# World Journal of *Gastroenterology*

World J Gastroenterol 2021 September 21; 27(35): 5796-5988





Published by Baishideng Publishing Group Inc

JG  $\mathcal{N}$ 

# World Journal of VVoriu jour. Gastroenterology

# Contents

Weekly Volume 27 Number 35 September 21, 2021

# **EDITORIAL**

5796 Induced pluripotent stem cells as an innovative model to study drug induced pancreatitis								
	Genova E, Stocco G, Decorti G							
	OPINION REVIEW							
5803	Screening for nonalcoholic fatty liver disease-when, who and how?							
	Dietrich CG, Rau M, Geier A							
	REVIEW							
5822	Environmental perspectives of COVID-19 outbreaks: A review							
	Samanta P, Ghosh AR							

5851 Pancreatic cancer in 2021: What you need to know to win

Tonini V, Zanni M

5890 Gastrinoma and Zollinger Ellison syndrome: A roadmap for the management between new and old therapies Rossi RE, Elvevi A, Citterio D, Coppa J, Invernizzi P, Mazzaferro V, Massironi S

#### **MINIREVIEWS**

5908 Optical diagnosis of colorectal polyps using convolutional neural networks Kader R, Hadjinicolaou AV, Georgiades F, Stoyanov D, Lovat LB

- 5919 Liver-spleen axis dysfunction in COVID-19 Cococcia S, Lenti MV, Santacroce G, Achilli G, Borrelli de Andreis F, Di Sabatino A
- 5932 Primary gastric non-Hodgkin lymphomas: Recent advances regarding disease pathogenesis and treatment Diamantidis MD, Papaioannou M, Hatjiharissi E

# **ORIGINAL ARTICLE**

#### **Basic Study**

5946 Proteomics identifies a novel role of fibrinogen-like protein 1 in Crohn's disease Sun XL, Qiao LC, Gong J, Wen K, Xu ZZ, Yang BL

#### **Retrospective Study**

5958 Effectiveness and safety of over-the-scope clip in closing perforations after duodenal surgery Wang ZZ, Zhou XB, Wang Y, Mao XL, Ye LP, Yan LL, Chen YH, Song YQ, Cai Y, Xu SW, Li SW



Contra	World Journal of Gastroenterology							
Conter	Weekly Volume 27 Number 35 September 21, 2021							
5967	Hepatic perivascular epithelioid cell tumor: Clinicopathological analysis of 26 cases with emphasis on disease management and prognosis							
	Zhang S, Yang PP, Huang YC, Chen HC, Chen DL, Yan WT, Yang NN, Li Y, Li N, Feng ZZ							
5978	Diagnosis of focal liver lesions with deep learning-based multi-channel analysis of hepatocyte-specific contrast-enhanced magnetic resonance imaging							
	Stollmayer R, Budai BK, Tóth A, Kalina I, Hartmann E, Szoldán P, Bérczi V, Maurovich-Horvat P, Kaposi PN							



# Contents

Weekly Volume 27 Number 35 September 21, 2021

# **ABOUT COVER**

Editorial Board Member of World Journal of Gastroenterology, Haruhiko Sugimura, PhD, MD, Vice President, Hamamatsu University School of Medicine; Professor and Chairman of the Department of Tumor Pathology, Hamamatsu University School of Medicine, 1-20-1 Handayama, Higashi-ku, Hamamatsu 431-3192, Japan. hsugimur@hama-med.ac.jp

# **AIMS AND SCOPE**

The primary aim of World Journal of Gastroenterology (WJG, World J Gastroenterol) is to provide scholars and readers from various fields of gastroenterology and hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WIG mainly publishes articles reporting research results and findings obtained in the field of gastroenterology and hepatology and covering a wide range of topics including gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, gastrointestinal oncology, and pediatric gastroenterology.

# **INDEXING/ABSTRACTING**

The WJG is now indexed in Current Contents<sup>®</sup>/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central, and Scopus. The 2021 edition of Journal Citation Report® cites the 2020 impact factor (IF) for WJG as 5.742; Journal Citation Indicator: 0.79; IF without journal self cites: 5.590; 5-year IF: 5.044; Ranking: 28 among 92 journals in gastroenterology and hepatology; and Quartile category: Q2. The WJG's CiteScore for 2020 is 6.9 and Scopus CiteScore rank 2020: Gastroenterology is 19/136.

# **RESPONSIBLE EDITORS FOR THIS ISSUE**

Production Editor: Yan-Xia Xing, Production Department Director: Xiang Li; Editorial Office Director: Ze-Mao Gong.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS						
World Journal of Gastroenterology	https://www.wjgnet.com/bpg/gerinfo/204						
ISSN	GUIDELINES FOR ETHICS DOCUMENTS						
ISSN 1007-9327 (print) ISSN 2219-2840 (online)	https://www.wjgnet.com/bpg/GerInfo/287						
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH						
October 1, 1995	https://www.wjgnet.com/bpg/gerinfo/240						
FREQUENCY	PUBLICATION ETHICS						
Weekly	https://www.wjgnet.com/bpg/GerInfo/288						
<b>EDITORS-IN-CHIEF</b>	PUBLICATION MISCONDUCT						
Andrzej S Tarnawski, Subrata Ghosh	https://www.wjgnet.com/bpg/gerinfo/208						
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE						
http://www.wjgnet.com/1007-9327/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242						
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS						
September 21, 2021	https://www.wjgnet.com/bpg/GerInfo/239						
COPYRIGHT	ONLINE SUBMISSION						
© 2021 Baishideng Publishing Group Inc	https://www.f6publishing.com						

© 2021 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



WJG

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5796-5802

DOI: 10.3748/wjg.v27.i35.5796

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

EDITORIAL

# Induced pluripotent stem cells as an innovative model to study drug induced pancreatitis

Elena Genova, Gabriele Stocco, Giuliana Decorti

ORCID number: Elena Genova 0000-0002-6692-6573; Gabriele Stocco 0000-0003-0964-5879; Giuliana Decorti 0000-0002-9714-6246.

Author contributions: Genova E, Stocco G and Decorti G designed the research study, reviewed and edited the manuscript; Genova E performed the research and wrote the manuscript; all authors have read and approved the final manuscript.

Supported by Italian Ministry of Health (IRCCS Burlo Garofolo), No. RC 7\_2014.

Conflict-of-interest statement:

Authors have nothing to disclose.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited

Elena Genova, Giuliana Decorti, Institute for Maternal and Child Health, IRCCS Burlo Garofolo, Trieste 34137, Italy

Gabriele Stocco, Department of Life Sciences, University of Trieste, Trieste 34127, Italy

Giuliana Decorti, Department of Medical, Surgical and Health Sciences, University of Trieste, Trieste 34127, Italy

Corresponding author: Giuliana Decorti, MD, Full Professor, Institute for Maternal and Child Health, IRCCS Burlo Garofolo, Via dell'Istria 65, Trieste 34137, Italy. decorti@units.it

# Abstract

Drug-induced pancreatitis is a gastrointestinal adverse effect concerning about 2% of drugs. The majority of cases are mild to moderate but severe episodes can also occur, leading to hospitalization or even death. Unfortunately, the mechanisms of this adverse reaction are still not clear, hindering its prevention, and the majority of data available of this potentially life-threatening adverse effect are limited to case reports leading to a probable underestimation of this event. In particular, in this editorial, special attention is given to thiopurine-induced pancreatitis (TIP), an idiosyncratic adverse reaction affecting around 5% of inflammatory bowel disease (IBD) patients taking thiopurines as immunosuppressants, with a higher incidence in the pediatric population. Validated biomarkers are not available to assist clinicians in the prevention of TIP, also because of the inaccessibility of the pancreatic tissue, which limits the possibility to perform dedicated cellular and molecular studies. In this regard, induced pluripotent stem cells (iPSCs) and the exocrine pancreatic differentiated counterpart could be a great tool to investigate the cellular and molecular mechanisms underlying the development of this undesirable event. This particular type of stem cells is obtained by reprogramming adult cells, including fibroblasts and leukocytes, with a set of transcription factors known as the Yamanaka's factors. Maintaining unaltered the donors' genetic heritage, iPSCs represent an innovative model to study the mechanisms of adverse drug reactions in individual patients' tissues not easily obtainable from human probands. Indeed, iPSCs can differentiate under adequate stimuli into almost any somatic lineage, opening a new world of opportunities for researchers. Several works are already available in the literature studying liver, central nervous system and cardiac cells derived from iPSCs and adverse drug effects. However, to our knowledge no studies have been performed on exocrine pancreas differentiated from iPSCs and drug-induced pancreatitis, so far. Hence, in



#### manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: Italy

#### Peer-review report's scientific quality classification

Grade A (Excellent): A Grade B (Very good): B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Received: March 17, 2021 Peer-review started: March 17, 2021 First decision: April 16, 2021 Revised: April 27, 2021 Accepted: August 30, 2021 Article in press: August 30, 2021 Published online: September 21, 2021

P-Reviewer: Maharshi S, Nayudu SK, Soliman YY S-Editor: Fan JR L-Editor: A P-Editor: Yuan YY



this editorial we focus specifically on the description of the study of the mechanisms of TIP by using IBD patient-specific iPSCs and exocrine pancreatic differentiated cells as innovative in vitro models.

Key Words: Induced pluripotent stem cells; Therapy personalization; Patient-specific cells; Drug-induced pancreatitis; Thiopurines; Inflammatory bowel disease

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core Tip:** About 5% of inflammatory bowel disease patients develop pancreatitis after thiopurine administration. The mechanism of this adverse effect is still not clear making it difficult to prevent. By differentiating induced pluripotent stem cells into their pancreatic exocrine counterpart, it is possible to set up innovative personalized in vitro models to study this adverse effect in a more effective way.

Citation: Genova E, Stocco G, Decorti G. Induced pluripotent stem cells as an innovative model to study drug induced pancreatitis. World J Gastroenterol 2021; 27(35): 5796-5802 URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5796.htm

DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5796

#### INTRODUCTION

Gastrointestinal adverse effects are common especially with orally absorbed drugs and may result in undesirable consequences leading to the reduction of treatment efficacy and, in the most serious cases, to therapy interruption with associated healthcare costs. To better study and prevent these adverse events there is the need for dedicated clinical investigation[1]. Over the past years, adverse drug reactions (ADRs) have been widely studied also for their negative effect on the development of new drugs[2,3].

Among the different ADRs, drug-induced pancreatitis has become increasingly recognized as an important cause of acute pancreatitis with a wide range of drug classes involved in its development[4]. Unfortunately, the majority of data available of this potentially life-threatening ADR are principally limited to case reports, leading to a probably underestimated incidence, reported to be around 2%[4]. Furthermore, the mechanisms of drug-induced pancreatitis of many drugs are still not clear, making it difficult to determine a definitive association of causality between specific medications and acute pancreatitis, and in only less than 10% of cases the real cause has been determined. Drugs known to induce pancreatitis have been classified considering the number of case reports, the recurrence of pancreatitis with a re-challenge with the drug, consistent latency between the drug assumption and the onset of acute pancreatitis and the exclusion of alternative causes such as alcohol assumption or gallstones [4,5] (Table 1).

Interestingly, certain types of ADRs are reported to be more frequent in patients affected by specific diseases. An important example is thiopurine-induced pancreatitis (TIP), an idiosyncratic ADR affecting more frequently inflammatory bowel disease (IBD) patients taking thiopurines, such as azathioprine and mercaptopurine[6]. In the vast majority of cases, TIP is manageable, however patients have to stop the treatment and to be sometimes hospitalized until the symptoms are resolved<sup>[7]</sup>. The higher incidence of this adverse event in IBD patients, especially in the pediatric population, suggests that molecular mechanisms involved in the disease may contribute to TIP predisposition[6]. However, mechanisms determining TIP predisposition are still unknown and only hypotheses have been postulated. In particular, the mechanisms proposed can be divided into three different groups: genetic predisposition[8,9], alteration in thiopurine biotransformation[7] and abnormalities in innate or adaptative immunity<sup>[10]</sup>.

The thiopurines azathioprine, mercaptopurine and thioguanine undergo an extensive biotransformation catalyzed by several enzymes[11]. Regarding genetic predisposition, TIP seems unrelated to candidate variants on important genes of the thiopurine biotransformation pathway, such as TPMT, ITPA and NUDT15, well-known to induce severe ADRs, including myelosuppression and hematologic toxicity[12,13].



Table 1 Classification system of drugs related to pancreatitis development[4,5]						
Class						
Class Ia	At least one case report with positive rechallenge, excluding other possible causes such as alcohol, gallstones and other drugs					
Class Ib	At least one case report with positive rechallenge but not excluding other possible causes					
Class II	At least four cases in the literature without rechallenge but with consistent latency in greater than 75% of cases					
Class III	At least two cases in the literature without rechallenge and consistent latency					
Class IV	Single case reported in the literature not fitting the previous described classed without rechallenge					

Recently, two different research groups have found a strong association between the Class II HLA gene region polymorphism rs2647087 and TIP[8,9], but more efforts are needed to translate these variants into clinical practice. TIP development may be also related to direct damage to the exocrine pancreatic cells or to an accumulation of toxic metabolites (biotransformation hypothesis). However, pancreatitis frequently occurs early after thiopurine administration, making the accumulation of toxic metabolites unlikely, while more probably immunological reactions are involved. However, direct toxicity of thiopurines or their metabolites on patients' pancreatic cells cannot be completely excluded[7,10].

To study and discover TIP mechanisms and predisposition, innovative patientspecific in vitro models could be helpful and decisive. In this regard, induced pluripotent stem cells (iPSCs) and their differentiated counterpart are widely used to set up groundbreaking personalized *in vitro* models representative of patients' genetic background. The peculiar characteristics of these cells allow to set up in vitro models to study disease mechanisms and ADRs with the purpose to personalize patients' therapy, improving the disease outcome. The iPSC model can be a great tool to better understand, and thus prevent, ADRs in particular in comparison to animal models and immortalized cells. Indeed, the predisposition to a specific ADR may be related to the individual genetic patients' background, leading to a wide range of toxicities of different severity[14]. Therefore, the iPSC technology, matching the donor's genetic background, can be extremely helpful for developing patient-specific assays. Indeed, by using iPSCs, it seems reasonable to precisely mimic the patients' susceptibility to an abnormal response to a specific drug, setting up powerful assays useful to identify predictive biomarkers. In the last years, many different models[15] have been developed using the iPSC technology, including the differentiation into pancreatic exocrine cells[16].

# PATIENT-SPECIFIC IPSCS AS AN IN VITRO MODEL TO STUDY DRUG-INDUCED PANCREATITIS

Patient-specific iPSCs can be obtained by reprogramming patients' fibroblasts or peripheral blood mononuclear cells using the four Yamanaka's factors OCT4, SOX2, KLF4 and MYC, forcing somatic cells to an embryonic-like state [17,18]. Differentiation of iPSCs allows to generate almost any kind of somatic cells using appropriate protocols. In the literature it is possible to find a wide range of differentiation possibilities including neural-like cells, hepatocytes, enterocytes, pancreatic endocrine cells and many others as recently reviewed by our group[15]. These cells, being patientspecific, have been frequently used to model and study individual susceptibility to develop ADRs. For example, regarding gastrointestinal toxicity, some groups have already tried to model hepatocytes[19-21] and enterocytes[22,23] to study drug-induced liver injury and intestinal toxicity, respectively. However, in comparison to other ADRs, drug-induced pancreatitis has not been deeply studied yet. A limited number of protocols[16,24-26] are available in the literature to generate pancreatic exocrine cells starting from iPSCs in comparison to the endocrine counterpart[15]. To the best of our knowledge, our group recently evaluated for the first time the mechanisms behind TIP predisposition using iPSCs and pancreatic differentiated cells of pediatric patients affected by IBD that developed or not TIP. Differentiation of iPSCs in pancreatic exocrine cells was performed using the protocol developed by Takizawa-Shirasawa et al[16]. Briefly, different stimuli were added to the culture medium in 4 different steps (Figure 1). To characterize cells obtained during each differentiation step, genetic expression of specific genetic markers was analyzed and confirmed:



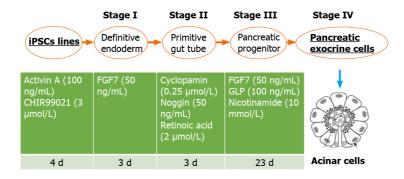


Figure 1 Differentiation of induced pluripotent stem cells into pancreatic cells towards a 4 steps protocol. iPSCs: Induced pluripotent stem cells; d: Days of culture.

*OCT4* for undifferentiated cells (iPSCs), *FOXA2* and *SOX17* for definitive endoderm (stage I), *PDX1* for pancreatic progenitors (stage III) and amylase, in particular its pancreatic isoforms *AMY2A* and *AMY2B* for pancreatic exocrine cells (stage IV).

The gold standard of cytotoxicity assay showed an almost double *in vitro* sensitivity of TIP cases cells to thiopurines, more marked in iPSCs rather than in the differentiated counterpart, after mercaptopurine and thioguanine exposure. *TPMT* variants (rs1142345, rs1800460 and rs1800462) were excluded as a possible cause of this different sensitivity because all patients resulted wild-type.

The results obtained are encouraging, however some limitations have to be overcome in the next future. For instance, the differentiation protocol to obtain exocrine pancreatic cells could be further improved in terms of efficiency based on the more recent studies performed by Hohwieler et al[24] and Ito et al[25] which used 3D culture methods and the distinction between acinar and ductal cell type, by analyzing the expression of different genetic and protein markers such as amylase and chymotrypsin C for acinar cells, and SOX9 and cytokeratin 19 for ductal cells[24,25]. An important point to consider is if the amylase markers are sufficient to reflect terminal differentiation. Beside studies considering the mRNA levels of these markers[24,25], more functional studies, evaluating the amylase protein concentrations and enzyme activity, should be implemented. These comparisons would allow to ensure that terminal differentiation is as representative as possible of the in vivo models. Another important point to focus, linked to pancreatic cell generation, is the time necessary that is too long for a clinical application of this in vitro model for TIP predisposition screening. Studies are now ongoing to partially resolve this limitation trying to develop more efficient and faster ready-to-use patient-specific pancreatic exocrine differentiated cells. The cost of hospitalization after a pancreatitis event has been recently calculated, resulting in around 8000 € per patient[27]. Considering an incidence of pancreatitis of 5%, we can estimate that every 20 patients treated with azathioprine one will be at risk of pancreatitis. Therefore, to be cost-effective, the analysis should amount to 400  $\epsilon$ , considering only the cost of the analysis, without evaluating the health benefit<sup>[28]</sup>. Current costs are still higher but there is a trend toward reduction; indeed, the iPSC technology is still expensive and costs have to be reduced before they can be introduced into clinical practice. In particular, characterization costs are high, but several suggestions to address this limitation have been already proposed such as SNP microarray technology for the routine karyotyping and cost-effective methods such as innovative flow cytometry analyses to assess cell surface expression of pluripotent markers<sup>[29]</sup>.

Beyond technical limitations, it is conceivable that thiopurines do not directly reach the pancreatic tissue unmodified, but rather as metabolites. Therefore, to improve the clinical relevance of the *in vitro* model, patient-specific pancreatic cells would need to be exposed to a representative mixture of thiopurine metabolites or to conditioned media of other thiopurine metabolizing cells such as hepatocytes[30]. Moreover, it is important to keep in mind that TIP predisposition could be influenced by the contribution of the immune system that, in predisposed patients, could be activated for unknown reasons after thiopurine administration attacking the pancreatic tissue. This aspect has to be considered, modeled and studied as well[7,31]. Finally, data obtained have to be confirmed in a larger cohort of patients that now includes 3 cases and 3 controls already analyzed while 2 cases and 2 controls still have to be analyzed.

# **CLINICAL IMPLICATIONS**

Drug-induced pancreatitis represents an important clinical issue for different reasons including therapy interruption, reduction of treatment efficacy, the need for unnecessary diagnostic procedures and treatment for the adverse effect resolution[1] with associated healthcare costs. Moreover, in recent years an increasing number of drugs have been associated with pancreatitis development although its recognition by clinicians is still limited because of the lack of biomarkers useful to prevent this ADR.

# CONCLUSION

Drug-induced pancreatitis is a growing problem related to several drugs and TIP recapitulates well all complications related to the development of this ADR. The possibility of studying TIP by an iPSC-based model seems a great opportunity to investigate TIP mechanisms that still remain not clear. The in vitro model established in our laboratory has proven to be suitable for studying and investigating TIP predisposition in a personalized way in pediatric IBD patients. Alongside thiopurines, several other drugs such as asparaginase, nilotinib and pazopanib can cause pancreatitis. Therefore, the *in vitro* model developed in this study could be applied also to study the sensitivity of other drugs with the purpose of pancreatitis prevention.

# REFERENCES

- Philpott HL, Nandurkar S, Lubel J, Gibson PR. Drug-induced gastrointestinal disorders. Frontline 1 Gastroenterol 2014; 5: 49-57 [PMID: 28839751 DOI: 10.1136/flgastro-2013-100316]
- Guengerich FP. Mechanisms of drug toxicity and relevance to pharmaceutical development. Drug Metab Pharmacokinet 2011; 26: 3-14 [PMID: 20978361 DOI: 10.2133/dmpk.dmpk-10-rv-062]
- 3 Timilsina M, Tandan M, d'Aquin M, Yang H. Discovering Links Between Side Effects and Drugs Using a Diffusion Based Method. Sci Rep 2019; 9: 10436 [PMID: 31320740 DOI: 10.1038/s41598-019-46939-6
- 4 Weissman S, Aziz M, Perumpail RB, Mehta TI, Patel R, Tabibian JH. Ever-increasing diversity of drug-induced pancreatitis. World J Gastroenterol 2020; 26: 2902-2915 [PMID: 32587438 DOI: 10.3748/wjg.v26.i22.2902]
- 5 Badalov N, Baradarian R, Iswara K, Li J, Steinberg W, Tenner S. Drug-induced acute pancreatitis: an evidence-based review. Clin Gastroenterol Hepatol 2007; 5: 648-61; quiz 644 [PMID: 17395548 DOI: 10.1016/j.cgh.2006.11.023]
- Ramos LR, Sachar DB, DiMaio CJ, Colombel JF, Torres J. Inflammatory Bowel Disease and 6 Pancreatitis: A Review. J Crohns Colitis 2016; 10: 95-104 [PMID: 26351384 DOI: 10.1093/ecco-jcc/jjv153]
- Stocco G, Lanzi G, Yue F, Giliani S, Sasaki K, Tommasini A, Pelin M, Martelossi S, Ventura A, 7 Decorti G. Patients' Induced Pluripotent Stem Cells to Model Drug Induced Adverse Events: A Role in Predicting Thiopurine Induced Pancreatitis? Curr Drug Metab 2015; 17: 91-98 [PMID: 26526832 DOI: 10.2174/1389200216666151103120220]
- Heap GA, Weedon MN, Bewshea CM, Singh A, Chen M, Satchwell JB, Vivian JP, So K, Dubois PC, Andrews JM, Annese V, Bampton P, Barnardo M, Bell S, Cole A, Connor SJ, Creed T, Cummings FR, D'Amato M, Daneshmend TK, Fedorak RN, Florin TH, Gaya DR, Greig E, Halfvarson J, Hart A, Irving PM, Jones G, Karban A, Lawrance IC, Lee JC, Lees C, Lev-Tzion R, Lindsay JO, Mansfield J, Mawdsley J, Mazhar Z, Parkes M, Parnell K, Orchard TR, Radford-Smith G, Russell RK, Reffitt D, Satsangi J, Silverberg MS, Sturniolo GC, Tremelling M, Tsianos EV, van Heel DA, Walsh A, Watermeyer G, Weersma RK, Zeissig S, Rossjohn J, Holden AL; International Serious Adverse Events Consortium; IBD Pharmacogenetics Study Group, Ahmad T. HLA-DQA1-HLA-DRB1 variants confer susceptibility to pancreatitis induced by thiopurine immunosuppressants. Nat Genet 2014; 46: 1131-1134 [PMID: 25217962 DOI: 10.1038/ng.3093]
- Wilson A, Jansen LE, Rose RV, Gregor JC, Ponich T, Chande N, Khanna R, Yan B, Jairath V, Khanna N, Sey M, Beaton M, McIntosh K, Teft WA, Kim RB. HLA-DQA1-HLA-DRB1 polymorphism is a major predictor of azathioprine-induced pancreatitis in patients with inflammatory bowel disease. Aliment Pharmacol Ther 2018; 47: 615-620 [PMID: 29270995 DOI: 10.1111/apt.14483]
- 10 Weersma RK, Batstra MR, Kleibeuker JH, van Dullemen HM. Are pancreatic autoantibodies associated with azathioprine-induced pancreatitis in Crohn's disease? JOP 2008; 9: 283-289 [PMID: 18469440]
- 11 Zaza G, Cheok M, Krynetskaia N, Thorn C, Stocco G, Hebert JM, McLeod H, Weinshilboum RM, Relling MV, Evans WE, Klein TE, Altman RB. Thiopurine pathway. Pharmacogenet Genomics 2010; 20: 573-574 [PMID: 19952870 DOI: 10.1097/FPC.0b013e328334338f]
- 12 Zabala-Fernández W, Barreiro-de Acosta M, Echarri A, Carpio D, Lorenzo A, Castro J, Martínez-



Ares D, Pereira S, Martin-Granizo I, Corton M, Carracedo A, Barros F. A pharmacogenetics study of TPMT and ITPA genes detects a relationship with side effects and clinical response in patients with inflammatory bowel disease receiving Azathioprine. J Gastrointestin Liver Dis 2011; 20: 247-253 [PMID: 21961091]

- 13 Moriyama T, Nishii R, Perez-Andreu V, Yang W, Klussmann FA, Zhao X, Lin TN, Hoshitsuki K, Nersting J, Kihira K, Hofmann U, Komada Y, Kato M, McCorkle R, Li L, Koh K, Najera CR, Kham SK, Isobe T, Chen Z, Chiew EK, Bhojwani D, Jeffries C, Lu Y, Schwab M, Inaba H, Pui CH, Relling MV, Manabe A, Hori H, Schmiegelow K, Yeoh AE, Evans WE, Yang JJ. NUDT15 polymorphisms alter thiopurine metabolism and hematopoietic toxicity. Nat Genet 2016; 48: 367-373 [PMID: 26878724 DOI: 10.1038/ng.3508]
- 14 Wilke RA, Lin DW, Roden DM, Watkins PB, Flockhart D, Zineh I, Giacomini KM, Krauss RM. Identifying genetic risk factors for serious adverse drug reactions: current progress and challenges. Nat Rev Drug Discov 2007; 6: 904-916 [PMID: 17971785 DOI: 10.1038/nrd2423]
- Genova E, Cavion F, Lucafò M, Leo L, Pelin M, Stocco G, Decorti G. Induced pluripotent stem cells 15 for therapy personalization in pediatric patients: Focus on drug-induced adverse events. World J Stem Cells 2019; 11: 1020-1044 [PMID: 31875867 DOI: 10.4252/wjsc.v11.i12.1020]
- Takizawa-Shirasawa S, Yoshie S, Yue F, Mogi A, Yokoyama T, Tomotsune D, Sasaki K. FGF7 and 16 cell density are required for final differentiation of pancreatic amylase-positive cells from human ES cells. Cell Tissue Res 2013; 354: 751-759 [PMID: 23996199 DOI: 10.1007/s00441-013-1695-6]
- Fusaki N, Ban H, Nishiyama A, Saeki K, Hasegawa M. Efficient induction of transgene-free human 17 pluripotent stem cells using a vector based on Sendai virus, an RNA virus that does not integrate into the host genome. Proc Jpn Acad Ser B Phys Biol Sci 2009; 85: 348-362 [PMID: 19838014 DOI: 10.2183/piab.85.348]
- 18 Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell 2006; 126: 663-676 [PMID: 16904174 DOI: 10.1016/j.cell.2006.07.024]
- Kondo Y, Iwao T, Nakamura K, Sasaki T, Takahashi S, Kamada N, Matsubara T, Gonzalez FJ, 19 Akutsu H, Miyagawa Y, Okita H, Kiyokawa N, Toyoda M, Umezawa A, Nagata K, Matsunaga T, Ohmori S. An efficient method for differentiation of human induced pluripotent stem cells into hepatocyte-like cells retaining drug metabolizing activity. Drug Metab Pharmacokinet 2014; 29: 237-243 [PMID: 24334537 DOI: 10.2133/dmpk.dmpk-13-rg-104]
- 20 Kang SJ, Lee HM, Park YI, Yi H, Lee H, So B, Song JY, Kang HG. Chemically induced hepatotoxicity in human stem cell-induced hepatocytes compared with primary hepatocytes and HepG2. Cell Biol Toxicol 2016; 32: 403-417 [PMID: 27287938 DOI: 10.1007/s10565-016-9342-0]
- Liu J, Brzeszczynska J, Samuel K, Black J, Palakkan A, Anderson RA, Gallagher R, Ross JA. 21 Efficient episomal reprogramming of blood mononuclear cells and differentiation to hepatocytes with functional drug metabolism. Exp Cell Res 2015; 338: 203-213 [PMID: 26256888 DOI: 10.1016/j.yexcr.2015.08.004]
- Kondo S, Mizuno S, Hashita T, Iwao T, Matsunaga T. Using human iPS cell-derived enterocytes as 22 novel in vitro model for the evaluation of human intestinal mucosal damage. Inflamm Res 2018; 67: 975-984 [PMID: 30317465 DOI: 10.1007/s00011-018-1193-0]
- 23 Ozawa T, Takayama K, Okamoto R, Negoro R, Sakurai F, Tachibana M, Kawabata K, Mizuguchi H. Generation of enterocyte-like cells from human induced pluripotent stem cells for drug absorption and metabolism studies in human small intestine. Sci Rep 2015; 5: 16479 [PMID: 26559489 DOI: 10.1038/srep16479
- 24 Hohwieler M, Illing A, Hermann PC, Mayer T, Stockmann M, Perkhofer L, Eiseler T, Antony JS, Müller M, Renz S, Kuo CC, Lin Q, Sendler M, Breunig M, Kleiderman SM, Lechel A, Zenker M, Leichsenring M, Rosendahl J, Zenke M, Sainz B Jr, Mayerle J, Costa IG, Seufferlein T, Kormann M, Wagner M, Liebau S, Kleger A. Human pluripotent stem cell-derived acinar/ductal organoids generate human pancreas upon orthotopic transplantation and allow disease modelling. Gut 2017; 66: 473-486 [PMID: 27633923 DOI: 10.1136/gutjnl-2016-312423]
- 25 Ito K, Matsuura K, Mihara Y, Sakamoto Y, Hasegawa K, Kokudo N, Shimizu T. Delivery of pancreatic digestive enzymes into the gastrointestinal tract by pancreatic exocrine tissue transplant. Sci Rep 2019; 9: 5922 [PMID: 30976035 DOI: 10.1038/s41598-019-42362-z]
- 26 Huang L, Holtzinger A, Jagan I, BeGora M, Lohse I, Ngai N, Nostro C, Wang R, Muthuswamy LB, Crawford HC, Arrowsmith C, Kalloger SE, Renouf DJ, Connor AA, Cleary S, Schaeffer DF, Roehrl M, Tsao MS, Gallinger S, Keller G, Muthuswamy SK. Ductal pancreatic cancer modeling and drug screening using human pluripotent stem cell- and patient-derived tumor organoids. Nat Med 2015; 21: 1364-1371 [PMID: 26501191 DOI: 10.1038/nm.3973]
- Wilson A, Wang Q, Choi YH, Ponich T, Gregor JC, Chande N, Yan B, Sey M, Beaton M, Kim RB. 27 Pretreatment HLADQA1-HLADRB1 Testing for the Prevention of Azathioprine-Induced Pancreatitis in Inflammatory Bowel Disease: A Prospective Cohort Study. Clin Transl Gastroenterol 2021; 12: e00332 [PMID: 33821842 DOI: 10.14309/ctg.00000000000332]
- Owens DK. Interpretation of cost-effectiveness analyses. J Gen Intern Med 1998; 13: 716-717 28 [PMID: 9798822 DOI: 10.1046/j.1525-1497.1998.00211.x]
- 29 Malihi G, Nikoui V, Elson EL. A review on qualifications and cost effectiveness of induced pluripotent stem cells (IPSCs)-induced cardiomyocytes in drug screening tests. Arch Physiol Biochem 2020; 1-12 [PMID: 32783745 DOI: 10.1080/13813455.2020.1802600]
- Vikingsson S, Carlsson B, Almer SH, Peterson C. Monitoring of thiopurine metabolites in patients 30



with inflammatory bowel disease-what is actually measured? Ther Drug Monit 2009; 31: 345-350 [PMID: 19363461 DOI: 10.1097/FTD.0b013e3181a1ea58]

31 Hung WY, Abreu Lanfranco O. Contemporary review of drug-induced pancreatitis: A different perspective. World J Gastrointest Pathophysiol 2014; 5: 405-415 [PMID: 25400984 DOI: 10.4291/wjgp.v5.i4.405]



WJG

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5803-5821

DOI: 10.3748/wjg.v27.i35.5803

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

OPINION REVIEW

# Screening for nonalcoholic fatty liver disease-when, who and how?

Christoph G Dietrich, Monika Rau, Andreas Geier

ORCID number: Christoph G Dietrich 0000-0001-6927-7970; Monika Rau 0000-0003-1219-4044; Andreas Geier 0000-0002-9626-5083.

Author contributions: Dietrich CG and Geier A developed the concept of this review; all authors contributed to the manuscript.

Conflict-of-interest statement: CGD reports personal speaker fees from Falk Foundation. MR declares no conflict of interest. AG serves as advisor and steering committee member for AbbVie, Alexion, Bayer, BMS, CSL Behring, Eisai, Gilead, Intercept, Ipsen, Merz, MSD, Novartis, Pfizer, Roche, Sanofi-Aventis, Sequana and as speaker for AbbVie, Alexion, BMS, CSL Behring, Falk, Gilead, Intercept, Merz, MSD, Novartis, Roche, Sequana. AG also received research support from Intercept and Falk (NAFLD CSG), Novartis.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and Christoph G Dietrich, Department of Internal Medicine, Bethlehem Health Center, Stolberg 52222, Germany

Monika Rau, Department of Internal Medicine II, University Hospital Würzburg, Würzburg 97080, Germany

Andreas Geier, Department of Medicine II, University Hospital Würzburg, Würzburg 97080, Germany

Corresponding author: Christoph G Dietrich, MD, PhD, Chief Doctor, Department of Internal Medicine, Bethlehem Health Center, Steinfeldstr. 5, Stolberg 52222, Germany. christoph.g.dietrich@googlemail.com

# Abstract

Nonalcoholic fatty liver disease (NAFLD) is becoming a frequent liver disease, especially in patients with metabolic syndrome and especially in Western countries. Complications of NAFLD comprise progressive fibrosis, cirrhosis and hepatocellular carcinoma. NAFLD also represents an independent risk factor for cardiovascular disease, extrahepatic neoplasia and other organ damage, such as renal insufficiency. Given the epidemiological importance of the disease, new developments in specific treatment of the disease and the wide availability of noninvasive techniques in estimating steatosis and fibrosis, NAFLD should be subject to screening programs, at least in countries with a high prevalence of the disease. The review discusses prerequisites for screening, cost-effectiveness, current guideline recommendations, suitability of techniques for screening and propositions for the following questions: Who should be screened? Who should perform screening? How should screening be performed? It is time for a screening program in patients at risk for NAFLD.

Key Words: Screening; Nonalcoholic fatty liver disease; Diabetes; Liver fibrosis; Cirrhosis

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core Tip:** Nonalcoholic fatty liver disease (NAFLD) is becoming more important in Western countries and leads to serious complications in patients with progressive disease. The epidemiological, clinical and technical requirements for screening for this disease are fulfilled and are outlaid in this review. It is time to consider a screening program for NAFLD.



the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: Germany

#### Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Received: March 16, 2021 Peer-review started: March 16, 2021 First decision: May 1, 2021 Revised: May 13, 2021 Accepted: August 30, 2021 Article in press: August 30, 2021 Published online: September 21, 2021

P-Reviewer: Maevskaya M S-Editor: Fan JR L-Editor: Y P-Editor: Yuan YY



Citation: Dietrich CG, Rau M, Geier A. Screening for nonalcoholic fatty liver disease-when, who and how? World J Gastroenterol 2021; 27(35): 5803-5821 URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5803.htm **DOI:** https://dx.doi.org/10.3748/wjg.v27.i35.5803

# INTRODUCTION

Nonalcoholic fatty liver disease (NAFLD) is the most common liver disease worldwide, with rising prevalence to an estimate of 25% in Western populations[1]. NAFLD is regarded as one component of metabolic syndrome, including obesity, insulin resistance or type 2 diabetes mellitus (T2DM), hypertension, and dyslipidemia. Recently, the new term metabolic dysfunction-associated fatty liver disease has been proposed to emphasize this association[2]. Over the next decade, the number of patients with advanced fibrosis stages is expected to rise further together with an increasing incidence of complications [nonalcoholic steato hepatitis (NASH)-related end stage liver disease, e.g. hepatic decompensation, liver cancer and mortality [3]. In this recent modeling, the number of NAFLD patients in the United States, the EU5 (France, Germany, Italy, Spain, United Kingdom) and China was estimated to be 85.3 million, 72.2 million and 211 million, respectively, whereby in the same countries, more than 17.3 million, 12.6 million and 32.6 million patients were predicted to have NASH[3]. The number of NASH patients with advanced fibrosis is expected to more than double until 2030. Similar but slightly more conservative calculations have been obtained with different modeling methodologies but confirm the extent of the clinical problem[4]. In addition to liver-related morbidity and mortality, it is important to emphasize that NAFLD patients have increased cardiovascular mortality, which together cause an enormous socioeconomic impact in industrialized countries<sup>[4]</sup>. The fact that NAFLD has become the most frequent disease entity on the liver transplant waiting list in the UNOS network documents the need for early detection and intervention in the future<sup>[5]</sup>. Given the sheer frequency of patients with obesity, metabolic syndrome and NAFLD worldwide, it is remarkable that this disease entity has been overlooked by clinicians and the pharmaceutical industry for a considerable period of time, and no widely established algorithms for screening exist. The global burden of disease documents the burning need to establish clinical care structures and diagnostic algorithms to cope with the increasing number of patients at risk.

A multistep diagnostic screening algorithm is recommended in current guidelines in Western countries and combines an initial ultrasound (US) examination with subsequent risk prediction tools such as the Fibrosis-4 (FIB-4) or NAFLD fibrosis score (NFS) followed by transient elastography (TE) stratification for liver biopsy[6,7]. Increasing public and professional awareness as well as the implementation of screening algorithms in primary and secondary care will lead to a more frequent diagnosis of NAFLD patients at different stages of the disease (NAFL, noncirrhotic NASH, NASH with cirrhosis) in the near future. For the histological assessment of NAFLD, different systems are used for scoring in clinical practice [e.g., NAFLD activity score (NAS)][8]. The definite histopathological diagnosis of NAFL vs NASH is based on the simultaneous presence of steatosis, ballooning and inflammation, which are required for the diagnosis of "NASH" in the European SAF/FLIP algorithm[9].

Of the different histologic features of NASH, fibrosis has been identified as the strongest predictor of adverse clinical outcomes, including decompensation and liverrelated death[10-14]. The latest meta-analysis showed that the stage of biopsy-confirmed liver fibrosis is a strong predictor of future all-cause mortality and morbidity in NAFLD with and without adjustment for key potential confounding variables[15]. It became clear that evaluation of the fibrosis stage is even more fundamental than scoring necroinflammation or diagnosing NASH. Several options for the noninvasive evaluation of liver fibrosis in NASH, such as elastography devices and blood tests, are available[16]. Despite recent progress in noninvasive tests (NITs) for the evaluation of liver fibrosis in NAFLD, the diagnosis of NASH is still often based on liver biopsy, an invasive procedure not suitable for the large proportion of the general population affected by NAFLD. To identify patients with an increased risk, the NFS was introduced in 2007 as a simple scoring system to distinguish NAFLD with and without advanced fibrosis (fibrosis stages 3 and 4)[17]. Subsequently, further fibrosis tests, including the FIB-4 index, Fibrotest/Fibrosure, enhanced liver fibrosis (ELF) test, and liver stiffness measurement (LSM) by vibration-controlled TE, have entered clinical practice[18-21]. Of relevance for fibrosis screening, these NITs show excellent



AUROCs for the diagnosis of advanced fibrosis and cirrhosis[22]. Furthermore, repeated testing of FIB-4 within 5 years improved the identification of individuals at an increased risk of severe liver disease in the general population[23]. In light of a multistep screening algorithm, the performance has been further improved by the sequential combination of different NITs for advanced fibrosis, thereby refining the patient referral pathway between primary care or diabetologists and liver specialists [24]. Sequential combinations of FIB-4 (or NFS) and TE with a lower cut-off to rule-out advanced fibrosis and a higher cut-off to rule-in cirrhosis can increase the specificity and thereby reduce the need for liver biopsies from 33% to 19%[25]. The ultimate goal of screening measures is to identify patients at high risk for liver-related events and unfavorable overall outcomes. Longitudinal retrospective studies have demonstrated that NITs calibrated on liver fibrosis are prognostic markers to stratify the risk of liver-related outcomes and mortality in NAFLD patients[26].

Comparative diagnostic accuracy studies for established and novel biomarkers and combinations thereof are ongoing in the European LITMUS and United States NIBLE consortia[27]. It will be interesting to learn whether and which of the novel biomarkers outperforms the established freely available routine scores NFS and FIB-4. At the same time, biomarker screening strategies are currently being tested to establish validated numbers of patients to test to identify NASH patients with advanced fibrosis suitable for specific treatment.

The following review gives an overview of current guideline recommendations and answers the question of when, whom and how to screen in the different clinical settings.

#### **RECOMMENDATIONS FOR NAFLD SCREENING IN RECENT GUIDELINES**

Several guidelines worldwide have already taken a position on screening for NAFLD. The consensus is that screening in the general population is not recommended[6,7,28, 29]. AASLD also discourages screening in high-risk groups because of the current lack of treatment options, unclear value of screening tests, and unclear cost-effectiveness. However, "a high index of suspicion" for the presence of NAFLD in diabetes mellitus type 2 patients is advised[7]. The Asian guideline takes a similarly noncommittal view, which also does not explicitly recommend screening in risk groups (here T2DM and obesity) but merely describes it as worth considering[29].

In contrast, specific screening recommendations can be found in the Latin American and European guidelines. Here, NAFLD screening is recommended for patients with repeatedly altered liver enzymes, features of metabolic syndrome, or obesity [body mass index (BMI) > 30] according to Latin American guidelines[28]. In the same direction, patients with insulin resistance and metabolic syndrome, especially manifest type 2 diabetes, should also be screened for the presence of NAFLD according to the European recommendation, regardless of the level of liver enzymes[6]. Both guidelines primarily recommend abdominal US as the initial examination to determine the presence of steatosis. Serum fibrosis tests are considered appropriate for further risk stratification[6,28], with the Latin American guideline decidedly recommending determination of FIB-4 and NFS. Elastography, as a more reliable method, is also mentioned[28] but is considered secondary due to its lack of availability in many places.

The guidelines differ in their treatment of patients in whom serum fibrosis scores indicate intermediate fibrosis risk. While the European algorithm recommends both high-risk and intermediate-risk patients for referral to the hepatologist[6], the Latin American guidelines suggest that this should only be the case for patients > 50 years of age with diabetes or obesity[28].

The basis of the differing recommendations is an ultimate lack of data on the efficacy and efficiency of structured screening and on the effectiveness of the therapeutic efforts that begin after NAFLD has been diagnosed in the context of screening. There are also discrepancies between the lack of widespread availability of specific examination procedures and the desire for screening results that are as sensitive and specific as possible and avoid overloading specialists by referring numerous false-positive screened patients.

### SCREENING-WHEN? IS IT TIME FOR A NAFLD SCREENING PROGRAM?

#### Prerequisites for a disease to justify screening

In 1968, Wilson and Jungner formulated basic criteria for the usefulness of screening procedures for a particular disease in a paper by the WHO[30,31]. These criteria include peculiarities of the disease (significant burden of disease in the population and knowledge of etiology and stages of disease) and of reaching a diagnosis (simple test acceptable to patients) as well as organizational requirements (available facilities for diagnosis and therapy). In general, these criteria already apply for NAFLD for some time.

However, the authors also point out that efficient therapy as well as cost-effectiveness of screening must be present[30]. Here, important new developments have occurred in recent years that make screening for NAFLD much more justified than in the past.

The general progress in diagnosing and treating liver disease led an expert group 2016 to the proposal that screening for liver fibrosis (independent from the underlying disease) may now be feasible even for the general population[32].

For a long time, missing therapeutic options were a major argument against NAFLD screening, since lifestyle changes could only be maintained in a minority of patients and NASH-specific drugs were not even developed. In the meantime, several new drugs acting on various pathophysiological processes in NASH have entered clinical development. Current drug classes being investigated for NASH treatment are agonists of nuclear receptors such as FXR agonists (including FGF19), peroxisome proliferator-activated receptors agonists, chemokine receptor inhibitors, thyroid hormone receptor- $\beta$  agonists and analogs of enterohepatic hormones such as GLP-1 and FGF21 or SGLT2 inhibitors[33]. Despite disappointment by negative interim results from three out of four recent phase 3 trials, the process of approval is ongoing for obeticholic acid as the only drug with a significant benefit in the phase 3 interim analysis. Obeticholic acid is an obvious candidate for the first conditional approval as a NASH therapeutic in the near future. However, even before approval of new drugs, NAFLD patients "at risk" should be offered to participate in ongoing clinical trials, particularly those with drug combinations, since the future will putatively be a more efficient combination therapy of two different drug classes with complementary effects[33].

#### Cost-effectiveness of NAFLD screening

Decisions on the target population for screening are mostly driven by cost-effectiveness and depend on the prevalence of the disease in the target population and health outcomes measured as quality-adjusted life-years (QALYs). Unfortunately, the cost-effectiveness of noninvasive liver tests in NAFLD is scarcely available in the literature.

However, the cost-effectiveness of noninvasive screening for alcohol-related liver fibrosis has been investigated in more detail[34]. For low prevalence populations, a screening strategy involving a blood-based noninvasive fibrosis test (ELF) in the first-line follow-up with LSM in intermediate- or high-risk individuals in the second-line follow-up was most cost-effective, both short- and long-term, depending on whether diagnostic testing had lasting or temporary effects on abstinence rates. The study documents that the effect of screening measures strongly depends on the therapeutic options and the size of the treatment effect. Moreover, for high-prevalence populations, direct referral to LSM was highly cost-effective.

In contrast to the growing burden of disease, a cross-sectional study of the public health response to NAFLD among experts in 29 European countries in 2018 and 2019 revealed a general lack of national policies, awareness campaigns and civil society involvement and only a few epidemiological registries[35]. Only one-third of the countries reported having national recommendations for NAFLD screening in all patients with diabetes, obesity and/or metabolic syndrome.

Data on cost-effectiveness need to be interpreted in the context of the national health system, economy and availability of treatment. Nevertheless, available data for certain diagnostic measures allow at least some general insight and can be used as part of evidence-informed decision making. As the most basic diagnostic method, ultrasono-graphy screening for NAFLD has been found to be cost-effective in Thailand for patients with metabolic syndrome participating in an intensive weight reduction program when compared with no screening[36]. Differences in the age of the target population have been observed, since screening before 45 years was cost saving, while screening at 45 to 64 years was cost-effective.

The cost-effectiveness of LSM by TE has only been assessed in comparison to liver biopsy as the invasive reference method. In a systematic analysis covering four costeffectiveness and four cost-utility studies[37], high-quality cost-effectiveness studies suggested that TE is less costly but also less accurate than liver biopsy (which is not surprising since histology is still regarded as the diagnostic gold standard). The incremental cost-effectiveness ratio (ICER) of TE improves with a greater level of diagnostic accuracy and a higher degree of liver fibrosis. Similar data have been obtained in a Canadian systematic review of existing TE cost-effectiveness studies from the perspective of the Ontario Ministry of Health and Long-Term Care[38]. For a primary economic evaluation, decision analytic models were used to compare shortterm costs and outcomes of TE compared to liver biopsy. Again, data suggested that TE leads to cost savings but is less effective than liver biopsy in the diagnosis of liver fibrosis. Of note, TE became more economically attractive in a high-risk population with a higher degree of liver fibrosis. No studies have assessed the cost-effectiveness of TE with controlled attenuation parameter (CAP)-based fat quantification for the diagnosis of liver steatosis.

It remains open whether NAFLD screening can become cost-effective in the near future with a further increasing number of at-risk NAFLD patients in Western countries. Investigators from six prospective cohorts in Europe and Asia used patients with mostly alcohol-related liver disease to explore the cost-effectiveness of TE as a screening method to detect liver fibrosis against standard of care in a primary care pathway[39]. In 6295 participants, TE with the proposed cutoffs for the diagnosis of significant fibrosis (≥ F2) of 9.1 kPa in general population settings and 9.5 kPa in at-risk populations outperformed fibrosis scores in terms of accuracy. Screening with TE was cost-effective, with mean ICER ranging from 2570 €/QALY for a population at risk of alcohol-related liver disease (age ≥ 45 years) to 6217 €/QALY in the general population [39]. Overall, there was a 12% chance of TE screening, even though it was cost saving across countries and populations. This study clearly documents that screening for liver fibrosis with TE can be a cost-effective intervention for European and Asian populations, even in primary care, and may even be cost saving.

For various other screening tools, a comparative cost-utility model analysis of different annual noninvasive screening strategies has been conducted in Canada using a third-party payer perspective in a general population compared to screening in a high-risk obese or diabetic population<sup>[40]</sup>. The investigated screening algorithms involved the NFS, cytokeratin-18, TE and acoustic radiation force impulse (ARFI) imaging for detecting advanced fibrosis (≥ F3). Liver biopsy and magnetic resonance elastography were compared as confirmation methods. Compared with no screening, screening in high-risk obese or diabetic populations was more cost-effective than in the unselected general population. Interestingly, liver biopsy confirmation was not found to be cost-effective. These data suggest that annual NASH screening can be costeffective in high-risk obese or diabetic populations in a Western country.

Using a different simulation model in the United States, the effectiveness and costeffectiveness of US screening for NAFLD followed by liver biopsy has been assessed for type 2 diabetic patients<sup>[41]</sup>. In this more basic NASH screening strategy, all patients received a one-time screening US, individuals with hyperechogenicity on US underwent subsequent liver biopsy, and those found to have NASH received medical therapy to decrease disease progression. Screening for NASH decreased the number of individuals who developed cirrhosis by 12.9% and resulted in an 11.9% reduction in liver-related deaths. However, the screening strategy resulted in only 0.02 fewer QALYs due to the disutility associated with treatment and was dominated by the "no screening" strategy<sup>[41]</sup>. The impact of treatment efficacy and treatment-related side effects became clear in this study because when the model excluded the treatmentrelated quality-of-life decrement, screening became cost-effective. This study documents that treatment-associated side effects are relevant for quality of life and impact QALYs and the suitability of screening.

Referral strategies between primary care and secondary care by specialists have also been investigated. Given the high prevalence of NAFLD in Western countries, the optimal evaluation of NAFLD likely involves triage by a primary care physician (PCP) with advanced disease managed by gastroenterologists or hepatologists. Screening in a cohort of 10000 simulated United States-American patients with NAFLD performed in either PCP or referral clinics was simulated<sup>[42]</sup>. Risk stratification by the PCP using the NFS alone costs approximately 20% more per QALY than usual care costs. In the microsimulation, at a willingness-to-pay threshold of \$100000, the NFS alone in the PCP setting was the most cost-effective strategy in 94.2% of samples, followed by the combination NFS/vibration-controlled transient elastography in the PCP setting (5.6%) and usual care in 0.2% [42]. This study indicates that risk stratification of pa-



tients with NAFLD in primary care is a cost-effective strategy that should be further explored in clinical practice.

Finally, the outcome of the entire diagnostic chain is relevant for decision making upon screening. This certainly includes the likelihood of referral to the specialist after obtaining a risk surrogate (which is often moderate at best), the availability of effective drugs for the target disease (in case of NASH to be established) and relevant side effects of the treatment impacting quality of life. Taking into account the emerging awareness campaigns among the public and PCPs and ongoing phase 3 treatment studies for NASH patients, it is likely that the impact of screening on the overall outcome could improve over the near future.

# WHO TO SCREEN?

NAFLD is an asymptomatic disease in the early phase, often leading to a late diagnosis [43]. In a large population-based, cross-sectional study from Barcelona, the authors found elevated liver stiffness (as defined with TE > 6.8 kPa) in 9% of the participants, and NAFLD was the leading etiology (followed by alcohol risk consumption)[44]. Risk factors for elevated liver stiffness included obesity, type 2 diabetes and the presence of metabolic syndrome (each with a prevalence of elevated liver stiffness in 20%-30%). This study convincingly underlines the importance of NAFLD in the general population but especially in the known risk groups. While the prevalence of NAFLD in the general population is quite high (20%-30%), only approximately 7%-10% of NAFLD patients develop relevant complications of this disease, such as advanced fibrosis, cirrhosis or hepatocellular carcinoma (HCC)[45,46] (Figure 1). Thus, screening the entire population cannot (yet) be justified because too many patients would suffer overdiagnosis and possibly overtherapy. For advanced testing or invasive diagnostic measures such as liver biopsy, which applies to a selected patient population of still 3%-5%, primary testing to rule out low-risk individuals appears mandatory.

These numbers from the general population, however, do not apply to patient groups with increased NAFLD prevalence and increased risk for advanced disease. In the presence of the risk factors diabetes and obesity, the prevalence of NAFLD increases to 75% [47,48]. Diabetes and obesity are clear independent risk factors for the development of NASH-related fibrosis[46,47] and other factors of the metabolic syndrome are closely associated [49]. In addition, patients with these underlying diseases are more likely to develop complications of NAFLD[48]. Consequently, screening in the group of patients with these risk factors for complications is particularly important<sup>[50]</sup>. Elevated liver enzymes alone are sufficient as a reason for screening but are not sufficient as a sole decision criterion, as relevant NAFLD with fibrosis or cirrhosis may be present even with normal transaminases[51-53].

These facts warrant screening of this risk population[54], especially at higher HbA1c levels[54]. In some cohorts, patients with NAFLD also had an older age > 50 years in addition to the above risk factors[55-57], and an increased prevalence of NAFLD and advanced fibrosis has been shown in men[57]. These risk factors reflect quite well the collective for which screening for NAFLD is repeatedly discussed in the current literature or even concrete recommendations exist[6,7,28,29].

NAFLD is linked to several other diseases and is connected to metabolic disturbances. It is straightforward to consider the presence of NAFLD in patients with such concomitant diseases, one of the most important being coronary heart disease. Additionally, NAFLD should also be considered, depending on the advancement of the respective disease, in diseases such as polycystic ovary syndrome, sleep apnea, hypothyroidism, depression, renal insufficiency or psoriasis[7,58-60]. Making a specific screening recommendation for these patients is probably not warranted at this time; further risk profiles are needed here to justify such screening in selected patient groups with these diseases.

General screening of close relatives is also not reasonable despite some familial clustering and genetic factors (e.g., PNPLA3[61]) that may influence the course of NAFLD. The penetrance of these genetic risk factors is too low to justify screening in the presence alone (RR 3.26 for the histological presence of NAFLD per effect allele [62]). However, relatives with the presence of the abovementioned risk factors should definitely be screened for the presence of NAFLD[6]. Screening with diabetes type 2 as a central risk factor again has very recently been shown to be cost effective in the United States by avoiding advanced liver-specific disease and endpoints (all calculated screening models based on US and AST, with an ICER between \$17000 and \$35000/ QALY[63], see also Cost-effectiveness of NAFLD screening).



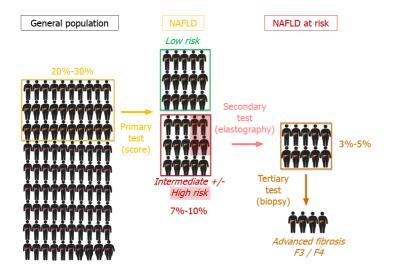


Figure 1 Nonalcoholic fatty liver disease patient proportions according to risk assessment. Stepwise enrichment of nonalcoholic fatty liver disease (NAFLD) patients at risk for advanced fibrosis using a three-step strategy with score-based primary testing in a subgroup of the general population at risk for NAFLD and elastometric secondary testing to identify candidate patients for liver biopsy represents the third and final step in most algorithms. Patients with a diagnosis of NAFLD by either surrogate scores or ultrasound (20%-30% of the general population) are divided into low-risk vs intermediate-to-high-risk subgroups (the latter 7%-10% of the general population). After elastometry testing, half of these subjects can be assigned to a high likelihood of advanced fibrosis F3/F4 and should be subjected to liver biopsy. NAFLD: Nonalcoholic fatty liver disease.

# WHO SCREENS?

A decision about who carries out screening is determined by the care structures of a particular health care system rather than by the efficacy of particular screening procedures. Even if certain diagnostic procedures proved to be cost-efficient for screening (e.g., LSM as shown above in section 3), the lack of a broad availability of LSM-determining procedures may preclude its application. Consequently, more broadly available blood-based tests are needed, and the design of a screening algorithm must then be aligned with the capabilities of those performing the screening[64-66].

In many countries, almost all patients are primarily cared for by PCPs. A certain proportion of patients defined in the at-risk population (see above) are assigned to specialists (diabetologists/endocrinologists, cardiologists), but numerous patients with diabetes mellitus, obesity, and arterial hypertension are also treated exclusively by PCPs (e.g., in the context of so-called disease management programs). In Europe, screening algorithms are implemented in a total of only 5 countries and are located in the primary health care sector in all of these countries (Belgium, Denmark, Czech Republic, Slovakia, and United Kingdom [35]). However, there are sometimes considerable structural differences in the health care systems of these countries.

Due to access to patients, comprehensive risk population screening in many countries can only be in the hands of PCPs, possibly supported by diabetologists and cardiologists. This group of physicians is particularly suited to broadly identify the major risk diseases for NAFLD and thus to determine the individual NAFLD risk in these patients<sup>[67]</sup>. This assessment is also in line with existing EASL recommendations [6] and a recently developed algorithm for general practitioners and diabetologists [68]. Direct referral of all patients at risk to hepatologists is not feasible. The need for a screening filter at the primary care level to prevent unnecessary referrals to specialists is shown by data from England ("Camden and Islington NAFLD pathway" [69]) and the United States [70]. In both studies, almost 90% of unnecessary referrals could be avoided by structured screening at the primary care provider level. On the other hand, in an American study, more than 25% of NAFLD patients referred to a hepatologist without screening already had advanced fibrosis (characterized as at least F3 with TE measurement[54]).

Data on awareness of NAFLD at GP level are rare. In the United States, data from the United States Veteran Affairs Database showed that NAFLD is significantly underdiagnosed in primary care patients<sup>[71]</sup>. Patients with abnormal alanine aminotransferase (ALT)/glutamate pyruvate transaminase (GPT) without other known liver disease (viral hepatitis and alcohol use were largely excluded by data analysis) were detected in only 40% of cases in this study, received a suspected diagnosis of NAFLD in only 21%, received therapeutic counseling in only 15% and were referred to a specialist in only 3% of cases. Initially, there is no reason to assume that the situation

in other countries differs significantly from these results. A study by the professional association of gastroenterologists in private practice in Germany (bng) showed for a cohort of NAFLD patients in secondary care that approximately 10% of these patients already had advanced fibrosis according to FIB-4 screening, but even these patients were not consistently counseled or guided regarding therapy [55]. Only 27% of patients with presumed advanced fibrosis in this study received nutritional counseling. In this respect, education and training activities for PCPs are definitely necessary to increase awareness of the presence and risks of NAFLD and to create acceptance for screening. Diabetologists and cardiologists should also be included by these measures, as they should also be involved in screening due to their spectrum of patients they treat.

Integration of primary care identification of patients at risk for the presence of NAFLD, particularly with advanced fibrosis, into secondary testing facilities at a specialist setting is a crucial issue for the overall efficacy of a screening algorithm (Figure 2). Dedicated elastography platforms have been established at several places, such as in our own center[72]. The likelihood of referral of "intermediate or high risk" individuals to secondary care, the proportion of subjects with "indeterminate" test results (the so-called "gray zone" of respective score-based tests) and the availability of advanced testing platforms for referral are relevant factors at this interface. As pointed out, existing or emerging networks between PCPs and specialists are key to optimizing a bidirectional transition into secondary testing and, in case of "low risk", back to long-term observation and basic treatment in a primary setting.

### HOW TO SCREEN?

#### Value of transabdominal ultrasonography of the liver in NAFLD

US is a widely available, cost-effective, radiation-free method that allows assessment of hepatic fatty degeneration [73]. Hepatic fatty degeneration results in an increase in the echogenicity of the liver parenchyma (e.g., compared with the renal parenchyma). US is thus suitable as a screening method for NAFLD. However, steatosis below 10% of hepatocytes is not detected, and up to 20% is unreliably detected [74] (especially with microvesicular fatty degeneration). In moderate and severe hepatic steatosis, good sensitivity (85%-96%) is achieved with specificity up to 98% [75]. The best results are seen above a liver fat content of 12.5%, where AUROC values under consideration of different echographic parameters reached comparable results to H-magnetic resonance spectroscopy (MRS)[76]. With the above referenced threshold, exclusion of steatosis by US is not completely possible. With regard to possible fibrosis of the liver, US diagnostics do not allow reliable determination and staging[73].

#### Noninvasive measurement of hepatic steatosis and fibrosis by elastography

US-based shear wave elastography techniques are well suited as a method for measuring liver stiffness to detect or exclude advanced liver fibrosis and cirrhosis in NASH. In addition, FibroScan, for example, now also offers the possibility of quantifying the fat content of the liver *via* the measurement of additional parameters.

The CAP measurement integrated in the FibroScan achieved AUROC values between 0.7[77] and 0.84[78] in studies with more than 400 patients each for (histologically confirmed) steatosis of > 33% and > 66%.

Different elastography techniques are now available on the market, and a differentiated overview cannot be given here but is available elsewhere [79]. While TE using FibroScan requires the purchase of a dedicated device, other techniques, such as ARFI imaging (Siemens), Elast-PQ (Philipps), and supersonic shear-wave elastography (SWE, Aixplorer), offer the advantage of being integrated into routine US equipment [73].

In large cohorts from Europe and Asia, the reliability of TE, its superiority over fibrosis scores, and even its cost-effectiveness have been demonstrated, at least for certain at-risk populations, in determining liver fibrosis of different origins[39]. TE is also well suited for quantifying fibrosis in NAFLD. Here, sensitivity, specificity, and AUROC values improve as fibrosis progresses, reaching values of approximately 92% and 0.89 for cirrhosis (F4), respectively [77,80]. Difficulties in estimating fibrosis in obese patients with the normal (M) probe[81] were countered by the company's introduction of an XL probe for particularly obese patients, which provides reliable values and is automatically chosen if the patient has appropriate physical conditions [82,83]

Apart from slight differences in patients with different body types, the diagnostic value of the different elastography methods in determining liver fibrosis in NAFLD



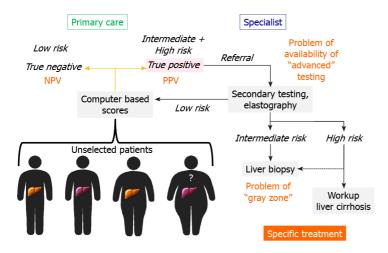


Figure 2 Linking primary care to hepatology. Unselected patients from the general population are most likely in contact with primary care. In primary care, patients at risk for the presence of nonalcoholic fatty liver disease and according to computer-based scores at risk for advanced fibrosis should be transferred into secondary testing facilities at a specialist setting. Critical for the overall efficacy of a screening algorithm are the likelihood of referral of "intermediate or high risk" individuals to secondary care, the proportion of subjects with "indeterminate" test results ("gray zone" of score-based tests) and the availability of advanced testing platforms for referral. NPV: Negative predictive value; PPV: Positive predictive value.

patients appears to be similar. Several studies with different populations and study designs yielded similar AUROC values for TE, SSI, ARFI, and 2D-SWE[84-86]. However, problems with a tendency to overestimate fibrosis occurred in bariatric, extremely obese (median BMI 47 kg/sqm) patients for both TE and ARFI, where the ELF score was actually superior to these two elastography methods[87]. Nevertheless, the procedures should also be well suited for screening most patients. The availability of the methods is very heterogeneous, so broad screening with elastography is currently not possible.

# Value of magnetic resonance imaging and computed tomography in the diagnosis and screening of NAFLD

The availability of computed tomography (CT) is bound to institutions with large medical devices but is well reproducible and reliably determines the fat content of the liver by measuring organ density[73]. In a meta-analysis comparing different radiological methods, CT performed rather modestly with a sensitivity of 46%-72%[88]. At least moderate hepatic fatty degeneration can be diagnosed if the density ratio of the liver and spleen on native CT has a cutoff value > 1.1[89]. Dual-energy CT has been able to show promising results for quantifying fat content in the liver in smaller cohorts, even in comparison with magnetic resonance imaging[90]. However, such techniques are poorly validated and not widely available. Overall, CT should not be used as a primary screening method for detecting NAFLD because of its cost, lack of broad availability, and substantial radiation exposure.

Magnetic resonance imaging (MRI), though also a large medical device, is a radiologic imaging modality without any radiation exposure. Certain modalities of MRI can be used to determine both the fat content of the liver and the fibrosis stage quite reliably[73]. MR-based quantification of liver fat content using proton density fat fraction (PDFF) has high linearity and precision with simple postprocessing[91], but it is also not suitable for screening large risk groups because of cost and effort[92]. Compared with histology as a reference standard and in comparison to CAP, PDFFbased determinations have a higher diagnostic accuracy for detecting steatosis (histological grade 1-3) with an AUROC of 0.96 up to 0.99, a sensitivity of 96%, and a specificity of 100%[93,94]. MRS has the highest accuracy for fat assessment in the literature[88,92,95] but is currently limited to research centers due to a lack of standardization of methodology and high costs for hardware and software requirements [73].

MR elastography measures liver stiffness significantly more reliably than US-based elastography techniques[85,96]. In a biopsy-controlled study of 100 patients, an AUROC of 0.98 was achieved at 40 Hz[97]. A joint analysis from 12 studies with over 900 patients still showed summary AUROC values of 0.93-0.95[98]. MR elastography also correlated better to clinical fibrosis parameters and scores than TE[99] but remains restricted to specialized centers[92].

Multiparametric MRI with determination of fat content (by PDFF or spectroscopy) and fibrosis (by MR elastography) was superior to the respective FibroScan-based non-MR methods (CAP for steatosis and TE for fibrosis) in a comprehensive new study [100] and cost-effective for risk stratification of NAFLD in a United Kingdom study [101]. Nevertheless, these methods are not (yet) suitable for broad screening due to lack of availability and high costs.

#### Laboratory chemistry scores

Because screening must be performed primarily by PCPs, screening tools must be widely available, inexpensive, and noninvasive[58,66,67]. This allows screening to be performed on a day-to-day basis and, more importantly, increases the acceptance of screening by the physicians performing it. The two-step design with the verification of steatosis and fibrosis risk improves the specificity (and in some cases even the sensitivity) of screening[65,102]. Positively screened patients must be transferred to a hepatologist for further evaluation. In this context, the proportion of positively screened patients should not be too large to avoid overloading hepatologists[65,103]. The extent of the diagnostic "gray zone" is of particular importance in this regard and can vary substantially from test to test. In any case, however, patients with prolonged or repeated elevations of GPT/ALT should be referred for further evaluation (as is usually the case), as they are generally at increased risk for liver disease or injury [51, 104.105].

There are significant differences between different countries and health care systems in the availability and cost-effectiveness of different screening tools. However, despite the limited sensitivity of US, this procedure is an attractive screening option for PCPs because of its ease of performance. More technically sophisticated and sensitive procedures such as CAP or elastography are generally not available at this level of care.

Steatosis scores correlate with insulin resistance. Their diagnostic performance for steatosis depends on the degree of fatty degeneration, fibrosis, and inflammation[106]. Assuming at least moderate steatosis is relevant, the performance of the fatty liver index (FLI) and NAFLD liver fat score is best, with the highest AUROC values with a positive predictive value of 99%, but without safe exclusion of steatosis below the cutoff[106-108]. Only the FLI can easily be obtained from routine values in family practice (see Table 1) and should therefore be used when US is not feasible[109].

Fibrosis scores also vary in both availability and quality of information. In this regard, the sensitivity and specificity of each score for significant fibrosis, advanced fibrosis, and cirrhosis are quite different and additionally vary depending on the population screened (population screening vs high-risk screening vs screening of confirmed NAFLD)[110]. Scores that require the determination of expensive specialty laboratory parameters are not suitable for primary care screening, nor are scores that include, at least in part, unavailable laboratory parameters or instrumental procedures. Although these special scores are superior to routine scores, as expected[111], and would also improve specificity in combination with them[112], the lack of availability and the lack of acceptance of these special scores by general practitioners, based in part on complicated determination, hinder their widespread use. This applies, for example, to the ELF test[113] (hyaluronic acid, TIMP-1, and procollagen peptide), which is of similar prognostic value to liver biopsy[114], and the fibrometer VCTE test (with elastography), which is also superior to purely laboratory chemistry-clinical indices [24].

Scores with readily available routine parameters for fibrosis risk include NFS, FIB-4 score, APRI score, Forns score, and BARD score. The first two (NFS, FIB-4) are superior to the last three (APRI, Forns, BARD) in screening fibrosis in the NAFLD cohort[115,116]. In a recent systematic review, this could be confirmed, especially for the hardest endpoint (mortality)[117]. These two scores (FIB-4 and NFS) are also suitable for screening patients with normal ALT[118] and can be easily determined via internet-based calculators.

In population screening, all scores have significant weaknesses and are therefore of limited use for this question[110]. However, the discriminatory performance of all tests is significantly better in high-risk collectives[110]. Although the FIB-4 score was initially developed for the detection of hepatitis C virus fibrosis[119], it has since been validated<sup>[120]</sup> and compared<sup>[121]</sup> in NAFLD collectives and may be considered suitable in principle for liver fibrosis of other etiologies. The FIB-4 score has an additional advantage over the NFS in that no albumin value is needed and that the proportion of intermediate tested patients is somewhat smaller[65,116]. However, both scores have lower specificity in patients > 65 years of age[122], which may increase the referral rate to the specialist due to a higher proportion of false-positive screened



Table 1 Scores for diagnosing steatosis and fibrosis with parameters used														
	Routine parameters								Spe	Special parameters				
Scores for Steatosis	AST	ALT	yGT	Platelets	TG	Bilirubin	BMI	Waist	Age	Sex	Diab.	A2 M	HA	Other
FLI			Х		Х		х	Х						
HSI	х	х					х			х	х			
Steato-Test		Х	Х		Х	Х			Х	х	Gluc	Х		Apo-A1, Haptoglobin, Cholesterol
NAFLD-LFS	х	х									Х			Insulin
VAI					Х		х	Х						
TyG					Х						Gluc			
Scores for fib	rosis													
NFS	Х	Х		х			х		х		Х			Albumin
FIB-4	Х	Х		х					х					
APRI	Х			х										
ELF													Х	PIIINP, TIMP-1
Fibrotest		х	Х			Х						Х		Haptoglobin,Apo- A1
Fibrometer (V2G) ((V3G))	х		((X)), for HA	Х					х	(X)		Х	Х	Prothrombin, Urea
NIKEI	х	х				х			х					

New fibrometer versions (V2G, V3G) and their respective parameters labeled with brackets: (V2G) and ((V3G)). AST: Aspartate-aminotransferase; ALT: Alanine-aminotransferase; yGT: gamma-glutamyltransferase; TG: triglycerides; BMI: Body mass index; Diab.: Diabetes; A2M: Alpha-2-microglobulin; HA: Hyaluronic acid; Gluc: Glucose; PIIINP: Procollagen-III-peptide; TIMP-1: Tissue inhibitor of metalloproteinases I; Apo-A1: Apo-A1-lipoprotein; FLI: Fatty liver index; HIS: Hepatic steatosis index; NAFLD-LFS: Nonalcoholic fatty liver-liver fat score; VAI: Visceral adiposity index; TyG: Triglyceride and glucose index; NFS: NAFLD fibrosis score; FIB-4: Fibrosis-4; APRI: AST-platelet-ratio index; ELF: Enhanced liver fibrosis; NIKEI: Noninvasive Koeln-Essen-index.

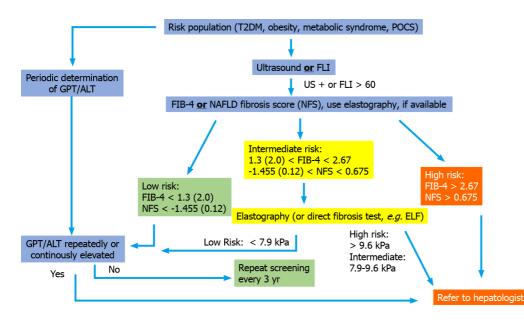
patients. Data from a screening study of type 2 diabetes patients show that the use of age-adjusted cutoffs on FIB-4 (in delineating negative *vs* intermediate) reduces the number of patients tested intermediate (from 38.3% to 15.4%[65]). Repeated measurements of laboratory scores could also help to identify patients at risk of severe liver disease in the general population, as was recently shown for repeated measurements of FIB-4 within 5 years[23].

The screening strategy proposed in Figure 3 relies on recent proposals and takes into account the aforementioned prerequisites of high-risk screening by PCPs but may not currently be evidence-based in several areas. In particular, this concerns the handling of the intermediate-risk group, the screening interval in low-risk patients, and the cost-effectiveness of the entire algorithm. In addition, the screening recommendation given requires further education and possibly training of PCPs about the prevalence and prognosis of NAFLD.

#### CONCLUSION

It is time for NAFLD screening. NAFLD is hard to diagnose in the early phase of the disease. The prevalence of this disease is increasing in countries with Western lifestyles, and the complication rate (inflammation, fibrosis, cirrhosis and HCC) is high in patients with metabolic dysfunction. Additionally, there are inexpensive noninvasive tools for the diagnosis of steatosis and fibrosis, leading to a reliable identification of persons at risk who can be referred to hepatologists. Apart from lifestyle modification, there are evolving drug treatments shortly before approval or in the late phases of clinical trials.

Boishidena® WJG https://www.wjgnet.com



#### Figure 3 Possible screening algorithm that can be modified according to availability but contains the two main elements (detection of steatosis and fibrosis risk) and can be performed in the primary care physician's office. The algorithm corresponds well to the so-called European

algorithm of the EASL-EASD-EASO Clinical Practice Guidelines[6] and to a recently proposed approach for family physicians and diabetologists[68] but is simpler to use. The sequences of fatty liver index and Fibrosis-4 (FIB-4) have been decisively studied for screening in a high-risk population of type 2 diabetes patients[65]. The use of age-adjusted cutoff values (in parentheses) is reasonable to reduce the high proportion of intermediate tested individuals. The sequential use of FIB-4, nonalcoholic fatty liver disease fibrosis score or enhanced liver fibrosis in the intermediate group has not been investigated in studies so far, but there are first studies on the basic sequential use of noninvasive fibrosis scores [123]. FLI: Fatty liver index; FIB-4: Fibrosis-4; T2DM: Type 2 diabetes mellitu; NFS: Nonalcoholic fatty liver disease fibrosis score; GPT: Glutamate pyruvate transaminase; ALT: Alanine aminotransferase.

> Studies show that screening for NAFLD, at least for a risk population, is cost effective and will help to prevent serious hepatic consequences of pandemic metabolic dysfunction. However, it will not be easy to implement comprehensive screening programs in all countries since there are large structural differences between national health systems. For example, the extent of availability of elastography will decide in each country, whether this promising technique can be used in broad screening approaches or whether US and lab scores will be necessary for PCPs to conduct screening for NAFLD. Therefore, each screening algorithm (as the one depicted in Figure 3) should be adapted locally depending on the broad availability of methods for detecting steatosis and fibrosis. Additionally, the screening population (i.e. the patients with an amount of risk factors high enough for qualifying for the screening program) has to be determined in each country individually depending on the epidemiology of NAFLD in this country.

> So what is to be done? We have to increase awareness for NAFLD and its consequences in the population and in primary care. National professional gastroenterology and hepatology societies have to develop guidelines for screening programs depending on the structure of the population and health care system of their respective country. National health systems must implement reimbursement for the tools needed for reliable screening. Hepatologists should prepare for rising numbers of patients referred for risk stratification and specific counseling.

# REFERENCES

- Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of 1 nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. Hepatology 2016; 64: 73-84 [PMID: 26707365 DOI: 10.1002/hep.28431]
- 2 Eslam M, Newsome PN, Sarin SK, Anstee QM, Targher G, Romero-Gomez M, Zelber-Sagi S, Wai-Sun Wong V, Dufour JF, Schattenberg JM, Kawaguchi T, Arrese M, Valenti L, Shiha G, Tiribelli C, Yki-Järvinen H, Fan JG, Grønbæk H, Yilmaz Y, Cortez-Pinto H, Oliveira CP, Bedossa P, Adams LA, Zheng MH, Fouad Y, Chan WK, Mendez-Sanchez N, Ahn SH, Castera L, Bugianesi E, Ratziu V, George J. A new definition for metabolic dysfunction-associated fatty liver disease: An international expert consensus statement. J Hepatol 2020; 73: 202-209 [PMID: 32278004 DOI: 10.1016/i.ihep.2020.03.039]
- 3 Estes C, Anstee QM, Arias-Loste MT, Bantel H, Bellentani S, Caballeria J, Colombo M, Craxi A,



Crespo J, Day CP, Eguchi Y, Geier A, Kondili LA, Kroy DC, Lazarus JV, Loomba R, Manns MP, Marchesini G, Nakajima A, Negro F, Petta S, Ratziu V, Romero-Gomez M, Sanyal A, Schattenberg JM, Tacke F, Tanaka J, Trautwein C, Wei L, Zeuzem S, Razavi H. Modeling NAFLD disease burden in China, France, Germany, Italy, Japan, Spain, United Kingdom, and United States for the period 2016-2030. J Hepatol 2018; 69: 896-904 [PMID: 29886156 DOI: 10.1016/j.jhep.2018.05.036]

- 4 Younossi ZM, Blissett D, Blissett R, Henry L, Stepanova M, Younossi Y, Racila A, Hunt S, Beckerman R. The economic and clinical burden of nonalcoholic fatty liver disease in the United States and Europe. *Hepatology* 2016; 64: 1577-1586 [PMID: 27543837 DOI: 10.1002/hep.28785]
- 5 Wong RJ, Aguilar M, Cheung R, Perumpail RB, Harrison SA, Younossi ZM, Ahmed A. Nonalcoholic steatohepatitis is the second leading etiology of liver disease among adults awaiting liver transplantation in the United States. Gastroenterology 2015; 148: 547-555 [PMID: 25461851 DOI: 10.1053/j.gastro.2014.11.039]
- European Association for the Study of the Liver (EASL); European Association for the Study of Diabetes (EASD); European Association for the Study of Obesity (EASO). EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease. Diabetologia 2016; **59**: 1121-1140 [PMID: 27053230 DOI: 10.1007/s00125-016-3902-y]
- Chalasani N, Younossi Z, Lavine JE, Charlton M, Cusi K, Rinella M, Harrison SA, Brunt EM, 7 Sanyal AJ. The diagnosis and management of nonalcoholic fatty liver disease: Practice guidance from the American Association for the Study of Liver Diseases. Hepatology 2018; 67: 328-357 [PMID: 28714183 DOI: 10.1002/hep.29367]
- Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW, Ferrell LD, Liu YC, 8 Torbenson MS, Unalp-Arida A, Yeh M, McCullough AJ, Sanyal AJ; Nonalcoholic Steatohepatitis Clinical Research Network. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatology* 2005; **41**: 1313-1321 [PMID: 15915461 DOI: 10.1002/hep.20701]
- 9 Bedossa P, Poitou C, Veyrie N, Bouillot JL, Basdevant A, Paradis V, Tordjman J, Clement K. Histopathological algorithm and scoring system for evaluation of liver lesions in morbidly obese patients. Hepatology 2012; 56: 1751-1759 [PMID: 22707395 DOI: 10.1002/hep.25889]
- 10 Angulo P, Kleiner DE, Dam-Larsen S, Adams LA, Bjornsson ES, Charatcharoenwitthaya P, Mills PR, Keach JC, Lafferty HD, Stahler A, Haflidadottir S, Bendtsen F. Liver Fibrosis, but No Other Histologic Features, Is Associated With Long-term Outcomes of Patients With Nonalcoholic Fatty Liver Disease. Gastroenterology 2015; 149: 389-97.e10 [PMID: 25935633 DOI: 10.1053/j.gastro.2015.04.043]
- 11 Dulai PS, Singh S, Patel J, Soni M, Prokop LJ, Younossi Z, Sebastiani G, Ekstedt M, Hagstrom H, Nasr P, Stal P, Wong VW, Kechagias S, Hultcrantz R, Loomba R. Increased risk of mortality by fibrosis stage in nonalcoholic fatty liver disease: Systematic review and meta-analysis. Hepatology 2017; 65: 1557-1565 [PMID: 28130788 DOI: 10.1002/hep.29085]
- 12 Ekstedt M, Hagström H, Nasr P, Fredrikson M, Stål P, Kechagias S, Hultcrantz R. Fibrosis stage is the strongest predictor for disease-specific mortality in NAFLD after up to 33 years of follow-up. Hepatology 2015; 61: 1547-1554 [PMID: 25125077 DOI: 10.1002/hep.27368]
- Hagström H, Nasr P, Ekstedt M, Hammar U, Stål P, Hultcrantz R, Kechagias S. Fibrosis stage but 13 not NASH predicts mortality and time to development of severe liver disease in biopsy-proven NAFLD. J Hepatol 2017; 67: 1265-1273 [PMID: 28803953 DOI: 10.1016/j.jhep.2017.07.027]
- 14 Hagström H, Nasr P, Ekstedt M, Kechagias S, Stål P, Bedossa P, Hultcrantz R. SAF score and mortality in NAFLD after up to 41 years of follow-up. Scand J Gastroenterol 2017; 52: 87-91 [PMID: 27616339 DOI: 10.1080/00365521.2016.1230779]
- 15 Taylor RS, Taylor RJ, Bayliss S, Hagström H, Nasr P, Schattenberg JM, Ishigami M, Toyoda H, Wai-Sun Wong V, Peleg N, Shlomai A, Sebastiani G, Seko Y, Bhala N, Younossi ZM, Anstee QM, McPherson S, Newsome PN. Association Between Fibrosis Stage and Outcomes of Patients With Nonalcoholic Fatty Liver Disease: A Systematic Review and Meta-Analysis. Gastroenterology 2020; 158: 1611-1625.e12 [PMID: 32027911 DOI: 10.1053/j.gastro.2020.01.043]
- 16 Geier A, Boursier J. Non-invasive diagnosis of patients with 'at-risk' NAFLD : only fibrosis counts? Gut 2020; 69: 1164-1165 [PMID: 32220903 DOI: 10.1136/gutjnl-2020-320785]
- 17 Angulo P, Hui JM, Marchesini G, Bugianesi E, George J, Farrell GC, Enders F, Saksena S, Burt AD, Bida JP, Lindor K, Sanderson SO, Lenzi M, Adams LA, Kench J, Therneau TM, Day CP. The NAFLD fibrosis score: a noninvasive system that identifies liver fibrosis in patients with NAFLD. Hepatology 2007; 45: 846-854 [PMID: 17393509 DOI: 10.1002/hep.21496]
- Ratziu V, Massard J, Charlotte F, Messous D, Imbert-Bismut F, Bonyhay L, Tahiri M, Munteanu M, 18 Thabut D, Cadranel JF, Le Bail B, de Ledinghen V, Poynard T; LIDO Study Group; CYTOL study group. Diagnostic value of biochemical markers (FibroTest-FibroSURE) for the prediction of liver fibrosis in patients with non-alcoholic fatty liver disease. BMC Gastroenterol 2006; 6: 6 [PMID: 16503961 DOI: 10.1186/1471-230X-6-6]
- Guha IN, Parkes J, Roderick P, Chattopadhyay D, Cross R, Harris S, Kaye P, Burt AD, Ryder SD, 19 Aithal GP, Day CP, Rosenberg WM. Noninvasive markers of fibrosis in nonalcoholic fatty liver disease: Validating the European Liver Fibrosis Panel and exploring simple markers. Hepatology 2008; 47: 455-460 [PMID: 18038452 DOI: 10.1002/hep.21984]
- 20 McPherson S, Stewart SF, Henderson E, Burt AD, Day CP. Simple non-invasive fibrosis scoring systems can reliably exclude advanced fibrosis in patients with non-alcoholic fatty liver disease. Gut 2010; 59: 1265-1269 [PMID: 20801772 DOI: 10.1136/gut.2010.216077]



- 21 Yoneda M, Yoneda M, Fujita K, Inamori M, Tamano M, Hiriishi H, Nakajima A. Transient elastography in patients with non-alcoholic fatty liver disease (NAFLD). Gut 2007; 56: 1330-1331 [PMID: 17470477 DOI: 10.1136/gut.2007.126417]
- 22 Anstee QM, Lawitz EJ, Alkhouri N, Wong VW, Romero-Gomez M, Okanoue T, Trauner M, Kersey K, Li G, Han L, Jia C, Wang L, Chen G, Subramanian GM, Myers RP, Djedjos CS, Kohli A, Bzowej N, Younes Z, Sarin S, Shiffman ML, Harrison SA, Afdhal NH, Goodman Z, Younossi ZM. Noninvasive Tests Accurately Identify Advanced Fibrosis due to NASH: Baseline Data From the STELLAR Trials. Hepatology 2019; 70: 1521-1530 [PMID: 31271665 DOI: 10.1002/hep.30842]
- Hagström H, Talbäck M, Andreasson A, Walldius G, Hammar N. Repeated FIB-4 measurements 23 can help identify individuals at risk of severe liver disease. J Hepatol 2020; 73: 1023-1029 [PMID: 32621944 DOI: 10.1016/j.jhep.2020.06.007]
- 24 Boursier J, Guillaume M, Leroy V, Irlès M, Roux M, Lannes A, Foucher J, Zuberbuhler F, Delabaudière C, Barthelon J, Michalak S, Hiriart JB, Peron JM, Gerster T, Le Bail B, Riou J, Hunault G, Merrouche W, Oberti F, Pelade L, Fouchard I, Bureau C, Calès P, de Ledinghen V. New sequential combinations of non-invasive fibrosis tests provide an accurate diagnosis of advanced fibrosis in NAFLD. J Hepatol 2019; 71: 389-396 [PMID: 31102719 DOI: 10.1016/j.jhep.2019.04.020]
- 25 Mózes FE, Lee JA, Selvaraj EA, Jayaswal ANA, Trauner M, Boursier J, Fournier C, Staufer K, Stauber RE, Bugianesi E, Younes R, Gaia S, Lupsor-Platon M, Petta S, Shima T, Okanoue T, Mahadeva S, Chan WK, Eddowes PJ, Hirschfield GM, Newsome PN, Wong VW, de Ledinghen V, Fan J, Shen F, Cobbold JF, Sumida Y, Okajima A, Schattenberg JM, Labenz C, Kim W, Lee MS, Wiegand J, Karlas T, Yılmaz Y, Aithal GP, Palaniyappan N, Cassinotto C, Aggarwal S, Garg H, Ooi GJ, Nakajima A, Yoneda M, Ziol M, Barget N, Geier A, Tuthill T, Brosnan MJ, Anstee QM, Neubauer S, Harrison SA, Bossuyt PM, Pavlides M; LITMUS Investigators. Diagnostic accuracy of non-invasive tests for advanced fibrosis in patients with NAFLD: an individual patient data metaanalysis. Gut 2021 [PMID: 34001645 DOI: 10.1136/gutjnl-2021-324243]
- Angulo P, Bugianesi E, Bjornsson ES, Charatcharoenwitthaya P, Mills PR, Barrera F, Haflidadottir 26 S, Day CP, George J. Simple noninvasive systems predict long-term outcomes of patients with nonalcoholic fatty liver disease. Gastroenterology 2013; 145: 782-9.e4 [PMID: 23860502 DOI: 10.1053/j.gastro.2013.06.057]
- Hardy T, Wonders K, Younes R, Aithal GP, Aller R, Allison M, Bedossa P, Betsou F, Boursier J, 27 Brosnan MJ, Burt A, Cobbold J, Cortez-Pinto H, Day CP, Dufour JF, Ekstedt M, Francque S, Harrison S, Miele L, Nasr P, Papatheodoridis G, Petta S, Tiniakos D, Torstenson R, Valenti L, Holleboom AG, Yki-Jarvinen H, Geier A, Romero-Gomez M, Ratziu V, Bugianesi E, Schattenberg JM, Anstee QM; LITMUS Consortium. The European NAFLD Registry: A real-world longitudinal cohort study of nonalcoholic fatty liver disease. Contemp Clin Trials 2020; 98: 106175 [PMID: 33045403 DOI: 10.1016/j.cct.2020.106175]
- Arab JP, Dirchwolf M, Álvares-da-Silva MR, Barrera F, Benítez C, Castellanos-Fernandez M, 28 Castro-Narro G, Chavez-Tapia N, Chiodi D, Cotrim H, Cusi K, de Oliveira CPMS, Díaz J, Fassio E, Gerona S, Girala M, Hernandez N, Marciano S, Masson W, Méndez-Sánchez N, Leite N, Lozano A, Padilla M, Panduro A, Paraná R, Parise E, Perez M, Poniachik J, Restrepo JC, Ruf A, Silva M, Tagle M, Tapias M, Torres K, Vilar-Gomez E, Costa Gil JE, Gadano A, Arrese M. Latin American Association for the study of the liver (ALEH) practice guidance for the diagnosis and treatment of non-alcoholic fatty liver disease. Ann Hepatol 2020; 19: 674-690 [PMID: 33031970 DOI: 10.1016/j.aohep.2020.09.006
- 29 Wong VW, Chan WK, Chitturi S, Chawla Y, Dan YY, Duseja A, Fan J, Goh KL, Hamaguchi M, Hashimoto E, Kim SU, Lesmana LA, Lin YC, Liu CJ, Ni YH, Sollano J, Wong SK, Wong GL, Chan HL, Farrell G. Asia-Pacific Working Party on Non-alcoholic Fatty Liver Disease guidelines 2017-Part 1: Definition, risk factors and assessment. J Gastroenterol Hepatol 2018; 33: 70-85 [PMID: 28670712 DOI: 10.1111/jgh.13857]
- 30 Andermann A, Blancquaert I, Beauchamp S, Déry V. Revisiting Wilson and Jungner in the genomic age: a review of screening criteria over the past 40 years. Bull World Health Organ 2008; 86: 317-319 [PMID: 18438522 DOI: 10.2471/blt.07.050112]
- Sturdy S, Miller F, Hogarth S, Armstrong N, Chakraborty P, Cressman C, Dobrow M, Flitcroft K, Grossman D, Harris R, Hoebee B, Holloway K, Kinsinger L, Krag M, Löblová O, Löwy I, Mackie A, Marshall J, O'Hallahan J, Rabeneck L, Raffle A, Reid L, Shortland G, Steele R, Tarini B, Taylor-Phillips S, Towler B, van der Veen N, Zappa M. Half a Century of Wilson & Jungner: Reflections on the Governance of Population Screening. Wellcome Open Res 2020; 5: 158 [PMID: 32923689 DOI: 10.12688/wellcomeopenres.16057.2]
- Ginès P, Graupera I, Lammert F, Angeli P, Caballeria L, Krag A, Guha IN, Murad SD, Castera L. 32 Screening for liver fibrosis in the general population: a call for action. Lancet Gastroenterol Hepatol 2016; 1: 256-260 [PMID: 28404098 DOI: 10.1016/S2468-1253(16)30081-4]
- 33 Rau M, Geier A. An update on drug development for the treatment of nonalcoholic fatty liver disease - from ongoing clinical trials to future therapy. Expert Rev Clin Pharmacol 2021; 14: 333-340 [PMID: 33535836 DOI: 10.1080/17512433.2021.1884068]
- Asphaug L, Thiele M, Krag A, Melberg HO. Cost-Effectiveness of Noninvasive Screening for 34 Alcohol-Related Liver Fibrosis. Hepatology 2020; 71: 2093-2104 [PMID: 31595545 DOI: 10.1002/hep.30979]
- 35 Lazarus JV, Ekstedt M, Marchesini G, Mullen J, Novak K, Pericàs JM, Roel E, Romero-Gómez M,



Ratziu V, Tacke F, Cortez-Pinto H, Anstee QM; EASL International Liver Foundation NAFLD Policy Review Collaborators. A cross-sectional study of the public health response to non-alcoholic fatty liver disease in Europe. J Hepatol 2020; 72: 14-24 [PMID: 31518646 DOI: 10.1016/j.jhep.2019.08.027]

- Phisalprapa P, Supakankunti S, Charatcharoenwitthaya P, Apisarnthanarak P, Charoensak A, 36 Washirasaksiri C, Srivanichakorn W, Chaiyakunapruk N. Cost-effectiveness analysis of ultrasonography screening for nonalcoholic fatty liver disease in metabolic syndrome patients. Medicine (Baltimore) 2017; 96: e6585 [PMID: 28445256 DOI: 10.1097/MD.00000000006585]
- 37 van Katwyk S, Coyle D, Cooper C, Pussegoda K, Cameron C, Skidmore B, Brener S, Moher D, Thavorn K. Transient elastography for the diagnosis of liver fibrosis: a systematic review of economic evaluations. Liver Int 2017; 37: 851-861 [PMID: 27699993 DOI: 10.1111/liv.13260]
- Thavorn K, Coyle D. Transient Elastography and Controlled Attenuation Parameter for Diagnosing 38 Liver Fibrosis and Steatosis in Ontario: An Economic Analysis. Ont Health Technol Assess Ser 2015; 15: 1-58 [PMID: 26664666]
- Serra-Burriel M, Graupera I, Torán P, Thiele M, Roulot D, Wai-Sun Wong V, Neil Guha I, Fabrellas N, Arslanow A, Expósito C, Hernández R, Lai-Hung Wong G, Harman D, Darwish Murad S, Krag A, Pera G, Angeli P, Galle P, Aithal GP, Caballeria L, Castera L, Ginès P, Lammert F; investigators of the LiverScreen Consortium. Transient elastography for screening of liver fibrosis: Cost-effectiveness analysis from six prospective cohorts in Europe and Asia. J Hepatol 2019; 71: 1141-1151 [PMID: 31470067 DOI: 10.1016/j.jhep.2019.08.019]
- Zhang E, Wartelle-Bladou C, Lepanto L, Lachaine J, Cloutier G, Tang A. Cost-utility analysis of 40 nonalcoholic steatohepatitis screening. Eur Radiol 2015; 25: 3282-3294 [PMID: 25994191 DOI: 10.1007/s00330-015-3731-2
- 41 Corey KE, Klebanoff MJ, Tramontano AC, Chung RT, Hur C. Screening for Nonalcoholic Steatohepatitis in Individuals with Type 2 Diabetes: A Cost-Effectiveness Analysis. Dig Dis Sci 2016; 61: 2108-2117 [PMID: 26825843 DOI: 10.1007/s10620-016-4044-2]
- Tapper EB, Hunink MG, Afdhal NH, Lai M, Sengupta N. Cost-Effectiveness Analysis: Risk 42 Stratification of Nonalcoholic Fatty Liver Disease (NAFLD) by the Primary Care Physician Using the NAFLD Fibrosis Score. PLoS One 2016; 11: e0147237 [PMID: 26905872 DOI: 10.1371/journal.pone.0147237]
- Bertot LC, Jeffrey GP, Wallace M, MacQuillan G, Garas G, Ching HL, Adams LA. Nonalcoholic 43 fatty liver disease-related cirrhosis is commonly unrecognized and associated with hepatocellular carcinoma. Hepatol Commun 2017; 1: 53-60 [PMID: 29404433 DOI: 10.1002/hep4.1018]
- Caballería L, Pera G, Arteaga I, Rodríguez L, Alumà A, Morillas RM, de la Ossa N, Díaz A, Expósito C, Miranda D, Sánchez C, Prats RM, Urquizu M, Salgado A, Alemany M, Martinez A, Majeed I, Fabrellas N, Graupera I, Planas R, Ojanguren I, Serra M, Torán P, Caballería J, Ginès P. High Prevalence of Liver Fibrosis Among European Adults With Unknown Liver Disease: A Population-Based Study. Clin Gastroenterol Hepatol 2018; 16: 1138-1145.e5 [PMID: 29452268 DOI: 10.1016/j.cgh.2017.12.048]
- Younossi Z, Anstee QM, Marietti M, Hardy T, Henry L, Eslam M, George J, Bugianesi E. Global burden of NAFLD and NASH: trends, predictions, risk factors and prevention. Nat Rev Gastroenterol Hepatol 2018; 15: 11-20 [PMID: 28930295 DOI: 10.1038/nrgastro.2017.109]
- Cohen JC, Horton JD, Hobbs HH. Human fatty liver disease: old questions and new insights. 46 Science 2011; 332: 1519-1523 [PMID: 21700865 DOI: 10.1126/science.1204265]
- Jarvis H, Craig D, Barker R, Spiers G, Stow D, Anstee QM, Hanratty B. Metabolic risk factors and 47 incident advanced liver disease in non-alcoholic fatty liver disease (NAFLD): A systematic review and meta-analysis of population-based observational studies. PLoS Med 2020; 17: e1003100 [PMID: 32353039 DOI: 10.1371/journal.pmed.1003100]
- Chen K, Sng WK, Quah JH, Liu J, Chong BY, Lee HK, Wang XF, Tan NC, Chang PE, Tan HC, 48 Bee YM, Goh GBB. Clinical spectrum of non-alcoholic fatty liver disease in patients with diabetes mellitus. PLoS One 2020; 15: e0236977 [PMID: 32822391 DOI: 10.1371/journal.pone.0236977]
- 49 Sookoian S, Pirola CJ. Systematic review with meta-analysis: risk factors for non-alcoholic fatty liver disease suggest a shared altered metabolic and cardiovascular profile between lean and obese patients. Aliment Pharmacol Ther 2017; 46: 85-95 [PMID: 28464369 DOI: 10.1111/apt.14112]
- 50 Caussy C. Should We Screen High-Risk Populations for NAFLD? Curr Hepatology Rep 2019; 18: 433-443 [DOI: 10.1007/s11901-019-00497-7]
- Mofrad P, Contos MJ, Haque M, Sargeant C, Fisher RA, Luketic VA, Sterling RK, Shiffman ML, 51 Stravitz RT, Sanyal AJ. Clinical and histologic spectrum of nonalcoholic fatty liver disease associated with normal ALT values. Hepatology 2003; 37: 1286-1292 [PMID: 12774006 DOI: 10.1053/jhep.2003.50229]
- Browning JD, Szczepaniak LS, Dobbins R, Nuremberg P, Horton JD, Cohen JC, Grundy SM, 52 Hobbs HH. Prevalence of hepatic steatosis in an urban population in the United States: impact of ethnicity. Hepatology 2004; 40: 1387-1395 [PMID: 15565570 DOI: 10.1002/hep.20466]
- Fracanzani AL, Valenti L, Bugianesi E, Andreoletti M, Colli A, Vanni E, Bertelli C, Fatta E, 53 Bignamini D, Marchesini G, Fargion S. Risk of severe liver disease in nonalcoholic fatty liver disease with normal aminotransferase levels: a role for insulin resistance and diabetes. Hepatology 2008; 48: 792-798 [PMID: 18752331 DOI: 10.1002/hep.22429]
- Shieh C, Halegoua-De Marzio DL, Hung ML, Fenkel JM, Herrine SK. Timely diagnosis and staging 54 of non-alcoholic fatty liver disease using transient elastography and clinical parameters. JGH Open



2020; 4: 1002-1006 [PMID: 33102776 DOI: 10.1002/jgh3.12385]

- Hofmann WP, Buggisch P, Schubert L, Dikopoulos N, Schwenzer J, Muche M, Felten G, Heyne R, 55 Ingiliz P, Schmidt A, Stein K, Wedemeyer H, Berg T, Wiegand J, Lammert F, Zeuzem S, Schattenberg JM. The Fatty Liver Assessment in Germany (FLAG) cohort study identifies large heterogeneity in NAFLD care. JHEP Rep 2020; 2: 100168 [PMID: 32964201 DOI: 10.1016/j.jhepr.2020.100168]
- Labenz C, Huber Y, Kalliga E, Nagel M, Ruckes C, Straub BK, Galle PR, Wörns MA, Anstee QM, 56 Schuppan D, Schattenberg JM. Predictors of advanced fibrosis in non-cirrhotic non-alcoholic fatty liver disease in Germany. Aliment Pharmacol Ther 2018; 48: 1109-1116 [PMID: 30288767 DOI: 10.1111/apt.14976]
- 57 Teeratorn N, Piyachaturawat P, Thanapirom K, Chaiteerakij R, Sonsiri K, Komolmit P, Tangkijvanich P, Rerknimitr R, Adams L, Treeprasertsuk S. Screening for non-alcoholic fatty liver disease in community setting: A cohort study using controlled attenuation parameter-transient elastography. JGH Open 2020; 4: 245-250 [PMID: 32280772 DOI: 10.1002/jgh3.12252]
- 58 Rinella ME. Screening for nonalcoholic fatty liver disease in patients with atherosclerotic coronary disease? Hepatology 2016; 63: 688-690 [PMID: 26566595 DOI: 10.1002/hep.28341]
- 59 Mantovani A, Gisondi P, Lonardo A, Targher G. Relationship between Non-Alcoholic Fatty Liver Disease and Psoriasis: A Novel Hepato-Dermal Axis? Int J Mol Sci 2016; 17: 217 [PMID: 26861300 DOI: 10.3390/ijms17020217]
- Heitmann J, Frings VG, Geier A, Goebeler M, Kerstan A. Non-alcoholic fatty liver disease and 60 psoriasis - is there a shared proinflammatory network? J Dtsch Dermatol Ges 2021; 19: 517-528 [PMID: 33768700 DOI: 10.1111/ddg.14425]
- 61 Romeo S, Kozlitina J, Xing C, Pertsemlidis A, Cox D, Pennacchio LA, Boerwinkle E, Cohen JC, Hobbs HH. Genetic variation in PNPLA3 confers susceptibility to nonalcoholic fatty liver disease. Nat Genet 2008; 40: 1461-1465 [PMID: 18820647 DOI: 10.1038/ng.257]
- Speliotes EK, Yerges-Armstrong LM, Wu J, Hernaez R, Kim LJ, Palmer CD, Gudnason V, 62 Eiriksdottir G, Garcia ME, Launer LJ, Nalls MA, Clark JM, Mitchell BD, Shuldiner AR, Butler JL, Tomas M, Hoffmann U, Hwang SJ, Massaro JM, O'Donnell CJ, Sahani DV, Salomaa V, Schadt EE, Schwartz SM, Siscovick DS; NASH CRN; GIANT Consortium; MAGIC Investigators, Voight BF, Carr JJ, Feitosa MF, Harris TB, Fox CS, Smith AV, Kao WH, Hirschhorn JN, Borecki IB; GOLD Consortium. Genome-wide association analysis identifies variants associated with nonalcoholic fatty liver disease that have distinct effects on metabolic traits. PLoS Genet 2011; 7: e1001324 [PMID: 21423719 DOI: 10.1371/journal.pgen.1001324]
- 63 Noureddin M, Jones C, Alkhouri N, Gomez EV, Dieterich DT, Rinella ME; NASHNET. Screening for Nonalcoholic Fatty Liver Disease in Persons with Type 2 Diabetes in the United States Is Costeffective: A Comprehensive Cost-Utility Analysis. Gastroenterology 2020; 159: 1985-1987.e4 [PMID: 32763241 DOI: 10.1053/j.gastro.2020.07.050]
- Nones RB, Ivantes CP, Pedroso MLA. Can FIB4 and NAFLD fibrosis scores help endocrinologists 64 refer patients with non-alcoholic fat liver disease to a hepatologist? Arch Endocrinol Metab 2017; 61: 276-281 [PMID: 28225987 DOI: 10.1590/2359-3997000000233]
- Ciardullo S, Muraca E, Perra S, Bianconi E, Zerbini F, Oltolini A, Cannistraci R, Parmeggiani P, 65 Manzoni G, Gastaldelli A, Lattuada G, Perseghin G. Screening for non-alcoholic fatty liver disease in type 2 diabetes using non-invasive scores and association with diabetic complications. BMJ Open Diabetes Res Care 2020; 8 [PMID: 32049637 DOI: 10.1136/bmjdrc-2019-000904]
- 66 Castera L. Non-invasive tests for liver fibrosis in NAFLD: Creating pathways between primary healthcare and liver clinics. Liver Int 2020; 40 Suppl 1: 77-81 [PMID: 32077617 DOI: 10.1111/liv.14347
- 67 Pandyarajan V, Gish RG, Alkhouri N, Noureddin M. Screening for Nonalcoholic Fatty Liver Disease in the Primary Care Clinic. Gastroenterol Hepatol (N Y) 2019; 15: 357-365 [PMID: 31391806
- Younossi ZM, Corey KE, Alkhouri N, Noureddin M, Jacobson I, Lam B, Clement S, Basu R, 68 Gordon SC, Ravendhra N, Puri P, Rinella M, Scudera P, Singal AK, Henry L; US Members of the Global Nash Council. Clinical assessment for high-risk patients with non-alcoholic fatty liver disease in primary care and diabetology practices. Aliment Pharmacol Ther 2020; 52: 513-526 [PMID: 32598051 DOI: 10.1111/apt.15830]
- Srivastava A, Gailer R, Tanwar S, Trembling P, Parkes J, Rodger A, Suri D, Thorburn D, Sennett K, Morgan S, Tsochatzis EA, Rosenberg W. Prospective evaluation of a primary care referral pathway for patients with non-alcoholic fatty liver disease. J Hepatol 2019; 71: 371-378 [PMID: 30965069 DOI: 10.1016/j.jhep.2019.03.033]
- Leung M, Piao C, Sarkar S. NAFLD referral patterns in a large US academic center. J Hepatol 70 2020; 73: 218-219 [PMID: 32273138 DOI: 10.1016/j.jhep.2019.12.026]
- Blais P, Husain N, Kramer JR, Kowalkowski M, El-Serag H, Kanwal F. Nonalcoholic fatty liver 71 disease is underrecognized in the primary care setting. Am J Gastroenterol 2015; 110: 10-14 [PMID: 24890441 DOI: 10.1038/ajg.2014.134]
- 72 Alsenbesy M, Rau M, Weiss J, Götze O, Geier A. A 2-step fast-track elastometry service for advanced workup of nonalcoholic fatty liver disease (NAFLD) patients - single-center real-world experience of outpatient clinical practice. Z Gastroenterol 2019; 57: 1209-1217 [PMID: 31610584 DOI: 10.1055/a-0981-64841
- Roeb E, Steffen HM, Bantel H, Baumann U, Canbay A, Demir M, Drebber U, Geier A, Hampe J, 73



Hellerbrand C, Pathil-Warth A, Schattenberg JM, Schramm C, Seitz HK, Stefan N, Tacke F, Tannapfel A, Lynen Jansen P, Bojunga J. [S2k Guideline non-alcoholic fatty liver disease]. Z Gastroenterol 2015; 53: 668-723 [PMID: 26167698 DOI: 10.1055/s-0035-1553193]

- 74 Ryan CK, Johnson LA, Germin BI, Marcos A. One hundred consecutive hepatic biopsies in the workup of living donors for right lobe liver transplantation. Liver Transpl 2002; 8: 1114-1122 [PMID: 12474149 DOI: 10.1053/jlts.2002.36740]
- 75 Dasarathy S, Dasarathy J, Khiyami A, Joseph R, Lopez R, McCullough AJ. Validity of real time ultrasound in the diagnosis of hepatic steatosis: a prospective study. J Hepatol 2009; 51: 1061-1067 [PMID: 19846234 DOI: 10.1016/j.jhep.2009.09.001]
- 76 Bril F, Ortiz-Lopez C, Lomonaco R, Orsak B, Freckleton M, Chintapalli K, Hardies J, Lai S, Solano F, Tio F, Cusi K. Clinical value of liver ultrasound for the diagnosis of nonalcoholic fatty liver disease in overweight and obese patients. Liver Int 2015; 35: 2139-2146 [PMID: 25847730 DOI: 10.1111/liv.12840]
- Eddowes PJ, Sasso M, Allison M, Tsochatzis E, Anstee QM, Sheridan D, Guha IN, Cobbold JF, 77 Deeks JJ, Paradis V, Bedossa P, Newsome PN. Accuracy of FibroScan Controlled Attenuation Parameter and Liver Stiffness Measurement in Assessing Steatosis and Fibrosis in Patients With Nonalcoholic Fatty Liver Disease. Gastroenterology 2019; 156: 1717-1730 [PMID: 30689971 DOI: 10.1053/j.gastro.2019.01.042]
- de Lédinghen V, Vergniol J, Capdepont M, Chermak F, Hiriart JB, Cassinotto C, Merrouche W, Foucher J, Brigitte le B. Controlled attenuation parameter (CAP) for the diagnosis of steatosis: a prospective study of 5323 examinations. J Hepatol 2014; 60: 1026-1031 [PMID: 24378529 DOI: 10.1016/i.ihep.2013.12.018
- 79 Honda Y, Yoneda M, Imajo K, Nakajima A. Elastography Techniques for the Assessment of Liver Fibrosis in Non-Alcoholic Fatty Liver Disease. Int J Mol Sci 2020; 21 [PMID: 32516937 DOI: 10.3390/iims211140391
- 80 Kwok R, Tse YK, Wong GL, Ha Y, Lee AU, Ngu MC, Chan HL, Wong VW. Systematic review with meta-analysis: non-invasive assessment of non-alcoholic fatty liver disease--the role of transient elastography and plasma cytokeratin-18 fragments. Aliment Pharmacol Ther 2014; 39: 254-269 [PMID: 24308774 DOI: 10.1111/apt.12569]
- 81 Castéra L, Foucher J, Bernard PH, Carvalho F, Allaix D, Merrouche W, Couzigou P, de Lédinghen V. Pitfalls of liver stiffness measurement: a 5-year prospective study of 13,369 examinations. Hepatology 2010; 51: 828-835 [PMID: 20063276 DOI: 10.1002/hep.23425]
- 82 Myers RP, Pomier-Layrargues G, Kirsch R, Pollett A, Duarte-Rojo A, Wong D, Beaton M, Levstik M, Crotty P, Elkashab M. Feasibility and diagnostic performance of the FibroScan XL probe for liver stiffness measurement in overweight and obese patients. Hepatology 2012; 55: 199-208 [PMID: 21898479 DOI: 10.1002/hep.24624]
- Wong VW, Irles M, Wong GL, Shili S, Chan AW, Merrouche W, Shu SS, Foucher J, Le Bail B, 83 Chan WK, Chan HL, de Ledinghen V. Unified interpretation of liver stiffness measurement by M and XL probes in non-alcoholic fatty liver disease. Gut 2019; 68: 2057-2064 [PMID: 30658997 DOI: 10.1136/gutjnl-2018-317334]
- 84 Lee MS, Bae JM, Joo SK, Woo H, Lee DH, Jung YJ, Kim BG, Lee KL, Kim W. Prospective comparison among transient elastography, supersonic shear imaging, and ARFI imaging for predicting fibrosis in nonalcoholic fatty liver disease. PLoS One 2017; 12: e0188321 [PMID: 29176844 DOI: 10.1371/journal.pone.0188321]
- 85 Furlan A, Tublin ME, Yu L, Chopra KB, Lippello A, Behari J. Comparison of 2D Shear Wave Elastography, Transient Elastography, and MR Elastography for the Diagnosis of Fibrosis in Patients With Nonalcoholic Fatty Liver Disease. AJR Am J Roentgenol 2020; 214: W20-W26 [PMID: 31714842 DOI: 10.2214/AJR.19.21267]
- 86 Jiang W, Huang S, Teng H, Wang P, Wu M, Zhou X, Ran H. Diagnostic accuracy of point shear wave elastography and transient elastography for staging hepatic fibrosis in patients with nonalcoholic fatty liver disease: a meta-analysis. BMJ Open 2018; 8: e021787 [PMID: 30139901 DOI: 10.1136/bmjopen-2018-0217871
- Karlas T, Dietrich A, Peter V, Wittekind C, Lichtinghagen R, Garnov N, Linder N, Schaudinn A, 87 Busse H, Prettin C, Keim V, Tröltzsch M, Schütz T, Wiegand J. Evaluation of Transient Elastography, Acoustic Radiation Force Impulse Imaging (ARFI), and Enhanced Liver Function (ELF) Score for Detection of Fibrosis in Morbidly Obese Patients. PLoS One 2015; 10: e0141649 [PMID: 26528818 DOI: 10.1371/journal.pone.0141649]
- 88 Bohte AE, van Werven JR, Bipat S, Stoker J. The diagnostic accuracy of US, CT, MRI and 1H-MRS for the evaluation of hepatic steatosis compared with liver biopsy: a meta-analysis. Eur Radiol 2011; 21: 87-97 [PMID: 20680289 DOI: 10.1007/s00330-010-1905-5]
- Iwasaki M, Takada Y, Hayashi M, Minamiguchi S, Haga H, Maetani Y, Fujii K, Kiuchi T, Tanaka 89 K. Noninvasive evaluation of graft steatosis in living donor liver transplantation. Transplantation 2004; 78: 1501-1505 [PMID: 15599315 DOI: 10.1097/01.tp.0000140499.23683.0d]
- Hyodo T, Yada N, Hori M, Maenishi O, Lamb P, Sasaki K, Onoda M, Kudo M, Mochizuki T, 90 Murakami T. Multimaterial Decomposition Algorithm for the Quantification of Liver Fat Content by Using Fast-Kilovolt-Peak Switching Dual-Energy CT: Clinical Evaluation. Radiology 2017; 283: 108-118 [PMID: 28212047 DOI: 10.1148/radiol.2017160130]
- 91 Yokoo T, Serai SD, Pirasteh A, Bashir MR, Hamilton G, Hernando D, Hu HH, Hetterich H, Kühn JP, Kukuk GM, Loomba R, Middleton MS, Obuchowski NA, Song JS, Tang A, Wu X, Reeder SB,



Sirlin CB; RSNA-QIBA PDFF Biomarker Committee. Linearity, Bias, and Precision of Hepatic Proton Density Fat Fraction Measurements by Using MR Imaging: A Meta-Analysis. Radiology 2018; 286: 486-498 [PMID: 28892458 DOI: 10.1148/radiol.2017170550]

- 92 Caussy C, Johansson L. Magnetic resonance-based biomarkers in nonalcoholic fatty liver disease and nonalcoholic steatohepatitis. Endocrinol Diabetes Metab 2020; 3: e00134 [PMID: 33102797 DOI: 10.1002/edm2.134]
- 93 Imajo K, Kessoku T, Honda Y, Tomeno W, Ogawa Y, Mawatari H, Fujita K, Yoneda M, Taguri M, Hyogo H, Sumida Y, Ono M, Eguchi Y, Inoue T, Yamanaka T, Wada K, Saito S, Nakajima A. Magnetic Resonance Imaging More Accurately Classifies Steatosis and Fibrosis in Patients With Nonalcoholic Fatty Liver Disease Than Transient Elastography. Gastroenterology 2016; 150: 626-637.e7 [PMID: 26677985 DOI: 10.1053/j.gastro.2015.11.048]
- 94 Park CC, Nguyen P, Hernandez C, Bettencourt R, Ramirez K, Fortney L, Hooker J, Sy E, Savides MT, Alquiraish MH, Valasek MA, Rizo E, Richards L, Brenner D, Sirlin CB, Loomba R. Magnetic Resonance Elastography vs Transient Elastography in Detection of Fibrosis and Noninvasive Measurement of Steatosis in Patients With Biopsy-Proven Nonalcoholic Fatty Liver Disease. Gastroenterology 2017; 152: 598-607.e2 [PMID: 27911262 DOI: 10.1053/j.gastro.2016.10.026]
- Cowin GJ, Jonsson JR, Bauer JD, Ash S, Ali A, Osland EJ, Purdie DM, Clouston AD, Powell EE, 95 Galloway GJ. Magnetic resonance imaging and spectroscopy for monitoring liver steatosis. J Magn Reson Imaging 2008; 28: 937-945 [PMID: 18821619 DOI: 10.1002/jmri.21542]
- 96 Hsu C, Caussy C, Imajo K, Chen J, Singh S, Kaulback K, Le MD, Hooker J, Tu X, Bettencourt R, Yin M, Sirlin CB, Ehman RL, Nakajima A, Loomba R, Magnetic Resonance vs Transient Elastography Analysis of Patients With Nonalcoholic Fatty Liver Disease: A Systematic Review and Pooled Analysis of Individual Participants. Clin Gastroenterol Hepatol 2019; 17: 630-637.e8 [PMID: 29908362 DOI: 10.1016/j.cgh.2018.05.059]
- 97 Loomba R, Cui J, Wolfson T, Haufe W, Hooker J, Szeverenyi N, Ang B, Bhatt A, Wang K, Aryafar H, Behling C, Valasek MA, Lin GY, Gamst A, Brenner DA, Yin M, Glaser KJ, Ehman RL, Sirlin CB. Novel 3D Magnetic Resonance Elastography for the Noninvasive Diagnosis of Advanced Fibrosis in NAFLD: A Prospective Study. Am J Gastroenterol 2016; 111: 986-994 [PMID: 27002798 DOI: 10.1038/ajg.2016.65]
- 98 Liang Y, Li D. Magnetic resonance elastography in staging liver fibrosis in non-alcoholic fatty liver disease: a pooled analysis of the diagnostic accuracy. BMC Gastroenterol 2020; 20: 89 [PMID: 32252641 DOI: 10.1186/s12876-020-01234-x]
- Choi SJ, Kim SM, Kim YS, Kwon OS, Shin SK, Kim KK, Lee K, Park IB, Choi CS, Chung DH, 99 Jung J, Paek M, Lee DH. Magnetic Resonance-Based Assessments Better Capture Pathophysiologic Profiles and Progression in Nonalcoholic Fatty Liver Disease. Diabetes Metab J 2020 [PMID: 33108854 DOI: 10.4093/dmj.2020.0137]
- Lee YS, Yoo YJ, Jung YK, Kim JH, Seo YS, Yim HJ, Kim IH, Lee SY, Kim BH, Kim JW, Lee CH, 100 Yeon JE, Kwon SY, Um SH, Byun KS. Multiparametric MR Is a Valuable Modality for Evaluating Disease Severity of Nonalcoholic Fatty Liver Disease. Clin Transl Gastroenterol 2020; 11: e00157 [PMID: 32251018 DOI: 10.14309/ctg.000000000000157]
- Eddowes PJ, McDonald N, Davies N, Semple SIK, Kendall TJ, Hodson J, Newsome PN, Flintham 101 RB, Wesolowski R, Blake L, Duarte RV, Kelly CJ, Herlihy AH, Kelly MD, Olliff SP, Hübscher SG, Fallowfield JA, Hirschfield GM. Utility and cost evaluation of multiparametric magnetic resonance imaging for the assessment of non-alcoholic fatty liver disease. Aliment Pharmacol Ther 2018; 47: 631-644 [PMID: 29271504 DOI: 10.1111/apt.14469]
- Grecian SM, McLachlan S, Fallowfield JA, Kearns PKA, Hayes PC, Guha NI, Morling JR, Glancy 102 S, Williamson RM, Reynolds RM, Frier BM, Zammitt NN, Price JF, Strachan MWJ. Non-invasive risk scores do not reliably identify future cirrhosis or hepatocellular carcinoma in Type 2 diabetes: The Edinburgh type 2 diabetes study. Liver Int 2020; 40: 2252-2262 [PMID: 32638496 DOI: 10.1111/liv.14590
- 103 Blond E, Disse E, Cuerq C, Drai J, Valette PJ, Laville M, Thivolet C, Simon C, Caussy C. EASL-EASD-EASO clinical practice guidelines for the management of non-alcoholic fatty liver disease in severely obese people: do they lead to over-referral? Diabetologia 2017; 60: 1218-1222 [PMID: 28352941 DOI: 10.1007/s00125-017-4264-9]
- Verma S, Jensen D, Hart J, Mohanty SR. Predictive value of ALT levels for non-alcoholic 104 steatohepatitis (NASH) and advanced fibrosis in non-alcoholic fatty liver disease (NAFLD). Liver Int 2013; 33: 1398-1405 [PMID: 23763360 DOI: 10.1111/liv.12226]
- 105 Lee TY, Wu JC, Yu SH, Lin JT, Wu MS, Wu CY. The occurrence of hepatocellular carcinoma in different risk stratifications of clinically noncirrhotic nonalcoholic fatty liver disease. Int J Cancer 2017; 141: 1307-1314 [PMID: 28509327 DOI: 10.1002/ijc.30784]
- 106 Fedchuk L, Nascimbeni F, Pais R, Charlotte F, Housset C, Ratziu V; LIDO Study Group. Performance and limitations of steatosis biomarkers in patients with nonalcoholic fatty liver disease. Aliment Pharmacol Ther 2014; 40: 1209-1222 [PMID: 25267215 DOI: 10.1111/apt.12963]
- 107 Cheung CL, Lam KS, Wong IC, Cheung BM. Non-invasive score identifies ultrasonographydiagnosed non-alcoholic fatty liver disease and predicts mortality in the USA. BMC Med 2014; 12: 154 [PMID: 25204761 DOI: 10.1186/s12916-014-0154-x]
- Lind L, Johansson L, Ahlström H, Eriksson JW, Larsson A, Risérus U, Kullberg J, Oscarsson J. 108 Comparison of four non-alcoholic fatty liver disease detection scores in a Caucasian population. World J Hepatol 2020; 12: 149-159 [PMID: 32685107 DOI: 10.4254/wjh.v12.i4.149]



- 109 Byrne CD, Targher G. EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease: is universal screening appropriate? *Diabetologia* 2016; 59: 1141-1144 [PMID: 27053232 DOI: 10.1007/s00125-016-3910-y]
- 110 Hagström H, Talbäck M, Andreasson A, Walldius G, Hammar N. Ability of Noninvasive Scoring Systems to Identify Individuals in the Population at Risk for Severe Liver Disease. *Gastroenterology* 2020; 158: 200-214 [PMID: 31563624 DOI: 10.1053/j.gastro.2019.09.008]
- 111 Staufer K, Halilbasic E, Spindelboeck W, Eilenberg M, Prager G, Stadlbauer V, Posch A, Munda P, Marculescu R, Obermayer-Pietsch B, Stift J, Lackner C, Trauner M, Stauber RE. Evaluation and comparison of six noninvasive tests for prediction of significant or advanced fibrosis in nonalcoholic fatty liver disease. *United European Gastroenterol J* 2019; 7: 1113-1123 [PMID: 31662868 DOI: 10.1177/2050640619865133]
- 112 Inadomi C, Takahashi H, Ogawa Y, Oeda S, Imajo K, Kubotsu Y, Tanaka K, Kessoku T, Okada M, Isoda H, Akiyama T, Fukushima H, Yoneda M, Anzai K, Aishima S, Nakajima A, Eguchi Y. Accuracy of the Enhanced Liver Fibrosis test, and combination of the Enhanced Liver Fibrosis and non-invasive tests for the diagnosis of advanced liver fibrosis in patients with non-alcoholic fatty liver disease. *Hepatol Res* 2020; 50: 682-692 [PMID: 32090397 DOI: 10.1111/hepr.13495]
- 113 Vali Y, Lee J, Boursier J, Spijker R, Löffler J, Verheij J, Brosnan MJ, Böcskei Z, Anstee QM, Bossuyt PM, Zafarmand MH; LITMUS systematic review team(†). Enhanced liver fibrosis test for the non-invasive diagnosis of fibrosis in patients with NAFLD: A systematic review and metaanalysis. *J Hepatol* 2020; **73**: 252-262 [PMID: 32275982 DOI: 10.1016/j.jhep.2020.03.036]
- 114 Parkes J, Roderick P, Harris S, Day C, Mutimer D, Collier J, Lombard M, Alexander G, Ramage J, Dusheiko G, Wheatley M, Gough C, Burt A, Rosenberg W. Enhanced liver fibrosis test can predict clinical outcomes in patients with chronic liver disease. *Gut* 2010; **59**: 1245-1251 [PMID: 20675693 DOI: 10.1136/gut.2009.203166]
- 115 Xiao G, Zhu S, Xiao X, Yan L, Yang J, Wu G. Comparison of laboratory tests, ultrasound, or magnetic resonance elastography to detect fibrosis in patients with nonalcoholic fatty liver disease: A meta-analysis. *Hepatology* 2017; 66: 1486-1501 [PMID: 28586172 DOI: 10.1002/hep.29302]
- 116 Boursier J, Vergniol J, Guillet A, Hiriart JB, Lannes A, Le Bail B, Michalak S, Chermak F, Bertrais S, Foucher J, Oberti F, Charbonnier M, Fouchard-Hubert I, Rousselet MC, Calès P, de Lédinghen V. Diagnostic accuracy and prognostic significance of blood fibrosis tests and liver stiffness measurement by FibroScan in non-alcoholic fatty liver disease. *J Hepatol* 2016; 65: 570-578 [PMID: 27151181 DOI: 10.1016/j.jhep.2016.04.023]
- 117 Lee J, Vali Y, Boursier J, Spijker R, Anstee QM, Bossuyt PM, Zafarmand MH. Prognostic accuracy of FIB-4, NAFLD fibrosis score and APRI for NAFLD-related events: A systematic review. *Liver Int* 2021; **41**: 261-270 [PMID: 32946642 DOI: 10.1111/liv.14669]
- 118 Yoneda M, Imajo K, Eguchi Y, Fujii H, Sumida Y, Hyogo H, Ono M, Suzuki Y, Kawaguchi T, Aoki N, Sata M, Kanemasa K, Kohgo Y, Saibara T, Chayama K, Itoh Y, Yoshikawa T, Anzai K, Fujimoto K, Okanoue T, Nakajima A; Japan Study Group of Nonalcoholic Fatty Liver Disease (JSG-NAFLD). Noninvasive scoring systems in patients with nonalcoholic fatty liver disease with normal alanine aminotransferase levels. *J Gastroenterol* 2013; 48: 1051-1060 [PMID: 23184095 DOI: 10.1007/s00535-012-0704-y]
- 119 Sterling RK, Lissen E, Clumeck N, Sola R, Correa MC, Montaner J, S Sulkowski M, Torriani FJ, Dieterich DT, Thomas DL, Messinger D, Nelson M; APRICOT Clinical Investigators. Development of a simple noninvasive index to predict significant fibrosis in patients with HIV/HCV coinfection. *Hepatology* 2006; 43: 1317-1325 [PMID: 16729309 DOI: 10.1002/hep.21178]
- 120 Sumida Y, Yoneda M, Hyogo H, Itoh Y, Ono M, Fujii H, Eguchi Y, Suzuki Y, Aoki N, Kanemasa K, Fujita K, Chayama K, Saibara T, Kawada N, Fujimoto K, Kohgo Y, Yoshikawa T, Okanoue T; Japan Study Group of Nonalcoholic Fatty Liver Disease (JSG-NAFLD). Validation of the FIB4 index in a Japanese nonalcoholic fatty liver disease population. *BMC Gastroenterol* 2012; 12: 2 [PMID: 22221544 DOI: 10.1186/1471-230X-12-2]
- 121 Sun W, Cui H, Li N, Wei Y, Lai S, Yang Y, Yin X, Chen DF. Comparison of FIB-4 index, NAFLD fibrosis score and BARD score for prediction of advanced fibrosis in adult patients with non-alcoholic fatty liver disease: A meta-analysis study. *Hepatol Res* 2016; 46: 862-870 [PMID: 26763834 DOI: 10.1111/hepr.12647]
- 122 McPherson S, Hardy T, Dufour JF, Petta S, Romero-Gomez M, Allison M, Oliveira CP, Francque S, Van Gaal L, Schattenberg JM, Tiniakos D, Burt A, Bugianesi E, Ratziu V, Day CP, Anstee QM. Age as a Confounding Factor for the Accurate Non-Invasive Diagnosis of Advanced NAFLD Fibrosis. *Am J Gastroenterol* 2017; 112: 740-751 [PMID: 27725647 DOI: 10.1038/ajg.2016.453]
- 123 Yang M, Jiang L, Wang Y, Li X, Zou Z, Han T, Nan Y, Lu F, Zhao J. Step layered combination of noninvasive fibrosis models improves diagnostic accuracy of advanced fibrosis in nonalcoholic fatty liver disease. J Gastrointestin Liver Dis 2019; 28: 289-296 [PMID: 31517325 DOI: 10.15403/jgld-420]

WJG | https://www.wjgnet.com

September 21, 2021 Volume 27 Issue 35

WJG

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5822-5850

DOI: 10.3748/wjg.v27.i35.5822

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

REVIEW

# **Environmental perspectives of COVID-19 outbreaks: A review**

Palas Samanta, Apurba Ratan Ghosh

ORCID number: Palas Samanta 0000-0001-9369-7502; Apurba Ratan Ghosh 0000-0003-1454-7720.

#### Author contributions: Samanta P contributed to the

conceptualization, writing- original draft preparation, software running; Ghosh AR contributed to the conceptualization, supervision, writing- reviewing and editing.

Conflict-of-interest statement: The authors have no conflicts of interest to declare.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Public, environmental and occupational health

Country/Territory of origin: India

Palas Samanta, Department of Environmental Science, Sukanta Mahavidyalaya, University of North Bengal, Dhupguri 735210, West Bengal, India

Apurba Ratan Ghosh, Department of Environmental Science, The University of Burdwan, Burdwan 713104, West Bengal, India

Corresponding author: Palas Samanta, PhD, Assistant Professor, Department of Environmental Science, Sukanta Mahavidyalaya, University of North Bengal, Dhupguri, Jalpaiguri, Dhupguri 735210, West Bengal, India. samanta.palas2010@gmail.com

# Abstract

The coronavirus disease 2019 (COVID-19) pandemic, caused by the novel virus severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), began in December 2019 in China and has led to a global public health emergency. Previously, it was known as 2019-nCoV and caused disease mainly through respiratory pathways. The COVID-19 outbreak is ranked third globally as the most highly pathogenic disease of the twenty-first century, after the outbreak of SARS-CoV and Middle East respiratory syndrome in 2002 and 2012, respectively. Clinical, laboratory, and diagnostic methodology have been demonstrated in some observational studies. No systematic reviews on COVID-19 have been published regarding the integration of COVID-19 outbreaks (monitoring, fate and treatment) with environmental and human health perspectives. Accordingly, this review systematically addresses environmental aspects of COVID-19 outbreak such as the origin of SARS-CoV-2, epidemiological characteristics, diagnostic methodology, treatment options and technological advancement for the prevention of COVID-19 outbreaks. Finally, we integrate COVID-19 outbreaks (monitoring, fate and treatment) with environmental and human health perspectives. We believe that this review will help to understand the SARS-CoV-2 outbreak as a multipurpose document, not only for the scientific community but also for global citizens. Countries should adopt emergency preparedness such as prepare human resources, infrastructure and facilities to treat severe COVID-19 as the virus spreads rapidly globally.

Key Words: COVID-19; SARS-CoV-2 virus; Environmental perspectives; Epidemiological characteristics; Public health; Emergency preparedness

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.



### Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Received: December 14, 2020 Peer-review started: December 14, 2020

First decision: January 7, 2021 Revised: January 10, 2021 Accepted: August 12, 2021 Article in press: August 12, 2021 Published online: September 21, 2021

P-Reviewer: Jiang T, Pan H S-Editor: Gong ZM L-Editor: Webster JR P-Editor: Ma YJ



**Core Tip:** This review is the first attempt to integrate coronavirus disease 2019 (COVID-19) outbreaks (monitoring, fate and treatment) with respect to environmental and human health perspectives. Briefly, the paper systematically addresses the environmental aspects of the COVID-19 outbreak such as the origin of severe acute respiratory syndrome coronavirus 2, epidemiological characteristics, diagnostic methodology, treatment options and technological advancement for the prevention of COVID-19 outbreaks.

Citation: Samanta P, Ghosh AR. Environmental perspectives of COVID-19 outbreaks: A review. World J Gastroenterol 2021; 27(35): 5822-5850

URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5822.htm DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5822

# INTRODUCTION

A series of patients with unidentified pneumonia, caused by  $\beta$ -coronavirus, was reported in late December 2019 in Wuhan (Hubei Province), China. Coronavirus disease 2019 (COVID-19) outbreaks are clinically very similar to viral pneumonia. A number of experts from the PRC Centers for Disease Control declared that this respiratory disorder (alternatively known as novel coronavirus pneumonia, NCP) was caused by a novel coronavirus[1]. The World Health Organization (WHO) initially named the disease as 2019-nCoV (2019-novel coronavirus) on January 12, 2020. It was officially later named COVID-19 on February 11, 2020 by the WHO. On the same date, the International Committee on Taxonomy of Viruses named the virus as severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) after developing the genome sequence from a COVID-19 patient in Wuhan on January 7, 2020. The virus belongs to the  $\beta$ -coronavirus family, which is very prevalent in nature among other families. Similar to other viruses, the SARS-CoV-2 also has many natural hosts including different intermediate and final hosts, which makes it challenging for scientific communities to treat and prevent COVID-19 outbreaks. It has higher transmission and infection potential but causes a lower mortality rate compared with SARS-CoV and Middle East respiratory syndrome (MERS-CoV)[2]. The genomic sequence of SARS-CoV-2 revealed that it has 79.5% and 96% similarity with SARS-CoV and bat coronavirus, respectively[1], which implies that bats might be the source of SARS-CoV-2. Although the COVID-19 outbreak started in China, the virus has spread to over 213 countries with the highest rate of infection in the United States, Italy, France, and Spain among others as per data published by the WHO on December 13, 2020 (Figure 1). There are approximately 202608306 confirmed SARS-CoV-2 cases and 4293591 deaths worldwide. Consequently, COVID-19 has emerged as a global threat to public health and is steadily growing due to human-to-human transmission. Moreover, this transmission also spreads in different environmental sectors such as water, air, soil, sewage and fecal matter[3]. Additionally, this process is accelerated by a number of meteorological factors namely temperature, weather, humidity and air quality parameters including particulate matter, SOx, NOx and carbon, etc. Therefore, a better understanding of the global consequences of COVID-19 is required with regard to environmental perspectives. Accordingly, this review will address the origin of SARS-CoV-2, route of transmission, pathogenesis, epidemiological characteristics, diagnostic methodology, treatment options and technological advancement for the prevention of COVID-19 outbreaks with regard to environmental perspectives in order to acquire the latest understanding of this new infectious disease of which certain immediate as well as long-term remedial measures can be explored.

# **EPIDEMIOLOGY OF THE COVID-19 OUTBREAK**

#### Origin of the COVID-19 outbreak

SARS-CoV-2 is a β-coronavirus and is enveloped with non-segmented Orthocoronavirinae subfamily RNA[4]. Among the four genera,  $\gamma$ - and  $\delta$ -CoV infect birds while  $\alpha$ - and  $\beta$ -CoV infect mammals including humans (Table 1). The  $\alpha$ - and  $\beta$ -CoV have six



Table 1 De	tails of coronavirus (genus, species and receptor)		
Genus	Species	Targets	Receptor
a-CoV	Alphacoronavirus 1:	Mammals	
	Feline coronavirus serotype 2		Aminopeptidase N
	Canine coronavirus serotype 2		Aminopeptidase N
	Transmissible gastroenteritis virus		Aminopeptidase N
	Human coronavirus 229E		Aminopeptidase N
	Human coronavirus NL63		ACE2
	Porcine epidemic diarrhea coronavirus		Aminopeptidase N
	Rhinolophus bat coronavirus HKU2		
	Scotophilus bat coronavirus 512/05		
	Miniopterus bat coronavirus 1		
	Miniopterus bat coronavirus HKU8		
β-CoV	Betacoronavirus 1:	Mammals	
	Bovine coronavirus		Neu 5,9 Ac2
	Human coronavirus OC43		Neu 5,9 Ac2
	Equine coronavirus		
	Human enteric coronavirus		
	Porcine haemagglutinating encephalomyelitis virus		
	Canine respiratory coronavirus		
	Murine coronavirus:		
	Mouse hepatitis virus		CEACAM1
	Rat coronavirus		
	Puffinosis virus		
	Hedgehog coronavirus 1		
	Human coronavirus HKU1		
	Middle East respiratory syndrome-related coronavirus		
	Pipistrellus bat coronavirus HKU5		
	Rousettus bat coronavirus HKU9		
	Severe acute respiratory syndrome-related coronavirus		
	SARS-CoV		
	SARS-CoV-2		ACE2
	Rhinolophus bat viruses		
	Tylonycteris bat coronavirus HKU4		
γ-CoV	Avian coronavirus:	Birds	
	IBV (turkey, pheasant, duck, goose and pigeon)		
	Beluga Whale coronavirus SW1		
δ-CoV	Bulbul coronavirus HKU11	Birds	
	Thrush coronavirus HKU12		
	Munia coronavirus HKU13		
	Porcine coronavirus HKU15		

ACE2: Angiotensin-converting enzyme 2; SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2.

variants. Among them  $\alpha\text{-}CoVs$  variants (HCoV-229E and HCoV-NL63), and  $\beta$ 



Jaisbideng® WJG | https://www.wjgnet.com

5824

September 21, 2021 Volume 27 Issue 35

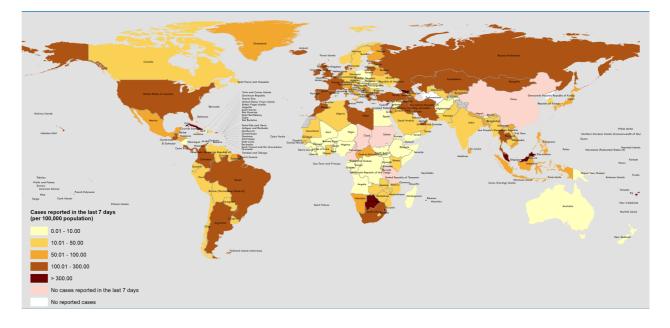


Figure 1 Geographical distribution of coronavirus disease 2019 outbreaks. Source: https://www.who.int/publications/m/item/weekly-epidemiologicalupdate-on-covid-19---10-august-2021 (data as reported at 4:58 pm CET on August 10, 2021).

-CoVs variants (HCoV-HKU1 and HCoV-OC43) have lower pathogenic capability in humans and cause mild respiratory symptoms similar to the common cold. Only  $\beta$ -CoVs variants (SARS-CoV and MERS-CoV) have severe pathogenic capability in humans. This pandemic started in Wuhan specifically in a seafood wet market, on December 12, 2019. Several studies have demonstrated that bats are natural hosts of SARS-CoV-2 and animals such as snakes, turtles and pangolins are intermediate hosts of SARS-CoV-2.

Previously, snakes were thought to be involved in COVID-19 outbreaks by Ji *et al*[5] but this hypothesis was rejected by Zhang *et al*[6] who did not find any similarity in genome sequence between snakes and COVID-19 patients. In another study, researchers found an approximately 96.2% genome sequence similarity between SARS-CoV-2 and bat coronavirus (CoV RaTG13)[7]. In addition, the genomic sequence of SARS-CoV-2 matched with 79.5% of the genome sequence of SARS-CoV[8]. These findings implied that bats were the suspected source of COVID-19 outbreaks as well as the natural host of this virus. The virus was finally transmitted to humans via unknown intermediate hosts from bats. However, few bats are sold in the Wuhan seafood market[9]. Accordingly, scientists are trying to determine the intermediate sources such as snakes, turtles and pangolins. Xu et al[10] found approximately 99% genomic similarity between SARS-CoV-2 and pangolins. Furthermore, they revealed that pangolins are the potential intermediate host of SARS-CoV-2. Apart from these studies, to date there is no adequate evidence on the virus origin regarding potential intermediate hosts and the natural host of SARS-CoV-2. Therefore, SARS-CoV-2 could use angiotensin-converting enzyme 2 (ACE2), similar to SARS-CoV receptor for human infection[7]. However, there is controversy regarding the infectious potential of COVID-19 patients to transmit the disease during the incubation period. Recently, the WHO reported that cats may be the carrier of this virus, whereas other domestic animals like ducks, hens and dogs may not be carriers of this deadly virus.

#### Transmission of COVID-19

The animal-human interface is not a new concept. Zoonotic diseases with a wildlife reservoir have long been recognized as significant public health problems. Indeed, up to three-quarters of infectious diseases that cause human infections are known to be zoonotic[11]. Apart from this, the complexity of animal, human, and environmental factors is thought to play a critical role in its emergence<sup>[12]</sup>. On the other hand, contact with infected patients and droplets are considered to be major transmission routes of COVID-19. Aerosol transmission is another important route of SARS-CoV-2 infection. By contrast, SARS-CoV and MERS-CoV transmission are mainly reported through nosocomial transmission. However, human-to-human SARS-CoV-2 transmission occurs mainly through close contact between COVID-19 patients or friends or carriers and between family members including relatives. It can be spread rapidly in



healthcare workers (up to 50%) and patients (62-79%) similar to SARS-CoV and MERS-CoV and is considered the most common route of infection[13]. It is also assumed that consumption of wild animals who are the hosts of SARS-CoV-2 and humans in close contact with these animals are suspected to be the route of entry of SARS-CoV-2 and its mode of transmission. However, this route of SARS-CoV-2 transmission remains controversial and requires further study.

To date, 1 million people around the world have tested positive for this virus, but only 4 cases have so far been reported in which pets showed positive for SARS-CoV-2. These involved 2 dogs and 2 cats, the owners had COVID-19 and are believed to be the most likely source of transmission to their pets. The dogs showed clinical signs, but one of the cats did not have signs of illness. In late March 2020, health officials in Belgium reported that a cat from Liège province had also tested positive for SARS-CoV-2. Nevertheless, the US Centers for Disease Control and Prevention (CDC), WHO, and key animal health organizations have all issued statements aiming to calm people's fears about their pets being a source of the novel virus [14-16]. In this regard, the World Organization for Animal Health has emphasized that "there is no justification in taking measures against companion animals which may compromise their welfare". Furthermore, given the speculation that wild live animal species may be linked to this pandemic, this collaborative approach will also require the expertise of wildlife forensic specialists.

SARS-CoV-2 has also been detected in saliva, the gastrointestinal tract, urine and stool. In particular, the gastrointestinal tract or digestive tract has been recognized as another route of SARS-CoV-2 infection based on a bioinformatics study[17]. SARS-CoV-2 has been detected in gastrointestinal mucosal tissue of COVID-19 patients[18]. In addition, it has also been detected in tears and conjunctival secretions of COVID-19 patients<sup>[19]</sup>. Intrauterine vertical transmission from pregnant women to the newborn is temporarily excluded due to a lack of adequate data on pregnant women infected with SARS-CoV-2[20].

#### Prevalence of COVID-19

A number of researchers estimated the basic reproduction number  $(R_0)$  to calculate the number of people affected by secondary infections. Generally, it represents the number of people with COVID-19 but in a completely susceptible population without intervention[21]. Using the SEIR model, Wu et al[22] recorded an R<sub>0</sub> value for SARS-CoV-2 in the range of 2.47-2.86, while Majumder and Kenneth [23] estimated the  $R_0$ value to be 2.0-3.3 based on the IDEA model. By contrast, other β-CoV viruses namely SARS-CoV and MERS-CoV showed an R<sub>0</sub> value in the range of 2.2-3.6 and 2.0-6.7, respectively<sup>[24,25]</sup>, which indicated that SARS-CoV-2 has higher transmissibility than SARS-CoV and MERS-CoV. In China, 87% of cases were in the age group 30 to 79 years and 3% cases were noted to be aged  $\geq$  80 years, while female cases were only 41.9% [26,27]. Additionally, 81% of cases were classified as mild, 14% cases were severe and 5% cases were very critical. In another study, it was reported that the overall casefatality rate (CFR) was 2.3%; however, in the age groups 70-79 and  $\geq$ 80 years, the CFRs were 8.0% and 14.8%, respectively<sup>[22]</sup>. These findings clearly indicated that elderly males are more susceptible to SARS-CoV-2 compared with other groups. In addition, the virus affected those elderly males with chronic diseases such as diabetes, hypertension, heart disease, etc.[20]. In summary, the prevalence of COVID-19 is very high, and it can spread very rapidly within countries and outside countries.

#### Virus susceptibility and incubation period

Generally, elderly people aged between 55 and 75 years are more susceptible to SARS-CoV-2 infection. Currently, it has been found that the virus is also infecting middleaged people aged between 25 and 50 years. The average age of patients across 18 studies was 51.97 years (95%CI: 46.06%-57.89%), 55.9% were male (95%CI: 51.6%-60.1%). Additionally, 36.8% cases showed comorbidities (95%CI: 24.7%-48.9%), the most significant being hypertension (18.6%; 95%CI, 8.1-29.0%), cardiovascular disease (14.4%; 95%CI: 5.7%-23.1%), and diabetes (11.9%; 95%CI: 9.1%-14.6%), among others[28]. Children account for 1% to 3% of COVID-19 cases across countries and likely experience an asymptomatic infection (mild or no symptoms on infection) compared with adults. Zhong et al[29] demonstrated that the virus has an average median incubation period of about 3 d but it can range between 0 and 24 d, and the average median time from symptomatic onset to death is 14 d. They also found that mortality rises in patients with comorbidities or a surgical history before virus infection. Generally, the average median latency period for SARS-CoV-2 infection was 4 d, the average interval to hospital admission after onset of symptoms was 3.8 d, and the average time to death after admission to hospital was 17.4 d[30]. Another study



reported that the time to appearance of COVID-19 symptoms to death ranged between 6 and 41 d with a median period 14 d[22]. They also showed that this period was agedependent and related to the patient's immune system status. The prevalence was greater in patients aged over 70 years compared with those less than 70 years. According to the WHO, the incubation period for COVID-19 ranged from 2 to 10 d. By contrast, for MERS-CoV infection the average median latency was 7 d[31]. However, in COVID-19, the maximum latency was observed to be 24 d, which was high compared with SARS and MERS. This indicated that SARS-CoV-2 has a higher risk of transmission. Accordingly, in comparison with SARS and MERS, SARS-CoV-2 has a shorter median incubation period. Recent data showed that elderly people (aged above 75 years) have a shorter median interval, i.e., 11.5 d from symptom onset to death in comparison to COVID-19 patients (20 d). This finding indicated that disease progression is more rapid in elderly people compared to younger people[1].

# GENOMIC STRUCTURE AND PATHOPHYSIOLOGY

#### Genomic structure

SARS-CoV-2, a  $\beta$ -coronavirus, is a single-stranded RNA virus with a diameter ranging between 80 nm and 120 nm. Currently, four types of coronavirus are present in nature:  $\alpha$ -,  $\beta$ -,  $\delta$ - and  $\gamma$ - coronavirus. The  $\gamma$ - and  $\delta$ -CoV infect birds, while  $\alpha$ - and  $\beta$ -CoV infect mammals. Details of these coronaviruses are presented in Table 1. There are six coronaviruses causing human infection including SARS-CoV and MERS-CoV. The complete genome sequence of SARS-CoV-2 is closest to SARS-like bat CoV (MG772933). There is approximately 79% homology in genome sequence between SARS-CoV-2 and SARS[9]. In addition, the complete genomic sequence of SARS-CoV-2 is approximately 29.9 kb, while SARS-CoV and MERS-CoV have a genome length of 27.9 kb and 30.1 kb, respectively [8,32]. The SARS-CoV-2 genome contains a variable number of open reading frames (ORFs) ranging between 6 and 11[33]. Two-thirds are located mainly in the first ORF (ORF1a/b) which encodes 16 non-structural proteins (NSP) and translates polyproteins (pp1a and pp1ab), while the remaining ORFs encode accessory and structural proteins. The remainder of the RNA virus encodes four essential structural proteins, including the spike (S) glycoprotein, small envelope (E) protein, matrix (M) protein, and nucleocapsid (N) protein, and several accessory proteins, that interfere with the host innate immune response[34]. Frameshift mutation between ORF1a and ORF1b is mainly responsible for the production of pp1a and pp1ab polypeptides that are regulated by chymotrypsin-like protease (3CLpro) or main protease (Mpro), and this process produces 16 non-structural proteins (NSPs) with the help of papain-like proteases[35]. Therefore, SARS-CoV-2 pathophysiology and virulence are thought to be linked with NSPs and structural protein functions.

#### Pathophysiology

The pathophysiology of COVID-19 produces pneumonia which seems to be very complex. The pathological mechanism is presented in Figure 2. A group of researchers claimed that viral infection is caused by an immune reaction through the "cytokine storm" [36,37]. The main protagonist of this "cytokine storm" is interleukin 6 (IL-6). Generally, activated leukocytes are primarily responsible for IL-6 production and IL-6 acts on a number of cells and tissues. It stimulates acute phase protein production and regulates thermoregulation, bone structure and central nervous system functions[36, 37]. However, its main role is pro-inflammatory actions. COVID-19 enhances IL-6 level, which is implicated in the pathogenesis of the cytokine release syndrome (CRS), which is an acute systemic inflammatory syndrome characterized by fever and multiple organ dysfunction[36,37].

Another group of researchers demonstrated that SARS-CoV-2 uses angiotensin converting enzyme 2 (ACE2) receptor for both cross-species and human-to-human transmission[1,38]. The virion S-glycoprotein present on the virus surface interacts with ACE2 receptors on human cells to spread the infection[39]. S-glycoprotein contains two subunits, S1 and S2. The S1 determines the virus-host range and cellular tropism in the key function domain - RBD (receptor-binding domain), while S2 is responsible for cell membrane-virus fusion by two tandem domains, heptad repeats 1 (HR1) and HR2[40,41]. Following membrane fusion, viral RNA is released into the cytoplasm, and the uncoated RNA is induced to produce pp1a and pp1ab polypeptides with the help of either chymotrypsin-like protease (3CLpro) or main protease (Mpro), which encode 16 non-structural proteins (NSPs) in the presence of papain-like proteases, and finally form a replication-transcription complex (RTC) in double-



Samanta P et al. Environmental aspects of COVID-19 outbreaks

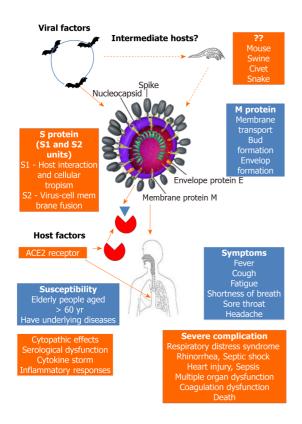


Figure 2 Pathogenesis of severe acute respiratory syndrome coronavirus 2 (viral and host factors). ACE2: Angiotensin-converting enzyme 2.

membrane vesicles<sup>[8]</sup>. Subsequently, the RTC replicates continuously and synthesizes sub-genomic RNAs[42] to encode accessory proteins and structural proteins. This newly formed genomic RNA, envelopes glycoproteins and nucleocapsid proteins mediated through the endoplasmic reticulum (ER) and Golgi<sup>[43]</sup> are assembled together to form viral buds. Finally, these newly formed virion-containing vesicles are fused with plasma membrane to release the virus and cause infection through mucous membranes, especially nasal and larynx mucosa, and then enter the lungs through the respiratory tract.

These ACE2 receptors are very important in the spread of COVID-19. They are mainly found in the lower respiratory tract of humans. After entry through mucous membranes, especially nasal and larynx mucosa, the virus enters directly into the lungs through the respiratory tract. In the next step, the virus attacks other target organs which contain ACE2 receptors, such as the lungs, heart, renal system and gastrointestinal tract[36,37]. Accordingly, the binding affinity of this virus-receptor has been intensively studied using different approaches. Systematic detection analysis showed that SARS-CoV-2 S-glycoprotein binding capacity with ACE2 was 10-fold higher than SARS-CoV as shown under cryo-electron microscopy of the SARS-CoV-2S protein in pre-fusion conformation[39]. Recently, Wu et al[9] demonstrated moderate genomic and phylogenetic similarity with SARS-CoV but higher similarity with bat CoV genome sequence, particularly in the S-glycoprotein and RBD. They also found that there were no amino acid substitutions occurring in the NSP7, NSP13, envelope, matrix, or accessory proteins p6 and 8b at the protein level, except in NSP2, NSP3, spike protein, underpinning the subdomain, i.e., RBD. Another recent study demonstrated that mutation of NSP2 and NSP3 plays an important role in infection and SARS-CoV-2 differentiation. However, this mechanism of SARS-CoV-2 infection in humans via S-protein binding with ACE2 is unclear, as is the interaction strength for risk transmission. Accordingly, the WHO was also unable to clarify the mechanism of COVID-19. This has led to further investigations regarding potential human-to-human transmission and the pathophysiological mechanisms of COVID-19 outbreaks.

# CLINICAL CHARACTERISTICS OF COVID-19 INFECTION

Being an acute respiratory infection, COVID-19 is initiated in the respiratory tract, primarily by droplets, respiratory secretions, and direct contact. After entry, the virus

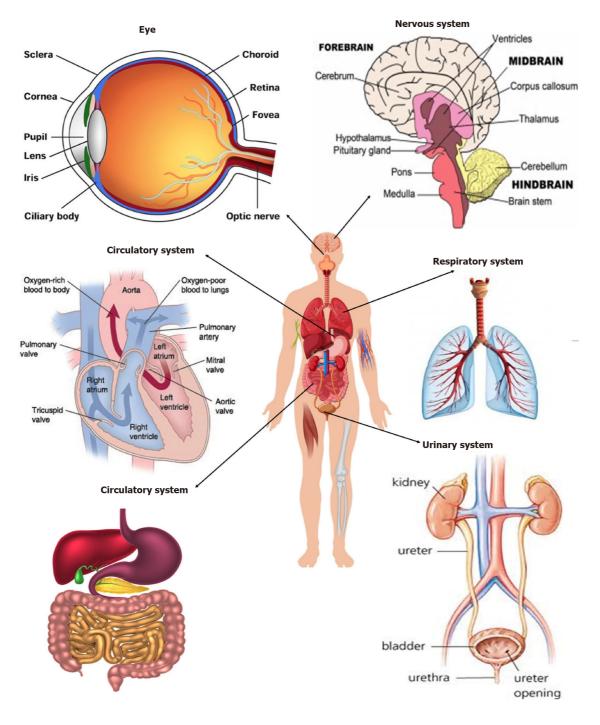


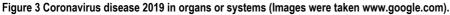
affects a number of organs or systems (Figure 3). The clinical symptoms of COVID-19 vary from asymptomatic or paucisymptomatic forms to clinical conditions. In particular, all patients are divided into general, severe, and critical patient groups. The most common clinical symptoms of COVID-19 are fever (87.9%), cough (67.7%), fatigue (38.1%), sputum production (33.4%), shortness of breath (18.6%), sore throat (13.9%), and headache (13.6%)[27,44]. The development of these symptoms may occur within 3 d of viral infection. On the other hand, other symptoms may occur 9 d after virus infection. Of these, fever and cough are the dominant COVID-19 symptoms. The incidence of diarrhea (3.7%) and vomiting (5.0%) is very rare [27,44]. However, it is very difficult to accurately distinguish COVID-19 from other viral respiratory infections. The CDC included loss of taste or smell, pink eye, muscle pain, intense chills, headache and sore throat as COVID symptoms. In severe cases, symptoms such as acute respiratory distress syndrome, rhinorrhea, dyspnea, gastrointestinal disorders, septic shock, mental stress, acute heart injury, sepsis, multiple organ dysfunction syndrome (MODS), secondary infection and even death may occur[8,34]. Critical COVID-19 patients with severe respiratory failure require an intensive care unit (ICU) or ventilation support. However, the occurrence of upper respiratory symptoms and gastrointestinal symptoms are very rare compared with other symptoms. In addition to this, the elderly and those who have underlying diseases ( i.e., chronic obstructive pulmonary disease, hypertension, diabetes, cardiovascular disease) are very prone to COVID-19 and develop symptoms such as metabolic acidosis, acute respiratory distress syndrome, coagulation dysfunction and even death [8,45]. Sometimes, COVID-19 patients experience acute heart injury, arrhythmia, impaired renal function and abnormal liver function such as the formation of microvesicular steatosis (50.7%) at the time of admission[1,45,46].

Hematological assays revealed that most patients had decreased white blood cell counts, and lymphocytopenia[27]. In the case of critical patients, neutrophil count, Ddimer, blood urea, creatinine and lymphocyte levels decreased markedly. In another study, a reduction in albumin level (75.8%; 95%CI, 30.5%-100.0%), higher C-reactive protein (58.3%; 95%CI: 21.8%-94.7%) and lactate dehydrogenase (LDH) levels (57.0%; 95%CI: 38.0%-76.0%), higher lymphopenia level (43.1%; 95%CI, 18.9%-67.3%), and higher erythrocyte sedimentation rate (ESR) (41.8%; 95%CI: 0.0-92.8%) and other clinical manifestations were recorded [28]. Additionally, inflammatory factors, which indicated the immune status of patients, namely IL-6, IL-10, and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) are also markedly increased. In critical patients (admitted to the ICU), higher IL-2, IL-7, IL-10, granulocyte colony-stimulating factor (GCSF), 10 kD interferon gamma-induced protein (IP-10), monocyte chemoattractant protein-1 (MCP-1), macrophage inflammatory protein 1- $\alpha$  (MIP-1 $\alpha$ ), and TNF- $\alpha$  levels in plasma were observed[8,45]. In patients with severe COVID-19 [admitted to the ICU; 20.3% cases (95%CI, 10.0-30.6%)], 32.8% of patients experienced ARDS (95%CI: 13.7%-51.8%), 13.0% patients had acute cardiac injury (95%CI: 4.1%-21.9%), 7.9% patients experienced acute kidney injury (95%CI: 1.8-14.0%), 6.2% cases (95%CI: 3.1%-9.3%) developed shock and 13.9% cases (95%CI 6.2%-21.5%) experienced fatal outcomes [28]. Furthermore, 96.8% of all patients (95%CI: 94.9%-98.7%) had RNAemia in blood and nasopharyngeal aspirates (NPA)[28].

#### IMMUNOPATHOLOGICAL RESPONSES

Immunological symptoms are generally caused due to binding of virus S proteins with ACE2 at the receptor, usually in the endosome Toll-like receptor (TLR) 3, TLR7, TLR8, and TLR9[8,47]. Retinoic-acid inducible gene I (RIG-I) of the virus, melanoma differentiation-associated gene 5 (MDA5) of the cytosol and nucleotidyltransferase cyclic GMP-AMP synthase (cGAS) are generally responsible for the spread of COVID-19[8, 48,49]. Viral infection activates nuclear factor-κB (NF-κB) and interferon regulatory factor 3 (IRF3) to produce type I interferons (IFN- $\alpha/\beta$ ) and pro-inflammatory cytokines as immune mediators (i.e., innate immunity) to prevent infection[8,50]. As a result, the plasma levels of some cytokines and chemokines are elevated in COVID-19 patients such as IL-1, IL-2, IL-4, IL-7, IL-10, IL-12, IL-13, IL-17, GCSF, macrophage colonystimulating factor (MCSF), IP-10, MCP-1, MIP-1a, hepatocyte growth factor (HGF), IFN- $\gamma$  and TNF- $\alpha$ [20,45,51]. Generally, these inflammatory responses were noted in the lower airway and lung[52]. Consequently, these trigger immune signaling and produce the "cytokine storm' within the body leading to a very critical condition in COVID-19 patients.





# **DIAGNOSIS OF COVID-19**

Since the outbreak of COVID-19, a number of diagnostic tools have been used to detect the infection. The classical Koch's postulates method was used to detect the infection in Wuhan<sup>[22]</sup>. This method is very expensive and time-consuming as it uses electron microscopy. In some countries, radiography was used to detect the viral infection such as a chest computed tomography (CT) scan. CT scan is an important tool in diagnosing COVID-19 pneumonia. Typical COVID-19 pneumonia features were observed by CT. and CT imaging showed ground-glass opacities (56.4%-65%), an air bronchogram (47%), bilateral patchy shadowing (51.8%), consolidations (50%), smooth or irregular interlobular septal thickening (35%), thickening of adjacent pleura (32%), sometimes rounded morphology, peripheral and lower lobe involvement and a peripheral lung distribution in COVID patients[27,53,54]. A very recent study recorded bilateral chest CT findings in 90% patients, and proved its sensitivity (97%) in detecting COVID-19 [55]. However, in another study clinical scientists found that some patients with



confirmed COVID-19 had normal CT scans[53]. Therefore, the diagnosis of COVID-19 is very confusing. Moreover, this technique mainly determines pneumonia. Accordingly, scientists are looking for an alternative method which is more reliable and confirmative. The detection of viral nucleic acid from nasal and throat swab samples, cough, sputum or other respiratory tract samples is the golden diagnostic method for COVID-19 detection. This method uses RT-PCR technology to detect viral infection. Although, this method has high specificity, false-negative results may occur due to low sensitivity and the testing time is too long. In the case of false-positive tests, the WHO recommends resampling and further testing. In this regard, serologic testing is an important diagnostic tool to detect patients who have either current or previous infection but have a negative PCR test[56,57]. In this technique, basic parameters are tested to detect the COVID-19, namely white blood cell count, neutrophil and lymphocyte count, D-dimer, blood urea, and creatinine estimation to identify the appearance of leukopenia, leukocytosis, and lymphopenia as COVID-19 symptoms [58, 59]. In another study, it was demonstrated that 82.1% of COVID patients are lymphopenic, 33.7% patients are leukopenic and 36.2% patients are thrombocytopenic [1]. In addition, another group of researchers recommended elevated plasma levels of C-reactive protein, lactate dehydrogenase, creatinine kinase, transaminase, abnormal myocardial enzyme spectrum or creatinine as COVID-19 indicators[27,45]. They also showed that cytokine release syndrome is an important vital indicator of disease progression. On the other hand, Wan et al[60] demonstrated higher IL-6 and IL-10 levels, and lower CD4+T and CD8+T levels as indicators of COVID-19.

Currently, a number of technological inventions are ongoing to detect COVID-19 in a simplistic pathway. Different technological inventions such as the more organized sequencing library (SHERRY) in China, SHERLOCK technology in China, FELUDA in India *etc.*, have been developed as testing tools for rapid detection of COVID-19[6,61]. However, clinical verification of these technological inventions has not been undertaken to date, and once approved, they will be a major breakthrough in technology to diagnose COVID-19 rapidly and economically.

#### GLOBAL SCENARIOS OF COVID-19 OUTBREAKS

Since its outbreak in Wuhan, China in late December 2019, SARS-CoV-2 infection is spreading very rapidly across the globe. COVID-19 has affected 202608306 people and caused around 4293591 deaths (Table 2). The inter-continental spread is described in Table 2. Figure 4 shows COVID-19 outbreaks in different countries. In the beginning, the Asian countries namely China and South Korea were the epicenter of COVID-19 until the first week of February. Up to August 10, 2021, there have been 93826 confirmed cases and 4636 deaths in China (WHO). In Korea the first COVID case was recorded on January 20, 2020. Since then, about 212448 cases have been confirmed and 2125 deaths recorded in Korea. The epicenter then moved from Asian countries to European countries mainly Italy and Spain. COVID-19 was recorded in Italy on January 30, 2020, and was found in France and Spain on January 24, 2020 and January 31, 2020, respectively. In particular, in Italy, the United Kingdom, France, Germany and Spain it affected people more seriously; approximately 4400617, 6094243, 6310933, 3800048, and 4627770 confirmed cases and 128242, 130357, 112288, 92291, and 82125 deaths were recorded in these countries, respectively, up to August 10, 2021. Among the European countries, mortality rate was highest in Italy due to its travel connection with China. In the middle of March, the virus epicenter moved to the United States and other American countries. The United States and Canada were the most affected countries during this phase. Although the first COVID-19 patient was recorded in late January, 2020 the first death was confirmed in February. In the USA, the first COVID-19 patient died in the middle of March. On August 10, 2021, the USA had recorded the greatest number of confirmed cases and deaths worldwide. The death rate is 206 per million people, which is the tenth highest rate globally. The first COVID-19 patient in Canada was reported on January 27, 2020. On August 10, 2021 there have been 36780480 and 1442087 confirmed cases in the USA and Canada, respectively, and 633799 and 26678 deaths, respectively. In the middle of April, the virus epicenter moved to Russia and India. As of August 10, 2021, there have been 6469910 and 31997017 confirmed cases in Russia and India, respectively, and the number of deaths is 165650 and 428715, respectively. However, the first confirmed COVID-19 case was recorded on January 30, 2020 in Kerala state and January 31, 2020 in Russia. The virus infection in these countries took a very long time to spread due to the implementation of different control measures. The details of COVID-19 cases in India are presented in







Figure 4 Coronavirus disease 2019 routes of transmission across countries. Figure modified after Ali and Alharbi (2020)[68], an Elsevier journal.

Table 3. However, according to fatality rate data, Belgium (15% fatality) is highest, followed by the United Kingdom (15%), France (14.7%), Italy (13.6%) and the Netherlands (12.3%) (John Hopkins Bulletin).

# **TREATMENT OF COVID-19**

#### Antiviral drug treatment

Presently, COVID-19 treatment is based on symptomatic findings. To date, there is no precise treatment method, but currently the WHO, CDC and Food and Drug Administration have recommended certain drugs for COVID-19 treatment. The effectiveness

Table 3 Coronavirus disease 2019 state-wise status in India (as on August 10, 2021; Ministry of Home Affairs, Gol)				
No.	Name of State / UT	Total confirmed cases*	Cured/discharged/migrated	Deaths**
1	Andaman and Nicobar Islands	7546	7412	129
2	Andhra Pradesh		1950623	13549
3	Arunachal Pradesh	50372	47520	246
4	Assam		558720	5404
5	Bihar		715303	9646
6	Chandigarh	61984	61146	811
7	Chhattisgarh		988004	13540
8	Dadar Nagar Haveli	10656	10612	4
9	Delhi		1411235	25067
10	Goa		167884	3164
11	Gujarat		814778	10077
12	Haryana		759769	9650
13	Himachal Pradesh		202569	3519
14	Jammu and Kashmir		316957	4390
15	Jharkhand		342074	5130
16	Karnataka		2859552	36817
17	Kerala		3377691	17852
18	Ladakh	20393	20117	207
19	Madhya Pradesh		781307	10514
20	Maharashtra		6151956	134064
21	Manipur		96128	1657
22	Meghalaya	69358	63450	1174
23	Mizoram	44520	32854	168
24	Odisha		971391	6554
25	Puducherry		119031	1800
26	Punjab		582753	16320
27	Rajasthan		944670	8954
28	Tamil Nadu		2522470	34340
29	Telengana		637789	3828
30	Tripura	80208	77230	767
31	Uttarakhand		328569	7368
32	Uttar Pradesh		1685449	22774
33	West Bengal		1505808	18240
34	Nagaland	28709	25906	585
35	Sikkim	27908	24544	355
36	Lakshadweep	10257	10112	51

and limitations of each drug are summarized in Table 4[62]. The existing drugs for treating COVID-19 patients are remdesivir, chloroquine, hydroxychloroquine, tocilizumab, lopinavir-ritonavir, azithromycin, baloxavir, favipiravir, etc.[63]. Remdesivir, is most prominent for treating COVID-19 patients[64]. The efficacy of remdesivir in treating patients has been reported globally[63-65]. Recently, the ChAdOx1 vaccine developed by the University of Oxford's Jenner Institute and the Oxford Vaccine Group has proved effective in combatting COVID-19. More recently,



Common drugs	Dose	Mechanism
Chloroquine; Antimalarial	50% for GFR < 10 mL/min	In vitro activity and has immunomodulating properties
· · · · ·		Inhibits viral enzymes or processes such as viral DNA and RNA polymerase, viral protein glycosylation, virus assembly, new virus particle transport, and virus release
		ACE2 inhibition due to acidification at cell membrane surface, inhibits fusion of virus, and cytokine release
<b>Hydroxychloroquine</b> ; Antimalarial	800 mg orally on day one, followed by 400 mg/d orally for four to seven days	Same as chloroquine
<b>Chloroquine phosphate</b> ; Antimalarial	1 g orally on day one, followed by 500 mg/d orally for four to seven days	Same as chloroquine
Remdesivir; Nucleoside	200 mg IV on day 1 followed by 100 mg IV daily on days two to five or 200 mg IV on day 1 followed by 100 mg IV daily on days two to ten	In vitro activity; Inhibitor of RNA-dependent RNA polymerases (RdRps)
Analogue		Remdesivir-TP competes with adenosine-triphosphate for incorporation into nascent viral RNA chains
		Once incorporated into the viral RNA at position i, RDV-TP terminates RNA synthesis at position i+3
		Because RDV-TP does not cause immediate chain termination (i.e., 3 additional nucleotides are incorporated after RDV-TP), the drug appears to evade proofreading by viral exoribonuclease (an enzyme thought to excise nucleotide analogue inhibitors)
<b>Azithromycin</b> ; Macrolide Antibacterial	500 mg on day one, followed by 250 mg daily for four days	Prevents bacterial superinfection, has immunomodulatory action on pulmonary inflammatory disorders
		Downregulates inflammatory responses and reduces excessive cytokine production associated with respiratory viral infections; however, its direct effects on viral clearance are uncertain
		Immunomodulatory mechanisms include reducing chemotaxis of neutrophils (PMNs) to lungs by inhibiting cytokines ( <i>i.e.</i> , IL-8), inhibition of mucus hypersecretion, decreased production of ROS, accelerating neutrophil apoptosis, blocking activation of nuclear transcription factors
<b>Lopinavir; Ritonavir</b> ; HIV protease inhibitor	400 mg/ritonavir 100 mg orally twice daily for up to 21 d	<i>In vitro</i> animal model studies show potential activity for other coronaviruses (SARS-CoV and MERS-CoV)
		Lopinavir and ritonavir may bind to Mpro, a key enzyme for virus replication and suppress virus activity
Tocilizumab; Interleukin-6 (IL- 6) Receptor- InhibitingMonoclonal Antibody	4-8 mg/kg infused over more than 60 min (additional dose after 12 h)	Cytokine release syndrome; Inhibits IL-6-mediated signaling by competitively binding to both soluble and membrane-bound IL-6 receptors. IL-6 involved in T- cell activation, immunoglobulin secretion induction, hepatic acute-phase protein synthesis initiation, and hematopoietic precursor cell proliferation and differentiation stimulation
Baloxavir; Antiviral	80 mg orally on day 1 and on day 4, and another dose of 80 mg on day 7 (as needed); not to exceed 3 total doses	Active against influenza viruses; <i>In vitro</i> antiviral activity against SARS-CoV-2 demonstrated in one trial
Favipiravir; Antiviral	1600 mg twice daily on day 1, then 600 mg twice daily for 7-10 d; Severe: 1600 mg every 12 h on day 1, then 600 mg every 12 h days 2-10	In vitro activity against Vero E6 cells

SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2.

Russia has reportedly developed a coronavirus vaccine named Sputnik V.

#### Chinese medicine treatment

A number of Chinese medicines have been used to treat COVID-19 patients. According to the Academy of Sciences, Shuanghuanglian oral liquid is most prominent and inhibits SARS-CoV-2. Several studies reported that baicalin, chlorogenic acid and forsythin present in Shuanghuanglian oral liquid have certain inhibitory effects on various viruses and bacteria including SARS-CoV-2[66]; however, the detailed mechanism is not yet known. Lianhuaqingwen capsules have also been used to treat



Baishideng® WJG | https://www.wjgnet.com

SARS-CoV-2 infected people as well as other diseases such as influenza viruses, including H7N9 by reducing inflammatory factors[1,17].

#### Unani medicine treatment

These are plant-based treatments, called Ayurvedic treatments, and these treatments are nontoxic and have no side effects. Different plant parts are used to treat anti-viral activities[67]. The most important plants are Glycyrrhiza glabra, Allium cepa, Allium sativum, Ocimum sanctum, Ocimum tenuiflorum, Piper nigrum, Cinnamomum verum, Daucus maritimus, Curcuma longa, etc. Administration of the aqueous extracts of these plants along with lemon juice and honey is very effective for flu and the common cold [68]. According to Fiore *et al*[69] *Glycyrrhiza glabra* plant extract is effective in treating viruses such as SARS related coronavirus, HIV-1, respiratory syncytial virus, varicella zoster, hepatitis A, B, C, and cytomegalovirus herpes. Similarly, Wang et al [70] indicated that Glycyrrhiza glabra also has antiviral and antimicrobial activities. Therefore, Glycyrrhiza glabra plant extract along with other plants may be useful in controlling COVID-19. Accordingly, the Government of India has recommended Ayurveda treatment methods to improve immunity (Table 5).

#### Homeopathic treatment

Arsenic album-30 is considered beneficial for viral infections. Recently, the Directorate of AYUSH, New Delhi, India has issued an order on January 30, 2020 to take prophylactic medicine to avoid coronavirus infection. Dr Rajan Sankaran has recommended Camphor 1M as a potential medicine for COVID-19 (https://www.boomlive.in/ coronavirus-outbreak/homoeopathy-can-be-used-as-adjuvant-to-covid-19-treatmentdr-anil-khurana-7997). They recommended 4 pills of Arsenic album-30 medicine once daily on an empty stomach for 3 d. It is highly diluted arsenic trioxide and works as a homeopathic prophylaxis. Accordingly, the Homeopathy Department of Kerala Government is administering Arsenicum Album 30C as a preventive medicine to boost immunity in COVID-19 patients and it was approved by the Department of AYUSH, GoI (https://gulfnews.com/world/asia/india/covid-19-kerala-governmentdistributes-homeopathy-medicine-to-boost-immunity-1.1588091249686). However, to date, there is no clinical evidence that Arsenic album-30 is an effective medicine. As a result, the use of these medicines to manage COVID-19 has been criticized globally. Mathie *et al*<sup>[71]</sup> reported that *Arsenicum album* medicine is effective in reducing fever, runny nose, headache, and sore throat in patients with swine flu. Therefore, the use of homeopathy in COVID-19 management is debatable and requires further scientific study.

#### Immuno-booster treatment

Boosting the body's immunity is a potential individual protocol as COVID-19 pathogenesis is caused by a disproportionate immune response. Therefore, it is important to take supplements to boost both innate and adaptive immune response. Interferon is reported to inhibit viral infection and in particular, recombinant interferon  $\alpha$  is effective for SARS-like viruses. Additionally, interferon was reported to be an effective inhibitor of MERS-CoV replication<sup>[72]</sup>. These findings indicated that interferon could be used to treat COVID-19 infection. Intravenous immunoglobulin might be the safest immune modulator for all age groups, and could help to inhibit pro-inflammatory cytokine production and to increase anti-inflammatory mediators [1, 73]. Moreover, thymosin alpha-1 (Ta1) is used as an immune booster for SARS patients to effective control the disease [74]. Accordingly, intravenous immunoglobulin and Ta1 may also be used for the treatment of COVID-19. Recently, different immune-booster drugs have been used to treat COVID-19 such as neuraminidase inhibitors (e.g., oseltamivir used to treat influenza). Apart from these, citrus fruits, dry fruits (almonds, walnuts, and dates) are very effective in improving the immune system. Vitamin A, C, D and E, and zinc supplements are effective in older patients. Additionally, adequate sleep, regular exercise and stress avoidance is essential to boost the immune system[68].

#### Plasma therapy

Due to lack of appropriate vaccines and specific drugs, plasma therapy could be an effective way to treat COVID-19. Previously, convalescent plasma therapy was proved to be an effective treatment option for SARS patients and those with H1N1 influenza [75,76]. From an immunological perspective, it was observed that recovered COVID-19 patients produced specific antibodies against SARS-CoV-2, and therefore their serum could be used to prevent re-infection. Additionally, these antibodies can limit the



Table 5 Unani drugs for coronavirus disease 2019 treatment (Source: Department of AYUSH, Government of India)				
Unani drugs Doses				
Symptomatic treatments				
SharbatUnnab	10-20 mL twice a day			
TiryaqArba	3-5 g twice a day			
TiryaqNazla	5 g twice a day			
KhamiraMarwareed	3-5 g once a day			
ArqAjeeb	4-8 drops in fresh water and four times a day			
Habb e IkseerBukhar (fever)	2 pills with lukewarm water twice daily			
SharbatNazla	10 mL mixed in 100 mL of lukewarm water twice daily			
Qurs e Suaal	2 tablets to be chewed twice daily			
Decoction				
Behidana	3 g			
Unnab	7 nos			
Sapistan	7 nos			
Darchini	3 g			
Banafsha	5 g			
Berg-e-Gaozabaan	7 g			
Sore throat				
Khashkhash; Bazrulbanj; Post Khashkhash; Barg e Moard (Habbulaas); Tukhm e kahuMukashar; GuleSurkh	Any of them @12 g (each)			

production of virus in the acute phase and help to clear the virus if injected during the first week of the viremia peak. Therefore, plasma globulin specific to SARS-CoV-2 has to be prepared from recovered COVID-19 patients. Recently, the Delhi Government successfully applied plasma therapy to treat COVID-19 patients.

In summary, in addition to the abovementioned treatments for COVID-19, auxiliary blood purification treatment (mainly used for severe NCP patients) could be used as an alternative therapy. According to Zarbock et al[77] the ACE2 receptor, the key receptor of SARS-CoV-2, is highly expressed in human kidney (100 times higher than in the lung). Kidney is one of the target organs for SARS-CoV-2; therefore, continuous blood purification could reduce renal recovery during COVID-19. Additionally, the kidney suffers from cytokine storms under severe COVID-19 infection. Therefore, blood purification technology could be an alternative method for removing inflammatory factors, eliminating cytokine storms, correcting electrolyte imbalances and maintaining acid-base status[1]. In addition, randomized double-blind clinical trials should be used as standard methodology for large sample sizes to determine antiviral drug efficacy in clinical practice. Currently, in India the discharge policy for COVID-19 recovered patients is based on 3 tier COVID-19 facilities and the categorization of patients is based on clinical severity. The revised discharge policy is indicated in Figure 5.

# PREVENTION OF COVID-19 OUTBREAKS

COVID-19 has affected all sectors of society. Therefore, prevention is the best practice to reduce the impact of COVID-19 considering the lack of effective treatments. This can be achieved through a variety of means as follows:

#### Individual measures

Individual measures are essential in reducing the spread of COVID-19 at the community level. Community level spread is mainly caused when an infected person is in close contact with other healthy individuals. According to the WHO, the following individual measures should be taken to reduce the contamination level such



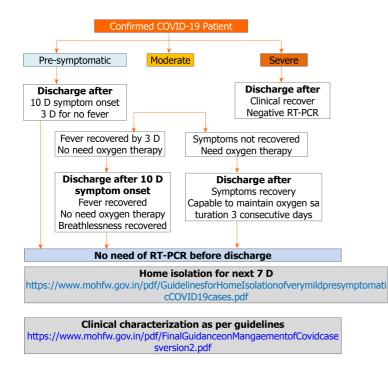


Figure 5 Discharge policies adopted by the Indian government. COVID-19: Coronavirus disease 2019.

as the use of face masks; respiratory hygiene by covering the mouth and nose with a bent elbow or tissue during coughing or sneezing; washing hands regularly with soap or disinfectant (containing at least 60% alcohol); avoiding contact with infected people, maintaining an appropriate distance (at least 2 m) from coughing or sneezing people; refraining from touching eyes, nose, and mouth with unwashed hands and finally, following advice from the healthcare provider.

#### Community level measures, social lockdown

Social lockdown is the restriction of inter-individual physical contact. Generally, it is a community level measure. The prime objective of social lockdown is to avoid two people from different families or nearby inhabitants coming in close contact with each other[78]. However, minimal and emergency movement of the general public is allowed under this condition. The emergency services (medical care, food security, general security and medicine supply) vary in different countries. However, in severe situations, emergency services such as the food and medical supply chain can also be closed as external or internal body fluid discharges such as coughs, sneezes, saliva *etc.* from COVID-19 patients infect healthy persons due to its easy transmissibility. Another objective of social lockdown is to allow the community to develop mild or full resistance to a mutated virus[78]. Moreover, it provides researchers more time to work on medicine or vaccines production. Considering the advantages of social lockdown, many nations across the globe have started different degrees of social lockdown to prevent SARS-CoV-2 infection.

#### International social lockdown progress

Some of the international social lockdown campaigns have been addressed here to understand COVID-19 preventive measures. Since the outbreak of COVID-19, China was the first country to implement social lockdown, which occurred in the last week of January 2020 in Wuhan city, the epicenter of the COVID-19 outbreak. During lockdown, buses and cars were allowed to run but domestic flights and trains were cancelled in various cities, and around 760 million people were under lockdown[29]. Accordingly, the WHO praised China as they had taken "perhaps the most ambitious, agile and aggressive disease containment effort in history" [79-81]. After China, Italy was the second country to adopt social lockdown. In Italy, social lockdown was declared on February 21, 2020 in northern Italy covering only 50000 people. Considering the disease incidence, the Federal government of Italy declared whole country lockdown on March 9, 2020. Only public transport was partially allowed, and a public pass system was initiated to ride buses or board flights on an emergency basis [82].



COVID-19 in the USA was spreading very rapidly with a high death rate since its first official COVID-19 case. Higher infection was mainly due to either higher migrant movement or a higher rate of clinical diagnosis[83]. Hence, following the high death and infection rate in the USA, the Trump government implemented the first lockdown on March 19, 2020 but to achieve total control of COVID-19, the American government extended the lockdown period to April 30, 2020 on March 30, 2020. The Trump government explained the second lockdown as follows "The better you do, the faster this whole nightmare will end. Therefore, we will be extending our guidelines to April 30th to slow the spread." Accordingly, the Director of NIH recommended the people of the USA to adapt to the lockdown voluntarily and stringently<sup>[84]</sup>. Most of the African countries had started to implement social distancing in the middle of March and ended it between May 10 and May 20, 2020. The same window was also used by most European countries. Social distancing in Bangladesh was implemented by Prime Minister Sheikh Hasina very late on March 25, 2020 and ended on May 16, 2020. Other countries such as Pakistan and Sri Lanka started to implement social distancing on March 24, 2020 which ended on May 9, 2020. Additionally, Sri Lanka declared a curfew to maintain strict social distancing.

#### Social lockdown status in India

Being a populous country, a large portion of the population lives in places of high density and their unhygienic lifestyle results in frequent infectious and epidemic diseases[85]. Therefore, as World Bank data have indicated India is still struggling to improve its health care system and is unable to provide sufficient hospital beds for its citizens. India can only afford 0.7 hospital beds per 1000 people, the doctor: population ratio is 1:1800 (standard is 1:1000), and the total number of ventilators available is 48000[86]. Considering this, the Government of India under Prime Minister Narendra Modi declared a Janata Curfew for 14-h (from 7 a.m. to 9 p.m.) on March 22, 2020 prior to total lockdown. Except for 'essential services' (police, medical services, media and home delivery) everyone took part in the curfew. According to Swiss firm IQAir, at least 75 Indian districts took part and helped to control the spread of SARS-CoV-2, which had an immediate positive effect, especially in Delhi, which is known as one of the world's most polluted capital cities. This resulted in a massive change in New Delhi's Air Quality Index (AQI). This was mainly due to a huge reduction in vehicular traffic; during lockdown there was a 70% reduction in the demand for petroleum oil. India is the third largest user of oil, after the USA and China. After that a nationwide lockdown for 21 days (except emergency services) was declared on March 24, 2020. The government implemented the following restrictions: (1) ban on people from stepping out of their homes; (2) closed all services and shops except pharmacies, hospitals, banks, grocery shops and other essential services; (3) closed all commercial and private establishments (only work-from-home allowed); (4) suspended all educational, training, and research institutions; (5) closed all places of worship; (6) suspended all non-essential public and private transport; (7) prohibited all social, political, sports, entertainment, academic, cultural, and religious activities; and (8) suspended entry of all international commercial flights from March 22. During the first phase of lockdown, the infection rate was not as high as that in the USA, Spain and Italy. It was previously reported that temperature may adversely affect virus infection [87]. Considering the influence of the upcoming Indian hot and humid summer, the health experts urged the Government to extend the lockdown. Many international news agencies described this strict lockdown by the Indian government as harsh, intensive and mismanaged[88,89]. However, the WHO declared that "the measures taken by India to break the community spread of COVID-19 by the lockdown was a very early, scientific and timely decision" [90]. In the words of Dr. David Nabarro, special envoy on the disease, WHO "The lockdown in India was quite early on, when there was relatively a small number of cases detected. This was really a far-sighted decision because it gave the whole country the opportunity to come to terms with the reality of this enemy. People understood that there is a virus in our midst. It gave time to develop capacities at the local level for interrupting transmission and sorting out hospitals. Of course, there is a lot of debate and criticism, and inevitably with a lot of frustration and anger that life is being disturbed in this way. It is very, very upsetting. I think it is courageous of the government, honestly, to take this step and provoke this enormous public debate and let the frustration come out, to accept that there will be hundreds of millions of people whose lives are being disrupted. For poor people on daily wages, this is a massive sacrifice they are making. And to do it now at an early stage as opposed to waiting three or four weeks later when the virus is much more widespread was very courageous [91]."

In the second phase, PM Modi extended the nationwide lockdown on April 14, 2002 until May 3, with a conditional relaxation after April 20. On April 16, lockdown areas



were classified as "red, orange and green zones", indicating the presence of infection hotspots, some infection, no infections, respectively. On April 20, the government announced relaxations in different sectors such as agriculture including dairy, aquaculture and plantations, selling of farming products, cargo transportation including trucks, trains and planes following social distancing norms[92]. On April 25, the government allowed the opening of small retail shops with half-staff following social distancing norms. On April 29, the Ministry of Home Affairs allowed inter-state movement of migrant people following the guidelines laid down by the government. An additional extension (May 4 - May 17) was granted by Government of India on May 1, 2020 with additional relaxation to curb the infection.

In this phase, the whole country was categorized into three zones namely red zones (130 districts), orange zones (284 districts) and green zones (319 districts). Red zones were areas with high infection and a high doubling rate, orange zones had comparatively fewer cases and green zones had no cases in the past 21 days. Normal movement was allowed in green zones with buses (50% capacity). In orange zones, only private and hired vehicles but no public transportation was allowed, while red zones were under complete lockdown. The government then implemented a fourth phase of lockdown to prevent COVID-19 between May 18 and May 31, 2020. On May 30, the government extended the ongoing lockdown until June 30 for only containment zones with services resumed in a phased-manner from 8 June. This was termed "Unlock 1.0". The second phase of unlock, called Unlock 2.0, was announced for the period of 1 to 31 July, followed by the easing of restrictions. Currently, Unlock 3.0 has been announced for August.

# ENVIRONMENTAL PERSPECTIVES: INFLUENCE AND IMPACTS

The lockdown period has greatly helped the environment to rejuvenate, simply due to a reduction in pollution level to a large extent.

#### Longevity of SARS-CoV-2 in the environment

SARS-CoV-2 can remain suspended for approximately 30 min as an aerosol (< 5 µm). SARS-CoV-2 remained viable in aerosols for up to 3 h, with a reduction in infectious titer from  $10^{35}$  to  $10^{27}$  TCID<sub>50</sub> per L of air. SARS-CoV-2 is more stable on plastic and stainless steel than on copper and cardboard[78]. The virus has the longest life on plastic and steel, surviving up to 72 h but the total number of virus particles decreases sharply over this time (10<sup>3.7</sup> to 10<sup>0.6</sup> TCID<sub>50</sub> per mL of medium after 72 h on plastic and 10<sup>3.7</sup> to 10<sup>0.6</sup> TCID<sub>50</sub> per mL after 48 h on stainless steel). On copper, it survives up to 4 h [78]. On cardboard, it survives up to 24 h, which suggests packages that arrived in the mail should have only low levels of the virus. On copper and cardboard, the virus is undetectable by 8 and 48 h, respectively [78]. The half-life of SARS-CoV-2 is similar to SARS-CoV-1 in aerosols, with a median of approximately 1.1 to 1.2 h and 95% credible intervals of 0.64 to 2.64 for SARS-CoV-2 and 0.78 to 2.43 for SARS-CoV-1[78]. The halflife of these two viruses is also similar on copper. On cardboard, the half-life of SARS-CoV-2 is longer than SARS-CoV-1. The longest viability was detected on stainless steel and plastic; the estimated median half-life of SARS-CoV-2 is 5.6 h on stainless steel and 6.8 h on plastic[78].

#### Meteorological influence

The COVID-19 pandemic is spreading globally irrespective of meteorological influence. Meteorological factors such as temperature, weather conditions and humidity are thought to play a vital role in COVID-19 transmission. At the beginning of the outbreak, it was speculated that COVID-19 may decrease with increasing air temperature as the outbreak occurred in the winter months [93]. Additionally, air temperature was relatively low in those months in comparison with Spring and/or Summer months. Accordingly, Zhou and Xie[94] demonstrated there is no concrete evidence of a decrease in COVID-19 when ambient temperature increases. Recently, Ma et al [95] indicated the positive influence of temperature and humidity on COVID-19 *i.e.*, increase in temperature and humidity decreases the number of COVID-19 deaths. This study was also conducted in same time period (January-February) as the study by Zhou and Xie[94]. A similar positive influence of meteorological factors on COVID-19 in various countries [96,97] was demonstrated. In addition to meteorological factors, Ramadhan[96] highlighted very high mobility and high density of people resulted in fast transmission of COVID-19 in Jakarta.



#### Influence on air quality

COVID-19 transmission has a direct impact on air quality namely particulate matter, SOx, NOx and carbon, etc. Standard air quality is essential in maintaining human health. However, almost 91% of the world's population lives in very poor air quality that exceeds the permissible limits [98], resulting in approximately 8% of deaths globally mainly in Asia, Africa and parts of Europe[98]. Coccia[99] demonstrated that cities (North Italy) with poor air quality ( $PM_{10}$  or ozone) increased the probability of COVID-19, mainly due to air pollution-to-human rather than human-to-human transmission. Another study from the same city indicated that prolonged exposure to poor air quality (PM<sub>10</sub>, PM<sub>2.5</sub>, O<sub>3</sub>, SOx and NO<sub>2</sub>) boosts COVID-19 incidence and even death in elderly people who have severe respiratory and cardiovascular disorders[97].

On the other hand, COVID-19 has significantly improved the air quality globally, particularly during lockdown periods due to the cessation of social activity, industrial activity, institutional activity, etc. Columbia University reported that the amount of carbon monoxide and carbon dioxide in New York City was reduced by 5% and 10%, respectively. During February 2020, carbon emission was decreased by 25% in China, which was last recorded during the economic crisis of 2008-2009. NASA's OMI instrument measured a 36% reduction in NO<sub>2</sub> concentration in China as well as in Italy, Spain, and France during February 2020 (these countries declared lockdown before other European nations). The level of particulate matter ( $PM_{2,5}$ ) in London, Cardiff, and Bristol was less following the implementation of lockdown. PM induces inflammation in lung cells and exposure to PM increases the susceptibility and severity of COVID-19 symptoms.

In China, there was a profound decline in air pollution (greenhouse gases) during January and February as recorded by NASA using satellite images due to the decrease in industrial, business and transportation activity. Accordingly, the China's Ministry of Ecology and Environment declared that it is 'good quality, air days'.

An approximately 43%, 31%, 10%, and 18% decrease in PM<sub>2.5</sub>, PM<sub>10</sub>, CO, and NO<sub>2</sub> levels, respectively, were observed in India during COVID-19 lockdown compared to previous years[100]. The AQI was reduced by 44%, 33%, 29%, 15% and 32% in north, south, east, central and western India, respectively. In New Delhi, the AQI was reduced to as low as 93, and in Mumbai it decreased to 90 from 161 and 153, respectively.

Due to quarantine, NO<sub>2</sub> level was reduced by 22.8  $\mu$ g/m<sup>3</sup> and 12.9  $\mu$ g/m<sup>3</sup> in Wuhan and China, respectively. PM25 level dropped by 1.4 µg/m3 in Wuhan but in another 367 cities it was decreased by 18.9 µg/m<sup>3</sup>[103]. After two weeks of lockdown in Spain, the black carbon and NO<sub>2</sub> level decreased markedly (-45 to -51%)[102]. However, O<sub>3</sub> level increased (+33 to +57%, 8 h daily), probably due to lower titration of O<sub>3</sub> by NO due to lower NOx level[102]. Additionally, the Copernicus Atmosphere Monitoring Service (CAMS) of the European Union observed a drop in PM<sub>25</sub> level during February 2020 in comparison with the previous three years. In China, according to CAMS[103], an approximately 20%-30% decrease in PM25 was recorded in different parts of China during February 2020 compared with monthly averages in February 2017, 2018 and 2019. It is likely that the improvement in air quality around the globe was recorded due to COVID-19 control measures mainly by lockdown and quarantine[104-108]. During this period the demand for petroleum oil was reduced by 20% worldwide.

Furthermore, different national and international media on 10th February reported increased SO<sub>2</sub> concentration of approximately 1,350 µg/m<sup>3</sup> in Wuhan and Chongqing cities due to mass cremation of COVID-19 victims based on a screenshot image from windy.com. These were the results of the GEOS-5 Model. On the other hand, The Sun showed that this was not certain but mainly due to the cremation of virus-infected victims. Accordingly, The Sun (https://archive.is/ShAfz), WION (https://archive.is/ Cdz4d) and IndiaTimes (https://timesofindia.indiatimes.com/times-fact-check/ news/fact-check-satellite-images-showing-high-levels-of-sulphur-dioxide-indicatemass-cremations-in-china/articleshow/74130633.cms) demonstrated that the mass cremations in Wuhan and Chongqing cities could be the prime reason for increased SO<sub>2</sub> concentration. Dr Arlindo M da Silva, from the Global Modeling and Assimilation Office, stated that GEOS-5 sulfur dioxide models do not "assimilate real satellite data" to confirm the image of *windy.com*. The China National Environmental Monitoring Center and the Center for Satellite Application on Environment and Ecology and the Chinese Academy of Sciences explained that the SO<sub>2</sub> data fluctuated between 4 and 8  $\mu$ g/m<sup>3</sup>, which was over 200 times less than the data shown on the website.

#### Influence on noise level and water quality

Environmental noise produced mainly by industrial or commercial operations, transit



vehicles, and many other sources cause serious health problems in the population [109]. The implementation of quarantine and lockdown due to COVID-19 preventive measures by most governments around the globe has compelled people to stay at home. The use of private and public transportation including trains and planes decreased significantly. Additionally, all commercial activities, shopping complexes and industrial operations stopped almost entirely. Accordingly, it is thought that noise level should have reduced; however, there are currently no studies on this issue. Most studies are confined to air quality assessment. Therefore, more attention should be focused on this environmental aspect.

Water quality in freshwater and marine ecosystems is also expected to improve globally. The lack of tourists, as a result of social distancing, has caused a significant change in beaches around the world. Coastal areas are important natural assets, which provide recreation and tourism, and fishing activities. These services are crucial for the nutrition and survival of coastal animals and human communities, and impart intrinsic values[110]. The lack of tourists has resulted in less pollution, especially plastics and wastes as well as reduced drainage volume into water bodies. A lower pollution level in aquatic ecosystems improves the health of the ecosystem by improving the health of aquatic organisms. In undisturbed habitats, olive ridley turtles were able to lay their eggs in Odisha's Gahirmatha beach and Rushikulya roockery. A number of dolphins were observed jumping in the water at the Marine Drive of Mumbai in the Arabian Sea, and the Canals of Venice are now full of fish and dolphins, as the water has sufficient time for sediments to settle to the bottom. According to Sunita Narain, the environmental activist, also the Director General of the Centre for Science and Environment (CSE), explained that, "Right after this health crisis subsides, it is imperative to get the economy back in shape. People need to get back to work and continue leading their lives. This is just a phase. People can learn from it. However, we require long-term solutions like that of the utilization of clean energy, conservation of forests, and efficient waste management systems in order to see real impact." According to R. Ramamurthy, COVID-19 is an eye-opener. For example, beaches such as those of Acapulco (Mexico), Barcelona (Spain), or Salinas (Ecuador) are now cleaner with crystal clear waters[101]. This aspect also needs further study to understand the impact of COVID-19.

#### Influence on waste generation and waste recycling

A number of environmental issues such as air and water pollution, soil erosion, and deforestation are responsible for direct or indirect generation of organic and inorganic waste[111]. Home quarantine measures, established across most countries as COVID-19 measures, have expanded online shopping dramatically. Accordingly, online procurement systems enhanced the generation of inorganic waste due to packaging, in addition to enhanced organic waste generation by households. Furthermore, medical waste generation is also high. In Wuhan, around 240 metric tons of medical waste is generated per day since the COVID-19 outbreak, which is too high compared with previous years (average 50 tons)[45]. Calma[112] reported that in countries like the USA garbage generation due to personal protective equipment such as masks and gloves have increased significantly compared with previous years.

Waste recycling is a common and effective way to prevent pollution, save energy, and conserve natural resources; simultaneously, it is a major environmental problem across the globe[113,114]. Although wastes are generated in high volume globally, at present it is impossible for all countries to recycle these wastes due to the further spread of SARS-CoV-2 infection. Accordingly, the USA has closed waste recycling totally due to COVID-19. Affected European countries have also restricted waste management during this outbreak [101]. For example, Italy totally prohibited infected residents from sorting their waste. Industry also seized the use of reusable bags, as single-use plastic can harbor viruses[115]. China has implemented the use of additional disinfectant in wastewater treatment plants to strengthen their disinfection process to prevent the new coronavirus spreading *via* wastewater. However, to date, there is no evidence of the survival of SARS-CoV-2 in drinking water or wastewater [116].

#### Other indirect influences on the environment

Wildlife is also affected by SARS-CoV-2. In a USA sanctuary, one tiger was reported to be coronavirus positive. In a Chinese sanctuary, two pangolins died due to the virus infection. It also affected the movement of migratory birds. Different migratory birds are now visiting places where they never visited before due to high pollution levels. It has also forced the UN organization to postpone the Annual Climate Change Conference, *i.e.*, COP-26, which was scheduled to be held at Glasgow in the UK in



November 2020.

# SOCIAL IMPACTS

COVID-19 outbreaks have adversely affected different sectors of society with big losses globally in terms of both monetary and personal loss, which cannot be accurately estimated. However, some aspects can be addressed here. Globalization is a chain process; therefore, it will collapse if a single chain stops working. In particular, the economy of countries is adversely affected. Functions, especially business meetings, sports events, scientific conferences, running educational institutes, fashion shows, and wedding parties are to be avoided, which has a big social impact on society. In the educational sector, many countries banned the running of schools, colleges and universities as well as students attending classes, which has deprived the students of a good quality education. This loss poses a large problem not only in monetary matters but also a big disadvantage to the students and their families mainly due to psychological stress. Apart from this, the tourism sector and industrial sectors are facing a major problem due to lack of labor. Prices of commodities are increasing, which has had a negative impact on poor people worldwide. Implementation of lockdown has had an enormous negative impact on poor people especially their daily wage as they are unable to earn. According to the ILO, half of permanent employees will be deprived of work, particularly in the Asia and Pacific regions. In India, 90% of workers from unorganized sectors were highly affected. In addition, production in eight major sectors was reduced by 6.5%, which obviously affected the industrial production index. According to an estimate by the IATA there was a loss of about \$113 billion during the lockdown period so far. However, the positive effect of social lockdown is spending more time with family members as well as friends but without physical meetings. It has positive effects on health and accordingly improves immunity.

This pandemic has had a serious impact on major festivals around the world, which may lead to secondary epidemic burnout and stress-related absenteeism. The Public Health Department of England has mentioned 14 ways to protect mental health during the pandemic. The WHO has recommended two most effective protocols, the R-TEP (Recent Traumatic Episode Protocol) and G-TEP (Group Traumatic Episode Protocol) to treat the invisible and psychological wounds of trauma in these situations. *'The Lancet'* documented the psychological impact of quarantine in people which included low mood, insomnia, stress, anxiety, anger, irritability, emotional exhaustion, depression and post-traumatic stress symptoms. Some people have a higher risk because of long-term absenteeism from work due to illness and burnout, which has led to a loss of productivity of approximately 35% in these workers (America's State of Mind Report). In the case of patients who are in quarantine with their children they are facing major mental disorders such as trauma-related health disorder.

It is obvious that this pandemic has both long- and short-term implications on public mental health. Poor mental health may be the result of social isolation and loneliness. It is reported that 47% cases showed negative mental health effects due to worry or stress related to coronavirus, in particular, the situation is very pronounced among older adults and households with adolescents. Research has shown that older adults are at higher risk of poor mental health due to loneliness and bereavement. It also showed that job loss enhances depression, anxiety, distress, and low self-esteem and a higher rate of disorders. In the USA, 30 million students and subsequently their families face physical, social, and mental health impairment. During this pandemic, mental health illness among adolescents has been exacerbated, and over 12% of adolescents aged between 12 and 17 years have depression and/or anxiety. Closures of non-essential businesses and disruption to livelihood have a negative impact on mental health. It has been observed that people with low incomes (about 26%) experience major negative mental health impacts (worry, 17% and stress, 14%) compared with high income groups. Presently, the Coronavirus Aid, Relief, and Economic Security Act (CARES Act) endorsed the need for emergency services to improve the mental health conditions of remote people. According to the CDC, people who suffer from chronic illness such as chronic lung disease, asthma, chronic cardiovascular disease, and diabetes are at high risk of severe illness due to COVID-19.

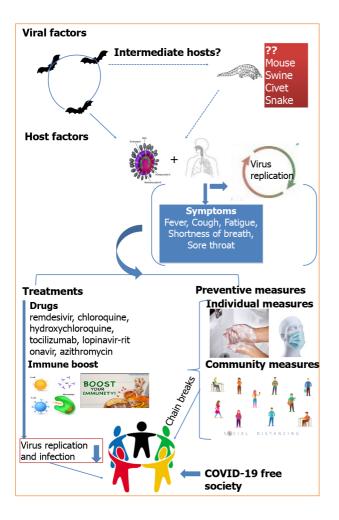


Figure 6 Schematic presentation of the management of coronavirus disease 2019 outbreaks. COVID-19: Coronavirus disease 2019.

# **RECURRENCE OF COVID19**

Although a large number of individuals recover from COVID-19, the incidence of SARS-CoV-2 RNA recurrence has been recorded in various countries. To date, the incidence of recurrent SARS-CoV-2 in recovered individuals ranges from 7.35 to 21.4% [117]. Bonifácio et al[118] reported the recurrence of COVID-19 in a female nurse from Brazil. Following her recovery, two family members developed flu-like symptoms and tested positive for COVID-19 by RT-PCR. The next day, the nurse experienced malaise, myalgia, severe headache, fatigue, weakness, feverish sensation, sore throat, anosmia and dysgeusia. Hoang[119] estimated that 15% (95%CI, 12% to 19%) of patients (among 3,644 patients, recovering from COVID-19) tested positive for SARS-CoV-2. In addition, Hoang[119] documented that the proportion was 14% (95%CI, 11% to 17%) in China and 31% (95%CI: 26%-37%) in Korea. Furthermore, he demonstrated that among recurrent cases, 39% (95%CI: 31%-48%) experienced at least one comorbidity. The estimates for times from disease onset to admission, from admission to discharge, and from discharge to RNA positive conversion were 4.8, 16.4, and 10.4 d, respectively [119]. Loconsole *et al*[120] reported the recurrence of COVID-19 in a 48-year-old man from Italy who developed dyspnea and chest pain. The recurrence of COVID-19 has been reported around the world, and raises questions about the durability and quality of immune protection from SARS-CoV-2 as well as the quality of treatment options.

# FUTURE PERSPECTIVES

COVID-19 has been an unprecedented disaster around the globe in every aspect, especially environmental health, social and economic aspects. This pandemic originated from bats. People worldwide are consuming different animals including bats, cats, snakes, mice, rats, pigs, dogs, etc., as food stuff. Accordingly, our future generation must be provided with substantial knowledge before consuming these



animals as food. Furthermore, people should be informed about the negative impact of these foods as they may harbor dangerous microbes. Emphasis should be given to providing adequate health care facilities to all people across countries including a greater number of health care systems, health insurance etc. This pandemic has highlighted the lack of health care facilities across the globe. Therefore, investment is needed in science and technology to establish specialized research centers to fight against such disasters in the future. In addition, more scientific studies are needed especially on viral diseases, mosquito-and insect-based diseases, bacterial infections, cancer, etc., to combat any future pandemics. Currently, no medicine or vaccines have been identified to treat or eradicate COVID-19. Therefore, efforts should be focused on developing effective medicine or vaccines to treat COVID-19 through technological advancements.

# CONCLUSION

This review provides an insight into the current status of COVID-19 (to date) from an environmental perspective. COVID-19 is a zoonotic disease, which originated from bats in Wuhan, China and was declared a pandemic by the WHO. The main symptoms are high fever, cough, shortness of breath and fatigue, which are similar to those of SARS. COVID-19 is highly infectious and transmissible through either aerosol droplets or close contact. The virus has spread to 213 countries/territories with approximately 202608306 confirmed cases and 4293591 deaths up to August 10, 2021. SARS-CoV-2 binds to human ACE2 and infects humans. Elderly people are more prone to SARS-CoV-2 compared to other age groups. To date, there is no specific medicine or vaccines for COVID-19. Currently, drugs such as remdesivir, chloroquine, hydroxychloroquine, tocilizumab, lopinavir-ritonavir, azithromycin, etc., are used to treat SARS-CoV-2 infection. However, no drug is able to induce full recovery in COVID-19 patients. Remdesivir is effective in treating the virus. Recently, the ChAdOx1 vaccine was developed by the University of Oxford's Jenner Institute and the Oxford Vaccine Group. More recently, Russia has developed a coronavirus vaccine, named Sputnik V but these are still in the testing phase. Therefore, boosting the immune response could be an effective way to improve viral resistance. Accordingly, prevention and management are currently the best solution to control COVID-19. Therefore, it is essential that we follow the preventive measures, management and quarantine strictly laid down by the concerned government (Figure 6). Source reduction as an individual protective measure is the best way to control the infection. Lockdown as a social strategy is considered an indirect, but effective alternative tool to control spread of the virus. Additionally, the pandemic has had a direct impact on the environment, society and economy. Therefore, we should promote science and technology to develop vaccines or specific drugs to combat COVID-19.

# ACKNOWLEDGEMENTS

The authors would like to thank the Department of Environmental Science of Sukanta Mahavidyalaya and The University of Burdwan for allowing working from home during the lockdown period.

# REFERENCES

- Wang L, Wang Y, Ye D, Liu Q. Review of the 2019 novel coronavirus (SARS-CoV-2) based on current evidence. Int J Antimicrob Agents 2020; 55: 105948 [PMID: 32201353 DOI: 10.1016/j.ijantimicag.2020.105948]
- 2 Liu Y, Gayle AA, Wilder-Smith A, Rocklöv J. The reproductive number of COVID-19 is higher compared to SARS coronavirus. J Travel Med 2020; 27 [PMID: 32052846 DOI: 10.1093/jtm/taaa021]
- 3 Núñez-Delgado A. What do we know about the SARS-CoV-2 coronavirus in the environment? Sci Total Environ 2020; 727: 138647 [PMID: 32315907 DOI: 10.1016/j.scitotenv.2020.138647]
- Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, Zhao X, Huang B, Shi W, Lu R, Niu P, Zhan F, Ma X, Wang D, Xu W, Wu G, Gao GF, Tan W; China Novel Coronavirus Investigating and Research Team. A Novel Coronavirus from Patients with Pneumonia in China, 2019. N Engl J Med 2020; 382: 727-733 [PMID: 31978945 DOI: 10.1056/NEJMoa2001017]
- 5 Ji W, Wang W, Zhao X, Zai J, Li X. Cross-species transmission of the newly identified coronavirus



2019-nCoV. J Med Virol 2020; 92: 433-440 [PMID: 31967321 DOI: 10.1002/jmv.25682]

- 6 Zhang C, Zheng W, Huang X, Bell EW, Zhou X, Zhang Y. Protein Structure and Sequence Reanalysis of 2019-nCoV Genome Refutes Snakes as Its Intermediate Host and the Unique Similarity between Its Spike Protein Insertions and HIV-1. J Proteome Res 2020; 19: 1351-1360 [PMID: 32200634 DOI: 10.1021/acs.jproteome.0c00129]
- 7 Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W, Si HR, Zhu Y, Li B, Huang CL, Chen HD, Chen J, Luo Y, Guo H, Jiang RD, Liu MQ, Chen Y, Shen XR, Wang X, Zheng XS, Zhao K, Chen QJ, Deng F, Liu LL, Yan B, Zhan FX, Wang YY, Xiao GF, Shi ZL. A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature 2020; 579: 270-273 [PMID: 32015507 DOI: 10.1038/s41586-020-2012-7]
- Guo YR, Cao QD, Hong ZS, Tan YY, Chen SD, Jin HJ, Tan KS, Wang DY, Yan Y. The origin, 8 transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak - an update on the status. Mil Med Res 2020; 7: 11 [PMID: 32169119 DOI: 10.1186/s40779-020-00240-0]
- 9 Wu A, Peng Y, Huang B, Ding X, Wang X, Niu P, Meng J, Zhu Z, Zhang Z, Wang J, Sheng J, Quan L, Xia Z, Tan W, Cheng G, Jiang T. Genome Composition and Divergence of the Novel Coronavirus (2019-nCoV) Originating in China. Cell Host Microbe 2020; 27: 325-328 [PMID: 32035028 DOI: 10.1016/j.chom.2020.02.001]
- 10 Xu X, Chen P, Wang J, Feng J, Zhou H, Li X, Zhong W, Hao P. Evolution of the novel coronavirus from the ongoing Wuhan outbreak and modeling of its spike protein for risk of human transmission. Sci China Life Sci 2020; 63: 457-460 [PMID: 32009228 DOI: 10.1007/s11427-020-1637-5]
- Kruse H, kirkemo AM, Handeland K. Wildlife as source of zoonotic infections. Emerg Infect Dis 11 2004; 10: 2067-2072 [PMID: 15663840 DOI: 10.3201/eid1012.040707]
- 12 NMA. COVID-19 and pets: When pandemic meets panic. Forensic Sci Int Rep 2020; 2: 100090 [DOI: 10.1016/j.fsir.2020.100090]
- 13 Chowell G, Abdirizak F, Lee S, Lee J, Jung E, Nishiura H, Viboud C. Transmission characteristics of MERS and SARS in the healthcare setting: a comparative study. BMC Med 2015; 13: 210 [PMID: 26336062 DOI: 10.1186/s12916-015-0450-0]
- 14 British Veterinary Association. Coronavirus and animals. [cited 3 April 2020]. Available from: https://www.bva.co.uk/news-and-blog/news-article/coronavirus-disease-covid-19-updates-for-theveterinary-profession/
- 15 Centers for Disease Control and Prevention. Coronavirus Disease 2019 (COVID-19). If You Have Animals, (2020). [cited 6 April 2020]. Available from: https://www.cdc.gov/coronavirus/2019ncov/daily-life-coping/animals.html
- 16 World Organisation for Animal Health (OIE). Questions and Answers on the 2019 Coronavirus Disease (COVID-19), (2020). [cited 3 April 2020]. Available from: https://www.oie.int/en/scientificexpertise/specific-information-and-recommendations/questions-andanswers-on-2019novelcoronavirus/
- 17 Wang W, Tang J, Wei F. Updated understanding of the outbreak of 2019 novel coronavirus (2019nCoV) in Wuhan, China. J Med Virol 2020; 92: 441-447 [PMID: 31994742 DOI: 10.1002/jmv.25689]
- 18 Xiao F, Tang M, Zheng X, Liu Y, Li X, Shan H. Evidence for Gastrointestinal Infection of SARS-CoV-2. Gastroenterology 2020; 158: 1831-1833.e3 [PMID: 32142773 DOI: 10.1053/j.gastro.2020.02.055]
- 19 Xia J, Tong J, Liu M, Shen Y, Guo D. Evaluation of coronavirus in tears and conjunctival secretions of patients with SARS-CoV-2 infection. J Med Virol 2020; 92: 589-594 [PMID: 32100876 DOI: 10.1002/jmv.25725
- Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, Qiu Y, Wang J, Liu Y, Wei Y, Xia J, Yu T, Zhang 20 X, Zhang L. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. Lancet 2020; 395: 507-513 [PMID: 32007143 DOI: 10.1016/S0140-6736(20)30211-7]
- 21 Remais J. Modelling environmentally-mediated infectious diseases of humans: transmission dynamics of schistosomiasis in China. Adv Exp Med Biol 2010; 673: 79-98 [PMID: 20632531 DOI: 10.1007/978-1-4419-6064-1 6
- 22 Wu JT, Leung K, Leung GM. Nowcasting and forecasting the potential domestic and international spread of the 2019-nCoV outbreak originating in Wuhan, China: a modelling study. Lancet 2020; **395**: 689-697 [PMID: 32014114 DOI: 10.1016/S0140-6736(20)30260-9]
- 23 Majumder MS, Mandl KD. Early Transmissibility Assessment of a Novel Coronavirus in Wuhan, China. SSRN 2020; 3524675 [PMID: 32714102 DOI: 10.2139/ssrn.3524675]
- 24 Lipsitch M, Cohen T, Cooper B, Robins JM, Ma S, James L, Gopalakrishna G, Chew SK, Tan CC, Samore MH, Fisman D, Murray M. Transmission dynamics and control of severe acute respiratory syndrome. Science 2003; 300: 1966-1970 [PMID: 12766207 DOI: 10.1126/science.1086616]
- Majumder MS, Rivers C, Lofgren E, Fisman D. Estimation of MERS-Coronavirus Reproductive 25 Number and Case Fatality Rate for the Spring 2014 Saudi Arabia Outbreak: Insights from Publicly Available Data. PLoS Curr 2014; 6 [PMID: 25685622 DOI: 10.1371/currents.outbreaks.98d2f8f3382d84f390736cd5f5fe133c]
- 26 Wu Z. McGoogan JM. Characteristics of and Important Lessons From the Coronavirus Disease 2019 (COVID-19) Outbreak in China: Summary of a Report of 72 314 Cases From the Chinese Center for Disease Control and Prevention. JAMA 2020; 323: 1239-1242 [PMID: 32091533 DOI: 10.1001/jama.2020.2648]



- 27 Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, Liu L, Shan H, Lei CL, Hui DSC, Du B, Li LJ, Zeng G, Yuen KY, Chen RC, Tang CL, Wang T, Chen PY, Xiang J, Li SY, Wang JL, Liang ZJ, Peng YX, Wei L, Liu Y, Hu YH, Peng P, Wang JM, Liu JY, Chen Z, Li G, Zheng ZJ, Qiu SQ, Luo J, Ye CJ, Zhu SY, Zhong NS; China Medical Treatment Expert Group for Covid-19. Clinical Characteristics of Coronavirus Disease 2019 in China. N Engl J Med 2020; 382: 1708-1720 [PMID: 32109013 DOI: 10.1056/NEJMoa2002032]
- 28 Rodriguez-Morales AJ, Cardona-Ospina JA, Gutiérrez-Ocampo E, Villamizar-Peña R, Holguin-Rivera Y. Escalera-Antezana JP, Alvarado-Arnez LE, Bonilla-Aldana DK, Franco-Paredes C, Henao-Martinez AF, Paniz-Mondolfi A, Lagos-Grisales GJ, Ramírez-Vallejo E, Suárez JA, Zambrano LI, Villamil-Gómez WE, Balbin-Ramon GJ, Rabaan AA, Harapan H, Dhama K, Nishiura H, Kataoka H, Ahmad T, Sah R; Latin American Network of Coronavirus Disease 2019-COVID-19 Research (LANCOVID-19). Electronic address: https://www.lancovid.org. Clinical, laboratory and imaging features of COVID-19: A systematic review and meta-analysis. Travel Med Infect Dis 2020; 34: 101623 [PMID: 32179124 DOI: 10.1016/j.tmaid.2020.101623]
- Zhong R, Mozur P, Tame TYT. Coronavirus, mao-style social control blankets china. The New Yorks Times. [cited 10 April 2020]. Available from: https://www.nytimes.com/2020/02/15/business/china-coronavirus-lockdown.html
- Lessler J, Reich NG, Brookmeyer R, Perl TM, Nelson KE, Cummings DA. Incubation periods of 30 acute respiratory viral infections: a systematic review. Lancet Infect Dis 2009; 9: 291-300 [PMID: 19393959 DOI: 10.1016/S1473-3099(09)70069-6]
- 31 Cho SY, Kang JM, Ha YE, Park GE, Lee JY, Ko JH, Kim JM, Kang CI, Jo IJ, Ryu JG, Choi JR, Kim S, Huh HJ, Ki CS, Kang ES, Peck KR, Dhong HJ, Song JH, Chung DR, Kim YJ. MERS-CoV outbreak following a single patient exposure in an emergency room in South Korea: an epidemiological outbreak study. Lancet 2016; 388: 994-1001 [PMID: 27402381 DOI: 10.1016/S0140-6736(16)30623-71
- 32 de Wit E, van Doremalen N, Falzarano D, Munster VJ. SARS and MERS: recent insights into emerging coronaviruses. Nat Rev Microbiol 2016; 14: 523-534 [PMID: 27344959 DOI: 10.1038/nrmicro.2016.81]
- 33 Belouzard S, Millet JK, Licitra BN, Whittaker GR. Mechanisms of coronavirus cell entry mediated by the viral spike protein. Viruses 2012; 4: 1011-1033 [PMID: 22816037 DOI: 10.3390/v4061011]
- 34 Lupia T, Scabini S, Mornese Pinna S, Di Perri G, De Rosa FG, Corcione S. 2019 novel coronavirus (2019-nCoV) outbreak: A new challenge. J Glob Antimicrob Resist 2020; 21: 22-27 [PMID: 32156648 DOI: 10.1016/j.jgar.2020.02.021]
- 35 Letko M, Marzi A, Munster V. Functional assessment of cell entry and receptor usage for SARS-CoV-2 and other lineage B betacoronaviruses. Nat Microbiol 2020; 5: 562-569 [PMID: 32094589 DOI: 10.1038/s41564-020-0688-y]
- Bennardo F, Buffone C, Giudice A. New therapeutic opportunities for COVID-19 patients with 36 Tocilizumab: Possible correlation of interleukin-6 receptor inhibitors with osteonecrosis of the jaws. Oral Oncol 2020; 106: 104659 [PMID: 32209313 DOI: 10.1016/j.oraloncology.2020.104659]
- Chen C, Zhang XR, Ju ZY, He WF. [Advances in the research of mechanism and related 37 immunotherapy on the cytokine storm induced by coronavirus disease 2019]. Zhonghua Shao Shang Za Zhi 2020; 36: 471-475 [PMID: 32114747 DOI: 10.3760/cma.j.cn501120-20200224-00088]
- Hoffmann M, Kleine-Weber H, Krüger N, Müller M, Drosten C, Pöhlmann S. The novel 38 coronavirus 2019 (2019-nCoV) uses the SARS-coronavirus receptor ACE2 and the cellular protease TMPRSS2 for entry into target cells. 2020 Preprint. Available from: bioRxiv [DOI: 10.1101/2020.01.31.929042]
- 39 Wrapp D, Wang N, Corbett KS, Goldsmith JA, Hsieh CL, Abiona O, Graham BS, McLellan JS. Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. Science 2020; 367: 1260-1263 [PMID: 32075877 DOI: 10.1126/science.abb2507]
- 40 Xia S, Zhu Y, Liu M, Lan Q, Xu W, Wu Y, Ying T, Liu S, Shi Z, Jiang S, Lu L. Fusion mechanism of 2019-nCoV and fusion inhibitors targeting HR1 domain in spike protein. Cell Mol Immunol 2020; 17: 765-767 [PMID: 32047258 DOI: 10.1038/s41423-020-0374-2]
- Yu F, Du L, Ojcius DM, Pan C, Jiang S. Measures for diagnosing and treating infections by a novel 41 coronavirus responsible for a pneumonia outbreak originating in Wuhan, China. Microbes Infect 2020; 22: 74-79 [PMID: 32017984 DOI: 10.1016/j.micinf.2020.01.003]
- 42 de Wilde AH, Snijder EJ, Kikkert M, van Hemert MJ. Host Factors in Coronavirus Replication. Curr Top Microbiol Immunol 2018; 419: 1-42 [PMID: 28643204 DOI: 10.1007/82 2017 25]
- 43 Perrier A, Bonnin A, Desmarets L, Danneels A, Goffard A, Rouillé Y, Dubuisson J, Belouzard S. The C-terminal domain of the MERS coronavirus M protein contains a trans-Golgi network localization signal. J Biol Chem 2019; 294: 14406-14421 [PMID: 31399512 DOI: 10.1074/ibc.RA119.008964
- 44 Yang Y, Lu Q, Liu M, Wang Y, Zhang A, Jalali N, Dean NE, Longini I, Halloran ME, Xu B, Zhang XA, Wang LP, Liu W, Fang LQ. Epidemiological and clinical features of the 2019 novel coronavirus outbreak in China. 2020 Preprint. Available from: medRxiv [DOI: 10.1101/2020.02.10.20021675
- 45 Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q, Wang J, Cao B. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet 2020; 395: 497-506 [PMID: 31986264 DOI: 10.1016/S0140-6736(20)30183-5]



- Li Z, Wu M, Guo J, Yao J, Liao X, Song S, et al Caution on kidney dysfunctions of 2019-nCoV 46 patients. 2020 Preprint. Available from: medRxiv [DOI: 10.1101/2020.02.08.20021212]
- 47 Wu J, Chen ZJ. Innate immune sensing and signaling of cytosolic nucleic acids. Annu Rev Immunol 2014; 32: 461-488 [PMID: 24655297 DOI: 10.1146/annurev-immunol-032713-120156]
- 48 Wu J, Sun L, Chen X, Du F, Shi H, Chen C, Chen ZJ. Cyclic GMP-AMP is an endogenous second messenger in innate immune signaling by cytosolic DNA. Science 2013; 339: 826-830 [PMID: 23258412 DOI: 10.1126/science.1229963]
- 49 Yoo JS, Kato H, Fujita T. Sensing viral invasion by RIG-I like receptors. Curr Opin Microbiol 2014; 20: 131-138 [PMID: 24968321 DOI: 10.1016/j.mib.2014.05.011]
- 50 Kawai T, Akira S. The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors. Nat Immunol 2010; 11: 373-384 [PMID: 20404851 DOI: 10.1038/ni.1863]
- Liu Y, Zhang C, Huang F, Yang Y, Wang F, Yuan J, Zhang Z, Qin Y, Li X, Zhao D, Li S, Tan S, 51 Wang Z, Li J, Shen C, Peng L, Wu W, Cao M, Xing L, Xu Z, Chen L, Zhou C, Liu WJ, Liu L, Jiang C. 2019-novel coronavirus (2019-nCoV) infections trigger an exaggerated cytokine response aggravating lung injury. 2020 Preprint. Available from: ChinaXiv
- Liu Q, Wang R, Qu G, Wang Y, Liu P, Zhu Y. General anatomy report of novel coronavirus 52 pneumonia death corpse. J Forensic Med 2020; 36: 19-21
- Chung M, Bernheim A, Mei X, Zhang N, Huang M, Zeng X, Cui J, Xu W, Yang Y, Fayad ZA, 53 Jacobi A, Li K, Li S, Shan H. CT Imaging Features of 2019 Novel Coronavirus (2019-nCoV). Radiology 2020; 295: 202-207 [PMID: 32017661 DOI: 10.1148/radiol.2020200230]
- Shi H, Han X, Jiang N, Cao Y, Alwalid O, Gu J, Fan Y, Zheng C. Radiological findings from 81 54 patients with COVID-19 pneumonia in Wuhan, China: a descriptive study. Lancet Infect Dis 2020; 20: 425-434 [PMID: 32105637 DOI: 10.1016/S1473-3099(20)30086-4]
- Ai T, Yang Z, Hou H, Zhan C, Chen C, Lv W, Tao Q, Sun Z, Xia L. Correlation of Chest CT and 55 RT-PCR Testing for Coronavirus Disease 2019 (COVID-19) in China: A Report of 1014 Cases. Radiology 2020; 296: E32-E40 [PMID: 32101510 DOI: 10.1148/radiol.2020200642]
- Lim CC, Tan CS, Kaushik M, Tan HK. Initiating acute dialysis at earlier Acute Kidney Injury 56 Network stage in critically ill patients without traditional indications does not improve outcome: a prospective cohort study. Nephrology (Carlton) 2015; 20: 148-154 [PMID: 25395245 DOI: 10.1111/nep.12364]
- 57 Liang W, Guan W, Chen R, Wang W, Li J, Xu K, Li C, Ai Q, Lu W, Liang H, Li S, He J. Cancer patients in SARS-CoV-2 infection: a nationwide analysis in China. Lancet Oncol 2020; 21: 335-337 [PMID: 32066541 DOI: 10.1016/S1470-2045(20)30096-6]
- Lagier JC, Colson P, Tissot Dupont H, Salomon J, Doudier B, Aubry C, Gouriet F, Baron S, 58 Dudouet P, Flores R, Ailhaud L, Gautret P, Parola P, La Scola B, Raoult D, Brouqui P. Testing the repatriated for SARS-Cov2: Should laboratory-based quarantine replace traditional quarantine? Travel Med Infect Dis 2020; 34: 101624 [PMID: 32179125 DOI: 10.1016/j.tmaid.2020.101624]
- 59 Lippi G, Simundic AM, Plebani M. Potential preanalytical and analytical vulnerabilities in the laboratory diagnosis of coronavirus disease 2019 (COVID-19). Clin Chem Lab Med 2020; 58: 1070-1076 [PMID: 32172228 DOI: 10.1515/cclm-2020-0285]
- Wan S, Yi Q, Fan S, Lv J, Zhang X, Guo L, Lang C, Xiao Q, Xiao K, Yi Z, Qiang M, Xiang J, 60 Zhang B, Chen Y. Characteristics of lymphocyte subsets and cytokines in peripheral blood of 123 hospitalized patients with 2019 novel coronavirus pneumonia (NCP). 2020 Preprint. Available from: medRxiv [DOI: 10.1101/2020.02.10.20021832]
- 61 Di L, Fu Y, Sun Y, Li J, Liu L, Yao J, Wang G, Wu Y, Lao K, Lee RW, Zheng G, Xu J, Oh J, Wang D, Xie XS, Huang Y, Wang J. RNA sequencing by direct tagmentation of RNA/DNA hybrids. Proc Natl Acad Sci USA 2020; 117: 2886-2893 [PMID: 31988135 DOI: 10.1073/pnas.1919800117]
- Smith T, Bushek J, LeClaire A, Prosser T. COVID-19 Drug Therapy. Available from: https://www.elsevier.com/ data/assets/pdf file/0007/988648/COVID-19-Drug-Therapy 2020-8-28.pdf
- 63 Di Gennaro F, Pizzol D, Marotta C, Antunes M, Racalbuto V, Veronese N, Smith L. Coronavirus Diseases (COVID-19) Current Status and Future Perspectives: A Narrative Review. Int J Environ Res Public Health 2020; 17 [PMID: 32295188 DOI: 10.3390/ijerph17082690]
- 64 Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M, Shi Z, Hu Z, Zhong W, Xiao G. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. Cell Res 2020; 30: 269-271 [PMID: 32020029 DOI: 10.1038/s41422-020-0282-0]
- 65 Holshue ML, DeBolt C, Lindquist S, Lofy KH, Wiesman J, Bruce H, Spitters C, Ericson K, Wilkerson S, Tural A, Diaz G, Cohn A, Fox L, Patel A, Gerber SI, Kim L, Tong S, Lu X, Lindstrom S, Pallansch MA, Weldon WC, Biggs HM, Uyeki TM, Pillai SK; Washington State 2019-nCoV Case Investigation Team. First Case of 2019 Novel Coronavirus in the United States. N Engl J Med 2020; 382: 929-936 [PMID: 32004427 DOI: 10.1056/NEJMoa2001191]
- Lu HT, Yang JC, Yuan ZC, Sheng WH, Yan WH. [Effect of combined treatment of 66 Shuanghuanglian and recombinant interferon alpha 2a on cossackievirus B3 replication in vitro]. Zhongguo Zhong Yao Za Zhi 2000; 25: 682-684 [PMID: 12525074]
- Kim HY, Eo EY, Park H, Kim YC, Park S, Shin HJ, Kim K. Medicinal herbal extracts of Sophorae 67 radix, Acanthopanacis cortex, Sanguisorbae radix and Torilis fructus inhibit coronavirus replication in vitro. Antivir Ther 2010; 15: 697-709 [PMID: 20710051 DOI: 10.3851/IMP1615]
- 68 Ali I, Alharbi OML. COVID-19: Disease, management, treatment, and social impact. Sci Total



Environ 2020; 728: 138861 [PMID: 32344226 DOI: 10.1016/j.scitotenv.2020.138861]

- 69 Fiore C, Eisenhut M, Krausse R, Ragazzi E, Pellati D, Armanini D, Bielenberg J. Antiviral effects of Glycyrrhiza species. Phytother Res 2008; 22: 141-148 [PMID: 17886224 DOI: 10.1002/ptr.2295]
- 70 Wang L, Yang R, Yuan B, Liu Y, Liu C. The antiviral and antimicrobial activities of licorice, a widely-used Chinese herb. Acta Pharm Sin B 2015; 5: 310-315 [PMID: 26579460 DOI: 10.1016/j.apsb.2015.05.005]
- 71 Mathie RT, Baitson ES, Frye J, Nayak C, Manchanda RK, Fisher P. Homeopathic treatment of patients with influenza-like illness during the 2009 A/H1N1 influenza pandemic in India. Homeopathy 2013; 102: 187-192 [PMID: 23870378 DOI: 10.1016/j.homp.2013.04.001]
- 72 Mustafa S, Balkhy H, Gabere MN. Current treatment options and the role of peptides as potential therapeutic components for Middle East Respiratory Syndrome (MERS): A review. J Infect Public Health 2018; 11: 9-17 [PMID: 28864360 DOI: 10.1016/j.jiph.2017.08.009]
- Gilardin L, Bayry J, Kaveri SV. Intravenous immunoglobulin as clinical immune-modulating 73 therapy. CMAJ 2015; 187: 257-264 [PMID: 25667260 DOI: 10.1503/cmaj.130375]
- Kumar V, Jung YS, Liang PH. Anti-SARS coronavirus agents: a patent review (2008 present). 74 Expert Opin Ther Pat 2013; 23: 1337-1348 [PMID: 23905913 DOI: 10.1517/13543776.2013.823159]
- Soo YO, Cheng Y, Wong R, Hui DS, Lee CK, Tsang KK, Ng MH, Chan P, Cheng G, Sung JJ. 75 Retrospective comparison of convalescent plasma with continuing high-dose methylprednisolone treatment in SARS patients. Clin Microbiol Infect 2004; 10: 676-678 [PMID: 15214887 DOI: 10.1111/j.1469-0691.2004.00956.x]
- Hung IF, To KK, Lee CK, Lee KL, Chan K, Yan WW, Liu R, Watt CL, Chan WM, Lai KY, Koo 76 CK, Buckley T, Chow FL, Wong KK, Chan HS, Ching CK, Tang BS, Lau CC, Li IW, Liu SH, Chan KH, Lin CK, Yuen KY. Convalescent plasma treatment reduced mortality in patients with severe pandemic influenza A (H1N1) 2009 virus infection. Clin Infect Dis 2011; 52: 447-456 [PMID: 21248066 DOI: 10.1093/cid/ciq106]
- 77 Zarbock A, Kellum JA, Schmidt C, Van Aken H, Wempe C, Pavenstädt H, Boanta A, Gerß J, Meersch M. Effect of Early vs Delayed Initiation of Renal Replacement Therapy on Mortality in Critically Ill Patients With Acute Kidney Injury: The ELAIN Randomized Clinical Trial. JAMA 2016; 315: 2190-2199 [PMID: 27209269 DOI: 10.1001/jama.2016.5828]
- 78 van Doremalen N, Bushmaker T, Morris DH, Holbrook MG, Gamble A, Williamson BN, Tamin A, Harcourt JL, Thornburg NJ, Gerber SI, Lloyd-Smith JO, de Wit E, Munster VJ. Aerosol and Surface Stability of SARS-CoV-2 as Compared with SARS-CoV-1. N Engl J Med 2020; 382: 1564-1567 [PMID: 32182409 DOI: 10.1056/NEJMc2004973]
- 79 World Health Organization. Clinical Management of Severe Acute Respiratory Infection (SARI) When COVID-19 Disease Is Suspected. Interim Guidance. 2020 [DOI: 10.15557/PiMR.2020.0003]
- 80 WHO. Report of the WHO-China joint mission on coronavirus disease 2019 (COVID-19). Available from: https://www.who.int/docs/default-source/coronaviruse/who-chinajoint-mission-on-COVID-19-final-report.pdf. 2020b
- WHO. Rolling updates on coronavirus disease (COVID-19). [Accessed April 10, 2020] Available 81 from: https://www.who.int/emergencies/diseases/novel-coronavirus-2019/events-as-they-happen
- 82 **BBC.** Coronavirus: Venice carnival closes as Italy imposes lockdown. Available from: https://www.bbc.com/news/world-europe-51602007,2020a
- Rasheed Z, Allahoum R, Siddiqui U. Trump extends US social distancing until April 30: live 83 updates. 2020. Available from: https://www.aljazeera.com/news/2020/03/trump-weighs-coronaviruslockdown-york-live-updates-200328234401911.html
- 84 Collins F. To beat COVID-19, social distancing is a must. 2020. Available from: https://directorsblog.nih.gov/2020/03/19/to-beat-COVID-19-social-distancing-is-a-must/
- Naidoo D, Schembri A, Cohen M. The health impact of residential retreats: a systematic review. 85 BMC Complement Altern Med 2018; 18: 8 [PMID: 29316909 DOI: 10.1186/s12906-017-2078-4]
- 86 World Bank. Hospital beds (per 1,000 people). [cited 10 April 2020]. Available from: https://data.worldbank.org/indicator/sh.med.beds.zs
- 87 Chin AWH, Chu JTS, Perera MRA, Hui KPY, Yen HL, Chan MCW, Peiris M, Poon LLM. Stability of SARS-CoV-2 in different environmental conditions. Lancet Microbe 2020; 1: e10 [PMID: 32835322 DOI: 10.1016/S2666-5247(20)30003-3]
- 88 Daniyal S. India is enforcing the harshest and most extensive COVID-19 Lockdown in the world. [cited 11 April 2020]. Available from: https://qz.com/india/1828915/indias-coronavirus-lockdownharsherthan-china-italy pakistan/
- Abidi A, Jacinto L. Lack of compassion, more than resources, marks india's deadly lockdown 89 mismanagement. [cited 11 April 2020]. Available from: https://www.france24.com/en/20200401lack-of-compassion-more-than-resources-marks-india-s-deadly-lockdown-mismanagement
- 90 Kumar A. Coronavirus: WHO lauds Modi government's social outreach during lockdown. [cited 11 April 2020]. Available from: https://www.indiatoday.in/india/story/who-coronaviruslockdownindia-economic-stimulus-package-1662392-2020-04-02
- Sharma S. Lockdown in India was early, far-sighted and courageous': WHO envoy. [cited 25 October 2020]. Available from: https://www.hindustantimes.com/india-news/Lockdown-in-indiawas-early-thiswas-far-sighted courageous-move-who-special-envoy-on-COVID-19/storywNdCkNVOqV5gCN8Du9jJ3N.html
- 92 BBC. Coronavirus deaths exceed SARS fatalities in 2003. [cited 11 April 2020]. Available from:



https://www.bbc.com/news/world-asia-china-51431087

- 93 Barcelo D. An environmental and health perspective for COVID-19 outbreak: Meteorology and air quality influence, sewage epidemiology indicator, hospitals disinfection, drug therapies and recommendations. J Environ Chem Eng 2020; 8: 104006 [PMID: 32373461 DOI: 10.1016/j.jece.2020.104006]
- 94 Xie J, Zhu Y. Association between ambient temperature and COVID-19 infection in 122 cities from China. Sci Total Environ 2020; 724: 138201 [PMID: 32408450 DOI: 10.1016/j.scitotenv.2020.138201
- 95 Ma Y, Zhao Y, Liu J, He X, Wang B, Fu S, Yan J, Niu J, Zhou J, Luo B. Effects of temperature variation and humidity on the death of COVID-19 in Wuhan, China. Sci Total Environ 2020; 724: 138226 [PMID: 32408453 DOI: 10.1016/j.scitotenv.2020.138226]
- Tosepu R, Gunawan J, Effendy DS, Ahmad OAI, Lestari H, Bahar H, Asfian P. Correlation between 96 weather and Covid-19 pandemic in Jakarta, Indonesia. Sci Total Environ 2020; 725: 138436 [PMID: 32298883 DOI: 10.1016/j.scitotenv.2020.138436]
- 97 Conticini E, Frediani B, Caro D. Can atmospheric pollution be considered a co-factor in extremely high level of SARS-CoV-2 lethality in Northern Italy? Environ Pollut 2020; 261: 114465 [PMID: 32268945 DOI: 10.1016/j.envpol.2020.114465]
- 98 WHO. Air pollution. [cited 5 April 2020]. Available from: https://www.who.int/health-topics/airpollution#tab=tab 1
- 99 Coccia M. Factors determining the diffusion of COVID-19 and suggested strategy to prevent future accelerated viral infectivity similar to COVID. Sci Total Environ 2020; 729: 138474 [PMID: 32498152 DOI: 10.1016/j.scitotenv.2020.138474]
- 100 Sharma S, Zhang M, Anshika, Gao J, Zhang H, Kota SH. Effect of restricted emissions during COVID-19 on air quality in India. Sci Total Environ 2020; 728: 138878 [PMID: 32335409 DOI: 10.1016/j.scitotenv.2020.138878]
- 101 Zambrano-Monserrate MA, Ruano MA, Sanchez-Alcalde L. Indirect effects of COVID-19 on the environment. Sci Total Environ 2020; 728: 138813 [PMID: 32334159 DOI: 10.1016/j.scitotenv.2020.138813]
- 102 Tobías A, Carnerero C, Reche C, Massagué J, Via M, Minguillón MC, Alastuey A, Querol X. Changes in air quality during the lockdown in Barcelona (Spain) one month into the SARS-CoV-2 epidemic. Sci Total Environ 2020; 726: 138540 [PMID: 32302810 DOI: 10.1016/j.scitotenv.2020.138540]
- 103 CAMS. [cited 5 April 2020]. Available from: https://atmosphere.copernicus.eu/amid-coronavirusoutbreak-copernicusmonitors-reduction-particulate-matter-pm25-over-china
- 104 Ceylan Z. Estimation of COVID-19 prevalence in Italy, Spain, and France. Sci Total Environ 2020; 729: 138817 [PMID: 32360907 DOI: 10.1016/j.scitotenv.2020.138817]
- Ibarra-Vega D. Lockdown, one, two, none, or smart. Modeling containing covid-19 infection. A 105 conceptual model. Sci Total Environ 2020; 730: 138917 [PMID: 32387821 DOI: 10.1016/j.scitotenv.2020.138917]
- Mahato S, Pal S, Ghosh KG. Effect of lockdown amid COVID-19 pandemic on air quality of the 106 megacity Delhi, India. Sci Total Environ 2020; 730: 139086 [PMID: 32375105 DOI: 10.1016/j.scitotenv.2020.139086]
- 107 Ogen Y. Assessing nitrogen dioxide (NO2) levels as a contributing factor to coronavirus (COVID-19) fatality. Sci Total Environ 2020; 726: 138605 [PMID: 32302812 DOI: 10.1016/j.scitotenv.2020.138605]
- 108 Prata DN, Rodrigues W, Bermejo PH. Temperature significantly changes COVID-19 transmission in (sub)tropical cities of Brazil. Sci Total Environ 2020; 729: 138862 [PMID: 32361443 DOI: 10.1016/j.scitotenv.2020.138862
- 109 Zambrano-Monserrate MA, Ruano MA. Does environmental noise affect housing rental prices in developing countries? Land Use Policy 2019; 87: 104059 [DOI: 10.1016/j.landusepol.2019.104059]
- Zambrano-Monserrate MA, Silva-Zambrano CA, Ruano MA. The economic value of natural 110 protected areas in Ecuador: a case of Villamil Beach National Recreation Area. Ocean Coast Manage 2018; 157: 193-202 [DOI: 10.1016/j.ocecoaman.2018.02.020]
- Schanes K, Dobernig K, Gözet B. Food waste matters-a systematic review of household food waste 111 practices and their policy implications. J Clean Prod 2018; 182: 978-991 [DOI: 10.1016/j.jclepro.2018.02.030]
- 112 Calma J. The COVID-19 pandemic is generating tons of medical waste. [cited 5 April 2020]. Available from: https://www.theverge.com/2020/3/26/21194647/the-covid-19-pandemic-isgenerating-tons-of-medical-waste
- 113 Liu M, Tan S, Zhang M, He G, Chen Z, Fu Z, Luan C. Waste paper recycling decision system based on material flow analysis and life cycle assessment: A case study of waste paper recycling from China. J Environ Manage 2020; 255: 109859 [PMID: 32063319 DOI: 10.1016/i.jenvman.2019.109859
- Ma B, Li X, Jiang Z, Jiang J. Recycle more, waste more? J Clean Prod 2019; 206: 870-877 [DOI: 114 10.1016/j.jclepro.2018.09.063
- Bir B. Single-use items not safest option amid COVID-19. [cited 5 April 2020]. Available from: 115 https://www.aa.com.tr/en/health/single-use-items-not-safest-option-amidcovid-19/1787067
- WHO. Water, sanitation, hygiene, and waste management for COVID-19. [cited 5 April 2020]. 116 Available from: https://www.who.int/publications-detail/water-sanitation-hygiene-andwaste



management-for-covid-19

- 117 Xiao AT, Tong YX, Zhang S. False negative of RT-PCR and prolonged nucleic acid conversion in COVID-19: Rather than recurrence. J Med Virol 2020; 92: 1755-1756 [PMID: 32270882 DOI: 10.1002/jmv.25855]
- 118 Bonifácio LP, Pereira APS, Araújo DCAE, Balbão VDMP, Fonseca BALD, Passos ADC, Bellissimo-Rodrigues F. Are SARS-CoV-2 reinfection and Covid-19 recurrence possible? Rev Soc Bras Med Trop 2020; 53: e20200619 [PMID: 32965458 DOI: 10.1590/0037-8682-0619-2020]
- Hoang T. Characteristics of COVID-19 recurrence: a systematic review and meta-analysis. 2020 119 Preprint. Available from: medRxiv [DOI: 10.1101/2020.09.05.20189134]
- 120 Loconsole D, Passerini F, Palmieri VO, Centrone F, Sallustio A, Pugliese S, Grimaldi LD, Portincasa P, Chironna M. Recurrence of COVID-19 after recovery: a case report from Italy. Infection 2020; 48: 965-967 [PMID: 32415334 DOI: 10.1007/s15010-020-01444-1]



WJG

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5851-5889

DOI: 10.3748/wjg.v27.i35.5851

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

REVIEW

# Pancreatic cancer in 2021: What you need to know to win

Valeria Tonini, Manuel Zanni

ORCID number: Valeria Tonini 0000-0003-3130-2928; Manuel Zanni 0000-0001-7732-7739.

Author contributions: Tonini V and Zanni M wrote the manuscript; both authors reviewed and approved the final version.

Conflict-of-interest statement: No conflict of interest.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: Italy

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B Valeria Tonini, Department of Medical Sciences and Surgery, University of Bologna-Emergency Surgery Unit, IRCCS Sant'Orsola Hospital, Bologna 40121, Italy

Manuel Zanni, University of Bologna, Emergency Surgery Unit, IRCCS Sant'Orsola Hospital, Bologna 40121, Italy

Corresponding author: Valeria Tonini, MD, PhD, Professor, Surgeon, Surgical Oncologist, Department of Medical Sciences and Surgery, University of Bologna- Emergency Surgery Unit, IRCCS Sant'Orsola Hospital, Via Massarenti 9, Bologna 40121, Italy. valeria.tonini@unibo.it

# Abstract

Pancreatic cancer is one of the solid tumors with the worst prognosis. Five-year survival rate is less than 10%. Surgical resection is the only potentially curative treatment, but the tumor is often diagnosed at an advanced stage of the disease and surgery could be performed in a very limited number of patients. Moreover, surgery is still associated with high post-operative morbidity, while other therapies still offer very disappointing results. This article reviews every aspect of pancreatic cancer, focusing on the elements that can improve prognosis. It was written with the aim of describing everything you need to know in 2021 in order to face this difficult challenge.

Key Words: Pancreatic cancer treatment; Advanced pancreatic cancer; Metastatic pancreatic cancer; Pancreatic cancer surgery; Pancreatic cancer chemotherapy; Pancreatic cancer screening

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Pancreatic cancer is a very dangerous enemy and the results are still very unsatisfactory. But we have not given up. Research is running fast on many paths, without losing its enthusiasm. The number of articles published on this subject in the last two years is impressive. I have tried to summarize all the most significant data from the different lines of research, ranging from screening and early diagnosis to new developments in surgery and associated therapies. I hope I have succeeded in the task of describing as comprehensively as possible the most promising fields of research available to us today, in order to achieve the improved results we desire.

Citation: Tonini V, Zanni M. Pancreatic cancer in 2021: What you need to know to win. World



Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Received: January 28, 2021 Peer-review started: January 28, 2021 First decision: June 14, 2021 Revised: July 14, 2021 Accepted: August 23, 2021

Accepted: August 23, 2021 Article in press: August 23, 2021 Published online: September 21, 2021

P-Reviewer: Sato H S-Editor: Gao CC L-Editor: A P-Editor: Li JH



*J Gastroenterol* 2021; 27(35): 5851-5889 URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5851.htm DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5851

# INTRODUCTION

Pancreatic cancer is currently the seventh leading cause of cancer death worldwide and the fourth following lung, colorectal and breast cancers in the United States and Europe. It will become the third by 2030. It is an age-related neoplasm and this trend is similar between males and females. In particular the number of both deaths and incident cases peaked at the ages of 65-69 years in males, whereas the peak in females was observed at the ages of 75-79 years [1-4]. The commonly used term "pancreatic cancer" usually refers to ductal adenocarcinoma (PDAC), which represents 85% of all pancreatic tumor<sup>[4]</sup>. Complete surgical resection significantly prolongs survival, but the tumor is often diagnosed at an advanced stage and only a small percentage of patients are therefore candidates for surgery. Moreover, surgery is still associated with high post-operative morbidity. Despite ongoing developments, PDAC remains one of the most difficult tumors to treat, and the five-year survival rate is less than 10%[5]. There are four fundamental challenges that underlie the high mortality. First, the retroperitoneal location of the pancreas, deep in the abdomen, protects growing tumors from detection. The symptoms are late and therefore the diagnosis is made when the tumor is already in an advanced stage. Second, PDAC has an aggressive biology characterized by early metastasis and 50% of patients has metastatic disease at presentation. In addition, a large number of patients undergoing surgery develop metastases within 4 years. This suggests the presence of micrometastasis in apparently localized cases[6]. Third, pancreatic cancer dramatically weakens patients, limiting their ability to withstand aggressive treatments. Finally, it shows resistance to many antineoplastic therapies [7,8]. Advances in prevention, screening, early detection, and therapy, particularly on new frontiers, are essential to improve outcomes. This article has been written with the aim of describing everything you need to know in 2021 in order to face this difficult challenge.

# NON-FAMILIAL RISK FACTORS AND PREVENTION

Identification of risk factors, high-risk populations and early detection markers is the first and crucial step to change the pancreatic cancer horizon[9]. PDAC incidence rates are nearly four times higher in high-income countries such as the United States and Western European countries than in middle- and low-income countries[3]. The different incidence seems to be related with different lifestyles.

Obesity, smoking, alcohol consumption and type 2 diabetes are considered nonfamilial risk factors for pancreatic cancer. Chronic pancreatitis, cystic fibrosis and intraductal papillary mucinous neoplasm (IPMN) should also be considered. An increased risk of pancreatic cancer has been observed following gastrectomy[10-17].

One-third of all cancers could have been prevented through lifestyle correction[18]. A 2020 European prospective study (EPIC) evaluated the association between the healthy lifestyle index score and PDAC[19-22]. Healthy lifestyle habits were inversely related to the risk of PDAC. Adherence to healthy behaviors, corresponding to a three-point increase in the score, was associated with a 16%-23% lower risk. The result summarizes many previous studies[23-29] and support the adoption of healthy lifestyles in PDAC prevention.

A recent nutrigenomic study has highlighted nutrients capable of preventing cancer through epigenetic modifications. An optimal diet should include omega 3 fatty acids, polyphenols, folic acid, selenium and zinc. Particularly important for PDAC prevention could be the epigallocatechin, a polyphenol from tea and green tea[30,31].

Data linking type 2 diabetes with pancreatic cancer suggest that the new onset of diabetes in a lean older adult should prompt consideration of PDAC. This is even more valid if new-onset diabetes is associated with unintentional weight loss[32-34]. A Mayo Clinic study evaluated the use of computed tomography (CT) at the time of diabetes diagnosis in otherwise asymptomatic patients. A higher likelihood of showing potentially resectable tumors was observed compared with scans performed six months later[32]. However, CT screening of all elderly subjects with new-onset

Zaishidene® WJG | https://www.wjgnet.com

diabetes is not feasible<sup>[33]</sup>. With the identification of these characteristics that differentiate pancreatic cancer-associated diabetes from other cases of new-onset diabetes, perhaps the guidelines will updacate[35].

# HEREDITARY RISKS FACTORS

PDAC can be hereditary. There are two categories of inherited risk for PDAC: Genetic syndromes (20% of cases) and familial pancreatic cancer (80%). Familial pancreatic cancer is defined as a predisposition that is based on familial clustering in families in which there is at least one pair of first-degree relative (FDR) relatives with PDAC in the absence of a known genetic syndrome. Genetic syndromes that predispose to pancreatic cancer are listed in Table 1. Table 1 also shows in parentheses the frequencies of mutated genes in PDAC patients[36-42].

Knowledge of inherited risk factors is important because it allows us an effective stratification and management of patients. According to American Society of Clinical Oncology and National Comprehensive Cancer Network guidelines, all patients diagnosed with PDAC should be evaluated to understand if there is a risk of familial predisposition to cancer. All patients should undergo risk assessment for syndromes associated with an increased risk of PDAC. Germline genetic testing is recommended for patients with PDAC and an unremarkable family history[43,44].

# SCREENING

Screening aims to detect preinvasive lesions (IPMNs and pancreatic intraepithelial neoplasias) with high-grade neoplastic changes and early invasive tumors that are more amenable to potentially curative resection[45-49].

#### Candidates for screening

(1) Patients with Peutz-Jeghers syndrome or CDKN2A mutation, regardless of family history; (2) BRCA2 mutation with at least one affected FDR or at least two affected relatives of any degree; (3) BRCA1, partner and localizer of BRCA2 (PALB2), ataxiatelangiectasia mutated (ATM), and Lynch syndrome mutation carriers with one or more affected FDRs; (4) Hereditary pancreatitis with a PRSS1 mutation; and (5) Regardless of gene mutation status: (a) At least three affected relatives on the same side of the family, of whom at least one is an FDR of the individual being considered for surveillance; (b) At least two affected relatives who are FDRs of each other, of whom at least one is an FDR of the individual being considered for surveillance; and (c) At least two affected relatives on the same side of the family, of whom at least one is an FDR of the individual being considered for surveillance.

General population-based screening for average-risk patients is not recommended [33] because the average lifetime risk for developing PDAC is too low[49].

#### Screening modality

The current recommendation provides for the execution of endoscopic ultrasonography (EUS) or magnetic resonance imaging (MRI)/magnetic resonance cholangiopancreatography (MRCP). It has been demonstrated that they detect more lesion as compared with CT scan[50]. Screening is recommended at age 50 years or 10 years younger than the youngest relative with PDAC in familial pancreatic cancer relatives. In other cases, screening is carried out between 35 and 45 years. For patients with a normal pancreas on imaging, repeat the procedure every year alternating EUS and MRCP. The age for stopping screening should be individualized based on each patient's medical status, life expectancy, and preferences.

# SURGICAL RESECTION FOR IPMNS AND OTHER CYSTIC LESIONS

Surgical resection is indicated in patients with any of the following[45]: (1) Solid pancreatic lesion  $\geq$  5 mm of indeterminate pathology or if additional evaluation does not yield a definitive preoperative diagnosis; (2) Any positive fine-needle aspiration (FNA) result, except for a pancreatic neuroendocrine tumor; (3) Main-duct IPMNs with any one of the following: (a) Main pancreatic duct dilation of  $\geq$  10 mm; (b) Main pancreatic duct stricture; or (c) Mural nodules; (4) Branch duct IPMNs (BD-IPMNs)



Table 1 Genetic syndromes predisposing	to pancreatic cancer (the frequency of mutated genes among patients with pancreatic ductal
adenocarcinoma is indicated in brackets	

Genetic syndrome	Mutated genes	
Hereditary breast/ovarian cancer syndrome[36,37]	BRCA1 (0.7%), BRCA2 (1.4%), PALB2 (1%)	
Familial atypical multiple mole melanoma syndrome[38]	CDKN2A (0.7%)	
Peutz-Jeghers syndrome[39]	STK11	
Familial adenomatous polyposis	APC (0.4%)	
Lynch syndrome[40]	MLH1, MSH2 (0.4%), PMS2 (0.3%)	
Hereditary pancreatitis[41]	PRSS1, SPINK1	
Ataxia telangectasia[42]	ATM (1.4%)	
Li-Fraumeni syndrome[42]	P53 (0.4%)	

with any one of the following: (a) Rapid growth (> 5 mm over six months); (b) Mural nodules or an enhancing solid component; (c) Abrupt main pancreatic duct caliber change with distal atrophy (even if no mass is visible); (d) Main pancreatic duct dilation of  $\geq$  10 mm; (e) Positive cytology; or (f) Associated symptoms of pancreatitis, jaundice, or pancreatic-type pain; or (5) Asymptomatic main pancreatic duct stricture with an associated suspicious mass.

For patients who do not meet these criteria for surgery, repeat imaging in three months if worrisome features are present[47,51]. Worrisome features include the following: (1) Solid lesion with main pancreatic duct size of 5 mm to 9 mm in diameter; (2) Main pancreatic duct stricture and/or dilation  $\geq 6$  mm of unknown etiology without an associated mass; and (3) Solid lesion < 5 mm of uncertain significance.

Repeat imaging in six months is recommended for patients who have the following imaging abnormalities: (1) Cystic lesion (presumed BD-IPMN) ≥ 3 cm in size; (2) Cystic lesion with associated main pancreatic duct 5 mm to 9 mm; (3) Cystic lesion associated with lymphadenopathy; (4) Cyst growth rate of  $\geq$  5 mm in two years; and (5) Increased serum carbohydrate antigen 19-9 (CA 19-9).

Individuals without worrisome features of malignancy should undergo repeat imaging in 12 mo[47,51].

Screening/surveillance should be continued until the patient is no longer a surgical candidate.

A 2020 paper analyzed the benefits of screening. Nine out of 10 screen-detected PDAC were resectable, with a three-year survival of 85%, compared with 25% in PDAC detected outside surveillance. With continued follow-up of patients with resectable PDAC, the five-year overall survival (OS) rate was 60%[49].

#### **BIOMARKERS AND EARLY DETECTION**

Different biomarkers are being evaluated to improve early diagnosis of tumor not detectable by imaging and to differentiate cancer and high-grade dysplasia from benign disease[52].

#### Blood tests

The most useful serum tumor marker for PDAC is CA 19-9. It is recommended adding this test when there are worrisome features on abdominal imaging. The sensitivity and the specificity of elevated CA 19-9 to detect PDAC are 79% and 82%, respectively [53-55]. It becomes more precise when used in combination with CA 125[56,57]. Other carbohydrate markers, such as CA 50, CA 72.4 and CA 242, were extensively analyzed in PDAC patients. Although they exhibited less sensitivity than CA 19-9 for the diagnosis, they improved specificity [58-61]. Satake and Takeuchi [62] also studied SPan-1 and DUPAN-2. SPan-1 has a high sensitivity for PDAC (81.4%), but the specificity (67.5%) and diagnostic accuracy (71%) are lower than those of CA19-9. SPan-1 may be considered as an additional useful serum marker, but it does not significantly improve the diagnostic accuracy obtained with CA 19-9. In contrast, DUPAN-2 has a high specificity (85.3%) and low sensitivity (47.7%). Furthermore, it seems that serum levels of DUPAN-2 are influenced by liver function. SPan-1 and DUPAN-2 unfortunately have not yet shown the sensitivity and specificity needed to



be used for early detection[62,63].

A huge step forward in the early detection of pancreatic cancer could come from studying cell-free DNA (cfDNA), which consists of circulating double-stranded DNA molecules that can be found in plasma or blood serum. From the analysis of these molecules, it is possible to understand if we are in the presence of a tumor DNA and to go back to the tissue of origin. By analyzing the methylation status of two genes in cfDNA, ADAMTS1 and BNC1, early stage cancer can be identified with a sensitivity of 94.8% and a specificity of 91.6% [64].

Innovative discoveries have also been made in the field of RNA. Abnormal microRNA expressions are potential diagnostic markers for several cancers, including PDAC. Multiple microRNA tests performed in combination with CA 19-9 can improve diagnostic accuracy, particularly miR-216[65-69]. Permuth et al[70] demonstrated that a combination of eight lncRNAs helps in the differential diagnosis between malignant and non-malignant IPMNs. Furthermore, three lncRNAs (HAND2-AS1, CTD-2033D15.2, and lncRNA-TGF) could be exploited as early diagnostic biomarkers of IPMN[71,72].

#### Pancreatic juice and pancreatic cyst fluid

Pancreatic juice collected at the time of ERCP and cyst fluid obtained by EUS-guided FNA can be analyzed for molecular markers. These procedures also have broad potential in terms of early diagnosis of PDAC. Next-generation sequencing can be performed at low cost to detect low-frequency mutations. Potential markers include mutant GNAS (specific for IPMNs) and mutant KRAS. TP53, SMAD4, PIK3CA, PTEN, and AKT1 mutants are also useful as they correlate with IPMN-associated tumors 73-75]. According to Suenaga *et al*[76], a pancreatic juice collection, to ensure optimal yield of mutations for pancreatic screening assays, should be performed 10 min after secretin administration. The authors detected 40 patients with KRAS mutations in pancreatic juice out of 45 undergoing surveillance with EUS, reconfirming the usefulness of these analysis<sup>[76]</sup>.

There are many other biomarkers that are currently being validated for clinical use, such as mucins (MUC). Normal pancreatic ductal epithelium expresses low levels of MUC, while an upregulation of MUC occurs in BD-IPMNs and more pronounced changes in expression in PDAC. Normal pancreatic ductal epithelium expresses low levels of MUC, while upregulation of MUC occurs in BD-IPMN and PDAC[77-83]. The analysis of mucin changes in the fluid of pancreatic cysts allows us to differentiate mucinous from non-mucinous pancreatic cysts with high sensitivity and specificity and to diagnose PDACs associated with IPMN at an early stage[84]. MUC4 and MUC16 have been reported to be 100% specific for PDAC, while associated with sensitivities of 63% and 67%, respectively[85].

Interesting data were reported about interleukins (IL). Higher concentrations of IL-1b, IL-5, and IL-8 have been identified in cystic lesions with high grade dysplasia or malignancy[86]. IL-1b is a potentially useful factor in differentiating high-risk from low-risk pancreatic cysts.

The Das-1 monoclonal antibody is also capable of detecting pancreatic cysts at risk of malignancy with high levels of sensitivity (88%) and specificity (98%)[87,88]. Das-1, IL and MUC could be used in conjunction with clinical guidelines to identify patients at risk for malignancy.

#### Saliva

Saliva is a suitable substance for screening because it is obtained in a simple and noninvasive manner. In addition, salivary mRNA is relatively stable and informative for disease diagnosis, including cancer. Zhang et al[89] identified 7 up-regulated genes (MBD3L2, KRAS, STIM2, DMXL2, ACRV1, DMD, and CABLES1) and 5 downregulated genes (TK2, GLTSCR2, CDKL3, TPT1, and DPM1) in subjects with PDAC compared with healthy controls or those with chronic pancreatitis. A combination of 4 mRNAs (MBD3L2, KRAS, ACRV1, and DPM1) can discriminate diseased patients from healthy ones with sensitivity and specificity over 90%[89]. Xie et al[90] worked on miR-3679-5p and miR-940. The former is down-regulated, while the latter is upregulated in PDAC patients compared to controls. The combination of the two miRNAs identifies diseased subjects with sensitivity and specificity of 70%. The same group evaluated the expression of salivary long non-coding RNAs (lincRNAs). They identified HOTAIR and PV1T as significantly up-regulated lincRNAs in the PDAC group compared with controls and benign pancreatic tumors. The combination of salivary HOTAIR and PVT1 differentiated PDAC from healthy controls with a sensitivity of 78.2% and specificity of 90.9% and PDAC from benign tumors with a sensitivity of 81.8% and specificity of 95% [90,91]. Another important mRNA studied in



serum, urine, and saliva is MIR1246. Salivary expression of miR-1246 is related to serum CA19-9 levels[92]. Significantly higher expression of MIR1246 in serum and urine was observed in patients with cancer compared with healthy controls. Ishige *et al* [93] observed an AUC for MIR1246 in serum of 0.87 (sensitivity, 92.3%; specificity, 73.3%), for MIR1246 in urine of 0.90 (sensitivity, 90.2%; specificity, 83.3%). Combining the expression of MIR1246 in serum and urine resulted in a sensitivity of 85%. These results indicate that MIR246 may be a useful diagnostic biomarker for pancreatic cancer. The accuracy further increases if we consider miR-1246 and miR-4644 simultaneously[92].

#### Urine

Several biomarkers have also been evaluated in urine. Radon et al[94] used three protein biomarkers (REG1A, TFF1 and LYVE1) to form a powerful urinary panel that can detect patients with stages I-II PDAC, with over 90% accuracy. Brezgyte et al[95] found four miRNAs (miR-143, miR-204 and miR-223) in significantly higher amounts and one miRNA (miR-30e) in lower amounts in the urine of PDAC Stage I patients compared to the healthy population. These miRNAs (except for miR-204) also showed a decreased expression in Stage II-IV compared to Stage I[95]. However, more studies are needed to validate the clinical utility of these biomarkers.

# CLINICAL FEATURES

The presenting symptoms in patients with PDAC varies according to location. Tumors in the body and tail present with pain and weight loss, while tumor of the head cause jaundice and steatorrhea[96]. Pain associated with PDAC is usually insidious, visceral, generally epigastric, radiating to the sides or straight through to the back. It is worse by eating or lying supine at night. Rarely, it develops acutely on account of acute pancreatitis due to tumoral occlusion of the main pancreatic duct[97]. Pancreatic cancer may result in an onset of diabetes mellitus[98,99]. The hypercoagulable state that accompanies PDAC can result in Trousseau syndrome, which consists of superficial, sometimes migratory thrombophlebitis[100]. Thromboembolic complications occur more commonly in patients with tumors arising in the tail or body of the pancreas[101]. Skin manifestations could occur as paraneoplastic phenomena[102]. Rarely, erythematous subcutaneous areas of nodular fat necrosis (pancreatic panniculitis), typically located on the legs, may be evident. It is more frequent in patients with the acinar cell variant of PDAC. It is not pathognomonic for an PDAC, because it has also been described in associated with pancreatic neuroendocrine tumors, IPMNs and chronic pancreatitis<sup>[103]</sup>.

When assessing symptoms, it should be borne in mind that PDAC tends to infiltrate nearby organs and structures and to give distant metastases very early. Local extension typically involves adjacent structures, such as the duodenum, the portal vein (PV), or the superior mesenteric vessels. PDAC also show a striking tendency toward perineural invasion, both within and beyond the pancreas. The difficulty in achieving a wide resection margin due to the proximity to the vessels accounts for the fact that the retroperitoneal tissue behind the head of the pancreas represents the most common site of disease recurrence. Sometimes the tumor extends to the spleen, adrenal glands, vertebral column, transverse colon, and/or stomach. In these cases, tumors are not resectable. Tumor may metastasize to regional peripancreatic lymph nodes or less often to distant lymph node, peri-gastric, mesenteric, omental or porta-hepatic nodes. Distant metastasis may affect the liver, peritoneum, lungs, and less frequently, bone. Signs of advanced, incurable disease include an abdominal mass, ascites, Virchow's node, Sister Mary Joseph's node or a palpable rectal shelf. Pancreatic cancer is the origin of a cutaneous metastasis to the umbilicus in 7% to 9% of cases[104].

# DIAGNOSIS

#### СТ

CT is considered the gold standard for pancreatic cancer's diagnosis. Protocol pancreatic CT is performed for evaluation of suspected PDAC or if a routine CT scan was not sufficient for initial staging[105,106]. This protocol consists of evaluating the patient at different stages of contrast injection. The arterial phase provides excellent opacification of the celiac axis, superior mesenteric artery (SMA), and peripancreatic



arteries. An attenuation difference between tumor and normal pancreas is best achieved after peak enhancement of the aorta in the arterial phase but before the one of the liver, in the portal venous phase. This is sometimes termed the "pancreatic phase". The portal venous phase provides better enhancement of the superior mesenteric vein (SMV), splenic and PVs. In addition, peak hepatic enhancement, which optimizes the detection of hepatic metastases, also occurs in the portal venous phase[107,108].

The typical CT appearance of a PDAC is an ill-defined hypoattenuating mass within the pancreas. Smaller lesions may be iso-attenuating, making difficult their identification[109]. Secondary signs of PDAC include a dilatation of the pancreatic duct or common bile duct, parenchymal atrophy, and contour abnormalities. Dilation of both the pancreatic duct and the common bile duct, commonly referred to as the "double duct sign" is not diagnostic for a pancreatic head malignancy [110]. Routine preoperative CT helps to identify hepatic vascular anatomy and prepares the surgeon for any potential vascular anomalies. It can detect hemodynamically significant arterial stenosis[111]. The contrast-enhanced CT scan is the best technique for PDAC staging [112] and it is essential to detect vascular invasion. CT criteria for vascular invasion include arterial embedment in the tumor mass or venous obliteration, tumor involvement exceeding one-half the circumference of the vessel, vessel wall irregularity, vessel caliber stenosis, or a "teardrop" sign of the SMV[113]. Classic CT criteria for vascular involvement are not reliable in patients who have undergone neoadjuvant therapy with a highly active chemotherapy combination such as mFOLFIRINOX (mFFX). In such cases, surgical exploration may be the only method to assess resectability[114].

#### MRI

Contrast-enhanced MRI of the pancreas may be useful in staging patients at initial presentation. MRI is the best technique for detection of small liver metastases[115]. The importance of MRI also lies in the ability to diagnose pancreatic cancer by identifying changes in the body that indicate systemic effects of PDAC. It has been well recognized that anorexia, sarcopenia, and weight loss are hallmarks of PDAC. Consequently, it can be used to measure adipose and muscle mass in high-risk populations to identify early disease[116-118].

#### EUS

EUS is considered the most sensitive method to detect early neoplasia in the pancreas. PDAC on EUS appears as a hypoechoic mass, typically with dilation of the proximal pancreatic duct and the border of the lesion may have an irregular contour. This is the best accurate technique for local T and N staging, and for predicting vascular invasion. However, EUS is inferior to CT for evaluation of distant metastases. In addition, the specificity of EUS for excluding vascular invasion in small tumors is limited, particularly when inflammatory changes are present[119].

EUS is mainly used as part of the workup to obtain fine needle aspiration or biopsy material in patients suspected of having a PDAC[120]. EUS is not readily accessible and as a result is considered a complementary modality to the pancreatic protocol CT. Emerging area for endoscopic ultrasound includes the incorporation of elastography. Elastography shows significantly lower elasticity values for PDAC than for normal pancreatic tissue<sup>[121]</sup>. Incorporation of elastography in the evaluation of solid pancreatic lesions improves diagnostic accuracy[122,123].

#### Endoscopic retrograde cholangiopancreatography

A meta-analysis demonstrated a 92% sensitivity and 96% specificity of endoscopic retrograde cholangiopancreatography (ERCP) in the diagnosis of PDAC[124]. Findings suggestive of a malignant tumor of the pancreatic head include stenosis or obstruction of the common and pancreatic bile ducts (the "double duct" sign), a pancreatic duct stenosis greater than 1 cm in length, and pancreatic duct obstruction. In addition, ERCP provides an opportunity to collect tissue samples for cytohistologic analysis [124].

Some early-stage pancreatic tumors are not detected by CT, MRI, or EUS. Especially for carcinoma in situ, localized stenosis of the main pancreatic duct is often the only imaging finding. Pancreatic duct imaging evaluation by ERCP and subsequent pancreatic juice cytology are critical for diagnosis.

On the other hand, ERCP is an invasive procedure that can cause acute pancreatitis, bleeding, and cholangitis. Consequently, it has purely therapeutic value for patients with cholestasis due to tumor obstruction of the biliary system and require placement of a biliary stent[125].



#### Positron emission tomography

The role of positron emission tomography (PET) is limited for PDAC due to the high number of false positives and false negatives [126]. However, the degree of fluorodeoxyglucose (FDG) uptake correlates with histopathology, aggressiveness, and metastatic potential [127,128]. According to a meta-analysis, PET/CT is more accurate than CT in detecting distant metastases. Preoperatively, it may therefore be useful in avoiding unnecessary resection if unexpected metastases are found [129,130]. After treatment, FDG-PET is instead used to detect residual or recurrent cancer. It can also be applied to assess and monitor response to therapy in unresectable or metastatic disease[127,131].

Other molecular imaging agents including overexpressed proteins, signaling pathways, and tumor stroma may also be used[132]. Among these, promising results appear to involve 68Ga-cicratide, an integrin  $\alpha\nu\beta6$ -specific radiotracer, which has favorable pharmacokinetics and is capable of detecting pancreatic cancer lesions and monitoring response to therapy [133]. Another molecular imaging method that is of interest for early detection is hyperpolarized MRI. It can identify metabolic aberrations in the pancreas that indicate preneoplasia<sup>[134]</sup>.

#### Staging laparoscopy

Sub-centimeter metastases of the liver or peritoneum that are rarely visible by CT, MRI or PET may be visualized laparoscopically. Up to one-third of patients thought to be resectable by imaging will be found to be unresectable based upon laparoscopic findings[135,136].

Some experts suggest a selective approach to staging laparoscopy, limiting the procedure to those with the highest likelihood of occult metastatic disease[137,138]. First, this includes tumors of the body or tail of the pancreas that appear potentially resectable by CT scan. Second, it includes large (> 3 cm) primary tumors and patients with a high initial CA 19-9 level (> 100 units/mL)[139].

#### Biopsy

Biopsy of a pancreatic mass can be performed either percutaneously or via EUS. EUSguided FNA is the best modality for obtaining a tissue diagnosis. EUS-FNA is a safe method with a 0.98% morbidity and a 0.02% mortality. Although the most common adverse events of EUS-FNA include pancreatitis and postprocedural pain, there is also some concern regarding tumor cell seeding[140]. According to a study by Yane et al [141] the cumulative needle tract seeding rate at five years was 3.8%. However the preoperative EUS-FNA has no negative effect on recurrence-free survival and OS.

In many cases, the diagnosis will not yet be histologically confirmed. Once PDAC is suspected on imaging studies, the next step is generally a staging evaluation rather than biopsy. Patients who are fit for major surgery and who appear to have potentially resectable PDAC, they do not necessarily need a biopsy before surgery. Biopsy could be indicated if there is evidence of systemic spread or local evidence of unresectability on staging studies. It is also indicated if the patient is unfit for major surgery or if other diagnoses need to be excluded [142,143].

#### Pancreatic incidentaloma

A 2014 systematic review[144] evaluated 5 studies enrolling patients with incidentalomas and concluded that most solid lesions are malignant. Histologic definition of a solid lesion of the pancreas should be the first option, as opposed to radiologic monitoring alone. It is important to avoid operating on benign solid lesions such as chronic focal pancreatitis or autoimmune pancreatitis.

In case of cystic lesion, surgery is the first option for cystadenomamucinous and IPMN with high-risk stigmata. A recent review defined high-risk stigmata as the presence of obstructive jaundice, vascularized mural nodules ≥ 5 mm, main duct diameter  $\geq 10 \text{ mm}[145]$ .

## STAGING

The goal of the staging workup is to delineate the extent of disease spread and to identify patients who are eligible for resection with curative intent. Patients with PDAC can be staged according to the eighth edition of TNM system of American Joint Committee on Cancer (AJCC). However, most clinicians use a four-tiered staging system including resectable, borderline resectable, locally advanced (LA), and



metastatic cancer[146,147] (Table 2). In 2017, a classification was published, by the International Association of Pancreatology, which redefines the concept of resectability in relation to biological risk and patient conditions[148]. Table 3 summarizes the different resectability criteria assumed by the different scientific societies.

# SURGERY

Surgical resection is the only potentially curative treatment. Unfortunately, PDAC is often diagnosed at an advanced stage and radical surgery could be performed in a very limited number of patients. The surgical interventions that can be performed are different depending on the tumour location and extension. In all cases the operation involves the removal of the tumour with free margins and at least twelve lymph nodes, which are necessary for staging. Tumors of the head require more complex operations, which still have a high operative morbidity. In high-frequency surgical centres mortality after pancreatoduodenectomy (PD) is now less than 2%, but post-operative morbidity remains high, 30%-50%. Anastomotic dehiscences, are the most serious post-operative complication. They are difficult to manage and are unfortunately associated with a still high mortality rate. Tumors of the tail and body require easier operations than head tumors, with a low operative morbidity and mortality. Unfortunately, because of their late symptomatology, they are more frequently unresectable.

#### Pancreaticoduodenectomy

PD is the classic operation performed for pancreatic tumors of the head or uncinate process. Conventional pancreaticoduodenectomy involves removal of the pancreatic head, duodenum, first 15 cm of the jejunum, common bile duct, gallbladder, and a partial gastrectomy. It is a complex procedure and patients may experience several complications. These complications could be intra-operative or post-operative[149, 150].

The most important intraoperative complication of PD is bleeding. Most patients undergoing PD for PDAC have an obstructive jaundice with associated coagulopathy. Bleeding can occur from multiple sites during the various phases of mobilization and resection, so hemostasis must be monitored and assured before reconstruction begins.

Postoperative complications can be further divided into short-term and long-term complications. The short-term ones are pancreatic fistula, delayed gastric emptying, and postoperative bleeding. The long-term ones are biliary stenosis and cholangitis, pancreatitis, peptic ulcer disease, small bowel obstruction, and incisional hernia[149, 150].

Modifications of the conventional PD procedure have been developed in an attempt to improve outcomes or minimize the morbidity associated with this operation. The pylorus-preserving pancreaticoduodenectomy preserves the gastric antrum, pylorus, and proximal 3 cm to 6 cm of the duodenum. It can decrease the incidence of postoperative dumping, marginal ulceration, and bile reflux gastritis, without negative effect on the morbidity, mortality and long-term survival[151]. Instead, the subtotal stomach-preserving pancreaticoduodenectomy is performed with the aims to preserve as much stomach as possible, minimizing the delayed gastric emptying that are associated with preserving the pyloric ring in the face of vagal denervation. In this procedure, the duodenum, pylorus, and 1 cm to 2 cm of stomach are resected with the pancreatic specimen. Although described, this modification has yet to be validated, and it is uncommonly performed[152].

The "Artery-first" approach is a surgical technique or set of techniques that have in common the dissection of the main arterial vasculature involved in pancreatic cancer, prior to performing any irreversible surgical step (transection of the pancreatic neck or bile duct division). The "Artery-first" approach has the potential to reduce blood loss and increase R0 resection rates and OS, as demonstrated in a recent meta-analysis[153].

Modified child reconstruction aims to reduce the incidence of cholangitis due to digestive reflux through hepatic-digiunal anastomosis. In case of pancreatic-digiunal anastomosis, the hepatic-digiunal anastomosis is made downstream of the previous one. In case of pancreatico-gastric anastomosis, the hepatico-digiunal anastomosis is made near the previously closed loop. Whatever the type of pancreatico-digestive anastomosis, the digestive anastomosis (gastro-digiunal or duodeno-digiunal) is made 60 cm downstream of the hepatico-digiunal anastomosis, to reduce digestive reflux into the biliary tract.

Zaishidene® WJG | https://www.wjgnet.com

Table 2 Resectability criteria				
Resectability status		Resectable	Borderline resectable	Locally advanced
Arterial Celiac artery involvement		None	$\leq 180^\circ; > 180^\circ,$ without involvement of a orta o GDA (body/tail)	>180° (head/uncinate); Solid tumor contact with CA and aorta
	SMA common hepatic artery	None	≤ 180°; Solit tumor contact without extension into CA or hepatic artery biforcation	> 180°
Venous involvement (portal vein/smv)		None; ≤ 180° contact without contour irregularity	> $180^\circ$ ; $\le 180^\circ$ with contour irregularity or thrombosis, with reconstructible PV/SMV; Solid tumor contact with IVC	Unreconstractible PV/SMV due to tumor involvement or occlusion

CA: Celiac artery; GDA: Gastroduodenal artery; IVC: Inferior vena cava; PV: Portal vein; SMA: Superior mesenteric artery; SMV: Superior mesenteric vein.

Table 3 Resectability criteria and societies				
Vessel involvement	NCCN 2019	MDACC	ACTO	AHPBA/SSAT/SSO
CA abutment (≤180°)	Borderline	Borderline	Borderline	Unresectable
CA encasement (> 180°)	Borderline (body/tail); locally advanced (head/uncinate)	Unresectable	Unresectable	Unresectable
SMA abutment (< 180°); SMA encasement (> 180°); CHA abutment or encasement	Borderline; Locally advanced; Borderline	Borderline; Unresectable; Borderline	Borderline; Unresectable; Borderline	Borderline; Unresectable; Borderline
PV/SMV encasement (> 180°) or abutment (≤ 180°) with contour abnormality	Borderline	Borderline	Borderline	Borderline

ACTO: Alliance for Clinical Trials in Oncology; AHPBA: American Hepato-Pancreato-Biliary Association; CA: Celiac artery; CHA: Common hepatic artery; MDACC: The University of Texas MD Anderson Cancer Center; NCCN: National Comprehensive Cancer Network; PV: Portal vein; SMA: Superior mesenteric artery; SMV: Superior mesenteric vein; SSAT: Society for Surgery of the Alimentary Tract; SSO: Society for Surgical Oncology.

> Post-operative pancreatic fistula (POPF) is the main and most frequent complication after pancreatic resection surgery. It is caused by leakage of pancreatic juice into the abdominal cavity, which is collected and conveyed to the outside by the drains normally placed at the end of surgery or during postoperative care if necessary. The diagnosis is made on the basis of the quality of the drainage fluid (varying from transparent to coffee-colored to brown) and the value of amylase in the fluid itself, greater than three times the normal limit of serum amylase[149,150].

> POPFs are classified into three grades based on clinical impact. Grade A fistulas do not involve any special intervention and do not significantly modify the postoperative hospital stay. Grade B fistulas require a longer postoperative stay, the retention of surgical drains, the possible placement of additional drains under radiological guidance, antibiotic therapy and the use of artificial nutrition (enteral or parenteral). In grade C fistulas, reoperation is required to resolve the complication.

> Several methods have been used to reduce the risk of pancreatic fistula, including the use of octreotide, pancreatic duct occlusion, pancreatic duct stenting, pancreaticojejunostomy, anastomosis modification, and pancreaticogastrostomy. The efficacy of octreotide in preventing POPF is still a hotly debated topic. According to a 2020 metaanalysis<sup>[154]</sup>, somatostatin analogs did not affect POPF after PD, but rather appeared to be associated with a lower rate of POPF after distal pancratectomy. Therefore, reconstruction technique is the most important factor in reducing the risk of this complication. Recently, interesting results concern the blumgart anastomosis (BA), which combines the duct-mucosal principle with the transpancreatic U-suture technique. Unlike other duct-mucosal anastomoses such as Cattell-Warren anastomosis and Kakita anastomosis, U-shaped sutures and horizontal mattress suture technique are used in BA. The difference is that Blumgart's technique involves the placement of 3 to 6 transpance atic and digestive seromuscular U-sutures to bring the pancreatic stump and jejunum closer together. A meta-analysis conducted by Ricci et al [155] demonstrated the ability of BA to reduce the risk of pancreatic fistula compared with non-blumgart duct-to-mucosal anastomoses (non-BA DtoM). The reduction seems clinically significant, with a number needed to treat of 9 which means that one pancreatic fistula can be avoided every ten patients treated with BA instead of non-BA



#### DtoM[155,156].

Indications for the preoperative treatment of jaundice in patients who are candidates for surgery are still under debate. It increases post-operative complications and should be reserved to patients with cholangitis or with bilirubin levels greater than 15 mg/dL[157].

#### Distal pancreasectomy

Distal pancreasectomy with splenectomy is the conventional operation for PDAC located in the body or tail of the pancreas. It can provide a margin-negative resection and ensure a sampling of at least 12 regional lymph nodes. A systematic review, that included 29 observational studies, found less blood loss and reduced length of hospital stay in patients operated with laparoscopic approach. However, the laparoscopic technique has some disadvantages that may lead to inadequate resection margins: Technical difficulties, inability to palpate the gland, difficulty in closing the pancreatic stump. Generally, surgeons advocate an open approach when the concern for malignancy is high, reserving laparoscopic resection for benign or premalignant indications[158-160].

Petrucciani et al[161] evaluated the prognosis of patients with positive surgical margin (R1). A better OS was observed in patients with R0 margin vs R1. However, an extension of the surgical resection following R1 pancreasectomy did not improve long term survival.

#### Total pancreasectomy

Sometimes, because of the extent or location of the tumor, a total pancreasectomy is required to achieve microscopically negative resection margins[162,163]. However, the metabolic consequences of this procedure, which include permanent exocrine insufficiency and brittle diabetes, have a detrimental impact on the quality of life and longterm survival [164]. A recent study showed a moderately reduced summary score of 76%, compared with a general population score of 86% using the EORTC QLQ-C30 questionnaire to evaluate the overall quality of life. Diarrhea is the most important symptom[165].

#### Lymphadenectomy

Tomlinson *et al*[166] evaluated the minimum number of lymph nodes removed during pancreasectomy that are essential for proper staging. They consider a number of 15 Lymph nodes as the optimal cut-off. Therefore, the cut-off of 12 lymph nodes reported by Schwarz, represents a more easily threshold value, but sufficient for correct staging.

Standard lymphadenectomy should strive to resect lymph node stations 5, 6, 8a, 12b1, 12b2, 12c, 13a, 13b, 14a, 14b, 17a, and 17b[167].

In some centres, mainly in Japan, surgeons routinely perform extensive lymph node dissection, including all 8, 9, all 12, all 14, 16a2, and 16b1 lymph nodes. A systematic review comparing standard vs extended lymphadenectomy demonstrated that there are no differences in OS between the two groups at one, three, or five years. However, the risk of complications was significantly increased after extended lymphadenectomy [168].

#### Vascular resection

If the pancreatic tumor involves the PV or SMV, pancreatic resection with PV or SMV resection may be considered: (1) When the vascular resection allows for adequate vascular flow; (2) When the tumor does not involve the SMA or hepatic artery; and (3) When an R0 resection can be accomplished. Nevertheless, many surgeons prefer to treat patients with PV or SMV involvement with neoadjuvant systemic chemotherapy before surgery.

A systematic review of 12 single-center reports concluded that pancreasectomy with PV/SMV resection is a safe and feasible procedure. It increases the number of patients who can undergo curative surgery and improves long term prognosis in a selected group of patients [169]. However, post-operative morbidity and mortality increase markedly when arterial resections are performed and few data are available to support these procedures [170-172].

#### Open vs minimally invasive approach

A systematic review identified 27 retrospective studies, including close to 7000 patients who underwent pancreasectomy (1306 minimally invasive, 5603 open)[173]. The laparoscopic approach was associated with longer operative times [mean difference (MD) 71 min], but lower intraoperative blood loss (MD -300 mL). The rate of



lymph node retrieval was significantly higher in the minimally invasive group (MD 1.34 nodes), and the likelihood of an R0 resection was also higher (odds ratio 1.45). Hospital stay, postoperative hemorrhage and wound infection were significantly lower in the laparoscopic group, while the rate of overall mortality, reoperations, vascular resection, pancreatic fistula, delayed gastric emptying and bile leak were similar between the two groups [174-176].

In some high-volume surgical centres, robotic-assisted pancreatic resection has been adopted. Experienced surgeon reported the same morbidity and mortality of open surgery. Decreased blood loss, higher number of adequate lymphadenectomy and improved gastric emptying are reported in some studies. These results may improve OS, but, because robotic-assisted pancreasectomy is still in its infancy, available longterm oncologic outcomes are limited[177-181].

# CHEMOTHERAPY FOR RESECTABLE AND BORDERLINE RESECTABLE PANCREATIC CANCER

The only treatment with curative potential for pancreatic cancer is surgery. Five-year survival ranges from 10% to 25%.

For patients with PDAC resectable or borderline resectable, surgical resection is followed by adjuvant chemotherapy. Some high-volume centers also use neoadjuvant therapy in these categories of patients [182,183].

#### Adjuvant chemotherapy

Several adjuvant chemotherapy regimens have been evaluated in randomized controlled trials[184-190]. Currently, mFFX is the recommended therapy for patients with a good performance status. Gemcitabine (+/- capecitabine) remains a treatment option for patients not sufficiently fit or with contraindications to mFFX[182,191]. Because mFFX has a high toxicity, Brown University Oncology Research Group suggests FOLFOX + nab-paclitaxel (FOLFOX-A) as an alternative[192].

According to a meta-analysis[193], S1 was ranked best for overall and disease-free survival followed by mFFX. Whilst there was no significant difference between S1 and mFFX for OS, S1 had significantly longer disease-free survival (MD 2.8 mo) and was ranked best for lowest overall and haematological grade 3/4 toxicities[194]. However, the results should be interpreted with care, as S-1 has shown good results in the Asian population, but its performance in Caucasians remains unclear due to the different expression of cytochrome P-450.

Adjuvant chemotherapy should be administered between 28 and 59 d after surgery. This timing appears to provide better survival than administering before 28 or after 59 d[182,194].

A 2020 study compared the efficacy between adjuvant chemotherapy and chemoradiation therapy in relation to AJCC stage. Monochemotherapy and combination chemotherapy + chemoradiotherapy (CRT) showed better OS and disease free survival than CRT alone in patients with AJCC stage III, whereas there was no significant difference in OS in patients with AJCC stage I/II[195].

#### Neoadjuvant chemotherapy

The main purpose of neoadjuvant chemotherapy (NACT) differs according to the stage. For patients with BR-PDAC the objective of the therapy is to decrease tumor size and to control the micro metastases. For patients with primary resectable PDAC the purpose is to increase the proportion of patients receiving chemotherapy, because half of patients undergoing surgery, do not receive adjuvant chemotherapy due to postoperative morbidity or poor general condition[196].

In 2020, important advances were made in this field. For patients with BR-PDAC several studies confirmed the benefits on R0 resection rates and survival of NACT with mFFX[197-200] or multi-agent gemcitabine[201]. Moreover, in the PREOPANC-1 trial, patients receiving neoadjuvant CRT with gemcitabine obtained the same benefits of mFFX[202]. A study of the University of Texas showed that patients who received neoadjuvant CRT had significantly improved R0 resection rates, lymph node resection rates, and locoregional recurrence rates, compared with those who received NACT [203]. Although early data suggest the importance of integrating both NACT and CRT into the treatment, large prospective trial data are lacking[204]. New evidence for a standard regimen for BR-PDAC will be established by the result of the ESPAC-5F trial (ISRCTN89500674)[205].



For primary resectable cancer, the potential benefit of NACT has been validated, particularly when initiated within 6 wk of diagnosis<sup>[206]</sup>. The SWOG S1505 study observed that patients who received gemcitabine and nab-paclitaxel had a greater pathologic response and median survival comparable to those who received mFFX [207]. Several chemotherapeutic agents for resectable pancreatic cancer are currently being studied in several RCTs[208]. The NorPACT-1 study[209] and the Panache-01 study<sup>[210]</sup> are evaluating the effect of NACT with mFFX, and the NEONAX study [211] of NACT with 2 cycles of nab-paclitaxel/ gemcitabine.

In the Asian population, treatment regimens differ. The Prep-02/JSAP-05 study study demonstrated, in patients with resectable PDAC, that NACT with gemcitabine plus S-1 (GS therapy) improves median OS compared with initial surgery (37 mo vs 27 mo). The resection rate and morbidity of surgery remain the same [212]

Based on these results, the latest Japanese guidelines recommend GS therapy as standard neoadjuvant therapy for patients with resectable PDAC. In this regimen, patients receive intravenous gemcitabine at a dose of 1000 mg/m2 on days 1 and 8, plus oral S-1, twice daily, at a dose based on body surface area (80, 100, 120 mg/d) on days 1-14 every 3 wk for 2 cycles. For patients with BR-PDAC, they recommend NACT, but have refrained from recommending any specific regimens[212-214]. Among several ongoing RCTs on treatments for borderline resectable pancreatic cancer, a Japanese trial is comparing neoadjuvant therapy with gemcitabine plus nabpaclitaxel and CRT therapy with S-1[215].

A subset of patients does not respond to NACT. There is therefore a need to find markers that can predict response to NACT. At the moment the best ones seem to be GRP78, CADM1, PGES2 and RUXF[216] (Table 4).

# CHEMOTHERAPY FOR LA PANCREATIC CANCER

Thirty to forty percent of patients with PDAC are initially diagnosed LA PDAC[182, 215]. LA PDAC is still nonmetastatic, but due to the local growth, curative resection is not possible at the time of diagnosis. Treatment involves chemotherapy with regimens that are also used in the metastatic setting, such as mFFX or gemcitabine plus nabpaclitaxel[217-219]. A small percentage of patients, with excellent response to chemotherapy, may become eligible for surgical resection. The majority have incurable disease. A systematic review of studies investigating mFFX in LA-PDAC revealed a median OS ranging from 10.0 mo to 32.7 mo[220], while in the LAPACT study, about the Nab-Paclitaxel + Gemcitabine regimen, OS 18.8 mo[221]. Recently, Kunzmann et al [222] compared two different NACT regimens, mFFX and gemcitabine plus nabpaclitaxel. The mFFX was superior in both the conversion rate to surgery (45.0% vs 30.6%) and the rate of R0 resections achieved (74% vs 68%). A subsequent study confirmed that mFFX patients had greater tumor size reduction, fewer positive lymph nodes, longer OS and distant metastasis-free survival compared to the nab-P/G patients[223].

The role of CRT for LA disease is still unclear. According to the LAP07 study, CRT improves the rate of local control but does not prolong survival in patients with LA PDAC after treatment with chemotherapy (gemcitabine with or without erlotinib) [224]. It is unclear whether these conclusions still hold true in the setting of newer combination chemotherapy regimens and improved radiation therapy techniques, such as stereotactic radiation therapy and proton therapy. The PAULA-1 study compared two cohorts of LAPDAC patients treated with stereotactic body radiotherapy (SBRT) ± chemotherapy vs CRT ± chemotherapy in terms of local control, distant metastases-free survival (DMFS), progression-free survival (PFS), OS, and toxicity. Patients treated with SBRT showed higher local control rate and similar OS, DMFS, PFS and toxicity compared to CRT[225].

## CHEMOTHERAPY FOR METASTATIC PDAC

Half of patients have metastatic disease at the time of diagnosis. The primary treatment is systemic chemotherapy, with the goal of increasing survival and palliating cancer-related symptoms. Both mFFX and gemcitabine plus nab-paclitaxel improve median OS compared to gemcitabine monotherapy [226,227]. In clinical practice, for patients who are fitter, mFFX is generally preferred, reserving gemcitabine plus nab-paclitaxel as a second-line option if they have adequate performance status[228,229]. For patients who have received first-line gemcitabine and



Table 4 Phase of trial and level of evidence of trial about chemotherapy for resectable and borderline resectable pancreatic ductal adenocarcinoma

Ref.	Phase of trial	Level of evidence
Neoptolemos et al[185]	III	II
Oettle <i>et al</i> [186]	III	Ι
Neoptolemos et al[187]	III	Ι
Neoptolemos et al[188]	III	Ι
Conroy et al[189]	III	Ι
You et al[195]	III	II
van Roessel <i>et al</i> [198]	IV	II
Versteijne et al[202]	III	II
Ghaneh et al[205]	Ш	II
Sohal et al[207]	IV	II
Labori <i>et al</i> [209]	III	II
Schwarz et al[210]	Ш	Ι
Ettrich <i>et al</i> [211]	Ш	II
Motoi et al[212]	III	II
UMIN-CTR Clinical Trial[215] (UMIN000026858)	III	П

have progressed, a good option might be the combination of fluorouracil plus leucovorin with nanoliposomal irinotecan<sup>[230]</sup>. Golan *et al*<sup>[231]</sup> evaluated patients with metastatic PDAC and BRCA1-2 germline mutation. In these patients, disease progression had not occurred during at least 4 mo of first-line platinum derivativebased chemotherapy. Patients were randomized to receive olaparib or placebo. Olaparib showed a benefit in terms of PFS and a relatively safe toxicity profile. Although AIFA has not yet approved the indication, this study suggests a role for olaparib as maintenance therapy[231].

Finally, we look forward to the results of the AVENGER 500 trial (NCT03504423) to evaluate the efficacy of mFFX with or without CPI-613. CPI613 (devimistat) is an inhibitor of pyruvate dehydrogenase and a-ketoglutarate, key enzymes of the Krebs cycle. It has already shown good results in a phase I study[232].

# STROMA-TARGETING THERAPY

Although chemotherapy is the recommended treatment for patients with advanced PDAC, its efficacy is not satisfactory. The major hurdle is considered the dense dysplastic stroma. The stroma components occupy more than 70% of the total tumor volume. The dense desmoplastic stroma of PDAC leads to vascular compression and a hypoxic microenvironment, which in turn influences drug pharmacokinetics/ pharmacodynamics. It also prevents proper action of immune system cells, which are unable to reach the target site. The result is a chemoresistant and immunoresistant tumor[233,234].

One of the major components of the PDAC stroma is hyaluronic acid (HA). HA promotes the survival, proliferation, and migration of tumor cells[235]. HA is a potential therapeutic target using pegylated hyaluronidase (PEGPH20). The HALO-109-202 study demonstrated that PEGPH20, combined with Abraxane (nab-paclitaxel) and gemcitabine, improves progression-free and OS in patients with high HA levels [236]. However, poor results were obtained from the subsequent HALO-109-301 study (NCT02715804). Another element to be acted upon is the Hedgehog signaling pathway, which is generally overactivated in pancreatic cancer. Vismodegib, in combination with gemcitabine or erlotinib, was studied for this purpose. It did not significantly affect survival compared with these two drugs administered as monotherapy[237,238].

WJG | https://www.wjgnet.com

In tumors, Angiotensin II activates transforming growth factor- $\beta$  through the AT1R and stimulates proliferation, so several angiotensin system inhibitors have been used to target PDAC stroma[233]. One study evaluated the efficacy of mFFX combined with losartan in a neoadjuvant regimen in patients with LA PDAC. The therapy was associated with an increased R0 resection rate[239].

A clinical trial evaluated the efficacy of focused ultrasound combined with gemcitabine microbubble delivery in PDAC patients. Patients treated with the combination tolerated multiple chemotherapy cycles of gemcitabine. A prolongation of median survival by almost 9 mo and, in 50% of cases, a reduction in tumor size were observed[240].

Poor results were obtained from stroma depletion in clinical settings. They are due to the fact that, although stroma-targeting therapy enhances the delivery of chemotherapeutic agents, it might also promote tumor chemoresistance and metastasis (a double-edged sword)[241]. According to several experts, future research should focus on the tumor ECM biology, biomarkers correlated with treatment benefit (as ADAM12)[242] and pharmacological agents able to alter the tumor microenvironment (TME). One of the most interesting discoveries in this regard involves clodronate liposomes. They prevent metastasis formation by inhibiting the activity of PDACassociated macrophages and altering the microenvironment of key organs that are sites of metastatic invasion. They are therefore valuable candidates to be evaluated in combination with target therapy against stroma<sup>[243]</sup>.

### IMMUNOTHERAPY

#### Immune checkpoint inhibitors

Checkpoint inhibitors activates the function "kill the tumor" of the immune system, targeting immune checkpoint molecules (PD-1, PD-L1, CTLA-4) that negatively regulate T-cell function. Although they resulted in remarkable successes in other cancers, ipilimumab, BMS-936559 and tremelimumab showed little efficacy in PDAC [244-247]. The reasons of failure of immune checkpoint inhibitors are the low baseline PD-1+ T-cell infiltration into the tumor and a paucity of neoepitopes[248,249]. Indeed, in a very small subset of PDAC patients with a high burden of microsatellite instability (MSI-high) PD-1 inhibitor is effective and was recently FDA approved[250,251].

Currently, the development of immune checkpoint inhibitors for PDAC is focused on combination therapy with chemotherapeutic agents [252-255].

#### Therapeutic cancer vaccines

Therapeutic cancer vaccines present of immunogenic tumor antigens to the immune system, resulting in activation of the anti-cancer response. GVAX is an allogeneic vaccine irradiated with tumor cells engineered to express GM-CSF. It was studied alone and in combination with CRS-207 and cyclophoshamide, however it didn't correlate with improved survival[256,257].

More promising results were instead obtained with KIF20A-66[258-260].

K-RAS vaccines have been tested in the past, but data remain unclear and with no prominent advantages in metastatic patients[261-264].

We are currently awaiting the results of some studies: (1) TLP0-001, a phase III study of a dendritic cell (DC) vaccine loaded with WT1 peptides in patients with advanced PDAC refractory to standard chemotherapy [265,266]; (2) A clinical trial using GV1001 with GM-CSF in patients with LA-PDAC in combination with gemcitabine chemotherapy, tadalafil and radiation therapy (NCT01342224); and (3) NCT01836432, NCT02405585 and NCT01072981 evaluating algenpantucel-L in combination with chemotherapy and CRT therapy. They involve patients with borderline resectable and LA unresectable PDAC.

#### CAR-T cell

CAR-T cell therapy is a type of adoptive cell therapy. CAR-T cells are T lymphocytes that are extracted from a patient's blood sample or from a donor by apheresis, genetically modified to express the receptor for chimeric antigen (CAR), and cultured in the laboratory. They are then re-infused into the patient. The resulting T cells are able to recognize tumor cells and activate the immune system response against the disease<sup>[267]</sup>. The target antigens of CAR-T cells include mesothelin, prostate stem cell antigen (PSCA), CEA, HER2, MUC-1, and CD133[268,269]. In a study of metastatic PDAC, autologous mesothelin-specific T lymphocytes improved PFS in two patients of the six examined. An additional patient had complete remission of all liver metastases



# [270].

Combination of immunotherapy drugs was experienced and showed good results over time. Le *et al*[271] compared the efficacy of Ipilimumab as monotherapy (arm 1) and Ipilimumab in combination with GVAX (arm 2) in patients with already treated PDAC. Combination therapy showed an increase in median OS (5.7 mo vs 3.6 mo) and 1-year OS (27% vs 7%). Chung et al[272] evaluated the combination of Pembrolizumab with modified p53-expressing Ankara vaccinia virus (p53MVA). Three of eleven patients experienced disease stabilization by 30, 32, and 49 wk. Good OS and PFS results were also obtained using DC and cytokine-induced killer cell immunotherapy in combination with S-1 chemotherapy, compared with chemotherapy or supportive care alone<sup>[256]</sup>.

Several trials of immunotherapy-based treatment combinations with targeted agents are ongoing for patients with pancreatic cancer [273-275].

#### **Oncolytic viruses**

Oncolytic viruses are modified therapeutic drugs that selectively infect and selfreplicate in tumor cells with tumor-dissolving effect. They also activate the anti-tumor immunity and change the TME from an immunosuppressed state to an immuneactivated state. Futhermore, oncolytic viruses have the advantages of specificity, low toxicity, and low drug resistance[276]. Adenovirus, Herpes Simplex Virus, Protoparvovirus, Reovirus and Vaccinia Virus have been tested. However most of the studies have shown unsatisfactory results. The only positive results derive from ParvOryx02 (NCT02653313). A single-arm study published in 2020 showed an encouraging efficacy of pembrolizumab in combination with Pelareorep and chemotherapy in patients progressed after first-line treatment[277-281] (Table 5).

### GENETIC MUTATION AND TARGET THERAPY

Some genetic alterations produce cellular changes in neoplastic cells that are potentially therapeutically targetable. BRAF mutations occur in 1%-3% of PDAC. They showed to be targetable in metastatic colon cancer where the combination of Encorafenib and Cetuximab has recently been approved [282,283]. Encorafenib and Cetuximab should also be evaluated in PDAC. Furthermore, pancreatic tumors with NTRK gene fusions can be treated with tropomyosin receptor kinase inhibitors [284, 285]. Similarly, some wild-type Kras pancreatic tumors hosting somatic NRG1 gene fusions respond to treatment with a kinase inhibitor of the HER family[286,287].

However, the results of the targeted therapies have been unsatisfactory, mainly due to the low life expectancy. There is no time to sequence the tumors and develop a treatment based on mutations[288].

The exceptions were the germline alterations. Patients with mutations of BRCA1, BRCA2 or PALB1 are remarkably sensitive to treatment with DNA cross-linking agents, such as platinum-based drugs, and poly(ADP-ribose) polymerase (PARP) inhibitors[289-291]. Patients with Lynch syndrome (MSI-high) respond well to treatment with immune checkpoint inhibitors[292-294] and those with ATM mutations could respond to the drugs, targeting the ATR-checkpoint kinase 1 (Chk1) pathway [295,296].

The elephant in the targeted therapy room remains Kras[297]. It has been considered "undrinkable"[297-299] because the protein lacks an efficient smallmolecule binding pocket and has a high affinity for cellular guanosine triphosphate (GTP), which is highly concentrated in the cytoplasm. Furthermore, other than the GTP/GDP binding pocket, KRAS has no other pockets for small-molecule inhibitor binding. A druggable variant of Kras appears to be G12C. Enormous progress has been made in this regard and several drugs (AMG 510, MRTX849, JNJ-74699157 and LY3499446) are currently in clinical trials[299]. The importance of these can be deduced from the fact that 95% of pancreatic cancers harbor mutations in the Kras gene (the four Kras mountains, TP53, CDKN2A and SMAD4 present in > 50% of tumors)[300,301]. Although Kras G12C mutations are only a small fraction of Kras mutations in PDAC, these drugs represent a chance to take down a previously thought invincible adversary.

#### PANCREATIC CANCER AND GUT MICROBIOTA

Recent studies have shown the gut microbiota (GM) may play a role in the



WJG https://www.wjgnet.com

Table 5 Phase and level of evidence of trials about immunotherapy for pancreatic ductal adenocarcinoma			
Ref.	Phase of trial	Level of evidence	
Royal <i>et a</i> [245]	II	II	
Brahmer <i>et al</i> [246]	Ι	Ι	
O'Reilly et al[247]	П	II	
Tumeh <i>et al</i> [248]	П	III	
Le <i>et al</i> [250]	П	II	
Le <i>et a</i> [ <mark>251</mark> ]	П	II	
Wainberg et al[252]	Ι	II	
Weiss <i>et al</i> [253]	Ib/II	II	
National Institute of Public Health[254] (JapicCTI-184230,ONO-4538)	П	II	
Wang-Gillam et al[255]	Ш	II	
Le <i>et al</i> [257]	IIb	Ι	
Asahara et al <mark>[258]</mark>	I/II	II	
Suzuki et al[259]	П	III	
Miyazawa et al[ <mark>260]</mark>	II	II	
Wedén <i>et al</i> [261]	IV	III	
Toubaji <i>et al</i> [ <mark>262</mark> ]	Ι	III	
Abou-Alfa <i>et al</i> [ <mark>263</mark> ]	I/II	III	
Cohn <i>et al</i> [264]	Ι	III	
Katsuda et al[ <mark>26</mark> 5]	III	Ι	
Katsuda et al[ <mark>266]</mark>	I/II	II	
Beatty et al[270]	Ι	III	
Le <i>et al</i> [271]	Ib	II	
Chung et al[272]	Ι	III	
Wang-Gillam et al[273]	Ι	III	
Reiss et al[274]	П	III	
Desai <i>et al</i> [275]	Ib/II	Ongoing trial	
Chang et al[278]	Ι	III	
Noonan et al[ <mark>279</mark> ]	П	II	
Mahalingam <i>et al</i> [280]	Ib	III	

development of PDAC and its response to therapy. GM alterations result in reduced mucus thickness, leading to decreased antimicrobial defenses and increased exposure to bacterial components such as LPS, flagellin, single or doubled DNA and CpG DNA. These agents activate Toll-like-receptors and trigger chronic inflammation that are related to carcinogenesis. Moreover, inflammation and dysbiosis lead to mutation of Kras, that accelerates carcinogenesis, activating nuclear factor-κB pathway[302-304].

Several bacterial products are considered potential carcinogens. Cyclomodulins promote tumorigenesis through active interference with host cell cycles. Colibactin and Bacteroides fragilis toxin act synergistically with Escherichia coli to create doublestranded DNA damage[305]. E. coli cytotoxic necrotizing factor and CagA lead to uncontrolled cell proliferation, while cytolytic distending toxin and cycle inhibitory factor participate in genetic alterations and induce hyperploidy even in the absence of cell division[306]. The presence of an Helicobacter pylori infection and high concentrations of Fusobacterium spp and Porphyromonas gingivalis (bacteria generally present in the oral cavity) are associated with an increased risk of pancreatic cancer [307-310].

WJG | https://www.wjgnet.com

Moreover, other studies correlated a large number of microbes with immune suppression, downregulation of tumor suppressive pathways and the upregulation of oncogenic pathways[311].

Dysbiosis is also related to obesity, chronic pancreatitis and diabetes, wellestablished risk factors of PDAC[312,313].

Because it participates in drug metabolism and biotransformation and immune regulation, the GM is implicated in the efficacy of chemotherapeutic agents [314]. The innate immune response activated by the GM potentiates the action of oxaliplatin [315]. Gentamicin activity may be reduced by the enzymes pyrimidine nucleoside phosphorylase and cytidine deaminase, which are produced by Gamma-proteobacteria and mycoplasmas within PDAC. Thus, these data suggest the possibility of modulating GM to counteract the chemoresistance characteristic of pancreatic cancer [316].

Intratumoral microorganisms can play a key role in anticancer therapy [317]. Indeed, they can stimulate host immune responses with positive or negative impacts on therapy. Gammaproteobacteria, Escherichia Coli and Fusobacteria are most commonly present in PDAC. Gamma proteobacteria contain the enzyme CDD which could be responsible for the ineffectiveness of gemcitabine[318]. Escheria Coli is capable of inducing chemical changes in the structure of gemcitabine, fludarabine, cladribine, and CB1954[319]. The desmoplastic response induced by tumor cells is dependent on MyD88. It is activated by Fusobacterium species.

The intratumoral microbiota thus emerges as a major proponent of the chemoimmunoresistant phenotype of pancreatic cancer and is related to long-term survival in PDAC patients.

# PROGNOSIS

The most important prognostic factor is tumor stage. The median survival time after resection for patients with stage IA, IB, IIA, IIB, and III was 38, 24, 18, 17, and 14 mo, respectively<sup>[320]</sup>. Other factors may influence the prognosis of PDAC after surgery: Surgical margin status, tumor grading, presence of lymphatic invasion, preoperative and postoperative serum levels of CA 19-9, and cigarette smoking[321-329]. Squamous subtypes have a poor prognosis. They are enriched with TP53 and KDM6A mutations, upregulation of TP63ΔN transcriptional network, hypermethylation of pancreatic endoderm cell fate determining genes[330].

Several studies have investigated novel factors influencing prognosis: (1) Increased expression of CDK1 and CCNA2 is associated with poor prognosis, although they may be potential therapeutic targets[331]; (2) The autophagy regulatory genes MET and RIPK2 play a prognostic role in PDAC[332]; (3) High expression of GPDAC2, GPDAC3 and GPDAC5 has been significantly associated with favorable survival[333]; (4) High expression of Hic-5 is negatively correlated with postoperative survival time, as Hic-5 stimulates tumor proliferation, migration, and invasion<sup>[334]</sup>; (5) PRMT1 promotes pancreatic cancer growth by increasing cellular β-catenin levels and predicts poor prognosis[335]; (6) Patients with first recurrence in the lung have a better prognosis than patients with first recurrence in the liver[336]; (7) Increased levels of ZIP4 correlate with poorer survival. ZIP4 inhibits the expression of the gemcitabine transporter ENT1, so that cells take up smaller amounts of the drug. Activation of this pathway participates in the chemoresistance of pancreatic cancers[337]; (8) The highly upregulated in liver cancer (HULC) lncRNA distinguishes patients with pancreatic cancer, patients with benign pancreatic disease, and healthy subjects and correlates with TNM stage. Subjects with low HULC expression have significantly higher 3- and 5-year OS than those with high expression. Therefore, HULC lncRNA could be considered an effective marker for the diagnosis and prognosis of PDAC[338]; (9) Upregulation of TYMS leads to unfavorable OS and RFS[339]; and (10) The GINS complex has four subunits, encoded by the GINS1, GINS2, GINS3, and GINS4 genes, all of which are overexpressed in PDAC. The expression of each member is associated with the histological grade of PDAC and is a negative prognostic marker[340].

# CONCLUSION

Pancreatic cancer is a very treacherous, dangerous enemy and the results are still very unsatisfactory. But we have not given up. Research is running fast on many paths, without losing its enthusiasm. It is proof that we are encircling it, and at the end, we



will win. The success of a fight is linked to the ability to move from one failure to another without losing one's enthusiasm.

# REFERENCES

- GBD 2017 Pancreatic Cancer Collaborators. The global, regional, and national burden of pancreatic cancer and its attributable risk factors in 195 countries and territories, 1990-2017: a systematic analysis for the Global Burden of Disease Study 2017. Lancet Gastroenterol Hepatol 2019; 4: 934-947 [PMID: 31648972 DOI: 10.1016/S2468-1253(19)30347-4]
- GBD 2015 Mortality and Causes of Death Collaborators. Global, regional, and national life 2 expectancy, all-cause mortality, and cause-specific mortality for 249 causes of death, 1980-2015: a systematic analysis for the Global Burden of Disease Study 2015. Lancet 2016; 388: 1459-1544 [PMID: 27733281 DOI: 10.1016/S0140-6736(16)31012-1]
- 3 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- 4 Ilic M, Ilic I. Epidemiology of pancreatic cancer. World J Gastroenterol 2016; 22: 9694-9705 [PMID: 27956793 DOI: 10.3748/wjg.v22.i44.9694]
- Oldfield LE, Connor AA, Gallinger S. Molecular Events in the Natural History of Pancreatic 5 Cancer. Trends Cancer 2017; 3: 336-346 [PMID: 28718411 DOI: 10.1016/j.trecan.2017.04.005]
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin 2020; 70: 7-30 [PMID: 6 31912902 DOI: 10.3322/caac.21590]
- 7 Grasso C, Jansen G, Giovannetti E. Drug resistance in pancreatic cancer: Impact of altered energy metabolism. Crit Rev Oncol Hematol 2017; 114: 139-152 [PMID: 28477742 DOI: 10.1016/j.critrevonc.2017.03.026]
- He J, Blair AB, Groot VP, Javed AA, Burkhart RA, Gemenetzis G, Hruban RH, Waters KM, Poling 8 J, Zheng L, Laheru D, Herman JM, Makary MA, Weiss MJ, Cameron JL, Wolfgang CL. Is a Pathological Complete Response Following Neoadjuvant Chemoradiation Associated With Prolonged Survival in Patients With Pancreatic Cancer? Ann Surg 2018; 268: 1-8 [PMID: 29334562 DOI: 10.1097/SLA.00000000002672]
- Singhi AD, Koay EJ, Chari ST, Maitra A. Early Detection of Pancreatic Cancer: Opportunities and 9 Challenges. Gastroenterology 2019; 156: 2024-2040 [PMID: 30721664 DOI: 10.1053/i.gastro.2019.01.259
- 10 Bosetti C, Lucenteforte E, Silverman DT, Petersen G, Bracci PM, Ji BT, Negri E, Li D, Risch HA, Olson SH, Gallinger S, Miller AB, Bueno-de-Mesquita HB, Talamini R, Polesel J, Ghadirian P, Baghurst PA, Zatonski W, Fontham E, Bamlet WR, Holly EA, Bertuccio P, Gao YT, Hassan M, Yu H, Kurtz RC, Cotterchio M, Su J, Maisonneuve P, Duell EJ, Boffetta P, La Vecchia C. Cigarette smoking and pancreatic cancer: an analysis from the International Pancreatic Cancer Case-Control Consortium (Panc4). Ann Oncol 2012; 23: 1880-1888 [PMID: 22104574 DOI: 10.1093/annonc/mdr541]
- 11 Carreras-Torres R, Johansson M, Gaborieau V, Haycock PC, Wade KH, Relton CL, Martin RM, Davey Smith G, Brennan P. The Role of Obesity, Type 2 Diabetes, and Metabolic Factors in Pancreatic Cancer: A Mendelian Randomization Study. J Natl Cancer Inst 2017; 109 [PMID: 28954281 DOI: 10.1093/jnci/djx012]
- 12 Li D, Morris JS, Liu J, Hassan MM, Day RS, Bondy ML, Abbruzzese JL. Body mass index and risk, age of onset, and survival in patients with pancreatic cancer. JAMA 2009; 301: 2553-2562 [PMID: 19549972 DOI: 10.1001/jama.2009.886]
- O'Rorke MA, Cantwell MM, Cardwell CR, Mulholland HG, Murray LJ. Can physical activity 13 modulate pancreatic cancer risk? Int J Cancer 2010; 126: 2957-2968 [PMID: 19856317 DOI: 10.1002/iic.24997]
- 14 Huang J, Magnusson M, Törner A, Ye W, Duberg AS. Risk of pancreatic cancer among individuals with hepatitis C or hepatitis B virus infection: a nationwide study in Sweden. Br J Cancer 2013; 109: 2917-2923 [PMID: 24178755 DOI: 10.1038/bjc.2013.689]
- 15 Amundadottir L, Kraft P, Stolzenberg-Solomon RZ, Fuchs CS, Petersen GM, Arslan AA, Buenode-Mesquita HB, Gross M, Helzlsouer K, Jacobs EJ, LaCroix A, Zheng W, Albanes D, Bamlet W, Berg CD, Berrino F, Bingham S, Buring JE, Bracci PM, Canzian F, Clavel-Chapelon F, Clipp S, Cotterchio M, de Andrade M, Duell EJ, Fox JW Jr, Gallinger S, Gaziano JM, Giovannucci EL, Goggins M, González CA, Hallmans G, Hankinson SE, Hassan M, Holly EA, Hunter DJ, Hutchinson A, Jackson R, Jacobs KB, Jenab M, Kaaks R, Klein AP, Kooperberg C, Kurtz RC, Li D, Lynch SM, Mandelson M, McWilliams RR, Mendelsohn JB, Michaud DS, Olson SH, Overvad K, Patel AV, Peeters PH, Rajkovic A, Riboli E, Risch HA, Shu XO, Thomas G, Tobias GS, Trichopoulos D, Van Den Eeden SK, Virtamo J, Wactawski-Wende J, Wolpin BM, Yu H, Yu K, Zeleniuch-Jacquotte A, Chanock SJ, Hartge P, Hoover RN. Genome-wide association study identifies variants in the ABO locus associated with susceptibility to pancreatic cancer. Nat Genet 2009; 41: 986-990 [PMID: 19648918 DOI: 10.1038/ng.429]
- 16 Yamada A, Komaki Y, Komaki F, Micic D, Zullow S, Sakuraba A. Risk of gastrointestinal cancers in patients with cystic fibrosis: a systematic review and meta-analysis. Lancet Oncol 2018; 19: 758-



767 [PMID: 29706374 DOI: 10.1016/S1470-2045(18)30188-8]

- 17 Pergolini I, Sahora K, Ferrone CR, Morales-Oyarvide V, Wolpin BM, Mucci LA, Brugge WR, Mino-Kenudson M, Patino M, Sahani DV, Warshaw AL, Lillemoe KD, Fernández-Del Castillo C. Long-term Risk of Pancreatic Malignancy in Patients With Branch Duct Intraductal Papillary Mucinous Neoplasm in a Referral Center. Gastroenterology 2017; 153: 1284-1294.e1 [PMID: 28739282 DOI: 10.1053/j.gastro.2017.07.019]
- 18 IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Personal habits and indoor combustions. Volume 100 E. A review of human carcinogens. IARC Monogr Eval Carcinog Risks Hum 2012; 100: 1-538 [PMID: 23193840]
- 19 Riboli E, Hunt KJ, Slimani N, Ferrari P, Norat T, Fahey M, Charrondière UR, Hémon B, Casagrande C, Vignat J, Overvad K, Tjønneland A, Clavel-Chapelon F, Thiébaut A, Wahrendorf J, Boeing H, Trichopoulos D, Trichopoulou A, Vineis P, Palli D, Bueno-De-Mesquita HB, Peeters PH, Lund E, Engeset D, González CA, Barricarte A, Berglund G, Hallmans G, Day NE, Key TJ, Kaaks R, Saracci R. European Prospective Investigation into Cancer and Nutrition (EPIC): study populations and data collection. Public Health Nutr 2002; 5: 1113-1124 [PMID: 12639222 DOI: 10.1079/PHN2002394]
- 20 McKenzie F, Biessy C, Ferrari P, Freisling H, Rinaldi S, Chajès V, Dahm CC, Overvad K, Dossus L, Lagiou P, Trichopoulos D, Trichopoulou A, Bueno-de-Mesquita HB, May A, Peeters PH, Weiderpass E, Sanchez MJ, Navarro C, Ardanaz E, Ericson U, Wirfält E, Travis RC, Romieu I. Healthy Lifestyle and Risk of Cancer in the European Prospective Investigation Into Cancer and Nutrition Cohort Study. Medicine (Baltimore) 2016; 95: e2850 [PMID: 27100409 DOI: 10.1097/MD.000000000028501
- 21 Chajès V, Biessy C, Byrnes G, Deharveng G, Saadatian-Elahi M, Jenab M, Peeters PH, Ocké M, Bueno-de-Mesquita HB, Johansson I, Hallmans G, Manjer J, Wirfält E, Jakszyn P, González CA, Huerta JM, Martinez C, Amiano P, Suárez LR, Ardanaz E, Tjønneland A, Halkjaer J, Overvad K, Jakobsen MU, Berrino F, Pala V, Palli D, Tumino R, Vineis P, de Magistris MS, Spencer EA, Crowe FL, Bingham S, Khaw KT, Linseisen J, Rohrmann S, Boeing H, Nöethlings U, Olsen KS, Skeie G, Lund E, Trichopoulou A, Zilis D, Oustoglou E, Clavel-Chapelon F, Riboli E, Slimani N. Ecologicallevel associations between highly processed food intakes and plasma phospholipid elaidic acid concentrations: results from a cross-sectional study within the European prospective investigation into cancer and nutrition (EPIC). Nutr Cancer 2011; 63: 1235-1250 [PMID: 22043987 DOI: 10.1080/01635581.2011.617530]
- 22 Saadatian-Elahi M, Slimani N, Chajès V, Jenab M, Goudable J, Biessy C, Ferrari P, Byrnes G, Autier P, Peeters PH, Ocké M, Bueno de Mesquita B, Johansson I, Hallmans G, Manjer J, Wirfält E, González CA, Navarro C, Martinez C, Amiano P, Suárez LR, Ardanaz E, Tjønneland A, Halkjaer J, Overvad K, Jakobsen MU, Berrino F, Pala V, Palli D, Tumino R, Vineis P, Santucci de Magistris M, Spencer EA, Crowe FL, Bingham S, Khaw KT, Linseisen J, Rohrmann S, Boeing H, Noethlings U, Olsen KS, Skeie G, Lund E, Trichopoulou A, Oustoglou E, Clavel-Chapelon F, Riboli E. Plasma phospholipid fatty acid profiles and their association with food intakes: results from a cross-sectional study within the European Prospective Investigation into Cancer and Nutrition. Am J Clin Nutr 2009; 89: 331-346 [PMID: 19056549 DOI: 10.3945/ajcn.2008.26834]
- 23 Jiao L, Mitrou PN, Reedy J, Graubard BI, Hollenbeck AR, Schatzkin A, Stolzenberg-Solomon R. A combined healthy lifestyle score and risk of pancreatic cancer in a large cohort study. Arch Intern Med 2009; 169: 764-770 [PMID: 19398688 DOI: 10.1001/archinternmed.2009.46]
- Ferrari P, Licaj I, Muller DC, Kragh Andersen P, Johansson M, Boeing H, Weiderpass E, Dossus 24 L, Dartois L, Fagherazzi G, Bradbury KE, Khaw KT, Wareham N, Duell EJ, Barricarte A, Molina-Montes E, Sanchez CN, Arriola L, Wallström P, Tjønneland A, Olsen A, Trichopoulou A, Benetou V, Trichopoulos D, Tumino R, Agnoli C, Sacerdote C, Palli D, Li K, Kaaks R, Peeters P, Beulens JW, Nunes L, Gunter M, Norat T, Overvad K, Brennan P, Riboli E, Romieu I. Lifetime alcohol use and overall and cause-specific mortality in the European Prospective Investigation into Cancer and nutrition (EPIC) study. BMJ Open 2014; 4: e005245 [PMID: 24993766 DOI: 10.1136/bmjopen-2014-005245]
- 25 Naudin S, Li K, Jaouen T, Assi N, Kyrø C, Tjønneland A, Overvad K, Boutron-Ruault MC, Rebours V, Védié AL, Boeing H, Kaaks R, Katzke V, Bamia C, Naska A, Trichopoulou A, Berrino F, Tagliabue G, Palli D, Panico S, Tumino R, Sacerdote C, Peeters PH, Bueno-de-Mesquita HBA, Weiderpass E, Gram IT, Skeie G, Chirlaque MD, Rodríguez-Barranco M, Barricarte A, Quirós JR, Dorronsoro M, Johansson I, Sund M, Sternby H, Bradbury KE, Wareham N, Riboli E, Gunter M, Brennan P, Duell EJ, Ferrari P. Lifetime and baseline alcohol intakes and risk of pancreatic cancer in the European Prospective Investigation into Cancer and Nutrition study. Int J Cancer 2018; 143: 801-812 [PMID: 29524225 DOI: 10.1002/ijc.31367]
- 26 Molina-Montes E, Sánchez MJ, Buckland G, Bueno-de-Mesquita HB, Weiderpass E, Amiano P, Wark PA, Kühn T, Katzke V, Huerta JM, Ardanaz E, Quirós JR, Affret A, His M, Boutron-Ruault MC, Peeters PH, Ye W, Sund M, Boeing H, Iqbal K, Ohlsson B, Sonestedt E, Tjønneland A, Petersen KE, Travis RC, Skeie G, Agnoli C, Panico S, Palli D, Tumino R, Sacerdote C, Freisling H, Huybrechts I, Overvad K, Trichopoulou A, Bamia C, Vasilopoulou E, Wareham N, Khaw KT, Cross AJ, Ward HA, Riboli E, Duell EJ. Mediterranean diet and risk of pancreatic cancer in the European Prospective Investigation into Cancer and Nutrition cohort. Br J Cancer 2017; 116: 811-820 [PMID: 28170373 DOI: 10.1038/bjc.2017.14]
- 27 Rawla P, Thandra KC, Sunkara T. Pancreatic cancer and obesity: epidemiology, mechanism, and



preventive strategies. Clin J Gastroenterol 2019; 12: 285-291 [PMID: 30788774 DOI: 10.1007/s12328-019-00953-3]

- Genkinger JM, Kitahara CM, Bernstein L, Berrington de Gonzalez A, Brotzman M, Elena JW, 28 Giles GG, Hartge P, Singh PN, Stolzenberg-Solomon RZ, Weiderpass E, Adami HO, Anderson KE, Beane-Freeman LE, Buring JE, Fraser GE, Fuchs CS, Gapstur SM, Gaziano JM, Helzlsouer KJ, Lacey JV Jr, Linet MS, Liu JJ, Park Y, Peters U, Purdue MP, Robien K, Schairer C, Sesso HD, Visvanathan K, White E, Wolk A, Wolpin BM, Zeleniuch-Jacquotte A, Jacobs EJ. Central adiposity, obesity during early adulthood, and pancreatic cancer mortality in a pooled analysis of cohort studies. Ann Oncol 2015; 26: 2257-2266 [PMID: 26347100 DOI: 10.1093/annonc/mdv355]
- 29 Song S, Wang B, Zhang X, Hao L, Hu X, Li Z, Sun S. Long-Term Diabetes Mellitus Is Associated with an Increased Risk of Pancreatic Cancer: A Meta-Analysis. PLoS One 2015; 10: e0134321 [PMID: 26222906 DOI: 10.1371/journal.pone.0134321]
- 30 Bimonte S, Cascella M, Leongito M, Palaia R, Caliendo D, Izzo F, Cuomo A. An overview of preclinical studies on the effects of (-)-epigallocatechin-3-gallate, a catechin found in green tea, in treatment of pancreatic cancer. Recenti Prog Med 2017; 108: 282-287 [PMID: 28631776 DOI: 10.1701/2715.27715
- Nasir A, Bullo MMH, Ahmed Z, Imtiaz A, Yaqoob E, Jadoon M, Ahmed H, Afreen A, Yaqoob S. 31 Nutrigenomics: Epigenetics and cancer prevention: A comprehensive review. Crit Rev Food Sci Nutr 2020; 60: 1375-1387 [PMID: 30729798 DOI: 10.1080/10408398.2019.1571480]
- 32 Pelaez-Luna M, Takahashi N, Fletcher JG, Chari ST. Resectability of presymptomatic pancreatic cancer and its relationship to onset of diabetes: a retrospective review of CT scans and fasting glucose values prior to diagnosis. Am J Gastroenterol 2007; 102: 2157-2163 [PMID: 17897335 DOI: 10.1111/j.1572-0241.2007.01480.x]
- 33 US Preventive Services Task Force, Owens DK, Davidson KW, Krist AH, Barry MJ, Cabana M, Caughey AB, Curry SJ, Doubeni CA, Epling JW Jr, Kubik M, Landefeld CS, Mangione CM, Pbert L, Silverstein M, Simon MA, Tseng CW, Wong JB. Screening for Pancreatic Cancer: US Preventive Services Task Force Reaffirmation Recommendation Statement. JAMA 2019; 322: 438-444 [PMID: 31386141 DOI: 10.1001/jama.2019.10232]
- 34 Yuan C, Babic A, Khalaf N, Nowak JA, Brais LK, Rubinson DA, Ng K, Aguirre AJ, Pandharipande PV, Fuchs CS, Giovannucci EL, Stampfer MJ, Rosenthal MH, Sander C, Kraft P, Wolpin BM. Diabetes, Weight Change, and Pancreatic Cancer Risk. JAMA Oncol 2020; 6: e202948 [PMID: 32789511 DOI: 10.1001/jamaoncol.2020.2948]
- 35 Naudin S, Viallon V, Hashim D, Freisling H, Jenab M, Weiderpass E, Perrier F, McKenzie F, Bueno-de-Mesquita HB, Olsen A, Tjønneland A, Dahm CC, Overvad K, Mancini FR, Rebours V, Boutron-Ruault MC, Katzke V, Kaaks R, Bergmann M, Boeing H, Peppa E, Karakatsani A, Trichopoulou A, Pala V, Masala G, Panico S, Tumino R, Sacerdote C, May AM, van Gils CH, Rylander C, Borch KB, Chirlaque López MD, Sánchez MJ, Ardanaz E, Quirós JR, Amiano Exezarreta P, Sund M, Drake I, Regnér S, Travis RC, Wareham N, Aune D, Riboli E, Gunter MJ, Duell EJ, Brennan P, Ferrari P. Healthy lifestyle and the risk of pancreatic cancer in the EPIC study. Eur J Epidemiol 2020; 35: 975-986 [PMID: 31564045 DOI: 10.1007/s10654-019-00559-6]
- Lal G, Liu G, Schmocker B, Kaurah P, Ozcelik H, Narod SA, Redston M, Gallinger S. Inherited predisposition to pancreatic adenocarcinoma: role of family history and germ-line p16, BRCA1, and BRCA2 mutations. Cancer Res 2000; 60: 409-416 [PMID: 10667595]
- 37 Holter S, Borgida A, Dodd A, Grant R, Semotiuk K, Hedley D, Dhani N, Narod S, Akbari M, Moore M, Gallinger S. Germline BRCA Mutations in a Large Clinic-Based Cohort of Patients With Pancreatic Adenocarcinoma. J Clin Oncol 2015; 33: 3124-3129 [PMID: 25940717 DOI: 10.1200/JCO.2014.59.7401
- Goldstein AM, Fraser MC, Struewing JP, Hussussian CJ, Ranade K, Zametkin DP, Fontaine LS, 38 Organic SM, Dracopoli NC, Clark WH Jr. Increased risk of pancreatic cancer in melanoma-prone kindreds with p16INK4 mutations. N Engl J Med 1995; 333: 970-974 [PMID: 7666916 DOI: 10.1056/NEJM199510123331504
- van Lier MG, Wagner A, Mathus-Vliegen EM, Kuipers EJ, Steyerberg EW, van Leerdam ME. 39 High cancer risk in Peutz-Jeghers syndrome: a systematic review and surveillance recommendations. Am J Gastroenterol 2010; 105: 1258-64; author reply 1265 [PMID: 20051941 DOI: 10.1038/ajg.2009.725]
- Park JG, Park YJ, Wijnen JT, Vasen HF. Gene-environment interaction in hereditary nonpolyposis 40 colorectal cancer with implications for diagnosis and genetic testing. Int J Cancer 1999; 82: 516-519 [PMID: 10404064 DOI: 10.1002/(sici)1097-0215(19990812)82:4<516::aid-ijc8>3.0.co;2-u]
- Whitcomb DC. Inflammation and Cancer V. Chronic pancreatitis and pancreatic cancer. Am J 41 Physiol Gastrointest Liver Physiol 2004; 287: G315-G319 [PMID: 15246966 DOI: 10.1152/ajpgi.00115.2004]
- 42 Yurgelun MB, Chittenden AB, Morales-Oyarvide V, Rubinson DA, Dunne RF, Kozak MM, Qian ZR, Welch MW, Brais LK, Da Silva A, Bui JL, Yuan C, Li T, Li W, Masuda A, Gu M, Bullock AJ, Chang DT, Clancy TE, Linehan DC, Findeis-Hosey JJ, Doyle LA, Thorner AR, Ducar MD, Wollison BM, Khalaf N, Perez K, Syngal S, Aguirre AJ, Hahn WC, Meyerson ML, Fuchs CS, Ogino S, Hornick JL, Hezel AF, Koong AC, Nowak JA, Wolpin BM. Germline cancer susceptibility gene variants, somatic second hits, and survival outcomes in patients with resected pancreatic cancer. Genet Med 2019; 21: 213-223 [PMID: 29961768 DOI: 10.1038/s41436-018-0009-5]
- Stoffel EM, McKernin SE, Brand R, Canto M, Goggins M, Moravek C, Nagarajan A, Petersen GM, 43



Simeone DM, Yurgelun M, Khorana AA. Evaluating Susceptibility to Pancreatic Cancer: ASCO Provisional Clinical Opinion. J Clin Oncol 2019; 37: 153-164 [PMID: 30457921 DOI: 10.1200/JCO.18.01489

- 44 Ohmoto A, Yachida S, Morizane C. Genomic Features and Clinical Management of Patients with Hereditary Pancreatic Cancer Syndromes and Familial Pancreatic Cancer. Int J Mol Sci 2019; 20 [PMID: 30699894 DOI: 10.3390/ijms20030561]
- Goggins M, Overbeek KA, Brand R, Syngal S, Del Chiaro M, Bartsch DK, Bassi C, Carrato A, 45 Farrell J, Fishman EK, Fockens P, Gress TM, van Hooft JE, Hruban RH, Kastrinos F, Klein A, Lennon AM, Lucas A, Park W, Rustgi A, Simeone D, Stoffel E, Vasen HFA, Cahen DL, Canto MI, Bruno M; International Cancer of the Pancreas Screening (CAPS) consortium. Management of patients with increased risk for familial pancreatic cancer: updated recommendations from the International Cancer of the Pancreas Screening (CAPS) Consortium. Gut 2020; 69: 7-17 [PMID: 31672839 DOI: 10.1136/gutjnl-2019-319352]
- Syngal S, Brand RE, Church JM, Giardiello FM, Hampel HL, Burt RW; American College of Gastroenterology. ACG clinical guideline: Genetic testing and management of hereditary gastrointestinal cancer syndromes. Am J Gastroenterol 2015; 110: 223-62; quiz 263 [PMID: 25645574 DOI: 10.1038/ajg.2014.435]
- 47 Brand RE, Lerch MM, Rubinstein WS, Neoptolemos JP, Whitcomb DC, Hruban RH, Brentnall TA, Lynch HT, Canto MI; Participants of the Fourth International Symposium of Inherited Diseases of the Pancreas. Advances in counselling and surveillance of patients at risk for pancreatic cancer. Gut 2007; 56: 1460-1469 [PMID: 17872573 DOI: 10.1136/gut.2006.108456]
- 48 Aslanian HR, Lee JH, Canto MI. AGA Clinical Practice Update on Pancreas Cancer Screening in High-Risk Individuals: Expert Review. Gastroenterology 2020; 159: 358-362 [PMID: 32416142 DOI: 10.1053/i.gastro.2020.03.088]
- 49 Hirono S, Yamaue H. Surgical strategy for intraductal papillary mucinous neoplasms of the pancreas. Surg Today 2020; 50: 50-55 [PMID: 31807871 DOI: 10.1007/s00595-019-01931-5]
- Canto MI, Hruban RH, Fishman EK, Kamel IR, Schulick R, Zhang Z, Topazian M, Takahashi N, 50 Fletcher J, Petersen G, Klein AP, Axilbund J, Griffin C, Syngal S, Saltzman JR, Mortele KJ, Lee J, Tamm E, Vikram R, Bhosale P, Margolis D, Farrell J, Goggins M; American Cancer of the Pancreas Screening (CAPS) Consortium. Frequent detection of pancreatic lesions in asymptomatic high-risk individuals. Gastroenterology 2012; 142: 796-804; quiz e14 [PMID: 22245846 DOI: 10.1053/j.gastro.2012.01.005]
- 51 Grover S, Syngal S. Hereditary pancreatic cancer. Gastroenterology 2010; 139: 1076-1080, 1080.e1 [PMID: 20727885 DOI: 10.1053/j.gastro.2010.08.012]
- 52 Matthaei H, Schulick RD, Hruban RH, Maitra A. Cystic precursors to invasive pancreatic cancer. Nat Rev Gastroenterol Hepatol 2011; 8: 141-150 [PMID: 21383670 DOI: 10.1038/nrgastro.2011.2]
- 53 Del Chiaro M, Verbeke CS, Kartalis N, Pozzi Mucelli R, Gustafsson P, Hansson J, Haas SL, Segersvärd R, Andren-Sandberg Å, Löhr JM. Short-term Results of a Magnetic Resonance Imaging-Based Swedish Screening Program for Individuals at Risk for Pancreatic Cancer. JAMA Surg 2015; 150: 512-518 [PMID: 25853369 DOI: 10.1001/jamasurg.2014.3852]
- 54 Tanaka M, Fernández-del Castillo C, Adsay V, Chari S, Falconi M, Jang JY, Kimura W, Levy P, Pitman MB, Schmidt CM, Shimizu M, Wolfgang CL, Yamaguchi K, Yamao K; International Association of Pancreatology. International consensus guidelines 2012 for the management of IPMN and MCN of the pancreas. Pancreatology 2012; 12: 183-197 [PMID: 22687371 DOI: 10.1016/j.pan.2012.04.004]
- 55 Canto MI, Kerdsirichairat T, Yeo CJ, Hruban RH, Shin EJ, Almario JA, Blackford A, Ford M, Klein AP, Javed AA, Lennon AM, Zaheer A, Kamel IR, Fishman EK, Burkhart R, He J, Makary M, Weiss MJ, Schulick RD, Goggins MG, Wolfgang CL. Surgical Outcomes After Pancreatic Resection of Screening-Detected Lesions in Individuals at High Risk for Developing Pancreatic Cancer. J Gastrointest Surg 2020; 24: 1101-1110 [PMID: 31197699 DOI: 10.1007/s11605-019-04230-z]
- 56 Lennon AM, Wolfgang CL, Canto MI, Klein AP, Herman JM, Goggins M, Fishman EK, Kamel I, Weiss MJ, Diaz LA, Papadopoulos N, Kinzler KW, Vogelstein B, Hruban RH. The early detection of pancreatic cancer: what will it take to diagnose and treat curable pancreatic neoplasia? Cancer Res 2014; 74: 3381-3389 [PMID: 24924775 DOI: 10.1158/0008-5472.CAN-14-0734]
- 57 Goonetilleke KS, Siriwardena AK. Systematic review of carbohydrate antigen (CA 19-9) as a biochemical marker in the diagnosis of pancreatic cancer. Eur J Surg Oncol 2007; 33: 266-270 [PMID: 17097848 DOI: 10.1016/j.ejso.2006.10.004]
- 58 Rawla P, Sunkara T, Gaduputi V. Epidemiology of Pancreatic Cancer: Global Trends, Etiology and Risk Factors. World J Oncol 2019; 10: 10-27 [PMID: 30834048 DOI: 10.14740/wjon1166]
- 59 Meng Q, Shi S, Liang C, Liang D, Xu W, Ji S, Zhang B, Ni Q, Xu J, Yu X. Diagnostic and prognostic value of carcinoembryonic antigen in pancreatic cancer: a systematic review and metaanalysis. Onco Targets Ther 2017; 10: 4591-4598 [PMID: 28979147 DOI: 10.2147/OTT.S145708]
- Ni XG, Bai XF, Mao YL, Shao YF, Wu JX, Shan Y, Wang CF, Wang J, Tian YT, Liu Q, Xu DK, 60 Zhao P. The clinical value of serum CEA, CA19-9, and CA242 in the diagnosis and prognosis of pancreatic cancer. Eur J Surg Oncol 2005; 31: 164-169 [PMID: 15698733 DOI: 10.1016/j.ejso.2004.09.007
- Khomiak A, Brunner M, Kordes M, Lindblad S, Miksch RC, Öhlund D, Regel I. Recent 61 Discoveries of Diagnostic, Prognostic and Predictive Biomarkers for Pancreatic Cancer. Cancers (Basel) 2020; 12 [PMID: 33147766 DOI: 10.3390/cancers12113234]



- 62 Satake K, Takeuchi T. Comparison of CA19-9 with other tumor markers in the diagnosis of cancer of the pancreas. Pancreas 1994; 9: 720-724 [PMID: 7846015 DOI: 10.1097/00006676-199411000-00008
- 63 Bussom S, Saif MW. Methods and rationale for the early detection of pancreatic cancer. Highlights from the "2010 ASCO Gastrointestinal Cancers Symposium". Orlando, FL, USA. January 22-24, 2010. JOP 2010; 11: 128-130 [PMID: 20208319]
- Eissa MAL, Lerner L, Abdelfatah E, Shankar N, Canner JK, Hasan NM, Yaghoobi V, Huang B, 64 Kerner Z, Takaesu F, Wolfgang C, Kwak R, Ruiz M, Tam M, Pisanic TR 2nd, Iacobuzio-Donahue CA, Hruban RH, He J, Wang TH, Wood LD, Sharma A, Ahuja N. Promoter methylation of ADAMTS1 and BNC1 as potential biomarkers for early detection of pancreatic cancer in blood. Clin Epigenetics 2019; 11: 59 [PMID: 30953539 DOI: 10.1186/s13148-019-0650-0]
- Shen SY, Singhania R, Fehringer G, Chakravarthy A, Roehrl MHA, Chadwick D, Zuzarte PC, 65 Borgida A, Wang TT, Li T, Kis O, Zhao Z, Spreafico A, Medina TDS, Wang Y, Roulois D, Ettayebi I, Chen Z, Chow S, Murphy T, Arruda A, O'Kane GM, Liu J, Mansour M, McPherson JD, O'Brien C, Leighl N, Bedard PL, Fleshner N, Liu G, Minden MD, Gallinger S, Goldenberg A, Pugh TJ, Hoffman MM, Bratman SV, Hung RJ, De Carvalho DD. Sensitive tumour detection and classification using plasma cell-free DNA methylomes. Nature 2018; 563: 579-583 [PMID: 30429608 DOI: 10.1038/s41586-018-0703-0]
- Cirmena G, Dameri M, Ravera F, Fregatti P, Ballestrero A, Zoppoli G. Assessment of Circulating 66 Nucleic Acids in Cancer: From Current Status to Future Perspectives and Potential Clinical Applications. Cancers (Basel) 2021; 13 [PMID: 34298675 DOI: 10.3390/cancers13143460]
- 67 Bloomston M, Frankel WL, Petrocca F, Volinia S, Alder H, Hagan JP, Liu CG, Bhatt D, Taccioli C, Croce CM. MicroRNA expression patterns to differentiate pancreatic adenocarcinoma from normal pancreas and chronic pancreatitis. JAMA 2007; 297: 1901-1908 [PMID: 17473300 DOI: 10.1001/jama.297.17.1901]
- Duell EJ, Lujan-Barroso L, Sala N, Deitz McElyea S, Overvad K, Tjonneland A, Olsen A, 68 Weiderpass E, Busund LT, Moi L, Muller D, Vineis P, Aune D, Matullo G, Naccarati A, Panico S, Tagliabue G, Tumino R, Palli D, Kaaks R, Katzke VA, Boeing H, Bueno-de-Mesquita HBA, Peeters PH, Trichopoulou A, Lagiou P, Kotanidou A, Travis RC, Wareham N, Khaw KT, Ramon Quiros J, Rodríguez-Barranco M, Dorronsoro M, Chirlaque MD, Ardanaz E, Severi G, Boutron-Ruault MC, Rebours V, Brennan P, Gunter M, Scelo G, Cote G, Sherman S, Korc M. Plasma microRNAs as biomarkers of pancreatic cancer risk in a prospective cohort study. Int J Cancer 2017; 141: 905-915 [PMID: 28542740 DOI: 10.1002/ijc.30790]
- Liu J, Gao J, Du Y, Li Z, Ren Y, Gu J, Wang X, Gong Y, Wang W, Kong X. Combination of plasma microRNAs with serum CA19-9 for early detection of pancreatic cancer. Int J Cancer 2012; 131: 683-691 [PMID: 21913185 DOI: 10.1002/ijc.26422]
- 70 Permuth JB, Chen DT, Yoder SJ, Li J, Smith AT, Choi JW, Kim J, Balagurunathan Y, Jiang K, Coppola D, Centeno BA, Klapman J, Hodul P, Karreth FA, Trevino JG, Merchant N, Magliocco A, Malafa MP, Gillies R. Linc-ing Circulating Long Non-coding RNAs to the Diagnosis and Malignant Prediction of Intraductal Papillary Mucinous Neoplasms of the Pancreas. Sci Rep 2017; 7: 10484 [PMID: 28874676 DOI: 10.1038/s41598-017-09754-5]
- 71 Ding J, Li Y, Zhang Y, Fan B, Li Q, Zhang J. Identification of key lncRNAs in the tumorigenesis of intraductal pancreatic mucinous neoplasm by coexpression network analysis. Cancer Med 2020; 9: 3840-3851 [PMID: 32239802 DOI: 10.1002/cam4.2927]
- Nasca V, Chiaravalli M, Piro G, Esposito A, Salvatore L, Tortora G, Corbo V, Carbone C. 72 Intraductal Pancreatic Mucinous Neoplasms: A Tumor-Biology Based Approach for Risk Stratification. Int J Mol Sci 2020; 21 [PMID: 32887490 DOI: 10.3390/ijms21176386]
- 73 Kanda M, Knight S, Topazian M, Syngal S, Farrell J, Lee J, Kamel I, Lennon AM, Borges M, Young A, Fujiwara S, Seike J, Eshleman J, Hruban RH, Canto MI, Goggins M. Mutant GNAS detected in duodenal collections of secretin-stimulated pancreatic juice indicates the presence or emergence of pancreatic cysts. Gut 2013; 62: 1024-1033 [PMID: 22859495 DOI: 10.1136/gutjnl-2012-302823]
- Kanda M, Sadakari Y, Borges M, Topazian M, Farrell J, Syngal S, Lee J, Kamel I, Lennon AM, 74 Knight S, Fujiwara S, Hruban RH, Canto MI, Goggins M. Mutant TP53 in duodenal samples of pancreatic juice from patients with pancreatic cancer or high-grade dysplasia. Clin Gastroenterol Hepatol 2013; 11: 719-30.e5 [PMID: 23200980 DOI: 10.1016/j.cgh.2012.11.016]
- Singhi AD, McGrath K, Brand RE, Khalid A, Zeh HJ, Chennat JS, Fasanella KE, Papachristou GI, 75 Slivka A, Bartlett DL, Dasyam AK, Hogg M, Lee KK, Marsh JW, Monaco SE, Ohori NP, Pingpank JF, Tsung A, Zureikat AH, Wald AI, Nikiforova MN. Preoperative next-generation sequencing of pancreatic cvst fluid is highly accurate in cvst classification and detection of advanced neoplasia. Gut 2018; 67: 2131-2141 [PMID: 28970292 DOI: 10.1136/gutjnl-2016-313586]
- 76 Suenaga M, Dudley B, Karloski E, Borges M, Irene Canto M, Brand RE, Goggins M. The Effect of Pancreatic Juice Collection Time on the Detection of KRAS Mutations. Pancreas 2018; 47: 35-39 [PMID: 29200129 DOI: 10.1097/MPA.000000000000956]
- 77 Garcia-Carracedo D, Chen ZM, Qiu W, Huang AS, Tang SM, Hruban RH, Su GH. PIK3CA mutations in mucinous cystic neoplasms of the pancreas. Pancreas 2014; 43: 245-249 [PMID: 24518503 DOI: 10.1097/MPA.00000000000034]
- 78 Garcia-Carracedo D, Turk AT, Fine SA, Akhavan N, Tweel BC, Parsons R, Chabot JA, Allendorf JD, Genkinger JM, Remotti HE, Su GH. Loss of PTEN expression is associated with poor prognosis



in patients with intraductal papillary mucinous neoplasms of the pancreas. Clin Cancer Res 2013; 19: 6830-6841 [PMID: 24132918 DOI: 10.1158/1078-0432.CCR-13-0624]

- 79 Kaur S, Kumar S, Momi N, Sasson AR, Batra SK. Mucins in pancreatic cancer and its microenvironment. Nat Rev Gastroenterol Hepatol 2013; 10: 607-620 [PMID: 23856888 DOI: 10.1038/nrgastro.2013.120]
- Nagata K, Horinouchi M, Saitou M, Higashi M, Nomoto M, Goto M, Yonezawa S. Mucin 80 expression profile in pancreatic cancer and the precursor lesions. J Hepatobiliary Pancreat Surg 2007; 14: 243-254 [PMID: 17520199 DOI: 10.1007/s00534-006-1169-2]
- Moniaux N, Chaturvedi P, Varshney GC, Meza JL, Rodriguez-Sierra JF, Aubert JP, Batra SK. 81 Human MUC4 mucin induces ultra-structural changes and tumorigenicity in pancreatic cancer cells. Br J Cancer 2007; 97: 345-357 [PMID: 17595659 DOI: 10.1038/sj.bjc.6603868]
- 82 Gold DV, Modrak DE, Ying Z, Cardillo TM, Sharkey RM, Goldenberg DM. New MUC1 serum immunoassay differentiates pancreatic cancer from pancreatitis. J Clin Oncol 2006; 24: 252-258 [PMID: 16344318 DOI: 10.1200/JCO.2005.02.8282]
- Haab BB, Porter A, Yue T, Li L, Scheiman J, Anderson MA, Barnes D, Schmidt CM, Feng Z, 83 Simeone DM. Glycosylation variants of mucins and CEACAMs as candidate biomarkers for the diagnosis of pancreatic cystic neoplasms. Ann Surg 2010; 251: 937-945 [PMID: 20395854 DOI: 10.1097/SLA.0b013e3181d7738d
- Sinha J, Cao Z, Dai J, Tang H, Partyka K, Hostetter G, Simeone DM, Feng Z, Allen PJ, Brand RE, 84 Haab BB. A Gastric Glycoform of MUC5AC Is a Biomarker of Mucinous Cysts of the Pancreas. PLoS One 2016; 11: e0167070 [PMID: 27992432 DOI: 10.1371/journal.pone.0167070]
- 85 Horn A, Chakraborty S, Dey P, Haridas D, Souchek J, Batra SK, Lele SM. Immunocytochemistry for MUC4 and MUC16 is a useful adjunct in the diagnosis of pancreatic adenocarcinoma on fineneedle aspiration cytology. Arch Pathol Lab Med 2013; 137: 546-551 [PMID: 23544943 DOI: 10.5858/arpa.2011-0229-OA]
- Maker AV, Katabi N, Qin LX, Klimstra DS, Schattner M, Brennan MF, Jarnagin WR, Allen PJ. 86 Cvst fluid interleukin-1beta (IL1beta) levels predict the risk of carcinoma in intraductal papillary mucinous neoplasms of the pancreas. Clin Cancer Res 2011; 17: 1502-1508 [PMID: 21266527 DOI: 10.1158/1078-0432.CCR-10-1561]
- Hao S, Takahashi C, Snyder RA, Parikh AA. Stratifying Intraductal Papillary Mucinous Neoplasms 87 by Cyst Fluid Analysis: Present and Future. Int J Mol Sci 2020; 21 [PMID: 32050465 DOI: 10.3390/ijms21031147]
- 88 Das KK, Geng X, Brown JW, Morales-Oyarvide V, Huynh T, Pergolini I, Pitman MB, Ferrone C, Al Efishat M, Haviland D, Thompson E, Wolfgang C, Lennon AM, Allen P, Lillemoe KD, Fields RC, Hawkins WG, Liu J, Castillo CF, Das KM, Mino-Kenudson M. Cross Validation of the Monoclonal Antibody Das-1 in Identification of High-Risk Mucinous Pancreatic Cystic Lesions. Gastroenterology 2019; 157: 720-730.e2 [PMID: 31175863 DOI: 10.1053/j.gastro.2019.05.014]
- Zhang L, Farrell JJ, Zhou H, Elashoff D, Akin D, Park NH, Chia D, Wong DT. Salivary 89 transcriptomic biomarkers for detection of resectable pancreatic cancer. Gastroenterology 2010; 138: 949-57.e1 [PMID: 19931263 DOI: 10.1053/j.gastro.2009.11.010]
- 90 Xie Z, Yin X, Gong B, Nie W, Wu B, Zhang X, Huang J, Zhang P, Zhou Z, Li Z. Salivary microRNAs show potential as a noninvasive biomarker for detecting resectable pancreatic cancer. Cancer Prev Res (Phila) 2015; 8: 165-173 [PMID: 25538087 DOI: 10.1158/1940-6207.CAPR-14-0192]
- Satoh K. Molecular Approaches Using Body Fluid for the Early Detection of Pancreatic Cancer. 91 Diagnostics (Basel) 2021; 11 [PMID: 33671729 DOI: 10.3390/diagnostics11020375]
- 92 Setti G, Pezzi ME, Viani MV, Pertinhez TA, Cassi D, Magnoni C, Bellini P, Musolino A, Vescovi P, Meleti M. Salivary MicroRNA for Diagnosis of Cancer and Systemic Diseases: A Systematic Review. Int J Mol Sci 2020; 21 [PMID: 32019170 DOI: 10.3390/ijms21030907]
- 93 Ishige F, Hoshino I, Iwatate Y, Chiba S, Arimitsu H, Yanagibashi H, Nagase H, Takayama W. MIR1246 in body fluids as a biomarker for pancreatic cancer. Sci Rep 2020; 10: 8723 [PMID: 32457495 DOI: 10.1038/s41598-020-65695-6]
- Radon TP, Massat NJ, Jones R, Alrawashdeh W, Dumartin L, Ennis D, Duffy SW, Kocher HM, 94 Pereira SP, Guarner posthumous L, Murta-Nascimento C, Real FX, Malats N, Neoptolemos J, Costello E, Greenhalf W, Lemoine NR, Crnogorac-Jurcevic T. Identification of a Three-Biomarker Panel in Urine for Early Detection of Pancreatic Adenocarcinoma. Clin Cancer Res 2015; 21: 3512-3521 [PMID: 26240291 DOI: 10.1158/1078-0432.CCR-14-2467]
- 95 Brezgyte G, Shah V, Jach D, Crnogorac-Jurcevic T. Non-Invasive Biomarkers for Earlier Detection of Pancreatic Cancer-A Comprehensive Review. Cancers (Basel) 2021; 13 [PMID: 34072842 DOI: 10.3390/cancers13112722]
- 96 Porta M, Fabregat X, Malats N, Guarner L, Carrato A, de Miguel A, Ruiz L, Jariod M, Costafreda S, Coll S, Alguacil J, Corominas JM, Solà R, Salas A, Real FX. Exocrine pancreatic cancer: symptoms at presentation and their relation to tumour site and stage. Clin Transl Oncol 2005; 7: 189-197 [PMID: 15960930 DOI: 10.1007/BF02712816]
- 97 Mujica VR, Barkin JS, Go VL. Acute pancreatitis secondary to pancreatic carcinoma. Study Group Participants. Pancreas 2000; 21: 329-332 [PMID: 11075985 DOI: 10.1097/00006676-200011000-00001]
- Chari ST, Leibson CL, Rabe KG, Ransom J, de Andrade M, Petersen GM. Probability of pancreatic 98 cancer following diabetes: a population-based study. Gastroenterology 2005; 129: 504-511 [PMID:



16083707 DOI: 10.1016/j.gastro.2005.05.007]

- 99 Aggarwal G, Kamada P, Chari ST. Prevalence of diabetes mellitus in pancreatic cancer compared to common cancers. Pancreas 2013; 42: 198-201 [PMID: 23000893 DOI: 10.1097/MPA.0b013e3182592c96]
- 100 Khorana AA, Fine RL. Pancreatic cancer and thromboembolic disease. Lancet Oncol 2004; 5: 655-663 [PMID: 15522652 DOI: 10.1016/S1470-2045(04)01606-7]
- 101 Pinzon R, Drewinko B, Trujillo JM, Guinee V, Giacco G. Pancreatic carcinoma and Trousseau's syndrome: experience at a large cancer center. J Clin Oncol 1986; 4: 509-514 [PMID: 3958764 DOI: 10.1200/JCO.1986.4.4.509]
- 102 Bravo-Piris J, Villaron LG, Martinez C, Garcia-Perez A. Pipillon-Lefèvre syndrome: report of two familial cases. Dermatologica 1976; 152: 168-176 [PMID: 133038 DOI: 10.1111/j.1365-2230.1992.tb02541.x
- Marcos P, Kieselova K, Cunha M. Pancreatic Panniculitis. Am J Gastroenterol 2017; 112: 1218 103 [PMID: 28766564 DOI: 10.1038/ajg.2017.161]
- 104 Galvañ VG. Sister Mary Joseph's nodule. Ann Intern Med 1998; 128: 410 [PMID: 9490607 DOI: 10.7326/0003-4819-128-5-199803010-00017
- 105 Lee ES, Lee JM. Imaging diagnosis of pancreatic cancer: a state-of-the-art review. World J Gastroenterol 2014; 20: 7864-7877 [PMID: 24976723 DOI: 10.3748/wjg.v20.i24.7864]
- 106 Garces-Descovich A, Beker K, Jaramillo-Cardoso A, James Moser A, Mortele KJ. Applicability of current NCCN Guidelines for pancreatic adenocarcinoma resectability: analysis and pitfalls. Abdom Radiol (NY) 2018; 43: 314-322 [PMID: 29392370 DOI: 10.1007/s00261-018-1459-6]
- 107 Wong JC, Raman S. Surgical resectability of pancreatic adenocarcinoma: CTA. Abdom Imaging 2010; 35: 471-480 [PMID: 19468791 DOI: 10.1007/s00261-009-9539-2]
- 108 Fletcher JG, Wiersema MJ, Farrell MA, Fidler JL, Burgart LJ, Koyama T, Johnson CD, Stephens DH, Ward EM, Harmsen WS. Pancreatic malignancy: value of arterial, pancreatic, and hepatic phase imaging with multi-detector row CT. Radiology 2003; 229: 81-90 [PMID: 14519871 DOI: 10.1148/radiol.2291020582
- 109 Yoon SH, Lee JM, Cho JY, Lee KB, Kim JE, Moon SK, Kim SJ, Baek JH, Kim SH, Lee JY, Han JK, Choi BI. Small (≤ 20 mm) pancreatic adenocarcinomas: analysis of enhancement patterns and secondary signs with multiphasic multidetector CT. Radiology 2011; 259: 442-452 [PMID: 21406627 DOI: 10.1148/radiol.11101133]
- Shukla PJ, Barreto SG, Kulkarni A, Nagarajan G, Fingerhut A. Vascular anomalies encountered 110 during pancreatoduodenectomy: do they influence outcomes? Ann Surg Oncol 2010; 17: 186-193 [PMID: 19838756 DOI: 10.1245/s10434-009-0757-1]
- Chu LC, Goggins MG, Fishman EK. Diagnosis and Detection of Pancreatic Cancer. Cancer J 2017; 111 23: 333-342 [PMID: 29189329 DOI: 10.1097/PPO.000000000000290]
- 112 Li H, Zeng MS, Zhou KR, Jin DY, Lou WH. Pancreatic adenocarcinoma: the different CT criteria for peripancreatic major arterial and venous invasion. J Comput Assist Tomogr 2005; 29: 170-175 [PMID: 15772532 DOI: 10.1097/01.rct.0000155060.73107.83]
- Xia BT, Fu B, Wang J, Kim Y, Ahmad SA, Dhar VK, Levinsky NC, Hanseman DJ, Habib DA, 113 Wilson GC, Smith M, Olowokure OO, Kharofa J, Al Humaidi AH, Choe KA, Abbott DE. Does radiologic response correlate to pathologic response in patients undergoing neoadjuvant therapy for borderline resectable pancreatic malignancy? J Surg Oncol 2017; 115: 376-383 [PMID: 28105634 DOI: 10.1002/jso.24538]
- Holzapfel K, Reiser-Erkan C, Fingerle AA, Erkan M, Eiber MJ, Rummeny EJ, Friess H, Kleeff J, 114 Gaa J. Comparison of diffusion-weighted MR imaging and multidetector-row CT in the detection of liver metastases in patients operated for pancreatic cancer. Abdom Imaging 2011; 36: 179-184 [PMID: 20563868 DOI: 10.1007/s00261-010-9633-5]
- 115 Motosugi U, Ichikawa T, Morisaka H, Sou H, Muhi A, Kimura K, Sano K, Araki T. Detection of pancreatic carcinoma and liver metastases with gadoxetic acid-enhanced MR imaging: comparison with contrast-enhanced multi-detector row CT. Radiology 2011; 260: 446-453 [PMID: 21693662 DOI: 10.1148/radiol.11103548]
- Carrara G, Pecorelli N, De Cobelli F, Cristel G, Damascelli A, Beretta L, Braga M. Preoperative 116 sarcopenia determinants in pancreatic cancer patients. Clin Nutr 2017; 36: 1649-1653 [PMID: 27789123 DOI: 10.1016/j.clnu.2016.10.014]
- 117 Ozola Zalite I, Zykus R, Francisco Gonzalez M, Saygili F, Pukitis A, Gaujoux S, Charnley RM, Lyadov V. Influence of cachexia and sarcopenia on survival in pancreatic ductal adenocarcinoma: a systematic review. Pancreatology 2015; 15: 19-24 [PMID: 25524484 DOI: 10.1016/j.pan.2014.11.006
- 118 Prado CM, Lieffers JR, McCargar LJ, Reiman T, Sawyer MB, Martin L, Baracos VE. Prevalence and clinical implications of sarcopenic obesity in patients with solid tumours of the respiratory and gastrointestinal tracts: a population-based study. Lancet Oncol 2008; 9: 629-635 [PMID: 18539529 DOI: 10.1016/S1470-2045(08)70153-0]
- 119 Schmocker RK, Vanness DJ, Greenberg CC, Havlena JA, LoConte NK, Weiss JM, Neuman HB, Leverson G, Smith MA, Winslow ER. Utilization of preoperative endoscopic ultrasound for pancreatic adenocarcinoma. HPB (Oxford) 2017; 19: 465-472 [PMID: 28237627 DOI: 10.1016/j.hpb.2017.01.017]
- 120 Hartwig W, Schneider L, Diener MK, Bergmann F, Büchler MW, Werner J. Preoperative tissue diagnosis for tumours of the pancreas. Br J Surg 2009; 96: 5-20 [PMID: 19016272 DOI:



#### 10.1002/bis.6407

- 121 Lee TH, Cho YD, Cha SW, Cho JY, Jang JY, Jeong SW, Choi HJ, Moon JH. Endoscopic ultrasound elastography for the pancreas in Korea: a preliminary single center study. Clin Endosc 2013; 46: 172-177 [PMID: 23614128 DOI: 10.5946/ce.2013.46.2.172]
- 122 Chantarojanasiri T, Kongkam P. Endoscopic ultrasound elastography for solid pancreatic lesions. World J Gastrointest Endosc 2017; 9: 506-513 [PMID: 29085561 DOI: 10.4253/wjge.v9.i10.506]
- Okasha H, Elkholy S, El-Sayed R, Wifi MN, El-Nady M, El-Nabawi W, El-Dayem WA, Radwan 123 MI, Farag A, El-Sherif Y, Al-Gemeie E, Salman A, El-Sherbiny M, El-Mazny A, Mahdy RE. Real time endoscopic ultrasound elastography and strain ratio in the diagnosis of solid pancreatic lesions. World J Gastroenterol 2017; 23: 5962-5968 [PMID: 28932088 DOI: 10.3748/wjg.v23.i32.5962]
- 124 Niederau C, Grendell JH. Diagnosis of pancreatic carcinoma. Imaging techniques and tumor markers. Pancreas 1992; 7: 66-86 [PMID: 1557348 DOI: 10.1097/00006676-199201000-00011]
- 125 Vozzo CF, Sanaka MR. Endoscopic Management of Pancreaticobiliary Disease. Surg Clin North Am 2020; 100: 1151-1168 [PMID: 33128885 DOI: 10.1016/j.suc.2020.08.006]
- 126 Alauddin MM. De Palatis L. Current and Future Trends in Early Detection of Pancreatic Cancer: Molecular Targets and PET Probes. Curr Med Chem 2015; 22: 3370-3389 [PMID: 26295468 DOI: 10.2174/0929867322666150821094015]
- Moradi F, Iagaru A. The Role of Positron Emission Tomography in Pancreatic Cancer and 127 Gallbladder Cancer. Semin Nucl Med 2020; 50: 434-446 [PMID: 32768007 DOI: 10.1053/j.semnuclmed.2020.04.002]
- 128 Li XX, Liu NB, Zhu L, Yuan XK, Yang CW, Ren P, Gong LL, Zhao LJ, Xu WG, Wang P. Consequences of additional use of contrast-enhanced (18)F-FDG PET/CT in target volume delineation and dose distribution for pancreatic cancer. Br J Radiol 2015; 88: 20140590 [PMID: 25939819 DOI: 10.1259/bjr.20140590]
- Wang L, Dong P, Wang WG, Tian BL. Positron emission tomography modalities prevent futile 129 radical resection of pancreatic cancer: A meta-analysis. Int J Surg 2017; 46: 119-125 [PMID: 28890410 DOI: 10.1016/j.ijsu.2017.09.003]
- Santhosh S, Mittal BR, Bhasin DK, Rana SS, Gupta R, Das A, Nada R. Fluorodeoxyglucose-130 positron emission tomography/computed tomography performs better than contrast-enhanced computed tomography for metastasis evaluation in the initial staging of pancreatic adenocarcinoma. Ann Nucl Med 2017; 31: 575-581 [PMID: 28689356 DOI: 10.1007/s12149-017-1193-0]
- 131 Daamen LA, Groot VP, Goense L, Wessels FJ, Borel Rinkes IH, Intven MPW, van Santvoort HC, Molenaar IQ. The diagnostic performance of CT vs FDG PET-CT for the detection of recurrent pancreatic cancer: a systematic review and meta-analysis. Eur J Radiol 2018; 106: 128-136 [PMID: 30150034 DOI: 10.1016/j.ejrad.2018.07.010]
- England CG, Hernandez R, Eddine SB, Cai W. Molecular Imaging of Pancreatic Cancer with 132 Antibodies. Mol Pharm 2016; 13: 8-24 [PMID: 26620581 DOI: 10.1021/acs.molpharmaceut.5b00626]
- Feng X, Wang Y, Lu D, Xu X, Zhou X, Zhang H, Zhang T, Zhu H, Yang Z, Wang F, Li N, Liu Z. 133 Clinical Translation of a <sup>68</sup>Ga-Labeled Integrin  $\alpha_{\nu}\beta_{6}$ -Targeting Cyclic Radiotracer for PET Imaging of Pancreatic Cancer. J Nucl Med 2020; 61: 1461-1467 [PMID: 32086242 DOI: 10.2967/jnumed.119.237347
- 134 Serrao EM, Kettunen MI, Rodrigues TB, Dzien P, Wright AJ, Gopinathan A, Gallagher FA, Lewis DY, Frese KK, Almeida J, Howat WJ, Tuveson DA, Brindle KM. MRI with hyperpolarised [1-13C]pyruvate detects advanced pancreatic preneoplasia prior to invasive disease in a mouse model. Gut 2016; 65: 465-475 [PMID: 26347531 DOI: 10.1136/gutjnl-2015-310114]
- 135 Mayo SC, Austin DF, Sheppard BC, Mori M, Shipley DK, Billingsley KG. Evolving preoperative evaluation of patients with pancreatic cancer: does laparoscopy have a role in the current era? J Am Coll Surg 2009; 208: 87-95 [PMID: 19228509 DOI: 10.1016/j.jamcollsurg.2008.10.014]
- 136 Allen VB, Gurusamy KS, Takwoingi Y, Kalia A, Davidson BR. Diagnostic accuracy of laparoscopy following computed tomography (CT) scanning for assessing the resectability with curative intent in pancreatic and periampullary cancer. Cochrane Database Syst Rev 2016; 7: CD009323 [PMID: 27383694 DOI: 10.1002/14651858.CD009323.pub3]
- 137 Pisters PW, Lee JE, Vauthey JN, Charnsangavej C, Evans DB. Laparoscopy in the staging of pancreatic cancer. Br J Surg 2001; 88: 325-337 [PMID: 11260096 DOI: 10.1046/j.1365-2168.2001.01695.x]
- 138 Fong ZV, Alvino DML, Fernández-Del Castillo C, Mehtsun WT, Pergolini I, Warshaw AL, Chang DC, Lillemoe KD, Ferrone CR. Reappraisal of Staging Laparoscopy for Patients with Pancreatic Adenocarcinoma: A Contemporary Analysis of 1001 Patients. Ann Surg Oncol 2017; 24: 3203-3211 [PMID: 28718038 DOI: 10.1245/s10434-017-5973-5]
- Karachristos A, Scarmeas N, Hoffman JP. CA 19-9 Levels predict results of staging laparoscopy in 139 pancreatic cancer. J Gastrointest Surg 2005; 9: 1286-1292 [PMID: 16332484 DOI: 10.1016/i.gassur.2005.06.008]
- 140 Wang KX, Ben QW, Jin ZD, Du YQ, Zou DW, Liao Z, Li ZS. Assessment of morbidity and mortality associated with EUS-guided FNA: a systematic review. Gastrointest Endosc 2011; 73: 283-290 [PMID: 21295642 DOI: 10.1016/j.gie.2010.10.045]
- 141 Yane K, Kuwatani M, Yoshida M, Goto T, Matsumoto R, Ihara H, Okuda T, Taya Y, Ehira N, Kudo T, Adachi T, Eto K, Onodera M, Sano I, Nojima M, Katanuma A. Non-negligible rate of needle tract seeding after endoscopic ultrasound-guided fine-needle aspiration for patients undergoing distal



pancreatectomy for pancreatic cancer. Dig Endosc 2020; 32: 801-811 [PMID: 31876309 DOI: 10.1111/den.13615]

- Zhang L, Sanagapalli S, Stoita A. Challenges in diagnosis of pancreatic cancer. World J 142 Gastroenterol 2018; 24: 2047-2060 [PMID: 29785074 DOI: 10.3748/wjg.v24.i19.2047]
- 143 Clarke DL, Clarke BA, Thomson SR, Garden OJ, Lazarus NG. The role of preoperative biopsy in pancreatic cancer. HPB (Oxford) 2004; 6: 144-153 [PMID: 18333068 DOI: 10.1080/13651820410030862]
- 144 Pezzilli R. Asymptomatic lesions of the pancreas: an overview. J Gastroenterol Hepatol Res 2014; 3: 1216-1219
- 145 Tanaka M, Fernández-Del Castillo C, Kamisawa T, Jang JY, Levy P, Ohtsuka T, Salvia R, Shimizu Y, Tada M, Wolfgang CL. Revisions of international consensus Fukuoka guidelines for the management of IPMN of the pancreas. Pancreatology 2017; 17: 738-753 [PMID: 28735806 DOI: 10.1016/j.pan.2017.07.007]
- 146 van Roessel S, Kasumova GG, Verheij J, Najarian RM, Maggino L, de Pastena M, Malleo G, Marchegiani G, Salvia R, Ng SC, de Geus SW, Lof S, Giovinazzo F, van Dam JL, Kent TS, Busch OR, van Eijck CH, Koerkamp BG, Abu Hilal M, Bassi C, Tseng JF, Besselink MG. International Validation of the Eighth Edition of the American Joint Committee on Cancer (AJCC) TNM Staging System in Patients With Resected Pancreatic Cancer. JAMA Surg 2018; 153: e183617 [PMID: 30285076 DOI: 10.1001/jamasurg.2018.3617]
- 147 Bockhorn M, Uzunoglu FG, Adham M, Imrie C, Milicevic M, Sandberg AA, Asbun HJ, Bassi C, Büchler M, Charnley RM, Conlon K, Cruz LF, Dervenis C, Fingerhutt A, Friess H, Gouma DJ, Hartwig W, Lillemoe KD, Montorsi M, Neoptolemos JP, Shrikhande SV, Takaori K, Traverso W, Vashist YK, Vollmer C, Yeo CJ, Izbicki JR; International Study Group of Pancreatic Surgery. Borderline resectable pancreatic cancer: a consensus statement by the International Study Group of Pancreatic Surgery (ISGPS). Surgery 2014; 155: 977-988 [PMID: 24856119 DOI: 10.1016/j.surg.2014.02.001]
- 148 Isaji S, Mizuno S, Windsor JA, Bassi C, Fernández-Del Castillo C, Hackert T, Hayasaki A, Katz MHG, Kim SW, Kishiwada M, Kitagawa H, Michalski CW, Wolfgang CL. International consensus on definition and criteria of borderline resectable pancreatic ductal adenocarcinoma 2017. Pancreatology 2018; 18: 2-11 [PMID: 29191513 DOI: 10.1016/j.pan.2017.11.011]
- Brown JA, Zenati MS, Simmons RL, Al Abbas AI, Chopra A, Smith K, Lee KKW, Hogg ME, Zeh HJ, Paniccia A, Zureikat AH. Long-Term Surgical Complications After Pancreatoduodenectomy: Incidence, Outcomes, and Risk Factors. J Gastrointest Surg 2020; 24: 1581-1589 [PMID: 32410174 DOI: 10.1007/s11605-020-04641-3]
- 150 Seiler CA, Wagner M, Bachmann T, Redaelli CA, Schmied B, Uhl W, Friess H, Büchler MW. Randomized clinical trial of pylorus-preserving duodenopancreatectomy vs classical Whipple resection-long term results. Br J Surg 2005; 92: 547-556 [PMID: 15800958 DOI: 10.1002/bjs.4881]
- 151 Hüttner FJ, Fitzmaurice C, Schwarzer G, Seiler CM, Antes G, Büchler MW, Diener MK. Pyloruspreserving pancreaticoduodenectomy (pp Whipple) vs pancreaticoduodenectomy (classic Whipple) for surgical treatment of periampullary and pancreatic carcinoma. Cochrane Database Syst Rev 2016; 2: CD006053 [PMID: 26905229 DOI: 10.1002/14651858.CD006053.pub6]
- 152 Karim SAM, Abdulla KS, Abdulkarim QH, Rahim FH. The outcomes and complications of pancreaticoduodenectomy (Whipple procedure): Cross sectional study. Int J Surg 2018; 52: 383-387 [PMID: 29438817 DOI: 10.1016/j.ijsu.2018.01.041]
- 153 Mora-Oliver I, Garcés-Albir M, Dorcaratto D, Muñoz-Forner E, Izquierdo Moreno A, Carbonell-Aliaga MP, Sabater L. Pancreatoduodenectomy with artery-first approach. Minerva Chir 2019; 74: 226-236 [PMID: 30600965 DOI: 10.23736/S0026-4733.18.07944-0]
- Li Z, Wei A, Xia N, Zheng L, Yang D, Ye J, Xiong J, Hu W. Blumgart anastomosis reduces the 154 incidence of pancreatic fistula after pancreaticoduodenectomy: a systematic review and metaanalysis. Sci Rep 2020; 10: 17896 [PMID: 33087777 DOI: 10.1038/s41598-020-74812-4]
- Ricci C, Ingaldi C, Alberici L, Pagano N, Mosconi C, Marasco G, Minni F, Casadei R. Blumgart 155 Anastomosis After Pancreaticoduodenectomy. A Comprehensive Systematic Review, Meta-Analysis, and Meta-Regression. World J Surg 2021; 45: 1929-1939 [PMID: 33721074 DOI: 10.1007/s00268-021-06039-x]
- 156 Fang Y, Gurusamy KS, Wang Q, Davidson BR, Lin H, Xie X, Wang C. Meta-analysis of randomized clinical trials on safety and efficacy of biliary drainage before surgery for obstructive jaundice. Br J Surg 2013; 100: 1589-1596 [PMID: 24264780 DOI: 10.1002/bjs.9260]
- 157 Schorn S, Vogel T, Demir IE, Demir E, Safak O, Friess H, Ceyhan GO. Do somatostatin-analogues have the same impact on postoperative morbidity and pancreatic fistula in patients after pancreaticoduodenectomy and distal pancreatectomy? Pancreatology 2020; 20: 1770-1778 [PMID: 33121847 DOI: 10.1016/j.pan.2020.10.043]
- 158 Ammori BJ, Ayiomamitis GD. Laparoscopic pancreaticoduodenectomy and distal pancreatectomy: a UK experience and a systematic review of the literature. Surg Endosc 2011; 25: 2084-2099 [PMID: 21298539 DOI: 10.1007/s00464-010-1538-4]
- 159 Mehrabi A, Hafezi M, Arvin J, Esmaeilzadeh M, Garoussi C, Emami G, Kössler-Ebs J, Müller-Stich BP, Büchler MW, Hackert T, Diener MK. A systematic review and meta-analysis of laparoscopic vs open distal pancreatectomy for benign and malignant lesions of the pancreas: it's time to randomize. Surgery 2015; 157: 45-55 [PMID: 25482464 DOI: 10.1016/j.surg.2014.06.081]
- 160 Pierce RA, Spitler JA, Hawkins WG, Strasberg SM, Linehan DC, Halpin VJ, Eagon JC, Brunt LM,



Frisella MM, Matthews BD. Outcomes analysis of laparoscopic resection of pancreatic neoplasms. Surg Endosc 2007; 21: 579-586 [PMID: 17180287 DOI: 10.1007/s00464-006-9022-x]

- 161 Petrucciani N. Nigri G. Debs T. Giannini G. Sborlini E. Antolino L. Aurello P. D'Angelo F. Gugenheim J, Ramacciato G. Frozen section analysis of the pancreatic margin during pancreaticoduodenectomy for cancer: Does extending the resection to obtain a secondary R0 provide a survival benefit? Pancreatology 2016; 16: 1037-1043 [PMID: 27697467 DOI: 10.1016/j.pan.2016.09.004]
- 162 Winter JM, Cameron JL, Campbell KA, Arnold MA, Chang DC, Coleman J, Hodgin MB, Sauter PK, Hruban RH, Riall TS, Schulick RD, Choti MA, Lillemoe KD, Yeo CJ. 1423 pancreaticoduodenectomies for pancreatic cancer: A single-institution experience. J Gastrointest Surg 2006; 10: 1199-210; discussion 1210 [PMID: 17114007 DOI: 10.1016/j.gassur.2006.08.018]
- Schmidt CM, Glant J, Winter JM, Kennard J, Dixon J, Zhao Q, Howard TJ, Madura JA, Nakeeb A, 163 Pitt HA, Cameron JL, Yeo CJ, Lillemoe KD. Total pancreatectomy (R0 resection) improves survival over subtotal pancreatectomy in isolated neck margin positive pancreatic adenocarcinoma. Surgery 2007; 142: 572-8; discussion 578 [PMID: 17950350 DOI: 10.1016/j.surg.2007.07.016]
- Karpoff HM, Klimstra DS, Brennan MF, Conlon KC. Results of total pancreatectomy for 164 adenocarcinoma of the pancreas. Arch Surg 2001; 136: 44-7; discussion 48 [PMID: 11146775 DOI: 10.1001/archsurg.136.1.44]
- Scholten L, Stoop TF, Del Chiaro M, Busch OR, van Eijck C, Molenaar IQ, de Vries JH, Besselink 165 MG; Dutch Pancreatic Cancer Group. Systematic review of functional outcome and quality of life after total pancreatectomy. Br J Surg 2019; 106: 1735-1746 [PMID: 31502658 DOI: 10.1002/bis.11296]
- Tomlinson JS, Jain S, Bentrem DJ, Sekeris EG, Maggard MA, Hines OJ, Reber HA, Ko CY. 166 Accuracy of staging node-negative pancreas cancer: a potential quality measure. Arch Surg 2007; 142: 767-723; discussion 773 [PMID: 17709731 DOI: 10.1001/archsurg.142.8.767]
- Tol JA, Gouma DJ, Bassi C, Dervenis C, Montorsi M, Adham M, Andrén-Sandberg A, Asbun HJ, 167 Bockhorn M, Büchler MW, Conlon KC, Fernández-Cruz L, Fingerhut A, Friess H, Hartwig W, Izbicki JR, Lillemoe KD, Milicevic MN, Neoptolemos JP, Shrikhande SV, Vollmer CM, Yeo CJ, Charnley RM; International Study Group on Pancreatic Surgery. Definition of a standard lymphadenectomy in surgery for pancreatic ductal adenocarcinoma: a consensus statement by the International Study Group on Pancreatic Surgery (ISGPS). Surgery 2014; 156: 591-600 [PMID: 25061003 DOI: 10.1016/j.surg.2014.06.016]
- 168 Sun J, Yang Y, Wang X, Yu Z, Zhang T, Song J, Zhao H, Wen J, Du Y, Lau WY, Zhang Y. Metaanalysis of the efficacies of extended and standard pancreatoduodenectomy for ductal adenocarcinoma of the head of the pancreas. World J Surg 2014; 38: 2708-2715 [PMID: 24912627 DOI: 10.1007/s00268-014-2633-9]
- 169 Zgliczynski S, Gietka-Czernel M, Gorowski T, Bednarski A, Chomicki O, Jastrzebska W, Makowska A, Niegowska E, Pucilowska J, Soszynski P. Results of 1311 theory for 2,000 thyrotoxic patients: do the effects depend on the dose? Exp Clin Endocrinol 1991; 97: 286-291 [PMID: 1915646 DOI: 10.1245/s10434-008-0281-8]
- 170 Gurusamy KS, Kumar S, Davidson BR, Fusai G. Resection vs other treatments for locally advanced pancreatic cancer. Cochrane Database Syst Rev 2014; CD010244 [PMID: 24578248 DOI: 10.1002/14651858.CD010244.pub2]
- 171 Yamada S, Fujii T, Sugimoto H, Nomoto S, Takeda S, Kodera Y, Nakao A. Aggressive surgery for borderline resectable pancreatic cancer: evaluation of National Comprehensive Cancer Network guidelines. Pancreas 2013; 42: 1004-1010 [PMID: 23532000 DOI: 10.1097/MPA.0b013e31827b2d7c]
- 172 Mollberg N, Rahbari NN, Koch M, Hartwig W, Hoeger Y, Büchler MW, Weitz J. Arterial resection during pancreatectomy for pancreatic cancer: a systematic review and meta-analysis. Ann Surg 2011; 254: 882-893 [PMID: 22064622 DOI: 10.1097/SLA.0b013e31823ac299]
- 173 Wang S, Shi N, You L, Dai M, Zhao Y. Minimally invasive surgical approach vs open procedure for pancreaticoduodenectomy: A systematic review and meta-analysis. Medicine (Baltimore) 2017; 96: e8619 [PMID: 29390259 DOI: 10.1097/MD.00000000008619]
- Palanivelu C, Senthilnathan P, Sabnis SC, Babu NS, Srivatsan Gurumurthy S, Anand Vijai N, 174 Nalankilli VP, Praveen Raj P, Parthasarathy R, Rajapandian S. Randomized clinical trial of laparoscopic vs open pancreatoduodenectomy for periampullary tumours. Br J Surg 2017; 104: 1443-1450 [PMID: 28895142 DOI: 10.1002/bjs.10662]
- 175 Poves I, Burdío F, Morató O, Iglesias M, Radosevic A, Ilzarbe L, Visa L, Grande L. Comparison of Perioperative Outcomes Between Laparoscopic and Open Approach for Pancreatoduodenectomy: The PADULAP Randomized Controlled Trial. Ann Surg 2018; 268: 731-739 [PMID: 30138162 DOI: 10.1097/SLA.00000000002893]
- 176 Croome KP, Farnell MB, Que FG, Reid-Lombardo KM, Truty MJ, Nagorney DM, Kendrick ML. Total laparoscopic pancreaticoduodenectomy for pancreatic ductal adenocarcinoma: oncologic advantages over open approaches? Ann Surg 2014; 260: 633-8; discussion 638 [PMID: 25203880 DOI: 10.1097/SLA.0000000000009371
- Boone BA, Zenati M, Hogg ME, Steve J, Moser AJ, Bartlett DL, Zeh HJ, Zureikat AH. Assessment 177 of quality outcomes for robotic pancreaticoduodenectomy: identification of the learning curve. JAMA Surg 2015; 150: 416-422 [PMID: 25761143 DOI: 10.1001/jamasurg.2015.17]
- 178 Wang SE, Shyr BU, Chen SC, Shyr YM. Comparison between robotic and open



pancreaticoduodenectomy with modified Blumgart pancreaticojejunostomy: A propensity scorematched study. Surgery 2018; 164: 1162-1167 [PMID: 30093277 DOI: 10.1016/j.surg.2018.06.031]

- 179 Zureikat AH, Postlewait LM, Liu Y, Gillespie TW, Weber SM, Abbott DE, Ahmad SA, Maithel SK, Hogg ME, Zenati M, Cho CS, Salem A, Xia B, Steve J, Nguyen TK, Keshava HB, Chalikonda S, Walsh RM, Talamonti MS, Stocker SJ, Bentrem DJ, Lumpkin S, Kim HJ, Zeh HJ 3rd, Kooby DA. A Multi-institutional Comparison of Perioperative Outcomes of Robotic and Open Pancreaticoduodenectomy. Ann Surg 2016; 264: 640-649 [PMID: 27433907 DOI: 10.1097/SLA.000000000001869]
- 180 Nassour I, Tohme S, Hoehn R, Adam MA, Zureikat AH, Alessandro P. Safety and oncologic efficacy of robotic compared to open pancreaticoduodenectomy after neoadjuvant chemotherapy for pancreatic cancer. Surg Endosc 2021; 35: 2248-2254 [PMID: 32440928 DOI: 10.1007/s00464-020-07638-w
- 181 Baimas-George M, Watson M, Murphy KJ, Iannitti D, Baker E, Ocuin L, Vrochides D, Martinie JB. Robotic pancreaticoduodenectomy may offer improved oncologic outcomes over open surgery: a propensity-matched single-institution study. Surg Endosc 2020; 34: 3644-3649 [PMID: 32328825 DOI: 10.1007/s00464-020-07564-x]
- 182 Mizrahi JD, Surana R, Valle JW, Shroff RT. Pancreatic cancer. Lancet 2020; 395: 2008-2020 [PMID: 32593337 DOI: 10.1016/S0140-6736(20)30974-0]
- Perri G, Prakash L, Qiao W, Varadhachary GR, Wolff R, Fogelman D, Overman M, Pant S, Javle 183 M, Koay EJ, Herman J, Kim M, Ikoma N, Tzeng CW, Lee JE, Katz MHG. Postoperative Chemotherapy Benefits Patients Who Received Preoperative Therapy and Pancreatectomy for Pancreatic Adenocarcinoma. Ann Surg 2020; 271: 996-1002 [PMID: 31895709 DOI: 10.1097/SLA.00000000003763
- 184 Müller PC, Frey MC, Ruzza CM, Nickel F, Jost C, Gwerder C, Hackert T, Z'graggen K, Kessler U. Neoadjuvant Chemotherapy in Pancreatic Cancer: An Appraisal of the Current High-Level Evidence. Pharmacology 2021; 106: 143-153 [PMID: 32966993 DOI: 10.1159/000510343]
- 185 Neoptolemos JP, Stocken DD, Friess H, Bassi C, Dunn JA, Hickey H, Beger H, Fernandez-Cruz L, Dervenis C, Lacaine F, Falconi M, Pederzoli P, Pap A, Spooner D, Kerr DJ, Büchler MW; European Study Group for Pancreatic Cancer. A randomized trial of chemoradiotherapy and chemotherapy after resection of pancreatic cancer. N Engl J Med 2004; 350: 1200-1210 [PMID: 15028824 DOI: 10.1056/NEJMoa032295
- 186 Oettle H, Neuhaus P, Hochhaus A, Hartmann JT, Gellert K, Ridwelski K, Niedergethmann M, Zülke C, Fahlke J, Arning MB, Sinn M, Hinke A, Riess H. Adjuvant chemotherapy with gemcitabine and long-term outcomes among patients with resected pancreatic cancer: the CONKO-001 randomized trial. JAMA 2013; 310: 1473-1481 [PMID: 24104372 DOI: 10.1001/jama.2013.279201]
- 187 Neoptolemos JP, Stocken DD, Bassi C, Ghaneh P, Cunningham D, Goldstein D, Padbury R, Moore MJ, Gallinger S, Mariette C, Wente MN, Izbicki JR, Friess H, Lerch MM, Dervenis C, Oláh A, Butturini G, Doi R, Lind PA, Smith D, Valle JW, Palmer DH, Buckels JA, Thompson J, McKay CJ, Rawcliffe CL, Büchler MW; European Study Group for Pancreatic Cancer. Adjuvant chemotherapy with fluorouracil plus folinic acid vs gemcitabine following pancreatic cancer resection: a randomized controlled trial. JAMA 2010; 304: 1073-1081 [PMID: 20823433 DOI: 10.1001/jama.2010.1275]
- 188 Neoptolemos JP, Palmer DH, Ghaneh P, Psarelli EE, Valle JW, Halloran CM, Faluyi O, O'Reilly DA, Cunningham D, Wadsley J, Darby S, Meyer T, Gillmore R, Anthoney A, Lind P, Glimelius B, Falk S, Izbicki JR, Middleton GW, Cummins S, Ross PJ, Wasan H, McDonald A, Crosby T, Ma YT, Patel K, Sherriff D, Soomal R, Borg D, Sothi S, Hammel P, Hackert T, Jackson R, Büchler MW; European Study Group for Pancreatic Cancer. Comparison of adjuvant gemcitabine and capecitabine with gemcitabine monotherapy in patients with resected pancreatic cancer (ESPAC-4): a multicentre, open-label, randomised, phase 3 trial. Lancet 2017; 389: 1011-1024 [PMID: 28129987 DOI: 10.1016/S0140-6736(16)32409-6
- Conroy T, Hammel P, Hebbar M, Ben Abdelghani M, Wei AC, Raoul JL, Choné L, Francois E, Artru P, Biagi JJ, Lecomte T, Assenat E, Faroux R, Ychou M, Volet J, Sauvanet A, Breysacher G, Di Fiore F, Cripps C, Kavan P, Texereau P, Bouhier-Leporrier K, Khemissa-Akouz F, Legoux JL, Juzyna B, Gourgou S, O'Callaghan CJ, Jouffroy-Zeller C, Rat P, Malka D, Castan F, Bachet JB; Canadian Cancer Trials Group and the Unicancer-GI-PRODIGE Group. FOLFIRINOX or Gemcitabine as Adjuvant Therapy for Pancreatic Cancer. N Engl J Med 2018; 379: 2395-2406 [PMID: 30575490 DOI: 10.1056/NEJMoa1809775]
- 190 Parmar A, Chaves-Porras J, Saluja R, Perry K, Rahmadian AP, Santos SD, Ko YJ, Berry S, Doherty M, Chan KKW. Adjuvant treatment for resected pancreatic adenocarcinoma: A systematic review and network meta-analysis. Crit Rev Oncol Hematol 2020; 145: 102817 [PMID: 31955005 DOI: 10.1016/j.critrevonc.2019.102817]
- 191 Galvano A, Castiglia M, Rizzo S, Silvestris N, Brunetti O, Vaccaro G, Gristina V, Barraco N, Bono M, Guercio G, Graceffa G, Fulfaro F, Gori S, Bazan V, Russo A. Moving the Target on the Optimal Adjuvant Strategy for Resected Pancreatic Cancers: A Systematic Review with Meta-Analysis. Cancers (Basel) 2020; 12 [PMID: 32110977 DOI: 10.3390/cancers12030534]
- 192 Raufi AG, Breakstone R, Leonard K, Charpentier K, Beard R, Renaud J, Cavanaugh L, Sturtevant A, MacKinnon K, Almhanna K, Olszewski A, Safran HP. Adjuvant FOLFOX+Nab-Paclitaxel (FOLFOX-A) for Pancreatic Cancer: A Brown University Oncology Research Group Phase II Study



(BrUOG295). Am J Clin Oncol 2020; 43: 857-860 [PMID: 32976178 DOI: 10.1097/COC.000000000000762]

- Kamarajah SK, Bundred JR, Alrawashdeh W, Manas D, White SA. A systematic review and 193 network meta-analysis of phase III randomised controlled trials for adjuvant therapy following resection of pancreatic ductal adenocarcinoma (PDAC). HPB (Oxford) 2020; 22: 649-659 [PMID: 31894014 DOI: 10.1016/j.hpb.2019.12.001]
- Ma SJ, Oladeru OT, Miccio JA, Iovoli AJ, Hermann GM, Singh AK. Association of Timing of 194 Adjuvant Therapy With Survival in Patients With Resected Stage I to II Pancreatic Cancer. JAMA Netw Open 2019; 2: e199126 [PMID: 31411712 DOI: 10.1001/jamanetworkopen.2019.9126]
- 195 You MS, Ryu JK, Huh G, Chun JW, Paik WH, Lee SH, Kim YT. Comparison of efficacy between adjuvant chemotherapy and chemoradiation therapy for pancreatic cancer: AJCC stage-based approach. World J Clin Oncol 2020; 11: 747-760 [PMID: 33033696 DOI: 10.5306/wjco.v11.i9.747]
- 196 Janssen QP, O'Reilly EM, van Eijck CHJ, Groot Koerkamp B. Neoadjuvant Treatment in Patients With Resectable and Borderline Resectable Pancreatic Cancer. Front Oncol 2020; 10: 41 [PMID: 32083002 DOI: 10.3389/fonc.2020.00041]
- Murphy JE, Wo JY, Ryan DP, Jiang W, Yeap BY, Drapek LC, Blaszkowsky LS, Kwak EL, Allen 197 JN, Clark JW, Faris JE, Zhu AX, Goyal L, Lillemoe KD, DeLaney TF, Fernández-Del Castillo C, Ferrone CR, Hong TS. Total Neoadjuvant Therapy With FOLFIRINOX Followed by Individualized Chemoradiotherapy for Borderline Resectable Pancreatic Adenocarcinoma: A Phase 2 Clinical Trial. JAMA Oncol 2018; 4: 963-969 [PMID: 29800971 DOI: 10.1001/jamaoncol.2018.0329]
- 198 van Roessel S, van Veldhuisen E, Klompmaker S, Janssen QP, Abu Hilal M, Alseidi A, Balduzzi A, Balzano G, Bassi C, Berrevoet F, Bonds M, Busch OR, Butturini G, Del Chiaro M, Conlon KC, Falconi M, Frigerio I, Fusai GK, Gagnière J, Griffin O, Hackert T, Halimi A, Klaiber U, Labori KJ, Malleo G, Marino MV, Mortensen MB, Nikov A, Lesurtel M, Keck T, Kleeff J, Pandé R, Pfeiffer P, Pietrasz D, Roberts KJ, Sa Cunha A, Salvia R, Strobel O, Tarvainen T, Bossuyt PM, van Laarhoven HWM, Wilmink JW, Groot Koerkamp B, Besselink MG; European-African Hepato-Pancreato-Biliary Association. Evaluation of Adjuvant Chemotherapy in Patients With Resected Pancreatic Cancer After Neoadjuvant FOLFIRINOX Treatment. JAMA Oncol 2020; 6: 1733-1740 [PMID: 32910170 DOI: 10.1001/jamaoncol.2020.3537]
- Yoo C, Hwang I, Song TJ, Lee SS, Jeong JH, Park DH, Seo DW, Lee SK, Kim MH, Byun JH, Park 199 JH, Hwang DW, Song KB, Lee JH, Lee W, Chang HM, Kim KP, Kim SC, Ryoo BY. FOLFIRINOX in borderline resectable and locally advanced unresectable pancreatic adenocarcinoma. Ther Adv Med Oncol 2020; 12: 1758835920953294 [PMID: 32983266 DOI: 10.1177/1758835920953294]
- 200 Oba A, Ho F, Bao QR, Al-Musawi MH, Schulick RD, Del Chiaro M. Neoadjuvant Treatment in Pancreatic Cancer. Front Oncol 2020; 10: 245 [PMID: 32185128 DOI: 10.3389/fonc.2020.00245]
- Giovinazzo F, Soggiu F, Jang JY, Versteijne E, van Tienhoven G, van Eijck CH, Han Y, Choi SH, 201 Kang CM, Zalupski M, Ahmad H, Yentz S, Helton S, Rose JB, Takishita C, Nagakawa Y, Abu Hilal M. Gemcitabine-Based Neoadjuvant Treatment in Borderline Resectable Pancreatic Ductal Adenocarcinoma: A Meta-Analysis of Individual Patient Data. Front Oncol 2020; 10: 1112 [PMID: 32850319 DOI: 10.3389/fonc.2020.01112]
- 202 Versteijne E, Suker M, Groothuis K, Akkermans-Vogelaar JM, Besselink MG, Bonsing BA, Buijsen J. Busch OR. Creemers GM, van Dam RM, Eskens FALM, Festen S, de Groot JWB, Groot Koerkamp B, de Hingh IH, Homs MYV, van Hooft JE, Kerver ED, Luelmo SAC, Neelis KJ. Nuyttens J, Paardekooper GMRM, Patijn GA, van der Sangen MJC, de Vos-Geelen J, Wilmink JW, Zwinderman AH, Punt CJ, van Eijck CH, van Tienhoven G; Dutch Pancreatic Cancer Group. Preoperative Chemoradiotherapy Versus Immediate Surgery for Resectable and Borderline Resectable Pancreatic Cancer: Results of the Dutch Randomized Phase III PREOPANC Trial. J Clin Oncol 2020; 38: 1763-1773 [PMID: 32105518 DOI: 10.1200/JCO.19.02274]
- 203 Cloyd JM, Chen HC, Wang X, Tzeng CD, Kim MP, Aloia TA, Vauthey JN, Lee JE, Katz MHG. Chemotherapy Versus Chemoradiation as Preoperative Therapy for Resectable Pancreatic Ductal Adenocarcinoma: A Propensity Score Adjusted Analysis. Pancreas 2019; 48: 216-222 [PMID: 30629022 DOI: 10.1097/MPA.00000000001231]
- 204 Grossberg AJ, Chu LC, Deig CR, Fishman EK, Hwang WL, Maitra A, Marks DL, Mehta A, Nabavizadeh N, Simeone DM, Weekes CD, Thomas CR Jr. Multidisciplinary standards of care and recent progress in pancreatic ductal adenocarcinoma. CA Cancer J Clin 2020; 70: 375-403 [PMID: 32683683 DOI: 10.3322/caac.21626]
- Ghaneh P, Palmer DH, Cicconi S, Halloran C, Psarelli EE, Rawcliffe CL, Sripadam R, Mukherjee 205 S, Wadsley J, Al-Mukhtar A, Jiao LR, Wasan HS, Carter R, Graham JS, Ammad F, Evans J, Tjaden C, Hackert T, Buchler MW, Neoptolemos JP; European Study Group for Pancreatic Cancer (ESPAC). ESPAC-5F: four-arm, prospective, multicenter, international randomized phase II trial of immediate surgery compared with neoadjuvant gemcitabine plus capecitabine (GEMCAP) or MFFX or chemoradiotherapy (CRT) in patients with borderline resectable pancreatic cancer. J Clin Oncol 2020; 38 (15\_Suppl): 4505 [DOI: 10.1200/JCO.2020.38.15\_suppl.4505]
- 206 Gamboa AC, Rupji M, Switchenko JM, Lee RM, Turgeon MK, Meyer BI, Russell MC, Cardona K, Kooby DA, Maithel SK, Shah MM. Optimal timing and treatment strategy for pancreatic cancer. J Surg Oncol 2020; 122: 457-468 [PMID: 32470166 DOI: 10.1002/jso.25976]
- 207 Sohal D, Duong MT, Ahmad SA, Gandhi N, Beg MS. SWOG S1505: Results of perioperative chemotherapy (peri-op CTx) with mFFX vs gemcitabine/nab-paclitaxel (Gem/nabP) for resectable pancreatic ductal adenocarcinoma (PRA). J Clin Oncol 2020; 38 (15 Suppl): 4504



- Motoi F, Unno M. Adjuvant and neoadjuvant treatment for pancreatic adenocarcinoma. Jpn J Clin 208 Oncol 2020; 50: 483-489 [PMID: 32083290 DOI: 10.1093/jjco/hyaa018]
- 209 Labori KJ, Lassen K, Hoem D, Grønbech JE, Søreide JA, Mortensen K, Smaaland R, Sorbve H, Verbeke C, Dueland S. Neoadjuvant chemotherapy vs surgery first for resectable pancreatic cancer (Norwegian Pancreatic Cancer Trial - 1 (NorPACT-1)) - study protocol for a national multicentre randomized controlled trial. BMC Surg 2017; 17: 94 [PMID: 28841916 DOI: 10.1186/s12893-017-0291-1]
- 210 Schwarz L, Vernerey D, Bachet JB, Tuech JJ, Portales F, Michel P, Cunha AS. Resectable pancreatic adenocarcinoma neo-adjuvant FOLF(IRIN)OX-based chemotherapy - a multicenter, noncomparative, randomized, phase II trial (PANACHE01-PRODIGE48 study). BMC Cancer 2018; 18: 762 [PMID: 30041614 DOI: 10.1186/s12885-018-4663-4]
- Ettrich TJ, Berger AW, Perkhofer L, Daum S, König A, Dickhut A, Wittel U, Wille K, Geissler M, 211 Algül H, Gallmeier E, Atzpodien J, Kornmann M, Muche R, Prasnikar N, Tannapfel A, Reinacher-Schick A, Uhl W, Seufferlein T. Neoadjuvant plus adjuvant or only adjuvant nab-paclitaxel plus gemcitabine for resectable pancreatic cancer - the NEONAX trial (AIO-PAK-0313), a prospective, randomized, controlled, phase II study of the AIO pancreatic cancer group. BMC Cancer 2018; 18: 1298 [PMID: 30594153 DOI: 10.1186/s12885-018-5183-y]
- 212 Motoi F, Kosuge T, Ueno H, Yamaue H, Satoi S, Sho M, Honda G, Matsumoto I, Wada K, Furuse J, Matsuyama Y, Unno M; Study Group of Preoperative Therapy for Pancreatic Cancer (Prep) and Japanese Study Group of Adjuvant Therapy for Pancreatic cancer (JSAP). Randomized phase II/III trial of neoadjuvant chemotherapy with gemcitabine and S-1 vs upfront surgery for resectable pancreatic cancer (Prep-02/JSAP05). Jpn J Clin Oncol 2019; 49: 190-194 [PMID: 30608598 DOI: 10.1093/jjco/hyy190]
- 213 Okusaka T, Nakamura M, Yoshida M, Kitano M, Uesaka K, Ito Y, Furuse J, Hanada K, Okazaki K; Committee for Revision of Clinical Guidelines for Pancreatic Cancer of the Japan Pancreas Society. Clinical Practice Guidelines for Pancreatic Cancer 2019 From the Japan Pancreas Society: A Synopsis. Pancreas 2020; 49: 326-335 [PMID: 32132516 DOI: 10.1097/MPA.00000000001513]
- 214 Okusaka T, Furuse J. Recent advances in chemotherapy for pancreatic cancer: evidence from Japan and recommendations in guidelines. J Gastroenterol 2020; 55: 369-382 [PMID: 31997007 DOI: 10.1007/s00535-020-01666-y
- 215 UMIN-CTR Clinical Trial. Randomized phase II/III study of gemcitabine and nab-paclitaxel therapy vs S-1 and concurrent radiotherapy as neoadjuvant treatment for Borderline resectable pancreatic cancer. [cited 21 Dec 2019]. In: UMIN-CTR Clinical Trial [Internet]. Available from: https://upload.umin.ac.jp/cgi-open-bin/ctr\_e/ctr\_view.cgi?recptno=R000030821
- 216 Sahni S, Nahm C, Krisp C, Molloy MP, Mehta S, Maloney S, Itchins M, Pavlakis N, Clarke S, Chan D, Gill AJ, Howell VM, Samra J, Mittal A. Identification of Novel Biomarkers in Pancreatic Tumor Tissue to Predict Response to Neoadjuvant Chemotherapy. Front Oncol 2020; 10: 237 [PMID: 32195182 DOI: 10.3389/fonc.2020.00237]
- 217 Hammel P, Lacy J, Portales F, Sobrero AF, Pazo Cid RA, Mozo JLM, Terrebonne E, Dowden SD, Li JS, Ong TJ, Nydam T, Philip PA. Phase II LAPACT trial of nab-paclitaxel (nab-P) plus gemcitabine (G) for patients with locally advanced pancreatic cancer (LAPDAC). J Clin Oncol 2018; 36 (suppl 4): 204
- 218 Marthey L. Sa-Cunha A, Blanc JF, Gauthier M, Cueff A, Francois E, Trouilloud I, Malka D, Bachet JB, Coriat R, Terrebonne E, De La Fouchardière C, Manfredi S, Solub D, Lécaille C, Thirot Bidault A, Carbonnel F, Taieb J. FOLFIRINOX for locally advanced pancreatic adenocarcinoma: results of an AGEO multicenter prospective observational cohort. Ann Surg Oncol 2015; 22: 295-301 [PMID: 25037971 DOI: 10.1245/s10434-014-3898-9]
- 219 Weniger M, Moir J, Damm M, Maggino L, Kordes M, Rosendahl J, Ceyhan GO, Schorn S; RESPECT-study group. Respect - A multicenter retrospective study on preoperative chemotherapy in locally advanced and borderline resectable pancreatic cancer. Pancreatology 2020; 20: 1131-1138 [PMID: 32739267 DOI: 10.1016/j.pan.2020.06.012]
- 220 Suker M, Beumer BR, Sadot E, Marthey L, Faris JE, Mellon EA, El-Rayes BF, Wang-Gillam A, Lacy J, Hosein PJ, Moorcraft SY, Conroy T, Hohla F, Allen P, Taieb J, Hong TS, Shridhar R, Chau I, van Eijck CH, Koerkamp BG. FOLFIRINOX for locally advanced pancreatic cancer: a systematic review and patient-level meta-analysis. Lancet Oncol 2016; 17: 801-810 [PMID: 27160474 DOI: 10.1016/S1470-2045(16)00172-8]
- 221 Philip PA, Lacy J, Portales F, Sobrero A, Pazo-Cid R, Manzano Mozo JL, Kim EJ, Dowden S, Zakari A, Borg C, Terrebonne E, Rivera F, Sastre J, Bathini V, López-Trabada D, Asselah J, Saif MW, Shiansong Li J, Ong TJ, Nydam T, Hammel P. Nab-paclitaxel plus gemcitabine in patients with locally advanced pancreatic cancer (LAPACT): a multicentre, open-label phase 2 study. Lancet Gastroenterol Hepatol 2020; 5: 285-294 [PMID: 31953079 DOI: 10.1016/S2468-1253(19)30327-9]
- 222 Kunzmann V, Algül H, Goekkurt E, Siegler GM, Martens UM, Waldschmidt D, Pelzer U, Hennes E, Fuchs M, Siveke J, Kullmann F, Boeck S, Ettrich TJ, Ferenczy P, Keller R, Germer C, Stein H, Hartlapp I, Klein I, Heinemann V. Conversion rate in locally advanced pancreatic cancer (LAPDAC) after nab-paclitaxel/gemcitabine- or MFFX-based induction chemotherapy (NEOLAP): final results of a multicenter randomised phase II AIO trial. Ann Oncol 2019; 30: v253
- 223 Wolfe AR, Prabhakar D, Yildiz VO, Cloyd JM, Dillhoff M, Abushahin L, Alexandra Diaz D, Miller ED, Chen W, Frankel WL, Noonan A, Williams TM. Neoadjuvant-modified FOLFIRINOX vs nabpaclitaxel plus gemcitabine for borderline resectable or locally advanced pancreatic cancer patients



who achieved surgical resection. Cancer Med 2020; 9: 4711-4723 [PMID: 32415696 DOI: 10.1002/cam4.3075]

- Hammel P, Huguet F, van Laethem JL, Goldstein D, Glimelius B, Artru P, Borbath I, Bouché O, 224 Shannon J, André T, Mineur L, Chibaudel B, Bonnetain F, Louvet C; LAP07 Trial Group. Effect of Chemoradiotherapy vs Chemotherapy on Survival in Patients With Locally Advanced Pancreatic Cancer Controlled After 4 Months of Gemcitabine With or Without Erlotinib: The LAP07 Randomized Clinical Trial. JAMA 2016; 315: 1844-1853 [PMID: 27139057 DOI: 10.1001/jama.2016.4324]
- 225 Arcelli A, Buwenge M, Macchia G, Bertini F, Guido A, Deodato F, Cilla S, Scotti V, Rosetto ME, Djan I, Parisi S, Mattiucci GC, Cellini F, Fiore M, Bonomo P, Belgioia L, Niespolo RM, Gabriele P, Di Marco M, Simoni N, Mazzarotto R, Morganti AG; AIRO (Italian Association of Radiation Oncology and Clinical Oncology) Gastrointestinal Study Group. Stereotactic body radiotherapy vs conventionally fractionated chemoradiation in locally advanced pancreatic cancer: A multicenter case-control study (PAULA-1). Cancer Med 2020; 9: 7879-7887 [PMID: 32910549 DOI: 10.1002/cam4.3330]
- 226 Conroy T, Desseigne F, Ychou M, Bouché O, Guimbaud R, Bécouarn Y, Adenis A, Raoul JL, Gourgou-Bourgade S, de la Fouchardière C, Bennouna J, Bachet JB, Khemissa-Akouz F, Péré-Vergé D, Delbaldo C, Assenat E, Chauffert B, Michel P, Montoto-Grillot C, Ducreux M; Groupe Tumeurs Digestives of Unicancer; PRODIGE Intergroup. FOLFIRINOX vs genetitabine for metastatic pancreatic cancer. N Engl J Med 2011; 364: 1817-1825 [PMID: 21561347 DOI: 10.1056/NEJMoa1011923
- 227 Von Hoff DD, Ervin T, Arena FP, Chiorean EG, Infante J, Moore M, Seay T, Tjulandin SA, Ma WW, Saleh MN, Harris M, Reni M, Dowden S, Laheru D, Bahary N, Ramanathan RK, Tabernero J, Hidalgo M, Goldstein D, Van Cutsem E, Wei X, Iglesias J, Renschler MF. Increased survival in pancreatic cancer with nab-paclitaxel plus gemcitabine. N Engl J Med 2013; 369: 1691-1703 [PMID: 24131140 DOI: 10.1056/NEJMoa1304369]
- Chan KKW, Guo H, Cheng S, Beca JM, Redmond-Misner R, Isaranuwatchai W, Qiao L, Earle C, 228 Berry SR, Biagi JJ, Welch S, Meyers BM, Mittmann N, Coburn N, Arias J, Schwartz D, Dai WF, Gavura S, McLeod R, Kennedy ED. Real-world outcomes of FOLFIRINOX vs gemcitabine and nab-paclitaxel in advanced pancreatic cancer: A population-based propensity score-weighted analysis. Cancer Med 2020; 9: 160-169 [PMID: 31724340 DOI: 10.1002/cam4.2705]
- 229 Wang Y, Camateros P, Cheung WY. A Real-World Comparison of FOLFIRINOX, Gemcitabine Plus nab-Paclitaxel, and Gemcitabine in Advanced Pancreatic Cancers. J Gastrointest Cancer 2019; 50: 62-68 [PMID: 29143916 DOI: 10.1007/s12029-017-0028-5]
- 230 Wang-Gillam A, Li CP, Bodoky G, Dean A, Shan YS, Jameson G, Macarulla T, Lee KH, Cunningham D, Blanc JF, Hubner RA, Chiu CF, Schwartsmann G, Siveke JT, Braiteh F, Moyo V, Belanger B, Dhindsa N, Bayever E, Von Hoff DD, Chen LT; NAPOLI-1 Study Group. Nanoliposomal irinotecan with fluorouracil and folinic acid in metastatic pancreatic cancer after previous gemcitabine-based therapy (NAPOLI-1): a global, randomised, open-label, phase 3 trial. Lancet 2016; 387: 545-557 [PMID: 26615328 DOI: 10.1016/S0140-6736(15)00986-1]
- Golan T, Hammel P, Reni M, Van Cutsem E, Macarulla T, Hall MJ, Park JO, Hochhauser D, Arnold D, Oh DY, Reinacher-Schick A, Tortora G, Algül H, O'Reilly EM, McGuinness D, Cui KY, Schlienger K, Locker GY, Kindler HL. Maintenance Olaparib for Germline BRCA-Mutated Metastatic Pancreatic Cancer. N Engl J Med 2019; 381: 317-327 [PMID: 31157963 DOI: 10.1056/NEJMoa1903387
- 232 Alistar A, Morris BB, Desnoyer R, Klepin HD, Hosseinzadeh K, Clark C, Cameron A, Leyendecker J, D'Agostino R Jr, Topaloglu U, Boteju LW, Boteju AR, Shorr R, Zachar Z, Bingham PM, Ahmed T, Crane S, Shah R, Migliano JJ, Pardee TS, Miller L, Hawkins G, Jin G, Zhang W, Pasche B. Safety and tolerability of the first-in-class agent CPI-613 in combination with modified FOLFIRINOX in patients with metastatic pancreatic cancer: a single-centre, open-label, doseescalation, phase 1 trial. Lancet Oncol 2017; 18: 770-778 [PMID: 28495639 DOI: 10.1016/S1470-2045(17)30314-5
- 233 Jiang B, Zhou L, Lu J, Wang Y, Liu C, You L, Guo J. Stroma-Targeting Therapy in Pancreatic Cancer: One Coin With Two Sides? Front Oncol 2020; 10: 576399 [PMID: 33178608 DOI: 10.3389/fonc.2020.576399]
- 234 von Ahrens D, Bhagat TD, Nagrath D, Maitra A, Verma A. The role of stromal cancer-associated fibroblasts in pancreatic cancer. J Hematol Oncol 2017; 10: 76 [PMID: 28351381 DOI: 10.1186/s13045-017-0448-5]
- 235 Provenzano PP, Hingorani SR. Hyaluronan, fluid pressure, and stromal resistance in pancreas cancer. Br J Cancer 2013; 108: 1-8 [PMID: 23299539 DOI: 10.1038/bjc.2012.569]
- 236 Hingorani SR, Zheng L, Bullock AJ, Seery TE, Harris WP, Sigal DS, Braiteh F, Ritch PS, Zalupski MM, Bahary N, Oberstein PE, Wang-Gillam A, Wu W, Chondros D, Jiang P, Khelifa S, Pu J, Aldrich C, Hendifar AE. HALO 202: Randomized Phase II Study of PEGPH20 Plus Nab-Paclitaxel/Gemcitabine Versus Nab-Paclitaxel/Gemcitabine in Patients With Untreated, Metastatic Pancreatic Ductal Adenocarcinoma. J Clin Oncol 2018; 36: 359-366 [PMID: 29232172 DOI: 10.1200/JCO.2017.74.9564]
- 237 Catenacci DV, Junttila MR, Karrison T, Bahary N, Horiba MN, Nattam SR, Marsh R, Wallace J, Kozloff M, Rajdev L, Cohen D, Wade J, Sleckman B, Lenz HJ, Stiff P, Kumar P, Xu P, Henderson L, Takebe N, Salgia R, Wang X, Stadler WM, de Sauvage FJ, Kindler HL. Randomized Phase Ib/II Study of Gemcitabine Plus Placebo or Vismodegib, a Hedgehog Pathway Inhibitor, in Patients With



Metastatic Pancreatic Cancer. J Clin Oncol 2015; 33: 4284-4292 [PMID: 26527777 DOI: 10.1200/JCO.2015.62.8719]

- McCleary-Wheeler AL, Carr RM, Palmer SR, Smyrk TC, Allred JB, Almada LL, Tolosa EJ, 238 Lamberti MJ, Marks DL, Borad MJ, Molina JR, Qi Y, Lingle WL, Grothey A, Pitot HC, Jatoi A, Northfelt DW, Bryce AH, McWilliams RR, Okuno SH, Haluska P, Kim GP, Colon-Otero G, Lowe VJ, Callstrom MR, Ma WW, Bekaii-Saab T, Hung MC, Erlichman C, Fernandez-Zapico ME. Phase 1 trial of Vismodegib and Erlotinib combination in metastatic pancreatic cancer. Pancreatology 2020; 20: 101-109 [PMID: 31787526 DOI: 10.1016/j.pan.2019.11.011]
- 239 Murphy JE, Wo JY, Ryan DP, Clark JW, Jiang W, Yeap BY, Drapek LC, Ly L, Baglini CV, Blaszkowsky LS, Ferrone CR, Parikh AR, Weekes CD, Nipp RD, Kwak EL, Allen JN, Corcoran RB, Ting DT, Faris JE, Zhu AX, Goyal L, Berger DL, Qadan M, Lillemoe KD, Talele N, Jain RK, DeLaney TF, Duda DG, Boucher Y, Fernández-Del Castillo C, Hong TS. Total Neoadjuvant Therapy With FOLFIRINOX in Combination With Losartan Followed by Chemoradiotherapy for Locally Advanced Pancreatic Cancer: A Phase 2 Clinical Trial. JAMA Oncol 2019; 5: 1020-1027 [PMID: 31145418 DOI: 10.1001/jamaoncol.2019.0892]
- 240 Dimcevski G, Kotopoulis S, Bjånes T, Hoem D, Schjøtt J, Gjertsen BT, Biermann M, Molven A, Sorbye H, McCormack E, Postema M, Gilja OH. A human clinical trial using ultrasound and microbubbles to enhance gemcitabine treatment of inoperable pancreatic cancer. J Control Release 2016; 243: 172-181 [PMID: 27744037 DOI: 10.1016/j.jconrel.2016.10.007]
- Cooke VG, LeBleu VS, Keskin D, Khan Z, O'Connell JT, Teng Y, Duncan MB, Xie L, Maeda G, 241 Vong S, Sugimoto H, Rocha RM, Damascena A, Brentani RR, Kalluri R. Pericyte depletion results in hypoxia-associated epithelial-to-mesenchymal transition and metastasis mediated by met signaling pathway. Cancer Cell 2012; 21: 66-81 [PMID: 22264789 DOI: 10.1016/j.ccr.2011.11.024]
- 242 Veenstra VL, Damhofer H, Waasdorp C, van Rijssen LB, van de Vijver MJ, Dijk F, Wilmink HW, Besselink MG, Busch OR, Chang DK, Bailey PJ, Biankin AV, Kocher HM, Medema JP, Li JS, Jiang R, Pierce DW, van Laarhoven HWM, Bijlsma MF. ADAM12 is a circulating marker for stromal activation in pancreatic cancer and predicts response to chemotherapy. Oncogenesis 2018; 7: 87 [PMID: 30442938 DOI: 10.1038/s41389-018-0096-9]
- Griesmann H, Drexel C, Milosevic N, Sipos B, Rosendahl J, Gress TM, Michl P. Pharmacological 243 macrophage inhibition decreases metastasis formation in a genetic model of pancreatic cancer. Gut 2017; 66: 1278-1285 [PMID: 27013602 DOI: 10.1136/gutjnl-2015-310049]
- 244 Morrison AH, Byrne KT, Vonderheide RH. Immunotherapy and Prevention of Pancreatic Cancer. Trends Cancer 2018; 4: 418-428 [PMID: 29860986 DOI: 10.1016/j.trecan.2018.04.001]
- 245 Royal RE, Levy C, Turner K, Mathur A, Hughes M, Kammula US, Sherry RM, Topalian SL, Yang JC, Lowy I, Rosenberg SA. Phase 2 trial of single agent Ipilimumab (anti-CTLA-4) for locally advanced or metastatic pancreatic adenocarcinoma. J Immunother 2010; 33: 828-833 [PMID: 20842054 DOI: 10.1097/CJI.0b013e3181eec14c]
- Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, Drake CG, Camacho LH, Kauh 246 J, Odunsi K, Pitot HC, Hamid O, Bhatia S, Martins R, Eaton K, Chen S, Salay TM, Alaparthy S, Grosso JF, Korman AJ, Parker SM, Agrawal S, Goldberg SM, Pardoll DM, Gupta A, Wigginton JM. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. N Engl J Med 2012; 366: 2455-2465 [PMID: 22658128 DOI: 10.1056/NEJMoa1200694]
- 247 O'Reilly EM, Oh DY, Dhani N, Renouf DJ, Lee MA, Sun W, Fisher G, Hezel A, Chang SC, Vlahovic G, Takahashi O, Yang Y, Fitts D, Philip PA. Durvalumab With or Without Tremelimumab for Patients With Metastatic Pancreatic Ductal Adenocarcinoma: A Phase 2 Randomized Clinical Trial. JAMA Oncol 2019; 5: 1431-1438 [PMID: 31318392 DOI: 10.1001/jamaoncol.2019.1588]
- 248 Tumeh PC, Harview CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, Chmielowski B, Spasic M, Henry G, Ciobanu V, West AN, Carmona M, Kivork C, Seja E, Cherry G, Gutierrez AJ, Grogan TR, Mateus C, Tomasic G, Glaspy JA, Emerson RO, Robins H, Pierce RH, Elashoff DA, Robert C, Ribas A. PD-1 blockade induces responses by inhibiting adaptive immune resistance. Nature 2014; 515: 568-571 [PMID: 25428505 DOI: 10.1038/nature13954]
- 249 Humphris JL, Patch AM, Nones K, Bailey PJ, Johns AL, McKay S, Chang DK, Miller DK, Pajic M, Kassahn KS, Quinn MC, Bruxner TJ, Christ AN, Harliwong I, Idrisoglu S, Manning S, Nourse C, Nourbakhsh E, Stone A, Wilson PJ, Anderson M, Fink JL, Holmes O, Kazakoff S, Leonard C, Newell F, Waddell N, Wood S, Mead RS, Xu Q, Wu J, Pinese M, Cowley MJ, Jones MD, Nagrial AM, Chin VT, Chantrill LA, Mawson A, Chou A, Scarlett CJ, Pinho AV, Rooman I, Giry-Laterriere M, Samra JS, Kench JG, Merrett ND, Toon CW, Epari K, Nguyen NQ, Barbour A, Zeps N, Jamieson NB, McKay CJ, Carter CR, Dickson EJ, Graham JS, Duthie F, Oien K, Hair J, Morton JP, Sansom OJ, Grützmann R, Hruban RH, Maitra A, Iacobuzio-Donahue CA, Schulick RD, Wolfgang CL, Morgan RA, Lawlor RT, Rusev B, Corbo V, Salvia R, Cataldo I, Tortora G, Tempero MA; Australian Pancreatic Cancer Genome Initiative, Hofmann O, Eshleman JR, Pilarsky C, Scarpa A, Musgrove EA, Gill AJ, Pearson JV, Grimmond SM, Waddell N, Biankin AV. Hypermutation In Pancreatic Cancer. Gastroenterology 2017; 152: 68-74.e2 [PMID: 27856273 DOI: 10.1053/j.gastro.2016.09.060]
- Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, Skora AD, Luber BS, Azad NS, 250 Laheru D, Biedrzycki B, Donehower RC, Zaheer A, Fisher GA, Crocenzi TS, Lee JJ, Duffy SM, Goldberg RM, de la Chapelle A, Koshiji M, Bhaijee F, Huebner T, Hruban RH, Wood LD, Cuka N, Pardoll DM, Papadopoulos N, Kinzler KW, Zhou S, Cornish TC, Taube JM, Anders RA, Eshleman JR, Vogelstein B, Diaz LA Jr. PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. N Engl



J Med 2015; 372: 2509-2520 [PMID: 26028255 DOI: 10.1056/NEJMoa1500596]

- Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, Lu S, Kemberling H, Wilt C, 251 Luber BS, Wong F, Azad NS, Rucki AA, Laheru D, Donehower R, Zaheer A, Fisher GA, Crocenzi TS, Lee JJ, Greten TF, Duffy AG, Ciombor KK, Eyring AD, Lam BH, Joe A, Kang SP, Holdhoff M, Danilova L, Cope L, Meyer C, Zhou S, Goldberg RM, Armstrong DK, Bever KM, Fader AN, Taube J, Housseau F, Spetzler D, Xiao N, Pardoll DM, Papadopoulos N, Kinzler KW, Eshleman JR, Vogelstein B, Anders RA, Diaz LA Jr. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. Science 2017; 357: 409-413 [PMID: 28596308 DOI: 10.1126/science.aan6733]
- Wainberg ZA, Hochster HS, Kim EJH, George B, Kalyan A, Chiorean EG, Waterhouse DM, 252 Gutierrez M, Parikh AR, Jain R, Carrizosa DR, Soliman HH, Bhore R, Banerjee S, Lyons L, Louis CU, Ong TJ, O'Dwyer PJ. Phase I study of nivolumab (Nivo) + nab-paclitaxel (nab-P) + gemcitabine (Gem) in advanced pancreatic cancer (APDAC). J Clin Oncol 2019; 37: 298
- 253 Weiss GJ, Blaydorn L, Beck J, Bornemann-Kolatzki K, Urnovitz H, Schütz E, Khemka V. Phase Ib/II study of gemcitabine, nab-paclitaxel, and pembrolizumab in metastatic pancreatic adenocarcinoma. Invest New Drugs 2018; 36: 96-102 [PMID: 29119276 DOI: 10.1007/s10637-017-0525-1]
- National Institute of Public Health. ONO-4538 Phase II Study (ONO-4538–83/TASUKI-83). 254 [cited 21 Dec 2019]. In: National Institute of Public Health [Internet]. Available from: https://rctportal.niph.go.jp/en/detail?trial id=JapicCTI-184230
- 255 Wang-Gillam A, O'Reilly EM, Bendell JC, Wainberg ZA, Borazanci EH, Bahary N, O'Hara MH, Beatty GL, Pant S, Cohen DJ, Leong S, Beg MS, Yu KH, Evans TRJ, Seufferlein T, Okusaka T, Phillips P, Liu X, Perna SK, Le DT. A randomized phase II study of cabiralizumab (cabira) + nivolumab (nivo)  $\pm$  chemotherapy (chemo) in advanced pancreatic ductal adenocarcinoma (PDAC). J Clin Oncol 2019: 37: TPS465
- 256 Schizas D, Charalampakis N, Kole C, Economopoulou P, Koustas E, Gkotsis E, Ziogas D, Psyrri A, Karamouzis MV. Immunotherapy for pancreatic cancer: A 2020 update. Cancer Treat Rev 2020; 86: 102016 [PMID: 32247999 DOI: 10.1016/j.ctrv.2020.102016]
- 257 Le DT, Picozzi VJ, Ko AH, Wainberg ZA, Kindler H, Wang-Gillam A, Oberstein P, Morse MA, Zeh HJ 3rd, Weekes C, Reid T, Borazanci E, Crocenzi T, LoConte NK, Musher B, Laheru D, Murphy A, Whiting C, Nair N, Enstrom A, Ferber S, Brockstedt DG, Jaffee EM. Results from a Phase IIb, Randomized, Multicenter Study of GVAX Pancreas and CRS-207 Compared with Chemotherapy in Adults with Previously Treated Metastatic Pancreatic Adenocarcinoma (ECLIPSE Study). Clin Cancer Res 2019; 25: 5493-5502 [PMID: 31126960 DOI: 10.1158/1078-0432.CCR-18-2992]
- 258 Asahara S, Takeda K, Yamao K, Maguchi H, Yamaue H. Phase I/II clinical trial using HLA-A24restricted peptide vaccine derived from KIF20A for patients with advanced pancreatic cancer. J Transl Med 2013; 11: 291 [PMID: 24237633 DOI: 10.1186/1479-5876-11-291]
- 259 Suzuki N, Hazama S, Iguchi H, Uesugi K, Tanaka H, Hirakawa K, Aruga A, Hatori T, Ishizaki H, Umeda Y, Fujiwara T, Ikemoto T, Shimada M, Yoshimatsu K, Shimizu R, Hayashi H, Sakata K, Takenouchi H, Matsui H, Shindo Y, Iida M, Koki Y, Arima H, Furukawa H, Ueno T, Yoshino S, Nakamura Y, Oka M, Nagano H. Phase II clinical trial of peptide cocktail therapy for patients with advanced pancreatic cancer: VENUS-PC study. Cancer Sci 2017; 108: 73-80 [PMID: 27783849 DOI: 10.1111/cas.13113]
- Miyazawa M, Katsuda M, Maguchi H, Katanuma A, Ishii H, Ozaka M, Yamao K, Imaoka H, Kawai 260 M, Hirono S, Okada KI, Yamaue H. Phase II clinical trial using novel peptide cocktail vaccine as a postoperative adjuvant treatment for surgically resected pancreatic cancer patients. Int J Cancer 2017; 140: 973-982 [PMID: 27861852 DOI: 10.1002/ijc.30510]
- 261 Wedén S, Klemp M, Gladhaug IP, Møller M, Eriksen JA, Gaudernack G, Buanes T. Long-term follow-up of patients with resected pancreatic cancer following vaccination against mutant K-ras. Int J Cancer 2011; 128: 1120-1128 [PMID: 20473937 DOI: 10.1002/ijc.25449]
- 262 Toubaii A. Achtar M. Provenzano M. Herrin VE. Behrens R. Hamilton M. Bernstein S. Venzon D. Gause B, Marincola F, Khleif SN. Pilot study of mutant ras peptide-based vaccine as an adjuvant treatment in pancreatic and colorectal cancers. Cancer Immunol Immunother 2008; 57: 1413-1420 [PMID: 18297281 DOI: 10.1007/s00262-008-0477-6]
- 263 Abou-Alfa GK, Chapman PB, Feilchenfeldt J, Brennan MF, Capanu M, Gansukh B, Jacobs G, Levin A, Neville D, Kelsen DP, O'Reilly EM. Targeting mutated K-ras in pancreatic adenocarcinoma using an adjuvant vaccine. Am J Clin Oncol 2011; 34: 321-325 [PMID: 20686403 DOI: 10.1097/COC.0b013e3181e84b1f]
- 264 Cohn A, Morse MA, O'Neil B, Whiting S, Coeshott C, Ferraro J, Bellgrau D, Apelian D, Rodell TC. Whole Recombinant Saccharomyces cerevisiae Yeast Expressing Ras Mutations as Treatment for Patients With Solid Tumors Bearing Ras Mutations: Results From a Phase 1 Trial. J Immunother 2018; **41**: 141-150 [PMID: 29528991 DOI: 10.1097/CJI.0000000000000219]
- 265 Katsuda M, Miyazawa M, Kawai M, Hirono S, Okada KI, Shimizu A, Kitahata Y, Yamaue H, A phase III, double-blind, randomized clinical trial comparing S-1 in combination with DC vaccine loaded with WT1 peptides (TLP0-001) or placebo for the patients with advanced pancreatic cancer refractory to standard chemotherapy. J Clin Oncol 2017; 35: TPS4153
- 266 Katsuda M, Miyazawa M, Ojima T, Katanuma A, Hakamada K, Sudo K, Asahara S, Endo I, Ueno M, Hara K, Yamada S, Fujii T, Satoi S, Ioka T, Ohira M, Akahori T, Kitano M, Nagano H, Furukawa M, Adachi T, Yamaue H. A double-blind randomized comparative clinical trial to



evaluate the safety and efficacy of dendritic cell vaccine loaded with WT1 peptides (TLP0-001) in combination with S-1 in patients with advanced pancreatic cancer refractory to standard chemotherapy. Trials 2019; 20: 242 [PMID: 31029154 DOI: 10.1186/s13063-019-3332-5]

- 267 Liu J, Zhong JF, Zhang X, Zhang C. Allogeneic CD19-CAR-T cell infusion after allogeneic hematopoietic stem cell transplantation in B cell malignancies. J Hematol Oncol 2017; 10: 35 [PMID: 28143567 DOI: 10.1186/s13045-017-0405-3]
- 268 June CH, O'Connor RS, Kawalekar OU, Ghassemi S, Milone MC. CAR T cell immunotherapy for human cancer. Science 2018; 359: 1361-1365 [PMID: 29567707 DOI: 10.1126/science.aar6711]
- Wu J, Cai J. Dilemma and Challenge of Immunotherapy for Pancreatic Cancer. Dig Dis Sci 2021; 269 66: 359-368 [PMID: 32140943 DOI: 10.1007/s10620-020-06183-9]
- 270 Beatty GL, O'Hara MH, Lacey SF, Torigian DA, Nazimuddin F, Chen F, Kulikovskaya IM, Soulen MC, McGarvey M, Nelson AM, Gladney WL, Levine BL, Melenhorst JJ, Plesa G, June CH. Activity of Mesothelin-Specific Chimeric Antigen Receptor T Cells Against Pancreatic Carcinoma Metastases in a Phase 1 Trial. Gastroenterology 2018; 155: 29-32 [PMID: 29567081 DOI: 10.1053/j.gastro.2018.03.029]
- Le DT, Lutz E, Uram JN, Sugar EA, Onners B, Solt S, Zheng L, Diaz LA Jr, Donehower RC, Jaffee 271 EM, Laheru DA. Evaluation of ipilimumab in combination with allogeneic pancreatic tumor cells transfected with a GM-CSF gene in previously treated pancreatic cancer. J Immunother 2013; 36: 382-389 [PMID: 23924790 DOI: 10.1097/CJI.0b013e31829fb7a2]
- Chung V, Kos FJ, Hardwick N, Yuan Y, Chao J, Li D, Waisman J, Li M, Zurcher K, Frankel P, 272 Diamond DJ. Evaluation of safety and efficacy of p53MVA vaccine combined with pembrolizumab in patients with advanced solid cancers. Clin Transl Oncol 2019; 21: 363-372 [PMID: 30094792 DOI: 10.1007/s12094-018-1932-2]
- 273 Wang-Gillam A, Lockhart AC, Tan BR, Suresh R, Lim KH, Ratner L, Morton A, Huffman J, Marquez S, Boice N, DeNardo DG. Phase I study of defactinib combined with pembrolizumab and gemcitabine in patients with advanced cancer. J Clin Oncol 2018; 36: 2561
- 274 Reiss KA, Mick R, O'Hara MH, Teitelbaum UR, Karasic TB, Schneider CJ, O'Dwyer PJ, Karlson D, Cowden S, Fuhrer MJ, Carpenter EL, Pantel AA, Makvandi M, Mankoff DA, Nathanson K, Maxwell KN, Beatty GL, Domchek SM. A randomized phase II trial of niraparib plus either nivolumab or ipilimumab in patients with advanced pancreatic cancer whose cancer has not progressed on platinum-based therapy. J Clin Oncol 2019; 37: TPS4161
- 275 Desai J, Kortmansky JS, Segal NH, Fakih M, Oh DY, Kim KP, Rahma OE, Ko AH, Chung HC, Alsina M, Yeh KH, Li S, Al-Sakaff NJA, Patel J, Barak H, Wang J, Zhang X, Bleul C, Cha E, Lee J. MORPHEUS: A phase Ib/II study platform evaluating the safety and clinical efficacy of cancer immunotherapy (CIT)-based combinations in gastrointestinal (GI) cancers. J Clin Oncol 2019; 37: **TPS467**
- 276 Rahal A, Musher B. Oncolytic viral therapy for pancreatic cancer. J Surg Oncol 2017; 116: 94-103 [PMID: 28407327 DOI: 10.1002/jso.24626]
- 277 Eissa IR, Bustos-Villalobos I, Ichinose T, Matsumura S, Naoe Y, Miyajima N, Morimoto D, Mukoyama N, Zhiwen W, Tanaka M, Hasegawa H, Sumigama S, Aleksic B, Kodera Y, Kasuya H. The Current Status and Future Prospects of Oncolytic Viruses in Clinical Trials against Melanoma, Glioma, Pancreatic, and Breast Cancers. Cancers (Basel) 2018; 10 [PMID: 30261620 DOI: 10.3390/cancers10100356
- Chang KJ, Senzer NN, Binmoeller K, Goldsweig H, Coffin R. Phase I dose-escalation study of 278 talimogene laherparepvec (T-VEC) for advanced pancreatic cancer (ca). J Clin Oncol 2012; 30: e14546
- 279 Noonan AM, Farren MR, Geyer SM, Huang Y, Tahiri S, Ahn D, Mikhail S, Ciombor KK, Pant S, Aparo S, Sexton J, Marshall JL, Mace TA, Wu CS, El-Rayes B, Timmers CD, Zwiebel J, Lesinski GB, Villalona-Calero MA, Bekaii-Saab TS. Randomized Phase 2 Trial of the Oncolytic Virus Pelareorep (Reolysin) in Upfront Treatment of Metastatic Pancreatic Adenocarcinoma. Mol Ther 2016; 24: 1150-1158 [PMID: 27039845 DOI: 10.1038/mt.2016.66]
- 280 Mahalingam D, Wilkinson GA, Eng KH, Fields P, Raber P, Moseley JL, Cheetham K, Coffey M, Nuovo G, Kalinski P, Zhang B, Arora SP, Fountzilas C. Pembrolizumab in Combination with the Oncolytic Virus Pelareorep and Chemotherapy in Patients with Advanced Pancreatic Adenocarcinoma: A Phase Ib Study. Clin Cancer Res 2020; 26: 71-81 [PMID: 31694832 DOI: 10.1158/1078-0432.CCR-19-2078
- 281 Haller SD, Monaco ML, Essani K. The Present Status of Immuno-Oncolytic Viruses in the Treatment of Pancreatic Cancer. Viruses 2020; 12 [PMID: 33213031 DOI: 10.3390/v12111318]
- 282 Aguirre AJ, Nowak JA, Camarda ND, Moffitt RA, Ghazani AA, Hazar-Rethinam M, Raghavan S, Kim J, Brais LK, Ragon D, Welch MW, Reilly E, McCabe D, Marini L, Anderka K, Helvie K, Oliver N, Babic A, Da Silva A, Nadres B, Van Seventer EE, Shahzade HA, St Pierre JP, Burke KP, Clancy T, Cleary JM, Doyle LA, Jajoo K, McCleary NJ, Meyerhardt JA, Murphy JE, Ng K, Patel AK, Perez K, Rosenthal MH, Rubinson DA, Ryou M, Shapiro GI, Sicinska E, Silverman SG, Nagy RJ, Lanman RB, Knoerzer D, Welsch DJ, Yurgelun MB, Fuchs CS, Garraway LA, Getz G, Hornick JL, Johnson BE, Kulke MH, Mayer RJ, Miller JW, Shyn PB, Tuveson DA, Wagle N, Yeh JJ, Hahn WC, Corcoran RB, Carter SL, Wolpin BM. Real-time Genomic Characterization of Advanced Pancreatic Cancer to Enable Precision Medicine. Cancer Discov 2018: 8: 1096-1111 [PMID: 29903880 DOI: 10.1158/2159-8290.CD-18-0275]
- Bernabe-Ramirez C, Patel R, Chahal J, Saif MW. Treatment options in BRAF-mutant metastatic



colorectal cancer. Anticancer Drugs 2020; 31: 545-557 [PMID: 32304411 DOI: 10.1097/CAD.00000000000940]

- 284 O'Reilly EM, Hechtman JF. Tumour response to TRK inhibition in a patient with pancreatic adenocarcinoma harbouring an NTRK gene fusion. Ann Oncol 2019; 30: viii36-viii40 [PMID: 31605106 DOI: 10.1093/annonc/mdz385]
- Solomon JP, Linkov I, Rosado A, Mullaney K, Rosen EY, Frosina D, Jungbluth AA, Zehir A, 285 Benayed R, Drilon A, Hyman DM, Ladanyi M, Sireci AN, Hechtman JF. NTRK fusion detection across multiple assays and 33,997 cases: diagnostic implications and pitfalls. Mod Pathol 2020; 33: 38-46 [PMID: 31375766 DOI: 10.1038/s41379-019-0324-7]
- 286 Aguirre AJ. Oncogenic NRG1 Fusions: A New Hope for Targeted Therapy in Pancreatic Cancer. Clin Cancer Res 2019; 25: 4589-4591 [PMID: 31164372 DOI: 10.1158/1078-0432.CCR-19-1280]
- 287 Jones MR, Williamson LM, Topham JT, Lee MKC, Goytain A, Ho J, Denroche RE, Jang G, Pleasance E, Shen Y, Karasinska JM, McGhie JP, Gill S, Lim HJ, Moore MJ, Wong HL, Ng T, Yip S, Zhang W, Sadeghi S, Reisle C, Mungall AJ, Mungall KL, Moore RA, Ma Y, Knox JJ, Gallinger S, Laskin J, Marra MA, Schaeffer DF, Jones SJM, Renouf DJ. NRG1 Gene Fusions Are Recurrent, Clinically Actionable Gene Rearrangements in KRAS Wild-Type Pancreatic Ductal Adenocarcinoma. Clin Cancer Res 2019; 25: 4674-4681 [PMID: 31068372 DOI: 10.1158/1078-0432.CCR-19-0191]
- 288 Thompson ED, Roberts NJ, Wood LD, Eshleman JR, Goggins MG, Kern SE, Klein AP, Hruban RH. The genetics of ductal adenocarcinoma of the pancreas in the year 2020: dramatic progress, but far to go. Mod Pathol 2020; 33: 2544-2563 [PMID: 32704031 DOI: 10.1038/s41379-020-0629-6]
- 289 Chantrill LA, Nagrial AM, Watson C, Johns AL, Martyn-Smith M, Simpson S, Mead S, Jones MD, Samra JS, Gill AJ, Watson N, Chin VT, Humphris JL, Chou A, Brown B, Morey A, Pajic M, Grimmond SM, Chang DK, Thomas D, Sebastian L, Sjoquist K, Yip S, Pavlakis N, Asghari R, Harvey S, Grimison P, Simes J, Biankin AV; Australian Pancreatic Cancer Genome Initiative (APGI); Individualized Molecular Pancreatic Cancer Therapy (IMPaCT) Trial Management Committee of the Australasian Gastrointestinal Trials Group (AGITG). Precision Medicine for Advanced Pancreas Cancer: The Individualized Molecular Pancreatic Cancer Therapy (IMPaCT) Trial. Clin Cancer Res 2015; 21: 2029-2037 [PMID: 25896973 DOI: 10.1158/1078-0432.CCR-15-0426
- Kuo KK, Hsiao PJ, Chang WT, Chuang SC, Yang YH, Wuputra K, Ku CC, Pan JB, Li CP, Kato K, 290 Liu CJ, Wu DC, Yokoyama KK. Therapeutic Strategies Targeting Tumor Suppressor Genes in Pancreatic Cancer. Cancers (Basel) 2021; 13 [PMID: 34359820 DOI: 10.3390/cancers13153920]
- Yarchoan M, Myzak MC, Johnson BA 3rd, De Jesus-Acosta A, Le DT, Jaffee EM, Azad NS, 291 Donehower RC, Zheng L, Oberstein PE, Fine RL, Laheru DA, Goggins M. Olaparib in combination with irinotecan, cisplatin, and mitomycin C in patients with advanced pancreatic cancer. Oncotarget 2017; 8: 44073-44081 [PMID: 28454122 DOI: 10.18632/oncotarget.17237]
- 292 Aung KL, Fischer SE, Denroche RE, Jang GH, Dodd A, Creighton S, Southwood B, Liang SB, Chadwick D, Zhang A, O'Kane GM, Albaba H, Moura S, Grant RC, Miller JK, Mbabaali F, Pasternack D, Lungu IM, Bartlett JMS, Ghai S, Lemire M, Holter S, Connor AA, Moffitt RA, Yeh JJ, Timms L, Krzyzanowski PM, Dhani N, Hedley D, Notta F, Wilson JM, Moore MJ, Gallinger S, Knox JJ. Genomics-Driven Precision Medicine for Advanced Pancreatic Cancer: Early Results from the COMPASS Trial. Clin Cancer Res 2018; 24: 1344-1354 [PMID: 29288237 DOI: 10.1158/1078-0432.CCR-17-2994]
- Singhi AD, George B, Greenbowe JR, Chung J, Suh J, Maitra A, Klempner SJ, Hendifar A, Milind 293 JM, Golan T, Brand RE, Zureikat AH, Roy S, Schrock AB, Miller VA, Ross JS, Ali SM, Bahary N. Real-Time Targeted Genome Profile Analysis of Pancreatic Ductal Adenocarcinomas Identifies Genetic Alterations That Might Be Targeted With Existing Drugs or Used as Biomarkers. Gastroenterology 2019; 156: 2242-2253.e4 [PMID: 30836094 DOI: 10.1053/j.gastro.2019.02.037]
- 294 Luchini C, Brosens LAA, Wood LD, Chatterjee D, Shin JI, Sciammarella C, Fiadone G, Malleo G, Salvia R, Kryklyva V, Piredda ML, Cheng L, Lawlor RT, Adsay V, Scarpa A. Comprehensive characterisation of pancreatic ductal adenocarcinoma with microsatellite instability: histology, molecular pathology and clinical implications. Gut 2021; 70: 148-156 [PMID: 32350089 DOI: 10.1136/gutjnl-2020-320726
- Choi M, Kipps T, Kurzrock R. ATM Mutations in Cancer: Therapeutic Implications. Mol Cancer 295 Ther 2016; 15: 1781-1791 [PMID: 27413114 DOI: 10.1158/1535-7163.MCT-15-0945]
- Nanda N, Roberts NJ. ATM Serine/Threonine Kinase and its Role in Pancreatic Risk. Genes (Basel) 296 2020; 11 [PMID: 31963441 DOI: 10.3390/genes11010108]
- Mukhopadhyay S, Goswami D, Adiseshaiah PP, Burgan W, Yi M, Guerin TM, Kozlov SV, Nissley 297 DV, McCormick F. Undermining Glutaminolysis Bolsters Chemotherapy While NRF2 Promotes Chemoresistance in KRAS-Driven Pancreatic Cancers. Cancer Res 2020; 80: 1630-1643 [PMID: 31911550 DOI: 10.1158/0008-5472.CAN-19-1363]
- Janes MR, Zhang J, Li LS, Hansen R, Peters U, Guo X, Chen Y, Babbar A, Firdaus SJ, Darjania L, 298 Feng J, Chen JH, Li S, Long YO, Thach C, Liu Y, Zarieh A, Ely T, Kucharski JM, Kessler LV, Wu T, Yu K, Wang Y, Yao Y, Deng X, Zarrinkar PP, Brehmer D, Dhanak D, Lorenzi MV, Hu-Lowe D, Patricelli MP, Ren P. Targeting KRAS Mutant Cancers with a Covalent G12C-Specific Inhibitor. Cell 2018; 172: 578-589.e17 [PMID: 29373830 DOI: 10.1016/j.cell.2018.01.006]
- 299 Nagasaka M, Li Y, Sukari A, Ou SI, Al-Hallak MN, Azmi AS. KRAS G12C Game of Thrones, which direct KRAS inhibitor will claim the iron throne? Cancer Treat Rev 2020; 84: 101974 [PMID:



#### 32014824 DOI: 10.1016/j.ctrv.2020.101974]

- 300 Jones S, Zhang X, Parsons DW, Lin JC, Leary RJ, Angenendt P, Mankoo P, Carter H, Kamiyama H, Jimeno A, Hong SM, Fu B, Lin MT, Calhoun ES, Kamiyama M, Walter K, Nikolskaya T, Nikolsky Y, Hartigan J, Smith DR, Hidalgo M, Leach SD, Klein AP, Jaffee EM, Goggins M, Maitra A, Iacobuzio-Donahue C, Eshleman JR, Kern SE, Hruban RH, Karchin R, Papadopoulos N, Parmigiani G, Vogelstein B, Velculescu VE, Kinzler KW. Core signaling pathways in human pancreatic cancers revealed by global genomic analyses. Science 2008; 321: 1801-1806 [PMID: 18772397 DOI: 10.1126/science.1164368]
- 301 Waddell N, Pajic M, Patch AM, Chang DK, Kassahn KS, Bailey P, Johns AL, Miller D, Nones K, Quek K, Quinn MC, Robertson AJ, Fadlullah MZ, Bruxner TJ, Christ AN, Harliwong I, Idrisoglu S, Manning S, Nourse C, Nourbakhsh E, Wani S, Wilson PJ, Markham E, Cloonan N, Anderson MJ, Fink JL, Holmes O, Kazakoff SH, Leonard C, Newell F, Poudel B, Song S, Taylor D, Waddell N, Wood S, Xu Q, Wu J, Pinese M, Cowley MJ, Lee HC, Jones MD, Nagrial AM, Humphris J, Chantrill LA, Chin V, Steinmann AM, Mawson A, Humphrey ES, Colvin EK, Chou A, Scarlett CJ, Pinho AV, Giry-Laterriere M, Rooman I, Samra JS, Kench JG, Pettitt JA, Merrett ND, Toon C, Epari K, Nguyen NO, Barbour A, Zeps N, Jamieson NB, Graham JS, Niclou SP, Bjerkvig R, Grützmann R, Aust D, Hruban RH, Maitra A, Iacobuzio-Donahue CA, Wolfgang CL, Morgan RA, Lawlor RT, Corbo V, Bassi C, Falconi M, Zamboni G, Tortora G, Tempero MA; Australian Pancreatic Cancer Genome Initiative, Gill AJ, Eshleman JR, Pilarsky C, Scarpa A, Musgrove EA, Pearson JV, Biankin AV, Grimmond SM. Whole genomes redefine the mutational landscape of pancreatic cancer. Nature 2015; 518: 495-501 [PMID: 25719666 DOI: 10.1038/nature14169]
- 302 Cani PD. Human gut microbiome: hopes, threats and promises. Gut 2018; 67: 1716-1725 [PMID: 29934437 DOI: 10.1136/gutjnl-2018-316723]
- 303 Maekawa T, Fukaya R, Takamatsu S, Itoyama S, Fukuoka T, Yamada M, Hata T, Nagaoka S, Kawamoto K, Eguchi H, Murata K, Kumada T, Ito T, Tanemura M, Fujimoto K, Tomita Y, Tobe T, Kamada Y, Miyoshi E. Possible involvement of Enterococcus infection in the pathogenesis of chronic pancreatitis and cancer. Biochem Biophys Res Commun 2018; 506: 962-969 [PMID: 30401562 DOI: 10.1016/j.bbrc.2018.10.169]
- 304 Wang S, Dong W, Liu L, Xu M, Wang Y, Liu T, Zhang Y, Wang B, Cao H. Interplay between bile acids and the gut microbiota promotes intestinal carcinogenesis. Mol Carcinog 2019; 58: 1155-1167 [PMID: 30828892 DOI: 10.1002/mc.22999]
- 305 Cougnoux A, Dalmasso G, Martinez R, Buc E, Delmas J, Gibold L, Sauvanet P, Darcha C, Déchelotte P, Bonnet M, Pezet D, Wodrich H, Darfeuille-Michaud A, Bonnet R, Bacterial genotoxin colibactin promotes colon tumour growth by inducing a senescence-associated secretory phenotype. Gut 2014; 63: 1932-1942 [PMID: 24658599 DOI: 10.1136/gutjnl-2013-305257]
- 306 Nougayrède JP, Taieb F, De Rycke J, Oswald E. Cyclomodulins: bacterial effectors that modulate the eukaryotic cell cycle. Trends Microbiol 2005; 13: 103-110 [PMID: 15737728 DOI: 10.1016/j.tim.2005.01.002
- 307 Bao Y, Spiegelman D, Li R, Giovannucci E, Fuchs CS, Michaud DS. History of peptic ulcer disease and pancreatic cancer risk in men. Gastroenterology 2010; 138: 541-549 [PMID: 19818786 DOI: 10.1053/j.gastro.2009.09.059
- 308 Mitsuhashi K, Nosho K, Sukawa Y, Matsunaga Y, Ito M, Kurihara H, Kanno S, Igarashi H, Naito T, Adachi Y, Tachibana M, Tanuma T, Maguchi H, Shinohara T, Hasegawa T, Imamura M, Kimura Y, Hirata K, Maruyama R, Suzuki H, Imai K, Yamamoto H, Shinomura Y. Association of Fusobacterium species in pancreatic cancer tissues with molecular features and prognosis. Oncotarget 2015; 6: 7209-7220 [PMID: 25797243 DOI: 10.18632/oncotarget.3109]
- 309 Fan X, Alekseyenko AV, Wu J, Peters BA, Jacobs EJ, Gapstur SM, Purdue MP, Abnet CC, Stolzenberg-Solomon R, Miller G, Ravel J, Haves RB, Ahn J, Human oral microbiome and prospective risk for pancreatic cancer: a population-based nested case-control study. Gut 2018; 67: 120-127 [PMID: 27742762 DOI: 10.1136/gutjnl-2016-312580]
- 310 Li Q, Jin M, Liu Y, Jin L. Gut Microbiota: Its Potential Roles in Pancreatic Cancer. Front Cell Infect Microbiol 2020; 10: 572492 [PMID: 33117731 DOI: 10.3389/fcimb.2020.572492]
- Chakladar J, Kuo SZ, Castaneda G, Li WT, Gnanasekar A, Yu MA, Chang EY, Wang XQ, 311 Ongkeko WM. The Pancreatic Microbiome is Associated with Carcinogenesis and Worse Prognosis in Males and Smokers. Cancers (Basel) 2020; 12 [PMID: 32962112 DOI: 10.3390/cancers12092672]
- 312 Rogers CJ, Prabhu KS, Vijay-Kumar M. The microbiome and obesity-an established risk for certain types of cancer. Cancer J 2014; 20: 176-180 [PMID: 24855004 DOI: 10.1097/PPO.000000000000049]
- Qin J, Li Y, Cai Z, Li S, Zhu J, Zhang F, Liang S, Zhang W, Guan Y, Shen D, Peng Y, Zhang D, Jie 313 Z, Wu W, Qin Y, Xue W, Li J, Han L, Lu D, Wu P, Dai Y, Sun X, Li Z, Tang A, Zhong S, Li X, Chen W, Xu R, Wang M, Feng Q, Gong M, Yu J, Zhang Y, Zhang M, Hansen T, Sanchez G, Raes J, Falony G, Okuda S, Almeida M, LeChatelier E, Renault P, Pons N, Batto JM, Zhang Z, Chen H, Yang R, Zheng W, Yang H, Wang J, Ehrlich SD, Nielsen R, Pedersen O, Kristiansen K. A metagenome-wide association study of gut microbiota in type 2 diabetes. Nature 2012; 490: 55-60 [PMID: 23023125 DOI: 10.1038/nature11450]
- 314 Sousa T, Paterson R, Moore V, Carlsson A, Abrahamsson B, Basit AW. The gastrointestinal microbiota as a site for the biotransformation of drugs. Int J Pharm 2008; 363: 1-25 [PMID: 18682282 DOI: 10.1016/j.ijpharm.2008.07.009]



- 315 Iida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, Molina DA, Salcedo R, Back T, Cramer S, Dai RM, Kiu H, Cardone M, Naik S, Patri AK, Wang E, Marincola FM, Frank KM, Belkaid Y, Trinchieri G, Goldszmid RS. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. Science 2013; 342: 967-970 [PMID: 24264989 DOI: 10.1126/science.1240527]
- 316 Vande Voorde J, Sabuncuoğlu S, Noppen S, Hofer A, Ranjbarian F, Fieuws S, Balzarini J, Liekens S. Nucleoside-catabolizing enzymes in mycoplasma-infected tumor cell cultures compromise the cytostatic activity of the anticancer drug gemcitabine. J Biol Chem 2014; 289: 13054-13065 [PMID: 24668817 DOI: 10.1074/jbc.M114.558924]
- 317 Pushalkar S, Hundeyin M, Daley D, Zambirinis CP, Kurz E, Mishra A, Mohan N, Aykut B, Usyk M, Torres LE, Werba G, Zhang K, Guo Y, Li Q, Akkad N, Lall S, Wadowski B, Gutierrez J, Kochen Rossi JA, Herzog JW, Diskin B, Torres-Hernandez A, Leinwand J, Wang W, Taunk PS, Savadkar S, Janal M, Saxena A, Li X, Cohen D, Sartor RB, Saxena D, Miller G. The Pancreatic Cancer Microbiome Promotes Oncogenesis by Induction of Innate and Adaptive Immune Suppression. Cancer Discov 2018; 8: 403-416 [PMID: 29567829 DOI: 10.1158/2159-8290.CD-17-1134]
- Geller LT, Barzily-Rokni M, Danino T, Jonas OH, Shental N, Nejman D, Gavert N, Zwang Y, 318 Cooper ZA, Shee K, Thaiss CA, Reuben A, Livny J, Avraham R, Frederick DT, Ligorio M, Chatman K, Johnston SE, Mosher CM, Brandis A, Fuks G, Gurbatri C, Gopalakrishnan V, Kim M, Hurd MW, Katz M, Fleming J, Maitra A, Smith DA, Skalak M, Bu J, Michaud M, Trauger SA, Barshack I, Golan T, Sandbank J, Flaherty KT, Mandinova A, Garrett WS, Thayer SP, Ferrone CR, Huttenhower C, Bhatia SN, Gevers D, Wargo JA, Golub TR, Straussman R. Potential role of intratumor bacteria in mediating tumor resistance to the chemotherapeutic drug gemcitabine. Science 2017; 357: 1156-1160 [PMID: 28912244 DOI: 10.1126/science.aah5043]
- 319 Lehouritis P, Cummins J, Stanton M, Murphy CT, McCarthy FO, Reid G, Urbaniak C, Byrne WL, Tangnev M. Local bacteria affect the efficacy of chemotherapeutic drugs. Sci Rep 2015: 5: 14554 [PMID: 26416623 DOI: 10.1038/srep14554]
- Kamarajah SK, Burns WR, Frankel TL, Cho CS, Nathan H. Validation of the American Joint 320 Commission on Cancer (AJCC) 8th Edition Staging System for Patients with Pancreatic Adenocarcinoma: A Surveillance, Epidemiology and End Results (SEER) Analysis. Ann Surg Oncol 2017; 24: 2023-2030 [PMID: 28213792 DOI: 10.1245/s10434-017-5810-x]
- 321 Raut CP, Tseng JF, Sun CC, Wang H, Wolff RA, Crane CH, Hwang R, Vauthey JN, Abdalla EK, Lee JE, Pisters PW, Evans DB. Impact of resection status on pattern of failure and survival after pancreaticoduodenectomy for pancreatic adenocarcinoma. Ann Surg 2007; 246: 52-60 [PMID: 17592291 DOI: 10.1097/01.sla.0000259391.84304.2b]
- Crippa S, Guarneri G, Belfiori G, Partelli S, Pagnanelli M, Gasparini G, Balzano G, Lena MS, 322 Rubini C, Doglioni C, Zamboni G, Falconi M. Positive neck margin at frozen section analysis is a significant predictor of tumour recurrence and poor survival after pancreatodudenectomy for pancreatic cancer. Eur J Surg Oncol 2020; 46: 1524-1531 [PMID: 32098733 DOI: 10.1016/j.ejso.2020.02.013]
- Meyer W, Jurowich C, Reichel M, Steinhäuser B, Wünsch PH, Gebhardt C. Pathomorphological 323 and histological prognostic factors in curatively resected ductal adenocarcinoma of the pancreas. Surg Today 2000; 30: 582-587 [PMID: 10930222 DOI: 10.1007/s005950070096]
- Sohn TA, Yeo CJ, Cameron JL, Koniaris L, Kaushal S, Abrams RA, Sauter PK, Coleman J, Hruban 324 RH, Lillemoe KD. Resected adenocarcinoma of the pancreas-616 patients: results, outcomes, and prognostic indicators. J Gastrointest Surg 2000; 4: 567-579 [PMID: 11307091 DOI: 10.1016/s1091-255x(00)80105-5]
- 325 Chang DK, Johns AL, Merrett ND, Gill AJ, Colvin EK, Scarlett CJ, Nguyen NQ, Leong RW, Cosman PH, Kelly MI, Sutherland RL, Henshall SM, Kench JG, Biankin AV. Margin clearance and outcome in resected pancreatic cancer. J Clin Oncol 2009; 27: 2855-2862 [PMID: 19398572 DOI: 10.1200/JCO.2008.20.5104]
- Helm J, Centeno BA, Coppola D, Melis M, Lloyd M, Park JY, Chen DT, Malafa MP. Histologic 326 characteristics enhance predictive value of American Joint Committee on Cancer staging in resectable pancreas cancer. Cancer 2009; 115: 4080-4089 [PMID: 19626671 DOI: 10.1002/cncr.24503]
- Kinsella TJ, Seo Y, Willis J, Stellato TA, Siegel CT, Harpp D, Willson JK, Gibbons J, Sanabria JR, 327 Hardacre JM, Schulak JP. The impact of resection margin status and postoperative CA19-9 Levels on survival and patterns of recurrence after postoperative high-dose radiotherapy with 5-FU-based concurrent chemotherapy for resectable pancreatic cancer. Am J Clin Oncol 2008; 31: 446-453 [PMID: 18838880 DOI: 10.1097/COC.0b013e318168f6c4]
- Pelucchi C, Galeone C, Polesel J, Manzari M, Zucchetto A, Talamini R, Franceschi S, Negri E, La 328 Vecchia C. Smoking and body mass index and survival in pancreatic cancer patients. Pancreas 2014; 43: 47-52 [PMID: 24177141 DOI: 10.1097/MPA.0b013e3182a7c74b]
- 329 Yuan C, Morales-Oyarvide V, Babic A, Clish CB, Kraft P, Bao Y, Qian ZR, Rubinson DA, Ng K, Giovannucci EL, Ogino S, Stampfer MJ, Gaziano JM, Sesso HD, Cochrane BB, Manson JE, Fuchs CS, Wolpin BM. Cigarette Smoking and Pancreatic Cancer Survival. J Clin Oncol 2017; 35: 1822-1828 [PMID: 28358654 DOI: 10.1200/JCO.2016.71.2026]
- 330 Jiang P, Zhang M, Gui L, Zhang K. Expression patterns and prognostic values of the cyclindependent kinase 1 and cyclin A2 gene cluster in pancreatic adenocarcinoma. J Int Med Res 2020; 48: 300060520930113 [PMID: 33290118 DOI: 10.1177/0300060520930113]



- 331 Bailey P, Chang DK, Nones K, Johns AL, Patch AM, Gingras MC, Miller DK, Christ AN, Bruxner TJ, Quinn MC, Nourse C, Murtaugh LC, Harliwong I, Idrisoglu S, Manning S, Nourbakhsh E, Wani S, Fink L, Holmes O, Chin V, Anderson MJ, Kazakoff S, Leonard C, Newell F, Waddell N, Wood S, Xu Q, Wilson PJ, Cloonan N, Kassahn KS, Taylor D, Quek K, Robertson A, Pantano L, Mincarelli L, Sanchez LN, Evers L, Wu J, Pinese M, Cowley MJ, Jones MD, Colvin EK, Nagrial AM, Humphrey ES, Chantrill LA, Mawson A, Humphris J, Chou A, Pajic M, Scarlett CJ, Pinho AV, Giry-Laterriere M, Rooman I, Samra JS, Kench JG, Lovell JA, Merrett ND, Toon CW, Epari K, Nguyen NQ, Barbour A, Zeps N, Moran-Jones K, Jamieson NB, Graham JS, Duthie F, Oien K, Hair J, Grützmann R, Maitra A, Iacobuzio-Donahue CA, Wolfgang CL, Morgan RA, Lawlor RT, Corbo V, Bassi C, Rusev B, Capelli P, Salvia R, Tortora G, Mukhopadhyay D, Petersen GM; Australian Pancreatic Cancer Genome Initiative, Munzy DM, Fisher WE, Karim SA, Eshleman JR, Hruban RH, Pilarsky C, Morton JP, Sansom OJ, Scarpa A, Musgrove EA, Bailey UM, Hofmann O, Sutherland RL, Wheeler DA, Gill AJ, Gibbs RA, Pearson JV, Waddell N, Biankin AV, Grimmond SM. Genomic analyses identify molecular subtypes of pancreatic cancer. Nature 2016; 531: 47-52 [PMID: 26909576 DOI: 10.1038/nature16965]
- 332 Li X, Li Z, Zhu H, Yu X. Autophagy Regulatory Genes MET and RIPK2 Play a Prognostic Role in Pancreatic Ductal Adenocarcinoma: A Bioinformatic Analysis Based on GEO and TCGA. Biomed Res Int 2020; 2020: 8537381 [PMID: 33204717 DOI: 10.1155/2020/8537381]
- Liu JQ, Liao XW, Wang XK, Yang CK, Zhou X, Liu ZQ, Han QF, Fu TH, Zhu GZ, Han CY, Su H, 333 Huang JL, Ruan GT, Yan L, Ye XP, Peng T. Prognostic value of Glypican family genes in earlystage pancreatic ductal adenocarcinoma after pancreaticoduodenectomy and possible mechanisms. BMC Gastroenterol 2020; 20: 415 [PMID: 33302876 DOI: 10.1186/s12876-020-01560-0]
- 334 Qian B, Wei L, Yang Z, He Q, Chen H, Wang A, Yang D, Li Q, Li J, Zheng S, Fu W. Hic-5 in pancreatic stellate cells affects proliferation, apoptosis, migration, invasion of pancreatic cancer cells and postoperative survival time of pancreatic cancer. Biomed Pharmacother 2020; 121: 109355 [PMID: 31683179 DOI: 10.1016/j.biopha.2019.109355]
- 335 Song C, Chen T, He L, Ma N, Li JA, Rong YF, Fang Y, Liu M, Xie D, Lou W. PRMT1 promotes pancreatic cancer growth and predicts poor prognosis. Cell Oncol (Dordr) 2020; 43: 51-62 [PMID: 31520395 DOI: 10.1007/s13402-019-00435-1]
- 336 Kurahara H, Maemura K, Mataki Y, Tanoue K, Iino S, Kawasaki Y, Idichi T, Arigami T, Mori S, Shinden Y, Higashi M, Ueno S, Shinchi H, Natsugoe S. Lung recurrence and its therapeutic strategy in patients with pancreatic cancer. Pancreatology 2020; 20: 89-94 [PMID: 31787525 DOI: 10.1016/j.pan.2019.11.015
- Liu M, Zhang Y, Yang J, Cui X, Zhou Z, Zhan H, Ding K, Tian X, Yang Z, Fung KA, Edil BH, 337 Postier RG, Bronze MS, Fernandez-Zapico ME, Stemmler MP, Brabletz T, Li YP, Houchen CW, Li M. ZIP4 Increases Expression of Transcription Factor ZEB1 to Promote Integrin a3β1 Signaling and Inhibit Expression of the Gemcitabine Transporter ENT1 in Pancreatic Cancer Cells. Gastroenterology 2020; 158: 679-692.e1 [PMID: 31711924 DOI: 10.1053/j.gastro.2019.10.038]
- Ou ZL, Luo Z, Lu YB. Long non-coding RNA HULC as a diagnostic and prognostic marker of 338 pancreatic cancer. World J Gastroenterol 2019; 25: 6728-6742 [PMID: 31857775 DOI: 10.3748/wjg.v25.i46.6728]
- Fu Z, Jiao Y, Li Y, Ji B, Jia B, Liu B. TYMS presents a novel biomarker for diagnosis and 339 prognosis in patients with pancreatic cancer. Medicine (Baltimore) 2019; 98: e18487 [PMID: 31861032 DOI: 10.1097/MD.00000000018487]
- 340 Bu F, Zhu X, Yi X, Luo C, Lin K, Zhu J, Hu C, Liu Z, Zhao J, Huang C, Zhang W, Huang J. Expression Profile of GINS Complex Predicts the Prognosis of Pancreatic Cancer Patients. Onco Targets Ther 2020; 13: 11433-11444 [PMID: 33192076 DOI: 10.2147/OTT.S275649]



WJG https://www.wjgnet.com

WJG

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5890-5907

DOI: 10.3748/wjg.v27.i35.5890

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

REVIEW

# Gastrinoma and Zollinger Ellison syndrome: A roadmap for the management between new and old therapies

Roberta Elisa Rossi, Alessandra Elvevi, Davide Citterio, Jorgelina Coppa, Pietro Invernizzi, Vincenzo Mazzaferro, Sara Massironi

ORCID number: Roberta Elisa Rossi 0000-0003-4208-4372: Alessandra Elvevi 0000-0001-9841-2051: Davide Citterio 0000-0002-0708-8733; Jorgelina Coppa 0000-0003-2466-0524; Pietro Invernizzi 0000-0003-3262-1998; Vincenzo Mazzaferro 0000-0002-4013-8085; Sara Massironi 0000-0003-3214-8192.

Author contributions: Rossi RE designed the research; Rossi RE, Elvevi A, Citterio D, and Massironi S performed the literature search and wrote the first draft of the paper; Rossi RE, Massironi S, Coppa J, Invernizzi P, and Mazzaferro V revised the manuscript for important intellectual content; Rossi RE and Massironi S wrote the final version of the paper; All of the authors approved the final version for publication.

Conflict-of-interest statement: The authors declare no conflict of interest for this article.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build

Roberta Elisa Rossi, Davide Citterio, Jorgelina Coppa, Vincenzo Mazzaferro, HPB Surgery, Hepatology and Liver Transplantation, ENETS Center of Excellence, Fondazione IRCCS Istituto Nazionale Tumori (INT, National Cancer Institute), Milan 20133, Italy

Roberta Elisa Rossi, Department of Pathophysiology and Transplantation, University of Milan, Milan 20122, Italy

Alessandra Elvevi, Pietro Invernizzi, Sara Massironi, Division of Gastroenterology and Center for Autoimmune Liver Diseases, Department of Medicine and Surgery, University of Milano-Bicocca, Monza 20900, Italy

Alessandra Elvevi, Pietro Invernizzi, Sara Massironi, European Reference Network on Hepatological Diseases (ERN RARE-LIVER), San Gerardo Hospital, Monza 20033, Italy

Vincenzo Mazzaferro, Department of Oncology and Hemato-Oncology, University of Milan, Milan 20122, Italy

Corresponding author: Roberta Elisa Rossi, MD, PhD, HPB Surgery, Hepatology and Liver Transplantation, ENETS Center of Excellence, Fondazione IRCCS Istituto Nazionale Tumori (INT, National Cancer Institute), via Giacomo Venezian, Milan 20133, Italy. robertaelisa.rossi@gmail.com

# Abstract

Zollinger-Ellison syndrome (ZES) associated with pancreatic or duodenal gastrinoma is characterized by gastric acid hypersecretion, which typically leads to gastroesophageal reflux disease, recurrent peptic ulcers, and chronic diarrhea. As symptoms of ZES are nonspecific and overlap with other gastrointestinal disorders, the diagnosis is often delayed with an average time between the onset of symptoms and final diagnosis longer than 5 years. The critical step for the diagnosis of ZES is represented by the initial clinical suspicion. Hypergastrinemia is the hallmark of ZES; however, hypergastrinemia might recognize several causes, which should be ruled out in order to make a final diagnosis. Gastrin levels > 1000 pg/mL and a gastric pH below 2 are considered to be diagnostic for gastrinoma; some specific tests, including esophageal pH-recording and secretin test, might be useful in selected cases, although they are not widely available. Endoscopic ultrasound is very useful for the diagnosis and the local staging of the primary tumor in patients with ZES, particularly in the setting of multiple endocrine neoplasia type 1. Some controversies about the management of these



WJG | https://www.wjgnet.com

upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Gastroenterology and hepatology

### Country/Territory of origin: Italy

# Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Received: March 15, 2021 Peer-review started: March 15, 2021 First decision: April 17, 2021 Revised: April 29, 2021 Accepted: August 10, 2021 Article in press: August 10, 2021 Published online: September 21, 2021

P-Reviewer: Prisciandaro M S-Editor: Gao CC L-Editor: Filipodia P-Editor: Xing YX



tumors also exist. For the localized stage, the combination of proton pump inhibitory therapy, which usually resolves symptoms, and surgery, whenever feasible, with curative intent represents the hallmark of gastrinoma treatment. The high expression of somatostatin receptors in gastrinomas makes them highly responsive to somatostatin analogs, supporting their use as anti-proliferative agents in patients not amenable to surgical cure. Other medical options for advanced disease are super-imposable to other neuroendocrine neoplasms, and studies specifically focused on gastrinomas only are scant and often limited to case reports or small retrospective series. The multidisciplinary approach remains the cornerstone for the proper management of this composite disease. Herein, we reviewed available literature about gastrinoma-associated ZES with a specific focus on differential diagnosis, providing potential diagnostic and therapeutic algorithms.

Key Words: Gastrinoma; Zollinger-Ellison syndrome; Neuroendocrine neoplasms; Pancreatic neuroendocrine neoplasm; Duodenal neuroendocrine neoplasm; Diagnosis; Therapy

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: As symptoms of Zollinger-Ellison syndrome are nonspecific and overlap with other gastrointestinal disorders, most of these patients are usually referred to general gastroenterologists, leading to a diagnostic delay. A better disease awareness together with the maintenance of a high index of suspicion are necessary to make the final diagnosis. The proper management of Zollinger-Ellison syndrome due to a gastrinoma include both the medical treatment for symptom's relief and surgery whenever feasible with curative intent; the multidisciplinary approach, with close cooperation between gastroenterologists and surgeons, and the referral to tertiary centers with great expertise in the neuroendocrine field are mandatory.

Citation: Rossi RE, Elvevi A, Citterio D, Coppa J, Invernizzi P, Mazzaferro V, Massironi S. Gastrinoma and Zollinger Ellison syndrome: A roadmap for the management between new and old therapies. World J Gastroenterol 2021; 27(35): 5890-5907 URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5890.htm

DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5890

# INTRODUCTION

Zollinger-Ellison syndrome (ZES) was firstly described in 1955 as associated with a neuroendocrine neoplasm (NEN) capable of ectopic gastrin secretion (namely gastrinoma)[1], resulting in gastric acid hypersecretion, which typically leads to gastroesophageal reflux disease (GERD), recurrent peptic ulcers, and chronic diarrhea. The terms gastrinoma and ZES have been frequently used as synonymous, although gastrinoma refers to the NEN secreting gastrin, whereas ZES refers to the clinical manifestations of the disease. ZES has an incidence of 1-1.5 cases/million per year[2]. Gastrinomas are NENs located in the duodenum (70%), pancreas (25%), and rarely (5%), in other sites, including stomach, liver, ovary, and lung. Gastrinoma is the most frequent functioning duodenal NEN and the second most frequently occurring functional pancreatic NEN (pNEN), following insulinoma; in turn, 15% of functioning pNENs is represented by gastrinoma. It may be sporadic, which is generally diagnosed between the ages of 50 and 70 years with a male to female ratio of 1.5-2:1 [3], whilst 20%-30% of the patients develop ZES in the context of a genetic syndrome known as multiple endocrine neoplasia type 1 (MEN-1)[4].

The diagnosis of ZES is not always straightforward due to both non-specific symptoms and confounding factors including proton pump inhibitor (PPI) therapy, which might temporarily relieve symptoms. Furthermore, as these patients tend to be referred to gastroenterologists because of diarrhea and/or reflux disease disorder, despite a better awareness of the disease, the diagnosis might be challenging for those



gastroenterologists with low experience in the neuroendocrine setting as well as for many oncologists who are less used to dealing with diarrhea and reflux disease. As a consequence, the average time between the onset of symptoms and the final diagnosis is often longer than 5 years [5,6], and nearly 25% of patients are metastatic at the first diagnosis and show a worse prognosis when compared to non-metastatic patients in whom the surgical management is associated with a promising 15-year survival rate of > 80%[7].

Furthermore, some controversies about the management of these tumors still exist, particularly regarding the exact role of surgery or medical treatment and the possible role of somatostatin analogs (SSAs)[3]. Given that gastrinoma and ZES need both a proper medical treatment for symptom relief and a surgical procedure whenever feasible, the multidisciplinary approach, with close cooperation between clinicians and surgeons, remains the cornerstone for proper management of this composite disease, which should be always referred to tertiary centers.

Herein, we review from a critical point of view current knowledge about gastrinoma-associated ZES, also providing potential diagnostic and therapeutic algorithms based on both evidence from literature and own personal experience.

# METHODOLOGY

Bibliographical searches were performed in PubMed using the following keywords: Gastrinoma; Zollinger Ellison syndrome; neuroendocrine neoplasms; pancreatic neuroendocrine neoplasm; duodenal neuroendocrine neoplasm; diagnosis; therapy; guidelines. We searched for all relevant articles published over the last 10 years. The reference lists from the studies returned by the electronic search were manually searched to identify further relevant reports. The reference lists from all available review articles, primary studies, and proceedings of major meetings were also considered. Articles published as abstracts were included, whereas non-English language papers were excluded.

# **CLINICAL PRESENTATION**

ZES is characterized by gastric acid hypersecretion and consequent hyperchlorhydria resulting in severe acid-related peptic disease and diarrhea. The symptoms usually resolve when gastric acid secretion is controlled pharmacologically with PPIs[8,9]; of note, the disappearance of diarrhea following PPI treatment is typical of ZES and represents one of the factors contributing to the diagnostic delay. According to data from the literature, common symptoms include abdominal pain (75%), diarrhea (73%), heartburn (44%), and weight loss (17%) [6,8,10]. As these symptoms are both not specific and often less severe due to concomitant PPI treatment, the final diagnosis is often delayed and patients are diagnosed with irritable bowel syndrome or reflux disease by gastroenterologists with low or no knowledge of the disease[8,11].

The endoscopic features are also not specific and might include erosions and ulcers [12], however, ZES patients often present with multiple ulcers located at unusual sites, e.g., beyond the first or second portion of the duodenum[8,13]. Furthermore, enlarged gastric folds can be present in more than 90% of patients with ZES[11].

One should keep in mind that approximately 25% of gastrinomas occur in the context of MEN-1, which is characterized by the presence of parathyroid, pancreaticduodenal, and pituitary tumors[14]; thus the occurrence of unexplained hypercalcemia might be a sign for possible MEN-1 syndrome-associated ZES[15,16], also taken into account that primary hyperparathyroidism is generally the presenting feature in the majority of cases of MEN-1 syndrome[8,16,17]. Of note, parathyroidectomy usually improves gastrin levels and basal acid output[16]. Finally, in ZES/MEN-1 patients, type 2 gastric NENs might occur[3].

# **DIFFERENTIAL DIAGNOSIS**

Symptoms of ZES are nonspecific and overlap with other gastrointestinal (GI) disorders, which explains the frequent diagnostic delay.

As concerns chronic diarrhea in ZES, this is sustained by hyperchlorhydria and sodium and water malabsorption due to hypergastrinemia[18]. As afore-mentioned,



diarrhea is one of the most frequent symptoms in ZES; up to 75% of patients manifest diarrhea<sup>[19]</sup>, and this could be the sole presenting symptom in 3%-10% of the patients [20]. Moreover, chronic diarrhea is one of the most frequent symptoms requiring gastroenterologist referral; its diagnostic workup could be challenging because many different causes could cooperate to diarrhea development and recurrence (e.g., dietary habits, drugs). Recent British Society of Gastroenterology (BSG) guidelines for chronic diarrhea<sup>[21]</sup> tried to classify different causes of chronic diarrhea (Table 1) and to standardize a diagnostic work-up in these patients. Since hormone-secreting tumors are considered rare causes of chronic diarrhea, BSG guidelines suggest testing patients for these tumors only when other causes of diarrhea have been excluded. From a clinical point of view, the association between chronic diarrhea with both other ZES suggestive symptoms (e.g., chronic peptic ulcer disease) and clinical response to PPIs may be helpful in diagnosing this challenging syndrome, taking into account that the delay in diagnosis of ZES remains between 6 to 9 years from the first clinical presentation[9,19].

Abdominal pain and heartburn are frequently reported as symptoms of ZES[9]. As well as diarrhea, they are sustained by hyperchlorhydria, which directly damages GI mucosa, causing ulcers and erosions. Abdominal pain could be associated with peptic ulcers, which, differently from Helicobacter pylori or non-steroidal anti-inflammatory drug-related ulcers, are multiple, located at unusual locations (e.g., the third part of the duodenum, small bowel) and complicated by bleeding, penetration, perforation, or strictures[8,13,19].

Similar to peptic ulcer disease, chronic GERD is one of the most frequent manifestations of ZES[13]. Heartburn and regurgitation are the most typical symptoms, which are super-imposable to symptoms associated with typical GERD; differently from the typical syndrome, patients with ZES often present esophageal strictures due to overexposition to acid reflux.

Again, the association between these symptoms and chronic diarrhea, after exclusion of other common GI etiologies, might raise the suspicion of ZES, which requires specific tests in order to get the final diagnosis.

# DIAGNOSIS

The diagnosis of ZES is quite challenging, also considered that the critical point is the initial suspicion of ZES. A suggested diagnostic algorithm is represented in Figure 1.

ZES is a clinical syndrome characterized by the following triad: (1) gastric acid hypersecretion, sustained by (2) fasting serum hypergastrinemia causing (3) peptic ulcer disease and diarrhea[1]. Hypergastrinemia is sustained by a gastrinoma, a rare NEN (located primarily in the duodenum or pancreas) that secretes gastrin.

Since ZES symptoms can be explained almost entirely by acid hypersecretion, PPIs, which significantly decrease acid secretion, can mitigate or resolve ZES symptoms, making ZES diagnosis even more challenging than in the past[9,22], but avoiding severe ZES complications.

Hypergastrinemia is the hallmark of ZES; however, hypergastrinemia might recognize several causes, which should be ruled out in order to make a final diagnosis of ZES[23]. In detail, it can be distinguished between (1) appropriate hypergastrinemia, due to atrophic gastritis (with or without pernicious anemia), anti-secretory therapy (PPIs or high-dose histamine H2-receptor antagonist, namely famotidine), chronic renal failure, Helicobacter pylori-related pan-gastritis, vagotomy, and (2) inappropriate hypergastrinemia that can be observed in ZES (sporadic or associated with MEN-1), antral-predominant Helicobacter pylori infection, retained-antrum syndrome, gastric-outlet obstruction, extensive small-bowel resection.

The diagnosis of ZES requires the demonstration of inappropriate gastrin secretion associated with gastric hyperchlorhydria, which corresponds to a gastric pH < 2[5]. Normal fasting gastrin levels are < 100 pg/mL; levels > 300 pg/mL are highly suspicious, and levels > 1000 pg/mL together with a gastric pH below 2 are considered to be diagnostic for gastrinoma[2,9,24]. Naso-gastric tube aspiration has classically been used to estimate gastric pH, but it can be uncomfortable for patients and can underestimate gastric acid output; alternatively, gastric pH can be measured during upper GI endoscopy, by aspiration of gastric juice for pH determination using either pH paper or a pH meter; while endoscopic sampling was shown to overestimate total acid volume, it provided more reproducible results and offered greater patient tolerance than nasogastric tube placement[5,23,25,26]. To avoid false-negative results, fasting serum gastrin levels and gastric pH should be measured after PPI withdrawal



Table 1 Differential diagnosis of chronic diarrhea[21]			
Common	Infrequent	Rare	
IBS-diarrhea	Small bowel bacterial overgrowth	Small bowel enteropathies ( <i>i.e.</i> Whipple's disease, tropical sprue, amyloid, <i>etc.</i> )	
Bile acid diarrhea	Mesenteric ischemia	Hypoparathyroidism	
Diet (artificial sweeteners, caffeine, FODMAP malabsorption, <i>etc.</i> )	Lymphoma	Addison's disease	
Colonic neoplasia	Surgical causes (small bowel resection, incontinence, <i>etc.</i> )	Hormone secreting tumors ( <i>i.e.</i> VIPoma, gastrinoma, carcinoid)	
IBD	Chronic pancreatitis	Autonomic neuropathy	
Celiac disease	Radiation enteropathy	Factitious diarrhea	
Drugs (antibiotics, NSAID, etc.)	Pancreatic carcinoma	Brainerd diarrhea	
Overflow diarrhea	Hyperthyroidism		
	Diabetes		
	Chronic infections (i.e. giardiasis)		
	Cystic fibrosis		

FODMAP: Fermentable Oligosaccharides, Disaccharides, Monosaccharides, and Polyols; IBS: Irritable bowel syndrome; NSAID: Nonsteroidal antiinflammatory drugs.

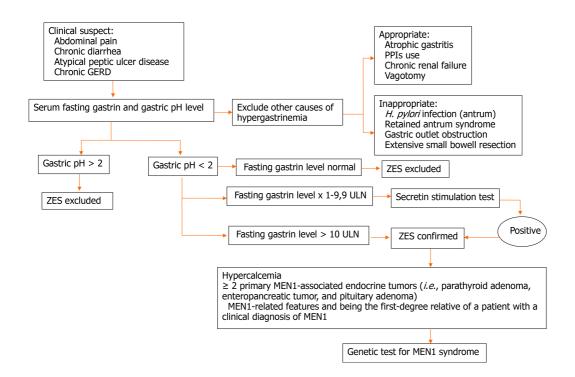


Figure 1 A suggested diagnostic algorithm is depicted. GERD: Gastroesophageal reflux disease; *H. pylori: Helicobacter pylori;* MEN-1: Multiple endocrine neoplasia type 1; PPIs: Proton pump inhibitors; ULN: Upper limit of normal; ZES: Zollinger Ellison syndrome.

[2,6,27]. However, PPI withdrawal could be dangerous for ZES patients, because it could bring a dramatic increase in gastric acid secretion, hence causing severe peptic ulcer disease and its complications[23], thus the decision to stop the treatment should be tailored to every single patient. Then, it is usually suggested to start histamine H2-receptor antagonists (*i.e.* famotidine) as soon as PPIs are stopped in order to prevent complications due to gastric acid hypersecretion. Having a shorter duration of action compared to PPIs, H2-antagonists could be used until the evening before serum gastrin and gastric pH tests[23].

WJG | https://www.wjgnet.com

#### Imaging and ultrasound endoscopy (endoscopic ultrasound)

Localization of the primary tumor and its metastases is the first diagnostic step when ZES associated with gastrinoma is suspected.

Contrast-enhanced computed tomography (CT) scan is useful to identify primary tumor > 1 cm, pancreatic head tumors, and liver metastases, with a sensitivity between 59% and 78% and a specificity between 95% and 98%, respectively. Conversely, sensitivity decreases for tumor size < 1 cm and extra-pancreatic locations[28,29].

Contrast-enhanced magnetic resonance imaging (MRI) showed high specificity (namely 100%) in detecting small pancreatic tumors and liver metastases, whereas sensibility is sub-optimal varying from 25% to 85%. Of note, MRI showed a higher sensibility for liver metastases detection when compared to CT scan[28,30].

Somatostatin receptor scintigraphy (Octreoscan<sup>®</sup>) has been used to localize gastrinomas<sup>[8,31]</sup>. This test involves the administration of indium radio-labeled octreotide, which binds selectively to somatostatin receptors found on gastrinoma cells. It showed quite good sensitivity (between 77% and 78%) and a good specificity (93%-94%) for primary tumor detection and its metastases, although sensitivity decreases for small tumors (< 1 cm)[32]. Diagnostic accuracy of somatostatin receptor scintigraphy (Octreoscan®) can be improved by performing it in combination with single-photon emission CT (SRS-SPECT)[28]. Different studies showed higher sensitivity and specificity in primary tumor detection, 78%-88% and 97%, respectively, when compared to Octreoscan<sup>®</sup> alone[33-35].

In more recent years, somatostatin receptor positron emission tomography (PET) techniques have shown great promise for improving the localization of gastrinomas as well as other NENs[36-39] and for the detection of distant metastases, including bone lesions. The radioisotope 68Ga can be ligated to peptides that bind to somatostatin receptors found in abundance on the NEN surface[36]. This technique showed a higher sensibility and specificity (72%-100% and 83%-100%, respectively) when compared to the aforementioned diagnostic techniques in localizing the primary tumor, especially small size tumors[36,37,40]. Combining 68Ga-radiotracers with traditional CT scans (PET/CT) further enhances diagnostic accuracy compared to PET alone, showing a sensitivity of 93% and a specificity of 96% in primary tumor detection [41]. Gallium-68PET-scan should be always included in the diagnostic pathway of all NENs, including gastrinoma, in order to both identify the primary tumor and stage the disease.

Endoscopic ultrasound (EUS) has become an important diagnostic tool to localize gastrinomas, particularly small (i.e. < 2 cm) pancreatic lesions; its sensitivity and specificity are 75%-100% and 95%, respectively, for pancreatic tumors. Unfortunately, its sensitivity dramatically decreases in cases of duodenal localization, ranging from 38% to 63% [28,42]. A further advantage of this technique is the possibility of taking cytologic/histologic samples through a fine needle aspiration/biopsy (FNA/B) to confirm the diagnosis of NEN, even if false-negative results are possible mainly due to poor sampling adequacy. EUS-FNA/B is now considered the primary sampling technique for pancreatic tumors, with a sensitivity ranging between 80% and 90%, specificity at 96% [43], and a sampling adequacy rate of 83%-93% [44].

When used as a screening modality in asymptomatic patients with MEN-1, EUS has been reported to be more accurate than CT scan to detect smaller tumors[45]. Therefore, its diagnostic ability has led experts to recommend it as an annual screening modality for all patients with MEN-1, although recent evidence suggests that the growth rate of small pNENs (*i.e.* < 2 cm) is low and that EUS screening frequency can likely be extended [14,46].

#### Esophageal pH-recording

Since one of the most common symptoms of ZES is GERD, it could be argued that esophageal pH-monitoring could be a useful tool to diagnose ZES. Recent BSG guidelines for esophageal manometry and esophageal pH monitoring[47] stated indications to perform esophageal pH-monitoring, also including as an indication GERD symptoms that did not respond to double dose of PPIs. This technique allows to diagnose an increased acid exposure, to evaluate the association between symptoms and acid or non-acid reflux, and to identify different phenotypes of upper symptoms ( i.e. non-erosive reflux disease, hypersensitive esophagus, and functional heartburn).

ZES is not usually included in diagnosis performed by esophageal pH-monitoring, and, consequently, ZES reference standard for esophageal pH-monitoring is lacking. However, evidence of a high number of acidic reflux episodes (*i.e.* esophageal pH < 4), a high number of long (*i.e.* > 5 min) reflux episodes, a high percentage of time with esophageal pH < 4, both on a double dose of PPIs and off PPIs, could raise the



suspicion of abnormal gastric acid secretion. This hypothesis should be confirmed by prospective studies; however, considering the rarity of this syndrome, it would be very difficult to obtain standard values to use in clinical practice; therefore, despite its potential utility, this test is not currently included in the standard diagnostic workup of gastrinoma.

#### Secretin provocative test

Secretin provocative test in ZES diagnosis founds its application in controversial cases, that is patients with suspected ZES, gastric pH < 2 but fasting serum gastrin < × 10 upper limit of normal[9]. To perform a secretin stimulation test, fasting gastrin levels are obtained before intravenous (IV) administration of secretin and then 2, 5, and 10 min after infusion[25]. Patients with gastrinomas exhibit an inappropriate increase in gastrin production in response to secretin infusion[9]. This mechanism can be explained in part by the fact that secretin receptors are expressed directly on the gastrinoma cell surface[48]. Different cut-offs for positive tests have been proposed, including an absolute increase in gastrin concentration  $\geq 110 \text{ pg/mL}$  or  $\geq 200 \text{ pg/mL}$ or a 50% increase in gastrin concentration<sup>[49]</sup>. However, previous data suggested that a positive secretin-provocative test ( $\geq$  120 pg/mL increase) has a sensitivity of 94% and specificity of 100%, respectively<sup>[50]</sup>. According to data from the literature, a falsenegative response can occur in 6% to 20% of patients[51,52], whereas false-positive responses, ranging from 15% to 39% in different studies[52,53], are found in patients with pernicious anemia or chronic PPI use.

In order to reduce the risk of false-positive results, PPI treatment should be withdrawn, but, again, the decision should be discussed in a case-by-case manner to limit the risk of severe complications (e.g., perforation or bleeding). This might partially explain the reason why the secretin test can be difficult to be performed and should be reserved for strictly selected cases when the diagnosis is not straightforward.

# MEN-1

MEN-1 is an autosomal dominant disorder, whose incidence has been estimated from random postmortem studies to be 0.25%, and to be 1%-18% in patients with primary hyperparathyroidism, 16%-38% in patients with gastrinomas, and less than 3% in patients with pituitary tumors [14]. From a clinical point of view, MEN-1 syndrome includes the occurrence of parathyroid adenoma (90%), entero-pancreatic tumor (30%-70%), being gastrinoma the most frequent (40%), and pituitary adenoma (30%-40%). Other tumors that might occur in MEN-1 patients are adrenal cortical tumor (40%), pheochromocytoma (< 1%), bronchopulmonary NEN (2%), thymic NEN (2%), gastric NEN (10%), lipomas, (30%), angiofibromas (85%), collagenomas (70%), and meningiomas (8%)[14].

In patients with an established diagnosis of gastrinoma-related ZES, MEN-1 syndrome might be present in approximately 25% of the cases. The presence of hypercalcemia due to hyperparathyroidism is one of the first signs. However, the diagnosis might be challenging in this specific setting as ZES does not usually develop in the absence of primary hyperparathyroidism, and hypergastrinemia has also been reported to be associated with hypercalcemia as a confounding factor[15]. Furthermore, parathyroidectomy leads to restoration of normocalcemia and improvement in clinical symptoms and biochemical abnormalities in as many as 20% of MEN-1 patients with ZES[14]. Moreover, staging and localization with CT or MRI is even more challenging in the setting of MEN-1 due to the presence of numerous small tumors < 1 cm in size[27,28]. A high index of suspicion must be maintained if a patient with chronic diarrhea and unexplained peptic ulcer disease presents with primary hyperparathyroidism. The genetic test for MEN-1 syndrome should be performed in a selected subgroup of patients, namely (1) in patients with two or more primary MEN-1-associated endocrine tumors (e.g., parathyroid adenoma, entero-pancreatic tumor, and pituitary adenoma) or hypercalcemia associated with an endocrine tumor; and (2) patients showing MEN-1-related features and being the first-degree relative of a patient with a clinical diagnosis of MEN-1[14].

# THERAPY

The management of gastrinoma and ZES includes both a proper medical treatment for symptom's relief and surgery with curative intent whenever feasible. A proposed therapeutic algorithm is represented in Figure 2.



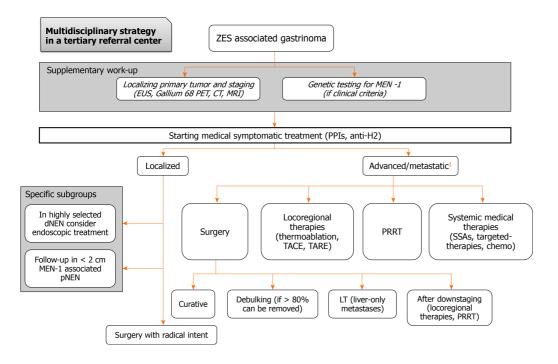


Figure 2 A proposed therapeutic algorithm is represented based on both evidence from literature and personal own experience. <sup>1</sup>Allocation driving prognostic factors are performance status, age, metastatic disease burden and pattern, comorbidities. CT: Computed tomography; dNEN: Duodenal neuroendocrine neoplasm; EUS: Endoscopic ultrasound; H2: Histamine receptor 2: LT: Orthotopic liver transplantation; MEN-1: Multiple endocrine neoplasia type 1; MRI: Magnetic resonance imaging; PET: Positron emission tomography; pNEN: Pancreatic NEN; PPIs: Proton pump inhibitors; PRRT: Peptide-radioreceptor therapy; SSAs: Somatostatin analogs; TACE: Transarterial chemoembolization; TARE: Transarterial radioembolization; ZES: Zollinger Ellison syndrome.

#### Surgery

The role of surgery in the treatment of gastrinoma has changed completely from the introduction of PPIs in the 1980s. In fact, before the advent of an effective antisecretory therapy, surgery was performed to control acid hypersecretion, mainly removing the target cells of gastrin through total gastrectomy. These operations were, by the way, affected by a high mortality rate due to acid-related complications in the postoperative course. With the use of PPIs, gastric hypersecretion was no longer a problem, and the main determinant of prognosis became the gastrinoma itself because of its malignant potential and surgical excision started to be proposed as a potentially curative therapy. From 1981, the National Institute of Health began a prospective study recruiting patients with ZES for surgical therapy, with a well-designed surgical protocol in order to capture the long-term results of the best available surgical approach. The study reported a 10-year overall survival (OS) and disease-free survival (DFS) of 94% and 34%, respectively[54]. Therefore, surgery has gradually changed its role and gastrinoma resection has started to be increasingly proposed to patients eligible for resection. Currently, across the most important guidelines, surgical excision is generally recommended either for sporadic gastrinoma or for MEN-1 associated gastrinoma if complete tumor removal is possible[2,55-58]. Subsequent studies reported a 20 year OS of 58%-71%, a 20-year disease-related survival of 73%-88% [59], and a 10-year DFS of 25%-50% [60]. Surgery of the primary tumor also demonstrated to reduce the occurrence of liver metastases[61-63], which are one of the main determinants of prognosis, and to improve DFS in comparison with non-surgical management[62].

The majority of gastrinomas (from 60% to 90% depending on the series)[42,60] occur in the duodenum, and, since these are often very small lesions (less than 1 cm) and located at the submucosal layer, tumor detection is not so straightforward. Therefore, the surgical technique should follow a stepwise approach to search for the tumor even in case of negative preoperative imaging. In this context, surgery has firstly a diagnostic purpose, which is quite uncommon in modern surgery and, given the peculiarity of this technique and the rarity of the disease, it should be performed by experienced surgeons in tertiary referral centers. After a complete abdominal exploration, the duodenum and the pancreatic head are mobilized (Kocher maneuver) and carefully palpated. Intra-operative ultrasound with a linear probe is then performed on the duodenum and pancreas looking for the primary tumor and on the liver in search for liver metastases. Intra-operative endoscopy is performed thereafter



advancing the scope into the duodenum; duodenal gastrinomas may be found through trans-illumination of the bowel wall as non-trans-illuminated spots. If a lesion is identified, it should be marked with a suture and the duodenum opened around it for a full-thickness excision. If the described steps fail to reveal any lesion, a 3 cm longitudinal incision is made on the anterior aspect of the second portion of the duodenum, and the entire duodenal wall is palpated. Suspicious lesions are excised with a fullthickness rim of normal tissue and sent for pathology. The duodenum is then closed transversally, if possible, to minimize the risk of strictures [64,65]. In the hands of an experienced surgeon, lesions could be found in 98% of imaging-negative ZES patients, with a 50% curative rate [59], similar to that of imaging-positive patients. These findings suggest that surgery should be performed as soon as possible in sporadic ZES, despite negative imaging findings. Pancreatic gastrinomas should be enucleated if located 3 mm or farther from the main pancreatic duct. Conversely, lesions that are closer to the pancreatic duct require distal pancreatectomy with or without splenectomy if located in the body or tail of the gland and pancreaticoduodenectomy if located in the head/neck. Pancreaticoduodenectomy or distal pancreatectomy may be necessary also for local recurrence after enucleation[66].

Regional lymph nodes should always be removed because nodal metastases are present in almost half of the patients[54,67] and lymphadenectomy has been associated with increased DFS[68], as reported also for other pNENs[69-71]. The presence of primary gastrinoma located in a lymph node is controversial, however, several studies reported long disease-free survivors after resection of only a positive lymph node[72, 73], and this supports the role of routine lymphadenectomy.

Since pancreaticoduodenectomy provides complete removal of the regional lymph nodes of the pancreatic head, the results in terms of DFS are better with respect to enucleation because of the higher chance of radicality[54,67]. However, given the high postoperative morbidity and the good prognosis also of patients with small residual disease, pancreaticoduodenectomy is not recommended as the standard operation for these patients[2,55-58]. Generally, the indication for surgery should always follow a thorough risk/benefit assessment within a multidisciplinary tumor board aiming at maximum radicality and minimum morbidity. This is particularly the case for MEN-1 patients; in these patients, who have generally an earlier age of onset, pNENs should be resected in low-risk patients, and surgery is generally recommended for tumors larger than 2 cm[14,58]. However, according to most authorities, as well as all guidelines, surgical resection for an attempted cure should be performed in ZES patients whenever possible[2,27,58]. This is particularly true for functioning duodenal NENs, including gastrinomas, which have been reported to express a high metastatic potential<sup>[74]</sup>, thus a radical surgical approach should be the first choice in this specific setting. However, in highly selected cases (*i.e.* duodenal lesions  $\leq 1$  cm, limited to the submucosal layer and without lymph nodal involvement), endoscopic resection might also be considered, although the risk of undetected micro-metastases might represent an issue.

Another controversial issue is laparoscopic surgery; while it is widely adopted for pNENs, its role for gastrinomas is limited to patients in whom preoperative imaging gives an accurate definition of tumor location. Unfortunately, as already mentioned, extensive exploration is often needed for diagnostic purposes. In these cases, laparoscopy is inadequate, and laparotomy is mandatory.

The role of surgical resection in ZES patients with advanced metastatic disease or even with extensive invasive localized disease is not well-defined. In this setting, the possibility of surgical removal of all resectable tumors (cytoreductive surgery, debulking surgery) should be considered, and surgery is generally recommended if  $\geq$ 80% of all disease can be removed (generally feasible in 5%-15% of all metastatic gastrinomas), although only a few reports containing primarily gastrinomas treated with this approach are currently available[10,72].

Finally, in highly selected metastatic gastrinomas, with liver-only metastases and fulfilling strict inclusion criteria, liver transplantation might be considered, even if its use remains controversial and the risk of tumor recurrence represents an issue<sup>[58]</sup>.

#### Liver-directed therapies

Studies specifically focused on liver-directed therapies in the context of gastrinomas are scant; however, as for other NENs, the embolization approaches in the setting of gastrinoma are generally reserved for patients with metastatic unresectable hepatic metastases either limited to the liver or with a liver-predominant disease, particularly if locally symptomatic[10,58]. Of note, liver-directed therapies are used less frequently in ZES than in other metastatic NENs, because in ZES, the hormone excess-state can be well-controlled medically.



#### Medical treatment

Among functioning NENs, gastrinoma is the most frequent type. There are two therapeutic goals in the management of patients with gastrinoma: The control of gastric acid hypersecretion and the treatment of the tumor itself.

#### Antisecretory medications

The therapy for syndrome control is based on PPI (e.g., omeprazole, esomeprazole, lansoprazole, pantoprazole, etc.), which are highly effective drugs and considered the drugs of choice for suppressing acid secretion. PPIs effectively block gastric acid secretion by irreversibly binding to and inhibiting the hydrogen-potassium ATPase pump on the luminal surface of the parietal cell membrane. Theoretically, the choice and titration of anti-secretory therapy should be guided by the parameters of gastric acid secretions such as basal acid output (to reduce it below 10 mEq/h)[75], since using symptoms alone as a signal of efficacy might be misleading, even if in many centers these methods are not available. Therefore, in most cases, PPI therapy is started at an empirical maximized dosage. The recommended initial dose of omeprazole is 60 mg/daily or esomeprazole 120 mg/daily, lansoprazole 45 mg/daily, rabeprazole 60 mg/daily, pantoprazole 120 mg/daily, divided, twice-a-day[75-78]. The type of PPI used seems not to be of relevance and a systematic review of 12 randomized trials examining the relative effectiveness of different PPI doses and dosing regimens found no consistent differences in symptom resolution and esophagitis healing rates [79]. IV PPIs are indicated in patients with clinically significant upper GI bleeding from a suspected peptic ulcer. Omeprazole, pantoprazole, and esomeprazole are the only PPIs available as an IV formulation. The other patients can be treated with oral preparation.

As concerns efficacy, PPIs have significantly decreased the morbidity and mortality resulting from severe ulcer disease<sup>[80]</sup>. In 60% of patients, ulcer healing occurs within 2 wk; in 90%-100% of patients, healing occurs within 4 wk. PPIs are generally safe, even when used in high doses.

Once an effective clinical control of the peptic disease has been achieved, a gradual dose reduction is generally suggested[81,82]. In a study by Metz et al[83], 37 patients received high-dose omeprazole for almost 2 years, and nearly 50% were able to lower the dose down to 20 mg once daily, with 95% of patients experiencing safe long-term reductions in their medication dose. PPIs are generally well tolerated and can control hypergastrinemia in ZES for > 10 years (although some patients experience low vitamin B12 levels)[84].

No tachyphylaxis has been described. Therefore, the long duration of action, the fewer adverse effects, and the high potency make them superior to H2 blockers.

Regarding the use of H2-receptor antagonists in ZES, the dose usually is 4-8 times higher than the dose administered to patients with peptic ulcer disease. Although a good success rate exists, this treatment has been reported to fail in 50% of patients. Therefore, these drugs are never the first choice.

Only when PPIs are unable to control gastric acid secretion, SSAs can be considered, as they reduce gastrin secretion, even if they do not represent a first-line treatment at least for symptom control.

Even if this is not a treatment currently approved in localized gastrinoma, it is worth mentioning that in animals, the cholecystokinin-2 receptor antagonist YF476 has been shown to inhibit the development of enterochromaffin-like cell-tumors in susceptible animals with induced hypergastrinemia. Therefore, this drug could represent a potential option in ZES, not only to inhibit hypergastrinemia but also to prevent gastric NEN type 2 (e.g., associated with ZES/MEN-1). Furthermore, there continues to be interest in the development of cholecystokinin-2 receptor antagonists as anti-secretory agents [85]. However, strong evidence supporting the role of these molecules in this specific setting is lacking.

#### Anti-proliferative treatment

Approximately one-third of ZES patients present with metastatic disease to the liver [10,86]. There are several systemic therapeutic options for advanced gastrinoma, not substantially different from the ones for other NENs, however, studies evaluating specific response rates in gastrinomas alone are limited.

SSAs like octreotide and lanreotide are highly effective in controlling the symptoms associated with hormone hypersecretion in all functioning tumors[87,88]; furthermore, they can reduce gastrin levels and their anti-proliferative effect has been demonstrated in PROMID and CLARINET studies [89,90]. However, in these studies only a few cases of gastrinoma were included, and, even if different case reports and case series suggested the role of SSAs in controlling gastrin secretion and symptoms in ZES

WJG | https://www.wjgnet.com

patients[91-94], to date only a few studies with a very low number of patients investigated specifically the role of SSAs in ZES[3].

The multitargeted tyrosine kinase inhibitor, sunitinib, has demonstrated an improved progression-free survival from 5.5 mo to 11.4 mo in metastatic pNENs[95]. Moreover, based on the results of two randomized, double-blind, prospective, placebo-controlled studies, the mammalian target of rapamycin-inhibitor everolimus has been approved in advanced both pancreatic[96] and extra-pNENs[96]. However, there are no specific studies on the effects of sunitinib/everolimus in the specific setting of gastrinomas.

Streptozocin, 5-fluorouracil, and doxorubicin have been used, with the response rate reported to be as high as 69% [97]. Despite these reported response rates, the true radiologic response rate is more probably between 10% and 40% [98,99]. More recently, anti-proliferative activity has also been shown for temozolomide. Data came from retrospective studies[100] as well as from a prospective randomized study comparing capecitabine plus temozolomide to temozolomide alone in pNENs that revealed a median progression-free survival longer in the combination arm (22.7 mo vs 14.4 mo, hazard ratio 0.58, P = 0.023), but satisfactory in both[101]. Moreover, a recent realworld analysis confirmed the combination of capecitabine and temozolomide as an active treatment for metastatic NENs[102]. Because of these studies, the use of capecitabine plus temozolomide has become routine for advanced pNENs, including gastrinomas.

Lastly, peptide receptor radionuclide therapy (PRRT) may be the most promising systemic therapy, and it has been repeatedly reported as particularly useful for symptom relief in functioning forms, even if this aspect might be less important in the setting of gastrinomas due to concomitant PPI treatment which is considered to be the first-line approach for symptoms' control[10]. Two different isotopes have been used in most studies: 90Yttrium (90Y)- or 177Lutetium (177Lu)-labeled SSAs[103]. The approval of PRRT treatment comes from the promising results of a double-blinded, control phase 3 trial (NETTER-1)[104] in patients with advanced unresectable, midgut carcinoids and the results of treatment of 510 patients with advanced pNENs and other NENs[105,106]. According to data from the literature, gastrinomas are one of the malignant pNENs that were most responsive to PRRT; however, they also had one of the highest recurrence rates leading to a poorer prognosis[103,105]. In detail, in one study including 11 patients with metastatic ZES[107] treated with either <sup>90</sup>Y-and/or <sup>177</sup> Lu-labeled SSAs, the mean serum gastrin decreased by 81%, complete response occurred in 9%, partial tumor response in 45%, tumor stabilization in 45%, with a persistence of the antitumor effect for a median period of 14 mo in 64% of the cases. Another study[108] involving 30 gastrinoma patients treated with <sup>90</sup>Y-labeled SSAs reported a partial response rate of 33% with a mean OS time of 40 mo.

#### CONCLUSION

As the diagnosis of ZES is challenging; the maintenance of a high index of suspicion is necessary to get the final diagnosis. Better disease awareness is useful to reduce the diagnostic delay, particularly due to the improper referral of patients to physicians with low or no expertise in the neuroendocrine field. The association between typical symptoms including chronic diarrhea, reflux disorder, and recurrent peptic disease particularly at unusual sites should raise the suspicion of ZES after exclusion of alternative and more common GI etiologies. The possibility of an underlying MEN-1 syndrome should be always considered, particularly in young patients with concomitant hypercalcemia suggestive of hyperparathyroidism and/or familiar history of MEN-1. A fasting gastrin level is generally the first step and confounding factors such as PPI use need to be considered. Gastric pH, esophageal pH-recording, and possibly a secretin stimulation test might be necessary as well, although the decision to perform them should be tailored to every single patient, considered both the need to withdraw PPI treatment and the limited availability of these tests in routine clinical practice. Tumor localization must be performed and EUS with the possibility of getting a sampling through FNA is considered to be a more accurate technique than conventional imaging for small lesions. Given the high expression of STTRs in gastrinomas, gallium-68PET-scan should be always included in the diagnostic pathway of all NENs, including gastrinoma, in order to both identify the primary tumor and to stage the disease.

Regarding the treatment of the localized disease, the two milestones are represented by PPIs for symptoms' control and surgery with curative intent. The role of surgery in



the treatment of gastrinoma has changed completely from the introduction of PPIs. In the past, total gastrectomy represented the sole effective treatment to treat ZES by removing the end-organ target of gastrin. With the use of PPIs, gastric hypersecretion was no longer considered a problem and surgical excision started to be proposed as a potentially curative therapy. Surgical removal of the primary tumor (and possibly its metastases) with curative intent should be, indeed, always performed. Unfortunately, the diagnosis is often made when the disease is too advanced for a surgical approach. The first step, again, is represented by syndrome control, based on PPIs, which are considered to be the drugs of choice for suppressing acid secretion. In order to achieve tumor growth control, SSAs constitute a viable option; studies specifically focused on advanced gastrinomas are scanty and often retrospective, however, according to data from the literature, treatments for the advanced disease are super-imposable to other NENs and include targeted therapies, chemotherapy, and PRRT. As there is a need for both a proper medical treatment for symptom's relief and a surgical procedure whenever feasible with curative intent, the multidisciplinary approach, with close cooperation between clinicians and surgeons, remains the cornerstone for proper management of this composite disease. Due to the risk of overlapping ZES with other GI common disorders, referral to tertiary centers with great expertise in the neuroendocrine field is mandatory.

#### ACKNOWLEDGEMENTS

Pietro Invernizzi and Sara Massironi are members of the European Reference Network on Hepatological Diseases (ERN RARE LIVER), and they thank AMAF Monza ONLUS and AIRCS for their support.

#### REFERENCES

- Zollinger RM, Ellison EH. Primary peptic ulcerations of the jejunum associated with islet cell tumors of the pancreas. Ann Surg 1955; 142: 709-23; discussion, 724 [PMID: 13259432 DOI: 10.1097/00000658-195510000-00015
- 2 Falconi M, Eriksson B, Kaltsas G, Bartsch DK, Capdevila J, Caplin M, Kos-Kudla B, Kwekkeboom D, Rindi G, Klöppel G, Reed N, Kianmanesh R, Jensen RT; Vienna Consensus Conference participants. ENETS Consensus Guidelines Update for the Management of Patients with Functional Pancreatic Neuroendocrine Tumors and Non-Functional Pancreatic Neuroendocrine Tumors. Neuroendocrinology 2016; 103: 153-171 [PMID: 26742109 DOI: 10.1159/000443171]
- Guarnotta V, Martini C, Davì MV, Pizza G, Colao A, Faggiano A; NIKE group. The Zollinger-3 Ellison syndrome: is there a role for somatostatin analogues in the treatment of the gastrinoma? Endocrine 2018; 60: 15-27 [PMID: 29019150 DOI: 10.1007/s12020-017-1420-4]
- Ito T, Igarashi H, Jensen RT. Zollinger-Ellison syndrome: recent advances and controversies. Curr Opin Gastroenterol 2013; 29: 650-661 [PMID: 24100728 DOI: 10.1097/MOG.0b013e328365efb1]
- 5 Ito T, Cadiot G, Jensen RT. Diagnosis of Zollinger-Ellison syndrome: increasingly difficult. World J Gastroenterol 2012; 18: 5495-5503 [PMID: 23112541 DOI: 10.3748/wjg.v18.i39.5495]
- Jensen RT, Niederle B, Mitry E, Ramage JK, Steinmuller T, Lewington V, Scarpa A, Sundin A, 6 Perren A, Gross D, O'Connor JM, Pauwels S, Kloppel G; Frascati Consensus Conference; European Neuroendocrine Tumor Society. Gastrinoma (duodenal and pancreatic). Neuroendocrinology 2006; 84: 173-182 [PMID: 17312377 DOI: 10.1159/000098009]
- 7 Keutgen XM, Nilubol N, Kebebew E. Malignant-functioning neuroendocrine tumors of the pancreas: A survival analysis. Surgery 2016; 159: 1382-1389 [PMID: 26704781 DOI: 10.1016/j.surg.2015.11.010]
- 8 Gibril F, Jensen RT. Zollinger-Ellison syndrome revisited: diagnosis, biologic markers, associated inherited disorders, and acid hypersecretion. Curr Gastroenterol Rep 2004; 6: 454-463 [PMID: 15527675 DOI: 10.1007/s11894-004-0067-5]
- Mendelson AH, Donowitz M. Catching the Zebra: Clinical Pearls and Pitfalls for the Successful Diagnosis of Zollinger-Ellison Syndrome. Dig Dis Sci 2017; 62: 2258-2265 [PMID: 28776139 DOI: 10.1007/s10620-017-4695-7
- Jensen RT, Ito T. Gastrinoma. 2020 Nov 21. In: Feingold KR, Anawalt B, Boyce A, Chrousos G, 10 de Herder WW, Dhatariya K, Dungan K, Grossman A, Hershman JM, Hofland J, Kalra S, Kaltsas G, Koch C, Kopp P, Korbonits M, Kovacs CS, Kuohung W, Laferrère B, McGee EA, McLachlan R, Morley JE, New M, Purnell J, Sahay R, Singer F, Stratakis CA, Trence DL, Wilson DP, editors. Endotext [Internet]. South Dartmouth (MA): MDText.com, Inc.; 2000 [PMID: 25905301]
- Wilcox CM, Seay T, Arcury JT, Mohnen J, Hirschowitz BI. Zollinger-Ellison syndrome: 11 presentation, response to therapy, and outcome. Dig Liver Dis 2011; 43: 439-443 [PMID: 21193359 DOI: 10.1016/j.dld.2010.11.007]



- 12 Hatta W, Iijima K, Koike T, Kondo Y, Ara N, Asanuma K, Uno K, Asano N, Imatani A, Shimosegawa T. Endoscopic findings for predicting gastric acid secretion status. Dig Endosc 2015; 27: 582-589 [PMID: 25556402 DOI: 10.1111/den.12427]
- 13 Roy PK, Venzon DJ, Shojamanesh H, Abou-Saif A, Peghini P, Doppman JL, Gibril F, Jensen RT. Zollinger-Ellison syndrome. Clinical presentation in 261 patients. Medicine (Baltimore) 2000; 79: 379-411 [PMID: 11144036 DOI: 10.1097/00005792-200011000-00004]
- 14 Thakker RV, Newey PJ, Walls GV, Bilezikian J, Dralle H, Ebeling PR, Melmed S, Sakurai A, Tonelli F, Brandi ML; Endocrine Society. Clinical practice guidelines for multiple endocrine neoplasia type 1 (MEN1). J Clin Endocrinol Metab 2012; 97: 2990-3011 [PMID: 22723327 DOI: 10.1210/jc.2012-1230
- 15 Gibril F, Schumann M, Pace A, Jensen RT. Multiple endocrine neoplasia type 1 and Zollinger-Ellison syndrome: a prospective study of 107 cases and comparison with 1009 cases from the literature. Medicine (Baltimore) 2004; 83: 43-83 [PMID: 14747767 DOI: 10.1097/01.md.0000112297.72510.32]
- 16 Norton JA, Venzon DJ, Berna MJ, Alexander HR, Fraker DL, Libutti SK, Marx SJ, Gibril F, Jensen RT. Prospective study of surgery for primary hyperparathyroidism (HPT) in multiple endocrine neoplasia-type 1 and Zollinger-Ellison syndrome: long-term outcome of a more virulent form of HPT. Ann Surg 2008; 247: 501-510 [PMID: 18376196 DOI: 10.1097/SLA.0b013e31815efda5]
- Davì MV, Boninsegna L, Dalle Carbonare L, Toaiari M, Capelli P, Scarpa A, Francia G, Falconi M. 17 Presentation and outcome of pancreaticoduodenal endocrine tumors in multiple endocrine neoplasia type 1 syndrome. Neuroendocrinology 2011; 94: 58-65 [PMID: 21464564 DOI: 10.1159/000326164]
- Aamar A, Madhani K, Virk H, Butt Z. Zollinger-Ellison Syndrome: A Rare Case of Chronic 18 Diarrhea. Gastroenterology Res 2016; 9: 103-104 [PMID: 28058079 DOI: 10.14740/gr734w]
- 19 Campana D, Piscitelli L, Mazzotta E, Bonora M, Serra C, Salomone L, Corinaldesi R, Tomassetti P. Zollinger-Ellison syndrome. Diagnosis and therapy. Minerva Med 2005; 96: 187-206 [PMID: 161751611
- 20 Simmons LH, Guimaraes AR, Zukerberg LR. Case records of the Massachusetts General Hospital. Case 6-2013. A 54-year-old man with recurrent diarrhea. N Engl J Med 2013; 368: 757-765 [PMID: 23425169 DOI: 10.1056/NEJMcpc1208149]
- 21 Arasaradnam RP, Brown S, Forbes A, Fox MR, Hungin P, Kelman L, Major G, O'Connor M, Sanders DS, Sinha R, Smith SC, Thomas P, Walters JRF. Guidelines for the investigation of chronic diarrhoea in adults: British Society of Gastroenterology, 3rd edition. Gut 2018; 67: 1380-1399 [PMID: 29653941 DOI: 10.1136/gutjnl-2017-315909]
- 22 Corleto VD, Annibale B, Gibril F, Angeletti S, Serrano J, Venzon DJ, Delle Fave G, Jensen RT. Does the widespread use of proton pump inhibitors mask, complicate and/or delay the diagnosis of Zollinger-Ellison syndrome? Aliment Pharmacol Ther 2001; 15: 1555-1561 [PMID: 11563994 DOI: 10.1046/j.1365-2036.2001.01085.x]
- Metz DC. Diagnosis of the Zollinger-Ellison syndrome. Clin Gastroenterol Hepatol 2012; 10: 126-23 130 [PMID: 21806955 DOI: 10.1016/j.cgh.2011.07.012]
- 24 Epelboym I, Mazeh H. Zollinger-Ellison syndrome: classical considerations and current controversies. Oncologist 2014; 19: 44-50 [PMID: 24319020 DOI: 10.1634/theoncologist.2013-0369]
- 25 Phan J, Benhammou JN, Pisegna JR. Gastric Hypersecretory States: Investigation and Management. Curr Treat Options Gastroenterol 2015; 13: 386-397 [PMID: 26342486 DOI: 10.1007/s11938-015-0065-8
- 26 Oh DS, Wang HS, Ohning GV, Pisegna JR. Validation of a new endoscopic technique to assess acid output in Zollinger-Ellison syndrome. Clin Gastroenterol Hepatol 2006; 4: 1467-1473 [PMID: 17101299 DOI: 10.1016/j.cgh.2006.08.015]
- Kulke MH, Anthony LB, Bushnell DL, de Herder WW, Goldsmith SJ, Klimstra DS, Marx SJ, 27 Pasieka JL, Pommier RF, Yao JC, Jensen RT; North American Neuroendocrine Tumor Society (NANETS). NANETS treatment guidelines: well-differentiated neuroendocrine tumors of the stomach and pancreas. Pancreas 2010; 39: 735-752 [PMID: 20664472 DOI: 10.1097/MPA.0b013e3181ebb168
- Krampitz GW, Norton JA. Current management of the Zollinger-Ellison syndrome. Adv Surg 2013; 47: 59-79 [PMID: 24298844 DOI: 10.1016/j.yasu.2013.02.004]
- 29 Reznek RH. CT/MRI of neuroendocrine tumours. Cancer Imaging 2006; 6: S163-S177 [PMID: 17114072 DOI: 10.1102/1470-7330.2006.9037]
- 30 Thoeni RF, Mueller-Lisse UG, Chan R, Do NK, Shyn PB. Detection of small, functional islet cell tumors in the pancreas: selection of MR imaging sequences for optimal sensitivity. Radiology 2000; 214: 483-490 [PMID: 10671597 DOI: 10.1148/radiology.214.2.r00fe32483]
- 31 Metz DC, Jensen RT. Gastrointestinal neuroendocrine tumors: pancreatic endocrine tumors. Gastroenterology 2008; 135: 1469-1492 [PMID: 18703061 DOI: 10.1053/j.gastro.2008.05.047]
- 32 Stokkel MP, Rietbergen DD, Korse CM, Taal BG. Somatostatin receptor scintigraphy and chromogranin A assay in staging and follow-up of patients with well-differentiated neuroendocrine tumors. Nucl Med Commun 2011; 32: 731-737 [PMID: 21633314 DOI: 10.1097/MNM.0b013e328347a895]
- Ruf J, von Wedel F, Furth C, Denecke T, Stelter L, Steffen IG, Schütte K, Arend J, Ulrich G, Klose 33 S, Bornschein J, Apostolova I, Amthauer H. Significance of a Single-Time-Point Somatostatin



Receptor SPECT/Multiphase CT Protocol in the Diagnostic Work-up of Gastroenteropancreatic Neuroendocrine Neoplasms. J Nucl Med 2016; 57: 180-185 [PMID: 26609177 DOI: 10.2967/jnumed.115.161117

- 34 Wong KK, Gandhi A, Viglianti BL, Fig LM, Rubello D, Gross MD. Endocrine radionuclide scintigraphy with fusion single photon emission computed tomography/computed tomography. World J Radiol 2016; 8: 635-655 [PMID: 27358692 DOI: 10.4329/wjr.v8.i6.635]
- 35 Sainz-Esteban A, Olmos R, González-Sagrado M, González ML, Ruiz MÁ, García-Talavera P, Gamazo C, Villanueva JG, Cobo A, de Luis D. Contribution of 111In-pentetreotide SPECT/CT imaging to conventional somatostatin receptor scintigraphy in the detection of neuroendocrine tumours. Nucl Med Commun 2015; 36: 251-259 [PMID: 25369750 DOI: 10.1097/MNM.00000000000239
- 36 Johnbeck CB, Knigge U, Kjær A. PET tracers for somatostatin receptor imaging of neuroendocrine tumors: current status and review of the literature. Future Oncol 2014; 10: 2259-2277 [PMID: 25471038 DOI: 10.2217/fon.14.139]
- 37 Gabriel M, Decristoforo C, Kendler D, Dobrozemsky G, Heute D, Uprimny C, Kovacs P, Von Guggenberg E, Bale R, Virgolini IJ. 68Ga-DOTA-Tyr3-octreotide PET in neuroendocrine tumors: comparison with somatostatin receptor scintigraphy and CT. JNucl Med 2007; 48: 508-518 [PMID: 17401086 DOI: 10.2967/jnumed.106.035667]
- Skoura E, Michopoulou S, Mohmaduvesh M, Panagiotidis E, Al Harbi M, Toumpanakis C, Almukhailed O, Kayani I, Syed R, Navalkissoor S, Ell PJ, Caplin ME, Bomanji J. The Impact of 68Ga-DOTATATE PET/CT Imaging on Management of Patients with Neuroendocrine Tumors: Experience from a National Referral Center in the United Kingdom. J Nucl Med 2016; 57: 34-40 [PMID: 26471695 DOI: 10.2967/jnumed.115.166017]
- 39 Sharma P, Arora S, Mukherjee A, Pal S, Sahni P, Garg P, Khadgawat R, Thulkar S, Bal C, Kumar R. Predictive value of 68Ga-DOTANOC PET/CT in patients with suspicion of neuroendocrine tumors: is its routine use justified? Clin Nucl Med 2014; 39: 37-43 [PMID: 24152621 DOI: 10.1097/RLU.00000000000257
- 40 Wild D, Bomanji JB, Benkert P, Maecke H, Ell PJ, Reubi JC, Caplin ME. Comparison of 68Ga-DOTANOC and 68Ga-DOTATATE PET/CT within patients with gastroenteropancreatic neuroendocrine tumors. J Nucl Med 2013; 54: 364-372 [PMID: 23297077 DOI: 10.2967/jnumed.112.111724]
- 41 Geijer H, Breimer LH. Somatostatin receptor PET/CT in neuroendocrine tumours: update on systematic review and meta-analysis. Eur J Nucl Med Mol Imaging 2013; 40: 1770-1780 [PMID: 23873003 DOI: 10.1007/s00259-013-2482-z]
- 42 Norton JA, Jensen RT. Resolved and unresolved controversies in the surgical management of patients with Zollinger-Ellison syndrome. Ann Surg 2004; 240: 757-773 [PMID: 15492556 DOI: 10.1097/01.sla.0000143252.02142.3e]
- 43 Zilli A, Arcidiacono PG, Conte D, Massironi S. Clinical impact of endoscopic ultrasonography on the management of neuroendocrine tumors: lights and shadows. Dig Liver Dis 2018; 50: 6-14 [PMID: 29102525 DOI: 10.1016/j.dld.2017.10.007]
- 44 Atiq M, Bhutani MS, Bektas M, Lee JE, Gong Y, Tamm EP, Shah CP, Ross WA, Yao J, Raju GS, Wang X, Lee JH. EUS-FNA for pancreatic neuroendocrine tumors: a tertiary cancer center experience. Dig Dis Sci 2012; 57: 791-800 [PMID: 21964743 DOI: 10.1007/s10620-011-1912-7]
- 45 Thomas-Marques L, Murat A, Delemer B, Penfornis A, Cardot-Bauters C, Baudin E, Niccoli-Sire P, Levoir D, Choplin Hdu B, Chabre O, Jovenin N, Cadiot G; Groupe des Tumeurs Endocrines (GTE). Prospective endoscopic ultrasonographic evaluation of the frequency of nonfunctioning pancreaticoduodenal endocrine tumors in patients with multiple endocrine neoplasia type 1. Am J Gastroenterol 2006; 101: 266-273 [PMID: 16454829 DOI: 10.1111/j.1572-0241.2006.00367.x]
- 46 Kappelle WF, Valk GD, Leenders M, Moons LM, Bogte A, Siersema PD, Vleggaar FP. Growth rate of small pancreatic neuroendocrine tumors in multiple endocrine neoplasia type 1: results from an endoscopic ultrasound based cohort study. Endoscopy 2017; 49: 27-34 [PMID: 27975336 DOI: 10.1055/s-0042-119402
- 47 Trudgill NJ, Sifrim D, Sweis R, Fullard M, Basu K, McCord M, Booth M, Hayman J, Boeckxstaens G, Johnston BT, Ager N, De Caestecker J. British Society of Gastroenterology guidelines for oesophageal manometry and oesophageal reflux monitoring. Gut 2019; 68: 1731-1750 [PMID: 31366456 DOI: 10.1136/gutjnl-2018-318115]
- 48 Long SH, Berna MJ, Thill M, Pace A, Pradhan TK, Hoffmann KM, Serrano J, Jensen RT. Secretinreceptor and secretin-receptor-variant expression in gastrinomas: correlation with clinical and tumoral features and secretin and calcium provocative test results. J Clin Endocrinol Metab 2007; 92: 4394-4402 [PMID: 17711922 DOI: 10.1210/jc.2007-0986]
- 49 Metz DC, Buchanan M, Purich E, Fein S, A randomized controlled crossover study comparing synthetic porcine and human secretins with biologically derived porcine secretin to diagnose Zollinger-Ellison Syndrome. Aliment Pharmacol Ther 2001; 15: 669-676 [PMID: 11328261 DOI: 10.1046/j.1365-2036.2001.00976.x
- Berna MJ, Hoffmann KM, Long SH, Serrano J, Gibril F, Jensen RT. Serum gastrin in Zollinger-50 Ellison syndrome: II. Prospective study of gastrin provocative testing in 293 patients from the National Institutes of Health and comparison with 537 cases from the literature. evaluation of diagnostic criteria, proposal of new criteria, and correlations with clinical and tumoral features. Medicine (Baltimore) 2006; 85: 331-364 [PMID: 17108779 DOI: 10.1097/MD.0b013e31802b518c]



- 51 Poitras P, Gingras MH, Rehfeld JF. Secretin stimulation test for gastrin release in Zollinger-Ellison syndrome: to do or not to do? Pancreas 2013; 42: 903-904 [PMID: 23851427 DOI: 10.1097/MPA.0b013e318298df75
- 52 Kuiper P, Biemond I, Masclee AA, Jansen JB, Verspaget HW, Lamers CB. Diagnostic efficacy of the secretin stimulation test for the Zollinger-Ellison syndrome: an intra-individual comparison using different dosages in patients and controls. Pancreatology 2010; 10: 14-18 [PMID: 20299818 DOI: 10.1159/000265936
- Shah P, Singh MH, Yang YX, Metz DC. Hypochlorhydria and achlorhydria are associated with 53 false-positive secretin stimulation testing for Zollinger-Ellison syndrome. Pancreas 2013; 42: 932-936 [PMID: 23851430 DOI: 10.1097/MPA.0b013e3182847b2e]
- Norton JA, Fraker DL, Alexander HR, Venzon DJ, Doppman JL, Serrano J, Goebel SU, Peghini 54 PL, Roy PK, Gibril F, Jensen RT. Surgery to cure the Zollinger-Ellison syndrome. N Engl J Med 1999; 341: 635-644 [PMID: 10460814 DOI: 10.1056/nejm199908263410902]
- 55 Jensen RT, Cadiot G, Brandi ML, de Herder WW, Kaltsas G, Komminoth P, Scoazec JY, Salazar R, Sauvanet A, Kianmanesh R; Barcelona Consensus Conference participants. ENETS Consensus Guidelines for the management of patients with digestive neuroendocrine neoplasms: functional pancreatic endocrine tumor syndromes. Neuroendocrinology 2012; 95: 98-119 [PMID: 22261919 DOI: 10.1159/0003355911
- Kunz PL, Reidy-Lagunes D, Anthony LB, Bertino EM, Brendtro K, Chan JA, Chen H, Jensen RT, 56 Kim MK, Klimstra DS, Kulke MH, Liu EH, Metz DC, Phan AT, Sippel RS, Strosberg JR, Yao JC; North American Neuroendocrine Tumor Society. Consensus guidelines for the management and treatment of neuroendocrine tumors. Pancreas 2013; 42: 557-577 [PMID: 23591432 DOI: 10.1097/MPA.0b013e31828e34a4
- 57 Howe JR, Merchant NB, Conrad C, Keutgen XM, Hallet J, Drebin JA, Minter RM, Lairmore TC, Tseng JF, Zeh HJ, Libutti SK, Singh G, Lee JE, Hope TA, Kim MK, Menda Y, Halfdanarson TR, Chan JA, Pommier RF, The North American Neuroendocrine Tumor Society Consensus Paper on the Surgical Management of Pancreatic Neuroendocrine Tumors. Pancreas 2020; 49: 1-33 [PMID: 31856076 DOI: 10.1097/MPA.00000000001454]
- 58 Pavel M, Öberg K, Falconi M, Krenning EP, Sundin A, Perren A, Berruti A; ESMO Guidelines Committee. Gastroenteropancreatic neuroendocrine neoplasms: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol 2020; 31: 844-860 [PMID: 32272208 DOI: 10.1016/j.annonc.2020.03.304]
- 59 Norton JA, Fraker DL, Alexander HR, Jensen RT. Value of surgery in patients with negative imaging and sporadic Zollinger-Ellison syndrome. Ann Surg 2012; 256: 509-517 [PMID: 22868363 DOI: 10.1097/SLA.0b013e318265f08d]
- Norton JA, Alexander HR, Fraker DL, Venzon DJ, Gibril F, Jensen RT. Does the use of routine 60 duodenotomy (DUODX) affect rate of cure, development of liver metastases, or survival in patients with Zollinger-Ellison syndrome? Ann Surg 2004; 239: 617-25; discussion 626 [PMID: 15082965 DOI: 10.1097/01.sla.0000124290.05524.5e]
- 61 Weber HC, Venzon DJ, Lin JT, Fishbein VA, Orbuch M, Strader DB, Gibril F, Metz DC, Fraker DL, Norton JA. Determinants of metastatic rate and survival in patients with Zollinger-Ellison syndrome: a prospective long-term study. Gastroenterology 1995; 108: 1637-1649 [PMID: 7768367 DOI: 10.1016/0016-5085(95)90124-8]
- Norton JA, Fraker DL, Alexander HR, Gibril F, Liewehr DJ, Venzon DJ, Jensen RT. Surgery 62 increases survival in patients with gastrinoma. Ann Surg 2006; 244: 410-419 [PMID: 16926567 DOI: 10.1097/01.sla.0000234802.44320.a5]
- Fraker DL, Norton JA, Alexander HR, Venzon DJ, Jensen RT. Surgery in Zollinger-Ellison 63 syndrome alters the natural history of gastrinoma. Ann Surg 1994; 220: 320-8; discussion 328 [PMID: 7916560 DOI: 10.1097/00000658-199409000-00008]
- 64 Sugg SL, Norton JA, Fraker DL, Metz DC, Pisegna JR, Fishbeyn V, Benya RV, Shawker TH, Doppman JL, Jensen RT. A prospective study of intraoperative methods to diagnose and resect duodenal gastrinomas. Ann Surg 1993; 218: 138-144 [PMID: 8342993 DOI: 10.1097/0000658-199308000-00004
- Norton JA, Doppman JL, Jensen RT. Curative resection in Zollinger-Ellison syndrome. Results of a 65 10-year prospective study. Ann Surg 1992; 215: 8-18 [PMID: 1531004 DOI: 10.1097/00000658-199201000-00012
- Norton JA, Krampitz GW, Poultsides GA, Visser BC, Fraker DL, Alexander HR, Jensen RT. 66 Prospective Evaluation of Results of Reoperation in Zollinger-Ellison Syndrome. Ann Surg 2018; 267: 782-788 [PMID: 29517561 DOI: 10.1097/SLA.00000000002122]
- Giovinazzo F, Butturini G, Monsellato D, Malleo G, Marchegiani G, Bassi C. Lymph nodes 67 metastasis and recurrences justify an aggressive treatment of gastrinoma. Updates Surg 2013; 65: 19-24 [PMID: 23417896 DOI: 10.1007/s13304-013-0201-8]
- 68 Bartsch DK, Waldmann J, Fendrich V, Boninsegna L, Lopez CL, Partelli S, Falconi M. Impact of lymphadenectomy on survival after surgery for sporadic gastrinoma. Br J Surg 2012; 99: 1234-1240 [PMID: 22864882 DOI: 10.1002/bjs.8843]
- 69 Liu P, Zhang X, Shang Y, Lu L, Cao F, Sun M, Tang Z, Vollmar B, Gong P. Lymph node ratio, but not the total number of examined lymph nodes or lymph node metastasis, is a predictor of overall survival for pancreatic neuroendocrine neoplasms after surgical resection. Oncotarget 2017; 8: 89245-89255 [PMID: 29179516 DOI: 10.18632/oncotarget.19184]



- 70 Conrad C, Kutlu OC, Dasari A, Chan JA, Vauthey JN, Adams DB, Kim M, Fleming JB, Katz MH, Lee JE. Prognostic Value of Lymph Node Status and Extent of Lymphadenectomy in Pancreatic Neuroendocrine Tumors Confined To and Extending Beyond the Pancreas. J Gastrointest Surg 2016; 20: 1966-1974 [PMID: 27714644 DOI: 10.1007/s11605-016-3243-7]
- 71 Curran T, Pockaj BA, Gray RJ, Halfdanarson TR, Wasif N. Importance of lymph node involvement in pancreatic neuroendocrine tumors: impact on survival and implications for surgical resection. J Gastrointest Surg 2015; 19: 152-60; discussion 160 [PMID: 25118642 DOI: 10.1007/s11605-014-2624-z]
- 72 Norton JA, Alexander HR, Fraker DL, Venzon DJ, Gibril F, Jensen RT. Possible primary lymph node gastrinoma: occurrence, natural history, and predictive factors: a prospective study. Ann Surg 2003; 237: 650-7; discussion 657 [PMID: 12724631 DOI: 10.1097/01.SLA.0000064375.51939.48]
- 73 Chen Y, Deshpande V, Ferrone C, Blaszkowsky LS, Parangi S, Warshaw AL, Lillemoe KD, Fernandez-Del Castillo C. Primary lymph node gastrinoma: A single institution experience. Surgery 2017; 162: 1088-1094 [PMID: 28705492 DOI: 10.1016/j.surg.2017.05.017]
- 74 Massironi S, Campana D, Partelli S, Panzuto F, Rossi RE, Faggiano A, Brighi N, Falconi M, Rinzivillo M, Delle Fave G, Colao AM, Conte D. Heterogeneity of Duodenal Neuroendocrine Tumors: An Italian Multi-center Experience. Ann Surg Oncol 2018; 25: 3200-3206 [PMID: 30054824 DOI: 10.1245/s10434-018-6673-5]
- Jensen RT, Fraker DL. Zollinger-Ellison syndrome. Advances in treatment of gastric hypersecretion 75 and the gastrinoma. JAMA 1994; 271: 1429-1435 [PMID: 7513768 DOI: 10.1001/jama.271.18.1429]
- Metz DC, Pisegna JR, Fishbeyn VA, Benya RV, Jensen RT. Control of gastric acid hypersecretion 76 in the management of patients with Zollinger-Ellison syndrome. World J Surg 1993; 17: 468-480 [PMID: 8362529 DOI: 10.1007/BF01655106]
- Hirschowitz BI, Simmons J, Mohnen J. Clinical outcome using lansoprazole in acid hypersecretors 77 with and without Zollinger-Ellison syndrome: a 13-year prospective study. Clin Gastroenterol Hepatol 2005; 3: 39-48 [PMID: 15645403 DOI: 10.1016/S1542-3565(04)00606-8]
- 78 Metz DC, Comer GM, Soffer E, Forsmark CE, Cryer B, Chey W, Pisegna JR. Three-year oral pantoprazole administration is effective for patients with Zollinger-Ellison syndrome and other hypersecretory conditions. Aliment Pharmacol Ther 2006; 23: 437-444 [PMID: 16423003 DOI: 10.1111/j.1365-2036.2006.02762.x
- 79 Ip S, Bonis P, Tatsioni A, Raman G, Chew P, Kupelnick B, Fu L, DeVine D, Lau J. Comparative Effectiveness of Management Strategies For Gastroesophageal Reflux Disease [Internet]. Rockville (MD): Agency for Healthcare Research and Quality (US); 2005 Dec. Report No.: 06-EHC003-EF [PMID: 21348043]
- Quatrini M, Castoldi L, Rossi G, Cesana BM, Peracchi M, Bardella MT. A follow-up study of 80 patients with Zollinger-Ellison syndrome in the period 1966-2002: effects of surgical and medical treatments on long-term survival. J Clin Gastroenterol 2005; 39: 376-380 [PMID: 15815204 DOI: 10.1097/01.mcg.0000159221.77913.ac
- Maton PN, Vinayek R, Frucht H, McArthur KA, Miller LS, Saeed ZA, Gardner JD, Jensen RT. 81 Long-term efficacy and safety of omeprazole in patients with Zollinger-Ellison syndrome: a prospective study. Gastroenterology 1989; 97: 827-836 [PMID: 2777040 DOI: 10.1016/0016-5085(89)91485-6]
- 82 Poitras P, Gingras MH, Rehfeld JF. The Zollinger-Ellison syndrome: dangers and consequences of interrupting antisecretory treatment. Clin Gastroenterol Hepatol 2012; 10: 199-202 [PMID: 21871248 DOI: 10.1016/j.cgh.2011.08.012]
- 83 Metz DC, Pisegna JR, Fishbeyn VA, Benya RV, Feigenbaum KM, Koviack PD, Jensen RT. Currently used doses of omeprazole in Zollinger-Ellison syndrome are too high. Gastroenterology 1992; 103: 1498-1508 [PMID: 1426868 DOI: 10.1016/0016-5085(92)91170-9]
- 84 Norton JA, Foster DS, Ito T, Jensen RT. Gastrinomas: Medical or Surgical Treatment. Endocrinol Metab Clin North Am 2018; 47: 577-601 [PMID: 30098717 DOI: 10.1016/j.ecl.2018.04.009]
- 85 Dockray GJ, Moore A, Varro A, Pritchard DM. Gastrin receptor pharmacology. Curr Gastroenterol Rep 2012; 14: 453-459 [PMID: 22983899 DOI: 10.1007/s11894-012-0293-1]
- 86 Ellison EC, Johnson JA. The Zollinger-Ellison syndrome: a comprehensive review of historical, scientific, and clinical considerations. Curr Probl Surg 2009; 46: 13-106 [PMID: 19059523 DOI: 10.1067/j.cpsurg.2008.09.001
- Tomassetti P, Migliori M, Gullo L. Slow-release lanreotide treatment in endocrine gastrointestinal tumors. Am J Gastroenterol 1998; 93: 1468-1471 [PMID: 9732927 DOI: 10.1111/j.1572-0241.1998.465\_q.x]
- Eriksson B, Renstrup J, Imam H, Oberg K. High-dose treatment with lanreotide of patients with 88 advanced neuroendocrine gastrointestinal tumors: clinical and biological effects. Ann Oncol 1997; 8: 1041-1044 [PMID: 9402179 DOI: 10.1023/A:1008205415035]
- 89 Rinke A, Müller HH, Schade-Brittinger C, Klose KJ, Barth P, Wied M, Mayer C, Aminossadati B, Pape UF, Bläker M, Harder J, Arnold C, Gress T, Arnold R; PROMID Study Group. Placebocontrolled, double-blind, prospective, randomized study on the effect of octreotide LAR in the control of tumor growth in patients with metastatic neuroendocrine midgut tumors: a report from the PROMID Study Group. J Clin Oncol 2009; 27: 4656-4663 [PMID: 19704057 DOI: 10.1200/JCO.2009.22.8510
- 90 Caplin ME, Pavel M, Ćwikła JB, Phan AT, Raderer M, Sedláčková E, Cadiot G, Wolin EM, Capdevila J, Wall L, Rindi G, Langley A, Martinez S, Blumberg J, Ruszniewski P; CLARINET



Investigators. Lanreotide in metastatic enteropancreatic neuroendocrine tumors. N Engl J Med 2014; 371: 224-233 [PMID: 25014687 DOI: 10.1056/NEJMoa1316158]

- 91 Saijo F. Naito H. Funavama Y. Fukushima K. Shibata C. Hashimoto A. Kitavama T. Nagao M. Matsuno S, Sasaki I. Octreotide in control of multiple liver metastases from gastrinoma. J Gastroenterol 2003; 38: 905-908 [PMID: 14564638 DOI: 10.1007/s00535-002-1170-8]
- 92 Granberg D, Jacobsson H, Oberg K, Gustavsson J, Lehtihet M. Regression of a large malignant gastrinoma on treatment with Sandostatin LAR: a case report. Digestion 2008; 77: 92-95 [PMID: 18376130 DOI: 10.1159/000122229]
- 93 Yamaguchi M, Yamada Y, Hosokawa Y, Iwamoto R, Tamba S, Ihara A, Yamamoto K, Hoshida Y, Matsuzawa Y. Long-term suppressive effect of octreotide on progression of metastatic gastrinoma with multiple endocrine neoplasia type 1: seven-year follow up. Intern Med 2010; 49: 1557-1563 [PMID: 20686291 DOI: 10.2169/internalmedicine.49.3607]
- 94 Ruszniewski P, Laucournet H, Elouaer-Blanc L, Mignon M, Bonfils S. Long-acting somatostatin (SMS 201-995) in the management of Zollinger-Ellison syndrome: evidence for sustained efficacy. Pancreas 1988; 3: 145-152 [PMID: 2897687 DOI: 10.1097/00006676-198804000-00006]
- 95 Raymond E, Dahan L, Raoul JL, Bang YJ, Borbath I, Lombard-Bohas C, Valle J, Metrakos P, Smith D, Vinik A, Chen JS, Hörsch D, Hammel P, Wiedenmann B, Van Cutsem E, Patyna S, Lu DR, Blanckmeister C, Chao R, Ruszniewski P. Sunitinib malate for the treatment of pancreatic neuroendocrine tumors. N Engl J Med 2011; 364: 501-513 [PMID: 21306237 DOI: 10.1056/NEJMoa1003825
- Yao JC, Shah MH, Ito T, Bohas CL, Wolin EM, Van Cutsem E, Hobday TJ, Okusaka T, Capdevila J, de Vries EG, Tomassetti P, Pavel ME, Hoosen S, Haas T, Lincy J, Lebwohl D, Öberg K; RAD001 in Advanced Neuroendocrine Tumors, Third Trial (RADIANT-3) Study Group. Everolimus for advanced pancreatic neuroendocrine tumors. N Engl J Med 2011; 364: 514-523 [PMID: 21306238 DOI: 10.1056/NEJMoa1009290]
- 97 Moertel CG, Hanley JA, Johnson LA. Streptozocin alone compared with streptozocin plus fluorouracil in the treatment of advanced islet-cell carcinoma. N Engl J Med 1980; 303: 1189-1194 [PMID: 6252466 DOI: 10.1056/nejm198011203032101]
- 98 Prakash L, Bhosale P, Cloyd J, Kim M, Parker N, Yao J, Dasari A, Halperin D, Aloia T, Lee JE, Vauthey JN, Fleming JB, Katz MH. Role of Fluorouracil, Doxorubicin, and Streptozocin Therapy in the Preoperative Treatment of Localized Pancreatic Neuroendocrine Tumors. J Gastrointest Surg 2017; 21: 155-163 [PMID: 27634306 DOI: 10.1007/s11605-016-3270-4]
- 99 Kouvaraki MA, Ajani JA, Hoff P, Wolff R, Evans DB, Lozano R, Yao JC. Fluorouracil, doxorubicin, and streptozocin in the treatment of patients with locally advanced and metastatic pancreatic endocrine carcinomas. J Clin Oncol 2004; 22: 4762-4771 [PMID: 15570077 DOI: 10.1200/JCO.2004.04.024]
- 100 Cives M, Ghayouri M, Morse B, Brelsford M, Black M, Rizzo A, Meeker A, Strosberg J. Analysis of potential response predictors to capecitabine/temozolomide in metastatic pancreatic neuroendocrine tumors. Endocr Relat Cancer 2016; 23: 759-767 [PMID: 27552969 DOI: 10.1530/ERC-16-0147
- Kunz PL, Catalano PJ, Nimeiri H, Fisher GA, Longacre TA, Suarez CJ, Yao JC, Kulke MH, 101 Hendifar AE, Shanks JC, Shah MH, Zalupski M, Schmulbach EL, Reidy DL, Strosberg JR, O'Dwyer PJ, Benson AB. A randomized study of temozolomide or temozolomide and capecitabine in patients with advanced pancreatic neuroendocrine tumors: A trial of the ECOG-ACRIN Cancer Research Group (E2211). J Clin Oncol 2018; 36: 4004-4004 [DOI: 10.1200/jco.2018.36.15\_suppl.4004]
- 102 Bongiovanni A, Liverani C, Foca F, Fausti V, Di Menna G, Mercatali L, De Vita A, Riva N, Calpona S, Miserocchi G, Spadazzi C, Cocchi C, Ibrahim T. Temozolomide Alone or Combined with Capecitabine for the Treatment of Metastatic Neuroendocrine Neoplasia: a "Real World" data analysis. Neuroendocrinology 2020 [PMID: 33221806 DOI: 10.1159/000513218]
- 103 Ito T, Igarashi H, Jensen RT. Therapy of metastatic pancreatic neuroendocrine tumors (pNETs): recent insights and advances. J Gastroenterol 2012; 47: 941-960 [PMID: 22886480 DOI: 10.1007/s00535-012-0642-8
- Strosberg J, El-Haddad G, Wolin E, Hendifar A, Yao J, Chasen B, Mittra E, Kunz PL, Kulke MH, 104 Jacene H, Bushnell D, O'Dorisio TM, Baum RP, Kulkarni HR, Caplin M, Lebtahi R, Hobday T, Delpassand E, Van Cutsem E, Benson A, Srirajaskanthan R, Pavel M, Mora J, Berlin J, Grande E, Reed N, Seregni E, Öberg K, Lopera Sierra M, Santoro P, Thevenet T, Erion JL, Ruszniewski P, Kwekkeboom D, Krenning E; NETTER-1 Trial Investigators. Phase 3 Trial of <sup>177</sup>Lu-Dotatate for Midgut Neuroendocrine Tumors. N Engl J Med 2017; 376: 125-135 [PMID: 28076709 DOI: 10.1056/NEJMoa1607427
- 105 Kwekkeboom DJ, de Herder WW, Kam BL, van Eijck CH, van Essen M, Kooij PP, Feelders RA, van Aken MO, Krenning EP. Treatment with the radiolabeled somatostatin analog [177 Lu-DOTA 0, Tyr3]octreotate: toxicity, efficacy, and survival. J Clin Oncol 2008; 26: 2124-2130 [PMID: 18445841 DOI: 10.1200/JCO.2007.15.2553]
- 106 Kwekkeboom DJ, Teunissen JJ, Bakker WH, Kooij PP, de Herder WW, Feelders RA, van Eijck CH, Esser JP, Kam BL, Krenning EP. Radiolabeled somatostatin analog [177Lu-DOTA0, Tyr3 octreotate in patients with endocrine gastroenteropancreatic tumors. J Clin Oncol 2005; 23: 2754-2762 [PMID: 15837990 DOI: 10.1200/JCO.2005.08.066]
- Grozinsky-Glasberg S, Barak D, Fraenkel M, Walter MA, Müeller-Brand J, Eckstein J, Applebaum 107



L, Shimon I, Gross DJ. Peptide receptor radioligand therapy is an effective treatment for the longterm stabilization of malignant gastrinomas. Cancer 2011; 117: 1377-1385 [PMID: 21425137 DOI: 10.1002/cncr.25646]

108 Dumont RA, Seiler D, Marincek N, Brunner P, Radojewski P, Rochlitz C, Müller-Brand J, Maecke HR, Briel M, Walter MA. Survival after somatostatin based radiopeptide therapy with (90)Y-DOTATOC vs. (90)Y-DOTATOC plus (177)Lu-DOTATOC in metastasized gastrinoma. Am J Nucl Med Mol Imaging 2015; 5: 46-55 [PMID: 25625026 DOI: 10.7892/boris.67183]



WJG

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5908-5918

DOI: 10.3748/wjg.v27.i35.5908

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

MINIREVIEWS

# Optical diagnosis of colorectal polyps using convolutional neural networks

Rawen Kader, Andreas V Hadjinicolaou, Fanourios Georgiades, Danail Stoyanov, Laurence B Lovat

ORCID number: Rawen Kader 0000-0001-9133-0838; Andreas V Hadjinicolaou 0000-0002-6520-443X; Fanourios Georgiades 0000-0003-0440-2720; Danail Stoyanov 0000-0002-0980-3227; Laurence B Lovat 0000-0003-4542-3915.

Author contributions: Kader R, Hadjinicolaou AV and Georgiades F performed the literature review and wrote the manuscript: Stoyanov D and Lovat LB revised the manuscript; All authors have read and approved the final manuscript.

#### Conflict-of-interest statement:

Rawen Kader is supported by the Wellcome/EPSRC Centre for Interventional and Surgical Sciences (WEISS) at UCL; [203145Z/16/Z]. Danail Stoyanov owns shares in Odin Vision and Digital Surgery Ltd. Laurence B Lovat owns shares in Odin Vision. The remaining authors declare no conflict of interest.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially,

Rawen Kader, Danail Stoyanov, Laurence B Lovat, Wellcome/EPSRC Centre for Interventional and Surgical Sciences, University College London, London W1W 7TY, United Kingdom

Rawen Kader, Laurence B Lovat, Division of Surgery and Interventional Sciences, University College London, London W1W 7TY, United Kingdom

Andreas V Hadjinicolaou, MRC Cancer Unit, Department of Gastroenterology, University of Cambridge, Cambridge CB2 0QQ, United Kingdom

Fanourios Georgiades, Department of Surgery, University of Cambridge, Cambridge CB2 0QQ, United Kingdom

Danail Stoyanov, Department of Computer Science, University College London, London W1W 7TY, United Kingdom

Corresponding author: Rawen Kader, BMed, MBBS, MRCP, Research Fellow, Wellcome/ EPSRC Centre for Interventional and Surgical Sciences, University College London, Charles Bell House, 43-45 Foley Street, Fitzrovia, London W1W 7TY, United Kingdom. r.kader@nhs.net

# Abstract

Colonoscopy remains the gold standard investigation for colorectal cancer screening as it offers the opportunity to both detect and resect pre-malignant and neoplastic polyps. Although technologies for image-enhanced endoscopy are widely available, optical diagnosis has not been incorporated into routine clinical practice, mainly due to significant inter-operator variability. In recent years, there has been a growing number of studies demonstrating the potential of convolutional neural networks (CNN) to enhance optical diagnosis of polyps. Data suggest that the use of CNNs might mitigate the inter-operator variability amongst endoscopists, potentially enabling a "resect and discard" or "leave in" strategy to be adopted in real-time. This would have significant financial benefits for healthcare systems, avoid unnecessary polypectomies of non-neoplastic polyps and improve the efficiency of colonoscopy. Here, we review advances in CNN for the optical diagnosis of colorectal polyps, current limitations and future directions.

Key Words: Artificial intelligence; Deep learning; Convolutional neural networks; Computer aided diagnosis; Optical diagnosis; Colorectal polyps



and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: United Kingdom

#### Peer-review report's scientific quality classification

Grade A (Excellent): A Grade B (Very good): 0 Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Received: February 27, 2021 Peer-review started: February 27, 2021 First decision: April 18, 2021

Revised: April 29, 2021 Accepted: August 24, 2021 Article in press: August 24, 2021 Published online: September 21, 2021

P-Reviewer: Cavdar SC S-Editor: Ma YJ L-Editor: A P-Editor: Li JH



©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core Tip:** A convolutional neural network (CNN) is a specific type of artificial intelligence deep learning. These networks may play an important role in the coming years in assisting endoscopists to optically diagnose colorectal polyps. CNNs can mitigate the inter-operator variability amongst endoscopists, potentially enabling a "resect and discard" or "leave in" strategy to be adopted. This would improve the efficiency of colonoscopy, reduce healthcare costs and reduce adverse events for patients by avoiding unnecessary resections of non-neoplastic polyps. In this article, we expand on the most relevant studies in this field and discuss limitations and future directions that will determine fulfilment of the potential of CNN in the optical diagnosis of colorectal polyps.

Citation: Kader R, Hadjinicolaou AV, Georgiades F, Stoyanov D, Lovat LB. Optical diagnosis of colorectal polyps using convolutional neural networks. World J Gastroenterol 2021; 27(35): 5908-5918

URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5908.htm DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5908

# INTRODUCTION

Colorectal cancer (CRC) is the third most commonly diagnosed cancer worldwide<sup>[1]</sup> and thus, a significant burden on global healthcare systems. Most CRCs develop in a relatively predictable, stepwise sequence from mutation-accumulating neoplastic polyps, such as adenomas and sessile serrated lesions (SSL)[2]. Current evidencebased societal guidelines unequivocally accept colonoscopy to be the gold standard tool for screening of CRC[3]. Colonoscopy offers the opportunity to both detect and resect neoplastic polyps<sup>[4]</sup> and its implementation, especially as part of bowel cancer screening programs, has been linked to a significant reduction in the incidence of the CRC and CRC-related mortality<sup>[5]</sup>.

Over 90% of polyps detected at colonoscopy are either small (6-9 mm) or diminutive  $(\leq 5 \text{ mm})$ , entities that are thought to harbour a very low risk for developing into CRC [6]. Furthermore, almost half of these polyps are non-neoplastic in nature; and frequently hyperplastic<sup>[7]</sup>. Accurate differentiation of neoplastic from non-neoplastic polyps can prevent the unnecessary resection of the latter, avoiding an intervention which is not cost-effective and which carries risks of significant morbidity[8].

Recent years have seen significant research activity in the use of artificial intelligence (AI), particularly convolutional neural networks (CNN), to optically diagnose colorectal polyps. The field is gaining increasing momentum. The aim of this review article is to summarise and critically appraise the available medical literature related to advances in CNN for optical diagnosis of colorectal polyps and highlight the field's current limitations and future directions.

# **OPTICAL DIAGNOSIS**

The term "optical diagnosis" refers to the use of advanced imaging techniques for realtime, *in-vivo* polyp characterisation and evaluation to guide therapeutic decisions[9]. Accurate optical diagnosis of diminutive polyps would enable identification of hyperplastic polyps in the rectosigmoid region, where they are commonly found, and allow the endoscopist to confidently take a "diagnose and leave" approach instead of resecting the lesion. Equally, for diminutive adenomas, accurate optical diagnosis would prompt the endoscopist to remove the lesion on the spot and discard the specimen without the need for histological assessment ("resect and discard" strategy) [9]

The American Society of Gastrointestinal Endoscopy established the Preservation and Incorporation of Valuable endoscopic Innovations (PIVI) to provide thresholds that are required of endoscopic technology in order to implement a "resect and discard" (PIVI 1) and "diagnose and leave" (PIVI 2) strategy [9]. PIVI 1 requires  $\geq 90\%$ 



concordance in post-polypectomy surveillance intervals when comparing the combination of optical diagnosis for diminutive adenomas with histopathology assessment of all other polyps against decisions based solely on histopathology evaluation of all identified polyps[10]. PIVI 2 requires a technology to achieve a negative predictive value (NPV) of  $\geq 90\%$  for diminutive adenomatous polyps in the rectosigmoid region[9].

There has been extensive research in image enhanced endoscopy (IEE), such as narrow band imaging (NBI), to assist endoscopists in optical diagnosis to characterise diminutive polyps[11-13]. Using IEE, expert endoscopists in academic centres have consistently demonstrated an optical diagnosis accuracy that exceeds PIVI thresholds [14-16], however, studies have often found community and non-expert endoscopists to fall short of these minimal thresholds<sup>[17]</sup>. An example is the multi-centre DISCARD-2 study which evaluated the optical diagnosis accuracy of 28 community endoscopists using NBI. Disappointingly, the endoscopists' optical diagnosis derived colonoscopy surveillance intervals only matched 68% of the histopathology derived intervals[18]. Although widely available, technologies for optical diagnosis has not been incorporated into routine clinical practice with one of the main barriers being the interoperator variability amongst endoscopists[19].

#### WHAT IS A CONVOLUTIONAL NEURAL NETWORK?

AI is the ability of computers to perform tasks that traditionally require human intelligence (Figure 1)[20]. Machine learning (ML) is a subset of AI, whereby computers continuously learn from data without explicit human programming[21]. This can be used to predicate a polyp's histology. ML models can be trained using unsupervised or supervised techniques. Unsupervised learning is when the input and output data are not paired. Supervised ML is more labour intensive as it requires paired input and output data for training. An example of a supervised ML model for optical diagnosis is to annotate a bounding box around a polyp (input data), commonly referred to as a region of interest, and label it with the histology of the polyp (output data). The model automatically learns to extract features that allow it to differentiate polyp subtypes and output a diagnosis based on the histology classification system it was trained with but the annotation process is time consuming for the clinician.

Deep learning is a subset of ML, whereby algorithms use multiple layers within a neural network<sup>[22]</sup>, mimicking the human brain, to extract high level features from input data. CNNs are the most commonly used network in the application of deep learning to optically diagnose polyps. They provide an objective output, bypassing the human inter and intra-operator variability, and can develop classification algorithms without exhaustive effort as they do not require human-crafted feature extraction or extensive pre-processing of data[23].

Building a CNN model typically involves three separate datasets; a training set, a validation set and a test set[24]. The training set is used to develop the model so that it predicts a label (e.g., adenomatous or hyperplastic polyp for polyp characterisation) based on features extracted from the endoscopic image by the algorithm itself. The validation set is used to avoid over-fitting into the training dataset through fine tuning of the hyperparameters of the model. Finally, the testing set is used as an independent dataset to evaluate the generalisability of the CNN. With smaller datasets, crossvalidation can be used to assess the model's robustness. In cross-validation, the data is split into equal parts (e.g., 4 parts), with one part held out as a validation dataset. This process is repeated multiple times, with the results of each split eventually pooled together to decide how robust the model is [24]. CNNs evaluated using crossvalidation should still be assessed against an independent test set to examine their generalisability[24].

#### CONVOLUTIONAL NEURAL NETWORKS AND OPTICAL DIAGNOSIS

It is only in the last few years that the use of CNNs in optical diagnosis of colorectal polyps has been extensively investigated, with various studies emerging (Table 1). Many of these studies have in fact demonstrated the capability of CNNs to surpass the PIVI 2 threshold in order to support a "leave in" strategy for rectosigmoid hyperplastic polyps (Table 2). This was first demonstrated by Chen et al[25], who used a single centre, retrospective, still image dataset of 2157 polyps to train a CNN and reported a



Ref.	Study design (training/testing)	Multi- centre study	Dataset	lmage quality	Classification system	Lesion number (training/testing)	SSL excluded	Endoscopic processor	Image modality (training)	Real-time capability
Komeda <i>et al</i> [37]	Retrospective	Single	Video	Not specified	Adenoma/non-adenoma	Not specified/10	No	Not specified	WLI, NBI, chromoendoscopy	Not specified
Chen et al[25]	Retrospective/prospective	Single	Still	HQ	Hyperplastic/neoplastic	2157/284	Yes	Olympus 260 + 290	Magnified NBI	Real-time (approximately 450 ms)
Byrne <i>et al</i> [23]	Retrospective/prospective	Single	Video	All images	NICE Type 1/NICE Type 2	220/125	Yes	Olympus 190	NBI-NF	Real-time ( approximately 50 ms)
Zachariah et al <mark>[26</mark> ]	Prospective	Two	Still	Adequate and HQ	Adenomatous/serrated polyp	5278/634	No	Olympus 190 (90%), 180 (7%), Pentax i10(3%)	WLI, NBI, i-SCAN	Real-time ( approximately 13 ms)
Ozawa et al [ <mark>38</mark> ]	Retrospective/prospective	Single	Still	HQ	Hyperplastic/adenomatous/SSL/CRC/other	WLI: 17566/783 NBI: 2865/290	No	Olympus 260 + 290	WLI, NBI	Real-time (approximately 20 ms)
Jin <i>et al</i> [ <mark>31</mark> ]	Retrospective/prospective	Single	Still	HQ	Hyperplastic/adenomatous	2150/300	Yes	Olympus 290	NBI-NF	Real-time (approximately 10 ms)
Song et al[ <mark>39</mark> ]	Retrospective/prospective	Single	Still	HQ	Serrated polyp/benign adenoma/MSM/DSMC	624/545	No	Olympus 290	NBI-NF	Real-time ( approximately 20- 40 ms)
Rodriguez- Diaz et al[ <mark>28</mark> ]	Retrospective/prospective	Two	Still	Not specified	Neoplastic (adenomas, CRC)/non-neoplastic (hyperplastic, normal)	607/280	Training: Yes Testing: No	Olympus 190	NBI-NF, NBI (digital magnification)	Real-time (approximately 100 ms)
van der Zander <i>et al</i> [ <mark>27</mark> ]	Retrospective/prospective	Not specified	Still	HQ	Benign (hyperplastic)/pre-malignant (adenomatous, SSL, T1 CRC)	398/60	No	Fujifilm, Pentax	WLI, BLI, i-SCAN	Real-time (approximately 14.8 ms)

Table 1 Summary of the studies on convolutional neural network algorithms for the optical diagnosis of colorectal polyps

SSL: Sessile serrated lesion; WLI: White light imaging; BLI: Blue light imaging; NBI: Narrow band imaging; NBI-NF: Narrow band imaging–near focus; NICE: NBI International Colorectal Endoscopic; HQ: High-quality; CRC: Colorectal cancer; MSMC: Mucosal or superficial submucosal cancer; DSMC: Deep submucosal cancer.

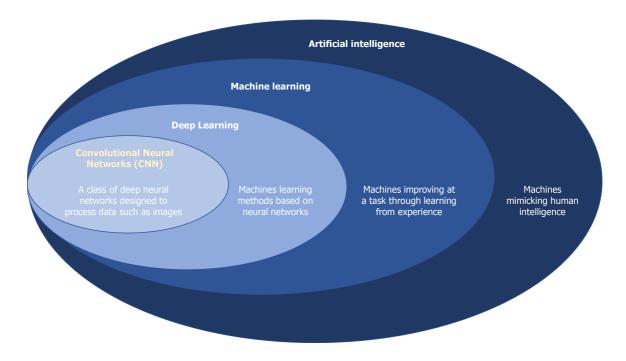
sensitivity for identifying adenomas of 96.3%, specificity 78.1%, and NPV of 91.5% when evaluating a test set of 284 colonic and rectal diminutive adenomatous and hyperplastic polyps. Using colonic diminutive polyps is a common strategy to assess against PIVI 2 due to difficulties in obtaining large datasets of diminutive rectosigmoid polyps. An important limitation of this study is that it used magnified narrow-band

Table 2 Summary of the per-polyp results of studies on convolutional neural network algorithms for the optical diagnosis of colorectal polyps (cross-validation results not included)

Ref.	Image Modality (testing)	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)	Accuracy for neoplasia (%)	PIVI 1 achieved (%)	PIVI 2 achieved (%)
Komeda et al[37]	Not specified	-	-	-	-	70	-	-
Chen et al[25]	Magnified NBI	96.3	78.1	89.6	91.5	90.1	-	Yes (91.5)
Byrne <i>et al</i> [23]	NBI-NF	98	83	90	97	94	-	Yes (97)
Zachariah et al	NBI	-	-	-	96.5	93.1	Yes (98.3)	Yes (96.5)
[26]	WLI	-	-	-	88.9	92.8	Yes (90.8)	No (88.9)
Ozawa et al[38] <sup>1</sup>	NBI	97	-	84	88	-	-	-
	WLI	98	-	85	88	-	-	-
Jin et al	NBI-NF	83.3	91.7	93.3	78.6	86.7	-	-
Song et al[39]	NBI-NF (test set 1)	84.1	74	88.3	67.7	-	-	-
	NBI-NF (test set 2)	88.5	72.1	88.6	84.7	-	-	-
Rodriguez-Diaz et al[ <mark>28</mark> ]	NBI-NF (90%) + NBI (10%)	95	88	-	93	-	Yes (94 (20/90 LC))	Yes (98 (6/68 LC))
van der Zander <i>et</i> al[ <mark>27</mark> ]	WLI + BLI	95.6	93.3	97.7	87.5	95.0	-	No (87.5)

<sup>1</sup>Per frame analysis reported only.

WLI: White light imaging; BLI: Blue light imaging; NBI: Narrow band imaging; NBI-NF: Narrow band imaging-near focus; PIVI: Preservation and Incorporation of Valuable endoscopic Innovations; PPV: Positive predictor value; NPV: Negative predictor value; LC: Low-confidence.



#### Figure 1 The relationship between convolutional neural networks, deep learning, machine learning and artificial intelligence.

imaging (NBI) data. This recently developed modality is not yet readily available in most endoscopy departments, although it will become more widely used with time.

Byrne et al<sup>[23]</sup> further advanced the field by training a CNN with NBI-near focus (NBI-NF) which is more commonly used in Europe and North America. It was trained with 220 polyp positive videos and when tested against 125 diminutive polyps which were collected prospectively, the model diagnosed 106 polyps with high confidence, achieving a sensitivity for identifying NBI International Colorectal Endoscopic (NICE)



type 1 polyps of 98%, specificity 83% and NPV of 97%. A novelty worth highlighting in this study was the use of images derived from videos, an approach that reduces selection bias compared to retrospective still images as endoscopists usually capture high quality polyp views that are free from motion blur and surface artifact. An additional advantage of this CNN is that it simplified the clinical workflow as it automatically diagnoses polyps without requiring a still image of the polyp to be captured. Limitations of the study are that SSLs, normal tissue and lymphoid aggregates were excluded from the final analysis and the videos used to train and test the CNN were captured from colonoscopies performed by a single expert endoscopist and hence, potentially less generalisable to novice users.

The most commonly used imaging modalities amongst community endoscopists are white light imaging (WLI) and NBI without magnification. Using a large retrospective still image training set of 5278 polyps and tested against 634 polyps, Zachariah et al[26] 's CNN fell short of PIVI 2 in WLI (NPV of 88.9% and accuracy 92.8%) but achieved the threshold in NBI without magnification (NPV of 90.8% and accuracy 93.1%). This study advanced the field as it demonstrated the capabilities of CNNs to optically diagnose polyps in standard NBI modality and also to differentiate adenomas from serrated polyps through the inclusion of SSLs in its dataset.

Whilst the majority of CNNs have been trained and tested using Olympus data, studies are emerging using data from other manufacturers. van der Zander et al[27] recently developed a CNN using Fujifilm data in high definition white light (HDWL) and blue light imaging (BLI). The CNN was more efficacious when it used a unique multimodal imaging approach where it combined both HDWL and BLI images of the same polyp in its decision process compared to a single imaging modality. When evaluated against 60 prospectively collected diminutive polyps, it did not reach the PIVI 2 threshold with a NPV of 87.5% but did achieve an optical diagnosis accuracy of 95% (sensitivity for identifying pre-malignant polyps 95.6% and specificity 93.3%) and demonstrated superiority to both expert and novice endoscopists in human benchmark testing.

In comparison to PIVI 2, there are fewer studies evaluating the performance of CNNs against PIVI 1. The CNN presented in Zachariah et al<sup>[26]</sup> reached PIVI 1 thresholds in both WLI and NBI with normal magnification, achieving concordance with histology-based colonoscopy surveillance intervals in 90.9% and 98.3% of patients, for each respective modality. Rodrigues-Diaz et al[28] used a single centre retrospective still image dataset to train a CNN with 607 polyps and tested against 90 diminutive polyps where it achieved a high confidence diagnosis in 78% of cases, with a 94% agreement with histology-based colonoscopy surveillance intervals. Tested against 68 rectosigmoid polyps, the model diagnosed 88% of polyps with high confidence, achieving PIVI 2 thresholds with a NPV of 97%.

There is also potential to expand the use of optical diagnosis CNNs outside of the "resect and discard" and "leave in strategy". A dilemma that can complicate issuing post-polypectomy surveillance intervals is discrepancies between endoscopic and histological diagnosis and classification of polyps with tissue fragmentation in the specimen retrieval process playing an important role. Shahidi et al[29]'s proof of concept study used a CNN to resolve discrepancies in polyps ≤ 3 mm in size. Tested against 900 polyps that were  $\leq 3$  mm and optically diagnosed as adenomatous by an expert endoscopist, the CNN diagnosed the adenomas with high confidence in 644 polyps, with 256 polyps deemed to be of sub-optimal imaging quality. However, of these high confidence diagnoses, the pathologists diagnosed 15.4% as normal mucosa, 13.2% as hyperplastic polyp and 0.3% as SSL. In this context, a CNN could help to mitigate against the risk of under-surveillance.

Whilst CNN's diagnostic accuracy excels in many studies, without real-time capabilities, they would have no clinical utility. Prior to the era of deep learning, computer aided diagnosis algorithms lacked real-time capability, but most CNNs do not share this problem and often process data at a rate that exceeds the 25 frames per second that is generated in a video recording of a colonoscopy procedure. Given the excellent performance in ex-vivo studies and the real-time capabilities displayed by CNNs, the future appears promising for their integration in colonoscopy.

#### TRANSPARENCY OF CONVOLUTIONAL NEURAL NETWORKS

The complexity of CNN models' decision process is often referred to as a "black box" and represents an important barrier to its acceptance by both clinicians and patients [30]. Opening the "black box" to display the raw features which informed the CNN's



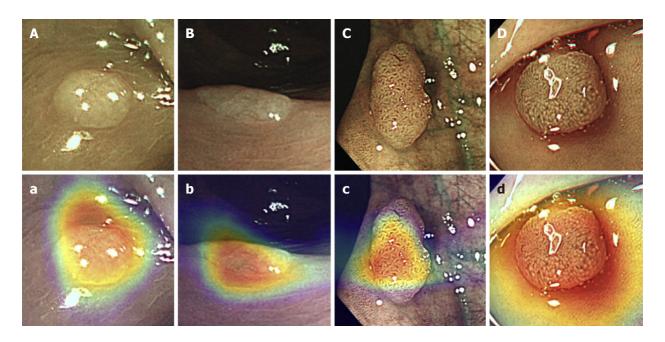


Figure 2 Illustration of coloured heatmaps, overlaid to the polyp, which demonstrates the regions that most likely contributed to the convolutional neural networks's diagnosis. A, B, C, D: Original narrow band imaging (NBI) of polyps; a, b, c, d: Coloured heatmap overlaid on the NBI image; Red: Higher probability that this region informed the convolutional neural networks (CNN)'s diagnosis; Blue: Lower probability that this region informed the CNN's diagnosis. Images adapted and modified with permission from the publisher[31]. Citation: Jin EH, Lee D, Bae JH, Kang HY, Kwak MS, Seo JY, Yang JI, Yang SY, Lim SH, Yim JY, Lim JH, Chung GE, Chung SJ, Choi JM, Han YM, Kang SJ, Lee J, Chan Kim H, Kim JS. Improved Accuracy in Optical Diagnosis of Colorectal Polyps Using Convolutional Neural Networks. Gastroenterology 2020; 158(8): 2169-2179. Copyright© The Authors 2020. Published by Elsevier.

decision is important for transparency especially from a safety standpoint[28]. Transparency can help identify biases within the neural network and aid root-cause analyses in cases of patient harm, for example, if a neoplastic polyp that subsequently develops into a CRC is originally misdiagnosed as non-neoplastic by the CNN model.

For polyp characterisation, important steps have been taken to open the black box. Jin et al[31] developed a CNN that generated a coloured heat map, overlaid to the polyp, to help the endoscopist comprehend the specific aspects of the image that contributed to the CNN's prediction (Figure 2). This could help the endoscopist to decide which information is relevant and which decisions are truly based on appropriate image analysis. If, for example, the heatmap is overlaid to normal mucosa, then the endoscopist would quickly be able to appreciate this and disregard the CNN's diagnosis.

More recently, in order to further enhance CNN transparency, Rodriguez-Diaz et al [28] developed a colour coded segmentation model (Figure 3). In this model, the CNN divides the polyp into distinct segments to allow the endoscopist to identify the specific regions within the image that is informing the CNN's decision. The CNN predicts the histology of each subregion of the segmented polyp, with high confidence neoplastic diagnoses coloured in red, high confidence non-neoplastic in green, and low confidence/indeterminate diagnoses in yellow, with the final predication resulting from an aggregate of all the analysed regions. The end result is a detailed spatial colour coded histology map of the polyp surface, which the endoscopist can visualise and incorporate into their decision process<sup>[28]</sup>, enhancing the interpretability of this CNN model in comparison to others. However, an important limitation to this advanced CNN is that it currently lacks the ability to operate at a video rate.

Further research in the interpretability of CNN models is required to improve its acceptance[32] and accelerate its translation to clinical practise.

### LIMITATIONS AND FUTURE DIRECTIONS

Despite the promise shown by CNNs this far, it is crucial to recognise that there are various limitations that need to be overcome before they can become part of the endoscopic clinical workflow. The most significant limitations are the reliance on retrospective datasets[33], which are inherently subject to selection bias, and the lack of prospective studies and randomised controlled trials[34]. Most studies train and test



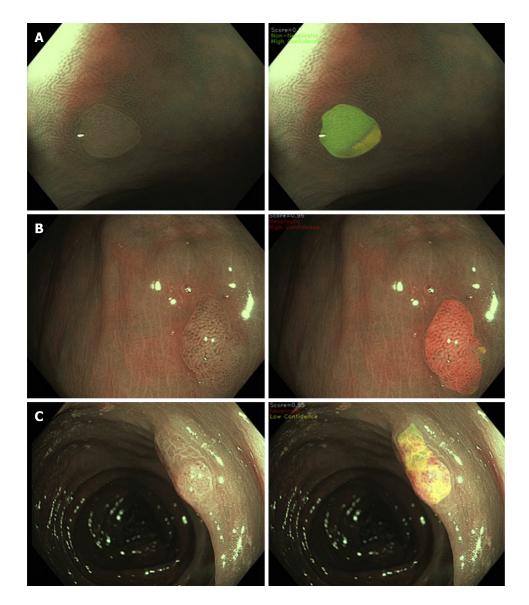


Figure 3 Spatial colour coded histology map which allows the user to visualise the sub-regions of the polyp surface that contributed to the convolutional neural networks's decision process. A: Hyperplastic polyps; B: Adenomatous polyps; C: Sessile serrated lesions; Red: High-confidence neoplastic diagnosis; Green: High-confidence non-neoplastic diagnosis; Yellow: Indeterminate or low-confidence diagnosis. Adapted from Ref. [28]. Citation: Rodriguez-Diaz E, Baffy G, Lo WK, Mashimo H, Vidyarthi G, Mohapatra SS, Singh SK. Real-time artificial intelligence-based histologic classification of colorectal polyps with augmented visualization. *Gastrointest Endosc* 2021; 93: 662-670. Copyright© The Authors 2021. Published by Elsevier.

CNNs using high quality images of polyps, free from "noise" such as motion blur and polyp surface artifact (*e.g.*, mucus, stool or blood). The extent to which CNNs preclinical results are reproducible in the real-world setting, where 'noise' is frequently encountered, remains to be seen.

To the best of our knowledge, there have been no prospective randomised controlled clinical trials evaluating optical diagnosis CNN *in-vivo*. This is partly due to clinical trials being time consuming and expensive, and an alternative pragmatic approach could be the use of a benchmark test in the form a publicly available external dataset to compare different CNN models[35]. No such datasets currently exist for polyp characterisation and therefore the generalisability of CNN models remains poorly understood. Generalisability refers to the CNN performance with different endoscope models and clinical settings from the site that the data was generated to train the CNN. To date, only one study[36] has evaluated generalisability, and this was limited to a small testing set of 69 polyp images from two population cohorts (Australian and Japanese) using two separate endoscope manufactures (Olympus and Fujifilm). Despite the small test-set, this study highlighted the concerns of generalisability as the operator area under the curve fell from 94.3% for the internal set, to 84.5% and 90.3% for the external testing sets (NBI and BLI respectively).

Beishideng® WJG https://www.wjgnet.com

Another important limitation is that studies often exclude polyps that are not adenomas or hyperplastic polyps, restricting the possible classification outputs of CNNs. This, in turn, limits their clinical utility as polyps such as SSL and inflammatory polyps would be misclassified due to limitations in the initial training phase of the CNNs when the categorisation system is established.

Research in this field is likely to continue to expand and future directions to consider include: (1) Guidelines to identify the role of CNNs in the clinical workflow, specifically, whether it is a second reader, a concurrent reader or a provider of an independent diagnosis[30]; (2) Prospective multi-centre randomised clinical trials; (3) Publicly available external datasets for benchmark testing and evaluation of the generalisability of CNN models in different clinical settings and population cohorts; and (4) Acquiring datasets inclusive of all polyp sub-types to advance CNN classification systems.

## CONCLUSION

In summary, this is an exciting time for the endoscopy community. CNNs diagnostic performance has excelled in ex-vivo studies and in human benchmarking testing. CNNs are likely to be a key adjunct in optically diagnosing polyps and have renewed optimism that implementation of a "resect and discard" and "leave in" strategy is feasible due to the potential to alleviate the inter-operator variability amongst endoscopists. This would bring significant financial benefits to healthcare systems, avoid unnecessary polypectomies of non-neoplastic polyps and improve the efficiency of colonoscopy. However, prospective multi-centre randomised controlled trials and publicly available datasets for benchmark testing are required to further evaluate the efficacy and generalisability of CNNs. Furthermore, with these models now emerging in endoscopy units, it's imperative that guidelines are developed to establish their role in the clinical workflow.

# REFERENCES

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- Song M, Emilsson L, Bozorg SR, Nguyen LH, Joshi AD, Staller K, Nayor J, Chan AT, Ludvigsson JF. Risk of colorectal cancer incidence and mortality after polypectomy: a Swedish record-linkage study. Lancet Gastroenterol Hepatol 2020; 5: 537-547 [PMID: 32192628 DOI: 10.1016/S2468-1253(20)30009-1]
- US Preventive Services Task Force. Bibbins-Domingo K, Grossman DC, Curry SJ, Davidson KW, 3 Epling JW Jr, García FAR, Gillman MW, Harper DM, Kemper AR, Krist AH, Kurth AE, Landefeld CS, Mangione CM, Owens DK, Phillips WR, Phipps MG, Pignone MP, Siu AL. Screening for Colorectal Cancer: US Preventive Services Task Force Recommendation Statement. JAMA 2016; 315: 2564-2575 [PMID: 27304597 DOI: 10.1001/jama.2016.5989]
- Rutter MD, East J, Rees CJ, Cripps N, Docherty J, Dolwani S, Kaye PV, Monahan KJ, Novelli MR, Plumb A, Saunders BP, Thomas-Gibson S, Tolan DJM, Whyte S, Bonnington S, Scope A, Wong R, Hibbert B, Marsh J, Moores B, Cross A, Sharp L. British Society of Gastroenterology/Association of Coloproctology of Great Britain and Ireland/Public Health England post-polypectomy and postcolorectal cancer resection surveillance guidelines. Gut 2020; 69: 201-223 [PMID: 31776230 DOI: 10.1136/gutjnl-2019-319858]
- Zauber AG, Winawer SJ, O'Brien MJ, Lansdorp-Vogelaar I, van Ballegooijen M, Hankey BF, Shi W, Bond JH, Schapiro M, Panish JF, Stewart ET, Waye JD. Colonoscopic polypectomy and longterm prevention of colorectal-cancer deaths. N Engl J Med 2012; 366: 687-696 [PMID: 22356322 DOI: 10.1056/NEJMoa11003701
- 6 Lieberman D, Moravec M, Holub J, Michaels L, Eisen G. Polyp size and advanced histology in patients undergoing colonoscopy screening: implications for CT colonography. Gastroenterology 2008; 135: 1100-1105 [PMID: 18691580 DOI: 10.1053/j.gastro.2008.06.083]
- 7 Rex DK, Overhiser AJ, Chen SC, Cummings OW, Ulbright TM. Estimation of impact of American College of Radiology recommendations on CT colonography reporting for resection of high-risk adenoma findings. Am J Gastroenterol 2009; 104: 149-153 [PMID: 19098863 DOI: 10.1038/ajg.2008.35]
- El Hajjar A, Rey JF. Artificial intelligence in gastrointestinal endoscopy: general overview. Chin 8 *Med J (Engl)* 2020; **133**: 326-334 [PMID: 31929362 DOI: 10.1097/CM9.00000000000623]
- 9 ASGE Technology Committee. Abu Dayyeh BK, Thosani N, Konda V, Wallace MB, Rex DK, Chauhan SS, Hwang JH, Komanduri S, Manfredi M, Maple JT, Murad FM, Siddiqui UD, Banerjee S.



ASGE Technology Committee systematic review and meta-analysis assessing the ASGE PIVI thresholds for adopting real-time endoscopic assessment of the histology of diminutive colorectal polyps. Gastrointest Endosc 2015; 81: 502.e1-502.e16 [PMID: 25597420 DOI: 10.1016/j.gie.2014.12.022]

- 10 Rex DK. Can we do resect and discard with AI-assisted colon polyp 'optical biopsy'? Tech Gastrointest Endosc 2019; 150638 [DOI: 10.1016/j.tgie.2019.150638]
- Ignjatovic A, East JE, Suzuki N, Vance M, Guenther T, Saunders BP. Optical diagnosis of small 11 colorectal polyps at routine colonoscopy (Detect InSpect ChAracterise Resect and Discard; DISCARD trial): a prospective cohort study. Lancet Oncol 2009; 10: 1171-1178 [PMID: 19910250 DOI: 10.1016/S1470-2045(09)70329-8]
- 12 Ignjatovic A, Thomas-Gibson S, East JE, Haycock A, Bassett P, Bhandari P, Man R, Suzuki N, Saunders BP. Development and validation of a training module on the use of narrow-band imaging in differentiation of small adenomas from hyperplastic colorectal polyps. Gastrointest Endosc 2011; 73: 128-133 [PMID: 21184878 DOI: 10.1016/j.gie.2010.09.021]
- 13 Rastogi A, Keighley J, Singh V, Callahan P, Bansal A, Wani S, Sharma P. High accuracy of narrow band imaging without magnification for the real-time characterization of polyp histology and its comparison with high-definition white light colonoscopy: a prospective study. Am J Gastroenterol 2009; 104: 2422-2430 [PMID: 19584829 DOI: 10.1038/ajg.2009.403]
- Repici A, Hassan C, Radaelli F, Occhipinti P, De Angelis C, Romeo F, Paggi S, Saettone S, Cisarò F, 14 Spaander M, Sharma P, Kuipers EJ. Accuracy of narrow-band imaging in predicting colonoscopy surveillance intervals and histology of distal diminutive polyps: results from a multicenter, prospective trial. Gastrointest Endosc 2013; 78: 106-114 [PMID: 23582472 DOI: 10.1016/j.gie.2013.01.035]
- 15 Wallace MB, Crook JE, Coe S, Ussui V, Staggs E, Almansa C, Patel MK, Bouras E, Cangemi J, Keaveny A, Picco M, Riegert-Johnson D. Accuracy of in vivo colorectal polyp discrimination by using dual-focus high-definition narrow-band imaging colonoscopy. Gastrointest Endosc 2014; 80: 1072-1087 [PMID: 24973171 DOI: 10.1016/j.gie.2014.05.305]
- Singh R, Jayanna M, Navadgi S, Ruszkiewicz A, Saito Y, Uedo N. Narrow-band imaging with dual 16 focus magnification in differentiating colorectal neoplasia. Dig Endosc 2013; 25 Suppl 2: 16-20 [PMID: 23617643 DOI: 10.1111/den.12075]
- 17 Rees CJ, Rajasekhar PT, Wilson A, Close H, Rutter MD, Saunders BP, East JE, Maier R, Moorghen M, Muhammad U, Hancock H, Jayaprakash A, MacDonald C, Ramadas A, Dhar A, Mason JM. Narrow band imaging optical diagnosis of small colorectal polyps in routine clinical practice: the Detect Inspect Characterise Resect and Discard 2 (DISCARD 2) study. Gut 2017; 66: 887-895 [PMID: 27196576 DOI: 10.1136/gutjnl-2015-310584]
- Kuiper T, Marsman WA, Jansen JM, van Soest EJ, Haan YC, Bakker GJ, Fockens P, Dekker E. 18 Accuracy for optical diagnosis of small colorectal polyps in nonacademic settings. Clin Gastroenterol Hepatol 2012; 10: 1016-20; quiz e79 [PMID: 22609999 DOI: 10.1016/j.cgh.2012.05.004]
- 19 Ladabaum U, Fioritto A, Mitani A, Desai M, Kim JP, Rex DK, Imperiale T, Gunaratnam N. Realtime optical biopsy of colon polyps with narrow band imaging in community practice does not yet meet key thresholds for clinical decisions. Gastroenterology 2013; 144: 81-91 [PMID: 23041328 DOI: 10.1053/j.gastro.2012.09.054]
- Sharma P, Pante A, Gross SA. Artificial intelligence in endoscopy. Gastrointest Endosc 2020; 91: 20 925-931 [PMID: 31874161 DOI: 10.1016/j.gie.2019.12.018]
- Alagappan M, Brown JRG, Mori Y, Berzin TM. Artificial intelligence in gastrointestinal endoscopy: 21 The future is almost here. World J Gastrointest Endosc 2018; 10: 239-249 [PMID: 30364792 DOI: 10.4253/wjge.v10.i10.239]
- 22 Vakli P, Deák-Meszlényi RJ, Hermann P, Vidnyánszky Z. Transfer learning improves resting-state functional connectivity pattern analysis using convolutional neural networks. Gigascience 2018; 7 [PMID: 30395218 DOI: 10.1093/gigascience/giy130]
- 23 Byrne MF, Chapados N, Soudan F, Oertel C, Linares Pérez M, Kelly R, Iqbal N, Chandelier F, Rex DK. Real-time differentiation of adenomatous and hyperplastic diminutive colorectal polyps during analysis of unaltered videos of standard colonoscopy using a deep learning model. Gut 2019; 68: 94-100 [PMID: 29066576 DOI: 10.1136/gutjnl-2017-314547]
- van der Sommen F, de Groof J, Struyvenberg M, van der Putten J, Boers T, Fockens K, Schoon EJ, 24 Curvers W, de With P, Mori Y, Byrne M, Bergman JJGHM. Machine learning in GI endoscopy: practical guidance in how to interpret a novel field. Gut 2020; 69: 2035-2045 [PMID: 32393540 DOI: 10.1136/gutjnl-2019-320466]
- 25 Chen PJ, Lin MC, Lai MJ, Lin JC, Lu HH, Tseng VS. Accurate Classification of Diminutive Colorectal Polyps Using Computer-Aided Analysis. Gastroenterology 2018; 154: 568-575 [PMID: 29042219 DOI: 10.1053/j.gastro.2017.10.010]
- Zachariah R, Samarasena J, Luba D, Duh E, Dao T, Requa J, Ninh A, Karnes W. Prediction of Polyp 26 Pathology Using Convolutional Neural Networks Achieves "Resect and Discard" Thresholds. Am J Gastroenterol 2020; 115: 138-144 [PMID: 31651444 DOI: 10.14309/ajg.00000000000429]
- 27 van der Zander QEW, Schreuder RM, Fonollà R, Scheeve T, van der Sommen F, Winkens B, Aepli P, Hayee B, Pischel AB, Stefanovic M, Subramaniam S, Bhandari P, de With PHN, Masclee AAM, Schoon EJ. Optical diagnosis of colorectal polyp images using a newly developed computer-aided diagnosis system (CADx) compared with intuitive optical diagnosis. Endoscopy 2020 [PMID: 33368056 DOI: 10.1055/a-1343-1597]



- Rodriguez-Diaz E, Baffy G, Lo WK, Mashimo H, Vidyarthi G, Mohapatra SS, Singh SK. Real-time 28 artificial intelligence-based histologic classification of colorectal polyps with augmented visualization. Gastrointest Endosc 2021; 93: 662-670 [PMID: 32949567 DOI: 10.1016/j.gie.2020.09.018]
- Shahidi N, Rex DK, Kaltenbach T, Rastogi A, Ghalehjegh SH, Byrne MF. Use of Endoscopic 29 Impression, Artificial Intelligence, and Pathologist Interpretation to Resolve Discrepancies Between Endoscopy and Pathology Analyses of Diminutive Colorectal Polyps. Gastroenterology 2020; 158: 783-785.e1 [PMID: 31863741 DOI: 10.1053/j.gastro.2019.10.024]
- 30 Ahmad OF, Mori Y, Misawa M, Kudo SE, Anderson JT, Bernal J, Berzin TM, Bisschops R, Byrne MF, Chen PJ, East JE, Eelbode T, Elson DS, Gurudu SR, Histace A, Karnes WE, Repici A, Singh R, Valdastri P, Wallace MB, Wang P, Stoyanov D, Lovat LB. Establishing key research questions for the implementation of artificial intelligence in colonoscopy: a modified Delphi method. Endoscopy 2021; 53: 893-901 [PMID: 33167043 DOI: 10.1055/a-1306-7590]
- Jin EH, Lee D, Bae JH, Kang HY, Kwak MS, Seo JY, Yang JI, Yang SY, Lim SH, Yim JY, Lim JH, Chung GE, Chung SJ, Choi JM, Han YM, Kang SJ, Lee J, Chan Kim H, Kim JS. Improved Accuracy in Optical Diagnosis of Colorectal Polyps Using Convolutional Neural Networks with Visual Explanations. Gastroenterology 2020; 158: 2169-2179.e8 [PMID: 32119927 DOI: 10.1053/j.gastro.2020.02.036]
- Yang YJ, Bang CS. Application of artificial intelligence in gastroenterology. World J Gastroenterol 32 2019; 25: 1666-1683 [PMID: 31011253 DOI: 10.3748/wjg.v25.i14.1666]
- Namikawa K, Hirasawa T, Yoshio T, Fujisaki J, Ozawa T, Ishihara S, Aoki T, Yamada A, Koike K, 33 Suzuki H, Tada T. Utilizing artificial intelligence in endoscopy: a clinician's guide. Expert Rev Gastroenterol Hepatol 2020; 14: 689-706 [PMID: 32500760 DOI: 10.1080/17474124.2020.1779058]
- 34 Pannala R, Krishnan K, Melson J, Parsi MA, Schulman AR, Sullivan S, Trikudanathan G, Trindade AJ, Watson RR, Maple JT, Lichtenstein DR. Artificial intelligence in gastrointestinal endoscopy. VideoGIE 2020; 5: 598-613 [PMID: 33319126 DOI: 10.1016/j.vgie.2020.08.013]
- Misawa M, Kudo SE, Mori Y, Hotta K, Ohtsuka K, Matsuda T, Saito S, Kudo T, Baba T, Ishida F, 35 Itoh H, Oda M, Mori K. Development of a computer-aided detection system for colonoscopy and a publicly accessible large colonoscopy video database (with video). Gastrointest Endosc 2021; 93: 960-967.e3 [PMID: 32745531 DOI: 10.1016/j.gie.2020.07.060]
- 36 Zorron Cheng Tao Pu L, Maicas G, Tian Y, Yamamura T, Nakamura M, Suzuki H, Singh G, Rana K, Hirooka Y, Burt AD, Fujishiro M, Carneiro G, Singh R. Computer-aided diagnosis for characterization of colorectal lesions: comprehensive software that includes differentiation of serrated lesions. Gastrointest Endosc 2020; 92: 891-899 [PMID: 32145289 DOI: 10.1016/j.gie.2020.02.042]
- Komeda Y, Handa H, Watanabe T, Nomura T, Kitahashi M, Sakurai T, Okamoto A, Minami T, Kono 37 M, Arizumi T, Takenaka M, Hagiwara S, Matsui S, Nishida N, Kashida H, Kudo M. Computer-Aided Diagnosis Based on Convolutional Neural Network System for Colorectal Polyp Classification: Preliminary Experience. Oncology 2017; 93 Suppl 1: 30-34 [PMID: 29258081 DOI: 10.1159/000481227]
- Ozawa T, Ishihara S, Fujishiro M, Kumagai Y, Shichijo S, Tada T. Automated endoscopic detection 38 and classification of colorectal polyps using convolutional neural networks. Therap Adv Gastroenterol 2020; 13: 1756284820910659 [PMID: 32231710 DOI: 10.1177/1756284820910659]
- 39 Song EM, Park B, Ha CA, Hwang SW, Park SH, Yang DH, Ye BD, Myung SJ, Yang SK, Kim N, Byeon JS. Endoscopic diagnosis and treatment planning for colorectal polyps using a deep-learning model. Sci Rep 2020; 10: 30 [PMID: 31913337 DOI: 10.1038/s41598-019-56697-0]



WŨ

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5919-5931

DOI: 10.3748/wjg.v27.i35.5919

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

MINIREVIEWS

# Liver-spleen axis dysfunction in COVID-19

Sara Cococcia, Marco Vincenzo Lenti, Giovanni Santacroce, Giovanna Achilli, Federica Borrelli de Andreis, Antonio Di Sabatino

ORCID number: Sara Cococcia 0000-0002-1507-6513; Marco Vincenzo Lenti 0000-0002-6654-4911: Giovanni Santacroce 0000-0002-0544-0414; Giovanna Achilli 0000-0002-4763-0945; Federica Borrelli de Andreis 0000-0003-2933-8825; Antonio Di Sabatino 0000-0002-0302-8645.

Author contributions: All authors significantly participated in the drafting of the manuscript or in its critical revision for important intellectual content and the approval of the final submitted version; Cococcia S, Borrelli de Andreis F. Santacroce G. and Achilli G wrote the manuscript; Di Sabatino A and Lenti MV supervised the other authors, reviewed the paper, and carried out the final critical revision for important intellectual content.

Conflict-of-interest statement: The authors declare that there are no conflicts of interest.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the

Sara Cococcia, Marco Vincenzo Lenti, Giovanni Santacroce, Giovanna Achilli, Federica Borrelli de Andreis, Antonio Di Sabatino, First Department of Internal Medicine, San Matteo Hospital Foundation, University of Pavia, Pavia 27100, Italy

Sara Cococcia, Department of Gastroenterology, Royal Free Hospital, London NW3 2QG, United Kingdom

Corresponding author: Marco Vincenzo Lenti, MD, Academic Research, Research Assistant Professor, First Department of Internal Medicine, San Matteo Hospital Foundation, University of Pavia, Viale Golgi 19, Pavia 27100, Italy. marco.lenti@unipv.it

# Abstract

Coronavirus disease 2019 (COVID-19), caused by the novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), is an acute infectious disease that spreads mainly through the respiratory route. Besides interstitial pneumonia, a number of other clinical manifestations were noticed in COVID-19 patients. In particular, liver and spleen dysfunctions have been described both as complications of COVID-19 and as potential predisposing factors for severe COVID-19. Liver damage is rather common in COVID-19 patients, and it is most likely multifactorial, caused by the direct insult of SARS-CoV-2 to the liver by the cytokine storm triggered by the virus, by the use of hepatotoxic drugs, and as a consequence of hypoxia. Although generally mild, liver impairment has been found to be associated with a higher rate of intensive care unit admission. A higher mortality rate was reported among chronic liver disease patients. Instead, spleen impairment in patients with COVID-19 has been poorly described. The main anatomical changes are the architectural derangement of the B cell compartment, white pulp atrophy, and reduction or absence of lymphoid follicles, while, from a functional point of view, the IgM memory B cell pool is markedly depleted. The outcome of COVID-19 in asplenic or hyposplenic patients is yet to be defined. In this review, we will summarise the current knowledge regarding the impact of SARS-CoV-2 on the liver and spleen function, as well as the outcome of patients with a pre-existent liver disease or defective spleen function.

Key Words: Asplenia; Chronic liver disease; IgM memory B cell; Liver transplantation; Transaminase

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.



original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: Italy

#### Peer-review report's scientific quality classification

Grade A (Excellent): A Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Received: March 21, 2021 Peer-review started: March 21, 2021 First decision: April 29, 2021 Revised: May 1, 2021 Accepted: August 17, 2021 Article in press: August 17, 2021 Published online: September 21, 2021

P-Reviewer: Guo WZ, Montasser IF S-Editor: Ma YJ L-Editor: Filipodia P-Editor: Wu RR



**Core Tip:** The severe acute respiratory syndrome coronavirus 2 has rapidly spread worldwide, primarily causing interstitial pneumonia, although many other organs can be involved. Here, we will discuss the current knowledge regarding the liver and spleen involvement caused by this infection.

Citation: Cococcia S, Lenti MV, Santacroce G, Achilli G, Borrelli de Andreis F, Di Sabatino A. Liver-spleen axis dysfunction in COVID-19. World J Gastroenterol 2021; 27(35): 5919-5931 URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5919.htm DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5919

# INTRODUCTION

In December 2019, a novel coronavirus-related pneumonia was detected in a Chinese group of patients<sup>[1]</sup>. The pathogen was later named severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)[2], and on 30th January 2020 the World Health Organization publicly declared the outbreak of the new virus-related disease, the socalled coronavirus disease 19 (COVID-19)[3].

The most common clinical manifestations of SARS-CoV-2 infection include fever, dry cough, dyspnoea, fatigue, and myalgia[4,5], but the increasing information in published literature reported a wide spectrum of extrapulmonary symptoms and signs, especially arising from the gastrointestinal tract[6]. Hepatic involvement in COVID-19 patients has been largely documented in several observational studies, highlighting a significant prevalence of liver impairment in hospitalized individuals and a correlation with the severity of the disease [7,8]. COVID-19 implications for individuals with a pre-existent chronic liver disease (CLD) have also been evaluated, and a few studies have focused on the management and prognosis of post-transplant patients[9,10].

Little is known about the splenic involvement in COVID-19 patients. The spleen plays a fundamental role in the immune system modulation, regulating the T and B cell responses to the antigenic targets in the blood, and the tropism of the coronaviruses for the spleen has been documented<sup>[11]</sup>. Although splenic alterations in autoptic specimens have already been shown, and these anatomical changes might contribute to the abnormal immune reaction occurring in COVID-19[12], data on prognosis of COVID-19 individuals with splenic function impairment have been poorly investigated so far.

In this review, we aim at elucidating the pathological role of SARS-CoV-2 in patients with hepatic and splenic involvement, ranging from specific biochemical alterations to any histopathological modifications. Secondly, our purpose is to evaluate the impact of COVID-19 in individuals with a pre-existent diagnosis of hepatic disease or defective spleen function or asplenia.

# MATERIALS AND METHODS

From January to March 2021 we searched on MEDLINE (PubMed) by using the medical subject heading terms "liver", "hepatic", "spleen", "splenectomy", "hyposplenic" matched with "coronavirus", "COVID-19", "SARS-CoV-2" for all articles published since database inception. More than 3000 papers were found with this search strategy, most of which were not strictly related to the subject of this review. Hence, we selected human studies exploring relationships between COVID-19 and liver or spleen function, as well as the outcomes of COVID-19 patients with CLD or spleen hypofunction/asplenia. Given the high number of papers and senior authors (SC, MVL, ADS), after a careful review, we selected the most important or representative ones, summarising current evidence. We also searched for additional papers in the reference lists of review articles, and they were included if deemed appropriate.

#### LIVER IMPAIRMENT IN COVID-19

#### Pathogenesis

Since the most recent studies reporting clinical manifestation of COVID-19 were carried out, the alteration of liver function tests (LFTs) has been reported[4,13-15]. These abnormalities, which still have an unclear clinical significance, have been repeatedly reported in patients suffering from a more severe disease[4,16-18]. The exact cause of liver damage during SARS-CoV-2 infection is partly unknown and most likely multifactorial (Figure 1)[18]. One of the possible explanations has been found in the direct insult of SARS-CoV-2 to the liver through the binding of the virus to the angiotensin-converting enzyme 2 (ACE2) receptor, which represents the main cell entry receptor for the virus [19-21]. ACE2 receptors, which are key players in regulating arterial blood pressure, are expressed in almost any tissue of the human body, especially in the lungs, kidneys, gut, liver and brain. Their polymorphisms may lead to a cardiovascular disease and a stroke<sup>[22]</sup>. However, the ACE2 receptor is highly expressed by cholangiocytes rather than hepatocytes; therefore, the hepatic damage would be channelled through the bile duct dysfunction, which might alter the immune responses and liver regeneration[20]. Nonetheless, it should be noted that alkaline phosphatase (ALP) is not constantly raised in these patients<sup>[23]</sup>.

In addition to the aforementioned mechanisms, the cytokine storm resulting from the excessive immune response triggered by the virus could be another factor leading to liver damage[23,24]. An excessive increase in pro-inflammatory cytokines has been found in a high percentage of critically-ill COVID-19 patients, alongside with a reduction in T cells and an increase in the neutrophilic count. The hypothesis that the lymphocytopenia and C-reactive protein (CRP) levels are independently correlated to the presence of liver damage has been proposed, suggesting a role of the cytokine storm in causing liver dysfunction[25]. This hypothesis has also been proved with regard to organs other than the liver, including heart and kidneys[26], supporting the idea that the cytokine storm may cause shock and tissue damage. Another contributing factor is the use of potentially hepatotoxic drugs, including antibiotics ( e.g., macrolides), antiviral agents especially used during the first wave of the pandemic, corticosteroids, and paracetamol<sup>[23,24]</sup>. Lastly, liver damage can be caused by hypoxia, as a result of severe respiratory failure [23,24,27].

#### Clinical findings

Liver abnormalities are rather common in COVID-19 patients. The proportion of COVID-19 inpatients with an elevated alanine aminotransferase (ALT) has been found to be as high as 36%, and a higher proportion (46%) also had raised aspartate aminotransferase (AST)[18,28]. On the contrary, ALP or gamma-glutamyl transpeptidase (GGT) alterations were reported more rarely[18,29]. Although rather common, in most cases liver injury is mild and it usually manifests in more critically-ill patients[18,30,31]. A mild-to-moderate increase of ALT was reported in 43/87 patients (49.4%), and a higher mortality rate was observed among those with deranged LFT who had developed acute respiratory distress syndrome (ARDS)[29]. Similarly, Richardson et al<sup>[7]</sup>, who enrolled more than 5000 patients with liver involvement, showed that acute hepatic injury, although rare, was associated with higher mortality. Liver involvement was reported in 2700 patients (39%), and 1% of the whole cohort developed acute liver injury. Another study enrolling more than 2000 patients confirmed these findings, reporting acute liver injury in a quarter of the included patients and severe liver injury in only 6.4% of the patients. However, this small proportion had a more complex clinical course, including intensive care admission and intubation need in more than 60% of the cases, renal replacement therapy in a third, and mortality as high as 42% [31]. A low incidence of severe liver injury (9%) was also reported by a German study enrolling 44 patients of which 6 had deranged ALT. Also, the German cohort reported AST to be more commonly deranged than ALT[28].

Although generally mild, liver impairment has been found to be associated to a higher rate of intensive care unit admission[30,32], as well as to a longer hospital stay [33]. Ponziani et al[30] reported liver involvement in 161 out of 515 patients enrolled (31.3%) and no cases of severe acute liver injury. Moreover, although liver involvement led to a higher need for intensive care, no increase in mortality was recorded among those patients. However, conflicting data have been published on the role of liver impairment in increasing mortality in patients without pre-existing liver disease. Medetalibeyoglu et al[32] reported that AST/ALT ratio was a good predictor of mortality (area under the curve [AUC]: 0.713; P = 0.0001) in a cohort of 554 individuals enrolled in Turkey, and that AST and ALT levels were independently



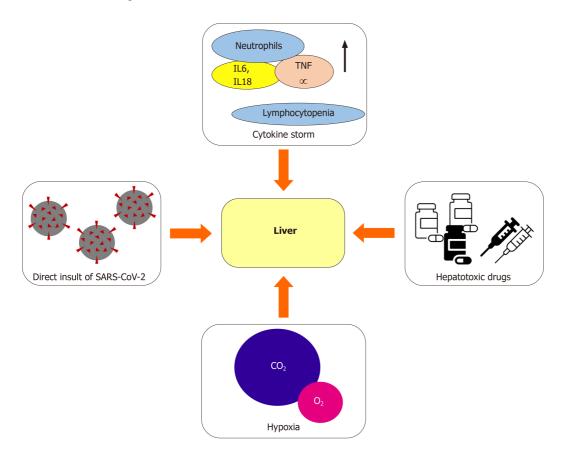


Figure 1 Putative mechanisms of liver damage in coronavirus disease 2019. IL: Interleukin; SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2; TNF: Tumour necrosis factor.

associated with an increased need for intensive care and with mortality (P = 0.001). Table 1 reports the main studies focusing on liver abnormalities in COVID-19 patients.

#### Histological features

Limited data are available about histological liver findings in COVID-19 patients. Lagana et al[34] reported the histological features of 40 patients who died of COVID-19-related complications and who had liver biochemical abnormalities. Two-thirds of the included patients presented macrovesicular steatosis, which was most commonly panlobular, while 2 patients (7%) showed active steatohepatitis. Half of the included patients had mild lobular necroinflammation and, therefore, active hepatitis, which was mild in 80% of the cases and moderate in the remaining 20%. Similarly, portal inflammation was reported in 20 patients, 3 of which had interface hepatitis. Lobular mild and focal cholestasis changes were observed in 15 (38%) cases. Although the ACE2 receptor is mainly expressed by cholangiocytes in the liver, ductopenia was not reported. Vascular alterations (i.e. phlebosclerosis, portal arteriolar muscular hyperplasia, focal fibrinoid necrosis, and sinusoidal thrombus) were less common (15%). Interestingly, no significant correlation was found between laboratory and histological findings. Wang et al[35] demonstrated, in 2 deceased COVID-19 patients, that SARS-CoV-2 can infect the liver causing direct cytopathy. They also reported massive hepatic apoptosis as well as binuclear hepatocytes. However, due to the small sample size, further studies are needed to confirm these preliminary findings.

# **COVID-19 IN PATIENTS WITH A PRE-EXISTING CLD**

Immune dysregulation is known to affect people with CLD or cirrhosis, leading to the concern that these patients are at higher risk of having a more severe form of COVID-19[36]. A limited number of studies have investigated the role of COVID-19 in patients with a pre-existing CLD and most of them only included a limited number of patients from a restricted geographical area. It is to be noted that all these studies reported a higher mortality rate among CLD patients[37-43]. Marjot et al[43] conducted one of the largest studies of CLD cases (745 patients) from 29 different countries. They showed



Table 1 Main studies reporting liver involvement in patients without pre-existing liver disease							
Ref.	Country	Patients	Liver involvement criteria	Patients with liver involvement, <i>n</i> (%)	Main findings		
Fan <i>et al</i> [ <mark>33</mark> ]	China	148	ALT > 40 U/L or AST > 35 U/L	55 (37.2)	Abnormal liver function is common in COVID-19 inpatients, leading to a longer hospital stay		
Goyal <i>et al</i> [17]	United States	375	ALT > 40 U/L	120 (32)	Mechanically ventilated patients more likely to have liver involvement.		
Lenti <i>et al</i> [29]	Italy	100	ALT or GGT > 50 U/L	58/93 (62.4)	Liver involvement correlate to higher mortality and ICU need in those who develop ARDS		
Medetalibeyoglu <i>et al</i> [32]	Turkey	554	ALT or AST > $40 \text{ U/L}$	153 (27.6)	Higher rate of moderate-to-severe pneumonia and ICU admission need in patients with liver involvement		
Phipps <i>et al</i> [31]	United States	2273	ALT > 50 U/L	537 (24)	Severe liver involvement was rare (6.4%) and led to worse outcomes (ICU admission, higher mortality)		
Ponziani et al[ <mark>30</mark> ]	Italy	515	AST > 45 U/I orALT > 45 U/I orGGT > 61	161 (31.3)	No cases of severe liver injury in this cohort. Liver involvement was generally mild and, although correlated to a higher need of ICU care, not associated to higher mortality		
Richardson <i>et al</i> [7]	United States	5700	ALT > 60	2176 (39)	Acute liver injury occurred in 1% of the included patients and was associated with higher mortality		
Schattenberg <i>et al</i> [28]	Germany	44	ALT >50 U/L	6/38 (15.8)	Severe liver involvement was rare (9%), with AST more commonly deranged than ALT		

ALT: Alanine aminotransferase; ARDS: Acute severe respiratory distress syndrome; AST: Aspartate aminotransferase; COVID-19: Coronavirus disease 2019; GGT; Gamma-glutamyl transpeptidase; ICU: Intensive care unit.

that CLD was associated with increased mortality according to the Child-Pugh class. They reported an increase in mortality, ranging from 19% in Child-Pugh-A patients to 51% in Child-Pugh-C patients. Although mortality has consistently reported to be increased in CLD patients, respiratory failure was found to be the main cause of death in these patients. Interestingly, alcohol-related liver disease was found to be independently associated to higher mortality. Liver decompensation was also reported to be common in cirrhotic patients (46%) with half of them having acute-on-chronic liver failure[44].

Among patients with CLD, liver transplant recipients were thought to represent a high risk category due to their frailty, comorbidities, and immunosuppressant therapy. Only few studies evaluated their clinical outcomes, showing conflicting results. Additionally, the majority of these studies are small case series in which patients did not always have the confirmation of SARS-CoV-2 infection[45-49]. Complying with preventive measures (*i.e.* frequent hand washing/sanitisation, use of surgical mask in public places and avoidance of public or crowded places) has been found effective to reduce the infection rate in this population[49]. A large multinational registry-based study[50], including 151 transplanted patients with laboratory-confirmed infection, showed that liver transplantation was not independently associated with higher mortality, hospitalisation rate or intensive care unit admission, whereas age and comorbidities were[47,50]. Tables 2 and 3 report the main studies focusing respectively on the outcome of COVID-19 in patients with CLD and in those with a transplanted liver.

#### **SPLEEN IMPAIRMENT IN COVID-19**

Spleen impairment in patients with COVID-19 has been poorly described. It is assumed that it may be driven by several mechanisms, including direct organ attack by the virus, cytokine-mediated immune pathogenesis, microvascular dysfunction, and lymphocyte apoptosis (Figure 2)[51].

Coronavirus detection in biopsies and autopsies has shown a tropism of this virus family for the spleen. The first available evidence is related to studies carried out on patients infected with SARS-CoV[11,52] and in experimental models of the Middle East respiratory syndrome[53]. In 2020, through immunohistochemistry techniques and the real-time reverse-transcript polymerase chain reaction assay, the SARS-CoV-2 nucleocapsid protein and the RNA were detected in the spleen tissue[54-56].

Zaishidene® WJG | https://www.wjgnet.com

#### Table 2 Main studies reporting outcomes in patients with pre-existing chronic liver disease

Ref.	Country	Patients	Patients with CLD, <i>n</i> (%)	Main findings
Bajaj et al[40]	United States	272	37 (13.6)	Higher mortality in cirrhotic COVID-19 positive patients
Hashemi <i>et al</i> [ <mark>41</mark> ]	United States	363	69 (19)	CLD patients had higher ICU admission and mechanical ventilation rate. CLD was a predictor of mortality
Iavarone <i>et al</i> [ <mark>42</mark> ]	Italy	50	50 (100)	COVID-19 infection led to liver function deterioration. CLD patients had increased mortality
Marjot <i>et al</i> [43]	International	1365	745 (54.6)	CLD correlate to higher mortality rate according to the CPT class. ALD was an independent risk factor for mortality
Qi et al[ <mark>39</mark> ]	China	21	21 (100)	Respiratory failure was the cause of death in most patients
Singh et al[37]	United States	250	60 (46.1)	Pre-existing CLD patients had higher hospitalisation and mortality rates
Sarin <i>et al</i> [38]	International	228	228 (100)	Decompensation of pre-existing CLD occurred in one fifth of cirrhotic patients

CLD: Chronic liver disease; COVID-19: Coronavirus disease 2019.

#### Table 3 Main studies reporting outcomes in liver transplant patients

Ref.	Country	Patients	Patients with LT	Main findings
Bhoori <i>et al</i> [45]	Italy	151 (COVID status unknown)	151 (100)	3 deaths recorded in long-term LT recipient on low immunosuppressant dose
Belli <i>et al</i> [47]	International	103	103 (100)	Mortality might correlate with age and longer time since LT
Donato <i>et al</i> [49]	Italy	640 (8 COVID positive)	640 (100)	Low prevalence of infection in LT patients who adhere to preventive measures
Lee <i>et al</i> [48]	United States	38	38 (100)	High mortality in LT patients regardless of time since transplant
Pereira <i>et al</i> [ <mark>46]</mark>	United States	90	14 (15)	Solid organ transplant recipient had more severe outcomes
Webb <i>et al</i> [ <mark>50</mark> ]	International	778	151 (19.4)	LT patients did not have a higher mortality, ICU admission or hospitalisation rate; age and comorbidities correlated with outcomes

COVID: Coronavirus disease; ICU: Intensive care unit; LT: Liver transplant.

This tropism of coronaviruses for the spleen, as for other organs, seems to be mediated by the presence of the ACE2 receptor. In fact, a study published in 2004 already described ACE2 receptors in the red pulp sinus endothelium[57]. More recent studies have confirmed the expression of ACE2 receptor in the splenic tissue, although at lower levels compared to others (i.e. small intestine, testis, kidneys, heart, thyroid, and adipose tissue)[58,59]. These studies also highlighted no difference, according to sex and age, in ACE2 receptor expression. Further immunohistochemical studies detected this receptor in tissue-resident CD169+ macrophages[54].

Autopsy studies have revealed interesting anatomical changes in the spleen during SARS-CoV-2 infection[60-63], including a reduction in the splenic cellular composition, with a specific depletion of T and B lymphocyte pools. Some authors assumed that this lymphocytopenia was linked to SARS-CoV-2-induced apoptosis, via Fas/Fas-ligand signalling, as well as increased interleukin (IL)-6 secretion by macrophages[54]. Furthermore, other frequent histopathological features were the white pulp atrophy and the reduction or absence of lymphoid follicles, with increased red pulp to white pulp proportion. In addition, spleen autopsies frequently showed a congested and haemorrhagic appearance. Microscopic studies of splenic vessels revealed, in many cases, a splenic infarction due to arterial thrombosis and proliferation of fibrous tissue in the sinuses.

Ref.	Country	Patients	Patients with spleen involvement, <i>n</i> (%)	Main findings
Feng et al[54]	China	6	6 (100)	ACE2 expression on tissue-resident CD169+ macrophages in spleen; viral NP antiger found in ACE2+ cells in spleen; direct damage of spleen tissue (lymph follicle depletion, splenic nodule atrophy, lymphocyte reduction, <i>etc.</i> )
Remmelink et al[55]	Belgium	17	11 (65)	SARS-CoV-2 RNA detected in spleen autopsy samples by RT-PCR assay
Sekulic <i>et al</i> [ <mark>56</mark> ]	United States	2	2 (100)	SARS-CoV-2 RNA detected at high level in spleen FFPE samples by RT-PCR assay
Han et al <mark>[58</mark> ]	China	7356	NA	Expression of ACE2 in spleen tissue (lower than in other tissues), without difference according to sex
Li et al[ <mark>59</mark> ]	China	31	NA	Expression of ACE2 in spleen tissue (lower than in other tissues), without difference according to sex and age
Xu et al[ <mark>60</mark> ]	China	10	10 (100)	Decrease in spleen cell composition with decrease in lymphocyte components, white pulp atrophied, lymphoid follicles decreased or absent, increase in red pulp to white pulp ratio
Menter <i>et al</i> [ <mark>61</mark> ]	Switzerland	21	6 (29)	Acute splenitis and/or septic neutrophilic leucocytosis of the red pulp, suggesting vascular disfunction in patients with COVID-19
Lax et al[ <mark>62</mark> ]	Austria	11	10 (90)	White pulp atrophy due to lymphocyte depletion, areas of haemorrhage with acute o chronic congestion
Duarte-Neto et al[63]	Brazil	5	5 (100)	Lymphoid hypoplasia in 100%, red pulp haemorrhages in 60%, splenitis in 40%, extramedullary haematopoiesis in 50%, endothelial changes in 80%, vasculitis and arterial thrombus in 20%
Lenti <i>et al</i> [ <mark>29</mark> ]	Italy	63	55 (87.3)	IgM memory B cell depletion that correlates with increased mortality and superimposed infections
Kaneko <i>et al</i> [65]	United States	11	11 (100)	Loss of spleen germinal centres due to depletion of Bcl-6+ germinal centre B cells and Bcl-6+ germinal centre T follicular helper cells, resulting in a dysregulated humoral immune response

COVID-19: Coronavirus disease 2019; FFPE: Formalin-fixed paraffin-embedded; NA: Not available; NP: Nucleocapsid protein; RT-PCR: Real-time reversetranscript polymerase chain reaction; SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2.

> The functional impact of these anatomical damages has been poorly investigated. A recent study<sup>[12]</sup> assessed the splenic immunological function through the detection of circulating IgM+ IgD+ CD27+ B lymphocytes, also known as IgM memory B cells, a unique B cell population in the marginal zone of the spleen which plays a major role in early inflammatory responses, including those caused by viral and bacterial infections [64]. A high prevalence of persistent IgM memory B cell depletion was demonstrated in patients with COVID-19, resulting in a higher mortality rate and an increased risk of developing superimposed bacterial infections. Other molecular studies have suggested that the loss of germinal centres may be due to the depletion of Bcl-6+ germinal centre B cells and Bcl-6+ germinal centre T follicular helper cells, resulting in a dysregulated immune response during the SARS-CoV-2 infection[65]. Although further studies are needed, it can be assumed that splenic involvement could be one of the causes of immune perturbations associated with severe COVID-19[66]. It still has to be ascertained whether the spleen immunological defect is reversible or not. Conversely, the haemocateretic function, assessed by counting pitted red cells (PRCs; red cells with membrane abnormalities [pits] visible under interference phase microscopy[67]) was preserved in patients with acute COVID-19, contrary to what happens in asplenia and spleen hypofunction[12]. The long average life span of circulating erythrocytes (approximately 120 d) might explain the lack of PRC increase in the acute phase of COVID-19.

### COVID-19 IN ASPLENIC OR HYPOSPLENIC PATIENTS

It is well known that asplenic or hyposplenic patients are predisposed to a greater risk of developing serious infections or overwhelming post-splenectomy infections, due to the defect in mounting the immune response against encapsulated bacteria[67,68].



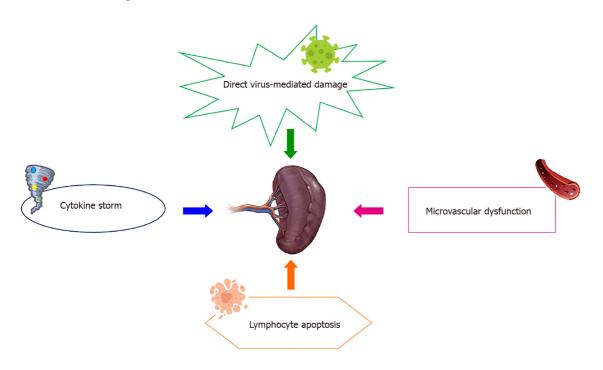


Figure 2 Putative mechanisms of spleen damage in coronavirus disease 2019.

Starting from these premises, it would be interesting to know whether patients with asplenia or spleen dysfunction could be more susceptible to Sars-CoV-2, both in terms of severity and incidence of the disease. Indeed, apart from a document drafted by the British Society of Haematology, stating that asplenic and hyposplenic patients are not exposed to a major risk of COVID-19, data regarding this population are completely missing[69]. Moreover, it is unknown whether patients who might develop spleen hypofunction as a consequence of COVID-19 could be more exposed to infections sustained by encapsulated bacteria and less responsive to vaccine immune-prophylaxis.

According to a single-centre, longitudinal, prospective, study conducted in an academic, tertiary referral hospital from Northern Italy, asplenic/hyposplenic patients did not seem to have an increased risk of developing COVID-19. The study had the purpose of characterising the spleen function, through circulating IgM memory B cell and PRC detection, in patients with COVID-19, in relation to their clinical outcome. Overall, 66 COVID-19 patients (mean age: 74 ± 16.6 years; 29 females) were enrolled; three patients had been splenectomised for trauma, all of them having IgM memory B cell depletion, and one of them died. Most COVID-19 patients had marked IgM memory B cell depletion, and this was associated to a higher mortality rate and a higher risk of developing superimposed infections[12]. Another important study conducted to identify, quantify, and analyse factors associated with COVID-19-related death in one of the largest cohort studies on this topic conducted so far (primary care records of 17278392 adults were linked to 10926 COVID-19-related deaths), considered asplenia as a comorbidity of interest. The results showed that 0.2% of the study population were affected by asplenia, and that the proportion of COVID-19-related death was 0.14%. In addition, asplenic COVID-19 patients had a 1.62 higher risk of death than individuals with a normal spleen function[70].

Indeed, further studies are needed to clarify the impact of SARS-CoV-2 in patients without a spleen or with spleen dysfunction. Table 4 reports the main studies reporting COVID-19 related spleen dysfunction.

### CONCLUSION

While the involvement of the respiratory system in SARS-CoV-2 infection is well established, the impact on the liver and spleen has not been explored much. Some studies have shown a direct tropism of the virus for these organs, and this may be one of the mechanisms underlying their damage, in association with the systemic inflammatory response. Regarding the liver, its involvement seems to be quite common, especially in more severe cases of infection, resulting in a worse prognosis for these



patients. The spleen involvement, on the other hand, has been poorly investigated. The splenic immune function appears to be defective in COVID-19 patients, resulting in a higher mortality rate and superimposed infections. Further studies may lead to a better diagnostic and therapeutic approach in SARS-CoV-2-infected patients, especially those with pre-existing liver and spleen diseases, who seem to be at higher risk of a worse outcome.

## ACKNOWLEDGEMENTS

Dr. Marco Vincenzo Lenti is grateful to the University of Pavia for supporting his research projects. We thank Intermediate SRL for having proofread the paper.

#### REFERENCES

- Wang C, Horby PW, Hayden FG, Gao GF. A novel coronavirus outbreak of global health concern. 1 Lancet 2020; 395: 470-473 [PMID: 31986257 DOI: 10.1016/S0140-6736(20)30185-9]
- Coronaviridae Study Group of the International Committee on Taxonomy of Viruses. The species Severe acute respiratory syndrome-related coronavirus: classifying 2019-nCoV and naming it SARS-CoV-2. Nat Microbiol 2020; 5: 536-544 [PMID: 32123347 DOI: 10.1038/s41564-020-0695-z]
- 3 Eurosurveillance Editorial Team. Note from the editors: World Health Organization declares novel coronavirus (2019-nCoV) sixth public health emergency of international concern. Euro Surveill 2020; 25 [PMID: 32019636 DOI: 10.2807/1560-7917.ES.2020.25.5.200131e]
- 4 Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q, Wang J, Cao B. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet 2020; 395: 497-506 [PMID: 31986264 DOI: 10.1016/S0140-6736(20)30183-5]
- Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, Wang B, Xiang H, Cheng Z, Xiong Y, Zhao Y, Li Y, 5 Wang X, Peng Z. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. JAMA 2020; 323: 1061-1069 [PMID: 32031570 DOI: 10.1001/jama.2020.1585]
- Marasco G, Lenti MV, Cremon C, Barbaro MR, Stanghellini V, Di Sabatino A, Barbara G. 6 Implications of SARS-CoV-2 infection for neurogastroenterology. Neurogastroenterol Motil 2021; 33: e14104 [PMID: 33591607 DOI: 10.1111/nmo.14104]
- Richardson S, Hirsch JS, Narasimhan M, Crawford JM, McGinn T, Davidson KW; the Northwell COVID-19 Research Consortium, Barnaby DP, Becker LB, Chelico JD, Cohen SL, Cookingham J, Coppa K, Diefenbach MA, Dominello AJ, Duer-Hefele J, Falzon L, Gitlin J, Hajizadeh N, Harvin TG, Hirschwerk DA, Kim EJ, Kozel ZM, Marrast LM, Mogavero JN, Osorio GA, Qiu M, Zanos TP. Presenting Characteristics, Comorbidities, and Outcomes Among 5700 Patients Hospitalized With COVID-19 in the New York City Area. JAMA 2020; 323: 2052-2059 [PMID: 32320003 DOI: 10.1001/jama.2020.6775]
- Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, Liu L, Shan H, Lei CL, Hui DSC, Du B, Li LJ, Zeng G, Yuen KY, Chen RC, Tang CL, Wang T, Chen PY, Xiang J, Li SY, Wang JL, Liang ZJ, Peng YX, Wei L, Liu Y, Hu YH, Peng P, Wang JM, Liu JY, Chen Z, Li G, Zheng ZJ, Qiu SQ, Luo J, Ye CJ, Zhu SY, Zhong NS; China Medical Treatment Expert Group for Covid-19. Clinical Characteristics of Coronavirus Disease 2019 in China. N Engl J Med 2020; 382: 1708-1720 [PMID: 32109013 DOI: 10.1056/NEJMoa2002032]
- Becchetti C, Zambelli MF, Pasulo L, Donato MF, Invernizzi F, Detry O, Dahlqvist G, Ciccarelli O, Morelli MC, Fraga M, Svegliati-Baroni G, van Vlierberghe H, Coenraad MJ, Romero MC, de Gottardi A, Toniutto P, Del Prete L, Abbati C, Samuel D, Pirenne J, Nevens F, Dufour JF; COVID-LT group. COVID-19 in an international European liver transplant recipient cohort. Gut 2020; 69: 1832-1840 [PMID: 32571972 DOI: 10.1136/gutjnl-2020-321923]
- 10 Fernández-Ruiz M, Andrés A, Loinaz C, Delgado JF, López-Medrano F, San Juan R, González E, Polanco N, Folgueira MD, Lalueza A, Lumbreras C, Aguado JM. COVID-19 in solid organ transplant recipients: A single-center case series from Spain. Am J Transplant 2020; 20: 1849-1858 [PMID: 32301155 DOI: 10.1111/ajt.15929]
- 11 Zhan J, Deng R, Tang J, Zhang B, Tang Y, Wang JK, Li F, Anderson VM, McNutt MA, Gu J. The spleen as a target in severe acute respiratory syndrome. FASEB J 2006; 20: 2321-2328 [PMID: 17077309 DOI: 10.1096/fj.06-6324com]
- Lenti MV, Aronico N, Pellegrino I, Boveri E, Giuffrida P, Borrelli de Andreis F, Morbini P, Vanelli 12 L, Pasini A, Ubezio C, Melazzini F, Rascaroli A, Antoci V, Merli S, Di Terlizzi F, Sabatini U, Cambiè G, Tenore A, Picone C, Vanoli A, Arcaini L, Baldanti F, Paulli M, Corazza GR, Di Sabatino A. Depletion of circulating IgM memory B cells predicts unfavourable outcome in COVID-19. Sci Rep 2020; 10: 20836 [PMID: 33257775 DOI: 10.1038/s41598-020-77945-8]
- 13 Yang J, Zheng Y, Gou X, Pu K, Chen Z, Guo Q, Ji R, Wang H, Wang Y, Zhou Y. Prevalence of comorbidities and its effects in patients infected with SARS-CoV-2: a systematic review and meta-



analysis. Int J Infect Dis 2020; 94: 91-95 [PMID: 32173574 DOI: 10.1016/j.ijid.2020.03.017]

- Grasselli G, Zangrillo A, Zanella A, Antonelli M, Cabrini L, Castelli A, Cereda D, Coluccello A, Foti 14 G, Fumagalli R, Iotti G, Latronico N, Lorini L, Merler S, Natalini G, Piatti A, Ranieri MV, Scandroglio AM, Storti E, Cecconi M, Pesenti A; COVID-19 Lombardy ICU Network. Baseline Characteristics and Outcomes of 1591 Patients Infected With SARS-CoV-2 Admitted to ICUs of the Lombardy Region, Italy. JAMA 2020; 323: 1574-1581 [PMID: 32250385 DOI: 10.1001/jama.2020.5394]
- 15 Borges do Nascimento IJ, Cacic N, Abdulazeem HM, von Groote TC, Jayarajah U, Weerasekara I, Esfahani MA, Civile VT, Marusic A, Jeroncic A, Carvas Junior N, Pericic TP, Zakarija-Grkovic I, Meirelles Guimarães SM, Luigi Bragazzi N, Bjorklund M, Sofi-Mahmudi A, Altujjar M, Tian M, Arcani DMC, O'Mathúna DP, Marcolino MS. Novel Coronavirus Infection (COVID-19) in Humans: A Scoping Review and Meta-Analysis. J Clin Med 2020; 9 [PMID: 32235486 DOI: 10.3390/jcm9040941]
- 16 Shi H, Han X, Jiang N, Cao Y, Alwalid O, Gu J, Fan Y, Zheng C. Radiological findings from 81 patients with COVID-19 pneumonia in Wuhan, China: a descriptive study. Lancet Infect Dis 2020; 20: 425-434 [PMID: 32105637 DOI: 10.1016/S1473-3099(20)30086-4]
- Goyal P, Choi JJ, Pinheiro LC, Schenck EJ, Chen R, Jabri A, Satlin MJ, Campion TR Jr, Nahid M, 17 Ringel JB, Hoffman KL, Alshak MN, Li HA, Wehmeyer GT, Rajan M, Reshetnyak E, Hupert N, Horn EM, Martinez FJ, Gulick RM, Safford MM. Clinical Characteristics of Covid-19 in New York City. N Engl J Med 2020; 382: 2372-2374 [PMID: 32302078 DOI: 10.1056/NEJMc2010419]
- 18 Bertolini A, van de Peppel IP, Bodewes FAJA, Moshage H, Fantin A, Farinati F, Fiorotto R, Jonker JW, Strazzabosco M, Verkade HJ, Peserico G. Abnormal Liver Function Tests in Patients With COVID-19: Relevance and Potential Pathogenesis. Hepatology 2020; 72: 1864-1872 [PMID: 32702162 DOI: 10.1002/hep.31480]
- 19 Du M, Cai G, Chen F, Christiani DC, Zhang Z, Wang M. Multiomics Evaluation of Gastrointestinal and Other Clinical Characteristics of COVID-19. Gastroenterology 2020; 158: 2298-2301.e7 [PMID: 32234303 DOI: 10.1053/j.gastro.2020.03.045]
- 20 Chai X, Hu L, Zhang Y, Han W, Lu Z, Ke A, Zhou J, Shi G, Fang N, Fan J, Cai J, Lan F. Specific ACE2 Expression in Cholangiocytes May Cause Liver Damage After 2019-nCoV Infection. bioRxiv 2020 [DOI: 10.1101/2020.02.03.931766]
- 21 Wu J, Song S, Cao HC, Li LJ. Liver diseases in COVID-19: Etiology, treatment and prognosis. World J Gastroenterol 2020; 26: 2286-2293 [PMID: 32476793 DOI: 10.3748/wjg.v26.i19.2286]
- 22 Samavati L, Uhal BD. ACE2, Much More Than Just a Receptor for SARS-COV-2. Front Cell Infect Microbiol 2020; 10: 317 [PMID: 32582574 DOI: 10.3389/fcimb.2020.00317]
- Cha MH, Regueiro M, Sandhu DS. Gastrointestinal and hepatic manifestations of COVID-19: A 23 comprehensive review. World J Gastroenterol 2020; 26: 2323-2332 [PMID: 32476796 DOI: 10.3748/wjg.v26.i19.2323]
- 24 Yang RX, Zheng RD, Fan JG. Etiology and management of liver injury in patients with COVID-19. World J Gastroenterol 2020; 26: 4753-4762 [PMID: 32921955 DOI: 10.3748/wjg.v26.i32.4753]
- 25 Lu L, Shuang L, Manman X, Yu P, Zheng S, Duan Z, Liu J, Chen Y, Li J. Risk factors related to hepatic injury in patients with corona virus disease 2019. medRxiv 2020
- 26 Cao X. COVID-19: immunopathology and its implications for therapy. Nat Rev Immunol 2020; 20: 269-270 [PMID: 32273594 DOI: 10.1038/s41577-020-0308-3]
- Zhang C, Shi L, Wang FS. Liver injury in COVID-19: management and challenges. Lancet 27 Gastroenterol Hepatol 2020; 5: 428-430 [PMID: 32145190 DOI: 10.1016/S2468-1253(20)30057-1]
- Schattenberg JM, Labenz C, Wörns MA, Menge P, Weinmann A, Galle PR, Sprinzl MF. Patterns of 28 liver injury in COVID-19 - a German case series. United European Gastroenterol J 2020; 8: 814-819 [PMID: 32588791 DOI: 10.1177/2050640620931657]
- 29 Lenti MV, Borrelli de Andreis F, Pellegrino I, Klersy C, Merli S, Miceli E, Aronico N, Mengoli C, Di Stefano M, Cococcia S, Santacroce G, Soriano S, Melazzini F, Delliponti M, Baldanti F, Triarico A, Corazza GR, Pinzani M, Di Sabatino A; Internal Medicine Covid-19 Team. Impact of COVID-19 on liver function: results from an internal medicine unit in Northern Italy. Intern Emerg Med 2020; 15: 1399-1407 [PMID: 32651938 DOI: 10.1007/s11739-020-02425-w]
- 30 Ponziani FR, Del Zompo F, Nesci A, Santopaolo F, Ianiro G, Pompili M, Gasbarrini A; "Gemelli against COVID-19" group. Liver involvement is not associated with mortality: results from a large cohort of SARS-CoV-2-positive patients. Aliment Pharmacol Ther 2020; 52: 1060-1068 [PMID: 32628793 DOI: 10.1111/apt.15996]
- Phipps MM, Barraza LH, LaSota ED, Sobieszczyk ME, Pereira MR, Zheng EX, Fox AN, Zucker J, 31 Verna EC. Acute Liver Injury in COVID-19: Prevalence and Association with Clinical Outcomes in a Large U.S. Cohort. Hepatology 2020; 72: 807-817 [PMID: 32473607 DOI: 10.1002/hep.31404]
- Medetalibeyoglu A, Catma Y, Senkal N, Ormeci A, Cavus B, Kose M, Bayramlar OF, Yildiz G, 32 Akyuz F, Kaymakoglu S, Tukek T. The effect of liver test abnormalities on the prognosis of COVID-19. Ann Hepatol 2020; 19: 614-621 [PMID: 32920162 DOI: 10.1016/j.aohep.2020.08.068]
- 33 Fan Z, Chen L, Li J, Cheng X, Yang J, Tian C, Zhang Y, Huang S, Liu Z, Cheng J. Clinical Features of COVID-19-Related Liver Functional Abnormality. Clin Gastroenterol Hepatol 2020; 18: 1561-1566 [PMID: 32283325 DOI: 10.1016/j.cgh.2020.04.002]
- Lagana SM, Kudose S, Iuga AC, Lee MJ, Fazlollahi L, Remotti HE, Del Portillo A, De Michele S, 34 de Gonzalez AK, Saqi A, Khairallah P, Chong AM, Park H, Uhlemann AC, Lefkowitch JH, Verna EC. Hepatic pathology in patients dying of COVID-19: a series of 40 cases including clinical,



histologic, and virologic data. Mod Pathol 2020; 33: 2147-2155 [PMID: 32792598 DOI: 10.1038/s41379-020-00649-x]

- Wang Y, Liu S, Liu H, Li W, Lin F, Jiang L, Li X, Xu P, Zhang L, Zhao L, Cao Y, Kang J, Yang J, 35 Li L, Liu X, Li Y, Nie R, Mu J, Lu F, Zhao S, Lu J, Zhao J. SARS-CoV-2 infection of the liver directly contributes to hepatic impairment in patients with COVID-19. J Hepatol 2020; 73: 807-816 [PMID: 32437830 DOI: 10.1016/j.jhep.2020.05.002]
- Albillos A, Lario M, Álvarez-Mon M. Cirrhosis-associated immune dysfunction: distinctive features 36 and clinical relevance. J Hepatol 2014; 61: 1385-1396 [PMID: 25135860 DOI: 10.1016/j.jhep.2014.08.010]
- 37 Singh S, Khan A. Clinical Characteristics and Outcomes of Coronavirus Disease 2019 Among Patients With Preexisting Liver Disease in the United States: A Multicenter Research Network Study. Gastroenterology 2020; 159: 768-771.e3 [PMID: 32376408 DOI: 10.1053/j.gastro.2020.04.064]
- Sarin SK, Choudhury A, Lau GK, Zheng MH, Ji D, Abd-Elsalam S, Hwang J, Qi X, Cua IH, Suh JI, Park JG, Putcharoen O, Kaewdech A, Piratvisuth T, Treeprasertsuk S, Park S, Wejnaruemarn S, Payawal DA, Baatarkhuu O, Ahn SH, Yeo CD, Alonzo UR, Chinbayar T, Loho IM, Yokosuka O, Jafri W, Tan S, Soo LI, Tanwandee T, Gani R, Anand L, Esmail ES, Khalaf M, Alam S, Lin CY, Chuang WL, Soin AS, Garg HK, Kalista K, Batsukh B, Purnomo HD, Dara VP, Rathi P, Al Mahtab M, Shukla A, Sharma MK, Omata M; APASL COVID Task Force, APASL COVID Liver Injury Spectrum Study (APCOLIS Study-NCT 04345640). Pre-existing liver disease is associated with poor outcome in patients with SARS CoV2 infection; The APCOLIS Study (APASL COVID-19 Liver Injury Spectrum Study). Hepatol Int 2020; 14: 690-700 [PMID: 32623632 DOI: 10.1007/s12072-020-10072-8
- 39 Qi X, Liu Y, Wang J, Fallowfield JA, Li X, Shi J, Pan H, Zou S, Zhang H, Chen Z, Li F, Luo Y, Mei M, Liu H, Wang Z, Li J, Yang H, Xiang H, Liu T, Zheng MH, Liu C, Huang Y, Xu D, Kang N, He Q, Gu Y, Zhang G, Shao C, Liu D, Zhang L, Kawada N, Jiang Z, Wang F, Xiong B, Takehara T, Rockey DC; COVID-Cirrhosis-CHESS Group. Clinical course and risk factors for mortality of COVID-19 patients with pre-existing cirrhosis: a multicentre cohort study. Gut 2021; 70: 433-436 [PMID: 32434831 DOI: 10.1136/gutjnl-2020-321666]
- Bajaj JS, Garcia-Tsao G, Biggins SW, Kamath PS, Wong F, McGeorge S, Shaw J, Pearson M, Chew 40 M, Fagan A, de la Rosa Rodriguez R, Worthington J, Olofson A, Weir V, Trisolini C, Dwyer S, Reddy KR. Comparison of mortality risk in patients with cirrhosis and COVID-19 compared with patients with cirrhosis alone and COVID-19 alone: multicentre matched cohort. Gut 2021; 70: 531-536 [PMID: 32660964 DOI: 10.1136/gutjnl-2020-322118]
- 41 Hashemi N, Viveiros K, Redd WD, Zhou JC, McCarty TR, Bazarbashi AN, Hathorn KE, Wong D, Njie C, Shen L, Chan WW. Impact of chronic liver disease on outcomes of hospitalized patients with COVID-19: A multicentre United States experience. Liver Int 2020; 40: 2515-2521 [PMID: 32585065 DOI: 10.1111/liv.14583]
- 42 Iavarone M, D'Ambrosio R, Soria A, Triolo M, Pugliese N, Del Poggio P, Perricone G, Massironi S, Spinetti A, Buscarini E, Viganò M, Carriero C, Fagiuoli S, Aghemo A, Belli LS, Lucà M, Pedaci M, Rimondi A, Rumi MG, Invernizzi P, Bonfanti P, Lampertico P. High rates of 30-day mortality in patients with cirrhosis and COVID-19. J Hepatol 2020; 73: 1063-1071 [PMID: 32526252 DOI: 10.1016/j.jhep.2020.06.001]
- 43 Marjot T, Moon AM, Cook JA, Abd-Elsalam S, Aloman C, Armstrong MJ, Pose E, Brenner EJ, Cargill T, Catana MA, Dhanasekaran R, Eshraghian A, García-Juárez I, Gill US, Jones PD, Kennedy J, Marshall A, Matthews C, Mells G, Mercer C, Perumalswami PV, Avitabile E, Qi X, Su F, Ufere NN, Wong YJ, Zheng MH, Barnes E, Barritt AS 4th, Webb GJ. Outcomes following SARS-CoV-2 infection in patients with chronic liver disease: An international registry study. J Hepatol 2021; 74: 567-577 [PMID: 33035628 DOI: 10.1016/j.jhep.2020.09.024]
- 44 Patrono D, Lupo F, Canta F, Mazza E, Mirabella S, Corcione S, Tandoi F, De Rosa FG, Romagnoli R. Outcome of COVID-19 in liver transplant recipients: A preliminary report from Northwestern Italy. Transpl Infect Dis 2020; 22: e13353 [PMID: 32500942 DOI: 10.1111/tid.13353]
- Bhoori S, Rossi RE, Citterio D, Mazzaferro V. COVID-19 in long-term liver transplant patients: 45 preliminary experience from an Italian transplant centre in Lombardy. Lancet Gastroenterol Hepatol 2020; 5: 532-533 [PMID: 32278366 DOI: 10.1016/S2468-1253(20)30116-3]
- Pereira MR, Mohan S, Cohen DJ, Husain SA, Dube GK, Ratner LE, Arcasoy S, Aversa MM, Benvenuto LJ, Dadhania DM, Kapur S, Dove LM, Brown RS Jr, Rosenblatt RE, Samstein B, Uriel N, Farr MA, Satlin M, Small CB, Walsh TJ, Kodiyanplakkal RP, Miko BA, Aaron JG, Tsapepas DS, Emond JC, Verna EC. COVID-19 in solid organ transplant recipients: Initial report from the US epicenter. Am J Transplant 2020; 20: 1800-1808 [PMID: 32330343 DOI: 10.1111/ajt.15941]
- Belli LS, Duvoux C, Karam V, Adam R, Cuervas-Mons V, Pasulo L, Loinaz C, Invernizzi F, Patrono 47 D, Bhoori S, Ciccarelli O, Morelli MC, Castells L, Lopez-Lopez V, Conti S, Fondevila C, Polak W. COVID-19 in liver transplant recipients: preliminary data from the ELITA/ELTR registry. Lancet Gastroenterol Hepatol 2020; 5: 724-725 [PMID: 32505228 DOI: 10.1016/S2468-1253(20)30183-7]
- Lee BT, Perumalswami PV, Im GY, Florman S, Schiano TD; COBE Study Group. COVID-19 in 48 Liver Transplant Recipients: An Initial Experience From the US Epicenter. Gastroenterology 2020; 159: 1176-1178.e2 [PMID: 32442561 DOI: 10.1053/j.gastro.2020.05.050]
- 49 Donato MF, Invernizzi F, Lampertico P, Rossi G. Health Status of Patients Who Underwent Liver Transplantation During the Coronavirus Outbreak at a Large Center in Milan, Italy. Clin Gastroenterol Hepatol 2020; 18: 2131-2133.e1 [PMID: 32334081 DOI: 10.1016/j.cgh.2020.04.041]



- Webb GJ, Marjot T, Cook JA, Aloman C, Armstrong MJ, Brenner EJ, Catana MA, Cargill T, 50 Dhanasekaran R, García-Juárez I, Hagström H, Kennedy JM, Marshall A, Masson S, Mercer CJ, Perumalswami PV, Ruiz I, Thaker S, Ufere NN, Barnes E, Barritt AS 4th, Moon AM. Outcomes following SARS-CoV-2 infection in liver transplant recipients: an international registry study. Lancet Gastroenterol Hepatol 2020; 5: 1008-1016 [PMID: 32866433 DOI: 10.1016/S2468-1253(20)30271-5]
- 51 Li H, Liu L, Zhang D, Xu J, Dai H, Tang N, Su X, Cao B. SARS-CoV-2 and viral sepsis: observations and hypotheses. Lancet 2020; 395: 1517-1520 [PMID: 32311318 DOI: 10.1016/S0140-6736(20)30920-X
- Tang JW, To KF, Lo AW, Sung JJ, Ng HK, Chan PK. Quantitative temporal-spatial distribution of 52 severe acute respiratory syndrome-associated coronavirus (SARS-CoV) in post-mortem tissues. J Med Virol 2007; 79: 1245-1253 [PMID: 17607787 DOI: 10.1002/jmv.20873]
- Chu H, Zhou J, Wong BH, Li C, Chan JF, Cheng ZS, Yang D, Wang D, Lee AC, Yeung ML, Cai JP, Chan IH, Ho WK, To KK, Zheng BJ, Yao Y, Qin C, Yuen KY. Middle East Respiratory Syndrome Coronavirus Efficiently Infects Human Primary T Lymphocytes and Activates the Extrinsic and Intrinsic Apoptosis Pathways. J Infect Dis 2016; 213: 904-914 [PMID: 26203058 DOI: 10.1093/infdis/jiv380]
- Feng Z, Diao B, Wang R, Wang G, Wang C, Tan Y, Liu L, Liu Y, Yuan Z, Ren L, Wu Y, Chen Y. 54 The Novel Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Directly Decimates Human Spleens and Lymph Nodes. medRxiv 2020
- 55 Remmelink M, De Mendonça R, D'Haene N, De Clercq S, Verocq C, Lebrun L, Lavis P, Racu ML, Trépant AL, Maris C, Rorive S, Goffard JC, De Witte O, Peluso L, Vincent JL, Decaestecker C, Taccone FS, Salmon I. Unspecific post-mortem findings despite multiorgan viral spread in COVID-19 patients. Crit Care 2020; 24: 495 [PMID: 32787909 DOI: 10.1186/s13054-020-03218-5]
- Sekulic M, Harper H, Nezami BG, Shen DL, Sekulic SP, Koeth AT, Harding CV, Gilmore H, Sadri 56 N. Molecular Detection of SARS-CoV-2 Infection in FFPE Samples and Histopathologic Findings in Fatal SARS-CoV-2 Cases. Am J Clin Pathol 2020; 154: 190-200 [PMID: 32451533 DOI: 10.1093/ajcp/agaa091]
- Hamming I, Timens W, Bulthuis ML, Lely AT, Navis G, van Goor H. Tissue distribution of ACE2 57 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. J Pathol 2004; 203: 631-637 [PMID: 15141377 DOI: 10.1002/path.1570]
- Han T, Kang J, Li G, Ge J, Gu J. Analysis of 2019-nCoV receptor ACE2 expression in different 58 tissues and its significance study. Ann Transl Med 2020; 8: 1077 [PMID: 33145296 DOI: 10.21037/atm-20-4281]
- Li MY, Li L, Zhang Y, Wang XS. Expression of the SARS-CoV-2 cell receptor gene ACE2 in a wide 59 variety of human tissues. Infect Dis Poverty 2020; 9: 45 [PMID: 32345362 DOI: 10.1186/s40249-020-00662-x
- Xu X, Chang XN, Pan HX, Su H, Huang B, Yang M, Luo DJ, Weng MX, Ma L, Nie X. [Pathological 60 changes of the spleen in ten patients with coronavirus disease 2019(COVID-19) by postmortem needle autopsy]. Zhonghua Bing Li Xue Za Zhi 2020; 49: 576-582 [PMID: 32340089 DOI: 10.3760/cma.j.cn112151-20200401-00278]
- Menter T, Haslbauer JD, Nienhold R, Savic S, Hopfer H, Deigendesch N, Frank S, Turek D, Willi N, 61 Pargger H, Bassetti S, Leuppi JD, Cathomas G, Tolnay M, Mertz KD, Tzankov A. Postmortem examination of COVID-19 patients reveals diffuse alveolar damage with severe capillary congestion and variegated findings in lungs and other organs suggesting vascular dysfunction. Histopathology 2020; 77: 198-209 [PMID: 32364264 DOI: 10.1111/his.14134]
- Lax SF, Skok K, Zechner P, Kessler HH, Kaufmann N, Koelblinger C, Vander K, Bargfrieder U, 62 Trauner M. Pulmonary Arterial Thrombosis in COVID-19 With Fatal Outcome : Results From a Prospective, Single-Center, Clinicopathologic Case Series. Ann Intern Med 2020; 173: 350-361 [PMID: 32422076 DOI: 10.7326/M20-2566]
- Duarte-Neto AN, Monteiro RAA, da Silva LFF, Malheiros DMAC, de Oliveira EP, Theodoro-Filho 63 J, Pinho JRR, Gomes-Gouvêa MS, Salles APM, de Oliveira IRS, Mauad T, Saldiva PHN, Dolhnikoff M. Pulmonary and systemic involvement in COVID-19 patients assessed with ultrasound-guided minimally invasive autopsy. Histopathology 2020; 77: 186-197 [PMID: 32443177 DOI: 10.1111/his.14160]
- Seifert M, Przekopowitz M, Taudien S, Lollies A, Ronge V, Drees B, Lindemann M, Hillen U, 64 Engler H, Singer BB, Küppers R. Functional capacities of human IgM memory B cells in early inflammatory responses and secondary germinal center reactions. Proc Natl Acad Sci USA 2015; 112: E546-E555 [PMID: 25624468 DOI: 10.1073/pnas.1416276112]
- Kaneko N, Kuo HH, Boucau J, Farmer JR, Allard-Chamard H, Mahajan VS, Piechocka-Trocha A, 65 Lefteri K, Osborn M, Bals J, Bartsch YC, Bonheur N, Caradonna TM, Chevalier J, Chowdhury F, Diefenbach TJ, Einkauf K, Fallon J, Feldman J, Finn KK, Garcia-Broncano P, Hartana CA, Hauser BM, Jiang C, Kaplonek P, Karpell M, Koscher EC, Lian X, Liu H, Liu J, Ly NL, Michell AR, Rassadkina Y, Seiger K, Sessa L, Shin S, Singh N, Sun W, Sun X, Ticheli HJ, Waring MT, Zhu AL, Alter G, Li JZ, Lingwood D, Schmidt AG, Lichterfeld M, Walker BD, Yu XG, Padera RF Jr, Pillai S; Massachusetts Consortium on Pathogen Readiness Specimen Working Group. Loss of Bcl-6-Expressing T Follicular Helper Cells and Germinal Centers in COVID-19. Cell 2020; 183: 143-157.e13 [PMID: 32877699 DOI: 10.1016/j.cell.2020.08.025]
- Kuri-Cervantes L, Pampena MB, Meng W, Rosenfeld AM, Ittner CAG, Weisman AR, Agyekum



RS, Mathew D, Baxter AE, Vella LA, Kuthuru O, Apostolidis SA, Bershaw L, Dougherty J, Greenplate AR, Pattekar A, Kim J, Han N, Gouma S, Weirick ME, Arevalo CP, Bolton MJ, Goodwin EC, Anderson EM, Hensley SE, Jones TK, Mangalmurti NS, Luning Prak ET, Wherry EJ, Meyer NJ, Betts MR. Comprehensive mapping of immune perturbations associated with severe COVID-19. Sci Immunol 2020; 5 [PMID: 32669287 DOI: 10.1126/sciimmunol.abd7114]

- Di Sabatino A, Carsetti R, Corazza GR. Post-splenectomy and hyposplenic states. Lancet 2011; 378: 67 86-97 [PMID: 21474172 DOI: 10.1016/S0140-6736(10)61493-6]
- Kruetzmann S, Rosado MM, Weber H, Germing U, Tournilhac O, Peter HH, Berner R, Peters A, 68 Boehm T, Plebani A, Quinti I, Carsetti R. Human immunoglobulin M memory B cells controlling Streptococcus pneumoniae infections are generated in the spleen. J Exp Med 2003; 197: 939-945 [PMID: 12682112 DOI: 10.1084/jem.20022020]
- 69 Ryan K, Cooper N, Eleftheriou P, Garg M, Grainger J, Hill Q, Howard J, Kesse-Adu R, Lugthart S, Laffan M, McDonald V, Misbah S, Pavord S. Guidance on shielding for Children and Adults with splenectomy or splenic dysfunction during the COVID-19 pandemic. British Society of Haematology. [cited 4 July 2020]. Available from: https://b-s-h.org.uk/media/18292/covid19-bsh-guidance-onsplenectomy-v2-fnal-6-may2020\_.pdf
- 70 Williamson EJ, Walker AJ, Bhaskaran K, Bacon S, Bates C, Morton CE, Curtis HJ, Mehrkar A, Evans D, Inglesby P, Cockburn J, McDonald HI, MacKenna B, Tomlinson L, Douglas IJ, Rentsch CT, Mathur R, Wong AYS, Grieve R, Harrison D, Forbes H, Schultze A, Croker R, Parry J, Hester F, Harper S, Perera R, Evans SJW, Smeeth L, Goldacre B. Factors associated with COVID-19-related death using OpenSAFELY. Nature 2020; 584: 430-436 [PMID: 32640463 DOI: 10.1038/s41586-020-2521-4]



WJG

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5932-5945

DOI: 10.3748/wjg.v27.i35.5932

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

MINIREVIEWS

# Primary gastric non-Hodgkin lymphomas: Recent advances regarding disease pathogenesis and treatment

Michael D Diamantidis, Maria Papaioannou, Evdoxia Hatjiharissi

**ORCID number: Michael D** Diamantidis 0000-0002-0041-5930; Maria Papaioannou 0000-0002-1999-9622; Evdoxia Hatjiharissi 0000-0002-8233-0542.

Author contributions: Diamantidis MD performed the literature review and wrote the manuscript; Hatjiharissi E wrote the manuscript, performed the literature review and corrected the manuscript; Papaioannou M wrote and corrected the manuscript.

Conflict-of-interest statement: All authors declare they have no conflicts of interest.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Michael D Diamantidis, Department of Hematology, Thalassemia and Sickle Cell Disease Unit, General Hospital of Larissa, Larissa 41221, Thessaly, Greece

Maria Papaioannou, Evdoxia Hatjiharissi, Division of Hematology, First Department of Internal Medicine, AHEPA General Hospital, Aristotle University of Thessaloniki, Thessaloniki 54636, Greece

Corresponding author: Evdoxia Hatjiharissi, MD, PhD, Consultant Hematologist, Division of Hematology, First Department of Internal Medicine, AHEPA General Hospital, Aristotle University of Thessaloniki, St. Kiriakidis 1, Thessaloniki 54636, Greece. ehatjiharissi@gmail.com

# Abstract

Primary gastric lymphomas (PGLs) are distinct lymphoproliferative neoplasms described as heterogeneous entities clinically and molecularly. Their main histological types are diffuse large B-cell lymphoma (DLBCL) or mucosaassociated lymphoma tissue. PGL has been one of the main fields of clinical research of our group in recent years. Although gastric DLBCLs are frequent, sufficient data to guide optimal care are scarce. Until today, a multidisciplinary approach has been applied, including chemotherapy, surgery, radiotherapy or a combination of these treatments. In this minireview article, we provide an overview of the clinical manifestations, diagnosis and staging of these diseases, along with their molecular pathogenesis and the most important related clinical published series. We then discuss the scientific gaps, perils and pitfalls that exist regarding the aforementioned studies, in parallel with the unmet need for future research and comment on the proper methodology for such retrospective studies. Aiming to fill this gap, we retrospectively evaluated the trends in clinical presentation, management and outcome among 165 patients with DLBCL PGL who were seen in our institutions in 1980-2014. The study cohort was divided into two subgroups, comparing the main 2 therapeutic options [cyclophosphamide doxorubicin vincristine prednisone (CHOP) vs rituximab-CHOP (R-CHOP)]. A better outcome with immunochemotherapy (R-CHOP) was observed. In the next 2 mo, we will present the update of our study with the same basic conclusion.

Key Words: Primary gastric lymphoma; Extranodal non-Hodgkin's lymphoma; Diffuse large B-cell lymphoma; Mucosa-associated lymphoid tissue; Immunochemotherapy; Rituximab-cyclophosphamide doxorubicin vincristine prednisone



Specialty type: Gastroenterology and hepatology

#### Country/Territory of origin: Greece

#### Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B Grade C (Good): 0 Grade D (Fair): D Grade E (Poor): 0

Received: March 22, 2021 Peer-review started: March 22, 2021 First decision: June 14, 2021 Revised: June 27, 2021 Accepted: August 30, 2021 Article in press: August 30, 2021 Published online: September 21, 2021

P-Reviewer: Ahmed M, Li G, Saito Μ S-Editor: Gao CC L-Editor: A P-Editor: Liu JH



©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: A few small, heterogeneous, retrospective studies have attempted to determine the optimal treatment for gastric diffuse large B-cell lymphoma, investigating the role of chemotherapy +/- rituximab, surgery and radiation in patient outcomes. Our retrospective research suggests that a better outcome is observed for these patients after the introduction of immunochemotherapy (rituximab-cyclophosphamide doxorubicin vincristine prednisone). Because statistical analysis might differ among various studies, it is crucial to correctly define the terms freedom from progression and lymphoma-specific survival. The latter provides information on whether the patients died from lymphoma or from other causes.

Citation: Diamantidis MD, Papaioannou M, Hatjiharissi E. Primary gastric non-Hodgkin lymphomas: Recent advances regarding disease pathogenesis and treatment. World J Gastroenterol 2021; 27(35): 5932-5945

URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5932.htm DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5932

# INTRODUCTION

Primary gastric lymphomas (PGLs) are a diverse group of lymphoproliferative disorders that originate from the stomach and comprise many different histologic types. Either of diffuse large B-cell lymphoma (DLBCL) subtype or mucosa-associated lymphoma tissue (MALT) histology, PGL is the second most common gastric malignancy globally, following the adenocarcinoma of the stomach[1,2]. The latter is the most common form of gastric cancer and the fifth most common malignancy in the world[3]. Despite the fact that prevention and treatment of *Helicobacter pylori* infection (H. pylori I) has led to a decrease in its overall incidence, gastric cancer remains the 3rd most deadly cancer, with an estimated 783000 deaths in 2018 worldwide[4,5]. Therefore, accurately recognizing and diagnosing gastric cancer from gastric lymphomas is important, as these diseases are treated differently, and any confusion may result in inappropriate treatment management.

The gastrointestinal tract (GIT) is the most common site for the development of extranodal lymphomas. The incidence of these neoplasms has been increasing in recent years [2,6]. The stomach represents 30%-40% of all extranodal lymphomas and 55%-65% of all GI lymphomas. The incidence of PGL varies from 4% to 20% of extranodal non-Hodgkin lymphomas (NHLs) and reaches up to 5% of primary gastric neoplasms<sup>[2]</sup>. The incidence of PGL is estimated to be 1 per 100000 in Western countries<sup>[7]</sup>. B-cell lymphomas are more frequent in these countries than in Eastern countries<sup>[1]</sup>.

To date, the term PGL was originally used to describe lymphomas that arise from the stomach. However, within the medical literature, controversy exists regarding the definition, staging and treatment of this entity. Most cases of PGLs are B-cell subtypes of NHLs. The majority of these subtypes have DLBCL histology and are classified as DLBCL of the stomach, not otherwise specified (NOS).

PGLs are histologically heterogeneous neoplasms. This contributes to a different biology, clinical presentation and prognosis and subsequently determines special therapeutic needs for each subtype[8,9]. For example, certain subtypes of PGLs, such as DLBCL, are more aggressive than others and require immediate therapy[8], whereas for patients with MALT histology, unique management is usually applied ranging from watch and wait to antibiotic-based treatment[9].

As stated above, PGLs are histologically, biologically and clinically heterogeneous neoplasms. Although gastric DLBCL is an extranodal high-grade lymphoma, it is considered less aggressive than its nodal counterpart and other extranodal DLBCL locations. Its appropriate treatment has not been satisfactorily determined, and treatment choices vary considerably. Human immunodeficiency virus, Epstein-Barr infection, hepatitis B virus, human T-cell lymphotropic virus 1, immunosuppression, celiac disease, inflammatory bowel disease, and H. pylori I have all been implicated in the factors predisposing patients to PGLs, increasing the risk of developing the disease [1,2,10]. PGL usually occurs in patients older than 50 years. There are many older



patients over 80 years of age. Males are more prone to be diagnosed with PGL with a 2-3-fold higher risk than females[2].

This review mainly focuses on DLBCL gastric lymphoma, which is one of the main fields of our clinical research and comments briefly on MALT lymphoma. The molecular etiology and pathophysiology of DLBCL gastric lymphomas and the available clinical data for their optimal management will be discussed. In parallel, a brief review of the MALT subtype that represents almost 50% of PGLs will also be provided. This review aims to meet the therapeutic needs of those who are involved and/or interested in the treatment of GI-DLBCL lymphomas and extensively focuses on the role of rituximab, the first in class anti-CD20 monoclonal antibody (mAb), in the outcome of patients with PGL of the DLBCL subtype.

#### CLINICAL MANIFESTATIONS AND DIAGNOSIS

The stomach is the most common site for the development of extranodal lymphomas in the GI tract, accounting for 60% of cases, followed by the small bowel, ileum, cecum, colon and rectum[7]. Distinguishing PGL from secondary dissemination of the stomach due to primary nodal lymphoma can be difficult. No peripheral and mediastinal lymphadenopathy at the time of diagnosis, no spleen or liver infiltration and normal blood counts are in contrast to the presence of a secondary gastric lymphoma[11].

The diagnosis of PGL can be delayed for many years due to the presence of nonspecific symptoms, mimicking peptic ulcer disease, gastritis, functional gastric or even pancreatic disorder. The main symptoms include nausea, vomiting, anorexia, abdominal distention, fullness or pain, indigestion, dyspepsia and weight loss, whereas weakness, night sweats, fever, jaundice, hematemesis or melena are less common[2,7]. An obvious epigastric mass or perforation is rare as an initial presentation[7,10,12].

An appropriate endoscopic evaluation with generously sized tissue samples is the hallmark of diagnosis. The diagnostic accuracy of endoscopic biopsy is very high, reaching 90%. Endoscopic ultrasonography can improve this diagnostic accuracy. The diagnosis becomes difficult when there is deep infiltration and preservation of the mucosa. Computed tomography (CT), magnetic resonance imaging (MRI) and 18Ffluorodeoxyglucose positron emission tomography (FDG-PET) assist in the diagnosis and staging of PGL[1,7]. Sporadically, PGL might present as multifocal, clonally identical foci surrounded by macroscopically unaffected tissue. Thus, gastric mapping of unaffected mucosa is strongly recommended[13]. Bone marrow infiltration, B symptoms and elevated lactate dehydrogenase (LDH) are more frequently encountered in nodal lymphomas than in gastric lymphomas[13].

Different staging systems have been proposed for PGLs. The Ann Arbor staging system, which is widely used for primary nodal lymphomas, is considered unsatisfactory as PGLs originate from the lining of the stomach instead of the lymph nodes[7, 13]. In recent years, a more specific Lugano staging system for PGLs was proposed and applied based on the Lugano score[14,15], which includes the following stages: Stage IE – Lymphoma is confined to the GIT (single lesion or multiple noncontiguous lesions): IE1 = mucosa, submucosa; IE2 = muscularis propria, serosa; Stage II -Lymphoma extends into the abdomen from the primary site within the GI tract: II1 = local nodal involvement; II2 = distant nodal involvement; Stage IIE – Penetration of serosa to involve adjacent organs or tissues; Stage IV - Disseminated extranodal involvement or concomitant supra diaphragmatic nodal involvement. Note: Stage III does not exist because gastric lymphoma is always below the diaphragm.

A complete staging work-up includes the following: Biochemical examinations, chest, abdomen and pelvis CT scan, bone marrow biopsy, thorough endoscopy including biopsies from the stomach, duodenum and gastroesophageal junction, endoscopic ultrasound, evaluation of the Waldever ring, investigation for H. pylori I, routine histology and immunohistochemistry. Cytogenetic studies and even fluorescence in situ hybridization (FISH) can all be used in biopsies to provide the appropriate information needed for optimal treatment. PET/CT scans have documented diagnostic and prognostic value only for DLBCL lymphomas, in contrast to MALT gastric lymphomas, which can be reported as false-negative because of the small tumor burden of the disease and their indolent behavior[13]. However, there is an unmet need regarding the use of PET scans in the clinical setting to guide treatment. Usually, this examination is performed before and after the end of treatment to guide therapeutic decisions as a standard of care. Because the stomach is

an abdominal organ, it is unclear how PET scans can assist in the aforementioned necessary clinical decisions.

In general, the prognosis of extranodal lymphomas varies according to the affected organ; it is poor in the testis, central nervous system (CNS) and intestine, whereas it is quite good in the stomach, mediastinum and bone. Nevertheless, PGL is an aggressive malignancy characterized by rapid growth. However, the prognosis of DLBCL PGL is relatively good, with a 5-year overall survival (OS) higher than 80% [7].

#### COMPARISON AMONG CLINICAL STUDIES/TREATMENT

The optimal treatment for DLBCL PGLs is not clear, because prospective clinical studies are missing. In the past, a spectrum of treatment approaches was applied, ranging from gastrectomy or radiotherapy alone to chemotherapy (cyclophosphamide doxorubicin vincristine prednisone, CHOP) or the combination of chemotherapy plus radiotherapy and surgery. Wang et al[16] compared surgery over conservative treatment in a retrospective study. Conservative treatment in this study included chemotherapy (CHOP) or radiotherapy alone, chemotherapy plus radiotherapy or H. pylori I eradication (HPE). The authors found superiority of surgery alone compared with conservative treatment in the DLBCL type regarding prognosis, but not in the MALT type[16]. Currently, the role of surgical resection has been minimized, even in cases of extreme intestinal obstruction, as immunochemotherapy can induce rapid and complete resolution of large obstructing tumor masses. Gastrectomy is restricted to the management of major complications, including perforation or hemorrhage of DLBCL PGLs.

In contrast, other studies demonstrated that DLBCL PGL is a potentially curable disease with rituximab-CHOP (R-CHOP)-like treatment, leading to long-term survival [17]. Investigators found that surgical treatment did not offer survival benefits when compared with chemotherapy for 5-year progression-free survival (PFS) and OS estimates and that no significant differences were noted in these endpoints for patients treated with R-CHOP or conventional chemotherapy [18].

Sohn *et al*[19] directly compared CHOP *vs* R-CHOP as a front-line approach in 93 patients with DLBCL PGL. With a median follow-up of 48 mo, no differences were noted among the 2 groups regarding OS, EFS and CR. High serum levels of  $\beta$ 2microglobulin were associated with worse OS and EFS in patients who received R-CHOP[19]. In a retrospective analysis of 95 Japanese patients, the clinical outcomes of gastric DLBCL were extremely favorable for localized-stage patients in the rituximab era. Conversely, these treatments were poor for advanced-stage patients [20]. Interestingly, an effective approach in treating deeply infiltrated DLBCL PGL patients by switching fractioned R-CHOP (rituximab d0, 50% dose of CHOP d1 and d5) to standard R-CHOP cycles guided by endoscopic ultrasonography has been proposed [21].

The following factors were identified as having a negative impact on survival: age above 65, Eastern Cooperative Oncology Group 2-3, B symptoms, bulky disease, IPI 3-4, more than 3 treatment lines, and absence of response to first-line treatment [17].

Conversely, other factors were considered negative for prognosis in the subsequent study: elevated LDH levels, chemotherapy or radiotherapy alone or the combination of chemotherapy plus radiotherapy[16]. The non-germinal center B-cell-like lymphoma (GCB) subtype has also been associated with shorter OS[18]. H. pylori I negativity, advanced Lugano stage and elevated LDH levels have been reported as adverse prognostic factors in gastric DLBCL[22].

Low serum albumin at diagnosis was the only risk factor for developing gastric complications, such as bleeding and stenosis, in patients with gastric DLBCL who received R-CHOP[23]. Furthermore, a low CD4:CD8 ratio at diagnosis is an independent poor prognostic factor for subsequent OS and EFS24 (24 mo after diagnosis) in patients with gastric DLBCL<sup>[24]</sup>. Finally, the microRNA miR-150 is reportedly a negative independent prognostic biomarker for primary GI DLBCL[25].

Some patients with DLBCL PGL also have a MALT component. The 5-year PFS and OS estimates were similar when de novo DLBCL patients were compared with DLBCL/MALT patients, suggesting that patients with a MALT component, along with DLBCL, might have the same biological type of lymphoma as de novo DLBCL patients [18]. In such DLBCL/MALT cases, an important deregulation of Bcl-2 and an upregulation of p53 protein of uncertain clinical significance have been observed[26]. A synopsis of the studies comparing R-CHOP vs CHOP for DLBCL PGLs is shown in Table 1. Indeed, there is a lack of a head-to-head comparison between CHOP and R-



Table 1 Studies comparing rituximab-cyclophosphamide doxorubicin vincristine prednisone vs cyclophosphamide doxorubicin vincristine prednisone for diffuse large B-cell lymphoma primary gastric lymphomas

Ref.		Number of Pts	R-CHOP OS	CHOP OS	R-CHOP PFS	CHOP PFS	Comments
Sohn <i>et al</i> [ <mark>19</mark> ], 2012	Double-arm Retrospective Study (R-CHOP <i>vs</i> CHOP as 1 <sup>st</sup> line treatment)	93 (55 R-CHOP, 38 CHOP)	3-yr 84.7% ( <i>P</i> > 0.05)	3-yr 94.7% (P > 0.05)	3-yr 81.7% (EFS) ( <i>P</i> > 0.05)	3-yr 86% (EFS) ( <i>P</i> > 0.05)	CR: (CHOP: 93.9%), (R- CHOP: 92.5%)
Liu <i>et al</i> [62], 2018	Double-arm Retrospective Study (diagnosis: 1973- 2000 era <i>vs</i> 2001-2014 era of immuno-CT)	SEER Database 7051 [(4186, 1973-2000), (2865, 2001-2014)	5-yr 53% ( <i>P</i> = 0.001)	5-yr 47% ( <i>P</i> = 0.001)			
Tanaka et al[20], 2012	Single-arm Retrospective Study (R-CHOP)	95	3-yr 91% (localized disease); 3-yr 95% (localized disease); 3- yr 64% (localized disease)		3-yr 91% (localized disease); 3-yr 92% (localized disease); 3- yr 43% (localized disease)		6c. R-CHOP; 3-4 c. R- CHOP plus radiotherapy; R-CHOP ± radiotherapy
Couto <i>et</i> <i>al</i> [17], 2021	Single-arm Retrospective Study (R-CHOP)	101	Not reached		Not reached		80% CR (after 1 <sup>st</sup> line); 54% CR (3 yrs FU)

R-CHOP: Rituximab-cyclophosphamide doxorubicin vincristine prednisone; CHOP: Cyclophosphamide doxorubicin vincristine prednisone; OS: Overall survival; PFS: Progression-free survival; EFS: Event free survival; CR: Complete remission; CT: Computed tomography; FU: Follow-up.

#### CHOP in PGLs.

Regarding the role of radiotherapy, more data are available for patients with gastric MALT lymphoma or early-stage gastric lymphoma. When there is an unsatisfactory response to HPE, recurrence after HPE or in MALT cases negative for *H. pylori* I, gastric radiotherapy of the entire stomach plus irradiation of the pathological and perigastric lymph nodes (30-440 Gy, 15-20 fractions) has been proposed. However, it is less clear whether radiotherapy should be applied in cases of DLBCL PGLs. However, involved-field radiotherapy has a role, especially for patients with DLBCL PGL of advanced stage who achieve partial remission (PR) after immunochemotherapy (R-CHOP)[27]. R-CHOP plus additional local treatment for gastric lesions (*e.g.*, consolidative radiotherapy or surgical resection) has also been recommended[28]. Alternatively, several studies have found that in the era of immunochemotherapy (R-CHOP), radiotherapy does not improve OS[29-31]. The side effects of radiotherapy should always be taken into account in clinical decision making[27].

Despite the presence of several clinical series involving primary gastric DLBCL lymphomas mainly addressing the issue of selecting the optimal treatment, there are sporadic single cases in the literature<sup>[22,32-34]</sup>. Some very rare, more aggressive cases of DLBCL lymphoma originating from the stomach and infiltrating the adrenals bilaterally have been reported [32,34]. The first patient presented with nausea, vomiting, abdominal pain and hypotension, was treated with glucocorticoids and died after developing respiratory failure, severe hypotension refractory to vasopressors and severe metabolic acidosis[34]. The second case was a DLBCL, PGL of the non-germinal center (non-GC) type. This patient received 8 cycles of rituximab therapy, 6 cycles of CHOP and 3 cycles of prophylactic intrathecal chemotherapy. The patient maintained a CR for approximately 14 mo after the completion of the aforementioned treatment. The latter is in favor of the hypothesis that DLBCL lymphomas of the stomach have a better prognosis than other DLBCL nodal and extranodal lymphomas. In contrast to the very dismal prognosis of primary adrenal lymphomas (PALs)[35], this patient survived, likely because the primary neoplasm was gastric DLBCL, which has better biological and clinical behavior for unknown molecular reasons (even though it is considered an aggressive neoplasm, being DLBCL).

Regarding the role of HPE, Nakamura *et al*[36] studied 420 patients with gastric MALT lymphoma and found a significant responsiveness to HPE therapy (77%), with treatment failure (relapse or progressive disease) occurring in only 9% of the patients. However, this primary refractory disease was not associated with a dismal outcome, as the subsequent therapy still yielded a 90% OS rate after 10 years[36].

Nevertheless, even though HPE has already been established as an optimal strategy for the management of gastric MALT lymphoma, there are conflicting results, either in favor of or against HPE for patients with DLBCL PGLs. Thus, HPE has been reported to be a suitable strategy for patients with DLBCL PGLs[37,38]. The concept of a less

aggressive biological behavior for *H. pylori* I-dependent gastric DLBCL has been proposed with the suggestion to apply HPE in such cases<sup>[39]</sup>. However, it is not clear how accurately these lymphomas can be distinguished. Alternatively, high-grade gastric lymphomas can rapidly progress if they do not respond to HPE. The loss of H. pylori I dependency and the possible high-grade lymphomatic evolution/ transformation are separate and distinct events in the natural history of PGL[38,40]. The description of defined molecular markers linked to *H. pylori* I dependency of PGLs is beyond the scope of this article.

Moreover, a substantial portion of early-stage *H. pylori* I-positive gastric de novo DLBCLs remain *H. pylori* I-dependent and respond to antibiotic treatment (HPE). Prospective studies to validate these findings are needed[41]. Our personal opinion is that HPE should not be applied as monotherapy, even in the early stage of *H. pylori* Ipositive DLBCL PGLs.

# **MOLECULAR PATHOGENESIS**

Extranodal lymphomas are distinct types of lymphomas that show a predilection for anatomical sites harboring extranodal lymphoid tissue, such as the CNS, testis, mediastinum, bone and GIT, in contrast to the typical pattern of the nodal counterpart in the lymph nodes for nodal lymphomas[42]. Extranodal lymphomas can even appear in immune-privileged (sanctuary) sites (CNS, testis) or arise in sites of chronic inflammation, effusions or other closed spaces within the body. The complex mechanisms of local immune evasion leading to extranodal lymphoproliferations have not been fully elucidated[43]. The capacity of mature lymphocytes to recirculate between blood and lymphoid tissue and to migrate to extranodal anatomical sites is crucial for the pathogenesis of the disease. During this process, lymphocytes interact with endothelial venules, mediated by receptor molecules (integrins and lymphocytes)[44].

The role of specific B-cell receptor (BCR) antigens has been proposed in the process of lymphomagenesis. Oncogenic translocations during BCR development and generation (VDJ rearrangement), the activation of mature B-cells and the germinal center reaction, the mechanisms of loss of immunological self-tolerance, and the role of infectious agents and autoantigens are all hallmarks and basic elements of lymphomagenesis, a complex multifactorial process, in both aggressive and indolent lymphomas<sup>[45]</sup>. Gastric DLBCL is a high-grade lymphoma compared to low-grade MALT lymphomas. Whether DLBCL transforms from low-grade MALT lymphoma or whether it arises de novo in the stomach is unknown. DLBCL gastric lymphoma has been associated with a lower CR and shorter survival than MALT lymphoma[2,46]. Nevertheless, transformed DLBCLs from MALTs are CD10- and Bcl-2-negative, while de novo DLBCLs are CD10- and Bcl-2-positive[31,46].

The oncogene Bcl-6 is located on chromosome 3q27 and is frequently present in the majority of extranodal high-grade lymphomas. Conversely, Bcl-2 oncogene expression was significantly lower in gastric lymphomas than in other primary extranodal highgrade B-cell lymphomas (HGBCLs). p53 protein expression did not differ significantly between these 2 groups[2].

#### Primary gastric DLBCL

DLBCL is described by diffuse proliferation of large, atypical cells, with vesicular nuclei, prominent nucleoli, and basophilic cytoplasm. These cells typically express CD19, CD20, CD22 and CD79a (pan-B-cell markers). Bcl-6 is expressed in 60% of cases. FISH can identify poor prognostic subtypes of DLBCL, such as double-hit (DH) or triple-hit (TH) lymphomas (high-grade, B-cells), characterized by translocations of MYC and Bcl-2 and/or Bcl-6[47,48]. DH or TH lymphomas are defined by their genetic aberrations, irrespective of their morphology. Genetic variability has been documented for DLBCL PGL[47]. Gene expression profiling distinguishes DLBCL into GCB and non-GCB or activated B-cell-like (ABC) subtypes based on the cell of origin profile. ABC lymphomas show a worse prognosis than GCB lymphomas<sup>[49]</sup>. In routine diagnostic practice, this screening is conducted by immunohistochemistry based on the assessment of three markers (CD10, bcl-6 and MUM1)[50] (Figure 1).

More analytically, ABC DLBCLs are characterized by nuclear factor kappa beta (NFκB) activation, showing a higher frequency of Bcl-2 amplifications, Bcl-6 rearrangements and recurrent mutations of MYD88, PRDM1 and CD79B, whereas GCBlike DLBCLs are enriched for activating EZH2 and Bcl-2 mutations, defined by perturbations/molecular defects in the JAK/STAT and PI3K/AKT signaling pathways [48]. EZH2 overexpression has been associated with inferior outcomes in patients with



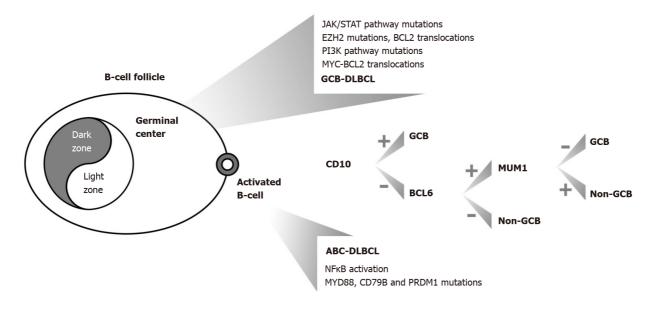


Figure 1 Primary gastric diffuse large B-cell lymphoma lymphomas and related molecular lesions. GCB: Germinal center B-cell lymphoma; ABC: Activated B-cell-like lymphoma; the combination of MYC plus BCL2 translocations corresponds to 'double hit lymphomas'; DLBCL: Diffuse large B-cell lymphoma; NFκβ: Nuclear factor κappa beta.

#### DLBCL PGL[51] (Figure 1).

Interestingly, 2 HGBCLs were included in the recent revised WHO classification of lymphoid neoplasms. These entities are clinically and biologically distinct from DLBCL NOS and Burkitt lymphoma (BL). The HGBCL, NOS entity includes cases previously termed 'unclassifiable, with features intermediate between DLBCL and BL', or showing blastoid morphology but lacking DH/TH translocations[49].

High levels of Bcl-6 expression were found in GCB gastric lymphomas, whereas in the non-GCB cases, a high Bcl-6 expression level correlated importantly with mutations producing Bcl-6 deregulation, even if in the latter cases no correlation was found between survival rates[2].

Clinical studies addressing the role of programmed cell death 1 (PD-1) and its ligand (PD-L1) have shown promising results. PD-1 blockade in patients with PD-L1 expression on tumor cells has been linked with clinical responses. Investigators from Japan evaluated the role of PD-L1 expression on neoplastic and non neoplastic immune cells in the microenvironment (miPD-L1) in a retrospective study of patients with GI DLBCL lymphoma. They found that elevated miPD-L1 expression had a favorable impact on the outcome of these DLBCL patients, regardless of the anatomical site of the disease[52].

#### Gastric MALT lymphoma

MALT lymphoma is a low-grade B-cell NHL, and the majority of cases (approximately 90%) are directly related to H. pylori I. However, 10% of gastric MALT lymphomas are H. pylori I negative[53]. Chronic H. pylori I of the gastric mucosa and the accompanying inflammation have been strongly linked to MALT lymphomagenesis. Moreover, abnormalities in the expression of various miRNAs contribute to the neoplastic gastric phenotype[54,55].

H. pylori I expresses proteins related to the corresponding genes, contributing to the related lymphomagenesis from the bacterium. These are cytotoxin-associated gene A (CagA), vacuolization cytotoxin A (VacA) and heat shock proteins (Hsps). The Cag pathogenicity island (a common gene sequence considered responsible for the pathophysiology of the infection) contains over 40 genes, which mainly code for a complex type IV secretion system. This pathogenicity island is usually absent from H. pylori I strains isolated from asymptomatic human carriers. The CagA protein is frequently co-expressed with the vacuolating cytotoxin VacA[56].

Hamoudi et al[57] established the connection between abnormal NF-KB signaling due to the chromosomal translocations, t(11;18)(q21;q21)/API2-MALT1, t(1;14) (p22;q32)/BCL10-IGH, t(14;18) (q32;q21)/IGH-MALT1 and t(3;14) (p13;q32)/FOXP1-IGH, in gastric MALT lymphomas [57,58] (Figure 2).

MALT1 and BCL10 proteins are involved in surface immune receptor-mediated activation of the NF-KB transcription factor; chromosomal translocations involving



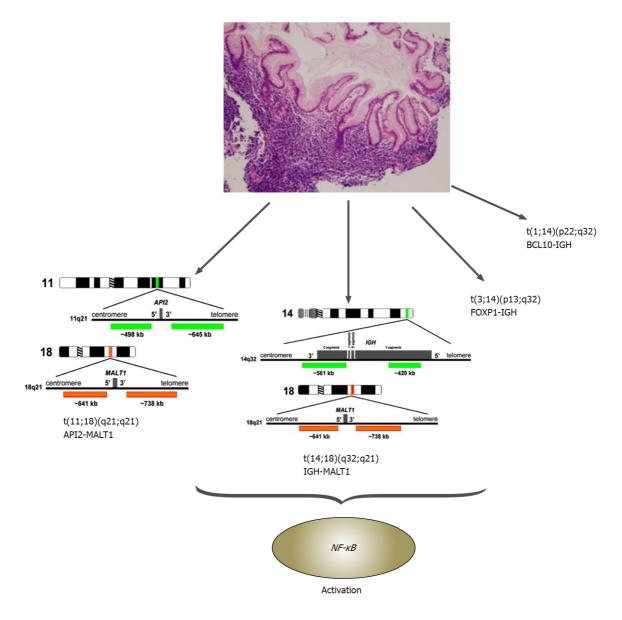


Figure 2 Gastric mucosa-associated lymphoid tissue lymphomas and related chromosomal translocations. BCL: B-cell lymphoma; FOXP: Forkhead box protein; IGH: Immunoglobulin heavy (chain); MALT: Mucosa-associated lymphoid tissue; NF-kB: Nuclear factor kappa beta.

these genes are believed to exert their oncogenic activities through constitutive activation of the NF- $\kappa$ B pathway, leading to the expression of numerous genes important for cell survival and proliferation[40,55,57,58] (Figure 2).

In gastric MALT lymphoma, t(11;18)/API2-MALT1 is the most frequent translocation, detected in 20% of cases. This translocation fuses the N-terminal region of *API2* to the C-terminal region of *MALT1* and generates a functional chimeric fusion, which can activate the NF- $\kappa$ B pathway. Clinically, t(11;18) is more frequently associated with the absence of *H. pylori* I, and the majority of translocation-positive cases do not respond to HPE therapy. Interestingly, t(11;18)-positive cases rarely transform to DLBCL[55,58].

Gastric MALT lymphoma is indirectly influenced by *H. pylori* I through T-cell stimulation, and recent studies have shown that *H. pylori*-triggering chemokines and their receptors, *H. pylori*-associated epigenetic changes, *H. pylori*-regulated miRNA expression and tumor infiltration by CD4<sup>+</sup> CD25<sup>+</sup> regulatory T cells contribute to lymphomagenesis of gastric MALT lymphoma (Figure 3). Recent studies have also demonstrated that the translocation of CagA into B lymphocytes inhibits apoptosis through p53 accumulation, BAD phosphorylation and the upregulation of Bcl-2 and Bcl-XL expression (Figure 3). In gastric MALT lymphoma, CagA may stimulate lymphomagenesis directly through the regulation of signal transduction, and intracellular CagA is associated with *H. pylori* I dependence. These findings represent a substantial paradigm shift compared with the classical theory of *H. pylori*-reactive T cells contributing indirectly to the development of MALT lymphoma[40].



Diamantidis MD et al. Optimal treatment for DLBCL PGL

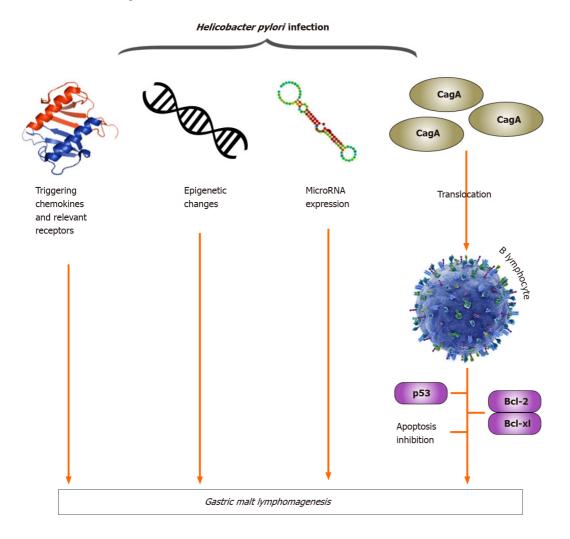


Figure 3 Helicobacter pylori infection, molecular mechanisms and gastric mucosa-associated lymphoid tissue lymphomagenesis. BCL: Bcell lymphoma; Bcl-XL: B-cell leukemia XL; CagA: Cytotoxin-associated gene A.

Other cytogenetic aberrations, often associated with one of the four main chromosomal translocations described above, include trisomies 3, 12 and/or 18, which can also present as a sole abnormality in one-fifth of the total cases. Somatic missense mutations in PIM1 and cMYC have been reported in 46% of MALT gastric lymphomas and in 30% of transformed MALT lymphomas. The majority of these genetic lesions are not MALT lymphoma specific. Aberrant somatic hypermutation can still be encountered in indolent lymphomas, such as MALT, but not at the extent noted in DLBCL lymphomas[40,55,58]. Interestingly, the loss of the chemokine receptor CXCR4 and the upregulation of CXCR7 have been associated with the progression of gastric MALT lymphoma to DLBCL lymphoma[59]. Furthermore, lower expression of the microRNA miR-34a has also been linked to the transition from MALT to DLBCL lymphoma[54]. Finally, among the proposed pathogenetic etiologies for H. pylorinegative MALT lymphoma cases, genetic alterations in NF-KB signaling are the main hypothesis<sup>[53]</sup>.

# SCIENTIFIC GAPS

While gastric DLBCLs are frequent, sufficient data to guide optimal care are still limited. In the past, gastrectomy was the treatment of choice for these patients. Nevertheless, due to the observed high morbidity rates linked with this procedure, novel therapeutic approaches have emerged, such as radiation and combination chemotherapy. Hence, until today, a multidisciplinary approach has been applied, including chemotherapy, surgery, radiotherapy or a combination of these modalities.

Today, immunochemotherapy with R-CHOP is the most acceptable option for treating gastric DLBCL, as for nodal DLBCL. R-CHOP was established as a standard approach for DLBCL patients; in the study of patients aged 60-80 years, the rate of



complete response (CR) was significantly higher in the group that received R-CHOP *vs* CHOP[60]. Since then, a few small, heterogeneous, retrospective studies have attempted to determine the optimal management of gastric DLBCL, investigating the role of immunochemotherapy, surgery and radiation in patient outcomes[16-21,23,61-63].

Significant advances in diagnosis, treatment and response assessment options over the last years have been made in the field of high-grade lymphomas. Molecular characterization of DLBCL has also described 3 major lymphoma subgroups that correlate with distinct biological and clinical behavior (ABCs, GCBs, double hit lymphomas), supporting the rationale for distinct therapeutic options[48]. However, these advances were extracted from nodal DLBCL, while the intrinsic pathogenesis of primary gastric DLBCL is unclear, and similar studies on this particular type of lymphoma are lacking.

The heterogeneity of the various clinical retrospective studies investigating the outcomes of patients with DLBCL PGLs is impressive. For example, these studies differ in the number of patients, in the time intervals when each therapeutic approach was applied, or in the type of therapeutic approaches compared. Other studies calculate surgery alone and other surgeries with chemotherapy and/or radiotherapy, without separating treatment subgroups of patients. Some researchers place all DLBCL patients together into the statistical analysis, regardless of the anatomical site (stomach, intestine). Hence, comparisons are difficult and not head-to-head. Thus, evidence-based conclusions cannot be drawn, and these results should be regarded with caution.

Finally, the use of various staging systems combined with the variability in the applied procedures for staging make the application of meaningful comparisons among the published series difficult.

# CURRENT AND FUTURE RESEARCH — FRONTIER PERSPECTIVE

We retrospectively evaluated the clinical profile and the patterns of outcome among patients who were treated after the diagnosis of aggressive, B-cell, primary endocrine lymphoma (another type of extranodal lymphoma). The patients were diagnosed with either primary testicular lymphoma, primary thyroid lymphoma (PTHL), or PAL. Better outcomes were observed in patients with PTHL for whom the median OS had not been reached until the end date of the study, whereas the PAL group had the worst prognosis[35].

To better understand the nature and outcome of extranodal DLBCL PGL, we described patients' and disease characteristics and assessed trends in treatment options, management and outcome among 159 newly diagnosed patients with primary gastric DLBCL who were seen in our institutions in the years 1971-2017.

Previously, we retrospectively evaluated the trends in clinical presentation, management and outcome among 165 consecutive patients with biopsy-proven primary gastric DLBCL who were seen in 1980-2014. The study cohort was divided into two subgroups based on the era of treatment (CHOP *vs* R-CHOP, before and after the initiation of rituximab). A better outcome after immunochemotherapy (R-CHOP) was observed comparatively[64].

Our novel manuscript and update of the same cohort of patients will be sent for peer review within the next 2 mo (under preparation). We have been preparing and analyzing it for years, focusing on the proper methodology and aiming to correct the perils and pitfalls seen in other relevant studies in the past. We will still have the same conclusion that a better outcome has been noted for the R-CHOP patient cohort, as in the past[64]. However, there are individual variations of the results regarding the OS and freedom from progression (FFP) time intervals, which will be analyzed accordingly, now that a longer follow-up of the patients has been achieved.

The term FFP is based on the strict scientific definition for this type of lymphoma and is preferable to define the aforementioned important endpoint for retrospective clinical studies. PFS has disadvantages in nonrandomized studies because in such studies, there is a lack of specific or concrete criteria for the comparison between time intervals (fixed check points), necessary for the re-evaluation of the disease and the definition of relapse in a similar way (for example, with CT or MRI). However, the term PFS is more widely used in the literature in an equivalent meaning for these lymphomas without being absolutely accurate or to the point in a strict scientific sense. We especially focused on defining FFP accurately, as this is crucial for this novel study. FFP for our novel update will be measured from the initiation of the first treatment until relapse or until death or until the last day of the study for the non

Zaishidena® WJG | https://www.wjgnet.com

relapsed patients or until the day of the last follow-up for the censored patients (lost to follow-up).

Per-protocol analysis will be used in our clinical research compared to intention-totreat analysis. The latter is considered a better marker of treatment efficacy for prospective, randomized studies and not for retrospective studies.

Finally, lymphoma-specific survival, another important endpoint, will be measured from diagnosis until the time of death from lymphoma. The number of patients who died from causes other than lymphoma was not calculated at this endpoint. As the long-year follow-up continued, we noted a proportion of our patients dying from lymphoma but also other patients dying from causes other than lymphoma. This analysis is important because it attributes the specific hazard ratio to DLBCL gastric lymphoma (death risk) and separates causes of death other than lymphoma for patients who have survived longer. Importantly, when a patient died from another cause in addition to lymphoma, there was no relapse because the patient was in follow-up. Thus, the possible drug might have protected the patient from relapse, and these patients contributed to the studied time-to-event analysis.

# CONCLUSION

In conclusion, retrospective studies, despite their limitations, if conducted with the correct methodology, can provide useful clinical information for treating patients. Our research in recent years has shown that immunochemotherapy (R-CHOP) is the optimal treatment for patients with DLBCL PGLs, as it is associated with a better outcome.

# ACKNOWLEDGEMENTS

The authors would like to thank Tsangalas E for the preparation of the figures of this work.

# REFERENCES

- **Peng JC**, Zhong L, Ran ZH. Primary lymphomas in the gastrointestinal tract. J Dig Dis 2015; 16: 1 169-176 [PMID: 25678011 DOI: 10.1111/1751-2980.12234]
- 2 Juárez-Salcedo LM, Sokol L, Chavez JC, Dalia S. Primary Gastric Lymphoma, Epidemiology, Clinical Diagnosis, and Treatment. Cancer Control 2018; 25: 1073274818778256 [PMID: 29779412 DOI: 10.1177/1073274818778256]
- Milano AF. 20-Year Comparative Survival and Mortality of Cancer of the Stomach by Age, Sex, 3 Race, Stage, Grade, Cohort Entry Time-Period, Disease Duration & Selected ICD-O-3 Oncologic Phenotypes: A Systematic Review of 157,258 Cases for Diagnosis Years 1973-2014: (SEER\*Stat 8.3.4). J Insur Med 2019; 48: 5-23 [PMID: 31609640 DOI: 10.17849/insm-48-1-1-19.1]
- Rawla P, Barsouk A. Epidemiology of gastric cancer: global trends, risk factors and prevention. Prz Gastroenterol 2019; 14: 26-38 [PMID: 30944675 DOI: 10.5114/pg.2018.80001]
- 5 Zhang X, Yang J, Huang Q, Lyu J. Prognostic factors in patients with gastric adenocarcinoma using competing-risk analysis: a study of cases in the SEER database. Scand J Gastroenterol 2019; 54: 1015-1021 [PMID: 31382800 DOI: 10.1080/00365521.2019.1649456]
- Violeta Filip P, Cuciureanu D, Sorina Diaconu L, Maria Vladareanu A, Silvia Pop C. MALT 6 lymphoma: epidemiology, clinical diagnosis and treatment. J Med Life 2018; 11: 187-193 [PMID: 30364585 DOI: 10.25122/jml-2018-0035]
- 7 Ferreri AJ, Montalbán C. Primary diffuse large B-cell lymphoma of the stomach. Crit Rev Oncol Hematol 2007; 63: 65-71 [PMID: 17339119 DOI: 10.1016/j.critrevonc.2007.01.003]
- 8 Ikoma N, Badgwell BD, Mansfield PF. Multimodality Treatment of Gastric Lymphoma. Surg Clin North Am 2017; 97: 405-420 [PMID: 28325194 DOI: 10.1016/j.suc.2016.11.012]
- Fischbach W. Long-term follow-up of gastric lymphoma after stomach conserving treatment. Best Pract Res Clin Gastroenterol 2010; 24: 71-77 [PMID: 20206110 DOI: 10.1016/j.bpg.2009.12.005]
- Shirwaikar Thomas A, Schwartz M, Quigley E. Gastrointestinal lymphoma: the new mimic. BMJ 10 Open Gastroenterol 2019; 6: e000320 [PMID: 31645987 DOI: 10.1136/bmjgast-2019-000320]
- Dawson IM, Cornes JS, Morson BC. Primary malignant lymphoid tumours of the intestinal tract. 11 Report of 37 cases with a study of factors influencing prognosis. Br J Surg 1961; 49: 80-89 [PMID: 13884035 DOI: 10.1002/bjs.18004921319]
- 12 Malipatel R, Patil M, Pritilata Rout P, Correa M, Devarbhavi H. Primary Gastric Lymphoma: Clinicopathological Profile. Euroasian J Hepatogastroenterol 2018; 8: 6-10 [PMID: 29963454 DOI:



#### 10.5005/jp-journals-10018-1250]

- 13 Psyrri A, Papageorgiou S, Economopoulos T. Primary extranodal lymphomas of stomach: clinical presentation, diagnostic pitfalls and management. Ann Oncol 2008; 19: 1992-1999 [PMID: 18647965 DOI: 10.1093/annonc/mdn525]
- 14 Cheson BD, Fisher RI, Barrington SF, Cavalli F, Schwartz LH, Zucca E, Lister TA; Alliance, Australasian Leukaemia and Lymphoma Group; Eastern Cooperative Oncology Group; European Mantle Cell Lymphoma Consortium; Italian Lymphoma Foundation; European Organisation for Research; Treatment of Cancer/Dutch Hemato-Oncology Group; Grupo Español de Médula Ósea; German High-Grade Lymphoma Study Group; German Hodgkin's Study Group; Japanese Lymphorra Study Group; Lymphoma Study Association; NCIC Clinical Trials Group; Nordic Lymphoma Study Group; Southwest Oncology Group; United Kingdom National Cancer Research Institute, Recommendations for initial evaluation, staging, and response assessment of Hodgkin and non-Hodgkin lymphoma: the Lugano classification. J Clin Oncol 2014; 32: 3059-3068 [PMID: 25113753 DOI: 10.1200/jco.2013.54.8800]
- 15 Barrington SF, Mikhaeel NG, Kostakoglu L, Meignan M, Hutchings M, Müeller SP, Schwartz LH, Zucca E, Fisher RI, Trotman J, Hoekstra OS, Hicks RJ, O'Doherty MJ, Hustinx R, Biggi A, Cheson BD. Role of imaging in the staging and response assessment of lymphoma: consensus of the International Conference on Malignant Lymphomas Imaging Working Group. J Clin Oncol 2014; 32: 3048-3058 [PMID: 25113771 DOI: 10.1200/jco.2013.53.5229]
- Wang YG, Zhao LY, Liu CQ, Pan SC, Chen XL, Liu K, Zhang WH, Yang K, Chen XZ, Zhang B, 16 Chen ZX, Chen JP, Zhou ZG, Hu JK. Clinical characteristics and prognostic factors of primary gastric lymphoma: A retrospective study with 165 cases. Medicine (Baltimore) 2016; 95: e4250 [PMID: 27495029 DOI: 10.1097/MD.00000000004250]
- 17 Couto ME, Oliveira I, Domingues N, Viterbo L, Martins Â, Moreira I, Espírito-Santo A, Chacim S, Moreira C, Pereira D, Henrique R, Mariz J. Gastric Diffuse Large B-Cell Lymphoma: A Single-Center 9-Year Experience. Indian J Hematol Blood Transfus 2021; 1-5 [PMID: 33424149 DOI: 10.1007/s12288-020-01391-91
- Li X, Xia B, Guo S, Zhan Z, Zhang L, Zhao D, Wu X, Zhang Y. A retrospective analysis of primary 18 gastric diffuse large B-cell lymphoma with or without concomitant mucosa-associated lymphoid tissue (MALT) lymphoma components. Ann Hematol 2013; 92: 807-815 [PMID: 23417758 DOI: 10.1007/s00277-013-1701-9]
- Sohn BS, Kim SM, Yoon DH, Kim S, Lee DH, Kim JH, Lee SW, Huh J, Suh C. The comparison 19 between CHOP and R-CHOP in primary gastric diffuse large B cell lymphoma. Ann Hematol 2012; 91: 1731-1739 [PMID: 22752193 DOI: 10.1007/s00277-012-1512-4]
- 20 Tanaka T, Shimada K, Yamamoto K, Hirooka Y, Niwa Y, Sugiura I, Kitamura K, Kosugi H, Kinoshita T, Goto H, Nakamura S. Retrospective analysis of primary gastric diffuse large B cell lymphoma in the rituximab era: a multicenter study of 95 patients in Japan. Ann Hematol 2012; 91: 383-390 [PMID: 21822617 DOI: 10.1007/s00277-011-1306-0]
- Liu Y, Liu Y, Zhao P, Zhang Q, Liu X, Lv F, Hong X, Cao J, Xue K. Switching Fractioned R-CHOP Cycles to Standard R-CHOP Cycles Guided by Endoscopic Ultrasonography in Treating Patients with Primary Gastric Diffuse Large B-Cell Lymphoma. Cancer Manag Res 2020; 12: 5041-5048 [PMID: 32612391 DOI: 10.2147/CMAR.S260974]
- 22 Goto A, Nishikawa J, Ito S, Hideura E, Ogawa R, Hashimoto S, Okamoto T, Sakaida I. A Rapidly Developing Diffuse Large B cell Lymphoma of the Stomach. J Gastrointest Cancer 2019; 50: 657-659 [PMID: 29623599 DOI: 10.1007/s12029-018-0098-z]
- 23 Kadota T, Seo S, Fuse H, Ishii G, Itoh K, Yano T, Kaneko K, Tsukasaki K. Complications and outcomes in diffuse large B-cell lymphoma with gastric lesions treated with R-CHOP. Cancer Med 2019; 8: 982-989 [PMID: 30730104 DOI: 10.1002/cam4.1982]
- Bai Z, Li Z, Guan T, Wang L, Wang J, Wu S, Su L. Primary Gastric Diffuse Large B-Cell 24 Lymphoma: Prognostic Factors in the Immuno-Oncology Therapeutics Era. Turk J Haematol 2020; 37: 193-202 [PMID: 32160735 DOI: 10.4274/tjh.galenos.2020.2019.0332]
- Wang X, Kan Y, Chen L, Ge P, Ding T, Zhai Q, Yu Y, Wang X, Zhao Z, Yang H, Liu X, Li L, Qiu 25 L, Qian Z, Zhang H, Wang Y, Zhao H. miR-150 is a negative independent prognostic biomarker for primary gastrointestinal diffuse large B-cell lymphoma. Oncol Lett 2020; 19: 3487-3494 [PMID: 32269622 DOI: 10.3892/ol.2020.114521
- Yonezumi M, Suzuki R, Suzuki H, Yoshino T, Oshima K, Hosokawa Y, Asaka M, Morishima Y, 26 Nakamura S, Seto M. Detection of AP12-MALT1 chimaeric gene in extranodal and nodal marginal zone B-cell lymphoma by reverse transcription polymerase chain reaction (PCR) and genomic long and accurate PCR analyses. Br J Haematol 2001; 115: 588-594 [PMID: 11736940 DOI: 10.1046/j.1365-2141.2001.03158.x]
- Aleman BM, Haas RL, van der Maazen RW. Role of radiotherapy in the treatment of lymphomas of 27 the gastrointestinal tract. Best Pract Res Clin Gastroenterol 2010; 24: 27-34 [PMID: 20206106 DOI: 10.1016/j.bpg.2009.12.002
- Kang HJ, Lee HH, Jung SE, Park KS, O JH, Jeon YW, Choi BO, Cho SG. Pattern of failure and 28 optimal treatment strategy for primary gastric diffuse large B-cell lymphoma treated with R-CHOP chemotherapy. PLoS One 2020; 15: e0238807 [PMID: 32960887 DOI: 10.1371/journal.pone.0238807
- 29 Avilés A, Nambo MJ, Neri N, Huerta-Guzmán J, Cuadra I, Alvarado I, Castañeda C, Fernández R, González M. The role of surgery in primary gastric lymphoma: results of a controlled clinical trial.



Ann Surg 2004; 240: 44-50 [PMID: 15213617 DOI: 10.1097/01.sla.0000129354.31318.f1]

- Cuccurullo R, Govi S, Ferreri AJ. De-escalating therapy in gastric aggressive lymphoma. World J 30 Gastroenterol 2014; 20: 8993-8997 [PMID: 25083073]
- Olszewska-Szopa M, Wróbel T. Gastrointestinal non-Hodgkin lymphomas. Adv Clin Exp Med 2019; 31 28: 1119-1124 [PMID: 31414733 DOI: 10.17219/acem/94068]
- 32 Wakabayashi M, Sekiguchi Y, Shimada A, Ichikawa K, Sugimoto K, Tomita S, Izumi H, Nakamura N, Sawada T, Ohta Y, Komatsu N, Noguchi M. Diffuse large B-cell lymphoma solely involving bilateral adrenal glands and stomach: report of an extremely rare case with review of the literature. Int J Clin Exp Pathol 2014; 7: 8190-8197 [PMID: 25550871]
- Ceniceros-Cabrales AP, Sánchez-Fernández P. Perforated gastric diffuse large B-cell lymphoma: A 33 case report and literature review. Rev Gastroenterol Mex (Engl Ed) 2019; 84: 412-414 [PMID: 30245175 DOI: 10.1016/j.rgmx.2018.07.004]
- 34 Hassan M, Mandal AK, Sidhu JS, Cardenas LM. Gastric diffuse large B-cell lymphoma with bilateral adrenal metastasis. BMJ Case Rep 2019; 12 [PMID: 31272994 DOI: 10.1136/bcr-2019-229758]
- 35 Hatjiharissi E, Diamantidis MD, Papaioannou M, Dimou T, Chrisoulidou A, Patakiouta F, Constantinou N, Pazaitou-Panayiotou K. Long-term outcome of primary endocrine non-Hodgkin lymphomas: does the site make the difference? QJM 2013; 106: 623-630 [PMID: 23426729 DOI: 10.1093/gjmed/hct048]
- 36 Nakamura S, Sugiyama T, Matsumoto T, Iijima K, Ono S, Tajika M, Tari A, Kitadai Y, Matsumoto H, Nagaya T, Kamoshida T, Watanabe N, Chiba T, Origasa H, Asaka M; JAPAN GAST Study Group. Long-term clinical outcome of gastric MALT lymphoma after eradication of Helicobacter pylori: a multicentre cohort follow-up study of 420 patients in Japan. Gut 2012; 61: 507-513 [PMID: 21890816 DOI: 10.1136/gutjnl-2011-300495]
- Ferreri AJ, Govi S, Ponzoni M. The role of Helicobacter pylori eradication in the treatment of 37 diffuse large B-cell and marginal zone lymphomas of the stomach. Curr Opin Oncol 2013; 25: 470-479 [PMID: 23942292 DOI: 10.1097/01.cco.0000432523.24358.15]
- 38 Paydas S. Helicobacter pylori eradication in gastric diffuse large B cell lymphoma. World J Gastroenterol 2015; 21: 3773-3776 [PMID: 25852262 DOI: 10.3748/wjg.v21.i13.3773]
- Kuo SH, Yeh KH, Chen LT, Lin CW, Hsu PN, Hsu C, Wu MS, Tzeng YS, Tsai HJ, Wang HP, 39 Cheng AL. Helicobacter pylori-related diffuse large B-cell lymphoma of the stomach: a distinct entity with lower aggressiveness and higher chemosensitivity. Blood Cancer J 2014; 4: e220 [PMID: 24949857 DOI: 10.1038/bcj.2014.40]
- 40 Kuo SH, Cheng AL. Helicobacter pylori and mucosa-associated lymphoid tissue: what's new. Hematology Am Soc Hematol Educ Program 2013; 2013: 109-117 [PMID: 24319171 DOI: 10.1182/asheducation-2013.1.109]
- Kuo SH, Yeh KH, Wu MS, Lin CW, Hsu PN, Wang HP, Chen LT, Cheng AL. Helicobacter pylori 41 eradication therapy is effective in the treatment of early-stage H pylori-positive gastric diffuse large B-cell lymphomas. Blood 2012; 119: 4838-44; quiz 5057 [PMID: 22403257 DOI: 10.1182/blood-2012-01-404194]
- 42 Ollila TA, Olszewski AJ. Extranodal Diffuse Large B Cell Lymphoma: Molecular Features, Prognosis, and Risk of Central Nervous System Recurrence. Curr Treat Options Oncol 2018; 19: 38 [PMID: 29931605 DOI: 10.1007/s11864-018-0555-8]
- 43 King RL, Goodlad JR, Calaminici M, Dotlic S, Montes-Moreno S, Oschlies I, Ponzoni M, Traverse-Glehen A, Ott G, Ferry JA. Lymphomas arising in immune-privileged sites: insights into biology, diagnosis, and pathogenesis. Virchows Arch 2020; 476: 647-665 [PMID: 31863183 DOI: 10.1007/s00428-019-02698-3]
- Taal BG, Burgers JM. Primary non-Hodgkin's lymphoma of the stomach: endoscopic diagnosis and 44 the role of surgery. Scand J Gastroenterol Suppl 1991; 188: 33-37 [PMID: 1775939 DOI: 10.3109/00365529109111227
- 45 Thurner L, Hartmann S, Neumann F, Hoth M, Stilgenbauer S, Küppers R, Preuss KD, Bewarder M. Role of Specific B-Cell Receptor Antigens in Lymphomagenesis. Front Oncol 2020; 10: 604685 [PMID: 33363034 DOI: 10.3389/fonc.2020.604685]
- Bautista-Quach MA, Ake CD, Chen M, Wang J. Gastrointestinal lymphomas: Morphology, 46 immunophenotype and molecular features. J Gastrointest Oncol 2012; 3: 209-225 [PMID: 22943012 DOI: 10.3978/j.issn.2078-6891.2012.024]
- Lin P, Medeiros LJ. The impact of MYC rearrangements and "double hit" abnormalities in diffuse 47 large B-cell lymphoma. Curr Hematol Malig Rep 2013; 8: 243-252 [PMID: 23892979 DOI: 10.1007/s11899-013-0169-y
- Abramson JS. Hitting back at lymphoma: How do modern diagnostics identify high-risk diffuse 48 large B-cell lymphoma subsets and alter treatment? Cancer 2019; 125: 3111-3120 [PMID: 31287161 DOI: 10.1002/cncr.32145]
- Foukas PG, Bisig B, de Leval L. Recent advances upper gastrointestinal lymphomas: molecular updates and diagnostic implications. *Histopathology* 2021; 78: 187-214 [PMID: 33382495 DOI: 10.1111/his.14289]
- 50 Hans CP, Weisenburger DD, Greiner TC, Gascoyne RD, Delabie J, Ott G, Müller-Hermelink HK, Campo E, Braziel RM, Jaffe ES, Pan Z, Farinha P, Smith LM, Falini B, Banham AH, Rosenwald A, Staudt LM, Connors JM, Armitage JO, Chan WC. Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a tissue microarray. *Blood* 2004; 103: 275-282 [PMID: 14504078 DOI: 10.1182/blood-2003-05-1545]



- 51 Liu Y, Yu K, Li M, Zeng K, Wei J, Li X, Liu Y, Zhao D, Fan L, Yu Z, Wang Y, Li Z, Zhang W, Bai Q, Yan Q, Guo Y, Wang Z, Guo S. EZH2 overexpression in primary gastrointestinal diffuse large Bcell lymphoma and its association with the clinicopathological features. Hum Pathol 2017; 64: 213-221 [PMID: 28438623 DOI: 10.1016/j.humpath.2017.04.011]
- 52 Ishikawa E, Nakamura M, Shimada K, Tanaka T, Satou A, Kohno K, Sakakibara A, Furukawa K, Yamamura T, Miyahara R, Nakamura S, Kato S, Fujishiro M. Prognostic impact of PD-L1 expression in primary gastric and intestinal diffuse large B-cell lymphoma. J Gastroenterol 2020; 55: 39-50 [PMID: 31493237 DOI: 10.1007/s00535-019-01616-3]
- Asano N, Iijima K, Koike T, Imatani A, Shimosegawa T. Helicobacter pylori-negative gastric 53 mucosa-associated lymphoid tissue lymphomas: A review. World J Gastroenterol 2015; 21: 8014-8020 [PMID: 26185372 DOI: 10.3748/wjg.v21.i26.8014]
- 54 Vasilatou D, Sioulas AD, Pappa V, Papanikolaou IS, Triantafyllou K, Dimitriadis GD, Papageorgiou SG. The role of miRNAs and epigenetic mechanisms in primary gastric mucosa-associated lymphoid tissue lymphoma. Future Oncol 2016; 12: 1587-1593 [PMID: 27079806 DOI: 10.2217/fon-2016-0038
- Troppan K, Wenzl K, Neumeister P, Deutsch A. Molecular Pathogenesis of MALT Lymphoma. 55 Gastroenterol Res Pract 2015; 2015: 102656 [PMID: 25922601 DOI: 10.1155/2015/102656]
- Amedei A, Cappon A, Codolo G, Cabrelle A, Polenghi A, Benagiano M, Tasca E, Azzurri A, D'Elios 56 MM, Del Prete G, de Bernard M. The neutrophil-activating protein of Helicobacter pylori promotes Th1 immune responses. J Clin Invest 2006; 116: 1092-1101 [PMID: 16543949 DOI: 10.1172/jci27177]
- 57 Hamoudi RA, Appert A, Ye H, Ruskone-Fourmestraux A, Streubel B, Chott A, Raderer M, Gong L, Wlodarska I, De Wolf-Peeters C, MacLennan KA, de Leval L, Isaacson PG, Du MQ. Differential expression of NF-kappaB target genes in MALT lymphoma with and without chromosome translocation: insights into molecular mechanism. Leukemia 2010; 24: 1487-1497 [PMID: 20520640 DOI: 10.1038/leu.2010.118]
- Nakamura S, Matsumoto T. Helicobacter pylori and gastric mucosa-associated lymphoid tissue 58 lymphoma: recent progress in pathogenesis and management. World J Gastroenterol 2013; 19: 8181-8187 [PMID: 24363507 DOI: 10.3748/wjg.v19.i45.8181]
- 59 Deutsch AJ, Steinbauer E, Hofmann NA, Strunk D, Gerlza T, Beham-Schmid C, Schaider H, Neumeister P. Chemokine receptors in gastric MALT lymphoma: loss of CXCR4 and upregulation of CXCR7 is associated with progression to diffuse large B-cell lymphoma. Mod Pathol 2013; 26: 182-194 [PMID: 22936065 DOI: 10.1038/modpathol.2012.134]
- Coiffier B, Lepage E, Briere J, Herbrecht R, Tilly H, Bouabdallah R, Morel P, Van Den Neste E, 60 Salles G, Gaulard P, Reyes F, Lederlin P, Gisselbrecht C. CHOP chemotherapy plus rituximab compared with CHOP alone in elderly patients with diffuse large-B-cell lymphoma. N Engl J Med 2002; 346: 235-242 [PMID: 11807147 DOI: 10.1056/NEJMoa011795]
- Ge Z, Liu Z, Hu X. Anatomic distribution, clinical features, and survival data of 87 cases primary 61 gastrointestinal lymphoma. World J Surg Oncol 2016; 14: 85 [PMID: 26988370 DOI: 10.1186/s12957-016-0821-9
- Liu PP, Xia Y, Bi XW, Wang Y, Sun P, Yang H, Li ZM, Jiang WQ. Trends in Survival of Patients 62 with Primary Gastric Diffuse Large B-Cell Lymphoma: An Analysis of 7051 Cases in the SEER Database. Dis Markers 2018; 2018: 7473935 [PMID: 30410635 DOI: 10.1155/2018/7473935]
- 63 Taal BG, Burgers JM, van Heerde P, Hart AA, Somers R. The clinical spectrum and treatment of primary non-Hodgkin's lymphoma of the stomach. Ann Oncol 1993; 4: 839-846 [PMID: 8117603 DOI: 10.1093/oxfordjournals.annonc.a058390]
- 64 Hatjiharissi E, Diamantidis M, Papadopoulou A, Chatzileontiadou S, Gerofotis A, Pouptsis A, Karabatzakis N, Pentidou K, Patakiouta F, Konstantinou N, Papaioannou M. Long-term follow-up of patients with non-Hodgkin primary diffuse large B-cell lymphoma of the stomach: Better outcome after immunochemotherapy. Hematol Oncol 2017; 35: 377 [DOI: 10.1002/hon.2439\_136]



WÜ

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5946-5957

DOI: 10.3748/wjg.v27.i35.5946

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

ORIGINAL ARTICLE

# **Basic Study** Proteomics identifies a novel role of fibrinogen-like protein 1 in Crohn's disease

Xue-Liang Sun, Li-Chao Qiao, Jing Gong, Ke Wen, Zhi-Zhong Xu, Bo-Lin Yang

ORCID number: Xue-Liang Sun 0000-0001-8520-6937; Li-Chao Qiao 0000-0002-0292-2936; Jing Gong 0000-0001-8423-3706; Ke Wen 0000-0003-3925-6567; Zhi-Zhong Xu 0000-0002-1725-7190; Bo-Lin Yang 0000-0002-2474-4085.

Author contributions: Sun XL and Yang BL contributed to designing the study; Sun XL, Qiao LC and Gong J contributed to performing the experiments; Wen K and Xu ZZ contributed to sample collection and statistical analysis; Sun XL and Qiao LC contributed to manuscript drafting; all authors made critical revisions to the manuscript and approved the final version of the article to be published.

Supported by National Natural Science Foundation of China, No. 82074431; The Open Projects of the Discipline of Chinese Medicine of Nanjing University of Chinese Medicine Supported by the Subject of Academic Priority Discipline of Jiangsu Higher Education Institutions, No. ZYX03KF034; and Suzhou Municipal Science and Technology Bureau, No. SYSD2020253 and No. SS202085.

#### Institutional review board

statement: The study was approved by the Ethics Committee of the Affiliated Hospital of Nanjing University of Chinese

Xue-Liang Sun, Li-Chao Qiao, Jing Gong, Bo-Lin Yang, First Clinical Medical College, The Affiliated Hospital of Nanjing University of Chinese Medicine, Nanjing 210029, Jiangsu Province, China

Xue-Liang Sun, Ke Wen, Zhi-Zhong Xu, Department of Colorectal Surgery, Suzhou TCM Hospital Affiliated to Nanjing University of Chinese Medicine, Suzhou 215000, Jiangsu Province, China

Bo-Lin Yang, Department of Colorectal Surgery, Jiangsu Province Hospital of Chinese Medicine, Nanjing 210029, Jiangsu Province, China

Corresponding author: Bo-Lin Yang, MD, Chief Doctor, First Clinical Medical College, The Affiliated Hospital of Nanjing University of Chinese Medicine, No. 155 Hanzhong Road, Nanjing 210029, Jiangsu Province, China. yfy0051@njucm.edu.cn

# Abstract

# BACKGROUND

Crohn's disease (CD) is an incurable intestinal disorder with unclear etiology and pathogenesis. Currently, there is a lack of specific biomarkers and drug targets for CD in clinical practice. It is essential to identify the precise pathophysiological mechanism of CD and investigate new therapeutic targets.

#### AIM

To explore a new biomarker and therapeutic target for CD and verify its role in the CD pathological mechanism.

# **METHODS**

Proteomics was performed to quantify the protein profile in the plasma of 20 CD patients and 20 matched healthy controls. Hub genes among the selected differentially expressed proteins (DEPs) were detected via the MCODE plugin in Cytoscape software. The expression level of one hub gene with an immunoregulatory role that interested us was verified in the inflamed intestinal tissues of 20 CD patients by immunohistochemical analysis. After that, the effects of the selected hub gene on the intestinal inflammation of CD were identified in a CD cell model by examining the levels of proinflammatory cytokines by enzymelinked immunosorbent assays and the expression of the NF-KB signalling pathway by quantitative real-time PCR analysis and Western blot assays.



Medicine (2018NL-171-02).

Conflict-of-interest statement: To the best of our knowledge, no conflict of interest exists.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Unsolicited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: China

#### Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Received: March 5, 2021 Peer-review started: March 5, 2021 First decision: April 5, 2021 Revised: April 7, 2021 Accepted: August 10, 2021 Article in press: August 10, 2021 Published online: September 21, 2021

P-Reviewer: Kumar S S-Editor: Gong ZM L-Editor: Wang TQ P-Editor: Xing YX



#### RESULTS

Thirty-five DEPs were selected from 393 credible proteins identified by proteomic analysis. Among the DEPs, fibrinogen-like protein 1 (FGL1), which attracted our attention due to its function in the regulation of the immune response, had 1.722fold higher expression in the plasma of CD patients and was identified as a hub gene by MCODE. Furthermore, the expression of *FGL1* in the intestinal mucosal and epithelial tissues of CD patients was also upregulated (P < 0.05). In vitro, the mRNA levels of FGL1 and NF- $\kappa B$ ; the protein expression levels of FGL1, IKK $\alpha$ , IKKβ, p-IKK $\alpha$ /β, p-I $\kappa$ B $\alpha$ , and p-p65; and the concentrations of the proinflammatory cytokines IL-1 $\beta$ , IL-6, IL-17, and TNF- $\alpha$  were increased (P < 0.05) after stimulation with lipopolysaccharide, which were reversed by knockdown of FGL1 with siRNA transfection (P < 0.05). Conversely, FGL1 overexpression enhanced the abovementioned results (P < 0.05).

#### **CONCLUSION**

FGL1 can induce intestinal inflammation by activating the canonical NF-κB signalling pathway, and it may be considered a potential biomarker and therapeutic target for CD.

**Key Words:** Crohn's disease; Fibrinogen-like protein 1; Proteomics; NF-κB pathway

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: In this study, fibrinogen-like protein 1 (FGL1) was identified to be significantly upregulated in the plasma and intestinal mucosa of Crohn's disease (CD) patients. In vitro, silencing FGL1 downregulated the levels of the proinflammatory cytokines IL-1 $\beta$ , IL-6, IL-17, and TNF- $\alpha$ . Furthermore, FGL1 knockdown suppressed the mRNA expression of NF- $\kappa B$  and the protein levels of IKK $\alpha$ , IKK $\beta$ , p-IKK $\alpha/\beta$ , p- $I\kappa B\alpha$ , and p-p65. These results could be reversed by the overexpression of FGL1. Taken together, these data suggest that FGL1 may induce intestinal inflammation by activating the canonical NF- $\kappa$ B signalling pathway and has the potential to be a therapeutic target for CD.

Citation: Sun XL, Qiao LC, Gong J, Wen K, Xu ZZ, Yang BL. Proteomics identifies a novel role of fibrinogen-like protein 1 in Crohn's disease. World J Gastroenterol 2021; 27(35): 5946-5957

URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5946.htm DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5946

# INTRODUCTION

Crohn's disease (CD) is a chronic, idiopathic intestinal inflammatory disease affecting any segment of the gastrointestinal tract. Although CD is believed to be a result of an imbalanced interaction among genetic susceptibility, environmental factors, the intestinal microflora, and the immune system, the precise pathogenesis is still not entirely clear<sup>[1]</sup>. Consequently, CD remains incurable even though great advancement has been achieved in medical therapy. Symptoms evolving in a relapsing and remitting manner indicate that CD has a progressive disease course that may induce complications, such as abscess, fistula, and stricture development. Eventually, up to 70% of CD patients require at least one intestinal surgery over their lifetime[2]. Targeted therapy is anticipated to change the natural course of CD, and even to cure it.

Currently, anti-tumor necrosis factor (TNF) agents (infliximab, adalimumab, and certolizumab pegol) are the most potent drugs for inducing and maintaining remission of CD. Unfortunately, anti-TNF treatment failure is common. Primary non-response occurs in 21.9% of infliximab-treated CD patients and 26.8% of adalimumab-treated patients<sup>[3]</sup>. More than 60% of patients treated with infliximab or adalimumab do not achieve deep remission[3]. These data indicate that increased TNF- $\alpha$  levels may be the result of an immunoinflammatory response instead of the cause. Vedolizumab blocking the  $\alpha 4\beta 7$  integrin can induce endoscopic remission in approximately one-

third of CD patients at week 52[4]. The decreased long-term efficacy of biologic medications makes it urgent to investigate new therapeutic targets for CD.

Omics techniques, including genomics, metabolomics, and proteomics, have been applied to explore potential biomarkers and targets for CD in recent years. It is widely known that cellular function and biological behaviour are primarily regulated by proteins. The protein domain is likely the most ubiquitously affected in disease development, treatment response, and physical recovery. Hence, it is promising to reveal the crucial changes in CD pathogenesis and discover novel drug targets by proteomics directly profiling protein expression. Proteomic techniques are classified into three major stages: Discovery, verification, and validation. Currently, the application of proteomics in CD remains in the initial discovery phase<sup>[5]</sup>

In the present study, we applied proteomics to identify differentially expressed proteins (DEPs) in the plasma of CD patients in an attempt to discover a potential biomarker and therapeutic target for CD. Our data showed that fibrinogen-like protein 1 (FGL1) was significantly upregulated in the plasma of CD patients. FGL1, also known as hepassocin or hepatocyte-derived fibrinogen-related protein 1 (HFREP1), is a hepatocyte-secreted protein that belongs to the fibrinogen family[6]. However, FGL1 lacks a platelet-binding site, a cross-linking region, and a thrombin-sensitive site, which are crucial for fibrin clot formation. Several studies have demonstrated that FGL1 can regulate immune systems to induce inflammatory response and tumor immune evasion[7,8]. To date, whether FGL1 is correlated with the development of CD remains unclear. Therefore, we further verified the expression of FGL1 in intestinal tissues of CD patients and validated its crucial role in the pathogenesis of CD in vitro.

#### MATERIALS AND METHODS

#### **Clinical samples**

Plasma samples were collected from 20 treatment-naive patients with CD and 20 ageand sex-matched healthy individuals between July 2017 and August 2018. The protein profiles in the plasma were analysed by tandem mass tag (TMT)-based quantitative proteomics. Paraffin-embedded mucosal biopsy specimens from an additional 20 treatment-naive patients with active CD and 20 matched healthy individuals undergoing colonoscopy screening were obtained for immunohistochemical examination. The protocols of this study were approved by the ethics committee of the Affiliated Hospital of Nanjing University of Chinese Medicine (2018NL-171-02). All patients provided informed consent.

#### TMT-based quantitative proteomics

Plasma samples were homogenized in sodium dodecyl sulfate (SDS) lysis buffer. Centrifugation was performed to collect the supernatant. Total protein concentrations were quantified using a bicinchoninic acid (BCA) assay (Thermo Scientific, United States). Protein extracts were reduced with reducing buffer (10 mmol/L dithiothreitol, 8 mol/L urea, and 100 mmol/L tetraethylammonium bromide (TEAB), pH 8.0) at 60 °C for 1 h. All samples were alkylated with iodoacetamide for 40 min at room temperature in the dark. After centrifugation, the protein pellets were digested with TEAB (100 mmol/L) and sequencing-grade trypsin (1  $\mu$ g/ $\mu$ L) at 37 °C for 12 h.

For TMT labelling, 100 µL of protein sample was incubated with a mixed solution of 41 µL of TMT labelling reagent (Thermo Fisher Scientific, United States) and 41 µL of anhydrous acetonitrile for 1 h at room temperature. The reaction was terminated with  $8 \ \mu L$  of 5% hydroxylamine. The samples from the CD patients were labelled with TMT-130 and TMT-131, while those from the healthy controls were labelled with TMT-126 and TMT-127.

The TMT-labelled peptides were eluted by using an Agilent Zorbax Extend-C18 column (2.1 mm × 150 mm, 5 µm) and fractionated with a high-performance liquid chromatography (HPLC) system at a flow rate of 300  $\mu$ L/min. The elution gradient was set to 98%, 95%, 75%, 60%, and 10%. The collected peptides were loaded on a reverse-phase trap column (C18, 100 µm × 20 mm, Thermo Fisher Scientific, United States) and enriched on an analysis column (C18, 75 µm × 150 mm, Thermo Fisher Scientific, United States) following redissolution in nano-HPLC buffer (HPLC water containing 0.1% formic acid). The flow rate was 300 nL/min, and the linear elution gradient was set as 5%, 30%, 50%, and 100%.

For mass spectrometry (MS) survey scans, the ion spray voltage, interface heating temperature, MS resolution, and ion population were set to 1,800 V, 250 °C, 70000, and  $1 \times 10^6$ , respectively. The precursor ion was acquired at 300-1600 m/z. A maximum of



10 precursors were selected for higher-energy collisional dissociation with analysis in an LTQ Orbitrap Velos Pro (Thermo Fisher Scientific, United States), and the normal chemical energy was 32%. For MS/MS detection, the tandem MS resolution, ion population, ion maximum injection time, and dynamic exclusion time were set to 17500, 2 × 10<sup>5</sup>, 80 ms, and 30 s, respectively.

#### Quantitative proteomic analysis

The raw proteomic data were analysed using Proteome Discoverer software (version 2.2, Thermo Fisher Scientific, United States) and searched against the UniProtKB database (Hunam, 2015-09, 88473 sequences). Andromeda was used as the search engine with the following parameters: (1) Homo sapiens taxonomy; (2) Q Exactive plus as instrument type; (3) Trypsin as the proteolytic enzyme, with two missed cleavages allowed; (4) TMT 6 plex and cysteine carbamidomethylations as fixed modifications; (5) Oxidation of methionine as a variable modification; (6) 20 ppm as the MS tolerance; and (7) Seven amino acids as minimum cut-off for peptide length. A false discovery rate (FDR) of less than 1% was set to refine the results.

For quantitative analysis, the TMT reporter ion intensity of each protein was analysed using Proteome Discoverer software. Proteins with empty values were discarded. Student's t test was performed to examine the difference in each protein between the two groups with Perseus software. Proteins with a fold change > 1.5 or < 0.67 and a *P* value < 0.05 were considered to be DEPs.

#### **Bioinformatics analysis**

Genes of DEPs were visualized in Cytoscape software (version 3.7.2), in which the MCODE plugin was used to select significant modules for identification of hub genes. Subsequently, the hub genes were input into the Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) database (https://string-db.org/) to construct a protein-protein interaction (PPI) network. The Database for Annotation, Visualization and Integrated Discovery (DAVID) database (https://david.ncifcrf.gov/) was applied for gene ontology (GO) enrichment analysis. Reactome pathway analysis ( https://www.reactome.org/) was performed for pathway enrichment analysis.

#### Immunohistochemical staining

Immunohistochemical staining was implemented to detect the expression of FGL1 in inflamed intestinal tissues of CD patients and normal intestinal biopsies. Mucosal biopsy specimens were fixed in 10% neutral formalin for 24 h. Afterwards, they were embedded in paraffin and cut into 5 µm sections. The sections were deparaffinized and rehydrated and then incubated in citrate buffer (pH 6.0) for antigen retrieval. After endogenous peroxidase activity was quenched with 3% hydrogen peroxide, the samples were incubated in 1% bovine serum albumin (BSA) to block non-specific immunoglobulin binding. Subsequently, the slides were incubated with an anti-FGL1 antibody (1:200 dilution, 16000-1-AP, Proteintech, United States) at 4 °C overnight. Following washing with phosphate buffered saline (PBS), the slides were incubated with a secondary IgG antibody (1:1000 dilution, ab6721, Abcam, United Kingdom) at room temperature for 1 h, counterstained with haematoxylin, and stained with a diaminobenzidine kit (DAB, Beyotime, China). All the sections were visualized under a light microscope (Nikon 80i, Japan). ImageJ software (version 1.52) was used to calculate the integrated optical density (IOD) values.

#### Cell culture and treatment

The human colonic adenoma cell line HT-29 (ATCC, United States) was cultured with Dulbecco's modified Eagle's medium, supplemented with 10% fetal bovine serum, 100 µg/mL penicillin, and 100 U/mL streptomycin at 37 °C with 5% CO<sub>2</sub>. The HT-29 cells were stimulated with 100 ng/mL lipopolysaccharide (LPS, Sigma, United States) to establish a cell model of intestinal inflammation. To uncover the impact of FGL1 on intestinal inflammation, the HT-29 cells were transfected with FGL1 siRNA and plasmid DNA (Nanjing KeyGen Biotech Co., Ltd., China) before stimulation with LPS. The transfection efficiency was determined by examining the mRNA expression of FGL1.

#### Quantitative real-time PCR analysis

Total RNA in HT-29 cells was extracted using a TRIzol reagent kit (Takara, Japan) according to the manufacturer's instructions. A PrimeScript RT reagent kit (Takara, Japan) was used for reverse transcription of the extracted RNA into cDNA. Quantitative real-time PCR (qRT-PCR) was conducted to detect the mRNA expression



of *FGL1* and *NF*- $\kappa$ *B* by using a SYBR green kit (Takara, Japan). The housekeeping gene  $\beta$ -actin was used for normalization to an endogenous reference. The relative gene expression was evaluated by using the  $2^{-\Delta\Delta Ct}$  method. The sequences of the PCR primers are as follows: FGL1-forward: 5'-ATGGCAAAGGTGTTCAGTTTCA-3', reverse: 5'-ACAATCTGCATACTGCCTCTTG-3'; NF-κB-forward: 5'-GAAGCACGAATGACAGAGGC-3', reverse: 5'-GCTTGGCGGATTAGCTCTTTT-3'; and β-actin-forward: 5'-CATGTACGTTGCTATCCAGGC-3', reverse: 5'-CTCCTTAAT-GTCACGCACGAT-3'.

#### Enzymelinked immunosorbent assay

The levels of the proinflammatory cytokines IL-1 $\beta$ , IL-6, IL-17, and TNF- $\alpha$  (Sigma, United States) in the culture medium collected after 48 h were examined by enzyme linked immunosorbent assay (ELISA) according to the manufacturer's protocol.

#### Western blot assay

Cells lysed with radioimmunoprecipitation assay (RIPA) lysis buffer were centrifuged at 12000 g for 20 min at 4 °C. Protein concentrations in the collected supernatant were quantified with a BCA assay kit. After equal amounts of protein (20 µg/well) were loaded and separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE), they were transferred onto polyvinylidene difluoride (PVDF) membranes and incubated in 5% BSA for 1 h at room temperature. The membranes were washed with Tris-borate saline containing 0.1% Tween-20 (TBST) and were incubated with primary antibodies against FGL1 (1:1000 dilution, 16000-1-AP, Proteintech, United States), IKKα (1:1000 dilution, ab32041, Abcam, United Kingdom), IKKβ (1:1000 dilution, ab32135, Abcam), p-IKKα/β (1:1000 dilution, ab194528, Abcam), ΙκΒα (1:1000 dilution, ab32518, Abcam), p-ΙκΒα (1:1000 dilution, ab133462, Abcam), NF-кB (p65, 1:1000 dilution, ab32536, Abcam), p-p65 (1:1000 dilution, ab76302, Abcam) and  $\beta$ -actin (1:1000 dilution, 20536-1-AP, Proteintech) at 4 °C overnight. The membranes were washed with TBST again and incubated with a horseradish peroxidase (HRP)-conjugated secondary antibody for 1 h at room temperature. The blots were imaged using enhanced chemiluminescence (ECL). ImageJ software was used to calculate the protein signal grey values.

#### Statistical analysis

All data were statistically analysed with SPSS 22.0 (SPSS Inc., United States). Continuous variables with a normal distribution are summarized using the mean ± SD, which in a skewness distribution are expressed as the median with range. The Mann-Whitney U test, Student's t tests, and chi-square test were performed to compare numerical variables and categorical variables as appropriate. One-way analysis of variance was used for multi-group comparisons. A two-sided P value < 0.05 was considered statistically significant.

#### RESULTS

#### Patients' characteristics

Twenty treatment-naive CD patients and 20 healthy controls were recruited for plasma proteomic analysis. The diagnostic criteria for CD referred to the clinical guidelines of the American College of Gastroenterology (ACG)[9]. Thirteen males and seven females with a median age of 20.5 (14-43) years were included in the CD group, and eleven males and nine females with a median age of 24.5 (18-46) years were included in the normal control group. Baseline demographic characteristics were comparable between the two groups (P > 0.05).

Colonoscopic biopsy specimens from an additional 20 treatment-naive patients with active CD and 20 healthy controls were used for immunohistochemical staining. There was no significant difference in sex distribution, age, or biopsy site between the two groups (P > 0.05). The baseline clinical characteristics of patients for plasma proteomic detection and immunohistochemical analysis are presented in Tables 1 and 2, respectively.

#### FGL1 is significantly upregulated in plasma proteomic analysis

Plasma samples in each group were randomly divided into four subclusters. A total of 393 credible proteins were identified by proteomic analysis, among which 35 had differential expression between the two groups (Figure 1A). Among the DEPs, FGL1



Table 1 Clinical characteristics of patients for plasma proteomic analysis						
Item	Crohn's disease ( <i>n</i> = 20)	Normal control ( <i>n</i> = 20)	P value			
Sex						
Male	13	11	0.519			
Female	7	9				
Median age (range), yr	20.5 (14-43)	24.5 (18-46)	0.069			
Disease location in the endoscopy						
Ileum	6	N/A				
Colon	6	N/A				
Ileocolon	8	N/A				

N/A: Not applicable.

Table 2 Clinical characteristics of patients for immunohistochemical assay						
Item	Crohn's disease ( <i>n</i> = 20)	Normal control ( <i>n</i> = 20)	P value			
Sex						
Male	15	13	0.731			
Female	5	7				
Age (mean ± SD), yr	27.1 ± 7.9	$25.6 \pm 4.5$	0.465			
Biopsy site						
Terminal ileum	8	5	0.832			
Ascending colon	2	1				
Transverse colon	2	2				
Descending colon	2	4				
Sigmoid colon	4	6				
Rectum	2	2				

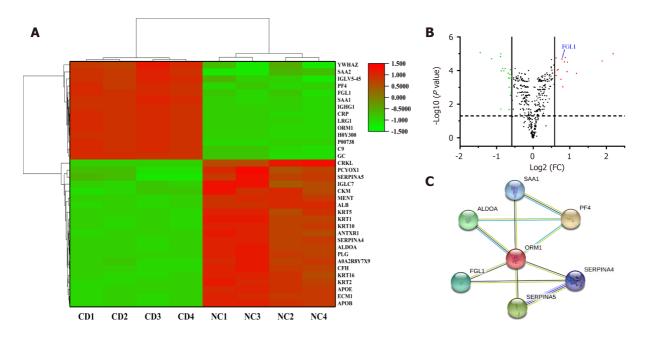
SD: Standard deviation.

attracted our attention because of its function in the regulation of the immune response. The expression level of FGL1 in the plasma of CD patients was 1.722-fold greater than that in healthy people (Figure 1B). Three MCODE modules were established to screen hub genes *via* Cytoscape software. As *FGL1* was contained in the 3rd module, the genes in this module were used for further bioinformatics analysis. Figure 1C shows the PPI network of the genes. GO enrichment analysis showed that the genes were involved in the biological processes of platelet degranulation, acute phase response, platelet activation, and negative regulation of endopeptidase activity, and the molecular functions of heparin binding and serine-type endopeptidase inhibitor activity. Reactome pathway analysis demonstrated that the genes were related to the common pathway of fibrin clot formation and the pathways of platelet degranulation, peptide ligand-binding receptors, haemostasis, G alpha (i) signalling events, and innate immune system.

#### FGL1 expression is increased in intestinal tissues of CD patients

Immunohistochemical analysis was performed to verify the expression of FGL1 in the intestinal tissues of CD patients. The results demonstrated that the FGL1 levels in the intestinal mucosal and epithelial tissues were higher than those in the normal intestinal tissues (P < 0.01, Figure 2).

Sun XL et al. FGL1 in Crohn's disease



**Figure 1 Fibrinogen-like protein 1 expression in the plasma of Crohn's disease patients.** A: Heat map showing 35 differentially expressed proteins between Crohn's disease (CD) patients and healthy individuals, among which fibrinogen-like protein 1 (FGL1) expression was upregulated in the CD group; B: The FGL1 expression level in the plasma of CD patients was 1.722-fold greater than that in healthy people; C: Protein-protein interaction network of an MCODE module containing *FGL1* as a hub gene. CD: Crohn's disease; NC: Normal control; FGL1: Fibrinogen-like protein 1.

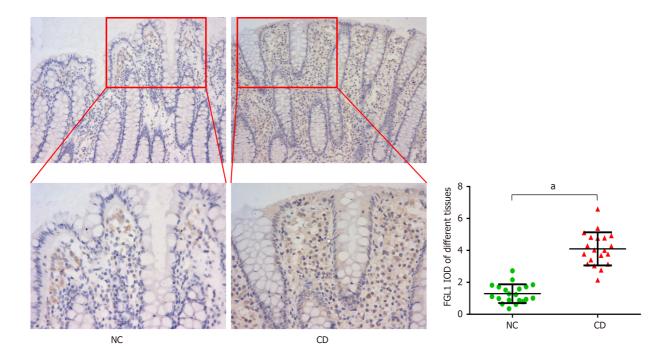
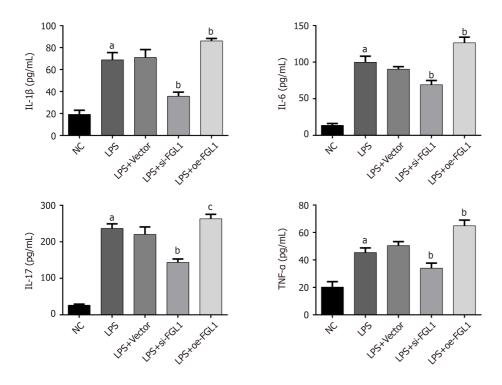


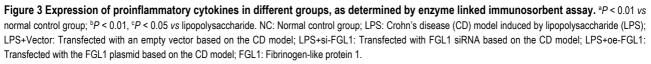
Figure 2 Fibrinogen-like protein 1 expression, as determined by immunohistochemical analysis (× 200, × 400), was increased in the intestinal mucosal and epithelial tissues of Crohn's disease patients. <sup>a</sup>P < 0.01 vs normal control group. NC: Normal control group; CD: Crohn's disease group; FGL1: Fibrinogen-like protein 1.

# FGL1 mediates the expression of proinflammatory cytokines in intestinal epithelial cells

To investigate the regulation of intestinal inflammation by FGL1, proinflammatory cytokines were detected by ELISA after *FGL1* siRNA and plasmids were transfected into HT-29 cells. After LPS stimulation, the IL-1 $\beta$ , IL-6, IL-17, and TNF- $\alpha$  levels were significantly upregulated. FGL1 knockdown reversed the expression of IL-1 $\beta$ , IL-6, IL-17, and TNF- $\alpha$ , while overexpression of FGL1 elevated the levels of the four proinflammatory cytokines (*P* < 0.05, Figure 3).

Zaisbidena® WJG | https://www.wjgnet.com





#### FGL1 activates the NF-kB signalling pathway

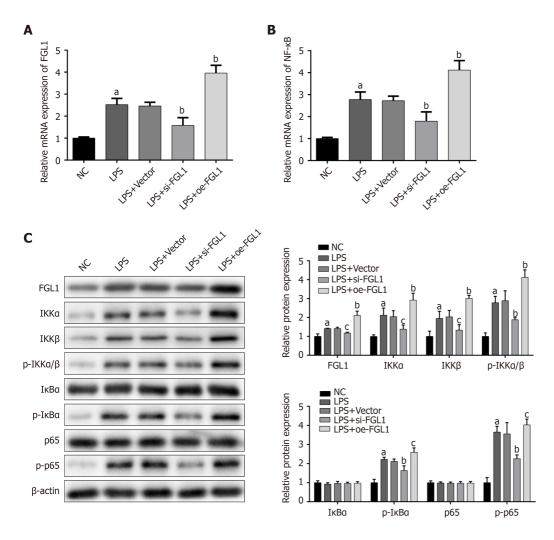
Given that NF- $\kappa$ B plays a fundamental role in the intestinal inflammation of CD, the modulation of the NF- $\kappa$ B signalling pathway by FGL1 was investigated. The mRNA expression levels of *FGL1* and *NF*- $\kappa$ B were increased by LPS stimulation. After intervention with *FGL1* siRNA, the mRNA expression levels of *FGL1* and *NF*- $\kappa$ B were both downregulated, while the mRNA levels were enhanced following FGL1 overexpression with plasmid transfection (*P* < 0.01, Figure 4A and B).

The exact mechanism by which FGL1 regulates the NF-κB signalling pathway was revealed by Western blot assay. The protein levels of FGL1, IKKα, IKKβ, p-IKKα/β, p-IκBα, and p-p65 were upregulated in HT-29 cells stimulated with LPS (P < 0.05). *FGL1* gene knockdown inhibited the protein expression of FGL1 and downregulated the protein expression of IKKα, IKKβ, p-IKKα/β, p-IκBα, and p-p65 (P < 0.05). Conversely, the overexpression of the *FGL1* gene enhanced the protein expression of FGL1, IKKα, IKKβ, p-IκBα, and p-p65 (P < 0.05). Conversely, the overexpression of the *FGL1* gene enhanced the protein expression of FGL1, IKKα, IKKβ, p-IκBα, and p-p65 (P < 0.05).

#### DISCUSSION

In the present study, we took advantage of proteomics for a large-scale screen of DEPs between the plasma of CD patients and healthy people. The expression of *FGL1*, a hub gene among the DEPs, was increased in the plasma of CD patients, which was verified in intestinal mucosal and epithelial tissues. Furthermore, FGL1 was validated to exacerbate the inflammatory response in intestinal epithelial cells by activating the NF- $\kappa$ B signalling pathway.

At present, knowledge about the etiology and pathogenesis of CD is limited, which makes it incurable. Hence, it is essential to detect potential drug targets for CD. As proteins are directly involved in nearly all pathophysiological processes, proteomics has become a hotspot tool for the discovery of novel biomarkers or therapeutic targets for CD, differential diagnosis between CD and ulcerative colitis, and disease stratification by examining and quantifying thousands of proteins encoded by the genome in a holistic manner[10-13]. To our knowledge, the present study revealed for the first time by proteomics that FGL1 may be a key contributor to CD onset and progression.



**Figure 4 Impact of fibrinogen-like protein 1 on the NF-kB signalling pathway.** A and B: Effect of knockdown or overexpression of fibrinogen-like protein 1 (FGL1) on the mRNA expression of *FGL1* and *NF-kB*, as determined by quantitative real-time PCR assay; C: Effect of FGL1 on the protein expression of related proteins in the canonical NF-kB signalling pathway, as determined by Western blot assay.  $^{a}P < 0.01$  vs NC;  $^{b}P < 0.01$ ,  $^{c}P < 0.05$  vs LPS. NC: Normal control group; LPS: Crohn's disease (CD) model induced by LPS; LPS+Vector: Transfected with an empty vector based on the CD model; LPS+si-FGL1: Transfected with the FGL1 plasmid based on the CD model; LPS: Lipopolysaccharide; FGL1: Fibrinogen-like protein 1.

Accumulating evidence has demonstrated that FGL1 plays a prominent role in the pathogenesis of various diseases, including hepatocellular carcinoma, gastric cancer, lung cancer, diabetes mellitus, and obesity[14-19]. Additionally, FGL1 is also considered a potential biomarker and drug target in certain inflammatory conditions. FGL1 may promote liver injury-induced inflammation *via* the IL-6/STAT3 signalling pathway[20]. Proteomics revealed that FGL1 is a specific biomarker for predicting the progression of rheumatoid arthritis[7]. This finding displays a fundamental role of FGL1 in regulating immune-mediated inflammation.

Pierre and colleagues have demonstrated that CD relapse is correlated with the innate immune response of the liver[21]. Given that FGL1 is a liver-derived protein and is involved in the innate immune system pathway, we hypothesize that FGL1 may influence the pathophysiology of CD based on the evidence of increased expression of FGL1 in the plasma and intestinal tissues of CD patients. Although FGL1 has been demonstrated to be a potent target for cancer immunotherapy, its precise role in CD therapy is unknown[8]. To unravel the mystery, a cell experiment was designed in the current study that focused on the FGL1-mediated regulation of signalling by NF-κB, an important proinflammatory transcription factor for inflammatory disorders.

Activation of NF- $\kappa$ B plays a central role in the induction and exacerbation of the intestinal inflammatory response of CD patients[22]. The NF- $\kappa$ B family consists of p65 (RELA), RELB, c-REL, p50/p105 (NF- $\kappa$ B1), and p52/p100 (NF- $\kappa$ B2). Activated p65 can translocate into the nucleus to upregulate the transcriptional expression of proinflammatory cytokines. In the present study, the mRNA and protein levels of FGL1 and NF- $\kappa$ B and the concentrations of IL-1 $\beta$ , IL-6, IL-17, and TNF- $\alpha$  were markedly upregulated



in HT-29 cells stimulated with LPS, and these effects were reversed by depleting FGL1 with specific siRNA. Correspondingly, the expression of NF-KB and the four proinflammatory cytokines was enhanced following overexpression of FGL1. These results indicate that FGL1 may promote the intestinal inflammatory response by activating NF-KB signalling. The canonical pathway of NF-KB activation involves the IKK complex, consisting of NEMO, IKK $\alpha$ , and IKK $\beta$ , and the I $\kappa$ B protein family, including IkB $\alpha$ , IkB $\beta$ , and IkBe. After stimulation, IKK $\alpha$  and IKK $\beta$  activation promotes phosphorylation of IkBa. Degradation of phosphorylated IkBa releases the p65-p50 dimer for nuclear translocation [23]. In this study, the protein expression levels of IKK $\alpha$ , IKK $\beta$ , p-IKK $\alpha/\beta$ , p-IkB $\alpha$ , and p-p65 were decreased after knockdown of FGL1 compared to those in the cell model, and the inverse effect was verified by overexpression of FGL1. Therefore, FGL1 may induce intestinal inflammation by activating the canonical NF-κB pathway.

# CONCLUSION

In summary, we found for the first time that the expression of FGL1 is considerably upregulated in the plasma and intestinal mucosal and epithelial tissues of CD patients. FGL1 might induce intestinal inflammation by activating the canonical NF-κB signalling pathway to stimulate the secretion of proinflammatory cytokines, such as IL-1 $\beta$ , IL-6, IL-17, and TNF- $\alpha$ . Hence, FGL1 may be considered a potential biomarker and therapeutic target for CD. However, given the exploratory design of our study, the precise role of FGL1 in the pathogenesis of CD needs to be deeply investigated and further validated.

## ACKNOWLEDGEMENT

The authors are deeply grateful to Fan Yang from the Faculty of Art and Science, St. George Campus, University of Toronto, for helping us analyze the data in the statistics of this study.

# ARTICLE HIGHLIGHTS

#### Research background

Currently, the etiology and pathogenesis of Crohn's disease (CD) are not completely known, which makes it incurable. It is urgent to reveal the pathophysiological mechanism of CD and investigate new therapeutic targets.

#### Research motivation

To explore a potential therapeutic target for CD and verify its role in the CD pathological mechanism.

#### Research objectives

In this study, we attempted to find a potential therapeutic target for CD and verify its role in the CD pathological mechanism in vitro.

#### Research methods

Proteomics was implemented to quantify the protein profile in the plasma of CD patients. Among the differentially expressed proteins, a hub gene that could regulate the immune response was selected for further study. The expression of the selected hub gene in the inflamed intestinal mucosa was verified by immunohistochemical staining. In vitro, the effects of the hub gene on the expression of proinflammatory cytokines and the NF-KB signalling pathway were evaluated by ELISA, qRT-PCR, and Western blot analysis.

#### Research results

Fibrinogen-like protein 1 (FGL1), as a hub gene of the differentially expressed proteins, was confirmed to be markedly upregulated in the plasma and intestinal mucosa of CD patients. Silencing FGL1 downregulated the levels of the proinflammatory cytokines IL-1β, IL-6, IL-17, and TNF-α. Furthermore, FGL1 knockdown repressed the mRNA



expression of *NF*- $\kappa B$  and the protein levels of IKK $\alpha$ , IKK $\beta$ , p-IKK $\alpha/\beta$ , p-I $\kappa B\alpha$ , and pp65. Overexpression of FGL1 enhanced these results.

#### Research conclusions

FGL1 may promote intestinal inflammation modulated by the canonical NF- $\kappa$ B signalling pathway and has the potential to be a therapeutic target for CD.

#### Research perspectives

Our findings indicate a critical role of FGL1 in the onset and progression of CD, which may serve as a potential prognostic biomarker and therapeutic target for CD.

## REFERENCES

- Torres J. Mehandru S, Colombel JF, Pevrin-Biroulet L. Crohn's disease. Lancet 2017; 389: 1741-1 1755 [PMID: 27914655 DOI: 10.1016/S0140-6736(16)31711-1]
- Patel KV, Darakhshan AA, Griffin N, Williams AB, Sanderson JD, Irving PM. Patient optimization for surgery relating to Crohn's disease. Nat Rev Gastroenterol Hepatol 2016; 13: 707-719 [PMID: 27780971 DOI: 10.1038/nrgastro.2016.158]
- Kennedy NA, Heap GA, Green HD, Hamilton B, Bewshea C, Walker GJ, Thomas A, Nice R, Perry 3 MH, Bouri S, Chanchlani N, Heerasing NM, Hendy P, Lin S, Gaya DR, Cummings JRF, Selinger CP, Lees CW, Hart AL, Parkes M, Sebastian S, Mansfield JC, Irving PM, Lindsay J, Russell RK, McDonald TJ, McGovern D, Goodhand JR, Ahmad T; UK Inflammatory Bowel Disease Pharmacogenetics Study Group. Predictors of anti-TNF treatment failure in anti-TNF-naive patients with active luminal Crohn's disease: a prospective, multicentre, cohort study. Lancet Gastroenterol Hepatol 2019; 4: 341-353 [PMID: 30824404 DOI: 10.1016/S2468-1253(19)30012-3]
- Löwenberg M, Vermeire S, Mostafavi N, Hoentjen F, Franchimont D, Bossuyt P, Hindryckx P, Rispens T, de Vries A, van der Woude CJ, Berends S, Ambarus CA, Mathot R, Clasquin E, Baert F, D'Haens G. Vedolizumab Induces Endoscopic and Histologic Remission in Patients With Crohn's Disease. Gastroenterology 2019; 157: 997-1006.e6 [PMID: 31175865 DOI: 10.1053/j.gastro.2019.05.067]
- Gisbert JP, Chaparro M. Clinical Usefulness of Proteomics in Inflammatory Bowel Disease: A Comprehensive Review. J Crohns Colitis 2019; 13: 374-384 [PMID: 30307487 DOI: 10.1093/ecco-jcc/jjy158]
- 6 Yamamoto T, Gotoh M, Sasaki H, Terada M, Kitajima M, Hirohashi S. Molecular cloning and initial characterization of a novel fibrinogen-related gene, HFREP-1. Biochem Biophys Res Commun 1993; 193: 681-687 [PMID: 8390249 DOI: 10.1006/bbrc.1993.1678]
- 7 Liu S, Guo Y, Lu L, Lu J, Ke M, Xu T, Lu Y, Chen W, Wang J, Kong D, Shen Q, Zhu Y, Tan W, Ji W, Zhou W. Fibrinogen-Like Protein 1 Is a Novel Biomarker for Predicting Disease Activity and Prognosis of Rheumatoid Arthritis. Front Immunol 2020; 11: 579228 [PMID: 33123164 DOI: 10.3389/fimmu.2020.579228]
- Wang J, Sanmamed MF, Datar I, Su TT, Ji L, Sun J, Chen L, Chen Y, Zhu G, Yin W, Zheng L, Zhou 8 T, Badri T, Yao S, Zhu S, Boto A, Sznol M, Melero I, Vignali DAA, Schalper K. Fibrinogen-like Protein 1 Is a Major Immune Inhibitory Ligand of LAG-3. Cell 2019; 176: 334-347.e12 [PMID: 30580966 DOI: 10.1016/j.cell.2018.11.010]
- Lichtenstein GR, Loftus EV, Isaacs KL, Regueiro MD, Gerson LB, Sands BE. ACG Clinical Guideline: Management of Crohn's Disease in Adults. Am J Gastroenterol 2018; 113: 481-517 [PMID: 29610508 DOI: 10.1038/ajg.2018.27]
- 10 Tyers M, Mann M. From genomics to proteomics. *Nature* 2003; 422: 193-197 [PMID: 12634792 DOI: 10.1038/nature015101
- Titz B, Gadaleta RM, Lo Sasso G, Elamin A, Ekroos K, Ivanov NV, Peitsch MC, Hoeng J. 11 Proteomics and Lipidomics in Inflammatory Bowel Disease Research: From Mechanistic Insights to Biomarker Identification. Int J Mol Sci 2018; 19 [PMID: 30223557 DOI: 10.3390/ijms19092775]
- Starr AE, Deeke SA, Ning Z, Chiang CK, Zhang X, Mottawea W, Singleton R, Benchimol EI, Wen 12 M, Mack DR, Stintzi A, Figeys D. Proteomic analysis of ascending colon biopsies from a paediatric inflammatory bowel disease inception cohort identifies protein biomarkers that differentiate Crohn's disease from UC. Gut 2017; 66: 1573-1583 [PMID: 27216938 DOI: 10.1136/gutjnl-2015-310705]
- 13 Townsend P, Zhang Q, Shapiro J, Webb-Robertson BJ, Bramer L, Schepmoes AA, Weitz KK, Mallette M, Moniz H, Bright R, Merrick M, Shah SA, Sands BE, Leleiko N. Serum Proteome Profiles in Stricturing Crohn's Disease: A Pilot Study. Inflamm Bowel Dis 2015; 21: 1935-1941 [PMID: 26199992 DOI: 10.1097/MIB.00000000000445]
- Wang J, Wei W, Tang Q, Lu L, Luo Z, Li W, Lu Y, Pu J. Oxysophocarpine suppresses hepatocellular 14 carcinoma growth and sensitizes the therapeutic blockade of anti-Lag-3 via reducing FGL1 expression. Cancer Med 2020; 9: 7125-7136 [PMID: 32810392 DOI: 10.1002/cam4.3151]
- Sun C, Gao W, Liu J, Cheng H, Hao J. FGL1 regulates acquired resistance to Gefitinib by inhibiting 15 apoptosis in non-small cell lung cancer. Respir Res 2020; 21: 210 [PMID: 32778129 DOI: 10.1186/s12931-020-01477-y



- 16 Bie F, Wang G, Qu X, Wang Y, Huang C, Du J. Loss of FGL1 induces epithelialmesenchymal transition and angiogenesis in LKB1 mutant lung adenocarcinoma. Int J Oncol 2019; 55: 697-707 [PMID: 31322182 DOI: 10.3892/ijo.2019.4838]
- Wu HT, Ou HY, Hung HC, Su YC, Lu FH, Wu JS, Yang YC, Wu CL, Chang CJ. A novel 17 hepatokine, HFREP1, plays a crucial role in the development of insulin resistance and type 2 diabetes. Diabetologia 2016; 59: 1732-1742 [PMID: 27221093 DOI: 10.1007/s00125-016-3991-7]
- Kang L, Li HY, Ou HY, Wu P, Wang SH, Chang CJ, Lin SY, Wu CL, Wu HT. Role of placental 18 fibrinogen-like protein 1 in gestational diabetes. Transl Res 2020; 218: 73-80 [PMID: 32006524 DOI: 10.1016/j.trsl.2020.01.001]
- 19 Wu HT, Chen SC, Fan KC, Kuo CH, Lin SY, Wang SH, Chang CJ, Li HY. Targeting fibrinogen-like protein 1 is a novel therapeutic strategy to combat obesity. FASEB J 2020; 34: 2958-2967 [PMID: 31908014 DOI: 10.1096/fj.201901925R]
- 20 Liu Z, Ukomadu C. Fibrinogen-like protein 1, a hepatocyte derived protein is an acute phase reactant. Biochem Biophys Res Commun 2008; 365: 729-734 [PMID: 18039467 DOI: 10.1016/j.bbrc.2007.11.069]
- 21 Pierre N, Baiwir D, Huynh-Thu VA, Mazzucchelli G, Smargiasso N, De Pauw E, Bouhnik Y, Laharie D, Colombel JF, Meuwis MA, Louis E; GETAID (Groupe d'Etude Thérapeutique des Affections Inflammatoires du tube Digestif). Discovery of biomarker candidates associated with the risk of short-term and mid/long-term relapse after infliximab withdrawal in Crohn's patients: a proteomics-based study. Gut 2020 [PMID: 33106355 DOI: 10.1136/gutjnl-2020-322100]
- 22 Atreya I, Atreya R, Neurath MF. NF-kappaB in inflammatory bowel disease. J Intern Med 2008; 263: 591-596 [PMID: 18479258 DOI: 10.1111/j.1365-2796.2008.01953.x]
- 23 Rius-Pérez S, Pérez S, Martí-Andrés P, Monsalve M, Sastre J. Nuclear Factor Kappa B Signaling Complexes in Acute Inflammation. Antioxid Redox Signal 2020; 33: 145-165 [PMID: 31856585 DOI: 10.1089/ars.2019.7975]



W U

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5958-5966

DOI: 10.3748/wjg.v27.i35.5958

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

ORIGINAL ARTICLE

# **Retrospective Study** Effectiveness and safety of over-the-scope clip in closing perforations after duodenal surgery

Zhen-Zhen Wang, Xian-Bin Zhou, Yi Wang, Xin-Li Mao, Li-Ping Ye, Ling-Ling Yan, Ya-Hong Chen, Ya-Qi Song, Yue Cai, Shi-Wen Xu, Shao-Wei Li

ORCID number: Zhen-Zhen Wang 0000-0002-6274-2646; Xian-Bin Zhou 0000-0003-0048-7456; Yi Wang 0000-0002-9817-1769; Xin-Li Mao 0000-0003-4548-1867; Li-Ping Ye 0000-0001-9839-2062; Ling-Ling Yan 0000-0001-5103-9886; Ya-Hong Chen 0000-0003-0004-0187; Ya-Qi Song 0000-0001-9648-8159; Yue Cai 0000-0002-7201-6525; Shi-Wen Xu 0000-0001-6774-1979; Shao-Wei Li 0000-0002-3276-1037.

Author contributions: Wang ZZ, Zhou XB, Wang Y, Mao XL, and Ye LP participated in the clinical treatment; Xu SW, Yan LL, and Li SW wrote the original draft; Song YQ, Cai Y, and Chen YH undertook validation, writing, review, and editing; all authors contributed to the article and approved the submitted version.

Supported by Program of Taizhou Science and Technology Grant, No. 20ywb29; Medical Health Science and Technology Project of Zhejiang Province, No. 2021PY083 and No. 2019KY239; Key Technology Research and Development Program of Zhejiang Province, No. 2019C03040; Major Research Program of Taizhou Enze Medical Center Grant, No. 19EZZDA2; and Open Fund of Key Laboratory of Key Laboratory of Minimally Invasive Techniques & Rapid

Zhen-Zhen Wang, Xian-Bin Zhou, Yi Wang, Xin-Li Mao, Li-Ping Ye, Ling-Ling Yan, Ya-Qi Song, Yue Cai, Shao-Wei Li, Key Laboratory of Minimally Invasive Techniques and Rapid Rehabilitation of Digestive System Tumor of Zhejiang Province, Taizhou Hospital of Zhejiang Province Affiliated to Wenzhou Medical University, Linhai 317000, Zhejiang Province, China

Zhen-Zhen Wang, Xian-Bin Zhou, Yi Wang, Xin-Li Mao, Li-Ping Ye, Ling-Ling Yan, Ya-Qi Song, Yue Cai, Shao-Wei Li, Department of Gastroenterology, Taizhou Hospital of Zhejiang Province Affiliated to Wenzhou Medical University, Linhai 317000, Zhejiang Province, China

Ya-Hong Chen, Health Management Center, Taizhou Hospital of Zhejiang Province Affiliated to Wenzhou Medical University, Linhai 317000, Zhejiang Province, China

Shi-Wen Xu, Taizhou Hospital of Zhejiang Province, Wenzhou Medical University, Linhai 317000, Zhejiang Province, China

Corresponding author: Shao-Wei Li, PhD, Academic Fellow, Assistant Professor, Associate Research Scientist, Instructor, Key Laboratory of Minimally Invasive Techniques and Rapid Rehabilitation of Digestive System Tumor of Zhejiang Province, Taizhou Hospital of Zhejiang Province Affiliated to Wenzhou Medical University, No. 150 Xinmen Street, Linhai 317000, Zhejiang Province, China. li shaowei81@hotmail.com

# Abstract

# BACKGROUND

Endoscopic resection of duodenal subepithelial lesions (SELs) is a difficult procedure with a high risk of perforation. At present, dealing with perforation after endoscopic resection of duodenal SELs is still considered a great challenge.

# AIM

To evaluate the effectiveness and safety of an over-the-scope clip (OTSC) in the treatment of perforation post-endoscopic resection of duodenal SELs.

# **METHODS**

From May 2015 to November 2019, 18 patients with perforation following endoscopic resection of duodenal SELs were treated with OTSCs. Data comprising the rate of complete resection, closure of intraprocedural perforation, delayed bleeding, delayed perforation, and postoperative infection were extracted.



Rehabilitation of Digestive System Tumor of Zhejiang Province, No. 21SZDSYS01 and No. 21SZDSYS09.

## Institutional review board

statement: The study was reviewed and approved by the Ethics Committee of Taizhou Hospital of Zhejiang Province affiliated to Wenzhou Medical University Institutional Review Board (approval No. K20210412).

#### Informed consent statement: All

study participants, or their legal guardian, provided informed written consent prior to study enrollment.

Conflict-of-interest statement: The authors declare that they have no conflict interests.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially. and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: China

## Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B, B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Received: May 10, 2021 Peer-review started: May 10, 2021 First decision: June 27, 2021

# RESULTS

The rate of complete removal of duodenal SELs and successful closure of the perforation was 100%. The median perforation size was 1 cm in diameter. Seventeen patients had minor intraoperative bleeding, while the remaining 1 patient had considerable amount of bleeding during the procedure. Seven patients had postoperative abdominal infections, of which 1 patient developed an abscess in the right iliac fossa and another patient developed septic shock. All 18 patients recovered and were discharged. No delayed bleeding or perforation was reported. The mean time taken to resume normal diet after the procedure was 6.5 d. The mean postoperative hospital stay was 9.5 d. No residual or recurrent lesions were detected during the follow-up period (15-66 mo).

## CONCLUSION

Closing a perforation after endoscopic resection of duodenal SELs with OTSCs seems to be an effective and reasonably safe therapeutic method.

Key Words: Over-the-scope clip; Duodenal subepithelial lesion; Endoscopic resection; Perforation; Effectiveness; Safety

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: This study presents the use of over-the-scope clip in closing duodenal perforation of 18 patients. We believe that our study makes a significant contribution to the literature because dealing with perforation after endoscopic resection of duodenal subepithelial lesions is challenging. This study aimed to evaluate the effectiveness and safety of over-the-scope clip in closing perforation after endoscopic resection of duodenal subepithelial lesions. The rate of successful closure was 100%. No delayed perforation occurred in any of the patients. Seven patients had postoperative infection, of which 1 patient developed septic shock and underwent surgery. All 18 patients recovered.

Citation: Wang ZZ, Zhou XB, Wang Y, Mao XL, Ye LP, Yan LL, Chen YH, Song YQ, Cai Y, Xu SW, Li SW. Effectiveness and safety of over-the-scope clip in closing perforations after duodenal surgery. World J Gastroenterol 2021; 27(35): 5958-5966 URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5958.htm DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5958

# INTRODUCTION

Duodenal subepithelial lesions (SELs) include Brunner's adenomas, lipomas, heterotopic pancreas, leiomyomas, neuroendocrine tumors, and gastrointestinal stromal tumors (GISTs). Most of these are benign, while some lesions, such as neuroendocrine tumors and GISTs, are potentially malignant[1-3]. Resection of these lesions may contribute to improvement in diagnosis and treatment outcomes.

Surgery, including pancreatoduodenectomy and limited resection, is the most basic treatment for duodenal lesions. However, due to the complexity of the operation, risk of trauma, high incidence of postoperative complications, poor quality of life of patients after surgery, and other difficulties, these surgeries are not easily consented by patients, which also puts the medical staff in a difficult position. With the recent development of minimally invasive endoscopic treatment technologies, such as endoscopic submucosal dissection (ESD), endoscopic muscularis excavation, and endoscopic full-thickness resection, endoscopic treatment has become increasingly popular, which brings hope for the use of minimally invasive treatment of duodenal SELs in the future.

However, endoscopic resection of duodenal SELs is still regarded as a challenging procedure due to a high risk of perforation. The incidence of perforations in duodenal ESD has been reported to range from 6.7%-36.6% during the procedure and 0%-14.3% during the postoperative period [1,4-7]. Management of perforations after endoscopic removal of duodenal SELs is particularly challenging. However, this may be achieved



Revised: July 8, 2021 Accepted: August 18, 2021 Article in press: August 18, 2021 Published online: September 21, 2021

P-Reviewer: Masaki S, Seicean A S-Editor: Gao CC L-Editor: Filipodia P-Editor: Liu JH



by using over-the-scope clips (OTSCs). An OTSC was developed as an endoscopic fullthickness gastrointestinal closure device and has become one of the treatment options for gastrointestinal perforation because it is less invasive compared to conventional surgical closure. At present, there are few reports on endoscopic resection of duodenal SELs and endoscopic methods for the management of perforations[1,2,8,9]. To explore further this area, this study aimed to assess the effectiveness and safety of OTSCs in the treatment of perforation after endoscopic resection of duodenal SELs.

# MATERIALS AND METHODS

#### Patients

This was a retrospective study and was approved by the ethics committee of Taizhou Hospital of Zhejiang Province (Linhai, China). The study included 18 consecutive patients who were treated with OTSCs to close perforations that resulted after endoscopic resection of duodenal SELs, from May 2015 to November 2019. Patients were recruited if they met all of the following criteria: (1) Patients with duodenal SELs diagnosed by computed tomography and endoscopic ultrasound (EUS) with a highfrequency miniprobe (UM-2R, 12 MHz; UM-3R, 20 MHz, Olympus Optical, Tokyo, Japan); (2) Patients who underwent endoscopic resection of duodenal SELs and had intraoperative or postoperative perforations; (3) The duodenal perforation was closed using an OTSC; and (4) Patients who were able to tolerate general anesthesia and had no blood coagulation disorders prior to the procedure.

Before the endoscopic procedure, informed consent was obtained from all 18 patients. Patients were also informed that an OTSC might be used, and surgical intervention might be required in case of unsuccessful resection of the lesion or the occurrence of severe complications that cannot be successfully managed by endoscopic methods and conservative treatment.

The main outcome measurements were as follows: (1) The rate of complete closure of intraprocedural perforation; (2) Delayed perforation rate; and (3) Postoperative infection rate. All endoscopic resection procedures were performed by an experienced endoscopist in a sterile operating room while the patients were under general anesthesia with tracheal intubation.

#### Endoscopic procedures

The main equipment and accessories used were as follows: A single-accessory channel endoscope (Q260J; Olympus) with a transparent cap (ND-201-11802; Olympus) attached to its tip, an argon plasma coagulation unit (APC 300; ERBE, Tübingen, Germany), a high-frequency electronic cutting device (ICC 200; ERBE), a hook knife (KD-620LR; Olympus), an insulated-tip knife (KD-611L, IT2; Olympus), hot biopsy forceps (FD-410LR; Olympus), foreign body forceps (FG-B-24, Kangjin, Changzhou, China), a snare (SD-230U-20; Olympus), a carbon dioxide insufflator (Olympus), twin graspers (Ovesco Endoscopy AG, Tuebingen, Germany), an OTSC (12/6 t-type, Ovesco Endoscopy AG,), a titanium clip (HX-600-135; Olympus and M00522600), and endoloop (Leo Medical Co., Ltd, Changzhou, China).

Endoscopic resection was performed as follows (Figure 1): (1) Several marking dots were initially made around the lesion using a needle-knife to define the border; (2) A submucosal elevation was made by injection of solution (100 mL saline plus 1 mL epinephrine and 2 mL indigo carmine); (3) Subsequently, the mucosa was incised with a hook knife outside the border to reveal the lesion; (4) A circumferential excavation was made as deep as the submucosa or muscularis propria layer around the lesion using an insulated tip knife; (5) After the lesion was completely resected, it was removed using a snare or foreign body forceps; and (6) Duodenal tissues adjacent to the perforation were clamped with twin graspers and then drawn into the transparent cap of the OTSC device until they were fully inhaled into the transparent cap following which the OTSC closure system was released to close the wound. If defect closure was not complete, several clip and/or endoloops were used to close the remaining portions. The mucosa defect was closed with several clips in a 'side to center' manner, and an endoloop was placed to trap all the clips. Finally, the endoloop was slowly tightened, and all the clips were tied together with the endoloop[8].

#### Postoperative management and follow-up

After the operation, all patients were treated with postoperative fasting, gastrointestinal decompression, proton-pump inhibitors, and antibiotics for infection prevention. Oral intake was gradually resumed depending on the speed of recovery.



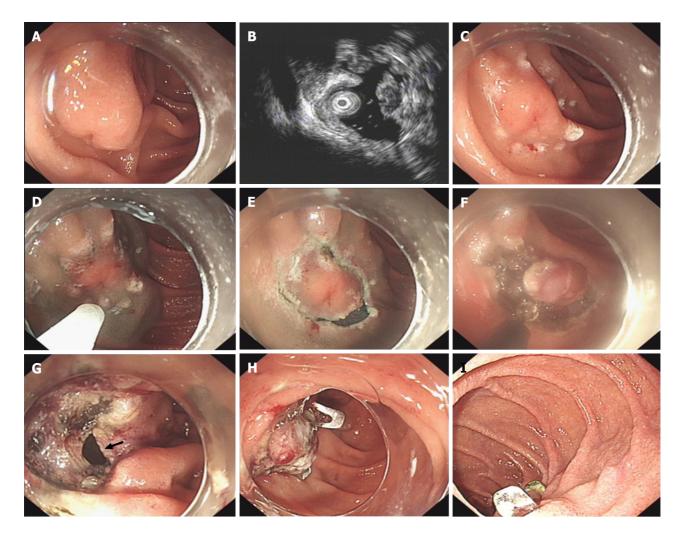


Figure 1 Endoscopic resection of a subepithelial lesion located in the descending duodenum with perforation closure using an over-thescope clip. A: Endoscopic view of a subepithelial lesion located in the descending duodenum; B: Endoscopic ultrasound evaluation of the same lesion; C: Several marking dots are made around the lesion; D: Injection solution used to elevate the submucosa; E: The mucosa is incised outside the marking dots; F: A circumferential excavation is made as deep as the submucosa around the lesion; G: A duodenal perforation is observed (black arrow) after removal of the lesion; H: The perforation is closed with an over-the-scope clip; I: Healed wound 9 mo after the procedure.

> Every patient underwent follow-up endoscopies to monitor wound healing at 3 mo and 6 mo after endoscopic resection. EUS was performed to check for residual lesions after 3 mo. Patients with potentially malignant lesions, such as neuroendocrine tumors and GISTs, were monitored by endoscopy and/or EUS to detect recurrent lesions, and abdominal US and/or computed tomography to detect distant metastasis every 12 mo.

#### Statistical analysis

Data were analyzed using SPSS software (version 20.0; SPSS Inc., Armonk, NY, United States). Descriptive statistics were used for this study. The median was used for variables with a skewed distribution, while the mean was used in the case of a normal distribution of variables. Enumeration data are expressed as case numbers and percentages (%).

# RESULTS

#### Clinical characteristics and therapeutic outcome

Patient information is summarized in Table 1 and therapeutic outcomes, are described in Table 2. The rate of successful *en bloc* resection was 100%. The vertical and horizontal margins of all specimens were tumor-free. Thus, the complete resection rate was 100%.

22eishidena® WJG https://www.wjgnet.com

#### Table 1 Clinical characteristics of the 18 patients with duodenal subepithelial lesions, n (%)

Patients	
Median age, yr (range)	53.5 (29-74)
Gender	
Male	8 (44.4)
Female	10 (55.6)
Symptom	
Upper abdominal pain	3 (16.7)
Abdominal distention	4 (22.2)
Melena	5 (27.8)
Asymptomatic	6 (33.3)
Lesions	
Median size, cm (range)	2.0 (1.3-5.0)
Location of lesion	
Duodenal bulb	11 (61.1)
Descending junction of duodenal bulb	4 (22.2)
Descending duodenum	3 (16.7)
Origination of lesion	
Submucosal layer	9 (50.0)
Muscularis propria layer	9 (50.0)

All 18 patients had intraoperative perforations. The median perforation size was 1 cm in diameter (range, 0.5-3.0 cm). The wound was closed with an OTSC in 6 cases, an OTSC + a titanium clip in 1 case, and an OTSC + a titanium clip + an endoloop in 5 cases. The rate of successful intraprocedural perforation closure was 100%.

Seventeen patients had minor intraoperative bleeding. The remaining 1 patient, who had a tumor originating from the lamina propria, growing mainly out of the lumen, with rich blood supply, had considerable amount of bleeding during the procedure. All patients were treated with hot biopsy forceps to achieve hemostasis during the procedure.

None of the patients developed delayed bleeding or perforation. Seven patients had postoperative abdominal infections and were administered intensive antibiotic therapy. Among the 7 patients, 1 patient developed an abscess in the right iliac fossa that improved after puncture and drainage, while another patient developed septic shock and received peritoneal lavage and underwent distal subtotal gastrectomy with duodenal bulb resection. All 18 patients recovered and were discharged. The mean time taken to resume normal diet after the procedure was 6.5 d. The mean postoperative hospital stay was 9.5 d.

#### Follow-up

The median follow-up period after the procedure was 27 mo (range, 15-66 mo). No residual or recurrent lesions, duodenal stenosis, or adhesions were detected during the follow-up period in any of the patients.

# DISCUSSION

Currently, endoscopic resection of duodenal SELs is a challenging procedure with a high risk of perforation. Published studies about endoscopic resection of duodenal SELs and endoscopic methods for management of perforations are limited [1,2,8,9]. In this study, we used OTSCs to close perforations in 18 patients. The rate of complete removal of duodenal SELs and successful perforation closure was 100%. No delayed bleeding or perforation occurred in any of the patients. This suggests that the use of OTSCs can effectively close perforations following endoscopic resection of duodenal



#### Table 2 Therapeutic outcome and adverse events of endoscopic resection for duodenal subepithelial lesions, n (%)

Therapeutic outcome and adverse events					
Complete resection	18 (100)				
Histology diagnosis					
Brunner's adenoma	1 (5.6)				
Heterotopic pancreas	7 (38.9)				
GIST	7 (38.9)				
Very low risk	1 (5.6)				
Low risk	6 (33.3)				
Neuroendocrine tumors	3 (16.6)				
Complication					
Delayed perforation	0 (0)				
Delayed bleeding	0 (0)				
Postoperative infection	7 (38.9)				
Mean time of diet recovery after the procedure, d (range)	6.5 (2-14)				
Mean hospital stay after the procedure, d (range)	9.5 (4-18)				
Median follow-up period, mo (range)	27 (15-66)				

GIST: Gastrointestinal stromal tumor.

SELs when performed by an experienced endoscopist.

The clinical manifestations of duodenal SELs are nonspecific and related to the location, size, growth pattern, presence of mucosal ulcers, and invasion or compression of adjacent organs. Most duodenal lesions have no symptoms and are usually found incidentally during endoscopic examinations. Clinical symptoms such as gastrointestinal bleeding, abdominal pain, and abdominal distention may occur when the lesion is very large or when an ulcer develops on the surface of the lesion.

Though most duodenal SELs, such as lipomas, Brunner's adenomas, heterotopic pancreas, and cysts, are benign, some including neuroendocrine tumors and GISTs are potentially malignant<sup>[1-3]</sup>. Endoscopy and EUS are of great value in the diagnosis of duodenal SELs; however, they may be difficult to diagnose on some occasions. Patients with duodenal SELs can be monitored by endoscopy, especially for asymptomatic tumors that lack high-risk features as identified by EUS[10]. However, surveillance using only endoscopy may increase the risk of delayed diagnosis of a malignancy[11]. Furthermore, the difficulty of the operation and risk of combined evisceration will increase if the lesion is large. In such cases, removal of the lesion is inevitable.

Traditional surgical approaches for duodenal lesions, including pancreatoduodenectomy and limited resection, are traumatic and may result in serious complications, such as bleeding, perforation, and infection. Considering these potential risks associated with surgical therapy, endoscopic treatment is used as an alternative choice, which may be safer, more effective, and is minimally invasive. However, endoscopic resection of duodenal SELs is still considered to be a challenging procedure because the duodenal lumen is narrow and the initial part (ball to lower part) is an anti-c loop, which renders the endoscope unstable. Moreover, the abundant blood vessels and Brunner glands in the submucosa of the duodenum make it difficult to lift the mucosa after injection. In addition, compared to other parts of the gastrointestinal tract, the muscularis propria layer of the duodenum is soft and thin, and the posterior wall lacks the serosal layer; therefore, perforation can occur easily during or after the endoscopic resection of duodenal lesions, especially duodenal SELs[8]. The incidence of intraprocedural perforations in duodenal ESD has been reported to range from 6.7%-36.6%, and is 0%-14.3% in delayed perforations[1,4-7]. Moreover, emergency operations have been performed in 3.3%-25.0% of patients due to intraprocedural uncontrollable perforation or delayed perforation[1,4-7]. Our previous study reported that the perforation rate of endoscopic resection of duodenal SELs in our hospital was 7.4%[8].



Perioperative perforation associated with endoscopic therapy was previously considered a serious complication that usually requires surgery. With the development of endoscopic suture instruments and techniques, patients with iatrogenic gastrointestinal perforation can be successfully managed using endoscopic methods and conservative treatment without surgical intervention[12,13]. Thus, most perforations related to endoscopic treatment are no longer life-threatening complications. However, endoscopic closure of perforations after endoscopic resection of duodenal SELs remains a great challenge.

In the past, titanium clips were used for endoscopic closure of gastrointestinal perforations, especially for small acute perforations (< 5 mm). However, a titanium clip has a narrow wingspan and lacks the ability to approximate adequately the margins of the defect. Consequently, the rate of leakage after repairing a large perforation of more than 1 cm is high as the seal is confined to the surface rather than the full-thickness of the mucosa[14-16]. An OTSC has a greater holding strength[16, 17]; it can clamp the entire wall of the lumen and grasp more tissue. The design can manage full-thickness perforations with diameters of up to 3 cm[14]. Moreover, the gap between the teeth of an OTSC allows blood to pass through to avoid tissue necrosis. The advantage of an OTSC lies in its ease of use, ability to close defects between 1 and 3 cm with a single clip, and safety, which allows endoscopists to deal effectively with acute perforations immediately after identification[18]. Thus, OTSCs are easy to operate and can effectively shorten operation times. Moreover, The European Society of Gastrointestinal Endoscopy recommends OTSCs for endoscopic closure of iatrogenic perforations[18]. According to a systematic review, the success rate of using OTSCs to manage perforations was 85.3%, while 9.4% of patients still required surgical intervention after an OTSC placement to achieve complete closure [19]. Voermans et al[14] reported 12 cases of duodenal perforation that were treated with OTSCs, nine of which were effectively closed, with an overall success rate of 75%. In our study, the rate of successful closure of intraprocedural perforations was 100%. However, we have also used a titanium clip in 1 case and a titanium clip along with an endoloop in 5 cases. It seems that if the perforation is larger than 1.5 cm, using an OTSC alone may fail to achieve complete closure. We speculate that the combination of OTSC, titanium clip, and endoloops may be more effective. Given that the duodenal lumen is narrow, caution should be exercised to avoid grasping too much tissue to avoid further narrowing of the lumen while deploying the OTSC in the duodenum. In our study, no duodenal stenosis was detected in any patient during the follow-up period.

The duodenum is exposed to pancreatic juices and bile, causing delayed perforations more likely to occur after endoscopic resection of duodenal lesions. Complete closure of the wound facilitates prevention of delayed perforation[6,7,17]. Due to its the strong tightening force and the gap between its teeth, an OTSC can manage to close full-thickness duodenal perforations and avoid tissue necrosis, which effectively reduces the occurrence of delayed perforations. A carbon dioxide pump is also recommended to use with endoscopic treatment, especially when a perforation occurs. The use of gastrointestinal decompression after endoscopic closure of perforation is helpful for the absorption of gas and liquid in the intestinal cavity. It also reduces tension in the wound, and promotes wound healing, which can reduce the incidence of delayed perforations. In this study, we placed a jejunal nutrition tube next to the wound and a gastrointestinal decompression tube to extract gas and digestive juice. Thereafter, none of the patients developed delayed perforations.

The duodenum is an interperitoneal organ, most of which is located in the retroperitoneum. After perforation or full-thickness resection, digestive fluid from the duodenum (mainly bile and pancreatic juice) flows into the peritoneal cavity or retroperitoneal cavity, which may cause serious abdominal or retroperitoneal infection. In our study, 7 patients (38.9%) had postoperative abdominal infection, including 1 who developed an abscess in the right iliac fossa and another who developed septic shock. Severe infection in the 2 cases were considered to be caused by long operation times and large amounts of digestive juice entering the abdominal cavity. Timely conversion of the endoscopic procedure to surgery or combining with laparoscopy when the resection is found to be difficult may help avoid such complications.

Due to their strong holding strength, OTSCs are more difficult to detach spontaneously from the mucosa than normal titanium clips. The OTSC is made of nitinol, which has favorable biocompatibility. Thus, this device is considered a permanent implanted material. However, OTSCs should be removed in the following circumstances: (1) Poor healing; (2) OTSC misplacement; (3) Repeat biopsy/therapy or further treatment; (4) Adverse events after OTSC implantation, such as ulcers and



stenosis of the digestive tract; (5) Removal after recovery; and (6) Patient's wishes<sup>[20]</sup>. In our study, there were no such indications for removal. During the follow-up period, OTSCs detached spontaneously in most cases.

This study has a few limitations. First, this was a single-center retrospective study and the sample size was relatively small; therefore, selection bias may have been present. Second, since this was a retrospective study, it lacked randomized and control samples. Third, our institution is a tertiary endoscopic center in Zhejiang Province where the procedures were performed by an experienced operator; thus, the results of this study may not be applicable to all other endoscopic centers.

# CONCLUSION

Closing of perforations after endoscopic resection of duodenal SELs with OTSCs is an effective and reasonably safe therapeutic method. However, this procedure should be performed by an experienced endoscopic team. If the endoscopic procedure fails or the postoperative complications are difficult to manage, the patient should be planned to undergo surgery immediately.

# ARTICLE HIGHLIGHTS

#### Research background

Currently, endoscopic resection of duodenal subepithelial lesions (SELs) is a challenging procedure with a high risk of perforation.

#### Research motivation

It is importance to deal with perforation after endoscopic resection of duodenal SELs. However, so far, there were few reports on endoscopic methods for management of perforations.

#### Research objectives

We aim to evaluate the effectiveness and safety of over-the-scope clip (OTSC) in the closing the perforation after endoscopic resection of duodenal SELs.

#### Research methods

This was a retrospective study. We collected data of 18 consecutive patients who were treated with OTSCs to close the perforation after endoscopic resection of duodenal SELs and analyzed the rate of complete resection, closure of intraprocedural perforation, delayed bleeding, delayed perforation, and postoperative infection.

#### Research results

All the perforations after endoscopic resection of duodenal SELs were successfully closed. No delayed bleeding or perforation occurred in any of the patients.

#### Research conclusions

OTSC can effectively and safely close the perforations after endoscopic resection of duodenal SELs by an experienced endoscopist.

#### Research perspectives

We need to expand the sample size to confirm further the effectiveness and safety of OTSC in closing the perforation after endoscopic resection of duodenal SELs. In addition, the long-term outcome of OTSC should be observed by extending the followup time.

# REFERENCES

- Matsumoto S, Miyatani H, Yoshida Y. Endoscopic submucosal dissection for duodenal tumors: a single-center experience. Endoscopy 2013; 45: 136-137 [PMID: 22930172 DOI: 10.1055/s-0032-1310123
- Musumba C, Sonson R, Tutticci N, Nanda K, Bourke MJ. Endoscopic submucosal dissection of a duodenal neuroendocrine tumor. Gastrointest Endosc 2014; 79: 716 [PMID: 24368076 DOI:



10.1016/j.gie.2013.11.011]

- 3 Xu GQ, Wu YQ, Wang LJ, Chen HT. Values of endoscopic ultrasonography for diagnosis and treatment of duodenal protruding lesions. *J Zhejiang Univ Sci B* 2008; 9: 329-334 [PMID: 18381809 DOI: 10.1631/jzus.B0710546]
- 4 Hoteya S, Yahagi N, Iizuka T, Kikuchi D, Mitani T, Matsui A, Ogawa O, Yamashita S, Furuhata T, Yamada A, Kimura R, Nomura K, Kuribayashi Y, Kaise M. Endoscopic submucosal dissection for nonampullary large superficial adenocarcinoma/adenoma of the duodenum: feasibility and long-term outcomes. *Endosc Int Open* 2013; 1: 2-7 [PMID: 26135505 DOI: 10.1055/s-0033-1359232]
- 5 Jung JH, Choi KD, Ahn JY, Lee JH, Jung HY, Choi KS, Lee GH, Song HJ, Kim DH, Kim MY, Bae SE, Kim JH. Endoscopic submucosal dissection for sessile, nonampullary duodenal adenomas. *Endoscopy* 2013; 45: 133-135 [PMID: 23364841 DOI: 10.1055/s-0032-1326178]
- 6 Nonaka S, Oda I, Tada K, Mori G, Sato Y, Abe S, Suzuki H, Yoshinaga S, Nakajima T, Matsuda T, Taniguchi H, Saito Y, Maetani I. Clinical outcome of endoscopic resection for nonampullary duodenal tumors. *Endoscopy* 2015; 47: 129-135 [PMID: 25314330 DOI: 10.1055/s-0034-1390774]
- 7 Yamamoto Y, Yoshizawa N, Tomida H, Fujisaki J, Igarashi M. Therapeutic outcomes of endoscopic resection for superficial non-ampullary duodenal tumor. *Dig Endosc* 2014; 26 Suppl 2: 50-56 [PMID: 24750149 DOI: 10.1111/den.12273]
- 8 Ye LP, Mao XL, Zheng HH, Zhang Y, Shen LY, Zhou XB, Zhu LH. Safety of endoscopic resection for duodenal subepithelial lesions with wound closure using clips and an endoloop: an analysis of 68 cases. *Surg Endosc* 2017; 31: 1070-1077 [PMID: 27387179 DOI: 10.1007/s00464-016-5065-9]
- 9 Schmidt A, Bauder M, Riecken B, von Renteln D, Muehleisen H, Caca K. Endoscopic full-thickness resection of gastric subepithelial tumors: a single-center series. *Endoscopy* 2015; 47: 154-158 [PMID: 25380509 DOI: 10.1055/s-0034-1390786]
- 10 Demetri GD, von Mehren M, Antonescu CR, DeMatteo RP, Ganjoo KN, Maki RG, Pisters PW, Raut CP, Riedel RF, Schuetze S, Sundar HM, Trent JC, Wayne JD. NCCN Task Force report: update on the management of patients with gastrointestinal stromal tumors. *J Natl Compr Canc Netw* 2010; 8 Suppl 2: S1-41; quiz S42 [PMID: 20457867 DOI: 10.6004/jnccn.2010.0116]
- 11 Białek A, Wiechowska-Kozłowska A, Pertkiewicz J, Polkowski M, Milkiewicz P, Karpińska K, Ławniczak M, Starzyńska T. Endoscopic submucosal dissection for treatment of gastric subepithelial tumors (with video). *Gastrointest Endosc* 2012; **75**: 276-286 [PMID: 22032850 DOI: 10.1016/j.gie.2011.08.029]
- 12 Yoshizumi F, Yasuda K, Kawaguchi K, Suzuki K, Shiraishi N, Kitano S. Submucosal tunneling using endoscopic submucosal dissection for peritoneal access and closure in natural orifice transluminal endoscopic surgery: a porcine survival study. *Endoscopy* 2009; **41**: 707-711 [PMID: 19670139 DOI: 10.1055/s-0029-1214959]
- 13 Guo J, Liu Z, Sun S, Liu X, Wang S, Ge N, Wang G, Qi Y. Endoscopic full-thickness resection with defect closure using an over-the-scope clip for gastric subepithelial tumors originating from the muscularis propria. *Surg Endosc* 2015; 29: 3356-3362 [PMID: 25701060 DOI: 10.1007/s00464-015-4076-2]
- 14 Voermans RP, Le Moine O, von Renteln D, Ponchon T, Giovannini M, Bruno M, Weusten B, Seewald S, Costamagna G, Deprez P, Fockens P; CLIPPER Study Group. Efficacy of endoscopic closure of acute perforations of the gastrointestinal tract. *Clin Gastroenterol Hepatol* 2012; 10: 603-608 [PMID: 22361277 DOI: 10.1016/j.cgh.2012.02.005]
- 15 Verlaan T, Voermans RP, van Berge Henegouwen MI, Bemelman WA, Fockens P. Endoscopic closure of acute perforations of the GI tract: a systematic review of the literature. *Gastrointest Endosc* 2015; 82: 618-28.e5 [PMID: 26005015 DOI: 10.1016/j.gie.2015.03.1977]
- 16 Singhal S, Changela K, Papafragkakis H, Anand S, Krishnaiah M, Duddempudi S. Over the scope clip: technique and expanding clinical applications. *J Clin Gastroenterol* 2013; 47: 749-756 [PMID: 23751852 DOI: 10.1097/MCG.0b013e318296ecb9]
- 17 Mori H, Shintaro F, Kobara H, Nishiyama N, Rafiq K, Kobayashi M, Nakatsu T, Miichi N, Suzuki Y, Masaki T. Successful closing of duodenal ulcer after endoscopic submucosal dissection with over-the-scope clip to prevent delayed perforation. *Dig Endosc* 2013; 25: 459-461 [PMID: 23368742 DOI: 10.1111/j.1443-1661.2012.01363.x]
- 18 Paspatis GA, Dumonceau JM, Barthet M, Meisner S, Repici A, Saunders BP, Vezakis A, Gonzalez JM, Turino SY, Tsiamoulos ZP, Fockens P, Hassan C. Diagnosis and management of iatrogenic endoscopic perforations: European Society of Gastrointestinal Endoscopy (ESGE) Position Statement. *Endoscopy* 2014; 46: 693-711 [PMID: 25046348 DOI: 10.1055/s-0034-1377531]
- 19 Bartell N, Bittner K, Kaul V, Kothari TH, Kothari S. Clinical efficacy of the over-the-scope clip device: A systematic review. *World J Gastroenterol* 2020; 26: 3495-3516 [PMID: 32655272 DOI: 10.3748/wjg.v26.i24.3495]
- 20 Ou YH, Kong WF, Li LF, Chen PS, Deng SH, He FJ, Peng QQ, Yue H. Methods for Endoscopic Removal of Over-the-Scope Clip: A Systematic Review. *Can J Gastroenterol Hepatol* 2020; 2020: 5716981 [PMID: 32908852 DOI: 10.1155/2020/5716981]

WÜ

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5967-5977

DOI: 10.3748/wjg.v27.i35.5967

**Retrospective Study** 

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

ORIGINAL ARTICLE

# Hepatic perivascular epithelioid cell tumor: Clinicopathological analysis of 26 cases with emphasis on disease management and prognosis

Shan Zhang, Pan-Pan Yang, Yu-Chen Huang, Hong-Chun Chen, De-Li Chen, Wen-Tian Yan, Ning-Ning Yang, Yuan Li, Nan Li, Zhen-Zhong Feng

ORCID number: Shan Zhang 0000-0001-7891-6906; Pan-Pan Yang 0000-0003-3633-6844; Yu-Chen Huang 0000-0002-7740-5634; Hong-Chun Chen 0000-0002-7751-5390; De-Li Chen 0000-0002-4593-9295; Wen-Tian Yan 0000-0001-5260-4701; Ning-Ning Yang 0000-0003-0691-7293; Yuan Li 0000-0002-8178-5131; Nan Li 0000-0002-5522-4252; Zhen-Zhong Feng 0000-0001-9385-3157.

Author contributions: Zhang S and Feng ZZ designed the study; Zhang S and Yang PP performed the experiments; Li Y prepared the slices; Zhang S, Huang YC, Chen HC, Yan WT, and Yang NN collected and analyzed the data; Feng ZZ and Li N reviewed the histopathology and IHC results; Chen DL provided critical comments on the manuscript; Zhang S, Yang PP, and Feng ZZ wrote the manuscript; all authors read and approved the final manuscript.

Supported by the Anhui Provincial Natural Science Foundation, No. 1908085MH275; Bengbu Medical College Key projects of Natural Science Foundation, No. BYKF1710; and Bengbu City-Bengbu Medical College Joint Science and Technology Project,

Shan Zhang, Department of Pathology, The Second People's Hospital of Hefei, Hefei 230011, Anhui Province, China

Pan-Pan Yang, Zhen-Zhong Feng, Department of Pathology, The Second Affiliated Hospital of Anhui Medical University, Hefei 230601, Anhui Province, China

Yu-Chen Huang, Hong-Chun Chen, Wen-Tian Yan, Ning-Ning Yang, Nan Li, Department of Pathology, The First Affiliated Hospital of Bengbu Medical College, Bengbu 233000, Anhui Province, China

De-Li Chen, Department of Gastrointestinal Surgery, The First Affiliated Hospital of Bengbu Medical College, Bengbu 233000, Anhui Province, China

Yuan Li, Department of Pathology, The First Affiliated Hospital of University of Science and Technology of China, Anhui Provincial Hospital, Hefei 230001, Anhui Province, China

Corresponding author: Zhen-Zhong Feng, PhD, Professor, Department of Pathology, The Second Affiliated Hospital of Anhui Medical University, No. 678 Furong Road, Hefei 230601, Anhui Province, China. fzz18297301626@163.com

# Abstract

# BACKGROUND

Perivascular epithelioid cell tumor (PEComa) is an uncommon tumor of mesenchymal origin. Cases of PEComa in the liver are extremely rare.

# AIM

To analyze the clinicopathological features and treatment of hepatic PEComa and to evaluate the prognosis after different treatments.

# **METHODS**

Clinical and pathological data of 26 patients with hepatic PEComa were collected. All cases were analyzed by immunohistochemistry and clinical follow-up.

# RESULTS

This study included 17 females and 9 males, with a median age of 50 years.



#### No. BYLK201812.

Institutional review board statement: This study was approved by the Ethics and Research Committees of the First Affiliated Hospital of Bengbu Medical College (Anhui Province, China).

Informed consent statement:

Informed written consent was obtained from all the patients.

Conflict-of-interest statement: The authors declare no conflicts of interest for this manuscript.

#### Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Unsolicited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: China

## Peer-review report's scientific quality classification

Grade A (Excellent): A Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Received: March 29, 2021 Peer-review started: March 29, 2021 First decision: May 28, 2021 Revised: May 29, 2021 Accepted: August 13, 2021 Article in press: August 13, 2021 Published online: September 21, 2021

Lesions were located in the left hepatic lobe in 13 cases, in the right lobe in 11, and in the caudate lobe in 2. The median tumor diameter was 6.5 cm. Light microscopy revealed that the tumor cells were mainly composed of epithelioid cells. The cytoplasm contained heterogeneous eosinophilic granules. There were thick-walled blood vessels, around which tumor cells were radially arranged. Immunohistochemical analysis of pigment-derived and myogenic markers in PEComas revealed that 25 cases were HMB45 (+), 23 were Melan-A (+), and 22 SMA (+). TFE3 and Desmin were negative in all cases. All the fluorescence in situ hybridization samples were negative for TFE3 gene break-apart probe. Tumor tissues were collected by extended hepatic lobe resection or simple hepatic tumor resection as the main treatments. Median follow-up was 62.5 mo. None of the patients had metastasis or recurrence, and there were no deaths due to the disease.

# CONCLUSION

Hepatic PEComa highly expresses melanin and smooth muscle markers, and generally exhibits an inert biological behavior. The prognosis after extended hepatic lobe resection and simple hepatic tumor resection is semblable.

Key Words: Hepatic tumor; Perivascular epithelioid cells; PEComa; Immunohistochemistry; Treatment; Prognosis

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Hepatic perivascular epithelioid cell tumor (PEComa) exhibits an inert biological behavior, and its diagnosis, treatment, and follow-up are challenging. Our study revealed that there was no difference in the prognosis between simple resection of liver tumor and extended resection of liver lobe. Optimal surgical resection currently is the best treatment option, and radiotherapy, chemotherapy, and immunotherapy may become more effective in future. The number of cases in the current retrospective study was limited by the rarity of hepatic PEComa. Therefore, further multicenter, largercohort studies are warranted to investigate the clinicopathological features and biological behavior of hepatic PEComa.

Citation: Zhang S, Yang PP, Huang YC, Chen HC, Chen DL, Yan WT, Yang NN, Li Y, Li N, Feng ZZ. Hepatic perivascular epithelioid cell tumor: Clinicopathological analysis of 26 cases with emphasis on disease management and prognosis. World J Gastroenterol 2021; 27(35): 5967-5977

URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5967.htm DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5967

# INTRODUCTION

Perivascular epithelioid cells were first described in 1992 by Bonetti et al[1]. In 2013, the World Health Organization<sup>[2]</sup> defined perivascular epithelioid cell tumor (PEComa) as "a mesenchymal tumor, which shows a local association with the vessel wall and usually expresses melanocyte markers and smooth muscle markers." Bonetti *et al*[1] were the first to propose the concept of a PEComa family, which includes angiomyolipoma, clear cell sugar tumor of the lung, lymphangioleiomyomatosis, and a group of histologically and immunophenotypically similar tumors that include primary extrapulmonary sugar tumor, clear cell myomelanocytic tumor, and abdominopelvic sarcoma of perivascular epithelioid cells. PEComas are mainly composed of eosinophilic and clear epithelioid cells, which are usually arranged in nests of different sizes associated with blood vessels[3,4]. The diagnosis of PEComa relies on its pathological features, including epithelioid cellular shapes with ample clear to eosinophilic cytoplasm, and in some cases, arrangement around thick-walled blood vessels and immunohistochemical phenotypes, including melanocyte and smooth muscle markers[1,4,5]. Cases of PEComa in the liver are extremely rare[6], and surgical resection currently is the most effective therapeutic strategy to cure patients or prolong





the survival period. In this study, the clinical and pathological features, immunohistochemical phenotypes, and information on treatment modalities of 26 cases of hepatic PEComa were collected, and the effects of different surgical methods on prognosis were evaluated to provide information for the guidance of clinical treatment.

# MATERIALS AND METHODS

#### Patient selection

The study included 17 women and 9 men who were diagnosed with hepatic PEComa for the first time. Tumor tissue samples were collected at the time of diagnosis between January 2010 and December 2018 at the First Affiliated Hospital of Bengbu Medical College (Anhui Province, China). None of the patients received preoperative radio- or chemo-therapy. Sixteen patients underwent extended hepatic lobe resection, eight underwent simple hepatic tumor resection, and two received the oral mTOR inhibitor sirolimus. None of the 26 patients had metastasis or recurrence, and there were no deaths due to the disease. Only two patients with extended liver lobectomy had a poor prognosis (one had postoperative pain in the liver area, and the other was diagnosed with liver cancer 2 years after surgery). Informed consent was obtained from all patients. The study protocol was approved by the ethics committees of the hospitals partaking in this study.

#### Imageological examination

Imaging data of all patients were collected and reviewed by two experienced physicians who analyzed the imaging characteristics of the patients.

## Histological observation and immunohistochemical analysis

Two experienced pathologists reviewed hematoxylin and eosin-stained sections of each tissue sample, marked the representative regions of tissue blocks, and assessed the following histological features: Tumor boundary (infiltration), tumor cell structure (trabecular and nested), tumor cell type (epithelial and fusiform), cytological features (cytoplasm and nucleus), nuclear features (atypical and pleomorphic), presence of pleomorphic tumor cells, and tumor necrosis.

Immunohistochemical staining was conducted on 4-µm-thick serial PEComa tissue sections using the standard ElivisionTM Plus/HRP detection system (Fuzhou Maixin Biotechnology, Fuzhou, China) and DAB substrate, generating a brown color. The antibodies, clones, dilutions, and pretreatment conditions used, as well as the positively stained sites, are listed in Table 1. Serial sections were incubated in parallel with rabbit IgG instead of the primary antibody as a negative control. Immunoreactivity was graded according to the percentage of positive tumor cells (0, negative; 1+, 1%-5%; 2+, 6%-25%; 3+, 26%-50%; 4+, 51%-100%), and tumor cell immunoreactivity was also semi-qualitatively graded: Weak, heterogeneous, or strong[7,8]. For calculation of IHC totals, a score of 1+ with weak, heterogeneous, or strong staining was considered positive for all antibodies except TFE3. A minimum of 3+ was required for TFE3 immunopositivity[8].

#### Fluorescence in situ hybridization

FISH was performed on paraffin-embedded tissue sections with a thickness of 4 µm and labeled with a TFE3 gene break-apart probe (Guangzhou Anbiping Medical, Guangzhou, Guangdong Province, China). For probe preparation, TFE3 gene was labeled with green fluorescence on the centromere side and red fluorescence on the telomere side. FISH interpretation criteria are as follows: The positive pattern for *TFE3* translocation should be 1 red, 1 green, and 1 fusion (yellow) signal in females, and 1 red, 1 green, and 1 negative signal in males; the pattern for intact *TFE3* alleles should be 2 fusion (yellow) signals in females and 1 fusion (yellow) signal in males. When the distance between the red and green signals exceeds 1 fusion signal size, it is interpreted as a red-green signal separation. A case was scored as positive if at least 10% of 100 scored nuclei showed a split signal pattern.

Zhang S et al. Clinicopathological features and prognosis of hepatic PEComa

Table 1 Antibodies used in this study					
Antigen	Clone	Dilution Antigen retrieval		Localization	
HMB-45	HMB-45	1:400	None	Cytoplasm	
Melan-A	A103	1:200	Citrate buffer pressure cook	Cytoplasm	
SMA	1A4	1:20000	None	Cytoplasm	
Desmin	D33	1:500	None	Cytoplasm	
S100 protein	Polyclonal	1:4000	Citrate buffer pressure cook	Cytoplasm/nucleus	
Hepatocyte	OCH1E5	1:1000	Citrate buffer pressure cook	Cytoplasm	
Vimentin	V9	1:200	Citrate buffer pressure cook	Cytoplasm	
CD34	QBEnd/10	1:500	Citrate buffer pressure cook	Cell membrane	
TFE-3	MRQ-0663	1:500	ETDA buffer pressure cook	Nucleus	
Ki-67	MX006	1:200	Citrate buffer pressure cook	Nucleus	

# RESULTS

#### **Clinical features**

The clinical and pathological data for all 26 cases are summarized in Table 2. We enrolled 26 patients, including 17 females and 9 males. The median patient age was 50 years (range, 26-77 years). Of the 26 patients, 23 had liver-occupying lesions, 2 had hepatic hemangioma, and 1 had hepatic hamartoma. Six patients had a history of liver disease (cysts, hamartoma, or hemangioma). The most common site of tumors was the left hepatic lobe. Sixteen patients underwent extended hepatic lobe resection, eight underwent simple hepatic tumor resection, and two were treated only with the mTOR inhibitor sirolimus (both patients were treated for 8 mo). The clinical symptoms of hepatic PEComa were non-specific. Most patients were admitted to one of our hospitals because of space-occupying lesions in the liver during medical examination, nausea, vomiting, loss of appetite, or weight loss. During physical examination, the abdomen was soft, with no tenderness or rebound tenderness, occasional contact with the ribs at the liver margin, and no pain in the liver area. Some patients experienced compression pain under the ribs and xiphoid, or in the right abdomen when the tumor involved the caudate lobe, or in the right kidney.

#### Imaging findings

B-ultrasound usually revealed strong echoes in the liver, the boundary was clear, and the internal echo was uneven, suggesting that the liver had substantial spaceoccupying lesions (data not shown). Plain computed tomography (CT) scans commonly revealed an irregular soft tissue density (Figure 1A). Enhanced scanning in the arterial phase revealed obvious enhancement of the mass edge and of central heterogeneity (Figure 1B). Portal vein scanning revealed a low mass density (Figure 1C). Magnetic resonance imaging (MRI) revealed a solid cystic space in the liver, and tumors had clear boundaries and uneven internal signal (data not shown).

#### Macroscopic features

The median tumor diameter was 6.5 cm (range, 0.5-13.0 cm). PEComa tumors were located in the liver parenchyma and were round or oval. The surface was smooth and occasionally highlighted the surface of the liver. The boundary was clear and appeared to be enveloped. Tumors did not invade the surrounding tissue. The cut surface was solid and grayish yellow, had a slightly hard texture, and showed loose necrotic tissue in the center. The liver tissue surrounding the tumor was normal, and the lymph nodes in the hilar region were not swollen. Focal hemorrhage and necrosis were seen in two cases.

#### Microscopic features

Microscopically, the tumor cells were clearly distinct from normal liver cells, and were largely composed of proliferating epithelioid cells and spindle cells, nested in trabeculae or lamellae. In most cases, the tumor cell nest was surrounded by capillaries. Tumor cells were arranged radially around the thick-walled blood vessels (Figure 2A). Tumor cells were polygonal and cytoplasm was translucent, with hetero-



Tab	le 2 Clinicop	athological fea	tures of the 26 c	cases of hepatic PEComa		
No.	Sex/age (yr)	Tumor location	Tumor size (cm)	First diagnosis	Treatment	Follow-up (mo) and prognosis
1	F/40	Left lobe	2.5	Left lobe occupying lesion	Left hepatic tumor simple resection	91, favorable prognosis
2	M/57	Left lobe	7.5	Left lobe occupying lesion	Left hepatic tumor simple resection	80, favorable prognosis
3	F/58	Left lobe	8.5	Left lobe occupying lesion	Left hepatic tumor simple resection	79, favorable prognosis
4	F/48	Right lobe	8.0	Right lobe occupying lesion	Right hepatic tumor simple resection	69, favorable prognosis
5	F/64	Right lobe	7.0	Right lobe occupying lesion	Right hepatic tumor simple resection	66, favorable prognosis
6	M/72	Right lobe	8.0	Right lobe occupying lesion	Right hepatic tumor simple resection	59, favorable prognosis
7	F/26	Right lobe	3.0	Right hepatic hamartoma	Extended hepatic lobe resection	55, favorable prognosis
8	M/47	Right lobe	6.5	Right lobe occupying lesion	mTOR inhibitor-sirolimus	51, favorable prognosis
9	F/47	Left lobe	5.5	Left lobe occupying lesion	Extended hepatic lobe resection	25, favorable prognosis
10	M/72	Right lobe	8.0	Right lobe occupying lesion	Extended right hepatic lobe resection	57, favorable prognosis
11	F/56	Right lobe	8.0	Right lobe occupying lesion	mTOR inhibitor-sirolimus	32, favorable prognosis
12	F/54	Right lobe	13.0	Left lobe occupying lesion	Extended left hepatic lobe resection	99, favorable prognosis
13	F/41	Caudate lobe	8.0	Caudate lobe occupying lesion	Caudate hepatic tumor simple resection	98, favorable prognosis
14	F/46	Left lobe	2.0	Left lobe occupying lesion	Extended left hepatic lobe resection	99, favorable prognosis
15	F/54	Right lobe	8.0	Right hepatic hemangioma	Extended Rright hepatic lobe resection	84, favorable prognosis
16	F/41	Caudate lobe	6.0	Caudate lobe occupying lesion	Extended caudate hepatic lobe resection	87, hepatic pain often occurs after discharge
17	M/45	Right lobe	0.5	Right hepatic hemangioma	Extended hepatic lobe resection	85, favorable prognosis
18	F/66	Right lobe	5.5	Right lobe occupying lesion	Extended hepatic lobe resection	59, favorable prognosis
19	F/43	Right lobe	2.8	Right lobe occupying lesion	Extended hepatic lobe resection	47, favorable prognosis
20	F/41	Left lobe	5.0	Left lobe occupying lesion	Extended hepatic lobe resection	49, reoperation for liver cancer in 2017
21	M/52	Left lobe	7.5	Left lobe occupying lesion	Left hepatic tumor simple resection	48, favorable prognosis
22	F/48	Right lobe	9.5	Right lobe occupying lesion	Extended right hepatic lobe resection	71, favorable prognosis
23	M/58	Left lobe	4.0	Left lobe occupying lesion	Left hepatic tumor simple resection	70, favorable prognosis
24	M/77	Left lobe	4.0	Left lobe occupying lesion	Extended left hepatic lobe resection	47, favorable prognosis
25	M/62	Left lobe	6.5	Left lobe occupying lesion	Extended left hepatic lobe resection	36, favorable prognosis
26	F/45	Left lobe	3.0	Left lobe occupying lesion	Extended left hepatic lobe resection	35, favorable prognosis

geneous eosinophilic particles; tumor nuclei were round or oval, nucleoli were



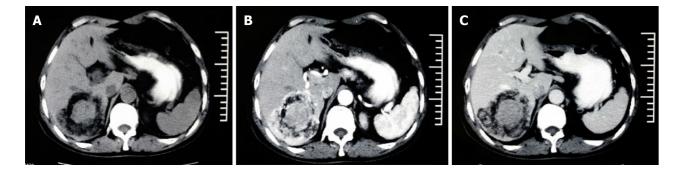


Figure 1 Computed tomography scans of the right hepatic lobe of a 72-year-old male patient with PEComa (patient 10). A: Plain computed tomography scan showing an irregular soft tissue density shadow; B: Enhanced scan showing obvious enhancement of the mass margin and of central heterogeneity in the arterial phase; C: Portal vein scan showing a low mass density.

obvious, chromatin was sparse, part of the cells were heteromorphic, and mitotic figures were not common. Collagen fibers were observed in the interstitium and were generally feathery, and a few fibers were accompanied by hemorrhage and necrosis (Figure 2B).

Immunohistochemistry findings are summarized in Table 3. Of the 26 cases, 25 were HMB45 (+), usually with multifocal or diffuse distribution and occasionally, with scattered distribution (Figure 2C), 23 were Melan-A (+) (Figure 2D), 22 were SMA (+) (Figure 2E), 20 were VIM (+), and 12 were S-100 (+). Only three cases showed focal staining (1%-5%) for TFE3. All tumors were desmin (–) (Figure 2F). The positive rate for Ki-67 was < 10%. All cases expressed at least one smooth muscle or melanocyte marker. FISH showed that no abnormal TFE3 separation signal was found in 26 cases of hepatic PEComa (Figure 3).

### Treatment and follow-up

Sixteen patients underwent extended hepatic lobe resection, eight underwent simple hepatic tumor resection, and two were treated with the mTOR inhibitor sirolimus. During a follow-up period of 25 mo to 99 mo, none of the 26 patients had metastasis or recurrence, and there were no deaths due to the disease. Only two patients with extended liver lobectomy had a poor prognosis (one had postoperative pain in the liver area, and the other was diagnosed with liver cancer 2 years after surgery). There was no difference in patient prognosis between the two surgical treatment methods, and long-term follow-up indicated that the patients went into remission.

## DISCUSSION

Hepatic PEComa is a rare mesenchymal tumor derived from pericytes. Ultrasound, CT, and MRI are commonly used for preoperative diagnosis of PEComa. On contrastenhanced CT, PEComa is characterized by vascular proliferation and arteriovenous connections[5,9,10]. MRI scans have revealed significant enhancement in PEComa in the arterial phase, but not in the portal venous and delayed phases[10]. Contrastenhanced ultrasonography is another commonly used diagnostic method, in which the contrast agent characteristically reaches the tumor rapidly and drains the arterial blood rapidly to the vein[11]. However, due to the different proportions of smooth muscle cells, adipose tissue, blood vessels, and rare tumors, the accuracy of preoperative diagnosis is currently low. In our study, only one patient was diagnosed with hepatic PEComa before undergoing surgery.

Martignoni et al[12] defined PEComa as a tumor that is composed mainly of epithelioid cells and is closely associated with dilated blood vessels and contains eosinophils, but not fat cells or disordered blood vessels. The final diagnosis of PEComa currently depends on pathological features and immunohistochemical analysis. Hepatic PEComa is mainly composed of proliferating epithelioid cells and spindle cells. The tumor cells are polygonal, have translucent cytoplasm, and contain eosinophilic particles, and thick-walled blood vessels are visible in the tumors. Epithelioid cells are arranged radially around thick-walled blood vessels. Feather-like collagen fibers are visible. Nearly all PEComas have specific immunological characteristics, with melanocyte markers (e.g., HMB-45 and/or melan-A) and smooth muscle markers (e.g., SMA) being strongly expressed [11,13], whereas desmin, hepatocyte-



Table 3 Immunohistochemical features of the 26 cases of hepatic PEComa					
Target protein	Positive cases ( <i>n</i> )/total	% Positive			
HMB45	25/26	96.2			
Melan-A	23/26	88.5			
SMA	22/26	83.6			
Desmin	1/26*	3.8			
S100	14/26	53.8			
Hepatocyte	9/26	34.6			
Vimentin	20/26	76.9			
CD34	18/26	69.2			
TFE3	0/26	0			
Ki-67 (> 10%)	1/26	3.8			

Weakly positive (1%-5%), only scattered cells.

specific antigen, and TFE3 are generally negative. In this study, 25 cases were HMB-45 (+), 17 were SMA (+), and only 3 showed focal staining (1%-5%) for TFE3.

TFE3 is a member of the MiTF family of transcription factors. A recent study[14] showed that TFE3 gene rearrangements occur in approximately 14% of PEComas. Similar to other TFE3 translocation-associated tumors, TFE3 (+) PEComa usually exhibits an acinar structure and epithelioid cell morphology, shows aggressive biological behavior, and has a poor prognosis. PSF-TFE3 gene fusion has been detected in gastrointestinal tract PEComa, but fusion partners in other cases remain unknown [15]. In this study, TFE3 expression was weak and detected in only three patients with small tumors and typical morphological PEComa images, and was associated with a low malignancy and good prognosis. Moreover, no break-apart of the TFE3 gene was detected by FISH method. Whether there is a TFE3 fusion gene still needs to be confirmed by subsequent studies. This suggests that liver PEComa may be less malignant than PEComas in other organs.

PEComas are mainly benign tumors[16] that usually do not recur after surgical resection; however, some are malignant, and their biological behavior has not been fully elucidated. In 2005, Folpe et al[17] reviewed 26 cases of PEComa of soft tissue and gynecological origin, and suggested to classify PEComa into benign, uncertain malignant potential, and malignant. Further, the authors proposed seven evaluation criteria for PEComa malignancies: (1) Tumor size > 5 cm; (2) Infiltration and growth into surrounding normal tissue; (3) High nuclear grade; (4) Excessive cells; (5) Mitotic figures in > 1/50 high-power fields; (6) Coagulative necrosis of tumor; and (7) Vascular invasion. PEComas with two or more of these features are considered to be malignant, and tumors with only nuclear polymorphism, multinucleated giant cells, or tumors > 5 cm in size are considered to have malignant potential[18].

Because of the rare disease types and the scarcity of cases, treatment plans for hepatic PEComa can only be developed based on statistical analysis of a small number of cases. Surgical resection currently is the main means of treating hepatic PEComa. In clinical practice, surgical methods are usually selected based on the tumor size and on whether the tumor is benign or malignant. Larger and malignant tumors are removed by extended hepatic lobe resection, whereas simple hepatic tumor resection is used for smaller or benign tumors. In this study, the 26 cases showed clinical and biological manifestations of inertness, and no morphological criteria for malignant PEComa. Sixteen patients underwent extended hepatic lobe resection, eight underwent simple hepatic tumor resection, and two received sirolimus. The survival rate of the patients treated with the three different modalities was good, and there was no significant difference among the treatments. Hepatic pain complications were reported only in a few cases with extended lobe resection. It has been reported that when the tumor diameter is less 5 cm, resection can be suspended or regular follow-up suffices[18].

Current data do not support that chemo- or radio-therapy improves the survival time in patients with PEComa<sup>[12]</sup>; however, sirolimus is expected to improve outcomes either when used alone or in combination with other treatments [4,10,19,20]. A 31-year-old woman with hepatic PEComa showed a significant reduction in tumor

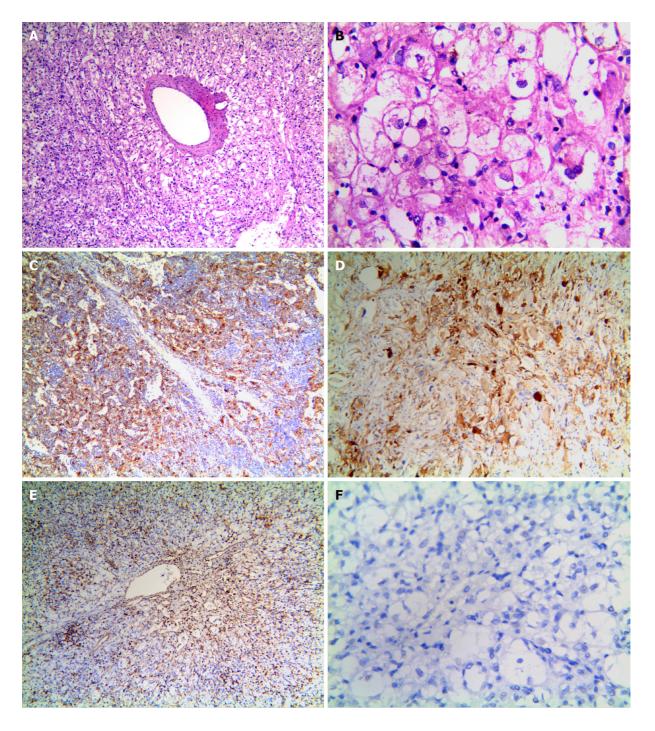


Figure 2 Morphologic appearance of hepatic PEComa. A: Tumor cells consists of proliferating epithelioid cells nested in trabeculae or lamellae and radially arranged around thick-walled vessels (HE, magnification: 100 ×); B: Tumor cells are polygonal, have translucent cytoplasm, and contain uneven eosinophilic granules. Nuclei are round or oval, with a clear nucleolus and sparse chromatin. Interstitial collagen fibers are feathery (HE, magnification, 400 ×); C: Immunoreactivity for HMB45 was detected in the cytoplasm of tumor cells in contrast to normal liver cells, which were negative for this marker (magnification, 100 ×); D: Increased expression of Melan-A was observed in both the cytoplasm and nuclei of carcinoma cells, whereas normal cells displayed lower expression of this marker (magnification, 100 ×); E: Vimentin was detected in the cytoplasm of tumor cells (magnification, 100 ×), whereas normal tissues were negative for this marker (magnification, 100 ×); F: Desmin immunoreactivity was not detected in tumor cells and normal tissues (magnification, 400 ×). Elivision<sup>TM</sup> Plus/HRP was used.

volume after 8 mo of treatment with sirolimus[19]. After subsequent surgical resection, there were no complications and the prognosis was favorable. This suggests that hepatic PEComa has a better prognosis when surgery is combined with chemotherapy [13,14]. In addition, Wagner *et al*[21] treated three patients with PEComa with sirolimus and found that the tumors responded to the drug, suggesting that sirolimus can be used alone or in combination to treat PEComa. Italiano *et al*[22] reported similar efficacy in a number of cases. However, large-scale clinical trials are needed. Numerous previous studies and this study showed that hepatic PEComa displays an inert biological behavior. However, due to the heterogeneous nature of PEComa, the

Baishidena® WJG | https://www.wjgnet.com

September 21, 2021 Volume 27 Issue 35

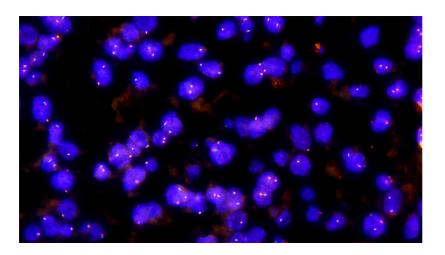


Figure 3 FISH detection of TFE3 gene break-apart in hepatic PEComa. Most of the tumor cells show fused (yellow) signals, and the distance between the red and green signals is less than 1 fusion signal. For each sample, 100 cells were counted. Only less than 10% of tumor cells showed break-apart signals (magnification, 1000 ×).

existing diagnostic criteria cannot accurately determine the nature of this tumor, which has led to overtreatment in some cases. In addition, because the nature of hepatic PEComa is not entirely clear, there is no standard treatment, and it is difficult to develop an optimal treatment plan. Therefore, clinical observation and follow-up of more cases, and the establishment of a clinical online registration system for hepatic PEComa are needed to provide clinical data for future exploration of the differentiation and distribution of the disease and the development of more accurate diagnostic criteria.

## CONCLUSION

Hepatic PEComa is a rare mesenchymal tumor that exhibits an inert biological behavior, and its diagnosis, treatment, and follow-up are challenging. Our study of 26 cases of hepatic PEComa revealed that there was no difference in the prognosis between simple resection of liver tumor and extended resection of liver lobe. Optimal surgical resection currently is the best treatment option, and radiotherapy, chemotherapy, and immunotherapy may become more effective in future[4,9]. The number of cases in the current retrospective study was limited by the rarity of hepatic PEComa. Therefore, further multicenter, larger-cohort studies are warranted to investigate the clinicopathological features and biological behavior of hepatic PEComa.

## ARTICLE HIGHLIGHTS

## Research background

Perivascular epithelioid cell tumor (PEComa) is an uncommon tumor of mesenchymal origin. Cases of PEComa in the liver are extremely rare.

## Research motivation

Cases of PEComa in the liver are extremely rare, and surgical resection currently is the most effective therapeutic strategy to cure patients or prolong the survival period. In this study, the clinical and pathological features, immunohistochemical phenotypes, and information on treatment modalities of 26 cases of hepatic PEComa were collected, and the effects of different surgical methods on prognosis were evaluated to provide information for the guidance of clinical treatment.

## Research objectives

We aimed to analyze the clinicopathological features and treatment of hepatic PEComa and to evaluate the prognosis after different treatments.



### Research methods

Clinical and pathological data of 26 patients with hepatic PEComa were collected. All cases were analyzed by immunohistochemistry and clinical follow-up.

#### Research results

This study included 17 females and 9 males, with a median age of 50 years. Lesions were located in the left hepatic lobe in 13 cases, in the right lobe in 11, and in the caudate lobe in 2. The median tumor diameter was 6.5 cm. Light microscopy revealed that the tumor cells were mainly composed of epithelioid cells. The cytoplasm contained heterogeneous eosinophilic granules. There were thick-walled blood vessels, around which tumor cells were radially arranged. Immunohistochemical analysis of pigment-derived and myogenic markers in PEComa tumors revealed that 25 cases were HMB45 (+), 23 were Melan-A (+), and 22 SMA (+). TFE3 and Desmin were negative in all cases. All the FISH samples were negative for TFE3 gene break-apart probe. Tumor tissues were collected by extended hepatic lobe resection or simple hepatic tumor resection as the main treatments. Median follow-up was 62.5 mo. None of the patients had metastasis or recurrence, and there were no deaths due to the disease.

## Research conclusions

Hepatic PEComa is a rare mesenchymal tumor that exhibits an inert biological behavior, and its diagnosis, treatment, and follow-up are challenging. Our study of 26 cases of hepatic PEComa revealed that there was no difference in the prognosis between simple resection of liver tumor and extended resection of liver lobe. Optimal surgical resection currently is the best treatment option, and radiotherapy, chemotherapy, and immunotherapy may become more effective in future.

## Research perspectives

The number of cases in the current retrospective study was limited by the rarity of hepatic PEComa. Therefore, further multicenter, larger-cohort studies are warranted to investigate the clinicopathological features and biological behavior of hepatic PEComa.

## REFERENCES

- Bonetti F, Pea M, Martignoni G, Zamboni G. PEC and sugar. Am J Surg Pathol 1992; 16: 307-308 [PMID: 1599021 DOI: 10.1097/00000478-199203000-00013]
- 2 Abhirup B, Kaushal K, Sanket M, Ganesh N. Malignant hepatic perivascular epithelioid cell tumor (PEComa) - Case report and a brief review. J Egypt Natl Canc Inst 2015; 27: 239-242 [PMID: 26071321 DOI: 10.1016/j.jnci.2015.05.004]
- 3 Hornick JL, Fletcher CD. PEComa: what do we know so far? *Histopathology* 2006; 48: 75-82 [PMID: 16359539 DOI: 10.1111/j.1365-2559.2005.02316.x]
- 4 Parfitt JR, Bella AJ, Izawa JI, Wehrli BM. Malignant neoplasm of perivascular epithelioid cells of the liver. Arch Pathol Lab Med 2006; 130: 1219-1222 [PMID: 16879028 DOI: 10.5858/2006-130-1219-MNOPEC]
- 5 Fang SH, Zhou LN, Jin M, Hu JB. Perivascular epithelioid cell tumor of the liver: a report of two cases and review of the literature. World J Gastroenterol 2007; 13: 5537-5539 [PMID: 17907305 DOI: 10.3748/wjg.v13.i41.5537]
- 6 Strzelczyk JM, Durczynski A, Szymanski D, Jablkowski M, Dworniak D, Sporny S. Primary perivascular epithelioid cell tumor (PEComa) of the liver: report of a case. Surg Today 2009; 39: 916-921 [PMID: 19784736 DOI: 10.1007/s00595-009-3945-5]
- Schoolmeester JK, Dao LN, Sukov WR, Wang L, Park KJ, Murali R, Hameed MR, Soslow RA. 7 TFE3 translocation-associated perivascular epithelioid cell neoplasm (PEComa) of the gynecologic tract: morphology, immunophenotype, differential diagnosis. Am J Surg Pathol 2015; 39: 394-404 [PMID: 25517951 DOI: 10.1097/PAS.00000000000349]
- Schoolmeester JK, Howitt BE, Hirsch MS, Dal Cin P, Quade BJ, Nucci MR. Perivascular epithelioid 8 cell neoplasm (PEComa) of the gynecologic tract: clinicopathologic and immunohistochemical characterization of 16 cases. Am J Surg Pathol 2014; 38: 176-188 [PMID: 24418852 DOI: 10.1097/PAS.00000000000133]
- Högemann D, Flemming P, Kreipe H, Galanski M. Correlation of MRI and CT findings with histopathology in hepatic angiomyolipoma. Eur Radiol 2001; 11: 1389-1395 [PMID: 11519547 DOI: 10.1007/s003300000750]
- Tan Y, Xiao EH. Hepatic perivascular epithelioid cell tumor (PEComa): dynamic CT, MRI, 10 ultrasonography, and pathologic features--analysis of 7 cases and review of the literature. Abdom Imaging 2012; 37: 781-787 [PMID: 22278345 DOI: 10.1007/s00261-012-9850-1]



- Akitake R, Kimura H, Sekoguchi S, Nakamura H, Seno H, Chiba T, Fujimoto S. Perivascular 11 epithelioid cell tumor (PEComa) of the liver diagnosed by contrast-enhanced ultrasonography. Intern Med 2009; 48: 2083-2086 [PMID: 20009396 DOI: 10.2169/internalmedicine.48.2133]
- 12 Martignoni G, Pea M, Reghellin D, Zamboni G, Bonetti F. PEComas: the past, the present and the future. Virchows Arch 2008; 452: 119-132 [PMID: 18080139 DOI: 10.1007/s00428-007-0509-1]
- 13 Bergamo F, Maruzzo M, Basso U, Montesco MC, Zagonel V, Gringeri E, Cillo U. Neoadjuvant sirolimus for a large hepatic perivascular epithelioid cell tumor (PEComa). World J Surg Oncol 2014; 12: 46 [PMID: 24575738 DOI: 10.1186/1477-7819-12-46]
- Argani P, Aulmann S, Illei PB, Netto GJ, Ro J, Cho HY, Dogan S, Ladanyi M, Martignoni G, 14 Goldblum JR, Weiss SW. A distinctive subset of PEComas harbors TFE3 gene fusions. Am J Surg Pathol 2010; 34: 1395-1406 [PMID: 20871214 DOI: 10.1097/PAS.0b013e3181f17ac0]
- 15 Tanaka M, Kato K, Gomi K, Matsumoto M, Kudo H, Shinkai M, Ohama Y, Kigasawa H, Tanaka Y. Perivascular epithelioid cell tumor with SFPQ/PSF-TFE3 gene fusion in a patient with advanced neuroblastoma. Am J Surg Pathol 2009; 33: 1416-1420 [PMID: 19606011 DOI: 10.1097/PAS.0b013e3181a9cd6c]
- Selvaggi F, Risio D, Claudi R, Cianci R, Angelucci D, Pulcini D, D'Aulerio A, Legnini M, Cotellese 16 R, Innocenti P. Malignant PEComa: a case report with emphasis on clinical and morphological criteria. BMC Surg 2011; 11: 3 [PMID: 21272348 DOI: 10.1186/1471-2482-11-3]
- Folpe AL, Mentzel T, Lehr HA, Fisher C, Balzer BL, Weiss SW. Perivascular epithelioid cell 17 neoplasms of soft tissue and gynecologic origin: a clinicopathologic study of 26 cases and review of the literature. Am J Surg Pathol 2005; 29: 1558-1575 [PMID: 16327428 DOI: 10.1097/01.pas.0000173232.22117.37
- 18 Tang D, Wang J, Tian Y, Li Q, Yan H, Wang B, Xiong L. Hepatic perivascular epithelioid cell tumor: Case report and brief literature review. Medicine (Baltimore) 2016; 95: e5572 [PMID: 28002331 DOI: 10.1097/MD.000000000005572]
- Khaja F, Carilli A, Baidas S, Sriharan A, Norford S. PEComa: A Perivascular Epithelioid Cell 19 Tumor in the Liver-A Case Report and Review of the Literature. Case Rep Med 2013; 2013: 904126 [PMID: 24489554 DOI: 10.1155/2013/904126]
- Bleeker JS, Quevedo JF, Folpe AL. "Malignant" perivascular epithelioid cell neoplasm: risk 20 stratification and treatment strategies. Sarcoma 2012; 2012: 541626 [PMID: 22619565 DOI: 10.1155/2012/541626
- Wagner AJ, Malinowska-Kolodziej I, Morgan JA, Qin W, Fletcher CD, Vena N, Ligon AH, 21 Antonescu CR, Ramaiya NH, Demetri GD, Kwiatkowski DJ, Maki RG. Clinical activity of mTOR inhibition with sirolimus in malignant perivascular epithelioid cell tumors: targeting the pathogenic activation of mTORC1 in tumors. J Clin Oncol 2010; 28: 835-840 [PMID: 20048174 DOI: 10.1200/JCO.2009.25.2981]
- Italiano A, Delcambre C, Hostein I, Cazeau AL, Marty M, Avril A, Coindre JM, Bui B. Treatment 22 with the mTOR inhibitor temsirolimus in patients with malignant PEComa. Ann Oncol 2010; 21: 1135-1137 [PMID: 20215136 DOI: 10.1093/annonc/mdq044]



WJG

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 September 21; 27(35): 5978-5988

DOI: 10.3748/wjg.v27.i35.5978

**Retrospective Study** 

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

ORIGINAL ARTICLE

# Diagnosis of focal liver lesions with deep learning-based multichannel analysis of hepatocyte-specific contrast-enhanced magnetic resonance imaging

Róbert Stollmayer, Bettina K Budai, Ambrus Tóth, Ildikó Kalina, Erika Hartmann, Péter Szoldán, Viktor Bérczi, Pál Maurovich-Horvat, Pál N Kaposi

ORCID number: Róbert Stollmayer 0000-0003-4673-7588; Bettina K Budai 0000-0002-3982-7887; Ambrus Tóth 0000-0002-1150-957X; Ildikó Kalina 0000-0002-2647-9123; Erika Hartmann 0000-0001-6073-9286: Péter Szoldán 0000-0002-3808-8541: Viktor Bérczi 0000-0003-4386-2527; Pál Maurovich-Horvat 0000-0003-0885-736X; Pál N Kaposi 0000-0002-7150-3495.

Author contributions: Stollmayer R designed and performed the research and wrote the paper; Budai BK and Tóth A contributed to data collection and analysis; Kalina I and Hartmann E provided clinical advice; Szoldán P contributed to the analysis; Bérczi V and Maurovich-Horvat P supervised the report; Kaposi PN designed the research, contributed to the analysis and supervised the report; all authors have read and approved the final manuscript.

## Institutional review board

statement: The present study has been approved by the institutional ethics committee of Semmelweis University (Semmelweis University Regional and Institutional Committee of Science and Research Ethics) according to

Róbert Stollmayer, Bettina K Budai, Ambrus Tóth, Ildikó Kalina, Viktor Bérczi, Pál Maurovich-Horvat, Pál N Kaposi, Department of Radiology, Medical Imaging Centre, Faculty of Medicine, Semmelweis University, Budapest 1083, Hungary

Erika Hartmann, Department of Transplantation and Surgery, Faculty of Medicine, Semmelweis University, Budapest 1082, Hungary

Péter Szoldán, MedInnoScan Research and Development Ltd., Budapest 1112, Hungary

Corresponding author: Bettina K Budai, MD, Department of Radiology, Medical Imaging Centre, Faculty of Medicine, Semmelweis University, Korányi Sándor st. 2., Budapest 1083, Hungary. budai.bettina@med.semmelweis-univ.hu

## Abstract

## BACKGROUND

The nature of input data is an essential factor when training neural networks. Research concerning magnetic resonance imaging (MRI)-based diagnosis of liver tumors using deep learning has been rapidly advancing. Still, evidence to support the utilization of multi-dimensional and multi-parametric image data is lacking. Due to higher information content, three-dimensional input should presumably result in higher classification precision. Also, the differentiation between focal liver lesions (FLLs) can only be plausible with simultaneous analysis of multisequence MRI images.

## AIM

To compare diagnostic efficiency of two-dimensional (2D) and three-dimensional (3D)-densely connected convolutional neural networks (DenseNet) for FLLs on multi-sequence MRI.

## **METHODS**

We retrospectively collected T2-weighted, gadoxetate disodium-enhanced arterial phase, portal venous phase, and hepatobiliary phase MRI scans from patients with focal nodular hyperplasia (FNH), hepatocellular carcinomas (HCC) or liver metastases (MET). Our search identified 71 FNH, 69 HCC and 76 MET. After volume registration, the same three most representative axial slices from all



the World Medical Association guidelines and Declaration of Helsinki, revised in 2000 in Edinburgh, No. SE-RKEB 136/2019.

## Informed consent statement: As

this is a retrospective study, in compliance with the Hungarian legal code, the need for written patient consent was waived by the ethics committee. Patients were not required to give informed consent to the study because the analysis used only anonymized clinical data that were obtained after each patient agreed to treatment and gave written informed consent to the MRI scan in compliance with our institutional protocol.

## Conflict-of-interest statement: The

authors have no financial relationships to disclose.

Data sharing statement: Additional anonymized data are available upon request from the corresponding author.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution-NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: Hungary

## Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0

sequences were combined into four-channel images to train the 2D-DenseNet264 network. Identical bounding boxes were selected on all scans and stacked into 4D volumes to train the 3D-DenseNet264 model. The test set consisted of 10-10-10 tumors. The performance of the models was compared using area under the receiver operating characteristic curve (AUROC), specificity, sensitivity, positive predictive values (PPV), negative predictive values (NPV), and f1 scores.

## RESULTS

The average AUC value of the 2D model (0.98) was slightly higher than that of the 3D model (0.94). Mean PPV, sensitivity, NPV, specificity and f1 scores (0.94, 0.93, 0.97, 0.97, and 0.93) of the 2D model were also superior to metrics of the 3D model (0.84, 0.83, 0.92, 0.92, and 0.83). The classification metrics of FNH were 0.91, 1.00, 1.00, 0.95, and 0.95 using the 2D and 0.90, 0.90, 0.95, 0.95, and 0.90 using the 3D models. The 2D and 3D networks' performance in the diagnosis of HCC were 1.00, 0.80, 0.91, 1.00, and 0.89 and 0.88, 0.70, 0.86, 0.95, and 0.78, respectively; while the evaluation of MET lesions resulted in 0.91, 1.00, 1.00, 0.95, and 0.95 and 0.75, 0.90, 0.94, 0.85, and 0.82 using the 2D and 3D networks, respectively.

## CONCLUSION

Both 2D and 3D-DenseNets can differentiate FNH, HCC and MET with good accuracy when trained on hepatocyte-specific contrast-enhanced multi-sequence MRI volumes.

Key Words: Artificial intelligence; Multi-parametric magnetic resonance imaging; Hepatocyte-specific contrast; Densely connected convolutional network; Hepatocellular carcinoma; Focal nodular hyperplasia

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Our study aimed to assess the performance of two-dimensional (2D) and three-dimensional (3D) densely connected convolutional neural networks (DenseNets) in the classification of focal liver lesions (FLLs) based on multi-parametric magnetic resonance imaging (MRI) with hepatocyte-specific contrast. We used multi-channel data input to train our networks and found that both 2D and 3D-DenseNets can differentiate between focal nodular hyperplasias, hepatocellular carcinomas or liver metastases with excellent accuracy. We conclude that DensNets can reliably classify FLLs based on multi-parametric and hepatocyte-specific post-contrast MRI. Meanwhile, multi-channel input is advantageous when the number of clinical cases available for model training is limited.

Citation: Stollmayer R, Budai BK, Tóth A, Kalina I, Hartmann E, Szoldán P, Bérczi V, Maurovich-Horvat P, Kaposi PN. Diagnosis of focal liver lesions with deep learning-based multi-channel analysis of hepatocyte-specific contrast-enhanced magnetic resonance imaging. World J Gastroenterol 2021; 27(35): 5978-5988

URL: https://www.wjgnet.com/1007-9327/full/v27/i35/5978.htm DOI: https://dx.doi.org/10.3748/wjg.v27.i35.5978

## INTRODUCTION

Artificial intelligence (AI)-based analysis is one of the fastest evolving fields in medical imaging, thanks to the rapid development of medical physics, electronic engineering, and computer science. The need for computer-aided diagnostics has been further amplified by the continuously increasing demand for imaging studies and the arrival of new modalities that put extra pressure on radiologists while also increasing the probability of diagnostic errors[1]. Meanwhile, deep learning (DL)-based algorithms have started to gain attention among medical researchers, since they provide excellent reproducibility and the ability to quantify aspects of imaging data unobservable to the human eye, resulting in automatically generated statistical reports and predictions, such as the potential of malignancy or metastatic spread and automated volume



Grade E (Poor): 0

Received: April 30, 2021 Peer-review started: April 30, 2021 First decision: June 23, 2021 Revised: July 7, 2021 Accepted: August 25, 2021 Article in press: August 25, 2021 Published online: September 21, 2021

P-Reviewer: Bork U S-Editor: Fan JR L-Editor: A P-Editor: Xing YX



assessment, among other uses. Nowadays, AI has become compatible with the full spectrum of imaging modalities and has evolved the capacity to diagnose lesions in various organ systems with greater accuracy than a human reader[2]. The processed data often include two-dimensional (2D) slices or three-dimensional (3D) image volumes; moreover, in the case of magnetic resonance imaging (MRI) studies, the different sequences are condensed into a multi-channel input. Due to their efficiency, convolutional neural networks (CNNs) have replaced other machine learning (ML) approaches in most image classification and segmentation tasks[3,4]. Recently, densely connected CNNs (DenseNets) have become more popular than plain CNN architectures. DenseNets use shortcut connections between the convolutional layers to facilitate gradient flow and optimize the number of trainable parameters. In return, these networks yield improved accuracy and efficiency in medical image classification tasks[5].

Focal liver lesions (FLLs) are common incidental findings during imaging studies, and the work-up often requires further diagnostic procedures, such as dynamic contrast-enhanced ultrasound, computed tomography and liver biopsy. Meanwhile, the excellent soft-tissue contrast, volumetric image acquisition and avoidance of ionizing radiation make multi-phase dynamic post-contrast MRI the primary tool for detection and characterization of liver lesions. The use of hepatocyte-specific contrast agents (HSAs), such as gadoxetic acid and gadobenate dimeglumine, further improves the sensitivity and specificity of the diagnosis of FLLs, as the enhancement characteristics of these lesions in the hepatobiliary phase (HBP) correlates with hepatocyte uptake[6,7]. Additionally, HSA-enhanced MRI is capable of detecting lesions smaller than 10 mm, making it an optimal modality for the early detection of liver metastases (METs)[8]

In the present study, we compared the performance of 2D and 3D-DenseNets in the classification of three types of FLLs, including focal nodular hyperplasia (FNH), hepatocellular carcinoma (HCC) and MET. To guarantee the highest possible prediction rate, we used HSA-enhanced multi-phase dynamic post-contrast MRI scans for the classification task. According to our knowledge, this is the first study to evaluate 2D and 3D-DenseNets for the diagnosis of FLLs and using multi-channel images combining four different MRI sequences. The reporting of this study follows the STROBE Statement checklist of items[9].

## MATERIALS AND METHODS

## Patient and MRI study selection

In our single-center study, we retrospectively collected multi-phasic MRI studies of patients with FNHs, HCCs or METs, that were acquired using Primovist (gadoxetate disodium), an HSA, from the picture archiving and communication system of the Medical Imaging Centre of our university. As this is a retrospective study, the need for written patient consent was waived by the Institutional Research Ethics Committee. The collected images were acquired between November 2017 and October 2020 using a Philips Ingenia 1.5 T scanner (Cambridge, MA, United States). T2-weighted (T2w) spectral-attenuated inversion recovery (commonly referred to as SPAIR), arterial phase (HAP), portal venous phase (PVP), and HBP scans were collected from each eligible patient for further analysis. Included lesions were either histologically confirmed or exhibited typical characteristics of the given lesion type with MRI. Patients younger than 18 years of age at the time of imaging were excluded from the study. Table 1 contains details of patient demographics, properties of each lesion class, and metastatic lesion origin.

## Data preparation and dataset creation

MRI scans were exported as DICOM files, that were then anonymized to remove the patients' social security numbers, birth date, sex, age, body weight, and date of the imaging study. Anonymized PVP and HBP files were resampled and spatially aligned to the corresponding T2w volume using BSpline as a non-rigid registration method via an open-source visualization and medical image computing software, called 3D Slicer ( www.slicer.org). 3D Slicer was also used for annotation cropping and file conversion [10,11]. Lesions were annotated by cubic regions of interest (referred to as ROIs). The lesions were then cropped from the aligned HAP, PVP, HBP, and T2w volumes using the same ROI. The cropped volumes were saved as NIfTI files, which were then combined into one four-dimensional (4D) file for each lesion (Figure 1). Cropped lesions were randomly sorted into datasets. After 10-10 lesions were added to the test



Table 1 Patient demographics, imaging properties of each lesion class, and details of metastatic lesion origin						
Patent properties	FNH	НСС	MET	Total		
Number of patients	42	13	14	69		
Age in years at imaging, mean $\pm$ SD	$45 \pm 12$	66 ± 5	$57 \pm 10$	$54 \pm 14$		
Sex						
Male	11	8	8	27		
Female	31	5	6	42		
Lesion properties						
Number	71	69	76	216		
Primary type						
CRC			21			
Leiomyosarcoma	18					
GI adenocc. or cholangiocc.			15			
Breast cc.			11			
Pancreas cc.			7			
Neuroendocrine ileum cc.			3			
Papillary thyroid cc.			1			

cc.: Carcinoma; CRC: Colorectal cancer; FNH: Focal nodular hyperplasia; GI: Gastrointestinal; HCC: Hepatocellular carcinoma; MET: Metastasis; SD: Standard deviation; T: Tesla.

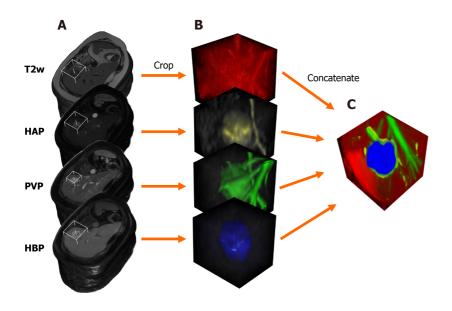
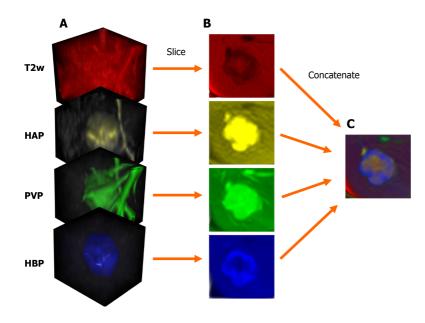
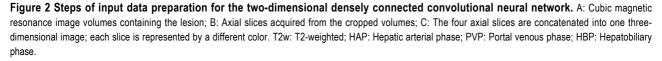


Figure 1 Steps of input data preparation for the three-dimensional densely connected convolutional neural network. A: Three-dimensionally rendered whole volumes at the level of the lesion (indicated by the white frame); B: Cropped cubic volumes containing the lesion; C: The four cubic volumes are concatenated into one four-dimensional file; each volume is represented by a different color. T2w: T2-weighted; HAP: Hepatic arterial phase; PVP: Portal venous phase; HBP: Hepatobiliary phase.

and validation dataset from each class, the remaining tumors were added to the training dataset. NIfTI files were sliced up into axial PNG images. The resulting T2w, HAP, PVP, and HBP PNG files were concatenated (Figure 2) using a custom-written computer program in Python. The training and validation datasets contained three axial slices of each lesion (*i.e.* three most representative axial slices of the NIfTI files), while the test set consisted of only one slice from each lesion.

Baishidena® WJG | https://www.wjgnet.com





### Data processing, training, and testing

Parameters of concatenated files were modified *via* transform functions. Image pixel intensity was scaled between -1.0 minimum and 1.0 maximum values. Data augmentation transforms were applied to the training samples, including random rotation (70° range along two axes) and zoom (0.7–1.4 scaling) to enrich training data. PNGs were resized to 64 × 64 resolution. Transformed images were converted to tensors (2D images were converted into 3D tensors, with the additional dimension equaling the number of network input channels), which were then fed to DenseNet264 that used 2D convolutional layers[5].

In the case of the 3D-DenseNet264 network, NIfTI voxels were resampled to isovolumetric shape, voxel intensities were rescaled between -1.0 minimum and 1.0 maximum value and NIfTI files were resized to 64 × 64 × 64 spatial resolution. The four NIfTI files were concatenated (T2w, HAP, PVP, HBP) to be used as multi-channel input for the 3D CNN. We used random 90° rotation (along two spatial axes), random 60° rotation (along x and y axes), random zoom (between 0.8 and 1.35), and random flipping on the training samples. MR volumes were converted to 4D tensors (number of channels, x-, y- and z-dimensions) that were used as network input. We used DenseNet264 models through the Pytorch-based open-source Medical Open Network For Artificial Intelligence (*i.e.* MONAI) framework[12]. We used categorical crossentropy loss to measure the prediction error of the network during training and an Adam optimizer to update model parameters[13]. Networks were trained for 70 epochs. Using a Tesla T4 graphical processing unit, the 2D network was trained for 18 min, while the 3D CNN was trained for 41 min. Validation set area under the receiver operating characteristic curve (AUROC) values were calculated after each epoch, and the model with the highest average AUC value was saved as the final model.

The trained models were used to make predictions on an independent test dataset consisting of 10 lesions from each class. The tumor type with the highest probability, according to the last softmax layer of the convolutional networks, was chosen as the predicted lesion type *via* an argmax function, encoding the predicted diagnosis as 1, while the predicted incorrect classes as 0. Specificity, sensitivity, f1 score, positive predictive value (PPV), negative predictive value (NPV) were calculated for each class based on these outputs.

Classification performance was also measured using AUC values of each class, calculated from the softmax layer probability outputs. DeLong's test was used to determine the statistical significance between the test performance of the 2D and 3D classifiers[14].

## RESULTS

The 2D model achieved the highest average validation set AUC after the 46<sup>th</sup> epoch, while the best average AUC value of the 3D network was reached after the 62<sup>nd</sup> epoch. These models were saved and then used to make test set predictions (Figure 3).

The finalized 2D and 3D networks were evaluated on the same independent test set, consisting of 10 lesions from each tumor type. On the independent test set, the finalized 2D model achieved 0.9900 [95% confidence interval (CI): 0.9664-1.0000], 0.9600 (95%CI: 0.8786-1.0000) and 0.9950 (95%CI: 0.9811-1.0000) AUC values for FNH, HCC and MET respectively, with an average AUC of 0.9783 (95%CI: 0.9492-1.0000). The finalized 3D model achieved 0.9700 (95%CI: 0.9077-1.0000), 0.9050 (95%CI: 0.7889-1.0000) and 0.9550 (95%CI: 0.8890-1.0000) AUC values for FNH, HCC and MET diagnosis, and an average AUC value of 0.9433 (95%CI: 0.8942-0.9924) on the test dataset (Figure 4). No statistically significant difference was found between the diagnostic performance of the 2D and 3D classifiers based on the ROC curve comparison for the three classes (Z = 0.7007, P = 0.4835 for FNH; Z = 0.7812, P = 0.4347for HCC; Z = 1.3069, P = 0.1913 for MET). The 2D input data achieved excellent results in the distinction between all three lesion classes, similar to the 3D network (Table 2). Both networks achieved excellent PPV, sensitivity, f1 score, NPV, and specificity values for all three classes. The highest diagnostic accuracy was achieved by both networks for FNH and MET, while both networks demonstrated lower AUC values for HCC (Table 2). PPV, sensitivity, f1 score, specificity and an NPV of 0.9091, 1.0000, 0.9524, 0.9500, 1.000 values were achieved by the 2D model for FNH diagnosis. The 3D network performed FNH classification with similar PPV (0.9000), sensitivity (0.9000), f1 score (0.9000), specificity (0.9500) and NPV (0.9500) values as the 2D network. During HCC classification both the 2D and 3D models reached acceptable metrics with PPVs of 1.000 and 0.8750, sensitivities of 0.8000 and 0.7000, f1 scores of 0.8889 and 0.7778, specificities of 1.000 and 0.9500, lastly NPVs of 0.9091 and 0.8636. For the differentiation of METs from FNHs and HCCs the use of the 2D DenseNet resulted in a PPV of 0.9091, sensitivity of 1.000, f1 score of 0.9524, specificity of 0.9500 and NPV of 1.000, while the 3D DenseNet achieved values of 0.7500, 0.9000, 0.8182, 0.8500 and 0.9444 for PPV, sensitivity, f1 score, specificity and NPV respectively. On average, both the 2D and 3D trained models could distinguish FNHs, HCCs and METs reliably with PPVs of 0.9394 and 0.8417, sensitivities of 0.9333 and 0.8333, f1 scores of 0.9312 and 0.8320, specificities of 0.9667 and 0.9167, NPVs of 0.9697 and 0.9194.

In addition, these results are supported by the extraction of attention maps from the trained models using test set images. We implemented an open-source software (M3d-CAM) to visualize the most important regions for diagnosis-making[15]. The extracted attention maps may correlate with the certainty with which a model classifies FLLs. By marking the areas within images, based on which the model makes a decision, attention maps form optimal bases of training dataset tailoring for certain radiological or other medical computer vision tasks by focusing on image regions that are difficult to analyze for the trained neural network (Figure 5).

## DISCUSSION

FLLs are common findings during liver imaging, and the differentiation of benign and malignant types of FLLs is a significant diagnostic challenge, as imaging signs may overlap between different pathologies which can substantially alter the therapeutic decision. Therefore, precise and reproducible differential diagnosis of FLLs is critical for optimal patient management.

Today, the most accurate imaging modality to diagnose FLLs is multi-phase dynamic contrast-enhanced MRI. Extracellular contrast agents (ECAs) are commonly used to perform multi-phase dynamic post-contrast MRI studies to differentiate between lesions based on their distinct contrast enhancement patterns, such as HAP hyper-enhancement or washout in the PVP[16]. In comparison to ECAs, HSAs are taken up by hepatocytes and (in part) excreted through the biliary tract; thus, they can better differentiate between those lesions that consist of functionally active and impaired hepatocytes or those that are extrahepatic in origin[7]. This behavior of HSAs is utilized for making a distinction between FNH and hepatocellular adenoma, or to detect small foci of HCC and MET within the surrounding liver parenchyma[17,18].

In the current study, we evaluated different AI models on liver MRI images for the prediction of 216 FLLs compiled from three different types of lesions, namely FNHs, HCCs and METs. To ensure that the models could achieve the highest possible



Table 2 Evaluation metrics of the two-dimensional and three-dimensional densely connected convolutional neural networks						
Input data	PPV	Sensitivity	F1 score	Specificity	NPV	
FNH 2D	0.9091	1.0000	0.9524	0.9500	1.0000	
3D	0.9000	0.9000	0.9000	0.9500	0.9500	
HCC 2D	1.0000	0.8000	0.8889	1.0000	0.9091	
3D	0.8750	0.7000	0.7778	0.9500	0.8636	
MET 2D	0.9091	1.0000	0.9524	0.9500	1.0000	
3D	0.7500	0.9000	0.8182	0.8500	0.9444	
Mean 2D	0.9394	0.9333	0.9312	0.9667	0.9697	
3D	0.8417	0.8333	0.8320	0.9167	0.9194	

2D: Two-dimensional; 3D: Three-dimensional; FNH: Focal nodular hyperplasia; HCC: Hepatocellular carcinoma; MET: Metastasis; NPV: Negative predictive value; PPV: Positive predictive value.

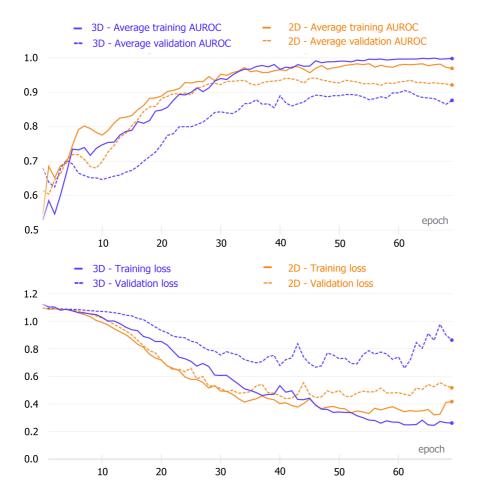


Figure 3 Comparison of the training evaluation metric curves and loss curves. The upper figure shows the area under the receiver operating characteristic curve (AUROC) values after each training epoch of the two-dimensional (2D) and three-dimensional (3D) densely connected convolutional neural networks (DenseNets). The best average AUC was obtained after the 46th (2D network) and 62nd (3D network) epochs. The lower figure indicates the loss values for each training epoch of the two networks. 2D: Two-dimensional; 3D: Three-dimensional; AUROC: Area under the receiver operating characteristic curve.

prediction rate, we narrowed down our data collection to only those four MRI sequences that provided the highest tissue contrast compared to the neighboring parenchyma or depicted distinctive imaging features of the lesion types. For the same reason, we used only HSA-enhanced scans for the analysis. We collected post-contrast images from HAP, PVP and HBP, and a T2w SPAIR image in the case of each lesion. A similar image analysis strategy was used by Hamm *et al*[19], who predicted 494 FLLs from six categories, including simple cyst, cavernous hemangioma, FNH, HCC,



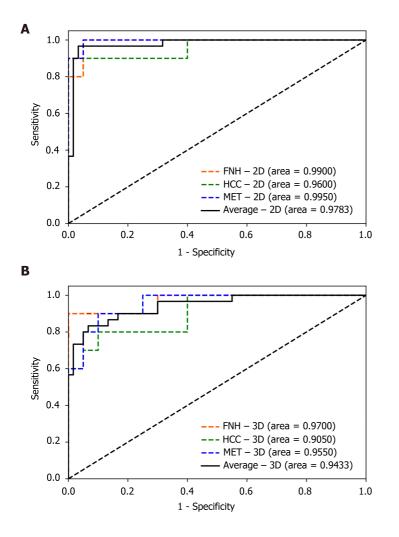


Figure 4 Receiver operating characteristic curves of the two-dimensional and three-dimensional densely connected convolutional neural network 264 models' performance on the test set. A: Two-dimensional; B: Three-dimensional. 2D: Two-dimensional; 3D: Three-dimensional; FNH: Focal nodular hyperplasia; HCC: Hepatocellular carcinoma; MET: Metastasis.

intrahepatic cholangiocarcinoma, and colorectal cancer METs using a 3D CNN model. The authors used HAP, PVP and delayed venous phase MRI images for the classification of the FLLs. They reported that the CNN model demonstrated 0.92 accuracy, 0.92 sensitivity and 0.98 specificity. The disadvantage of this study compared to ours was that it did not include HBP images, with only ECA images used for the MRI scans.

There are a handful of studies that included conventional ML methods and achieved reasonably good results. Wu *et al*[20], for example, extracted radiomics features from non-enhanced multi-parametric MRI images of FLLs and used them in ML models to differentiate between hepatic haemangioma and HCC. The final classifier achieved an AUC of 0.89, a sensitivity of 0.822 and a specificity of 0.714. Jansen *et al*[21], in their 2019 paper, used traditional ML methods for the same problem achieving an average accuracy of 0.77 for five major FLL types.

Our models' performance in the test set was comparable to or even surpassed those from previous publications, as the AUC, sensitivity and specificity were excellent for both the 2D (0.9783, 0.9333 and 0.9667 respectively) and 3D (0.9433, 0.8333 and 0.9167 respectively) architectures, which demonstrates the robustness of our data collection and analysis.

The quality and quantity of input data are pivotal when training neural networks. MRI liver tumor analysis using DL methods has steeply increased, but there is evidence lacking to support the use of 2D or 3D data. The additional dimension in 3D network inputs makes them computationally more demanding and the different data augmentation methods and hyperparameters must be well chosen to avoid artifacts. The 2D neural networks have the advantage of pretraining, which may improve classification accuracy[16,22,23]. Our study supports the results of Wang *et al*[24] and Hamm *et al*[19], emphasizing the need for multi-channel input volumes in order to achieve better accuracy. In contrast to these approaches, we have also utilized HBP



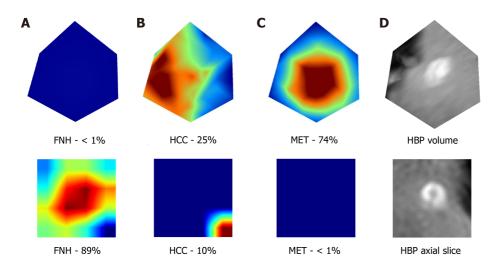


Figure 5 Visualization of the attention maps extracted from the two-dimensional and three-dimensional densely connected convolutional neural networks compared to the hepatobiliary phase input images. Two-dimensional (lower row) and three-dimensional (upper row) attention maps (column A-C) and hepatobiliary phase (column D) images were extracted from the 3rd dense block of the trained network. A-C: Two-dimensional (lower row) and three-dimensional (upper row) attention maps; D: Hepatobiliary phase images. Column A contains the attention maps for focal nodular hyperplasia (FNH), column B for hepatocellular carcinoma, and column C for metastasis diagnosis. The correct diagnosis is FNH in this case. Probabilities for different lesion classes are annotated below each attention map. The red areas are more important for the classification than other image regions. FNH: Focal nodular hyperplasia; HCC: Hepatocellular carcinoma; MET: Metastasis; HBP: Hepatobiliary phase.

> images, thereby increasing the number of input channels to four in order to improve accuracy and additionally trained 2D CNNs, proving them to be just as effective classifiers as 3D models.

> The selected architecture of the DL model can substantially alter classification accuracy. It is a novelty of our analysis that compared to previous examinations we utilized a DenseNet architecture. DenseNets contain multiple dense blocks, where each layer is connected with the residuals from previous layers. DenseNets require fewer trainable parameters at the same depth than conventional CNNs, as newly learned features are shared through all layers<sup>[5]</sup>. Our results are among the first to indicate that this highly efficient network design can enhance the performance of AI models for the classification of multi-parametric MRI images of FLLs.

> Our study's limitations are the low number of patients involved, the retrospective nature of the study, and that it was conducted within a single institute. Further improvement may be achieved by additional data collection (including additional lesion classes) and the use of more MRI volumes and different data augmentation methods as well as the use of pre-trained networks.

## CONCLUSION

Based on our study, we can say that routinely acquired radiological image materials can be used for analysis with AI methods, such as CNNs. According to our results, densely connected CNNs trained on multi-sequence MRI scans can be promising new alternatives to single-phase approaches; furthermore, the use of multi-dimensional input volumes can help the AI-based diagnosis of FLLs. According to our results, 3D and 2D DenseNets can reach similar performance in the differentiation of FLLs based on a small dataset of MRI images. The use of gadoxetate disodium-enhanced MRI scans can also enhance the diagnostic performance of MRI-based hepatic lesion classification.

## ARTICLE HIGHLIGHTS

## Research background

Interest in medical applications of artificial intelligence (AI) has steeply risen in the last few years. As one of the most obvious beneficiaries of the advances in computer vision, radiology research has also put AI in a prominent position. Convolutional



neural networks are the state-of-the-art methods used in computer vision. Focal liver lesions (FLLs) are common findings during imaging, which can best be evaluated via hepatocyte-specific contrast-enhanced magnetic resonance imaging (MRI).

## Research motivation

Though convolutional neural networks are widely used for medical image research purposes, the effect of input, such as data dimensionality and the effect of multiple input channels, has not yet been widely examined in this area. MRI volumes presumably hold more complex information about each lesion; as such, threedimensional inputs may be more difficult to process and properly use for classification tasks in comparison to two-dimensional axial slices. The combination of multiple MRI sequences in addition to the use of hepatocyte-specific contrast agents (HSAs) may also affect diagnostic accuracy.

## Research objectives

Our research aimed to compare two- and three-dimensional DenseNets264 networks for the multi-phasic hepatocyte-specific contrast-enhanced MRI-based classification of FLLs.

## Research methods

T2-weighted, arterial phase, portal venous phase, and hepatobiliary phase volumes of focal nodular hyperplasias, hepatocellular carcinomas and liver metastases were used to train the two models. Diagnostic performance was evaluated on an independent test set, based on area under the curve, positive and negative predictive values (NPVs), sensitivity, specificity and f1 score.

## Research results

The study found that *via* the use of either two- or three-dimensional convolutional neural networks and the combination of multiple MRI sequences, the average area under the curve, sensitivity, specificity, NPV, positive predictive value and f1 scores of comparable level can be achieved.

## Research conclusions

According to our findings, two- and three-dimensional networks can both be used for highly accurate differentiation of multiple classes of FLLs by combining multiple MRI phases and using HSAs.

## **Research perspectives**

This study's findings can help to clarify the potential applicability of two- and threedimensional multi-channel MRI images for the convolutional neural network-based classification of FLLs using HSAs.

## ACKNOWLEDGEMENTS

The authors would like to express their gratitude to Dr. Endre Szabó, mathematician from the Alfréd Rényi Institute of Mathematics of The Hungarian Academy of Sciences, for expert review of the manuscript and discussion of the AI analysis and statistical methods. The authors also thank Tamás Wentzel and Lilla Petovsky, technicians of the MRI unit of the Medical Imaging Center, Semmelweis University, for their enthusiasm for the current research and professionalism during patient examinations.

## REFERENCES

- Bhargavan M, Kaye AH, Forman HP, Sunshine JH. Workload of radiologists in United States in 2006-2007 and trends since 1991-1992. Radiology 2009; 252: 458-467 [PMID: 19508987 DOI: 10.1148/radiol.2522081895]
- 2 Bi WL, Hosny A, Schabath MB, Giger ML, Birkbak NJ, Mehrtash A, Allison T, Arnaout O, Abbosh C, Dunn IF, Mak RH, Tamimi RM, Tempany CM, Swanton C, Hoffmann U, Schwartz LH, Gillies RJ, Huang RY, Aerts HJWL. Artificial intelligence in cancer imaging: Clinical challenges and applications. CA Cancer J Clin 2019; 69: 127-157 [PMID: 30720861 DOI: 10.3322/caac.21552]
- 3 Kim J, Min JH, Kim SK, Shin SY, Lee MW. Detection of Hepatocellular Carcinoma in Contrast-



Enhanced Magnetic Resonance Imaging Using Deep Learning Classifier: A Multi-Center Retrospective Study. *Sci Rep* 2020; **10**: 9458 [PMID: 32527998 DOI: 10.1038/s41598-020-65875-4]

- 4 Singh SP, Wang L, Gupta S, Goli H, Padmanabhan P, Gulyás B. 3D Deep Learning on Medical Images: A Review. Sensors (Basel) 2020; 20 [PMID: 32906819 DOI: 10.3390/s20185097]
- 5 Huang G, Liu Z, Pleiss G, Van Der Maaten L, Weinberger K. Convolutional Networks with Dense Connectivity. *IEEE Trans Pattern Anal Mach Intell* 2019 [PMID: 31135351 DOI: 10.1109/TPAMI.2019.2918284]
- 6 Kim YY, Park MS, Aljoqiman KS, Choi JY, Kim MJ. Gadoxetic acid-enhanced magnetic resonance imaging: Hepatocellular carcinoma and mimickers. *Clin Mol Hepatol* 2019; 25: 223-233 [PMID: 30661336 DOI: 10.3350/cmh.2018.0107]
- 7 Thian YL, Riddell AM, Koh DM. Liver-specific agents for contrast-enhanced MRI: role in oncological imaging. *Cancer Imaging* 2013; 13: 567-579 [PMID: 24434892 DOI: 10.1102/1470-7330.2013.0050]
- 8 Coenegrachts K. Magnetic resonance imaging of the liver: New imaging strategies for evaluating focal liver lesions. *World J Radiol* 2009; 1: 72-85 [PMID: 21160723 DOI: 10.4329/wjr.v1.i1.72]
- 9 von Elm E, Altman DG, Egger M, Pocock SJ, Gøtzsche PC, Vandenbroucke JP; STROBE Initiative. The Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement: guidelines for reporting observational studies. *J Clin Epidemiol* 2008; **61**: 344-349 [PMID: 18313558 DOI: 10.1016/j.jclinepi.2007.11.008]
- 10 Fedorov A, Beichel R, Kalpathy-Cramer J, Finet J, Fillion-Robin JC, Pujol S, Bauer C, Jennings D, Fennessy F, Sonka M, Buatti J, Aylward S, Miller JV, Pieper S, Kikinis R. 3D Slicer as an image computing platform for the Quantitative Imaging Network. *Magn Reson Imaging* 2012; 30: 1323-1341 [PMID: 22770690 DOI: 10.1016/j.mri.2012.05.001]
- 11 Klein S, Staring M, Pluim JP. Evaluation of optimization methods for nonrigid medical image registration using mutual information and B-splines. *IEEE Trans Image Process* 2007; 16: 2879-2890 [PMID: 18092588 DOI: 10.1109/tip.2007.909412]
- 12 Zenodo. The MONAI Consortium. Project MONAI. [cited 10 March 2021]. Available from: https://www.f6publishing.com/Forms/Manuscript/Editorial/ReviewAndEditProcess.aspx?id=WJG-27-5978
- 13 Kingma DP, Ba J. Adam: A method for stochastic optimization. 2014 Preprint. Available from: arXiv:1412.6980v9
- 14 DeLong ER, DeLong DM, Clarke-Pearson DL. Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. *Biometrics* 1988; 44: 837-845 [PMID: 3203132]
- 15 Gotkowski K, Gonzalez C, Bucher A, Mukhopadhyay A. M3d-CAM: A PyTorch library to generate 3D data attention maps for medical deep learning. 2020 Preprint. Available from: arXiv:2007.00453v1
- 16 Matos AP, Velloni F, Ramalho M, AlObaidy M, Rajapaksha A, Semelka RC. Focal liver lesions: Practical magnetic resonance imaging approach. *World J Hepatol* 2015; 7: 1987-2008 [PMID: 26261689 DOI: 10.4254/wjh.v7.i16.1987]
- 17 Granata V, Fusco R, de Lutio di Castelguidone E, Avallone A, Palaia R, Delrio P, Tatangelo F, Botti G, Grassi R, Izzo F, Petrillo A. Diagnostic performance of gadoxetic acid-enhanced liver MRI vs multidetector CT in the assessment of colorectal liver metastases compared to hepatic resection. *BMC Gastroenterol* 2019; **19**: 129 [PMID: 31340755 DOI: 10.1186/s12876-019-1036-7]
- 18 Grieser C, Steffen IG, Kramme IB, Bläker H, Kilic E, Perez Fernandez CM, Seehofer D, Schott E, Hamm B, Denecke T. Gadoxetic acid enhanced MRI for differentiation of FNH and HCA: a single centre experience. *Eur Radiol* 2014; 24: 1339-1348 [PMID: 24658870 DOI: 10.1007/s00330-014-3144-7]
- 19 Hamm CA, Wang CJ, Savic LJ, Ferrante M, Schobert I, Schlachter T, Lin M, Duncan JS, Weinreb JC, Chapiro J, Letzen B. Deep learning for liver tumor diagnosis part I: development of a convolutional neural network classifier for multi-phasic MRI. *Eur Radiol* 2019; 29: 3338-3347 [PMID: 31016442 DOI: 10.1007/s00330-019-06205-9]
- 20 Wu J, Liu A, Cui J, Chen A, Song Q, Xie L. Radiomics-based classification of hepatocellular carcinoma and hepatic haemangioma on precontrast magnetic resonance images. *BMC Med Imaging* 2019; 19: 23 [PMID: 30866850 DOI: 10.1186/s12880-019-0321-9]
- 21 Jansen MJA, Kuijf HJ, Veldhuis WB, Wessels FJ, Viergever MA, Pluim JPW. Automatic classification of focal liver lesions based on MRI and risk factors. *PLoS One* 2019; 14: e0217053 [PMID: 31095624 DOI: 10.1371/journal.pone.0217053]
- 22 Liu Z, Tang H, Lin Y, Han SJapa. Point-voxel cnn for efficient 3d deep learning. 2020 Preprint. Available from: arXiv:1907.03739v2
- 23 Morid MA, Borjali A, Del Fiol G. A scoping review of transfer learning research on medical image analysis using ImageNet. *Comput Biol Med* 2021; **128**: 104115 [PMID: 33227578 DOI: 10.1016/j.compbiomed.2020.104115]
- 24 Wang C, Hamm C, Letzen B, Duncan J. A probabilistic approach for interpretable deep learning in liver cancer diagnosis. SPIE Medical Imaging. United States, California, San Diego: SPIE, 2019

Zaishideng® WJG | https://www.wjgnet.com



## Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

