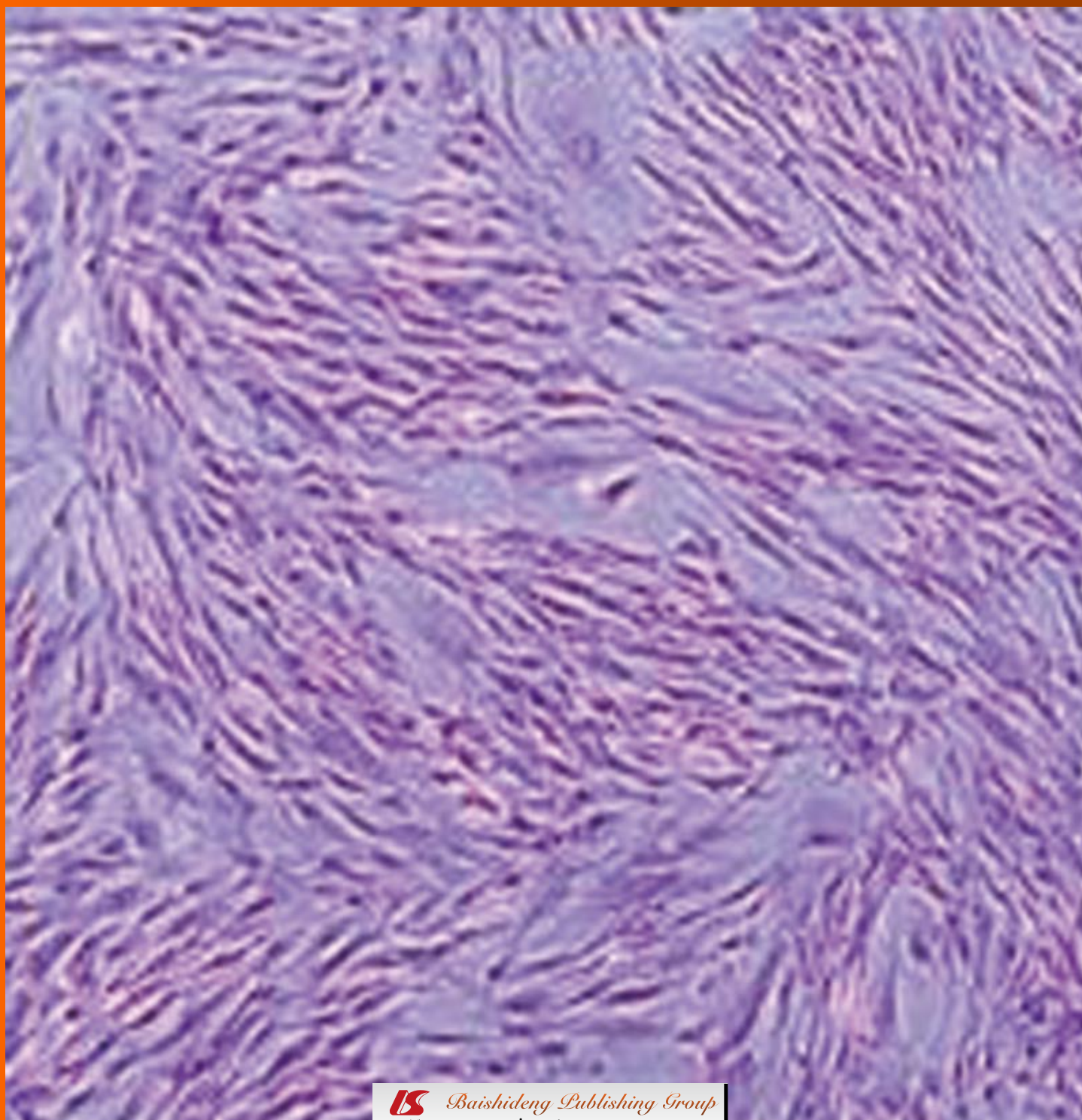


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Characterization of mouse clonal mesenchymal stem cell lines established by subfractionation culturing method

Jeon MS, Yi TG, Lim HJ, Moon SH, Lee MH, Kang JS, Kim CS, Lee DH, Song SU

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Characterization of mouse clonal mesenchymal stem cell lines established by subfractionation culturing method

Myung-Shin Jeon, Tac-Ghee Yi, Hyun-Ja Lim, Sun-Hwa Moon, Moon-Hee Lee, Joon-Soon Kang, Chul-Soo Kim, Dae-Hyun Lee, Sun U Song

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Author contributions: Jeon MS and Yi TG performed the majority of experiments; Lim HJ, Moon SH and Lee DH performed some experiments; Lee MH, Kang JS and Kim CS provided the materials and financial support for this work; Song SU designed the study and wrote the manuscript.

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Abstract

AIM: To characterize single-cell-derived mouse clonal mesenchymal stem cells (mcMSCs) established with bone marrow samples from three different mouse strains.

METHODS: We established mcMSC lines using subfractionation culturing method from bone marrow samples obtained from long bones. These lines were characterized by measuring cell growth, cell surface epitopes, differentiation potential, lineage-specific gene expression and T-cell suppression capability. Nonclonal MSCs isolated by the conventional gradient centrifugation method were used as controls.

RESULTS: All mcMSC lines showed typical nonclonal MSC-like spindle shape morphology. Lines differed in

optimal growth density requirement. Cell surface epitope profiles of these mcMSC lines were similar to those of nonclonal MSCs. However, some lines exhibited different expression levels in a few epitopes, such as CD44 and CD105. Differentiation assays showed that 90% of the mcMSC lines were capable of differentiating into adipogenic and/or chondrogenic lineages, but only 20% showed osteogenic lineage differentiation. T-cell suppression analysis showed that 75% of the lines exhibited T-cell suppression capability.

CONCLUSION: mcMSC lines have similar cell morphology and cell growth rate but exhibit variations in their cell surface epitopes, differentiation potential, lineage-specific gene expression and T-cell suppression capability.

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Key words: Clonal stem cells; Bone marrow stem cells; Multilineage differentiation; Clonal stromal cell isolation; T-cell suppression

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INTRODUCTION

Mesenchymal stem cells (MSCs) were first isolated from the bone marrow by Friedenstein *et al*^[1] in the 1960s and they are now known to be present in a variety of tis-

sues, including adipose tissue, umbilical cord blood and muscle. MSCs can be readily obtained from humans and animals, isolated by means of their ability to adhere to plastic culture plates and can be expanded for many generations in culture.

The culture-expanded MSCs appear to have multi-lineage differentiation potential, including at least adipogenic, chondrogenic and osteogenic differentiation under appropriate induction conditions^[2-4]. Recently, there have been reports regarding additional potential areas for lineage differentiation, such as neurogenic, cardiogenic, myogenic and hepatogenic differentiation^[5-10]. Therefore, these cells are now also called multipotent stem/stromal cells since they are capable of differentiating into mesodermal, ectodermal and/or endodermal origin cells. In addition, more experimental data have become available about the immune modulation and hypoinmunogenic property of MSCs^[11-16] and there has been a surge of interest in both the basic biology and the potential clinical applications of these cells.

The conventional isolation method of MSCs relies on the fractionation of mononuclear cells by gradient centrifugation and selection of fibroblast-like cells adhering to the culture plate surface by removing nonadherent floating cells. Experimental and clinical data over the last decade have indicated that cultured adherent MSCs are heterogeneous and exhibit variations in terms of their differentiation potentials and clinical outcomes^[17,18]. In order to obtain more homogeneous MSC populations, fluorescence-activated cell sorting (FACS) isolation^[19] and specific cell-surface antibody selection^[20,21] have been applied, improving the purity of the final MSC population. However, although significant improvements have been made in purification techniques, these isolation methods have not yet been sufficient to isolate completely homogeneous populations of MSCs. The limiting dilution method has been the only option to generate single-cell-derived clonal MSC lines. Since a mononuclear cell fraction produced by gradient centrifugation or a culture-expanded population of MSCs was the usual source for the limiting dilution method, there is still a possibility of losing some “real” MSCs in the isolation of mononuclear cells and culture expansion of the cells.

Recently, we developed a new protocol, called the sub-fractionation culturing method (SCM), to generate single-cell-derived clonal MSC lines from whole bone marrow aspirate without any centrifugation step for mononuclear cells or enzyme treatment process. Using this method, we identified human bone marrow-derived, colony-forming fibroblastic cells as MSCs from relatively small amounts of bone marrow aspirates. This method allowed us to rapidly establish single-cell-derived human clonal MSC (hcMSC) lines from raw bone marrow aspirates and to establish a library of these hcMSC lines^[22]. The rationale behind the SCM is that different types of mesenchymal stem or stromal cells can be isolated based on their cell densities and/or adherence to a plastic culture plate produced by a series of transfers of the culture supernatant

to subsequent dishes; thus, these low-density stem or stromal cells may have a higher proliferation and differentiation potential.

In the present work, we established single-cell-derived mouse clonal MSC (mcMSC) lines from mouse bone marrow samples from 3 different mouse strains and investigated their cell growth rate, cell surface epitopes, differentiation potentials, lineage-specific gene expression and T-cell suppression capability. The newly established mcMSC lines showed characteristics of progenitors and varied in terms of cell growth rate, cell surface epitopes, differentiation potential, lineage-specific gene expression and T-cell suppression ability, suggesting that variations of progenitor cells exist in mouse bone marrow. These kinds of highly homogeneous, single-cell-derived cMSCs will be very useful for both basic research and clinical applications of such cells.

MATERIALS AND METHODS

Isolation and establishment of mcMSC lines

Mouse bone marrow samples were harvested from tibiae and femurs of 5-wk-old BALB/c, C3H and C57/BL6 mice by flushing out the bone marrow cavity with 5-mL culture medium that comprised Dulbecco's modified Eagle's Medium (DMEM)-low glucose (GIBCO-BRL, Life Technologies, Gaithersburg, MD), 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin. Then 15-mL of the complete growth medium was added to the mixture. This mixture was then incubated in a 100-mm culture dish. As shown in Figure 1, after incubation for 2 h at 37°C with 50 mL/L CO₂, the supernatant only was transferred to a new, intermediate 100-mm dish. After the second 2-h incubation, the supernatant was transferred from the intermediate dish to a new dish (D1) and incubated for another 2 h. The supernatant was then transferred from D1 to a new dish (D2), incubated for 1 d, then transferred from D2 to a new dish (D3) and incubated for 1 d. This process was repeated 2 more times with 1- and 2-d incubation sequentially (for D4 and D5, respectively). After incubation for 7-14 d, the well-separated single-cell-derived colonies that appeared in the culture dishes were detached and isolated after a 2-3 min treatment with trypsin/ethylene diamine tetraacetic acid (EDTA) in cloning cylinders (GIBCO-BRL), transferred to a six-well plate, and then to larger culture flasks where they continued to expand. Once the cells reached approximately 80%-90% confluence, they were recovered with trypsin/EDTA and passaged at 50-100 cells/cm². We obtained ten to twenty colonies on D1 to D3 dishes using bone marrow samples out of five mice and then were able to establish a total of seven, four and six mcMSC lines from each C3H, C57/BL6, Balb/c strain respectively. Out of these mcMSC lines, two lines from each strain were selected for this study.

Cell growth in vitro

For analysis of proliferation ability, the established mc-

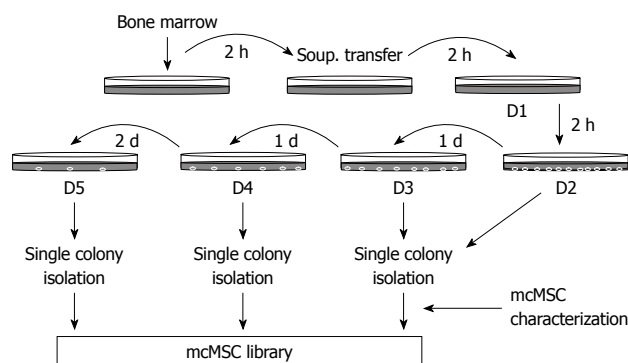


Figure 1 Flow diagram of the “subfractionation culturing method” used to establish mouse clonal mesenchymal stem cell lines. Mouse bone marrow aspirate was mixed with Dulbecco’s modified Eagle’s Medium-low glucose and plated onto a 100-mm cell culture dish. After 2-h incubation, the supernatant only was transferred to a new dish. This procedure was repeated two more times, and then the supernatant was subsequently transferred to cell culture dishes with a 1- or 2-d interval as shown. Each dish was then incubated until single-cell-derived colonies appeared. When colonies of cells were large enough, they were transferred to a six-well plate or 100-mm cell culture dish and then expanded to larger flasks for freezing and further study. Unique mouse clonal mesenchymal stem cell (mcMSC) lines were saved in a mcMSC library.

MSC lines and nonclonal MSCs were seeded at a density of 1×10^3 , 5×10^3 , or 1×10^4 cells/60-mm dish and cultured in complete growth medium for 3, 7, 10 and 14 d. The culture medium was replaced every 3 d. The cell numbers were counted at each time point with a hemocytometer.

Immunostaining and flow cytometry

For phenotyping of cell surface antigens, the established mcMSC lines at 5–7 passages were harvested from the 175-cm² flasks by treatment with trypsin/EDTA and washed twice with phosphate-buffered saline (PBS). The cells were incubated with fluorescein isothiocyanate (FITC)-conjugated antibodies for 30 min at 4°C. The cells were washed twice with PBS consisting of 0.1% bovine serum albumin (BSA). The following antibodies were used for flow cytometry: CD34 (Serotec, Oxford, UK), CD44 (Serotec), CD45 (BD Biosciences Pharmingen, San Diego, CA, USA), CD103 (BD Biosciences Pharmingen), CD105 (R&D Systems, Minneapolis, MN, USA), CD117 (BD Biosciences Pharmingen) and Sca-1 (BD Biosciences Pharmingen). The cells were washed twice with PBS consisting of 0.1% BSA and analyzed using flow cytometry with a 525-nm filter for green FITC fluorescence.

In vitro differentiation

To compare the differentiation potential, the established mcMSC lines and nonclonal MSCs were induced to differentiate into adipogenic, osteogenic or chondrogenic lineage by using the following method.

For chondrogenic differentiation, a pellet culture system was used. Approximately 2×10^5 mcMSCs or nonclonal MSCs were placed in a 15-mL polypropylene tube (Falcon, Bedford, MA, USA) and centrifuged to a pellet. The pellet was cultured at 37°C with 50 mL/L CO₂ in

500 μ L chondrogenic differentiation medium composed of α -MEM supplemented with 10 ng/mL TGF- β 1 (R&D Systems), 10 ng/mL TGF- β 3 (R&D Systems), 50 μ g/mL ascorbic acid (Sigma Chemical Co., St Louis, MO, USA), 10^{-7} mol/L dexamethasone (Sigma Chemical Co.), 40 μ g/mL proline (Sigma Chemical Co.) and 1% ITS⁺ Premix (Becton Dickinson, Bedford, MA, USA; 6.25 μ g/mL insulin, 6.25 μ g/mL transferrin, 6.25 ng/mL selenious acid, 1.25 mg/mL BSA and 5.25 mg/mL linoleic acid). The chondrogenic differentiation medium was changed every 3 d for 3 wk. For microscopy, the pellets were embedded with an OCT compound (Sakura Finetek, Torrance, CA, USA), frozen sectioned into 8- μ m sections and stained with toluidine blue or safranin O.

For investigating osteogenic and adipogenic differentiation, a monolayer culture system was used. For osteogenic differentiation, mcMSCs or nonclonal MSCs were plated at 50 cells/cm² in a 12-well plate and incubated until they reached confluence. The cultures were then incubated in osteogenic differentiation medium composed of α -MEM supplemented with 10% FBS, 50 μ g/mL ascorbic acid (Sigma Chemical Co.), 10^{-8} mol/L dexamethasone (Sigma Chemical Co.), 10 mmol/L β -glycerophosphate (Sigma Chemical Co.), and 1 mmol/L dibutyl- γ -cyclic AMP (db-cAMP). The db-cAMP was added only for the first 4 d. The medium was changed every 3 d for 3 wk. The cells were fixed with 10% formalin (Sigma Chemical Co.) for 10 min at room temperature and stained with 0.1% alizarin red S (Sigma Chemical Co.).

To evaluate adipogenic differentiation, mcMSCs or nonclonal MSCs were plated at 50 cells/cm² in a 12-well plate and incubated in α -MEM (20% FBS) until they reached confluence. The cultures were then incubated in adipogenic differentiation medium composed of DMEM-high glucose supplemented with 10% newborn calf serum, 10^{-7} mol/L dexamethasone (Sigma Chemical Co.), 10 μ g/mL insulin (Sigma Chemical Co.), 0.5 μ mol/L 1-methyl-3-isobutylxanthine (IBMX; Sigma Chemical Co.) and 50 μ g/mL indomethacin (Sigma Chemical Co.). The culture was incubated at 37°C with 50 mL/L CO₂ for 4 d. The cells were fixed with 10% formalin (Sigma Chemical Co.) for 10 min and stained with oil-red-O for 10 min at room temperature.

RNA extraction and reverse transcriptase-polymerase chain reaction analysis

The total RNA was extracted from mcMSC lines grown either in complete growth medium or differentiation medium using easyBLUE[®] (Intron Biotechnology, Sungnam, Korea) reagent. The complementary DNA was synthesized using an AccuPower Reverse-Transcription kit (Bioneer, Daejeon, Korea) by following the manufacturer’s instructions. Polymerase chain reaction (PCR) was carried out using specific primers designed for each gene as follows: type I collagen α 1 chain [ColA1, 587 base pair (bp)], forward primer 5'-ACGTCCTGGT-GAAGTTGGTC-3' and reverse primer 5'-AGCCAC-GATGACCCCTTTATG-3'; type II collagen α 1 chain

(Col2A1, 418 bp), forward primer 5'-CTACACTCAAGTCACTGAACAACCAGAT-3' and reverse primer 5'-CTGTTCTTACAGTGGTAGGTGATGTTCT-3'; type X collagen α 1 chain (Col10A1, 382 bp), forward primer 5'-GTCCAAGAGGTGAACCTGGA-3' and reverse primer 5'-GCACCTACTGCTGGGTAAGC-3'; aggrecan (Acan, 564 bp), forward primer 5'-GCGACATCTGAGTGACTGA-3' and reverse primer 5'-AGACACAGTGGGGAACCTG-3'; lipoprotein lipase (LPL, 421 bp), forward primer 5'-GCGTAGCAGGAAGTC TGACC-3' and reverse primer 5'-CTACAACTCAGGCAGAGCCC-3'; fatty acid-binding protein 4 (FABP4, 319 bp), forward primer 5'-AAAGAAGTGGGAGTGGGCTT-3' and reverse primer 5'-CTCTTGTGGAAGTCACGCCT-3'; peroxisome proliferator-activated receptor- γ 2 (PPAR- γ 2, 377 bp), forward primer 5'-CTTATTTATGATAGGTGTGATCTTAACTGC-3' and reverse primer 5'-GTGATATGTTTGAACCTTGATTTTATCTTCT-3'; alkaline phosphatase (ALP, 555 bp), forward primer 5'-AACCCAGACACAAGCATTC-3' and reverse primer 5'-ACTGGGCCTGGTAGTTGTTG-3'; Distal-less homeobox 5 (Dlx5, 350 bp), forward primer 5'-CAGAAGAGTCCCAAGCATCC-3' and reverse primer 5'-CCACTTCTTCTCTGGCTGG-3'; Runx2 (469 bp), forward primer 5'-TGC GTATTCCTGTAGATCCG-3' and reverse primer 5'-TAGTTCTCATCATTCCTGGC-3'; and glyceraldehyde phosphate dehydrogenase (GAPDH; 350 bp), forward primer 5'-AACTCCCTCAAGATTGTCAGCA-3' and reverse primer 5'-TCCACACCCTGTTGCTTGTA-3'. PCR was performed for 30-40 cycles with each cycle involving denaturation at 94°C for 30 s, annealing at 58-60°C for 30 s and extension at 72°C for 1 min. The amplified PCR products were run on a 1% agarose gel.

T-cell suppression analysis

Mouse splenocytes were cultured in RPMI supplemented with 10% FBS, 10 mmol/L HEPES, 2 mmol/L L-glutamine, 1 mmol/L sodium pyruvate, 50 μ mol/L β -mercaptoethanol, 100 g/mL penicillin and 100 g/mL streptomycin. Splenocytes were isolated from BALB/c spleen. 2×10^5 cells were stimulated with 1 μ g/mL of anti-CD3 and anti-CD28 antibodies (BD Pharmingen, Franklin Lakes, NJ, USA) together with 1×10^4 of indicated mcMSCs for 72 h. Proliferation of T cells was determined by incorporation of [3 H]-thymidine (1 μ Ci/well) for the last 12-16 h of culture. After 48-h stimulation, interferon- γ (IFN- γ) was measured by ELISA according to the protocol of BD Pharmingen.

RESULTS

Establishment of mcMSC lines from mouse bone marrow aspirates

In order to establish mcMSC lines with mouse bone marrow samples from BALB/c, C3H and C57/BL6 strains, as described in the Materials and Methods, approximately 15-mL of each bone marrow mixture with

the complete culture medium mixture were plated in a 100-mm culture dish. Subsequently, the supernatant only was transferred to new culture dishes as described in Figure 1. Representative images of two single-cell-derived colonies from each strain and pictures of mcMSC lines and nonclonal MSCs are shown in Figure 2. The nonclonal MSCs isolated by the gradient centrifugation method (GCM) from the strains were used as control cells. These mcMSC lines established by the SCM are spindle-shaped fibroblast-like cells, which are similar to those of nonclonal MSCs.

Cell proliferation of the established mcMSC lines

The cell growth rates of the established mcMSC lines and nonclonal MSCs were measured with 3 different starting cell densities, 1×10^3 , 5×10^3 , and 1×10^4 per 60-mm plate. As shown in Figure 3, all the lines except C57/BL6 D3-#1 showed exponential growth with starting populations of 5×10^3 and 1×10^4 cells. There was no significant difference between the GCM- or SCM-isolated cells in the proliferation rate for the 16 d of culture. No exponential growth was observed with the starting population of 1×10^3 cells, suggesting that a certain initial number of cells are required for attaining proper growth phase of these cells.

Cell surface epitopes of the established mcMSC lines

The cell surface epitopes of the mcMSC lines and nonclonal MSCs were compared by FACS analysis of a panel of eight cell surface proteins (Table 1). The results showed that all the mcMSC lines and nonclonal MSCs were strongly positive for Sca-1 and negative for CD34, HLA-DR, CD45, CD103 and CD117, as expected. One intriguing finding was that the mcMSC lines showed variations in their expressions of CD44 and CD105, as shown in Table 1. For example, BALB/c D1-#2, C3H D3-#2, C57/BL6 D3-#1 and C57/BL6 D3-#2 showed modest expression of CD44, whereas BALB/s GCM, C3H GCM and C3H D2-#1 were negative. In addition, BALB/c D2-#1 and C57/BL6 D3-#2 were strongly positive and BALB/c GCM, C3H D2-#1 and C3H D3-#2 were modestly positive for the expression of CD105, whereas C3H GCM, C57/BL6 GCM and C57/BL6 D3-#1 were negative. One observation we noticed was that one of the two mcMSC lines, C3H D3-#2, showed both CD44 and CD105 positive exhibited a high level of osteogenic differentiation potential. Other mcMSC lines and nonclonal MSCs showed either CD44 or CD105 positive.

These data demonstrate that the mcMSC lines established by the SCM exhibit variations in the expression of certain cell surface epitopes compared to those of nonclonal MSCs. The significance of these variations remains to be studied.

Adipogenic differentiation potential of the established mcMSC lines

In order to examine the differentiation potential of the

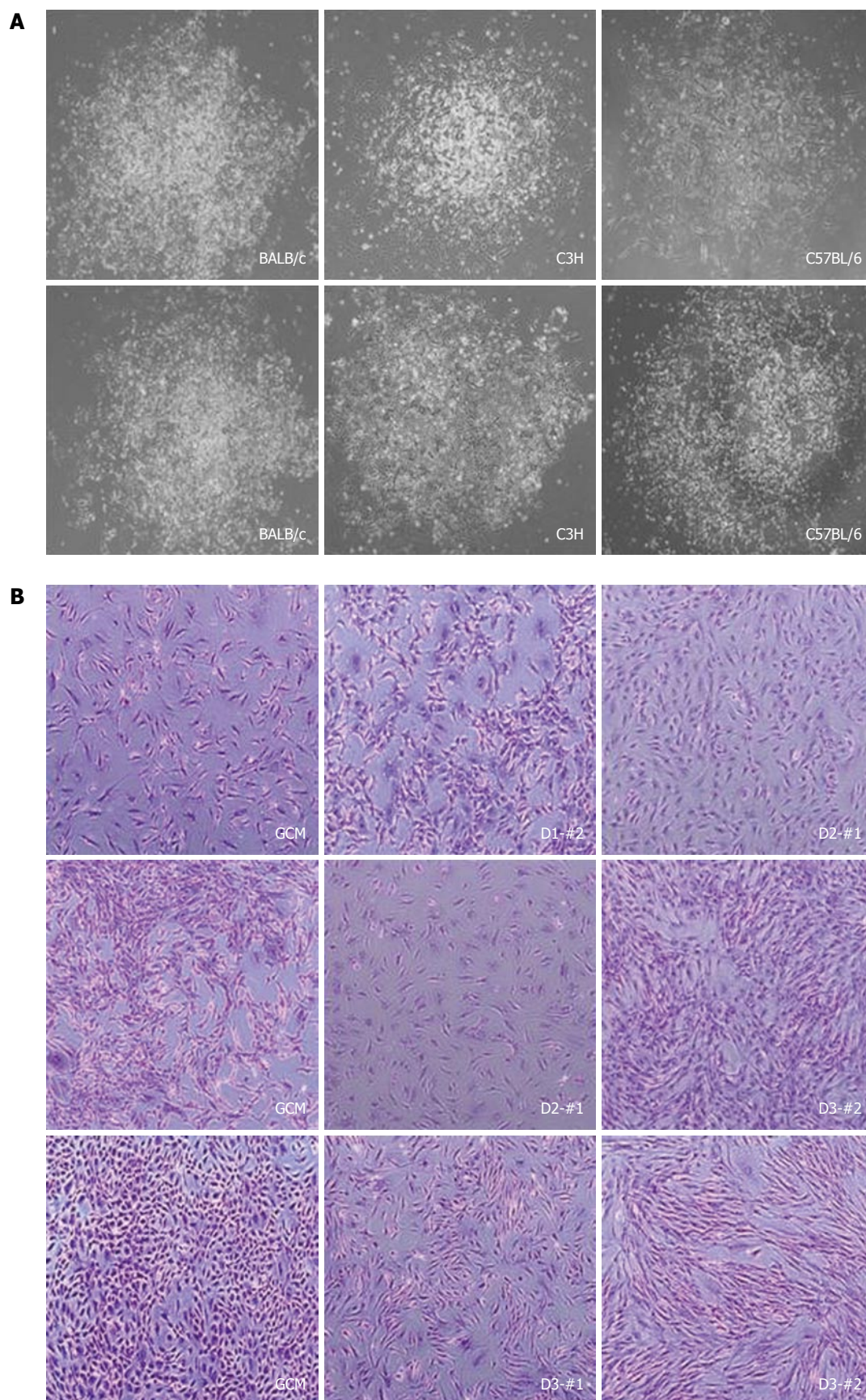


Figure 2 Pictures of the individual colonies and cell morphology of the established mouse clonal mesenchymal stem cell lines. A: Representative pictures of single colonies obtained from different strains; B: Pictures of mouse clonal mesenchymal stem cells (mcMSCs) established from single colonies at about 70%-80% confluence. Nonclonal MSCs isolated by the conventional method, labeled as the gradient centrifugation method, from the three strains are also shown. Every mc-MSCs in the right panel have a fibroblast-like morphology. The cells were fixed with 4% paraformaldehyde for 10 min and stained with 0.1% crystal violet for 5 min. Magnification: 100 \times .

Table 1 Cell surface epitopes of the established mouse clonal mesenchymal stem cell lines and nonclonal mesenchymal stem cells

	Sca-1	CD44	CD105	CD34	HLA-DR	CD45	CD103	CD117
BALB/c GCM	+++	-	+	-	-	-	-	-
BALB/c D1-#2	+++	+	-	-	-	-	-	-
BALB/c D2-#1	+++	-	++	-	-	-	-	-
C3H GCM	+++	-	-	-	-	-	-	-
C3H D2-#1	+++	-	+	-	-	-	-	-
C3H D3-#2	+++	+	+	-	-	-	-	-
C57BL/6 GCM	+++	+	-	-	-	-	-	-
C57BL/6 D3-#1	+++	++	-	-	-	-	-	-
C57BL/6 D3-#2	+++	+	++	-	-	-	-	-

-: Negative; +: 2%-20%; ++: 20%-70%; +++: 70%-100%.

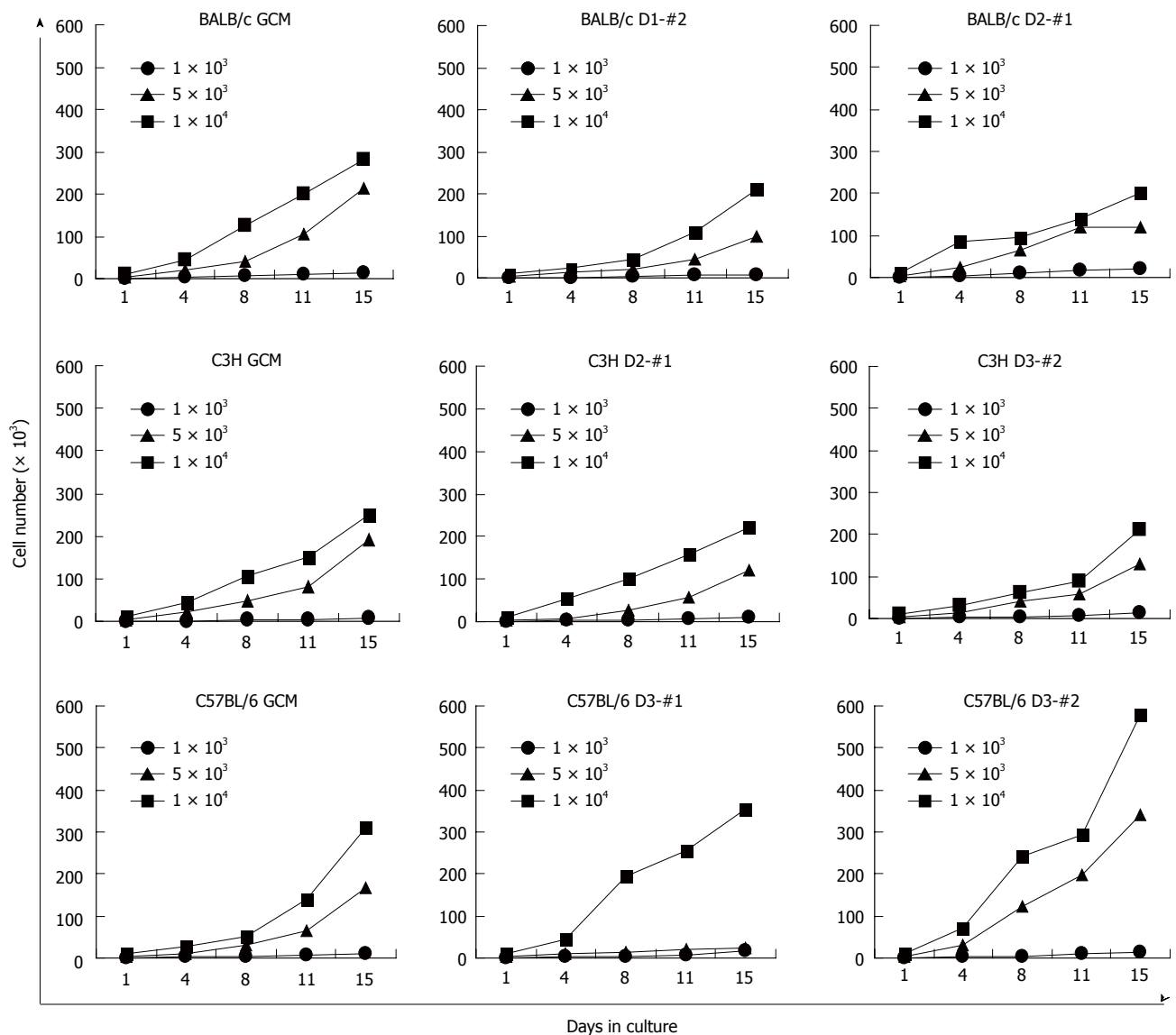


Figure 3 Cell growth of the established mouse clonal mesenchymal stem cell lines and nonclonal mesenchymal stem cells. Cells were plated at 1×10^3 , 5×10^3 , or 1×10^4 cells in a 60-mm dish and incubated in Dulbecco's modified Eagle's Medium-low glucose. Cells were trypsinized by incubation with trypsin/ethylene diamine tetraacetic acid and counted twice a week with a haemocytometer for 15 d. GCM: Gradient centrifugation method.

established mcMSC lines and nonclonal MSCs, *in vitro* adipogenic differentiation analysis was performed using

a monolayer culture system. Passage 6 mcMSCs from each of the cell lines were induced to differentiate into

adipocytes under the conditions described in Materials and Methods. As shown in Figure 4A, every mcMSC line showed a high level of adipogenic differentiation potential except C3H D3-#2. In contrast, the monolayer cells of both nonclonal and mcMSCs grown in the normal control medium showed no oil-red-O staining.

To further confirm the adipogenic differentiation potentials of the established mcMSC lines and nonclonal MSCs, reverse transcriptase-PCR (RT-PCR) assays using total mRNA from the cells were performed before and after induction of differentiation. The lineage-specific expression of adipogenic marker genes, *PPAR-γ2*, *FABP4*, *LPL* and *Col1A1* was measured at 0 and 4 d after induction of differentiation. As shown in Figure 4B, the expression of *PPAR-γ2*, *FABP4* and *LPL* was upregulated in most of the cells, whereas the expression of *Col1A1* was downregulated as expected. These results suggest that both mcMSCs and nonclonal MSCs have a high potential for adipogenic differentiation.

Osteogenic differentiation potential of the established mcMSC lines

To examine the osteogenic differentiation potential of the established mcMSC lines and nonclonal MSCs, *in vitro* differentiation analyses were performed using a monolayer culture system for osteogenic differentiation. Passage 6 mcMSCs from each cell line were induced to differentiate into osteoblasts under the conditions described in Materials and Methods. As shown in Figure 5A, among the twelve tested lines, only one nonclonal MSC line, C57/BL6 GCM, and one mcMSC line, C3H D3-#2, exhibited a high level of osteogenic differentiation potential, whereas other nonclonal and mcMSCs showed no differentiation potential. The monolayer cells of both nonclonal and mcMSCs grown in the normal control medium were negative for Alizarin Red S.

To further confirm the osteogenic differentiation capabilities of the mcMSC lines and nonclonal MSCs, RT-PCR assays were performed using total mRNA from cells before and after induction of differentiation. The lineage-specific expression of osteogenic marker genes, *ALP*, *Runx2*, *Colα1* and *Dlx5* were measured at 0 and 7 d after differentiation induction. As seen in Figure 5B, the expression of *ALP*, *Runx2*, *Colα1* and *Dlx5* was upregulated in C3H D3-#2 and C57/BL6 GCM cells.

Chondrogenic differentiation potential of the established mcMSC lines

To examine the chondrogenic differentiation potential of the established mcMSC lines and nonclonal MSCs, *in vitro* chondrogenic differentiation analysis was performed using a pellet culture system. Passage 6 mcMSCs from each cell line were induced to differentiate into chondrocytes under the conditions described in Materials and Methods. As shown in Figure 6A, every mcMSC line showed a high level of chondrogenic differentiation potential except BALB/c GCM and D2-#1. In contrast, the control monolayer cells grown in the normal medium exhibited only slight safranin O staining.

layer cells grown in the normal medium exhibited only slight safranin O staining.

To further confirm the chondrogenic differentiation capabilities of the mcMSC lines and nonclonal MSCs, RT-PCR assays were performed using total mRNA from cells before and after induction of differentiation. The lineage-specific expression of chondrogenic marker genes, *Col1A1*, *Acan* and *Col10A1* were measured at 0 and 21 d after differentiation induction. As seen in Figure 6B, the expressions of *Col1A1*, *Acan* and *Col10A1* were observed to be upregulated.

T-cell suppression potential of the established mcMSC lines

To test the immunosuppressive potential of mcMSC lines and nonclonal MSCs, mixed lymphocyte reaction analysis was performed with the cells. The formation of T-cell receptor (TCR)-activated T-cell clusters, IFN-γ production suppression and T-cell proliferation inhibition were measured with antibody-activated splenocytes in the presence of each mcMSC line or nonclonal MSCs. As shown in Figure 7A, the number of TCR activated T-cell clusters were reduced in BALB/c D2-#1, C57/BL6 GCM, D3-#1, D3-#2 and C3H D3-#2 cells, whereas other MSCs showed less or no reduction. The results of the IFN-γ production suppression and T-cell proliferation inhibition experiments were highly comparable to the T-cell cluster results except for C57/BL6 GCM cells. These data suggest that the T-cell suppression potential may vary from mcMSC line to mcMSC line.

DISCUSSION

In this paper, single-cell-derived mcMSC lines were established using the SCM and characterized for their proliferation, cell surface epitopes, differentiation potential, lineage-specific gene expression and T-cell suppression capability. The results suggest that mcMSC lines are readily established using the SCM protocol and that some of the newly established mcMSCs have stem/progenitor cell characteristics with variations in their cell surface epitopes, differentiation potential, lineage-specific gene expression and T-cell suppression capability.

In spite of recent high levels of experimental and clinical interest, a standard protocol for producing homogeneous populations of MSCs has not yet been available. The homogeneity of MSCs has become an important issue in terms of understanding the molecular characterization of MSCs and also for applying them in clinical settings. For example, several studies have investigated the gene expression profiles of MSCs to understand the molecular properties of MSCs such as stemness, self-renewal and differentiation potential. Although very informative, most of these studies have limitations due to the heterogeneity of MSCs used in the experiments. In addition, heterogeneous populations of MSCs might possibly generate various efficacies in the treatment of disease. A mixed population of mononuclear cells has been

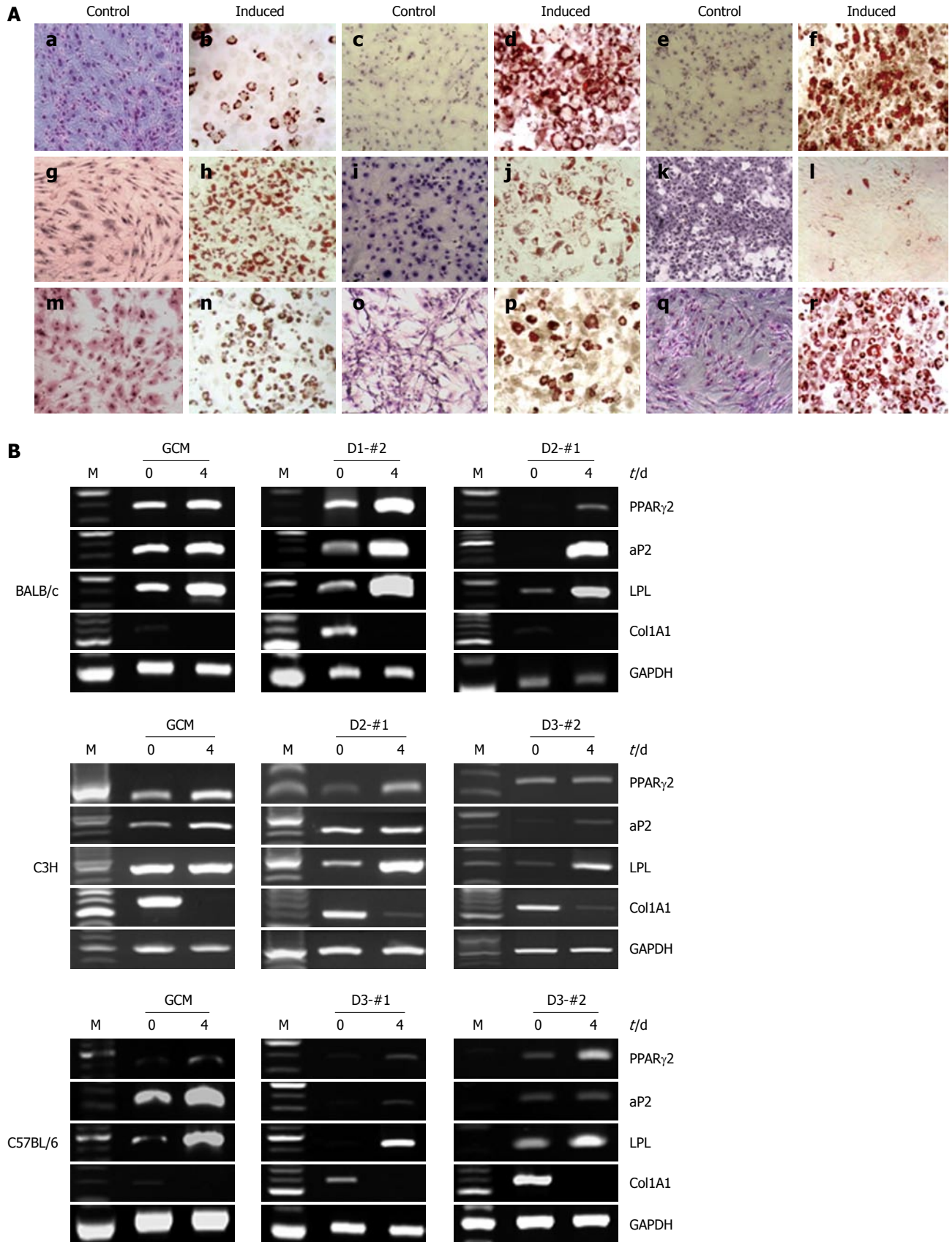


Figure 4 Adipogenic differentiation potential of the established mouse clonal mesenchymal stem cell lines and nonclonal mesenchymal stem cells. A: Oil-red-O staining showed that adipogenically differentiated mouse clonal mesenchymal stem cell lines were positive, tested 4 d after adipogenic induction: a and b: BALB/c gradient centrifugation method (GCM); c and d: BALB/c D1-#2; e and f: BALB/c D2-#1; g and h: C3H GCM; i and j: C3H D2-#1; k and l: C3H D3-#2; m and n: C57BL/6 GCM; o and p: C57BL/6 D3-#1; q and r: C57BL/6 D3-#2; B: Reverse transcriptase-polymerase chain reaction analysis of the adipogenic markers peroxisome proliferator-activated receptor γ 2 (PPAR γ 2), fatty acid-binding protein 4, lipoprotein lipase (LPL), and Col1A1 at days 0 and 4 after adipogenic induction. glyceraldehyde phosphate dehydrogenase was used as an internal control.

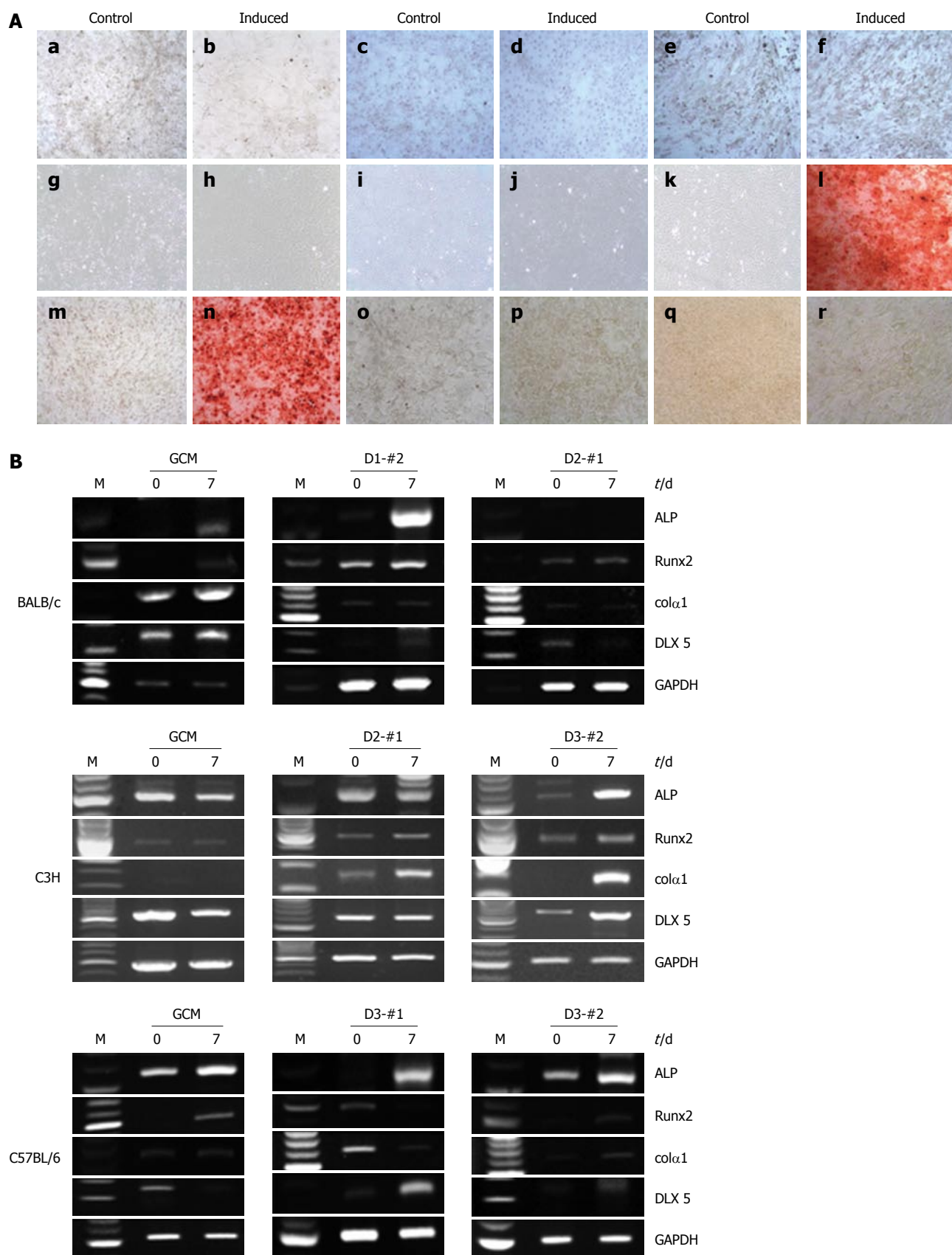


Figure 5 Osteogenic differentiation potentials of the established mouse clonal mesenchymal stem cell lines and nonclonal mesenchymal stem cells. A: Alizarin Red S staining showed matrix mineralization in the osteogenically differentiated mouse clonal mesenchymal stem cell lines 21 d after osteogenic induction: a and b: BALB/c gradient centrifugation method (GCM); c and d: BALB/c D1-#2; e and f: BALB/c D2-#1; g and h: C3H GCM; i and j: C3H D2-#1; k and l: C3H D3-#2; m and n: C57BL/6 GCM; o and p: C57BL/6 D3-#1; q and r: C57BL/6 D3-#2; B: Reverse transcriptase-polymerase chain reaction analysis of osteogenic markers, alkaline phosphatase (ALP), Runx-2, Col1A1, and Distal-less homeobox 5 at days 0 and 7 after osteogenic induction. Glyceraldehyde phosphate dehydrogenase was used as an internal control.

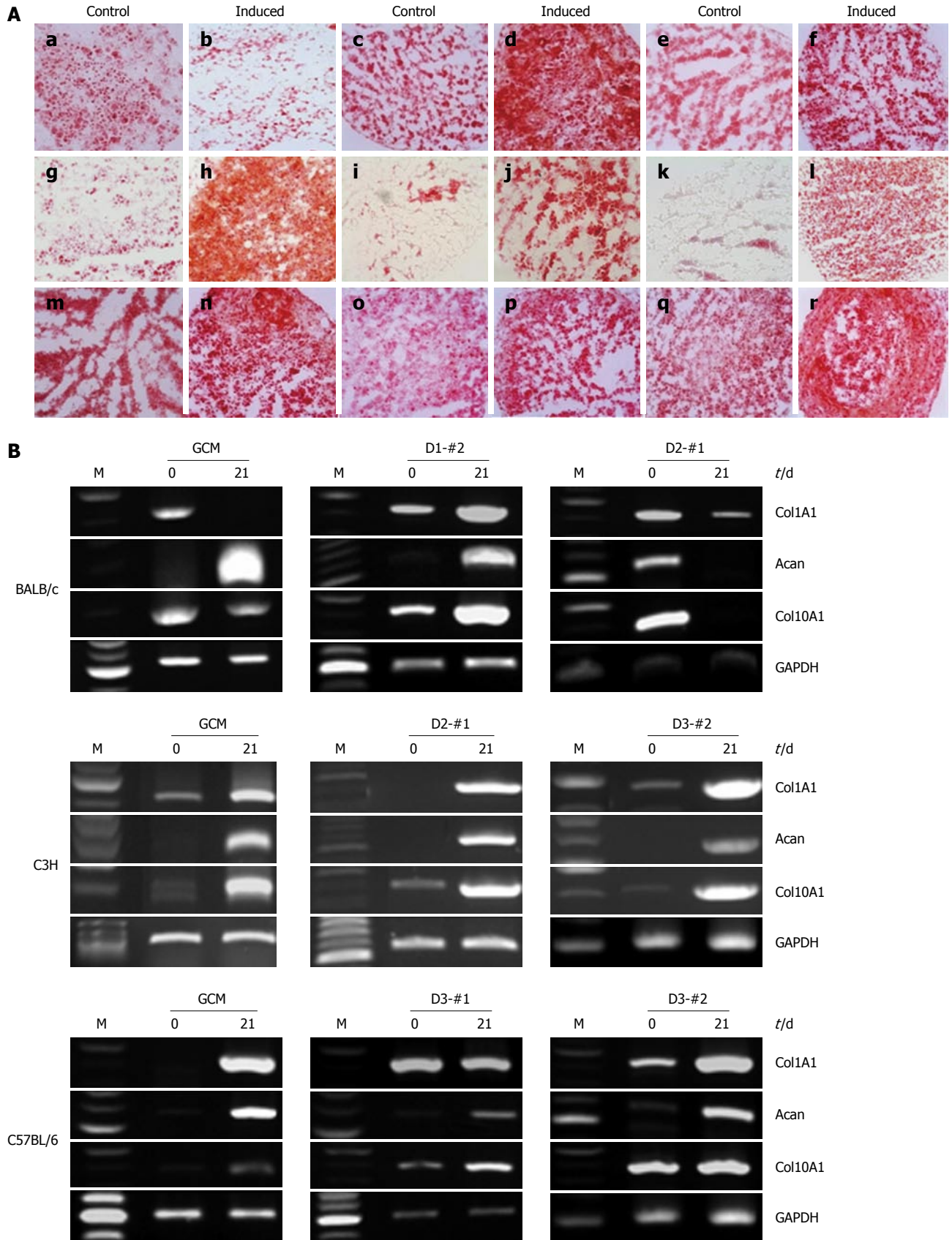


Figure 6 Chondrogenic differentiation potential of the established mouse clonal mesenchymal stem cell lines and nonclonal mesenchymal stem cells. A: Histochemical staining with Safranin-O showed chondrogenically differentiated mouse clonal mesenchymal stem cell lines, tested 21 d after chondrogenic induction: a and b: BALB/c gradient centrifugation method (GCM); c and d: BALB/c D1-#2; e and f: BALB/c D2-#1; g and h: C3H GCM; i and j: C3H D2-#1; k and l: C3H D3-#2; m and n: C57BL/6 GCM; o and p: C57BL/6 D3-#1; q and r: C57BL/6 D3-#2; B: Reverse transcriptase-polymerase chain reaction analysis of chondrogenic markers, type II collagen, aggrecan, and type X collagen at days 0 and 21 after chondrogenic induction. Glyceraldehyde phosphate dehydrogenase was used as an internal control.

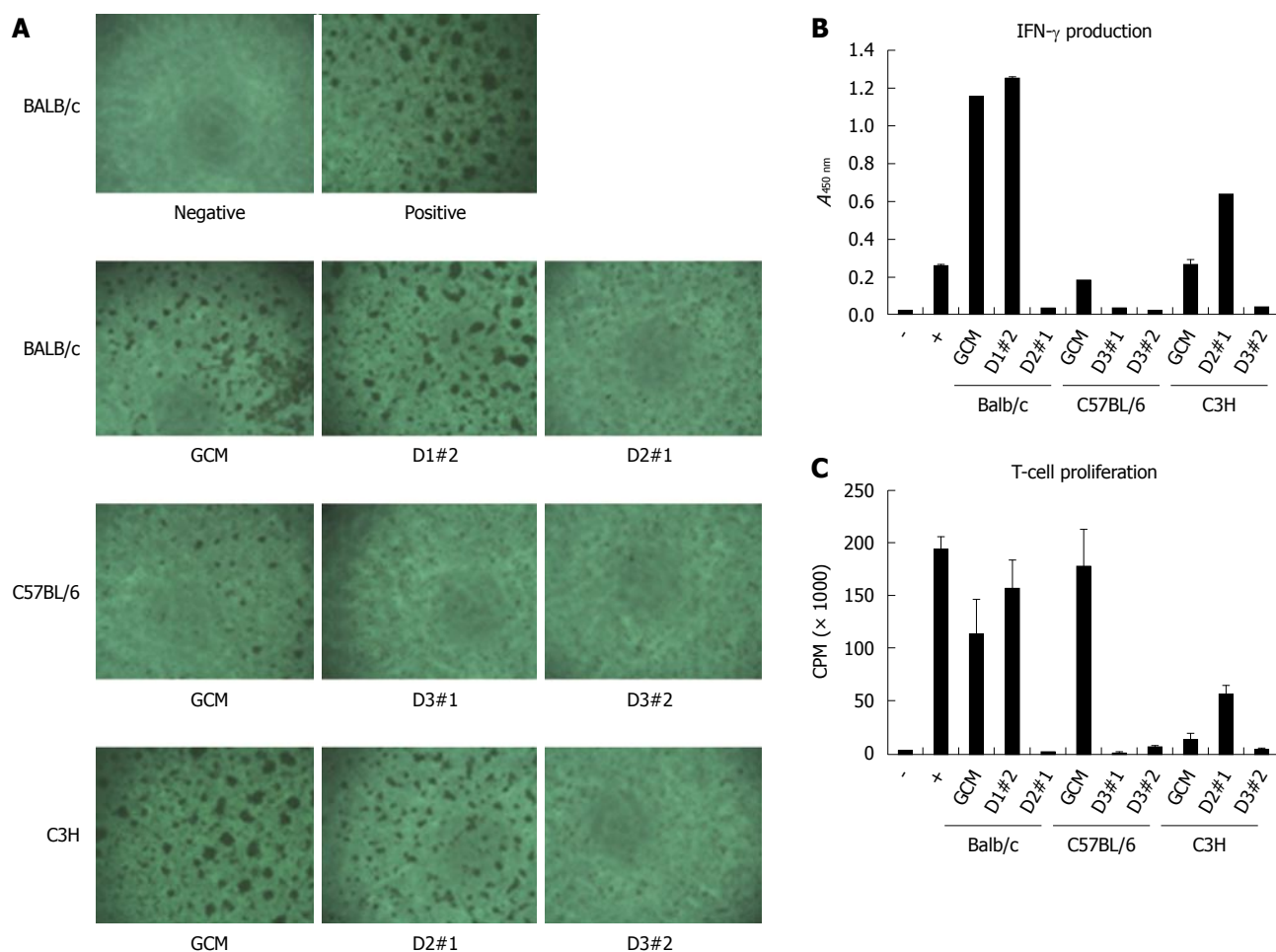


Figure 7 Immunosuppressive effect of mouse clonal mesenchymal stem cell lines or nonclonal mesenchymal stem cells on T cells. Splenocytes (2×10^5) from BALB/c were stimulated with soluble anti-CD3 ($1 \mu\text{g/mL}$) and anti-CD28 ($1 \mu\text{g/mL}$) in the presence of 1×10^4 different mesenchymal stem cells. A: After 48 h, T-cell receptor-activated T-cell clusters were observed by microscopy; B: Interferon (IFN)- γ production was determined by ELISA; C: T-cell proliferation was determined by [^3H]-thymidine incorporation after 3 d culture. GCM: Gradient centrifugation method.

used in most preclinical and clinical studies so far^[18]. The benefit of using a mixed population of MSCs *vs* clonally expanded MSCs from bone marrow for a clinical trial has yet to be adequately investigated. In order to obtain more homogeneous populations of MSCs, antibody selection of specific surface antigen- expressing MSCs^[20,21], FACS sorting of specific sizes of MSCs^[19] and clonal selection by limiting dilution have been utilized. Recently, we developed the SCM and reported how to establish human bone marrow-derived cMSC lines. This method allows a rapid establishment of single-cell-derived clonal MSC lines with a small amount of bone marrow aspirate and could be a very useful protocol with which to obtain highly homogeneous population of MSCs^[22]. The establishment of mcMSC lines from three different mouse strains, as shown in the present study, and our previous success with human bone marrow strongly suggests that the SCM is universally applicable in establishing clonal stem cell lines from relatively small amounts of bone marrow samples regardless of species. We have also succeeded in establishing cMSC lines with rat and rabbit bone marrow aspirates (unpublished data). Therefore, the advantages of the SCM are as follows: Firstly, this

method does not require any centrifugation step in the whole procedure. Secondly, it does not require any enzymatic process or filtering procedure. Thirdly, it can generate single-cell-derived homogeneous clonal MSC lines. Fourthly, the protocol works well for bone marrow samples from different species, including human, mouse, rat and rabbit. These advantages suggest that the SCM can generate a homogeneous population of MSCs in a simple, effective and economic way and may allow safer applications in therapeutic settings.

One difference we have observed while establishing and characterizing mouse and human cMSC lines was the number of lines with osteogenic differentiation potential. The frequency of obtaining cMSC lines with osteogenic differentiation potential was approximately five times lower with mouse bone marrow samples than that with human bone marrow. This difference may be just a simple species difference or might be caused by the source of mouse and human bone marrow; the mouse bone marrow was from long bones whereas the human bone marrow was from the iliac crest. Enzymatic treatment of bone marrow, such as with collagenase, might improve the number of mouse lines with osteogenic differentia-

tion potential. Although there was no correlation between the cell surface marker expression and differentiation potential, one observation we noticed was that one of the two mcMSC lines, C3H D3-#2, showed both CD44 and CD105 positive exhibited a high level of osteogenic differentiation potential. Other mcMSC lines and nonclonal MSCs showed either CD44 or CD105 positive.

The rapid establishment of mcMSC lines and variations of the lines in phenotype, differentiation potential, gene expression and T-cell suppression capacity encouraged us to establish a mcMSC library of a variety of different mcMSC lines. This kind of library will be very useful for both the basic study of bone-marrow-derived stem cells and for clinical study of specific target disease treatment. For example, a cMSC line exhibiting great capability for inhibition of T-cell proliferation would be an excellent candidate stem cell for the development of a stem cell product to treat graft-versus-host disease. One potential concern of using clonal MSC lines is their tumorigenicity potential. Thus, we have tested the transformation potential of seven hcMSC lines by *in vitro*-transformation assay with human HeLa tumor cell line as a positive control and mouse NIH3T3 cell line as a negative control. The results showed that our hcMSC lines produces a smaller number of colonies than do NIH3T3 cells (data not shown), suggesting that the hcMSC lines are probably not tumorigenic.

In conclusion, this study reports a protocol that allows the rapid establishment of mcMSC lines with a small amount of mouse bone marrow. Some of the newly established mcMSC lines exhibit stem/progenitor cell characteristics with some variations in their cell surface epitopes, differentiation potential, lineage-specific gene expression and T cell suppression capability, indicating the presence of different types of stem/progenitor cells in the bone marrow. This protocol will allow easy construction of a mcMSC library containing a variety of different types of bone marrow mcMSC lines. These types of different homogeneous populations of cMSC lines will likely have useful applications in both basic research and clinical applications.

COMMENTS

Background

The conventional isolation method of mesenchymal stem cells (MSCs) relies on the fractionation of mononuclear cells by gradient centrifugation and selection of fibroblast-like cells adhering to the culture plate surface by removing nonadherent floating cells. Experimental and clinical data over the last decade have indicated that cultured adherent MSCs are heterogeneous and exhibit variations in terms of their differentiation potentials and clinical outcomes. In order to obtain more homogeneous MSC populations, fluorescence-activated cell sorting (FACS) isolation and specific cell-surface antibody selection have been applied, improving the purity of the final MSC population. However, although significant improvements have been made in purification techniques, these isolation methods have not yet been sufficient to isolate completely homogeneous populations of MSCs.

Research frontiers

In order to obtain more homogeneous MSC populations, FACS isolation and specific cell-surface antibody selection have been applied, improving the purity of the final MSC population. However, although significant improvements have

been made in purification techniques, these isolation methods have not yet been sufficient to isolate completely homogeneous populations of MSCs.

Innovations and breakthroughs

The advantages of the subfractionation culturing method (SCM) are as follows: Firstly, this method does not require any centrifugation step in the whole procedure. Secondly, it does not require any enzymatic process or filtering procedure. Thirdly, it can generate single-cell-derived homogeneous clonal MSC lines. Fourthly, the protocol works well for bone marrow samples from different species, including human, mouse, rat and rabbit. These advantages suggest that the SCM can generate a homogeneous population of MSCs in a simple, effective and economic way and may allow safer applications in therapeutic settings.

Applications

The results in this study suggest that SCM allows the rapid establishment of mouse clonal MSC (mcMSC) lines with a small amount of mouse bone marrow. This protocol will allow easy construction of a mcMSC library containing a variety of different types of bone marrow mcMSC lines. These types of different homogeneous populations of cMSC lines will likely have useful applications in both basic research and clinical applications.

Terminology

MSC: These stem cells were first isolated from the bone marrow in the 1960s and they are now known to be present in a variety of tissues, including adipose tissue, umbilical cord blood and muscle. They can be differentiated into adipocytes, chondrocytes, osteoblasts, neurons, cardiocytes and hepatocytes. SCM: A newly developed method to isolate clonal MSCs which are derived from a single stem cell so that these cells can be highly homogeneous.

Peer review

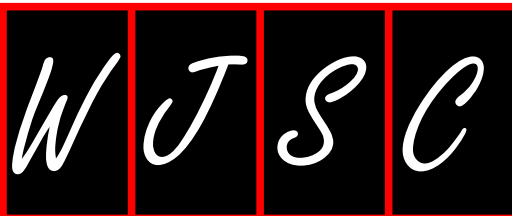
The authors described a simplified protocol to obtain clonal mouse MSCs. The paper is of interest since a lot of data have been collected on the set up of a proper method to isolate MSCs. The paper is well written and the study well conducted.

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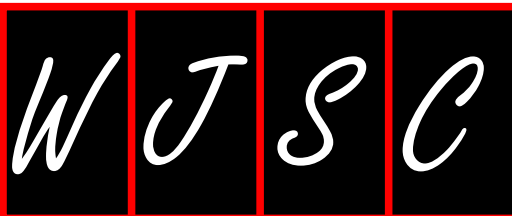
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Events Calendar 2011

March 26, 2011

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Burlingame, CA,
United States

January 29-February 2, 2011
LabAutomation2011,
Palm Springs, CA, United States

February 4, 2011
7th annual Swiss Stem Cell Network
meeting, Swiss Federal Institute
of Technology in Lausanne,
Switzerland

March 1, 2011
The 6th Annual Stem Cell Summit,

11 Fulton Street, New York City, NY,
United States

March 22, 2011
StemCONN 2011, Farmington, CT,
United States

March 27-31, 2011
SBS 17th Annual Conference and
Exhibition, Orlando, FL, United States

April 6-8, 2011
EMBO Conference-Advances in
Stem Cell Research: Development,
Regeneration and Disease,
Institut Pasteur, Paris,
France

April 7-10, 2011
2011 CSHL Meeting on Stem Cell
Engineering & Cell Therapy, Cold

Spring Harbor Laboratory, Cold
Spring Harbor, NY, United States

April 25-26, 2011
International Conference on Stem
Cell Research, Hotel Equatorial
Penang, Malaysia

April 27, 2011
6th Annual Wisconsin Stem Cell
Symposium, BioPharmaceutical
Technology Center, Madison, WI,
United States

May 9-11, 2011
The World Stem Cells and
Regenerative Medicine Congress
2011, Victoria Park Plaza, London,
United Kingdom

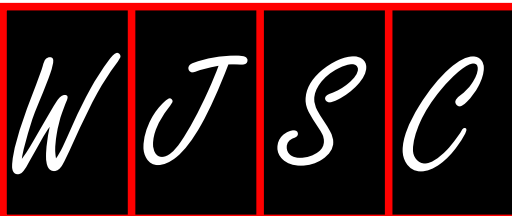
May 23-24, 2011

The 4th Annual Israeli Stem Cell
Meeting, Beit Sourasky,
Chaim Sheba Medical Center,
Israel

May 26-27, 2011
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Therapeutics Conference, Boston,
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September 20-24, 2011
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Cell Biology, Cold Spring
Harbor Laboratory, Cold Spring
Harbor, NY, United States

October 2011
3rd Annual World Stem Cells &
Regenerative Medicine
Congress Asia 2011, Seoul,
South Korea



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- 2 **Lin GZ**, Wang XZ, Wang P, Lin J, Yang FD. Immunologic effect of Jianpi Yishen decoction in treatment of Pixu-diarhoea. *Shijie Huaren Xiaobua Zazhi* 1999; **7**: 285-287

In press

- 3 **Tian D**, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. *Proc Natl Acad Sci USA* 2006; In press

Organization as author

- 4 **Diabetes Prevention Program Research Group**. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; **40**: 679-686 [PMID: 12411462 PMCID:2516377 DOI:10.1161/01.HYP.0000035706.28494.09]

Both personal authors and an organization as author

- 5 **Vallancien G**, Emberton M, Harving N, van Moorselaar RJ; Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; **169**: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

- 6 21st century heart solution may have a sting in the tail. *BMJ* 2002; **325**: 184 [PMID: 12142303 DOI:10.1136/bmj.325.7357.184]

Volume with supplement

- 7 **Geraud G**, Spierings EL, Keywood C. Tolerability and safety of frovatriptan with short- and long-term use for treatment of migraine and in comparison with sumatriptan. *Headache* 2002; **42** Suppl 2: S93-99 [PMID: 12028325 DOI:10.1046/j.1526-4610.42.s2.7.x]

Issue with no volume

- 8 **Banit DM**, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (**401**): 230-238 [PMID: 12151900 DOI:10.1097/00003086-200208000-00026]

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- 9 Outreach: Bringing HIV-positive individuals into care. *HRSA Careaction* 2002; 1-6 [PMID: 12154804]

Books

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- 10 **Sherlock S**, Dooley J. Diseases of the liver and biliary system. 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296

Chapter in a book (list all authors)

- 11 **Lam SK**. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450

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Patent (list all authors)

- 16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1

Statistical data

Write as mean \pm SD or mean \pm SE.

Statistical expression

Express *t* test as *t* (in italics), *F* test as *F* (in italics), chi square test as χ^2 (in Greek), related coefficient as *r* (in italics), degree of freedom as ν (in Greek), sample number as *n* (in italics), and probability as *P* (in italics).

Units

Use SI units. For example: body mass, *m* (B) = 78 kg; blood pressure, *p* (B) = 16.2/12.3 kPa; incubation time, *t* (incubation) = 96 h, blood glucose concentration, *c* (glucose) 6.4 ± 2.1 mmol/L; blood CEA mass concentration, *p* (CEA) = 8.6 ± 24.5 μ g/L; CO₂ volume fraction, 50 mL/L CO₂, not 5% CO₂; likewise for 40 g/L formaldehyde, not 10% formalin; and mass fraction, 8 ng/g, etc. Arabic numerals such as 23, 243, 641 should be read 23 243 641.

The format for how to accurately write common units and quantums can be found at: http://www.wjgnet.com/1948-0210/g_info_20100313172144.htm.

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Standard abbreviations should be defined in the abstract and on first mention in the text. In general, terms should not be abbreviated unless they are used repeatedly and the abbreviation is helpful to the reader. Permissible abbreviations are listed in Units, Symbols and Abbreviations: A Guide for Biological and Medical Editors and

Authors (Ed. Baron DN, 1988) published by The Royal Society of Medicine, London. Certain commonly used abbreviations, such as DNA, RNA, HIV, LD50, PCR, HBV, ECG, WBC, RBC, CT, ESR, CSF, IgG, ELISA, PBS, ATP, EDTA, mAb, can be used directly without further explanation.

Italics

Quantities: *t* time or temperature, *c* concentration, *A* area, *l* length, *m* mass, *V* volume.

Genotypes: *gyrA*, *arg 1*, *c myc*, *c fos*, etc.

Restriction enzymes: *EcoRI*, *HindII*, *BamHI*, *Kbo I*, *Kpn I*, etc.

Biology: *H. pylori*, *E. coli*, etc.

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