strong provisional diagnosis by clinical examination, biochemical results and positive endoscopic or imaging [i.e., endoscopic retrograde cholangiopancreatography (ERCP)/magnetic resonance cholangiopancreatography (MRCP) or computed tomography (CT) of the abdomen] results^[7] were included. Patients with HCC, tumor at the head of the pancreas and gallbladder carcinoma were excluded.

Combinations of ultrasonography, CT, MRCP, ERCP and percutaneous cholangiography (PTC) were used for tumor diagnosis and staging, and for assessment of resectability. Metastatic disease was evaluated by CT of the thorax, abdomen and pelvis and/or chest radiography (CXR).

Patients were classified into three groups based on the anatomic location of the primary lesion, specifically intrahepatic, perihilar and distal types as proposed by the guidelines for the diagnosis and treatment of cholangiocarcinoma^[5]. Intrahepatic tumors were defined as those confined to the liver and not involving the extrahepatic biliary tree. Perihilar tumors were defined as those involving or requiring resection of the hepatic duct bifur-

cation and were typically located in the extrahepatic biliary tree proximal to the origin of the cystic duct. Distal tumors were defined as extrahepatic lesions located in the peripancreatic region^[9].

Patients were staged according to the tumor-node-metastasis (TNM) system^[6,10] and assessed for resectability. Variables assessed included therapeutic options (surgical or palliative treatment), operative data, and 30-d postoperative morbidity and mortality. Survival was measured from the date of first presentation to the date of death or last follow-up visit.

Statistical calculations were performed using SPSS version 13.0. Categorical variables were compared using χ^2 tests for association. One-way analysis of variance was used to compare continuous variables among the three groups of patients classified by tumor location. Results were presented as means \pm SD, unless otherwise specified. Survival curves were calculated using the Kaplan-Meier method and compared using log-rank tests (Mantel-Cox). Cox proportional hazard models were used to calculate adjusted hazard ratios. A *P* value $<$ 0.05 was considered significant.

RESULTS

Demography

Of the 69 patients included in this study, 12 (17%) had intrahepatic, 38 (55%) had perihilar and 19 (28%) had distal tumors. Thirty-eight patients were male (55%) and 31 (45%) were female (a male to female ratio, 1.2:1). Mean \pm SD patient age was 61 \pm 14.2 years (range, 18-91 years), although patients with intrahepatic tumors were younger than those with perihilar and distal tumors (*P* $<$ 0.05). When subdivided ethnically, 42 patients (61%) were Chinese, 20 (29%) were Malay and 7 (10%) were Indian.

Risk factors

Of the 69 patients, 22 (29%) had a history of chronic cigarette smoking, 13 (19%) each had diabetes mellitus and regular alcohol intake, and 5 (7%) had chronic hepatitis B, with only 1 each (1.4%) having hepatolithiasis and choledochal cyst. None of our patients had other strong risk factors, such as inflammatory bowel disease, primary sclerosing cholangitis or liver fluke infestation.

Clinical evaluation

The most common symptoms among our patients were jaundice (78%), anorexia (57%), weight loss (52%), abdominal pain (44%), abdominal mass (44%), itchiness (25%), vomiting (9%) and fever (7%). Ninety-eight percent of the patients with extrahepatic cholangiocarcinoma were jaundiced (63% of perihilar tumor and 35% of distal tumor patients), whereas only 2% of intrahepatic patients was jaundiced (*P* $<$ 0.001). No specific symptom was significantly related to any of the three types of cholangiocarcinoma. The median duration of complaints prior to medical consultation was 30 d (range 1-365 d). However, patients with intrahepatic lesions present somewhat later (*P*

Table 1 Laboratory data on admission and location of tumours

	Total (<i>n</i> = 69)	Intrahepatic (<i>n</i> = 12)	Perihilar (<i>n</i> = 38)	Distal (<i>n</i> = 19)	<i>P</i> value ¹
Total bilirubin (mmol/L)	178 (158)	45 (109)	216 (163)	188 (131)	0.003
Conjugated bilirubin (mmol/L)	140 (122)	31 (87)	170 (123)	151 (102)	0.002
INR	1.2 (0.7)	1.1 (0.1)	1.0 (0.1)	1.6 (1.1)	0.005

¹Analysis of variance. Values in mean (SD); *P* value < 0.05 is significant. INR: International normalized ratio.

< 0.05) than those with perihilar or distal lesions.

Investigations

When we assessed laboratory variables in these patients, we found that total bilirubin concentration was significantly higher in patients with perihilar than in those with distal or intrahepatic lesions (*P* = 0.003) (Table 1). The international normalized ratio (INR) value was also predictive of lesion site, as it was higher in patients with distal than with perihilar or intrahepatic lesions (*P* = 0.005).

The tumor markers carcinoembryonic antigen, alpha-fetoprotein and carbohydrate antigen 19-9 were measured in about 50% of these patients, but none of them significantly correlated with tumor location.

Imaging

Ultrasound was the first line non-invasive imaging method used in 45 (65%) patients, detecting 50%-76% of obstructed biliary systems. Almost 91% of these patients underwent CT scans to further assess the extent of pathology, including liver masses, lymph nodes and the involvement of major vessels. We found that about 20% of the abdominal lesions were not detected by ultrasound alone, but were detected on CT scan. Only four patients underwent MRCP.

In 58 (84%) patients, ERCP was the first invasive method used to assess the obstructed biliary system. ERCP showed that 51 (74%) patients had strictures at various levels of their biliary trees. Plastic stents were inserted successfully into 44 (76%) of these patients to relieve their biliary obstructions, whereas the other seven (14%) failed in stent insertion; five later underwent PTC drainage. Cytology brushing of suspicious strictures was performed in 22 (38%) patients, with 10 (45%) being positive for cancer.

Preoperative staging for resectability was predicted based on ultrasound, CT scan, MRCP and CXR. None of these patients underwent staging laparoscopy because it was not a routine practice in our center. Based on the American Joint Committee on Cancer TNM staging system, 23 (33%) patients had stage 1 tumors, 7 (10%) had stage 2, 17 (25%) had stage 3 and 22 (32%) had stage 4. Stages 3 and 4 were considered unresectable.

Surgical treatment

Although preoperative staging indicated that 30 (43%)

Table 2 Types of surgical resection

Types of surgery	Patients (<i>n</i> = 12)
Left hemihepatectomy	2
Right hemihepatectomy	1
Segmental hepatic resections	2
Extrahepatic bile duct resection	5
Whipple's procedure	2

patients were candidates for surgical resection, only 22 (32%) patients underwent laparotomy with curative intent. Three patients (and their relatives) refused surgery, citing age as their primary concern (mean age 78 years), whereas five were excluded from surgery due to age (mean 71 years) and/or associated comorbidity. At laparotomy, 10 patients had extensive local disease and/or hepatic metastasis that precluded resection. The remaining 12 patients underwent potentially curative resection (resectability rate, 55%). All except one with evidence of biliary obstruction underwent preoperative biliary drainage either by endoscopic stenting or percutaneous transhepatic biliary drainage (PTBD). The type of surgery depended on the location of the tumor (Table 2). In general, intrahepatic tumors were treated by hepatic resection; perihilar lesions by excision of the extrahepatic biliary tree and lymph node dissection, with or without hepatic resection; and distal tumors by Whipple pancreaticoduodenectomy. Biliary reconstructions were mostly by Roux-ex-Y hepaticojejunostomy. All operations were performed by trained hepatobiliary surgeons.

Surgical morbidity and mortality

Complications occurred in patients with all three tumor types, but the differences were not statistically significant. Three patients developed post-operative ileus, which resolved after conservative treatment. Two patients had intra-abdominal hemorrhage. One had undergone a Whipple procedure and was re-explored within 24 h because of portovenous bleeding; unfortunately, this patient died the next day. The second patient did not require intervention and was treated with blood transfusion and correction of coagulopathy. Three patients developed intra-abdominal collection, later complicated by abscesses, including one caused by bile leakage from the anastomosis. None of these patients, however, required surgical or percutaneous intervention. Two patients had surgical site infections, including one with anastomotic stenosis and the other with deep venous thrombosis (DVT) despite DVT prophylaxis. Only one patient died of complications within 30 postoperative days; hence the perioperative mortality rate was 8%.

Of the 10 patients who underwent surgery but were found to have advanced inoperable disease at laparotomy, 3 underwent a palliative bypass procedure, consisting of hepaticojejunostomy and gastrojejunostomy, whereas four underwent gastrojejunostomy alone. Two patients underwent cholecystectomy and biliary stent insertion, whereas one underwent only laparotomy and biopsy.

Table 3 Tumour histology, degree of differentiation, diameter, margin, perineural and lymph node involvements by tumour location *n* (%)

	Total (<i>n</i> = 12)	Intrahepatic (<i>n</i> = 3)	Perihilar (<i>n</i> = 6)	Distal (<i>n</i> = 3)	<i>P</i> value ¹
Tumour histology					
Adenocarcinoma	11 (92)	2 (18%)	6 (55%)	3 (27)	
Other	1 (8)	1 (100%)	0	0	
Degree of differentiation					
Well	5 (42)	2 (40%)	2 (40%)	1 (20)	
Moderate	6 (50)	1 (17%)	3 (50%)	2 (34)	
Poor	1 (8)	0	1 (100%)	0	
Tumour diameter, cm					
Size, mean ± SD	9.6 ± 15.2	27.1 ± 24.3	2.8 ± 2.2	5.7 ± 15.2	0.048 ¹
Margin					
Negative	7 (58)	3 (43)	3 (43)	1 (14)	
Positive	5 (42)	0	3 (60)	2 (40)	
Lymph node involvement					
Negative	8 (67)	2 (25)	4 (50)	2 (25)	
Positive	4 (33)	1 (25)	2 (50)	1 (25)	
Perineural involvement					
Negative	5 (42)	3 (60)	2 (40)	0	
Positive	7 (58)	0	4 (57)	3 (43)	
Lymphovascular invasion					
Negative	7 (58)	2 (29)	4 (57)	1 (14)	
Positive	5 (42)	1 (20)	2 (40)	2 (40)	

¹Analysis of variance. *P* value < 0.05 is significant.

Tumor characteristics

Of the 12 patients who underwent curative resection, 11 (92%) had adenocarcinoma and one had a papillary adenocarcinoma. The mean ± SD tumor diameter was 9.6 ± 15.2 cm (range, 0.5 cm-55 cm). Intrahepatic tumors were larger than perihilar and distal tumors (*P* < 0.05). Five (42%) patients had well-differentiated adenocarcinoma and the others had moderately or poorly differentiated adenocarcinoma. Seven (58%) patients had perineural involvement, 5 (42%) had lymphovascular invasion and 4 (33%) had regional lymph node metastases. Seven (58%) patients were resected with negative margins (R0 resection), whereas the other 5 (42%) had microscopically positive margins. Table 3 summarizes the characteristics of these tumors.

Palliation

Forty-seven (68%) patients did not undergo surgery but received palliative treatment, including 31 (66%) who underwent palliative biliary drainage by endoscopic stenting, 23 (49%) who underwent PTBD with or without concurrent stenting and 13 who underwent both. Sixteen patients had their stents changed on subsequent follow-up, of whom, nine had self-expanding metal stents. Three (6%) patients received palliative chemotherapy alone, using a variety of chemotherapeutic agents (5-fluorouracil, cisplatin, gemcitabine). Three (6%) patients were too ill and hence received best supportive care.

DISCUSSION

We have described our experience managing patients

with cholangiocarcinoma, a rare type of tumor. The incidence of this tumor is likely increasing in Malaysia. The latest National Cancer Registry 2003-2005 Peninsular Malaysia has classified cholangiocarcinoma into the category of liver and gallbladder cancers rather than as a separate tumor. The incidence of liver cancer in Malaysia has been reported to be 3.6% for males and 1.2% for females, while the rates of gallbladder cancer were 0.8% and 0.7%, respectively. Morphologically, about 2.2% of these liver cancers and 33.9% of these gallbladder cancers were cholangiocarcinomas. The incidence of both liver and gallbladder cancers was higher for Chinese than for Malays and Indians.

Cholangiocarcinoma is best classified according to its anatomical location into intrahepatic, perihilar and distal tumors^[9,11,12]. Most (40%-60%) tumors are perihilar or Klatskin tumors, with 20%-30% being distal and 10% being intrahepatic tumors^[9,13]. Our findings were similar to these rates, in that 17% of tumors were intrahepatic, 54% were perihilar and 29% were distal.

The demographic characteristics of our patients were comparable to those of patients in other larger series. Mean patient age was 61 years (range, 18-91 years), while a review of 294 patients with cholangiocarcinoma by Nakeeb *et al*^[9] showed a mean age of 62.2 years and a review by DeOliveira *et al*^[13] of 564 patients showed a median age of 65 years. We found, however, that patients with intrahepatic tumors were significantly younger than those with perihilar or distal tumors. We observed a slight male predominance, with a male to female ratio of 1.2:1, similar to the ratios of 1.2:1^[9] and 1.38:1 reported previously^[13].

Of these patients, 32% presented in 1997-2002, whereas 68% presented in 2003-2007. We observed similar increases in the number of patients with primary tumors, suggesting that these higher rates may be due to improvement in tumor detection, ERCP proficiency, or increased awareness of the availability of local surgical expertise.

Only a few patients in our series had strong risk factors such as chronic hepatitis B, hepatolithiasis and choledochal cyst^[14-16]. None had a history of primary sclerosing cholangitis^[14] or liver fluke infestation^[17], the latter of which is not endemic in Malaysia. Chronic cigarette smoking, regular alcohol intake and diabetes mellitus were among the risk factors^[18,19] observed in our population, but none of these risk factors was associated with any particular tumor site.

The most common symptom observed in these patients was jaundice. Jaundice was significantly more common in patients with extrahepatic (i.e., perihilar and distal) than with intrahepatic cholangiocarcinoma (*P* < 0.001). This was confirmed by laboratory results showing that the concentration of bilirubin was significantly higher in patients with perihilar (extrahepatic) than in those with intrahepatic tumors (*P* = 0.002). Similar results have been reported previously^[13,20]. We also found that INR was higher in patients with distal tumors (*P* =

Table 4 Univariate analysis of different variables in relation to survival

Variables	Survival rates (%)			P value
	1-year	2-year	3-year	
Jaundice				0.038
Absent	40	19	9	
Present	19	6	2	
Staging				0.016
Stage 1	43	17	9	
Stage 2	43	14	0	
Stage 3	12	4	0	
Stage 4	5	0	0	
Surgery (curative or palliative)				0.017
No	13	10	0	
Yes	45	9	9	
Curative resection				0.002
No	14	8	0	
Yes	67	17	17	
Type (among resected patient)				0.003
Intrahepatic	100	33	33	
Perihilar	67	17	17	
Distal	33	0	0	
Type (among all patient)				0.027
Intrahepatic	50	23	11	
Perihilar	18	6	3	
Distal	16	5	0	
Histology				0.032
Well	80	40	40	
Moderate and poor	57	0	0	
Margin involvement				0.013
Negative	86	29	29	
Positive	40	0	0	

0.005). Obstructive jaundice may decrease the concentrations of vitamin K-dependent coagulation factors, resulting in aberrant coagulation profiles^[5]. However, none of the other symptoms or blood parameters we assayed was significantly related to tumor location.

Many of our patients initially underwent ERCP, not MRCP. ERCP enables cytological brushing and can decompress the obstructed biliary system^[6]. Although we found that the success rate of internal stent placement for drainage was high, PTBD may also play a role, especially when the endoscopic approach has failed. PTBD can also be used to visualize proximal biliary tumors and anatomy^[5]. Failure of biliary decompression may result in an infected biliary system and further risks of liver failure and sepsis^[21]. These risks may be prevented by decompression of an obstructed biliary system in patients with potentially resectable cholangiocarcinoma, although a series by Figueras *et al*^[22] demonstrated no significant differences in morbidity and mortality between patients with and without preoperative biliary drainage. None of our patients who underwent hepatic resection developed postoperative liver failure.

Despite improved diagnostic methods and a relatively early presentation (median duration of symptoms, 30 d), only about one-third of our patients (32%) underwent surgery, and only 12 underwent curative resection. More than half of our patients (58%) were considered to have advanced unresectable disease, whereas the remaining

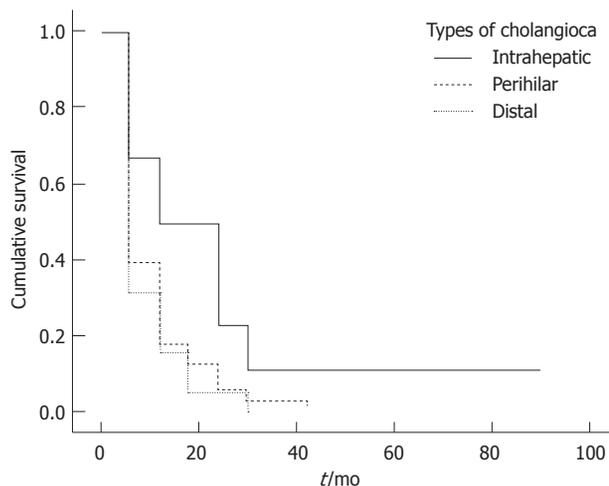


Figure 1 Overall survival by types of cholangiocarcinoma regardless of surgery or not, $P = 0.027$.

patients were considered unsuitable for surgery because of comorbidity and/or advanced age, findings similar to those observed previously^[7]. Our overall resectability, 55% (12/22), was similar to previously reported rates 18%-70%^[23]. Postoperative complications were not associated with tumor location. In high volume centers with considerable experience, the operative mortality and morbidity rates varied from 6%-14% and 32%-65%, respectively^[24-26]. Our 30-d postoperative mortality and morbidity rates were similar, 8% and 67%, respectively.

Patients who underwent surgical resection had a definite survival advantage over those who did not, confirming that surgical resection is the best treatment available for patients with cholangiocarcinoma and providing further evidence that potentially resectable patients should be referred early to a specialized surgical team^[7,10]. The median survival time for the 12 patients who underwent curative resection was 16 mo, compared with 3 mo for the 57 patients who did not undergo curative resection ($P = 0.002$). The 1-, 2- and 3-year cumulative survival rates in patients who underwent resection were 67%, 17% and 17%, respectively, significantly higher than the 14%, 8% and 0%, respectively, in those who did not. Furthermore, complete surgical resection with histologically negative margins offers the best chance for cure and long-term survival^[27-30].

Univariate log-rank analyses of tumor related variables in Table 4 showed that absence of jaundice ($P = 0.038$), tumor location ($P = 0.027$) (Figure 1)^[17], curative resection ($P = 0.002$) (Figure 2)^[31], early tumor stage ($P = 0.016$)^[32], negative margin status ($P = 0.013$) (Figure 3)^[17,32,33] and higher degree of tumor differentiation ($P = 0.032$) (Figure 4)^[17] were significant predictors of longer survival. In contrast to previous reports, regional lymph node metastasis^[28] and perineural invasion^[34] were not associated with survival in resected patients. Moreover, we found that age, gender, race and risk factors were not predictors of survival.

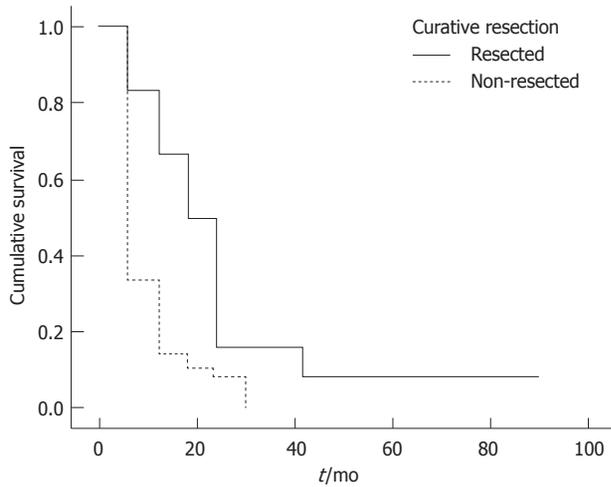


Figure 2 Cumulative survival curves after curative resection, $P = 0.002$.

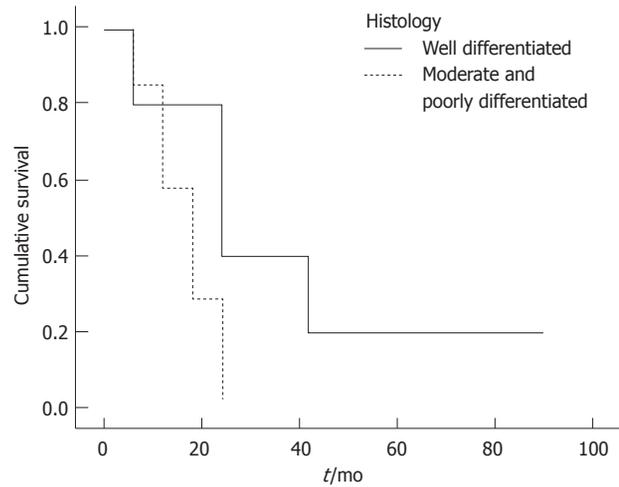


Figure 4 Overall survival according to degree of tumour differentiation, $P = 0.032$.

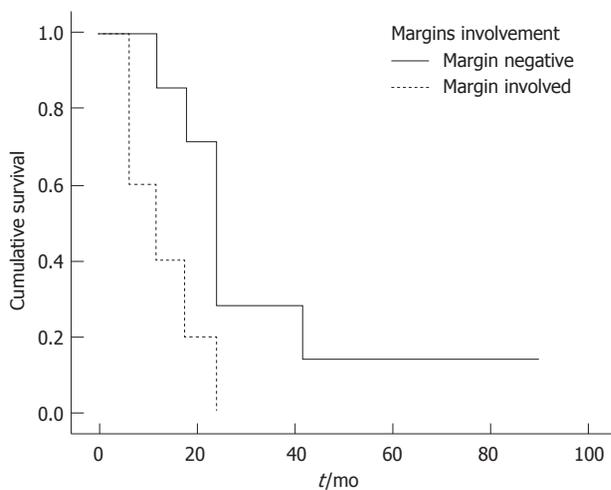


Figure 3 Cumulative survival curves after resection according to histologic margin clearance, $P = 0.013$.

Multivariate analysis showed that tumor location was the only independent predictor of long-term survival. Survival was significantly lower in patients with perihilar than in those with intrahepatic and distal tumors (HR = 0.016, 95% CI: 0.01-0.607; $P = 0.026$), in agreement with the findings of other series showing that patients with intrahepatic tumors had the best survival^[17,2]. This could be explained by the fact that, compared with extrahepatic tumors, intrahepatic tumors are characterized by different epidemiology and tumor biology, younger age ($P < 0.05$), and lower rate of tumor negative margins^[10,17].

For the majority of patients with cholangiocarcinoma who cannot undergo curative resection, palliative treatment to relieve jaundice, pruritus and cholangitis and to avoid liver failure becomes the priority. This can be achieved surgically *via* biliary-enteric bypass or stent placement *via* PTBD or ERCP^[33]. We found however, that neither palliative surgical bypass nor biliary drainage had significant survival benefits, in agreement with previous findings by Prat *et al*^[34] Both groups had a median

survival of 3 mo.

A novel palliative therapy for unresectable cholangiocarcinoma, photodynamic therapy (PDT), has shown promising benefits in terms of patient survival, cholestasis and quality of life^[35]. PDT utilizes the intravenous photosensitizer sodium porfimer, which accumulates in tumor tissues. Upon illumination of the tumor bed by a specific endoscopic light, the porfimer becomes activated and forms oxygen free radicals, resulting in tumor necrosis^[36].

The major limitation of our study was its involvement of patients at a single center. Therefore, our findings may not be representative of patients with cholangiocarcinoma throughout Malaysia.

The incidence of cholangiocarcinoma is increasing throughout Malaysia, as shown by the increase in the number of patients diagnosed per year throughout our study period. This may be due to an increased detection rate and to increases in referrals to our center. Curative surgical resection with clear histological margins of early stage well-differentiated intrahepatic tumors is associated with improved long-term survival. Further prospective randomized studies involving multiple centers are warranted.

ACKNOWLEDGMENTS

The authors would like to thank Dr. Aqel Mohd Dhafer of Universiti Teknologi MARA and Dr. Azlan Darus of the University of Malaya for their assistance with the data analysis.

COMMENTS

Background

Cholangiocarcinoma is a rare malignant cancer of the bile duct with poor prognosis. This cancer is difficult to diagnose and often the patient presented late when curative surgical resection that can provide the chance for cure, is not feasible. Any person in his or her fifth decade with jaundice and significant

weight loss should raise a suspicion of this cancer. Despite its rarity, the incidence of this cancer has been steadily increasing worldwide.

Research frontiers

Many studies have concurred that early detection together with radical surgery will increase the survival period of these patients. A research article to be published in the *World Journal of Gastroenterology* has further emphasized the importance of early diagnosis and early referral to a specialized surgical centre with vast experience in the management of this cancer.

Innovations and breakthroughs

The authors have analysed various factors that affect the survival of their cholangiocarcinoma patients treated over a 10-year period. It was found that the survival period will improve if complete tumour excision is performed whereby the margins are tumour-free on histology. The outcome is also favourable if surgery is performed early when the abnormal cells are still at their early grade and have not spread elsewhere. Despite a range of palliative procedures available for inoperable cancer, none could surpass the result of a successful surgical treatment. These have further consolidated similar findings and recommendations of early aggressive surgery from other larger centres in the West and Far East.

Applications

The study also shows that cholangiocarcinoma is an important health problem in Malaysia. Although the increase in number of patient being treated may not truly reflect a true increase in incidence, this would mean more need to be done to address such problem. This study provides a framework for future studies in Malaysia and hopefully stimulates other groundbreaking research in this region especially concerning the epidemiology and pathophysiology of this fatal cancer.

Peer review

This 10-year retrospective review defines treatment modalities and survival statistics for cholangiocarcinoma in a Malaysian Referral Hospital. Not surprisingly, patients who underwent R-O resections had higher cumulative survival rates than patients palliated surgically, endoscopically or by percutaneous transhepatic biliary drainage. This manuscript is worthy of publication.

REFERENCES

- 1 **Parker SL**, Tong T, Bolden S, Wingo PA. Cancer statistics, 1996. *CA Cancer J Clin* 1996; **46**: 5-27
- 2 **Khan SA**, Taylor-Robinson SD, Toledano MB, Beck A, Elliott P, Thomas HC. Changing international trends in mortality rates for liver, biliary and pancreatic tumours. *J Hepatol* 2002; **37**: 806-813
- 3 **Olnes MJ**, Erlich R. A review and update on cholangiocarcinoma. *Oncology* 2004; **66**: 167-179
- 4 **Carriaga MT**, Henson DE. Liver, gallbladder, extrahepatic bile ducts, and pancreas. *Cancer* 1995; **75**: 171-190
- 5 **Khan SA**, Davidson BR, Goldin R, Pereira SP, Rosenberg WM, Taylor-Robinson SD, Thillainayagam AV, Thomas HC, Thursz MR, Wasan H. Guidelines for the diagnosis and treatment of cholangiocarcinoma: consensus document. *Gut* 2002; **51** Suppl 6: VI1-VI9
- 6 **Khan SA**, Thomas HC, Davidson BR, Taylor-Robinson SD. Cholangiocarcinoma. *Lancet* 2005; **366**: 1303-1314
- 7 **Lazaridis KN**, Gores GJ. Cholangiocarcinoma. *Gastroenterology* 2005; **128**: 1655-1667
- 8 **Liu CL**, Fan ST, Lo CM, Tso WK, Lam CM, Wong J. Improved operative and survival outcomes of surgical treatment for hilar cholangiocarcinoma. *Br J Surg* 2006; **93**: 1488-1494
- 9 **Nakeeb A**, Pitt HA, Sohn TA, Coleman J, Abrams RA, Piantadosi S, Hruban RH, Lillemoe KD, Yeo CJ, Cameron JL. Cholangiocarcinoma. A spectrum of intrahepatic, perihilar, and distal tumors. *Ann Surg* 1996; **224**: 463-473; discussion 473-475
- 10 **Greene FL**. The American Joint Committee on Cancer: updating the strategies in cancer staging. *Bull Am Coll Surg* 2002; **87**: 13-15
- 11 **de Groen PC**, Gores GJ, LaRusso NF, Gunderson LL, Nargorney DM. Biliary tract cancers. *N Engl J Med* 1999; **341**: 1368-1378
- 12 **Singh P**, Patel T. Advances in the diagnosis, evaluation and management of cholangiocarcinoma. *Curr Opin Gastroenterol* 2006; **22**: 294-299
- 13 **DeOliveira ML**, Cunningham SC, Cameron JL, Kamangar F, Winter JM, Lillemoe KD, Choti MA, Yeo CJ, Schulick RD. Cholangiocarcinoma: thirty-one-year experience with 564 patients at a single institution. *Ann Surg* 2007; **245**: 755-762
- 14 **Chapman RW**. Risk factors for biliary tract carcinogenesis. *Ann Oncol* 1999; **10** Suppl 4: 308-311
- 15 **Chen MF**, Jan YY, Wang CS, Hwang TL, Jeng LB, Chen SC, Chen TJ. A reappraisal of cholangiocarcinoma in patient with hepatolithiasis. *Cancer* 1993; **71**: 2461-2465
- 16 **Donato F**, Gelatti U, Tagger A, Favret M, Ribero ML, Callea F, Martelli C, Savio A, Trevisi P, Nardi G. Intrahepatic cholangiocarcinoma and hepatitis C and B virus infection, alcohol intake, and hepatolithiasis: a case-control study in Italy. *Cancer Causes Control* 2001; **12**: 959-964
- 17 **Parkin DM**, Srivatanakul P, Khlut M, Chenvidhya D, Chotivan P, Insiripong S, L'Abbé KA, Wild CP. Liver cancer in Thailand. I. A case-control study of cholangiocarcinoma. *Int J Cancer* 1991; **48**: 323-328
- 18 **Bergquist A**, Glaumann H, Persson B, Broomé U. Risk factors and clinical presentation of hepatobiliary carcinoma in patients with primary sclerosing cholangitis: a case-control study. *Hepatology* 1998; **27**: 311-316
- 19 **Welzel TM**, Graubard BI, El-Serag HB, Shaib YH, Hsing AW, Davila JA, McGlynn KA. Risk factors for intrahepatic and extrahepatic cholangiocarcinoma in the United States: a population-based case-control study. *Clin Gastroenterol Hepatol* 2007; **5**: 1221-1228
- 20 **Alexopoulou A**, Soultati A, Dourakis SP, Vasilieva L, Archimandritis AJ. Cholangiocarcinoma: a 7-year experience at a single center in Greece. *World J Gastroenterol* 2008; **14**: 6213-6217
- 21 **Anderson CD**, Pinson CW, Berlin J, Chari RS. Diagnosis and treatment of cholangiocarcinoma. *Oncologist* 2004; **9**: 43-57
- 22 **Figueras J**, Llado L, Valls C, Serrano T, Ramos E, Fabregat J, Rafecas A, Torras J, Jaurrieta E. Changing strategies in diagnosis and management of hilar cholangiocarcinoma. *Liver Transpl* 2000; **6**: 786-794
- 23 **Hammill CW**, Wong LL. Intrahepatic cholangiocarcinoma: a malignancy of increasing importance. *J Am Coll Surg* 2008; **207**: 594-603
- 24 **Nimura Y**, Kamiya J, Kondo S, Nagino M, Uesaka K, Oda K, Sano T, Yamamoto H, Hayakawa N. Aggressive preoperative management and extended surgery for hilar cholangiocarcinoma: Nagoya experience. *J Hepatobiliary Pancreat Surg* 2000; **7**: 155-162
- 25 **Lillemoe KD**, Cameron JL. Surgery for hilar cholangiocarcinoma: the Johns Hopkins approach. *J Hepatobiliary Pancreat Surg* 2000; **7**: 115-121
- 26 **Jarnagin WR**, Fong Y, DeMatteo RP, Gonen M, Burke EC, Bodniewicz BS J, Youssef BA M, Klimstra D, Blumgart LH. Staging, resectability, and outcome in 225 patients with hilar cholangiocarcinoma. *Ann Surg* 2001; **234**: 507-517; discussion 517-519
- 27 **Jarnagin WR**. Cholangiocarcinoma of the extrahepatic bile ducts. *Semin Surg Oncol* 2000; **19**: 156-176
- 28 **Su CH**, Tsay SH, Wu CC, Shyr YM, King KL, Lee CH, Lui WY, Liu TJ, P'eng FK. Factors influencing postoperative morbidity, mortality, and survival after resection for hilar cholangiocarcinoma. *Ann Surg* 1996; **223**: 384-394
- 29 **Silva MA**, Tekin K, Aytekin F, Bramhall SR, Buckels JA, Mirza DF. Surgery for hilar cholangiocarcinoma; a 10 year experience of a tertiary referral centre in the UK. *Eur J Surg Oncol* 2005; **31**: 533-539

- 30 **Lai EC**, Lau WY. Aggressive surgical resection for hilar cholangiocarcinoma. *ANZ J Surg* 2005; **75**: 981-985
- 31 **Bhuiya MR**, Nimura Y, Kamiya J, Kondo S, Fukata S, Hayakawa N, Shionoya S. Clinicopathologic studies on perineural invasion of bile duct carcinoma. *Ann Surg* 1992; **215**: 344-349
- 32 **Madariaga JR**, Iwatsuki S, Todo S, Lee RG, Irish W, Starzl TE. Liver resection for hilar and peripheral cholangiocarcinomas: a study of 62 cases. *Ann Surg* 1998; **227**: 70-79
- 33 **Abu-Hamda EM**, Baron TH. Endoscopic management of cholangiocarcinoma. *Semin Liver Dis* 2004; **24**: 165-175
- 34 **Prat F**, Chapat O, Ducot B, Ponchon T, Fritsch J, Choury AD, Pelletier G, Buffet C. Predictive factors for survival of patients with inoperable malignant distal biliary strictures: a practical management guideline. *Gut* 1998; **42**: 76-80
- 35 **Ortner ME**, Caca K, Berr F, Liebetruh J, Mansmann U, Huster D, Voderholzer W, Schachschal G, Mössner J, Lochs H. Successful photodynamic therapy for nonresectable cholangiocarcinoma: a randomized prospective study. *Gastroenterology* 2003; **125**: 1355-1363
- 36 **Berr F**. Photodynamic therapy for cholangiocarcinoma. *Semin Liver Dis* 2004; **24**: 177-187

S- Editor Sun H L- Editor Ma JY E- Editor Li JY

Thalidomide-based multidisciplinary treatment for patients with advanced hepatocellular carcinoma: A retrospective analysis

Yang-Yuan Chen, Hsu-Heng Yen, Kun-Ching Chou, Shun-Sheng Wu

Yang-Yuan Chen, Department of Gastroenterology, China Medical University Hospital, China Medical University, Taichung, 40000 Taiwan, China

Hsu-Heng Yen, Kun-Ching Chou, Shun-Sheng Wu, Department of Gastroenterology, Changhua Christian Medical Center, Changhua, 50000 Taiwan, China

Author contributions: Chou KC designed the study; Wu SS analyzed the data; Yen HH wrote the manuscript; Chen YY wrote and gave final approval to this manuscript.

Correspondence to: Yang-Yuan Chen, MD, Department of Gastroenterology, China Medical University Hospital, China Medical University, 2 Yude road, Taichung, 40000 Taiwan, China. ychen02@gmail.com

Telephone: +886-4-7359253 Fax: +886-4-7228289

Received: March 23, 2011 Revised: May 27, 2011

Accepted: June 3, 2011

Published online: February 7, 2012

Abstract

AIM: To evaluate the efficacy of thalidomide in combination with other therapies to treat patients with advanced hepatocellular carcinoma (HCC).

METHODS: We performed a retrospective analysis of all patients with HCC who were treated with thalidomide for at least two months. The medical records of patients with HCC who were treated at our institution between April 2003 and March 2008 were reviewed. Image studies performed before and after treatment, tumor response, overall survival, and the decrease in α -fetoprotein (AFP) levels were evaluated.

RESULTS: A total of 53 patients with HCC received either 100 or 200 mg/d of thalidomide. The patient population consisted of 9 women and 44 men with a median age of 61 years. Thirty patients (56.6%) were classified as Child-Pugh A, and 12 patients (22.6%) were classified as Child-Pugh B. Twenty-six patients had portal vein thrombosis (49.1%), and 25 patients had extrahepatic metastasis (47.1%). The median duration

of thalidomide treatment was 6.0 mo. Six of the 53 patients achieved a confirmed response (11.3%), one achieved a complete response (1.9%) and 5 achieved a partial response (9.4%). The disease control rate (CR + PR + SD) was 28.3% (95% CI: 17.8-42.4), and the median overall survival rate was 10.5 mo. The 1- and 2-year survival rates were 45% and 20%, respectively. Only one complete response patient showed an improved overall survival rate of 66.8 mo. Sixteen patients (30.2%) showed more than a 50% decrease in their serum AFP levels from baseline, indicating a better response rate (31.3%), disease control rate (43.8%), and overall survival time (20.7 mo). The therapy was well tolerated, and no significant toxicities were observed.

CONCLUSION: Thalidomide was found to be safe for advanced HCC patients, demonstrating anti-tumor activity including response, survival, and AFP decreases of greater than 50% from baseline.

© 2012 Baishideng. All rights reserved.

Key words: Thalidomide; Hepatocellular carcinoma

Peer reviewer: Dr. Thomas Kietzmann, Professor, Department of Biochemistry, University of Oulu, Oulu 90014, Finland

Chen YY, Yen HH, Chou KC, Wu SS. Thalidomide-based multidisciplinary treatment for patients with advanced hepatocellular carcinoma: A retrospective analysis. *World J Gastroenterol* 2012; 18(5): 466-471 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/466.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.466>

INTRODUCTION

Hepatocellular carcinoma (HCC) is the most frequent primary liver cancer, the fifth most common malignancy worldwide (with over 700 000 new cases per year), and the third most common cause of cancer deaths^[1]. In

Taiwan, HCC, which ranks second among the major types of cancer in the list of cancer-related mortalities, is responsible for approximately 7000 to 8000 deaths per year^[2]. Unfortunately, most patients seek treatment when the disease is beyond curative treatment (surgery or percutaneous ablation), and palliative care is the only alternative. According to the Barcelona Clinic Liver Cancer (BCLC) staging classification^[1] and treatment schedule, chemoembolization is the best option for intermediate-stage patients. However, for advanced-stage patients, no standard treatment was established until 2007. Systemic chemotherapy is generally ineffective and is associated with significant toxicity because hepatic function is often impaired by underlying cirrhosis that is often accompanied by hypersplenism and peripheral cytopenia^[3]. Fortunately, after the positive results of the Study of Heart and Renal Protection (SHARP) trials^[4], a new treatment, sorafenib, was approved for advanced-stage patients, which offers major improvements in overall survival and time to progression compared to placebo. There are many new modalities of treatment with more favorable therapeutic indices that are suitable for patients with advanced HCC. HCC is a hypervascular tumor that is one of the most antiangiogenic and angiogenesis-dependent tumors^[5,6]. Consequently, it is reasonable to hypothesize that antiangiogenesis therapy may inhibit the growth of HCC. A number of antiangiogenic agents have been developed, including thalidomide, which is a glutamic acid derivative that was first described in 1953 when it was labeled as a sedative and anti-emetic agent. However, it was withdrawn from the European market 30 years ago because of its teratogenic effects^[7]. Recently, oral thalidomide has been shown to inhibit basic fibroblast growth factor- and vascular endothelial growth factor-induced angiogenesis of cancer cells^[8,9]. Studies published on the efficacy of thalidomide in advanced hepatocellular carcinoma have reported modest responses to therapy with acceptable toxicity^[10-12]. Treatment of patients with HCC continues to present a major challenge. We retrospectively analyzed our records of HCC patients who received thalidomide in combination with other therapies to determine whether thalidomide was effective.

MATERIALS AND METHODS

Patients

Between April 2003 and March 2008, 53 patients with HCC were treated for at least two months with either 100 or 200 mg/d of thalidomide (50 mg/capsule, TTY Biopharm Co. Ltd., Taipei, Taiwan) at Changhua Christian Medical Center in Taiwan. HCC was diagnosed by histological examination and imaging findings. The diagnosis of HCC was confirmed by histological examination or the presence of all of the following criteria: (1) pathological diagnosis of HCC; (2) cirrhotic liver with a tumor size greater than 2 cm plus one dynamic image [computed tomography (CT) or magnetic resonance image (MRI)] or alpha fetoprotein (AFP) > 200 ng/mL;

(3) cirrhotic liver with a tumor size of 1-2 cm plus two dynamic images (CT + MRI); and (4) non-cirrhotic liver greater than 2 cm plus one dynamic image (CT or MRI) and AFP > 200 ng/mL. The inclusion criteria were as follows: (1) advanced HCC (surgically unresectable); (2) failed previous local therapy, such as radiotherapy, hepatic arterial chemoembolization, radiofrequency ablation, or percutaneous interventional therapy; and (3) distant metastasis (lung, lymph node, or bone) that is not eligible for curative surgery and radiotherapy or locoregional therapy failure [e.g., transarterial chemoembolization (TACE), recurrence-free interval or percutaneous ethanol injection (PEI)]. All patients had bidimensionally measurable disease that was staged by the pathological tumor-node-metastasis (TNM) system, the Okuda system, and the BCLC parameters for HCC. The demographic data, details of the primary tumors, serum AFP levels, dates of recurrence, length of survival, and last follow-up dates were analyzed retrospectively. The responses to thalidomide were determined by CT performed according to the Response Evaluation Criteria in Solid Tumor Guidelines^[13], and the AFP levels were also analyzed before and after thalidomide treatment. Overall survival was calculated from the date of the start of chemotherapy and analyzed by the Kaplan-Meier method. Follow-up data were obtained for all patients until the time of their death or the last follow-up.

RESULTS

Patient and demographic characteristics

A total of 53 patients with HCC were available for analysis, and their demographic characteristics are shown in Table 1. The patient population included 9 females (17.0%) and 44 males (83.0%) with a median age of 61 years (range, 29-88 years). Of the 53 patients, 10 had not received prior treatment or therapy. Pretreatment curative surgery had been performed on 12 patients (22.6%), transarterial embolization on 13 patients (24.5%), TACE on 16 patients (30.2%), radio frequency ablation on 10 patients (18.9%), and radiotherapy (RT) on 10 patients (18.9%). Twenty-six patients had portal vein thrombosis (49.1%), and 25 patients had extrahepatic metastasis (47.2%). The prevalence of hepatitis B was 56.5% (30/53), that of hepatitis C was 37.7% (20/53), and that of concomitant hepatitis was 1.9% (1/53). Of the 53 patients, most patients had TNM stage IV (45.3%), Okuda stage I (51.9%), and BCLC stage C (71.2%). There were 22 patients (41.5%) whose serum AFP levels were greater than 400 ng/mL above the baseline. The liver functions of the majority of patients were classified as Child-Pugh A (56.6%), and the median duration of treatment was 6.0 mo (range, 1.5-53.9 mo) (Table 1).

Efficacy

Of the 53 patients, one had a complete response (CR, 2.9%) to thalidomide, five had a partial response (PR, 9.4%) and nine were classified as stable disease (SD,

Table 1 The clinical characteristics of 53 patients with hepatocellular carcinoma

Characteristic	n (%)
Age, yr (median)	61 (range, 29-88)
Sex	
Male	44 (83.0)
Female	9 (17.0)
Type of hepatitis	
Hepatitis B	30 (56.6)
Hepatitis C	20 (37.7)
Hepatitis B + C	1 (1.9)
Child-Pugh classification	
A	30 (56.6)
B	12 (22.6)
C	1 (1.8)
TNM stage	
I	0 (0)
II	6 (11.2)
III A	11 (20.8)
III B	3 (5.7)
III C	6 (11.2)
IV	24 (45.3)
Okuda stage	
I	27 (51.9)
II	16 (30.8)
III	1 (1.9)
BCLC stage	
A	2 (3.9)
B	7 (13.5)
C	37 (71.2)
D	1 (1.9)
Extrahepatic metastasis	
Yes	25 (47.2)
No	28 (52.8)
Portal vein thrombosis	
Yes	26 (49.1)
No	26 (49.1)
Unknown	1 (1.8%)
Site of extrahepatic metastasis	
Lung	11 (21.2)
Bone	6 (11.5)
Brain	1 (1.9)
Others	7 (13.5)
Prior therapy	
Surgery	12 (22.6)
TACE	16 (30.2)
TAE	13 (24.5)
Radiation therapy	10 (18.9)
RFA	10 (18.9)
No therapy	10 (18.9)
Duration of treatment, mo	
Median	6.0 (range, 1.5-53.9)
AFP level	
> 400 ng/mL	31 (58.5)
< 400 ng/mL	22 (41.5)

BCLC: Barcelona Clinic Liver Cancer; TACE: Transarterial chemoembolization; TAE: Transarterial embolization; RFA: Radiofrequency ablation; AFP: Alpha fetoprotein; TNM: Tumor node metastasis.

17.0%). The remaining 38 patients had disease that continued to progress after the thalidomide treatments. The objective response rate was 11.3% (95% CI: 4.3-23.0), and the disease control rate (CR + PR + SD) was 28.3% (95% CI: 17.8-42.4). The median overall survival rate was 10.5 mo (95% CI: 6.9-23.3). The 1- and 2-year sur-

Table 2 Efficacy results of thalidomide

Overall objective response, n = 53, (%)	
CR	1 (2.9)
PR	5 (9.4)
SD	9 (17.0)
PD	38 (44.1)
Objective response rate	6 (11.3), 95% CI: 4.3-23.0
Disease control rate	15 (28.3), 95% CI: 17.8-42.4
Overall survival, mo	
Median	10.5, 95% CI: 6.9-23.3
1-year survival	(45)
2-year survival	(20)
A decrease in AFP > 50% after treatment	
Yes	16 (30.2)
No	37 (69.8)

CR: Complete response; PR: Partial response; SD: Stable disease; PD: Progressive disease; AFP: Alpha fetoprotein.

vival rates were 45% and 20%, respectively. Sixteen patients (30.2%) showed more than a 50% decrease in their serum AFP levels below the baseline and showed a better response rate (31.3%), disease control rate (43.8%), and overall survival time (20.7 mo) (Table 2, Figure 1). The prognostic factors for the response rate, disease control rate, and overall survival in HCC patients receiving thalidomide are listed in Table 3. Multivariate analysis showed that almost all of these patients qualified as having independent prognostic factors for the efficacy analysis. The only significant difference in the efficacy activity was an AFP decrease of > 50% after treatment. The median overall survival time of the patients who registered > 50% AFP decrease was 20.7 mo, with a response rate of 31.3% and a disease control rate of 43.8%. The median overall survival time of those patients with a < 50% AFP decrease was 7.1 mo, with a response rate of 2.7% and a disease control rate of 21.6% (Table 3, Figure 1). Table 4 is a comparison of the patients who responded and the patients whose disease progressed. Patients in the CR + PR + SD group had a significantly longer survival time (33.3 mo) than those in the progressive disease (PD) group (6.9 mo, $P < 0.003$) (Figure 1).

DISCUSSION

Thalidomide has been used in the treatment of advanced HCC patients. Hsu *et al.*^[10] reported an overall response rate of 6.3% with an overall survival time of 18.7 wk when an escalating dose (100-600 mg/d) of thalidomide was used for the treatment of advanced HCC. Patt *et al.*^[14] also showed a 5% overall response rate with a 6.8-mo overall survival time when a high dose (400-1000 mg/d) of thalidomide was used. In a phase II study^[12], high-dose (200-800 mg/d) single-agent thalidomide demonstrated a response rate of 3.9% with an overall survival time of 123 d. The first retrospective study to analyze the efficacy and tolerability of fixed low-dose thalidomide in the treatment of advanced HCC patients^[15]

Table 3 Prognostic factors for efficacy analysis in hepatocellular carcinoma patients receiving thalidomide

Variables		P value
Overall response rate n (%)		
Child-Pugh classification		
A	3/30 (10.0)	1.000 ¹
B and C	3/23 (13.0)	
Okuda staging		
Stage 1	2/27 (7.4)	0.344 ¹
Stage 2	3/16 (18.8)	
AFP level		
> 400 ng/mL	1/31 (3.2)	0.071 ¹
< 400 ng/mL	5/22 (22.7)	
A decrease in AFP > 50% after treatment		
Yes	5/16 (31.3)	0.007 ¹
No	1/37 (2.7)	
Disease control rate n (%)		
Child-Pugh Classification		
A	6/30 (20.0)	0.218 ¹
B and C	9/23 (39.1)	
Okuda staging		
Stage 1	6/27 (22.2)	0.719 ¹
Stage 2	5/16 (31.3)	
AFP level		
> 400 ng/mL	7/31 (22.6)	0.357 ¹
< 400 ng/mL	8/22 (36.4)	
A decrease in AFP > 50% after treatment		
Yes	7/16 (43.8)	0.071 ¹
No	8/37 (21.6)	
Overall survival, mo		
Child-Pugh Classification		
A	8.8	0.922 ²
B and C	10.8	
Okuda staging		
Stage 1	22.2	0.075 ²
Stage 2	6.9	
AFP level		
> 400 ng/mL	10.8	0.679 ²
< 400 ng/mL	6.5	
A decrease in AFP > 50% after treatment		
Yes	20.7 (95% CI: 1.7-NA)	0.307 ²
No	7.1 (95% CI: 6.3-24.3)	

¹P value was calculated by Fisher's exact test; ²P value was calculated by Log-rank test. AFP: Alpha fetoprotein; NA: Not assessable.

showed that low-dose thalidomide has a comparable single-agent activity (response rate of 5%, with an overall survival time of 4.3 mo) but fewer treatment-related toxicities than high-dose thalidomide when treating advanced HCC patients. Patients treated with low-dose thalidomide have similar overall survival times compared to patients treated with chemotherapeutic agents, with a far better toxicity profile and less hematological toxicity (no grade 3/4 neutropenia or thrombocytopenia)^[15,16]. The largest randomized phase III trial for HCC (the SHARP trial) showed better progression free survival and overall survival times with sorafenib than with placebo^[4]. The primary drug-related adverse events were dermatological (constitutional and hand-foot skin reactions) and gastrointestinal^[4,17]. The toxicity of sorafenib is a serious problem because approximately 50% of the patients had to interrupt or stop their treatment because of sorafenib-induced toxicity. The tolerance of low-dose thalidomide in HCC patients may be worth further investigation.

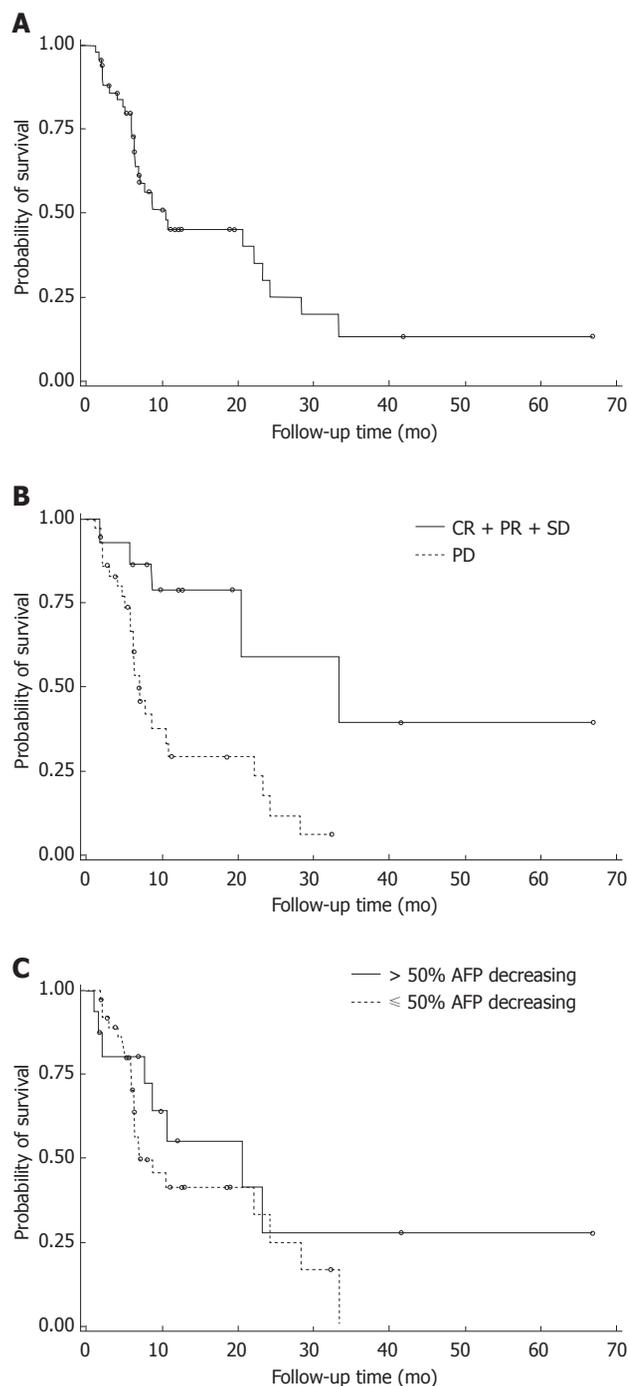


Figure 1 Kaplan-Meier analysis of the survival time in all advanced hepatocellular carcinoma patients (A), in the subgroup of disease stabilization (B), and in the subgroup of > 50% decrease in alpha fetoprotein (C). CR: Complete response; PR: Partial response; SD: Stable disease; PD: Progressive disease; AFP: Alpha fetoprotein.

The treatment of hepatoma with thalidomide appears to be feasible. A complete response was rare with thalidomide treatment of HCC; the PR rate was 5%-10%, and the SD rate was approximately 37%^[10,12,14], depending on the duration of observation, cancer stage, and the definition of stability. In our study, one patient had complete remission; the PR rate was 9.4%, and the SD rate was 17%. One CR patient received thalidomide alone after a TACE therapy failure; the duration of the treatment

Table 4 Comparison of patients who responded and patients with progressive disease

Characteristic	CR	PR	SD	PD	CR + PR + SD	P value ^a
AFP level						0.357
> 400 ng/mL	0 (0)	1 (3.2)	6 (19.4)	24 (77.4)	7 (22.6)	
< 400 ng/mL	1 (4.6)	4 (18.2)	3 (13.6)	14 (63.6)	8 (36.4)	
A decrease in AFP > 50% after treatment	1 (6.3)	4 (25.0)	2 (12.5)	9 (56.3)	7 (43.8)	0.182
Overall survival, mo	66.8	NA	20.7 (95% CI: 20.7-33.3)	6.9 (95% CI: 6.3-10.8)	33.3 (95% CI: 20.7-NA)	0.003 ^b

AFP: Alpha fetoprotein; CR: Complete response; PR: Partial response; SD: Stable disease; PD: Progressive disease; NA: Not assessable. ^a $P < 0.05$ between the two groups (CR + PR + SD vs PD); ^b $P = 0.0003$ (less than 0.005) between progress disease and CR + PR + SD.

was 53.9 mo, the patient had no recurrence, and he is still alive (66.8 mo post-treatment). The most interesting finding was the AFP decrease from 11 005.3 ng/mL at diagnosis to < 20 ng/mL (Table 4). Of the 5 patients with partial responses, 2 had prior TACE treatments, 2 had RT, 1 had PEI and 1 had systemic therapy. The median survival time among these patients was 502 d (range, 248-1263 d), and 3 of them are still alive. The median survival time of patients with stable disease was 412 d (range, 60-1013 d).

Patients in the CR + PR + SD group had a significantly longer survival time (33.3 mo) than those in the PD group (6.9 mo, $P < 0.003$). Thalidomide may offer HCC stabilization and prolong survival, especially in patients with stabilization. Survival time should be the focus of future clinical trials of thalidomide therapy. In this study, we evaluated the clinical implication of the AFP tumor marker response in assessing the therapeutic effects of thalidomide in HCC. The results showed that the AFP response was an independent prognostic factor for the response rate, disease control rate, and overall survival time. We also identified patients with more than or less than a 50% decrease in serum AFP levels from the baseline, which made a significant difference in their response rates (31.3% vs 2.7%, $P = 0.007$). There was a better trend in the disease control rate (43.8%) and overall survival time (20.7 mo) when there was greater than a 50% AFP decrease (Table 3). The AFP response may correlate with the biological response and, consequently, predict the survival benefits of thalidomide in HCC.

In conclusion, thalidomide has shown modest clinical activity, including response and survival, and was safely administered to patients with advanced HCC. Because the present study is retrospective in nature with a relatively small number of patients, a larger, randomized phase II/III study is needed to clearly define the role of single-agent thalidomide in the treatment of HCC as an alternative to the expensive molecular-targeted therapies.

COMMENTS

Background

Hepatocellular carcinoma (HCC) is the most frequent primary liver cancer, the fifth most common malignancy worldwide (with over 700 000 new cases per year), and the third most common cause of cancer deaths. However, for advanced-stage patients, no standard treatment was established until the positive result of the Study of Heart and Renal Protection study. However, there are many new modalities of treatment with more favorable therapeutic indices that

are suitable for patients with advanced HCC.

Research frontiers

HCC is a hypervascular tumor that is one of the most antiangiogenic and angiogenesis-dependent tumors. Recently, thalidomide was shown to inhibit the angiogenesis of cancer cells and studies have reported modest responses to this therapy in advanced HCC. The authors retrospectively analyzed the records of HCC patients who received thalidomide in combination with other therapies to determine whether thalidomide was effective.

Innovations and breakthroughs

Studies published on the efficacy of thalidomide in advanced hepatocellular carcinoma have reported modest responses to the therapy with acceptable toxicity. Some of them highlighted the alpha fetoprotein (AFP) tumor marker response in assessing the therapeutic effects of thalidomide in HCC. In this study, the authors concluded thalidomide showed modest clinical activity, including response and survival, and was safely administered to patients with advanced HCC. Furthermore, they also identified patients with more than or less than a 50% decrease in serum AFP levels from the baseline, which made a significant difference in their response rates.

Applications

The results showed that the AFP response was an independent prognostic factor for the response rate, disease control rate, and overall survival time. The AFP response may correlate with the biological response and, consequently, predict the survival benefits of thalidomide in HCC.

Peer review

This is an interesting and well written manuscript summarising the effects of thalidomide on HCC patients in a retrospective study.

REFERENCES

- Llovet JM, Burroughs A, Bruix J. Hepatocellular carcinoma. *Lancet* 2003; **362**: 1907-1917
- Taiwan cancer registration report 2007. Available from: URL: <http://cph.ntu.edu.tw/>
- Venook AP. Treatment of hepatocellular carcinoma: too many options? *J Clin Oncol* 1994; **12**: 1323-1334
- Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, de Oliveira AC, Santoro A, Raoul JL, Forner A, Schwartz M, Porta C, Zeuzem S, Bolondi L, Greten TF, Galle PR, Seitz JF, Borbath I, Häussinger D, Giannaris T, Shan M, Moscovici M, Voliotis D, Bruix J. Sorafenib in advanced hepatocellular carcinoma. *N Engl J Med* 2008; **359**: 378-390
- Ikeda K, Saitoh S, Koida I, Tsubota A, Arase Y, Chayama K, Kumada H. Diagnosis and follow-up of small hepatocellular carcinoma with selective intraarterial digital subtraction angiography. *Hepatology* 1993; **17**: 1003-1007
- Yamaguchi R, Yano H, Iemura A, Ogasawara S, Haramaki M, Kojiro M. Expression of vascular endothelial growth factor in human hepatocellular carcinoma. *Hepatology* 1998; **28**: 68-77
- McBride WG. Thalidomide and congenital abnormalities. *Lancet* 1961; **2**: 1358
- D'Amato RJ, Loughnan MS, Flynn E, Folkman J. Thalidomide is an inhibitor of angiogenesis. *Proc Natl Acad Sci USA* 1994; **91**: 4082-4085
- Kruse FE, Jousen AM, Rohrschneider K, Becker MD, Völcker HE. Thalidomide inhibits corneal angiogenesis in

- duced by vascular endothelial growth factor. *Graefes Arch Clin Exp Ophthalmol* 1998; **236**: 461-466
- 10 **Hsu C**, Chen CN, Chen LT, Wu CY, Yang PM, Lai MY, Lee PH, Cheng AL. Low-dose thalidomide treatment for advanced hepatocellular carcinoma. *Oncology* 2003; **65**: 242-249
 - 11 **Wang TE**, Kao CR, Lin SC, Chang WH, Chu CH, Lin J, Hsieh RK. Salvage therapy for hepatocellular carcinoma with thalidomide. *World J Gastroenterol* 2004; **10**: 649-653
 - 12 **Lin AY**, Brophy N, Fisher GA, So S, Biggs C, Yock TI, Levitt L. Phase II study of thalidomide in patients with unresectable hepatocellular carcinoma. *Cancer* 2005; **103**: 119-125
 - 13 **Therasse P**, Arbuck SG, Eisenhauer EA, Wanders J, Kaplan RS, Rubinstein L, Verweij J, Van Glabbeke M, van Oosterom AT, Christian MC, Gwyther SG. New guidelines to evaluate the response to treatment in solid tumors. European Organization for Research and Treatment of Cancer, National Cancer Institute of the United States, National Cancer Institute of Canada. *J Natl Cancer Inst* 2000; **92**: 205-216
 - 14 **Patt YZ**, Hassan MM, Lozano RD, Nooka AK, Schnirer II, Zeldis JB, Abbruzzese JL, Brown TD. Thalidomide in the treatment of patients with hepatocellular carcinoma: a phase II trial. *Cancer* 2005; **103**: 749-755
 - 15 **Yau T**, Chan P, Wong H, Ng KK, Chok SH, Cheung TT, Lam V, Epstein RJ, Fan ST, Poon RT. Efficacy and tolerability of low-dose thalidomide as first-line systemic treatment of patients with advanced hepatocellular carcinoma. *Oncology* 2007; **72** Suppl 1: 67-71
 - 16 **Chuah B**, Lim R, Boyer M, Ong AB, Wong SW, Kong HL, Millward M, Clarke S, Goh BC. Multi-centre phase II trial of Thalidomide in the treatment of unresectable hepatocellular carcinoma. *Acta Oncol* 2007; **46**: 234-238
 - 17 **Cheng AL**, Kang YK, Chen Z, Tsao CJ, Qin S, Kim JS, Luo R, Feng J, Ye S, Yang TS, Xu J, Sun Y, Liang H, Liu J, Wang J, Tak WY, Pan H, Burock K, Zou J, Voliotis D, Guan Z. Efficacy and safety of sorafenib in patients in the Asia-Pacific region with advanced hepatocellular carcinoma: a phase III randomised, double-blind, placebo-controlled trial. *Lancet Oncol* 2009; **10**: 25-34

S- Editor Tian L L- Editor O'Neill M E- Editor Li JY

A pharmacodynamic model of portal hypertension in isolated perfused rat liver

Tao Zhang, Xue-Yan Xu, Hang Zhou, Xin Zhao, Meng Song, Tao-Tao Zhang, He Yin, Ting Li, Peng-Tao Li, Da-Yong Cai

Tao Zhang, Xue-Yan Xu, Hang Zhou, Xin Zhao, Da-Yong Cai, Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences, Peking Union Medical College, Beijing 100193, China

Meng Song, Tao-Tao Zhang, He Yin, Ting Li, Peng-Tao Li, Beijing University of Chinese Medicine, Beijing 100029, China

Author contributions: Zhang T, Xu XY, Zhou H, Zhao X, Song M, Zhang TT, Yin H and Li T took part in the experiments; Cai DY and Li PT designed the study and wrote the manuscript.

Supported by The Major State Creative New Drug Project, No. 2009ZX09502-017; Education Ministry Science Foundation of China, No. 108019

Correspondence to: Dr. Da-Yong Cai, Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences, Peking Union Medical College, Beijing 100193, China. dycal@implad.ac.cn

Telephone: +86-10-62899771 Fax: +86-10-62899715

Received: April 22, 2011 Revised: June 30, 2011

Accepted: July 7, 2011

Published online: February 7, 2012

Abstract

AIM: To develop a pharmacodynamic model of portal hypertension from chronic hepatitis.

METHODS: Pathological changes and collagen depositions were analyzed using morphometry to confirm CCl₄-induced chronic hepatitis. At d₀, d₂₈, d₅₆ and d₈₄ of the process, the portal perfused velocities ($\mu\text{L}/\text{min}$) in isolated rat livers were exactly controlled with a quantified pump. The pressure (mmHg) was monitored with a Physiological System. The geometric concentrations of phenylephrine or acetylcholine were added to a fixed volume (300 mL) of the circulating perfusate. The equation, the median effective concentration and its 95% confidence intervals of phenylephrine or acetylcholine were regressed with Prism-4 software in non-

linear fit and various slopes. In the isolated perfused rat livers with chronic hepatitis, both median effective concentrations were defined as the pharmacodynamic model of portal hypertension.

RESULTS: At d₀, d₂₈, d₅₆ and d₈₄, the equations of portal pressure potency from the concentrations of phenylephrine used to constrict the portal vein in isolated perfused rat livers were $Y = 0.1732 + 0.3970/[1 + 10^{-(4.3061-0.4407 X)}]$, $Y = -0.004934 + 0.12113/[1 + 10^{-(3.1247-0.3262 X)}]$, $Y = 0.0104 + 0.2643/[1 + 10^{-(8.8462-0.9579 X)}]$, and $Y = 0.01603 + 0.12107/[1 + 10^{-(5.1134-0.563 X)}]$; the median effective concentrations were 1.69×10^{-10} mol/L, 2.64×10^{-10} mol/L, 5.82×10^{-10} mol/L, and 8.24×10^{-10} mol/L, respectively. The equations from the concentrations of acetylcholine used to relax the portal vein were $Y = -0.4548 + 0.3274/[1 + 10^{-(6.1538 + 0.5554 X)}]$, $Y = -0.05391 + 0.06424/[1 + 10^{-(3.8541 + 0.3469 X)}]$, $Y = -0.2733 + 0.22978/[1 + 10^{-(3.0472 + 0.3008 X)}]$, and $Y = -0.0559 + 0.053178/[1 + 10^{-(5.6336 + 0.5883 X)}]$; the median effective concentrations were 8.40×10^{-10} mol/L, 7.73×10^{-12} mol/L, 5.98×10^{-11} mol/L, and 2.66×10^{-10} mol/L, respectively.

CONCLUSION: A pharmacodynamic model of portal hypertension in isolated perfused rat livers with chronic hepatitis was defined as the median effective concentrations of phenylephrine and acetylcholine.

© 2012 Baishideng. All rights reserved.

Key words: Chronic hepatitis; Isolated portal perfused rat liver; Pharmacodynamic model; Portal hypertension

Peer reviewers: Dr. Orhan Sezgin, Professor, Gastroenteroloji Bilim Dalı, Mersin Üniversitesi Tıp Fakültesi, Mersin 33190, Turkey; Yoshiaki Iwasaki, MD, PhD, Associate Professor, Health Service Center, Okayama University, 2-1-1, Tsushima-Naka, Kita-ku, Okayama 700-8530, Japan

Zhang T, Xu XY, Zhou H, Zhao X, Song M, Zhang TT, Yin H, Li T, Li PT, Cai DY. A pharmacodynamic model of portal hypertension in isolated perfused rat liver. *World J Gastroenterol* 2012; 18(5): 472-478 Available from: URL: <http://www.wjg-net.com/1007-9327/full/v18/i5/472.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.472>

INTRODUCTION

Patients with portal hypertension have significant mortality^[1]. A lack of drugs^[2] to treat this disease is derived from the failure to use the reversible mechanisms in its pathogenesis^[3]. Being similar to amiloride, a candidate drug for portal hypertension^[4], molecules from medical plants have been demonstrated to affect portal hypertension in rats^[5-7] and in patients^[8] with chronic hepatitis. The effect of these molecules on relaxation of the extrahepatic portal rings did not account for the efficacy of these therapies *in vivo*^[9]. A novel mode of portal perfusion has been characterized with both controlled velocity and monitored pressure in the isolated portal perfused rat liver (IPPRL)^[10-12]. With the primary velocity and preload at the various advanced stages of CCl₄-induced chronic hepatitis in rats, constriction with phenylephrine (PE) and relaxation with acetylcholine (Ach) were more sensitive than those reported previously^[13,14]. With standardization of the IPPRL^[15], both median effective concentrations of Ach and PE were defined as the pharmacodynamic model of portal hypertension. Both the controlled velocity and monitored pressure made the model sensitive enough for the basis of systems biology in portal regulation^[16].

MATERIALS AND METHODS

Materials

Thirty two healthy male Wistar rats weighing 200-220 g were supplied by the Animal Center of the Chinese Academy of Medical Sciences. Standard rodent pellets for rats were prepared by Beijing Scientific Animal Feed-stuff Company. The study was approved by the Animal Study Committee of the Chinese Academy of Medical Sciences. All experimental procedures were performed in accordance with the Guidelines of Animal Experiments from the Committee of Medical Ethics, National Health Department of China. All rats were maintained in a temperature-controlled room (25.0 °C ± 0.2 °C) in the SPF laboratory, with a 12-h/12-h light/dark photoperiod and 45% ± 2% humidity. The rats were fed standard rodent pellets and allowed free access to tap water throughout the experiment.

Carbon tetrachloride (CCl₄, MW 153.84, CAS 56-23-5), Olive oil (CAS 8001-25-0) and Heparin sodium (MW 12 000, CAS 9041-08-1) were purchased from Sinopharm Chemical Reagent Company to induce chronic hepatic hepatitis or for anticoagulation.

As the perfusate in portal perfusion, Krebs-Henseleit solution consisted of KCl 4.7, KH₂PO₄ 1.2, CaCl₂ 2.5,

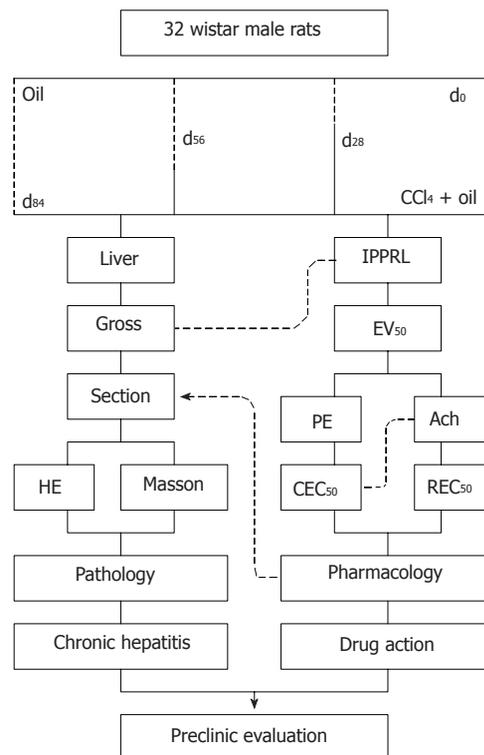


Figure 1 Group design of pharmacological model of portal hypertension. Oil indicated was administered subcutaneously with 3 mL/kg olive oil twice weekly for 84 d, as was CCl₄ + Oil 40% (v/v) CCl₄ in olive oil; IPPRL: Isolated portal perfused rat livers; EV₅₀: Median effective velocity; PE: Phenylephrine; Ach: Acetylcholine; CEC₅₀: Median effective concentration of PE constriction; REC₅₀: Median effective concentration of Ach relaxation; HE: Hematoxylin and eosin.

MgSO₄ 1.2, NaCl 118, and Glucose 11.0 mmol/L in the final concentration at pH 7.35-7.45 equilibrated with 95% O₂-5% CO₂ and warmed to 37.8 °C before being added to the livers.

Phenylephrine hydrochloride (PE, MW 203.67, CAS 61-76-7) and Acetylcholine chloride (Ach, MW 181.67, CAS 60-31-1) were purchased from Sigma (United States) as the α₁-adrenoceptor and M₃-cholinoceptor agonist, respectively, used to elevate or reduce portal pressure.

Experimental design

Male Wistar rats were randomly divided into four groups. In four rats, PE was used to constrict the portal vein and in the other rats Ach was used to relax the portal vein in each group. Group 1 was the vehicle control without CCl₄. In this group, rats were subcutaneously administered 3 mL/kg olive oil twice weekly for 84 d. Groups 2, 3 and 4 were model groups with CCl₄-induced chronic hepatitis, the rats in these groups were subcutaneously administered the same volume of a mixture of 40% (v/v) CCl₄ in olive oil twice weekly for 28 d, 56 d and 84 d, beginning at d₅₈, d₂₈, and d₀, respectively (Figure 1). Forty-eight hours after the last CCl₄ injection, rats were anesthetized with 50 mg/kg pentobarbital sodium subcutaneously; a midline incision was made to expose the liver and its vessels. The hepatic artery, portal vein and hepatic vein were cannalized. The remaining blood in the IPPRLs

Table 1 Hepatic pathological changes in model rats (mean \pm SE, $n = 8$)

Advanced	Lobule ratio	Collagen ratio
d ₀	0.38 \pm 0.05	0.0000700 \pm 0.0001180
d ₂₈	0.33 \pm 0.04 ^b	0.0019658 \pm 0.0024864 ^b
d ₅₆	0.11 \pm 0.04 ^{b,d}	0.0043315 \pm 0.0048768 ^{b,d}
d ₈₄	0.06 \pm 0.01 ^{b,d,f}	0.0143996 \pm 0.0143860 ^{b,d,f}

Lobule ratio is the average percentage obtained from ten random fields using Image-Pro Plus v 5.1 software, and each field had a ratio of lobule area per total analyzed field area in hematoxylin-eosin stained sections under a Digital Pathology System at $\times 20$ magnification from isolated portal perfused rat livers. Collagen ratio is the average percentage obtained from ten random fields using Image-Pro Plus v 5.1 software, and each field had a ratio of collagen area per total analyzed field area in Masson-stained sections under a Digital Pathology System at $\times 40$ magnification from isolated portal perfused rat livers. a and b denote significant (^b $P < 0.01$) differences between those at d₀ and at d₂₈, d₅₆, or d₈₄. c and d denote significant (^c $P < 0.05$ and ^d $P < 0.01$) differences at d₂₈ and at d₅₆, or d₈₄. e and f denote significant (^e $P < 0.05$ and ^f $P < 0.01$) differences at d₅₆ and at d₈₄.

was eliminated using Krebs-Henseleit perfusate through the hepatic artery. When portal perfusion was complete, a small portion of the liver was removed for pathological examination following fixation with 40 g/L formaldehyde solution and subsequent embedding in paraffin.

Protocol for portal perfusion

When CCl₄-induced chronic hepatitis was complete, eight rats from each group were randomized into two subgroups, one for PE constriction and the other for Ach relaxation of the portal vein. Each IPPRL was instrumented for portal pressure measurement.

Each IPPRL was perfused in a recirculation at a fixed temperature of 37.8 °C and equilibrated with 95% O₂-5% CO₂ mixed gas (Beijing Specialized Mixed Gas Institute), portal velocity was precisely controlled by a quantified BTO₁ pump (Beijing Yidaxk Technical Company), and 3935.50, 4720.63, 4753.35, and 5164.16 (μ L/min) at d₀, d₂₈, d₅₆, and d₈₄, were chosen, respectively, the equation of portal perfusion median velocity (Y) from the day (x) of chronic hepatitis was $Y = 13.28x + 4085$ ($r = 0.935$, $P < 0.01$)^[12].

The portal pressure of perfusion (mmHg) was continuously monitored and recorded with a strain-gauge transducer connected to the portal inflow cannula 6 cm proximal to the perfusion cannula with BL-420S Physiological Systems (Taimeng Instruments, Chengdu) according to a previously published method^[10-14]. The global viability of livers was assessed by gross appearance and perfusate stable pH.

Pharmacodynamic actions

Perfusions were performed in the recirculating system containing 300 mL Krebs-Henseleit solution. Each preparation was allowed to stabilize for 15 min. The flow rate during each individual perfusion was maintained at a constant rate equalized to the portal perfusion median velocity at d₀, d₂₈, d₅₆, and d₈₄, respectively, the average portal pressure during this condition had been designated as the

baseline. With a fixed volume of the recirculating perfusate in the portal perfusion system, cumulative geometric concentrations of PE (10^{-12} - 10^{-6} mol/L) were added to elevate portal pressure.

After the median effective concentration of PE to constrict the portal vein was added, cumulative geometric concentrations of Ach (10^{-13} - 10^{-7} mol/L) were added to reduce portal pressure.

Concentration-response curves were obtained following the addition of PE and Ach, and the changes in intra-hepatic resistance expressed as the percentage increase or decrease in perfusion pressure from baseline in the various portal perfused velocities were obtained.

Pathological changes due to chronic hepatitis

To observe pathological changes after portal perfusion, a portion of the left liver lobe (40 mg) from each liver was fixed in 40 g/L formaldehyde solution for 48 h, embedded in paraffin, sectioned (6 μ m), and stained with hematoxylin-eosin and Masson according to standard procedures.

Images were acquired with a Nano Zoomer Digital Pathology system (Hamamatsu, Japan), at a low magnification ($\times 20$); all the compartments of the liver were analyzed. At high magnification ($\times 40$), the collagen density in the liver section was quantified using a computerized image analysis system (Image-Pro Plus v 5.1). The density of collagen in blinded specimens was expressed as a percentage (the ratio of collagen area per total analyzed field area). The average of the score taken from ten random fields was used to generate a single score for each IPPRL.

Statistical analysis

All primary data are presented as means \pm SE for each dosage in each group. Statistical significance was calculated using Student's *t* test between groups, $P < 0.05$ was significant. (1) Dose-effect relationship: The equation, the median effective concentration and its 95% confidence intervals of PE or Ach were calculated by regression analysis using Graph-Pad Prism 4 in non-linear fit and various slopes, to express the dose-effect relationship; and (2) Time-effect relationship: The median effective concentrations of PE or Ach were calculated by linear regression analysis with the duration (0 d, 28 d, 56 d, and 84 d) of chronic hepatitis, to express the time-effect relationship of pathological conditions affecting the portal response to both molecules.

RESULTS

Pathological changes due to chronic hepatitis

Lobule or pseudo-lobule ratio: Hepatic tissues were clear in hematoxylin and eosin-stained sections. When compared with those in control rats at d₀, the lobule ratios (Table 1) in the model rats at d₂₈, d₅₆ and d₈₄ were significantly decreased by 4.04%, 70.22%, and 83.82%, respectively ($P < 0.01$). When compared with those at d₂₈, the pseudo-lobule ratios in the model rats at d₅₆ and d₈₄ were significantly decreased by 65.35% and 81.00%, respectively ($P < 0.01$). In addition,

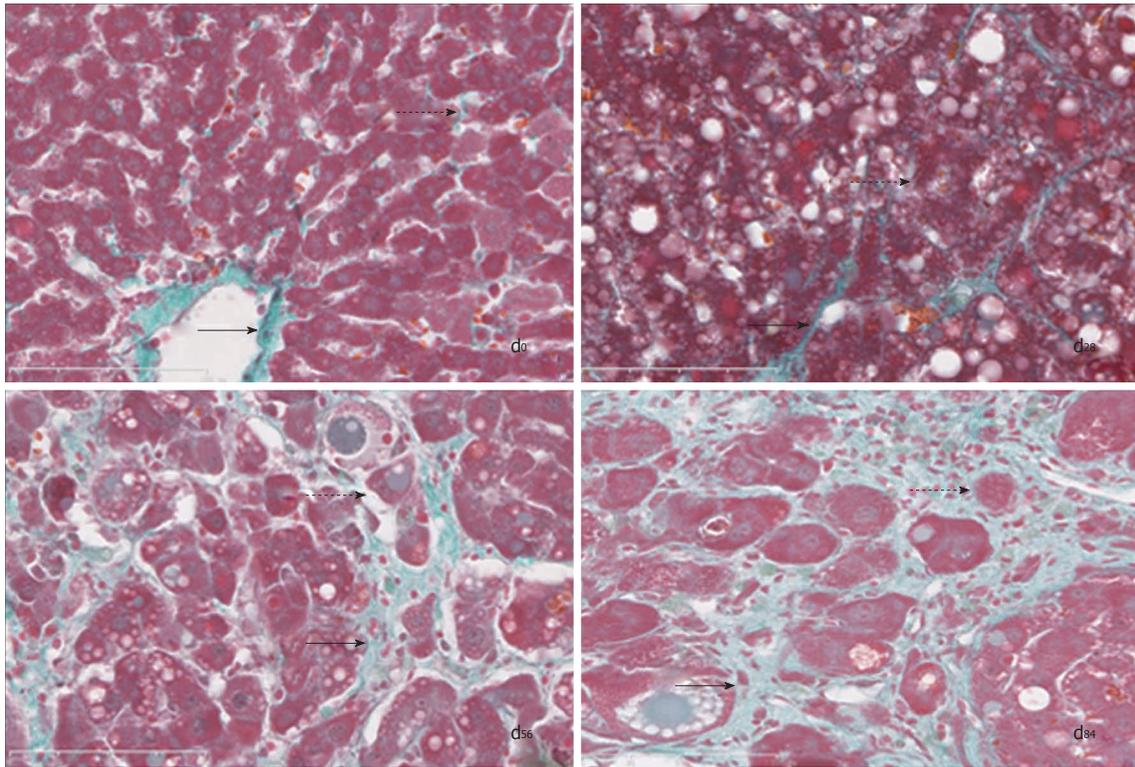


Figure 2 Collagen distributions in rat model livers (Masson $\times 40$). Normal hepatic structure (d_0) was demonstrated in control rats administered 3 mL/kg olive oil subcutaneously twice weekly for 84 d. Hepatic degeneration (d_{28}), fibrosis (d_{56}) and cirrhosis (d_{84}) were seen in the chronic hepatitis model rats administered 3 mL/kg 40% (v/v) CCl_4 in olive oil subcutaneously twice weekly for 28 d, 56 d, and 84 d, respectively. There was less collagen mainly around the central veins (solid arrow) and the portal areas, with some staining noise signals in lobules (dashed arrow) in normal hepatic structure (d_0). The collagen fiber bundles (solid arrow) had partly spread into the lobules from the portal areas, the hepatic cells had obvious watery and fatty degeneration (dashed arrow) and had not been isolated by collagen during hepatic degeneration (d_{28}). The collagen fiber bundles (solid arrow) had completely separated some of the lobules to form many pseudo-lobules, several hepatic cells with obvious fatty degeneration (dashed arrow) had been completely isolated by collagen in hepatic fibrosis (d_{56}). The collagen fiber bundles (solid arrow) had limited smaller pseudo-lobules, some single hepatic cells (dashed arrow) were completely isolated by collagen in hepatic cirrhosis (d_{84}).

these ratios were significantly decreased by 45.67% at d_{84} compared with those at d_{56} ($P < 0.01$).

Hepatic collagen distribution: Hepatic histological changes in Masson-stained sections showed collagen depositions along with CCl_4 -induced chronic hepatitis (Figure 2). (1) Normal structure at d_0 : The histological structure in control rats showed normal hepatic architecture with some fatty degeneration and less collagen located at the lobules; (2) Degeneration at d_{28} : The pathological changes in the model rats at d_{28} showed mainly hepatic fatty degeneration and cellular swelling, collagen was deposited around the center veins, thus the enlarged hepatic cords severely narrowed the hepatic sinusoid; (3) Hepatic fibrosis at d_{56} : The pathological changes in the model rats at d_{56} showed more collagen deposited in the lobules, thus the enlarged hepatic cords led to significant widening of the hepatic sinusoid; and the collagen in interlobular area extended into the lobules, some separating the lobules completely, therefore the direction of the circulating blood did not change in the hepatic sinusoid of the lobules; and (4) Hepatic cirrhosis at d_{84} : The pathological changes in the model rats at d_{84} showed extensive collagen deposited in the lobules, which were all pseudo-lobules instead of normal lobules, thus the direction of the circulating

blood had completely changed in the hepatic sinusoid.

Deposited collagen ratio: Compared with the control rats (Table 1), the collagen ratio in the model rats at 28 d, 56 d and 84 d was significantly increased by 2707.65%, 60 860.51%, and 20 466.49%, respectively ($P < 0.01$). Compared with the model rats at 28 d, the collagen ratio in the model rats at 56 d and 84 d was significantly increased by 120.34% and 632.52%, respectively ($P < 0.01$). The collagen ratio in the model rats at 84 d increased by 232.44% compared to that at 56 d ($P < 0.01$).

Pharmacodynamic actions on the portal vein

Phenylephrine elevated portal pressure: Geometric concentrations of PE to activate the α_1 receptor were added to the recirculating perfusate to elevate perfused portal pressure (Table 2 and Figure 3). The equation, the median effective concentration of PE and its 95% confidence intervals were regressed: (1) dose-effect at d_0 : The data showed that the equation of PE was $Y = 0.1732 + 0.3970/[1 + 10^{-(4.3061-0.4407 \cdot x)}]$ ($r = 0.9701$, $P < 0.01$); the median effective concentration with its 95% confidence intervals was 1.69×10^{-10} ($4.9769 \times 10^{-12} - 5.7599 \times 10^{-9}$) mol/L; (2) dose-effect at d_{28} : The data showed that the equation of PE was $Y = -0.004934 + 0.121134/[1 +$

Table 2 Phenylephrine elevates portal pressure (mean ± SE, n = 4)

Log [PE (mol/L)]	d ₀	d ₂₈	d ₅₆	d ₈₄
-12	0.210 ± 0.19	0.013 ± 0.02	0.013 ± 0.02	0.019 ± 0.03
-11	0.260 ± 0.16	0.025 ± 0.03	0.017 ± 0.02	0.024 ± 0.04
-10	0.360 ± 0.18	0.044 ± 0.06	0.046 ± 0.06	0.047 ± 0.07
-9	0.420 ± 0.24	0.072 ± 0.09	0.182 ± 0.25	0.076 ± 0.09
-8	0.550 ± 0.37	0.090 ± 0.12	0.247 ± 0.27	0.119 ± 0.10
-7	0.520 ± 0.37	0.093 ± 0.12	0.269 ± 0.27	0.123 ± 0.11
-6	0.570 ± 0.24	0.113 ± 0.13	0.286 ± 0.28	0.138 ± 0.12

Primary data were used to calculate the dose-effect relationship between phenylephrine and portal vein constriction in the isolated portal perfused rat livers without chronic hepatitis, where rats were administered 3 mL/kg olive oil subcutaneously twice per week for 84 d (control rats) (d₀). Rats with chronic hepatitis at three stages, hepatic degeneration (d₂₈), fibrosis (d₅₆) and cirrhosis (d₈₄), were administered 3 mL/kg 40% (v/v) CCl₄ in olive oil subcutaneously twice per week for 28 d, 56 d and 84 d, respectively. PE: Phenylephrine.

Table 3 Acetylcholine reduces portal pressure (mean ± SE, n = 4)

Log [Ach (mol/L)]	d ₀	d ₂₈	d ₅₆	d ₈₄
-13	-0.16 ± 0.12	-0.002 ± 0.01	-0.076 ± 0.08	-0.001 ± 0.01
-12	-0.19 ± 0.10	-0.009 ± 0.01	-0.091 ± 0.10	-0.006 ± 0.01
-11	-0.31 ± 0.07	-0.025 ± 0.01	-0.125 ± 0.15	-0.012 ± 0.01
-10	-0.39 ± 0.08	-0.035 ± 0.18	-0.176 ± 0.23	-0.019 ± 0.01
-9	-0.42 ± 0.08	-0.043 ± 0.04	-0.203 ± 0.26	-0.041 ± 0.02
-8	-0.44 ± 0.12	-0.049 ± 0.05	-0.225 ± 0.26	-0.050 ± 0.03
-7	-0.47 ± 0.14	-0.052 ± 0.07	-0.256 ± 0.28	-0.054 ± 0.03

Primary data were used to calculate the dose-effect relationship between acetylcholine and portal vein relaxation in the isolated portal perfused rat livers without chronic hepatitis, where rats were administered 3 mL/kg olive oil subcutaneously twice per week for 84 d (control rats) (d₀). Rats with chronic hepatitis at three stages, hepatic degeneration (d₂₈), fibrosis (d₅₆) and cirrhosis (d₈₄), were administered 3 mL/kg 40% (v/v) CCl₄ in olive oil subcutaneously twice per week for 28 d, 56 d and 84 d, respectively. Ach: Acetylcholine.

$10^{(-3.1247-0.3262 x)}$ ($r = 0.9937, P < 0.01$); the median effective concentration with its 95% confidence intervals was 2.64×10^{-10} ($7.1864 \times 10^{-12} - 9.6834 \times 10^{-9}$) mol/L; (3) dose-effect at d₅₆: The data showed that the equation of PE was $Y = 0.0104 + 0.2643/[1 + 10^{(-8.8462-0.9579 x)}]$ ($r = 0.9980, P < 0.01$); the median effective concentration with its 95% confidence intervals was 5.82×10^{-10} ($3.0691 \times 10^{-10} - 1.1031 \times 10^{-9}$) mol/L; (4) dose-effect at d₈₄: The data showed that the equation of PE was $Y = 0.01603 + 0.12107/[1 + 10^{(-5.1134+0.563 x)}]$ ($r = 0.9963, P < 0.01$); the median effective concentration with its 95% confidence intervals was 8.24×10^{-10} ($2.2476 \times 10^{-10} - 3.0207 \times 10^{-9}$) mol/L; and (5) time-effect: The linear regression equation was $Y = 0.081x + 1.173$ ($r = 0.981, P < 0.01$) between the median effective concentrations of PE (1.69, 2.64, 5.82 and 8.24) $\times 10^{-10}$ mol/L and the durations (0 d, 28 d, 56 d and 84 d) of chronic hepatic hepatitis.

Acetylcholine reduced portal pressure: Geometric con-

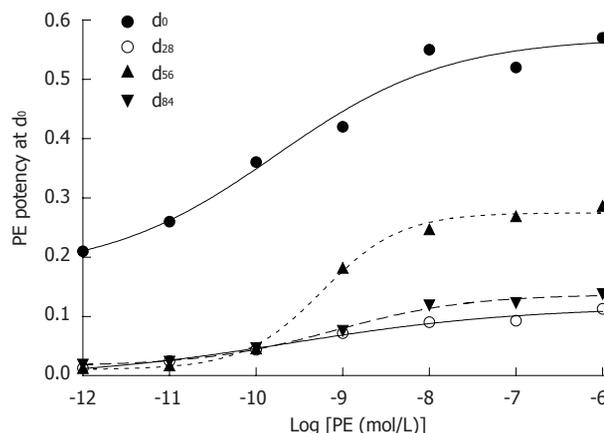


Figure 3 Phenylephrine elevates portal pressure (mean ± SE, n = 4). The dose-effect relationship between phenylephrine and portal vein constriction in isolated portal perfused rat livers without chronic hepatitis, where rats were administered 3 mL/kg olive oil subcutaneously twice per week for 84 d (control rats) (d₀). Rats with chronic hepatitis at three stages, hepatic degeneration (d₂₈), fibrosis (d₅₆) and cirrhosis (d₈₄), were administered 3 mL/kg 40% (v/v) CCl₄ in olive oil subcutaneously twice per week for 28 d, 56 d and 84 d, respectively. PE: Phenylephrine.

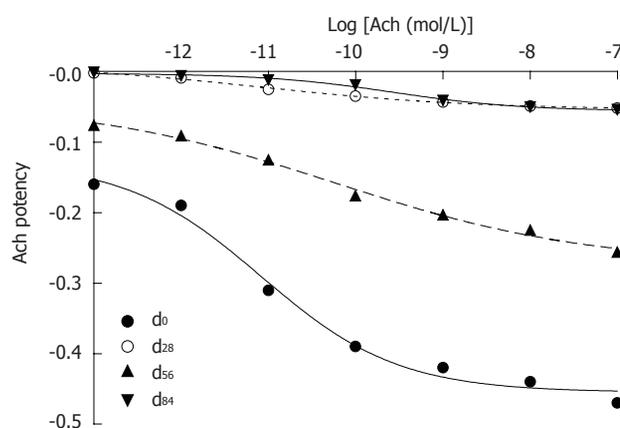


Figure 4 Acetylcholine reduces portal pressure (mean ± SE, n = 4). The dose-effect relationship between Acetylcholine and portal vein relaxation in isolated portal perfused rat livers without chronic hepatitis, where rats were administered 3 mL/kg olive oil subcutaneously twice per week for 84 d (control rats) (d₀). Rats with chronic hepatitis at three stages, hepatic degeneration (d₂₈), fibrosis (d₅₆) and cirrhosis (d₈₄), were administered 3 mL/kg 40% (v/v) CCl₄ in olive oil subcutaneously twice per week for 28 d, 56 d and 84 d, respectively. Ach: Acetylcholine.

centrations of Ach to activate the M₃ receptor were added to the circulating perfusate to reduce perfused portal pressure. The equation, the median effective concentration of Ach and its 95% confidence intervals of Ach were regressed (Table 3 and Figure 4): (1) dose-effect at d₀: The data showed that the equation of Ach was $Y = -0.4548 + 0.3274/[1 + 10^{(6.1538 + 0.5554 x)}]$ ($r = 0.9950, P < 0.01$); the median effective concentration with its 95% confidence intervals was 8.40×10^{-10} ($1.3263 \times 10^{-12} - 5.3240 \times 10^{-11}$) mol/L; (2) dose-effect at d₂₈: The data showed that the equation of Ach was $Y = -0.05391 + 0.06424/[1 + 10^{(3.8541 + 0.3469 x)}]$ ($r = 0.9982, P < 0.01$); the median effective concentration with its 95% confidence intervals was 7.73×10^{-12} ($7.3614 \times 10^{-13} - 8.1095 \times$

10^{-11}) mol/L; (3) dose-effect at d_{56} : The data showed that the equation of Ach was $Y = -0.2733 + 0.22978/[1 + 10^{(3.0472 + 0.3008 x)}]$ ($r = 0.9964$, $P < 0.01$); the median effective concentration with its 95% confidence intervals was 5.98×10^{-11} ($4.2797 \times 10^{-12} - 8.3556 \times 10^{-11}$) mol/L; (4) dose-effect at d_{84} : The data showed that the equation of Ach was $Y = -0.0559 + 0.053178/[1 + 10^{(5.6336 + 0.5883 x)}]$ ($r = 0.9956$, $P < 0.01$); the median effective concentration with its 95% confidence intervals was 2.66×10^{-10} ($6.5887 \times 10^{-11} - 1.0701 \times 10^{-9}$) mol/L; and (5) time-effect: The linear regression equation was $Y = 0.046 X - 1.470$ ($r = 0.945$, $P < 0.05$) between the median effective concentrations of Ach (0.0773 , 0.598 and 2.66) $\times 10^{-10}$ mol/L and the durations (28 d, 56 d and 84 d) of chronic hepatitis.

DISCUSSION

Patients with portal hypertension have significant morbidity and mortality^[1] without special drugs^[2] based on the reversible pathogenesis of this disease^[3]. Some candidate drugs from chemicals and medical plants have demonstrated effects on portal hypertension in animal experiments and in clinical trials^[4-8]. Data from the extra-hepatic portal rings failed to account for these effects^[9]. Consequently, sensitive portal perfusion for intra-hepatic portal resistance has been developed with both controlled velocity and monitored pressure in IPPRLs^[10-14]. The pharmacodynamic model of portal hypertension has further been defined as the median effective concentrations of Ach and PE in the IPPRLs at various stages of CCl₄-induced chronic hepatitis.

At d_0 , d_{28} , d_{56} , and d_{84} in CCl₄-induced chronic hepatitis, there were similar portal pressure potency equations with various coefficients due to the concentrations of PE and Ach in the IPPRLs. The median effective concentrations of PE increased geometrically during the process, suggesting that the function of portal smooth muscle cells gradually decreased. A similar effect was noted with the median effective concentrations of Ach in advanced stages, which suggested that portal endothelia were gradually damaged. During portal perfusion with both controlled pressure and monitored velocity, as reported previously, the effective range of PE and Ach concentrations was from 10^{-3} mol/L to 10^{-8} mol/L^[15,16]. In this novel model of portal perfusion with both controlled velocity and monitored pressure, the effective range was from 10^{-6} mol/L to 10^{-12} mol/L, which indicated that this novel mode was more sensitive than the previous mode by 10^3 - 10^6 times in IPPRLs.

Hepatocyte injuries originate from the free radicals of CCl₄ metabolites^[3]. Amiloride reduced intra-hepatic portal resistance through inhibition of the Rho kinase pathway in hepatic stellate cells^[4]. Glycyrrhizinate and Salvianolic acid B are representative molecules from medical plants used for portal hypertension in rats^[5-7] and patients^[8] with chronic hepatitis, however, their biomolecular mechanisms are not yet clear. PE, as a α_1 -adrenoceptor agonist, constricts vascular smooth muscle^[13] and Ach, as a M₃-cholinoceptor agonist in endothelia, relaxes vascular

smooth muscle^[14]. Due to these mechanisms in IPPRLs, the median perfused velocity in portal pressure has been defined as the primary flow rate of portal perfusion in this novel mode, the median effective concentration of PE for elevating portal pressure as the preload, the median effective concentration of Ach for reducing the portal pressure as the positive action at the classic stages of the pathological process in chronic hepatitis.

The pharmacodynamic model of portal hypertension has been defined as both the median effective concentrations of PE and Ach in the IPPRLs with advanced chronic hepatitis in this study. This model may be used to evaluate the preclinical effects of candidate drugs for the treatment of portal hypertension. Both controlled velocity and monitored pressure^[10-12] made this model more sensitive than previous models^[13-14]. Based on the standardization^[15] of the IPPRL, this sensitive model is considered the basis of systems biology for portal regulation in an isolated setting^[16].

COMMENTS

Background

Portal hypertension results in significant mortality without the administration of special drugs. Candidate drugs for this condition require serious pre-clinical evaluation using suitable methods.

Research frontiers

The recently identified reversible pathogenesis of portal hypertension may allow the development of new drugs for this disease. Candidate drugs derived from medical plants used in Chinese medical practices have confirmed its reversible pathogenesis. A sensitive pressure transducer was used here for exploiting the reversible pathogenesis as the pharmacological models. The optimal conditions for each step of the procedure can be defined as an available model.

Innovations and breakthroughs

Reversible portal hypertension was replicated in the advanced stages of chronic hepatitis in rats using CCl₄. A pharmacological model was developed using the median primary velocity of perfused flow as the anatomical preload, median effective concentrations of phenylephrine to constrict portal veins as the physiological preload, and the median effective concentrations of acetylcholine to relax portal veins in IPPRLs with chronic hepatitis.

Applications

This novel pharmacological model can be used to evaluate candidate drugs for the treatment of portal hypertension.

Terminology

This novel mode of portal perfusion is characterized by both controlled velocity and monitored pressure in the isolated portal perfused rat livers. The controlled velocity creates the optimal conditions for research purposes, and the monitored pressure gives exact data from vascular smooth muscle or endothelia.

Peer review

The authors investigated to develop a pharmacodynamic model for portal hypertension from chronic hepatitis. They have developed a pharmacodynamic model for portal hypertension in rats with chronic hepatitis and demonstrated that the model had been defined as the median effective concentrations of phenylephrine and acetylcholine. The results are clear and informative for the study on portal hypertension.

REFERENCES

- 1 Roberts SE, Goldacre MJ, Yeates D. Trends in mortality after hospital admission for liver cirrhosis in an English population from 1968 to 1999. *Gut* 2005; **54**: 1615-1621
- 2 Bosch J, Abrales JG, Fernández M, García-Pagán JC. Hepatic endothelial dysfunction and abnormal angiogenesis: new targets in the treatment of portal hypertension. *J Hepa-*

- tol* 2010; **53**: 558-567
- 3 **Xu YL**, Cai DY, Tang CS. Mechanism of CCl₄-induced hepatocirrhosis and portal hypertension. *Shijie Huaren Xiaohua Zazhi* 2005; **13**: 235-238
 - 4 **Steib CJ**, Hennenberg M, Beiting F, Hartmann AC, Bystrom M, De Toni EN, Gerbes AL. Amiloride reduces portal hypertension in rat liver cirrhosis. *Gut* 2010; **59**: 827-836
 - 5 **Liu SF**, Cai DY, Li PT, Xiang PR. Study on compatibility of Ganshen decoction in moderate Hepatic Fibrosis. *Zhonghua Zhongyiyao Zazhi* 2005; **20**: 373-375
 - 6 **Deng XL**, Wang QQ, Zhang XJ, Liu YN, Zeng YM, Jia L, Li PT, Cai DY. Effective mechanisms of Glycyrrhetic acid with Salvianolic acid B on immunological hepatic fibrosis in rat. *Zhongguo Yaoshi* 2007; **10**: 741-744
 - 7 **Du QH**, Li PT. Pathophysiology and clinical practice analysis on endothelin system and portal hypertension. *Shijie Huaren Xiaohua Zazhi* 2008; **16**: 1092-1097
 - 8 **Qin G**, Shi GF, Song YY, Chen MQ. Meta-analysis of document on diammonium Glycyrrhizinate in treatment of patients with chronic hepatitis B. *Zhonghua Chuanranbing Zazhi* 2005; **23**: 333-337
 - 9 **Ren LW**, Zang XJ, Deng XL, Liu SF, Liu YN, Cai DY. Wave characteristics of portal pressure and their affected factors. *Beijing Zhongyiyao Daxue Xuebao* 2006; **29**: 840-843
 - 10 **Wu K**, Xu XY, Wang SX, Zhou H, Zhang WL, Zhang WT, Lu AN, Song M, Zhang BC, Li PT, Cai DY. A rat model for monitoring isolated perfused hepatic artery or portal vein pressure in vitro. *Zhongguo Yaoshi* 2010; **13**: 1390-1393
 - 11 **Zhou H**, Zhang T, Xu XY, Wang SX, Wu K, Xu J, Chen M, Li PT, Cai DY. Salvianolic acid B or Glycyrrhizinate acting on rat infused portal pressure. *Zhangguo Zhongxiyi Jiehe Zazhi* 2010; **30**: 1084-1086
 - 12 **Xu XY**, Zhang T, Zhou H, Zhao X, Zhang TT, Yin H, Li T, Li PT, Cai DY. Portal pressure determined by perfusion velocity in isolated rat with chronic injury liver in vitro. *Shijie Huaren Xiaohua Zazhi* 2010; **18**: 2745-2749
 - 13 **Laviña B**, Gracia-Sancho J, Rodríguez-Vilarrupla A, Chu Y, Heistad DD, Bosch J, García-Pagán JC. Superoxide dismutase gene transfer reduces portal pressure in CCl₄ cirrhotic rats with portal hypertension. *Gut* 2009; **58**: 118-125
 - 14 **Hennenberg M**, Trebicka J, Sauerbruch T, Heller J. Mechanisms of extrahepatic vasodilation in portal hypertension. *Gut* 2008; **57**: 1300-1314
 - 15 **Bessemers M**, 't Hart NA, Tolba R, Doorschodt BM, Leuvenink HG, Ploeg RJ, Minor T, van Gulik TM. The isolated perfused rat liver: standardization of a time-honoured model. *Lab Anim* 2006; **40**: 236-246
 - 16 **Hood L**, Rowen L, Galas DJ, Aitchison JD. Systems biology at the Institute for Systems Biology. *Brief Funct Genomic Proteomic* 2008; **7**: 239-248

S- Editor Zhang SJ L- Editor Webster JR E- Editor Zhang DN

Safety assessment of *Bifidobacterium longum* JDM301 based on complete genome sequences

Yan-Xia Wei, Zhuo-Yang Zhang, Chang Liu, Pradeep K Malakar, Xiao-Kui Guo

Yan-Xia Wei, Zhuo-Yang Zhang, Chang Liu, Xiao-Kui Guo, Department of Medical Microbiology and Parasitology, Institutes of Medical Sciences, Shanghai Jiao Tong University School of Medicine, Shanghai 200025, China

Pradeep K Malakar, Institute of Food Research, Norwich Research Park, NR4 7UA, Norwich, United Kingdom

Author contributions: Guo XK and Wei YX designed the study; Wei YX and Liu C analyzed the data; Wei YX and Zhang ZY carried out the experiments; Wei YX wrote the paper; and Malakar PK edited the paper.

Supported by The National Key Program for Infectious Diseases of China, No. 2008ZX10004 and 2009ZX10004; the Program of Shanghai Subject Chief Scientist, No. 09XD1402700; the Program of Shanghai Research and Development, No. 10JC1408200; and a China Partnering Award from the Biotechnology and Biological Sciences Research Council, United Kingdom

Correspondence to: Xiao-Kui Guo, Professor, Department of Medical Microbiology and Parasitology, Institutes of Medical Sciences, Shanghai Jiao Tong University School of Medicine, Shanghai 200025, China. microbiology@sjtu.edu.cn

Telephone: +86-21-64453285 Fax: +86-21-64453285

Received: May 25, 2011 Revised: July 31, 2011

Accepted: August 7, 2011

Published online: February 7, 2012

Abstract

AIM: To assess the safety of *Bifidobacterium longum* (*B. longum*) JDM301 based on complete genome sequences.

METHODS: The complete genome sequences of JDM301 were determined using the GS 20 system. Putative virulence factors, putative antibiotic resistance genes and genes encoding enzymes responsible for harmful metabolites were identified by blast with virulence factors database, antibiotic resistance genes database and genes associated with harmful metabolites in previous reports. Minimum inhibitory concentration of 16 common antimicrobial agents was

evaluated by *E*-test.

RESULTS: JDM301 was shown to contain 36 genes associated with antibiotic resistance, 5 enzymes related to harmful metabolites and 162 nonspecific virulence factors mainly associated with transcriptional regulation, adhesion, sugar and amino acid transport. *B. longum* JDM301 was intrinsically resistant to ciprofloxacin, amikacin, gentamicin and streptomycin and susceptible to vancomycin, amoxicillin, cephalothin, chloramphenicol, erythromycin, ampicillin, cefotaxime, rifampicin, imipenem and trimethoprim-sulphamethoxazol. JDM301 was moderately resistant to bacitracin, while an earlier study showed that bifidobacteria were susceptible to this antibiotic. A tetracycline resistance gene with the risk of transfer was found in JDM301, which needs to be experimentally validated.

CONCLUSION: The safety assessment of JDM301 using information derived from complete bacterial genome will contribute to a wider and deeper insight into the safety of probiotic bacteria.

© 2012 Baishideng. All rights reserved.

Key words: *Bifidobacterium longum*; Safety assessment; Genome; Antibiotic resistance; Harmful metabolite; Virulence factor

Peer reviewer: Tauseef Ali, MD, Assistant Professor, Section of Digestive Diseases and Nutrition, University of Oklahoma Health Sciences Center, 920 SL Young Blvd, Oklahoma City, OK 73104, United States

Wei YX, Zhang ZY, Liu C, Malakar PK, Guo XK. Safety assessment of *Bifidobacterium longum* JDM301 based on complete genome sequences. *World J Gastroenterol* 2012; 18(5): 479-488 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/479.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.479>

INTRODUCTION

Bifidobacteria spp are high-GC content, Gram-positive bacteria which belong to the *Actinobacteria* branch and these species naturally colonize the gastrointestinal tract (GIT) of mammals, birds and insects^[1]. Scientists have determined the major probiotic properties of *Bifidobacteria* spp isolated from the human intestine and these properties include the strengthening of the intestinal barrier, modulation of the immune response and antagonism of pathogens^[2].

Bifidobacterium spp has been reported to possess various glycosyl hydrolases (GH) and these hydrolases metabolize plant- or milk-derived oligosaccharides including nondigestible ones such as galacto-oligosaccharides (GOS) and fructo-oligosaccharides (FOS)^[3,4]. The capability to utilize nondigestible oligosaccharides confers a competitive advantage to *Bifidobacterium* spp in the human gut.

Bifidobacterium longum (*B. longum*) and various other bifidobacteria strains are often added to probiotic products in combination with other lactic acid bacteria (LAB). Through their long and safe history of application, LAB have acquired the status of “Generally Regarded As Safe” (GRAS), but the safety of *Bifidobacteria* and other LAB strains selected for probiotics still need to be carefully evaluated. The key safety aspects for use of bifidobacteria and other LAB strains in probiotics include antibiotic resistance, production of harmful metabolites and the potential for virulence. Antibiotic resistance in potential probiotic strains is not considered a risk factor unless resistance is transferred to pathogens or it renders the *probiotic untreatable* in very rare cases of infection^[5]. Biogenic amines, D-lactic acid, azoreductases and nitroreductases produced by **bifidobacteria and other LAB** strains are potential health hazards^[6,7] and the safety of some of these compounds have been evaluated^[8]. Virulence genes may be present in commensal bacteria and absence of virulence in these bacteria needs to be proved on a case by case basis.

Probiotic agents are widely used in the food and drug industry and as more commercial probiotic products are being introduced in the market, it is timely to reassess the safety of these probiotic products using the latest technology. Information from the complete genome sequences of *Bifidobacteria* will provide additional insight into the genetic basis for their safety. We sequenced the complete genome sequences of *B. longum* JDM301 (GenBank accession number CP002010), a commercial strain used widely in China with several probiotic functions, for this purpose^[9].

The aim of the present work was to assess the safety of *B. longum* JDM301 based on complete genome sequences. The criteria used were the potential to transfer antibiotic resistance to pathogens, the potential for production of harmful metabolites and the potential for virulence.

MATERIALS AND METHODS

Bacterial strains and growth conditions

JDM301 was isolated from commercial probiotic product

and identified using a sequence analysis of its 16S rRNA gene. De Man-Rogosa-Sharpe (MRS) broth (Difco) supplemented with 0.05% L-cysteine-HCl (Sigma) was used for cultivating JDM301. Cultures were incubated at 37 °C under anaerobic conditions.

Genome sequencing and assembly

We determined the complete genome sequence of JDM301 at the Chinese National Human Genome Center in Shanghai using the GS 20 system (454 Life Science Corporation, Branford, Connecticut). A total of 192 888 reads with an average length of 210 bps were assembled into 112 contigs by the 454 assembly tool. The order of most large contigs, which were larger than 500 bp, was determined through the basic local alignment search tool (BLAST) analysis with the reference strain *B. longum* ATCC15697 (GenBank accession number CP001095) and the others were arranged by multiplex polymerase chain reaction (PCR). Gap closure was carried out by sequencing gap-spanning PCR products or clones using ABI 3730 xl DNA sequencers. Primer design and sequence assembly were performed by the Phred/Phrap/Consed software package^[9]. The locations of low-quality sequences in genome were verified by directly resequencing the PCR products spanning the low-quality sequences using the ABI 3730 xl DNA sequencers.

Statistical analysis

The genome sequences of *Bifidobacteria* except JDM301 were retrieved from GenBank at the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>)^[10]. Potential open reading frames (ORF) were identified using Glimmer^[11] and ZCURVE^[12] 1.0 using default settings. Clusters of orthologous group (COG) functional categories were used for functional classification of all genes in the genome sequences of JDM301 and the COGs. A BLAST analysis of the translations with GenBank’s nonredundant database was performed, which was followed by manual curation. The best matches were chosen for preliminary product assignments. Insertion sequences (IS) elements, prophage sequences and clustered regularly interspaced short palindromic repeats (CRISPR) were identified by IS finder (<http://www-is.biotoul.fr/is.html>), Prophage Finder^[13] and CRISPRFinder (<http://crispr.u-psud.fr/crispr/>)^[14] respectively. Putative orthologues were determined by Omics Explorer (<http://omics.biosino.org:14000/kweb/about.jsp>) using default values. Ribosomal RNA genes were detected on the basis of BLASTN searches and transfer RNA genes were identified using tRNAscan-SE^[15]. The atlas of genome was drawn using GenomeViz1.1^[16]. Putative virulence factors and putative antibiotic resistance genes were identified by blast with virulence factors database (VFDB) (<http://www.mgc.ac.cn/VFs/main.htm>)^[17] and antibiotic resistance genes database (ARDB) (<http://arbd.cbcb.umd.edu/>)^[18] respectively.

Antibiotic susceptibility

Minimum inhibitory concentration (MIC) of 16 common

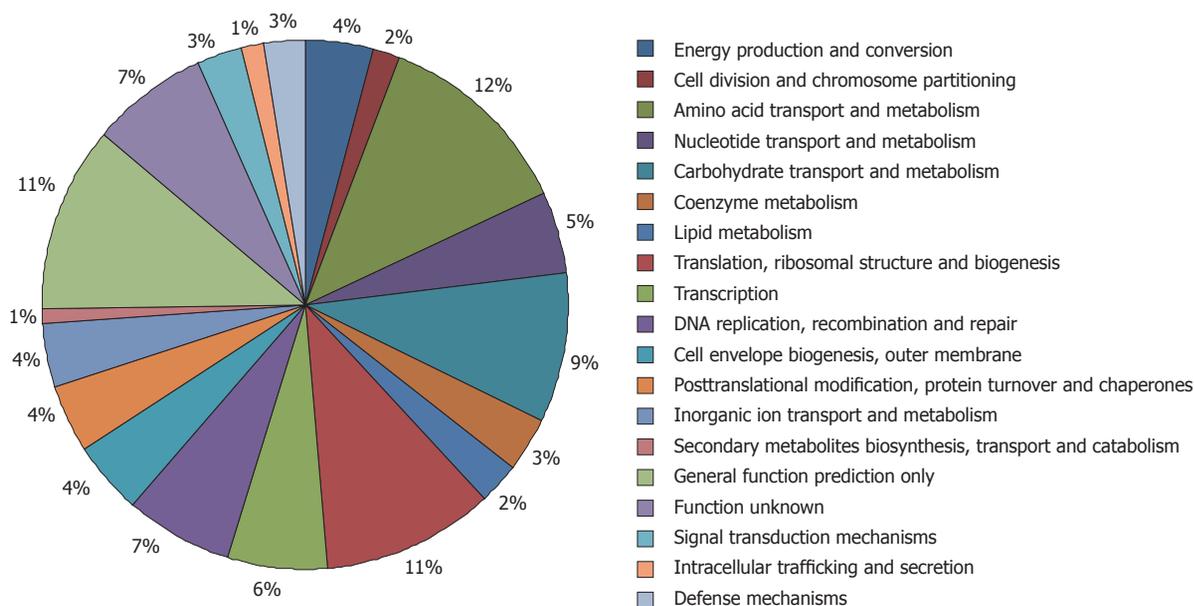


Figure 1 Functional distribution of *Bifidobacterium longum* core proteins. A total of 1265 proteins were conserved in all four *Bifidobacterium longum* (*B. longum*) strains (*B. longum* JDM301, *B. longum* NCC2705, *B. longum* DJO10A and *B. longum* ATCC15697), representing the "core" genome of *B. longum*.

antimicrobial agents was evaluated by *E*-test (AB Biodisk, Solna, Sweden) including amoxicillin (0.016-256 mg/L), amikacin (0.016-256 mg/L), ampicillin (0.016-256 mg/L), bacitracin (0.016-256 mg/L), cephalothin (0.016-256 mg/L), ciprofloxacin (0.002-32 mg/L), cefotaxime (0.016-256 mg/L), chloramphenicol (0.016-256 mg/L), erythromycin (0.016-256 mg/L), gentamicin (0.016-256 mg/L), imipenem (0.002-32 mg/L), rifampicin (0.016-256 mg/L), streptomycin (0.016-256 mg/L), tetracycline (0.016-256 mg/L), trimethoprim-sulphamethoxazol (0.002-32 mg/L), and vancomycin (0.016-256 mg/L). Tests were done with MRS agar supplemented with 0.05% L-cysteine ·HCl (Sigma) and were conducted in triplicate for each antibiotics. Cultures sub-inoculated into the MRS agar supplemented with 0.05% L-cysteine ·HCl were incubated anaerobically at 37 °C for 24 h.

RESULTS

Comparative genomic analysis of *Bifidobacteria*

The predicted proteins of *B. longum* JDM301 were functionally categorized. The functional distribution of genes assigned to clusters of orthologous groups of proteins was relatively similar to the other *Bifidobacteria*, e.g., *B. longum* and *B. adolescentis* in the GIT and *B. dentium* in the oral cavity^[3,4,19]. The top four functional categories in *B. longum* JDM301, namely, carbohydrate transport and metabolism, amino acid transport and metabolism, were identical with other *Bifidobacteria*^[20].

Putative orthologues among *B. longum* strains were determined in a comparative study (Figure 1). Overall, 1265 proteins were conserved in all four *B. longum* strains (*B. longum* JDM301, *B. longum* NCC2705, *B. longum* DJO10A and *B. longum* ATCC15697). These proteins represent the "core" genome of *B. longum*, whereas 219 proteins

are unique to *B. longum* JDM301. The most common functional distributions of the core proteins were these involved in housekeeping functions including amino acid transport and metabolism, translation, ribosomal structure and biogenesis, carbohydrate transport and metabolism and DNA replication, recombination and repair. Twenty-one percent of the core proteins were dedicated to carbohydrate and amino acid transport and metabolism, indicating the important roles of these proteins in *Bifidobacteria*.

Stability of the genome of *B. longum* JDM301

Horizontal gene transfer (HGT) events are responsible for introduction of alien genes, which may reinforce the adaptation of bacteria in their specific niches. Genes on plasmids, bacteriophages, genomic islands and IS are sensitive to HGT^[21]. Twelve phage-related fragments were identified in the genome of *B. longum* JDM301^[9], but no complete prophages were found. The JDM301 chromosome also possesses 15 complete or disrupted IS elements^[9]. The number of IS element in JDM301 is relatively smaller than the other sequenced *B. longum* spp^[3,4]. Another set of genes disseminated by HGT in *Bifidobacteria* is the CRISPR-related system. No CRISPR was discovered in the genome.

One complete type II restriction-modification (R-M) system and one type III R-M system were present in the genome of JDM301. A complete and incomplete type I R-M system was also identified in this genome. Two complete type II R-M systems and one type I R-M system were present in the genome of *B. longum* NCC2705, while one complete type II R-M system and type I R-M system were found in *B. longum* DJO10A.

Antibiotic resistance determinants

The antibiotic resistance genes in JDM301 were identified

Table 1 Putative antibiotic resistance genes identified in the genome of *Bifidobacterium longum* JDM301

Antibiotics	Antibiotic resistance genes	Product name
Bacitracin	BLJ_1636	ABC transporter-related protein
	BLJ_0984	ABC transporter-related protein
	BLJ_0923	ABC transporter-related protein
	BLJ_1055	Undecaprenyl pyrophosphate phosphatase
	BLJ_1119	Bacitracin transport ATP-binding protein bcrA
Vancomycin	BLJ_0853	VanU
	BLJ_1764	Dehydrogenase VanH
	BLJ_1084	Sensor protein vanSB
	BLJ_0707	VanSD5
	BLJ_0343	Histidine kinase VanSc3
	BLJ_0287	D-Ala: D-Lac ligase VanD
Multiple drugs	BLJ_1090	ATP-binding protein
	BLJ_1650	Lsa
	BLJ_1437	LmrB
	BLJ_0618	Multidrug export protein MepA
	BLJ_0769	Efflux transporter, RND family, MFP subunit
Chloramphenicol	BLJ_0181	Multidrug efflux protein QacB
	BLJ_1062	Multidrug export protein MepA
	BLJ_1672	Chloramphenicol resistance protein
Thiostrepton	BLJ_1322	Chloramphenicol resistance protein
Penicillin	BLJ_0885	Thiostrepton-resistance methylase
Kasugamycin	BLJ_1301	Penicillin binding protein
	BLJ_2030	S-adenosylmethionine-6-N', N'-adenosyl (rRNA) dimethyltransferase
Tetracycline	BLJ_0814	Tetracycline-resistance determinant tetV
	BLJ_1245	TetW
	BLJ_0594	Tetracycline resistance protein
	BLJ_1401	TetQ
Carbomycin	BLJ_1625	Carbomycin resistance protein
Sulfonamide	BLJ_1629	Dihydropteroate synthase
Tetracenomycin C	BLJ_1624	Tetracenomycin C efflux protein
Trimethoprim	BLJ_1657	dihydrofolate reductase
Macrolide	BLJ_0925	Macrolide-efflux protein
	BLJ_1936	Macrolide-efflux protein
	BLJ_0819	Macrolide-efflux protein
	BLJ_0042	Macrolide-efflux protein
	BLJ_1154	Macrolide-efflux protein variant

ABC: ATP-binding cassette; RND: Resistance-nodulation-cell division.

using ARDB ($E < 1e-2$, coverage $> 70\%$)^[18]. Homologs of the antibiotic resistance determinants for vancomycin, methicillin, tetracycline, chloramphenicol and trimethoprim were found in the genome of JDM301 (Table 1) and **6 putative resistance genes for vancomycin**. *B. longum* JDM301 also possessed **5 putative bacitracin efflux pumps**, 5 homologs of macrolide efflux proteins. Additionally, **7 putative multidrug resistance efflux pumps** belonging to an ATP-binding cassette (ABC)-type transport system, a major facilitator superfamily transporter and resistance-nodulation-cell division (RND) family were found in the genome. The genome of *B. longum* JDM301 also contains 4 tetracycline resistance genes encoding for TetV, TetW, TetPB and TetQ. The gene for TetW shows a strong difference in G + C content (53.0%) compared to the average value of *B. longum* JDM301 (59.8%) genome

Table 2 Minimum inhibitory concentration values of 16 antibiotics for *Bifidobacterium longum* JDM301

Antibiotics	Minimum inhibitory concentration (mg/L)
Ciprofloxacin	> 32
Amikacin	> 256
Gentamicin	> 256
Bacitracin	26.67
Streptomycin	170.67
Vancomycin	0.9
Amoxicillin	0.064
Cephalothin	1.33
Chloramphenicol	0.25
Erythromycin	0.04
Ampicillin	0.058
Cefotaxime	0.19
Rifampicin	0.074
Tetracycline	8
Imipenem	0.19
Trimethoprim-sulphamethoxazol	1.83

and it is flanked by genes encoding for integrases, indicating that this region may have been acquired by HGT.

The antibiotic susceptibility of *B. longum* JDM301 to 16 antibiotics was determined by an *E*-test to probe the *in silico* analyses of the complete genome sequence. The results of the *E*-test are summarized in Table 2. The breakpoints for determining susceptibility were determined using accepted protocols^[22-25]. *B. longum* JDM301 showed a **high resistance to ciprofloxacin, amikacin and gentamicin**, moderate resistance to streptomycin and bacitracin and were sensitive to tetracycline, vancomycin, amoxicillin, cephalothin, chloramphenicol, erythromycin, ampicillin, cefotaxime, rifampicin, imipenem and an antimicrobial compound, trimethoprim-sulphamethoxazol.

Putative enzymes for harmful metabolites

Genes encoding enzymes responsible for harmful metabolites, including beta-glucosidase (GS), arylsulphatase (AS), beta-glucuronidase (GN), nitroreductase (NR), azoreductase (AR), D-lactate dehydrogenase (DLD), amino acid decarboxylase (AD) and conjugated bile salt hydrolase (CBSH) were searched for in the genome of *B. longum* JDM301. Two GS genes (BLJ_1280, BLJ_1540) and one CBSH gene (BLJ_0948) were found in the chromosome of *B. longum* JDM301. Homologs of DLD (BLJ_1306, BLJ_1436) and NR (BLJ_1980) were also discovered in the genome. Enzymes involved in putatively harmful metabolites, AR, GN, AD and AS were not found in JDM301 genome.

Putative virulence factors

Published reports of rare infections involving *Lactobacilli* or *Bifidobacteria* are available and the potential virulence of *Lactobacilli* or *Bifidobacteria* used as probiotics should be assessed^[5]. Putative virulence genes of *B. longum* JDM301 were determined by BLAST analysis of the VFDB^[17]. A total of 141 homologs of virulence factors were identified in the genome of JDM301, including 28 sugar-binding transcriptional regulators, 20 genes associated

Table 3 Putative virulence factors identified in the genome of *Bifidobacterium longum* JDM301

Query	Identity	Subject	Predicted functions
BLJ_1089	24.9	VFG0934	2,3-dihydro-2,3-dihydroxybenzoate dehydrogenase
BLJ_1835	26.36	VFG0934	2,3-dihydro-2,3-dihydroxybenzoate dehydrogenase
BLJ_0323	29.3	VFG0934	2,3-dihydro-2,3-dihydroxybenzoate dehydrogenase
BLJ_1476	22.11	VFG2378	6 kDa early secretory antigenic target <i>esxA</i>
BLJ_0992	32.98	VFG0869	AatC ATB binding protein of ABC transporter
BLJ_1080	34.81	VFG0869	AatC ATB binding protein of ABC transporter
BLJ_1968	37.3	VFG0869	AatC ATB binding protein of ABC transporter
BLJ_0770	37.43	VFG0869	AatC ATB binding protein of ABC transporter
BLJ_0026	35.71	VFG1404	<i>ahpC</i>
BLJ_0136	28.73	VFG2218	ATPase VirB11 homolog
BLJ_0880	24.18	VFG1042	ATP-binding protein <i>FecE</i>
BLJ_0787	47.92	VFG0077	ATP-dependent Clp protease proteolytic subunit
BLJ_0786	53.8	VFG0077	ATP-dependent Clp protease proteolytic subunit
BLJ_0948	37.66	VFG2162	Bile salt hydrolase
BLJ_1243	22.97	VFG2242	Conjugal transfer protein <i>trag</i>
BLJ_0551	26.54	VFG1108	Conserved hypothetical protein
BLJ_1951	29.85	VFG1269	Cyclolysin secretion ATP-binding protein
BLJ_1925	32.31	VFG1269	Cyclolysin secretion ATP-binding protein
BLJ_1863	45.5	VFG0079	Endopeptidase Clp ATP-binding chain C
BLJ_1465	56.77	VFG0079	Endopeptidase Clp ATP-binding chain C
BLJ_0713	30.12	VFG0925	Ferric enterobactin transport ATP-binding protein <i>fepC</i>
BLJ_1872	25.51	VFG2225	GDP-mannose 4,6-dehydratase
BLJ_1324	32.49	VFG1399	<i>glnA1</i>
BLJ_0624	62.11	VFG1399	<i>glnA1</i>
BLJ_1834	29.47	VFG0313	Glucose/galactose transporter
BLJ_1926	30.02	VFG1557	HlyB protein
BLJ_1477	56.12	VFG1855	Hsp60, 60K heat shock protein <i>HtpB</i>
BLJ_0064	26.21	VFG1397	<i>hspX</i>
BLJ_1444	40.85	VFG1563	Hypothetical protein
BLJ_1606	27.78	VFG1593	Hypothetical protein
BLJ_1640	30.81	VFG1593	Hypothetical protein
BLJ_0011	22.16	VFG1604	Hypothetical protein
BLJ_1513	26.3	VFG1604	Hypothetical protein
BLJ_1846	27.67	VFG1604	Hypothetical protein
BLJ_0337	44.25	VFG1630	Hypothetical protein
BLJ_0336	44.38	VFG1630	Hypothetical protein
BLJ_1500	23.53	VFG1963	Hypothetical protein Cj1435c
BLJ_1169	24.64	VFG1390	Hypothetical protein Rv0981
BLJ_0708	36.8	VFG1390	Hypothetical protein Rv0981
BLJ_0802	28.83	VFG1824	Hypothetical protein Rv3133c
BLJ_1357	30.46	VFG1824	Hypothetical protein Rv3133c
BLJ_1113	32.41	VFG1824	Hypothetical protein Rv3133c
BLJ_0835	32.42	VFG1824	Hypothetical protein Rv3133c
BLJ_0859	27.93	VFG1206	Iron(III) ABC transporter, ATP-binding protein
BLJ_0348	28.13	VFG1206	Iron(III) ABC transporter, ATP-binding protein
BLJ_0530	29.29	VFG1206	Iron(III) ABC transporter, ATP-binding protein
BLJ_2016	35.81	VFG1206	Iron(III) ABC transporter, ATP-binding protein
BLJ_1875	36.19	VFG1627	IS100 transposase; transposase ORFA
BLJ_1249	37.55	VFG1627	IS100 transposase; transposase ORFA
BLJ_1252	39.22	VFG1627	IS100 transposase; transposase ORFA
BLJ_0930	42.29	VFG1627	IS100 transposase; transposase ORFA
BLJ_1966	30.68	VFG1485	L7045
BLJ_1850	59.7	VFG1411	<i>leuD</i>
BLJ_0379	39.24	VFG0320	Lipopolysaccharide core biosynthesis protein (<i>kdtB</i>)
BLJ_1549	22.02	VFG1817	<i>mbtA</i>
BLJ_1204	25.8	VFG0574	Mg ²⁺ transport protein
BLJ_2010	30.62	VFG0574	Mg ²⁺ transport protein
BLJ_1270	28.62	VFG1116	N-acetylglucosamine-6-phosphate deacetylase
BLJ_1832	21.89	VFG1109	N-acetylneuraminase lyase, putative
BLJ_0490	25.95	VFG1109	N-acetylneuraminase lyase, putative
BLJ_0021	26.83	VFG0307	Neutrophil activating protein (bacterioferritin)
BLJ_1889	24.14	VFG2227	O-antigen export system permease protein
BLJ_1251	26.05	VFG1461	ORF A protein
BLJ_0214	30.5	VFG0594	Pathogenicity island encoded protein: SPI3
BLJ_0159	33.25	VFG0594	Pathogenicity island encoded protein: SPI3
BLJ_1474	57.32	VFG1386	<i>phoP</i>
BLJ_1703	25.65	VFG2220	Phosphoglucosyltransferase
BLJ_0497	28.35	VFG2362	Phosphomannomutase
BLJ_1137	25.1	VFG1983	ABC-type amino-acid transporter periplasmic solute-binding protein
BLJ_0508	25.93	VFG1983	ABC-type amino-acid transporter periplasmic solute-binding protein
BLJ_1453	29.27	VFG1983	ABC-type amino-acid transporter periplasmic solute-binding protein
BLJ_0408	38.22	VFG2059	ATP-binding component of ABC transporter
BLJ_0480	27.04	VFG2061	Phosphoprotein phosphatase
BLJ_0805	28.09	VFG1384	<i>proC</i>
BLJ_1396	31.06	VFG1384	<i>proC</i>
BLJ_0584	26.09	VFG1387	<i>purC</i>
BLJ_1772	22.28	VFG0480	Putative amino acid permease
BLJ_0538	25.17	VFG0480	Putative amino acid permease
BLJ_1329	24.42	VFG1965	Putative aminotransferase
BLJ_0025	30.45	VFG2301	Putative carbonic anhydrase
BLJ_0922	23.51	VFG0031	Putative glycosyl transferase
BLJ_1670	38.88	VFG1668	Putative lysyl-tRNA synthetase <i>LysU</i>
BLJ_0563	25	VFG1498	Putative periplasmic solute binding protein
BLJ_1171	28.48	VFG0483	Putative regulatory protein, <i>deoR</i> family
BLJ_1517	29.25	VFG0483	Putative regulatory protein, <i>deoR</i> family
BLJ_0344	37.02	VFG1702	Putative response regulator
BLJ_0040	27.91	VFG1746	Putative two-component response regulator
BLJ_0740	29.13	VFG1746	Putative two-component response regulator
BLJ_1105	24.49	VFG0168	Pyochelin biosynthesis protein <i>PchD</i>
BLJ_0409	25.56	VFG0168	Pyochelin biosynthesis protein <i>PchD</i>
BLJ_0720	41.04	VFG0479	Pyruvate kinase I (formerly F), fructose stimulated
BLJ_1163	55.32	VFG1826	<i>relA</i>
BLJ_0995	25.84	VFG1889	Response regulator <i>GacA</i>
BLJ_1679	28.89	VFG1889	Response regulator <i>GacA</i>
BLJ_1083	40.89	VFG0473	Response regulator in two-component regulatory system with <i>BasS</i>
BLJ_1273	26.57	VFG1115	ROK family protein
BLJ_1620	26.62	VFG1115	ROK family protein
BLJ_1622	31.35	VFG1115	ROK family protein
BLJ_1796	27.31	VFG0526	Salmonella iron transporter: <i>fur</i> regulated
BLJ_0662	29.06	VFG0526	Salmonella iron transporter: <i>fur</i> regulated

BLJ_0712	25.4	VFG0528	Salmonella iron transporter: fur regulated
BLJ_1174	51.39	VFG1405	sigA
BLJ_1258	41.15	VFG1412	sigH
BLJ_1342	33.11	VFG2161	Signal peptidase II
BLJ_0906	21.73	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1923	22.38	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1360	22.88	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1421	23.24	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0611	23.31	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1836	23.32	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0459	23.33	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1278	23.43	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1998	23.51	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1522	23.6	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0109	23.63	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0418	23.69	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0118	23.7	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0520	23.85	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1976	24.27	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0099	24.31	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1605	24.34	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0132	24.53	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1912	24.58	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0912	24.69	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0318	24.71	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1515	24.93	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1933	25	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1718	25	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1997	25.36	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0400	25.37	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_0515	27.08	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1321	28.21	VFG2197	Sugar-binding transcriptional regulator, LacI family
BLJ_1232	29.83	VFG1028	Tn21 integrase Intf1
BLJ_1160	43.1	VFG2168	Transcriptional regulator, Cro/CI family
BLJ_0747	28.98	VFG1122	Transposase ORFAB, subunit B
BLJ_1180	43.64	VFG1398	trpD
BLJ_1871	39.62	VFG1967	UDP-galactopyranose mutase
BLJ_1644	39	VFG2361	UDP-glucose 4-epimerase
BLJ_1680	54.49	VFG2361	UDP-glucose 4-epimerase
BLJ_1891	52.63	VFG0963	UDP-glucose 6-dehydrogenase
BLJ_0697	46.15	VFG1414	whiB3

MIC: Minimum inhibitory concentration; ABC: ATP-binding cassette.

Table 4 Putative genes associated with adhesion identified in the genome of *Bifidobacterium longum* JDM301

Locus_tag	Pfam number	Product name
BLJ_1932	pfam01547	Family 1 extracellular solute-binding protein
BLJ_0112	pfam01547	Family 1 extracellular solute-binding protein
BLJ_1284	pfam01547	Family 1 extracellular solute-binding protein
BLJ_1420	pfam01547	Family 1 extracellular solute-binding protein
BLJ_0131	pfam01547	Family 1 extracellular solute-binding protein
BLJ_1604	pfam01547	Family 1 extracellular solute-binding protein
BLJ_1686	pfam01547	Family 1 extracellular solute-binding protein
BLJ_1964	pfam01547	Family 1 extracellular solute-binding protein
BLJ_1994	pfam01547	Family 1 extracellular solute-binding protein
BLJ_1996	pfam01547	Family 1 extracellular solute-binding protein
BLJ_2001	pfam01547	Family 1 extracellular solute-binding protein
BLJ_0288	pfam01547	Family 1 extracellular solute-binding protein
BLJ_0321	pfam01547	Family 1 extracellular solute-binding protein
BLJ_0345	pfam01547	phosphate ABC transporter periplasmic phosphate-binding protein
BLJ_0414	pfam01547	Family 1 extracellular solute-binding protein
BLJ_0522	pfam01547	Family 1 extracellular solute-binding protein
BLJ_0523	pfam01547	Family 1 extracellular solute-binding protein
BLJ_0524	pfam01547	Family 1 extracellular solute-binding protein
BLJ_0012	pfam07174	Hypothetical protein BLJ_0012
BLJ_1801	pfam05738	LPXTG-motif protein cell wall anchor domain-containing protein
BLJ_0140	pfam07811	TadE family protein

with iron, amino acid and sugar transport, 5 transposases, and 2 glutamine synthetase related to plasminogen (Plg)-binding (Table 3).

Although the ability to adhere to the intestinal wall has been one of the selection criteria for probiotics and also a characteristic of commensal bacteria in the intestine, adhesion is also considered to be a significant step in the initial pathogen infections^[26]. Thus, predicted proteins for adhesion of JDM301 were also included in the analysis of virulence. A total of 21 predicted proteins for adhesion were identified in JDM301 (Table 4). A large number of predicted surface and extracellular proteins were identified in JDM301, which may be involved in the bacterium-host interaction as in other LAB^[27]. A total of 217 proteins with probable Sec-type signal peptides were identified by the tool, Signal P^[28]. The genome of JDM301 also harbors 18 copies of extracellular solute-binding protein (SBP, pfam01547) which is predicted to bind oligosaccharides (SBP family 1) as a component of the ABC transporter complex.

DISCUSSION

As more probiotic strains are used in the food and drug industry, more attentions should be paid to the safety of strains used as probiotics. Thus, the safety of LAB used as probiotics need to be reassessed using the latest technology. *B. longum* JDM301, is a commercial probiotic strain used in many probiotic products sold in China. Analysis of the genome of JDM301 reveals several potential risk factors needing further experimental validation, including a tetracycline resistance gene (*tetW*) with the risk of transfer, and the genes associated with harmful metabolites.

Bifidobacteria were considered free of phage infection until prophage-like elements were identified in the genomes of *B. longum* NCC2705, *B. longum* DJO10A and *B. breve* UCC2003^[29]. Absence of complete prophages is important for the stability of genomes and for industrial applications of probiotic bacteria^[21,30]. Absence of complete prophages and scarcity of IS element may play important roles in promoting genome stability of JDM301^[31]. Another set of genes disseminated by HGT in *Bifidobacteria* is the CRISPR-related system (CASS), which is involved in defense against phages and plasmids^[32]. No CRISPR was discovered in the genome. R-M systems are diverse and widespread in nature and they are considered as barriers to HGT, e.g., **in transformation and phage infection**^[33]. The diversity of R-M systems in *B. longum* JDM301 may be significant to the stability of genome and its use in industry compared with the other two *B. longum* strains.

B. longum JDM301 was not resistant to tetracycline as the minimum inhibitory concentration (8.0 mg/L) was not higher than the breakpoint value (8.0 mg/L)^[34]. However, the MIC for *B. longum* strains **ranges from 0.5 to 2 mg/L in a report**^[35]. Thus, further experiments may be needed to determine the microbiological breakpoint. The *tetW* (BLJ_1245) gene encodes for a ribosomal protection protein and *tetW* genes were responsible for acquired tetracycline resistance in human *B. longum* strains^[36]. The rest of the tetracycline resistance genes found in *B. longum* JDM301 were *tetV* (BLJ_0814), *tetQ* (BLJ_1401) and *tetPB* (BLJ_0594). The gene *tetV* encodes for a tetracycline efflux pump and the genes *tetQ* and *tetPB* encode for ribosomal protection proteins. Further experiments are needed to confirm whether the *tetW* gene in the chromosome of *B. longum* JDM301 is a transferable antibiotic resistance determinant and responsible for resistance to tetracycline in human *B. longum* strains.

The MIC of *B. longum* JDM301 to bacitracin was 26.7 mg/L, **which indicated a moderate resistance. A previous report**^[25] indicated that *B. longum* strains were susceptible to bacitracin. A total of 7 putative bacitracin resistance genes were identified, including 6 genes encoding for ABC transporters and 1 for an uncharacterized bacitracin resistance protein. These genes may be responsible for the resistance to bacitracin.

The resistances to ciprofloxacin, amikacin, gentamicin and streptomycin and susceptibility of JDM301 to vancomycin, amoxicillin, cephalothin, chloramphenicol, erythromycin, ampicillin, cefotaxime, rifampicin, imipenem and an antimicrobial compound, trimethoprim-sulphamethoxazol were consistent with reported findings^[22-25,36]. However, **there are discrepancies between the phenotype and the genotype.** *B. longum* JDM301 was sensitive to vancomycin and chloramphenicol but the genome contained vancomycin and chloramphenicol resistance genes. Further analysis will be needed to determine this discrepancy.

Several cases of D-lactic acidosis associated with consumption of LAB in patients with short bowel syndrome were reported^[37,38], **implying that bacteria used as probiotics should be screened for the ability to generate D-lac-**

tate. In this study, two homologs of DLD genes were identified in the genome of JDM301. Since there were no reported cases of D-lactic acidosis caused by bifidobacteria^[37-39], the activities of these homologous DLDs in bifidobacteria may be low so that the amount of lactate produced is insufficient to cause D-lactic acidosis.

Although biogenic amines (BA) play an important physiological role in mammals, **a high amount of BA in the diet** may have a variety of toxic effects^[40]. The main BA contained in food and beverage includes **histamine, tyramine, putrescine, and cadaverine**, some of which are associated with toxicological characteristics of food poisoning^[41]. The decarboxylase activities of histidine, tyrosine and ornithine were reported in lactobacilli and the capabilities might be strain-dependent rather than species-dependent^[42]. Therefore, **BA production, especially thylamine and tyramine, must be carefully evaluated for individual strains.**

Bacterial enzymes, such as GN, GS, NR, AR and AS, play important roles in the metabolism of carcinogens and other toxicants in the intestine. Homologs of GS are common in sequenced *Bifidobacteria* genomes where GS and GN facilitate the absorption of a variety of toxicants and may contribute to the development of colon cancer. The link between *Bifidobacteria* and the genotoxic enzyme activities of intestinal microflora has been reported^[43,44], with *Bifidobacteria* inhibiting the activity of some genotoxic enzymes^[45]. NR activity is common in oral bacteria and it plays an important role in bacterial nitrate reduction. Although NR activities have **been reported in *Bifidobacteria***, the activity of this enzyme is lower than the NR activity of other gut bacteria^[6].

CBSH mediates microbial bile tolerance and enhances microbial survival in the intestine^[46]. Metagenomic analyses demonstrated that CBSH activity is enriched in the human gut microbiome, and has the potential to greatly influence host physiology^[46]. In *Bifidobacterium* spp. and *Lactobacillus* spp., CBSH activity is also common and nearly all *Bifidobacteria* species and strains have bile salt hydrolase activities^[47]. However, **bile salt hydrolase activity releases free bile acids** which are harmful to the human body and may act as mutagens^[48,49]. Recommendations have been made for absence of bile salt transformation capacity in bacteria added to food^[50]. However, it is noteworthy that the evidence for harmful effects is inconclusive so far and bile salt deconjugation activity may play a role in reducing human serum cholesterol^[51]. Given the huge CBSH pool in intestinal microflora, the CBSH activities of the small number of additional bacteria consumed as probiotics can be ignored^[48].

Putative genes for Plg-binding proteins, DnaK (BLJ_0123) and glutamine synthetase (BLJ_0624 and BLJ_1324) were found in the JDM301 genome, where these proteins play a role in the interaction with human epithelial cells. The protein DnaK has been shown to be present on the surface of pathogens, such as *Neisseria meningitidis*^[52]. The glutamine synthetases BLJ_0624 and BLJ_1324 had a **62.11% and 32.49% similarity to the glutamine synthetases in *Mycobacterium tuberculosis* H37 Rv.** In

the presence of Plg activators, Plg binding to the bacterial surface is converted to plasmin, which is a broad-spectrum serine protease involved in degradation of fibrin and noncollagenous proteins of extracellular matrices and activates latent procollagenases^[53]. It is believed that the capability to intervene with the Plg/plasmin system of a host is a strategy for host colonization and bacterial metastasis shared by several pathogens and commensals of the human intestinal tract^[53,54]. The plasminogen-dependent proteolytic activity of *B. lactis* BI07 and *B. longum* was shown to be dose-dependent^[55,56].

A homolog (BLJ_0880, 24.18% identity) of a gene encoding a component in ferric dicitrate uptake system (Fec) of *Shigella flexneri* serotype 2a, FecE, was identified in the genome of JDM301. As an iron uptake system, Fec is critical for bacterial survival and plays an important role in bacterial virulence^[57]. In addition, BLJ_1105 and BLJ_0409 proteins associated with iron acquisition in JDM301 were 24.49% and 25.56% similar to pyochelin biosynthesis protein in *Pseudomonas aeruginosa*, and BLJ_0712, BLJ_1796 and BLJ_0662 proteins were 25.4%, 27.31 and 29.06% similar to iron transporters of *Salmonella enterica*.

The human pathogen, *Helicobacter pylori*, produces a neutrophil activating protein (NAP) which activate human leukocytes and induces an inflammation, which facilitates the growth of the pathogen^[58]. A homolog (BLJ_0021; 26.83% identity) of the gene encoding a NAP was identified in the genome of JDM301.

In JDM301, BLJ_0012 encodes a protein harboring fibronectin-binding motif (Pfam number 07174) that allows mycobacteria to bind to fibronectin in the extracellular matrix and may mediate the adhesion of JDM301 to its host^[59]. A potential protein for *Bifidobacteria* adhesion to intestinal cells is the putative LPXTG-motif protein with collagen binding motifs (Cna_B, pfam05738) encoded by BLJ_1801, which shows a 34% identity to a predicted fimbrial subunit in the genome of *B. dentium* Bd1. This protein may be involved in the recognition of and adhesion to mucosal epithelial cell surfaces^[19]. Its homologous proteins were also identified in the genome sequences of both *B. longum* NCC2705 and *B. longum* DJO10A genomes^[3,60]. *B. longum* subsp. *infantis* 15697, *B. longum* NCC2705 and *B. adolescentis* contains 21, 10 and 11 copies of extracellular solute-binding protein, respectively^[3,4]. Comparably, the SBP family 1 proteins are more abundant in JDM301 than the three other *Bifidobacteria* strains due to the genome size.

Finally, JDM301 encodes a number of proteases and peptidase that may contribute to virulence owing their ability to degrade host proteins for bacterial nutrition sources^[61]. However, not all the genes associated with virulence have been known until now. Thus, despite the evaluation based on the whole genome sequences, it is recommended that the rat endocarditis and the immunocompromised mouse model should be used for *in vivo* assessment of safety for the low pathogenicity of LAB^[48].

Recently, there has been more interest in using probiotic products to promote health and treat diseases.

Probiotics have been investigated in clinical trials, such as treatment for diarrhea, D-lactic acidosis, necrotizing enterocolitis, inflammatory bowel disease and so on^[59,62-64]. The mechanisms by which probiotics exert their effects are still obscure, which may include modification of gut pH, antagonism of pathogens, modulation of immunity as well as supplements of some nutrients^[65]. However, safety issues of probiotics have been discussed in many reports^[5,48]. There are reported cases of infections associated with probiotic strains^[5]. Although the strain is safe based on phenotype, the information derived from complete bacterial genome sequences reveals some putative unfavorable genes, such as genes encoding for Plg-binding proteins, proteases and genes associated with production of D-lactate. In addition, patients are generally more susceptible to infection and harmful metabolites, such as D-lactate than healthy persons. Thus, the biosafety of probiotics, especially strains used in therapy, must be assessed more carefully and comprehensively.

In conclusion, this study compared the genome of JDM301 with other *Bifidobacteria* and assessed the genomic stability, the potential for antibiotic resistance, the potential for virulence and the potential production of harmful metabolites of this strain. The core genome of *B. longum* is composed of 1265 genes, and 219 genes are unique in JDM301. Our data showed putative virulence genes in the genomes of JDM301 as well as putative genes associated with production of harmful metabolites. In addition, a potentially transferable antibiotic resistance gene was detected in the chromosome of JDM301, which needs to be experimentally validated. This assessment provides information on potential risk factors, which should be further evaluated experimentally, e.g., *in vivo* assessment using animal models.

ACKNOWLEDGMENTS

The authors thank Dr. Hua-Jun Zheng from Shanghai-MOST Key Laboratory of Health and Disease Genomics, Chinese National Human Genome Center at Shanghai for kindly providing us assistance in data analysis.

COMMENTS

Background

Bifidobacterium longum JDM301 is a commercial strain used widely in China with several probiotic functions. Recently, there has been more interest in using probiotic products to promote health and treat diseases. As model probiotic bacteria, *Bifidobacteria* are often added to probiotic products in combination with other lactic acid bacteria. The biosafety of probiotic bacteria is attracting more attentions with its enlarged applications. As more commercial probiotic products are being introduced in the market, it is necessary to reassess the safety of these probiotic products using the latest technology.

Research frontiers

With a long and safe history of application, lactic acid bacteria have acquired the status of "Generally Regarded As Safe". However, published reports of rare infections involving *Lactobacilli* or *Bifidobacteria* are available. The strains selected as probiotics are needed to be assessed carefully and comprehensively. This study may contribute to a better biosafety assessment of probiotic bacteria.

Innovations and breakthroughs

This is the first study to assess the biosafety of probiotic bacteria based on

complete genome sequences. Through bioinformatics analysis of the genome sequences, the authors found that although the strain was safe based on phenotype, the information derived from complete bacterial genome sequences revealed some putative unfavourable genes that should be paid attention to.

Applications

The study provides a comprehensive assessment on potential risk factors of a probiotic strain based on complete genome sequences. The information related to biosafety derived from the genome of JDM301 will contribute to a wider and deeper insight into the safety of probiotic bacteria.

Peer review

This is a very nice and comprehensive study assessing the genomic stability, potential of antibiotic resistance, virulence and production of harmful metabolites. This adds valuable information to current knowledge about probiotics.

REFERENCES

- Ventura M, Canchaya C, Tauch A, Chandra G, Fitzgerald GF, Chater KF, van Sinderen D. Genomics of Actinobacteria: tracing the evolutionary history of an ancient phylum. *Microbiol Mol Biol Rev* 2007; **71**: 495-548
- Marco ML, Pavan S, Kleerebezem M. Towards understanding molecular modes of probiotic action. *Curr Opin Biotechnol* 2006; **17**: 204-210
- Schell MA, Karmirantzou M, Snel B, Vilanova D, Berger B, Pessi G, Zwahlen MC, Desiere F, Bork P, Delley M, Pridmore RD, Arigoni F. The genome sequence of *Bifidobacterium longum* reflects its adaptation to the human gastrointestinal tract. *Proc Natl Acad Sci USA* 2002; **99**: 14422-14427
- Sela DA, Chapman J, Adeuya A, Kim JH, Chen F, Whitehead TR, Lapidus A, Rokhsar DS, Lebrilla CB, German JB, Price NP, Richardson PM, Mills DA. The genome sequence of *Bifidobacterium longum* subsp. *infantis* reveals adaptations for milk utilization within the infant microbiome. *Proc Natl Acad Sci USA* 2008; **105**: 18964-18969
- Borriello SP, Hammes WP, Holzapfel W, Marteau P, Schrezenmeier J, Vaara M, Valtonen V. Safety of probiotics that contain lactobacilli or bifidobacteria. *Clin Infect Dis* 2003; **36**: 775-780
- O'Brien J, Crittenden R, Arthur C. Ouwehand and Seppo Salminen. Safety evaluation of probiotics. *Trends in Food Sci and Technol* 1999; **10**: 418-424
- McBain AJ, Macfarlane GT. Investigations of bifidobacterial ecology and oligosaccharide metabolism in a three-stage compound continuous culture system. *Scand J Gastroenterol Suppl* 1997; **222**: 32-40
- Ruiz-Moyano S, Martín A, Benito MJ, Casquete R, Seradilla MJ, Córdoba MD. Safety and functional aspects of pre-selected lactobacilli for probiotic use in Iberian dry-fermented sausages. *Meat Sci* 2009; Epub ahead of print
- Wei YX, Zhang ZY, Liu C, Zhu YZ, Zhu YQ, Zheng H, Zhao GP, Wang S, Guo XK. Complete genome sequence of *Bifidobacterium longum* JDM301. *J Bacteriol* 2010; **192**: 4076-4077
- Benson DA, Karsch-Mizrachi I, Lipman DJ, Ostell J, Wheeler DL. GenBank. *Nucleic Acids Res* 2008; **36**: D25-D30
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 1999; **27**: 4636-4641
- Guo FB, Ou HY, Zhang CT. ZCURVE: a new system for recognizing protein-coding genes in bacterial and archaeal genomes. *Nucleic Acids Res* 2003; **31**: 1780-1789
- Bose M, Barber RD. Prophage Finder: a prophage loci prediction tool for prokaryotic genome sequences. *In Silico Biol* 2006; **6**: 223-227
- Grissa I, Vergnaud G, Pourcel C. The CRISPRdb database and tools to display CRISPRs and to generate dictionaries of spacers and repeats. *BMC Bioinformatics* 2007; **8**: 172
- Lowe TM, Eddy SR. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 1997; **25**: 955-964
- Ghai R, Hain T, Chakraborty T. GenomeViz: visualizing microbial genomes. *BMC Bioinformatics* 2004; **5**: 198
- Chen L, Yang J, Yu J, Yao Z, Sun L, Shen Y, Jin Q. VFDB: a reference database for bacterial virulence factors. *Nucleic Acids Res* 2005; **33**: D325-D328
- Liu B, Pop M. ARDB--Antibiotic Resistance Genes Database. *Nucleic Acids Res* 2009; **37**: D443-D447
- Ventura M, Turrone F, Zomer A, Foroni E, Giubellini V, Bottacini F, Canchaya C, Claesson MJ, He F, Mantzourani M, Mulas L, Ferrarini A, Gao B, Delledonne M, Henrissat B, Coutinho P, Oggioni M, Gupta RS, Zhang Z, Beighton D, Fitzgerald GF, O'Toole PW, van Sinderen D. The *Bifidobacterium dentium* Bd1 genome sequence reflects its genetic adaptation to the human oral cavity. *PLoS Genet* 2009; **5**: e1000785
- Barrangou R, Briczinski EP, Traeger LL, Loquasto JR, Richards M, Horvath P, Coûté-Monvoisin AC, Leyer G, Rendulic S, Steele JL, Broadbent JR, Oberg T, Dudley EG, Schuster S, Romero DA, Roberts RF. Comparison of the complete genome sequences of *Bifidobacterium animalis* subsp. *lactis* DSM 10140 and BI-04. *J Bacteriol* 2009; **191**: 4144-4151
- Philippe H, Douady CJ. Horizontal gene transfer and phylogenetics. *Curr Opin Microbiol* 2003; **6**: 498-505
- Ammor MS, Flórez AB, Mayo B. Antibiotic resistance in non-enterococcal lactic acid bacteria and bifidobacteria. *Food Microbiol* 2007; **24**: 559-570
- Delgado S, Flórez AB, Mayo B. Antibiotic susceptibility of *Lactobacillus* and *Bifidobacterium* species from the human gastrointestinal tract. *Curr Microbiol* 2005; **50**: 202-207
- Moubareck C, Gavini F, Vaugien L, Butel MJ, Doucet-Populaire F. Antimicrobial susceptibility of bifidobacteria. *J Antimicrob Chemother* 2005; **55**: 38-44
- D'Aimmo MR, Modesto M, Biavati B. Antibiotic resistance of lactic acid bacteria and *Bifidobacterium* spp. isolated from dairy and pharmaceutical products. *Int J Food Microbiol* 2007; **115**: 35-42
- Wilson JW, Schurr MJ, LeBlanc CL, Ramamurthy R, Buchanan KL, Nickerson CA. Mechanisms of bacterial pathogenicity. *Postgrad Med J* 2002; **78**: 216-224
- von Ossowski I, Reunanen J, Satokari R, Vesterlund S, Kankainen M, Huhtinen H, Tynkkyinen S, Salminen S, de Vos WM, Palva A. Mucosal adhesion properties of the probiotic *Lactobacillus rhamnosus* GG SpaCBA and SpaFED pilin subunits. *Appl Environ Microbiol* 2010; **76**: 2049-2057
- Emanuelsson O, Brunak S, von Heijne G, Nielsen H. Locating proteins in the cell using TargetP, SignalP and related tools. *Nat Protoc* 2007; **2**: 953-971
- Ventura M, Lee JH, Canchaya C, Zink R, Leahy S, Moreno-Munoz JA, O'Connell-Motherway M, Higgins D, Fitzgerald GF, O'Sullivan DJ, van Sinderen D. Prophage-like elements in bifidobacteria: insights from genomics, transcription, integration, distribution, and phylogenetic analysis. *Appl Environ Microbiol* 2005; **71**: 8692-8705
- Brussow H. Phages of dairy bacteria. *Annu Rev Microbiol* 2001; **55**: 283-303
- Touchon M, Rocha EP. Causes of insertion sequences abundance in prokaryotic genomes. *Mol Biol Evol* 2007; **24**: 969-981
- Godde JS, Bickerton A. The repetitive DNA elements called CRISPRs and their associated genes: evidence of horizontal transfer among prokaryotes. *J Mol Evol* 2006; **62**: 718-729
- O'Driscoll J, Heiter DF, Wilson GG, Fitzgerald GF, Roberts R, van Sinderen D. A genetic dissection of the LlaJI restriction cassette reveals insights on a novel bacteriophage resistance system. *BMC Microbiol* 2006; **6**: 40
- FEEDAP. Prepared by the Panel on Additives and Products or Substances used in Animal Feed on the Update of the criteria used in the assessment of bacterial resistance to antibiotics of human or veterinary importance. *EFSA* 2008; **732**: 1-15
- Masco L, Van Hoorde K, De Brandt E, Swings J, Huys G.

- Antimicrobial susceptibility of Bifidobacterium strains from humans, animals and probiotic products. *J Antimicrob Chemother* 2006; **58**: 85-94
- 36 **Scott KP**, Melville CM, Barbosa TM, Flint HJ. Occurrence of the new tetracycline resistance gene tet(W) in bacteria from the human gut. *Antimicrob Agents Chemother* 2000; **44**: 775-777
- 37 **Munakata S**, Arakawa C, Kohira R, Fujita Y, Fuchigami T, Mugishima H. A case of D-lactic acid encephalopathy associated with use of probiotics. *Brain Dev* 2010; **32**: 691-694
- 38 **Bongaerts G**, Bakkeren J, Severijnen R, Sperl W, Willems H, Naber T, Wevers R, van Meurs A, Tolboom J. Lactobacilli and acidosis in children with short small bowel. *J Pediatr Gastroenterol Nutr* 2000; **30**: 288-293
- 39 **Uchida H**, Yamamoto H, Kisaki Y, Fujino J, Ishimaru Y, Ikeda H. D-lactic acidosis in short-bowel syndrome managed with antibiotics and probiotics. *J Pediatr Surg* 2004; **39**: 634-636
- 40 **Garai G**, Dueñas MT, Irastorza A, Martín-Alvarez PJ, Moreno-Arribas MV. Biogenic amines in natural ciders. *J Food Prot* 2006; **69**: 3006-3012
- 41 **Silla Santos MH**. Biogenic amines: their importance in foods. *Int J Food Microbiol* 1996; **29**: 213-231
- 42 **Garai G**, Dueñas MT, Irastorza A, Moreno-Arribas MV. Biogenic amine production by lactic acid bacteria isolated from cider. *Lett Appl Microbiol* 2007; **45**: 473-478
- 43 **Benno Y**, Mitsuoka T. Impact of Bifidobacterium longum on human fecal microflora. *Microbiol Immunol* 1992; **36**: 683-694
- 44 **Kim Y**, Lee D, Kim D, Cho J, Yang J, Chung M, Kim K, Ha N. Inhibition of proliferation in colon cancer cell lines and harmful enzyme activity of colon bacteria by Bifidobacterium adolescentis SPM0212. *Arch Pharm Res* 2008; **31**: 468-473
- 45 **Choi SS**, Kang BY, Chung MJ, Kim SD, Park SH, Kim JS, Kang CY, Ha NJ. Safety assessment of potential lactic acid bacteria Bifidobacterium longum SPM1205 isolated from healthy Koreans. *J Microbiol* 2005; **43**: 493-498
- 46 **Jones BV**, Begley M, Hill C, Gahan CG, Marchesi JR. Functional and comparative metagenomic analysis of bile salt hydrolase activity in the human gut microbiome. *Proc Natl Acad Sci USA* 2008; **105**: 13580-13585
- 47 **Tanaka H**, Doesburg K, Iwasaki T, Mierau I. Screening of lactic acid bacteria for bile salt hydrolase activity. *J Dairy Sci* 1999; **82**: 2530-2535
- 48 **Vankerckhoven V**, Huys G, Vancanneyt M, Vael C, Klare I, Romond MB, Entenza JM, Moreillon P, Wind RD, Knol J, Wiertz E, Pot B, Vaughan EE, Kahlmeter G, Goossens H. Biosafety assessment of probiotics used for human consumption: recommendations from the EU-PROSAFE project. *Trends in Food Sci and Technol* 2008; **19**: 102-114
- 49 **Nagengast FM**, Grubben MJ, van Munster IP. Role of bile acids in colorectal carcinogenesis. *Eur J Cancer* 1995; **31A**: 1067-1070
- 50 **Marteau P**, Gerhardt MF, Myara A, Bouvier E, Trivin F, Rambaud JC. Metabolism of Bile Salts by Alimentary Bacteria During Transit in the Human Small Intestine. *Micro Ecol in Heal and Dis* 1995; **8**: 151-157
- 51 **Jones ML**, Chen H, Ouyang W, Metz T, Prakash S. Microencapsulated Genetically Engineered Lactobacillus plantarum 80 (pCBH1) for Bile Acid Deconjugation and Its Implication in Lowering Cholesterol. *J Biomed Biotechnol* 2004; **2004**: 61-69
- 52 **Knaust A**, Weber MV, Hammerschmidt S, Bergmann S, Frosch M, Kurzai O. Cytosolic proteins contribute to surface plasminogen recruitment of Neisseria meningitidis. *J Bacteriol* 2007; **189**: 3246-3255
- 53 **Lähteenmäki K**, Kuusela P, Korhonen TK. Bacterial plasminogen activators and receptors. *FEMS Microbiol Rev* 2001; **25**: 531-552
- 54 **Candela M**, Miccoli G, Bergmann S, Turrone S, Vitali B, Hammerschmidt S, Brigidi P. Plasminogen-dependent proteolytic activity in Bifidobacterium lactis. *Microbiology* 2008; **154**: 2457-2462
- 55 **Candela M**, Bergmann S, Vici M, Vitali B, Turrone S, Eikmanns BJ, Hammerschmidt S, Brigidi P. Binding of human plasminogen to Bifidobacterium. *J Bacteriol* 2007; **189**: 5929-5936
- 56 **Candela M**, Centanni M, Fiori J, Biagi E, Turrone S, Orrico C, Bergmann S, Hammerschmidt S, Brigidi P. DnaK from Bifidobacterium animalis subsp. lactis is a surface-exposed human plasminogen receptor upregulated in response to bile salts. *Microbiology* 2010; **156**: 1609-1618
- 57 **Luck SN**, Turner SA, Rajakumar K, Sakellaris H, Adler B. Ferric dicitrate transport system (Fec) of Shigella flexneri 2a YSH6000 is encoded on a novel pathogenicity island carrying multiple antibiotic resistance genes. *Infect Immun* 2001; **69**: 6012-6021
- 58 **Satin B**, Del Giudice G, Della Bianca V, Dusi S, Laudanna C, Tonello F, Kelleher D, Rappuoli R, Montecucco C, Rossi F. The neutrophil-activating protein (HP-NAP) of Helicobacter pylori is a protective antigen and a major virulence factor. *J Exp Med* 2000; **191**: 1467-1476
- 59 **Verbelen C**, Dufrêne YF. Direct measurement of Mycobacterium-fibronectin interactions. *Integr Biol (Camb)* 2009; **1**: 296-300
- 60 **Lee JH**, Karamychev VN, Kozyavkin SA, Mills D, Pavlov AR, Pavlova NV, Polouchine NN, Richardson PM, Shakhova VV, Slesarev AI, Weimer B, O'Sullivan DJ. Comparative genomic analysis of the gut bacterium Bifidobacterium longum reveals loci susceptible to deletion during pure culture growth. *BMC Genomics* 2008; **9**: 247
- 61 **Gaillot O**, Pellegrini E, Bregenholt S, Nair S, Berche P. The ClpP serine protease is essential for the intracellular parasitism and virulence of Listeria monocytogenes. *Mol Microbiol* 2000; **35**: 1286-1294
- 62 **Alfaleh K**, Anabrees J, Bassler D. Probiotics reduce the risk of necrotizing enterocolitis in preterm infants: a meta-analysis. *Neonatology* 2010; **97**: 93-99
- 63 **McFarland LV**. Meta-analysis of probiotics for the prevention of antibiotic associated diarrhea and the treatment of Clostridium difficile disease. *Am J Gastroenterol* 2006; **101**: 812-822
- 64 **Mach T**. Clinical usefulness of probiotics in inflammatory bowel diseases. *J Physiol Pharmacol* 2006; **57** Suppl 9: 23-33
- 65 **Parvez S**, Malik KA, Ah Kang S, Kim HY. Probiotics and their fermented food products are beneficial for health. *J Appl Microbiol* 2006; **100**: 1171-1185

S- Editor Tian L L- Editor Ma JY E- Editor Zhang DN

Potential implications of *Helicobacter pylori*-related neutrophil-activating protein

Jannis Kountouras, Christos Zavos, Georgia Deretzi, Emmanuel Gavalas, Dimitrios Chatzopoulos, Panagiotis Katsinelos, Elena Tsiaousi, Stergios Gagalis, Stergios A Polyzos, Ioannis Venizelos

Jannis Kountouras, Christos Zavos, Georgia Deretzi, Emmanuel Gavalas, Dimitrios Chatzopoulos, Panagiotis Katsinelos, Elena Tsiaousi, Stergios Gagalis, Stergios A Polyzos, Ioannis Venizelos, Department of Medicine, Second Medical Clinic, Aristotle University of Thessaloniki, Ippokration Hospital, Thessaloniki, Greece

Author contributions: All authors generated the ideas and contributed to the writing of this paper.

Correspondence to: Jannis Kountouras, MD, PhD, Professor of Medicine, Gastroenterologist, Department of Medicine, Second Medical Clinic, Aristotle University of Thessaloniki, Ippokration Hospital, 8 Fanariou St, Byzantio, 55133 Thessaloniki, Macedonia, Greece. jannis@auth.gr

Telephone: +30-2310-892238 Fax: +30-2310-992794

Received: June 14, 2011 Revised: August 18, 2011

Accepted: August 27, 2011

Published online: February 7, 2012

Abstract

Helicobacter pylori (*H. pylori*) virulence factors promote the release of various chemoattractants/inflammatory mediators, including mainly the neutrophil-attractant chemokine interleukin-8 and neutrophil-activating protein (NAP), involved in *H. pylori*-induced gastric pathologies. Co-administration of Chios mastic gum (CMG), which inhibits *H. pylori* NAP, with an *H. pylori* eradication regimen might add clinical benefits against *H. pylori*-related gastric pathologies, but possibly not CMG as main therapy. Although *H. pylori* NAP and other *H. pylori*-related cytotoxins [i.e., vaculating cytotoxin (VacA)] appear to play a major role in generating and maintaining the *H. pylori*-associated gastric inflammatory response and *H. pylori* NAP is a promising vaccine candidate against *H. pylori* infection (*H. pylori*-I), concerns regarding its potential drawbacks, particularly neurogenic ones, due to possible cross-mimicry, should be considered. Possible cross-mimicry between *H. pylori* NAP and/or bacterial aquaporin (AQP) and neural tissues may be associated with the anti-AQP-4 antibody-related neural damage in multiple

sclerosis (MS)/neuromyelitis optica patients. Moreover, the sequence homology found between *H. pylori* VacA and human Na⁺/K⁺-ATPase A subunit suggests that antibodies to VacA involve ion channels in abaxonal Schwann cell plasmalemma resulting in demyelination in some patients. A series of factors have been implicated in inducing blood-brain barrier (BBB) disruption, including inflammatory mediators (e.g., cytokines and chemokines induced by *H. pylori*-I) and oxidative stress. BBB disruption permits access of AQP4-specific antibodies and T lymphocytes to the central nervous system, thereby playing a major role in multiple sclerosis pathogenesis. Relative studies show a strong association between *H. pylori*-I and MS. *H. pylori*-I induces humoral and cellular immune responses that, owing to the sharing of homologous epitopes (molecular mimicry), cross-react with components of nerves, thereby contributing and perpetuating neural tissue damage. Finally, *H. pylori* NAP also plays a possible pathogenetic role in both gastric and colon oncogenesis.

© 2012 Baishideng. All rights reserved.

Key words: *Helicobacter pylori*; Neutrophil-activating protein; Chios mastic gum; Cross-mimicry; Multiple sclerosis; Demyelination; Gastric carcinogenesis

Peer reviewers: Francesco Franceschi, MD, PhD, Assistant Professor, Internal Medicine, Catholic University of Rome, Gemelli Hospital, Largo A. Gemelli, 8, 00168 Rome, Italy; Dr. Orhan Sezgin, Professor, Gastroenteroloji Bilim Dalı, Mersin Üniversitesi Tıp Fakültesi, 33190 Mersin, Turkey; Da-Jun Deng, Professor, Department of Cancer Etiology, Peking University School of Oncology, 1 Da-Hong-Luo-Chang Street, Western District, Beijing 100034, China

Kountouras J, Zavos C, Deretzi G, Gavalas E, Chatzopoulos D, Katsinelos P, Tsiaousi E, Gagalis S, Polyzos SA, Venizelos I. Potential implications of *Helicobacter pylori*-related neutrophil-activating protein. *World J Gastroenterol* 2012; 18(5): 489-490 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v18/i5/489.htm> DOI: <http://dx.doi.org/10.3748/wjg.v18.i5.489>

TO THE EDITOR

In their recent paper published in this journal, Choli-Papadopoulou *et al*^[1] consider the development of new drugs targeting *Helicobacter pylori* (*H. pylori*) neutrophil-activating protein (NAP) and this raises some concerns.

With reference to a study^[2] focusing on *H. pylori* NAP-mediated neutrophil activation before and 2 mo after *per os* administration of Chios mastic gum (CMG), the authors claimed that “these results indicate a substantial down-regulation of the innate cellular immune effectors, which, according to unpublished clinical data in the context of this study, are accompanied by a significant clinical improvement of the patients’ complaints (dyspepsia, epigastric discomfort, distention)”^[1]. However, such clinical benefits cannot be deduced from this study^[2] and, as mentioned, relative clinical data on CMG as treatment for *H. pylori* and peptic ulcer are controversial^[2]. Although *H. pylori* virulence factors promote the release of various chemoattractants/inflammatory mediators including mainly the neutrophil-attractant chemokine interleukin-8 and *H. pylori* NAP involved in *H. pylori*-induced gastric pathologies^[3], our clinical experience suggests that only co-administration of CMG with an *H. pylori* eradication regimen might add clinical benefits against *H. pylori*-related gastric pathologies, but possibly not CMG as main therapy, as the authors claimed^[1,2]. In particular, co-administration of CMG might be a potential therapy to reduce damage of gastric mucosa induced by *H. pylori* NAP. However, large-scale relative prospective studies are needed to elucidate this field.

The authors, further considering data on the safety and immunogenicity of a vaccine comprising *H. pylori*-induced vaculating cytotoxin (VacA), cytotoxin associated gene and *H. pylori* NAP, suggested that the obtained neutrophil activation by the C-terminal region of *H. pylori* NAP opens new pathways for drug design directed at *H. pylori* inflammation^[1]. In particular, both VacA and *H. pylori* NAP play a major role in generating and maintaining the *H. pylori*-associated gastric inflammatory response, and *H. pylori* NAP is a promising vaccine candidate against *H. pylori* infection (*H. pylori*-I). However, concerns regarding potential drawbacks of *H. pylori* NAP, particularly neurogenic ones, should be considered. For instance, possible cross-mimicry between *H. pylori* NAP and/or bacterial aquaporin (AQP) and neural tissues may be associated with the anti-AQP-4 antibody-related neural damage in multiple sclerosis (MS)/neuromyelitis optica (NMO) patients. In this regard, by using histology, the practical gold standard for the diagnosis of *H. pylori*-I, we have shown a strong association between *H. pylori*-I and MS^[4]. *H. pylori*-I induces humoral and cellular immune responses that, owing to the sharing of homologous epitopes (molecular mimicry), cross-react with components of nerves, thereby contributing

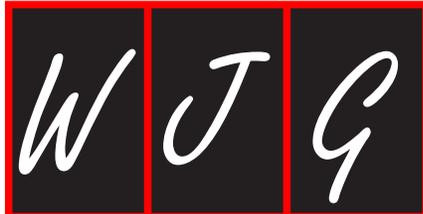
and perpetuating neural tissue damage^[4]. In this respect, *H. pylori* NAP, as a virulence factor, recruits leukocytes from the vascular lumen, and activates neutrophils, monocytes and mast cells, as mentioned by the authors. Besides, the sequence homology found between *H. pylori* VacA and human Na⁺/K⁺-ATPase A subunit suggests that antibodies to VacA involve ion channels in axonal Schwann cell plasmalemma resulting in demyelination in some patients^[5]. Moreover, VacA exhibits chemotactic activities to the bone marrow-derived mast cells (BMDMCs) and induces BMDMCs to produce pro-inflammatory cytokines^[5]. A series of factors have been implicated in inducing blood-brain barrier (BBB) disruption, including the aforementioned inflammatory mediators (e.g., cytokines and chemokines induced by *H. pylori*-I) and oxidative stress. BBB disruption permits access of AQP4-specific antibodies and T lymphocytes to the central nervous system, thereby playing a major role in MS/NMO pathogenesis^[6]. Therefore, *H. pylori* NAP and *HP*-I itself, by inducing several mediators, may influence MS/NMO (including relapsing type) pathophysiology, thereby raising possible concerns regarding even the C-terminal region of *H. pylori* NAP use as a candidate vaccine. Accordingly, relative studies are also needed to clarify the aforementioned concerns.

Finally, the possible *H. pylori* NAP pathogenetic role in gastric carcinogenesis, mentioned by the authors, may also apply to colon oncogenesis^[2,7].

REFERENCES

- 1 Choli-Papadopoulou T, Kottakis F, Papadopoulos G, Pendas S. Helicobacter pylori neutrophil activating protein as target for new drugs against H. pylori inflammation. *World J Gastroenterol* 2011; **17**: 2585-2591
- 2 Kottakis F, Kouzi-Koliakou K, Pendas S, Kountouras J, Choli-Papadopoulou T. Effects of mastic gum Pistacia lentiscus var. Chia on innate cellular immune effectors. *Eur J Gastroenterol Hepatol* 2009; **21**: 143-149
- 3 Kountouras J, Chatzopoulos D, Zavos C. Reactive oxygen metabolites and upper gastrointestinal diseases. *Hepatogastroenterology* 2001; **48**: 743-751
- 4 Gavalas E, Kountouras J, Deretzi G, Boziki M, Grigoriadis N, Zavos C, Venizelos I. Helicobacter pylori and multiple sclerosis. *J Neuroimmunol* 2007; **188**: 187-189; author reply 190
- 5 Kountouras J, Deretzi G, Grigoriadis N, Zavos C, Boziki M, Gavalas E, Katsinelos P, Tzilves D, Giouleme O, Lazaraki G. Guillain-Barré syndrome. *Lancet Neurol* 2008; **7**: 1080-1081; author reply 1083-1085
- 6 Kountouras J, Gavalas E, Deretzi G, Boziki M, Zavos C, Chatzopoulos D, Katsinelos P, Giartzia-Taxidou E, Grigoriadis N, Venizelos I. Helicobacter pylori with or without its neutrophil-activating protein may be the common denominator associated with multiple sclerosis and neuromyelitis optica. *Mult Scler* 2010; **16**: 376-377; author reply 378-379
- 7 Kountouras J, Touloumis L, Karatzoglou P, Zavos C, Chatzopoulos D, Venizelos I, Lazaraki G. In situ H. pylori infection and oncogenes' expression in patients with colorectal cancer. *Gut* 2004; **53**: A270

S- Editor Lv S L- Editor Logan S E- Editor Li JY



Acknowledgments to reviewers of World Journal of Gastroenterology

Many reviewers have contributed their expertise and time to the peer review, a critical process to ensure the quality of *World Journal of Gastroenterology*. The editors and authors of the articles submitted to the journal are grateful to the following reviewers for evaluating the articles (including those published in this issue and those rejected for this issue) during the last editing time period.

Huijie Bian, Professor, Vice-Director, Department of Cell Biololy/Cell Engineering Research Center, Fourth Military Medical University, Xi'an 710032, Shaanxi Province, China

Metin Basaranoglu, MD, PhD, Associate Professor, Gastroenterology Division, Ankara Hospital, Sihhiye, Tr-06000 Ankara, Turkey

Chi-Hin Cho, B.Pharm, PhD, Professor of Pharmacology and Chairman, Department of Pharmacology, Faculty of Medicine, The Chinese University of Hong Kong, Shatin, Hong Kong, China

Lin-Feng Chen, Assistant Professor, Department of Biochemistry, COM 190 MSB, MC-714, University of Illinois at Urbana-Champaign, Urbana, IL 61801, United States

Dr. Seyed Mohsen Dehghani, MD, Associate Professor, Department of Pediatrics, Nemazee Hospital, Shiraz University of Medical Sciences, Shiraz, Iran

Antonio Gasbarrini, MD, Professor, Department of Internal Medicine, Gemelli Hospital, Catholic University of Rome, Largo A. Gemelli 8, 00168 Rome, Italy

Dr. Pankaj Garg, Consultant, Department of General Surgery, Fortis Super Speciality Hospital, Mohali, Punjab, Panchkula 134112, India

Suryakanth R Gurudu, MD, Division of Gastroenterology, 13400 East Shea Blvd, Scottsdale, AZ 85259, United States

Ming-Chih Hou, MD, Professor of Medicine and Director, Department of Internal Medicine, National Yang-Ming University, Taipei 11217, Taiwan, China

Jae J Kim, MD, PhD, Associate Professor, Department of Medicine, Samsung Medical Center, Sungkyunkwan University School of Medicine, 50, Irwon-dong, Gangnam-gu, Seoul 135-710, South Korea

Espen Melum, MD, Medical Department, Rikshospitalet University hospital, Sognsvannsavn 20, Oslo 0027, Norway

Finlay A Macrae, MD, Professor, Department of Colorectal Medicine and Genetics, Royal Melbourne Hospital, PO Box 2010, Victoria 3050, Australia

Kotaro Miyake, MD, PhD, Department of Surgery, Institute of Health Biosciences, The University of Tokushima Graduate School, 3-18-15 Kuramoto, Tokushima 770-8503, Japan

Ross McManus, PhD, Senior Lecturer in Molecular Medicine, Department of Clinical Medicine, Institute of Molecular Medicine, Trinity Centre for Health Science, St. James's Hospital, Dublin 8, Ireland

Zenichi Morise, MD, PhD, Professor, Chairman, Department of Surgery Banbuntane Houtokukai Hospital, Fujita Health University School of Medicine, 3-6-10 Otobashi Nakagawa-ku, Nagoya, Aichi 454-8509, Japan

George Papatheodoridis, MD, Associate Professor in Medicine and Gastroenterology, 2nd Department of Internal Medicine, Athens University Medical School, Hippokraton General Hospital of Athens, 114 Vas. Sophias Ave., 115 27 Athens, Greece

Dr. Stefan Riss, Department of General Surgery, Medical University of Vienna, Währinger Gürtel 18-20, 1090 Vienna, Austria

George Sgourakis, MD, PhD, FACS, 2nd Surgical Department and Surgical Oncology Unit, Red Cross Hospital, 11 Mantzarou Str, Neo Psychiko, Athens 15451, Greece

Venkatesh Shanmugam, MD, MBBS, MS (Gen. Surg.), Dip. NB (Gen. Surg.), FRCS (Glasg.), Specialist Registrar (Trent Deanery), Royal Derby Hospital, Uttoxeter Road, Derby, DE22 3NE, United Kingdom

Hiroaki Takeuchi, MD, PhD, Kochi Medical School, Nankoku-City, Kochi 783-8505, Japan

Dr. Wei-Dong Tong, MD, PhD, Associate Professor, Daping hospital, Third Military Medical University, Chongqing 400042, China

Korkut Uygun, PhD, Assistant Professor in Surgery, Massachusetts General Hospital, Harvard Medical School, 51 Blossom St. Boston, MA 02114, United States

Ruben Zamora, PhD, Assistant Professor of Surgery, Department of Surgery, University of Pittsburgh, W1540 Biomedical Science Tower 200 Lothrop St., Pittsburgh, PA 15213, United States

Events Calendar 2012

January 13-15, 2012
Asian Pacific *Helicobacter pylori*
Meeting 2012
Kuala Lumpur, Malaysia

January 19-21, 2012
American Society of Clinical
Oncology 2012 Gastrointestinal
Cancers Symposium
San Francisco, CA 3000,
United States

January 19-21, 2012
2012 Gastrointestinal Cancers
Symposium
San Francisco, CA 94103,
United States

January 20-21, 2012
American Gastroenterological
Association Clinical Congress of
Gastroenterology and Hepatology
Miami Beach, FL 33141,
United States

February 3, 2012
The Future of Obesity Treatment
London, United Kingdom

February 16-17, 2012
4th United Kingdom Swallowing
Research Group Conference
London, United Kingdom

February 23, 2012
Management of Barretts
Oesophagus: Everything you need
to know
Cambridge, United Kingdom

February 24-27, 2012
Canadian Digestive Diseases Week
2012
Montreal, Canada

March 1-3, 2012
International Conference on
Nutrition and Growth 2012
Paris, France

March 7-10, 2012
Society of American Gastrointestinal
and Endoscopic Surgeons Annual
Meeting
San Diego, CA 92121, United States

March 12-14, 2012
World Congress on
Gastroenterology and Urology
Omaha, NE 68197, United States

March 17-20, 2012
Mayo Clinic Gastroenterology and
Hepatology
Orlando, FL 32808, United States

March 26-27, 2012
26th Annual New Treatments in
Chronic Liver Disease
San Diego, CA 92121, United States

March 30-April 2, 2012
Mayo Clinic Gastroenterology and
Hepatology
San Antonio, TX 78249,
United States

March 31-April 1, 2012
27th Annual New Treatments in
Chronic Liver Disease
San Diego, CA 92121, United States

April 8-10, 2012
9th International Symposium on
Functional GI Disorders
Milwaukee, WI 53202, United States

April 13-15, 2012
Asian Oncology Summit 2012
Singapore, Singapore

April 15-17, 2012
European Multidisciplinary
Colorectal Cancer Congress 2012
Prague, Czech

April 18-20, 2012
The International Liver Congress
2012
Barcelona, Spain

April 19-21, 2012
Internal Medicine 2012
New Orleans, LA 70166,
United States

April 20-22, 2012
Diffuse Small Bowel and Liver
Diseases
Melbourne, Australia

April 22-24, 2012
EUROSON 2012 EFSUMB Annual

Meeting
Madrid, Spain

April 28, 2012
Issues in Pediatric Oncology
Kiev, Ukraine

May 3-5, 2012
9th Congress of The Jordanian
Society of Gastroenterology
Amman, Jordan

May 7-10, 2012
Digestive Diseases Week
Chicago, IL 60601, United States

May 17-21, 2012
2012 ASCRS Annual Meeting-
American Society of Colon and
Rectal Surgeons
Hollywood, FL 1300, United States

May 18-19, 2012
Pancreas Club Meeting
San Diego, CA 92101, United States

May 18-23, 2012
SGNA: Society of Gastroenterology
Nurses and Associates Annual
Course
Phoenix, AZ 85001, United States

May 19-22, 2012
2012-Digestive Disease Week
San Diego, CA 92121, United States

June 2-6, 2012
American Society of Colon and
Rectal Surgeons Annual Meeting
San Antonio, TX 78249,
United States

June 18-21, 2012
Pancreatic Cancer: Progress and
Challenges
Lake Tahoe, NV 89101, United States

July 25-26, 2012
PancreasFest 2012
Pittsburgh, PA 15260, United States

September 1-4, 2012
OESO 11th World Conference
Como, Italy

September 6-8, 2012
2012 Joint International

Neurogastroenterology and Motility
Meeting
Bologna, Italy

September 7-9, 2012
The Viral Hepatitis Congress
Frankfurt, Germany

September 8-9, 2012
New Advances in Inflammatory
Bowel Disease
La Jolla, CA 92093, United States

September 8-9, 2012
Florida Gastroenterologic Society
2012 Annual Meeting
Boca Raton, FL 33498, United States

September 15-16, 2012
Current Problems of
Gastroenterology and Abdominal
Surgery
Kiev, Ukraine

September 20-22, 2012
1st World Congress on Controversies
in the Management of Viral Hepatitis
Prague, Czech

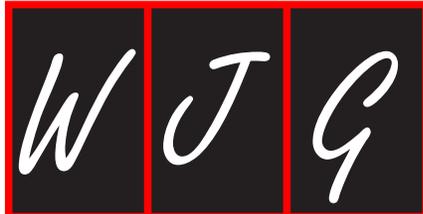
October 19-24, 2012
American College of
Gastroenterology 77th Annual
Scientific Meeting and Postgraduate
Course
Las Vegas, NV 89085, United States

November 3-4, 2012
Modern Technologies in
Diagnosis and Treatment of
Gastroenterological Patients
Dnepropetrovsk, Ukraine

November 4-8, 2012
The Liver Meeting
San Francisco, CA 94101,
United States

November 9-13, 2012
American Association for the Study
of Liver Diseases
Boston, MA 02298, United States

December 1-4, 2012
Advances in Inflammatory Bowel
Diseases
Hollywood, FL 33028, United States



GENERAL INFORMATION

World Journal of Gastroenterology (*World J Gastroenterol*, *WJG*, print ISSN 1007-9327, online ISSN 2219-2840, DOI: 10.3748) is a weekly, open-access (OA), peer-reviewed journal supported by an editorial board of 1361 experts in gastroenterology and hepatology from 64 countries.

The biggest advantage of the OA model is that it provides free, full-text articles in PDF and other formats for experts and the public without registration, which eliminates the obstacle that traditional journals possess and usually delays the speed of the propagation and communication of scientific research results. The open access model has been proven to be a true approach that may achieve the ultimate goal of the journals, i.e. the maximization of the value to the readers, authors and society.

Maximization of personal benefits

The role of academic journals is to exhibit the scientific levels of a country, a university, a center, a department, and even a scientist, and build an important bridge for communication between scientists and the public. As we all know, the significance of the publication of scientific articles lies not only in disseminating and communicating innovative scientific achievements and academic views, as well as promoting the application of scientific achievements, but also in formally recognizing the “priority” and “copyright” of innovative achievements published, as well as evaluating research performance and academic levels. So, to realize these desired attributes of *WJG* and create a well-recognized journal, the following four types of personal benefits should be maximized. The maximization of personal benefits refers to the pursuit of the maximum personal benefits in a well-considered optimal manner without violation of the laws, ethical rules and the benefits of others. (1) Maximization of the benefits of editorial board members: The primary task of editorial board members is to give a peer review of an unpublished scientific article *via* online office system to evaluate its innovativeness, scientific and practical values and determine whether it should be published or not. During peer review, editorial board members can also obtain cutting-edge information in that field at first hand. As leaders in their field, they have priority to be invited to write articles and publish commentary articles. We will put peer reviewers’ names and affiliations along with the article they reviewed in the journal to acknowledge their contribution; (2) Maximization of the benefits of authors: Since *WJG* is an open-access journal, readers around the world can immediately download and read, free of charge, high-quality, peer-reviewed articles from *WJG* official website, thereby realizing the goals and significance of the communication between authors and peers as well as public reading; (3) Maximization of the benefits of readers: Readers can read or use, free of charge, high-quality peer-reviewed articles without any limits, and cite the arguments, viewpoints, concepts, theories, methods, results, conclusion or facts and data of pertinent literature so as to validate the innovativeness, scientific and practical values of their own research achievements, thus ensuring that their articles have novel arguments or viewpoints, solid

evidence and correct conclusion; and (4) Maximization of the benefits of employees: It is an iron law that a first-class journal is unable to exist without first-class editors, and only first-class editors can create a first-class academic journal. We insist on strengthening our team cultivation and construction so that every employee, in an open, fair and transparent environment, could contribute their wisdom to edit and publish high-quality articles, thereby realizing the maximization of the personal benefits of editorial board members, authors and readers, and yielding the greatest social and economic benefits.

Aims and scope

The major task of *WJG* is to report rapidly the most recent results in basic and clinical research on esophageal, gastrointestinal, liver, pancreas and biliary tract diseases, *Helicobacter pylori*, endoscopy and gastrointestinal surgery, including: gastroesophageal reflux disease, gastrointestinal bleeding, infection and tumors; gastric and duodenal disorders; intestinal inflammation, microflora and immunity; celiac disease, dyspepsia and nutrition; viral hepatitis, portal hypertension, liver fibrosis, liver cirrhosis, liver transplantation, and metabolic liver disease; molecular and cell biology; geriatric and pediatric gastroenterology; diagnosis and screening, imaging and advanced technology.

Columns

The columns in the issues of *WJG* will include: (1) Editorial: To introduce and comment on major advances and developments in the field; (2) Frontier: To review representative achievements, comment on the state of current research, and propose directions for future research; (3) Topic Highlight: This column consists of three formats, including (A) 10 invited review articles on a hot topic, (B) a commentary on common issues of this hot topic, and (C) a commentary on the 10 individual articles; (4) Observation: To update the development of old and new questions, highlight unsolved problems, and provide strategies on how to solve the questions; (5) Guidelines for Basic Research: To provide guidelines for basic research; (6) Guidelines for Clinical Practice: To provide guidelines for clinical diagnosis and treatment; (7) Review: To review systematically progress and unresolved problems in the field, comment on the state of current research, and make suggestions for future work; (8) Original Article: To report innovative and original findings in gastroenterology; (9) Brief Article: To briefly report the novel and innovative findings in gastroenterology and hepatology; (10) Case Report: To report a rare or typical case; (11) Letters to the Editor: To discuss and make reply to the contributions published in *WJG*, or to introduce and comment on a controversial issue of general interest; (12) Book Reviews: To introduce and comment on quality monographs of gastroenterology and hepatology; and (13) Guidelines: To introduce consensus and guidelines reached by international and national academic authorities worldwide on basic research and clinical practice gastroenterology and hepatology.

Name of journal

World Journal of Gastroenterology

Instructions to authors

ISSN and EISSN

ISSN 1007-9327 (print)
ISSN 2219-2840 (online)

Editor-in-chief

Ferruccio Bonino, MD, PhD, Professor of Gastroenterology, Director of Liver and Digestive Disease Division, Department of Internal Medicine, University of Pisa, Director of General Medicine 2 Unit University Hospital of Pisa, Via Roma 67, 56124 Pisa, Italy

Myung-Hwan Kim, MD, PhD, Professor, Head, Department of Gastroenterology, Director, Center for Biliary Diseases, University of Ulsan College of Medicine, Asan Medical Center, 388-1 Pungnap-2dong, Songpa-gu, Seoul 138-736, South Korea

Kjell Öberg, MD, PhD, Professor, Department of Endocrine Oncology, Uppsala University Hospital, SE-751 85 Uppsala, Sweden

Matt D Rutter, MBBS, MD, FRCP, Consultant Gastroenterologist, Senior Lecturer, Director, Tees Bowel Cancer Screening Centre, University Hospital of North Tees, Durham University, Stockton-on-Tees, Cleveland TS19 8PE, United Kingdom

Andrzej S Tarnawski, MD, PhD, DSc (Med), Professor of Medicine, Chief of Gastroenterology, VA Long Beach Health Care System, University of California, Irvine, CA, 5901 E. Seventh Str., Long Beach, CA 90822, United States

Editorial office

World Journal of Gastroenterology
Editorial Department: Room 903, Building D,
Ocean International Center,
No. 62 Dongsihuan Zhonglu,
Chaoyang District, Beijing 100025, China
E-mail: wjg@wjgnet.com
<http://www.wjgnet.com>
Telephone: +86-10-59080039
Fax: +86-10-85381893

Indexed and abstracted in

Current Contents[®]/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch[®]), Journal Citation Reports[®], Index Medicus, MEDLINE, PubMed, PubMed Central, Digital Object Identifier, and Directory of Open Access Journals. ISI, Thomson Reuters, 2010 Impact Factor: 2.240 (35/71 Gastroenterology and Hepatology).

Published by

Baishideng Publishing Group Co., Limited

SPECIAL STATEMENT

All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

Biostatistical editing

Statistical review is performed after peer review. We invite an expert in Biomedical Statistics from to evaluate the statistical method used in the paper, including *t*-test (group or paired comparisons), chi-squared test, Ridit, probit, logit, regression (linear, curvilinear, or stepwise), correlation, analysis of variance, analysis of covariance, *etc.* The reviewing points include: (1) Statistical methods should be described when they are used to verify the results; (2) Whether the statistical techniques are suitable or correct; (3) Only

homogeneous data can be averaged. Standard deviations are preferred to standard errors. Give the number of observations and subjects (*n*). Losses in observations, such as drop-outs from the study should be reported; (4) Values such as ED50, LD50, IC50 should have their 95% confidence limits calculated and compared by weighted probit analysis (Bliss and Finney); and (5) The word ‘significantly’ should be replaced by its synonyms (if it indicates extent) or the *P* value (if it indicates statistical significance).

Conflict-of-interest statement

In the interests of transparency and to help reviewers assess any potential bias, *WJG* requires authors of all papers to declare any competing commercial, personal, political, intellectual, or religious interests in relation to the submitted work. Referees are also asked to indicate any potential conflict they might have reviewing a particular paper. Before submitting, authors are suggested to read “Uniform Requirements for Manuscripts Submitted to Biomedical Journals: Ethical Considerations in the Conduct and Reporting of Research: Conflicts of Interest” from International Committee of Medical Journal Editors (ICMJE), which is available at: http://www.icmje.org/ethical_4conflicts.html.

Sample wording: [Name of individual] has received fees for serving as a speaker, a consultant and an advisory board member for [names of organizations], and has received research funding from [names of organization]. [Name of individual] is an employee of [name of organization]. [Name of individual] owns stocks and shares in [name of organization]. [Name of individual] owns patent [patent identification and brief description].

Statement of informed consent

Manuscripts should contain a statement to the effect that all human studies have been reviewed by the appropriate ethics committee or it should be stated clearly in the text that all persons gave their informed consent prior to their inclusion in the study. Details that might disclose the identity of the subjects under study should be omitted. Authors should also draw attention to the Code of Ethics of the World Medical Association (Declaration of Helsinki, 1964, as revised in 2004).

Statement of human and animal rights

When reporting the results from experiments, authors should follow the highest standards and the trial should conform to Good Clinical Practice (for example, US Food and Drug Administration Good Clinical Practice in FDA-Regulated Clinical Trials; UK Medicines Research Council Guidelines for Good Clinical Practice in Clinical Trials) and/or the World Medical Association Declaration of Helsinki. Generally, we suggest authors follow the lead investigator’s national standard. If doubt exists whether the research was conducted in accordance with the above standards, the authors must explain the rationale for their approach and demonstrate that the institutional review body explicitly approved the doubtful aspects of the study.

Before submitting, authors should make their study approved by the relevant research ethics committee or institutional review board. If human participants were involved, manuscripts must be accompanied by a statement that the experiments were undertaken with the understanding and appropriate informed consent of each. Any personal item or information will not be published without explicit consents from the involved patients. If experimental animals were used, the materials and methods (experimental procedures) section must clearly indicate that appropriate measures were taken to minimize pain or discomfort, and details of animal care should be provided.

SUBMISSION OF MANUSCRIPTS

Manuscripts should be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Number all pages consecutively, and start each of the following sections on a new page: Title Page, Abstract, Introduction, Materials and Methods, Results, Discussion, Acknowledgements, References, Tables, Figures, and Figure Legends. Neither the editors nor the publisher are responsible for the opinions expressed by contributors. Manuscripts formally accepted for publication become the permanent property of Baishideng Publishing Group Co., Limited, and may not be reproduced by any means, in whole or in part, without the written permission of both the authors and the publisher. We reserve the right to copy-edit and put onto our website accepted manuscripts. Authors should follow the relevant guidelines for the care and use of laboratory animals of their institution or national animal welfare committee. For the sake of transparency in regard to the performance and reporting of clinical trials, we endorse the policy of the ICMJE to refuse to publish papers on clinical trial results if the trial was not recorded in a publicly-accessible registry at its outset. The only register now available, to our knowledge, is <http://www.clinicaltrials.gov> sponsored by the United States National Library of Medicine and we encourage all potential contributors to register with it. However, in the case that other registers become available you will be duly notified. A letter of recommendation from each author's organization should be provided with the contributed article to ensure the privacy and secrecy of research is protected.

Authors should retain one copy of the text, tables, photographs and illustrations because rejected manuscripts will not be returned to the author(s) and the editors will not be responsible for loss or damage to photographs and illustrations sustained during mailing.

Online submissions

Manuscripts should be submitted through the Online Submission System at: <http://www.wjgnet.com/1007-9327/office>. Authors are highly recommended to consult the ONLINE INSTRUCTIONS TO AUTHORS (http://www.wjgnet.com/1007-9327/g_info_20100315215714.htm) before attempting to submit online. For assistance, authors encountering problems with the Online Submission System may send an email describing the problem to wjg@wjgnet.com, or by telephone: +86-10-5908-0039. If you submit your manuscript online, do not make a postal contribution. Repeated online submission for the same manuscript is strictly prohibited.

MANUSCRIPT PREPARATION

All contributions should be written in English. All articles must be submitted using word-processing software. All submissions must be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Style should conform to our house format. Required information for each of the manuscript sections is as follows:

Title page

Title: Title should be less than 12 words.

Running title: A short running title of less than 6 words should be provided.

Authorship: Authorship credit should be in accordance with the standard proposed by ICMJE, based on (1) substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; (2) drafting the article or revising it critically

for important intellectual content; and (3) final approval of the version to be published. Authors should meet conditions 1, 2, and 3.

Institution: Author names should be given first, then the complete name of institution, city, province and postcode. For example, Xu-Chen Zhang, Li-Xin Mei, Department of Pathology, Chengde Medical College, Chengde 067000, Hebei Province, China. One author may be represented from two institutions, for example, George Sgourakis, Department of General, Visceral, and Transplantation Surgery, Essen 45122, Germany; George Sgourakis, 2nd Surgical Department, Korgialenio-Benakio Red Cross Hospital, Athens 15451, Greece.

Author contributions: The format of this section should be: Author contributions: Wang CL and Liang L contributed equally to this work; Wang CL, Liang L, Fu JF, Zou CC, Hong F and Wu XM designed the research; Wang CL, Zou CC, Hong F and Wu XM performed the research; Xue JZ and Lu JR contributed new reagents/analytic tools; Wang CL, Liang L and Fu JF analyzed the data; and Wang CL, Liang L and Fu JF wrote the paper.

Supportive foundations: The complete name and number of supportive foundations should be provided, e.g. Supported by National Natural Science Foundation of China, No. 30224801

Correspondence to: Only one corresponding address should be provided. Author names should be given first, then author title, affiliation, the complete name of institution, city, postcode, province, country, and email. All the letters in the email should be in lower case. A space interval should be inserted between country name and email address. For example, Montgomery Bissell, MD, Professor of Medicine, Chief, Liver Center, Gastroenterology Division, University of California, Box 0538, San Francisco, CA 94143, United States. montgomery.bissell@ucsf.edu

Telephone and fax: Telephone and fax should consist of +, country number, district number and telephone or fax number, e.g. Telephone: +86-10-59080039 Fax: +86-10-85381893

Peer reviewers: All articles received are subject to peer review. Normally, three experts are invited for each article. Decision for acceptance is made only when at least two experts recommend an article for publication. Reviewers for accepted manuscripts are acknowledged in each manuscript, and reviewers of articles which were not accepted will be acknowledged at the end of each issue. To ensure the quality of the articles published in *WJG*, reviewers of accepted manuscripts will be announced by publishing the name, title/position and institution of the reviewer in the footnote accompanying the printed article. For example, reviewers: Professor Jing-Yuan Fang, Shanghai Institute of Digestive Disease, Shanghai, Affiliated Renji Hospital, Medical Faculty, Shanghai Jiaotong University, Shanghai, China; Professor Xin-Wei Han, Department of Radiology, The First Affiliated Hospital, Zhengzhou University, Zhengzhou, Henan Province, China; and Professor Anren Kuang, Department of Nuclear Medicine, Huaxi Hospital, Sichuan University, Chengdu, Sichuan Province, China.

Abstract

There are unstructured abstracts (no less than 256 words) and structured abstracts (no less than 480). The specific requirements for structured abstracts are as follows:

An informative, structured abstracts of no less than 480 words should accompany each manuscript. Abstracts for original contributions should be structured into the following sections.

Instructions to authors

AIM (no more than 20 words): Only the purpose should be included. Please write the aim as the form of "To investigate/study/..."; MATERIALS AND METHODS (no less than 140 words); RESULTS (no less than 294 words): You should present *P* values where appropriate and must provide relevant data to illustrate how they were obtained, e.g. 6.92 ± 3.86 vs 3.61 ± 1.67 , $P < 0.001$; CONCLUSION (no more than 26 words).

Key words

Please list 5-10 key words, selected mainly from *Index Medicus*, which reflect the content of the study.

Text

For articles of these sections, original articles and brief articles, the main text should be structured into the following sections: INTRODUCTION, MATERIALS AND METHODS, RESULTS and DISCUSSION, and should include appropriate Figures and Tables. Data should be presented in the main text or in Figures and Tables, but not in both. The main text format of these sections, editorial, topic highlight, case report, letters to the editors, can be found at: http://www.wjgnet.com/1007-9327/g_info_20100315215714.htm.

Illustrations

Figures should be numbered as 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each figure on a separate page. Detailed legends should not be provided under the figures. This part should be added into the text where the figures are applicable. Figures should be either Photoshop or Illustrator files (in tiff, eps, jpeg formats) at high-resolution. Examples can be found at: <http://www.wjgnet.com/1007-9327/13/4520.pdf>; <http://www.wjgnet.com/1007-9327/13/4554.pdf>; <http://www.wjgnet.com/1007-9327/13/4891.pdf>; <http://www.wjgnet.com/1007-9327/13/4986.pdf>; <http://www.wjgnet.com/1007-9327/13/4498.pdf>. Keeping all elements compiled is necessary in line-art image. Scale bars should be used rather than magnification factors, with the length of the bar defined in the legend rather than on the bar itself. File names should identify the figure and panel. Avoid layering type directly over shaded or textured areas. Please use uniform legends for the same subjects. For example: Figure 1 Pathological changes in atrophic gastritis after treatment. A...; B...; C...; D...; E...; F...; G: ...*etc.* It is our principle to publish high resolution-figures for the printed and E-versions.

Tables

Three-line tables should be numbered 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each table. Detailed legends should not be included under tables, but rather added into the text where applicable. The information should complement, but not duplicate the text. Use one horizontal line under the title, a second under column heads, and a third below the Table, above any footnotes. Vertical and italic lines should be omitted.

Notes in tables and illustrations

Data that are not statistically significant should not be noted. ^a $P < 0.05$, ^b $P < 0.01$ should be noted ($P > 0.05$ should not be noted). If there are other series of *P* values, ^c $P < 0.05$ and ^d $P < 0.01$ are used. A third series of *P* values can be expressed as ^e $P < 0.05$ and ^f $P < 0.01$. Other notes in tables or under illustrations should be expressed as ¹F, ²F, ³F; or sometimes as other symbols with a superscript (Arabic numerals) in the upper left corner. In a multi-curve illustration, each curve should be la-

beled with ●, ○, ■, □, ▲, △, *etc.*, in a certain sequence.

Acknowledgments

Brief acknowledgments of persons who have made genuine contributions to the manuscript and who endorse the data and conclusions should be included. Authors are responsible for obtaining written permission to use any copyrighted text and/or illustrations.

REFERENCES

Coding system

The author should number the references in Arabic numerals according to the citation order in the text. Put reference numbers in square brackets in superscript at the end of citation content or after the cited author's name. For citation content which is part of the narration, the coding number and square brackets should be typeset normally. For example, "Crohn's disease (CD) is associated with increased intestinal permeability^[1,2]". If references are cited directly in the text, they should be put together within the text, for example, "From references^[19,22-24], we know that...".

When the authors write the references, please ensure that the order in text is the same as in the references section, and also ensure the spelling accuracy of the first author's name. Do not list the same citation twice.

PMID and DOI

Please provide PubMed citation numbers to the reference list, e.g. PMID and DOI, which can be found at <http://www.ncbi.nlm.nih.gov/sites/entrez?db=pubmed> and <http://www.crossref.org/SimpleTextQuery/>, respectively. The numbers will be used in E-version of this journal.

Style for journal references

Authors: the name of the first author should be typed in bold-faced letters. The family name of all authors should be typed with the initial letter capitalized, followed by their abbreviated first and middle initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR). The title of the cited article and italicized journal title (journal title should be in its abbreviated form as shown in PubMed), publication date, volume number (in black), start page, and end page [PMID: 11819634 DOI: 10.3748/wjg.13.5396].

Style for book references

Authors: the name of the first author should be typed in bold-faced letters. The surname of all authors should be typed with the initial letter capitalized, followed by their abbreviated middle and first initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR) Book title. Publication number. Publication place: Publication press, Year: start page and end page.

Format

Journals

English journal article (list all authors and include the PMID where applicable)

- 1 **Jung EM**, Clevert DA, Schreyer AG, Schmitt S, Rennert J, Kubale R, Feuerbach S, Jung F. Evaluation of quantitative contrast harmonic imaging to assess malignancy of liver tumors: A prospective controlled two-center study. *World J Gastroenterol* 2007; **13**: 6356-6364 [PMID: 18081224 DOI: 10.3748/wjg.13.6356]

Chinese journal article (list all authors and include the PMID where applicable)

- 2 **Lin GZ**, Wang XZ, Wang P, Lin J, Yang FD. Immunolog-

ic effect of Jianpi Yishen decoction in treatment of Pixu-diarrhoea. *Shijie Huaren Xiaohua Zazhi* 1999; 7: 285-287

In press

- 3 **Tian D**, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. *Proc Natl Acad Sci USA* 2006; In press

Organization as author

- 4 **Diabetes Prevention Program Research Group**. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; 40: 679-686 [PMID: 12411462 PMID:2516377 DOI:10.1161/01.HYP.0000035706.28494.09]

Both personal authors and an organization as author

- 5 **Vallancien G**, Emberton M, Harving N, van Moorselaar RJ; Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; 169: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

- 6 21st century heart solution may have a sting in the tail. *BMJ* 2002; 325: 184 [PMID: 12142303 DOI:10.1136/bmj.325.7357.184]

Volume with supplement

- 7 **Geraud G**, Spierings EL, Keywood C. Tolerability and safety of frovatriptan with short- and long-term use for treatment of migraine and in comparison with sumatriptan. *Headache* 2002; 42 Suppl 2: S93-99 [PMID: 12028325 DOI:10.1046/j.1526-4610.42.s2.7.x]

Issue with no volume

- 8 **Banit DM**, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (401): 230-238 [PMID: 12151900 DOI:10.1097/00003086-200208000-00026]

No volume or issue

- 9 Outreach: Bringing HIV-positive individuals into care. *HRS A Careaction* 2002; 1-6 [PMID: 12154804]

Books

Personal author(s)

- 10 **Sherlock S**, Dooley J. Diseases of the liver and biliary system. 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296

Chapter in a book (list all authors)

- 11 **Lam SK**. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450

Author(s) and editor(s)

- 12 **Breedlove GK**, Schorfheide AM. Adolescent pregnancy. 2nd ed. Wiczorek RR, editor. White Plains (NY): March of Dimes Education Services, 2001: 20-34

Conference proceedings

- 13 **Harnden P**, Joffe JK, Jones WG, editors. Germ cell tumours V. Proceedings of the 5th Germ cell tumours Conference; 2001 Sep 13-15; Leeds, UK. New York: Springer, 2002: 30-56

Conference paper

- 14 **Christensen S**, Oppacher F. An analysis of Koza's computational effort statistic for genetic programming. In: Foster JA, Lutton E, Miller J, Ryan C, Tettamanzi AG, editors. Genetic programming. EuroGP 2002: Proceedings of the 5th European Conference on Genetic Programming; 2002 Apr 3-5; Kinsdale, Ireland. Berlin: Springer, 2002: 182-191

Electronic journal (list all authors)

- 15 Morse SS. Factors in the emergence of infectious dis-

eases. *Emerg Infect Dis* serial online, 1995-01-03, cited 1996-06-05; 1(1): 24 screens. Available from: URL: <http://www.cdc.gov/ncidod/eid/index.htm>

Patent (list all authors)

- 16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1

Statistical data

Write as mean \pm SD or mean \pm SE.

Statistical expression

Express *t* test as *t* (in italics), *F* test as *F* (in italics), chi square test as χ^2 (in Greek), related coefficient as *r* (in italics), degree of freedom as ν (in Greek), sample number as *n* (in italics), and probability as *P* (in italics).

Units

Use SI units. For example: body mass, *m* (B) = 78 kg; blood pressure, *p* (B) = 16.2/12.3 kPa; incubation time, *t* (incubation) = 96 h; blood glucose concentration, *c* (glucose) 6.4 ± 2.1 mmol/L; blood CEA mass concentration, *p* (CEA) = 8.6 24.5 μ g/L; CO₂ volume fraction, 50 mL/L CO₂, not 5% CO₂; likewise for 40 g/L formaldehyde, not 10% formalin; and mass fraction, 8 ng/g, etc. Arabic numerals such as 23, 243, 641 should be read 23 243 641.

The format for how to accurately write common units and quantum can be found at: http://www.wjgnet.com/1007-9327/g_info_20100315223018.htm.

Abbreviations

Standard abbreviations should be defined in the abstract and on first mention in the text. In general, terms should not be abbreviated unless they are used repeatedly and the abbreviation is helpful to the reader. Permissible abbreviations are listed in Units, Symbols and Abbreviations: A Guide for Biological and Medical Editors and Authors (Ed. Baron DN, 1988) published by The Royal Society of Medicine, London. Certain commonly used abbreviations, such as DNA, RNA, HIV, LD50, PCR, HBV, ECG, WBC, RBC, CT, ESR, CSF, IgG, ELISA, PBS, ATP, EDTA, mAb, can be used directly without further explanation.

Italics

Quantities: *t* time or temperature, *c* concentration, *A* area, *l* length, *m* mass, *V* volume.

Genotypes: *gyrA*, *arg 1*, *c myc*, *c fos*, etc.

Restriction enzymes: *EcoRI*, *HindI*, *BamHI*, *Kho I*, *Kpn I*, etc.

Biology: *H. pylori*, *E. coli*, etc.

Examples for paper writing

Editorial: http://www.wjgnet.com/1007-9327/g_info_20100315220036.htm

Frontier: http://www.wjgnet.com/1007-9327/g_info_20100315220305.htm

Topic highlight: http://www.wjgnet.com/1007-9327/g_info_20100315220601.htm

Observation: http://www.wjgnet.com/1007-9327/g_info_20100312232427.htm

Guidelines for basic research: http://www.wjgnet.com/1007-9327/g_info_20100315220730.htm

Instructions to authors

Guidelines for clinical practice: http://www.wjgnet.com/1007-9327/g_info_20100315221301.htm

Review: http://www.wjgnet.com/1007-9327/g_info_20100315221554.htm

Original articles: http://www.wjgnet.com/1007-9327/g_info_20100315221814.htm

Brief articles: http://www.wjgnet.com/1007-9327/g_info_20100312231400.htm

Case report: http://www.wjgnet.com/1007-9327/g_info_20100315221946.htm

Letters to the editor: http://www.wjgnet.com/1007-9327/g_info_20100315222254.htm

Book reviews: http://www.wjgnet.com/1007-9327/g_info_20100312231947.htm

Guidelines: http://www.wjgnet.com/1007-9327/g_info_20100312232134.htm

RESUBMISSION OF THE REVISED MANUSCRIPTS

Please revise your article according to the revision policies of *WJG*. The revised version including manuscript and high-resolution image figures (if any) should be re-submitted online (<http://www.wjgnet.com/1007-9327/office/>). The author should send the copyright transfer letter, responses to the reviewers, English language Grade B certificate (for non-native speakers of English) and final manuscript checklist to wjg@wjgnet.com.

Language evaluation

The language of a manuscript will be graded before it is sent for revision. (1) Grade A: priority publishing; (2) Grade B: minor language polishing; (3) Grade C: a great deal of language polishing needed; and (4) Grade D: rejected. Revised articles should reach Grade A or B.

Copyright assignment form

Please download a Copyright assignment form from http://www.wjgnet.com/1007-9327/g_info_20100315222818.htm.

Responses to reviewers

Please revise your article according to the comments/suggestions provided by the reviewers. The format for responses to the reviewers' comments can be found at: http://www.wjgnet.com/1007-9327/g_info_20100315222607.htm.

Proof of financial support

For paper supported by a foundation, authors should provide a copy of the document and serial number of the foundation.

Links to documents related to the manuscript

WJG will be initiating a platform to promote dynamic interactions between the editors, peer reviewers, readers and authors. After a manuscript is published online, links to the PDF version of the submitted manuscript, the peer-reviewers' report and the revised manuscript will be put on-line. Readers can make comments on the peer reviewer's report, authors' responses to peer reviewers, and the revised manuscript. We hope that authors will benefit from this feedback and be able to revise the manuscript accordingly in a timely manner.

Science news releases

Authors of accepted manuscripts are suggested to write a science news item to promote their articles. The news will be released rapidly at EurekAlert/AAAS (<http://www.eurekalert.org>). The title for news items should be less than 90 characters; the summary should be less than 75 words; and main body less than 500 words. Science news items should be lawful, ethical, and strictly based on your original content with an attractive title and interesting pictures.

Publication fee

WJG is an international, peer-reviewed, Open-Access, online journal. Articles published by this journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license. Authors of accepted articles must pay a publication fee. The related standards are as follows. Publication fee: 1300 USD per article. Editorial, topic highlights, book reviews and letters to the editor are published free of charge.