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EDITORIAL

Importance of BRCA mutation for the current treatment of pancreatic cancer beyond maintenance

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Abstract

In this editorial, we comment on pancreatic cancer (PC), one of the most aggressive and lethal cancers. Only minimal improvements in survival rates have been achieved over recent years. Available chemotherapeutic regimens have little impact, and surgical resection remains the only reliable curative approach. We address current treatment options for these patients, focusing on the usefulness of breast cancer (BRCA) gene mutation as a prognostic biomarker and predictor of response to chemotherapy. Superior survival outcomes have been reported in patients with PC and mutant BRCA gene treated with first-line platinum-based chemotherapy. Therefore, it appears appropriate to include BRCA gene status among clinical criteria used to select the chemotherapy regimen. In addition, maintenance treatment with poly(ADP-ribose) polymerase inhibitors has been found to improve progression-free survival in patients with PC and mutated BRCA whose disease does not progress after first-line platinum-based chemotherapy. This combination has therefore been proposed as the optimal treatment regimen for these patients.

Key Words: Pancreatic cancer; Treatment; BRCA; Mutation; Poly(ADP-ribose) polymerase inhibitor; Maintenance

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Core Tip: Pancreatic cancer remains one of the most lethal malignant neoplasms, and available treatments have several limitations. Genetic studies are not currently recommended to support treatment selection. However, breast cancer (BRCA) gene



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mutation has been associated with superior survival outcomes in patients treated with platinum-based chemotherapy. Hence, it appears appropriate to consider the BRCA gene status of patients with this cancer among clinical criteria for the selection of firstline chemotherapy regimen.

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INTRODUCTION

Pancreatic cancer (PC) continues to be one of the most lethal malignant neoplasms, with a 5-year survival rate of only 5%[1]. It currently represents the third cause of cancer-related mortality and is expected to be the second by 2030[2]. Surgery is considered the sole potentially curative treatment; however, only 20% of patients diagnosed with PC are candidates for surgery at the time of diagnosis, and surgical resection is frequently followed by recurrence and therapeutic resistance[3].

There have been only limited advances in PC treatment over the past few decades. However, progress in basic research has recently generated increased molecular information on PC, improving knowledge of its biology and helping to explain the poor effectiveness of current therapies and the therapeutic resistance observed[4]. For instance, KRAS^[5], breast cancer (BRCA)^[6], and ataxia-telangiectasia mutated^[7] genes play a major role in the prognosis and response to treatment of patients with advanced PC. Hence, the identification of patients with mutations in these genes can support the design of individualized therapies that may improve survival outcomes.

In this article, we evaluate the usefulness of *BRCA* gene mutation as a prognostic and predictive biomarker of the response to chemotherapy in PC patients, beyond their maintenance treatment.

CURRENT ADVANCED PC TREATMENT

Two first-line chemotherapy options are currently available for advanced PC, FOLFIRINOX and gemcitabine+nab-paclitaxel (GEM+Nab-P)[8]. These have both demonstrated superior overall survival (OS), progression-free survival (PFS), and response rates (RRs) compared to patients receiving monotherapy with gemcitabine. Specifically, the PRODIGE4/ACCORD11 study reported improved OS (11.1 vs 6.8 mo, respectively), PFS (6.4 vs 3.3 mo), and RR (31.6% vs 9.4%) in the FOLFIRINOX vs gemcitabine arm[9]. The MPACT study also reported improved OS (8.7 vs 6.6 mo), PFS (5.5 vs 3.7 mo), and RR (23% vs 7%) with GEM+Nab-P vs gemcitabine alone[10]. The higher percentage improvements obtained in the PRODIGE4/ACCORD11 study may be explained by the more favorable prognosis of the participants, who were less representative of the real-life clinical setting compared to those in the MPACT study. Specifically, the functional Eastern Cooperative Oncology Group score was 0 in 37% of PRODIGE4/ACCORD11 study participants vs 16% of MPACT study participants, the pancreatic head was tumor site in < 40% of the former vs 44% of the latter (60%-65% in clinical practice), the mean number of metastatic sites was two in the former vs three in the latter, the carbohydrate antigen 19-9 marker was elevated in 42% of the former vs52% of the latter, and no patient over the age of 76 years participated in the former study. It should be noted that the higher survival and RRs in the PRODIGE4/ ACCORD11 study were accompanied by a significant increase in hematologic and non-hematologic toxicity. This explains why FOLFIRINOX is frequently administered at a reduced dose or in modified form in the clinical setting. Finally, no randomized trials have been undertaken to compare these options, hampering evaluation of the optimal first-line treatment of PC. The only published studies have a retrospective or non-randomized prospective design, and the results have been contradictory[11-13]. Consequently, the choice of chemotherapy regimen largely depends on clinical variables, such as the performance status and previous comorbidities of patients[14,



15].

There are currently no recommendations for genetic studies to support the selection of PC treatments. One promising approach is the identification of mutations in genes involved in response mechanisms to DNA damage, such as BRCA, whose mutation has been associated with superior OS outcomes in PC patients treated with platinumbased chemotherapy[16]. This evidence is based on multiple in vitro studies and is supported by the longer OS observed in patients with advanced PC treated with platinum-based regimens who were BRCA mutation carriers than in those who were not (14 mo vs 5 mo; hazard ratio [HR] = 0.58; P = 0.08); however, this clinical trial was retrospective and only included 12 patients[17].

ROLE OF POLY(ADP-RIBOSE) POLYMERASE AND ITS IMPORTANCE IN MUTATED BRCA

Mutations in the genetic code must be detected and repaired to preserve genome integrity, avoiding the uncontrolled proliferation of healthy cells and possible development of cancer^[18]. One DNA repair pathway detects single-strand DNA breaks. If defective, another pathway is involved in the detection of double-strand DNA breaks followed by their homologous recombination repair (HRR), using sister chromatids to restore the original DNA sequence in a high-fidelity mechanism[19]. Nuclear enzyme poly(ADP-ribose) polymerase (PARP) is responsible for detecting DNA damage and facilitating its repair. Specifically, PARP1, the main member of the PARP family, binds to and repairs both single- and double-strand DNA breaks^[20]. Conversely, PARP1 inhibition results in persistent single-strand DNA breaks that lead to replication bifurcations and double-strand DNA breaks^[21].

About 7% of PC patients possess BRCA mutations^[22]. In these patients, the inhibition of PARP and resulting loss by the tumor of functional DNA repair pathways can synergically interact and produce the specific death of tumor cells. Studies in patients with ovarian, prostate, or breast cancer found that PARP inhibition enhances the activity of cytotoxic DNA agents including alkylating agents, topoisomerase inhibitors, and radiotherapeutic agents[23]. Hence, it appears plausible to assume distinct biological behaviors and responses to therapy in patients with advanced PC who have BRCA mutations, especially germline mutations. This has implications for the treatment selection and suggests that BRCA mutations may be a useful biomarker to predict the response to first-line treatment with platinum.

PLATINUM-BASED CHEMOTHERAPY IN PATIENTS WITH PC AND MUTATED BRCA GENE

Tumors with BRCA mutations are phenotypically characterized by their susceptibility to platinum-based chemotherapy, as noted above. BRCA-deficient cells accumulate double-strand DNA breaks, generating genomic instability and a greater predisposition to malignant transformation and progression. This is because the loss of HRR and PARP1 pathways leads to so-called synthetic lethality during DNA replication [24].

Patients with ovarian cancer who had mutated BRCA, either of somatic or germline origin, respond better to platinum-based chemotherapy regimens, with a superior prognosis and survival rate compared to those without this mutation[25]. In a study of 549 patients with metastatic PC, 78% of whom had at least one family member with a history of cancer, a median OS (mOS) of 8.1 mo (95% confidence interval [CI]: 7.5-9.0) was achieved by platinum-based chemotherapy, and 31% remained alive at 1 year. The mOS was higher in the patients with a family history of breast or ovarian cancer (8.5 mo; HR = 0.76; P = 0.042) and even higher in those with a family history of pancreatic and breast or ovarian cancer (14.8 mo; HR = 0.43; P = 0.0003)[17]. According to these findings, a substantial subpopulation of patients with PC could benefit from platinum-based regimens. However, the underlying molecular mechanisms have not yet been elucidated, and further research is warranted in patients with BRCA mutant/deficient profiles. Other studies of PC patients receiving platinum-based chemotherapy have described a longer OS in those with a family history of breast, ovarian, or PC than in those with no family history of these cancers [26].

Cells with mutated BRCA are more susceptible to platinum and anthracyclines, which are selectively lethal in cells with HRR defects^[27]. In a retrospective study of 36 PC patients treated with FOLFIRINOX, multivariate analyses confirmed a significantly longer mOS in patients with vs without homologous repair gene mutations (odds ratio [OR] = 1.47; 95% CI: 1.04-2.06; P = 0.04)[17]. In a study by Lowery et al[28] of 15 patients with advanced PC and germline BRCA mutation (BRCA1 in 4 [27%] and BRCA2 in 11 [73%]), 6 received platinum chemotherapy as first-line treatment, and 5 of these had a radiological partial response according to RECIST criteria, while the remaining patient had a complete response to the infusion of 5-fluorouracil, leucovorin, oxaliplatin, and irinotecan[28]. A study by Golan et al[26] of 71 patients with PC and a mutation in BRCA1 (n = 21), BRCA2 (n = 49), or both (n = 1) reported a longer mOS in patients treated with platinum than in those receiving other agents (22 vs 4.4 mo, respectively); the authors concluded that outcomes are more favorable in patients with PC who have BRCA1 or BRCA2 mutations than in those who do not[26].

USEFULNESS OF IPARP IN PC WITH MUTATED BRCA

In the highly influential POLO study [29], 154 patients with PC and germline BRCA mutation who showed no disease progression after at least 16 wk with FOLFIRINOX were randomly assigned to a group receiving maintenance therapy with olaparib, a PARP inhibitor (iPARP), or to a group receiving no maintenance treatment. Patients in the olaparib group showed a statistically significant improvement in PFS (7.4 mo vs 3.8 mo; HR = 0.53; 95%CI: 0.35-0.82), and 22% of them remained progression-free at 2 years compared to 9.6% in the untreated group, although there was no between-group difference in OS (18.9 vs 18.1 mo, respectively). Accordingly, the United States Food and Drug Administration approved olaparib as maintenance therapy for patients with advanced PC and germline BRCA mutation who show no disease progression after at least 16 wk of first-line treatment with platinum-based chemotherapy. In this regard, it has been reported that the iPARP-associated response does not depend on the germline or somatic origin of the BRCA mutation. Thus, one meta-analysis^[22] describes eight studies that reported a response to PARPi in 24/43 (55.8%) patients with somatic BRCA mutation vs 69/157 (43.9%) patients with germline BCRA mutation, a non-significant difference (P = 0.399). In addition, five studies in the metaanalysis found no difference in PFS between patients with somatic vs germline BCRA mutations. The authors concluded that the response to iPARP therapy is similar between these types of patient.

Platinum-based chemotherapy has been combined with the administration of iPARP. An open-label, randomized, multicenter phase II trial was conducted on the efficacy of cisplatin plus gemcitabine with vs without veliparib in 50 patients with PC and germline-mutated BRCA. The RR was 74.1% for cisplatin plus gemcitabine with veliparib vs 65.2% for cisplatin plus gemcitabine alone (P = 0.55), obtaining a disease control rate of 100% with the former regimen vs 78.3% with the latter (P = 0.02). According to the authors, cisplatin plus gemcitabine is effective in advanced germlinemutated BRCA PC, and the addition of veliparib offers no improvement in therapeutic response[30-32]. These results support the selection of platinum-based chemotherapy as first-line treatment for patients with PC and germline BRCA mutation.

Various clinical trials are currently exploring the combination of iPARP with different chemotherapy and immunotherapy regimens (Table 1). It has been proposed that PARP inhibition induces tumor immunogenicity by increasing the tumor antigen load and the expression of programmed death-ligand 1 in tumor tissue, thereby increasing the susceptibility of patients with BRCA mutations to immunotherapy, as already demonstrated in breast cancer[33], small cell lung cancer[34], and ovarian cancer[35].

In summary, current studies suggest that BRCA mutation status may be a useful prognostic and predictive biomarker of the response to platinum in patients with PC, identifying those who may benefit from platinum-based chemotherapy as standard first-line treatment.

CLINICAL IMPLICATIONS

PC is associated with a poor prognosis and high resistance to chemotherapy. Few cytotoxic agents have demonstrated activity against this tumor, including platinumbased (FOLFIRINOX) and gemcitabine-based (GEM+Nab-P) regimens, and they



Table 1 Clinical trials on the combination of poly(ADP-ribose) polymerase inhibitor with chemotherapy and immunotherapy				
Identifier	Phase	iPARP	Title	Status
NCT04548752	II	Olaparib	Randomized Phase II Clinical Trial of Olaparib + Pembrolizumab vs Olaparib Alone as Maintenance Therapy in Metastatic Pancreatic Cancer Patients with Germline BRCA1 or BRCA2 Mutations	Recruiting
NCT02890355	Π	Veliparib	Randomized Phase II Study of 2 nd Line FOLFIRI <i>vs</i> Modified FOLFIRI With PARP Inhibitor ABT-888 (Veliparib) (NSC-737664) in Metastatic Pancreatic Cancer	Active, not recruiting
NCT01585805	П	Veliparib	A Randomized Phase II Study of Gemcitabine, Cisplatin +/- Veliparib in Patients with Pancreas Adenocarcinoma and known BRCA/ PALB2 Mutation (Part I) and a Phase II Single Arm Study of Single-Agent Veliparib in Previously Treated Pancreas Adenocarcinoma (Part II)	Active, not recruiting
NCT01489865	I/II	ABT-888	A Phase I/II Study of ABT-888 in combination with 5-fluorouracil and Oxaliplatin (Modified FOLFOX-6) in Patients with Metastatic Pancreatic Cancer	Active, not recruiting
NCT03404960	I/II	Niraparib + Nivolumab Niraparib + Ipilimumab	PARPVAX: A Phase 1b/2, Open Label Study of Niraparib Plus either Ipilimumab or Nivolumab in Patients with Advanced Pancreatic Cancer whose disease has not progressed on Platinum-based Therapy	Recruiting
NCT03553004	II	Niraparib	Niraparib in Metastatic Pancreatic Cancer after previous Chemotherapy (NIRA-PANC)	Recruiting

iPARP: Poly(ADP-ribose) polymerase inhibitor.

deliver very modest benefits to the patient. There has been no comparative study of these agents to determine which is more appropriate as a first-line treatment, and this decision relies on the clinical characteristics and comorbidities of the patients. Two important issues must still be resolved: the best regimen for the personalization and optimization of first-line chemotherapy in patients with PC; and the ideal sequencing of chemotherapy lines, taking into account the accumulated toxicity and the molecular profile of the cancer.

As noted above, the BRCA gene encodes proteins essential for repairing doublestrand DNA damage via the HRR pathway, and its mutation has been found to predict the response to first-line chemotherapy with platinum plus iPARP in patients with PC [26]. Thus, patients with advanced PC and germline BRCA mutation lived significantly longer when treated with platinum vs other cytotoxic agents[17]. In addition, maintenance treatment with iPARP has been found to improve the PFS of patients with PC and mutated BRCA whose disease does not progress after first-line platinumbased chemotherapy. Taken together, these findings support the selection of platinumbased regimens as first-line treatment of patients with PC and germline BRCA mutation[30-32].

Given the lack of evidence on the optimal treatment of patients with PC, it appears appropriate to consider the presence/absence of BRCA mutation among clinical criteria for the selection of first-line chemotherapy regimen.

CONCLUSION

An appreciable number of patients with PC have a mutated BRCA gene, and the ongoing development of drugs that target DNA repair pathways may offer relevant therapeutic benefits to this little-studied but clinically important sub-population. This defect in DNA repair pathways has the potential to improve outcomes in patients undergoing platinum-based chemotherapy, assisting individualized selection of the optimal first-line regimen.

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REFERENCES

1 Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin 2020; 70: 7-30 [PMID:



31912902 DOI: 10.3322/caac.21590]

- Rahib L, Smith BD, Aizenberg R, Rosenzweig AB, Fleshman JM, Matrisian LM. Projecting cancer 2 incidence and deaths to 2030: the unexpected burden of thyroid, liver, and pancreas cancers in the United States. Cancer Res 2014; 74: 2913-2921 [PMID: 24840647 DOI: 10.1158/0008-5472.CAN-14-0155]
- Martin-Perez E, Domínguez-Muñoz JE, Botella-Romero F, Cerezo L, Matute Teresa F, Serrano T, 3 Vera R. Multidisciplinary consensus statement on the clinical management of patients with pancreatic cancer. Clin Transl Oncol 2020; 22: 1963-1975 [PMID: 32318964 DOI: 10.1007/s12094-020-02350-6]
- 4 Zeng S, Pöttler M, Lan B, Grützmann R, Pilarsky C, Yang H. Chemoresistance in Pancreatic Cancer. Int J Mol Sci 2019; 20 [PMID: 31514451 DOI: 10.3390/ijms20184504]
- 5 Buscail L, Bournet B, Cordelier P. Role of oncogenic KRAS in the diagnosis, prognosis and treatment of pancreatic cancer. Nat Rev Gastroenterol Hepatol 2020; 17: 153-168 [PMID: 32005945 DOI: 10.1038/s41575-019-0245-4]
- Luo G, Lu Y, Jin K, Cheng H, Guo M, Liu Z, Long J, Liu C, Ni Q, Yu X. Pancreatic cancer: BRCA mutation and personalized treatment. Expert Rev Anticancer Ther 2015; 15: 1223-1231 [PMID: 26402249 DOI: 10.1586/14737140.2015.1086271]
- 7 Armstrong SA, Schultz CW, Azimi-Sadjadi A, Brody JR, Pishvaian MJ. ATM Dysfunction in Pancreatic Adenocarcinoma and Associated Therapeutic Implications. Mol Cancer Ther 2019; 18: 1899-1908 [PMID: 31676541 DOI: 10.1158/1535-7163.MCT-19-0208]
- Neoptolemos JP, Kleeff J, Michl P, Costello E, Greenhalf W, Palmer DH. Therapeutic developments in pancreatic cancer: current and future perspectives. Nat Rev Gastroenterol Hepatol 2018; 15: 333-348 [PMID: 29717230 DOI: 10.1038/s41575-018-0005-x]
- Conroy T, Desseigne F, Ychou M, Bouché O, Guimbaud R, Bécouarn Y, Adenis A, Raoul JL, Gourgou-Bourgade S, de la Fouchardière C, Bennouna J, Bachet JB, Khemissa-Akouz F, Péré-Vergé D, Delbaldo C, Assenat E, Chauffert B, Michel P, Montoto-Grillot C, Ducreux M; Groupe Tumeurs Digestives of Unicancer; PRODIGE Intergroup. FOLFIRINOX vs genetiabine for metastatic pancreatic cancer. N Engl J Med 2011; 364: 1817-1825 [PMID: 21561347 DOI: 10.1056/NEJMoa1011923
- 10 Von Hoff DD, Ervin T, Arena FP, Chiorean EG, Infante J, Moore M, Seay T, Tjulandin SA, Ma WW, Saleh MN, Harris M, Reni M, Dowden S, Laheru D, Bahary N, Ramanathan RK, Tabernero J, Hidalgo M, Goldstein D, Van Cutsem E, Wei X, Iglesias J, Renschler MF. Increased survival in pancreatic cancer with nab-paclitaxel plus gemcitabine. N Engl J Med 2013; 369: 1691-1703 [PMID: 24131140 DOI: 10.1056/NEJMoa1304369]
- 11 Kim S, Signorovitch JE, Yang H, Patterson-Lomba O, Xiang CQ, Ung B, Parisi M, Marshall JL. Comparative Effectiveness of nab-Paclitaxel Plus Gemcitabine vs FOLFIRINOX in Metastatic Pancreatic Cancer: A Retrospective Nationwide Chart Review in the United States. Adv Ther 2018; 35: 1564-1577 [PMID: 30209750 DOI: 10.1007/s12325-018-0784-z]
- 12 Tempero MA, Malafa MP, Chiorean EG, Czito B, Scaife C, Narang AK, Fountzilas C, Wolpin BM, Al-Hawary M, Asbun H, Behrman SW, Benson AB, Binder E, Cardin DB, Cha C, Chung V, Dillhoff M, Dotan E, Ferrone CR, Fisher G, Hardacre J, Hawkins WG, Ko AH, LoConte N, Lowy AM, Moravek C, Nakakura EK, O'Reilly EM, Obando J, Reddy S, Thayer S, Wolff RA, Burns JL, Zuccarino-Catania G. Pancreatic Adenocarcinoma, Version 1.2019. J Natl Compr Canc Netw 2019; 17: 202-210 [PMID: 30865919 DOI: 10.6004/jnccn.2019.0014]
- Williet N, Saint A, Pointet AL, Tougeron D, Pernot S, Pozet A, Bechade D, Trouilloud I, Lourenco 13 N, Hautefeuille V, Locher C, Desrame J, Artru P, Thirot Bidault A, Le Roy B, Pezet D, Phelip JM, Taieb J. Folfirinox versus gemcitabine/nab-paclitaxel as first-line therapy in patients with metastatic pancreatic cancer: a comparative propensity score study. Therap Adv Gastroenterol 2019; 12: 1756284819878660 [PMID: 31598136 DOI: 10.1177/1756284819878660]
- 14 Pavel M, Öberg K, Falconi M, Krenning EP, Sundin A, Perren A, Berruti A; ESMO Guidelines Committee. Gastroenteropancreatic neuroendocrine neoplasms: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol 2020; 31: 844-860 [PMID: 32272208 DOI: 10.1016/j.annonc.2020.03.304]
- 15 Hidalgo M, Álvarez R, Gallego J, Guillén-Ponce C, Laquente B, Macarulla T, Muñoz A, Salgado M, Vera R, Adeva J, Alés I, Arévalo S, Blázquez J, Calsina A, Carmona A, de Madaria E, Díaz R, Díez L, Fernández T, de Paredes BG, Gallardo ME, González I, Hernando O, Jiménez P, López A, López C, López-Ríos F, Martín E, Martínez J, Martínez A, Montans J, Pazo R, Plaza JC, Peiró I, Reina JJ, Sanjuanbenito A, Yaya R, Carrato A. Consensus guidelines for diagnosis, treatment and follow-up of patients with pancreatic cancer in Spain. Clin Transl Oncol 2017; 19: 667-681 [PMID: 27995549 DOI: 10.1007/s12094-016-1594-x]
- Mylavarapu S, Das A, Roy M. Role of BRCA Mutations in the Modulation of Response to Platinum 16 Therapy. Front Oncol 2018; 8: 16 [PMID: 29459887 DOI: 10.3389/fonc.2018.00016]
- Sehdev A, Gbolahan O, Hancock BA, Stanley M, Shahda S, Wan J, Wu HH, Radovich M, O'Neil 17 BH. Germline and Somatic DNA Damage Repair Gene Mutations and Overall Survival in Metastatic Pancreatic Adenocarcinoma Patients Treated with FOLFIRINOX. Clin Cancer Res 2018; 24: 6204-6211 [PMID: 30131383 DOI: 10.1158/1078-0432.CCR-18-1472]
- 18 Lord CJ, Ashworth A. PARP inhibitors: Synthetic lethality in the clinic. Science 2017; 355: 1152-1158 [PMID: 28302823 DOI: 10.1126/science.aam7344]
- Hoeijmakers JH. Genome maintenance mechanisms for preventing cancer. Nature 2001; 411: 366-19



374 [PMID: 11357144 DOI: 10.1038/35077232]

- Alemasova EE, Lavrik OI. Poly(ADP-ribosyl)ation by PARP1: reaction mechanism and regulatory 20 proteins. Nucleic Acids Res 2019; 47: 3811-3827 [PMID: 30799503 DOI: 10.1093/nar/gkz120]
- Mateo J, Lord CJ, Serra V, Tutt A, Balmaña J, Castroviejo-Bermejo M, Cruz C, Oaknin A, Kaye SB, 21 de Bono JS. A decade of clinical development of PARP inhibitors in perspective. Ann Oncol 2019; 30: 1437-1447 [PMID: 31218365 DOI: 10.1093/annonc/mdz192]
- Mohyuddin GR, Aziz M, Britt A, Wade L, Sun W, Baranda J, Al-Rajabi R, Saeed A, Kasi A. Similar 22 response rates and survival with PARP inhibitors for patients with solid tumors harboring somatic vs Germline BRCA mutations: a Meta-analysis and systematic review. BMC Cancer 2020; 20: 507 [PMID: 32493233 DOI: 10.1186/s12885-020-06948-5]
- Kamel D, Gray C, Walia JS, Kumar V. PARP Inhibitor Drugs in the Treatment of Breast, Ovarian, 23 Prostate and Pancreatic Cancers: An Update of Clinical Trials. Curr Drug Targets 2018; 19: 21-37 [PMID: 28699513 DOI: 10.2174/1389450118666170711151518]
- 24 Hu Y, Guo M. Synthetic lethality strategies: Beyond BRCA1/2 mutations in pancreatic cancer. Cancer Sci 2020; 111: 3111-3121 [PMID: 32639661 DOI: 10.1111/cas.14565]
- 25 Ray-Coquard I, Pautier P, Pignata S, Pérol D, González-Martín A, Berger R, Fujiwara K, Vergote I, Colombo N, Mäenpää J, Selle F, Sehouli J, Lorusso D, Guerra Alía EM, Reinthaller A, Nagao S, Lefeuvre-Plesse C, Canzler U, Scambia G, Lortholary A, Marmé F, Combe P, de Gregorio N, Rodrigues M, Buderath P, Dubot C, Burges A, You B, Pujade-Lauraine E, Harter P; PAOLA-1 Investigators. Olaparib plus Bevacizumab as First-Line Maintenance in Ovarian Cancer. N Engl J Med 2019; 381: 2416-2428 [PMID: 31851799 DOI: 10.1056/NEJMoa1911361]
- 26 Golan T, Kanji ZS, Epelbaum R, Devaud N, Dagan E, Holter S, Aderka D, Paluch-Shimon S, Kaufman B, Gershoni-Baruch R, Hedley D, Moore MJ, Friedman E, Gallinger S. Overall survival and clinical characteristics of pancreatic cancer in BRCA mutation carriers. Br J Cancer 2014; 111: 1132-1138 [PMID: 25072261 DOI: 10.1038/bjc.2014.418]
- 27 Tung NM, Garber JE. BRCA1/2 testing: therapeutic implications for breast cancer management. Br J Cancer 2018; 119: 141-152 [PMID: 29867226 DOI: 10.1038/s41416-018-0127-5]
- Lowery MA, Kelsen DP, Stadler ZK, Yu KH, Janjigian YY, Ludwig E, D'Adamo DR, Salo-Mullen 28 E, Robson ME, Allen PJ, Kurtz RC, O'Reilly EM. An emerging entity: pancreatic adenocarcinoma associated with a known BRCA mutation: clinical descriptors, treatment implications, and future directions. Oncologist 2011; 16: 1397-1402 [PMID: 21934105 DOI: 10.1634/theoncologist.2011-0185]
- 29 Golan T, Locker GY, Kindler HL. Maintenance Olaparib for Metastatic Pancreatic Cancer. Reply. N Engl J Med 2019; 381: 1492-1493 [PMID: 31597029 DOI: 10.1056/NEJMc1911185]
- 30 O'Reilly EM, Lee JW, Zalupski M, Capanu M, Park J, Golan T, Tahover E, Lowery MA, Chou JF, Sahai V, Brenner R, Kindler HL, Yu KH, Zervoudakis A, Vemuri S, Stadler ZK, Do RKG, Dhani N, Chen AP, Kelsen DP. Randomized, Multicenter, Phase II Trial of Gemcitabine and Cisplatin With or Without Veliparib in Patients With Pancreas Adenocarcinoma and a Germline BRCA/PALB2 Mutation. J Clin Oncol 2020; 38: 1378-1388 [PMID: 31976786 DOI: 10.1200/JCO.19.02931]
- Fazio N. Cisplatin Plus Gemcitabine as Standard of Care for Germline BRCA/PALB2-Mutated 31 Pancreatic Adenocarcinoma: Are We Moving Too Fast? J Clin Oncol 2020; 38: 2466-2467 [PMID: 32407212 DOI: 10.1200/JCO.20.004191
- 32 O'Reilly EM, Park W, Kelsen DP. Reply to N. Fazio. J Clin Oncol 2020; 38: 2467-2468 [PMID: 32407214 DOI: 10.1200/JCO.20.00833]
- Jiao S, Xia W, Yamaguchi H, Wei Y, Chen MK, Hsu JM, Hsu JL, Yu WH, Du Y, Lee HH, Li CW, 33 Chou CK, Lim SO, Chang SS, Litton J, Arun B, Hortobagyi GN, Hung MC. PARP Inhibitor Upregulates PD-L1 Expression and Enhances Cancer-Associated Immunosuppression. Clin Cancer Res 2017; 23: 3711-3720 [PMID: 28167507 DOI: 10.1158/1078-0432.CCR-16-3215]
- 34 Sen T, Rodriguez BL, Chen L, Corte CMD, Morikawa N, Fujimoto J, Cristea S, Nguyen T, Diao L, Li L, Fan Y, Yang Y, Wang J, Glisson BS, Wistuba II, Sage J, Heymach JV, Gibbons DL, Byers LA. Targeting DNA Damage Response Promotes Antitumor Immunity through STING-Mediated T-cell Activation in Small Cell Lung Cancer. Cancer Discov 2019; 9: 646-661 [PMID: 30777870 DOI: 10.1158/2159-8290.CD-18-1020
- Rocconi RP, Monk BJ, Walter A, Herzog TJ, Galanis E, Manning L, Bognar E, Wallraven G, 35 Stanbery L, Aaron P, Senzer N, Coleman RL, Nemunaitis J. Gemogenovatucel-T (Vigil) immunotherapy demonstrates clinical benefit in homologous recombination proficient (HRP) ovarian cancer. Gynecol Oncol 2021; 161: 676-680 [PMID: 33715892 DOI: 10.1016/j.ygyno.2021.03.009]



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FRONTIER

Acetyl-CoA carboxylase inhibitors in non-alcoholic steatohepatitis: Is there a benefit?

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Abstract

De novo lipogenesis (DNL) plays an important role in the pathogenesis of hepatic steatosis and also appears to be implicated in hepatic inflammation and fibrosis. Accordingly, the inhibition of acetyl-CoA carboxylase, which catalyzes the ratelimiting step of DNL, might represent a useful approach in the management of patients with nonalcoholic fatty liver disease (NAFLD). Animal studies and preliminary data in patients with NAFLD consistently showed an improvement in steatosis with the use of these agents. However, effects on fibrosis were variable and an increase in plasma triglyceride levels was observed. Therefore, more longterm studies are needed to clarify the role of these agents in NAFLD and to determine their risk/benefit profile.

Key Words: Acetyl-CoA carboxylase inhibitors; Non-alcoholic steatohepatitis; Fibrosis; Steatosis; Firsocostat

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Core Tip: Acetyl-CoA carboxylase inhibitors suppress de novo lipogenesis resulting in improvement in hepatic steatosis in both animal models and in patients with nonalcoholic fatty liver disease. However, the effects of these agents on hepatic fibrosis are inconsistent and they increase plasma triglyceride levels, casting doubt on their risk/ benefit profile.



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INTRODUCTION

Non-alcoholic fatty liver disease (NAFLD) is the commonest chronic liver disease in high-income countries, affecting 17%-46% of the general population[1]. NAFLD includes non-alcoholic fatty liver, characterized by isolated hepatic steatosis, and nonalcoholic steatohepatitis (NASH), where variable degrees of hepatic inflammation and fibrosis coexist with steatosis[2]. NASH is associated with increased risk for cirrhosis, hepatocellular cancer (HCC) and cardiovascular disease[3,4]. Diet and exercise, aiming at weight loss, is the cornerstone of management of NAFLD, but only a minority of patients achieves and maintains weight loss > 5%, which is essential for improvement in liver histology [2,5]. Several pharmacological agents have been evaluated in patients with NAFLD but none is currently licensed for use in this disease^[2]. Therefore, there is an unmet need for safe and effective treatments in patients with NASH.

NON-ALCOHOLIC FATTY LIVER DISEASE

The pathogenesis of NASH is complex and multiple pathways, including insulin resistance, inflammation, oxidative stress and apoptosis are implicated [6]. De novo lipogenesis (DNL), defined as the synthesis of fatty acids from non-lipid sources, is pivotal in the development and progression of NASH. DNL is increased in patients with NAFLD and appears to be responsible for up to 38% of intrahepatic triglyceride content in this population[7]. In addition to its contribution to the development of hepatic steatosis, DNL also promotes fibrosis by activating hepatic stellate cells (HSC), which are the principal contributors to liver fibrosis[8,9]. Acetyl-CoA carboxylase (ACC) catalyzes the ATP-dependent carboxylation of acetyl-coenzyme A (CoA) to form malonyl-CoA, which is the rate-limiting and key regulatory step in DNL[10]. ACC exists as two isoenzymes that are encoded by two different genes; ACC1 is cytosolic whereas ACC2 is located at the mitochondrial membrane[10].

Given the central role of ACC in DNL and the implication of the latter in the pathogenesis of NAFLD, ACC might represent an attractive therapeutic target in this disease. Indeed, early studies showed that liver-specific, genetic inactivation of ACC protects against the development of hepatic steatosis[11,12]. More recently, several orally available, liver-specific, dual ACC1/ACC2 inhibitors have been developed and are being evaluated in the management of NAFLD (Table 1). Perhaps the most promising is firsocostat, formerly known as GS-0976. In mice with NASH, this agent improved hepatic steatosis and also reduced hepatic inflammation[13,14]. However, an increase in serum triglyceride, glucose and insulin levels as well in total body fat mass was observed[13,14]. In another study, a structural analog of GS-0976 reduced hepatic steatosis and hepatic insulin resistance in high-fructose-fed rats[15]. However, a 30%-130% increase in plasma triglyceride levels was again observed, which was attributed to an increase in very low density lipoprotein production and a decrease in triglyceride clearance by lipoprotein lipase[15]. Other ACC inhibitors also showed promise in ameliorating hepatic steatosis in rodent models of NASH. ND-630 reduced hepatic steatosis in Zucker diabetic fatty rats[16]. In addition, PF-05221304 not only improved liver steatosis in a rat model of NASH but also reduced hepatic inflammation[17].

In addition to the reduction in hepatic steatosis, ACC inhibition also appears to ameliorate hepatic fibrosis (Table 1), which is the strongest predictor of mortality in NASH[18-20]. In recent studies, firsocostat and a structural analog of this agent inhibited the activation of HSCs and reduced hepatic fibrosis both in vitro and in animal models of NASH[9,13,14]. PF-05221304 also prevented the activation of primary HSCs to myofibroblasts in vitro and reduced fibrosis in choline-deficient, high-fat-fed rats[17]. In contrast, MK-4074 did not affect fibrosis in a rat model of NASH, suggesting that the effect of ACC inhibition on fibrosis might be agent-specific [21]. On the other hand, another liver-specific, dual ACC1/ACC2 inhibitor, ND-654,

Table 1 Major findings of preclinical and clinical studies that evaluated the effects of acetyl-CoA carboxylase inhibitors in non-alcoholic steatohepatitis

Population	ACC inhibitor	Major findings	Ref.
Mice with NASH	Firsocostat (GS-0976)	\downarrow Hepatic steatosis, inflammation and fibrosis	[13, 14]
High-fructose-fed rats	A structural analog of firsocostat	\downarrow Hepatic steatosis; \downarrow hepatic insulin resistance	[15]
Zucker diabetic fatty rats	ND-630	↓ Hepatic steatosis	[<mark>16</mark>]
Rat model of NASH	PF-05221304	\downarrow Hepatic steatosis, inflammation and fibrosis	[17]
Rat model of NASH	MK-4074	No effect on hepatic fibrosis	[<mark>21</mark>]
Rat model of NASH	ND-654	↓ Hepatic steatosis; Delayed progression of hepatocellular cancer	[22]
10 patients with NASH	Firsocostat	\downarrow Hepatic steatosis and fibrosis	[23]
126 patients with NASH	Firsocostat	↓ Hepatic steatosis and tissue inhibitor of metalloproteinase-1 levels	[24]
392 patients with NASH and bridging fibrosis or compensated cirrhosis (F3-F4)	Firsocostat	\downarrow Hepatic steatosis and stiffness	[25]
Healthy subjects	PF-05221304	Dose-dependent suppression of de novo lipogenesis	[<mark>26</mark>]
Overweight and/or obese adult males	ND-630	Suppression of de novo lipogenesis	[27]
30 patients with non-alcoholic fatty liver	MK-4074	↓ Hepatic steatosis	[28]

not only reduced hepatic steatosis but also delayed the progression of HCC in a rat model[22].

Preliminary studies suggest that ACC inhibition might also be effective in patients with NAFLD (Table 1). In a pilot, open-label, prospective study in 10 patients with NASH, administration of firsocostat for 12 wk reduced hepatic steatosis, assessed with magnetic resonance imaging (MRI), and fibrosis, assessed with both magnetic resonance elastography (MRE) and serum levels of tissue inhibitor of metalloproteinase 1 (TIMP-1)[23]. However, serum alanine aminotransferase levels did not change[23]. In a phase 2, randomized study in 126 patients with NASH, treatment with GS-0976 for 12 wk reduced hepatic steatosis, assessed with MRI, and TIMP-1 Levels more than placebo^[24]. However, changes in MRE-measured liver stiffness did not differ among groups and an 11%-13% increase in serum triglyceride levels was observed in patients treated with GS-0976[24]. In a larger, phase 2b, randomized trial in 392 patients with NASH and bridging fibrosis or compensated cirrhosis (F3-F4), the incidence of the primary endpoint (a \geq 1-stage improvement in fibrosis without worsening of NASH) did not differ between firsocostat and placebo[25]. However, firsocostat improved steatosis, increased the proportion of patients with \geq 1-grade improvement in liver histology and improved liver stiffness evaluated by transient elastography and the Enhanced Liver Fibrosis Test compared with placebo[25]. Notably, serum glucose and insulin levels as well as body weight did not change in patients treated with firsocostat [25]. On the other hand, a mean increase in serum triglyceride levels by 42 mg/dL was observed in the firsocostat group[25].

Other ACC inhibitors also showed promising results in pilot clinical studies (Table 1). In healthy subjects, PF-05221304 dose-dependently suppressed DNL and was well-tolerated[26]. With doses yielding \geq 90% DNL inhibition, asymptomatic increases in serum triglyceride levels and declines in platelet count occurred but these were not observed at \leq 80% DNL inhibition[26]. A single dose of ND-630 was also shown to suppress DNL in overweight and/or obese but otherwise healthy adult males and was well tolerated[27]. Finally, in a randomized study in 30 patients with NAFL, treatment with MK-4074 for 4 wk decreased hepatic fat more than pioglitazone and placebo[28]. However, a 2-fold increase in plasma triglyceride levels was observed in patients treated with MK-4074 and not in the other groups[28]. It was shown that inhibition of ACC results in reduced intrahepatic content of polyunsaturated fatty acids, which in turn activates sterol regulatory element-binding protein-1c that increases hepatic production of very low density lipoprotein and therefore plasma triglyceride levels[28].

CONCLUSION

In conclusion, ACC inhibitors appear to represent a promising tool for ameliorating hepatic steatosis. The effect of these agents on hepatic fibrosis is less consistent and more studies are needed to assess their impact on NASH. In addition, given the high cardiovascular risk of patients with NASH, the increase in triglyceride levels during treatment with ACC inhibitors is a cause of concern and should be also be factored in the decision to administer them in this population.

REFERENCES

- 1 Vernon G, Baranova A, Younossi ZM. Systematic review: the epidemiology and natural history of non-alcoholic fatty liver disease and non-alcoholic steatohepatitis in adults. Aliment Pharmacol Ther 2011; **34**: 274-285 [PMID: 21623852 DOI: 10.1111/j.1365-2036.2011.04724.x]
- European Association for the Study of the Liver (EASL); European Association for the Study of 2 Diabetes (EASD); European Association for the Study of Obesity (EASO). EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease. J Hepatol 2016; 64: 1388-1402 [PMID: 27062661 DOI: 10.1016/j.jhep.2015.11.004]
- 3 Haflidadottir S, Jonasson JG, Norland H, Einarsdottir SO, Kleiner DE, Lund SH, Björnsson ES. Long-term follow-up and liver-related death rate in patients with non-alcoholic and alcoholic related fatty liver disease. BMC Gastroenterol 2014; 14: 166 [PMID: 25260964 DOI: 10.1186/1471-230X-14-166
- Targher G, Day CP, Bonora E. Risk of cardiovascular disease in patients with nonalcoholic fatty liver disease. N Engl J Med 2010; 363: 1341-1350 [PMID: 20879883 DOI: 10.1056/NEJMra0912063
- Vilar-Gomez E, Martinez-Perez Y, Calzadilla-Bertot L, Torres-Gonzalez A, Gra-Oramas B, 5 Gonzalez-Fabian L, Friedman SL, Diago M, Romero-Gomez M. Weight Loss Through Lifestyle Modification Significantly Reduces Features of Nonalcoholic Steatohepatitis. Gastroenterology 2015; 149: 367-78.e5; quiz e14-5 [PMID: 25865049 DOI: 10.1053/j.gastro.2015.04.005]
- Robinson KE, Shah VH. Pathogenesis and pathways: nonalcoholic fatty liver disease & alcoholic 6 liver disease. Transl Gastroenterol Hepatol 2020; 5: 49 [PMID: 33073044 DOI: 10.21037/tgh.2019.12.05]
- Smith GI, Shankaran M, Yoshino M, Schweitzer GG, Chondronikola M, Beals JW, Okunade AL, 7 Patterson BW, Nyangau E, Field T, Sirlin CB, Talukdar S, Hellerstein MK, Klein S. Insulin resistance drives hepatic de novo lipogenesis in nonalcoholic fatty liver disease. J Clin Invest 2020; 130: 1453-1460 [PMID: 31805015 DOI: 10.1172/JCI134165]
- Mederacke I, Hsu CC, Troeger JS, Huebener P, Mu X, Dapito DH, Pradere JP, Schwabe RF. Fate 8 tracing reveals hepatic stellate cells as dominant contributors to liver fibrosis independent of its aetiology. Nat Commun 2013; 4: 2823 [PMID: 24264436 DOI: 10.1038/ncomms3823]
- Bates J, Vijayakumar A, Ghoshal S, Marchand B, Yi S, Kornyeyev D, Zagorska A, Hollenback D, 9 Walker K, Liu K, Pendem S, Newstrom D, Brockett R, Mikaelian I, Kusam S, Ramirez R, Lopez D, Li L, Fuchs BC, Breckenridge DG. Acetyl-CoA carboxylase inhibition disrupts metabolic reprogramming during hepatic stellate cell activation. J Hepatol 2020; 73: 896-905 [PMID: 32376414 DOI: 10.1016/j.jhep.2020.04.037]
- 10 Carotti S, Aquilano K, Valentini F, Ruggiero S, Alletto F, Morini S, Picardi A, Antonelli-Incalzi R, Lettieri-Barbato D, Vespasiani-Gentilucci U. An overview of deregulated lipid metabolism in nonalcoholic fatty liver disease with special focus on lysosomal acid lipase. Am J Physiol Gastrointest Liver Physiol 2020; 319: G469-G480 [PMID: 32812776 DOI: 10.1152/ajpgi.00049.2020]
- 11 Mao J, DeMayo FJ, Li H, Abu-Elheiga L, Gu Z, Shaikenov TE, Kordari P, Chirala SS, Heird WC, Wakil SJ. Liver-specific deletion of acetyl-CoA carboxylase 1 reduces hepatic triglyceride accumulation without affecting glucose homeostasis. Proc Natl Acad Sci USA 2006; 103: 8552-8557 [PMID: 16717184 DOI: 10.1073/pnas.0603115103]
- 12 Choi CS, Savage DB, Abu-Elheiga L, Liu ZX, Kim S, Kulkarni A, Distefano A, Hwang YJ, Reznick RM, Codella R, Zhang D, Cline GW, Wakil SJ, Shulman GI. Continuous fat oxidation in acetyl-CoA carboxylase 2 knockout mice increases total energy expenditure, reduces fat mass, and improves insulin sensitivity. Proc Natl Acad Sci U S A 2007; 104: 16480-16485 [PMID: 17923673 DOI: 10.1073/pnas.0706794104]
- 13 Gapp B, Jourdain M, Bringer P, Kueng B, Weber D, Osmont A, Zurbruegg S, Knehr J, Falchetto R, Roma G, Dietrich W, Valdez R, Beckmann N, Nigsch F, Sanyal AJ, Ksiazek I. Farnesoid X Receptor Agonism, Acetyl-Coenzyme A Carboxylase Inhibition, and Back Translation of Clinically Observed Endpoints of De Novo Lipogenesis in a Murine NASH Model. Hepatol Commun 2020; 4: 109-125 [PMID: 31909359 DOI: 10.1002/hep4.1443]
- Matsumoto M, Yashiro H, Ogino H, Aoyama K, Nambu T, Nakamura S, Nishida M, Wang X, Erion DM, Kaneko M. Acetyl-CoA carboxylase 1 and 2 inhibition ameliorates steatosis and hepatic fibrosis in a MC4R knockout murine model of nonalcoholic steatohepatitis. PLoS One 2020; 15: e0228212 [PMID: 31990961 DOI: 10.1371/journal.pone.0228212]
- Goedeke L, Bates J, Vatner DF, Perry RJ, Wang T, Ramirez R, Li L, Ellis MW, Zhang D, Wong KE, 15



Beysen C, Cline GW, Ray AS, Shulman GI. Acetyl-CoA Carboxylase Inhibition Reverses NAFLD and Hepatic Insulin Resistance but Promotes Hypertriglyceridemia in Rodents. Hepatology 2018; 68: 2197-2211 [PMID: 29790582 DOI: 10.1002/hep.30097]

- 16 Harriman G, Greenwood J, Bhat S, Huang X, Wang R, Paul D, Tong L, Saha AK, Westlin WF, Kapeller R, Harwood HJ Jr. Acetyl-CoA carboxylase inhibition by ND-630 reduces hepatic steatosis, improves insulin sensitivity, and modulates dyslipidemia in rats. Proc Natl Acad Sci USA 2016; 113: E1796-E1805 [PMID: 26976583 DOI: 10.1073/pnas.1520686113]
- Ross TT, Crowley C, Kelly KL, Rinaldi A, Beebe DA, Lech MP, Martinez RV, Carvajal-Gonzalez S, 17 Boucher M, Hirenallur-Shanthappa D, Morin J, Opsahl AC, Vargas SR, Bence KK, Pfefferkorn JA, Esler WP. Acetyl-CoA Carboxylase Inhibition Improves Multiple Dimensions of NASH Pathogenesis in Model Systems. Cell Mol Gastroenterol Hepatol 2020; 10: 829-851 [PMID: 32526482 DOI: 10.1016/j.jcmgh.2020.06.001]
- Ekstedt M, Hagström H, Nasr P, Fredrikson M, Stål P, Kechagias S, Hultcrantz R. Fibrosis stage is 18 the strongest predictor for disease-specific mortality in NAFLD after up to 33 years of follow-up. Hepatology 2015; 61: 1547-1554 [PMID: 25125077 DOI: 10.1002/hep.27368]
- 19 Vilar-Gomez E, Calzadilla-Bertot L, Wai-Sun Wong V, Castellanos M, Aller-de la Fuente R, Metwally M, Eslam M, Gonzalez-Fabian L, Alvarez-Quiñones Sanz M, Conde-Martin AF, De Boer B, McLeod D, Hung Chan AW, Chalasani N, George J, Adams LA, Romero-Gomez M. Fibrosis Severity as a Determinant of Cause-Specific Mortality in Patients With Advanced Nonalcoholic Fatty Liver Disease: A Multi-National Cohort Study. Gastroenterology 2018; 155: 443-457.e17 [PMID: 29733831 DOI: 10.1053/j.gastro.2018.04.034]
- Dulai PS, Singh S, Patel J, Soni M, Prokop LJ, Younossi Z, Sebastiani G, Ekstedt M, Hagstrom H, 20 Nasr P, Stal P, Wong VW, Kechagias S, Hultcrantz R, Loomba R. Increased risk of mortality by fibrosis stage in nonalcoholic fatty liver disease: Systematic review and meta-analysis. Hepatology 2017; 65: 1557-1565 [PMID: 28130788 DOI: 10.1002/hep.29085]
- Zhang J, Muise ES, Han S, Kutchukian PS, Costet P, Zhu Y, Kan Y, Zhou H, Shah V, Huang Y, 21 Saigal A, Akiyama TE, Shen XL, Cai TQ, Shah K, Carballo-Jane E, Zycband E, Yi L, Tian Y, Chen Y, Imbriglio J, Smith E, Devito K, Conway J, Ma LJ, Hoek M, Sebhat IK, Peier AM, Talukdar S, McLaren DG, Previs SF, Jensen KK, Pinto S. Molecular Profiling Reveals a Common Metabolic Signature of Tissue Fibrosis. Cell Rep Med 2020; 1: 100056 [PMID: 33205063 DOI: 10.1016/j.xcrm.2020.100056]
- Lally JSV, Ghoshal S, DePeralta DK, Moaven O, Wei L, Masia R, Erstad DJ, Fujiwara N, Leong V, 22 Houde VP, Anagnostopoulos AE, Wang A, Broadfield LA, Ford RJ, Foster RA, Bates J, Sun H, Wang T, Liu H, Ray AS, Saha AK, Greenwood J, Bhat S, Harriman G, Miao W, Rocnik JL, Westlin WF, Muti P, Tsakiridis T, Harwood HJ Jr, Kapeller R, Hoshida Y, Tanabe KK, Steinberg GR, Fuchs BC. Inhibition of Acetyl-CoA Carboxylase by Phosphorylation or the Inhibitor ND-654 Suppresses Lipogenesis and Hepatocellular Carcinoma. Cell Metab 2019; 29: 174-182.e5 [PMID: 30244972 DOI: 10.1016/j.cmet.2018.08.020]
- Lawitz EJ, Coste A, Poordad F, Alkhouri N, Loo N, McColgan BJ, Tarrant JM, Nguyen T, Han L, 23 Chung C, Ray AS, McHutchison JG, Subramanian GM, Myers RP, Middleton MS, Sirlin C, Loomba R, Nyangau E, Fitch M, Li K, Hellerstein M. Acetyl-CoA Carboxylase Inhibitor GS-0976 for 12 Weeks Reduces Hepatic De Novo Lipogenesis and Steatosis in Patients With Nonalcoholic Steatohepatitis. Clin Gastroenterol Hepatol 2018; 16: 1983-1991.e3 [PMID: 29705265 DOI: 10.1016/j.cgh.2018.04.042]
- Loomba R, Kayali Z, Noureddin M, Ruane P, Lawitz EJ, Bennett M, Wang L, Harting E, Tarrant JM, 24 McColgan BJ, Chung C, Ray AS, Subramanian GM, Myers RP, Middleton MS, Lai M, Charlton M, Harrison SA. GS-0976 Reduces Hepatic Steatosis and Fibrosis Markers in Patients With Nonalcoholic Fatty Liver Disease. Gastroenterology 2018; 155: 1463-1473.e6 [PMID: 30059671 DOI: 10.1053/j.gastro.2018.07.027
- Loomba R, Noureddin M, Kowdley KV, Kohli A, Sheikh A, Neff G, Bhandari BR, Gunn N, 25 Caldwell SH, Goodman Z, Wapinski I, Resnick M, Beck AH, Ding D, Jia C, Chuang JC, Huss RS, Chung C, Subramanian GM, Myers RP, Patel K, Borg BB, Ghalib R, Kabler H, Poulos J, Younes Z, Elkhashab M, Hassanein T, Iyer R, Ruane P, Shiffman ML, Strasser S, Wong VW, Alkhouri N; ATLAS Investigators. Combination Therapies Including Cilofexor and Firsocostat for Bridging Fibrosis and Cirrhosis Attributable to NASH. Hepatology 2021; 73: 625-643 [PMID: 33169409 DOI: 10.1002/hep.31622]
- 26 Bergman A, Carvajal-Gonzalez S, Tarabar S, Saxena AR, Esler WP, Amin NB. Safety, Tolerability, Pharmacokinetics, and Pharmacodynamics of a Liver-Targeting Acetyl-CoA Carboxylase Inhibitor (PF-05221304): A Three-Part Randomized Phase 1 Study. Clin Pharmacol Drug Dev 2020; 9: 514-526 [PMID: 32065514 DOI: 10.1002/cpdd.782]
- 27 Stiede K, Miao W, Blanchette HS, Beysen C, Harriman G, Harwood HJ Jr, Kelley H, Kapeller R, Schmalbach T, Westlin WF. Acetyl-coenzyme A carboxylase inhibition reduces de novo lipogenesis in overweight male subjects: A randomized, double-blind, crossover study. Hepatology 2017; 66: 324-334 [PMID: 28470676 DOI: 10.1002/hep.29246]
- Kim CW, Addy C, Kusunoki J, Anderson NN, Deja S, Fu X, Burgess SC, Li C, Ruddy M, 28 Chakravarthy M, Previs S, Milstein S, Fitzgerald K, Kelley DE, Horton JD. Acetyl CoA Carboxylase Inhibition Reduces Hepatic Steatosis but Elevates Plasma Triglycerides in Mice and Humans: A Bedside to Bench Investigation. Cell Metab 2017; 26: 394-406.e6 [PMID: 28768177 DOI: 10.1016/j.cmet.2017.07.009]



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REVIEW

Therapeutic resistance in pancreatic ductal adenocarcinoma: Current challenges and future opportunities

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Abstract

Pancreatic ductal adenocarcinoma (PDAC) is the third leading cause of cancerrelated deaths in the United States. Although chemotherapeutic regimens such as gemcitabine+ nab-paclitaxel and FOLFIRINOX (FOLinic acid, 5-Fluroruracil, IRINotecan, and Oxaliplatin) significantly improve patient survival, the prevalence of therapy resistance remains a major roadblock in the success of these agents. This review discusses the molecular mechanisms that play a crucial role in PDAC therapy resistance and how a better understanding of these mechanisms has shaped clinical trials for pancreatic cancer chemotherapy. Specifically, we have discussed the metabolic alterations and DNA repair mechanisms observed in PDAC and current approaches in targeting these mechanisms. Our discussion also includes the lessons learned following the failure of immunotherapy in PDAC and current approaches underway to improve tumor's immunological response.

Key Words: Pancreatic cancer; Metabolism; DNA repair; Therapy-resistance; Immunotherapy

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Core Tip: With a five-year survival rate of 10%, pancreatic adenocarcinomas are one of the most aggressive forms of cancer. Despite extensive efforts, only a few drug combinations have been found to be effective in improving patient outcomes. The drug-resistant mechanisms active in pancreatic ductal adenocarcinoma contribute to the



quality classification

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ineffectiveness of therapies. Through this review, we discuss key mechanisms that contribute to the development of resistant phenotype in pancreatic tumors and how these mechanisms are being sought as a target to treat this cancer.

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INTRODUCTION

Pancreatic ductal adenocarcinoma (PDAC) is a highly aggressive tumor, with a 5-year overall survival of 10%. As the cause of approximately 47000 deaths annually, it is the third leading cause of cancer-related mortality in the United States and is expected to be the second primary cause of cancer-related deaths by 2030[1,2]. Surgical resection of the tumor remains the only curative option for patients with PDAC. However, due to late diagnosis, only a limited number of patients qualify for it. Relapse is common and often observed as early as two months post-surgery. Therefore, adjuvant chemotherapy is often prescribed to improve patient outcomes. For over a decade, gemcitabine was the mainstay for chemotherapy for resectable PDACs. The drug advanced the patient survival to 5.65 mo compared with 4.41 mo with 5-fluorouracil[3]. Recently, a combination therapy FOLFIRINOX (FOLinic acid, 5-Fluorouracil, IRINotecan, and Oxaliplatin) displayed better patient outcomes than gemcitabine[4]. The four-drug cocktail, although toxic, significantly improved survival in PDAC patients and is currently approved for both resectable and metastatic PDAC[5-9] (Table 1).

The complex pancreatic cancer biology is often attributed as the underlying cause of the poor chemotherapeutic response. This review will highlight the current knowledge of the therapeutic resistance mechanisms prevalent in PDAC and the opportunities PDAC tumor biology provides for its efficient targeting.

CURRENT THERAPIES IN PDAC

Gemcitabine

Gemcitabine has been a mainstay for PDAC treatment since 1997, when it was found to improve median and overall survival compared to 5-fluorouracil[3]. Gemcitabine (2', 2'- difluorodeoxycytidine) is a difluoro analog of deoxycytidine which inhibits DNA synthesis through (1) inhibition of ribonucleotide reductase (RR), (2) inhibition of DNA polymerase (*via* diphosphate analog), or (3) mis-incorporation into the DNA, thus preventing chain elongation (*via* triphosphate analog)[10,11]. The inhibition of RR by the diphosphate analog depletes the deoxy-ribonucleotide pool essential for DNA synthesis.

Numerous mechanisms for gemcitabine inactivity have been demonstrated. Although resistance can be divided into innate and acquired forms, we will present evidence referring to both as "resistance" for this review.

The first interaction of gemcitabine with the cells occurs at the nucleotide transporter level. These transporters-concentrative nucleoside transporters (hCNTs) and equilibrative nucleoside transporters (hENTs) allow the transport of gemcitabine into the cells[12]. Evidence of the importance of nucleotide transporters for gemcitabine activity includes the observation that, in the absence of hENT1, PDAC patients treated with gemcitabine have reduced survival[13]. The enzyme deoxycytidine kinase (dCK) is the rate-limiting enzyme that converts gemcitabine-induced cytotoxicity[14]. Acquired resistant models demonstrate reduced expression of dCK in cells that do not respond to gemcitabine[14,15]. However, a recent analysis of the patient-derived xenograft PDAC model found no change in dCK levels in the gemcitabine-resistant tumors[16], indicating that mechanisms independent of dCK contribute to poor response to gemcitabine.

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Table 1 Landmark trials for approved pancreatic ductal adenocarcinoma therapies				
Treatment	Tumor characteristic	Primary endpoint	Ref.	
Gemcitabine	Advanced PDAC	Median survival, 5.65 mo	Burris <i>et al</i> [3]	
Gemcitabine + Erlotinib vs Gemcitabine	Locally Advanced or metastatic PDAC	Overall survival (OS), 6.24 mo vs 5.91 mo	Hoffmann et al[59]	
FOLFIRINOX vs Gemcitabine	Metastatic PDAC	OS, 11.1 mo vs 6.8 mo	Conroy et al[4]	
Gemcitabine + nab-paclitaxel vs Gemcitabine	Metastatic PDAC	OS, 8.5 mo <i>vs</i> 6.7 mo	Couvelard <i>et al</i> [60]	
Gemcitabine + Capacitabine vs Gemcitabine	Resectable PDAC	OS, 28 mo <i>vs</i> 25.5 mo	Neoptolemos et al[8]	

PDAC: Pancreatic ductal adenocarcinoma.

As mentioned earlier, when gemcitabine inhibits RR, the deoxy-ribonucleotide pool of the cells becomes depleted, leading to cell death. Overexpression of M1 and M2 isoforms, namely RRM1 and RRM2, is associated with reduced cellular response to gemcitabine[16-18]. Micro RNAs such as miR20a-5 and miR211 have been shown to downregulate RR, enhancing pancreatic cancer's sensitivity to gemcitabine and inhibiting cellular invasion[19,20]. Similarly, natural product, small molecule, and miRNA-based inhibition of RR sensitizes PDAC cells to gemcitabine[19-21-24]. Although strong in vitro data indicate RRM1/RRM2 play a key role in gemcitabine sensitivity, conflicting clinical outcomes have limited the utility of these enzymes for PDAC prognosis[25-28].

Other cellular processes such as epithelial-mesenchymal transition (EMT), mitogenic signaling, and tumor-stroma interaction also contribute to gemcitabine resistance [29]. Analysis of PDAC lines revealed that the EMT gene expression profile differs considerably between drug-sensitive and -resistant cells[30]. The drug-resistant cells showed reduced response to gemcitabine, 5-fluorouracil, and cisplatin, and expressed elevated levels of EMT marker Zeb1[30]. In addition, suppression of EMT enhanced the sensitivity of PDAC to gemcitabine by regulating the expression of nucleoside transporters[31].

5-Fluorouracil

Similar to gemcitabine, 5-fluorouracil belongs to the antimetabolite class of anti-cancer agents. 5-Fluorouracil inhibits the enzyme thymidylate synthetase (TS), which is responsible for methylation of deoxyuridine mono-phosphate to deoxythymidine mono-phosphate, a precursor for DNA synthesis. 5-Fluorouracil was the first drug to be approved as PDAC adjuvant therapy [32,33]. Although no longer used as monotherapy, 5-fluorouracil forms a part of the PDAC chemotherapeutic regimen FOL-FIRINOX. Compared to gemcitabine therapy, combination therapy with FOLFIRINOX improved the overall survival and median progression-free survival of patients with metastatic PDAC^[4]. Although any improvement in PDAC patient outcomes should be observed as a positive sign, the high toxicity of the drug regimen, limited patient eligibility for FOLFIRINOX, and prevalence of 5-fluorouracil resistant mechanisms may further limit the use this combination therapy in PDAC[34-38]. Multiple mechanisms have demonstrated to contribute to 5-fluorouracil resistance, such as alteration in (1) 5-fluorouracil metabolizing enzymes, (2) membrane transporters, and (3) pro-survival/ pro-apoptotic pathways. High TS expression is associated with poor survival in PDAC patients, however, the difference in survival is more significant in patients that received 5-fluorouracil based therapy[39,40]. The enzyme dihydropyrimidine dehydrogenase (DPD) catabolizes the 5-fluorouracil in the liver. In colorectal cancer patients receiving 5-fluorouracil based therapy, high DPD levels was associated with significantly shorter disease-free survival and overall survival^[41]. In vitro analysis of PDAC cells lines and 5-fluorouracil-resistant sub-lines revealed that high expression of TS and DPDY is associated with poor 5-fluorouracil response[42].

Targeted therapies in PDAC

Comprehensive genetic analysis has revealed that pancreatic cancers are a host of numerous genetic mutations^[43]. Mutation of K-ras is the most frequent genetic alteration observed in more than 90% of pancreatic cancer cases [44]. K-ras protein is a downstream signaling molecule activated by various transmembrane receptor tyrosine kinases, such as the epidermal growth factor receptor (EGFR), insulin-like growth factor receptor, and c-met. EGFR, overexpressed in more than 40% of pancreatic



cancers, is associated with poor disease prognosis, invasion, and aggressive clinical behavior [45,46]. Given its importance, therapies targeting EGFR have been tested to determine their ability to improve the outcomes of PDAC patients. In one phase III trial, the addition of erlotinib (EGFR tyrosine kinase inhibitor) to gemcitabine-based therapy significantly improved the overall survival of PDAC patients[47]. A recent clinical trial compared the efficacy of gemcitabine + erlotinib in rash-positive pancreatic cancer patients and found similar one-year survival and better quality of life compared to patients on FOLFIRINOX[48]. Some trials however, have failed to show the clinical benefit of adding EGFR targeting drugs to PDAC chemotherapy [49-52]. Therapies targeting other molecular mechanisms active in pancreatic cancer have not shown beneficial effects, and EGFR targeting may have a place in PDAC therapy as precision medicine[53-57].

FUTURE OPPORTUNITIES TO TARGET PDAC

Pancreatic tumor metabolism

Pancreatic cancer is characterized by a dense stroma surrounding the tumor. This dense stromal region limits vascularization, creating an environment limiting oxygen and nutrient supply [58,59]. Limited oxygen gives rise to hypoxia that is associated with poor patient prognosis[59-61]. In an abundance of oxygen, the non-malignant cells produce most of their energy from mitochondrial oxidative phosphorylation (OXPHOS) while cancer cells exhibit an altered metabolism, first observed in the 1920s by Warburg[62], in which they produce most of their energy from glycolysis. Further, Warburg^[62] observed that the majority of the glucose taken up by the cancer cells is converted to lactate rather than CO_{2} an observation that has since been witnessed and verified by various researchers in various tumors, including PDAC[63-70]. Pancreatic cancer shows upregulation in glycolysis, pentose phosphate pathway (PPP), fatty acid synthesis, and purine/pyrimidine synthesis, and downregulation of enzymes involved in Kreb's cycle and the OXPHOS.

Analysis of the pancreatic cancer progression model revealed that the metabolic alterations precede tumor formation[71]. Metabolic rewiring in the early stages involves upregulated glycolytic and PPP. The altered metabolic profile allows quick ATP production and provides nucleotides and other metabolic intermediates required for proliferating cancer cells^[72]. However, the suppression of OXPHOS can lead to excessive acid build-up within the cancer cells in the form of lactate. To circumvent this, pancreatic cancers express monocarboxylate transporters (MCT1 and MCT4) to efflux out lactate[73,74]. These metabolic adaptations, aided by the upregulation of glucose transporters GLUT1, allow the cancer cells to utilize glucose for their energy and biosynthetic needs. In addition, the molecular biology of pancreatic cancers, such as mutation of KRAS and P53, contribute to the so-called "glycolytic switch" in the PDACs by regulating genes like hexokinase-2, glucose transporters GLUT-1, and PKM2, and by promoting anabolic processes[75-78].

Altered tumor metabolism is also associated with poor therapy response in pancreatic tumors. Acquired gemcitabine-resistant models of pancreatic cancer show a marked increase in aerobic glycolysis that maintains the EMT phenotype and reduced responsiveness to the therapeutic agent[79]. The resistant cells exhibit elevated glycolytic enzymes HK2, LDHA and PKM2, and glucose transporter GLUT1. Below we discuss the central carbon metabolic pathways – namely, glycolysis, tricarboxylic acid (TCA) cycle, and the PPP – as therapeutic targets in pancreatic cancer.

Glycolysis as therapeutic target: Analysis of pancreatic tumors reveals that HK2 expression is upregulated in localized tumors as well as metastatic tumors compared to non-malignant tissues[80]. Since HK2 plays a crucial role in pancreatic tumors, efforts have been made to evaluate HK2 as a therapeutic target for pancreatic cancers. We were among the first to show that inhibition of glycolytic enzymes HK2 inhibits the growth and pro-survival signaling in pancreatic cancers[81]. In addition, inhibition of HK2 in pancreatic cancer cells suppresses their anchorage-independent growth and invasion[80]. The role of HK2 has also been implicated in gemcitabine resistance, as HK2 dimerization is enhanced in cells that do not respond to gemcitabine[82]. In vitro and in vivo analysis revealed that inhibition of HK2 enhanced the sensitivity of PDAC to gemcitabine. Similarly, in another study, inhibition of HK2 using chemical inhibitor 2-deoxyglucose enhanced resistant cells' sensitivity to gemcitabine [79].

PKM2: Pyruvate kinase (PK) is a glycolytic enzyme that catalyzes the conversion of phosphoenol pyruvate and ADP into pyruvate and ATP. Four isoforms of the enzyme



exist in vertebrates: PKR in erythrocytes; PKL in liver and kidney; PKM1 in adult muscle, brain, and heart; and PKM2 in most adult tissues and fetal tissues[83]. Phosphorylation of PKM2 at tyrosine residue 105 (Y105) is associated with reduced PKM2 activity and enhanced tumor growth [84,85]. Analyses of PKM isoform show abundance of isoform M2 in tumor cells compared to high levels of M1 in normal tissues[52,53]. In cancer cell lines, high PKM2 Levels are associated with proliferation, metastasis, and angiogenesis[54-56]. The role of PKM2 in pancreatic tumors is, however, controversial. Using the mice model of PDAC, a recent report demonstrated that although PKM2 expression is elevated in PDAC, the loss of PKM2 does not significantly affect the size of tumors or the survival of mice bearing PDAC[86]. Surgical specimens from 115 PDAC patients show that PKM2 expression is associated with better overall survival [87]. However, others have shown that high PKM2 expression correlates with poor patient outcomes [88,89]. Considering several observations demonstrating a vital role of PKM2 in pancreatic cancer survival, invasion, angiogenesis, metastasis, and drug resistance, we believe the PKM2 serves as an attractive target for the treatment of PDAC, even though its role in pancreatic cancer tumorigenesis is still unproven[90-95].

Lactate dehydrogenase (LDH): LDH is an enzyme that exists as a tetramer and catalyzes the conversion of pyruvate to lactate and *vice versa*. LDHA (LDH gene product) regulates pyruvate's conversion to lactate, thus preventing the entry of pyruvate into the TCA cycle. Deregulated expression of LDHA is observed in various tumors, including pancreatic, gastric, bladder, cholangiocarcinoma, lung, and endometrial cancers[96-102]. Numerous oncogenic signaling molecules, namely, HIF1 alpha, myc, FOXM1, and tyrosine kinase receptors, can regulate the level or the activity of LDH[96,103-106]. Elevated levels of LDH are associated with unfavorable prognoses for PDAC patient survival, chemotherapy response, and recurrence[107-112]. Preclinical studies have revealed that inhibition of LDH reduces the survival of PDAC cells[113,114].

PPP as therapeutic target: The PPP branches from glycolysis and contributes to the cancer phenotype through (1) synthesis of NADPH (oxidative PPP), which is important for redox regulation and fatty acid synthesis, and (2) supplying the proliferating cells with pentose sugar (non-oxidative PPP) for nucleic acid biosynthesis [115]. Accumulating evidence indicates that PPP plays a vital role in pancreatic tumor survival, metastasis, and therapy resistance. Our lab and others have shown that MYC regulates the activity of both oxidative and non-oxidative PPP through the regulation of G6PD and the RPIA (non-oxidative PPP) gene[78,116,117]. The regulation of RPIA via MYC appears to be under the directive of KRAS. The MAPK-MYC-RPIA-nucleotide biosynthesis pathway is shown to be important for KRAS-mediated maintenance of PDAC[78,116]. Considering that most PDAC patients (90%) express mutant KRAS, inhibition of PPP is an attractive strategy for developing more efficient pancreatic cancer therapies that would target KRAS-induced metabolic abnormalities. Our recent results found that pancreatic cancer cells resistant to erlotinib express elevated levels of G6PD. The upregulated G6PD prevents the induction of ROS in response to erlotinib, thus protecting the cells from drug-induced cytotoxicity[117]. The non-oxidative PPP has also been implicated in PDAC therapy resistance. Shukla et al[118] found that gemcitabine-resistant cells express enhanced carbon flux into the non-oxidative PPP, aided by elevated non-oxidative PPP enzyme levels. This alteration in metabolic flux allows elevated pyrimidine synthesis that contributes to gemcitabine resistance[118].

TCA cycle and OXPHOS as therapeutic target: Although cancer cells exhibit an elevated flux of glycolytic intermediate into branched pathways, the TCA cycle is still functional. The TCA cycle continues to provide proliferating cancer cells with energy, macromolecules and maintain the cellular redox balance. Recent reports have demonstrated the importance of the TCA cycle and OXPHOS in pancreatic cancer survival[119-123]. Due to their critical roles, the TCA cycle and OXPHOS have been tested as a therapeutic target for PDAC therapy. Three major approaches have been sought to this end: (1) Targeting TCA cycle enzyme/intermediates; (2) Targeting glutamine-dependent anaplerosis; and (3) Targeting the OXPHOS.

Glutamine, a non-essential amino acid, is considered an important energy source for PDAC along with glucose[124,125]. Accumulating evidence demonstrates that glutamine plays a vital role in PDAC proliferation, invasion, maintenance of redox balance, chemotherapy, and radiotherapy resistance, underlining glutamine metabolism as a potential therapeutic target[126-132]. However, conflicting results show that the presence of glutamine suppresses PDAC growth and invasion, dampening

enthusiasm for targeting glutamine metabolism[133-135]. A current clinical trial (NCT04634539) is analyzing whether adding glutamine improves efficacy and reduces the toxicity of PDAC chemotherapy. The results from this trial will shed light on the effect of glutamine on PDAC chemotherapy.

Two additional approaches, targeting the OXPHOS and the TCA cycle, have shown promise in preclinical evaluations, and agents targeting them are currently in clinical trials (Table 2). IACS-010759 inhibits mitochondrial complex one and has recently completed a phase I study in different tumor types, including advanced pancreatic cancers (Table 2). Although the preclinical data regarding the effect of IACS-010759 on pancreatic tumors is lacking, inhibition of OXPHOS complex one appears to be a promising strategy for overcoming drug resistance[136-139]. The anti-diabetic drug metformin has been tested and continues to be tested for its efficacy in PDAC (NCT01210911, NCT02336087, and NCT01666730). Although the experience with metformin in clinical settings has not resulted in improved patient outcomes, a recent meta-analysis indicated survival benefits in patients with PDAC and concurrent diabetes mellitus, highlighting a need for a personalized therapeutic approach for the success of this therapy[140-142].

CPI-613 or Demivistat (Table 2) is a TCA cycle targeting agent that inhibits the activity of pyruvate dehydrogenase and a- ketoglutarate dehydrogenase. In a phase 1 trial, 61% of patients achieved an objective response, and 3 (17%) patients achieved a complete response after receiving CPI-613[143].

Targeting PDAC DNA repair

Activating KRAS mutations are major drivers of malignant growth in PDAC and have remained undruggable until recent promising developments. Oncogenic KRASinduced DNA replication stress drives genomic instability and tumorigenesis in PDAC. Genomic analysis have also revealed that modifications in "DNA damage control" is a prominent genetic alteration observed in PDAC[43]. Recently, genetic alterations in PDAC have been classified into four sub-types by Waddell et al[144]: (1) Stable; (2) Locally rearranged; (3) Scattered; and (4) Unstable. The "unstable" phenotype harbors mutations in the DNA damage repair (DDR), such as BRCA1, BRCA2, PALB2, and ATM. Mutations in ATM account for the most frequently occurring somatic mutations in approximately 4% of PDAC cases, followed by BRCA2, STK11, and BRCA1[144-147]. Given the important role these DDR genes play in a significant proportion of human PDACs, patients are likely to benefit from tailored, targeted therapies, including platinums, directed against specific DDR (Table 3). The following paragraphs will discuss these therapies.

Platinums: Platinum agents (cisplatin, oxaliplatin) cause DNA damage by forming platinum adducts on the DNA and causing DNA interstrand crosslinks[148]. Oxaliplatin is a component of the standard of care FOLFIRINOX, and platinum compounds alone are well suited in cancers that have a deficiency in the homologous repair (HR) pathway. Many studies have highlighted the advantageous use of platinum compounds for HR-deficient PDAC. Golan et al[149] showed a survival benefit (22 mo vs 9 mo) in platinum-treated vs platinum-naïve BRCA1/2 mutated advanced PDAC. Similarly, platinum improved overall survival in patients with HR-deficient PDACs and in patients with germline BRCA1, BRCA2, and PALB2 mutations[150,151]. Hence careful patient selection depending on the genetic make-up of the tumor would be essential for platinums to succeed.

Poly (ADP-ribose) glycohydrolase: Poly (ADP-ribose) glycohydrolase (PARG) is a macrodomain protein with exo- and endo-glycohydrolase activity[152,153]. It critically regulates DNA damage responses by removing poly (ADP-ribose) molecules (PARylation) on modified proteins during the DNA repair process. It is the primary PAR degrading enzyme and reverses poly (ADP ribose) polymerase (PARP) functions by hydrolyzing the ribose-ribose bonds present in PAR molecules. By preventing cytoplasmic PAR accumulation, PARG prevents PAR-mediated apoptosis, termed as parthanatos[154]. Inhibiting PARG causes DNA replication fork collapse, which leads to irreparable DNA damage and cell death. Recent studies have highlighted the benefits of selectively targeting PARG as an anti-cancer therapeutic strategy alone or in combination with other genotoxic therapies[155-157]. Targeting PARG was shown to enhance chemotherapeutic effects of DNA damaging agents, like oxaliplatin and 5fluorouracil in PDAC, and was also synergistic with mitotic kinase, Wee-1 inhibition. In a siRNA screen with DNA replication factors, PARG inhibition was shown to be synergistic with TIMELESS, HUS1, MCM2, CHK1, and RFC2 proteins in an ovarian cancer model, indicating that combinations of PARGi and DNA replication stress



Table 2 Pancreatic ductal adenocarcinoma trials involving agents that target tumor metabolism			
Drug	Target	Trial description	NCI trial number
IACS-010759	OXPHOS inhibitor	Phase I, in advanced cancers	NCT03291938
CPI-613	PDH/alpha KDH inhibitor	Phase I, combination with Gem + nab-paclitaxel	NCT03435289
CPI-613	PDH/alpha KDH inhibitor	Phase II, combination with FOLFIRINOX	NCT03699319
CPI-613	PDH/alpha KDH inhibitor	Phase III, combination with modified FOLFIRINOX	NCT03504423
Metformin and atorvastatin	Metabolic inhibitors	Metformin + Atorvastatin + Doxycycline + Mebendazole in cancers	NCT02201381
L-glutamine	Glutamine analog	Phase I, combination with Gem + nab-paclitaxel	NCT04634539

OXPHOS: Oxidative phosphorylation; PDH: Pyruvate dehydrogenase; KDH: Ketoglutarate dehydrogenase.

Table 3 Pancreatic ductal adenocarcinoma trials involving agents that target DNA repair

Drug	Target	Trial description	NCI trial number
M6620 (VX-970)	ATR	Phase I, M6620 and irinotecan hydrochloride in treating patients with solid tumors that are metastatic or cannot be removed by surgery	NCT02595931
AZD6738/olaparib	ATR/PARP	Phase II, Phase II trial of AZD6738 alone and in combination with olaparib	NCT03682289
BAY1895344	ATR	Phase I, testing the addition of an anti-cancer drug, BAY 1895344 ATR inhibitor, to the chemotherapy treatment (Gemcitabine) for advanced solid tumors, pancreatic cancer, and ovarian cancer	NCT04616534
Olaparib	PARP	Phase II, a study of pembrolizumab and olaparib for people with metastatic pancreatic ductal adenocarcinoma and homologous recombination deficiency or exceptional treatment response to platinum-based therapy	NCT04666740
Olaparib	PARP	Phase I, targeted PARP or MEK/ERK inhibition in patients with pancreatic cancer	NCT04005690
Olaparib	PARP	Phase II, a phase 2 study of cediranib in combination with olaparib in advanced solid tumors	NCT02498613
Olaparib	PARP	Phase II, olaparib in treating patients with stage IV pancreatic cancer	NCT02677038
Talazoparib	PARP	Phase II, measuring the effects of talazoparib in patients with advanced cancer and DNA repair variations	NCT04550494
Talazoparib	PARP	Phase I/II, a study of avelumab, binimetinib and talazoparib in patients with locally advanced or metastatic RAS-mutant solid tumors	NCT03637491
Niraparib	PARP	Phase II, niraparib in metastatic pancreatic cancer after previous chemotherapy (NIRA-PANC): A phase 2 trial	NCT03553004
Niraparib	PARP	Phase II, niraparib in patients with pancreatic cancer	NCT03601923
Rucaparib	PARP	Phase II, maintenance rucaparib in BRCA1, BRCA2 or PALB2 mutated pancreatic cancer that has not progressed on platinum-based therapy	NCT03140670
MK1775	WEE1	Phase I/II, a phase i and randomized phase II study of nab-paclitaxel/gemcitabine with or without AZD1775 for treatment of metastatic adenocarcinoma of the pancreas	NCT02194829

PARP: Poly (ADP ribose) polymerase.

inducers should be evaluated as potential therapeutic strategies for PDAC treatment [158]. A synthetic lethal relationship with PARG inhibition and DDR proteins like BRCA1, BRCA2, ABRAXAS, BARD1, and PALB2 was reported in an MCF7 breast cancer model[159]. Since genomic screens in PDAC have revealed alterations/mutations in similar DDR proteins, it is valuable to target PARG in such DDR-deficient PDAC tumors.

Wee-1: WEE1 kinase is an important cell cycle regulator of the G2-M checkpoint and is overexpressed in various cancers, including glioblastoma, breast cancer, osteosarcoma, and hepatocellular carcinoma[160-163]. It phosphorylates and inactivates CDK1 to allow for the repair of damaged DNA before entering mitosis. Wee-1 has regulatory roles in DNA replication stress and HR mechanisms[164-166]. In PDAC, Wee-1 expression is upregulated by a post-transcriptional mechanism regulated by RNA



binding protein, HuR[167], and its inhibition has been found to be effective in DNA repair-deficient PDAC cells[168]. In one study, Wee-1 inhibition was found to sensitize PDAC cells to gemcitabine chemo-radiation therapy[165]. Another study showed Wee-1 inhibition was synergistic with gemcitabine in p53-deficient PDAC xenografts[169]. Co-targeting WEE1 and ATM was shown to synergistically reduce cell proliferation and migration via downregulation of PDL-1 expression in pancreatic cancers[170]. Recently, it was also published that a combination of Wee-1 with another DNA repair target, PARG, enhances DNA damage and decreases cell survival in PDAC cells[171].

PARP: PARP is a DNA repair enzyme that plays a role in inflammation, regulation of cell death, transcription, and modulation of post-transcriptional gene expression. In response to DNA damage, PARP-1 could either promote cell survival and DNA repair or cause cell death when the damage is high[172]. PARP covalently adds Poly (ADP ribose) (PAR) chains onto its target proteins by consuming beta nicotinamide adenine dinucleotide (β NAD+). PAR further recruits other DNA repair proteins in the process of damage repair. Chemical competitive inhibitors of PARP enzymatic activity have gained interest as treatment options for many cancers, like ovarian, breast, uterine, and prostate[173], specifically for patients with tumors harboring somatic or germline defects/mutations in HR genes like BRCA1/2. Recent whole-genome sequencing studies done in patients with familial pancreatic cancer show that mutations in BRCA2 gene accounts for 5%-10% of familial pancreatic cancers. In the Ashkenazi Jewish population with PDAC, this percentage increases to 13.7% and represents a major subgroup of PDAC cases that could benefit from PARP inhibitor (PARPi) therapy. In the context of synthetic lethality, impairment of two DNA repair pathways induces cell death and thus targeting HR deficient cells (BRCA1/2 mutants or others) with PARP inhibitors was found to be lethal [174,175]. Following the success of POLO trial (Pancreas Cancer Olaparib Ongoing), in 2019 FDA approved olaparib (PARPi) as a maintenance therapy in patients with a germline BRCA mutated metastatic PDAC that had not progressed on first-line platinum therapy[174]. An increasing amount of ongoing preclinical and clinical studies suggest that PARPi in combination with either conventional chemotherapeutics (gemcitabine/nab-paclitaxel) or radiation therapy could benefit patients in the long run[176]. However, recent research suggests that although these respond greatly to PARP inhibitors, there is still 40%-70% of BRCA1/2mutated cancers that fail to respond to PARPi therapy and in those settings PARPi cannot be used. Novel efforts to create a 'BRCAness-tumors harboring mutations in HR beyond BRCA1/2' phenotype in the cells by use of other small molecule inhibitors and their combination with PARPi is now being exploited. Bagnolini et al[174] discovered a small molecule disruptor of RAD51-BRCA2 interaction synergizes with olaparib in pancreatic cancer cells. Another study showed synthetic lethaility with PARPi therapy and FGFR1 blockade in pancreatic cancer[177]. Failure of PARPi therapy can also be attributed to acquired resistance mechanisms[178]. A study in pancreatic cancer showed a secondary mutation in BRCA2 emerged after the patient's exceptional response to platinum and PARPi therapy, which likely restored BRCA2 function in PARP inhibitor-resistant tumor cells[179]. Thus, careful evaluation and design of PARPi therapy should be pursued, and novel targets for PARPi beyond BRCA1/2 should be explored.

Other inhibitors of DDR pathway: Ataxia telangiectasia mutated (ATM) and RAD-3 related (ATR) are serine/threonine protein kinases that are involved in double/singlestrand break repair and modulate DNA replication stress and DDR signaling[180-182]. ATM is one of the most commonly mutated DDR genes, and many whole genomic sequencing studies in PDAC have reported both somatic or germline ATM loss-of-function mutations. ATM loss drives pancreatic cancer progression, angiogenesis, epithelial-to-mesenchymal transition, and stemness[183]. Radiosensitization of cells with ATM loss/inhibition has been well documented in many tumor types, including pancreatic cancers[184-186]. ATM loss can also synergize with platinums and PARP inhibitor therapies, emphasizing its role in DNA repair. Specific to PDAC, two studies have shown that patients with ATM/ATR mutated tumors respond well to oxaliplatin-based chemotherapy, experiencing either improved progression-free survival or a stable disease[187,188]. Based on these data, multiple ongoing clinical trials (Phase I/II) involving ATM-deficient solid tumors have been initiated with DNA damage agents like PARP inhibitor therapies (olaparib, talazoparib, and niraparib), some of which accept pancreatic cancer patients. Chemical inhibition of ATM via small molecule inhibitors (AZD0156, AZD1390) is also being tested in combination with other agents in early stage clinical trials in patients with advanced solid tumors and brain tumors (NCT02588105, NCT03423628). Lack of ATM function may lead to



increased dependence on ATR for DDR, and thus ATR inhibition may be particularly potent in PDACs with somatic mutations in ATM. A recent study employing a multi-DDR interference strategy that included an ATR inhibitor and PARP and DNA-PKC inhibitor was shown to inhibit FOLFIRINOX-induced invasive clones in ATMdeficient PDAC tumors[189]. In 2012, a study tested VX-970, an ATR inhibitor, and found it sensitizes PDAC cells to radiation therapy in vivo and in vitro[190]. Another study found that a combination treatment of AZD6738 (ATR inhibitor) and gemcitabine induces PDAC regression by preventing checkpoint activation by gemcitabine[191]. The ATR inhibitors (VX-970, AZD6738, BAY18953[43]) are currently in the early stages of clinical development, like ATM inhibitors in patients with advanced solid tumors and lymphomas (NCT03188965, NCT03682289, NCT02595931, and NCT03718091), with or without other chemotherapeutic agents. Although these appear to be promising therapies, their clinical activity in PDAC patients is yet to be shown[183].

Immunotherapy

Immunotherapy has achieved promising outcomes in certain cancers, however is yet to be realized in PDAC[192-194]. Tumors with high tumor mutation burden (TMB, approximate mutations per megabase), such as melanomas and NSCLC, have shown to respond better to immunotherapy [195-197]. These TMBs are generally associated with mismatch repair (MMR) deficiency. PDACs intrinsically have low MMR deficiencies, which may explain the lower response to immunotherapy approaches such as immune checkpoint inhibitors (ICI)[198]. The immunosuppressive nature and "T cell exhaustion" further contributes to the poor response of PDAC to immunotherapy.

The PDAC is characterized by the presence of dense stroma in the tumor microenvironment. The stromal components include T cells (cytotoxic and regulatory) and myeloid cells such as tumor-associated macrophages (TAM). Infiltration with macrophages is observed in early PDAC tumor development stages and is associated with poor prognosis in PDAC patients [199-201]. These macrophages secrete immunosuppressive factors such as arginase and TGF β , and thereby regulate T-cell mediated cytotoxicity and surveillance[200]. The myeloid-derived suppressor cells are immature myeloid cells that suppress T cell proliferation and promote ROS-induced T cell apoptosis[202,203]. The term "T cell exhaustion" is used for T cells' differentiation state in chronic antigen exposure. The exhaustion stage is driven by persistent T cell receptor signaling leading to ineffective T cell functioning[204-206]. Recent evidence has shown that the T cells present in the PDAC tumor microenvironment are defective in the production of interferon and tumor necrosis factors following peptide recognition[207,208]. However, the T cells with identical peptide specificity in the spleen retain functionality in tumor-bearing animals[209].

Some approaches that are currently under investigation for improving the immunological response of PDAC include as follow.

Cancer vaccines and immune checkpoint blockade: Monotherapies targeting programmed cell death 1 (PD-1) and programmed cell death ligand 1 (PD-L1) have not shown promising responses in PDAC. However, the therapy showed tumor regression and disease stabilization in other advanced cancers such as NSCLC, melanoma, and renal cancers[193]. Similarly, inhibition of PD-1 or PD-L1 failed to demonstrate a positive response in PDAC animal models[207,210-212]. Similar to ICI inhibitors, vaccine trials using vaccine-GVAX pancreas (granulocyte-macrophage colonystimulating factor-secreting allogeneic pancreatic tumor cells) failed to improve overall survival in PDAC patients compared to single-agent chemotherapy[213]. Since the vaccines were able to recruit T cells, one approach to improve their efficacy would be to promote the activation of T cells, which may be achieved through the combination of vaccines with ICI[214]. Currently, clinical trials are underway for establishing the safety and efficacy of these GVAX with ICIs (NCT03153410, NCT02451982, and NCT02648282).

Targeting tumor associated macrophages: Another way to improve the efficacy of immunotherapies is to inhibit the immunosuppressive signaling that originates from the tumor microenvironment. For this, one strategy being tested is to inhibit myeloid cells. Researchers found that CD11b agonist reduces the total number of myeloid cells and improves survival in PDAC mice. In addition, when CD11b was combined with anti-PD-1, anti-CLTA-4, and gemcitabine, enhanced infiltration of tumor with CD8 T cells was observed^[212]. Similarly, other studies have confirmed that targeting TAMs improves therapeutic and T-cell checkpoint immunotherapy response in PDAC models[215-217]. Blockade of Csf1/Csf1R (macrophage colony-stimulating factor



1/receptor) reduces collagen deposits and enhances CD8 T cell infiltration in the PDAC mice model[218]. Currently, a phase II trial is underway to determine the efficacy of cabralizumab (CSF1R inhibitor) in combination with nivolumab and chemotherapy in PDAC (NCT03336216).

Adoptive T cell therapy

Adoptive T cell therapy involves isolating T cells from tumors and then engineering, expanding, and infusing them back into the patients^[219]. The chimeric antigen receptor (CAR) T cell therapy is an example of adoptive T cell therapy wherein the T cells are manipulated to express CAR to assist tumor recognition [220]. Antigen targets that are being tested for PDAC include mesothelin, prostate stem cell antigen, CEA, MUC1, and HER2[221]. However, the immunosuppressive microenvironment remains a hindrance in CAR-T cell therapy's success in PDAC[222,223]. Other barrier to the success of adoptive T cell therapy in PDAC include antigen selection and toxicities [224-226]. Still, a few promising outcomes have sustained hope for the use of this approach in PDAC. A phase 1 trial found that treatment of PDAC patients with mesothelin-targeting-CART-T cells stabilized disease in 2 out of 6 patients[227]. Similarly, analysis of efficacy and safety of MUC1-targeting CART-T cells found the therapy to be safe and successfully elevated the levels of CD4+ and CD8+ T cells at the tumor [228]. Currently, clinical trials are underway to determine MUC-1-targeted CAR-T cell therapy's efficacy and safety in patients with solid tumors, including PDAC (NCT02587689 and NCT02617134).

CONCLUSION

The PDAC remains an intractable disease that is slated to be the second leading cause of cancer-related deaths by 2030. Although surgical resection remains the only curative treatment option, late diagnosis, in addition to the patient's performance status, limits the scope of surgical intervention. Chemotherapeutic regimens such as gemcitabine+ nab-paclitaxel and FOLFIRINOX has shown promise in improving patient survival; however, drug resistance remains a continuing challenge that has limited their efficacy. Two approaches that may improve PDAC patient outcomes include inhibiting the mechanism(s) that promote therapy resistance and targeting the key pathways essential for PDAC survival. The altered metabolism provides the PDAC cells with energy (ATP) and macromolecules essential for tumor growth. Additionally, studies have shown that metabolism plays a key role in PDAC therapy resistance. Similarly, PARP targeting therapies' success has once again brought the importance of DNA repair mechanisms in PDAC into the center. The limited success of immunotherapy has dampened the enthusiasm for targeting PDAC using this approach. However, the uncovering of mechanisms contributing to poor PDAC's response to immunotherapy has provided opportunities to test newer approaches. Even though the strategies mentioned above have shown promising pre-clinical results individually, a regimen targeting multiple aspects of PDAC will likely deliver a better clinical outcome in this deadly disease.

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REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin 2020; 70: 7-30 [PMID: 1 31912902 DOI: 10.3322/caac.21590]
- 2 Rahib L, Smith BD, Aizenberg R, Rosenzweig AB, Fleshman JM, Matrisian LM. Projecting cancer incidence and deaths to 2030: the unexpected burden of thyroid, liver, and pancreas cancers in the United States. Cancer Res 2014; 74: 2913-2921 [PMID: 24840647 DOI: 10.1158/0008-5472.CAN-14-0155]
- Burris HA 3rd, Moore MJ, Andersen J, Green MR, Rothenberg ML, Modiano MR, Cripps MC, 3 Portenoy RK, Storniolo AM, Tarassoff P, Nelson R, Dorr FA, Stephens CD, Von Hoff DD. Improvements in survival and clinical benefit with gencitabine as first-line therapy for patients with



advanced pancreas cancer: a randomized trial. J Clin Oncol 1997; 15: 2403-2413 [PMID: 9196156 DOI: 10.1200/JCO.1997.15.6.2403]

- 4 Conroy T, Desseigne F, Ychou M, Bouché O, Guimbaud R, Bécouarn Y, Adenis A, Raoul JL, Gourgou-Bourgade S, de la Fouchardière C, Bennouna J, Bachet JB, Khemissa-Akouz F, Péré-Vergé D, Delbaldo C, Assenat E, Chauffert B, Michel P, Montoto-Grillot C, Ducreux M; Groupe Tumeurs Digestives of Unicancer; PRODIGE Intergroup. FOLFIRINOX vs gemcitabine for metastatic pancreatic cancer. N Engl J Med 2011; 364: 1817-1825 [PMID: 21561347 DOI: 10.1056/NEJMoa1011923]
- Labori KJ, Katz MH, Tzeng CW, Bjørnbeth BA, Cvancarova M, Edwin B, Kure EH, Eide TJ, Dueland S, Buanes T, Gladhaug IP. Impact of early disease progression and surgical complications on adjuvant chemotherapy completion rates and survival in patients undergoing the surgery first approach for resectable pancreatic ductal adenocarcinoma - A population-based cohort study. Acta Oncol 2016; 55: 265-277 [PMID: 26213211 DOI: 10.3109/0284186X.2015.1068445]
- Oettle H, Post S, Neuhaus P, Gellert K, Langrehr J, Ridwelski K, Schramm H, Fahlke J, Zuelke C, Burkart C, Gutberlet K, Kettner E, Schmalenberg H, Weigang-Koehler K, Bechstein WO, Niedergethmann M, Schmidt-Wolf I, Roll L, Doerken B, Riess H. Adjuvant chemotherapy with gemcitabine vs observation in patients undergoing curative-intent resection of pancreatic cancer: a randomized controlled trial. JAMA 2007; 297: 267-277 [PMID: 17227978 DOI: 10.1001/jama.297.3.267]
- Neoptolemos JP, Stocken DD, Bassi C, Ghaneh P, Cunningham D, Goldstein D, Padbury R, Moore MJ, Gallinger S, Mariette C, Wente MN, Izbicki JR, Friess H, Lerch MM, Dervenis C, Oláh A, Butturini G, Doi R, Lind PA, Smith D, Valle JW, Palmer DH, Buckels JA, Thompson J, McKay CJ, Rawcliffe CL, Büchler MW; European Study Group for Pancreatic Cancer. Adjuvant chemotherapy with fluorouracil plus folinic acid vs gemcitabine following pancreatic cancer resection: a randomized controlled trial. JAMA 2010; 304: 1073-1081 [PMID: 20823433 DOI: 10.1001/jama.2010.1275
- Neoptolemos JP, Palmer DH, Ghaneh P, Psarelli EE, Valle JW, Halloran CM, Faluyi O, O'Reilly DA, Cunningham D, Wadsley J, Darby S, Meyer T, Gillmore R, Anthoney A, Lind P, Glimelius B, Falk S, Izbicki JR, Middleton GW, Cummins S, Ross PJ, Wasan H, McDonald A, Crosby T, Ma YT, Patel K, Sherriff D, Soomal R, Borg D, Sothi S, Hammel P, Hackert T, Jackson R, Büchler MW; European Study Group for Pancreatic Cancer. Comparison of adjuvant gemcitabine and capecitabine with gemcitabine monotherapy in patients with resected pancreatic cancer (ESPAC-4): a multicentre, open-label, randomised, phase 3 trial. Lancet 2017; 389: 1011-1024 [PMID: 28129987 DOI: 10.1016/S0140-6736(16)32409-6]
- Dreyer SB, Chang DK, Bailey P, Biankin AV. Pancreatic Cancer Genomes: Implications for Clinical Management and Therapeutic Development. Clin Cancer Res 2017; 23: 1638-1646 [PMID: 28373362 DOI: 10.1158/1078-0432.CCR-16-2411]
- 10 Mini E, Nobili S, Caciagli B, Landini I, Mazzei T. Cellular pharmacology of gemcitabine. Ann Oncol 2006; 17 Suppl 5: v7-12 [PMID: 16807468 DOI: 10.1093/annonc/mdj941]
- 11 Heinemann V, Hertel LW, Grindey GB, Plunkett W. Comparison of the cellular pharmacokinetics and toxicity of 2',2'-difluorodeoxycytidine and 1-beta-D-arabinofuranosylcytosine. Cancer Res 1988; 48: 4024-4031 [PMID: 3383195]
- 12 Mackey JR, Mani RS, Selner M, Mowles D, Young JD, Belt JA, Crawford CR, Cass CE. Functional nucleoside transporters are required for gemcitabine influx and manifestation of toxicity in cancer cell lines. Cancer Res 1998; 58: 4349-4357 [PMID: 9766663]
- 13 Spratlin J, Sangha R, Glubrecht D, Dabbagh L, Young JD, Dumontet C, Cass C, Lai R, Mackey JR. The absence of human equilibrative nucleoside transporter 1 is associated with reduced survival in patients with gemcitabine-treated pancreas adenocarcinoma. Clin Cancer Res 2004; 10: 6956-6961 [PMID: 15501974 DOI: 10.1158/1078-0432.CCR-04-0224]
- 14 Ohhashi S, Ohuchida K, Mizumoto K, Fujita H, Egami T, Yu J, Toma H, Sadatomi S, Nagai E, Tanaka M. Down-regulation of deoxycytidine kinase enhances acquired resistance to gemcitabine in pancreatic cancer. Anticancer Res 2008; 28: 2205-2212 [PMID: 18751396]
- 15 Saiki Y, Yoshino Y, Fujimura H, Manabe T, Kudo Y, Shimada M, Mano N, Nakano T, Lee Y, Shimizu S, Oba S, Fujiwara S, Shimizu H, Chen N, Nezhad ZK, Jin G, Fukushige S, Sunamura M, Ishida M, Motoi F, Egawa S, Unno M, Horii A. DCK is frequently inactivated in acquired gemcitabine-resistant human cancer cells. Biochem Biophys Res Commun 2012; 421: 98-104 [PMID: 22490663 DOI: 10.1016/j.bbrc.2012.03.122]
- Miller AL, Garcia PL, Gamblin TL, Vance RB, Yoon KJ. Development of gemcitabine-resistant 16 patient-derived xenograft models of pancreatic ductal adenocarcinoma. Cancer Drug Resist 2020; 3: 572-585 [PMID: 33073205 DOI: 10.20517/cdr.2020.35]
- 17 Wang C, Zhang W, Fu M, Yang A, Huang H, Xie J. Establishment of human pancreatic cancer gemcitabineresistant cell line with ribonucleotide reductase overexpression. Oncol Rep 2015; 33: 383-390 [PMID: 25394408 DOI: 10.3892/or.2014.3599]
- 18 Nakahira S, Nakamori S, Tsujie M, Takahashi Y, Okami J, Yoshioka S, Yamasaki M, Marubashi S, Takemasa I, Miyamoto A, Takeda Y, Nagano H, Dono K, Umeshita K, Sakon M, Monden M. Involvement of ribonucleotide reductase M1 subunit overexpression in gemcitabine resistance of human pancreatic cancer. Int J Cancer 2007; 120: 1355-1363 [PMID: 17131328 DOI: 10.1002/ijc.22390]
- Maftouh M, Avan A, Funel N, Frampton AE, Fiuji H, Pelliccioni S, Castellano L, Galla V, Peters 19



GJ, Giovannetti E. miR-211 modulates gemcitabine activity through downregulation of ribonucleotide reductase and inhibits the invasive behavior of pancreatic cancer cells. Nucleosides Nucleotides Nucleic Acids 2014; 33: 384-393 [PMID: 24940696 DOI: 10.1080/15257770.2014.891741]

- Lu H, Lu S, Yang D, Zhang L, Ye J, Li M, Hu W. MiR-20a-5p regulates gemcitabine 20 chemosensitivity by targeting RRM2 in pancreatic cancer cells and serves as a predictor for gemcitabine-based chemotherapy. Biosci Rep 2019; 39 [PMID: 30777929 DOI: 10.1042/BSR20181374]
- 21 Xia G, Wang H, Song Z, Meng Q, Huang X. Gambogic acid sensitizes gemcitabine efficacy in pancreatic cancer by reducing the expression of ribonucleotide reductase subunit-M2 (RRM2). J Exp Clin Cancer Res 2017; 36: 107 [PMID: 28797284 DOI: 10.1186/s13046-017-0579-0]
- 22 Vena F, Li Causi E, Rodriguez-Justo M, Goodstal S, Hagemann T, Hartley JA, Hochhauser D. The MEK1/2 Inhibitor Pimasertib Enhances Gemcitabine Efficacy in Pancreatic Cancer Models by Altering Ribonucleotide Reductase Subunit-1 (RRM1). Clin Cancer Res 2015; 21: 5563-5577 [PMID: 26228206 DOI: 10.1158/1078-0432.CCR-15-0485]
- 23 Roman NO, Samulitis BK, Wisner L, Landowski TH, Dorr RT. Imexon enhances gemcitabine cytotoxicity by inhibition of ribonucleotide reductase. Cancer Chemother Pharmacol 2011; 67: 183-192 [PMID: 20339847 DOI: 10.1007/s00280-010-1306-0]
- Mitsuno M, Kitajima Y, Ohtaka K, Kai K, Hashiguchi K, Nakamura J, Hiraki M, Noshiro H, Miyazaki K. Tranilast strongly sensitizes pancreatic cancer cells to gemcitabine via decreasing protein expression of ribonucleotide reductase 1. Int J Oncol 2010; 36: 341-349 [PMID: 20043067 DOI: 10.3892/ijo 000005051
- 25 Hwang DW, Shin E, Cho JY, Han HS, Yoon YS. Human equilibrative nucleoside transporter-1 (hENT1) and ribonucleotide reductase regulatory subunit M1 (RRM1) expression; do they have survival impact to pancreatic cancer? Ann Hepatobiliary Pancreat Surg 2020; 24: 127-136 [PMID: 32457256 DOI: 10.14701/ahbps.2020.24.2.127]
- 26 Han QL, Zhou YH, Lyu Y, Yan H, Dai GH. Effect of ribonucleotide reductase M1 expression on overall survival in patients with pancreatic cancer receiving gemcitabine chemotherapy: A literaturebased meta-analysis. J Clin Pharm Ther 2018; 43: 163-169 [PMID: 29214667 DOI: 10.1111/jcpt.12655]
- Aoyama T, Miyagi Y, Murakawa M, Yamaoku K, Atsumi Y, Shiozawa M, Ueno M, Morimoto M, 27 Oshima T, Yukawa N, Yoshikawa T, Rino Y, Masuda M, Morinaga S. Clinical implications of ribonucleotide reductase subunit M1 in patients with pancreatic cancer who undergo curative resection followed by adjuvant chemotherapy with gemcitabine. Oncol Lett 2017; 13: 3423-3430 [PMID: 28521448 DOI: 10.3892/ol.2017.5935]
- 28 Elander NO, Aughton K, Ghaneh P, Neoptolemos JP, Palmer DH, Cox TF, Campbell F, Costello E, Halloran CM, Mackey JR, Scarfe AG, Valle JW, McDonald AC, Carter R, Tebbutt NC, Goldstein D, Shannon J, Dervenis C, Glimelius B, Deakin M, Charnley RM, Anthoney A, Lerch MM, Mayerle J, Oláh A, Büchler MW, Greenhalf W; European Study Group for Pancreatic Cancer. Intratumoural expression of deoxycytidylate deaminase or ribonuceotide reductase subunit M1 expression are not related to survival in patients with resected pancreatic cancer given adjuvant chemotherapy. Br J Cancer 2018; 118: 1084-1088 [PMID: 29523831 DOI: 10.1038/s41416-018-0005-1]
- 29 Zeng S, Pöttler M, Lan B, Grützmann R, Pilarsky C, Yang H. Chemoresistance in Pancreatic Cancer. Int J Mol Sci 2019; 20 [PMID: 31514451 DOI: 10.3390/ijms20184504]
- 30 Arumugam T, Ramachandran V, Fournier KF, Wang H, Marquis L, Abbruzzese JL, Gallick GE, Logsdon CD, McConkey DJ, Choi W. Epithelial to mesenchymal transition contributes to drug resistance in pancreatic cancer. Cancer Res 2009; 69: 5820-5828 [PMID: 19584296 DOI: 10.1158/0008-5472.CAN-08-2819]
- 31 Zheng X, Carstens JL, Kim J, Scheible M, Kaye J, Sugimoto H, Wu CC, LeBleu VS, Kalluri R. Epithelial-to-mesenchymal transition is dispensable for metastasis but induces chemoresistance in pancreatic cancer. Nature 2015; 527: 525-530 [PMID: 26560028 DOI: 10.1038/nature16064]
- Kalser MH, Ellenberg SS. Pancreatic cancer. Adjuvant combined radiation and chemotherapy 32 following curative resection. Arch Surg 1985; 120: 899-903 [PMID: 4015380 DOI: 10.1001/archsurg.1985.01390320023003]
- Moertel CG, Frytak S, Hahn RG, O'Connell MJ, Reitemeier RJ, Rubin J, Schutt AJ, Weiland LH, 33 Childs DS, Holbrook MA, Lavin PT, Livstone E, Spiro H, Knowlton A, Kalser M, Barkin J, Lessner H, Mann-Kaplan R, Ramming K, Douglas HO Jr, Thomas P, Nave H, Bateman J, Lokich J, Brooks J, Chaffey J, Corson JM, Zamcheck N, Novak JW. Therapy of locally unresectable pancreatic carcinoma: a randomized comparison of high dose (6000 rads) radiation alone, moderate dose radiation (4000 rads + 5-fluorouracil), and high dose radiation + 5-fluorouracil: The Gastrointestinal Tumor Study Group. *Cancer* 1981; **48**: 1705-1710 [PMID: 7284971 DOI: 10.1002/1097-0142(19811015)48:8<1705::aid-cncr2820480803>3.0.co;2-4]
- 34 Chevalier H, Vienot A, Lièvre A, Edeline J, El Hajbi F, Peugniez C, Vernerey D, Meurisse A, Hammel P, Neuzillet C, Borg C, Turpin A. FOLFIRINOX De-Escalation in Advanced Pancreatic Cancer: A Multicenter Real-Life Study. Oncologist 2020; 25: e1701-e1710 [PMID: 32886823 DOI: 10.1634/theoncologist.2020-0577]
- Foschini F, Napolitano F, Servetto A, Marciano R, Mozzillo E, Carratù AC, Santaniello A, De 35 Placido P, Cascetta P, Butturini G, Frigerio I, Regi P, Silvestris N, Delcuratolo S, Vasile E, Vivaldi C, Bianco C, De Placido S, Formisano L, Bianco R. FOLFIRINOX after first-line gemcitabine-based



chemotherapy in advanced pancreatic cancer: a retrospective comparison with FOLFOX and FOLFIRI schedules. Ther Adv Med Oncol 2020; 12: 1758835920947970 [PMID: 33062062 DOI: 10.1177/1758835920947970

- 36 Jung JH, Shin DW, Kim J, Lee JC, Hwang JH. Primary Granulocyte Colony-Stimulating Factor Prophylaxis in Metastatic Pancreatic Cancer Patients Treated with FOLFIRINOX as the First-Line Treatment. Cancers (Basel) 2020; 12 [PMID: 33120908 DOI: 10.3390/cancers12113137]
- 37 Kang SP, Saif MW. Pharmacogenomics and pancreatic cancer treatment. Optimizing current therapy and individualizing future therapy. JOP 2008; 9: 251-266 [PMID: 18469437]
- 38 Zhang N, Yin Y, Xu SJ, Chen WS. 5-Fluorouracil: mechanisms of resistance and reversal strategies. Molecules 2008; 13: 1551-1569 [PMID: 18794772 DOI: 10.3390/molecules13081551]
- 39 Hu YC, Komorowski RA, Graewin S, Hostetter G, Kallioniemi OP, Pitt HA, Ahrendt SA. Thymidylate synthase expression predicts the response to 5-fluorouracil-based adjuvant therapy in pancreatic cancer. Clin Cancer Res 2003; 9: 4165-4171 [PMID: 14519641]
- 40 Jenh CH, Geyer PK, Baskin F, Johnson LF. Thymidylate synthase gene amplification in fluorodeoxyuridine-resistant mouse cell lines. Mol Pharmacol 1985; 28: 80-85 [PMID: 2991733]
- 41 Ciaparrone M, Quirino M, Schinzari G, Zannoni G, Corsi DC, Vecchio FM, Cassano A, La Torre G, Barone C. Predictive role of thymidylate synthase, dihydropyrimidine dehydrogenase and thymidine phosphorylase expression in colorectal cancer patients receiving adjuvant 5-fluorouracil. Oncology 2006; 70: 366-377 [PMID: 17179731 DOI: 10.1159/000098110]
- 42 Kurata N, Fujita H, Ohuchida K, Mizumoto K, Mahawithitwong P, Sakai H, Onimaru M, Manabe T, Ohtsuka T, Tanaka M. Predicting the chemosensitivity of pancreatic cancer cells by quantifying the expression levels of genes associated with the metabolism of genetiabine and 5-fluorouracil. Int J Oncol 2011; 39: 473-482 [PMID: 21617862 DOI: 10.3892/ijo.2011.1058]
- 43 Jones S, Zhang X, Parsons DW, Lin JC, Leary RJ, Angenendt P, Mankoo P, Carter H, Kamiyama H, Jimeno A, Hong SM, Fu B, Lin MT, Calhoun ES, Kamiyama M, Walter K, Nikolskaya T, Nikolsky Y, Hartigan J, Smith DR, Hidalgo M, Leach SD, Klein AP, Jaffee EM, Goggins M, Maitra A, Iacobuzio-Donahue C, Eshleman JR, Kern SE, Hruban RH, Karchin R, Papadopoulos N, Parmigiani G, Vogelstein B, Velculescu VE, Kinzler KW. Core signaling pathways in human pancreatic cancers revealed by global genomic analyses. Science 2008; 321: 1801-1806 [PMID: 18772397 DOI: 10.1126/science.1164368]
- 44 Smit VT, Boot AJ, Smits AM, Fleuren GJ, Cornelisse CJ, Bos JL. KRAS codon 12 mutations occur very frequently in pancreatic adenocarcinomas. Nucleic Acids Res 1988; 16: 7773-7782 [PMID: 3047672 DOI: 10.1093/nar/16.16.7773]
- 45 Lemoine NR, Hughes CM, Barton CM, Poulsom R, Jeffery RE, Klöppel G, Hall PA, Gullick WJ. The epidermal growth factor receptor in human pancreatic cancer. J Pathol 1992; 166: 7-12 [PMID: 1538276 DOI: 10.1002/path.1711660103]
- 46 Boeck S, Jung A, Laubender RP, Neumann J, Egg R, Goritschan C, Vehling-Kaiser U, Winkelmann C, Fischer von Weikersthal L, Clemens MR, Gauler TC, Märten A, Klein S, Kojouharoff G, Barner M, Geissler M, Greten TF, Mansmann U, Kirchner T, Heinemann V. EGFR pathway biomarkers in erlotinib-treated patients with advanced pancreatic cancer: translational results from the randomised, crossover phase 3 trial AIO-PK0104. Br J Cancer 2013; 108: 469-476 [PMID: 23169292 DOI: 10.1038/bjc.2012.495
- 47 Moore MJ, Goldstein D, Hamm J, Figer A, Hecht JR, Gallinger S, Au HJ, Murawa P, Walde D, Wolff RA, Campos D, Lim R, Ding K, Clark G, Voskoglou-Nomikos T, Ptasynski M, Parulekar W; National Cancer Institute of Canada Clinical Trials Group. Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. J Clin Oncol 2007; 25: 1960-1966 [PMID: 17452677 DOI: 10.1200/JCO.2006.07.9525]
- 48 Haas M, Siveke JT, Schenk M, Lerch MM, Caca K, Freiberg-Richter J, Fischer von Weikersthal L, Kullmann F, Reinacher-Schick A, Fuchs M, Kanzler S, Kunzmann V, Ettrich TJ, Kruger S, Westphalen CB, Held S, Heinemann V, Boeck S. Efficacy of gemcitabine plus erlotinib in rashpositive patients with metastatic pancreatic cancer selected according to eligibility for FOLFIRINOX: A prospective phase II study of the 'Arbeitsgemeinschaft Internistische Onkologie'. Eur J Cancer 2018; 94: 95-103 [PMID: 29549862 DOI: 10.1016/j.ejca.2018.02.008]
- 49 Van Cutsem E, Li CP, Nowara E, Aprile G, Moore M, Federowicz I, Van Laethem JL, Hsu C, Tham CK, Stemmer SM, Lipp R, Zeaiter A, Fittipaldo A, Csutor Z, Klughammer B, Meng X, Ciuleanu T. Dose escalation to rash for erlotinib plus gemcitabine for metastatic pancreatic cancer: the phase II RACHEL study. Br J Cancer 2014; 111: 2067-2075 [PMID: 25247318 DOI: 10.1038/bjc.2014.494]
- Philip PA, Benedetti J, Corless CL, Wong R, O'Reilly EM, Flynn PJ, Rowland KM, Atkins JN, 50 Mirtsching BC, Rivkin SE, Khorana AA, Goldman B, Fenoglio-Preiser CM, Abbruzzese JL, Blanke CD. Phase III study comparing gencitabine plus cetuximab vs gencitabine in patients with advanced pancreatic adenocarcinoma: Southwest Oncology Group-directed intergroup trial S0205. J Clin Oncol 2010; 28: 3605-3610 [PMID: 20606093 DOI: 10.1200/JCO.2009.25.7550]
- 51 Katopodis O, Souglakos J, Stathopoulos E, Christopoulou A, Kontopodis E, Kotsakis A, Kalbakis K, Kentepozidis N, Polyzos A, Hatzidaki D, Georgoulias V. Frontline treatment with gemcitabine, oxaliplatin and erlotinib for the treatment of advanced or metastatic pancreatic cancer: a multicenter phase II study of the Hellenic Oncology Research Group (HORG). Cancer Chemother Pharmacol 2014; 74: 333-340 [PMID: 24930058 DOI: 10.1007/s00280-014-2509-6]



- 52 Heinemann V, Vehling-Kaiser U, Waldschmidt D, Kettner E, Märten A, Winkelmann C, Klein S, Kojouharoff G, Gauler TC, von Weikersthal LF, Clemens MR, Geissler M, Greten TF, Hegewisch-Becker S, Rubanov O, Baake G, Höhler T, Ko YD, Jung A, Neugebauer S, Boeck S. Gemcitabine plus erlotinib followed by capecitabine vs capecitabine plus erlotinib followed by gemcitabine in advanced pancreatic cancer: final results of a randomised phase 3 trial of the 'Arbeitsgemeinschaft Internistische Onkologie' (AIO-PK0104). Gut 2013; 62: 751-759 [PMID: 22773551 DOI: 10.1136/gutjnl-2012-302759]
- Rich TA, Winter K, Safran H, Hoffman JP, Erickson B, Anne PR, Myerson RJ, Cline-Burkhardt VJ, 53 Perez K, Willett C. Weekly paclitaxel, gemcitabine, and external irradiation followed by randomized farnesyl transferase inhibitor R115777 for locally advanced pancreatic cancer. Onco Targets Ther 2012; 5: 161-170 [PMID: 22977306 DOI: 10.2147/OTT.S33560]
- Kindler HL, Niedzwiecki D, Hollis D, Sutherland S, Schrag D, Hurwitz H, Innocenti F, Mulcahy 54 MF, O'Reilly E, Wozniak TF, Picus J, Bhargava P, Mayer RJ, Schilsky RL, Goldberg RM. Gemcitabine plus bevacizumab compared with gemcitabine plus placebo in patients with advanced pancreatic cancer: phase III trial of the Cancer and Leukemia Group B (CALGB 80303). J Clin Oncol 2010; 28: 3617-3622 [PMID: 20606091 DOI: 10.1200/JCO.2010.28.1386]
- 55 Kindler HL, Ioka T, Richel DJ, Bennouna J, Létourneau R, Okusaka T, Funakoshi A, Furuse J, Park YS, Ohkawa S, Springett GM, Wasan HS, Trask PC, Bycott P, Ricart AD, Kim S, Van Cutsem E. Axitinib plus gemcitabine vs placebo plus gemcitabine in patients with advanced pancreatic adenocarcinoma: a double-blind randomised phase 3 study. Lancet Oncol 2011; 12: 256-262 [PMID: 21306953 DOI: 10.1016/S1470-2045(11)70004-3]
- 56 Infante JR, Somer BG, Park JO, Li CP, Scheulen ME, Kasubhai SM, Oh DY, Liu Y, Redhu S, Steplewski K, Le N. A randomised, double-blind, placebo-controlled trial of trametinib, an oral MEK inhibitor, in combination with gemcitabine for patients with untreated metastatic adenocarcinoma of the pancreas. Eur J Cancer 2014; 50: 2072-2081 [PMID: 24915778 DOI: 10.1016/j.ejca.2014.04.024]
- 57 Chung V, McDonough S, Philip PA, Cardin D, Wang-Gillam A, Hui L, Tejani MA, Seery TE, Dy IA, Al Baghdadi T, Hendifar AE, Doyle LA, Lowy AM, Guthrie KA, Blanke CD, Hochster HS. Effect of Selumetinib and MK-2206 vs Oxaliplatin and Fluorouracil in Patients With Metastatic Pancreatic Cancer After Prior Therapy: SWOG S1115 Study Randomized Clinical Trial. JAMA Oncol 2017; 3: 516-522 [PMID: 27978579 DOI: 10.1001/jamaoncol.2016.5383]
- 58 Mahadevan D, Von Hoff DD. Tumor-stroma interactions in pancreatic ductal adenocarcinoma. Mol Cancer Ther 2007; 6: 1186-1197 [PMID: 17406031 DOI: 10.1158/1535-7163.MCT-06-0686]
- 59 Hoffmann AC, Mori R, Vallbohmer D, Brabender J, Klein E, Drebber U, Baldus SE, Cooc J, Azuma M, Metzger R, Hoelscher AH, Danenberg KD, Prenzel KL, Danenberg PV. High expression of HIF1a is a predictor of clinical outcome in patients with pancreatic ductal adenocarcinomas and correlated to PDGFA, VEGF, and bFGF. Neoplasia 2008; 10: 674-679 [PMID: 18592007 DOI: 10.1593/neo.08292]
- Couvelard A, O'Toole D, Leek R, Turley H, Sauvanet A, Degott C, Ruszniewski P, Belghiti J, 60 Harris AL, Gatter K, Pezzella F. Expression of hypoxia-inducible factors is correlated with the presence of a fibrotic focus and angiogenesis in pancreatic ductal adenocarcinomas. Histopathology 2005; 46: 668-676 [PMID: 15910598 DOI: 10.1111/j.1365-2559.2005.02160.x]
- 61 Büchler P, Reber HA, Lavey RS, Tomlinson J, Büchler MW, Friess H, Hines OJ. Tumor hypoxia correlates with metastatic tumor growth of pancreatic cancer in an orthotopic murine model. J Surg Res 2004; 120: 295-303 [PMID: 15234226 DOI: 10.1016/j.jss.2004.02.014]
- 62 Warburg O. On the origin of cancer cells. Science 1956; 123: 309-314 [PMID: 13298683 DOI: 10.1126/science.123.3191.309]
- 63 Cairns RA, Harris IS, Mak TW. Regulation of cancer cell metabolism. Nat Rev Cancer 2011; 11: 85-95 [PMID: 21258394 DOI: 10.1038/nrc2981]
- Christofk HR, Vander Heiden MG, Harris MH, Ramanathan A, Gerszten RE, Wei R, Fleming MD, 64 Schreiber SL, Cantley LC. The M2 splice isoform of pyruvate kinase is important for cancer metabolism and tumour growth. Nature 2008; 452: 230-233 [PMID: 18337823 DOI: 10.1038/nature06734]
- 65 Dong X, Li Y, Chang P, Tang H, Hess KR, Abbruzzese JL, Li D. Glucose metabolism gene variants modulate the risk of pancreatic cancer. Cancer Prev Res (Phila) 2011; 4: 758-766 [PMID: 21411499 DOI: 10.1158/1940-6207.CAPR-10-0247]
- Dong X, Tang H, Hess KR, Abbruzzese JL, Li D. Glucose metabolism gene polymorphisms and 66 clinical outcome in pancreatic cancer. Cancer 2011; 117: 480-491 [PMID: 20845477 DOI: 10.1002/cncr.25612]
- Markovets AA, Herman D. Analysis of cancer metabolism with high-throughput technologies. 67 BMC Bioinformatics 2011; 12 Suppl 10: S8 [PMID: 22166000 DOI: 10.1186/1471-2105-12-S10-S8]
- Wei Z, Cui L, Mei Z, Liu M, Zhang D. miR-181a mediates metabolic shift in colon cancer cells via 68 the PTEN/AKT pathway. FEBS Lett 2014; 588: 1773-1779 [PMID: 24685694 DOI: 10.1016/j.febslet.2014.03.037]
- Mikuriya K, Kuramitsu Y, Ryozawa S, Fujimoto M, Mori S, Oka M, Hamano K, Okita K, Sakaida 69 I, Nakamura K. Expression of glycolytic enzymes is increased in pancreatic cancerous tissues as evidenced by proteomic profiling by two-dimensional electrophoresis and liquid chromatographymass spectrometry/mass spectrometry. Int J Oncol 2007; 30: 849-855 [PMID: 17332923 DOI: 10.3892/ijo.30.4.849]



- 70 Zhou W, Capello M, Fredolini C, Piemonti L, Liotta LA, Novelli F, Petricoin EF. Proteomic analysis of pancreatic ductal adenocarcinoma cells reveals metabolic alterations. J Proteome Res 2011; **10**: 1944-1952 [PMID: 21309613 DOI: 10.1021/pr101179t]
- 71 Vernucci E, Abrego J, Gunda V, Shukla SK, Dasgupta A, Rai V, Chaika N, Buettner K, Illies A, Yu F, Lazenby AJ, Swanson BJ, Singh PK. Metabolic Alterations in Pancreatic Cancer Progression. Cancers (Basel) 2019; 12 [PMID: 31861288 DOI: 10.3390/cancers12010002]
- 72 Martinez-Outschoorn UE, Peiris-Pagés M, Pestell RG, Sotgia F, Lisanti MP. Cancer metabolism: a therapeutic perspective. Nat Rev Clin Oncol 2017; 14: 11-31 [PMID: 27141887 DOI: 10.1038/nrclinonc.2016.60]
- 73 Wu DH, Liang H, Lu SN, Wang H, Su ZL, Zhang L, Ma JQ, Guo M, Tai S, Yu S. miR-124 Suppresses Pancreatic Ductal Adenocarcinoma Growth by Regulating Monocarboxylate Transporter 1-Mediated Cancer Lactate Metabolism. Cell Physiol Biochem 2018; 50: 924-935 [PMID: 30355947 DOI: 10.1159/000494477]
- 74 Kong SC, Nøhr-Nielsen A, Zeeberg K, Reshkin SJ, Hoffmann EK, Novak I, Pedersen SF. Monocarboxylate Transporters MCT1 and MCT4 Regulate Migration and Invasion of Pancreatic Ductal Adenocarcinoma Cells. Pancreas 2016; 45: 1036-1047 [PMID: 26765963 DOI: 10.1097/MPA.00000000000571
- 75 Feldmann G, Beaty R, Hruban RH, Maitra A. Molecular genetics of pancreatic intraepithelial neoplasia. J Hepatobiliary Pancreat Surg 2007; 14: 224-232 [PMID: 17520196 DOI: 10.1007/s00534-006-1166-5
- 76 Shen L, Sun X, Fu Z, Yang G, Li J, Yao L. The fundamental role of the p53 pathway in tumor metabolism and its implication in tumor therapy. Clin Cancer Res 2012; 18: 1561-1567 [PMID: 22307140 DOI: 10.1158/1078-0432.CCR-11-3040]
- 77 Schwartzenberg-Bar-Yoseph F, Armoni M, Karnieli E. The tumor suppressor p53 down-regulates glucose transporters GLUT1 and GLUT4 gene expression. Cancer Res 2004; 64: 2627-2633 [PMID: 15059920 DOI: 10.1158/0008-5472.can-03-0846]
- 78 Ying H, Kimmelman AC, Lyssiotis CA, Hua S, Chu GC, Fletcher-Sananikone E, Locasale JW, Son J, Zhang H, Coloff JL, Yan H, Wang W, Chen S, Viale A, Zheng H, Paik JH, Lim C, Guimaraes AR, Martin ES, Chang J, Hezel AF, Perry SR, Hu J, Gan B, Xiao Y, Asara JM, Weissleder R, Wang YA, Chin L, Cantley LC, DePinho RA. Oncogenic Kras maintains pancreatic tumors through regulation of anabolic glucose metabolism. Cell 2012; 149: 656-670 [PMID: 22541435 DOI: 10.1016/j.cell.2012.01.058]
- 79 Zhao H, Duan Q, Zhang Z, Li H, Wu H, Shen Q, Wang C, Yin T. Up-regulation of glycolysis promotes the stemness and EMT phenotypes in gemcitabine-resistant pancreatic cancer cells. J Cell Mol Med 2017; 21: 2055-2067 [PMID: 28244691 DOI: 10.1111/jcmm.13126]
- 80 Anderson M, Marayati R, Moffitt R, Yeh JJ. Hexokinase 2 promotes tumor growth and metastasis by regulating lactate production in pancreatic cancer. Oncotarget 2017; 8: 56081-56094 [PMID: 28915575 DOI: 10.18632/oncotarget.9760]
- Bhardwaj V, Rizvi N, Lai MB, Lai JC, Bhushan A. Glycolytic enzyme inhibitors affect pancreatic cancer survival by modulating its signaling and energetics. Anticancer Res 2010; 30: 743-749 [PMID: 20392992]
- 82 Fan K, Fan Z, Cheng H, Huang Q, Yang C, Jin K, Luo G, Yu X, Liu C. Hexokinase 2 dimerization and interaction with voltage-dependent anion channel promoted resistance to cell apoptosis induced by gemcitabine in pancreatic cancer. Cancer Med 2019; 8: 5903-5915 [PMID: 31426130 DOI: 10.1002/cam4.2463
- 83 Dayton TL, Jacks T, Vander Heiden MG. PKM2, cancer metabolism, and the road ahead. EMBO Rep 2016; 17: 1721-1730 [PMID: 27856534 DOI: 10.15252/embr.201643300]
- 84 Anastasiou D, Poulogiannis G, Asara JM, Boxer MB, Jiang JK, Shen M, Bellinger G, Sasaki AT, Locasale JW, Auld DS, Thomas CJ, Vander Heiden MG, Cantley LC. Inhibition of pyruvate kinase M2 by reactive oxygen species contributes to cellular antioxidant responses. Science 2011; 334: 1278-1283 [PMID: 22052977 DOI: 10.1126/science.1211485]
- 85 Hitosugi T, Kang S, Vander Heiden MG, Chung TW, Elf S, Lythgoe K, Dong S, Lonial S, Wang X, Chen GZ, Xie J, Gu TL, Polakiewicz RD, Roesel JL, Boggon TJ, Khuri FR, Gilliland DG, Cantley LC, Kaufman J, Chen J. Tyrosine phosphorylation inhibits PKM2 to promote the Warburg effect and tumor growth. Sci Signal 2009; 2: ra73 [PMID: 19920251 DOI: 10.1126/scisignal.2000431]
- 86 Hillis AL, Lau AN, Devoe CX, Dayton TL, Danai LV, Di Vizio D, Vander Heiden MG. PKM2 is not required for pancreatic ductal adenocarcinoma. Cancer Metab 2018; 6: 17 [PMID: 30386596 DOI: 10.1186/s40170-018-0188-1]
- Lockney NA, Zhang M, Lu Y, Sopha SC, Washington MK, Merchant N, Zhao Z, Shyr Y, 87 Chakravarthy AB, Xia F. Pyruvate Kinase Muscle Isoenzyme 2 (PKM2) Expression Is Associated with Overall Survival in Pancreatic Ductal Adenocarcinoma. J Gastrointest Cancer 2015; 46: 390-398 [PMID: 26385349 DOI: 10.1007/s12029-015-9764-6]
- 88 Mohammad GH, Olde Damink SW, Malago M, Dhar DK, Pereira SP. Pyruvate Kinase M2 and Lactate Dehydrogenase A Are Overexpressed in Pancreatic Cancer and Correlate with Poor Outcome. PLoS One 2016; 11: e0151635 [PMID: 26989901 DOI: 10.1371/journal.pone.0151635]
- Ogawa H, Nagano H, Konno M, Eguchi H, Koseki J, Kawamoto K, Nishida N, Colvin H, Tomokuni A, Tomimaru Y, Hama N, Wada H, Marubashi S, Kobayashi S, Mori M, Doki Y, Ishii H. The combination of the expression of hexokinase 2 and pyruvate kinase M2 is a prognostic marker in patients with pancreatic cancer. Mol Clin Oncol 2015; 3: 563-571 [PMID: 26137268 DOI:



10.3892/mco.2015.490]

- 90 Azoitei N, Becher A, Steinestel K, Rouhi A, Diepold K, Genze F, Simmet T, Seufferlein T. PKM2 promotes tumor angiogenesis by regulating HIF-1a through NF-kB activation. Mol Cancer 2016; 15: 3 [PMID: 26739387 DOI: 10.1186/s12943-015-0490-2]
- 91 Calabretta S, Bielli P, Passacantilli I, Pilozzi E, Fendrich V, Capurso G, Fave GD, Sette C. Modulation of PKM alternative splicing by PTBP1 promotes gemcitabine resistance in pancreatic cancer cells. Oncogene 2016; 35: 2031-2039 [PMID: 26234680 DOI: 10.1038/onc.2015.270]
- Chen S, Chen X, Shan T, Ma J, Lin W, Li W, Kang Y. MiR-21-mediated Metabolic Alteration of 92 Cancer-associated Fibroblasts and Its Effect on Pancreatic Cancer Cell Behavior. Int J Biol Sci 2018; 14: 100-110 [PMID: 29483829 DOI: 10.7150/ijbs.22555]
- 93 Cheng TY, Yang YC, Wang HP, Tien YW, Shun CT, Huang HY, Hsiao M, Hua KT. Pyruvate kinase M2 promotes pancreatic ductal adenocarcinoma invasion and metastasis through phosphorylation and stabilization of PAK2 protein. Oncogene 2018; 37: 1730-1742 [PMID: 29335522 DOI: 10.1038/s41388-017-0086-y]
- Kim DJ, Park YS, Kang MG, You YM, Jung Y, Koo H, Kim JA, Kim MJ, Hong SM, Lee KB, Jang 94 JJ, Park KC, Yeom YI. Pyruvate kinase isoenzyme M2 is a therapeutic target of gemcitabineresistant pancreatic cancer cells. Exp Cell Res 2015; 336: 119-129 [PMID: 26112218 DOI: 10.1016/j.yexcr.2015.05.017]
- Li C, Zhao Z, Zhou Z, Liu R. PKM2 Promotes Cell Survival and Invasion Under Metabolic Stress by Enhancing Warburg Effect in Pancreatic Ductal Adenocarcinoma. Dig Dis Sci 2016; 61: 767-773 [PMID: 26500118 DOI: 10.1007/s10620-015-3931-2]
- 96 Cui J, Shi M, Xie D, Wei D, Jia Z, Zheng S, Gao Y, Huang S, Xie K. FOXM1 promotes the warburg effect and pancreatic cancer progression via transactivation of LDHA expression. Clin Cancer Res 2014; 20: 2595-2606 [PMID: 24634381 DOI: 10.1158/1078-0432.CCR-13-2407]
- Giatromanolaki A, Sivridis E, Gatter KC, Turley H, Harris AL, Koukourakis MI; Tumour and Angiogenesis Research Group. Lactate dehydrogenase 5 (LDH-5) expression in endometrial cancer relates to the activated VEGF/VEGFR2(KDR) pathway and prognosis. Gynecol Oncol 2006; 103: 912-918 [PMID: 16837029 DOI: 10.1016/j.ygyno.2006.05.043]
- Jiang W, Zhou F, Li N, Li Q, Wang L. FOXM1-LDHA signaling promoted gastric cancer glycolytic 98 phenotype and progression. Int J Clin Exp Pathol 2015; 8: 6756-6763 [PMID: 26261559]
- 99 Kobari M, Hisano H, Matsuno S, Sato T, Kan M, Tachibana T. Establishment of six human pancreatic cancer cell lines and their sensitivities to anti-tumor drugs. Tohoku J Exp Med 1986; 150: 231-248 [PMID: 3547771 DOI: 10.1620/tjem.150.231]
- 100 Koukourakis MI, Kakouratos C, Kalamida D, Bampali Z, Mavropoulou S, Sivridis E, Giatromanolaki A. Hypoxia-inducible proteins HIF1a and lactate dehydrogenase LDH5, key markers of anaerobic metabolism, relate with stem cell markers and poor post-radiotherapy outcome in bladder cancer. Int J Radiat Biol 2016; 92: 353-363 [PMID: 27010533 DOI: 10.3109/09553002.2016.1162921]
- 101 Thonsri U, Seubwai W, Waraasawapati S, Sawanyawisuth K, Vaeteewoottacharn K, Boonmars T, Cha'on U. Overexpression of lactate dehydrogenase A in cholangiocarcinoma is correlated with poor prognosis. Histol Histopathol 2017; 32: 503-510 [PMID: 27615379 DOI: 10.14670/HH-11-819]
- 102 Yu C, Hou L, Cui H, Zhang L, Tan X, Leng X, Li Y. LDHA upregulation independently predicts poor survival in lung adenocarcinoma, but not in lung squamous cell carcinoma. Future Oncol 2018; 14: 2483-2492 [PMID: 29756998 DOI: 10.2217/fon-2018-0177]
- 103 Semenza GL, Jiang BH, Leung SW, Passantino R, Concordet JP, Maire P, Giallongo A. Hypoxia response elements in the aldolase A, enolase 1, and lactate dehydrogenase A gene promoters contain essential binding sites for hypoxia-inducible factor 1. J Biol Chem 1996; 271: 32529-32537 [PMID: 8955077 DOI: 10.1074/jbc.271.51.32529]
- Shim H, Dolde C, Lewis BC, Wu CS, Dang G, Jungmann RA, Dalla-Favera R, Dang CV. c-Myc 104 transactivation of LDH-A: implications for tumor metabolism and growth. Proc Natl Acad Sci USA 1997; 94: 6658-6663 [PMID: 9192621 DOI: 10.1073/pnas.94.13.6658]
- Li SS, Pan YE, Sharief FS, Evans MJ, Lin MF, Clinton GM, Holbrook JJ. Cancer-associated lactate 105 dehydrogenase is a tyrosylphosphorylated form of human LDH-A, skeletal muscle isoenzyme. Cancer Invest 1988; 6: 93-101 [PMID: 3365574 DOI: 10.3109/07357908809077032]
- 106 Liu J, Chen G, Liu Z, Liu S, Cai Z, You P, Ke Y, Lai L, Huang Y, Gao H, Zhao L, Pelicano H, Huang P, McKeehan WL, Wu CL, Wang C, Zhong W, Wang F. Aberrant FGFR Tyrosine Kinase Signaling Enhances the Warburg Effect by Reprogramming LDH Isoform Expression and Activity in Prostate Cancer. Cancer Res 2018; 78: 4459-4470 [PMID: 29891507 DOI: 10.1158/0008-5472.CAN-17-3226
- 107 Faloppi L, Bianconi M, Giampieri R, Sobrero A, Labianca R, Ferrari D, Barni S, Aitini E, Zaniboni A, Boni C, Caprioni F, Mosconi S, Fanello S, Berardi R, Bittoni A, Andrikou K, Cinquini M, Torri V, Scartozzi M, Cascinu S; Italian Group for the Study of Digestive Tract Cancer (GISCAD). The value of lactate dehydrogenase serum levels as a prognostic and predictive factor for advanced pancreatic cancer patients receiving sorafenib. Oncotarget 2015; 6: 35087-35094 [PMID: 26397228 DOI: 10.18632/oncotarget.5197]
- 108 Gan J, Wang W, Yang Z, Pan J, Zheng L, Yin L. Prognostic value of pretreatment serum lactate dehydrogenase level in pancreatic cancer patients: A meta-analysis of 18 observational studies. Medicine (Baltimore) 2018; 97: e13151 [PMID: 30431587 DOI: 10.1097/MD.00000000013151]
- Ji F, Fu SJ, Guo ZY, Pang H, Ju WQ, Wang DP, Hua YP, He XS. Prognostic value of combined



preoperative lactate dehydrogenase and alkaline phosphatase levels in patients with resectable pancreatic ductal adenocarcinoma. Medicine (Baltimore) 2016; 95: e4065 [PMID: 27399091 DOI: 10.1097/MD.000000000004065

110 Mann JR, Pearson D, Barrett A, Raafat F, Barnes JM, Wallendszus KR. Results of the United Kingdom Children's Cancer Study Group's malignant germ cell tumor studies. Cancer 1989; 63: 1657-1667 [PMID: 2467734 DOI:

10.1002/1097-0142(19900501)63:9<1657::aid-cncr2820630902>3.0.co;2-8

- 111 Tas F, Aykan F, Alici S, Kaytan E, Aydiner A, Topuz E. Prognostic factors in pancreatic carcinoma: serum LDH levels predict survival in metastatic disease. Am J Clin Oncol 2001; 24: 547-550 [PMID: 11801751 DOI: 10.1097/00000421-200112000-00003]
- Xiao Y, Chen W, Xie Z, Shao Z, Xie H, Qin G, Zhao N. Prognostic relevance of lactate 112 dehydrogenase in advanced pancreatic ductal adenocarcinoma patients. BMC Cancer 2017; 17: 25 [PMID: 28056913 DOI: 10.1186/s12885-016-3012-8]
- 113 Annas D, Cheon SY, Yusuf M, Bae SJ, Ha KT, Park KH. Synthesis and initial screening of lactate dehydrogenase inhibitor activity of 1,3-benzodioxole derivatives. Sci Rep 2020; 10: 19889 [PMID: 33199724 DOI: 10.1038/s41598-020-77056-4]
- 114 Moir JAG, Long A, Haugk B, French JJ, Charnley RM, Manas DM, Wedge SR, Mann J, Robinson SM, White SA. Therapeutic Strategies Toward Lactate Dehydrogenase Within the Tumor Microenvironment of Pancreatic Cancer. Pancreas 2020; 49: 1364-1371 [PMID: 33122526 DOI: 10.1097/MPA.000000000001689
- 115 Stanton RC. Glucose-6-phosphate dehydrogenase, NADPH, and cell survival. *IUBMB Life* 2012; 64: 362-369 [PMID: 22431005 DOI: 10.1002/iub.1017]
- 116 Santana-Codina N, Roeth AA, Zhang Y, Yang A, Mashadova O, Asara JM, Wang X, Bronson RT, Lyssiotis CA, Ying H, Kimmelman AC. Oncogenic KRAS supports pancreatic cancer through regulation of nucleotide synthesis. Nat Commun 2018; 9: 4945 [PMID: 30470748 DOI: 10.1038/s41467-018-07472-8]
- 117 Sharma N, Bhushan A, He J, Kaushal G, Bhardwaj V. Metabolic plasticity imparts erlotinibresistance in pancreatic cancer by upregulating glucose-6-phosphate dehydrogenase. Cancer Metab 2020; 8: 19 [PMID: 32974013 DOI: 10.1186/s40170-020-00226-5]
- 118 Shukla SK, Purohit V, Mehla K, Gunda V, Chaika NV, Vernucci E, King RJ, Abrego J, Goode GD, Dasgupta A, Illies AL, Gebregiworgis T, Dai B, Augustine JJ, Murthy D, Attri KS, Mashadova O, Grandgenett PM, Powers R, Ly QP, Lazenby AJ, Grem JL, Yu F, Matés JM, Asara JM, Kim JW, Hankins JH, Weekes C, Hollingsworth MA, Serkova NJ, Sasson AR, Fleming JB, Oliveto JM, Lyssiotis CA, Cantley LC, Berim L, Singh PK. MUC1 and HIF-1alpha Signaling Crosstalk Induces Anabolic Glucose Metabolism to Impart Gemcitabine Resistance to Pancreatic Cancer. Cancer Cell 2017; 32: 71-87.e7 [PMID: 28697344 DOI: 10.1016/j.ccell.2017.06.004]
- Viale A, Pettazzoni P, Lyssiotis CA, Ying H, Sánchez N, Marchesini M, Carugo A, Green T, Seth S, 119 Giuliani V, Kost-Alimova M, Muller F, Colla S, Nezi L, Genovese G, Deem AK, Kapoor A, Yao W, Brunetto E, Kang Y, Yuan M, Asara JM, Wang YA, Heffernan TP, Kimmelman AC, Wang H, Fleming JB, Cantley LC, DePinho RA, Draetta GF. Oncogene ablation-resistant pancreatic cancer cells depend on mitochondrial function. Nature 2014; 514: 628-632 [PMID: 25119024 DOI: 10.1038/nature13611
- 120 Sancho P, Burgos-Ramos E, Tavera A, Bou Kheir T, Jagust P, Schoenhals M, Barneda D, Sellers K, Campos-Olivas R, Graña O, Viera CR, Yuneva M, Sainz B Jr, Heeschen C. MYC/PGC-1a Balance Determines the Metabolic Phenotype and Plasticity of Pancreatic Cancer Stem Cells. Cell Metab 2015; 22: 590-605 [PMID: 26365176 DOI: 10.1016/j.cmet.2015.08.015]
- 121 Kovalenko I, Glasauer A, Schöckel L, Sauter DR, Ehrmann A, Sohler F, Hägebarth A, Novak I, Christian S. Identification of KCa3.1 Channel as a Novel Regulator of Oxidative Phosphorylation in a Subset of Pancreatic Carcinoma Cell Lines. PLoS One 2016; 11: e0160658 [PMID: 27494181 DOI: 10.1371/journal.pone.0160658]
- 122 Dey P, Baddour J, Muller F, Wu CC, Wang H, Liao WT, Lan Z, Chen A, Gutschner T, Kang Y, Fleming J, Satani N, Zhao D, Achreja A, Yang L, Lee J, Chang E, Genovese G, Viale A, Ying H, Draetta G, Maitra A, Wang YA, Nagrath D, DePinho RA. Genomic deletion of malic enzyme 2 confers collateral lethality in pancreatic cancer. Nature 2017; 542: 119-123 [PMID: 28099419 DOI: 10.1038/nature21052]
- Zarei M, Lal S, Parker SJ, Nevler A, Vaziri-Gohar A, Dukleska K, Mambelli-Lisboa NC, Moffat C, 123 Blanco FF, Chand SN, Jimbo M, Cozzitorto JA, Jiang W, Yeo CJ, Londin ER, Seifert EL, Metallo CM, Brody JR, Winter JM. Posttranscriptional Upregulation of IDH1 by HuR Establishes a Powerful Survival Phenotype in Pancreatic Cancer Cells. Cancer Res 2017; 77: 4460-4471 [PMID: 28652247 DOI: 10.1158/0008-5472.CAN-17-0015]
- 124 Nishi K, Suzuki M, Yamamoto N, Matsumoto A, Iwase Y, Yamasaki K, Otagiri M, Yumita N. Glutamine Deprivation Enhances Acetyl-CoA Carboxylase Inhibitor-induced Death of Human Pancreatic Cancer Cells. Anticancer Res 2018; 38: 6683-6689 [PMID: 30504377 DOI: 10.21873/anticanres.13036
- 125 Son J, Lyssiotis CA, Ying H, Wang X, Hua S, Ligorio M, Perera RM, Ferrone CR, Mullarky E, Shyh-Chang N, Kang Y, Fleming JB, Bardeesy N, Asara JM, Haigis MC, DePinho RA, Cantley LC, Kimmelman AC. Glutamine supports pancreatic cancer growth through a KRAS-regulated metabolic pathway. Nature 2013; 496: 101-105 [PMID: 23535601 DOI: 10.1038/nature12040]
- Chakrabarti G, Moore ZR, Luo X, Ilcheva M, Ali A, Padanad M, Zhou Y, Xie Y, Burma S, 126



Scaglioni PP, Cantley LC, DeBerardinis RJ, Kimmelman AC, Lyssiotis CA, Boothman DA. Targeting glutamine metabolism sensitizes pancreatic cancer to PARP-driven metabolic catastrophe induced by B-lapachone. Cancer Metab 2015; 3: 12 [PMID: 26462257 DOI: 10.1186/s40170-015-0137-1]

- 127 Chen R, Lai LA, Sullivan Y, Wong M, Wang L, Riddell J, Jung L, Pillarisetty VG, Brentnall TA, Pan S. Disrupting glutamine metabolic pathways to sensitize gemcitabine-resistant pancreatic cancer. Sci Rep 2017; 7: 7950 [PMID: 28801576 DOI: 10.1038/s41598-017-08436-6]
- Jia C, Li H, Fu D, Lan Y. GFAT1/HBP/O-GlcNAcylation Axis Regulates β-Catenin Activity to 128 Promote Pancreatic Cancer Aggressiveness. Biomed Res Int 2020; 2020: 1921609 [PMID: 32149084 DOI: 10.1155/2020/1921609]
- 129 Li D, Fu Z, Chen R, Zhao X, Zhou Y, Zeng B, Yu M, Zhou Q, Lin Q, Gao W, Ye H, Zhou J, Li Z, Liu Y. Inhibition of glutamine metabolism counteracts pancreatic cancer stem cell features and sensitizes cells to radiotherapy. Oncotarget 2015; 6: 31151-31163 [PMID: 26439804 DOI: 10.18632/oncotarget.5150
- 130 Lyssiotis CA, Son J, Cantley LC, Kimmelman AC. Pancreatic cancers rely on a novel glutamine metabolism pathway to maintain redox balance. Cell Cycle 2013; 12: 1987-1988 [PMID: 23759579 DOI: 10.4161/cc.25307]
- Raho S, Capobianco L, Malivindi R, Vozza A, Piazzolla C, De Leonardis F, Gorgoglione R, Scarcia 131 P, Pezzuto F, Agrimi G, Barile SN, Pisano I, Reshkin SJ, Greco MR, Cardone RA, Rago V, Li Y, Marobbio CMT, Sommergruber W, Riley CL, Lasorsa FM, Mills E, Vegliante MC, De Benedetto GE, Fratantonio D, Palmieri L, Dolce V, Fiermonte G. KRAS-regulated glutamine metabolism requires UCP2-mediated aspartate transport to support pancreatic cancer growth. Nat Metab 2020; 2: 1373-1381 [PMID: 33230296 DOI: 10.1038/s42255-020-00315-1]
- 132 Wang J, Wang B, Ren H, Chen W. miR-9-5p inhibits pancreatic cancer cell proliferation, invasion and glutamine metabolism by targeting GOT1. Biochem Biophys Res Commun 2019; 509: 241-248 [PMID: 30591220 DOI: 10.1016/j.bbrc.2018.12.114]
- Jeong SM, Hwang S, Park K, Yang S, Seong RH. Enhanced mitochondrial glutamine anaplerosis 133 suppresses pancreatic cancer growth through autophagy inhibition. Sci Rep 2016; 6: 30767 [PMID: 27477484 DOI: 10.1038/srep30767]
- 134 Recouvreux MV, Moldenhauer MR, Galenkamp KMO, Jung M, James B, Zhang Y, Lowy A, Bagchi A, Commisso C. Glutamine depletion regulates Slug to promote EMT and metastasis in pancreatic cancer. J Exp Med 2020; 217 [PMID: 32510550 DOI: 10.1084/jem.20200388]
- 135 Roux C, Riganti C, Borgogno SF, Curto R, Curcio C, Catanzaro V, Digilio G, Padovan S, Puccinelli MP. Isabello M. Aime S. Cappello P. Novelli F. Endogenous glutamine decrease is associated with pancreatic cancer progression. Oncotarget 2017; 8: 95361-95376 [PMID: 29221133 DOI: 10.18632/oncotarget.20545]
- 136 Bao B, Wang Z, Ali S, Ahmad A, Azmi AS, Sarkar SH, Banerjee S, Kong D, Li Y, Thakur S, Sarkar FH. Metformin inhibits cell proliferation, migration and invasion by attenuating CSC function mediated by deregulating miRNAs in pancreatic cancer cells. Cancer Prev Res (Phila) 2012; 5: 355-364 [PMID: 22086681 DOI: 10.1158/1940-6207.CAPR-11-0299]
- Cheng G, Zielonka J, Ouari O, Lopez M, McAllister D, Boyle K, Barrios CS, Weber JJ, Johnson 137 BD, Hardy M, Dwinell MB, Kalyanaraman B. Mitochondria-Targeted Analogues of Metformin Exhibit Enhanced Antiproliferative and Radiosensitizing Effects in Pancreatic Cancer Cells. Cancer Res 2016; 76: 3904-3915 [PMID: 27216187 DOI: 10.1158/0008-5472.CAN-15-2534]
- Masoud R, Reyes-Castellanos G, Lac S, Garcia J, Dou S, Shintu L, Abdel Hadi N, Gicquel T, El 138 Kaoutari A, Diémé B, Tranchida F, Cormareche L, Borge L, Gayet O, Pasquier E, Dusetti N, Iovanna J, Carrier A. Targeting Mitochondrial Complex I Overcomes Chemoresistance in High OXPHOS Pancreatic Cancer. Cell Rep Med 2020; 1: 100143 [PMID: 33294863 DOI: 10.1016/j.xcrm.2020.100143]
- 139 Rajeshkumar NV, Yabuuchi S, Pai SG, De Oliveira E, Kamphorst JJ, Rabinowitz JD, Tejero H, Al-Shahrour F, Hidalgo M, Maitra A, Dang CV. Treatment of Pancreatic Cancer Patient-Derived Xenograft Panel with Metabolic Inhibitors Reveals Efficacy of Phenformin. Clin Cancer Res 2017; 23: 5639-5647 [PMID: 28611197 DOI: 10.1158/1078-0432.CCR-17-1115]
- 140 Kordes S. Pollak MN, Zwinderman AH, Mathôt RA, Weterman MJ, Beeker A, Punt CJ, Richel DJ, Wilmink JW. Metformin in patients with advanced pancreatic cancer: a double-blind, randomised, placebo-controlled phase 2 trial. Lancet Oncol 2015; 16: 839-847 [PMID: 26067687 DOI: 10.1016/S1470-2045(15)00027-3]
- Reni M, Dugnani E, Cereda S, Belli C, Balzano G, Nicoletti R, Liberati D, Pasquale V, Scavini M, 141 Maggiora P, Sordi V, Lampasona V, Ceraulo D, Di Terlizzi G, Doglioni C, Falconi M, Piemonti L. (Ir)relevance of Metformin Treatment in Patients with Metastatic Pancreatic Cancer: An Open-Label, Randomized Phase II Trial. Clin Cancer Res 2016; 22: 1076-1085 [PMID: 26459175 DOI: 10.1158/1078-0432.CCR-15-1722
- 142 Shi YQ, Zhou XC, Du P, Yin MY, Xu L, Chen WJ, Xu CF. Relationships are between metformin use and survival in pancreatic cancer patients concurrent with diabetes: A systematic review and meta-analysis. Medicine (Baltimore) 2020; 99: e21687 [PMID: 32925714 DOI: 10.1097/MD.00000000021687
- Alistar A, Morris BB, Desnoyer R, Klepin HD, Hosseinzadeh K, Clark C, Cameron A, Leyendecker 143 J, D'Agostino R Jr, Topaloglu U, Boteju LW, Boteju AR, Shorr R, Zachar Z, Bingham PM, Ahmed T, Crane S, Shah R, Migliano JJ, Pardee TS, Miller L, Hawkins G, Jin G, Zhang W, Pasche B.



Safety and tolerability of the first-in-class agent CPI-613 in combination with modified FOLFIRINOX in patients with metastatic pancreatic cancer: a single-centre, open-label, doseescalation, phase 1 trial. Lancet Oncol 2017; 18: 770-778 [PMID: 28495639 DOI: 10.1016/S1470-2045(17)30314-5]

- 144 Waddell N, Pajic M, Patch AM, Chang DK, Kassahn KS, Bailey P, Johns AL, Miller D, Nones K, Quek K, Quinn MC, Robertson AJ, Fadlullah MZ, Bruxner TJ, Christ AN, Harliwong I, Idrisoglu S, Manning S, Nourse C, Nourbakhsh E, Wani S, Wilson PJ, Markham E, Cloonan N, Anderson MJ, Fink JL, Holmes O, Kazakoff SH, Leonard C, Newell F, Poudel B, Song S, Taylor D, Waddell N, Wood S, Xu Q, Wu J, Pinese M, Cowley MJ, Lee HC, Jones MD, Nagrial AM, Humphris J, Chantrill LA, Chin V, Steinmann AM, Mawson A, Humphrey ES, Colvin EK, Chou A, Scarlett CJ, Pinho AV, Girv-Laterriere M, Rooman I, Samra JS, Kench JG, Pettitt JA, Merrett ND, Toon C, Epari K, Nguyen NQ, Barbour A, Zeps N, Jamieson NB, Graham JS, Niclou SP, Bjerkvig R, Grützmann R, Aust D, Hruban RH, Maitra A, Iacobuzio-Donahue CA, Wolfgang CL, Morgan RA, Lawlor RT, Corbo V, Bassi C, Falconi M, Zamboni G, Tortora G, Tempero MA; Australian Pancreatic Cancer Genome Initiative, Gill AJ, Eshleman JR, Pilarsky C, Scarpa A, Musgrove EA, Pearson JV, Biankin AV, Grimmond SM. Whole genomes redefine the mutational landscape of pancreatic cancer. Nature 2015; 518: 495-501 [PMID: 25719666 DOI: 10.1038/nature14169]
- 145 Bailey P, Chang DK, Nones K, Johns AL, Patch AM, Gingras MC, Miller DK, Christ AN, Bruxner TJ, Quinn MC, Nourse C, Murtaugh LC, Harliwong I, Idrisoglu S, Manning S, Nourbakhsh E, Wani S, Fink L, Holmes O, Chin V, Anderson MJ, Kazakoff S, Leonard C, Newell F, Waddell N, Wood S, Xu Q, Wilson PJ, Cloonan N, Kassahn KS, Taylor D, Quek K, Robertson A, Pantano L, Mincarelli L, Sanchez LN, Evers L, Wu J, Pinese M, Cowley MJ, Jones MD, Colvin EK, Nagrial AM, Humphrey ES, Chantrill LA, Mawson A, Humphris J, Chou A, Pajic M, Scarlett CJ, Pinho AV, Giry-Laterriere M, Rooman I, Samra JS, Kench JG, Lovell JA, Merrett ND, Toon CW, Epari K, Nguyen NQ, Barbour A, Zeps N, Moran-Jones K, Jamieson NB, Graham JS, Duthie F, Oien K, Hair J, Grützmann R, Maitra A, Iacobuzio-Donahue CA, Wolfgang CL, Morgan RA, Lawlor RT, Corbo V, Bassi C, Rusev B, Capelli P, Salvia R, Tortora G, Mukhopadhyay D, Petersen GM; Australian Pancreatic Cancer Genome Initiative, Munzy DM, Fisher WE, Karim SA, Eshleman JR, Hruban RH, Pilarsky C, Morton JP, Sansom OJ, Scarpa A, Musgrove EA, Bailey UM, Hofmann O, Sutherland RL, Wheeler DA, Gill AJ, Gibbs RA, Pearson JV, Waddell N, Biankin AV, Grimmond SM. Genomic analyses identify molecular subtypes of pancreatic cancer. Nature 2016; 531: 47-52 [PMID: 26909576 DOI: 10.1038/nature16965]
- 146 Hoadley KA, Yau C, Hinoue T, Wolf DM, Lazar AJ, Drill E, Shen R, Taylor AM, Cherniack AD, Thorsson V, Akbani R, Bowlby R, Wong CK, Wiznerowicz M, Sanchez-Vega F, Robertson AG, Schneider BG, Lawrence MS, Noushmehr H, Malta TM; Cancer Genome Atlas Network, Stuart JM, Benz CC, Laird PW. Cell-of-Origin Patterns Dominate the Molecular Classification of 10,000 Tumors from 33 Types of Cancer. Cell 2018; 173: 291-304.e6 [PMID: 29625048 DOI: 10.1016/j.cell.2018.03.022]
- 147 Witkiewicz AK, McMillan EA, Balaji U, Baek G, Lin WC, Mansour J, Mollaee M, Wagner KU, Koduru P, Yopp A, Choti MA, Yeo CJ, McCue P, White MA, Knudsen ES. Whole-exome sequencing of pancreatic cancer defines genetic diversity and therapeutic targets. Nat Commun 2015; 6: 6744 [PMID: 25855536 DOI: 10.1038/ncomms7744]
- 148 Johnstone TC, Park GY, Lippard SJ. Understanding and improving platinum anticancer drugs-phenanthriplatin. Anticancer Res 2014; 34: 471-476 [PMID: 24403503]
- 149 Golan T, Kanji ZS, Epelbaum R, Devaud N, Dagan E, Holter S, Aderka D, Paluch-Shimon S, Kaufman B, Gershoni-Baruch R, Hedley D, Moore MJ, Friedman E, Gallinger S. Overall survival and clinical characteristics of pancreatic cancer in BRCA mutation carriers. Br J Cancer 2014; 111: 1132-1138 [PMID: 25072261 DOI: 10.1038/bjc.2014.418]
- 150 Pishvaian MJ, Blais EM, Brody JR, Rahib L, Lyons E, De Arbeloa P, Hendifar A, Mikhail S, Chung V, Sohal DPS, Leslie S, Mason K, Tibbets L, Madhavan S, Matrisian LM, Petricoin E. Outcomes in Patients With Pancreatic Adenocarcinoma With Genetic Mutations in DNA Damage Response Pathways: Results From the Know Your Tumor Program. JCO Precis Oncol 2019: 1-10 [DOI: 10.1200/po.19.00115]
- Yu S, Agarwal P, Mamtani R, Symecko H, Spielman K, O'Hara M, O'Dwyer PJ, Schneider C, Teitelbaum U, Nathanson KL, Domchek SM, Reiss KA. Retrospective Survival Analysis of Patients With Resected Pancreatic Ductal Adenocarcinoma and a Germline BRCA or PALB2 Mutation. JCO Precis Oncol 2018; 1-11 [DOI: 10.1200/po.18.00271]
- Rack JG, Perina D, Ahel I. Macrodomains: Structure, Function, Evolution, and Catalytic Activities. 152 Annu Rev Biochem 2016; 85: 431-454 [PMID: 26844395 DOI: 10.1146/annurev-biochem-060815-014935]
- 153 Barkauskaite E, Jankevicius G, Ladurner AG, Ahel I, Timinszky G. The recognition and removal of cellular poly(ADP-ribose) signals. FEBS J 2013; 280: 3491-3507 [PMID: 23711178 DOI: 10.1111/febs.12358
- 154 Harrision D, Gravells P, Thompson R, Bryant HE. Poly(ADP-Ribose) Glycohydrolase (PARG) vs. Poly(ADP-Ribose) Polymerase (PARP) - Function in Genome Maintenance and Relevance of Inhibitors for Anti-cancer Therapy. Front Mol Biosci 2020; 7: 191 [PMID: 33005627 DOI: 10.3389/fmolb.2020.00191
- 155 Lear AL, Perkins HR. O-acetylation of peptidoglycan in Neisseria gonorrhoeae. Investigation of lipid-linked intermediates and glycan chains newly incorporated into the cell wall. J Gen Microbiol



1986; 132: 2413-2420 [PMID: 3098911 DOI: 10.1099/00221287-132-9-2413]

- Shirai H, Poetsch AR, Gunji A, Maeda D, Fujimori H, Fujihara H, Yoshida T, Ogino H, Masutani 156 M. PARG dysfunction enhances DNA double strand break formation in S-phase after alkylation DNA damage and augments different cell death pathways. Cell Death Dis 2013; 4: e656 [PMID: 23744356 DOI: 10.1038/cddis.2013.133]
- 157 Gravells P, Neale J, Grant E, Nathubhai A, Smith KM, James DI, Bryant HE. Radiosensitization with an inhibitor of poly(ADP-ribose) glycohydrolase: A comparison with the PARP1/2/3 inhibitor olaparib. DNA Repair (Amst) 2018; 61: 25-36 [PMID: 29179156 DOI: 10.1016/j.dnarep.2017.11.004]
- 158 Pillay N, Tighe A, Nelson L, Littler S, Coulson-Gilmer C, Bah N, Golder A, Bakker B, Spierings DCJ, James DI, Smith KM, Jordan AM, Morgan RD, Ogilvie DJ, Foijer F, Jackson DA, Taylor SS. DNA Replication Vulnerabilities Render Ovarian Cancer Cells Sensitive to Poly(ADP-Ribose) Glycohydrolase Inhibitors. Cancer Cell 2019; 35: 519-533.e8 [PMID: 30889383 DOI: 10.1016/j.ccell.2019.02.004]
- 159 Gravells P, Grant E, Smith KM, James DI, Bryant HE. Specific killing of DNA damage-response deficient cells with inhibitors of poly(ADP-ribose) glycohydrolase. DNA Repair (Amst) 2017; 52: 81-91 [PMID: 28254358 DOI: 10.1016/j.dnarep.2017.02.010]
- Iorns E, Lord CJ, Grigoriadis A, McDonald S, Fenwick K, Mackay A, Mein CA, Natrajan R, 160 Savage K, Tamber N, Reis-Filho JS, Turner NC, Ashworth A. Integrated functional, gene expression and genomic analysis for the identification of cancer targets. PLoS One 2009; 4: e5120 [PMID: 19357772 DOI: 10.1371/journal.pone.0005120]
- Masaki T, Shiratori Y, Rengifo W, Igarashi K, Yamagata M, Kurokohchi K, Uchida N, Miyauchi Y, 161 Yoshiji H, Watanabe S, Omata M, Kuriyama S. Cyclins and cyclin-dependent kinases: comparative study of hepatocellular carcinoma vs cirrhosis. Hepatology 2003; 37: 534-543 [PMID: 12601350 DOI: 10.1053/jhep.2003.50112]
- 162 Mir SE, De Witt Hamer PC, Krawczyk PM, Balaj L, Claes A, Niers JM, Van Tilborg AA, Zwinderman AH, Geerts D, Kaspers GJ, Peter Vandertop W, Cloos J, Tannous BA, Wesseling P, Aten JA, Noske DP, Van Noorden CJ, Würdinger T. In silico analysis of kinase expression identifies WEE1 as a gatekeeper against mitotic catastrophe in glioblastoma. Cancer Cell 2010; 18: 244-257 [PMID: 20832752 DOI: 10.1016/j.ccr.2010.08.011]
- 163 Wang H, Huang M, Zhang DY, Zhang F. Global profiling of signaling networks: study of breast cancer stem cells and potential regulation. Oncologist 2011; 16: 966-979 [PMID: 21665913 DOI: 10.1634/theoncologist.2010-0230
- 164 Dreyer SB, Upstill-Goddard R, Paulus-Hock V, Paris C, Lampraki EM, Dray E, Serrels B, Caligiuri G, Rebus S, Plenker D, Galluzzo Z, Brunton H, Cunningham R, Tesson M, Nourse C, Bailey UM, Jones M, Moran-Jones K, Wright DW, Duthie F, Oien K, Evers L, McKay CJ, McGregor GA, Gulati A, Brough R, Bajrami I, Pettitt S, Dziubinski ML, Candido J, Balkwill F, Barry ST, Grützmann R, Rahib L; Glasgow Precision Oncology Laboratory,; Australian Pancreatic Cancer Genome Initiative, Johns A, Pajic M, Froeling FEM, Beer P, Musgrove EA, Petersen GM, Ashworth A, Frame MC, Crawford HC, Simeone DM, Lord C, Mukhopadhyay D, Pilarsky C, Tuveson DA, Cooke SL, Jamieson NB, Morton JP, Sansom OJ, Bailey PJ, Biankin AV, Chang DK. Targeting DNA Damage Response and Replication Stress in Pancreatic Cancer. Gastroenterology 2021; 160: 362-377.e13 [PMID: 33039466 DOI: 10.1053/j.gastro.2020.09.043]
- 165 Kausar T, Schreiber JS, Karnak D, Parsels LA, Parsels JD, Davis MA, Zhao L, Maybaum J, Lawrence TS, Morgan MA. Sensitization of Pancreatic Cancers to Gemcitabine Chemoradiation by WEE1 Kinase Inhibition Depends on Homologous Recombination Repair. Neoplasia 2015; 17: 757-766 [PMID: 26585231 DOI: 10.1016/j.neo.2015.09.006]
- Saini P, Li Y, Dobbelstein M. Wee1 is required to sustain ATR/Chk1 signaling upon replicative 166 stress. Oncotarget 2015; 6: 13072-13087 [PMID: 25965828 DOI: 10.18632/oncotarget.3865]
- 167 Lal S, Burkhart RA, Beeharry N, Bhattacharjee V, Londin ER, Cozzitorto JA, Romeo C, Jimbo M, Norris ZA, Yeo CJ, Sawicki JA, Winter JM, Rigoutsos I, Yen TJ, Brody JR. HuR posttranscriptionally regulates WEE1: implications for the DNA damage response in pancreatic cancer cells. Cancer Res 2014; 74: 1128-1140 [PMID: 24536047 DOI: 10.1158/0008-5472.CAN-13-1915]
- 168 Lal S, Zarei M, Chand SN, Dylgjeri E, Mambelli-Lisboa NC, Pishvaian MJ, Yeo CJ, Winter JM, Brody JR. WEE1 inhibition in pancreatic cancer cells is dependent on DNA repair status in a context dependent manner. Sci Rep 2016; 6: 33323 [PMID: 27616351 DOI: 10.1038/srep33323]
- 169 Rajeshkumar NV, De Oliveira E, Ottenhof N, Watters J, Brooks D, Demuth T, Shumway SD, Mizuarai S, Hirai H, Maitra A, Hidalgo M. MK-1775, a potent Wee1 inhibitor, synergizes with gemcitabine to achieve tumor regressions, selectively in p53-deficient pancreatic cancer xenografts. Clin Cancer Res 2011; 17: 2799-2806 [PMID: 21389100 DOI: 10.1158/1078-0432.CCR-10-2580]
- 170 Jin MH, Nam AR, Park JE, Bang JH, Bang YJ, Oh DY. Therapeutic Co-targeting of WEE1 and ATM Downregulates PD-L1 Expression in Pancreatic Cancer. Cancer Res Treat 2020; 52: 149-166 [PMID: 31291716 DOI: 10.4143/crt.2019.183]
- Agostini LC, Jain A, Shupp A, Nevler A, McCarthy G, Bussard KM, Yeo CJ, Brody JR. Combined 171 Targeting of PARG and Wee1 Causes Decreased Cell Survival and DNA Damage in an S-Phase-Dependent Manner. Mol Cancer Res 2021; 19: 207-214 [PMID: 33257507 DOI: 10.1158/1541-7786.MCR-20-0708
- 172 Krishnakumar R, Kraus WL. The PARP side of the nucleus: molecular actions, physiological



outcomes, and clinical targets. Mol Cell 2010; 39: 8-24 [PMID: 20603072 DOI: 10.1016/j.molcel.2010.06.017]

- Rouleau M, Patel A, Hendzel MJ, Kaufmann SH, Poirier GG. PARP inhibition: PARP1 and 173 beyond. Nat Rev Cancer 2010; 10: 293-301 [PMID: 20200537 DOI: 10.1038/nrc2812]
- 174 Bagnolini G, Milano D, Manerba M, Schipani F, Ortega JA, Gioia D, Falchi F, Balboni A, Farabegoli F, De Franco F, Robertson J, Pellicciari R, Pallavicini I, Peri S, Minucci S, Girotto S, Di Stefano G, Roberti M, Cavalli A. Svnthetic Lethality in Pancreatic Cancer: Discovery of a New RAD51-BRCA2 Small Molecule Disruptor That Inhibits Homologous Recombination and Synergizes with Olaparib. J Med Chem 2020; 63: 2588-2619 [PMID: 32037829 DOI: 10.1021/acs.jmedchem.9b01526]
- 175 Farmer H, McCabe N, Lord CJ, Tutt AN, Johnson DA, Richardson TB, Santarosa M, Dillon KJ, Hickson I, Knights C, Martin NM, Jackson SP, Smith GC, Ashworth A. Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. Nature 2005; 434: 917-921 [PMID: 15829967 DOI: 10.1038/nature03445]
- 176 Miller AL, Garcia PL, Yoon KJ. Developing effective combination therapy for pancreatic cancer: An overview. Pharmacol Res 2020; 155: 104740 [PMID: 32135247 DOI: 10.1016/j.phrs.2020.104740
- Lai SW, Bamodu OA, Chen JH, Wu AT, Lee WH, Chao TY, Yeh CT. Targeted PARP Inhibition 177 Combined with FGFR1 Blockade is Synthetically Lethal to Malignant Cells in Patients with Pancreatic Cancer. Cells 2020; 9 [PMID: 32276472 DOI: 10.3390/cells9040911]
- Noordermeer SM, van Attikum H. PARP Inhibitor Resistance: A Tug-of-War in BRCA-Mutated 178 Cells. Trends Cell Biol 2019; 29: 820-834 [PMID: 31421928 DOI: 10.1016/j.tcb.2019.07.008]
- 179 Pishvaian MJ, Biankin AV, Bailey P, Chang DK, Laheru D, Wolfgang CL, Brody JR. BRCA2 secondary mutation-mediated resistance to platinum and PARP inhibitor-based therapy in pancreatic cancer. Br J Cancer 2017; 116: 1021-1026 [PMID: 28291774 DOI: 10.1038/bjc.2017.40]
- 180 Hsieh HJ, Peng G. Cellular responses to replication stress: Implications in cancer biology and therapy. DNA Repair (Amst) 2017; 49: 9-20 [PMID: 27908669 DOI: 10.1016/j.dnarep.2016.11.002]
- 181 Maréchal A, Zou L. DNA damage sensing by the ATM and ATR kinases. Cold Spring Harb Perspect Biol 2013; 5 [PMID: 24003211 DOI: 10.1101/cshperspect.a012716]
- 182 Ubhi T, Brown GW. Exploiting DNA Replication Stress for Cancer Treatment. Cancer Res 2019; 79: 1730-1739 [PMID: 30967400 DOI: 10.1158/0008-5472.CAN-18-3631]
- 183 Russell R, Perkhofer L, Liebau S, Lin Q, Lechel A, Feld FM, Hessmann E, Gaedcke J, Güthle M, Zenke M, Hartmann D, von Figura G, Weissinger SE, Rudolph KL, Möller P, Lennerz JK, Seufferlein T. Wagner M. Kleger A. Loss of ATM accelerates pancreatic cancer formation and epithelial-mesenchymal transition. Nat Commun 2015; 6: 7677 [PMID: 26220524 DOI: 10.1038/ncomms8677]
- 184 Moding EJ, Lee CL, Castle KD, Oh P, Mao L, Zha S, Min HD, Ma Y, Das S, Kirsch DG. Atm deletion with dual recombinase technology preferentially radiosensitizes tumor endothelium. J Clin Invest 2014; 124: 3325-3338 [PMID: 25036710 DOI: 10.1172/JCI73932]
- Cowell IG, Durkacz BW, Tilby MJ. Sensitization of breast carcinoma cells to ionizing radiation by 185 small molecule inhibitors of DNA-dependent protein kinase and ataxia telangiectsia mutated. Biochem Pharmacol 2005; 71: 13-20 [PMID: 16293233 DOI: 10.1016/j.bcp.2005.09.029]
- 186 Ayars M, Eshleman J, Goggins M. Susceptibility of ATM-deficient pancreatic cancer cells to radiation. Cell Cycle 2017; 16: 991-998 [PMID: 28453388 DOI: 10.1080/15384101.2017.1312236]
- 187 Kondo T, Kanai M, Kou T, Sakuma T, Mochizuki H, Kamada M, Nakatsui M, Uza N, Kodama Y, Masui T, Takaori K, Matsumoto S, Miyake H, Okuno Y, Muto M. Association between homologous recombination repair gene mutations and response to oxaliplatin in pancreatic cancer. Oncotarget 2018; 9: 19817-19825 [PMID: 29731985 DOI: 10.18632/oncotarget.24865]
- 188 Aguirre AJ, Nowak JA, Camarda ND, Moffitt RA, Ghazani AA, Hazar-Rethinam M, Raghavan S, Kim J, Brais LK, Ragon D, Welch MW, Reilly E, McCabe D, Marini L, Anderka K, Helvie K, Oliver N, Babic A, Da Silva A, Nadres B, Van Seventer EE, Shahzade HA, St Pierre JP, Burke KP, Clancy T, Cleary JM, Doyle LA, Jajoo K, McCleary NJ, Meyerhardt JA, Murphy JE, Ng K, Patel AK, Perez K, Rosenthal MH, Rubinson DA, Ryou M, Shapiro GI, Sicinska E, Silverman SG, Nagy RJ, Lanman RB, Knoerzer D, Welsch DJ, Yurgelun MB, Fuchs CS, Garraway LA, Getz G, Hornick JL, Johnson BE, Kulke MH, Mayer RJ, Miller JW, Shyn PB, Tuveson DA, Wagle N, Yeh JJ, Hahn WC, Corcoran RB, Carter SL, Wolpin BM. Real-time Genomic Characterization of Advanced Pancreatic Cancer to Enable Precision Medicine. Cancer Discov 2018; 8: 1096-1111 [PMID: 29903880 DOI: 10.1158/2159-8290.CD-18-0275]
- 189 Roger E, Gout J, Arnold F, Beutel AK, Müller M, Abaei A, Barth TFE, Rasche V, Seufferlein T, Perkhofer L, Kleger A. Maintenance Therapy for ATM-Deficient Pancreatic Cancer by Multiple DNA Damage Response Interferences after Platinum-Based Chemotherapy. Cells 2020; 9 [PMID: 32948057 DOI: 10.3390/cells9092110]
- Fokas E, Prevo R, Pollard JR, Reaper PM, Charlton PA, Cornelissen B, Vallis KA, Hammond EM, 190 Olcina MM, Gillies McKenna W, Muschel RJ, Brunner TB. Targeting ATR in vivo using the novel inhibitor VE-822 results in selective sensitization of pancreatic tumors to radiation. Cell Death Dis 2012; 3: e441 [PMID: 23222511 DOI: 10.1038/cddis.2012.181]
- Wallez Y, Dunlop CR, Johnson TI, Koh SB, Fornari C, Yates JWT, Bernaldo de Quirós Fernández 191 S, Lau A, Richards FM, Jodrell DI. The ATR Inhibitor AZD6738 Synergizes with Gemcitabine In Vitro and In Vivo to Induce Pancreatic Ductal Adenocarcinoma Regression. Mol Cancer Ther 2018;



17: 1670-1682 [PMID: 29891488 DOI: 10.1158/1535-7163.MCT-18-0010]

- 192 Rosenberg SA, Restifo NP. Adoptive cell transfer as personalized immunotherapy for human cancer. Science 2015; 348: 62-68 [PMID: 25838374 DOI: 10.1126/science.aaa4967]
- 193 Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, Drake CG, Camacho LH, Kauh J, Odunsi K, Pitot HC, Hamid O, Bhatia S, Martins R, Eaton K, Chen S, Salay TM, Alaparthy S, Grosso JF, Korman AJ, Parker SM, Agrawal S, Goldberg SM, Pardoll DM, Gupta A, Wigginton JM. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. N Engl J Med 2012; 366: 2455-2465 [PMID: 22658128 DOI: 10.1056/NEJMoa1200694]
- 194 Royal RE, Levy C, Turner K, Mathur A, Hughes M, Kammula US, Sherry RM, Topalian SL, Yang JC, Lowy I, Rosenberg SA. Phase 2 trial of single agent Ipilimumab (anti-CTLA-4) for locally advanced or metastatic pancreatic adenocarcinoma. J Immunother 2010; 33: 828-833 [PMID: 20842054 DOI: 10.1097/CJI.0b013e3181eec14c]
- 195 Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, Skora AD, Luber BS, Azad NS, Laheru D, Biedrzycki B, Donehower RC, Zaheer A, Fisher GA, Crocenzi TS, Lee JJ, Duffy SM, Goldberg RM, de la Chapelle A, Koshiji M, Bhaijee F, Huebner T, Hruban RH, Wood LD, Cuka N, Pardoll DM, Papadopoulos N, Kinzler KW, Zhou S, Cornish TC, Taube JM, Anders RA, Eshleman JR, Vogelstein B, Diaz LA Jr. PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. N Engl J Med 2015; 372: 2509-2520 [PMID: 26028255 DOI: 10.1056/NEJMoa1500596]
- 196 Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SA, Behjati S, Biankin AV, Bignell GR, Bolli N, Borg A, Børresen-Dale AL, Boyault S, Burkhardt B, Butler AP, Caldas C, Davies HR, Desmedt C. Eils R. Evfjörd JE. Foekens JA. Greaves M. Hosoda F. Hutter B. Ilicic T. Imbeaud S. Imielinski M, Jäger N, Jones DT, Jones D, Knappskog S, Kool M, Lakhani SR, López-Otín C, Martin S, Munshi NC, Nakamura H, Northcott PA, Pajic M, Papaemmanuil E, Paradiso A, Pearson JV, Puente XS, Raine K, Ramakrishna M, Richardson AL, Richter J, Rosenstiel P, Schlesner M, Schumacher TN, Span PN, Teague JW, Totoki Y, Tutt AN, Valdés-Mas R, van Buuren MM, van 't Veer L, Vincent-Salomon A, Waddell N, Yates LR; Australian Pancreatic Cancer Genome Initiative; ICGC Breast Cancer Consortium; ICGC MMML-Seq Consortium; ICGC PedBrain, Zucman-Rossi J, Futreal PA, McDermott U, Lichter P, Meyerson M, Grimmond SM, Siebert R, Campo E, Shibata T, Pfister SM, Campbell PJ, Stratton MR. Signatures of mutational processes in human cancer. Nature 2013; 500: 415-421 [PMID: 23945592 DOI: 10.1038/nature12477]
- 197 **Varchoan M.** Hopkins A. Jaffee EM. Tumor Mutational Burden and Response Rate to PD-1 Inhibition. N Engl J Med 2017; 377: 2500-2501 [PMID: 29262275 DOI: 10.1056/NEJMc1713444]
- 198 Humphris JL, Patch AM, Nones K, Bailey PJ, Johns AL, McKay S, Chang DK, Miller DK, Pajic M, Kassahn KS, Quinn MC, Bruxner TJ, Christ AN, Harliwong I, Idrisoglu S, Manning S, Nourse C, Nourbakhsh E, Stone A, Wilson PJ, Anderson M, Fink JL, Holmes O, Kazakoff S, Leonard C, Newell F, Waddell N, Wood S, Mead RS, Xu Q, Wu J, Pinese M, Cowley MJ, Jones MD, Nagrial AM, Chin VT, Chantrill LA, Mawson A, Chou A, Scarlett CJ, Pinho AV, Rooman I, Giry-Laterriere M, Samra JS, Kench JG, Merrett ND, Toon CW, Epari K, Nguyen NQ, Barbour A, Zeps N, Jamieson NB, McKay CJ, Carter CR, Dickson EJ, Graham JS, Duthie F, Oien K, Hair J, Morton JP, Sansom OJ, Grützmann R, Hruban RH, Maitra A, Iacobuzio-Donahue CA, Schulick RD, Wolfgang CL, Morgan RA, Lawlor RT, Rusev B, Corbo V, Salvia R, Cataldo I, Tortora G, Tempero MA; Australian Pancreatic Cancer Genome Initiative, Hofmann O, Eshleman JR, Pilarsky C, Scarpa A, Musgrove EA, Gill AJ, Pearson JV, Grimmond SM, Waddell N, Biankin AV. Hypermutation In Pancreatic Cancer. Gastroenterology 2017; 152: 68-74.e2 [PMID: 27856273 DOI: 10.1053/j.gastro.2016.09.060]
- Tsujikawa T, Kumar S, Borkar RN, Azimi V, Thibault G, Chang YH, Balter A, Kawashima R, Choe G, Sauer D, El Rassi E, Clayburgh DR, Kulesz-Martin MF, Lutz ER, Zheng L, Jaffee EM, Leyshock P, Margolin AA, Mori M, Gray JW, Flint PW, Coussens LM. Quantitative Multiplex Immunohistochemistry Reveals Myeloid-Inflamed Tumor-Immune Complexity Associated with Poor Prognosis. Cell Rep 2017; 19: 203-217 [PMID: 28380359 DOI: 10.1016/j.celrep.2017.03.037]
- 200 Clark CE, Hingorani SR, Mick R, Combs C, Tuveson DA, Vonderheide RH. Dynamics of the immune reaction to pancreatic cancer from inception to invasion. Cancer Res 2007; 67: 9518-9527 [PMID: 17909062 DOI: 10.1158/0008-5472.CAN-07-0175]
- 201 Diaz-Montero CM, Salem ML, Nishimura MI, Garrett-Mayer E, Cole DJ, Montero AJ. Increased circulating myeloid-derived suppressor cells correlate with clinical cancer stage, metastatic tumor burden, and doxorubicin-cyclophosphamide chemotherapy. Cancer Immunol Immunother 2009; 58: 49-59 [PMID: 18446337 DOI: 10.1007/s00262-008-0523-4]
- 202 Siret C, Collignon A, Silvy F, Robert S, Cheyrol T, André P, Rigot V, Iovanna J, van de Pavert S, Lombardo D, Mas E, Martirosyan A. Deciphering the Crosstalk Between Myeloid-Derived Suppressor Cells and Regulatory T Cells in Pancreatic Ductal Adenocarcinoma. Front Immunol 2019; 10: 3070 [PMID: 32038621 DOI: 10.3389/fimmu.2019.03070]
- 203 Stromnes IM, DelGiorno KE, Greenberg PD, Hingorani SR. Stromal reengineering to treat pancreas cancer. Carcinogenesis 2014; 35: 1451-1460 [PMID: 24908682 DOI: 10.1093/carcin/bgu115]
- 204 Blank CU, Haining WN, Held W, Hogan PG, Kallies A, Lugli E, Lynn RC, Philip M, Rao A, Restifo NP, Schietinger A, Schumacher TN, Schwartzberg PL, Sharpe AH, Speiser DE, Wherry EJ, Youngblood BA, Zehn D. Defining 'T cell exhaustion'. Nat Rev Immunol 2019; 19: 665-674 [PMID: 31570879 DOI: 10.1038/s41577-019-0221-9]
- 205 Martinez GJ, Pereira RM, Äijö T, Kim EY, Marangoni F, Pipkin ME, Togher S, Heissmeyer V, Zhang YC, Crotty S, Lamperti ED, Ansel KM, Mempel TR, Lähdesmäki H, Hogan PG, Rao A. The



transcription factor NFAT promotes exhaustion of activated CD8+ T cells. Immunity 2015; 42: 265-278 [PMID: 25680272 DOI: 10.1016/j.immuni.2015.01.006]

- 206 Pauken KE, Sammons MA, Odorizzi PM, Manne S, Godec J, Khan O, Drake AM, Chen Z, Sen DR, Kurachi M, Barnitz RA, Bartman C, Bengsch B, Huang AC, Schenkel JM, Vahedi G, Haining WN, Berger SL, Wherry EJ. Epigenetic stability of exhausted T cells limits durability of reinvigoration by PD-1 blockade. Science 2016; 354: 1160-1165 [PMID: 27789795 DOI: 10.1126/science.aaf2807
- Burrack AL, Spartz EJ, Raynor JF, Wang I, Olson M, Stromnes IM. Combination PD-1 and PD-L1 207 Blockade Promotes Durable Neoantigen-Specific T Cell-Mediated Immunity in Pancreatic Ductal Adenocarcinoma. Cell Rep 2019; 28: 2140-2155.e6 [PMID: 31433988 DOI: 10.1016/j.celrep.2019.07.059
- Stromnes IM, Schmitt TM, Hulbert A, Brockenbrough JS, Nguyen H, Cuevas C, Dotson AM, Tan 208 X, Hotes JL, Greenberg PD, Hingorani SR. T Cells Engineered against a Native Antigen Can Surmount Immunologic and Physical Barriers to Treat Pancreatic Ductal Adenocarcinoma. Cancer Cell 2015; 28: 638-652 [PMID: 26525103 DOI: 10.1016/j.ccell.2015.09.022]
- 209 Burrack AL, Rollins MR, Spartz EJ, Mesojednik TD, Schmiechen ZC, Raynor JF, Wang IX, Kedl RM, Stromnes IM. CD40 Agonist Overcomes T Cell Exhaustion Induced by Chronic Myeloid Cell IL-27 Production in a Pancreatic Cancer Preclinical Model. J Immunol 2021; 206: 1372-1384 [PMID: 33558374 DOI: 10.4049/jimmunol.2000765]
- 210 Ma Y, Li J, Wang H, Chiu Y, Kingsley CV, Fry D, Delaney SN, Wei SC, Zhang J, Maitra A, Yee C. Combination of PD-1 Inhibitor and OX40 Agonist Induces Tumor Rejection and Immune Memory in Mouse Models of Pancreatic Cancer. Gastroenterology 2020; 159: 306-319.e12 [PMID: 32179091 DOI: 10.1053/j.gastro.2020.03.018]
- 211 Mirlekar B, Michaud D, Searcy R, Greene K, Pylayeva-Gupta Y. IL35 Hinders Endogenous Antitumor T-cell Immunity and Responsiveness to Immunotherapy in Pancreatic Cancer. Cancer Immunol Res 2018; 6: 1014-1024 [PMID: 29980536 DOI: 10.1158/2326-6066.CIR-17-0710]
- 212 Panni RZ, Herndon JM, Zuo C, Hegde S, Hogg GD, Knolhoff BL, Breden MA, Li X, Krisnawan VE, Khan SQ, Schwarz JK, Rogers BE, Fields RC, Hawkins WG, Gupta V, DeNardo DG. Agonism of CD11b reprograms innate immunity to sensitize pancreatic cancer to immunotherapies. Sci Transl Med 2019; 11 [PMID: 31270275 DOI: 10.1126/scitranslmed.aau9240]
- 213 Le DT, Picozzi VJ, Ko AH, Wainberg ZA, Kindler H, Wang-Gillam A, Oberstein P, Morse MA, Zeh HJ 3rd, Weekes C, Reid T, Borazanci E, Crocenzi T, LoConte NK, Musher B, Laheru D, Murphy A, Whiting C, Nair N, Enstrom A, Ferber S, Brockstedt DG, Jaffee EM. Results from a Phase IIb, Randomized, Multicenter Study of GVAX Pancreas and CRS-207 Compared with Chemotherapy in Adults with Previously Treated Metastatic Pancreatic Adenocarcinoma (ECLIPSE Study). Clin Cancer Res 2019; 25: 5493-5502 [PMID: 31126960 DOI: 10.1158/1078-0432.CCR-18-2992
- Kinkead HL, Hopkins A, Lutz E, Wu AA, Yarchoan M, Cruz K, Woolman S, Vithayathil T, 214 Glickman LH, Ndubaku CO, McWhirter SM, Dubensky TW Jr, Armstrong TD, Jaffee EM, Zaidi N. Combining STING-based neoantigen-targeted vaccine with checkpoint modulators enhances antitumor immunity in murine pancreatic cancer. JCI Insight 2018; 3 [PMID: 30333318 DOI: 10.1172/jci.insight.122857]
- 215 Nywening TM, Belt BA, Cullinan DR, Panni RZ, Han BJ, Sanford DE, Jacobs RC, Ye J, Patel AA, Gillanders WE, Fields RC, DeNardo DG, Hawkins WG, Goedegebuure P, Linehan DC. Targeting both tumour-associated CXCR2⁺ neutrophils and CCR2⁺ macrophages disrupts myeloid recruitment and improves chemotherapeutic responses in pancreatic ductal adenocarcinoma. Gut 2018; 67: 1112-1123 [PMID: 29196437 DOI: 10.1136/gutjnl-2017-313738]
- 216 Steele CW, Karim SA, Leach JDG, Bailey P, Upstill-Goddard R, Rishi L, Foth M, Bryson S, McDaid K, Wilson Z, Eberlein C, Candido JB, Clarke M, Nixon C, Connelly J, Jamieson N, Carter CR, Balkwill F, Chang DK, Evans TRJ, Strathdee D, Biankin AV, Nibbs RJB, Barry ST, Sansom OJ, Morton JP. CXCR2 Inhibition Profoundly Suppresses Metastases and Augments Immunotherapy in Pancreatic Ductal Adenocarcinoma. Cancer Cell 2016; 29: 832-845 [PMID: 27265504 DOI: 10.1016/j.ccell.2016.04.014]
- 217 Zhu Y, Knolhoff BL, Meyer MA, Nywening TM, West BL, Luo J, Wang-Gillam A, Goedegebuure SP, Linehan DC, DeNardo DG. CSF1/CSF1R blockade reprograms tumor-infiltrating macrophages and improves response to T-cell checkpoint immunotherapy in pancreatic cancer models. Cancer Res 2014; 74: 5057-5069 [PMID: 25082815 DOI: 10.1158/0008-5472.CAN-13-3723]
- 218 Candido JB, Morton JP, Bailey P, Campbell AD, Karim SA, Jamieson T, Lapienyte L, Gopinathan A, Clark W, McGhee EJ, Wang J, Escorcio-Correia M, Zollinger R, Roshani R, Drew L, Rishi L, Arkell R, Evans TRJ, Nixon C, Jodrell DI, Wilkinson RW, Biankin AV, Barry ST, Balkwill FR, Sansom OJ. CSF1R⁺ Macrophages Sustain Pancreatic Tumor Growth through T Cell Suppression and Maintenance of Key Gene Programs that Define the Squamous Subtype. Cell Rep 2018; 23: 1448-1460 [PMID: 29719257 DOI: 10.1016/j.celrep.2018.03.131]
- 219 Stromnes IM, Schmitt TM, Chapuis AG, Hingorani SR, Greenberg PD. Re-adapting T cells for cancer therapy: from mouse models to clinical trials. Immunol Rev 2014; 257: 145-164 [PMID: 24329795 DOI: 10.1111/imr.121411
- 220 Liu J, Zhong JF, Zhang X, Zhang C. Allogeneic CD19-CAR-T cell infusion after allogeneic hematopoietic stem cell transplantation in B cell malignancies. J Hematol Oncol 2017; 10: 35 [PMID: 28143567 DOI: 10.1186/s13045-017-0405-3]



- 221 Ali AI, Oliver AJ, Samiei T, Chan JD, Kershaw MH, Slaney CY. Genetic Redirection of T Cells for the Treatment of Pancreatic Cancer. Front Oncol 2019; 9: 56 [PMID: 30809507 DOI: 10.3389/fonc.2019.00056
- 222 Akce M, Zaidi MY, Waller EK, El-Rayes BF, Lesinski GB. The Potential of CAR T Cell Therapy in Pancreatic Cancer. Front Immunol 2018; 9: 2166 [PMID: 30319627 DOI: 10.3389/fimmu.2018.02166]
- Wu AA, Jaffee E, Lee V. Current Status of Immunotherapies for Treating Pancreatic Cancer. Curr 223 Oncol Rep 2019; 21: 60 [PMID: 31101991 DOI: 10.1007/s11912-019-0811-5]
- 224 Morgan RA, Yang JC, Kitano M, Dudley ME, Laurencot CM, Rosenberg SA. Case report of a serious adverse event following the administration of T cells transduced with a chimeric antigen receptor recognizing ERBB2. Mol Ther 2010; 18: 843-851 [PMID: 20179677 DOI: 10.1038/mt.2010.24]
- 225 Parkhurst MR, Yang JC, Langan RC, Dudley ME, Nathan DA, Feldman SA, Davis JL, Morgan RA, Merino MJ, Sherry RM, Hughes MS, Kammula US, Phan GQ, Lim RM, Wank SA, Restifo NP, Robbins PF, Laurencot CM, Rosenberg SA. T cells targeting carcinoembryonic antigen can mediate regression of metastatic colorectal cancer but induce severe transient colitis. Mol Ther 2011; 19: 620-626 [PMID: 21157437 DOI: 10.1038/mt.2010.272]
- Thistlethwaite FC, Gilham DE, Guest RD, Rothwell DG, Pillai M, Burt DJ, Byatte AJ, Kirillova N, 226 Valle JW, Sharma SK, Chester KA, Westwood NB, Halford SER, Nabarro S, Wan S, Austin E, Hawkins RE. The clinical efficacy of first-generation carcinoembryonic antigen (CEACAM5)specific CAR T cells is limited by poor persistence and transient pre-conditioning-dependent respiratory toxicity. Cancer Immunol Immunother 2017; 66: 1425-1436 [PMID: 28660319 DOI: 10.1007/s00262-017-2034-7]
- 227 Beatty GL, O'Hara MH, Lacey SF, Torigian DA, Nazimuddin F, Chen F, Kulikovskaya IM, Soulen MC, McGarvey M, Nelson AM, Gladney WL, Levine BL, Melenhorst JJ, Plesa G, June CH. Activity of Mesothelin-Specific Chimeric Antigen Receptor T Cells Against Pancreatic Carcinoma Metastases in a Phase 1 Trial. Gastroenterology 2018; 155: 29-32 [PMID: 29567081 DOI: 10.1053/j.gastro.2018.03.029]
- 228 You F, Jiang L, Zhang B, Lu Q, Zhou Q, Liao X, Wu H, Du K, Zhu Y, Meng H, Gong Z, Zong Y, Huang L, Lu M, Tang J, Li Y, Zhai X, Wang X, Ye S, Chen D, Yuan L, Qi L, Yang L. Phase 1 clinical trial demonstrated that MUC1 positive metastatic seminal vesicle cancer can be effectively eradicated by modified Anti-MUC1 chimeric antigen receptor transduced T cells. Sci China Life Sci 2016; 59: 386-397 [PMID: 26961900 DOI: 10.1007/s11427-016-5024-7]



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REVIEW

Evaluation of botanicals as potential COVID-19 symptoms terminator

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Abstract

Information about the coronavirus disease 2019 (COVID-19) pandemic is still evolving since its appearance in December 2019 and has affected the whole world. Particularly, a search for an effective and safe treatment for COVID-19 continues. Botanical mixtures contain secondary metabolites (such as flavonoids, phenolics, alkaloids, essential oils etc.) with many therapeutic effects. In this study, the use of herbal treatments against COVID-19 was evaluated. Medical synthetic drugs focus mainly on respiratory symptoms, however herbal therapy with plant extracts may be useful to relieve overall symptoms of COVID-19 due to the variety of bioactive ingredients. Since COVID-19 is a virus that affects the respiratory tract, the antiviral effects of botanicals/plants against respiratory viruses have been examined through clinical studies. Data about COVID-19 patients revealed that the virus not only affects the respiratory system but different organs including the gastrointestinal (GI) system. As GI symptoms seriously affect quality of life, herbal options that might eliminate these problems were also evaluated. Finally, computer modeling studies of plants and their active compounds on COVID-19 were included. In summary, herbal therapies were identified as potential options for both antiviral effects and control of COVID-19 symptoms. Further data will be needed to enlighten all aspects of COVID-19 pathogenesis, before determining the effects of plants on severe acute respiratory syndrome coronavirus 2.

Key Words: COVID-19; Herbal therapies; Plant; SARS-CoV-2; Antiviral; Symptom

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Core Tip: To stop the coronavirus disease 2019 (COVID-19) pandemic, extensive search is ongoing to develop effective and safe drugs against severe acute respiratory syndrome coronavirus 2. COVID-19 in a major way affects the respiratory system, but many patients also have gastrointestinal (GI) symptoms. Plants have beneficial effects



Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

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on various systems with their varied array of metabolites. In our study, the potential effects of herbal treatments against COVID-19 were examined. Their antiviral effects, their effects on the respiratory system, GI system, and other COVID-19 symptoms were investigated.

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INTRODUCTION

New coronavirus disease 2019 (COVID-19), which emerged in Wuhan in December 2019, spread rapidly and affected the whole world. The emergence, epidemiology, origin and evolution of COVID-19 has been extensively studied by Sun et al[1].

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has been shown to carry out viral replication in the human host mainly through three main proteins and enzymes: 3-chymotrypsin-like protease (3CLpro), angiotensin-converting enzyme-2 (ACE2) and spike protein (TMPRSS2)[2,3]. ACE2 receptors are found in the body not only in the lungs but also in tissues such as the endothelium, heart, kidney and intestine^[2]. This distribution makes many organs a target of COVID-19. The significance of ACE2, which is found in intestinal tissues, especially for amino acid uptake from foods, has been emphasized and it has been suggested that the intestine may be an important entry site for SARS-CoV-2[2-4] Azithromycin, chloroquine, lopinavir, remdesivir, ritonavir are options used in treatment and whose effects are evaluated^[5]. Effective and safe drugs and vaccines are sought all over the world to prevent novel coronavirus. Minerals, herbs, herbal products, probiotics and vitamins are the main natural resources, whose effectiveness and also the usability of herbal medicines in COVID-19 were investigated and benefit, risk assessments were evaluated [6-8]. Truly, since the beginning of the COVID-19, herbal medicines have been used in China. A study has shown that 90% of the 214 patients were treated with the traditional herbal medicine, moreover, it is reported that some of them prevented COVID-19 infection in healthy individuals and enhanced the health state of patients with mild or severe symptoms[9,10]. Health scientists from the Zhongnan Hospital of Wuhan University included the use of traditional medicines in the guidelines for the treatment and prevention of COVID-19. The experts recommended using medicinal plants for the prevention of COVID-19, additionally, the use of different herbal mixtures were recommended according to the disease-stage[11].

Herbs and herbal products provide generous sources of primary and mostly secondary metabolites, which are valuable compounds (phenolics, flavonoids, tannins, alkaloids, essential oils, etc.) for prophylactic and chronical therapeutic purposes. Some of these metabolites in herbs and herbal mixtures have high chemical variety than the synthetics in stopping viral proliferations, and having antiviral activities[12]. Thus, botanicals can both show antiviral effects and relieve the symptoms of COVID-19 thanks to the different substance groups, which demonstrate different biological effects that will not be possible to achieve with a single synthetic drug. Based on this understanding, in this review, we offer all the potential interventions for COVID-19 infection according to previous and recently found antiviral effects of herbals. Considering the major transmission routes of COVID-19, where mostly ACE2 receptors found and the symptoms, the plants have been handled especially with their effects on the mostly respiratory and also gastrointestinal (GI) systems. Although ACE2, is typically expressed in epithelial cells of the airways, various GI symptoms in COVID-19 might be explained by the high expression of ACE2 in the digestive tract. Additionally, liver tests abnormalities, active viral replication in GI tract and patients' manifestations with GI symptoms (abdominal pain, diarrhea, nausea, vomiting) and possible fecal-oral transmission reveal the GI involvement in COVID-19[13].

Recent findings demonstrated that early blocking of COVID-19 with ACE2 inhibitors was one of the mechanisms used by novel drugs[14], on the other hand diabetes mellitus and hypertension enhanced the risk of COVID-19 infection, in spite of using ACE2 inhibitors[15-17]. Furthermore, unpredicted ACE2 upregulation by



ACE2 inhibitors, ibuprofen and angiotensin II type-I receptor blockers lead to need of identifying/using alternative ACE2 blockers[18]. Consequently, botanicals or natural products might be alternatively and selectively might block the ACE2 receptors without inhibiting the enzyme activity in order to treat and/or prevent COVID-19 spread in humans without increasing ACE2 expression in patients and therefore increased risk for COVID-19[19].

Clinical human studies showing the effect of plants on respiratory infections are presented as a table. Based on the pharmacological properties of plants, their practicality on COVID-19 symptoms have been evaluated. In the last part of the article, plants that inhibit ACE receptors, the research studies and their active compounds on COVID-19 also included and it is aimed to examine the plants from a broad perspective.

ANTIVIRAL EFFECTS OF HERBAL THERAPIES

Most of the respiratory diseases (approximately 80%) are caused by viral agents[20]. Viral respiratory diseases are responsible for high mortality and morbidity, especially in disadvantaged and sensitive elderlies and immunocompromised individuals[21, 22]. The main respiratory viruses are adenovirus, coronavirus, influenza virus, respiratory syncytial virus and rhinovirus^[20]. Plants with antiviral effects and studies showing the effects of these on respiratory viruses are given in Table 1. Human clinical studies showing the effects of plants on respiratory tract infections are presented in Table 2.

EFFECTS OF HERBAL TREATMENT ON COVID-19 SYMPTOMS

Cough and fever are common symptoms in patients with COVID-19, including fatigue, shortness of breath, headache, muscle pain, sore throat, sputum, hemoptysis, diarrhea, dyspnea, rhinorrhea, chest pain, nausea, and vomiting[23]. COVID-19 symptoms in children are similar to those in adults and are relatively mild^[24].

Although, the current synthetic drugs focus on mainly respiratory symptoms, herbal therapy can be used to relieve overall symptoms of COVID-19 with their bioactive ingredients^[25]. The meta-analysis study, which included randomized controlled trial studies, found significant effects of the combination of western medicine and herbal therapies. Combined treatment has been effective in cough, fever, dry and sore throat, fatigue and overall GI symptoms. The combined therapy significantly improved the disappearance rate of cough and sputum production[26]. In another meta-analysis, it was found that the addition of Chinese herbal medicine for standard care improved the symptoms and signs of COVID-19 as well as decreased levels of C-reactive protein[27]. The effects of plants that can alleviate the symptoms of COVID-19 are summarized in Table 3. In addition, plants regarded as ACE inhibitors are shown in Table 4.

THE EFFECTS OF HERBS AND THEIR ACTIVE COMPOUNDS ON COVID-19

In recent years, artificial intelligence has often been used to discover natural products as medicine^[28,29]. After the outbreak of COVID-19, computer models were used to investigate the effect of many plants and their components on SARS-CoV-2. Khaerunnisa et al[30], determined the COVID-19 Main Protease (Mpro) inhibitor effects of medicinal plant components in a molecular docking study. They suggested apigenin-7-glucoside, curcumin, catechin, demethoxycurcumin, epicatechin-gallate, luteolin-7-glucoside, and oleuropein, as potential inhibitors of COVID-19 Mpro. In a similar molecular docking study using sixty-seven molecules of natural origin, crocin, digitoxigenin and b-eudesmol were proposed as inhibitors against coronavirus[31]. Another study was carried out using one hundred seventy-one essential oil components. The study determined the best docking ligands for the SARS-CoV target proteins were (E)--farnesene, (E,E)--farnesene and (E,E)-farnesol, thereby suggesting essential oil components may act synergistically with other antiviral agents, or they may provide some relief of COVID-19 symptoms[32]. Computer modeling studies and clinical studies against SARS-CoV-2 in some prominent plants/products and their



Plant name	Preparation	Susceptible viruses	Ref.
Allium sativum (Garlic)	Aqueous extracts	Influenza A (H9N2)	Rasool <i>et al</i> [53], 2017
	Extract	Infectious bronchitis virus	Mohajer Shojai <i>et al</i> [<mark>54</mark>], 2016
	Ethanolic extract	Influenza A (H1N1)	Chavan <i>et al</i> [55], 2016
	Garlic oil	Influenza A (H1N1)	Choi[<mark>56</mark>], 2018
	Fresh extract	Influenza A (H1N1)	Mehrbod <i>et al</i> [57], 2013
	Aqueous extract	Adenovirus (ADV3 and ADV41)	Chen <i>et al</i> [58], 2011
Aloe vera (Aloe)	Aloe anthraquinones and several derivatives (3-O- tetraacetoglupiranosil)	Influenza A	Borges-Argáez <i>et al</i> [<mark>59</mark>], 2019
	Aloe-emodin	Influenza A	Li et al[<mark>60</mark>], 2014
Astragalus mongholicus Astragalus)	Astragalus polysaccharides	Avian infectious bronchitis virus	Zhang <i>et al</i> [61], 2018
	Astragalus polysaccharide	Influenza A (H9N2)	Kallon <i>et al</i> [62], 2013
Camellia sinensis (Green tea)	Catechins -EGCG	Adenovirus	Weber <i>et al</i> [63], 2003
	Catechin	Influenza A	Kuzuhara <i>et al</i> [<mark>64</mark>], 2009
	Catechins	Influenza A (H5N1)	Liu et al[65], 2012
	Polyphenols	Influenza A; Influenza B	Yang et al[66], 2014
Curcuma longa (Turmeric)	Curcumin	Influenza A virus	Chen <i>et al</i> [67], 2013
			Dai et al[68], 2018
	Curcumin	Influenza A (H1N1, H6N1)	Chen <i>et al</i> [69], 2010
	Curcumin	RSV	Obata et al[70], 2013
Echinacea purpurea (Purple coneflower)	<i>E. purpurea</i> fresh herb and root tinctures	Influenza	Vimalanathan <i>et al</i> [<mark>71</mark>], 2013
	Standardized E. purpurea extract	Influenza A (H5N1, H7N7, H1N1)	Pleschka <i>et al</i> [72] , 2009
	Standardized E. purpurea extract	Rhinoviruses, RSV	Hudson <i>et al</i> [73], 2011
Eucalyptus globulus (Eucalyptus)	Essential oil- vapor phase	Influenza	Vimalanathan <i>et al</i> [74], 2014
Ginkgo biloba (Ginkgo)	Leaf extract	Influenza A (H1N1, H3N2)	Haruyama et al[75], 2013
<i>Glycyrrhiza</i> sp. (Licorice)	Water extract of licorice (Glycyrrhiza uralensis)	RSV	Feng Yeh et al[76], 2013
	Glycyrrhizic acid derivatives	SARS-CoV	Hoever <i>et al</i> [77], 2005
	Extract of Glycyrrhiza inflata	Influenza A (H1N1)	Dao et al[78], 2011
	Glycyrrhizin	Influenza A	Wolkerstorfer <i>et al</i> [79], 20
	Glycyrrhizin	Influenza A (H5N1)	Michaelis <i>et al</i> [80], 2010
epidium meyenii (Maca)	Extracted with methanol	Influenza A; Influenza B	Del Valle Mendoza <i>et al</i> [<mark>81]</mark> , 2014
Melaleuca alternifolia (Tea tree)	Tea tree oil	Influenza A (H1N1)	Garozzo <i>et al</i> [<mark>82</mark>], 2011
	Aerosol and vapor of tea tree oil	Influenza A (H11N9)	Usachev <i>et al</i> [83], 2013
	Tea tree oil	Influenza A (H11N9)	Pyankov <i>et al</i> [<mark>84</mark>], 2012
Melissa officinalis (Lemon balm)	Essential oil	Influenza A (H9N2)	Pourghanbari et al[<mark>85</mark>], 20
	Extract	Avian infectious bronchitis	Lelešius <i>et al</i> [<mark>86</mark>], 2019
Mentha piperita (Peppermint)	Ethanol extract	RSV	Li et al[<mark>87]</mark> , 2017
	Extract	Avian infectious bronchitis	Lelešius <i>et al</i> [86], 2019



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Nigella sativa (Black cumin)	Ethanol extracts of	Influenza A (H5N1)	Dorra <i>et al</i> [88], 2019
	Ethanol extracts of	Influenza A (H9N2)	Umar <i>et al</i> [<mark>89</mark>], 2016
	Extract	Coronavirus	Ulasli <i>et al</i> [90], 2014
Panax ginseng (Ginseng)	Root of plant Panax ginseng	RSV	Lee <i>et al</i> [91], 2014
	Panax Korean red ginseng extract	RSV	Lee <i>et al</i> [92], 2014
	Red ginseng extract and polysaccharide and saponin fractions	Influenza A (H1N1)	Yin et al[93], 2013
	Korean red ginseng extract	Influenza A (H1N1, H3N2)	Yoo et al[<mark>94</mark>], 2012
Pelargonium sidoides	Pelargonium sidoides radix extract EPs®7630	Rhinovirus	Roth <i>et al</i> [95], 2019
(Pelargonium)	EPs [®] 7630	Respiratory viruses	Michaelis <i>et al</i> [96], 2011
	EPs [®] 7630	Influenza A (H1N1, H3N2)	Theisen <i>et al</i> [97], 2012
Sambucus nigra (Black elder)	Extract	Infectious bronchitis virus	Chen <i>et al</i> [98], 2014
	Standardized elderberry liquid extract	Influenza A; Influenza B	Krawitz et al <mark>[99]</mark> , 2011
	Concentrated juice of elderberry	Influenza A	Kinoshita <i>et al</i> [100], 2012
	Elderberry flavonoids	Influenza A (H1N1)	Roschek et al[101], 2009
Scutellaria baicalensis (Chinese	Chemical constituents	Influenza A (H1N1)	Ji <i>et al</i> [102], 2015
skullcap)	Baicalin	SARS-CoV	Chen <i>et al</i> [103], 2004
<i>Torreya nucifera</i> (Japanese nutmeg yew)	Ethanol extract	SARS-CoV	Ryu et al[104], 2010
Thymus vulgaris (Thyme)	Essential oil- liquid phase	Influenza	Vimalanathan <i>et al</i> [74], 2014
	Extract	Avian infectious bronchitis	Lelešius <i>et al</i> [<mark>86</mark>], 2019
Withania somnifera (Ashwagandha)	Withaferin A	Influenza A (H1N1)	Cai et al[105], 2015
Zingiber officinalis (Ginger)	Aqueous extracts	Influenza A (H9N2)	Rasool <i>et al</i> [53], 2017
	Ethanol extracts	Influenza A- (H5N1)	Dorra <i>et al</i> [<mark>88</mark>], 2019
	Fresh ginger	RSV	Chang et al[106], 2013

Influenza A strains: H1N1, H3N2, H5N1, H6N1, H7N7, H9N2, H11N9; RSV: Respiratory syncytial virus; H1N1: Influenza A; SARS-CoV: Severe acute respiratory syndrome coronavirus.

metabolites are given below.

Curcuma longa

Utomo and Meiyanto[33] revealed the potential of several compounds of Curcuma longa against SARS-CoV-2 by binding to three protein receptors (RBD-S, PD-ACE2, SARS-CoV-2 protease). They showed that Curcuma sp. compounds can bind to target receptors, thus, have potential inhibitory effects on SARS-CoV-2 infectivity. Rajagopal et al[34] showed in their in silico docking study that Curcuma longa components could be effective against COVID-19 by inhibiting the SARS-CoV-2 Mpro enzyme. Morever, cyclocurcumin and curcumin possess significant binding at the active site of SARS-CoV-2 Mpro when compared to hydroxychloroquine and nelfinavir. When compared to remdesivir, cyclocurcumin is significantly more active [Glide score: Cyclocurcumin (-6.77); remdesivir (-6.38); curcumin (-6.13); nelfinavir (-5.93); hydroxychloroquine (- 5.47)]. In a similar study, diacetylcurcuminin was more effective on COVID-19 (Mpro) than nelfinavir[35]. Another study suggested the use of curcumin with hydroxychloroquine to destabilize the SARS-CoV2 receptor proteins[36]. Gonzalez-Paz et al[37] showed that curcumin strongly binds to 3CL-protease of COVID-19 Curcumin caused enzyme folding and structural changes in viral protease. Moreover, curcumin bound more strongly to the enzyme than chloroquine.

Eucalyptus globulus

Sharma[38] suggested that eucalyptus essential oil active compounds are potential inhibitors of COVID-19 Mpro. They conducted a molecular docking study to evaluate



Table 2 Huma	an clinical studies she	owing the effect of plants on re	spiratory infections		
Plant	Disease state	Participant	Dosage	Study design	Results
Aged garlic extract[107]	Cold andflu illness	120 healthy subjects, 2 groups (21-50 yr)	4 capsules/d (2.56 g); 90 d	Double-blind, randomized, placebo- controlled parallel intervention	Increase in γδ-T cell and NK cell. Reduction in cold and flu severity; decrease in symptom days
E. purpurea and E. angustifolia root[108]	New-onset common cold	719 patients, 4 parallel groups (12-80 yr)	First 24 h: Equivalent of 10.2 g of root. Next 4 d: 5.1 g	Randomized, controlled trial	Disease duration and severity are not statistically significantly changed
Echinacea purpurea alcohol extract (Echinaforce [®]) [109]	Common cold	755 healthy subjects, 2 groups (≥ 18 yr)	Illness prevention: 3 × 0.9 mL. Acute stages of colds: 5 × 0.9 mL	Randomized, double-blind, placebo- controlled trial	Reduction of the total number of cold episodes, cumulated episode days, and pain-killer medicated episodes. Inhibited virally confirmed colds and especially prevented enveloped virus infections. Maximal effects on recurrent infections. Prophylactic intake of <i>E. purpurea</i> over a period of 4 mo to provide a positive risk/benefit ratio
<i>Echinacea</i> root extract[110]	Respiratory symptoms	175 adults, 2 groups (18–65 yr)	Tablets: 112.5 mg <i>E.</i> <i>purpurea</i> 6:1 extract (equivalent to 675 mg dry root) and 150 mg <i>E. angustifolia</i> 4:1 extract (equivalent to 600 mg dry root) 3 × 1 tablet, if required: 3 × 2 tablets	Randomized, double blind, placebo- controlled trial	Lower respiratory symptom scores. Preventive effect against the development of respiratory symptoms during travel, including long-haul flights
Green tea catechins and theanine[111]	Influenza	200 healthcare workers, 2 groups	Capsules: Green tea catechins (378 mg/d) and theanine (210 mg/d). 5 m	Randomized, double-blind, placebo- controlled trial	Lower incidence of influenza infection in the catechin/ theanine group
Ivy leaf extract[112]	Acute or chronic bronchial inflammatory disease	9657 patients (5181 children)	Ivy leaves extract [drug-to-extract ratio: 5-7.5:1; extraction solvent: ethanol 30% (w/w)]. 0–5 yr: 3 × 2.5 mL; 6–12 yr: 3 × 5 mL; 12 yr and adults: 3 × 5–7.5 mL. 7 d	Prospective, open, multicenter post marketing study	Healing or improvement in 95% of symptoms. Effective and well tolerated
Ivy extract (Hedelix [®]) [113]	Acute respiratory catarrh and/or chronic recidivating inflammatory bronchial disease	268 children, 2 groups (syrup and drops groups) (0-12 yr)	0-1 yr: 1 × 2.5 mL syrup or 3 × 5 drops, 1- 4 yr: 3 × 2.5 mL syrup or 3 × 16 drops, 4-10 yr: 4 × 2.5 mL syrup or 3 × 21 drops, 10-12 yr: 3 × 5 mL syrup or 3 × 31 drops. 14 d	Independent open, non- interventional studies	Effective and safe treatment of cough. Reduction in symptoms (especially rhinitis, cough and viscous mucus)
Ivy leaves dry extract (Prospan [®]) [114]	Bronchial asthma	30 children (suffering from partial or uncontrolled mild persistent allergic asthma despite long-term treatment with 400 µg budesonide equivalent), 2 groups (6–11 yr)	2 × 5 mL (corresponding to 70 mg extract) 28–30 d	Randomized, double blind, placebo- controlled, cross- over study	Improvement of MEF75-25, MEF25 and VC
Korean red ginseng extract[115]	Influenza-like illness	100 healthy adults, 2 groups (30- 70 yr)	9 capsules/d. 3 m	Placebo- controlled trial	Reduced the incidence of influenza-like illness
Modified ginseng extracts (GS- 3K8 and GINST)[116]	Acute respiratory illness	45 healthy applicants, 3 groups (39-65 yr)	Capsules: 500 mg; 6 capsules/d; 8 wk	Randomized, double-blind, placebo- controlled pilot study	Reduction in acute respiratory illness development and symptom duration
Panax quinquefolius extract CVT- E002[117]	Acute respiratory illness and Chronic Lymphocytic Leukemia	293 patients, 2 groups (≥ 18 yr)	2 × 200 mg extract. 3 m	Randomized, double-blind, placebo- controlled study	Reduction intense acute respiratory illness and moderately-severe sore throat. Increased antibody responses.



Panax ginseng [118]	Chronic obstructive pulmonary disease	14 participants, 2 groups (57–73 yr)	2 × 200 mg 4 wk	Clinical trial protocol and pilot study	One participant in <i>P. ginseng</i> group reported events of sore throat, cough and fever
Panax ginseng root extract [119]	Chronic obstructive pulmonary disease	168 participants, 2 groups	2 × 100 mg capsules. 24 wk	Randomized, multi-center, double-blind, placebo controlled	Reduction in symptoms
Pelargonium sidoides extract EPs [®] 7630 [120]	Chronic obstructive pulmonary disease	199 adults, 2 groups (18 yr and older)	30 drops. 24 wk	Randomized, double-blind, placebo- controlled, parallel group trial	Improvement in HRQoL (health- related quality-of-life) and PRO (Patient-reported outcomes)
Pelargonium sidoides extract EPs [®] 7630 [121]	Acute bronchitis	220 patients (1-18 yr)	1-6 yr: 3 × 10 drops; 6-12 yr: 3 × 20 drops; 12-18 yr: 3 × 30 drops; 7 d	Randomized, double-blind, placebo- controlled clinical trial	Reduction in the total score of bronchitis-specific symptoms (especially cough and rales at auscultation)
Pelargonium sidoides extract EPs [®] 7630 [122]	Upper respiratory tract infections	28 children with a diagnosed transient hypogammaglobulinemia of infancy (1-5 yr)	3 × 10 drops; 7 d	Randomized, placebo controlled, prospective, monocentric pilot study	Increased appetite. Reduction of nasal congestion
Pelargonium sidoides root extract EPs® 7630[123]	Upper respiratory tract- asthma attacks	61 children (1–14 yr)	1–5 yr: 3 × 10 drops; 6–12 yr: 3 × 20 drops; 12 yr and above: 3 × 30 drops; 5 d	Randomized, placebo controlled	Reduction the severity of symptoms (especially cough and nasal congestion). Shortening of the duration of upper respiratory viral infections. Reduction asthma attack frequency
Pelargonium sidoides preparation EPs [®] 7630 [124]	Acute non- streptococcal tonsillopharyngitis	126 children, 2 groups (6-10 yr)	3 × 20 drops. 6 d	Double-blind, placebo- controlled clinical trial	Decrease in tonsillitis severity score compared to placebo in the EPs [®] 7630 group after 4 d of treatment
Pelargonium sidoides extract EPs® 7630 [125]	Common cold	207 adults (18-55 yr)	SD: 3 × 30 drops; HD: 3 × 60 drops; 10 d	Prospective, double-blind, parallel-group, placebo- controlled, phase 3, 2 parts, 2-arm, clinical trial	After 10 d, clinical treatment in 90.4% of the active drug group. Reduction the severity of symptoms and short the duration of the disease. Higher full recovery rates or greater recovery for HD treatment on day 5
Sambucus nigra extract [126]	Influenza	64 patients (16-60 yr)	Lozenge: 175 mg extract; 4 lozenges/d; 2 d	Randomized, double-blind, placebo- controlled, pilot clinical trials	Significant improvement in most symptoms within 24 h (fever, headache, muscle aches and nasal congestion). Significant improvement in all investigated symptoms within 48 h (cough and mucus discharge)
Sambucus nigra extract [127]	Respiratory health	312 adults, 2 groups	Capsules: 300 mg. Before travel: 2 capsules/d. During travel and after arrival: 3 capsules/d. 14 d	Randomized, double-blind placebo- controlled clinical trial	Reduction of cold duration and severity in air travelers. Low symptom score

SD: Standard dose; HD: High dose.

the effect of eucalyptol (1.8 cineol), which is a component of eucalyptus essential oil, on Mpro. They showed that eucalyptol/Mpro complexes produce hydrophobic interactions, strong ionic interactions, hydrogen bond interactions, and eucalyptol may be a potential inhibitor of COVID-19 Mpro. Similarly, M pro/3CL pro/eucalyptol complexes have been shown to form hydrophobic interactions[39]. In another study, Sharma and Kaur[40] suggested jensenone, the component of eucalyptus essential oil, as a potential COVID-19 Mpro inhibitor. In a molecular docking study of 12 active ingredients of eucalyptus essential oil, all of these ingredients were found to bind effectively to the COVID-19 S-protein. Especially the toruatone component was effectively bound and the Spike (S) protein/Toruatone complexes formed hydrogen and hydrophobic interactions[41]. Muhammad et al[42], in a study of the molecular

Table 3 Plants that can have an impact on coronavirus disease 2019 symptoms

Plant name	Effects	Ref.
Allium sativum (Garlic)	Analgesic	Dehghani <i>et al</i> [<mark>128</mark>], 2018
	Anti-inflammatory	Arreola <i>et al</i> [129], 2015
	Anti-platelet	Hiyasat <i>et al</i> [130], 2009
	Heart protection	Sultana <i>et al</i> [131], 2016
	Hepatic protection	Aprioku <i>et al</i> [132], 2017
	Improving GI function	Chen <i>et al</i> [133], 2018
	Renal protection	Seckiner <i>et al</i> [134], 2014
Curcuma longa (Turmeric)	Analgesic	Henrotin <i>et al</i> [135], 2020
		Eke-Okoro <i>et al</i> [136], 2018
	Antiemetic	Liu <i>et al</i> [137] , 2018
	Antifatigue	Huang <i>et al</i> [138], 2015
	Anti-inflammatory	Shimizu <i>et al</i> [139], 2019
	Antifibrotic	Gouda <i>et al</i> [140], 2019
	Antipyretic	Haider <i>et al</i> [141], 2013
	Bronchodilator	Ram <i>et al</i> [142], 2003
	GI protection	Haider <i>et al</i> [141], 2013
		Dulbecco and Savarino[143], 2013
	Hepatic protection	Dulbecco and Savarino[143], 2013
Glycyrrhiza glabra (Licorice)	Antitussives	Nosalova <i>et al</i> [144], 2013
		Kuang <i>et al</i> [145], 2018
	Anti-inflammatory	Kao <i>et al</i> [<mark>146</mark>], 2010
	Respiratory system protection	Shi <i>et al</i> [147], 2011
Nigella sativa (Black cumin)	Analgesic	Rushmi <i>et al</i> [148], 2017
	Anticoagulant	Muralidharan-Chari et al[149], 2016
	Antihistaminic	Ansari <i>et al</i> [150], 2010
		Alsamarai <i>et al</i> [151], 2014
	Anti-inflammatory	Majdalawieh and Fayyad[152], 2015
		Mahdavi <i>et al</i> [153], 2016
	Bronchodilation	Boskabady <i>et al</i> [154], 2010
		Salem <i>et al</i> [155], 2017
Panax ginseng (Ginseng)	Adaptogenic	Ratan <i>et al</i> [156], 2021
Pelargonium sidoides (Pelargonium)	Antitussives	Bao <i>et al</i> [157], 2015
	Secretolytic activity	Bao <i>et al</i> [157], 2015
Scutellaria baicalensis (Chinese skullcap)	Antiemetic	Aung <i>et al</i> [158], 2005
	Anti-inflammatory	Hong <i>et al</i> [159], 2013
	GI protection	Mehendale <i>et al</i> [160], 2007
		Cui et al[161], 2021
	Hepatic protection	Thanh <i>et al</i> [162], 2015
	Neuroprotective	Dai <i>et al</i> [163], 2013
	Regulation of histamine release-Anti allergic	Bui <i>et al</i> [164], 2017
Thymus vulgaris (Thyme)	Analgesic	Laub[<mark>165</mark>], 2018



		Salmalian et al[166], 2014
	Anticoagulant	Okazaki <i>et al</i> [167], 2002
	Anti-inflammatory	Habashy <i>et al</i> [168], 2018
Withania somnifera (Ashwagandha)	Adaptogenic	Salve <i>et al</i> [169], 2019
	Analgesic	Murthy <i>et al</i> [170], 2019
	Anticoagulant, antithrombotic	Ku et al <mark>[171]</mark> , 2014
	Anti-inflammatory	Gupta and Singh[172], 2014
	Antitussives	Nosalova <i>et al</i> [<mark>144]</mark> , 2013
	Stress-relieving	Lopresti <i>et al</i> [173], 2019
Zingiber officinale (Ginger)	Analgesic	Maghbooli <i>et al</i> [174], 2014
		Bartels <i>et al</i> [175], 2015
	Antiemetic	Tóth <i>et al</i> [<mark>176</mark>], 2018
	Anti-inflammatory	Khan <i>et al</i> [177], 2015
	Antiplatelet, antithrombotic	Lee et al[178], 2017
	Antitussives	Bera <i>et al</i> [179], 2016
	GI protection	Nanjundaiah <i>et al</i> [<mark>180</mark>], 2011
	Hepatic protection	Ajith <i>et al</i> [181], 2007
	Nephroprotective	Ajith <i>et al</i> [182], 2007

insertion of eucalyptus active ingredients into Mpro, showed that the a-gurjune of eucalyptus, aromadene and allo-aromadene components have strong binding energy.

Glycyrrhiza glabra

Sinha et al[43] conducted molecular docking simulation studies of two antiviral drugs (lopinavir and ribavirin) and 20 compounds of Glycyrrhiza glabra. Two protein targets from COVID-19 have been identified: Non-structural protein-15 endoribonuclease and spike glycoprotein. Glycyrrhizic acid prevented the virus from entering the host cell, due to its bulky structure. Gliasperin A showed high affinity to Nsp15 endoribonuclease and inhibited its activity. The authors suggested that glycyrrhizic acid disrupts the connection of the virus with the ACE2 receptor at the input level, and Gliasperin A inhibits the replication process of the virus after it enters the host cell. Another study showed that glycyrrhizin can be highly bound to Mpro[44].

Scutellaria baicalensis

Liu et al^[45] investigated the in vitro effect of Scutellaria baicalensis and its components on COVID-19. Baicalein (its main ingredient) and the ethanol extract of the plant inhibited the 3CLpro activity and replication of COVID-19. The ethanol extract also inhibited viral entry. Udrea et al[46] suggested the benefit Scutellaria baicalensis flavones (especially baicalein) against respiratory damage caused by COVID-19. Flavones bound to 3CLpro. strongly bound to wogonin flavone, nitric oxide synthase and cyclooxygenase 2. In addition, norwogonin and baicalein arachidonate modulated 15-lipoxygenase and lysine-specific demethylase 4D analogue.

Thymus vulgaris

In a randomized clinical study conducted on patients suffering from COVID-19, it was found that *Thymus vulgaris* strengthens the immune system and can be used to reduce COVID-19 symptoms. In the study, 83 COVID-19 patients were randomly divided into the control group and the group receiving thyme (TRG). TRG was given as thyme essential oil three times a day for seven days. A questionnaire asking about symptoms such as fever, cough, fatigue, and loss of appetite was completed before and at the end of treatment to determine the effect of thyme on symptoms. Thyme essential oil significantly reduced the severity of symptoms such as fever, cough, shortness of breath, dizziness, muscle pain, anorexia, weakness and lethargy and fatigue. Additionally, thyme increased lymphocyte count and calcium while decreasing blood urea nitrogen and neutrophil count^[47]. Carvacrol, a component of thyme, has been



Plants	The compound under study	Results	Ref.
Ammoides verticillata essential oil	Isothymol	SARS-CoV-2/ACE2 inhibition	Abdelli <i>et al</i> [<mark>183</mark>], 2021
Allium sativum essential oil	Organosulfur compounds (99.4% of its essential oil SARS-CoV-2/ACE2 inhibition. Garlic essential oil can prevent protein maturation of the virus and the spread of infection		Thuy <i>et al</i> [<mark>184</mark>], 2020
Apium graveolens	Apigenin	Kidneys of spontaneous hypertensive rats/Regulation in ACE2 expression	Sui et al[185], 2010
Camellia sinensis	Black tea; Dark tea; Green tea; Oolong tea; White tea	ACE inhibition: Green < oolong < white < black < dark teas	Dong et al[186], 2011
Citrus aurantium Erigeron breviscapus	Hesperetin. Scutellarin. Nicotianamine. Glycyrrhizin. Baicalin	SARS-CoV-2/Connecting to ACE2 and blocking the SARS-CoV-2 input	Chen and Du[187], 2020
Glycine max			
Glycyrrhiza radix			
Scutellaria baicalensis			
Geranium and lemon essential oils	Citronellol and limonene	SARS-CoV-2/ACE2 inhibition	Senthil Kumar <i>et al</i> [188], 2020
Ginseng Glycyrrhiza uralensis	Ginsenoside Rg6; Ginsenoside F1; Monoammonium glycyrrhizinate; Glycyrrhizic acid methyl ester	SARS-CoV-2/ACE2 kinase inhibition	Zi et al [189] , 2020
<i>Glycine max</i> (soybean)	Nicotianamine	ACE2 inhibition	Takahashi <i>et al</i> [<mark>190</mark>], 2015
Glycyrrhiza glabra	Glycyrrhizic acid	SARS-CoV-2/Glycyrrhizic acid disrupts the connection of the virus with the ACE2 receptor at the entry level	Sinha et al[43], 2021
<i>Hibiscus sabdariffa</i> anthocyanins	Delphinidin- and cyanidin-3-O-sambubiosides	ACE inhibition	Ojeda <i>et al</i> [<mark>191</mark>], 2010
Linum usitatissimum (Flaxseed)	Secoisolariciresinol diglucoside	ACE inhibition	Prasad <i>et al</i> [192], 2013
<i>Melaleuca cajuputi</i> essential oil	Components (70.9% of the oil)	SARS-CoV-2/ACE2 and PDB6LU7 proteins inhibition	My et al[193], 2020
Nicotiana benthamiana	Recombinant ACE2-Fc fusion protein produced from <i>N. benthamiana</i>	SARS-CoV-2/Strong binding to the RBD of SARS-CoV-2 and inhibition	Siriwattananon <i>et al</i> [194], 2020
Withania somnifera	Withanone	SARS-CoV-2/Docking to the connector interface of the AEC2-RBD complex	Balkrishna <i>et al</i> [<mark>51</mark>], 2020

ACE: Angiotensin-converting enzyme; RBD: Receptor binding domain; SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2; ACE2: Angiotensin-converting enzyme-2.

> shown to inhibit Mpro by in silico study. It can be a potential inhibitor of controlling viral replication[48].

Withania somnifera

W. somnifera components withanolides have potential antiviral properties on COVID-19[49]. Patel et al[50] demonstrated that W. somnifera's Withanoside VI components have positive interactions at the binding site of protein targets of SARS-CoV-2. Withanone reduced the electrostatic interaction between ACE2 and receptor binding domain[51]. Withaferin A, which is found in the W. somnifera plant, has been shown to interact with Mpro and Glucose regulated protein 78 (GRP78) receptor[52].

CONCLUSION

In this study, the concept of "being effective against COVID-19" for herbal treatments was discussed from the angles of antiviral effect and control of symptoms, specifically related to GI system.

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Antiviral effects on COVID-19

Since COVID-19 is a virus that mainly affects the respiratory tract, the antiviral effects of medicinal plants against respiratory viruses have been examined firstly. The structure similarities of SARS-CoV-2 have been found with SARS-CoV and Middle East respiratory syndrome coronavirus. Therefore, it can be suggested that plants and their compounds affecting these viruses may also be potential treatment options for COVID-19. Here firstly, clinical studies supporting antiviral effects of 22 plant on respiratory viruses has been reviewed which determined that glycyrrhizic acid derivatives obtained from Glycyrrhiza sp, Nigella sativa, Scutellaria baicalensis and Torreya nucifera have anti-COVID-19 effects. Plants such as Allium sativum, Glycyrrhiza glabra, Melaleuca sp, Withania somnifera have been shown to bind to ACE2 receptors that are imperative for COVID-19 replication. Focusing on these plants might be a logical way to go for herbal treatment against COVID-19.

This review also showed the antiviral effects of essential oils obtained from plants have the potential to affect COVID-19. The treatment involves using inhaled steam supplemented by essential oils possessing natural antimicrobial properties, oropharyngeal sanitization, as well as they are remedies for symptomatic relief. Inhalation of antimicrobial essential oils may help attenuate the virus in the nasal cavity, nasopharynx, oropharynx, and laryngopharynx. Antiseptic mouthwashes and gargles can also help to sanitize the oral cavity and oropharynx, whereas antiseptic lozenges can help to sanitize the oro- and laryngopharynx as well. The steam will carry the tiny particles of the antimicrobial constituents from these essential oils into the respiratory tract and is likely to improve the efficacy of the steam treatment. The steam supplemented by antimicrobial volatile oils may help to provide a local antimicrobial effect within the airways.

There are computer model studies showing that some botanicals and active ingredients are effective in COVID-19. Allium sativum, Curcuma longa, Eucalyptus globulus, Glycyrrhiza glabra, Melaleuca sp, Thymus vulgaris, Withania somnifera is among these plants. These studies with commonly found plants will guide future studies to develop effective supplements or drugs for COVID-19.

Symptomatic treatment of COVID-19

Since the symptoms of COVID-19 seriously affect the quality of life, herbal options to eliminate them were also evaluated in this review. Previously, herbs such as garlic, echinacea and ginseng were found to reduce the symptoms of cold in healthy individuals. Plants with their pharmacological effects are natural options for eliminating the symptoms of COVID-19. Based on the effects described in Table 3, Allium sativum, Curcuma longa, Scutellaria baicalensis and Zingiber officinale are easily found as prominent plants to eliminate the GI symptoms of COVID-19. For example, ginger can eliminate the negative effects of COVID-19 on the GI system with its antiemetic and hepatic protective properties. A clinical study was conducted with thyme essential oil on COVID-19. Thyme essential oil was found to significantly reduce COVID-19 symptoms. This revealed an option that thyme and essential oil have potential effects for consideration in treatment of COVID-19. Studies on more essential oils of eucalyptus reveal more effects of eucalyptus on respiratory system symptoms. Eucalyptus globulus, Hedera helix, Pelargonium sidoides, Sambucus nigra, Thymus vulgaris can be recommended for relief of respiratory symptoms. ACE2 receptors are found in tissues other than the lung, such as the intestine. Based on this fact, we concluded that the use of herbs binding to ACE2 receptors can eliminate the side effects that may occur in variety of organs including GI tract. As shown in Table 4 these plants are Ammoides verticillate, Allium sativum, Apium graveolens, Camellia sinensis, Citrus aurantium, Erigeron breviscapus, Glycine max, Glycyrrhiza glabra, Hibiscus sabdariffa, Linum usitatissimum, Melaleuca sp., Nicotiana benthamiana, Withania somnifera.

Based on these studies, herbal treatments offer several potential treatments of COVID-19. Plants may be an option for the treatment of COVID-19 and its symptoms, as well as protection from COVID-19. Even though these data point to good outcomes there is always the possibility of interaction between drugs used and these herbs. For instance, herbs such as ginger with antithrombotic effects can be beneficial on COVID-19 symptoms, but one might be cautious about escalated risk of bleeding when it is used together with antithrombotic or anticoagulant drugs. Therefore, it is extremely important to avoid the indiscriminate use of plants.

For a plant to be used as a medicine, its effect must be supported by clinical studies. COVID-19 is just emerging, and more research are needed for its treatment. Yet, herbal therapies are potential options for both antiviral effects and the control of COVID-19 symptoms. Since plants with multiple pharmacological effects can affect many systems

(respiratory, GI, and nervous), herbs might be more effective against COVID-19 than synthetic drugs. But first, all aspects of SARS-CoV-2 need to be examined. Then, the effects of plants on this virus should be determined by further studies.

The strengths and weaknesses of this review

Unlike other studies, in this report, the effect of plants on COVID-19 was evaluated in several ways. Preclinical studies, clinical studies and silico studies are included in this review. Moreover, the efficacy on COVID-19 symptoms has been addressed by including different systems. On the other hand, the focus is on the respiratory and GI systems. The effects, not only of botanicals but also active metabolites of have been studied.

The biggest limitation of this study is the lack of sufficient studies on the efficacy of botanicals. Since botanical studies are generally preclinical studies, results may vary due to conducting and including clinical studies. In clinical studies showing the effects of the plants in Table 2 on respiratory tract infections, the results were generally obtained with questionnaire studies. Placebo effects and breadth of study may be effective in positive results.

REFERENCES

- Sun J, He WT, Wang L, Lai A, Ji X, Zhai X, Li G, Suchard MA, Tian J, Zhou J, Veit M, Su S. 1 COVID-19: Epidemiology, Evolution, and Cross-Disciplinary Perspectives. Trends Mol Med 2020; 26: 483-495 [PMID: 32359479 DOI: 10.1016/j.molmed.2020.02.008]
- 2 Zhang H, Penninger JM, Li Y, Zhong N, Slutsky AS. Angiotensin-converting enzyme 2 (ACE2) as a SARS-CoV-2 receptor: Molecular mechanisms and potential therapeutic target. Intensive Care Med 2020; 46: 586-590 [PMID: 32125455 DOI: 10.1007/s00134-020-05985-9]
- 3 Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, Schiergens TS, Herrler G, Wu NH, Nitsche A, Müller MA, Drosten C, Pöhlmann S. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. Cell 2020; 181: 271-280.e8 [PMID: 32142651 DOI: 10.1016/j.cell.2020.02.052]
- 4 Hashimoto T, Perlot T, Rehman A, Trichereau J, Ishiguro H, Paolino M, Sigl V, Hanada T, Hanada R, Lipinski S, Wild B, Camargo SM, Singer D, Richter A, Kuba K, Fukamizu A, Schreiber S, Clevers H, Verrey F, Rosenstiel P, Penninger JM. ACE2 links amino acid malnutrition to microbial ecology and intestinal inflammation. Nature 2012; 487: 477-481 [PMID: 22837003 DOI: 10.1038/nature11228]
- 5 Felsenstein S, Herbert JA, McNamara PS, Hedrich CM. COVID-19: Immunology and treatment options. Clin Immunol 2020; 215: 108448 [PMID: 32353634 DOI: 10.1016/j.clim.2020.108448]
- 6 Singh R, Shaik L, Mehra I, Kashyap R, Surani S. Novel and Controversial Therapies in COVID-19. Open Respir Med J 2020; 14: 79-86 [PMID: 33717367 DOI: 10.2174/1874306402014010079]
- 7 Silveira D, Prieto-Garcia JM, Boylan F, Estrada O, Fonseca-Bazzo YM, Jamal CM, Magalhães PO, Pereira EO, Tomczyk M, Heinrich M. COVID-19: Is There Evidence for the Use of Herbal Medicines as Adjuvant Symptomatic Therapy? Front Pharmacol 2020; 11: 581840 [PMID: 33071794 DOI: 10.3389/fphar.2020.581840]
- 8 Panyod S, Ho CT, Sheen LY. Dietary therapy and herbal medicine for COVID-19 prevention: A review and perspective. J Tradit Complement Med 2020; 10: 420-427 [PMID: 32691006 DOI: 10.1016/j.jtcme.2020.05.004]
- DU HZ, Hou XY, Miao YH, Huang BS, Liu DH. Traditional Chinese Medicine: an effective 9 treatment for 2019 novel coronavirus pneumonia (NCP). Chin J Nat Med 2020; 18: 206-210 [PMID: 32245590 DOI: 10.1016/S1875-5364(20)30022-4]
- 10 Xu K, Cai H, Shen Y, Ni Q, Chen Y, Hu S, Li J, Wang H, Yu L, Huang H, Qiu Y, Wei G, Fang Q, Zhou J, Sheng J, Liang T, Li L. [Management of corona virus disease-19 (COVID-19): the Zhejiang experience]. Zhejiang Da Xue Xue Bao Yi Xue Ban 2020; 49: 147-157 [PMID: 32096367 DOI: 10.3785/j.issn.1008-9292.2020.02.02
- 11 Jin YH, Cai L, Cheng ZS, Cheng H, Deng T, Fan YP, Fang C, Huang D, Huang LQ, Huang Q, Han Y, Hu B, Hu F, Li BH, Li YR, Liang K, Lin LK, Luo LS, Ma J, Ma LL, Peng ZY, Pan YB, Pan ZY, Ren XQ, Sun HM, Wang Y, Wang YY, Weng H, Wei CJ, Wu DF, Xia J, Xiong Y, Xu HB, Yao XM, Yuan YF, Ye TS, Zhang XC, Zhang YW, Zhang YG, Zhang HM, Zhao Y, Zhao MJ, Zi H, Zeng XT, Wang XH; , for the Zhongnan Hospital of Wuhan University Novel Coronavirus Management and Research Team, Evidence-Based Medicine Chapter of China International Exchange and Promotive Association for Medical and Health Care (CPAM). A rapid advice guideline for the diagnosis and treatment of 2019 novel coronavirus (2019-nCoV) infected pneumonia (standard version). Mil Med Res 2020; 7: 4 [PMID: 32029004 DOI: 10.1186/s40779-020-0233-6
- Naithani R, Mehta RG, Shukla D, Chandersekera SN, Moriarty RM. Antiviral activity of 12 phytochemicals: A current perspective. In Dietary Components and Immune Function. Totowa, NJ, Humana Press, 2010: 421-468



- 13 Vespa E, Pugliese N, Colapietro F, Aghemo A. Stay (GI) Healthy: COVID-19 and Gastrointestinal Manifestations. Tech Innov Gastrointest Endosc 2021; 23: 179-189 [PMID: 33521703 DOI: 10.1016/j.tige.2021.01.006
- 14 Adedeji AO, Severson W, Jonsson C, Singh K, Weiss SR, Sarafianos SG. Novel inhibitors of severe acute respiratory syndrome coronavirus entry that act by three distinct mechanisms. J Virol 2013; 87: 8017-8028 [PMID: 23678171 DOI: 10.1128/JVI.00998-13]
- 15 Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, Liu L, Shan H, Lei CL, Hui DSC, Du B, Li LJ, Zeng G, Yuen KY, Chen RC, Tang CL, Wang T, Chen PY, Xiang J, Li SY, Wang JL, Liang ZJ, Peng YX, Wei L, Liu Y, Hu YH, Peng P, Wang JM, Liu JY, Chen Z, Li G, Zheng ZJ, Qiu SQ, Luo J, Ye CJ, Zhu SY, Zhong NS; China Medical Treatment Expert Group for Covid-19. Clinical Characteristics of Coronavirus Disease 2019 in China. N Engl J Med 2020; 382: 1708-1720 [PMID: 32109013 DOI: 10.1056/NEJMoa2002032]
- 16 Yang X, Yu Y, Xu J, Shu H, Xia J, Liu H, Wu Y, Zhang L, Yu Z, Fang M, Yu T, Wang Y, Pan S, Zou X, Yuan S, Shang Y. Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study. Lancet Respir Med 2020; 8: 475-481 [PMID: 32105632 DOI: 10.1016/S2213-2600(20)30079-5]
- 17 Zhang JJ, Dong X, Cao YY, Yuan YD, Yang YB, Yan YQ, Akdis CA, Gao YD. Clinical characteristics of 140 patients infected with SARS-CoV-2 in Wuhan, China. Allergy 2020; 75: 1730-1741 [PMID: 32077115 DOI: 10.1111/all.14238]
- Fang L, Karakiulakis G, Roth M. Are patients with hypertension and diabetes mellitus at increased 18 risk for COVID-19 infection? Lancet Respir Med 2020; 8: e21 [PMID: 32171062 DOI: 10.1016/S2213-2600(20)30116-8
- 19 Benarba B, Pandiella A. Medicinal Plants as Sources of Active Molecules Against COVID-19. Front Pharmacol 2020; 11: 1189 [PMID: 32848790 DOI: 10.3389/fphar.2020.01189]
- 20 Mahony JB, Petrich A, Smieja M. Molecular diagnosis of respiratory virus infections. Crit Rev Clin Lab Sci 2011; 48: 217-249 [PMID: 22185616 DOI: 10.3109/10408363.2011.640976]
- 21 Talbot HK, Falsey AR. The diagnosis of viral respiratory disease in older adults. Clin Infect Dis 2010; **50**: 747-751 [PMID: 20121411 DOI: 10.1086/650486]
- Englund J, Feuchtinger T, Ljungman P. Viral infections in immunocompromised patients. Biol 22 Blood Marrow Transplant 2011; 17: S2-S5 [PMID: 21195305 DOI: 10.1016/j.bbmt.2010.11.008]
- 23 Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, Qiu Y, Wang J, Liu Y, Wei Y, Xia J, Yu T, Zhang X, Zhang L. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. Lancet 2020; 395: 507-513 [PMID: 32007143 DOI: 10.1016/S0140-6736(20)30211-7]
- Xia W, Shao J, Guo Y, Peng X, Li Z, Hu D. Clinical and CT features in pediatric patients with 24 COVID-19 infection: Different points from adults. Pediatr Pulmonol 2020; 55: 1169-1174 [PMID: 32134205 DOI: 10.1002/ppul.24718]
- Geier MR, Geier DA. Respiratory conditions in coronavirus disease 2019 (COVID-19): Important 25 considerations regarding novel treatment strategies to reduce mortality. Med Hypotheses 2020; 140: 109760 [PMID: 32344310 DOI: 10.1016/j.mehy.2020.109760]
- 26 Ang L, Song E, Lee HW, Lee MS. Herbal Medicine for the Treatment of Coronavirus Disease 2019 (COVID-19): A Systematic Review and Meta-Analysis of Randomized Controlled Trials. J Clin Med 2020; 9 [PMID: 32456123 DOI: 10.3390/jcm9051583]
- Fan AY, Gu S, Alemi SF; Research Group for Evidence-based Chinese Medicine. Chinese herbal 27 medicine for COVID-19: Current evidence with systematic review and meta-analysis. J Integr Med 2020; 18: 385-394 [PMID: 32792254 DOI: 10.1016/j.joim.2020.07.008]
- 28 Chen Y, de Bruyn Kops C, Kirchmair J. Data Resources for the Computer-Guided Discovery of Bioactive Natural Products. J Chem Inf Model 2017; 57: 2099-2111 [PMID: 28853576 DOI: 10.1021/acs.jcim.7b00341]
- 29 Yang YJ, Bang CS. Application of artificial intelligence in gastroenterology. World J Gastroenterol 2019; 25: 1666-1683 [PMID: 31011253 DOI: 10.3748/wjg.v25.i14.1666]
- 30 Khaerunnisa S, Kurniawan H, Awaluddin R, Suhartati S, Soetjipto S. Potential inhibitor of COVID-19 main protease (Mpro) from several medicinal plant compounds by molecular docking study. 2020 Preprint. Available from: 20944 [DOI: 10.20944/preprints202003.0226.v1]
- 31 Aanouz I, Belhassan A, El-Khatabi K, Lakhlifi T, El-Ldrissi M, Bouachrine M. Moroccan Medicinal plants as inhibitors against SARS-CoV-2 main protease: Computational investigations. J Biomol Struct Dyn 2021; 39: 2971-2979 [PMID: 32306860 DOI: 10.1080/07391102.2020.1758790]
- 32 Silva JKRD, Figueiredo PLB, Byler KG, Setzer WN. Essential Oils as Antiviral Agents. Potential of Essential Oils to Treat SARS-CoV-2 Infection: An In-Silico Investigation. Int J Mol Sci 2020; 21 [PMID: 32408699 DOI: 10.3390/ijms21103426]
- Utomo RY, Meiyanto E. Revealing the potency of citrus and galangal constituents to halt SARS-33 CoV-2 infection. 2020 Preprint. Available from: 2020030214 [DOI: 10.20944/preprints202003.0214.v1]
- 34 Rajagopal K, Varakumar P, Baliwada A, Byran G. Activity of phytochemical constituents of Curcuma longa (turmeric) and Andrographis paniculata against coronavirus (COVID-19): an in silico approach. Futur J Pharm Sci 2020; 6: 104 [PMID: 33215042 DOI: 10.1186/s43094-020-00126-x
- Adem S, Eyupoglu V, Sarfraz I, Rasul A, Ali M. Identification of potent COVID-19 main protease 35 (Mpro) inhibitors from natural polyphenols: An in silico strategy unveils a hope against corona. 2020



Preprint. Available from: 2020030333 [DOI: 10.20944/preprints202003.0333.v1]

- Srivastava A, Singh D. Destabilizing the structural integrity of SARS-CoV2 receptor proteins by 36 curcumin along with hydroxychloroquine: An in silico approach for a combination therapy. 2020 Preprint. Available from: chemrxiv:12090438 [DOI: 10.26434/chemrxiv.12090438]
- 37 Gonzalez-Paz LA, Lossada CA, Moncayo LS, Romero F, Paz JL, Vera-Villalobos J, Pérez AE, San-Blas E, Alvarado YJ. Theoretical molecular docking study of the structural disruption of the viral 3CL-protease of COVID19 induced by binding of capsaicin, piperine and curcumin part 1: A comparative study with chloroquine and hydrochloroquine two antimalaric drugs. 2020 Preprint. Available from: rs-21206 [DOI: 10.21203/rs.3.rs-21206/v1]
- 38 Sharma AD. Eucalyptol (1, 8 cineole) from eucalyptus essential oil a potential inhibitor of COVID 19 corona virus infection by molecular docking studies. 2020 Preprint. Available from: 2020030455
- 39 Sharma AD, Inderjeet KAUR. Molecular docking and pharmacokinetic screening of eucalyptol (1, 8 cineole) from eucalyptus essential oil against SARS-CoV-2. Not Sci Biol 2020; 12: 536-545 [DOI: 10.15835/nsb12210711]
- 40 Sharma AD, Kaur I. Molecular docking studies on jensenone from eucalyptus essential oil as a potential inhibitor of COVID 19 corona virus infection. 2020 Preprint. Available from: arXiv:2004.00217
- 41 Sharma AD, Kaur I. Eucalyptus essential oil bioactive molecules from against SARS-CoV-2 spike protein: Insights from computational studies. 2021 Preprint. Available from: rs-140069 [DOI: 10.21203/rs.3.rs-140069/v1]
- Muhammad IA, Muangchoo K, Muhammad A, Ajingi YUS, Muhammad IY, Umar ID, 42 Muhammad AB. A computational study to identify potential inhibitors of SARS-CoV-2 main protease (Mpro) from Eucalyptus active compounds. Computation 2020; 8: 79 [DOI: 10.3390/computation8030079
- 43 Sinha SK, Prasad SK, Islam MA, Gurav SS, Patil RB, AlFaris NA, Aldayel TS, AlKehayez NM, Wabaidur SM, Shakya A. Identification of bioactive compounds from Glycyrrhiza glabra as possible inhibitor of SARS-CoV-2 spike glycoprotein and non-structural protein-15: a pharmacoinformatics study. J Biomol Struct Dyn 2021; 39: 4686-4700 [PMID: 32552462 DOI: 10.1080/07391102.2020.1779132
- Narkhede RR, Pise AV, Cheke RS, Shinde SD. Recognition of Natural Products as Potential 44 Inhibitors of COVID-19 Main Protease (Mpro): In-Silico Evidences. Nat Prod Bioprospect 2020; 10: 297-306 [PMID: 32557405 DOI: 10.1007/s13659-020-00253-1]
- 45 Liu H, Ye F, Sun Q, Liang H, Li C, Li S, Lu R, Huang B, Tan W, Lai L. Scutellaria baicalensis extract and baicalein inhibit replication of SARS-CoV-2 and its 3C-like protease in vitro. J Enzyme Inhib Med Chem 2021; 36: 497-503 [PMID: 33491508 DOI: 10.1080/14756366.2021.1873977]
- 46 Udrea AM, Mernea M, Buiu C, Avram S. Scutellaria baicalensis flavones as potent drugs against acute respiratory injury during SARS-CoV-2 infection: Structural biology approaches. Processes 2020; 8: 1468 [DOI: 10.3390/pr8111468]
- Sardari S, Mobaiend A, Ghassemifard L, Kamali K, Khavasi N. Therapeutic effect of thyme (47 Thymus vulgaris) essential oil on patients with COVID19: A randomized clinical trial. J Adv Med 2021; 29: 83-91 [DOI: 10.30699/jambs.29.133.83]
- Kumar A, Choudhir G, Shukla SK, Sharma M, Tyagi P, Bhushan A, Rathore M. Identification of 48 phytochemical inhibitors against main protease of COVID-19 using molecular modeling approaches. J Biomol Struct Dyn 2021; 39: 3760-3770 [PMID: 32448034 DOI: 10.1080/07391102.2020.1772112
- 49 Dhawan M, Parmar M, Sharun K, Tiwari R, Bilal M, Dhama K. Medicinal and therapeutic potential of withanolides from Withania somnifera against COVID-19. J Appl Pharm Sci 2021; 11: 6-13 [DOI: 10.7324/JAPS.2021.110402]
- 50 Patel CN, Goswami D, Jaiswal DG, Parmar RM, Solanki HA, Pandya HA. Pinpointing the potential hits for hindering interaction of SARS-CoV-2 S-protein with ACE2 from the pool of antiviral phytochemicals utilizing molecular docking and molecular dynamics (MD) simulations. J Mol Graph Model 2021; 105: 107874 [PMID: 33647752 DOI: 10.1016/j.jmgm.2021.107874]
- Balkrishna A, Pokhrel S, Singh J, Varshney A. Withanone from Withania somnifera may inhibit 51 novel coronavirus (COVID-19) entry by disrupting interactions between viral S-protein receptor binding domain and host ACE2 receptor. 2020 Preprint. Available from: rs-17806 [DOI: 10.21203/rs.3.rs-17806/v1]
- 52 Sudeep HV, Gouthamchandra K, Shyamprasad K. Molecular docking analysis of Withaferin A from Withania somnifera with the Glucose regulated protein 78 (GRP78) receptor and the SARS-CoV-2 main protease. Bioinformation 2020; 16: 411-417 [PMID: 32831523 DOI: 10.6026/97320630016411
- 53 Rasool A, Khan MU, Ali MA, Anjum AA, Ahmed I, Aslam A, Mustafa G, Masood S, Nawaz M. Anti-avian influenza virus H9N2 activity of aqueous extracts of Zingiber officinalis (Ginger) and Allium sativum (Garlic) in chick embryos. Pak J Pharm Sci 2017; 30: 1341-1344 [PMID: 29039335]
- 54 Mohajer Shojai T, Ghalyanchi Langeroudi A, Karimi V, Barin A, Sadri N. The effect of Allium sativum (Garlic) extract on infectious bronchitis virus in specific pathogen free embryonic egg. Avicenna J Phytomed 2016; 6: 458-267 [PMID: 27516987]
- 55 Chavan RD, Shinde P, Girkar K, Madage R, Chowdhary A. Assessment of Anti-Influenza Activity and Hemagglutination Inhibition of Plumbago indica and Allium sativum Extracts. Pharmacognosy Res 2016; 8: 105-111 [PMID: 27034600 DOI: 10.4103/0974-8490.172562]



- 56 Choi HJ. Chemical Constituents of Essential Oils Possessing Anti-Influenza A/WS/33 Virus Activity. Osong Public Health Res Perspect 2018; 9: 348-353 [PMID: 30584499 DOI: 10.24171/j.phrp.2018.9.6.09]
- 57 Mehrbod P, Aini I, Amini E, Eslami M, Torabi A, Bande F, Kheiri MT. Assessment of direct immunofluorescence assay in detection of antiviral effect of garlic extract on influenza virus. Afr J Microbiol Res 2013; 7: 2608-2612 [DOI: 10.5897/ajmr12.2329]
- 58 Chen CH, Chou TW, Cheng LH, Ho CW. In vitro anti-adenoviral activity of five Allium plants. Taiwan Huaxuegongchengshi Xuehui Xuebao 2011; 42: 228-232 [DOI: 10.1016/j.jtice.2010.07.011]
- 59 Borges-Argáez R, Chan-Balan R, Cetina-Montejo L, Ayora-Talavera G, Sansores-Peraza P, Gómez-Carballo J, Cáceres-Farfán M. In vitro evaluation of anthraquinones from Aloe vera (Aloe barbadensis Miller) roots and several derivatives against strains of influenza virus. Ind Crops Prod 2019; 132: 468-475 [PMID: 32288269 DOI: 10.1016/j.indcrop.2019.02.056]
- 60 Li SW, Yang TC, Lai CC, Huang SH, Liao JM, Wan L, Lin YJ, Lin CW. Antiviral activity of aloeemodin against influenza A virus via galectin-3 up-regulation. Eur J Pharmacol 2014; 738: 125-132 [PMID: 24877694 DOI: 10.1016/j.ejphar.2014.05.028]
- Zhang P, Liu X, Liu H, Wang W, Li X, Wu X. Astragalus polysaccharides inhibit avian infectious 61 bronchitis virus infection by regulating viral replication. Microb Pathog 2018; 114: 124-128 [PMID: 29170045 DOI: 10.1016/j.micpath.2017.11.026]
- Kallon S, Li X, Ji J, Chen C, Xi Q, Chang S, Xue C, Ma J, Xie Q, Zhang Y. Astragalus 62 polysaccharide enhances immunity and inhibits H9N2 avian influenza virus in vitro and in vivo. J Anim Sci Biotechnol 2013; 4: 22 [PMID: 23786718 DOI: 10.1186/2049-1891-4-22]
- Weber JM, Ruzindana-Umunyana A, Imbeault L, Sircar S. Inhibition of adenovirus infection and adenain by green tea catechins. Antiviral Res 2003; 58: 167-173 [PMID: 12742577 DOI: 10.1016/s0166-3542(02)00212-7]
- 64 Kuzuhara T, Iwai Y, Takahashi H, Hatakeyama D, Echigo N. Green tea catechins inhibit the endonuclease activity of influenza A virus RNA polymerase. PLoS Curr 2009; 1: RRN1052 [PMID: 20025206 DOI: 10.1371/currents.rrn1052]
- 65 Liu J, Yang Z, Wang S, Liu L, Chen G, Wang L. Exploring the molecular basis of H5N1 hemagglutinin binding with catechins in green tea: A flexible docking and molecular dynamics study. J Theor Comput Chem 2012; 11: 111-125 [DOI: 10.1142/s0219633612500071]
- Yang ZF, Bai LP, Huang WB, Li XZ, Zhao SS, Zhong NS, Jiang ZH. Comparison of in vitro 66 antiviral activity of tea polyphenols against influenza A and B viruses and structure-activity relationship analysis. Fitoterapia 2014; 93: 47-53 [PMID: 24370660 DOI: 10.1016/i.fitote.2013.12.0111
- Chen TY, Chen DY, Wen HW, Ou JL, Chiou SS, Chen JM, Wong ML, Hsu WL. Inhibition of 67 enveloped viruses infectivity by curcumin. PLoS One 2013; 8: e62482 [PMID: 23658730 DOI: 10.1371/journal.pone.0062482]
- Dai J, Gu L, Su Y, Wang Q, Zhao Y, Chen X, Deng H, Li W, Wang G, Li K. Inhibition of curcumin 68 on influenza A virus infection and influenzal pneumonia via oxidative stress, TLR2/4, p38/JNK MAPK and NF-KB pathways. Int Immunopharmacol 2018; 54: 177-187 [PMID: 29153953 DOI: 10.1016/j.intimp.2017.11.009
- 69 Chen DY, Shien JH, Tiley L, Chiou SS, Wang SY, Chang TJ, Lee YJ, Chan KW, Hsu WL. Curcumin inhibits influenza virus infection and haemagglutination activity. Food Chem 2010; 119: 1346-1351 [DOI: 10.1016/j.foodchem.2009.09.011]
- 70 Obata K, Kojima T, Masaki T, Okabayashi T, Yokota S, Hirakawa S, Nomura K, Takasawa A, Murata M, Tanaka S, Fuchimoto J, Fujii N, Tsutsumi H, Himi T, Sawada N. Curcumin prevents replication of respiratory syncytial virus and the epithelial responses to it in human nasal epithelial cells. PLoS One 2013; 8: e70225 [PMID: 24058438 DOI: 10.1371/journal.pone.0070225]
- 71 Vimalanathan S, Schoop R, Hudson J. High-potency anti-influenza therapy by a combination of Echinacea purpurea fresh herb and root tinctures. J App Pharmac Sci 2013; 3: 001-005 [DOI: 10.1055/s-0033-1352301
- Pleschka S, Stein M, Schoop R, Hudson JB. Anti-viral properties and mode of action of 72 standardized Echinacea purpurea extract against highly pathogenic avian influenza virus (H5N1, H7N7) and swine-origin H1N1 (S-OIV). Virol J 2009; 6: 197 [PMID: 19912623 DOI: 10.1186/1743-422X-6-197]
- 73 Hudson J, Vimalanathan S. Echinacea—A source of potent antivirals for respiratory virus infections. Pharmaceuticals 2011; 4: 1019-1031 [DOI: 10.3390/ph4071019]
- 74 Vimalanathan S, Hudson J. Anti-influenza virus activity of essential oils and vapors. Am J Essent Oil 2014; 2: 47-53 [DOI: 10.7324/japs.2012.2734]
- Haruyama T, Nagata K. Anti-influenza virus activity of Ginkgo biloba leaf extracts. J Nat Med 75 2013; 67: 636-642 [PMID: 23179317 DOI: 10.1007/s11418-012-0725-0]
- Feng Yeh C, Wang KC, Chiang LC, Shieh DE, Yen MH, San Chang J. Water extract of licorice had 76 anti-viral activity against human respiratory syncytial virus in human respiratory tract cell lines. J Ethnopharmacol 2013; 148: 466-473 [PMID: 23643542 DOI: 10.1016/j.jep.2013.04.040]
- Hoever G, Baltina L, Michaelis M, Kondratenko R, Tolstikov GA, Doerr HW, Cinatl J Jr. Antiviral activity of glycyrrhizic acid derivatives against SARS-coronavirus. J Med Chem 2005; 48: 1256-1259 [PMID: 15715493 DOI: 10.1021/jm0493008]
- 78 Dao TT, Nguyen PH, Lee HS, Kim E, Park J, Lim SI, Oh WK. Chalcones as novel influenza A (H1N1) neuraminidase inhibitors from Glycyrrhiza inflata. Bioorg Med Chem Lett 2011; 21: 294-



298 [PMID: 21123068 DOI: 10.1016/j.bmcl.2010.11.016]

- 79 Wolkerstorfer A, Kurz H, Bachhofner N, Szolar OH. Glycyrrhizin inhibits influenza A virus uptake into the cell. Antiviral Res 2009; 83: 171-178 [PMID: 19416738 DOI: 10.1016/j.antiviral.2009.04.012
- 80 Michaelis M, Geiler J, Naczk P, Sithisarn P, Ogbomo H, Altenbrandt B, Leutz A, Doerr HW, Cinatl J Jr. Glycyrrhizin inhibits highly pathogenic H5N1 influenza A virus-induced pro-inflammatory cytokine and chemokine expression in human macrophages. Med Microbiol Immunol 2010; 199: 291-297 [PMID: 20386921 DOI: 10.1007/s00430-010-0155-0]
- 81 Del Valle Mendoza J, Pumarola T, Gonzales LA, Del Valle LJ. Antiviral activity of maca (Lepidium meyenii) against human influenza virus. Asian Pac J Trop Med 2014; 7S1: S415-S420 [PMID: 25312160 DOI: 10.1016/S1995-7645(14)60268-6]
- 82 Garozzo A, Timpanaro R, Stivala A, Bisignano G, Castro A. Activity of Melaleuca alternifolia (tea tree) oil on Influenza virus A/PR/8: Study on the mechanism of action. Antiviral Res 2011; 89: 83-88 [PMID: 21095205 DOI: 10.1016/j.antiviral.2010.11.010]
- Usachev EV, Pyankov OV, Usacheva OV, Agranovski IE. Antiviral activity of tea tree and 83 eucalyptus oil aerosol and vapour. J Aerosol Sci 2013; 59: 22-30 [DOI: 10.1016/j.jaerosci.2013.01.004]
- Pyankov OV, Usachev EV, Pyankova O, Agranovski IE. Inactivation of airborne influenza virus by 84 tea tree and eucalyptus oils. Aerosol Sci Technol 2012; 46: 1295-1302 [DOI: 10.1080/02786826.2012.708948]
- 85 Pourghanbari G, Nili H, Moattari A, Mohammadi A, Iraji A. Antiviral activity of the oseltamivir and Melissa officinalis L. essential oil against avian influenza A virus (H9N2). Virusdisease 2016; 27: 170-178 [PMID: 27366768 DOI: 10.1007/s13337-016-0321-0]
- 86 Lelešius R, Karpovaitė A, Mickienė R, Drevinskas T, Tiso N, Ragažinskienė O, Kubilienė L, Maruška A, Šalomskas A. In vitro antiviral activity of fifteen plant extracts against avian infectious bronchitis virus. BMC Vet Res 2019; 15: 178 [PMID: 31142304 DOI: 10.1186/s12917-019-1925-6]
- Li Y, Liu Y, Ma A, Bao Y, Wang M, Sun Z. In vitro antiviral, anti-inflammatory, and antioxidant 87 activities of the ethanol extract of Mentha piperita L. Food Sci Biotechnol 2017; 26: 1675-1683 [PMID: 30263705 DOI: 10.1007/s10068-017-0217-9]
- 88 Dorra N, El-Berrawy M, Sallam S, Mahmoud R. Evaluation of antiviral and antioxidant activity of selected herbal extracts. Public Health 2019; 49: 36-40 [DOI: 10.21608/jhiph.2019.29464]
- 89 Umar S, Munir MT, Subhan S, Azam T, Nisa Q, Khan MI, Umar W, Rehman Z, Saqib AS, Shah MA. Protective and antiviral activities of Nigella sativa against avian influenza (H9N2) in Turkeys. J Saudi Soc Agric Sci 2016; 10 [DOI: 10.1016/j.jssas.2016.09.004]
- Ulasli M, Gurses SA, Bayraktar R, Yumrutas O, Oztuzcu S, Igci M, Igci YZ, Cakmak EA, Arslan A. 90 The effects of Nigella sativa (Ns), Anthemis hyalina (Ah) and Citrus sinensis (Cs) extracts on the replication of coronavirus and the expression of TRP genes family. Mol Biol Rep 2014; 41: 1703-1711 [PMID: 24413991 DOI: 10.1007/s11033-014-3019-7]
- 91 Lee JS, Ko EJ, Hwang HS, Lee YN, Kwon YM, Kim MC, Kang SM. Antiviral activity of ginseng extract against respiratory syncytial virus infection. Int J Mol Med 2014; 34: 183-190 [PMID: 24756136 DOI: 10.3892/ijmm.2014.1750]
- Lee JS, Cho MK, Hwang HS, Ko EJ, Lee YN, Kwon YM, Kim MC, Kim KH, Lee YT, Jung YJ, 92 Kang SM. Ginseng diminishes lung disease in mice immunized with formalin-inactivated respiratory syncytial virus after challenge by modulating host immune responses. J Interferon Cytokine Res 2014; 34: 902-914 [PMID: 25051168 DOI: 10.1089/jir.2013.0093]
- 93 Yin SY, Kim HJ. A comparative study of the effects of whole red ginseng extract and polysaccharide and saponin fractions on influenza A (H1N1) virus infection. Biol Pharm Bull 2013; 36: 1002-1007 [PMID: 23727921 DOI: 10.1248/bpb.b13-00123]
- 94 Yoo DG, Kim MC, Park MK, Song JM, Quan FS, Park KM, Cho YK, Kang SM. Protective effect of Korean red ginseng extract on the infections by H1N1 and H3N2 influenza viruses in mice. J Med Food 2012; 15: 855-862 [PMID: 22856395 DOI: 10.1089/jmf.2012.0017]
- Roth M, Fang L, Stolz D, Tamm M. Pelargonium sidoides radix extract EPs 7630 reduces 95 rhinovirus infection through modulation of viral binding proteins on human bronchial epithelial cells. PLoS One 2019; 14: e0210702 [PMID: 30707726 DOI: 10.1371/journal.pone.0210702]
- 96 Michaelis M, Doerr HW, Cinatl J Jr. Investigation of the influence of EPs® 7630, a herbal drug preparation from Pelargonium sidoides, on replication of a broad panel of respiratory viruses. Phytomedicine 2011; 18: 384-386 [PMID: 21036571 DOI: 10.1016/j.phymed.2010.09.008]
- Theisen LL, Muller CP. EPs® 7630 (Umckaloabo®), an extract from Pelargonium sidoides roots, 97 exerts anti-influenza virus activity in vitro and in vivo. Antiviral Res 2012; 94: 147-156 [PMID: 22475498 DOI: 10.1016/j.antiviral.2012.03.006]
- 98 Chen C, Zuckerman DM, Brantley S, Sharpe M, Childress K, Hoiczyk E, Pendleton AR. Sambucus nigra extracts inhibit infectious bronchitis virus at an early point during replication. BMC Vet Res 2014; 10: 24 [PMID: 24433341 DOI: 10.1186/1746-6148-10-24]
- 99 Krawitz C, Mraheil MA, Stein M, Imirzalioglu C, Domann E, Pleschka S, Hain T. Inhibitory activity of a standardized elderberry liquid extract against clinically-relevant human respiratory bacterial pathogens and influenza A and B viruses. BMC Complement Altern Med 2011; 11: 16 [PMID: 21352539 DOI: 10.1186/1472-6882-11-16]
- 100 Kinoshita E, Hayashi K, Katayama H, Hayashi T, Obata A. Anti-influenza virus effects of elderberry juice and its fractions. Biosci Biotechnol Biochem 2012; 76: 1633-1638 [PMID: 22972323



DOI: 10.1271/bbb.120112]

- 101 Roschek B Jr, Fink RC, McMichael MD, Li D, Alberte RS. Elderberry flavonoids bind to and prevent H1N1 infection in vitro. Phytochemistry 2009; 70: 1255-1261 [PMID: 19682714 DOI: 10.1016/j.phytochem.2009.06.003
- 102 Ji S, Li R, Wang Q, Miao WJ, Li ZW, Si LL, Qiao X, Yu SW, Zhou DM, Ye M. Anti-H1N1 virus, cytotoxic and Nrf2 activation activities of chemical constituents from Scutellaria baicalensis. J Ethnopharmacol 2015; 176: 475-484 [PMID: 26578185 DOI: 10.1016/j.jep.2015.11.018]
- 103 Chen F, Chan KH, Jiang Y, Kao RY, Lu HT, Fan KW, Cheng VC, Tsui WH, Hung IF, Lee TS, Guan Y, Peiris JS, Yuen KY. In vitro susceptibility of 10 clinical isolates of SARS coronavirus to selected antiviral compounds. J Clin Virol 2004; 31: 69-75 [PMID: 15288617 DOI: 10.1016/j.jcv.2004.03.003
- Ryu YB, Jeong HJ, Kim JH, Kim YM, Park JY, Kim D, Nguyen TT, Park SJ, Chang JS, Park KH, 104 Rho MC, Lee WS. Biflavonoids from Torreya nucifera displaying SARS-CoV 3CL(pro) inhibition. Bioorg Med Chem 2010; 18: 7940-7947 [PMID: 20934345 DOI: 10.1016/j.bmc.2010.09.035]
- 105 Cai Z, Zhang G, Tang B, Liu Y, Fu X, Zhang X. Promising Anti-influenza Properties of Active Constituent of Withania somnifera Ayurvedic Herb in Targeting Neuraminidase of H1N1 Influenza: Computational Study. Cell Biochem Biophys 2015; 72: 727-739 [PMID: 25627548 DOI: 10.1007/s12013-015-0524-9
- 106 Chang JS, Wang KC, Yeh CF, Shieh DE, Chiang LC. Fresh ginger (Zingiber officinale) has antiviral activity against human respiratory syncytial virus in human respiratory tract cell lines. J Ethnopharmacol 2013; 145: 146-151 [PMID: 23123794 DOI: 10.1016/j.jep.2012.10.043]
- 107 Nantz MP, Rowe CA, Muller CE, Creasy RA, Stanilka JM, Percival SS. Supplementation with aged garlic extract improves both NK and $\gamma\delta$ -T cell function and reduces the severity of cold and flu symptoms: A randomized, double-blind, placebo-controlled nutrition intervention. Clin Nutr 2012; **31**: 337-344 [PMID: 22280901 DOI: 10.1016/j.clnu.2011.11.019]
- Barrett B, Brown R, Rakel D, Mundt M, Bone K, Barlow S, Ewers T. Echinacea for treating the 108 common cold: A randomized trial. Ann Intern Med 2010; 153: 769-777 [PMID: 21173411 DOI: 10.7326/0003-4819-153-12-201012210-00003
- 109 Jawad M, Schoop R, Suter A, Klein P, Eccles R. Safety and Efficacy Profile of Echinacea purpurea to Prevent Common Cold Episodes: A Randomized, Double-Blind, Placebo-Controlled Trial. Evid Based Complement Alternat Med 2012; 2012: 841315 [PMID: 23024696 DOI: 10.1155/2012/841315
- 110 Tiralongo E, Lea RA, Wee SS, Hanna MM, Griffiths LR. Randomised, double blind, placebocontrolled trial of echinacea supplementation in air travellers. Evid Based Complement Alternat Med 2012; 2012: 417267 [PMID: 22229040 DOI: 10.1155/2012/417267]
- Matsumoto K, Yamada H, Takuma N, Niino H, Sagesaka YM. Effects of green tea catechins and 111 theanine on preventing influenza infection among healthcare workers: A randomized controlled trial. BMC Complement Altern Med 2011; 11: 15 [PMID: 21338496 DOI: 10.1186/1472-6882-11-15]
- Fazio S, Pouso J, Dolinsky D, Fernandez A, Hernandez M, Clavier G, Hecker M. Tolerance, safety 112 and efficacy of Hedera helix extract in inflammatory bronchial diseases under clinical practice conditions: A prospective, open, multicentre postmarketing study in 9657 patients. Phytomedicine 2009; 16: 17-24 [PMID: 16860549 DOI: 10.1016/j.phymed.2006.05.003]
- 113 Schmidt M, Thomsen M, Schmidt U. Suitability of ivy extract for the treatment of paediatric cough. Phytother Res 2012; 26: 1942-1947 [PMID: 22532491 DOI: 10.1002/ptr.4671]
- 114 Zeil S, Schwanebeck U, Vogelberg C. Tolerance and effect of an add-on treatment with a cough medicine containing ivy leaves dry extract on lung function in children with bronchial asthma. Phytomedicine 2014; 21: 1216-1220 [PMID: 24916707 DOI: 10.1016/j.phymed.2014.05.006]
- 115 Ha KC, Kim MG, Oh MR, Choi EK, Back HI, Kim SY, Park EO, Kwon DY, Yang HJ, Kim MJ, Kang HJ, Lee JH, Choi KM, Chae SW, Lee CS. A placebo-controlled trial of Korean red ginseng extract for preventing influenza-like illness in healthy adults. BMC Complement Altern Med 2012; 12: 10 [PMID: 22314101 DOI: 10.1186/1472-6882-12-10]
- 116 Hwang JH, Park SH, Choi EK, Jung SJ, Pyo MK, Chae SW. A randomized, double-blind, placebocontrolled pilot study to assess the effects of protopanaxadiol saponin-enriched ginseng extract and pectinase-processed ginseng extract on the prevention of acute respiratory illness in healthy people. J Ginseng Res 2020; 44: 697-703 [PMID: 32913399 DOI: 10.1016/j.jgr.2019.01.002]
- High KP, Case D, Hurd D, Powell B, Lesser G, Falsey AR, Siegel R, Metzner-Sadurski J, Krauss JC, Chinnasami B, Sanders G, Rousey S, Shaw EG. A randomized, controlled trial of Panax quinquefolius extract (CVT-E002) to reduce respiratory infection in patients with chronic lymphocytic leukemia. J Support Oncol 2012; 10: 195-201 [PMID: 22266154 DOI: 10.1016/j.suponc.2011.10.005]
- Wu L, Zhang AL, Di YM, Shergis JL, Chen Y, Guo X, Wen Z, Thien F, Worsnop C, Lin L, Xue 118 CC. Panax ginseng therapy for chronic obstructive pulmonary disease: A clinical trial protocol and pilot study. Chin Med 2014; 9: 20 [PMID: 25161696 DOI: 10.1186/1749-8546-9-20]
- 119 Xue CC, Shergis JL, Zhang AL, Worsnop C, Fong H, Story D, Da Costa C, Thien FC. Panax ginseng C.A Meyer root extract for moderate chronic obstructive pulmonary disease (COPD): Study protocol for a randomised controlled trial. Trials 2011; 12: 164 [PMID: 21718484 DOI: 10.1186/1745-6215-12-164]
- Matthys H, Funk P. Pelargonium sidoides preparation EPs 7630 in COPD: Health-related quality-120 of-life and other patient-reported outcomes in adults receiving add-on therapy. Curr Med Res Opin



2018; 34: 1245-1251 [PMID: 29231073 DOI: 10.1080/03007995.2017.1416344]

- Kamin W, Ilyenko LI, Malek FA, Kieser M. Treatment of acute bronchitis with EPs 7630: 121 Randomized, controlled trial in children and adolescents. Pediatr Int 2012; 54: 219-226 [PMID: 22360575 DOI: 10.1111/j.1442-200X.2012.03598.x]
- 122 Patiroglu T, Tunc A, Eke Gungor H, Unal E. The efficacy of Pelargonium sidoides in the treatment of upper respiratory tract infections in children with transient hypogammaglobulinemia of infancy. Phytomedicine 2012; 19: 958-961 [PMID: 22809962 DOI: 10.1016/j.phymed.2012.06.004]
- 123 Tahan F, Yaman M. Can the Pelargonium sidoides root extract EPs® 7630 prevent asthma attacks during viral infections of the upper respiratory tract in children? Phytomedicine 2013; 20: 148-150 [PMID: 23142309 DOI: 10.1016/j.phymed.2012.09.022]
- Berezhnoi VV, Heger M, Lehmacher W, Seifert G. Clinical efficacy and safety of liquid 124 Pelargonium sidoides preparation (EPs 7630) in children with acute non-streptococcal tonsillopharyngitis. J Compr Ped 2016; 7 [DOI: 10.17795/compreped-42158]
- 125 Riley DS, Lizogub VG, Zimmermann A, Funk P, Lehmacher W. Efficacy and Tolerability of Highdose Pelargonium Extract in Patients with the Common Cold. Altern Ther Health Med 2018; 24: 16-26 [PMID: 29055287]
- Kong FK. Pilot clinical study on a proprietary elderberry extract: Efficacy in addressing influenza 126 symptoms. J Pharmacokinet Pharmacodyn 2009; 5: 32-43 [DOI: 10.1007/s10928-014-9365-1]
- 127 Tiralongo E, Wee SS, Lea RA. Elderberry Supplementation Reduces Cold Duration and Symptoms in Air-Travellers: A Randomized, Double-Blind Placebo-Controlled Clinical Trial. Nutrients 2016; 8: 182 [PMID: 27023596 DOI: 10.3390/nu8040182]
- 128 Dehghani S, Alipoor E, Salimzadeh A, Yaseri M, Hosseini M, Feinle-Bisset C, Hosseinzadeh-Attar MJ. The effect of a garlic supplement on the pro-inflammatory adipocytokines, resistin and tumor necrosis factor-alpha, and on pain severity, in overweight or obese women with knee osteoarthritis. Phytomedicine 2018; 48: 70-75 [PMID: 30195882 DOI: 10.1016/j.phymed.2018.04.060]
- Arreola R, Quintero-Fabián S, López-Roa RI, Flores-Gutiérrez EO, Reyes-Grajeda JP, Carrera-129 Quintanar L, Ortuño-Sahagún D. Immunomodulation and anti-inflammatory effects of garlic compounds. J Immunol Res 2015; 2015: 401630 [PMID: 25961060 DOI: 10.1155/2015/401630]
- 130 Hiyasat B, Sabha D, Grotzinger K, Kempfert J, Rauwald JW, Mohr FW, Dhein S. Antiplatelet activity of Allium ursinum and Allium sativum. Pharmacology 2009; 83: 197-204 [PMID: 19174616 DOI: 10.1159/0001968111
- 131 Sultana MR, Bagul PK, Katare PB, Anwar Mohammed S, Padiya R, Banerjee SK. Garlic activates SIRT-3 to prevent cardiac oxidative stress and mitochondrial dysfunction in diabetes. Life Sci 2016; 164: 42-51 [PMID: 27590611 DOI: 10.1016/j.lfs.2016.08.030]
- 132 Aprioku JS, Amah-Tariah FS. Garlic (Allium sativum L.) protects hepatic and renal toxicity of alloxan in rats. Br J Pharm Res 2017; 1-7 [DOI: 10.9734/JPRI/2017/34909]
- Chen YA, Tsai JC, Cheng KC, Liu KF, Chang CK, Hsieh CW. Extracts of black garlic exhibits 133 gastrointestinal motility effect. Food Res Int 2018; 107: 102-109 [PMID: 29580467 DOI: 10.1016/j.foodres.2018.02.003
- Seckiner I, Bayrak O, Can M, Mungan AG, Mungan NA. Garlic supplemented diet attenuates 134 gentamicin nephrotoxicity in rats. Int Braz J Urol 2014; 40: 562-567 [PMID: 25251961 DOI: 10.1590/S1677-5538.IBJU.2014.04.17]
- 135 Henrotin Y, Donneau AF, de Vlam K, Wittoek R, Luyten F. Responses to "Bio-optimized Curcuma longa extract is efficient on knee osteoarthritis pain: A double-blind multicenter randomized placebo controlled three-arm study": authors' reply. Arthritis Res Ther 2020; 22: 23 [PMID: 32046787 DOI: 10.1186/s13075-020-2109-2]
- Eke-Okoro UJ, Raffa RB, Pergolizzi JV Jr, Breve F, Taylor R Jr; NEMA Research Group. 136 Curcumin in turmeric: Basic and clinical evidence for a potential role in analgesia. J Clin Pharm Ther 2018; 43: 460-466 [PMID: 29722036 DOI: 10.1111/jcpt.12703]
- 137 Liu Z, Huang P, Law S, Tian H, Leung W, Xu C. Preventive Effect of Curcumin Against Chemotherapy-Induced Side-Effects. Front Pharmacol 2018; 9: 1374 [PMID: 30538634 DOI: 10.3389/fphar.2018.01374]
- 138 Huang WC, Chiu WC, Chuang HL, Tang DW, Lee ZM, Wei L, Chen FA, Huang CC. Effect of curcumin supplementation on physiological fatigue and physical performance in mice. Nutrients 2015; 7: 905-921 [PMID: 25647661 DOI: 10.3390/nu7020905]
- 139 Shimizu K, Funamoto M, Sunagawa Y, Shimizu S, Katanasaka Y, Miyazaki Y, Wada H, Hasegawa K, Morimoto T. Anti-inflammatory Action of Curcumin and Its Use in the Treatment of Lifestylerelated Diseases. Eur Cardiol 2019; 14: 117-122 [PMID: 31360234 DOI: 10.15420/ecr.2019.17.2]
- 140 Gouda MM, Bhandary YP. Acute Lung Injury: IL-17A-Mediated Inflammatory Pathway and Its Regulation by Curcumin. Inflammation 2019; 42: 1160-1169 [PMID: 31011925 DOI: 10.1007/s10753-019-01010-4
- 141 Haider S, Naqvi F, Tabassum S, Saleem S, Batool Z, Sadir S, Rasheed S, Saleem D, Nawaz A, Ahmad S. Preventive effects of curcumin against drug- and starvation-induced gastric erosions in rats. Sci Pharm 2013; 81: 549-558 [PMID: 23833720 DOI: 10.3797/scipharm.1207-17]
- 142 Ram A, Das M, Ghosh B. Curcumin attenuates allergen-induced airway hyperresponsiveness in sensitized guinea pigs. Biol Pharm Bull 2003; 26: 1021-1024 [PMID: 12843631 DOI: 10.1248/bpb.26.1021
- Dulbecco P, Savarino V. Therapeutic potential of curcumin in digestive diseases. World J 143 Gastroenterol 2013; 19: 9256-9270 [PMID: 24409053 DOI: 10.3748/wjg.v19.i48.9256]



- Nosalova G, Fleskova D, Jurecek L, Sadlonova V, Ray B. Herbal polysaccharides and cough reflex. 144 Respir Physiol Neurobiol 2013; 187: 47-51 [PMID: 23597834 DOI: 10.1016/j.resp.2013.03.015]
- 145 Kuang Y, Li B, Fan J, Qiao X, Ye M. Antitussive and expectorant activities of licorice and its major compounds. Bioorg Med Chem 2018; 26: 278-284 [PMID: 29224994 DOI: 10.1016/j.bmc.2017.11.046]
- Kao TC, Shyu MH, Yen GC. Glycyrrhizic acid and 18beta-glycyrrhetinic acid inhibit inflammation 146 via PI3K/Akt/GSK3beta signaling and glucocorticoid receptor activation. J Agric Food Chem 2010; 58: 8623-8629 [PMID: 20681651 DOI: 10.1021/jf101841r]
- 147 Shi Q, Hou Y, Yang Y, Bai G. Protective effects of glycyrrhizin against β2-adrenergic receptor agonist-induced receptor internalization and cell apoptosis. Biol Pharm Bull 2011; 34: 609-617 [PMID: 21532146 DOI: 10.1248/bpb.34.609]
- 148 Rushmi ZT, Akter N, Mow RJ, Afroz M, Kazi M, de Matas M, Rahman M, Shariare MH. The impact of formulation attributes and process parameters on black seed oil loaded liposomes and their performance in animal models of analgesia. Saudi Pharm J 2017; 25: 404-412 [PMID: 28344496 DOI: 10.1016/j.jsps.2016.09.011]
- Muralidharan-Chari V, Kim J, Abuawad A, Naeem M, Cui H, Mousa SA. Thymoquinone 149 Modulates Blood Coagulation in Vitro via Its Effects on Inflammatory and Coagulation Pathways. Int J Mol Sci 2016; 17: 474 [PMID: 27043539 DOI: 10.3390/ijms17040474]
- Ansari MA, Ansari NA, Junejo SA. Montelukast vs Nigella sativa for management of seasonal 150 allergic rhinitis: A single blind comparative clinical trial. Pak J Med Sci 2010; 26: 249-254 [DOI: 10.1046/j.1365-2222.2002.01422.x]
- 151 Alsamarai AM, Abdulsatar M, Ahmed Alobaidi AH. Evaluation of topical black seed oil in the treatment of allergic rhinitis. Antiinflamm Antiallergy Agents Med Chem 2014; 13: 75-82 [PMID: 23855426 DOI: 10.2174/18715230113129990014]
- 152 Majdalawieh AF, Fayyad MW. Immunomodulatory and anti-inflammatory action of Nigella sativa and thymoquinone: A comprehensive review. Int Immunopharmacol 2015; 28: 295-304 [PMID: 26117430 DOI: 10.1016/j.intimp.2015.06.023]
- 153 Mahdavi R, Namazi N, Alizadeh M, Farajnia S. Nigella sativa oil with a calorie-restricted diet can improve biomarkers of systemic inflammation in obese women: A randomized double-blind, placebo-controlled clinical trial. J Clin Lipidol 2016; 10: 1203-1211 [PMID: 27678438 DOI: 10.1016/j.jacl.2015.11.019
- 154 Boskabady MH, Mohsenpoor N, Takaloo L. Antiasthmatic effect of Nigella sativa in airways of asthmatic patients. Phytomedicine 2010; 17: 707-713 [PMID: 20149611 DOI: 10.1016/j.phymed.2010.01.002
- Salem AM, Bamosa AO, Qutub HO, Gupta RK, Badar A, Elnour A, Afzal MN. Effect of Nigella 155 sativa supplementation on lung function and inflammatory mediatorsin partly controlled asthma: a randomized controlled trial. Ann Saudi Med 2017; 37: 64-71 [PMID: 28151459 DOI: 10.5144/0256-4947.2017.64
- Ratan ZA, Youn SH, Kwak YS, Han CK, Haidere MF, Kim JK, Min H, Jung YJ, Hosseinzadeh H, 156 Hyun SH, Cho JY. Adaptogenic effects of Panax ginseng on modulation of immune functions. J Ginseng Res 2021; 45: 32-40 [PMID: 33437154 DOI: 10.1016/j.jgr.2020.09.004]
- 157 **Bao Y.** Gao Y. Koch E. Pan X. Jin Y. Cui X. Evaluation of pharmacodynamic activities of EPs[®] 7630, a special extract from roots of Pelargonium sidoides, in animals models of cough, secretolytic activity and acute bronchitis. Phytomedicine 2015; 22: 504-509 [PMID: 25925973 DOI: 10.1016/j.phymed.2015.03.004]
- 158 Aung H, Mehendale S, Chang WT, Wang CZ, Xie JT, Yuan CS. Scutellaria baicalensis decreases ritonavir-induced nausea. AIDS Res Ther 2005; 2: 12 [PMID: 16368007 DOI: 10.1186/1742-6405-2-12
- Hong GE, Kim JA, Nagappan A, Yumnam S, Lee HJ, Kim EH, Lee WS, Shin SC, Park HS, Kim 159 GS. Flavonoids Identified from Korean Scutellaria baicalensis Georgi Inhibit Inflammatory Signaling by Suppressing Activation of NF- K B and MAPK in RAW 264.7 Cells. Evid Based Complement Alternat Med 2013; 2013: 912031 [PMID: 24348728 DOI: 10.1155/2013/912031]
- 160 Mehendale S, Aung H, Wang CZ, Tong R, Foo A, Xie JT, Yuan CS. Scutellaria baicalensis and a constituent flavonoid, baicalein, attenuate ritonavir-induced gastrointestinal side-effects. J Pharm Pharmacol 2007; 59: 1567-1572 [PMID: 17976269 DOI: 10.1211/jpp.59.11.0015]
- Cui L, Guan X, Ding W, Luo Y, Wang W, Bu W, Song J, Tan X, Sun E, Ning Q, Liu G, Jia X, Feng 161 L. Scutellaria baicalensis Georgi polysaccharide ameliorates DSS-induced ulcerative colitis by improving intestinal barrier function and modulating gut microbiota. Int J Biol Macromol 2021; 166: 1035-1045 [PMID: 33157130 DOI: 10.1016/j.ijbiomac.2020.10.259]
- Thanh HN, Minh HPT, Le TA, Ly HDT, Huu TN, Duc LV, Kim TD, Thanh TB. Ethanol extracts of 162 Scutellaria baicalensis protect against lipopolysaccharide-induced acute liver injury in mice. Asian Pac J Trop Biomed 2015; 5: 761-767 [DOI: 10.1016/j.apjtb.2015.07.007]
- Dai J, Chen L, Qiu YM, Li SQ, Xiong WH, Yin YH, Jia F, Jiang JY. Activations of GABAergic 163 signaling, HSP70 and MAPK cascades are involved in baicalin's neuroprotection against gerbil global ischemia/reperfusion injury. Brain Res Bull 2013; 90: 1-9 [PMID: 23041106 DOI: 10.1016/j.brainresbull.2012.09.014]
- 164 Bui TT, Piao CH, Song CH, Lee CH, Shin HS, Chai OH. Baicalein, wogonin, and Scutellaria baicalensis ethanol extract alleviate ovalbumin-induced allergic airway inflammation and mast cellmediated anaphylactic shock by regulation of Th1/Th2 imbalance and histamine release. Anat Cell



Biol 2017; 50: 124-134 [PMID: 28713616 DOI: 10.5115/acb.2017.50.2.124]

- Laub A. Using species of the Lamiaceae family for musculoskeletal pain. 2018. [cited 10 January 165 2021]. Available from: https://www.researchgate.net/publication/335156154 Using Species of the Lamiaceae Family for Musculoskeletal Pain
- 166 Salmalian H, Saghebi R, Moghadamnia AA, Bijani A, Faramarzi M, Nasiri Amiri F, Bakouei F, Behmanesh F, Bekhradi R. Comparative effect of Thymus vulgaris and ibuprofen on primary dysmenorrhea: A triple-blind clinical study. Caspian J Intern Med 2014; 5: 82-88 [PMID: 24778782
- 167 Okazaki K, Kawazoe K, Takaishi Y. Human platelet aggregation inhibitors from thyme (Thymus vulgaris L.). Phytother Res 2002; 16: 398-399 [PMID: 12112303 DOI: 10.1002/ptr.979]
- 168 Habashy NH, Serie MM, Attia WE, Abdelgaleil SA. Chemical characterization, antioxidant and anti-inflammatory properties of Greek Thymus vulgaris extracts and their possible synergism with Egyptian Chlorella vulgaris. J Funct Foods 2018; 40: 317-328 [DOI: 10.1016/j.jff.2017.11.022]
- Salve J, Pate S, Debnath K, Langade D. Adaptogenic and Anxiolytic Effects of Ashwagandha Root 169 Extract in Healthy Adults: A Double-blind, Randomized, Placebo-controlled Clinical Study. Cureus 2019; 11: e6466 [PMID: 32021735 DOI: 10.7759/cureus.6466]
- Murthy MNK, Gundagani S, Nutalapati C, Pingali U. Evaluation of analgesic activity of 170 standardised aqueous extract of Withania somnifera in healthy human volunteers using mechanical pain model. J Clin Diagn 2019; 13: 1-4 [DOI: 10.7860/jcdr/2019/37590.12441]
- Ku SK, Bae JS. Antiplatelet, anticoagulant, and profibrinolytic activities of withaferin A. Vascul 171 Pharmacol 2014; 60: 120-126 [PMID: 24534482 DOI: 10.1016/j.vph.2014.01.009]
- 172 Gupta A, Singh S. Evaluation of anti-inflammatory effect of Withania somnifera root on collageninduced arthritis in rats. Pharm Biol 2014; 52: 308-320 [PMID: 24188460 DOI: 10.3109/13880209.2013.835325
- 173 Lopresti AL, Smith SJ, Malvi H, Kodgule R. An investigation into the stress-relieving and pharmacological actions of an ashwagandha (Withania somnifera) extract: A randomized, doubleblind, placebo-controlled study. Medicine (Baltimore) 2019; 98: e17186 [PMID: 31517876 DOI: 10.1097/MD.000000000017186
- Maghbooli M, Golipour F, Moghimi Esfandabadi A, Yousefi M. Comparison between the efficacy 174 of ginger and sumatriptan in the ablative treatment of the common migraine. Phytother Res 2014; 28: 412-415 [PMID: 23657930 DOI: 10.1002/ptr.4996]
- 175 Bartels EM, Folmer VN, Bliddal H, Altman RD, Juhl C, Tarp S, Zhang W, Christensen R. Efficacy and safety of ginger in osteoarthritis patients: A meta-analysis of randomized placebo-controlled trials. Osteoarthritis Cartilage 2015; 23: 13-21 [PMID: 25300574 DOI: 10.1016/j.joca.2014.09.024]
- Tóth B, Lantos T, Hegyi P, Viola R, Vasas A, Benkő R, Gyöngyi Z, Vincze Á, Csécsei P, Mikó A, 176 Hegyi D, Szentesi A, Matuz M, Csupor D. Ginger (Zingiber officinale): An alternative for the prevention of postoperative nausea and vomiting. A meta-analysis. Phytomedicine 2018; 50: 8-18 [PMID: 30466995 DOI: 10.1016/j.phymed.2018.09.007]
- 177 Khan AM, Shahzad M, Raza Asim MB, Imran M, Shabbir A. Zingiber officinale ameliorates allergic asthma via suppression of Th2-mediated immune response. Pharm Biol 2015; 53: 359-367 [PMID: 25420680 DOI: 10.3109/13880209.2014.920396]
- 178 Lee W. Ku SK, Kim MA, Bae JS, Anti-factor Xa activities of zingerone with anti-platelet aggregation activity. Food Chem Toxicol 2017; 105: 186-193 [PMID: 28414123 DOI: 10.1016/j.fct.2017.04.012]
- 179 Bera K, Nosalova G, Sivova V, Ray B. Structural Elements and Cough Suppressing Activity of Polysaccharides from Zingiber officinale Rhizome. Phytother Res 2016; 30: 105-111 [PMID: 26522239 DOI: 10.1002/ptr.5508]
- Nanjundaiah SM, Annaiah HN, Dharmesh SM. Gastroprotective Effect of Ginger Rhizome (180 Zingiber officinale) Extract: Role of Gallic Acid and Cinnamic Acid in H(+), K(+)-ATPase/H. pylori Inhibition and Anti-Oxidative Mechanism. Evid Based Complement Alternat Med 2011; 2011: 249487 [PMID: 19570992 DOI: 10.1093/ecam/nep060]
- 181 Ajith TA, Hema U, Aswathy MS. Zingiber officinale Roscoe prevents acetaminophen-induced acute hepatotoxicity by enhancing hepatic antioxidant status. Food Chem Toxicol 2007; 45: 2267-2272 [PMID: 17637489 DOI: 10.1016/j.fct.2007.06.001]
- 182 Ajith TA, Nivitha V, Usha S. Zingiber officinale Roscoe alone and in combination with alphatocopherol protect the kidney against cisplatin-induced acute renal failure. Food Chem Toxicol 2007; 45: 921-927 [PMID: 17210214 DOI: 10.1016/j.fct.2006.11.014]
- Abdelli I, Hassani F, Bekkel Brikci S, Ghalem S. In silico study the inhibition of angiotensin 183 converting enzyme 2 receptor of COVID-19 by Ammoides verticillata components harvested from Western Algeria. J Biomol Struct Dyn 2021; 39: 3263-3276 [PMID: 32362217 DOI: 10.1080/07391102.2020.1763199
- Thuy BTP, My TTA, Hai NTT, Hieu LT, Hoa TT, Thi Phuong Loan H, Triet NT, Anh TTV, Quy 184 PT, Tat PV, Hue NV, Quang DT, Trung NT, Tung VT, Huynh LK, Nhung NTA. Investigation into SARS-CoV-2 Resistance of Compounds in Garlic Essential Oil. ACS Omega 2020; 5: 8312-8320 [PMID: 32363255 DOI: 10.1021/acsomega.0c00772]
- 185 Sui H, Yu Q, Zhi Y, Geng G, Liu H, Xu H. [Effects of apigenin on the expression of angiotensinconverting enzyme 2 in kidney in spontaneously hypertensive rats]. Wei Sheng Yan Jiu 2010; 39: 693-696, 700 [PMID: 21351633]
- Dong J, Xu X, Liang Y, Head R, Bennett L. Inhibition of angiotensin converting enzyme (ACE) 186



activity by polyphenols from tea (Camellia sinensis) and links to processing method. Food Funct 2011; 2: 310-319 [PMID: 21779569 DOI: 10.1039/c1fo10023h]

- 187 Chen H, Du Q. Potential natural compounds for preventing SARS-CoV-2 (2019-nCoV) infection. 2020 Preprint. Available from: 2020010358 [DOI: 10.20944/preprints202001.0358.v3]
- Senthil Kumar KJ, Gokila Vani M, Wang CS, Chen CC, Chen YC, Lu LP, Huang CH, Lai CS, 188 Wang SY. Geranium and Lemon Essential Oils and Their Active Compounds Downregulate Angiotensin-Converting Enzyme 2 (ACE2), a SARS-CoV-2 Spike Receptor-Binding Domain, in Epithelial Cells. Plants (Basel) 2020; 9 [PMID: 32575476 DOI: 10.3390/plants9060770]
- 189 Zi CT, Zhang N, Yang L, Wang LX, Wu YL, Su YS, Wang XJ. Discovery of a potent angiotensin converting enzyme 2 inhibitor from Chinese medicinal and edible plant via docking-based virtual screening. 2020 Preprint. Available from: rs-32515 [DOI: 10.21203/rs.3.rs-32515/v1]
- 190 Takahashi S, Yoshiya T, Yoshizawa-Kumagaye K, Sugiyama T. Nicotianamine is a novel angiotensin-converting enzyme 2 inhibitor in soybean. Biomed Res 2015; 36: 219-224 [PMID: 26106051 DOI: 10.2220/biomedres.36.219]
- Ojeda D, Jiménez-Ferrer E, Zamilpa A, Herrera-Arellano A, Tortoriello J, Alvarez L. Inhibition of 191 angiotensin convertin enzyme (ACE) activity by the anthocyanins delphinidin- and cyanidin-3-Osambubiosides from Hibiscus sabdariffa. J Ethnopharmacol 2010; 127: 7-10 [PMID: 19808084 DOI: 10.1016/j.jep.2009.09.059]
- 192 Prasad K. Secoisolariciresinol Diglucoside (SDG) Isolated from Flaxseed, an Alternative to ACE Inhibitors in the Treatment of Hypertension. Int J Angiol 2013; 22: 235-238 [PMID: 24436618 DOI: 10.1055/s-0033-1351687
- My TTA, Loan HTP, Hai NTT, Hieu LT, Hoa TT, Thuy BTP, Quang DT, Triet NT, Anh TTV, Dieu 193 NTX, Trung NT, Hue NV, Tat PV, Tung VT, Nhung NTA. Evaluation of the Inhibitory Activities of COVID-19 of Melaleuca cajuputi Oil Using Docking Simulation. Chemistry Select 2020; 5: 6312-6320 [PMID: 32572383 DOI: 10.1002/slct.202000822]
- 194 Siriwattananon K, Manopwisedjaroen S, Kanjanasirirat P, Budi Purwono P, Rattanapisit K, Shanmugaraj B, Smith DR, Borwornpinyo S, Thitithanyanont A, Phoolcharoen W. Development of Plant-Produced Recombinant ACE2-Fc Fusion Protein as a Potential Therapeutic Agent Against SARS-CoV-2. Front Plant Sci 2020; 11: 604663 [PMID: 33584747 DOI: 10.3389/fpls.2020.604663]



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MINIREVIEWS

Current and emerging therapeutic strategies in pancreatic cancer: Challenges and opportunities

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Abstract

Pancreatic carcinoma (PC) is one of the leading causes of cancer-related deaths worldwide. Despite early detection and advances in therapeutics, the prognosis remains dismal. The outcome and therapeutic approach are dependent on the stage of PC at the time of diagnosis. The standard of care is surgery, followed by adjuvant chemotherapy. The advent of newer drugs has changed the landscape of adjuvant therapy. Moreover, recent trials have highlighted the role of neoadjuvant therapy and chemoradiotherapy for resectable and borderline resectable PC. As we progress towards a better understanding of tumor biology, genetics, and microenvironment, novel therapeutic strategies and targeted agents are now on the horizon. We have described the current and emerging therapeutic strategies in PC.

Key Words: Resectable pancreatic carcinoma; Borderline resectable pancreatic carcinoma; Locally advanced pancreatic carcinoma; Adjuvant therapy; Neoadjuvant therapy; Newer advances in pancreatic carcinoma

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Core Tip: An improved understanding of the natural history of pancreatic carcinoma, genetics, and tumor biology has highlighted the role of novel therapeutic strategies. However, despite recent advances in the management of pancreatic carcinoma, the

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prognosis remains poor. We have attempted to conceptualize the current therapeutic strategies in light of recent advances.

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INTRODUCTION

World over, new cases of pancreatic carcinoma (PC) add up close to three lakh each year[1,2]. There hasn't been a significant increase in the long-term survival rates, with the 5-year survival rates increasing to 5%-6% over the last 30 years, despite early detection and advances in therapeutics for pancreatic cancer[3,4]. The estimates of leading causes of cancer deaths suggest that PC may become the second, next only to lung cancer in the United States over the next decade[1].

Pancreatic ductal adenocarcinoma (PDAC) is exemplified by abundant genetic mutations, germline or acquired. Among the common ones are CDK2NA and KRAS seen in nearly 90%, TP53 in 75%-90%, and SMAD4/DPC4 in about 50% [5,6]. Additionally, genomic and epigenetic alterations are present, which have ignited research for targeted therapy. The desmoplastic stroma and the tumor microenvironment have been the focus of clinical explorations.

The outcomes of PC depend on the stage at diagnosis. Nearly half the cases are diagnosed as metastatic, wherein the survival ranges from 7-11 mo, at best[7,8]. In cases where the disease is non-metastatic but unresectable, there is a modest increase of survival, of nearly 6 months over the metastatic disease. The peculiarity of resectable PC lies in the poor overall survival, of approximately 2 years with adjuvant therapy. This is in stark contrast to most of the other resectable cancers.

The standard of care of resectable PC is surgery followed by adjuvant chemotherapy (CT). The benefit for this approach was established by the European Study Group for Pancreatic Cancer 1 (ESPAC-1) and the CONKO-001 trials, using 5-fluorouracil (5-FU)/leucovorin and gemcitabine respectively[7-9]. The phase III randomized PRODIGE 24 trial using 5-FU/leucovorin with irinotecan and oxaliplatin (FOLFIRINOX) and APACT trial with nab-paclitaxel with gemcitabine changed the landscape of adjuvant therapy following their success noticed in the metastatic setting [10,11]. The role of chemoradiotherapy (CRT) in the adjuvant setting is yet to see the final statement based on the existing literature. Following the lack of survival benefit with CRT in the ESPAC-1 and European Organization for Research and Treatment of Cancer (EORTC) trials and contrasting results with two registry data showing survival benefit of CRT compared to CT, one large series compared the three modalities of systemic CT, CRT or CRT followed by CT. There was a significant survival benefit with CT and CRT followed by CT than in the CRT in patients with stage III disease. This benefit was however not seen in patients with stage I/II disease[12-16].

The management of borderline resectable pancreatic adenocarcinomas (BRPCs) has seen the emergence of adjuvant regimes in the 'neoadjuvant' or 'induction therapy' role with FOLFIRINOX and nab-paclitaxel with gemcitabine[17,18]. There is no robust data to suggest the survival benefit of these protocols so far; however, there has been demonstrable tolerability and increased resection rates. A clinical challenge has been to offer adjuvant therapy to patients receiving induction therapy. The results of phase II ESPAC-5F trial, presented at the 2020 virtual ASCO meeting, comparing four armsfrontline surgery, induction therapy with gemcitabine and capecitabine, or modified FOLFIRINOX and CRT. The study revealed similar outcomes between the frontline and induction treatment[19].

The metastatic setting is seeing numerous trials with conventional CT as well as targeted agents as the biology and tumor microenvironment, genetics, and molecular concepts are being better understood. This has led to a search for novel therapeutic strategies for managing PC. This review attempts to address the challenge faced by the practicing clinician in optimal sequencing of the available modalities in the various stages of the illness.



MANAGEMENT OF RESECTABLE PANCREATIC CANCER

A resectable adenocarcinoma does not have metastases to a distant organ or distant lymph nodes; there is no vascular involvement [characterized by absence of superior mesenteric vein (SMV) or portal vein (PV) involvement], tumor thrombosis, or venous encasement $> 180^{\circ}$. Also, the fat planes around the celiac axis (CA), hepatic artery (HA), and superior mesenteric artery (SMA) ought to be clear^[20].

Surgery

Surgery is the only treatment option that offers a cure for PDAC. Surgery aims to completely resect the tumor and achieve a microscopically negative tumor margin (R0). R0 dissection is defined as clearance of > 1 mm *i.e.*, the margin of healthy tissue around the removed tumor should be > 1 mm. The various surgical options include pancreaticoduodenectomy (Whipple's procedure) and distal pancreatectomy. Pancreaticoduodenectomy with SMA first approach is the standard of care for adenocarcinoma localized to the head of the pancreas (HOP). The surgery should involve dissection of greater than 15 lymph nodes and skeletonization of SMA down to adventitia of anterior, left lateral and posterior borders[21,22]. A sampling of paraaortic lymph nodes with an examination of the frozen section is an additional option. For PDAC involving the body and tail of the pancreas, distal pancreatectomy along with splenectomy is the treatment option. This involves dissection of greater than 15 lymph nodes[23,24].

Minimally invasive techniques for pancreatic resection beginning with laparoscopic distal pancreatectomy have been attempted. They offer advantages in the form of reduced blood loss and decreased hospital stay. However, the rate of achieving positive resection margin, morbidity, and mortality of the procedure remains the same as that of an open procedure. The use of robotic techniques in Whipple's procedure has shown reduced rates of post-procedure complications[25]. Traditionally preoperative biliary drainage has been advised for patients who present with obstructive jaundice. Recent evidence, however, points towards a higher rate of perioperative complications among those undergoing pre-operative drainage vs those undergoing upfront surgery^[26].

The risk of developing tumor recurrence among those patients who undergo curative resection for PC varies from 69%-75% at 2 years to 80%-90% at 5 years postsurgery^[27]. Tumor recurrence occurs secondary to locoregional occurrence in a majority of cases. This led to the hypothesis that the use of adjuvant therapy may reduce locoregional tumor recurrence.

Post-operative complications may reduce a patients' access to adjuvant CT and overall survival. Hence, it becomes imperative to screen patients who are at high risk of post-operative complications, like elderly patients, patients with poor performance status, or higher comorbidity profiles. Preoperative pancreatic resection score (PREPARE) and Surgical results analysis and search (SOAR) are validated and useful scoring systems for assessing the risk of developing complications post-operatively [28, 29].

Adjuvant CT

The gold standard treatment for resectable PC is surgery followed by adjuvant CT. The era of adjuvant CT gained prominence when the results of the European Group for Pancreatic Cancer (ESPAC-1) trial showed significant improved median survival and 5-year survival in patients who received adjuvant CT of fluorouracil and folinic acid vs those who underwent surgery alone (20.1 mo vs 15.5 mo, respectively; P = 0.009). This was followed by the CONKO-001 (Charité Onkologietrial) trial, using adjuvant gemcitabine, which showed a median disease-free survival of 13.4 mo and 5-year survival of 20.7% vs 10.4% vs 6.9 mo respectively in the surgery alone group. The efficacy of these two treatment regimens was compared in the ESPAC-3 trial. Results of this study showed no survival benefit of one treatment regimen over the other, however, the treatment-related adverse effects were higher in the fluorouracil and folinic acid group.

To further improve the therapeutic outcome with adjuvant CT, a concept of combination systemic therapy has evolved. Several agents have been studied in various trials (Table 1). Among those of note are the ESPAC-4, PRODIGE and APACT studies. The ESPAC-4 trial carried out a comparison of gemcitabine vs a combination of gemcitabine plus capecitabine, which showed favorable overall survival benefit while using combination therapy [hazard ratio (HR): 0.82, 95% confidence interval (CI): 0.68-0.98; *P* = 0.032]. However, no significant recurrence-free survival benefit was seen in 2 years of follow-up of these patients (HR: 0.86, 95%CI: 0.73-1.02; *P* = 0.082).



Table 1 Landmark trials on adjuvant treatment in pancreatic adenocarcinoma					
Study	No. of patients	Treatment arms	Median DFS in mo	Median OS in mo	
GITSG[33]	43	Observation	NR	20.0	
		Radiotherapy + 5-FU f/b adjuvant 5-FU	NR	10.9	
ESPAC-1[34]	289	Observation	NR	15.5	
		Chemoradiotherapy	NR	13.9	
		5-FU/folinic acid	NR	20.1	
		Chemoradiotherapy + 5-FU/folinic acid	NR	19.9	
CONKO-001[9]	354	Observation	6.7	20.2	
		Gemcitabine	13.4	22.8	
ESPAC-3[35]	1088	5-FU/folinic acid	14.1	23.0	
		Gemcitabine	14.3	23.6	
ESPAC-4[36]	730	Gemcitabine	13.1	25.5	
		Gemcitabine + Capecitabine	13.9	28.0	
CONKO-005[30]	436	Gemcitabine	11.4	26.5	
		Gemcitabine + Erlotinib	11.4	24.6	
PRODIGE 24-PA6[37]	493	Gemcitabine	12.8	35.0	
		FOLFIRINOX	21.6	54.4	
APACT[11]	866	Gemcitabine	18.8	36.2	
		Gemcitabine + nab-paclitaxel	19.4	40.5	

5-FU: 5-Fluorouracil; CONKO: Charité Onkologie; DFS: Disease-free survival; ESPAC: European Group for Pancreatic Cancer; GITSG: Gastrointestinal Tumor Study Group; NR: Not reported; OS: Overall survival; PRODIGE: Partenariat de Recherche en Oncologie DIGEstive.

> The Partenariat de Recherche en Oncologie Digestive (PRODIGE 24-PA6) trial highlighted the successful use of FOLFIRINOX (oxaliplatin + irinotecan + leucovorin) vs gemcitabine in patients with good performance status (Eastern Cooperative Oncology Group, ECOG: 0-1). The median disease-free survival in patients with combination therapy was 21.6 mo vs 12.8 mo in the gemcitabine group. The latest in series is the Nab-paclitaxel and Gemcitabine vs Gemcitabine Alone as Adjuvant Therapy for Patients with Resected Pancreatic Cancer (APACT) study, which has shown encouraging results of using combination therapy of gemcitabine plus nanoparticle albumin-bound paclitaxel (nab-paclitaxel). Median disease-free survival was not statistically significant in the two arms (19.4 mo in combination arm vs 18.8 mo in gemcitabine only arm). Overall survival favored the combination CT group (HR: 0.82, 95% CI: 0.68-0.996; P = 0.045)[27].

> Another concept is the use of targeted agents (erlotinib or sorafenib) or immunotherapy (algenpantucel-L) in combination with gemcitabine. However, to date, none of these agents have shown any favorable survival outcome vs the use of gemcitabine alone[30-32].

Adjuvant CRT

Adjuvant CRT aims to prevent locoregional tumor recurrence. Amongst the first trials to assess the efficacy of CRT in PC was the EORTC trial, which showed no significant survival benefit amongst patients of PDAC who received CRT (40 Gy + continuous infusion of fluorouracil)[13]. This was followed by the ESPAC-1 trial, which employed the following three different adjuvant therapy designs: CRT, CT alone, and CRT followed by adjuvant CT. Patients undergoing adjuvant CT were found to have poor survival benefits after a median follow-up of 47 mo (HR: 1.28, 95%CI: 0.99-1.66; P = 0.05)[34]. The RTOG 9704 trial was carried out to assess the benefit of adding gemcitabine to postoperative radiation + fluorouracil vs adjuvant therapy with fluorouracil. The trial showed no survival benefit in either of the two treatment groups [38].

Although these trials failed to prove any survival benefit, they nevertheless provided useful information on the feasibility and tolerability of these treatment options. Analysis of the US National Cancer Database which included patients of pancreatic adenocarcinoma who underwent curative resection followed by adjuvant CT showed survival benefit (median overall survival with CT + radiation: 22.3 mo and adjuvant CT alone: 20.0 mo; P = 0.001)[14].

Timing of adjuvant CT

There are definite gaps in our knowledge regarding the optimal timing of initiating CT following surgery and follow-up of patients undergoing adjuvant therapy for curable PC. Delay in initiating as well as non-initiation of adjuvant treatment is not uncommon. Post-operative morbidity adversely impacts the initiation of adjuvant treatment. It has been estimated that approximately 20% of patients become ineligible for adjuvant CT. The ESPAC-3 trial aimed to analyze the outcome among those who were initiated on CT within 8 wk of surgery and those who were initiated on CT after 8 wk of surgery. There was no survival benefit in either of the two arms, while successful completion of six cycles of CT was found to be an independent predictor of survival^[39]. Similar results have been put forward by analyzing multi-institutional retrospective data of patients who had undergone curative resection for PDAC. Thus, the evidence so far suggests that patients who receive adjuvant therapy more than 12 wk following surgery are still good candidates for adjuvant CT[40].

Therefore, in summary, the standard of care in resectable pancreatic adenocarcinoma is surgical resection followed by adjuvant CT. Single-agent (5-FU) is preferred for periampullary tumors of pancreatic origin. Patients with adenocarcinoma in the head, body, or tail of the pancreas may be treated with FOLFIRINOX (patients with good post-operative performance status, ECOG: 0-1) and combination therapy of gemcitabine+ capecitabine in those with poor performance status postoperatively (Figure 1).

Neoadjuvant therapy

Conceptually, the use of neoadjuvant therapy in resectable PC gained prominence when this approach was shown to have improved survival benefit in other gastrointestinal malignancies^[41]. It offers the following theoretical advantages: reduction of circulating tumor cells and micrometastasis before surgery, and avoiding surgery in those who experience disease progression while on neoadjuvant therapy, thus, reducing surgical mortality and morbidity. Of particular interest is the ability of neoadjuvant treatment to achieve tumor downsizing or downstaging (lower T and N stages, reducing the rates of vascular, lymphatic, and perineural invasion, and the ability to achieve better R0 resection post-surgery). Moreover, individuals who receive neoadjuvant CT are more likely to be able to access the entire therapeutic sequence[42, 43]

The preoperative or postoperative CT for resectable pancreatic adenocarcinoma (PACT-15) trial has shown the efficacy of neoadjuvant CT in patients with resectable PDAC[44]. One of the first published phase III trials assessing the role of neoadjuvant gemcitabine followed by gemcitabine and radiation before surgery is the PREOPANC trial. Although it has not shown statistically significant survival benefit in patients using neoadjuvant CT + CRT (overall survival of 16 mo for neoadjuvant therapy vs 14.3 mo for initial surgery), better R0 resection rates, locoregional disease-free survival, and lower rates of vascular and perineural invasion favor neoadjuvant CT[45]. Analyses of 'US National Cancer Database data have shown a favorable survival using perioperative CT in patients with early PC as compared with upfront surgery. Another similar cohort study by Mokdad and colleagues^[46] demonstrated statistically significant median overall survival in the neoadjuvant group vis a vis upfront surgery group (26 mo vs 23 mo respectively; P = 0.01). Many other phase III trials are being conducted to have a better understanding of this therapeutic aspect[27] (Table 2). Neoadjuvant treatment is well tolerated. The risk of postoperative pancreatic fistula formation (3%-11%), risk of postoperative infections (3%-7%) and mortality (0%-4%) compared to those who have undergone surgery alone[47].

MANAGEMENT OF BRPC

The National Comprehensive Cancer Network has defined BRPC based on the following radiological criteria: Contrast-enhanced computerized tomographic (CECT) scan using the pancreatic protocol, the relationship of tumor with the surrounding



Table 2 Landmark trails on use of neoadjuvant chemotherapy							
Study	No. of patients	Treatment arms	Resection rate, %	R0	Median DFS in mo	Median OS in mo	
PACT-15[44]	93 Resectable	Surgery + gemcitabine	85	27	4.7	20.4	
		Surgery + 6 PEXG	90	37	12.4	26.4	
		3PEXG + surgery + 3 PEXG	84	63	16.9	16.9	
PREOPANC-01[48]	248 Resectable + BRPC	26Gy/15fr + gemcitabine	60	63	9.9	17.1	
		Surgery	72	31	7.9	13.7	

BRPC: Borderline resectable pancreatic cancer; DFS: Disease-free survival; OS: Overall survival; PEXG: Cisplatin, epirubicin, gemcitabine, and capecitabine.

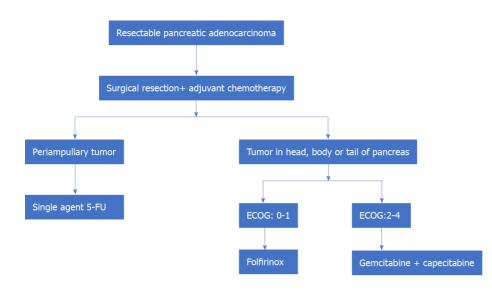


Figure 1 Management of resectable pancreatic adenocarcinoma. 5-FU: 5-Fluorouracil; ECOG: Eastern Cooperative Oncology Group.

vasculature, and the absence/presence of metastasis.

Thus, a BRPC localized to the HOP is one where the tumor is in contact with the common HA but without extension to the CA or artery bifurcation and variant arterial anatomy, contact with the SMA < 180°, contact with the SMV or PV < 180° without venous contour irregularity or thrombosis, which allows for safe and complete arterial and venous resection and reconstruction[20].

Borderline resectable PDAC in the pancreatic body or tail is 'the solid tumor in contact with the CA of $< 180^{\circ}$ or contact with the CA $> 180^{\circ}$ without the involvement of the aorta and gastroduodenal artery to allow a 'modified Appleby surgery' [20]. Another definition, the Anderson classification for BRPC, classifies it into the following three different groups: Group A includes patients with a tumor that abuts visceral arteries or causes short-segment occlusion of SMV; group B, have findings suggestive of metastasis; and group C patients are those who have marginal performance status[49].

Historically, therapeutic options for BRPC consist of upfront surgery, surgery followed by adjuvant CT/CRT, and neoadjuvant CT. The standard surgical options remain Whipple procedure, total pancreatectomy, or distal pancreatectomy, based on the tumor localization. An approach favoring upfront surgery carries with itself the risk of early failure, which has often been attributed to the poor pre-operative staging of tumor radiologically, inability to carry out a radical surgery, and the aggressive tumor behavior owing to variations in tumor biology. This has brought about an interest in considering neoadjuvant CT for patients with BRPC.

A meta-analysis carried out by the Dutch Pancreatic Cancer Group has shown a median survival of 19.2 mo in patients undergoing neoadjuvant CT vs 12.8 mo in those undergoing upfront surgery^[50]. A peculiar problem in comparing different groups arises when different CT/CRT regimens are being compared. A recent patient-level meta-analysis has analyzed the efficacy of neoadjuvant FOLFIRINOX in BRPC, finding a favorable trend using FOLFIRINOX (median overall survival of 22 mo and median disease progression-free survival of 18.0 mo)[18].



A recent randomized control trial has assessed the use of CRT as neoadjuvant therapy for BRPC. The investigators combined 54 Gy in 30 fractions + weekly gemcitabine followed by adjuvant gemcitabine. Results have shown a better resection rate in the neoadjuvant group *vs* the surgery only group (51.8% *vs* 26.1% respectively) as well as a statistically significant 2-year survival (40.7% *vs* 26%; *P* = 0.028)[51]. The preoperative CRT *vs* immediate surgery for resectable and borderline resectable pancreatic cancer (PREOPANC-1) trial has used intention-to-treat analysis to study the efficacy of neoadjuvant CRT (gemcitabine-based) *vs* upfront surgery in BRPC. Patients in the neoadjuvant CRT group had a lower resection rate (60%) *vs* those in the upfront surgery group (72%; *P* = 0.065). R0 resection rate was statistically higher in the neoadjuvant group *vs* the upfront surgery group (61% *vs* 31%; *P* < 0.001). To add to the benefits, patients receiving neoadjuvant CRT had a longer median time till recurrence *vs* the surgery only group (9.9 mo *vs* 7.9 mo; *P* = 0.023)[48].

Results of these trials have shown the efficacy of both neoadjuvant CT as well as for CRT in BRPC. Several ongoing trials are being carried out to study the efficacy of induction therapy in improving overall survival in BRPC and which of the two therapeutic strategies (CT/CRT) is better suited for the same.

MANAGEMENT OF LOCALLY ADVANCED PC

The concept of locally advanced PC (LAPC) has evolved based on the development of radiological criteria of resectability and the availability of neoadjuvant therapy. Practically, the pancreatic tumors which are not metastatic and are unresectable due to 'irreversible' vascular invasion (encasement of aorta, invasion of PV or SMV, involvement of the SMA or celiac trunk by > 180°) are considered as LAPC[52,53]. The median survival has been variably reported from 10 mo to 30 mo.

The standard treatment for LAPC is gemcitabine-based CT. For patients with good performance status (ECOG0-1), the FOLFIRINOX-based regimen and for those with ECOG0-2, the nab-paclitaxel-gemcitabine-based regimen may be considered[53]. Some observational studies and pooled analyses of different approaches have advocated induction treatment with either FOLFIRINOX or nab-paclitaxel-gemcitabine followed by CRT. Using this approach median survival of 24.2 mo and disease progression-free survival of 15 mo has been reported[54]. However, the use of either upfront radiation therapy or following induction treatment with gemcitabine is not beneficial for patients with LAPC. Recently published meta-analysis has reported similar overall survival and drug-related side effects of CRT and CT in the setting of LAPC[55]. CRT using capecitabine as a radiosensitizer, however, has been tried for improvement of local control of disease in a subset of patients with LAPC. Since this is not a standard treatment approach, it has been proposed that this may be offered to only a select group of individuals[56].

Ongoing trials in LAPC are trying to assess the efficacy of FOLFIRINOC *vs* gemcitabine as induction therapy (NEOPAN; NCT02539537), use of nab-paclitaxel-gemcitabine for induction regimen + radiation therapy *vs* continuous CT, and use of activation of the *DPC4* gene (RTOG 1201; NCT01921751).

An interesting recent concept that has emerged in the management of LAPC is the role of surgery. This was proposed by researchers from the Medical College of Wisconsin based on the observation that those with an initially unresectable disease as per radiological criteria may convert to the resectable tumor after induction $CT \pm CRT$. They have proposed the classification of patients with LAPC into the following two distinct categories: Type A, tumors that may be considered for resection following induction CT; and type B: Definitively unresectable tumors (> 270° encasement of the SMA, $> 180^{\circ}$ encasement of the CA, encasement of the aorta, and $> 180^{\circ}$ encasement of the HA with extension beyond the bifurcation of the proper HA into right and left HA) [52]. Thus, patients with LAPC type A, who have completed induction CT should be considered for surgical exploration. This is especially pertinent as the specificity of a CECT scan to determine tumor staging, operability, and R0 resectability for HOP carcinoma decrease following induction therapy. In such a setting, surgical exploration is the ideal modality to prove/ rule out vascular involvement[57]. Resected patients who have received CT preoperatively can also be considered for an additional course of CT following surgery[58] (Figure 2).

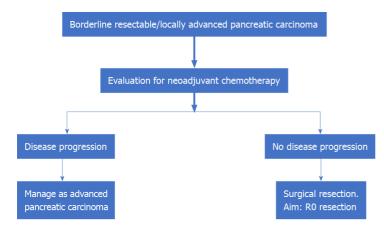


Figure 2 Management of borderline resectable/locally advance pancreatic carcinoma.

MANAGEMENT OF ADVANCED METASTATIC PC

CT forms the backbone of therapeutic regimens for the management of metastatic PC. Till the advent of gemcitabine, fluorouracil was the only approved drug in the management of APC. The use of gemcitabine has brought about benefits of disease progression-free survival, and overall survival (from 4.4 mo to 5.7 mo, P = 0.0025) with similar drug-related side effects as compared to fluorouracil^[59]. This was followed by trials that analyzed the survival benefit of adding another cytotoxic drug or targeted therapy to gemcitabine. Use of erlotinib to gemcitabine-based regimen has shown median overall survival benefit (5.91 mo to 6.24 mo, P = 0.038) and 1-year survival benefit (17% to 23%, P = 0.023)[60]. A combination regimen consisting of cisplatin, epirubicin, fluorouracil, and gemcitabine (PEFG) vs gemcitabine has shown 'fourmonth progression-free survival' of 60% vs 28% in gemcitabine alone arm, P = 0.001with no difference in overall survival [44]. A combination of gemcitabine + nabpaclitaxel in patients with ECOG0-2 has shown an improvement of 'median overall survival' vs gemcitabine alone (8.5 mo vs 6.7 mo, P < 0.001). The PRODIGE 4-ACCORD11 trial is a landmark trial that has shown the overall survival benefit of using FOLFIRINOX (median survival: 11.1 mo) in patients with APC vs gemcitabine (6.8 mo, P < 0.001) [10]. FOLFIRINOX has not been compared to nab-paclitaxel + gemcitabine in any prospective trial to date. Thus, the FOLFIRINOX regimen is now the standard of care for patients of APC with ECOG: 0-1, normal serum bilirubin, and no underlying cardiac pathology. The limiting step in the FOLFIRINOX regimen happens to be the performance status of the individual and comorbidity profile with the elderly or low-profile patients likely to have a poorer outcome. Other prognostic factors are the number of metastases, and liver metastases, presence of genetic mutations such as DNA damage response (DDR) gene mutations and BRCA tumor suppressor gene mutations[61-63]. Modified FOLFIRINOX has been tried with similar efficacy and better tolerance profile as compared to the standard FOLFIRINOX regimen. This regimen includes fluorouracil bolus suppression or a dose reduction of irinotecan (or both)[64].

Progress of disease in patients with first-line CT regimens presents a particular challenge with around 50% of patients being eligible for second-line CT[65]. Combination regimens like gemcitabine-platinum and fluoropyrimidine-platinum have shown disease progression-free survival of 2.5 mo vs 1.9 mo for single agents (P =0.169) but no improvement in overall survival (5.1 mo vs 4.3 mo, P = 0.169)[66]. The various combination regimens that have been tried on the failure of first- and secondline regimens are oxaliplatin, folinic acid, and 5-FU (OFF regimen), and nanoliposomal irinotecan (MM-398), 5-FU, and folinic acid regimen. The CONKO-003 trial has shown overall survival benefit of using the OFF regimen over the 5-FU-folinic acid (FF regimen); the median survival was 5.9 mo vs 3.3 mo respectively (P = 0.01)[67]. The disease progression-free survival with use of nanoliposomal irinotecan (MM-398), 5-FU, and folinic acid regimen after failure of the FOLFIRINOX regimen has been reported to be 5.1 mo with overall survival of 8.8 mo[68]. The results of using targeted agents, namely the Jak1 and Jak2 tyrosine kinase inhibitor ruxolitinib and glufosfamide, have been rather disappointing. However, patients with metastatic solid tumors (including 8 patients with pancreatic tumors), with deficient mismatch repair and failed first-line therapy have shown response to the PD-1 immune checkpoint



inhibitor (ICI) pembrolizumab (disease control rate: 77%, objective response: 53% and complete radiological recovery: 21%)[69]. Mutation in the BRCA gene has been reported in around 5% of patients with APC. Targeted therapy in patients with BRCA gene mutation using 'poly ADP-ribose polymerase (PARP) inhibitors' is being actively investigated[70] (Figure 3).

Palliative treatment

Palliative treatment aims to allay patients' symptoms and improve their quality of life. Pain management, symptomatic relief, and psychological support are the pillars of this strategy.

Gastric outlet obstruction (GOO), extrahepatic biliary obstruction (EHBO), and abdominal pain are the three most common disabling symptoms in APC which adversely affect an individual's quality of life besides being a major source of 'caregiver fatigue'. GOO, presenting as nausea, vomiting, dehydration, and weight loss, is seen in 10%-25% of all APC cases. Palliative surgery with open gastrojejunostomy (GJ) is the traditional approach to managing a malignant GOO. Placement of endoscopic duodenal stents and laparoscopic GJ has been tried, with varying degrees of success. Surgical procedures offer good functional outcomes at the cost of increased mortality[71]. EHBO can present with obstructive jaundice. Endoscopic retrograde cholangiopancreatography (ERCP)-guided biliary stent placement is the accepted gold standard approach for the management of malignant EHBO. Both plastic and selfexpanding metal stents (SEMSs) have been used, with literature favoring the use of covered SEMSs. Failure of ERCP-guided biliary drainage may warrant drainage through the percutaneous route or an endoscopic ultrasonography-guided biliary drainage (EUS BD)[72]. Hepaticojejunostomy with a Roux-en-Y reconstruction and cholecystectomy is the favored palliative surgical procedure for palliation of EHBO secondary to PC. Laparoscopic biliary bypass and robot-assisted laparoscopic hepaticojejunostomy have been technically successful with satisfactory surgical outcomes[73,74]. Thus, patients with good performance status may benefit from a palliative surgical procedure while those with poor performance status warrant endoscopic biliary drainage.

Malignant infiltration of celiac or mesenteric nerve plexus in patients of APC may cause abdominal and back pain. This may adversely affect an individual's quality of life. Multimodal drug therapy encompassing the use of non-steroidal anti-inflammatory drugs and or opioid analgesics in various combinations form the bedrock of pain management. Intraoperative celiac block using ethanol or a local anesthetic using either laparoscopic or open approach and EUS-guided neurolysis of celiac plexus, have all been tried[71] (Figure 4).

NEWER MODALITIES FOR PANCREATIC CANCER

Immunotherapy for pancreatic cancer

The tumor microenvironment of PC is responsible for aggressiveness as well as chemoresistance. This makes the case for utilizing immunotherapy in the advanced and metastatic settings. However, the approval for ICI is currently for patients with mismatch repair-deficient cases[69]. The updated results of the Keynote-158, using pembrolizumab in patients with advanced PDAC revealed an overall response rate (ORR) of 18.2%, median progression-free survival of 2.1 mo, and median overall survival of 4.0 mo[75]. The results are not very encouraging despite this, due to fewer driver mutations, variable expression of PD-1/PD-L1 and neoantigen burden in the tumor tissue, and absence of DDR, which are hallmarks of this malignancy [76].

Targeted therapy

The highly actionable mutations detected in molecular profiling of the Know Your Tumor initiative were 27%, of which the common ones involved the KRAS, TP53, MLL3, CDKN2A, SMAD4, TGFBR2, ARID1A, and SF3B1 genes. These mutations, however, do not have any therapeutic modality to target[77,78]. Neurotrophic receptor tyrosine kinase (NTRK) gene fusions have been detected in about 6% of pancreatic adenocarcinomas. Larotrectinib and entrectinib are two newer agents receiving accelerated approval for use in metastatic tumors with NTRK gene biomarker with tissue agnostic indication[79,80]. Three single-arm trials, namely LOXO-TRK-14001, SCOUT and NAVIGATE, had a total of 55 patients with NTRK fusions and had an ORR of 55% with Larotrectinib[81]. The median progression-free survival was not achieved after a median duration of 9.9 mo. Similarly, in three other single-arm trials,



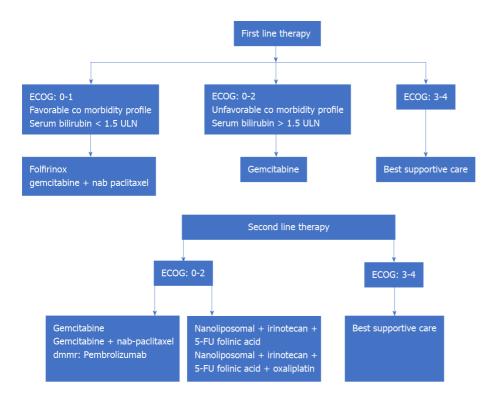


Figure 3 Management of advanced metastatic pancreatic carcinoma. ECOG: Eastern Cooperative Oncology Group; ULN: Upper limit of normal.

including ALKA-372-001, STARTRK-1 and STARTRK-2, with 60 patients in total, the patients with *NTRK* fusion had an ORR of 100%[82].

Germline *BRCA1/2* mutations and homologous recombination deficiency enable the utility of platinum agents as well as poly (adenosine diphosphate-ribose) polymerase (PARP) inhibition as a novel therapeutic modality. Olaparib, a PARP inhibitor was approved by the United States' Food and Drug Administration in December 2019 for patients with advanced metastatic malignancy having germline *BRCA1/2* mutation. The phase III POLO trial utilizing olaparib in PAC, progressing after platinum-based therapy revealed median progression-free survival of 7.4 mo *vs* 3.8 in the control arm (HR: 0.53, 95%CI: 0.35-0.82; *P* = 0.004). This did not translate into an overall survival benefit[83].

Macrophage-targeted therapy

Macrophages residing in the tumor environment are labeled as tumor-associated macrophages. CD 51 is a marker of macrophages that promotes the stemness of PDAC cells by regulating the TGF- β 1/smad2/3 pathway. As a result, CD 51- targeted therapy is evolving as a newer therapeutic modality. Similarly, CD 40 activation, which promotes anti-tumor T-cell responses has been targeted by using anti-CD 40 antibody, CP-870893 along with gencitabine, providing initial results of response[84].

Cancer vaccines

The ability of cancer vaccines to stimulate dendritic cell responses and activate the adaptive immune responses has been harnessed for many cancers, including PC. The expression of murine enzyme alpha-1,3-galactosyltransferase (alpha-GT) by genetic engineering on PC cell lines HAPa1 and HAPa2 lead to anti-alpha-Gal antibody responses in humans[85]. Algenpanteucel-L was used in a phase II study and phase III (IMPRESS Trial). When algenpanteucel-L was given after gemcitabine and 5-FU based CRT, 81%-86% 1-year disease-free survival and 96% 1-year overall survival were observed[86,87].

GVAX, a line of engineered pancreatic tumor cells secreting GM-CSF has been tested in phase I study along with cyclophosphamide (Cy) with early results of tolerability and survival. In a phase II study, GVAX/Cy was tested against a GVAX/Cy followed by CRS 207 (a live-attenuated *Listeria* strain that induces tumor-associated antigens) and resulted in better overall survival in the latter arm (6.1 mo *vs* 3.9 mo, P = 0.02)[88].

Manrai M et al. Current and emerging therapeutic strategies: Pancreatic cancer

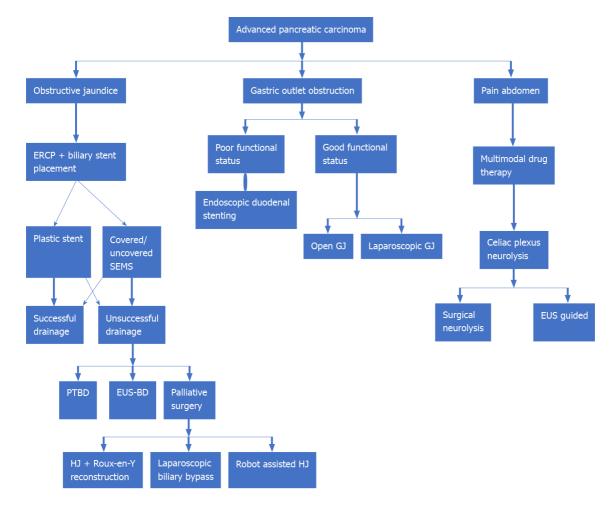


Figure 4 Palliative management in metastatic pancreatic carcinoma. ERCP: Endoscopic retrograde cholangiopancreatography; EUS: Endoscopic ultrasound; EUS-BD: Endoscopic ultrasound guided biliary drainage; GJ: Gastrojejunostomy; HJ: Hepaticojejunostomy; PTBD: Percutaneous transhepatic biliary drainage; SEMS: Self-expanding metal stent.

Other membranous and intracytoplasmic targets

There have been numerous trials using targets like vascular endothelial growth factor (VEGF) and VEGF-receptor, RAS-RAF-MEK-ERK pathway, etc. but none have shown robust survival data[89,90]. The rapamycin-insensitive companion of mTOR (RICTOR) expression was found to have a survival benefit in resected PAC patients, with those having a lower expression doubling the survival as compared to the high expressers [91].

CONCLUSION

We have attempted to provide an inclusive version of the management of PC, with special emphasis on current strategies and the road ahead with emerging modalities of therapy. Of course, there are certain gaps in the understanding of this disease and the evolution of treatment options is always challenging. For times to come, newer modalities appear promising; however, there is no substitute for early diagnosis and management for disease-free survival.

REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. CA Cancer J Clin 2019; 69: 7-34 [PMID: 1 30620402 DOI: 10.3322/caac.21551]
- 2 Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer 2015; 136: E359-E386 [PMID: 25220842 DOI: 10.1002/ijc.29210]
- 3 Bosetti C, Bertuccio P, Malvezzi M, Levi F, Chatenoud L, Negri E, La Vecchia C. Cancer mortality



in Europe, 2005-2009, and an overview of trends since 1980. Ann Oncol 2013; 24: 2657-2671 [PMID: 23921790 DOI: 10.1093/annonc/mdt301]

- 4 Bosetti C, Bertuccio P, Negri E, La Vecchia C, Zeegers MP, Boffetta P. Pancreatic cancer: overview of descriptive epidemiology. Mol Carcinog 2012; 51: 3-13 [PMID: 22162227 DOI: 10.1002/mc.20785]
- 5 Rozenblum E, Schutte M, Goggins M, Hahn SA, Panzer S, Zahurak M, Goodman SN, Sohn TA, Hruban RH, Yeo CJ, Kern SE. Tumor-suppressive pathways in pancreatic carcinoma. Cancer Res 1997; 57: 1731-1734 [PMID: 9135016]
- Moore PS, Orlandini S, Zamboni G, Capelli P, Rigaud G, Falconi M, Bassi C, Lemoine NR, Scarpa 6 A. Pancreatic tumours: molecular pathways implicated in ductal cancer are involved in ampullary but not in exocrine nonductal or endocrine tumorigenesis. Br J Cancer 2001; 84: 253-262 [PMID: 11161385 DOI: 10.1054/bjoc.2000.1567]
- 7 Oettle H, Post S, Neuhaus P, Gellert K, Langrehr J, Ridwelski K, Schramm H, Fahlke J, Zuelke C, Burkart C, Gutberlet K, Kettner E, Schmalenberg H, Weigang-Koehler K, Bechstein WO, Niedergethmann M, Schmidt-Wolf I, Roll L, Doerken B, Riess H. Adjuvant chemotherapy with gemcitabine vs observation in patients undergoing curative-intent resection of pancreatic cancer: a randomized controlled trial. JAMA 2007; 297: 267-277 [PMID: 17227978 DOI: 10.1001/jama.297.3.267
- Abdel-Rahman O, Elsayed Z, Elhalawani H. Gemcitabine-based chemotherapy for advanced biliary tract carcinomas. Cochrane Database Syst Rev 2018; 4: CD011746 [PMID: 29624208 DOI: 10.1002/14651858.CD011746.pub2]
- Oettle H, Neuhaus P, Hochhaus A, Hartmann JT, Gellert K, Ridwelski K, Niedergethmann M, Zülke C, Fahlke J, Arning MB, Sinn M, Hinke A, Riess H. Adjuvant chemotherapy with gemcitabine and long-term outcomes among patients with resected pancreatic cancer: the CONKO-001 randomized trial. JAMA 2013; 310: 1473-1481 [PMID: 24104372 DOI: 10.1001/jama.2013.279201]
- 10 Conroy T, Desseigne F, Ychou M, Bouché O, Guimbaud R, Bécouarn Y, Adenis A, Raoul JL, Gourgou-Bourgade S, de la Fouchardière C, Bennouna J, Bachet JB, Khemissa-Akouz F, Péré-Vergé D, Delbaldo C, Assenat E, Chauffert B, Michel P, Montoto-Grillot C, Ducreux M; Groupe Tumeurs Digestives of Unicancer; PRODIGE Intergroup. FOLFIRINOX vs genetiabine for metastatic pancreatic cancer. N Engl J Med 2011; 364: 1817-1825 [PMID: 21561347 DOI: 10.1056/NEJMoa1011923
- 11 Tempero MA, Reni M, Riess H, Pelzer U, O'Reilly EM, Winter JM, Oh DY, Li CP, Tortora G, Chang HM, Lopez CD, Tabernero J, Van Cutsem E, Philip PA, Goldstein D, Berlin J, Ferrara S, Li M, Lu BD, Biankin A. APACT: phase III, multicenter, international, open-label, randomized trial of adjuvant nab-paclitaxel plus gemcitabine (nab-P/G) vs gemcitabine (G) for surgically resected pancreatic adenocarcinoma. J Clin Oncol 2019; 37: 15
- 12 Neoptolemos JP, Stocken DD, Friess H, Bassi C, Dunn JA, Hickey H, Beger H, Fernandez-Cruz L, Dervenis C, Lacaine F, Falconi M, Pederzoli P, Pap A, Spooner D, Kerr DJ, Büchler MW; European Study Group for Pancreatic Cancer. A randomized trial of chemoradiotherapy and chemotherapy after resection of pancreatic cancer. N Engl J Med 2004; 350: 1200-1210 [PMID: 15028824 DOI: 10.1056/NEJMoa032295
- 13 Klinkenbijl JH, Jeekel J, Sahmoud T, van Pel R, Couvreur ML, Veenhof CH, Arnaud JP, Gonzalez DG, de Wit LT, Hennipman A, Wils J. Adjuvant radiotherapy and 5-fluorouracil after curative resection of cancer of the pancreas and periampullary region: phase III trial of the EORTC gastrointestinal tract cancer cooperative group. Ann Surg 1999; 230: 776-782; discussion 782 [PMID: 10615932 DOI: 10.1097/00000658-199912000-00006]
- 14 Rutter CE, Park HS, Corso CD, Lester-Coll NH, Mancini BR, Yeboa DN, Johung KL. Addition of radiotherapy to adjuvant chemotherapy is associated with improved overall survival in resected pancreatic adenocarcinoma: An analysis of the National Cancer Data Base. Cancer 2015; 121: 4141-4149 [PMID: 26280559 DOI: 10.1002/cncr.29652]
- Hsieh MC, Chang WW, Yu HH, Lu CY, Chang CL, Chow JM, Chen SU, Cheng Y, Wu SY. 15 Adjuvant radiotherapy and chemotherapy improve survival in patients with pancreatic adenocarcinoma receiving surgery: adjuvant chemotherapy alone is insufficient in the era of intensity modulation radiation therapy. Cancer Med 2018; 7: 2328-2338 [PMID: 29665327 DOI: 10.1002/cam4.1479
- You MS, Ryu JK, Huh G, Chun JW, Paik WH, Lee SH, Kim YT. Comparison of efficacy between adjuvant chemotherapy and chemoradiation therapy for pancreatic cancer: AJCC stage-based approach. World J Clin Oncol 2020; 11: 747-760 [PMID: 33033696 DOI: 10.5306/wjco.v11.i9.747]
- 17 Cascinu S, Berardi R, Bianco R, Bilancia D, Zaniboni A, Ferrari D, Mosconi S, Spallanzani A, Cavanna L, Leo S, Negri F, Beretta GD, Sobrero A, Banzi M, Morabito A, Bittoni A, Marciano R, Ferrara D, Noventa S, Piccirillo MC. Nab-paclitaxel (Nab) plus gemcitabine (G) is more effective than G alone in locally advanced, unresectable pancreatic cancer (LAUPC): The GAP trial, a GISCAD phase II comparative randomized trial. Ann Oncol 2019; 30: v253-v254 [DOI: 10.1093/annonc/mdz247.001]
- Janssen QP, Buettner S, Suker M, Beumer BR, Addeo P, Bachellier P, Bahary N, Bekaii-Saab T, 18 Bali MA, Besselink MG, Boone BA, Chau I, Clarke S, Dillhoff M, El-Rayes BF, Frakes JM, Grose D, Hosein PJ, Jamieson NB, Javed AA, Khan K, Kim KP, Kim SC, Kim SS, Ko AH, Lacy J, Margonis GA, McCarter MD, McKay CJ, Mellon EA, Moorcraft SY, Okada KI, Paniccia A, Parikh PJ, Peters NA, Rabl H, Samra J, Tinchon C, van Tienhoven G, van Veldhuisen E, Wang-Gillam A, Weiss MJ,



Wilmink JW, Yamaue H, Homs MYV, van Eijck CHJ, Katz MHG, Groot Koerkamp B. Neoadjuvant FOLFIRINOX in Patients With Borderline Resectable Pancreatic Cancer: A Systematic Review and Patient-Level Meta-Analysis. J Natl Cancer Inst 2019; 111: 782-794 [PMID: 31086963 DOI: 10.1093/jnci/djz073]

- 19 Ghaneh P, Palmer HP, Cicconi S, Halloran C, Psarelli EE, Rawcliffe CL, Sripadam R, Mukherjee S, Wadsley J, Al-Mukhtar A, Jiao LR, Wasan HS, Carter R, Graham JS, Ammad F, Evans J, Tjaden C, Hackert T, Buchler MW, Neoptolemos JP; European Study Group for Pancreatic Cancer (ESPAC). ESPAC-5F: Four-arm, prospective, multicenter, international randomized phase II trial of immediate surgery compared with neoadjuvant gemcitabine plus capecitabine (GEMCAP) or FOLFIRINOX or chemoradiotherapy (CRT) in patients with borderline resectable pancreatic cancer. J Clin Oncol 2020; 15: 4505 [DOI: 10.1200/JCO.2020.38.15 suppl.4505]
- Tempero MA, Malafa MP, Chiorean EG, Czito B, Scaife C, Narang AK, Fountzilas C, Wolpin BM, 20 Al-Hawary M, Asbun H, Behrman SW, Benson AB, Binder E, Cardin DB, Cha C, Chung V, Dillhoff M, Dotan E, Ferrone CR, Fisher G, Hardacre J, Hawkins WG, Ko AH, LoConte N, Lowy AM, Moravek C, Nakakura EK, O'Reilly EM, Obando J, Reddy S, Thayer S, Wolff RA, Burns JL, Zuccarino-Catania G. Pancreatic Adenocarcinoma, Version 1.2019. J Natl Compr Canc Netw 2019; 17: 202-210 [PMID: 30865919 DOI: 10.6004/jnccn.2019.0014]
- Strasberg SM, Linehan DC, Hawkins WG. Radical antegrade modular pancreatosplenectomy 21 procedure for adenocarcinoma of the body and tail of the pancreas: ability to obtain negative tangential margins. J Am Coll Surg 2007; 204: 244-249 [PMID: 17254928 DOI: 10.1016/i.jamcollsurg.2006.11.002]
- 22 Malleo G, Maggino L, Ferrone CR, Marchegiani G, Mino-Kenudson M, Capelli P, Rusev B, Lillemoe KD, Bassi C, Fernàndez-Del Castillo C, Salvia R. Number of Examined Lymph Nodes and Nodal Status Assessment in Distal Pancreatectomy for Body/Tail Ductal Adenocarcinoma. Ann Surg 2019; 270: 1138-1146 [PMID: 29672406 DOI: 10.1097/SLA.00000000002781]
- Delpero JR, Bachellier P, Regenet N, Le Treut YP, Paye F, Carrere N, Sauvanet A, Autret A, Turrini 23 O, Monges-Ranchin G, Boher JM. Pancreaticoduodenectomy for pancreatic ductal adenocarcinoma: a French multicentre prospective evaluation of resection margins in 150 evaluable specimens. HPB (Oxford) 2014; 16: 20-33 [PMID: 23464850 DOI: 10.1111/hpb.12061]
- 24 Negoi I, Hostiuc S, Runcanu A, Negoi RI, Beuran M. Superior mesenteric artery first approach vs standard pancreaticoduodenectomy: a systematic review and meta-analysis. Hepatobiliary Pancreat Dis Int 2017; 16: 127-138 [PMID: 28381375 DOI: 10.1016/s1499-3872(16)60134-0]
- McGuigan A, Kelly P, Turkington RC, Jones C, Coleman HG, McCain RS. Pancreatic cancer: A 25 review of clinical diagnosis, epidemiology, treatment and outcomes. World J Gastroenterol 2018; 24: 4846-4861 [PMID: 30487695 DOI: 10.3748/wjg.v24.i43.4846]
- van der Gaag NA, Rauws EA, van Eijck CH, Bruno MJ, van der Harst E, Kubben FJ, Gerritsen JJ, 26 Greve JW, Gerhards MF, de Hingh IH, Klinkenbijl JH, Nio CY, de Castro SM, Busch OR, van Gulik TM, Bossuyt PM, Gouma DJ. Preoperative biliary drainage for cancer of the head of the pancreas. NEngl J Med 2010; 362: 129-137 [PMID: 20071702 DOI: 10.1056/NEJMoa0903230]
- Lambert A, Schwarz L, Borbath I, Henry A, Van Laethem JL, Malka D, Ducreux M, Conroy T. An 27 update on treatment options for pancreatic adenocarcinoma. Ther Adv Med Oncol 2019; 11: 1758835919875568 [PMID: 31598142 DOI: 10.1177/1758835919875568]
- Uzunoglu FG, Reeh M, Vettorazzi E, Ruschke T, Hannah P, Nentwich MF, Vashist YK, Bogoevski 28 D, König A, Janot M, Gavazzi F, Zerbi A, Todaro V, Malleo G, Uhl W, Montorsi M, Bassi C, Izbicki JR, Bockhorn M. Preoperative Pancreatic Resection (PREPARE) score: a prospective multicenterbased morbidity risk score. Ann Surg 2014; 260: 857-863; discussion 863 [PMID: 25243549 DOI: 10.1097/SLA.000000000000946
- Ragulin-Coyne E, Carroll JE, Smith JK, Witkowski ER, Ng SC, Shah SA, Zhou Z, Tseng JF. 29 Perioperative mortality after pancreatectomy: a risk score to aid decision-making. Surgery 2012; 152: S120-S127 [PMID: 22766367 DOI: 10.1016/j.surg.2012.05.018]
- Sinn M, Bahra M, Liersch T, Gellert K, Messmann H, Bechstein W, Waldschmidt D, Jacobasch L, 30 Wilhelm M, Rau BM, Grützmann R, Weinmann A, Maschmeyer G, Pelzer U, Stieler JM, Striefler JK, Ghadimi M, Bischoff S, Dörken B, Oettle H, Riess H. CONKO-005: Adjuvant Chemotherapy With Gemcitabine Plus Erlotinib Versus Gemcitabine Alone in Patients After R0 Resection of Pancreatic Cancer: A Multicenter Randomized Phase III Trial. J Clin Oncol 2017; 35: 3330-3337 [PMID: 28817370 DOI: 10.1200/JCO.2017.72.6463]
- Sinn M, Liersch T, Gellert K, Riess H, Stübs P, Waldschmidt DT, Pelzer U, Stieler J, Striefler JK, Bahra M, Dörken B, Oettle H. LBA18 - Conko-006: a Randomized Double-Blinded Phase Iib-Study of Adjuvant Therapy with Gemcitabine + Sorafenib/Placebo for Patients with R1-Resection of Pancreatic Cancer. Ann Oncol 2014; 25: v1 [DOI: 10.1093/annonc/mdu438.18]
- Yoshitomi H, Togawa A, Kimura F, Ito H, Shimizu H, Yoshidome H, Otsuka M, Kato A, Nozawa S, 32 Furukawa K, Miyazaki M; Pancreatic Cancer Chemotherapy Program of the Chiba University Department of General Surgery Affiliated Hospital Group. A randomized phase II trial of adjuvant chemotherapy with uracil/tegafur and gemcitabine vs gemcitabine alone in patients with resected pancreatic cancer. Cancer 2008; 113: 2448-2456 [PMID: 18823024 DOI: 10.1002/cncr.23863]
- Further evidence of effective adjuvant combined radiation and chemotherapy following curative 33 resection of pancreatic cancer. Gastrointestinal Tumor Study Group. Cancer 1987; 59: 2006-2010 [PMID: 3567862 DOI: 10.1002/1097-0142(19870615)59:12<2006::aid-cncr2820591206>3.0.co;2-b]
- Neoptolemos JP, Dunn JA, Stocken DD, Almond J, Link K, Beger H, Bassi C, Falconi M, Pederzoli 34



P, Dervenis C, Fernandez-Cruz L, Lacaine F, Pap A, Spooner D, Kerr DJ, Friess H, Büchler MW; European Study Group for Pancreatic Cancer. Adjuvant chemoradiotherapy and chemotherapy in resectable pancreatic cancer: a randomised controlled trial. Lancet 2001; 358: 1576-1585 [PMID: 11716884 DOI: 10.1016/s0140-6736(01)06651-x]

- Neoptolemos JP, Stocken DD, Bassi C, Ghaneh P, Cunningham D, Goldstein D, Padbury R, Moore 35 MJ, Gallinger S, Mariette C, Wente MN, Izbicki JR, Friess H, Lerch MM, Dervenis C, Oláh A, Butturini G, Doi R, Lind PA, Smith D, Valle JW, Palmer DH, Buckels JA, Thompson J, McKay CJ, Rawcliffe CL, Büchler MW; European Study Group for Pancreatic Cancer. Adjuvant chemotherapy with fluorouracil plus folinic acid vs gemcitabine following pancreatic cancer resection: a randomized controlled trial. JAMA 2010; 304: 1073-1081 [PMID: 20823433 DOI: 10.1001/jama.2010.1275]
- 36 Neoptolemos JP, Palmer DH, Ghaneh P, Psarelli EE, Valle JW, Halloran CM, Faluyi O, O'Reilly DA, Cunningham D, Wadsley J, Darby S, Meyer T, Gillmore R, Anthoney A, Lind P, Glimelius B, Falk S, Izbicki JR, Middleton GW, Cummins S, Ross PJ, Wasan H, McDonald A, Crosby T, Ma YT, Patel K, Sherriff D, Soomal R, Borg D, Sothi S, Hammel P, Hackert T, Jackson R, Büchler MW; European Study Group for Pancreatic Cancer. Comparison of adjuvant gemcitabine and capecitabine with gemcitabine monotherapy in patients with resected pancreatic cancer (ESPAC-4): a multicentre, open-label, randomised, phase 3 trial. Lancet 2017; 389: 1011-1024 [PMID: 28129987 DOI: 10.1016/S0140-6736(16)32409-6]
- Conroy T, Hammel P, Hebbar M, Ben Abdelghani M, Wei AC, Raoul JL, Choné L, Francois E, Artru P, Biagi JJ, Lecomte T, Assenat E, Faroux R, Ychou M, Volet J, Sauvanet A, Breysacher G, Di Fiore F, Cripps C, Kavan P, Texereau P, Bouhier-Leporrier K, Khemissa-Akouz F, Legoux JL, Juzyna B, Gourgou S, O'Callaghan CJ, Jouffroy-Zeller C, Rat P, Malka D, Castan F, Bachet JB; Canadian Cancer Trials Group and the Unicancer-GI-PRODIGE Group. FOLFIRINOX or Gemcitabine as Adjuvant Therapy for Pancreatic Cancer. N Engl J Med 2018; 379: 2395-2406 [PMID: 30575490 DOI: 10.1056/NEJMoa1809775]
- Regine WF, Winter KA, Abrams RA, Safran H, Hoffman JP, Konski A, Benson AB, Macdonald JS, 38 Kudrimoti MR, Fromm ML, Haddock MG, Schaefer P, Willett CG, Rich TA. Fluorouracil vs gemcitabine chemotherapy before and after fluorouracil-based chemoradiation following resection of pancreatic adenocarcinoma: a randomized controlled trial. JAMA 2008; 299: 1019-1026 [PMID: 18319412 DOI: 10.1001/jama.299.9.1019]
- 39 Valle JW, Palmer D, Jackson R, Cox T, Neoptolemos JP, Ghaneh P, Rawcliffe CL, Bassi C, Stocken DD, Cunningham D, O'Reilly D, Goldstein D, Robinson BA, Karapetis C, Scarfe A, Lacaine F, Sand J, Izbicki JR, Mayerle J, Dervenis C, Oláh A, Butturini G, Lind PA, Middleton MR, Anthoney A, Sumpter K, Carter R, Büchler MW. Optimal duration and timing of adjuvant chemotherapy after definitive surgery for ductal adenocarcinoma of the pancreas: ongoing lessons from the ESPAC-3 study. J Clin Oncol 2014; 32: 504-512 [PMID: 24419109 DOI: 10.1200/JCO.2013.50.7657]
- 40 Xia BT, Ahmad SA, Al Humaidi AH, Hanseman DJ, Ethun CG, Maithel SK, Kooby DA, Salem A, Cho CS, Weber SM, Stocker SJ, Talamonti MS, Bentrem DJ, Abbott DE. Time to Initiation of Adjuvant Chemotherapy in Pancreas Cancer: A Multi-Institutional Experience. Ann Surg Oncol 2017; 24: 2770-2776 [PMID: 28600732 DOI: 10.1245/s10434-017-5918-z]
- 41 Newton AD, Datta J, Loaiza-Bonilla A, Karakousis GC, Roses RE. Neoadjuvant therapy for gastric cancer: current evidence and future directions. J Gastrointest Oncol 2015; 6: 534-543 [PMID: 26487948 DOI: 10.3978/j.issn.2078-6891.2015.047]
- Gillen S, Schuster T, Meyer Zum Büschenfelde C, Friess H, Kleeff J. Preoperative/neoadjuvant 42 therapy in pancreatic cancer: a systematic review and meta-analysis of response and resection percentages. PLoS Med 2010; 7: e1000267 [PMID: 20422030 DOI: 10.1371/journal.pmed.1000267]
- 43 Schorn S, Demir IE, Reyes CM, Saricaoglu C, Samm N, Schirren R, Tieftrunk E, Hartmann D, Friess H, Ceyhan GO. The impact of neoadjuvant therapy on the histopathological features of pancreatic ductal adenocarcinoma - A systematic review and meta-analysis. Cancer Treat Rev 2017; 55: 96-106 [PMID: 28342938 DOI: 10.1016/j.ctrv.2017.03.003]
- Reni M, Balzano G, Zanon S, Zerbi A, Rimassa L, Castoldi R, Pinelli D, Mosconi S, Doglioni C, 44 Chiaravalli M, Pircher C, Arcidiacono PG, Torri V, Maggiora P, Ceraulo D, Falconi M, Gianni L. Safety and efficacy of preoperative or postoperative chemotherapy for resectable pancreatic adenocarcinoma (PACT-15): a randomised, open-label, phase 2-3 trial. Lancet Gastroenterol Hepatol 2018; **3**: 413-423 [PMID: 29625841 DOI: 10.1016/S2468-1253(18)30081-5]
- O'Reilly EM, Ferrone C. Neoadjuvant or Adjuvant Therapy for Resectable or Borderline Resectable 45 Pancreatic Cancer: Which Is Preferred? J Clin Oncol 2020; 38: 1757-1759 [PMID: 32119598 DOI: 10.1200/JCO.19.03318]
- 46 Mokdad AA, Minter RM, Zhu H, Augustine MM, Porembka MR, Wang SC, Yopp AC, Mansour JC, Choti MA, Polanco PM. Neoadjuvant Therapy Followed by Resection Versus Upfront Resection for Resectable Pancreatic Cancer: A Propensity Score Matched Analysis. J Clin Oncol 2017; 35: 515-522 [PMID: 27621388 DOI: 10.1200/JCO.2016.68.5081]
- Verma V, Li J, Lin C. Neoadjuvant Therapy for Pancreatic Cancer: Systematic Review of Postoperative Morbidity, Mortality, and Complications. Am J Clin Oncol 2016; 39: 302-313 [PMID: 26950464 DOI: 10.1097/COC.000000000000278]
- Versteijne E, Suker M, Groothuis K, Akkermans-Vogelaar JM, Besselink MG, Bonsing BA, Buijsen 48 J. Busch OR, Creemers GM, van Dam RM, Eskens FALM, Festen S, de Groot JWB, Groot Koerkamp B, de Hingh IH, Homs MYV, van Hooft JE, Kerver ED, Luelmo SAC, Neelis KJ, Nuyttens J, Paardekooper GMRM, Patijn GA, van der Sangen MJC, de Vos-Geelen J, Wilmink JW, Zwinderman



AH, Punt CJ, van Eijck CH, van Tienhoven G; Dutch Pancreatic Cancer Group. Preoperative Chemoradiotherapy Versus Immediate Surgery for Resectable and Borderline Resectable Pancreatic Cancer: Results of the Dutch Randomized Phase III PREOPANC Trial. J Clin Oncol 2020; 38: 1763-1773 [PMID: 32105518 DOI: 10.1200/JCO.19.02274]

- Katz MH, Pisters PW, Evans DB, Sun CC, Lee JE, Fleming JB, Vauthey JN, Abdalla EK, Crane CH, 49 Wolff RA, Varadhachary GR, Hwang RF. Borderline resectable pancreatic cancer: the importance of this emerging stage of disease. J Am Coll Surg 2008; 206: 833-846; discussion 846 [PMID: 18471707 DOI: 10.1016/j.jamcollsurg.2007.12.020]
- Versteijne E, Vogel JA, Besselink MG, Busch ORC, Wilmink JW, Daams JG, van Eijck CHJ, Groot 50 Koerkamp B, Rasch CRN, van Tienhoven G; Dutch Pancreatic Cancer Group. Meta-analysis comparing upfront surgery with neoadjuvant treatment in patients with resectable or borderline resectable pancreatic cancer. Br J Surg 2018; 105: 946-958 [PMID: 29708592 DOI: 10.1002/bjs.10870]
- Jang JY, Han Y, Lee H, Kim SW, Kwon W, Lee KH, Oh DY, Chie EK, Lee JM, Heo JS, Park JO, Lim DH, Kim SH, Park SJ, Lee WJ, Koh YH, Park JS, Yoon DS, Lee IJ, Choi SH. Oncological Benefits of Neoadjuvant Chemoradiation With Gemcitabine Versus Upfront Surgery in Patients With Borderline Resectable Pancreatic Cancer: A Prospective, Randomized, Open-label, Multicenter Phase 2/3 Trial. Ann Surg 2018; 268: 215-222 [PMID: 29462005 DOI: 10.1097/SLA.00000000002705]
- Evans DB, George B, Tsai S. Non-metastatic Pancreatic Cancer: Resectable, Borderline Resectable, 52 and Locally Advanced-Definitions of Increasing Importance for the Optimal Delivery of Multimodality Therapy. Ann Surg Oncol 2015; 22: 3409-3413 [PMID: 26122369 DOI: 10.1245/s10434-015-4649-2]
- 53 Neuzillet C, Gaujoux S, Williet N, Bachet JB, Bauguion L, Colson Durand L, Conroy T, Dahan L, Gilabert M, Huguet F, Marthey L, Meilleroux J, de Mestier L, Napoléon B, Portales F, Sa Cunha A, Schwarz L, Taieb J, Chibaudel B, Bouché O, Hammel P; Thésaurus National de Cancérologie Digestive (TNCD); Société Nationale Française de Gastroentérologie (SNFGE); Fédération Francophone de Cancérologie Digestive (FFCD); Groupe Coopérateur multidisciplinaire en Oncologie (GERCOR); Fédération Nationale des Centres de Lutte Contre le Cancer (UNICANCER); Société Française de Chirurgie Digestive (SFCD); Société Française d'Endoscopie Digestive (SFED); Société Française de Radiothérapie Oncologique (SFRO); Association de Chirurgie Hépato-Bilio-Pancréatique et Transplantation (ACHBT); Association Française de Chirurgie (AFC). Pancreatic cancer: French clinical practice guidelines for diagnosis, treatment and follow-up (SNFGE, FFCD, GERCOR, UNICANCER, SFCD, SFED, SFRO, ACHBT, AFC). Dig Liver Dis 2018; 50: 1257-1271 [PMID: 30219670 DOI: 10.1016/j.dld.2018.08.008]
- 54 Suker M, Beumer BR, Sadot E, Marthey L, Faris JE, Mellon EA, El-Rayes BF, Wang-Gillam A, Lacy J, Hosein PJ, Moorcraft SY, Conroy T, Hohla F, Allen P, Taieb J, Hong TS, Shridhar R, Chau I, van Eijck CH, Koerkamp BG. FOLFIRINOX for locally advanced pancreatic cancer: a systematic review and patient-level meta-analysis. Lancet Oncol 2016; 17: 801-810 [PMID: 27160474 DOI: 10.1016/S1470-2045(16)00172-8
- 55 Wang C, Liu X, Wang X, Wang Y, Cha N. Effects of chemoradiotherapy and chemotherapy on survival of patients with locally advanced pancreatic cancer: A meta-analysis of randomized controlled trials. Medicine (Baltimore) 2018; 97: e12260 [PMID: 30200163 DOI: 10.1097/MD.00000000012260
- Mukherjee S, Hurt CN, Bridgewater J, Falk S, Cummins S, Wasan H, Crosby T, Jephcott C, Roy R, 56 Radhakrishna G, McDonald A, Ray R, Joseph G, Staffurth J, Abrams RA, Griffiths G, Maughan T. Gemcitabine-based or capecitabine-based chemoradiotherapy for locally advanced pancreatic cancer (SCALOP): a multicentre, randomised, phase 2 trial. Lancet Oncol 2013; 14: 317-326 [PMID: 23474363 DOI: 10.1016/S1470-2045(13)70021-4]
- Bockhorn M, Uzunoglu FG, Adham M, Imrie C, Milicevic M, Sandberg AA, Asbun HJ, Bassi C, 57 Büchler M, Charnley RM, Conlon K, Cruz LF, Dervenis C, Fingerhutt A, Friess H, Gouma DJ, Hartwig W, Lillemoe KD, Montorsi M, Neoptolemos JP, Shrikhande SV, Takaori K, Traverso W, Vashist YK, Vollmer C, Yeo CJ, Izbicki JR; International Study Group of Pancreatic Surgery. Borderline resectable pancreatic cancer: a consensus statement by the International Study Group of Pancreatic Surgery (ISGPS). Surgery 2014; 155: 977-988 [PMID: 24856119 DOI: 10.1016/j.surg.2014.02.001]
- Khorana AA, Mangu PB, Berlin J, Engebretson A, Hong TS, Maitra A, Mohile SG, Mumber M, 58 Schulick R, Shapiro M, Urba S, Zeh HJ, Katz MHG. Potentially Curable Pancreatic Cancer: American Society of Clinical Oncology Clinical Practice Guideline Update. J Clin Oncol 2017; 35: 2324-2328 [PMID: 28398845 DOI: 10.1200/JCO.2017.72.4948]
- Burris HA 3rd, Moore MJ, Andersen J, Green MR, Rothenberg ML, Modiano MR, Cripps MC, 59 Portenoy RK, Storniolo AM, Tarassoff P, Nelson R, Dorr FA, Stephens CD, Von Hoff DD. Improvements in survival and clinical benefit with gencitabine as first-line therapy for patients with advanced pancreas cancer: a randomized trial. J Clin Oncol 1997; 15: 2403-2413 [PMID: 9196156 DOI: 10.1200/JCO.1997.15.6.2403]
- Moore K. Endothelin and vascular function in liver disease. Gut 2004; 53: 159-161 [PMID: 60 14724140 DOI: 10.1136/gut.2003.024703]
- Tabernero J, Chiorean EG, Infante JR, Hingorani SR, Ganju V, Weekes C, Scheithauer W, 61 Ramanathan RK, Goldstein D, Penenberg DN, Romano A, Ferrara S, Von Hoff DD. Prognostic factors of survival in a randomized phase III trial (MPACT) of weekly nab-paclitaxel plus



gemcitabine vs gemcitabine alone in patients with metastatic pancreatic cancer. Oncologist 2015; 20: 143-150 [PMID: 25582141 DOI: 10.1634/theoncologist.2014-0394]

- 62 Sehdev A, Gbolahan O, Hancock BA, Stanley M, Shahda S, Wan J, Wu HH, Radovich M, O'Neil BH. Germline and Somatic DNA Damage Repair Gene Mutations and Overall Survival in Metastatic Pancreatic Adenocarcinoma Patients Treated with FOLFIRINOX. Clin Cancer Res 2018; 24: 6204-6211 [PMID: 30131383 DOI: 10.1158/1078-0432.CCR-18-1472]
- 63 Fogelman D, Sugar EA, Oliver G, Shah N, Klein A, Alewine C, Wang H, Javle M, Shroff R, Wolff RA, Abbruzzese JL, Laheru D, Diaz LA Jr. Family history as a marker of platinum sensitivity in pancreatic adenocarcinoma. Cancer Chemother Pharmacol 2015; 76: 489-498 [PMID: 26126726 DOI: 10.1007/s00280-015-2788-6]
- 64 Kang H, Jo JH, Lee HS, Chung MJ, Bang S, Park SW, Song SY, Park JY. Comparison of efficacy and safety between standard-dose and modified-dose FOLFIRINOX as a first-line treatment of pancreatic cancer. World J Gastrointest Oncol 2018; 10: 421-430 [PMID: 30487953 DOI: 10.4251/wjgo.v10.i11.421]
- Walker EJ, Ko AH. Beyond first-line chemotherapy for advanced pancreatic cancer: an expanding array of therapeutic options? World J Gastroenterol 2014; 20: 2224-2236 [PMID: 24605022 DOI: 10.3748/wjg.v20.i9.2224]
- Nagrial AM, Chin VT, Sjoquist KM, Pajic M, Horvath LG, Biankin AV, Yip D. Second-line 66 treatment in inoperable pancreatic adenocarcinoma: A systematic review and synthesis of all clinical trials. Crit Rev Oncol Hematol 2015; 96: 483-497 [PMID: 26481952 DOI: 10.1016/j.critrevonc.2015.07.007
- 67 Oettle H, Riess H, Stieler JM, Heil G, Schwaner I, Seraphin J, Görner M, Mölle M, Greten TF, Lakner V, Bischoff S, Sinn M, Dörken B, Pelzer U. Second-line oxaliplatin, folinic acid, and fluorouracil vs folinic acid and fluorouracil alone for gemcitabine-refractory pancreatic cancer: outcomes from the CONKO-003 trial. J Clin Oncol 2014; 32: 2423-2429 [PMID: 24982456 DOI: 10.1200/JCO.2013.53.6995
- 68 Portal A, Pernot S, Tougeron D, Arbaud C, Bidault AT, de la Fouchardière C, Hammel P, Lecomte T, Dréanic J, Coriat R, Bachet JB, Dubreuil O, Marthey L, Dahan L, Tchoundjeu B, Locher C, Lepère C, Bonnetain F, Taieb J. Nab-paclitaxel plus gemcitabine for metastatic pancreatic adenocarcinoma after Folfirinox failure: an AGEO prospective multicentre cohort. Br J Cancer 2015; 113: 989-995 [PMID: 26372701 DOI: 10.1038/bjc.2015.328]
- Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, Lu S, Kemberling H, Wilt C, Luber BS, Wong F, Azad NS, Rucki AA, Laheru D, Donehower R, Zaheer A, Fisher GA, Crocenzi TS, Lee JJ, Greten TF, Duffy AG, Ciombor KK, Eyring AD, Lam BH, Joe A, Kang SP, Holdhoff M, Danilova L, Cope L, Meyer C, Zhou S, Goldberg RM, Armstrong DK, Bever KM, Fader AN, Taube J, Housseau F, Spetzler D, Xiao N, Pardoll DM, Papadopoulos N, Kinzler KW, Eshleman JR, Vogelstein B, Anders RA, Diaz LA Jr. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. Science 2017; 357: 409-413 [PMID: 28596308 DOI: 10.1126/science.aan6733]
- 70 Domchek SM, Hendifar AE, McWilliams RR, Geva R, Epelbaum R, Biankin A, Vonderheide RH, Wolff RA, Alberts SR, Giordano H, Goble S, Lin KK, Shroff RT. RUCAPANC: An open-label, phase 2 trial of the PARP inhibitor rucaparib in patients (pts) with pancreatic cancer (PC) and a known deleterious germline or somatic BRCA mutation. J Clin Oncol 2016; 15: 34
- 71 Perinel J, Adham M. Palliative therapy in pancreatic cancer-palliative surgery. Transl Gastroenterol Hepatol 2019; 4: 28 [PMID: 31231695 DOI: 10.21037/tgh.2019.04.03]
- 72 Dumonceau JM, Tringali A, Papanikolaou IS, Blero D, Mangiavillano B, Schmidt A, Vanbiervliet G, Costamagna G, Devière J, García-Cano J, Gyökeres T, Hassan C, Prat F, Siersema PD, van Hooft JE. Endoscopic biliary stenting: indications, choice of stents, and results: European Society of Gastrointestinal Endoscopy (ESGE) Clinical Guideline - Updated October 2017. Endoscopy 2018; 50: 910-930 [PMID: 30086596 DOI: 10.1055/a-0659-9864]
- Berti S, Ferrarese A, Feleppa C, Francone E, Martino V, Bianchi C, Falco E. Laparoscopic 73 perspectives for distal biliary obstruction. Int J Surg 2015; 21 Suppl 1: S64-S67 [PMID: 26118614 DOI: 10.1016/j.ijsu.2015.04.092]
- 74 Lai EC, Tang CN. Robot-assisted laparoscopic hepaticojejunostomy for advanced malignant biliary obstruction. Asian J Surg 2015; 38: 210-213 [PMID: 25797562 DOI: 10.1016/j.asjsur.2015.01.010]
- 75 Marabelle A, Le DT, Ascierto PA, Di Giacomo AM, De Jesus-Acosta A, Delord JP, Geva R, Gottfried M, Penel N, Hansen AR, Piha-Paul SA, Doi T, Gao B, Chung HC, Lopez-Martin J, Bang YJ, Frommer RS, Shah M, Ghori R, Joe AK, Pruitt SK, Diaz LA Jr. Efficacy of Pembrolizumab in Patients With Noncolorectal High Microsatellite Instability/Mismatch Repair-Deficient Cancer: Results From the Phase II KEYNOTE-158 Study. J Clin Oncol 2020; 38: 1-10 [PMID: 31682550 DOI: 10.1200/JCO.19.02105]
- 76 Banerjee K, Kumar S, Ross KA, Gautam S, Poelaert B, Nasser MW, Aithal A, Bhatia R, Wannemuehler MJ, Narasimhan B, Solheim JC, Batra SK, Jain M. Emerging trends in the immunotherapy of pancreatic cancer. Cancer Lett 2018; 417: 35-46 [PMID: 29242097 DOI: 10.1016/j.canlet.2017.12.012
- Pishvaian MJ, Bender RJ, Halverson D, Rahib L, Hendifar AE, Mikhail S, Chung V, Picozzi VJ, 77 Sohal D, Blais EM, Mason K, Lyons EE, Matrisian LM, Brody JR, Madhavan S, Petricoin EF 3rd. Molecular Profiling of Patients with Pancreatic Cancer: Initial Results from the Know Your Tumor Initiative. Clin Cancer Res 2018; 24: 5018-5027 [PMID: 29954777 DOI: 10.1158/1078-0432.CCR-18-0531



- 78 Biankin AV, Waddell N, Kassahn KS, Gingras MC, Muthuswamy LB, Johns AL, Miller DK, Wilson PJ, Patch AM, Wu J, Chang DK, Cowley MJ, Gardiner BB, Song S, Harliwong I, Idrisoglu S, Nourse C, Nourbakhsh E, Manning S, Wani S, Gongora M, Pajic M, Scarlett CJ, Gill AJ, Pinho AV, Rooman I, Anderson M, Holmes O, Leonard C, Taylor D, Wood S, Xu Q, Nones K, Fink JL, Christ A, Bruxner T, Cloonan N, Kolle G, Newell F, Pinese M, Mead RS, Humphris JL, Kaplan W, Jones MD, Colvin EK, Nagrial AM, Humphrey ES, Chou A, Chin VT, Chantrill LA, Mawson A, Samra JS, Kench JG, Lovell JA, Daly RJ, Merrett ND, Toon C, Epari K, Nguyen NQ, Barbour A, Zeps N; Australian Pancreatic Cancer Genome Initiative, Kakkar N, Zhao F, Wu YQ, Wang M, Muzny DM, Fisher WE, Brunicardi FC, Hodges SE, Reid JG, Drummond J, Chang K, Han Y, Lewis LR, Dinh H, Buhay CJ, Beck T, Timms L, Sam M, Begley K, Brown A, Pai D, Panchal A, Buchner N, De Borja R, Denroche RE, Yung CK, Serra S, Onetto N, Mukhopadhyay D, Tsao MS, Shaw PA, Petersen GM, Gallinger S, Hruban RH, Maitra A, Iacobuzio-Donahue CA, Schulick RD, Wolfgang CL, Morgan RA, Lawlor RT, Capelli P, Corbo V, Scardoni M, Tortora G, Tempero MA, Mann KM, Jenkins NA, Perez-Mancera PA, Adams DJ, Largaespada DA, Wessels LF, Rust AG, Stein LD, Tuveson DA, Copeland NG, Musgrove EA, Scarpa A, Eshleman JR, Hudson TJ, Sutherland RL, Wheeler DA, Pearson JV, McPherson JD, Gibbs RA, Grimmond SM. Pancreatic cancer genomes reveal aberrations in axon guidance pathway genes. Nature 2012; 491: 399-405 [PMID: 23103869 DOI: 10.1038/nature11547]
- National Institutes of Health's National Library of Medicine. DailyMed VITRAKVI-79 larotrectinib capsule VITRAKVI- larotrectinib solution, concentrate. In: National Institutes of Health's National Library of Medicine [Internet]. Available from: https://NLM/NIH/HHS/USA.gov
- 80 Genentech. RozlytrekTM (entrectinib) - Information for Healthcare Providers. In: Genentech [Internet]. Available from: www.gene.com
- Drilon A, Laetsch TW, Kummar S, DuBois SG, Lassen UN, Demetri GD, Nathenson M, Doebele 81 RC, Farago AF, Pappo AS, Turpin B, Dowlati A, Brose MS, Mascarenhas L, Federman N, Berlin J, El-Deiry WS, Baik C, Deeken J, Boni V, Nagasubramanian R, Taylor M, Rudzinski ER, Meric-Bernstam F, Sohal DPS, Ma PC, Raez LE, Hechtman JF, Benayed R, Ladanyi M, Tuch BB, Ebata K, Cruickshank S, Ku NC, Cox MC, Hawkins DS, Hong DS, Hyman DM. Efficacy of Larotrectinib in TRK Fusion-Positive Cancers in Adults and Children. N Engl J Med 2018; 378: 731-739 [PMID: 29466156 DOI: 10.1056/NEJMoa1714448]
- Drilon A, Siena S, Ou SI, Patel M, Ahn MJ, Lee J, Bauer TM, Farago AF, Wheler JJ, Liu SV, 82 Doebele R, Giannetta L, Cerea G, Marrapese G, Schirru M, Amatu A, Bencardino K, Palmeri L, Sartore-Bianchi A, Vanzulli A, Cresta S, Damian S, Duca M, Ardini E, Li G, Christiansen J, Kowalski K, Johnson AD, Patel R, Luo D, Chow-Maneval E, Hornby Z, Multani PS, Shaw AT, De Braud FG. Safety and Antitumor Activity of the Multitargeted Pan-TRK, ROS1, and ALK Inhibitor Entrectinib: Combined Results from Two Phase I Trials (ALKA-372-001 and STARTRK-1). Cancer Discov 2017; 7: 400-409 [PMID: 28183697 DOI: 10.1158/2159-8290.CD-16-1237]
- Golan T, Hammel P, Reni M, Van Cutsem E, Macarulla T, Hall MJ, Park JO, Hochhauser D, Arnold 83 D, Oh DY, Reinacher-Schick A, Tortora G, Algül H, O'Reilly EM, McGuinness D, Cui KY, Schlienger K, Locker GY, Kindler HL. Maintenance Olaparib for Germline BRCA-Mutated Metastatic Pancreatic Cancer. N Engl J Med 2019; 381: 317-327 [PMID: 31157963 DOI: 10.1056/NEJMoa1903387]
- Beatty GL, Torigian DA, Chiorean EG, Saboury B, Brothers A, Alavi A, Troxel AB, Sun W, 84 Teitelbaum UR, Vonderheide RH, O'Dwyer PJ. A phase I study of an agonist CD40 monoclonal antibody (CP-870,893) in combination with gemcitabine in patients with advanced pancreatic ductal adenocarcinoma. Clin Cancer Res 2013; 19: 6286-6295 [PMID: 23983255 DOI: 10.1158/1078-0432.CCR-13-1320]
- 85 Rossi GR, Mautino MR, Unfer RC, Seregina TM, Vahanian N, Link CJ. Effective treatment of preexisting melanoma with whole cell vaccines expressing alpha(1,3)-galactosyl epitopes. Cancer Res 2005; 65: 10555-10561 [PMID: 16288048 DOI: 10.1158/0008-5472.CAN-05-0627]
- Hardacre JM, Mulcahy M, Small W, Talamonti M, Obel J, Krishnamurthi S, Rocha-Lima CS, 86 Safran H, Lenz HJ, Chiorean EG. Addition of algenpantucel-L immunotherapy to standard adjuvant therapy for pancreatic cancer: a phase 2 study. J Gastrointest Surg 2013; 17: 94-100; discussion p. 100 [PMID: 23229886 DOI: 10.1007/s11605-012-2064-6]
- 87 Coveler AL, Rossi GR, Vahanian NN, Link C, Chiorean EG. Algenpantucel-L immunotherapy in pancreatic adenocarcinoma. Immunotherapy 2016; 8: 117-125 [PMID: 26787078 DOI: 10.2217/imt.15.113
- 88 Sahin IH, Askan G, Hu ZI, O'Reilly EM. Immunotherapy in pancreatic ductal adenocarcinoma: an emerging entity? Ann Oncol 2017; 28: 2950-2961 [PMID: 28945842 DOI: 10.1093/annonc/mdx503]
- 89 Kindler HL, Niedzwiecki D, Hollis D, Sutherland S, Schrag D, Hurwitz H, Innocenti F, Mulcahy MF, O'Reilly E, Wozniak TF, Picus J, Bhargava P, Mayer RJ, Schilsky RL, Goldberg RM. Gemcitabine plus bevacizumab compared with gemcitabine plus placebo in patients with advanced pancreatic cancer: phase III trial of the Cancer and Leukemia Group B (CALGB 80303). J Clin Oncol 2010; 28: 3617-3622 [PMID: 20606091 DOI: 10.1200/JCO.2010.28.1386]
- 90 Lemstrova R, Brynychova V, Hughes DJ, Hlavac V, Dvorak P, Doherty JE, Murray HA, Crockard M, Oliverius M, Hlavsa J, Honsova E, Mazanec J, Kala Z, Lovecek M, Havlik R, Ehrmann J, Strouhal O, Soucek P, Melichar B, Mohelnikova-Duchonova B. Dysregulation of KRAS signaling in pancreatic cancer is not associated with KRAS mutations and outcome. Oncol Lett 2017; 14: 5980-5988 [PMID: 29113235 DOI: 10.3892/ol.2017.6946]



Manrai M et al. Current and emerging therapeutic strategies: Pancreatic cancer

91 Schmidt KM, Hellerbrand C, Ruemmele P, Michalski CW, Kong B, Kroemer A, Hackl C, Schlitt HJ, Geissler EK, Lang SA. Inhibition of mTORC2 component RICTOR impairs tumor growth in pancreatic cancer models. Oncotarget 2017; 8: 24491-24505 [PMID: 28445935 DOI: 10.18632/oncotarget.15524]



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MINIREVIEWS

Cathepsin L, transmembrane peptidase/serine subfamily member 2/4, and other host proteases in COVID-19 pathogenesis – with impact on gastrointestinal tract

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Abstract

Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) seems to employ two routes of entrance to the host cell; via membrane fusion (with the cells expressing both angiotensin converting enzyme 2 (ACE2) and transmembrane peptidase/serine subfamily member 2/4 (TMPRSS2/4)) or via receptor-mediated endocytosis (to the target cells expressing only ACE2). The second mode is associated with cysteine cathepsins (probably cathepsin L) involvement in the virus spike protein (S protein) proteolytic activation. Also furin might activate the virus S protein enabling it to enter cells. Gastrointestinal tract (GIT) involvement in SARS-CoV-2 infection is evident in a subset of coronavirus disease 2019 (COVID-19) patients exhibiting GIT symptoms, such as diarrhea, and presenting viral-shedding in feces. Considering the abundance and co-localization of ACE2 and TMPRSS2 in the lower GIT (especially brush-border enterocytes), these two receptors seem to be mainly involved in SARS-CoV-2 invasion of the digestive tract. Additionally, in vitro studies have demonstrated the virions capability of infection and replication in the human epithelial cells lining GIT. However, also furin and cysteine cathepsins (cathepsin L) might participate in the activation of SARS-CoV-2 spike protein contributing to the virus invasiveness within GIT. Moreover, cathepsin L (due to its involvement in extracellular matrix components degradation and remodeling, the processes enhanced during SARS-CoV-2induced inflammation) might be responsible for the dysregulation of absorption/ digestion functions of GIT, thus adding to the observed in some COVID-19 patients symptoms such as diarrhea.

Key Words: COVID-19; SARS-CoV-2; Angiotensin converting enzyme 2; Transmembrane peptidase/serine subfamily member 2/4; Cathepsin L; Gastrointestinal tract

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Core Tip: Gastrointestinal tract (GIT) is believed to participate in severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) dissemination. The current research shows the abundance and co-localization of angiotensin converting enzyme 2 (ACE2) and transmembrane peptidase/serine subfamily member 2 receptors in the lower GIT. Furthermore, about half of coronavirus disease 2019 patients present with GIT symptoms, such as diarrhea, and exhibit viral-shedding in feces. Additionally, in vitro studies have demonstrated the virions capability of infection and replication in the human epithelial cells lining GIT. This paper reviews the possible routes of the virus infection with respect to the host-enzymatic systems responsible for the proteolytic priming of SARS-CoV-2.

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INTRODUCTION

Coronaviruses may employ several host proteases for their invasion into target cells. The enzymes participating in the viruses activation include: proprotein convertases (PCs) (mainly furin), transmembrane serine proteases, especially transmembrane peptidase/serine subfamily member 2 (TMPRSS2), the lysosomal cathepsins (mainly cathepsin L), elastase, and coagulation factor Xa^[1].

Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) virus responsible for coronavirus disease 2019 (COVID-19) pandemic exhibits many similarities when compared to SARS-CoV. It employs a similar mechanism of host cells invasion; recognizes and binds the same type of angiotensin converting enzyme 2 (ACE2) receptors to enter host cells (but with higher affinity[2]), and comparable processing of spike protein (S protein) seems to be necessary for the virion fusion with the host cell membrane[3-7] (Figure 1).

Two proteolytic events need to be conducted for SARS-CoV-2 activation; initially spike protein is cut in the specific cleavage site between S1 and S2 domain, then the second cleavage within S2 domain (S2' site) allows for the exposition of the fusion peptide, which enables membrane fusion. The first proteolytic step can happen in the producer cell, in the extracellular space, or within the host cell's endosome. This cleavage site in SARS-CoV-2 spike protein is recognized by various proteases, including furin (unlike in SARS-CoV lacking furin-cleavage site between S1 and S2)[6, 8] and TMPRSS2[3]. The second cleavage can be either performed by TMPRSS2 on the surface of the host cell, or in the endolysosomes by lysosomal proteases, most probably cathepsin L[9].

Therefore, as proposed by Pislar *et al*[10], two routes of SARS-CoV-2 entry to the host cell are likely; via membrane fusion with the host cells which expose both ACE2 and TMPRSS2 (and/or other transmembrane serine proteases such as TMPRSS4[11]) proteins, or via receptor-mediated endocytosis (RME) to the target cells expressing only ACE2 receptors. In the first case, both processing steps performed by TMPRSS2 before the virus entry enable membrane fusion, whereas in the second mechanism, the virion binding with ACE2 receptors induces endocytosis followed by spike protein activation by cathepsin L (and/or other cysteine cathepsins)[4,10]. As a result of either of these pathways, viral RNA is released in the host cell and undergoes the processes of replication (Figure 2).

TYPES OF HOST PROTEASES IN SARS-CoV-2 INVASION

The findings of several studies support the notion that, apart from ACE2 receptors, the main host peptidases involved in SARS-CoV-2 spike protein processing include: TMPRSS2 (and/or TMPRSS4), lysosomal cysteine cathepsins (mainly cathepsin L), as well as furin-like PCs. They may participate in SARS-CoV-2 activation independently,



Berdowska I et al. Proteases in SARS-CoV-2 gastrointestinal tract invasion

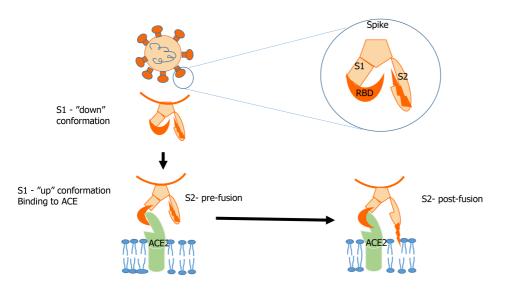


Figure 1 Alternations in spike proteins conformation upon binding to ACE 2. Spike S1 subunit contains receptor binding domain, that has to change from "down-conformation" state to "up-conformation" state to be accessible for ACE2. Changes in S1 subunit trigger conformational changes in S2 subunit, causing exposition of hydrophobic domain, changing it from "pre-fusion" to "post-fusion" state. This enables fusion of the virus with host membrane (after Zhu et al[7]).

or their actions may overlap or complete one another, depending on the pattern of the virion-recognized proteins exposed on the host cells (e.g. expressing or not TMPRSS2).

Cysteine cathepsins (CCs) in pathology

Cysteine cathepsins belonging to the papain-like family of cysteine proteases (containing cysteine in their catalytic center) comprise 11 cathepsins (B, C, F, H, K, L, O, S, V, X and W) in the human organism[12,13]. They belong to lysosomal proteases involved mainly in intracellular protein breakdown, antigen processing, MHC-II mediated immune response, and apoptosis. However, their functions go far beyond this; they participate in various physio-pathological processes not only intracellularly, but also in the extracellular matrix (ECM), because, except for their endolysosomal sequestration, they have been observed in the nucleus, cytosol, mitochondria, at the plasma membranes and in the extracellular milieu[13,14]. Their secretion is observed in physiological conditions (e.g. in bone remodeling - conducted by osteoclastssecreted cathepsin K, in wound healing performed by keratinocytes-secreted cathepsin B, in prohormone processing – thyroid hormones released from thyroglobulin by cathepsins B, L and K secreted from the thyroid epithelial cells). However, an excessive secretion of cysteine cathepsins is mostly observed in pathological states associated with inflammatory processes, such as cancer diseases (cathepsins B, C, K, L, S, H, X), cardiovascular diseases (cathepsins C, K, L, S, V), joint and bone diseases (cathepsins K, B, L, S, H), inflammatory bowel disease (cathepsin L), and many other disorders (summarized in[12-15]). In these pathological states CCs typically act extracellularly, where they participate in collagen, elastin, and other ECM components degradation directly or indirectly (activating other proteases) after being secreted from recruited immune cells (mostly), as well as from inflamed tissue cells (to the lesser extent). Macrophages and other immune cells infiltrating tissues seem to be the main extracellular source of CCs whose secretion is stimulated by inflammatory factors like cytokines. However, also other cell types secrete excessive amounts of CCs, including osteoclasts or chondrocytes (oversecreting cathepsin K and/or S in arthritis and osteoporosis), or cancer cells and tumor-associated fibroblasts (oversecreting cathepsin S, L and/or B in cancer invasion)[14]. Apart from degradation of main components of ECM, also more refined processing of ECM (undergoing both intra-, and extracellularly) is ascribed to CCs. This comprises modifying and shedding cell adhesion molecules and cell membrane receptors, which affects signal transduction pathways, as well as processing cytokines and chemokines, which upregulates immune response [14]. The resulting augmentation of inflammatory processes further induces the secretion of CCs, enhancing the destruction of ECM, which eventually leads to the acceleration of the processes observed in the aforementioned disorders.

Due to their roles in multiple inflammatory-based disorders, CCs have been considered for a long time as a target for medicinal drugs design. However, because of ubiquitous expression of most CCs, their constitutive, overlapping functions, broad substrate specificities, most of the studied so far medicines have exhibited unfavorable



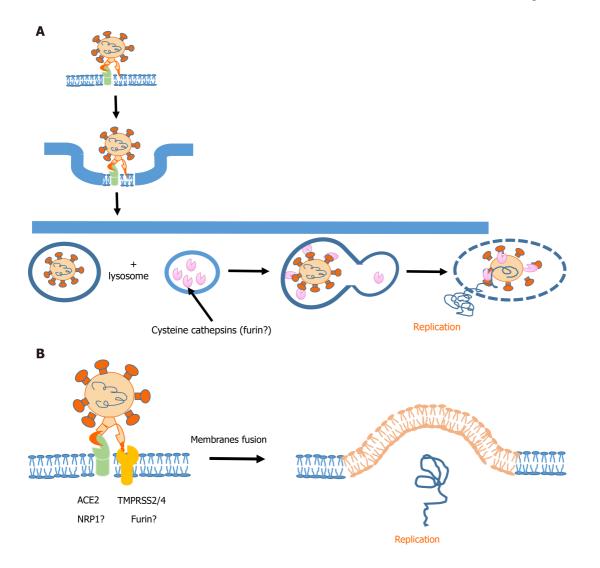


Figure 2 Two modes of virus entry. A: Receptor-mediated endocytosis of severe acute respiratory syndrome coronavirus-2. After binding to ACE2 and formation of endosome, lysosomal cathepsins activate spike protein, which leads to the release of the viral RNA into host cell. B: Membrane fusion mechanism - priming of the spike proteins is mediated by transmembrane peptidase/serine subfamily member 2/4, which leads to fusion of viral and host membranes and release of the viral RNA into host cell.

side effects, thus not surviving clinical trials. However, the attempts to construct CCsaimed drugs, taking advantage of the newest technology, are underway. The most clinical trials have been conducted on cathepsins K and S inhibitors, also cathepsin C seems to be a promising target, whereas the remaining cathepsins inhibitors have either got stuck in the initial stages of clinical trials or their trials have been discontinued (reviewed in[14]).

Cathepsin L in inflammatory processes

Similarly to other cysteine cathepsins, cathepsin L exhibits pleiotropic activities in the human organism. One of the most evident actions of this enzyme is (beside cathepsins S, K and V) its participation in inflammatory processes associated with various pathological conditions[14-16]. For example Menzel *et al*[17] have demonstrated the up to 10-fold induction of cathepsin L expression (mRNA) in intestinal macrophages derived from inflammatory bowel disease (IBD) patients, and the clear improvement of the disorder symptoms in DSS (dextran-sulphate-sodium)-induced colitis mice mode, when a simultaneous application of cathepsins L and B inhibitors was investigated. Xu *et al*[18] have exhibited the stimulatory function of cathepsin L in microglia-mediated neuroinflammation, which accompanies many neurological disorders including Parkinson's disease. Cao *et al*[19] have shown the correlation between serum cathepsin L activity and the markers of inflammation (such as neutrophile counts and hs-CRP) in the patients with chronic kidney disease.

Cysteine cathepsins and TMPRSS2 in SARS-CoV-2 invasion

Cell line experimental systems creating the environment aimed at the inhibition of CCs have substantiated the function of these enzymes in the processes of SARS-CoV-2 activation. Raising pH in the endolysosomal compartments (with ammonium chloride and/or bafilomycin A), which inactivates lysosomal proteases working in acidic environment, or application of cysteine proteases inhibitors (such as E-64d inhibitor inactivating cysteine cathepsins L, B, H, as well as cytosolic calpain) have significantly limited entry of SARS-CoV-2 into chosen cell lines[4]. Ou et al[20] (applying lentiviral pseudotype system) have demonstrated that the treatment of HEK-293/hACE2 cells with E-64d inhibitor reduced entry of SARS-CoV-2 S pseudovirions by over 90%. Further, the Authors compared the effect of two specific inhibitors of cathepsin L (SID 26681509), and cathepsin B (CA-074). Whereas the first inhibitor limited the pseudovirions entry by over 76%, the second one did not exhibit any significant effect, which suggests a prevalent function of cathepsin L in the receptor-mediated endocytosis mechanism of the virus invasion. RME mechanism in HEK- 293/hACE2 cells has been confirmed by the Authors in the experiments showing the inhibition of the SARS-CoV-2 S protein entry by blocking the factors inevitable in the process of endolysosomal trafficking (PI(3)P 5-kinase (PIKfyve) and two-pore channel subtype 2 (TPC2)[20]. The involvement of CCs has been also observed by Hoffmann *et al*[3] who demonstrated that the increase in pH (with ammonium chloride) almost completely inhibited SARS-2-S-driven entry into 293T cells expressing ACE2 but devoid of TMPRSS2. This observation is in agreement with Ou et al[20] findings confirming that the virus entry into these cells undergoes via RME with the involvement of cathepsin L in spike protein activation. In their experiments on the human colon cell line - Caco-2 cells overexpressing TMPRSS2, Hoffmann et al[3] exhibited the participation of both TMPRSS2 and CCs in the mechanism of the virus entry into these cells. Alkalization of the environment caused around 90% reduction in SARS-2-S-driven entry, whereas incubation with E-64d inhibited the virus entry by around 40%. On the other hand, the application of camostat mesylate (inhibitor of TMPRSS2 and other serine proteases) reduced the virus entry by about 90%, and when the Authors used both inhibitors simultaneously, they achieved nearly complete virus-entry inhibition to Caco-2 TMPRSS2 (+) cells. They also found that E-64 inhibitor significantly reduced the virus entry into Vero and 293T cells not expressing TMPRSS2, which indicates the endolysosomal pathway. However, the transduction of these cell lines with TMPRSS2 markedly reversed the effect of CCs inhibition, emphasizing the function of TMPRSS2 in the S protein priming, agreeably with Ou et al^[20] findings that the expression of TMPRSS 2, 4, 11 A, 11D, and 11E on 293/hACE2 cells enhanced SARS-CoV-2 S protein-mediated cell-cell fusion. Moreover, the Authors noted that, however the addition of trypsin (like TMPRSS belonging to serine proteases) stimulated the formation of syncytia in 293/hACE2 cells (indicative of activated by trypsin SARS-2-S protein-stimulated cell-cell fusion), this process was also noticed in the experiments without trypsin. These findings indicate that binding with ACE2 receptors may be a sufficient event inducing cell-cell fusion (without the proteolytic priming with the extracellular protease). Nevertheless, it is possible that other host extracellular peptidases are involved in spike protein activation. The findings derived from Hoffmann *et al*[3] and Ou *et al*[20] experiments are depicted in Table 1.

Furin in SARS-CoV-2 invasion

As determined by Shang et al[8], another enzyme involved in SARS-CoV-2 spike protein priming is furin belonging to proprotein convertases (PCs). PCs are eukaryotic serine proteases, and ubiquitously expressed furin belongs to the PCs subfamily present in the organelles of the constitutive protein secretion pathway. These enzymes participate in the proteolytic post-translational modification of a variety of functionally important peptides and proteins, such as growth factors and hormones, both intraand extracellularly (in the trans-Golgi network, endosomes, and pericellular environment). The amino acid sequence specifically recognized and cleaved by PCs including furin, is found in many viral surface proteins, so different viruses (like MERS-CoV) are activated by these enzymes[1]. Unlike SARS-CoV (exhibiting PC cleavage site motif only in the S2' site) [1], SARS-CoV-2 spike protein includes PCs specific motif at the S1/S2 boundary[8]. Shang et al[8] have performed experiments to examine whether this sequence is cut by furin. They demonstrated that PCs inhibitors reduced SARS-CoV-2 pseudovirus entry into three cell lines expressing hACE2 receptors; HeLa cells (human cervical cells), Calu-3 cells (human lung epithelial cells), and MRC-5 cells (human lung fibroblast cells). Moreover, the mutation of the PCs specific motif significantly reduced the pseudovirions entry to the studied cells. The



Table 1 Receptors/proteases involved in Severe acute respiratory syndrome coronavirus-2 invasion of human cells					
Receptor/protease	Experimental model	Observation	Ref.		
ACE2 and TMPRSS2/4	Human small intestinal enteroids; HEK-293T cell line transfected with ACE2, TMPRSS2, or TMPRSS4	Productive infection of SARS-CoV-2 in ACE2 (+) mature enterocytes; Correlation of ACE2, TMPRSS2 and 4 with SARS-CoV-2 invasiveness	Zang et al[<mark>11</mark>]		
ACE2 and TMPRSS2	ACE2 and TMPRSS2 expressing C2BBe1, Caco-2, and Calu-3 cell lines	Persistent invasion and replication of SARS-CoV-2 in the cells; Correlation of TMPRSS2 (but not ACE2) with SARS-CoV-2 RNA	Lee <i>et al</i> [<mark>33</mark>]		
ACE2, TMPRSS2, and CCs	HEK-293T cell line transfected with ACE2; Caco-2 cells overexpressing TMPRSS2	Inhibition of SARS-CoV-2 pseudovirus entry into HEK-293T ACE2(+)/TMPRSS2(-) by pH increase and E-64 inhibitor; Inhibition of SARS-CoV-2 pseudovirus entry into Caco-2 TMPRSS2(+) by pH increase, CCs inhibitor (E-64d), and TMPRSS2 inhibitor (camostat mesylate)	Hoffmann <i>et al</i> [3]		
ACE2, TMPRSS 2, 4, 11A, 11D, 11E, and CCs (cathepsin L)	HEK-293T cell line transfected with ACE2	Inhibition of SARS-CoV-2 pseudovirus entry into the cells with CCs inhibitor (E-64d) and cathepsin L inhibitor (but not cathepsin B inhibitor); Intensification of SARS-CoV-2 S protein-mediated cell-cell fusion, caused by expression of TMPRSS 2, 4, 11A, 11D, and 11E on 293/hACE2 cells	Ou et al[20]		
ACE2 and TMPRSS2	ACE2 and TMPRSS2 expressing Caco-2 and T84 cell lines	Persistent invasion and replication of SARS-CoV-2 in the cells	Stanifer <i>et al</i> [35]		
ACE2, furin, TMPRSS2, and CCs	ACE2-expressing HeLa, Calu-3, and MRC-5 cell lines	Reduction of SARS-CoV-2 pseudovirus entry into cells by inhibitors of PCs, CCs and TMPRSS2	Shang et al[8]		
NRP1, ACE2, and TMPRSS2	HEK-293T cell line transfected with ACE2, TMPRSS2, or NRP1	Augmentation of SARS-CoV-2 infectivity when NRP1 was coexpressed with ACE2 and TMPRSS2	Cantuti- Castelvetri <i>et al</i> [21]		

ACE2: Angiotensin converting enzyme 2; NRP1: Neuropilin 1 (receptor which binds furin-cleaved substrates); TMPRSS2/4: Transmembrane peptidase/serine subfamily member 2/4; CCs: Cysteine cathepsins; HEK-293T: Human embryonic kidney 293T cells; Caco-2: Human colorectal adenocarcinoma cell line; C2BBe1: A subclone of Caco-2; T84: Human colon carcinoma cell line; Human cervical cell line; Calu-3: Human lung epithelial cell line; MRC-5: Human lung fibroblast cell line; PCs: Proprotein convertases.

> Authors detected no cleavage within the spike protein, when they packaged the pseudoviruses to HEK293T cells pretreated with furin-targeting siRNA. Additionally, they excluded the participation of matrix metalloproteinases (MMPs) in the experiment with the application of MMP inhibitor. Therefore, they confirmed the involvement of furin in SARS-CoV-2 entry into chosen cells. Additionally, furin priming of SARS-CoV-2 spike protein has been associated with the virions recognition and binding by neuropilin 1 (NRP1 - receptor which binds furin-cleaved substrates). Cantuti-Castelvetri et al [21] have shown that, except for ACE2, also NRP1 receptors are involved in SARS-CoV-2 invasion. Although the exact mechanism of NRP1 participation in this process is not elucidated, the Authors found the receptors to markedly enhance the virus infectivity. Apart from furin involvement, Shang et al[8] also observed the association of other aforementioned peptidases with SARS-CoV-2 invasion. They noticed the reduction in the pseudovirus entry to the three studied cell lines, caused by the application of both serine proteases (camostat) and cysteine cathepsins (E64) inhibitors. Furthermore, the pseudovirus entry to HeLa cells was more markedly reduced when either camostat or E64d was applied in the cells pretreated with proprotein convertases inhibitor[8]. Therefore, it might be concluded that depending on the type of target cells, TMPRSS2, lysosomal cathepsins, and furin might be involved in the activation of SARS-CoV-2 entry exhibiting cumulative/ overlapping final effect (Figure 2). The observations of Shang et al[8] and Cantuti-Castelvetri *et al*[21] are collected in Table 1.

Cysteine cathepsins as a target in search for COVID-19 therapy

Focusing on cathepsin L expression as a target in search for an anti-Covid-19 drug, Smieszek et al[22] have tested an array of medications applied in clinical practice. They found amantadine (a drug used previously to treat influenza A, and now applied in neurological diseases including Parkinson's disease) to be a promising compound. Being able to accumulate in lysosomes and alkalize them, amantadine belongs to lysosomotropic agents. Such compounds inactivate lysosomal enzymes including CCs whose optimal pH lies below 5. The Authors demonstrated a significant reduction in cathepsin L gene expression using amantadine, however also other cysteine cathepsin

genes expression was inhibited, including cathepsins B and K, with the most pronounced effect observed for cathepsin H. Therefore, it might be hypothesized that this is also cathepsin H which plays an important role in the processing of the SARS-2-S spike protein. The activity of cathepsin H would have been inhibited similarly to other lysosomal cathepsins in the aforementioned experiments (raising pH and/or using E64 inhibitor), which demonstrated a significant reduction of SARS-2-Smediated entry to the studied cells. However, in comparison with thoroughly studied cathepsins B, L, S, and K, there is much less scientific data referring to cathepsin H.

Amantadine efficiency in COVID-19 treatment has been suggested by Rejdak et al [23] who documented no clinical manifestations of COVID-19 infection in 22 patients in spite of the confirmation of SARS-CoV-2 presence with rRT-PCR testing in all of these individuals. The patients had been treated with either amantadine or memantine, for at least 3 months prior to the infection exposure, due to their conditions (multiple sclerosis, Parkinson's disease or cognitive impairment). Therefore, amantadine seems to be a promising treatment for COVID-19 patients. The effect of amantadine may be associated with the down-regulation of cysteine cathepsins (L and/or H) in the endolysosomal compartment and/or the disturbance of viroporin protein channel probably involved in the viral RNA release, as suggested for SARS-CoV[24].

As discussed before, the generation of anti-CCs medicinal drugs is a problematic issue (associated with the observation of unfavorable side effects). Hence, the attitude aimed at screening the already existing therapeutics, which would lower the activity of proteases involved in SARS-CoV-2 activation, seems a rational approach.

EFFECT OF SARS-CoV-2 ON GASTROINTESTINAL TRACT

Gastrointestinal symptoms and fecal virus shedding in COVID-19 patients

A significant amount of scientific evidence accumulated so far points to gastrointestinal tract (GIT), especially its lower part, as a target organ affected by SARS-CoV-2, beside the respiratory system[25]. Apart from the typical pulmonary symptoms (cough, fever, shortness of breath), some of the patients (around 4%-50% individuals) present with digestive symptoms like diarrhea, nausea, vomiting and abdominal pain[26]. Moreover, the virus mRNA presence in stool samples has been observed in some patients, often persisting long after its disappearance from the respiratory tract. Wu et al [27] have documented SARS-CoV-2 mRNA presence in the feces of more than half of the studied patients, with duration for up to 5 wk after its vanishing from the respiratory tract specimens. It might suggest active proliferation of the virus in the gastrointestinal tract of some patients. Similarly, Xiao et al[28] have observed fecal virus shedding in more than 50% of the studied patients, and in over 20% of them the duration of positive results in stool exceeded the virus presence in the respiratory samples. Additionally, the Authors detected the protein parts of the virus (as well as the presence of ACE2 receptors) in gastrointestinal epithelial cells. The virus replication in rectal tissue derived from a COVID-19 patient has been noted by Qian et al[29] who detected SARS-CoV-2 components in the intestinal epithelial cells (but mainly in intestinal lymphocytes and macrophages). The Authors hypothesize that, like in the case of influenza virus, it is possible for SARS-CoV-2 virions to be transported from the respiratory tract to the GIT via the immune cells. In the metaanalysis of 60 studies comprising over 4000 COVID-19 patients, performed by Cheung et al[30], 17.6% of the patients exhibited gastrointestinal symptoms, and in almost 50% (30%-70%) the virus mRNA was detected in the feces. Most of these positive stool samples (above 70%) were collected after the loss of the virus from the respiratory specimens.

However, it is still not clear, whether the virus is transmittable *via* fecal-oral route. Whereas Wang *et al*[31] detected live virus in the fecal samples, Zang *et al*[11] demonstrated the inactivation of the virions released into the intestinal lumen in the environment simulating human colonic fluid. Moreover, the Authors did not manage to recover infectious virus from the stool specimens.

Ability of SARS-CoV-2 to productively infect human GIT cells via ACE2 and TMPRSS

Several studies have demonstrated the invasion and replication of SARS-CoV-2 in the human GIT cells. Chu et al^[32] in their ex-vivo experiments on human intestinal tissues have evidenced the ability of SARS-CoV-2 to infect, proliferate and release infectious virus particles from intestinal cells. In comparison with SARS-CoV, SARS-CoV-2 replicated less efficiently and brought about less damages in the human intestinal



epithelium, but evoked greater response of innate immune system (inducing the expression of proinflammatory mediators such as interferons and interleukins). Hence, the Authors suggested that the gastrointestinal tract might serve as an alternative route of virus dissemination. The vulnerability of the human GIT epithelial cells to SARS-CoV-2 invasion via ACE2 and TMPRSS-2 receptors has been confirmed in other studies with the application of intestinal cell lines models, as well as human small intestinal organoids - hSIOs[11,33-35]. Lee et al[33] investigated the growth of SARS-CoV-2 in a human GIT cell line model; C2BBe1 (a subclone of human epithelial colorectal adenocarcinoma cells: Caco-2). C2BBe1 (genetically and structurally resembling the brush border epithelial cells in the human GIT[36,37], and expressing moderate level of ACE2 and high level of TMPRSS2[33]) exhibited the greatest susceptibility to the virus. SARS-CoV-2 virions invaded and replicated in these cells, as well as in Caco-2[33,35] and T84 (human colon carcinoma) cells[35]. Furthermore, Stanifer et al[35] demonstrated SARS-CoV-2 infection of human colon organoids followed by active virions replication. These observations are in agreement with other studies on hSIOs[11,34]. Zang et al[11] reported productive infection of SARS-CoV-2 in ACE2(+) mature enterocytes, dependent on TMPRSS2 and TMPRSS4 receptors in human small intestinal enteroids. The Authors noted the role of an additional serine protease: TMPRSS4 which heightened the effect of TMPRSS2. Also, the two serine proteases enhanced SARS-CoV-2 spike protein-induced cell-cell fusion observed by the Authors.

ACE2 receptors (except for the respiratory system) are present in a variety of other organs including the gastrointestinal tract, where a great number of these proteins has been detected in the lower part of GIT[38]. Unlike in the upper segment (oral cavity, esophagus, stomach), high expression of ACE2 (both mRNA and protein) is observed in the small intestine (the greatest level), colon and rectum, as well as in the gall bladder[38,39]. Actually, ACE2 expression in the small intestine is much higher in comparison with all other organs in the human organism including the respiratory tract[11]. Specifically, ACE2 is present in the enterocyte cytoplasm and in the apical brush border, as well as in the glandular cells (in the lining epithelium of the lower GIT)[37]. The expression of ACE2 receptors has been exhibited to increase upon enterocytes differentiation. Lee et al [33] observed that (unlike constitutively expressed TMPRSS2), the expression of ACE2 receptors was significantly stimulated in the experiment inducing C2BBe1 enterocytes differentiation, associated with the generation of more pronounced features typical for brush border cells. Similarly, Zang et al[11] detected the greatest expression of ACE2 in mature brush border enterocytes, and Lamers et al[34] noticed around 1000-fold increase in ACE2 mRNA expression upon enterocytes differentiation. Zang et al[11] observed all studied receptors (ACE2, as well as TMPRSS2 and TMPRSS4) to be correlated with the virus invasiveness, similarly as Lee *et al*[33] noted a strong correlation between TMPRSS2 (although not ACE2) and viral RNA levels in the studied human epithelial cell lines (including Caco-2 and C2BBe1). Lee et al^[33] found that the ectopic coexpression of ACE2 and TMPRSS2 in RPMI 2650 cells enhanced viral dissemination by 56.7 times (over 10-fold more in comparison with the sole ACE2 effect - 4.9 times, whereas TMPRSS2 transfection alone did not enhance the level of infectivity). It might be supposed that an effective level of ACE2 receptors is a prerequisite for the virions invasion of epithelial cells, but abundant expression of TMPRSS2 (and possibly TMPRSS4) greatly facilitates ACE2-mediated SARS-CoV-2 dissemination in the human GIT.

The involvement of both ACE2 and TMPRSS2 in SARS-CoV-2 invasion of GIT epithelial cells is in accordance with the reported co-localization of the two receptors in the lower GIT[38-40]. The most abundant expression of both proteins has been detected in the small intestine epithelial cells[38]; especially in the brush border cells [33]. Lee *et al*[39] evaluated single-cell RNA-sequencing datasets from the GIT in search for these two genes co-expression, and found the small intestine enterocytes as well as colonocytes to display the highest proportions of cells co-expression of ACE2, TMPRSS2. Additionally, the Authors checked for the co-expression of ACE2, TMPRSS2 and TMPRSS4, and demonstrated the highest proportions of the three genes co-expression in the progenitor and stem-like epithelial cells in the small intestine. TMPRSS4 is an extra serine protease involved in SARS-CoV-2 activation and invasion, enhancing TMPRSS2 priming effect[11]. Therefore, both ACE2 and TMPRSS2 seem to be the main receptors responsible for SARS-CoV-2 invasion of GIT. The experimental data coming from Lee *et al*[33], Zang *et al*[11], and Stanifer *et al*[35] investigations are displayed in Table 1.

Putative association between SARS-CoV-2-mediated ACE2 disturbance and gastrointestinal symptoms development in COVID-19 patients

The known function of angiotensin converting enzyme 2 (ACE2) is the regulation of systemic arterial blood pressure (renin-angiotensin system). ACE2 catalyzes the conversion of Ang I to angiotensin (1-9) and angiotensin II (Ang II) to angiotensin (1-7), what counteracts the effects of Ang II, leading to decrease in blood pressure and inflammatory processes attenuation (reviewed in[41]). However, the role of ACE2 in GIT (where it is abundantly expressed) seems to be rather associated with the processes occurring in this organ. The analysis of the digestive system specific functional enrichment map for ACE2 gene suggests the involvement of ACE2 in digestion (with reference to its proteolytic activity) and transport of metabolites (the regulation of amino acid transport)[38]. In fact, ACE-2 proteins have been demonstrated to be coupled with sodium-dependent amino acids and glucose transporters. ACE2 is a chaperone for the sodium-dependent amino acid transporter B0AT1 which is involved in transport of neutral amino acids[42]. Moreover, ACE2 participates in the regulation of gut microbiota homeostasis[43,44].

Therefore, as proposed by Kumar et al[38], it might be hypothesized that SARS-CoV-2-associated dysregulation of ACE2 receptors in the human GIT may be involved in the mechanism of GIT symptoms development in COVID-19 patients.

CONCLUSION

In light of the presented studies, the impact of SARS-CoV-2 virus on GIT is evident, with the most substantiated involvement of ACE2 and TMPRSS2. However, except for these two receptors (and probably TMPRSS4), also other proteases might be implicated in SARS-CoV-2 invasion of GIT, as well as the development of the observed symptoms. Ubiquitously expressed furin and cathepsin L may be involved in spike protein processing, contributing to the virus invasiveness. Cathepsin L (and other CCs) might participate in the endolysosomal processing of the spike protein following ACE2-mediated endocytosis of the virion particles. Additionally, SARS-CoV-2 induced inflammatory cytokines could stimulate the secretion of cathepsin L. Therefore, extracellular cathepsin L may contribute both to the spike protein processing, as well as degradation/remodeling of the ECM components and membrane-bound receptors including TMPRSS2/4 and ACE2. The resulting events might accelerate the inflammatory processes disturbing the digestion/absorption of nutrients yielding the observed symptoms such as diarrhea. However, more research is required, since the participation of furin and lysosomal cathepsins in SARS-CoV-2 GIT-invasion is more speculative.

REFERENCES

- 1 Izaguirre G. The Proteolytic Regulation of Virus Cell Entry by Furin and Other Proprotein Convertases. Viruses 2019; 11 [PMID: 31505793 DOI: 10.3390/v11090837]
- Wrapp D, Wang N, Corbett KS, Goldsmith JA, Hsieh CL, Abiona O, Graham BS, McLellan JS. 2 Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. Science 2020; 367: 1260-1263 [PMID: 32075877 DOI: 10.1126/science.abb2507]
- Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, Schiergens TS, 3 Herrler G, Wu NH, Nitsche A, Müller MA, Drosten C, Pöhlmann S. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. Cell 2020; **181**: 271-280.e8 [PMID: 32142651 DOI: 10.1016/j.cell.2020.02.052]
- Blaess M, Kaiser L, Sauer M, Csuk R, Deigner HP. COVID-19/SARS-CoV-2 Infection: Lysosomes and Lysosomotropism Implicate New Treatment Strategies and Personal Risks. Int J Mol Sci 2020; 21 [PMID: 32668803 DOI: 10.3390/ijms21144953]
- Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W, Si HR, Zhu Y, Li B, Huang CL, Chen HD, 5 Chen J, Luo Y, Guo H, Jiang RD, Liu MQ, Chen Y, Shen XR, Wang X, Zheng XS, Zhao K, Chen QJ, Deng F, Liu LL, Yan B, Zhan FX, Wang YY, Xiao GF, Shi ZL. A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature 2020; 579: 270-273 [PMID: 32015507 DOI: 10.1038/s41586-020-2012-71
- 6 Walls AC, Park YJ, Tortorici MA, Wall A, McGuire AT, Veesler D. Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein. Cell 2020; 181: 281-292.e6 [PMID: 32155444 DOI: 10.1016/j.cell.2020.02.058]
- Zhu G, Zhu C, Zhu Y, Sun F. Minireview of progress in the structural study of SARS-CoV-2 7 proteins. Curr Res Microb Sci 2020; 1: 53-61 [PMID: 33236001 DOI: 10.1016/j.crmicr.2020.06.003]



- Shang J, Wan Y, Luo C, Ye G, Geng Q, Auerbach A, Li F. Cell entry mechanisms of SARS-CoV-2. 8 Proc Natl Acad Sci U S A 2020; 117: 11727-11734 [PMID: 32376634 DOI: 10.1073/pnas.2003138117
- 9 Wei J, Alfajaro MM, DeWeirdt PC, Hanna RE, Lu-Culligan WJ, Cai WL, Strine MS, Zhang SM, Graziano VR, Schmitz CO, Chen JS, Mankowski MC, Filler RB, Ravindra NG, Gasque V, de Miguel FJ, Patil A, Chen H, Oguntuyo KY, Abriola L, Surovtseva YV, Orchard RC, Lee B, Lindenbach BD, Politi K, van Dijk D, Kadoch C, Simon MD, Yan Q, Doench JG, Wilen CB. Genome-wide CRISPR Screens Reveal Host Factors Critical for SARS-CoV-2 Infection. Cell 2021; 184: 76-91.e13 [PMID: 33147444 DOI: 10.1016/j.cell.2020.10.028]
- 10 Pišlar A, Mitrović A, Sabotič J, Pečar Fonović U, Perišić Nanut M, Jakoš T, Senjor E, Kos J. The role of cysteine peptidases in coronavirus cell entry and replication: The therapeutic potential of cathepsin inhibitors. PLoS Pathog 2020; 16: e1009013 [PMID: 33137165 DOI: 10.1371/journal.ppat.1009013]
- Zang R, Gomez Castro MF, McCune BT, Zeng Q, Rothlauf PW, Sonnek NM, Liu Z, Brulois KF, 11 Wang X, Greenberg HB, Diamond MS, Ciorba MA, Whelan SPJ, Ding S. TMPRSS2 and TMPRSS4 promote SARS-CoV-2 infection of human small intestinal enterocytes. Sci Immunol 2020; 5 [PMID: 32404436 DOI: 10.1126/sciimmunol.abc3582]
- Berdowska I. Cysteine proteases as disease markers. Clin Chim Acta 2004; 342: 41-69 [PMID: 12 15026265 DOI: 10.1016/j.cccn.2003.12.016]
- 13 Vizovišek M, Fonović M, Turk B. Cysteine cathepsins in extracellular matrix remodeling: Extracellular matrix degradation and beyond. Matrix Biol 2019; 75-76: 141-159 [PMID: 29409929 DOI: 10.1016/j.matbio.2018.01.024]
- Vizovišek M, Vidak E, Javoršek U, Mikhaylov G, Bratovš A, Turk B. Cysteine cathepsins as 14 therapeutic targets in inflammatory diseases. Expert Opin Ther Targets 2020; 24: 573-588 [PMID: 32228244 DOI: 10.1080/14728222.2020.1746765]
- Vidak E, Javoršek U, Vizovišek M, Turk B. Cysteine Cathepsins and their Extracellular Roles: 15 Shaping the Microenvironment. Cells 2019; 8 [PMID: 30897858 DOI: 10.3390/cells8030264]
- 16 Gomes CP, Fernandes DE, Casimiro F, da Mata GF, Passos MT, Varela P, Mastroianni-Kirsztajn G, Pesquero JB. Cathepsin L in COVID-19: From Pharmacological Evidences to Genetics. Front Cell Infect Microbiol 2020; 10: 589505 [PMID: 33364201 DOI: 10.3389/fcimb.2020.589505]
- 17 Menzel K, Hausmann M, Obermeier F, Schreiter K, Dunger N, Bataille F, Falk W, Scholmerich J, Herfarth H, Rogler G. Cathepsins B, L and D in inflammatory bowel disease macrophages and potential therapeutic effects of cathepsin inhibition in vivo. Clin Exp Immunol 2006; 146: 169-180 [PMID: 16968411 DOI: 10.1111/j.1365-2249.2006.03188.x]
- 18 Xu S, Zhang H, Yang X, Qian Y, Xiao Q. Inhibition of cathepsin L alleviates the microglia-mediated neuroinflammatory responses through caspase-8 and NF-KB pathways. Neurobiol Aging 2018; 62: 159-167 [PMID: 29154036 DOI: 10.1016/j.neurobiolaging.2017.09.030]
- Cao Y, Liu X, Li Y, Lu Y, Zhong H, Jiang W, Chen AF, Billiar TR, Yuan H, Cai J. Cathepsin L 19 activity correlates with proteinuria in chronic kidney disease in humans. Int Urol Nephrol 2017; 49: 1409-1417 [PMID: 28534128 DOI: 10.1007/s11255-017-1626-7]
- Ou X, Liu Y, Lei X, Li P, Mi D, Ren L, Guo L, Guo R, Chen T, Hu J, Xiang Z, Mu Z, Chen X, Chen J, Hu K, Jin Q, Wang J, Qian Z. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. Nat Commun 2020; 11: 1620 [PMID: 32221306 DOI: 10.1038/s41467-020-15562-91
- 21 Cantuti-Castelvetri L, Ojha R, Pedro LD, Djannatian M, Franz J, Kuivanen S, van der Meer F, Kallio K, Kaya T, Anastasina M, Smura T, Levanov L, Szirovicza L, Tobi A, Kallio-Kokko H, Österlund P, Joensuu M, Meunier FA, Butcher SJ, Winkler MS, Mollenhauer B, Helenius A, Gokce O, Teesalu T, Hepojoki J, Vapalahti O, Stadelmann C, Balistreri G, Simons M. Neuropilin-1 facilitates SARS-CoV-2 cell entry and infectivity. Science 2020; 370: 856-860 [PMID: 33082293 DOI: 10.1126/science.abd2985]
- Smieszek SP, Przychodzen BP, Polymeropoulos MH. Amantadine disrupts lysosomal gene 22 expression: A hypothesis for COVID19 treatment. Int J Antimicrob Agents 2020; 55: 106004 [PMID: 32361028 DOI: 10.1016/j.ijantimicag.2020.106004]
- 23 Rejdak K, Grieb P. Adamantanes might be protective from COVID-19 in patients with neurological diseases: multiple sclerosis, parkinsonism and cognitive impairment. Mult Scler Relat Disord 2020; 42: 102163 [PMID: 32388458 DOI: 10.1016/j.msard.2020.102163]
- Torres J, Maheswari U, Parthasarathy K, Ng L, Liu DX, Gong X. Conductance and amantadine 24 binding of a pore formed by a lysine-flanked transmembrane domain of SARS coronavirus envelope protein. Protein Sci 2007; 16: 2065-2071 [PMID: 17766393 DOI: 10.1110/ps.062730007]
- Sahu T, Mehta A, Ratre YK, Jaiswal A, Vishvakarma NK, Bhaskar LVKS, Verma HK. Current 25 understanding of the impact of COVID-19 on gastrointestinal disease: Challenges and openings. World J Gastroenterol 2021; 27: 449-469 [PMID: 33642821 DOI: 10.3748/wjg.v27.i6.449]
- Pan L, Mu M, Yang P, Sun Y, Wang R, Yan J, Li P, Hu B, Wang J, Hu C, Jin Y, Niu X, Ping R, Du 26 Y, Li T, Xu G, Hu Q, Tu L. Clinical Characteristics of COVID-19 Patients With Digestive Symptoms in Hubei, China: A Descriptive, Cross-Sectional, Multicenter Study. Am J Gastroenterol 2020; 115: 766-773 [PMID: 32287140 DOI: 10.14309/ajg.0000000000000620]
- 27 Wu Y, Guo C, Tang L, Hong Z, Zhou J, Dong X, Yin H, Xiao Q, Tang Y, Qu X, Kuang L, Fang X, Mishra N, Lu J, Shan H, Jiang G, Huang X. Prolonged presence of SARS-CoV-2 viral RNA in faecal samples. Lancet Gastroenterol Hepatol 2020; 5: 434-435 [PMID: 32199469 DOI: 10.1016/S2468-1253(20)30083-2]



- 28 Xiao F, Tang M, Zheng X, Liu Y, Li X, Shan H. Evidence for Gastrointestinal Infection of SARS-CoV-2. *Gastroenterology* 2020; 158: 1831-1833.e3 [PMID: 32142773 DOI: 10.1053/j.gastro.2020.02.055]
- 29 Qian Q, Fan L, Liu W, Li J, Yue J, Wang M, Ke X, Yin Y, Chen Q, Jiang C. Direct evidence of active SARS-CoV-2 replication in the intestine. *Clin Infect Dis* 2020 [PMID: 32638022 DOI: 10.1093/cid/ciaa925]
- 30 Cheung KS, Hung IFN, Chan PPY, Lung KC, Tso E, Liu R, Ng YY, Chu MY, Chung TWH, Tam AR, Yip CCY, Leung KH, Fung AY, Zhang RR, Lin Y, Cheng HM, Zhang AJX, To KKW, Chan KH, Yuen KY, Leung WK. Gastrointestinal Manifestations of SARS-CoV-2 Infection and Virus Load in Fecal Samples From a Hong Kong Cohort: Systematic Review and Meta-analysis. *Gastroenterology* 2020; **159**: 81-95 [PMID: 32251668 DOI: 10.1053/j.gastro.2020.03.065]
- 31 Wang W, Xu Y, Gao R, Lu R, Han K, Wu G, Tan W. Detection of SARS-CoV-2 in Different Types of Clinical Specimens. *JAMA* 2020; **323**: 1843-1844 [PMID: 32159775 DOI: 10.1001/jama.2020.3786]
- 32 Chu H, Chan JF, Wang Y, Yuen TT, Chai Y, Shuai H, Yang D, Hu B, Huang X, Zhang X, Hou Y, Cai JP, Zhang AJ, Zhou J, Yuan S, To KK, Hung IF, Cheung TT, Ng AT, Hau-Yee Chan I, Wong IY, Law SY, Foo DC, Leung WK, Yuen KY. SARS-CoV-2 Induces a More Robust Innate Immune Response and Replicates Less Efficiently Than SARS-CoV in the Human Intestines: An Ex Vivo Study With Implications on Pathogenesis of COVID-19. *Cell Mol Gastroenterol Hepatol* 2021; 11: 771-781 [PMID: 33010495 DOI: 10.1016/j.jcmgh.2020.09.017]
- 33 Lee S, Yoon GY, Myoung J, Kim SJ, Ahn DG. Robust and persistent SARS-CoV-2 infection in the human intestinal brush border expressing cells. *Emerg Microbes Infect* 2020; 9: 2169-2179 [PMID: 32969768 DOI: 10.1080/22221751.2020.1827985]
- 34 Lamers MM, Beumer J, van der Vaart J, Knoops K, Puschhof J, Breugem TI, Ravelli RBG, Paul van Schayck J, Mykytyn AZ, Duimel HQ, van Donselaar E, Riesebosch S, Kuijpers HJH, Schipper D, van de Wetering WJ, de Graaf M, Koopmans M, Cuppen E, Peters PJ, Haagmans BL, Clevers H. SARS-CoV-2 productively infects human gut enterocytes. *Science* 2020; **369**: 50-54 [PMID: 32358202 DOI: 10.1126/science.abc1669]
- 35 Stanifer ML, Kee C, Cortese M, Zumaran CM, Triana S, Mukenhirn M, Kraeusslich HG, Alexandrov T, Bartenschlager R, Boulant S. Critical Role of Type III Interferon in Controlling SARS-CoV-2 Infection in Human Intestinal Epithelial Cells. *Cell Rep* 2020; **32**: 107863 [PMID: 32610043 DOI: 10.1016/j.celrep.2020.107863]
- 36 Peterson MD, Mooseker MS. Characterization of the enterocyte-like brush border cytoskeleton of the C2BBe clones of the human intestinal cell line, Caco-2. *J Cell Sci* 1992; 102 (Pt 3): 581-600 [PMID: 1506435 DOI: 10.1242/jcs.102.3.581]
- 37 Klijn C, Durinck S, Stawiski EW, Haverty PM, Jiang Z, Liu H, Degenhardt J, Mayba O, Gnad F, Liu J, Pau G, Reeder J, Cao Y, Mukhyala K, Selvaraj SK, Yu M, Zynda GJ, Brauer MJ, Wu TD, Gentleman RC, Manning G, Yauch RL, Bourgon R, Stokoe D, Modrusan Z, Neve RM, de Sauvage FJ, Settleman J, Seshagiri S, Zhang Z. A comprehensive transcriptional portrait of human cancer cell lines. *Nat Biotechnol* 2015; 33: 306-312 [PMID: 25485619 DOI: 10.1038/nbt.3080]
- 38 Kumar A, Faiq MA, Pareek V, Raza K, Narayan RK, Prasoon P, Kumar P, Kulandhasamy M, Kumari C, Kant K, Singh HN, Qadri R, Pandey SN, Kumar S. Relevance of SARS-CoV-2 related factors ACE2 and TMPRSS2 expressions in gastrointestinal tissue with pathogenesis of digestive symptoms, diabetes-associated mortality, and disease recurrence in COVID-19 patients. *Med Hypotheses* 2020; 144: 110271 [PMID: 33254575 DOI: 10.1016/j.mehy.2020.110271]
- 39 Lee JJ, Kopetz S, Vilar E, Shen JP, Chen K, Maitra A. Relative Abundance of SARS-CoV-2 Entry Genes in the Enterocytes of the Lower Gastrointestinal Tract. *Genes (Basel)* 2020; 11 [PMID: 32545271 DOI: 10.3390/genes11060645]
- 40 Gkogkou E, Barnasas G, Vougas K, Trougakos IP. Expression profiling meta-analysis of ACE2 and TMPRSS2, the putative anti-inflammatory receptor and priming protease of SARS-CoV-2 in human cells, and identification of putative modulators. *Redox Biol* 2020; 36: 101615 [PMID: 32863223 DOI: 10.1016/j.redox.2020.101615]
- 41 Wiese O, Zemlin AE, Pillay TS. Molecules in pathogenesis: angiotensin converting enzyme 2 (ACE2). J Clin Pathol 2021; 74: 285-290 [PMID: 32759311 DOI: 10.1136/jclinpath-2020-206954]
- 42 **Singer D**, Camargo SM. Collectrin and ACE2 in renal and intestinal amino acid transport. *Channels* (*Austin*) 2011; **5**: 410-423 [PMID: 21814048 DOI: 10.4161/chan.5.5.16470]
- 43 Camargo SM, Singer D, Makrides V, Huggel K, Pos KM, Wagner CA, Kuba K, Danilczyk U, Skovby F, Kleta R, Penninger JM, Verrey F. Tissue-specific amino acid transporter partners ACE2 and collectrin differentially interact with hartnup mutations. *Gastroenterology* 2009; 136: 872-882 [PMID: 19185582 DOI: 10.1053/j.gastro.2008.10.055]
- 44 Hashimoto T, Perlot T, Rehman A, Trichereau J, Ishiguro H, Paolino M, Sigl V, Hanada T, Hanada R, Lipinski S, Wild B, Camargo SM, Singer D, Richter A, Kuba K, Fukamizu A, Schreiber S, Clevers H, Verrey F, Rosenstiel P, Penninger JM. ACE2 Links amino acid malnutrition to microbial ecology and intestinal inflammation. *Nature* 2012; 487: 477-481 [PMID: 22837003 DOI: 10.1038/nature11228]

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MINIREVIEWS

Endoscopic anti-reflux therapy for gastroesophageal reflux disease

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Abstract

Gastroesophageal reflux disease has an increasing incidence and prevalence worldwide. A significant proportion of patients have a suboptimal response to proton pump inhibitors or are unwilling to take lifelong medication due to concerns about long-term adverse effects. Endoscopic anti-reflux therapies offer a minimally invasive option for patients unwilling to undergo surgical treatment or take lifelong medication. The best candidates are those with a good response to proton pump inhibitors and without a significant sliding hiatal hernia. Transoral incisionless fundoplication and nonablative radiofrequency are the techniques with the largest body of evidence and that have been tested in several randomized clinical trials. Band-assisted ligation techniques, anti-reflux mucosectomy, antireflux mucosal ablation, and new plication devices have yielded promising results in recent noncontrolled studies. Nonetheless, the role of endoscopic procedures remains controversial due to limited long-term and comparative data, and no consensus exists in current clinical guidelines. This review provides an updated summary focused on the patient selection, technical details, clinical success, and safety of current and future endoscopic anti-reflux techniques.

Key Words: Treatment; Gastroesophageal reflux; Transoral incisionless fundoplication; Anti-reflux mucosectomy; Anti-reflux mucosal ablation; Stretta

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Core Tip: Gastroesophageal reflux disease is a common disorder that impacts quality of life. Endoscopic anti-reflux therapies are intended to offer an alternative for patients unwilling to undergo surgical treatment or take lifelong medication. Several techniques, such as transoral incisionless fundoplication, nonablative radiofrequency, plication methods, and anti-reflux mucosectomy, have shown encouraging results, but their role in the management of gastroesophageal reflux disease remains controversial. Careful patient selection and awareness of the advantages and disadvantages of each technique are essential to optimize outcomes. We herein provide an updated review of the technical aspects, clinical success, and safety of the principle endoscopic anti-reflux procedures.

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INTRODUCTION

Gastroesophageal reflux disease (GERD) is a condition that develops when reflux of stomach contents causes troublesome symptoms or complications in the esophagus or beyond [1,2]. GERD is very frequent worldwide, with a prevalence ranging from 7.4% in Southern Asia to 19.6% in Central America, and it affects both sexes similarly[3]. The increment in aging and obesity, both predisposing factors for GERD, may increase its impact in the near future even further[4]. Many other factors also favor GERD exacerbation, including tobacco and certain drugs, such as calcium blockers and tricyclic antidepressants[5,6]. GERD negatively affects quality of life and imposes economic and productivity loss burdens[7].

Although the cause of GERD is still incompletely understood, several underlying predisposing pathophysiological mechanisms have been described. While low esophageal sphincter (LES) basal pressure may facilitate reflux after abdominal strain or during swallowing, a more pertinent mechanism is transient LES relaxation (TLESR), which can be associated with esophageal shortening[8,9]. Gastroesophageal junction (GEJ) disruption due to a hiatal hernia (HH) constitutes an additional factor because it contributes to LES incompetence and also displaces the acid pocket closer to the esophageal mucosa[10,11]. Altered visceral sensitivity has a bidirectional effect in GERD, magnifying symptoms in patients without mucosal injury and reducing symptom awareness in Barrett's esophagus patients[12]. Esophageal hypomotility, low saliva production, and other mechanisms such as certain breathing patterns may also contribute to GERD[13].

The management of GERD is multimodal. Lifestyle modifications such as weight loss, tobacco cessation, and, in selected cases, postural advice[14] have proven efficacy and may be sufficient in mild cases. Drug therapy occupies the next level, with proton pump inhibitors (PPIs) having a huge impact on GERD treatment due to high esophagitis healing rates, surpassing the performance of histamine receptor antagonists and exhibiting high cost-efficacy [15,16]. They are the cornerstone of medical GERD treatment. Anti-reflux surgery (ARS), namely laparoscopic fundoplication, is the last step in GERD management. Its objectives are as follows: (1) LES fixation to the hiatus and intraabdominal segment length augmentation; (2) LES basal pressure increase; and (3) hiatal repair. The latter aspect appears crucial because hiatal repair itself impacts the length and pressure of the LES more than fundoplication[17]. Randomized controlled trials (RCTs) have failed to demonstrate a clear long-term superiority of ARS over PPIs[18]. Consequently, ARS is reserved for patients who do not respond to PPIs, do not tolerate them due to adverse effects, or are unwilling to maintain them in the long term.

PPI refractoriness probably constitutes the most frequent indication for surgery, although it is a confusing term and thus deserves further consideration. The same concept frequently encompasses vastly different realities. Refractoriness can be partial or complete, a distinction that is clinically relevant. Recent and major trials have defined the grade of refractoriness needed to meet inclusion criteria^[19]. Subsequently,



symptoms can persist for very different reasons, such as poor adherence to medical therapies, absence of a relationship with reflux (e.g., functional heartburn), or objectively proven reflux persistence despite proper medical treatment. Therefore, guidelines advise a full diagnostic workup before surgery to demonstrate as consistently as possible that the symptoms, whether refractory or not, are objectively secondary to GERD[2,20-27].

In the last 30 years, effort has been made to design endoscopic anti-reflux therapies that serve as a valuable option for GERD management, either as an alternative to ARS or as bridge therapy between pharmacological treatment and surgery. They do not thus far allow hiatal repair and constrain candidate selection to individuals without a HH. In 1979, Angelchik^[28] used a silicon prosthesis as the first endoscopic treatment for GERD. Since then, numerous other treatments have emerged, with many, such as GEJ injections of bulking agents and several plication techniques, disappearing because of low efficacy or unacceptable adverse effects [29-31]. Here, we present a comprehensive review of the endoscopic approaches for the treatment of GERD that have survived the test of time or have recently been designed (Table 1).

INDICATIONS FOR ENDOSCOPIC ANTI-REFLUX THERAPY

Endoscopic therapies should be considered at least in the same scenarios as surgery and should offer some advantages over ARS. Specifically, endoscopic anti-reflux therapy should be considered in PPI nonresponders, in patients who have a contraindication to PPIs or have concerns regarding their long-term adverse effects, and in those who either do not qualify for ARS or refuse it. Ideally, endoscopic techniques should demonstrate noninferior efficacy, alongside a shorter operation time, lower complication rate, and lower secondary long-term morbidity. Finally, they should not preclude a future fundoplication in case of failure.

Laparoscopic fundoplication performed by skilled surgeons has a low short-term morbidity and mortality but can cause significant adverse effects in the medium term, such as dysphagia (in up to 24% of patients), gas-bloat syndrome, and incisional hernia, and revision surgeries are not infrequent^[22]. It fails in 10%-15% of patients in the short term, and long-term studies have shown that more than 30% of patients are still on PPIs years after surgery [22,32]. This constitutes the scenario against which endoscopic therapies should be compared.

The guidelines of the main medical and surgical societies and expert consensus documents published in the last 10 years have addressed the endoscopic alternatives as well as the surgical option. Their recommendations and the level of evidence or consensus that they are based upon are summarized in Table 2[2,20-26,33-35]. Transoral incisionless fundoplication (TIF) and nonablative radiofrequency are considered appropriate in well-selected patients and situations according to recent guidelines.

CURRENT ENDOSCOPIC THERAPIES

Transoral incisionless fundoplication

The aim of TIF is to perform an endoscopic fundoplication by reestablishing the flap valve mechanism with a 3-cm high-pressure zone at the distal esophagus to durably restore LES function[36]. This procedure mirrors ARS by using an endoscopic suturing device with T-fasteners, the EsophyXâ device (EndoGastric Solutions, Inc., Redmond, WA, United States)[37]. These devices have evolved from a longitudinally oriented gastrogastric plication to one with a greater degree of rotational movement, 200° to 300° in circumference and a 2-3-cm length wrap over the distal esophagus below the diaphragm to create full-thickness serosa-to-serosa esophagogastric plications. This easier to use and more automated device can deploy about 20 fasteners without the need for visualization of the stylet/fastener deployment. The objective of the technique is to restore the integrity of the angle of His by firing stabilizing T-fasteners, deployed 2 to 3 cm above the GEJ, with a 270° esophagogastric wrap, to mimic a Toupet surgical fundoplication. The EsophyXâ device was approved in 2007 by the United States Food and Drug Administration as a single-use, two-operator device comprising a tip (tissue retractor, tissue mold and chassis, fasteners over a stylet, and the invaginator) and body (H-fasteners, helix retractor lock, vacuum connection, fastener pusher, helix retractor control, tissue mold knob, gastroscope point of insertion).



Table 1 Comparison of current endoscopic therapies for gastroesophageal reflux disease						
	TIF	MUSE	Stretta®	GERDx™	ARMS/ARMA	Band ligation
Efficacy	++	+	+ -	+	+	+
Safety	+	+	++	+	+	+
Technical difficulty	++	++	+	++	+	+
Add-on device	+	+	+	+	-	-
RCT available	+	-	+	-	-	-
Maximum follow-up (yr)	10	5	10	0.25	3	1
Cost	++	++	++	++	+	+

++: Indicates the highest score; +: Indicates a moderate score or yes; -: Indicates uncertainty; TIF: Transoral incisionless fundoplication; MUSE: Medigus ultrasonic surgical endostapler; GERDxTM: Endoscopic full-thickness plication device; ARMS: Anti-reflux mucosectomy; ARMA: Anti-reflux mucosal ablation; RCT: Randomized controlled trial.

> Optimal candidates for TIF are patients who demonstrate LES incompetence (Hill grade II) without a concomitant HH. TIF 1.0 has been discontinued because TIF 2.0 achieves much better results[36]. The improved procedure has been evaluated in nine noncomparative studies[38-46] and in five RCTs[47-51] comprising 886 patients with moderate GERD without a large HH, Los Angeles grade C or D esophagitis, or Barrett's esophagus (Table 3). Clinical success rates ranged from a modest 50% at 12 mo to as high as 92% at 10 years. Severe adverse events (SAEs) have been reported in 2.4% of patients^[52]. A recent network meta-analysis suggested that the TIF 2.0 procedure manages symptoms and allows PPI discontinuation at rates similar to those of ARS with an improved safety profile and fewer long-term adverse events[53]. A clinical response, defined by an improvement of at least 50% in GERD health-related quality of life (GERD-HRQL) score or remission of heartburn and regurgitation, was observed in 66% of patients treated with TIF. Moreover, TIF had the highest probability of improving GERD-HRQL (0.96), followed by ARS (0.66) and PPIs (0.042). In contrast, ARS had the highest probability of increasing the percent time at pH < 4(0.99), followed by PPIs (0.64) and TIF (0.32)[53]. A review of the published evidence supports the belief that most selected patients undergoing TIF 2.0 experience a longterm elimination of GERD symptoms with no SAEs and that this procedure is a costeffective alternative to ARS.

Medigus ultrasonic surgical endostapler

The Medigus ultrasonic surgical endostapler (MUSE), or MUSE[™] system (Medigus, Omer, Israel), combines microvisual, ultrasonic, and surgical stapling capabilities into one device, which enables a single endoscopist to perform a transoral anterior fundoplication. This flexible surgical endostapler resembles an endoscope with a rigid section holding a cartridge with five standard 4.8-mm titanium surgical staples. The distal tip contains an anvil for bending the staples, two small 21-gauge screws, and an ultrasonic transducer to measure the distance to the cartridge. This method is a threestep procedure: (1) The stapler is advanced into the stomach through an overtube and retroflex; (2) The system is retracted to 3 cm proximal to the GEJ for clamping when the tissue thickness is 1.4-1.6 mm, and the stapler is then fired; and (3) The procedure is repeated to add quintuplets of staples to create an anti-reflux barrier.

This endoscopic stapling system has been evaluated in four noncomparative studies [46,54-56] and in one two-arm case series study [57] including 209 patients with GERD without a HH larger than 3 cm (Table 3). Clinical success rates ranged from 69% to 92% with follow-up durations from 6 mo to 5 years. The risk of SAEs (empyema, hemorrhage, esophageal perforation) was 3.5%. Overall, data on the efficacy and safety of MUSE are scarce and evidence from RCTs is lacking.

Nonablative radiofrequency treatment (Stretta®)

This endoscopic-guided method involves the application of radiofrequency energy to the muscle fibers of the LES and the gastric cardia, through the Stretta® system (Restech, Houston, TX, United States). The Stretta® catheter is introduced over the guidewire and positioned sequentially at three levels: 0.5 cm proximal to the GEJ, at the GEJ, and 0.5 cm below the GEJ. At each level, the balloon basket assembly is



Table 2 Summary of guidelines and consensus recommendations and invasive gastroesophageal reflux disease therapies

Society guidelines		Strength of recommendation,	Endoscopic anti-	Guideline recommendation on	Strength of recommendation	
and year of publication	Indication for surgery	level of evidence, and grade of consensus	reflux therapy addressed	endoscopic anti-reflux therapy	and level of evidence	
ACG guidelines for diagnosis and	Option for long-term treatment	Quality: High. Strength: Strong	Radiofrequency, bulking agents,	Not recommended	Quality: Moderate. Strength: Conditional	
management of GERD, 2013[2]	Generally not recommended in PPI- unresponsive patients	Quality: High. Strength: Strong	endoscopic suturing			
	Refractory patients with objective evidence of ongoing reflux as the cause of symptoms	Quality: Low. Strength: Conditional				
EAES recommendations, 2014[22]	Good response but dependent on long-term PPI therapy, after optimal risk-benefit discussion	Grade: C. Consensus: 100%	Radiofrequency (Stretta [®]), bulking agent injection (Enteryx [®]), plication (EndoCinch [®] , full-	Not enough evidence available to recommend any as an alternative option to surgery	Grade of recommendation: B. Expert consensus: 100%	
	Total or partial refractoriness despite adequate PPI therapy in terms of dosage and intake	Grade: A. Consensus: 100%	thickness plication, EsophyX [®]			
	Well-selected NERD patients and those with hypersensitive esophagus	Grade: C. Consensus: 100%				
American Society of Gastrointestinal Endoscopy: The role of endoscopy in the management of GERD, 2015[95]	Not provided	Not provided	Radiofrequency (Stretta [®]) and transoral incisionless fundoplication	Consider in highly selected patients. No details on selection criteria	Low quality	
Asia-Pacific consensus on refractory GERD management, 2016[23]	Refractory symptoms with objectively documented GERD	Quality: Moderate. Strength: Strong. Consensus: 100%	None	Not applicable	Not applicable	
World Gastroenterology Organisation Global Guidelines, 2017[24]	Large hiatal hernia with volume-related reflux symptoms. Refractory esophagitis. Refractory symptoms documented as caused by GERD. Medication adverse effects	Not specified	Endoscopic therapies in general	Only in the context of clinical trials	Not specified	
SAGES Guidelines on GERD surgical treatment, 2010, and on endoluminal anti-reflux treatments, 2017[21,34]	Appropriately selected GERD patients	Grade A	Transoral incisionless fundoplication	Control of symptoms in appropriately selected patients in the short term; appears to lose effectiveness	Quality: Moderate. Strength: Strong	
			Radiofrequency	Control of symptoms in appropriately selected patients; long-term effect in appropriately selected patients	Quality: Moderate. Strength: Strong	
USA expert panel (surgeons and advanced therapeutic	PPI responders (complete ed or partial)	Appropriate. Consensus: 87%-100%	Transoral incisionless fundoplication	PPI responders (complete or partial), no hernia, any other scenario	Appropriate. Consensus: 93%	
endoscopists) recommendations on GERD management, 2020[25]				PPI responders (complete or partial) or nonresponders, significant hernia, any other scenario	Not appropriate	
	PPI nonresponder, no hernia, heartburn- hypersensitivity, or negative pH-impedance	Appropriateness uncertain		PPI nonresponder, no hernia and acid breakthrough, hypersensitivity or	Appropriate. Consensus: 80%–93%	

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	study negative pH-impedance study for heartburn				
	PPI nonresponder, any other scenario	Appropriate. Consensus: 80%-100%		PPI nonresponder, regurgitation, negative pH-impedance study	Appropriateness uncertain
			Radiofrequency	PPI responders (complete or partial) or nonresponders, no hernia, any scenario	Appropriateness uncertain
				PPI responders (complete or partial) or nonresponders, significant hernia	Not appropriate
ESGE guidelines on endoscopic management of gastrointestinal motility disorders, 2020 [35]	Not applicable	Not applicable	Transoral incisionless fundoplication	Possible role in mild GERD patients who are unwilling to take PPIs or undergo surgery. Against widespread use	Quality: Moderate. Strength: Strong. Consensus: 92.8%
			Medigus Ultrasonic Surgical Endostapler	Insufficient data. Use only in clinical trials	Quality: Low. Strength: Strong. Consensus: 100%
			Radiofrequency	Can be considered in selected patients only, without erosive esophagitis and hiatal hernia	Quality: Moderate. Strength: Weak. Consensus: 92.9%
			Anti-reflux mucosectomy	Against routine use in clinical practice	Quality: Low. Strength: Strong. Consensus: 100%
ESNM/ASNM consensus paper on management of refractory GERD, 2020	Refractory GERD symptoms in patients with proven GERD	Consensus: 100%	Transoral incisionless fundoplication	Short-term benefit in improving regurgitation in carefully selected patients	Consensus: 100%
[26]			Radiofrequency	Variable symptom improvement, limited objective improvement in acid burden or manometric esophagogastric junction features	Consensus: 100%

ACG: American College of Gastroenterology; EAES: European Association of Endoscopic Surgery; SAGES: Society of the Americans Gastrointestinal and Endoscopic Surgeons; GERD: Gastroesophageal reflux disease; ESGE: European Society of Gastrointestinal Endoscopy; ESNM: European Society of Neurogastroenterology and Motility; ASNM: American Society of Neurogastroenterology and Motility; PPIs: Proton pump inhibitors; NERD: Nonerosive reflux disease

> inflated and then four nitinol needle electrodes (22-gauge, 5.5-mm) are extended into the muscular layer to deliver a radiofrequency current and induce a thermal reaction. Next, to deliver radiofrequency energy to four additional points, the catheter is rotated 45° clockwise[58]. The pathophysiological mechanism is not fully understood, but the thermal injury is thought to promote submucosal fibrosis and muscularis propria hypertrophy, which would decrease the frequency of TLESR and GEJ compliance while increasing LES and gastric yield pressures[58].

> The Stretta® procedure has been evaluated in numerous cohort studies and in five RCTs, three with sham therapy and two with PPI use[59] (Tables 1 and 3). The RCT results did not show significant changes in esophageal acid exposure at 6 mo following Stretta[®], compared with the PPI group[60]. Likewise, patients treated with Stretta® presented significant improvements in heartburn symptoms and quality of life in only the short term, compared with a sham procedure, with no long-term data[61-63]. A meta-analysis including 159 patients, limited to four RCTs, confirmed the absence of significant changes in patients with GERD[64]. More recently, a second meta-analysis that included both RCTs and 24 other cohort studies with 2468 evaluated patients^[65] showed a significant postprocedural improvement in quality of life and in heartburn score but no improvement in basal LES pressure. The procedure is safe and well-tolerated, and SAEs are very rare. RCTs and cohort studies reported erosions, mucosal lacerations, gastroparesis, mediastinal inflammation, pneumonia,

Table 3 Clinical success and safety of endoscopic therapies						
Technique	Study design and population	Clinical success, range	Major adverse events, range			
Transoral incisionless fundoplication	No. of RCTs: 5; <i>n</i> = 343	50%-92%	0%-4.4%			
	No. of nonrandomized case series: 9; <i>n</i> = 543					
Medigus ultrasonic surgical endostapler	No. of RCTs: 0	69%-92%	0%-9%			
encostapler	No. of nonrandomized case series: 5; <i>n</i> = 199					
Nonablative radiofrequency (Stretta [®])	No. of RCTs: 5; <i>n</i> = 173	15%-100%	0%-1%			
	No. of nonrandomized case series: 29; <i>n</i> = 2571					
Endoscopic plication device (GERDx™)	No. of RCTs: 0	19 out of 40 patients were off PPIs	10%			
(GERDX ^{***})	No. of nonrandomized case series: 1; $n = 40$					
Band ligation techniques	No. of RCTs: 1; <i>n</i> = 150	43%-54% ¹	0%			
	No. of nonrandomized case series: 2; <i>n</i> = 73					
Anti-reflux mucosectomy	No. of RCTs: 0	58%-100%	0%-17%			
	No. of nonrandomized case series: 12; <i>n</i> = 331					
Anti-reflux mucosal ablation	No. of RCTs: 0	58%-89%	0%-13%			
	No. of nonrandomized case series: 3; <i>n</i> = 130					

¹Clinical success not defined in the randomized controlled trial. There was a significant reduction in gastroesophageal reflux disease health-related quality of life score and 24-h pH-metry outcomes. RCT: Randomized controlled trial; PPIs: Proton pump inhibitors.

and pleural effusion[66].

Endoscopic plication device (GERDx[™])

The GERDxTM device (G-SURG GmbH, Seeon-Seebruck, Germany) uses hydraulic elements for control and requires a slim gastroscope that works as a light source. It is the advanced single-use product of the company that has acquired the Plicator technology after withdrawal of the Plicator device (Ethicon Endo-Surgery, Sommerville, NJ) from the market. The experience with GERDxTM is still minimal, with only two publications in this regard, one of which is an interim analysis by the same authors (Tables 1 and 3).

In a single-center, single-arm trial, Weitzendorfer *et al*[67,68] prospectively assessed the outcomes of 40 patients with refractory GERD treated with the GERDxTM device. Of the 40 patients, 7 underwent LARS before the 3-mo follow-up. The mean De-Meester score was reduced from 46.48 to 20.03 in the 30 patients who completed the follow-up. Of these 30 patients, 18 (60.0%) achieved normal DeMeester score levels. In addition, 3 (10.0%) stated that they were on daily PPI medication after the plication, with 8 (26.7%) taking on-demand medication and 19 (63.3%) off medication. Moderate SAEs were reported by 10% of the patients (a hematoma at the GEJ, a case of pneumonia, a suture passing through the left hepatic lobe, pleural empyema, a severe Mallory-Weiss tear). The single-study evidence, lack of a comparator arm, and the very short follow-up make this endoscopic treatment experimental at this time, necessitating new RCTs to corroborate improvements in quality of life and acid exposure and confirm procedural safety.

Anti-reflux mucosectomy and anti-reflux mucosal ablation

Anti-reflux mucosectomy (ARMS) was first devised in a patient with a Barrett's esophagus-related lesion treated by endoscopic submucosal dissection. The resulting scar improved GERD symptoms and normalized the DeMeester score[69]. This observation led to the first case series, published by Inoue *et al*[69] in 2014. In ARMS, endoscopic resection of the gastric cardiac mucosa is performed to reduce the opening

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of the GEJ. Initial ARMS cases were performed by endoscopic submucosal dissection, but subsequent reports indicated that cap- or band-assisted mucosal resection is faster, easier to perform, and equally effective [70-72]. ARMS has been suggested to suppress the backflow of gastric content and enhance the GEJ flap valve mechanism, but the underlying anti-reflux mechanism is poorly understood [72]. A RCT conducted in animals found that ARMS increased the pressure and volume required to induce fluid passage from the gastric cavity to the esophagus[73]. One clinical study revealed that ARMS increased the integrated relaxation pressure and LES resting pressure but decreased GEJ distensibility, which could hypothetically reduce the frequency of TLESR[72,74].

In 2020, Inoue *et al*^[75] and Hernández Mondragón *et al*^[76] proposed that ablation of the gastric cardiac mucosa by argon plasma coagulation (forced mode 100 W) or a coagulation current applied by an endoknive (spray coagulation 50 W, effect 2) can also induce scar formation and yield similar clinical outcomes. This approach, named anti-reflux mucosal ablation (ARMA), is intended to simplify the procedure, reduce the risk of perforation, and facilitate the retreatment of patients who have failed ARMS

In addition to their technical simplicity, ARMS and ARMA do not require costly add-on devices and can be performed in a standard endoscopy room [72,76]. Key points during ARMS and ARMA are adequate submucosal injection to prevent perforation and the sparing of a rim of healthy mucosa to minimize the risk of GEJ stenosis. The procedure is not standardized, but most authors spare the esophageal mucosa and perform a gastric cardia 270°-320° treatment or mimic a "butterfly" shape by sparing 1 cm of normal mucosa along the greater and lesser curvature[72,75-77].

In total, 15 nonrandomized studies (12 on ARMS[69-72,74,77-83] and three on ARMA[75,76,84]) comprising 461 patients have evaluated the safety and effectiveness of these techniques (Tables 1 and 3). Follow-up ranged from 2 mo to a maximum of 3 years (in two studies[72,76]). Clinical success ranged from 58% to 100% at 2-6 mo[81, 83] and from 72% to 76% at 3 years[72,76]. Dysphagia was the most common adverse event, occurring in about 5% to 10% of the patients. In contrast to what occurs with dysphagia associated with ARS[85], ARMS- and ARMA-associated dysphagia can be easily treated by small-caliber balloon dilation and does not necessarily compromise clinical success^[72,76]. Gastrointestinal perforation is the most feared complication and has been reported in four patients treated with ARMS[72,77,78] and in none treated with ARMA. Given the lack of RCT and long-term data, these techniques should be viewed as experimental and reserved for patients included in research protocols.

Band ligation techniques

Three studies have assessed the outcomes of rubber band placement at the GEJ to reduce the width of the opening of the gastric cardia. Seleem et al [86] performed a RCT that included 150 patients with refractory GERD. The number of bands applied and the frequency of endoscopic sessions were determined according to the narrowing of the GEJ during banding. A maximum of four bands per session were allowed. Followup at 1 year showed a significant improvement in GERD-HRQL score and the number of reflux episodes. Mild dysphagia (25.3%) and epigastric pain (40%) were the most common adverse events, but no SAEs were recorded[86]. Hu et al[87] also reported favorable subjective and 24-h pH-metry outcomes in a case series of 13 patients and named the procedure "peroral endoscopic cardial constriction". The authors placed two single-band ligation devices (Fujinon, Tokyo, Japan) at the greater and lesser curvatures, close to the Z line. The first band was placed approximately 1.0 cm above the cardia along the lesser curvature, whereas the second band was delivered 1.0 cm above the greater curvature^[87]. Finally, a clip was placed at the base of the bands to minimize the risk of band slippage. In 2020, another Chinese group reported favorable results with this technique in a nonrandomized study of 60 patients, with the approach now named "clip band ligation anti-reflux therapy (C-BLART)" [88] (Tables 1 and 3).

Because the above-mentioned RCT does not adhere to the Consolidated Standards of Reporting Trials quality requirements and the two case series were noncontrolled and included a limited number of patients, the technique should currently be viewed as experimental.

FUTURE DIRECTIONS

The history of endoscopic therapies for GERD is replete with encouraging preclinical studies and case series that fail to clear the hurdle of long-term and well-designed



RCTs. The main underlying reasons are the complex and multifactorial pathophysiology of GERD and the often short-lived anatomical changes induced by endoscopic therapies. Moreover, many endoscopic techniques require expensive add-on devices and cumbersome technical steps that have limited their popularization. To complicate further this issue, patient selection has been heterogeneous, and we lack consensus regarding the definition of clinical success or the admissible thresholds of cost and adverse events. Future endoscopic therapies and GERD research should bear all of this in mind.

The first consideration is that only a subset of well-selected GERD patients are good candidates for endoscopic therapies because current techniques remain unable to fix the hiatus, enhance esophageal motility, or normalize LES competence. Artificial intelligence through knowledge-based clinical decision support systems could be of help in the future for improving patient selection. Combined approaches that consider more than one GERD mechanism have been proposed to address this issue, such as a combination of ARMS with a plication method[89] or of TIF with laparoscopic HH repair[90]. Second, technical feasibility is critical for introducing a procedure into clinical practice. The learning curve of anti-reflux endoscopic therapies has not been well-described, and scientific societies have not published curricula documents to guide training. Band ligation, ARMS, and, more recently, ARMA are at very early stages but represent an attractive option from this perspective. Our group is currently performing a double-blind RCT to assess the clinical success and safety of ARMA[91]. Third, patient-reported outcomes are increasingly being recognized by clinicians, regulatory agencies, and patients as highly valuable tools to assess the impact of new interventions. Thus, we believe that studies should place symptoms and GERD-related quality of life as primary endpoints. A "black or white" perspective for clinical success does not reflect the complexity of GERD patients, and partial but significant improvements should also be taken into account. This makes anti-reflux endoscopy not only an alternative to PPIs, but also a complementary tool that can reduce their consumption and partially improve quality of life. A > 50% drop in the GERD-HRQL score or in other validated clinical questionnaires has been used in recent RCTs and appears to be a reasonable approach [52,53]. In addition, more objective GERD parameters (24-h pH-impedance testing, endoscopic esophagitis) and sham/placebo arms are needed to support subjective improvements. Outcome definitions should be in line with recent international consensus [26,27,92,93]. RCTs should include longterm follow-up as part of the trial or as a post-RCT prospective observational phase to assess durability. Finally, endoscopic therapies seem cost-effective, but we need more comparative data with PPI and surgery[94].

CONCLUSION

Endoscopic therapy for GERD aims to offer an alternative to PPIs and ARS in patients without significant diaphragmatic crura impairment. TIF, the technique with the largest body of evidence, has been proven to improve GERD symptoms and acid exposure time and reduce PPI consumption. Nonablative radiofrequency (Stretta®) is the method with the lowest rate of SAEs, but its efficacy has been called into question in recent meta-analyses. Band ligation techniques, ARMS, ARMA, and new plication devices have shown promising results in initial reports and RCTs are eagerly awaited. Careful patient selection, ongoing technical refinements, and RCTs with long-term data are the roadmap to unveil the potential of minimally invasive anti-reflux endoscopic techniques.

REFERENCES

- Vakil N, van Zanten SV, Kahrilas P, Dent J, Jones R; Global Consensus Group. The Montreal 1 definition and classification of gastroesophageal reflux disease: a global evidence-based consensus. Am J Gastroenterol 2006; 101: 1900-1920; quiz 1943 [PMID: 16928254 DOI: 10.1111/j.1572-0241.2006.00630.x]
- Katz PO, Gerson LB, Vela MF. Guidelines for the diagnosis and management of gastroesophageal 2 reflux disease. Am J Gastroenterol 2013; 108: 308-328; quiz 329 [PMID: 23419381 DOI: 10.1038/ajg.2012.444]
- 3 Eusebi LH, Ratnakumaran R, Yuan Y, Solaymani-Dodaran M, Bazzoli F, Ford AC. Global prevalence of, and risk factors for, gastro-oesophageal reflux symptoms: a meta-analysis. Gut 2018; 67: 430-440 [PMID: 28232473 DOI: 10.1136/gutjnl-2016-313589]



- 4 Diaz-Rubio M, Moreno-Elola-Olaso C, Rey E, Locke GR 3rd, Rodriguez-Artalejo F. Symptoms of gastro-oesophageal reflux: prevalence, severity, duration and associated factors in a Spanish population. Aliment Pharmacol Ther 2004; 19: 95-105 [PMID: 14687171 DOI: 10.1046/j.1365-2036.2003.01769.x]
- van Soest EM, Dieleman JP, Siersema PD, Schoof L, Sturkenboom MC, Kuipers EJ. Tricyclic 5 antidepressants and the risk of reflux esophagitis. Am J Gastroenterol 2007; 102: 1870-1877 [PMID: 17511756 DOI: 10.1111/j.1572-0241.2007.01320.x]
- Boeckxstaens G, El-Serag HB, Smout AJ, Kahrilas PJ. Republished: symptomatic reflux disease: the 6 present, the past and the future. Postgrad Med J 2015; 91: 46-54 [PMID: 25583739 DOI: 10.1136/postgradmedj-2013-306393rep]
- Gisbert JP, Cooper A, Karagiannis D, Hatlebakk J, Agréus L, Jablonowski H, Nuevo J. Impact of 7 gastroesophageal reflux disease on work absenteeism, presenteeism and productivity in daily life: a European observational study. Health Qual Life Outcomes 2009; 7: 90 [PMID: 19835583 DOI: 10.1186/1477-7525-7-90]
- Holloway RH, Dent J. Pathophysiology of gastroesophageal reflux. Lower esophageal sphincter 8 dysfunction in gastroesophageal reflux disease. Gastroenterol Clin North Am 1990; 19: 517-535 [PMID: 2228162 DOI: 10.1016/S0889-8553(21)00654-3]
- Iovino P, Theron B, Prew S, Menon S, Trudgill N. The mechanisms associated with reflux episodes in ambulant subjects with gastro-esophageal reflux disease. Neurogastroenterol Motil 2021; 33: e14023 [PMID: 33112052 DOI: 10.1111/nmo.14023]
- 10 Ciriza-de-los-Ríos C, Canga-Rodríguez-Valcárcel F, Castel-de-Lucas I, Lora-Pablos D, de-la-Cruz-Bértolo J, Castellano-Tortajada G. How useful is esophageal high resolution manometry in diagnosing gastroesophageal junction disruption: causes affecting this disruption and its relationship with manometric alterations and gastroesophageal reflux. Rev Esp Enferm Dig 2014; 106: 22-29 [PMID: 24689712 DOI: 10.4321/s1130-010820140001000041
- van Herwaarden MA, Samsom M, Smout AJ. Excess gastroesophageal reflux in patients with hiatus 11 hernia is caused by mechanisms other than transient LES relaxations. Gastroenterology 2000; 119: 1439-1446 [PMID: 11113064 DOI: 10.1053/gast.2000.20191]
- 12 Aziz Q, Fass R, Gyawali CP, Miwa H, Pandolfino JE, Zerbib F. Functional Esophageal Disorders. Gastroenterology 2016 [PMID: 27144625 DOI: 10.1053/j.gastro.2016.02.012]
- 13 Tack J, Pandolfino JE. Pathophysiology of Gastroesophageal Reflux Disease. Gastroenterology 2018; 154: 277-288 [PMID: 29037470 DOI: 10.1053/j.gastro.2017.09.047]
- Kaltenbach T, Crockett S, Gerson LB. Are lifestyle measures effective in patients with 14 gastroesophageal reflux disease? Arch Intern Med 2006; 166: 965-971 [PMID: 16682569 DOI: 10.1001/archinte.166.9.965
- 15 Dent J, Hetzel DJ, MacKinnon MA, Reed WD, Narielvala FM. Evaluation of omeprazole in reflux oesophagitis. Scand J Gastroenterol Suppl 1989; 166: 76-82; discussion 94 [PMID: 2690334 DOI: 10.3109/00365528909091249
- Weijenborg PW, Cremonini F, Smout AJ, Bredenoord AJ. PPI therapy is equally effective in well-16 defined non-erosive reflux disease and in reflux esophagitis: a meta-analysis. Neurogastroenterol Motil 2012; 24: 747-757, e350 [PMID: 22309489 DOI: 10.1111/j.1365-2982.2012.01888.x]
- 17 Louie BE, Kapur S, Blitz M, Farivar AS, Vallières E, Aye RW. Length and pressure of the reconstructed lower esophageal sphincter is determined by both crural closure and Nissen fundoplication. J Gastrointest Surg 2013; 17: 236-243 [PMID: 23188217 DOI: 10.1007/s11605-012-2074-4]
- 18 Garg SK, Gurusamy KS. Laparoscopic fundoplication surgery versus medical management for gastro-oesophageal reflux disease (GORD) in adults. Cochrane Database Syst Rev 2015; CD003243 [PMID: 26544951 DOI: 10.1002/14651858.CD003243.pub3]
- 19 Spechler SJ, Hunter JG, Jones KM, Lee R, Smith BR, Mashimo H, Sanchez VM, Dunbar KB, Pham TH, Murthy UK, Kim T, Jackson CS, Wallen JM, von Rosenvinge EC, Pearl JP, Laine L, Kim AW, Kaz AM, Tatum RP, Gellad ZF, Lagoo-Deenadayalan S, Rubenstein JH, Ghaferi AA, Lo WK, Fernando RS, Chan BS, Paski SC, Provenzale D, Castell DO, Lieberman D, Souza RF, Chey WD, Warren SR, Davis-Karim A, Melton SD, Genta RM, Serpi T, Biswas K, Huang GD. Randomized Trial of Medical versus Surgical Treatment for Refractory Heartburn. N Engl J Med 2019; 381: 1513-1523 [PMID: 31618539 DOI: 10.1056/NEJMoa1811424]
- 20 Kahrilas PJ, Shaheen NJ, Vaezi MF, Hiltz SW, Black E, Modlin IM, Johnson SP, Allen J, Brill JV; American Gastroenterological Association. American Gastroenterological Association Medical Position Statement on the management of gastroesophageal reflux disease. Gastroenterology 2008; 135: 1383-1391, 1391.e1 [PMID: 18789939 DOI: 10.1053/j.gastro.2008.08.045]
- Stefanidis D, Hope WW, Kohn GP, Reardon PR, Richardson WS, Fanelli RD; SAGES Guidelines Committee. Guidelines for surgical treatment of gastroesophageal reflux disease. Surg Endosc 2010; 24: 2647-2669 [PMID: 20725747 DOI: 10.1007/s00464-010-1267-8]
- 22 Fuchs KH, Babic B, Breithaupt W, Dallemagne B, Fingerhut A, Furnee E, Granderath F, Horvath P, Kardos P, Pointner R, Savarino E, Van Herwaarden-Lindeboom M, Zaninotto G; European Association of Endoscopic Surgery (EAES). EAES recommendations for the management of gastroesophageal reflux disease. Surg Endosc 2014; 28: 1753-1773 [PMID: 24789125 DOI: 10.1007/s00464-014-3431-z
- 23 Fock KM, Talley N, Goh KL, Sugano K, Katelaris P, Holtmann G, Pandolfino JE, Sharma P, Ang TL, Hongo M, Wu J, Chen M, Choi MG, Law NM, Sheu BS, Zhang J, Ho KY, Sollano J, Rani AA,



Kositchaiwat C, Bhatia S. Asia-Pacific consensus on the management of gastro-oesophageal reflux disease: an update focusing on refractory reflux disease and Barrett's oesophagus. Gut 2016; 65: 1402-1415 [PMID: 27261337 DOI: 10.1136/gutjnl-2016-311715]

- 24 Hunt R, Armstrong D, Katelaris P, Afihene M, Bane A, Bhatia S, Chen MH, Choi MG, Melo AC, Fock KM, Ford A, Hongo M, Khan A, Lazebnik L, Lindberg G, Lizarzabal M, Myint T, Moraes-Filho JP, Salis G, Lin JT, Vaidya R, Abdo A, LeMair A; Review Team:. World Gastroenterology Organisation Global Guidelines: GERD Global Perspective on Gastroesophageal Reflux Disease. J Clin Gastroenterol 2017; 51: 467-478 [PMID: 28591069 DOI: 10.1097/MCG.0000000000854]
- Gawron AJ, Bell R, Abu Dayyeh BK, Buckley FP, Chang K, Dunst CM, Edmundowicz SA, Jobe B, 25 Lipham JC, Lister D, Canto MI, Smith MS, Starpoli AA, Triadafilopoulos G, Watson TJ, Wilson E, Pandolfino JE, Kaizer A, Van De Voorde Z, Yadlapati R. Surgical and endoscopic management options for patients with GERD based on proton pump inhibitor symptom response: recommendations from an expert U.S. panel. Gastrointest Endosc 2020; 92: 78-87.e2 [PMID: 32007519 DOI: 10.1016/j.gie.2020.01.037
- 26 Zerbib F, Bredenoord AJ, Fass R, Kahrilas PJ, Roman S, Savarino E, Sifrim D, Vaezi M, Yadlapati R, Gyawali CP. ESNM/ANMS consensus paper: Diagnosis and management of refractory gastroesophageal reflux disease. Neurogastroenterol Motil 2021; 33: e14075 [PMID: 33368919 DOI: 10.1111/nmo.14075
- Pauwels A, Boecxstaens V, Andrews CN, Attwood SE, Berrisford R, Bisschops R, Boeckxstaens GE, 27 Bor S, Bredenoord AJ, Cicala M, Corsetti M, Fornari F, Gyawali CP, Hatlebakk J, Johnson SB, Lerut T, Lundell L, Mattioli S, Miwa H, Nafteux P, Omari T, Pandolfino J, Penagini R, Rice TW, Roelandt P, Rommel N, Savarino V, Sifrim D, Suzuki H, Tutuian R, Vanuytsel T, Vela MF, Watson DI, Zerbib F, Tack J. How to select patients for antireflux surgery? Gut 2019; 68: 1928-1941 [PMID: 31375601 DOI: 10.1136/gutjnl-2019-318260]
- 28 Angelchik JP, Cohen R. A new surgical procedure for the treatment of gastroesophageal reflux and hiatal hernia. Surg Gynecol Obstet 1979; 148: 246-248 [PMID: 154176]
- Yew KC, Chuah SK. Antireflux endoluminal therapies: past and present. Gastroenterol Res Pract 29 2013; 2013: 481417 [PMID: 23935608 DOI: 10.1155/2013/481417]
- Filipi CJ, Lehman GA, Rothstein RI, Raijman I, Stiegmann GV, Waring JP, Hunter JG, Gostout CJ, 30 Edmundowicz SA, Dunne DP, Watson PA, Cornet DA. Transoral, flexible endoscopic suturing for treatment of GERD: a multicenter trial. Gastrointest Endosc 2001; 53: 416-422 [PMID: 11275879 DOI: 10.1067/mge.2001.113502]
- Fockens P, Cohen L, Edmundowicz SA, Binmoeller K, Rothstein RI, Smith D, Lin E, Nickl N, 31 Overholt B, Kahrilas PJ, Vakil N, Abdel Aziz Hassan AM, Lehman GA. Prospective randomized controlled trial of an injectable esophageal prosthesis vs a sham procedure for endoscopic treatment of gastroesophageal reflux disease. Surg Endosc 2010; 24: 1387-1397 [PMID: 20198491 DOI: 10.1007/s00464-009-0784-9]
- Oor JE, Roks DJ, Broeders JA, Hazebroek EJ, Gooszen HG. Seventeen-year Outcome of a 32 Randomized Clinical Trial Comparing Laparoscopic and Conventional Nissen Fundoplication: A Plea for Patient Counseling and Clarification. Ann Surg 2017; 266: 23-28 [PMID: 28294958 DOI: 10.1097/SLA.000000000002106
- Falk GW, Fennerty MB, Rothstein RI. AGA Institute medical position statement on the use of 33 endoscopic therapy for gastroesophageal reflux disease. Gastroenterology 2006; 131: 1313-1314 [PMID: 17030198 DOI: 10.1053/j.gastro.2006.08.018]
- 34 Pearl J, Pauli E, Dunkin B, Stefanidis D. SAGES endoluminal treatments for GERD. Surg Endosc 2017; 31: 3783-3790 [PMID: 28643067 DOI: 10.1007/s00464-017-5639-1]
- Weusten BLAM, Barret M, Bredenoord AJ, Familiari P, Gonzalez JM, van Hooft JE, Lorenzo-35 Zúñiga V, Louis H, Martinek J, van Meer S, Neumann H, Pohl D, Prat F, von Renteln D, Savarino E, Sweis R, Tack J, Tutuian R, Ishaq S. Endoscopic management of gastrointestinal motility disorders part 2: European Society of Gastrointestinal Endoscopy (ESGE) Guideline. Endoscopy 2020; 52: 600-614 [PMID: 32462649 DOI: 10.1055/a-1171-3174]
- 36 Chang KJ, Bell R. Transoral Incisionless Fundoplication. Gastrointest Endosc Clin N Am 2020; 30: 267-289 [PMID: 32146946 DOI: 10.1016/j.giec.2019.12.008]
- 37 Ihde GM. The evolution of TIF: transoral incisionless fundoplication. Therap Adv Gastroenterol 2020; **13**: 1756284820924206 [PMID: 32499834 DOI: 10.1177/1756284820924206]
- 38 Testoni PA, Corsetti M, Di Pietro S, Castellaneta AG, Vailati C, Masci E, Passaretti S. Effect of transoral incisionless fundoplication on symptoms, PPI use, and ph-impedance refluxes of GERD patients. World J Surg 2010; 34: 750-757 [PMID: 20091308 DOI: 10.1007/s00268-010-0394-7]
- 39 Ihde GM, Besancon K, Deljkich E. Short-term safety and symptomatic outcomes of transoral incisionless fundoplication with or without hiatal hernia repair in patients with chronic gastroesophageal reflux disease. Am J Surg 2011; 202: 740-746; discussion 746-747 [PMID: 22014853 DOI: 10.1016/j.amjsurg.2011.06.035]
- Narsule CK, Burch MA, Ebright MI, Hess DT, Rivas R Jr, Daly BD, Fernando HC. Endoscopic 40 fundoplication for the treatment of gastroesophageal reflux disease: initial experience. J Thorac Cardiovasc Surg 2012; 143: 228-234 [PMID: 22070927 DOI: 10.1016/j.jtcvs.2011.10.008]
- 41 Bell RC, Barnes WE, Carter BJ, Sewell RW, Mavrelis PG, Ihde GM, Hoddinott KM, Fox MA, Freeman KD, Gunsberger T, Hausmann MG, Dargis D, DaCosta Gill B, Wilson E, Trad KS. Transoral incisionless fundoplication: 2-year results from the prospective multicenter U.S. study. Am Surg 2014; 80: 1093-1105 [PMID: 25347499 DOI: 10.1177/000313481408001124]



- Wilson EB, Barnes WE, Mavrelis PG, Carter BJ, Bell RC, Sewell RW, Ihde GM, Dargis D, 42 Hoddinott KM, Shughoury AB, Gill BD, Fox MA, Turgeon DG, Freeman KD, Gunsberger T, Hausmann MG, Leblanc KA, Deljkich E, Trad KS. The effects of transoral incisionless fundoplication on chronic GERD patients: 12-month prospective multicenter experience. Surg Laparosc Endosc Percutan Tech 2014; 24: 36-46 [PMID: 24487156 DOI: 10.1097/SLE.0b013e3182a2b05c]
- 43 Testoni PA, Testoni S, Mazzoleni G, Vailati C, Passaretti S. Long-term efficacy of transoral incisionless fundoplication with Esophyx (Tif 2.0) and factors affecting outcomes in GERD patients followed for up to 6 years: a prospective single-center study. Surg Endosc 2015; 29: 2770-2780 [PMID: 25480624 DOI: 10.1007/s00464-014-4008-6]
- Stefanidis G, Viazis N, Kotsikoros N, Tsoukalas N, Lala E, Theocharis L, Fassaris A, 44 Manolakopoulos S. Long-term benefit of transoral incisionless fundoplication using the esophyx device for the management of gastroesophageal reflux disease responsive to medical therapy. Dis Esophagus 2017; 30: 1-8 [PMID: 27868281 DOI: 10.1111/dote.12525]
- Chimukangara M, Jalilvand AD, Melvin WS, Perry KA. Long-term reported outcomes of transoral 45 incisionless fundoplication: an 8-year cohort study. Surg Endosc 2019; 33: 1304-1309 [PMID: 30167944 DOI: 10.1007/s00464-018-6403-x]
- 46 Testoni PA, Testoni S, Mazzoleni G, Pantaleo G, Cilona MB, Distefano G, Fanti L, Antonelli M, Passaretti S. Transoral incisionless fundoplication with an ultrasonic surgical endostapler for the treatment of gastroesophageal reflux disease: 12-month outcomes. Endoscopy 2020; 52: 469-473 [PMID: 32187630 DOI: 10.1055/a-1124-3187]
- Rinsma NF, Farré R, Bouvy ND, Masclee AA, Conchillo JM. The effect of endoscopic 47 fundoplication and proton pump inhibitors on baseline impedance and heartburn severity in GERD patients. Neurogastroenterol Motil 2015; 27: 220-228 [PMID: 25348594 DOI: 10.1111/nmo.12468]
- 48 Witteman BP, Conchillo JM, Rinsma NF, Betzel B, Peeters A, Koek GH, Stassen LP, Bouvy ND. Randomized controlled trial of transoral incisionless fundoplication vs. proton pump inhibitors for treatment of gastroesophageal reflux disease. Am J Gastroenterol 2015; 110: 531-542 [PMID: 25823768 DOI: 10.1038/ajg.2015.28]
- Hunter JG, Kahrilas PJ, Bell RC, Wilson EB, Trad KS, Dolan JP, Perry KA, Oelschlager BK, Soper 49 NJ, Snyder BE, Burch MA, Melvin WS, Reavis KM, Turgeon DG, Hungness ES, Diggs BS. Efficacy of transoral fundoplication vs omeprazole for treatment of regurgitation in a randomized controlled trial. Gastroenterology 2015; 148: 324-333.e5 [PMID: 25448925 DOI: 10.1053/j.gastro.2014.10.009]
- 50 Håkansson B, Montgomery M, Cadiere GB, Rajan A, Bruley des Varannes S, Lerhun M, Coron E, Tack J, Bischops R, Thorell A, Arnelo U, Lundell L. Randomised clinical trial: transoral incisionless fundoplication vs. sham intervention to control chronic GERD. Aliment Pharmacol Ther 2015; 42: 1261-1270 [PMID: 26463242 DOI: 10.1111/apt.13427]
- Trad KS, Barnes WE, Prevou ER, Simoni G, Steffen JA, Shughoury AB, Raza M, Heise JA, Fox 51 MA, Mavrelis PG. The TEMPO Trial at 5 Years: Transoral Fundoplication (TIF 2.0) Is Safe, Durable, and Cost-effective. Surg Innov 2018; 25: 149-157 [PMID: 29405886 DOI: 10.1177/1553350618755214]
- Huang X, Chen S, Zhao H, Zeng X, Lian J, Tseng Y, Chen J. Efficacy of transoral incisionless 52 fundoplication (TIF) for the treatment of GERD: a systematic review with meta-analysis. Surg Endosc 2017; **31**: 1032-1044 [PMID: 27495332 DOI: 10.1007/s00464-016-5111-7]
- 53 Richter JE, Kumar A, Lipka S, Miladinovic B, Velanovich V. Efficacy of Laparoscopic Nissen Fundoplication vs Transoral Incisionless Fundoplication or Proton Pump Inhibitors in Patients With Gastroesophageal Reflux Disease: A Systematic Review and Network Meta-analysis. Gastroenterology 2018; 154: 1298-1308.e7 [PMID: 29305934 DOI: 10.1053/j.gastro.2017.12.021]
- Roy-Shapira A, Bapaye A, Date S, Pujari R, Dorwat S. Trans-oral anterior fundoplication: 5-year 54 follow-up of pilot study. Surg Endosc 2015; 29: 3717-3721 [PMID: 25783833 DOI: 10.1007/s00464-015-4142-9
- 55 Kim HJ, Kwon CI, Kessler WR, Selzer DJ, McNulty G, Bapaye A, Bonavina L, Lehman GA. Longterm follow-up results of endoscopic treatment of gastroesophageal reflux disease with the MUSE™ endoscopic stapling device. Surg Endosc 2016; 30: 3402-3408 [PMID: 26537905 DOI: 10.1007/s00464-015-4622-y]
- 56 Zacherl J, Roy-Shapira A, Bonavina L, Bapaye A, Kiesslich R, Schoppmann SF, Kessler WR, Selzer DJ, Broderick RC, Lehman GA, Horgan S. Endoscopic anterior fundoplication with the Medigus Ultrasonic Surgical Endostapler (MUSETM) for gastroesophageal reflux disease: 6-month results from a multi-center prospective trial. Surg Endosc 2015; 29: 220-229 [PMID: 25135443 DOI: 10.1007/s00464-014-3731-3]
- Danalioglu A, Cipe G, Toydemir T, Kocaman O, Ince AT, Muslumanoglu M, Senturk H. Endoscopic 57 stapling in comparison to laparoscopic fundoplication for the treatment of gastroesophageal reflux disease. Dig Endosc 2014; 26: 37-42 [PMID: 23560891 DOI: 10.1111/den.12081]
- 58 Sowa P, Samarasena JB. Nonablative Radiofrequency Treatment for Gastroesophageal Reflux Disease (STRETTA). Gastrointest Endosc Clin N Am 2020; 30: 253-265 [PMID: 32146945 DOI: 10.1016/j.giec.2019.12.006]
- Xie P, Yan J, Ye L, Wang C, Li Y, Chen Y, Li G. Efficacy of different endoscopic treatments in 59 patients with gastroesophageal reflux disease: a systematic review and network meta-analysis. Surg Endosc 2021; 35: 1500-1510 [PMID: 33650003 DOI: 10.1007/s00464-021-08386-1]
- 60 Coron E, Sebille V, Cadiot G, Zerbib F, Ducrotte P, Ducrott F, Pouderoux P, Arts J, Le Rhun M,



Piche T, Bruley des Varannes S, Galmiche JP; Consortium de Recherche Indépendant sur le Traitement et L'exploration du Reflux Gastro-oesophagien et de L'endobrachyoesophage (CRITERE). Clinical trial: Radiofrequency energy delivery in proton pump inhibitor-dependent gastro-oesophageal reflux disease patients. Aliment Pharmacol Ther 2008; 28: 1147-1158 [PMID: 18616516 DOI: 10.1111/j.1365-2036.2008.03790.x]

- Corley DA, Katz P, Wo JM, Stefan A, Patti M, Rothstein R, Edmundowicz S, Kline M, Mason R, 61 Wolfe MM. Improvement of gastroesophageal reflux symptoms after radiofrequency energy: a randomized, sham-controlled trial. Gastroenterology 2003; 125: 668-676 [PMID: 12949712 DOI: 10.1016/s0016-5085(03)01052-7
- Aziz AM, El-Khayat HR, Sadek A, Mattar SG, McNulty G, Kongkam P, Guda MF, Lehman GA. A 62 prospective randomized trial of sham, single-dose Stretta, and double-dose Stretta for the treatment of gastroesophageal reflux disease. Surg Endosc 2010; 24: 818-825 [PMID: 19730952 DOI: 10.1007/s00464-009-0671-4]
- Arts J, Bisschops R, Blondeau K, Farré R, Vos R, Holvoet L, Caenepeel P, Lerut A, Tack J. A 63 double-blind sham-controlled study of the effect of radiofrequency energy on symptoms and distensibility of the gastro-esophageal junction in GERD. Am J Gastroenterol 2012; 107: 222-230 [PMID: 22108449 DOI: 10.1038/ajg.2011.395]
- Lipka S, Kumar A, Richter JE. No evidence for efficacy of radiofrequency ablation for treatment of 64 gastroesophageal reflux disease: a systematic review and meta-analysis. Clin Gastroenterol Hepatol 2015; 13: 1058-67.e1 [PMID: 25459556 DOI: 10.1016/j.cgh.2014.10.013]
- 65 Fass R, Cahn F, Scotti DJ, Gregory DA. Systematic review and meta-analysis of controlled and prospective cohort efficacy studies of endoscopic radiofrequency for treatment of gastroesophageal reflux disease. Surg Endosc 2017; 31: 4865-4882 [PMID: 28233093 DOI: 10.1007/s00464-017-5431-2]
- 66 Vaezi MF, Shaheen NJ, Muthusamy VR. State of Evidence in Minimally Invasive Management of Gastroesophageal Reflux: Findings of a Scoping Review. Gastroenterology 2020; 159: 1504-1525 [PMID: 32621903 DOI: 10.1053/j.gastro.2020.05.097]
- Weitzendorfer M, Spaun GO, Antoniou SA, Tschoner A, Schredl P, Emmanuel K, Koch OO. 67 Interim Report of a Prospective Trial on the Clinical Efficiency of a New Full-thickness Endoscopic Plication Device for Patients With GERD: Impact of Changed Suture Material. Surg Laparosc Endosc Percutan Tech 2017; 27: 163-169 [PMID: 28383316 DOI: 10.1097/SLE.00000000000396]
- Weitzendorfer M, Spaun GO, Antoniou SA, Witzel K, Emmanuel K, Koch OO. Clinical feasibility of a new full-thickness endoscopic plication device (GERDxTM) for patients with GERD: results of a prospective trial. Surg Endosc 2018; 32: 2541-2549 [PMID: 29602998 DOI: 10.1007/s00464-018-6153-9
- Inoue H, Ito H, Ikeda H, Sato C, Sato H, Phalanusitthepha C, Hayee B, Eleftheriadis N, Kudo SE. 69 Anti-reflux mucosectomy for gastroesophageal reflux disease in the absence of hiatus hernia: a pilot study. Ann Gastroenterol 2014; 27: 346-351 [PMID: 25330784]
- 70 Monino L, Gonzalez JM, Vitton V, Barthet M. Antireflux mucosectomy band in treatment of refractory gastroesophageal reflux disease: a pilot study for safety, feasibility and symptom control. Endosc Int Open 2020; 8: E147-E154 [PMID: 32010747 DOI: 10.1055/a-1038-4012]
- Debourdeau A, Vitton V, Monino L, Barthet M, Gonzalez JM. Antireflux Mucosectomy Band 71 (ARM-b) in Treatment of Refractory Gastroesophageal Reflux Disease After Bariatric Surgery. Obes Surg 2020; 30: 4654-4658 [PMID: 32676843 DOI: 10.1007/s11695-020-04753-4]
- Sumi K, Inoue H, Kobayashi Y, Iwaya Y, Abad MRA, Fujiyoshi Y, Shimamura Y, Ikeda H, Onimaru 72 M. Endoscopic treatment of proton pump inhibitor-refractory gastroesophageal reflux disease with anti-reflux mucosectomy: Experience of 109 cases. Dig Endosc 2021; 33: 347-354 [PMID: 32415898 DOI: 10.1111/den.13727]
- 73 Li X, Zhang W, Chen M, Wei S, Zhao X, Zhang G. A Prospective Randomized Trial to Assess the Antireflux Effect of Antireflux Mucosectomy in the Porcine Model. Gastroenterol Res Pract 2019; 2019: 3286738 [PMID: 30944560 DOI: 10.1155/2019/3286738]
- Yoo IK, Ko WJ, Kim HS, Kim HK, Kim JH, Kim WH, Hong SP, Yeniova AÖ, Cho JY. Anti-reflux 74 mucosectomy using a cap-assisted endoscopic mucosal resection method for refractory gastroesophageal disease: a prospective feasibility study. Surg Endosc 2020; 34: 1124-1131 [PMID: 31139995 DOI: 10.1007/s00464-019-06859-y]
- Inoue H, Tanabe M, de Santiago ER, Abad MRA, Shimamura Y, Fujiyoshi Y, Ueno A, Sumi K, 75 Tomida H, Iwaya Y, Ikeda H, Onimaru M. Anti-reflux mucosal ablation (ARMA) as a new treatment for gastroesophageal reflux refractory to proton pump inhibitors: a pilot study. Endosc Int Open 2020; 8: E133-E138 [PMID: 32010745 DOI: 10.1055/a-1031-9436]
- 76 Hernández Mondragón OV, Zamarripa Mottú RA, García Contreras LF, Gutiérrez Aguilar RA, Solórzano Pineda OM, Blanco Velasco G, Murcio Perez E. Clinical feasibility of a new antireflux ablation therapy on gastroesophageal reflux disease (with video). Gastrointest Endosc 2020; 92: 1190-1201 [PMID: 32343977 DOI: 10.1016/j.gie.2020.04.046]
- 77 Wong HJ, Su B, Attaar M, Kuchta K, Stearns S, Linn JG, Haggerty SP, Denham W, Ujiki MB. Antireflux mucosectomy (ARMS) results in improved recovery and similar reflux quality of life outcomes compared to laparoscopic Nissen fundoplication. Surg Endosc 2020 [PMID: 33237465 DOI: 10.1007/s00464-020-08144-9
- Patil G, Dalal A, Maydeo A. Feasibility and outcomes of anti-reflux mucosectomy for proton pump 78 inhibitor dependent gastroesophageal reflux disease: First Indian study (with video). Dig Endosc



2020; 32: 745-752 [PMID: 31834663 DOI: 10.1111/den.13606]

- 79 Bapaye A, Mahadik M, Pujari R, Bharadwaj T, Vare S, Date S, Dubale N, Bapaye J, Kulkarni A. Anti-reflux mucosectomy (ARMS) for refractory GERD-Initial clinical experience. J Gastroenterol Hepatol 2017; 32: 255
- 80 Vasilevskiy DI, Bagnenko SF, Smirnov A, Lapshin AS, Dvoretskiy SU, Pryadko AS. Antireflux mucosectomy (Arms) in the treatment of patients with gerd and columnar-cell lined (barrett's) esophagus. First experiences. *Surg Endosc* 2017; 31: S405 [PMID: 28488176 DOI: 10.1007/s00464-017-5565-2]
- 81 Shah R, Maydeo AP, Dhir V. Anti reflux mucosectomy (ARMS) for refractory gastro esophageal reflux disease (GERD)-are we there yet? *United European Gastroenterol J* 2017; 5: A354-A355 [DOI: 10.1177/2050640617725676]
- 82 Ota K, Takeuchi T, Harada S, Edogawa S, Kojima Y, Inoue T, Higuchi K. A novel endoscopic submucosal dissection technique for proton pump inhibitor-refractory gastroesophageal reflux disease. *Scand J Gastroenterol* 2014; 49: 1409-1413 [PMID: 25384555 DOI: 10.3109/00365521.2014.978815]
- 83 Ortega A, Rosón P, Fern, ez F, Angeles Romero M, Angeles Perez Aisa M, Cotta J, Lozano M. Antireflux mucosectomy. preliminary results of a prospective study. *Endoscopy* 2019; 51: S240-S241
- 84 Mondragón OVH, Pintor JC, Aguilar RAG, Garcia-Contreras L, Pineda OS, Mottú RAZ, Blanco-Velasco G, Murcio-Pérez E. Sa1247 Antireflux ablation therapy (ARAT), for reflux disease after poem procedure. Early clinical experience. *Gastrointest Endosc* 2020; **91**: AB130
- 85 Schuitenmaker JM, van Hoeij FB, Schijven MP, Tack J, Conchillo JM, Hazebroek EJ, Smout AJPM, Bredenoord AJ. Pneumatic dilation for persistent dysphagia after antireflux surgery, a multicentre single-blind randomised sham-controlled clinical trial. *Gut* 2021 [PMID: 33452179 DOI: 10.1136/gutjnl-2020-322355]
- 86 Seleem WM, Hanafy AS, Mohamed SI. Endoscopic management of refractory gastroesophageal reflux disease. *Scand J Gastroenterol* 2018; 53: 390-397 [PMID: 29488430 DOI: 10.1080/00365521.2018.1445775]
- 87 Hu HQ, Li HK, Xiong Y, Zhang XB, Zhi JL, Wang XX, Ling-Hu EQ. Peroral endoscopic cardial constriction in gastroesophageal reflux disease. *Medicine (Baltimore)* 2018; 97: e0169 [PMID: 29642142 DOI: 10.1097/MD.00000000010169]
- 88 Liu S, Chai N, Zhai Y, Zou J, Feng X, Li Z, Li L, Zhang X, Wang X, Wang S, Linghu EQ. New treatment method for refractory gastroesophageal reflux disease (GERD): C-BLART (clip band ligation anti-reflux therapy)-a short-term study. *Surg Endosc* 2020; **34**: 4516-4524 [PMID: 31728750 DOI: 10.1007/s00464-019-07238-3]
- 89 Benias PC, D'Souza L, Lan G, Gluckman C, Inamdar S, Trindade AJ, Miller LS, Carr-Locke DL. Initial experience with a novel resection and plication (RAP) method for acid reflux: a pilot study. *Endosc Int Open* 2018; 6: E443-E449 [PMID: 29607397 DOI: 10.1055/s-0044-101453]
- 90 Janu P, Shughoury AB, Venkat K, Hurwich D, Galouzis T, Siatras J, Streeter D, Korman K, Mavrelis G, Mavrelis P. Laparoscopic Hiatal Hernia Repair Followed by Transoral Incisionless Fundoplication With EsophyX Device (HH + TIF): Efficacy and Safety in Two Community Hospitals. *Surg Innov* 2019; 26: 675-686 [PMID: 31431138 DOI: 10.1177/1553350619869449]
- 91 Fundacion para la Investigacion Biomedica del Hospital Universitario Ramon y Cajal. Doubleblind, Placebo-controlled Clinical Trial on the Efficacy of Antireflux Ablation of the Cardiac Mucosa for the Treatment of Gastroesophageal Reflux Disease. Available from: https://clinicaltrials.gov/ct2/show/NCT04711655
- 92 Gyawali CP, Kahrilas PJ, Savarino E, Zerbib F, Mion F, Smout AJPM, Vaezi M, Sifrim D, Fox MR, Vela MF, Tutuian R, Tack J, Bredenoord AJ, Pandolfino J, Roman S. Modern diagnosis of GERD: the Lyon Consensus. *Gut* 2018; 67: 1351-1362 [PMID: 29437910 DOI: 10.1136/gutjnl-2017-314722]
- 93 Yadlapati R, Kahrilas PJ, Fox MR, Bredenoord AJ, Prakash Gyawali C, Roman S, Babaei A, Mittal RK, Rommel N, Savarino E, Sifrim D, Smout A, Vaezi MF, Zerbib F, Akiyama J, Bhatia S, Bor S, Carlson DA, Chen JW, Cisternas D, Cock C, Coss-Adame E, de Bortoli N, Defilippi C, Fass R, Ghoshal UC, Gonlachanvit S, Hani A, Hebbard GS, Wook Jung K, Katz P, Katzka DA, Khan A, Kohn GP, Lazarescu A, Lengliner J, Mittal SK, Omari T, Park MI, Penagini R, Pohl D, Richter JE, Serra J, Sweis R, Tack J, Tatum RP, Tutuian R, Vela MF, Wong RK, Wu JC, Xiao Y, Pandolfino JE. Esophageal motility disorders on high-resolution manometry: Chicago classification version 4.0[®]. *Neurogastroenterol Motil* 2021; 33: e14058 [PMID: 33373111 DOI: 10.1111/nmo.14058]
- Funk LM, Zhang JY, Drosdeck JM, Melvin WS, Walker JP, Perry KA. Long-term cost-effectiveness of medical, endoscopic and surgical management of gastroesophageal reflux disease. *Surgery* 2015; 157: 126-136 [PMID: 25262216 DOI: 10.1016/j.surg.2014.05.027]
- 95 ASGE Standards of Practice Committee, Muthusamy VR, Lightdale JR, Acosta RD, Chandrasekhara V, Chathadi KV, Eloubeidi MA, Fanelli RD, Fonkalsrud L, Faulx AL, Khashab MA, Saltzman JR, Shaukat A, Wang A, Cash B, DeWitt JM. The role of endoscopy in the management of GERD. *Gastrointest Endosc* 2015; 81: 1305-1310 [PMID: 25863867 DOI: 10.1016/j.gie.2015.02.021]

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Basic Study

ORIGINAL ARTICLE

Cold exposure and capsaicin promote 1,2-dimethylhyrazine-induced colon carcinogenesis in rats correlates with extracellular matrix remodeling

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Abstract

BACKGROUND

Extracellular matrix (ECM) remodeling and stiffening, which are correlated with tumor malignancy, drives tumor development. However, the relationship between ECM remodeling and rat experimental model of 1,2-dimethylhyrazine (DMH)-induced colorectal cancer (CRC) imposed by cold and capsaicin exposure remains unclear.

AIM

To explore the effects of cold exposure and capsaicin on ECM remodeling and ECM enzymes in DMH-induced CRC.

METHODS

For histopathological analysis, the sections of colon tissues were stained with hematoxylin and eosin, Masson's trichrome, Picrosirius red, and Weigert's Resorcin-Fuchsin to observe the remodeling of collagen and elastin. Additionally, the protein expression level of type I collagen (COL I), type 3 collagen (COL III0, elastin, matrix metalloproteinase (MMP) 1, MMP2, MMP9, and tissue-specific



nature or kind in any product, service and/or company that could be construed as influencing the position presented in, or the review of, the manuscript entitled "Cold exposure and Capsaicin promote 1,2-dimethylhydrazineinduced colon carcinogenesis in rats correlates with extracellular matrix remodeling".

Data sharing statement: Technical appendix, statistical code, and dataset available from the

corresponding author at

wenbin@gzucm.edu.cn. Participants gave informed consent

for data sharing.

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matrix metalloproteinase 1 (TIMP1) was assessed by immunohistochemistry. The messenger RNA (mRNA) levels of COL I, COL III, elastin, and lysyl oxidase-like-2 (LOXL2) in the colon tissues of rats was measured by reverse-transcriptase quantitative polymerase chain reaction.

RESULTS

Although no differences were observed in the proportion of adenomas, a trend towards the increase of invasive tumors was observed in the cold and capsaicin group. The cold exposure group had a metastasis rate compared with the other groups. Additionally, abnormal accumulation of both collagen and elastin was observed in the cold exposure and capsaicin group. Specifically, collagen quantitative analysis showed increased length, width, angle, and straightness compared with the DMH group. Collagen deposition and straightness were significantly increased in the cold exposure group compared with the capsaicin group. Cold exposure and capsaicin significantly increased the protein levels of COL I, elastin, and LOXL2 along with increases in their mRNA levels in the colon tissues compared with the DMH group, while COL III did not show a significant difference. Furthermore, in immunohistochemical evaluations, MMP1, MMP2, MMP9, and TIMP1 staining increased in the cold exposure and capsaicin group compared with the DMH group.

CONCLUSION

These results suggest that chronic cold and capsaicin exposure further increased the deposition of collagen and elastin in the colonic tissue. Increased COL I and elastin mRNA and protein levels expression may account for the enhanced ECM remodel and stiffness variations of colon tissue. The upregulated expression of the LOXL2 and physiological imbalance between MMP/TIMP activation and deactivation could contribute to the progression of the CRC resulting from cold and capsaicin exposure.

Key Words: Colon cancer; Cold exposure; Capsaicin; Extracellular matrix remodeling; Extracellular matrix enzymes

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Core Tip: In this study, we discovered that remodeling of extracellular matrix (ECM) plays an important role in the progression of colorectal cancer (CRC). These results suggest that increased stiffness of colonic tissue and the remodeling of ECM mediated by ECM enzymes resulting from cold and capsaicin exposure predisposes an environment suitable for CRC development and progression. To target ECM in CRC tumor tissue could represent a novel potential therapeutic strategy.

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INTRODUCTION

Colorectal cancer (CRC), a common cause of cancer deaths in the world, is a multifactorial disease driven by genetic predisposition, epigenetic alterations, and environmental factors[1]. Only a minority of CRC is caused by the accumulation of genetic epigenetic alterations, while the majority is linked to environmental factors such as dietary intake, alcohol consumption, and ambient environment[2,3]. Increasing epidemiological data have indicated that cold weather might be associated with an increased occurrence of cancer[4]. Additionally, the consumption of chili-pepper and cancer incidence have a positive correlation [5,6]. However, the mechanisms under-



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lying the effects of cold exposure and capsaicin in 1,2-dimethylhyrazine (DMH)induced CRC tumorigenesis and progression remain poorly understood. Extracellular matrix (ECM) is a non-cellular structure that is essential for the maintenance of normal tissue and organ function and disease pathophysiology[7]. Collagen is a major component of the ECM. It provides cellular components with physical support and is an important contributor to tumor growth and progression[8]. Tumor progression is accompanied by the dysregulation of collagen structure and deposition. Tumor associated-collagen is usually compacted to thick collagen bundles and the anisotropic arrangement of relatively straight in the matrix of malignancies compared with healthy tissues[9,10]. In clinical samples of breast tumors, collagen deposition increased, and linearization and thickening of collagen occurred; these processes can be linked to poor prognosis and high risk of mortality[11,12].

Elastin is another important fibrous ECM protein that provides elastic recoil to tissue. Importantly, excessive accumulation of ECM, particularly collagen and elastin, gradually leads to progressive organ fibrosis[13]. The fibrosis results in tissue stiffness and can predispose tissue to malignancy. Several human studies indicated that patients with liver fibrosis and stiffness are positively correlated with the risk hepatocellular carcinoma[14]. ECM remodeling is mainly orchestrated by ECM modifying enzymes such as lysyl oxidase-like-2 (LOXL2), matrix metalloproteinases (MMP), and tissue-specific matrix metalloproteinase inhibitors (TIMPs)[15]. LOXL2 is a key factor in ECM remodeling and is a copper-dependent amine oxidase that catalyzes the crosslinking of collagen or elastin in the ECM and thus regulates the tensile strength of tissues. LOXL2 causes disorganization and composition of ECM, resulting in many pathological conditions, including fibrosis and cancer[16]. Active LOXL2 is involved in stiffness-associated cancer progression, whereas inhibition of LOXL2 result in less collagen cross-linking and impeded cancer progression[17]. Moreover, LOXL2 expression is overexpressed in many types of tumors and is associated with poor prognosis[18-20].

MMP has been implicated in cancer development, progression, invasiveness, and dissemination by promoting a protumorigenic microenvironment and modulating the ECM and intercellular junctions^[21]. MMPs and TIMPs are the main enzymes involved in the regulation of ECM remodeling and collagen degradation process[22]. Their expression and activity are upregulated in almost all human cancers with disparate changes, and this phenomenon is associated with advanced tumor stage, poor prognosis, and decreased overall survival rate[23,24]. Increased MMP expression/ activity or decreased TIMPs could lead to MMP/TIMP imbalance, resulting in various pathological conditions including fibrosis and cancers[25].

Limited information, however, is available about ECM remolding and ECM enzyme activity in the progression of experimental colorectal malignancy. We have previously shown that cold exposure and long-term administration of capsaicin at a low dose further promote the development and progression of CRC[26]. However, the specific mechanisms underlying cold and capsaicin exposure tumor promotion remained unknown. This study aimed to investigate the effects of cold exposure and capsaicin on ECM remodeling and ECM enzymes in DMH-induced CRC. Moreover, we determined whether excessive ECM deposition, particular whether collagen and elastin and dysregulation of ECM enzymes expression and/or secretion in rat treatment with cold exposure, could further stiffen the colon tissues and disrupt the intestinal morphogenesis to exacerbate the experimental colorectal malignancy.

MATERIALS AND METHODS

Experimental design in adult male rats

Wistar rats weighing 200-250 g (6-wk-old) were obtained from Experimental Animal Center in Guangzhou University of Chinese Medicine. Animals were housed in plastic cages under a controlled environment (24 ± 2 °C, 50% ± 5% humidity, 12 h/12 h lightdark cycle) with ad libitum food and water access. All the experimental protocols were approved by the Institutional Animal Ethics Committee of the Guangzhou University of Chinese Medicine (No. 20130001). Briefly, after 3 d of acclimation, the animals were randomly assigned into four groups (n = 10). Rats in group A received no treatment and served as control. Five weeks later, rats in groups B-D received subcutaneous injection of DMH (25 mg/kg) once a week for 12 wk. In addition to DMH, Group C rats received cold distilled water (10 mg/kg) until the end of 38 wk. Group D rats were given capsaicin (0.9 mg/mL) every day throughout the experiment. By the end of the week, 10 rats from each group were sacrificed. For macroscopic evaluation of the



incidence of polyps at the end of the experimental period, rats were sacrificed and colons were incised and washed with physiological saline. Then cleaned colons were cut opened longitudinally and the total number of polyps/tumors was carefully counted and later verified with histopathological examination. The counting and histopathological analysis of gross macroscopic neoplastic lesions was carried out by two investigators from this study. If the histopathological types of these two investigators were different, then tumor histology was classified as adenomas and adenocarcinomas by one pathologist under blinded conditions from the Pathology Department in Guangzhou University of Chinese Medicine. Microscope findings were classified as adenomas and adenocarcinomas according to previous criteria described by Jikihara et *al*[27]. Tumor incidence is the percentage of rat bearing the indicated type of tumor.

Histopathological staining

All specimens were fixed in 4% paraformaldehyde solution for 24 h and embedded in paraffin and processed by standard histological processing techniques. Serial issue sections (8-µm thick) were obtained from each sample with the microtome and then were stained with hematoxylin and eosin (HE), picrosirius red, Masson's trichrome (MT), and Weigert's Resorcin-Fuschin (WRF). For Picrosirius red staining, sections were stained in picrosirius red solution 0.1) (Sirius red F3B; Sigma-Aldrich Co., St Louis, MO, United States) in a saturated aqueous solution of picric acid for 1 h at room temperature for collagen bundle staining. Images were subsequently analyzed using ImageJ to calculate the fiber density, which was measured as image % area coverage. MT was performed according to the manufacture's protocol including Weigert's Iron Hematoxylin Solution, Ponceau acid fuchsin, and Aniline Blue as reagents. The collagen volume fraction was measured by ImageJ software and calculated as the proportion of blue positive areas in the total section areas. The process of WRF used reagents and kits from Solarbio (Beijing, China). Sections were then mounted for observation under polarized light microscopy (NIKON Eclipse Ci, Tokyo, Japan) and light microscopy, respectively. Three microphotographs of the reticular dermis were taken with a 400 × magnification with light microscopy and polarized microscopy, respectively. Digitized images of histological sections obtained under final magnification of × 400 were analyzed using the Image-Pro Plus 4.5 software.

Collagen fiber analysis

CT-FIRE, an open-resource software (http://Loci.wisc.edu/software/cifire), was used as previously described to quantify automatically collagen fibers^[28]. The quantitative parameters included alignment of collagen fibers as well as individual length, straightness, and width. These features of collagen fibers are widely used to investigate collagen organization in a various of cancer^[29]. All the picrosirius red images were converted to 8-bit images and threshold values between 10-255 to eliminate background noise using FIJI ImageJ[30]. These images were uploaded to CT-FIRE; collagen fiber extraction parameters were set to default parameters.

Real-time quantitative reverse-transcriptase polymerase chain reaction

Total RNA extracts of colon tissues were prepared using TRIZOL reagent (TaKaRa, Kusatsu, Japan). RNA (1 μ g) was reverse transcribed in a 20 μ L reaction mixture using Prime Script RT Master Mix (TaKaRa). The purity of total RNA was evaluated by measuring the concentration and OD_{260:280} values with a Nanodrop 2000 spectrophotometer (Thermo Fisher Scientific, Waltham, MA, United States). The mRNA levels of collagen type 1, alpha1 (COL1A1), collagen type 3, alpha1 (COL3A1), LOXL2, and elastin in colon mucosa were assessed using a Step One Plus real-time polymerase chain reaction system (CFX384TM Real-time System; Thermo Fisher Scientific). The relative levels of gene expression were enumerated using the comparative formula 2- $\Delta\Delta$ Ct. The primer sequences used in this polymerase chain reaction amplification were as follows: 5'-AGCCATGTACGTAGCCATCC-3'/3'-ACCCTCATAGATGGGCACAG-5' for β-actin; 5'-AGGCATAAAGGGTCATCGTGGCTT-3'/3'-AGTCCATCTTTGC-CAGGAGAACCA-5' for COL1a1; 5'-GGTTTGGAGAATCTATGAATGGTGG-3'/3'-GCTGGAAAGAAGTCTGAGGAAGG-5' for Col3a1; 5'-AGCCTATAAGCCG-GAGCAAC-3'/3'-GTCCCACTTGTCATCGCAGA-5 'for LOXL2; 5'-CGCCTGTAAT-GCCTCCAATC-3'/3'-AGCAGCTAAAGCAGCGAAGT-5' for elastin.

Immunohistochemistry

Colonic tissue sections were deparaffinized and rehydrated through a series of xylene and ethanol/water. The sections were placed in a 95 °C antigen retrieval solution (citrate buffer; PH 6.0) for 15 min. After cooling in retrieval solutions for 20 min at



Table 1 Incidence of various tumors induced in different treatment groups

Group	Total number _ of tumors	Adenc	oma inciden	ce, %	Adenocarcinon		With lymphytic metastasis (%)		
		Mild	Moderate	Severe	Well- differentiated	Moderate- differentiated	Poor- differentiated	Mucinous	Metastasis rate
Control	-	-	-	-	-	-	-	-	-
DMH	23	2/23 (8.7)	2/23 (8.7)	3/23 (13)	12/23 (52.2)	4/23 (17.4)	3/23 (13.0)	-	-
Cold exposure	38	1/38 (2.6)	2/38 (5.3)	3/38 (7.8)	5/38 (15.8) ^b	8/38 (21.1)	15/38 (36.8) ^a	4/38 (10.5)	20.0
Capsaicin	34	2/31 (5.9)	1/34 (2.9)	4/31 (12.9)	7/31(22.6) ^a	12/31 (38.7)	7/31(22.6)	1/31 (3.2)	-

Values are expressed as the proportion of lesions-bearing rats. n = 10 rats/group. Incidence data was analyzed by using chi-square or Fisher's exact test. $^{a}P < 0.05$.

 $^{b}P < 0.01.$

1,2-dimethylhyrazine compared with cold exposure and capsaicin-treated group. DMH: 1,2-Dimethylhyrazine.

room temperature, the slides were treated with hydrogen peroxide for 10 min to block endogenous peroxidase activity. Primary rabbit anti-histone polyclonal antibodies were applied for 14 h at 4 °C overnight at the following dilutions: Type I collagen (COL I) (1:500; ab34710; Abcam, Cambridge, United Kingdom), type III collagen (COL III) (1:200, ab7778; Abcam), LOXL2 (1:400), elastin (1:600; ab217356; Abcam), MMP1 (1:500; 10371-2-AP; Proteintech, Rosemont, IL, United States), MMP2 (1:200; ab86607; Abcam), MMP9 (1:800; ab38898; Abcam), and TIMP1 (1:600, ab61224; Abcam). The next day, biotin-conjugated secondary antibody and streptavidin-biotin peroxidase were applied each for 20 min. 3,3'-Diaminobenzidine tetrahydrochloride (0.05%) was used as the substrate, and nuclear contrast was performed using hematoxylin counterstaining. Each section was analyzed in three different fields using Image Pro Plus software. The density of yellow reflects the expression levels of target proteins. Integral optical density sum / area SUM was applied to quantify the relative expression of COL I, COL III, LOXL2, elastin, MMP1, MMP2, MMP9, and TIMP1.

Statistical analysis

All the data are summarized as mean \pm SD, and data were analyzed using SPSS 23.0 statistical software. We performed the data with a one-way analysis of variance with the post-hoc comparison by the L.S.D. method and Fisher's exact test was employed to compare tumor incidence. Differences with values of P < 0.05 were considered statistically significant.

RESULTS

Macroscopic and pathological observation study of colon tumor

No visible colon tumor was found in the normal control. We observed findings such as colonic mucosal thickening, stiffness, and not tiled completely on filter paper in the majority of rats of the cold exposure group (Figure 1C). As shown in Figure 1B, the length of colon in the DMH and normal group was not significantly different. However, the length of the colon in the cold exposure and capsaicin group was shorter than that of DMH group. The pathological classification of colonic tumors in each group is shown in Table 1. No difference was observed in the proportion of adenomas among groups. In the DMH-induced group of animals, most tumors had well-differentiated tubular adenocarcinomas. Invasive tumors increased in the cold and capsaicin group. Histopathological analysis showed in the cold exposure group that an evident malignant transformation occurred in the colon with the features of poor-differentiated mucinous adenocarcinomas, and some of the glands were filled with mucinous material (Figure 2C). In addition, the mesenteric lymph nodes of rats in each group were stained with hematoxylin and eosin, and the lymphatic metastasis was observed under light microscope (Figure 2B). No lymphatic metastasis was observed in the DMH and capsaicin group, while a mesocolic lymph node was totally replaced by

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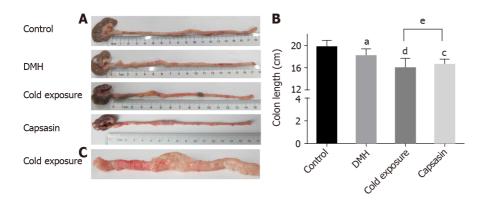


Figure 1 Colonic morphology of rats in different groups. A and B: Changes in colon length and colonic morphology; C: Stiff colonic tissues in cold exposure group. ${}^{a}P < 0.05$, control compared with 1,2-dimethylhyrazine (DMH); ${}^{c}P < 0.05$, ${}^{d}P < 0.01$, DMH compared with cold exposure and capsaicin-treated group; ${}^{e}P < 0.05$, cold exposure compared with capsaicin-treated group.

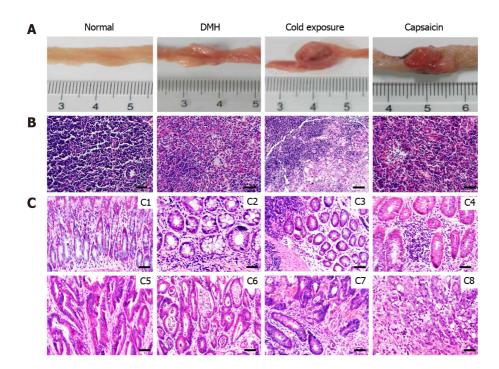


Figure 2 Pathological observation and lymph node metastases of different groups. A: Macroscopic image of the colonic tumors; B: Representative sections stained with hematoxylin and eosin (HE) showing the histopathology of the mesocolic lymph node in the different groups; C: Representative sections stained with HE showing the histopathology of the colonic mucosa in the different groups. Normal architecture of colon was observed in the control groups (C1), adenoma with mild dysplasia with massive infiltration of inflammatory cells (C2). Histology of adenoma with moderate dysplasia in cold exposure groups (C3). Histology of adenoma with severe dysplasia (C4). Histology of well-differentiated tubular adenocarcinomas (C5). Histology of Moderately differentiated adenocarcinomas (C6). Histology of Poorly differentiated adenocarcinomas (C7). Histology of Mucinous adenocarcinoma with signet ring cells (C8) (HE staining, × 400, scalar bar 20 µm). DMH: 1,2-Dimethylhyrazine.

metastatic cancer tissue in the cold exposure group; the lymph node metastasis rate was 20.0% (Table 1, Figures 1 and 2).

Alterations in collagens after cold exposure and capsaicin

Colon tissue sections were stained by MT and picrosirius red to identify the total collagen in the colon mucosa. As shown in Figure 3A, there were few collagen fibers in normal colonic mucosa. After DMH treatment, wave shape collagens stained blue were markedly increased around the glands, and this was increased further in the cold exposure and capsaicin treatment group. The collagen density was quantified using ImageJ, and it was significantly increased in the colonic tissue of cold exposure and capsaicin group. This excessive collagen deposition was further confirmed by picrosirius red staining. As shown in Figure 3B, picrosirius red staining revealed in normal tissue the collagen fibers with sparse deposition composed of thin collagen fibers. The collagen fibers in the DMH group were denser than that in normal collagen



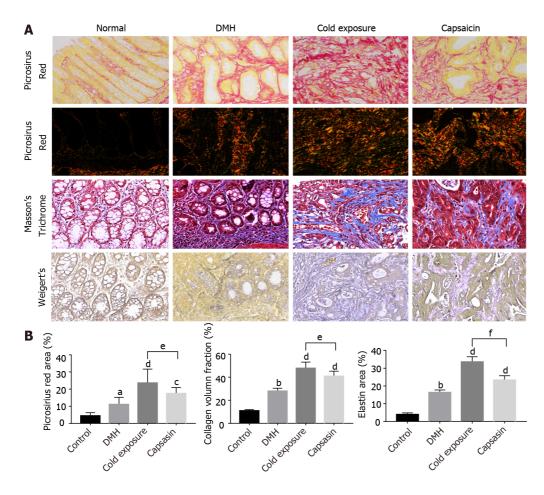


Figure 3 Changes in extracellular matrix components (collagen fibers and elastin) in colonic mucosa of different treatment groups. A: Representative photographs of colonic tissues in rats of normal, 1,2-dimethylhyrazine (DMH), cold exposure and capsaicin groups using Masson's trichrome: collagen (blue), nuclei and cytoplasm (red); picrosirius red in bright-field: collagen (red); polarized light: collagen (yellow-orange to green birefringence) and Weigert's Resorcin-Fuschin: elastin (blue-black), myofibers (yellow). Magnification, × 400, scalar bar 20 µm; B: Quantitative analysis of picrosirius red staining, trichrome and Weigert's staining as a measure of collagen and elastin density.

fibers. In the capsaicin treatment group, collagen fibers showed an evident increase and were crosslinked into bundles. On the other hand, the cold exposure group apparently displayed an increased amount of collagen fibers with heterogeneous thickness and alignment. The collagen in the cold exposure and capsaicin group exhibited a predominant reddish or yellow-orange. The structure and organization of collagen fibers were evaluated in colon tissue sections by quantifying the polarization microscopy images. As shown in Figure 4A, visualized collagen fibers were extracted and analyzed for fiber width, angle, length, and straightness using CT-FIRE software. As shown in Figure 4B, compared with the DMH group, collagen fibers in the cold exposure and capsaicin group showed a significant increase in angle, length, width, and straightness. These results revealed that cold exposure and capsaicin induced a progressive increase in the content and orientation of collagen fibers in CRC as a function of malignancy.

Alterations in elastin after cold exposure and capsaicin

Treatment WRF was used to identify the elastin fibers, which were stained black. As shown in Figure 3A, elastin was hardly expressed in the colonic mucosa of the normal rats. After treatment with DMH, the elastin fibers aligned surrounding the epithelium and stroma. After cold exposure and capsaicin treatment, the amount of elastin fibers increased, and thick elastic fibers was found highly disorganized between the gland compared with their respective control and DMH groups. Alterations in the mRNA levels of COL I, COL III, LOXL2, and elastin. The expression levels of COL I, COL III, LOXL2, and elastin mRNAs in colonic tissue are shown in Figure 5. The COL I, LOXL2, and elastin mRNA levels were higher in the DMH-induced cancer group than in the control group. In comparison with the DMH group, a significant increase was detected both in the cold exposure and capsaicin treatment group, but the mRNA levels of COL III were not significantly different between DMH and capsaicin ex-



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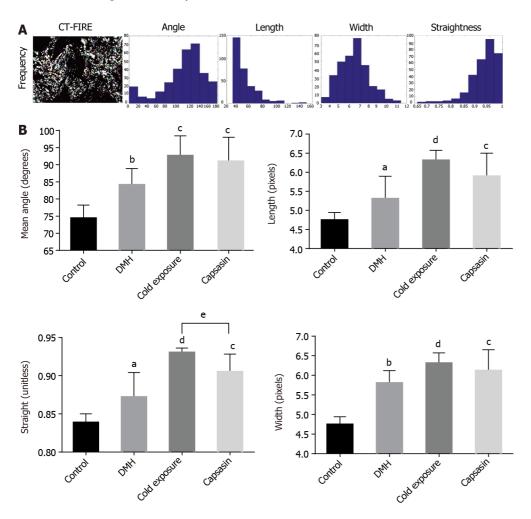


Figure 4 Collagen fibers were automatically extracted for analysis using open-source software CT-FIRE. A: Histograms were generated to show the distribution of various parameters in each polarized light microscopy imaging; B: Quantitative analysis of collagen fibers from polarized light microscopy imaging in the colonic mucosa of different treatment groups. Data are mean \pm SE of three images per tissues region. ^a*P* < 0.05, ^b*P* < 0.01, control compared with 1,2-dimethylhyrazine (DMH); ^c*P* < 0.05, ^d*P* < 0.01, DMH compared with cold exposure and capsaicin-treated group; ^e*P* < 0.05, Cold exposure compared with capsaicin-treated group.

posure group.

Alterations in the protein levels of COL I, COL III, LOXL2, and elastin

The expression levels of COL I, COL III, LOXL2, and elastin in colonic tissue are shown in Figure 6A and B. The protein expression levels of collagen type I, III, LOXL2, and elastin were significantly elevated in colonic tissue from DMH-treated rats in comparison to the control group. The expression level of proteins in the cold exposure and capsaicin treatment group increased. COL I and LOXL2 levels were significantly higher in the cold exposure group, but no statistical difference was observed in the change of COL III and elastin between cold exposure and capsaicin group.

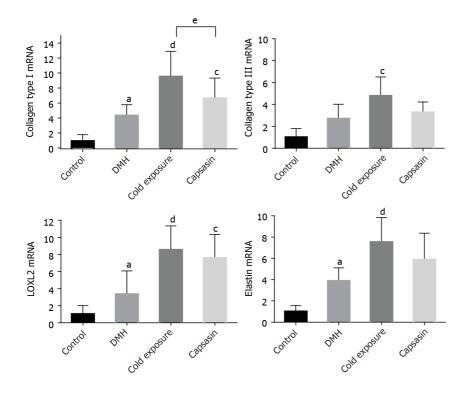
Alterations in the protein levels of MMP1, MMP2, MMP9, and TIMP1

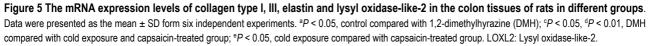
The expression levels of MMP1, MMP2, MMP9, and TIMP1 in colonic tissue are shown in Figure 7. Significantly elevated MMP1, MMP2, MMP9, and TIMP1 immunoreactivity was observed in DMH-treated rats compared with the control group. The expression levels of proteins in the cold and capsaicin group increased compared with the DMH group. In comparison with the capsaicin group, the expression of MMP2, MMP9, and TIMP1 increased in the cold exposure group.

DISCUSSION

ECM has been increasingly considered as an important regulator at diverse aspects of tumor initiation, promotion, neoplastic transformation, invasion, and metastasis[31].







Furthermore, ECM remodeling is a consequence of or increases risk for malignant transformation of colonic, hepatic, pulmonary, and pancreatic cells[32,33]. Collagen and elastin are the major components of ECM, and their excessive deposition has been implicated in a number of diseases, particularly fibrosis and cancer. However, the morphology and structure of collagen and elastin fibers in the animal models of CRC remains unclear. In the present study, we analyzed the morphology and structure of collagen and elastin fibers in rat experimental model of DMH-induced CRC imposed by cold and capsaicin exposure. Results showed an association between collagen expression or ECM modifying enzymes and CRC development, thus supporting ECM remodeling is highly relevant to CRC cancer progression. Tumor tissue often exhibits fibrosis, and this fibrotic state is characterized by the excessive deposition of collagen and elastin[34].

Fibrosis can develop in nearly any organ, and it is an important driver of tissue stiffness and increases the risk of malignancy[35]. In fibrotic kidney biopsy specimens or multiple experimental kidney fibrosis rodent models, the accumulation of elastin can be observed in renal tissue[36]. In human fibrosis of the liver, kidney, and pancreas, the ECM on average becomes stiffer than normal. Our previous study indicated in human CRC that the collagen development features numerous changes in composition and organization compared with normal colonic tissue[37]. In the present study, we found that collagen components were quantitatively and qualitatively changed in the rat experimental model of CRC. By using picrosirius red, MT, and WRF staining, we revealed a marked increase in collagen and elastin deposition in rats exposed to cold and capsaicin treatment. Furthermore, they were more orderly organized based on the collagen fibers being more aligned with each other, longer, wider, and slightly straighter.

The structure, orientation, and physical properties of collagen regulate the aggressive behavior of cancer. For example, in glioblastoma, Pointer *et al*[38] showed that patients with more organized glioblastoma multiforme collagen survive longer than patients with less organized glioblastoma multiforme collagen. Zhou *et al*[39] also demonstrated that the increased density, length, or width of collagen negatively affects patients with gastric cancer prognosis. The stromal tissue in CRC has also shown that an increase in the collagen content of the ECM increases mechanical stiffness, which predisposes to aggressive CRC[40]. In human breast cancer, linear organization and relatively straight collagen that facilitates migration of tumor cells indicate poor cancer outcome[41]. Similarly, our data indicated that cold and capsaicin exposure increased



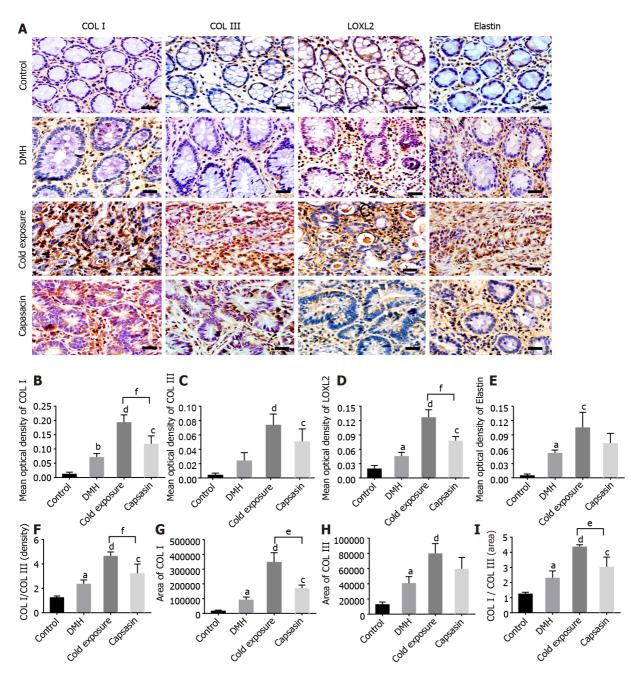


Figure 6 Changes in collagen, elastin and lysyl oxidase-like-2 proteins in the colonic tissues of different treatment groups. A: Protein expressions of type I collagen (COL I), type III collagen (COL III), lysyl oxidase-like-2 (LOXL2), and elastin in the colonic tissues *via* immunohistochemical staining. Magnification, × 400, scalar bar 20 μ m; B-I: Densitometric analysis of COL I (B), COL III (C), LOXL2 (D), elastin(E), COL I/COL III (F), COL I area (G), COL III area (H), and COL I area/COL III area (I) during immunohistochemical staining. ^eP < 0.05, ^bP < 0.01, control compared with 1,2-dimethylhyrazine (DMH); ^eP < 0.05, ^dP < 0.01, COL I, cold exposure and capsaicin-treated group; ^eP < 0.05, ^fP < 0.01, cold exposure compared with capsaicin-treated group.

collagen and elastin deposition, thus triggering alterations in the ECM architecture and organization in the DMH-induced CRC for further tumor development and progression. Furthermore, all parameters of collagen fibers (*e.g.*, density, angle, length, width, and straightness) significantly increased in the cold exposure group and could accurately explain the cold-induced CRC more seriously.

Recently, the relationship between ECM remodeling and malignant transformation of cancer has attracted much attention[42]. COL I is the most abundant protein present in the body. COL I, a major component of collagen, was significantly up-regulated in CRC tissues and showed enhanced CRC migratory capabilities through the overexpression of WNT/planar cell polarity signaling pathway[43]. Moreover, the elevated expression of type I collagen in CRC tissues is correlated to patients with high metastasis that was due to activation of phosphatidylinositol-3-kinase/AKT signaling [44]. Bode *et al*[45] also showed that the expression of COL was increased in malignant colon tissue compared with COL III. Moreover, the elevated expression of COL I has

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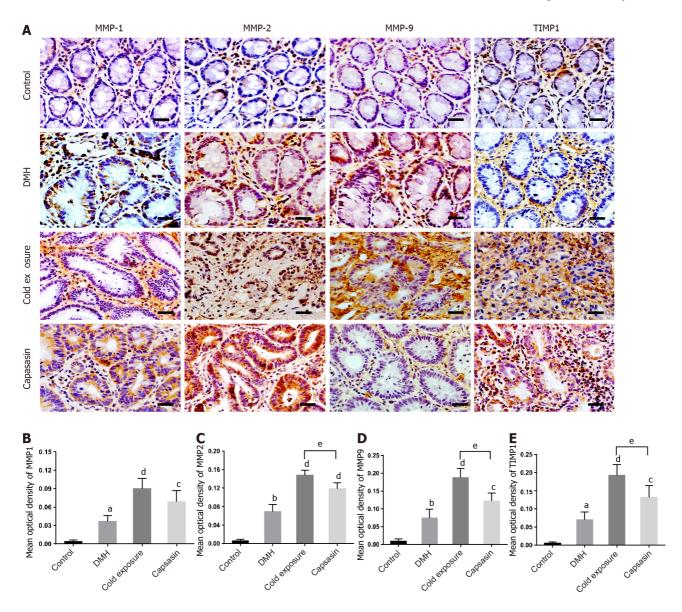


Figure 7 Changes in matrix metalloproteinase 1, matrix metalloproteinase 2, matrix metalloproteinase 9, and tissue-specific matrix metalloproteinase 1 proteins in the colonic tissues of different treatment groups. A: Protein expressions of matrix metalloproteinase (MMP) 1, MMP2, MMP9 and tissue-specific matrix metalloproteinase 1 (TIMP1) in the colonic tissues *via* immunohistochemical staining. Magnification, × 400, scalar bar 20 µm; B-E: Densitometric analysis of MMP1 (B), MMP2 (C), MMP9 (D) and TIMP1 (E) during immunohistochemical staining. $^{e}P < 0.05$, $^{b}P < 0.01$, control compared with 1,2-dimethylhyrazine (DMH); $^{c}P < 0.05$, $^{d}P < 0.01$, DMH compared with cold exposure and capsaicin-treated group; $^{e}P < 0.05$, cold exposure compared with capsaicin-treated group.

been linked to the invasive and aggressive behavior of CRC[46]. In the present study, we also evaluated the expression of COL I in cold exposure and capsaicin treatment CRC colonic tissue. The increase in expression levels of COL I in our study is consistent with other reports in CRC. However, the mRNA level of COL III was not significantly different between the DMH and capsaicin group. In addition, compared with other groups, the COL I/ COL III in cold exposure group was significantly increased. With the increase in collagen expression, distribution area, and collagen ratio, the degree of fibrosis in ECM pathological characteristics was aggravated[47,48]. Therefore, the colonic tissue stiffness was significantly higher than that of the other groups, and the degree of ECM fibrosis in CRC with cold exposure was more serious than that in other groups. ECM remodeling is regulated by ECM enzymes such as LOXL2, MMPs, and TIMPs. ECM-crosslinking enzyme LOXL2 has been implicated in stiffness-associated tumor progression[49]. The LOXL2-mediated collagen cross-linking, both *in vitro* and *in vivo* models of CRC, results in increased tissue stiffness and activation of the focal adhesion kinase/SRC signaling[50].

MMP1, MMP-2, and MMP-9 play a fundamental role in many pathophysiological processes such as cell migration, angiogenesis, and the invasion and metastasis of malignant tumors[51]. TIMPs are the most important physiological inhibitors of MMP,



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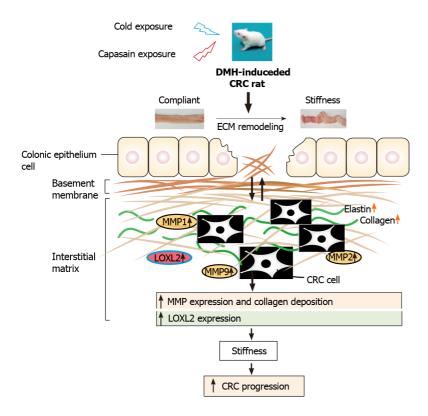


Figure 8 Schematic diagram depicting the role of extracellular matrix and extracellular matrix enzymes in promoting colorectal cancer pathogenesis. Comparison with the normal rats, rats exposed to cold and capsaicin with profound remodeling of extracellular matrix (ECM) in the colonic tissue, which was mediated by ECM enzymes. These results implicate a crucial role of ECM remodeling on cold and capsaicin exposure colorectal cancer development and progression. CRC: Colorectal cancer; DMH: 1,2-Dimethylhyrazine; MMP: Matrix metalloproteinase.

and they are also commonly expressed in tumor sites[52]. The expression of both MMP-2 MMP9 and TIMP-2 is higher in invasive tumors and is strongly associated with angiogenesis in DMH-induced CRC[24]. A previous study indicated that the cross-linking of collagen is known to activate enzymes involved in matrix remodeling, such as LOXL2, MMPs, and TIMPs[53,54]. MMPs are responsible for the degradation of ECM; LOXL2 mediate ECM cross-linking and stiffening[55]. However, recent studies indicated that LOXL2 activity promotes breast cancer metastasis by regulating the expression of MMPs and TIMPs involved in matrix remodeling[56]. LOXL2, TIMP1, and MMP9 are co-expressed during mammary metastasis, suggesting that they function together in glandular remodeling. Our previous study also found that expression levels of LOXL2, MMP1, MMP2, and MMP9 are positively correlated in CRC tissues, and they play synergistic roles in ECM remodeling of human CRC[37].

In the present study, the LOXL2, MMP1, MMP2, MMP9, and TIMP parameters were analyzed, and the results indicated a significant increase in the expression of these proteins in the cold exposure and capsaicin group accompanied by the enhancement of collagen and elastin deposition. Therefore, they may act together in regulating ECM remodeling. Growing insights from experimental studies on the roles of the ECM in CRC suggest that the quantitative and qualitative changes in ECM mediated by specific enzymes promote numerous cellular functions that steer cancer progression and metastasis[57,58]. In the present study, the ECM remodeling in the colonic tissue under cold and capsaicin exposure was more serious, thus increasing the exacerbation severity of CRC. Therefore, environmental factors, such as diet, will affect the internal and external constitutions of organism, causing different manifestations and disease progression in the organism. In the cold exposure and capsaicin treatment group, remodeling of the ECM and stromal stiffness is associated with increased propensity for progression to invasive CRC. Therefore, the levels of ECM remodeling can distinguish different organism characteristics and evaluate the CRC progression successfully, thus providing a novel pathological direction of analysis for clinicians. Furthermore, nanoscale mechanical imaging can be used to observe patients with heterogenous features of ECM, who would benefit most from ECM target therapies.

CONCLUSION

In summary, as shown in Figure 8, the present study revealed profound remodeling of the ECM in cold exposure and long-term administration of capsaicin at a low dose in rats. Collagen signatures including angle, length, width, and straightness have a great impact on CRC progression. Additionally, our results show that higher colonic tissue stiffness might result from ECM enzymes-mediated ECM crosslinking and excessive deposition of collagen and elastin, and such changes are strongly associated with the tumor progression of cold and capsaicin exposure CRC. A better understanding of the role of ECM remodeling and ECM enzymes on the pathogenic mechanisms of colon cancer may help in determining the molecular mechanism of CRC progression and could afford a novel therapeutic intervention in the treatment of this disease.

ARTICLE HIGHLIGHTS

Research background

Colorectal cancer (CRC) is a cancer with high prevalence and mortality in the world. Extracellular matrix (ECM) is a dynamic compartment that regulates tissue development and homeostasis, and its remodeling contributes to neoplastic progression. The cancerous ECM can change cell phenotype and has profound influence on the colonization of metastatic cancer cells. However, the relationship between ECM remodeling and progression and aggression CRC from imposed by cold and capsaicin exposure remains unclear.

Research motivation

To identify the effect of cold exposure and capsaicin on ECM remodeling, ECM enzymes, and the underlying mechanism.

Research objectives

To explore the role of ECM remodeling and ECM enzymes in the 1,2-dimethylhydrazine (DMH)-induced CRC progression and the underlying mechanism.

Research methods

The CRC rat model was conducted by adding DMH and examining the role of ECM remodeling and ECM enzymes on DMH-induced CRC in the model. We investigated the morphology and structure of collagen and elastin using Masson's trichrome, Picrosirius red, and Weigert's Resorcin-Fuchsin stains. Additionally, we evaluated the protein expression level of type I collagen (COL I), type III collagen (COL III), elastin, lysyl oxidase-like 2 (LOXL2), matrix metalloproteinase (MMP) 1, MMP2, MMP9, and tissue-specific matrix metalloproteinase 1 by immunohistochemistry and observed the expression of COL I, COL III, elastin, and LOXL2 in the colon tissues of rats by reverse-transcriptase quantitative polymerase chain reaction.

Research results

We found that although there were no differences in the proportion of adenomas, a trend towards the increase of invasive tumors was observed in the cold and capsaicin group. Cold exposure group had a metastasis rate comparative with the other groups. Additionally, abnormal accumulation of both collagen and elastin was observed in the cold exposure and capsaicin group. Specifically, collagen quantitative analysis showed increased length, width, angle, and straightness compared with the DMH group. Collagen deposition and straightness were significantly increased in the cold exposure group compared with the capsaicin group. Cold exposure and capsaicin significantly increased the protein levels of COL I, elastin, and LOXL2 along with increases in their messenger RNA levels in the colon tissues compared with the DMH group, while COL III did not show a significant difference. Furthermore, in immunohistochemical evaluations, MMP1, MMP2, MMP9, and tissue-specific matrix metalloproteinase 1 staining increased in the cold exposure and capsaicin group compared with the DMH group.

Research conclusions

Increased stiffness of colonic tissue and the remodeling of ECM mediated by ECM enzymes resulted from cold and capsaicin exposure, predisposing an environment suitable for CRC development and progression.



Research perspectives

To target ECM in CRC tumor tissue could represent a novel potential therapeutic strategy.

ACKNOWLEDGEMENTS

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REFERENCES

- Okugawa Y, Grady WM, Goel A. Epigenetic Alterations in Colorectal Cancer: 1 Emerging Biomarkers. Gastroenterology 2015; 149: 1204-1225.e12 [PMID: 26216839 DOI: 10.1053/j.gastro.2015.07.011]
- Dionigi G, Bianchi V, Rovera F, Boni L, Annoni M, Castano P, Villa F, Dionigi R. Genetic alteration 2 in hereditary colorectal cancer. Surg Oncol 2007; 16 Suppl 1: S11-S15 [PMID: 18023570 DOI: 10.1016/j.suronc.2007.10.020]
- Murphy N, Moreno V, Hughes DJ, Vodicka L, Vodicka P, Aglago EK, Gunter MJ, Jenab M. Lifestyle and dietary environmental factors in colorectal cancer susceptibility. Mol Aspects Med 2019; 69: 2-9 [PMID: 31233770 DOI: 10.1016/j.mam.2019.06.005]
- 4 Lehrer S, Rosenzweig KE. Cold Climate Is a Risk Factor for Thyroid Cancer. Clin Thyroidol 2014; 26: 273-276 [PMID: 25558467 DOI: 10.1089/ct.2014;26.273-276]
- 5 López-Carrillo L, López-Cervantes M, Robles-Díaz G, Ramírez-Espitia A, Mohar-Betancourt A, Meneses-García A, López-Vidal Y, Blair A. Capsaicin consumption, Helicobacter pylori positivity and gastric cancer in Mexico. Int J Cancer 2003; 106: 277-282 [PMID: 12800206 DOI: 10.1002/ijc.11195]
- 6 Serra I, Yamamoto M, Calvo A, Cavada G, Báez S, Endoh K, Watanabe H, Tajima K. Association of chili pepper consumption, low socioeconomic status and longstanding gallstones with gallbladder cancer in a Chilean population. Int J Cancer 2002; 102: 407-411 [PMID: 12402311 DOI: 10.1002/ijc.10716]
- Frantz C, Stewart KM, Weaver VM. The extracellular matrix at a glance. J Cell Sci 2010; 123: 4195-7 4200 [PMID: 21123617 DOI: 10.1242/jcs.023820]
- Nissen NI, Karsdal M, Willumsen N. Collagens and Cancer associated fibroblasts in the reactive 8 stroma and its relation to Cancer biology. J Exp Clin Cancer Res 2019; 38: 115 [PMID: 30841909 DOI: 10.1186/s13046-019-1110-6]
- Malik R, Lelkes PI, Cukierman E. Biomechanical and biochemical remodeling of stromal 9 extracellular matrix in cancer. Trends Biotechnol 2015; 33: 230-236 [PMID: 25708906 DOI: 10.1016/j.tibtech.2015.01.004
- 10 Drifka CR, Tod J, Loeffler AG, Liu Y, Thomas GJ, Eliceiri KW, Kao WJ. Periductal stromal collagen topology of pancreatic ductal adenocarcinoma differs from that of normal and chronic pancreatitis. Mod Pathol 2015; 28: 1470-1480 [PMID: 26336888 DOI: 10.1038/modpathol.2015.97]
- Esbona K, Yi Y, Saha S, Yu M, Van Doorn RR, Conklin MW, Graham DS, Wisinski KB, Ponik SM, 11 Eliceiri KW, Wilke LG, Keely PJ. The Presence of Cyclooxygenase 2, Tumor-Associated Macrophages, and Collagen Alignment as Prognostic Markers for Invasive Breast Carcinoma Patients. Am J Pathol 2018; 188: 559-573 [PMID: 29429545 DOI: 10.1016/j.ajpath.2017.10.025]
- 12 Provenzano PP, Eliceiri KW, Campbell JM, Inman DR, White JG, Keely PJ. Collagen reorganization at the tumor-stromal interface facilitates local invasion. BMC Med 2006; 4: 38 [PMID: 17190588 DOI: 10.1186/1741-7015-4-38]
- 13 Herrera J, Henke CA, Bitterman PB. Extracellular matrix as a driver of progressive fibrosis. J Clin Invest 2018; 128: 45-53 [PMID: 29293088 DOI: 10.1172/JCI93557]
- Kendall TJ, Dolman GE, Duff CM, Paish EC, Zaitoun A, Irving W, Fallowfield JA, Guha IN. 14 Hepatic elastin content is predictive of adverse outcome in advanced fibrotic liver disease. Histopathology 2018; 73: 90-100 [PMID: 29464815 DOI: 10.1111/his.13499]
- 15 Schmelzer CEH, Heinz A, Troilo H, Lockhart-Cairns MP, Jowitt TA, Marchand MF, Bidault L, Bignon M, Hedtke T, Barret A, McConnell JC, Sherratt MJ, Germain S, Hulmes DJS, Baldock C, Muller L. Lysyl oxidase-like 2 (LOXL2)-mediated cross-linking of tropoelastin. FASEB J 2019; 33: 5468-5481 [PMID: 30676771 DOI: 10.1096/fj.201801860RR]
- 16 Wang TH, Hsia SM, Shieh TM. Lysyl Oxidase and the Tumor Microenvironment. Int J Mol Sci 2016; 18 [PMID: 28036074 DOI: 10.3390/ijms18010062]
- Rachman-Tzemah C, Zaffryar-Eilot S, Grossman M, Ribero D, Timaner M, Mäki JM, Myllyharju J, 17 Bertolini F, Hershkovitz D, Sagi I, Hasson P, Shaked Y. Blocking Surgically Induced Lysyl Oxidase Activity Reduces the Risk of Lung Metastases. Cell Rep 2017; 19: 774-784 [PMID: 28445728 DOI: 10.1016/j.celrep.2017.04.005
- 18 Cui X, Wang G, Shen W, Huang Z, He H, Cui L. Lysyl oxidase-like 2 is highly expressed in colorectal cancer cells and promotes the development of colorectal cancer. Oncol Rep 2018; 40: 932-942 [PMID: 29845296 DOI: 10.3892/or.2018.6452]



- Zhan P, Lv XJ, Ji YN, Xie H, Yu LK. Increased lysyl oxidase-like 2 associates with a poor prognosis 19 in non-small cell lung cancer. Clin Respir J 2018; 12: 712-720 [PMID: 27860390 DOI: 10.1111/crj.12584]
- 20 Choi J, Chung T, Rhee H, Kim YJ, Jeon Y, Yoo JE, Noh S, Han DH, Park YN. Increased Expression of the Matrix-Modifying Enzyme Lysyl Oxidase-Like 2 in Aggressive Hepatocellular Carcinoma with Poor Prognosis. Gut Liver 2019; 13: 83-92 [PMID: 29938458 DOI: 10.5009/gn117569]
- Cui N, Hu M, Khalil RA. Biochemical and Biological Attributes of Matrix Metalloproteinases. Prog 21 Mol Biol Transl Sci 2017; 147: 1-73 [PMID: 28413025 DOI: 10.1016/bs.pmbts.2017.02.005]
- 22 Page-McCaw A, Ewald AJ, Werb Z. Matrix metalloproteinases and the regulation of tissue remodelling. Nat Rev Mol Cell Biol 2007; 8: 221-233 [PMID: 17318226 DOI: 10.1038/nrm2125]
- 23 Li BH, Zhao P, Liu SZ, Yu YM, Han M, Wen JK. Matrix metalloproteinase-2 and tissue inhibitor of metallo-proteinase-2 in colorectal carcinoma invasion and metastasis. World J Gastroenterol 2005; 11: 3046-3050 [PMID: 15918187 DOI: 10.3748/wjg.v11.i20.3046]
- Gungor H, Ilhan N, Eroksuz H. The effectiveness of cyclooxygenase-2 inhibitors and evaluation of 24 angiogenesis in the model of experimental colorectal cancer. Biomed Pharmacother 2018; 102: 221-229 [PMID: 29562216 DOI: 10.1016/j.biopha.2018.03.066]
- Xu GF, Li PT, Wang XY, Jia X, Tian DL, Jiang LD, Yang JX. Dynamic changes in the expression of 25 matrix metalloproteinases and their inhibitors, TIMPs, during hepatic fibrosis induced by alcohol in rats. World J Gastroenterol 2004; 10: 3621-3627 [PMID: 15534918 DOI: 10.3748/wjg.v10.i24.3621]
- Li HX, Tan JQ, Lv ZH, Liang YQ, Wen B. Colorectal Cancer Modeling and Difference Analysis in 26 Cold and Heat Conditions. Zhongguo Shiyan Fangji Xue Zazhi 2020; 26: 109-117 [DOI: 10.13422/j.cnki.syfjx.20201964]
- 27 Jikihara H, Qi G, Nozoe K, Hirokawa M, Sato H, Sugihara Y, Shimamoto F. Aged garlic extract inhibits 1,2-dimethylhydrazine-induced colon tumor development by suppressing cell proliferation. Oncol Rep 2015; 33: 1131-1140 [PMID: 25573280 DOI: 10.3892/or.2014.3705]
- 28 Liu Y, Keikhosravi A, Mehta GS, Drifka CR, Eliceiri KW. Methods for Quantifying Fibrillar Collagen Alignment. Methods Mol Biol 2017; 1627: 429-451 [PMID: 28836218 DOI: 10.1007/978-1-4939-7113-8 28
- Keikhosravi A, Bredfeldt JS, Sagar AK, Eliceiri KW. Second-harmonic generation imaging of 29 cancer. Methods Cell Biol 2014; 123: 531-546 [PMID: 24974046 DOI: 10.1016/B978-0-12-420138-5.00028-8
- Schindelin J, Arganda-Carreras I, Frise E, Kaynig V, Longair M, Pietzsch T, Preibisch S, Rueden C, 30 Saalfeld S. Schmid B. Tinevez JY, White DJ, Hartenstein V, Eliceiri K, Tomancak P, Cardona A, Fiji: an open-source platform for biological-image analysis. Nat Methods 2012; 9: 676-682 [PMID: 22743772 DOI: 10.1038/nmeth.2019]
- Walker C, Mojares E, Del Río Hernández A. Role of Extracellular Matrix in Development and 31 Cancer Progression. Int J Mol Sci 2018; 19 [PMID: 30287763 DOI: 10.3390/ijms19103028]
- 32 Crotti S, Piccoli M, Rizzolio F, Giordano A, Nitti D, Agostini M. Extracellular Matrix and Colorectal Cancer: How Surrounding Microenvironment Affects Cancer Cell Behavior? J Cell Physiol 2017; 232: 967-975 [PMID: 27775168 DOI: 10.1002/jcp.25658]
- Mohan V, Das A, Sagi I. Emerging roles of ECM remodeling processes in cancer. Semin Cancer Biol 33 2020; 62: 192-200 [PMID: 31518697 DOI: 10.1016/j.semcancer.2019.09.004]
- Pickup MW, Mouw JK, Weaver VM. The extracellular matrix modulates the hallmarks of cancer. 34 EMBO Rep 2014; 15: 1243-1253 [PMID: 25381661 DOI: 10.15252/embr.201439246]
- Cernaro V, Lacquaniti A, Donato V, Fazio MR, Buemi A, Buemi M. Fibrosis, regeneration and 35 cancer: what is the link? Nephrol Dial Transplant 2012; 27: 21-27 [PMID: 22102616 DOI: 10.1093/ndt/gfr567
- Sun Q, Baues M, Klinkhammer BM, Ehling J, Djudjaj S, Drude NI, Daniel C, Amann K, Kramann R, 36 Kim H, Saez-Rodriguez J, Weiskirchen R, Onthank DC, Botnar RM, Kiessling F, Floege J, Lammers T, Boor P. Elastin imaging enables noninvasive staging and treatment monitoring of kidney fibrosis. Sci Transl Med 2019; 11 [PMID: 30944168 DOI: 10.1126/scitranslmed.aat4865]
- 37 Liang Y, Lv Z, Huang G, Qin J, Li H, Nong F, Wen B. Prognostic significance of abnormal matrix collagen remodeling in colorectal cancer based on histologic and bioinformatics analysis. Oncol Rep 2020; 44: 1671-1685 [PMID: 32945508 DOI: 10.3892/or.2020.7729]
- Pointer KB, Clark PA, Schroeder AB, Salamat MS, Eliceiri KW, Kuo JS. Association of collagen 38 architecture with glioblastoma patient survival. J Neurosurg 2017; 126: 1812-1821 [PMID: 27588592 DOI: 10.3171/2016.6.JNS152797]
- Zhou ZH, Ji CD, Xiao HL, Zhao HB, Cui YH, Bian XW. Reorganized Collagen in the Tumor 39 Microenvironment of Gastric Cancer and Its Association with Prognosis. J Cancer 2017; 8: 1466-1476 [PMID: 28638462 DOI: 10.7150/jca.18466]
- 40Nebuloni M, Albarello L, Andolfo A, Magagnotti C, Genovese L, Locatelli I, Tonon G, Longhi E, Zerbi P, Allevi R, Podestà A, Puricelli L, Milani P, Soldarini A, Salonia A, Alfano M. Insight On Colorectal Carcinoma Infiltration by Studying Perilesional Extracellular Matrix. Sci Rep 2016; 6: 22522 [PMID: 26940881 DOI: 10.1038/srep22522]
- Conklin MW, Eickhoff JC, Riching KM, Pehlke CA, Eliceiri KW, Provenzano PP, Friedl A, Keely PJ. Aligned collagen is a prognostic signature for survival in human breast carcinoma. Am J Pathol 2011; 178: 1221-1232 [PMID: 21356373 DOI: 10.1016/j.ajpath.2010.11.076]
- Lu P, Weaver VM, Werb Z. The extracellular matrix: a dynamic niche in cancer progression. J Cell 42 Biol 2012; 196: 395-406 [PMID: 22351925 DOI: 10.1083/jcb.201102147]



- 43 Zhang Z, Wang Y, Zhang J, Zhong J, Yang R. COL1A1 promotes metastasis in colorectal cancer by regulating the WNT/PCP pathway. Mol Med Rep 2018; 17: 5037-5042 [PMID: 29393423 DOI: 10.3892/mmr.2018.8533
- Wu X, Cai J, Zuo Z, Li J. Collagen facilitates the colorectal cancer stemness and metastasis through 44 an integrin/PI3K/AKT/Snail signaling pathway. Biomed Pharmacother 2019; 114: 108708 [PMID: 30913493 DOI: 10.1016/j.biopha.2019.108708]
- Bode MK, Karttunen TJ, Mäkelä J, Risteli L, Risteli J. Type I and III collagens in human colon 45 cancer and diverticulosis. Scand J Gastroenterol 2000; 35: 747-752 [PMID: 10972180 DOI: 10.1080/003655200750023435]
- 46 Wei B, Zhou X, Liang C, Zheng X, Lei P, Fang J, Han X, Wang L, Qi C, Wei H. Human colorectal cancer progression correlates with LOX-induced ECM stiffening. Int J Biol Sci 2017; 13: 1450-1457 [PMID: 29209148 DOI: 10.7150/ijbs.21230]
- Beam J, Botta A, Ye J, Soliman H, Matier BJ, Forrest M, MacLeod KM, Ghosh S. Excess Linoleic Acid Increases Collagen I/III Ratio and "Stiffens" the Heart Muscle Following High Fat Diets. J Biol Chem 2015; 290: 23371-23384 [PMID: 26240151 DOI: 10.1074/jbc.M115.682195]
- Gilkes DM, Semenza GL, Wirtz D. Hypoxia and the extracellular matrix: drivers of tumour 48 metastasis. Nat Rev Cancer 2014; 14: 430-439 [PMID: 24827502 DOI: 10.1038/nrc3726]
- 49 Piersma B, Bank RA. Collagen cross-linking mediated by lysyl hydroxylase 2: an enzymatic battlefield to combat fibrosis. Essays Biochem 2019; 63: 377-387 [PMID: 31324706 DOI: 10.1042/EBC20180051
- Baker AM, Bird D, Lang G, Cox TR, Erler JT. Lysyl oxidase enzymatic function increases stiffness 50 to drive colorectal cancer progression through FAK. Oncogene 2013; 32: 1863-1868 [PMID: 22641216 DOI: 10.1038/onc.2012.202]
- 51 Hojilla CV, Mohammed FF, Khokha R. Matrix metalloproteinases and their tissue inhibitors direct cell fate during cancer development. Br J Cancer 2003; 89: 1817-1821 [PMID: 14612884 DOI: 10.1038/sj.bjc.6601327]
- Deryugina EI, Quigley JP. Matrix metalloproteinases and tumor metastasis. Cancer Metastasis Rev 52 2006; 25: 9-34 [PMID: 16680569 DOI: 10.1007/s10555-006-7886-9]
- 53 Dittmore A, Silver J, Sarkar SK, Marmer B, Goldberg GI, Neuman KC. Internal strain drives spontaneous periodic buckling in collagen and regulates remodeling. Proc Natl Acad Sci USA 2016; 113: 8436-8441 [PMID: 27402741 DOI: 10.1073/pnas.1523228113]
- Erler JT, Bennewith KL, Cox TR, Lang G, Bird D, Koong A, Le QT, Giaccia AJ. Hypoxia-induced 54 lysyl oxidase is a critical mediator of bone marrow cell recruitment to form the premetastatic niche. Cancer Cell 2009; 15: 35-44 [PMID: 19111879 DOI: 10.1016/j.ccr.2008.11.012]
- 55 Afratis NA, Klepfish M, Karamanos NK, Sagi I. The apparent competitive action of ECM proteases and cross-linking enzymes during fibrosis: Applications to drug discovery. Adv Drug Deliv Rev 2018; 129: 4-15 [PMID: 29627371 DOI: 10.1016/j.addr.2018.03.004]
- Barker HE, Chang J, Cox TR, Lang G, Bird D, Nicolau M, Evans HR, Gartland A, Erler JT. 56 LOXL2-mediated matrix remodeling in metastasis and mammary gland involution. Cancer Res 2011; 71: 1561-1572 [PMID: 21233336 DOI: 10.1158/0008-5472.CAN-10-2868]
- Bonnans C, Chou J, Werb Z. Remodelling the extracellular matrix in development and disease. Nat 57 Rev Mol Cell Biol 2014; 15: 786-801 [PMID: 25415508 DOI: 10.1038/nrm3904]
- Brauchle E, Kasper J, Daum R, Schierbaum N, Falch C, Kirschniak A, Schäffer TE, Schenke-Layland K. Biomechanical and biomolecular characterization of extracellular matrix structures in human colon carcinomas. Matrix Biol 2018; 68-69: 180-193 [PMID: 29605717 DOI: 10.1016/j.matbio.2018.03.016



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ORIGINAL ARTICLE

Basic Study Detection and analysis of common pathogenic germline mutations in **Peutz-Jeghers syndrome**

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Abstract

BACKGROUND

Different types of pathogenic mutations may produce different clinical phenotypes, but a correlation between Peutz-Jeghers syndrome (PJS) genotype and clinical phenotype has not been found. Not all patients with PJS have detectable mutations of the STK11/LKB1 gene, what is the genetic basis of clinical phenotypic heterogeneity of PJS? Do PJS cases without STK11/LKB1 mutations have other pathogenic genes? Those are clinical problems that perplex doctors.

AIM

The aim was to investigate the specific gene mutation of PJS, and the correlation between the genotype and clinical phenotype of PJS.

METHODS

A total of 24 patients with PJS admitted to the Air Force Medical Center, PLA (formerly the Air Force General Hospital, PLA) from November 1994 to January 2020 were randomly selected for inclusion in the study. One hundred thirty-nine common hereditary tumor-related genes including STK11/LKB1 were screened and analyzed for pathogenic germline mutations by high-throughput nextgeneration sequencing (NGS). The mutation status of the genes and their relationship with clinical phenotypes of PJS were explored.

RESULTS



patients (legal guardians of minors) understood the process and purpose of this study and signed an informed consent form. In the process of sample collection, follow the principles of informed consent in the Declaration of Helsinki, the Universal Declaration of Human Genome and Human Rights, and the Declaration of the Human Genome Ethics Committee on DNA Sampling, Control, and Acquisition. No additional data are available.

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Twenty of the 24 PJS patients in this group (83.3%) had STK11/LKB1 gene mutations, 90% of which were pathogenic mutations, and ten had new mutation sites. Pathogenic mutations in exon 7 of STK11/LKB1 gene were significantly lower than in other exons. Truncation mutations are more common in exons 1 and 4 of *STK11/LKB1*, and their pathogenicity was significantly higher than that of missense mutations. We also found SLX4 gene mutations in PJS patients.

CONCLUSION

PJS has a relatively complicated genetic background. Changes in the sites responsible for coding functional proteins in exon 1 and exon 4 of STK11/LKB1 may be one of the main causes of PJS. Mutation of the SLX4 gene may be a cause of genetic heterogeneity in PJS.

Key Words: Peutz-Jeghers syndrome; Genotype; Phenotype; STK11; Mutation

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Core Tip: It is currently believed that Peutz-Jeghers syndrome (PJS) is an autosomal dominant genetic disease predominantly caused by germline mutations in the STK11/LKB1 gene. No correlation of the PJS genotype and clinical phenotype has been found so far. The correlation of genotype and clinical phenotype and exploration of the internal molecular mechanism of different clinical phenotypes were studied in 24 treated PJS patients with different clinical phenotypes. Peripheral venous blood or normal tissue adjacent to polyps were collected for high-throughput next-generation sequencing (NGS) of 139 hereditary colorectal tumor-related genes including STK11/LKB1. A newly discovered likely pathogenic gene (SLX4) provided new data explaining the genetic heterogeneity of PJS.

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INTRODUCTION

It is currently believed that Peutz-Jeghers syndrome (PJS) is an autosomal dominant genetic disease predominantly caused by germline mutations in the STK11/LKB1 gene. PJS is characterized by multiple hamartoma polyps in the gastrointestinal tract, pigmentation at specific sites, and hereditary tumors[1-4]. Pathogenic mutations of STK11/LKB1 lead to inactivation of its expression product and loss of inhibition of mammalian target of rapamycin (mTOR) activity, which leads to abnormal activation of the LKB1/mTOR signal pathway and the occurrence of black spots on the skin and gastrointestinal hamartoma polyps[5]. More than 400 different pathogenic STK11/LKB1 gene mutations are included in the Human Gene Mutation Database (HGMD), most of which are microminiature. Different types of pathogenic mutations may produce different clinical phenotypes, but no correlations of PJS genotype and clinical phenotype has been found so far[6], Not all patients with PJS have detectable mutations in the *STK11/LKB1* gene. What is the genetic basis of clinical phenotypic heterogeneity in PJS? Do PJS patients without STK11/LKB1 mutations have other pathogenic genes? These are clinical problems that perplex doctors [7,8]. We enrolled 24 patients treated for PJS. Peripheral venous blood and normal tissue adjacent to polyps were collected for high-throughput next-generation sequencing (NGS) of 139 hereditary colorectal tumor-related genes including STK11/LKB1 to study the correlation between genotype and clinical phenotype of PJS and explore the internal molecular mechanism of the clinical phenotypes.





MATERIALS AND METHODS

Study participants

Patients with PJS, from 18-70 years of age, met the clinical diagnostic criteria of PJs, had complete clinicopathological data, well preserved specimens, were eligible for inclusion. All participants gave their signed informed consent. Patients who could not provide experimental specimens or did not agree to participate in the study were excluded. Twenty-four PJS patients admitted to the Air Force Medical Center (formerly the Air Force General Hospital) from November 1994 to January 2020 met the above criteria and were enrolled. Their clinical information is shown in Table 1. Twenty-three were inpatients, one was an outpatient, 11 had family histories, and 12 had early onset pigment spots that had appeared when they were younger than 3 years of age. All patients met the PJS diagnostic criteria recommended by the National Comprehensive Cancer Network (NCCN)[9]. The experimental samples included 5 mL peripheral venous blood samples collected from 19 patients into tubes containing EDTA-2Na, and paraffin-embedded normal tissue surgically removed from areas adjacent to polyps in five patients. The study was reviewed and approved by the Ethics Committee of the Air Force Medical Center and the Second Affiliated Hospital of Zhejiang University School of Medicine. All patients or the legal guardians of minors, understood the process and purpose of this study and signed an informed consent form. Sample collection followed the ethical principles of the Declaration of Helsinki, the Universal Declaration of Human Genome and Human Rights, and the Declaration of the Human Genome Ethics Committee on DNA Sampling, Control, and Acquisition.

Methods

DNA was extracted from peripheral venous blood samples with TGuide Blood Genomic DNA Kits (CHI-TIANGEN) following the manufacturer's instructions. DNA was extracted from paraffin-embedded tissue specimens with QIAamp DNA FFPE micro sample tissue kits (GER-QIAGEN). Nucleic acids were broken into small, random 150-200 bp fragments by ultrasonic fragmentation (Covaris S220) and separated and evaluated with a Tapestation 2200 electrophoresis working platform (Agilent) to check whether the fragments met the requirements for library construction. A standard gene library was constructed using KAPA HyperPlus Kit (Illumina). A panel of 139 common tumor genetic susceptibility genes including colorectal cancer (Table 2) was selected and provided by Genetron Health Co.(Beijing). The specific gene capture probe was hybridized with the library in the environment of a hybridization buffer, and purified by the magnetic bead method. High-throughput NGS was performed with a Novaseq 6000 sequencer (Illumina, United States). Trimmomatic (version 0.33) was used to crop and filter the original data, which was stored in FastQ format, after sequencing. The reads at the end of each pair were aligned with the human reference sequence GRCh37 (hg19) using the BWA-MEM algorithm (BWA version 0.7.10-r789) and the default parameters. The Picard tool (version 1.103 http://broadinstitute.github.io/picard/) was used to delete duplicate readings, and GATK (version 3.1-0-g72492bb) was used to realign the sequences around the known insertion loss at the single sample level and to recalibrate the base quality. Integrative Genomics Viewer version 2.3.34 (https://software.broadins titute.org/software/igv/) was used to check the mutations in the coding region.

The Chinese (1000 CN), general population (1000 MAF). and dbSNP (https://www.ncbi.nlm.nih.gov/) at 1000 Genome Project (http://ftp.ncbi.nih.gov/) Snip/), ESP6500 AA/EA (NHLBI GO Exome Sequencing Project https://evs gs.washington.edu/EVS/), ExAC MAF (The Exome Aggregation Consortium) and other population databases were searched for the mutation frequency of this gene. The location of genes with a mutation frequency < 0.01 in the HGMD database (HGMD-PUBLIC version 20152) were used for pathogenicity analysis.

The diseases that the variant gene was related to were searched in the OMIM disease database (https://omim.org/) by ClinVar (https://www.ncbi.nlm. nih.gov/clinvar/). HGMD https://www.hgmd.cf.ac.uk) retrieved the description of the mutation. SIFT[10] (http://sift.jcvi.org), PolyPhen2[11] (http://genetics.bwh. harvard.edu/pph2), and Mutation Assessor (http://mutationassessor.org) make conservative predictions of amino acid sequences. The results were used to evaluate the pathogenicity of the mutations[12,13].

SPSS 24.0 was used for statistical analysis of the acquired data. Qualitative results were reported as numbers and percentages. The chi-square test or Fisher's exact probability method was used for between-group comparisons. P < 0.05 was considered



Table 1 Clinical characteristics of 24 enrolled Peutz-Jeghers syndrome patients

No.	Gender	Specimen	Time since onset of pigment spots (yr)	Early or late onset	Family history (members)	Number of hospitalizations	Number of operations	Stomach and enteroscopy times	Age at initial diagnosis of polyps	Age at first treatment	Polyp pathology	Load of Gastric polyps/Max. diameter (mm)	Load of small intestinal polyps/Max. diameter (mm)	Load of colorectal polyps/Max. diameter (mm)
1	Male	Paraffin section	20	Late	No	2	1	6	20	15	1	/	20/30	/
2	Male	Paraffin section	6	Late	Yes (mother and sister)	1	2	3	9	9	1	2/16	20/40	1/8
3	Female	Paraffin section	4	Late	No	2	1	4	9	9	1	/	3/28	/
4	Male	Paraffin section	5	Late	No	1	2	1	21	21	3	20/4	6/50	/
5	Male	Paraffin section	1	Early	Yes (mother)	4	2	1	4	4	1	2/12	2/60	/
6	Female	Blood	5	Late	Yes (father)	1	0	1	29	29	1	/	/	/
7	Female	Blood	1	Early	Yes (father and sister)	4	0	11	7	7	1	1/8	2/30	3/40
8	Male	Blood	0	Early	Yes (father and sister)	1	0	1	10	10	1	/	10/50	/
9	Male	Blood	6	Late	Yes (mother and grandmother)	4	1	7	6	7	1	5/12	2/30	3/35
10	Female	Blood	2	Early	No	1	0	3	7	7	1	2/15	/	1/30
11	Male	Blood	3	Late	No	1	4	0	22	32	1	/	1/30	/
12	Male	Blood	2	Early	No	2	1	10	4	4	1	1/6	2/50	/
13	Male	Blood	2	Early	No	1	2	1	25	24	1	/	10/20	/
14	Female	Blood	3	Late	No	8	2	8	6	6	1	1/10	8/80	1/20
15	Male	Blood	5	Late	No	1	2	3	20	19	2	1/6	1/80	2/30
16	Male	Blood	1	Early	Yes (mother)	3	0	2	10	9	1	/	1/25	/
17	Male	Blood	1	Early	No	3	1	4	6	6	1	8/40	10/30	/
18	Female	Blood	1	Early	No	6	2	9	11	10	1	1/15	3/35	1/50
19	Female	Blood	3	Late	Yes (mother)	2	0	4	15	15	1	1/12	2/12	1/25

20	Female	Blood	3	Late	Yes (father, uncle, and grandmother)	2	2	5	7	7	1	/	18/50	/
21	Female	Blood	1	Early	Yes (mother, uncle, and aunt)	2	0	4	31	31	1	/	10/50	10/40
22	Female	Blood	2	Early	Yes (father and brother)	1	0	1	6	6	1	10/10	8/50	/
23	Male	Blood	5	Late	No	1	0	2	11	11	1	1/30	5/70	1/30
24	Male	Blood	2	Early	No	1	0	4	5	4	1	10/15	/	/

(1) *STK11* mutation, *SLX4* mutation, other gene mutation groups: 0: None 1: Yes; (2) Early onset: Pigment spots appeared at < 3 years of age; Late onset: Pigment spots appeared at \geq 3 years of age; (3) Polyp pathology: 1 hamartoma, 2 hamartoma with adenoma, 3 hamartoma with cancer; (4) Polyp load is the number of polyps, the largest diameter unit is mm; and (5) 6 was an outpatient, the results of previous endoscopy are unknown.

statistically significant.

RESULTS

STK11/LKB1 gene detection results and pathogenicity analysis

Twenty of the 24 PJS patients (83.3%) in this group had STK11/LKB1 gene mutations (Table 3). All were heterozygous and ten were newly discovered mutation sites not included in the dbSNP database. There were eight frameshift mutations, five splicesite mutations, four missense mutations and three nonsense mutations. The mutations occurred in eight of the ten exons in the STK11/LKB1 gene, mutations in exons 1 and 4 and 4 each in exon 7, two in each exons 5 and 8, and one in exons 2, 3, and 6. Frameshift mutations, splice-site mutations, and nonsense mutations were all related to pathogenicity. Frameshift mutations accounted for 62.5% (5/8) that were clearly pathogenic, and 37.5% (3/8) that might cause disease. Splice-site mutations accounted for 40% (2/5) that are clearly pathogenic, and 60% (3/5) that might cause disease. All three nonsense mutations were clearly pathogenic, and the missense mutations were related to and might cause disease. Sites of unclear clinical significance accounted for 50% (2/4); of the 11 truncated mutations, eight cases were clearly pathogenic and three were likely to cause disease. The pathogenicity of STK11 gene mutations in exon 7 was significantly lower than that of other exons (P = 0.000). Truncation mutations were significantly more pathogenic than missense mutations (P = 0.012). The prediction results of bioinformatics tools for missense mutations are shown in Table 4, and the relevant database records and the pathogenicity judgment of all mutations are shown in Table 5.

Table 2 Cancer gen	etic susceptibility 139	gene panel coverage			
AIP	CYLD	FANCL	MLH3	PRSS1	SMARCA4
ALK	DDB2	FANCM	MRE11A	PTCH1	SMARCB1
APC	DICER1	FAS	MSH2	РТСН2	SMARCE1
ATM	DIS3L2	FH	MSH6	PTEN	SOS1
ATR	EGFR	FLCN	MTAP	PTPN11	STAT3
AXIN2	ELANE	GALNT12	MTUS1	RAD50	STK11
BAP1	EPCAM	GATA2	МИТҮН	RAD51B	SUFU
BARD1	ERCC1	GEN1	NBN	RAD51C	TERT
BLM	ERCC2	GJB2	NF1	RAD51D	TGFBR1
BMPR1A	ERCC3	GPC3	NF2	RB1	TMEM127
BRCA1	ERCC4	GREM1	NSD1	RECQL	TP53
BRCA2	ERCC5	HMBS	NTRK1	RECQL4	TSC1
BRIP1	EXT1	HNF1A	PALB2	RET	TSC2
BUB1B	EXT2	HOXB13	PALLD	RHBDF2	UROD
CBL	EZH2	HRAS	PDGFRA	RUNX1	USHBP1
CDC73	FANCA	KIT	РНОХ2В	SBDS	VEGFA
CDH1	FANCB	LASP1	PMS1	SDHA	VHL
CDK4	FANCC	MAX	PMS2	SDHAF2	WRN
CDKN1B	FANCD2	MC1R	POLD1	SDHB	WT1
CDKN1C	FANCE	MEN1	POLE	SDHC	XPA
CDKN2A	FANCF	MET	POLH	SDHD	ХРС
CEBPA	FANCG	MTTF	PPM1D	SLX4	XRCC2
CHEK1	FANCI	MLH1	PRKAR1A	SMAD4	ZMAT3
СНЕК2					

Considering that the type of specimen may impact on the detection rate of *STK11/LKB1* gene mutations, we analyzed the paraffin-embedded tissue and blood samples separately. The detection rate of *STK11/LKB1* mutations in 60 patients with paraffin samples was 60% (3/5), slightly less than the 89.4% (17/19) of the blood samples from 19 patients. The difference in mutation detection rate of this gene in the two types of sample was not statistically different (P = 0.116).

SLX4 gene detection results and pathogenicity analysis

SLX4 gene mutation (Table 6) was detected in 5 PJS patient samples in this group, with a total detection rate of 20.83% (5/24), all of which were heterozygous mutations. The mutation occurred in 4 of 15 exons of SLX4 gene. Mutation types include: 3 missense mutations, one splice-site mutation, and one non-frameshift mutation. No truncation mutation was found. The SLX4 gene is a tumor suppressor gene, and there are three newly discovered mutation sites. The prediction results of three cases of missense mutations by bioinformatics tools (Table 7), the collection of relevant databases and the judgment of the pathogenicity of all mutations (Table 8) are as follows.

Other gene detection results and pathogenicity analysis

A total of 55 mutations of 46 genes other than *STK11/LKB1* and *SLX4* were detected in 21 cases (Table 9), f a detection rate of 87.5% (21/24). Twenty-three of the genes were related to cancer suppression and had 32 different mutation sites. Two mismatch repair *MMR* genes were detected, *MSH2*, *MSH6*. Except for a frameshift mutation (frameshift deletion) in the *BRIP1* gene detected in one patient (No. 18), the rest were missense mutations (Table 10).

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Table	• 3 Characteristics of S7	<i>K11/LKB1</i> gene muta	ations			
No.	Mutation type	dbSNP RS	Mutation site	Amino acid change	Exon	Variant type
2	Frameshift	rs372511774	c.357delC	p.N119Kfs	2 10	SNV
4	Splice-site variant	rs398123406	c.921-1G>A	/	8 10	SNP
5	Frameshift	rs1060499961	c.131dupA	p.L45Afs	1 10	INS
6	Missense	/	c.869T>C	p.L290P	7 10	SNP
7	Nonsense	/	c.658C>T	p.Q220X	5 10	SNP
8	Frameshift	/	c.548del	p.L183Rfs	4 10	DEL
9	Splice-site variant	rs398123406	c.921-1G>C	/	8 10	SNP
10	Frameshift	/	c.471_472del	p.F157Lfs	4 10	DEL
12	Frameshift	/	c.180del	p.Y60X	1 10	DEL
13	Missense	/	c.869T>A	p.L290H	7 10	SNP
14	Splice-site variant	/	c.598-2A>G	/	5 10	SNP
15	Missense	rs121913315	c.580G>A	p.D194N	4 10	SNP
16	Missense	rs730881978	c.890G>A	p.R297K	7 10	SNP
17	Frameshift	/	c.577_578del	p.S193Rfs	4 10	DEL
18	Splice-site variant	/	c.863-2A>G	/	7 10	SNP
19	Splice-site variant	rs1555735080	c.290+1G>T	/	1 10	SNP
20	Nonsense	/	c.179dup	p.Y60X	1 10	INS
21	Frameshift	rs587782584	c.842dup	p.L282Afs	6 10	INS
23	Frameshift	rs786203886	c.228dup	p.V77Rfs	1 10	INS
24	Nonsense	rs730881970	c.409C>T	p.Q137X	3 10	SNP

DEL; Deletion; INS: Insertion; SNP: Single nucleotide polymorphism; SNV: Single nucleotide variation.

Table 4 Prediction of protein function change caused by STK11/LKB1 mutation

Na	PolyPhen		Mutation Assesso	r	SIFT	SIFT		
No.	Score	Prediction	Score	Prediction	Score	Prediction		
6	1	Probably damaging	0.98351; 4.21	High	0	Deleterious		
13	1	Probably damaging	0.99415; 4.555	High	0	Deleterious		
15	1	Probably damaging	0.98178; 4.165	High	0	Deleterious		
16	1	Probably damaging	0.98818; 4.34	High	0.01	Deleterious		
23	0.022	Benign	0.56769; 1.78	Low	0.26	Tolerated		

STK11/LKB1 genotype-phenotype correlation analysis

Investigation of the relationship between genotype and family history found that the proportion of patients with truncated mutations was slightly higher in those with a family history than in those without a history (60% vs 50%). The proportion of splicesite mutations was lower in those with a family history (20% vs 30%), and the proportion of nonsense mutations was higher in patients with a family history (20.0% vs 11.1%). The proportions of missense mutations were the same (20% vs 20%), and the proportion of frameshift mutations were also equal (40% vs 10%). There were no significant difference between-group differences in $P_{truncation mutation} = 0.653$, $P_{splice site mutation} =$ 0.606, $P_{nonsense mutation} = 0.371$, $P_{missense mutation} = 1.000$, and $P_{frameshift mutation} = 1.000$.

Evaluation of the relationship between genotype and early onset/late onset found that the proportion of truncated mutations in patients with early onset was higher than that in patients with late onset (72.7% vs 33.3%). In patients with early onset, the

Table 5 STK11/LKB1 mutation-related databases and pathogenicity analysis

No.	cDNA/protein –	Disease databas	se		- Dethogonic judgmont
NO.	CDNA/protein	HGMD	ClinVar	OMIM	 Pathogenic judgment
2	p.N119Kfs	/	(1/1) pathogenic	/	Pathogenic
4	c.921-1G>A	\checkmark	/	PJS	Pathogenic
5	p.L45Afs	/	/	/	Pathogenic
6	p.L290P	\checkmark	(1/1) pathogenic	PJS	Clinical significance unknown
7	p.Q220X	/	(3/3) pathogenic	PJS	Pathogenic
8	p.L183Rfs	/	/	PJS	Pathogenic
9	c.921-1G>C	\checkmark	(2/2) pathogenic	PJS	Pathogenic
10	p.F157Lfs	\checkmark	/	PJS	Likely pathogenic
12	p.Y60X	\checkmark	\checkmark	PJS	Pathogenic
13	p.L290H	/	/	PJS	Clinical significance unknown
14	c.598-2A>G	/	(1/1) pathogenic	PJS	Likely pathogenic
15	p.D194N	\checkmark	(4/6) likely pathogenic; (2/6) pathogenic	PJS	Likely pathogenic
16	p.R297K	\checkmark	(1/2) pathogenic; (1/2) unknown	PJS	Likely pathogenic
17	p.S193Rfs	/	/	PJS	Likely pathogenic
18	c.863-2A>G	/	(1/1) pathogenic	PJS	Likely pathogenic
19	c.290+1G>T	Pathogenic	/	PJS	Likely pathogenic
20	p.Y60X	Pathogenic	(2/2) pathogenic	PJS	Pathogenic
21	p.L282Afs	Pathogenic	(1/1) pathogenic	PJS	Pathogenic
23	p.V77Rfs	/	/	PJS	Likely pathogenic
24	p.Q137X	Pathogenic	(1/1) pathogenic	PJS	Pathogenic

(4/6) likely pathogenic: A total of six institutions have judged this mutation, four of which are judged as probably pathogenic, the same below. PJS: Peutz-Jeghers syndrome.

Table 6 Characteristics of SLX4 gene mutations											
No.	Mutation type	dbSNP RS	Mutation site	Amino acid changes	Exon	Variant type					
1	Missense	rs551385115	c.5072A>G	p.N1691S	14 15	SNP					
2	Splice-site variant	/	c.1683+1G>A	splice	7 15	SNP					
3	Missense	rs774243118	c.2990C>T	p.P997L	12 15	SNP					
18	Missense	/	c.2425G>C	p.E809Q	12 15	SNP					
22	Non-frameshift	/	c.568_570del	p.P190del	3 15	DEL					

DEL: Deletion; SNP: Single nucleotide polymorphism.

percentages of frameshift mutations (54.5% vs 22.2%) and sense mutations (18.2% vs 11.1%) were higher than those in late onset patients. The percentages of splice-site mutations (9% vs 44.4%) and missense mutations were lower (18.2% vs 22.2%). There were no significant between-group differences in $P_{truncation mutation} = 0.078$, $P_{frameshift mutation} = 0.142$, $P_{nonsense mutation} = 0.660$, $P_{splice site mutation} = 0.069$, $P_{missense mutation} = 0.822$.

DISCUSSION

The STK11/LKB1 gene located on chromosome 19p13.3 is considered to be a tumor



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Table 7	Table 7 Prediction of protein function change caused by SLX4 mutation										
No.	PolyPhen		Mutation assessor		SIFT	SIFT					
NO.	Score	Prediction	Score	Prediction	Score	Prediction					
1	0	Benign	0.08118; 0	Neutral	0.16	Tolerated					
3	0.004	Benign	0.05510; -0.035	Neutral	1	Tolerated /					
18	0.341	Benign	0.59436; 1.845	Low	0.04	Deleterious					

Table 8 SLX4 mutation-related databases and pathogenicity analysis

No	cDNA/Protein	Diseas	e database		Dethogonic judgment	
No.	CDNA/Protein	HGMD	ClinVar	ОМІМ	 Pathogenic judgment 	
1	p.N1691S	/	(1/1)Uncertain Significance	BTB/POZ domain containing 12\SLX4 structure-specific	Clinical significance unknown	
2	c.1683+1G>A	/	/	BTB/POZ domain containing 12\SLX4 structure-specific	Likely pathogenic	
3	p.P997L	/	/	BTB/POZ domain containing 12\SLX4 structure-specific	Clinical significance unknown	
18	p.E809Q	\checkmark	/	BTB (POZ) domain containing 12\SLX4 structure-specific	Clinical significance unknown	
22	p.P190del	/ /		BTB (POZ) domain containing 12\SLX4 structure-specific	Clinical significance unknown	

suppressor gene^[14] and is widely expressed in human tissues. Pathogenic mutation of STK11 can inactivate its expressed product, which results in the loss of its inhibitory effect on the activity of mammalian target of rapamycin (mTOR), leading to the occurrence of skin and mucous membrane black spots and gastrointestinal polyps^[5]. Methylation of the STK11/LKB1 gene promoter has an important role in the process of malignant transformation of gastrointestinal polyps[15]. At present, the comprehensive mutation rate of STK11/LKB1 gene in PJS patients detected by multiple sequencing methods is about 80%-94% [8,15,16]. The detection rate of STK11/LKB1 gene mutation in PJS patients in this study was 83.3% (20/24), 90% of which are related to pathogenicity. Analysis of the pathogenicity of all the detected mutation sites included in the Mendelian Inheritance in Man (OMIM) database found that about 90% of the STK11/LKB1 mutations were related to PJS. Except for the STK11/LKB1 gene and one case of SLX4 gene mutation, no other gene mutations related to the disease or the possibility of disease were found.

Research on whether there is a correlation between the PJS genotype and clinical phenotype is ongoing. Although the correlation is currently unclear[6,17], some studies have reported positive results. For example, Forcet et al[18] reported that patients often present with only black spots and without gastrointestinal polyps when heterozygous mutations occur in exon 8 of the STK11 gene. Amos et al[19] found that PJS patients with missense mutations had a first episode of polypectomy and appearance of other symptoms significantly later than those with truncated mutations or no detectable mutations. In a study including 116 PJS patients in 52 families, Wang et al[20] found that nearly 30% of the mutations occurred in exon 7, and some of those mutations affected the protein Kinase domain XI region, which is associated with 90% of cases with gastrointestinal polyp dysplasia. An analysis of the start region of the STK11/LKB1 coding sequence by Hearle et al[21] found that a change in promoter sequence was unlikely to be the cause of PJS. In this study the time that dark spots first appeared, which is a relatively objective indicator, was the basis of clinical classification, and was used to determine whether there was a correlation between the appearance of the spots and any of the genotypes. Spots that appear in early childhood will be noticed. On the other hand, unless there are obvious clinical symptoms, it is extremely difficult to know about gastrointestinal polyps that appear in early childhood. Also, PJS is an autosomal dominant genetic disease and does not completely follow Mendelian inheritance[6]. In clinical practice, it is often found that neither parent has a family history but their child has the disease. This is difficult to fully explain if the disease is caused by a single gene. Therefore, whether the patient has a family history was also included in the basis of clinical classification.

This study did not found that patients with different clinical phenotypes (early onset/late onset and with or without a family history) had statistically significant differences in their STK11/LKB1 gene mutations and loci. However, we found that the



Table 9 Other gene mutations and inclusion in relevant database

Ne	Gana	Ture	Mutation	Amino acid	Ever		e database	
NO.	Gene	Туре	site	changes	Exon	HGMD	ClinVar	OMIM
1	BARD1	TSG	c.556A>G	p.S186G	4 11	/	(6/6)Uncertain Significance	/
	EGFR	/	c.61G>A	p.A21T	1 28	/	/	Epidermal growth factor receptor
2	GEN1	/	c.181T>A	p.S61T	3 14	/	/	Gen endonuclease homolog 1
	BRCA1	TSG	c.2387C>T	p.T796I	10 23	/	(8/8)Uncertain Significance	/
4	NTRK1	/	c.1604A>G	p.E535G	13 17	/	/	/
	PDGFRA	/	c.1423G>A	p.E475K	10 23	/	/	/
	TSC2	TSG	c.521C>T	p.S174L	6 42	/	(2/2)Uncertain Significance	/
	MSH6	/	c.1063G>A	p.G355S	4 10		(4/7)Uncertain Significance(3/7)likely benign	/
5	EGFR	/	c.3040G>A	p.D1014N	25 28	/	/	Epidermal growth factor receptor
	MTUS1	TSG	c.2282G>A	p.S761N	3 15	/	/	Mitochondrial tumor suppressor 1
	PTCH1	TSG	c.2222C>T	p.A741V	14 24	/	(3/4)benign, (1/4)likely benign	/
6	SDHA	TSG	c.715A>G	p.I239V	6 15	\checkmark	(2/2)Uncertain significance	/
	MTUS1	TSG	c.1866C>G	p.N622K	2 15	\checkmark	\checkmark	Mitochondrial tumor suppressor 1
7	RECQL4	/	c.1048A>G	p.R350G	5 21	/	(1/1)Uncertain Significance	/
	RECQL4	/	c.236G>A	p.G79E	4 21	/	/	/
8	ATM	TSG	c.6503C>T	p.S2168L	45 63	/	(7/7)Uncertain Significance	Ataxia telangiectasia mutated
10	TSC2	TSG	c.3475C>T	p.R1159W	30 42	/	(2/4)benign, (2/4)likely benign	/
	FANCG	TSG	c.458C>G	p.A153G	4 14	/	(1/1)Uncertain Significance	/
11	SBDS	/	c.98A>G	p.K33R	1 5	/	/	/
12	VHL	TSG	c.134C>T	p.P45L	1 3	/	/	Von Hippel-Lindau syndrome
	FANCA	/	c.3031C>T	p.R1011C	31 43	/	(1/1)likely benign	/
	TP53	TSG	c.620A>G	p.D207G	6 11	\checkmark	/	/
13	FANCA	/	c.2944A>G	p.T982A	30 43	/	(2/2)Uncertain Significance	/
14	PALLD	/	c.1011C>A	p.D337E	3 21	/	/	/
	MLH3	TSG	c.1519A>G	p.M507V	2 13	/	(1/1)Uncertain Significance	Mutl (E. Coli) homolo 3
	SMARCA4	TSG	c.3791C>T	p.T1264M	28 36	/	(3/3)Uncertain Significance	/
	NF1	TSG	c.3940T>C	p.W1314R	29 58	/	(1/1)Uncertain Significance	/
15	PTCH1	TSG	c.2222C>T	p.A741V	14 24	/	(1/1)likely benign	/
	GALNT12	/	c.148C>A	p.P50T	1 10	/	/	/
16	ATR	TSG	c.325C>T	p.R109W	4 47	/	(1/1)Uncertain Significance	Ataxia telangiectasia and Rad3 related
	VEGFA	TSG	c.1039G>A	p.V347I	6 8	/	/	Vascular endothelial growth factor
	DIS3L2	/	c.1642G>A	p.A548T	13 21	/	/	/
17	TSC1	TSG	c.2693C>G	p.T898S	21 23	\checkmark	(3/5)likely benign, (1/5)benign, (1/5)Uncertain significance	/



	BRIP1	/	c.3072del	p.S1025Hfs	20 20	\checkmark	(1/2)likely pathogenic, (1/2)Uncertain significance	/
	WRN	/	c.3778G>A	p.A1260T	32 35	/	(2/2)Uncertain significance	werner syndrome
	RECQL	/	c.166G>A	p.G56R	4 16	/	/	/
19	BARD1	TSG	c.1148T>G	p.M383R	4 11	/	/	/
	USHBP1	/	c.1358C>T	p.P453L	9 13	/	/	/
	APC	TSG	c.2882A>G	p.N961S	16 16	/	(1/1)Uncertain Significance	Adenomatosis polyposis coli
20	DICER1	TSG	c.2113A>G	p.I705V	13 27	/	/	Multinodular goiter
	FANCM	/	c.2762G>A	p.C921Y	14 23	/	/	/
	APC	TSG	c.5257G>C	p.A1753P	16 16	/	(3/3)Uncertain Significance	Adenomatosis polyposis coli
	NSD1	/	c.5493T>G	p.D1831E	16 23	/	/	Sotos syndrome
	SDHA	TSG	c.739A>G	p.I247V	6 15	/	(4/4)Uncertain Significance	/
	MTUS1	TSG	c.908A>G	p.N303S	2 15	/	/	Mitochondrial tumor suppressor 1
22	EXT2	TSG	c.896G>A	p.R299H	5 14	\checkmark	(1/2)likely benign, (1/2)uncategorized	/
	ATM	TSG	c.1555G>A	p.V519I	10 63	\checkmark	(3/3)Uncertain Significance	Ataxia telangiectasia mutated
	BRCA2	TSG	c.1568A>G	p.H523R	10 27	\checkmark	(1/12)benign, (9/12)likely benign, (2/12)Uncertain Significance	Fanconi anemia
	TP53	TSG	c.214C>G	p.P72A	4 11	\checkmark	(5/5)Uncertain Significance	/
23	FLCN	TSG	c.1366G>C	p.D456H	12 14	/	/	
	MSH2	TSG	c.1789G>A	p.D597N	12 16	/	(1/1)Uncertain Significance	Colon cancer, nonpolyposis type 1
	KIT	/	c.2263G>A	p.A755T	16 21	/	(1/2)Uncertain Significance,(1/2)uncategorized	Piebald trait
24	BAP1	TSG	c.1154G>A	p.R385Q	12 17	/	(2/2)Uncertain Significance	/
	TSC2	TSG	c.1609C>T	p.R537C	16 42	\checkmark	(1/5)benign, (2/5)likely benign; (1/5)Uncertain Significance; (1/5)uncategorized	/

HGMD: Human Gene Mutation Database; OMIM: Online Mendelian Inheritance in Man; TSG: Tumor suppressor gene.

most truncation mutations of the STK11/LKB1 gene mostly occurred in exons 1 and 4, most missense mutations occurred in exon 7, and that truncation mutations were significantly more pathogenic than missense mutations. The results indicate that changes in the sites encoding functional proteins in exon regions 1 and 4 may be among the main causes of PJS. Also, the percentage of STK11/LKB1 truncation mutations in patients with early onset PJS was higher than that in patients with late onset PJS, and the between-group difference in the percentage of missense mutations was not significant. Because the evidence of a correlation with missense mutations was not strong, it suggests that early onset PJS is more likely to be caused by pathogenic mutations in STK11/LKB1, while late onset disease is likely to be clinically heterogeneous. The study results also suggest that analysis of the age of appearance of dark spots in a large sample of PJS patients would yield some interesting findings.

For the first time, we detected more concentrated mutations in the SLX4 gene in PJS patients. The SLX4 (FANCP) gene is a tumor suppressor gene located on chromosome 16p13.3[21]. It serves as a key scaffold element for the assembly of multiprotein complexes containing enzymes involved in DNA maintenance and repair[22] and has low to moderate expression in all adult and fetal tissues and specific adult brain regions [23]. It has been reported that [24] truncated mutations in the SLX4 gene were detected in families with Fanconi anemia, and it was determined that SLX4 mutations are clearly related to one of the subtypes of the disease. Fanconi anemia is a rare autosomal recessive genetic disease^[25]. In addition to blood system-related manifestations, the clinical manifestations of FA include multiple congenital malformations, brown pigmentation of the skin, and tumor susceptibility [26]. There are many similarities with PJS, mutations in the SLX4 gene have been detected in patients with PJS in previous studies, the first of which was found in this group. SLX4 is considered



Table 10 Prediction of protein function changes caused by other gene mutations

C	SIFT		PolyPhe	n	Mutation Assess	Mutation Assessor	
Gene	Score	Prediction	Score	Prediction	Score	Prediction	
BARD1	0	Deleterious	0.144	Benign	0.66939; 2.045	Medium	
EGFR	0.4	Tolerated	0.956	Probably damaging	0.33485; 1.01	Low	
GEN1	0	Deleterious	0.999	Probably damaging	0.34521; 1.04	Low	
BRCA1	0.02	Deleterious	0.775	Probably damaging	0.78223; 2.4	Medium	
NTRK1	0.01	Deleterious	0.639	Probably damaging	0.02685; -0.53	Neutral	
PDGFRA	0.1	Tolerated	0.05	Benign	0.38838; 1.175	Low	
TSC2	0.15	Tolerated	0.327	Benign	0.57536; 1.79	Low	
MSH6	0.45	Tolerated	0.176	Benign	0.08118; 0	Neutral	
EGFR	0	Deleterious	0.814	Possibly damaging	0.83953; 2.67	Medium	
MTUS1	0.09	Tolerated	0.044	Benign	0.27053; 0.805	Low	
PTCH1	0	Deleterious	0.7	Possibly damaging	0.88377; 2.95	Medium	
SDHA	0.01	Deleterious low confidence	0.078	Benign	0.49699; 1.58	Low	
MTUS1	0.01	Deleterious	0.096	Benign	0.29908; 0.895	Low	
RECQL4	/	/	/	/	/	/	
RECQL4	/	/	/	/	/	/	
ATM	0	Deleterious	0.294	Benign	0.67953; 2.075	Medium	
TSC2	0.01	Deleterious	0.226	Benign	0.08118; 0	Neutral	
FANCG	0.03	Deleterious	0.018	Benign	0.14661; 0.345	Neutral	
SBDS	0.12	Tolerated	0.051	Benign	0.71920; 2.185	Medium	
/HL	0.06	Tolerated	0.012	Benign	0.19112; 0.55	Neutral	
FANCA	0.24	Tolerated	0	Benign	0.02315; -0.6	Neutral	
TP53	0.03	Deleterious	0.386	Benign	0.45228; 1.405	Low	
FANCA	0.79	Tolerated	0.007	Benign	0.52573; 1.65	Low	
PALLD	0.7	Tolerated	0.159	Benign	0.00602; -1.34	Neutral	
MLH3	0.47	Tolerated	0	Benign	0.55103; 1.725	Low	
SMARCA4	0.05	Deleterious	0.007	Benign	0.29908; 0.895	Low	
NF1	0.62	Tolerated	0.015	Benign	0.08118; 0	Neutral	
PTCH1	0	Deleterious	0.626	Possibly damaging	0.88377; 2.95	Medium	
GALNT12	0.11	Tolerated	0.007	Benign	0.51422; 1.61	Low	
ATR	0	Deleterious	0.998	Probably damaging	0.65975; 2.015	Medium	
/EGFA	0.25	Tolerated low confidence	0.695	Probably damaging	0.08118; 0	Neutral	
DIS3L2	0.05	Tolerated	0.996	Probably damaging	0.87328; 2.875	Medium	
TSC1	/	/		/	0.00621; -1.32	Neutral	
PTCH1	0.03	Deleterious low confidence	0.259	Benign	0.36672; 1.1	Low	
3RIP1	/	/	/	/	/	/	
VRN	0.59	Tolerated	0.164	Benign	0.70595; 2.14	Medium	
RECQL	0.5	Tolerated	0.005	Benign	0.41079; 1.255	Low	
BARD1	0.4	Tolerated	0	Benign	0.08118; 0	Neutral	
JSHBP1	0.05	Tolerated	0.521	Possibly damaging	0.56769; 1.78	Low	
APC	0.16	Tolerated	0.82	Possibly damaging	0.46157; 1.445	Low	

DICER1	0.29	Tolerated	0.664	Possibly damaging	0.34521; 1.04	Low
FANCM	1	Tolerated	0	Benign	0.40543; 1.245	Low
APC	0.57	Tolerated low confidence	0.003	Benign	0.14661; 0.345	Neutral
NSD1	0.03	Deleterious	0.684	Possibly damaging	0.66939; 2.045	Medium
SDHA	0.02	Deleterious low confidence	0.02	Benign	0.20574; 0.59	Neutral
MTUS1	0.87	Tolerated	0	Benign	0.12746; 0.255	Neutral
EXT2	0.03	Deleterious	0.993	Possibly damaging	0.82323; 2.585	Medium
ATM	0.58	Tolerated	0.007	Benign	0.56769; 1.78	Low
BRCA2	0.09	Tolerated	0.003	Benign	0.08118; 0	Neutral
TP53	0.94	Tolerated	0	Benign	0.03608; -0.345	Neutral
FLCN	0.03	Deleterious	0	Benign	0.47716; 1.5	Low
MSH2	0.25	Tolerated	0.023	Benign	0.39692;1.235	Low
KIT	0.15	Tolerated	0.472	Possibly damaging	0.03608; -0.345	Neutral
BAP1	0	Deleterious low confidence	0.968	Possibly damaging	0.59436; 1.845	Low
TSC2	0.02	Deleterious	0.446	Possibly damaging	0.75777; 2.31	Medium

to be an important regulator of DNA repair. Studies have shown that repairing specific types of DNA damage requires *SLX4* and other endonucleases to participate together [22]. At present, it is believed that[27-29] the loss of DNA MMR genes causes the accumulation of mismatches in the process of DNA replication, resulting in the occurrence of microsatellite instability and partial junctions. Colorectal cancer has obvious genetic characteristics. We also detected mutations in some MMR genes (*MSH2* and *MSH6*) in PJS, and the role of *SLX4* gene is highly similar to that. Perhaps the mutation of the *SLX4* gene may explain the genetic heterogeneity of PJS to some extent.

CONCLUSION

In conclusion, we discovered a series of new gene mutation sites, analyzed their pathogenicity, and enriched the mutation spectrum of PJS pathogenic genes. And through the summary of the clinical phenotypes with different *STK11* genotypes, to explore whether they are related, and get some tendentious research results. The detection of *SLX4* gene mutations in patients with PJS was reported for the first time. The relationship between *SLX4* gene mutations and the occurrence of PJS is still unclear, but may help to explain the genetic heterogeneity of PJS.

ARTICLE HIGHLIGHTS

Research background

Different types of pathogenic mutations may produce different clinical phenotypes, but no exact correlation between Peutz-Jeghers syndrome (PJS) genotype and clinical phenotype has been found so far. So it is necessary to study the correlation between genotype and clinical phenotype of PJS, and explore the internal molecular mechanism of different clinical phenotypes.

Research motivation

The authors included 24 cases of treated PJS cases as study participants, collected peripheral venous blood or normal tissue adjacent to polyps for high-throughput next-generation sequencing (NGS) of 139 hereditary colorectal tumor-related genes including *STK11/LKB1* to study the correlation between genotype and clinical phenotype of PJS.

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Research objectives

To investigate the correlation between the genotype and clinical phenotype of PJS.

Research methods

Twenty-four patients with PJS were randomly selected for study inclusion. A total of 139 common hereditary tumor-related genes including STK11/LKB1 were screened and analyzed for pathogenic germline mutations by high-throughput next-generation sequencing (NGS), and the pathogenicity of these mutations was evaluated.

Research results

STK11/LKB1 gene mutations were identified in 20 PJS patients, 90% of which were pathogenic mutations. 10 cases had new mutation sites. Pathogenic mutations were significantly less frequent in exon 7 of the STK11/LKB1 gene than in other exons. Truncation mutations were more common in exons 1 and 4, and their pathogenicity was significantly higher than that of missense mutations. We also identified SLX4 gene mutations in PJS patients.

Research conclusions

PJS has a relatively complicated genetic background. Changes in the sites responsible for coding functional proteins in exon 1 and exon 4 of STK11/LKB1 may be one of the main causes of PJS. Mutation of the SLX4 gene may help to explain the genetic heterogeneity of PJS.

Research perspectives

Exploration of the relationships of clinical phenotypes with different STK11 genotypes, may help to interpret some controversial research results. The detection of SLX4 gene mutations in patients with PJS was reported for the first time.

REFERENCES

- 1 van Lier MG, Wagner A, Mathus-Vliegen EM, Kuipers EJ, Steyerberg EW, van Leerdam ME. High cancer risk in Peutz-Jeghers syndrome: a systematic review and surveillance recommendations. Am J Gastroenterol 2010; 105: 1258-64; author reply 1265 [PMID: 20051941 DOI: 10.1038/ajg.2009.725]
- Hearle N, Schumacher V, Menko FH, Olschwang S, Boardman LA, Gille JJ, Keller JJ, Westerman AM, Scott RJ, Lim W, Trimbath JD, Giardiello FM, Gruber SB, Offerhaus GJ, de Rooij FW, Wilson JH, Hansmann A, Möslein G, Royer-Pokora B, Vogel T, Phillips RK, Spigelman AD, Houlston RS. Frequency and spectrum of cancers in the Peutz-Jeghers syndrome. Clin Cancer Res 2006; 12: 3209-3215 [PMID: 16707622 DOI: 10.1158/1078-0432.CCR-06-0083]
- Lim W, Olschwang S, Keller JJ, Westerman AM, Menko FH, Boardman LA, Scott RJ, Trimbath J, 3 Giardiello FM, Gruber SB, Gille JJ, Offerhaus GJ, de Rooij FW, Wilson JH, Spigelman AD, Phillips RK, Houlston RS. Relative frequency and morphology of cancers in STK11 mutation carriers. Gastroenterology 2004; 126: 1788-1794 [PMID: 15188174 DOI: 10.1053/j.gastro.2004.03.014]
- Hemminki A, Markie D, Tomlinson I, Avizienyte E, Roth S, Loukola A, Bignell G, Warren W, Aminoff M, Höglund P, Järvinen H, Kristo P, Pelin K, Ridanpää M, Salovaara R, Toro T, Bodmer W, Olschwang S, Olsen AS, Stratton MR, de la Chapelle A, Aaltonen LA. A serine/threonine kinase gene defective in Peutz-Jeghers syndrome. Nature 1998; 391: 184-187 [PMID: 9428765 DOI: 10.1038/34432]
- 5 Jia Y, Fu H, Li N, Kang Q, Sheng J. [Diagnosis and treatment for 46 cases of Peutz-Jeghers syndrome]. Zhong Nan Da Xue Xue Bao Yi Xue Ban 2018; 43: 1323-1327 [PMID: 30643048 DOI: 10.11817/j.issn.1672-7347.2018.12.007]
- Beggs AD, Latchford AR, Vasen HF, Moslein G, Alonso A, Aretz S, Bertario L, Blanco I, Bülow S, Burn J, Capella G, Colas C, Friedl W, Møller P, Hes FJ, Järvinen H, Mecklin JP, Nagengast FM, Parc Y, Phillips RK, Hyer W, Ponz de Leon M, Renkonen-Sinisalo L, Sampson JR, Stormorken A, Tejpar S, Thomas HJ, Wijnen JT, Clark SK, Hodgson SV. Peutz-Jeghers syndrome: a systematic review and recommendations for management. Gut 2010; 59: 975-986 [PMID: 20581245 DOI: 10.1136/gut.2009.198499]
- Riegert-Johnson DL, Westra W, Roberts M. High cancer risk and increased mortality in patients with Peutz-Jeghers syndrome. Gut 2012; 61: 322; author reply 322-322; author reply 323 [PMID: 21330574 DOI: 10.1136/gut.2011.238642]
- de Leng WW, Jansen M, Carvalho R, Polak M, Musler AR, Milne AN, Keller JJ, Menko FH, de Rooij FW, Iacobuzio-Donahue CA, Giardiello FM, Weterman MA, Offerhaus GJ. Genetic defects underlying Peutz-Jeghers syndrome (PJS) and exclusion of the polarity-associated MARK/Par1 gene family as potential PJS candidates. Clin Genet 2007; 72: 568-573 [PMID: 17924967 DOI: 10.1111/j.1399-0004.2007.00907.x
- 9 Williams CD, Grady WM, Zullig LL. Use of NCCN Guidelines, Other Guidelines, and Biomarkers



for Colorectal Cancer Screening. J Natl Compr Canc Netw 2016; 14: 1479-1485 [PMID: 27799515 DOI: 10.6004/inccn.2016.0154]

- 10 Kumar P, Henikoff S, Ng PC. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. Nat Protoc 2009; 4: 1073-1081 [PMID: 19561590 DOI: 10.1038/nprot.2009.86]
- Adzhubei IA, Schmidt S, Peshkin L, Ramensky VE, Gerasimova A, Bork P, Kondrashov AS, 11 Sunyaev SR. A method and server for predicting damaging missense mutations. Nat Methods 2010; 7: 248-249 [PMID: 20354512 DOI: 10.1038/nmeth0410-248]
- 12 Thompson BA, Spurdle AB, Plazzer JP, Greenblatt MS, Akagi K, Al-Mulla F, Bapat B, Bernstein I, Capellá G, den Dunnen JT, du Sart D, Fabre A, Farrell MP, Farrington SM, Frayling IM, Frebourg T, Goldgar DE, Heinen CD, Holinski-Feder E, Kohonen-Corish M, Robinson KL, Leung SY, Martins A, Moller P, Morak M, Nystrom M, Peltomaki P, Pineda M, Qi M, Ramesar R, Rasmussen LJ, Royer-Pokora B, Scott RJ, Sijmons R, Tavtigian SV, Tops CM, Weber T, Wijnen J, Woods MO, Macrae F, Genuardi M. Application of a 5-tiered scheme for standardized classification of 2,360 unique mismatch repair gene variants in the InSiGHT locus-specific database. Nat Genet 2014; 46: 107-115 [PMID: 24362816 DOI: 10.1038/ng.2854]
- MacArthur DG, Manolio TA, Dimmock DP, Rehm HL, Shendure J, Abecasis GR, Adams DR, 13 Altman RB, Antonarakis SE, Ashley EA, Barrett JC, Biesecker LG, Conrad DF, Cooper GM, Cox NJ, Daly MJ, Gerstein MB, Goldstein DB, Hirschhorn JN, Leal SM, Pennacchio LA, Stamatoyannopoulos JA, Sunyaev SR, Valle D, Voight BF, Winckler W, Gunter C. Guidelines for investigating causality of sequence variants in human disease. Nature 2014; 508: 469-476 [PMID: 24759409 DOI: 10.1038/nature13127]
- Yoo LI, Chung DC, Yuan J. LKB1--a master tumour suppressor of the small intestine and beyond. 14 Nat Rev Cancer 2002; 2: 529-535 [PMID: 12094239 DOI: 10.1038/nrc843]
- 15 Chen C, Zhang X, Wang D, Wang F, Pan J, Wang Z, Liu C, Wu L, Lu H, Li N, Wei J, Shi H, Wan H, Zhu M, Chen S, Zhou Y, Zhou X, Yang L, Liu J. Genetic Screening and Analysis of LKB1 Gene in Chinese Patients with Peutz-Jeghers Syndrome. Med Sci Monit 2016; 22: 3628-3640 [PMID: 27721366 DOI: 10.12659/msm.897498]
- Aretz S, Stienen D, Uhlhaas S, Loff S, Back W, Pagenstecher C, McLeod DR, Graham GE, Mangold 16 E, Santer R, Propping P, Friedl W. High proportion of large genomic STK11 deletions in Peutz-Jeghers syndrome. Hum Mutat 2005; 26: 513-519 [PMID: 16287113 DOI: 10.1002/humu.20253]
- 17 Forcet C, Etienne-Manneville S, Gaude H, Fournier L, Debilly S, Salmi M, Baas A, Olschwang S, Clevers H, Billaud M. Functional analysis of Peutz-Jeghers mutations reveals that the LKB1 Cterminal region exerts a crucial role in regulating both the AMPK pathway and the cell polarity. Hum Mol Genet 2005; 14: 1283-1292 [PMID: 15800014 DOI: 10.1093/hmg/ddi139]
- Amos CI, Keitheri-Cheteri MB, Sabripour M, Wei C, McGarrity TJ, Seldin MF, Nations L, Lynch 18 PM, Fidder HH, Friedman E, Frazier ML. Genotype-phenotype correlations in Peutz-Jeghers syndrome. J Med Genet 2004; 41: 327-333 [PMID: 15121768 DOI: 10.1136/jmg.2003.010900]
- Wang Z, Wu B, Mosig RA, Chen Y, Ye F, Zhang Y, Gong W, Gong L, Huang F, Wang X, Nie B, 19 Zheng H, Cui M, Wang Y, Wang J, Chen C, Polydorides AD, Zhang DY, Martignetti JA, Jiang B. STK11 domain XI mutations: candidate genetic drivers leading to the development of dysplastic polyps in Peutz-Jeghers syndrome. Hum Mutat 2014; 35: 851-858 [PMID: 24652667 DOI: 10.1002/humu.22549
- Hearle NC, Tomlinson I, Lim W, Murday V, Swarbrick E, Lim G, Phillips R, Lee P, O'Donohue J, 20 Trembath RC, Morrison PJ, Norman A, Taylor R, Hodgson S, Lucassen A, Houlston RS. Sequence changes in predicted promoter elements of STK11/LKB1 are unlikely to contribute to Peutz-Jeghers syndrome. BMC Genomics 2005; 6: 38 [PMID: 15774015 DOI: 10.1186/1471-2164-6-38]
- 21 Fekairi S, Scaglione S, Chahwan C, Taylor ER, Tissier A, Coulon S, Dong MQ, Ruse C, Yates JR 3rd, Russell P, Fuchs RP, McGowan CH, Gaillard PHL. Human SLX4 is a Holliday junction resolvase subunit that binds multiple DNA repair/recombination endonucleases. Cell 2009; 138: 78-89 [PMID: 19596236 DOI: 10.1016/j.cell.2009.06.029]
- 22 Svendsen JM, Smogorzewska A, Sowa ME, O'Connell BC, Gygi SP, Elledge SJ, Harper JW. Mammalian BTBD12/SLX4 assembles a Holliday junction resolvase and is required for DNA repair. Cell 2009; 138: 63-77 [PMID: 19596235 DOI: 10.1016/j.cell.2009.06.030]
- Nagase T, Kikuno R, Ohara O. Prediction of the coding sequences of unidentified human genes. 23 XXII. The complete sequences of 50 new cDNA clones which code for large proteins. DNA Res 2001; 8: 319-327 [PMID: 11853319 DOI: 10.1093/dnares/8.6.319]
- Stoepker C, Hain K, Schuster B, Hilhorst-Hofstee Y, Rooimans MA, Steltenpool J, Oostra AB, 24 Eirich K, Korthof ET, Nieuwint AW, Jaspers NG, Bettecken T, Joenje H, Schindler D, Rouse J, de Winter JP. SLX4, a coordinator of structure-specific endonucleases, is mutated in a new Fanconi anemia subtype. Nat Genet 2011; 43: 138-141 [PMID: 21240277 DOI: 10.1038/ng.751]
- 25 Jacquemont C, Taniguchi T. The Fanconi anemia pathway and ubiquitin. BMC Biochem 2007; 8 Suppl 1: S10 [PMID: 18047734 DOI: 10.1186/1471-2091-8-S1-S10]
- Kutler DI, Singh B, Satagopan J, Batish SD, Berwick M, Giampietro PF, Hanenberg H, Auerbach 26 AD. A 20-year perspective on the International Fanconi Anemia Registry (IFAR). Blood 2003; 101: 1249-1256 [PMID: 12393516 DOI: 10.1182/blood-2002-07-2170]
- Picard E, Verschoor CP, Ma GW, Pawelec G. Relationships Between Immune Landscapes, Genetic 27 Subtypes and Responses to Immunotherapy in Colorectal Cancer. Front Immunol 2020; 11: 369 [PMID: 32210966 DOI: 10.3389/fimmu.2020.00369]



- 28 Bourhis A, De Luca C, Cariou M, Vigliar E, Barel F, Conticelli F, Marcorelles P, Nousbaum JB, Robaszkiewicz M, Samaison L, Badic B, Doucet L, Troncone G, Uguen A. Evaluation of KRAS, NRAS and BRAF mutational status and microsatellite instability in early colorectal carcinomas invading the submucosa (pT1): towards an in-house molecular prognostication for pathologists? J Clin Pathol 2020; 73: 741-747 [PMID: 32273401 DOI: 10.1136/jclinpath-2020-206496]
- Vageli DP, Doukas SG, Markou A. Mismatch DNA repair mRNA expression profiles in oral melanin 29 pigmentation lesion and hamartomatous polyp of a child with Peutz-Jeghers syndrome. Pediatr Blood Cancer 2013; 60: E116-E117 [PMID: 23677888 DOI: 10.1002/pbc.24579]



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ORIGINAL ARTICLE

Clinical and Translational Research

Validation of the Italian translation of the perceived stigma scale and resilience assessment in inflammatory bowel disease patients

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Abstract

BACKGROUND

Stigmatization is the separation of an individual from a group due to aspects that make them different. Resilience may in turn influence the perception of stigma. Patients with inflammatory bowel disease (IBD) are susceptible to stigma, although data are very limited.

AIM

To validate an Italian translation of the IBD perceived stigma scale (PSS) in relation to patients' resilience.

METHODS

Consecutive IBD outpatients were prospectively enrolled (December 2018-September 2019) in an Italian, tertiary referral, IBD center. Clinical and demographic data were collected. Stigma and resilience were evaluated through the IBD-PSS and the 25-item Connor-Davidson Resilience Scale, respectively. The International Quality of Life Assessment Project approach was followed to translate the IBD-PSS into Italian and to establish data quality. Higher scores represent greater perceived stigma and resilience. Multivariable analysis for factors associated with greater stigma was computed.

Institutional review board

RESULTS



statement: The study was approved by the local Ethics Committee (Protocol Number 20190003611). The study protocol conforms to the ethical guidelines of the 1975 Declaration of Helsinki as reflected in a priori approval by the institution's human research committee.

Informed consent statement: All

participants gave their informed written consent to take part to the study and for the anonymized publication of data.

Conflict-of-interest statement: The authors of this manuscript having no conflicts of interest to disclose.

Data sharing statement: There is no additional data available.

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Overall, 126 IBD patients (mean age 46.1 ± 16.9) were enrolled. The International Quality of Life Assessment criteria for acceptable psychometric properties of the scale were satisfied, with optimal data completeness. There was no ceiling effect, whilst floor effect was present (7.1%). The discriminant validity and the internal consistency reliability were good (Cronbach alpha = 0.87). The overall internal consistency was 95%, and the test-retest reliability was excellent 0.996. The median PSS score was 0.45 (0.20-0.85). Resilience negatively correlated with perceived stigma (Spearman's correlation = -0.18, 95% confidence intervals: -0.42-0.08, P = 0.03).

CONCLUSION

We herein validated the Italian translation of the PSS scale, also demonstrating that resilience negatively impacts perceived stigma.

Key Words: Crohn's disease; Quality of life; Stress; Ulcerative colitis

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Core Tip: We have here validated an Italian version of the Perceived Stigma Scale for patients with inflammatory bowel disease. We have also found that resilience levels negatively correlated with perceived stigma. This is the first study assessing this issue in patients with inflammatory bowel disease.

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INTRODUCTION

Stigmatization is defined as the societal identification of an individual as abnormal and worthy of separation from the group, leading to discrimination and loss of their social status^[1]. It has been reported that inflammatory bowel disease (IBD) is susceptible to stigmatization, not only because of the *taboo* around its symptoms, but also due to the assumption of being a psychosomatic condition affecting people because of their "obsessive behavior" [2] and because it affects sexual life[3]. Stigmatization in IBD patients was reported to be as high as 84%, regardless of disease activity **[4**].

An important aim of taking care of chronic patients should be the improvement of their quality of life (QoL), taking into account the social context and their needs[5]. Nonetheless, it emerged from a recent review that the burden of stigmatization in IBD, and the ability to positively cope with the disease (*i.e.* resilience), are not adequately addressed by clinicians^[6]. In IBD patients, resilience has been found to be influenced by individual characteristics, including age, sex, and employment status and to influence positively the disease prognosis [7-9]. Stigma can be evaluated through the use of different scales, including the IBD perceived stigma scale (PSS)[10], which has been adapted and used in IBD patients[11]. Similarly, resilience can be measured through the Connor-Davidson resilience scale (CD-RISC), a 25-item self-administrated scale exploring different aspects related to the individual ability to cope with adversity and stress[12]. The CD-RISC was initially designed for psychiatric American patients, and it has now been translated into more than 70 languages, being the most widely used resilience scale in a variety of conditions[13].

The first study looking into perceived stigma in IBD showed that functional impairment was mainly due to IBD patients' psychological dimension rather than to their physical one. They also reported that patients with Crohn's disease (CD) had a higher degree of perceived stigma than patients with ulcerative colitis (UC)[14]. While perceived stigma is difficult to address and modify, resilience is responsive to



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behavioral intervention and is independently associated with better QoL and lower disease activity in IBD[9].

There are very limited data regarding perceived stigma in IBD, and no validated translation of the PSS into Italian is available. As a consequence, perceived stigma in Italian IBD patients has never been assessed. Therefore, we aimed to validate the Italian version of the PSS in IBD patients in order to obtain a meaningful instrument for assessing stigmatization and compare with international studies. We also assessed resilience and its relation with stigmatization.

MATERIALS AND METHODS

Study population

All IBD patients followed-up at the IBD Clinical & Research Centre of the San Matteo Hospital Foundation were consecutively enrolled between December 2018 and September 2019. IBD diagnosis was established according to internationally agreed criteria^[15]. Patients were eligible for inclusion if they had at least a 3-mo history of IBD, were aged \geq 18, were able to complete a questionnaire, and were willing to give written informed consent. Patients with an inconclusive or uncertain diagnosis of IBD or those diagnosed less than 3 mo before or unwilling to provide informed consent were excluded. Demographic and clinical characteristics were gathered, including IBD type, disease activity and duration, comorbidities, and previous IBD-related surgery. Clinical activity was assessed using the Harvey-Bradshaw index (HBI)[16,17] for CD and the partial Mayo score (pMayo)[18] for UC. For CD patients, HBI < 5 was defined as clinical remission, HBI 5-7 as mild disease, HBI 8-16 as moderate disease, and HBI > 16 as severe disease [16,17]. For UC patients, pMayo < 2 was defined as remission, pMayo 2-4 was defined as mild activity, pMayo 5-7 was defined as moderate activity, and pMayo > 7 was defined as severe activity [18]. The study was approved by the local Ethics Committee (Protocol Number 20190003611), and all participants gave their informed written consent to take part to the study and for the anonymized publication of data. The study protocol conforms to the ethical guidelines of the 1975 Declaration of Helsinki as reflected in a priori approval by the institution's human research committee.

Assessment of the PSS

The PSS was initially designed to assess stigma in irritable bowel syndrome (IBS) patients and was validated in IBD in 2009 in a cohort of patients from the United States [4]. The PSS is a self-administered questionnaire designed to measure perceived stigma through 10 items on a five-point Likert scale (ranging from 0 = never to 4 = always), with a higher score reflecting a greater level of perceived stigma. Each item is assessed on two different domains: Significant others (SO) and healthcare professionals (HP), leading to a total of 20 items. The two-domain PSS version has already been used for different gastrointestinal disease and was found to have an excellent internal consistency and split-half reliability (≥ 0.89) [19].

The score indicating the perceived stigma ranges from 0 to 4 and is obtained by calculating the mean of all the values and the values within each domain.

Translation and cultural validation

We aimed to create a version that was easy to understand and complete by Italian IBD patients, without losing the original English version's equivalence and psychometric validity. The translation and adaptation were made in accordance with the International QoL Assessment (IQOLA) Project approach, which consists of three steps: A forward translation, a backward translation, and a cognitive testing [20,21].

Step 1-Forward translation: Two bilingual physicians (Lenti MV and Cococcia S) blindly translated the questionnaire from English into Italian. The two versions were compared, and discrepancies reconciled. Difficulty and degree of agreement of the translation were rated on a 1 to 100 scale (lowest-highest). For each item, the agreed forward translation was accepted if the scores were \geq 75, otherwise retranslation was independently performed and scoring repeated.

Step 2 - Backward translation: The Italian translation was blindly translated back into British English by two mother-tongue English people with a high educational level (graduated). The same reconciliation process reported for the forward translation was applied. The equivalence of the agreed backward translation to the original version



was rated and expected to be \geq 75. If that threshold was not reached, the four translators had to agree on a new forward translation.

Step 3-Cognitive testing: Cognitive testing of the agreed Italian version was performed on 10 individuals with different age, sex, and educational background to verify that the translation was clear and understandable by a range of different people. Finally, a panel discussion was held to approve the final version (see Supplementary data).

Resilience assessment

Resilience was assessed through the Italian validated translation of the CD-RISC scale, a self-administered questionnaire assessing resilience through 25 items on a five-point Likert scale (ranging from 0 = totally disagree to 4 = totally agree)[12]. The score is calculated by summing the score of each item (ranging from 0 to 100) with higher scores meaning higher resilience.

Statistical analysis and psychometric evaluation

The sample size was computed based on the primary endpoint. A sample of 100 subjects responding to 20 items would achieve 80% power to detect the difference between the coefficient alpha under the null hypothesis of 0.70 and the coefficient alpha under the alternative hypothesis of 0.81, using a two-sided F-test with a significance level of 0.05. Twenty-six extra patients were enrolled to account for possible dropouts.

The PSS scoring was performed according to the scoring manuals, meaning that higher scores represent a higher level of perceived stigma. The psychometric evaluation of the Italian version of the PSS questionnaire included evaluation of data quality, including completeness (Table 1). Results were described as mean and standard deviation, and ceiling and floor effect were evaluated. Reliability was assessed and expressed by means of Cronbach's alpha for internal consistency, alpha = $k \times r/[1+(k-1) \times r]$; with k = number of items and r = mean correlation. Item internal consistency (correlation of item and corresponding scale, corrected for overlap), equality of item-scale correlations, and item discriminant validity (correlation of item with the corresponding scale vs correlation of item with other scales) were evaluated through the multi-trait/multi-item correlation matrix. Means of Pearson's correlation coefficient and intraclass correlation coefficient were used to evaluate the test-retest correlation for temporal stability (within 1 mo), with 95% confidence intervals (95% CI). External validity was assessed through the comparison of PSS scores in patients with different characteristics by means of the Kruskall Wallis test, the test for trend; the correlation with continuous variables was assessed with the Spearman R. Stata 16 (StataCorp, College Station, TX, United States) was used for all computations. All tests were two-sided, and a *P* value < 0.05 was considered statistically significant. In the presence of missing data, missing items were replaced by the median of the corresponding scale, unless more than 50% of the items were missing, in which instance the questionnaire was dropped.

RESULTS

Demographic and clinical characteristics

Overall, 146 IBD patients were screened for inclusion in the study. Of these, 20 patients did not participate because denied consent (15 patients) or because were due to be followed up in another hospital. Hence, 126 IBD patients (mean age 46.1 ± 16.9, male 56.4%), 57 with CD and 69 with UC, were consecutively enrolled in the study. The demographic and clinical characteristics of the enrolled patients are reported in Table 2. CD patients were significantly younger than UC patients ($42.3 \pm 15.7 vs 49.3 \pm$ 17.4; P = 0.03). Psychiatric disorders, including anxiety and depression, were the most common concomitant diseases (25.4%), followed by hypertension (22.2%) and cardiomyopathy (11.9%), which was significantly more common among UC patients (17.4% vs 5.26%; P = 0.05). Overall, the median disease duration was 8 years [interquartile range (IQR) 3-16]. The majority of the CD patients had a disease with an inflammatory behavior (56.1%), 43.9% had a structuring behavior, and 28.1% had a penetrating disease. Almost half of the CD patients (49.1%) had ileo-colonic involvement, and 33.3% had perianal disease. Among UC patients, half (52.2%) had an extensive disease, 37.7% had a left UC, 10.1% had an ulcerative proctitis, and 4.4% had



Table 1 Thresholds defining acceptable psychometric properties according to the International Quality of Life Assessment project						
	Definition	Threshold				
Data quality						
Missing items	Unanswered items	< 5%-10%				
	Incomplete scales (< 50% of items answered)	< 5%-10%				
Floor and ceiling effect	Extreme scores (either on the lower- or higher-end)	< 10%				
Scaling assumption						
Internal consistency						
Item	Correlation among items of the same scale (Pearson correlation ≥ 0.4)	> 90%				
Reliability	Overall consistency of the scale (Cronbach's alpha coefficient)	> 0.7				
Discriminant validity	Items whose Pearson correlation with other scales is higher than with their scale	0%				
Test-retest evaluation	Correlation between the results scales filled in twice by the same patients at defined time points (Pearson correlation)	> 0.7				

a pouch. The proportion of patients with severe disease activity was higher among UC patients (10.1% vs 0%), while two-thirds were in remission in both groups. A quarter of the enrolled patients had an extraintestinal manifestation (28.6%), including anemia, arthritis, uveitis, and dermatological manifestations. When available, calprotectin and C-reactive protein (CRP) were used as inflammatory markers. Overall, 34.1% of the patients had a calprotectin < 50 mg/kg, 16.2% between 51-250 mg/kg, and 12.7% > 250 mg/kg with no difference according to the disease (P = 0.54). Similarly, 61.1% of the patients had a normal CRP, whilst roughly a third had raised levels of CRP (31.0%), with no difference between UC and CD patients (P = 1.00).

Translation and cultural validation

The PSS was translated according to the IQOLA project guidelines[20,21]. For the forward translation, the median difficulty was rated as 10 (range: 10-60) and the agreement was found to be 95 (range: 70-100). Items 1, 5, and 10 were adjusted after discussion. The backward translation equivalence was rated at 95 (range: 80-100). Minor changes were made to item 2 and 4 of the Italian translation to improve the original version's equivalence. A cognitive testing of the agreed Italian version was performed on 10 individuals with different ages (median 48-years-old, range: 29-88), sex (5 female), and educational background (5 graduated), which did not lead to any adjustment of the scale. Supplementary data show the validated Italian version of the PSS-IBD, while the questions of the original English version have already been published elsewhere[4].

Psychometric evaluation

The majority of the IQOLA criteria for acceptable psychometric properties of the scale were satisfied in our cohort as reported in Table 3. We reached an optimal data completeness, and we did not have any ceiling effect, whilst a floor effect was present in 7.1% of the cases (overall domain). The floor effect was greater for the HP domain when compared to the SO domain (42.1% vs 8.7%). When looking at scaling assumption, the internal consistency reliability of the Italian version of the PSS was good, with an overall Cronbach alpha coefficient of 0.87 (0.83 for SO and 0.81 for HP). Although an excellent item, internal consistency was found in each domain, with a Pearson correlation ranging from 0.4 to 0.6, and one item (item 8, SO domain) did not reach the predetermined threshold of 0.4, determining an overall item internal consistency of 95% (still indicative of an excellent item internal consistency). The discriminant validity of the scale was good for items 1 to 7 in both domains, whereas items 8 to 10 had exactly the same Pearson correlation with their domain and the other one for both SO and HP.

The test-retest reliability was excellent, being 0.999 (0.997-1.000) overall, 0.99 (0.997-1.000) in the SO domain and 0.994 (0.979-0.998) in the HP domain. The median PSS score was 0.45 (0.20-0.85) with a significantly higher score for the SO domain (0.70 IQR 0.40-1.40 vs 0.10 IQR 0.00-0.40, P < 0.001), whilst the median resilience score was 64 (IQR 53-78). The level of perceived stigma did not differ according to sex (P = 0.51), IBD type (P = 0.33), disease activity, age (P = 0.11), or disease duration (P = 0.49) (See

Table 2 Demographic and clinica				
	Overall (<i>n</i> = 126)	CD (<i>n</i> = 57)	UC (<i>n</i> = 69)	P value
Age (mean ± SD)	46.13 (± 16.95)	42.29 (± 15.7)	49.29 (± 17.38)	0.03
Male	71 (56.4%)	33 (57.9%)	38 (55.1%)	0.86
3MI	23.9 (± 4.1)	23.9 (± 4.3)	23.9 (± 3.9)	0.67
Disease duration (median, IQR)	8 (3-16)	10 (3-17)	8 (3.5-13)	0.44
Disease characteristics (CD)				
ocation (CD)	-		-	
Terminal ileum (L1)		13 (22.8%)		
Colon (L2)		8 (14.0%)		
Ileo-colon (L3)		28 (49.1%)		
Upper GI (L4)		2 (3.5%)		
Perianal disease (p)		19 (33.3%)		
ehavior (CD)	-	-		
Inflammatory (B1)		32 (56.1%)		
Stricturing (B2)		25 (43.9%)		
Penetrating (B3)		16 (28.1%)		
isease activity (HBI)	-	-		
5		38 (66.7%)		
7		14 (24.6%)		
16		5 (8.8%)		
16	-	0 (0%)		
sease characteristics (UC)			-	
cation	-	-	7 (10.1%)	
Proctitis (E1)			26 (37.7%)	
Left sided (E2)			36 (52.2%)	
Extensive (E3)				
sease activity (pMayo)	-			
2			45 (65.2%)	
4			13 (18.8%)	
7			4 (5.8%)	
7			7 (10.1%)	
buch			3 (4.4%)	
traintestinal manifestations	36 (28.6%)	17 (29.8%)	19 (27.5%)	0.84
revious abdominal surgery	38 (30.2%)	22 (38.6%)	16 (42.1%)	0.07
alprotectin				0.54
50	43(34.1%)	15 (26.3%)	28 (40.6%)	
-250	33 (16.2%)	17 (29.8%)	16 (23.2.%)	
250	16 (12.7%)	7 (12.3%)	9 (13.4%)	
fissing	34 (27.0%)	18 (31.6%)	16 (23.2%)	
RP				1.00
Iormal	77 (61.1)	35 (62.4%)	42 (60.9%)	
aised	39 (31.0%)	18 (31.6%)	21 (30.4%)	



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Missing	10 (7.9%)	4 (7.0%)	6 (8.7%)	
Comorbidities	40 (31.8%)	16 (28.1%)	24 (34.8%)	0.45
Cardiomyopathy	15 (11.9%)	3 (5.26%)	12 (17.4%)	0.05
Hypertension	28 (22.2%)	12 (42.9%)	16 (23.2%)	0.83
Diabetes	11 (8.73%)	4 (7.0%)	7 (10.1%)	0.75
Hepatic failure	1 (0.8%)	1 (1.8%)	0 (0.0%)	0.45
Kidney failure	4 (3.17%)	3 (5.26%)	1 (1.45%)	0.32
Respiratory failure	2 (1.6%)	1 (1.8%)	1 (1.5%)	1.00
Neurologic diseases	5 (4.0%)	1 (1.8%)	4 (5.8%)	0.37
Psychiatric disorder	15 (11.9%)	7 (12.3%)	8 (11.6%)	1.00
Onco-hematological diseases	12 (9.5%)	2 (3.5%)	10 (14.5%)	0.06

BMI: Body mass index; CD: Crohn's disease; CRP: C-reactive protein; GI: Gastrointestinal; HBI: Harvey-Bradshaw Index; IQR: Interquartile range; PMS: Partial Mayo Score; SD: Standard deviation; UC: Ulcerative colitis.

Table 3 Psychometric characteristics of perceived stigma scale and its sub-scales

	Overall	Significant others	Healthcare professionals
Median score	0.45 (0.20-0.85)	0.70 (0.30-1.40)	0.10 (0-0.40)
Data quality			
Missing items	0 (0%)	0 (0%)	0 (0%)
Floor effect	9 (7.1%)	11 (8.7%)	53 (42.1%)
Ceiling effect	0 (0%)	0 (0%)	0 (0%)
Scaling assumption			
Internal consistency			
Item	19/20 (95.0%)	10/10 (100%)	10/10 (100%)
Reliability (Cronbach alpha)	0.87	0.83	0.81
Discriminant validity	-	30.0%	30.0%
Test-retest	0.99 (0.99-1.00)	0.999 (0.99-1.00)	0.99 (0.97-0.99)
Evaluation ^a			

^aIntraclass correlation coefficient (95% confidence interval).

Table 4). On the contrary, disease activity was found to significantly reduce resilience (Spearman's correlation -0.18, 95% CI: -0.42-0.08, P = 0.03) in CD patients, whilst no significant difference was found in UC patients according to the disease activity (P =0.23). When exploring the relations between perceived stigma and resilience, a significant negative Spearman's correlation was found (-0.20, 95%CI: -0.36 to -0.02; P =0.03).

DISCUSSION

Stigmatization is an important, though often unattended, issue in clinical medicine. Whilst for other conditions (e.g., human immunodeficiency virus, mental illness, and lung cancer) stigmatization has been widely studied[22-25], IBD data are scant and fragmentary. This might be partly due to the lack of a validated tool to be used for this purpose. We herein validated an Italian version of the PSS questionnaire that performs well, has good psychometric properties, and is easily understandable. The psychometric evaluation of the Italian PSS version showed an excellent Cronbach alpha coefficient, item internal consistency, and test-retest reliability. Our results are in line

Table 4 Correlation between inflammatory bowel disease perceived stigma scale scores and demographic or clinical characteristics									cal characteristics
	PSS	P	Spearman's correlation (95%Cl)	PSS SO	P	Spearman's correlation (95%Cl)	PSS HP	P	Spearman's correlation (95%CI)
Median score (IQR)	0.45 (0.20 to 0.85)			0.70 (0.30 to 1.40)			0.10 (0.00 to 0.40)		
Age		0.11	-0.14 (-0.31 to 0.03)		0.08	-0.157 (-0.33 to 0.02)		0.20	-0.116 (-0.29 to 0.06)
Sex		0.51			0.26			0.29	
Female	0.45 (0.30 to 0.85)			0.70 (0.50 to 1.40)			0.10 (0.00 to 0.40)		
Male	0.45 (0.15 to 0.90)			0.70 (0.20 to 1.40)			0.10 (0.00 to 0.50)		
Diagnosis		0.33			0.35			0.34	
CD	0.55 (0.25 to 0.95)			0.80 (0.40 to 1.40)			0.10 (0.00 to 0.50)		
UC	0.45 (0.20 to 0.85)			0.70 (0.30 to 1.30)			0.10 (0.00 to 0.40)		
HBI		0.91	0.05 (-0.22 to 0.30)		0.91	0.05 (-0.21 to 0.31)		0.70	0.11 (-0.16 to 0.36)
< 5	0.53 (0.25 to 0.80)			0.75 (0.50 to 1.30)			0.10 (0.00 to 0.50)		
5-7	0.60 (0.10 to 1.15)			0.90 (0.20 to 1.60)			0.30 (0.00 to 0.50)		
8-16	0.40 (0.10 to 1.40)			0.70 (0.20 to 1.80)			0.10 (0.00 to 0.80)		
pMS		0.52	0.06 (-0.18 to 0.29)		0.44	0.03 (-0.21 to 0.26)		0.81	0.11 (-0.13 to 0.34)
< 2	0.40 (0.20 to 0.85)			0.60 (0.40 to 1.30)			0.10 (0.00 to 0.30)		
2-4	0.45 (0.05 to 0.75)			0.60 (0.10 to 1.20)			0.20 (0.00 to 0.40)		
5-7	0.75 (0.48 to 1.22)			1.40 (0.90 to 1.75)			0.15 (0.05 to 0.70)		
>7	0.35 (0.15 to 1.50)			0.70 (0.00 to 2.70)			0.10 (0.00 to 0.70		
CD-RISC25	-	0.03	-0.20 (-0.36 to -0.02)	-	0.02	-0.20 (-0.36 to -0.03)	-	0.12	-0.14 (-0.31 to 0.04)

CD: Crohn's disease; CD-RISC25: 25-item Connor-Davidson Resilience Scale; CI: Confidence interval; HBI: Harvey-Bradshaw Index; HP: Healthcare professionals; IQR: Interquartile range; pMS: Partial Mayo Score; SO: Significant others; UC: Ulcerative colitis.

> with previous literature validating stigma scales in different settings[4,10,26,27], showing that our translation is reliable and offers a tool to assess stigma in Italian IBD patients. In our cohort, the main concern is the high floor effect recorded, especially in the HP domain. However, this result was partially expected since all the included patients were followed-up at a tertiary IBD center. These patients likely experience lower levels of perceived stigma since they are looked after by IBD dedicated HP, whereas a different result might be obtained if the questionnaire would be administered in different settings, such as community centers or private practices. Even considering the setting bias, the level of perceived stigma was found to be lower than expected (median 0.45, IQR 0.20-0.85), when compared to previous literature showing a low-to-moderate level of perceived stigma among IBD patients, using the PSS[4]. This might be explained by the fact that the PSS has been originally designed to address perceived stigma in patients affected by a functional disorder rather than by an organic disease, such as IBD. In the PSS questionnaire, there are no questions addressing some of IBD patients' main concerns, including the fear of relapsing or of incontinence. Therefore, even if this scale offers a useful tool to assess stigma in IBD, it is our opinion that some adjustments are needed.

> Additionally, levels of perceived stigma in IBD patients tend to decrease in longstanding disease, in contrast with what happens for IBS[11]. Since IBD is an organic disease not associated with unhealthy or socially unacceptable vices, it is therefore



more likely to be recognized and accepted as a "real" organic disease over time, especially when compared to IBS[11]. Most of the included patients have a longstanding history, which could have led to lower levels of perceived stigma in our sample.

Stigmatization is multifactorial and is influenced by both patient-dependent and environment-dependent factors [28-30]. Among these factors, we have previously speculated that a relation between stigma and resilience may exist[6]. In line with previous literature, we found a moderate level of resilience in our cohort[9]. We have here shown for the first time that, in IBD patients, higher levels of resilience correlate with lower levels of perceived stigma (overall and for SO) and, conceivably, to better QoL. Such correlation was not found for the HP domain, which might be due to the high floor effect reported in this domain. In case of adversity, resilience can modulate catecholamine and cortisol production reducing long-term effects on the body[31] and leading to better outcome also in IBD, which requires continuous adaptation to the unpredictable course of the disease. This hypothesis is supported by a recent study in which higher levels of resilience are associated with lower disease activity, although it is unclear if this result was due to reverse causation[9]. Our findings suggest that downstream public health intervention that focus on patients' resilience may reduce the level of perceived stigma and consequently may improve the patients' QoL. Follow-up data are being gathered to support this hypothesis, since resilience is easily influenced by other events and a single assessment might be misleading. In addition to intervention focused on building individual resilience, upstream public health interventions are needed to reduce stigma around IBD improving the awareness on the disease

This study has some limitations. Firstly, the sample size was calculated to validate the Italian translation of the PSS and was not adequate to draw firm conclusions about the level of perceived stigma among IBD patients. Larger prospective studies are needed to explore this aspect. Secondly, the majority of the included patients had a low disease activity and that could represent a bias in interpreting the results. Stigma is internalized over time and, since IBD is a chronic disease, the level of perceived stigma is influenced mostly by the social environment rather than by acute events. On the contrary, resilience is strongly influenced by contextual events and can be improved through behavioral intervention, and this is why the ongoing follow-up of these patients will be useful to better assess the relation between stigma and resilience in IBD. Additionally, the level of resilience of the sampled patients ranged from average to good, and this might have played a role in lowering the stigma scores.

CONCLUSION

To conclude, we have herein developed a validated Italian version of the PSS. Also, we have assessed for the first-time stigmatization and its relation with resilience in a cohort of IBD patients. Interventions aimed at building a stronger resilience may reduce perceived stigma. The follow-up data on the variation of stigma and resilience levels over time are being collected.

ARTICLE HIGHLIGHTS

Research background

Patients living with inflammatory bowel disease (IBD) often experience a poor quality of life due to stigmatization that can be assessed through the IBD perceived stigma scale (PSS). Resilience is the ability to cope positively with a specific disease or situation.

Research motivation

Stigmatization in IBD patients, especially in relation to one's own resilience, has been poorly characterized. A validated Italian version of the IBD-PSS is not available.

Research objectives

To validate an Italian version of the PSS in IBD patients and to assess patients' resilience and its relation with stigmatization.

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Research methods

We enrolled 126 consecutive IBD patients (mean age 46.1 ± 16.9 , male 56.4%), 57 with CD and 69 with UC, in an Italian, tertiary referral, IBD center. Clinical and demographic data were collected, and stigma and resilience were evaluated through the IBD-PSS and the 25-item Connor-Davidson Resilience Scale, respectively. Psychometric validity of the IBD-PSS was assessed, and a multivariable analysis for factors associated with greater stigma was computed.

Research results

We found that the Italian version of the IBD-PSS had an acceptable reliability, having a Cronbach alpha of 0.87, with an excellent test-retest score. The median PSS score was 0.45 (0.20-0.85), and resilience negatively correlated with perceived stigma (Spearman's correlation -0.18, 95% CI: -0.42-0.08, *P* = 0.03).

Research conclusions

We have developed a reliable tool to be used in clinical practice for assessing stigmatization in Italian IBD patients. Also, we found that resilience may have an influence on stigmatization, possibly improving patients' illness perception.

Research perspectives

The Italian IBD-PSS should be used extensively in order to assess this important endpoint in the care of IBD patients. More prospective, long-term studies looking at more detailed factors influencing stigmatization and resilience are urgently needed.

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REFERENCES

- Link BG, Phelan JC. Conceptualizing stigma. Annu Rev Sociol 2001; 27: 363-385 [DOI: 1 10.1146/annurev.soc.27.1.363]
- Sheffield BF, Carney MW. Crohn's disease: a psychosomatic illness? Br J Psychiatry 1976; 128: 446-2 450 [PMID: 1276548 DOI: 10.1192/bjp.128.5.446]
- 3 Taft TH, Keefer L. A systematic review of disease-related stigmatization in patients living with inflammatory bowel disease. Clin Exp Gastroenterol 2016; 9: 49-58 [PMID: 27022294 DOI: 10.2147/CEG.S83533
- Taft TH, Keefer L, Leonhard C, Nealon-Woods M. Impact of perceived stigma on inflammatory bowel disease patient outcomes. Inflamm Bowel Dis 2009; 15: 1224-1232 [PMID: 19180581 DOI: 10.1002/ibd.20864]
- 5 Levenstein S, Li Z, Almer S, Barbosa A, Marquis P, Moser G, Sperber A, Toner B, Drossman DA. Cross-cultural variation in disease-related concerns among patients with inflammatory bowel disease. Am J Gastroenterol 2001; 96: 1822-1830 [PMID: 11419836 DOI: 10.1111/j.1572-0241.2001.03878.x]
- 6 Lenti MV, Cococcia S, Ghorayeb J, Di Sabatino A, Selinger CP. Stigmatisation and resilience in inflammatory bowel disease. Intern Emerg Med 2020; 15: 211-223 [PMID: 31893346 DOI: 10.1007/s11739-019-02268-0]
- 7 Luo D, Lin Z, Shang XC, Li S. "I can fight it! Int J Nurs Sci 2019; 6: 127-133 [PMID: 31406881 DOI: 10.1016/j.ijnss.2018.12.008]
- 8 Acciari AS, Leal RF, Coy CSR, Dias CC, Ayrizono MLS. Relationship among psychological wellbeing, resilience and coping with social and clinical features in crohn's disease patients. Arq Gastroenterol 2019; 56: 131-140 [PMID: 31460575 DOI: 10.1590/S0004-2803.201900000-27]
- Sehgal P, Ungaro RC, Foltz C, Iacoviello B, Dubinsky MC, Keefer L. High Levels of Psychological Resilience Associated With Less Disease Activity, Better Quality of Life, and Fewer Surgeries in Inflammatory Bowel Disease. Inflamm Bowel Dis 2021; 27: 791-796 [PMID: 32696966 DOI: 10.1093/ibd/izaa196]
- 10 Jones MP, Keefer L, Bratten J, Taft TH, Crowell MD, Levy R, Palsson O. Development and initial validation of a measure of perceived stigma in irritable bowel syndrome. Psychol Health Med 2009;



14: 367-374 [PMID: 19444714 DOI: 10.1080/13548500902865956]

- Taft TH, Keefer L, Artz C, Bratten J, Jones MP. Perceptions of illness stigma in patients with 11 inflammatory bowel disease and irritable bowel syndrome. Qual Life Res 2011; 20: 1391-1399 [PMID: 21424542 DOI: 10.1007/s11136-011-9883-x]
- 12 Connor KM, Davidson JR. Development of a new resilience scale: the Connor-Davidson Resilience Scale (CD-RISC). Depress Anxiety 2003; 18: 76-82 [PMID: 12964174 DOI: 10.1002/da.10113]
- Connor KM, Davidson JR. Translations of the CD-RISC [cited February 14, 2021]. In: Connor-13 Davidson Resilience Scale. Available from: http://www.connordavidsonresiliencescale.com/translations.php
- Drossman DA, Patrick DL, Mitchell CM, Zagami EA, Appelbaum MI. Health-related quality of life 14 in inflammatory bowel disease. Functional status and patient worries and concerns. Dig Dis Sci 1989; 34: 1379-1386 [PMID: 2766905 DOI: 10.1007/BF01538073]
- 15 Maaser C, Sturm A, Vavricka SR, Kucharzik T, Fiorino G, Annese V, Calabrese E, Baumgart DC, Bettenworth D, Borralho Nunes P, Burisch J, Castiglione F, Eliakim R, Ellul P, González-Lama Y, Gordon H, Halligan S, Katsanos K, Kopylov U, Kotze PG, Krustinš E, Laghi A, Limdi JK, Rieder F, Rimola J, Taylor SA, Tolan D, van Rheenen P, Verstockt B, Stoker J; European Crohn's and Colitis Organisation [ECCO] and the European Society of Gastrointestinal and Abdominal Radiology [ESGAR]. ECCO-ESGAR Guideline for Diagnostic Assessment in IBD Part 1: Initial diagnosis, monitoring of known IBD, detection of complications. J Crohns Colitis 2019; 13: 144-164 [PMID: 30137275 DOI: 10.1093/ecco-jcc/jjy113]
- 16 Harvey RF, Bradshaw JM. A simple index of Crohn's-disease activity. Lancet 1980; 1: 514 [PMID: 6102236 DOI: 10.1016/s0140-6736(80)92767-1]
- Sandborn WJ, Feagan BG, Hanauer SB, Lochs H, Löfberg R, Modigliani R, Present DH, Rutgeerts 17 P, Schölmerich J, Stange EF, Sutherland LR. A review of activity indices and efficacy endpoints for clinical trials of medical therapy in adults with Crohn's disease. Gastroenterology 2002; 122: 512-530 [PMID: 11832465 DOI: 10.1053/gast.2002.31072]
- Rutgeerts P, Sandborn WJ, Feagan BG, Reinisch W, Olson A, Johanns J, Travers S, Rachmilewitz D, 18 Hanauer SB, Lichtenstein GR, de Villiers WJ, Present D, Sands BE, Colombel JF. Infliximab for induction and maintenance therapy for ulcerative colitis. N Engl J Med 2005; 353: 2462-2476 [PMID: 16339095 DOI: 10.1056/NEJMoa050516]
- Guadagnoli L, Taft TH, Keefer L. Stigma perceptions in patients with eosinophilic gastrointestinal 19 disorders. Dis Esophagus 2017; 30: 1-8 [PMID: 28475723 DOI: 10.1093/dote/dox014]
- 20 Bullinger M, Alonso J, Apolone G, Leplège A, Sullivan M, Wood-Dauphinee S, Gandek B, Wagner A, Aaronson N, Bech P, Fukuhara S, Kaasa S, Ware JE Jr. Translating health status questionnaires and evaluating their quality: the IQOLA Project approach. International Quality of Life Assessment. J Clin Epidemiol 1998; 51: 913-923 [PMID: 9817108 DOI: 10.1016/s0895-4356(98)00082-1]
- 21 Ware JE Jr, Gandek B. Methods for testing data quality, scaling assumptions, and reliability: the IQOLA Project approach. International Quality of Life Assessment. J Clin Epidemiol 1998; 51: 945-952 [PMID: 9817111 DOI: 10.1016/s0895-4356(98)00085-7]
- Rössler W. The stigma of mental disorders: A millennia-long history of social exclusion and 22 prejudices. EMBO Rep 2016; 17: 1250-1253 [PMID: 27470237 DOI: 10.15252/embr.201643041]
- 23 Henderson C, Noblett J, Parke H, Clement S, Caffrey A, Gale-Grant O, Schulze B, Druss B, Thornicroft G. Mental health-related stigma in health care and mental health-care settings. Lancet Psychiatry 2014; 1: 467-482 [PMID: 26361202 DOI: 10.1016/S2215-0366(14)00023-6]
- 24 Rueda S, Mitra S, Chen S, Gogolishvili D, Globerman J, Chambers L, Wilson M, Logie CH, Shi Q, Morassaei S, Rourke SB. Examining the associations between HIV-related stigma and health outcomes in people living with HIV/AIDS: a series of meta-analyses. BMJ Open 2016; 6: e011453 [PMID: 27412106 DOI: 10.1136/bmjopen-2016-011453]
- 25 Chapple A, Ziebland S, McPherson A. Stigma, shame, and blame experienced by patients with lung cancer: qualitative study. BMJ 2004; 328: 1470 [PMID: 15194599 DOI: 10.1136/bmj.38111.639734.7C
- Zhu M, Zhou H, Zhang W, Deng Y, Wang X, Bai X, Li M, Hu R, Hou J, Liu Y. The Stroke Stigma 26 Scale: a reliable and valid stigma measure in patients with stroke. Clin Rehabil 2019; 33: 1800-1809 [PMID: 31307214 DOI: 10.1177/0269215519862329]
- 27 Pourmarzi D, Khoramirad A, Ahmari Tehran H, Abedini Z. Validity and Reliability of Persian Version of HIV/AIDS Related Stigma Scale for People Living With HIV/AIDS in Iran. J Family Reprod Health 2015; 9: 164-171 [PMID: 27047562]
- 28 Bifftu BB, Dachew BA. Perceived Stigma and Associated Factors among People with Schizophrenia at Amanuel Mental Specialized Hospital, Addis Ababa, Ethiopia: A Cross-Sectional Institution Based Study. Psychiatry J 2014; 2014: 694565 [PMID: 24967300 DOI: 10.1155/2014/694565]
- Chandra, A Minkovitz CS. Factors that Influence Mental Health Stigma Among 8th Grade 29 Adolescents. J Youth Adolescence 2007; 36: 763-774 [DOI: 10.1007/s10964-006-9091-0]
- 30 Brown RL. Perceived stigma among people with chronic health conditions: the influence of age, stressor exposure, and psychosocial resources. Res Aging 2015; 37: 335-360 [PMID: 25651574 DOI: 10.1177/0164027514533133
- 31 Bowirrat A, Chen TJ, Blum K, Madigan M, Bailey JA, Chuan Chen AL, Downs BW, Braverman ER, Radi S, Waite RL, Kerner M, Giordano J, Morse S, Oscar-Berman M, Gold M. Neuropsychopharmacogenetics and Neurological Antecedents of Posttraumatic Stress Disorder: Unlocking the Mysteries of Resilience and Vulnerability. Curr Neuropharmacol 2010; 8: 335-358 [PMID:



21629442 DOI: 10.2174/157015910793358123]



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ORIGINAL ARTICLE

Retrospective Cohort Study

Prognostic factors of minimally invasive surgery for gastric cancer: Does robotic gastrectomy bring oncological benefit?

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Abstract

BACKGROUND

Gastric cancer is the third leading cause of cancer-related death worldwide and surgical resection remains the sole curative treatment for gastric cancer. Minimally invasive gastrectomy including laparoscopic and robotic approaches has been increasingly used in a few decades. Thus far, only a few reports have investigated the oncological outcomes following minimally invasive gastrectomy.

AIM

To determine the 5-year survival following minimally invasive gastrectomy for gastric cancer and identify prognostic predictors.

METHODS

This retrospective cohort study identified 939 patients who underwent gastrectomy for gastric cancer during the study period. After excluding 125 patients with non-curative surgery (n = 77), other synchronous cancer (n = 2), remnant gastric cancer (n = 25), insufficient physical function (n = 13), and open gastrectomy (n = 8), a total of 814 consecutive patients with primary gastric cancer who underwent minimally invasive R0 gastrectomy at our institution between 2009 and 2014 were retrospectively examined. Accordingly, 5-year overall and recurrence-free survival were analyzed using the Kaplan-Meier method with the log-rank test and Cox regression analyses, while factors associated with survival were determined using multivariate analysis.

RESULTS

Our analysis showed that age > 65 years, American Society of Anesthesiologists



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(ASA) physical status 3, total or proximal gastrectomy, and pathological T4 and N positive status were independent predictors of both 5-year overall and recurrencefree survival. Accordingly, the included patients had a 5-year overall and recurrence-free survival of 80.3% and 78.2%, respectively. Among the 814 patients, 157 (19.3%) underwent robotic gastrectomy, while 308 (37.2%) were diagnosed with pathological stage II or III disease. Notably, our findings showed that robotic gastrectomy was an independent positive predictor for recurrence-free survival in patients with pathological stage II/III [hazard ratio: 0.56 (0.33-0.96), *P* = 0.035]. Comparison of recurrence-free survival between the robotic and laparoscopic approach using propensity score matching analysis verified that the robotic group had less morbidity (P = 0.005).

CONCLUSION

Age, ASA status, gastrectomy type, and pathological T and N status were prognostic factors of minimally invasive gastrectomy, with the robot approach possibly improving long-term outcomes of advanced gastric cancer.

Key Words: Laparoscopy; Gastric cancer; Minimally invasive surgery; Prognostic factor; Stomach neoplasms

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Core Tip: This retrospective cohort study on 814 patients undergoing minimally invasive surgery for primary gastric cancer revealed a 5-year overall and recurrencefree survival of 80.3% and 78.2%, respectively. Moreover, our analysis identified age, American Society of Anesthesiologists status, type of gastrectomy, and pathological T and N status as prognostic predictors for overall and recurrence-free survival. The robotic approach was also identified as an independent positive predictor for recurrence-free survival in patients with pathological stage II/III disease, confirmed by the lesser morbidity in the robotic group following propensity score analysis.

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INTRODUCTION

Gastric cancer is the fifth most common malignancy and the third leading cause of cancer-related death worldwide[1]. Surgical resection remains the sole curative treatment for gastric cancer, with regional lymphadenectomy being recommended as a component of radical gastrectomy^[2]. Laparoscopic gastrectomy has been increasingly used, considering its better short-term effects and comparable long-term outcomes compared to open gastrectomy[2].

The da Vinci surgical system (DVSS; Intuitive Surgical, Sunnyvale, CA, United States) had been developed to overcome several disadvantages identified for standard minimally invasive laparoscopic surgery[2]. Most laparoscopic surgeons expect that utilizing the DVSS for gastric surgery would allow them to overcome the technical difficulties of laparoscopic gastrectomy, thereby improving its safety, reproducibility, teachability, and long-term outcomes. However, only one large, nonrandomized prospective study (NCT01309256) has compared DVSS with laparoscopic gastrectomy. Accordingly, the study results mentioned above demonstrated that DVSS had higher operative time and cost than laparoscopic gastrectomy with no difference in morbidity, suggesting that DVSS might reduce cost-effectiveness³. Concurrently, robotic gastrectomy, which has been actively used for operable patients with resectable gastric cancer at the patient's own expense^[2], was introduced at our institution in 2009. Analysis of patient outcomes following robotic gastrectomy had demonstrated that its morbidity was approximately one-fifth of that observed with



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laparoscopic gastrectomy, with such a reduction in morbidity, including decreased incidences of postoperative pancreatic fistula, certainly improving the short-term postoperative course[2]. Moreover, our previous study had compared the oncological outcomes, particularly 3-year survival rates, between robotic gastrectomy and laparoscopic gastrectomy^[2]. Thus far, only a few reports have investigated the oncological outcomes following robotic gastrectomy, considering that DVSS remains a relatively new technology. Therefore, the current study aimed to determine the prognostic factors of minimally invasive gastrectomy, including laparoscopic and robotic procedures.

MATERIALS AND METHODS

Patients

This single-center retrospective cohort study included patients who underwent curative gastrectomy for gastric cancer at our institution between January 2009 and September 2014. The inclusion criteria were patients with primary gastric adenocarcinoma who underwent curative resection using minimally invasive surgery (MIS). The exclusion criteria were patients with other synchronous cancer and those whose resection was limited due to poor physical functioning. Among the 939 patients who underwent gastrectomy for gastric cancer during the study period, 125 were excluded due to non-curative surgery (n = 77), other synchronous cancer (n = 2), remnant gastric cancer (n = 25), insufficient physical function (n = 13), and open gastrectomy (n = 8). Thus, the 814 patients who satisfied the study criteria were ultimately analyzed. The clinicopathological variables collected included age, sex, body mass index (BMI), American Society of Anesthesiologists (ASA) physical status classification, date of surgery, type of approach, histologic type, lymphovascular invasion status, TNM staging (Japanese Gastric Cancer Association classification, 14th edition), number of harvested lymph nodes, postoperative complications determined by Clavien-Dindo (C-D) classification[4], date of the first recurrence, and date and status of the last follow-up. The extent of gastrectomy and lymphadenectomy was defined based on Japanese gastric cancer treatment guidelines^[5]. Overall survival (OS) was calculated from the date of resection to the date of the last follow-up or death of any cause. Recurrence-free survival (RFS) was calculated from the date of resection to the date of first recurrence, last follow-up, or death of any cause, whichever occurred first. Details regarding indications for radical gastrectomy, including the selection of laparoscopic or robotic approach, surgical procedures, perioperative management, adjuvant chemotherapy, and oncologic follow-up, have been previously reported[2]. Neoadjuvant chemotherapy (NAC) (S-1 80 mg/m² days 1-21 + CDDP 60 mg/m² day 8 or S-1 80 mg/m² days 1-28) was offered to patients with clinical T \ge 2, tumor size \ge 5 cm, and/or swollen locoregional lymph nodes \geq 1.5 cm[2]. All patients were uniformly offered robotic surgery without considering their backgrounds, including physical and oncological status. Patients who agreed to the uninsured use of the surgical robot underwent robotic gastrectomy, whereas those who wished for insured treatment underwent laparoscopic gastrectomy[2]. All patients were completely involved in the decision-making process and provided informed consent prior to participation. All surgical procedures were performed or guided by surgeons qualified by the Japanese Society for Endoscopic Surgical Skill Qualification System, initiated in 2004 by the Japanese Society for Endoscopic Surgery to develop a tool for the reliable and reproducible evaluation of trainees' surgical techniques[6]. All procedures were supervised by an expert gastric surgeon (I.U.) who had performed more than 1500 Laparoscopic gastrectomies and 400 robotic gastrectomies. This study was approved by the institutional review board of Fujita Health University.

Statistical analysis

All statistical analyses were conducted using IBM SPSS Statistics 25 (IBM Corporation, Armonk, NY, United States). Long-term outcomes were analyzed using the Kaplan-Meier method with the log-rank test and Cox regression analyses. Considering our relatively small sample size, multivariate analysis was conducted using all variables determined to be significant (P < 0.1) during univariate analysis as independent variables. Data were expressed as median, interquartile range, or hazard ratio (HR) with the 95% confidence interval (CI) unless otherwise stated. A P value of < 0.05 (two-tailed) was considered statistically significant. Propensity score matching analysis was used to reduce selection bias with regard to potential confounding factors when establishing the laparoscopic and robotic groups. Possible confounders were



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selected based on their potential association with the outcome of interest according to clinical knowledge. Therefore, clinicopathological characteristics (age, BMI, sex, ASA status, pathological T and N factor, type of surgery, tumor size, and NAC) were used to adjust differences between the laparoscopic and robotic groups through one-to-one pair matching using optimal match without replacement. Propensity scores were matched using a caliper width 1/5 Logit of the standard deviation. The absolute standardized difference was used to measure covariate balance, in which an absolute standardized mean difference above represented a meaningful imbalance^[7]. Independent continuous variables were compared using the Mann–Whitney U test or Kruskal-Wallis test. Categorical variables were compared using the χ^2 test or Fisher's exact test.

RESULTS

Patient characteristics

Table 1 summarizes the characteristics of all patients included herein. Accordingly, the included patients had a median age of 68 years, among whom 31.4% (*n* = 256) were diagnosed with clinical stage II or more disease, while 14.6% (n = 119) underwent NAC. Laparoscopic and robotic gastrectomy was performed in 657 (80.7%) and 157 (19.3%) patients, respectively. None of the patients required intraoperative conversion to open procedure from MIS. Pathological stage II and III disease was diagnosed in 160 (19.7%) and 148 (18.2%) patients, respectively. Morbidity of C-D grade \geq III was observed in 72 patients (8.8%).

Survival outcomes

The median follow-up period was 59.5 mo, while the 5-year OS and RFS were 80.3% and 78.2%, respectively (Figure 1A and B). Patients with pStage I, II, and III had a 5year OS of 91.9%, 76.3%, and 43.7%, and a 5-year RFS of 91.6%, 74.7%, and 36.0%, respectively.

Factors related to survival

Univariate analysis identified age > 65 years, ASA status 3, total or proximal gastrectomy, D2 lymphadenectomy, tumor size > 30 mm, lymphovascular invasion, C-D grade ≥ III morbidity, NAC administration, adjuvant chemotherapy administration, and higher pT and pN status as factors significantly associated with OS (Table 2). However, multivariate analysis revealed that only age > 65 years [HR: 1.62 (1.09-2.40), *P* = 0.017], ASA status 3 [HR: 1.91 (1.18-3.10), *P* = 0.009], total or proximal gastrectomy [HR: 1.45 (1.03-2.05), P = 0.036], pT4 [HR: 4.31 (2.37-7.82), P < 0.001], and pN positive status were significantly and independently associated with OS (Table 2). Similarly, multivariate analysis identified age > 65 years [HR: 1.48 (1.02-2.14), P =0.038], ASA status 3 [HR: 1.62 (1.02-2.60), P = 0.043], total or proximal gastrectomy [HR: 1.55 (1.12–2.15), P = 0.009], pT4 [HR: 4.20 (2.38–7.41), P < 0.001], and pN positive status as factors significantly and independently associated with RFS (Table 3). Moreover, multivariate analysis showed that robotic approach could likely be a positive predictor for RFS, although no significant association was observed [HR 0.68 (0.44-1.06), P = 0.088] (Table 3).

Survival outcomes following the laparoscopic and robotic approach

The laparoscopic and robotic approach had a 5-year OS of 79.4% and 83.4% (P = 0.243) and a 5-year RFS of 76.9% and 84.2% (P = 0.085), respectively. No significant difference in the 5-year OS and RFS was noted between both groups for patients with pStage I (91.6% vs 93.4%, P = 0.471 and 91.4% vs 92.7%, P = 0.634) (Figure 2A and B). Notably, among patients with pStage II/III, those in the robotic group had significantly better RFS compared to those in the laparoscopic group (74.1% vs 51.7%, P = 0.006) (Figure 2D), although no significant difference in the 5-year OS was observed (P =0.071) (Figure 2C).

Factors associated with survival in pStage II/III diseases

Our analysis showed that pT4 [HR: 4.02 (1.21-13.42), P = 0.024] and pN positive status were significantly and independently associated with OS. Notably, univariate analysis showed that robotic gastrectomy (P = 0.007), total or proximal gastrectomy (P = 0.004), tumor size > 30 mm (P = 0.014), pT4 (P = 0.007), and pN positive status were significantly associated with RFS. Meanwhile, multivariate analysis found that robotic



Variables	n = 814
Age, yr [IQR]	68[61-74]
ъде, уг [юск] Бех, n (%)	00[01-74]
Male	562 (69.0)
Female	
BMI, kg/m ² [IQR]	252 (31.0) 22.2 [20.0-24.1]
ASA, n (%)	22.2 [20.0-24.1]
1	314 (38.6)
2	396 (48.6)
3	104 (12.8)
	10* (12.0)
Clinical stage, n (%)	EE9 (69 6)
I	558 (68.6)
Ш	125 (15.3)
III	121 (14.9)
IV	10 (1.2)
Neoadjuvant chemotherapy, n (%)	119 (14.6)
Neoadjuvant radiotherapy, n (%)	0 (0)
Approach, n (%)	
Laparoscopic	657 (80.7)
Robotic	157 (19.3)
Гуре of gastrectomy, n (%)	
Distal	559 (68.7)
Total	238 (29.2)
Proximal	16 (2.0)
Pylorus preserving	1 (0.1)
Lymphadenectomy, n (%)	
D1+	378 (46.4)
D2	436 (53.6)
Dissected nodes, <i>n</i> [IQR]	38[28-48]
Гumor size, mm [IQR]	30[20-50]
pT, n (%)	
1	469 (57.6)
2	87 (10.7)
3	112 (13.8)
4	138 (17.0)
CR	8 (1.0)
DN, n (%)	
0	559 (68.7)
1	98 (12.0)
2	79 (9.7)
3	78 (9.6)
pStage, n (%)	

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I	498 (61.2)
П	160 (19.7)
ш	148 (18.2)
TCRNany	8 (1.0)
WHO histologic type, <i>n</i> (%)	
Tub/pap	402 (49.4)
Por/sig	352 (43.2)
Mixed/other	60 (7.4)
Lymphovascular invasion, <i>n</i> (%)	531 (65.2)
Adjuvant chemotherapy, n (%)	242 (29.7)
Adjuvant radiotherapy, n (%)	0 (0)
Morbidity (C–D grade \geq III), <i>n</i> (%)	72 (8.8)
Anastomotic leakage	22 (2.7)
Pancreatic fistula	30 (3.7)

Categorical and continuous data are presented as n (%) and median [IQR], respectively. IQR: Interquartile range; ASA: American Society of Anesthesiologists; BMI: Body mass index; CR: Complete response at the primary site; C-D: Clavien-Dindo classification.

> gastrectomy was independently and positively associated with RFS [HR: 0.56 (0.33-0.96), P = 0.035]. Apart from robotic gastrectomy, only pT4 and pN positive status were identified as factors independently associated with RFS (Table 4).

Comparison between robotic and laparoscopic gastrectomy in pStage II/III diseases

To account for confounding factors between both groups, propensity score matching was performed (Table 5). In the prematched cohort of 308 patients with pStage II/III disease, 67 and 241 patients belonged to the robotic and laparoscopic groups, respectively. After matching, each group comprised 61 patients. The matched cohort had a considerably better balance of covariates, with < 0.245 of the cutoff value of an absolute standardized difference. In the postmatched cohort, no differences in clinicopathological variables were observed between the laparoscopic and robotic groups, although the robotic group had lower morbidity (4.9% vs 16.4%, P = 0.04) (Table 5). Furthermore, the robotic group had significantly better 5-year OS (70.4% vs 50.2%, *P* = 0.039) and RFS (74.1% *vs* 44.5%, *P* = 0.005) than the laparoscopic group in the postmatched cohort (Figure 3A and B).

DISCUSSION

The current study clearly identified factors related to survival in patients with gastric cancer who underwent MIS, subsequently presenting three significant findings.

First, the present study highlighted the feasibility and safety of MIS for gastric cancer as determined by the 5-year outcomes. While the long-term outcomes of laparoscopic surgery have been increasingly reported in recent years, only a few studies have investigated the long-term outcomes of the robotic approach [2,8,9]. Consistent with previous studies, including those from our group, the current study demonstrated no significant difference in OS and RFS between the laparoscopic and robotic approaches [2,8,9]. However, among patients with pStage II/III, those in the robotic group demonstrated significantly better RFS than those in the laparoscopic group (P = 0.006).

Second, our results showed that pT and pN status was independently associated with both OS and RFS. Currently, multidisciplinary treatment for gastric cancer utilizing various chemotherapeutic options has been developed worldwide. In Western countries, neoadjuvant and adjuvant chemotherapy combined with curative resection has been the standard treatment for advanced gastric cancer[10,11], whereas adjuvant chemotherapy following curative resection remains the standard approach in Japan^[5]. Regardless of treatment options, however, evidence has shown that the pN factor is consistently strongly associated with survival following gastric cancer treatment[12-14]. The results of the current study are consistent with those presented



Table 2 Factors associated with overall survival for the entire cohort (<i>n</i> = 814)							
	Univariat	e		Multivari	ate		
	HR	95%CI	Р	HR	95%CI	Р	
Age > 65 yr	1.46	1.04-2.06	0.031	1.62	1.09-2.40	0.017	
Female sex	0.75	0.52-1.09	0.129				
BMI > 23 kg/m ²	0.77	0.55-1.07	0.123				
ASA							
1	1			1			
2	0.96	0.68-1.38	0.837	1.06	0.72-1.57	0.753	
3	1.97	1.27-3.05	0.003	1.91	1.18-3.10	0.009	
Neoadjuvant chemotherapy	1.84	1.27-2.67	0.001	1.34	0.88-2.04	0.166	
Robotic approach	0.77	0.50-1.21	0.258				
Type of gastrectomy							
Distal/pylorus-preserving	1			1			
Total/proximal	2.17	1.58-2.99	< 0.001	1.45	1.03-2.05	0.036	
D2 lymphadenectomy	1.86	1.32-2.61	< 0.001	0.87	0.57-1.33	0.528	
Tumor > 30 mm	3.23	2.20-4.75	< 0.001	1.05	0.66-1.69	0.832	
WHO histologic type							
Tub/pap	1			1			
Por/sig/mixed/other	1.54	1.12-2.13	0.009	1.26	0.89-1.78	0.190	
Lymphovascular invasion	4.38	2.68-7.17	< 0.001	1.17	0.60-2.26	0.651	
pT							
1	1			1			
2	2.82	1.62-4.91	< 0.001	1.72	0.90-3.27	0.099	
3	3.03	1.82-5.03	< 0.001	1.54	0.82-2.91	0.184	
4	9.78	6.54-14.60	< 0.001	4.31	2.37-7.82	< 0.001	
CR	1.67	0.23-12.17	0.613	1.34	0.16-10.97	0.784	
pN							
0	1			1			
1	2.76	1.74-4.39	< 0.001	2.02	1.22-3.34	0.007	
2	4.05	2.56-6.41	< 0.001	1.97	1.15-3.36	0.013	
3	8.08	5.40-12.10	< 0.001	2.92	1.79-4.78	< 0.001	
Adjuvant chemotherapy	3.27	2.37-4.50	< 0.001	1.21	0.80-1.82	0.371	
Morbidity (C-D grade ≥ III)	1.85	1.18-2.91	0.008	1.27	0.79-2.05	0.325	

HR: Hazard ratio; CI: Confidence interval; ASA: American Society of Anesthesiologists; BMI: Body mass index; CR: Complete response at the primary site; C-D: Clavien-Dindo classification.

in previous studies.

Third, the use of the surgical robot was significantly associated with improved RFS among propensity score-matched patients with pStage II/III disease. This could have been attributed to lower morbidity in the robotic gastrectomy group, a causal relationship between morbidity and survival, and higher morbidity in patients undergoing surgery for advanced disease. First, a few studies have shown that robotic gastrectomy was technically safe and feasible but did not have superior morbidity compared to the laparoscopic approach[3]. However, Wang et al[15] who compared morbidity between robotic and laparoscopic gastrectomy using propensity score-

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		Univariate		Multivar	iate	
	HR	95%CI	Р	HR	95%CI	Р
Age > 65 yr	1.33	0.97-1.84	0.076	1.48	1.02-2.14	0.038
Female sex	0.76	0.54-1.06	0.108			
BMI > 23 kg/m ²	0.77	0.56-1.05	0.100			
ASA						
1	1			1		
2	0.95	0.68-1.32	0.761	1.08	0.75-1.55	0.692
3	1.67	1.09-2.55	0.018	1.62	1.02-2.60	0.043
Neoadjuvant chemotherapy	1.91	1.34-2.71	< 0.001	1.39	0.93-2.08	0.104
Robotic approach	0.69	0.45-1.06	0.087	0.68	0.44-1.06	0.088
Гуре of gastrectomy						
Distal/pylorus-preserving	1			1		
Total/proximal	2.24	1.66-3.02	< 0.001	1.55	1.12-2.15	0.009
D2 lymphadenectomy	2.08	1.50-2.86	< 0.001	0.99	0.66-1.48	0.957
Гитог > 30 mm	3.19	2.23-4.56	< 0.001	0.95	0.61-1.48	0.827
VHO histologic type						
Tub/pap	1			1		
Por/sig/mixed/other	0.54	0.14-2.08	0.005	1.20	0.86-1.66	0.284
ymphovascular invasion	4.93	3.06-7.94	< 0.001	1.29	0.69-2.43	0.430
ρT						
1	1			1		
2	2.87	1.69-4.85	< 0.001	1.57	0.85-2.89	0.148
3	3.42	2.13-5.48	< 0.001	1.60	0.89-2.89	0.120
4	10.62	7.26-15.53	< 0.001	4.20	2.38-7.41	< 0.001
CR	1.41	0.19–10.24	0.737	0.96	0.12-7.77	0.967
bN						
0	1			1		
1	3.08	1.99-4.77	< 0.001	2.23	1.39-3.58	0.001
2	5.01	3.29-7.62	< 0.001	2.24	1.36-3.70	0.002
3	8.92	6.07-13.11	< 0.001	3.32	2.06-5.34	< 0.001
Adjuvant chemotherapy	3.53	2.62-4.77	< 0.001	1.18	0.80-1.73	0.410
Morbidity (C-D grade≥III)	1.69	1.09-2.62	0.019	1.04	0.65-1.67	0.868

ASA: American Society of Anesthesiologists; BMI: Body mass index; CR: Complete response at the primary site; C-D: Clavien-Dindo classification; HR: Hazard ratio; CI: Confidence interval.

> matched analysis, reported that the robotic group exhibited significantly lower morbidity, particularly with regard to infectious complications (e.g., anastomotic leakage and intra-abdominal abscess)[15]. Furthermore, a multicenter, prospective, single-arm study by our group recently reported that robotic gastrectomy promoted lesser morbidity than laparoscopic gastrectomy among historical controls[2]. Similarly, the present study showed that the robotic group had significantly lesser morbidity compared to the laparoscopic in the postmatched cohort. Second, several studies have demonstrated that morbidity was associated with worse survival in gastric cancer[16-19]. In fact, Jin et al[16] reported that patients with and without postoperative complic-

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Table 4 Factors associated with recurrence-free survival for patients with pathological stage II/III disease (<i>n</i> = 308)						
	Univariate			Multivariate		
	HR	95%CI	Р	HR	95%CI	Р
Age > 65 yr	1.04	0.73-1.48	0.848			
Female sex	1.08	0.75-1.58	0.673			
BMI > 23 kg/m ²	0.69	0.47-1.00	0.052	0.92	0.62-1.35	0.657
ASA						
1	1					
2	0.82	0.56-1.19	0.297			
3	1.07	0.63-1.80	0.809			
Neoadjuvant chemotherapy	1.37	0.93-2.01	0.114			
Robotic approach	0.50	0.30-0.83	0.007	0.56	0.33-0.96	0.035
Type of gastrectomy						
Distal/pylorus-preserving	1			1		
Total/proximal	1.67	1.18-2.37	0.004	1.32	0.91-1.90	0.145
D2 lymphadenectomy	1.26	0.80-2.00	0.320			
Tumor > 30 mm	2.17	1.17-4.03	0.014	1.34	0.69–2.60	0.303
WHO histologic type						
Tub/pap	1			1		
Por/sig/mixed/other	1.38	0.95-2.00	0.089	1.29	0.88-1.90	0.197
pT						
1	1			1		
2	1.82	0.60-5.53	0.292	1.33	0.42-4.23	0.628
3	1.32	0.47-3.75	0.6	1.45	0.49-4.30	0.505
4	3.96	1.45-10.83	0.007	3.52	1.23-10.07	0.019
pN						
0	1			1		
1	2.07	1.16-3.69	0.014	2.86	1.57-5.24	0.001
2	2.24	1.29-3.90	0.004	2.45	1.38-4.34	0.002
3	3.74	2.21-6.32	< 0.001	3.25	1.88-5.61	< 0.001
Adjuvant chemotherapy	1.37	0.92-2.04	0.119			
Morbidity (C-D grade \geq III)	1.58	0.97-2.58	0.066	1.22	0.73-2.05	0.453

ASA: American Society of Anesthesiologists; BMI: Body mass index; C-D: Clavien-Dindo classification; HR: Hazard ratio; CI: Confidence interval.

ations had RFS rates of 23% and 40%, respectively (P < 0.001). Third, advanced gastric cancer requires complicated procedures, which can cause more complications. Notably, studies have reported that patient with advanced disease had morbidity rates of 8.3%-15.2% following minimally invasive gastrectomy, respectively[20,21]. The aforementioned findings therefore indicate that utilizing surgical robots, which cause less morbidity, might at least partly contribute to the better RFS in patients with advanced gastric cancer, suggesting that surgical robots may be more beneficial for patients with advanced disease. However, although univariate analysis found morbidity to be significantly associated with RFS, multivariate analysis did not identify the same as a significant independent factor associated with RFS in the entire cohort. As such, further investigations are warranted to confirm such findings.

The current study has several limitations worth noting. First, this study was retrospective in nature and involved only a single institution. Moreover, the sample

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Nakauchi M et al. Prognostic factors of MIS for gastric cancer

	Prematched				Postmatched			
	Lap (<i>n</i> = 241)	Robotic (<i>n</i> = 67)	D	ASD	Lap (<i>n</i> = 61)	Robotic (<i>n</i> = 61)	D	ASD
Sex, n (%)	Lap (11 - 241)		0.132	0.204			0.580	0.100
Male	174 (72.2)	42 (62.7)			35 (57.4)	38 (62.3)		
Female	67 (27.8)	25 (37.3)			26 (42.6)	23 (37.7)		
Age, yr [IQR]	69 [61-75]	65 [60-77]	0.134	0.235	68 [61-75]	65 [60-77]	0.824	0.042
BMI, kg/m ² [IQR]	21.6 [19.2-23.7]	23.1 [20.0-24.8]	0.008	0.329	22.6 [20.4-24.9]	23.0 [20.0-24.9]	0.810	0.007
ASA, n (%)			0.074	0.315			0.959	0.052
1	89 (36.9)	35 (52.2)			31 (50.8)	30 (49.2)		
2	118 (49.0)	24 (35.8)			23 (37.7)	23 (37.7)		
3	34 (14.1)	8 (11.9)			7 (11.5)	8 (13.1)		
Neoadjuvant chemotherapy, n (%)	61 (25.3)	11 (16.4)	0.128	0.220	10 (16.4)	11 (18.0)	0.810	0.043
Type of gastrectomy, <i>n</i> (%)			0.075	0.329			1	< 0.001
Distal	136 (56.4)	48 (71.6)			42 (68.9)	42 (68.9)		
Total	104 (43.2)	19 (28.4)			19 (31.1)	19 (31.1)		
Proximal	1 (0.4)	0 (0)			0 (0)	0 (0)		
Tumor size, mm [IQR]	50[35-70]	40[30-63]	0.026	0.265	50 [35-77]	43 [30-65]	0.192	0.187
рТ, и (%)			0.042	0.391			0.860	0.158
1	11 (4.6)	8 (11.9)			4 (6.6)	6 (9.8)		
2	35 (14.5)	4 (6.0)			4 (6.6)	4 (6.6)		
3	85 (35.3)	27 (40.3)			21 (34.4)	23 (37.7)		
4	110 (45.6)	28 (41.8)			32 (52.5)	28 (45.9)		
pN, n (%)			0.15	0.338			0.617	0.244
0	65 (27.0)	24 (35.8)			16 (26.2)	22 (36.1)		
1	48 (19.9)	15 (22.4)			17 (27.9)	13 (21.3)		
2	68 (28.2)	10 (14.9)			9 (14.8)	10 (16.4)		
3	60 (24.9)	18 (26.9)			19 (31.1)	16 (26.2)		
pStage, n (%)			0.246				0.716	
Ш	121 (50.2)	39 (58.2)			32 (52.5)	34 (55.7)		
III	120 (49.8)	28 (41.8)			29 (47.5)	27 (44.3)		
Dissected nodes, n [IQR]	44 [35-53]	43 [35-51]	0.858		45 [35-54]	43 [30-65]	0.556	
WHO histological type, n (%	5)		0.667				0.229	
Tub/pap	88 (36.5)	27 (41.8)			17 (27.9)	26 (42.6)		
Por/sig	129 (53.5)	34 (50.7)			37 (60.7)	30 (49.2)		
Mixed/other	24 (10.0)	5 (7.5)			7 (11.5)	5 (8.2)		
Lymphovascular invasion, n (%)	241 (100)	66 (98.5)	0.218		61 (100)	60 (98.4)	0.5	
Adjuvant chemotherapy, <i>n</i> (%)	161 (66.8)	47 (70.1)	0.605		38 (62.3)	43 (70.5)	0.338	
Morbidity (C-D grade ≥ III), n (%)	31 (12.9)	3 (4.5)	0.053		10 (16.4)	3 (4.9)	0.04	

Categorical and continuous data are presented as n (%) and median [IQR], respectively. ASA: American Society of Anesthesiologists; BMI: Body mass



index; CR: Complete response at the primary site; C-D: Clavien-Dindo classification; ASD: Absolute standardized mean difference.

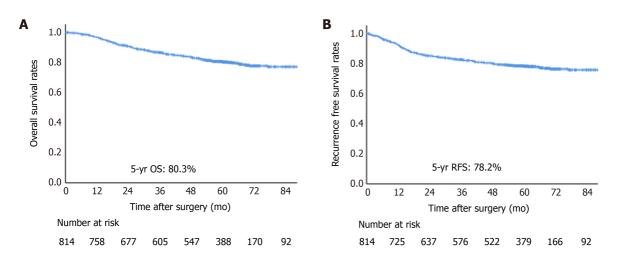


Figure 1 Kaplan-Meier curves. Kaplan-Meier estimates in the entire cohort A: Overall survival probability; B: Recurrence-free survival probability. OS: Overall survival; RFS: Recurrence-free survival.

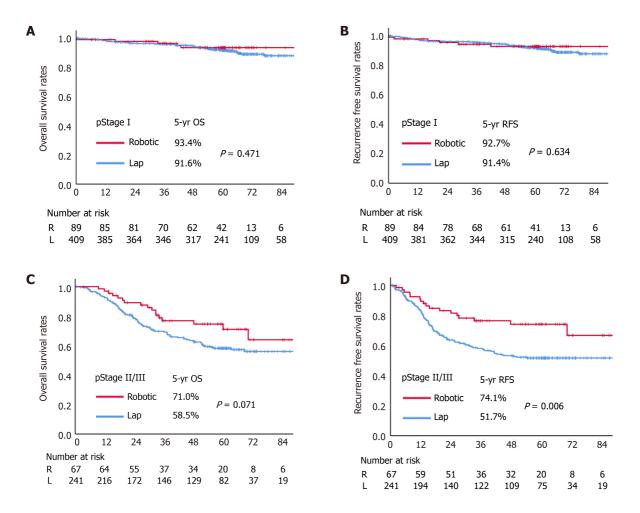


Figure 2 Kaplan-Meier curves. A and C: Kaplan-Meier estimates of overall survival probability for pathological stage I and II/III, B and D: Kaplan-Meier estimates of recurrence-free survival probability for pathological stage I and II/III. OS: Overall survival; RFS: Recurrence-free survival.

size, particularly that of the robotic group, was relatively small. Therefore, given that biases may exist in our data, the overall results should be interpreted with caution. As described in our previous reports[2,6], patients were selected according to whether the they agreed to the uninsured use of robot-assisted surgery, which may have caused

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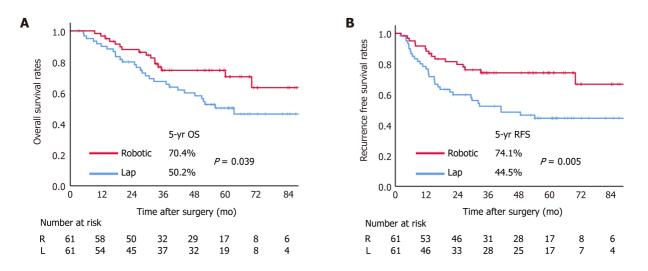


Figure 3 Kaplan-Meier curves. Kaplan-Meier estimates in the postmatched cohort. A: Overall survival probability; B: Recurrence-free survival probability. OS: Overall survival; RFS: Recurrence-free survival.

selection bias due to a possible preference for robotic gastrectomy in patients of higher economic status. However, this was an inherent limitation at the time of study enrollment considering that the DVSS was not covered by the medical insurance in Japan at the time the enrolled patients underwent gastrectomy, whereas conventional laparoscopic gastrectomy was covered. Second, propensity score matching between the laparoscopic and robotic group did not account for adjuvant chemotherapy administration given that, similarly to postoperative complications, adjuvant chemotherapy was determined after robotic or laparoscopic gastrectomy was conducted. Considering that both adjuvant chemotherapy and postoperative complications may affect prognosis[2,6,21], well-designed prospective trials are needed to determine a cause-effect relationship between robotic or laparoscopic gastrectomy and postoperative complications, as well as adjuvant chemotherapy administration.

CONCLUSION

In conclusion, the current study identified age, ASA status, type of gastrectomy, and pathological T and N status are prognostic factors of minimally invasive gastrectomy for gastric cancer. Moreover, the use of robotic assistance was associated with reduced early morbidity, as well as potentially better oncological outcomes in advanced gastric cancer.

ARTICLE HIGHLIGHTS

Research background

Minimally invasive surgery (MIS) including laparoscopic and robotic approaches for gastric cancer has been increasingly used because of its beneficial short-term effects over the open approach. However, oncological outcomes are not established.

Research motivation

There have been few reports on the oncological outcomes of MIS for gastric cancer patients, especially for the robotic approach, because a surgical robot remains a relatively new technology. Therefore, this study aimed to determine the prognostic factors of minimally invasive gastrectomy, including laparoscopic and robotic approaches.

Research objectives

This study aimed to determine the prognostic factors of minimally invasive gastrectomy, including laparoscopic and robotic approaches.

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Research methods

This single-institutional retrospective cohort study included 814 consecutive patients with primary gastric cancer who underwent minimally invasive R0 gastrectomy between 2009 and 2014. We retrospectively examined 5-year overall survival and recurrence-free survival and investigated factors related to survival.

Research results

Age > 65 years, American Society of Anesthesiologists (ASA) physical status 3, total or proximal gastrectomy, and pathological T4 and N positive status were independent predictors of overall survival and recurrence-free survival. The five-year overall survival and recurrence-free survival were 80.3% and 78.2%, respectively. Of all 814 patients, 157 patients (19.3%) underwent robotic gastrectomy and 308 (37.2%) were diagnosed with pathological stage II or III disease. Robotic gastrectomy was an independent positive predictor for recurrence-free survival in pathological stage II/III patients (hazard ratio: 0.56 [0.33-0.96], P = 0.035). Comparison of recurrence-free survival between robotic and laparoscopic approach using propensity score matching analysis verified that with less morbidity in the robotic group (P = 0.005).

Research conclusions

Age, ASA status, type of gastrectomy, and pathological T and N status were prognostic factors of minimally invasive gastrectomy for gastric cancer, and the use of a surgical robot may improve its long-term outcomes for advanced gastric cancer.

Research perspectives

Future studies to better prove the efficacy of robotic gastrectomy for advanced gastric cancer patients are warranted.

REFERENCES

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- Shibasaki S, Suda K, Obama K, Yoshida M, Uyama I. Should robotic gastrectomy become a standard 2 surgical treatment option for gastric cancer? Surg Today 2020; 50: 955-965 [PMID: 31512060 DOI: 10.1007/s00595-019-01875-w]
- Kim HI, Han SU, Yang HK, Kim YW, Lee HJ, Ryu KW, Park JM, An JY, Kim MC, Park S, Song 3 KY, Oh SJ, Kong SH, Suh BJ, Yang DH, Ha TK, Kim YN, Hyung WJ. Multicenter Prospective Comparative Study of Robotic Versus Laparoscopic Gastrectomy for Gastric Adenocarcinoma. Ann Surg 2016; 263: 103-109 [PMID: 26020107 DOI: 10.1097/SLA.000000000001249]
- 4 Clavien PA, Barkun J, de Oliveira ML, Vauthey JN, Dindo D, Schulick RD, de Santibañes E, Pekolj J, Slankamenac K, Bassi C, Graf R, Vonlanthen R, Padbury R, Cameron JL, Makuuchi M. The Clavien-Dindo classification of surgical complications: five-year experience. Ann Surg 2009; 250: 187-196 [PMID: 19638912 DOI: 10.1097/SLA.0b013e3181b13ca2]
- 5 Japanese Gastric Cancer Association. Japanese gastric cancer treatment guidelines 2014 (ver. 4). Gastric Cancer 2017; 20: 1-19 [PMID: 27342689 DOI: 10.1007/s10120-016-0622-4]
- Mori T, Kimura T, Kitajima M. Skill accreditation system for laparoscopic gastroenterologic 6 surgeons in Japan. Minim Invasive Ther Allied Technol 2010; 19: 18-23 [PMID: 20095893 DOI: 10.3109/136457009034929691
- Austin PC. Balance diagnostics for comparing the distribution of baseline covariates between 7 treatment groups in propensity-score matched samples. Stat Med 2009; 28: 3083-3107 [PMID: 19757444 DOI: 10.1002/sim.3697]
- Jiang Y, Zhao Y, Qian F, Shi Y, Hao Y, Chen J, Li P, Yu P. The long-term clinical outcomes of robotic gastrectomy for gastric cancer: a large-scale single institutional retrospective study. Am J Transl Res 2018; 10: 3233-3242 [PMID: 30416664]
- Okumura N, Son T, Kim YM, Kim HI, An JY, Noh SH, Hyung WJ. Robotic gastrectomy for elderly gastric cancer patients: comparisons with robotic gastrectomy in younger patients and laparoscopic gastrectomy in the elderly. Gastric Cancer 2016; 19: 1125-1134 [PMID: 26541766 DOI: 10.1007/s10120-015-0560-6
- 10 Cunningham D, Allum WH, Stenning SP, Thompson JN, Van de Velde CJ, Nicolson M, Scarffe JH, Lofts FJ, Falk SJ, Iveson TJ, Smith DB, Langley RE, Verma M, Weeden S, Chua YJ, MAGIC Trial Participants. Perioperative chemotherapy vs surgery alone for resectable gastroesophageal cancer. N Engl J Med 2006; 355: 11-20 [PMID: 16822992 DOI: 10.1056/NEJMoa055531]
- 11 Al-Batran SE, Homann N, Pauligk C, Goetze TO, Meiler J, Kasper S, Kopp HG, Mayer F, Haag GM, Luley K, Lindig U, Schmiegel W, Pohl M, Stoehlmacher J, Folprecht G, Probst S, Prasnikar N, Fischbach W, Mahlberg R, Trojan J, Koenigsmann M, Martens UM, Thuss-Patience P, Egger M,



Block A, Heinemann V, Illerhaus G, Moehler M, Schenk M, Kullmann F, Behringer DM, Heike M, Pink D, Teschendorf C, Löhr C, Bernhard H, Schuch G, Rethwisch V, von Weikersthal LF, Hartmann JT, Kneba M, Daum S, Schulmann K, Weniger J, Belle S, Gaiser T, Oduncu FS, Güntner M, Hozaeel W, Reichart A, Jäger E, Kraus T, Mönig S, Bechstein WO, Schuler M, Schmalenberg H, Hofheinz RD; FLOT4-AIO Investigators. Perioperative chemotherapy with fluorouracil plus leucovorin, oxaliplatin, and docetaxel vs fluorouracil or capecitabine plus cisplatin and epirubicin for locally advanced, resectable gastric or gastro-oesophageal junction adenocarcinoma (FLOT4): a randomised, phase 2/3 trial. Lancet 2019; 393: 1948-1957 [PMID: 30982686 DOI: 10.1016/S0140-6736(18)32557-1]

- 12 Kodera Y, Yamamura Y, Shimizu Y, Torii A, Hirai T, Yasui K, Morimoto T, Kato T, Kito T. Metastatic gastric lymph node rate is a significant prognostic factor for resectable stage IV stomach cancer. J Am Coll Surg 1997; 185: 65-69 [PMID: 9208963 DOI: 10.1016/s1072-7515(97)00006-9]
- Zhao BW, Chen YM, Jiang SS, Chen YB, Zhou ZW, Li YF. Lymph Node Metastasis, a Unique 13 Independent Prognostic Factor in Early Gastric Cancer. PLoS One 2015; 10: e0129531 [PMID: 26154617 DOI: 10.1371/journal.pone.0129531]
- Smyth EC, Fassan M, Cunningham D, Allum WH, Okines AF, Lampis A, Hahne JC, Rugge M, 14 Peckitt C, Nankivell M, Langley R, Ghidini M, Braconi C, Wotherspoon A, Grabsch HI, Valeri N. Effect of Pathologic Tumor Response and Nodal Status on Survival in the Medical Research Council Adjuvant Gastric Infusional Chemotherapy Trial. J Clin Oncol 2016; 34: 2721-2727 [PMID: 27298411 DOI: 10.1200/JCO.2015.65.7692]
- 15 Wang WJ, Li HT, Yu JP, Su L, Guo CA, Chen P, Yan L, Li K, Ma YW, Wang L, Hu W, Li YM, Liu HB. Severity and incidence of complications assessed by the Clavien-Dindo classification following robotic and laparoscopic gastrectomy for advanced gastric cancer: a retrospective and propensity score-matched study. Surg Endosc 2019; 33: 3341-3354 [PMID: 30560498 DOI: 10.1007/s00464-018-06624-7]
- Jin LX, Sanford DE, Squires MH 3rd, Moses LE, Yan Y, Poultsides GA, Votanopoulos KI, Weber 16 SM, Bloomston M, Pawlik TM, Hawkins WG, Linehan DC, Schmidt C, Worhunsky DJ, Acher AW, Cardona K, Cho CS, Kooby DA, Levine EA, Winslow E, Saunders N, Spolverato G, Colditz GA, Maithel SK, Fields RC. Interaction of Postoperative Morbidity and Receipt of Adjuvant Therapy on Long-Term Survival After Resection for Gastric Adenocarcinoma: Results From the U.S. Gastric Cancer Collaborative. Ann Surg Oncol 2016; 23: 2398-2408 [PMID: 27006126 DOI: 10.1245/s10434-016-5121-7]
- 17 Tokunaga M, Tanizawa Y, Bando E, Kawamura T, Terashima M. Poor survival rate in patients with postoperative intra-abdominal infectious complications following curative gastrectomy for gastric cancer. Ann Surg Oncol 2013; 20: 1575-1583 [PMID: 23076557 DOI: 10.1245/s10434-012-2720-9]
- Wang S, Xu L, Wang Q, Li J, Bai B, Li Z, Wu X, Yu P, Li X, Yin J. Postoperative complications and 18 prognosis after radical gastrectomy for gastric cancer: a systematic review and meta-analysis of observational studies. World J Surg Oncol 2019; 17: 52 [PMID: 30885211 DOI: 10.1186/s12957-019-1593-9]
- 19 Yuan P, Wu Z, Li Z, Bu Z, Wu A, Wu X, Zhang L, Shi J, Ji J. Impact of postoperative major complications on long-term survival after radical resection of gastric cancer. BMC Cancer 2019; 19: 833 [PMID: 31443699 DOI: 10.1186/s12885-019-6024-3]
- 20 Hu Y, Huang C, Sun Y, Su X, Cao H, Hu J, Xue Y, Suo J, Tao K, He X, Wei H, Ying M, Hu W, Du X, Chen P, Liu H, Zheng C, Liu F, Yu J, Li Z, Zhao G, Chen X, Wang K, Li P, Xing J, Li G. Morbidity and Mortality of Laparoscopic Versus Open D2 Distal Gastrectomy for Advanced Gastric Cancer: A Randomized Controlled Trial. J Clin Oncol 2016; 34: 1350-1357 [PMID: 26903580 DOI: 10.1200/JCO.2015.63.7215]
- Lee JH, Son SY, Lee CM, Ahn SH, Park DJ, Kim HH. Morbidity and mortality after laparoscopic 21 gastrectomy for advanced gastric cancer: results of a phase II clinical trial. Surg Endosc 2013; 27: 2877-2885 [PMID: 23404155 DOI: 10.1007/s00464-013-2848-0]



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Observational Study

ORIGINAL ARTICLE

Diagnostic usefulness of selected proteases and acute phase factors in patients with colorectal adenocarcinoma

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Informed consent statement: All study participants, or their legal guardian, provided informed written consent prior to study enrollment.

Conflict-of-interest statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Data sharing statement: The raw data supporting the conclusions of this article will be made available

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Abstract

BACKGROUND

Uncontrolled growth and loss of control over basic metabolic functions, leading to invasive proliferation and metastases, are the salient traits of malignant tumors in general and colorectal cancer in particular. Invasion and metastases hinder effective tumor treatment. While surgical techniques and radiotherapy can be used to remove tumor focus, only chemotherapy can eliminate dispersed neoplastic cells. However, the efficacy of the latter method is limited in the advanced stages of the disease. Therefore, recognition of the mechanisms involved in neoplastic cell spreading is indispensable for developing effective therapies.

AIM

To use a number of biomarkers involved in cancer progression and identify a panel that could be used for effective early diagnosis.

METHODS

We recruited 185 patients with colorectal adenocarcinoma (98 men, 87 women with median age 63). Thirty-five healthy controls were sex and age-matched. Dukes' staging was as follows: A = 22, B = 52, C = 72, D = 39. We analyzed patients' blood serum before surgery. We determined: (1) Cathepsin B (CB) with Barrett's method (fluorogenic substrate); (2) Leukocytic elastase (LE) in a complex



by the authors, without undue reservation, to any qualified researcher.

STROBE statement: The authors have read the STROBE Statementchecklist of items, and the manuscript was prepared and revised according to the STROBE Statement-checklist of items.

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with alpha 1 trypsin inhibitor (AAT) using the immunoenzymatic MERCK test; (3) Total sialic acid (TSA) with the colorimetric periodate-resorcinol method; (4) Lipid-bound sialic acid (LASA) with the colorimetric Taut's method; and (5) The antitrypsin activity (ATA) employing the colorimetric test.

RESULTS

In patients, the values of the five biochemical parameters were as follows: CB = $16.1 \pm 8.8 \text{ mU/L}$, LE = $875 \pm 598 \mu \text{g/L}$, TSA = $99 \pm 31 \text{ mg\%}$, LASA = 0.68 ± 0.33 mg%, and ATA = $3211 \pm 1504 \text{ U/mL}$. Except for LASA, they were significantly greater than those of controls: CB = $11.4 \pm 6.5 \text{ mU/L}$, LE = $379 \pm 187 \mu g/L$, TSA = $71.4 \pm 15.1 \text{ mg\%}$, LASA = $0.69 \pm 0.28 \text{ mg\%}$, and ATA = $2016 \pm 690 \text{ U/mL}$. For CB and LASA, the differences between the four Dukes' stages and controls were not statistically significant. The inter-stage differences for CB and LASA were also absent. The receiver operating characteristic (ROC) analysis revealed the potential diagnostic value of CB, TSA, and ATA. The area under ROC, sensitivity, and specificity for these three parameters were: 0.85, 72%, 90%; 0.75, 66%, 77%; and 0.77, 63%, 84%, respectively. The sensitivity and specificity for the threeparameter panel CB-TSA-ATA were equal to 88.2% and 100%, respectively.

CONCLUSION

The increased value of CB, TSA, and ATA parameters are associated with tumor biology, invasion, and metastasis of colorectal cancer. The presented evidence suggests the potential value of the CB-TSA-ATA biochemical marker panel in early diagnostics.

Key Words: Colorectal cancer; Cathepsin B; Acute phase reactants; Colorectal adenocarcinoma; Acute phase factor

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Core Tip: We searched for biomarkers applicable to the early detection of colorectal adenocarcinoma. Five parameters were determined in sera of patients and healthy individuals: Cathepsin B activity, total sialic acids concentration, lipid-associated sialic acids concentration, elastase concentration, and alpha 1 antitrypsin activity. We performed receiver operating characteristic analysis for single and multiple parameters. While the sensitivity and specificity were not very high for single parameters, the combined analysis of cathepsin B, alpha 1 antitrypsin, and total sialic acids concentration yielded 88% sensitivity and 100% specificity. We believe that this set of markers can be useful in clinical practice.

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INTRODUCTION

The process of neoplastic invasion consists of two main stages: Penetration of tissues surrounding cancer and creation of metastases in places distant from the original location. During migration, cancer-transformed cells encounter anatomical barriers: The basement membrane and connective tissue. Cathepsins play an important role in the process of overcoming them.

Under normal conditions, cathepsins do not occur extracellularly or appear outside the cell only in small quantities. Their main function is to participate in processes related to the "turn-over" of endogenous proteins and degradation of exogenous proteins absorbed in the process of endocytosis[1-4]. In smaller concentrations, *e.g.*, in extralysosomal spaces, the enzymes can catalyze – by means of limited proteolysis – posttranslational processes of conversion of many peptides and proteins, including



growth factors and hormones such as albumins, insulins, endorphins, and enkephalins [5,6]. The release of cathepsins from cancer cells and their expression in the plasma membrane of cancer cells facilitates the dissolution of the basement membrane and participation in the proteolytic metastatic cascade. In clinical practice, a significant increase in the activity of cathepsin B (CB) is observed in the serum of patients with malignant tumors. This phenomenon occurs regardless of the location of the tumor and is closely related to the severity of ovarian, cervical, breast, laryngeal and colorectal cancer. It was also found that the activation of CB, which takes place with the participation of elastase coming from the neutrophil intumescence and tumor tissue, has an impact on its invasiveness and metastatic capacity. Studies have also confirmed that an increase in cathepsin expression in colorectal tumor tissue homogenates may be a sensitive marker for cancer progression[7-16].

Leukocytic elastase (LE) belongs to the group of serine proteases. It is located mainly in the azurophilic granules of neutrophils, where it is an active component of the phagocytic system along with other hydrolyses and reactive oxygen species. The enzyme is also cytochemically detected in the nuclear membrane, the Golgi complex, endoplasmic reticulum, and mitochondria[17,18]. It also participates in remodeling and tissue repair processes and modulates the activity of cytokines and their receptors (e.g., mitogen-activated protein kinase 3). It can degrade elements of connective tissue by hydrolysis of elastin, various types of collagen, and other extracellular matrix proteins such as fibronectin, laminin, or proteoglycans. The physiological regulation of the activity and prevention of potentially destructive effects of elastase in pathological states is caused by protein inhibitors present in the blood serum. They include alfa 1 trypsin inhibitor (AAT), alpha 2-macroglobulin (alpha-2-MG), a secretory leukocytic protease inhibitor (SLPI), and elaphin[19-21]. In a healthy organism, LE remains in balance with them, and after the secretion from the cells, it is directly bound by inhibitors. The inhibitor molecules are thermally very stable and do not undergo proteolysis. Increased levels of LE in blood plasma, most often determined as LE-AAT complexes, have been found in many associated inflammatory conditions and are therefore considered acute-phase factors. Many authors regard the concentration of the above complexes to be a measure of the activity of the inflammatory process itself and be a marker of stimulation of neutrophils in the inflammatory focus. Lowering the level of AAT, caused by genetic or environmental factors, enables uncontrolled elastase activity and leads to many serious pathological conditions. This is caused by the enzyme-inhibitor imbalance and overexpression of the enzyme or a decrease in inhibitor concentration.

Sialic acid (N-acetyloneuraminic acid, NANA) is an organic compound from the sugar group, a derivative of neuraminic acid. It is a sugar component of glycoproteins and glycolipids. It occupies a terminal position in carbohydrate glycoprotein residues and has an important function in cell physiology as well as in the metabolism and maintenance of the proper concentration of glycoproteins in serum. It is also part of the ligand for the selectin and lectin receptors on leukocytes, T lymphocytes, platelets, and endothelium. It plays a key role in the immune response and hemostasis[22].

In the blood of patients suffering from metabolic disorders (e.g., diabetes mellitus, atherosclerosis), a clear increase in the glycoprotein fraction is observed, while the sialic acid content remains low. In the case of cancer, the level of glycoproteins that contain normal or increased amounts of sialic acid increases in the blood. Increased sialisation is associated with the development of a neoplastic tumor and malignant cell metastasis. An increase in sialic acid concentration is observed in the case of malignant melanoma, lung, larynx, breast, ovary, prostate, liver, or colorectal cancer. It has been shown that the sialyltransferase activity in blood serum taken from people with cancer is increased and reaches its highest concentration at advanced stages of cancer development. The main areas of protein glycosylation include the endoplasmic reticulum and Golgi apparatus^[23]. During the transformation of normal cells into cancer cells, there are significant differences in the biosynthesis of the sugar parts of proteins and membrane lipids. These changes are primarily of a qualitative nature. The external part of the plasma membrane of cancer cells has an increased number of sialic acid molecules compared to normal cells. Glycoproteins rich in sialic acids are more frequently found in case of metastatic cancer. The increased level of sialoglycoproteins and sialoglycolipids in neoplasms is mainly due to increased disintegration of cancer cells, increased synthesis and secretion of glycoconjugates (glycoproteins and glycolipids) containing sialic acid[24-32].

Recently, many works have been published on the activity of proteolytic enzymes and their inhibitors in blood serum. However, they do not solve all the problems related to the diagnostic and prognostic usefulness of these parameters in neoplastic diseases.



In this study, a statistically consistent group of colorectal cancer patients was collected: Women and men. The division of colorectal cancer into the colon and rectal cancer, histopathological criteria (adenocarcinoma), a clinical division system based on Dukes' cancer staging were considered.

The study presented below has both basic and diagnostic-clinical nature. The results of the study may be used in the future in the complementary diagnosis of colorectal adenocarcinoma to determine the severity of the disease and in further monitoring of patients under outpatient control and prognosis.

MATERIALS AND METHODS

Patients

One hundred and eighty-five patients were recruited from the Lower Silesian Oncology Center and the Provincial Specialist Hospital in Wroclaw. The study material presented in this paper was blood serum from patients with colorectal adenocarcinoma. The patients' blood serum was examined before surgery.

The examined patients were evaluated in terms of their age, sex, location of neoplastic lesions (colon, sigmoid colon, rectum), histopathological differentiation of neoplastic cells (G), and their clinical stages were based on the Dukes' classification. The characteristics of the examined patients are summarized in Table 1.

Biochemical measurements

The following tests were performed in blood serum according to the methods given below: (1) CB was determined with the use of fluorogenic substrate using the Barrett method[33]; (2) LE in a complex with AAT was determined immunoenzymatically using the MERCK test; (3) Total sialic acid (TSA) was determined colorimetrically using the periodate-resorcinol method, according to Jourdian et al[34]; (4) Lipid-bound sialic acid (LASA) was determined colorimetrically using Tautu *et al*[35]; and (5) The antitrypsin activity (ATA) in blood plasma (in the study referred to as antitrypsin capacity – ATA) was determined colorimetrically against trypsin using the method proposed by Warwas et al[36] and Dietz et al[37].

Statistical analysis

The examined continuous features were characterized by the distribution parameters of these features, i.e., mean value (M), standard deviation (SD), and the number of patients (*n*). For the analysis of the statistical material, the following were used: For continuous features – single-factor analysis of variance (ANOVA), using Tukey's post hoc tests and multi-factor analysis of variance (MANOVA). The t-Student's t-test was also used for dependent samples. For features deviating from the normal distribution, which were also characterized by a median value, non-parametric tests were used: For independent samples - the Mann-Whitney U test, and for dependent samples - the Wilcoxon test. For categorized or dichotomous features, the χ^2 test and the nonparametric Kruskal-Wallis test were used. The relationship between these features was also studied by determining the Spearman correlation coefficient. The receiver operating characteristic (ROC) curve method was used to determine the threshold values of clinical markers (continuous variables) with optimal precision. The significance threshold *P* for all statistical tests was set to 0.05. The statistical analysis was conducted using Statistica v. 10.

RESULTS

Table 2 presents the results of biochemical parameters examined, their M, SD, n, including the group of colorectal cancer patients and the control group of healthy individuals.

In the group of patients with colorectal cancer, differences were observed in the values of examined biochemical parameters in blood serum compared to the control group. Despite clear changes in the levels and activity of the studied factors, not in all groups, these differences were statistically significant. Relevance was observed between patient groups suffering from colorectal cancers and the control group for CB, LE, TSA, and ATA.

Table 3 presents statistically significant differences between the examined groups of patients according to Dukes' classification and the control group. There were no



Table 1 Characteristics of patients with colorectal cancer					
Number of patients with colorectal cancer	185				
Age, median (age range)	63 (18-86)				
Sex, n (%)					
Men	98 (55)				
Women	87 (45)				
Anatomical location, n (%)					
Colon	77 (42)				
Sigmoid colon	37 (20)				
Rectum	71 (38)				
Histological differentiation of cells, <i>n</i> (%)					
G1	8 (4)				
G2	103 (56)				
G3	74 (40)				
Division of patients according to Dukes' classification, <i>n</i> (%)					
Α	22 (12)				
В	52 (28)				
C	72 (39)				
D	39 (21)				
Number of patients in the control group	35				
Age, median (age range)	61 (19-85)				
Sex, n (%)					
Men	19 (54)				
Women	16 (46)				

significant differences for CB and LASA (P > 0.05), while the differences for other parameters were: LE (*P* < 0.001), TSA (*P* < 0.001), ATA (*P* < 0.01).

Table 4 presents a comparison of statistically significant differences between individual parameters in the patient groups (A, B, C, D, according to Dukes' classification) and control group. There were statistically significant differences for all groups except for group A. Statistically significant differences between patient groups were also being observed. For LE, the most statistically significant difference was observed between groups C and D (P < 0.002). For TSA, the difference was most strongly pronounced for A and D (P < 0.05). There were no stage differences for ATA.

In Table 5 the correlations between in main parameters are summarized. The values of all the above-mentioned correlation coefficients were positive except for CB and ATA and LASA and ATA. The highest correlations were observed between CB and ATA; LE and TSA; TSA and ATA (P < 0.001).

No statistically significant differences in the examined parameters in respect to histopathological differentiation of tumor cells (G1, G2, G3) were observed (data not shown).

The examined biochemical factors in the blood serum were also examined using ROC analysis. The analyses are summarized in Figure 1. The highest diagnostic value was observed for a single value for CB in blood serum, with a threshold value of 11.22 mU/L. The sensitivity of the method was 72%, and its specificity was 90%. Then, respectively: for TSA, with a threshold value of 75.34 mg% - 66% and 77%; for LASA with a threshold value of 0.562 mg% - 80% and 41%; for LE with a threshold value of 543 μ g/L – 49% and 89% and ATA in blood serum with a threshold value of 2400 U/mL - 63% and 84%.

The analysis of the ROC curves of the two associated parameters has shown that the connection between CB and TSA gives a sensitivity of 73% and a specificity of 100%, with a value under the curve reaching 95%.



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Sebzda T et al. Proteases usefulness in colorectal adenocarcinoma diagnosis

Table 2 Char	Table 2 Characteristics of the tested biochemical parameters in blood serum in all patients and in the control group							
No.	Biochemical parameters tested	Groups	ABCD	К	Significance level <i>P</i> value			
1	CB, mU/L	М	16.1	11.4	< 0.050			
		n	185	35				
		SD	8.8	6.5				
2	LE, µg/L	М	875.1	379.1	< 0.001			
		n	51	30				
		SD	597.9	187.3				
3	TSA, mg%	М	98.9	71.4	< 0.001			
		n	71	31				
		SD	30.8	15.1				
4	LASA, mg%	М	0.68	0.69	NS			
		n	68	35				
		SD	0.33	0.28				
5	ATA, U/mL	М	3211.4	2015.9	< 0.001			
		n	74	29				
		SD	1504.1	689.6				

ABCD: All patients with colorectal adenocarcinoma; ATA: Antitrypsin activity; CB: Cathepsin B; K: Control group; LASA: Lipid-bound sialic acid; LE: Leukocytic elastase; M: Mean value; NS: Not significant; TSA: Total sialic acid.

Combining three parameters for the ROC curve: CB, TSA, and ATA, in blood serum, gives a sensitivity of 88% and a specificity of 100%, respectively, with a value of area under the curve of 95%.

In conclusion, ROC analysis has a high diagnostic value and can be helpful, especially in the combined analysis (biomarker panel determination).

DISCUSSION

The ability of the tumor to invade and metastasize is associated, among other pathogenic issues, with an increase in the expression of cysteine peptides. The source of these enzymes may be, in addition to the neoplastic tissue, neutrophils infiltrating the tumor. It is believed that CB determination may be helpful in the diagnosis and monitoring of colon and rectal cancer therapy[38-40].

In our study, the average CB activity determined in the blood serum of patients with colorectal cancers using a synthetic Z-Arg-Arg-N-MC substrate was about 1.5 times higher than that of healthy individuals (P < 0.05). Statistically significant results were obtained in the group of patients with different degrees of clinical advancement (ABCD: P < 0.05) (Table 2 and Figure 1A). On the other hand, CB did not statistically significantly differentiate patients according to Dukes' staging of colorectal cancer (A, B, C, D), although the average values in patients exceeded the average values in the control group (Table 3). Among other parameters, CB correlated only with ATA (P < 0.001) (Table 5).

The activity of CB in serums of patients with colon and rectal cancers was also studied by other authors. Dufek *et al*[41] observed a fivefold increase in this parameter. It was found that the CB activity in patients with colon and rectal cancers was significantly higher than in patients with polyps and in healthy individuals. This difference may be due to the use of the other substrate (Z-Ala-Arg-Arg-N-MC), which may also have been hydrolyzed by other proteases. These studies have also shown a high level of alkali-stable form of CB. In people with mild lesions (polyps) and in healthy people, the level of this form did not reach the threshold of determination. After the effective treatment, CB levels decreased significantly and increased again in case of metastases or resistant chemotherapy.

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Table 3 Characteristics of biochemical parameters in patients' blood serum in relation to the severity according to the Dukes' classification

No.	Biochemical parameters tested	Groups	Α	В	С	D	К	Significance level <i>P</i> value
1	CB, mU/L	М	19.9	15.4	16.8	15.5	11.4	NS
		n	22	52	72	39	35	
		SD	19.3	9.4	8.4	7.6	6.5	
2	LE, µg/L	М	386.1	929.9	602.1	1129.5	379.1	< 0.001
		n	5	16	15	15	30	
		SD	190.2	637.5	389.6	651.7	187.3	
3	TSA, mg%	М	67.8	100.8	92.9	107.2	71.4	< 0.001
		n	6	21	24	20	31	
		SD	7.4	39.3	25.1	26.3	15.1	
4	LASA, mg%	М	0.44	0.8	0.6	0.6	0.7	NS
		n	6	21	24	21	37	
		SD	0.12	0.51	0.23	0.20	0.31	
5	ATA, U/mL	М	3100.0	2925.0	3152.1	3576.2	2015.9	< 0.010
		n	2	21	24	21	29	
		SD	424.3	1294.5	1684.7	1548.8	689.6	

Dukes' classification system for colorectal cancer: A, B, C, D: All patients with colorectal adenocarcinoma; ATA: Antitrypsin activity; CB: Cathepsin B; K: Control group; LASA: Lipid-bound sialic acid; LE: Leukocytic elastase; M: Mean value; NS: Not significant; TSA: Total sialic acid.

Kos *et al*[40] also observed the fivefold increase in CB concentration using immuneenzymatic method. The CB level correlated well with stage C and D of Dukes' staging system. As in our research, no correlation between CB and age or gender was observed. However, it should be stressed that the antibodies used in this method detected both the active and non-active precursor form of this enzyme and their complexes with inhibitors, such as cystatins.

Similar studies on the proteolytic activity of blood serum with colon and rectal cancer were performed by Amiguet *et al*[42]. Similar to our research, the proteolytic activities of CB and elastase, were determined. Patients in Dukes' B and D stages were examined. The activities of the examined proteases were increased in relation to the control group of healthy individuals. In Dukes' stage D, the increase in CB levels was directly proportional to the weight of the tumor. In metastatic carcinomas, the increase in CB was accompanied by an increase in AAT concentration.

Padilla *et al*[43] showed, with the use of the immunoreactive method, that CB levels in colorectal cancer patients were different from the control group.

Clinical and pathological evaluation of patients with the use of serum CB and cathepsin D, based on the TNM system before and after the operation, was performed by Skrzydlewska *et al*[44]. The CB activity before the cancer tumor resection was significantly higher. However, in relation to the control group, both before and after the procedure, the CB activity was approximately 8.4 times lower. The authors concluded that the postoperative level of CB was associated with the involvement of the surrounding lymph nodes and higher when not accompanied by lymph node involvement. However, a relatively non-specific Z-Arg-pNA substrate was used to test the CB activity, and thus the observations might have been biased by the activity of other proteases.

Zore *et al*[45] examined CB levels in the complex with cystatin C using the ELISA method. Their observations show that CB in the Dukes' CD stage was significantly lower than in the AB stage (P = 0.02). Our study did not observe any significant differences between these groups (the results are not presented). The inhibitory capacity of cystatin C does not compensate for the increase in CB levels in patients suffering from colorectal cancers. This supports the hypothesis that inhibitory capacity might have been impaired during colorectal cancer progression.

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Table 4 Statistical analysis of differences in the activity or concentration of examined parameters in serum of patients with adenocarcinoma and the control group

Patients' Groups							
LE	Α	В	C	D	К		
А	NS	NS	NS	NS	NS		
В	NS	NS	P < 0.05	NS	P < 0.001		
С	NS	P < 0.05	NS	P < 0.002	P < 0.004		
D	NS	NS	<i>P</i> < 0.002	NS	P < 0.001		
К	NS	P < 0.001	P < 0.004	NS	NS		
TSA	Α	В	С	D	K		
А	NS	NS	NS	NS	NS		
В	NS	NS	NS	NS	P < 0.001		
С	NS	NS	NS	NS	<i>P</i> < 0.003		
D	P < 0.05	NS	NS	NS	P < 0.001		
К	NS	P < 0.001	P < 0.003	P < 0.001	NS		
ATA	Α	В	С	D	K		
А	NS	NS	NS	NS	NS		
В	NS	NS	NS	NS	P < 0.02		
С	NS	NS	NS	NS	P < 0.002		
D	NS	NS	NS	NS	P < 0.001		
К	NS	<i>P</i> < 0.02	<i>P</i> < 0.002	P < 0.001	NS		

Significance levels of differences in the expected values of the analyzed biochemical parameters in patients divided into colorectal cancer patients (Dukes' stage A, B, C, D) and control group (K). The values of statistical significance – P were provided. A, B, C, D: All patients with colorectal adenocarcinoma; ATA: Antitrypsin activity; K: Control group; LE: Leukocytic elastase; NS: Not significant; TSA: Total sialic acid.

Table 5 Statistical analysis of correlations between the biochemical parameters examined in blood serum in all patients								
Studied parameters	CB, mU/L	LE, µg/L	TSA, mg%	LASA, mg%	ATA, U/mL			
CB, mU/L					-0.24 (187), $P < 0.001$			
LE, µg/L			0.37 (140), $P < 0.001$	0.17~(141), P < 0.05				
TSA, mg%		0.37 (140), $P < 0.001$		0.19 (212), $P < 0.01$	0.33 (209), $P < 0.001$			
LASA, mg%		0.17 (141), $P < 0.05$	0.19~(212), P < 0.006		-0.14 (210), $P < 0.05$			
ATA, U/mL	-0.24 (187), $P < 0.001$		0.33 (209), $P < 0.001$	-0.14 (210), $P < 0.05$				

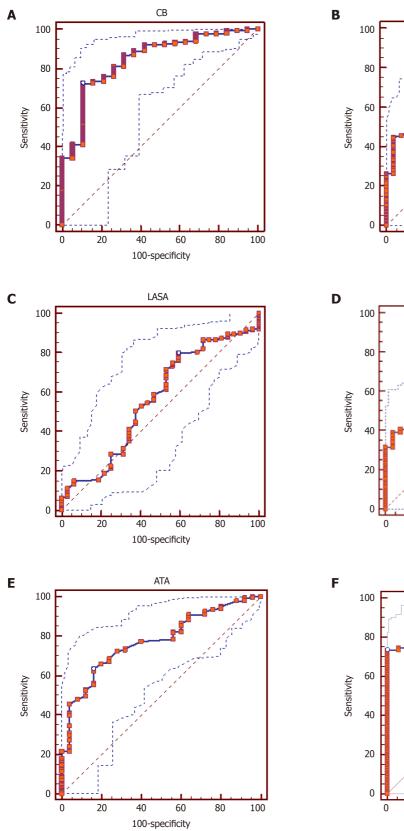
Values of linear correlation coefficients between concentrations or activities of the studied biochemical parameters (number of the examined). Description of other parts as in Table 2. ATA: Antitrypsin activity; CB: Cathepsin B; LASA: Lipid-bound sialic acid; LE: Leukocytic elastase; TSA: Total sialic acid.

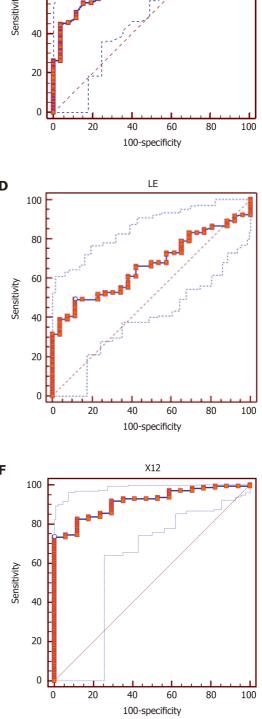
> Cysteine proteases - CB and cathepsin L levels in blood serum in patients with colorectal cancer were studied by Herszényi et al[46], who used the ELISA method for this purpose. CB correlated with the progressive Dukes' scale, reaching a 2.3 times higher level in patients compared to the control group. Analysis of the ROC curve confirmed the diagnostic importance of the examined factors, including CB. The sensitivity and specificity in the ROC analysis of CB were similar to the results obtained in our study (72 and 89% respectively vs 82 and 88%, and the areas under the ROC curve 0.85 vs 0.87), thus confirming the high diagnostic value of the studied parameter. Comparing CB with other biochemical parameters such as TSA and ATA in the ROC analysis results in an even higher level of sensitivity and specificity. Also, other researchers noticed that proteolytic enzymes are excellent indicators for colorectal cancers, often better than the commonly used tumor markers^[47].



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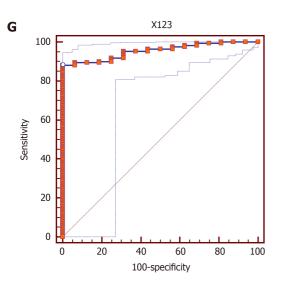


Figure 1 Diagram of receiver operating characteristic curve. A: For cathepsin B (CB). Threshold value of > 11.22 mU/L yielded sensitivity of 72.3% and specificity of 90%. Area under the receiver operating characteristic curve = 0.85; B: For total sialic acid (TSA). Threshold value > 75.34 mg% yielded sensitivity of 65.8% and specificity of 76.9%. Area under the receiver operating characteristic curve = 0.75; C: For lipid-bound sialic acid (LASA). Threshold value > 0.739 mg% yielded sensitivity of 79.7% and specificity of 40.6%. Area under the receiver operating characteristic curve = 0.56; D: For leukocytic elastase (LE). Threshold value > 543 μ g/L yielded sensitivity of 49.2% and specificity of 88.5%. Area under the receiver operating characteristic curve = 0.56; E: For antitrypsin activity (ATA). Threshold value > 2400 U/mL yielded sensitivity of 63.2% and specificity of 84.0%. Area under the receiver operating characteristic curve = 0.77; F: For the combined two biochemical parameters. Threshold value > 0.4757 yielded sensitivity of 73.4% and specificity of 100%. Area under the receiver operating characteristic curve = 0.91; G: The three parameters combined (X123): CB, TSA, and ATA. Threshold value > 1.3457 yielded sensitivity of 88.2% and specificity of 100%. Area under the receiver operating characteristic curve = 0.95.

The imbalance of protease/inhibitors ratio is particularly relevant for LE and AAT involvement in the pathogenesis of cancer. The level of AAT, an acute-phase protein, increases in cancer patients in response to the increased levels of proteolytic enzymes released from leukocytes into circulation. In patients, despite an increase in their levels, the functional activity of the inhibitor decreases, thus disturbing the LE/AAT balance. This imbalance is further exacerbated by the fact that membrane forms of proteases, such as CB or elastase are more resistant to these inhibitors. Moreover, LE has itself ability to degrade those inhibitors[48]. A disturbed balance between LE and AAT may be associated with an increased risk of liver, cholecystitis, bladder, lymphoma, or lung cancer[49-55]. Apart from AAT, other protein inhibitors present in the blood serum are also responsible for the physiological regulation of LE: α 2MG, SLPI, or elaphin. SLPI, as it is clear from the work by Sugino *et al*[54], performed in various types of cancer, including colorectal cancer, has a dual effect. On the one hand, it suppresses the invasion of neoplastic cells, and on the other hand, it promotes metastasis that transmits through blood circulation.

The inclusion of serum ATA and LE, as indicated by the results of our research, may be a factor informing about the balance of serine proteases.

LE in all listed patient groups according to the Dukes' classification: Combined ABCD stages and A, B, C, D stages separately (Table 2 and Figure 1D), shows a clear statistically significant difference when compared to the control group. Its activity reached the highest values in groups B and D. It is possible that it is associated with subsequent stages of cancer spread and the presence of metastatic foci rich in granulocytic intumescence, increased elastase-induced adhesion of cancer cells to the endothelium of vessels, or through mitogenic activity[56,57].

The different activity of serum elastase, in successive stages of the disease according to the Dukes' classification eliminates the possibility of using this parameter for early diagnosis in colorectal cancers. In addition, serum elastase is characterized by relatively low sensitivity in the ROC analysis (49%) (Figure 1D). On the other hand, some authors have found it as a putative diagnostic biomarker and also a potential therapeutic target[58].

AAT is a blood plasma protein belonging to α_1 -globulin fraction, one of the strongest inhibitors of circulating serine proteases (serpins). It is also an acute-phase protein, synthesized mainly in the liver but also by macrophages. AAT has the ability to inactivate many proteolytic enzymes, but its most important effect is the inactivation of LE released by neutrophils as a result of an inflammatory reaction.

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In our study, statistically significant (P < 0.001) elevated ATA levels were observed in the cumulative group of patients, regardless of the Dukes' classification stage (ABCD) in relation to the control group. In the B, C, and D stages, in relation to the control, these levels were highly statistically significant (P < 0.001; P < 0.004; P < 0.001). Also, ATA differentiated very well patients with Dukes' stages B against C and C against D (P < 0.05 and P < 0.002, respectively). In our ROC analysis, with the cut-off value > 2400 U/mL, the ATA value reached the sensitivity of 63% and specificity of 84%, respectively. The listed sensitivity and specificity parameters are lower than in the ROC analysis for CB (72% and 90%, respectively).

There is little clinical work on the contribution of AAT to the diagnosis or monitoring of the treatment of colorectal cancer [59-64]. Yüceyar et al [65] and Gallardo-Valverde *et al*[66] did not find relationship between the AAT and the severity of colorectal cancer and showed a statistically significant correlation between AAT and other biochemical factors such as acute-phase protein, carcinoembryonic antigen (CEA), or tumor-associated trypsin inhibitor (TATI).

Bernacka et al[67] studied plasma levels of AAT in patients with gastrointestinal cancers: Stomach and colorectal cancers of adenocarcinoma type. This marker did not differentiate colorectal cancer patients in terms of local and metastatic lesions. However, it had the highest level in stage C and differentiated patients in terms of the histological degree of tumor stage (G). The author postulates that AAT levels are associated with increased production by liver cells in response to the increased release of lysosomal proteases of tumor cells or from mononuclear inflammatory tumor-infiltrating cells. It seems that tumor cells may be the third source of antiproteases.

Ward *et al*[68], using proteomic profiling, identified AAT as having the potential to classify the colorectal patients with 95% sensitivity and 91% specificity.

Interesting conclusions were drawn from the work of Bujanda et al[64], who studied a group of 42 colorectal cancer patients using combined AAT, matrix metalloproteinase 7 (MMP-7), urokinase-type plasmin activator receptor (uPAR), and cyclooxygenase-2 (COX-2). Compared to the control group, AAT levels were about 1.4 times higher at stages B and C, and the AAT level was high and reached its value under the ROC curve (0.88). The above results are similar to those obtained in our study. The level under the ROC curve was 0.77. In patients in stages B and C, ATA levels were 1.5 and 1.6 times higher, respectively, than in the control group. AAT has a promising diagnostic profile and, most importantly, at the early stages of colorectal cancer.

The neoplastic process is associated not only with the activation of the cascade system of proteolytic enzymes, the activation of which mainly takes place with the participation of CB and LE, but also induces the activation of acute-phase proteins.

The interest in the sialic acids (TSA, LASA) as markers useful in diagnosing and monitoring the course of many diseases, including colorectal cancer, is reflected in the publications briefly reviewed in[69-72]. Increased levels of sialic acid expression are associated with changes in biosynthesis and posttranslational processes of acute-phase proteins glycosylation in the liver. This phenomenon is associated with increased expression of sialyltransferases by cancer cells[73]. The mechanism of increased TSA levels in serum takes the following into account: (1) Spontaneous release of the compounds from the surface of cancer cells; (2) Increase in concentration and/or glycosylation of serum glycoproteins; and (3) Secondary inflammatory reaction associated with the increase in acute-phase proteins[32,74]. Increased sialyltransferases activities, observed in cancer cells, results in increased glycoprotein secretion as well as the secretion of cell membrane components into the culture medium. The cancer cell hypoxia may also contribute to the above [75]. Increased sialisation of glycosphingolipids leads to abnormal adhesion and a disturbed premembranous signal exchange [75]. Determination of sialic acids is a laboratory marker of many pathological lesions. A significant increase in serum levels of sialic acids was observed in many malignant diseases^[76]. Elevated levels of TSA or LASA were observed in malignant melanoma, lung, breast, ovary, and laryngeal cancers [28,32,77], as well as in colorectal cancer [26, 31,70,78-82].

In our study, TSA in blood serum was elevated and statistically significantly different from the control group. The above observation concerns the cumulative group of patients (ABCD: P < 0.001) as well as individual groups (B, C, D in relation to the control: P < 0.001; P < 0.003; P < 0.001 respectively). The lowest level among patients was found in group A, and the highest in group D. The above clearly shows differences between particular patient groups. TSA was only less effective than ATA. The TSA level moderately but statistically significantly correlates with LE and ATA. The results of the ROC analysis are interesting. With a cut-off value for TSA > 75.34mg%, the sensitivity of the method was 66%, and its specificity was 77%. In ROC analysis TSA clearly benefits when combined CB with the same cut-off values)



sensitivity 74% and specificity 100% were obtained. The sensitivity further increases if we also take the ATA into account, which is also studied by the authors and reflects the AAT level. In the TSA test, the main component determined is the sialic acid associated with proteins. The level of sialic acids increases in the sera of cancer patients as a result of increased concentration of acute-phase proteins rather than gangliosides from decaying cancer cells[65,74,75,83].

CONCLUSION

We have found a statistically significant increase in the CB activity in the blood serum of the examined individuals suffering from colorectal cancers. The highest CB level was observed in Dukes' stage A patients, and in stages B, C and D, it was lower and comparable to each other.

Concordantly, the following increased concentrations in blood serum of investigated markers were observed: LE, TSA, and ATA. According to Dukes' classification, these values were statistically significantly increased with respect to the control group, gradually rising from the A to D stage.

The ROC analysis showed the high diagnostic value of CB, TSA, ATA determinations in blood serum, both in the single and combined analysis (biomarker panels) with two biochemical parameters: CB and TSA, and with three parameters: CB, TSA, and ATA. The above results suggest high diagnostic usefulness in determining these 2 or 3 combined parameters in relation to single determinations, obtaining sensitivity and specificity of 88.2% and 100% for three parameters.

ARTICLE HIGHLIGHTS

Research background

Recognition of the mechanisms involved in neoplastic cell spreading is indispensable for the early diagnosis and detection of colorectal cancer.

Research motivation

Colorectal cancer is the third most common type of cancer, making up about 10% of all cases. In 2018, there were 1.09 million new cases and 551000 deaths from the disease. Consequently, early diagnosis of colorectal cancer remains a significant medical and economic problem.

Research objectives

Using several biomarkers involved in cancer progression, we have tried to identify a panel that could be used for effective early diagnosis.

Research methods

Before surgery, we analyzed the blood serum of 185 patients with colorectal cancer and determined: Cathepsin B (CB), leukocytic elastase (LE), total sialic acid (TSA), lipid-bound sialic acid (LASA), and antitrypsin activity (ATA).

Research results

The receiver operating characteristic analysis revealed the potential diagnostic value of CB, TSA, and ATA. The sensitivity and specificity for the three-parameter panel CB-TSA-ATA were equal to 88.2% and 100%, respectively.

Research conclusions

The increased value of CB, TSA, and ATA parameters are associated with tumor biology, invasion, and metastasis of colorectal cancer.

Research perspectives

The presented evidence suggests the potential diagnostic and prognostic value of the CB-TSA-ATA biochemical marker panel.

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REFERENCES

- Guinec N, Dalet-Fumeron V, Pagano M. "In vitro" study of basement membrane degradation by the 1 cysteine proteinases, cathepsins B, B-like and L. Digestion of collagen IV, laminin, fibronectin, and release of gelatinase activities from basement membrane fibronectin. Biol Chem Hoppe Seyler 1993; 374: 1135-1146 [PMID: 8129860 DOI: 10.1515/bchm3.1993.374.7-12.1135]
- Korzus G, Turyna B. [Cysteine proteinases and their endogenous inhibitors]. Postepy Biochem 1988; 2 34: 23-46 [PMID: 3064060]
- Calkins CC, Sameni M, Koblinski J, Sloane BF, Moin K. Differential localization of cysteine 3 protease inhibitors and a target cysteine protease, cathepsin B, by immuno-confocal microscopy. J Histochem Cytochem 1998; 46: 745-751 [PMID: 9603786 DOI: 10.1177/002215549804600607]
- Olszewska D, Drewa T, Makarewicz R, Drewa J, Woźniak A, Maciak R. [Significance of cathepsin B and D in physiologic and pathologic processes]. Pol Merkur Lekarski 2001; 10: 65-70 [PMID: 11320558]
- Qian F, Chan SJ, Gong QM, Bajkowski AS, Steiner DF, Frankfater A. The expression of cathepsin B and other lysosomal proteinases in normal tissues and in tumors. Biomed Biochim Acta 1991; 50: 531-540 [PMID: 1801719]
- Howie AJ, Burnett D, Crocker J. The distribution of cathepsin B in human tissues. J Pathol 1985; 6 145: 307-314 [PMID: 3889245 DOI: 10.1002/path.1711450404]
- 7 Nomura T, Katunuma N. Involvement of cathepsins in the invasion, metastasis and proliferation of cancer cells. J Med Invest 2005; 52: 1-9 [PMID: 15751268 DOI: 10.2152/jmi.52.1]
- Talieri M, Papadopoulou S, Scorilas A, Xynopoulos D, Arnogianaki N, Plataniotis G, Yotis J, Agnanti N. Cathepsin B and cathepsin D expression in the progression of colorectal adenoma to carcinoma. Cancer Lett 2004; 205: 97-106 [PMID: 15036666 DOI: 10.1016/j.canlet.2003.09.033]
- 9 Sheahan K, Shuja S, Murnane MJ. Cysteine protease activities and tumor development in human colorectal carcinoma. Cancer Res 1989; 49: 3809-3814 [PMID: 2544258]
- Troy AM, Sheahan K, Mulcahy HE, Duffy MJ, Hyland JM, O'Donoghue DP. Expression of 10 Cathepsin B and L antigen and activity is associated with early colorectal cancer progression. Eur J Cancer 2004; 40: 1610-1616 [PMID: 15196548 DOI: 10.1016/j.ejca.2004.03.011]
- 11 Shuja S, Sheahan K, Murnane MJ. Cysteine endopeptidase activity levels in normal human tissues, colorectal adenomas and carcinomas. Int J Cancer 1991; 49: 341-346 [PMID: 1917131 DOI: 10.1002/ijc.2910490305]
- Kruszewski WJ, Rzepko R, Wojtacki J, Skokowski J, Kopacz A, Jaśkiewicz K, Drucis K. 12 Overexpression of cathepsin B correlates with angiogenesis in colon adenocarcinoma. Neoplasma 2004; 51: 38-43 [PMID: 15004658]
- 13 Campo E, Muñoz J, Miquel R, Palacín A, Cardesa A, Sloane BF, Emmert-Buck MR. Cathepsin B expression in colorectal carcinomas correlates with tumor progression and shortened patient survival. Am J Pathol 1994; 145: 301-309 [PMID: 7519824]
- 14 Berquin IM, Sloane BF. Cathepsin B expression in human tumors. Adv Exp Med Biol 1996; 389: 281-294 [PMID: 8861022 DOI: 10.1007/978-1-4613-0335-0 35]
- Hirai K, Yokoyama M, Asano G, Tanaka S. Expression of cathepsin B and cystatin C in human 15 colorectal cancer. Hum Pathol 1999; 30: 680-686 [PMID: 10374777 DOI: 10.1016/s0046-8177(99)90094-1
- Adenis A, Huet G, Zerimech F, Hecquet B, Balduyck M, Peyrat JP. Cathepsin B, L, and D activities 16 in colorectal carcinomas: relationship with clinico-pathological parameters. Cancer Lett 1995; 96: 267-275 [PMID: 7585467 DOI: 10.1016/0304-3835(95)03930-u]
- Travis J, Dubin A, Potempa J, Watorek W, Kurdowska A. Neutrophil proteinases. Caution signs in 17 designing inhibitors against enzymes with possible multiple functions. Ann NY Acad Sci 1991; 624: 81-86 [PMID: 1905897 DOI: 10.1111/j.1749-6632.1991.tb17008.x]
- Watorek W, Farley D, Salvesen G, Travis J. Neutrophil elastase and cathepsin G: structure, function, 18 and biological control. Adv Exp Med Biol 1988; 240: 23-31 [PMID: 3266707 DOI: 10.1007/978-1-4613-1057-0 3
- Imamura T, Maeda H. [Alpha1-proteinase inhibitor: structure and functions]. Nihon Rinsho 2005; 63 19 Suppl 4: 81-87 [PMID: 15861638]
- 20 Okada Y, Watanabe S, Nakanishi I, Kishi J, Hayakawa T, Watorek W, Travis J, Nagase H. Inactivation of tissue inhibitor of metalloproteinases by neutrophil elastase and other serine proteinases. FEBS Lett 1988; 229: 157-160 [PMID: 3162216 DOI: 10.1016/0014-5793(88)80817-2]
- Potempa J, Dubin A, Watorek W, Travis J. An elastase inhibitor from equine leukocyte cytosol 21 belongs to the serpin superfamily. Further characterization and amino acid sequence of the reactive center. J Biol Chem 1988; 263: 7364-7369 [PMID: 3366785]
- Varki NM, Varki A. Diversity in cell surface sialic acid presentations: implications for biology and 22 disease. Lab Invest 2007; 87: 851-857 [PMID: 17632542 DOI: 10.1038/Labinvest.3700656]
- 23 Sillanaukee P, Pönniö M, Jääskeläinen IP. Occurrence of sialic acids in healthy humans and different



disorders. Eur J Clin Invest 1999; 29: 413-425 [PMID: 10354198 DOI: 10.1046/j.1365-2362.1999.00485.x]

- Akcay F, Taysi S, Uslu C, Doğru Y, Gümüştekin K. Levels of soluble intercellular adhesion 24 molecule-1 and total sialic acid in serum of patients with laryngeal cancer. Jpn J Clin Oncol 2001; 31: 584-588 [PMID: 11902488 DOI: 10.1093/jjco/hye128]
- 25 de Albuquerque Garcia Redondo P, Nakamura CV, de Souza W, Morgado-Díaz JA. Differential expression of sialic acid and N-acetylgalactosamine residues on the cell surface of intestinal epithelial cells according to normal or metastatic potential. J Histochem Cytochem 2004; 52: 629-640 [PMID: 15100240 DOI: 10.1177/002215540405200507]
- 26 Feijoo-Carnero C, Rodríguez-Berrocal FJ, Páez de la Cadena M, Ayude D, de Carlos A, Martínez-Zorzano VS. Clinical significance of preoperative serum sialic acid levels in colorectal cancer: utility in the detection of patients at high risk of tumor recurrence. Int J Biol Markers 2004; 19: 38-45 [PMID: 15077925 DOI: 10.5301/ibm.2008.576]
- Katopodis N, Hirshaut Y, Geller NL, Stock CC. Lipid-associated sialic acid test for the detection of 27 human cancer. Cancer Res 1982; 42: 5270-5275 [PMID: 7139630]
- 28 López Sáez JJ, Senra-Varela A. Evaluation of lipid-bound sialic acid (LSA) as a tumor marker. Int J Biol Markers 1995; 10: 174-179 [PMID: 8551061]
- 29 Miyazaki K, Ohmori K, Izawa M, Koike T, Kumamoto K, Furukawa K, Ando T, Kiso M, Yamaji T, Hashimoto Y, Suzuki A, Yoshida A, Takeuchi M, Kannagi R. Loss of disialyl Lewis(a), the ligand for lymphocyte inhibitory receptor sialic acid-binding immunoglobulin-like lectin-7 (Siglec-7) associated with increased sialyl Lewis(a) expression on human colon cancers. Cancer Res 2004; 64: 4498-4505 [PMID: 15231659 DOI: 10.1158/0008-5472.CAN-03-3614]
- Shen Y, Tiralongo J, Kohla G, Schauer R. Regulation of sialic acid O-acetylation in human colon 30 mucosa. Biol Chem 2004; 385: 145-152 [PMID: 15101557 DOI: 10.1515/BC.2004.033]
- Shen Y, Kohla G, Lrhorfi AL, Sipos B, Kalthoff H, Gerwig GJ, Kamerling JP, Schauer R, Tiralongo 31 J. O-acetylation and de-O-acetylation of sialic acids in human colorectal carcinoma. Eur J Biochem 2004; 271: 281-290 [PMID: 14717696 DOI: 10.1046/j.1432-1033.2003.03927.x]
- 32 Uslu C, Taysi S, Akcay F, Sutbeyaz MY, Bakan N. Serum free and bound sialic acid and alpha-1-acid glycoprotein in patients with laryngeal cancer. Ann Clin Lab Sci 2003; 33: 156-159 [PMID: 128176191
- Barrett AJ. Fluorimetric assays for cathepsin B and cathepsin H with methylcoumarylamide 33 substrates. Biochem J 1980; 187: 909-912 [PMID: 6897924 DOI: 10.1042/bj1870909]
- 34 Jourdian GW, Dean L, Roseman S. The sialic acids. XI. A periodate-resorcinol method for the quantitative estimation of free sialic acids and their glycosides. J Biol Chem 1971; 246: 430-435 [PMID: 5542012]
- Tautu C, Verazin G, Prorok JJ, Alhadeff JA. Improved procedure for determination of serum lipid-35 associated sialic acid: application for early diagnosis of colorectal cancer. J Natl Cancer Inst 1988; 80: 1333-1337 [PMID: 3172258 DOI: 10.1093/jnci/80.16.1333]
- Warwas M, Knapik-Kordecka M, Kowal-Gierczak B. Stężenie i aktywność alfal-inhibitora proteaz 36 oraz alfa-2 makroglobuliny w osoczu krwi chorych na cukrzycę typu II. Diagn Lab 1991; 44: 552-556
- Dietz AA, Rubinstein HM, Hodges LK. Use of alpha-N-benzoyl-L-arginine-p-nitroanilide as trypsin 37 substrate in estimation of alpha 1-antitrypsin. Clin Chem 1976; 22: 1754-1755 [PMID: 10097]
- Herszényi L, Farinati F, Plebani M, Carraro P, Roveroni G, De Paoli M, Cardin R, Naccarato R, 38 Tulassay Z. [Prognostic role of cisteine and serin proteases in gastriC cancer]. Orv Hetil 1996; 137: 1637-1641 [PMID: 9019701]
- Herszényi L, Farinati F, Plebani M, István G, Sápi Z, Carraro P, De Paoli M, Naccarato R, Tulassay Z. [The role of cathepsins and the plasminogen activator/inhibitor system in colorectal cancer]. Orv Hetil 1999; 140: 1833-1836 [PMID: 10489782]
- 40 Kos J, Nielsen HJ, Krasovec M, Christensen IJ, Cimerman N, Stephens RW, Brünner N. Prognostic values of cathepsin B and carcinoembryonic antigen in sera of patients with colorectal cancer. Clin Cancer Res 1998; 4: 1511-1516 [PMID: 9626470]
- 41 Dufek V, Jirásek V, Král V, Matous B, Drazná E. Changes in serum cathepsin B-like activity in patients with colorectal cancer. Neoplasma 1985; 32: 51-54 [PMID: 3982561]
- 42 Amiguet JA, Jiménez J, Monreal JI, Hernández MJ, López-Vivanco G, Vidán JR, Conchillo F, Liso P. Serum proteolytic activities and antiproteases in human colorectal carcinoma. J Physiol Biochem 1998; 54: 9-13 [PMID: 9732103]
- 43 Padilla D, Cubo T, Molina JM, García M, De la Osa G, Palomino T, Pardo R, Martín J, Arévalo E, Hernández Calvo J. [Prognostic significance and clinic utility of serum and immunohistochemical cathepsin B levels in colorectal cancer]. An Med Interna 2003; 20: 521-525 [PMID: 14585038]
- Skrzydlewska E, Sulkowska M, Wincewicz A, Koda M, Sulkowski S. Evaluation of serum cathepsin 44 B and D in relation to clinicopathological staging of colorectal cancer. World J Gastroenterol 2005; 11: 4225-4229 [PMID: 16015694 DOI: 10.3748/wjg.v11.i27.4225]
- Zore I, Krasovec M, Cimerman N, Kuhelj R, Werle B, Nielsen HJ, Brünner N, Kos J. Cathepsin 45 B/cystatin C complex levels in sera from patients with lung and colorectal cancer. Biol Chem 2001; 382: 805-810 [PMID: 11517934 DOI: 10.1515/BC.2001.097]
- Herszényi L, Plebani M, Carraro P, De Paoli M, Roveroni G, Cardin R, Foschia F, Tulassay Z, Naccarato R, Farinati F. Proteases in gastrointestinal neoplastic diseases. Clin Chim Acta 2000; 291:



171-187 [PMID: 10675722 DOI: 10.1016/s0009-8981(99)00227-2]

- Herszényi L, Farinati F, Cardin R, István G, Molnár LD, Hritz I, De Paoli M, Plebani M, Tulassay Z. 47 Tumor marker utility and prognostic relevance of cathepsin B, cathepsin L, urokinase-type plasminogen activator, plasminogen activator inhibitor type-1, CEA and CA 19-9 in colorectal cancer. BMC Cancer 2008; 8: 194 [PMID: 18616803 DOI: 10.1186/1471-2407-8-194]
- Geraghty P, Rogan MP, Greene CM, Boxio RM, Poiriert T, O'Mahony M, Belaaouaj A, O'Neill SJ, 48 Taggart CC, McElvaney NG. Neutrophil elastase up-regulates cathepsin B and matrix metalloprotease-2 expression. J Immunol 2007; 178: 5871-5878 [PMID: 17442971 DOI: 10.4049/jimmunol.178.9.5871]
- 49 Sun Z, Yang P. Role of imbalance between neutrophil elastase and alpha 1-antitrypsin in cancer development and progression. Lancet Oncol 2004; 5: 182-190 [PMID: 15003202 DOI: 10.1016/S1470-2045(04)01414-7
- Xu Y, Zhang J, Han J, Pan X, Cao Y, Guo H, Pan Y, An Y, Li X. Curcumin inhibits tumor 50 proliferation induced by neutrophil elastase through the upregulation of α1-antitrypsin in lung cancer. Mol Oncol 2012; 6: 405-417 [PMID: 22507634 DOI: 10.1016/j.molonc.2012.03.005]
- Sato T, Takahashi S, Mizumoto T, Harao M, Akizuki M, Takasugi M, Fukutomi T, Yamashita J. 51 Neutrophil elastase and cancer. Surg Oncol 2006; 15: 217-222 [PMID: 17320378 DOI: 10.1016/j.suronc.2007.01.003
- Kataoka H, Nabeshima K, Komada N, Koono M. New human colorectal carcinoma cell lines that 52 secrete proteinase inhibitors in vitro. Virchows Arch B Cell Pathol Incl Mol Pathol 1989; 57: 157-165 [PMID: 2570482 DOI: 10.1007/BF02899077]
- 53 Keppler D, Markert M, Carnal B, Berdoz J, Bamat J, Sordat B. Human colon carcinoma cells synthesize and secrete alpha 1-proteinase inhibitor. Biol Chem Hoppe Seyler 1996; 377: 301-311 [PMID: 8828821 DOI: 10.1515/bchm3.1996.377.5.301]
- Sugino T, Yamaguchi T, Ogura G, Kusakabe T, Goodison S, Homma Y, Suzuki T. The secretory 54 leukocyte protease inhibitor (SLPI) suppresses cancer cell invasion but promotes blood-borne metastasis via an invasion-independent pathway. J Pathol 2007; 212: 152-160 [PMID: 17455170 DOI: 10.1002/path.2156]
- 55 Korkmaz B, Attucci S, Jourdan ML, Juliano L, Gauthier F. Inhibition of neutrophil elastase by alpha1-protease inhibitor at the surface of human polymorphonuclear neutrophils. J Immunol 2005; 175: 3329-3338 [PMID: 16116225 DOI: 10.4049/jimmunol.175.5.3329]
- 56 Meyer-Hoffert U, Wiedow O. Neutrophil serine proteases: mediators of innate immune responses. Curr Opin Hematol 2011; 18: 19-24 [PMID: 21042214 DOI: 10.1097/MOH.0b013e32834115d1]
- Nozawa F, Hirota M, Okabe A, Shibata M, Iwamura T, Haga Y, Ogawa M. Elastase activity 57 enhances the adhesion of neutrophil and cancer cells to vascular endothelial cells. J Surg Res 2000; 94: 153-158 [PMID: 11104655 DOI: 10.1006/jsre.2000.6002]
- 58 Ho AS, Chen CH, Cheng CC, Wang CC, Lin HC, Luo TY, Lien GS, Chang J. Neutrophil elastase as a diagnostic marker and therapeutic target in colorectal cancers. Oncotarget 2014; 5: 473-480 [PMID: 24457622 DOI: 10.18632/oncotarget.1631]
- 59 Kovcić V, Jelić S, Filipović I, Tomasević Z. [Serum haptoglobin and alpha-1 antitrysin levels as biological evolution markers in patients with gastric and colorectal cancer]. Srp Arh Celok Lek 1994; 122: 311-313 [PMID: 17974404]
- 60 Millán Núñez-Cortés J. [Biological and clinical aspects of alpha 1-antitrypsin with special reference to malignant tumor processes]. Rev Esp Oncol 1981; 28: 591-641 contd [PMID: 6764960]
- Simpson WG, Heys SD, Whiting PH, Eremin O, Broom J. Acute phase proteins and recombinant IL-61 2 therapy: prediction of response and survival in patients with colorectal cancer. Clin Exp Immunol 1995; 99: 143-147 [PMID: 7531626 DOI: 10.1111/j.1365-2249.1995.tb05524.x]
- Stamatiadis AP, St Toumanidou M, Vyssoulis GP, Manouras AJ, Apostolidis NS. Value of serum 62 acute-phase reactant proteins and carcinoembryonic antigen in the preoperative staging of colorectal cancer. A multivariate analysis. Cancer 1990; 65: 2055-2057 [PMID: 1695546 DOI: 10.1002/1097-0142(19900501)65:9<2055::aid-cncr2820650927>3.0.co;2-6]
- Jaberie H, Hosseini SV, Naghibalhossaini F. Evaluation of Alpha 1-Antitrypsin for the Early 63 Diagnosis of Colorectal Cancer. Pathol Oncol Res 2020; 26: 1165-1173 [PMID: 31183614 DOI: 10.1007/s12253-019-00679-0
- Bujanda L, Sarasqueta C, Cosme A, Hijona E, Enríquez-Navascués JM, Placer C, Villarreal E, 64 Herreros-Villanueva M, Giraldez MD, Gironella M, Balaguer F, Castells A, Evaluation of alpha 1antitrypsin and the levels of mRNA expression of matrix metalloproteinase 7, urokinase type plasminogen activator receptor and COX-2 for the diagnosis of colorectal cancer. PLoS One 2013; 8: e51810 [PMID: 23300952 DOI: 10.1371/journal.pone.0051810]
- 65 Yüceyar S, Ertürk S, Dirican A, Cengiz A, Saner H. The role of acute-phase reactant proteins, carcinoembryonic antigen and CA 19-9 as a marker in the preoperative staging of colorectal cancer: a prospective clinical study. Int Surg 1996; 81: 136-139 [PMID: 8912078]
- Gallardo-Valverde JM, Calañas-Continente A, Baena-Delgado E, Zurera-Tendero L, Vázquez-66 Martínez C, Membrives-Obrero A, Muntané J, Arévalo-Jiménez E. Obstruction in patients with colorectal cancer increases morbidity and mortality in association with altered nutritional status. Nutr Cancer 2005; 53: 169-176 [PMID: 16573378 DOI: 10.1207/s15327914nc5302_6]
- 67 Bernacka K, Kuryliszyn-Moskal A, Sierakowski S. The levels of alpha 1-antitrypsin and alpha 1antichymotrypsin in the sera of patients with gastrointestinal cancers during diagnosis. Cancer 1988; 62: 1188-1193 [PMID: 3261623 DOI:



10.1002/1097-0142(19880915)62:6<1188::aid-cncr2820620624>3.0.co;2-e]

- 68 Ward DG, Suggett N, Cheng Y, Wei W, Johnson H, Billingham LJ, Ismail T, Wakelam MJ, Johnson PJ, Martin A. Identification of serum biomarkers for colon cancer by proteomic analysis. Br J Cancer 2006; 94: 1898-1905 [PMID: 16755300 DOI: 10.1038/sj.bjc.6603188]
- 69 Kątnik-Prastowska I. Struktura i biologia kwasów sjalowych. Adv Clin Exp Med 2003; 5: 653-663
- 70 Painbeni T, Gamelin E, Cailleux A, Le Bouil A, Boisdron-Celle M, Daver A, Larra F, Allain P.
- Plasma sialic acid as a marker of the effect of the treatment on metastatic colorectal cancer. Eur J Cancer 1997; 33: 2216-2220 [PMID: 9470809 DOI: 10.1016/s0959-8049(97)00318-3]
- Romppanen J, Eskelinen M, Tikanoja S, Mononen I. Total and lipid-bound serum sialic acid in 71 benign and malignant breast disease. Anticancer Res 1997; 17: 1249-1253 [PMID: 9137482]
- 72 Varki A. Sialic acids in human health and disease. Trends Mol Med 2008; 14: 351-360 [PMID: 18606570 DOI: 10.1016/j.molmed.2008.06.002]
- 73 Brockhausen I, Yang JM, Burchell J, Whitehouse C, Taylor-Papadimitriou J. Mechanisms underlying aberrant glycosylation of MUC1 mucin in breast cancer cells. Eur J Biochem 1995; 233: 607-617 [PMID: 7588808 DOI: 10.1111/j.1432-1033.1995.607 2.x]
- 74 Inagaki Y, Tang W, Guo Q, Kokudo N, Sugawara Y, Karako H, Konishi T, Nakata M, Nagawa H, Makuuchi M. Sialoglycoconjugate expression in primary colorectal cancer and metastatic lymph node tissues. Hepatogastroenterology 2007; 54: 53-57 [PMID: 17419230]
- 75 Miyagi T, Wada T, Yamaguchi K. Roles of plasma membrane-associated sialidase NEU3 in human cancers. Biochim Biophys Acta 2008; 1780: 532-537 [PMID: 18023981 DOI: 10.1016/j.bbagen.2007.09.016
- Schutter EM, Visser JJ, van Kamp GJ, Mensdorff-Pouilly S, van Dijk W, Hilgers J, Kenemans P. 76 The utility of lipid-associated sialic acid (LASA or LSA) as a serum marker for malignancy. A review of the literature. Tumour Biol 1992; 13: 121-132 [PMID: 1626178 DOI: 10.1159/000217755]
- Cylwik B, Chrostek L, Szmitkowski M. [Diagnostic value of total and lipid-bound sialic acid in 77 malignancies]. Pol Merkur Lekarski 2005; 19: 237-241 [PMID: 16245443]
- 78 Feijoo C, Páez de la Cadena M, Rodríguez-Berrocal FJ, Martínez-Zorzano VS. Sialic acid levels in serum and tissue from colorectal cancer patients. Cancer Lett 1997; 112: 155-160 [PMID: 9066722 DOI: 10.1016/s0304-3835(96)04564-8]
- 79 Sebzda T, Saleh Y, Gburek J, Warwas M, Andrzejak R, Siewinski M, Rudnicki J. Total and lipidbound plasma sialic acid as diagnostic markers in colorectal cancer patients: correlation with cathepsin B expression in progression to Dukes stage. J Exp Ther Oncol 2006; 5: 223-229 [PMID: 165289721
- 80 İliklerden ÜH, Peksen C, Kalayci T, Kemik O. Evaluation of preoperative and postoperative total serum sialic acid levels in patients with colon cancer. Ann Ital Chir 2020; 91: 649-647 [PMID: 335549431
- 81 Zhang Z, Wuhrer M, Holst S. Serum sialylation changes in cancer. *Glycoconj J* 2018; 35: 139-160 [PMID: 29680984 DOI: 10.1007/s10719-018-9820-0]
- 82 Herszényi L, Barabás L, Hritz I, István G, Tulassay Z. Impact of proteolytic enzymes in colorectal cancer development and progression. World J Gastroenterol 2014; 20: 13246-13257 [PMID: 25309062 DOI: 10.3748/wjg.v20.i37.13246]
- Yin J, Hashimoto A, Izawa M, Miyazaki K, Chen GY, Takematsu H, Kozutsumi Y, Suzuki A, 83 Furuhata K, Cheng FL, Lin CH, Sato C, Kitajima K, Kannagi R. Hypoxic culture induces expression of sialin, a sialic acid transporter, and cancer-associated gangliosides containing non-human sialic acid on human cancer cells. Cancer Res 2006; 66: 2937-2945 [PMID: 16540641 DOI: 10.1158/0008-5472.CAN-05-2615]



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ORIGINAL ARTICLE

Observational Study Impact of a colorectal cancer screening program implantation on delays and prognosis of non-screening detected colorectal cancer

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Abstract

BACKGROUND

The implementation of a colorectal cancer (CRC) screening programme may increase the awareness of Primary Care Physicians, reduce the diagnostic delay in CRC detected outside the scope of the screening programme and thus improve prognosis.

AIM

To determine the effect of implementation of a CRC screening programme on diagnostic delays and prognosis of CRC detected outside the scope of a screening programme.

METHODS

We performed a retrospective intervention study with a pre-post design. We identified 322 patients with incident and confirmed CRC in the pre-implantation cohort (June 2014 - May 2015) and 285 in the post-implantation cohort (June 2017 -May 2018) in the Cancer Registry detected outside the scope of a CRC screening programme. In each patient we calculated the different healthcare diagnostics delays: global, primary and secondary healthcare, referral and colonoscopyrelated delays. In addition, we collected the initial healthcare that evaluated the patient, the home location (urban/rural), and the CRC stage at diagnosis. We



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conducted in accordance with the Declaration of Helsinki, and the protocol was approved by the Ethics Committee of Galicia, Spain (code 2016/274). As long as the study was based on database use, no informed consent was required. The information was accessed according to prevailing European and Spanish legislation.

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determined the two-year survival and we performed a multivariate proportional hazard regression analysis to determine the variables associated with survival.

RESULTS

We did not detect any differences in the patient or CRC baseline-related variables. A total of 20.1% of patients was detected with metastatic disease. There was a significant increase in direct referral to colonoscopy from primary healthcare (25.5%, 35.8%; P = 0.04) in the post-implantation cohort. Diagnostic delay was reduced by 24 d (106.64 \pm 148.84 days, 82.84 \pm 109.31 d; P = 0.02) due to the reduction in secondary healthcare delay (46.01 \pm 111.65 d; 29.20 \pm 60.83 d; P = 0.02). However, we did not find any differences in CRC stage at diagnosis or in two-year survival (70.3%; P = 0.9). Variables independently associated with twoyear risk of death were age (Hazard Ratio-HR: 1.06, 95% CI: 1.04-1.07), CRC stage (II HR: 2.17, 95% CI: 1.07-4.40; III HR: 3.07, 95% CI: 1.56-6.08; IV HR: 19.22, 95% CI: 9.86-37.44; unknown HR: 9.24, 95% CI: 4.27-19.99), initial healthcare consultation (secondary HR: 2.93, 95%CI: 1.01-8.55; emergency department HR: 2.06, 95%CI: 0.67-6.34), hospitalization during the diagnostic process (HR: 1.67, 95%CI: 1.17-2.38) and urban residence (HR: 1.44, 95%CI: 1.06-1.98).

CONCLUSION

Although implementation of a CRC screening programme can reduce diagnostic delays for CRC detected in symptomatic patients, this has no effect on CRC stage or survival.

Key Words: Colorectal cancer; Population based screening; Primary healthcare; Diagnostic delay; Prognosis

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Core Tip: We have designed a retrospective intervention study with a pre-post design to confirm the hypothesis that the implementation of a colorectal cancer (CRC) screening program may increase the awareness of primary care physicians and, thus, reduce the diagnostic delays in CRC detected outside the screening program and improve prognosis. Our results confirm that the implementation of the CRC screening program reduced the diagnostic delays due to an increase in the direct referrals to colonoscopy from primary healthcare. However, this reduction in the delays had no effect on the stage at diagnosis or in the two year survival. These later results were confirmed in a multivariable Cox regression analysis.

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INTRODUCTION

Colorectal cancer (CRC) is one of the most important health problems in the Western world. In 2018, almost half a million new cases were diagnosed in Europe and 250,000 patients died due to CRC[1]. In order to reduce the disease burden, population-based CRC screening programmes have been established in the Western world. This strategy has demonstrated its efficacy to reduce CRC mortality and incidence in randomized controlled trials. Furthermore, we have real data showing that implementation of CRC screening programmes has achieved its expected efficiency in reducing both CRC mortality and incidence[2,3].

In spite of the implementation of CRC screening programmes, most CRC are detected among symptomatic patients outside the scope of CRC screening mainly due to the limited participation and the detection in age cohorts that are not candidates for CRC screening[4,5]. However, as in breast cancer screening, the implementation of



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CRC screening may have an additional positive effect on these patients due to increased awareness and creation of multidisciplinary teams[6]. In this sense, CRC screening may increase the CRC awareness of patients and primary care physicians (PCPs) and promote use of faecal immunochemical test (FIT) as a triage test to refer patients to colonoscopy[7].

The delay to diagnosis in cancer is due to factors related to the patient and health system. The period from initial symptoms until final diagnosis is made can be highly variable. Although the common belief is that a longer delay can lead to an advanced stage at diagnosis and worse prognosis, evidence on CRC is controversial[8]. Patients seeking assistance with more severe symptoms are diagnosed in a shorter period and have more advanced disease[9]. In contrast, there is no evidence that a health system delay lower than six months worsens prognosis in the context of an outpatient diagnosis[10].

Based on the hypothesis that implementation of a mass CRC screening programme could raise awareness of patients and PCPs, we decided to design a retrospective intervention study to determine whether implementation of a CRC screening programme could reduce health system delays and, secondarily, improve CRC staging at diagnosis and long term survival.

MATERIALS AND METHODS

Study design

We designed a retrospective intervention study with a pre-post design without a control group.

Description of the intervention

The intervention was the first round of the Galician CRC screening programme that took place between 1 July 2015 and 30 June 2017 in Ourense, Spain. Galician CRC mass screening is based on biennial FIT with a 20 μ g haemoglobin/g of faeces threshold. FIT is offered to subjects aged 50 to 69 years. It is coordinated by the Public Health Department of the Galician Regional Health Department. They are in charge of the identification of subjects, invitation to participate, reception of FIT results, citation of patients with a positive result to perform a colonoscopy and final evaluation of the endoscopic and histological results. Primary healthcare clinics are in charge of promoting participation in the screening programme, collecting FIT kits and evaluation of subjects with a positive FIT prior to colonoscopy. The hospitals in each health area are responsible for FIT analysis, colonoscopies, histological analysis and evaluation and treatment of patients with a CRC. Finally, personnel at the Coordination Unit key in data into the screening programme's information system regarding CRC stage according to the AJCC classification,[11]. the final classification of patients with a positive result^[12]. as well as several quality endoscopist indicators according to the Spanish guideline on quality in screening colonoscopy^[13]. During the implantation of the CRC screening program no change was performed in the diagnostic pathways for CRC diagnosis in symptomatic patients.

Inclusion criteria and definition of the cohorts

Pre cohort: We included all invasive incident CRC histologically confirmed detected in the natural year before implementation of the CRC screening programme (1 July 2014 - 30 June 2015) in Ourense.

Post cohort: We included all invasive incident CRC histologically confirmed and detected outside the scope of the CRC screening programme in the natural year after the first round: (1 July 2017- 30 June 2018).

Identification of the incident CRC

We identified the incident using the case identification structure developed and validated by the project for implementation of the Galician Tumour Registry (Project REGAT). REGAT uses the topographic codes ICD-O-3.1 C18-C19-C20 to identify the CRC[14]. Codes C18.1 (appendix), C21 (anus and anal canal) were excluded. REGAT data were crosslinked with the Galician CRC screening information system to exclude those patients with a CRC diagnosed within the screening programme.

Variables analyzed

We collected information regarding: (1) Demographics (age and sex); and (2) Tumour



location in relation to the splenic flexure: proximal (caecum, ascending colon, hepatic flexure and transverse colon) and distal (rectum, sigma, descending colon and splenic flexure).

Cancer stage at diagnosis according to the TNM classification (AJCC 7th edition) [11]. We used the following data to determine the stage at diagnosis: clinical or anatomo-pathological stage for metastatic disease, imaging tests for the local rectal cancer stage (T and N), anatomo-pathological evaluation for the remaining situations (colon cancer T and N).

We searched in IANUS, the unified clinical record database of the Galician Health Department, for information regarding the contacts and referrals in the healthcare system. IANUS includes all information regarding attendance in primary and secondary healthcare as well as emergency departments and hospitalization. We determined the first contact in the health system (primary, secondary, emergency), whether the patient required hospitalization during the diagnostic process and the diagnostic delays. We defined five diagnostic delays (Figure 1): (1) Global diagnostic delay: Overall delay from the first consultation to definitive diagnosis; (2) Primary healthcare delay: Delay from the initial evaluation in primary healthcare until the decision to refer to secondary healthcare. In the event of colonoscopy being directly requested from primary healthcare, this date was considered as the referral date; (3) Referral delay: Delay from the primary healthcare referral to the first attendance in secondary healthcare (either clinical consultation or performing of colonoscopy); (4) Secondary healthcare delay: Delay from the first attendance in secondary healthcare to final diagnosis; and (5) Colonoscopy delay: Delay from the colonoscopy request to the performing of colonoscopy.

Statistical analysis

First, we performed a descriptive analysis of the variables included: number and frequencies in the qualitative variables and mean and standard deviation in the quantitative variables. We determined whether there were differences between both cohorts in the diagnostic pathways (hospitalization, direct referral to colonoscopy from primary healthcare) using the Chi-square test. In order to detect whether there were differences in the referral delays between both cohorts we used the Student t test. We analyzed whether there were differences in two-year survival between both cohorts in the Kaplan-Meier analysis using the log-rank test. Finally, to control confounding variables we performed a Cox multivariate regression analysis and we determined which variables were independently associated with survival after diagnosis. The study was statistically reviewed by a biomedical statistician.

Ethics aspects

The study was conducted in accordance with the Declaration of Helsinki, and the protocol was approved by the Ethics Committee of Galicia, Spain (code 2016/274). As long as the study was based on database use, no informed consent was required. The information was accessed according to prevailing European and Spanish legislation.

RESULTS

Description of the sample

We identified records from 757 patients in the two periods analyzed in the cancer registry. We excluded 92 patients that did not meet the inclusion criteria and 58 patients with CRC detected within the CRC screening programme in the postimplantation cohort. Finally, the pre-implantation and post-implantation cohort consisted of 322 and 285 patients, respectively (Figure 2).

As we show in Table 1, we did not detect baseline differences between both cohorts. CRC was detected more commonly in males (59.6%) with a mean age of 74.5 ± 11.5 years and more than two thirds were distal to the splenic flexure. There were no differences with respect to the place of residence either. Most patients were initially evaluated in primary healthcare but up to 41.0% required hospitalization before reaching final diagnosis. Diagnosis was made through colonoscopy in 89.8% of detected CRC. In this sense, we detected a significant increase in the colonoscopy directly requested from primary healthcare in the post-implantation cohort (P = 0.04). When we limited the analysis to those patients initially seen in primary healthcare (522), the results were similar. In this sense, we only found differences in the rate of colonoscopy directly referred from primary healthcare (29.2%, 42.3%; P = 0.005).



Table 1 Characteristics of the patients included						
	Pre-implantation cohort (<i>n</i> = 322)	Post-implantation cohort (<i>n</i> = 285)	P value ¹			
Sex			0.1			
Male	184 (57.1%)	178 (62.5%)				
Female	138 (42.9%)	107 (37.5%)				
Age (yr)	74.1 ± 11.8	74.8 ± 11.1	0.4			
Colorectal location			0.8			
Distal to splenic	221 (68.6%)	197 (69.1%)				
Proximal to splenic	101 (31.4%)	88 (30.9%)				
TNM						
Ι	45 (14.0%)	47 (16.5%)				
П	93 (26.9%)	71 (24.9%)				
III	101 (31.4%)	99 (34.7%)	0.5			
IV	65 (20.2%)	57 (20.0%)				
Unknown	18 (5.6%)	11 (3.9%)				
Rural/Urban						
Rural	218 (67.9%)	195 (68.4%)	0.8			
Urban	103 (32.1%)	90 (31.6%)				
Initial consultation						
Primary healthcare	281 (87.3%)	241 (84.6%)				
Secondar y healthcare	33 (10.2%)	37 (13.0%)	0.5			
Emergency department	8 (2.5%)	7 (2.5%)				
Hospitalization						
Yes	135 (41.9%)	114 (40.0%)	0.6			
No	187 (52.2%)	171 (60.0%)				
Colonoscopy request						
Primary healthcare	82 (25.5%)	102 (35.8%)				
Secondary healthcare			0.04			
After referral	160 (49.7%)	116 (40.7%)				
Direct request	45 (14.0%)	40 (14.0%)				
No colonoscopy	35 (10.9%)	27 (9.5%)				

¹Statistical significance in the univariate analysis using the Chi-square test for qualitative variables and the Student *t* test for quantitative variables.

Delay to diagnosis

The delay to diagnosis was reduced in 24 d after implantation of the CRC screening programme (P = 0.02). We did not detect any differences in the primary healthcare, referral or colonoscopy delay. The reduction was due to a secondary healthcare delay in relation to an increased rate of direct referral to colonoscopy from primary healthcare, as we show in Table 2 and Figure 3.

The global delay was also reduced by 27 d in patients evaluated initially in primary healthcare (117.66 \pm 154.08 days, 90.06 \pm 111.31 days; *P* = 0.02) also due to a reduction in secondary healthcare delay ($48.17 \pm 116.42 \text{ d}$, $26.89 \pm 54.50 \text{ d}$; P = 0.02). There were no differences in primary healthcare, referral or colonoscopy delay.

Factors associated with survival

The incidence of metastatic CRC remained stable (20.1%) in both cohorts and overall survival after one and two years was 71.3% and 70.3% without differences in the log-

Table 2 Delay to colorectal cancer diagnosis in the pre and post-implantation cohorts						
	Pre-implantation cohort (n = 322)	Post-implantation cohort (n = 285)	P value ¹			
Global diagnostic delay (d)	106.64 ± 148.84	82.84 ± 109.31	0.02			
Primary healthcare delay (d)	35.88 ± 84.47	39.28 ± 98.03	0.7			
Referral delay (d)	13.18 ± 25.77	16.02 ± 41.63	0.4			
Secondary healthcare delay (d)	46.01 ± 111.65	29.20 ± 60.83	0.02			
Colonoscopy delay (d)	43.71 ± 78.22	37.75 ± 53.37	0.3			

¹Statistical significance in the univariate analysis using the Student *t* test.

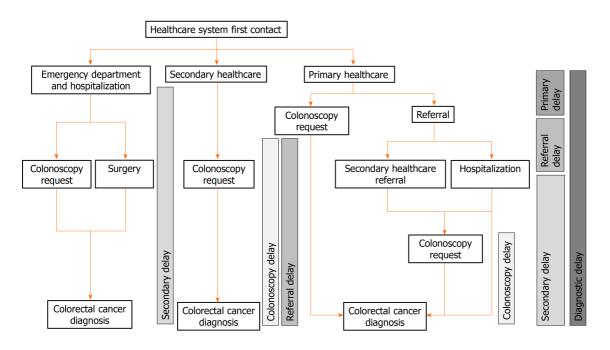


Figure 1 Flowchart of the referral and diagnostic pathways.

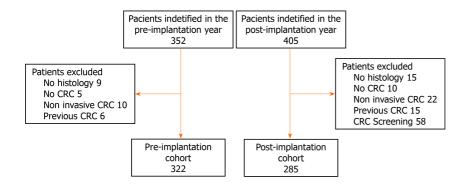


Figure 2 Flowchart of the patients included in the analysis.

rank test (P = 0.9) as we show in Figure 4. These results were confirmed in the Cox multivariate regression analysis and there were no differences in the survival between both cohorts (post-implantation cohort HR: 1.12, 95%CI: 0.83-1.51). As we show in Table 3, only age, CRC staging according to TNM classification, initial healthcare consultation, hospitalization during the diagnostic process and residence were independently associated with death after CRC diagnosis.

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Table 3 Factors associated with survival	
	Hazard ratio ¹ (95%CI)
Sex	
Male	1
Female	1.18 (0.89-1.58)
Age (yr)	1.06 (1.04-1.07)
Colorectal location	
Distal to splenic	1
Proximal to splenic	0.84 (0.62-1.13)
Cohort	
Pre-implantation	1
Post-implantation	1.12 (0.83-1.51)
TNM	
I	1
П	2.17 (1.07-4.40)
ш	3.07 (1.56-6.08)
IV	19.22 (9.86-37.44)
Unknown	9.24 (4.27-19.99)
Initial consultation	
Primary healthcare	1
Secondary healthcare	2.93 (1.01-8.55)
Emergency department	2.06 (0.67-6.34)
Hospitalization	
Yes	1.67 (1.17-2.38)
No	1
Colonoscopy request	
Primary healthcare	1.79 (0.96-3.35)
Secondary healthcare	1.54 (0.92-2.58)
After referral from Primary Healthcare	0.74 (0.25-2.21)
Direct request	1
No colonoscopy	
Rural/Urban	
Rural	1
Urban	1.44 (1.06-1.98)
Diagnostic delay (d)	1.001 (1.00-1.002)

¹Hazard Ratio and its 95% confidence interval calculated using a multivariate proportional hazard regression analysis.

DISCUSSION

Our study shows that implementation of the CRC screening programme reduced healthcare referral delays due to the direct request of colonoscopy from primary healthcare. Unfortunately, this reduction in referral delay had no effect on CRC staging at diagnosis nor on the two-year survival. Finally, although we detected several variables associated with overall survival, multivariate logistic analysis confirms that neither implantation of the CRC screening nor diagnostic delay were related to the prognosis of CRC detected outside the scope of CRC screening.

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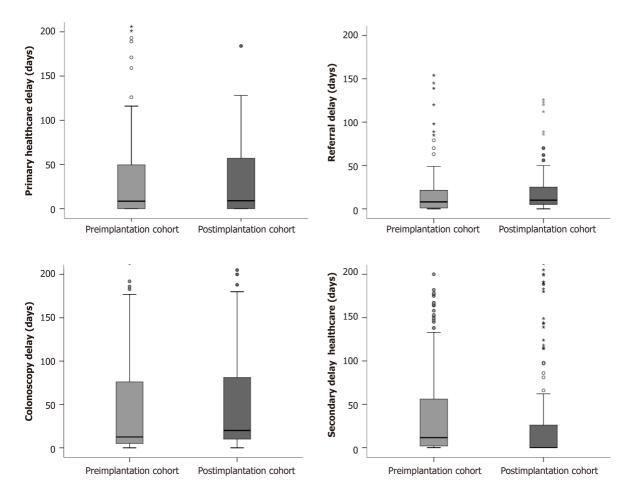


Figure 3 Healthcare diagnostic delays. We show the distribution of the primary and secondary healthcare, referral and colonoscopy delays expressed in days.

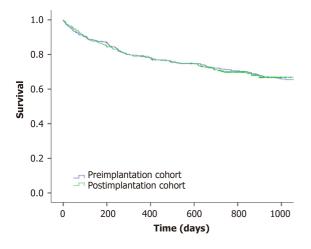


Figure 4 Survival curves of the pre and post-implantation cohorts. Survival curves were calculated using the Kaplan-Meier method.

PCPs play an important role in CRC care, from encouraging screening and accurate diagnosis to providing care during and after treatment for cancer and any comorbid complications. The implication of PCPs on CRC screening is variable according to the screening programme. Participation rates are increased when PCPs are involved in the invitation process. However, in the European population-based programmes in Europe PCPs play a rather supportive, informative or facilitating role[15]. In our case, PCPs receive full information on the screening programme organization and they are in charge of promoting participation as well as resolving any doubts. Within the training, PCPs are reminded which symptoms may lead to suspicion of CRC as well as the established referral pathways, including direct referral criteria for colonoscopy evaluation from primary healthcare[8,16].

We designed this analysis under the hypothesis that increased awareness on CRC and training in the diagnosis of CRC and the established protocols could reduce delays attributed to the health system. In this sense, our results confirm that implementation of the screening programme enabled a reduction in the diagnostic delay due to an increase in direct referrals to colonoscopy from primary healthcare. PCPs, as demonstrated in our study, are the main gateway and responsible for a significant part of the delay [17-19]. The role of PCPs in CRC diagnosis is complex since gastrointestinal symptoms that may suggest CRC are very common, the CRC prevalence low and the diagnostic performance of available symptom-based tools is very limited [20]. Recently, implementation of the faecal immunochemical test to triage patients with gastrointestinal symptoms in primary healthcare has improved diagnostic referral pathways[20,21]. In the health area of Ourense, faecal immunochemical test was implemented as a triage test seven years ago, so we cannot attribute the decreases in delay to this modification[16].

However, it is relevant that, despite the reduction in delay, we have not detected any changes in the stage at diagnosis or in the prognosis of CRC. These data are in accordance with results previously published by our group[8]. and with the data in the available meta-analysis on the effect of diagnostic delays in CRC prognosis[9]. In this sense, the prognosis of patients with shorter diagnostic delay is worse due to presentation with urgent symptoms that require hospitalization or more serious systemic symptoms^[19]. In fact, in our study, hospitalization during admission was associated with a higher risk of mortality after diagnosis. In our research, although initial urgent presentation was rare, up to 40% of patients required hospitalization during the diagnostic process, similar to the information available on literature[22-24]. This lack of relationship between delay and prognosis may be related to different forms of presentation. In this sense, a prospective study on patients that met the National Institute for Health and Care Excellence referral criteria demonstrated that a delay of more than six months was associated with a worse prognosis compared to patients with the same symptoms diagnosed in an interval of less than one month[10].

Our study has two main strengths. We had the opportunity to evaluate the effect of the CRC screening programme on the diagnostic delays of CRC detected in symptomatic patients. This is the first study that evaluates additional impacts of the implementation of CRC screening on CRC diagnosis. No study has evaluated whether a CRC screening programme can increase the awareness of patients and PCPs, reduce delays and improve prognosis. However, we could identify all the CRC through the Galician cancer registry, confirm the diagnosis in IANUS, the centralized clinical record and determine when the patient was evaluated in the health system and thus calculate all the referral delays[25].

There are several limitations. Due to the design of the study, we could not evaluate the effect of CRC screening on the patient delays to seek assistance. Patient delay accounts for a relevant proportion of the delay between the onset of symptoms and the final diagnosis^[26]. Moreover, we did not collect the initial symptoms as long as they were not collected uniformly in the clinical records.

CONCLUSION

To conclude, the implementation of a CRC screening programme enabled reduction of health system diagnostic delay by means of increased patients referred directly by PCPs to colonoscopy. However, this reduction in referral delay did not modify either CRC stage at diagnosis or two-year survival.

ARTICLE HIGHLIGHTS

Research background

In spite of the implementation of colorectal cancer (CRC) screening programmes, most CRC are detected among symptomatic patients outside the scope of CRC screening. However, they may increase the CRC awareness of patients and primary care physicians (PCP).

Research motivation

The implementation of a mass CRC screening programme could raise awareness of patients and PCPs, we decided to design a retrospective intervention study to



determine whether implementation of a CRC screening programme could reduce health system delays and, secondarily, improve CRC staging at diagnosis and long term survival.

Research objectives

To determine the effect of implementation of a CRC screening programme on diagnostic delays and prognosis of CRC detected outside the scope of a screening programme.

Research methods

We designed a retrospective intervention study with a pre-post design without a control group. We compared diagnostic delays, CRC stage and two year survival of a yearly CRC diagnosed before the implementation of a CRC screening programa with a CRC cohort diagnosed the year after the first round.

Research results

There was a significant increase in direct referral to colonoscopy from primary healthcare (25.5%, 35.8%; P = 0.04) in the post-implantation cohort. Diagnostic delay was reduced by 24 d (106.64 ± 148.84 d, 82.84 ± 109.31 d; P = 0.02) due to the reduction in secondary healthcare delay ($46.01 \pm 111.65 \text{ d}$; $29.20 \pm 60.83 \text{ d}$; P = 0.02). However, we did not find any differences in CRC stage at diagnosis or in two-year survival (70.3%; P = 0.9).

Research conclusions

Although implementation of a CRC screening programme can reduce diagnostic delays for CRC detected in symptomatic patients, this has no effect on CRC stage or survival.

Research perspectives

We need more research on the motivations and perspectives of patients seeking help in primary healthcare.

REFERENCES

- World Health Organization. Cancer Today. International Agency for Research on Cancer 1 [Internet]. [cited 23 May 2020]. Available from: https://gco.iarc.fr/today/home
- 2 Zorzi M, Fedeli U, Schievano E, Bovo E, Guzzinati S, Baracco S, Fedato C, Saugo M, Dei Tos AP. Impact on colorectal cancer mortality of screening programmes based on the faecal immunochemical test. Gut 2015; 64: 784-790 [PMID: 25179811 DOI: 10.1136/gutjnl-2014-307508]
- Levin TR, Corley DA, Jensen CD, Schottinger JE, Quinn VP, Zauber AG, Lee JK, Zhao WK, 3 Udaltsova N, Ghai NR, Lee AT, Quesenberry CP, Fireman BH, Doubeni CA. Effects of Organized Colorectal Cancer Screening on Cancer Incidence and Mortality in a Large Community-Based Population. Gastroenterology 2018; 155: 1383-1391.e5 [PMID: 30031768 DOI: 10.1053/j.gastro.2018.07.017
- 4 Mansouri D, McMillan DC, Crearie C, Morrison DS, Crighton EM, Horgan PG. Temporal trends in mode, site and stage of presentation with the introduction of colorectal cancer screening: a decade of experience from the West of Scotland. Br J Cancer 2015; 113: 556-561 [PMID: 26158422 DOI: 10.1038/bjc.2015.230]
- 5 Gutierrez-Stampa MA, Aguilar V, Sarasqueta C, Cubiella J, Portillo I, Bujanda L. Impact of the faecal immunochemical test on colorectal cancer survival. BMC Cancer 2020; 20: 616 [PMID: 32611328 DOI: 10.1186/s12885-020-07074-y]
- 6 Kalager M, Zelen M, Langmark F, Adami HO. Effect of screening mammography on breast-cancer mortality in Norway. N Engl J Med 2010; 363: 1203-1210 [PMID: 20860502 DOI: 10.1056/NEJMoa1000727]
- 7 Gutierrez-Stampa MA, Aguilar V, Sarasqueta C, Cubiella J, Portillo I, Bujanda L. Colorectal Cancer Survival in 50- to 69-Year-Olds after Introducing the Faecal Immunochemical Test. Cancers (Basel) 2020; 12 [PMID: 32854370 DOI: 10.3390/cancers12092412]
- Fernández-de Castro JD, Baiocchi Ureta F, Fernández González R, Pin Vieito N, Cubiella 8 Fernández J. The effect of diagnostic delay attributable to the healthcare system on the prognosis of colorectal cancer. Gastroenterol Hepatol 2019; 42: 527-533 [PMID: 31421857 DOI: 10.1016/j.gastrohep.2019.03.012
- Tørring ML, Murchie P, Hamilton W, Vedsted P, Esteva M, Lautrup M, Winget M, Rubin G. Evidence of advanced stage colorectal cancer with longer diagnostic intervals: a pooled analysis of seven primary care cohorts comprising 11 720 patients in five countries. Br J Cancer 2017; 117: 888-897 [PMID: 28787432 DOI: 10.1038/bjc.2017.236]



- Alonso-Abreu I, Alarcón-Fernández O, Gimeno-García AZ, Romero-García R, Carrillo-Palau M, 10 Nicolás-Pérez D, Jiménez A, Quintero E. Early Colonoscopy Improves the Outcome of Patients With Symptomatic Colorectal Cancer. Dis Colon Rectum 2017; 60: 837-844 [PMID: 28682969 DOI: 10.1097/DCR.00000000000863]
- 11 Edge SB, Byrd DR, Compton CC, Fritz AG, Greene FL. In: Trotti A, editor. AJCC Cancer Staging Manual [Internet]. 7th edition. Springer, 2010. [cited 23 May 2020]. Available from: http://www.springer.com/medicine/surgery/book/978-0-387-88440-0
- 12 European Colorectal Cancer Screening Guidelines Working Group, von Karsa L, Patnick J, Segnan N, Atkin W, Halloran S, Lansdorp-Vogelaar I, Malila N, Minozzi S, Moss S, Quirke P, Steele RJ, Vieth M, Aabakken L, Altenhofen L, Ancelle-Park R, Antoljak N, Anttila A, Armaroli P, Arrossi S, Austoker J, Banzi R, Bellisario C, Blom J, Brenner H, Bretthauer M, Camargo Cancela M, Costamagna G, Cuzick J, Dai M, Daniel J, Dekker E, Delicata N, Ducarroz S, Erfkamp H, Espinàs JA, Faivre J, Faulds Wood L, Flugelman A, Frkovic-Grazio S, Geller B, Giordano L, Grazzini G, Green J, Hamashima C, Herrmann C, Hewitson P, Hoff G, Holten I, Jover R, Kaminski MF, Kuipers EJ, Kurtinaitis J, Lambert R, Launoy G, Lee W, Leicester R, Leja M, Lieberman D, Lignini T, Lucas E, Lynge E, Mádai S, Marinho J, Maučec Zakotnik J, Minoli G, Monk C, Morais A, Muwonge R, Nadel M, Neamtiu L, Peris Tuser M, Pignone M, Pox C, Primic-Zakelj M, Psaila J, Rabeneck L, Ransohoff D, Rasmussen M, Regula J, Ren J, Rennert G, Rey J, Riddell RH, Risio M, Rodrigues V, Saito H, Sauvaget C, Scharpantgen A, Schmiegel W, Senore C, Siddiqi M, Sighoko D, Smith R, Smith S, Suchanek S, Suonio E, Tong W, Törnberg S, Van Cutsem E, Vignatelli L, Villain P, Voti L, Watanabe H, Watson J, Winawer S, Young G, Zaksas V, Zappa M, Valori R. European guidelines for quality assurance in colorectal cancer screening and diagnosis: overview and introduction to the full supplement publication. Endoscopy 2013; 45: 51-59 [PMID: 23212726 DOI: 10.1055/s-0032-13259971
- Jover R, Herráiz M, Alarcón O, Brullet E, Bujanda L, Bustamante M, Campo R, Carreño R, Castells 13 A, Cubiella J, García-Iglesias P, Hervás AJ, Menchén P, Ono A, Panadés A, Parra-Blanco A, Pellisé M, Ponce M, Quintero E, Reñé JM, Sánchez del Río A, Seoane A, Serradesanferm A, Soriano Izquierdo A, Vázquez Sequeiros E; Spanish Society of Gastroenterology; Spanish Society of Gastrointestinal Endoscopy Working Group. Clinical practice guidelines: quality of colonoscopy in colorectal cancer screening. Endoscopy 2012; 44: 444-451 [PMID: 22438159 DOI: 10.1055/s-0032-1306690]
- 14 World Health Organization. The International Classification of Diseases for Oncology (ICD-O) - 3 rd edition, 1st revision [Internet]. 2013. [cited 31 January 2021]. Available from: https://apps.who.int/iris/handle/10665/96612
- 15 Triantafillidis JK, Vagianos C, Gikas A, Korontzi M, Papalois A. Screening for colorectal cancer: the role of the primary care physician. Eur J Gastroenterol Hepatol 2017; 29: e1-e7 [PMID: 27676092 DOI: 10.1097/MEG.000000000000759]
- Vega-Villaamil P, Salve-Bouzo M, Cubiella J, Valentín-Gómez F, Sánchez-Hernández E, Gómez-16 Fernández I, Fernández-Seara J. Evaluation of the implementation of Galician Health Service indications and priority levels for colonoscopy in symptomatic patients: prospective, cross-sectional study. Rev Esp Enferm Dig 2013; 105: 600-608 [PMID: 24641457 DOI: 10.4321/s1130-01082013001000005
- 17 van Erp NF, Helsper CW, Olyhoek SM, Janssen RRT, Winsveen A, Peeters PHM, de Wit NJ. Potential for Reducing Time to Referral for Colorectal Cancer Patients in Primary Care. Ann Fam Med 2019; 17: 419-427 [PMID: 31501203 DOI: 10.1370/afm.2446]
- 18 Brandenbarg D, Groenhof F, Siewers IM, van der Voort A, Walter FM, Berendsen AJ. Possible missed opportunities for diagnosing colorectal cancer in Dutch primary care: a multimethods approach. Br J Gen Pract 2018; 68: e54-e62 [PMID: 29203683 DOI: 10.3399/bjgp17X693905]
- Esteva M, Leiva A, Ramos M, Pita-Fernández S, González-Luján L, Casamitjana M, Sánchez MA, 19 Pértega-Díaz S, Ruiz A, Gonzalez-Santamaría P, Martín-Rabadán M, Costa-Alcaraz AM, Espí A, Macià F, Segura JM, Lafita S, Arnal-Monreal F, Amengual I, Boscá-Watts MM, Hospital A, Manzano H, Magallón R; DECCIRE GROUP. Factors related with symptom duration until diagnosis and treatment of symptomatic colorectal cancer. BMC Cancer 2013; 13: 87 [PMID: 23432789 DOI: 10.1186/1471-2407-13-87
- Cubiella J, Marzo-Castillejo M, Mascort-Roca JJ, Amador-Romero FJ, Bellas-Beceiro B, Clofent-20 Vilaplana J, Carballal S, Ferrándiz-Santos J, Gimeno-García AZ, Jover R, Mangas-Sanjuán C, Moreira L, Pellisè M, Quintero E, Rodríguez-Camacho E, Vega-Villaamil P; Sociedad Española de Medicina de Familia y Comunitaria y Asociación Española de Gastroenterología. Clinical practice guideline. Diagnosis and prevention of colorectal cancer. 2018 Update. Gastroenterol Hepatol 2018; 41: 585-596 [PMID: 30245076 DOI: 10.1016/j.gastrohep.2018.07.012]
- 21 Pin Vieito N, Zarraquiños S, Cubiella J. High-risk symptoms and quantitative faecal immunochemical test accuracy: Systematic review and meta-analysis. World J Gastroenterol 2019; 25: 2383-2401 [PMID: 31148909 DOI: 10.3748/wjg.v25.i19.2383]
- Renzi C, Lyratzopoulos G, Card T, Chu TP, Macleod U, Rachet B. Do colorectal cancer patients 22 diagnosed as an emergency differ from non-emergency patients in their consultation patterns and symptoms? Br J Cancer 2016; 115: 866-875 [PMID: 27537389 DOI: 10.1038/bjc.2016.250]
- Esteva M, Ruidíaz M, Sánchez MA, Pértega S, Pita-Fernández S, Macià F, Posso M, González-Luján 23 L, Boscá-Wats MM, Leiva A, Ripoll J; DECCIRE GROUP. Emergency presentation of colorectal patients in Spain. PLoS One 2018; 13: e0203556 [PMID: 30273339 DOI:



10.1371/journal.pone.0203556]

- Borowski DW, Cawkwell S, Zaidi SM, Toward M, Maguire N, Gill TS. Primary care referral 24 practice, variability and socio-economic deprivation in colorectal cancer. Colorectal Dis 2016; 18: 1072-1079 [PMID: 27110954 DOI: 10.1111/codi.13360]
- 25 Leiva A, Esteva M, Llobera J, Macià F, Pita-Fernández S, González-Luján L, Sánchez-Calavera MA, Ramos M. Time to diagnosis and stage of symptomatic colorectal cancer determined by three different sources of information: A population based retrospective study. Cancer Epidemiol 2017; 47: 48-55 [PMID: 28126583 DOI: 10.1016/j.canep.2016.10.021]
- 26 Weller D, Menon U, Zalounina Falborg A, Jensen H, Barisic A, Knudsen AK, Bergin RJ, Brewster DH, Cairnduff V, Gavin AT, Grunfeld E, Harland E, Lambe M, Law RJ, Lin Y, Malmberg M, Turner D, Neal RD, White V, Harrison S, Reguilon I; ICBP Module 4 Working Group, Vedsted P. Diagnostic routes and time intervals for patients with colorectal cancer in 10 international jurisdictions; findings from a cross-sectional study from the International Cancer Benchmarking Partnership (ICBP). BMJ Open 2018; 8: e023870 [PMID: 30482749 DOI: 10.1136/bmjopen-2018-023870]



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ORIGINAL ARTICLE

Prospective Study Standard liver weight model in adult deceased donors with fatty liver: A prospective cohort study

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Author contributions: Li B and Chen PY contributed equally to this work; Li B collected all the clinical data, analyzed the data, drafted the manuscript and prepared the revised materials; Chen PY collected the data of donor liver weight in the operating room and assisted with data analysis; Tan YF analyzed the data, drafted the manuscript and prepared the revised materials; Huang H and Luo Y assisted with ultrasound examination and data processing; Wu ZR and Shi YJ helped with tissue staining and histological assessment; Zheng DF, He D and Jiang CH assisted with donor liver weight measurement and tissue Sampling; Jiang M participated in design of the study, data processing and data analysis; Yang JY participated in design and oversight of the study and was

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Abstract

BACKGROUND

Standard liver weight (SLW) is frequently used in deceased donor liver transplantation to avoid size mismatches with the recipient. However, some deceased donors (DDs) have fatty liver (FL). A few studies have reported that FL could impact liver size. To the best of our knowledge, there are no relevant SLW models for predicting liver size.

AIM

To demonstrate the relationship between FL and total liver weight (TLW) in detail



involved with drafting of the manuscript; all authors have read and approve the final manuscript.

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Institutional review board

statement: This study was reviewed and approved by the West China Hospital of Sichuan University Institutional Review Board.

Clinical trial registration statement:

This study was registered at http://www.chictr.org.cn. The registration identification number is ChiCTR2000041406.

Informed consent statement: All study participants, or their legal guardians, provided informed written consent prior to study enrollment.

Conflict-of-interest statement: The authors of this manuscript have no conflicts of interest to disclose.

Data sharing statement: No additional data are available.

CONSORT 2010 statement: The authors have read the CONSORT 2010 Statement, and the manuscript was prepared and revised according to the CONSORT 2010 Statement.

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and present a related SLW formula.

METHODS

We prospectively enrolled 212 adult DDs from West China Hospital of Sichuan University from June 2019 to February 2021, recorded their basic information, such as sex, age, body height (BH) and body weight (BW), and performed abdominal ultrasound (US) and pathological biopsy (PB). The chi-square test and kappa consistency score were used to assess the consistency in terms of FL diagnosed by US relative to PB. Simple linear regression analysis was used to explore the variables related to TLW. Multiple linear regression analysis was used to formulate SLW models, and the root mean standard error and interclass correlation coefficient were used to test the fitting efficiency and accuracy of the model, respectively. Furthermore, the optimal formula was compared with previous formulas.

RESULTS

Approximately 28.8% of DDs had FL. US had a high diagnostic ability (sensitivity and specificity were 86.2% and 92.9%, respectively; kappa value was 0.70, P < 0.001) for livers with more than a 5% fatty change. Simple linear regression analysis showed that sex (R^2 , 0.226; P < 0.001), BH (R^2 , 0.241; P < 0.001), BW (R^2 , 0.441; *P* < 0.001), BMI (R², 0.224; *P* < 0.001), BSA (R², 0.454; *P* < 0.001) and FL (R², 0.130; P < 0.001) significantly impacted TLW. In addition, multiple linear regression analysis showed that there was no significant difference in liver weight between the DDs with no steatosis and those with steatosis within 5%. Furthermore, in the context of hepatic steatosis, TLW increased positively (nonlinear); compared with the TLW of the non-FL group, the TLW of the groups with hepatic steatosis within 5%, between 5% and 20% and more than 20% increased by 0 g, 90 g, and 340 g, respectively. A novel formula, namely, -348.6 + (110.7 x Sex $[0 = \text{Female}, 1 = \text{Male}]) + 958.0 \times \text{BSA} + (179.8 \times \text{FL}_{US} [0 = \text{No}, 1 = \text{Yes}])$, where FL was diagnosed by US, was more convenient and accurate than any other formula for predicting SLW.

CONCLUSION

FL is positively correlated with TLW. The novel formula deduced using sex, BSA and FL_{US} is the optimal formula for predicting SLW in adult DDs.

Key Words: Standard liver weight; Body surface area; Fatty liver; Sex; Deceased donors

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Core Tip: This study was the first to explore the relationship between fatty liver (FL) and total liver weight (TLW) in detail using pathological biopsy based on adult deceased donors (DDs) and developed a new standard liver weight (SLW) formula. Moreover, to conveniently apply the SLW formula to the clinic, we introduced ultrasound (US). Notably, we found that FL was positively correlated with TLW and that US had a high diagnostic ability for mild to severe FL, which could increase liver weight significantly. The formula deduced using sex, BSA and FL_{us} is the optimal formula for predicting SLW in adult DDs.

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INTRODUCTION

Standard liver weight (SLW) is a key parameter in liver surgery. Its accurate evaluation is the basis for patient safety in both hepatectomy and liver transplantation



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(LT). In hepatectomy, the underestimation of SLW may lead to residual liver failure[1, 2], and in living donor liver transplantation (LDLT)/split liver transplantation (SLT), the underestimation of SLW can lead to small-for-size syndrome (SFSS)[3-5]. Since the establishment of Urata's standard liver volume (SLV) model[6], approximately 14 SLV models have been published worldwide, most of which are based on healthy people, living donors and autopsy donors from various medical centres. Deceased donor liver transplantation (DDLT) is a crucial donor liver source for alleviating the shortage of donor livers. Subsequently, SLT was established and further expanded the donor liver pool. Previous studies [7-10] have reported that SLT is not inferior to whole liver transplantation in terms of patient prognosis, which has encouraged the extensive use of SLT and necessitated an urgent demand for an SLW formula for DDLT to avoid severe mismatches, large-for-size syndrome[11,12] or SFSS. Moreover, deceased donors (DDs) and living donors (LDs) are from the general population and may have hepatic steatosis, which has a reported global incidence of 15%-30% [13,14]. To our knowledge, fatty liver (FL) may be associated with marginal grafts, as severe steatosis is a risk factor related to graft survival^[15] and may affect liver size^[16,17]. However, these associations have not been quantified conclusively. To the best of our knowledge, only one model[18] has been published for DDs, and it was based on a Western population and did not address FL. Therefore, this study prospectively collected adult DDs' clinical data combined with FL parameters to develop an SLW model

MATERIALS AND METHODS

The present study prospectively enrolled consecutive deceased liver donors from West China Hospital of Sichuan University from June 2019 to February 2021 and recorded basic patient information, such as sex, age, body height (BH) and body weight (BW). This study was reviewed and approved by the West China Hospital of Sichuan University Institutional Review Board and registered at http://www.chictr.org.cn. The registration identification number is ChiCTR2000041406. All the study participants, or their legal guardians, provided informed written consent prior to study enrollment, and the study protocol conformed to the ethical guidelines of the 1975 Declaration of Helsinki as reflected in a priori approval by the ethics committee. No executed prisoners were included in the study. A total of 212 DDs were enrolled, and brain death was confirmed in all of them before organ procurement. Advanced life support was maintained in an intensive care unit (ICU); moreover, abdominal ultrasound (US) examinations, liver function tests and kidney function tests were completed for each donor. Pathological biopsy (PB) was performed for all enrolled donor livers after they were obtained.

US examination

A US examination was carried out for all DDs before organ procurement. Scanning and diagnosis were conducted by 2 experienced (> 5 years) US doctors who were blinded to the final PB diagnosis. The examinations were performed by using a MultiWave ultrasound system (Aixplorer, France) equipped with an SC6-1 (1-6 MHz) transducer. FL was identified as a diffuse increase in fine echoes in the liver parenchyma. Representative images[19] are presented in Figure 1.

Donor liver weight measurement, tissue sampling and histological assessment

Donor livers were procured and trimmed in the operating room and were then weighed with a precision electronic balance (unit: kg, accurate to 0.001 kg, Figure 2) on a back table.

A single tissue wedge of approximately 1.0 cm × 1.0 cm × 1.0 cm was excised from the left lateral lobe surface of the donor liver, fixed in formalin and embedded in paraffin. Each donor liver was stained with haematoxylin and eosin (HE) and Masson's trichrome. The histological degree of liver pathology, including hepatic steatosis, ballooning of hepatocytes, lobular inflammation, necrosis, and fibrosis, was evaluated by two expert liver pathologists blinded to any other clinical information and laboratory data. The extent of hepatic steatosis was assessed by the percentage of hepatocytes containing large- and medium-sized intracytoplasmic lipid droplets (but not foamy microvesicles). The definition of ballooning of hepatocytes and lobular inflammation was as described by Kleiner *et al*[20] and Bedossa *et al*[21]. The definition of necrosis is described in Table 1. Fibrosis was scored according to the standard grading (inflammation) and staging (fibrosis) method based on the modified Scheuer



Characteristic	Total, <i>n</i> = 212	
Sex, male, <i>n</i> (%)	167 (78.8)	
Age, median (range), yr	49 (18-68)	
BH, median (range), cm	168 (150-185)	
BW, median (range), kg	65 (45-90)	
BMI, median (range), kg/m ²	23.35 (15.57-30.48)	
BSA, median (range), m ²	1.73 (1.37-2.10)	
ILW, median (range), g	1400 (830–2100)	
Cause of death, n (%)		
Trauma	106 (50.0)	
Cerebrovascular	97 (45.8)	
Other	9 (4.2)	
Degree of fatty change, median (range)	0 (0-40%)	
0, n (%)	151 (71.2)	
> 0, < 5%, n (%)	32 (15.1)	
5%-33%, n (%)	22 (10.4)	
> 33%, n (%)	7 (3.3)	
Ballooning of hepatocytes		
None	24 (11.1)	
Ballooned hepatocyte with normal size	116 (54.9)	
Enlarged ballooned hepatocyte	72 (34.0)	
Lobular inflammation		
None	66 (30.9)	
< 2 foci per lobule	131 (61.7)	
> 2 foci per lobule	15 (7.4)	
Necrosis		
None	200 (94.4)	
Focal or unicellular necrosis	8 (3.7)	
More extensive necrosis and above	4 (1.9)	
Stage of fibrosis ¹		
0	72 (33.8)	
1	88 (41.6)	
2	47 (22.1)	
3	4 (1.9)	
4	1 (0.6)	

¹According to the modified Scheuer system[22]. BH: Body height; BW: Body weight; BMI: Body mass index; BSA: Body surface area; TLW: Total liver weight.

system[22].

Estimating SLW using previous formulas

According to previous studies at our centre^[23] and other centres^[24-26], the density of the liver was determined to be 1 g/cm³; that is, the weight and volume of the donor liver were equal. For comparison, we calculated the estimated SLW according to

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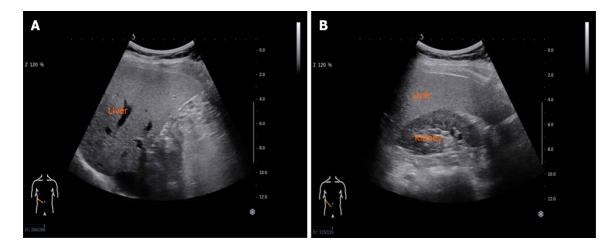


Figure 1 Diagram of fatty liver diagnosed by ultrasound from the view of the liver and kidney. A: Diffuse increase in fine echoes in liver parenchyma with normal visualization of intrahepatic vessel borders; B: Diffuse increase in fine echoes in liver parenchyma. There was an increase in echogenicity of the liver compared with the echogenicity of the renal cortex.



Figure 2 Actual liver weight measurement by electronic balance. A: Zero correction of electronic balance; B: Donor liver weighing. The arrow indicates that a single tissue wedge of approximately 1.0 cm × 1.0 cm vas excised from the left lateral lobe surface of the donor liver.

previous formulas for adults[6-19]. Body mass index (BMI) = BW/BH^2 and body surface area (BSA) = $BW^{0.425} \times BH^{0.725} \times 0.007184$ using the Dubois formula[27] were also calculated.

Statistical analysis

In this study, simple linear regression analysis was used to explore the variables related to TLW. Multiple linear regression analysis was used to formulate the SLW. As BH, BW, BMI and BSA are collinear variables, each was applied in a different prediction model. The root mean standard error (RMSE) and interclass correlation coefficient (ICC) were used to test the fitting efficiency and accuracy of the model, respectively. The chi-square test and kappa consistency score were used to assess the consistency in terms of FL diagnosed by US relative to PB. Continuous variables were analysed by a paired-samples t test. Two-tailed statistical analysis was used, and P values less than 0.05 were considered to be statistically significant. SPSS, version 25.0 (IBM, Armonk, NY, United States) was used for all statistical analyses. GraphPad Prism 7.0 (GraphPad Software, Inc.) was used for drawing.

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RESULTS

Baseline data

This study included 167 males (78.8%). The median age was 49 years, ranging from 18 to 68 years. The median BH, BW, BMI, BSA and TLW were 1.68 m, 65 kg, 23.35 kg/m², 1.73 m² and 1400 g, respectively. The main causes of death of the DDs were trauma (50%), cerebrovasculature (45.8%), and other (4.2%), which included brain tumours and hypoxic-ischaemic encephalopathy. There were 151 DDs (71.2%) with no steatosis, 32 (15.1%) with steatosis within 5%, 22 (10.4%) with steatosis between 5% and 33%, and 7 (3.3%) with steatosis greater than 33%. Moreover, hepatocyte ballooning was observed in 88.9% of DDs. Lobular inflammation was observed in approximately 69.1% of DDs. Necrosis (focal or unicellular necrosis, in 3.7% of DDs samples, and more extensive necrosis, in 1.9% of DDs samples) was observed in only a few DDs liver tissue samples. Stage 0-2 Liver fibrosis was observed in approximately 97.5% of DDs (Table 1).

Impact factors related to the TLW of deceased donors

Simple linear regression analysis showed that sex, BH, BW, BMI, BSA and FL significantly impacted TLW (P < 0.001) (Table 2). BSA was the most influential factor related to liver size [R², 0.454; 95% confidence interval (CI): 1024.56–1383.79]. Multiple linear regression analysis showed that there was no significant difference in TLW between no steatosis and steatosis within 5% (P = 0.147, Figure 3A). Furthermore, in the context of hepatic steatosis, TLW increased positively (non-linear); compared with the TLW of the non-FL group, the TLW of the groups with hepatic steatosis within 5%, between 5% and 20% and more than 20% increased by 0 g, 90 g, and 340 g, respectively (Figure 3B).

Consistency test for FL diagnosis between US and PB

This study investigated 61 hepatic steatosis cases, which accounted for 28.8% of all cases, and moderate and severe steatosis cases, which accounted for 3.3%. The cases of hepatic steatosis and non-hepatic steatosis diagnosed by US were 38 and 174, respectively. The sensitivity and specificity of US were 55.7% and 97.4%, respectively, and the kappa value was 0.598 (P < 0.001). That is, its diagnostic consistency was good (Supplementary Table 1). Furthermore, when setting 5% as the cut-off value for diagnosing FL by PB, there were 174 cases within a 5% fatty change and 38 cases with more than a 5% fatty change diagnosed by US, with a sensitivity and specificity of 86.2% and 92.9%, respectively, and a kappa value of 0.70 (P < 0.001). Therefore, the diagnostic consistency between US and PB was high (Table 3).

Current formulas for estimating SLW

The SLW models were separately formulated based on four collinear variables, namely, BH, BW, BMI and BSA. Subsequently, three prediction model groups were established, two of which were used to assess the presence of FL based on US or PB; the third group did not include FL as an indicator. The present study showed that the SLW models based on BSA, FL and sex had the best fitness, and the adjusted R² and RMSE for PB and US were 0.546 and 169.985 and 0.546 and 169.913, respectively. The fitting efficiency of these two models was almost equal and better than that of the traditional method (adjusted R², 0.485; RMSE, 181.095) (Table 4).

Comparison between the current formula and previous formulas

Previously reported formulas were used to assess our DDs cohort, and the results showed that the fitting efficiency and accuracy of the SLW model introducing FL diagnosed by US were 168.3 (RMSE) and 0.71 (ICC), with a non-significant difference (P = 1.00) between the SLW and TLW of 1.5 g. The RMSE and ICC of Yu *et al*[25]'s and Lin et al[28]'s models were 187.5 and 0.61 and 188.0 and 0.63, respectively. There were no significant differences between the SLW and actual TLW for these two formulas, but those of the remaining formulas were significantly different (Table 5)[6,18,25-37].

DISCUSSION

The shortage of donor livers is a problem worldwide and has become a major obstacle hindering the development of LT. To date, experts in the LT field have explored expanding the donor liver pool, including via SLT, marginal donor LT, domino LT and



Table 2 Factors related to the total liver weight of the deceased donors						
Factor	R ²	P value	95%CI			
Sex	0.226	< 0.001	220.89-369.68			
ВН	0.241	< 0.001	13.92-22.78			
BW	0.441	< 0.001	15.25-20.77			
BSA	0.454	< 0.001	1024.56-1383.79			
BMI	0.224	< 0.001	32.28-54.18			
Degree of fatty change (< 5%, 5%–20%, > 20%)	0.130	< 0.001	116.89–244.17			
Hepatic steatosis ¹	0.125	< 0.001	149.67-318.33			

¹Diagnosed by ultrasound. BH: Body height; BW: Body weight; BSA: Body surface area; BMI: Body mass index.

Table 3 Results for livers with more than 5% fatty change diagnosed by ultrasound and pathological biopsy in the deceased donors

Ultrasound	Pathological biopsy	— Total	
Ulliasoullu	+	•	TOLAI
+	25	13	38
-	4	170	174
Total	29	183	212

According to the table above, livers with a fatty change of more than 5% were diagnosed by ultrasound, and the sensitivity and specificity were 86.2% and 92.9%, respectively. The chi-square test showed that the kappa value was 0.70, P < 0.001.

> so on. These schemes have successfully and significantly expanded the donor liver pool, and SLT has become one of the most valuable means of promotion. Graft weight (GW) plays a key role in recipients, especially in DDLT and LDLT. Therefore, it is necessary to evaluate the donor liver size in LT.

> DDs are patients with brain death caused by non-liver diseases. This study illustrated that 95.8% of DDs died from trauma or cardiovascular and cerebrovascular accidents. Biopsies showed that many donor livers had hepatocyte oedema and lobular inflammation, which can be explained by the cause of death. Trauma and cardiovascular and cerebrovascular accidents can cause instability of the circulatory system, leading to long-term ICU stays and the requirement for resuscitative therapy, which may cause unstable organ perfusion (hypoperfusion or hyperperfusion) and reperfusion injury. In addition, the use of a large number of vasoactive drugs may aggravate organ microcirculation disorder. Thus, the graft may have acute injury, such as lobular inflammation, hepatocyte oedema and even necrosis. The present study found that 28.8% of DDs had hepatic steatosis and that 2.5% had stage 3-4 Liver fibrosis. Unlike DDs, LDs screened from healthy populations rarely have FL or other acute liver injuries. In addition, it was unclear whether there was a difference in the SLW between DDs and LDs. To the best of our knowledge, there have been few relevant reports. Therefore, we explored the SLW model based on DDs data derived from West China Hospital.

> Simple linear regression analysis showed that liver size was correlated with sex. The liver size of males was larger than that of females, which was in line with previous studies[30,33]. We speculated that this might be related to the fact that the body size of men is generally larger than that of women and that men have a larger skeletal muscle system and higher daily consumption and metabolic requirements. Therefore, a larger liver mass is needed to meet physiological needs[38,39]. In addition, the present study found that BH, BW, BMI and BSA were closely related to liver size, which was similar to previous studies[6,25,31,40]. Indeed, multiple linear regression analysis revealed that the above four variables were collinear. From the perspective of morphology, liver size and physical indicators are supposed to be positively correlated. Moreover, in terms of energy requirements, to meet metabolic needs, a larger body size needs more organ support. Furthermore, the current study found that BSA was the most influential factor impacting TLW, which was consistent with previous studies[6,29,



Table 4 Results of multiple linear regression analysis performed to predict the total liver weight using each of the body anthropometric measures divided into groups of the traditional method and two new methods, which introduce the parameter of fatty liver diagnosed by ultrasound and pathological biopsy

Groups	Formulas	Adjusted R ²	RMSE
Traditional method			
ВН	- 809.4 + 167.3 x Sex + 12.6 x BH	0.29	212.0
BW	322.1 + 147.0 x Sex + 15.2 x BW	0.49	181.1
BSA	- 466.9 + 99.0 x Sex + 1051.0 x BSA	0.48	182.8
BMI	329.2 + 264.5 x Sex + 37.8 x BMI	0.39	196.5
Ultrasound method			
ВН	- 1011.9 + 149.7 x Sex + 13.6 x BH + 240.7 x ${\rm FL}_{\rm US}$	0.43	191.1
BW	392.7 + 158.3 x Sex + 13.5 x BW + 158.6 x ${\rm FL}_{\rm US}$	0.54	171.4
BSA	- 348.6 + 110.7 x Sex + 958.0 x BSA + 179.8 x $\rm FL_{US}$	0.55	169.9
BMI	453.7 + 264.5 x Sex + 31.2 x BMI + 162.9 x ${\rm FL}_{\rm US}$	0.45	187.5
Pathological biopsy method (<	5%, 5%-20%, > 20%)		
ВН	- 803.7 + 178.5 x sex + 12.3 x BH + FL _{PB} (0 = 0, 1 = 163.5, 2 = 393.0)	0.43	190.0
BW	414.5 + 172.6 x sex + 13.1 x BW + FL_{\rm PB} (0 = 0, 1 = 79.8, 2 = 280.7)	0.54	170.8
BSA	- 288.8 + 129.5 x sex + 919.6 x BSA + $\mathrm{FL}_{\mathrm{PB}}$ (0 = 0, 1 = 93.9, 2 = 304.5)	0.55	170.0
BMI	478.1 + 276.5 x Sex + 30.0 x BMI + FL_{PB} (0 = 0, 1 = 105.3, 2 = 299.1)	0.46	185.4

Sex and FL_{US} are binary variables; FL_{PB} is a dummy variable. Sex: 0 = Female, 1 = Male; FL_{US} : 0 = No, 1 = Yes; FL_{PB} : 0 < 5%, 1 = 5%–20%, 2 > 20%. BH: Body height; BW: Body weight; BSA: Body surface area; BMI: Body mass index; FL_{US}; Fatty liver diagnosed by ultrasound; FL_{PB}: Fatty liver diagnosed by pathological biopsy; RMSE: Root mean standard error.

> 31]. BSA is a widely used parameter in physiology and clinical medicine for normalizing biological function with respect to variations in body size and conformation. Thus, we believe that the liver size required to meet the metabolic demands of the individual may correlate more closely with BSA than with any other parameter. Additionally, previous studies [30,34] reported that age was associated with TLW; however, similar to Poovathumkadavil's study[35], we failed to identify an association between age and TLW. Several previous studies[31,40] reported that the partial regression coefficient of age was very small, and the authors considered the effect of this variable in adults to be negligible. Therefore, our negative result may be explained by the age distribution of patients in our study and the sample size, and further studies with larger sample sizes are needed to confirm the relation between age and TLW.

> Interestingly, this study found that more than a quarter of DDs from the general population had hepatic steatosis, which was similar to Zhou *et al*[41]'s report (29.2%). To our knowledge, an increasing number of individuals, especially those who are obese, suffer from FL worldwide[42,43]. Furthermore, the present study also found that 10.4% and 3.3% of livers had mild and moderate steatosis, respectively, while no liver was detected to have severe steatosis. Several studies have confirmed that mild steatosis grafts (< 33%) can be used safely in LT. However, the eligibility of livers with moderate steatosis is controversial, while livers with severe steatosis are generally discarded because of the increased probability of primary non-function[15,44,45]. Importantly, in the current study, simple linear regression analysis demonstrated that FL was correlated with TLW. Moreover, multivariate analysis showed that steatosis significantly affected TLW, and the degree of steatosis was positively correlated with liver size, which was consistent with previous studies[16,46,47]. Multiple linear regression analysis showed that compared with non-FLs, the presence of hepatic steatosis within 5%, 5%-20% and over 20% resulted in an increase in liver weight by 0 g, 93.9 g, and 304.5 g, respectively. In LT, we generally evaluate the feasibility of SLT



Table 5 Differences between the estimated and actual liver weights calculated using previous formulas in our deceased donor cohort.

Ref.	Formula	Difference ¹ (g)	RMSE	ICC	P value ²
Autopsy					
DeLand et al[29]	1020 × BSA - 220	135.5 (-366–632)	221.2	0.52	< 0.01
Heinemann <i>et al</i> [26]	1072.8 × BSA - 345.7	95 (-421-556)	202.5	0.56	< 0.01
Yu et al <mark>[25</mark>]	$21.585 \times BW^{0.732} \times BH^{0.225}$	34.5 (-490-576)	187.5	0.61	0.102
Choukèr et al[30]	[16–50 yr] 452 + 16.34 x BW + 11.85 × age - 166 × sex (1 = female, 0 = male) 51–70 yr] 1390 + 15.94 × BW - 12.86 × age	435 (-301–1000)	484.0	0.24	< 0.01
General population/livin	g donor				
Urata[<mark>6</mark>]	$706.2 \times BSA + 2.4$	-185 (-713–337)	278.1	0.32	< 0.01
Lin <i>et al</i> [28]	13 × BH + 12 × BW - 1530	11.5 (-546-445)	188.0	0.63	0.472
Vauthey <i>et al</i> $[31]^3$	1267.28 × BSA - 794.41	-15 (-544-421)	188.1	0.64	< 0.01
Hashimoto <i>et al</i> [32]	961.3 × BSA - 404.8	-161 (-668-317)	253.4	0.42	< 0.01
Chan et al[<mark>33</mark>]	218 + BW × 12.3 + sex × 51 (0 = female, 1 = male)	-356.5 (- 859–175)	411.1	0.21	< 0.01
Yuan et al[34]	949.7 × BSA - 247.4–48.3 x age factor (1, < 40; 2, 41–60; 3, > 60)	-106 (-646-359)	228.0	0.48	< 0.01
Fu-Gui et al[23]	11.508 × BW + 334.024	-319 (-845-241)	393.6	0.19	< 0.01
Poovathumkadavil <i>et al</i> [<mark>35</mark>]	12.26 × BW + 555.65	-57 (-572–510)	207.5	0.47	< 0.01
Um et al[<mark>36</mark>]	893.485 x BSA - 439.169	-312.5 (- 816-173)	372.8	0.24	< 0.01
Cadaveric population					
Yoshizumi <i>et al</i> [18] ³	772 × BSA	-79 (-602-416)	214.6	0.45	< 0.01
Current	- 348.6 + 110.7 x Sex (0 = Female, 1 = Male) + 958.0 x BSA + 179.8 x FL _{US} (0 = No, 1 = Yes)	1.5 (- 477.0-450.0)	168.3	0.71	1

¹Difference between estimated and actual liver weight using previous formulas.

²Paired-samples *t* test.

³Mosteller's formula[37] was adopted for BSA, and the remaining formulas used the Dubois formula[27].

BH: Body height; BW: Body weight; BSA: Body surface area; BMI: Body mass index; FL_{US}: Fatty liver diagnosed by ultrasound; ICC: Interclass correlation coefficient: RMSE: Root mean standard error.

> based on the criteria of GW/SLW (30%-40%) or GW/BW (0.8%)[11]. Thus, for FL, the GW required for recipients would be underestimated if calculated according to the traditional SLV method, leading to an increased risk of SFSS. Therefore, the current study introduced the FL variable for the first time to develop an SLW model. To diagnose FL before organ procurement, US was performed for all DDs. Notably, for a diagnosis of mild steatosis and greater (\geq 5%), the sensitivity and specificity of US were 86.2% and 92.9%, respectively, and the ICC was 0.70 (P < 0.001). That is, US had a higher diagnostic consistency with PB. In addition, this study revealed that the size of livers with a fatty change less than 5% was not different from that of livers without fatty change but was different from that of livers with a fatty change of 5% or greater. The gap of liver size between these two hepatic steatosis categories was significant (180 g, P < 0.001), which laid a solid theoretical foundation to apply US in the diagnosis of FL and develop the SLW model, highlighting its clinical practical value.

> In this study, the deduced best fit formula based on US had equivalence with that based on PB and was better than the best fit traditional model. Furthermore, the present study showed that the formulas of Deland et al[29], Heinemann et al[26], and Chouker *et al*[30] overestimated liver size, while the formulas of Urata *et al*[6], Vauthey *et al*[31], Yoshizumi *et al*[18], Hashimoto *et al*[32], Chan *et al*[33], Yuan *et al*[34], Fu Gui et al[23], Poovathumkadavil et al[35], and Um et al[36] underestimated liver size. On the other hand, there was no significant difference between the actual liver weight and the predicted liver weight calculated by Yu *et al*[25]'s and Lin *et al*[28]'s formulas. This was speculated to be related to the characteristics of the study samples. Deland et al [29]'s, Heinemann *et al*[26]'s and Chouker *et al*[30]'s cohorts were autopsy samples. To



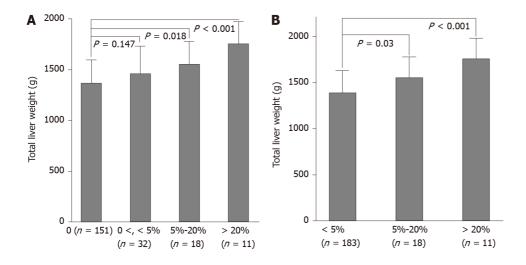


Figure 3 Total liver weight comparison of different groups according to the degree of fatty change of donor livers. A: Groups according to the degree of fatty change of 0, (0 <, < 5%), (5%-20%) and > 20%; B: Groups according to the degree of fatty change of < 5%, (5%-20%) and > 20%. Multiple linear regression analysis including parameters of sex, BSA, and FL_{PB}, which were dummy variables divided into groups according to the degree of fatty change in donor livers, was used. FL_{PB}, fatty liver diagnosed by pathological biopsy.

our knowledge, data from autopsy studies^[29] includes the weight of the gallbladder, the attached ligaments, and the hepatic vena cava. In addition, various causes of death, *i.e.*, cardiac failure and traffic accidents, might increase liver weight through mechanisms associated with shock-related hepatic congestion. On the other hand, due to long-term immersion in the fixed solution, the weight of the specimen may exceed the actual size in vivo. However, the autopsy study of Yu et al^[25] was not consistent with the other three autopsy studies but was similar to our study, which may be explained by racial differences. Additionally, the cohorts of Vauthey et al[31], Hashimoto *et al*[32], Chan *et al*[33], Yuan *et al*[34], Fu Gui *et al*[23], Poovathumkadavil et al[35], and Um et al[36] were based on healthy populations without liver disease. However, Lin et al[28]'s study cohort comprised 44 (57.1%) patients with chronic liver disease (alcoholic hepatitis, 9; hepatitis B, 24; and hepatitis C, 11), which may explain the difference from other studies based on the general population. Notably, the difference was significant between actual liver weight and estimated liver weight using the formula of Yoshizumi *et al*[18], which was the only previous study based on a cadaveric population. Their study included DDs of several races, most of which were Western, and subjects under 18 years were enrolled. These confounding factors may explain the difference. Therefore, for different study populations, the model for predicting liver size is supposed to be different, which highlights the need for this study for adult DDs. In addition, this study shows the practicability and rationality of the current SLW model in DDLT. Theoretically, it suggests that the current formula is the most suitable for recipients assigned with FL in SLT, and use of this formula is anticipated to reduce the risk of SFSS.

However, the sample size of this study was relatively small, especially in regard to cases of moderate to severe hepatic steatosis. Therefore, studies with larger sample sizes are warranted to optimize the SLW model. Additionally, the extrapolation and clinical practicability of the current SLW model need to be further verified.

CONCLUSION

In conclusion, this study was the first to demonstrate the positive correlation between the degree of hepatic steatosis and liver size based on pathological findings. Furthermore, this study creatively proposed and verified the equivalent value of FL diagnosed by US instead of that diagnosed by PB in terms of the FL variable in the SLW model as follows: SLW (g)= -348.6 + [110.7 x Sex (0 = Female, 1 = Male)] + 958.0 x BSA + [179.8 x FL_{US} (0 = No, 1 = Yes)]. This formula can be used to estimate the liver weight before liver procurement. Additionally, our formula lays a theoretical and practical basis for the further application of donor livers with fatty changes in SLT.

ARTICLE HIGHLIGHTS

Research background

Standard liver weight (SLW) is frequently used in liver transplantation, especially for living donor liver transplantation/split liver transplantation (SLT). However, some deceased donors (DDs) have fatty liver (FL). There have been a few studies to report that FL could impact liver size. This study was to develop a new formula including FL to predict liver size.

Research motivation

To explore SLW model in adult DDs with FL and help transplant doctors make allocation decisions, especially for recipients assigned with FL in SLT to reduce the risk of small-for-size syndrome.

Research objectives

To explore the liver pathology of DDs, such as hepatic steatosis, and diagnostic ability of ultrasound for FL, as well as the relationship between FL and total liver weight. Furthermore, to develop an SLW formula, combined with FL parameter, used to predict graft weight required for recipients in SLT.

Research methods

This study prospectively enrolled consecutive DDs from West China Hospital of Sichuan University from June 2019 to February 2021 and recorded basic patient information, and abdominal ultrasound (US) examination and pathological biopsy (PB) were performed for them. Furthermore, the chi-square test and kappa consistency score were used to assess the consistency in terms of FL diagnosed by US relative to PB. Simple linear regression analysis was used to explore the variables related to TLW. Multiple linear regression analysis was used to formulate SLW models.

Research results

More than a quarter of DDs had hepatic steatosis, and US had a high diagnostic ability for mild to severe FL. Furthermore, this study found that FL was positively correlated with liver size and deduced an optimal SLW formula in adult DDs with FL. However, the extrapolation and clinical practicability of the current SLW model need to be further verified in the future.

Research conclusions

FL is positively correlated with liver size. Our novel formula deduced using sex, BSA and FL_{US} is the optimal formula for predicting SLW in adult DDs with FL.

Research perspectives

To verify the extrapolation of the current SLW model using multicentre data and its clinical practicability in SLT.

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REFERENCES

- 1 Ribero D, Amisano M, Bertuzzo F, Langella S, Lo Tesoriere R, Ferrero A, Regge D, Capussotti L. Measured versus estimated total liver volume to preoperatively assess the adequacy of the future liver remnant: which method should we use? Ann Surg 2013; 258: 801-806; discussion 806 [PMID: 24045451 DOI: 10.1097/SLA.000000000000213]
- Mullen JT, Ribero D, Reddy SK, Donadon M, Zorzi D, Gautam S, Abdalla EK, Curley SA, Capussotti L, Clary BM, Vauthey JN. Hepatic insufficiency and mortality in 1,059 noncirrhotic



patients undergoing major hepatectomy. J Am Coll Surg 2007; 204: 854-862; discussion 862 [PMID: 17481498 DOI: 10.1016/j.jamcollsurg.2006.12.032]

- 3 Dahm F, Georgiev P, Clavien PA. Small-for-size syndrome after partial liver transplantation: definition, mechanisms of disease and clinical implications. Am J Transplant 2005; 5: 2605-2610 [PMID: 16212618 DOI: 10.1111/j.1600-6143.2005.01081.x]
- 4 Soejima Y, Taketomi A, Yoshizumi T, Uchiyama H, Harada N, Ijichi H, Yonemura Y, Shimada M, Maehara Y. Feasibility of left lobe living donor liver transplantation between adults: an 8-year, singlecenter experience of 107 cases. Am J Transplant 2006; 6: 1004-1011 [PMID: 16611337 DOI: 10.1111/j.1600-6143.2006.01284.x]
- 5 Emond JC, Renz JF, Ferrell LD, Rosenthal P, Lim RC, Roberts JP, Lake JR, Ascher NL. Functional analysis of grafts from living donors. Implications for the treatment of older recipients. Ann Surg 1996; **224**: 544-552; discussion 552 [PMID: 8857858 DOI: 10.1097/00000658-199610000-00012]
- Urata K, Kawasaki S, Matsunami H, Hashikura Y, Ikegami T, Ishizone S, Momose Y, Komiyama A, 6 Makuuchi M. Calculation of child and adult standard liver volume for liver transplantation. Hepatology 1995; 21: 1317-1321 [PMID: 7737637]
- Park GC, Hwang S, Song GW, Jung DH, Ha TY, Ahn CS, Moon DB, Kim KH, Yoon YI, Kang WH, 7 Cho HD, Choi JU, Kim M, Na BG, Kim SH, Lee SG. Prognosis of Split Liver Transplantation Compared with Whole Liver Transplantation in Adult Patients: Single-center Results under the Korean MELD Score-based Allocation Policy. J Korean Med Sci 2020; 35: e304 [PMID: 32959541 DOI: 10.3346/jkms.2020.35.e304]
- Cherukuru R, Reddy MS, Shanmugam NP, Rajalingam R, Kota V, Gunasekaran V, Narasimhan G, Kaliamoorthy I, Rela M. Feasibility and Safety of Split-Liver Transplantation in a Nascent Framework of Deceased Donation. Liver Transpl 2019; 25: 450-458 [PMID: 30586233 DOI: 10.1002/lt.25405]
- Valentino PL, Emre S, Geliang G, Li L, Deng Y, Mulligan D, Rodriguez-Davalos MI. Frequency of 9 whole-organ in lieu of split-liver transplantation over the last decade: Children experienced increased wait time and death. Am J Transplant 2019; 19: 3114-3123 [PMID: 31152483 DOI: 10.1111/ajt.15481]
- 10 Chul Yoon K, Song S, Jwa EK, Lee S, Man Kim J, Kim OK, Kyun Hong S, Yi NJ, Lee KW, Soo Kim M, Hwang S, Suh KS, Lee SK. Survival Outcomes in Split Compared With Whole Liver Transplantation. Liver Transpl 2018; 24: 1411-1424 [PMID: 29747216 DOI: 10.1002/lt.25196]
- Kiuchi T, Kasahara M, Uryuhara K, Inomata Y, Uemoto S, Asonuma K, Egawa H, Fujita S, Hayashi 11 M. Tanaka K. Impact of graft size mismatching on graft prognosis in liver transplantation from living donors. Transplantation 1999; 67: 321-327 [PMID: 10075602 DOI: 10.1097/00007890-199901270-00024
- 12 Tanaka A, Tanaka K, Tokuka A, Kitai T, Shinohara H, Hatano E, Sato S, Inomoto T, Takada Y, Higashiyama H, Nakamura Y, Yamamoto Y, Egawa H, Uemoto S, Ikai I, Ozaki N, Inomata Y, Yamaoka Y. Graft size-matching in living related partial liver transplantation in relation to tissue oxygenation and metabolic capacity. Transpl Int 1996; 9: 15-22 [PMID: 8748406 DOI: 10.1007/BF00336807
- 13 Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. Hepatology 2016; 64: 73-84 [PMID: 26707365 DOI: 10.1002/hep.28431]
- 14 Gao X, Fan JG; Study Group of Liver and Metabolism, Chinese Society of Endocrinology. Diagnosis and management of non-alcoholic fatty liver disease and related metabolic disorders: consensus statement from the Study Group of Liver and Metabolism. Chinese Society of Endocrinology, J Diabetes 2013; 5: 406-415 [PMID: 23560695 DOI: 10.1111/1753-0407.12056]
- 15 Spitzer AL, Lao OB, Dick AA, Bakthavatsalam R, Halldorson JB, Yeh MM, Upton MP, Reyes JD, Perkins JD. The biopsied donor liver: incorporating macrosteatosis into high-risk donor assessment. Liver Transpl 2010; 16: 874-884 [PMID: 20583086 DOI: 10.1002/lt.22085]
- 16 Kromrey ML, Ittermann T, vWahsen C, Plodeck V, Seppelt D, Hoffmann RT, Heiss P, Kühn JP. Reference values of liver volume in Caucasian population and factors influencing liver size. Eur J Radiol 2018; 106: 32-37 [PMID: 30150048 DOI: 10.1016/j.ejrad.2018.07.005]
- Bian H, Hakkarainen A, Zhou Y, Lundbom N, Olkkonen VM, Yki-Järvinen H. Impact of non-17 alcoholic fatty liver disease on liver volume in humans. Hepatol Res 2015; 45: 210-219 [PMID: 24698021 DOI: 10.1111/hepr.12338]
- 18 Yoshizumi T, Gondolesi GE, Bodian CA, Jeon H, Schwartz ME, Fishbein TM, Miller CM, Emre S. A simple new formula to assess liver weight. Transplant Proc 2003; 35: 1415-1420 [PMID: 12826175 DOI: 10.1016/s0041-1345(03)00482-2]
- Saadeh S, Younossi ZM, Remer EM, Gramlich T, Ong JP, Hurley M, Mullen KD, Cooper JN, 19 Sheridan MJ. The utility of radiological imaging in nonalcoholic fatty liver disease. Gastroenterology 2002; 123: 745-750 [PMID: 12198701 DOI: 10.1053/gast.2002.35354]
- 20 Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW, Ferrell LD, Liu YC, Torbenson MS, Unalp-Arida A, Yeh M, McCullough AJ, Sanyal AJ; Nonalcoholic Steatohepatitis Clinical Research Network. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatology* 2005; **41**: 1313-1321 [PMID: 15915461 DOI: 10.1002/hep.20701]
- 21 Bedossa P; FLIP Pathology Consortium. Utility and appropriateness of the fatty liver inhibition of progression (FLIP) algorithm and steatosis, activity, and fibrosis (SAF) score in the evaluation of biopsies of nonalcoholic fatty liver disease. *Hepatology* 2014; **60**: 565-575 [PMID: 24753132 DOI:



10.1002/hep.27173]

- 22 Batts KP, Ludwig J. Chronic hepatitis. An update on terminology and reporting. Am J Surg Pathol 1995; **19**: 1409-1417 [PMID: 7503362 DOI: 10.1097/00000478-199512000-00007]
- Fu-Gui L, Lu-Nan Y, Bo L, Yong Z, Tian-Fu W, Ming-Qing X, Wen-Tao W, Zhe-Yu C. Estimation 23 of standard liver volume in Chinese adult living donors. Transplant Proc 2009; 41: 4052-4056 [PMID: 20005340 DOI: 10.1016/j.transproceed.2009.08.079]
- Van Thiel DH, Hagler NG, Schade RR, Skolnick ML, Heyl AP, Rosenblum E, Gavaler JS, Penkrot 24 RJ. In vivo hepatic volume determination using sonography and computed tomography. Validation and a comparison of the two techniques. Gastroenterology 1985; 88: 1812-1817 [PMID: 3888769 DOI: 10.1016/0016-5085(85)90005-8]
- 25 Yu HC, You H, Lee H, Jin ZW, Moon JI, Cho BH. Estimation of standard liver volume for liver transplantation in the Korean population. Liver Transpl 2004; 10: 779-783 [PMID: 15162473 DOI: 10.1002/Lt.20188]
- 26 Heinemann A, Wischhusen F, Püschel K, Rogiers X. Standard liver volume in the Caucasian population. Liver Transpl Surg 1999; 5: 366-368 [PMID: 10477836 DOI: 10.1002/Lt.500050516]
- Du Bois D, Du Bois EF. A formula to estimate the approximate surface area if height and weight be 27 known. 1916. Nutrition 1989; 5: 303-311; discussion 312 [PMID: 2520314]
- Lin XZ, Sun YN, Liu YH, Sheu BS, Cheng BN, Chen CY, Tsai HM, Shen CL. Liver volume in 28 patients with or without chronic liver diseases. Hepatogastroenterology 1998; 45: 1069-1074 [PMID: 9756008
- 29 DeLand FH, North WA. Relationship between liver size and body size. Radiology 1968; 91: 1195-1198 [PMID: 5699624 DOI: 10.1148/91.6.1195]
- Choukèr A, Martignoni A, Dugas M, Eisenmenger W, Schauer R, Kaufmann I, Schelling G, Löhe F, 30 Jauch KW, Peter K, Thiel M. Estimation of liver size for liver transplantation: the impact of age and gender. Liver Transpl 2004; 10: 678-685 [PMID: 15108261 DOI: 10.1002/Lt.20113]
- 31 Vauthey JN, Abdalla EK, Doherty DA, Gertsch P, Fenstermacher MJ, Loyer EM, Lerut J, Materne R, Wang X, Encarnacion A, Herron D, Mathey C, Ferrari G, Charnsangavej C, Do KA, Denys A. Body surface area and body weight predict total liver volume in Western adults. Liver Transpl 2002; 8: 233-240 [PMID: 11910568 DOI: 10.1053/jlts.2002.31654]
- 32 Hashimoto T, Sugawara Y, Tamura S, Hasegawa K, Kishi Y, Kokudo N, Makuuchi M. Estimation of standard liver volume in Japanese living liver donors. J Gastroenterol Hepatol 2006; 21: 1710-1713 [PMID: 16984594 DOI: 10.1111/j.1440-1746.2006.04433.x]
- 33 Chan SC, Liu CL, Lo CM, Lam BK, Lee EW, Wong Y, Fan ST. Estimating liver weight of adults by body weight and gender. World J Gastroenterol 2006; 12: 2217-2222 [PMID: 16610024 DOI: 10.3748/wjg.v12.i4.2217
- Yuan D, Lu T, Wei YG, Li B, Yan LN, Zeng Y, Wen TF, Zhao JC. Estimation of standard liver 34 volume for liver transplantation in the Chinese population. Transplant Proc 2008; 40: 3536-3540 [PMID: 19100432 DOI: 10.1016/j.transproceed.2008.07.135]
- Poovathumkadavil A, Leung KF, Al Ghamdi HM, Othman Iel H, Meshikhes AW. Standard formula 35 for liver volume in Middle Eastern Arabic adults. Transplant Proc 2010; 42: 3600-3605 [PMID: 21094823 DOI: 10.1016/j.transproceed.2010.07.098]
- Um EH, Hwang S, Song GW, Jung DH, Ahn CS, Kim KH, Moon DB, Park GC, Lee SG. Calculation 36 of standard liver volume in Korean adults with analysis of confounding variables. Korean J Hepatobiliary Pancreat Surg 2015; 19: 133-138 [PMID: 26693231 DOI: 10.14701/kjhbps.2015.19.4.133]
- Mosteller RD. Simplified calculation of body-surface area. N Engl J Med 1987; 317: 1098 [PMID: 37 3657876 DOI: 10.1056/NEJM198710223171717]
- 38 Midorikawa T, Kondo M, Beekley MD, Koizumi K, Abe T. High REE in Sumo wrestlers attributed to large organ-tissue mass. Med Sci Sports Exerc 2007; 39: 688-693 [PMID: 17414807 DOI: 10.1249/mss.0b013e31802f58f6]
- Abe T, Kearns CF, Fukunaga T. Sex differences in whole body skeletal muscle mass measured by 39 magnetic resonance imaging and its distribution in young Japanese adults. Br J Sports Med 2003; 37: 436-440 [PMID: 14514537 DOI: 10.1136/bjsm.37.5.436]
- Feng LM, Wang PQ, Yu H, Chen RT, Wang J, Sheng X, Yuan ZL, Shi PM, Xie WF, Zeng X. New 40 formula for predicting standard liver volume in Chinese adults. World J Gastroenterol 2017: 23: 4968-4977 [PMID: 28785151 DOI: 10.3748/wjg.v23.i27.4968]
- 41 Zhou F, Zhou J, Wang W, Zhang XJ, Ji YX, Zhang P, She ZG, Zhu L, Cai J, Li H. Unexpected Rapid Increase in the Burden of NAFLD in China From 2008 to 2018: A Systematic Review and Meta-Analysis. Hepatology 2019; 70: 1119-1133 [PMID: 31070259 DOI: 10.1002/hep.30702]
- 42 Loomba R, Sanyal AJ. The global NAFLD epidemic. Nat Rev Gastroenterol Hepatol 2013; 10: 686-690 [PMID: 24042449 DOI: 10.1038/nrgastro.2013.171]
- Li J, Zou B, Yeo YH, Feng Y, Xie X, Lee DH, Fujii H, Wu Y, Kam LY, Ji F, Li X, Chien N, Wei M, 43 Ogawa E, Zhao C, Wu X, Stave CD, Henry L, Barnett S, Takahashi H, Furusyo N, Eguchi Y, Hsu YC, Lee TY, Ren W, Qin C, Jun DW, Toyoda H, Wong VW, Cheung R, Zhu Q, Nguyen MH. Prevalence, incidence, and outcome of non-alcoholic fatty liver disease in Asia, 1999-2019: a systematic review and meta-analysis. Lancet Gastroenterol Hepatol 2019; 4: 389-398 [PMID: 30902670 DOI: 10.1016/S2468-1253(19)30039-1]
- McCormack L, Dutkowski P, El-Badry AM, Clavien PA. Liver transplantation using fatty livers: 44 always feasible? J Hepatol 2011; 54: 1055-1062 [PMID: 21145846 DOI: 10.1016/j.jhep.2010.11.004]



- 45 Wong TC, Fung JY, Chok KS, Cheung TT, Chan AC, Sharr WW, Dai WC, Chan SC, Lo CM. Excellent outcomes of liver transplantation using severely steatotic grafts from brain-dead donors. Liver Transpl 2016; 22: 226-236 [PMID: 26359934 DOI: 10.1002/lt.24335]
- 46 Chen TY, Chen CL, Tsang LL, Huang TL, Wang CC, Concejero AM, Lu CH, Cheng YF. Correlation between hepatic steatosis, hepatic volume, and spleen volume in live liver donors. Transplant Proc 2008; 40: 2481-2483 [PMID: 18929772 DOI: 10.1016/j.transproceed.2008.08.045]
- 47 Busetto L, Tregnaghi A, De Marchi F, Segato G, Foletto M, Sergi G, Favretti F, Lise M, Enzi G. Liver volume and visceral obesity in women with hepatic steatosis undergoing gastric banding. Obes Res 2002; 10: 408-411 [PMID: 12006641 DOI: 10.1038/oby.2002.56]



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SYSTEMATIC REVIEWS

Microbiota shaping — the effects of probiotics, prebiotics, and fecal microbiota transplant on cognitive functions: A systematic review

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Abstract

BACKGROUND

Dementia is a chronic progressive neurological disease affecting millions of people worldwide, and represents a relevant economic burden for healthcare systems. Although its pathogenesis is still unknown, recent findings have reported that a dysregulated gut-brain axis communication, a fundamental relationship mediated by several host and microbial molecules, is associated with cognitive disorders. In addition, gut microbiota manipulation reduces neuroinflammation, improving cognitive function by restoring the functional gut-brain axis.

AIM

To better define the effects of probiotics, prebiotics, synbiotics, and fecal microbiota transplant (FMT) on cognitive function.

METHODS

We performed a literature search of human randomized clinical trials to examine the effects of the administration of probiotics, prebiotics, synbiotics, or FMT on cognition outcomes in healthy or sick people of every age, sex, and nationality. We systematically searched Embase, Medline/PubMed, Cochrane Library, central and clinicaltrials.gov databases with a combination of comprehensive terms related to cognition and gut microbiota manipulation. Then we carefully reviewed and synthesized the data by type of study design and setting, characteristics of the studied population, kind of intervention (strain type or mixture type, dosage, and frequency of administration), control treatment, inclusion and exclusion criteria, follow-up duration, and cognitive or memory outcomes.

RESULTS

After examining the titles and abstracts, the initial literature screening identified



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995 articles, but we added 23 papers in our systematic review. The analyses of these selected studies highlighted that both probiotic supplementation and FMT improved cognitive function regardless of the type and posology of administration and the adopted cognitive tests and questionnaires. We found that most of the studies conducted in healthy people showed a significant positive effect of the intervention on at least one of the performed cognitive tests. Regarding unhealthy subjects, while FMT and especially probiotic administration had multiple beneficial effects on different cognitive functions, supplementation with prebiotics did not provide any cognitive improvement.

CONCLUSION

Probiotic supplementation and FMT may represent a promising strategy to restore gut eubiosis and enhance the cognitive functions of healthy people and patients with neurological disorders.

Key Words: Dementia; Cognitive disorders; Gut microbiota; Probiotics; Prebiotics; Fecal microbiota transplant

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Core Tip: Dementia and cognitive impairment are age-related conditions that are on the rise worldwide. Recent studies have demonstrated the existence of a gut-brain axis and that the manipulation of gut microbiota composition can exert positive effects on cognition. The administration of probiotics, prebiotics, and fecal microbiota transplant may represent a good strategy to counteract gut dysbiosis and ameliorate cognitive dysfunction by reducing neuroinflammation and brain damage.

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INTRODUCTION

Global population ageing, defined as the increasing proportion of older people around the globe, represents a deep shift in society and a considerable challenge for the sustainability of healthcare systems due to the rise of geriatric illnesses[1,2]. Currently, the prevalence of cognition impairment, particularly dementia, is estimated worldwide in 50 million people with an economic burden of 818 billion dollars in 2016 and a forecast of about 115 million people by 2050[3,4].

Dementia is an acquired, gradual, and progressive disorder involving multiple adverse neurocognitive changes that can affect learning processes, memory, executive function, language, complex attention, mood, perceptual-motor function or social cognition. Moreover, although its detailed pathological mechanism is still not well understood, dementia often occurs in association with advanced age or the presence of contributing causes, usually Alzheimer's disease (AD), Parkinson's disease (PD), or cerebrovascular pathology[5-7]. Unfortunately, current therapies are only symptomatic, and notably, no treatment stops the disease progression[8].

Recent studies have shown that that the gut microbiota, with more than 100 trillion microorganisms carrying three times of human genes, plays a pivotal role in human health; manipulation of the intestinal microbiota can modify the release of neuroactive metabolites, which affect brain health[9,10]. This role can be further explained by the documented existence of the gut-brain axis, a complex bidirectional system in which communication occurs through three parallel and interplaying pathways that involve nervous, endocrine, and immune signals[11]. Therefore, different preclinical and observational studies have demonstrated that the gut dysbiosis is responsible for increased intestinal permeability, which correlates with both neuroinflammation and a decline of cognitive abilities[12-15].



Dietary interventions (in nutritional supplements or specific diets) have often been applied in clinical practice to restore intestinal eubiosis and prevent and treat cognitive disorders. For example, a Mediterranean diet and/or a healthy diet based on fruits, vegetables, and fish seems to stabilize or slow cognitive decline[16].

Nevertheless, the most promising strategy to counteract gut dysbiosis and to maintain cognitive function seems represented by the administration of probiotics, prebiotics, and fecal microbiota transplant (FMT).

Interestingly, administration in animal models of an adequate posology of multistrain probiotics reduces both Firmicutes/Bacteroidetes ratio and intestinal permeability, slowing cognitive decline and reducing neuroinflammation[17,18]. Moreover, using specific prebiotics seems to ameliorate cognitive performance with a direct effect on gut microbiota[19].

Moreover, even FMT has demonstrated remarkable efficacy in healthy subjects and people affected by various diseases caused by gut microbiota perturbation, particularly Clostridium difficile infection. It could represent a promising therapy for cognitive impairment improvement because of its capability to re-establish a healthy gut microbial community[20,21].

Therefore, since the evidence derived from human randomized clinical trials (RCTs) is currently limited, this systematic review identified the available RCTs and better defined the effects of probiotics, prebiotics, synbiotics, and FMT on cognitive functions.

MATERIALS AND METHODS

Literature search

Our study followed the PRISMA statement guidelines. A computerized search of the articles published until 24 October 2019 was conducted in Embase, Medline/PubMed, Cochrane Library, central and clinicaltrials.gov databases and other individual journal sources, using the following search string: (memory OR cognition OR dementia) AND (lactobacillus OR bifidobacteria OR streptococcus OR enterococcus OR probiotic OR prebiotic OR symbiotic OR fecal, transplantation). In the PubMed database, we activated the filter "Humans"; in Embase, we selected the filter "Research articles"; in Cochrane Library, we activated the filter "Trials"; and in clinicaltrials.gov, we selected the filter "recruitment: terminated or completed." The search did not apply filters for language, country, duration of follow-up, and participants' characteristics (age and sex).

Study selection

Two authors independently reviewed the titles and abstracts of the collected articles, applying predefined inclusion/exclusion criteria. The inclusion criteria were as follows: RCTs; availability of full text; patients regardless of age, nationality, sex, and health status; comparison between oral intake of probiotics, prebiotics, or symbiotic and control treatment or placebo; and outcome as cognitive or memory evaluation. The adopted exclusion criteria were as follows: Studies with fewer than 10 participants; reviews, articles, and case reports; or studies with incomplete outcomes.

Data extraction

The same two authors performed analyses of the full text and data extraction with the intervention of a third author in case of poor agreement or discrepancies. Each reviewer independently recorded the data in a predefined data extraction form. The following data, if reported, were obtained from each selected trial: First author name, year of publication, study design, setting (institution, city, and country), characteristics of the studied population (mainly age and health status), number of total participants and their gender, number of subjects in both treatment and control groups, characteristics of the intervention (strain type or mixture type, dosage and frequency of administration), control treatment, inclusion and exclusion criteria, follow-up duration, cognitive or memory outcomes and compliance data.

Outcome assessment

For each selected study, cognitive functions were assessed through specific tests which evaluated the eight main cognitive skills: sustained attention, speed of information processing, cognitive flexibility and control, multiple simultaneous attention, working memory (short-term memory), category formation, pattern recognition, and response



inhibition[22]. A detailed description of all cognitive tests performed in the selected papers for this systematic review is annexed in Supplementary material.

RESULTS

Study selection

The initial literature screening identified 995 papers. Eight studies were excluded for duplication and another 964 papers were removed after the title and abstract screening because they did not respect inclusion criteria. The selection process, in accordance with the PRISMA statement 2009, is illustrated in Figure 1.

Characteristics of the included studies

An overview of the 23 studies included in this systematic review is reported in Table 1. All 23 included papers were RCTs published from 2007 to 2019[23-45]. The total number of participants was 1285 (491 males and 650 females); unfortunately, both articles published by Tamtaji *et al*[44,45] did not report the gender of the participants. Regarding the age of the enrolled subjects, one study was conducted in healthy scholars (7-9 years)[31], four studies enrolled young adults (19-30 years)[25,36,40,43], and most of the studies involved adults or older people (48-95 years)[23,24,26-30,32-35, 37-39,40,42,44,45]. Most studies (four) were performed in Iran[22,24,44,45]; three in the United States[26-28] and Japan[34,37,39]; two in the United Kingdom[29,31], South Korea[33,35], and Spain[30,41]; and one in Austria[25], Italy[32], Ireland[36], Malaysia [38], Poland[42], Wales[43] and the Netherlands[40].

Concerning the patients' health state, most studies enrolled healthy people[25,29,31, 33-36,40,43], whereas three studies involved patients with AD[23,24,44] or cirrhotic subjects with recurrent encephalopathy[26-28]. The other studies were focused on stressed adults[38], patients with PD[45], human immunodeficiency virus (HIV)-1-infected individuals[32], subjects affected by fibromyalgia syndrome[41], people with major depression[42], elderly with frailty syndrome[30], and adults with forgetfulness [39] and mild cognitive impairment[37].

In the trials, subjects were administered probiotics[22-25,26,29,32-42,44,45], prebiotics[30,31,43], or FMT[27,28] and its duration lasted a maximum of 24 wk[32] and a minimum of 4 h[43]; however, the trials continued for 12 wk for most of the studies[23,24,31,33-35,37,38,44,45]. No studies have reported the administration of synbiotics. In the studies examining the effects of probiotics, a total of 21 different bacterial species were administered (alone or in combination) in a dosage ranging from 1×10^{9} CFU/mL to 2.5×10^{10} CFU/mL; the most represented species were *Lactobacillus plantarum*, *L. acidophilus*, and *Bifidobacterium bifidum*. On the other side, the administered prebiotics was composed of inulin or galacto-oligosaccharides (GOS), in a dosage that ranged from 5 g/d to 7.5 g/d. In FMT studies, subjects were administered a capsule containing 550 µL stool and buffer solution or enema infusion of 90 mL FMT solution. Lastly, only five studies reported their compliance[31,33-35,44,45], and it was generally considered high because it ranged from 82.69% to 100%.

Effects of probiotics and prebiotics on the cognitive functions of healthy people

Regarding the healthy subjects, three studies showed no significant difference between probiotic and placebo groups[29,31,36]. In comparison, five studies showed a significant positive effect of the intervention on at least one of the performed cognitive tests[25,33,35,40,43].

In Benton *et al*[29], the healthy enrolled subjects ingested fermented milk containing *L. casei* Shirota daily for 3 wk. However, no significant differences between the probiotic and placebo groups were reported regarding episodic and long-term memory, assessed with the Wechsler Memory Scale test and the ability to remember the capitals of 30 countries. Moreover, the healthy people treated with *L. rhamnosus* supplement in Kelly *et al*[36] did not report any cognitive improvement, as assessed with the Paired Associates Learning, Attention Switching Task, Rapid Visual Information-Processing task (RVIP), Emotion Recognition Task and electroencephalography tests.

Considering the five studies reporting a significant cognitive improvement, Bagga *et al*[25] found that 4 wk administration of a multistrain probiotic increased Positive and Negative Affect Schedule (PANAS) score (paired with the response accuracy to unpleasant stimuli test) and showed the activation of the cingulum, pre-cuneum and cerebellum areas, involved in decision making and memory process.

Table 1 Summ	Table 1 Summarizing of all selected studies									
Ref.	Study design	Setting	Characteristics of the studied population	Number of participants (M/F)	Intervention	Comparison	Duration of intervention	Outcomes	Compliance	
Agahi <i>et al</i> [<mark>23</mark>], 2018	RCT	Cities: Emam Ali, Golabchi, Miad, Barekat; Country: Iran	Patients with Alzheimer disease; Age: $65-90 \text{ yr}$; Control group: 80.57 ± 1.79 yr; Intervention group: $79.70 \pm 1.72 \text{ yr}$	Total: 48; Control group = 23 (10/13); Intervention group = 25 (7/18)	1 capsule with <i>L. fermentum</i> , <i>L. plantarum</i> , <i>B. lactis</i> and 1 capsule with <i>L. acidophilus</i> , <i>B. bifidum</i> , and <i>B. longum</i> (3 × 10 ⁹ CFU)	Placebo	12 wk	ТҮМ	-	
Akbari <i>et al</i> [<mark>24</mark>], 2016	RCT	Cities: Golabchi, Sadeghyeh; Country: Iran	Patients with Alzheimer disease; Age: 60-95 yr; Control group: 82.00 ± 1.69 yr; Intervention group: 77.67 ± 2.62 yr	Total: 60; Control group = 30 (24/6); Intervention group = 30 (24/6)	200 mL/d probiotic milk containing <i>L. acidophilus, L. casei, B.</i> <i>bifidum,</i> and <i>L. fermentum</i> (2 × 10 ⁹ CFU each)	Placebo	12 wk	MMSE	-	
Bagga <i>et al</i> [25], 2018	RCT	City: Graz Country: Austria	Healthy volunteers; Age: 20- 40 yr; Control group (placebo): 27.25 ± 5.78 yr; No intervention group: 23.87 ± 4.97 yr; Intervention group: 28.27 ± 4.2 yr	Total: 45; Control group = 15 (9/6); No intervention group = 15 (7/8); Intervention group = 15 (7/8)	1 sachet/d with 3 g freeze-dried powder containing <i>L. casei</i> W56, <i>L.</i> <i>acidophilus</i> W22, <i>L. paracasei</i> W20, <i>B. lactis</i> W51, <i>L. salivarius</i> W24, <i>L.</i> <i>lactis</i> W19, <i>B. lactis</i> W52, <i>L.</i> <i>plantarum</i> W62 and <i>B. bifidum</i> W23(7.5 × 10 ⁶ CFU/g)	Placebo or no intervention	4 wk	PANAS, SCL-90, ADS, LEIDS, RM task, ED task	-	
Bajaj et al[<mark>26</mark>], 2014	RCT	City: Richmond, Virginia Country: United States	Patients with hepatic encephalopathy; Age: 18-65 yr; Control group: 58.5 ± 4.5 yr; Intervention group: $58.4 \pm$ 3.8 yr	Total: 30; Control group = 16 (12/4); Intervention group = 14 (10/4)	L. rhamnosus GG (ATCC 53103)(> 50 billion CFU/gm)	Placebo	8 wk	NCT-A, NCT-B, DS, BDT	-	
Bajaj et al[<mark>27</mark>], 2017	RCT	City: Richmond, Virginia Country: United States	Patients with hepatic encephalopathy; Mean age: 62 yr; Control group: 62.9 ± 9.8 yr; Intervention group: 64.5 ± 5.1 yr	Total: 20; Control group = 10 (10/0); Intervention group = 10 (10/0)	FMT units (90 mL total) instilled by enema and retained for 30 min	Standard of care	20 wk	EncephalApp-Stroop, PHES	-	
Bajaj et al[<mark>28</mark>], 2019	RCT	City: Richmond, Virginia Country: United States	Patients with hepatic encephalopathy; Control group: 64.2 ± 6.2 yr; Intervention group: 63.3 ± 4.2 yr	Total: 20; Control group = 10 (8/2); Intervention group = 10 (8/2)	FMT capsules (550 µL of stool and buffer solution)	Placebo	20 wk	EncephalApp-Stroop, PHES	-	
Benton <i>et al</i> [29], 2007	RCT	City: Swansea Country: Wales	Healthy volunteers; Age: 48- 79 yr; Average age 61.8 ±7.3 yr	Total: 126 (51/75)	65 mL of milk drink containing <i>L. casei</i> Shirota (10 ⁸ /mL)	Placebo	3 wk	POMS, WMS, VFT, NART, Ability to recall the capital cities of countries	-	
Buigues <i>et al</i> [30], 2016	RCT	City: Valencia Country: Spain	People with frailty syndrome; Age: 66-90 yr; Control group: 73.4 ± 1.8 yr; Intervention group: 74.2 ± 1.6 yr	Total: 50; Control group = 22 (6 /16); Intervention group = 28 (9/19)	7.5 g/d of Darmocare Pre® (Inulin 3375 mg, FOS 3488)	Placebo	13 wk	MMSE	-	

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Capitão <i>et al</i> [<mark>31</mark>], 2020	RCT	Cities: Swindon, Milton Keynes, London Country: United Kingdom	Healthy scholars; Age: 7-9 yr; Control group: 9.12 ± 1.02 yr; Intervention group: 8.54 ± 0.79 yr		5.5 g/d of Bimuno (B-GOS, Lactose, Glucose, Galactose)	Placebo	12 wk	BAS-III, CogTrackTM battery, STAIC, MFQ	High (> 80%)
Ceccarelli <i>et al</i> [32], 2017	RCT	City: Rome Country: Italy	HIV-1 infected individuals; Median age: 48 (IQR: 38-54) yr; Intervention group: 45 (35-52.5) yr; Control group: 43 (38.2-53) yr	Total: 35; Control group = 26 (24/2); Intervention group = 9 (9/0)	Sachet containing <i>L. plantarum</i> DSM 24730 <i>S. thermophilus</i> DSM 24731, <i>B. breve</i> DSM 24732, <i>L.</i> <i>paracasei</i> DSM 24733, <i>L. delbrueckii</i> <i>subsp. bulgaricus</i> DSM 24734, <i>L.</i> <i>acidophilus</i> DSM 24735 <i>B. longum</i> DSM 24736, and <i>B. infantis</i> DSM 24737(450 × 10 ⁹ bacteria)	Control group	24 wk	ROCF, RAVLT, STEP, VST, PVF, SVF, SPM, DS, CBTT, AAT, TMT A, TMT B	-
Chung <i>et al</i> [33], 2014	RCT	City: Jeonju Country: Korea	Healthy volunteers; Age: 60- 75 yr; Control group: $64.50 \pm$ 4.84 yr; Intervention group (500 mg): 64.50 ± 2.17 yr; Intervention group (1000 mg): 64.43 ± 4.47 yr; Intervention group (2000 mg): 66.56 ± 4.98 yr	Total: 36; Control group = 10 (4/6); Intervention group (500 mg) = 10 (9/1); Intervention group (1000 mg) = 7 (2/5); Intervention group (2000 mg) = 9 (5/4)	Daily doses of 500, 1000, or 2000 mg. of tablet containing <i>L.</i> <i>helveticus</i> IDCC3801	Placebo	12 wk	DS, SRT, VLT, RVIP, SCWT	> 70%
Inoue <i>et al</i> [<mark>34]</mark> , 2018	RCT	City; Hyogo prefecture, Country: Japan	Healthy volunteers; Average age: 70.3 ± 3.1 yr; Control group: 70.9 ± 3.2 yr; Intervention group: 69.9 ± 3.0 yr	Total: 38; Control group = 18 (7/11); Intervention group = 20 (7/13)	Sachet containing lyophilised powder of <i>B. longum</i> BB536, <i>B. infantis</i> M-63, <i>B. breve</i> M-16V and <i>B.breve</i> B-3 (1.25 × 10 ¹⁰ CFU each)	Placebo	12 wk	MoCA, Modified flanker task, PHQ-9, GAD-7	> 99%
Hwang <i>et al</i> [35], 2019	RCT	City: Jeonju Country: South Korea	People with mild cognitive impairment; Age: 55-85 yr; Control group: 69.2 ± 7.00 yr; Intervention group: $68.0 \pm$ 5.12 yr	Total: 100; Control group = 50 (14/36); Intervention group = 50 (20/30)	Mixture of fermented soybean powder and <i>L. plantarum</i> C29 (1.25 \times 10 ¹⁰ CFU/g)	Placebo	12 wk	VLT, DS, ACPT	> 90%
Kelly <i>et al</i> [<mark>36</mark>], 2017	RCT	City: Cork Country: Ireland	Healthy volunteers; Age: 20- 33 yr; Placebo/Probiotic group: 23.6 ± 0.97 yr; Probiotic/Placebo group: 25.64 ± 1.14 yr	Total: 29; Placebo/Probiotic group = 15 (15/0); Probiotic/Placebo group = 14 (14/0)	Active capsules contained corn starch, magnesium stearate, silicon dioxide and <i>L. Rhamnosus</i> (1×10^9 CFU)	Placebo	8 wk	MOT, PAL, AST, RVIP, ERT, Emotional Stroop	-
Kobayashi <i>et al</i> [37], 2019	RCT	City: Tokyo Country: Japan	People with memory complaints; Age: 50-80 yr; Control group: 61.6 ± 6.37 yr; Intervention group: 61.5 ± 6.83 yr	Total: 117; Control group = 58 (29/29); Intervention group = 59 (29/30)	1 capsule per day with <i>B. breve</i> A1 (> 2 × 10^{10} CFU)	Placebo	12 wk	RBANS, MMSE	-
Lew <i>et al</i> [<mark>38</mark>], 2019	RCT	Cities: Penang, Kubang Kerian Country: Malaysia	Stressed adults; Age: 18-60 yr; Control group: 32.1 ± 11.4 yr; Intervention group: 31.3 ± 10.8 yr	51 (12/39); Probiotic group	<i>L. plantarum</i> P8 (10 ¹⁰ CFU/sachet per day)	Placebo	12 wk	PSS-10, DASS-42, CBB	-
Ohsawa et al [<mark>39</mark>], 2018	RCT	Country: Japan	People with forgetfulness; 50-70 yr; Control group: 57.8	Total: 60; Control group = 29 (13/16); Intervention	One bottle per day (190 g per bottle) of a <i>L. helveticus</i> -fermented	Placebo	8 wk	RBANS, POMS	-

			± 5.9 yr; Intervention group: 58.5 ± 6.5 yr	group = 31 (13/18)	milk contained 2.4 mg of lactononadecapeptide				
Papalini <i>et al</i> [<mark>40</mark>], 2019	I RCT	City: Nijmegen Country: The Netherlands	Healthy volunteers; Age:18- 40 yr; Control group: 22 yr (SE = 0.5); Intervention group: 21 yr (SE = 0.4)	Total: 58; Control group = 29 (0/29); Intervention group = 29 (0/29)	2 g/d of powder diluted in water or milk containing <i>B. bifidum</i> W23, <i>B. lactis</i> W51, <i>B. lactis</i> W52, <i>L.</i> <i>acidophilus</i> W37, <i>L. brevis</i> W63, <i>L.</i> <i>casei</i> W56, <i>L. salivarius</i> W24, <i>L.</i> <i>lactis</i> W19 <i>L. lactis</i> W58(5 × 10 ⁹ CFU)	Placebo	4 wk	BDI, LEIDS-r, Emotional face-word Stroop task, Emotional face-matching paradigm, SCWT, DS, SECPT	-
Roman <i>et al</i> [<mark>41]</mark> , 2018	RCT	City: Almerìa Country: Spain	Fibromyalgia patients; Control group: 50.27 ± 2.03 yr; Intervention group: 55.00 ± 2.09 yr	Total: 31; Control group = 15 (2/13); Intervention group = 16 (1/15)	4 pills/d containing L. Rhamnosus GG^{\oplus} , L. casei, L. acidophilus, and B. Bifidus (6 × 10 ⁶ bacteria per capsule)	Placebo	8 wk	MMSE, BDI, IGT, Two-choice Task	-
Rudzki <i>et al</i> [42], 2019	RCT	City: Bialystok Country: Poland	People with major depression; Control group: 38.90 (12) yr (SD); Intervention group: 39.13 (9.96) yr	Total: 60; Control group = 30 (10/20); Intervention group = 30 (7/23)	2 capsules/d containing <i>L.</i> <i>plantarum</i> 299v (10 × 10 ⁹ CFU per capsule)	Placebo	8 wk	HAM-D 17, SCL-90, PSS-10, APT, RFFT, TMT A, TMT B, CVLT Stroop Test parts A and B	-
Smith <i>et al</i> [4 3 2015	3], RCT	City: Cardiff Country: Galles	Healthy volunteers; Age: 19- 30 yr; Mean age 23.0 yr	Total: 47 (19/28)	One sachet of Inulin per day (5 mg)	Placebo	4 h	Mood, Performance Tasks, Memory Tasks, Psychomotor Tasks, Selective Attention Tasks, Sustained Attention Task	-
Tamtaji et al [44], 2019	RCT	City: Kashan, Shahrekord Country: Iran	Patients with Alzheimer disease; Age: 55-100 yr; Control group: 78.5 ± 8.0 yr; Intervention group (Selenium): 78.8 ± 10.2 yr; Intervention group (Selenium + probiotic): 76.2 ± 8.1 yr	Total: 79; Control group = 26; Intervention group (Selenium) = 26; Intervention group (Selenium + probiotic) = 27	Selenium (200 μ g/d) and probiotic containing <i>L. acidophilus</i> , <i>B. bifidum</i> , and <i>B. longum</i> (2 × 10 ⁹ CFU/d each)	Placebo or only selenium (200 µg/d)	12 wk	MMSE	100%
Tamtaji <i>et al</i> [45], 2019	RCT	City: Kashan Country: Iran	Patients with Parkinson disease; Age: 50-90 yr; Control group: 67.7 ± 10.2 yr; Intervention group: 68.2 ± 7.8 yr	Total: 60; Control group = 30; Intervention group = 30	Probiotic containing <i>L. acidophilus</i> , <i>B. bifidum</i> , <i>L. reuteri</i> , and <i>L.</i> <i>fermentum</i> (each 2×10^9 CFU/g)	Placebo	12 wk	MDS-UPDRS	90%

AAT: Aachener Aphasia Test; ACPT: Auditory Continuous Performance Test; ADS: Allgemeine Depressionsskala; APT: Attention and Perceptivity Test; AST: Attention Switching Task; BAS-III: British Ability Scales III; BDI: Beck Depression Inventory; BDT: Block Design Test; CBB: Cogstate Brief Battery; CBT: Corsi Block Tapping Test; CFU: Colony-forming unit; CVLT: California Verbal Learning Test; DASS-42: Depression Anxiety and Stress Scale questionnaire; DS: Digit Symbol Test; ED: Emotional Decision Making; ERT: Emotion Recognition Task; F: Female; GAD-7: Generalised Anxiety Disorder Questionnaire-7; HAM-D 17: Hamilton Depression Rating-17; IGT: Iowa Gambling Task; LEIDS: Leiden Index of Depression Severity; M: Male; MDS-UPDRS: Movement Disorders Society-Unified Parkinson's Disease Rating Scale; MMSE: Mini Mental State Evaluation; MoCA: Montreal Cognitive Assessment instrument; MOT: Motor Screening Test; NART: National Adult Reading Test; NCT-A: Number Connection Test A; NCT-B: Number Connection Test B; PAL: Paired Associates Learning; PANAS: Positive and Negative Affect Schedule; PHES: Psychometric Hepatic Encephalopathy Score; PHQ-9: Patient Health Questionnaire-9; POMS: Profile of Mood States; PSS-10: Perceived Stress Scale-10; PVFT: Phonological Verbal Fluency (PVF) test; RAVLT: Rey Auditory Verbal Learning Test; RBANS: Repeatable Battery for the Assessment of Neuropsychological Status; RCT: Randomized controlled trial; RFFT: Ruff Figural Fluency Test; RM: Emotional Recognition Memory; ROCF: Rey-Osterrieth Complex Figure Test; RVIP: Rapid Visual Information-Processing task; SCL-90: Symptom Checklist 90; SCWT: Stroop Color and Word Test; SECPT: Socially Evaluated Cold Pressor Test; SPM: Raven's Standard Progressive Matrices; SRT: Story Recall Test; STAIC: State-Trait Anxiety Inventory for Children; STEP: Test of Time and Weights Estimation; SVF: Semantic Verbal Fluency test; TMT-A: Trail Making Test A; TMT-B: Trail Making Test B; TYM: Test your memory; VFT: Verbal Fluency test; WMS: Wechsler Memory Scale MFQ: Children Mood and Feelings Questionnaire; VLT: Verbal Learning Test; VST: Visual Search Test.

Papalini *et al*[40] tested a probiotic multistrain mixture in women who underwent a stressful condition for 4 wk. The results showed that the trial reduced the unfavorable stress effect on working memory performance measured by the DS backward test. In addition, Chung et al^[33] demonstrated a significant improvement in Verbal Learning Test, Story Recall Test, RVIP and Stroop Color and Word Test after 12 wk administration of *L. helveticus* in healthy subjects compared to placebo. Finally, Inoue *et al*[34] demonstrated that intervention with *Bifidobacterium spp.* for 12 wk, added to resistance training, significantly improved response accuracy and reaction time tests in healthy elderly subjects. Regarding the cognitive effects of prebiotic administration, the study conducted in healthy children by Capitão et al^[31] reported that 12 wk GOS supplement only improved memory retrieval speed assessed with the CogTrackTM test battery. By contrast, Smith *et al*[43] investigated the acute effects of inulin intake on healthy volunteers and reported improving memory tasks, especially immediate free and delayed recall. No FMT intervention has been carried out in healthy subjects. Hence, although three of eight studies conducted in healthy subjects showed no significant difference between intervention and placebo groups, probiotics resulted were more effective in improving cognitive function than prebiotics.

Patients with different pathologies and the impact of probiotics/prebiotics/FMT on cognitive functions

The effects of probiotics, prebiotics, and FMT on cognitive functions were also assessed in different diseases, of which the most represented were hepatic encephalopathy (HE) and AD. Bajaj *et al*[26] conducted three studies on HE. In particular, the authors first investigated the effect of *Lactobacillus GG* administration on HE but did not report changes in cognition. However, they also treated HE patients with FMT *via* enema and reported a significant improvement in PHES and EncephalApp Stroop tests[27]. Moreover, Bajaj *et al*[28] evaluated the treatment with FMT capsules effects on HE patients, and they reported only a significant improvement in the EncephalApp Stroop test.

Regarding AD, Agahi *et al*[23] administered two different multistrain probiotic capsules to patients affected by severe disease for 12 wk, but no effect on TYM cognitive tests were reported. By contrast, Akbari *et al*[24] found that daily administration of probiotic milk enriched with *Lactobacillus spp*. led to a decline in Mini Mental State Evaluation (MMSE) score in AD patients compared to placebo. Moreover, Tamtaji *et al*[44] found that a probiotic and selenium co-supplement in AD patients was responsible for a significant increase in MMSE score. In addition, Hwang *et al*[35] found that people with mild cognitive impairment showed an improvement in a

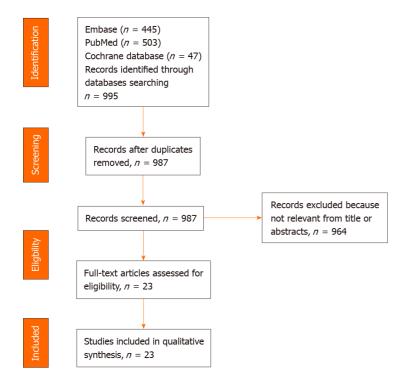


Figure 1 PRISMA flow diagram.

battery of tests related to verbal memory and attention domains after ingesting a mixture of L. plantarum C29 and fermented soybean powder. Finally, Lew et al[38] reported that daily administration of L. plantarum P8 for 12 wk in stressed adults led to a reduction of stress score and enhanced cognition and verbal learning memory, assessed through the CBB. Another study, conducted by Roman et al[41], explored the effect of a multispecies probiotic on fibromyalgia patients and found a significantly reduced number of impulsive choices. Moreover, Kobayashi et al[37] carried out a 12wk treatment with Bifidobacterium breve A1 in elderly subjects with memory complaints, documenting a significant decline of total scores of both Repeatable Battery for the Assessment of Neuropsychological Status (RBANS) and MMSE tests. Regarding patients affected by PD, Tamtaji et al[45] highlighted a favorable reduction of the Movement Disorders Society-Unified PD Rating Scale (MDS-UPDRS) after ingesting a probiotic mixture for 12 wk. In addition, Ohsawa et al[39] reported improved attention, coding, and delayed memory scores (assessed with RBANS) in people with forgetfulness after 8 wk intake of a *L. helveticus* fermented milk drink. Also, Rudzki et al[42] reported a significant improvement in CVLT and APT in people with major depression treated with L. plantarum 299v. Moreover, Ceccarelli et al [32] demonstrated a significant improvement in several cognitive functions in HIV-1 infected patients ingesting a multistrain probiotic for 24 wk (primarily in the following neurocognitive tests: Rey-Osterrieth Complex Figure, Rey Auditory Verbal Learning Test, Test of Time and Weights Estimation, Phonological Verbal Fluency Test, Corsi Block Tapping Test and Trail Making Test A). Finally, the only study which assessed the effectiveness of a prebiotic intake (inulin and fructooligosaccharides), conducted by Buigues *et al*[30] on elderly affected by frailty syndrome, the MMSE did not report significant cognitive improvement. As a result, while the FMT and especially probiotics played multiple beneficial effects on different cognitive functions of unhealthy subjects, the prebiotics' supplementation did not provide any cognitive improvement, maybe because of their short-term administration.

DISCUSSION

Dementia is a chronic, gradual, and progressive neurological disease that affects millions of people in both industrialized and rural countries. Cognitive decline and daily activities impairment limits patients' self-care and causes a severe burden to parents, friends, caregivers, and especially to the healthcare systems [46,47]. Increasing evidence suggests that the prevalence of dementia rises with age and is strongly



associated with other comorbidities, including AD and cardiovascular risk factors such as hypertension and hypercholesterolaemia[48].

Although the specific dementia pathogenesis is not yet understood and current therapies only attempt to counterbalance the disturbance, several studies recently highlighted the central role of the gut microbiota in brain health and the onset and persistence of neurodegenerative diseases[49,50]. Nevertheless, our systematic review of human RCTs reported contradictory results due to the diverse type, posology, and duration of interventions and the different responses of healthy or diseased people to the treatment.

In general, supplementation with probiotics and prebiotics determined the positive effects on healthy subjects. Five (63%) out[25,33,34,40,43] of the eight studies conducted in volunteers reported beneficial effects on the cognitive functions, while the other three [29,31,36] studies did not find any difference between the intervention and control groups.

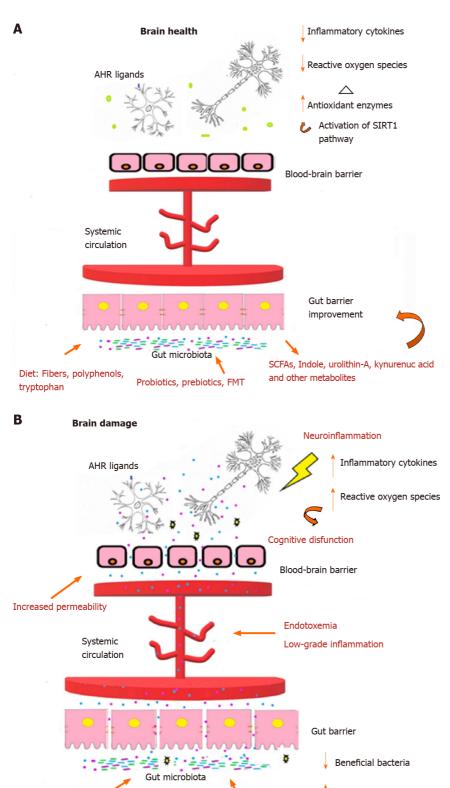
Regarding the different evaluated patients, only 3 (20%)[23,26,30] of the 15 studies did not report an amelioration of cognitive functions for other possible reasons. For example, the probiotics' administration performed by Bajaj et al[26] probably did not last for a sufficient time to obtain cognitive improvement in patients with HE. In contrast, with 13 wk of prebiotics' supplementation, Buigues et al[30] did not observe effects on cognitive behaviour because MMSE does not represent a sensitive tool to detect the small changes in cognition that may occur after inulin and FOS supplementation. In addition, in the study conducted by Agahi et al[23], the 12 wk probiotic administration did not lead to cognitive amelioration in patients with AD; a probable explanation could be the enrollment of only patients with advanced disease.

Probiotic supplementation improves cognitive functions in many different diseases such as HIV[32], PD[45], fibromyalgia, major depression[42], AD[24,44], and other mild cognitive deficits[35,37-39]. Furthermore, studies evaluating the effects of FMT on patients with HE highlighted a significant amelioration in cognitive functionality[27, 28]. It is well established that a balanced gut microbiota composition (eubiosis condition) plays a crucial role in our health; a dysbiotic status (meaning a reduced gut microbiota diversity) is related to many human gastrointestinal, immunological, and neurodegenerative diseases[51]. Concerning neurological impairments, recent findings elucidated the importance of the gut microbiota in the bidirectional communication between the central and enteric nervous systems, called the gut-brain axis[52]. Hence, the main factors responsible for intestinal dysbiosis such as stress, unbalanced diet, and drug abuse also determine an alteration of the gut-brain axis by causing a loss of epithelial integrity. The loss of this barrier functionality allows microbial-derived molecules to enter the systemic circulation, promoting endotoxemia, oxidative stress and low-grade inflammation responsible for the blood-brain barrier disruption[53,54] (Figure 2); these factors represent a signature for neurodegenerative disorders, especially AD. Consequently, given the importance of the intestinal barrier integrity for the prevention of neuroinflammation and brain damage, gut microbiota modulation by psychobiotics, namely beneficial bacteria (probiotics) or support for such bacteria (prebiotics) and FMT, represent an excellent strategy to restore the intestinal permeability and prevent the consequences of a leaky gut[55-57].

However, although several studies have highlighted the local beneficial effects of probiotics, prebiotics and FMT (e.g., modification of the gut microbiota composition, strengthening of the gut epithelial barrier and modulation of the local (mucosal) immune system), they also exerted systemic effects, in particular on the central nervous system[58,60]. More specifically, recent studies have reported that the intestinal microbiota affects neurodevelopment and diverse brain functions by regulating the gut-brain axis, for example, by acting on the electrophysiological thresholds of the enteric nervous system neurons, which interact via neurotransmitters (adrenaline, noradrenaline, and acetylcholine) with the central nervous system[61].

Another important neuronal pathway in gut-brain communication involves the vagus nerve, and many effects of probiotics strains influence its activity [62]. Furthermore, since the gut houses the most extensive collection of lymphoid tissues in the human body and various intestinal immune cells can cross the blood-brain barrier, gut microbiota manipulation represents a key indirect route for communication between the gut microbiota and the central nervous system^[63]. Intriguingly, specific probiotic formulations have also been shown an ability to stimulate the production of neurotransmitters (e.g., GABA, serotonin, and dopamine) or are even microbially neuroactive. These microbial metabolites can trigger epigenetic signals on human brain genes involved in various complex networks or act as a ligand for specific human receptors[64].





Stress, aging, drugs, unhealthy diet

Pathogenic bacteria

Figure 2 Gut-brain axis in eubiosis and dysbiosis condition. A: Eubiosis; B: Dysbiosis condition. AHR: Aryl hydrocarbon receptor; FMT: Fecal microbiota transplant; SCFA: Short-chain fatty acid; SIRT1: Sirtuin 1.

> For instance, the probiotic activated Sirtuin 1 pathway, which regulates the brain antioxidant enzymes such as superoxide dismutase and glutathione peroxidase, could favor cognitive improvements by preventing oxidative stress and deposition of betaamyloid in the brain[65]. Even the modulation of kynurenine metabolism, the primary route for tryptophan catabolism, which is closely related to the structural and functional dynamics of the gut microbiota, could positively affect brain health[66]. Indeed, in vivo L. plantarum administration demonstrated a beneficial reduction of

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Increased permeability

kynurenines as most act as neurotoxic compounds [42,67,68]. Notably, although L. plantarum was administered to healthy or ill people in most of our selected RCTs, its positive effects have been probably underestimated because of the unknown impact of the other components of the probiotic formula that include it.

By contrast, indole-3-lactic acid (ILA) is an interesting neuroprotective tryptophan metabolite mainly produced by *Bifidobacterium spp.* acting as an aryl hydrocarbon receptor (AhR) agonist, expressed by intestinal and neuronal cells [69,70]. In detail, microbial agonists produced by L. bulgaricus and L. reuteri could activate microglia and astrocytes AhRs, suppressing pro-inflammatory signals and preventing neuronal damage[71-73]. Furthermore, the administration of some probiotics (especially L. helveticus, L. casei and L. rhamnosus) and prebiotics could also improve cognitive functions by stimulating the production of short-chain fatty acids as they enhance the transcription of the brain-derived neurotrophic factor that stimulates neuronal plasticity, protecting against neuroinflammation and neuronal apoptosis[74-78]. Moreover, the FMT represents a very promising strategy to re-establish gut eubiosis and improve cognitive functions. For instance, in transgenic mice, FMT significantly improved cognitive deficits, beta-amyloid accumulation, and neuroinflammation while reducing UPDRS score and tremor in people with Parkinson disease[79].

Finally, our recent study demonstrated that age-associated shifts of the microbiota have a detrimental impact on the central nervous system's protein expression and critical functions. Still, FMT represents an excellent strategy to restore a young-like microbiota and improve cognitive functions[80]. Therefore, although the modulation of intestinal microbiota represents a new precious therapeutic opportunity, it also shows some restrictions and risks. In particular, even if probiotics are generally considered safe and have many advantages such as a tolerated mode of administration (orally) and the possibility to integrate them with other pharmacological/non-pharmacological approaches, they displayed some limitations mainly due to potential side effects, especially in some patients (including immunocompromised people), or to their long-term safety[81]. Besides, even if probiotics can promote the production of several compounds such as lactic acid, bioamines, bile salts and other molecules that could play detrimental effects on the host, most of them are sold as dietetic supplements, and the regulatory agencies do not require safety studies in humans before their commercialization[82,83].

Although reported to be fairly safe in most clinical trials, FMT can be responsible for acute or prolonged adverse effects such as diarrhea, abdominal pain, nausea, headaches, and fatigue[84]. In particular, immunological concerns have been raised regarding safety assessments for both probiotics and the FMT because either indigenous or transient microorganisms could impact the immune system's functionality. Hence, the FMT application or the administration of probiotics to specific vulnerable populations and stressed or aged people, immunocompromised patients, newborns or pregnant women must be well evaluated to prevent microbial translocation and sepsis[85-87]. Moreover, the current literature lacks information about the long-term administration of probiotics; therefore, the possible horizontal transfer of antibiotic resistance genes favored by their supplementation cannot be excluded. Likewise, because stool contains thousands of microorganisms and a vast number of metabolites, FMT represents a constant risk of pathogens or commensals transfer to donors that may harmfully affect them [88].

CONCLUSION

As a final note, the different defects found in the evaluated studies highlighted some methodical limitations such as small sample sizes, the limited sampling time and the wide range of other cognitive tests. Supplementation of probiotics and FMT could represent a non-invasive successful strategy to restore gut eubiosis and enhance cognitive functions in healthy people and patients with different neurological/neurodegenerative diseases. Of course, further specific and clinical studies with numerous patients are needed to confirm this encouraging hypothesis.

ARTICLE HIGHLIGHTS

Research background

Due to the global population aging, cognitive impairments will affect approximately



115 million people by 2050. Since current therapies only attempt to counterbalance cognitive disorders, many recent studies recently highlighted the central role of the gut microbiota in brain health.

Research motivation

The pathogenesis of several cognitive disorders is still not fully understood; however, it has been recently established that a dysregulated gut-brain axis communication is associated with the onset and persistence of neurodegenerative diseases. Thus, gut microbiota manipulation could restore a functional gut-brain axis improving cognitive functions.

Research objectives

Since the evidence derived from human randomized clinical trials (RCTs) is currently limited, the main purpose of this systematic review was to detect the currently available RCTs, to define better the effects of probiotics, prebiotics, and fecal microbiota transplant (FMT) on cognitive functions.

Research methods

We systematically searched Embase, Medline/PubMed, Cochrane Library, central and clinicaltrials.gov databases with a combination of comprehensive terms related to cognition and gut microbiota manipulation. Then, we carefully reviewed and synthesized the data by types of study design and setting, characteristics of the studied population, kind of the intervention (strain type or mixture type, dosage and frequency of administration), control treatment, inclusion and exclusion criteria, follow-up duration, and cognitive or memory outcomes.

Research results

The analysis of the 23 included in our systematic review highlighted that, although the different type and posology of administration and the various cognitive tests and questionnaires adopted, both probiotics supplementation and FMT improved the cognitive functions in most of healthy people and patients affected by different neurological pathologies.

Research conclusions

The gut microbiota manipulation could represent a good strategy to counteract gut dysbiosis and so ameliorate cognitive dysfunction.

Research perspectives

The supplementation of probiotics and FMT could represent a non-invasive successful strategy to restore gut eubiosis and enhance cognitive functions in healthy people and patients with different neurological/neurodegenerative diseases.

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REFERENCES

- Land KC, Lamb VL. Demography of Aging. In: Heggenhougen K, Stella Q. International Encyclopedia of Public Health. San Diego: Academic Press, 2008: 89-95
- 2 Brayne C, Miller B. Dementia and aging populations-A global priority for contextualized research and health policy. PLoS Med 2017; 14: e1002275 [PMID: 28350794 DOI: 10.1371/journal.pmed.1002275
- 3 Tomaskova H, Kuhnova J, Cimler R, Dolezal O, Kuca K. Prediction of population with Alzheimer's disease in the European Union using a system dynamics model. *Neuropsychiatr Dis Treat* 2016; 12: 1589-1598 [PMID: 27418826 DOI: 10.2147/NDT.S107969]
- 4 Prince MJ, Wu F, Guo Y, Gutierrez Robledo LM, O'Donnell M, Sullivan R, Yusuf S. The burden of disease in older people and implications for health policy and practice. Lancet 2015; 385: 549-562 [PMID: 25468153 DOI: 10.1016/S0140-6736(14)61347-7]
- Sacuiu SF. Dementias. Handb Clin Neurol 2016; 138: 123-151 [PMID: 27637956 DOI: 5 10.1016/B978-0-12-802973-2.00008-2]
- Snyder HM, Corriveau RA, Craft S, Faber JE, Greenberg SM, Knopman D, Lamb BT, Montine TJ, 6



Nedergaard M, Schaffer CB, Schneider JA, Wellington C, Wilcock DM, Zipfel GJ, Zlokovic B, Bain LJ, Bosetti F, Galis ZS, Koroshetz W, Carrillo MC. Vascular contributions to cognitive impairment and dementia including Alzheimer's disease. Alzheimers Dement 2015; 11: 710-717 [PMID: 25510382 DOI: 10.1016/j.jalz.2014.10.008]

- Raz L, Knoefel J, Bhaskar K. The neuropathology and cerebrovascular mechanisms of dementia. J 7 Cereb Blood Flow Metab 2016; 36: 172-186 [PMID: 26174330 DOI: 10.1038/jcbfm.2015.164]
- 8 Yiannopoulou KG, Papageorgiou SG. Current and future treatments for Alzheimer's disease. Ther Adv Neurol Disord 2013; 6: 19-33 [PMID: 23277790 DOI: 10.1177/1756285612461679]
- Boem F, Amedei A. Healthy axis: Towards an integrated view of the gut-brain health. World J 9 Gastroenterol 2019; 25: 3838-3841 [PMID: 31413521 DOI: 10.3748/wjg.v25.i29.3838]
- Cussotto S, Sandhu KV, Dinan TG, Cryan JF. The Neuroendocrinology of the Microbiota-Gut-Brain 10 Axis: A Behavioural Perspective. Front Neuroendocrinol 2018; 51: 80-101 [PMID: 29753796 DOI: 10.1016/j.yfrne.2018.04.002]
- Cryan JF, Dinan TG. Mind-altering microorganisms: the impact of the gut microbiota on brain and 11 behaviour. Nat Rev Neurosci 2012; 13: 701-712 [PMID: 22968153 DOI: 10.1038/nrn3346]
- 12 Mangiola F, Ianiro G, Franceschi F, Fagiuoli S, Gasbarrini G, Gasbarrini A. Gut microbiota in autism and mood disorders. World J Gastroenterol 2016; 22: 361-368 [PMID: 26755882 DOI: 10.3748/wjg.v22.i1.361
- Noble EE, Hsu TM, Kanoski SE. Gut to Brain Dysbiosis: Mechanisms Linking Western Diet 13 Consumption, the Microbiome, and Cognitive Impairment. Front Behav Neurosci 2017; 11:9 [PMID: 28194099 DOI: 10.3389/fnbeh.2017.00009]
- Ticinesi A, Tana C, Nouvenne A, Prati B, Lauretani F, Meschi T. Gut microbiota, cognitive frailty 14 and dementia in older individuals: a systematic review. Clin Interv Aging 2018; 13: 1497-1511 [PMID: 30214170 DOI: 10.2147/CIA.S139163]
- 15 Bonfili L, Cecarini V, Berardi S, Scarpona S, Suchodolski JS, Nasuti C, Fiorini D, Boarelli MC, Rossi G, Eleuteri AM. Microbiota modulation counteracts Alzheimer's disease progression influencing neuronal proteolysis and gut hormones plasma levels. Sci Rep 2017; 7: 2426 [PMID: 28546539 DOI: 10.1038/s41598-017-02587-2]
- 16 Risk Reduction of Cognitive Decline and Dementia: WHO Guidelines. Geneva: World Health Organization; 2019 [PMID: 31219687]
- 17 Chen D, Yang X, Yang J, Lai G, Yong T, Tang X, Shuai O, Zhou G, Xie Y, Wu Q. Prebiotic Effect of Fructooligosaccharides from Morinda officinalis on Alzheimer's Disease in Rodent Models by Targeting the Microbiota-Gut-Brain Axis. Front Aging Neurosci 2017; 9: 403 [PMID: 29276488 DOI: 10.3389/fnagi.2017.00403]
- 18 Larsen N, Vogensen FK, Gøbel RJ, Michaelsen KF, Forssten SD, Lahtinen SJ, Jakobsen M. Effect of Lactobacillus salivarius Ls-33 on fecal microbiota in obese adolescents. Clin Nutr 2013; 32: 935-940 [PMID: 23510724 DOI: 10.1016/j.clnu.2013.02.007]
- Cammarota G, Ianiro G, Tilg H, Rajilić-Stojanović M, Kump P, Satokari R, Sokol H, Arkkila P, 19 Pintus C, Hart A, Segal J, Aloi M, Masucci L, Molinaro A, Scaldaferri F, Gasbarrini G, Lopez-Sanroman A, Link A, de Groot P, de Vos WM, Högenauer C, Malfertheiner P, Mattila E, Milosavljević T, Nieuwdorp M, Sanguinetti M, Simren M, Gasbarrini A; European FMT Working Group. European consensus conference on faecal microbiota transplantation in clinical practice. Gut 2017; 66: 569-580 [PMID: 28087657 DOI: 10.1136/gutjnl-2016-313017]
- Sun J, Xu J, Ling Y, Wang F, Gong T, Yang C, Ye S, Ye K, Wei D, Song Z, Chen D, Liu J. Fecal 20 microbiota transplantation alleviated Alzheimer's disease-like pathogenesis in APP/PS1 transgenic mice. Transl Psychiatry 2019; 9: 189 [PMID: 31383855 DOI: 10.1038/s41398-019-0525-3]
- Goloshchapov OV, Olekhnovich EI, Sidorenko SV, Moiseev IS, Kucher MA, Fedorov DE, Pavlenko 21 AV, Manolov AI, Gostev VV, Veselovsky VA, Klimina KM, Kostryukova ES, Bakin EA, Shvetcov AN, Gumbatova ED, Klementeva RV, Shcherbakov AA, Gorchakova MV, Egozcue JJ, Pawlowsky-Glahn V, Suvorova MA, Chukhlovin AB, Govorun VM, Ilina EN, Afanasyev BV. Long-term impact of fecal transplantation in healthy volunteers. BMC Microbiol 2019; 19: 312 [PMID: 31888470 DOI: 10.1186/s12866-019-1689-y]
- Neisser U. Cognitive psychology. Englewood Cliffs: Prentice-Hall II, Inc., 1967 22
- 23 Agahi A, Hamidi GA, Daneshvar R, Hamdieh M, Soheili M, Alinaghipour A, Esmaeili Taba SM, Salami M. Does Severity of Alzheimer's Disease Contribute to Its Responsiveness to Modifying Gut Microbiota? Front Neurol 2018; 9: 662 [PMID: 30158897 DOI: 10.3389/fneur.2018.00662]
- Akbari E, Asemi Z, Daneshvar Kakhaki R, Bahmani F, Kouchaki E, Tamtaji OR, Hamidi GA, 24 Salami M. Effect of Probiotic Supplementation on Cognitive Function and Metabolic Status in Alzheimer's Disease: A Randomized, Double-Blind and Controlled Trial. Front Aging Neurosci 2016; 8: 256 [PMID: 27891089 DOI: 10.3389/fnagi.2016.00256]
- 25 Bagga D, Reichert JL, Koschutnig K, Aigner CS, Holzer P, Koskinen K, Moissl-Eichinger C, Schöpf V. Probiotics drive gut microbiome triggering emotional brain signatures. Gut Microbes 2018; 9: 486-496 [PMID: 29723105 DOI: 10.1080/19490976.2018.1460015]
- 26 Bajaj JS, Heuman DM, Hylemon PB, Sanyal AJ, Puri P, Sterling RK, Luketic V, Stravitz RT, Siddiqui MS, Fuchs M, Thacker LR, Wade JB, Daita K, Sistrun S, White MB, Noble NA, Thorpe C, Kakiyama G, Pandak WM, Sikaroodi M, Gillevet PM. Randomised clinical trial: Lactobacillus GG modulates gut microbiome, metabolome and endotoxemia in patients with cirrhosis. Aliment Pharmacol Ther 2014; 39: 1113-1125 [PMID: 24628464 DOI: 10.1111/apt.12695]
- 27 Bajaj JS, Kassam Z, Fagan A, Gavis EA, Liu E, Cox IJ, Kheradman R, Heuman D, Wang J, Gurry T,



Williams R, Sikaroodi M, Fuchs M, Alm E, John B, Thacker LR, Riva A, Smith M, Taylor-Robinson SD, Gillevet PM. Fecal microbiota transplant from a rational stool donor improves hepatic encephalopathy: A randomized clinical trial. Hepatology 2017; 66: 1727-1738 [PMID: 28586116 DOI: 10.1002/hep.29306]

- Bajaj JS, Salzman NH, Acharya C, Sterling RK, White MB, Gavis EA, Fagan A, Hayward M, Holtz 28 ML, Matherly S, Lee H, Osman M, Siddiqui MS, Fuchs M, Puri P, Sikaroodi M, Gillevet PM. Fecal Microbial Transplant Capsules Are Safe in Hepatic Encephalopathy: A Phase 1, Randomized, Placebo-Controlled Trial. Hepatology 2019; 70: 1690-1703 [PMID: 31038755 DOI: 10.1002/hep.30690
- 29 Benton D, Williams C, Brown A. Impact of consuming a milk drink containing a probiotic on mood and cognition. Eur J Clin Nutr 2007; 61: 355-361 [PMID: 17151594 DOI: 10.1038/sj.ejcn.1602546]
- Buigues C, Fernández-Garrido J, Pruimboom L, Hoogland AJ, Navarro-Martínez R, Martínez-30 Martínez M, Verdejo Y, Mascarós MC, Peris C, Cauli O. Effect of a Prebiotic Formulation on Frailty Syndrome: A Randomized, Double-Blind Clinical Trial. Int J Mol Sci 2016; 17 [PMID: 27314331 DOI: 10.3390/ijms17060932]
- Capitão LP, Baião R, Baek HK, Kappelmann N, Sharman R, Harvey CJ, Montgomery P, Burnet PW. 31 Prebiotic supplementation does not affect reading and cognitive performance in children: A randomised placebo-controlled study. J Psychopharmacol 2020; 34: 148-152 [PMID: 31342840 DOI: 10.1177/0269881119862534]
- Ceccarelli G, Brenchley JM, Cavallari EN, Scheri GC, Fratino M, Pinacchio C, Schietroma I, Fard 32 SN, Scagnolari C, Mezzaroma I, Vullo V, d'Ettorre G. Impact of High-Dose Multi-Strain Probiotic Supplementation on Neurocognitive Performance and Central Nervous System Immune Activation of HIV-1 Infected Individuals. Nutrients 2017; 9 [PMID: 29160817 DOI: 10.3390/nu9111269]
- 33 Chung YC, Jin HM, Cui Y, Kim DS, Jung JM, Park JI, Jung ES, Choi EK, Chae SW. Fermented milk of Lactobacillus helveticus IDCC3801 improves cognitive functioning during cognitive fatigue tests in healthy older adults. J Funct Foods 2014; 10: 465-474 [DOI: 10.1016/j.jff.2014.07.007]
- Inoue T, Kobayashi Y, Mori N, Sakagawa M, Xiao JZ, Moritani T, Sakane N, Nagai N. Effect of 34 combined bifidobacteria supplementation and resistance training on cognitive function, body composition and bowel habits of healthy elderly subjects. Benef Microbes 2018; 9: 843-853 [PMID: 30198326 DOI: 10.3920/BM2017.0193]
- 35 Hwang YH, Park S, Paik JW, Chae SW, Kim DH, Jeong DG, Ha E, Kim M, Hong G, Park SH, Jung SJ, Lee SM, Na KH, Kim J, Chung YC. Efficacy and Safety of Lactobacillus Plantarum C29-Fermented Soybean (DW2009) in Individuals with Mild Cognitive Impairment: A 12-Week, Multi-Center, Randomized, Double-Blind, Placebo-Controlled Clinical Trial. Nutrients 2019; 11 [PMID: 30717153 DOI: 10.3390/nu110203051
- 36 Kelly JR, Allen AP, Temko A, Hutch W, Kennedy PJ, Farid N, Murphy E, Boylan G, Bienenstock J, Cryan JF, Clarke G, Dinan TG. Lost in translation? Brain Behav Immun 2017; 61: 50-59 [PMID: 27865949 DOI: 10.1016/j.bbi.2016.11.018]
- 37 Kobayashi Y, Kuhara T, Oki M, Xiao JZ. Effects of *Bifidobacterium breve* A1 on the cognitive function of older adults with memory complaints: a randomised, double-blind, placebo-controlled trial. Benef Microbes 2019; 10: 511-520 [PMID: 31090457 DOI: 10.3920/BM2018.0170]
- Lew LC, Hor YY, Yusoff NAA, Choi SB, Yusoff MSB, Roslan NS, Ahmad A, Mohammad JAM, 38 Abdullah MFIL, Zakaria N, Wahid N, Sun Z, Kwok LY, Zhang H, Liong MT. Probiotic Lactobacillus plantarum P8 alleviated stress and anxiety while enhancing memory and cognition in stressed adults: A randomised, double-blind, placebo-controlled study. Clin Nutr 2019; 38: 2053-2064 [PMID: 30266270 DOI: 10.1016/j.clnu.2018.09.010]
- Ohsawa K, Nakamura F, Uchida N, Mizuno S, Yokogoshi H. Lactobacillus helveticus-fermented 39 milk containing lactononadecapeptide (NIPPLTQTPVVVPPFLQPE) improves cognitive function in healthy middle-aged adults: a randomised, double-blind, placebo-controlled trial. Int J Food Sci Nutr 2018; 69: 369-376 [PMID: 28819993 DOI: 10.1080/09637486.2017.1365824]
- Papalini S, Michels F, Kohn N, Wegman J, van Hemert S, Roelofs K, Arias-Vasquez A, Aarts E. 40 Stress matters: Randomized controlled trial on the effect of probiotics on neurocognition. Neurobiol Stress 2019; 10: 100141 [PMID: 30937347 DOI: 10.1016/j.ynstr.2018.100141]
- 41 Roman P, Estévez AF, Miras A, Sánchez-Labraca N, Cañadas F, Vivas AB, Cardona D. A Pilot Randomized Controlled Trial to Explore Cognitive and Emotional Effects of Probiotics in Fibromyalgia. Sci Rep 2018; 8: 10965 [PMID: 30026567 DOI: 10.1038/s41598-018-29388-5]
- 42 Rudzki L, Ostrowska L, Pawlak D, Małus A, Pawlak K, Waszkiewicz N, Szulc A. Probiotic Lactobacillus Plantarum 299v decreases kynurenine concentration and improves cognitive functions in patients with major depression: A double-blind, randomized, placebo controlled study. Psychoneuroendocrinology 2019; 100: 213-222 [PMID: 30388595 DOI: 10.1016/j.psyneuen.2018.10.010
- 43 Smith AP, Sutherland D, Hewlett P. An Investigation of the Acute Effects of Oligofructose-Enriched Inulin on Subjective Wellbeing, Mood and Cognitive Performance. Nutrients 2015; 7: 8887-8896 [PMID: 26516908 DOI: 10.3390/nu7115441]
- Tamtaji OR, Heidari-Soureshjani R, Mirhosseini N, Kouchaki E, Bahmani F, Aghadavod E, 44 Tajabadi-Ebrahimi M, Asemi Z. Probiotic and selenium co-supplementation, and the effects on clinical, metabolic and genetic status in Alzheimer's disease: A randomized, double-blind, controlled trial. Clin Nutr 2019; 38: 2569-2575 [PMID: 30642737 DOI: 10.1016/j.clnu.2018.11.034]
- Tamtaji OR, Taghizadeh M, Daneshvar Kakhaki R, Kouchaki E, Bahmani F, Borzabadi S, Oryan S, 45



Mafi A, Asemi Z. Clinical and metabolic response to probiotic administration in people with Parkinson's disease: A randomized, double-blind, placebo-controlled trial. Clin Nutr 2019; 38: 1031-1035 [PMID: 29891223 DOI: 10.1016/j.clnu.2018.05.018]

- 46 Lee MT, Jang Y, Chang WY. How do impairments in cognitive functions affect activities of daily living functions in older adults? PLoS One 2019; 14: e0218112 [PMID: 31173607 DOI: 10.1371/journal.pone.0218112
- Bunn F, Burn AM, Goodman C, Rait G, Norton S, Robinson L, Schoeman J, Brayne C. Comorbidity 47 and dementia: a scoping review of the literature. BMC Med 2014; 12: 192 [PMID: 25358236 DOI: 10.1186/s12916-014-0192-4]
- 48 Qiu C, Kivipelto M, von Strauss E. Epidemiology of Alzheimer's disease: occurrence, determinants, and strategies toward intervention. Dialogues Clin Neurosci 2009; 11: 111-128 [PMID: 19585947 DOI: 10.31887/DCNS.2009.11.2/cgiu]
- Vuotto C, Battistini L, Caltagirone C, Borsellino G. Gut Microbiota and Disorders of the Central Nervous System. Neuroscientist 2020; 26: 487-502 [PMID: 32441219 DOI: 10.1177/1073858420918826
- Sarkar A, Lehto SM, Harty S, Dinan TG, Cryan JF, Burnet PWJ. Psychobiotics and the Manipulation 50 of Bacteria-Gut-Brain Signals. Trends Neurosci 2016; 39: 763-781 [PMID: 27793434 DOI: 10.1016/j.tins.2016.09.002
- 51 Cryan JF, O'Riordan KJ, Cowan CSM, Sandhu KV, Bastiaanssen TFS, Boehme M, Codagnone MG, Cussotto S, Fulling C, Golubeva AV, Guzzetta KE, Jaggar M, Long-Smith CM, Lyte JM, Martin JA, Molinero-Perez A, Moloney G, Morelli E, Morillas E, O'Connor R, Cruz-Pereira JS, Peterson VL, Rea K, Ritz NL, Sherwin E, Spichak S, Teichman EM, van de Wouw M, Ventura-Silva AP, Wallace-Fitzsimons SE, Hyland N, Clarke G, Dinan TG. The Microbiota-Gut-Brain Axis. Physiol Rev 2019; 99: 1877-2013 [PMID: 31460832 DOI: 10.1152/physrev.00018.2018]
- Martin CR, Osadchiy V, Kalani A, Mayer EA. The Brain-Gut-Microbiome Axis. Cell Mol 52 Gastroenterol Hepatol 2018; 6: 133-148 [PMID: 30023410 DOI: 10.1016/j.jcmgh.2018.04.003]
- 53 Liu S, Gao J, Zhu M, Liu K, Zhang HL. Gut Microbiota and Dysbiosis in Alzheimer's Disease: Implications for Pathogenesis and Treatment. Mol Neurobiol 2020; 57: 5026-5043 [PMID: 32829453 DOI: 10.1007/s12035-020-02073-3]
- Cerovic M, Forloni G, Balducci C. Neuroinflammation and the Gut Microbiota: Possible Alternative 54 Therapeutic Targets to Counteract Alzheimer's Disease? Front Aging Neurosci 2019; 11: 284 [PMID: 31680937 DOI: 10.3389/fnagi.2019.00284]
- Rao RK, Samak G. Protection and Restitution of Gut Barrier by Probiotics: Nutritional and Clinical 55 Implications. Curr Nutr Food Sci 2013; 9: 99-107 [PMID: 24353483 DOI: 10.2174/1573401311309020004
- Cheng S, Ma X, Geng S, Jiang X, Li Y, Hu L, Li J, Wang Y, Han X. Fecal Microbiota 56 Transplantation Beneficially Regulates Intestinal Mucosal Autophagy and Alleviates Gut Barrier Injury. mSystems 2018; 3 [PMID: 30320222 DOI: 10.1128/mSystems.00137-18]
- Li LL, Wang YT, Zhu LM, Liu ZY, Ye CQ, Qin S. Inulin with different degrees of polymerization 57 protects against diet-induced endotoxemia and inflammation in association with gut microbiota regulation in mice. Sci Rep 2020; 10: 978 [PMID: 31969646 DOI: 10.1038/s41598-020-58048-w]
- 58 Bermudez-Brito M, Plaza-Díaz J, Muñoz-Ouezada S, Gómez-Llorente C, Gil A, Probiotic mechanisms of action. Ann Nutr Metab 2012; 61: 160-174 [PMID: 23037511 DOI: 10.1159/000342079]
- Plaza-Diaz J, Ruiz-Ojeda FJ, Gil-Campos M, Gil A. Mechanisms of Action of Probiotics. Adv Nutr 59 2019; 10: S49-S66 [PMID: 30721959 DOI: 10.1093/advances/nmy063]
- Ma Q, Xing C, Long W, Wang HY, Liu Q, Wang RF. Impact of microbiota on central nervous 60 system and neurological diseases: the gut-brain axis. J Neuroinflammation 2019; 16: 53 [PMID: 30823925 DOI: 10.1186/s12974-019-1434-3]
- Wang Y, Kasper LH. The role of microbiome in central nervous system disorders. Brain Behav 61 Immun 2014; 38: 1-12 [PMID: 24370461 DOI: 10.1016/j.bbi.2013.12.015]
- 62 Bonaz B, Bazin T, Pellissier S. The Vagus Nerve at the Interface of the Microbiota-Gut-Brain Axis. Front Neurosci 2018; 12: 49 [PMID: 29467611 DOI: 10.3389/fnins.2018.00049]
- 63 Kim N, Yun M, Oh YJ, Choi HJ. Mind-altering with the gut: Modulation of the gut-brain axis with probiotics. J Microbiol 2018; 56: 172-182 [PMID: 29492874 DOI: 10.1007/s12275-018-8032-4]
- 64 Qin Y, Wade PA. Crosstalk between the microbiome and epigenome: messages from bugs. J Biochem 2018; 163: 105-112 [PMID: 29161429 DOI: 10.1093/jb/mvx080]
- 65 Bonfili L, Cecarini V, Cuccioloni M, Angeletti M, Berardi S, Scarpona S, Rossi G, Eleuteri AM. SLAB51 Probiotic Formulation Activates SIRT1 Pathway Promoting Antioxidant and Neuroprotective Effects in an AD Mouse Model. Mol Neurobiol 2018; 55: 7987-8000 [PMID: 29492848 DOI: 10.1007/s12035-018-0973-4]
- Agus A, Planchais J, Sokol H. Gut Microbiota Regulation of Tryptophan Metabolism in Health and 66 Disease. Cell Host Microbe 2018; 23: 716-724 [PMID: 29902437 DOI: 10.1016/j.chom.2018.05.003]
- Birner A, Platzer M, Bengesser SA, Dalkner N, Fellendorf FT, Queissner R, Pilz R, Rauch P, Maget 67 A, Hamm C, Herzog-Eberhard S, Mangge H, Fuchs D, Moll N, Zelzer S, Schütze G, Schwarz M, Reininghaus B, Kapfhammer HP, Reininghaus EZ. Increased breakdown of kynurenine towards its neurotoxic branch in bipolar disorder. PLoS One 2017; 12: e0172699 [PMID: 28241062 DOI: 10.1371/journal.pone.0172699]
- 68 Bansal Y, Singh R, Parhar I, Kuhad A, Soga T. Quinolinic Acid and Nuclear Factor Erythroid 2-



Related Factor 2 in Depression: Role in Neuroprogression. Front Pharmacol 2019; 10: 452 [PMID: 31164818 DOI: 10.3389/fphar.2019.00452]

- 69 Wong CB, Tanaka A, Kuhara T, Xiao JZ. Potential Effects of Indole-3-Lactic Acid, a Metabolite of Human Bifidobacteria, on NGF-induced Neurite Outgrowth in PC12 Cells. Microorganisms 2020; 8 [PMID: 32178456 DOI: 10.3390/microorganisms8030398]
- 70 Cuartero MI, Ballesteros I, de la Parra J, Harkin AL, Abautret-Daly A, Sherwin E, Fernández-Salguero P, Corbí AL, Lizasoain I, Moro MA. L-kynurenine/aryl hydrocarbon receptor pathway mediates brain damage after experimental stroke. Circulation 2014; 130: 2040-2051 [PMID: 25359166 DOI: 10.1161/CIRCULATIONAHA.114.011394]
- Rothhammer V, Quintana FJ. The aryl hydrocarbon receptor: an environmental sensor integrating 71 immune responses in health and disease. Nat Rev Immunol 2019; 19: 184-197 [PMID: 30718831 DOI: 10.1038/s41577-019-0125-8]
- Takamura T, Harama D, Fukumoto S, Nakamura Y, Shimokawa N, Ishimaru K, Ikegami S, Makino 72 S, Kitamura M, Nakao A. Lactobacillus bulgaricus OLL1181 activates the aryl hydrocarbon receptor pathway and inhibits colitis. Immunol Cell Biol 2011; 89: 817-822 [PMID: 21321579 DOI: 10.1038/icb.2010.165]
- Lamas B, Hernandez-Galan L, Galipeau HJ, Constante M, Clarizio A, Jury J, Breyner NM, Caminero 73 A, Rueda G, Hayes CL, McCarville JL, Bermudez Brito M, Planchais J, Rolhion N, Murray JA, Langella P, Loonen LMP, Wells JM, Bercik P, Sokol H, Verdu EF. Aryl hydrocarbon receptor ligand production by the gut microbiota is decreased in celiac disease leading to intestinal inflammation. Sci Transl Med 2020; 12 [PMID: 33087499 DOI: 10.1126/scitranslmed.aba0624]
- Liang S, Wang T, Hu X, Luo J, Li W, Wu X, Duan Y, Jin F. Administration of Lactobacillus 74 helveticus NS8 improves behavioral, cognitive, and biochemical aberrations caused by chronic restraint stress. Neuroscience 2015; 310: 561-577 [PMID: 26408987 DOI: 10.1016/j.neuroscience.2015.09.033]
- Gu F, Wu Y, Liu Y, Dou M, Jiang Y, Liang H. Lactobacillus casei improves depression-like behavior 75 in chronic unpredictable mild stress-induced rats by the BDNF-TrkB signal pathway and the intestinal microbiota. Food Funct 2020; 11: 6148-6157 [PMID: 32578646 DOI: 10.1039/d0fo00373e]
- 76 Orlando A, Chimienti G, Lezza AMS, Pesce V, Gigante I, D'Attoma B, Russo F. Lactobacillus Rhamnosus GG Affects the BDNF System in Brain Samples of Wistar Rats with Pepsin-Trypsin-Digested Gliadin (PTG)-Induced Enteropathy. Nutrients 2020; 12 [PMID: 32120967 DOI: 10.3390/nu12030629]
- McLoughlin RF, Berthon BS, Jensen ME, Baines KJ, Wood LG. Short-chain fatty acids, prebiotics, 77 synbiotics, and systemic inflammation: a systematic review and meta-analysis. Am J Clin Nutr 2017; 106: 930-945 [PMID: 28793992 DOI: 10.3945/ajcn.117.156265]
- 78 Silva YP, Bernardi A, Frozza RL. The Role of Short-Chain Fatty Acids From Gut Microbiota in Gut-Brain Communication. Front Endocrinol (Lausanne) 2020; 11: 25 [PMID: 32082260 DOI: 10.3389/fendo.2020.00025]
- Vendrik KEW, Ooijevaar RE, de Jong PRC, Laman JD, van Oosten BW, van Hilten JJ, Ducarmon 79 QR, Keller JJ, Kuijper EJ, Contarino MF. Fecal Microbiota Transplantation in Neurological Disorders. Front Cell Infect Microbiol 2020; 10: 98 [PMID: 32266160 DOI: 10.3389/fcimb.2020.00098]
- D'Amato A, Di Cesare Mannelli L, Lucarini E, Man AL, Le Gall G, Branca JJV, Ghelardini C, 80 Amedei A, Bertelli E, Regoli M, Pacini A, Luciani G, Gallina P, Altera A, Narbad A, Gulisano M, Hoyles L, Vauzour D, Nicoletti C. Faecal microbiota transplant from aged donor mice affects spatial learning and memory via modulating hippocampal synaptic plasticity- and neurotransmission-related proteins in young recipients. Microbiome 2020; 8: 140 [PMID: 33004079 DOI: 10.1186/s40168-020-00914-w]
- Sanders ME, Akkermans LM, Haller D, Hammerman C, Heimbach J, Hörmannsperger G, Huys G, 81 Levy DD, Lutgendorff F, Mack D, Phothirath P, Solano-Aguilar G, Vaughan E. Safety assessment of probiotics for human use. Gut Microbes 2010; 1: 164-185 [PMID: 21327023 DOI: 10.4161/gmic.1.3.12127
- Lerner A, Shoenfeld Y, Matthias T. Probiotics: If It Does Not Help It Does Not Do Any Harm. 82 Really? Microorganisms 2019; 7 [PMID: 30979072 DOI: 10.3390/microorganisms7040104]
- Zawistowska-Rojek A, Tyski S. Are Probiotic Really Safe for Humans? Pol J Microbiol 2018; 67: 83 251-258 [PMID: 30451441 DOI: 10.21307/pjm-2018-044]
- Sbahi H, Di Palma JA. Faecal microbiota transplantation: applications and limitations in treating 84 gastrointestinal disorders. BMJ Open Gastroenterol 2016; 3: e000087 [PMID: 27239328 DOI: 10.1136/bmjgast-2016-000087]
- 85 Gill HS, Rutherfurd KJ, Cross ML. Dietary probiotic supplementation enhances natural killer cell activity in the elderly: an investigation of age-related immunological changes. J Clin Immunol 2001; 21: 264-271 [PMID: 11506196 DOI: 10.1023/a:1010979225018]
- 86 Snydman DR. The safety of probiotics. Clin Infect Dis 2008; 46 Suppl 2: S104-11; discussion S144 [PMID: 18181712 DOI: 10.1086/523331]
- Wardill HR, Secombe KR, Bryant RV, Hazenberg MD, Costello SP. Adjunctive fecal microbiota 87 transplantation in supportive oncology: Emerging indications and considerations in immunocompromised patients. EBioMedicine 2019; 44: 730-740 [PMID: 30940601 DOI: 10.1016/j.ebiom.2019.03.070]
- Merrick B, Allen L, Masirah M Zain N, Forbes B, Shawcross DL, Goldenberg SD. Regulation, risk



and safety of Faecal Microbiota Transplant. Infect Prev Pract 2020; 2: 100069 [PMID: 34316559 DOI: 10.1016/j.infpip.2020.100069]



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LETTER TO THE EDITOR

Impact of COVID-19 pandemic on the neuropsychiatric status of Wilson's disease

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Ferri R declared a personal Participation on a Data Safety Monitoring Board or Advisory Board for Jazz in the past 36 mo. Drs. Lanza G, Godani M, and Raggi A declared no conflict of interest.

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Abstract

We have read with interest the Letter to the Editor by Drs. Zhuang and Zhong, who presented the clinical data of 68 patients with Wilson's disease (WD) who were admitted to the hospital before and during the coronavirus disease 2019 (COVID-19) pandemic, and appreciated their findings on hepatic and some extrahepatic manifestations. Nevertheless, given the strong impact of the pandemic on patients with neurological and psychiatric disorders, we would have expected a worsening of the psychiatric and/or neurological impairments in these patients. In contrast, according to the authors, these manifestations remained, somewhat unexpectedly, unchanged. This finding is in contrast with most of the current literature that highlights not only an increased incidence of mental health disorders in the general population but also an exacerbation of neurological and psychiatric symptoms in patients with chronic diseases, especially in those with pre-existing neuropsychiatric disorders, such as WD. Although the study was mainly focused on the hepatic features of WD patients taking anti-copper treatment, a generic and cumulative definition of neurological and psychiatric manifestations, as in this study, does not allow for further considerations. Future studies during and after the pandemic are necessary to clarify the real impact, either direct or indirect, of the COVID-19 pandemic on the neurological and psychiatric symptoms of WD patients.



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Core Tip: In the interesting letter by Drs. Zhuang and Zhong, the psychiatric and neurological manifestations of 68 patients with Wilson's disease who were treated with anti-copper therapy unexpectedly remained unchanged after the pandemic. Given the impact of the pandemic on patients with neurological and psychiatric disorders, a worsening in the severity or frequency of these manifestations could have been expected. The possible reasons underlying this finding, including the relatively small sample size, the effect of therapy, and the patients' resilience, are discussed.

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TO THE EDITOR

We have read with interest the Letter to the Editor by Zhuang and Zhong[1], who presented the clinical data of 68 patients with Wilson's disease (WD) who were admitted to the hospital before and during the coronavirus disease 2019 (COVID-19) pandemic in Guangzhou (China). Of note, none of the patients had COVID-19. As WD is a rare chronic systemic disease, the impact of the pandemic on the multiorgan clinical status of these patients is still unclear and, therefore, certainly worth investigating. Indeed, we have appreciated the findings that showed a marked shortage of medical resources for the clinical management of patients with WD during the pandemic, as well as the findings that patients who consistently took anti-copper medications showed no significant difference in hepatic and some extrahepatic features, although the incidence of their complications (especially those related to infections) significantly increased. For these reasons, we fully support the authors' recommendations to strictly adhere to the anti-copper therapy and closely monitor these patients to prevent complications[1].

However, given the strong psychological and socio-behavioral impacts of the pandemic on the clinical symptoms of different neuropsychiatric disorders[2,3], a worsening, or at least an increase in the frequency or severity, of psychiatric and/or neurological manifestations in WD patients could have been expected. Conversely, according to the authors, before the COVID-19 pandemic, 50 out of the 68 patients had neurological involvement, and three had psychiatric manifestations, which remained, somewhat unexpectedly, unchanged after the pandemic (49 and 4 out of the 68 patients, respectively)[1]. Although this study was mainly focused on the hepatic features of WD, only a generic and cumulative definition of neurological and psychiatric manifestations was adopted without additional specifications (e.g., type, onset, severity, and duration), which did not allow for any further considerations; indeed, these apparently negative results were not discussed. Moreover, it was not specified how neurological and psychiatric manifestations were evaluated (e.g., were specific scales used?), both at the study entry and at the end of the study. A more detailed stratification, for instance, by type of manifestation (such as cognitive or motor deficits, among the neurological aspects, and anxiety or mood disorders, among the psychiatric conditions), would have likely disclosed additional findings.

In this context, COVID-19, being the major infectious outbreak in the 21st century, has led to an unprecedented global hazard to mental health. A recent systematic review^[4] on the impact of the pandemic on mental health in the general population found significantly higher rates of symptoms of anxiety (6.3%-50.9%), depression (14.6%-48.3%), post-traumatic stress disorder (7.0%-53.8%), psychological distress (34.4%-38.0%), and stress in general (8.1%-81.9%) in several countries worldwide, including China. Although a certain degree of heterogeneity was noted across the studies, the risk factors associated with these manifestations included, among others, a



younger age (≤ 40 years) and comorbid chronic or neuropsychiatric illnesses[4], such as WD.

On the other hand, the negative effect of the COVID-19 outbreak on mental health and health care services has been and will likely continue to be significant because of the unpredictability and uncertainty of the pandemic, the associated lockdowns, physical distancing, and other containment strategies, and the resulting economic breakdown[5]. Reasonably, as also observed in the Letter discussed here[1], the impact of the COVID-19 pandemic on the utilization of health care services, in terms of outpatient visits, hospital admissions, diagnostic exams, and therapeutic interventions, decreased by approximately one-third during the pandemic, with considerable variations and greater reductions among people with less severe illness[6]. Throughout the pandemic and even still, there has also been evidence of increased levels of relapse, in people with pre-existing mental health conditions and even in people with no previous history of a mental health disorder[7]. In particular, patients with pre-existing anxiety, depression, panic, delirium, psychosis, and suicidality appear to be extremely vulnerable[8].

Nevertheless, the matter is still debated, since it has also been observed that some individuals with severe mental illnesses, such as schizophrenia and affective disorders, may not report a worsening of symptoms, thus appearing to be resilient to the negative effects of the pandemic[9]. However, frequent assessments and periodic follow-up are needed to determine whether this resilience will persist as the pandemic progresses or after its end. In this context, in addition to the relatively small sample size, the potential effect of anti-copper therapy in this cohort, and the fact that none of the patients were affected by COVID-19, the patients' resilience might also represent a possible reason that supports the findings reported by Zhuang and Zhong[1]. However, their patients did not seem to be affected by severe psychopathologies, thus making this possibility less likely to justify the authors' conclusions. Moreover, although WD is a rare pathology, it is worth mentioning some methodological issues in the study^[1] that may be appropriate and relevant for discussion. In particular, the relatively small sample size, the fact that not all patients were admitted to the hospital during the pandemic, and the possibility of the patients' resilience raise crucial questions that need to be addressed in the near future. Further multicenter prospective cohort studies, retrospective studies, and case-control studies on inpatients and outpatients with WD, both before and after anti-copper treatment, should be performed.

Less is known about the effects of the COVID-19 pandemic on neurological disorders, although a recent systematic review concluded that patients with preexisting conditions (including those characterized by cognitive impairments or parkinsonism, which may be part of the clinical spectrum of WD) can develop exacerbation of their neurological symptoms, thus encouraging clinicians to be aware of this risk and to focus on its prevention and early management^[10]. In this context, it is known that concomitant infections, especially infections of the respiratory and urinary tracts (such as those reported in the letter[1]), frequently worsen the symptoms and the course of several neurological diseases, including parkinsonism and dementia[11,12]. Therefore, the fact that the increased infection incidence detected in the study^[1] did not induce an even transient worsening of the patients' clinical status remains to be explained or at least briefly commented on.

In conclusion, the study by Zhuang and Zhong^[1] provides a valid clinical basis for the proper management of WD patients during the pandemic, thus representing an advance in this field of clinical and research interest. However, further independent multicenter studies during and after the pandemic are necessary to clarify the real impact, either direct (i.e., infected patients) or indirect (i.e., psychological and sociobehavioral consequences), of the COVID-19 pandemic on the neurological and psychiatric symptoms of WD.

REFERENCES

- Zhuang YP, Zhong HJ. Impact of COVID-19 on the clinical status of patients with Wilson disease. World J Gastroenterol 2021; 27: 4248-4251 [PMID: 34326624 DOI: 10.3748/wjg.v27.i26.4248]
- 2 Fisicaro F, Di Napoli M, Liberto A, Fanella M, Di Stasio F, Pennisi M, Bella R, Lanza G, Mansueto G. Neurological Sequelae in Patients with COVID-19: A Histopathological Perspective. Int J Environ Res Public Health 2021; 18 [PMID: 33546463 DOI: 10.3390/ijerph18041415]
- Pennisi M, Lanza G, Cantone M, Ricceri R, Ferri R, D'Agate CC, Pennisi G, Di Lazzaro V, Bella R. 3 Cortical involvement in celiac disease before and after long-term gluten-free diet: A Transcranial Magnetic Stimulation study. PLoS One 2017; 12: e0177560 [PMID: 28489931 DOI:



10.1371/journal.pone.0177560]

- 4 Xiong J, Lipsitz O, Nasri F, Lui LMW, Gill H, Phan L, Chen-Li D, Iacobucci M, Ho R, Majeed A, McIntyre RS. Impact of COVID-19 pandemic on mental health in the general population: A systematic review. J Affect Disord 2020; 277: 55-64 [PMID: 32799105 DOI: 10.1016/j.jad.2020.08.001]
- Moreno C, Wykes T, Galderisi S, Nordentoft M, Crossley N, Jones N, Cannon M, Correll CU, Byrne 5 L, Carr S, Chen EYH, Gorwood P, Johnson S, Kärkkäinen H, Krystal JH, Lee J, Lieberman J, López-Jaramillo C, Männikkö M, Phillips MR, Uchida H, Vieta E, Vita A, Arango C. How mental health care should change as a consequence of the COVID-19 pandemic. Lancet Psychiatry 2020; 7: 813-824 [PMID: 32682460 DOI: 10.1016/S2215-0366(20)30307-2]
- Moynihan R, Sanders S, Michaleff ZA, Scott AM, Clark J, To EJ, Jones M, Kitchener E, Fox M, 6 Johansson M, Lang E, Duggan A, Scott I, Albarqouni L. Impact of COVID-19 pandemic on utilisation of healthcare services: a systematic review. BMJ Open 2021; 11: e045343 [PMID: 33727273 DOI: 10.1136/bmjopen-2020-045343]
- Gavin B, Lyne J, McNicholas F. Mental health and the COVID-19 pandemic: looking back and 7 moving forward. Ir J Psychol Med 2020; 37: 247-249 [PMID: 33323135 DOI: 10.1017/ipm.2020.128]
- 8 Tsamakis K, Tsiptsios D, Ouranidis A, Mueller C, Schizas D, Terniotis C, Nikolakakis N, Tyros G, Kympouropoulos S, Lazaris A, Spandidos DA, Smyrnis N, Rizos E. COVID-19 and its consequences on mental health (Review). Exp Ther Med 2021; 21: 244 [PMID: 33603852 DOI: 10.3892/etm.2021.9675
- 9 Pinkham AE, Ackerman RA, Depp CA, Harvey PD, Moore RC. A Longitudinal Investigation of the Effects of the COVID-19 Pandemic on the Mental Health of Individuals with Pre-existing Severe Mental Illnesses. Psychiatry Res 2020; 294: 113493 [PMID: 33038789 DOI: 10.1016/j.psychres.2020.113493
- 10 Kubota T, Kuroda N. Exacerbation of neurological symptoms and COVID-19 severity in patients with preexisting neurological disorders and COVID-19: A systematic review. Clin Neurol Neurosurg 2021; 200: 106349 [PMID: 33172719 DOI: 10.1016/j.clineuro.2020.106349]
- 11 Lanza G, Ferri R. The neurophysiology of hyperarousal in restless legs syndrome: Hints for a role of glutamate/GABA. Adv Pharmacol 2019; 84: 101-119 [PMID: 31229167 DOI: 10.1016/bs.apha.2018.12.002]
- 12 Deleidi M, Isacson O. Viral and inflammatory triggers of neurodegenerative diseases. Sci Transl Med 2012; 4: 121ps3 [PMID: 22344685 DOI: 10.1126/scitranslmed.3003492]





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