World Journal of *Gastrointestinal Oncology*

World J Gastrointest Oncol 2021 December 15; 13(12): 1850-2222





Published by Baishideng Publishing Group Inc

World Journal of Gastrointestinal Oncology

Contents

Monthly Volume 13 Number 12 December 15, 2021

FRONTIER

1850 Management of obstructive colon cancer: Current status, obstacles, and future directions Yoo RN, Cho HM, Kye BH

REVIEW

- 1863 Role of endoscopic ultrasound in anticancer therapy: Current evidence and future perspectives Bratanic A, Bozic D, Mestrovic A, Martinovic D, Kumric M, Ticinovic Kurir T, Bozic J
- 1880 Pancreatic intraductal papillary mucinous neoplasms: Current diagnosis and management Jabłońska B, Szmigiel P, Mrowiec S
- Combined treatments in hepatocellular carcinoma: Time to put them in the guidelines? 1896 Sparchez Z, Radu P, Bartos A, Nenu I, Craciun R, Mocan T, Horhat A, Spârchez M, Dufour JF
- 1919 Unique situation of hepatocellular carcinoma in Egypt: A review of epidemiology and control measures Ezzat R, Eltabbakh M, El Kassas M
- 1939 Moving forward in the treatment of cholangiocarcinoma Manzia TM, Parente A, Lenci I, Sensi B, Milana M, Gazia C, Signorello A, Angelico R, Grassi G, Tisone G, Baiocchi L
- 1956 Solid extraintestinal malignancies in patients with inflammatory bowel disease Mala A, Foteinogiannopoulou K, Koutroubakis IE
- 1981 Mesenchymal stem cell-derived exosomes for gastrointestinal cancer Zhao LX, Zhang K, Shen BB, Li JN
- 1997 Diabetes mellitus contribution to the remodeling of the tumor microenvironment in gastric cancer Rojas A, Lindner C, Schneider I, Gonzàlez I, Araya H, Morales E, Gómez M, Urdaneta N, Araya P, Morales MA
- 2013 Macrophages play a role in inflammatory transformation of colorectal cancer Lu L, Liu YJ, Cheng PQ, Hu D, Xu HC, Ji G

MINIREVIEWS

- 2029 Advancement of chimeric antigen receptor-natural killer cells targeting hepatocellular carcinoma Dai K, Wu Y, She S, Zhang Q
- 2038 Current status of first-line therapy, anti-angiogenic therapy and its combinations of other agents for unresectable hepatocellular carcinoma

Alqahtani SA, Colombo MG



. .	World Journal of Gastrointestinal Onco				
Conter	Monthly Volume 13 Number 12 December 15, 2021				
2050	Endoscopic or percutaneous biliary drainage in hilar cholangiocarcinoma: When and how?				
	Mocan T, Horhat A, Mois E, Graur F, Tefas C, Craciun R, Nenu I, Spârchez M, Sparchez Z				
2064	Current status of non-surgical treatment of locally advanced pancreatic cancer				
	Spiliopoulos S, Zurlo MT, Casella A, Laera L, Surico G, Surgo A, Fiorentino A, de'Angelis N, Calbi R, Memeo R, Inchingolo R				
2076	Prospect of lenvatinib for unresectable hepatocellular carcinoma in the new era of systemic chemotherapy				
	Sho T, Morikawa K, Kubo A, Tokuchi Y, Kitagataya T, Yamada R, Shigesawa T, Kimura M, Nakai M, Suda G, Natsuizaka M, Ogawa K, Sakamoto N				
	ORIGINAL ARTICLE				
	Basic Study				
2088	Dysbiosis of the duodenal microbiota as a diagnostic marker for pancreaticobiliary cancer				
	Sugimoto M, Abe K, Takagi T, Suzuki R, Konno N, Asama H, Sato Y, Irie H, Watanabe K, Nakamura J, Kikuchi H, Takasum M, Hashimoto M, Kato T, Kobashi R, Hikichi T, Ohira H				
2101	MutL homolog 1 methylation and microsatellite instability in sporadic colorectal tumors among Filipinos				
	Cabral LKD, Mapua CA, Natividad FF, Sukowati CHC, Cortez ER, Enriquez MLD				
2114	Propofol induces ferroptosis and inhibits malignant phenotypes of gastric cancer cells by regulating miR 125b-5p/STAT3 axis				
	Liu YP, Qiu ZZ, Li XH, Li EY				
2129	BRAF ^{V600E} mutant colorectal cancer cells mediate local immunosuppressive microenvironment through exosomal long noncoding RNAs				
	Zhi J, Jia XJ, Yan J, Wang HC, Feng B, Xing HY, Jia YT				
	Retrospective Cohort Study				
2149	Hepatocellular carcinoma surveillance and quantile regression for determinants of underutilisation in at- risk Australian patients				
	Low ES, Apostolov R, Wong D, Lin S, Kutaiba N, Grace JA, Sinclair M				
2161	Comparison of tumor regression grading systems for locally advanced gastric adenocarcinoma after neoadjuvant chemotherapy				
	Liu ZN, Wang YK, Zhang L, Jia YN, Fei S, Ying XJ, Zhang Y, Li SX, Sun Y, Li ZY, Ji JF				
	Retrospective Study				
2180	Clinical features of intracerebral hemorrhage in patients with colorectal cancer and its underlying pathogenesis				
	Deng XH, Li J, Chen SJ, Xie YJ, Zhang J, Cen GY, Song YT, Liang ZJ				
	Prospective Study				
2190	Anatomic resection improved the long-term outcome of hepatocellular carcinoma patients with microvascular invasion: A prospective cohort study				
	Zhou JM, Zhou CY, Chen XP, Zhang ZW				



Contents

Monthly Volume 13 Number 12 December 15, 2021

SYSTEMATIC REVIEWS

2203 Minimally invasive surgical treatment of intrahepatic cholangiocarcinoma: A systematic review Patrone R, Izzo F, Palaia R, Granata V, Nasti G, Ottaiano A, Pasta G, Belli A

LETTER TO THE EDITOR

2216 Gender differences in the relationship between alcohol consumption and gastric cancer risk are uncertain and not well-delineated

Verma HK, Bhaskar L

2219 Critical biomarkers of hepatocellular carcinoma in body fluids and gut microbiota Nath LR, Murali M, Nair B



World Journal of Gastrointestinal Oncology

Contents

Monthly Volume 13 Number 12 December 15, 2021

ABOUT COVER

Editorial Board Member of World Journal of Gastrointestinal Oncology, Ladan Teimoori-Toolabi, MD, PhD, Associate Professor, Department of Molecular Medicine, Pasteur Institute of Iran, Tehran 1316943551, Iran. iteimoori@pasteur.ac.ir

AIMS AND SCOPE

The primary aim of World Journal of Gastrointestinal Oncology (WJGO, World J Gastrointest Oncol) is to provide scholars and readers from various fields of gastrointestinal oncology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJGO mainly publishes articles reporting research results and findings obtained in the field of gastrointestinal oncology and covering a wide range of topics including liver cell adenoma, gastric neoplasms, appendiceal neoplasms, biliary tract neoplasms, hepatocellular carcinoma, pancreatic carcinoma, cecal neoplasms, colonic neoplasms, colorectal neoplasms, duodenal neoplasms, esophageal neoplasms, gallbladder neoplasms, etc.

INDEXING/ABSTRACTING

The WJGO is now indexed in Science Citation Index Expanded (also known as SciSearch®), PubMed, PubMed Central, and Scopus. The 2021 edition of Journal Citation Reports® cites the 2020 impact factor (IF) for WJGO as 3.393; IF without journal self cites: 3.333; 5-year IF: 3.519; Journal Citation Indicator: 0.5; Ranking: 163 among 242 journals in oncology; Quartile category: Q3; Ranking: 60 among 92 journals in gastroenterology and hepatology; and Quartile category: Q3. The WJGO's CiteScore for 2020 is 3.3 and Scopus CiteScore rank 2020: Gastroenterology is 70/136.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Ying-Yi Yuan; Production Department Director: Xiang Li; Editorial Office Director: Ya-Juan Ma.

NAME OF JOURNAL World Journal of Gastrointestinal Oncology	INSTRUCTIONS TO AUTHORS https://www.wjgnet.com/bpg/gerinfo/204
ISSN	GUIDELINES FOR ETHICS DOCUMENTS
ISSN 1948-5204 (online)	https://www.wjgnet.com/bpg/GerInfo/287
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
February 15, 2009	https://www.wjgnet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Monthly	https://www.wjgnet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Rosa M Jimenez Rodriguez, Pashtoon M Kasi, Monjur Ahmed, Florin Burada	https://www.wjgnet.com/bpg/gerinfo/208
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
https://www.wjgnet.com/1948-5204/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS
December 15, 2021	https://www.wjgnet.com/bpg/GerInfo/239
COPYRIGHT	ONLINE SUBMISSION
© 2021 Baishideng Publishing Group Inc	https://www.f6publishing.com

© 2021 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



 \mathcal{O} WŰ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 1850-1862

DOI: 10.4251/wjgo.v13.i12.1850

ISSN 1948-5204 (online)

FRONTIER

Management of obstructive colon cancer: Current status, obstacles, and future directions

Ri-Na Yoo, Hyeon-Min Cho, Bong-Hyeon Kye

ORCID number: Ri-Na Yoo 0000-0002-7597-5182; Hyeon-Min Cho 0000-0002-7183-2838; Bong-Hyeon Kye 0000-0002-5251-990X.

Author contributions: Yoo RN, Cho HM and Kye BH were involved equally and have read and approved the final manuscript; Yoo R, Cho HM, and Kye BH met the criteria for authorship established by the International Committee of Medical Journal Editors and verified the validity of the results reported.

Conflict-of-interest statement: The authors declare no conflict of interest

Country/Territory of origin: South Korea

Specialty type: Surgery

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C, C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and Ri-Na Yoo, Hyeon-Min Cho, Bong-Hyeon Kye, Division of Colorectal Surgery, Department of Surgery, St. Vincent's Hospital, The Catholic University of Korea, Suwon 442-723, South Korea

Corresponding author: Bong-Hyeon Kye, MD, PhD, Associate Professor, Surgeon, Surgical Oncologist, Division of Colorectal Surgery, Department of Surgery, St. Vincent's Hospital, The Catholic University of Korea, 222 Banpo-daero, Suwon 442-723, South Korea. ggbong@catholic.ac.kr

Abstract

Approximately 10%-18% of patients with colon cancer present with obstruction at the initial diagnosis. Despite active screening efforts, the incidence of obstructive colon cancer remains stable. Traditionally, emergency surgery has been indicated to treat patients with obstructive colon cancer. However, compared to patients undergoing elective surgery, the morbidity and mortality rates of patients requiring emergency surgery for obstructive colon cancer are high. With the advancement of colonoscopic techniques and equipment, a self-expandable metal stent (SEMS) was introduced to relieve obstructive symptoms, allowing the patient's general condition to be restored and for them undergo elective surgery. As the use of SEMS placement is growing, controversies about its application in potentially curable diseases have been raised. In this review, the short- and longterm outcomes of different treatment strategies, particularly emergency surgery vs SEMS placement followed by elective surgery in resectable, locally advanced obstructive colon cancer, are described based on the location of the obstructive cancer lesion. Controversies regarding each treatment strategy are discussed. To overcome current obstacles, a potential diagnostic method using circulating tumor DNA and further research directions incorporating neoadjuvant chemotherapy are introduced.

Key Words: Colonic neoplasms; Self-expandable metallic stents; Intestinal obstruction; Survival rate; Morbidity; Mortality

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: The optimal management of obstructive colon cancer remains intricate and



fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: March 20, 2021 Peer-review started: March 20, 2021 First decision: July 3, 2021 Revised: July 7, 2021 Accepted: September 22, 2021 Article in press: September 22, 2021 Published online: December 15, 2021

P-Reviewer: Casella C, Pravisani R S-Editor: Ma YJ L-Editor: A P-Editor: Liu JH



controversial, particularly now that the use of colonoscopic stents is expanding. Several studies have demonstrated the better short-term outcome achieved by delayed surgery after self-expandable metal stent (SEMS) placement than emergency surgery. However, concerns that SEMS placement diminishes oncologic outcomes have been increasing. This article summarizes the impact of SEMS placement in managing obstructive colon cancer and suggests future research to resolve current problems.

Citation: Yoo RN, Cho HM, Kye BH. Management of obstructive colon cancer: Current status, obstacles, and future directions. World J Gastrointest Oncol 2021; 13(12): 1850-1862 URL: https://www.wjgnet.com/1948-5204/full/v13/i12/1850.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.1850

INTRODUCTION

Obstructive colon cancer requiring emergency surgical or procedural intervention accounts for 10%-18% of patients initially diagnosed with colon cancer[1-3]. With obstructive symptoms of nausea, vomiting, abdominal pain, and severe distension, patients often present with physical deterioration resulting from the catabolic state of muscle wasting and poor oral intake. Occasionally, patients with obstructive colon cancer have concomitant spontaneous colon perforation, a frightening complication associated with high postoperative morbidity and mortality, at either the cancer site or proximal colonic segment^[4]. Moreover, the obstructive feature of colon cancer itself is an independent high-risk feature of recurrence, and patients with obstructive colon cancer tend to have an advanced cancer stage with poor prognostic factors[5]. Compared to patients undergoing elective surgery, patients requiring emergency surgery for obstructive colon cancer have worse short-term and long-term oncologic outcomes[1,2,5].

Unfortunately, despite active screening efforts, the incidence of obstructive colon cancer remains stable [3,6]. The management of obstructive colon cancer is complex and challenging due to multiple factors, including the patient's index presentation of a poor general condition, limited information regarding the cancer stage, and the need for immediate intervention in an emergency setting. This article addresses the current treatment optionsr, difficulties, and future directions for managing obstructive colon cancer to achieve an optimal outcome.

LOCATION OF OBSTRUCTIVE CANCER LESION

It is well known that the most common cause of colonic obstruction in adults is colorectal cancer^[7]. Abdominopelvic computerized tomography scans are an excellent diagnostic modality that is readily available and accurately localizes obstructive cancer lesions with high sensitivity and specificity[8]. Obstructive colon cancer is usually classified as right-sided or left-sided according to proximal or distal to the splenic flexure. Previous studies indicate that more than half of acute obstructive colon cancer occurs on the left side, most commonly in the sigmoid colon[1,6,8]. The anatomic characteristic of a narrow luminal diameter in the left-sided colon explains the higher incidence of obstructive colon cancer compared to the right-sided colon. on the other hand, a large diameter of the cecum and ascending colon allows a bulky characteristic of the tumor. Indeed, right-sided obstructive colon cancer seems more locally advanced than left-sided[3]. An obstructive cancer lesion in the rectum is the least frequent due to the sizeable luminal diameter of the rectum and the symptoms of bleeding and defecation difficulty caused by rectal cancer itself[3].

The anatomic characteristics of the colon and rectum have led to different treatment approaches depending on the obstructive lesion location. Primary tumor resection with ileocolic anastomosis has been preferred to treat right-sided cancer obstruction [9]. Generally, oncologic resection of the right-sided colon is considered less arduous. Additionally, the relatively low anastomotic complication rate, from 2.8% to 4.6%, supports primary resection and anastomosis[10]. Double-barrel entero-colostomy after primary tumor resection is an alternative for patients with a high risk of anastomotic leakage. Compared to right-sided obstructive colon cancer, left-sided colon cancer has



variable treatment options, primarily comprised of emergency primary tumor resection with or without stoma formation or decompression followed by delayed tumor resection. The choice is usually made based on the patient's general condition, availability of resources, and complete oncologic resection feasibility. In obstructive rectal cancer, optimal oncologic resection with total mesorectal excision is unobtainable. Rectal cancer that causes acute obstruction is usually locally far advanced and highly likely to invade adjacent urogenital organs, large neurovascular structures, and even bony structures. Therefore, for obstructive rectal cancer, decompression to relieve acute symptoms is more desirable than primary tumor resection to avoid serious intraoperative morbidity and suboptimal surgical outcomes. This article discusses the details regarding treatment options and the outcomes of obstructive colon cancer.

TREATMENTS AND THEIR OUTCOMES

Right-sided cancer obstruction

Traditionally, primary oncologic resection via right hemicolectomy or extended right hemicolectomy with ileocolic anastomosis has been advocated for right-sided obstructive colon cancer^[11]. A systematic review of studies related to treating rightsided obstructive colon cancer demonstrated that 86% of patients underwent emergency resection^[12]. Less strenuous surgical techniques for mobilization and resection of the right-sided colon lead many surgeons to prefer primary tumor resection in an emergency setting. Compared to colocolic or colorectal anastomosis, many surgeons pursue primary ileocolic anastomosis, even in frail patients, due to the abundant blood supply and relatively simple manipulation of the dilated proximal bowel with enough length. A recently published multicentered retrospective study on the outcomes of elderly patients treated for obstructive colon cancer revealed that 97% of patients received upfront surgery for proximal colon cancer obstruction[13]. In the study, 54% of patients were over 75 years of age. The rate of resection with primary anastomosis among the elderly patients was not different from that of the younger patients.

Not surprisingly, the short-term outcome after emergency right hemicolectomy with ileocolic anastomosis for obstructive right-sided cancer is worse than that after elective surgery for right-sided colon cancer. Postoperative morbidity after emergency surgery is reported to range from 46% to 54% [13,14]. Compared to the morbidity rate of 30% after elective right hemicolectomy for colon cancer[15], the postoperative morbidity rate after emergency surgery is much higher. The rate of anastomotic leakage after emergency surgery is reported to range from 12% to 16.4% [13,14], which is also higher than the leakage rate of 4.1% after elective surgery [16]. As expected, the postoperative mortality rate after emergency surgery is 14.5%, higher than the rate of 2.6% after elective surgery. Risk factor analysis for anastomotic leakage supports the notion that emergency surgery imposes a greater risk of anastomotic leakage and leakage-related mortality^[17]. Such short-term outcomes indicate that oncologic resection with primary anastomosis for right-sided obstructive colon cancer in an emergency setting may not be as uncomplicated as anticipated.

Other surgical treatment options in an emergency setting may include the creation of loop ileostomy after resection and anastomosis, the creation of end ileostomy after resection only, the creation of loop ileostomy without resection, or enterocolic bypass surgery. For resectable right-sided obstructive colon cancer, primary tumor resection may be an optimal treatment option. In cases of unstable hemodynamics or severe intraperitoneal contamination by bowel perforation, loop ileostomy or end ileostomy is unavoidable. However, high output from ileostomy and related morbidities, such as dehydration, electrolyte imbalance, and acute renal failure, frequently occurs. Distressingly, surgeons encounter the dilemma of risking anastomotic leakage or stoma-related morbidity. For unresectable right-sided obstructive colon cancer, the creation of loop ileostomy or enterocolic bypass surgery is recommended to decompress proximal bowel dilatation, alleviating bowel obstruction symptoms[11]. Relevant studies on the treatment options for unresectable right-sided obstructive colon cancer are scarce and limited to the formation of percutaneous cecostomy, which is currently not performed due to malfunctions and complications or is reserved only for a small number of patients with very high morbidity[11,18].

Staged operation after endoscopic placement of the SEMS may offer a possible treatment option for both resectable and unresectable right-sided obstructive colon cancer. However, the technical difficulty seems to hamper the wide application of



colonoscopic stenting for right-sided colonic obstruction. Additionally, due to a lack of evidence, the use of the SEMS as a bridge to elective surgery is currently not recommended, except for patients with high morbidity[11]. Nevertheless, a newly updated systematic review comparing the treatment outcomes of staged operations after SEMS placement in curable right-sided obstructive colon cancer demonstrated that the technical success rate reached 96%, disputing the previous belief of technical difficulty[12]. Furthermore, the postoperative complication rate is approximately 30%, ranging from 7% to 44% [12,14], similar to the morbidity rate after elective surgery. After the staged operation, the rate of anastomotic leakage is estimated to be approximately 5.5% [12], comparable with the rate observed after elective surgery. The mortality rate of 1.2% after the staged operation is comparable to elective surgery [12]. The long-term oncologic outcome of right-sided obstructive colon cancer appears to be equivalent or better in patients who underwent the staged operation after SEMS insertion than in patients who received emergency surgery^[19].

Current evidence provides safety and feasibility of the staged operation after SEMS insertion to treat right-sided obstructive colon cancer. However, the studies were mostly designed retrospectively with small sample sizes. Patients with unstable hemodynamics would have been excluded from the staged operation, leading to selection bias. The short-term outcome might have been related to the staged operation. On the other hand, such evidence indicates that the staged operation after SEMS placement can achieve a better short-term outcome in a certain group of patients. The selection criteria of an appropriate patient for the staged operation after SEMS placement can be suggested. Because SEMS insertion is impossible if an obstructing cancer is located in the cecum or ileocolic valve, SEMS insertion could be considered when the obstructive cancer is located beyond the cecum in the abdominopelvic computed tomography (CT) scan. Additionally, experts in colonoscopy seem to handle a long colonic length of the distal segment. Therefore, if an expert in colonoscopy is available and there is no sign of perforation on CT imaging, colonoscopic SEMS insertion may be an option before undergoing emergency surgery. After all, optimal treatment for right-sided obstructive colon cancer should be determined based on the resectability of the tumor, presence of perforation and peritonitis, and hemodynamic stability of the patient. Large-sized prospective comparative studies are necessary to obtain concrete evidence.

Left-sided cancer obstruction

Compared to those for right-sided obstructive colon cancer, treatment options for leftsided obstructive colon cancer are diverse and controversial. Conventionally, primary tumor resection with end stoma formation has been highly preferred to treat obstructive left-sided colon cancer in an emergency setting[20,21]. However, emergency surgery itself has been identified as an independent risk factor for mortality^[22]. The postoperative complication rate is higher than the rate after elective surgery^[8,23]. A significant number of patients end up with temporary or permanent stoma after emergency surgery. Furthermore, subsequent surgery for stoma reversal is associated with a high morbidity rate of 21% to 36% [24,25]. Up to 71% of patients never undergo surgery for stoma reversal, significantly affecting the quality of life[25, 26]. The risk factors related to nonreversal of the stoma include advanced age, a postoperative complication that occurred after emergency surgery, comorbidity, and advanced cancer stage[24,26]. Although the operative approach with primary tumor resection with end stoma is considered the safest option due to the absence of anastomotic complications^[20], the two-stage operation is complex and may significantly reduce patient quality of life.

Diverting stoma is another operative procedure for damage control in acute leftsided colonic obstruction. The formation of a diverting stoma is followed by the second-stage operation of primary tumor resection with or without colostomy closure. Stoma closure can take place at a third stage. A diverting stoma may stabilize the patient's general condition and allow bowel preparation and proper staging before oncologic resection. However, patients for whom a three-stage procedure is planned may not undergo subsequent operations, even if they are considered fit. When the morbidity and mortality associated with each surgical stage are considered cumulatively, the staged operation does not provide any advantage. Furthermore, a randomized controlled trial comparing emergency colostomy followed by a staged operation to emergency Hartmann's procedure showed similar morbidity and mortality rates between two procedures; thus, the authors did not support the use of colostomy in frail patients[27]. One advantage of the staged operation is a lower rate of permanent stoma. However, patients who undergo staged operations require a more extended hospital stay for additional surgical procedures following initial colostomy



formation. Nevertheless, due to a lack of evidence, a Cochrane systematic review in 2004 could not conclude which of the two alternative approaches was the most beneficial in acute left-sided obstructive colon cancer^[28].

In an emergency setting, the reconstruction of bowel continuity has been avoided in the management of acute left-sided colonic obstruction due to the risk of anastomotic leakage. However, a two-stage operation followed by Hartmann's operation has the significant disadvantage of cumulative morbidity and mortality associated with a second operation and reduced quality of life when the stoma is kept. Nonetheless, some colorectal surgeons attempted to create a primary anastomosis after emergent primary resection and demonstrated its feasibility and safety in selected patients^{[29,} 30]. In a systematic review, Breitenstein *et al*[31] evaluated the superiority of the onestage procedure compared to multistage procedures for left-sided obstructive colon cancer. The authors demonstrated that primary resection and anastomosis were superior to two- or three-stage operations in terms of mortality, with a relative risk difference from -2% to -27% [31]. It appeared that selection bias strongly affected the study result, in which patients with better prognoses were more likely to have a onestage operation. Previous studies extensively investigated risk factors related to mortality and anastomotic leakage and identified high-risk conditions, such as advanced age, presence of comorbidity, advanced tumor stage, malnutrition, and presence of peritoneal contamination[22,32]. Currently, primary resection and anastomosis are the preferred options for uncomplicated left-sided obstructive colon cancer without risk factors for anastomotic leakage[11].

There are two main methods of resection, and the optimal procedure type is still debated. Segmental colonic resection with intraoperative colonic irrigation is one option, and the other is subtotal or total colectomy. Segmental resection can preserve the proximal colonic segment, but on-table lavage is time-consuming and may result in fecal spillage[33]. Subtotal or total colectomy is advocated for low risk of anastomotic leakage in ileocolic or ileorectal anastomosis[34,35]. It can eliminate the distended proximal colon with ischemic lesions and serosal tears on the cecum, reducing the risk of fecal spillage and contamination. Subtotal or total colectomy can also effectively manage synchronous tumors in the proximal colon[36]. Negative aspects of subtotal or total colectomy include the need for an experienced surgeon or subspecialist, a prolonged operation time, and a decreased bowel function[37]. In terms of morbidity and mortality, a multicenter randomized controlled trial demonstrated no differences when two different surgical procedures - total/subtotal colectomy vs segmental colectomy with on-table lavage - were compared[37]. Therefore, the current 2017 WSES guideline states that total colectomy is not preferred to segmental colectomy in the absence of impending perforation in the cecum, evidence of bowel ischemia, or synchronous right colonic cancers[11].

As in right-sided obstructive colon cancer, endoscopic stent placement followed by the staged operation is an alternative option frequently applied in left-sided obstructive colon cancer. Although SEMS was first introduced for palliative treatment in unresectable colorectal cancer^[38], its use has been expanded to relieve colonic obstruction to avoid emergency surgery in resectable disease^[39]. Its role as a "bridge to surgery" for patients with curable disease has been widely accepted because it allows planned curative resection after the adequate restoration of the patient's general condition and bowel preparation^[40]. The use of SEMS appeared to decrease morbidity and mortality and the rate of stoma formation compared to emergency surgery^[41]. Ultimately, its efficacy and effectiveness in reducing medical costs and improving quality of life were shown to be associated with shorter hospital stays, fewer stays in the intensive care unit, and fewer surgical procedures[41-43].

However, other studies comparing the SEMS decompression outcome to emergency surgery demonstrated controversial results in terms of morbidity, mortality, and the stoma rate. An observational study by Kavanagh et al^[44] comparing emergency surgery to SEMS placement reported no difference in postoperative morbidity and mortality and the stoma formation rate, although technical and clinical success with SEMS placement was achieved in 91% and 83% of patients. A systematic review and meta-analysis including a total of 197 patients, 97 with SEMS placement vs 100 with emergency surgery, reported a clinical success rate of only 52.5% in the SEMS group in contrast to a rate of 99% in the emergency surgery group[45]. The overall complication rate and 30-d postoperative mortality rates of both groups were similar. The two groups also showed no significant difference in the permanent stoma rate or anastomotic leakage rate. In a systematic review and meta-analysis based on seven different randomized controlled trials, Huang et al [43] reported that the mean technical success rate was 77%. In contrast, the permanent stoma rate, primary anastomosis rate, and overall mortality rate were 9%, 67%, and 11%, respectively [43].



It appears that the location and length of an obstructing cancer lesion influence technical and clinical success^[46]. The European Society of Gastrointestinal Endoscopy (ESGE) guidelines, updated in 2020, suggests that by an experienced endoscopist or under the supervision of an expert, colonic stenting should be attempted with an individually tailored method to the length of the stenosis and location of the tumor [47].

Despite the controversy, the use of SEMS is endorsed by surgeons pursuing minimally invasive elective surgery with laparoscopy[48,49]. The multimodal approach using endoscopic and laparoscopic procedures demonstrated favorable short-term outcomes in SEMS placement as a bridge to surgery, offering less invasive treatment than multistage open surgery. In a meta-analysis including eight different randomized controlled trials comparing SEMS decompression as a bridge to surgery to emergency surgery from 2009 to 2016, Arezzo et al[50] confirmed similar rates of morbidity and mortality in both treatment approaches. Nonetheless, SEMS decompression showed a significantly lower rate of temporary and permanent stoma and a higher success rate of primary anastomosis than emergency surgery [50]. Based on many clinical studies and other randomized controlled trials, the 2017 WSES and ESGE guidelines concluded that SEMS decompression as a bridge to elective surgery offers a better short-term outcome than immediate emergency surgery [11,47].

The long-term oncological outcome remains uncertain and rather suboptimal when SEMS decompression was compared to emergency surgery, as shown in Table 1. Several meta-analyses evaluating the oncologic outcome of patients who underwent SEMS decompression followed by elective surgery demonstrated that disease-free survival and overall survival were not significantly different from those who received emergency surgery [51]. Additionally, in the recent update of the randomized controlled trial by Arezzo et al[52], the 3-year overall survival, time to progression, and disease-free survival of the patients with SEMS decompression as a bridge to surgery were not significantly different from those of the patients with emergency surgery. However, the meta-analysis of eight different randomized controlled trials by Yang et al[53] showed that patients with SEMS decompression presented a higher tumor recurrence rate than patients with emergency surgery, with an odds ratio of 1.79 (95% confidence interval 1.09-2.93). Another meta-analysis of seven randomized controlled trials by Foo et al[54] demonstrated that the overall recurrence rate in the SEMS decompression group was higher than that in the emergency surgery group, at 37.0% and 25.9%, respectively. Although the 3-year overall survival and disease-free survival were not different between the two groups, the risk ratio of systemic recurrence was 1.627 for the SEMS decompression group[54]. Nevertheless, available data are limited, and the sample size is inadequate to draw a firm conclusion on the long-term oncological outcome of SEMS decompression as a bridge to surgery. The ESGE guidelines currently recommend that decision-making for individual patients be influenced by the relative importance of particular endpoints, either short-term outcomes or long-term outcomes[47]. Patients should be informed within a shared decision-making process to use SEMS decompression as a bridge to surgery in potentially curable left-sided obstructive colon cancer regarding a potentially higher risk of recurrence^[47].

PITFALLS OF THE SEMS DECOMPRESSION FOLLOWED BY DELAYED OPERATION

Stent-related complications are a vital issue to address. A previous randomized controlled trial comparing SEMS placement to emergency surgery in left-sided obstructive colon cancer was terminated prematurely due to a high technical failure rate of 53.3%[49]. As the most lethal complication, colonic perforation during the procedure was the main reason for the premature closure of the trial. Other complications include stent migration, failure to expand within the colonic lumen, bleeding, and subsequent reobstruction[46]. The rate of colonic perforation related to stent placement was reported to be as high as 12.8% [55]. Stent migration and bleeding occur less frequently at a rate of 0% to less than 5% [55,56]. As expected, colonic perforation during the procedure, failure to self-expand, or bleeding requires emergency surgery, which may lead to mortality directly related to SEMS placement. Currently, the 30-d stent-related mortality is estimated to be 4% [46].

Stent-related perforation has not only an imminent mortality risk but also existing concerns about its implication in worsening oncological outcomes. In a recent systematic review and meta-analysis evaluating the oncological outcome of patients



Table 1 Oncological outcome after self-expandable metal stent placement as a bridge to surgery vs emergency surgery in malignant colonic obstruction

Ref.	Year	Study population	Study design	Location of obstructive cancer	Survival outcome
Matsuda <i>et al</i> [75]	2015	<i>n</i> = 1136: (1) BTS = 432; and (2) ES = 704	Meta-analysis: (1) 2 RCTs; (2) 2 prospective nonrandomized comparative studies; and (3) 7 retrospective comparative studies	Right- and left- sided	(1) No difference in disease-free survival and overall survival; and (2) No difference in recurrence
Ceresoli <i>et al</i> [76]	2017	n = 1333: (1) BTS = 688; (2) ES = 655	Meta-analysis: (1) 5 RCTs; (2) 3 prospective nonrandomized comparative studies; and (4) 9 retrospective comparative studies	Left-sided	(1) No difference in local recurrence and overall recurrence; (2) No difference in 3-yr and 5-yr recurrence; and (3) No difference in 3-yr and 5-yr mortality
Yang et al[53]	2018	<i>n</i> = 497: (1) BTS = 251; and (2) ES = 246	Meta-analysis: 8 RCTs	Left-sided	Higher tumor recurrence rate in BTS with an odds ratio of 1.79, 95%CI: 1.09–2.93
Amelung <i>et al</i> [51]	2018	<i>n</i> = 1919: (1) BTS = 938; and (2) ES = 981	Meta-analysis: (1) 5 RCTs; (2) 4 prospective nonrandomized comparative studies; and (3) 12 retrospective comparative studies	Left-sided	(1) No difference in locoregional recurrence and overall recurrence; (2) No difference in 3-yr and 5-yr disease-free survival; and (3) No difference in 3-yr and 5-yr overall survival
Foo <i>et al</i> [54]	2019	<i>n</i> = 448: (1) BTS = 222; and (2) ES = 226	Meta-analysis: 7 RCTs	Left-sided	(1) Overall recurrence rate: 37.0% in BTS <i>vs</i> 25.9% in ES; (2) The risk ratio of systemic recurrence 1.627 for BTS; and (3) No difference in 3-yr overall survival and disease-free survival
Arezzo <i>et al</i> [52] (ESCO trial)	2020	<i>n</i> = 115: (1) BTS = 56; and (2) ES = 59	RCT	Left-sided	No difference in 3-yr overall survival, time to progression, and disease-free survival

BTS: Bridge to surgery; ES: Emergency surgery; RCT: Randomized controlled trial; CI: Confidence interval.

who experienced stent-related perforation in an attempt at SEMS decompression, patients with stent-related perforation demonstrated significantly higher rates of global recurrence and locoregional recurrence than patients without perforation, although stent-related perforation did not influence the survival outcome[57]. A significantly increased risk of systemic and locoregional spread of the tumor was also observed in a previous meta-analysis by Foo et al[54,57]. Indeed, the current guidelines do not state SEMS decompression as a bridge to surgery as the treatment of choice in a potentially curable disease but consider it only as an alternative to emergency surgery in patients with an increased risk of perioperative morbidity and mortality, despite its better short-term outcome[11,47].

Several background theories explain the increased risk of tumor recurrence. First, the manipulation of a tumor during colonoscopy can cause the dissemination of cancer cells into peripheral circulation by mechanical compression of the guidewire and air insufflation, violating the principle of oncologic treatment[58]. Additionally, an increase in interstitial pressure within the tumor may cause tumor embolism in the lymphatic channels, resulting in lymphatic invasion[59]. Moreover, clinical or silent, stent-related perforation would promote cancer spread inside the peritoneal cavity, leading to locoregional and peritoneal metastasis[60]. Such arguments are supported by clinical correlations with pathologic findings, of which surgical specimens from patients with SEMS decompression presented a higher rate of perineural and lymphatic invasion^[61]. In a recent systematic review and meta-analysis by Balciscueta et al [60], patients who underwent SEMS decompression had a significantly higher risk of perineural and lymphatic invasion, with odds ratios of 1.98 and 1.45, respectively. The authors claim that stent placement as a bridge to surgery modifies the pathologic characteristics, including perineural and lymphatic invasion, which may worsen the long-term prognosis, raising the risk of global recurrence by 1.7 times the locoregional recurrence and carcinomatosis by 2.4 times[60]. However, pathologic findings of perineural and lymphatic invasion are often observed in obstructive colon cancer. From the available data, the significant influence of pathologic changes on survival outcomes is ambiguous. However, data on survival outcomes still lack a firm conclusion. Further translational research on how disruption in the microenvironment of tumors affects locoregional and systemic metastasis may delineate SEMS decompression's effect on survival outcome.



OPTIMAL INTERVAL BETWEEN SEMS PLACEMENT AND ELECTIVE SURGERY

The time interval between SEMS placement and elective surgery remains uncertain in the current management of obstructive colon cancer. At present, prospective comparative data on how the different intervals affect the short- and long-term outcomes are not available. Two retrospective studies have reported conflicting results regarding resection timing after decompression for postoperative morbidity and oncologic outcomes[62,63]. A retrospective study using the Dutch nationwide cohort demonstrated that surgery within 5-10 d resulted in a longer hospital stay, a lower rate of laparoscopic resection, and a higher rate of stoma creation than surgery after 11 d[63]. Additionally, stent-related complications were most frequently observed in patients who underwent surgery after 17 d[63]. On the other hand, a multicenter retrospective study on the optimal timing of elective surgery after SEMS placement by Kye et al[62] supports the concept that early elective surgery within seven days after SEMS placement correlates with better oncological outcomes than elective surgery after seven days. Currently, the ESGE guidelines state only that the time interval for surgery after SEMS placement should balance stent-related adverse events and surgical outcomes[47]. Further investigation is necessary to determine the optimal time interval between SEMS placement and elective surgery.

ROLE OF CIRCULATING TUMOR DNA

The development of genomic sequencing technology and molecular diagnostic testing has allowed the detection of tumor-specific DNA in peripheral blood samples, suggesting the new diagnostic concept of "liquid biopsy" [64]. Circulating cell-free DNA (cfDNA) is derived and released from apoptotic or necrotic cells, and circulating tumor DNA (ctDNA) with tumor-specific DNA from tumor cells undergoing apoptosis or necrosis is released into the systemic circulation[65]. Diagnostic strategies measuring ctDNA are under active investigation for clinical application in screening, diagnosis, and predicting tumor response or resistance to treatment[65]. The concentration of ctDNA in the bloodstream represents the tumor burden in individuals; thus, its use is highly accurate and valued as a biomarker for therapeutic monitoring[65,66].

Applying the concept of therapeutic monitoring, Takahashi *et al*^[67] evaluated ctDNA concentration changes after SEMS decompression to test whether SEMS placement disturbs the tumor microenvironment, causing cancer spread. In a prospective observational study, the authors observed that SEMS placement increased the ctDNA concentration in 83% of cases, indicating that SEMS placement inherently induces tumor manipulation and disruption[67]. The authors suggested that stentinduced tumor manipulation may worsen the prognosis of patients with obstructive colon cancer[67]. However, the effect of an increased ctDNA concentration on prognosis is still under investigation. A long-term follow-up study with a larger cohort is required to determine the effect of increased ctDNA on oncological outcomes.

Furthermore, the ctDNA concentration change pattern may help decide the timing and type of subsequent treatment. In the study by Takahashi et al[67], the ctDNA concentration changes after SEMS placement showed an increase in concentration over time. This finding implies that the longer time interval between SEMS placement and elective surgery may result in a more unsatisfactory oncological outcome. Since stentrelated complications occur mostly within seven days, a short interval after SEMS placement should be considered to minimize its impact on the survival outcome[47]. On the other hand, a long interval may optimize the patient's general condition and reduce the risk of postoperative complications[47]. Serial measurement of the ctDNA concentration may help decide the optimal timing for elective surgery for individual patients considering the long-term outcome.

APPLICATION OF NEOADJUVANT SYSTEMIC CHEMOTHERAPY IN **OBSTRUCTIVE COLON CANCER**

Neoadjuvant chemoradiotherapy has proven its benefit of downstaging tumors and reducing the local recurrence rate in locally advanced rectal cancer[68]. Recently, there has been a growing body of literature on the use of neoadjuvant chemotherapy before and after neoadjuvant concomitant chemoradiotherapy in locally advanced rectal



cancer for early control of micrometastasis, an increase in the complete response rate, conservative surgery with organ preservation, and an increase in adherence to chemotherapy^[69]. In colon cancer, since the mainstay of treatment for the potentially curable disease is complete oncologic resection, it is rare to find studies evaluating the effect of neoadjuvant chemotherapy, and only a few studies have been conducted to evaluate its safety [70-72]. The use of neoadjuvant chemotherapy after SEMS placement in patients with potentially curable obstructive colon cancer is rare. Only one retrospective analysis with a small sample size (n = 9) was found [73]. In this study, the efficacy and safety of SEMS insertion followed by neoadjuvant chemotherapy and elective surgery were evaluated, and the study results revealed relatively low toxicity, high adherence to two to three cycles of neoadjuvant chemotherapy before elective surgery and no evidence of perineural invasion in resected specimens^[73]. The authors suggested that neoadjuvant chemotherapy may lower the risk of perineural invasion, possibly improving survival outcomes[73]. In the FOXTROT trial, a randomized controlled trial evaluating the efficacy of neoadjuvant chemotherapy in locally advanced colon cancer, a few patients underwent SEMS placement as a bridge to surgery, and the results showed a significant decrease in the R1 resection rate and a nonsignificant trend toward better oncological outcomes at two years^[70]. However, it is difficult to draw meaningful conclusions about the effect of neoadjuvant chemotherapy after SEMS placement from these studies.

In obstructive colon cancer, the ESGE guidelines updated in 2020 recommend that the treatment strategy of SEMS placement followed by neoadjuvant chemotherapy is occasionally used in patients with stage IV disease for the early and safe introduction of systemic chemotherapy[47]. In a review article, Matsuda et al[74] suggested that systemic chemotherapy after SEMS placement may be an optimal treatment for patients with unresectable metastatic disease who are unfit for emergency surgery. However, the authors warned of delayed stent-related complications that require strict monitoring[74]. Additionally, the use of bevacizumab should be avoided because bevacizumab is a risk factor for stent-related perforation[74]. Nevertheless, for patients with stage IV obstructive colon cancer, an optimal outcome of upfront treatment for colonic obstruction would require immediate symptomatic control and minimizing complications related to emergent treatment for subsequent treatment for systemic disease. A personalized treatment approach should be implemented to achieve the ultimate result in an individual patient.

Perhaps an observational study utilizing a serial measurement of ctDNA concentration may help determine the efficacy of this treatment approach by decreasing the ctDNA concentration following neoadjuvant chemotherapy. A well-designed, randomized controlled trial evaluating the effect of neoadjuvant chemotherapy on the short- and long-term outcomes of SEMS placement followed by neoadjuvant chemotherapy and elective surgery in potentially resectable colon cancer would provide an ample amount of information regarding the management of obstructive colon cancer.

CONCLUSION

This article summarizes the current treatment strategies for obstructive colon cancer. Clinicians and surgeons often encounter complicated decision-making processes with complex and controversial treatment strategies. Decisions should be made based on the patient's index presentation of the general condition and risk factors that affect the short-term outcome. For the long-term outcome, it would be wise to implement a treatment approach that may induce a better oncological response in individual patients. Nevertheless, a diagnostic method for monitoring treatment response is still lacking. Clinical application of ctDNA analysis may offer new insights into individualized therapeutic strategies. Although several obstacles and preconditions should be resolved, incorporating neoadjuvant chemotherapy may be a viable option for effective systemic control of micrometastasis in selected patients. After all, further investigation on the implementation of neoadjuvant chemotherapy in the treatment of obstructive colon cancer is required.

REFERENCES



Chen TM, Huang YT, Wang GC. Outcome of colon cancer initially presenting as colon perforation and obstruction. World J Surg Oncol 2017; 15: 164 [PMID: 28841901 DOI:

10.1186/s12957-017-1228-y]

- 2 Dahdaleh FS, Sherman SK, Poli EC, Vigneswaran J, Polite BN, Sharma MR, Catenacci DV, Maron SB, Turaga KK. Obstruction predicts worse long-term outcomes in stage III colon cancer: A secondary analysis of the N0147 trial. Surgery 2018; 164: 1223-1229 [PMID: 30297240 DOI: 10.1016/j.surg.2018.06.044]
- 3 Decker KM, Lambert P, Nugent Z, Biswanger N, Samadder J, Singh H. Time Trends in the Diagnosis of Colorectal Cancer With Obstruction, Perforation, and Emergency Admission After the Introduction of Population-Based Organized Screening. JAMA Netw Open 2020; 3: e205741 [PMID: 32453385 DOI: 10.1001/jamanetworkopen.2020.5741]
- 4 Otani K, Kawai K, Hata K, Tanaka T, Nishikawa T, Sasaki K, Kaneko M, Murono K, Emoto S, Nozawa H. Colon cancer with perforation. Surg Today 2019; 49: 15-20 [PMID: 29691659 DOI: 10.1007/s00595-018-1661-8]
- 5 Cortet M, Grimault A, Cheynel N, Lepage C, Bouvier AM, Faivre J. Patterns of recurrence of obstructing colon cancers after surgery for cure: a population-based study. Colorectal Dis 2013; 15: 1100-1106 [PMID: 23634749 DOI: 10.1111/codi.12268]
- Høydahl Ø, Edna TH, Xanthoulis A, Lydersen S, Endreseth BH. Long-term trends in colorectal 6 cancer: incidence, localization, and presentation. BMC Cancer 2020; 20: 1077 [PMID: 33167924 DOI: 10.1186/s12885-020-07582-x]
- 7 Walls R, Hockberger R, Gausche-Hill M. Rosen's emergency medicine: Concepts and clinical practice. Philadelphia, PA: Elsevier Health Sciences; 2017
- Frago R, Ramirez E, Millan M, Kreisler E, del Valle E, Biondo S. Current management of acute 8 malignant large bowel obstruction: a systematic review. Am J Surg 2014; 207: 127-138 [PMID: 24124659 DOI: 10.1016/j.amjsurg.2013.07.027]
- Baer C, Menon R, Bastawrous S, Bastawrous A. Emergency Presentations of Colorectal Cancer. Surg Clin North Am 2017; 97: 529-545 [PMID: 28501245 DOI: 10.1016/j.suc.2017.01.004]
- 10 Benizri EI, Rahili A, Benchimol D. Emergency management of acute colonic cancer obstruction. J Visc Surg 2012; 149: e366-e367 [PMID: 22739385 DOI: 10.1016/j.jviscsurg.2012.05.006]
- Pisano M, Zorcolo L, Merli C, Cimbanassi S, Poiasina E, Ceresoli M, Agresta F, Allievi N, 11 Bellanova G, Coccolini F, Coy C, Fugazzola P, Martinez CA, Montori G, Paolillo C, Penachim TJ, Pereira B, Reis T, Restivo A, Rezende-Neto J, Sartelli M, Valentino M, Abu-Zidan FM, Ashkenazi I, Bala M, Chiara O, De' Angelis N, Deidda S, De Simone B, Di Saverio S, Finotti E, Kenji I, Moore E, Wexner S, Biffl W, Coimbra R, Guttadauro A, Leppäniemi A, Maier R, Magnone S, Mefire AC, Peitzmann A, Sakakushev B, Sugrue M, Viale P, Weber D, Kashuk J, Fraga GP, Kluger I, Catena F, Ansaloni L. 2017 WSES guidelines on colon and rectal cancer emergencies: obstruction and perforation. World J Emerg Surg 2018; 13: 36 [PMID: 30123315 DOI: 10.1186/s13017-018-0192-3]
- Boeding JRE, Ramphal W, Rijken AM, Crolla RMPH, Verhoef C, Gobardhan PD, Schreinemakers JMJ. A Systematic Review Comparing Emergency Resection and Staged Treatment for Curable Obstructing Right-Sided Colon Cancer. Ann Surg Oncol 2021; 28: 3545-3555 [PMID: 33067743 DOI: 10.1245/s10434-020-09124-v]
- Manceau G, Mege D, Bridoux V, Lakkis Z, Venara A, Voron T, Sielezneff I, Karoui M; French 13 Surgical Association Working Group. Emergency Surgery for Obstructive Colon Cancer in Elderly Patients: Results of a Multicentric Cohort of the French National Surgical Association. Dis Colon Rectum 2019; 62: 941-951 [PMID: 31283592 DOI: 10.1097/DCR.000000000001421]
- 14 Frago R, Biondo S, Millan M, Kreisler E, Golda T, Fraccalvieri D, Miguel B, Jaurrieta E. Differences between proximal and distal obstructing colonic cancer after curative surgery. Colorectal Dis 2011; 13: e116-e122 [PMID: 21564463 DOI: 10.1111/j.1463-1318.2010.02549.x]
- Frasson M, Granero-Castro P, Ramos Rodríguez JL, Flor-Lorente B, Braithwaite M, Martí Martínez 15 E, Álvarez Pérez JA, Codina Cazador A, Espí A, Garcia-Granero E; ANACO Study Group. Risk factors for anastomotic leak and postoperative morbidity and mortality after elective right colectomy for cancer: results from a prospective, multicentric study of 1102 patients. Int J Colorectal Dis 2016; 31: 105-114 [PMID: 26315015 DOI: 10.1007/s00384-015-2376-6]
- Voron T. Bruzzi M, Ragot E, Zinzindohoue F, Chevallier JM, Douard R, Berger A, Anastomotic 16 Location Predicts Anastomotic Leakage After Elective Colonic Resection for Cancer. J Gastrointest Surg 2019; 23: 339-347 [PMID: 30076589 DOI: 10.1007/s11605-018-3891-x]
- Alves A, Panis Y, Trancart D, Regimbeau JM, Pocard M, Valleur P. Factors associated with clinically 17 significant anastomotic leakage after large bowel resection: multivariate analysis of 707 patients. World J Surg 2002; 26: 499-502 [PMID: 11910487 DOI: 10.1007/s00268-001-0256-4]
- Lim CH, McDonald NM, Freeman ML, Amateau SK, Percutaneous cecostomy with fully covered 18 self-expandable metal stent for initial management of severe malignant colon obstruction. Endoscopy 2017; 49: E313-E315 [PMID: 28992638 DOI: 10.1055/s-0043-119973]
- 19 Suzuki Y, Moritani K, Seo Y, Takahashi T. Comparison of decompression tubes with metallic stents for the management of right-sided malignant colonic obstruction. World J Gastroenterol 2019; 25: 1975-1985 [PMID: 31086465 DOI: 10.3748/wjg.v25.i16.1975]
- 20 Meyer F, Marusch F, Koch A, Meyer L, Führer S, Köckerling F, Lippert H, Gastinger I; German Study Group "Colorectal Carcinoma (Primary Tumor)". Emergency operation in carcinomas of the left colon: value of Hartmann's procedure. Tech Coloproctol 2004; 8 Suppl 1: s226-s229 [PMID: 15655630 DOI: 10.1007/s10151-004-0164-3]
- 21 Timmermans DR, Gooszen AW, Geelkerken RH, Tollenaar RA, Gooszen HG. Analysis of the variety in surgeons' decision strategies for the management of left colonic emergencies. Med Care



1997; 35: 701-713 [PMID: 9219497 DOI: 10.1097/00005650-199707000-00004]

- Alves A, Panis Y, Mathieu P, Mantion G, Kwiatkowski F, Slim K; Association Française de 22 Chirurgie. Postoperative mortality and morbidity in French patients undergoing colorectal surgery: results of a prospective multicenter study. Arch Surg 2005; 140: 278-283, discussion 284 [PMID: 15781793 DOI: 10.1001/archsurg.140.3.278]
- 23 Irvin GL 3rd, Horsley JS 3rd, Caruana JA Jr. The morbidity and mortality of emergent operations for colorectal disease. Ann Surg 1984; 199: 598-603 [PMID: 6609686 DOI: 10.1097/00000658-198405000-00015
- 24 Hallam S, Mothe BS, Tirumulaju R. Hartmann's procedure, reversal and rate of stoma-free survival. Ann R Coll Surg Engl 2018; 100: 301-307 [PMID: 29484943 DOI: 10.1308/rcsann.2018.0006]
- 25 Kang JH, Kang BM, Yoon SN, Kim JY, Park JH, Oh BY, Kim JW. Analysis of factors affecting reversal of Hartmann's procedure and post-reversal complications. Sci Rep 2020; 10: 16820 [PMID: 33033297 DOI: 10.1038/s41598-020-73791-w]
- Whitney S, Gross BD, Mui A, Hahn S, Read B, Bauer J. Hartmann's reversal: factors affecting 26 complications and outcomes. Int J Colorectal Dis 2020; 35: 1875-1880 [PMID: 32504334 DOI: 10.1007/s00384-020-03653-4]
- Kronborg O. Acute obstruction from tumour in the left colon without spread. A randomized trial of 27 emergency colostomy versus resection. Int J Colorectal Dis 1995; 10: 1-5 [PMID: 7745314 DOI: 10.1007/bf00337576
- De Salvo GL, Gava C, Pucciarelli S, Lise M. Curative surgery for obstruction from primary left 28 colorectal carcinoma: primary or staged resection? Cochrane Database Syst Rev 2004; CD002101 [PMID: 15106167 DOI: 10.1002/14651858.CD002101.pub2]
- 29 Zorcolo L, Covotta L, Carlomagno N, Bartolo DC. Safety of primary anastomosis in emergency colorectal surgery. Colorectal Dis 2003; 5: 262-269 [PMID: 12780890 DOI: 10.1046/j.1463-1318.2003.00432.x]
- Villar JM, Martinez AP, Villegas MT, Muffak K, Mansilla A, Garrote D, Ferron JA. Surgical options 30 for malignant left-sided colonic obstruction. Surg Today 2005; 35: 275-281 [PMID: 15815842 DOI: 10.1007/s00595-004-2931-1]
- Breitenstein S, Rickenbacher A, Berdajs D, Puhan M, Clavien PA, Demartines N. Systematic 31 evaluation of surgical strategies for acute malignant left-sided colonic obstruction. Br J Surg 2007; 94: 1451-1460 [PMID: 17968980 DOI: 10.1002/bjs.6007]
- Biondo S, Parés D, Frago R, Martí-Ragué J, Kreisler E, De Oca J, Jaurrieta E. Large bowel 32 obstruction: predictive factors for postoperative mortality. Dis Colon Rectum 2004; 47: 1889-1897 [PMID: 15622582 DOI: 10.1007/s10350-004-0688-7]
- 33 Awotar GK, Guan G, Sun W, Yu H, Zhu M, Cui X, Liu J, Chen J, Yang B, Lin J, Deng Z, Luo J, Wang C, Nur OA, Dhiman P, Liu P, Luo F. Reviewing the Management of Obstructive Left Colon Cancer: Assessing the Feasibility of the One-stage Resection and Anastomosis After Intraoperative Colonic Irrigation. Clin Colorectal Cancer 2017; 16: e89-e103 [PMID: 28254356 DOI: 10.1016/j.clcc.2016.12.001
- 34 Hsu TC. Comparison of one-stage resection and anastomosis of acute complete obstruction of left and right colon. Am J Surg 2005; 189: 384-387 [PMID: 15820447 DOI: 10.1016/j.amjsurg.2004.06.046]
- Kluger Y, Shiloni E, Jurim O, Katz E, Rivkind A, Ayalon A, Durst A. Subtotal colectomy with 35 primary ileocolonic anastomosis for obstructing carcinoma of the left colon: valid option for elderly high risk patients. Isr J Med Sci 1993; 29: 726-730 [PMID: 8270407 DOI: 10.1007/BF02942191]
- 36 Gainant A. Emergency management of acute colonic cancer obstruction. J Visc Surg 2012; 149: e3e10 [PMID: 22189474 DOI: 10.1016/j.jviscsurg.2011.11.003]
- 37 Single-stage treatment for malignant left-sided colonic obstruction: a prospective randomized clinical trial comparing subtotal colectomy with segmental resection following intraoperative irrigation. The SCOTIA Study Group. Subtotal Colectomy versus On-table Irrigation and Anastomosis. Br J Surg 1995; 82: 1622-1627 [PMID: 8548221 DOI: 10.1002/bjs.1800821211]
- Rupp KD, Dohmoto M, Meffert R, Holzgreve A, Hohlbach G. Cancer of the rectum--palliative 38 endoscopic treatment. Eur J Surg Oncol 1995; 21: 644-647 [PMID: 8631413 DOI: 10.1016/s0748-7983(95)95563-1]
- Schwenter F, Morel P, Gervaz P. Management of obstructive and perforated colorectal cancer. -39 Expert Rev Anticancer Ther 2010; 10: 1613-1619 [PMID: 20942632 DOI: 10.1586/era.10.147]
- Tung KL, Cheung HY, Ng LW, Chung CC, Li MK. Endo-laparoscopic approach versus conventional 40 open surgery in the treatment of obstructing left-sided colon cancer: long-term follow-up of a randomized trial. Asian J Endosc Surg 2013; 6: 78-81 [PMID: 23601995 DOI: 10.1111/ases.12030]
- 41 Sebastian S, Johnston S, Geoghegan T, Torreggiani W, Buckley M. Pooled analysis of the efficacy and safety of self-expanding metal stenting in malignant colorectal obstruction. Am J Gastroenterol 2004; **99**: 2051-2057 [PMID: 15447772 DOI: 10.1111/j.1572-0241.2004.40017.x]
- 42 Xinopoulos D, Dimitroulopoulos D, Theodosopoulos T, Tsamakidis K, Bitsakou G, Plataniotis G, Gontikakis M, Kontis M, Paraskevas I, Vassilobpoulos P, Paraskevas E. Stenting or stoma creation for patients with inoperable malignant colonic obstructions? Surg Endosc 2004; 18: 421-426 [PMID: 14735348 DOI: 10.1007/s00464-003-8109-x]
- Huang X, Lv B, Zhang S, Meng L. Preoperative colonic stents versus emergency surgery for acute 43 left-sided malignant colonic obstruction: a meta-analysis. J Gastrointest Surg 2014; 18: 584-591 [PMID: 24170606 DOI: 10.1007/s11605-013-2344-9]



- Kavanagh DO, Nolan B, Judge C, Hyland JM, Mulcahy HE, O'Connell PR, Winter DC, Doherty 44 GA. A comparative study of short- and medium-term outcomes comparing emergent surgery and stenting as a bridge to surgery in patients with acute malignant colonic obstruction. Dis Colon Rectum 2013; 56: 433-440 [PMID: 23478610 DOI: 10.1097/DCR.0b013e3182760506]
- Cirocchi R, Farinella E, Trastulli S, Desiderio J, Listorti C, Boselli C, Parisi A, Noya G, Sagar J. 45 Safety and efficacy of endoscopic colonic stenting as a bridge to surgery in the management of intestinal obstruction due to left colon and rectal cancer: a systematic review and meta-analysis. Surg Oncol 2013; 22: 14-21 [PMID: 23183301 DOI: 10.1016/j.suronc.2012.10.003]
- Lee JM, Byeon JS. Colorectal Stents: Current Status. Clin Endosc 2015; 48: 194-200 [PMID: 46 26064818 DOI: 10.5946/ce.2015.48.3.194]
- 47 van Hooft JE, Veld JV, Arnold D, Beets-Tan RGH, Everett S, Götz M, van Halsema EE, Hill J, Manes G, Meisner S, Rodrigues-Pinto E, Sabbagh C, Vandervoort J, Tanis PJ, Vanbiervliet G, Arezzo A. Self-expandable metal stents for obstructing colonic and extracolonic cancer: European Society of Gastrointestinal Endoscopy (ESGE) Guideline - Update 2020. Endoscopy 2020; 52: 389-407 [PMID: 32259849 DOI: 10.1055/a-1140-3017]
- 48 Cheung HY, Chung CC, Tsang WW, Wong JC, Yau KK, Li MK. Endolaparoscopic approach vs conventional open surgery in the treatment of obstructing left-sided colon cancer: a randomized controlled trial. Arch Surg 2009; 144: 1127-1132 [PMID: 20026830 DOI: 10.1001/archsurg.2009.216
- Pirlet IA, Slim K, Kwiatkowski F, Michot F, Millat BL. Emergency preoperative stenting versus 49 surgery for acute left-sided malignant colonic obstruction: a multicenter randomized controlled trial. Surg Endosc 2011; 25: 1814-1821 [PMID: 21170659 DOI: 10.1007/s00464-010-1471-6]
- 50 Arezzo A, Passera R, Lo Secco G, Verra M, Bonino MA, Targarona E, Morino M. Stent as bridge to surgery for left-sided malignant colonic obstruction reduces adverse events and stoma rate compared with emergency surgery: results of a systematic review and meta-analysis of randomized controlled trials. Gastrointest Endosc 2017; 86: 416-426 [PMID: 28392363 DOI: 10.1016/j.gie.2017.03.1542]
- Amelung FJ, Burghgraef TA, Tanis PJ, van Hooft JE, Ter Borg F, Siersema PD, Bemelman WA, 51 Consten ECJ. Critical appraisal of oncological safety of stent as bridge to surgery in left-sided obstructing colon cancer; a systematic review and meta-analysis. Crit Rev Oncol Hematol 2018; 131: 66-75 [PMID: 30293707 DOI: 10.1016/j.critrevonc.2018.08.003]
- 52 Arezzo A, Forcignanò E, Bonino MA, Balagué C, Targarona E, Borghi F, Giraudo G, Ghezzo L, Passera R, Morino M; collaborative ESCO study group. Long-term Oncologic Results After Stenting as a Bridge to Surgery Versus Emergency Surgery for Malignant Left-sided Colonic Obstruction: A Multicenter Randomized Controlled Trial (ESCO Trial). Ann Surg 2020; 272: 703-708 [PMID: 32833762 DOI: 10.1097/SLA.00000000004324]
- Yang P, Lin XF, Lin K, Li W. The Role of Stents as Bridge to Surgery for Acute Left-Sided 53 Obstructive Colorectal Cancer: Meta-Analysis of Randomized Controlled Trials. Rev Invest Clin 2018; 70: 269-278 [PMID: 30532112 DOI: 10.24875/RIC.18002516]
- Foo CC, Poon SHT, Chiu RHY, Lam WY, Cheung LC, Law WL. Is bridge to surgery stenting a safe 54 alternative to emergency surgery in malignant colonic obstruction: a meta-analysis of randomized control trials. Surg Endosc 2019; 33: 293-302 [PMID: 30341649 DOI: 10.1007/s00464-018-6487-3]
- Yoon JY, Jung YS, Hong SP, Kim TI, Kim WH, Cheon JH. Clinical outcomes and risk factors for 55 technical and clinical failures of self-expandable metal stent insertion for malignant colorectal obstruction. Gastrointest Endosc 2011; 74: 858-868 [PMID: 21862005 DOI: 10.1016/j.gie.2011.05.044]
- Meisner S, González-Huix F, Vandervoort JG, Goldberg P, Casellas JA, Roncero O, Grund KE, 56 Alvarez A, García-Cano J, Vázquez-Astray E, Jiménez-Pérez J; WallFlex Colonic Registry Group. Self-expandable metal stents for relieving malignant colorectal obstruction: short-term safety and efficacy within 30 days of stent procedure in 447 patients. Gastrointest Endosc 2011; 74: 876-884 [PMID: 21855868 DOI: 10.1016/j.gie.2011.06.019]
- Balciscueta I, Balciscueta Z, Uribe N, García-Granero E. Long-term outcomes of stent-related 57 perforation in malignant colon obstruction: a systematic review and meta-analysis. Int J Colorectal Dis 2020; 35: 1439-1451 [PMID: 32572603 DOI: 10.1007/s00384-020-03664-1]
- Maruthachalam K, Lash GE, Shenton BK, Horgan AF. Tumour cell dissemination following 58 endoscopic stent insertion. Br J Surg 2007; 94: 1151-1154 [PMID: 17541987 DOI: 10.1002/bjs.5790]
- 59 Hayashi K, Jiang P, Yamauchi K, Yamamoto N, Tsuchiya H, Tomita K, Moossa AR, Bouvet M, Hoffman RM. Real-time imaging of tumor-cell shedding and trafficking in lymphatic channels. Cancer Res 2007; 67: 8223-8228 [PMID: 17804736 DOI: 10.1158/0008-5472.Can-07-1237]
- 60 Balciscueta I, Balciscueta Z, Uribe N, García-Granero E. Perineural invasion is increased in patients receiving colonic stenting as a bridge to surgery: a systematic review and meta-analysis. Tech Coloproctol 2021; 25: 167-176 [PMID: 33200308 DOI: 10.1007/s10151-020-02350-2]
- 61 Hu Y, Fan J, Xv Y, Hu Y, Ding Y, Jiang Z, Tao Q. Comparison of safety between self-expanding metal stents as a bridge to surgery and emergency surgery based on pathology: a meta-analysis. BMC Surg 2020; 20: 255 [PMID: 33109142 DOI: 10.1186/s12893-020-00908-3]
- Kye BH, Kim JH, Kim HJ, Lee YS, Lee IK, Kang WK, Cho HM, Ahn CH, Oh ST. The optimal time 62 interval between the placement of self-expandable metallic stent and elective surgery in patients with obstructive colon cancer. Sci Rep 2020; 10: 9502 [PMID: 32528099 DOI: 10.1038/s41598-020-66508-6



- Veld JV, Kumcu A, Amelung FJ, Borstlap WAA, Consten ECJ, Dekker JWT, van Westreenen HL, 63 Siersema PD, Ter Borg F, Kusters M, Bemelman WA, de Wilt JHW, van Hooft JE, Tanis PJ; Dutch Snapshot Research Group. Time interval between self-expandable metal stent placement or creation of a decompressing stoma and elective resection of left-sided obstructive colon cancer. Endoscopy 2021; 53: 905-913 [PMID: 33339059 DOI: 10.1055/a-1308-1487]
- 64 Osumi H, Shinozaki E, Yamaguchi K, Zembutsu H. Clinical utility of circulating tumor DNA for colorectal cancer. Cancer Sci 2019; 110: 1148-1155 [PMID: 30742729 DOI: 10.1111/cas.13972]
- Moati E, Taly V, Didelot A, Perkins G, Blons H, Taieb J, Laurent-Puig P, Zaanan A. Role of 65 circulating tumor DNA in the management of patients with colorectal cancer. Clin Res Hepatol Gastroenterol 2018; 42: 396-402 [PMID: 29627453 DOI: 10.1016/j.clinre.2018.03.002]
- Khakoo S, Georgiou A, Gerlinger M, Cunningham D, Starling N. Circulating tumour DNA, a 66 promising biomarker for the management of colorectal cancer. Crit Rev Oncol Hematol 2018; 122: 72-82 [PMID: 29458792 DOI: 10.1016/j.critrevonc.2017.12.002]
- Takahashi G, Yamada T, Iwai T, Takeda K, Koizumi M, Shinji S, Uchida E. Oncological 67 Assessment of Stent Placement for Obstructive Colorectal Cancer from Circulating Cell-Free DNA and Circulating Tumor DNA Dynamics. Ann Surg Oncol 2018; 25: 737-744 [PMID: 29235008 DOI: 10.1245/s10434-017-6300-x]
- Benson AB, Venook AP, Al-Hawary MM, Arain MA, Chen YJ, Ciombor KK, Cohen S, Cooper HS, 68 Deming D, Garrido-Laguna I, Grem JL, Gunn A, Hoffe S, Hubbard J, Hunt S, Kirilcuk N, Krishnamurthi S, Messersmith WA, Meyerhardt J, Miller ED, Mulcahy MF, Nurkin S, Overman MJ, Parikh A, Patel H, Pedersen K, Saltz L, Schneider C, Shibata D, Skibber JM, Sofocleous CT, Stoffel EM, Stotsky-Himelfarb E, Willett CG, Johnson-Chilla A, Gurski LA. NCCN Guidelines Insights: Rectal Cancer, Version 6.2020. J Natl Compr Canc Netw 2020; 18: 806-815 [PMID: 32634771 DOI: 10.6004/inccn.2020.0032]
- Yoo RN, Kim HJ. Total neoadjuvant therapy in locally advanced rectal cancer: Role of systemic 69 chemotherapy. Ann Gastroenterol Surg 2019; 3: 356-367 [PMID: 31346574 DOI: 10.1002/ags3.12253]
- Foxtrot Collaborative Group. Feasibility of preoperative chemotherapy for locally advanced, 70 operable colon cancer: the pilot phase of a randomised controlled trial. Lancet Oncol 2012; 13: 1152-1160 [PMID: 23017669 DOI: 10.1016/S1470-2045(12)70348-0]
- Arredondo J, Baixauli J, Pastor C, Chopitea A, Sola JJ, González I, A-Cienfuegos J, Martínez P, 71 Rodriguez J, Hernández-Lizoain JL. Mid-term oncologic outcome of a novel approach for locally advanced colon cancer with neoadjuvant chemotherapy and surgery. Clin Transl Oncol 2017; 19: 379-385 [PMID: 27496023 DOI: 10.1007/s12094-016-1539-4]
- Jakobsen A, Andersen F, Fischer A, Jensen LH, Jørgensen JC, Larsen O, Lindebjerg J, Pløen J, 72 Rafaelsen SR, Vilandt J. Neoadjuvant chemotherapy in locally advanced colon cancer. A phase II trial. Acta Oncol 2015; 54: 1747-1753 [PMID: 25920359 DOI: 10.3109/0284186X.2015.1037007]
- Han JG, Wang ZJ, Zeng WG, Wang YB, Wei GH, Zhai ZW, Zhao BC, Yi BQ. Efficacy and safety 73 of self-expanding metallic stent placement followed by neoadjuvant chemotherapy and scheduled surgery for treatment of obstructing left-sided colonic cancer. BMC Cancer 2020; 20: 57 [PMID: 31992260 DOI: 10.1186/s12885-020-6560-x]
- Matsuda A, Yamada T, Matsumoto S, Shinji S, Ohta R, Sonoda H, Takahashi G, Iwai T, Takeda K, 74 Sekiguchi K, Yoshida H. Systemic Chemotherapy is a Promising Treatment Option for Patients with Colonic Stents: A Review. J Anus Rectum Colon 2021; 5: 1-10 [PMID: 33537495 DOI: 10.23922/jarc.2020-061]
- 75 Matsuda A, Miyashita M, Matsumoto S, Matsutani T, Sakurazawa N, Takahashi G, Kishi T, Uchida E. Comparison of long-term outcomes of colonic stent as "bridge to surgery" and emergency surgery for malignant large-bowel obstruction: a meta-analysis. Ann Surg Oncol 2015; 22: 497-504 [PMID: 25120255 DOI: 10.1245/s10434-014-3997-7]
- Ceresoli M, Allievi N, Coccolini F, Montori G, Fugazzola P, Pisano M, Sartelli M, Catena F, 76 Ansaloni L. Long-term oncologic outcomes of stent as a bridge to surgery versus emergency surgery in malignant left side colonic obstructions: a meta-analysis. J Gastrointest Oncol 2017; 8: 867-876 [PMID: 29184691 DOI: 10.21037/jgo.2017.09.04]



0 W J

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 1863-1879

DOI: 10.4251/wjgo.v13.i12.1863

ISSN 1948-5204 (online)

REVIEW

Role of endoscopic ultrasound in anticancer therapy: Current evidence and future perspectives

Andre Bratanic, Dorotea Bozic, Antonio Mestrovic, Dinko Martinovic, Marko Kumric, Tina Ticinovic Kurir, Josko Bozic

ORCID number: Andre Bratanic 0000-0002-3261-183X; Dorotea Bozic 0000-0001-9234-4203: Antonio Mestrovic 0000-0002-0156-2748; Dinko Martinovic 0000-0003-2060-5130; Marko Kumric 0000-0002-9696-3359; Tina Ticinovic Kurir 0000-0003-1695-9235; Josko Bozic 0000-0003-1634-0635.

Author contributions: Bratanic A, Bozic D, Kumric M, Ticinovic Kurir T, and Bozic J conceptualized, wrote the original draft, and supervised the work; Bratanic A, Martinovic D. and Mestrovic A reviewed the literature: all authors contributed to the final draft of the manuscript and approved of the final version of the manuscript.

Conflict-of-interest statement: The authors declare no conflicts of interest.

Country/Territory of origin: Croatia

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0

Andre Bratanic, Dorotea Bozic, Antonio Mestrovic, Department of Gastroenterology and Hepatology, University Hospital of Split, Split 21000, Croatia

Dinko Martinovic, Marko Kumric, Tina Ticinovic Kurir, Josko Bozic, Department of Pathophysiology, University of Split School of Medicine, Split 21000, Croatia

Tina Ticinovic Kurir, Department of Endocrinology, University Hospital of Split, Split 21000, Croatia

Corresponding author: Josko Bozic, MD, PhD, Associate Professor, Department of Pathophysiology, University of Split School of Medicine, Soltanska 2, Split 21000, Croatia. josko.bozic@mefst.hr

Abstract

The digestive system is one of the most common sites of malignancies in humans. Since gastrointestinal tumors represent a massive global health burden both in terms of morbidity and health care expenditures, scientists continuously develop novel diagnostic and therapeutic methods to ameliorate the detrimental effects of this group of diseases. Apart from the well-established role of the endoscopic ultrasound (EUS) in the diagnostic course of gastrointestinal and hepatobiliary malignancies, we have recently become acquainted with a vast array of its therapeutic possibilities. A multitude of previously established, evidence-based methods that might now be guided by the EUS emerged: Radiofrequency ablation, brachytherapy, fine needle injection, celiac plexus neurolysis, and endoscopic submucosal dissection. In this review we endeavored to provide a comprehensive overview of the role of these methods in different malignancies of the digestive system, primarily in the treatment and symptom control in pancreatic cancer, and additionally in the management of hepatic, gastrointestinal tumors, and pancreatic cysts.

Key Words: Pancreatic cancer; Endoscopic ultrasound; Endoscopic ultrasound-guided fine needle injection; Pancreatic cyst; Gastrointestinal tumor; Portal vein

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.



Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: February 21, 2021 Peer-review started: February 21, 2021 First decision: May 8, 2021 Revised: May 17, 2021 Accepted: August 27, 2021 Article in press: August 27, 2021 Published online: December 15, 2021

P-Reviewer: Jagielski M S-Editor: Ma YJ L-Editor: Filipodia P-Editor: Ma YJ



Core Tip: Apart from the well-established role of the endoscopic ultrasound (EUS) along the diagnostic path of gastrointestinal and hepatobiliary malignancies, the EUS recently emerged as a carrier of various therapeutic modalities. In this review we sought to give a comprehensive overview of the role of various established methods that might now be guided by the EUS in different malignancies of the digestive system, primarily regarding the treatment and symptom control in pancreatic cancer, and additionally in the management of hepatic, gastrointestinal tumors, and pancreatic cysts.

Citation: Bratanic A, Bozic D, Mestrovic A, Martinovic D, Kumric M, Ticinovic Kurir T, Bozic J. Role of endoscopic ultrasound in anticancer therapy: Current evidence and future perspectives. World J Gastrointest Oncol 2021; 13(12): 1863-1879 URL: https://www.wjgnet.com/1948-5204/full/v13/i12/1863.htm

DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.1863

INTRODUCTION

Endoscopic ultrasound (EUS) is one of the principal tools in cancer screening and staging. Apart from its use in the diagnostic course of gastrointestinal and hepatobiliary malignancies, it has an entire range of other therapeutic possibilities (Figure 1). The ability of the EUS to obtain tissue samples using either fine needle aspiration (FNA) or fine needle biopsy makes it a unique method in the screening of pancreatic cystic lesions, as well as in the assessment of regional lymph node involvement in esophageal, gastric, and rectal cancer (Figures 2-4). The EUS is therefore essential in the concurrent pancreatic cancer diagnosis[1]. Over the last few years, the implementation of the EUS has expanded from the diagnostic into the therapeutic field. Endoscopic ultrasound-guided fine needle injection (EUS-FNI) of chemotherapeutics, immunotherapy and gene therapy, tissue ablation, stereotactic radiation therapy, brachytherapy and celiac plexus neurolysis have been thoroughly investigated and steadily introduced into the clinical practice [1,2]. Multiple studies suggest that some of these methods might be crucial in overcoming the problem of drug distribution to tumorous tissue in patients with pancreatic cancer, whereas other methods could be of aid in pain management of the same population. In this review we endeavored to give a comprehensive overview of the role of the EUS in anticancer therapy: primarily in the treatment and symptom control in pancreatic cancer, and, additionally, in the management of hepatic and gastrointestinal tumors.

THE EUS IN TREATMENT OF PANCREATIC CANCER

As hypovascularity and abundant desmoplasia are landmarks of pancreatic cancer, delivery of chemotherapeutic medications to the tumor-affected area is insufficient[3]. Consequently, effects of chemotherapeutics on pancreatic cancer are mitigated, resulting in higher required therapeutic doses which increases the incidence of adverse effects[4]. Therefore, several local strategies that could possibly overcome these issues were developed. Among a number of EUS-based therapeutic interventions, radiofrequency ablation (RFA), brachytherapy, EUS-FNI, and EUS-guided celiac plexus neurolysis (EUS-CPN) emerged as viable strategies for improvement of poor outcomes regarding the pancreatic cancer.

RFA

RFA is an invasive antitumor method which works by generating heat from highfrequency alternating current that induces frictional heating, leading to thermal coagulative necrosis of the target tissue[5]. In addition, several authors demonstrated that RFA may trigger immunomodulatory activity, and in this way, further dampening the cancer development[6-9]. RFA is already a well-established therapeutic modality in management of other cancers, particularly for patients with hepatocellular carcinoma, and has been included in the latest guidelines for management of hepatocellular carcinoma by a major European organization[10].





Figure 1 Overview of endoscopic ultrasound-guided methods. EUS-FNI: Endoscopic ultrasound-fine needle injection; HCC: Hepatocellular carcinoma; GI: Gastrointestinal; Nd:YAG: Neodymium-doped yttrium aluminum garnet; SBRT: Stereotactic body radiotherapy.



Figure 2 Endoscopic ultrasound-fine needle aspiration. Fine needle aspiration of inhomogeneous oval lesion located on the border between head and corpus of the pancreas (26.6 mm × 21.5 mm).

Owing to the anatomical positioning of the pancreas, suitable approaches for the inclusion of the RFA in pancreatic cancer are multiple, including open surgery, laparoscopic approach, percutaneous approach, and an EUS-guided approach. Initially, safety and effectiveness of EUS-guided RFA of the pancreatic tissue were evaluated in porcine models[11-14]. Although results of the animal studies were promising, clinicians were doubtful, as thermal-induced pancreatitis and thermal injuries of the adjoining structures emerged as adverse effects in the animal studies and in early intraoperative studies[11,15-17]. Early on, surgeons observed that detrimental effects of intraoperative RFA could be reduced by using lower temperatures, maintaining a margin from the major adjacent vessels, and by using stepwise approach in bigger, poorly demarcated lesions[18,19]. Intraoperative studies have also provided an insight into the effectiveness of the RFA. Multiple studies have demonstrated that intraoperative RFA leads to tumor necrosis and a decrease of tumor volume, as well as reduction in CA 19-9 plasma levels, the main pancreatic cancer marker[16,20-25]. Unfortunately, in all of the aforementioned studies, all patients that underwent RFA



Figure 3 Unsuccessful endoscopic ultrasound-fine needle aspiration. Unsuccessful attempt of fine needle aspiration of the cystic lesion with thick, calcified border (37 mm × 26 mm) located in the head of the pancreas.



Figure 4 Endoscopic ultrasound-fine needle biopsy. Fine needle biopsy of the focal lesion in the pancreatic head (42 mm × 38 mm).

eventually developed disease progression [16,20-25]. Of note, in a small sample (n = 25) study by Spiliotis et al[16], authors have shown a significant prolongation in survival in patients treated with both palliative care and RFA in opposition to patients that received palliative care exclusively, an observation that Matsui et al[25] failed to demonstrate. In addition, Cantore et al[26] argue that combined multiple-treatment followed by RFA can prolong survival in patients with unresectable pancreatic cancer.

Together, these findings paved the way for the investigation of the EUS-guided RFA. In a pilot study by Song et al[27], 6 patients with unresectable pancreatic cancer underwent EUS-guided RFA. Two out of six patients suffered abdominal pain, whereas other patients reported no adverse, effects implying the safety of the procedure. However, as the purpose of the study was to assess the technical feasibility and safety of the procedure, long-term survival of the patients was not studied. Multiple similar studies were performed following this study, and their results with regard to effectiveness and adverse events were summarized in a recent meta-analysis by Dhaliwal et al[28]. Meta-analysis showed that technical success, defined as the



successful placement of the needle within the pancreatic lesions with safe margins from the surrounding vital structure, and clinical success, defined as decrease in lesion size and presence of necrosis on CT scan after the procedure, were 100% and 91.5%, respectively. Adverse events were observed in 15% of the patients, with abdominal pain being the most common (10%) and only 2 cases of pancreatitis and 1 perforation in total. Overall, the EUS-guided RFA appears to be safe, but multicenter, randomized control trials are needed in order to clearly define the utility of this method.

EUS-FNI

EUS-FNI is an antitumor agent delivery method in which the EUS serves as a guide for needle placement into target lesions. Aside from injection of various therapeutic agents, EUS-FNI is also suitable for implantation of fiducial markers that enables targeted radiation therapy and for injection of dyes to tattoo the tumor [29,30]. The first human clinical trial (phase I) using the EUS-FNI was performed by Chang *et al*[31] in 2000. Authors used EUS-FNI to deliver allogeneic mixed lymphocyte culture (cytoimplant) into the tumor tissue. The median survival of the 8 patients enrolled in the study was 13.2 mo, with 3 partial responses and no adverse events reported. Since then, the largest conducted trial of this sort was a phase III trial in which effects of TNFerade[™]biologic, in combination with fluorouracil and radiotherapy, were assessed on 304 patients with advanced pancreatic cancer[32]. Unfortunately, Herman et al[32] failed to demonstrate better progression-free survival or overall survival in comparison to controls. Recently Lee et al[33] conducted a phase I trial in which authors combined adenovirus-mediated double-suicide gene therapy with gemcitabine in patients with locally advanced pancreatic cancer. Nonetheless, despite the fact that the trial proved the safety of the procedure, pioneers of EUS-FNI argued in a recent paper that a lot of hurdles have to be overcome in order to develop a clinically functional EUS-FNI method, especially in terms of oncolytic viruses[34]. Among recently conducted trials, we have highlighted the following two: (1) Nishimura et al[35] injected a double-stranded RNA oligonucleotide that repressed tumor growth in 6 patients with unresectable pancreatic cancer and reported no adverse effects, as well as reduction in plasma levels of the target molecule and tumor size reduction; and (2) A prospective study by Levy et al[36] tested the use of EUS-FNI for the guided delivery of gemcitabine, a chemotherapeutic whose role in treatment of pancreatic cancer has been well-established. Authors administered gemcitabine in 36 patients with various stages of pancreatic cancer (II-IV), and no adverse events were reported. Overall survival of the patients was 10.4 mo, and more importantly, 20% of patients with stage III unresectable disease were down-staged and underwent an R0 resection. In conclusion, as pancreatic cancer is a systemic disease, it is practically impossible to assume that EUS-FNI will fully replace current therapeutic modalities of pancreatic cancer, but, given the safety and feasibility of the method, as well as expansion of translational medicine, it could emerge as a viable adjuvant method in the future.

Brachytherapy

EUS-guided brachytherapy involves implantation of radioactive seeds directly into or adjacent to the tumor-affected tissue. The target tissue is then exposed to the emission of low-energy gamma, X-rays, or beta particles, leading to localized tissue injury and tumor ablation. The main advantage of brachytherapy is its ability to deliver a markedly higher dose of radiation to the tumor mass in comparison to external beam radiation therapy. In the latter, radiation beams pass through other non-tumorous tissues in reaching the target mass, thus resulting in collateral toxicity and more damage to healthy tissue. Although there is abundant evidence to suggest that brachytherapy can deliver a higher dose of radiation and provide local control as well as palliative benefits, currently no brachytherapy device is approved for the treatment of patients with pancreatic cancer [37-42]. There are multiple approaches for implantation of radioactive seeds, one of which is the EUS-guided brachytherapy, and a variety of chemical elements that could be used in this manner (phosphorus-32 (P-32), iodine, gold, iridium, etc.)[43,44]. Two pilot trials that tested the potential of EUS-guided brachytherapy in patients with pancreatic cancer had rather disappointing results with no benefits with regard to overall survival rate (27%) and partial response rate (13.6%), respectively^[45,46]. Nonetheless, authors of one of the trials argued that such results might be due to an insufficient radiation dose to local lesions[47]. They performed another study in which they implemented a novel computer-aided treatment-planning system (TPS). Under the support of the new TPS, partial remission rate was 80% and expected median survival time of the 42 patients was 9.0 mo (24 patients were in stage IV). At present, there is an ongoing open-label, single-arm pilot study of EUS-guided



brachytherapy with P-32 microparticles in combination with gemcitabine and/or nabpaclitaxel in unresectable locally advanced pancreatic cancer (OncoPaC-1 study)[48]. However, the role of the EUS-guided brachytherapy in pancreatic cancer is yet to be determined. Hopefully, conjunction of brachytherapy and chemotherapeutics will increase the share of patients with pancreatic cancer that convert to resectable and provide us with more durable, local control in comparison to conventional treatments.

EUS-guided celiac plexus neurolysis

Among a multitude of options, the EUS has also found its place in the palliative treatment of patients with pancreatic cancer. Given the fact that most patients present in the advanced stages of the disease, palliative care is often the primary goal of care. Pain, the most significant and most common complication of pancreatic cancer, has traditionally been managed by nonsteroidal and opioid analgesic, often with numerous side effects, including constipation, sedation, nausea, vomiting, and delirium^[49,50]. In order to overcome these issues and while trying to improve quality of life, endoscopic ultrasound-guided celiac plexus neurolysis (EUS-CPN) was introduced by Wierseme and Wierseme in 1996 (Table 1)[51].

EUS-CPN is a minimally invasive method for the treatment of pain through the chemical ablation of the celiac plexus under the control of the EUS. The current National Comprehensive Cancer Network guidelines recommend EUS-CPN for treatment of severe pancreatic cancer-associated pain in the case of failure of achieving adequate analgesia and/or intolerable adverse effects [52].

The EUS-guided neurolysis is indicated for patients with chronic abdominal pain caused by an upper GI tract malignancy, mostly pancreatic cancer. Patients with unresectable disease whose pain affects their quality of life are considered good candidates for this approach. Furthermore, there are pivotal studies of the EUS-CPN efficacy in patients with gallbladder carcinoma[53]. However, besides pancreatic carcinoma, the main indication for EUS-CPN still remains chronic pancreatitis[50,54].

The optimal timing in which the EUS neurolysis should be applied is still unclear. However, in randomized, double-blind, controlled trial, Wyse et al[55] found that early EUS-CPN, performed during the diagnostic EUS, provided better pain management and prevented an increase in morphine consumption than EUS-CPN later in the course of the disease. However, further studies are needed to define the optimal timing for CPN. CPN consists of injecting a neurolytic agent (absolute alcohol or phenol) into the area around the celiac plexus monitored by an echo endoscope with prior administration of a local anesthetic (bupivacaine or lidocaine). Application of the neurolytic agent can be unilateral (just above the celiac trunk) or bilateral (on both sides of the celiac trunk)[56-58].

Several studies have shown pain improvement after CPN[50,51,54,55]. In the initial study, pain improvement was achieved at 2, 4, 8, and 12 wk after CPN in 79%-88% patients[51]. Additionally, a Cochrane Review of six studies (358 patients), that showed that the EUS-CPN was superior at 4 and 8 wk compared to drug-based management, with significant drug consumption reduction^[5]. Whether the application of the EUS-CPN affects survival remains unclear. A study by Fujii-Lau et al [59] indicates that the EUS-guided neurolysis is associated with longer survival than the non-EUS guided approach. However, further studies are needed to assess the potential impact of the neurolysis on survival. In order to improve the efficiency of pain treatment, the following EUS-CPN modifications have been presented: EUSguided direct celiac ganglia neurolysis (EUS-CGN); and EUS-guided broad plexus neurolysis[60-62]. In a randomized study by Doi *et al*[60], authors indicated better pain improvement in CGN over CPN. Levy et al^[61] have successfully demonstrated the modification of the neurolysis of the celiac plexus in the form of direct injection of the agent into the celiac ganglion, requiring prior visualization of the ganglia itself. Sakamoto et al[62] in 2010 described the injection of a neurolytic agent in the area around the origin of the superior mesenteric artery, introducing a method of broad plexus neurolysis, which demonstrated better pain relief compared to the CPN.

A recent review of 20 studies comprising 1142 patients, revealed that complications of EUS-CPN occurred in 21% of 661 EUS-guided CPN interventions[63]. The most frequent complications included diarrhea, transient pain exacerbation, and hypotension. Most of the complications seem to be a consequence of a sympatholytic reaction and are self-limited (< 48 h). However, in 0.2% of cases, major complications were observed, including paraplegia, retroperitoneal abscess, and ischemia with visceral injury^[63].

Given the unequivocal beneficial effects of EUS-CPN in the palliative care of the patients with advance pancreatic cancer, we can expect further increase of its usage in routine clinical practice.



Table 1 Overview of the endoscopic ultrasound-guided celiac plexus neurolysis characteristics					
Procedure	Chemical ablation of the celiac plexus				
Indications	Chronic abdominal pain caused by: Pancreatic cancer; Chronic pancreatitis; Gallbladder carcinoma ¹				
Contraindications[57]	Coagulation disorders; INR > 1.5; Platelet count < $50000/\mu$ L; Retroperitoneal abscess; Disturbed anatomy (difficulties in visualizing the celiac trunk and ganglia); Malformations of the celiac or superior mesenteric artery				
Serious complications (observed in 0.2% of the patients[63])	Paraplegia; Retroperitoneal abscess; Ischemia with visceral injury				
Supporting evidence	Kaufman et al[50]; Wiersema et al[51]; Puli et al[54]; Wyse et al[55]; Arcidiacono et al[58]				

¹Only pivotal studies were so far conducted. INR: International normalized ratio.

EUS IN PANCREATIC CYSTIC NEOPLASMS

Pancreatic cystic neoplasms (PCN) represent a heterogeneous group of pancreatic cysts with significant differences in their pathological and clinical features, the most important one being the difference in malignant potential among subtypes. These differences are important as they determine the approach for both the treatment and the surveillance of PCN[64-66]. A prevalence of all PCN varies from 2%-45% in general population)[67-70], with constant increase in the PCN detection rate. The explanation for the observed increase lies in the improved modalities of non-invasive (computed tomography (CT) scan, magnetic resonance imaging- cholangiopancreatography (MRI-MRCP)) and minimally invasive (endoscopic ultrasound-fine needle aspiration (EUS-FNA)) imaging methods, and their broader use in preventive check-ups, as well as an awareness of true nature of PCN and the necessity for their close follow-up. According to different studies, abdominal ultrasonography detects PCN in 0.21% of individuals [71], CT in 2.6% [72], and MRI-MRCP in 2.4% to 49.1% of tested individuals [65,72-74]. The management of PCN may be quite challenging, with identification of specific PCN type being a crucial step since malignant potential varies significantly between different types of PCN. Timely and correct management of PCN is vital, as it may prevent progression to pancreatic cancer and decrease the need for lifelong follow-up [71].

Due to the challenges in differentiation between the various types of PCN and its implications on therapeutic approach, guidelines on the management of PCN were proposed by expert groups, most notably by the Association of Pancreatology, the American Gastroenterological Association, and by the European Study Group on Cystic Tumors of the Pancreas[65,66,73,75]. Various approaches, including surgical resection, endoscopic techniques, and surveillance, are covered in all of the abovenoted guidelines. Surgical resection is the golden standard for management of premalignant and malignant cystic lesions. Indications for resection depend on the presence of symptoms, probability of malignancy, location of the lesion, and surgical risk of the patient. On the other hand, surgery carries a considerable risk, with perioperative complication rates from 20% to 40%, and mortality rates up to 2% [76-79]. Therefore, endoscopic techniques represent an important alternative to surgery, especially in patients with significant comorbidities or in cases of indeterminate cystic lesions.

The main advantage of the EUS in this setting is the fact that EUS-guided pancreatic cyst ablation using ethanol and/or paclitaxel enables organ preservation, leaving endocrine and exocrine function intact[80-83]. However, there is concern about the use of ethanol, as the rate of reported complications was as high as 2%-10%[84]. Another disadvantage of this method is the inability to obtain a sample for histopathological analysis. The long-term effects of ablation and prevention of malignant alteration are yet to be evaluated in future studies [84-87]. The current diagnostic workup of PCN includes CT or MRI with the addition of MRCP and EUS when appropriate[64-67,88]. EUS is indicated in addition to other imaging modalities if there are worrisome clinical or radiological features present (nodules, dilated pancreatic duct, thickened wall), or if there is a need for obtaining the cystic fluid for cytological and/or biochemical analysis.

The reported accuracy of EUS imaging for differentiating mucinous from nonmucinous PCN is relatively low (48%-94%), with sensitivity of 36%-91%, and specificity of 45%-81% [89-93]. Cytopathological analysis of cystic fluid may reveal dysplasia or clear malignancy. Although cytology is highly specific (83%-100%), it is relatively insensitive (27%-48%), resulting in relatively low diagnostic accuracy of this



procedure (8%-59%)[89-92,94]. However, sensitivity could be increased by an additional puncture of the cystic wall. Additional biochemical markers, including carcinoembryonic antigen (CEA), which has been proved to be useful in distinction between mucinous and non-mucinous PCN, and amylase, which strongly suggests a connection between the cyst and the pancreatic ductal system, may also be obtained from cystic fluid[95,96]. Combination of multiple EUS-guided methods, such as EUS morphology, cytology, and cyst fluid CEA, provide us with greater accuracy in detecting malignant PCN than any of the methods individually[97,98]. Recently, DNA testing of pancreatic cyst fluid emerged as a promising additional tool for the differentiation between mucinous and non-mucinous PCN, between mucinous PCN subtypes, and between premalignant PCN and advanced neoplasia[99].

According to the latest recommendations, surgically fit patients with asymptomatic cysts that are presumed to be premalignant (intraductal papillary mucinous neoplasms (IPMN) or mucinous cystic neoplasms(MCN)), but which possess no concerning features, should be monitored, preferably using MRI-MRCP or EUS if MRCP is not available[100,101]. In cases in which any of the worrisome features emerges, the EUS-FNA should be used in the cyst follow-up. For both IPMN and MCN, surveillance should continue as long as the patient is fit for surgery[102]. On the other hand, current guidelines do not address the need for surveillance in asymptomatic patients with serous cystic neoplasms (SCN), since malignant progression of SCN is very rare[102-104].

EUS-GUIDED PORTAL VEIN INTERVENTIONS

The portal vein (PV) can be accessed *via* trans-gastric or trans-duodenal access under the EUS guidance using an 18G-25G needle with low risk of complications. Multiple studies have demonstrated that EUS is successful in sampling of the PV to reveal circulating tumor cells, as well as obtaining the portal vein thrombus specimen[105], which has an utmost significance in the staging of hepatocellular liver cancer (HCC) [106-108]. The diagnostic yield of the EUS is confirmed by the reported cases of HCC detection using the EUS-FNA of radiologically suspected malignant thrombi without an evident liver mass[108,109].

As a significant step forward, Park *et al*[110] have even demonstrated the technical feasibility of the EUS guided transhepatic PV stenting in porcine models, without any immediate or late complications[110]. However, patients who undergo PV stenting due to malignant thrombosis or stenosis may bear certain procedural risk factors, such as coagulopathy and risk of rapid clinical deterioration if biliary leak or bacteremia occurs.

The EUS-guided PV injection of chemotherapy (EPIC) is another novel therapeutic possibility with a few significant advantages in comparison to the current methods [106]. EPIC uses drug eluting microbeads that eventually lodges in hepatic sinusoids and results in the prolonged hepatic drug exposure[111]. Studies suggest that EPIC achieves appropriate intrahepatic drug levels, while simultaneously bypassing the systemic side effects and avoiding the ischemic bile duct injury that occurs during the transarterial approach[106]. Faigel *et al*[111] compared the EPIC administration of irinotecan loaded liquid chromatography beads with the systemic unloaded irinotecan application in animal models and revealed EPIC-associated higher irinotecan intrahepatic concentration, as well as lower plasma, bone marrow, and skeletal muscle drug concentrations. Two years later, the same group confirmed their findings on a greater number of animal models using irinotecan, doxorubicin, and albumin-bound paclitaxel nanoparticles[112].

Preoperative selective embolization of the PV branch that feeds the tumor-affected liver lobe has been utilized in clinical practice since 1986 using the percutaneous transhepatic approach[106,113]. Embolization leads to the atrophy of the involved liver segment and hypertrophy of the remnant liver parenchyma, thus preventing the postoperative liver failure[113]. Recently, Park *et al*[113] used nine porcine models to successfully prove the efficacy and safety of the EUS guided embolization of the PV using coil and cyanoacrylate. Furthermore, Matthes *et al*[114] demonstrated efficient selective PV embolization using ethylene-vinyl alcohol copolymer, known as EVAL (Enteryx), in an animal model. Unfortunately, to our knowledge, studies including PV interventions in anticancer treatment have been so far limited to animal models. Still, exciting advances in the field are revealed, and prospective studies involving humans are eagerly awaited.

Zaishideng® WJGO | https://www.wjgnet.com

ROLE OF THE EUS IN LIVER TUMOR MANAGEMENT

EUS-guided tumor ablation is a safe and effective treatment modality for tumor lesions of the caudate and left liver lobe. Multiple studies have successfully demonstrated the benefits of ethanol administration via EUS-FNI in the treatment of both HCC and liver metastases[115]. Carrara et al[116] described the EUS guided transgastric bipolar hybrid cryotherm ablation on a porcine model without complications. Varadarajulu et al[117] reported RFA in animal models with effective coagulation necrosis of large areas and without damage to the surrounding liver parenchyma. Multiple studies have also demonstrated success of the EUS-guided neodymiumdoped yttrium aluminum garnet (Nd:YAG) laser ablation in patients with HCC and colorectal cancer metastases[118,119]. In addition, several other methods, including injection of sclerosants and chemotherapeutics, represent viable future therapeutic options[115].

In the management of the HCC-related complications, one of the most common and disastrous is the variceal bleeding incident. With respect to secondary prevention of this complication in patients with inoperable HCC, Tang et al[120] used EUS guided cyanoacrilate injection, which led to reduced rebleeding rates, as well as improved variceal bleeding free survival.

ROLE OF EUS IN ENDOSCOPIC SUBMUCOSAL DISSECTION

The development of gastrointestinal endoscopic tissue resection techniques demands precise diagnostic tools in the preoperative evaluation. Accurate information about the depth of tumor invasion of the gastrointestinal wall and the nodal involvement are necessary for determining the appropriate intervention.

Endoscopic submucosal dissection (ESD), the newest and most invasive method, has become standard of care of precancerous and some early cancer lesions in gastrointestinal tract, allowing curative resection of the lesions[121]. Depending on the proximity of the GI tract wall during the procedure, the EUS enables clear image of the lesion depth and vital surrounding structures, especially of the lymph nodes[121].

Endoscopic resection is indicated in early esophageal cancer with minimal or no lymph node invasion[122]. According to the latest guidelines for the treatment of esophageal cancer of the Japan Esophageal Society, the absolute indication for endoscopic resection is defined as flat lesion (Paris 0-II), with m1 (intraepithelial) - m2 (invading lamina propria) invasion, and circumferential extent of $\leq 2/3$ [123]. A systematic review and meta-analysis has demonstrated that sensitivity and specificity for T1a staging were 85% and 87%, respectively, and 86% for both sensitivity and specificity for T1b staging[124]. However, the ability of the EUS to predict endoscopic resectability by discrimination between T1 and T2 lesions is still intensively studied. Available data suggests that 15%-25% of cases are under-staged compared with endoscopic mucosal resection staging, while about 4%-12% of cases are over-staged [125,126].

Conventional EUS has limited accuracy in the detection of submucosal invasion in early esophageal cancer [127,128]. It remains questionable whether the EUS should be routinely performed prior to ESD of esophageal superficial lesions. European Society of Gastrointestinal Endoscopy (ESGE) suggests that EUS should be considered in esophageal superficial carcinomas with suspicious features for submucosal invasion or lymph node metastasis[122].

Esophageal submucosal tumors are becoming more common indication for ESD. EUS allows for the evaluation of size, echo pattern, layer of origin, and eventual surrounding nodal involvement[121,129]. The biggest setback in EUS evaluation is still interobserver variation. Notwithstanding, endoscopic ultrasonography has become the most valuable tool in diagnostics of esophageal submucosal tumors^[129].

The role of EUS in establishing the feasibility of endoscopic resection of superficial gastric lesions is still controversial. ESD is indicated as the treatment of choice for most gastric superficial neoplastic lesions, including low- or high-grade non-invasive neoplasia and adenocarcinoma with no evidence of deep submucosal invasion[130-133]. Although EUS is considered a reliable method for locoregional staging, endoscopic evaluation is still favored over EUS for predicting endoscopic resectability [134]. We should also highlight different approaches in the use of EUS before intervention. Although favored in the prior planning of endoscopic resection in Western countries, in the Eastern countries (in which the incidence of gastric cancer is notably higher), it is not considered necessary to perform EUS in the preoperative

evaluation prior to the planned intervention [131,135,136].

Since ESD has been achieving similar results compared with surgery in treatment of gastric submucosal tumors (< 50 mm in size), the role of the EUS in preoperative management has recently evolved[137]. Today, it is the main tool in preoperative assessment, including evaluation of size, layer of origin, and echo pattern. Furthermore, the use of EUS has extended to marking the lesion with EUS-assisted injection into the muscularis propria, providing a deeper safety cushion for submucosal dissection procedure[138].

A randomized study by Fernandez-Esparrach et al [139] concluded that EUS and MRI have similar accuracy in T and N staging for rectal cancer. The ESGE recommends using one of these methods for staging of rectal cancer, but not for colon cancer^[122]. However, the role of EUS and MRI for superficial lesions has been undefined.

A prospective study comparing high frequency EUS vs magnifying chromoendoscopy in early colorectal neoplasia showed that high frequency EUS was superior to chromoendoscopy in determining the depth of invasion, showing an accuracy of 93% vs 59% [140]. Nevertheless, endoscopic resection will probably remain the best staging tool for superficial rectal lesions, and if the endoscopist feels the lesion is endoscopically resectable, it will probably not require preoperative EUS[122]. On the other hand, according to the ESGE recommendations, the use of EUS or MRI should be considered for rectal lesions with endoscopic features suspected for submucosal invasion, since the finding of suspicious lymph nodes could be an indication for neoadjuvant treatment[122].

To summarize, the role of EUS in ESD is still evolving. The main goal of the endoscopic ultrasonography remains to evaluate a potential submucosal invasion and locoregional staging of the disease. Future research on the role of EUS in ESD should be concentrated in reduction of interobserver variations and alleviating possible complications.

CONCLUSION

Even though recent technological advancements in endoscopic approaches led to an improvement of outcomes of tumors in the abdominal cavity, as indicated by reduced mortality, morbidity, and palliative care, we are still far from having an optimal treatment, particularly for pancreatic cancer. Nonetheless, the novel EUS-guided approaches (RFA, brachytherapy, FNI, etc.) for pancreatic cancer might be crucial in overcoming the problem of drug distribution to tumorous tissue and reducing the required therapeutic doses and incidence of adverse effects. In addition, alleviating extreme pain that many patients with pancreatic cancer endure seems to be achievable through celiac plexus neurolysis. Furthermore, EUS is already a part of the algorithm in management of PCN, where EUS-FNA is the method of choice in case of appearance of certain alarming features of cysts.

The utility of EUS in PV interventions is most prominent in relation to the staging of HCC. The role of EUS in ESD with respect to management of precancerous and some early cancer lesions in GI tract, as well as EUS-guided treatment of various hepatic cancers, is yet to be determined, as current data is insufficient to recommend these techniques as standards of care.

REFERENCES

- Yan BM, Van Dam J. Endoscopic ultrasound-guided intratumoral therapy for pancreatic cancer. 1 Can J Gastroenterol 2008; 22: 405-410 [PMID: 18414717 DOI: 10.1155/2008/104398]
- Cazacu IM, Singh BS, Saftoiu A, Bhutani MS. Endoscopic Ultrasound-Guided Treatment of Pancreatic Cancer. Curr Gastroenterol Rep 2020; 22: 27 [PMID: 32350629 DOI: 10.1007/s11894-020-00767-1]
- 3 Aguirre AJ, Collisson EA. Advances in the Genetics and Biology of Pancreatic Cancer. Cancer J 2017; 23: 315-320 [PMID: 29189326 DOI: 10.1097/PPO.000000000000286]
- Hajatdoost L, Sedaghat K, Walker EJ, Thomas J, Kosari S. Chemotherapy in Pancreatic Cancer: A Systematic Review. Medicina (Kaunas) 2018; 54 [PMID: 30344279 DOI: 10.3390/medicina54030048
- 5 LeVeen R. Laser hyperthermia and radiofrequency ablation of hepatic lesions. Semin Interv Radiol 1997; 205: 313-324
- Haen SP, Pereira PL, Salih HR, Rammensee HG, Gouttefangeas C. More than just tumor



destruction: immunomodulation by thermal ablation of cancer. Clin Dev Immunol 2011; 2011: 160250 [PMID: 22242035 DOI: 10.1155/2011/160250]

- 7 Waitz R, Solomon SB. Can local radiofrequency ablation of tumors generate systemic immunity against metastatic disease? Radiology 2009; 251: 1-2 [PMID: 19332838 DOI: 10.1148/radiol.2511082215]
- 8 Teng LS, Jin KT, Han N, Cao J. Radiofrequency ablation, heat shock protein 70 and potential antitumor immunity in hepatic and pancreatic cancers: a minireview. Hepatobiliary Pancreat Dis Int 2010; 9: 361-365 [PMID: 20688598 DOI: 10.1016/S0006-3223(01)01255-0]
- Giardino A, Innamorati G, Ugel S, Perbellini O, Girelli R, Frigerio I, Regi P, Scopelliti F, Butturini G, Paiella S, Bacchion M, Bassi C. Immunomodulation after radiofrequency ablation of locally advanced pancreatic cancer by monitoring the immune response in 10 patients. Pancreatology 2017; 17: 962-966 [PMID: 29037917 DOI: 10.1016/j.pan.2017.09.008]
- 10 European Association for the Study of the Liver. EASL Clinical Practice Guidelines: Management of hepatocellular carcinoma. J Hepatol 2018; 69: 182-236 [PMID: 29628281 DOI: 10.1016/j.jhep.2018.03.019]
- Goldberg SN, Mallery S, Gazelle GS, Brugge WR. EUS-guided radiofrequency ablation in the 11 pancreas: results in a porcine model. Gastrointest Endosc 1999; 50: 392-401 [PMID: 10462663 DOI: 10.1053/ge.1999.v50.98847]
- 12 Kim HJ, Seo DW, Hassanuddin A, Kim SH, Chae HJ, Jang JW, Park DH, Lee SS, Lee SK, Kim MH. EUS-guided radiofrequency ablation of the porcine pancreas. Gastrointest Endosc 2012; 76: 1039-1043 [PMID: 23078928 DOI: 10.1016/j.gie.2012.07.015]
- Lee JM, Han JK, Kim HC, Choi YH, Kim SH, Choi JY, Choi BI. Switching monopolar 13 radiofrequency ablation technique using multiple, internally cooled electrodes and a multichannel generator: ex vivo and in vivo pilot study. Invest Radiol 2007; 42: 163-171 [PMID: 17287646 DOI: 10.1097/01.rli.0000252495.44818.b3]
- 14 Gaidhane M, Smith I, Ellen K, Gatesman J, Habib N, Foley P, Moskaluk C, Kahaleh M. Endoscopic Ultrasound-Guided Radiofrequency Ablation (EUS-RFA) of the Pancreas in a Porcine Model. Gastroenterol Res Pract 2012; 2012: 431451 [PMID: 23049547 DOI: 10.1155/2012/431451]
- 15 Wu Y, Tang Z, Fang H, Gao S, Chen J, Wang Y, Yan H. High operative risk of cool-tip radiofrequency ablation for unresectable pancreatic head cancer. J Surg Oncol 2006; 94: 392-395 [PMID: 16967436 DOI: 10.1002/jso.20580]
- 16 Spiliotis JD, Datsis AC, Michalopoulos NV, Kekelos SP, Vaxevanidou A, Rogdakis AG, Christopoulou AN. Radiofrequency ablation combined with palliative surgery may prolong survival of patients with advanced cancer of the pancreas. Langenbecks Arch Surg 2007; 392: 55-60 [PMID: 17089173 DOI: 10.1007/s00423-006-0098-5]
- Elias D, Baton O, Sideris L, Lasser P, Pocard M. Necrotizing pancreatitis after radiofrequency 17 destruction of pancreatic tumors. Eur J Surg Oncol 2004; 30: 85-87 [PMID: 14736529 DOI: 10.1016/j.ejso.2003.10.013]
- 18 Girelli R, Frigerio I, Giardino A, Regi P, Gobbo S, Malleo G, Salvia R, Bassi C. Results of 100 pancreatic radiofrequency ablations in the context of a multimodal strategy for stage III ductal adenocarcinoma. Langenbecks Arch Surg 2013; 398: 63-69 [PMID: 23053459 DOI: 10.1007/s00423-012-1011-z]
- 19 Paiella S, Salvia R, Ramera M, Girelli R, Frigerio I, Giardino A, Allegrini V, Bassi C. Local Ablative Strategies for Ductal Pancreatic Cancer (Radiofrequency Ablation, Irreversible Electroporation): A Review. Gastroenterol Res Pract 2016; 2016: 4508376 [PMID: 26981115 DOI: 10.1155/2016/4508376]
- 20 Varshney S, Sewkani A, Sharma S, Kapoor S, Naik S, Sharma A, Patel K. Radiofrequency ablation of unresectable pancreatic carcinoma: feasibility, efficacy and safety. JOP 2006; 7: 74-78 [PMID: 16407624]
- 21 Siriwardena AK. Radiofrequency ablation for locally advanced cancer of the pancreas. JOP 2006; 7: 1-4 [PMID: 16407612 DOI: 10.1016/S0344-0338(97)80076-3]
- 22 Casadei R, Ricci C, Pezzilli R, Serra C, Calculli L, Morselli-Labate AM, Santini D, Minni F. A prospective study on radiofrequency ablation locally advanced pancreatic cancer. Hepatobiliary Pancreat Dis Int 2010; 9: 306-311 [PMID: 20525559]
- Giardino A, Girelli R, Frigerio I, Regi P, Cantore M, Alessandra A, Lusenti A, Salvia R, Bassi C, 23 Pederzoli P. Triple approach strategy for patients with locally advanced pancreatic carcinoma. HPB (Oxford) 2013; 15: 623-627 [PMID: 23458679 DOI: 10.1111/hpb.12027]
- 24 Tang Z, Wu YL, Fang HQ, Xu J, Mo GQ, Chen XM, Gao SL, Li JT, Liu YB, Wang Y. [Treatment of unresectable pancreatic carcinoma by radiofrequency ablation with 'cool-tip needle': report of 18 cases]. Zhonghua Yi Xue Za Zhi 2008; 88: 391-394 [PMID: 18581892]
- 25 Matsui Y, Nakagawa A, Kamiyama Y, Yamamoto K, Kubo N, Nakase Y. Selective thermocoagulation of unresectable pancreatic cancers by using radiofrequency capacitive heating. Pancreas 2000; 20: 14-20 [PMID: 10630378 DOI: 10.1097/00006676-200001000-00002]
- Cantore M, Girelli R, Mambrini A, Frigerio I, Boz G, Salvia R, Giardino A, Orlandi M, Auriemma 26 A, Bassi C. Combined modality treatment for patients with locally advanced pancreatic adenocarcinoma. Br J Surg 2012; 99: 1083-1088 [PMID: 22648697 DOI: 10.1002/bjs.8789]
- Song TJ, Seo DW, Lakhtakia S, Reddy N, Oh DW, Park DH, Lee SS, Lee SK, Kim MH. Initial 27 experience of EUS-guided radiofrequency ablation of unresectable pancreatic cancer. Gastrointest



Endosc 2016; 83: 440-443 [PMID: 26344883 DOI: 10.1016/j.gie.2015.08.048]

- 28 Dhaliwal A, Kolli S, Dhindsa BS, Choa J, Mashiana HS, Ramai D, Chandan S, Bhogal N, Sayles H, Bhat I, Singh S, Adler DG. Efficacy of EUS-RFA in pancreatic tumors: Is it ready for prime time? Endosc Int Open 2020; 8: E1243-E1251 [PMID: 33015325 DOI: 10.1055/a-1221-5012]
- 29 DiMaio CJ, Nagula S, Goodman KA, Ho AY, Markowitz AJ, Schattner MA, Gerdes H. EUSguided fiducial placement for image-guided radiation therapy in GI malignancies by using a 22gauge needle (with videos). Gastrointest Endosc 2010; 71: 1204-1210 [PMID: 20598247 DOI: 10.1016/j.gie.2010.01.003]
- 30 Lennon AM, Newman N, Makary MA, Edil BH, Shin EJ, Khashab MA, Hruban RH, Wolfgang CL, Schulick RD, Giday S, Canto MI. EUS-guided tattooing before laparoscopic distal pancreatic resection (with video). Gastrointest Endosc 2010; 72: 1089-1094 [PMID: 21034909 DOI: 10.1016/j.gie.2010.07.023
- 31 Chang KJ, Nguyen PT, Thompson JA, Kurosaki TT, Casey LR, Leung EC, Granger GA. Phase I clinical trial of allogeneic mixed lymphocyte culture (cytoimplant) delivered by endoscopic ultrasound-guided fine-needle injection in patients with advanced pancreatic carcinoma. Cancer 2000; 88: 1325-1335 [PMID: 10717613 DOI: 10.1002/(sici)1097-0142(20000315)88:6<1325::aid-cncr8>3.0.co;2-t]
- 32 Herman JM, Wild AT, Wang H, Tran PT, Chang KJ, Taylor GE, Donehower RC, Pawlik TM, Ziegler MA, Cai H, Savage DT, Canto MI, Klapman J, Reid T, Shah RJ, Hoffe SE, Rosemurgy A, Wolfgang CL, Laheru DA. Randomized phase III multi-institutional study of TNFerade biologic with fluorouracil and radiotherapy for locally advanced pancreatic cancer: final results. J Clin Oncol 2013; 31: 886-894 [PMID: 23341531 DOI: 10.1200/JCO.2012.44.7516]
- 33 Lee JC, Shin DW, Park H, Kim J, Youn Y, Kim JH, Hwang JH. Tolerability and safety of EUSinjected adenovirus-mediated double-suicide gene therapy with chemotherapy in locally advanced pancreatic cancer: a phase 1 trial. Gastrointest Endosc 2020; 92: 1044-1052.e1 [PMID: 32084409 DOI: 10.1016/j.gie.2020.02.012]
- 34 Nakai Y, Chang KJ. EUS-guided fine-needle injection for pancreatic cancer: back to the future. Gastrointest Endosc 2020; 92: 1053-1054 [PMID: 33160487 DOI: 10.1016/j.gie.2020.06.025]
- 35 Nishimura M, Matsukawa M, Fujii Y, Matsuda Y, Arai T, Ochiai Y, Itoi T, Yahagi N. Effects of EUS-guided intratumoral injection of oligonucleotide STNM01 on tumor growth, histology, and overall survival in patients with unresectable pancreatic cancer. Gastrointest Endosc 2018; 87: 1126-1131 [PMID: 29122598 DOI: 10.1016/j.gie.2017.10.030]
- Levy MJ, Alberts SR, Bamlet WR, Burch PA, Farnell MB, Gleeson FC, Haddock MG, Kendrick 36 ML, Oberg AL, Petersen GM, Takahashi N, Chari ST. EUS-guided fine-needle injection of gemcitabine for locally advanced and metastatic pancreatic cancer. Gastrointest Endosc 2017; 86: 161-169 [PMID: 27889543 DOI: 10.1016/j.gie.2016.11.014]
- 37 Handley WS. PANCREATIC CANCER AND ITS TREATMENT BY IMPLANTED RADIUM. Ann Surg 1934; 100: 215-223 [PMID: 17856333 DOI: 10.1097/00000658-193407000-00021]
- Shipley WU, Nardi GL, Cohen AM, Ling CC. Iodine-125 implant and external beam irradiation in 38 patients with localized pancreatic carcinoma: a comparative study to surgical resection. Cancer 1980; 45: 709-714 [PMID: 6244074 DOI: 10.1002/1097-0142(19800215)45:4<709::aid-cncr2820450416>3.0.co;2-5]
- Westlin JE, Andersson-Forsman C, Garske U, Linné T, Aas M, Glimelius B, Lindgren PG, Order 39 SE, Nilsson S. Objective responses after fractionated infusional brachytherapy of unresectable pancreatic adenocarcinomas. Cancer 1997; 80: 2743-2748 [PMID: 9406733 DOI: 10.1002/(sici)1097-0142(19971215)80:12+<2743::aid-cncr54>3.3.co;2-4]
- 40 DeNittis AS, Stambaugh MD, Lang P, Wallner PE, Lustig RA, Dillman RO, Order SE. Complete remission of nonresectable pancreatic cancer after infusional colloidal phosphorus-32 brachytherapy, external beam radiation therapy, and 5-fluorouracil: a preliminary report. Am J Clin Oncol 1999; 22: 355-360 [PMID: 10440189 DOI: 10.1097/00000421-199908000-00006]
- 41 Mutignani M, Shah SK, Morganti AG, Perri V, Macchia G, Costamagna G. Treatment of unresectable pancreatic carcinoma by intraluminal brachytherapy in the duct of Wirsung. Endoscopy 2002; 34: 555-559 [PMID: 12170409 DOI: 10.1055/s-2002-33214]
- Peretz T, Nori D, Hilaris B, Manolatos S, Linares L, Harrison L, Anderson LL, Fuks Z, Brennan 42 MF. Treatment of primary unresectable carcinoma of the pancreas with I-125 implantation. Int J Radiat Oncol Biol Phys 1989; 17: 931-935 [PMID: 2808054 DOI: 10.1016/0360-3016(89)90138-7]
- 43 Rosemurgy A, Luzardo G, Cooper J, Bowers C, Zervos E, Bloomston M, Al-Saadi S, Carroll R, Chheda H, Carey L, Goldin S, Grundy S, Kudryk B, Zwiebel B, Black T, Briggs J, Chervenick P. 32P as an adjunct to standard therapy for locally advanced unresectable pancreatic cancer: a randomized trial. J Gastrointest Surg 2008; 12: 682-688 [PMID: 18266048 DOI: 10.1007/s11605-007-0430-6]
- 44 Goertz SR, Ali MM, Parker GA. Local management of pancreatic carcinoma: iodine-125 implantation. Clin Oncol (R Coll Radiol) 1990; 2: 22-26 [PMID: 1702011 DOI: 10.1016/s0936-6555(05)80214-6]
- Sun S, Xu H, Xin J, Liu J, Guo Q, Li S. Endoscopic ultrasound-guided interstitial brachytherapy of 45 unresectable pancreatic cancer: results of a pilot trial. Endoscopy 2006; 38: 399-403 [PMID: 16680642 DOI: 10.1055/s-2006-925253]
- 46 Jin Z, Du Y, Li Z, Jiang Y, Chen J, Liu Y. Endoscopic ultrasonography-guided interstitial implantation of iodine 125-seeds combined with chemotherapy in the treatment of unresectable



pancreatic carcinoma: a prospective pilot study. Endoscopy 2008; 40: 314-320 [PMID: 18283622 DOI: 10.1055/s-2007-995476]

- 47 Sun X, Lu Z, Wu Y, Min M, Bi Y, Shen W, Xu Y, Li Z, Jin Z, Liu Y. An endoscopic ultrasonography-guided interstitial brachytherapy based special treatment-planning system for unresectable pancreatic cancer. Oncotarget 2017; 8: 79099-79110 [PMID: 29108290 DOI: 10.18632/oncotarget.15763
- 48 Bhutani MS, Klapman JB, Tuli R, El-Haddad G, Hoffe S, Wong FCL, Chasen B, Fogelman DR, Lo SK, Nissen NN, Hendifar AE, Varadhachary G, Katz MHG, Erwin WD, Koay EJ, Tamm EP, Singh BS, Mehta R, Wolff RA, Soman A, Cazacu IM, Herman JM. An open-label, single-arm pilot study of EUS-guided brachytherapy with phosphorus-32 microparticles in combination with gemcitabine +/- nab-paclitaxel in unresectable locally advanced pancreatic cancer (OncoPaC-1): Technical details and study protocol. Endosc Ultrasound 2020; 9: 24-30 [PMID: 31670288 DOI: 10.4103/eus.eus_44_19]
- 49 World Health Organization. Cancer Pain Relief, 2nd ed. WHO: Geneva, Switzerland, 2006
- 50 Kaufman M, Singh G, Das S, Concha-Parra R, Erber J, Micames C, Gress F. Efficacy of endoscopic ultrasound-guided celiac plexus block and celiac plexus neurolysis for managing abdominal pain associated with chronic pancreatitis and pancreatic cancer. J Clin Gastroenterol 2010; 44: 127-134 [PMID: 19826273 DOI: 10.1097/MCG.0b013e3181bb854d]
- Wiersema MJ, Wiersema LM. Endosonography-guided celiac plexus neurolysis. Gastrointest 51 Endosc 1996; 44: 656-662 [PMID: 8979053 DOI: 10.1016/s0016-5107(96)70047-0]
- 52 NCCN Guidelines for Adult Cancer Pain. Version 1. 2020. [cited 10 February 2021] Available from: https://www.nccn.org/professionals/physician_gls/pdf/pain.pdf
- 53 Rai P, Cr L, Kc H. Endoscopic ultrasound-guided celiac plexus neurolysis improves pain in gallbladder cancer. Indian J Gastroenterol 2020; 39: 171-175 [PMID: 32065352 DOI: 10.1007/s12664-019-01003-z]
- 54 Puli SR, Reddy JB, Bechtold ML, Antillon MR, Brugge WR. EUS-guided celiac plexus neurolysis for pain due to chronic pancreatitis or pancreatic cancer pain: a meta-analysis and systematic review. Dig Dis Sci 2009; 54: 2330-2337 [PMID: 19137428 DOI: 10.1007/s10620-008-0651-x]
- 55 Wyse JM, Carone M, Paquin SC, Usatii M, Sahai AV. Randomized, double-blind, controlled trial of early endoscopic ultrasound-guided celiac plexus neurolysis to prevent pain progression in patients with newly diagnosed, painful, inoperable pancreatic cancer. J Clin Oncol 2011; 29: 3541-3546 [PMID: 21844506 DOI: 10.1200/JCO.2010.32.2750]
- Sahai AV, Lemelin V, Lam E, Paquin SC. Central vs. bilateral endoscopic ultrasound-guided celiac 56 plexus block or neurolysis: a comparative study of short-term effectiveness. Am J Gastroenterol 2009; 104: 326-329 [PMID: 19174816 DOI: 10.1038/ajg.2008.64]
- Penman ID. State of the art: putting EUS-guided block/neurolysis into perspective. Gastrointest 57 Endosc 2009; 69: S174-S175 [PMID: 19179151 DOI: 10.1016/j.gie.2008.12.023]
- 58 Arcidiacono PG, Calori G, Carrara S, McNicol ED, Testoni PA. Celiac plexus block for pancreatic cancer pain in adults. Cochrane Database Syst Rev 2011; CD007519 [PMID: 21412903 DOI: 10.1002/14651858.CD007519.pub2
- Fujii-Lau LL, Bamlet WR, Eldrige JS, Chari ST, Gleeson FC, Abu Dayyeh BK, Clain JE, Pearson 59 RK, Petersen BT, Rajan E, Topazian MD, Vege SS, Wang KK, Wiersema MJ, Levy MJ. Impact of celiac neurolysis on survival in patients with pancreatic cancer. Gastrointest Endosc 2015; 82: 46-56.e2 [PMID: 25800661 DOI: 10.1016/j.gie.2014.12.036]
- 60 Doi S, Yasuda I, Kawakami H, Hayashi T, Hisai H, Irisawa A, Mukai T, Katanuma A, Kubota K, Ohnishi T, Ryozawa S, Hara K, Itoi T, Hanada K, Yamao K. Endoscopic ultrasound-guided celiac ganglia neurolysis vs. celiac plexus neurolysis: a randomized multicenter trial. Endoscopy 2013; 45: 362-369 [PMID: 23616126 DOI: 10.1055/s-0032-1326225]
- Levy MJ, Topazian MD, Wiersema MJ, Clain JE, Rajan E, Wang KK, de la Mora JG, Gleeson FC, 61 Pearson RK, Pelaez MC, Petersen BT, Vege SS, Chari ST. Initial evaluation of the efficacy and safety of endoscopic ultrasound-guided direct Ganglia neurolysis and block. Am J Gastroenterol 2008; 103: 98-103 [PMID: 17970834 DOI: 10.1111/j.1572-0241.2007.01607.x]
- Sakamoto H, Kitano M, Kamata K, Komaki T, Imai H, Chikugo T, Takeyama Y, Kudo M. EUS-62 guided broad plexus neurolysis over the superior mesenteric artery using a 25-gauge needle. Am J Gastroenterol 2010; 105: 2599-2606 [PMID: 20823834 DOI: 10.1038/ajg.2010.339]
- Alvarez-Sánchez MV, Jenssen C, Faiss S, Napoléon B. Interventional endoscopic ultrasonography: 63 an overview of safety and complications. Surg Endosc 2014; 28: 712-734 [PMID: 24196551 DOI: 10.1007/s00464-013-3260-5]
- 64 Del Chiaro M, Verbeke C, Salvia R, Klöppel G, Werner J, McKay C, Friess H, Manfredi R, Van Cutsem E, Löhr M, Segersvärd R; European Study Group on Cystic Tumors of the Pancreas. European experts consensus statement on cystic tumors of the pancreas. Dig Liver Dis 2013; 45: 703-711 [PMID: 23415799 DOI: 10.1016/j.dld.2013.01.010]
- Tanaka M, Fernández-Del Castillo C, Kamisawa T, Jang JY, Levy P, Ohtsuka T, Salvia R, Shimizu Y, Tada M, Wolfgang CL. Revisions of international consensus Fukuoka guidelines for the management of IPMN of the pancreas. Pancreatology 2017; 17: 738-753 [PMID: 28735806 DOI: 10.1016/j.pan.2017.07.007
- 66 European Study Group on Cystic Tumors of the Pancreas. European evidence-based guidelines on pancreatic cystic neoplasms. Gut 2018; 67: 789-804 [PMID: 29574408 DOI: 10.1136/gutjnl-2018-316027]



- 67 Ip IK, Mortele KJ, Prevedello LM, Khorasani R. Focal cystic pancreatic lesions: assessing variation in radiologists' management recommendations. Radiology 2011; 259: 136-141 [PMID: 21292867 DOI: 10.1148/radiol.10100970]
- 68 Girometti R, Intini S, Brondani G, Como G, Londero F, Bresadola F, Zuiani C, Bazzocchi M. Incidental pancreatic cysts on 3D turbo spin echo magnetic resonance cholangiopancreatography: prevalence and relation with clinical and imaging features. Abdom Imaging 2011; 36: 196-205 [PMID: 20473669 DOI: 10.1007/s00261-010-9618-4]
- Chang YR, Park JK, Jang JY, Kwon W, Yoon JH, Kim SW. Incidental pancreatic cystic neoplasms 69 in an asymptomatic healthy population of 21,745 individuals: Large-scale, single-center cohort study. Medicine (Baltimore) 2016; 95: e5535 [PMID: 28002329 DOI: 10.1097/MD.00000000005535
- de Jong K, Nio CY, Hermans JJ, Dijkgraaf MG, Gouma DJ, van Eijck CH, van Heel E, Klass G, 70 Fockens P, Bruno MJ. High prevalence of pancreatic cysts detected by screening magnetic resonance imaging examinations. Clin Gastroenterol Hepatol 2010; 8: 806-811 [PMID: 20621679 DOI: 10.1016/j.cgh.2010.05.017]
- Del Chiaro M, Segersvärd R, Lohr M, Verbeke C. Early detection and prevention of pancreatic 71 cancer: is it really possible today? World J Gastroenterol 2014; 20: 12118-12131 [PMID: 25232247 DOI: 10.3748/wjg.v20.i34.12118]
- Del Chiaro M, Segersvärd R, Pozzi Mucelli R, Rangelova E, Kartalis N, Ansorge C, Arnelo U, 72 Blomberg J, Löhr M, Verbeke C. Comparison of preoperative conference-based diagnosis with histology of cystic tumors of the pancreas. Ann Surg Oncol 2014; 21: 1539-1544 [PMID: 24385209 DOI: 10.1245/s10434-013-3465-91
- 73 Vege SS, Ziring B, Jain R, Moavyedi P; Clinical Guidelines Committee; American Gastroenterology Association. American gastroenterological association institute guideline on the diagnosis and management of asymptomatic neoplastic pancreatic cysts. Gastroenterology 2015; 148: 819-22; quize12 [PMID: 25805375 DOI: 10.1053/j.gastro.2015.01.015]
- 74 Jaeschke R, Guyatt GH, Dellinger P, Schünemann H, Levy MM, Kunz R, Norris S, Bion J; GRADE Working Group. Use of GRADE grid to reach decisions on clinical practice guidelines when consensus is elusive. BMJ 2008; 337: a744 [PMID: 18669566 DOI: 10.1136/bmj.a744]
- 75 van Huijgevoort NCM, Del Chiaro M, Wolfgang CL, van Hooft JE, Besselink MG. Diagnosis and management of pancreatic cystic neoplasms: current evidence and guidelines. Nat Rev Gastroenterol Hepatol 2019; 16: 676-689 [PMID: 31527862 DOI: 10.1038/s41575-019-0195-x]
- 76 Allen PJ, D'Angelica M, Gonen M, Jaques DP, Coit DG, Jarnagin WR, DeMatteo R, Fong Y, Blumgart LH, Brennan MF. A selective approach to the resection of cystic lesions of the pancreas: results from 539 consecutive patients. Ann Surg 2006; 244: 572-582 [PMID: 16998366 DOI: 10.1097/01.sla.0000237652.84466.54
- 77 Kiely JM, Nakeeb A, Komorowski RA, Wilson SD, Pitt HA. Cystic pancreatic neoplasms: enucleate or resect? J Gastrointest Surg 2003; 7: 890-897 [PMID: 14592663 DOI: 10.1007/s11605-003-0035-7
- Horvath KD, Chabot JA. An aggressive resectional approach to cystic neoplasms of the pancreas. 78 Am J Surg 1999; 178: 269-274 [PMID: 10587182 DOI: 10.1016/s0002-9610(99)00186-5]
- 79 Goh BK, Tan YM, Cheow PC, Chung YF, Chow PK, Wong WK, Ooi LL. Cystic lesions of the pancreas: an appraisal of an aggressive resectional policy adopted at a single institution during 15 years. Am J Surg 2006; 192: 148-154 [PMID: 16860621 DOI: 10.1016/j.amjsurg.2006.02.020]
- 80 Oh HC, Seo DW, Song TJ, Moon SH, Park DH, Soo Lee S, Lee SK, Kim MH, Kim J. Endoscopic ultrasonography-guided ethanol lavage with paclitaxel injection treats patients with pancreatic cysts. Gastroenterology 2011; 140: 172-179 [PMID: 20950614 DOI: 10.1053/j.gastro.2010.10.001]
- DeWitt J, McGreevy K, Schmidt CM, Brugge WR. EUS-guided ethanol vs saline solution lavage 81 for pancreatic cysts: a randomized, double-blind study. Gastrointest Endosc 2009; 70: 710-723 [PMID: 19577745 DOI: 10.1016/j.gie.2009.03.1173]
- 82 Moyer MT, Sharzehi S, Mathew A, Levenick JM, Headlee BD, Blandford JT, Heisey HD, Birkholz JH, Ancrile BB, Maranki JL, Gusani NJ, McGarrity TJ, Dye CE. The Safety and Efficacy of an Alcohol-Free Pancreatic Cyst Ablation Protocol. Gastroenterology 2017; 153: 1295-1303 [PMID: 28802565 DOI: 10.1053/j.gastro.2017.08.009]
- 83 Choi JH, Seo DW, Song TJ, Park DH, Lee SS, Lee SK, Kim MH. Long-term outcomes after endoscopic ultrasound-guided ablation of pancreatic cysts. Endoscopy 2017; 49: 866-873 [PMID: 28511236 DOI: 10.1055/s-0043-110030]
- Oh HC, Seo DW. Endoscopic ultrasonography-guided pancreatic cyst ablation (with video). J Hepatobiliary Pancreat Sci 2015; 22: 16-19 [PMID: 25376091 DOI: 10.1002/jhbp.179]
- 85 Ho KY, Brugge WR; EUS 2008 Working Group. EUS 2008 Working Group document: evaluation of EUS-guided pancreatic-cyst ablation. Gastrointest Endosc 2009; 69: S22-S27 [PMID: 19179162 DOI: 10.1016/j.gie.2008.10.059]
- Gómez V, Takahashi N, Levy MJ, McGee KP, Jones A, Huang Y, Chari ST, Clain JE, Gleeson FC, 86 Pearson RK, Petersen BT, Rajan E, Vege SS, Topazian MD. EUS-guided ethanol lavage does not reliably ablate pancreatic cystic neoplasms (with video). Gastrointest Endosc 2016; 83: 914-920 [PMID: 26363331 DOI: 10.1016/j.gie.2015.08.069]
- 87 DeWitt J, DiMaio CJ, Brugge WR. Long-term follow-up of pancreatic cysts that resolve radiologically after EUS-guided ethanol ablation. Gastrointest Endosc 2010; 72: 862-866 [PMID: 20883866 DOI: 10.1016/j.gie.2010.02.039]



- Tanaka M, Fernández-del Castillo C, Adsay V, Chari S, Falconi M, Jang JY, Kimura W, Levy P, 88 Pitman MB, Schmidt CM, Shimizu M, Wolfgang CL, Yamaguchi K, Yamao K; International Association of Pancreatology. International consensus guidelines 2012 for the management of IPMN and MCN of the pancreas. Pancreatology 2012; 12: 183-197 [PMID: 22687371 DOI: 10.1016/j.pan.2012.04.004]
- 89 de Jong K, van Hooft JE, Nio CY, Gouma DJ, Dijkgraaf MG, Bruno MJ, Fockens P. Accuracy of preoperative workup in a prospective series of surgically resected cystic pancreatic lesions. Scand J Gastroenterol 2012; 47: 1056-1063 [PMID: 22571417 DOI: 10.3109/00365521.2012.674970]
- 90 Brugge WR, Lewandrowski K, Lee-Lewandrowski E, Centeno BA, Szydlo T, Regan S, del Castillo CF, Warshaw AL. Diagnosis of pancreatic cystic neoplasms: a report of the cooperative pancreatic cyst study. Gastroenterology 2004; 126: 1330-1336 [PMID: 15131794 DOI: 10.1053/j.gastro.2004.02.013]
- 91 Cizginer S, Turner BG, Bilge AR, Karaca C, Pitman MB, Brugge WR. Cyst fluid carcinoembryonic antigen is an accurate diagnostic marker of pancreatic mucinous cysts. Pancreas 2011; 40: 1024-1028 [PMID: 21775920 DOI: 10.1097/MPA.0b013e31821bd62f]
- Sedlack R, Affi A, Vazquez-Sequeiros E, Norton ID, Clain JE, Wiersema MJ. Utility of EUS in the 92 evaluation of cystic pancreatic lesions. Gastrointest Endosc 2002; 56: 543-547 [PMID: 12297771 DOI: 10.1067/mge.2002.128106]
- Koito K, Namieno T, Nagakawa T, Shyonai T, Hirokawa N, Morita K. Solitary cystic tumor of the 93 pancreas: EUS-pathologic correlation. Gastrointest Endosc 1997; 45: 268-276 [PMID: 9087833 DOI: 10.1016/s0016-5107(97)70269-4]
- Morris-Stiff G, Lentz G, Chalikonda S, Johnson M, Biscotti C, Stevens T, Matthew Walsh R. 94 Pancreatic cyst aspiration analysis for cystic neoplasms: mucin or carcinoembryonic antigen--which is better? Surgery 2010; 148: 638-44; discussion 644 [PMID: 20797749 DOI: 10.1016/j.surg.2010.07.023]
- Thornton GD, McPhail MJ, Nayagam S, Hewitt MJ, Vlavianos P, Monahan KJ. Endoscopic 95 ultrasound guided fine needle aspiration for the diagnosis of pancreatic cystic neoplasms: a metaanalysis. Pancreatology 2013; 13: 48-57 [PMID: 23395570 DOI: 10.1016/j.pan.2012.11.313]
- 96 Dumonceau JM, Deprez PH, Jenssen C, Iglesias-Garcia J, Larghi A, Vanbiervliet G, Aithal GP, Arcidiacono PG, Bastos P, Carrara S, Czakó L, Fernández-Esparrach G, Fockens P, Ginès À, Havre RF, Hassan C, Vilmann P, van Hooft JE, Polkowski M. Indications, results, and clinical impact of endoscopic ultrasound (EUS)-guided sampling in gastroenterology: European Society of Gastrointestinal Endoscopy (ESGE) Clinical Guideline - Updated January 2017. Endoscopy 2017; **49**: 695-714 [PMID: 28511234 DOI: 10.1055/s-0043-109021]
- Ohno E, Hirooka Y, Itoh A, Ishigami M, Katano Y, Ohmiya N, Niwa Y, Goto H. Intraductal 97 papillary mucinous neoplasms of the pancreas: differentiation of malignant and benign tumors by endoscopic ultrasound findings of mural nodules. Ann Surg 2009; 249: 628-634 [PMID: 19300203 DOI: 10.1097/SLA.0b013e3181a189a8]
- 98 Nara S, Onaya H, Hiraoka N, Shimada K, Sano T, Sakamoto Y, Esaki M, Kosuge T. Preoperative evaluation of invasive and noninvasive intraductal papillary-mucinous neoplasms of the pancreas: clinical, radiological, and pathological analysis of 123 cases. Pancreas 2009; 38: 8-16 [PMID: 18665010 DOI: 10.1097/MPA.0b013e318181b90d]
- 99 Wu J, Jiao Y, Dal Molin M, Maitra A, de Wilde RF, Wood LD, Eshleman JR, Goggins MG, Wolfgang CL, Canto MI, Schulick RD, Edil BH, Choti MA, Adsay V, Klimstra DS, Offerhaus GJ, Klein AP, Kopelovich L, Carter H, Karchin R, Allen PJ, Schmidt CM, Naito Y, Diaz LA Jr, Kinzler KW, Papadopoulos N, Hruban RH, Vogelstein B. Whole-exome sequencing of neoplastic cysts of the pancreas reveals recurrent mutations in components of ubiquitin-dependent pathways. Proc Natl Acad Sci USA 2011; 108: 21188-21193 [PMID: 22158988 DOI: 10.1073/pnas.1118046108]
- 100 Canto MI, Hruban RH. Managing pancreatic cysts: less is more? Gastroenterology 2015; 148: 688-691 [PMID: 25724460 DOI: 10.1053/j.gastro.2015.02.033]
- 101 Fernández-Del Castillo C, Tanaka M. Management of pancreatic cysts: the evidence is not here yet. Gastroenterology 2015; 148: 685-687 [PMID: 25724457 DOI: 10.1053/j.gastro.2015.02.034]
- Crippa S, Pezzilli R, Bissolati M, Capurso G, Romano L, Brunori MP, Calculli L, Tamburrino D, 102 Piccioli A, Ruffo G, Fave GD, Falconi M. Active Surveillance Beyond 5 Years Is Required for Presumed Branch-Duct Intraductal Papillary Mucinous Neoplasms Undergoing Non-Operative Management. Am J Gastroenterol 2017; 112: 1153-1161 [PMID: 28244498 DOI: 10.1038/ajg.2017.43]
- 103 Lawrence SA, Attiyeh MA, Seier K, Gönen M, Schattner M, Haviland DL, Balachandran VP, Kingham TP, D'Angelica MI, DeMatteo RP, Brennan MF, Jarnagin WR, Allen PJ. Should Patients With Cystic Lesions of the Pancreas Undergo Long-term Radiographic Surveillance? Ann Surg 2017; 266: 536-544 [PMID: 28657939 DOI: 10.1097/SLA.00000000002371]
- 104 Jais B, Rebours V, Malleo G, Salvia R, Fontana M, Maggino L, Bassi C, Manfredi R, Moran R, Lennon AM, Zaheer A, Wolfgang C, Hruban R, Marchegiani G, Fernández Del Castillo C, Brugge W, Ha Y, Kim MH, Oh D, Hirai I, Kimura W, Jang JY, Kim SW, Jung W, Kang H, Song SY, Kang CM, Lee WJ, Crippa S, Falconi M, Gomatos I, Neoptolemos J, Milanetto AC, Sperti C, Ricci C, Casadei R, Bissolati M, Balzano G, Frigerio I, Girelli R, Delhaye M, Bernier B, Wang H, Jang KT, Song DH, Huggett MT, Oppong KW, Pererva L, Kopchak KV, Del Chiaro M, Segersvard R, Lee LS, Conwell D, Osvaldt A, Campos V, Aguero Garcete G, Napoleon B, Matsumoto I, Shinzeki M, Bolado F, Fernandez JM, Keane MG, Pereira SP, Acuna IA, Vaquero EC, Angiolini MR, Zerbi A,



Tang J, Leong RW, Faccinetto A, Morana G, Petrone MC, Arcidiacono PG, Moon JH, Choi HJ, Gill RS, Pavey D, Ouaïssi M, Sastre B, Spandre M, De Angelis CG, Rios-Vives MA, Concepcion-Martin M, Ikeura T, Okazaki K, Frulloni L, Messina O, Lévy P. Serous cystic neoplasm of the pancreas: a multinational study of 2622 patients under the auspices of the International Association of Pancreatology and European Pancreatic Club (European Study Group on Cystic Tumors of the Pancreas). Gut 2016; 65: 305-312 [PMID: 26045140 DOI: 10.1136/gutjnl-2015-309638]

- 105 Chapman CG, Waxman I. EUS-guided portal vein sampling. Endosc Ultrasound 2018; 7: 240-245 [PMID: 30117486 DOI: 10.4103/eus.eus_28_18]
- Kayar Y, Turkdogan KA, Baysal B, Unver N, Danalioglu A, Senturk H. EUS-guided FNA of a 106 portal vein thrombus in hepatocellular carcinoma. Pan Afr Med J 2015; 21: 86 [PMID: 26491529 DOI: 10.11604/pamj.2015.21.86.6991]
- 107 Garg R, Rustagi T. Endoscopic Ultrasound-guided Portal Venous Access: Diagnostic and Therapeutic Implications. J Clin Gastroenterol 2017; 51: 677-682 [PMID: 28742731 DOI: 10.1097/MCG.00000000000897
- Moreno M, Gimeno-García AZ, Corriente MM, Nicolás-Pérez D, Brito-García A, García-Castro C, 108 Quintero E. EUS-FNA of a portal vein thrombosis in a patient with a hidden hepatocellular carcinoma: confirmation technique after contrast-enhanced ultrasound. Endoscopy 2014; 46 Suppl 1 UCTN: E590-E591 [PMID: 25502254 DOI: 10.1055/s-0034-1390734]
- 109 Gimeno Garcia AZ, Aparicio JR, Barturen A, Moreno M, Nicolas-Perez D, Quintero E. Short article: Endoscopic ultrasound-guided fine-needle aspiration of portal vein thrombosis in patients with chronic liver disease and suspicion of hepatocellular carcinoma. Eur J Gastroenterol Hepatol 2018; 30: 418-423 [PMID: 29420366 DOI: 10.1097/MEG.0000000000001094]
- 110 Park TY, Seo DW, Kang HJ, Cho MK, Song TJ, Park DH, Lee SS, Lee SK, Kim MH. Endoscopic ultrasonography-guided placement of a transhepatic portal vein stent in a live porcine model. Endosc Ultrasound 2016; 5: 315-319 [PMID: 27803904 DOI: 10.4103/2303-9027.191611]
- 111 Faigel D, Lake D, Landreth T, Kelman C, Marler R. Endoscopic ultrasonography-guided portal injection chemotherapy for hepatic metastases. Endosc Ultrasound 2014; 3: S1 [PMID: 26425503]
- Faigel DO, Lake DF, Landreth TL, Kelman CC, Marler RJ. EUS-guided portal injection 112 chemotherapy for treatment of hepatic metastases: feasibility in the acute porcine model. Gastrointest Endosc 2016; 83: 444-446 [PMID: 26358330 DOI: 10.1016/j.gie.2015.08.064]
- Park TY, Seo DW, Kang HJ, Song TJ, Park DH, Lee SS, Lee SK, Kim MH. Feasibility and safety 113 of EUS-guided selective portal vein embolization with a coil and cyanoacrylate in a live porcine model. Endosc Ultrasound 2018; 7: 389-394 [PMID: 30246708 DOI: 10.4103/eus.eus_18_18]
- 114 Matthes K, Sahani D, Holalkere NS, Mino-Kenudson M, Brugge WR. Feasibility of endoscopic ultrasound-guided portal vein embolization with Enteryx. Acta Gastroenterol Belg 2005; 68: 412-415 [PMID: 16432991]
- Chua T, Faigel DO. Endoscopic Ultrasound-Guided Ablation of Liver Tumors. Gastrointest Endosc 115 Clin N Am 2019; 29: 369-379 [PMID: 30846159 DOI: 10.1016/j.giec.2018.11.007]
- 116 Carrara S, Arcidiacono PG, Albarello L, Addis A, Enderle MD, Boemo C, Neugebauer A, Campagnol M, Doglioni C, Testoni PA. Endoscopic ultrasound-guided application of a new internally gas-cooled radiofrequency ablation probe in the liver and spleen of an animal model: a preliminary study. Endoscopy 2008; 40: 759-763 [PMID: 18702032 DOI: 10.1055/s-2008-1077520]
- 117 Varadarajulu S, Jhala NC, Drelichman ER. EUS-guided radiofrequency ablation with a prototype electrode array system in an animal model (with video). Gastrointest Endosc 2009; 70: 372-376 [PMID: 19560138 DOI: 10.1016/j.gie.2009.03.008]
- Jiang T, Tian G, Bao H, Chen F, Deng Z, Li J, Chai W. EUS dating with laser ablation against the 118 caudate lobe or left liver tumors: a win-win proposition? Cancer Biol Ther 2018; 19: 145-152 [PMID: 29303406 DOI: 10.1080/15384047.2017.1414760]
- 119 Di Matteo F, Grasso R, Pacella CM, Martino M, Pandolfi M, Rea R, Luppi G, Silvestri S, Zardi E, Costamagna G. EUS-guided Nd:YAG laser ablation of a hepatocellular carcinoma in the caudate lobe. Gastrointest Endosc 2011; 73: 632-636 [PMID: 21030019 DOI: 10.1016/j.gie.2010.08.019]
- 120 Tang RSY, Kyaw MH, Teoh AYB, Lui RNS, Tse YK, Lam TYT, Chan SL, Wong VWS, Wu JCY, Lau JYW, Sung JJY. Endoscopic ultrasound-guided cyanoacrylate injection to prevent rebleeding in hepatocellular carcinoma patients with variceal hemorrhage. J Gastroenterol Hepatol 2020; 35: 2192-2201 [PMID: 32602261 DOI: 10.1111/jgh.15168]
- Han Y, Sun S, Guo J, Ge N, Wang S, Liu X, Wang G, Hu J. Is endoscopic ultrasonography useful 121 for endoscopic submucosal dissection? Endosc Ultrasound 2016; 5: 284-290 [PMID: 27803900 DOI: 10.4103/2303-9027.191606]
- 122 Pimentel-Nunes P, Dinis-Ribeiro M, Ponchon T, Repici A, Vieth M, De Ceglie A, Amato A, Berr F, Bhandari P, Bialek A, Conio M, Haringsma J, Langner C, Meisner S, Messmann H, Morino M, Neuhaus H, Piessevaux H, Rugge M, Saunders BP, Robaszkiewicz M, Seewald S, Kashin S, Dumonceau JM, Hassan C, Deprez PH. Endoscopic submucosal dissection: European Society of Gastrointestinal Endoscopy (ESGE) Guideline. Endoscopy 2015; 47: 829-854 [PMID: 26317585 DOI: 10.1055/s-0034-1392882]
- 123 Japan Esophageal Society. Japanese Classification of Esophageal Cancer, 11th Edition: part I. Esophagus 2017; 14: 1-36 [PMID: 28111535 DOI: 10.1007/s10388-016-0551-7]
- 124 Thosani N, Singh H, Kapadia A, Ochi N, Lee JH, Ajani J, Swisher SG, Hofstetter WL, Guha S, Bhutani MS. Diagnostic accuracy of EUS in differentiating mucosal vs submucosal invasion of superficial esophageal cancers: a systematic review and meta-analysis. Gastrointest Endosc 2012;



75: 242-253 [PMID: 22115605 DOI: 10.1016/j.gie.2011.09.016]

- 125 May A, Günter E, Roth F, Gossner L, Stolte M, Vieth M, Ell C. Accuracy of staging in early oesophageal cancer using high resolution endoscopy and high resolution endosonography: a comparative, prospective, and blinded trial. Gut 2004; 53: 634-640 [PMID: 15082579 DOI: 10.1136/gut.2003.029421]
- 126 Larghi A, Lightdale CJ, Memeo L, Bhagat G, Okpara N, Rotterdam H. EUS followed by EMR for staging of high-grade dysplasia and early cancer in Barrett's esophagus. Gastrointest Endosc 2005; 62: 16-23 [PMID: 15990814 DOI: 10.1016/s0016-5107(05)00319-6]
- 127 Pech O, Günter E, Dusemund F, Ell C. Value of high-frequency miniprobes and conventional radial endoscopic ultrasound in the staging of early Barrett's carcinoma. Endoscopy 2010; 42: 98-103 [PMID: 20140826 DOI: 10.1055/s-0029-1243839]
- 128 Pech O, May A, Günter E, Gossner L, Ell C. The impact of endoscopic ultrasound and computed tomography on the TNM staging of early cancer in Barrett's esophagus. Am J Gastroenterol 2006; **101**: 2223-2229 [PMID: 17032186 DOI: 10.1111/j.1572-0241.2006.00718.x]
- Ko WJ, Song GW, Cho JY. Evaluation and Endoscopic Management of Esophageal Submucosal 129 Tumor. Clin Endosc 2017; 50: 250-253 [PMID: 27817183 DOI: 10.5946/ce.2016.109]
- Chung IK, Lee JH, Lee SH, Kim SJ, Cho JY, Cho WY, Hwangbo Y, Keum BR, Park JJ, Chun HJ, 130 Kim HJ, Kim JJ, Ji SR, Seol SY. Therapeutic outcomes in 1000 cases of endoscopic submucosal dissection for early gastric neoplasms: Korean ESD Study Group multicenter study. Gastrointest Endosc 2009; 69: 1228-1235 [PMID: 19249769 DOI: 10.1016/j.gie.2008.09.027]
- 131 Gotoda T, Jung HY. Endoscopic resection (endoscopic mucosal resection/ endoscopic submucosal dissection) for early gastric cancer. Dig Endosc 2013; 25 Suppl 1: 55-63 [PMID: 23362925 DOI: 10.1111/den.12003]
- 132 Lian J, Chen S, Zhang Y, Qiu F. A meta-analysis of endoscopic submucosal dissection and EMR for early gastric cancer. Gastrointest Endosc 2012; 76: 763-770 [PMID: 22884100 DOI: 10.1016/j.gie.2012.06.014]
- Park YM, Cho E, Kang HY, Kim JM. The effectiveness and safety of endoscopic submucosal 133 dissection compared with endoscopic mucosal resection for early gastric cancer: a systematic review and metaanalysis. Surg Endosc 2011; 25: 2666-2677 [PMID: 21424201 DOI: 10.1007/s00464-011-1627-z]
- 134 Choi J, Kim SG, Im JP, Kim JS, Jung HC, Song IS. Comparison of endoscopic ultrasonography and conventional endoscopy for prediction of depth of tumor invasion in early gastric cancer. Endoscopy 2010; 42: 705-713 [PMID: 20652857 DOI: 10.1055/s-0030-1255617]
- 135 Gotoda T, Yamamoto H, Soetikno RM. Endoscopic submucosal dissection of early gastric cancer. J Gastroenterol 2006; 41: 929-942 [PMID: 17096062 DOI: 10.1007/s00535-006-1954-3]
- 136 Ajani JA, Bentrem DJ, Besh S, D'Amico TA, Das P, Denlinger C, Fakih MG, Fuchs CS, Gerdes H, Glasgow RE, Hayman JA, Hofstetter WL, Ilson DH, Keswani RN, Kleinberg LR, Korn WM, Lockhart AC, Meredith K, Mulcahy MF, Orringer MB, Posey JA, Sasson AR, Scott WJ, Strong VE, Varghese TK Jr, Warren G, Washington MK, Willett C, Wright CD, McMillian NR, Sundar H; National Comprehensive Cancer Network. Gastric cancer, version 2.2013: featured updates to the NCCN Guidelines. J Natl Compr Canc Netw 2013; 11: 531-546 [PMID: 23667204 DOI: 10.6004/jnccn.2013.0070
- 137 Meng FS, Zhang ZH, Hong YY, Li DJ, Lin JQ, Chen X, Ji F. Comparison of endoscopic submucosal dissection and surgery for the treatment of gastric submucosal tumors originating from the muscularis propria layer: a single-center study (with video). Surg Endosc 2016; 30: 5099-5107 [PMID: 27005293 DOI: 10.1007/s00464-016-4860-7]
- Fujii LL, Gomez V, Song LM, Levy MJ. Endoscopic ultrasound-assisted endoscopic submucosal 138 dissection of a gastric subepithelial tumor. Endoscopy 2013; 45 Suppl 2 UCTN: E225-E226 [PMID: 23945921 DOI: 10.1055/s-0033-1344157]
- Fernández-Esparrach G, Ayuso-Colella JR, Sendino O, Pagés M, Cuatrecasas M, Pellisé M, 139 Maurel J, Ayuso-Colella C, González-Suárez B, Llach J, Castells A, Ginès A. EUS and magnetic resonance imaging in the staging of rectal cancer: a prospective and comparative study. Gastrointest Endosc 2011; 74: 347-354 [PMID: 21802588 DOI: 10.1016/j.gie.2011.03.1257]
- Hurlstone DP, Brown S, Cross SS, Shorthouse AJ, Sanders DS. High magnification chromoscopic 140 colonoscopy or high frequency 20 MHz mini probe endoscopic ultrasound staging for early colorectal neoplasia: a comparative prospective analysis. Gut 2005; 54: 1585-1589 [PMID: 15964906 DOI: 10.1136/gut.2005.069849]



 \mathcal{O} W J

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 1880-1895

DOI: 10.4251/wjgo.v13.i12.1880

ISSN 1948-5204 (online)

REVIEW

Pancreatic intraductal papillary mucinous neoplasms: Current diagnosis and management

Beata Jabłońska, Paweł Szmigiel, Sławomir Mrowiec

ORCID number: Beata Jabłońska 0000-0002-5495-2969; Paweł Szmigiel 0000-0003-2973-1758; Sławomir Mrowiec 0000-0003-2206-3144.

Author contributions: Jabłońska B reviewed the literature and drafted the manuscript; Szmigiel P reviewed the literature; Mrowiec S revised the manuscript.

Conflict-of-interest statement: No conflict of interest.

Country/Territory of origin: Poland

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0)

Beata Jabłońska, Paweł Szmigiel, Sławomir Mrowiec, Department of Digestive Tract Surgery, Medical University of Silesia, Katowice 40-752, Poland

Corresponding author: Beata Jabłońska, MD, PhD, Adjunct Professor, Department of Digestive Tract Surgery, Medical University of Silesia, Medyków 14, Katowice 40-752, Poland. bjablonska@poczta.onet.pl

Abstract

Intraductal papillary mucinous neoplasms (IPMNs) represent approximately 1% of all pancreatic neoplasms and 25% of cystic neoplasms. They are divided into three types: main duct-IPMN (MD-IPPMN), branch duct-IPMN (BD-IPMN), and mixed type-IPMN. In this review, diagnostics, including clinical presentation and radiological investigations, were described. Magnetic resonance imaging is the most useful for most IPMNs. Management depends on the type and radiological features of IPMNs. Surgery is recommended for MD-IPMN. For BD-IPMN, management involves surgery or surveillance depending on the tumor size, cyst growth rate, solid components, main duct dilatation, high-grade dysplasia in cytology, the presence of symptoms (jaundice, new-onset diabetes, pancreatitis), and CA 19.9 serum level. The patient's age and comorbidities should also be taken into consideration. Currently, there are different guidelines regarding the diagnosis and management of IPMNs. In this review, the following guidelines were presented: Sendai International Association of Pancreatology guidelines (2006), American Gastroenterological Association guidelines, revised international consensus Fukuoka guidelines (2012), revised international consensus Fukuoka guidelines (2017), and European evidence-based guidelines according to the European Study Group on Cystic Tumours of the Pancreas (2018). The Verona Evidence-Based Meeting 2020 was also presented and discussed.

Key Words: Pancreatic cyst; Pancreatic cystic neoplasm; Intraductal papillary mucinous neoplasm; Pancreatic cancer; Pancreatectomy; Guidelines

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Intraductal papillary mucinous neoplasms (IPMNs) account about 1% of all pancreatic neoplasms and 25% of cystic neoplasms. We can distinguish three IPMN


license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: February 22, 2021 Peer-review started: February 22, 2021 First decision: June 4, 2021 Revised: June 17, 2021 Accepted: October 18, 2021 Article in press: October 18, 2021 Published online: December 15, 2021

P-Reviewer: Ausania F S-Editor: Wu YXJ L-Editor: A P-Editor: Wu YXJ



types: main duct-IPMN (MD-IPPMN), branch duct-IPMN (BD-IPMN), and mixed type-IPMN. Magnetic resonance imaging is the most useful approach for most IPMNs. Management depends on the type and radiological features of IPMNs. MD-IPMN is recommended for surgery. In BD-IPMN, management involves surgery or surveillance depending on the tumor size, cyst growth rate, solid components, main duct dilatation, high-grade dysplasia in cytology, the presence of symptoms (jaundice, new-onset diabetes, pancreatitis), and CA 19.9 serum level. The patient's age and comorbidities should also be taken into consideration. Currently, there are different guidelines regarding the diagnostics and management of IPMNs: Sendai International Association of Pancreatology guidelines (2006), American Gastroenterological Association guidelines, revised international consensus Fukuoka guidelines (2012), revised international consensus Fukuoka guidelines (2017), and European evidence-based guidelines based on the European Study Group on Cystic Tumors of the Pancreas (2018). The experts of Verona Evidence-Based Meeting 2020 determined the most important further directions regarding guidelines on IPMN management.

Citation: Jabłońska B, Szmigiel P, Mrowiec S. Pancreatic intraductal papillary mucinous neoplasms: Current diagnosis and management. World J Gastrointest Oncol 2021; 13(12): 1880-1895

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/1880.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.1880

INTRODUCTION

Pancreatic cystic neoplasms represent about 10%-13% of pancreatic cysts, 25% of cystic neoplasms and 1% of pancreatic carcinomas[1,2]. Pancreatic intraductal papillary mucinous neoplasms (IPMNs) are one of the two types of mucin-producing pancreatic cystic tumors (PCTs)[1,2]. According to World Health Organization, IPMNs are neoplasms which grow within the pancreatic ducts and produce mucin. They contain epithelial cells that can create papillary projections[2]. In 1982, Ohhashi et al[3], for the first time, reported four cases of mucin-producing pancreatic cancer. The term "intraductal papillary neoplasm" was introduced by Morohoshi[4] in a report of six cases in 1989. It should be added that numerous different terms were used for IPMNs before establishing the current nomenclature. The earlier names used were as follows: mucinous ductal ectasia, ductectatic mucinous cystadenoma and cystadenocarcinoma, intraductal mucin-hypersecreting neoplasm, intraductal papillary adenocarcinoma, intraductal mucin-producing tumor, and mucin-producing tumor^[1].

At this time, the number of pancreatic IPMNs has significantly increased, and there are many reports on these tumors. The aim of this study is to review and present most of the current important literature regarding the etiopathogenesis, classification, diagnostics and treatment of pancreatic IPMNs.

ETIOLOGY AND PATHOGENESIS OF IPMNS

The etiology of pancreatic IPMNs is not clear. A main feature of many IPMNs is excessive mucin production. It has been reported that mucin 2 (MUC2) is procuded by most IPMNs, while there is no expression of mucin 1 (MUC1) in IPMNs, except of components of ductal cancer[5,6]. Adsay et al[6] noted that invasive ductal adenocarcinomas develop from intraepithelial neoplasms of the pancreas (PanINs) (5-y survival is less than 15%), whereas IPMNs are often associated with colloid carcinoma (5-y survival is better of more than 55%). It is known that an associated invasive carcinoma is reported in approximately 30% of patients with IPMN. Adsay et al[6] described an association of mentioned above pancreatic pathologies by investigating the expression of MUC1 and MUC2 glycoproteins as "aggressive" and "indolent" phenotypes in pancreatic carcinoma, respectively. In fact, MUC1 (mammary-type mucin) and MUC2 (intestinal-type mucin) have been reported as markers of "aggressive" and "indolent" phenotypes in pancreatic cancer, respectively. IPMN and colloid (mucinous noncystic) carcinoma form a distinct pathway of carcinogenesis in the pancreas, and MUC2 may



be the marker of this pathway. Furthermore, ordinary ductal carcinoma of the pancreas was found to lack expression of this marker but showed MUC1 expression instead^[6]. In conclusion, the results of this study supported a dichotomial nature of the dysplasia-carcinoma in situ (CIS) sequence in the pancreas. Authors analyzed 2 routes leading to different types of invasive cancers. They noted that MUC2 is a marker of the "indolent" pathway (IPMN and colloid cancer), and MUC1 is a marker of the "aggressive" pathway (PanIN to ductal adenocarcinoma)[6].

In IPMNs, a classic "adenoma-carcinoma sequence" is observed. The duration of developing invasive carcinoma from low-grade dysplasia is approximately from 4 to 6 years. Various somatic mutations in the oncogenes KRAS and GNAS are reported in up to 90% of IPMNs. Other mutated genes are as follows: CDKN2A/p16, TP53, SMAD4, and less commonly STK11, BRAF, PIK3CA, PTEN. It has been noted that inactivated CDKN2A/p16, absent SMAD4 and mutation in TP53 are associated with progression from IPMN to carcinoma. They are almost exclusively reported in malignant IPMNs[7].

CLASSIFICATION OF IPMNS

IPMN is an exocrine neoplasm of the pancreas consisting of epithelial cells growing within the pancreatic ducts [main pancreatic duct (MPD) or its major branches and producing mucin[1]. There is no ovarian-type stroma in IPMNs in contrast to mucinous cystic neoplasms^[2]. According to the revised international consensus Fukuoka guidelines (2017)[8], IPMNs are divided into the following three types: MD-IPMN, BD-IPMN, and MT-IPMN diagnosed in radiological/histological investigations. In MD-IPMN, MPD segmental or diffuse dilation of > 5 mm without other obstruction reasons is noted. Although MPD dilation of 5-9 mm is not an absolute indication for surgery, it is one of the "worrisome features". MPD diameter ≥ 10 mm is one of the "high-risk stigmata". BD-IPMNs are cystic lesions of the pancreas measuring > 5 mm which communicate with MPD. They need differential diagnosis with pseudocysts in patients followingacute pancreatitis. In MT-IPMN, the features of both MD-IPMN and BD-IPMN are present[7].

HISTOPATHOLOGY OF IPMNS

Histologically, pancreatic IPMNs are noninvasive epithelial neoplasms arising from cells which produce mucin located within the MPD or its branches[9]. According to the degree of cytological atypia and abnormal crowding of the epithelium, low-grade, intermediate-grade and high-grade dysplasia IPMNs are distinguished^[10]. The four histopathological IPMN types are distinguished such as gastric type (49%-63%), intestinal type (18%-36%), pancreaticobiliary type (7%-18%), and oncocytic type (1%-8%). The gastric type is observed the most commonly. It is typically of low grade, rarely leading to cancer. Pancreatic cancer developing from this IPMN type is usually of the tubular type and is similar to ordinary pancreatic ductal adenocarcinoma. The intestinal type is reported in numerous MD-IPMNs. The pancreaticobiliary type is not well characterized and is uncommon. According to some authors, it is a high-grade dysplasia variation of the IPMN gastric type. Ductal and aggressive invasive cancer is commonly related to this IPMN type. The oncocytic type is the less frequent variant consisting of complex aborising papillae with delicate cores, oncocytic cells, and intraepithelial lumina formation. These lesions are uncommon and have limited invasion capability. Histological types correlate with the immunohistochemical phenotype of IPMN. This correlation was presented in Table 1[7,9].

DIAGNOSTICS OF IPMNS

Diagnostics of IPMNs involve analysis of clinical presentation, radiological imaging, and laboratory investigations, including biochemical and cytological tests.

Clinical presentation

The following clinical symptoms have been reported in patients with IPMNs: Epigastric discomfort or pain (70%-80%), loss of weight (20%-40%), nausea and vomiting (11%-21%), backache (10%), diabetes, and jaundice[5]. The mucin, which is



Table 1 Histological types and immunohistochemical profiles of intraductal papillary mucinous neoplasms[7,9]						
Туре	Percentage	Immunohistochemical profile				
		MUC1	MUC2	MUC5AC	MUC6	
Gastric	49-63	(-)	(-)	(+)	(+)	
Intestinal	18-36	(-)	(+)	(+)	(±)	
Pancreatobiliary	7-18	(+)	(-)	(+)	(±)	
Oncocytis	1-8	(+)	(-)	(±)	(+)	

MUC: Mucin.

hyperproduced, can obstruct normal secretion in the pancreas, that is a reason of meals-related pain. In this case, a patient does not eat to avoid pain. In advanced tumors, loss of appetite is related to neoplastic cachexia. Jaundice is a consequence of obstruction of the common bile duct by viscid mucin, mural nodules, or direct compression due to the size of the IPMN. Persistent occlusion of the MPD with mucin can lead to exocrine and/or endocrine pancreatic insufficiency, and persistent hyperamylasemia[5]. Regarding clinical presentation, an association between IPMNs and recurrent acute pancreatitis (AP) should be emphasized. According to Venkatesh et al [10], the AP is reported in 12%-67% of IPMN patients. Both MD-IPMN and BD-IPMN may lead to AP, with a similar risk. AP in IPMN patients is usually mild and does not need treatment. There is no difference in AP occurence between benign and malignant IPMNs. AP occurs more frequently in IPMN patients compared to cancer patients, possibly because of obstruction of the MPD by mucin. It is important to remember the above mentioned association in patients with recurrent AP. Frequently, in patients following AP, pancreatic pseudocysts or fluid collections are diagnosed and IPMNs are less frequently considered in the differential diagnosis. In our opinion, oncological vigilance is very important in patients with pancreatic cystic lesions and recurrent pancreatitis in medical history because the prognosis and management of patients with IPMNs and pancreatic pseudocysts are different[10]. Jang et al[11] analyzed IPMN patients with AP or acute recurrent pancreatitis (ARP) (AP/ARP) treated in the period of 2000-2008 in a single tertiary referral center. IPMN-associated AP/ARP was noted 34 (7%) of 488 IPMN patients, and the MD/MT-IPMN more frequently was associated with AP/ARP compared to the BD-type (14% vs 5%; P = 0.002). The mild AP was diagnosed in analyzed patients. Histological findings of 24 surgically treated tumors were as follows: Adenomas (n = 4) (17%), borderline malignancies (n = 17)(71%), CIS (n = 2) (8%), and invasive carcinoma (n = 1) (4%). There was no AP/ARP recurrence in any patients during the follow-up period (median 52 mo, range 38-115 mo). The authors concluded that, though uncommon, AP/ARP could be an initial clinical IPMN manifestation, which is helpful in the diagnostic process[11].

Regarding ARP as a clinical IPMN manifestation, Bernardoni *et al*[12], in their preliminary report, assessed the efficacy of pancreatic sphincterotomy (PS) in patients with IPMN-associated ARP. A prerequisite for treatment was the fact that IPMNassociated ARP may lead to a lower quality of life and chronic pancreatitis. In IPMN manifested as AP, a higher cancer risk is reported. According to Fukuoka consensus [13], pancreatitis may be an indication for surgery despite of no signs of malignancy in radiological and cytological investigations[12,13]. However, pancreatic surgery is associated with an increased morbidity and mortality risk even when performed at high volume surgical centers. Higher surgical risks are reported in old patients with numerous comorbidities. According to the IPMN-associated AP pathophysiology, the hypothesis regarding the falicitated mucin outflow into the duodenum by PS has developed. According to this theory, reduction of intraductal pressure could lead to reduction of AP episodes[12]. The authors retrospectively analyzed patients with ARP and IPMN undergoing PS in 2010-2015. Patients were divided into two different groups: (1) MD/MT-IPMN; and (2) BD-IPMN with or without worrisome features/high-risk stigmata. In this study, complete, partial (reduction of pancreatitis episodes > 50%), and no response were reported in 11 (68.7%), 3 (18.7%), and 2 (12.5%) patients, respectively. In 1 (6.25%) patient, mild pancreatitis was observed following endoscopic retrograde cholangiopancreatography (ERCP). There was no cancer in resected patients. Additionally, during follow-up, there were no worrisome features/ high-risk stigmata^[12]. The authors concluded that PS was effective for reduction of the number of AP and it should be taking into condideration as a treatment option in



selected IPMN patients. It is important that systematic follow-up should be performed in this patients' group due to the malignant IPMN potential^[12].

Apart from typical IPMN clinical presentation regarding abdominal symptoms and jaundice, skin lesions named pancreatic panniculitis have been reported. Yamashita et al[14] described a case of a 68-year-old man presenting pancreatic panniculitis on his trunk coexisted with IPMN-associated AP. A skin biopsy of the lesion histologically showed lobular panniculitis with characteristic "ghost cells" (pancreatic panniculitis). The authors concluded that clinicians should take into account IPMN in patients with in orderto avoid a missed or delayed diagnosis^[14]. Similar cases of AP and IPMNrelated panniculitis have also been reported by other authors[15,16]. Furthermore, IPMN-related panniculitis has been reported[17,18]. Therefore, we also recommend oncological vigilance in patients with panniculitis.

Infrequently, IPMN can form a fistula into the adjoining organs, including the stomach, duodenum, common bile duct, large and small bowel. The fistula may be related to benign IPMN (low-grade dysplasia). This fistula may occur as a consequence of mechanical penetration as a result of pressure by the mucin-filled ducts or due to inflammation or autodigestion by enzyme-rich fluids, or it could be a result of direct invasion due to malignancy, as in malignant IPMN (high-grade dysplasia)[5,9].

Some clinical symptoms, such as jaundice and new-onset diabetes, are more frequently associated with IPMN malignancy[5,19]. Additionally, according to Weisenauer et al [19], new-onset diabetes mellitus and jaundice suggest malignant IPMN. The authors noted that the absence of these features did not predict benign disease^[19].

Imaging diagnostics of IPMNs

Currently, there are several different guidelines on diagnostic and therapeutic management in IPMN, including Sendai International consensus guidelines for the management of pancreatic IPMNs and mucinous cystic neoplasms according to the International Association of Pancreatology (IAP) (2006)[20], American Gastroenterological Association Institute guidelines on the diagnosis and management of asymptomatic neoplastic pancreatic cysts according to the American Gastroenterological Association (AGA) (2015)[21], revised international consensus Fukuoka guidelines for the management of IPMN of the pancreas (2012)[13], revised international consensus Fukuoka guidelines for the management of IPMN of the pancreas (2017)[8], and European evidence-based guidelines on pancreatic cystic neoplasms according to the European Study Group on Cystic Tumors of the Pancreas (2018)[22]. Diagnostic investigations are performed to select IPMN patients indicated for surgical resection. Therefore, diagnostic investigations should show alarming symptoms for malignant transformation in IPMN. As such, indications for surgery according to different guidelines should be known. They are presented in Table 2[5,8,13,20-22,41].

Computed tomography of the abdominal cavity

According to the most recent European guidelines for pancreatic cystic neoplasms (PCNs) (2018)[22], the accuracy of abdominal CT for identifying the specific PCN type is 40%-81% [22]. Multidetector row computed tomography (MDCT) for IPMN diagnosis should be performed according to a special standardized protocol[22]. Takeshita et al[23] evaluated predictive factors for discriminating benign from malignant pancreatic IPMN on MDCT. The study included 53 patients. Tumors were classified as MD-type (n = 7) and BD-type (n = 46). All MD-IPMNs weremalignant, while 8 of 46 BD-IPMNs were malignant, and 38 were benign. In additionn, MPD dilatation and mural nodules or large cystic diameter combined were significant risk factors of malignancy in BD-IPMN. According to the authors, MD-IPMN is strongly associated with malignancy[24]. Nakagawa et al[24] retrospectively evaluated the utylity of MDCT with multiplanar reformations and curved planar reformations in diagnosis of protruding lesions in IPMNs compared to single-detector CT (SDCT) and endoscopic ultrasonography (EUS). This study showed that MDCT was more useful than SDCT and similar to EUS in diagnosis of protruding lesions in IPMNs[24]. Tan et al[25] also retrospectively evaluated the imaging features of IPMNs in MDCT. Comparison with the pathological diagnosis revealed that the sensitivity, specificity, and accuracy of MDCT in assessing the IPMN were 100%, 87.5% and 95%, respectively. Thus, MDCT can be used to predict the IPMN malignancy[26]. Murayama et al[26], compared CT and MRI in assessment of IPMN malignancy. There was a statistical difference in MPD diameter (P = 0.017) and intraductal volume (P = 0.0013) inadenoma, CIS, and invasive cancer. This study showed that intraductal volume (≥ 10 cm) was helpful in the malignant IPMN diagnosis[26].



Table 2 Indications for surgery in intraductal papillary mucinous neoplasms according to the International, European and American Gastroenterological Association guidelines[5,8,13,20-22,41]

Guidelines	Indications for surgery				
IAP (2006)	Symptoms; Cyst size \geq 3 cm; Mural nodule; MPD \geq 5 mm; Positive cytology				
AGA (2015)	High risk features: Cyst size \geq 3 cm; Presence of solid component; Dilated MPD				
	HGD or cancer on cytology				
IAP (2017)	High risk stigmata: Jaundice; Enhancing mural nodule \geq 5 mm; MPD \geq 10 mm				
	HGD or cancer on cytology				
	Worrisome features: Cyst size ≥ 3 cm; Acute pancreatitis (due to IPMN)				
	Enhancing mural nodule \geq 5 mm; Thickened and enhancing cyst wall				
	MPD dilation 5-9 mm; Abrupt change of MPD caliber with distal pancreatic atrophy; Presence of lymphadenopathy; Elevated serum CA 19-9; Cyst growth rate > 5 mm/2 yr				
European	Absolute indications: Jaundice; Enhancing mural nodule ≥ 5 mm; MPD ≥ 10 mm; HGD or cancer on cytology; Solid mass				
(2018)	Relative indications: Cyst size \geq 4 cm; Enhancing mural nodule \geq 5 mm/years; Acute pancreatitis (due to IPMN); New onset of diabetes; Rapidly increasing cyst size; Elevated serum levels of CA19-9				

IPMN: Intraductal papillary mucinous neoplasm; IAP: International Association of Pancreatology; AGA: American Gastroenterological Association; MPD: Main pancreatic duct; HGD: High grade dysplasia.

> Monnings et al[27] analyzed preoperative CT scans in IPMN patients. Benign (bIPMN; n = 28) and malignant (mIPMN; n = 19) tumors were compared. The MPD diameter was greater in patients with mIPMN (P < 0.0001). Obstruction of the bile duct, solid tumor components, contrast enhancement in walls of the cyst, peripancreatic lymph nodes, and abrupt MPD diameter changes were observed in more mIPMN patients (P < 0.01). In addition, in mIPMN, the CT cyst density was higher (P= 0.0063). The summary diagnostic accuracy was higher than all single CT parameters [27].

> Apart from the numerous above mentioned benefits, CT also has a disadvantage, which is most important in IPMN patients requiring systematic control imaging diagnostics. It has been reported that repeated exposure to ionizing radiation following CT increases the cancer risk[22,28]. Sodicson et al[28] estimated the cumulative radiation exposure and lifetime attributable risk (LAR) of radiation-induced cancer from CT scanning of adult patients at a tertiary care academic medical center. The analysis showed that 33% of patients had \geq 5 lifetime CT investigations, and 5% had 22-132 examinations. Cumulative effective doses > 100 mSv in 15%, and 250-1375 mSv in 4% of patients, respectively, were reported. In 7 % of patients, LAR > 1% was noted. It should be added that assigned effective doses per CT examination are as follows: for CT of the abdomen (without pelvis), 7.5 mSv, and for CT of the abdomen and pelvis, 15 mSv[28].

MRI and magnetic resonance cholangiopancreatography

According to European guidelines for PCNs[22], the accuracy of MRI/magnetic resonance cholangiopancreatography (MRCP) for identifying the special PCN type is 40%-95%. These guidelines recommend MRI as the preferred method for the investigation of patients with PCN. The higher sensivity of MRI/MRCP compared to CT for detection of communication between a PCN and the pancreatic ducts and presence of mural nodules or internal septations has been noted. MRI/MRCP is also good in the differential diagnosis of single and multiple PCNs, including multifocal BD-IPMN. Moreover, IPMN patients frequently require long-life control investigations, and MRI is less invasive than CT^[24]. According to the same guidelines, MDCT is helpful for diagnosis of calcification, tumor staging assessment, or for diagnosing postoperative recurrent disease^[22].

Min et al^[29] retrospectively analyzed patients undergoing surgery for IPMN followingpreoperative CT and MRI in 2009-2019. There were 88 (50.3%) malignant IPMNs in this study. All 3 high-risk stigmata (MPD \ge 10 mm, mural nodule \ge 5 mm, and obstructive jaundice) and 2 worrisome features (MPD 5-9 mm and increased level of CA 19.9) were related to malignant IPMN on CT and MRI (P < 0.05). A mural nodule < 5 mm on MRI was also related to malignant IPMN (P < 0.01). This study showed that CT and MRI were comparable for diagnosis of high risk stigmata (73.7% vs 75.4%; P =



0.505). In addition MRI was superior to CT for diagnosis of mural nodules, and similar to CT for differentiation ofmalignant from benign IPMNs[29].

Liu *et al*[30], in a meta-analysis, assessed the diagnostic properties of CT, PET/CT, MRI/MRCP, DWI, and EUS in differential IPMN diagnosis (benign *vs* malignant tumors). Twenty eight studies were included. This study showed the highest diagnostic accuracy results for PET/CT, and the use of MRI/MRCP, PET/CT was recommended as a first-line investigation in the diagnosis of malignant IPMN, and DWI, EUS and CT were additional for MRI/MRCP in IPMN diagnosis[30].

Jeon *et al*[31] investigated the MRI utility to predict the malignant IPMN potential. In this study, enhancing mural nodule size ≥ 5 mm, MPD ≥ 10 mm / MPD of 5-9 mm, and MPD abrupt changes significantly predicted to malignant IPMNs (P < 0.05). In multivariate analysis, enhancing mural nodules ≥ 5 mm, MPDs ≥ 10 mm or MPDs of 5-9 mm, larger entropy, smaller compactness were significant predictors for malignant IPMNs (P < 0.05)[31].

Boraschi *et al*[32] retrospectively in their retrospective study, showed the MRI utility in the diagnosis of worrisome features and high-risk stigmata in patients with BD-IPMNs during 10 years of observation from the tumor diagnosis[32].

Endoscopic ultrasound

According to European guidelines[22], EUS is recommended as additional to other radiological investigations. It is helpful for diagnosing PCN indicated for surgery. Similar to MRI and CT, EUS is not perfect in diagnosis of the exact PCN type of EUS is recommended in patients with PCNs with concern clinical or radiological features[22].

Contrast harmonic enhanced EUS (CH-EUS) is recommended for assessment of mural nodules. CH-EUS is also useful in assessment of presence of vessels and septations within the cyst. Hyperenhancement of a mural nodule, solid mass, or septations on CH-EUS predict malignancy, that is indication for EUS-fine needle aspiration (FNA) of the tumor[22].

Choi *et al*[33] compared EUS, CT and MR in the diagnosis of IPMN malignant transformation. All compared investigations were similar in this analysis. In the multivariable analysis, enhanced solid components on contrast-enhanced CT and MRI and mural nodules on EUS, MPD diameters \geq 10 mm, MPD diameters of 5-9 mm and thickened septa or walls were significant (*P* < 0.05). Thus, the diagnostic performance of CT, MRI, and EUS for prediction of malignant IPMNs was comparable[33].

The diagnostic accuracy of EUS increases if biopsy is performed and pancreatic cyst fluid is collected for analysis during EUS. EUS-FNA increases diagnostic accuracy for differential diagnosis of mucinous from nonmucinous PCN and malignant from benign PCN in patients in whom CT or MRI are unclear. A combined analysis of cyst fluid CEA, lipase levels, and cytology has the highest accuracy for differential diagnosis of mucinous from nonmucinous PCNs. It is important that EUS-FNA is recommended only when the results can modify management and EUS-FNA should not be performed if the diagnosis is already made using radiological investigations and in patients with clear indications for surgical treatment. Relative contraindications for this investigation are as follows: A distance of > 10 mm between the cyst and the transducer, a high hemorrhage risk, and the use of dual antiplatelet drugs[22]. Assessment of cyst fluid CEA, combined with cytology, or KRAS/GNAS mutation analyses may be considered for differentiating an IPMN or MCN from other PCNs[22].

Mc Carty *et al*[34] published a systematic review and meta-analysis including 6 studies (785 tumors) to asses the diagnostic utility of K-ras and G-nas mutations in EUS-acquired pancreatic cyst fluid for the diagnosis of IPMNs and mucinous cystic lesions. It should be added that molecular cyst fluid diagnostics are not yet a standard. There was a significantly higher accuracy of combined K-ras + G-nas compared to K-ras alone and G-nas alone in the differential diagnosis (P < 0.001). The pooled sensitivity, specificity, and diagnostic accuracy of K-ras + G-nas mutations in the IPMN diagnosis were 94%, 91% and 97%, respectively. They were significantly higher compared to CEA alone (all P < 0.001)[34].

Kadayifci *et al*[35] investigated the value of GNAS investigation in addition to KRAS and CEA tests of pancreatic cystic fluid (PCF) for the IPMN diagnosis. There were 108 IPMN and 89 non-IPMN patients in the analyzed group. GNAS was noted in 51 (47.2%) IPMN patients, and a KRAS mutation was noted in 42 (82.3%) patients. The diagnostic accuracy increased from 76.6% to 79.1% (P > 0.05), when GNAS to KRAS was added and from 66.4% to 80.7% (P < 0.05) when GNAS to CEA was added. It should be noted that the diagnostic accuracy of the combined all tests was significantly higher compared to all single investigations (P < 0.05)[35].

Zaishidena® WJGO | https://www.wjgnet.com

Lee et al[36] published a meta-analysis to analyze KRAS and GNAS mutations in pancreatic cystic lesions. In this study, KRAS and GNAS mutations were more common in IPMNs compared to mucinous and serous cystic neoplasms, respectively. KRAS and GNAS mutations were frequently reported in the gastric (P < 0.001) and intestinal (P < 0.001) types, respectively. KRAS mutation was not common in highgrade dysplasia IPMNs (P = 0.032). This meta-analysis confirmed that KRAS and GNAS mutations are useful for diagnostic tools for IPMN[36].

Gillis et al[37], in their meta-analysis, noted 42% sensitivity and 99% specificity of PCF cytological analysis for differential diagnosis of mucinous vs nonmucinous PCNs [22]. According to most authors, a cyst fluid CEA cutoff level of \geq 192 ng/mL can differentiate mucinous cysts from nonmucinous cysts, with a sensitivity of 52%-78% and specificity of 63%-91%[22].

Indications for EUS-FNA are different depending on International Consensus Guidelines (ICG), AGA, and European guidelines. According to ICG, this investigation is indicated in patients with pancreatitis, tumor diameter > 30 mm, thickened or enhanced wall of the cyst, MPD 5-9 mm, nonenhancing mural nodules, abrupt tapering of the pancreatic duct and atrophy of the distal tail. AGA recommends EUS-FNA in the presence of two of the following risk factors: cyst diameter > 30 mm, the presence of a solid component in the cyst, and MPD dilatation. The European guidelines recommend the use of EUS as part of a multimodality diagnostic assessment[38,39].

ERCP and/or pancreatoscopy

The role of ERCP in IPMN diagnostics is limited. According to European guidelines [22], pancreatoscopy may be used in selected patients to assess the MD-IPMN location and extent and can help to differentiate MD-IPMN from chronic pancreatitis. The diagnostic accuracy of pancreatoscopy was higher in MD-IPMN (88%) compared to BD-IPMN (67%). Intraoperative MPD pancreatoscopy made with frozen sections of intraductal biopsies may be used in assessment of the IPMN extent and MPD involvement, which is important for surgeons' decisions regarding the extent of surgical resection[22].

Blood tests

The role of blood tests in IPMN diagnostics is also limited. According to current guidelines on IPMNs^[22], molecular blood tests are not used in PCNs diagnostics. Only serum cancer antigen CA 19.9 can be useful in IPMN in patients with malignant transformation suspected[22].

MANAGEMENT OF IPMNS

Indications for surgery

Management of IPMNs is still controversial because of different recommendations of the ICG, AGA, and European guidelines. The earliest (2006) Sendai ICG guidelines were the most restrictive. In 2006, Tanaka et al[20] recommended resecting all MD- and MT-IPMNs as long as the patient is a good candidate for surgery. Patients with BD-IPMNs, with no symptoms, require surgery not only to relief the signs but also due to a n increased risk of malignant transformation. Moreover, according to these guidelines, BD IPMNs > 30 mm in diameter and without MPD dilation or mural nodules should be assessed if all BD-IPMNs > 30 mm in diameter require surgery immediately. The Sendai recommendations have resulted in a high rate of "unnecessary" pancreatic surgeries. This is important because pancreatectomy is a complex procedure associated with relatively high morbidity and mortality rates[38]. The original Sendai group published revised ICG, commonly known as the Fukuoka guidelines in 2012. According to the IAP Fukuoka 2012 guidelines, revised in 2017, surgery is strongly recommended for all MD-IPMNs with a MPD of diameter > 10 mm or with "high-risk stigmata" (HR), such as an enhancing solid component or jaundice. Dilatation of the MPD 5-9 mm is considered a "worrisome feature," and it is not recommended for immediate resection but requiring further assessment using EUS[8, 13]. In 2015, AGA recommended surgical treatment for patients, with no symptoms, only in the presence of two of three "concerning features" (presence of nodule, diameter > 30 m, or duct dilation) and malignant transformation in EUS-FNA[21].

Authors of the European guidelines^[22] recommended surgery in IPMNs with jaundice, an enhancing mural nodule (\geq 5 mm) or a solid component, positive cytology, or MPD diameter \geq 10 mm. Surgical management was also recommended for



IPMNs with MPD dilatation 5-9.9 mm, cystic growth rate \geq 5 mm/year, elevated serum CA 19.9 concentration (> 37 U/mL), signs, enhancing mural nodules, and IPMNs > 40 mm regardless of the presence of other high-risk factors[22]. In BD-IPMNs, jaundice, high-grade dysplasia or cancer in cytology, a contrast-enhancing mural nodule (\geq 5 mm) or solid mass are absolute indications for surgery. The relative indications for surgery are the following: Growth rate \geq 5 mm/year, elevated serum CA 19.9 concentration (in the absence of jaundice), MPD diameter 5-9.9 mm, IPMN size \geq 40 mm, clinical manifestation (new-onset diabetes mellitus or AP), and contrastenhancing mural nodules[22].

In conclusion, according to all current guidelines, surgical treatment is recommended in all IPMNs involving the MPD, but there is still no consensus regarding MPD dilation. In the absence of other "high-risk stigmata", MPD dilatation alone is considered as a risk of misdiagnosis and possible overtreatment. Therefore, some authors suggested radiologic surveillance in patients with no symptoms and with "worrisome" MPD dilatation (5-9 mm) and without other HR stigmata[40]. All guidelines regarding current management in IPMN patients are presented in Table 3 [5,8,13,20-22,41].

Extent of surgical resection

According to Sendai guidelines[21], pancreatectomy with lymphadenectomy is necessary when invasive cancer is suspected. The type and extent of surgery depend on the IPMN location and extent[22]. The pancreatic head is the most frequent IPMN location. Therefore, pancreaticoduodenectomy (PD) is recommended in IPMNs located within the pancreatic head, uncinate process, and neck. Distal pancreatectomy (DP) is indicated for IPMNs located within the pancreatic body and tail. Total pancreatectomy (TP) is performed in exceptional cases when IPMN diffusely involves the whole pancreas or when a proximal IPMN extends through the distal pancreas. It is associated with the long-term consequences of TP, such as exocrine and endocrine pancreatic insufficiency requiring supplementation of pancreatic enzymes and diabetes treatment with insulin use. In each partial pancreatic resection, an assessment of the margin by frozen section is needed to confirm R0 resection with negative margins, and the resection should be extended in cases with cancer-positive surgical margins^[5].

According to the revised Fukuoka guidelines^[8], PD, DP, or TP according to the IPMN location and extent with lymphadenectomy should be the standard surgical treatment. Limited resections or even focal nonanatomic resections (excision, enucleation, uncinatectomy) can be performed in BD-IPMN not suspected for invasive cancer^[8]. The authors added that nonanatomic resections could be associated with infrequent but possible mucin leakage followed by peritoneal pseudomyxoma, a higher risk of postoperative pancreatic fistula and a risk of neoplasm recurrence. Standard pancreatectomy and lymphadenectomy should be performed if the cancer possibility is present[8]. We recommend using the European guidelines in decision making regarding the extent of IPMN surgery. According to the European guidelines [22], PD with frozen section investigations of the resection margins is recommended for patients with MPD dilatation comprising the entire pancreas. TP can be taken into consideration in patients with mural nodules within the MPD, and a higher cancer risk (familial pancreatic cancer). For BD-IPMNs, the authors recommend oncological resection with standard lymphadenectomy. It should be emphasized that parenchymasparing pancreatectomy is not an oncological procedure that can be performed only in lesions with a very low malignancy probability-for example, in patients without risk factors strongly wishing to be surgically treated. Due to a high malignancy risk, oncologic resection including standard lymphadenectomy is the recommended for IPMN with an absolute indication for resection. In multifocal BD-IPMN, each tumor should be assessed individually for the presence of malignancy-associated features. Patients with IPMNs with no concerning features can be observed [22].

Surveillance in IPMN patients

Patients with IPMNs lacking HRS/absolute indications should undergo nonoperative management. The surveillance strategies according to different guidelines are presented in Table 4[5,8,13,20-22,41].

According to the revised Fukuoka guidelines, surveillance is determined by IPMN diameter. The revised guidelines are more restrictive compared to the Fukuoka (2012) and Sendai guidelines (2006) and recommend initial surveillance performed at a shorter interval (within 6 mo for cysts < 20 mm and within 3-6 mo for cysts 2-3 cm). Following initial risk stratification, cysts < 10 mm should be radiologically monitored every 2 years in cysts with no changes. Cysts 10-20 mm should also be controlled



Table 3 Management of intraductal papillary mucinous neoplasm patients regarding indications for surgery according to the International, European and American Gastroenterological Association guidelines[5,8,13,20-22,41]

Guidelines	Management
IAP (2006)	Indications: Surgery
AGA (2015)	Indications: Surgery
IAP (2017)	High risk stigmata: Surgery
	Worrisome features: Surgery versus close surveillance based on: Patients' age/comorbidities: More aggressive management (surgery) in young patients
	EUS findings: Surgery indicated in clear MPD involvement and/or high-risk features
European (2018)	Absolute indications: Surgery
	Relative indications: Surgery according to criteria count, depending on comorbidities
	In fit patients: surgery for 1 criterion
	In patients with significant comorbidities: surgery for 2 criteria

IAP: International Association of Pancreatology; AGA: American Gastroenterological Association; EUS: Endoscopic ultrasonography; MPD: Main pancreatic duct.

Table 4 Surveillance in intraductal papillary mucinous neoplasm patients regarding indications for surgery according to the International, European and American Gastroenterological Association guidelines[5,8,13,20-22,41]

Guidelines	Indications	Investigations	Algorithm of follow-up
IAP (2006)	BD-IPMNs ≤ 30 mm; Without: Symptoms, mural nodules, positive cytology	MRI/MRCP or CT	Size ≤ 20 mm: every 6-12 mo; Size 20-30 mm: every 3-6 mo; The interval can be longer after 2 yr without changes
AGA (2015)	BD-IPMNs ≤ 30 mm; Without: Solid component, dilated MPD, HGD/cancer	MRI	Years 1, 2, 5 from initial diagnosis; It can be considered to discontinue; If there is no changes after years
IAP (2017)	No HRS/WF	MRI/MRCP, CT	Size < 10 mm: At 6 mo from diagnosis every 2 yr (if no change)
	No HRS/WF	MRI/MRCP, CT	Size 10-20 mm: At 6 mo from diagnosis yearly per 2 yr
	No HRS/WF	MRI/MRCP, EUS	Size 20-30 mm: EUS in 3-6 mo, yearly EUS or MRI
	No HRS, WF present and size < 30 mm	MRI/MRCPEUS	Every 3-6 mo EUS or MRI
European (2018)	No AI	MRI/MRCP or EUS, CA 19.9	Every 6 mo for the first year; Yearly after first year
	No AI, 1 RI in patient, with comorbidities	MRI/MRCP or EUS, CA 19.9	Every 6 mo

IPMN: Intraductal papillary mucinous neoplasm; IAP: International Association of Pancreatology; AGA: American Gastroenterological Association, MRI: Magnetic resonance imaging; MRCP: Magnetic resonance cholangiopancreatography; MPD: Main pancreatic duct; HGD: High grade dysplasia; EUS: Endoscopic ultrasonography; HRS: High risk stigmata; WF: Worrisome features; AI: Absolute indications for surgery; RI: Relative indications for surgery.

> radiologically every 2 years, EUS or MRI should be performed every 1 year in cysts 20-30 mm. A diameter change alone (\geq 5 mm growth in 2 years), in addition to the presence of any worrisome features, is sufficient to recommend systematic EUS[8,13, 20,41].

> The AGA guidelines[21] recommend surveillance for patients with BD-IPMNs < 30 mm, with no a solid component, dilated MPD, HGD or cancer in cytologic findings. In these patients, MRI should be performed in years 1, 2, and 5 from initial diagnosis. If no significant change occurs, surveillance discontinuation should be considered. Other patients should be referred to surgery[8,21].

> The authors for the European guidelines recommend a 6-mo follow-up (using MRI/MRCP and/or EUS and serum CA 19.9) in the first year and then yearly followup, in patients with a suspected IPMN that does not meet the indication for surgery.



Baishidena® WJGO | https://www.wjgnet.com

The guidelines recommend to continue observation as long as the patient remains surgically fit[8,22,39].

Follow-up after surgery

According to the revised Fukuoka guidelines[8], all IPMN patients, including those with noninvasive IPMNs with negative surgical margins, need follow-up after surgery to diagnose a new IPMN requiring surgery or pancreatic cancer. Tanaka et al[8] recommend continuing surveillance as long as the patient remains fit. In patients with higher risks, such as a family history of pancreatic cancer, HGD in surgical margins, and nonintestinal IPMN histological type, radiological investigations at least twice a year are recommended, and in others investigations every 6-12 mo should be performed. The follow-up of invasive IPMN should be the same as in pancreatic cancer [8].

The European guidelines are similar, and according to them[22], lifelong follow-up is recommended after IPMN resection as long as the patient is fit for surgery. Patients with IPMN-associated invasive cancer should be followed up in the same manner as those with resected pancreatic cancer. In HGD IPMN and MD-IPMN, follow-up every 6 mo for the first 2 years, followed by yearly surveillance is recommended. LGD IPMN should be observed in the same manner as nonresected IPMN. Patients with IPMN in the remnant pancreas with no HGD or MD-IPMN should be observed as nonresected BD-IPMN. In a postoperative observation, MRI or EUS are recommended^[22].

The AGA guidelines are very liberal. The authors recommend postoperative surveillance only for patients following surgery due to invasive IPMN. According to the AGA guidelines, patients with invasive cancer or dysplasia in the cyst after surgery should undergo MRI every 2 years. Moreover, the AGA did not recommend routine follow-up of IPMNs with no HGD or malignancy in the surgical specimen[21].

The clinical utility of the current guidelines regarding the management of IPMNs

Hsiao et al[42] evaluated the utility of the 2006 Sendai and 2012 Fukuoka guidelines in the differential diagnosis malignant and benign IPMNs. The study included 138 IPMN patients operated on between January 2000 and March 2015. Patients were "Sendai positive" if the tumor diameter was \geq 30 mm, with no symptoms, with mural nodules or a thickened wall, or with a dilated MPD of \geq 6 mm. Patients without above mentioned criteria were classified as "Sendai negative". Patients were characterized as "Fukuoka high risk" in the presence of: obstructive jaundice, or enhancing solid component, or MPD of ≥ 10 mm. "Fukuoka worrisome" were IPMNs with the presence of any worrisome features (pancreatitis, a tumor diamater of \geq 30 mm, a thickened/enhancing cyst wall, nonenhancing mural nodules, an abrupt MPD diamater change with distal pancreatic atrophy, and an MPD of 5-9 mm). The positive predictive value (PPV) and negative predictive value (NPV) of the Sendai and Fukuoka guidelines for HGD/IC were 35.1%, 43.3%, 100%, and 85.4%, respectively. According to the multivariate analysis, jaundice, tumors of ≥ 30 mm, presence of mural nodules, and age < 65 years were associated with HGD/invasive cancer in IPMN patients. There was a better NPV in the Sendai guidelines, but a better PPV in the Fukuoka guidelines. In the authors' opinion, a more aggressive management in patients with Fukuoka worrisome features couled be considered. The study showed that IPMNs of \geq 30 mm, but not pancreatitis, are associated with malignancy [42].

Pérez-Cuadrado-Robles *et al*[43] assessed the accuracy of the European guidelines in BD-IPMN patients indicated for surgery in a multicenter, observational, retrospective study including 91 patients with absolute (n = 21), relative (n = 60), or no formal indications (n = 10) for surgery. There were 60 patients with one (n = 35) or ≥ 2 relative indications (n = 25) for surgery in this study. The global advanced tumor and invasive cancer rates were 40% and 13.3%, respectively. There were not risk factors for GHD or invasive cancer. A lower risk of invasive cancer was reported in patients with one relative indication compared to patients with \geq 2 relative indications (5.7% vs 24%, respectively; P = 0.048). The advanced IPMN incidences were similar in the compared groups (37.1% *vs* 44%; *P* = 0.593)[43].

Jan et al[44] also validated the European guidelines for the management of IPMNs. The study included 158 patients with resected IPMNs between January 1994 and December 2016. All patients were stratified into three groups according to the European guidelines: Absolute, relative indications, and conservative approach. The missed rate for HGD/IC by the European guidelines was 1.9% (3 of 158). The sensitivity, specificity, positive and negative predictive values, and accuracy of the absolute or relative indications for resecting IPMN according to these criteria were 94.1%, 28.0%, 38.4%, 90.9%, and 49.4%, respectively. Jaundice, enhancing mural nodules < 5 mm, cyst diameter > 40 mm, elevated serum CA 19.9 concentration, new-



onset diabetes, and MPD dilation were associated with HGD/IC. Thus, the missed rate for HGD/IC was low using the European guidelines. Increased serum CA 19.9 and new-onset diabetes in European recommendations were verified as indications for the surgical resection of IPMNs[44].

Correa-Gallego et al[45] analyzed two independent nomograms to predict the findings of adenoma, high-grade dysplasia (HGD-CIS), and invasive carcinoma separately in both MD- and BD-IPMN. This study involved 219 patients including 56% of BD-IPMN in resected specimens. The significantly higher proportion of HGD-CIS was reported in MD-IPMN (33%) compared to BD-IPMN (15%) (P = 0.003). Invasive cancer was significantly more frequent in MD-IPMNs (41%) compared to BD-IPMNs (15%) (P < 0.001). In addition patient sex, history of prior malignancy, presence of a solid component, and weight loss were significantly associated with the ordinal outcome for MD-IPMN patients and were included in the nomogram (concordance index 0.74). For BD-IPMN patients, weight loss, solid component, and lesion diameter were associated with the outcome (concordance index 0.74)[45].

Capurso et al[46] investigated patient- and cyst-related factors associated with progression into WF or HRS categories of BD-IPMNs. This study included 540 patients diagnosed from 2009 to 2018 with at least 12 mo of surveillance until February 28, 2020. The revised Fukuoka criteria were used. Disease progression was noted in 130 (24.1%) patients. The probability of progression was 3.7% during 1 year, 23.4% during 5 years, and 43.3% during 10 years. Surgical treatment was performed in 15 (2.8%) patients. In 7 (1.3%) patients, cancer was found, and 3 (0.56%) patients died of pancreatic-associated disease. Initial cyst size > 15 mm, body mass index > 26.4 and heavy smoking were independent progression risk factors. The authors analyzed the association between AB0 blood group and progression risk. The higher association of AA group compared to 00 group with progression was also associated. The authors concluded that IPMN diameter alone is not a sufficient for the assessment of progression risk; however, it is useful in correlation with correlated with other features in observation of BD-IPMN patients[46].

Kwon et al^[47] validated the current guidelines on BD-IPMNs in a meta-analysis including 40 studies (6301 patients). In this meta-analysis, HGD or pancreatic cancer was significantly associated with clinical manifestation, cyst diameter \geq 30 mm, thickening of the cystic wall, mural nodules, MPD dilatation, abrupt MPD diameter changes, lymphadenopathy, increased CA 19.9 and increased CEA[47].

Srinivasan et al [48] published a systematic review to assess the clinical utility of the Sendai Consensus Guidelines and Fukuoka Consensus Guidelines for IPMNs. This review included 10 studies assessing the Fukuoka guidelines, 8 assessing the Sendai criteria and 4 assessing both guidelines. Pooled analysis showed that 751 of 1801 (42%) Fukuoka-positive neoplasms were malignant, and 599 of 697 (86%) Fukuoka-negative neoplasms were benign. The PPVs of the high-risk and worrisome-risk groups were 465/986 (47%) and 239/520 (46%), respectively, while 265 of 802 (33%) Sendai-positive neoplasms were malignant and 238 of 266 Sendai-negative (90%) neoplasms were benign. In conclusion, a higher PPV was noted in the Fukuoka compared to the Sendai criteria. However, the NPV of the Fukuoka guidelines was slightly lower compared to the Sendai guidelines. A higher PPV and lower NPV was reported in the Fukuoka compared to the Sendai criteria. Thus, malignant and even invasive IPMNs may be missed using both guidelines[48].

The participants of the Verona Evidence-Based Meeting on IPMN[49] assessed and compared the dissemination, use in clinical practice, and reliability of current guidelines for the management of PCNs. PCN classification as well as clinical and radiologic features were based on the IAP, European guidelines, and AGA recommendations. The answers to 47 questions were collected from 259 international responders, including participants from Europe (86%), Asia (8%), and the United States (6%). Among the responders, 58% were surgeons and 38% were gastroenterologists. The European guidelines were the best-known (79%), followed by IAP (69%) and AGA (61%) recommendations. The diagnostic investigations (MRI, CT, EUS, and cyst fluid analysis) were known by all participants; however, contrast-enhanced EUS was available only for 41% of responders. The analysis showed that guidelines were the most widely disseminated among surgeons and gastroenterologists, but the clinical application was decreased by the limited availability of diagnostic examinations. For example, contrast-enhanced EUS examination is not available for > 50% of physicians. Although enhancing mural nodules \geq 5 mm, considered high-risk stigmata, are absolute indications for surgery, according to >30% of physicians, this feature was not a sufficient indication for surgery. Therefore, according to Verona EBM experts, some questions (including the role of mural nodes in patients during follow-up, the correlation between imaging and histopathological findings, the optimal diameter



cutoff for the optimal assessment of the risk malignancy, and the most accurate imaging for optimal diagnosis) should be resolved. Despite of knowledge of the increased rate of malignant transformation in resected IPMNs with an MPD of diameter 5.0-9.9 mm, according to > 80% of responders, this feature was not a sufficient indication for surgery. Without prospective observational data on the observed IPMN, moderate MPD dilatation alone was not associated with an increased perception of cancer risk by clinicians. According to > 60% of responders, IPMN diameter and cyst growth rate were not enough indications for surgery. According to Verona EBM participants, further studies regarding IPMN-related symptoms as indications for surgery are needed. The guidelines should be more detailed to identify patients requiring surgery due to clinical presentation to avoid unnecessary surgery. The length of follow-up is also questionable. According to the AGA guidelines, surveillance should be discontinued after 5 years in patients with a stable pancreatic cystic neoplasm. Only 18% of responders would consider to discontinue observation after 5 years, but according to 54% of them, there is not enough evidence to recommend lifetime observation. Therefore, further studies assessing the most costeffective surveillance protocols and identifying the most suitable population for surveillance discontinuation are required. In addition, further studies, including randomized controlled trials, should identify patients requiring adjuvant treatment after surgery for invasive IPMNs. The authors of Verona EBM pointed to three levels of discrepancies regarding recommendations in pancreatic cystic neoplasms: among the 3 existing guidelines themselves, between guidelines and available evidence, and between guidelines and clinical practice. The role of MPD dilatation, mural nodules, tumor diameter and growth rate, tumor-associated clinical signs, and discontinuation of observation are the most important issues. According to experts, the current guidelines should be updated and unified to facilitate their use in clinical practice. The goal of Verona EBM participants was to define future research directions to increase the level of available evidence[49].

Prognosis of IPMN patients following surgery

The overall 5-year survival is reported to be 36%-77%. It depends on tumor advancement and the presence of malignant transformation in the resected tumor. The best prognosis is in benign IPMNs. The 5-year survival following surgery for noninvasive IPMN is 77%-100%. In malignant IPMNs, the prognosis is poorer. The 5-year survival rate following surgery for IPMN with invasive cancer is 27%-60%[5].

CONCLUSION

Clinical decision making for patients with pancreatic IPMNs is still challenging. While the management of MD-IPMN does not raise doubts and all guidelines require resection due to the high risk of malignant transformation, the management of BD-IPMN is controversial. The most important is the correct selection of patients requiring surgery at the right time, without unnecessarily exposing patients who do not require surgical treatment to complications related to pancreatic resection. It is known that pancreatectomy performed even in the most experienced centers is associated with the risk of complications. The correct algorithm of observation of patients not qualified for resection is also important. This review of the literature showed that the current guidelines are indeed useful in managing patients with IPMNs but are not ideal. Further prospective multicenter studies are needed to optimally select surgical candidates so that only those patients who need surgery are operated on and that treatment is avoided for the remaining patients who can be safely monitored.

REFERENCES

- Shyr YM, Su CH, Tsay SH, Lui WY. Mucin-producing neoplasms of the pancreas. Intraductal 1 papillary and mucinous cystic neoplasms. Ann Surg 1996; 223: 141-146 [PMID: 8597507 DOI: 10.1097/00000658-199602000-00005]
- 2 Jablońska B. Pancreatic cysts: etiology, diagnosis and management. Cent Eur J Med 2014; 9: 92-107 [DOI: 10.2478/s11536-013-0244-8]
- Ohashi K, Murakami Y, Maruyama M, Takekoshi T, Ohta H, Ohashi I. Four cases of mucussecreting pancreatic cancer. Prog Digest Endosc 1982; 20: 348-351 [DOI: 10.1111/j.1443-1661.2006.00656.x
- Morohoshi T, Kanda M, Asanuma K, Klöppel G. Intraductal papillary neoplasms of the pancreas. A



clinicopathologic study of six patients. Cancer 1989; 64: 1329-1335 [PMID: 2548703 DOI: 10.1002/1097-0142(19890915)64:6<1329::aid-cncr2820640627>3.0.co;2-s]

- Machado NO, Al Qadhi H, Al Wahibi K. Intraductal Papillary Mucinous Neoplasm of Pancreas. N 5 Am J Med Sci 2015; 7: 160-175 [PMID: 26110127 DOI: 10.4103/1947-2714.157477]
- 6 Adsay NV, Merati K, Andea A, Sarkar F, Hruban RH, Wilentz RE, Goggins M, Iocobuzio-Donahue C, Longnecker DS, Klimstra DS. The dichotomy in the preinvasive neoplasia to invasive carcinoma sequence in the pancreas: differential expression of MUC1 and MUC2 supports the existence of two separate pathways of carcinogenesis. Mod Pathol 2002; 15: 1087-1095 [PMID: 12379756 DOI: 10.1097/01.MP.0000028647.98725.8B]
- Crippa S, Arcidiacono PG, De Cobelli F, Falconi M. Review of the diagnosis and management of 7 intraductal papillary mucinous neoplasms. United European Gastroenterol J 2020; 8: 249-255 [PMID: 32213017 DOI: 10.1177/2050640619894767]
- Tanaka M, Fernández-Del Castillo C, Kamisawa T, Jang JY, Levy P, Ohtsuka T, Salvia R, Shimizu Y, Tada M, Wolfgang CL. Revisions of international consensus Fukuoka guidelines for the management of IPMN of the pancreas. Pancreatology 2017; 17: 738-753 [PMID: 28735806 DOI: 10.1016/j.pan.2017.07.007
- Castellano-Megías VM, Andrés CI, López-Alonso G, Colina-Ruizdelgado F. Pathological features and diagnosis of intraductal papillary mucinous neoplasm of the pancreas. World J Gastrointest Oncol 2014; 6: 311-324 [PMID: 25232456 DOI: 10.4251/wjgo.v6.i9.311]
- 10 Venkatesh PG, Navaneethan U, Vege SS. Intraductal papillary mucinous neoplasm and acute pancreatitis. J Clin Gastroenterol 2011; 45: 755-758 [PMID: 21602701 DOI: 10.1097/MCG.0b013e31821b1081
- 11 Jang JW, Kim MH, Jeong SU, Kim J, Park DH, Lee SS, Seo DW, Lee SK, Kim JH. Clinical characteristics of intraductal papillary mucinous neoplasm manifesting as acute pancreatitis or acute recurrent pancreatitis. J Gastroenterol Hepatol 2013; 28: 731-738 [PMID: 23301513 DOI: 10.1111/jgh.12121]
- 12 Bernardoni L, Crinò SF, De Conti G, Conti Bellocchi MC, De Pretis N, Amodio A, Frulloni L, Gabbrielli A. Preliminary experience with pancreatic sphincterotomy as treatment for intraductal papillary mucinous neoplasm-associated recurrent pancreatitis. Endosc Int Open 2017; 5: E1144-E1150 [PMID: 29124124 DOI: 10.1055/s-0043-119753]
- 13 Tanaka M. International consensus on the management of intraductal papillary mucinous neoplasm of the pancreas. Ann Transl Med 2015; 3: 286 [PMID: 26697446 DOI: 10.3978/j.issn.2305-5839.2015.11.09]
- Yamashita Y, Joshita S, Ito T, Maruyama M, Wada S, Umemura T. A case report of pancreatic 14 panniculitis due to acute pancreatitis with intraductal papillary mucinous neoplasm. BMC Gastroenterol 2020; 20: 286 [PMID: 32831035 DOI: 10.1186/s12876-020-01430-9]
- 15 Menzies S, McMenamin M, Barnes L, O'Toole D. Pancreatic panniculitis preceding acute pancreatitis and subsequent detection of an intraductal papillary mucinous neoplasm: A case report. JAAD Case *Rep* 2016; **2**: 244-246 [PMID: 27408933 DOI: 10.1016/j.jdcr.2016.05.001]
- Warndorf M, Hu H, Papachristou G, Zureikat A, Dasyam A, Yadav D. Intraductal papillary 16 mucinous neoplasm causing recurrent acute pancreatitis, necrotizing pancreatitis, and multifocal adenocarcinoma. Gastrointest Endosc 2014; 80: 1181-1182; discussion 1182 [PMID: 25434666 DOI: 10.1016/j.gie.2014.09.022]
- 17 Qian DH, Shen BY, Zhan X, Peng C, Cheng D. Liquefying panniculitis associated with intraductal papillary mucinous neoplasm. JRSM Short Rep 2011; 2: 38 [PMID: 21637399 DOI: 10.1258/shorts.2011.010141
- Gahr N, Technau K, Ghanem N. Intraductal papillary mucinous adenoma of the pancreas presenting 18 with lobular panniculitis. Eur Radiol 2006; 16: 1397-1398 [PMID: 16273371 DOI: 10.1007/s00330-005-0058-4]
- Wiesenauer CA, Schmidt CM, Cummings OW, Yiannoutsos CT, Howard TJ, Wiebke EA, Goulet RJ 19 Jr, McHenry L, Sherman S, Lehman GA, Cramer H, Madura JA. Preoperative predictors of malignancy in pancreatic intraductal papillary mucinous neoplasms. Arch Surg 2003; 138: 610-617; discussion 617-618 [PMID: 12799331 DOI: 10.1001/archsurg.138.6.610]
- Tanaka M, Chari S, Adsay V, Fernandez-del Castillo C, Falconi M, Shimizu M, Yamaguchi K, 20 Yamao K, Matsuno S; International Association of Pancreatology. International consensus guidelines for management of intraductal papillary mucinous neoplasms and mucinous cystic neoplasms of the pancreas. Pancreatology 2006; 6: 17-32 [PMID: 16327281 DOI: 10.1159/000090023]
- Vege SS, Ziring B, Jain R, Moayyedi P; Clinical Guidelines Committee; American Gastroenterology 21 Association. American gastroenterological association institute guideline on the diagnosis and management of asymptomatic neoplastic pancreatic cysts. Gastroenterology 2015; 148: 819-822; quiz e12-13 [PMID: 25805375 DOI: 10.1053/j.gastro.2015.01.015]
- 22 European Study Group on Cystic Tumours of the Pancreas. European evidence-based guidelines on pancreatic cystic neoplasms. Gut 2018; 67: 789-804 [PMID: 29574408 DOI: 10.1136/gutjnl-2018-316027]
- Takeshita K, Kutomi K, Takada K, Haruyama T, Fukushima J, Aida R, Takada T, Furui S. 23 Differential diagnosis of benign or malignant intraductal papillary mucinous neoplasm of the pancreas by multidetector row helical computed tomography: evaluation of predictive factors by logistic regression analysis. J Comput Assist Tomogr 2008; 32: 191-197 [PMID: 18379300 DOI: 10.1097/RCT.0b013e3180676d97]



- 24 Nakagawa A, Yamaguchi T, Ohtsuka M, Ishihara T, Sudo K, Nakamura K, Hara T, Denda T, Miyazaki M. Usefulness of multidetector computed tomography for detecting protruding lesions in intraductal papillary mucinous neoplasm of the pancreas in comparison with single-detector computed tomography and endoscopic ultrasonography. Pancreas 2009; 38: 131-136 [PMID: 18981954 DOI: 10.1097/MPA.0b013e31818b0040
- 25 Tan L, Zhao YE, Wang DB, Wang QB, Hu J, Chen KM, Deng XX. Imaging features of intraductal papillary mucinous neoplasms of the pancreas in multi-detector row computed tomography. World J Gastroenterol 2009; 15: 4037-4043 [PMID: 19705500 DOI: 10.3748/wjg.15.4037]
- 26 Murayama S, Kimura W, Hirai I, Takasu N, Takeshita A, Moriya T. Volumetric and morphological analysis of intraductal papillary mucinous neoplasm of the pancreas using computed tomography and magnetic resonance imaging. Pancreas 2011; 40: 876-882 [PMID: 21747312 DOI: 10.1097/MPA.0b013e31821fdcff
- Mönnings P, Belyaev O, Uhl W, Giese A, Tannapfel A, Köster O, Meier JJ. Criteria for Determining 27 Malignancy in Pancreatic Intraductal Papillary Mucinous Neoplasm Based on Computed Tomography. Digestion 2016; 94: 230-239 [PMID: 28030856 DOI: 10.1159/000452738]
- 28 Sodickson A, Baeyens PF, Andriole KP, Prevedello LM, Nawfel RD, Hanson R, Khorasani R. Recurrent CT, cumulative radiation exposure, and associated radiation-induced cancer risks from CT of adults. Radiology 2009; 251: 175-184 [PMID: 19332852 DOI: 10.1148/radiol.2511081296]
- 29 Min JH, Kim YK, Kim SK, Kim H, Ahn S. Intraductal papillary mucinous neoplasm of the pancreas: diagnostic performance of the 2017 international consensus guidelines using CT and MRI. Eur Radiol 2021 [DOI: 10.1007/s00330-020-07583-1]
- 30 Liu H, Cui Y, Shao J, Shao Z, Su F, Li Y. The diagnostic role of CT, MRI/MRCP, PET/CT, EUS and DWI in the differentiation of benign and malignant IPMN: A meta-analysis. Clin Imaging 2021; 72: 183-193 [PMID: 33321460 DOI: 10.1016/j.clinimag.2020.11.018]
- Jeon SK, Kim JH, Yoo J, Kim JE, Park SJ, Han JK. Assessment of malignant potential in intraductal 31 papillary mucinous neoplasms of the pancreas using MR findings and texture analysis. Eur Radiol 2021; **31**: 3394-3404 [PMID: 33140171 DOI: 10.1007/s00330-020-07425-0]
- Boraschi P, Tarantini G, Donati F, Scalise P, Cervelli R, Caramella D. Side-branch intraductal 32 papillary mucinous neoplasms of the pancreas: outcome of MR imaging surveillance over a 10 years follow-up. Eur J Radiol Open 2020; 7: 100250 [PMID: 32884981 DOI: 10.1016/j.ejro.2020.100250]
- 33 Choi SY, Kim JH, Yu MH, Eun HW, Lee HK, Han JK. Diagnostic performance and imaging features for predicting the malignant potential of intraductal papillary mucinous neoplasm of the pancreas: a comparison of EUS, contrast-enhanced CT and MRI. Abdom Radiol (NY) 2017; 42: 1449-1458 [PMID: 28144718 DOI: 10.1007/s00261-017-1053-3]
- McCarty TR, Paleti S, Rustagi T. Molecular analysis of EUS-acquired pancreatic cyst fluid for 34 KRAS and GNAS mutations for diagnosis of intraductal papillary mucinous neoplasia and mucinous cystic lesions: a systematic review and meta-analysis. Gastrointest Endosc 2021; 93: 1019-1033.e5 [PMID: 33359054 DOI: 10.1016/j.gie.2020.12.014]
- 35 Kadayifci A, Atar M, Wang JL, Forcione DG, Casey BW, Pitman MB, Brugge WR. Value of adding GNAS testing to pancreatic cyst fluid KRAS and carcinoembryonic antigen analysis for the diagnosis of intraductal papillary mucinous neoplasms. Dig Endosc 2017; 29: 111-117 [PMID: 27514845 DOI: 10.1111/den.12710
- Lee JH, Kim Y, Choi JW, Kim YS. KRAS, GNAS, and RNF43 mutations in intraductal papillary 36 mucinous neoplasm of the pancreas: a meta-analysis. Springerplus 2016; 5: 1172 [PMID: 27512631 DOI: 10.1186/s40064-016-2847-4]
- Gillis A, Cipollone I, Cousins G, Conlon K. Does EUS-FNA molecular analysis carry additional 37 value when compared to cytology in the diagnosis of pancreatic cystic neoplasm? HPB (Oxford) 2015; 17: 377-386 [PMID: 25428782 DOI: 10.1111/hpb.12364]
- 38 Jin DX SA, Vollmer CM, Jhala N, Furth E, Ginsberg G, Kochman M, Ahmad N, Chandrasekhara V. A lower cyst fluid CEA cut-off increases diagnostic accuracy in identifying mucinous pancreatic cystic lesions. J Pancreas 2015; 16: 271-277 [DOI: 10.1016/s0016-5085(13)62938-8]
- 39 Farrell JJ. Pancreatic Cysts and Guidelines. Dig Dis Sci 2017; 62: 1827-1839 [PMID: 28528374 DOI: 10.1007/s10620-017-4571-5]
- 40 Dal Borgo C, Perri G, Borin A, Marchegiani G, Salvia R, Bassi C. The Clinical Management of Main Duct Intraductal Papillary Mucinous Neoplasm of the Pancreas. Dig Surg 2019; 36: 104-110 [PMID: 29421807 DOI: 10.1159/000486869]
- 41 Hasan A, Visrodia K, Farrell JJ, Gonda TA. Overview and comparison of guidelines for management of pancreatic cystic neoplasms. World J Gastroenterol 2019; 25: 4405-4413 [PMID: 31496620 DOI: 10.3748/wjg.v25.i31.4405]
- 42 Hsiao CY, Yang CY, Wu JM, Kuo TC, Tien YW. Utility of the 2006 Sendai and 2012 Fukuoka guidelines for the management of intraductal papillary mucinous neoplasm of the pancreas: A singlecenter experience with 138 surgically treated patients. Medicine (Baltimore) 2016; 95: e4922 [PMID: 27661043 DOI: 10.1097/MD.000000000004922]
- Pérez-Cuadrado-Robles E, Uribarri-González L, Borbath I, Vila JJ, López-López S, Deprez PH. Risk of advanced lesions in patients with branch-duct IPMN and relative indications for surgery according to European evidence-based guidelines. Dig Liver Dis 2019; 51: 882-886 [PMID: 30591368 DOI: 10.1016/j.dld.2018.11.028]
- 44 Jan IS, Chang MC, Yang CY, Tien YW, Jeng YM, Wu CH, Chen BB, Chang YT. Validation of Indications for Surgery of European Evidence-Based Guidelines for Patients with Pancreatic



Intraductal Papillary Mucinous Neoplasms. J Gastrointest Surg 2020; 24: 2536-2543 [PMID: 31745906 DOI: 10.1007/s11605-019-04420-9]

- 45 Correa-Gallego C, Do R, Lafemina J, Gonen M, D'Angelica MI, DeMatteo RP, Fong Y, Kingham TP, Brennan MF, Jarnagin WR, Allen PJ. Predicting dysplasia and invasive carcinoma in intraductal papillary mucinous neoplasms of the pancreas: development of a preoperative nomogram. Ann Surg Oncol 2013; 20: 4348-4355 [PMID: 24046103 DOI: 10.1245/s10434-013-3207-z]
- Capurso G, Crippa S, Vanella G, Traini M, Zerboni G, Zaccari P, Belfiori G, Gentiluomo M, 46 Pessarelli T, Petrone MC, Campa D, Falconi M, Arcidiacono PG. Factors Associated With the Risk of Progression of Low-Risk Branch-Duct Intraductal Papillary Mucinous Neoplasms. JAMA Netw Open 2020; 3: e2022933 [PMID: 33252689 DOI: 10.1001/jamanetworkopen.2020.22933]
- 47 Kwon W, Han Y, Byun Y, Kang JS, Choi YJ, Kim H, Jang JY. Predictive Features of Malignancy in Branch Duct Type Intraductal Papillary Mucinous Neoplasm of the Pancreas: A Meta-Analysis. Cancers (Basel) 2020; 12 [PMID: 32937809 DOI: 10.3390/cancers12092618]
- Srinivasan N, Teo JY, Chin YK, Hennedige T, Tan DM, Low AS, Thng CH, Goh BKP. Systematic 48 review of the clinical utility and validity of the Sendai and Fukuoka Consensus Guidelines for the management of intraductal papillary mucinous neoplasms of the pancreas. HPB (Oxford) 2018; 20: 497-504 [PMID: 29486917 DOI: 10.1016/j.hpb.2018.01.009]
- 49 Marchegiani G, Salvia R; Verona EBM 2020 on IPMN. Guidelines on Pancreatic Cystic Neoplasms: Major Inconsistencies With Available Evidence and Clinical Practice- Results From an International Survey. Gastroenterology 2021; 160: 2234-2238 [PMID: 33609506 DOI: 10.1053/j.gastro.2021.02.026]



0 WJ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 1896-1918

DOI: 10.4251/wjgo.v13.i12.1896

ISSN 1948-5204 (online)

REVIEW

Combined treatments in hepatocellular carcinoma: Time to put them in the guidelines?

Zeno Sparchez, Pompilia Radu, Adrian Bartos, Iuliana Nenu, Rares Craciun, Tudor Mocan, Adelina Horhat, Mihaela Spârchez, Jean-François Dufour

ORCID number: Zeno Sparchez 0000-0002-3813-1677; Pompilia Radu 0000-0001-7208-1477: Adrian Bartos 0000-0003-3177-9232; Iuliana Nenu 0000-0002-1690-6689; Rares Craciun 0000-0002-5872-8630; Tudor Mocan 0000-0001-7785-6403; Adelina Horhat 0000-0002-8701-8750; Mihaela Spârchez 0000-0001-8620-9160; Jean-François Dufour 0000-0002-8062-1346.

Author contributions: Sparchez Z, Radu P, Bartos A, Nenu I, Craciun R, Mocan T, and Horhat A wrote the manuscript; Sparchez Z, Nenu I, Bartos A, and Radu P performed the literature search; Sparchez Z, Dufour JF critically reviewed the review, the manuscript; all authors have read and approve the final manuscript.

Conflict-of-interest statement: Jean-

François Dufour: Advisory committees: Abbvie, Bayer, BMS, Falk, Genfit, Genkyotex, Gilead Science, HepaRegenix, Intercept, Lilly, Merck, Novartis. Speaking and teaching: Abbvie, Bayer, BMS, Genfit, Gilead Science, Novartis.

Country/Territory of origin: Switzerland

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer Zeno Sparchez, Iuliana Nenu, Rares Craciun, Tudor Mocan, Adelina Horhat, 3rd Medical Department, "Iuliu Hatieganu" University of Medicine and Pharmacy, Institute for Gastroenterology and Hepatology, Cluj-Napoca 400162, Romania

Pompilia Radu, Department of Visceral Surgery and Medicine, Hepatology, Inselspital, Bern University Hospital, University of Bern, Bern 3010, Switzerland

Adrian Bartos, Department of Surgery, "Ïuliu Hatieganu" University of Medicine and Pharmacy, Institute for Gastroenterology and Hepatology, Cluj-Napoca 400162, Romania

Mihaela Spârchez, Department of Mother and Child, 2nd Paediatric Clinic, "Iuliu Hatieganu" University of Medicine and Pharmacy, Cluj-Napoca 400177, Romania

Jean-François Dufour, Department for BioMedical Research, Hepatology, University of Bern, Bern 3008, Switzerland

Corresponding author: Pompilia Radu, MD, PhD, Staff Physician, Department of Visceral Surgery and Medicine, Hepatology, Inselspital, Bern University Hospital, University of Bern, Freiburgstrasse 18, Bern 3010, Switzerland. radupompilia@yahoo.com

Abstract

The time for battling cancer has never been more suitable than nowadays and fortunately against hepatocellular carcinoma (HCC) we do have a far-reaching arsenal. Moreover, because liver cancer comprises a plethora of stages-from very early to advanced disease and with many treatment options-from surgery to immunotherapy trials-it leaves the clinician a wide range of options. The scope of our review is to throw light on combination treatments that seem to be beyond guidelines and to highlight these using evidence-based analysis of the most frequently used combination therapies, discussing their advantages and flaws in comparison to the current standard of care. One particular combination therapy seems to be in the forefront: Transarterial chemoembolization plus ablation for medium-size non-resectable HCC (3-5 cm), which is currently at the frontier between Barcelona Clinic Liver Cancer classification A and B. Not only does it improve the outcome in contrast to each individual therapy, but it also seems to have similar results to surgery. Also, the abundance of immune checkpoint inhibitors that have appeared lately in clinical trials are bringing promising results against HCC. Although the path of combination therapies in HCC is still filled with uncertainty and caveats, in the following years the hepatology and oncology



reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): A Grade B (Very good): 0 Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: March 3, 2021 Peer-review started: March 3, 2021 First decision: April 19, 2021 **Revised:** May 3, 2021 Accepted: November 5, 2021 Article in press: November 5, 2021 Published online: December 15, 2021

P-Reviewer: Aoki H S-Editor: Fan JR L-Editor: A P-Editor: Fan JR



fields could witness an HCC guideline revolution.

Key Words: Hepatocellular carcinoma; Transarterial chemoembolization; Radiofrequency ablation; Microwave ablation; Systemic therapy; Immunotherapy combined treatments

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: The field of hepatocellular carcinoma has become highly interesting in recent years, given the emergence of a large amount of data on immunotherapy and combination treatment strategies. In this light, the current clinical practice guidelines may appear restrictive, especially in borderline cases, which have become a significant challenge in tumor boards across the world. The current review is designed to provide an exhaustive analysis of the most notable advances in the field, focusing on combination therapies and their role in the therapeutic algorithm, with the ultimate goal of aiding clinicians to navigate the Barcelona clinic liver cancer classification maze.

Citation: Sparchez Z, Radu P, Bartos A, Nenu I, Craciun R, Mocan T, Horhat A, Spârchez M, Dufour JF. Combined treatments in hepatocellular carcinoma: Time to put them in the guidelines? World J Gastrointest Oncol 2021; 13(12): 1896-1918 URL: https://www.wjgnet.com/1948-5204/full/v13/i12/1896.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.1896

INTRODUCTION

Hepatocellular carcinoma (HCC) is the most common primary liver cancer, ranking sixth overall among malignancies in incidence and, disproportionately, fourth in mortality[1]. It is a multifaceted disease, atypical among cancers due to its intricate and non-linear prognostic indicators, as survival is closely intertwined with the tumor extension, the severity of the underlying liver disease, and overall fitness. To this point, there is a wide array of available techniques in the therapeutic arsenal against HCC. The options range from curative-intent solutions such as surgery or transplantation to local ablation, interventional radiology, and systemic therapies[2,3]. However, despite recent advances and potentially game-changing developments, HCC is still associated with a poor prognosis, with an incidence to mortality ratio dismally approaching number one[4].

Currently, the most frequently employed algorithm for standardizing care is the Barcelona Clinic Liver Cancer Classification (BCLC) and its subsequent updates[5]. Arguably the best algorithm to this point and backed by extensive validation, it still poses significant clinical dilemmas, especially for cases that do not fit perfectly in its boxes. In such scenarios, the comfort of evidence-based guideline recommendations tends to fade, leaving both the clinicians and the patients in uncharted waters, seeking the best path forward.

Consequently, multiple approaches have been attempted, combining available techniques in various shapes and forms with the ultimate goal of improving the overall outcome. These combinations try to negate the individual deficiencies of each method while augmenting their strengths, hoping to provide a perfect match for any specific clinical scenario. Ranging from an early tumor in an advanced, decompensated liver disease, to an advanced tumor in an otherwise relatively normal liver and anything in-between, the severity spectrum of HCC leaves room for epistemic, databased improvisation. However, as the range of therapies is ever-increasing, there is a thin line between being too conservative and overtreating, as both extremes could lead to additional harm and cost. The current review aims to provide an evidence-based analysis of the most frequently used combination therapies, discussing their advantages and caveats in comparison to the current standard of care.

COMBINED TREATMENT TO FACILITATE CURE

Hepatic resection plus intraoperative ablation

Although surgical resection (SR) still represents the ideal and best option as treatment, having curative potential, unfortunately for patients diagnosed with liver malignancies, this treatment is feasible in only 10%-20% of cases[6]. The remaining 80%-90% of patients, which are not suitable for radical intervention, include patients with multiple tumors, located in both hepatic lobes with insufficient hepatic reserve[7]. In the case of HCC, according to the guidelines used in clinical practice (BCLC staging system), the patients with unresectable multicentric neoplasia and preserved liver function fall into stage B, being candidates only for transarterial chemoembolization (TACE)[8]. However, the results of various studies from the literature are beginning to support the use of combined techniques for this category of patients[9]. According to the literature, there are several radical options for bilobar localization, including two-stage resections, ALPPS technique (associating liver partition and portal vein ligation for staged hepatectomy), and combined techniques: Hepatic resections and ablative techniques[9].

Thereby, this complex treatment, which involves the combination of hepatectomies with simultaneous tumor ablations, has the role of increasing the proportion of patients who can become candidates for a radical, potentially curative treatment. However, although there are numerous reports in the literature underlining the clinical outcomes, a standard conduit and a therapeutic consensus in this field have not been yet established[6,10].

General indications for combined therapy are patients with multicentric, bilobar, unresectable neoplasms who are not candidates for curative resection but present with a compensated liver disease[11,12]. Other indications include inoperable tumors due to proximity to major vascular structures and/or the presence of liver cirrhosis with functional liver parenchyma, but not being able to tolerate a major resection[6,10]. Ablative techniques are indicated even if the tumors are in the proximity of a main portal branch, hepatic vein, or inferior vena cava[12].

The extrahepatic presence of neoplasia represents a contraindication for combined treatment, although there are authors who advocate for this treatment in the case of the associated resectable lung tumors or local invasion from liver tumors (in the diaphragm or adrenal gland)[9]. The association of ablation is not indicated when the tumor involves the right or left liver duct[12]. Incontestably, patients with decompensated liver disease, refractory ascites, coagulation disorders, and/or low-performance status (PS) cannot benefit from the combined resection-ablation treatment.

The intraoperative technique involves performing simultaneously, under general anesthesia and by laparotomy, both liver resections, and HCC ablation sessions. Most authors recommend performing resection first, followed by ultrasound-guided ablation, most commonly by tissue destruction by radiofrequency ablation (RFA)[6]. Microwave ablation (MWA) comes with some advantages, these being cited by some authors, but with a lower usage than RFA[9]. Although feasible, with comparable outcomes with "conventional" open surgery, the combined treatment performed by a laparoscopic approach is not a standardized technique, with only a few reports being found in the literature[13,14].

To exclude extrahepatic neoplasia it is mandatory to do a complete exploration of the entire abdominal cavity and intraoperative hepatic ultrasound (IOUS) to assess the topography and tumor relationships[15,16]. Anatomical resections are preferred whenever possible[17]. The indication for atypical, minor (1-2 segments), or major (> 3 segments) hepatectomy is determined by the preoperative assessment of the liver function, of the patient status, and by tumor extent. Also, to prevent blood loss, intermittent clamping of the afferent hepatic pedicle (Pringle maneuver) may be necessary. According to Qiu *et al*[6], it is required in less than 50% of cases[6]. Another advantage of this maneuver is the reduction of the cooling effect induced by the proximity of tumors to main vascular branches ("heat sink"), the rate of achieving a complete ablation being higher[18]. Evaluation of the necrotic area and the efficacy of RFA can be performed immediately by contrast-enhanced-IOUS (CE-IOUS), making possible repeated ablation sessions immediately[19].

Doing a literature survey guided by the words "hepatocellular carcinoma", "combined", "liver resection" and "ablative treatment", using the PubMed database for titles in English published from 2010 to 2020, we found that the mortality reported for the surgery-ablative combined technique is around 1%-2% with a general incidence of complications in the range 22%-70% [6,20-25] (Table 1).

Zaishideng® WJGO | https://www.wjgnet.com

Table 1 Morbidity, mortality, recurrence and survival after hepatic resection plus intraoperative ablation							
Ref.	Patients, <i>n</i> (%)	MO, <i>n</i> (%)	Mo, <i>n</i> (%)	Recurrence	Survival rates (1 yr/3 yr/5 yr)		
Qiu et al[6], 2014	112	2	22.3	72.3%	67.5%/32.5%/12.5%		
Hou <i>et al</i> [23], 2016	51	0	70.6	54.9%	88.2%/66.7%/52.9%		
Zhang et al[24], 2014	114	0.9	-	-	34.4%/70.7%/40.7%		
Huang et al[25], 2020	51	-	-	-	86.3%/66.6%/34.2%		

MO: Mortality; Mo: Morbidity.

In general, the literature supports the combined technique as feasible and safe, although the cited recurrence rate is high[6,20,23] (Table 1). The rate of major complications is reported to be around 15%, with a rate of acute liver failure of 1.8% and postoperative bleeding of 0.9%[6,21]. Other specific complications are biliary leaks (8.9%), postoperative ascites (11.6%), perihepatic abscesses (1.8%). 0.9% of patients with post-operative complications may require reinterventions[6,21]. No significant differences were reported between the rates of complications after combined techniques and conventional liver resections. Doing a literature survey guided by the words "hepatocellular carcinoma", "combined", "liver resection" and "ablative treatment", using the PubMed database for titles in English published from 2010 to 2020, we found that the mortality reported for the surgery-ablative combined technique is around 1%-2% with a general incidence of complications in the range 22%-70%[6,20,21].

In regards to long-term survival, we found a range between 12% and 88%, depending on 1-, 3- and 5-year survival reports (Table 1). As can be seen, there is a wide range of results, most likely explained by the lack of standardization of the combined procedures and by the contribution of the case selection bias.

Most of the authors concluded that whenever resection can be performed, it must be chosen instead of ablation. However, RFA remains a feasible alternative in cases that are not suitable for resection, except for large tumors, over 5 cm[22]. Moreover, whenever possible, combined therapy should be indicated to the detriment of TACE, the latter being followed by a shorter 5-years survival: 52.9% (combined treatment) vs 9.8% (TACE)[23].

The results are more optimistic when comparing combined treatment with resections only, with overall survival (OS) at 1-, 3- and 5-years being comparable between the two groups: 86.3%, 66.6% and 34.2% vs 92.8%, 67% and 37%, respectively (P = 0.4)[6,21,24,26].

Supported by the data mentioned in the literature above, the combined treatment (resection plus ablation) is a feasible alternative to the therapeutic options already existing in current guidelines. The fact that more and more studies highlight the increase in the number of curative resections by using these techniques will certainly support their integration into the standard conducting algorithm for HCC.

Combination of locoregional therapies for early and intermediate stage including comparison with other therapies

Locoregional therapies (LRT) include TACE and local ablation techniques. Local ablation including percutaneous ethanol injections, RFA, MWA, laser- and cryoablation (CA) are considered alternative curative methods for early-stage HCC[27,28]. As 75% of HCCs nodules are inoperable at the time of the diagnosis, TACE plays an important role in the management of unresectable HCC and is considered the first-line therapy for BCLC stage B HCC based on Barcelona Clinic Liver Cancer guidelines 27, 28].

Limits of LRT and rationale for combination therapy: For some patients undergoing TACE procedure the tumor necrosis rate is low with consequent frequent tumoral residue and high intrahepatic recurrence^[27,29]. Along these lines there are several reasons for this limited efficacy: (1) The difficulty to embolize all the feeding arteries of the tumor; (2) Recanalization and angiogenesis which may occur after TACE with consequent tumor recurrence and metastasis; and (3) The re-establish of collateral circulation^[27]. Ablations techniques like RFA present a high performance in tumors below 2-3 cm where complete necrosis may be achieved in up to 90% of cases[21]. However, RFA is less effective in medium-sized (3-5 cm) or large tumors (5-9 cm)



where the efficacy dismally drops to 61% and 24% respectively^[21]. When RFA was performed for lesions located near major vessels a heat sink effect was reported leading to an increased recurrence rate as well.

In this regard, the location of the lesion is a crucial factor when considering ablation on the grounds that some lesions cannot be successfully treated with thermal ablation without damaging adjacent structures (bile ducts, colon, diaphragm)[30]. Thereby, one intention-to-treat analysis found that 9% of small HCCs were not amenable to percutaneous ablation because of their location[30].

The rationale for combining TACE with ablation is to maximize the percentage of complete tumor response rate and thereby to reduce local recurrence rate due to incomplete or inadequate treatment of the adjacent hepatic parenchyma[9]. As follows, the synergy of the therapies leads to larger volumes of destructed tumoral tissue with consequent efficient treatment of presumed microsatellite nodules and microvascular invasion[9,27]. The sequencing of combined therapies is controversial, with most authors preferring TACE followed by RFA, although some prefer RFA followed by TACE or both techniques in the same session[9,27]. The theoretical advantages of performing TACE before ablation include (1) TACE reduces hepatic artery blood flow, thus diminishing heat sink effects and maximizing the size of the ablation zone; and (2) TACE can detect satellite lesions not seen on cross-sectional imaging[9,27].

TACE combined with RFA vs TACE: There are several papers published on this combination with different clinical scenarios. The most important ones are presented in Table 2.

By analyzing these studies, it was discovered that combination therapy might be beneficial compared to TACE alone, or RFA alone in several clinical scenarios.

BCLC-A patients: Song et al^[31] have compared the results of 71 patients with HCC within Milan criteria treated by TACE to 87 and 43 patients treated by TACE + RFA and respectively RFA. The combination therapy yielded a significantly higher complete response rate according to mRECIST criteria in comparison to TACE (96.5% vs 81.6%, P = 0.019) and a lower rate of local tumor progression at 1, 3 and 5 years (6%, 33%, and 45% vs 17%, 58% and 78%). Nonetheless, what is surprising is the fact that OS was significantly higher for TACE + RFA vs TACE or RFA alone for lesions below 3 cm but not for lesions larger than 3 cm[31]. Similar results were reported as well by Lee et al[32] with lower recurrence rate at 1 and 3 years 7.3% and 16.5% vs 12.5% and 31% but no difference in OS[32].

BCLC-B patients: Liu *et al*[33] have compared a B1 HCC population treated by TACE (195 pts) vs TACE + RFA (209 pts) and found a significant difference in favor of combination therapy regarding progression-free survival (PFS) and OS[33]. Also, the same results [significantly higher median OS-840 vs 2466 d and median time-to-tumor progression (TTP)-140 vs 1148 d] were communicated by Hirooka et al[34] although on a small sample of patients[34]. Likewise, Ren et al[35] have investigated a large BCLC-A and B population treated either by TACE (271 pts) or TACE + RFA (128 pts) and concluded by all means that the combination therapy is significantly superior to TACE in terms of PFS and both median and cumulative OS[35]. The superiority of TACE + RFA over TACE in terms of longer OS and PFS was demonstrated also for medium and large nonresectable HCCs in 2 large recent trials[36,37].

The combination therapy was assessed also in patients with small lesions not feasible for RFA and lesions in unreachable locations (e.g., near the hepatic hilum, subdiaphragmatic, subcapsular). TACE procedures resolve this dilemma due to intratumoral accumulation of radio-opaque iodized oil used and give radiographic contrast to a small tumor either of poor conspicuity or even ultrasound (US) blind and also difficult to target spots such as hepatic dome^[27]. An undoubtedly higher complete response rate (100% vs 54%, P < 0.01) and lower TTP rate were revealed by Hyun *et al* [38] in a series of patients with HCC not feasible for US-guided RFA treated by TACE (54 pts) or TACE + RFA (37 pts)[38]. However, Yang et al[39] compared the efficacy of combination therapy in 37 patients with HCC in special locations to 85 patients with HCC in convenient locations and found no differences in PFS and OS[39]. Three metaanalyses concerning the comparison of TACE + RFA vs TACE have been already published[40-42]. The last one issued in 2017 found that for intermediate-stage HCC, TACE plus RFA attained higher tumor response rates (OR = 6.08, 95% CI: 4.00-9.26, P < 0.00001), achieved longer recurrence-free survival (RFS) rates (ORRFS = 3.78, 95%CI: 2.38-6.02, *P* < 0.00001) and OS rates (OR1-year = 3.92, 95%CI: 2.41-6.39, *P* < 0.00001; OR3-year = 2.56; 95%CI: 1.81-3.60; *P* < 0.00001; OR5-year = 2.78; 95%CI: 1.77-4.38; *P* < 0.0001) when comparing to TACE alone. Unfortunately, as expected the number of

Table 2 Comparison of transarterial chemoembolization plus radiofrequency ablation to transarterial chemoembolization

Ref.	Treatment type, <i>n</i> (%)	Clinical scenario	Response rate mRECIST	Outcome
Morimoto <i>et al</i> [111], 2013	TACE + RFA (132)	HCC 1-5 cm, subcapsular	98.5% CR	LTP (3 yr) 9.7%. OS (3, 5, 7 yr): 79.3%, 60.6%, 50.9%
Song <i>et al</i> [31], 2016	TACE (71) vs TACE + RFA (87) vs RFA (43)	HCC within Milan	81.6% vs 96.5% vs 97.6% (TACE vs TACE + RFA P = 0.019)	LTR (1, 3, 5 yr): 17%, 58%, 78% vs 6%, 33%, 54% vs 10%, 31%, 48% (TACE + RFA vs TACE P = 0.015; RFA vs TACE P = 0.005). OS (1, 3, 5 yr): 98%, 90%, 83% vs 98%, 95%, 90% vs 94%, 84%, 71% OS significantly higher (P = 0.019) for TACE + RFA vs TACE or RFA for lesions < 3 cm but not for lesions > 3 cm
Lee <i>et al</i> [<mark>32</mark>], 2018	TACE (85) <i>vs</i> TACE + RFA (<i>n</i> = 82)	HCC BCLC 0 or A invisible for ultrasound	97.6% vs 100% (CR)	LTP (1, 3, 5, 7 yr): 12.5%, 31%, 37% <i>vs</i> 7.3%, 16.5%, 16.5% (<i>P</i> = 0.013). Median TTP: 18 mo <i>vs</i> 24 mo (<i>P</i> = 0.037). OS (1, 3, 5 yr): 100%, 93.2%, 87.7% <i>vs</i> 100%, 96.6%, 87.4% (<i>P</i> = 0.686)
Liu et al <mark>[33</mark>], 2019	TACE (195) vs TACE + RFA (209)	HCC B1	N/A	Median PFS: 14 mo vs 20 mo. PFS (1, 3, 5 yr): 59.1%, 11.0%, 2.2% vs 71.8%, 26.6%, 13.0% ($P < 0.001$). OS (1, 3, 5 yr): 80.7%, 26.4%, 6.7% vs 83.7%, 45.8%, 24.8% ($P = 0.003$)
Hiraoka <i>et al</i> [73], 2017	TACE (32) <i>vs</i> TACE + RFA (32)	HCC BCLC B1 + B2	N/A	Median OS: 840 d vs 2466 d. OS (1, 3, 5 yr): 86.3%, 43.5%, 15.8% vs 100%, 78.6%, 62.3% (P < 0.001). Median TTP: 140 d vs 1148 d (P < 0.0001)
Ren <i>et al</i> [35], 2019	TACE (271) <i>vs</i> TACE + RFA (128)	HCC BCLB A and B	44.7% vs 85.9% (CR)	Median OS: 16 mo <i>vs</i> 59 mo (<i>P</i> < 0.001). Median PFS: 4 mo <i>vs</i> 45 mo. OS (1, 3, 5, 8 yr): 64.5%, 15.1%, 10.8%, 10.8% <i>vs</i> 90.6%, 76.6%, 68.0%, 68.0%
Chu et al [<mark>36</mark>], 2019	TACE (314) <i>vs</i> TACE + RFA (109) <i>vs</i> RFA (115)	HCC 3.1-10 cm	84.7% vs 95.4% vs 94.8% (CR)	RFS (5, 10, 15 yr): 59.1%, 11.0%, 2.2% vs 25.5%, 13.3%, 7.9% vs 9.2%, 2.9%, and 2.9% (<i>P</i> = 0.002). OS (5, 10, 15 yr): 16.2%, 10.9%, 7.7% vs 57.8%, 41.8%, 30.9% vs 35.2%, 11.9%, 11.9% (<i>P</i> = 0.022)
Liu et al[<mark>37</mark>], 2020	TACE (124) vs TACE + RFA (77)	HCC 3-10 cm	N/A	Median PFS: 4 mo <i>vs</i> 9.13 mo (<i>P</i> < 0.001). PFS (1, 3, 5 yr): 11.9%, 0%, 0% <i>vs</i> 43%, 18%, 9.5%. Median OS: 12 mo <i>vs</i> 27.57 mo (<i>P</i> < 0.001). OS (1, 3, 5 yr): 48%, 6.5%, 0% <i>vs</i> 76.2%, 37.1%, 16.4%
Hyun <i>et al</i> [38], 2016	TACE (54) <i>vs</i> TACE + RFA (37)	HCC not feasible for RFA	57% <i>vs</i> 100% <i>P</i> < 0.01 (CR)	Median TTP: 29.7 mo <i>vs</i> 34.9 mo (<i>P</i> = 0.014). OS (1, 2, 3 yr): 91%, 79%, 71% <i>vs</i> 100%, 97%, 93%
Yang <i>et al</i> [<mark>39]</mark> , 2020	TACE + RFA special location (n = 37) vs TACE + RFA conventional location (n = 85)	HCC special locations	91.9% vs 85.9% (CR) (NS)	Median PFS: 14 mo <i>vs</i> 17 mo (NS). Median OS: 32 mo <i>vs</i> 28 mo (NS). OS (1, 2 yr): 96.3%, 65% <i>vs</i> 89.9%, 63.3% (NS)
Hyun <i>et al</i> [<mark>112</mark>], 2016	TACE + RFA (14)	HCC < 2 cm caudate lobe	90.9% CR	LTP (1, 3, 5 yr): 0%, 12.5%, 12.5%. PFS (1, 3, 5 yr): 81.8%, 51.9%, 26%. OS (1, 3, 5 yr): 100%, 80.8%, 80.8%
Hyun <i>et al</i> [<mark>113</mark>], 2018	TACE +RFA (69)	HCC < 3 cm not feasible for RFA	100% CR	LTP (1, 3, 5, 7 yr): 4.4%, 6.8%, 8.2%, 9.5%, 9.5%. OS (1, 3, 5, 7 yr): 100%, 95%, 89%, 80%, 80%
Yan <i>et al</i> [114], 2018	TACE + RFA single session (87)	HCC < 7 cm not resectable	87.4% CR	LTP (1, 3, 5 yr): 0%, 29.9%, 55.2%. Median OS: 39 mo. OS (1, 3, 5 yr): 100%, 65.5%, 47.5%
Kim <i>et al</i> [<mark>115</mark>], 2019	TACE + RFA (67)	BCLC A, non- surgical	N/A	PFS (1, 3, 5 yr): 86.8%, 55.9%, 29.7%. OS (1, 3, 5 yr): 100%, 93.4%, 83.5%
Duan <i>et al</i> [<mark>116</mark>], 2020	TACE + RFA, one session (46)	HCC > 8 cm	N/A	PFS (2, 3 yr): 9.4 mo and 10.2 mo. OS (2, 3 yr): 18.4 mo and 26.4 mo
Zhang et al [117], 2020	TACE + RFA (1) naive (40); (2) recurrent (36); and (3) hepatectomy	1 tumor < 7 cm, up to 3 tumors < 3 cm, Child A or B	62.5% <i>vs</i> 70% (CR + PR)	OS (1, 2, 3 yr): 97.5%, 84%, 66% (A) vs 90%, 82%, 66% (B) vs 90%, 79%, 63% (C) (A vs B vs C NS). DFS: 75%, 51%, 35% (A) vs 50%, 31%, 17% (B) vs 80%, 59%, 40% (C) (A vs B P = 0.013)
Wang <i>et al</i> [118], 2018	TACE (13) <i>vs</i> TACE + RFA (13)	HCC with hepatic vein thrombus	0% + 92.3% vs 46.2% + 53.7% (CR + PR)	Median OS: 6.5 mo <i>vs</i> 18 mo (<i>P</i> = 0.02)
Song <i>et al</i> [119], 2020	TACE (63) <i>vs</i> TACE + RFA (96)	Recurrent HCC < 5 cm after HR	N/A	DFS (1, 3, 5 yr): 41.1%, 9.9%, 4.9% vs 55.1%, 22.5%, 9.7%. OS (1, 3, 5 yr): 75.9%, 30.7%, 11.3% vs 82.3%, 42.7%, 16.5% (NS)

P value less than 0.05 (typically \leq 0.05) is statistically significant. HCC: Hepatocellular carcinoma; RFA: Radiofrequency ablation; TACE: Transarterial chemoembolization; OS: Overall survival; PFS: Progression free survival; RFS: Recurrence free survival; TTP: Time to progression; LTP: Local tumor progression; LTR: Local tumor recurrence; DFS: Disease free survival; TR: Tumor recurrence; CR: Complete response.

> complications were higher in the TACE plus RFA group than in the TACE alone group (OR = 2.74, 95% CI: 1.07-7.07, P = 0.04)[40].

> The combination therapy TACE + RFA was also compared to RFA in several other meta-analyses[43-46]. The last one including 8 trials and 648 patients showed that RFA

Zaishideng® WJGO | https://www.wjgnet.com

plus TACE is associated with a significant advantage in RFS [hazard ratio (HR) = 0.58; 95%CI: 0.42-0.80, *P* = 0.001], and OS (HR = 0.60; 95%CI: 0.47-0.76, *P* < 0.001). The authors concluded that TACE combined with RFA is more competent and appealing than RFA alone, especially for intermediate and large-size hepatic tumors or younger patients with HCC[47].

TACE combined with RFA vs hepatic resection: According to the BCLC guidelines published in 2018, the mainstay of curative therapy for early HCC is RFA or surgeryeither HR or liver transplantation. As mentioned earlier, even though HR is a cornerstone treatment in non-metastatic disease, some patients are considered inappropriate candidates due to underlying liver disease[48]. We have emphasized in the previous lines that the combination therapy of TACE and RFA is superior and beneficial for the treatment of early and intermediate HCC vs the therapies used individually. Along these lines, the comparison of the outcome of TACE + RFA vs HR is of paramount importance as some of the patients are not ideal surgical candidates. Several comparative studies are presented in Table 3.

For small single HCC (2-3 cm) TACE + RFA have similar outcomes to HR in terms of disease free survival (DFS) and OS but with significantly lower complications rate and hospital stay^[49]. However, as expected for medium size and larger lesions tumor recurrence (75% vs 35.4%, P = 0.005) and local tumor progression (LTP) (55.7% vs 16%, P = 0.013) are significantly lower for HR[49]. Moreover, it seems that for patients fitting the up-to seven criteria both treatment options provide similar DFS and OS. However for patients outside the Milan criteria but within up to seven criteria there was an increased median OS when HR was performed[50]. For patients within the Milan criteria, Takuma and colleague reported similar DFS and OS, whereas Liu et al [51] found a significantly increased DFS and OS for patients who had undergone HR [51]. In a recent study, Lin *et al*[52] demonstrated a significantly higher 5 years OS (61.2% *vs* 38.2%, *P* = 0.009) for HR in patients with HCC BCLC-B[52].

Beyond a shadow of a doubt, the association of TACE with RFA has proved its benefit in terms of controlling much more suitably early and intermediate primary liver cancer compared to the therapies used alone and we consider that it has gained its place in the treatment of HCC. Moreover, combination therapy is also an important alternative when surgery is not feasible.

TACE combined with MWA: MWA is a dielectric heating technique that generates an electromagnetic field surrounding the needle tip, consequently producing the coagulation necrosis of the target area. Along with RFA, MWA is an established thermal ablation method, best-suited for treating early-stage HCC with curative intent. Being non-inferior to RFA in the standard guideline setting, MWA boosts some theoretical advantages: Higher efficacy in larger nodules, quicker heating, lower procedural time, and higher heating temperatures [53]. Therefore, like RFA, MWA has often been combined with TACE in a multitude of clinical scenarios. These situations either fit outside the standardized BCLC boxes or aim to improve the relatively dim prognosis of the well-established path in intermediate or advanced HCC.

As addressed by Renzulli et al[54], intermediate-and advanced-stage HCC often leads the clinicians in the uncertain waters of an imperfect solution[54]. The typical case resembles the following: Curative intent solution off the board, unsatisfactory gold-standard (TACE), and technically treatable nodules within ablative reach. This has led to a recent surge of interest with regards to combining TACE and MWA, with numerous papers published on this topic in the past decade. The most important articles are comprised in Table 4, which will provide the cornerstone for the upcoming discussion.

As shown in Table 4, there is a wide array of clinical scenarios in which the TACE + MWA approach was tested. However, the large majority of the data comes from the most heterogeneous class: Intermediate-stage, BCLC-B. However, most of the HCC disease spectrum stood for trial, ranging from small, solitary unresectable nodules, up to 10 nodules and more advanced BCLC-C tumors[55-58]. The overall results were promising, if not always definitely positive. Therefore, both the empirical and the epistemological conclusions suggest that TACE + MWA can be safely employed whenever it is technically feasible with regards to tumor characteristics, vascularization, and percutaneous approach.

Three key aspects define therapeutic efficacy in HCC: Treatment response according to the mRECIST criteria, PFS, and OS. In this regard, the TACE + MWA combination appeared to exceed the standard of care in most of the clinical scenarios, in all three aspects (Table 4). Of course, the precise survival data varies widely, as the study designs were extremely heterogeneous, even within the same BCLC class. In



Table 5 Coll	nparison transarteriai c	nemoempolizatio	on + radiofrequency ablation	on to other curative therapies
Ref.	Treatment type, <i>n</i> (%)	Clinical scenario	Response rate mRECIST	Outcome
Saviano <i>et al</i> [49], 2017	TACE + RFA $(n = 25) vs$ HR $(n = 29)$	HCC 3.0-8.8 cm, solitary HCC 3-5 cm	N/A	OS (1, 3 yr): 89.4%, 48.2% vs 91.8%, 79.3% ($P = 0.117$). TR (1, 3 yr): 42.4%, 76.0% vs 29.5%, 45.0% ($P = 0.034$); LTP (3 yr): 58.1% vs 21.8% ($P = 0.005$). TR: 75.1% vs 35.4% ($P = 0.016$); LTP: 55.7% vs 16.0% $P = 0.013$)
Pan <i>et al</i> [50], 2017	TACE + RFA ($n = 154$) vs HR ($n = 176$)	Within Up-To Seven criteria	N/A	Median OS: 56 mo <i>vs</i> 58 mo (NS). OS (1, 3, 5 yr): 96.1%, 76.7%, 41.3% <i>vs</i> 96.1%, 86.4%, 46.2% ($P = 0.138$). Median OS (beyond Milan): 52 mo <i>vs</i> 45 mo ($P = 0.023$)
Liu <i>et al</i> [<mark>51</mark>], 2016	TACE + RFA $(n = 100)$ vs HR $(n = 100)$	Within Milan	N/A	OS (1, 3, 5 yr): 96%, 67.2%, 45.7% vs 97%, 83.7%, 61.9% (P = 0.007). RFS (1, 3, 5 yr): 83%, 44.9%, 35.5% vs 94%, 68.2%, 48.4% (P = 0.026). Complications rate: 11% vs 23%, P = 0.024)
Lin et al[<mark>52</mark>], 2020	TACE $(n = 231) vs$ TACE + RFA $(n = 57) vs$ HR $(n = 140)$	BCLC-B	N/A	OS (1, 3, 5 yr): 69.5%, 37.0%, 15.2% vs 86.0%, 57.9%, 38.2% vs 89.2%, 69.4%, 61.2%. OS higher HR vs TACE + RFA (P = 0.009), HR vs TACE (P < 0.001) and TACE + RFA vs TACE (P = 0.004)
Wei <i>et al</i> [63], 2020	TACE + RFA (<i>n</i> = 107) <i>vs</i> HR (<i>n</i> = 79)	Recurrent HCC < 5 cm after HR	N/A	DFS (1, 3, 5 yr): 58.2%, 35.2%, 29.6% vs 64.8%, 41.6%, 38.3% ($P = 0.258$). OS (1, 3, 5 yr): 84.6%, 66.9%, 49.1% vs 84.8%, 60.2%, 51.9% ($P = 0.871$). Lower major complication rates ($P = 0.009$) and shorter hospital stay ($P < 0.001$) for TACE + RFA
Sheta <i>et al</i> [64], 2016	TACE $(n = 20) vs$ TACE + RFA $(n = 20) vs$ TACE + MWA $(n = 10)$	Non resectable single lesion HCC > 4 cm	50% <i>vs</i> 70% <i>vs</i> 80% (CR at 6 mo)	LTR (1, 3, 6 mo): 30% vs 5% vs 0% (P = 0.027); 14.3% vs 15.8% vs 10% (NS); 16.7% vs 12.5% vs 11.1% (NS). Complications rate: 40% vs 10% vs 10%
Yuan <i>et al</i> [65], 2019	TACE + RFA $(n = 41)$ vs TACE + MWA $(n = 34)$	HCC > 3 cm. HCC 3-5 cm. HCC > 5 cm	68.3 vs 85.3% (NS). 73.5 vs 88.5% (NS). 42.9 vs 75% (P = 0.041)	DFS (1, 2, 3 yr): 53%, 29%, 12% vs 58%, 38%, 29% ($P = 0.07$). OS (1, 2, 3 yr): 68%, 36%, 14% vs 79%, 53%, 38% ($P = 0.393$)
Thornton <i>et al</i> [120], 2017	TAE/TACE + RFA ($n =$ 15) vs TAE/TACE + MWA ($n = 20$)	BCLC 0 and A	80% vs 95% (NS)	LTR: 30% vs 0%
Vasnani <i>et al</i> [67], 2016	TACE + RFA $(n = 11) vs$ TACE + MWA $(n = 31)$	HCC within Milan	91% vs 67% (CR) 45% vs 35% (rates of complete tumor coagulation on pathology)	

mRECIST: Modified RECIST; n: Number of patients; BCLC: Barcelona Clinic Liver Cancer; TACE: Transarterial chemoembolization; RFA: Radiofrequency ablation; MWA: Microwave ablation; HR: Hepatic resection; OS: Overall survival; PFS: Progression free survival; TTP: Time to progression; LTP: Local tumor progression; DFS: Disease free survival; TR: Tumor recurrence; CR: Complete response; NS: Not significative; N/A: Not applicable.

> intermediate-stage HCC, OS ranged from 17.1 mo[57] to 35 mo[59] as it reliably extended OS compared to TACE alone by 4 to 12 mo[56,57,60,61]. To objectivize the apparent benefit, a meta-analysis was recently performed by Liu et al[62], comparing TACE with TACE + MWA in single and up to three HCC nodules exceeding 5 cm[62]. The analysis included over 1700 patients, all from Chinese-conducted studies, and showed a significantly higher OS for the latter (1-year OS rate: RR = 1.36, 95%CI: 1.28-1.44; 2-year OS rate: RR = 1.56, 95% CI: 1.40-1.74, and 3-year OS rate: RR = 2.07, 95% CI: 1.67–2.57, P < 0.001). However, when compared to other non-standard therapies, such as radiation segmentectomy^[55] or CA^[63] the differences in outcome were not statistically significant, which might further suggest the rather suboptimal standard of care for intermediate HCC. Other clinical scenarios have shown a benefit of combination therapy, as TACE + MWA + Sorafenib outperformed TACE + Sorafenib in off-guideline advanced HCC[58]. However, it is unclear whether the available reports provide sufficient grounding for altering the current time-tested recommendations since no major randomized controlled trials are available. Furthermore, the overall quality of the data can be improved, as most studies are retrospective.

> There are only a few small sample studies directly comparing MWA to RFA in combination with TACE, with no significant differences in major outcomes such as PFS and OS[64-66]. However, subgroup analysis suggests a higher response rate in larger tumors for MWA.

> Per available data, TACE + MWA appears to laterally exceed the BCLC-B stage. On one hand, it might secure substantial survival benefits for nodules not amenable to curative intent solutions. One such scenario might be large, borderline BCLC-A nodules, unfit for resection due to portal hypertension, yet fit for transplantation, but

Table 4 Available studies on the transarterial chemoembolization plus microwave ablation

Ref.	Treatment type, <i>n</i> (%)	Clinical scenario	Response rate (CR + PR) mRECIST	Outcome
Ni et al[<mark>59</mark>], 2020	TACE + MWA (546)	BCLC B	N/A	Median PFS: 6.5 mo. Median OS: 35 mo
Ni et al[<mark>121</mark>], 2019	TACE + MWA (349)	Up to 3 nodules, 5-8 cm diameter	77.1%	Median PFS: 4.8 mo. Median OS: 28 mo
Chen <i>et al</i> [47], 2017	TACE (96) <i>vs</i> TACE + MWA (48)	$HCC \le 5 \text{ cm}$	46.3% vs 92.1%	2-yr PFS: 57.3% <i>vs</i> 10.4%; 2-yr OS: NS
Smolock <i>et al</i> [56], 2018	TACE (16) <i>vs</i> TACE + MWA (22)	HCC 3-5 cm	76% vs 95% (NS)	Median PFS: 4.2 mo <i>vs</i> 22.3 mo. Median OS: 14.8 mo <i>vs</i> 18.5 mo. 3-yr OS: 42.1% <i>vs</i> 79%
Zheng <i>et al</i> [<mark>57]</mark> , 2018	TACE (166) vs TACE + MWA (92)	Solitary HCC > 5 cm; 2-3 nodules > 3 cm; 4-10 nodules regardless of size	55.4% <i>vs</i> 81.5%	Median PFS: 12.5 mo <i>vs</i> 26.6 mo. Median OS: 6.7 mo <i>vs</i> 17.1 mo. 3-yr OS: 11.4% <i>vs</i> 32.6%
Zhang <i>et al</i> [<mark>60</mark>], 2018	TACE (100) <i>vs</i> TACE + MWA (50)	BCLC-B	55% vs 74%. At 6-mo, including stable disease	Median PFS: 6.1 mo <i>vs</i> 10.1 mo. Median OS: 14.4 mo <i>vs</i> 18.5 mo. 3-yr OS: 42.1% <i>vs</i> 79%. 5-yr OS: 21% <i>vs</i> 67.7%
Wang et al <mark>[61</mark>], 2020	TACE (111) vs TACE + MWA (72)	Recurrent (post-surgery) BCLC-B	N/A	Median PFS: N/A. Median OS: 14.4 mo vs 26.7 mo. 5-yr PFS: 13.0% vs 21.7%. 5- yr OS: 27.9% vs 43.3%
Li et al[<mark>29</mark>], 2020	MWA (88) <i>vs</i> TACE + MWA (62)	BCLC-B	N/A	3-yr PFS: 34.5% vs 32.5% (NS). 3-yr OS: 47.6% vs 49.2% (NS)
Biederman <i>et al</i> [55], 2017	TACE + MWA (80) vs Radiation segmentectomy (41)	Unresectable, solitary, ≤ 3 cm	CR 82.5% vs 82.9% (NS)	Median PFS: 12.1 mo <i>vs</i> 11.1 mo (NS). 90-d mortality: 0% all groups. Median OS: N/A
Ni et al[<mark>59</mark>], 2020	TACE + Sorafenib (n = 75) vs TACE + Sorafenib + MWA (77)	BCLC C	12% vs 46.7%	Median PFS: 3 mo <i>vs</i> 6 mo. Median OS: 13 mo <i>vs</i> 19 mo
Sheta <i>et al</i> [<mark>64</mark>], 2016	TACE (20) <i>vs</i> TACE + RFA (20) <i>vs</i> TACE + MWA (10)	Unrsesectable, solitary	6-mo CR-50% vs 70% vs 80%	Median PFS: N/A. Median OS: N/A
Wei <i>et al</i> [<mark>63</mark>], 2020	TACE + MWA (48) vs TACE + Cryoablation (60)	BCLC B	73.3% vs 33.4%	Median PFS: 8.8 mo <i>vs</i> 9.3 mo (NS). Median OS: 20.9 <i>vs</i> 13 mo (NS)

TACE: Transarterial chemoembolization; MWA: Microwave ablation; RFA: Radiofrequency ablation; HCC: Hepatocellular carcinoma; BCLC: Barcelona Clinic Liver Cancer classification system; CR: Complete response; PR: Partial response according to the mRECIST criteria; PFS: Progression-free survival; OS: Overall survival; NS: Non-significant; N/A: Not available.

living in a low-transplantation rate medical system. On the other hand, for BCLC-C patients, TACE + MWA appears to bring survival benefits, especially when adding sorafenib into the mix. The further one navigates beyond BCLC-B, the lesser the strength of the data. Within BCLC-B though, TACE + MWA shines the brightest, appearing to add a substantial survival benefit[67].

TACE combined with other local therapies: In a RCT comparing CA + drug-eluting bead (DEB)-TACE to CA monotherapy in large tumors (mean: 7.2 ± 4.5 cm and 6.5 ± 3.8 cm, respectively), the combination group demonstrated superior OS [16.8 mo *vs* 13.4 mo, respectively (P = 0.0493)] and significantly increased PFS [8.1 mo *vs* 6.0 mo, respectively (P = 0.0089)]. Interstitial laser therapy (ILT) uses optical fibers to create cytotoxic temperatures *via* the conversion of absorbed infrared light to heat with consequent coagulative necrosis[68]. TACE followed by ILT was used in patients with tumors up to 8 cm that has obtained a median OS of 36 mo (95%CI: 29.3–42.6)[69].

Stereotactic body radiation therapy (SBRT) can provide satisfactory local control with a low incidence of radiation-induced liver disease in patients with unresectable HCC that are not amenable to thermal ablation[68]. *Via* radiosensitization of tissue TACE/TAE may serve as targeting fiducials for SBRT. TACE + SBRT was compared to SBRT in a retrospective study including patients with tumors with a median size of 8.5 cm. An increased 5-year OS was reported in the combination arm *vs* SBRT arm (46.9 *vs* 32.9%, *P* = 0.047)[70]. In addition to cytotoxic properties, SBRT seems to be a potent activator of the adaptive immune system, and thus might stand as an interesting player in the field of immunotherapy[68].

Zaisbidena® WJGO | https://www.wjgnet.com

COMBINED TREATMENT TO ENHANCE PALLIATION AND INCREASE SURVIVAL

TACE + systemic therapies–the downfall of an era

As mentioned previously, current guidelines recommend local ablative therapies (RFA/MWA) for patients with early-stage HCC (BCLC-0) and TACE for patients with intermediate-stage BCLC-B or BCLC-A patients not suitable for resection or ablation [2]. Unfortunately, many of these patients experience a progression of the disease and a worsening of the liver function after repeated sessions of percutaneous ablation or TACE^[71]. The OPTIMIS study, which assessed the outcomes of HCC patients treated with TACE alone, or with TACE followed by sorafenib found that the proportion of patients with a progressive disease increases with each subsequent TACE, while the objective response rates decline as the number of TACE sessions increased (first TACE: 40%; second TACE: 26%; third TACE: 24%; and fourth TACE: 25%)[72]. Moreover, up to 30% of these patients experienced a deterioration of liver function. Similarly, Hiraoka *et al*^[73] reported that the liver function deteriorated with repeated TACE^[73]. These findings emphasize the importance of the appropriate timing of switching from local therapy to systemic therapies to obtain the maximal benefits of other therapies and consequently improve the OS.

As a result of an exclusively arterial vascularization of HCC tumors and comprising the fact that the normal surrounding liver parenchyma is vascularized from branches of the portal vein, TACE and other image-guided transcatheter treatments were born to destruct arterial tumoral vessels and hence inducing tumor necrosis^[74]. TACE procedure is based on an intra-arterial infusion of a chemotherapy agent such as doxorubicin or cisplatin, frequently embedded in lipiodol as a vehicle to increase the availability of the drug. Furthermore, the tumoral blood vessels could be embolized with different agents such as gelatine sponge particles, metallic coils, polyvinyl alcohol, starch microspheres and autologous blood clots leading to an increased tumoricidal and ischemic effect[27]. According to the European Association for the Study of the Liver (EASL), the median survival for untreated patients at an intermediate-stage [BCLC-B-multinodular disease, good PS, without vascular invasion or extrahepatic spread] is around 16 mo and in rigorously selected candidates TACE can increase the survival up to 3 years[2].

TACE causes local hypoxia in the tumor, building up an expression of hypoxia response genes in tumor cells regulated by hypoxia-inducible factor-1 alpha (HIF-1 α). The response triggers vascular endothelial growth factor (VEGF) expression and thus leading to the formation of neovascularization, and thereby forming a vicious cycle leading to tumor recurrence and metastasis^[75]. In this regard, studies have conceded that dynamic changes in serum HIF-1 α and VEGF levels occur after TACE in HCC patients[76-78]. Along these lines, Jia et al[76] investigated the expression levels of serum HIF-1a and VEGF before and after TACE and analyzed the correlations between prognosis factors and serum HIF-1α as well as VEGF levels[76]. The serum HIF-1a and VEGF levels of HCC patients pre-TACE, 1 d, 1-wk, 1-mo post-TACE were analyzed using ELISA and compared with that of 20 healthy volunteers[76]. The study revealed that the expression levels of serum HIF-1α and VEGF in HCC patients were significantly higher than those in the control group. One day after TACE, both serum HIF-1a and VEGF levels reached the peak values. One-week post-TACE, expression levels of them were decreased, but still significantly higher than those before TACE. The levels of both HIF-1a and VEGF incomplete response group 1-mo post-TACE were significantly lower than those in partial response, stable disease, or progressive disease groups. Thus, HIF-1α and VEGF might be important predictors of TACE efficacy.

Along these lines, hepatologists and oncologists hypothesized whether the combination of systemic therapy and TACE might be beneficial in terms of survival in HCC patients. In an effort to address this problem, several trials with TACE and antiangiogenic therapies have emerged. Nevertheless, some challenges have arisen. Firstly, TACE is addressed to BCLC-B class patients which is a heterogeneous group due to the wide range of liver function (Child-Pugh A or B cirrhosis) and variable lesion number and dimension. Secondly, the use of chemotherapy, degree of selectivity and management of adverse effects have to be considered. The GIDEON trial, the first observational trial of more than 3000 patients with HCC BCLC A to C treated with sorafenib or in combination with TACE reported that sorafenib could be safely associated or used sequentially with TACE^[79]. Thereby, taking this assumption into account, several randomized controlled trials have been reported, as seen in Table 5.

Table 5 Chemoembolization plus systemic therapies							
Trial	Experimental arms, <i>n</i> (%)	Outcomes					
TACE + sorafenib							
SPACE trial (Lencioni <i>et al</i> [80], 2016)	DEB-TACE plus sorafenib (154) <i>vs</i> DEB-TACE plus placebo (153)	5.6 mo <i>vs</i> 5.5 mo; HR: 0.797 (95%CI: 0.588–1.080); <i>P</i> = 0.072					
TACE 2 trial (Meyer <i>et al</i> [81], 2017)	DEB-TACE plus sorafenib (157) vs DEB-TACE plus placebo (156)	7.8 mo vs 7.7 mo; HR: 1.03 (95%CI: 0.75–1.42); $P=0.85$					
STAH trial (Park <i>et al</i> [82], 2019)	cTACE plus sorafenib (170) vs sorafenib (169)	12.8 mo <i>vs</i> 10.8 mo; HR: 0.91 (95%CI: 0.69–1.21); <i>P</i> = 0.290					
TACTICS trial (Kudo et al[83], 2020)	cTACE plus sorafenib (80) vs cTACE (76)	25.2 mo vs 13.5 mo; HR: 0.59 (95%CI: 0.41–0.87); P = 0.006					
TACE + other therapies							
BRISK-TA trial (Kudo <i>et al</i> [85], 2014)	cTACE or DEB-TACE plus brivanib (249) vs cTACE plus placebo (253)	26.4 mo <i>vs</i> 26.1 mo; HR: 0.90 (95%CI: 0.66- 1.23); <i>P</i> = 0.53					
ORIENTAL trial (Kudo <i>et al</i> [86], 2018)	cTACE plus orantinib (445) <i>vs</i> cTACE plus placebo (444)	31.1 mo <i>vs</i> 32.3 mo; HR: 1.090 (95%CI: 0.878-1.352); <i>P</i> = 0.435					
TACE combined with celecoxib and lanreotide (Tong <i>et al</i> [89], 2017)	TACE ($n = 35$) vs TACE + C + L (36)	7.5 mo <i>vs</i> 15.0 mo; HR: 0.534 (95%CI: 0.321- 0.888); <i>P</i> = 0.016					
TACE combined with thalidomide (Wu <i>et al</i> [87], 2014)	TACE + thalidomide (56)	21 mo (95%CI: 16-28 mo)					
TACE plus bevacizumab (Pinter <i>et al</i> [88], 2015)	TACE + bevacizumab (20) vs TACE + placebo (20)	5.3 mo <i>vs</i> 13.7 mo; HR: 1.7 (95%CI: 0.8-3.6); <i>P</i> = 0.195					

HR: Hepatic resection; TACE: Transarterial chemoembolization.

SPACE prospective randomized phase II trial included 307 patients with BCLC-B HCC randomly allocated to DEB-TACE with sorafenib 400 mg twice daily and DEB-TACE with placebo. Unfortunately, there was no difference in TTP between the 2 arms (169 d vs 166 d in the sorafenib and placebo arms, respectively, P = 0.072) and no impact on OS (P = 0.29) was observed[80].

A year later a phase III trial of TACE with sorafenib (TACE-2) from the United Kingdom emerged and included 313 patients randomized to sorafenib or placebo with DEB-TACE 2-5 wk later and additional TACE on demand[81]. The study aimed to reduce the adverse effects induced by combination treatment and to increase the prospect of continuing the drug at the time of the TACE procedure. Sadly, as in the SPACE trial, this RCT was also negative with a median PFS of 7.9 vs only 7.8 mo in the sorafenib and placebo arms, respectively (P = 0.94), and median OS of 21.1 and 19.7 mo in the sorafenib and placebo groups, respectively (P = 0.57). Moreover, discouraging results were also reported by the STAH trial in 2019 when comparing the combined treatment with sorafenib alone. The authors justify their results due to delays in starting sorafenib after TACE and/or low daily sorafenib doses[82].

TACTICS trial was the only phase II RCT that attested to the benefits of TACEsorafenib synergy and met its primary endpoint for the treatment of intermediate stage HCC[83]. The authors reported a median PFS significantly longer in the TACE plus sorafenib group vs TACE alone group (25.2 vs 13.5 mo; P = 0.006). Moreover, the innovation of the trial stands in the modification of PFS, defined as time-tounTACEable progression (TTUP), characterized as untreatable tumor progression, transient deterioration to Child-Pugh C, or appearance of vascular invasion/extrahepatic spread. Patients in the combination group received sorafenib 400 mg once daily for 2-3 wk before TACE, followed by 800 mg once daily during on-demand conventional TACE sessions until time to untreatable TTUP. Howbeit, a further analysis conceded that the TACTICS trial did not show an improved OS in the combination group as compared with TACE alone although significantly better PFS was consistently observed. However, the OS in TACE plus sorafenib arm showed the longest OS (36.2 mo) with the longest Δ OS (5.4 mo) as compared with the previous TACE combination trials[84]. The authors explain that the major reason for the negative OS result was due to many post-trial active treatments (other systemic treatments and immunotherapy agents) performed in the sorafenib group (76.3%), which implies that the OS endpoint in the TACE combination trial may not be feasible anymore in the current era of personalized medicine and immunotherapy.



It seems likely that other TKIs (brivanib, orantinib) and thalidomide derivatives as well combined with conventional TACE failed to meet the primary endpoint of OS[85-87]. Moreover, the addition of bevacizumab to TACE raised some safety concerns related to sepsis and vascular complications of the combination treatment[88]. Nevertheless, encouraging results were published by Tong *et al*^[89] which have compared TACE alone with TACE combined with the selective COX-2 inhibitor, celecoxib and the somatostatin analog, lanreotide in advanced HCC[89]. The patients receiving the combination therapy had a median OS of 15 mo compared to 7.5 mo for those receiving TACE alone, and a subgroup analysis of advanced patients demonstrated an OS of 13 mo for the combination and 4.5 mo for TACE alone (P = 0.013). Likewise, encouraging results published in 2020 confirmed that the use of lenvatinib-TACE sequential treatment after progression during lenvatinib therapy was associated with better post-progression survival (HR = 0.08; 95% CI: 0.01–0.71; *P* = 0.023)[90].

Overall, up to this point, the literature has failed to support the use of multi-kinase inhibitors in combination with TACE. However, with the emergence of immunotherapy, combined strategies are encouraged. Moreover, because intermediate stage liver cancer is very heterogeneous a personalized approach is the key to a better outcome for patients.

TACE + immunotherapy–a new beginning?

It has been noted for several years that LRTs result in the release of tumor antigens, which are taken up by antigen-presenting cells (mainly dendritic cells) and which have been shown to activate a tumor-specific immune response a[91,92]. This evidence suggests that I LRTs may boost the response to immune-oncology drugs. Preliminary results of the phase I/II PETAL clinical trial (NCT03397654) showed good tolerability of pembrolizumab after TACE without cumulative side effects. Additionally, several clinical trials testing immune checkpoint inhibitors (ICI) as neoadjuvant or adjuvant therapy in patients treated with LRTs are currently running (Table 6).

However, despite the strong antitumor response induced by ICI, not all patients experienced an objective response^[93]. Currently there is no biomarker to predict response or resistance to immunotherapy in HCC. Emerging evidence revealed that VEGF is not only a proangiogenic factor but that VEGF also plays an important role in the development of the immunosuppressive tumor microenvironment (e.g. inhibition of dendritic cell maturation, accumulation of dendritic cell maturation, accumulation of myeloid-derived suppressor cells and induction of T reg cells). Voron et al[94] showed that targeting VEGF-A can decrease the VEGF-induced expression of inhibitory receptors mediating CD8+T cell exhaustion[94]. Given these results, the association of anti-angiogenic therapy [e.g. inhibitors of VEGF (bevacizumab) or TKIs (lenvatinib)] with ICI seems to overcome tumor-intrinsic resistance to immune checkpoint blockade.

LRTs are minimally invasive therapies; however, it has become clear that the treatment regimens adopted 15 years ago will change in the next few years. In the light of new evidence, three groups might benefit from the combined therapy (locoregional therapy and immunotherapy), including (1) Patients with a high risk of recurrence after a complete response by local ablation; (2) Patients who progressed under TACE; and (3) Patients with poor predictors of response to immunotherapy, when such predictors will be validated, possibly including NAFLD as underlying liver disease.

Currently, there are several trials underway evaluating different combinations (ICI \pm anti-angiogenic therapy) as options for patients treated with LRTs (Table 6). It is important to remember that immunotherapies represent a two-edged sword, thus we must find the right timing, dose and combination of immunotherapy for a robust response and minimal side effects.

Ablation + other treatments

HCC is an attractive target for immunotherapy due to several reasons: (1) Usually, HCC develops on a background of chronic inflammation (cirrhosis or chronic hepatitis); (2) In the context of cirrhosis there is an immunosuppressive environment; and (3) Immune evasion was described in patients with liver cancer[95].

Combining thermal ablation with immunotherapy is a very appealing approach. Thermally-induced necrosis can act as a permanent source of tumor antigens, the sublethal zone around the necrotic zone can generate inflammatory cytokines, and the thermal stress is capable of making HCC cells more sensitive to immune therapies[48, 96,97]. The field of immunotherapy in HCC (different from other cancer entities) was only recently unraveled. However, some preliminary studies with RFA and immunotherapy combinations (immune-ablation) have been already published. When used in a palliative setting, tremelimumab (anti-CTLA4) in combination with RFA or TACE in



Table 6 Summary of ongoing clinical trials evaluating combination therapy of immune checkpoint inhibitors with locoregional therapies

BCLC stage	Estimated/included patients	Clinical trial identifier	Phase	Arm
0	530	NCT03383458	III	Arm 1: RFA/MWA/curative resection + nivolumab (neoadjuvant) vs Arm 2: RFA/MWA/curative resection
В	26	NCT03397654 (PETAL)	Ib	Single arm: TACE followed by pembrolizumab
В	950	NCT04246177 LEAP-012	III	Arm 1: TACE + lenvatinib + pembrolizumab vs Arm 2: TACE
В	49	NCT03572582 (IMMUTACE)	II	Single arm: TACE + Nivolumab
В	522	NCT04268888 TACE-3	II/III	Arm 1: DEB-TACE + Nivolumab vs Arm 2: DEB-TACE
В	765	NCT04340193, CheckMate 74W	III	Arm 1: TACE + nivolumab + ipilimumab vs Arm 2: TACE + nivolumab + placebo
А	50	NCT03939975	Π	Single arm: Pembrolizumab or nivolumab or toripalimab. For participants with stable disease or atypical progression to immunotherapy therapy, RFA or MWA is performed additionally
В	130	NCT03864211	I/II	Single arm: RFA or MWA followed by Toripalimab
В	61	NCT01853618	I/II	Single arm: Tremelimumab + RFA or TACE
В	30	NCT03638141	Π	Single arm: Initial DEB-TACE followed by Durvalumab + tremelimumab
В	22	NCT03937830	ΙΙ	Single arm: Durvalumab and bevacizumab + TACE
B/C	600	NCT03778957 EMERALD-1	III	Arm 1: TACE + durvalumab vs Arm 2: TACE +bevacizumab + durvalumab
A/B	662	NCT04102098 IMbrave050	III	Atezolizumab plus bevacizumab in HCC patients at high risk of recurrence after surgical resection or ablation <i>vs</i> Active surveillance in HCC patients at high risk of recurrence after surgical resection or ablation

BCLC: Barcelona Clinic Liver Cancer Classification; RFA: Radiofrequency ablation; MWA: Microwave ablation; HCC: Hepatocellular carcinoma.

32 HCC patients showed indeed interesting results. The patients received a total of six doses of tremelimumab at a 4-wk interval followed by an intentionally incomplete RFA or TACE to induce anti-tumor response at the ablation tumor junction. Patients with clinical response had an increase in CD8+ T cells in tumor biopsies obtained 6 wk after treatment. More interestingly, some patients experienced tumor responses in untreated lesions[98]. In another study, Ma et al[99] injected RetroNectin activated killer cells 14 d after RFA in patients with an HCC less than 4 cm[99]. They reported no severe adverse events, recurrences, or deaths during a seven-month follow-up. Using a similar approach, Cui et al[100] studied the combination of RFA and cellular therapy. Mononuclear cells from 30-HCC patients (early, intermediate and advanced stage) were harvested and induced into natural killer cells, y\deltaT cells and cytokine-induced killer (CIK) cells which were subsequently infused back into RFA-treated patients for three or six courses. The combination improved PFS and reduced HCC recurrence compared to RFA alone[100].

Adjuvant treatment for HCC patients is an unsolved medical need. Using cellularbased immunotherapy Lee et al[101] studied the use of CIK cells injected after RFA (n = 69), ethanol injection (n = 13), or surgery (32) in patients with early-stage HCC[101, 102]. They reported a better OS and cancer-specific survival in patients treated with a combined approach vs those treated with RFA, percutaneous ethanol injection (PEI), or surgery alone (P = 0.006 and P = 0.02). Similar findings were also reported in one multicentre randomized open-label phase 3 trial of adjuvant immunotherapy with CIK cells. The study included 230 patients with HCC treated by SR, RFA, or PEI. Patients were assigned randomly to receive immunotherapy or no adjuvant therapy. Adjuvant CIK cell therapy increased both recurrence-free survival and OS[103].

The use of monoclonal antibodies in combination with RFA was also studied in HCC. Either injected during RFA (131I-chTNT) or after RFA (131I metuximab) in an adjuvant setting both combinations showed improved PFS or OS[103,104]. More data about studies investigating the combination of RFA with different immunotherapy strategies are depicted in Table 7.



Table 7 Ra	Table 7 Radiofrequency ablation combined with immunotherapy							
Ref.	BCLC, n (%)	Treatment, <i>n</i> (%)	Results	Level of evidence				
Cui <i>et al</i> [100], 2014	A (10); B (10); C (10)	RFA and cellular immunotherapy 8-11 d after RFA <i>vs</i> RFA alone	Higher PFS ($P < 0.001$). Six courses had better survival prognosis than three courses	III				
Ma et al [99], 2010	A (7)	RFA and autologous RAK cells 14 d after RFA	No severe adverse events, recurrences or deaths during a seven month follow-up	IV				
Duffy et al [98], 2017	C (21)	Tremelimumab every 4 wk and subtotal RFA on day 36	Median OS-12.3 mo. Median time to progression-7.4 mo. A significant increase of CD3+ and CD8+ immune cells infiltrates in lesions not treated by RFA	III				
Lee <i>et al</i> [<mark>102</mark>], 2015	A (114)	PEI (13); RFA (69); Surgery (32) and adjuvant CIK cells <i>vs</i> PEI, RFA or Surgery alone	OS was significantly longer in the immunotherapy group than in control group (P = 0.006). CSS was significantly longer in the immunotherapy group (P = 0.02)	Π				
Tu <i>et al</i> [<mark>103</mark>], 2014	A and B	RFA and monoclonal antibody (131I- chTNT) injection during ablation <i>vs</i> RFA alone	Increased OS. Improved progression-free survival. Increased circulating white blood cells	IV				
Bian <i>et al</i> [104], 2014	0 + A (94); B (33)	RFA and adjuvant 1311 metuximab vs RFA alone	Prevention of tumor recurrence	Π				
Lee <i>et al</i> [101], 2019	0 and A (239)	RFA or PEI or Surgery plus CIK vs RFA or PEI or surgery alone	Increased recurrence-free survival and OS	Ι				

BCLC: Barcelona Clinic Liver Cancer Classification; RFA: Radiofrequency ablation; PEI: Percutaneous ethanol injection; CIK: Cytokine-induced Killer; OS: Overall survival; CSS: Cyberchondria severity scale; PFS: Progression-free survival.

Nevertheless, the treatment of HCC will change over the following years. Not much has changed in the last 20 years but the era of immunotherapy has started and we will probably witness groundbreaking changes in the years that will come. New treatments, new guidelines, from single option to multiple options and from RFA to "immune-ablation" the burden has moved from scientist to clinicians: It is an interesting world out there.

Assuming similar suppositions with the TACE procedure, RFA was also combined with VEGF inhibitors. One study showed that Bevacizumab is useful in preventing the rapid progression of residual HCC following RFA in a rat model[105,106]. EMERALD-2 is an ongoing A Phase III, multicenter study of Durvalumab monotherapy or in combination with Bevacizumab as adjuvant therapy in patients with HCC who are at high risk of recurrence after resection or RFA (NCT03847428). Last but not least, nanoparticle-mediated drug delivery systems have also gained ground in oncology. The lyso-thermosensitive liposomal doxorubicin (LTLD) treatment aims to deliver doxorubicin at the peripheral thermal ablation zone, where the thermal elevation is suboptimal. When heated to 40 °C, LTLD releases a 25-fold greater concentration of Doxorubicin. The HEAT study is a global randomized, double-blind, dummycontrolled trial comparing RFA plus LTLD vs RFA alone that enrolled 701 patients with ≤ 4 unresectable up to seven HCC lesions. No differences in PFS and OS were found. The subgroup post hoc analysis showed improved efficacy when the thermal ablation indwell time for a solitary lesion was ≥ 45 min and increased treatment time per tumor volume was associated with better OS in the RFA + LTLD group[107,108]. The subsequent phase III OPTIMA study (NCT02112656) was halted in the interim analysis for futility reasons.

An exhaustive report about the combination of different tyrosine kinase inhibitors, distinctive ICI, or even their combo in the advanced liver cancer setting is beyond the scope of our review. However, we consider it far-reaching to mention one of 2020's revolutions-the association of bevacizumab (VEGF inhibitor) with atezolizumab (PD-L1 inhibitor) that brings encouraging data for unresectable HCC patients[109].

Although the combination of TACE and TKI's seemed promising in terms of inhibiting hypoxia-activated tumoral growth factors, studies do not appear to benefit any amalgam therapy compared to TACE monotherapy. To such a degree, one might say that the association of TACE and TKI's might have seen its downfall. Hence, the attention of the hepatology and oncology community was diverted to a new starimmunotherapy. In both the association with TACE and ablation, ICI have quietly demonstrated their benefit in trials. Although this path is still filled with uncertainty and caveats, in the following years we will witness an HCC guideline revolution.

Zaishideng® WJGO | https://www.wjgnet.com



Figure 1 The place of combined therapy in the Barcelona clinic liver cancer classification algorithm. ¹Lesion not seen at ultrasound or in inappropiate positions; ²Lesion > 3 cm; ³Within Up-to-seven criteria. ABL: Ablation; HR: Hepatic resection; IOP: Intraoperatiove ablation; IT: Immunotherapy; LT: Liver transplantation; LRT: Locoregional therapy; OLT: Orthotopic liver transplantation; TACE: Transarterial chemoembolization; TARE: Transarterial radioembolization; TKI: Tyrosine kinase inhibitors.

THE PLACE OF COMBINED THERAPY IN THE BCLC/EASL-HCC GUIDELINES

In tumor boards across the world, the most debatable section of the BCLC classification appears to be ranging from non-resectable BCLC-A to the less severe spectrum of the BCLC-C class, which is, by excellence, the appanage of interventional therapies [110]. Conventionally, the available treatments are dichotomized in curative intent and, possibly mislabelled, palliative therapies. The first group comprises RFA, MWA, PEI, CA, irreversible electroporation, and the latter includes bland trans-arterial embolization, conventional TACE, DEB-TACE, and endovascular radiotherapyselective internal radiation therapy [2,3].

However, the basis for the aforementioned dichotomy might be fading, as research published in the past decade has shown that combination therapy is at least technically feasible, with the most relevant results being discussed in the previous sections. This has prompted a discussion with regards to the place of combination therapy in the therapeutic algorithm, as some of the approaches might be suited for second-, or even first-line choices for a select group of patients. On the other hand, it might be important to recognize that over-complicating a relatively straightforward algorithm could lead to disputable therapeutic choices and widespread heterogeneous interpretation, rendering data collection difficult.

As discussed earlier, one particular combination therapy appears to stand-out among other approaches: TACE-ablation for small, non-resectable HCC (3-5 cm), which is currently at the threshold between BCLC-A and -B. Not only does it improve outcomes in comparison to each individual therapy alone, but, according to limited data, it also appears to generate outcomes similar to surgery, which otherwise would have not been available[30]. A proposed alteration of the BCLC classification, which speculates on the potential role of combination therapies based on the available data previously discussed, is shown in Figure 1.

CONCLUSION

Of course, our proposal is based on the best available data and still needs further consensus validation, but might provide a foundation for future recommendations, as well as hinting towards potential areas of future development. There is a great need for well-designed, large-scale randomized controlled trials to adequately assess the benefits of combination therapy. Moreover, there are multiple nuances open for debate. Which is the best radiological method for combination therapy: Bland TAE, TACE, or DEB-TACE? Which ablative technique has the most benefits? Should treatments be applied in the same session or sequential? Which is the best sequence? The authors strongly believe that methodically addressing these questions could



ultimately lead to a truly personalized approach, hoping to improve the quality of life and OS of HCC patients.

REFERENCES

- GBD 2015 Mortality and Causes of Death Collaborators. Global, regional, and national life expectancy, all-cause mortality, and cause-specific mortality for 249 causes of death, 1980-2015: a systematic analysis for the Global Burden of Disease Study 2015. Lancet 2016; 388: 1459-1544 [PMID: 27733281 DOI: 10.1016/S0140-6736(16)31012-1]
- 2 Forner A, Da Fonseca LG, Díaz-González Á, Sanduzzi-Zamparelli M, Reig M, Bruix J. Controversies in the management of hepatocellular carcinoma. JHEP Rep 2019; 1: 17-29 [PMID: 32039350 DOI: 10.1016/j.jhepr.2019.02.003]
- 3 Vogel A, Cervantes A, Chau I, Daniele B, Llovet JM, Meyer T, Nault JC, Neumann U, Ricke J, Sangro B, Schirmacher P, Verslype C, Zech CJ, Arnold D, Martinelli E; ESMO Guidelines Committee. Hepatocellular carcinoma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol 2018; 29: iv238-iv255 [PMID: 30285213 DOI: 10.1093/annonc/mdy308]
- Global Burden of Disease Liver Cancer Collaboration, Akinyemiju T, Abera S, Ahmed M, Alam N, Alemayohu MA, Allen C, Al-Raddadi R, Alvis-Guzman N, Amoako Y, Artaman A, Ayele TA, Barac A, Bensenor I, Berhane A, Bhutta Z, Castillo-Rivas J, Chitheer A, Choi JY, Cowie B, Dandona L, Dandona R, Dey S, Dicker D, Phuc H, Ekwueme DU, Zaki MS, Fischer F, Fürst T, Hancock J, Hay SI, Hotez P, Jee SH, Kasaeian A, Khader Y, Khang YH, Kumar A, Kutz M, Larson H, Lopez A, Lunevicius R, Malekzadeh R, McAlinden C, Meier T, Mendoza W, Mokdad A, Moradi-Lakeh M, Nagel G, Nguyen Q, Nguyen G, Ogbo F, Patton G, Pereira DM, Pourmalek F, Qorbani M, Radfar A, Roshandel G, Salomon JA, Sanabria J, Sartorius B, Satpathy M, Sawhney M, Sepanlou S, Shackelford K, Shore H, Sun J, Mengistu DT, Topór-Madry R, Tran B, Ukwaja KN, Vlassov V, Vollset SE, Vos T, Wakayo T, Weiderpass E, Werdecker A, Yonemoto N, Younis M, Yu C, Zaidi Z, Zhu L, Murray CJL, Naghavi M, Fitzmaurice C. The Burden of Primary Liver Cancer and Underlying Etiologies From 1990 to 2015 at the Global, Regional, and National Level: Results From the Global Burden of Disease Study 2015. JAMA Oncol 2017; 3: 1683-1691 [PMID: 28983565 DOI: 10.1001/jamaoncol.2017.3055]
- 5 Llovet JM, Brú C, Bruix J. Prognosis of hepatocellular carcinoma: the BCLC staging classification. Semin Liver Dis 1999; 19: 329-338 [PMID: 10518312 DOI: 10.1055/s-2007-1007122]
- Qiu J, Chen S, Wu H. Long-term outcomes after hepatic resection combined with radiofrequency ablation for initially unresectable multiple and bilobar liver malignancies. J Surg Res 2014; 188: 14-20 [PMID: 24387841 DOI: 10.1016/j.jss.2013.11.1120]
- Schindl MJ, Redhead DN, Fearon KC, Garden OJ, Wigmore SJ; Edinburgh Liver Surgery and 7 Transplantation Experimental Research Group (eLISTER). The value of residual liver volume as a predictor of hepatic dysfunction and infection after major liver resection. Gut 2005; 54: 289-296 [PMID: 15647196 DOI: 10.1136/gut.2004.046524]
- 8 Forner A, Reig ME, de Lope CR, Bruix J. Current strategy for staging and treatment: the BCLC update and future prospects. Semin Liver Dis 2010; 30: 61-74 [PMID: 20175034 DOI: 10.1055/s-0030-1247133]
- 9 Arellano RS. What's New in Percutaneous Ablative Strategies for Hepatocellular Carcinoma and Colorectal Hepatic Metastases? Curr Oncol Rep 2020; 22: 105 [PMID: 32725433 DOI: 10.1007/s11912-020-00967-y]
- 10 de Jong KP, Wertenbroek MW. Liver resection combined with local ablation: where are the limits? Dig Surg 2011; 28: 127-133 [PMID: 21540598 DOI: 10.1159/000323823]
- Eisele RM, Zhukowa J, Chopra S, Schmidt SC, Neumann U, Pratschke J, Schumacher G. Results of 11 liver resection in combination with radiofrequency ablation for hepatic malignancies. Eur J Surg Oncol 2010; 36: 269-274 [PMID: 19726155 DOI: 10.1016/j.ejso.2009.07.188]
- 12 Pawlik TM, Izzo F, Cohen DS, Morris JS, Curley SA. Combined resection and radiofrequency ablation for advanced hepatic malignancies: results in 172 patients. Ann Surg Oncol 2003; 10: 1059-1069 [PMID: 14597445 DOI: 10.1245/aso.2003.03.026]
- 13 Ledoux G, Amroun K, Rhaiem R, Cagniet A, Aghaei A, Bouche O, Hoeffel C, Sommacale D, Piardi T, Kianmanesh R. Fully laparoscopic thermo-ablation of liver malignancies with or without liver resection: tumor location is an independent local recurrence risk factor. Surg Endosc 2021; 35: 845-853 [PMID: 32076859 DOI: 10.1007/s00464-020-07456-0]
- 14 Belli G, D'Agostino A, Fantini C, Cioffi L, Belli A, Russolillo N, Langella S. Laparoscopic radiofrequency ablation combined with laparoscopic liver resection for more than one HCC on cirrhosis. Surg Laparosc Endosc Percutan Tech 2007; 17: 331-334 [PMID: 17710062 DOI: 10.1097/SLE.0b013e31806d9c65
- 15 Donadon M, Costa G, Torzilli G. State of the art of intraoperative ultrasound in liver surgery: current use for staging and resection guidance. Ultraschall Med 2014; 35: 500-511; quiz 512-513 [PMID: 25474100 DOI: 10.1055/s-0034-1385515]
- 16 Mathuram Thiyagarajan U, Brown R, Dasari BVM. Liver Resection and Role of Extended Cytology and Histology: Response to: Viganò L, Costa G, Cimino MM, Procopio F, Donadon M, Del Fabbro D, Belghiti J, Kokudo N, Makuuchi M, Vauthey JN, Torzilli G. R1 Resection for



Colorectal Liver Metastases: a Survey Questioning Surgeons about Its Incidence, Clinical Impact, and Management. J Gastrointest Surg. 2018 Oct; 22(10):1752-1763. J Gastrointest Surg 2019; 23: 1283-1284 [PMID: 30891660 DOI: 10.1007/s11605-019-04189-x]

- 17 Ju M, Yopp AC. The Utility of Anatomical Liver Resection in Hepatocellular Carcinoma: Associated with Improved Outcomes or Lack of Supportive Evidence? Cancers (Basel) 2019; 11 [PMID: 31561585 DOI: 10.3390/cancers11101441]
- 18 Poch FGM, Neizert CA, Gemeinhardt O, Geyer B, Eminger K, Rieder C, Niehues SM, Vahldiek J, Thieme SF, Lehmann KS. Intermittent Pringle maneuver may be beneficial for radiofrequency ablations in situations with tumor-vessel proximity. Innov Surg Sci 2018; 3: 245-251 [PMID: 31579788 DOI: 10.1515/iss-2018-0008]
- 19 Bartoş A, Bartos D, Spârchez Z, Iancu I, Ciobanu L, Iancu C, Breazu C. Laparoscopic Contrast-Enhanced Ultrasonography for Real Time Monitoring of Laparoscopic Radiofrequency Ablation for Hepatocellular Carcinoma: an Observational Pilot Study. J Gastrointestin Liver Dis 2019; 28: 457-462 [PMID: 31826072 DOI: 10.15403/jgld-263]
- 20 Abdalla EK, Vauthey JN, Ellis LM, Ellis V, Pollock R, Broglio KR, Hess K, Curley SA. Recurrence and outcomes following hepatic resection, radiofrequency ablation, and combined resection/ablation for colorectal liver metastases. Ann Surg 2004; 239: 818-825; discussion 825-827 [PMID: 15166961 DOI: 10.1097/01.sla.0000128305.90650.71]
- 21 Xu LL, Zhang M, Yi PS, Zheng XB, Feng L, Lan C, Tang JW, Ren SS, Xu MQ. Hepatic resection combined with radiofrequency ablation vs hepatic resection alone for multifocal hepatocellular carcinomas: A meta-analysis. J Huazhong Univ Sci Technolog Med Sci 2017; 37: 974-980 [PMID: 29270762 DOI: 10.1007/s11596-017-1836-3]
- Chiappa A, Bertani E, Zbar AP, Foschi D, Fazio N, Zampino M, Belluco C, Orsi F, Della Vigna P, 22 Bonomo G, Venturino M, Ferrari C, Biffi R. Optimizing treatment of hepatic metastases from colorectal cancer: Resection or resection plus ablation? Int J Oncol 2016; 48: 1280-1289 [PMID: 26782649 DOI: 10.3892/ijo.2016.3324]
- 23 Hou YF, Wei YG, Yang JY, Wen TF, Xu MQ, Yan LN, Li B. Combined hepatectomy and radiofrequency ablation vs TACE in improving survival of patients with unresectable BCLC stage B HCC. Hepatobiliary Pancreat Dis Int 2016; 15: 378-385 [PMID: 27498577 DOI: 10.1016/s1499-3872(16)60089-9]
- 24 Zhang T, Zeng Y, Huang J, Liao M, Wu H. Combined resection with radiofrequency ablation for bilobar hepatocellular carcinoma: a single-center experience. J Surg Res 2014; 191: 370-378 [PMID: 24766727 DOI: 10.1016/j.jss.2014.03.048]
- 25 Huang Y, Song J, Zheng J, Jiang L, Yan L, Yang J, Zeng Y, Wu H. Comparison of Hepatic Resection Combined with Intraoperative Radiofrequency Ablation, or Hepatic Resection Alone, for Hepatocellular Carcinoma Patients with Multifocal Tumors Meeting the University of California San Francisco (UCSF) Criteria: A Propensity Score-Matched Analysis. Ann Surg Oncol 2020; 27: 2334-2345 [PMID: 32016632 DOI: 10.1245/s10434-020-08231-0]
- 26 Philips P, Scoggins CR, Rostas JK, McMasters KM, Martin RC. Safety and advantages of combined resection and microwave ablation in patients with bilobar hepatic malignancies. Int J Hyperthermia 2017; 33: 43-50 [PMID: 27405728 DOI: 10.1080/02656736.2016.1211751]
- Li W, Ni CF. Current status of the combination therapy of transarterial chemoembolization and local 27 ablation for hepatocellular carcinoma. Abdom Radiol (NY) 2019; 44: 2268-2275 [PMID: 31016345 DOI: 10.1007/s00261-019-01943-2]
- 28 Xu Z, Xie H, Zhou L, Chen X, Zheng S. The Combination Strategy of Transarterial Chemoembolization and Radiofrequency Ablation or Microwave Ablation against Hepatocellular Carcinoma. Anal Cell Pathol (Amst) 2019; 2019: 8619096 [PMID: 31534899 DOI: 10.1155/2019/8619096]
- 29 Li X, Chen B, An C, Cheng Z, Han Z, Liu F, Yu J, Liang P. Transarterial chemoembolization combined with microwave ablation vs microwave ablation only for Barcelona clinic liver cancer Stage B hepatocellular carcinoma: A propensity score matching study. J Cancer Res Ther 2020; 16: 1027-1037 [PMID: 33004744 DOI: 10.4103/jcrt.JCRT_380_19]
- 30 Young S, Golzarian J. Locoregional Therapies in the Treatment of 3- to 5-cm Hepatocellular Carcinoma: Critical Review of the Literature. AJR Am J Roentgenol 2020; 215: 223-234 [PMID: 32255691 DOI: 10.2214/AJR.19.22098]
- 31 Song MJ, Bae SH, Lee JS, Lee SW, Song DS, You CR, Choi JY, Yoon SK. Combination transarterial chemoembolization and radiofrequency ablation therapy for early hepatocellular carcinoma. Korean J Intern Med 2016; 31: 242-252 [PMID: 26874512 DOI: 10.3904/kjim.2015.112
- 32 Lee H, Yoon CJ, Seong NJ, Jeong SH, Kim JW. Comparison of Combined Therapy Using Conventional Chemoembolization and Radiofrequency Ablation Versus Conventional Chemoembolization for Ultrasound-Invisible Early-Stage Hepatocellular Carcinoma (Barcelona Clinic Liver Cancer Stage 0 or A). Korean J Radiol 2018; 19: 1130-1139 [PMID: 30386144 DOI: 10.3348/kjr.2018.19.6.1130]
- 33 Liu F, Chen M, Mei J, Xu L, Guo R, Lin X, Zhang Y, Peng Z. Transarterial Chemoembolization Combined with Radiofrequency Ablation in the Treatment of Stage B1 Intermediate Hepatocellular Carcinoma. J Oncol 2019; 2019: 6298502 [PMID: 31636667 DOI: 10.1155/2019/6298502]
- Hirooka M, Hiraoka A, Ochi H, Kisaka Y, Joko K, Michitaka K, Hiasa Y. Transcatheter Arterial 34 Chemoembolization With or Without Radiofrequency Ablation: Outcomes in Patients With



Barcelona Clinic Liver Cancer Stage B Hepatocellular Carcinoma. AJR Am J Roentgenol 2018; 210: 891-898 [PMID: 29412017 DOI: 10.2214/AJR.17.18177]

- 35 Ren Y, Cao Y, Ma H, Kan X, Zhou C, Liu J, Shi Q, Feng G, Xiong B, Zheng C. Improved clinical outcome using transarterial chemoembolization combined with radiofrequency ablation for patients in Barcelona clinic liver cancer stage A or B hepatocellular carcinoma regardless of tumor size: results of a single-center retrospective case control study. BMC Cancer 2019; 19: 983 [PMID: 31640620 DOI: 10.1186/s12885-019-6237-5]
- Chu HH, Kim JH, Yoon HK, Ko HK, Gwon DI, Kim PN, Sung KB, Ko GY, Kim SY, Park SH. 36 Chemoembolization Combined with Radiofrequency Ablation for Medium-Sized Hepatocellular Carcinoma: A Propensity-Score Analysis. J Vasc Interv Radiol 2019; 30: 1533-1543 [PMID: 31471190 DOI: 10.1016/j.jvir.2019.06.006]
- 37 Liu W, Xu H, Ying X, Zhang D, Lai L, Wang L, Tu J, Ji J. Radiofrequency Ablation (RFA) Combined with Transcatheter Arterial Chemoembolization (TACE) for Patients with Medium-to-Large Hepatocellular Carcinoma: A Retrospective Analysis of Long-Term Outcome. Med Sci Monit 2020; 26: e923263 [PMID: 32667906 DOI: 10.12659/MSM.923263]
- 38 Hyun D, Cho SK, Shin SW, Park KB, Park HS, Choo SW, Do YS, Choo IW, Lee MW, Rhim H, Lim HK. Early Stage Hepatocellular Carcinomas Not Feasible for Ultrasound-Guided Radiofrequency Ablation: Comparison of Transarterial Chemoembolization Alone and Combined Therapy with Transarterial Chemoembolization and Radiofrequency Ablation. Cardiovasc Intervent Radiol 2016; 39: 417-425 [PMID: 26246215 DOI: 10.1007/s00270-015-1194-0]
- Yang BS, Liu LX, Yuan M, Hou YB, Li QT, Zhou S, Shi YX, Gao BL. Multiple imaging modality-39 guided radiofrequency ablation combined with transarterial chemoembolization for hepatocellular carcinoma in special locations. Diagn Interv Radiol 2020; 26: 131-139 [PMID: 32071022 DOI: 10.5152/dir.2019.18540]
- 40 Yang DJ, Luo KL, Liu H, Cai B, Tao GQ, Su XF, Hou XJ, Ye F, Li XY, Tian ZQ. Meta-analysis of transcatheter arterial chemoembolization plus radiofrequency ablation vs transcatheter arterial chemoembolization alone for hepatocellular carcinoma. Oncotarget 2017; 8: 2960-2970 [PMID: 27936465 DOI: 10.18632/oncotarget.13813]
- 41 Cao JH, Zhou J, Zhang XL, Ding X, Long QY. Meta-analysis on radiofrequency ablation in combination with transarterial chemoembolization for the treatment of hepatocellular carcinoma. J Huazhong Univ Sci Technolog Med Sci 2014; 34: 692-700 [PMID: 25318879 DOI: 10.1007/s11596-014-1338-5]
- 42 Wang Y, Deng T, Zeng L, Chen W. Efficacy and safety of radiofrequency ablation and transcatheter arterial chemoembolization for treatment of hepatocellular carcinoma: A meta-analysis. Hepatol Res 2016; 46: 58-71 [PMID: 26265000 DOI: 10.1111/hepr.12568]
- 43 Lu Z, Wen F, Guo Q, Liang H, Mao X, Sun H. Radiofrequency ablation plus chemoembolization vs radiofrequency ablation alone for hepatocellular carcinoma: a meta-analysis of randomizedcontrolled trials. Eur J Gastroenterol Hepatol 2013; 25: 187-194 [PMID: 23134976 DOI: 10.1097/MEG.0b013e32835a0a07]
- Ni JY, Liu SS, Xu LF, Sun HL, Chen YT. Meta-analysis of radiofrequency ablation in combination 44 with transarterial chemoembolization for hepatocellular carcinoma. World J Gastroenterol 2013; 19: 3872-3882 [PMID: 23840128 DOI: 10.3748/wjg.v19.i24.3872]
- 45 Liu Z, Gao F, Yang G, Singh S, Lu M, Zhang T, Zhong Z, Zhang F, Tang R. Combination of radiofrequency ablation with transarterial chemoembolization for hepatocellular carcinoma: an upto-date meta-analysis. *Tumour Biol* 2014; 35: 7407-7413 [PMID: 24777334 DOI: 10.1007/s13277-014-1976-z
- 46 Chen L, Sun J, Yang X. Radiofrequency ablation-combined multimodel therapies for hepatocellular carcinoma: Current status. Cancer Lett 2016; 370: 78-84 [PMID: 26472630 DOI: 10.1016/j.canlet.2015.09.020]
- 47 Chen QF, Jia ZY, Yang ZQ, Fan WL, Shi HB. Transarterial Chemoembolization Monotherapy Versus Combined Transarterial Chemoembolization-Microwave Ablation Therapy for Hepatocellular Carcinoma Tumors ≤5 cm: A Propensity Analysis at a Single Center. Cardiovasc Intervent Radiol 2017; 40: 1748-1755 [PMID: 28681222 DOI: 10.1007/s00270-017-1736-8]
- Gui CH, Baey S, D'cruz RT, Shelat VG. Trans-arterial chemoembolization + radiofrequency 48 ablation vs surgical resection in hepatocellular carcinoma - A meta-analysis. Eur J Surg Oncol 2020; 46: 763-771 [PMID: 31937433 DOI: 10.1016/j.ejso.2020.01.004]
- 49 Saviano A, Iezzi R, Giuliante F, Salvatore L, Mele C, Posa A, Ardito F, De Gaetano AM, Pompili M; HepatoCATT Study Group. Liver Resection vs Radiofrequency Ablation plus Transcatheter Arterial Chemoembolization in Cirrhotic Patients with Solitary Large Hepatocellular Carcinoma. J Vasc Interv Radiol 2017; 28: 1512-1519 [PMID: 28734848 DOI: 10.1016/j.jvir.2017.06.016]
- Pan T, Mu LW, Wu C, Wu XQ, Xie QK, Li XS, Lyu N, Li SL, Deng HJ, Jiang ZB, Lin AH, Zhao 50 M. Comparison of Combined Transcatheter Arterial Chemoembolization and CT-guided Radiofrequency Ablation with Surgical Resection in Patients with Hepatocellular Carcinoma within the Up-to-seven Criteria: A Multicenter Case-matched Study. J Cancer 2017; 8: 3506-3513 [PMID: 29151935 DOI: 10.7150/jca.19964]
- Liu H, Wang ZG, Fu SY, Li AJ, Pan ZY, Zhou WP, Lau WY, Wu MC. Randomized clinical trial of 51 chemoembolization plus radiofrequency ablation vs partial hepatectomy for hepatocellular carcinoma within the Milan criteria. Br J Surg 2016; 103: 348-356 [PMID: 26780107 DOI: 10.1002/bjs.10061]



- 52 Lin CW, Chen YS, Lo GH, Hsu YC, Hsu CC, Wu TC, Yeh JH, Hsiao P, Hsieh PM, Lin HY, Shu CW, Hung CM. Comparison of overall survival on surgical resection vs transarterial chemoembolization with or without radiofrequency ablation in intermediate stage hepatocellular carcinoma: a propensity score matching analysis. BMC Gastroenterol 2020; 20: 99 [PMID: 32272898 DOI: 10.1186/s12876-020-01235-w]
- 53 Facciorusso A, Di Maso M, Muscatiello N. Microwave ablation vs radiofrequency ablation for the treatment of hepatocellular carcinoma: A systematic review and meta-analysis. Int J Hyperthermia 2016; 32: 339-344 [PMID: 26794414 DOI: 10.3109/02656736.2015.1127434]
- Renzulli M, Tovoli F, Clemente A, Ierardi AM, Pettinari I, Peta G, Marasco G, Festi D, Piscaglia F, 54 Cappabianca S, Carrafiello G, Golfieri R. Ablation for hepatocellular carcinoma: beyond the standard indications. Med Oncol 2020; 37: 23 [PMID: 32166482 DOI: 10.1007/s12032-020-01348-v
- Biederman DM, Titano JJ, Bishay VL, Durrani RJ, Dayan E, Tabori N, Patel RS, Nowakowski FS, 55 Fischman AM, Kim E. Radiation Segmentectomy vs TACE Combined with Microwave Ablation for Unresectable Solitary Hepatocellular Carcinoma Up to 3 cm: A Propensity Score Matching Study. Radiology 2017; 283: 895-905 [PMID: 27930089 DOI: 10.1148/radiol.2016160718]
- Smolock AR, Cristescu MM, Hinshaw A, Woo KM, Wells SA, Ziemlewicz TJ, Lubner MG, Dalvie 56 PS, Louis Hinshaw J, Brace CL, Ozkan OS, Lee FT Jr, Laeseke P. Combination transarterial chemoembolization and microwave ablation improves local tumor control for 3- to 5-cm hepatocellular carcinoma when compared with transarterial chemoembolization alone. Abdom Radiol (NY) 2018; 43: 2497-2504 [PMID: 29450606 DOI: 10.1007/s00261-018-1464-9]
- 57 Zheng L, Li HL, Guo CY, Luo SX. Comparison of the Efficacy and Prognostic Factors of Transarterial Chemoembolization Plus Microwave Ablation vs Transarterial Chemoembolization Alone in Patients with a Large Solitary or Multinodular Hepatocellular Carcinomas. Korean J Radiol 2018; 19: 237-246 [PMID: 29520181 DOI: 10.3348/kjr.2018.19.2.237]
- Ni JY, Sun HL, Luo JH, Jiang XY, Chen D, Wang WD, Chen YT, Huang JH, Xu LF. Transarterial 58 Chemoembolization and Sorafenib Combined with Microwave Ablation for Advanced Primary Hepatocellular Carcinoma: A Preliminary Investigation of Safety and Efficacy. Cancer Manag Res 2019; 11: 9939-9950 [PMID: 32063720 DOI: 10.2147/CMAR.S224532]
- Ni JY, Fang ZT, Sun HL, An C, Huang ZM, Zhang TQ, Jiang XY, Chen YT, Xu LF, Huang JH. A 59 nomogram to predict survival of patients with intermediate-stage hepatocellular carcinoma after transarterial chemoembolization combined with microwave ablation. Eur Radiol 2020; 30: 2377-2390 [PMID: 31900694 DOI: 10.1007/s00330-019-06438-8]
- 60 Zhang R, Shen L, Zhao L, Guan Z, Chen Q, Li W. Combined transarterial chemoembolization and microwave ablation vs transarterial chemoembolization in BCLC stage B hepatocellular carcinoma. Diagn Interv Radiol 2018; 24: 219-224 [PMID: 29792289 DOI: 10.5152/dir.2018.17528]
- Wang C, Liao Y, Qiu J, Yuan Y, Zhang Y, Li K, Zou R, Wang Y, Zuo D, He W, Zheng Y, Li B. 61 Transcatheter arterial chemoembolization alone or combined with ablation for recurrent intermediate-stage hepatocellular carcinoma: a propensity score matching study. J Cancer Res Clin Oncol 2020; 146: 2669-2680 [PMID: 32449005 DOI: 10.1007/s00432-020-03254-2]
- 62 Liu C, Li T, He JT, Shao H. TACE combined with microwave ablation therapy vs. TACE alone for treatment of early- and intermediate-stage hepatocellular carcinomas larger than 5 cm: a metaanalysis. Diagn Interv Radiol 2020; 26: 575-583 [PMID: 32965220 DOI: 10.5152/dir.2020.19615]
- Wei J, Cui W, Fan W, Wang Y, Li J. Unresectable Hepatocellular Carcinoma: Transcatheter Arterial Chemoembolization Combined With Microwave Ablation vs. Combined With Cryoablation. Front Oncol 2020; 10: 1285 [PMID: 32850395 DOI: 10.3389/fonc.2020.01285]
- 64 Sheta E, El-Kalla F, El-Gharib M, Kobtan A, Elhendawy M, Abd-Elsalam S, Mansour L, Amer I. Comparison of single-session transarterial chemoembolization combined with microwave ablation or radiofrequency ablation in the treatment of hepatocellular carcinoma: a randomized-controlled study. Eur J Gastroenterol Hepatol 2016; 28: 1198-1203 [PMID: 27362551 DOI: 10.1097/MEG.000000000000688]
- 65 Yuan P, Zhang Z, Kuai J. Analysis on efficacy and safety of TACE in combination with RFA and MWA in the treatment of middle and large primary hepatic carcinoma. J BUON 2019; 24: 163-170 [PMID: 30941966]
- Abdelaziz AO, Abdelmaksoud AH, Nabeel MM, Shousha HI, Cordie AA, Mahmoud ShH, Medhat 66 E, Omran D, Elbaz TM. Transarterial Chemoembolization Combined with Either Radiofrequency or Microwave Ablation in Management of Hepatocellular Carcinoma. Asian Pac J Cancer Prev 2017; 18: 189-194 [PMID: 28240516 DOI: 10.22034/APJCP.2017.18.1.189]
- Vasnani R, Ginsburg M, Ahmed O, Doshi T, Hart J, Te H, Van Ha TG. Radiofrequency and 67 microwave ablation in combination with transarterial chemoembolization induce equivalent histopathologic coagulation necrosis in hepatocellular carcinoma patients bridged to liver transplantation. Hepatobiliary Surg Nutr 2016; 5: 225-233 [PMID: 27275464 DOI: 10.21037/hbsn.2016.01.05]
- Lewis AR, Padula CA, McKinney JM, Toskich BB. Ablation plus Transarterial Embolic Therapy for Hepatocellular Carcinoma Larger than 3 cm: Science, Evidence, and Future Directions. Semin Intervent Radiol 2019; 36: 303-309 [PMID: 31680721 DOI: 10.1055/s-0039-1697641]
- Zangos S, Eichler K, Balzer JO, Straub R, Hammerstingl R, Herzog C, Lehnert T, Heller M, 69 Thalhammer A, Mack MG, Vogl TJ. Large-sized hepatocellular carcinoma (HCC): a neoadjuvant treatment protocol with repetitive transarterial chemoembolization (TACE) before percutaneous



MR-guided laser-induced thermotherapy (LITT). Eur Radiol 2007; 17: 553-563 [PMID: 16896704 DOI: 10.1007/s00330-006-0343-x]

- 70 Su TS, Lu HZ, Cheng T, Zhou Y, Huang Y, Gao YC, Tang MY, Jiang HY, Lian ZP, Hou EC, Liang P. Long-term survival analysis in combined transarterial embolization and stereotactic body radiation therapy vs stereotactic body radiation monotherapy for unresectable hepatocellular carcinoma >5 cm. BMC Cancer 2016; 16: 834 [PMID: 27809890 DOI: 10.1186/s12885-016-2894-9]
- Wang B, Xu H, Gao ZQ, Ning HF, Sun YQ, Cao GW. Increased expression of vascular endothelial 71 growth factor in hepatocellular carcinoma after transcatheter arterial chemoembolization. Acta Radiol 2008; 49: 523-529 [PMID: 18568538 DOI: 10.1080/02841850801958890]
- 72 Peck-Radosavljevic M, Kudo M, Raoul J-L, Lee HC, Decaens T, Heo J, Lin S-M, Shan H, Yang Y, Bayh I, Nakajima K, Cheng A-L. Outcomes of patients with hepatocellular carcinoma (HCC) treated with transarterial chemoembolization (TACE): Global OPTIMIS final analysis. J Clin Oncol 2018; **36**: 4018 [DOI: 10.1200/jco.2018.36.15_suppl.4018]
- Hiraoka A, Kumada T, Kudo M, Hirooka M, Koizumi Y, Hiasa Y, Tajiri K, Toyoda H, Tada T, 73 Ochi H, Joko K, Shimada N, Deguchi A, Ishikawa T, Imai M, Tsuji K, Michitaka K; Real-life Practice Experts for HCC (RELPEC) Study Group and HCC 48 Group (hepatocellular carcinoma experts from 48 clinics). Hepatic Function during Repeated TACE Procedures and Prognosis after Introducing Sorafenib in Patients with Unresectable Hepatocellular Carcinoma: Multicenter Analysis. Dig Dis 2017; 35: 602-610 [PMID: 29040999 DOI: 10.1159/000480256]
- 74 Lencioni R. Loco-regional treatment of hepatocellular carcinoma. Hepatology 2010; 52: 762-773 [PMID: 20564355 DOI: 10.1002/hep.23725]
- 75 Unruh A, Ressel A, Mohamed HG, Johnson RS, Nadrowitz R, Richter E, Katschinski DM, Wenger RH. The hypoxia-inducible factor 1α is a negative factor for tumor therapy. Oncogene 2003; 22: 3213-20 [DOI: 10.1038/sj.onc.1206385]
- 76 Jia ZZ, Jiang GM, Feng YL. Serum HIF-1alpha and VEGF levels pre- and post-TACE in patients with primary liver cancer. Chin Med Sci J 2011; 26: 158-162 [PMID: 22207924 DOI: 10.1016/s1001-9294(11)60041-2]
- Li X, Feng GS, Zheng CS, Zhuo CK, Liu X. Expression of plasma vascular endothelial growth 77 factor in patients with hepatocellular carcinoma and effect of transcatheter arterial chemoembolization therapy on plasma vascular endothelial growth factor level. World J Gastroenterol 2004; 10: 2878-2882 [PMID: 15334691 DOI: 10.3748/wjg.v10.i19.2878]
- Jia ZZ, Huang YQ, Feng YL, Jiang GM. [Correlations between serum hypoxia inducible factor-1a, 78 vascular endothelial growth factor and computed tomography perfusion imaging at pre-and post-TACE in patients with primary hepatic carcinoma]. Zhonghua Yi Xue Za Zhi 2013; 93: 1472-1475 [PMID: 24029570]
- Geschwind JF, Kudo M, Marrero JA, Venook AP, Chen XP, Bronowicki JP, Dagher L, Furuse J, 79 Ladrón de Guevara L, Papandreou C, Sanyal AJ, Takayama T, Ye SL, Yoon SK, Nakajima K, Lehr R, Heldner S, Lencioni R. TACE Treatment in Patients with Sorafenib-treated Unresectable Hepatocellular Carcinoma in Clinical Practice: Final Analysis of GIDEON. Radiology 2016; 279: 630-640 [PMID: 26744927 DOI: 10.1148/radiol.2015150667]
- 80 Lencioni R, Llovet JM, Han G, Tak WY, Yang J, Guglielmi A, Paik SW, Reig M, Kim DY, Chau GY, Luca A, Del Arbol LR, Leberre MA, Niu W, Nicholson K, Meinhardt G, Bruix J. Sorafenib or placebo plus TACE with doxorubicin-eluting beads for intermediate stage HCC: The SPACE trial. J Hepatol 2016; 64: 1090-1098 [PMID: 26809111 DOI: 10.1016/j.jhep.2016.01.012]
- Meyer T, Fox R, Ma YT, Ross PJ, James MW, Sturgess R, Stubbs C, Stocken DD, Wall L, Watkinson A, Hacking N, Evans TRJ, Collins P, Hubner RA, Cunningham D, Primrose JN, Johnson PJ, Palmer DH. Sorafenib in combination with transarterial chemoembolisation in patients with unresectable hepatocellular carcinoma (TACE 2): a randomised placebo-controlled, double-blind, phase 3 trial. Lancet Gastroenterol Hepatol 2017; 2: 565-575 [PMID: 28648803 DOI: 10.1016/S2468-1253(17)30156-5]
- 82 Park JW, Kim YJ, Kim DY, Bae SH, Paik SW, Lee YJ, Kim HY, Lee HC, Han SY, Cheong JY, Kwon OS, Yeon JE, Kim BH, Hwang J. Sorafenib with or without concurrent transarterial chemoembolization in patients with advanced hepatocellular carcinoma: The phase III STAH trial. J Hepatol 2019; 70: 684-691 [PMID: 30529387 DOI: 10.1016/j.jhep.2018.11.029]
- Kudo M, Ueshima K, Ikeda M, Torimura T, Tanabe N, Aikata H, Izumi N, Yamasaki T, Nojiri S, 83 Hino K, Tsumura H, Kuzuya T, Isoda N, Yasui K, Aino H, Ido A, Kawabe N, Nakao K, Wada Y, Yokosuka O, Yoshimura K, Okusaka T, Furuse J, Kokudo N, Okita K, Johnson PJ, Arai Y; TACTICS study group. Randomised, multicentre prospective trial of transarterial chemoembolisation (TACE) plus sorafenib as compared with TACE alone in patients with hepatocellular carcinoma: TACTICS trial. Gut 2020; 69: 1492-1501 [PMID: 31801872 DOI: 10.1136/gutjnl-2019-318934]
- Kudo M, Ueshima K, Ikeda M, Torimura T, Aikata H, Izumi N, Yamasaki T, Hino K, Kuzuya T, Isoda N, Yasui K, Aino H, Ido A, Kawabe N, Nakao K, Wada Y, Yoshimura K, Okusaka T, Furuse J, Arai Y. TACTICS: Final overall survival (OS) data from a randomized, open label, multicenter, phase II trial of transcatheter arterial chemoembolization (TACE) therapy in combination with sorafenib as compared with TACE alone in patients (pts) with hepatocellular. J Clin Oncol 2021; 39: 270 [DOI: 10.1200/jco.2021.39.3 suppl.270]
- 85 Kudo M, Han G, Finn RS, Poon RT, Blanc JF, Yan L, Yang J, Lu L, Tak WY, Yu X, Lee JH, Lin SM, Wu C, Tanwandee T, Shao G, Walters IB, Dela Cruz C, Poulart V, Wang JH. Brivanib as



adjuvant therapy to transarterial chemoembolization in patients with hepatocellular carcinoma: A randomized phase III trial. Hepatology 2014; 60: 1697-1707 [PMID: 24996197 DOI: 10.1002/hep.27290]

- 86 Kudo M, Cheng AL, Park JW, Park JH, Liang PC, Hidaka H, Izumi N, Heo J, Lee YJ, Sheen IS, Chiu CF, Arioka H, Morita S, Arai Y. Orantinib vs placebo combined with transcatheter arterial chemoembolisation in patients with unresectable hepatocellular carcinoma (ORIENTAL): a randomised, double-blind, placebo-controlled, multicentre, phase 3 study. Lancet Gastroenterol Hepatol 2018; 3: 37-46 [PMID: 28988687 DOI: 10.1016/S2468-1253(17)30290-X]
- 87 Wu J, Ng J, Christos PJ, Goldenberg AS, Sparano J, Sung MW, Hochster HS, Muggia FM. Chronic thalidomide and chemoembolization for hepatocellular carcinoma. Oncologist 2014; 19: 1229-1230 [PMID: 25361625 DOI: 10.1634/theoncologist.2014-0283]
- Pinter M, Ulbrich G, Sieghart W, Kölblinger C, Reiberger T, Li S, Ferlitsch A, Müller C, Lammer 88 J, Peck-Radosavljevic M. Hepatocellular Carcinoma: A Phase II Randomized Controlled Double-Blind Trial of Transarterial Chemoembolization in Combination with Biweekly Intravenous Administration of Bevacizumab or a Placebo. Radiology 2015; 277: 903-912 [PMID: 26131911 DOI: 10.1148/radiol.2015142140]
- 89 Tong H, Wei B, Chen S, Xie YM, Zhang MG, Zhang LH, Huang ZY, Tang CW. Adjuvant celecoxib and lanreotide following transarterial chemoembolisation for unresectable hepatocellular carcinoma: a randomized pilot study. Oncotarget 2017; 8: 48303-48312 [PMID: 28430638 DOI: 10.18632/oncotarget.15684]
- 90 Kawamura Y, Kobayashi M, Shindoh J, Kobayashi Y, Okubo S, Tominaga L, Kajiwara A, Kasuya K. Iritani S. Fujiyama S. Hosaka T. Saitoh S. Sezaki H. Akuta N. Suzuki F. Suzuki Y. Ikeda K. Arase Y, Hashimoto M, Kozuka T, Kumada H. Lenvatinib-Transarterial Chemoembolization Sequential Therapy as an Effective Treatment at Progression during Lenvatinib Therapy for Advanced Hepatocellular Carcinoma. Liver Cancer 2020; 9: 756-770 [PMID: 33442544 DOI: 10.1159/000510299]
- 91 Hiroishi K, Eguchi J, Baba T, Shimazaki T, Ishii S, Hiraide A, Sakaki M, Doi H, Uozumi S, Omori R, Matsumura T, Yanagawa T, Ito T, Imawari M. Strong CD8(+) T-cell responses against tumorassociated antigens prolong the recurrence-free interval after tumor treatment in patients with hepatocellular carcinoma. J Gastroenterol 2010; 45: 451-458 [PMID: 19936602 DOI: 10.1007/s00535-009-0155-2
- 92 Greten TF, Duffy AG, Korangy F. Hepatocellular carcinoma from an immunologic perspective. Clin Cancer Res 2013; 19: 6678-6685 [PMID: 24030702 DOI: 10.1158/1078-0432.CCR-13-1721]
- 93 Xu F, Jin T, Zhu Y, Dai C. Immune checkpoint therapy in liver cancer. J Exp Clin Cancer Res 2018; 37: 110 [PMID: 29843754 DOI: 10.1186/s13046-018-0777-4]
- Voron T, Colussi O, Marcheteau E, Pernot S, Nizard M, Pointet AL, Latreche S, Bergaya S, 94 Benhamouda N, Tanchot C, Stockmann C, Combe P, Berger A, Zinzindohoue F, Yagita H, Tartour E, Taieb J, Terme M. VEGF-A modulates expression of inhibitory checkpoints on CD8+ T cells in tumors. J Exp Med 2015; 212: 139-148 [PMID: 25601652 DOI: 10.1084/jem.20140559]
- 95 Yoo SY, Badrinath N, Woo HY, Heo J. Oncolytic Virus-Based Immunotherapies for Hepatocellular Carcinoma. Mediators Inflamm 2017; 2017: 5198798 [PMID: 28512387 DOI: 10.1155/2017/5198798
- 96 Fagnoni FF, Zerbini A, Pelosi G, Missale G. Combination of radiofrequency ablation and immunotherapy. Front Biosci 2008; 13: 369-381 [PMID: 17981554 DOI: 10.2741/2686]
- Gameiro SR, Higgins JP, Dreher MR, Woods DL, Reddy G, Wood BJ, Guha C, Hodge JW. 97 Combination therapy with local radiofrequency ablation and systemic vaccine enhances antitumor immunity and mediates local and distal tumor regression. PLoS One 2013; 8: e70417 [PMID: 23894654 DOI: 10.1371/journal.pone.0070417]
- 98 Duffy AG, Ulahannan SV, Makorova-Rusher O, Rahma O, Wedemeyer H, Pratt D, Davis JL, Hughes MS, Heller T, ElGindi M, Uppala A, Korangy F, Kleiner DE, Figg WD, Venzon D, Steinberg SM, Venkatesan AM, Krishnasamy V, Abi-Jaoudeh N, Levy E, Wood BJ, Greten TF. Tremelimumab in combination with ablation in patients with advanced hepatocellular carcinoma. J Hepatol 2017; 66: 545-551 [PMID: 27816492 DOI: 10.1016/j.jhep.2016.10.029]
- Ma H, Zhang Y, Wang Q, Li Y, He J, Wang H, Sun J, Pan K, Chen M, Xia J. Therapeutic safety and 99 effects of adjuvant autologous RetroNectin activated killer cell immunotherapy for patients with primary hepatocellular carcinoma after radiofrequency ablation. Cancer Biol Ther 2010; 9: 903-907 [PMID: 20364106 DOI: 10.4161/cbt.9.11.11697]
- Cui J, Wang N, Zhao H, Jin H, Wang G, Niu C, Terunuma H, He H, Li W. Combination of 100 radiofrequency ablation and sequential cellular immunotherapy improves progression-free survival for patients with hepatocellular carcinoma. Int J Cancer 2014; 134: 342-351 [PMID: 23825037 DOI: 10.1002/iic.28372]
- Lee JH, Lee JH, Lim YS, Yeon JE, Song TJ, Yu SJ, Gwak GY, Kim KM, Kim YJ, Lee JW, Yoon 101 JH. Sustained efficacy of adjuvant immunotherapy with cytokine-induced killer cells for hepatocellular carcinoma: an extended 5-year follow-up. Cancer Immunol Immunother 2019; 68: 23-32 [PMID: 30232520 DOI: 10.1007/s00262-018-2247-4]
- 102 Lee JH, Lee JH, Lim YS, Yeon JE, Song TJ, Yu SJ, Gwak GY, Kim KM, Kim YJ, Lee JW, Yoon JH. Adjuvant immunotherapy with autologous cytokine-induced killer cells for hepatocellular carcinoma. Gastroenterology 2015; 148: 1383-91.e6 [PMID: 25747273 DOI: 10.1053/j.gastro.2015.02.055]


- 103 Tu J, Ji J, Wu F, Zhang D, Ying X, Zhao Z. [Efficacies of ¹³¹I-chTNT plus radiofrequency ablation for the treatment of advanced hepatocellular carcinoma]. Zhonghua Yi Xue Za Zhi 2014; 94: 3586-3588 [PMID: 25622840]
- 104 Bian H, Zheng JS, Nan G, Li R, Chen C, Hu CX, Zhang Y, Sun B, Wang XL, Cui SC, Wu J, Xu J, Wei D, Zhang X, Liu H, Yang W, Ding Y, Li J, Chen ZN. Randomized trial of [1311] metuximab in treatment of hepatocellular carcinoma after percutaneous radiofrequency ablation. J Natl Cancer Inst 2014; 106 [PMID: 25210200 DOI: 10.1093/jnci/dju239]
- 105 Guan Q, Gu J, Zhang H, Ren W, Ji W, Fan Y. Correlation between vascular endothelial growth factor levels and prognosis of hepatocellular carcinoma patients receiving radiofrequency ablation. Biotechnol Biotechnol Equip 2015; 29: 119-123 [PMID: 26019624 DOI: 10.1080/13102818.2014.981776
- 106 Kong J, Kong J, Pan B, Ke S, Dong S, Li X, Zhou A, Zheng L, Sun WB. Insufficient radiofrequency ablation promotes angiogenesis of residual hepatocellular carcinoma via HIF-1α /VEGFA. PLoS One 2012; 7: e37266 [PMID: 22615958 DOI: 10.1371/journal.pone.0037266]
- 107 Tak WY, Lin SM, Wang Y, Zheng J, Vecchione A, Park SY, Chen MH, Wong S, Xu R, Peng CY, Chiou YY, Huang GT, Cai J, Abdullah BJJ, Lee JS, Lee JY, Choi JY, Gopez-Cervantes J, Sherman M, Finn RS, Omata M, O'Neal M, Makris L, Borys N, Poon R, Lencioni R. Phase III HEAT Study Adding Lyso-Thermosensitive Liposomal Doxorubicin to Radiofrequency Ablation in Patients with Unresectable Hepatocellular Carcinoma Lesions. Clin Cancer Res 2018; 24: 73-83 [PMID: 29018051 DOI: 10.1158/1078-0432.CCR-16-2433]
- Celik H, Wakim P, Pritchard WF, Castro M, Leonard S, Karanian JW, Dewhirst MW, Lencioni R, 108 Wood BJ. Radiofrequency Ablation Duration per Tumor Volume May Correlate with Overall Survival in Solitary Hepatocellular Carcinoma Patients Treated with Radiofrequency Ablation Plus Lyso-Thermosensitive Liposomal Doxorubicin. J Vasc Interv Radiol 2019; 30: 1908-1914 [PMID: 31409568 DOI: 10.1016/j.jvir.2019.04.023]
- 109 Finn RS, Qin S, Ikeda M, Galle PR, Ducreux M, Kim TY, Kudo M, Breder V, Merle P, Kaseb AO, Li D, Verret W, Xu DZ, Hernandez S, Liu J, Huang C, Mulla S, Wang Y, Lim HY, Zhu AX, Cheng AL; IMbrave150 Investigators. Atezolizumab plus Bevacizumab in Unresectable Hepatocellular Carcinoma. N Engl J Med 2020; 382: 1894-1905 [PMID: 32402160 DOI: 10.1056/NEJMoa1915745
- 110 Kudo M, Chung H, Osaki Y. Prognostic staging system for hepatocellular carcinoma (CLIP score): its value and limitations, and a proposal for a new staging system, the Japan Integrated Staging Score (JIS score). J Gastroenterol 2003; 38: 207-215 [PMID: 12673442 DOI: 10.1007/s005350300038]
- Morimoto M, Numata K, Kondo M, Moriya S, Morita S, Maeda S, Tanaka K. Radiofrequency 111 ablation combined with transarterial chemoembolization for subcapsular hepatocellular carcinoma: a prospective cohort study. Eur J Radiol 2013; 82: 497-503 [PMID: 23068563 DOI: 10.1016/j.ejrad.2012.09.014]
- 112 Hyun D, Cho SK, Shin SW, Rhim H, Koh KC, Paik SW. Treatment of Small Hepatocellular Carcinoma (≤2 cm) in the Caudate Lobe with Sequential Transcatheter Arterial Chemoembolization and Radiofrequency Ablation. Cardiovasc Intervent Radiol 2016; 39: 1015-1022 [PMID: 26975761 DOI: 10.1007/s00270-016-1314-5]
- 113 Hyun D, Cho SK, Shin SW, Park KB, Lee SY, Park HS, Do YS. Combined transarterial chemoembolization and radiofrequency ablation for small treatment-naïve hepatocellular carcinoma infeasible for ultrasound-guided radiofrequency ablation: long-term outcomes. Acta Radiol 2018; 59: 773-781 [PMID: 29034691 DOI: 10.1177/0284185117735349]
- 114 Yan JY, Zhang JL, Wang MQ, Yuan K, Bai YH, Wang Y, Xin HN, Wang ZJ, Liu FY, Duan F, Fu JX. Combined transcatheter arterial chemoembolization and radiofrequency ablation in singlesession for solitary hepatocellular carcinoma larger than 7 cm. Asia Pac J Clin Oncol 2018; 14: 300-309 [PMID: 29124894 DOI: 10.1111/ajco.12817]
- 115 Kim AR, Park E, Kwon SY, Park SJ, Kim YJ, Yoo BC, Choe WH, Kim JH, Hwang JH, Park SW, Park HS, Yu MH, Jeon HJ. Efficacy and Safety of Combined Radiofrequency Ablation with Transarterial Chemoembolization in Patients with Barcelona Clinic Liver Cancer Stage A Hepatocellular Carcinoma Ineligible for Curative Treatment. Korean J Gastroenterol 2019; 73: 167-176 [PMID: 31013560 DOI: 10.4166/kjg.2019.73.3.167]
- 116 Duan F, Bai YH, Cui L, Li XH, Yan JY, Wang MQ. Simultaneous transarterial chemoembolization and radiofrequency ablation for large hepatocellular carcinoma. World J Gastrointest Oncol 2020; 12: 92-100 [PMID: 31966917 DOI: 10.4251/wjgo.v12.i1.92]
- 117 Zhang Y, Zhang MW, Fan XX, Mao DF, Ding QH, Zhuang LH, Lv SY. Drug-eluting beads transarterial chemoembolization sequentially combined with radiofrequency ablation in the treatment of untreated and recurrent hepatocellular carcinoma. World J Gastrointest Surg 2020; 12: 355-368 [PMID: 32903981 DOI: 10.4240/wjgs.v12.i8.355]
- Wang Y, Ma L, Yuan Z, Zheng J, Li W. Percutaneous thermal ablation combined with TACE vs 118 TACE monotherapy in the treatment for liver cancer with hepatic vein tumor thrombus: A retrospective study. PLoS One 2018; 13: e0201525 [PMID: 30063737 DOI: 10.1371/journal.pone.0201525
- 119 Song Q, Ren W, Fan L, Zhao M, Mao L, Jiang S, Zhao C, Cui Y. Long-Term Outcomes of Transarterial Chemoembolization Combined with Radiofrequency Ablation Versus Transarterial Chemoembolization Alone for Recurrent Hepatocellular Carcinoma After Surgical Resection. Dig Dis Sci 2020; 65: 1266-1275 [PMID: 31312995 DOI: 10.1007/s10620-019-05733-0]



- 120 Thornton LM, Cabrera R, Kapp M, Lazarowicz M, Vogel JD, Toskich BB. Radiofrequency vs Microwave Ablation After Neoadjuvant Transarterial Bland and Drug-Eluting Microsphere Chembolization for the Treatment of Hepatocellular Carcinoma. Curr Probl Diagn Radiol 2017; 46: 402-409 [PMID: 28392205 DOI: 10.1067/j.cpradiol.2017.02.006]
- 121 Ni JY, Fang ZT, An C, Sun HL, Huang ZM, Zhang TQ, Jiang XY, Chen YT, Xu LF, Huang JH. Comparison of albumin-bilirubin grade, platelet-albumin-bilirubin grade and Child-Turcotte-Pugh class for prediction of survival in patients with large hepatocellular carcinoma after transarterial chemoembolization combined with microwave ablation. Int J Hyperthermia 2019; 36: 841-853 [PMID: 31452408 DOI: 10.1080/02656736.2019.1646927]



 \mathcal{O} WJ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 1919-1938

DOI: 10.4251/wjgo.v13.i12.1919

ISSN 1948-5204 (online)

REVIEW

Unique situation of hepatocellular carcinoma in Egypt: A review of epidemiology and control measures

Reem Ezzat, Mohamed Eltabbakh, Mohamed El Kassas

ORCID number: Reem Ezzat 0000-0001-8657-6796; Mohamed Eltabbakh 0000-0003-2836-807X; Mohamed El Kassas 0000-0002-3396-6894.

Author contributions: All authors equally contributed to this paper with conception and design of the work, literature review, drafting and critical revision, editing, and final approval of the final version of the manuscript.

Conflict-of-interest statement: None related to this work.

Country/Territory of origin: Egypt

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0)

Reem Ezzat, Internal Medicine Department, Faculty of Medicine, Assiut University, Assiut 71515, Egypt

Mohamed Eltabbakh, Tropical Medicine Department, Faculty of Medicine, Ain Shams University, Cairo 11566, Egypt

Mohamed El Kassas, Endemic Medicine Department, Faculty of Medicine, Helwan University, Cairo 11795, Cairo, Egypt

Corresponding author: Mohamed El Kassas, MD, Associate Professor, Endemic Medicine Department, Faculty of Medicine, Helwan University, Ain Helwan, Cairo 11795, Cairo, Egypt. m_elkassas@yahoo.com

Abstract

Hepatocellular carcinoma (HCC) is the sixth most common primary malignancy worldwide, and the third most common cause of death among cancers worldwide. HCC occurs in several pre-existing conditions, including hepatitis C, hepatitis B virus, and non-alcoholic cirrhosis. Egypt used to be the country with the heaviest hepatitis C virus (HCV) burden. The relationship between HCV and HCC is an important research area. In Egypt, HCC is a significant public health problem. A possible cause for the increasing rates of detection of HCC in Egypt is the mass screening program that was carried by the government for detecting and treating HCV. A multidisciplinary approach is now widely applied to HCC management in health centers all over Egypt. Different treatment modalities are available in Egypt, with success rates comparable to global rates. The Egyptian health authorities have made the elimination of HCV from Egypt a special priority, and this approach should lead to a decrease in number of HCC cases in the near future. In this article we review the current situation of HCC in Egypt, including epidemiological aspects, relevant risk factors for HCC development, strategies, and efforts established by health authorities for the screening and prevention of both HCV and HCC in Egypt. We highlight the different modalities for HCC treatment.

Key Words: Hepatocellular carcinoma; Liver cancer; Hepatitis C virus; Hepatitis B virus; Screening; Egypt

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.



license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: March 12, 2021 Peer-review started: March 12, 2021 First decision: April 6, 2021 Revised: April 17, 2021 Accepted: October 18, 2021 Article in press: October 18, 2021 Published online: December 15, 2021

P-Reviewer: Zhu Y S-Editor: Gao CC L-Editor: A P-Editor: Gao CC



Core Tip: Hepatocellular carcinoma (HCC) is the sixth most common cancer worldwide, and the fourth most common in Egypt. Many risk factors may lead to the development of HCC, and the relationship between hepatitis C virus (HCV) and HCC in Egypt is an important research area. Major screening programs for HCV in Egypt, such as the national initiative for screening 65 million citizens, have produced high success rates on the way for eliminating the main risk factor for HCC in the country. It is now an appropriate time for principled guidance and screening programs for HCC in Egypt.

Citation: Ezzat R, Eltabbakh M, El Kassas M. Unique situation of hepatocellular carcinoma in Egypt: A review of epidemiology and control measures. World J Gastrointest Oncol 2021; 13(12): 1919-1938

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/1919.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.1919

INTRODUCTION

Hepatocellular carcinoma (HCC) is the sixth most common primary malignancy worldwide[1] with higher incidence and prevalence in Africa and Asia[2]. The incidence of HCC worldwide has changed over the past few years, with some areas showing decreased rates, and others showing the opposite[3].

In Egypt, the relation between hepatitis C virus (HCV) and HCC is an important research area. Firstly, Egypt has a high recorded HCV transmission rate, with around 416000 new infections each year[4]. Secondly, there is known to be a relationship between HCV and HCC development. Thirdly, the programmed screening and follow up that was initiated by the government increased the number of known cases of individuals having both diseases. According to a study carried out by Ziada *et al*[5], 108 out of 514 patients diagnosed with HCV infection (21%) had focal lesions detected by ultrasound. In another study carried out by Abd-Elsalam *et al*[6], HCC occurred more frequently in patients with HCV than in those with hepatitis B virus (HBV) infection. These results may indicate the main predisposing factor for the development of HCC in Egypt.

A possible cause for the increase in detection of HCC in Egypt is the mass screening program that was implemented by the government for detecting and treating HCV. Due to this program, many patients were diagnosed and treated for HCC. According to a study carried out by Shaker et al[7], 75% of identified HCC cases came from rural areas in Egypt, with 45.7% of individuals ranging in age between 51-60 years.

According to the global cancer observatory, liver cancer represented 19% of all newly diagnosed cases in all ages and both sexes in 2018, with an incidence rate of 32% and a mortality rate of 31%[8].

RISK FACTORS FOR HCC IN EGYPT

HCV

HCV protein expression in infected hepatic cells causes mutation and malignant transformation leading to the development of HCC[9-11]. Repeated inflammation, damage and regeneration are believed to be the main cause of malignant transformation[12]. HCV infection increases the risk of HCC development up to 20-fold[13]. About 0.5%10% of HCV-related cirrhosis leads to HCC annually [14]. There are other factors that increase the risk of developing HCC with HCV, such as male gender, smoking, obesity, diabetes, and HBV or human immunodeficiency virus co-infection [14,15]. During the era of interferon-based therapy, sustained virologic response (SVR) and HCV eradication was associated with decreased incidence of HCC[16]. This outcome was hoped for from the direct-acting antiviral agents (DAAs), but researchers could not reach an agreement on that point. Reig *et al*[17] found an early recurrence of HCC in patients receiving DAA (27.6%). This was not the case in other studies, that found no increase in recurrence after DAA therapy[18-21].



Another review was published by Reig *et al*[17], debating about revising the published data, and they concluded that no solid evidence could be reached about the relationship between HCC recurrence and DAA therapy[22]. El Kassas *et al*[23] concluded that there is a possible role of DAAs in HCC recurrence.

Egypt recorded the highest prevalence of HCV worldwide, as a consequence of unsafe IV treatment of schistosomiasis in 1950s until the 1980s[24]. A decline was recorded in the prevalence of HCV infection from 14.7% in 2008 to 10% in 2015. This was attributed to the aging of the group who received antischistosomal treatment[25, 26].

In Egypt, genotype 4 is the main genotype, occurring in up to 92.5% of infected patients, followed by genotype 1 (3.6%)[27-30]. A study demonstrated that at least in Egypt, the lymphotoxin alpha gene mutation may have a role in susceptibility to HCV infection, and the subsequent development of clinical manifestations[31].

HBV

DNA viruses can be incorporated into a host genome[32], inducing malignant transformation by downregulating tumor suppressor genes and activating oncogenes[9]. The annual incidence of HCC is 0.42%[33] which differs according to the presence of HBV infection or cirrhosis[34], with the lifetime risk of HCC development among HBV carriers being from 10% to 25%[32]. Antiviral treatment for HBV can decrease HBV-DNA levels[35], with improved liver function and histology. There is increasing evidence that nucleos(t)ide analogs (NAs) decrease, but do not eliminate, the risk of HCC development[36,37].

In Egypt, the population prevalence of HBV was 1.4%, with an HBV-HCV coinfection rate of 0.06%[38]. The nationwide vaccination program has decreased the prevalence of HBV infection considerably[39,40]. The HBeAg negative variant was found to be highly prevalent in Egypt, and represents a late phase of HBV infection with persistent viral replication. This situation will lead to early development of cirrhosis[41]. However, 16% of patients with HCV have an occult B infection[42] A study carried out by Fouad *et al*[43] found that 81.9% of their chronic HBV cohort were HBeAg negative.

Of patients with liver cirrhosis, 3%-5% develop HCC annually[44]. In Egypt, HCC represents nearly 70% of all liver tumors[45]. The increased incidence in Egypt may be related to the increased screening carried by the government. and a greater focus on HBV and HCV as predisposing factors in the past few years[46].

Environmental toxins

The liver is the main organ involved in the metabolism of chemical agents[47]. It has a characteristic blood supply, and is involved in many metabolic and excretory processes. This causes damage to the liver ranging from fatty liver, hepatocellular injury, cirrhosis, and HCC.

In Egypt, nearly 26% of the population works in agriculture[48], and thus have a high risk of exposure to pesticides. A study carried out by Abou El Azm *et al*[49] found that 13.87 % of the total HCC in Egypt was associated with risk factors other than HVB or HCV, predominantly pesticides, and superphosphate and ammonium sulfate fertilizers (94.87%, P < 0.001) with significant exposure occurring in industry, farming, and residences. The HCC in these cases had specific criteria, being solitary, of smaller size, and having lower alpha fetoprotein (AFP) titers[49].

Aflatoxins are known to have a major role in the development of HCC in Egypt. They are known carcinogenic metabolites of molds, mainly *Aspergillus flavus*, and parasites that contaminate many agricultural products, such as peanuts, maize, and cotton seed[50].

Beside molds, a study conducted on desserts in Egypt showed that aflatoxin B1 (AFB1) was detected at above the acceptable limits of 2 ppb in 70% of samples of one of the dairy desserts, and Aflatoxin M1 exceeded the limits in 10% of each type of sample[51]. High serum levels were detected in Egyptians with HCC by a study that was carried out by Dilber *et al*[52]. AFB1 is the main metabolite produced, and is the most carcinogenic, teratogenic, and mutagenic metabolite[53]. It was present in high levels in those presenting with multiple hepatic focal lesions over 5 cm in diameter [54]. Anwar *et al*[55] found that presence of Aflatoxins and HCV is connected to hepatic disease progression to G3S3 which indicates HCC. Aflatoxin levels were found to be significantly higher in HCC patients than in cirrhotic individuals and controls in a study conducted by Sharaf-Eldin *et al*[56].

Zaishideng® WJGO | https://www.wjgnet.com

Non-alcoholic fatty liver disease

Non-alcoholic fatty liver disease (NAFLD) produces abnormal fat accumulation in the liver, without significant alcohol ingestion. NAFLD includes a broad spectrum of liver conditions ranging from steatosis and reaching up to cirrhosis. It is considered to be the most common liver disease related to obesity [57], and is a condition that can progress to HCC[58]. HCC development is related to disease progression from NAFLD to non-alcoholic steatohepatitis (NASH). NAFLD can accelerate the disease burden of HCV in terms of morbidity and mortality[59]. A study that was carried out on school children in Egypt, fatty liver was prevalent in 15.8% of the study group, and increased significantly with age (P = 0.004)[60]. NAFLD (56.8%) was a predominant feature among the study population in a study that was conducted by Abd El-Wahab EW et al[61] on 190 adults seeking health check-ups at the outpatient clinic of a tertiary care hospital in Alexandria, Egypt. Fatty liver was detected in 47 (65.3%) children, and in 52 (62.7%) adults in another Egyptian study by Wafaa *et al*[59]. A study concluded that NASH is present in 5.3% of Egyptian patients presenting with HCC[62]. This finding reflects the high prevalence of the condition in Egypt, and the subsequent increased risk of HCC transformation. Screening and early detection of the condition indicates the importance of avoiding further burdens on public health, as in the campaign carried out by the Egyptian government last year with respect to the detection of obesity, diabetes, and hypertension as predisposing causes for NAFLD.

Lifestyle factors (alcohol consumption, smoking, and dietary factors)

Excessive alcohol consumption is a well-known risk factor for developing HCC[62]. In the European Union, 60%-80% of liver-related mortality is caused by excessive drinking[63] and alcohol-related chronic disease is considered to be the second most common indication for liver transplantation, accounting for approximately 40% of all primary liver transplants[64]. In Egypt, this risk is low[48,65-67]. Heavy alcohol consumption increases the risk of HCC by up to 16% [68]. The risk is increased by 5- to 7-fold with heavy ethanol consumption for more than 10 years [69,70].

Smoking is another factor that may lead to HCC, due to the tobacco [71]. A Korean study reported a connection between primary liver cancer and smoking with the risk increased by up to 50% compared with non-smokers [72]. Bakir and Ali-Eldin [73] concluded that 64% of Egyptian patients with HCC are smokers. Abou El Azm et al [49] reported that heavy smoking is one of the primary risk factors for non-B non-C HCC in Egypt. Another study mentioned smoking as one of the main causative agents for HCC in Egypt[5]. Another Egyptian study documented an increased risk of HCC development in patients with a smoking pattern of 20 cigarettes per day for more than 29 years[74].

Obesity

Around 1.9 billion people around the world are overweight, and 600 million are suffering from obesity [75]. Obesity is related to the development of many metabolic disorders, including diabetes mellitus and hypertension, with an increased burden of HCC development. Premorbid obesity is associated with up to a two-fold risk of HCC related mortality^[75]. It has been suggested that for every 5 unit increase in body mass index (BMI), there is a 39% increased risk of HCC[76]. In another study, carried out by Calle *et al*[77], the HCC related mortality in obese men (BMI, 30-34.9 kg/m²) was 1.9 times the number in men with normal BMI (BMI 18.5-24.9 kg/m²).

In Egypt, a study on primary school students showed that the overall prevalence of obesity and overweight was 13.9% and 16.2% respectively [78]. In adults, it is estimated to be present in 61%70% of the whole population aged 20 and above, with a prevalence of 18%22% in men and 39%48% in women[79]. Aitsi-Selmi et al[80] investigated the relationship between wealth, education, and obesity among 49058 Egyptian women using the Demographic and Health Surveys' datasets. Obesity was mainly recorded among women with a primary education or less, and whether they are poor or wealthy. A survey of young people in Egypt[81] found that consuming more white bread and carbonated drinks is directly related to their economic state.

Genetic factors

Some hereditary liver diseases with genetic mutations are believed to carry a risk for HCC development. These diseases are Wilson disease, hemochromatosis, alpha-1 antitrypsin deficiency, tyrosinemia, glycogen storage diseases, and porphyrias. The same is true for polymorphisms with increased risk for HCC. Polymorphisms in *UGT1A7*, *MnSOD*, and *IL-1B* were reported to be significantly associated with risk [82]. HCV and HBV infection are reported to increase the risk of gene mutation,



leading to the development of HCC[83-87].

In an Egyptian study, the TNF- α -308 G > A polymorphism was associated with increased HCC risk in an Egyptian population, but no significant difference was found for cytokines interleukin (IL)-1β and IL-10[88]. In another study on Egyptian patients, XRCC1 G28152A (rs25487) and XRCC7 G6721T (rs7003908) polymorphisms were found to have a role in susceptibility to HCC in the Egyptian population[89].

Epidermal growth factor gene polymorphism 61*G was found to be positively associated with HCC risk in Egyptians. Uncreased concentration of EGF was associated with the G/G genotype[90].

The prevalence of hereditary hemochromatosis in Egypt is reported to be 0.5%[50]. This indicates that hereditary disorders are not a major cause of HCC.

Preventive measures: HCV control in Egypt

In 2015, 10% of the population tested positive for HCV antibodies, which would amount to around 5.5 million persons at that time[91]. As a major cause of HCC in Egypt, after the World Health Assembly's decision in 2016 to eliminate HCV, the Egyptian government decided to begin a nationwide campaign for the detection and treatment of HCV in Egypt[92]. More than two million individuals were treated by the year 2018 under the umbrella of this campaign, with cure rates reaching 90%. Disease elimination was achieved mostly by the decrease in the cost of direct-acting antiviral drugs implemented by the Egyptian government. This process was first applied to identified chronic patients. The government then began mass screening of the population, to facilitate rapid and effective elimination of the disease. Screening was done in all of the hospitals related to universities, military hospitals, rural health units, and police hospitals. This was achieved by moving teams to other areas, using gathering spaces, factories, and open places to aid in the screening. Finger prick rapid diagnostic tests were used. Patients reported positive were scheduled for evaluation and treatment plans. Between October 2018 and April 2019, 79.4% of the targeted population participated spontaneously in the screening, with higher female than male participation (84.5% vs 74.6%). By the end of September 2019, 1148346 (76.5%) of screened individuals were reported to have viremia, and treatment was started in 91.8% of them. Out of these people, 465992 reached 12-wk follow up after ending treatment. At this stage, 386103 (82.9%) had a known treatment outcome, and 381491 (98.8%) of those with a known outcome had a SVR. Of the 93651 patients with viremia who did not show up for treatment, 53445 who were reached reported having treatment in private[93].

There have been no screening programs for HCC in Egypt until now. Because HCC in Egypt is mostly diagnosed early, as more patients are diagnosed under surveillance, the survival duration is longer than in other African countries[94]. The effect of treatment itself is controversial. A study carried by El Kassas *et al*^[23] reported: "Our data point to a high (*i.e.*, almost 4 times) increased rate of recurrence after DAA treatment for patients with a history of successfully treated HCC, when compared to similar patients who were not given DAAs".

After HCV elimination, decreased rates of HCC were expected, but Reig et al[17] found exactly the opposite tendency after using direct-acting antiviral drugs for HCV treatment.

This work was followed by a paper that emphasized the early occurrence of HCC in patients receiving DAAs for HCV[18]. Another study produced different results, in which no difference in the cumulative incidence was found in developing de novo HCC in patients with HCV and those treated by DAAs or interferon-based therapy [95]. Similar results were reached by Cabibbo et al[96]. A study on patients with HCVrelated cirrhosis treated with DAAs and subsequently developing HCC reported a relation between age, Child-Pugh classification, liver stiffness, history of HCC, and the development of HCC[18]. In 2019, a study on 7344 patients concluded that DAAs decrease the risk of developing HCC[97].

An Egyptian study concluded that DAAs do not increase the risk of HCC recurrence, but still did not recommend abolishing it, rather implementing close follow up[98]. Another study denied the occurrence of HCC after DAAs although a high incidence of recurrence was still found. This study also suggested that high AFP before treatment is a good predictor for developing HCC[99].

Immunization for HBV and protection against HCC was discussed in a study on an analysis of 1509 patients with HCC in Taiwan. The study concluded that risk reduction of HCC is obvious after immunization of infants against HBV[100]. The HBV vaccination program in Egypt began in 1992 with a schedule of 2, 4, and 6 mo of age. This program was not associated with simultaneous screening for pregnant women [101]. A multicenter study was carried on 3600 children aging from 9 mo to 16 years

old to assess the effectiveness of the Egyptian vaccination program. The study concluded that the vaccination is protective from 1 years to 16 years post vaccination [40]. Another study assessed the benefit of follow up post vaccination response and seroprotection persistence, to determine the importance of booster doses in healthy subjects. A protective level of HBsAb was found (> 10 IU/mL) among 66.7% of all individuals studied[102]. The risk of HCC danger is escalated by co-infection with occult HBV in HCV patients[103].

HCC SCREENING

Screening programs gain value when the benefits from screening are greater than the expected harm. A large randomized controlled trial showed benefits for screening noncirrhotic HBV patients for the development of HCC, leading to improved early detection, better treatment, and better survival rates[104]. An association between screening for HCC and improvement in three-year survival rates is well established [105]. A study observed the difference between the survival rates of HCC in Japan and in Hong Kong. Japan has an intensive screening program unlike Hong Kong. The survival rate was 52 vs 17.8 mo[106]. In spite of the psychological or financial harm that could result from screening for HCC, the benefits overweigh the harm. Cirrhotic patients show an annual risk of 2%-4% of developing HCC which makes screening highly recommended in all cirrhotic patients whatever the etiology [107,108].

The risk of progression to HCC in non-cirrhotic patients has ranges from 7% to 54%, varying according to etiology and geographic distribution [109]. The most common etiological factors for this condition are obesity, aflatoxins, NAFLD, genetic mutations, smoking, inherited diseases, and sex hormones[107,109-112]. Non-cirrhotic liver HCC has a better prognosis and better results following surgical intervention than cirrhotic liver HCC[113]. In the European Association for the Study of the Liver 2018 report, a risk stratification model was recommended for non-cirrhotic HCC patients, namely PAGE-B (platelet, age, gender, hepatitis B), that is currently used in non-cirrhotic HBV patients[114,115].

The risk of HCC development in cirrhotic patients is from 2% to 4% annually. This high risk makes screening an obligation for all cirrhotic patients, whatever their etiology[107,111]. Screening is mainly to be done for compensated cirrhosis with Child-Pugh class A and B, while class C is to be offered liver transplantation[116].

Screening methods

Ultrasound is the most widely used imaging technique for regular screening for HCC. It has many advantages, being easy, readily available, non-invasive, and inexpensive. The sensitivity of ultrasound in detecting HCC is not more than 45% [117], especially in lesions less than 1 cm in diameter[118]. It is affected by the operator, the patient ability to hold their breath during examination, and the nodularity of the liver, which makes the detection of new lesions difficult, with some areas unreachable, like the dome of the liver. Obesity and NASH renders examination difficult which, decreasing the efficacy of the procedure[119]. In such cases, magnetic resonance imaging (MRI) and computed tomography (CT) scanning can replace ultrasound[120,121], but they are not cost effective, so they are not considered as first-line screening methods for HCC[119, 121].

Biomarkers

AFP is the biomarker most widely used in screening for HCC[122]. Although it is readily available, inexpensive, and easy to perform, its addition in the guidelines along with ultrasound was debatable. The American Association for the Study of Liver Diseases recommends using ultrasound, with the use of AFP to be judged by the clinician according to the patient's condition[107]. However, European guidelines recommend using ultrasound with no AFP needed[123].

In Egypt, HCV is the main etiological factor for liver cirrhosis, followed by HCC. Liver elastography is a documented method for assessing liver stiffness. A study investigated its role in the early detection of HCC in HCV cirrhotic patients. It recorded cutoff value of 24 kPa for diagnostic prediction of HCC produced sensitivity 100%, specificity 83.3%, PPV 94.5%, NPV 77.3%, and AUC 89%[124]. Another study discussed the superiority of an abbreviated MRI protocol over AFP and ultrasound in detecting small hepatic focal lesions in post HCV cirrhotic patients[125].

A scoring system was suggested by Abdelaziz *et al*[126]. The HCC Multidisciplinary Clinic-Cairo University (HMC-CU) score (Logit probability of HCC = -2.524 + 0.152 ×



age -0.121 × Hb -0.696 × INR -1.059 × Alb + 0.022 × AFP + 0.976 × Sex. Male = 1, Female = 0), with a cutoff of 0.559 was superior to other scores for predicting HCC, having a sensitivity of 90% and a specificity of 80.6%. In 2010, El-Zayadi et al[127] investigated the effect of surveillance of HCC on tumor staging and treatment options in Egypt. The study divided the patients into two groups: (1) For those who followed screening regularly; and (2) Who were diagnosed as HCC as first presentation with no screening program followed. They produced variable results three months after interval screening was suggested, as the doubling time of the tumor size is from 1 mo to 19 mo, and as HCV is the main predisposing factor in Egypt. The study reported that surveillance increased the detection of small lesions in the absence of vascular invasion

A prospective study carried out by Gomaa et al[46] on 2000 patients diagnosed with HCC reported that BCLC has the best prognostic stratification for Egyptians with HCC. Salama et al[128] suggested adding leptin to AFP for HCC screening in Egyptians. All of these studies were trials from separate centers to detect and screen for HCC in Egypt.

TREATMENT OF HCC IN EGYPT

HCC is a disease with different modalities of treatment. Surgical resection comes in the first place, followed by liver transplantation. Ablative techniques come next, including ethanol (percutaneous ethanol injection), microwave (MWA) or radiofrequency (RFA), catheter-directed trans arterial chemoembolization (TACE) or radioembolization (TARE). Last comes external beam radiation therapy in the form of stereotactic body radiation therapy or proton beam therapy, systemic targeted small molecule tyrosine kinase inhibitors (TKIs), check-point inhibitor immunotherapy, and investigational agents.

A multidisciplinary approach has been now widely recognized and is the mainstay in managing HCC in different health centers all over Egypt. This approach includes a scientific committee with the patient of HCC presented to it, and through discussion is performed, along with counseling the patient with different treatment options.

Surgical intervention

Surgery for HCC includes tumor resection or liver transplantation. Liver transplantation is the best choice, as the whole organ is replaced by a new one, and the underlying pathology is ended forever. However, this is not possible in all cases. Milan criteria were developed to diagnose a patient's suitability as a candidate for liver transplantation[129]. When it is inconvenient to do transplantation, surgical resection of HCC comes next. In non-cirrhotic patients, tumors less than 5 cm are best offered resection as the best treatment modality from an oncological point of view [130]. However, partial resection carries the risk of tumor recurrence[131]. Tumor size is not a contraindication for partial hepatectomy, but other factors such as extrahepatic metastasis, vascular invasion, main bile duct affection and portal hypertension may affect the decision[132]. In an Egyptian study carried out by Zakaria et al[132], the researchers concluded that total tumor volume is an appropriate prognostic measure to evaluate the tumor burden in HCC patients. Assessment of the hepatic function and future remnant liver are cornerstones in the liver resection decision[133]. A study carried out by Senbel et al[134] concluded that hepatic resection is an effective treatment for Child-Pugh A patient candidates for liver transplant. A study at Assuit university hospital reported 28 cases that underwent hepatic resection for HCC from 2013-2017. The study defined low serum albumin, high MELD score and high Child score to be risk factors for developing post-resection liver failure[135] in 268 patients who had undergone hepatic resection between the years 2010 and 2019 in Mansoura University, Egypt[136].

In Arab countries, 3804 liver transplants were done between the period 1990-2013, of which living donor liver transplantation (LDLT) was 80%, and deceased donor liver transplantation was 20%. Fifty-six percent of the reported cases were in Egypt[137]. In Egypt, the only source for a liver graft is from a living donor. From 2001 to 2019, 1230 cases of liver transplantation were reported from three transplantation centers in Egypt. Of them, 394 cases were HCC transplanted patients. In a retrospective study done by the surgical team in Dar ALfouad, Egypt, 60 patients with HCC who had undergone liver transplant within and beyond the Milan criteria were investigated for their clinical outcome. The results were as follows: "Overall 1-, 3-, and 5-year survival rates were 98.3%, 93.5%, and 71.4%. Overall disease-free survival rates at 1, 3, and 5



years were 96.6%, 93.5%, and 64.2%. There was no statistically significant difference in overall survival time between patients within and beyond the Milan criteria. Factors affecting recurrence were the tumor grade, lobar distribution, size of the largest nodule, and the total tumor burden in the explanted liver" [138]. In a study done by Galal *et al*[139], the researchers concluded that AFP may predict HCC recurrence after LDLT (area under the curve = 0.806) at cutoff values of more than 66 ng/mL, with 60% sensitivity, 94.3% specificity, 42.9% positive predictive value, and 97.1% negative predictive value.

We in Egypt have certain constants regarding liver transplantation as an option for HCC treatment, the major issue being the high cost of the operation, and the difficulty of finding a proper matched donor, as only living donor transplant is allowed in Egypt. Nevertheless, the success rate of liver transplant in Egypt is comparable to international results. So, it became of importance to allow health insurance coverage for liver transplantation operations in public health centers as a better treatment option for Egyptian HCC patients.

Local ablation techniques

Using thermal ablation for hepatic focal lesions has many advantages, such as the ability to repeating the maneuver, low morbidity and very few complications[140]. MWA ablation provides better results in areas with high blood flow, or near vessels, because it is not affected by the heat sink effect[141]. An Egyptian study carried out by Soliman *et al*[142] aimed to investigate the efficacy of MWA ablation in risky areas adjacent to other organs, near the diaphragm, and near blood vessels. In the study group, MWA reached ablation rates of 100%, 75%, and 87.5% for lesions close to the gall bladder, perivascular lesions, and subcapsular lesions, respectively. Another study done at Menoufia university, Egypt, compared single local ablative and combined techniques in HCC. The combined locoregional method provided better results[143]. However, Kamal et al[144] found no difference between MWA ablation and RFA ablation in treating HCC. Due to the high incidence of HCC related HCV in Egypt, the high risk of recurrence in those patients was investigated by Sharaf-Eldin et al[145]. The study concluded that in those patients, the presence of hepatomegaly, heterogenous liver, and splenomegaly, a sign of portal hypertension, together with tumor factors such as large size, bilobar affliction, and lesions near the liver capsule, showed a significant association with tumor recurrence.

TACE

TACE is the treatment of choice for patients with intermediate stage HCC, according to BCLC[123]. It is also the standard treatment in non-resectable HCC[107]. It is considered to be a palliative treatment, with positive impacts on survival and quality of life[146]. Since Seldinger described his technique in 1953, many intravascular procedures have been used[147]. This was followed by percutaneous selective angiography and arterial infusion of vasopressin by catheterization for controlling gastrointestinal bleeding. For more than a decade there was a debate about the use of chemotherapy to support TACE over trans arterial embolization[148]. Many studies supported TACE for providing both embolectomy and chemotherapy, and for keeping a good hepatic reserve for better survival[149-151]. TACE is not used only in nonresectable HCC, but also for downstaging before liver transplantation[152], and has good outcomes and overall survival [153]. Farouk Ahmed et al [154] found that the main etiology for HCC in Egypt is HCV. Patients who were inappropriate for transplantation, being outside of the Milan criteria, were chosen for downstaging by TACE before transplant. The study showed that good selection of patients for downstaging by TACE has good outcomes on liver transplantation. The patients' quality of life post TACE was evaluated by Fouad et al[155] in a study on 99 patients with HCC. The study showed improved quality of life after three months. In another Egyptian pilot study, RFA ablation showed better results with respect to quality of life than TACE [156].

TARE

Guidelines recommend TACE as the standard line of treatment for BCLC-B, but the results are still not very satisfactory [157]. Radiation from external beams to the liver is not effective in delivering lethal doses, as HCC is radio-resistant[158]. Radioembolization with Yttrium-90 microspheres is a recently used catheter-based treatment for HCC. It can be performed safely in patients with portal vein thrombosis, due to its low embolic effect[159]. TARE has the advantages of short hospital stay[160], prolonged time until progression[148], and long progression free survival[161].



Hamed et al[162] investigated the efficacy of Yttrium-90 on 20 Egyptian patients with intermediate and advanced HCC, with good outcomes even in the presence of compromised liver functions. Similar results were produced by Hetta *et al*[163], in a study in which TARE was investigated in advanced HCC with or without portal vein thrombosis. TARE Y90 showed the best results, especially in advanced stage disease, when compared to TACE in a study on 86 Egyptian patients with intermediate HCC [164].

Systemic therapies

Treatment for advanced HCC is now based on systemic therapy relying on TKIs, antiangiogenesis agents, and immunotherapy [123]. Before the development of sorafenib, no drug was available that could provide this improved overall survival in such patients[165]. Sorafenib is an oral multi-kinase inhibitor with anti-proliferative and anti-angiogenic properties. It acts by inhibiting vascular endothelial growth factor receptor (VEGFR) -2 and -3 tyrosine kinases, platelet-derived growth factor receptor (PDGFR)- β tyrosine kinases, and rapidly accelerated fibrosarcoma kinases[166]. Sorafenib was first used in cases with well-preserved liver function, but results from the Global Investigation of Therapeutic Decisions in Hepatocellular Carcinoma and of its treatment with sorafenib (GIDEON) found a similar safety profile, irrespective of Child-Pugh staging[167]. Routine use of sorafenib in patients with underlying liver dysfunction is not recommended.

Lenvatinib (Lenvima, Eisai) is an oral TKI of fibroblast growth factor receptor (FGFR), VEGFR, PDGFR-α, rearranged during transfection, and KIT. It has been accepted as a first-line therapy for unresectable HCC since August 2018[168]. Regorafenib (Stivarga, Bayer) came next. It is a potent oral inhibitor of angiopoietin-1 receptor (Tie2), VEGFR, PDGFR, and FGFR, and was studied by Bruix and colleagues in patients who did not respond to sorafenib. It was approved by the FDA based on this multinational study [169].

A study on sorafenib in Egypt claimed that it cannot be used except in patients with Child A and low disease burden[170]. The same recommendation was made by Abdel-Rahman et al[171]. When sorafenib was studied in Egyptian patients with advanced HCC, it gave better outcomes, overall survival, and progression free survival when compared to no treatment^[172]. It is, however, considered to be a costly treatment for the Egyptian patients, as was found in a study carried out by Hamdy Elsisi *et al*[173], in which they concluded that "sorafenib does offer increased survival and quality of life at an increased cost but at an incremental cost effective ratio that exceeds the nationally accepted cost-effectiveness threshold". Hanafy showed that a combination of sorafenib and low dose capecitabine is effective in advanced HCC in an Egyptian population[174].

A comprehensive summary of studies discussing the results of different treatment modalities for HCC in Egypt is presented in Table 1.

NATIONAL POPULATION-BASED CANCER REGISTRY PROGRAM

The Egyptian National Cancer Registry Program (NCRP) was launched in 2008 to represent a source for cancer incidence figures in Egypt[175]. NCRP stratified Egypt into 3 geographical areas: lower, middle, and upper. Data are regularly collected from specialized cancer treatment centers that are scattered all over the country map. Results of NCRP showed that HCC was the first among the most frequently observed cancers in lower and middle Egypt and the 2nd in upper Egypt (Figure 1).

HCC SCREENING AFTER HCV TREATMENT WITH DAAS

A major breakthrough was noted after the national campaigns of fighting and screening HCV, in which all of the population was screened for HCV, and basic laboratory results and ultrasonography were performed [93,176]. Many HCC patients were discovered and provided with treatment options. Despite the high safety profile of DAAs therapy, which enabled treatment of advanced cases and with expected lower incidence rate of HCC post-treatment, there were some contradictory reports on HCC incidence rates post SVR[177].

The major drawback in our campaign in Egypt was lack of a program after achieving SVR for continued screening for HCC after cure of HCV, with a resultant faulty impression of the patient that they were completely cured, with no need for



Table 1 Summary of studies discussing the results of different treatment modalities for hepatocellular carcinoma in Egypt

Treatment modality	Ref.	Design	Sample size	Summary of the most important results
Resection	Senbel <i>et al</i> [<mark>134</mark>]	Retrospective	84	Median OS was 50 mo
	Zakaria et al [<mark>132</mark>]	Retrospective	204	Predictors of decreased survival: serum AFP level > 400 ng/mL, TTV > 65.5 cm ³ , microvascular invasion, postoperative decompensation
	Makhlouf et al[135]	Retrospective	28	Predictors for developing post-resection liver failure: low serum albumin-higher child score
Liver transplant	Kamal <i>et al</i> [<mark>144</mark>]	Retrospective	60	Overall disease-free survival rates at 1, 3, and 5 yr were 96.6%, 93.5%, and 64.2%; Overall, 1-, 3-, and 5-yr survival rates were 98.3%, 93.5%, and 71.4%. Factors affecting recurrence were the tumor grade, lobar distribution, size of the largest nodule, and the total tumor burden in the explanted liver
	Galal <i>et al</i> [<mark>139</mark>]	Retrospective	75	AFP may predict HCC recurrence after LDLT (area under the curve = 0.806) at cutoff values of more than 66 ng/mL
MWA	Soliman et al [<mark>142</mark>]	Prospective	88	MWA reached ablation rates of 100%, 75%, and 87.5% for lesions close to the GB, perivascular lesions, and subcapsular lesions, respectively
Radio frequency	Sharaf-Eldin <i>et al</i> [145]	Retrospective	45	Hepatomegaly, heterogenous liver, and splenomegaly, a sign of portal hypertension, together with tumor factors such as large size, bilobar affliction, and lesions near the liver capsule, showed a significant association with tumor recurrence
	Nouh <i>et al</i> [<mark>143</mark>]	Prospective	60	Combined techniques (RFA and percutaneous ethanol injection) give the best results for management of HCCs in comparison with individual techniques
TACE	Farouk <i>et al</i> [<mark>154</mark>]	Retrospective	27	Successful TACE for down-staging of HCC can be achieved in the majority of carefully selected patients and is associated with excellent post transplantation outcome
	Fouad <i>et al</i> [<mark>155</mark>]	Prospective	99	Improved quality of life after three months of TACE
TARE	Hamed <i>et al</i> [<mark>162</mark>]	Prospective	20	The complete response, partial response, stable disease and disease progression rates for the study sample after 3 mo using the conventional RECIST criteria was 0%, 55%, 30% and 10%, while after 6 mo it became 0, 50%, 20% and 25% respectively
	Hetta <i>et al</i> [163]	Prospective	40	The overall response (complete or partial response) was exhibited by 9% of patients, stable disease exhibited by 80% of patients, progressive disease seen in 11% of patients after one month of TARE
	El Fouly <i>et al</i> [<mark>164</mark>]	Prospective	86	The median OS (TACE: 18 mo vs TARE Y-90: 16.4 mo) and the median TTP (TACE: 6.8 mo vs TARE Y-90: 13.3 mo) were not statistically different between TACE and TARE group
Systemic therapy	Nada <i>et al</i> [<mark>170</mark>]	Retrospective	130	The median overall survival of patients with HCC treated with sorafenib was 5 mo (CI: 4.166-5.834), and progression free survival was 4 mo (CI: 3.479-4.521)
	El Baghdady et al[<mark>172</mark>]	Prospective	55	The one-year OS was 0.0% vs 75.5% ($P = 0.008$) in control and sorafenib respectively. Median PFS was 5 mo vs 12 mo in control group and sorafenib respectively ($P = 0.008$). Sorafenib treatment showed a better outcome OS, PFS and QOL as compared to no- treatment in Egyptian patients with advanced Hepatocellular Carcinoma

HCC: Hepatocellular carcinoma; LDLT: Living donor liver transplantation; MWA: Microwave; OS: Overall survival; RFA: Radiofrequency; TACE: Trans arterial chemoembolization; TARE: Trans arterial radioembolization; RECIST: Response Evaluation Criteria in Solid Tumors; CI: Confidence interval; PFS: Progression-free survival; QOL: Quality of life; AFP: Alpha fetoprotein; TTV: Total tumor volume; TTP: Time to progression.

lifetime follow up and screening for HCC.

This is why it is important to highlight the importance of screening for HCC for all individuals with SVR for cirrhotic features for life. Increasing public awareness of the importance of the screening is warranted just as in the national screening campaign for breast cancer in Egypt 2020[178].

Major screening programs in Egypt, like the National Initiative of 100 Million Healthy Individuals and Breast Cancer 2020 have produced high success rates [178]. Now it is time for proper guidance and screening programs for HCC in Egypt.

CONCLUSION

HCC is a disease posing a rising burden in Egyptian society. HCV is the main etiology in our country, with an expected decline following the decline in HCV incidence. HBV is the second most important etiology in Egypt. Mass vaccination campaigns are the





Figure 1 Proportion and age standardized rate of liver cancer in lower, middle, and upper Egypt (results of the National Population-Based Cancer Registry Program). ¹Lower Egypt: Damietta National Cancer Registry [liver cancer has the highest proportion among the most frequently observed cancers (29.6%)]. Males: Proportion and age standardized rate (ASR): 41.7% and 81.0/100000. Females: Proportion and ASR:16.3% and 32.6/100000. ²Middle Egypt: Minya National Cancer Registry [Liver cancer has the highest proportion among the most frequently observed cancers (15.2%)]. Male: Proportion and ASR: 20.4% and 37.6/100000. Females: Proportion and ASR:8.9% and 13.7/100000. ³Upper Egypt: Aswan National Cancer Registry [Liver cancer has the 2nd highest proportion among the most frequently observed cancers (8.2%)]. Male: Proportion and ASR: 11.8% and 17.5/100000. Females: Proportion and ASR: 5.1% and 8.7/100000.

> only way to stop the disease and ameliorate its effects. A registry of the different modalities for management for HCC is still lacking in Egypt, and will require a more systematized effort between different centers. A national campaign is crucial for early diagnosis and management.

REFERENCES

- Forner A, Reig M, Bruix J. Hepatocellular carcinoma. Lancet 2018; 391: 1301-1314 [PMID: 1 29307467 DOI: 10.1016/S0140-6736(18)30010-2]
- 2 Petrick JL, Florio AA, Znaor A, Ruggieri D, Laversanne M, Alvarez CS, Ferlay J, Valery PC, Bray F, McGlynn KA. International trends in hepatocellular carcinoma incidence, 1978-2012. Int J Cancer 2020; 147: 317-330 [PMID: 31597196 DOI: 10.1002/ijc.32723]
- Valery PC, Laversanne M, Clark PJ, Petrick JL, McGlynn KA, Bray F. Projections of primary liver 3 cancer to 2030 in 30 countries worldwide. Hepatology 2018; 67: 600-611 [PMID: 28859220 DOI: 10.1002/hep.29498]
- 4 Kandeel AM, Talaat M, Afifi SA, El-Sayed NM, Abdel Fadeel MA, Hajjeh RA, Mahoney FJ. Case control study to identify risk factors for acute hepatitis C virus infection in Egypt. BMC Infect Dis 2012; 12: 294 [PMID: 23145873 DOI: 10.1186/1471-2334-12-294]
- 5 Ziada DH, El Sadany S, Soliman H, Abd-Elsalam S, Salama M, Hawash N, Selim A, Hamisa M, Elsabagh HM. Prevalence of hepatocellular carcinoma in chronic hepatitis C patients in Mid Delta, Egypt: A single center study. J Egypt Natl Canc Inst 2016; 28: 257-262 [PMID: 27378258 DOI: 10.1016/j.jnci.2016.06.001]
- Abd-Elsalam S, Elwan N, Soliman H, Ziada D, Elkhalawany W, Salama M, Hawash N, Arafa M, Badawi R, Shehata WM, Khalil HS, Elmashad N. Epidemiology of liver cancer in Nile delta over a decade: A single-center study. South Asian J Cancer 2018; 7: 24-26 [PMID: 29600229 DOI: 10.4103/sajc.sajc_82_17]
- Shaker MK, Abdella HM, Khalifa MO, El Dorry AK. Epidemiological characteristics of 7 hepatocellular carcinoma in Egypt: a retrospective analysis of 1313 cases. Liver Int 2013; 33: 1601-1606 [PMID: 23714212 DOI: 10.1111/liv.12209]
- 8 Ferlay J, Colombet M, Soerjomataram I, Mathers C, Parkin DM, Piñeros M, Znaor A, Bray F. Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. Int J Cancer 2019; 144: 1941-1953 [PMID: 30350310 DOI: 10.1002/ijc.31937]
- Tang A, Hallouch O, Chernyak V, Kamaya A, Sirlin CB. Epidemiology of hepatocellular carcinoma: target population for surveillance and diagnosis. Abdom Radiol (NY) 2018; 43: 13-25 [PMID: 28647765 DOI: 10.1007/s00261-017-1209-1]
- 10 El-Houseini ME, Ismail A, Abdelaal AA, El-Habashy AH, Abdallah ZF, Mohamed MZ, El-Hadidi M, Cho WCS, Ahmed H, Al-Shafie TA. Role of TGF-B1 and C-Kit Mutations in the Development of Hepatocellular Carcinoma in Hepatitis C Virus-Infected Patients: in vitro Study. Biochemistry (Mosc) 2019; 84: 941-953 [PMID: 31522676 DOI: 10.1134/S0006297919080108]



- 11 Neamatallah M, El-Bendary M, Elalfy H, Besheer T, El-Maksoud MA, Elhammady D, Abed S, Elegezy M, Kandeel L, Eldeib D, Mousa N, Abd El-Hafeez M, El-Gilany AH, Esmat G. Impact of Toll-like Receptors 2(TLR2) and TLR 4 Gene Variations on HCV Susceptibility, Response to Treatment and Development of Hepatocellular Carcinoma in Cirrhotic HCV Patients. Immunol Invest 2020; 49: 462-476 [PMID: 31615295 DOI: 10.1080/08820139.2019.1673772]
- 12 Borgia M, Dal Bo M, Toffoli G. Role of Virus-Related Chronic Inflammation and Mechanisms of Cancer Immune-Suppression in Pathogenesis and Progression of Hepatocellular Carcinoma. Cancers (Basel) 2021; 13 [PMID: 34503196 DOI: 10.3390/cancers13174387]
- Doi AM, Hill G, Seely J, Hailey JR, Kissling G, Bucher JR. alpha 2u-globulin nephropathy and 13 renal tumors in national toxicology program studies. Toxicol Pathol 2007; 35: 533-540 [PMID: 17562486 DOI: 10.1080/01926230701338941]
- 14 Samant H, Amiri HS, Zibari GB. Addressing the worldwide hepatocellular carcinoma: epidemiology, prevention and management. J Gastrointest Oncol 2021; 12: S361-S373 [PMID: 34422400 DOI: 10.21037/jgo.2020.02.08]
- 15 Chang KC, Wu YY, Hung CH, Lu SN, Lee CM, Chiu KW, Tsai MC, Tseng PL, Huang CM, Cho CL, Chen HH, Hu TH. Clinical-guide risk prediction of hepatocellular carcinoma development in chronic hepatitis C patients after interferon-based therapy. Br J Cancer 2013; 109: 2481-2488 [PMID: 24084770 DOI: 10.1038/bjc.2013.564]
- 16 El Kassas M, Elbaz T, Salaheldin M, Abdelsalam L, Kaseb A, Esmat G. Impact of treating chronic hepatitis C infection with direct-acting antivirals on the risk of hepatocellular carcinoma: The debate continues - A mini-review. J Adv Res 2019; 17: 43-48 [PMID: 31193326 DOI: 10.1016/j.jare.2019.03.001]
- 17 Reig M, Mariño Z, Perelló C, Iñarrairaegui M, Ribeiro A, Lens S, Díaz A, Vilana R, Darnell A, Varela M, Sangro B, Calleja JL, Forns X, Bruix J. Unexpected high rate of early tumor recurrence in patients with HCV-related HCC undergoing interferon-free therapy. J Hepatol 2016; 65: 719-726 [PMID: 27084592 DOI: 10.1016/j.jhep.2016.04.008]
- 18 Conti F, Buonfiglioli F, Scuteri A, Crespi C, Bolondi L, Caraceni P, Foschi FG, Lenzi M, Mazzella G, Verucchi G, Andreone P, Brillanti S. Early occurrence and recurrence of hepatocellular carcinoma in HCV-related cirrhosis treated with direct-acting antivirals. J Hepatol 2016; 65: 727-733 [PMID: 27349488 DOI: 10.1016/j.jhep.2016.06.015]
- 19 Ohki T, Sato K, Kondo M, Goto E, Sato T, Kondo Y, Akamatsu M, Sato S, Yoshida H, Koike Y, Obi S. Effectiveness of direct acting antiviral agents for hepatitis C virus related recurrent hepatocellular carcinoma patients who had multiple courses of recurrence. J Viral Hepat 2021 [PMID: 34312954 DOI: 10.1111/jvh.13579]
- 20 Imai K, Takai K, Hanai T, Suetsugu A, Shiraki M, Shimizu M. Sustained virological response by direct-acting antivirals reduces the recurrence risk of hepatitis C-related hepatocellular carcinoma after curative treatment. Mol Clin Oncol 2020; 12: 111-116 [PMID: 31929880 DOI: 10.3892/mco.2019.1956
- 21 Lui FH, Moosvi Z, Patel A, Hussain S, Duong A, Duong J, Nguyen DL. Decreased risk of hepatocellular carcinoma recurrence with direct-acting antivirals compared with no treatment for hepatitis C: a meta-analysis. Ann Gastroenterol 2020; 33: 293-298 [PMID: 32382233 DOI: 10.20524/aog.2020.0470]
- 22 Sapena V, Enea M, Torres F, Celsa C, Rios J, Rizzo GEM, Nahon P, Mariño Z, Tateishi R, Minami T, Sangiovanni A, Forns X, Toyoda H, Brillanti S, Conti F, Degasperi E, Yu ML, Tsai PC, Jean K, El Kassas M, Shousha HI, Omar A, Zavaglia C, Nagata H, Nakagawa M, Asahina Y, Singal AG, Murphy C, Kohla M, Masetti C, Dufour JF, Merchante N, Cavalletto L, Chemello LL, Pol S, Crespo J, Calleja JL, Villani R, Serviddio G, Zanetto A, Shalaby S, Russo FP, Bielen R, Trevisani F, Cammà C, Bruix J, Cabibbo G, Reig M. Hepatocellular carcinoma recurrence after direct-acting antiviral therapy: an individual patient data meta-analysis. Gut 2021 [PMID: 33741640 DOI: 10.1136/gutjnl-2020-323663]
- 23 El Kassas M, Funk AL, Salaheldin M, Shimakawa Y, Eltabbakh M, Jean K, El Tahan A, Sweedy AT, Afify S, Youssef NF, Esmat G, Fontanet A. Increased recurrence rates of hepatocellular carcinoma after DAA therapy in a hepatitis C-infected Egyptian cohort: A comparative analysis. J Viral Hepat 2018; 25: 623-630 [PMID: 29274197 DOI: 10.1111/jvh.12854]
- 24 El Kassas M, Elbaz T, Elsharkawy A, Omar H, Esmat G. HCV in Egypt, prevention, treatment and key barriers to elimination. Expert Rev Anti Infect Ther 2018; 16: 345-350 [PMID: 29506418 DOI: 10.1080/14787210.2018.1448709
- 25 Gomaa A, Allam N, Elsharkawy A, El Kassas M, Waked I. Hepatitis C infection in Egypt: prevalence, impact and management strategies. Hepat Med 2017; 9: 17-25 [PMID: 28553150 DOI: 10.2147/HMER.S113681]
- Kandeel A, Genedy M, El-Refai S, Funk AL, Fontanet A, Talaat M. The prevalence of hepatitis C 26 virus infection in Egypt 2015: implications for future policy on prevention and treatment. Liver Int 2017; 37: 45-53 [PMID: 27275625 DOI: 10.1111/liv.13186]
- 27 Roudot-Thoraval F. Epidemiology of hepatitis C virus infection. Clin Res Hepatol Gastroenterol 2021; 45: 101596 [PMID: 33610022 DOI: 10.1016/j.clinre.2020.101596]
- Leumi S, El Kassas M, Zhong J. Hepatitis C virus genotype 4: A poorly characterized endemic 28 genotype. J Med Virol 2021; 93: 6079-6088 [PMID: 34185316 DOI: 10.1002/jmv.27165]
- 29 Ghaderi-Zefrehi H. Gholami-Fesharaki M, Sharafi H, Sadeghi F, Alavian SM. The Distribution of Hepatitis C Virus Genotypes in Middle Eastern Countries: A Systematic Review and Meta-Analysis.



Hepat Mon 2016; 16: e40357 [PMID: 27826320 DOI: 10.5812/hepatmon.40357]

- Kouyoumjian SP, Chemaitelly H, Abu-Raddad LJ. Characterizing hepatitis C virus epidemiology in 30 Egypt: systematic reviews, meta-analyses, and meta-regressions. Sci Rep 2018; 8: 1661 [PMID: 29374178 DOI: 10.1038/s41598-017-17936-4]
- 31 Elsammak MY, Al-Sharkaweey RM, Ragab MS, Amin GM, Kandil MH. In Egyptians, a mutation in the lymphotoxin-alpha gene may increase susceptibility to hepatitis C virus but not that to schistosomal infection. Ann Trop Med Parasitol 2008; 102: 709-716 [PMID: 19000388 DOI: 10.1179/136485908X337599]
- 32 McGlynn KA, Petrick JL, London WT. Global epidemiology of hepatocellular carcinoma: an emphasis on demographic and regional variability. Clin Liver Dis 2015; 19: 223-238 [PMID: 25921660 DOI: 10.1016/j.cld.2015.01.001]
- 33 Gordon SC, Lamerato LE, Rupp LB, Li J, Holmberg SD, Moorman AC, Spradling PR, Teshale EH, Vijayadeva V, Boscarino JA, Henkle EM, Oja-Tebbe N, Lu M; CHeCS Investigators. Antiviral therapy for chronic hepatitis B virus infection and development of hepatocellular carcinoma in a US population. Clin Gastroenterol Hepatol 2014; 12: 885-893 [PMID: 24107395 DOI: 10.1016/j.cgh.2013.09.062
- 34 El-Serag HB. Epidemiology of viral hepatitis and hepatocellular carcinoma. *Gastroenterology* 2012; 142: 1264-1273.e1 [PMID: 22537432 DOI: 10.1053/j.gastro.2011.12.061]
- 35 Singal AG, El-Serag HB. Hepatocellular Carcinoma From Epidemiology to Prevention: Translating Knowledge into Practice. Clin Gastroenterol Hepatol 2015; 13: 2140-2151 [PMID: 26284591 DOI: 10.1016/j.cgh.2015.08.014]
- 36 Hsu YC, Wu CY, Lane HY, Chang CY, Tai CM, Tseng CH, Lo GH, Perng DS, Lin JT, Mo LR. Determinants of hepatocellular carcinoma in cirrhotic patients treated with nucleos(t)ide analogues for chronic hepatitis B. J Antimicrob Chemother 2014; 69: 1920-1927 [PMID: 24576950 DOI: 10.1093/jac/dku041]
- Nahon P, Vo Quang E, Ganne-Carrié N. Stratification of Hepatocellular Carcinoma Risk Following 37 HCV Eradication or HBV Control. J Clin Med 2021; 10 [PMID: 33477752 DOI: 10.3390/jcm10020353]
- Alavian SM, Haghbin H. Relative Importance of Hepatitis B and C Viruses in Hepatocellular 38 Carcinoma in EMRO Countries and the Middle East: A Systematic Review. Hepat Mon 2016; 16: e35106 [PMID: 27226803 DOI: 10.5812/hepatmon.35106]
- 39 Allison RD, Teleb N, Al Awaidy S, Ashmony H, Alexander JP, Patel MK. Hepatitis B control among children in the Eastern Mediterranean Region of the World Health Organization. Vaccine 2016; 34: 2403-2409 [PMID: 27043863 DOI: 10.1016/j.vaccine.2016.03.063]
- Salama II, Sami SM, Said ZN, El-Sayed MH, El Etreby LA, Rabah TM, Elmosalami DM, Abdel 40 Hamid AT, Salama SI, Abdel Mohsen AM, Emam HM, Elserougy SM, Hassanain AI, Abd Alhalim NF, Shaaban FA, Hemeda SA, Ibrahim NA, Metwally AM. Effectiveness of hepatitis B virus vaccination program in Egypt: Multicenter national project. World J Hepatol 2015; 7: 2418-2426 [PMID: 26464758 DOI: 10.4254/wjh.v7.i22.2418]
- 41 Fung J, Lai CL, Yuen MF. New paradigms for the treatment of chronic hepatitis B. J Gastroenterol Hepatol 2008; 23: 1182-1192 [PMID: 18637060 DOI: 10.1111/j.1440-1746.2008.05400.x]
- 42 Atti EA. HCC Burden in Egypt. Gastroenterol Hepatol 2015; 2: 00045 [DOI: 10.15406/ghoa.2015.02.00045
- 43 Fouad R, Abdo M, Eldeen HG, Sabry D, Atef M, Ahmed R, Zayed N. Influence of delta virus infection on the virologic status in Egyptian patients with chronic hepatitis B virus genotype D. J Med Virol 2016; 88: 837-842 [PMID: 26488214 DOI: 10.1002/jmv.24412]
- 44 Fares N, Péron JM. [Epidemiology, natural history, and risk factors of hepatocellular carcinoma]. Rev Prat 2013; 63: 216-217, 220 [PMID: 23513788]
- 45 Mokhtar N, Gouda I, Adel I. Cancer pathology registry 2003-2004 and time trend analysis. Malign Digest Sys Tumors 2007; 55-67
- Gomaa AI, Hashim MS, Waked I. Comparing staging systems for predicting prognosis and survival 46 in patients with hepatocellular carcinoma in Egypt. PLoS One 2014; 9: e90929 [PMID: 24603710 DOI: 10.1371/journal.pone.00909291
- Ledda C, Loreto C, Zammit C, Marconi A, Fago L, Matera S, Costanzo V, Fuccio Sanzà G, 47 Palmucci S, Ferrante M, Costa C, Fenga C, Biondi A, Pomara C, Rapisarda V. Noninfective occupational risk factors for hepatocellular carcinoma: A review (Review). Mol Med Rep 2017; 15: 511-533 [PMID: 28000892 DOI: 10.3892/mmr.2016.6046]
- 48 Omar A, Abou-Alfa GK, Khairy A, Omar H. Risk factors for developing hepatocellular carcinoma in Egypt. Chin Clin Oncol 2013; 2: 43 [PMID: 25841922 DOI: 10.3978/j.issn.2304-3865.2013.11.07
- 49 Abou El Azm AR, Yousef M, Mansour N, Awad A, El Dardiry S, Abdel Aziz I. New insights on non-B non-C hepatocellular carcinoma in mid Delta Region, Egypt. J Gastrointest Cancer 2014; 45: 276-283 [PMID: 24488435 DOI: 10.1007/s12029-013-9573-8]
- 50 Massarweh NN, El-Serag HB. Epidemiology of Hepatocellular Carcinoma and Intrahepatic Cholangiocarcinoma. Cancer Control 2017; 24: 1073274817729245 [PMID: 28975830 DOI: 10.1177/1073274817729245
- Khalifa MI, Shata RR. Mycobiota and Aflatoxins B1 and M1 Levels in Commercial and 51 Homemade Dairy Desserts in Aswan City, Egypt. J Adv Vet Res 2018; 8: 43-48
- 52 Dilber MS, Phelan A, Aints A, Mohamed AJ, Elliott G, Smith CI, O'Hare P. Intercellular delivery of



thymidine kinase prodrug activating enzyme by the herpes simplex virus protein, VP22. Gene Ther 1999; 6: 12-21 [PMID: 10341871 DOI: 10.1038/sj.gt.3300838]

- 53 Ismaiel A, Papenbrock J. Mycotoxins: producing fungi and mechanisms of phytotoxicity. Agriculture 2015; 5: 492-537 [DOI: 10.3390/agriculture5030492]
- 54 El-Farrash MA, Abdel-Wahab M, Rizk MS. Serum Aflatoxin level as a predictor of Hepatocarcinogenesis in HCV-infected Egyptians. Egypt J Med Microbiol 2008; 17: 83-90
- Anwar WA, Khaled HM, Amra HA, El-Nezami H, Loffredo CA. Changing pattern of 55 hepatocellular carcinoma (HCC) and its risk factors in Egypt: possibilities for prevention. Mutat Res 2008; 659: 176-184 [PMID: 18346933 DOI: 10.1016/j.mrrev.2008.01.005]
- Sharaf-Eldin M, Salah R, Soliman HH, Abdou SH, Abd-Elsalam S, Elkhalawany W, Mansour L, 56 Elsabagh HM, Khalil H. Aflatoxin As An Environmental Risk Factor Attributable To Liver Cancer In Nile Delta. Indian J Med Res Pharm Sci 2016; 3: 19-26 [DOI: 10.5281/zenodo.49353]
- 57 Hazlehurst JM, Tomlinson JW. Non-alcoholic fatty liver disease in common endocrine disorders. Eur J Endocrinol 2013; 169: R27-R37 [PMID: 23653455 DOI: 10.1530/EJE-13-0296]
- 58 Michelotti A, de Scordilli M, Palmero L, Guardascione M, Masala M, Roncato R, Foltran L, Ongaro E, Puglisi F. NAFLD-Related Hepatocarcinoma: The Malignant Side of Metabolic Syndrome. Cells 2021: 10 [PMID: 34440803 DOI: 10.3390/cells10082034]
- Wafaa ME, Shadia R, Nagwa AI, Yasser AE, Abeer M. Nour EA, Hebatallah F, Inas AR. 59 Frequency of non-alcoholic fatty liver disease in overweight/obese children and adults: clinical, sonographic picture and biochemical assessment. J Genet Eng Biotechnol 2012; 10: 221-227
- Alkassabany YM, Farghaly AG, El-Ghitany EM. Prevalence, risk factors, and predictors of 60 nonalcoholic fatty liver disease among schoolchildren: a hospital-based study in Alexandria, Egypt. Arab J Gastroenterol 2014; 15: 76-81 [PMID: 25097051 DOI: 10.1016/j.ajg.2014.05.002]
- 61 Abd El-Wahab EW, Zein El-Abedin RA, Ahmed WM, Shatat HZ. Validation of a Non-Laboratory Based Screening Tool for Predicting Non-Alcoholic Fatty Liver Disease in an Egyptian Setting. Am J Med Sci 2020; 360: 662-677 [PMID: 32739036 DOI: 10.1016/j.amjms.2020.06.020]
- 62 Bagnardi V, Rota M, Botteri E, Tramacere I, Islami F, Fedirko V, Scotti L, Jenab M, Turati F, Pasquali E, Pelucchi C, Galeone C, Bellocco R, Negri E, Corrao G, Boffetta P, La Vecchia C. Alcohol consumption and site-specific cancer risk: a comprehensive dose-response meta-analysis. Br J Cancer 2015; 112: 580-593 [PMID: 25422909 DOI: 10.1038/bjc.2014.579]
- 63 Cojocariu CE, Trifan AV, Gîrleanu I, Stanciu C. Alcoholic liver disease--epidemiology and risk factors. Rev Med Chir Soc Med Nat Iasi 2014; 118: 910-917 [PMID: 25581947]
- Burra P, Senzolo M, Adam R, Delvart V, Karam V, Germani G, Neuberger J; ELITA; ELTR Liver 64 Transplant Centers. Liver transplantation for alcoholic liver disease in Europe: a study from the ELTR (European Liver Transplant Registry). Am J Transplant 2010; 10: 138-148 [PMID: 19951276 DOI: 10.1111/j.1600-6143.2009.02869.x]
- Ezzat S, Abdel-Hamid M, Eissa SA, Mokhtar N, Labib NA, El-Ghorory L, Mikhail NN, Abdel-65 Hamid A, Hifnawy T, Strickland GT, Loffredo CA. Associations of pesticides, HCV, HBV, and hepatocellular carcinoma in Egypt. Int J Hyg Environ Health 2005; 208: 329-339 [PMID: 16217918 DOI: 10.1016/j.ijheh.2005.04.003]
- 66 Badawi AF, Michael MS. Risk factors for hepatocellular carcinoma in Egypt: the role of hepatitis-B viral infection and schistosomiasis. Anticancer Res 1999; 19: 4565-4569 [PMID: 10650811]
- Lehman EM, Soliman AS, Ismail K, Hablas A, Seifeldin IA, Ramadan M, El-Hamzawy H, 67 Shoushtari CS, Wilson ML. Patterns of hepatocellular carcinoma incidence in Egypt from a population-based cancer registry. Hepatol Res 2008; 38: 465-473 [PMID: 18042228 DOI: 10.1111/i.1872-034X.2007.00299.x
- Turati F, Galeone C, Rota M, Pelucchi C, Negri E, Bagnardi V, Corrao G, Boffetta P, La Vecchia 68 C. Alcohol and liver cancer: a systematic review and meta-analysis of prospective studies. Ann Oncol 2014; 25: 1526-1535 [PMID: 24631946 DOI: 10.1093/annonc/mdu020]
- Blonski W, Kotlyar DS, Forde KA. Non-viral causes of hepatocellular carcinoma. World J 69 Gastroenterol 2010; 16: 3603-3615 [PMID: 20677332 DOI: 10.3748/wjg.v16.i29.3603]
- 70 El-Serag HB, Rudolph KL. Hepatocellular carcinoma: epidemiology and molecular carcinogenesis. Gastroenterology 2007; 132: 2557-2576 [PMID: 17570226 DOI: 10.1053/j.gastro.2007.04.061]
- Rashed WM, Kandeil MAM, Mahmoud MO, Ezzat S. Hepatocellular Carcinoma (HCC) in Egypt: 71 A comprehensive overview. J Egypt Natl Canc Inst 2020; 32: 5 [PMID: 32372179 DOI: 10.1186/s43046-020-0016-x]
- 72 Yun YH, Jung KW, Bae JM, Lee JS, Shin SA, Min Park S, Yoo T, Yul Huh B. Cigarette smoking and cancer incidence risk in adult men: National Health Insurance Corporation Study. Cancer Detect Prev 2005; 29: 15-24 [PMID: 15734213 DOI: 10.1016/j.cdp.2004.08.006]
- 73 Bakir AS, Ali-Eldin ZA. Is diabetes mellitus a risk factor for hepatocellular carcinoma in Egyptian patients? J Am Sci 2012; 8: 353-358
- 74 Abdou Moustafa EF, Galal GM, Aly A, Mohammed K. Smoking and the risk of hepatocellular carcinoma among Egyptian patients. A preliminary case-control study. Arab J Gastroenterol 2009; 10: AB53-AB60 [DOI: 10.1016/j.ajg.2009.07.103]
- Gupta A, Das A, Majumder K, Arora N, Mayo HG, Singh PP, Beg MS, Singh S. Obesity is 75 Independently Associated With Increased Risk of Hepatocellular Cancer-related Mortality: A Systematic Review and Meta-Analysis. Am J Clin Oncol 2018; 41: 874-881 [PMID: 28537989 DOI: 10.1097/COC.00000000000388]
- Wang Y, Wang B, Shen F, Fan J, Cao H. Body mass index and risk of primary liver cancer: a meta-76



analysis of prospective studies. Oncologist 2012; 17: 1461-1468 [PMID: 22956536 DOI: 10.1634/theoncologist.2012-0066]

- 77 Calle EE, Rodriguez C, Walker-Thurmond K, Thun MJ. Overweight, obesity, and mortality from cancer in a prospectively studied cohort of U.S. adults. N Engl J Med 2003; 348: 1625-1638 [PMID: 12711737 DOI: 10.1056/NEJMoa021423]
- 78 Hamed A, Hassan A, Younis M, Kamal A. Prevalence of Obesity and Overweight among Primary Schools Children in Qena, Egypt. Egypt J Hosp Med 2019; 77: 4899-4905
- 79 Ellabany E, Abdel Nasser MA. Non-Communicable Disease Surveillance System, Egypt 2006. Ministry of Health and Population. Preventive and Primary Health Care Sector Preventive Sector. [cited 10 March 2021]. In: World Health Organization [Internet]. Available from: https://www.who.int/ncds/surveillance/steps/EgyptSTEPSPresentation.pdf
- 80 Aitsi-Selmi A, Chandola T, Friel S, Nouraei R, Shipley MJ, Marmot MG. Interaction between education and household wealth on the risk of obesity in women in Egypt. PLoS One 2012; 7: e39507 [PMID: 22761807 DOI: 10.1371/journal.pone.0039507]
- Population Council. Survey of Young People in Egypt. West Asia and North Africa Office. [cited 81 12 February 2021]. In: Population Council [Internet]. Available from: https://www.popcouncil.org/uploads/pdfs/2010PGY_SYPEFinalReport.pdf
- 82 Jin F, Xiong WJ, Jing JC, Feng Z, Qu LS, Shen XZ. Evaluation of the association studies of single nucleotide polymorphisms and hepatocellular carcinoma: a systematic review. J Cancer Res Clin Oncol 2011; 137: 1095-1104 [PMID: 21240526 DOI: 10.1007/s00432-010-0970-0]
- 83 Yang HI, Yuen MF, Chan HL, Han KH, Chen PJ, Kim DY, Ahn SH, Chen CJ, Wong VW, Seto WK; REACH-B Working Group. Risk estimation for hepatocellular carcinoma in chronic hepatitis B (REACH-B): development and validation of a predictive score. Lancet Oncol 2011; 12: 568-574 [PMID: 21497551 DOI: 10.1016/S1470-2045(11)70077-8]
- 84 Clifford RJ, Zhang J, Meerzaman DM, Lyu MS, Hu Y, Cultraro CM, Finney RP, Kelley JM, Efroni S, Greenblum SI, Nguyen CV, Rowe WL, Sharma S, Wu G, Yan C, Zhang H, Chung YH, Kim JA, Park NH, Song IH, Buetow KH. Genetic variations at loci involved in the immune response are risk factors for hepatocellular carcinoma. Hepatology 2010; 52: 2034-2043 [PMID: 21105107 DOI: 10.1002/hep.23943]
- Jiang DK, Sun J, Cao G, Liu Y, Lin D, Gao YZ, Ren WH, Long XD, Zhang H, Ma XP, Wang Z, 85 Jiang W, Chen TY, Gao Y, Sun LD, Long JR, Huang HX, Wang D, Yu H, Zhang P, Tang LS, Peng B, Cai H, Liu TT, Zhou P, Liu F, Lin X, Tao S, Wan B, Sai-Yin HX, Qin LX, Yin J, Liu L, Wu C, Pei Y, Zhou YF, Zhai Y, Lu PX, Tan A, Zuo XB, Fan J, Chang J, Gu X, Wang NJ, Li Y, Liu YK, Zhai K, Hu Z, Liu J, Yi Q, Xiang Y, Shi R, Ding Q, Zheng W, Shu XO, Mo Z, Shugart YY, Zhang XJ, Zhou G, Shen H, Zheng SL, Xu J, Yu L. Genetic variants in STAT4 and HLA-DQ genes confer risk of hepatitis B virus-related hepatocellular carcinoma. Nat Genet 2013; 45: 72-75 [PMID: 23242368 DOI: 10.1038/ng.2483]
- 86 Kumar V, Kato N, Urabe Y, Takahashi A, Muroyama R, Hosono N, Otsuka M, Tateishi R, Omata M, Nakagawa H, Koike K, Kamatani N, Kubo M, Nakamura Y, Matsuda K. Genome-wide association study identifies a susceptibility locus for HCV-induced hepatocellular carcinoma. Nat Genet 2011; 43: 455-458 [PMID: 21499248 DOI: 10.1038/ng.809]
- 87 Li S, Qian J, Yang Y, Zhao W, Dai J, Bei JX, Foo JN, McLaren PJ, Li Z, Yang J, Shen F, Liu L, Li S, Pan S, Wang Y, Li W, Zhai X, Zhou B, Shi L, Chen X, Chu M, Yan Y, Wang J, Cheng S, Shen J, Jia W, Liu J, Wen Z, Li A, Zhang Y, Zhang G, Luo X, Qin H, Chen M, Wang H, Jin L, Lin D, Shen H, He L, de Bakker PI, Zeng YX, Wu M, Hu Z, Shi Y, Zhou W. GWAS identifies novel susceptibility loci on 6p21.32 and 21q21.3 for hepatocellular carcinoma in chronic hepatitis B virus carriers. PLoS Genet 2012; 8: e1002791 [PMID: 22807686 DOI: 10.1371/journal.pgen.1002791]
- 88 Abdel-Azyem H, Abdel-Aziz A, Elbaz R, Eldesoky A And Abdel-Mageed WS. Single Nucleotide Polymorphism In Cytokines And Risk Of Hepatocellular Carcinoma In Egyptian Patients. Egypt J Genet Cytol 2016; 45: 245-259
- 89 Khaled IA, Zahran N, Saeed ME, Abdel-Aziz OA. Study of the relation between Egyptian patients with hepatocellular carcinoma and the genetic variations in DNA repair genes. J Blood Disord Transfus 2019; 10
- 90 EI Sergany HF, Mohamed AM, Madkour NK, Elsebeaey MA, Fared AM, Elshaer SS, Zahran FE, EI Deeb HH. Epidermal Growth Factor Gene Polymorphism in Egyptian Patients with Hepatocellular carcinoma related to Hepatitis C. J Gastroenterol Hepatol Res 2017; 6: 2481-2485
- Ministry of Health and Population; El-Zanaty and Associates; The DHS Program ICF 91 International. Egypt health issues survey 2015. Rockville, MD: Ministry of Health and Population, ICF International, October 2015. [cited 9 March 2021]. In: Dhsprogram [Internet]. Available from: https://dhsprogram.com/pubs/pdf/FR313/FR313.pdf
- 92 World Health Organization. Global health sector strategy on viral hepatitis 2016–2021: towards ending viral hepatitis. 2016. [cited 15 February 2021]. In: World Health Organization [Internet]. Available from: https://www.who.int/hepatitis/strategy2016-2021/ghss-hep/en
- 93 Waked I, Esmat G, Elsharkawy A, El-Serafy M, Abdel-Razek W, Ghalab R, Elshishiney G, Salah A, Abdel Megid S, Kabil K, El-Sayed MH, Dabbous H, El Shazly Y, Abo Sliman M, Abou Hashem K, Abdel Gawad S, El Nahas N, El Sobky A, El Sonbaty S, El Tabakh H, Emad E, Gemeah H, Hashem A, Hassany M, Hefnawy N, Hemida AN, Khadary A, Labib K, Mahmoud F, Mamoun S, Marei T, Mekky S, Meshref A, Othman A, Ragab O, Ramadan E, Rehan A, Saad T, Saeed R, Sharshar M, Shawky H, Shawky M, Shehata W, Soror H, Taha M, Talha M, Tealaab A, Zein M,



Hashish A, Cordie A, Omar Y, Kamal E, Ammar I, AbdAlla M, El Akel W, Doss W, Zaid H. Screening and Treatment Program to Eliminate Hepatitis C in Egypt. N Engl J Med 2020; 382: 1166-1174 [PMID: 32187475 DOI: 10.1056/NEJMsr1912628]

- 94 Yang JD, Altekruse SF, Nguyen MH, Gores GJ, Roberts LR. Impact of country of birth on age at the time of diagnosis of hepatocellular carcinoma in the United States. Cancer 2017; 123: 81-89 [PMID: 27571320 DOI: 10.1002/cncr.30246]
- Yoo SH, Kwon JH, Nam SW, Kim HY, Kim CW, You CR, Choi SW, Cho SH, Han JY, Song DS, 95 Chang UI, Yang JM, Lee HL, Lee SW, Han NI, Kim SH, Song MJ, Hwang S, Sung PS, Jang JW, Bae SH, Choi JY, Yoon SK. Early development of de novo hepatocellular carcinoma after directacting agent therapy: Comparison with pegylated interferon-based therapy in chronic hepatitis C patients. J Viral Hepat 2018; 25: 1189-1196 [PMID: 29660199 DOI: 10.1111/jvh.12918]
- 96 Cabibbo G, Petta S, Calvaruso V, Cacciola I, Cannavò MR, Madonia S, Distefano M, Larocca L, Prestileo T, Tinè F, Bertino G, Giannitrapani L, Benanti F, Licata A, Scalisi I, Mazzola G, Cartabellotta F, Alessi N, Barbàra M, Russello M, Scifo G, Squadrito G, Raimondo G, Craxì A, Di Marco V, Cammà C; Rete Sicilia Selezione Terapia - HCV (RESIST-HCV). Is early recurrence of hepatocellular carcinoma in HCV cirrhotic patients affected by treatment with direct-acting antivirals? Aliment Pharmacol Ther 2017; 46: 688-695 [PMID: 28791711 DOI: 10.1111/apt.14256]
- 97 Carrat F, Fontaine H, Dorival C, Simony M, Diallo A, Hezode C, De Ledinghen V, Larrey D, Haour G, Bronowicki JP, Zoulim F, Asselah T, Marcellin P, Thabut D, Leroy V, Tran A, Habersetzer F, Samuel D, Guyader D, Chazouilleres O, Mathurin P, Metivier S, Alric L, Riachi G, Gournay J, Abergel A, Cales P, Ganne N, Loustaud-Ratti V, D'Alteroche L, Causse X, Geist C, Minello A, Rosa I, Gelu-Simeon M, Portal I, Raffi F, Bourliere M, Pol S; French ANRS CO22 Hepather cohort. Clinical outcomes in patients with chronic hepatitis C after direct-acting antiviral treatment: a prospective cohort study. Lancet 2019; 393: 1453-1464 [PMID: 30765123 DOI: 10.1016/S0140-6736(18)32111-1]
- 98 Musa NI, Mohamed IE, Abohalima AS. Impact of treating chronic hepatitis C infection with directacting antivirals on the risk of hepatocellular carcinoma recurrence. Egypt Liver J 2020; 10: 26 [DOI: 10.1186/s43066-020-00035-x]
- Lashen SA, Shamseya MM, Madkour MA. Hepatocellular Carcinoma Occurrence/Recurrence after 99 Direct-Acting Antivirals for Hepatitis C in Egyptian Cohort: Single-Center Experience. Dig Dis 2019; 37: 488-497 [PMID: 31216532 DOI: 10.1159/000501072]
- 100 Chang MH, You SL, Chen CJ, Liu CJ, Lai MW, Wu TC, Wu SF, Lee CM, Yang SS, Chu HC, Wang TE, Chen BW, Chuang WL, Soon MS, Lin CY, Chiou ST, Kuo HS, Chen DS; Taiwan Hepatoma Study Group. Long-term Effects of Hepatitis B Immunization of Infants in Preventing Liver Cancer. Gastroenterology 2016; 151: 472-480.e1 [PMID: 27269245 DOI: 10.1053/j.gastro.2016.05.048]
- 101 Mansour E, Abdul-Rahim S, Batouty G, Zaghloul I, Abdel-Hadi S. Integration of hepatitis B immunization in the Expanded Program on Immunization of the Child Survival Project. J Egypt Public Health Assoc 1993; 68: 487-494 [PMID: 7775876]
- 102 El-Deen Mohamed NM, Abuo-El-Yazed AH, El-Deen Mohamed HM. Follow up of hepatitis b virus vaccine response in healthy individuals. Sci J Al-Azhar Med Fac Girls 2018; 2: 58-63
- Gaballah A, Shawky S, Elsawaf G, Shamsia M, Al Makdad A, Abd El Rahman M, Osman NA, 103 Islim H, Alhaifi A, Kader O. Virological profiles of HBV and HCV in hepatocellular carcinoma in Egypt and Yemen. Egypt J Med Microbiol 2018; 27: 7-17
- 104 Zhang BH, Yang BH, Tang ZY. Randomized controlled trial of screening for hepatocellular carcinoma. J Cancer Res Clin Oncol 2004; 130: 417-422 [PMID: 15042359 DOI: 10.1007/s00432-004-0552-0]
- 105 Singal AG, Pillai A, Tiro J. Early detection, curative treatment, and survival rates for hepatocellular carcinoma surveillance in patients with cirrhosis: a meta-analysis. PLoS Med 2014; 11: e1001624 [PMID: 24691105 DOI: 10.1371/journal.pmed.1001624]
- 106 Johnson P, Berhane S, Kagebayashi C, Satomura S, Teng M, Fox R, Yeo W, Mo F, Lai P, Chan SL, Tada T, Toyoda H, Kumada T. Impact of disease stage and aetiology on survival in hepatocellular carcinoma: implications for surveillance. Br J Cancer 2017; 116: 441-447 [PMID: 28081537 DOI: 10.1038/bjc.2016.422]
- 107 Heimbach JK, Kulik LM, Finn RS, Sirlin CB, Abecassis MM, Roberts LR, Zhu AX, Murad MH, Marrero JA. AASLD guidelines for the treatment of hepatocellular carcinoma. Hepatology 2018; 67: 358-380 [PMID: 28130846 DOI: 10.1002/hep.29086]
- 108 Kokudo N, Hasegawa K, Akahane M, Igaki H, Izumi N, Ichida T, Uemoto S, Kaneko S, Kawasaki S, Ku Y, Kudo M, Kubo S, Takayama T, Tateishi R, Fukuda T, Matsui O, Matsuyama Y, Murakami T. Arii S. Okazaki M. Makuuchi M. Evidence-based Clinical Practice Guidelines for Hepatocellular Carcinoma: The Japan Society of Hepatology 2013 update (3rd JSH-HCC Guidelines). Hepatol Res 2015; 45 [PMID: 25625806 DOI: 10.1111/hepr.12464]
- 109 Trevisani F, Frigerio M, Santi V, Grignaschi A, Bernardi M. Hepatocellular carcinoma in noncirrhotic liver: a reappraisal. Dig Liver Dis 2010; 42: 341-347 [PMID: 19828388 DOI: 10.1016/j.dld.2009.09.002]
- 110 Global Burden of Disease Liver Cancer Collaboration, Akinyemiju T, Abera S, Ahmed M, Alam N, Alemayohu MA, Allen C, Al-Raddadi R, Alvis-Guzman N, Amoako Y, Artaman A, Ayele TA, Barac A, Bensenor I, Berhane A, Bhutta Z, Castillo-Rivas J, Chitheer A, Choi JY, Cowie B, Dandona L, Dandona R, Dey S, Dicker D, Phuc H, Ekwueme DU, Zaki MS, Fischer F, Fürst T,



Hancock J, Hay SI, Hotez P, Jee SH, Kasaeian A, Khader Y, Khang YH, Kumar A, Kutz M, Larson H, Lopez A, Lunevicius R, Malekzadeh R, McAlinden C, Meier T, Mendoza W, Mokdad A, Moradi-Lakeh M, Nagel G, Nguyen Q, Nguyen G, Ogbo F, Patton G, Pereira DM, Pourmalek F, Qorbani M, Radfar A, Roshandel G, Salomon JA, Sanabria J, Sartorius B, Satpathy M, Sawhney M, Sepanlou S, Shackelford K, Shore H, Sun J, Mengistu DT, Topór-Mądry R, Tran B, Ukwaja KN, Vlassov V, Vollset SE, Vos T, Wakayo T, Weiderpass E, Werdecker A, Yonemoto N, Younis M, Yu C, Zaidi Z, Zhu L, Murray CJL, Naghavi M, Fitzmaurice C. The Burden of Primary Liver Cancer and Underlying Etiologies From 1990 to 2015 at the Global, Regional, and National Level: Results From the Global Burden of Disease Study 2015. JAMA Oncol 2017; 3: 1683-1691 [PMID: 28983565 DOI: 10.1001/jamaoncol.2017.3055]

- 111 Omata M, Cheng AL, Kokudo N, Kudo M, Lee JM, Jia J, Tateishi R, Han KH, Chawla YK, Shiina S, Jafri W, Payawal DA, Ohki T, Ogasawara S, Chen PJ, Lesmana CRA, Lesmana LA, Gani RA, Obi S, Dokmeci AK, Sarin SK. Asia-Pacific clinical practice guidelines on the management of hepatocellular carcinoma: a 2017 update. Hepatol Int 2017; 11: 317-370 [PMID: 28620797 DOI: 10.1007/s12072-017-9799-9
- Sastre J, Díaz-Beveridge R, García-Foncillas J, Guardeño R, López C, Pazo R, Rodriguez-Salas N, 112 Salgado M, Salud A, Feliu J. Clinical guideline SEOM: hepatocellular carcinoma. Clin Transl Oncol 2015; 17: 988-995 [PMID: 26607931 DOI: 10.1007/s12094-015-1451-3]
- 113 Schütte K, Schulz C, Poranzke J, Antweiler K, Bornschein J, Bretschneider T, Arend J, Ricke J, Malfertheiner P. Characterization and prognosis of patients with hepatocellular carcinoma (HCC) in the non-cirrhotic liver. BMC Gastroenterol 2014; 14: 117 [PMID: 24990270 DOI: 10.1186/1471-230X-14-117
- Fateen W, Ryder SD. Screening for hepatocellular carcinoma: patient selection and perspectives. J 114 Hepatocell Carcinoma 2017; 4: 71-79 [PMID: 28553624 DOI: 10.2147/JHC.S105777]
- 115 Seo YS, Jang BK, Um SH, Hwang JS, Han KH, Kim SG, Lee KS, Kim SU, Kim YS, Lee JI. Validation of risk prediction models for the development of HBV-related HCC: a retrospective multi-center 10-year follow-up cohort study. Oncotarget 2017; 8: 113213-113224 [PMID: 29348900 DOI: 10.18632/oncotarget.22375]
- 116 Frenette CT, Isaacson AJ, Bargellini I, Saab S, Singal AG. A Practical Guideline for Hepatocellular Carcinoma Screening in Patients at Risk. Mayo Clin Proc Innov Qual Outcomes 2019; 3: 302-310 [PMID: 31485568 DOI: 10.1016/j.mayocpiqo.2019.04.005]
- Marrero JA. Surveillance for Hepatocellular Carcinoma. Clin Liver Dis 2020; 24: 611-621 [PMID: 117 33012448 DOI: 10.1016/j.cld.2020.07.013]
- Franca AV, Elias Junior J, Lima BL, Martinelli AL, Carrilho FJ. Diagnosis, staging and treatment 118 of hepatocellular carcinoma. Braz J Med Biol Res 2004; 37: 1689-1705 [PMID: 15517086 DOI: 10.1590/s0100-879x2004001100015
- 119 Simmons O, Fetzer DT, Yokoo T, Marrero JA, Yopp A, Kono Y, Parikh ND, Browning T, Singal AG. Predictors of adequate ultrasound quality for hepatocellular carcinoma surveillance in patients with cirrhosis. Aliment Pharmacol Ther 2017; 45: 169-177 [PMID: 27862091 DOI: 10.1111/apt.13841
- 120 Pocha C, Dieperink E, McMaken KA, Knott A, Thuras P, Ho SB. Surveillance for hepatocellular cancer with ultrasonography vs. computed tomography -- a randomised study. Aliment Pharmacol Ther 2013; 38: 303-312 [PMID: 23750991 DOI: 10.1111/apt.12370]
- Kim SY, An J, Lim YS, Han S, Lee JY, Byun JH, Won HJ, Lee SJ, Lee HC, Lee YS. MRI With 121 Liver-Specific Contrast for Surveillance of Patients With Cirrhosis at High Risk of Hepatocellular Carcinoma. JAMA Oncol 2017; 3: 456-463 [PMID: 27657493 DOI: 10.1001/jamaoncol.2016.3147]
- Harding JJ, Khalil DN, Abou-Alfa GK. Biomarkers: What Role Do They Play (If Any) for 122 Diagnosis, Prognosis and Tumor Response Prediction for Hepatocellular Carcinoma? Dig Dis Sci 2019; 64: 918-927 [PMID: 30838478 DOI: 10.1007/s10620-019-05517-6]
- 123 European Association for the Study of the Liver. EASL Clinical Practice Guidelines: Management of hepatocellular carcinoma. J Hepatol 2018; 69: 182-236 [PMID: 29628281 DOI: 10.1016/i.ihep.2018.03.019
- Ebrahim AE, Shehata MAH, Abou-saif S, Hamisa MF, Abd-Elsalam S & Yousef M. Role of 124 Fibroscan for early detection of hepatocellular carcinoma (HCC) in hepatitis C cirrhotic patients. Egypt J Radiol Nucl Med 2020; 51: 134 [DOI: 10.1186/s43055-020-00258-3]
- 125 Ahmed NNA, El Gaafary SM, Elia RZ, Abdulhafiz EM. Role of abbreviated MRI protocol for screening of HCC in HCV related cirrhotic patients prior to direct-acting antiviral treatment. Egypt J Radiol Nucl Med 2020; 51: 102 [DOI: 10.1186/s43055-020-00199-x]
- Abdelaziz AO, Nabil MM, Omran DA, Abdelmaksoud AH, Asem N, Shousha HI, Elbaz TM, 126 Leithy R. Hepatocellular Carcinoma Multidisciplinary Clinic-Cairo University (HMC-CU) score: A new simple score for diagnosis of HCC. Arab J Gastroenterol 2020; 21: 102-105 [PMID: 32439235 DOI: 10.1016/j.ajg.2020.04.001]
- 127 El-Zayadi AR, Badran HM, Shawky S, Emara S, El-Bareedy A, Sobhi M. Effect of surveillance for hepatocellular carcinoma on tumor staging and treatment decisions in Egyptian patients. Hepatol Int 2010; 4: 500-506 [PMID: 20827407 DOI: 10.1007/s12072-010-9170-x]
- 128 Salama MM, Allam AS, Nasser HM, Kabiel YWA, Elsayed EH. Role of Serum Leptin in the Diagnosis and Prognosis of Hepatocellular Carcinoma in Egyptian Cirrhotic Patients. Med J Cairo Univ 2020; 88: 259-266
- 129 Bryant R, Laurent A, Tayar C, van Nhieu JT, Luciani A, Cherqui D. Liver resection for



hepatocellular carcinoma. Surg Oncol Clin N Am 2008; 17: 607-633, ix [PMID: 18486886 DOI: 10.1016/j.soc.2008.02.002]

- Hu RH, Lee PH, Chang YC, Ho MC, Yu SC. Treatment of centrally located hepatocellular 130 carcinoma with central hepatectomy. Surgery 2003; 133: 251-256 [PMID: 12660635 DOI: 10.1067/msv.2003.102
- 131 Shimozawa N, Hanazaki K. Longterm prognosis after hepatic resection for small hepatocellular carcinoma. J Am Coll Surg 2004; 198: 356-365 [PMID: 14992736 DOI: 10.1016/j.jamcollsurg.2003.10.017]
- 132 Zakaria HM, Macshut M, Gaballa NK, Sherif AE, Abdel-Samea ME, Abdel-Samiee M, Marwan I, Yassein T. Total tumor volume as a prognostic value for survival following liver resection in patients with hepatocellular carcinoma. Retrospective cohort study. Ann Med Surg (Lond) 2020; 54: 47-53 [PMID: 32368340 DOI: 10.1016/j.amsu.2020.04.001]
- 133 Kauffmann R, Fong Y. Post-hepatectomy liver failure. Hepatobiliary Surg Nutr 2014; 3: 238-246 [PMID: 25392835 DOI: 10.3978/j.issn.2304-3881.2014.09.01]
- 134 Senbel A, Elmahdy Y, Roshdy S, Khater A, Shehatoo F, Farouk O, Fathi A, Hamed E, Kotb S, Denwer A. Role of Hepatic Resection for HCC in the era of Transplantation; an Experience of Two Tertiary Egyptian Centers. Indian J Surg Oncol 2017; 8: 514-518 [PMID: 29203983 DOI: 10.1007/s13193-017-0679-5
- 135 Makhlouf NA, Abdel-Malek MO, Hassany SM, Abd-Elmawgood AM, Taha AM, Ibraheem TM, Fadel BA. Risk of liver failure after major hepatectomy for patients with hepatocellular carcinoma. Egypt J Surg 2020; 39: 81-85
- Shehta A, Farouk A, Fouad A, Aboelenin A, Elghawalby AN, Said R, Elshobary M, El Nakeeb A. 136 Post-hepatectomy liver failure after hepatic resection for hepatocellular carcinoma: a single center experience. Langenbecks Arch Surg 2021; 406: 87-98 [PMID: 32778915 DOI: 10.1007/s00423-020-01956-2
- 137 Amer KE, Marwan I. Living donor liver transplantation in Egypt. Hepatobiliary Surg Nutr 2016; 5: 98-106 [PMID: 27115003 DOI: 10.3978/j.issn.2304-3881.2015.10.03]
- 138 Kamel R, Hatata Y, Hosny K, Nabil A, El-Deen Abd-Allah A, Mostafa A, Abdel-Aal A, Elganzoury MZ, Elmalt O, Marwan I, Hosny A. Outcome of Living-Donor Liver Transplant for Hepatocellular Carcinoma: 15-Year Single-Center Experience in Egypt. Exp Clin Transplant 2017; 15: 12-20 [PMID: 28301993 DOI: 10.6002/ect]
- Galal M, Bahaa M, Ebrahim WA, El-Shafei AE, Sedrak CR. Pretransplantation alpha fetoprotein 139 level as a predictor of hepatocellular carcinoma recurrence after adult living donor liver transplantation within milan criteria in Egyptian patients. Egypt J Intern Med 2019; 31: 203-207 [DOI: 10.4103/ejim.ejim 106 18]
- 140 Ahmed M, Solbiati L, Brace CL, Breen DJ, Callstrom MR, Charboneau JW, Chen MH, Choi BI, de Baère T, Dodd GD 3rd, Dupuy DE, Gervais DA, Gianfelice D, Gillams AR, Lee FT Jr, Leen E, Lencioni R, Littrup PJ, Livraghi T, Lu DS, McGahan JP, Meloni MF, Nikolic B, Pereira PL, Liang P, Rhim H, Rose SC, Salem R, Sofocleous CT, Solomon SB, Soulen MC, Tanaka M, Vogl TJ, Wood BJ, Goldberg SN; International Working Group on Image-guided Tumor Ablation; Interventional Oncology Sans Frontières Expert Panel; Technology Assessment Committee of the Society of Interventional Radiology; Standard of Practice Committee of the Cardiovascular and Interventional Radiological Society of Europe. Image-guided tumor ablation: standardization of terminology and reporting criteria--a 10-year update. Radiology 2014; 273: 241-260 [PMID: 24927329 DOI: 10.1148/radiol.14132958]
- 141 Brace CL. Microwave tissue ablation: biophysics, technology, and applications. Crit Rev Biomed Eng 2010; 38: 65-78 [PMID: 21175404 DOI: 10.1615/critrevbiomedeng.v38.i1.60]
- 142 Soliman AF, Abouelkhair MM, Hasab Allah MS, El-Kady NM, Ezzat WM, Gabr HA, Elsayed EH, Saleh AI, Kamel A. Efficacy and Safety of Microwave Ablation (MWA) for Hepatocellular Carcinoma (HCC) in Difficult Anatomical Sites in Egyptian Patients with Liver Cirrhosis. Asian Pac J Cancer Prev 2019; 20: 295-301 [PMID: 30678453 DOI: 10.31557/APJCP.2019.20.1.295]
- 143 Nouh MA, El Sharkawy MK, El Deeb GS, Badawy AM, Azab HM. Comparative study of radiofrequency ablation combined with either percutaneous ethanol injection or percutaneous acetic acid injection in the management of hepatocellular carcinoma. Menoufia Med J 2020; 33: 819-823
- Kamal A, Elmoety AAA, Rostom YAM, Shater MS, Lashen SA. Percutaneous radiofrequency vs 144 microwave ablation for management of hepatocellular carcinoma: a randomized controlled trial. J Gastrointest Oncol 2019; 10: 562-571 [PMID: 31183208 DOI: 10.21037/jgo.2019.01.34]
- 145 Sharaf-Eldin MA, El-Yamany SA, Salah RA, Kohla M, Habba E, Fattah HA, Ghazy MS. Risk factors for recurrence of hepatocellular carcinoma after radiofrequency ablation in a cohort of Egyptian patients with hepatitis C virus-induced cirrhosis: a multicenter analysis. Egypt Liver J 2014; 4: 13-19 [DOI: 10.1097/01.ELX.0000440962.37421.c8]
- Salem R, Gilbertsen M, Butt Z, Memon K, Vouche M, Hickey R, Baker T, Abecassis MM, Atassi 146 R, Riaz A, Cella D, Burns JL, Ganger D, Benson AB 3rd, Mulcahy MF, Kulik L, Lewandowski R. Increased quality of life among hepatocellular carcinoma patients treated with radioembolization, compared with chemoembolization. Clin Gastroenterol Hepatol 2013; 11: 1358-1365.e1 [PMID: 23644386 DOI: 10.1016/j.cgh.2013.04.028]
- 147 Seldinger technique. Reprint from Acta Radiologica 1953. AJR Am J Roentgenol 1984; 142: 5-7 [PMID: 6362375 DOI: 10.2214/ajr.142.1.5]
- 148 Miraglia R, Pietrosi G, Maruzzelli L, Petridis I, Caruso S, Marrone G, Mamone G, Vizzini G, Luca



A, Gridelli B. Efficacy of transcatheter embolization/chemoembolization (TAE/TACE) for the treatment of single hepatocellular carcinoma. World J Gastroenterol 2007; 13: 2952-2955 [PMID: 17589945 DOI: 10.3748/wjg.v13.i21.2952]

- 149 Kang IK, Kim SW, Hahn SH, Cho SC, Gham CW, Lee DH. [A comparison of patients with hepatocellular carcinoma between a short-term (less than 6 mo) survival group and a long-term (over 24 mo) survival group after treatment with transcatheter arterial chemoembolization]. Taehan Kan Hakhoe Chi 2002; 8: 189-200 [PMID: 12499805]
- 150 Huang YH, Wu JC, Chau GY, Lui WY, King KL, Chiang JH, Yen SH, Sheng WY, Hou MC, Lu CL, Chang FY, Lee SD. Supportive treatment, resection and transcatheter arterial chemoembolization in resectable hepatocellular carcinoma: an analysis of survival in 419 patients. Eur J Gastroenterol Hepatol 1999; 11: 315-321 [PMID: 10333206 DOI: 10.1097/00042737-199903000-00017
- 151 Eltawil KM, Berry R, Abdolell M, Molinari M. Analysis of survival predictors in a prospective cohort of patients undergoing transarterial chemoembolization for hepatocellular carcinoma in a single Canadian centre. HPB (Oxford) 2012; 14: 162-170 [PMID: 22321034 DOI: 10.1111/j.1477-2574.2011.00420.x
- Salhab M, Canelo R. An overview of evidence-based management of hepatocellular carcinoma: a 152 meta-analysis. J Cancer Res Ther 2011; 7: 463-475 [PMID: 22269411 DOI: 10.4103/0973-1482.92023
- 153 Chapman WC, Majella Doyle MB, Stuart JE, Vachharajani N, Crippin JS, Anderson CD, Lowell JA, Shenoy S, Darcy MD, Brown DB. Outcomes of neoadjuvant transarterial chemoembolization to downstage hepatocellular carcinoma before liver transplantation. Ann Surg 2008; 248: 617-625 [PMID: 18936575 DOI: 10.1097/SLA.0b013e31818a07d4]
- 154 Farouk Ahmed AL, Nasser HM, Abo-Elmaaty ME, Montasser IF. Role of trans arterial chemoembolization (TACE) in down staging of hepatocellular carcinoma (HCC) before liver transplantation. Egypt J Hosp Med 2018; 72: 5578-5583
- 155 Fouad YM, Aboalelal AS, Mokarrab H, abdelghany W, Abdelhamid W, Essawy MG. Prospective Evaluation of Health-Related Quality of Life in Patients with Hepatocellular Carcinoma after Radiofrequency or TACE. Indian J Public Health Res Dev 2020; 11: 882-887
- 156 Hassan H, Eman MB, El-Folly RF, Amr MA, El-Hariri H, Abdelghany RS, El-Fouly NF. Assessment of health-related quality of life after hepatocellular carcinoma management (radiofrequency ablation or transarterial chemoembolization): a pilot Egyptian study. Egypt Liver J 2017; 7: 51-57 [DOI: 10.1097/01.ELX.0000528001.78130.1c]
- 157 Dufour JF, Bargellini I, De Maria N, De Simone P, Goulis I, Marinho RT. Intermediate hepatocellular carcinoma: current treatments and future perspectives. Ann Oncol 2013; 24 Suppl 2: ii24-ii29 [PMID: 23715940 DOI: 10.1093/annonc/mdt054]
- 158 Kim YH, Kim DY. Yttrium-90 radioembolization for hepatocellular carcinoma: what we know and what we need to know. Oncology 2013; 84 Suppl 1: 34-39 [PMID: 23428856 DOI: 10.1159/000345887
- 159 Zhang T, Ding X, Wei D, Cheng P, Su X, Liu H, Wang D, Gao H. Sorafenib improves the survival of patients with advanced hepatocellular carcinoma: a meta-analysis of randomized trials. Anticancer Drugs 2010; 21: 326-332 [PMID: 20016366 DOI: 10.1097/CAD.0b013e3283350e26]
- 160 Moreno-Luna LE, Yang JD, Sanchez W, Paz-Fumagalli R, Harnois DM, Mettler TA, Gansen DN, de Groen PC, Lazaridis KN, Narayanan Menon KV, Larusso NF, Alberts SR, Gores GJ, Fleming CJ, Slettedahl SW, Harmsen WS, Therneau TM, Wiseman GA, Andrews JC, Roberts LR. Efficacy and safety of transarterial radioembolization vs chemoembolization in patients with hepatocellular carcinoma. Cardiovasc Intervent Radiol 2013; 36: 714-723 [PMID: 23093355 DOI: 10.1007/s00270-012-0481-2]
- 161 Padia SA, Johnson GE, Horton KJ, Ingraham CR, Kogut MJ, Kwan S, Vaidya S, Monsky WL, Park JO, Bhattacharya R, Hippe DS, Harris WP. Segmental Yttrium-90 Radioembolization vs Segmental Chemoembolization for Localized Hepatocellular Carcinoma: Results of a Single-Center, Retrospective, Propensity Score-Matched Study. J Vasc Interv Radiol 2017; 28: 777-785.e1 [PMID: 28365172 DOI: 10.1016/j.jvir.2017.02.018]
- 162 Hamed MM, Abdelhay AA, Abd Alfattah MH, Gameel GA. Efficacy of Transarterial Y90 Radioembolization in Management for Unresectable-Intermediate and Locally Advanced-HCC. Med *J Cairo Univ* 2019; **87**: 3147-3156 [DOI: 10.21608/mjcu.2019.59518]
- 163 Hetta MO, Hetta MW, Shebrya NH, El Ghazaly HA. Radioembolization with Yttrium-90 resin microspheres in treatment of HCC with or without PVT: Initial Egyptian experience. Egypt J Radiol Nuc Med 2013; 44: 215-222
- 164 El Fouly A, Ertle J, El Dorry A, Shaker MK, Dechêne A, Abdella H, Mueller S, Barakat E, Lauenstein T, Bockisch A, Gerken G, Schlaak JF. In intermediate stage hepatocellular carcinoma: radioembolization with yttrium 90 or chemoembolization? Liver Int 2015; 35: 627-635 [PMID: 25040497 DOI: 10.1111/liv.12637]
- 165 Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, de Oliveira AC, Santoro A, Raoul JL, Forner A, Schwartz M, Porta C, Zeuzem S, Bolondi L, Greten TF, Galle PR, Seitz JF, Borbath I, Häussinger D, Giannaris T, Shan M, Moscovici M, Voliotis D, Bruix J; SHARP Investigators Study Group. Sorafenib in advanced hepatocellular carcinoma. N Engl J Med 2008; 359: 378-390 [PMID: 18650514 DOI: 10.1056/NEJMoa07088571
- 166 Wilhelm SM, Carter C, Tang L, Wilkie D, McNabola A, Rong H, Chen C, Zhang X, Vincent P,



McHugh M, Cao Y, Shujath J, Gawlak S, Eveleigh D, Rowley B, Liu L, Adnane L, Lynch M, Auclair D, Taylor I, Gedrich R, Voznesensky A, Riedl B, Post LE, Bollag G, Trail PA. BAY 43-9006 exhibits broad spectrum oral antitumor activity and targets the RAF/MEK/ERK pathway and receptor tyrosine kinases involved in tumor progression and angiogenesis. Cancer Res 2004; 64: 7099-7109 [PMID: 15466206 DOI: 10.1158/0008-5472.CAN-04-1443]

- 167 Keating GM. Sorafenib: A Review in Hepatocellular Carcinoma. Target Oncol 2017; 12: 243-253 [PMID: 28299600 DOI: 10.1007/s11523-017-0484-7]
- Javan H, Dayyani F, Abi-Jaoudeh N. Therapy in Advanced Hepatocellular Carcinoma. Semin 168 Intervent Radiol 2020; 37: 466-474 [PMID: 33328702 DOI: 10.1055/s-0040-1719187]
- Bruix J, Qin S, Merle P, Granito A, Huang YH, Bodoky G, Pracht M, Yokosuka O, Rosmorduc O, 169 Breder V, Gerolami R, Masi G, Ross PJ, Song T, Bronowicki JP, Ollivier-Hourmand I, Kudo M, Cheng AL, Llovet JM, Finn RS, LeBerre MA, Baumhauer A, Meinhardt G, Han G; RESORCE Investigators. Regorafenib for patients with hepatocellular carcinoma who progressed on sorafenib treatment (RESORCE): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet 2017; 389: 56-66 [PMID: 27932229 DOI: 10.1016/S0140-6736(16)32453-9]
- 170 Nada Y, Rashad N, Eissa M, Ghonaim A, Farag K, Saadawi I, Sheha A, El Gewaity M, Abdel-Rahman O. Outcomes of treatment with sorafenib in Egyptian patients with hepatocellular carcinoma: a retrospective cohort study. Expert Rev Gastroenterol Hepatol 2018; 12: 99-107 [PMID: 29124987 DOI: 10.1080/17474124.2018.1403898]
- Abdel-Rahman O, Abdelwahab M, Shaker M, Abdelwahab S, Elbassiony M, Ellithy M. Sorafenib 171 for Egyptian patients with advanced hepatocellular carcinoma; single center experience. J Egypt Nat Cancer Insti 2014; 26: 9-13 [DOI: 10.1016/j.jnci.2013.08.003]
- 172 El Baghdady NS, El Wakeel L, Ellithy MA, Eltohamy N, Shaheen SM, El Naggar AER. Assessment of efficacy and safety of sorafenib vs no treatment in Egyptian hepatocellular carcinoma patients. Ann Oncol 2019; 30: ix42-ix67 [DOI: 10.21608/APS.2020.45180.1043]
- Hamdy Elsisi G, Nada Y, Rashad N, Carapinha J. Cost-effectiveness of sorafenib vs best supportive 173 care in advanced hepatocellular carcinoma in Egypt. J Med Econ 2019; 22: 163-168 [PMID: 30479174 DOI: 10.1080/13696998.2018.1552432]
- 174 Hanafy AS. Efficacy of low dose capecitabine and sorafenib in patients with advanced alfafetoprotein secreting hepatocellular carcinoma: a 1 year experience. Springerplus 2016; 5: 1675 [PMID: 27733977 DOI: 10.1186/s40064-016-3376-x]
- 175 Ibrahim AS, Khaled HM, Mikhail NN, Baraka H, Kamel H. Cancer incidence in egypt: results of the national population-based cancer registry program. J Cancer Epidemiol 2014; 2014: 437971 [PMID: 25328522 DOI: 10.1155/2014/437971]
- 176 Omran D, Alboraie M, Zayed RA, Wifi MN, Naguib M, Eltabbakh M, Abdellah M, Sherief AF, Maklad S, Eldemellawy HH, Saad OK, Khamiss DM, El Kassas M. Towards hepatitis C virus elimination: Egyptian experience, achievements and limitations. World J Gastroenterol 2018; 24: 4330-4340 [PMID: 30344418 DOI: 10.3748/wjg.v24.i38.4330]
- El Kassas M. Tawheed A. Eltabbakh M. Kaseb A. Hepatitis C Antiviral Therapy In Patients With 177 Successfully Treated Hepatocellular Carcinoma: Dancing With Wolves. J Hepatocell Carcinoma 2019; 6: 183-191 [PMID: 31819865 DOI: 10.2147/JHC.S206668]
- Wahdan IH. Cost-Effectiveness of National Breast Cancer Screening Programs in Developing 178 Countries, with Reference to the Recent Egyptian Initiative. J High Institute Public Health 2020; 50: 1-9



0 WJ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 1939-1955

DOI: 10.4251/wjgo.v13.i12.1939

ISSN 1948-5204 (online)

REVIEW

Moving forward in the treatment of cholangiocarcinoma

Tommaso M Manzia, Alessandro Parente, Ilaria Lenci, Bruno Sensi, Martina Milana, Carlo Gazia, Alessandro Signorello, Roberta Angelico, Giuseppe Grassi, Giuseppe Tisone, Leonardo Baiocchi

ORCID number: Tommaso M Manzia 0000-0002-4636-3478; Alessandro Parente 0000-0001-5506-224X: Ilaria Lenci 0000-0001-5704-9890; Bruno Sensi 0000-0003-1912-2414; Martina Milana 0000-0003-2027-0481; Carlo Gazia 0000-0002-3543-4170; Alessandro Signorello 0000-0002-3831-7244; Roberta Angelico 0000-0002-3439-7750; Giuseppe Grassi 0000-0001-9182-8759; Giuseppe Tisone 0000-0001-8860-5909; Leonardo Baiocchi 0000-0003-3672-4505.

Author contributions: Manzia TM contributed to acquisition of data, analysis and interpretation, drafting of manuscript and critical revision; Parente A, Lenci I, Sensi B, Milana M, Gazia C, Signorello A, Angelico R, Grassi G and Tisone G contributed to acquisition of data and critical revision; Baiocchi L contribute to proposal of study, study conception, correction of manuscript and critical revision.

Conflict-of-interest statement: The authors declare no conflict of interest

Country/Territory of origin: Italy

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer reviewed

Peer-review report's scientific quality classification

Tommaso M Manzia, Bruno Sensi, Carlo Gazia, Roberta Angelico, Giuseppe Tisone, Hepato-Pancreato-Biliary and Transplant, Department of Surgery, University of Rome Tor Vergata, Rome 00133, Italy

Alessandro Parente, The Liver Unit, Queen Elizabeth Hospital Birmingham, Birmingham B15 2TH, United Kingdom

Ilaria Lenci, Martina Milana, Alessandro Signorello, Giuseppe Grassi, Leonardo Baiocchi, Hepatology Unit, University of Tor Vergata, Rome 00133, Italy

Corresponding author: Leonardo Baiocchi, MD, PhD, Associate Professor, Hepatology Unit, University of Tor Vergata, Viale Oxford 31, Rome 00133, Italy. baiocchi@uniroma2.it

Abstract

Despite being the second most frequent primary liver tumor in humans, early diagnosis and treatment of cholangiocarcinoma (CCA) are still unsatisfactory. In fact, survival after 5 years is expected in less than one fourth of patients diagnosed with this disease. Rare incidence, late appearance of symptoms and heterogeneous biology are all factors contributing to our limited knowledge of this cancer and determining its poor prognosis in the clinical setting. Several efforts have been made in the last decades in order to achieve an improved classification/understanding with regard to the diverse CCA forms. Location within the biliary tree has helped to distinguish between intrahepatic, perihilar and distal CCA types. Sequence analysis contributed to identifying several characteristic genetic aberrations in CCA that may also serve as possible targets for therapy. Novel findings are expected to significantly improve the management of this malignancy in the near future. In this changing scenario our review focuses on the current and future strategies for CCA treatment. Both systemic and surgical treatments are discussed in detail. The results of the main studies in this field are reported, together with the ongoing trials. The current findings suggest that an integrated multidisciplinary approach to this malignancy would be helpful to improve its outcome.

Key Words: Cholangiocarcinoma; Treatment; Genetic aberration; Immunotherapy; Liver resection; Liver transplantation

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.



Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: March 16, 2021 Peer-review started: March 16, 2021 First decision: May 3, 2021 Revised: May 14, 2021 Accepted: October 14, 2021 Article in press: October 14, 2021 Published online: December 15, 2021

P-Reviewer: Titapun A S-Editor: Chang KL L-Editor: Filipodia P-Editor: Chang KL



Core Tip: Cholangiocarcinoma is a lethal malignancy characterized by a poor survival. In this review we discuss in detail the actual treatment and the future therapeutic perspectives for this cancer. Systemic and surgical strategies are reported with the corresponding results. Improved knowledge of this malignancy and a multidisciplinary therapeutic approach are likely to improve the cholangiocarcinoma outcome in the future.

Citation: Manzia TM, Parente A, Lenci I, Sensi B, Milana M, Gazia C, Signorello A, Angelico R, Grassi G, Tisone G, Baiocchi L. Moving forward in the treatment of cholangiocarcinoma. World J Gastrointest Oncol 2021; 13(12): 1939-1955

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/1939.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.1939

INTRODUCTION

Cholangiocarcinoma (CCA) is a primary malignancy of the biliary system and represents the second most common primary hepatic malignancy after hepatocellular carcinoma (HCC), constituting around 15% of primary liver tumors and 3% of gastrointestinal malignancies[1,2]. It is a rare tumor with a global incidence of 0.3-6 per 100000 inhabitants per year, displaying an increasing trend in the last decades[1]. However, in some Asian countries, such as Thailand, Cambodia and Laos, rates can be as high as 85 per 100000 due to infection with liver flukes[2].

Distinction into subgroups of CCA is anatomical: intrahepatic CCA (iCCA) arises in the liver above the second order bile ducts; perihilar CCA (pCCA), also known as Klatskin tumor, arises in the first order or main bile duct above the junction with the cystic duct; and distal CCA (dCCA) originates distally to the cystic duct (Figure 1). This classification is crucial as each subtype has distinct clinical characteristics and therapeutic strategies. pCCA accounts for the majority of diagnoses (50%-60%), with dCCA (20%-30%) and iCCA (10-20%) being less frequent[3]. iCCA can be further classified on the basis of the cells of origin as large and small duct types, with chronic biliary inflammation and chronic hepatis as risk factors, respectively[4]. On top of this, a recent interesting study involved the epigenomic and transcriptomic analysis of CCAs from 10 different countries in order to further understand and classify the genetic basis of CCA. The authors performed the analysis on CCA samples associated with liver flukes (mainly Opisthorchis viverrine and Clonorchis sinensis) and non-fluke cases. Four CCA clusters were likely driven by distinct etiologies, with separate genetic, epigenetic and clinical features found, highlighting how distinct cancer subtypes in the same organ may arise through different carcinogenic pathways^[5].

Unfortunately, symptoms often appear when the disease is already advanced, resulting in a poor prognosis. In fact, this malignancy has an overall survival rate at 5 years of 5%-20%[1,3]. Nonetheless, many promising new approaches are currently under investigation.

Several issues have been encountered in the pursuit of a curative treatment for CCA in humans. Despite the evidence of different biological and epidemiological risk factors and genetic aberrations between diverse types of CCAs, these tumors are still frequently pooled together (also with gallbladder cancer) or misclassified in studies focusing on natural history or treatment[6,7]. On the other hand, histological classification (in particular for iCCA forms) remains suboptimal and also relies on heterogeneous genetic aberrations identified in this cancer^[2]. The difficulties in CCA classification and in the comprehension of its biology therefore affect both clinical and basic research in this field. For instance, despite next generation models now attempting the construction of complex 3D CCA systems in culture (such as organoids or spheroids), an adequate reproduction of this tumor remains difficult in the preclinical experimental setting[8].

From the clinical side, CCA symptoms are generally not specific and share similarities with inflammatory diseases of the biliary tract. Moreover, general biomarkers used in medical practice, such as carbohydrate antigen 19-9 exhibit a sensitivity and specificity lower than 70%, underscoring the importance of the identification of possible novel genomic or proteomic biomarkers^[9]. Also the appropriate surveillance of CCA-predisposing conditions, such as primary sclerosing cholangitis,





Figure 1 The anatomical location of intrahepatic, perihilar and distal cholangiocarcinoma is depicted.

remains undefined, leading to disappointing late-stage tumor identification in the majority of patients[10].

Furthermore, CCA remains an infrequent cancer in the majority of countries, several cases arise in the absence of recognized risk factors, and when some intraductal papillary or tubular forms are excluded[11], there is usually a short-term poor prognosis. Due to all of the above, clinical investigations and trials remain complicated and of partial impact. Framed in this perspective, this paper summarizes and critically reviews existing therapeutic strategies (both drug-based and surgical) for CCA and provides an overview of future perspectives in the treatment of this malignancy.

CCA PHARMACOLOGICAL TREATMENT: THE PRESENT

As described in detail in the dedicated paragraphs, the opportunity for a complete CCA cure should be offered in rare cases just employing surgical techniques. On the other hand, despite the fact that current drug therapy for this cancer is unsatisfactory, the pharmacological approach may present a larger margin of improvement in the future in comparison with operative methods.

Palliative treatments

At present, in subjects with unresectable, advanced disease, the best option is represented by cisplatin/gemcitabine first-line treatment. Confirmation of the utility of this treatment was obtained by a large study comparing this association with gemcitabine alone^[12]. Two-hundred and four patients affected by biliary tumor (nearly one third with gallbladder cancer) treated with the gemcitabine/cisplatin regimen had an increased tumor response (81.4% vs 71.8%, P = 0.049) and median survival (11.7 vs 8.1 mo, P < 0.001) in comparison with a similar group treated with gemcitabine alone. In the absence of an adequate second-line treatment, a recent Phase 3, open-label, randomized trial (ABC 06) was published on patients with CCA progression under gemcitabine/cisplatin comparing folinic acid/fluorouracil/oxaliplatin therapy to active symptom control[13]. Overall survival was longer in the folinic acid/fluorouracil/oxaliplatin group (6.2 vs 5.3 mo, P = 0.03), thus demonstrating the possible feasibility of second level therapy and possibly changing our clinical approach to these patients in the near future.

Adjuvant treatments

With regard to adjuvant therapy in subjects amenable to surgical resection, the major indication came from the BILCAP trial[14]. In this study, patients undergoing surgical treatment of biliary cancer (n = 447) were allocated to receive capecitabine or just observation after a macroscopically complete tumor resection. Capecitabine increased survival by almost one third. This difference was statistically significant in the perprotocol (53 mo vs 36 mo, P = 0.02) but not in the intention-to-treat analysis. Serious adverse events occurred in the two groups at a similar rate. A randomized Phase 3 clinical trial conducted with adjuvant gemcitabine chemotherapy did not show



significant improvement in overall survival or relapse-free survival in comparison with untreated control^[15]. An attempt was also conducted with adjuvant gemcitabine/oxaliplatin in the PRODIGE 12 study[16], and again no improvements were observed in comparison with supportive care.

In conclusion, excluding the modest, above-described, therapeutic options, physicians and patients are lacking any further pharmacological strategy. Also, radiation therapy gave inconclusive results in this setting[17], meaning that current national guidelines are not able to give an unequivocal indication on this approach[18]. In conclusion, the scarce results of systemic therapy have prompted extensive research in recent decades in order to find a more satisfactory pharmacological approach for this cancer. Current preclinical models and results together with ongoing trials are reported in the following paragraph.

CCA PHARMACOLOGICAL TREATMENT: THE FUTURE

The possible evolution of systemic therapy for CCA is largely dependent on the resolution of some issues with regard to this cancer[19]. First, scientists are still searching for an appropriate preclinical model of CCA[20]. CCA cell culture and tumor xenotransplantation in nude mice are the most commonly used strategies, but they do not adequately reproduce the neoplastic microenvironment^[21]. From the clinical experimental side, the rarity of this neoplasm and competition between new molecules do not facilitate the performance of trials with an adequate number and homogeneous type of CCAs. While exploring this undefined horizon, research efforts are oriented in some main fronts, as reported in the following subparagraphs.

Trying to overcome chemoresistance

One of the main issues greatly limiting chemotherapy effectiveness in CCA is represented by chemoresistance[22]. Chemoresistance describes the capacity of cancer cells to escape or attenuate therapeutic drug effects[23]. Several mechanisms have been identified as the basis of chemoresistance, some opposing drug uptake or increasing its extracellular export and others reducing cellular necrosis/apoptosis or stimulating tumoral phenotypic changes. For instance, the reduced expression of organic cation transporter 1, as observed in both CCA and HCC, has been related to a poor response to tyrosine kinase inhibitors such as sorafenib[24]. On the other hand, the phenotypic CCA evolution from an epithelial to a mesenchymal trait (so-called epithelialmesenchymal-transition) not only counteracts chemotherapy effects but also seems to favor metastatic progression[25]. Several strategies have been attempted in preclinical experimental studies to improve therapeutic response to chemotherapy, such as drug transporter induction or export pump inhibition in CCA cells or targeting cells with specific organic molecules such as bile acids or vesicles. With regard to human trials, a gemcitabine analogue (NUC-1031)[26] not requiring nucleoside cellular transport or intracellular kinase activation is currently being tested in a Phase 3 trial (NCT 04163900).

Targeting genetic aberrations

Several genetic aberrations have been identified in CCA, with a different distribution among intrahepatic, perihilar or distal CCA[27]. Kirsten rat sarcoma gene mutations are frequently encountered, ranging from 9%-40% of cases according to CCA location within the biliary tract^[28]. A specific molecule (AMG 510) targeting the Kirsten rat sarcoma/G12C mutation is currently being tested in a Phase1/2 trial (NCT03600883); however, downstream pathway suppression, obtained by kinase inhibition (such as those of the Raf or MEK family) also may be attempted. In this perspective, the dual suppression of BRAF and MEK, obtained with dabrafenib and trametinib, gave excellent results in anecdotal cases^[29], thus stimulating the Phase 2 ROAR study in patients with the BRAFV600E solid tumor mutation (NCT02034110). In an interim analysis of this trial of 43 patients with biliary tract cancer, the overall response rate (after external data review) accounted for 20% of cases[30].

The fibroblast growth factor (FGF) family comprises a group of proteins that react with their specific receptors (FGFRs) to stimulate several developmental and proliferative processes, also involving stem cell differentiation[31]. FGFR (subtype 2) genetic alterations, characterized by fusion with other genes, have been observed in nearly 15% of iCCAs, so FGF/FGFR signaling has emerged as a possible target to cure this cancer[32]. Among the FGFR inhibitors, infigratinib and pemigatinib have been evaluated in Phase 2 trials (NCT02150967 and NCT02924376, respectively) on advanced



iCCA harboring FGFR aberrations[33,34]. Progression-free survival was slightly better with pemigatinib, accounting for 62% at 6 mo for patients with an FGFR2 mutation. On the basis of these results, this drug was approved by the FDA in April 2020 for the treatment of advanced iCCA harboring this genetic aberration. Other molecules, such as derazantinib, futibatinib and Debio 1347, have been registered for evaluation in clinical trials (NCT03230318, NCT04093362 and NCT03834220), but the results are not vet available.

Since isocitrate dehydrogenase 1 mutations have been identified in approximately 13% of iCCA and 0.8% of other CCAs and the impairment of this enzyme may lead to the accumulation of the pro-oncogenic metabolite D-2-hydroxyglutarate, isocitrate dehydrogenase 1 inhibitors have been suggested for treatment of this cancer. The ClarIDHy phase 3 trial tested the isocitrate dehydrogenase 1 inhibitor ivosidenib in CCAs with a mutation of this enzyme and refractory to previous systemic therapy [35]. Six-month progression-free survival was 32% in the ivosidenib group in comparison with 0% in the placebo group. Other inhibitors are currently being examined in different trials, as summarized in a recent review on this issue[36]. Other genetic aberrations, such as those involving the ERRB family and proto-oncogene tyrosineprotein kinase 1, represent possible targets for CCA therapy; some drugs are under evaluation[37].

Immune checkpoint targeting

The activation of immune checkpoint (IC) pathways seems to be involved, under normal conditions, in tolerance and the prevention of autoimmune diseases[38]; however, tumor-mediated stimulation, hindering immune surveillance, may favor cancer proliferation and spread^[39]. In this perspective, IC inhibitors have recently gained major importance with regard to cancer therapy, achieving a complete response in 20% of melanoma patients[40]. Among diverse IC pathways, the cytotoxic T-lymphocyte antigen 4 and programmed cell death protein 1/programmed cell death protein ligand 1 are those that are mainly recognized and targeted in oncology. In another study, 22 patients harboring CCA characterized by microsatellite instability and mismatch repair reduced protein (findings related to IC upregulation) were treated with the programmed cell death protein ligand 1 inhibitor pembrolizumab, obtaining a median progression free survival of 4.2 mo and a median overall survival of 24.3 mo[41]. However, these results might be improved with careful patient selection since an increased response has been observed as a function of programmed cell death protein ligand 1 expression[42]. Several trials with IC inhibitors alone or in combination and including CCA patients are ongoing.

Newly identified pathways as possible targets for therapy

The neuroendocrine regulation of CCA expansion (as shown by preclinical experimental studies) might be an important factor to consider while searching for a therapy for this cancer[43]. Secretin, somatostatin and melatonin have all been demonstrated to decrease CCA growth, as observed in cancer cell lines or in animal models such as tumor xenotransplantation in nude mice[44-46]. At present, however, no clinical data are available with either secretin or melatonin for CCA treatment, while a trial with somatostatin gave negative results[47]. Also, angiogenic factors such as vascular endothelial growth factor are considered possible targets for CCA therapy. Vascular endothelial growth factor in fact seems to be increased in half of human biliary tract cancers^[48]. A trial using the anti- vascular endothelial growth factor antibody bevacizumab, in association with standard chemotherapy (gemcitabine, oxaliplatin), however, gave modest results[49].

SURGICAL TREATMENT: THE PRESENT

Surgery remains the best treatment option for long-term patient survival in CCA, and it is recommended to undertake surgical treatments in highly specialized centers to minimize morbidity and mortality[50].

Preoperative considerations

Preoperative workup and biliary drainage have been widely discussed in recent decades. The current consensus is that preoperative biliary drainage is required in cases of concomitant cholangitis, need for neoadjuvant therapy, malnutrition, hepatic or renal failure and need for portal vein embolization (PVE)[1]. When jaundice is the only indication, need for decompression is still a matter of debate. Asian guidelines



recommend preoperative drainage because of the higher risk of patients with cholangitis[51,52]. Furthermore, drainage may help restore liver function, decreasing the chance of postoperative liver failure^[52]. On the other hand some studies have shown that, while biliary drainage is beneficial by reducing morbidity and mortality in patients with small future liver remnant (FLR), it is equally detrimental when FLR is large enough[53,54]. In Western countries, many centers prefer to use selective biliary drainage when FLR is less than 30%-40% [55]. When stenting is required, both endoscopic and percutaneous methods are used. Percutaneous transhepatic biliary drainage has some advantages, such as reducing the need for re-intervention, reducing the time to achieving a therapeutic effect and fewer procedural risks. However, a recent randomized trial of percutaneous vs endoscopic stenting was terminated early due to excess mortality in the percutaneous group (41% vs 11%), mandating further prospective studies and a reconsideration of drainage strategies^[55]. Alternatively, nasobiliary drainage may be a valid option, showing good success rates and low morbidity despite greater patient discomfort[56,57]. The optimal timing of surgery in drained patients is currently unknown. A recent study identified a preoperative bilirubin level of $< 75 \,\mu$ mol/L (2.9 mg/dL) to be correlated with fewer complications, less mortality and longer 5-year overall survival[58].

Surgical considerations for iCCA

Patients are considered eligible for surgery whenever complete resection of the tumor with negative margins (R0) can be achieved, providing sufficient FLR. Bilateral multifocal or multicentric disease is associated in many studies to a significantly shorter overall survival (OS)[59,60]. In practice, only 32% of iCCAs satisfy resectability criteria at presentation. On top of this, around 30% of iCCAs will be deemed inoperable on the operating table. Staging laparoscopy can detect unresectable disease in around 36% of patients with minimal costs[61] and is advocated by current guidelines[62].

Principles: The established principles of surgery for iCCA are to achieve R0 resections and to provide adequate staging with hilar lymphadenectomy, sparing at the same time as much parenchyma as possible to avoid post-hepatectomy liver failure. Margin status is the primary objective in iCCA surgery. Evidence mainly derives from large single center and multicenter studies, which have demonstrated a significant survival impact of R0 resection. Overall survival at 5 years for R0, R1 and R2 resections are reported to be 28.7%, 13.9% and 0%, respectively[63], with an increased survival benefit for > 5 mm margins[64].

Lymphadenectomy and nodal disease: Nodal disease is recognized as the most important prognostic factor in most studies[59,63-66]. In fact, some authors have reported that margin status may have limited impact in the presence of nodal metastases[64]. Most guidelines suggest routine consideration of regional lymphadenectomy and a minimum of six lymph nodes are needed for accurate staging[2,62,67]. Nonetheless, the role of lymphadenectomy remains controversial in Western countries, where the practice is not widespread, and almost 50% of patients have no lymph nodes examined[68]. Regional lymph nodes include cystic, bile duct, hepatic artery and portal vein. Right and left hemi-livers have distinct lymphatic drainage: for right liver iCCA, the retropancreatic nodes along the common bile duct are considered regional nodes and should be removed, while for left liver iCCAs, the same considerations are true for the lesser curvature and inferior phrenic nodes. Lymph nodes may be positive in as many as 30% of cases, but with current adjuvant therapy, survival is acceptable, and this should not refrain the surgeon from resection[68]. On the contrary, distant nodes such as celiac, superior mesenteric, paraaortic or caval should be considered as distant metastatic disease and contraindicate extensive surgery as patients are unlikely to gain any benefit^[2,62,67].

Extended procedures: Given the poor prognosis (0% 5-year OS) of unresectable disease or R2 resection[63,65], in recent years, some groups have explored the benefits of major vascular resections to obtain R0 resection, resulting in up to 84% of patients [66] with morbidity and mortality rates comparable to standard resection[69]. Overall survival of these patients is also comparable to patients who did not undergo vascular resection[66,69,70]. In general, all patients with localized iCCA should be considered for resection even if this implies major hepatectomy or vascular resection[62]. In recent decades, based on the principles of liver regeneration, some authors have pushed the boundaries for resectability in liver surgery by introducing the concept of two-stage hepatectomies, namely portal vein ligation and PVE. The latter can enhance the resectability rates of liver tumors, allowing extensive resection with adequate FLR and are



usually well-tolerated by the patient. However, a major drawback is the long waiting time for the second stage procedure, which can take up to several weeks, carrying the risk of tumor progression. To solve these problems, a German group of authors developed a new technique, known as associating liver partition and portal vein ligation for staged hepatectomy (ALPPS), which was found to allow rapid growth of the FLR, with a median period of 9 d[71]. Another study investigated benchmark outcomes in ALPPS, demonstrating that it has a comparable standard outcome as other types of major liver surgery[72]. ALPPS for iCCA has been evaluated in an international multicenter study in which 102 patients underwent first-stage ALPPS; 99 completed the second procedure, and R0 resection was obtained in 85% of cases with 29% major morbidity and 7% mortality[60]. When disease is considered unresectable, neoadjuvant chemotherapy can convert as many as 53% of cases to secondary resectable disease[73].

Recurrent disease: Recurrence of iCCA is frequent. Most recurrences are intrahepatic and therefore potentially amenable to re-resection[74], with satisfactory outcomes when repeated resections are undertaken. These results lead to the recommendation that the same principles for resectability should be applied in consideration of primary and secondary resection[75].

Surgical considerations for pCCA

pCCA represents a surgical challenge due to its intrinsic anatomical location. Nonetheless, its higher prevalence (50% of CCAs) has translated into more extensive literature and pioneering advances in surgical treatment.

In pCCA, the main criteria that define surgical unresectability are inadequate FLR, absence of a suitable field for biliary reconstruction (i.e. bilateral segmental ductal extension) and major vascular infiltration[51]. Growth of FLR may be induced with two-stage hepatectomy techniques, broadening indications for resection. Nonetheless, 20%-50% of patients are deemed to be unresectable upon surgical exploration, making explorative laparoscopy a useful tool to avoid unnecessary laparotomies.

Principles: Surgery for pCCA routinely involves en bloc hemi-hepatectomy and bile duct resection to achieve negative biliary and parenchymal margins, with additional resection of the caudate lobe, regional lymphadenectomy [51,76] and biliary reconstruction. Negative margins are paramount. The caudate lobe usually drains directly into the biliary confluence, hence the necessity of its resection for curative intent is advised by current guidelines as it improves OS[51,77]. A number of studies have demonstrated that intraoperative additional resection to achieve R0 confers a significant survival advantage with few complications and a prognosis comparable to primary R0[78,79]. In particular, aggressive approaches such as pancreaticoduodenectomy seem to offer improved results[78]. Lately, a new concept of isolated circumferential margin has been introduced for pCCA. Stremitzer et al[80] identified a group of patients who, despite being classified as R1, did not have distal or proximal margin positivity but only focal circumferential positivity. These patients had better survival than those with surgical resection margin positivity, although inferior when compared to their R0 counterparts. Finally, a recent study has challenged these surgical dogmas, arguing that with current adjuvant therapies R1 patients may have similar survival to R0[77].

Lymphadenectomy and nodal disease: European guidelines affirm that lymphadenectomy should be considered the standard of care, but there is no consensus on the extent of lymphadenectomy for pCCA[81]. A recent systematic review identified a minimum of seven lymph nodes to convey sufficient information avoiding understaging, with no benefit coming from higher lymph node counts (\geq 15) which could only be achieved with extended lymphadenectomy [82]. The regional nodes for pCCA are cystic, biliary, hepatic artery, portal vein and retropancreatic. The impact of extended lymphadenectomy of N2 nodes (dissection of celiac, superior mesenteric and paraaortic nodes) on survival has not been established, but trials are ongoing[83,84]. For known N2 positive disease, current expert consensus suggests no benefit of resection[2].

Extended procedures: Extended resections have been explored for pCCA, including two-stage hepatectomies such as portal vein ligation/PVE and ALPPS with acceptable outcomes. The first reports of 29 ALPPS procedures for this indication featured a strikingly high mortality rate, although statistically comparable to results of 29 matched patients who underwent non-ALPPS resection[85]. These poor initial results



have dramatically improved for most ALPPS indications with better patient selection and inter-stage management and will hopefully improve for hCCA as well[86]. As of 2020, ALPPS should only be considered in highly experienced institutions. Hepatopancreaticoduodenectomy entails resection of the entire extrahepatic biliary tree, thus necessitating resection of the pancreatic head and duodenum. It is used for tumors with concomitant distal bile duct spread. This procedure is associated with high major morbidity rates of up to 37%. Nonetheless, the latest reports from highly specialized centers have been encouraging and suggest that hepatopancreaticoduodenectomy could be considered in young, fit patients when it represents the only chance of a cure [87]. Vascular resection can be adopted to increase R0 rates. Long-term oncological results are in the range of 25%-45%[88,89].

Surgical considerations for dCCA

dCCA affects the third portion of the extrahepatic biliary duct, which lies in a retro/intra pancreatic position. This particular anatomical configuration translates into a completely different surgical approach compared to iCCA and pCCA. In particular, resection involves pancreaticoduodenectomy, as for cancer of the pancreatic head. Negative margin status is imperative, as positive margins increase anastomotic recurrence rates and herald poor survival. An aggressive approach is justified in cases with vascular infiltration. Resection of the superior mesenteric or portal vein and reconstruction to obtain R0 obtains survival comparable to patients without vascular resection with no additional morbidity and mortality[90]. Data on arterial resection is more limited [91]. Specific for dCCA is the need to resect the bile duct high in the liver hilum as well as a lymphadenectomy of the porta hepatis and gastroduodenal ligament^[76]. Unfortunately, dCCA diagnosis is not always defined preoperatively, and these steps may be omitted, increasing the chance of R1 if the tumor has prominent intraductal spread.

SURGICAL TREATMENT: FUTURE PERSPECTIVES

Progress in the field of surgery for CCA has been limited for many years, yet the coming decade harbors great promise, with numerous innovations on the horizon. The main ongoing surgical trials are reported in Table 1.

Preoperative care

Resectability for CCA is limited mainly by inadequate FLR, especially when extensive resections are required. Portal vein ligation, PVE and ALPPS are compelling procedures for enhancing resectability with adequate FLR. On top of this, recently Guiu et al[92] described an interesting new technique, named liver venous deprivation, which involves PVE with simultaneous embolization of one or two hepatic veins. In a subsequent study, the same group demonstrated that liver venous deprivation permits a significantly greater increase in both FLR volume and function compared to PVE [93]. A randomized trial is ongoing with the aim of establishing the superiority of this technique (NCT03841305). Liver venous deprivation could represent an important advancement in liver surgery, combining the low morbidity of PVE with the greater efficacy and rapidity of ALPPS.

Minimally invasive surgery and enhanced recovery protocols

Minimally invasive approaches have developed slowly in liver surgery. Few studies specifically address the use of minimally invasive surgery for CCA with comparable outcomes, although no benefit has been clearly demonstrated so far[94]. For pCCA, the literature is discordant, but nevertheless it is possible that it could develop further in the near future[94].

Liver transplantation for unresectable CCA

Liver transplantation (LT) for hCCA has been investigated for many years, but the practice was abandoned due to very poor results compared to other indications, in the setting of the ongoing organ shortage. Initial experiences featured 5-year OS survival rates of 23%-38%, mainly due to early recurrence[95].

In the early 2000s, the idea of LT for unresectable iCCA changed thanks to the work of Vreede et al[96] at the Mayo Clinic. They developed a very rigorous protocol to optimize the selection of patients who were most likely to benefit from LT. In particular, patients with a diagnosis of unresectable, non-metastatic hCCA were



Table 1 Surgical ongoing trials for cholangiocarcinoma							
ССА Туре	Domain	Trial name	Summary				
iCCA/pCCA	Hepatic venous deprivation	NCT03841305	Randomized trial of portal vein embolization <i>vs</i> hepatic venous deprivation. Primary endpoint: future liver remnant at 3 wk				
iCCA	Liver transplantation	NCT02878473	Liver transplantation for early (< 3 cm) iCCA. Single group assignment				
iCCA	Liver transplantation	NCT04556214	Liver transplantation for stable (> 6 mo), advanced (unresectable) iCCA. Single group assignment				
iCCA	Liver transplantation	NCT04195503	Liver transplantation for stable (> 6 mo), advanced (unresectable) iCCA. Single group assignment				
pCCA	Lymphadenectomy	ChiCTR1800015688	Randomized trial of extended <i>vs</i> regional lymphadenectomy for resectable pCCA. Primary endpoint: overall survival				
pCCA	Liver transplantation	NCT02232932	Randomized trial of liver transplantation <i>vs</i> resection for resectable pCCA (< 3 cm). Primary endpoint: overall survival at 5 yr				

iCCA: Intrahepatic cholangiocarcinoma; pCCA: Perihilar cholangiocarcinoma.

treated with external beam radiotherapy (4500 cGy in 30 fractions) with concomitant intravenous 5 fluorouracil followed 3 wk later by transcatheter brachytherapy with an iridium-193 wire and finally maintenance oral capecitabine (as tolerated) until transplantation. Before LT, patients underwent staging laparotomy to exclude any intra-abdominal disease, including distant lymph node sampling. With this protocol, they reported a 5-year survival of 82% for patients undergoing LT[97]. Of note, almost half of the patients who enrolled in the protocol were not transplanted due to death or disease progression. Surgical exploration resulted in findings that precluded transplantation in 23% of cases.

Sahai *et al*[98] reported similar efficacy with a different neoadjuvant protocol consisting of higher brachytherapy doses and the omission of external beam radiotherapy. These successful experiences have been replicated in other studies[99-101]. Other studies investigated risk factors for drop-out or recurrence. Elevation of carbohydrate antigen 19-9 above 500 U/mL, a mass larger than 3 cm and Model End-stage Liver Disease score above 20 points predicted protocol drop-out before LT[102]. On the other hand, predictors for recurrence were elevated carbohydrate antigen 19-9, portal vein encasement and incomplete response to neoadjuvant therapy defined as residual tumor on the hepatectomy specimen as well as pathologic stage and perineural and perivascular invasion[99,100,102,103].

Notably, patients with hCCA developing in the setting of primary sclerosing cholangitis had a significantly better outlook than sporadic hCCA[104]. Given the technical and management complexity of this surgery, outcomes are influenced by center experience, with centers having performed at least six procedures providing the best results[104,105]. These experiences have led neoadjuvant therapy followed by LT to become the current standard of care for locally advanced non-metastatic unresectable hCCA, with both cadaveric and living donor programs active in highly specialized centers worldwide.

To date, iCCA is generally considered a contraindication to LT due to poor results in initial experiences[106]. Vilchez analyzed 440 patients with iCCA from the UNOS database and reported a significantly reduced OS with respect to HCC patients undergoing LT[107]. Yet, it may not be correct to generalize these poor results as analysis of the National Cancer Data Base revealed that only 2.2% of patients with iCCA underwent LT[107]. Furthermore, none of the studies cited so far have investigated the benefits of preoperative neoadjuvant therapy. The success and implementation of LT programs for pCCA compels consideration of this strategy for iCCA. Lunsford *et al*[108] in 2018 first reported results of their single center LT program for iCCA involving neoadjuvant chemotherapy[108]. Twelve patients were enrolled in the program, and six were transplanted. OS and disease-free survival were 83% and 50% at 5 years, respectively. Two large randomized trials are currently evaluating this approach (NCT04556214 and NCT04195503).

Whether mixed HCC-iCCA should be considered for LT is also debated. The literature is conflicting, with some studies reporting outcomes similar to HCC and others to iCCA[107,109].

Table 2 The new systemic, surgical and combined approaches to cholangiocarcinoma					
	Approaches				
Systemic therapy	(1) Overcoming chemoresistance; (2) Genetic aberration targeted therapy; (3) Immune checkpoint inhibitors; and (4) Neuroendocrine modulation of cancer growth				
Surgical therapy	(1) Liver venous deprivation; (2) Minimally invasive surgery; and (3) Liver transplantation				
Combined therapy	Liver transplantation or surgical resection after radiotherapy and/or neoadjuvant treatment				

Liver transplantation for resectable CCA

Successful results of LT after neoadjuvant therapy have induced investigators to compare them with conventional resection for resectable hCCA. Rea et al[97] reported a significantly improved OS at 5 years for patients undergoing LT compared to those undergoing resection. Ethun et al[110] showed similar results in an intention-to-treat analysis as well. They also went further and analyzed results for a subgroup of patients that were selected to be more comparable to patients in the resection group (*i.e.* hCCA not associated with primary sclerosing cholangitis, < 3 cm and lymph node negative). Even in this case, results were significantly better in the LT group. The authors suggest that the available data should prompt consideration of LT for hCCA patients with resectable disease. Indeed, this may be the new frontier in hCCA surgery. Nonetheless, some obstacles remain before the implementation of this strategy becomes widespread, the main one being the scarcity of allograft availability. In fact, critics of this approach argue that the benefit of LT (14% 5-year survival increase) is too little compared to the minimum benefit commonly applied to LT (50% at 5 years) and does not justify use of a deceased or living donor allograft. Better identification of patients who would benefit most from LT (e.g., patients who are less likely to undergo an R0 resection) could maximize the benefit and justify an LT program. In any case, a randomized trial is currently ongoing (NCT02232932).

Regarding resectable iCCA, Facciuto et al[111] recently published a small series of patients transplanted for HCC or iCCA. Their analysis showed that when iCCA features were within the Milan Criteria survival was comparable to that achieved for HCC. Further insights have come in recent years. Sapisochin et al[109] reported that the subgroup of patients transplanted for small iCCA (< 2 cm) had similar survival to HCC. In two subsequent studies, these results were confirmed with OS being significantly different between small (< 2 cm) and large tumors (> 2 cm)[71,112], 65%-73% vs 40%-45% respectively. Trials of LT for small iCCA are currently ongoing (NCT02878473).

Neoadjuvant therapy for resectable CCA

Experience with neoadjuvant therapy followed by LT has shown that disease can be stabilized in more than 50% of patients and that 57% of patients who ultimately undergo LT benefit from a complete response[97,103]. While LT seems to offer superior survival compared to resection, it is unknown to what extent neoadjuvant therapy or strict selection criteria contribute to the effect [113]. Neoadjuvant therapy may therefore prove useful in cases of resectable disease as well to increase chances of R0 resection. Consideration should be given to the risk of disease progression and loss of chance of resection. To date, there is little data available on this possible approach [114].

CONCLUSION

Poor CCA prognosis requires important therapeutic improvements in the next few years. Table 2 summarizes the new approaches in CCA therapy. Several attempts are being made or hypothesized at present, as described above in this review, with regard to systemic and/or surgical treatment for this cancer. The heterogeneity and rare occurrence of this tumor, however, impede the design of large trials with homogeneous patients. An increased understanding of the genetic changes occurring in CCA and the institution of collaborative international studies may improve this picture. The results of these efforts would be the possible definition of a model integrating different resources (diagnostic, radiological, surgical and chemotherapeutic) in order to achieve an early diagnosis and the best outcome, according to patient and tumor hallmarks. This integrated model should be implemented over time,



maintaining a strict relationship with new findings on CCA in order to adopt best practice for this lethal cancer.

REFERENCES

- Bertuccio P, Malvezzi M, Carioli G, Hashim D, Boffetta P, El-Serag HB, La Vecchia C, Negri E. 1 Global trends in mortality from intrahepatic and extrahepatic cholangiocarcinoma. J Hepatol 2019; 71: 104-114 [PMID: 30910538 DOI: 10.1016/j.jhep.2019.03.013]
- 2 Banales JM, Marin JJG, Lamarca A, Rodrigues PM, Khan SA, Roberts LR, Cardinale V, Carpino G, Andersen JB, Braconi C, Calvisi DF, Perugorria MJ, Fabris L, Boulter L, Macias RIR, Gaudio E, Alvaro D, Gradilone SA, Strazzabosco M, Marzioni M, Coulouarn C, Fouassier L, Raggi C, Invernizzi P, Mertens JC, Moncsek A, Rizvi S, Heimbach J, Koerkamp BG, Bruix J, Forner A, Bridgewater J, Valle JW, Gores GJ. Cholangiocarcinoma 2020: the next horizon in mechanisms and management. Nat Rev Gastroenterol Hepatol 2020; 17: 557-588 [PMID: 32606456 DOI: 10.1038/s41575-020-0310-z
- Nakeeb A, Pitt HA, Sohn TA, Coleman J, Abrams RA, Piantadosi S, Hruban RH, Lillemoe KD, Yeo CJ, Cameron JL. Cholangiocarcinoma. A spectrum of intrahepatic, perihilar, and distal tumors. Ann Surg 1996; 224: 463-473; discussion 473-475 [PMID: 8857851 DOI: 10.1097/00000658-199610000-00005
- 4 Aishima S, Oda Y. Pathogenesis and classification of intrahepatic cholangiocarcinoma: different characters of perihilar large duct type vs peripheral small duct type. J Hepatobiliary Pancreat Sci 2015; 22: 94-100 [PMID: 25181580 DOI: 10.1002/jhbp.154]
- Jusakul A, Cutcutache I, Yong CH, Lim JQ, Huang MN, Padmanabhan N, Nellore V, Kongpetch S, 5 Ng AWT, Ng LM, Choo SP, Myint SS, Thanan R, Nagarajan S, Lim WK, Ng CCY, Boot A, Liu M, Ong CK, Rajasegaran V, Lie S, Lim AST, Lim TH, Tan J, Loh JL, McPherson JR, Khuntikeo N, Bhudhisawasdi V, Yongvanit P, Wongkham S, Totoki Y, Nakamura H, Arai Y, Yamasaki S, Chow PK, Chung AYF, Ooi LLPJ, Lim KH, Dima S, Duda DG, Popescu I, Broet P, Hsieh SY, Yu MC, Scarpa A, Lai J, Luo DX, Carvalho AL, Vettore AL, Rhee H, Park YN, Alexandrov LB, Gordân R, Rozen SG, Shibata T, Pairojkul C, Teh BT, Tan P. Whole-Genome and Epigenomic Landscapes of Etiologically Distinct Subtypes of Cholangiocarcinoma. Cancer Discov 2017; 7: 1116-1135 [PMID: 28667006 DOI: 10.1158/2159-8290.CD-17-0368]
- 6 Cholangiocarcinoma Working Group. Italian Clinical Practice Guidelines on Cholangiocarcinoma - Part I: Classification, diagnosis and staging. Dig Liver Dis 2020; 52: 1282-1293 [PMID: 32893173 DOI: 10.1016/j.dld.2020.06.045]
- Cardinale V. Classifications and misclassification in cholangiocarcinoma. Liver Int 2019; 39: 260-262 [PMID: 30694026 DOI: 10.1111/liv.13998]
- Sato K, Zhang W, Safarikia S, Isidan A, Chen AM, Li P, Francis H, Kennedy L, Baiocchi L, Alvaro D, Glaser S, Ekser B, Alpini G. Organoids and spheroids as novel models for studying cholestatic liver injury and cholangiocarcinoma. *Hepatology* 2020 [DOI: 10.1002/hep.31653]
- Macias RIR, Banales JM, Sangro B, Muntané J, Avila MA, Lozano E, Perugorria MJ, Padillo FJ, Bujanda L, Marin JJG. The search for novel diagnostic and prognostic biomarkers in cholangiocarcinoma. Biochim Biophys Acta Mol Basis Dis 2018; 1864: 1468-1477 [PMID: 28782657 DOI: 10.1016/j.bbadis.2017.08.002]
- Song J, Li Y, Bowlus CL, Yang G, Leung PSC, Gershwin ME. Cholangiocarcinoma in Patients with 10 Primary Sclerosing Cholangitis (PSC): a Comprehensive Review. Clin Rev Allergy Immunol 2020; 58: 134-149 [PMID: 31463807 DOI: 10.1007/s12016-019-08764-7]
- 11 Tsukahara T, Shimoyama Y, Ebata T, Yokoyama Y, Igami T, Sugawara G, Mizuno T, Yamaguchi J, Nakamura S, Nagino M. Cholangiocarcinoma with intraductal tubular growth pattern vs intraductal papillary growth pattern. Mod Pathol 2016; 29: 293-301 [PMID: 26769137 DOI: 10.1038/modpathol.2015.152]
- 12 Valle J, Wasan H, Palmer DH, Cunningham D, Anthoney A, Maraveyas A, Madhusudan S, Iveson T, Hughes S, Pereira SP, Roughton M, Bridgewater J; ABC-02 Trial Investigators. Cisplatin plus gemcitabine vs gemcitabine for biliary tract cancer. N Engl J Med 2010; 362: 1273-1281 [PMID: 20375404 DOI: 10.1056/NEJMoa0908721]
- 13 Lamarca A, Palmer DH, Wasan HS, Ross PJ, Ma YT, Arora A, Falk S, Gillmore R, Wadsley J, Patel K, Anthoney A, Maraveyas A, Iveson T, Waters JS, Hobbs C, Barber S, Ryder WD, Ramage J, Davies LM, Bridgewater JA, Valle JW; Advanced Biliary Cancer Working Group. Second-line FOLFOX chemotherapy vs active symptom control for advanced biliary tract cancer (ABC-06): a phase 3, open-label, randomised, controlled trial. Lancet Oncol 2021; 22: 690-701 [PMID: 33798493 DOI: 10.1016/S1470-2045(21)00027-9]
- 14 Primrose JN, Fox RP, Palmer DH, Malik HZ, Prasad R, Mirza D, Anthony A, Corrie P, Falk S, Finch-Jones M, Wasan H, Ross P, Wall L, Wadsley J, Evans JTR, Stocken D, Praseedom R, Ma YT, Davidson B, Neoptolemos JP, Iveson T, Raftery J, Zhu S, Cunningham D, Garden OJ, Stubbs C, Valle JW, Bridgewater J; BILCAP study group. Capecitabine compared with observation in resected biliary tract cancer (BILCAP): a randomised, controlled, multicentre, phase 3 study. Lancet Oncol 2019; 20: 663-673 [PMID: 30922733 DOI: 10.1016/S1470-2045(18)30915-X]
- 15 Ebata T, Hirano S, Konishi M, Uesaka K, Tsuchiya Y, Ohtsuka M, Kaneoka Y, Yamamoto M,



Ambo Y, Shimizu Y, Ozawa F, Fukutomi A, Ando M, Nimura Y, Nagino M; Bile Duct Cancer Adjuvant Trial (BCAT) Study Group. Randomized clinical trial of adjuvant geneitabine chemotherapy vs observation in resected bile duct cancer. Br J Surg 2018; 105: 192-202 [PMID: 29405274 DOI: 10.1002/bjs.10776]

- 16 Edeline J, Benabdelghani M, Bertaut A, Watelet J, Hammel P, Joly JP, Boudjema K, Fartoux L, Bouhier-Leporrier K, Jouve JL, Faroux R, Guerin-Meyer V, Kurtz JE, Assénat E, Seitz JF, Baumgaertner I, Tougeron D, de la Fouchardière C, Lombard-Bohas C, Boucher E, Stanbury T, Louvet C, Malka D, Phelip JM. Gemcitabine and Oxaliplatin Chemotherapy or Surveillance in Resected Biliary Tract Cancer (PRODIGE 12-ACCORD 18-UNICANCER GI): A Randomized Phase III Study. J Clin Oncol 2019; 37: 658-667 [PMID: 30707660 DOI: 10.1200/JCO.18.00050]
- 17 Lischalk JW, Repka MC, Unger K. Radiation therapy for hepatobiliary malignancies. J Gastrointest Oncol 2017; 8: 279-292 [PMID: 28480067 DOI: 10.21037/jgo.2016.08.02]
- Cholangiocarcinoma Working Group. Italian Clinical Practice Guidelines on Cholangiocarcinoma 18 - Part II: Treatment. Dig Liver Dis 2020; 52: 1430-1442 [PMID: 32952071 DOI: 10.1016/j.dld.2020.08.030
- 19 Baiocchi L, Sato K, Ekser B, Kennedy L, Francis H, Ceci L, Lenci I, Alvaro D, Franchitto A, Onori P, Gaudio E, Wu C, Chakraborty S, Glaser S, Alpini G. Cholangiocarcinoma: bridging the translational gap from preclinical to clinical development and implications for future therapy. Expert Opin Investig Drugs 2021; 30: 365-375 [PMID: 33226854 DOI: 10.1080/13543784.2021.1854725]
- 20 Vicent S, Lieshout R, Saborowski A, Verstegen MMA, Raggi C, Recalcati S, Invernizzi P, van der Laan LJW, Alvaro D, Calvisi DF, Cardinale V. Experimental models to unravel the molecular pathogenesis, cell of origin and stem cell properties of cholangiocarcinoma. Liver Int 2019; 39 Suppl 1: 79-97 [PMID: 30851232 DOI: 10.1111/liv.14094]
- 21 Fabris L, Sato K, Alpini G, Strazzabosco M. The Tumor Microenvironment in Cholangiocarcinoma Progression. Hepatology 2021; 73 Suppl 1: 75-85 [PMID: 32500550 DOI: 10.1002/hep.31410]
- 22 Marin JJG, Lozano E, Herraez E, Asensio M, Di Giacomo S, Romero MR, Briz O, Serrano MA, Efferth T, Macias RIR. Chemoresistance and chemosensitization in cholangiocarcinoma. Biochim Biophys Acta Mol Basis Dis 2018; 1864: 1444-1453 [PMID: 28600147 DOI: 10.1016/j.bbadis.2017.06.005]
- 23 Yeldag G, Rice A, Del Río Hernández A. Chemoresistance and the Self-Maintaining Tumor Microenvironment. Cancers (Basel) 2018; 10 [PMID: 30487436 DOI: 10.3390/cancers10120471]
- 24 Herraez E, Lozano E, Macias RI, Vaquero J, Bujanda L, Banales JM, Marin JJ, Briz O. Expression of SLC22A1 variants may affect the response of hepatocellular carcinoma and cholangiocarcinoma to sorafenib. Hepatology 2013; 58: 1065-1073 [PMID: 23532667 DOI: 10.1002/hep.26425]
- Vaquero J, Guedj N, Clapéron A, Nguyen Ho-Bouldoires TH, Paradis V, Fouassier L. Epithelial-25 mesenchymal transition in cholangiocarcinoma: From clinical evidence to regulatory networks. J Hepatol 2017; 66: 424-441 [PMID: 27686679 DOI: 10.1016/j.jhep.2016.09.010]
- 26 Kapacee ZA, Knox JJ, Palmer D, Blagden SP, Lamarca A, Valle JW, McNamara MG. NUC-1031, use of ProTide technology to circumvent gemcitabine resistance: current status in clinical trials. Med Oncol 2020; 37: 61 [PMID: 32529264 DOI: 10.1007/s12032-020-01386-6]
- 27 Kayhanian H, Smyth EC, Braconi C. Emerging molecular targets and therapy for cholangiocarcinoma. World J Gastrointest Oncol 2017; 9: 268-280 [PMID: 28808500 DOI: 10.4251/wjgo.v9.i7.268]
- Massironi S, Pilla L, Elvevi A, Longarini R, Rossi RE, Bidoli P, Invernizzi P. New and Emerging 28 Systemic Therapeutic Options for Advanced Cholangiocarcinoma. Cells 2020; 9 [PMID: 32168869 DOI: 10.3390/cells9030688]
- Lavingia V, Fakih M. Impressive response to dual BRAF and MEK inhibition in patients with 29 BRAF mutant intrahepatic cholangiocarcinoma-2 case reports and a brief review. J Gastrointest Oncol 2016; 7: E98-E102 [PMID: 28078132 DOI: 10.21037/jgo.2016.09.13]
- 30 Subbiah V, Lassen U, Élez E, Italiano A, Curigliano G, Javle M, de Braud F, Prager GW, Greil R, Stein A, Fasolo A, Schellens JHM, Wen PY, Viele K, Boran AD, Gasal E, Burgess P, Ilankumaran P, Wainberg ZA. Dabrafenib plus trametinib in patients with BRAF^{V600E}-mutated biliary tract cancer (ROAR): a phase 2, open-label, single-arm, multicentre basket trial. Lancet Oncol 2020; 21: 1234-1243 [PMID: 32818466 DOI: 10.1016/S1470-2045(20)30321-1]
- Mossahebi-Mohammadi M, Quan M, Zhang JS, Li X. FGF Signaling Pathway: A Key Regulator 31 of Stem Cell Pluripotency. Front Cell Dev Biol 2020; 8: 79 [PMID: 32133359 DOI: 10.3389/fcell.2020.00079]
- 32 Mahipal A, Tella SH, Kommalapati A, Anaya D, Kim R. FGFR2 genomic aberrations: Achilles heel in the management of advanced cholangiocarcinoma. Cancer Treat Rev 2019; 78: 1-7 [PMID: 31255945 DOI: 10.1016/j.ctrv.2019.06.003]
- 33 Javle M, Lowery M, Shroff RT, Weiss KH, Springfeld C, Borad MJ, Ramanathan RK, Goyal L, Sadeghi S, Macarulla T, El-Khoueiry A, Kelley RK, Borbath I, Choo SP, Oh DY, Philip PA, Chen LT, Reungwetwattana T, Van Cutsem E, Yeh KH, Ciombor K, Finn RS, Patel A, Sen S, Porter D, Isaacs R, Zhu AX, Abou-Alfa GK, Bekaii-Saab T. Phase II Study of BGJ398 in Patients With FGFR-Altered Advanced Cholangiocarcinoma. J Clin Oncol 2018; 36: 276-282 [PMID: 29182496 DOI: 10.1200/JCO.2017.75.5009]
- 34 Abou-Alfa GK, Sahai V, Hollebecque A, Vaccaro G, Melisi D, Al-Rajabi R, Paulson AS, Borad MJ, Gallinson D, Murphy AG, Oh DY, Dotan E, Catenacci DV, Van Cutsem E, Ji T, Lihou CF, Zhen H, Féliz L, Vogel A. Pemigatinib for previously treated, locally advanced or metastatic



cholangiocarcinoma: a multicentre, open-label, phase 2 study. Lancet Oncol 2020; 21: 671-684 [PMID: 32203698 DOI: 10.1016/S1470-2045(20)30109-1]

- 35 Abou-Alfa GK, Macarulla T, Javle MM, Kelley RK, Lubner SJ, Adeva J, Cleary JM, Catenacci DV, Borad MJ, Bridgewater J, Harris WP, Murphy AG, Oh DY, Whisenant J, Lowery MA, Goyal L, Shroff RT, El-Khoueiry AB, Fan B, Wu B, Chamberlain CX, Jiang L, Gliser C, Pandya SS, Valle JW, Zhu AX. Ivosidenib in IDH1-mutant, chemotherapy-refractory cholangiocarcinoma (ClarIDHy): a multicentre, randomised, double-blind, placebo-controlled, phase 3 study. Lancet Oncol 2020; 21: 796-807 [PMID: 32416072 DOI: 10.1016/S1470-2045(20)30157-1]
- Crispo F, Pietrafesa M, Condelli V, Maddalena F, Bruno G, Piscazzi A, Sgambato A, Esposito F, 36 Landriscina M. IDH1 Targeting as a New Potential Option for Intrahepatic Cholangiocarcinoma Treatment-Current State and Future Perspectives. *Molecules* 2020; 25 [PMID: 32824685 DOI: 10.3390/molecules25163754]
- Jin W. ErBb Family Proteins in Cholangiocarcinoma and Clinical Implications. J Clin Med 2020; 9 37 [PMID: 32708604 DOI: 10.3390/jcm9072255]
- 38 Paluch C, Santos AM, Anzilotti C, Cornall RJ, Davis SJ. Immune Checkpoints as Therapeutic Targets in Autoimmunity. Front Immunol 2018; 9: 2306 [PMID: 30349540 DOI: 10.3389/fimmu.2018.02306]
- 39 Darvin P, Toor SM, Sasidharan Nair V, Elkord E. Immune checkpoint inhibitors: recent progress and potential biomarkers. Exp Mol Med 2018; 50: 1-11 [PMID: 30546008 DOI: 10.1038/s12276-018-0191-1]
- 40 Robert C. A decade of immune-checkpoint inhibitors in cancer therapy. Nat Commun 2020; 11: 3801 [PMID: 32732879 DOI: 10.1038/s41467-020-17670-y]
- 41 Marabelle A, Le DT, Ascierto PA, Di Giacomo AM, De Jesus-Acosta A, Delord JP, Geva R, Gottfried M, Penel N, Hansen AR, Piha-Paul SA, Doi T, Gao B, Chung HC, Lopez-Martin J, Bang YJ, Frommer RS, Shah M, Ghori R, Joe AK, Pruitt SK, Diaz LA Jr. Efficacy of Pembrolizumab in Patients With Noncolorectal High Microsatellite Instability/Mismatch Repair-Deficient Cancer: Results From the Phase II KEYNOTE-158 Study. J Clin Oncol 2020; 38: 1-10 [PMID: 31682550 DOI: 10.1200/JCO.19.02105]
- 42 Ahn S, Lee JC, Shin DW, Kim J, Hwang JH. High PD-L1 expression is associated with therapeutic response to pembrolizumab in patients with advanced biliary tract cancer. Sci Rep 2020; 10: 12348 [PMID: 32704067 DOI: 10.1038/s41598-020-69366-4]
- 43 Sato K, Francis H, Zhou T, Meng F, Kennedy L, Ekser B, Baiocchi L, Onori P, Mancinelli R, Gaudio E, Franchitto A, Glaser S, Alpini G. Neuroendocrine Changes in Cholangiocarcinoma Growth. Cells 2020; 9 [PMID: 32069926 DOI: 10.3390/cells9020436]
- 44 Onori P, Wise C, Gaudio E, Franchitto A, Francis H, Carpino G, Lee V, Lam I, Miller T, Dostal DE, Glaser SS. Secretin inhibits cholangiocarcinoma growth via dysregulation of the cAMPdependent signaling mechanisms of secretin receptor. Int J Cancer 2010; 127: 43-54 [PMID: 19904746 DOI: 10.1002/ijc.25028]
- Tan CK, Podila PV, Taylor JE, Nagorney DM, Wiseman GA, Gores GJ, LaRusso NF. Human 45 cholangiocarcinomas express somatostatin receptors and respond to somatostatin with growth inhibition. Gastroenterology 1995; 108: 1908-1916 [PMID: 7768398 DOI: 10.1016/0016-5085(95)90157-4
- 46 Han Y, Demorrow S, Invernizzi P, Jing Q, Glaser S, Renzi A, Meng F, Venter J, Bernuzzi F, White M, Francis H, Lleo A, Marzioni M, Onori P, Alvaro D, Torzilli G, Gaudio E, Alpini G. Melatonin exerts by an autocrine loop antiproliferative effects in cholangiocarcinoma: its synthesis is reduced favoring cholangiocarcinoma growth. Am J Physiol Gastrointest Liver Physiol 2011; 301: G623-G633 [PMID: 21778461 DOI: 10.1152/ajpgi.00118.2011]
- Fiebiger WC, Scheithauer W, Traub T, Kurtaran A, Gedlicka C, Kornek GV, Virgolini I, Raderer 47 M. Absence of therapeutic efficacy of the somatostatin analogue lanreotide in advanced primary hepatic cholangiocellular cancer and adenocarcinoma of the gallbladder despite in vivo somatostatinreceptor expression. Scand J Gastroenterol 2002; 37: 222-225 [PMID: 11843061 DOI: 10.1080/0036552027534169111
- 48 Yoshikawa D, Ojima H, Iwasaki M, Hiraoka N, Kosuge T, Kasai S, Hirohashi S, Shibata T. Clinicopathological and prognostic significance of EGFR, VEGF, and HER2 expression in cholangiocarcinoma. Br J Cancer 2008; 98: 418-425 [PMID: 18087285 DOI: 10.1038/sj.bjc.6604129]
- Zhu AX, Meyerhardt JA, Blaszkowsky LS, Kambadakone AR, Muzikansky A, Zheng H, Clark JW, 49 Abrams TA, Chan JA, Enzinger PC, Bhargava P, Kwak EL, Allen JN, Jain SR, Stuart K, Horgan K, Sheehan S, Fuchs CS, Ryan DP, Sahani DV. Efficacy and safety of gemcitabine, oxaliplatin, and bevacizumab in advanced biliary-tract cancers and correlation of changes in 18-fluorodeoxyglucose PET with clinical outcome: a phase 2 study. Lancet Oncol 2010; 11: 48-54 [PMID: 19932054 DOI: 10.1016/S1470-2045(09)70333-X
- 50 Idrees JJ, Merath K, Gani F, Bagante F, Mehta R, Beal E, Cloyd JM, Pawlik TM. Trends in centralization of surgical care and compliance with National Cancer Center Network guidelines for resected cholangiocarcinoma. HPB (Oxford) 2019; 21: 981-989 [PMID: 30591307 DOI: 10.1016/j.hpb.2018.11.013
- 51 Mansour JC, Aloia TA, Crane CH, Heimbach JK, Nagino M, Vauthey JN. Hilar cholangiocarcinoma: expert consensus statement. HPB (Oxford) 2015; 17: 691-699 [PMID: 26172136 DOI: 10.1111/hpb.12450]



- 52 Miyazaki M, Yoshitomi H, Miyakawa S, Uesaka K, Unno M, Endo I, Ota T, Ohtsuka M, Kinoshita H, Shimada K, Shimizu H, Tabata M, Chijiiwa K, Nagino M, Hirano S, Wakai T, Wada K, Isayama H, Okusaka T, Tsuyuguchi T, Fujita N, Furuse J, Yamao K, Murakami K, Yamazaki H, Kijima H, Nakanuma Y, Yoshida M, Takayashiki T, Takada T. Clinical practice guidelines for the management of biliary tract cancers 2015: the 2nd English edition. J Hepatobiliary Pancreat Sci 2015; 22: 249-273 [PMID: 25787274 DOI: 10.1002/jhbp.233]
- 53 Wiggers JK, Groot Koerkamp B, Cieslak KP, Doussot A, van Klaveren D, Allen PJ, Besselink MG, Busch OR, D'Angelica MI, DeMatteo RP, Gouma DJ, Kingham TP, van Gulik TM, Jarnagin WR. Postoperative Mortality after Liver Resection for Perihilar Cholangiocarcinoma: Development of a Risk Score and Importance of Biliary Drainage of the Future Liver Remnant. J Am Coll Surg 2016; 223: 321-331.e1 [PMID: 27063572 DOI: 10.1016/j.jamcollsurg.2016.03.035]
- 54 Kennedy TJ, Yopp A, Qin Y, Zhao B, Guo P, Liu F, Schwartz LH, Allen P, D'Angelica M, Fong Y, DeMatteo RP, Blumgart LH, Jarnagin WR. Role of preoperative biliary drainage of liver remnant prior to extended liver resection for hilar cholangiocarcinoma. HPB (Oxford) 2009; 11: 445-451 [PMID: 19768150 DOI: 10.1111/j.1477-2574.2009.00090.x]
- Lidsky ME, Jarnagin WR. Surgical management of hilar cholangiocarcinoma at Memorial Sloan 55 Kettering Cancer Center. Ann Gastroenterol Surg 2018; 2: 304-312 [PMID: 30003193 DOI: 10.1002/ags3.12181
- Kawakami H, Kuwatani M, Onodera M, Haba S, Eto K, Ehira N, Yamato H, Kudo T, Tanaka E, 56 Hirano S, Kondo S, Asaka M. Endoscopic nasobiliary drainage is the most suitable preoperative biliary drainage method in the management of patients with hilar cholangiocarcinoma. J Gastroenterol 2011; 46: 242-248 [PMID: 20700608 DOI: 10.1007/s00535-010-0298-1]
- 57 Kawashima H, Itoh A, Ohno E, Itoh Y, Ebata T, Nagino M, Goto H, Hirooka Y. Preoperative endoscopic nasobiliary drainage in 164 consecutive patients with suspected perihilar cholangiocarcinoma: a retrospective study of efficacy and risk factors related to complications. Ann Surg 2013; 257: 121-127 [PMID: 22895398 DOI: 10.1097/SLA.0b013e318262b2e9]
- 58 She WH, Cheung TT, Ma KW, Tsang SHY, Dai WC, Chan ACY, Lo CM. Defining the optimal bilirubin level before hepatectomy for hilar cholangiocarcinoma. BMC Cancer 2020; 20: 914 [PMID: 32967634 DOI: 10.1186/s12885-020-07385-0]
- Li J, Moustafa M, Linecker M, Lurje G, Capobianco I, Baumgart J, Ratti F, Rauchfuss F, Balci D, 59 Fernandes E, Montalti R, Robles-Campos R, Bjornsson B, Topp SA, Fronek J, Liu C, Wahba R, Bruns C, Brunner SM, Schlitt HJ, Heumann A, Stüben BO, Izbicki JR, Bednarsch J, Gringeri E, Fasolo E, Rolinger J, Kristek J, Hernandez-Alejandro R, Schnitzbauer A, Nuessler N, Schön MR, Voskanyan S, Petrou AS, Hahn O, Soejima Y, Vicente E, Castro-Benitez C, Adam R, Tomassini F, Troisi RI, Kantas A, Oldhafer KJ, Ardiles V, de Santibanes E, Malago M, Clavien PA, Vivarelli M, Settmacher U, Aldrighetti L, Neumann U, Petrowsky H, Cillo U, Lang H, Nadalin S. ALPPS for Locally Advanced Intrahepatic Cholangiocarcinoma: Did Aggressive Surgery Lead to the Oncological Benefit? Ann Surg Oncol 2020; 27: 1372-1384 [PMID: 32002719 DOI: 10.1245/s10434-019-08192-z]
- Goere D, Wagholikar GD, Pessaux P, Carrère N, Sibert A, Vilgrain V, Sauvanet A, Belghiti J. Utility of staging laparoscopy in subsets of biliary cancers : laparoscopy is a powerful diagnostic tool in patients with intrahepatic and gallbladder carcinoma. Surg Endosc 2006; 20: 721-725 [PMID: 16508808 DOI: 10.1007/s00464-005-0583-x]
- 61 Weber SM, Ribero D, O'Reilly EM, Kokudo N, Miyazaki M, Pawlik TM. Intrahepatic cholangiocarcinoma: expert consensus statement. HPB (Oxford) 2015; 17: 669-680 [PMID: 26172134 DOI: 10.1111/hpb.12441]
- 62 Endo I, Gonen M, Yopp AC, Dalal KM, Zhou Q, Klimstra D, D'Angelica M, DeMatteo RP, Fong Y, Schwartz L, Kemeny N, O'Reilly E, Abou-Alfa GK, Shimada H, Blumgart LH, Jarnagin WR. Intrahepatic cholangiocarcinoma: rising frequency, improved survival, and determinants of outcome after resection. Ann Surg 2008; 248: 84-96 [PMID: 18580211 DOI: 10.1097/SLA.0b013e318176c4d3]
- 63 Luo X, Yuan L, Wang Y, Ge R, Sun Y, Wei G. Survival outcomes and prognostic factors of surgical therapy for all potentially resectable intrahepatic cholangiocarcinoma: a large single-center cohort study. J Gastrointest Surg 2014; 18: 562-572 [PMID: 24395070 DOI: 10.1007/s11605-013-2447-3]
- 64 Farges O, Fuks D, Boleslawski E, Le Treut YP, Castaing D, Laurent A, Ducerf C, Rivoire M, Bachellier P, Chiche L, Nuzzo G, Regimbeau JM. Influence of surgical margins on outcome in patients with intrahepatic cholangiocarcinoma: a multicenter study by the AFC-IHCC-2009 study group. Ann Surg 2011; 254: 824-829; discussion 830 [PMID: 22042474 DOI: 10.1097/SLA.0b013e318236c21d]
- Nakagawa T, Kamiyama T, Kurauchi N, Matsushita M, Nakanishi K, Kamachi H, Kudo T, Todo S. 65 Number of lymph node metastases is a significant prognostic factor in intrahepatic cholangiocarcinoma. World J Surg 2005; 29: 728-733 [PMID: 15880276 DOI: 10.1007/s00268-005-7761-9
- Ribero D, Pinna AD, Guglielmi A, Ponti A, Nuzzo G, Giulini SM, Aldrighetti L, Calise F, Gerunda 66 GE, Tomatis M, Amisano M, Berloco P, Torzilli G, Capussotti L; Italian Intrahepatic Cholangiocarcinoma Study Group. Surgical Approach for Long-term Survival of Patients With Intrahepatic Cholangiocarcinoma: A Multi-institutional Analysis of 434 Patients. Arch Surg 2012; 147: 1107-1113 [PMID: 22910846 DOI: 10.1001/archsurg.2012.1962]
- Bridgewater J, Galle PR, Khan SA, Llovet JM, Park JW, Patel T, Pawlik TM, Gores GJ. Guidelines 67


for the diagnosis and management of intrahepatic cholangiocarcinoma. J Hepatol 2014; 60: 1268-1289 [PMID: 24681130 DOI: 10.1016/j.jhep.2014.01.021]

- 68 Nathan H, Aloia TA, Vauthey JN, Abdalla EK, Zhu AX, Schulick RD, Choti MA, Pawlik TM. A proposed staging system for intrahepatic cholangiocarcinoma. Ann Surg Oncol 2009; 16: 14-22 [PMID: 18987916 DOI: 10.1245/s10434-008-0180-z]
- 69 Reames BN, Ejaz A, Koerkamp BG, Alexandrescu S, Marques HP, Aldrighetti L, Maithel SK, Pulitano C, Bauer TW, Shen F, Poultsides GA, Martel G, Marsh JW, Pawlik TM. Impact of major vascular resection on outcomes and survival in patients with intrahepatic cholangiocarcinoma: A multi-institutional analysis. J Surg Oncol 2017; 116: 133-139 [PMID: 28411373 DOI: 10.1002/jso.24633]
- 70 Lang H, Sotiropoulos GC, Sgourakis G, Schmitz KJ, Paul A, Hilgard P, Zöpf T, Trarbach T, Malagó M, Baba HA, Broelsch CE. Operations for intrahepatic cholangiocarcinoma: singleinstitution experience of 158 patients. J Am Coll Surg 2009; 208: 218-228 [PMID: 19228533 DOI: 10.1016/j.jamcollsurg.2008.10.017]
- Sapisochin G, Rodríguez de Lope C, Gastaca M, Ortiz de Urbina J, Suarez MA, Santoyo J, 71 Castroagudín JF, Varo E, López-Andujar R, Palacios F, Sanchez Antolín G, Perez B, Guiberteau A, Blanco G, González-Diéguez ML, Rodriguez M, Varona MA, Barrera MA, Fundora Y, Ferron JA, Ramos E, Fabregat J, Ciria R, Rufian S, Otero A, Vazquez MA, Pons JA, Parrilla P, Zozaya G, Herrero JI, Charco R, Bruix J. "Very early" intrahepatic cholangiocarcinoma in cirrhotic patients: should liver transplantation be reconsidered in these patients? Am J Transplant 2014; 14: 660-667 [PMID: 24410861 DOI: 10.1111/ajt.12591]
- 72 Raptis DA, Linecker M, Kambakamba P, Tschuor C, Müller PC, Hadjittofi C, Stavrou GA, Fard-Aghaie MH, Tun-Abraham M, Ardiles V, Malagó M, Campos RR, Oldhafer KJ, Hernandez-Alejandro R, de Santibañes E, Machado MA, Petrowsky H, Clavien PA. Defining Benchmark Outcomes for ALPPS. Ann Surg 2019; 270: 835-841 [PMID: 31592812 DOI: 10.1097/SLA.00000000003539]
- 73 Le Roy B, Gelli M, Pittau G, Allard MA, Pereira B, Serji B, Vibert E, Castaing D, Adam R, Cherqui D, Sa Cunha A. Neoadjuvant chemotherapy for initially unresectable intrahepatic cholangiocarcinoma. Br J Surg 2018; 105: 839-847 [PMID: 28858392 DOI: 10.1002/bjs.10641]
- Spolverato G, Kim Y, Alexandrescu S, Marques HP, Lamelas J, Aldrighetti L, Clark Gamblin T, 74 Maithel SK, Pulitano C, Bauer TW, Shen F, Poultsides GA, Tran TB, Wallis Marsh J, Pawlik TM. Management and Outcomes of Patients with Recurrent Intrahepatic Cholangiocarcinoma Following Previous Curative-Intent Surgical Resection. Ann Surg Oncol 2016; 23: 235-243 [PMID: 26059651 DOI: 10.1245/s10434-015-4642-9]
- 75 Yoh T, Hatano E, Seo S, Okuda Y, Fuji H, Ikeno Y, Taura K, Yasuchika K, Okajima H, Kaido T, Uemoto S. Long-Term Survival of Recurrent Intrahepatic Cholangiocarcinoma: The Impact and Selection of Repeat Surgery. World J Surg 2018; 42: 1848-1856 [PMID: 29218465 DOI: 10.1007/s00268-017-4387-7]
- Valle JW, Borbath I, Khan SA, Huguet F, Gruenberger T, Arnold D; ESMO Guidelines Committee. 76 Biliary cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol 2016; 27: v28-v37 [PMID: 27664259 DOI: 10.1093/annonc/mdw324]
- 77 Watson MD, Baimas-George MR, Passeri MJ, Sulzer JK, Baker EH, Ocuin LM, Martinie JB, Iannitti DA, Vrochides D. Effect of Margin Status on Survival After Resection of Hilar Cholangiocarcinoma in the Modern Era of Adjuvant Therapies. Am Surg 2020; 3134820973401 [PMID: 33345594 DOI: 10.1177/0003134820973401]
- Otsuka S, Ebata T, Yokoyama Y, Mizuno T, Tsukahara T, Shimoyama Y, Ando M, Nagino M. 78 Clinical value of additional resection of a margin-positive distal bile duct in perihilar cholangiocarcinoma. Br J Surg 2019; 106: 774-782 [PMID: 30889275 DOI: 10.1002/bjs.11125]
- 79 Zhang XF, Squires MH 3rd, Bagante F, Ethun CG, Salem A, Weber SM, Tran T, Poultsides G, Son AY, Hatzaras I, Jin L, Fields RC, Weiss M, Scoggins C, Martin RCG, Isom CA, Idrees K, Mogal HD, Shen P, Maithel SK, Schmidt CR, Pawlik TM. The Impact of Intraoperative Re-Resection of a Positive Bile Duct Margin on Clinical Outcomes for Hilar Cholangiocarcinoma. Ann Surg Oncol 2018; 25: 1140-1149 [PMID: 29470820 DOI: 10.1245/s10434-018-6382-0]
- Stremitzer S, Stift J, Laengle J, Schwarz C, Kaczirek K, Jones RP, Quinn LM, Fenwick SW, Diaz-Nieto R, Poston GJ, Malik HZ. Prognosis and Circumferential Margin in Patients with Resected Hilar Cholangiocarcinoma. Ann Surg Oncol 2021; 28: 1493-1498 [PMID: 32914390 DOI: 10.1245/s10434-020-09105-11
- Cai Y, Cheng N, Ye H, Li F, Song P, Tang W. The current management of cholangiocarcinoma: A 81 comparison of current guidelines. Biosci Trends 2016; 10: 92-102 [PMID: 27026485 DOI: 10.5582/bst.2016.01048]
- 82 Kambakamba P, Linecker M, Slankamenac K, DeOliveira ML. Lymph node dissection in resectable perihilar cholangiocarcinoma: a systematic review. Am J Surg 2015; 210: 694-701 [PMID: 26212390 DOI: 10.1016/j.amjsurg.2015.05.015]
- 83 He M, Xu X, Feng H, Chen W, Liu H, Zhang Y, Wang J, Geng Z, Qiu Y, Duan W, Li X, Zhi X, Zhu W, Li F, Li J, Li S, He Y, Quan Z. Regional lymphadenectomy vs. extended lymphadenectomy for hilar cholangiocarcinoma (Relay-HC trial): study protocol for a prospective, multicenter, randomized controlled trial. Trials 2019; 20: 528 [PMID: 31443731 DOI: 10.1186/s13063-019-3605-z]
- 84 Ma WJ, Wu ZR, Hu HJ, Wang JK, Yin CH, Shi YJ, Li FY, Cheng NS. Extended Lymphadenectomy Versus Regional Lymphadenectomy in Resectable Hilar Cholangiocarcinoma. J



Gastrointest Surg 2020; 24: 1619-1629 [PMID: 31147975 DOI: 10.1007/s11605-019-04244-7]

- Olthof PB, Coelen RJS, Wiggers JK, Groot Koerkamp B, Malago M, Hernandez-Alejandro R, Topp 85 SA, Vivarelli M, Aldrighetti LA, Robles Campos R, Oldhafer KJ, Jarnagin WR, van Gulik TM. High mortality after ALPPS for perihilar cholangiocarcinoma: case-control analysis including the first series from the international ALPPS registry. HPB (Oxford) 2017; 19: 381-387 [PMID: 28279621 DOI: 10.1016/j.hpb.2016.10.008]
- 86 Balci D, Sakamoto Y, Li J, Di Benedetto F, Kirimker EO, Petrowsky H. Associating liver partition and portal vein ligation for staged hepatectomy (ALPPS) procedure for cholangiocarcinoma. Int J Surg 2020; 82S: 97-102 [PMID: 32645441 DOI: 10.1016/j.ijsu.2020.06.045]
- 87 Mizuno T, Ebata T, Nagino M. Advanced hilar cholangiocarcinoma: An aggressive surgical approach for the treatment of advanced hilar cholangiocarcinoma: Perioperative management, extended procedures, and multidisciplinary approaches. Surg Oncol 2020; 33: 201-206 [PMID: 31301935 DOI: 10.1016/j.suronc.2019.07.002]
- Aoki T, Sakamoto Y, Kohno Y, Akamatsu N, Kaneko J, Sugawara Y, Hasegawa K, Makuuchi M, 88 Kokudo N. Hepatopancreaticoduodenectomy for Biliary Cancer: Strategies for Near-zero Operative Mortality and Acceptable Long-term Outcome. Ann Surg 2018; 267: 332-337 [PMID: 27811506 DOI: 10.1097/SLA.000000000002059]
- 89 Sakamoto Y, Nara S, Kishi Y, Esaki M, Shimada K, Kokudo N, Kosuge T. Is extended hemihepatectomy plus pancreaticoduodenectomy justified for advanced bile duct cancer and gallbladder cancer? Surgery 2013; 153: 794-800 [PMID: 23415082 DOI: 10.1016/j.surg.2012.11.024]
- Riediger H, Makowiec F, Fischer E, Adam U, Hopt UT. Postoperative morbidity and long-term 90 survival after pancreaticoduodenectomy with superior mesenterico-portal vein resection. J Gastrointest Surg 2006; 10: 1106-1115 [PMID: 16966029 DOI: 10.1016/j.gassur.2006.04.002]
- 91 Stitzenberg KB, Watson JC, Roberts A, Kagan SA, Cohen SJ, Konski AA, Hoffman JP. Survival after pancreatectomy with major arterial resection and reconstruction. Ann Surg Oncol 2008; 15: 1399-1406 [PMID: 18320285 DOI: 10.1245/s10434-008-9844-y]
- 92 Guiu B, Chevallier P, Denys A, Delhom E, Pierredon-Foulongne MA, Rouanet P, Fabre JM, Quenet F, Herrero A, Panaro F, Baudin G, Ramos J. Simultaneous trans-hepatic portal and hepatic vein embolization before major hepatectomy: the liver venous deprivation technique. Eur Radiol 2016; 26: 4259-4267 [PMID: 27090112 DOI: 10.1007/s00330-016-4291-9]
- Guiu B, Quenet F, Panaro F, Piron L, Cassinotto C, Herrerro A, Souche FR, Hermida M, Pierredon-93 Foulongne MA, Belgour A, Aho-Glele S, Deshayes E. Liver venous deprivation vs portal vein embolization before major hepatectomy: future liver remnant volumetric and functional changes. Hepatobiliary Surg Nutr 2020; 9: 564-576 [PMID: 33163507 DOI: 10.21037/hbsn.2020.02.06]
- Shiraiwa DK, Carvalho PFDC, Maeda CT, Silva LC, Forones NM, Lopes-Filho GJ, Linhares MM, 94 Araujo RLC. The role of minimally invasive hepatectomy for hilar and intrahepatic cholangiocarcinoma: A systematic review of the literature. J Surg Oncol 2020; 121: 863-872 [PMID: 31902142 DOI: 10.1002/jso.25821]
- 95 Meyer CG, Penn I, James L. Liver transplantation for cholangiocarcinoma: results in 207 patients. Transplantation 2000; 69: 1633-1637 [PMID: 10836374 DOI: 10.1097/00007890-200004270-00019]
- 96 De Vreede I, Steers JL, Burch PA, Rosen CB, Gunderson LL, Haddock MG, Burgart L, Gores GJ. Prolonged disease-free survival after orthotopic liver transplantation plus adjuvant chemoirradiation for cholangiocarcinoma. Liver Transpl 2000; 6: 309-316 [PMID: 10827231 DOI: 10.1053/Lv.2000.6143]
- Rea DJ, Heimbach JK, Rosen CB, Haddock MG, Alberts SR, Kremers WK, Gores GJ, Nagorney 97 DM. Liver transplantation with neoadjuvant chemoradiation is more effective than resection for hilar cholangiocarcinoma. Ann Surg 2005; 242: 451-458; discussion 458-461 [PMID: 16135931 DOI: 10.1097/01.sla.0000179678.13285.fa]
- Sahai P, Kumar S. External radiotherapy and brachytherapy in the management of extrahepatic and 98 intrahepatic cholangiocarcinoma: available evidence. Br J Radiol 2017; 90: 20170061 [PMID: 28466653 DOI: 10.1259/bjr.20170061]
- Duignan S, Maguire D, Ravichand CS, Geoghegan J, Hoti E, Fennelly D, Armstrong J, Rock K, 99 Mohan H, Traynor O. Neoadjuvant chemoradiotherapy followed by liver transplantation for unresectable cholangiocarcinoma: a single-centre national experience. HPB (Oxford) 2014; 16: 91-98 [PMID: 23600750 DOI: 10.1111/hpb.12082]
- 100 Sio TT, Martenson JA Jr, Haddock MG, Novotny PJ, Gores GJ, Alberts SR, Miller RC, Heimbach JK, Rosen CB. Outcome of Transplant-fallout Patients With Unresectable Cholangiocarcinoma. Am J Clin Oncol 2016; 39: 271-275 [PMID: 24921218 DOI: 10.1097/COC.00000000000056]
- Darwish Murad S, Kim WR, Harnois DM, Douglas DD, Burton J, Kulik LM, Botha JF, Mezrich 101 JD, Chapman WC, Schwartz JJ, Hong JC, Emond JC, Jeon H, Rosen CB, Gores GJ, Heimbach JK. Efficacy of neoadjuvant chemoradiation, followed by liver transplantation, for perihilar cholangiocarcinoma at 12 US centers. Gastroenterology 2012; 143: 88-98.e3; quiz e14 [PMID: 22504095 DOI: 10.1053/j.gastro.2012.04.008]
- Darwish Murad S, Kim WR, Therneau T, Gores GJ, Rosen CB, Martenson JA, Alberts SR, 102 Heimbach JK. Predictors of pretransplant dropout and posttransplant recurrence in patients with perihilar cholangiocarcinoma. Hepatology 2012; 56: 972-981 [PMID: 22290335 DOI: 10.1002/hep.25629]



- Lehrke HD, Heimbach JK, Wu TT, Jenkins SM, Gores GJ, Rosen CB, Mounajjed T. Prognostic 103 Significance of the Histologic Response of Perihilar Cholangiocarcinoma to Preoperative Neoadjuvant Chemoradiation in Liver Explants. Am J Surg Pathol 2016; 40: 510-518 [PMID: 26752544 DOI: 10.1097/PAS.000000000000588]
- Tan EK, Taner T, Heimbach JK, Gores GJ, Rosen CB. Liver Transplantation for Peri-hilar 104 Cholangiocarcinoma. J Gastrointest Surg 2020; 24: 2679-2685 [PMID: 32671802 DOI: 10.1007/s11605-020-04721-4]
- 105 Kitajima T, Hibi T, Moonka D, Sapisochin G, Abouljoud MS, Nagai S. Center Experience Affects Liver Transplant Outcomes in Patients with Hilar Cholangiocarcinoma. Ann Surg Oncol 2020; 27: 5209-5221 [PMID: 32495286 DOI: 10.1245/s10434-020-08682-5]
- 106 O'Grady JG, Polson RJ, Rolles K, Calne RY, Williams R. Liver transplantation for malignant disease. Results in 93 consecutive patients. Ann Surg 1988; 207: 373-379 [PMID: 2451484 DOI: 10.1097/00000658-198804000-00002
- 107 Vilchez V, Shah MB, Daily MF, Pena L, Tzeng CW, Davenport D, Hosein PJ, Gedaly R, Maynard E. Long-term outcome of patients undergoing liver transplantation for mixed hepatocellular carcinoma and cholangiocarcinoma: an analysis of the UNOS database. HPB (Oxford) 2016; 18: 29-34 [PMID: 26776848 DOI: 10.1016/j.hpb.2015.10.001]
- 108 Lunsford KE, Javle M, Heyne K, Shroff RT, Abdel-Wahab R, Gupta N, Mobley CM, Saharia A, Victor DW, Nguyen DT, Graviss EA, Kaseb AO, McFadden RS, Aloia TA, Conrad C, Li XC, Monsour HP, Gaber AO, Vauthey JN, Ghobrial RM; Methodist-MD Anderson Joint Cholangiocarcinoma Collaborative Committee (MMAJCCC). Liver transplantation for locally advanced intrahepatic cholangiocarcinoma treated with neoadjuvant therapy: a prospective caseseries. Lancet Gastroenterol Hepatol 2018; 3: 337-348 [PMID: 29548617 DOI: 10.1016/S2468-1253(18)30045-1]
- 109 Sapisochin G, de Lope CR, Gastaca M, de Urbina JO, López-Andujar R, Palacios F, Ramos E, Fabregat J, Castroagudín JF, Varo E, Pons JA, Parrilla P, González-Diéguez ML, Rodriguez M, Otero A, Vazquez MA, Zozaya G, Herrero JI, Antolin GS, Perez B, Ciria R, Rufian S, Fundora Y, Ferron JA, Guiberteau A, Blanco G, Varona MA, Barrera MA, Suarez MA, Santoyo J, Bruix J, Charco R. Intrahepatic cholangiocarcinoma or mixed hepatocellular-cholangiocarcinoma in patients undergoing liver transplantation: a Spanish matched cohort multicenter study. Ann Surg 2014; 259: 944-952 [PMID: 24441817 DOI: 10.1097/SLA.00000000000494]
- 110 Ethun CG, Lopez-Aguiar AG, Anderson DJ, Adams AB, Fields RC, Doyle MB, Chapman WC, Krasnick BA, Weber SM, Mezrich JD, Salem A, Pawlik TM, Poultsides G, Tran TB, Idrees K, Isom CA, Martin RCG, Scoggins CR, Shen P, Mogal HD, Schmidt C, Beal E, Hatzaras I, Shenoy R, Cardona K, Maithel SK. Transplantation Versus Resection for Hilar Cholangiocarcinoma: An Argument for Shifting Treatment Paradigms for Resectable Disease. Ann Surg 2018; 267: 797-805 [PMID: 29064885 DOI: 10.1097/SLA.00000000002574]
- 111 Facciuto ME, Singh MK, Lubezky N, Selim MA, Robinson D, Kim-Schluger L, Florman S, Ward SC, Thung SN, Fiel M, Schiano TD. Tumors with intrahepatic bile duct differentiation in cirrhosis: implications on outcomes after liver transplantation. Transplantation 2015; 99: 151-157 [PMID: 25029385 DOI: 10.1097/TP.000000000000286]
- 112 Sapisochin G, Facciuto M, Rubbia-Brandt L, Marti J, Mehta N, Yao FY, Vibert E, Cherqui D, Grant DR, Hernandez-Alejandro R, Dale CH, Cucchetti A, Pinna A, Hwang S, Lee SG, Agopian VG, Busuttil RW, Rizvi S, Heimbach JK, Montenovo M, Reyes J, Cesaretti M, Soubrane O, Reichman T, Seal J, Kim PT, Klintmalm G, Sposito C, Mazzaferro V, Dutkowski P, Clavien PA, Toso C, Majno P, Kneteman N, Saunders C, Bruix J; iCCA International Consortium. Liver transplantation for "very early" intrahepatic cholangiocarcinoma: International retrospective study supporting a prospective assessment. Hepatology 2016; 64: 1178-1188 [PMID: 27481548 DOI: 10.1002/hep.28744]
- 113 Moris D, Kostakis ID, Machairas N, Prodromidou A, Tsilimigras DI, Ravindra KV, Sudan DL, Knechtle SJ, Barbas AS, Comparison between liver transplantation and resection for hilar cholangiocarcinoma: A systematic review and meta-analysis. PLoS One 2019; 14: e0220527 [PMID: 31365594 DOI: 10.1371/journal.pone.0220527]
- 114 Baltatzis M, Jegatheeswaran S, Siriwardena AK. Neoadjuvant chemoradiotherapy before resection of perihilar cholangiocarcinoma: A systematic review. Hepatobiliary Pancreat Dis Int 2020; 19: 103-108 [PMID: 32147487 DOI: 10.1016/j.hbpd.2020.02.007]



C D WŰ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 1956-1980

DOI: 10.4251/wjgo.v13.i12.1956

ISSN 1948-5204 (online)

REVIEW

Solid extraintestinal malignancies in patients with inflammatory bowel disease

Anastasia Mala, Kalliopi Foteinogiannopoulou, Ioannis E Koutroubakis

ORCID number: Anastasia Mala 0000-0003-4289-7840; Kalliopi Foteinogiannopoulou 0000-0003-0554-5256; Ioannis E Koutroubakis 0000-0002-2773-2709.

Author contributions: Mala A, Foteinogiannopoulou K and Koutroubakis IE wrote the paper.

Conflict-of-interest statement: Mala

A and Foteinogiannopoulou K have no conflict of interest. Koutroubakis IE is an Advisory board member for Abbvie, Astellas, Genesis, Janssen, MSD, Pharmacosmos, Pfizer, Shire and Takeda; Speaker for AbbVie, Astellas, Genesis, Janssen, MSD, Takeda and Mylan; research support Abbvie and Ferrin.

Country/Territory of origin: Greece

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was

Anastasia Mala, Department of Medical Oncology, University Hospital of Heraklion, Heraklion 71110, Crete, Greece

Kalliopi Foteinogiannopoulou, loannis E Koutroubakis, Department of Gastroenterology, University Hospital of Heraklion, Heraklion 71110, Crete, Greece

Corresponding author: Ioannis E Koutroubakis, MD, PhD, Professor, Department of Gastroenterology, University Hospital of Heraklion, Stavrakia-Voutes, PO BOX:1352, Heraklion 71110, Crete, Greece. ikoutroubakis@gmail.com

Abstract

Malignancies constitute the second cause of death in patients with inflammatory bowel diseases (IBD), after cardiovascular diseases. Although it has been postulated that IBD patients are at greater risk of colorectal cancer compared to the general population, lately there has been evidence supporting that this risk is diminishing over time as a result of better surveillance, while the incidence of extraintestinal cancers (EICs) is increasing. This could be attributed either to systemic inflammation caused by IBD or to long-lasting immunosuppression due to IBD treatments. It seems that the overall risk of EICs is higher for Crohn's disease patients and it is mainly driven by skin cancers, and liver-biliary cancers in patients with IBD and primary sclerosing cholangitis. The aims of this review were first to evaluate the prevalence, characteristics, and risk factors of EICs in patients with IBD and second to raise awareness regarding a proper surveillance program resulting in early diagnosis, better prognosis and survival, especially in the era of new IBD treatments that are on the way.

Key Words: Extraintestinal malignancies; Crohn's disease; Ulcerative colitis; Thiopurines; Anti-tumor necrosis factor

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Patients with inflammatory bowel disease (IBD) are at risk of malignancies. The incidence of colorectal cancer is decreasing over time as a result of screening surveillance and better endoscopic techniques. Although IBD patients are not at high risk of overall extraintestinal malignancies compared to the general population, they



selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: April 9, 2021 Peer-review started: April 9, 2021 First decision: June 24, 2021 Revised: July 6, 2021 Accepted: August 13, 2021 Article in press: August 13, 2021 Published online: December 15, 2021

P-Reviewer: Sassaki LY S-Editor: Gao CC L-Editor: Webster JR P-Editor: Yu HG



have an increased risk of certain cancers. This should be kept in mind in the management of IBD patients especially in the context of immunosuppressive treatment that constitutes the cornerstone of treatment. Guidelines should incorporate all the preventive measures and be applied to everyday clinical practice for early diagnosis and better prognosis.

Citation: Mala A, Foteinogiannopoulou K, Koutroubakis IE. Solid extraintestinal malignancies in patients with inflammatory bowel disease. World J Gastrointest Oncol 2021; 13(12): 1956-1980

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/1956.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.1956

INTRODUCTION

Inflammatory bowel diseases (IBD), encompassing Crohn's disease (CD) and ulcerative colitis (UC), are chronic diseases of unknown etiology. It has been proposed that in genetically predisposed patients, several environmental factors and altered gut microbiota interact, resulting in immune system dysregulation and finally to chronic intestinal inflammation[1]. IBD mainly affects the gastrointestinal tract but not exclusively, as 50% of patients experience at least one extraintestinal manifestation. In Western countries, IBD is not uncommon as the prevalence of CD is 246.7/100000 and UC is 286.3/10000[2].

Cancer is the second cause of death in IBD patients, after cardiovascular diseases[3] and 30% of IBD patients are diagnosed with malignancy at some point during the course of IBD[4]. A multicenter European study showed that the prevalence of cancer in IBD patients, during a 15-year follow-up period, was 9.1%, the majority of them were extraintestinal cancers (EICs) and the overall cancer frequency was equal to the background population^[5].

However, it was recognized that patients with IBD are at increased risk of certain EICs and was also shown in other reports[6,7].

The risk of colorectal cancer (CRC) in IBD patients is 1.7-fold higher than that in the general population[3]. Lately, a reduction in the incidence of CRC in IBD has been reported that could be attributed not only to successful inflammation control due to new IBD treatments, but also to better surveillance strategies, colonoscopy techniques and implementation of guidelines supporting colectomy for high-grade dysplasia[3,4, 8]. On the other hand, there is evidence that IBD patients have a higher risk of certain EICs^[9-11]. EICs in IBD patients could be related either to chronic inflammation or to longstanding immunosuppressive treatment. As the incidence of EICs is increasing among IBD patients, awareness should be raised for more scrupulous surveillance for early diagnosis and curative treatment.

The aim of this review was to evaluate the prevalence, characteristics, and risk factors of EICs in patients with IBD, and to provide possible measures for prevention and early diagnosis.

METHODOLOGY

A review of the literature in PubMed using the MESH terms Inflammatory Bowel Disease AND neoplasms NOT colorectal cancer NOT intestinal neoplasms was conducted. Only studies in English language involving humans were included.

HEAD AND NECK CANCERS

Head and neck cancers (HNCs) include mainly squamous cancers of the oral cavity (OSCC), oropharynx (OPSCC), nasopharynx, hypopharynx and larynx (LSCC). Approximately 75% of HNCs are caused by alcohol and tobacco use and the remaining are mainly human papilloma virus (HPV) 16 related (HPV + HNCs)[12-17].

Only a few studies on HNCs and IBD have been published [18-21]. Danish cohorts showed no increased risk for OSCC and OPSCC[22,23], while a recent US retrospective cohort by Katsanos et al[18] showed for the first time an increased risk for OSCC [standardized incidence ratio (SIR) 9.77 (95% confidence interval (CI): 5.14-16.98)] in IBD patients. The risk was more pronounced in females (12-fold increase) compared to males (8-fold increase) and for tongue cancer (22-fold and 17-fold increase in females and males, respectively). A population-based cohort study by Mosher et al[24] also showed an increased 20-year risk of OSCC and OPSCC [20 yr relative risk (RR) 2.00 (0.95-4.22)] but not of LSCC [20 yr RR 0.48 (0.07-3.43)]. Indeed, a retrospective casecontrol Dutch study showed that IBD patients had a higher rate of HPV + HNCs (52.2%) compared to the general population and an impaired OSCC survival (P = 0.018), not related to immunosuppression[19]. Older age and UC diagnosis were risk factors for OSCC and OPSCC. A recent Dutch study showed that older age at IBD diagnosis (P < 0.001) as well as male sex (P < 0.001) were risk factors for LSCC in UC patients. On the other hand, tobacco use (P < 0.001), structuring (P = 0.006) and penetrating (P = 0.008) disease were identified as risk factors in CD patients who developed LSCC, while immunosuppressive medication did not influence survival [20]. Currently, studies on the prevalence of HNCs, risk factors and outcome in IBD patients are limited. Education on modifiable risk factors, such as smoking cessation, and safe sexual behavior, is the cornerstone of prevention of HNCs. In addition, given that IBD patients may have reduced immunosurveillance resulting in persisting HPV infections and cancer, oral screening and prophylactic HPV vaccines could be considered.

THYROID CANCER

Thyroid cancer (TC) is the most common endocrine cancer, and is more common in Caucasians, and young adults, predominantly females. Thyroid dysfunction, radiation exposure at a young age and heredity are known risk factors. Over 80% of all cases are of the papillary type^[25].

A recent prospective multicenter Italian study showed that the prevalence of TC in IBD patients was 5.2% [26]. Several studies, have shown that TC risk was not significantly higher in IBD patients compared with the healthy population [27-29]. On the other hand, two studies reported an increased risk of TC in UC patients, the first in both sexes and the latter only in males[30,31]. A recent meta-analysis of 8 case-control studies with 334015 patients, confirmed that patients with UC had an increased risk of TC, while patients with CD did not, independent of sex and race[32]. On the contrary, a case-control study in Ohio, United States with 289935 IBD patients[33] and an Italian cohort study of 3664 IBD patients[9], reported a higher risk for TC only in CD patients [odds ratio (OR) 2.3 (1.06-5.1), P = 0.034 and SIR 5.58 (95% CI: 2.41-11.00), respectively].

Regarding the risk of immunosuppressive treatment for the development of TC in IBD, data are conflicting, with two studies reporting an increased risk in patients treated with immunosuppressants[29,31] and another two studies did not observe this relationship[28,32].

Most of the evidence is based on population-based studies[9,30-32] with many potential confounding factors, and limited studies are focused especially on the risk of TC in IBD patients[29,32,33]. There is not enough evidence to show that IBD is an independent risk factor for TC, but IBD might provide an appropriate inflammatory environment and promote development of the cancer. Further well-adjusted and population-based studies should be conducted to confirm this speculation, in order that screening strategies for TC can be recommended.

LUNG CANCER

Lung cancer is one of the most common cancers and the major cause of mortality worldwide. Several population-based studies[10,34-36] and a recent meta-analysis by Lo in 2020 showed an increased risk [incidence rate ratio (IRR), 1.53 (95%CI: 1.23-1.91), P = 0.00[37] of lung cancer in CD patients. Masala *et al*[38] also reported increased mortality only in CD patients [SMR 4.00 (95% CI: 1.60-8.24)]. On the contrary, in UC patients no significant risk was found in any study [31,34,36,37,39] and actually a significantly lower risk of lung cancers was reported in two studies[29,40]. The difference between CD and UC could be explained by the different association of the two diseases with smoking. In conclusion, further studies focused on lung cancer are



needed and efforts should be made to guide patients to quit smoking, an important risk factor for both lung cancer and CD.

CHOLANGIOCARCINOMA

Cholangiocarcinoma (CCA) is a relatively rare EIC that mainly affects IBD patients in the context of primary sclerosing cholangitis (PSC)[41]. PSC is a chronic inflammatory, immune mediated, liver disease causing fibrosis of the intrahepatic and extrahepatic bile ducts finally leading to strictures[42]. The mechanisms of carcinogenesis in PSC are not well understood but it seems that CCA is the result of DNA damage caused by chronic biliary inflammation and bile acids in IBD patients with altered DNA repair ability[39].

Several studies have shown that IBD patients have an increased risk of biliary cancer (Table 1). A meta-analysis that included 17052 IBD patients showed that CD patients had a borderline significant increased risk of liver-biliary cancer (SIR 2.47, 95%CI: 0.95-6.46), while UC patients are at significantly increased risk (SIR 2.58, 95%CI: 1.58-4.22) and this is attributed to the occurrence of PSC in patients with IBD [10]. The incidence of CCA in PSC patients ranges between 0.5-1.5/100 person-years [42,45,46]. The risk of CCA is increased by 160-fold and PSC patients have a cumulative life-time risk of 5%-10% [44,47]. Approximately 30%-50% of CCAs are diagnosed within the first year of PSC diagnosis[45,46,48]. In another meta-analysis, it was shown that IBD patients were at increased risk of CCA (RR 2.63; 95% CI: 1.47-4.72, CD 2.69, 95% CI: 1.59-4.55 and UC 3.40, 95% CI: 2.50-4.62). In addition, further analyses concerning the site of CCA revealed that IBD patients are notably at increased risk of intrahepatic (RR 2.61, 95% CI: 1.72-3.95) and to a lesser extent extrahepatic CCA (RR 1.47, 95%CI: 1.10-1.97)[49].

As for the risk factors of developing CCA in patients with PSC, those with prolonged IBD duration and those who underwent colectomy due to CRC or colonic dysplasia have an increased risk of CCA [50-52]. On the other hand, in a Scandinavian cohort, although IBD duration was associated with increased risk of CCA, colectomy and CRC were not [48]. CCA develops in chronic inflammation in patients with PSC-IBD and particularly in those with dominant biliary stenosis, something that seems to be a predisposing factor^[41]. The definition of a dominant stenosis (DS) in PSC patients is a stricture less than 1.5 mm in diameter in the common bile duct or less than 1 mm in the left or right main hepatic duct^[53]. Approximately 10%-62% of PSC patients develop a DS at some point during their disease course [54,55]. In a 25-year study of 128 PSC patients in the United Kingdom, the mean survival of the patients with DS was worst (13.7 years) than for those without a DS (23 years) and this difference was related to a 26% risk of CCA, which developed only in patients with DS. In 50% of patients with CCA, the diagnosis of CCA was made within 4 mo of the diagnosis of PSC[54]. A German study of 171 PSC patients, who were followed prospectively for 20 years, confirmed that the presence of DS in PSC is associated with a worse prognosis due to increased risk of CCA and CRC. Furthermore, this risk was directly related to the presence of underlying IBD. In total, 97 patients had DS, 20 at entry and 77 developed DS over the follow-up period. In patients with DS without IBD, no CCA developed and the survival free of transplantation was 77.8% at 18 years. On the contrary, the 18-year survival was only 23% in the PSC with a DS and IBD. On the other hand, the presence of IBD had no impact on survival in those without a DS[56, 57]. The finding that the risk of CCA is related to the presence of IBD was not confirmed in a cohort of 241 Dutch PSC patients followed for a mean of 6 years[58]. As far as small duct PSC is concerned, it has not been associated with an increased risk of both malignancies (CRC, CCA)[59].

CCA is considered to result in a poor prognosis and the surveillance strategy has not been proven to be beneficial. However, it has been suggested that patients with IBD and PSC should undergo magnetic resonance cholangiopancreatography (MRCP) annually, serum CA 19-9 testing periodically and annual colonoscopy as they also have a higher risk of CRC.

HEPATOCELLULAR CARCINOMA

Hepatocellular carcinoma (HCC) mostly occurs in IBD on the background of PSC liver cirrhosis[60] or other established chronic liver diseases such as viral hepatitis. The annual incidence of HCC in cirrhotic patients is 5%, and justifies the 6-mo surveillance



Table 1 Studies on cholangiocarcinoma in patients with inflammatory bowel disease

Ref.	Country	Type of study	Patients	Follow up time	Results	Limitations
Jussila <i>et al</i> [<mark>30</mark>], 2013	Finland	Population- based study	21964 IBD <i>vs</i> general population	Mean 10.8 yr	Biliary Ca UC (SIR 7.26, 95%CI: 4.37-11.1); CD (SIR 4.93, 95%CI: 1.02-14.4)	Patients diagnosed in 1987-1993 and 2000-2007 were only included
Kappelman <i>et al</i> [35], 2014	Denmark	Population- based cohort	13756 CD, 35152 UC <i>vs</i> general population	CD 7.6 yr, UC 7.8 yr	CD: GBC-biliary Ca (SIR 2.4, 95% CI: 1.1- 4.5); UC: liver Ca (SIR 1.6, 95% CI: 1.1-2.2) GBC (SIR 2.5, 95% CI: 1.8-3.5), IBD-PSC: Liver Ca 80.0 (95% CI: 32.1-164.8), GBC 129.1 (95% CI: 47.4-281.5), IBD-non-PSC: Liver Ca 1.3 (95% CI: 0.9-1.9), GBC 2.1 (1.4- 3.0)	Exclusion of patients with very mild disease, no inpatient encounters prior to 1995
Ananthakrishnan <i>et al</i> [<mark>43</mark>], 2014	United States	Multi- institutional IBD cohort of IBD	5506 CD 5522 UC 224 IBD-PSC	NA	CCA in IBD-PSC patient (OR 55.31, 95%CI: 22.20-137.80) compared to IBD non-PSC patients	PSC diagnosis was predicted with a model. Did not separately examine the risk of large- duct over small-duct PSC
Bernstein <i>et al</i> [<mark>39</mark>], 2001	Canada	Population- based study	2857 CD and 2672 UC patients <i>vs</i> with non-IBD (1:10)	14 yr	Liver and biliary Ca: CD (IRR 5.22; 95%CI: 0.96-28.5, <i>P</i> = 0.06), UC (IRR 3.96; 95%CI: 1.05-14.9)	Low percentage of the patients received IMMs
Sørensen <i>et al</i> [25], 2018	Denmark	Population- based study	222 PSC-IBD patients <i>vs</i> 8.231 IBD controls	PSC-IBD 7.4 yr, non- PSC IBD 8.4 yr	CCA PSC-IBD (HR; 190; 95%CI: 54.8-660)	Small number of PSC-IBD patients. Small duct PSC (8%) were included
Scharl <i>et al</i> [7] , 2019		Cohort study (IBD-Ca <i>vs</i> IBD-no Ca)	3119 IBD patients	5 yr	IBD Biliary Ca (SIR 6.3, 95%CI: 1.27-18.41)	Did not compute multivariate regression for Ca subtypes, IBD phenotypes

IBD: Inflammatory bowel disease; Ca: Cancer; UC: Ulcerative colitis; CD: Crohn's disease; SIR: Standardized incidence ratio; CI: Confidence interval; GBC: Gallbladder cancer; PSC: Primary sclerosing cholangitis; CCA: Cholangiocarcinoma; IRR: Incidence rate ratio; HR: Hazard ratio; IMMs: Immunomodulators.

with liver ultrasound \pm alpha-fetoprotein. Nevertheless, there have been sporadic case reports of HCC in non-cirrhotic IBD patients, mostly in CD patients treated with azathioprine[61]. Furthermore, it is of note that HCC develops more frequently in PSC-IBD patients compared to IBD non-PSC [hazard ratio (HR), 21.00][62]. However, most population-based studies address the risk of liver cancer without separating the histologic type (HCC or CCA) under the term liver or liver-biliary cancers.

GALLBLADDER CANCER

The risk of gallbladder cancer (GBC) mostly affects IBD patients in the context of PSC. Many studies have confirmed that PSC patients are at increased risk of GBC[62-70], and the prevalence is 3%-14% *vs* 0.35% in the general population[63]. The 10-year cumulative risk was 3% (95%CI: 1-7) in PSC-IBD compared to 0% in non-PSC IBD patients[45]. Furthermore, a study that evaluated 72 gallbladders from 100 liver explants-o (OLT) due to PSC showed that GBC was associated with intrahepatic bile duct dysplasia (P = 0.001), CCA (P = 0.023), and IBD (P = 0.03) and gallbladder dysplasia was associated with hilar/intrahepatic bile duct dysplasia (P = 0.0014), and older age at OLT (P = 0.007)[71].

A Danish study showed that CD patients had a significant increased risk of liver-GBC and UC patients had a more notable risk (SIR 2.5, 95%CI: 1.8-3.5) compared to the general population. Furthermore, this was profound for UC-PSC (SIR 129.1; 95%CI: 47.4-281.5) than in those without PSC (SIR 2.1; 95%CI: 1.4-3.0)[35]. This indicates the need for annual surveillance with ultrasound of the gallbladder in PSC-IBD patients.

Zaisbidena® WJGO | https://www.wjgnet.com

PANCREATIC CANCER

Pancreatic cancer (PC) is the seventh leading cause of cancer death worldwide with increasing incidence and mortality, and is considered to have an unfavorable outcome and prognosis.

In a recent large Scandinavian study with 161926 IBD patients, 442 (0.27%) were diagnosed with PC compared with 3386 (0.21%) of the 1599024 IBD-free individuals. The 20-year cumulative incidence was 0.34% (95%CI: 0.30-0.38) vs 0.29% (95%CI: 0.28-0.30), while the IR was 22.1 (20.1-24.2)/100000 person-years in IBD patients. The overall HR was 1.43 (1.30-1.58) [CD 1.44 (1.18-1.74), UC 1.35 (1.19-1.53), IBD unclassified was 1.99 (1.50-2.64)], whereas in IBD-PSC it was 7.55 (4.94-11.5)[72]. In a Korean study, the risk was only increased in females with CD (SIR 8.58; CI: 1.04-31.00)[30]. On the contrary, a previous meta-analysis and several population-based cohort studies have shown that the risk of PC is non-significantly increased in IBD patients [pooled SIR 0.51 (0.06-4.57) and 0.75 (0.30-1.87) accordingly][7-10]. IBD-PSC patients should be referred separately since they are at a higher risk of PC[62]. A Swedish study, which included PSC patients (79% had concurrent IBD), showed that they had a 14-fold greater risk of PC than the background population [46]. In another study of 224 PSC-IBD patients, a significant increased risk was confirmed (OR 11.22, 95% CI: 4.11-30.62) compared to IBD non-PSC patients[43]. Given that higher CCA risk in PSC is well established, we could assume that the increased PC risk might be due to misdiagnosis of periampullary cancer.

GASTRIC MALIGNANCIES

The cause of gastric cancer (GC) in IBD patients is uncertain. GC usually develops on the background of intestinal metaplasia and dysplasia due to Helicobacter pylori (H. *pylori*) infection and chronic inflammation^[73]. Nevertheless, the prevalence of *H*. *pylori* in IBD patients is low[74], while a possible causative relation could be attributed to the upper gastrointestinal involvement of CD. A previous review and meta-analysis study, found that CD patients had a significantly increased risk of cancer of the upper gastrointestinal tract (SIR 2.87, 95%CI: 1.66-4.96) and of the stomach per se (SIR 2.05, 95%CI: 1.06-3.97)[10]. However, most recent population-based studies[7,11,30,35,39,75] did not confirm this finding. With regard to the risk factors and survival of IBD patients with GC, a study that included 59 GC cases in IBD individuals, showed that UC was more frequent among these patients (69.5% vs 51.4%; P < 0.01) compared with 177 IBD controls, and IBD patients with GC showed reduced survival (P = 0.035; HR 1.385, CI: 1.023-1.875) compared with 1534 individuals of the general population with the same malignancy [76]. Studies on GC in IBD patients are limited and no secure conclusions can be drawn.

BREAST CANCER

Breast cancer (BC) is the second most common malignancy diagnosed worldwide and its mortality is decreasing over time due to screening protocols and improved therapy. Increasing age, prolonged estrogen exposure, obesity in postmenopausal women, reproductive and genetic factors, western life-style, smoking and alcohol are known risk factors^[77].

Limited data are available on the pathogenesis of BC in IBD[78]. IBD patients seem to have a shorter period of estrogen exposure than the general population, with later onset of menarche and earlier menopause, that could be related to a lower BC risk 78, 791

No studies primarily investigating the risk of BC in IBD are available. Data, generally not adjusted for confounding factors, can be derived mainly from population-based cohort studies investigating the general and type-specific risk of malignancies[6,15,30,36,39,75,80-88]. In 2012, Hemminki et al[84] showed a decreased risk for BC only for CD patients (SIR 0.85, 95%CI: 0.75-0.97) and a decreased mortality rate (HR 0.75, 95% CI: 0.58-0.98), and similar results were seen in the Dutch IBD-SL cohort (*n* = 1157; SIR 0.11 (95%CI: 0.00-0.64)[15] and the TREAT registry cohort study [n = 2975; SIR 0.28 (95%CI: 0.08-0.72)][86]. On the other hand, a Swedish cohort study conducted in 21788 CD patients showed a 30% higher BC risk in hospitalized patients between 45 and 64 years, possibly reflecting the advanced risk at older age or the exposure to IBD treatment or unregistered confounders[83]. Similarly, a retrospective



Taiwanese cohort study by Tsai in 2014 indicated that IBD is not associated with increased BC risk (aHR 0.95 (95%CI: 0.66-1.36); however, there was an association between the frequency of IBD-related hospitalizations and BC risk in patients less than 65 years old (aHR 8.45; 95% CI: 4.64-15.4) [87]. An increased risk of BC in both UC and CD was reported by the IBSEN study in 2016[88]. A large cohort study[36], a Finish register study[30], and two meta-analyses, by Pedersen *et al*[10] and by Lo *et al*[37], showed no difference in the occurrence of BC in both UC, CD and the general population, but data on treatment modalities were lacking.

No association between the use of thiopurines and BC has been reported by cohort studies and small series [22,75,89-93]. The correlation between BC and the use of biologics in IBD patients has been addressed in several reports[94-98]. A large nationwide Danish register-based cohort study with 56146 IBD patients and a long followup, showed no increased BC risk for patients treated with infliximab, adalimumab or certolizumab[94]. In the TREAT Registry cohort study consisting of CD patients, a decrease in the occurrence of BC was observed in both patients exposed and nonexposed to infliximab [SIR 0.50 (95%CI: 0.24-0.92) and SIR 0.32 (95%CI: 0.12-0.70), respectively], compared to the SEER database of the general United States population [86]. No association for vedolizumab was also reported in a study involving 2830 IBD patients with a follow-up period up to 5 years[95]. In addition, no increased risk of BC in IBD patients exposed to adalimumab monotherapy or combination therapy with thiopurine or methotrexate was suggested in a pooled analysis of 1594 CD patients [96]. Similar results were shown in a pooled analysis of 5 small studies and 5 landmark trials comprising 2385 IBD patients[99] and by a meta-analysis, involving 22 randomized controlled trials comparing anti-tumor necrosis factor (TNF) therapy vs placebo, even if only 4 trials had a low risk of bias[100].

In conclusion, the risk of BC is probably decreased in CD patients and seems to be similar to the background population in UC patients. IBD and anti-TNF therapy do not seem to affect BC incidence; however, for thiopurines and combination therapy, this is still uncertain. Future studies on the pathophysiological association between BC and IBD may help identify populations at high risk for BC, who might therefore benefit from close surveillance.

CERVICAL NEOPLASIA

Cervical neoplasia (CN) consists of dysplasia or cervical intra-epithelial neoplasia (CIN) and invasive cervical cancer (CC)[101]. High-risk HPV infection (types 16, 18, 45, 31) is considered the causal agent of CN, whereas smoking, sexual, socioeconomic, and immunologic co-factors may contribute to persistent infection[102]. Most low-grade lesions regress spontaneously but most high-grade dysplasia and CIN 2/3 do not[103, 104]. While HPV vaccines and Papanicolaou (Pap) smear have substantially reduced CC incidence and mortality, these are still high in developing countries and CC remains the second most common cancer in women worldwide[105-107].

The risk of CN in patients with IBD remains controversial and the role of immunosuppressants, a known risk factor for cervical dysplasia in other immunemediated diseases, is not clear [108,109]. It may be hypothesized that the underlying immunologic changes in IBD or immunosuppressive drugs may reactivate HPV from a latent status or decrease HPV clearance and CIN regression or make HPV vaccination less effective.

Data on the association between IBD, treatment and CN are conflicting[31,75,91,110-119] (Table 2). Also, non-conclusive are the results of two meta-analyses. Allegretti in 2015, in a meta-analysis of 8 studies with 77116 IBD patients but also heterogeneity, found that IBD patients had an increased risk of high-grade dysplasia/cancer compared to healthy controls (OR 1.34, 95% CI: 1.23-1.46) and the risk was greater with the use of immunomodulators, corticosteroids and 5-aminosalicylic acids, but not with anti-TNF[120]. A recent meta-analysis by Lo et al[37] in 2020 reported no statistically increased risk for CC in IBD patients.

Regarding screening adherence some studies indicated that this is influenced by various factors such as the state system, increased age and use of immunosuppressants and there is no significant difference with non-IBD patients[121-123]. Singh showed that women with IBD in Canada had a low (54%) screening adherence and older age, lower socioeconomic status, lower intensity of healthcare utilization, CD and exposure to immunosuppressant medications were independent predictors of lower use of Pap testing[121]. In the United States, Long et al[122] and Xu et al[123] showed higher adherence (70%).



Table 2 Studies on cervical neoplasia in patients with inflammatory bowel disease						
Ref.	Country	Type of study	Patients	Results	Limitations	
Connell <i>et</i> <i>al</i> [91], 1994	United Kingdom	Single centre- registry study, Prospective (1962-1991)	755 IBD patients taking AZA	CC: SIR 4.00	Small power of the study to detect an increased risk (expected 0.5). No variables analysed	
Bhatia <i>et al</i> [<mark>110</mark>], 2006	United States	Single centre- cohort study, Retrospective	116 IBD patients 116 age- matched healthy controls	Abnormal Pap smears: $18\% vs 5\%$; $P = 0.004$. Non association with IBD type or treatment exposure	Inter-observer variability in the smear interpretation. Recall bias (questionnaires)	
Kane <i>et al</i> [111], 2008	United States	Single centre- cohort study (2004-2005)	8 UC; 32 CD. 120 controls (age, race, parity-matched). 58% exposed to ≥ 1 treatment (prednisone, AZA, 6-MP and/or IFX)	Abnormal Pap smear 42.5% IBD vs 7% controls; OR 3.4 (95%CI: 1.7-12.1, ${}^{a}P < 0.001$), low-grade lesions OR 2.2 (95%CI: 1.7-4.4, ${}^{a}P < 0.001$), high-grade lesions OR 3.1 (95%CI: 1.3-8.7, ${}^{a}P < 0.001$). Exposed vs non-exposed: Abnormal Pap smear OR 1.5 (95%CI: 1.2-7.1, ${}^{b}P = 0.02$), High-grade lesions OR 6.5 (95%CI: 1.43-30.1, ${}^{b}P < 0.05$), IMMs > 6 mo OR 1.9 (1.1-12.1, ${}^{b}P < 0.001$)	Small population study. Level of immune suppression not assessed	
Hutfless <i>et</i> <i>al</i> [112], 2008	United States	Nested case- control study (1996-2006)	UC 778; CD 476; Controls 12124. TP monotherapy (AZA, 6-MP, MTX)	CC: aOR 1.45 (95%CI: 0.74-2.84), ASA: OR 1.65 (0.34-7.98), corticosteroids: OR 2.79 (0.71-11.00), IMMs: OR 3.45 (0.82-14.45)	Missing data for race, ethnicity, smoking status. No adjustment for therapy or disease severity. Diagnostic or screening Pap smears are not distinguishable	
Marehbian <i>et al</i> [<mark>113</mark>], 2009	United States	Insurance claims-based United States population (2002-2005)	CD 22310; controls 111550	IBD vs controls: Cervical dysplasia/HPV: RR 1.35 (1.28, 1.43). Exposed vs non exposed to treatment: aHR Steroids 1.11 (0.59, 2.09), IS 1.77 (1.26, 2.49), anti-TNF 1.30 (0.68, 2.51), Combination 1.81 (1.10, 3.10)	Disease severity is a possible confounding factor. Limited amount of person-time on various treatments	
Lees <i>et al</i> [114], 2009	Scotland	Tertiary centre, case-control study Retrospective	UC 178; CD 184; Healthy controls 1448	Abnormal Pap smear: IBD patients OR 0.82 (95%CI: 0.59-1.1.3), IBD patients current smokers vs ex or no smokers OR 2.95 (95%CI: 1.55-5.50); $P = 0.001$, Women < 20 yr at IBD diagnosis vs 20-39 yr vs > 40 yr: 27% vs 13.4% vs 5.0% ($P = 0.001$). No effect of IS therapy	No data on HPV status or exposure to corticosteroids. Small number of patients on MTX or anti-TNF. Median time exposed to IS 2.4 yr. Smoking may be a confounding factor	
Singh <i>et al</i> [115], 2009	Canada	Population- nested case- control (2002- 2006)	UC 233; CD 292; Controls 57898	Cervical abnormalities: OR 1.41 (1.09-1.81), IS+ steroids 1.41 (1.09-1.81). High risk lesions: IS monotherapy aOR 1.23 (0.57- 2.63), IS + steroids aOR 1.28 (0.77-2.12), CD patients exposed to > 10 prescriptions of oral contraceptives OR, 1.66 (1.08-2.54), UC OR 1.03 (0.77-1.38)	Administrative databases. Possible bias is smoking, parity and sexual factors. Small number of patients exposed to IMMs alone. No data on HPV infection. CN confirmed histologically in 19% of cases	
Jess <i>et al</i> [75], 2013	Denmark	Population- nested case- control (2002- 2006)	1437 UC; 774 CD. UC median F.U. 15 yr (0-33); CD 14 yr (0- 33). 1978-2010 Population-based cohort study (1978-2010). 25176 IBD patients; UC 1437; CD 774(UC median 15yrs/22 582 pt- yrsCD median 14yrs/11 261 pt- yrs)TP (18% of UC pts,45% of CD pts everused TP)	Cervical dysplasia/CC: SIR 1.65 (95%CI: 1.10-2.37), CD diagnosed at age 0-19 yr: SIR 2.52 (95%CI: 1.26-4.51), Smokers: SIR, 2.15 (95%CI: 1.27-3.40), 5-ASA: SIR, 1.69 (95%CI: 1.08-2.51), Thiopurines: SIR, 2.47; (95%CI: 1.54-3.73)	No detailed pharmaco- epidemiological analyses. Cervical dysplasia was analysed along with CC resulting in higher CC incidence	
Rungoe <i>et</i> <i>al</i> [116], 2015	Denmark	Nationwide population- based cohort (1979-2011)	27408 IBD patients (UC 18691; CD 8717), controls 1508334, median F.U: UC. 7.8 yr; CD 8.3 yr	CD: CC IRR 1.53 (95%CI: 1.04-2.27), High grade lesions IRR, 1.28 (95%CI: 1.13-1.45), low grade lesions IRR, 1.26 (95%CI: 1.07- 1.48), CN significantly higher risk in CD patients diagnosed at young age and treated with AZA. UC: CC IRR 0.78 (0.53- 1.13), High-grade lesions IRR 1.12 (95%CI: 1.01-1.25), Low-grade lesions: IRR 1.15 (95%CI: 1.00-1.32)	Data for smoking not available. Possible confounding factor is disease severity. Vaccination policies and screening may influence risk estimation	
Kim <i>et al</i> [119], 2015	United States	Cohort U.S. insurance data (2001-2008 and 2003-2012)	133333 SID patients, including 25176 IBD	High grade dysplasia/CC: aHR 1.72 (0.66-4.45). IS: aHR 1.72 (95%CI: 0.66-4.45)	Confounding factors (race, ethnicity, socioeconomic status, sexual behavioural, gynaecologic history). Study not designed to determine the comparative effect of IS drugs. Short follow-up (mean 2.1 yr)	



Jung <i>et al</i> [<mark>31</mark>], 2017	Korea	National Health Insurance claims (2011-2014)	IBD (5595 CD and 10049 UC)	CC: UC 5.65 (2.44-11.13)	Did not focus on cancer occurred during IBD treatment. Missing data (disease diagnosis, phenotype). Short follow-up
Segal <i>et al</i> [117], 2021	United Kingdom	Hospital Episode Statistics database (1997- 2012)	837 with IBD, 61648 control patients	IBD <i>vs</i> controls: CC: 5.2 of 100000 <i>vs</i> 4.6 of 100000; <i>P</i> = 0.042	Other possible mechanism of carcinogenesis (other than HPV) not evaluated. Database accuracy. HPV- related cancers were not considered separately in CD and UC
Li <i>et al</i> [<mark>118</mark>], 2019	China	Prospective study (2014- 2017)	124 IBD patients and 372 controls	HPV 16/18 infection OR 29.035 (3.64- 210.988) <i>P</i> = 0.001, HPV-infection rate: MTX OR 4.76 (1.471-15.402) <i>P</i> = 0.005, > 2 IS OR 3.64 (1.255-10.562) <i>P</i> = 0.013, CIN prevalence: 3.2 vs 0.0%, <i>P</i> = 0.004	No data on sexual behaviour in control group

^aP: Compared to controls

^bP: Compared to non-exposed.

IBD: Inflammatory bowel disease; AZA: Azathioprine; Pap: Papanicolaou; CC: Cervical cancer; SIR: Standardized incidence ratio; IFX: Infliximab; 6-MP: 6-Mercaptopurine; UC: Ulcerative colitis; CD: Crohn's disease; OR: Odds ratio; aOR: Adjusted odds ratio; CI: Confidence interval; IS: Immunosuppressant; TP: Thiopurine; MTX: Methotrexate; ASA: Amino-salicylate; IMMs: Immunomodulators; anti-TNF: Anti-tumour necrosis factor; SID: Systemic inflammatory disease; HPV: Human papilloma virus; HR: Hazard ratio; IRR: Incidence rate ratio; CIN: Cervical intraepithelial neoplasia.

> In many health care systems, it is not clear who has the primary responsibility for CC prevention. As incidence and mortality of CN are highly dependent on screening and treatment of precursor lesions, an inter-specialist cooperation is suggested to guide IBD patients to follow all preventive measures such as HPV vaccination, between 9 and 26 years and prior to the initiation of the sexual activity, in both men and women, annual screening testing for chronically immunocompromised patients and smokers starting at age 21, safe sexual practices and smoking cessation[124].

OTHER GYNAECOLOGICAL CANCERS

No studies primarily investigating the incidence and mortality risk of endometrial, ovarian, or vulvar-vaginal carcinoma in IBD have been identified. Data extracted from large cohort studies investigating the general risk of malignancies, an older metaanalysis and a recent meta-analysis in 2020 showed no increased risk of these cancers even when immunosuppressive medication was used [10,29,31,34,35,37,39,40,80-86, 125-127].

URINARY TRACT CANCERS

The urinary tract cancers (UTC) include bladder cancer, that occurs at advanced age, predominantly in males, smokers or in association with chronic inflammation, and renal cell carcinoma (RCC), that is related to smoking, obesity, and hypertension[128-131]. Elevated levels of TNF, a key mediator of cancer-related inflammation, have been reported in the early stages of RCC due to loss of the Von Hippel-Lindau tumorsuppressor gene[132,133].

The prevalence of UTC in IBD patients in a recent, multicenter, prospective Italian study was 9.6% (6.3% in CD and 13% in UC) with main risk factors being the disease duration (in UC) and the use of immunomodulators[134,135]. There are some reports (Table 3) and a recent meta-analysis of 15 studies by Lo et al [37] of the non-association between IBD and UTC [Urological: CD IRR 1.34 (0.91-1.98) P = 0.14, UC IRR 1.01 (0.82-1.25) *P* = 0.92; RCC: CD IRR 1.93 (0.80-4.65) *P* = 0.14, UC IRR 0.84 (0.33-2.10) *P* = 0.71] [30,35,37,39,75]. However, a case-control study conducted in veteran patients in Texas found an increased risk of RCC in IBD patients^[24]. Young IBD patients with a severe disease course also had an increased risk of RCC in a large Dutch cohort study[136]. These patients had a better outcome compared to the general population, probably related to incidental RCC diagnosis at lower stage and younger age due to frequent abdominal imaging. Another key finding of this study was that the use of immunosuppressive or anti-TNFa therapy did not adversely affect RCC disease-free

Table 3 Studies on urinary tract cancer in patients with inflammatory bowel disease						
Ref.	Country	Type of study	Patients	Results	Limitations	
Bernstein <i>et al</i> [39], 2001	Canada	Population- based-cohort study (1984-1997)	5529 IBD patients; Median F.U. 7.9 yr (3.5- 12)	Bladder Ca: CD IRR 1.30 (0.51-3.30) UC IRR 0.67 (0.24-1.85) IBD IRR 0.92 (0.47- 1.82). RCC: CD IRR 1.02 (0.31-3.34) UC IRR O.8 (0.25-2.58) IBD IRR 0.89 (0.39-2.06)	Possible confounding factor is socioeconomic status. Maximum F.U 12 yr. Data not analysed by extent of disease	
Pasternak <i>et al</i> [22], 2013	Denmark	Cohort study (1997-2008)	45986 patients	AZA: Current users RR 2.8 (1.24-6.51), former users: RR 1.73 (0.70-4.24)	Possibly confounded by indication	
Jess et al[75], 2013	Denmark	Population- nested case- control (2002- 2006)	1437 UC; 774 CD; Median F.U. UC 15 yr (0-33); CD 14 yr (0-33)	CD SIR 1.69 (0.68-3.49); UC SIR 1.08 (0.56- 1.89)	No detailed pharmaco- epidemiological analyses	
Jussila <i>et al</i> [<mark>30]</mark> , 2013	Finland	Cohort study (1987-1993 and 2000-2007 and followed up to 2010)	21964 patients with IBD	Urological CD IRR 1.56 (0.58-4.21), RCC CD IRR 1.61 (0.62-4.17)	Possibility of misclassification of IBD, CD, UC, and Ca	
Kappelman <i>et al</i> [35], 2014	Denmark	Population- based-cohort study (1978-2010)	48908 IBD patients	Bladder Ca CD SIR 1.1 (0.8-1.6), RCC CD SIR 0.98 (0.77-1.23)	Not age-and sex specific estimates of absolute Ca risk. Detection bias. Data possible lacking. No inpatient encounters prior to 1995	
Algaba <i>et al</i> [<mark>85]</mark> , 2015	Spain	Prospective- cohort study (2005-2011)	590 IBD patients; Controls 222219	Bladder Ca RR 5.23 (1.95-13.87)	Small number of cases and limited period of follow-up	
Nyboe Andersen <i>et al</i> [94], 2014	Denmark	Cohort study (1999-2012)	56146 IBD patients; Median F.U. 9.3 yr (4.2- 14)	Anti-TNF: aHR 1.60 (0.61-4.19)	Confounding by indication, smoking, missing data. Short median F.U. of anti-TNF exposed (3.7 yr). Small number of Ca did not permit subgroup analysis	
Bourrier <i>et al</i> [138], 2016 (CESAME study)	France	Prospective- cohort study (2004-2005)	19486 IBD patients (30.1% receiving TP)	Bladder Ca SIR 1.20 (0.44-2.61); RCC SIR 2.05 (0.98-3.77), AZA > 65 yr v_5 < 50 yr HR 13.26 (3.52-50.03, P = 0.0001), Current users: SIR 3.40 (1.47-6.71; P = 0.006), Ex- users: SIR 0.64 (0.01-3.56)	Smoking is a possible confounding factor. Risk of anti-TNFs not assessed. Short follow-up	
Wauters <i>et al</i> [139], 2017	Belgium	Retrospective case-control study (1990-2014)	RCC; Exposed to anti- TNF: 2083 IBD patients (952 men and 1131 women); Un-exposed to anti-TNF: 1952 (977 men and 975 women)	Un-exposed to anti-TNF males SIR 5.4 (2.9- 9.2), females SIR 8.5 (3.7-16.8) Exposed to anti-TNF males SIR 7.1(2.3-16.5), females SIR 4.8 (0.6-17.3)	Potential confounding factors were not adjusted. Disease type, severity, and drug exposure of hospitalized patients may not be comparable with the global patient population. Different agents and dose-response for anti-TNF were not studied	
Mosher <i>et al</i> [<mark>24</mark>], 2018	United States	Case-control study Veteran population (1996- 2015)	2080 patients with IBD; 271898 without IBD	Bladder Ca 20 yr RR 1.72 (0.86-3.45); RCC 20 yr RR 2.90 (1.46-5.84)	Administrative data. Possible underestimation of Ca incidence (newly diagnosed Ca treated in other centres)	
Derikx <i>et al</i> [136], 2015	Holland	Case-control study (1991-2013)	Case control study A: 180 IBD patients with RCC vs 1800 IBD patients; Case control study B: 180 IBD patients with RCC vs 4388 patients with RCC in the general population	Case control A: Montreal E3 UC OR 1.8-2.5 (95% CI: 1.0-5.3), penetrating IBD-CD OR 2.8 (95% CI: 1.3-5.8), IBD related surgery OR 3.7-4.5 (95% CI: 1.6-8.2), male gender OR 3.2-5.0 (95% CI: 1.7-13.2). Case control B: lower age at diagnosis RCC ($P < 0.001$), lower N-stage ($P = 0.025$), lower M-stage ($P = 0.020$), more frequent surgical treatment for RCC ($P < 0.001$), better survival ($P = 0.026$; HR 0.7)	Retrospective data collection. Selection bias (different registries and databases)	
Biancone <i>et al</i> [26], 2020	Italy	Multi-centre nested case- control study prospective (2011-2017)	403 IBD patients; 806 IBD controls	UC: OR 3.79 (1.27-16.2)	Referral IBD centres included more severe patients compared with community-based centres	

IBD: Inflammatory bowel disease; CD: Crohn's disease; IRR: Incidence rate ratio; CI: Confidence interval; F.U. : Follow-up; Ca: Cancer; UC: Ulcerative colitis; RR: Relative risk; SIR: Standardized incidence ratio; UTC: Urinary tract cancer; RCC: Renal cell carcinoma; aHR: Adjusted hazard ratio; AZA: Azathioprine; TP: Thiopurines; anti-TNF; Anti-tumour necrosis factor; TNF-: Not exposed to anti-TNF; TNF+: Exposed to TNF; N-stage: Lymph node stage; M-stage: Metastatic stage.

Baisbideng® WJGO | https://www.wjgnet.com

survival and overall survival[136].

With regard to bladder cancer, an increased risk in IBD patients was clearly demonstrated by a meta-analysis of eight population-based cohort studies with 17052 IBD patients in 2010 (SIR 2.03, 1.14-3.63)[10] and a prospective Spanish cohort study [85]. Moreover, long-standing IBD and use of immunosuppressive medication were found to be associated with the development of bladder cancer[137]. It is noteworthy that a reversible increased risk of UTC in patients treated with azathioprine has been reported by two large studies[22,138].

Cancer risk assessment in patients under anti-TNF therapy is problematic due to frequent combined treatment with thiopurines. Anti-TNF monotherapy does not increase either UTC risk, according to the results of a population-based prospective cohort nationwide Danish study of more than 56000 IBD patients[94] or RCC risk, independent of diagnosis age, type, and duration of IBD or coexistence of known RCC risk factors[139].

There are currently no screening guidelines for UTC in any population, while incidental detection of RCC has been identified as a positive prognostic index[140, 141]. Whether patients with a long IBD duration may require different surveillance programs for preventing UTC needs to be confirmed, but all CD patients should be encouraged to quit smoking as tobacco could be a key risk factor[128,142,143]. Limited evidence is available on the role of immunosuppressive therapy in the development of UTC; however, elderly men on thiopurine therapy, should be closely evaluated for UTC.

PROSTATE CANCER

Prostate cancer (PC) is the second most common cancer in men worldwide, with screening tools available for early diagnosis[144]. The most important risk factors are age, African American ethnicity, genetic and possibly dietary factors.

Some studies have indicated an increased risk of PC in IBD, especially in UC, while others did not confirm this[24,29-31,35,36,39,40,75,83,84,144-147] (Table 4). Intensive surveillance in IBD patients with digital rectal and prostate examination may be a reason for the high incidence of PC, but also potential pathophysiological mechanisms of IBD and PC association have been recognized. In principle, the distinct microbiome level between UC and CD may be related to the elevated risk of PC in UC[148-150]. In addition, simultaneous elevation of IBD-related inflammatory markers, such as C-reactive protein, and of prostate-specific antigen (PSA), known prostatic inflammation or PC index has been reported[151-155]. Receptors of pro-inflammatory cytokines, such as interleukin 6 (IL-6)[156-158], and folate hydrolase 1 (FOLH1)/prostate specific membrane antigen (PSMA) are up-regulated in both IBD and PC and in fact FOLH1/PSMA appears overexpressed in cases of biochemical recurrence with an increase of PSA and metastatic disease[159-163].

A meta-analysis of 9 studies involving 17052 IBD patients by Pedersen et al[10] in 2010 showed no association between IBD and PC [IBD: SIR 1.16 (0.88-1.52), UC: SIR 1.14 (0.85-1.42) CD: SIR 0.77 (0.41-1.45)]. These results were not confirmed by recent studies and on the other hand, an increased risk of PC in patients with UC was reported (Table 4). Actually, an increased risk worldwide only for UC patients was shown in another meta-analysis of nine population-based studies by Ge et al[164] in 2020, with a large sample and a long follow-up of patients [Cohort studies: IBD RR 1.33 (1.03-1.71); UC RR 1.58 (1.08-2.30); CD RR 1.12 (0.97-1.31); Case control studies: RR 1.81 (1.43-2.29)]. In addition, a recent meta-analysis of eight studies by Chen et al[165] in 2020 reported two interesting results. For the first time a higher risk was also found for CD patients [IBD RR 1.78 (1.32-2.41); UC RR 1.76 (1.06-2.91); CD RR 1.29 (1.04-1.61)] and a higher risk in Asian patients with IBD compared to Caucasians was shown [RR 3.02 (95%CI: 2.03-4.48) and 1.6 (95%CI: 1.17-2.19), respectively], in contrast with the known lower risk of PC in the Asian general population[165]. It is noteworthy that two other meta-analyses in 2020, by Carli et al[166] [IBD RR 1.71 (1.16-2.51) P = 0.007; UC RR 1.21 (0.98-1.51) P = 0.07; CD RR 1.10 (0.98-1.25) P = 0.12] and by Lo et al[37] [UC SIR 1.08 (0.9-1.3); CD SIR 1.04 (0.8-1.35)] did not demonstrate a higher risk of PC in the subgroup analysis of UC or CD.

In conclusion, data for PC risk in IBD patients are conflicting. Further prospective studies are needed to evaluate the impact of concomitant medications on the risk of developing PC and the possible utility of screening programs for early diagnosis of PC in patients with IBD.

Zaishidenq® WJGO | https://www.wjgnet.com

Table 4 Studies on prostate cancer in patients with inflammatory bowel disease						
Ref.	Country	Type of study	Patients	Results	Limitations	
Karlén <i>et al</i> [<mark>40]</mark> , 1999	Sweden	Cohort (1955-1989)	1547 UC	UC 7/1547; SIR 0.7 (0.3-1.5)	Missing data. Closer monitoring of UC patients may lead to higher frequency and early detection	
Bernstein <i>et al</i> [39], 2001	Canada	Cohort (1984 -1997)	5529 IBD- 1151000 controls	IBD 26/5529 SIR 0.86 (0.59-1.26), 6293/1151000 controls	Possible confounding factor is socioeconomic status. Maximum F.U 14 yr. Data not analyzed by extend of disease	
Winther <i>et al</i> [145], 2004	Denmark	Cohort (1962-1997)	1160 UC patients; F.U. median 19 yr	4/1160 UC; UC SMR 0.74 (0.20-1.88)	The treatment principles remained unchanged during the entire follow-up period	
Hemminki <i>et</i> al[84], 2012	Sweden	Cohort (1964-2004)	27606 UC patients	UC 277/27606 SIR 1.14 (1.01-1.28), All + 1; SIR 1.08 (0.95-1.22)	Possible incidental finding of PC in older UC patients	
Hemminki et al[83], 2009	Sweden	Cohort (1964-2004)	21788 CD patients	CD 152/21788; SIR 1.19 (1.01-1.4), All + 1 SIR 1.12 (0.94-1.32)	The sparseness of individual cancers did not allow conclusions about the trends	
Jess <i>et al</i> [75], 2013	Denmark	Cohort (1978-2010)	1437 UC; 774 CD	UC SIR 1.82 (1.17-2.71)	No detailed pharmaco- epidemiological analysis	
Jussila <i>et al</i> [<mark>30]</mark> , 2013	Finland	Cohort (1987-1993 and 2000-2007)	21964 IBD (16649 UC; 5315 CD); 5351000 controls	IBD 176/21964; 51045/5351000 controls; IBD SIR 0.84 (0.73-0.97) UC 150/16649 SIR 0.85 (0.72-0.99) $P < 0.05$; CD 26/5315 SIR 0.79 (0.52-1.16)	Possibility of mis-classification of IBD, CD, UC, and Ca. Patients diagnosed 1987-1993 and 2000-2007 were only included	
Kappelman <i>et al</i> [35], 2014	Denmark	Cohort (1978-2010)	42717 IBD (35152 UC; 13756 CD); 5554844 controls. F.U. CD for 7.6 yr, UC for 7.8 yr	IBD 316/42717; controls 33960/5554844. IBD SIR 1.21 (1.08-1.35); UC 258/35152 SIR 1.2 (1.1- 1.4); CD 58/13756 SIR 1.2 (0.9-1.6)	No age-estimates of absolute cancer risk. Detection bias. Data possibly missing. No inpatient encounters prior to 1995	
Wilson <i>et al</i> [<mark>36</mark>], 2016	Switzerland	Case-control (1995- 2012)	19647 IBD (7850 CD; 11797 UC); 19647 controls	IBD 79/19647; 67/19647 controls. IBD aHR 1.19 (0.86-1.65); CD 17/7850; 16/7850 controls; CD aHR 1.08 (0.54-2.15); UC 62/11797; 51/11797 controls UC aHR 1.22 (0.84-1.77)	Exposure misclassification. Potential bias in multivariate analysis (smoking, alcohol, BMI)	
Jung et al [<mark>31</mark>], 2017	Korea	Cohort (2011-2014)	9785 UC; 5506 CD; 50750000 controls	19/15291 IBD; 20607/50750000 controls. IBD SIR 3.5 (2.1-5.5); UC SIR 3.47 (2.06-5.48); CD SIR O.99 (0.03-5.54)	The study did not focus on IBD treatment. Data for disease diagnosis, phenotype not available. Short follow-up	
So et al <mark>[29]</mark> , 2017	China	Cohort (1990-2016)	2621 IBD; 1603 UC; 7392000 controls. Median F.U. 8 yr CD, 10 yr UC	8/2621 IBD; 11115/7392000 controls. IBD SIR 2.03 (1.03-4.06); 8/1603 UC; UC SIR 2.47 (1.24- 4.95)	The 25% of the cohort was followed up for < 5 yr. Small size of PC cases. Lead-time and detection bias. Exposure not evaluated	
Mosher <i>et al</i> [24], 2018	United States	Case-control study Veteran population (1996-2015)	2080 IBD patients; 271898 without IBD	574/2080 IBD; 337/271898 IBD free; 20 yr RR 1.70 (1.28-2.27)	Administrative data. Heterogeneity of the Ca types. Ca incidence rates may be underestimated	
Burns <i>et al</i> [<mark>146]</mark> , 2019	United States	Cohort (1996-2017)	1033 IBD; 9306 IBD free	IBD 30/1033; IBD free 29/9306, 10 yr HR 4.44 (2.98-6.62) <i>P</i> < 0.001; clinically significant PC: 10 yr HR 3.72 (2.15-6.42) <i>P</i> < 0.001; RR 9.32 (5.62-15.46)	Variables for IBD missing. Academic medical centre. PC morbidity, mortality, IBD treatments and healthcare utilization not assessed	
Meyers <i>et al</i> [147], 2020	United States	Prospective Population-based United Kingdom Biobank cohort (2006 and 2010, with follow- up through mid-2015)	2311 IBD (1488 UC; 643 CD); 215773 IBD free; Men aged 40 to 69 at study entry	UC 49/1488; aHR 1.47 (1.11-1.95) $P = 0.0070, \le 20$ yr: 1.29 (0.89, 1.85), > 20 yr: 1.87 (1.21-2.91) $P = 0.0052$, BMI ≤ 30 : 1.48 (1.07, 2.03), BM1 > 30: 1.35 (0.72, 2.51); CD 14/643 aHR 1.06 (0.63-1.80), ≤ 20 yr: 1.11 (0.58, 2.14), > 20 yr: 0.98 (0.41, 2.37), BMI ≤ 30 : 0.83 (0.43, 1.59), BMI > 30: 2.25 (0.93, 5.41)	Number of prior PSA test, DRE, PC morbidity and mortality and IBD treatments were not reported. No data for PC grade or stage. Selection bias	

IBD: Inflammatory bowel disease; Ca: Cancer; PC: Prostate cancer; UC: Ulcerative colitis; SIR: Standardized incidence ratio; CD: Crohn's disease; SMR: Standardized mortality ratio; All + 1: Diagnosis of cancer later than the first year of IBD diagnosis; F.U.: Follow-up; AHR: Adjusted hazard ratio; DRE: Digital rectal examination; BMI: Body mass index; PSA: Prostate-specific antigen.

Baisbideng® WJGO | https://www.wjgnet.com

December 15, 2021 Volume 13 Issue 12

SKIN MALIGNANCIES

Several population-based studies have shown an increased prevalence of skin malignancies among patients with IBD[10,167-169]. Furthermore, the incidence of skin cancer among those receiving immunosuppressants is rising over time[170]. On the contrary, other population-based studies showed that there is no per se increased risk of skin cancer in IBD patients[34,39,75]. The skin cancers include melanoma (MSC) and non-melanoma skin cancers (NMSC) with the latter further divided into squamous cell carcinoma (SCC) and basal cell carcinoma (BCC). It is well established that anti-TNF exposure is associated with the development of MSC[169], whereas NMSC is associated with thiopurines exposure (azathioprine, 6-mercaptopurine)[171]. With regard to the mechanism by which thiopurines may predispose patients to skin cancer the most prominent theory supports that thiopurines increase the vulnerability to UVA radiation. In detail, they cause the accumulation of 6-thioguanine in the DNA of patients and further UVA radiation causes production of reactive oxygen species that leads to DNA mutations and oxidative stress, resulting in oncogenesis[172]. On the other hand, MSC is an immunogenic aggressive tumor, in which suppression of the immune response allows these tumors to grow and metastasize[173]. For this reason, immunosuppressants may cause down-regulation of the tumor surveillance mechanisms, increased susceptibility to infection with oncogenic viruses (e.g., melanoma associated retroviruses), or direct pharmacologic effects of these medications on DNA metabolism[174].

There is evidence from several studies of an increased risk of NMSC in IBD associated with thiopurines use (Table 5). These findings were not confirmed by two other studies that found no increased NMSC risk among IBD thiopurine users[89,175]. A meta-analysis that pooled 60351 IBD patients to assess the risk of NMSC in those treated with thiopurines, showed that the risk was only modestly elevated (aHR 2.28; 95% CI: 1.50-3.45)[184]. There is also evidence of an increased risk of MSC in IBD and this risk is associated with anti-TNF use (Table 5). In a review and meta-analysis that included 172837 IBD patients, 179 MSC cases were reported (pooled crude IR 27.5/100000 person-years; 95% CI: 19.9-37.0) and a 37% increased risk of MSC, independent of biologic therapy[185]. On the contrary, a Danish Cochrane review and meta-analysis (56146 IBD patients 8.1% anti-TNF exposed) did not show increased risk of MSC in those treated with anti-TNF (HR 0.62; CI: 0.08-4.57)[94], which was further confirmed by a recent systematic review and meta-analysis (comprising 34029 biologic-treated and 135370 biologic-naive patients), where biologics did not significantly increase the risk of MSC[186]. In a recent study that included IBD patients with MSC, IBD extent was found to be a risk factor, both in UC (pancolitis OR: 3.09; 95%CI: 1.670-5.727) and CD (ileocolonic disease: OR: 1.98; 95%CI: 1.009-3.882), corticosteroids were risk factors in UC (OR: 1.41-3.72) whereas anti-TNFs were protective factors in UC (OR: 0.15-0.88) and CD (0.27-0.92), but this was attributed to in-situ melanoma only. Moreover, no association between survival of IBD-MSC patients and anti-TNF or immunosuppressants after MSC diagnosis was found[181].

The known risk factors for the development of skin cancers include smoking, older age, male sex, fair skin type, cumulative sun exposure (mostly for NMSC), sun-burn (mostly for MSC), family history of skin cancer, Caucasian race, geographical area and certain genetic factors (e.g., p53 polymorphisms). All these factors should be taken into account when immunosuppressant therapy is being considered for a patient with IBD. Given these findings, drugs other than anti-TNFs might be safer for use in patients after a melanoma diagnosis or in those at high risk for these tumors, whereas thiopurines should also be avoided in patients with a history of SCC, multiple BCCs, or premalignant skin lesions (e.g., solar keratosis)[187].

With regard to the maintenance of immunosuppressive treatment after the initial diagnosis of skin cancer, the results from various studies have been controversial. A large cohort study showed that thiopurines did not increase the risk of second NMSC when used in combination treatment compared with anti-TNF monotherapy (HR 0.79, 95%CI: 0.30-2.08)[180]. In addition, a previous study by Beaugerie concluded that exposure to immunosuppressants did not increase the risk of new or recurrent malignancy in individuals with previous cancer[188]. On the contrary, another recent cohort study with 54919 IBD patients, of whom 518 developed BCC and maintained their treatment, it was shown that repeated BCC occurrences were associated with active thiopurine use[183]. In line with this, a large meta-analysis that included 3706 IBD patients with 10332 person-years of follow-up after a cancer diagnosis showed, in a subgroup on skin cancer alone, that the risk was significantly higher in immunosuppressant-users (71.6 per 1000 person-year, 95% CI: 58.9-84.2, P = 0.035) compared to those not receiving immunosuppressants (50.8 per 1000 person-year, 95% CI: 43.7-57.8)



Table 5 Studies on skin cancer in patients with inflammatory bowel disease							
Ref.	Type of study	Country	Patients	Follow up time	Results	Limitations	
Armstrong <i>et al</i> [89], 2010	Nested case control	United Kingdom	16663 IBD patients; 392 developed Ca vs 1914 IBD controls	6.4 yr	NMSC with AZA use (OR 0.99, CI: 0.35-2.81)	AZA users were included but not 6MP	
Long <i>et al</i> [167], 2010	Retrospective cohort; nested case control	United States	53377 IBD patients vs 160037 non-IBD; 742 IBD NMSC cases vs 2968 IBD controls	1.32 yr	NMSC (IRR, 1.64; 95%CI: 1.51-1.78), NMSC recent TP use (OR, 3.56; 95%CI: 2.81-4.50), recent biologics in CD (OR, 2.07; 95%CI: 1.28- 3.33), persistent TP use (OR, 4.27; 95%CI: 3.08- 5.92), persistent biologic use in CD (OR, 2.18; 95%CI: 1.07-4.46)	Patients aged < 64 yr, no exposure dose, short follow-up	
Singh <i>et al</i> [<mark>168]</mark> , 2011	Retrospective cohort; case control	Canada	9618 IBD patients vs 91378 non-IBD; 237 IBD NMSC cases vs 948 IBD controls	11.7 yr	BCC (HR, 1.20; 95%CI: 1.03-1.40). TP use SCC (HR, 5.40; 95%CI: 2.00-14.56) BCC (HR, 1.12; CI 0.68-1.85). Case-control: TP use SCC (OR, 20.52; 95%CI: 2.42-173.81), BCC (OR: 2.07; 95%CI: 1.10-3.87)	Do not include use of IMMs before 1995	
Peyrin- Biroulet <i>et al</i> [171], 2011	Prospective observational cohort study (CESAME)	France	19486 IBD patients	2.55 yr	NMSC (SIR 2.89, 95%CI: 1.98-4.08) MSC (SIR 0.64, 95%CI: 0.17-1.63). NMSC: ongoing TP use (HR, 5.9; 95%CI: 2.1-16.4; <i>P</i> = 0.0006), past TP use (HR, 3.9; 95%CI: 1.3-12.1; <i>P</i> = 0.02), age per 1-yr increase (HR, 1.08; 95%CI: 1.05-1.11; <i>P</i> < 0.0001)	Younger patients	
van Schaik <i>et al</i> [<mark>175]</mark> , 2011	Retrospective cohort	Holland	2887 IBD patients	6.46 yr	NMSC AZA use (HR 0.85, 95%CI: 0.51-1.41)	Small study sample size	
Long <i>et al</i> [169], 2012	Retrospective cohort; nested case-control	United States	108579 IBD vs 434 233 non-IBD controls; 209 MSC cases vs 823 IBD non-MSC controls, 3288 NMSC cases vs 12945 IBD non- NMSC controls	2 yr	MSC (HR, 1.15; 95% CI: 0.97-1.36) NMSC (HR, 1.34; 95% CI: 1.28-1.40). MSC anti-TNF (OR, 1.88; 95% CI: 1.08-3.29), long-term vs non-long- term use (OR 3.93, 95% CI: 1.82-8.50), no association with TP or 5-ASA. NMSC any TP use (OR, 1.85; 95% CI: 1.66-2.05), anti-TNF (OR, 1.14; 95% CI: 0.95-1.36), combination treatment (OR, 3.89; 95% CI: 2.33-6.46)	Study population aged < 64 yr, no dose information about treatments, short mean follow-up	
Peyrin- Biroulet <i>et al</i> [176], 2012	Prospective observational cohort study (CESAME)	France	19486 IBD patients	2.55 yr	MSC previously TP treated (SIR: 0; 95%CI: 0- 3.11), current TP users (SIR: 1.09; 95%CI: 0.13- 3.94)	Younger patient population	
Abbas <i>et al</i> [177], 2014	Retrospective cohort; nested case control	United States	14527 patients; 421 NMSC and 45 MSC cases	8.1 yr	NMSC current AZA use (HR 2.1, 95%CI: 1.6- 2.6), previous AZA use (HR 0.7, 95%CI: 0.5- 1.0). MSC current AZA use (HR 1.5, 95%CI: 0.6-3.4), previous AZA use (HR 0.5, 95%CI: 90.1-1.8)	Patient population limited to VA health care system (older, white, male)	
McKenna <i>et</i> al[178], 2014	Database inquiry (AE- (FAERS)	United States	315 skin Ca	NA	PRR, increased odds of MSC and NMSC for anti-TNF (P = 0.035 and 0.03, respectively) and combination treatment (P < 0.001 and P < 0.001)	AE database (reporting bias) skewed towards CD	
Kopylov <i>et</i> <i>al</i> [179], 2015	Nested case control	Canada	19582 patients; (MSC 102 vs IBD Controls 1014) (NMSC 474 IBD vs Controls 4684)	No reported mean	NMSC: TP treatment \geq 3 yr (OR 1.41; 95%CI: 1.11-1.79), TP treatment \geq 5 yr (OR: 2.07; 95%CI: 1.36-3.7), combination treatment (OR: 3.11; 95%CI: 1.33-7.27). After stopping TP, OR: 1.04 (0.69-1.55). IMMs-anti-TNF were not associated with MSC	Younger, employed patients are underrepresented, not mentioned disease severity	
Scott <i>et al</i> [180], 2016	Retrospective cohort	United States	2788 IBD patients	2.24 yr	Second NMSC with short-term TP treatment (HR 1.53, 95%CI: 0.87-2.70), with > 1 yr of TP therapy (HR 1.49, 95%CI: 0.98-2.27)	Older patient population	
Nissen <i>et al</i> [181], 2017	2 Retrospective case-control studies	The Netherlands	304 IBD patients with MSC, 1800 IBD controls, 8177 MSC non-IBD controls		MSC: UC (pancolitis OR 3.09; 95%CI: 1.670- 5.727), CD (ileocolonic disease: OR 1.98; 95%CI: 1.009-3.882). Corticosteroids (OR 1.41- 3.72), anti-TNF UC (OR 0.15-0.88), CD (0.27- 0.92). (only attributed to the <i>in situ</i> MSC). Survival with anti-TNF (HR 0.32; 95%CI: 0.08- 1.27) and TP (HR 0.72; 95%CI: 0.37-1.31). Survival after MSC diagnosis anti-TNF (HR 0.16, 95%CI: 0.02-1.21) and TP (HR 0.55, 95%CI: 0.25-1.23)	Medication of patients after 1990 was included. Not informed about skin type, number of sun burns	
Clowry <i>et al</i> [182], 2017	Retrospective cohort	Ireland	2053 patients with IBD	9.8 yr	NMSC under IMMs SIR 1.8 (95%CI: 1.0-2.7), TP exposure (OR: 5.26, 95%CI: 2.15-12.93, P <	Small sample size, hospital database mostly	

Raisbideng® WJGO | https://www.wjgnet.com

				0.001), TP and/or anti-TNF (OR: 6.45, 95%CI: 2.69-15.95, <i>P</i> <0.001)	severe IBD
Khan <i>et al</i> Retrospec [183], 2020 cohort	ctive United States	54919 patients with IBD; VAHS 518 patients with BCC	5.71 yr	Repeated BCC occurrences, compared with 5- ASA, under active TP use (HR 1.65, 95%CI: 1.24-2.19, $P = 0.0005$), 6 mo after TP discontinuation (HR 1.22, 95%CI: 0.86-1.74, P = 0.26), for anti-TNF use (HR 1.27, 95%CI: 0.84-1.90, $P = 0.26$), for combination treatment (HR 1.37, 95%CI: 0.90-2.08, $P = 0.14$)	Study population mostly males. Prescriptions outside VAHS not included

IBD: Inflammatory bowel disease; Ca: Cancer; NMSC: Non-melanoma skin cancer; AZA: Azathioprine; OR: Odds ratio; CI: Confidence interval; 6MP: 6-Mercaptopurine; IRR: Incidence rate ratio; TP: Thiopurine; CD: Crohn's disease; BCC: Basal cell cancer ; SCC: Squamous cell carcinoma; HR: Hazard ratio; IMMs: Immunomodulators; MSC: Melanoma skin cancer; SIR: Standardized incidence ratio; anti-TNF: Anti-tumor necrosis factor; NA: Not applicable; PRR: Projection pursuit regression; AE: Adverse events; 5-ASA: 5-Aminosalicylic acid; VAHS: Veterans affairs healthcare system.

> and numerically higher than anti-TNFα therapy (55.5 per 1000 person-year, 95%CI: 44.7-66.3, P = 0.22 [189].

> With regard to prevention, a recent study showed that only a small proportion of IBD patients (8.3% during the 7-year study period) were seeking dermatologic care [190].

> In conclusion, it seems that there is an increased risk of NMSC and a slightly lower risk of MSC in IBD patients, which justifies the recommendations for routine screening for skin malignancies. Although there is some evidence of second NMSC after an index diagnosis with azathioprine, overall data are conflicting regarding maintaining immunosuppressive treatment after the diagnosis of a skin malignancy and the risk of recurrence or a second skin malignancy, so the decision should be individualized. Conclusively, primary prevention measures, such as sun-protection, annual dermatologic examination and smoking abstinence should be encouraged in all IBD patients, regardless of thiopurine and/or anti-TNF α use.

CONCLUSION

Cancer is the second cause of death in patients with IBD, after cardiovascular disease. Lately there has been evidence that the incidence of intestinal cancers is decreasing over time in IBD patients, as a result of the implementation of a tight screening strategy. Although recent data supporting that there is no excess overall risk of extraintestinal cancers, it seems that IBD patients are in higher risk of certain extraintestinal cancers. This indicates the need for close monitoring, as it seems that chronic systemic inflammation but also other factors such as older age, smoking, specific virus infections (e.g., HPV) and concurrent diseases (PSC) play a role in carcinogenesis. The IBD treatments, for both CD and UC, are aimed at clinical improvement, endoscopic remission and sustained mucosal healing. Although immunosuppressants, including azathioprine, methotrexate and anti-TNFs, constitute the cornerstone of IBD treatment, these drugs might have a carcinogenic effect either by directly altering cellular DNA or reducing tumor immunosurveillance. Lately there are more treatment choices available, such as anti-integrins, anti-IL-12/23 and JAK inhibitors. Although these new treatments are considered to have a better safety profile as far as cancer is concerned, data are limited and future studies are required.

Specialists who deal with the management of IBD patients must bear in mind the possibility of extraintestinal malignancies, especially in those with long standing immunosuppressive therapies and persistent systemic or local inflammation. Guidelines and recommendations should incorporate all the preventative measures [sun protection, dermatologic examination, vaccinations (HPV, hepatitis B virus), annual Pap smear, mammography, PSA testing, annual MRCP where appropriate] and applied in everyday clinical practice to diminish the risk of IBD-related extraintestinal malignancies, achieve an early diagnosis and improve the prognosis and overall outcome.

REFERENCES



¹ de Souza HS, Fiocchi C. Immunopathogenesis of IBD: current state of the art. Nat Rev Gastroenterol Hepatol 2016; 13: 13-27 [PMID: 26627550 DOI: 10.1038/nrgastro.2015.186]

- 2 Nieminen U, Färkkilä M. Malignancies in inflammatory bowel disease. Scand J Gastroenterol 2015; 50: 81-89 [PMID: 25523559 DOI: 10.3109/00365521.2014.992041]
- 3 Lutgens MW, van Oijen MG, van der Heijden GJ, Vleggaar FP, Siersema PD, Oldenburg B. Declining risk of colorectal cancer in inflammatory bowel disease: an updated meta-analysis of population-based cohort studies. Inflamm Bowel Dis 2013; 19: 789-799 [PMID: 23448792 DOI: 10.1097/MIB.0b013e31828029c0
- Jess T, Simonsen J, Jørgensen KT, Pedersen BV, Nielsen NM, Frisch M. Decreasing risk of 4 colorectal cancer in patients with inflammatory bowel disease over 30 years. Gastroenterology 2012; 143: 375-81.e1; quiz e13 [PMID: 22522090 DOI: 10.1053/j.gastro.2012.04.016]
- 5 Katsanos KH, Tatsioni A, Pedersen N, Shuhaibar M, Ramirez VH, Politi P, Rombrechts E, Pierik M, Clofent J, Beltrami M, Bodini P, Freitas J, Mouzas I, Fornaciari G, Moum B, Lakatos PL, Vermeire S, Langholz E, Odes S, Morain CO, Stockbrügger R, Munkholm P, Tsianos EV. Cancer in inflammatory bowel disease 15 years after diagnosis in a population-based European Collaborative follow-up study. J Crohns Colitis 2011; 5: 430-442 [PMID: 21939917 DOI: 10.1016/j.crohns.2011.04.013
- 6 Algaba A, Guerra I, Castaño A, de la Poza G, Castellano VM, López M, Bermejo F. Risk of cancer, with special reference to extra-intestinal malignancies, in patients with inflammatory bowel disease. World J Gastroenterol 2013; 19: 9359-9365 [PMID: 24409063 DOI: 10.3748/wjg.v19.i48.9359]
- Scharl S, Barthel C, Rossel JB, Biedermann L, Misselwitz B, Schoepfer AM, Straumann A, Vavricka SR, Rogler G, Scharl M, Greuter T. Malignancies in Inflammatory Bowel Disease: Frequency, Incidence and Risk Factors-Results from the Swiss IBD Cohort Study. Am J Gastroenterol 2019; 114: 116-126 [PMID: 30333538 DOI: 10.1038/s41395-018-0360-9]
- 8 Loo SY, Vutcovici M, Bitton A, Lakatos PL, Azoulay L, Suissa S, Brassard P. Risk of Malignant Cancers in Inflammatory Bowel Disease. J Crohns Colitis 2019; 13: 1302-1310 [PMID: 30874294 DOI: 10.1093/ecco-jcc/jjz058]
- 9 Taborelli M, Sozzi M, Del Zotto S, Toffolutti F, Montico M, Zanier L, Serraino D. Risk of intestinal and extra-intestinal cancers in patients with inflammatory bowel diseases: A populationbased cohort study in northeastern Italy. PLoS One 2020; 15: e0235142 [PMID: 32574216 DOI: 10.1371/journal.pone.0235142
- 10 Pedersen N, Duricova D, Elkjaer M, Gamborg M, Munkholm P, Jess T. Risk of extra-intestinal cancer in inflammatory bowel disease: meta-analysis of population-based cohort studies. Am JGastroenterol 2010; 105: 1480-1487 [PMID: 20332773 DOI: 10.1038/ajg.2009.760]
- van den Heuvel TR, Wintjens DS, Jeuring SF, Wassink MH, Romberg-Camps MJ, Oostenbrug LE, 11 Sanduleanu S, Hameeteman WH, Zeegers MP, Masclee AA, Jonkers DM, Pierik MJ. Inflammatory bowel disease, cancer and medication: Cancer risk in the Dutch population-based IBDSL cohort. Int J Cancer 2016; 139: 1270-1280 [PMID: 27170593 DOI: 10.1002/ijc.30183]
- 12 Aupérin A. Epidemiology of head and neck cancers: an update. Curr Opin Oncol 2020; 32: 178-186 [PMID: 32209823 DOI: 10.1097/CCO.00000000000629]
- 13 Hashibe M, Brennan P, Chuang SC, Boccia S, Castellsague X, Chen C, Curado MP, Dal Maso L, Daudt AW, Fabianova E, Fernandez L, Wünsch-Filho V, Franceschi S, Hayes RB, Herrero R, Kelsey K, Koifman S, La Vecchia C, Lazarus P, Levi F, Lence JJ, Mates D, Matos E, Menezes A, McClean MD, Muscat J, Eluf-Neto J, Olshan AF, Purdue M, Rudnai P, Schwartz SM, Smith E, Sturgis EM, Szeszenia-Dabrowska N, Talamini R, Wei Q, Winn DM, Shangina O, Pilarska A, Zhang ZF, Ferro G, Berthiller J, Boffetta P. Interaction between tobacco and alcohol use and the risk of head and neck cancer: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium. Cancer Epidemiol Biomarkers Prev 2009; 18: 541-550 [PMID: 19190158 DOI: 10.1158/1055-9965.EPI-08-0347]
- 14 Whiteman DC, Wilson LF. The fractions of cancer attributable to modifiable factors: A global review. Cancer Epidemiol 2016; 44: 203-221 [PMID: 27460784 DOI: 10.1016/j.canep.2016.06.013]
- de Martel C, Plummer M, Vignat J, Franceschi S. Worldwide burden of cancer attributable to HPV 15 by site, country and HPV type. Int J Cancer 2017; 141: 664-670 [PMID: 28369882 DOI: 10.1002/iic.30716]
- 16 Sabatini ME, Chiocca S. Human papillomavirus as a driver of head and neck cancers. Br J Cancer 2020; **122**: 306-314 [PMID: 31708575 DOI: 10.1038/s41416-019-0602-7]
- 17 Ndiaye C, Mena M, Alemany L, Arbyn M, Castellsagué X, Laporte L, Bosch FX, de Sanjosé S, Trottier H. HPV DNA, E6/E7 mRNA, and p16INK4a detection in head and neck cancers: a systematic review and meta-analysis. Lancet Oncol 2014; 15: 1319-1331 [PMID: 25439690 DOI: 10.1016/S1470-2045(14)70471-1]
- 18 Katsanos KH, Roda G, McBride RB, Cohen B, Colombel JF. Increased Risk of Oral Cancer in Patients With Inflammatory Bowel Diseases. Clin Gastroenterol Hepatol 2016; 14: 413-420 [PMID: 26499929 DOI: 10.1016/j.cgh.2015.09.041]
- 19 Nissen LHC, Derikx LAAP, Jacobs AME, van Herpen CM, Kievit W, Verhoeven R, van den Broek E, Bekers E, van den Heuvel T, Pierik M, Rahamat-Langendoen J, Takes RP, Melchers WJG, Nagtegaal ID, Hoentjen F; Dutch Initiative on Crohn and Colitis (ICC); Dutch Head and Neck Society, PALGA group; IBD/HNC group. Risk Factors and Clinical Outcomes of Head and Neck Cancer in Inflammatory Bowel Disease: A Nationwide Cohort Study. Inflamm Bowel Dis 2018; 24: 2015-2026 [PMID: 30759216 DOI: 10.1093/ibd/izy096]
- 20 van de Ven SEM, Derikx LAAP, Nagtegaal ID, van Herpen CM, Takes RP, Melchers WJG, Pierik M, van den Heuvel T, Verhoeven RHA, Hoentjen F, Nissen LHC. Laryngeal Carcinoma in Patients



With Inflammatory Bowel Disease: Clinical Outcomes and Risk Factors. Inflamm Bowel Dis 2020; 26: 1060-1067 [PMID: 31559415 DOI: 10.1093/ibd/izz210]

- 21 Giagkou E. Christodoulou DK. Katsanos KH. Mouth cancer in inflammatory bowel diseases. Oral Dis 2016; 22: 260-264 [PMID: 26671147 DOI: 10.1111/odi.12420]
- 22 Pasternak B, Svanström H, Schmiegelow K, Jess T, Hviid A. Use of azathioprine and the risk of cancer in inflammatory bowel disease. Am J Epidemiol 2013; 177: 1296-1305 [PMID: 23514635 DOI: 10.1093/aje/kws375]
- 23 Nyboe Andersen N, Pasternak B, Friis-Møller N, Andersson M, Jess T. Association between tumour necrosis factor- α inhibitors and risk of serious infections in people with inflammatory bowel disease: nationwide Danish cohort study. BMJ 2015; 350: h2809 [PMID: 26048617 DOI: 10.1136/bmj.h2809]
- 24 Mosher CA, Brown GR, Weideman RA, Crook TW, Cipher DJ, Spechler SJ, Feagins LA. Incidence of Colorectal Cancer and Extracolonic Cancers in Veteran Patients With Inflammatory Bowel Disease. Inflamm Bowel Dis 2018; 24: 617-623 [PMID: 29390104 DOI: 10.1093/ibd/izx046]
- National Cancer Institute. SEER cancer stat facts: thyroid cancer. [cited 21 August 2018]. In: 25 National Cancer Institute [Internet]. Available from: https://seer.cancer.gov/statfacts/html/thyro.html
- Biancone L, Armuzzi A, Scribano ML, Castiglione F, D'Incà R, Orlando A, Papi C, Daperno M, 26 Vecchi M, Riegler G, Fries W, Alvisi P, Meucci G, Mocciaro F, Rogai F, Festa S, Guidi L, Testa A, Spina L, Renna S, Viola A, Patturelli M, Di Mitri R, Frankovic I, Calabrese E, Petruzziello C, De Cristofaro E, Sena G, Ruffa A, Neri B, Rossi A. Cancer Risk in Inflammatory Bowel Disease: A 6-Year Prospective Multicenter Nested Case-Control IG-IBD Study. Inflamm Bowel Dis 2020; 26: 450-459 [PMID: 31498388 DOI: 10.1093/ibd/izz155]
- 27 Yano Y, Matsui T, Hirai F, Okado Y, Sato Y, Tsurumi K, Ishikawa S, Beppu T, Koga A, Yoshizawa N, Higashi D, Futami K. Cancer risk in Japanese Crohn's disease patients: investigation of the standardized incidence ratio. J Gastroenterol Hepatol 2013; 28: 1300-1305 [PMID: 23488881 DOI: 10.1111/jgh.12189
- 28 Sonu IS, Blonski W, Lin MV, Lewis J, Aberra F, Lichtenstein GR. Papillary thyroid cancer and inflammatory bowel disease: is there a relationship? World J Gastroenterol 2013; 19: 1079-1084 [PMID: 23467027 DOI: 10.3748/wjg.v19.i7.1079]
- So J, Tang W, Leung WK, Li M, Lo FH, Wong MTL, Sze ASF, Leung CM, Tsang SWC, Shan 29 EHS, Chan KH, Lam BCY, Hui AJ, Chow WH, Lam TY, Lam V, Lee TW, Lo HHH, Tang CM, Wong CL, Wu JCY, Chan FKL, Sung JJY, Harbord M, Ng SC. Cancer Risk in 2621 Chinese Patients with Inflammatory Bowel Disease: A Population-based Cohort Study. Inflamm Bowel Dis 2017; 23: 2061-2068 [PMID: 28991855 DOI: 10.1097/MIB.00000000001240]
- 30 Jussila A, Virta LJ, Pukkala E, Färkkilä MA. Malignancies in patients with inflammatory bowel disease: a nationwide register study in Finland. Scand J Gastroenterol 2013; 48: 1405-1413 [PMID: 24131389 DOI: 10.3109/00365521.2013.846402]
- 31 Jung YS, Han M, Park S, Kim WH, Cheon JH. Cancer Risk in the Early Stages of Inflammatory Bowel Disease in Korean Patients: A Nationwide Population-based Study. J Crohns Colitis 2017; 11: 954-962 [PMID: 28333358 DOI: 10.1093/ecco-jcc/jjx040]
- 32 Cao L. Assessment of thyroid cancer risk in more than 334,000 patients with inflammatory bowel disease: a case-control study and a meta-analysis. World J Surg Oncol 2018; 16: 182 [PMID: 30200972 DOI: 10.1186/s12957-018-1485-4]
- Wadhwa V, Lopez R, Shen B. Crohn's Disease Is Associated with the Risk for Thyroid Cancer. Inflamm Bowel Dis 2016; 22: 2902-2906 [PMID: 27846192 DOI: 10.1097/MIB.000000000000963
- 34 Jess T, Winther KV, Munkholm P, Langholz E, Binder V. Intestinal and extra-intestinal cancer in Crohn's disease: follow-up of a population-based cohort in Copenhagen County, Denmark. Aliment Pharmacol Ther 2004; 19: 287-293 [PMID: 14984375 DOI: 10.1111/j.1365-2036.2004.01858.x]
- 35 Kappelman MD, Farkas DK, Long MD, Erichsen R, Sandler RS, Sørensen HT, Baron JA. Risk of cancer in patients with inflammatory bowel diseases: a nationwide population-based cohort study with 30 years of follow-up evaluation. Clin Gastroenterol Hepatol 2014; 12: 265-73.e1 [PMID: 23602821 DOI: 10.1016/j.cgh.2013.03.034]
- 36 Wilson JC, Furlano RI, Jick SS, Meier CR. A population-based study examining the risk of malignancy in patients diagnosed with inflammatory bowel disease. J Gastroenterol 2016; 51: 1050-1062 [PMID: 27056729 DOI: 10.1007/s00535-016-1199-8]
- Lo B, Zhao M, Vind I, Burisch J. The Risk of Extraintestinal Cancer in Inflammatory Bowel 37 Disease: A Systematic Review and Meta-analysis of Population-based Cohort Studies. Clin Gastroenterol Hepatol 2021; 19: 1117-1138.e19 [PMID: 32801010 DOI: 10.1016/j.cgh.2020.08.015]
- 38 Masala G, Bagnoli S, Ceroti M, Saieva C, Trallori G, Zanna I, D'Albasio G, Palli D. Divergent patterns of total and cancer mortality in ulcerative colitis and Crohn's disease patients: the Florence IBD study 1978-2001. Gut 2004; 53: 1309-1313 [PMID: 15306591 DOI: 10.1136/gut.2003.031476]
- 39 Bernstein CN, Blanchard JF, Kliewer E, Wajda A. Cancer risk in patients with inflammatory bowel disease: a population-based study. Cancer 2001; 91: 854-862 [PMID: 11241255 DOI: 10.1002/1097-0142(20010215)91:4<854::aid-cncr1073>3.0.co;2-z]
- Karlén P, Löfberg R, Broström O, Leijonmarck CE, Hellers G, Persson PG. Increased risk of cancer 40 in ulcerative colitis: a population-based cohort study. Am J Gastroenterol 1999; 94: 1047-1052 [PMID: 10201481 DOI: 10.1111/j.1572-0241.1999.01012.x]



- 41 Chapman RW, Williamson KD. Are Dominant Strictures in Primary Sclerosing Cholangitis a Risk Factor for Cholangiocarcinoma? Curr Hepatol Rep 2017; 16: 124-129 [PMID: 28706774 DOI: 10.1007/s11901-017-0341-2
- 42 Bonato G, Cristoferi L, Strazzabosco M, Fabris L. Malignancies in Primary Sclerosing Cholangitis-A Continuing Threat. Dig Dis 2015; 33 Suppl 2: 140-148 [PMID: 26641079 DOI: 10.1159/000440826
- Ananthakrishnan AN, Cagan A, Gainer VS, Cheng SC, Cai T, Szolovits P, Shaw SY, Churchill S, 43 Karlson EW, Murphy SN, Kohane I, Liao KP. Mortality and extraintestinal cancers in patients with primary sclerosing cholangitis and inflammatory bowel disease. J Crohns Colitis 2014; 8: 956-963 [PMID: 24559536 DOI: 10.1016/j.crohns.2014.01.019]
- Sørensen JØ, Nielsen OH, Andersson M, Ainsworth MA, Ytting H, Bélard E, Jess T. Inflammatory 44 bowel disease with primary sclerosing cholangitis: A Danish population-based cohort study 1977-2011. Liver Int 2018; 38: 532-541 [PMID: 28796371 DOI: 10.1111/liv.13548]
- Fevery J, Verslype C, Lai G, Aerts R, Van Steenbergen W. Incidence, diagnosis, and therapy of 45 cholangiocarcinoma in patients with primary sclerosing cholangitis. Dig Dis Sci 2007; 52: 3123-3135 [PMID: 17431781 DOI: 10.1007/s10620-006-9681-4]
- 46 Bergquist A, Ekbom A, Olsson R, Kornfeldt D, Lööf L, Danielsson A, Hultcrantz R, Lindgren S, Prytz H, Sandberg-Gertzén H, Almer S, Granath F, Broomé U. Hepatic and extrahepatic malignancies in primary sclerosing cholangitis. J Hepatol 2002; 36: 321-327 [PMID: 11867174 DOI: 10.1016/s0168-8278(01)00288-4]
- 47 Singh S, Talwalkar JA. Primary sclerosing cholangitis: diagnosis, prognosis, and management. Clin Gastroenterol Hepatol 2013; 11: 898-907 [PMID: 23454027 DOI: 10.1016/j.cgh.2013.02.016]
- 48 Boberg KM, Bergquist A, Mitchell S, Pares A, Rosina F, Broomé U, Chapman R, Fausa O, Egeland T, Rocca G, Schrumpf E. Cholangiocarcinoma in primary sclerosing cholangitis: risk factors and clinical presentation. Scand J Gastroenterol 2002; 37: 1205-1211 [PMID: 12408527 DOI: 10.1080/003655202760373434]
- Huai JP, Ding J, Ye XH, Chen YP. Inflammatory bowel disease and risk of cholangiocarcinoma: 49 evidence from a meta-analysis of population-based studies. Asian Pac J Cancer Prev 2014; 15: 3477-3482 [PMID: 24870743 DOI: 10.7314/apjcp.2014.15.8.3477]
- 50 Gulamhusein AF, Eaton JE, Tabibian JH, Atkinson EJ, Juran BD, Lazaridis KN. Duration of Inflammatory Bowel Disease Is Associated With Increased Risk of Cholangiocarcinoma in Patients With Primary Sclerosing Cholangitis and IBD. Am J Gastroenterol 2016; 111: 705-711 [PMID: 27002801 DOI: 10.1038/ajg.2016.55]
- 51 Burak K, Angulo P, Pasha TM, Egan K, Petz J, Lindor KD. Incidence and risk factors for cholangiocarcinoma in primary sclerosing cholangitis. Am J Gastroenterol 2004; 99: 523-526 [PMID: 15056096 DOI: 10.1111/j.1572-0241.2004.04067.x]
- 52 Broomé U, Löfberg R, Veress B, Eriksson LS. Primary sclerosing cholangitis and ulcerative colitis: evidence for increased neoplastic potential. Hepatology 1995; 22: 1404-1408 [PMID: 7590655 DOI: 10.1002/hep.1840220511]
- European Association for the Study of the Liver. EASL Clinical Practice Guidelines: 53 management of cholestatic liver diseases. J Hepatol 2009; 51: 237-267 [PMID: 19501929 DOI: 10.1016/j.jhep.2009.04.009]
- 54 Chapman MH, Webster GJ, Bannoo S, Johnson GJ, Wittmann J, Pereira SP. Cholangiocarcinoma and dominant strictures in patients with primary sclerosing cholangitis: a 25-year single-centre experience. Eur J Gastroenterol Hepatol 2012; 24: 1051-1058 [PMID: 22653260 DOI: 10.1097/MEG.0b013e3283554bbf
- 55 Gotthardt DN, Rudolph G, Klöters-Plachky P, Kulaksiz H, Stiehl A. Endoscopic dilation of dominant stenoses in primary sclerosing cholangitis: outcome after long-term treatment. Gastrointest Endosc 2010; 71: 527-534 [PMID: 20189511 DOI: 10.1016/j.gie.2009.10.041]
- 56 Rudolph G, Gotthardt D, Klöters-Plachky P, Kulaksiz H, Rost D, Stiehl A. Influence of dominant bile duct stenoses and biliary infections on outcome in primary sclerosing cholangitis. J Hepatol 2009; 51: 149-155 [PMID: 19410324 DOI: 10.1016/j.jhep.2009.01.023]
- Rudolph G, Gotthardt D, Kloeters-Plachky P, Rost D, Kulaksiz H, Stiehl A. In PSC with dominant 57 bile duct stenosis, IBD is associated with an increase of carcinomas and reduced survival. J Hepatol 2010; 53: 313-317 [PMID: 20472317 DOI: 10.1016/j.jhep.2010.02.030]
- 58 Janse M, Lamberts LE, Verdonk RC, Weersma RK. IBD is associated with an increase in carcinoma in PSC irrespective of the presence of dominant bile duct stenosis. J Hepatol 2012; 57: 473-4; author reply 475 [PMID: 22537688 DOI: 10.1016/j.jhep.2012.02.034]
- Björnsson E, Olsson R, Bergquist A, Lindgren S, Braden B, Chapman RW, Boberg KM, Angulo P. 59 The natural history of small-duct primary sclerosing cholangitis. Gastroenterology 2008; 134: 975-980 [PMID: 18395078 DOI: 10.1053/j.gastro.2008.01.042]
- 60 Fevery J, Henckaerts L, Van Oirbeek R, Vermeire S, Rutgeerts P, Nevens F, Van Steenbergen W. Malignancies and mortality in 200 patients with primary sclerosering cholangitis: a long-term singlecentre study. Liver Int 2012; 32: 214-222 [PMID: 21745316 DOI: 10.1111/j.1478-3231.2011.02575.x
- 61 Ishida M, Naka S, Shiomi H, Tsujikawa T, Andoh A, Nakahara T, Saito Y, Kurumi Y, Takikita-Suzuki M, Kojima F, Hotta M, Tani T, Fujiyama Y, Okabe H. Hepatocellular carcinoma occurring in a Crohn's disease patient. World J Gastroenterol 2010; 16: 3215-3218 [PMID: 20593510 DOI: 10.3748/wjg.v16.i25.3215]



- 62 Trivedi PJ, Crothers H, Mytton J, Bosch S, Iqbal T, Ferguson J, Hirschfield GM. Effects of Primary Sclerosing Cholangitis on Risks of Cancer and Death in People With Inflammatory Bowel Disease, Based on Sex, Race, and Age. Gastroenterology 2020; 159: 915-928 [PMID: 32445859 DOI: 10.1053/j.gastro.2020.05.049]
- 63 Brandt DJ, MacCarty RL, Charboneau JW, LaRusso NF, Wiesner RH, Ludwig J. Gallbladder disease in patients with primary sclerosing cholangitis. AJR Am J Roentgenol 1988; 150: 571-574 [PMID: 3277348 DOI: 10.2214/ajr.150.3.571]
- Ahrendt SA, Rashid A, Chow JT, Eisenberger CF, Pitt HA, Sidransky D. p53 overexpression and 64 K-ras gene mutations in primary sclerosing cholangitis-associated biliary tract cancer. J Hepatobiliary Pancreat Surg 2000; 7: 426-431 [PMID: 11180865 DOI: 10.1007/s005340070039]
- 65 Bergquist A, Glaumann H, Persson B, Broomé U. Risk factors and clinical presentation of hepatobiliary carcinoma in patients with primary sclerosing cholangitis: a case-control study. Hepatology 1998; 27: 311-316 [PMID: 9462625 DOI: 10.1002/hep.510270201]
- Brandsaeter B, Isoniemi H, Broomé U, Olausson M, Bäckman L, Hansen B, Schrumpf E, Oksanen 66 A, Ericzon BG, Höckerstedt K, Mäkisalo H, Kirkegaard P, Friman S, Bjøro K. Liver transplantation for primary sclerosing cholangitis; predictors and consequences of hepatobiliary malignancy. J Hepatol 2004; 40: 815-822 [PMID: 15094230 DOI: 10.1016/j.jhep.2004.01.002]
- Buckles DC, Lindor KD, Larusso NF, Petrovic LM, Gores GJ. In primary sclerosing cholangitis, 67 gallbladder polyps are frequently malignant. Am J Gastroenterol 2002; 97: 1138-1142 [PMID: 12014717 DOI: 10.1111/j.1572-0241.2002.05677.x]
- Campbell WL, Ferris JV, Holbert BL, Thaete FL, Baron RL. Biliary tract carcinoma complicating primary sclerosing cholangitis: evaluation with CT, cholangiography, US, and MR imaging. Radiology 1998; 207: 41-50 [PMID: 9530297 DOI: 10.1148/radiology.207.1.9530297]
- 69 Campbell WL, Peterson MS, Federle MP, Siqueira ES, Slivka A, Grazioli L, Ichikawa T, Oliver JH 3rd, Kim T, Li W. Using CT and cholangiography to diagnose biliary tract carcinoma complicating primary sclerosing cholangitis. AJR Am J Roentgenol 2001; 177: 1095-1100 [PMID: 11641179 DOI: 10.2214/ajr.177.5.1771095]
- 70 Dorudi S, Chapman RW, Kettlewell MG. Carcinoma of the gallbladder in ulcerative colitis and primary sclerosing cholangitis. Report of two cases. Dis Colon Rectum 1991; 34: 827-828 [PMID: 1914750 DOI: 10.1007/BF02051079]
- 71 Lewis JT, Talwalkar JA, Rosen CB, Smyrk TC, Abraham SC. Prevalence and risk factors for gallbladder neoplasia in patients with primary sclerosing cholangitis: evidence for a metaplasiadysplasia-carcinoma sequence. Am J Surg Pathol 2007; 31: 907-913 [PMID: 17527079 DOI: 10.1097/01.pas.0000213435.99492.8a
- 72 Everhov ÅH, Erichsen R, Sachs MC, Pedersen L, Halfvarson J, Askling J, Ekbom A, Ludvigsson JF, Sørensen HT, Olén O. Inflammatory bowel disease and pancreatic cancer: a Scandinavian register-based cohort study 1969-2017. Aliment Pharmacol Ther 2020; 52: 143-154 [PMID: 32412143 DOI: 10.1111/apt.15785]
- Correa P. Human gastric carcinogenesis: a multistep and multifactorial process--First American 73 Cancer Society Award Lecture on Cancer Epidemiology and Prevention. Cancer Res 1992; 52: 6735-6740 [PMID: 1458460]
- 74 Papamichael K, Konstantopoulos P, Mantzaris GJ. Helicobacter pylori infection and inflammatory bowel disease: is there a link? World J Gastroenterol 2014; 20: 6374-6385 [PMID: 24914359 DOI: 10.3748/wjg.v20.i21.6374]
- 75 Jess T, Horváth-Puhó E, Fallingborg J, Rasmussen HH, Jacobsen BA. Cancer risk in inflammatory bowel disease according to patient phenotype and treatment: a Danish population-based cohort study. Am J Gastroenterol 2013; 108: 1869-1876 [PMID: 23978954 DOI: 10.1038/ajg.2013.249]
- Nissen LH, Assendorp EL, van der Post RS, Derikx LA, de Jong DJ, Kievit W, Pierik M, van den 76 Heuvel T, Verhoeven R, Overbeek LI, Hoentjen F, Nagtegaal ID. Impaired Gastric Cancer Survival in Patients with Inflammatory Bowel Disease. J Gastrointestin Liver Dis 2016; 25: 431-440 [PMID: 27981298 DOI: 10.15403/jgld.2014.1121.254.nis]
- Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. 77 CA Cancer J Clin 2015; 65: 87-108 [PMID: 25651787 DOI: 10.3322/caac.21262]
- 78 Lichtarowicz A, Norman C, Calcraft B, Morris JS, Rhodes J, Mayberry J. A study of the menopause, smoking, and contraception in women with Crohn's disease. Q J Med 1989; 72: 623-631 [PMID: 2608881]
- 79 Bharadwaj S, Kulkarni G, Shen B. Menstrual cycle, sex hormones in female inflammatory bowel disease patients with and without surgery. J Dig Dis 2015; 16: 245-255 [PMID: 25851437 DOI: 10.1111/1751-2980.12247
- 80 Ekbom A, Helmick C, Zack M, Adami HO. Extracolonic malignancies in inflammatory bowel disease. Cancer 1991; 67: 2015-2019 [PMID: 2004319 DOI: 10.1002/1097-0142(19910401)67:7<2015::aid-cncr2820670731>3.0.co;2-r]
- 81 Mellemkjaer L, Olsen JH, Frisch M, Johansen C, Gridley G, McLaughlin JK. Cancer in patients with ulcerative colitis. Int J Cancer 1995; 60: 330-333 [PMID: 7829239 DOI: 10.1002/ijc.2910600309
- 82 Mellemkjaer L, Johansen C, Gridley G, Linet MS, Kjaer SK, Olsen JH. Crohn's disease and cancer risk (Denmark). Cancer Causes Control 2000; 11: 145-150 [PMID: 10710198 DOI: 10.1023/a:1008988215904
- Hemminki K, Li X, Sundquist J, Sundquist K. Cancer risks in Crohn disease patients. Ann Oncol 83



2009; 20: 574-580 [PMID: 18765463 DOI: 10.1093/annonc/mdn595]

- 84 Hemminki K, Liu X, Ji J, Försti A, Sundquist J, Sundquist K. Effect of autoimmune diseases on risk and survival in female cancers. Gynecol Oncol 2012; 127: 180-185 [PMID: 22819787 DOI: 10.1016/j.ygyno.2012.07.100
- 85 Algaba A, Guerra I, Marín-Jiménez I, Quintanilla E, López-Serrano P, García-Sánchez MC, Casis B, Taxonera C, Moral I, Chaparro M, Martín-Rodríguez D, Martín-Arranz MD, Manceñido N, Menchén L, López-Sanromán A, Castaño Á, Bermejo F. Incidence, management, and course of cancer in patients with inflammatory bowel disease. J Crohns Colitis 2015; 9: 326-333 [PMID: 25687203 DOI: 10.1093/ecco-jcc/jjv032]
- 86 Lichtenstein GR, Feagan BG, Cohen RD, Salzberg BA, Diamond RH, Langholff W, Londhe A, Sandborn WJ. Drug therapies and the risk of malignancy in Crohn's disease: results from the TREAT™ Registry. Am J Gastroenterol 2014; 109: 212-223 [PMID: 24394749 DOI: 10.1038/ajg.2013.441]
- 87 Tsai MS, Chen HP, Hung CM, Lee PH, Lin CL, Kao CH. Hospitalization for Inflammatory Bowel Disease is Associated with Increased Risk of Breast Cancer: A Nationwide Cohort Study of an Asian Population. Ann Surg Oncol 2015; 22: 1996-2002 [PMID: 25354573 DOI: 10.1245/s10434-014-4198-0
- 88 Hovde Ø, Høivik ML, Henriksen M, Solberg IC, Småstuen MC, Moum BA. Malignancies in Patients with Inflammatory Bowel Disease: Results from 20 Years of Follow-up in the IBSEN Study. J Crohns Colitis 2017; 11: 571-577 [PMID: 28453756 DOI: 10.1093/ecco-jcc/jjw193]
- 89 Armstrong RG, West J, Card TR. Risk of cancer in inflammatory bowel disease treated with azathioprine: a UK population-based case-control study. Am J Gastroenterol 2010; 105: 1604-1609 [PMID: 20104215 DOI: 10.1038/ajg.2009.745]
- 90 Fraser AG, Orchard TR, Robinson EM, Jewell DP. Long-term risk of malignancy after treatment of inflammatory bowel disease with azathioprine. Aliment Pharmacol Ther 2002; 16: 1225-1232 [PMID: 12144571 DOI: 10.1046/j.1365-2036.2002.01297.x]
- 91 Connell WR, Kamm MA, Dickson M, Balkwill AM, Ritchie JK, Lennard-Jones JE. Long-term neoplasia risk after azathioprine treatment in inflammatory bowel disease. Lancet 1994; 343: 1249-1252 [PMID: 7910274 DOI: 10.1016/s0140-6736(94)92150-4]
- 92 Setshedi M, Epstein D, Winter TA, Myer L, Watermeyer G, Hift R. Use of thiopurines in the treatment of inflammatory bowel disease is associated with an increased risk of non-melanoma skin cancer in an at-risk population: a cohort study. J Gastroenterol Hepatol 2012; 27: 385-389 [PMID: 21793904 DOI: 10.1111/j.1440-1746.2011.06865.x]
- 93 Gómez-García M, Cabello-Tapia MJ, Sánchez-Capilla AD, De Teresa-Galván J, Redondo-Cerezo E. Thiopurines related malignancies in inflammatory bowel disease: local experience in Granada, Spain. World J Gastroenterol 2013; 19: 4877-4886 [PMID: 23946592 DOI: 10.3748/wjg.v19.i30.4877]
- Nyboe Andersen N, Pasternak B, Basit S, Andersson M, Svanström H, Caspersen S, Munkholm P, 94 Hviid A, Jess T. Association between tumor necrosis factor-α antagonists and risk of cancer in patients with inflammatory bowel disease. JAMA 2014; 311: 2406-2413 [PMID: 24938563 DOI: 10.1001/jama.2014.5613
- 95 Colombel JF, Sands BE, Rutgeerts P, Sandborn W, Danese S, D'Haens G, Panaccione R, Loftus EV Jr, Sankoh S, Fox I, Parikh A, Milch C, Abhyankar B, Feagan BG. The safety of vedolizumab for ulcerative colitis and Crohn's disease. Gut 2017; 66: 839-851 [PMID: 26893500 DOI: 10.1136/gutinl-2015-311079
- 96 Osterman MT, Sandborn WJ, Colombel JF, Robinson AM, Lau W, Huang B, Pollack PF, Thakkar RB, Lewis JD. Increased risk of malignancy with adalimumab combination therapy, compared with monotherapy, for Crohn's disease. Gastroenterology 2014; 146: 941-949 [PMID: 24361468 DOI: 10.1053/j.gastro.2013.12.025
- 97 Beigel F, Steinborn A, Schnitzler F, Tillack C, Breiteneicher S, John JM, Van Steen K, Laubender RP, Göke B, Seiderer J, Brand S, Ochsenkühn T. Risk of malignancies in patients with inflammatory bowel disease treated with thiopurines or anti-TNF alpha antibodies. Pharmacoepidemiol Drug Saf 2014; 23: 735-744 [PMID: 24788825 DOI: 10.1002/pds.3621]
- 98 Fidder H, Schnitzler F, Ferrante M, Noman M, Katsanos K, Segaert S, Henckaerts L, Van Assche G, Vermeire S, Rutgeerts P. Long-term safety of infliximab for the treatment of inflammatory bowel disease: a single-centre cohort study. Gut 2009; 58: 501-508 [PMID: 18832524 DOI: 10.1136/gut.2008.163642]
- Lichtenstein GR, Rutgeerts P, Sandborn WJ, Sands BE, Diamond RH, Blank M, Montello J, Tang 99 L, Cornillie F, Colombel JF. A pooled analysis of infections, malignancy, and mortality in infliximab- and immunomodulator-treated adult patients with inflammatory bowel disease. Am J Gastroenterol 2012; 107: 1051-1063 [PMID: 22613901 DOI: 10.1038/ajg.2012.89]
- Williams CJ, Peyrin-Biroulet L, Ford AC. Systematic review with meta-analysis: malignancies with 100 anti-tumour necrosis factor- α therapy in inflammatory bowel disease. Aliment Pharmacol Ther 2014; 39: 447-458 [PMID: 24444171 DOI: 10.1111/apt.12624]
- Waggoner SE. Cervical cancer. Lancet 2003; 361: 2217-2225 [PMID: 12842378 DOI: 101 10.1016/S0140-6736(03)13778-6
- Burd EM. Human papillomavirus and cervical cancer. Clin Microbiol Rev 2003; 16: 1-17 [PMID: 102 12525422 DOI: 10.1128/cmr.16.1.1-17.2003]
- 103 Wheeler CM. Natural history of human papillomavirus infections, cytologic and histologic



abnormalities, and cancer. Obstet Gynecol Clin North Am 2008; 35: 519-36; vii [PMID: 19061814 DOI: 10.1016/j.ogc.2008.09.006]

- 104 Dalstein V, Riethmuller D, Prétet JL, Le Bail Carval K, Sautière JL, Carbillet JP, Kantelip B, Schaal JP, Mougin C. Persistence and load of high-risk HPV are predictors for development of high-grade cervical lesions: a longitudinal French cohort study. Int J Cancer 2003; 106: 396-403 [PMID: 12845680 DOI: 10.1002/ijc.11222]
- 105 US Preventive Services Task Force, Curry SJ, Krist AH, Owens DK, Barry MJ, Caughey AB, Davidson KW, Doubeni CA, Epling JW Jr, Kemper AR, Kubik M, Landefeld CS, Mangione CM, Phipps MG, Silverstein M, Simon MA, Tseng CW, Wong JB. Screening for Cervical Cancer: US Preventive Services Task Force Recommendation Statement. JAMA 2018; 320: 674-686 [PMID: 30140884 DOI: 10.1001/jama.2018.10897]
- 106 FUTURE II Study Group. Quadrivalent vaccine against human papillomavirus to prevent highgrade cervical lesions. N Engl J Med 2007; 356: 1915-1927 [PMID: 17494925 DOI: 10.1056/NEJMoa061741
- Shrestha AD, Neupane D, Vedsted P, Kallestrup P. Cervical Cancer Prevalence, Incidence and 107 Mortality in Low and Middle Income Countries: A Systematic Review Asian Pac J Cancer Prev 2018; 19: 319-324 [PMID: 29479954 DOI: 10.22034/APJCP.2018.19.2.319]
- Strickler HD, Burk RD, Fazzari M, Anastos K, Minkoff H, Massad LS, Hall C, Bacon M, Levine 108 AM, Watts DH, Silverberg MJ, Xue X, Schlecht NF, Melnick S, Palefsky JM. Natural history and possible reactivation of human papillomavirus in human immunodeficiency virus-positive women. J Natl Cancer Inst 2005; 97: 577-586 [PMID: 15840880 DOI: 10.1093/jnci/dji073]
- García-Piñeres AJ, Hildesheim A, Herrero R, Trivett M, Williams M, Atmetlla I, Ramírez M, 109 Villegas M, Schiffman M, Rodríguez AC, Burk RD, Hildesheim M, Freer E, Bonilla J, Bratti C, Berzofsky JA, Pinto LA. Persistent human papillomavirus infection is associated with a generalized decrease in immune responsiveness in older women. Cancer Res 2006; 66: 11070-11076 [PMID: 17108147 DOI: 10.1158/0008-5472.CAN-06-2034]
- 110 Bhatia J, Bratcher J, Korelitz B, Vakher K, Mannor S, Shevchuk M, Panagopoulos G, Ofer A, Tamas E, Kotsali P, Vele O. Abnormalities of uterine cervix in women with inflammatory bowel disease. World J Gastroenterol 2006; 12: 6167-6171 [PMID: 17036389 DOI: 10.3748/wjg.v12.i38.6167
- 111 Kane S, Khatibi B, Reddy D. Higher incidence of abnormal Pap smears in women with inflammatory bowel disease. Am J Gastroenterol 2008; 103: 631-636 [PMID: 17941962 DOI: 10.1111/j.1572-0241.2007.01582.x
- 112 Hutfless S, Fireman B, Kane S, Herrinton LJ. Screening differences and risk of cervical cancer in inflammatory bowel disease. Aliment Pharmacol Ther 2008; 28: 598-605 [PMID: 18549465 DOI: 10.1111/j.1365-2036.2008.03766.x
- Marehbian J, Arrighi HM, Hass S, Tian H, Sandborn WJ. Adverse events associated with common 113 therapy regimens for moderate-to-severe Crohn's disease. Am J Gastroenterol 2009; 104: 2524-2533 [PMID: 19532125 DOI: 10.1038/ajg.2009.322]
- Lees CW, Critchley J, Chee N, Beez T, Gailer RE, Williams AR, Shand AG, Arnott ID, Satsangi J. Lack of association between cervical dysplasia and IBD: a large case-control study. Inflamm Bowel Dis 2009; 15: 1621-1629 [PMID: 19618462 DOI: 10.1002/ibd.20959]
- Singh H, Demers AA, Nugent Z, Mahmud SM, Kliewer EV, Bernstein CN. Risk of cervical 115 abnormalities in women with inflammatory bowel disease: a population-based nested case-control study. Gastroenterology 2009; 136: 451-458 [PMID: 18996382 DOI: 10.1053/j.gastro.2008.10.021]
- 116 Rungoe C, Simonsen J, Riis L, Frisch M, Langholz E, Jess T. Inflammatory bowel disease and cervical neoplasia: a population-based nationwide cohort study. Clin Gastroenterol Hepatol 2015; 13: 693-700.e1 [PMID: 25086189 DOI: 10.1016/j.cgh.2014.07.036]
- 117 Segal JP, Askari A, Clark SK, Hart AL, Faiz OD, The Incidence and Prevalence of Human Papilloma Virus-associated Cancers in IBD. Inflamm Bowel Dis 2021; 27: 34-39 [PMID: 32080713 DOI: 10.1093/ibd/izaa035]
- Li M, Yang QF, Cao Q, Tang J, Gao Y, Zhi M, Chao K, Su ML, Huang WM, Yi Y, Xia SY, Huang 118 LJ, Zhao Y, Wang XH, Liu XY, Lin L, Hu PJ, Gao X. High-risk human papilloma virus infection and cervical neoplasm in female inflammatory bowel disease patients: a cross-sectional study. Gastroenterol Rep (Oxf) 2019; 7: 338-344 [PMID: 31687153 DOI: 10.1093/gastro/goy053]
- 119 Kim SC, Glynn RJ, Giovannucci E, Hernández-Díaz S, Liu J, Feldman S, Karlson EW, Schneeweiss S, Solomon DH. Risk of high-grade cervical dysplasia and cervical cancer in women with systemic inflammatory diseases: a population-based cohort study. Ann Rheum Dis 2015; 74: 1360-1367 [PMID: 24618265 DOI: 10.1136/annrheumdis-2013-204993]
- 120 Allegretti JR, Barnes EL, Cameron A. Are patients with inflammatory bowel disease on chronic immunosuppressive therapy at increased risk of cervical high-grade dysplasia/cancer? Inflamm Bowel Dis 2015; 21: 1089-1097 [PMID: 25895005 DOI: 10.1097/MIB.00000000000338]
- Singh H, Nugent Z, Demers AA, Bernstein CN. Screening for cervical and breast cancer among 121 women with inflammatory bowel disease: a population-based study. Inflamm Bowel Dis 2011; 17: 1741-1750 [PMID: 21744429 DOI: 10.1002/ibd.21567]
- 122 Long MD, Porter CQ, Sandler RS, Kappelman MD. Suboptimal rates of cervical testing among women with inflammatory bowel disease. Clin Gastroenterol Hepatol 2009; 7: 549-553 [PMID: 18996498 DOI: 10.1016/j.cgh.2008.10.0071
- 123 Xu F, Dahlhamer JM, Terlizzi EP, Wheaton AG, Croft JB. Receipt of Preventive Care Services



Among US Adults with Inflammatory Bowel Disease, 2015-2016. Dig Dis Sci 2019; 64: 1798-1808 [PMID: 30746631 DOI: 10.1007/s10620-019-05494-w]

- 124 **Parian A.** Lazarev M. Who and how to screen for cancer in at-risk inflammatory bowel disease patients. Expert Rev Gastroenterol Hepatol 2015; 9: 731-746 [PMID: 25592672 DOI: 10.1586/17474124.2015.1003208]
- Goldacre MJ, Wotton CJ, Yeates D, Seagroatt V, Collier J. Liver cirrhosis, other liver diseases, 125 pancreatitis and subsequent cancer: record linkage study. Eur J Gastroenterol Hepatol 2008; 20: 384-392 [PMID: 18403939 DOI: 10.1097/MEG.0b013e3282f4489f]
- 126 Cheddani H, Dauchet L, Fumery M, Charpentier C, Marie Bouvier A, Dupas JL, Pariente B, Peyrin-Biroulet L, Savoye G, Gower-Rousseau C. Cancer in Elderly Onset Inflammatory Bowel Disease: A Population-Based Study. Am J Gastroenterol 2016; 111: 1428-1436 [PMID: 27481308 DOI: 10.1038/ajg.2016.304]
- 127 Yadav S, Singh S, Harmsen WS, Edakkanambeth Varayil J, Tremaine WJ, Loftus EV Jr. Effect of Medications on Risk of Cancer in Patients With Inflammatory Bowel Diseases: A Population-Based Cohort Study from Olmsted County, Minnesota. Mayo Clin Proc 2015; 90: 738-746 [PMID: 25963756 DOI: 10.1016/j.mayocp.2015.03.024]
- Boffetta P. Tobacco smoking and risk of bladder cancer. Scand J Urol Nephrol Suppl 2008; 45-54 128 [PMID: 18815916 DOI: 10.1080/03008880802283664]
- Burger M, Catto JW, Dalbagni G, Grossman HB, Herr H, Karakiewicz P, Kassouf W, Kiemeney 129 LA, La Vecchia C, Shariat S, Lotan Y. Epidemiology and risk factors of urothelial bladder cancer. Eur Urol 2013; 63: 234-241 [PMID: 22877502 DOI: 10.1016/j.eururo.2012.07.033]
- 130 Rini BI, Campbell SC, Escudier B. Renal cell carcinoma. Lancet 2009; 373: 1119-1132 [PMID: 19269025 DOI: 10.1016/S0140-6736(09)60229-4]
- 131 Chow WH, Dong LM, Devesa SS. Epidemiology and risk factors for kidney cancer. Nat Rev Urol 2010; 7: 245-257 [PMID: 20448658 DOI: 10.1038/nrurol.2010.46]
- Balkwill F. Tumour necrosis factor and cancer. Nat Rev Cancer 2009; 9: 361-371 [PMID: 19343034 132 DOI: 10.1038/nrc26281
- 133 Galbán S, Fan J, Martindale JL, Cheadle C, Hoffman B, Woods MP, Temeles G, Brieger J, Decker J, Gorospe M. von Hippel-Lindau protein-mediated repression of tumor necrosis factor alpha translation revealed through use of cDNA arrays. Mol Cell Biol 2003; 23: 2316-2328 [PMID: 12640117 DOI: 10.1128/mcb.23.7.2316-2328.2003]
- 134 Biancone L, Orlando A, Kohn A, Colombo E, Sostegni R, Angelucci E, Rizzello F, Castiglione F, Benazzato L, Papi C, Meucci G, Riegler G, Petruzziello C, Mocciaro F, Geremia A, Calabrese E, Cottone M, Pallone F. Infliximab and newly diagnosed neoplasia in Crohn's disease: a multicentre matched pair study. Gut 2006; 55: 228-233 [PMID: 16120759 DOI: 10.1136/gut.2005.075937]
- 135 Biancone L, Petruzziello C, Orlando A, Kohn A, Ardizzone S, Daperno M, Angelucci E, Castiglione F, D'Incà R, Zorzi F, Papi C, Meucci G, Riegler G, Sica G, Rizzello F, Mocciaro F, Onali S, Calabrese E, Cottone M, Pallone F. Cancer in Crohn's Disease patients treated with infliximab: a long-term multicenter matched pair study. Inflamm Bowel Dis 2011; 17: 758-766 [PMID: 20684009 DOI: 10.1002/ibd.21416]
- Derikx LA, Nissen LH, Drenth JP, van Herpen CM, Kievit W, Verhoeven RH, Mulders PF, 136 Hulsbergen-van de Kaa CA, Boers-Sonderen MJ, van den Heuvel TR, Pierik M, Nagtegaal ID, Hoentjen F; Dutch Initiative on Crohn and Colitis; PALGA Group; IBD/RCC Group. Better survival of renal cell carcinoma in patients with inflammatory bowel disease. Oncotarget 2015; 6: 38336-38347 [PMID: 26447542 DOI: 10.18632/oncotarget.5186]
- Madanchi M, Zeitz J, Barthel C, Samaras P, Scharl S, Sulz MC, Biedermann L, Frei P, Vavricka 137 SR, Rogler G, Scharl M. Malignancies in Patients with Inflammatory Bowel Disease: A Single-Centre Experience. Digestion 2016; 94: 1-8 [PMID: 27318857 DOI: 10.1159/000447259]
- 138 Bourrier A, Carrat F, Colombel JF, Bouvier AM, Abitbol V, Marteau P, Cosnes J, Simon T, Peyrin-Biroulet L, Beaugerie L; CESAME study group. Excess risk of urinary tract cancers in patients receiving thiopurines for inflammatory bowel disease: a prospective observational cohort study. Aliment Pharmacol Ther 2016; 43: 252-261 [PMID: 26549003 DOI: 10.1111/apt.13466]
- 139 Wauters L, Billiet T, Papamichael K, Ballet V, Joniau S, Verschueren P, Silversmit G, Van Assche G, Vermeire S, Ferrante M. Incidence of renal cell carcinoma in inflammatory bowel disease patients with and without anti-TNF treatment. Eur J Gastroenterol Hepatol 2017; 29: 84-90 [PMID: 27603297 DOI: 10.1097/MEG.000000000000735]
- 140 Ficarra V, Prayer-Galetti T, Novella G, Bratti E, Maffei N, Dal Bianco M, Artibani W, Pagano F. Incidental detection beyond pathological factors as prognostic predictor of renal cell carcinoma. Eur Urol 2003; 43: 663-669 [PMID: 12767368 DOI: 10.1016/s0302-2838(03)00142-8]
- Palsdottir HB, Hardarson S, Petursdottir V, Jonsson A, Jonsson E, Sigurdsson MI, Einarsson GV, 141 Gudbjartsson T. Incidental detection of renal cell carcinoma is an independent prognostic marker: results of a long-term, whole population study. J Urol 2012; 187: 48-53 [PMID: 22088336 DOI: 10.1016/j.juro.2011.09.025]
- 142 Lakatos PL, Szamosi T, Lakatos L. Smoking in inflammatory bowel diseases: good, bad or ugly? World J Gastroenterol 2007; 13: 6134-6139 [PMID: 18069751 DOI: 10.3748/wjg.v13.i46.6134]
- 143 Mahid SS, Minor KS, Soto RE, Hornung CA, Galandiuk S. Smoking and inflammatory bowel disease: a meta-analysis. Mayo Clin Proc 2006; 81: 1462-1471 [PMID: 17120402 DOI: 10.4065/81.11.1462
- 144 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018:



GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]

- 145 Winther KV, Jess T, Langholz E, Munkholm P, Binder V. Long-term risk of cancer in ulcerative colitis: a population-based cohort study from Copenhagen County. Clin Gastroenterol Hepatol 2004; 2: 1088-1095 [PMID: 15625654 DOI: 10.1016/s1542-3565(04)00543-9]
- 146 Burns JA, Weiner AB, Catalona WJ, Li EV, Schaeffer EM, Hanauer SB, Strong S, Burns J, Hussain MHA, Kundu SD. Inflammatory Bowel Disease and the Risk of Prostate Cancer. Eur Urol 2019; 75: 846-852 [PMID: 30528221 DOI: 10.1016/j.eururo.2018.11.039]
- Meyers TJ, Weiner AB, Graff RE, Desai AS, Cooley LF, Catalona WJ, Hanauer SB, Wu JD, 147 Schaeffer EM, Abdulkadir SA, Kundu SD, Witte JS. Association between inflammatory bowel disease and prostate cancer: A large-scale, prospective, population-based study. Int J Cancer 2020; 147: 2735-2742 [PMID: 32399975 DOI: 10.1002/ijc.33048]
- 148 Sfanos KS, Yegnasubramanian S, Nelson WG, De Marzo AM. The inflammatory microenvironment and microbiome in prostate cancer development. Nat Rev Urol 2018; 15: 11-24 [PMID: 29089606 DOI: 10.1038/nrurol.2017.167]
- Sfanos KS, Joshu CE. IBD as a risk factor for prostate cancer: what is the link? Nat Rev Urol 2019; 149 16: 271-272 [PMID: 30742047 DOI: 10.1038/s41585-019-0157-7]
- 150 Porter CM, Shrestha E, Peiffer LB, Sfanos KS. The microbiome in prostate inflammation and prostate cancer. Prostate Cancer Prostatic Dis 2018; 21: 345-354 [PMID: 29795140 DOI: 10.1038/s41391-018-0041-1
- De Marzo AM, Platz EA, Sutcliffe S, Xu J, Grönberg H, Drake CG, Nakai Y, Isaacs WB, Nelson 151 WG. Inflammation in prostate carcinogenesis. Nat Rev Cancer 2007; 7: 256-269 [PMID: 17384581 DOI: 10.1038/nrc2090]
- 152 Lippi G, Montagnana M, Guidi GC. Epidemiological association between C-reactive protein and prostate-specific antigen. Cancer 2009; 115: 1132 [PMID: 19156910 DOI: 10.1002/cncr.24116]
- 153 Lehrer S, Diamond EJ, Mamkine B, Droller MJ, Stone NN, Stock RG. C-reactive protein is significantly associated with prostate-specific antigen and metastatic disease in prostate cancer. BJU Int 2005; 95: 961-962 [PMID: 15839913 DOI: 10.1111/j.1464-410X.2005.05447.x]
- 154 Beer TM, Lalani AS, Lee S, Mori M, Eilers KM, Curd JG, Henner WD, Ryan CW, Venner P, Ruether JD, Chi KN; ASCENT Investigators. C-reactive protein as a prognostic marker for men with androgen-independent prostate cancer: results from the ASCENT trial. Cancer 2008; 112: 2377-2383 [PMID: 18428198 DOI: 10.1002/cncr.23461]
- 155 Sevcenco S, Mathieu R, Baltzer P, Klatte T, Fajkovic H, Seitz C, Karakiewicz PI, Rouprêt M, Rink M, Kluth L, Trinh QD, Loidl W, Briganti A, Scherr DS, Shariat SF. The prognostic role of preoperative serum C-reactive protein in predicting the biochemical recurrence in patients treated with radical prostatectomy. Prostate Cancer Prostatic Dis 2016; 19: 163-167 [PMID: 26810014 DOI: 10.1038/pcan.2015.60]
- 156 Nguyen DP, Li J, Tewari AK. Inflammation and prostate cancer: the role of interleukin 6 (IL-6). BJU Int 2014; 113: 986-992 [PMID: 24053309 DOI: 10.1111/bju.12452]
- Culig Z, Puhr M. Interleukin-6 and prostate cancer: Current developments and unsolved questions. 157 Mol Cell Endocrinol 2018; 462: 25-30 [PMID: 28315704 DOI: 10.1016/j.mce.2017.03.012]
- Shariat SF, Kattan MW, Traxel E, Andrews B, Zhu K, Wheeler TM, Slawin KM. Association of 158 pre- and postoperative plasma levels of transforming growth factor beta(1) and interleukin 6 and its soluble receptor with prostate cancer progression. Clin Cancer Res 2004; 10: 1992-1999 [PMID: 15041717 DOI: 10.1158/1078-0432.ccr-0768-03]
- Rais R, Jiang W, Zhai H, Wozniak KM, Stathis M, Hollinger KR, Thomas AG, Rojas C, Vornov JJ, 159 Marohn M, Li X, Slusher BS. FOLHI/GCPII is elevated in IBD patients, and its inhibition ameliorates murine IBD abnormalities. JCI Insight 2016; 1 [PMID: 27536732 DOI: 10.1172/ici.insight.88634]
- 160 Date AA, Rais R, Babu T, Ortiz J, Kanvinde P, Thomas AG, Zimmermann SC, Gadiano AJ, Halpert G, Slusher BS, Ensign LM. Local enema treatment to inhibit FOLH1/GCPII as a novel therapy for inflammatory bowel disease. J Control Release 2017; 263: 132-138 [PMID: 28159515 DOI: 10.1016/j.jconrel.2017.01.036
- 161 Zhang T, Song B, Zhu W, Xu X, Gong QQ, Morando C, Dassopoulos T, Newberry RD, Hunt SR, Li E. An ileal Crohn's disease gene signature based on whole human genome expression profiles of disease unaffected ileal mucosal biopsies. PLoS One 2012; 7: e37139 [PMID: 22606341 DOI: 10.1371/journal.pone.0037139
- 162 Yao V, Parwani A, Maier C, Heston WD, Bacich DJ. Moderate expression of prostate-specific membrane antigen, a tissue differentiation antigen and folate hydrolase, facilitates prostate carcinogenesis. Cancer Res 2008; 68: 9070-9077 [PMID: 18974153 DOI: 10.1158/0008-5472.CAN-08-2328
- Kaittanis C, Andreou C, Hieronymus H, Mao N, Foss CA, Eiber M, Weirich G, Panchal P, Gopalan 163 A, Zurita J, Achilefu S, Chiosis G, Ponomarev V, Schwaiger M, Carver BS, Pomper MG, Grimm J. Prostate-specific membrane antigen cleavage of vitamin B9 stimulates oncogenic signaling through metabotropic glutamate receptors. J Exp Med 2018; 215: 159-175 [PMID: 29141866 DOI: 10.1084/jem.20171052
- 164 Ge Y, Shi Q, Yao W, Cheng Y, Ma G. The association between inflammatory bowel disease and prostate cancer risk: a meta-analysis. Prostate Cancer Prostatic Dis 2020; 23: 53-58 [PMID: 31591455 DOI: 10.1038/s41391-019-0177-7]



- 165 Chen M, Yuan C, Xu T. An increase in prostate cancer diagnosis during inflammatory bowel disease: A systematic review and meta-analysis. Clin Res Hepatol Gastroenterol 2020; 44: 302-309 [PMID: 31447293 DOI: 10.1016/j.clinre.2019.07.003]
- 166 Carli E, Caviglia GP, Pellicano R, Fagoonee S, Rizza S, Astegiano M, Saracco GM, Ribaldone DG. Incidence of Prostate Cancer in Inflammatory Bowel Disease: A Meta-Analysis. Medicina (Kaunas) 2020; 56 [PMID: 32545154 DOI: 10.3390/medicina56060285]
- 167 Long MD, Herfarth HH, Pipkin CA, Porter CQ, Sandler RS, Kappelman MD. Increased risk for non-melanoma skin cancer in patients with inflammatory bowel disease. Clin Gastroenterol Hepatol 2010; 8: 268-274 [PMID: 20005977 DOI: 10.1016/j.cgh.2009.11.024]
- Singh H, Nugent Z, Demers AA, Bernstein CN. Increased risk of nonmelanoma skin cancers among 168 individuals with inflammatory bowel disease. Gastroenterology 2011; 141: 1612-1620 [PMID: 21806945 DOI: 10.1053/j.gastro.2011.07.039]
- 169 Long MD, Martin CF, Pipkin CA, Herfarth HH, Sandler RS, Kappelman MD. Risk of melanoma and nonmelanoma skin cancer among patients with inflammatory bowel disease. Gastroenterology 2012; 143: 390-399.e1 [PMID: 22584081 DOI: 10.1053/j.gastro.2012.05.004]
- 170 Brin L, Zubair AS, Brewer JD. Optimal management of skin cancer in immunosuppressed patients. Am J Clin Dermatol 2014; 15: 339-356 [PMID: 25015705 DOI: 10.1007/s40257-014-0085-5]
- 171 Peyrin-Biroulet L, Khosrotehrani K, Carrat F, Bouvier AM, Chevaux JB, Simon T, Carbonnel F, Colombel JF, Dupas JL, Godeberge P, Hugot JP, Lémann M, Nahon S, Sabaté JM, Tucat G, Beaugerie L; Cesame Study Group. Increased risk for nonmelanoma skin cancers in patients who receive thiopurines for inflammatory bowel disease. Gastroenterology 2011; 141: 1621-28.e1 [PMID: 21708105 DOI: 10.1053/j.gastro.2011.06.050]
- Magro F, Peyrin-Biroulet L, Sokol H, Aldeger X, Costa A, Higgins PD, Joyce JC, Katsanos KH, 172 Lopez A, de Xaxars TM, Toader E, Beaugerie L. Extra-intestinal malignancies in inflammatory bowel disease: results of the 3rd ECCO Pathogenesis Scientific Workshop (III). J Crohns Colitis 2014; 8: 31-44 [PMID: 23721759 DOI: 10.1016/j.crohns.2013.04.006]
- Kubica AW, Brewer JD. Melanoma in immunosuppressed patients. Mayo Clin Proc 2012; 87: 991-173 1003 [PMID: 23036673 DOI: 10.1016/j.mayocp.2012.04.018]
- 174 Dillon P, Thomas N, Sharpless N, Collichio F. Regression of advanced melanoma upon withdrawal of immunosuppression: case series and literature review. Med Oncol 2010; 27: 1127-1132 [PMID: 19890737 DOI: 10.1007/s12032-009-9348-z]
- 175 van Schaik FD, van Oijen MG, Smeets HM, van der Heijden GJ, Siersema PD, Oldenburg B. Risk of nonmelanoma skin cancer in patients with inflammatory bowel disease who use thiopurines is not increased. Clin Gastroenterol Hepatol 2011; 9: 449-50.e1; author reply 450 [PMID: 21296187 DOI: 10.1016/j.cgh.2011.01.021]
- 176 Peyrin-Biroulet L, Chevaux JB, Bouvier AM, Carrat F, Beaugerie L. Risk of melanoma in patients who receive thiopurines for inflammatory bowel disease is not increased. Am J Gastroenterol 2012; 107: 1443-1444 [PMID: 22951883 DOI: 10.1038/ajg.2012.181]
- Abbas AM, Almukhtar RM, Loftus EV Jr, Lichtenstein GR, Khan N. Risk of melanoma and non-177 melanoma skin cancer in ulcerative colitis patients treated with thiopurines: a nationwide retrospective cohort. Am J Gastroenterol 2014; 109: 1781-1793 [PMID: 25244964 DOI: 10.1038/ajg.2014.298]
- McKenna MR, Stobaugh DJ, Deepak P. Melanoma and non-melanoma skin cancer in inflammatory 178 bowel disease patients following tumor necrosis factor- α inhibitor monotherapy and in combination with thiopurines: analysis of the Food and Drug Administration Adverse Event Reporting System. J Gastrointestin Liver Dis 2014; 23: 267-271 [PMID: 25267954 DOI: 10.15403/jgld.2014.1121.233.mrmk]
- 179 Kopylov U, Vutcovici M, Kezouh A, Seidman E, Bitton A, Afif W. Risk of Lymphoma, Colorectal and Skin Cancer in Patients with IBD Treated with Immunomodulators and Biologics: A Quebec Claims Database Study. Inflamm Bowel Dis 2015; 21: 1847-1853 [PMID: 25993693 DOI: 10.1097/MIB.00000000000457]
- Scott FI, Mamtani R, Brensinger CM, Haynes K, Chiesa-Fuxench ZC, Zhang J, Chen L, Xie F, Yun 180 H, Osterman MT, Beukelman T, Margolis DJ, Curtis JR, Lewis JD. Risk of Nonmelanoma Skin Cancer Associated With the Use of Immunosuppressant and Biologic Agents in Patients With a History of Autoimmune Disease and Nonmelanoma Skin Cancer. JAMA Dermatol 2016: 152: 164-172 [PMID: 26510126 DOI: 10.1001/jamadermatol.2015.3029]
- 181 Nissen LHC, Pierik M, Derikx LAAP, de Jong E, Kievit W, van den Heuvel TRA, van Rosendael AR, Plasmeijer EI, Dewint P, Verhoeven RHA, Overbeek LIH, Nagtegaal ID, Hoentjen F, van der Meulen-de Jong AE. Risk Factors and Clinical Outcomes in Patients with IBD with Melanoma. Inflamm Bowel Dis 2017; 23: 2018-2026 [PMID: 28837522 DOI: 10.1097/MIB.00000000001191
- 182 Clowry J, Sheridan J, Healy R, Deady S, Keegan D, Byrne K, Cullen G, Mulcahy H, Comber H, Parnell AC, Doherty G, Lally A. Increased non-melanoma skin cancer risk in young patients with inflammatory bowel disease on immunomodulatory therapy: a retrospective single-centre cohort study. J Eur Acad Dermatol Venereol 2017; 31: 978-985 [PMID: 28045204 DOI: 10.1111/jdv.14105]
- Khan N, Patel D, Trivedi C, Kavani H, Medvedeva E, Pernes T, Xie D, Lewis J, Yang YX. 183 Repeated Occurrences of Basal Cell Cancer in Patients With Inflammatory Bowel Disease Treated With Immunosuppressive Medications. Am J Gastroenterol 2020; 115: 1246-1252 [PMID:



32453047 DOI: 10.14309/ajg.0000000000000679]

- 184 Ariyaratnam J, Subramanian V. Association between thiopurine use and nonmelanoma skin cancers in patients with inflammatory bowel disease: a meta-analysis. Am J Gastroenterol 2014; 109: 163-169 [PMID: 24419479 DOI: 10.1038/ajg.2013.451]
- 185 Singh S, Nagpal SJ, Murad MH, Yadav S, Kane SV, Pardi DS, Talwalkar JA, Loftus EV Jr. Inflammatory bowel disease is associated with an increased risk of melanoma: a systematic review and meta-analysis. Clin Gastroenterol Hepatol 2014; 12: 210-218 [PMID: 23644389 DOI: 10.1016/j.cgh.2013.04.033]
- 186 Esse S, Mason KJ, Green AC, Warren RB. Melanoma Risk in Patients Treated With Biologic Therapy for Common Inflammatory Diseases: A Systematic Review and Meta-analysis. JAMA Dermatol 2020; 156: 787-794 [PMID: 32432649 DOI: 10.1001/jamadermatol.2020.1300]
- 187 Annese V, Beaugerie L, Egan L, Biancone L, Bolling C, Brandts C, Dierickx D, Dummer R, Fiorino G, Gornet JM, Higgins P, Katsanos KH, Nissen L, Pellino G, Rogler G, Scaldaferri F, Szymanska E, Eliakim R; ECCO. European Evidence-based Consensus: Inflammatory Bowel Disease and Malignancies. J Crohns Colitis 2015; 9: 945-965 [PMID: 26294789 DOI: 10.1093/ecco-jcc/jjv141]
- Beaugerie L, Carrat F, Colombel JF, Bouvier AM, Sokol H, Babouri A, Carbonnel F, Laharie D, 188 Faucheron JL, Simon T, de Gramont A, Peyrin-Biroulet L; CESAME Study Group. Risk of new or recurrent cancer under immunosuppressive therapy in patients with IBD and previous cancer. Gut 2014; 63: 1416-1423 [PMID: 24162591 DOI: 10.1136/gutjnl-2013-305763]
- Shelton E, Laharie D, Scott FI, Mamtani R, Lewis JD, Colombel JF, Ananthakrishnan AN. Cancer 189 Recurrence Following Immune-Suppressive Therapies in Patients With Immune-Mediated Diseases: A Systematic Review and Meta-analysis. Gastroenterology 2016; 151: 97-109.e4 [PMID: 27039969 DOI: 10.1053/j.gastro.2016.03.037]
- 190 Anderson A, Ferris LK, Click B, Ramos-Rivers C, Koutroubakis IE, Hashash JG, Dunn M, Barrie A, Schwartz M, Regueiro M, Binion DG. Low Rates of Dermatologic Care and Skin Cancer Screening Among Inflammatory Bowel Disease Patients. Dig Dis Sci 2018; 63: 2729-2739 [PMID: 29713987 DOI: 10.1007/s10620-018-5056-x]



0 WŨ

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 1981-1996

DOI: 10.4251/wjgo.v13.i12.1981

ISSN 1948-5204 (online)

REVIEW

Mesenchymal stem cell-derived exosomes for gastrointestinal cancer

Lin-Xian Zhao, Kai Zhang, Bing-Bing Shen, Jian-Nan Li

ORCID number: Lin-Xian Zhao 0000-0003-2436-6966; Kai Zhang 0000-0002-4499-7186; Bing-Bing Shen 0000-0002-9358-3844; Jian-Nan Li 0000-0001-9744-7666.

Author contributions: Zhao LX wrote the paper; Zhang K and Shen BB collected the data; Li JN designed the review and revised the paper; Zhao LX, Zhang K, Shen BB, and Li JN performed the approval of final revision; Zhao LX and Zhang K contributed equally to this work.

Conflict-of-interest statement: The authors declare that there is no conflict of interest for this manuscript.

Supported by Science and Technology Development Project of Jilin Province, No. 3D5197434429; and Youth Program of the National Natural Science Foundation of China, No. 3A4205367429.

Country/Territory of origin: China

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Unsolicited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Lin-Xian Zhao, Kai Zhang, Jian-Nan Li, Department of General Surgery, The Second Hospital of Jilin University, Changchun 130041, Jilin Province, China

World Journal of **Gastrointestinal**

Oncology

Bing-Bing Shen, Department of Hepatobiliary Surgery, Renmin Hospital of Wuhan University, Wuhan 430060, Hubei Province, China

Corresponding author: Jian-Nan Li, MD, PhD, Assistant Professor, Department of General Surgery, The Second Hospital of Jilin University, No. 218 Ziqiang Street, Changchun 130041, Jilin Province, China. jnli@ciac.ac.cn

Abstract

Gastrointestinal (GI) malignancies, a series of malignant conditions originating from the digestive system, include gastric cancer, hepatocellular carcinoma, pancreatic cancer, and colorectal cancer. GI cancers have been regarded as the leading cancer-related cause of death in recent years. Therefore, it is essential to develop effective treatment strategies for GI malignancies. Mesenchymal stem cells (MSCs), a type of distinct non-hematopoietic stem cells and an important component of the tumor microenvironment, play important roles in regulating GI cancer development and progression through multiple mechanisms, such as secreting cytokines and direct interactions. Currently, studies are focusing on the anti-cancer effect of MSCs on GI malignancies. However, the effects and functional mechanisms of MSC-derived exosomes on GI cancer are less studied. MSC-derived exosomes can regulate GI tumor growth, drug response, metastasis, and invasion through transplanting proteins and miRNA to tumor cells to activate the specific signal pathway. Besides, the MSC-derived exosomes are also seen as an important drug delivery system and have shown potential in anti-cancer treatment. This study aims to summarize the effect and biological functions of MSC-derived exosomes on the development of GI cancers and discuss their possible clinical applications for the treatment of GI malignancies.

Key Words: Mesenchymal stem cells; Exosomes; Gastrointestinal cancer; Cancer treatment; Drug delivery system; Transplanting miRNA

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.



Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C, C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: April 29, 2021 Peer-review started: April 29, 2021 First decision: June 5, 2021 Revised: August 15, 2021 Accepted: September 8, 2021 Article in press: September 8, 2021 Published online: December 15, 2021

P-Reviewer: Kida YS, Saleh F S-Editor: Yan JP L-Editor: Wang TQ P-Editor: Guo X



Core Tip: Mesenchymal stem cells (MSCs) have shown potential for anti-cancer therapy. As an important content of MSCs, MSC-derived exosomes are attracting more and more researchers for anti-cancer studies. We herein summarize the effect of MSCderived exosomes on gastrointestinal malignancies and discuss their therapeutic potential.

Citation: Zhao LX, Zhang K, Shen BB, Li JN. Mesenchymal stem cell-derived exosomes for gastrointestinal cancer. World J Gastrointest Oncol 2021; 13(12): 1981-1996 URL: https://www.wjgnet.com/1948-5204/full/v13/i12/1981.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.1981

INTRODUCTION

Gastrointestinal (GI) cancer is one of the most common malignancies in the digestive system, such as the stomach, liver, pancreas, and colorectum^[1]. Based on the latest global epidemiological data, GI cancer accounts for 26% of all kinds of cancers and 35% of cancer patients died from GI cancer with approximately 4.8 million new cases and 3.4 million deaths each year [2,3]. Current therapeutic strategies for GI malignancies mainly include surgery, endoscopy, radiotherapy, chemotherapy, targeted therapy, and immunotherapy[4-6]. Despite that treatment strategies are becoming mature and diverse, the prognosis of GI cancer is still very poor due to the fact that most patients miss the therapeutic window[7]. If detected at an early stage, GI cancers are highly curable with traditional treatment methods^[8]. However, the early diagnosis of GI cancer is a significant challenge. Therefore, more promising treatment strategies are needed to cut down the mortality of GI malignancies.

Mesenchymal stem cells (MSCs), a type of distinct non-hematopoietic stem cells, possess the capacity of self-renewal and multipotentiality differentiation [9,10]. It has been shown that MSCs are involved in tumor development, including tumorigenesis, tumor growth and metastasis, as well as regulation of tumor microenvironment[11-13]. Therefore, MSCs have been commonly used in anti-cancer studies. However, the underlying mechanisms of how MSCs affect tumor development are still controversial [14]. A series of literature has reported that MSCs are capable of promoting tumor progression through secreting pro-tumorigenic factors[15] and differentiating into cancer-associated fibroblasts[16,17]. Nevertheless, other evidence suggests that MSCs could suppress tumor proliferation via secreting cycle inhibitor P21 and antitumorigenic factors such as interleukins, IFN- γ , Dkk-1, and promote tumor cell apoptosis through secreting apoptotic executor caspase 3[18,19]. Although the underlying mechanisms of how MSCs regulate tumor cells are still unclear, there is no doubt that MSC-derived exosomes play a key role in the interaction between MSCs and tumor cells^[20].

Exosomes, a distinct population of extracellular vesicles, are vital for cell-to-cell communication^[21]. Exosomes can be derived from mesenchymal cells, immune cells, and tumor cells and the effects of exosomes of different sources are distinct^[22]. MSCderived exosomes show many similar effects with MSCs, and have also been seen as an important component of the tumor microenvironment[13,23]. More importantly, compared to MSCs, MSC-derived exosomes are safer and show better penetrability, biocompatibility, and stability during the interaction with tumor cells[24,25]. In recent years, the functions of MSC-derived exosomes in GI treatment have been studied with in vitro experiments and animal models[21]. However, a systematic review is rare. This paper summarizes the features of exosomes and the effects of MSC-derived exosomes on GI cancers. Besides, the therapeutic potential of MSC-derived exosomes on GI malignancies is highlighted and future research opportunities related to MSC-derived exosomes are also proposed.

MESENCHYMAL STEM CELLS

MSCs, with anticancer, angiogenic, anti-apoptotic, and multi-differentiation capacity, have been commonly used in oncotherapy, and tissue regeneration and restoration[6, 26]. Based on the sources, MSCs can be divided into bone marrow-derived MSCs,



embryo-derived MSCs, human umbilical cord-derived MSCs, adipose tissue-derived MSCs, dental MSCs, and menstrual blood-derived MSCs[27-29]. It has also been shown that MSCs are key mediators of inflammation and the tumor microenvironment [30]. Based on these discoveries, a new clinical treatment strategy called cell therapy has been developed, which works *via* transplanting MSCs into human bodies to treat related diseases. At present, cell therapy based on MSCs is still in the clinical trial phase. Although MSC transplantation has shown huge potential in clinical application, more and more side effects and limitations have been found. For example, it has been proposed that MSC transplantation could increase the risk of tumorigenicity and cell death[31]. Besides, MSCs are limited by the lung barrier[32]. To solve these problems, researchers have proposed to replace MSCs with MSC-derived exosomes for cell therapies, because these exosomes show many similar functions with MSCs, are safer and more stable, and can be used as a vehicle to deliver anti-tumor drugs and bioactive factors[33]. In addition, it has been discovered that the MSC-derived exosomes could regulate tumor progression via changing the microenvironment of tumors[34].

EXOSOMES

Exosomes, cell-derived membranous structures, originate from the invagination of the endosomal system or segregation of the plasma membrane [35]. Abundant biomolecules such as biomarker proteins, regulatory RNA and DNA, functional cytokines, growth factors, etc. [36,37], are included in exosomes. To date, it has been discovered that MSC-derived exosomes contain more than 300 miRNAs and at least 730 proteins[38,39]. The sizes of exosomes are from 50 nm to 200 nm, which play a central role in cell-to-cell signaling networks[36,40]. One study has reported that exosomes are capable of regulating the pathway of downstream signals of recipient cells via releasing a variety of biomolecules and transporting the genetic material to downstream cells^[28]. Interestingly, exosomes can play a dual role in tumorigenesis, both anti-tumor and pro-tumor, which may be because the exosomes can be derived from different tissues [41]. For instance, one study has proposed that normal tissue MSC-derived exosomes are capable of suppressing tumor development through blocking carcinogenic reprogramming signaling pathways. In contrast, tumor cellderived exosomes can drive recipient cells to establish malignancy, resulting in tumorigenesis[42]. The main sources of exosomes include MSCs, immune cells, tumor cells, etc. [22]. MSC-derived exosomes have been commonly used in the studies of cancer development. Yang et al[43] have proposed that MSC-derived exosomes could promote tumor growth through secreting matrix metalloproteinase-2 (MMP-2) or MMP-2 enzyme to alter the tumor microenvironment and cellular functionalities. It has also been found that MSC-derived exosomes are capable of supporting tumor growth via transporting tumor-supportive factors such as proteins, miRNA, and metabolites to recipient tumor cells[44]. Besides, MSC-derived exosomes can suppress tumor growth by carrying tumor-inhibiting factors into tumor cells and decreasing the expression of vascular endothelial growth factor (VEGF)[45].

Because of the special properties and biological functions, exosomes have been used as natural nanocarriers to transport drugs and specific factors to tumor sites [46]. For example, one study has indicated that MSC-derived exosomes could reach a higher cell-target specificity by delivering paclitaxel (PTX) to tumor sites[47]. In addition, the glioma-associated MSC-derived exosomes could deliver miR-1587 to recipient glioma stem-like cells, increasing the proliferation and aggressiveness of glioblastoma through down-regulating the expression of the tumor-suppressor NCOR1[48]. Furthermore, it has also been demonstrated that MSC-derived exosomes could improve anti-cancer therapeutic efficacy through regulating immune response and reversing the chemoresistance^[49,50]. The following section describes the effect of MSC-derived exosomes on gastric cancer (GC), hepatoma, pancreatic cancer (PC), and colorectal cancer (CRC) (Table 1).

MSC-DERIVED EXOSOMES FOR GC

GC is the fourth most common malignant neoplasm and the third prominent cause of cancer death globally[51]. Despite routine gastroscopy increasing the rate of early diagnosis, the 5-year survival rate of GC patients is still less than 30% [52]. In the current treatments for GC, perioperative or adjuvant chemotherapy can significantly



Table 1 Effect of mesenchymal stem cells-derived exosomes on gastrointestinal cancer

Tumor type	Exosomes source	Cell lines	Function	Mechanism	Ref.
Gastric cancer	hUCMSCs	HGC-27; MGC- 803; SGC-7901	Conferring tumor chemoresistance	(1) Upregulating the expression of multi-drug resistance-associated genes and proteins; (2) Activating calcium/calmodulin-dependent protein kinases (CaMKs) and Raf/MEK/ERK pathway; and (3) Enhancing the functionality of P-gp/MDR	[13]
	hBMSCs	Animal model	Promoting tumor development	(1) Activating ERK1/2 and p38 MAPK pathways; and (2) Enhancing the expression of VEGF	[55]
	hBMSCs	SGC-7901	No effect	NA	[<mark>55</mark>]
	hUCMSCs	HGC-27	Promoting tumor development	(1) Activating the Akt signal pathway; (2) Inducing the epithelial- mesenchymal transition (EMT); and (3) Enhancing the tumorigenicity and stemness	[<mark>61</mark>]
	hBMSCs	SGC-7901	Promoting tumor development	Secreting miR-221 to activate Hedgehog signaling pathway	[64]
	GC-MSCs	HGC-27	Promoting tumor development	Increasing the expression of miR-214, miR-221, and miR-222	[<mark>66</mark>]
	mBMSCs	MFC	Promoting tumor development	Delivering UBR2 to activate Wnt/β -catenin signaling pathway	[<mark>69</mark>]
Liver cancer	hBMSCs	HepG-2	Inhibiting tumor development	(1) Blocking the cell cycle progression; and (2) Inducing tumor cells apoptosis	[<mark>93</mark>]
	AMSCs	HepG-2	Inhibiting tumor development and increasing tumor chemosensitivity	Secreting miR-122 to improve chemosensitivity of HepG2 HCC cells and inhibiting tumor development	[83]
	AMSCs	Huh-7; SMMC- 7721	Increasing tumor chemosensitivity	Delivering miR-199a-3p to improve liver cancer cell line chemosensitivity	[87]
	mBMSCs	Animal model	Inhibiting tumor development	(1) Promoting tumor cells apoptosis; and (2) Inhibiting angiogenetic activity, metastasis, and invasiveness	[96]
	AMSCs	Animal model	Inhibiting tumor development	Upregulating local and systemic NK cells	[95]
	AMSCs	Huh-7; SMMC- 7721	Increasing tumor chemosensitivity	Delivering miR-199a-3p to tumor sites	[87]
Pancreatic cancer	hBMSCs	BxPC-3; PANC-1	Inhibiting tumor development	Secreting miR-1231 to suppress tumor development	[107]
	hBMSCs	PANC-1	Inhibiting tumor development and promoting tumor cells apoptosis	Downregulating the expression of a disintegrin and a metalloproteinase-9 (ADAM9)	[108]
	mBMSCs	AsPC-1; PANC-1	Inhibiting tumor development and promoting tumor cells apoptosis	Delivering miR-124 to regulate the expression of EZH2	[109]
	mBMSCs	CFPAC-1	Inhibiting tumor development	Delivering anticancer agents	[47]
	Normal fibroblast-like MSCs	PANC-1	Inhibiting tumor development	Delivering short interfering RNA or short hairpin RNA to target oncogenic KRAS	[105]
	BMSCs	MiaPaca-2	Inhibiting tumor development	Loading PTX and gemcitabine monophosphate (GEMP) to pancreatic cancer	[100]
	BMSCs	Tumor model	Enhancing tumor immunotherapy	Constructing a dual delivery biosystem to achieve the combined therapy	[106]
Colorectal cancer	hBMSCs	SW-480	Promoting tumor development	Activating ERK1/2, p38, and JNK pathways	[55]
	BMSCs	Caco-2; SW- 480; SW-620; LoVo; HT-29	Inhibiting tumor development and promoting tumor cells apoptosis	Upregulating the expression of miR-16-5p to downregulate integrin $\alpha 2$ (ITGA2)	[124]
	BMSCs	DLD-1; HCT-	Inhibiting tumor	Secreting miR-4461 to downregulate the expression of COPB2	[125]



	116; SW-480	development		
BMSCs	SW-1116; Caco- 2	Inhibiting tumor development and promoting CSCs phenotype	Secreting miR-142-3p to decrease the expression of Numb. (1) Increasing the expression of Notch target genes; and (2) Secreting miR-142-3p to target CD133 and Lgr5	[126] [127]
hUCMSCs	HT-29; DLD-1	Inhibiting tumor development	(1) Downregulating the expression of Integrin alpha6 (ITGA6); and (2) Inhibiting the activity of transforming growth factor-beta1 (TGF- β 1) signaling pathway	[128]
mBMSCs	C-26; MCF-7	Inhibiting tumor development	Loading doxorubicin (DOX) to tumor cells	[<mark>132</mark>]

MSCs: Mesenchymal stem cells; hBMSCs: Human bone marrow-derived mesenchymal stem cells; AMSC: Adipose-derived mesenchymal stem cells; mBMSCs: Murine bone marrow-derived mesenchymal stem cells; hUCMSCs: Human umbilical cord mesenchymal stem cells; GC-MSCs: Gastric cancer tissue-derived mesenchymal stem cells; MFC: Murine foregastric carcinoma; CSCs: Cancer stem cells; NA: Not available.

> improve the therapeutic effect on advanced GC[53]. However, chemoresistance is one of the major obstacles[54]. One recent study has reported that human umbilical cord MSC-derived exosomes could confer chemoresistance to GC cells (HGC-27, MGC-803, and SGC-7901) through upregulating the expression of multi-drug resistance genes and proteins, activating calcium/calmodulin-dependent protein kinases and the Raf/MEK/ERK pathway, and enhancing the functionality of P-gp/MDR. In this way, GC cells are protected from chemotherapy-induced apoptosis[13]. In other words, the efficacy of chemotherapy in GC treatment can be improved by targeting the interaction between MSC-derived exosomes and tumor cells. For example, chemoresistance in GC can be overcome by blocking the CaM-Ks/Raf/MEK/ERK pathway. In conclusion, the therapeutic potential and efficacy of GC treatment can be improved based on the effects of MSC-derived exosomes on drug resistance.

> The effect of MSC-derived exosomes on GC development remains controversial. In a mouse model experiment, researchers have observed that human bone barrowderived MSCs (hBMSCs)-derived exosomes could promote the growth of SGC-7901 gastric tumor cells[55]. Further studies have indicated that MSC-derived exosomes are capable of promoting the incidence and growth of tumors via activating angiogenesis and facilitating tumor cell proliferation in vivo[56]. After the co-implantation with hBMSC-derived exosomes in vivo, MSC-derived exosomes show a tumor-promoting effect in these rat models, and significant up-expression of Bcl-2, phosphorylated ERK1/2, α-smooth muscle actin (a-SMA), CXCR4, VEGF, and MDM2 mRNA, all of which are very essential for tumor growth, metastasis, and angiogenesis, has been detected in the tumor microenvironment^[55]. In contrast, it has been discovered that hBMSC-exosomes do not affect SGC-7901 cell proliferation in vitro, suggesting that the effect of MSC-derived exosomes on the incidence and growth of tumor is exerted through indirect mechanisms^[55]. To further illustrate the mechanism of how hBMSCexosomes affect tumor growth, researchers have examined the expression levels of VEGF and CXCR4 in vivo. They discovered that hBMSC-exosomes could promote tumor growth via activating the ERK1/2 and p38 MAPK pathways, and therefore the expression of VEGF is upregulated, which, in turn, activates tumor angiogenesis[55-59]. Previous studies have also shown MSC-derived exosomes could increase the expression of octamer-binding transcription factor 4, ex deter mining region Y-box 2, and Lin28B, and therefore promote the formation of tumor blood vessels and potentiate gastric tumor growth[55,60,61]. Further studies have discovered that hUCMSC-derived exosomes can promote HGC-27 gastric tumor cell invasion and metastasis through increasing the expression of mesenchymal indicators, activating the Akt signaling pathway, and decreasing the expression of epithelial indicators, and therefore the epithelial-mesenchymal transition (EMT) of gastric tumor cells is induced [61]. EMT, an initial stage of tumor metastasis, can stimulate tumor cells to lose epithelial cell polarity, render mesenchymal features, infiltrate into adjacent tissues, and increase self-renewal capacity [62,63]. In addition to contributing to obtaining the EMT, hUCMSC-derived exosomes also contribute to enhancing the tumorigenicity and stemness of HGC-27 cells. After the treatment with hUCMSC-derived exosomes, the expression of Oct4, Sox2, and Lin28B is increased, all of which are stemness-relevant indicators^[61].

> Other studies have proposed that BMSC-derived exosomes could secret miR-221 as a pro-tumor molecule to activate the Hedgehog signaling pathway, promoting the proliferation and progression of gastric tumors[64,65]. Furthermore, the miR-221 level in the peripheral blood could also be seen as a GC diagnostic marker, and the high expression level of miR-221 is reckoned as an indicator of poor clinical prognosis of



gastric tumors^[34]. Another study has found that GC tissue-derived MSCs (GC-MSCs) are capable of increasing the expression of miR-214, miR-221, and miR-222, all of which are positively correlated with the development of GC[66]. For instance, the upregulation of miR-214 can be seen as a sign of venous invasion and unfavorable outcome of GC[67]. The high expression of miR-222 is mainly associated with serosal invasion and lymph node metastasis[66] and miR-221 is mainly involved in advanced stages of node metastasis, local invasion, and lymphatic metastasis of GC[68]. Therefore, it has been reported that the tumor-promoting effects of GC-MSCs could be impaired by using a miRNA inhibitor to downregulate the expression of miR-221[66]. Based on the above discoveries, the exosomal miR-214, miR-221 and miR-222 can be used for the early diagnosis and treatment of gastric tumors in the future.

In a recent preclinical study, Mao et al [69] have used the p53 deficient mouse BM-MSC exosomes to deliver UBR2 into murine foregastric carcinoma cells. They found that UBR2 enriched by exosomes could promote the proliferation and migration of these tumor cells through activating the Wnt/ β -catenin signaling pathway. Previous studies have demonstrated that the Wnt/ β -catenin pathway plays a key role in regulating the growth and metastasis of GC cells, and the maintenance of cancer stem cells (CSCs)[70].

MSC-DERIVED EXOSOMES FOR LIVER CANCER

Liver cancer is the second leading cause of cancer-related death worldwide^[71]. Hepatocellular carcinoma (HCC) is the most common type of liver cancer, accounting for more than 90% of cases[72]. Especially, the prognosis of patients with advanced HCC is poor due to the lack of an effective treatment strategy [73,74]. Currently, although many clinical treatments for HCC such as surgical techniques, conventional chemotherapy, transarterial chemoembolization, radiotherapy, targeted therapy, and liver transplantation have been applied, the 5-year survival rate of liver cancer patients is still not more than 20% [75,76]. In recent years, with more studies focusing on exosomes, it has been proposed that exosomes, especially MSC-derived exosomes, have shown substantial anticancer potential in the clinical application, especially in the treatment of HCC[37]

Chemotherapy has been regarded as the most common curative measure for HCC. However, HCC shows high resistance to conventional chemotherapeutic drugs and agents [77]. Therefore, new therapeutic approaches are needed to enhance HCC chemosensitivity. Previous studies have indicated that miR-122 plays a key role in diagnosing and prognosis of hepatoma[19]. For example, it has been proposed that the loss or downregulation of miR-122 could be reckoned as the sign of poor prognosis and metastasis of HCC[78], and is closely related hepatocarcinogenesis and HCC development^[79]. On the other hand, it has been found that miR-122 could increase the sensitivity of liver cancer cells to chemotherapeutic drugs such as 5-fluorouracil (5-FU) and doxorubicin[37,80]. Recent studies have demonstrated that upregulated miR-122 could inhibit the formation and development of HCC, and increase the chemotherapeutic sensitivity of these tumor cells[81,82]. Moreover, another study also reported that adipose MSC (AMSC)-derived exosomes could secret miR-122 to improve chemosensitivity of HepG2 HCC cells[83]. MiR-199a-3p, a highly expressed miRNA in normal liver cells, can increase HCC chemosensitivity by downregulating the gene expression of YAP1, CD151, and mTOR[84-86]. It has been proposed that AMSCderived exosomes could be used to deliver miR-199a-3p to improve the chemosensitivity of Huh7 and SMMC-7721 liver cancer cell lines[87]. According to the above discoveries, there is no doubt that miR-122-modified exosomes and miR-199amodified exosomes are two effective liver cancer treatment alternatives. However, finding a safe and effective vehicle for miR-122 and miR-199a-3p delivery is a challenge and researchers have studied how to use exosomes as a biological delivery vehicle for miRNA transfer[88]. Compared to other vehicles, MSC-derived exosomes possess less immunogenicity, higher biocompatibility, and less toxicity[89]. Besides, as the most prolific producers among exosome-producing cells, MSCs are suited for the mass production of exosomes[90]. All in all, MSC-derived exosomes can be used as a new nanocarrier of miRNAs and drugs[37].

Previous studies have indicated that MSCs can both suppress and promote liver cancer progression[6]. The function of MSC-derived exosomes on HCC progression, similar to that of MSCs, is not determined [91-94]. For example, one study has reported that BM-MSC-derived exosomes can inhibit HepG2 cell growth via blocking the cell cycle progression and inducing apoptosis in vitro[93]. In another study, AMSC-derived



exosomes have been directly injected into nude mice bearing HepG2 cells, and no significant differences compared to the control group have been observed[83]. Moreover, another rat model study has reported that AMSC-derived exosomes could inhibit HCC development through upregulating local and systemic NK cells[95]. The above studies show that mechanisms of the regulation of different MSC-derived exosomes on HCC progression are distinct. Although the experimental results may be interfered by experimental models, tumor types, MSC sources, as well as exosome injection administration, there is no doubt that AMSC-derived exosomes can be effectively used to transfer miR-122 to increase HepG2 liver cancer cell chemosensitivity and inhibit HCC growth and progression, providing a new treatment strategy for HCC[83].

Alzahrani et al[96] have conducted a long-term model study and found that BMSCderived exosomes could inhibit the development of diethylnitrosamine-induced HCC in vivo. After BM-MSC-derived exosomes being injected into established HCC, the overexpression of apoptotic genes, Bax and p53, and the downregulated antiapoptotic gene, Bcl2, were observed. In contrast, CSC-derived exosomes are capable of suppressing apoptosis, increasing angiogenetic activity, promoting metastasis and invasiveness, and inducing EMT.

Liver cancer is a highly angiogenic cancer, whose growth requires sufficient blood supply as nourishment. It is acknowledged that VEGF plays an important role during angiogenesis. MSC-derived exosomes can inhibit tumor angiogenesis by downregulating VEGF[45] and suppressing liver cancer cell progression. Some studies have found that MSC-derived exosomes are beneficial for acute liver injury and liver fibrosis via activating the proliferative and regenerative responses[97,98]. Moreover, it has been reported that tumor-derived exosomes could work with BM-MSCs to inhibit HCC cell growth through arresting these cells in the G0/G1 phase[99]. All in all, MSCderived exosomes have shown unlimited potential in liver cancer treatment, but there is still a long way to go before clinical application.

MSC-DERIVED EXOSOMES FOR PANCREATIC CANCER

PC, especially pancreatic ductal adenocarcinoma (PDAC), is a highly fatal malignancy with a 5-year survival rate less than 6% [100]. To date, despite an increasing number of clinical treatments, surgery remains the only curative treatment for PC. However, the surgical resection rate is only approximately 20% as most patients present transforming diseases upon diagnosis, and therefore chemotherapy remains the main strategy for clinical PC treatment[101]. However, traditional chemotherapy is not effective enough due to chemotherapy resistance, abnormally abundant extracellular matrix, and extremely deficient neovascularization in the tumor microenvironment [102,103]. To overcome the pathophysiological barrier of PC, an increasing number of nanotechnology-based drug delivery strategies have been proposed.

Previous studies have shown that MSCs could regulate the tumorous microenvironment and the development of PDAC[104]. With the deepening studies, it has been discovered that MSC-derived exosomes are capable of circumventing the tumor extracellular matrix barrier, overcoming chemoresistance, and efficiently targeting and penetrating tumor cells[100,105]. Therefore, the MSC-derived exosomes can be seen as novel systems to load chemotherapeutics to target PC. For example, one study has used the MSC-derived exosomes to load PTX and gemcitabine monophosphate homing to PC, and these exosomes show more preferable penetration and superior anti-tumor efficacy than the control group both in vivo and in vitro[100]. In a recent preclinical study, Zhou et al[106] have used the BMSC-derived exosomes to construct a dual delivery biosystem, which is capable of carrying both oxaliplatin (OXA) and siRNA for enhancing PDAC immunotherapy. The siRNA-exosomes-OXA nanoparticles can elicit anti-tumor immunity and exert significant therapeutic effects while showing better stability and fewer side effects than traditional synthetic delivery systems. More specifically, the combined therapy of iEXO-OXA could activate innate and adaptive anti-PDAC immunity by inducing the immunogenic cell death of tumor cells, initiating dendritic cell maturation and antigen presentation, and reversing immunosuppression and recruiting antitumoral cytotoxic T lymphocytes. Based on the findings above, it can be concluded that MSC-derived exosomes can serve as a promising nanoscale drug delivery platform for PC over the long run.

In addition to functioning as a carrier for drug delivery, MSC-derived exosomes can also affect PC progression through secreting multiple miRNAs[107]. For example, the expression level of miR-1231 in exosomes derived from the peripheral blood is

correlated with the pathological stage of PC, suggesting that miR-1231 may benefit PC diagnosis. Further studies have proposed that miR-1231 is capable of inhibiting the growth and development of BxPC-3 and PANC-1 pancreatic tumor cells[107]. Based on the above discoveries, it can be concluded that BMSC-derived exosomes with a high expression level of miR-1231 can be efficiently used in anti-cancer medicines, especially medicines for PC. Wu et al[108] have transfected miR-126-3p into the exosomes of BMSCs and found that the exosomes could downregulate the expression of a disintegrin and a metalloproteinase-9 and promote the apoptosis while suppressing the proliferation, invasion, and metastasis of PANC-1 pancreatic tumor cells. Therefore, the miR-126-3p can be reckoned as a novel biomarker for PC treatment. In a recent study, Xu et al [109] have indicated that miR-124-carried BMSCderived exosomes could inhibit the proliferation, metastasis, and invasion, and induce apoptosis of PC cells (AsPC-1 and PANC1) by regulating the expression of EZH2, which is a target of miR124[110]. Previous studies have also demonstrated that miR-124 serves as a tumor suppressor for many cancers, such as HR-HPV-positive cervical cancer[111], breast cancer[112], and bladder cancer[113]. The above findings suggest that MSC-derived exosomes can be considered as a potential vehicle to transport miR-124 in PC treatment.

MSC-DERIVED EXOSOMES FOR COLORECTAL CANCER

CRC ranks as the third most commonly diagnosed cancer and the most common in GI cancers[71,114]. In recent years, with the improvement of screening tests and therapeutic strategies, the 5-year survival rate of CRC in China has increased to 31% [115]. However, the incidence rates of CRC, especially those in most developing countries, increase sharply due to the lifestyle changes, growing population, and aging of the population[116]. In 2020, the new cases of CRC were more than 1.9 million in 185 countries[71]. Therefore, it is very necessary to explore more effective diagnostic and therapeutic strategies for CRC. Previous studies have pointed out different effects of MSCs on CRC. For example, it has been proposed that hBMSCs could promote the growth of the low-malignancy CRC cell line HT29, but could not affect the progression of the high-malignancy CRC cell line HCT 116[117]. With the discovery of anti-tumor and tumor homing properties, MSCs have been widely used in CRC studies. Despite that MSC therapy in CRC remains controversial due to MSCs can promote immune evasion of tumor cells in the tumor microenvironment, which might be caused by the powerful immunosuppression function of MSCs[118], the application of MSCs is still a promising strategy to ameliorate CRC. First, MSCs are capable of depressing tumor metastasis and complications[119]. For example, one study has reported that MSCs could inhibit CRC metastasis and decrease the formation of malignant ascites by suppressing VEGF expression[120]. It has also been shown that MSCs could inhibit the proliferation of colonic cancer via depressing the expression of proinflammatory factors, ERK, STAT3 phosphorylation, and Smad2, and blocking PI3K/AKT signaling pathway[121-123].

On the other hand, MSC-derived exosomes are also involved in CRC proliferation, migration, and invasion. For example, it has been reported that BMSC-derived exosomes are capable of overexpressing miR-16-5p to downregulate integrin α2 (ITGA2), and inhibiting the growth and progression but promoting the apoptosis of CRC cells (Caco-2, SW480, SW620, LoVo, and HT29)[124]. Therefore, miR-16-5p derived from MSC-derived exosomes can be developed as an effective therapy for CRC. Besides, Chen et al[125] have transfected BMSC-derived exosomes into CRC cells (DLD1, HCT116, and SW480) and found that the proliferation of these cells is inhibited and the content of miR-4461 increases significantly, indicating that exosomic miR-4461 might inhibit the growth of CRC cells. Further studies have proposed that the expression level of miR-4461 is lower in CRC cells than that in normal cells and miR-4461 is capable of downregulating the expression of coatomer protein complex subunit beta 2 (COPB2). Based on these discoveries, it can be concluded that miR-4461 derived from BMSCs exosomes can inhibit CRC tumorigenesis by downregulating the expression of COPB2, a target gene of miR-4461. In the future, miR-4461 can be applied for the diagnosis and treatment of CRC.

Li et al[126] have treated SW1116 and Caco2 colorectal tumor cells with miR-142-3p and found that miR-142-3p could inhibit the proliferation and invasion of CRC cells but increase the population of CSCs of colon cancer. Further studies have found that miR-142-3p promotes colon CSC-like traits by decreasing the expression of Numb while increasing the expression of Notch target genes, such as Hes1, P21, and Cyclin


D3. On the other hand, the underlying mechanism of inhibited tumor proliferation could be that miR-142-3p can target *CD133* and *Lgr5*[127].

Li et al[128] have transfected hUCMSC-derived exosomes containing miR-3940-5p into HT-29 and DLD-1 colorectal tumor cells and found that the exosomes suppress EMT, metastasis, progression, and invasion of these CRC cells by downregulating the expression of Integrin alpha6 (ITGA6) and inhibiting the activity of transforming growth factor-beta1 (TGF- β 1) signaling pathway. Previous studies have indicated that overexpression of ITGA6 could trigger CRC progression and migration via upregulating transforming growth factor-beta1 (TGF-β1)[129,130].

Significant progress has been made in the development of an efficient vehicle for the delivery of anticancer agents to tumor tissue. Similar to many kinds of natural exosomes, MSC-derived exosomes also possess many distinctive characteristics such as good stability, low toxicity and immunogenicity, good biocompatibility, and long circulation[131]. Therefore, Bagheri et al[132] have loaded doxorubicin into MSCderived exosomes using the electroporation method and found that MSC-derived exosomes inhibit the growth of C26 and MCF7 colon tumor cells more significantly and have proposed that MSC-derived exosomes can be used to construct a novel biomanufacturing drug delivery platform for CRC therapy. One study has proposed that MSC-derived exosomes can also be used in inflammatory bowel disease (IBD) treatment[133] as treatment with MSC-exosomes substantially mitigates IBD through inhibiting inflammatory responses, maintaining intestinal barrier integrity, and polarizing M2b macrophages.

DISCUSSION

It can be concluded from the above findings that MSC-derived exosomes have shown unlimited therapeutic potential for GI cancer treatment. The main methods for developing new treatment strategies are summarized as: (1) To use the nature contents of MSC-derived exosomes to inhibit tumor proliferation and invasion. To date, it has been shown that the main inhibitory factors are some miRNAs and proteins, but the specific mechanisms have not been found out [44,134]. Therefore, in future studies, more efforts are needed to illustrate the possible mechanism; (2) to target the interaction between MSC-derived exosomes and tumor cells. It has been shown that MSC-derived exosomes could promote the growth of some GI cell lines and increase the chemoresistance of these cell lines through upregulating the expression of the factors and proteins or activating some special signal pathways^[13]. Therefore, relative receptors can be targeted and the relative pathway can be blocked to improve therapeutic effectiveness; and (3) to modify MSC-derived exosomes as a drug delivery carrier. After being modified with special anti-cancer drugs, these exosomes are capable of homing to tumor sites with less immunogenicity.

Increasing studies have demonstrated that MSC-derived exosomes could exert both anti-tumor and pro-tumor effects on GI malignancies[42-44]. The reasons why MSCderived exosomes can play different functions in the development of GI cancers are concluded as: (1) The tumor cell lines chosen for the experimental research are different. For example, different types of CRC cells lines, such as Caco-2, SW-480, SW-620, HT-29, HCT-116, and DLD-1, are used in CRCs studies and the experimental results are different, which may be due to that different types of tumor cell lines show different invasion, metastasis, and proliferation capability[135]; (2) the sources of exosomes are different, and the contents, such as factors, signaling lipids, proteins, and miRNAs, of different types of MSC-derived exosomes are distinct. Therefore MSCderived exosomes can crosstalk with tumor cells through different mechanisms and exert different effects on tumor development; and (3) the experimental methods and models are different. The in vitro and in vivo studies can show different and even opposite results. Besides, the tumor microenvironment and cell cultivation conditions can both influence the experimental results.

Despite that both MSCs and MSC-derived exosomes can be used in anti-cancer research, MSC-derived exosomes show many potential advantages. First, MSC transplantation may result in the transfer of mutated or damaged DNA into normal cells, and an increasing risk of a new disease[136]. Fortunately, if MSC-derived exosomes are directly transferred into the body, these problems can be effectively avoided. Second, with smaller sizes, MSC-derived exosomes can circulate and pass through various barriers, such as capillary bed and lung barriers easily. Third, with the same infusion dose, the effect of MSCs-derived exosomes can be kept for a longer time than MSCs post-transplant, which can achieve a greater circulation extent[137].

To date, despite that MSC-derived exosomes have showing substantial therapeutic potential in GI treatment, many challenges and obstacles need to be overcome. The most common obstacle is to achieve large-scale production of MSC-derived exosomes. In addition, isolating these exosomes from MSCs without modification of the cargos of these MSCs is also a big challenge. Furthermore, because the sources or donors of MSCs are different, MSC-derived exosomes show heterogeneity and even the exosomes derived from the same type of MSCs can exert opposite effects on tumor development, which might be due to the fact that these exosomes carry di \Box erent molecules. Therefore, before applying MSC-derived exosomes in clinical trials, researchers need to improve the methods for mass-production, isolation, and homogeneity maintenance of MSC-derived exosomes[138]. The internal living conditions of MSCs can be simulated to achieve a function-specific and large-scale production of MSC-derived exosomes. Besides, more methods for storing and recovering these MSC-derived exosomes and a potency assay for therapeutic efficacy evaluation of exosomes are needed. Based on MSC-based clinical trials, MSC-derived exosome therapies can be developed more rapidly.

CONCLUSION

This review analyzes the effects of MSCs-derived, hBMSC-derived exosomes, mBMSCderived exosomes, hUCMSC-derived exosomes, and GC-MSC-derived exosomes on GI malignancy development. However, the reasons why different MSC-derived exosomes exert distinct effects on GI malignancies are not determined. In the future, a better understanding of the mechanisms of how MSC-derived exosomes regulate GI cancer development is needed, which will help to develop more promising treatment methods for GI cancer.

REFERENCES

- 1 Arnold M, Abnet CC, Neale RE, Vignat J, Giovannucci EL, McGlynn KA, Bray F. Global Burden of 5 Major Types of Gastrointestinal Cancer. Gastroenterology 2020; 159: 335-349.e15 [PMID: 32247694 DOI: 10.1053/j.gastro.2020.02.068]
- 2 Ferlay J, Colombet M, Soerjomataram I, Parkin DM, Piñeros M, Znaor A, Bray F. Cancer statistics for the year 2020: An overview. Int J Cancer 2021 [PMID: 33818764 DOI: 10.1002/ijc.33588]
- Ramai D, Heaton J, Ghidini M, Chandan S, Barakat M, Dhindsa B, Dhaliwal A, Facciorusso A. Population-Based Long-term Cardiac-Specific Mortality Among Patients With Major Gastrointestinal Cancers. JAMA Netw Open 2021; 4: e2112049 [PMID: 34137831 DOI: 10.1001/jamanetworkopen.2021.12049]
- 4 Rao D, Parakrama R, Augustine T, Liu Q, Goel S, Maitra R. Immunotherapeutic advances in gastrointestinal malignancies. NPJ Precis Oncol 2019; 3: 4 [PMID: 30729176 DOI: 10.1038/s41698-018-0076-8
- Gottumukkala S, Tumati V, Hrycushko B, Folkert M. Endoluminal and Interstitial Brachytherapy 5 for the Treatment of Gastrointestinal Malignancies: a Systematic Review. Curr Oncol Rep 2017; 19: 2 [PMID: 28110462 DOI: 10.1007/s11912-017-0561-1]
- 6 Li JN, Li W, Cao LQ, Liu N, Zhang K. Efficacy of mesenchymal stem cells in the treatment of gastrointestinal malignancies. World J Gastrointest Oncol 2020; 12: 365-382 [PMID: 32368316 DOI: 10.4251/wjgo.v12.i4.365]
- 7 Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. CA Cancer J Clin 2019; 69: 7-34 [PMID: 30620402 DOI: 10.3322/caac.21551]
- Harmsen S, Rogalla S, Huang R, Spaliviero M, Neuschmelting V, Hayakawa Y, Lee Y, Tailor Y, Toledo-Crow R, Kang JW, Samii JM, Karabeber H, Davis RM, White JR, van de Rijn M, Gambhir SS, Contag CH, Wang TC, Kircher MF. Detection of Premalignant Gastrointestinal Lesions Using Surface-Enhanced Resonance Raman Scattering-Nanoparticle Endoscopy. ACS Nano 2019; 13: 1354-1364 [PMID: 30624916 DOI: 10.1021/acsnano.8b06808]
- 9 Baksh D, Song L, Tuan RS. Adult mesenchymal stem cells: characterization, differentiation, and application in cell and gene therapy. J Cell Mol Med 2004; 8: 301-316 [PMID: 15491506 DOI: 10.1111/j.1582-4934.2004.tb00320.x]
- 10 Kolf CM, Cho E, Tuan RS. Mesenchymal stromal cells. Biology of adult mesenchymal stem cells: regulation of niche, self-renewal and differentiation. Arthritis Res Ther 2007; 9: 204 [PMID: 17316462 DOI: 10.1186/ar2116]
- Ho IA, Toh HC, Ng WH, Teo YL, Guo CM, Hui KM, Lam PY. Human bone marrow-derived 11 mesenchymal stem cells suppress human glioma growth through inhibition of angiogenesis. Stem Cells 2013; 31: 146-155 [PMID: 23034897 DOI: 10.1002/stem.1247]
- Kidd S, Spaeth E, Dembinski JL, Dietrich M, Watson K, Klopp A, Battula VL, Weil M, Andreeff 12



M, Marini FC. Direct evidence of mesenchymal stem cell tropism for tumor and wounding microenvironments using in vivo bioluminescent imaging. Stem Cells 2009; 27: 2614-2623 [PMID: 19650040 DOI: 10.1002/stem.187]

- 13 Ji R, Zhang B, Zhang X, Xue J, Yuan X, Yan Y, Wang M, Zhu W, Qian H, Xu W. Exosomes derived from human mesenchymal stem cells confer drug resistance in gastric cancer. Cell Cycle 2015; 14: 2473-2483 [PMID: 26091251 DOI: 10.1080/15384101.2015.1005530]
- 14 Ridge SM, Sullivan FJ, Glynn SA. Mesenchymal stem cells: key players in cancer progression. Mol Cancer 2017; 16: 31 [PMID: 28148268 DOI: 10.1186/s12943-017-0597-8]
- Mognetti B, La Montagna G, Perrelli MG, Pagliaro P, Penna C. Bone marrow mesenchymal stem 15 cells increase motility of prostate cancer cells via production of stromal cell-derived factor-1a. J Cell Mol Med 2013; 17: 287-292 [PMID: 23301946 DOI: 10.1111/jcmm.12010]
- 16 Quante M, Tu SP, Tomita H, Gonda T, Wang SS, Takashi S, Baik GH, Shibata W, Diprete B, Betz KS, Friedman R, Varro A, Tycko B, Wang TC. Bone marrow-derived myofibroblasts contribute to the mesenchymal stem cell niche and promote tumor growth. Cancer Cell 2011; 19: 257-272 [PMID: 21316604 DOI: 10.1016/j.ccr.2011.01.020]
- Djouad F, Plence P, Bony C, Tropel P, Apparailly F, Sany J, Noël D, Jorgensen C. 17 Immunosuppressive effect of mesenchymal stem cells favors tumor growth in allogeneic animals. Blood 2003; 102: 3837-3844 [PMID: 12881305 DOI: 10.1182/blood-2003-04-1193]
- 18 Zhu Y, Sun Z, Han Q, Liao L, Wang J, Bian C, Li J, Yan X, Liu Y, Shao C, Zhao RC. Human mesenchymal stem cells inhibit cancer cell proliferation by secreting DKK-1. Leukemia 2009; 23: 925-933 [PMID: 19148141 DOI: 10.1038/leu.2008.384]
- Lu YR, Yuan Y, Wang XJ, Wei LL, Chen YN, Cong C, Li SF, Long D, Tan WD, Mao YQ, Zhang 19 J, Li YP, Cheng JQ. The growth inhibitory effect of mesenchymal stem cells on tumor cells in vitro and in vivo. Cancer Biol Ther 2008; 7: 245-251 [PMID: 18059192 DOI: 10.4161/cbt.7.2.5296]
- Keshtkar S, Azarpira N, Ghahremani MH. Mesenchymal stem cell-derived extracellular vesicles: 20 novel frontiers in regenerative medicine. Stem Cell Res Ther 2018; 9: 63 [PMID: 29523213 DOI: 10.1186/s13287-018-0791-7
- 21 Zhou J, Tan X, Tan Y, Li Q, Ma J, Wang G. Mesenchymal Stem Cell Derived Exosomes in Cancer Progression, Metastasis and Drug Delivery: A Comprehensive Review. J Cancer 2018; 9: 3129-3137 [PMID: 30210636 DOI: 10.7150/jca.25376]
- 22 Sun F, Wang JZ, Luo JJ, Wang YQ, Pan Q. Exosomes in the Oncobiology, Diagnosis, and Therapy of Hepatic Carcinoma: A New Player of an Old Game. Biomed Res Int 2018; 2018: 2747461 [PMID: 30148162 DOI: 10.1155/2018/2747461]
- Zhang X, Yang Y, Chen H, Tu H, Li J. Exosomes from Bone Marrow Microenvironment-Derived 23 Mesenchymal Stem Cells Affect CML Cells Growth and Promote Drug Resistance to Tyrosine Kinase Inhibitors. Stem Cells Int 2020; 2020: 8890201 [PMID: 33414831 DOI: 10.1155/2020/8890201
- 24 Lee JR, Park BW, Kim J, Choo YW, Kim HY, Yoon JK, Kim H, Hwang JW, Kang M, Kwon SP, Song SY, Ko IO, Park JA, Ban K, Hyeon T, Park HJ, Kim BS. Nanovesicles derived from iron oxide nanoparticles-incorporated mesenchymal stem cells for cardiac repair. Sci Adv 2020; 6: eaaz0952 [PMID: 32494669 DOI: 10.1126/sciadv.aaz0952]
- Zagrean AM, Hermann DM, Opris I, Zagrean L, Popa-Wagner A. Multicellular Crosstalk Between 25 Exosomes and the Neurovascular Unit After Cerebral Ischemia, Therapeutic Implications, Front Neurosci 2018; 12: 811 [PMID: 30459547 DOI: 10.3389/fnins.2018.00811]
- 26 Wang M, Xu X, Lei X, Tan J, Xie H. Mesenchymal stem cell-based therapy for burn wound healing. Burns Trauma 2021; 9: tkab002 [PMID: 34212055 DOI: 10.1093/burnst/tkab002]
- 27 Kim S, Kim TM. Generation of mesenchymal stem-like cells for producing extracellular vesicles. World J Stem Cells 2019; 11: 270-280 [PMID: 31171955 DOI: 10.4252/wjsc.v11.i5.270]
- Dalirfardouei R, Jamialahmadi K, Jafarian AH, Mahdipour E. Promising effects of exosomes 28 isolated from menstrual blood-derived mesenchymal stem cell on wound-healing process in diabetic mouse model. J Tissue Eng Regen Med 2019; 13: 555-568 [PMID: 30656863 DOI: 10.1002/term.2799
- 29 Stanko P, Altanerova U, Jakubechova J, Repiska V, Altaner C. Dental Mesenchymal Stem/Stromal Cells and Their Exosomes. Stem Cells Int 2018; 2018: 8973613 [PMID: 29760738 DOI: 10.1155/2018/8973613]
- 30 Fitzsimmons REB, Mazurek MS, Soos A, Simmons CA. Mesenchymal Stromal/Stem Cells in Regenerative Medicine and Tissue Engineering. Stem Cells Int 2018; 2018: 8031718 [PMID: 30210552 DOI: 10.1155/2018/8031718]
- Trounson A, McDonald C. Stem Cell Therapies in Clinical Trials: Progress and Challenges. Cell 31 Stem Cell 2015; 17: 11-22 [PMID: 26140604 DOI: 10.1016/j.stem.2015.06.007]
- 32 Fischer UM, Harting MT, Jimenez F, Monzon-Posadas WO, Xue H, Savitz SI, Laine GA, Cox CS Jr. Pulmonary passage is a major obstacle for intravenous stem cell delivery: the pulmonary firstpass effect. Stem Cells Dev 2009; 18: 683-692 [PMID: 19099374 DOI: 10.1089/scd.2008.0253]
- 33 Cai J, Wu J, Wang J, Li Y, Hu X, Luo S, Xiang D. Extracellular vesicles derived from different sources of mesenchymal stem cells: therapeutic effects and translational potential. Cell Biosci 2020; 10: 69 [PMID: 32483483 DOI: 10.1186/s13578-020-00427-x]
- 34 Huang Y, Liu K, Li Q, Yao Y, Wang Y. Exosomes Function in Tumor Immune Microenvironment. Adv Exp Med Biol 2018; 1056: 109-122 [PMID: 29754177 DOI: 10.1007/978-3-319-74470-4_7]
- 35 van Niel G, D'Angelo G, Raposo G. Shedding light on the cell biology of extracellular vesicles. Nat



Rev Mol Cell Biol 2018; 19: 213-228 [PMID: 29339798 DOI: 10.1038/nrm.2017.125]

- Phinney DG, Pittenger MF. Concise Review: MSC-Derived Exosomes for Cell-Free Therapy. Stem 36 Cells 2017; 35: 851-858 [PMID: 28294454 DOI: 10.1002/stem.2575]
- 37 Li X, Li C, Zhang L, Wu M, Cao K, Jiang F, Chen D, Li N, Li W. The significance of exosomes in the development and treatment of hepatocellular carcinoma. Mol Cancer 2020; 19: 1 [PMID: 31901224 DOI: 10.1186/s12943-019-1085-0]
- 38 Record M, Carayon K, Poirot M, Silvente-Poirot S. Exosomes as new vesicular lipid transporters involved in cell-cell communication and various pathophysiologies. Biochim Biophys Acta 2014; 1841: 108-120 [PMID: 24140720 DOI: 10.1016/j.bbalip.2013.10.004]
- Collino F, Deregibus MC, Bruno S, Sterpone L, Aghemo G, Viltono L, Tetta C, Camussi G. 39 Microvesicles derived from adult human bone marrow and tissue specific mesenchymal stem cells shuttle selected pattern of miRNAs. PLoS One 2010; 5: e11803 [PMID: 20668554 DOI: 10.1371/journal.pone.0011803]
- 40 Simons M, Raposo G. Exosomes--vesicular carriers for intercellular communication. Curr Opin Cell Biol 2009; 21: 575-581 [PMID: 19442504 DOI: 10.1016/j.ceb.2009.03.007]
- 41 Sharma A. Role of stem cell derived exosomes in tumor biology. Int J Cancer 2018; 142: 1086-1092 [PMID: 28983919 DOI: 10.1002/ijc.31089]
- 42 Brinton LT, Sloane HS, Kester M, Kelly KA. Formation and role of exosomes in cancer. Cell Mol Life Sci 2015; 72: 659-671 [PMID: 25336151 DOI: 10.1007/s00018-014-1764-3]
- 43 Yang Y, Bucan V, Baehre H, von der Ohe J, Otte A, Hass R. Acquisition of new tumor cell properties by MSC-derived exosomes. Int J Oncol 2015; 47: 244-252 [PMID: 25963929 DOI: 10.3892/ijo.2015.3001
- 44 Vallabhaneni KC, Penfornis P, Dhule S, Guillonneau F, Adams KV, Mo YY, Xu R, Liu Y, Watabe K, Vemuri MC, Pochampally R. Extracellular vesicles from bone marrow mesenchymal stem/stromal cells transport tumor regulatory microRNA, proteins, and metabolites. Oncotarget 2015; 6: 4953-4967 [PMID: 25669974 DOI: 10.18632/oncotarget.3211]
- 45 Lee JK, Park SR, Jung BK, Jeon YK, Lee YS, Kim MK, Kim YG, Jang JY, Kim CW. Exosomes derived from mesenchymal stem cells suppress angiogenesis by down-regulating VEGF expression in breast cancer cells. *PLoS One* 2013; 8: e84256 [PMID: 24391924 DOI: 10.1371/journal.pone.0084256
- 46 Luan X, Sansanaphongpricha K, Myers I, Chen H, Yuan H, Sun D. Engineering exosomes as refined biological nanoplatforms for drug delivery. Acta Pharmacol Sin 2017; 38: 754-763 [PMID: 28392567 DOI: 10.1038/aps.2017.12]
- 47 Pascucci L, Coccè V, Bonomi A, Ami D, Ceccarelli P, Ciusani E, Viganò L, Locatelli A, Sisto F, Doglia SM, Parati E, Bernardo ME, Muraca M, Alessandri G, Bondiolotti G, Pessina A, Paclitaxel is incorporated by mesenchymal stromal cells and released in exosomes that inhibit in vitro tumor growth: a new approach for drug delivery. J Control Release 2014; 192: 262-270 [PMID: 25084218 DOI: 10.1016/j.jconrel.2014.07.042]
- 48 Figueroa J, Phillips LM, Shahar T, Hossain A, Gumin J, Kim H, Bean AJ, Calin GA, Fueyo J, Walters ET, Kalluri R, Verhaak RG, Lang FF. Exosomes from Glioma-Associated Mesenchymal Stem Cells Increase the Tumorigenicity of Glioma Stem-like Cells via Transfer of miR-1587. Cancer Res 2017; 77: 5808-5819 [PMID: 28855213 DOI: 10.1158/0008-5472.CAN-16-2524]
- 49 Munoz JL, Bliss SA, Greco SJ, Ramkissoon SH, Ligon KL, Rameshwar P. Delivery of Functional Anti-miR-9 by Mesenchymal Stem Cell-derived Exosomes to Glioblastoma Multiforme Cells Conferred Chemosensitivity. Mol Ther Nucleic Acids 2013; 2: e126 [PMID: 24084846 DOI: 10.1038/mtna.2013.60
- 50 Taylor DD, Gercel-Taylor C. Exosomes/microvesicles: mediators of cancer-associated immunosuppressive microenvironments. Semin Immunopathol 2011; 33: 441-454 [PMID: 21688197 DOI: 10.1007/s00281-010-0234-8]
- Smyth EC, Nilsson M, Grabsch HI, van Grieken NC, Lordick F. Gastric cancer. Lancet 2020; 396: 51 635-648 [PMID: 32861308 DOI: 10.1016/S0140-6736(20)31288-5]
- Machlowska J, Baj J, Sitarz M, Maciejewski R, Sitarz R. Gastric Cancer: Epidemiology, Risk 52 Factors, Classification, Genomic Characteristics and Treatment Strategies. Int J Mol Sci 2020; 21 [PMID: 32512697 DOI: 10.3390/ijms21114012]
- Lee JH, Chang KK, Yoon C, Tang LH, Strong VE, Yoon SS. Lauren Histologic Type Is the Most 53 Important Factor Associated With Pattern of Recurrence Following Resection of Gastric Adenocarcinoma. Ann Surg 2018; 267: 105-113 [PMID: 27759618 DOI: 10.1097/SLA.000000000002040]
- 54 Guan J, Chen J. Mesenchymal stem cells in the tumor microenvironment. Biomed Rep 2013; 1: 517-521 [PMID: 24648978 DOI: 10.3892/br.2013.103]
- 55 Zhu W, Huang L, Li Y, Zhang X, Gu J, Yan Y, Xu X, Wang M, Qian H, Xu W. Exosomes derived from human bone marrow mesenchymal stem cells promote tumor growth in vivo. Cancer Lett 2012; 315: 28-37 [PMID: 22055459 DOI: 10.1016/j.canlet.2011.10.002]
- 56 Hood JL, Pan H, Lanza GM, Wickline SA; Consortium for Translational Research in Advanced Imaging and Nanomedicine (C-TRAIN). Paracrine induction of endothelium by tumor exosomes. Lab Invest 2009; 89: 1317-1328 [PMID: 19786948 DOI: 10.1038/labinvest.2009.94]
- 57 Yoshino Y, Aoyagi M, Tamaki M, Duan L, Morimoto T, Ohno K. Activation of p38 MAPK and/or JNK contributes to increased levels of VEGF secretion in human malignant glioma cells. Int J Oncol 2006; 29: 981-987 [PMID: 16964394 DOI: 10.3892/ijo.29.4.981]



- 58 Walczak C, Gaignier F, Gilet A, Zou F, Thornton SN, Ropars A. Aldosterone increases VEGF-A production in human neutrophils through PI3K, ERK1/2 and p38 pathways. Biochim Biophys Acta 2011; 1813: 2125-2132 [PMID: 21803079 DOI: 10.1016/j.bbamcr.2011.07.010]
- 59 Essafi-Benkhadir K, Pouysségur J, Pagès G. Implication of the ERK pathway on the posttranscriptional regulation of VEGF mRNA stability. Methods Mol Biol 2010; 661: 451-469 [PMID: 20812001 DOI: 10.1007/978-1-60761-795-2_28]
- 60 Zhu W, Huang L, Li Y, Qian H, Shan X, Yan Y, Mao F, Wu X, Xu WR. Mesenchymal stem cellsecreted soluble signaling molecules potentiate tumor growth. Cell Cycle 2011; 10: 3198-3207 [PMID: 21900753 DOI: 10.4161/cc.10.18.17638]
- Gu H, Ji R, Zhang X, Wang M, Zhu W, Qian H, Chen Y, Jiang P, Xu W. Exosomes derived from 61 human mesenchymal stem cells promote gastric cancer cell growth and migration via the activation of the Akt pathway. Mol Med Rep 2016; 14: 3452-3458 [PMID: 27513187 DOI: 10.3892/mmr.2016.5625
- Tsai JH, Yang J. Epithelial-mesenchymal plasticity in carcinoma metastasis. Genes Dev 2013; 27: 62 2192-2206 [PMID: 24142872 DOI: 10.1101/gad.225334.113]
- Savagner P. The epithelial-mesenchymal transition (EMT) phenomenon. Ann Oncol 2010; 21 Suppl 63 7: vii89-vii92 [PMID: 20943648 DOI: 10.1093/annonc/mdq292]
- 64 Qi J, Zhou Y, Jiao Z, Wang X, Zhao Y, Li Y, Chen H, Yang L, Zhu H. Exosomes Derived from Human Bone Marrow Mesenchymal Stem Cells Promote Tumor Growth Through Hedgehog Signaling Pathway. Cell Physiol Biochem 2017; 42: 2242-2254 [PMID: 28817816 DOI: 10.1159/000479998]
- 65 Ma M, Chen S, Liu Z, Xie H, Deng H, Shang S, Wang X, Xia M, Zuo C. miRNA-221 of exosomes originating from bone marrow mesenchymal stem cells promotes oncogenic activity in gastric cancer. Onco Targets Ther 2017; 10: 4161-4171 [PMID: 28860826 DOI: 10.2147/OTT.S143315]
- 66 Wang M, Zhao C, Shi H, Zhang B, Zhang L, Zhang X, Wang S, Wu X, Yang T, Huang F, Cai J, Zhu Q, Zhu W, Qian H, Xu W. Deregulated microRNAs in gastric cancer tissue-derived mesenchymal stem cells: novel biomarkers and a mechanism for gastric cancer. Br J Cancer 2014; 110: 1199-1210 [PMID: 24473397 DOI: 10.1038/bjc.2014.14]
- Ueda T, Volinia S, Okumura H, Shimizu M, Taccioli C, Rossi S, Alder H, Liu CG, Oue N, Yasui 67 W, Yoshida K, Sasaki H, Nomura S, Seto Y, Kaminishi M, Calin GA, Croce CM. Relation between microRNA expression and progression and prognosis of gastric cancer: a microRNA expression analysis. Lancet Oncol 2010; 11: 136-146 [PMID: 20022810 DOI: 10.1016/S1470-2045(09)70343-21
- 68 Liu K, Li G, Fan C, Diao Y, Wu B, Li J. Increased Expression of MicroRNA-221 in gastric cancer and its clinical significance. J Int Med Res 2012; 40: 467-474 [PMID: 22613407 DOI: 10.1177/147323001204000208
- Mao J, Liang Z, Zhang B, Yang H, Li X, Fu H, Zhang X, Yan Y, Xu W, Qian H. UBR2 Enriched in 69 p53 Deficient Mouse Bone Marrow Mesenchymal Stem Cell-Exosome Promoted Gastric Cancer Progression via Wnt/β-Catenin Pathway. Stem Cells 2017; 35: 2267-2279 [PMID: 28895255 DOI: 10.1002/stem.2702]
- Mao J, Fan S, Ma W, Fan P, Wang B, Zhang J, Wang H, Tang B, Zhang Q, Yu X, Wang L, Song B, 70 Li L. Roles of Wnt/β-catenin signaling in the gastric cancer stem cells proliferation and salinomycin treatment. Cell Death Dis 2014; 5: e1039 [PMID: 24481453 DOI: 10.1038/cddis.2013.515]
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer 71 Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin 2021; 71: 209-249 [PMID: 33538338 DOI: 10.3322/caac.21660]
- Llovet JM, Kelley RK, Villanueva A, Singal AG, Pikarsky E, Roayaie S, Lencioni R, Koike K, 72 Zucman-Rossi J, Finn RS. Hepatocellular carcinoma. Nat Rev Dis Primers 2021; 7: 6 [PMID: 33479224 DOI: 10.1038/s41572-020-00240-3]
- Jemal A, Ward EM, Johnson CJ, Cronin KA, Ma J, Ryerson B, Mariotto A, Lake AJ, Wilson R, 73 Sherman RL, Anderson RN, Henley SJ, Kohler BA, Penberthy L, Feuer EJ, Weir HK. Annual Report to the Nation on the Status of Cancer, 1975-2014, Featuring Survival. J Natl Cancer Inst 2017; 109 [PMID: 28376154 DOI: 10.1093/jnci/djx030]
- Villanueva A. Hepatocellular Carcinoma. N Engl J Med 2019; 380: 1450-1462 [PMID: 30970190 74 DOI: 10.1056/NEJMra1713263]
- Kim SY, An J, Lim YS, Han S, Lee JY, Byun JH, Won HJ, Lee SJ, Lee HC, Lee YS. MRI With 75 Liver-Specific Contrast for Surveillance of Patients With Cirrhosis at High Risk of Hepatocellular Carcinoma. JAMA Oncol 2017; 3: 456-463 [PMID: 27657493 DOI: 10.1001/jamaoncol.2016.3147]
- 76 Katona BW, Weiss JM. Chemoprevention of Colorectal Cancer. Gastroenterology 2020; 158: 368-388 [PMID: 31563626 DOI: 10.1053/j.gastro.2019.06.047]
- 77 Lohitesh K, Chowdhury R, Mukherjee S. Resistance a major hindrance to chemotherapy in hepatocellular carcinoma: an insight. Cancer Cell Int 2018; 18: 44 [PMID: 29568237 DOI: 10.1186/s12935-018-0538-7]
- 78 Coulouarn C, Factor VM, Andersen JB, Durkin ME, Thorgeirsson SS. Loss of miR-122 expression in liver cancer correlates with suppression of the hepatic phenotype and gain of metastatic properties. Oncogene 2009; 28: 3526-3536 [PMID: 19617899 DOI: 10.1038/onc.2009.211]
- 79 Tsai WC, Hsu SD, Hsu CS, Lai TC, Chen SJ, Shen R, Huang Y, Chen HC, Lee CH, Tsai TF, Hsu MT, Wu JC, Huang HD, Shiao MS, Hsiao M, Tsou AP. MicroRNA-122 plays a critical role in liver homeostasis and hepatocarcinogenesis. J Clin Invest 2012; 122: 2884-2897 [PMID: 22820290 DOI:



10.1172/JCI63455]

- 80 Zhang H, Chen Z, Wang X, Huang Z, He Z, Chen Y. Long non-coding RNA: a new player in cancer. J Hematol Oncol 2013; 6: 37 [PMID: 23725405 DOI: 10.1186/1756-8722-6-37]
- 81 Fornari F, Gramantieri L, Giovannini C, Veronese A, Ferracin M, Sabbioni S, Calin GA, Grazi GL, Croce CM, Tavolari S, Chieco P, Negrini M, Bolondi L. MiR-122/cyclin G1 interaction modulates p53 activity and affects doxorubicin sensitivity of human hepatocarcinoma cells. Cancer Res 2009; 69: 5761-5767 [PMID: 19584283 DOI: 10.1158/0008-5472.CAN-08-4797]
- 82 Bai S, Nasser MW, Wang B, Hsu SH, Datta J, Kutay H, Yadav A, Nuovo G, Kumar P, Ghoshal K. MicroRNA-122 inhibits tumorigenic properties of hepatocellular carcinoma cells and sensitizes these cells to sorafenib. J Biol Chem 2009; 284: 32015-32027 [PMID: 19726678 DOI: 10.1074/jbc.M109.016774]
- Lou G, Song X, Yang F, Wu S, Wang J, Chen Z, Liu Y. Exosomes derived from miR-122-modified 83 adipose tissue-derived MSCs increase chemosensitivity of hepatocellular carcinoma. J Hematol Oncol 2015; 8: 122 [PMID: 26514126 DOI: 10.1186/s13045-015-0220-7]
- Ren K, Li T, Zhang W, Ren J, Li Z, Wu G. miR-199a-3p inhibits cell proliferation and induces 84 apoptosis by targeting YAP1, suppressing Jagged1-Notch signaling in human hepatocellular carcinoma. J Biomed Sci 2016; 23: 79 [PMID: 27832779 DOI: 10.1186/s12929-016-0295-7]
- 85 Kim JH, Badawi M, Park JK, Jiang J, Mo X, Roberts LR, Schmittgen TD. Anti-invasion and antimigration effects of miR-199a-3p in hepatocellular carcinoma are due in part to targeting CD151. Int J Oncol 2016; 49: 2037-2045 [PMID: 27599545 DOI: 10.3892/ijo.2016.3677]
- Fornari F, Milazzo M, Chieco P, Negrini M, Calin GA, Grazi GL, Pollutri D, Croce CM, Bolondi L, Gramantieri L. MiR-199a-3p regulates mTOR and c-Met to influence the doxorubicin sensitivity of human hepatocarcinoma cells. Cancer Res 2010; 70: 5184-5193 [PMID: 20501828 DOI: 10.1158/0008-5472.CAN-10-0145
- 87 Lou G, Chen L, Xia C, Wang W, Qi J, Li A, Zhao L, Chen Z, Zheng M, Liu Y. MiR-199a-modified exosomes from adipose tissue-derived mesenchymal stem cells improve hepatocellular carcinoma chemosensitivity through mTOR pathway. J Exp Clin Cancer Res 2020; 39: 4 [PMID: 31898515 DOI: 10.1186/s13046-019-1512-5]
- 88 Hu G, Drescher KM, Chen XM. Exosomal miRNAs: Biological Properties and Therapeutic Potential. Front Genet 2012; 3: 56 [PMID: 22529849 DOI: 10.3389/fgene.2012.00056]
- 89 Ha D, Yang N, Nadithe V. Exosomes as therapeutic drug carriers and delivery vehicles across biological membranes: current perspectives and future challenges. Acta Pharm Sin B 2016; 6: 287-296 [PMID: 27471669 DOI: 10.1016/j.apsb.2016.02.001]
- 90 Yeo RW, Lai RC, Zhang B, Tan SS, Yin Y, Teh BJ, Lim SK. Mesenchymal stem cell: an efficient mass producer of exosomes for drug delivery. Adv Drug Deliv Rev 2013; 65: 336-341 [PMID: 22780955 DOI: 10.1016/j.addr.2012.07.001]
- Akyurekli C, Le Y, Richardson RB, Fergusson D, Tay J, Allan DS. A systematic review of 91 preclinical studies on the therapeutic potential of mesenchymal stromal cell-derived microvesicles. Stem Cell Rev Rep 2015; 11: 150-160 [PMID: 25091427 DOI: 10.1007/s12015-014-9545-9]
- 92 Roccaro AM, Sacco A, Maiso P, Azab AK, Tai YT, Reagan M, Azab F, Flores LM, Campigotto F, Weller E, Anderson KC, Scadden DT, Ghobrial IM. BM mesenchymal stromal cell-derived exosomes facilitate multiple myeloma progression. J Clin Invest 2013; 123: 1542-1555 [PMID: 23454749 DOI: 10.1172/JCI66517]
- 93 Bruno S, Collino F, Deregibus MC, Grange C, Tetta C, Camussi G. Microvesicles derived from human bone marrow mesenchymal stem cells inhibit tumor growth. Stem Cells Dev 2013; 22: 758-771 [PMID: 23034046 DOI: 10.1089/scd.2012.0304]
- 94 Wu S, Ju GQ, Du T, Zhu YJ, Liu GH. Microvesicles derived from human umbilical cord Wharton's jelly mesenchymal stem cells attenuate bladder tumor cell growth in vitro and in vivo. PLoS One 2013; 8: e61366 [PMID: 23593475 DOI: 10.1371/journal.pone.0061366]
- 95 Ko SF, Yip HK, Zhen YY, Lee CC, Huang CC, Ng SH, Lin JW. Adipose-Derived Mesenchymal Stem Cell Exosomes Suppress Hepatocellular Carcinoma Growth in a Rat Model: Apparent Diffusion Coefficient, Natural Killer T-Cell Responses, and Histopathological Features. Stem Cells Int 2015; 2015: 853506 [PMID: 26345219 DOI: 10.1155/2015/853506]
- 96 Alzahrani FA, El-Magd MA, Abdelfattah-Hassan A, Saleh AA, Saadeldin IM, El-Shetry ES, Badawy AA, Alkarim S. Potential Effect of Exosomes Derived from Cancer Stem Cells and MSCs on Progression of DEN-Induced HCC in Rats. Stem Cells Int 2018; 2018: 8058979 [PMID: 30224923 DOI: 10.1155/2018/8058979]
- 97 Tan CY, Lai RC, Wong W, Dan YY, Lim SK, Ho HK. Mesenchymal stem cell-derived exosomes promote hepatic regeneration in drug-induced liver injury models. Stem Cell Res Ther 2014; 5: 76 [PMID: 24915963 DOI: 10.1186/scrt465]
- 98 Li T, Yan Y, Wang B, Qian H, Zhang X, Shen L, Wang M, Zhou Y, Zhu W, Li W, Xu W. Exosomes derived from human umbilical cord mesenchymal stem cells alleviate liver fibrosis. Stem Cells Dev 2013; 22: 845-854 [PMID: 23002959 DOI: 10.1089/scd.2012.0395]
- 99 Korashy HM, El Gendy MA, Alhaider AA, El-Kadi AO. Camel milk modulates the expression of aryl hydrocarbon receptor-regulated genes, Cyp1a1, Nqo1, and Gsta1, in murine hepatoma Hepa 1c1c7 cells. J Biomed Biotechnol 2012; 2012: 782642 [PMID: 22570534 DOI: 10.1155/2012/782642]
- 100 Zhou Y, Zhou W, Chen X, Wang Q, Li C, Chen Q, Zhang Y, Lu Y, Ding X, Jiang C. Bone marrow mesenchymal stem cells-derived exosomes for penetrating and targeted chemotherapy of pancreatic



cancer. Acta Pharm Sin B 2020; 10: 1563-1575 [PMID: 32963950 DOI: 10.1016/j.apsb.2019.11.013]

- Guler GD, Ning Y, Ku CJ, Phillips T, McCarthy E, Ellison CK, Bergamaschi A, Collin F, Lloyd P, 101 Scott A, Antoine M, Wang W, Chau K, Ashworth A, Quake SR, Levy S. Detection of early stage pancreatic cancer using 5-hydroxymethylcytosine signatures in circulating cell free DNA. Nat Commun 2020; 11: 5270 [PMID: 33077732 DOI: 10.1038/s41467-020-18965-w]
- Lunardi S, Muschel RJ, Brunner TB. The stromal compartments in pancreatic cancer: are there any 102 therapeutic targets? Cancer Lett 2014; 343: 147-155 [PMID: 24141189 DOI: 10.1016/j.canlet.2013.09.039]
- 103 Samulitis BK, Pond KW, Pond E, Cress AE, Patel H, Wisner L, Patel C, Dorr RT, Landowski TH. Gemcitabine resistant pancreatic cancer cell lines acquire an invasive phenotype with collateral hypersensitivity to histone deacetylase inhibitors. Cancer Biol Ther 2015; 16: 43-51 [PMID: 25485960 DOI: 10.4161/15384047.2014.986967]
- 104 Kabashima-Niibe A, Higuchi H, Takaishi H, Masugi Y, Matsuzaki Y, Mabuchi Y, Funakoshi S, Adachi M, Hamamoto Y, Kawachi S, Aiura K, Kitagawa Y, Sakamoto M, Hibi T. Mesenchymal stem cells regulate epithelial-mesenchymal transition and tumor progression of pancreatic cancer cells. Cancer Sci 2013; 104: 157-164 [PMID: 23121112 DOI: 10.1111/cas.12059]
- 105 Kamerkar S, LeBleu VS, Sugimoto H, Yang S, Ruivo CF, Melo SA, Lee JJ, Kalluri R. Exosomes facilitate therapeutic targeting of oncogenic KRAS in pancreatic cancer. Nature 2017; 546: 498-503 [PMID: 28607485 DOI: 10.1038/nature22341]
- 106 Zhou W, Zhou Y, Chen X, Ning T, Chen H, Guo Q, Zhang Y, Liu P, Li C, Chu Y, Sun T, Jiang C. Pancreatic cancer-targeting exosomes for enhancing immunotherapy and reprogramming tumor microenvironment. Biomaterials 2021; 268: 120546 [PMID: 33253966 DOI: 10.1016/j.biomaterials.2020.120546]
- 107 Shang S, Wang J, Chen S, Tian R, Zeng H, Wang L, Xia M, Zhu H, Zuo C. Exosomal miRNA-1231 derived from bone marrow mesenchymal stem cells inhibits the activity of pancreatic cancer. Cancer Med 2019; 8: 7728-7740 [PMID: 31642612 DOI: 10.1002/cam4.2633]
- 108 Wu DM, Wen X, Han XR, Wang S, Wang YJ, Shen M, Fan SH, Zhang ZF, Shan Q, Li MQ, Hu B, Lu J, Chen GQ, Zheng YL. Bone Marrow Mesenchymal Stem Cell-Derived Exosomal MicroRNA-126-3p Inhibits Pancreatic Cancer Development by Targeting ADAM9. Mol Ther Nucleic Acids 2019; 16: 229-245 [PMID: 30925451 DOI: 10.1016/j.omtn.2019.02.022]
- 109 Xu Y, Liu N, Wei Y, Zhou D, Lin R, Wang X, Shi B. Anticancer effects of miR-124 delivered by BM-MSC derived exosomes on cell proliferation, epithelial mesenchymal transition, and chemotherapy sensitivity of pancreatic cancer cells. Aging (Albany NY) 2020; 12: 19660-19676 [PMID: 33040049 DOI: 10.18632/aging.103997]
- 110 Neo WH, Yap K, Lee SH, Looi LS, Khandelia P, Neo SX, Makeyev EV, Su IH. MicroRNA miR-124 controls the choice between neuronal and astrocyte differentiation by fine-tuning Ezh2 expression. J Biol Chem 2014; 289: 20788-20801 [PMID: 24878960 DOI: 10.1074/jbc.M113.525493
- 111 Liu S, Song L, Zeng S, Zhang L. MALAT1-miR-124-RBG2 axis is involved in growth and invasion of HR-HPV-positive cervical cancer cells. Tumour Biol 2016; 37: 633-640 [PMID: 26242259 DOI: 10.1007/s13277-015-3732-4]
- Wang Y, Chen L, Wu Z, Wang M, Jin F, Wang N, Hu X, Liu Z, Zhang CY, Zen K, Chen J, Liang 112 H, Zhang Y, Chen X. miR-124-3p functions as a tumor suppressor in breast cancer by targeting CBL. BMC Cancer 2016; 16: 826 [PMID: 27842510 DOI: 10.1186/s12885-016-2862-4]
- Xiong Y, Wang L, Li Y, Chen M, He W, Qi L. The Long Non-Coding RNA XIST Interacted with 113 MiR-124 to Modulate Bladder Cancer Growth, Invasion and Migration by Targeting Androgen Receptor (AR). Cell Physiol Biochem 2017; 43: 405-418 [PMID: 28869948 DOI: 10.1159/000480419]
- 114 Wang F, Lau JKC, Yu J. The role of natural killer cell in gastrointestinal cancer: killer or helper. Oncogene 2021; 40: 717-730 [PMID: 33262461 DOI: 10.1038/s41388-020-01561-z]
- Edwards BK, Ward E, Kohler BA, Eheman C, Zauber AG, Anderson RN, Jemal A, Schymura MJ, 115 Lansdorp-Vogelaar I, Seeff LC, van Ballegooijen M, Goede SL, Ries LA. Annual report to the nation on the status of cancer, 1975-2006, featuring colorectal cancer trends and impact of interventions (risk factors, screening, and treatment) to reduce future rates. Cancer 2010; 116: 544-573 [PMID: 19998273 DOI: 10.1002/cncr.24760]
- Tsoi KKF, Hirai HW, Chan FCH, Griffiths S, Sung JJY. Predicted Increases in Incidence of 116 Colorectal Cancer in Developed and Developing Regions, in Association With Ageing Populations. Clin Gastroenterol Hepatol 2017; 15: 892-900.e4 [PMID: 27720911 DOI: 10.1016/j.cgh.2016.09.155]
- 117 Fu X, Xie F, Gong F, Yang Z, Lv X, Li X, Jiao H, Wang Q, Liu X, Yan L, Xiao R. Suppression of PTBP1 signaling is responsible for mesenchymal stem cell induced invasion of low malignancy cancer cells. Biochim Biophys Acta Mol Cell Res 2018; 1865: 1552-1565 [PMID: 30327198 DOI: 10.1016/j.bbamcr.2018.08.002
- 118 Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. CA Cancer J Clin 2015; 65: 87-108 [PMID: 25651787 DOI: 10.3322/caac.21262]
- 119 Kang J, Zhang L, Luo X, Ma X, Wang G, Yang Y, Yan Y, Qian H, Zhang X, Xu W, Mao F. Systematic Exposition of Mesenchymal Stem Cell for Inflammatory Bowel Disease and Its Associated Colorectal Cancer. Biomed Res Int 2018; 2018: 9652817 [PMID: 30687760 DOI:



10.1155/2018/9652817]

- 120 Yang L, Zhang Y, Cheng L, Yue D, Ma J, Zhao D, Hou X, Xiang R, Cheng P. Mesenchymal Stem Cells Engineered to Secrete Pigment Epithelium-Derived Factor Inhibit Tumor Metastasis and the Formation of Malignant Ascites in a Murine Colorectal Peritoneal Carcinomatosis Model. *Hum Gene Ther* 2016; 27: 267-277 [PMID: 26756933 DOI: 10.1089/hum.2015.135]
- 121 Tang RJ, Shen SN, Zhao XY, Nie YZ, Xu YJ, Ren J, Lv MM, Hou YY, Wang TT. Mesenchymal stem cells-regulated Treg cells suppress colitis-associated colorectal cancer. *Stem Cell Res Ther* 2015; 6: 71 [PMID: 25889203 DOI: 10.1186/s13287-015-0055-8]
- 122 Chen Z, He X, Chen X, Lin X, Zou Y, Wu X, Lan P. Bone marrow mesenchymal stem cells ameliorate colitis-associated tumorigenesis in mice. *Biochem Biophys Res Commun* 2014; 450: 1402-1408 [PMID: 25010644 DOI: 10.1016/j.bbrc.2014.07.002]
- 123 Feng H, Zhao JK, Schiergens TS, Wang PX, Ou BC, Al-Sayegh R, Li ML, Lu AG, Yin S, Thasler WE. Bone marrow-derived mesenchymal stromal cells promote colorectal cancer cell death under low-dose irradiation. *Br J Cancer* 2018; 118: 353-365 [PMID: 29384527 DOI: 10.1038/bjc.2017.415]
- 124 Xu Y, Shen L, Li F, Yang J, Wan X, Ouyang M. microRNA-16-5p-containing exosomes derived from bone marrow-derived mesenchymal stem cells inhibit proliferation, migration, and invasion, while promoting apoptosis of colorectal cancer cells by downregulating ITGA2. *J Cell Physiol* 2019; 234: 21380-21394 [PMID: 31102273 DOI: 10.1002/jcp.28747]
- 125 Chen HL, Li JJ, Jiang F, Shi WJ, Chang GY. MicroRNA-4461 derived from bone marrow mesenchymal stem cell exosomes inhibits tumorigenesis by downregulating COPB2 expression in colorectal cancer. *Biosci Biotechnol Biochem* 2020; 84: 338-346 [PMID: 31631786 DOI: 10.1080/09168451.2019.1677452]
- 126 Li H, Li F. Exosomes from BM-MSCs increase the population of CSCs *via* transfer of miR-142-3p. *Br J Cancer* 2018; 119: 744-755 [PMID: 30220706 DOI: 10.1038/s41416-018-0254-z]
- 127 Shen WW, Zeng Z, Zhu WX, Fu GH. MiR-142-3p functions as a tumor suppressor by targeting CD133, ABCG2, and Lgr5 in colon cancer cells. *J Mol Med (Berl)* 2013; **91**: 989-1000 [PMID: 23619912 DOI: 10.1007/s00109-013-1037-x]
- 128 Li T, Wan Y, Su Z, Li J, Han M, Zhou C. Mesenchymal Stem Cell-Derived Exosomal microRNA-3940-5p Inhibits Colorectal Cancer Metastasis by Targeting Integrin α6. *Dig Dis Sci* 2021; 66: 1916-1927 [PMID: 32671583 DOI: 10.1007/s10620-020-06458-1]
- 129 Guo L, Fu J, Sun S, Zhu M, Zhang L, Niu H, Chen Z, Zhang Y, Guo L, Wang S. MicroRNA-143-3p inhibits colorectal cancer metastases by targeting ITGA6 and ASAP3. *Cancer Sci* 2019; 110: 805-816 [PMID: 30536996 DOI: 10.1111/cas.13910]
- 130 Cui M, Chang Y, Du W, Liu S, Qi J, Luo R, Luo S. Upregulation of IncRNA-ATB by Transforming Growth Factor β1 (TGF-β1) Promotes Migration and Invasion of Papillary Thyroid Carcinoma Cells. *Med Sci Monit* 2018; 24: 5152-5158 [PMID: 30042377 DOI: 10.12659/MSM.909420]
- 131 Zhang Y, Chen Y, Lo C, Zhuang J, Angsantikul P, Zhang Q, Wei X, Zhou Z, Obonyo M, Fang RH, Gao W, Zhang L. Inhibition of Pathogen Adhesion by Bacterial Outer Membrane-Coated Nanoparticles. *Angew Chem Int Ed Engl* 2019; 58: 11404-11408 [PMID: 31206942 DOI: 10.1002/anie.201906280]
- 132 Bagheri E, Abnous K, Farzad SA, Taghdisi SM, Ramezani M, Alibolandi M. Targeted doxorubicinloaded mesenchymal stem cells-derived exosomes as a versatile platform for fighting against colorectal cancer. *Life Sci* 2020; 261: 118369 [PMID: 32882265 DOI: 10.1016/j.lfs.2020.118369]
- 133 Liu H, Liang Z, Wang F, Zhou C, Zheng X, Hu T, He X, Wu X, Lan P. Exosomes from mesenchymal stromal cells reduce murine colonic inflammation via a macrophage-dependent mechanism. JCI Insight 2019; 4 [PMID: 31689240 DOI: 10.1172/jci.insight.131273]
- 134 Penfornis P, Vallabhaneni KC, Whitt J, Pochampally R. Extracellular vesicles as carriers of microRNA, proteins and lipids in tumor microenvironment. *Int J Cancer* 2016; 138: 14-21 [PMID: 25559768 DOI: 10.1002/ijc.29417]
- 135 Reiter JG, Baretti M, Gerold JM, Makohon-Moore AP, Daud A, Iacobuzio-Donahue CA, Azad NS, Kinzler KW, Nowak MA, Vogelstein B. An analysis of genetic heterogeneity in untreated cancers. *Nat Rev Cancer* 2019; 19: 639-650 [PMID: 31455892 DOI: 10.1038/s41568-019-0185-x]
- 136 Wang L, Zhang F, Peng W, Zhang J, Dong W, Yuan D, Wang Z, Zheng Y. Preincubation with a low-dose hydrogen peroxide enhances anti-oxidative stress ability of BMSCs. *J Orthop Surg Res* 2020; 15: 392 [PMID: 32907609 DOI: 10.1186/s13018-020-01916-y]
- 137 Kourembanas S. Exosomes: vehicles of intercellular signaling, biomarkers, and vectors of cell therapy. Annu Rev Physiol 2015; 77: 13-27 [PMID: 25293529 DOI: 10.1146/annurev-physiol-021014-071641]
- 138 Kalimuthu S, Gangadaran P, Rajendran RL, Zhu L, Oh JM, Lee HW, Gopal A, Back SH, Jeong SY, Lee SW, Lee J, Ahn BC. A New Approach for Loading Anticancer Drugs Into Mesenchymal Stem Cell-Derived Exosome Mimetics for Cancer Therapy. *Front Pharmacol* 2018; 9: 1116 [PMID: 30319428 DOI: 10.3389/fphar.2018.01116]

Zaishidena® WJGO | https://www.wjgnet.com

0 WJ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 1997-2012

DOI: 10.4251/wjgo.v13.i12.1997

ISSN 1948-5204 (online)

REVIEW

Diabetes mellitus contribution to the remodeling of the tumor microenvironment in gastric cancer

Armando Rojas, Cristian Lindner, Iván Schneider, Ileana Gonzàlez, Hernan Araya, Erik Morales, Milibeth Gómez, Nelson Urdaneta, Paulina Araya, Miguel Angel Morales

ORCID number: Armando Rojas 0000-0001-9911-7142; Cristian Lindner 0000-0002-2642-4288: Iván Schneider 0000-0001-5294-5995; Ileana Gonzàlez 0000-0002-2488-9380; Hernan Araya 0000-0002-0758-4040; Erik Morales 0000-0002-0271-2787; Milibeth Gómez 0000-0002-7548-2337; Nelson Urdaneta 0000-0003-2626-6010; Paulina Araya 0000-0002-6797-3056; Miguel Angel Morales 0000-0001-7698-9669.

Author contributions: All authors contributed to the original ideas and writing of this paper; Rojas A designed the report and wrote the paper; Lindner C contributed artwork and data acquisition, drafting and revising the manuscript; Schneider I, Gonzàlez I, Araya H, Morales E, Gómez M, Urdaneta N, Araya P and Morales MA contributed data acquisition, drafting and revising the manuscript.

Conflict-of-interest statement: The authors declare that there is no conflict of interest.

Country/Territory of origin: Chile

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review report's scientific

Armando Rojas, Cristian Lindner, Iván Schneider, Ileana Gonzàlez, Erik Morales, Paulina Araya, Biomedical Research Lab., Medicine Faculty, Catholic University of Maule, Talca 34600000, Chile

Hernan Araya, Milibeth Gómez, Nelson Urdaneta, Department of Clinical Sciences, Medicine Faculty, Catholic University of Maule, Talca 34600000, Chile

Hernan Araya, Milibeth Gómez, Nelson Urdaneta, Servicio de Oncología, Hospital Regional de Talca, Talca 34600000, Chile

Erik Morales, Servicio de Anatomía Patologica, Hospital Regional de Talca, Talca 34600000, Chile

Miguel Angel Morales, Department of Molecular and Clinical Pharmacology Program, Institute of Biomedical Sciences, University of Chile, Santiago 8320000, Chile

Corresponding author: Armando Rojas, PhD, Full Professor, Biomedical Research Lab., Medicine Faculty, Catholic University of Maule, 3605 San Miguel Ave., Talca 34600000, Chile. arojasr@ucm.cl

Abstract

Compelling pieces of evidence derived from both clinical and experimental research has demonstrated the crucial contribution of diabetes mellitus (DM) as a risk factor associated with increased cancer incidence and mortality in many human neoplasms, including gastric cancer (GC). DM is considered a systemic inflammatory disease and therefore, this inflammatory status may have profound effects on the tumor microenvironment (TME), particularly by driving many molecular mechanisms to generate a more aggressive TME. DM is an active driver in the modification of the behavior of many cell components of the TME as well as altering the mechanical properties of the extracellular matrix (ECM), leading to an increased ECM stiffening. Additionally, DM can alter many cellular signaling mechanisms and thus favoring tumor growth, invasion, and metastatic potential, as well as key elements in regulating cellular functions and cross-talks, such as the microRNAs network, the production, and cargo of exosomes, the metabolism of cell stroma and resistance to hypoxia. In the present review, we intend to highlight the mechanistic contributions of DM to the remodeling of TME in GC.



quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: May 13, 2021 Peer-review started: May 13, 2021 First decision: June 5, 2021 Revised: June 10, 2021 Accepted: October 27, 2021 Article in press: October 27, 2021 Published online: December 15, 2021

P-Reviewer: Ugo O S-Editor: Gao CC L-Editor: A P-Editor: Gao CC



Key Words: Diabetes mellitus; Gastric cancer; Tumor microenvironment; Hyperglycemia; Chronic inflammation

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Compelling shreds of evidence support that diabetes mellitus (DM) is a crucial risk factor in human cancers. Due to its contribution to systemic inflammation, DM can sculpture the gastric tumor microenvironment through different mechanisms, which in turn, may generate highly malignant phenotypes in gastric cancer (GC). We herein discuss the contribution of DM in the remodeling tumor microenvironment in GC, which may then leads to more aggressive tumor phenotypes.

Citation: Rojas A, Lindner C, Schneider I, Gonzàlez I, Araya H, Morales E, Gómez M, Urdaneta N, Araya P, Morales MA. Diabetes mellitus contribution to the remodeling of the tumor microenvironment in gastric cancer. World J Gastrointest Oncol 2021; 13(12): 1997-2012

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/1997.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.1997

INTRODUCTION

At present, a compelling body of evidence suggests that diabetes mellitus (DM) patients have not only increased incidence but also worse outcomes when they develop malignant neoplasm, especially those originating from gastrointestinal (GI) organs, such as the pancreas, colon, liver, and stomach[1].

Gastric cancer (GC) is the fifth most common cancer and the fourth most common cause of cancer death globally, with a poor 5-year survival < 20% for advanced stages [2].

Strikingly, data from different epidemiological data suggest that DM and chronic hyperglycemia may even increase the risk and mortality of GC patients[3,4]. Although there is compelling clinical data supporting this association, the molecular mechanisms underlying this association are not fully understood.

DM is considered a systemic inflammatory disorder^[5], which triggers a dysregulated metabolism, and is characterized by sustained hyperglycemia[6]. However, the systemic pro-inflammatory effects induced by DM are not only mediated by chronic hyperglycemia but also enhanced by insulin resistance[7]. All these features drive critical modifications in extracellular elements such as ECM and favor the dysregulation of many intracellular signaling pathways[7-9].

Furthermore, DM not only interferes in intercellular communication increasing the biogenesis of exosomes but also altering the delivery of biomolecules to recipient cells and favors a pro-angiogenic and proliferative cross-talk between stromal cells, which could act as a key element in defining the fate of tumor development in these patients [<mark>9</mark>].

A crucial role in the tumor biology of gastric carcinoma plays the complex network established between cellular and non-cellular elements that composed the tumor microenvironment (TME), which drives the cancer cell fate and plays a critical role in the initiation, progression, and immune evasion[10].

A growing body of evidence suggests that these TME remodeling mechanisms can be crucial in favoring the development of highly malignant phenotypes in DM patients who develop GC[5,11].

In the present review, we intend to highlight the different mechanisms of the contribution of DM to the remodeling of TME in GC, which leads to more aggressive tumors.

EPIDEMIOLOGICAL ASSOCIATION

DM is considered an established risk factor for either higher incidence and increased long-term all-cause mortality rates in many cancer types[12,13], especially for those



originating in the digestive tract[14,15].

Although several observational studies have demonstrated a controversial association between DM and CG[4,16] or restricted only to gender differences[17] a growing body of evidence including both meta-analyses of wide-population cohorts and case-control studies demonstrate an increased risk and mortality of GC in DM patients[3,18-21].

Furthermore, a recent study with a prospective endoscopic follow-up shows that DM is an independent risk factor for GC[3]. Noteworthy, this positive association remains significant even in patients who only present pre-diabetes and hyperglycemic events[22,23].

Additionally, several reports suggest that DM and hyperglycemia are not only associated with a higher incidence of GC but also increased mortality [21,24-26], and may even lead to drug resistance and tumor progression in GC patients [27-29].

At present, Helicobacter pylori (H. pylori) colonization is a crucial risk factor in the pathogenesis of GC. H. pylori infection leads to chronic inflammation of gastric mucosa and then leads to atrophy of the glands, intestinal metaplasia, and GC[30]. In 1994, H. pylori was classified as a Group 1 carcinogen by the International Agency for Research on Cancer^[31].

Noteworthy, recent reports have not only demonstrated an association between DM and incidence of *H. pylori* infection[32-34] but also, as a higher risk of failure in eradication therapy[35-37]. Furthermore, sustained hyperglycemia influences the expressions of several H. pylori virulence factors, leading to promote carcinogenesis [38].

During the last decade, a growing body of evidence has shed light on the mechanisms underlying this epidemiological association (Figure 1).

THE CONTRIBUTION OF DM IN REMODELING THE TME IN GC

Changing stroma cells behavior

The TME is a complex tissue niche with a diverse repertoire of infiltrating host cells, mainly recruited by cancer cells, together with many secreted factors and components of the extracellular matrix, which profoundly influence tumor growth, and dissemination[39].

A convincing body of evidence supports that chronic inflammation caused by *H*. pylori infection is the major risk factor for the development of GC, and thus various types of cells in the gastric mucosa are exposed to an inflammatory environment for long periods. The robust inflammatory response triggered by infection, together with bacterial and host factors determines the transit from the early stages of inflammation through the development of metaplasia, dysplasia, and finally to invasive carcinoma [40].

In the GC microenvironment, the behavior of many cell types of the tumor stroma is influenced by diabetes or hyperglycemia. Noteworthy, all cellular components of tumor stroma express the receptor of advanced glycation end products (RAGE), including tumor cells, and a growing body of pieces of evidence supports the role of the RAGE/advanced glycation end products (AGEs) (RAGE/AGEs) axis on tumor growth. This important modifier of the TME will be covered in the cell signaling disturbances section.

Gastric epithelial homeostasis is maintained by long-lived stem cells surrounded by a supportive niche. Therefore, GC may arise from mutated stem cells that have been accumulating gene mutations during cell half-life, and the subsequent expansion of mutated clones[41]. In this context, the chronic inflammation induced by H. pylori infection may then damage gastric epithelial mucosa, followed by the recruitment of bone marrow-derived cells (BMDCs), which may then lead to tissue remodeling, transformation, and potential progression to malignancy[42].

Of note, the diabetic condition has been described to force BMDCs to express tumor necrosis factor-alpha (TNF- α) and thus they become a contributor to fuel inflammation instead of repairing the damaged gastric mucosa[43]. Tumor-associated mast cells are also part of the cell stroma in GC[44]. These infiltrating cells play crucial roles in remodeling the TME[45], by the release of large amounts of preformed and preactivated inflammatory mediators through degranulation, and supporting tumor progression, immunosuppression, and angiogenesis[46,47].

Hyperglycemia and advanced glycation end-products are known to activate mast cells and increase the expression of proinflammatory cytokines such as $TNF-\alpha$ and favor the degranulation of mast cells[48].

Changes in stroma cell behavior	ECM remodeling	Cellular signaling disturbance	Disturbance in miRNAs network	Altered exosomes production and cargo	Altered metabolism
Involved mechanisms					
Impaired cytokines production; CAFs formation of fibrotic TME; Alternative M2 polarization of TAMs; TAMCs release of pro- inflammatory mediators	ECM stiffening; Overexpression of LOX; Activation of mechanotransduction- dependent pathways	O-GlcNac modification; Cell cycle dysregulation; Activation of TGF-β pathway; Proangiogenic transcriptional profile; Hyperactivation of RAGE/ AGE axis; Hyperactivation of Ins-R/IGF — R system	Overexpression of oncomiRs; Downregulation of tumor supressor miRNAs	Increased releasing rate of exosomes; Proinflammatory and proangiogenic molecular cargo	RAGE upregulation; HIF-1a dependent gene-transcription; Enhanced expression of ChREBP; Increased glycolytic- related gene transcription
Phenotypical changes					
Supporting tumor growth; Reduced anti-tumor immunity; Desmoplastic tumoral stroma	Highly invasive phenotypes; Enhanced cancer cell invasion	Cell death resistance; Supporting tumor angiogenesis; Supporting tumor cell proliferation	Support tumor proliferation; Evasion of growth suppressor mechamisms	Favoring tumor metastasis; Supporting tumor angiogenesis	Proangiogenic cancer cells; Hypoxic resistant phenotype; Tumor growth and progression

Diabetes contribution to the remodeling of TME in gastric cancer

Figure 1 Diabetes mellitus can sculpture the tumor microenvironment of gastric cancer through a myriad of different molecular mechanisms ranging from dysregulation of cellular signaling pathways to marked metabolic disturbances. TME: Tumor microenvironment; ECM: Extracellular matrix; miRNAs: MicroRNAs; CAFs: Cancer-associated fibroblasts; TAMCs: Tumor-associated mast cells; LOX: Lysyl oxidase; TGF-B: Transforming growth factor β; RAGE: Receptor of advanced glycation end products; AGE: Advanced glycation end product; ChREBP: Carbohydrate-responsive element-binding protein.

> Cancer-associated fibroblasts (CAFs) are prominent components of the TME, and play important roles in GC, such as tumor growth and progression, matrix remodeling, promoting angiogenesis as well as fueling inflammation[49].

> CAFs are crucial cells in the production of a desmoplastic stroma, characterized by the formation of dense fibrosis and increased remodeling and deposition of ECM components. CAFs not only produce fibrillar collagens and other interstitial ECM components but also release matrix metalloproteinases[49,50].

> Of note, desmoplasia is commonly found in patients with diabetes, where the hyperglycemic condition activates fibrogenic pathways, not only through direct stimulation of the synthesis of ECM components but also by triggering epithelial and endothelial cell conversion to a fibroblast-like phenotype[51].

> Cancer cells can recruit and activate fibroblasts in the TME by the induction of their trans-differentiation into CAFs. Recently, the serine/threonine homeodomaininteracting protein kinase 2, has been reported as a crucial regulator of this process, and its downregulation favors tumor progression^[52]. Noteworthy, hyperglycemia produces a sustained degradation of this protein[53] and thus favoring the transdifferentiating process.

> CAFs play an important role in the progression of GC, by promoting migration and epithelial to mesenchymal transition (EMT) of GC cells, and EMT is fully potentiated by hyperglycemia^[54].

> Tumor-associated macrophages (TAMs) are crucial cells in sculpturing the TME[55, 56]. Furthermore, TAMs have a prognostic significance for GC patients, when combined with the TNM staging system[57].

> TAMs are crucial cells in tuning the machinery of inflammatory and host immune responses in TME. Once infiltrated, macrophages undergo a polarization process rendering distinct functional phenotypes, and where classically activated (M1) and alternatively activated (M2) macrophages represent two extreme phenotypes [55,58]. In GC, TAMs are predominantly bearing an M2 phenotype, which is associated with cancer metastasis and a worse prognosis in patients[56-59].

> During diabetes, macrophages and other innate immune cells are known to have a pro-inflammatory phenotype, which is believed to contribute to the pathogenesis of various diabetic complications[60]. Noteworthy, hyperglycemia acts in synergy with hypoxia and sensitizes macrophage responses to cytokine stimuli[61], and generates particular M1/M2 cytokine profiles[62].



Recently, hyperglycemia is reported to induce an increased flux through the hexosamine biosynthetic pathway in TAMS resulting in an upregulation of O-GlcNAcylation, which in turn favors the alternative M2 polarization of TAMs and reduced anti-tumor immunity[63].

Tumor-infiltrating neutrophils are very abundant in the GC microenvironment, where they promote GC cell migration and invasion as well as the induction of EMT through the interleukin (IL)-17-mediated JAK2/STAT3 signaling activation, indicating that neutrophils may play an important role in GC metastasis[64,65].

Hyperglycemia is reported to impair granulocyte-colony stimulating factor secretion, thereby hindering the mobilization of antitumor neutrophils, which in turn, leads to increased survival of disseminated tumor cells and consequently increasing the metastatic burden[66].

Obesity is a common comorbidity of diabetes, and some authors have associated obesity with an increased risk of GC[67]. Obesity can impact the TME both locally, and systemically through many signals associated with visceral adipose tissue inflammation, as reported for adipokines, growth factors, and cytokines[68,69]. In this context, the activation of the STAT3 gastric signaling pathway, which is crucial in promoting the malignant transformation of epithelial cells[70], is induced by leptin and IL-6 in obese subjects[71,72].

Modification of extracellular matrix

ECM is known to be a complex non-cellular network composed mainly of glycosaminoglycans and fibrous proteins, such as collagens, fibronectin, elastin, and laminin, which give structural support to tissues and regulate diverse cellular functions such as survival, growth, migration, adhesion, and differentiation^[73].

During these cellular-ECM interactions, a complex network of signaling pathways is activated through mechanotransduction receptors, which are capable of sensing changes in the stiffness of the ECM[74,75]. At present, ECM is considered a highly dynamic element that continuously undergoes remodeling induced by several conditions^[76].

Compelling evidence support that tumor-associated ECM remodeling and stiffening, are key elements behind the TME of highly invasive phenotypes of several neoplastic cells[77-79]. Strikingly, tumor-associated ECM remodeling and the subsequent stiffening play a critical role in the behavior of cancer cells in the TME[80], and thus, supporting cancer cell survival, progression, and metastatic invasion[81].

Fibronectin and type I collagen are the most common and abundant fibrillar ECM proteins found in cancer-associated ECM[82,83]. Their increase is a result of excessive fibrotic remodeling, also referred to as desmoplasia, which is largely mediated by alpha-smooth muscle actin-expressing myofibroblasts[83,84].

Most ECM fibrous proteins are long-live potential targets for the higher rate of AGEs formation observed in DM and chronic hyperglycemic-state^[85]. AGEs crosslinks of load-bearing protein lead to ECM stiffening which favors not only tumor cell survival, but also high rates of proliferation, and metastatic cancer cell interaction with the endothelium[86,87].

Furthermore, the chronic hyperglycemia state not only mediates mechanical changes in ECM but also can generate a reservoir of AGEs with the potential to trigger a multitude of RAGE-dependent mechanisms^[85].

In addition, in vivo studies support that these posttranslational modifications of ECM produced during hyperglycemia also favor cancer cell invasion by activation of mechanotransduction-dependent epidermal growth factor receptor (EGFR) signaling pathway[88,89]. The interplay between highly glycated-ECM and increased EGFR activity could be a key element in the enhanced cancer cell invasion in DM patients.

In this context, lysyl oxidase (LOX) plays a central role in modulating the formation of molecular cross-linkages of ECM components[90]. LOX is known to play a significant role in the GC microenvironment[91]. Interestingly, the DM milieu favors the overexpression of LOX[92], which is associated with increased ECM modifications and high invasion activity of GC in DM patients[93,94].

Cellular signaling disturbances

One of the earliest pieces of evidence supporting that DM represents an active landscape for cellular signaling disturbances, coming either from the complex network of mechanisms underlying diabetes complications and the altered insulin sensitivity observed in obesity and type 2 diabetes [95,96].

At present, a growing body of evidence supports that the hyperglycemic condition can activate different signaling mechanisms, such as the polyol pathway, the advanced-glycation end-product formation, and the subsequent activation of the



RAGE/AGEs axis, as well as the activation of Protein Kinase C and hexosamine pathway. All these activated pathways may then lead to the over-expression of reactive oxygen species (ROS), activation of the transcriptional factors nuclear factorкарра beta, and consequently the increased production of proinflammatory mediators, the activation of leukocytes, as well as increased apoptosis, and a desmoplastic reaction[97,98].

These pro-inflammatory signaling pathways have supported the new concept called meta-inflammation, which is characterized by a low-grade systemic and chronic inflammation and is associated with the pathogenesis of diabetic complications[99, 100]

Hyperglycemia results in calpain-1 upregulation in the mitochondria, which in turn leads to a reduction in ATP synthase activity and increased mitochondrial ROS formation[101]. Mitochondrial ROS is crucial for the stabilization of hypoxia-inducible transcription and thus leading to an activation of a transcriptional profile supporting tumor angiogenesis[102]. Additionally, ROS is a well-known driver of myofibroblast differentiation, through the activation of the TGF-B pathway^[103], and myofibroblasts are recognized as a major source of the CAFs[104].

Chronic hyperglycemia is the hallmark of DM. This condition leads to an accelerated formation of AGEs, a heterogeneous group of compounds resulting from the non-enzymatic reaction of reducing sugars with the free amino group of proteins lipids and nucleic acids[105].

The high rate of AGEs formation in DM patients favors the overexpression of RAGE, and the hyperactivation of the RAGE/AGE axis[98,106,107]. This receptor is involved not only in the adhesion of H. pylori to gastric epithelial cells but also in the inflammatory response to infection[108].

The activation of this signaling pathway is an important contributor to inflammation-related tumorigenesis through different signaling mechanisms, including the resistance to apoptotic insults and hypoxia, interfering with antitumor immunity, stimulating angiogenesis, and supporting invasiveness^[109].

Interestingly, hyperglycemia can disturb cell cycle regulation not only in normal cells[110] but also in cancer cells[111]. High levels of glucose and insulin are reported to enhance cyclin D, cyclin-dependent kinase 4 (Cdk4), and Cdk2 expression and suppress cyclin-dependent kinase inhibitors p21 and p15/16[112].

On the other hand, the protein O-linked β-N-acetylglucosamine (O-GlcNAc) modification is a dynamic post-translational modification affecting a wide variety of proteins involved in cell cycle regulation, and this modification is considered as a major contributor to the deleterious effects of hyperglycemia [113]. This post-translational modification relies on the addition of a single N-acetyl-glucosamine molecule to the OH residues of serine or threonine by the action of the O-GlcNAc-transferase, and where some oncogenic factors, such as p53, Myc, and β -catenin, as well as other cell cycle regulators are O-GlcNAcylated[114,115] and is increased in many human neoplasias, including GC[116,117].

A growing body of evidence supports the role of the Wnt/ β -catenin pathway in the development, progression, and metastasis of GC[118,119]. High glucose levels can produce profound effects on Wnt/ β -catenin signaling in cancer cells, leading to an increased expression of WNT target genes[120] by either a sustained increment of the p300 acetyltransferase activity or decreased sirtuin 1 deacetylase activity. In this way, hyperglycemia renders high levels of β -catenin acetylation, which in turn, allows nuclear accumulation and transcriptional activation of Wnt-target genes[121].

An overactive TGF-B1 signaling pathway has been reported in diabetes patients [122] and its role as a critical profibrotic factor in the progression of chronic kidney disease in diabetes is widely documented [27,123]. The role of TGF- β in the biology of GI cancers has been extensively studied showing crucial roles in regulating processes such as tumor progression, evasion of growth suppressors, and resistance to cell death, angiogenesis, invasion, and metastasis[124].

TGF-β1 is overexpressed in GCs and the stromal tissues surrounding the cancer cells [125,126]. The main source of TGF- β 1 is stromal cells, such as fibroblasts, lymphocytes, and macrophages[127], and therefore, the coexistence of the burden due to diabetes reinforces its impact on the TME.

Another key element in the signaling network in the gastric TME is the EGFR family, which consists of four related receptor tyrosine kinases (ErbB1 to ErbB4)[128], and all members of the family are expressed in gastric tumors[129].

On the other hand, diabetic kidney disease is a common microvascular complication of DM and the leading cause of end-stage renal disease [130]. Activation of the EGFR signaling pathway is linked to the onset and progression of renal damage in DM, by promoting cell proliferation, inflammatory processes, and ECM modification[131].



Interestingly, these signaling pathways can be activated by several ligands, but also by other biological mediators such as ROS, TGF- β , and PKC, all of which are upregulated in DM[122,132,133].

The COX-2/PGE2 signaling pathway plays a critical role in the inflammatory nature of gastric tumors. COX-2 is upregulated in GC and its precursor lesions, and it provides valuable clinical information as a prognostic factor[134]. Of note, the high levels of COX-2 expression are an earlier event reported during the *H. pylori* infection of gastric mucosa^[135].

Many PGE2-mediated mechanisms supporting tumor growth, new vessel formation, and enhancing metastasis have been described [136]. Additionally, COX-2 /PGE2-mediated signaling pathways are key contributors to many diabetes complications[137,138].

Noteworthy, the activation of the RAGE/AGEs axis, which is highly expressed in diabetic tissues and cells, can significantly increase both COX-2 messenger RNA and protein expression, together with a rise in PGE2 Levels[139].

Hyperinsulinemia is strongly associated with type 2 diabetes and it has been recently postulated to be a risk factor for GC[140] Emerging data suggest that insulin may be a crucial regulator in some human neoplasias, including GC[141-145].

In addition, hyperinsulinemia also increases the hepatic production and systemic bioavailability of IGF1 but also reduces the hepatic protein production of the insulinlike growth factor binding proteins 1 (IGFBP-1) and 2 (IGFBP-2). These two coordinated actions may, in turn, hyperactivate the Ins-R/IGF1-R system, and thus triggering their proliferative and anti-apoptotic programs in cancer cells[142]. Additionally, the activation of the RAGE/AGEs axis also upregulates the expression of both IGF1 mRNA and protein levels[143].

Disturbances in microRNAs network

MicroRNAs (miRNAs) are a class of small non-coding RNAs, which act as posttranscriptional regulators of gene expression[146], and are involved in several cellular activities such as cell growth, differentiation, development, and apoptosis in many cancer types [147]. Notably, recent research has demonstrated that dysregulation in the miRNAs network has crucial consequences in the cellular behavior of neoplastic cells [147], by modulating multiple signaling pathways, especially within the gastric TME [148].

Noteworthy, hyperglycemia, and hyperinsulinemia in DM patients are two conditions that induce major changes in miRNA expression profile, especially those that are involved in gastric carcinogenesis[149].

DM patients have a particularly pro-tumoral miRNA profile, characterized by downregulation of tumor suppressor miRNAs such miR-497, miR-495p, and miR-203, which can inhibit tumor cell proliferation and migration [150,151], and its decreased expression has been associated with poor prognosis in GC patients [152,153].

In addition, the expression of some members of the family of Let-7 miRNAs, which are known by their roles in regulating oncogenes and controlling cellular differentiation and apoptosis[154], is often downregulated in several cancers, and thus derepressing some relevant oncogenic targets in GC, such as K-ras, and c-Myc[155]. Noteworthy, Let-7 miRNAs are downregulated under DM conditions not only by hyperglycemia but also by insulin resistance[151].

Conversely, a diabetogenic milieu and chronic hyperglycemia favor the overexpression of some oncomiRs, such as miR-17-5p[150,156]. Furthermore, recent research support that increased stiffness of ECM significantly induces the expression of miR-17-5p and thus rendering a loop towards the support of tumor growth and invasion[157].

Altered exosomes production and cargo

Exosomes are submicron-sized extracellular vesicles that are involved in cell-to-cell and organ-to-organ communication[158]. Recently, exosomes are involved in the pathogenesis of various disorders, including inflammatory diseases and cancer[159].

In addition, these small vesicles are now emerging as important modulators of the interchange of bioactive molecules within the TME[148].

At present, a growing body of evidence supports that either chronic hyperglycemic or hyperinsulinemic state alters not only the molecular cargo of exosomes but also their production in DM patients. These changes consequently induce critical changes in cellular function that enhance the cross-talk communication between neoplastic and non-neoplastic stromal cells[9,160].

Noteworthy, recent studies suggest that exosome release from individuals with diabetes expresses a skewed profile of pro-inflammatory molecular cargo[160,161], which influences the neoplastic transformation and favors cancer cell dissemination,





Figure 2 The molecular mechanisms involved in the contribution of diabetes mellitus to the remodeling of gastric cancer microenvironment determine crucial phenotypical changes not only on tumor cells but also in many other infiltrating cells. All changes may then result in a supporting tumor-growth niche that favors angiogenesis, invasion, and metastasis, as well as interference with anti-tumor immunity and thus generating more aggressive tumor phenotypes. ECM: Extracellular matrix.

especially in gastric tumors[162].

In addition, the altered exosomes-profile in a hyperglycemic milieu drives an increased expression of pro-angiogenic factors such as VEGF and HIF-1[160,161], which give rise to an increased intercellular cross-talk between endothelial cells, and subsequently leading to a highly angiogenic gastric microenvironment phenotype, thus promoting cancer growth and progression[163].

Besides the alterations in the molecular cargo of exosomes in the DM milieu, the increased release rate of exosomes due to hyperinsulinemia could enhance the exosome-dependent molecular transfer associated with the peritoneal dissemination of GC[160,161,164], and thus affecting the prognosis of DM patients who develop GC [165].

Altered metabolism

The Warburg effect refers to the enhanced glucose uptake and lactate production observed in cancer cells, even in the presence of oxygen and fully functioning mitochondria, also known as aerobic glycolysis[166]. Aerobic glycolysis provides glycolytic intermediates, which function as important precursors required for the synthesis of carbohydrates, fats, and proteins by cancer cells[167].

In the context of hyperglycemia, all these requirements are covered; however, hyperglycemia also promotes glycolysis by inducing the expression of glycolytic-related genes[168-170]. However, the metabolic effects of hyperglycemia on cancer cells can go further than the Warburg effect, considering that the activation of some oncogenes can subsequently proceed to an increase in ATP production[171].

Hyperglycemia also enhances the expression of the carbohydrate-responsive element-binding protein (ChREBP) in cancer cells, a well-known promoter of lipogenesis[172]. This is particularly interesting considering many tumor cells produce *de novo* almost the total of the monounsaturated and saturated fatty acids required, which are used in many cellular events crucial for tumor growth and progression[173].

In tumor cells, high glucose levels can promote HIF-1α expression under both normoxic and hypoxic conditions[168,174]. In the GC microenvironment, the HIF-1 complex activates the transcription of crucial target genes in conferring the adaptation to the hypoxic milieu and its expression correlates with an aggressive tumor phenotype and a poor prognosis[175].

In summary, DM contributes through a myriad of molecular mechanisms to the remodeling of the GC microenvironment and thus renders crucial phenotypical changes, which in turn generates tumors that are more aggressive (Figure 2).

Zaishidena® WJGO | https://www.wjgnet.com

CONCLUSION

At present, a compelling body of evidence supports the contribution of DM not only to higher cancer incidence but also to an increased mortality rate of DM patients who develops GC.

In recent years, considerable efforts in experimental research have allowed elucidating the molecular mechanisms underlying this association. In this regard, growing data suggest that the mechanical alterations induced in ECM by AGEsmediated cross-linking and the profound changes in stromal cell behavior influenced by diabetes are pivotal elements in supporting tumor growth and progression.

Furthermore, the underlying pro-inflammatory signaling supporting the metainflammation in DM patients, favors several disturbances in intra- and intercellular signaling pathways, which ultimately converge in favor of the development, progression, and dissemination of GC. In addition, the chronic metabolic dysregulation observed in DM patients favors crucial changes in tumor cell metabolism that will ultimately contribute to highly hypoxic-resistant neoplastic cells as well as to a more aggressive tumoral phenotype.

Although in recent years crucial advances have been made in the knowledge of the mechanisms induced by DM in generating a TME that is supportive of tumor growth and spread, the strengthening of clinical research is essential to achieving a better understanding of the mechanisms underlying this epidemiological association.

REFERENCES

- Ling S, Brown K, Miksza JK, Howells L, Morrison A, Issa E, Yates T, Khunti K, Davies MJ, Zaccardi F. Association of Type 2 Diabetes With Cancer: A Meta-analysis With Bias Analysis for Unmeasured Confounding in 151 Cohorts Comprising 32 Million People. Diabetes Care 2020; 43: 2313-2322 [PMID: 32910779 DOI: 10.2337/dc20-0204]
- 2 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- 3 Yang HJ, Kang D, Chang Y, Ahn J, Ryu S, Cho J, Guallar E, Sohn CI. Diabetes mellitus is associated with an increased risk of gastric cancer: a cohort study. Gastric Cancer 2020; 23: 382-390 [PMID: 31853749 DOI: 10.1007/s10120-019-01033-8]
- Miao ZF, Xu H, Xu YY, Wang ZN, Zhao TT, Song YX, Xu HM. Diabetes mellitus and the risk of 4 gastric cancer: a meta-analysis of cohort studies. Oncotarget 2017; 8: 44881-44892 [PMID: 28415651 DOI: 10.18632/oncotarget.16487]
- Donath MY, Shoelson SE. Type 2 diabetes as an inflammatory disease. Nat Rev Immunol 2011; 11: 98-107 [PMID: 21233852 DOI: 10.1038/nri2925]
- Hameed I, Masoodi SR, Mir SA, Nabi M, Ghazanfar K, Ganai BA. Type 2 diabetes mellitus: From 6 a metabolic disorder to an inflammatory condition. World J Diabetes 2015; 6: 598-612 [PMID: 25987957 DOI: 10.4239/wjd.v6.i4.598]
- Gross B, Pawlak M, Lefebvre P, Staels B. PPARs in obesity-induced T2DM, dyslipidaemia and 7 NAFLD. Nat Rev Endocrinol 2017; 13: 36-49 [PMID: 27636730 DOI: 10.1038/nrendo.2016.135]
- 8 Yaribeygi H, Sathyapalan T, Atkin SL, Sahebkar A. Molecular Mechanisms Linking Oxidative Stress and Diabetes Mellitus. Oxid Med Cell Longev 2020; 2020: 8609213 [PMID: 32215179 DOI: 10.1155/2020/8609213]
- Noren Hooten N, Evans MK. Extracellular vesicles as signaling mediators in type 2 diabetes mellitus. Am J Physiol Cell Physiol 2020; 318: C1189-C1199 [PMID: 32348178 DOI: 10.1152/ajpcell.00536.2019
- Oya Y, Hayakawa Y, Koike K. Tumor microenvironment in gastric cancers. Cancer Sci 2020; 111: 10 2696-2707 [PMID: 32519436 DOI: 10.1111/cas.14521]
- Tseng CH, Tseng FH. Diabetes and gastric cancer: the potential links. World J Gastroenterol 2014; 11 20: 1701-1711 [PMID: 24587649 DOI: 10.3748/wjg.v20.i7.1701]
- 12 Barone BB, Yeh HC, Snyder CF, Peairs KS, Stein KB, Derr RL, Wolff AC, Brancati FL. Long-term all-cause mortality in cancer patients with preexisting diabetes mellitus: a systematic review and meta-analysis. JAMA 2008; 300: 2754-2764 [PMID: 19088353 DOI: 10.1001/jama.2008.824]
- Harding JL, Andes LJ, Gregg EW, Cheng YJ, Weir HK, Bullard KM, Burrows NR, Imperatore G. 13 Trends in cancer mortality among people with vs without diabetes in the USA, 1988-2015. Diabetologia 2020; 63: 75-84 [PMID: 31511931 DOI: 10.1007/s00125-019-04991-x]
- 14 Goto A, Yamaji T, Sawada N, Momozawa Y, Kamatani Y, Kubo M, Shimazu T, Inoue M, Noda M, Tsugane S, Iwasaki M. Diabetes and cancer risk: A Mendelian randomization study. Int J Cancer 2020; 146: 712-719 [PMID: 30927373 DOI: 10.1002/ijc.32310]
- 15 Ling S, Brown K, Miksza JK, Howells LM, Morrison A, Issa E, Yates T, Khunti K, Davies MJ, Zaccardi F. Risk of cancer incidence and mortality associated with diabetes: A systematic review with trend analysis of 203 cohorts. Nutr Metab Cardiovasc Dis 2021; 31: 14-22 [PMID: 33223399



DOI: 10.1016/j.numecd.2020.09.023]

- 16 Khan M, Mori M, Fujino Y, Shibata A, Sakauchi F, Washio M, Tamakoshi A; Japan Collaborative Cohort Study Group. Site-specific cancer risk due to diabetes mellitus history: evidence from the Japan Collaborative Cohort (JACC) Study. Asian Pac J Cancer Prev 2006; 7: 253-259 [PMID: 16839219]
- 17 Inoue M, Iwasaki M, Otani T, Sasazuki S, Noda M, Tsugane S. Diabetes mellitus and the risk of cancer: results from a large-scale population-based cohort study in Japan. Arch Intern Med 2006; 166: 1871-1877 [PMID: 17000944 DOI: 10.1001/archinte.166.17.1871]
- 18 Hong SH, Noh E, Kim J, Hwang SY, Kim JA, Lee YB, Roh E, Choi KM, Baik SH, Cho GJ, Yoo HJ. Fasting Plasma Glucose Variability and Gastric Cancer Risk in Individuals Without Diabetes Mellitus: A Nationwide Population-Based Cohort Study. Clin Transl Gastroenterol 2020; 11: e00221 [PMID: 32858572 DOI: 10.14309/ctg.00000000000221]
- 19 Ge Z, Ben Q, Qian J, Wang Y, Li Y. Diabetes mellitus and risk of gastric cancer: a systematic review and meta-analysis of observational studies. Eur J Gastroenterol Hepatol 2011; 23: 1127-1135 [PMID: 21934509 DOI: 10.1097/MEG.0b013e32834b8d73]
- 20 Shimoyama S. Diabetes mellitus carries a risk of gastric cancer: a meta-analysis. World J Gastroenterol 2013; 19: 6902-6910 [PMID: 24187468 DOI: 10.3748/wjg.v19.i40.6902]
- 21 Tian T, Zhang LQ, Ma XH, Zhou JN, Shen J. Diabetes mellitus and incidence and mortality of gastric cancer: a meta-analysis. Exp Clin Endocrinol Diabetes 2012; 120: 217-223 [PMID: 22187293 DOI: 10.1055/s-0031-1297969]
- 22 Ikeda F, Doi Y, Yonemoto K, Ninomiya T, Kubo M, Shikata K, Hata J, Tanizaki Y, Matsumoto T, Iida M, Kiyohara Y. Hyperglycemia increases risk of gastric cancer posed by Helicobacter pylori infection: a population-based cohort study. Gastroenterology 2009; 136: 1234-1241 [PMID: 19236964 DOI: 10.1053/j.gastro.2008.12.045]
- 23 Huang Y, Cai X, Qiu M, Chen P, Tang H, Hu Y, Huang Y. Prediabetes and the risk of cancer: a meta-analysis. Diabetologia 2014; 57: 2261-2269 [PMID: 25208757 DOI: 10.1007/s00125-014-3361-2
- 24 Tseng CH. Diabetes conveys a higher risk of gastric cancer mortality despite an age-standardised decreasing trend in the general population in Taiwan. Gut 2011; 60: 774-779 [PMID: 21193459 DOI: 10.1136/gut.2010.226522]
- 25 Li PF, Chen WL. Are the Different Diabetes Subgroups Correlated With All-Cause, Cancer-Related, and Cardiovascular-Related Mortality? J Clin Endocrinol Metab 2020; 105 [PMID: 32893854 DOI: 10.1210/clinem/dgaa628]
- 26 Population-based cohort study of diabetes mellitus and mortality in gastric adenocarcinoma. Br J Surg 2020; 107: 1012 [PMID: 32539215 DOI: 10.1002/bjs.11884]
- 27 Zhao W, Chen R, Zhao M, Li L, Fan L, Che XM. High glucose promotes gastric cancer chemoresistance in vivo and in vitro. Mol Med Rep 2015; 12: 843-850 [PMID: 25815791 DOI: 10.3892/mmr.2015.3522]
- 28 Li W, Zhang X, Sang H, Zhou Y, Shang C, Wang Y, Zhu H. Effects of hyperglycemia on the progression of tumor diseases. J Exp Clin Cancer Res 2019; 38: 327 [PMID: 31337431 DOI: 10.1186/s13046-019-1309-6]
- 29 Xu X, Chen B, Zhu S, Zhang J, He X, Cao G. Hyperglycemia promotes Snail-induced epithelialmesenchymal transition of gastric cancer via activating ENO1 expression. Cancer Cell Int 2019; 19: 344 [PMID: 31889896 DOI: 10.1186/s12935-019-1075-8]
- 30 Correa P. Human gastric carcinogenesis: a multistep and multifactorial process--First American Cancer Society Award Lecture on Cancer Epidemiology and Prevention. Cancer Res 1992; 52: 6735-6740 [PMID: 1458460]
- Schistosomes, liver flukes and Helicobacter pylori. IARC Working Group on the Evaluation of 31 Carcinogenic Risks to Humans. Lyon, 7-14 June 1994. IARC Monogr Eval Carcinog Risks Hum 1994; 61: 1-241 [PMID: 7715068]
- 32 Chen J, Xing Y, Zhao L, Ma H. The Association between Helicobacter pylori Infection and Glycated Hemoglobin A in Diabetes: A Meta-Analysis. J Diabetes Res 2019; 2019: 3705264 [PMID: 31583248 DOI: 10.1155/2019/3705264]
- 33 Youn Nam S, Park BJ, Nam JH, Ryu KH, Kook MC, Kim J, Lee WK. Association of current Helicobacter pylori infection and metabolic factors with gastric cancer in 35,519 subjects: A crosssectional study. United European Gastroenterol J 2019; 7: 287-296 [PMID: 31080613 DOI: 10.1177/2050640618819402]
- 34 Mansori K, Dehghanbanadaki H, Naderpour S, Rashti R, Moghaddam AB, Moradi Y. A systematic review and meta-analysis of the prevalence of Helicobacter pylori in patients with diabetes. Diabetes Metab Syndr 2020; 14: 601-607 [PMID: 32417710 DOI: 10.1016/j.dsx.2020.05.009]
- Nam SJ, Park SC, Lee SH, Choi DW, Lee SJ, Bang CS, Baik GH, Park JK. Helicobacter pylori 35 eradication in patients with type 2 diabetes mellitus: Multicenter prospective observational study. SAGE Open Med 2019; 7: 2050312119832093 [PMID: 30815260 DOI: 10.1177/2050312119832093
- 36 Yao CC, Kuo CM, Hsu CN, Yang SC, Wu CK, Tai WC, Liang CM, Wu KL, Huang CF, Bi KW, Lee CH, Chuah SK. First-line Helicobacter pylori eradication rates are significantly lower in patients with than those without type 2 diabetes mellitus. Infect Drug Resist 2019; 12: 1425-1431 [PMID: 31239721 DOI: 10.2147/IDR.S194584]
- 37 Ojetti V, Pitocco D, Bartolozzi F, Danese S, Migneco A, Lupascu A, Pola P, Ghirlanda G,



Gasbarrini G, Gasbarrini A. High rate of helicobacter pylori re-infection in patients affected by type 1 diabetes. Diabetes Care 2002; 25: 1485 [PMID: 12145262 DOI: 10.2337/diacare.25.8.1485]

- 38 Sheu SM, Cheng H, Kao CY, Yang YJ, Wu JJ, Sheu BS. Higher glucose level can enhance the H. pylori adhesion and virulence related with type IV secretion system in AGS cells. J Biomed Sci 2014; 21: 96 [PMID: 25296847 DOI: 10.1186/s12929-014-0096-9]
- 39 Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell 2011; 144: 646-674 [PMID: 21376230 DOI: 10.1016/j.cell.2011.02.013]
- 40 Correa P, Piazuelo MB. The gastric precancerous cascade. J Dig Dis 2012; 13: 2-9 [PMID: 22188910 DOI: 10.1111/j.1751-2980.2011.00550.x]
- Brungs D, Aghmesheh M, Vine KL, Becker TM, Carolan MG, Ranson M. Gastric cancer stem cells: 41 evidence, potential markers, and clinical implications. J Gastroenterol 2016; 51: 313-326 [PMID: 26428661 DOI: 10.1007/s00535-015-1125-5]
- 42 Houghton J, Stoicov C, Nomura S, Rogers AB, Carlson J, Li H, Cai X, Fox JG, Goldenring JR, Wang TC. Gastric cancer originating from bone marrow-derived cells. Science 2004; 306: 1568-1571 [PMID: 15567866 DOI: 10.1126/science.1099513]
- 43 Nobuta H, Katagi M, Kume S, Terashima T, Araki SI, Maegawa H, Kojima H, Nakagawa T. A role for bone marrow-derived cells in diabetic nephropathy. FASEB J 2019; 33: 4067-4076 [PMID: 30496699 DOI: 10.1096/fj.201801825R]
- Ribatti D, Guidolin D, Marzullo A, Nico B, Annese T, Benagiano V, Crivellato E. Mast cells and angiogenesis in gastric carcinoma. Int J Exp Pathol 2010; 91: 350-356 [PMID: 20412338 DOI: 10.1111/j.1365-2613.2010.00714.x
- 45 Liu J, Zhang Y, Zhao J, Yang Z, Li D, Katirai F, Huang B. Mast cell: insight into remodeling a tumor microenvironment. Cancer Metastasis Rev 2011; 30: 177-184 [PMID: 21267769 DOI: 10.1007/s10555-011-9276-1]
- Zhong B, Li Y, Liu X, Wang D. Association of mast cell infiltration with gastric cancer progression. 46 Oncol Lett 2018; 15: 755-764 [PMID: 29422964 DOI: 10.3892/ol.2017.7380]
- 47 Lv Y, Zhao Y, Wang X, Chen N, Mao F, Teng Y, Wang T, Peng L, Zhang J, Cheng P, Liu Y, Kong H, Chen W, Hao C, Han B, Ma Q, Zou Q, Chen J, Zhuang Y. Increased intratumoral mast cells foster immune suppression and gastric cancer progression through TNF-α-PD-L1 pathway. J Immunother Cancer 2019; 7: 54 [PMID: 30808413 DOI: 10.1186/s40425-019-0530-3]
- Nagai K, Fukushima T, Oike H, Kobori M. High glucose increases the expression of 48 proinflammatory cytokines and secretion of TNF α and β -hexosaminidase in human mast cells. Eur J Pharmacol 2012; 687: 39-45 [PMID: 22575517 DOI: 10.1016/j.ejphar.2012.04.038]
- 49 Ping Q, Yan R, Cheng X, Wang W, Zhong Y, Hou Z, Shi Y, Wang C, Li R. Cancer-associated fibroblasts: overview, progress, challenges, and directions. Cancer Gene Ther 2021; 28: 984-999 [PMID: 33712707 DOI: 10.1038/s41417-021-00318-4]
- 50 Kalluri R. The biology and function of fibroblasts in cancer. Nat Rev Cancer 2016; 16: 582-598 [PMID: 27550820 DOI: 10.1038/nrc.2016.73]
- 51 Tuleta I, Frangogiannis NG. Diabetic fibrosis. Biochim Biophys Acta Mol Basis Dis 2021; 1867: 166044 [PMID: 33378699 DOI: 10.1016/j.bbadis.2020.166044]
- Garufi A, Traversi G, Cirone M, D'Orazi G. HIPK2 role in the tumor-host interaction: Impact on 52 fibroblasts transdifferentiation CAF-like. IUBMB Life 2019; 71: 2055-2061 [PMID: 31414572 DOI: 10.1002/jub.2144]
- Baldari S, Garufi A, Granato M, Cuomo L, Pistritto G, Cirone M, D'Orazi G. Hyperglycemia 53 triggers HIPK2 protein degradation. Oncotarget 2017; 8: 1190-1203 [PMID: 27901482 DOI: 10.18632/oncotarget.13595
- 54 Osório H, Silva C, Ferreira M, Gullo I, Máximo V, Barros R, Mendonça F, Oliveira C, Carneiro F. Proteomics Analysis of Gastric Cancer Patients with Diabetes Mellitus. J Clin Med 2021; 10 [PMID: 33494396 DOI: 10.3390/jcm10030407]
- 55 Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. Trends Immunol 2002; 23: 549-555 [PMID: 12401408 DOI: 10.1016/s1471-4906(02)02302-5]
- Gambardella V, Castillo J, Tarazona N, Gimeno-Valiente F, Martínez-Ciarpaglini C, Cabeza-56 Segura M, Roselló S, Roda D, Huerta M, Cervantes A, Fleitas T. The role of tumor-associated macrophages in gastric cancer development and their potential as a therapeutic target. Cancer Treat Rev 2020; 86: 102015 [PMID: 32248000 DOI: 10.1016/j.ctrv.2020.102015]
- 57 Zhang QW, Liu L, Gong CY, Shi HS, Zeng YH, Wang XZ, Zhao YW, Wei YQ. Prognostic significance of tumor-associated macrophages in solid tumor: a meta-analysis of the literature. PLoS One 2012; 7: e50946 [PMID: 23284651 DOI: 10.1371/journal.pone.0050946]
- 58 Biswas SK, Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. Nat Immunol 2010; 11: 889-896 [PMID: 20856220 DOI: 10.1038/ni.1937]
- 59 Yamaguchi T, Fushida S, Yamamoto Y, Tsukada T, Kinoshita J, Oyama K, Miyashita T, Tajima H, Ninomiya I, Munesue S, Harashima A, Harada S, Yamamoto H, Ohta T. Tumor-associated macrophages of the M2 phenotype contribute to progression in gastric cancer with peritoneal dissemination. Gastric Cancer 2016; 19: 1052-1065 [PMID: 26621525 DOI: 10.1007/s10120-015-0579-8]
- Tesch GH. Role of macrophages in complications of type 2 diabetes. Clin Exp Pharmacol Physiol 2007; **34**: 1016-1019 [PMID: 17714088 DOI: 10.1111/j.1440-1681.2007.04729.x]
- 61 Pavlou S, Lindsay J, Ingram R, Xu H, Chen M. Sustained high glucose exposure sensitizes



macrophage responses to cytokine stimuli but reduces their phagocytic activity. BMC Immunol 2018; 19: 24 [PMID: 29996768 DOI: 10.1186/s12865-018-0261-0]

- 62 Moganti K, Li F, Schmuttermaier C, Riemann S, Klüter H, Gratchev A, Harmsen MC, Kzhyshkowska J. Hyperglycemia induces mixed M1/M2 cytokine profile in primary human monocyte-derived macrophages. Immunobiology 2017; 222: 952-959 [PMID: 27492721 DOI: 10.1016/j.imbio.2016.07.006
- 63 Rodrigues Mantuano N, Stanczak MA, Oliveira IA, Kirchhammer N, Filardy AA, Monaco G, Santos RC, Fonseca AC, Fontes M, Bastos CS Jr, Dias WB, Zippelius A, Todeschini AR, Läubli H. Hyperglycemia Enhances Cancer Immune Evasion by Inducing Alternative Macrophage Polarization through Increased O-GlcNAcylation. Cancer Immunol Res 2020; 8: 1262-1272 [PMID: 32819969 DOI: 10.1158/2326-6066.CIR-19-0904]
- Zhang W, Gu J, Chen J, Zhang P, Ji R, Qian H, Xu W, Zhang X. Interaction with neutrophils 64 promotes gastric cancer cell migration and invasion by inducing epithelial-mesenchymal transition. Oncol Rep 2017; 38: 2959-2966 [PMID: 28901479 DOI: 10.3892/or.2017.5942]
- 65 Li S, Cong X, Gao H, Lan X, Li Z, Wang W, Song S, Wang Y, Li C, Zhang H, Zhao Y, Xue Y. Tumor-associated neutrophils induce EMT by IL-17a to promote migration and invasion in gastric cancer cells. J Exp Clin Cancer Res 2019; 38: 6 [PMID: 30616627 DOI: 10.1186/s13046-018-1003-0]
- Fainsod-Levi T, Gershkovitz M, Völs S, Kumar S, Khawaled S, Sagiv JY, Sionov RV, Grunewald M, Keshet E, Granot Z. Hyperglycemia Impairs Neutrophil Mobilization Leading to Enhanced Metastatic Seeding. Cell Rep 2017; 21: 2384-2392 [PMID: 29186678 DOI: 10.1016/j.celrep.2017.11.010]
- 67 Yang P, Zhou Y, Chen B, Wan HW, Jia GQ, Bai HL, Wu XT. Overweight, obesity and gastric cancer risk: results from a meta-analysis of cohort studies. Eur J Cancer 2009; 45: 2867-2873 [PMID: 19427197 DOI: 10.1016/j.ejca.2009.04.019]
- Nieman KM, Romero IL, Van Houten B, Lengyel E. Adipose tissue and adipocytes support 68 tumorigenesis and metastasis. Biochim Biophys Acta 2013; 1831: 1533-1541 [PMID: 23500888 DOI: 10.1016/j.bbalip.2013.02.010]
- 69 Dumas JF, Brisson L. Interaction between adipose tissue and cancer cells: role for cancer progression. Cancer Metastasis Rev 2021; 40: 31-46 [PMID: 33009650 DOI: 10.1007/s10555-020-09934-2]
- 70 Iliopoulos D, Jaeger SA, Hirsch HA, Bulyk ML, Struhl K. STAT3 activation of miR-21 and miR-181b-1 via PTEN and CYLD are part of the epigenetic switch linking inflammation to cancer. Mol Cell 2010; 39: 493-506 [PMID: 20797623 DOI: 10.1016/j.molcel.2010.07.023]
- 71 Zheng J, Zhao M, Li J, Lou G, Yuan Y, Bu S, Xi Y. Obesity-associated digestive cancers: A review of mechanisms and interventions. Tumour Biol 2017; 39: 1010428317695020 [PMID: 28351315 DOI: 10.1177/1010428317695020]
- O'Sullivan J, Lysaght J, Donohoe CL, Reynolds JV. Obesity and gastrointestinal cancer: the 72 interrelationship of adipose and tumour microenvironments. Nat Rev Gastroenterol Hepatol 2018; 15: 699-714 [PMID: 30323319 DOI: 10.1038/s41575-018-0069-7]
- 73 McKee TJ, Perlman G, Morris M, Komarova SV. Extracellular matrix composition of connective tissues: a systematic review and meta-analysis. Sci Rep 2019; 9: 10542 [PMID: 31332239 DOI: 10.1038/s41598-019-46896-0]
- 74 Romani P, Valcarcel-Jimenez L, Frezza C, Dupont S. Crosstalk between mechanotransduction and metabolism. Nat Rev Mol Cell Biol 2021; 22: 22-38 [PMID: 33188273 DOI: 10.1038/s41580-020-00306-w
- 75 Winkler J, Abisoye-Ogunniyan A, Metcalf KJ, Werb Z. Concepts of extracellular matrix remodelling in tumour progression and metastasis. Nat Commun 2020; 11: 5120 [PMID: 33037194 DOI: 10.1038/s41467-020-18794-x]
- Manou D, Caon I, Bouris P, Triantaphyllidou IE, Giaroni C, Passi A, Karamanos NK, Vigetti D, 76 Theocharis AD. The Complex Interplay Between Extracellular Matrix and Cells in Tissues. Methods Mol Biol 2019; 1952: 1-20 [PMID: 30825161 DOI: 10.1007/978-1-4939-9133-4 1]
- Najafi M, Farhood B, Mortezaee K. Extracellular matrix (ECM) stiffness and degradation as cancer 77 drivers. J Cell Biochem 2019; 120: 2782-2790 [PMID: 30321449 DOI: 10.1002/jcb.27681]
- Poltavets V, Kochetkova M, Pitson SM, Samuel MS. The Role of the Extracellular Matrix and Its 78 Molecular and Cellular Regulators in Cancer Cell Plasticity. Front Oncol 2018; 8: 431 [PMID: 30356678 DOI: 10.3389/fonc.2018.00431]
- Walker C, Mojares E, Del Río Hernández A. Role of Extracellular Matrix in Development and 79 Cancer Progression. Int J Mol Sci 2018; 19 [PMID: 30287763 DOI: 10.3390/ijms19103028]
- Rigoglio NN, Rabelo ACS, Borghesi J, de Sá Schiavo Matias G, Fratini P, Prazeres PHDM, 80 Pimentel CMMM, Birbrair A, Miglino MA. The Tumor Microenvironment: Focus on Extracellular Matrix. Adv Exp Med Biol 2020; 1245: 1-38 [PMID: 32266651 DOI: 10.1007/978-3-030-40146-7 1]
- Dandia H, Makkad K, Tayalia P. Glycated collagen a 3D matrix system to study pathological cell 81 behavior. Biomater Sci 2019; 7: 3480-3488 [PMID: 31282511 DOI: 10.1039/c9bm00184k]
- Spada S, Tocci A, Di Modugno F, Nisticò P. Fibronectin as a multiregulatory molecule crucial in 82 tumor matrisome: from structural and functional features to clinical practice in oncology. J Exp Clin Cancer Res 2021; 40: 102 [PMID: 33731188 DOI: 10.1186/s13046-021-01908-8]
- 83 Kobayashi H, Enomoto A, Woods SL, Burt AD, Takahashi M, Worthley DL. Cancer-associated



fibroblasts in gastrointestinal cancer. Nat Rev Gastroenterol Hepatol 2019; 16: 282-295 [PMID: 30778141 DOI: 10.1038/s41575-019-0115-0]

- 84 Mohan V, Das A, Sagi I. Emerging roles of ECM remodeling processes in cancer. Semin Cancer Biol 2020; 62: 192-200 [PMID: 31518697 DOI: 10.1016/j.semcancer.2019.09.004]
- 85 Rojas A, Añazco C, González I, Araya P. Extracellular matrix glycation and receptor for advanced glycation end-products activation: a missing piece in the puzzle of the association between diabetes and cancer. Carcinogenesis 2018; 39: 515-521 [PMID: 29373651 DOI: 10.1093/carcin/bgy012]
- 86 Moreira AM, Pereira J, Melo S, Fernandes MS, Carneiro P, Seruca R, Figueiredo J. The Extracellular Matrix: An Accomplice in Gastric Cancer Development and Progression. Cells 2020; 9 [PMID: 32046329 DOI: 10.3390/cells9020394]
- Suh YJ, Hall MS, Huang YL, Moon SY, Song W, Ma M, Bonassar LJ, Segall JE, Wu M. Glycation 87 of collagen matrices promotes breast tumor cell invasion. Integr Biol (Camb) 2019 [PMID: 31041443 DOI: 10.1093/intbio/zyz011]
- 88 Grasset EM, Bertero T, Bozec A, Friard J, Bourget I, Pisano S, Lecacheur M, Maiel M, Bailleux C, Emelyanov A, Ilie M, Hofman P, Meneguzzi G, Duranton C, Bulavin DV, Gaggioli C. Matrix Stiffening and EGFR Cooperate to Promote the Collective Invasion of Cancer Cells. Cancer Res 2018; 78: 5229-5242 [PMID: 30026329 DOI: 10.1158/0008-5472.CAN-18-0601]
- Wei SC, Fattet L, Tsai JH, Guo Y, Pai VH, Majeski HE, Chen AC, Sah RL, Taylor SS, Engler AJ, Yang J. Matrix stiffness drives epithelial-mesenchymal transition and tumour metastasis through a TWIST1-G3BP2 mechanotransduction pathway. Nat Cell Biol 2015; 17: 678-688 [PMID: 25893917 DOI: 10.1038/ncb3157]
- Laczko R, Csiszar K. Lysyl Oxidase (LOX): Functional Contributions to Signaling Pathways. 90 Biomolecules 2020; 10 [PMID: 32708046 DOI: 10.3390/biom10081093]
- Añazco C, Delgado-López F, Araya P, González I, Morales E, Pérez-Castro R, Romero J, Rojas A. 91 Lysyl oxidase isoforms in gastric cancer. Biomark Med 2016; 10: 987-998 [PMID: 27564724 DOI: 10.2217/bmm-2016-0075]
- 92 Chronopoulos A, Tang A, Beglova E, Trackman PC, Roy S. High glucose increases lysyl oxidase expression and activity in retinal endothelial cells: mechanism for compromised extracellular matrix barrier function. Diabetes 2010; 59: 3159-3166 [PMID: 20823103 DOI: 10.2337/db10-0365]
- 93 Li Q, Zhu CC, Ni B, Zhang ZZ, Jiang SH, Hu LP, Wang X, Zhang XX, Huang PQ, Yang Q, Li J, Gu JR, Xu J, Luo KQ, Zhao G, Zhang ZG. Lysyl oxidase promotes liver metastasis of gastric cancer via facilitating the reciprocal interactions between tumor cells and cancer associated fibroblasts. EBioMedicine 2019; 49: 157-171 [PMID: 31678002 DOI: 10.1016/j.ebiom.2019.10.037]
- 94 Trackman PC. Lysyl Oxidase Isoforms and Potential Therapeutic Opportunities for Fibrosis and Cancer. Expert Opin Ther Targets 2016; 20: 935-945 [PMID: 26848785 DOI: 10.1517/14728222.2016.1151003
- Zand H, Morshedzadeh N, Naghashian F. Signaling pathways linking inflammation to insulin 95 resistance. Diabetes Metab Syndr 2017; 11 Suppl 1: S307-S309 [PMID: 28365222 DOI: 10.1016/j.dsx.2017.03.006]
- 96 Cheng F, Carroll L, Joglekar MV, Januszewski AS, Wong KK, Hardikar AA, Jenkins AJ, Ma RCW. Diabetes, metabolic disease, and telomere length. Lancet Diabetes Endocrinol 2021; 9: 117-126 [PMID: 33248477 DOI: 10.1016/S2213-8587(20)30365-X]
- 97 Brownlee M. Biochemistry and molecular cell biology of diabetic complications. Nature 2001; 414: 813-820 [PMID: 11742414 DOI: 10.1038/414813a]
- Yan SF, Ramasamy R, Naka Y, Schmidt AM. Glycation, inflammation, and RAGE: a scaffold for 98 the macrovascular complications of diabetes and beyond. Circ Res 2003; 93: 1159-1169 [PMID: 14670831 DOI: 10.1161/01.RES.0000103862.26506.3D]
- 99 Hotamisligil GS. Inflammation and metabolic disorders. Nature 2006; 444: 860-867 [PMID: 17167474 DOI: 10.1038/nature05485]
- 100 Diedisheim M, Carcarino E, Vandiedonck C, Roussel R, Gautier JF, Venteclef N. Regulation of inflammation in diabetes: From genetics to epigenomics evidence. Mol Metab 2020; 41: 101041 [PMID: 32603690 DOI: 10.1016/j.molmet.2020.101041]
- Ni R, Zheng D, Xiong S, Hill DJ, Sun T, Gardiner RB, Fan GC, Lu Y, Abel ED, Greer PA, Peng T. 101 Mitochondrial Calpain-1 Disrupts ATP Synthase and Induces Superoxide Generation in Type 1 Diabetic Hearts: A Novel Mechanism Contributing to Diabetic Cardiomyopathy. Diabetes 2016; 65: 255-268 [PMID: 26470784 DOI: 10.2337/db15-0963]
- Weinberg F, Ramnath N, Nagrath D. Reactive Oxygen Species in the Tumor Microenvironment: 102 An Overview. Cancers (Basel) 2019; 11 [PMID: 31426364 DOI: 10.3390/cancers11081191]
- 103 Jain M, Rivera S, Monclus EA, Synenki L, Zirk A, Eisenbart J, Feghali-Bostwick C, Mutlu GM, Budinger GR, Chandel NS. Mitochondrial reactive oxygen species regulate transforming growth factor-β signaling. *J Biol Chem* 2013; **288**: 770-777 [PMID: 23204521 DOI: 10.1074/jbc.M112.431973
- 104 Schmitt-Gräff A, Desmoulière A, Gabbiani G. Heterogeneity of myofibroblast phenotypic features: an example of fibroblastic cell plasticity. Virchows Arch 1994; 425: 3-24 [PMID: 7921410 DOI: 10.1007/BF00193944
- 105 Vlassara H, Palace MR. Diabetes and advanced glycation endproducts. J Intern Med 2002; 251: 87-101 [PMID: 11905595 DOI: 10.1046/j.1365-2796.2002.00932.x]
- 106 Yan SF, Ramasamy R, Schmidt AM. The RAGE axis: a fundamental mechanism signaling danger to the vulnerable vasculature. Circ Res 2010; 106: 842-853 [PMID: 20299674 DOI:



10.1161/CIRCRESAHA.109.212217]

- 107 Yan SF, Ramasamy R, Schmidt AM. Mechanisms of disease: advanced glycation end-products and their receptor in inflammation and diabetes complications. Nat Clin Pract Endocrinol Metab 2008; 4: 285-293 [PMID: 18332897 DOI: 10.1038/ncpendmet0786]
- 108 Rojas A, González I, Rodríguez B, Romero J, Figueroa H, Llanos J, Morales E, Pérez-Castro R. Evidence of involvement of the receptor for advanced glycation end-products (RAGE) in the adhesion of Helicobacter pylori to gastric epithelial cells. Microbes Infect 2011; 13: 818-823 [PMID: 21609778 DOI: 10.1016/j.micinf.2011.04.005]
- 109 Rojas A, Figueroa H, Morales E. Fueling inflammation at tumor microenvironment: the role of multiligand/RAGE axis. Carcinogenesis 2010; 31: 334-341 [PMID: 20028726 DOI: 10.1093/carcin/bgp322]
- Wolf G. Cell cycle regulation in diabetic nephropathy. Kidney Int Suppl 2000; 77: S59-S66 [PMID: 110 10997692 DOI: 10.1046/j.1523-1755.2000.07710.x]
- 111 Li X, Li J, Cai Y, Peng S, Wang J, Xiao Z, Wang Y, Tao Y, Leng Q, Wu D, Yang S, Ji Z, Han Y, Li L, Gao X, Zeng C, Wen X. Hyperglycaemia-induced miR-301a promotes cell proliferation by repressing p21 and Smad4 in prostate cancer. Cancer Lett 2018; 418: 211-220 [PMID: 29331421 DOI: 10.1016/j.canlet.2018.01.031]
- 112 Kim D, Ahn BN, Kim Y, Hur DY, Yang JW, Park GB, Jang JE, Lee EJ, Kwon MJ, Kim TN, Kim MK, Park JH, Rhee BD, Lee SH. High Glucose with Insulin Induces Cell Cycle Progression and Activation of Oncogenic Signaling of Bladder Epithelial Cells Cotreated with Metformin and Pioglitazone. J Diabetes Res 2019; 2019: 2376512 [PMID: 30729133 DOI: 10.1155/2019/2376512]
- 113 Nagy T, Fisi V, Frank D, Kátai E, Nagy Z, Miseta A. Hyperglycemia-Induced Aberrant Cell Proliferation; A Metabolic Challenge Mediated by Protein O-GlcNAc Modification. Cells 2019; 8 [PMID: 31466420 DOI: 10.3390/cells8090999]
- 114 Sakabe K, Wang Z, Hart GW. Beta-N-acetylglucosamine (O-GlcNAc) is part of the histone code. *Proc Natl Acad Sci U S A* 2010; **107**: 19915-19920 [PMID: 21045127 DOI: 10.1073/pnas.1009023107
- Fardini Y, Dehennaut V, Lefebvre T, Issad T. O-GlcNAcylation: A New Cancer Hallmark? Front 115 Endocrinol (Lausanne) 2013; 4: 99 [PMID: 23964270 DOI: 10.3389/fendo.2013.00099]
- 116 Forma E, Jóźwiak P, Bryś M, Krześlak A. The potential role of O-GlcNAc modification in cancer epigenetics. Cell Mol Biol Lett 2014; 19: 438-460 [PMID: 25141978 DOI: 10.2478/s11658-014-0204-6
- Jiang M, Qiu Z, Zhang S, Fan X, Cai X, Xu B, Li X, Zhou J, Zhang X, Chu Y, Wang W, Liang J, 117 Horvath T, Yang X, Wu K, Nie Y, Fan D. Elevated O-GlcNAcylation promotes gastric cancer cells proliferation by modulating cell cycle related proteins and ERK 1/2 signaling. Oncotarget 2016; 7: 61390-61402 [PMID: 27542217 DOI: 10.18632/oncotarget.11359]
- 118 **Chiurillo MA**. Role of the Wnt/ β -catenin pathway in gastric cancer: An in-depth literature review. World J Exp Med 2015; 5: 84-102 [PMID: 25992323 DOI: 10.5493/wjem.v5.i2.84]
- Zhan T, Rindtorff N, Boutros M. Wnt signaling in cancer. Oncogene 2017; 36: 1461-1473 [PMID: 119 27617575 DOI: 10.1038/onc.2016.304]
- 120 Chocarro-Calvo A, García-Martínez JM, Ardila-González S, De la Vieja A, García-Jiménez C. Glucose-induced β-catenin acetylation enhances Wnt signaling in cancer. Mol Cell 2013; 49: 474-486 [PMID: 23273980 DOI: 10.1016/j.molcel.2012.11.022]
- 121 García-Jiménez C, García-Martínez JM, Chocarro-Calvo A, De la Vieja A. A new link between diabetes and cancer: enhanced WNT/β-catenin signaling by high glucose. J Mol Endocrinol 2014; 52: R51-R66 [PMID: 24049067 DOI: 10.1530/JME-13-0152]
- 122 Ma X, Cui Z, Du Z, Lin H. Transforming growth factor- β signaling, a potential mechanism associated with diabetes mellitus and pancreatic cancer? J Cell Physiol 2020; 235: 5882-5892 [PMID: 32017070 DOI: 10.1002/jcp.29605]
- 123 Zhao L, Zou Y, Liu F. Transforming Growth Factor-Beta1 in Diabetic Kidney Disease. Front Cell Dev Biol 2020; 8: 187 [PMID: 32266267 DOI: 10.3389/fcell.2020.00187]
- 124 Achyut BR, Yang L. Transforming growth factor- β in the gastrointestinal and hepatic tumor microenvironment. Gastroenterology 2011; 141: 1167-1178 [PMID: 21839702 DOI: 10.1053/j.gastro.2011.07.048]
- 125 Ebert MP, Yu J, Miehlke S, Fei G, Lendeckel U, Ridwelski K, Stolte M, Bayerdörffer E, Malfertheiner P. Expression of transforming growth factor beta-1 in gastric cancer and in the gastric mucosa of first-degree relatives of patients with gastric cancer. Br J Cancer 2000; 82: 1795-1800 [PMID: 10839293 DOI: 10.1054/bjoc.1999.1107]
- 126 Naef M, Ishiwata T, Friess H, Büchler MW, Gold LI, Korc M. Differential localization of transforming growth factor-beta isoforms in human gastric mucosa and overexpression in gastric carcinoma. Int J Cancer 1997; 71: 131-137 [PMID: 9139831 DOI: 10.1002/(sici)1097-0215(19970410)71:2<131::aid-ijc1>3.0.co;2-1]
- 127 Ikushima H, Miyazono K. TGFbeta signalling: a complex web in cancer progression. Nat Rev Cancer 2010; 10: 415-424 [PMID: 20495575 DOI: 10.1038/nrc2853]
- 128 Hynes NE, MacDonald G. ErbB receptors and signaling pathways in cancer. Curr Opin Cell Biol 2009; 21: 177-184 [PMID: 19208461 DOI: 10.1016/j.ceb.2008.12.010]
- 129 Arienti C, Pignatta S, Tesei A. Epidermal Growth Factor Receptor Family and its Role in Gastric Cancer. Front Oncol 2019; 9: 1308 [PMID: 31850207 DOI: 10.3389/fonc.2019.01308]
- Thomas MC, Brownlee M, Susztak K, Sharma K, Jandeleit-Dahm KA, Zoungas S, Rossing P, 130



Groop PH, Cooper ME. Diabetic kidney disease. Nat Rev Dis Primers 2015; 1: 15018 [PMID: 27188921 DOI: 10.1038/nrdp.2015.18]

- 131 Rayego-Mateos S, Rodrigues-Diez R, Morgado-Pascual JL, Valentijn F, Valdivielso JM, Goldschmeding R, Ruiz-Ortega M. Role of Epidermal Growth Factor Receptor (EGFR) and Its Ligands in Kidney Inflammation and Damage. Mediators Inflamm 2018; 2018: 8739473 [PMID: 30670929 DOI: 10.1155/2018/8739473]
- 132 Dos Santos JM, Tewari S, Mendes RH. The Role of Oxidative Stress in the Development of Diabetes Mellitus and Its Complications. J Diabetes Res 2019; 2019: 4189813 [PMID: 31192263 DOI: 10.1155/2019/41898131
- Yuan T, Yang T, Chen H, Fu D, Hu Y, Wang J, Yuan Q, Yu H, Xu W, Xie X. New insights into 133 oxidative stress and inflammation during diabetes mellitus-accelerated atherosclerosis. Redox Biol 2019; 20: 247-260 [PMID: 30384259 DOI: 10.1016/j.redox.2018.09.025]
- 134 Wang Z, Chen JQ, Liu JL. COX-2 Inhibitors and Gastric Cancer. Gastroenterol Res Pract 2014; 2014: 132320 [PMID: 25371669 DOI: 10.1155/2014/132320]
- Sung JJ, Leung WK, Go MY, To KF, Cheng AS, Ng EK, Chan FK. Cyclooxygenase-2 expression 135 in Helicobacter pylori-associated premalignant and malignant gastric lesions. Am J Pathol 2000; 157: 729-735 [PMID: 10980112 DOI: 10.1016/S0002-9440(10)64586-5]
- 136 Oshima H, Oshima M. The role of PGE2-associated inflammatory responses in gastric cancer development. Semin Immunopathol 2013; 35: 139-150 [PMID: 23053397 DOI: 10.1007/s00281-012-0353-5
- Nasrallah R, Hassouneh R, Hébert RL. PGE2, Kidney Disease, and Cardiovascular Risk: Beyond 137 Hypertension and Diabetes. J Am Soc Nephrol 2016; 27: 666-676 [PMID: 26319242 DOI: 10.1681/ASN.2015050528
- Wang Y, Tao J, Yao Y. Prostaglandin E2 Activates NLRP3 Inflammasome in Endothelial Cells to 138 Promote Diabetic Retinopathy. Horm Metab Res 2018; 50: 704-710 [PMID: 30142638 DOI: 10.1055/a-0664-0699]
- 139 Shanmugam N, Kim YS, Lanting L, Natarajan R. Regulation of cyclooxygenase-2 expression in monocytes by ligation of the receptor for advanced glycation end products. J Biol Chem 2003; 278: 34834-34844 [PMID: 12837757 DOI: 10.1074/jbc.M302828200]
- 140 Kwon HJ, Park MI, Park SJ, Moon W, Kim SE, Kim JH, Choi YJ, Lee SK. Insulin Resistance Is Associated with Early Gastric Cancer: A Prospective Multicenter Case Control Study. Gut Liver 2019; 13: 154-160 [PMID: 30400721 DOI: 10.5009/gnl17556]
- 141 Yi HK, Hwang PH, Yang DH, Kang CW, Lee DY. Expression of the insulin-like growth factors (IGFs) and the IGF-binding proteins (IGFBPs) in human gastric cancer cells. Eur J Cancer 2001; 37: 2257-2263 [PMID: 11677116 DOI: 10.1016/S0959-8049(01)00269-6]
- Choi YJ. Insulin Resistance: A Hidden Risk Factor for Gastric Cancer? Gut Liver 2019; 13: 133-134 142 [PMID: 30893982 DOI: 10.5009/gnl19060]
- Tsujimoto T, Kajio H, Sugiyama T. Association between hyperinsulinemia and increased risk of 143 cancer death in nonobese and obese people: A population-based observational study. Int J Cancer 2017; 141: 102-111 [PMID: 28390156 DOI: 10.1002/ijc.30729]
- 144 Garay-Sevilla ME, Gomez-Oieda A, González I, Luévano-Contreras C, Rojas A, Contribution of RAGE axis activation to the association between metabolic syndrome and cancer. Mol Cell Biochem 2021; 476: 1555-1573 [PMID: 33398664 DOI: 10.1007/s11010-020-04022-z]
- 145 Pollak M. Insulin and insulin-like growth factor signalling in neoplasia. Nat Rev Cancer 2008; 8: 915-928 [PMID: 19029956 DOI: 10.1038/nrc2536]
- Saliminejad K, Khorram Khorshid HR, Soleymani Fard S, Ghaffari SH. An overview of 146 microRNAs: Biology, functions, therapeutics, and analysis methods. J Cell Physiol 2019; 234: 5451-5465 [PMID: 30471116 DOI: 10.1002/jcp.27486]
- 147 Ali Syeda Z, Langden SSS, Munkhzul C, Lee M, Song SJ. Regulatory Mechanism of MicroRNA Expression in Cancer. Int J Mol Sci 2020; 21 [PMID: 32138313 DOI: 10.3390/ijms21051723]
- 148 Rojas A, Araya P, Gonzalez I, Morales E. Gastric Tumor Microenvironment. Adv Exp Med Biol 2020; 1226: 23-35 [PMID: 32030673 DOI: 10.1007/978-3-030-36214-0 2]
- 149 Link A, Schirrmeister W, Langner C, Varbanova M, Bornschein J, Wex T, Malfertheiner P. Differential expression of microRNAs in preneoplastic gastric mucosa. Sci Rep 2015; 5: 8270 [PMID: 25652892 DOI: 10.1038/srep08270]
- 150 Kim M, Zhang X. The Profiling and Role of miRNAs in Diabetes Mellitus. J Diabetes Clin Res 2019; 1: 5-23 [PMID: 32432227 DOI: 10.33696/diabetes.1.003]
- 151 Chen B, Li J, Chi D, Sahnoune I, Calin S, Girnita L, Calin GA. Non-Coding RNAs in IGF-1R Signaling Regulation: The Underlying Pathophysiological Link between Diabetes and Cancer. Cells 2019; 8 [PMID: 31847392 DOI: 10.3390/cells8121638]
- Liang M, Shi B, Liu J, He L, Yi G, Zhou L, Yu G, Zhou X. Downregulation of miR203 induces 152 overexpression of PIK3CA and predicts poor prognosis of gastric cancer patients. Drug Des Devel Ther 2015; 9: 3607-3616 [PMID: 26213461 DOI: 10.2147/DDDT.S85525]
- 153 Feng L, Cheng K, Zang R, Wang Q, Wang J. miR-497-5p inhibits gastric cancer cell proliferation and growth through targeting PDK3. Biosci Rep 2019; 39 [PMID: 31409724 DOI: 10.1042/BSR20190654
- Wang BG, Jiang LY, Xu Q. A comprehensive evaluation for polymorphisms in let-7 family in 154 cancer risk and prognosis: a system review and meta-analysis. Biosci Rep 2018; 38 [PMID: 29717029 DOI: 10.1042/BSR20180273]



- 155 Zhu P, Liu J, Lu M, Wu G, Lin X, Cai L, Zhang X. Influence and mechanism of miR-99a suppressing development of colorectal cancer (CRC) with diabetes mellitus (DM). Onco Targets Ther 2019; 12: 10311-10321 [PMID: 31819515 DOI: 10.2147/OTT.S190998]
- 156 Coucha M, Mohamed IN, Elshaer SL, Mbata O, Bartasis ML, El-Remessy AB. High fat diet dysregulates microRNA-17-5p and triggers retinal inflammation: Role of endoplasmic-reticulumstress. World J Diabetes 2017; 8: 56-65 [PMID: 28265343 DOI: 10.4239/wjd.v8.i2.56]
- 157 Gao X, Qiao X, Xing X, Huang J, Qian J, Wang Y, Zhang Y, Zhang X, Li M, Cui J, Yang Y. Matrix Stiffness-Upregulated MicroRNA-17-5p Attenuates the Intervention Effects of Metformin on HCC Invasion and Metastasis by Targeting the PTEN/PI3K/Akt Pathway. Front Oncol 2020; 10: 1563 [PMID: 32974191 DOI: 10.3389/fonc.2020.01563]
- 158 Zhang L, Yu D. Exosomes in cancer development, metastasis, and immunity. Biochim Biophys Acta Rev Cancer 2019; 1871: 455-468 [PMID: 31047959 DOI: 10.1016/j.bbcan.2019.04.004]
- 159 Kalluri R, LeBleu VS. The biology, function, and biomedical applications of exosomes. Science 2020; 367 [PMID: 32029601 DOI: 10.1126/science.aau6977]
- Wu SF, Noren Hooten N, Freeman DW, Mode NA, Zonderman AB, Evans MK. Extracellular 160 vesicles in diabetes mellitus induce alterations in endothelial cell morphology and migration. J Transl Med 2020; 18: 230 [PMID: 32517700 DOI: 10.1186/s12967-020-02398-6]
- 161 Freeman DW, Noren Hooten N, Eitan E, Green J, Mode NA, Bodogai M, Zhang Y, Lehrmann E, Zonderman AB, Biragyn A, Egan J, Becker KG, Mattson MP, Ejiogu N, Evans MK. Altered Extracellular Vesicle Concentration, Cargo, and Function in Diabetes. Diabetes 2018; 67: 2377-2388 [PMID: 29720498 DOI: 10.2337/db17-1308]
- 162 Huang T, Song C, Zheng L, Xia L, Li Y, Zhou Y. The roles of extracellular vesicles in gastric cancer development, microenvironment, anti-cancer drug resistance, and therapy. Mol Cancer 2019; 18: 62 [PMID: 30925929 DOI: 10.1186/s12943-019-0967-5]
- Hsieh HL, Tsai MM. Tumor progression-dependent angiogenesis in gastric cancer and its potential 163 application. World J Gastrointest Oncol 2019; 11: 686-704 [PMID: 31558974 DOI: 10.4251/wigo.v11.i9.686]
- 164 Chen KB, Chen J, Jin XL, Huang Y, Su QM, Chen L. Exosome-mediated peritoneal dissemination in gastric cancer and its clinical applications. Biomed Rep 2018; 8: 503-509 [PMID: 29774141 DOI: 10.3892/br.2018.1088
- Deng G, Qu J, Zhang Y, Che X, Cheng Y, Fan Y, Zhang S, Na D, Liu Y, Qu X. Gastric cancer-165 derived exosomes promote peritoneal metastasis by destroying the mesothelial barrier. FEBS Lett 2017; 591: 2167-2179 [PMID: 28643334 DOI: 10.1002/1873-3468.12722]
- 166 WARBURG O. On the origin of cancer cells. Science 1956; 123: 309-314 [PMID: 13298683 DOI: 10.1126/science.123.3191.3091
- Burns JS, Manda G. Metabolic Pathways of the Warburg Effect in Health and Disease: Perspectives 167 of Choice, Chain or Chance. Int J Mol Sci 2017; 18 [PMID: 29257069 DOI: 10.3390/ijms18122755]
- Liu Z, Jia X, Duan Y, Xiao H, Sundqvist KG, Permert J, Wang F. Excess glucose induces hypoxia-168 inducible factor-1a in pancreatic cancer cells and stimulates glucose metabolism and cell migration. Cancer Biol Ther 2013; 14: 428-435 [PMID: 23377827 DOI: 10.4161/cbt.23786]
- Huang YL, Lin YC, Lin CC, Chen WM, Chen BPC, Lee H. High Glucose Induces VEGF-C 169 Expression via the LPA1/3-Akt-ROS-LEDGF Signaling Axis in Human Prostate Cancer PC-3 Cells. Cell Physiol Biochem 2018; 50: 597-611 [PMID: 30317243 DOI: 10.1159/000494177]
- Yang W, Lu Z. Regulation and function of pyruvate kinase M2 in cancer. Cancer Lett 2013; 339: 153-158 [PMID: 23791887 DOI: 10.1016/j.canlet.2013.06.008]
- Ramteke P, Deb A, Shepal V, Bhat MK. Hyperglycemia Associated Metabolic and Molecular 171 Alterations in Cancer Risk, Progression, Treatment, and Mortality. Cancers (Basel) 2019; 11 [PMID: 31546918 DOI: 10.3390/cancers11091402]
- Lei Y, Zhou S, Hu O, Chen X, Gu J. Carbohydrate response element binding protein (ChREBP) 172 correlates with colon cancer progression and contributes to cell proliferation. Sci Rep 2020; 10: 4233 [PMID: 32144313 DOI: 10.1038/s41598-020-60903-9]
- Kuhajda FP. Fatty acid synthase and cancer: new application of an old pathway. Cancer Res 2006; 173 66: 5977-5980 [PMID: 16778164 DOI: 10.1158/0008-5472.CAN-05-4673]
- Vordermark D, Kraft P, Katzer A, Bölling T, Willner J, Flentje M. Glucose requirement for 174 hypoxic accumulation of hypoxia-inducible factor-1alpha (HIF-1alpha). Cancer Lett 2005; 230: 122-133 [PMID: 16253768 DOI: 10.1016/j.canlet.2004.12.040]
- 175 Kitajima Y, Miyazaki K. The Critical Impact of HIF-1a on Gastric Cancer Biology. Cancers (Basel) 2013; 5: 15-26 [PMID: 24216696 DOI: 10.3390/cancers5010015]

C D WĴ

Oncology

World Journal of **Gastrointestinal**

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2013-2028

DOI: 10.4251/wjgo.v13.i12.2013

ISSN 1948-5204 (online)

REVIEW

Macrophages play a role in inflammatory transformation of colorectal cancer

Lu Lu, Yu-Jing Liu, Pei-Qiu Cheng, Dan Hu, Han-Chen Xu, Guang Ji

ORCID number: Lu Lu 0000-0003-2777-5992; Yu-Jing Liu 0000-0002-9879-4413; Pei-Qiu Cheng 0000-0002-4244-2224; Dan Hu 0000-0002-5331-3276; Han-Chen Xu 0000-0003-2335-5421; Guang Ji 0000-0003-0842-3676.

Author contributions: Lu L, Liu YJ, Cheng PQ, and Hu D wrote the manuscript; Ji G and Xu HC are cocorresponding authors, and they contributed to editing the manuscript; all authors wrote, read, and approved the final manuscript.

Conflict-of-interest statement:

There is no conflict of interest associated with any of the senior author or other coauthors who contributed their efforts in this manuscript.

Supported by The National Nature Science Foundation of China, No. 81874206; Shanghai Rising-Star Program, No. 20QA1409300; and the Program for Young Eastern Scholar at Shanghai Institutions of Higher Learning, No. QD2019034.

Country/Territory of origin: China

Specialty type: Oncology

Provenance and peer review: Unsolicited article; Externally peer reviewed.

Peer-review report's scientific

Lu Lu, Yu-Jing Liu, Pei-Qiu Cheng, Han-Chen Xu, Guang Ji, Institute of Digestive Diseases, Longhua Hospital, Shanghai University of Traditional Chinese Medicine, Shanghai 200032, China

Dan Hu, Han-Chen Xu, Shanghai Pudong New Area Hospital of Traditional Chinese Medicine, Shanghai 200120, China

Corresponding author: Guang Ji, MD, PhD, Chief Doctor, Professor, Institute of Digestive Diseases, Longhua Hospital, Shanghai University of Traditional Chinese Medicine, No. 725 South Wanping Road, Shanghai 200032, China. jiliver@vip.sina.com

Abstract

Colorectal cancer (CRC) is one of the most common and fatal cancers worldwide, and it is also a typical inflammatory cancer. The function of macrophages is very important in the tissue immune microenvironment during inflammatory and carcinogenic transformation. Here, we evaluated the function and mechanism of macrophages in intestinal physiology and in different pathological stages. Furthermore, the role of macrophages in the immune microenvironment of CRC and the influence of the intestinal population and hypoxic environment on macrophage function are summarized. In addition, in the era of tumor immunotherapy, CRC currently has a limited response rate to immune checkpoint inhibitors, and we summarize potential therapeutic strategies for targeting tumorassociated macrophages.

Key Words: Colorectal cancer; Macrophages; Inflammatory transformation; Tumor microenvironment

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: In this review, we provide a comprehensive review of the research progress of macrophages in intestinal inflammation and colorectal cancer. It is of great significance to discuss the intestinal macrophages under steady-state and inflammatory conditions and tumor-associated macrophages in the immune microenvironment. With the research on macrophages in intestinal inflammation and tumor diseases, targeted macrophage therapy will benefit patients with intestinal inflammation or colorectal



quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: May 17, 2021 Peer-review started: May 17, 2021 First decision: July 14, 2021 Revised: July 21, 2021 Accepted: August 25, 2021 Article in press: August 25, 2021 Published online: December 15, 2021

P-Reviewer: Lieto E S-Editor: Wang LL L-Editor: Wang TQ P-Editor: Yuan YY



cancer.

Citation: Lu L, Liu YJ, Cheng PQ, Hu D, Xu HC, Ji G. Macrophages play a role in inflammatory transformation of colorectal cancer. World J Gastrointest Oncol 2021; 13(12): 2013-2028

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2013.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2013

INTRODUCTION

Colorectal cancer (CRC) is a common malignant tumor. Changes in bowel habits and stool characteristics, abdominal discomfort, thigh lumps, intestinal obstruction, anemia, and other systemic symptoms can be related to disease progression, but no obvious clinical manifestations are present in the early stage[1]. According to the latest statistics by the American Cancer Society, the incidence and mortality of CRC rank third among all malignant tumors[2]. The latest statistics on cancer from China show that CRC has become the third most common cancer in terms of incidence, with the fifth highest mortality rate[1]. Most patients are already in a moderate or advanced stage when they are diagnosed, which imposes a great burden on their family and society. Therefore, early detection and screening, correct diagnosis of CRC, and early intervention and treatment to slow down the progression of the disease are particularly important. In recent years, an increasing number of studies have shown that macrophages play an important role in the occurrence and development of CRC. This article will review the research status of intestinal macrophages, the role and regulatory factors of tumor-associated macrophages (TAMs), and the research progress related to targeted TAM therapy to provide new ideas for the clinical diagnosis and treatment of inflammatory bowel disease and CRC.

INTESTINAL MACROPHAGES

Macrophages play an important role in intestinal inflammatory immunity, injury repair, epithelial-mesenchymal transition, and tumor development. Traditionally, macrophages differentiate from monocytes and play an immunomodulatory role^[3]. Further study found that there are two main sources of intestinal macrophages: Gutresident macrophages (gMacs) and monocytes (monocyte-derived macrophages). Resident tissue macrophages (RTMs) are derived from embryonic precursors, which accumulate in tissues before birth and are maintained by renewal in adulthood[4]. In contrast to the self-renewal and self-maintenance of Kupffer cells and microglia, whether the gMac population is maintained by contributions from mononuclear macrophages is not clear. Although traditional studies have concluded that embryonic macrophages in the intestinal tract are replaced by bone marrow-derived Ly6Chi monocytes in a microorganism-dependent manner, an experiment evaluating intestinal macrophage heterogeneity determined that the self-sustaining population of macrophages is produced by embryonic precursors and adult bone marrow-derived monocytes, which persist throughout adulthood, and that these cells settle in specific niches, including the vascular system, submucosa, muscular plexus, sites of Pan's cells, and Peyer's patches. Single-cell analysis has shown that gMacs have a unique transcriptional profile, which supports the vascular structure and permeability in the lamina propria (LP) and also regulates neuronal function and intestinal peristalsis in the LP and muscularis externa^[5].

Origin and differentiation of intestinal macrophages

The gene expression profiles of macrophages in tissues and sites vary[6]. Although no study has shown that the origin changes the macrophage life span or biological functions[7], recent studies have shown that macrophage origin influences the gene expression profile[8,9]. After treatment with chlorophosphate liposomes, mice with a monocyte-derived Kupffer cell population reacted more acutely to excessive paracetamol than mice with an intact embryonic Kupffer cell population. However, this functional difference might also be attributed to tissues because the difference



disappeared after monocyte-derived Kupffer cells were placed in the liver for 60 d without an overdose of acetaminophen[10]. Epigenetic analyses show that macrophages of different cell origins are relatively similar and are mainly influenced by living tissues. There are some epigenetic differences among macrophages derived from different precursors, which may be related to the changes in the local tissue environment caused by whole body irradiation[8]. To date, it is necessary to explore the differences in epigenetics and function, not only origin, among different macrophages in detail.

Surface markers of intestinal macrophages

The unique transcriptome of tissue macrophages endows different functions to these cells and allows them to play specific biological functions in the microenvironment[11-13].

Some of the main challenges in this field are to identify intestinal macrophages and their subgroup markers and determine how to regulate these cells to meet the biological functional requirements of their living environment. In mice, F4/80 is the best and most commonly used marker to identify macrophages[14]. However, conventional dendritic cells (cDCs) and eosinophils can also express F4/80[15,16]. Intestinal macrophages highly express CD11C and MHCII, which can identify cDCs and are related to the polarization of M1 macrophages[17]. However, intestinal macrophages also express CD206 and CD163 but do not express arginine[18]. Therefore, intestinal macrophages are not suitable for M1 and M2 typing. The identification of intestinal macrophages requires a multiparameter method.

gMacs and cDCs can be distinguished by CX3CR1 and CD64 in combination with CD11C and MHCII. Compared with cDCs, gMacs highly express the chemokine receptor CX3CR1[18,19], which is mainly located in the LP of the intestine, connective tissue under the skin, intestinal wall, submucosa, and muscle[19-22]. CX3CR1 is a key regulator of macrophage function in the inflammatory state [23,24], while the CX3CR1⁺ myeloid cell-Treg axis plays a central role in maintaining intestinal homeostasis^[25]. CX3CR1⁺ macrophages resident in the mucosa can recruit and activate antigenpresenting cells displaying epitopes to CD4⁺T cells and B cells at an invasion site[26], effectively inhibiting the production of IL-17 by CD4⁺ T cells by promoting Treg activity dependent on IFN- β [27]. Although there are reports that IFN- β can inhibit the production of IL-17 in mouse and human CD4⁺ T cells, the mechanism is not clear[28, 29]. The expression of CD11c differs among gMacs at different sites; CD11c⁺ gMacs are enriched in the LP, while CD11^{c-/lo}CX3CR1^{hi} gMacs are enriched in the muscle[29,30]. LP gMacs actively participate in host defense, maintain the integrity of the barrier, have high phagocytic activity, promote the constitutive secretion of interleukin-10 (IL-10), maintain FoxP3⁺ T cells, and protect mucous membranes[31]. The development and survival of CD64⁺ mononuclear phagocytes are highly dependent on colonystimulating factor 1 (CSF1), while CD64-CD11c⁺ MHC II⁺ mononuclear phagocytes, which are highly dependent on the CDC-specific growth factor FLT3[32], migrate to the mesenteric lymph nodes (MLNs) and participate in the initiation of T cell responses in a CCR7-dependent manner [33,34]. An experiment evaluating Tim-4- and CD4-labeled gMacs also provided evidence for the development and heterogeneity of intestinal macrophages[35]. However, the function of these cells is not clear. Tracking CD64⁺ gMacs with YFP in hybrid offspring from Cx3cr1^{CreERT2} mice and Rosa26-LSL-YFP mice successfully identified self-sustaining gMac subsets[5].

Intestinal macrophages under steady-state and inflammatory conditions

Intestinal macrophages are the main participants in establishing and maintaining intestinal homeostasis. gMacs produce a variety of cytokines and mediators (PGE2, BMP2, WNT ligand, etc.) to maintain the proliferation of intestinal epithelial cells and the physiology of intestinal neurons and endothelial cells[36]. gMacs also promote the expansion of antigen-specific CD4⁺ CD25⁺ regulatory T cells by producing IL-10, prevent inflammatory reactions in the microbial environment, and support intestinal tolerance[37]. Intrinsic receptors (including LPS (CD14), fca (CD89), fcy (CD64, CD32, and CD16), Cr3 (CD11b/CD18), and Cr4 (CD11c/CD18)) are not expressed in gMacs [38]. gMacs also lack trigger receptors expressed on myeloid cells 1 (TREM-1)[39], which is a cell-surface molecule expressed on neutrophils and monocytes/macrophages in the peripheral blood. The activation reaction mediated by TREM-1 can increase the expression of proinflammatory mediators (such as TNF, IL-1β, and IL-6) and upregulate the levels of cell-surface molecules (CD40, CD86, and CD32)[40], leading to oxidative stress. Therefore, when intestinal macrophages play an effective scavenging role, they usually do not induce inflammation or damage intestinal homeostasis.



Monocytes and macrophages can induce cytotoxicity and proinflammatory mediators, eliminate apoptotic and damaged cells, and promote tumor progression when tissue is damaged[41,42]. The CCL2-CCR2 axis plays an important role in the migration of monocytes from the bone marrow to the peripheral blood. CCR2-deficient and CCR2-positive mice have been widely used in the study of monocytes and monocyte-derived cells in the development of tissue damage and elimination of pathogens[43,44]. During inflammation, the transportation of CCR2^{-/-} monocytes to the small intestine is obviously decreased, but interestingly, the recruitment of circulating monocytes to other tissues, such as the liver and spleen, is not affected by CCR2 deficiency^[45]. Silencing CCR2 also significantly reduces repaglinide tolerance, which may be related to the stability of β -catenin regulated by AKT/GSK3[46]. Recent studies have shown that the exogenous antiaging factor Klotho can inhibit the progression of CRC by inhibiting the expression of CCL2[47]. The chemokines CCL2 and CXCL12 synergistically induce M2 macrophage polarization^[19]. Targeting CCL2/CCR2 without affecting transport to other tissues provides new hope for the treatment of CRC.

Under steady-state conditions, monocytes gradually differentiate into CX3CR1^{hi} macrophages that express genes related to the function of tolerant macrophages. According to the expression of Ly6C and MHCII, monocytes and macrophages in the small intestine can be divided into three subgroups: Ly6C⁺ MHCII⁺, Ly6C⁺ MHCII⁺, and Ly6C[·]MHCII⁺. Based on the expression of CX3CR1, Ly6C[·]MHCII⁺ cells can be divided into CX3CR1^{int} and CX3CR1^{hi} cells, which can reflect the different stages of monocyte differentiation in the small intestine and colon[18,48,49]. Transcriptomic analysis also shows significant differences in gene expression among different stages. In addition to CX3CR1, the expression of CD64, CD11c, and CD206 increases with the development of Ly6C⁺ MHCII monocytes into small intestinal Ly6C⁻MHCII⁺ CX3CR1^{hi} macrophages. In contrast, monocytes immediately adapt to different expression patterns in a TREM-1-dependent manner after they enter the intestine in an inflammatory state. Inflammation fundamentally changes the kinetics and mode of monocyte differentiation in tissues^[45]. In contrast to intestinal homeostasis, inflammatory injury results in the accumulation of Ly6C⁺monocytes in large numbers. In a study, the expression of CD64 was high, while that of CX3CR1 was always low. On the third day of inflammation, CD64+ Ly6C-MHCII^{int} monocytes were divided into two subsets: MHCII^{hi}CX3CR1^{int} (seen in the inflamed colon)[50]) and MHCII. In the Ly6C⁻MHCII^{int} population, the CX3CR1 expression level was slightly higher than that in the Ly6C⁺ MHCII⁻ and Ly6C⁺MHCII^{int} populations but lower than that in Ly6C⁻MHCII^{hi} macrophages. These cells may represent the intermediate stage of monocyte differentiation in intestinal inflammation. However, there was no differential expression of genes with enhanced expression during homeostasis in the inflammatory intestinal environment. The levels of some inflammation-related genes gradually decreased, while that of CD169 increased significantly.

Studies have shown that macrophages play a key role in the pathogenesis of IBD and that these cells are present throughout the occurrence, progression, and recovery of intestinal inflammation in both humans[18,51] and mice[52,53]. Macrophages regulate the progression of colitis by producing proinflammatory factors, such as TNF, IL-1β, IL-23, IL-6, reactive oxygen species (ROS), and NO[50]. Intestinal macrophages release IL-1 β , IL-6, IL-23, and TGF- β and mediate the Th17 immune response, which plays an important role in the pathogenesis of IBD[54].

Intestinal flora and intestinal macrophages

The intestinal flora maintains the integrity of the epithelial barrier, shapes the mucosal system, and balances host defense through metabolites, its own components, and adhesion to host cells. The metabolites and bacterial components of intestinal microorganisms can send signals to immune cells and regulate intestinal immunity.

Dietary fiber can directly enter the cecum and colon, where it can be fermented and metabolized by microorganisms to produce short-chain fatty acids (SCFAs)[55]. SCFAs are the energy source of colon cells and regulate the physiological functions of intestinal epithelial cells and intestinal immune cells. SCFA-mediated histone deacetylase (HDAC) inhibition has anti-inflammatory effects. Butyrate inhibits the differentiation of dendritic cells and proinflammatory macrophage effectors from bone marrow stem cells in the LP through HDACs and reduces the immune system response to beneficial symbionts[56]. In addition, macrophages and dendritic cells develop anti-inflammatory properties under the stimulation of butyrate-mediated GPR109A signaling. Foxp3⁺ Tregs and CD4⁺T cells accumulate in the colon, activating immunosuppressive mechanisms and maintaining intestinal homeostasis[57].

CX3CR1^{hi} mononuclear phagocytes do not migrate during intestinal homeostasis [58]. Symbiotic bacteria and pathogenic bacteria can regulate the host immune response by activating TLR pathways in the intestine. TLR/MyD88 signal transduction limits the transport of CX3CR1^{hi}monocytic phagocytes from the LP to the MLNs^[59]. MyD88 deficiency and malnutrition lead to the migration of CX3CR1^{hi} mononuclear phagocytes to the MLNs, enhance the Th1 response to noninvasive pathogens in the MLNs, and increase IgA. TLR signaling mediated by the intestinal microbiota can regulate IL-10 production by intestinal macrophages[60]. The probiotic Clostridium butyricum promotes the accumulation of F4/80⁺CD11b⁺CD11c macrophages in the inflamed intestinal mucosa through the TLR2/MyD88 signaling pathway and the production of IL-10 and prevents colitis in mice^[52]. It has also been shown that the LPS/TLR4 pathway can trigger CCL2 and promote the accumulation of monocyte-like macrophages (MLMs)[61], which can produce IL-1 β , promote Th17 cell expansion, aggravate malnutrition and inflammation, and lead to tumor progression tumor formation[62].

TUMOR-ASSOCIATED MACROPHAGES

Peripheral mononuclear cells or RTMs infiltrate near tumor masses or into tumor tissue to form TAMs, which are the main inflammatory cells in the tumor matrix[63].

Recent studies have shown that TAMs originate from RTMs and newly recruited monocytes[64]. The evolution of cells was inferred by the RNA velocity of single cells, and it was confirmed that FCN1⁺ monocyte-like cells with tumor enrichment may be the precursors of TAMs and have a tumor-promoting transcriptional program. Transcriptional tracking of macrophages[65,66] indicated that FCN1⁺ monocyte-like cells produce C1QC⁺ TAMs and SPP1⁺ TAMs from different RTMs. C1QC⁺ TAMs may develop through IL1B⁺ RTMs and express genes involved in phagocytosis and antigen presentation. SPP1⁺ TAMs are linked to NLRP3⁺ RTMs, which are rich in angiogenesisregulating factors and have specific enrichment of rectal adenocarcinoma and metastatic liver cancer pathways, suggesting that SPP1⁺ TAMs can promote tumor development and metastasis[67]. However, these subsets do not conform to the M1 and M2 classification of TAMs[68].

Dual role of tumor-associated macrophages

The plasticity of macrophages determines the polarization state, and the function of macrophages varies with the macrophage phenotype and tumor type[69,70]. The phenotype of polarized TAMs depends on the stage of tumor progression: In the early stage of cancer, that is, the stage of tumor elimination with local chronic inflammation in the tumor, cytokines and chemokines induce TAM polarization to the M1 type[71], which can induce an inflammatory response and phagocytosis^[72]. Subsequently, M2 polarization occurs, and these cells secrete cytokines or chemokines and inhibit the antitumor immune response with changes in the tumor microenvironment (TME) and external stimuli as the tumor progresses[73].

In most human cancers, a large number of TAMs are significantly related to a poor disease prognosis, and basic research also shows that macrophages have a tumorpromoting function^[74,75]. A study of 120 CRC patients with liver metastasis showed that M1 macrophages were negatively correlated with tumor metastasis, while M2 macrophages were positively correlated with lymph node and liver metastasis and the degree of tumor differentiation. M2 macrophages and the M2/M1 ratio can be used as accurate predictors of liver metastasis in CRC patients^[76]. Based on an analysis of peripheral blood mononuclear cell samples from 360 CRC patients at the European Oncology Center, polarized circulating mononuclear cells can be used as biomarkers for CRC diagnosis and may be useful for follow-up and treatment evaluation[77].

M1 macrophages have high expression of major histocompatibility complex-II (MHC-II), exhibiting an effective antigen-presenting ability, and secrete proinflammatory factors and immunostimulatory cytokines, such as IL-12, IL-23, CXCL9, and CXCL10; thus, these cells function to kill bacteria and viruses, promote TH1 cell polarization and recruitment, and enhance the type 1 immune response [78]. M2 macrophages express a large number of anti-inflammatory cytokines (IL-10), immune mediators (TGF- β), prostaglandins, indoleamines, growth factors (VEGF), chemokines (CCL2, CCL17, and CCL22), and matrix metallopeptidases; thus, M2 macrophages participate in anti-inflammatory activity, tissue remodeling, wound healing, angiogenesis, and tumor development^[79]. Prior research and a meta-analysis showed that M1 macrophages prevent the occurrence and development of tumors, while M2



macrophages promote tumor cell proliferation and invasion, enhance angiogenesis, and accelerate tumor growth and metastasis[80,81]. However, CRC exhibits a paradox in the function of specific groups of immune cells. A study of 205 CRC patients showed that there were a large number of infiltrating CD163⁺ macrophages in the CRC patients with less lymphatic metastasis and a lower tumor grade, and the patients with more CD163⁺ macrophages exhibited a survival benefit. Unexpectedly, iNOS⁺ macrophages did not show any advantage[82]. In CRC stage III patients, high TAM levels are related to a better prognosis in patients who receive chemotherapy but not to the prognosis of patients who do not receive chemotherapy^[83].

The type 1 immune response can inhibit the progression of CRC[84]. However, the molecular mechanism regulating antitumor activity and promoting tumor inflammation in CRC is still unclear. NF- κ B is a key regulator of inflammation, and its activation and inhibition are controlled at a variety of regulatory levels, which can regulate the function of macrophages[85]. NF-kB p50 promotes the transcriptional program of M2 macrophages[86]. In a model of colitis-associated cancer (CAC) induced by AOM combined with DSS[87], the number of tumor lesions was significantly decreased in p50-/- mice, accompanied by increases in Th1/M1 inflammatory genes (Il12b, Il27, Ebi3, Cxcl9, Cxcl10, Nos2, and Ifng) and gene products (TNFα, IL12, and iNOS). An analysis of CRC stage II/III patients showed that nuclear accumulation of p50 in TAMs inhibited Th1 cell/M1 macrophage-dependent antitumor reactions, which was related to the expression of M2 macrophage-related genes (IL10, TGF- β , Ccl17, and Ccl22) and increases in tumor-promoting genes (TNF- α and IL23). The expression of NF-KB p50 plays important roles in the development of colitis and CAC, but negative regulators (including p50) that only block inflammatory reactions also cause adverse reactions[88]. Type 1 proinflammatory factors (IL-12 and CXCL-10) can offset adverse reactions and restore antitumor immunity, which still needs to be evaluated in large-scale clinical studies.

Tumor microenvironment and tumor-related macrophages

The TME is composed of cellular components and noncellular components. The cellular components include cancer cells, mesenchymal cells, infiltrating immune cells, and tumor-related fibroblasts, while the noncellular components are composed of cytokines and chemokines[89]. The TME can regulate the infiltration of macrophages and promote the development of CRC through the synergistic effects of cytokines and cells.

Chemotactic factors

The chemokine family includes important signaling molecules in the TME. CCL3, CCL4, CCL5, CCL8, and CCL22 are highly expressed in various tumors and participate in the action of TAMs^[90]. Recent studies have shown that CCL5 plays an important role in the development of CRC and that CD8⁺T cell infiltration is significantly increased in the primary colorectal tumor site of CCL5^{-/-}mice[91]. In vivo and in vitro experiments show that CCL5 secreted by macrophages mediates the formation of the p65/STAT3 complex, induces upregulation of PD-L1, inhibits the CD8⁺T cell response, and promotes immune escape and CRC development in cancer cells. Macrophage infiltration decreases significantly after anti-CCL5 and C-15 treatment [92]. Inhibition of the CCL5-CCR5 axis is expected to be a new cancer treatment strategy^[93].

CCL2 plays an important role in regulating the TME[94]. CCL22 secreted by tumor cells plays a pivotal role in immunosuppression in the tumor microenvironment by binding with Foxp3⁺Tregs, which highly express CCR4[95]. CCL22 was recently identified to have potential as a molecular biomarker for evaluating chemotherapy and tumor progression. Moreover, M2 macrophages transfer CCL22 to cancer cells and contribute to the development of 5-FU resistance and the epithelial-mesenchymal transition (EMT) program in CRC cells[96]. CCL22 and its receptor CCR4 can also promote the migration and invasion of gastric cancer cells[97], and M2 macrophagederived CCL22 can enhance the migration of tumor cells in patients with liver cancer [98].

Hypoxia

Hypoxia in the TME can lead to angiogenesis, EMT, TGF-β signal transduction, and increases in tumor cell migration and metastasis[99,100]. Tissue hypoxia affects TAMs in two ways: Hypoxia can induce tumor cells and the stroma to produce monocyterecruiting factors (CCL2, CCL5, CXCL12, CSF1, and VEGF). After monocytes are recruited into hypoxic areas, the expression of cytokine receptors is downregulated, and TAMs are trapped in the hypoxic microenvironment[101]. Furthermore, macro-



phages capture oxygen through hypoxia inducible factors (HIFs), and decreased expression of ARG1 and immunosuppressive activity occur in vitro in the absence of HIF1α[102]. HIF2α deficiency weakens macrophage infiltration and cytokine production[103]. TAMs secrete "vascular factors" (VEGF, Sema3A, MMP2, and MMP9) [104]. TAMs in Nrp^{L/L} mice fail to enter the hypoxic tumor area, resulting in decreased angiogenesis and a weakened immunosuppressive ability, which leads to decreased vascular branches and a Th1 cell/CTL-mediated antitumor immune response[105]. Th1 cells release TAMs recruited by IFN-y and other cytokines, initiating feed-forward circulation and enhancing antitumor immunity[106]. Reduced angiogenesis and tumor perfusion also trigger feed-forward circulation, resulting in hypoxia and recruitment of more TAMs[107]. However, when Nrp1 is absent, these TAMs will not enter the hypoxic area and thus maintain the antitumor phenotype, which may explain observations made with clinical tumor biopsies: A higher number of TAMs are not necessarily related to a poor prognosis, and the clinical correlations between TAMs in different locations and the prognosis and survival of tumor patients are different[108].

Cluster analysis showed that the degree of M2 macrophage infiltration increased obviously under hypoxia but that the degree of M1 macrophage infiltration did not increase. The levels of CD163+ and CD206+ macrophages in the hypoxic subgroup were much higher than those in the normoxic subgroup. Hypoxia activates the RAS signaling pathway independently of KRAS mutation and activates the IL-6/ JAK/STAT3 signaling pathway by increasing the infiltration of M2 macrophages, thus regulating the progression of CRC[109]. The effect of lactic acid on macrophages under normoxic conditions is weak, but the combination of hypoxia and lactic acid can significantly promote the M2 polarization of macrophages through HIF-1, Hedgehog, and mTOR pathways[110].

Metabolism of tumor-related macrophages

Tumor metabolism plays important roles in promoting tumor growth and metastasis [111,112]. Amino acids and fatty acids provide substrates for tumor cells to produce metabolites and energy to meet the metabolic needs for proliferation and TME development. M1 macrophages mainly produce ATP through glycolysis, while M2 macrophages preferentially obtain energy through the oxidative TCA cycle coupled with oxidative phosphorylation. Compared with M1 macrophages, M2 macrophages have opposing arginine metabolism[113]. Increasing evidence shows that the lipid metabolism of immune cells, especially that of TAMs, plays important roles in the occurrence and development of tumors. In recent years, research on the process of lipid metabolism in TAMs has focused on the regulatory mechanisms of lipid metabolism-related enzymes.

In vitro and in vivo mouse experiments have shown that [114] the level of the lipolytic coactivator ABHD5 in CRC-associated macrophages is increased significantly, while that of monoacylcerolipase (MGLL) is decreased. ABHD5 can promote the growth of CRC by inhibiting the production of spermidine, which depends on SRM in TAMs. MGLL deficiency may lead to an increase in fatty acid glycerides[115]. The upregulation of ABHD5 may lead to a decrease in triglycerides and an increase in diglyceride[116]. A transplanted tumor model including mouse myeloid cells overexpressing ABHD5 showed that TAM ABHD5 could inhibit peritoneal and pulmonary metastasis of tumor cells (MC-38 and B-16 cells) and that macrophage ABHD5 regulated the migration and metastasis of tumor cells through the IL-1 β /NF- κ B/MMP pathway. The MMTV-PyMT mouse model of spontaneous breast cancer also verified that macrophage ABHD5 could inhibit lung metastasis of spontaneous breast cancer [114].

Phospholipid metabolism can affect the TME by regulating tumor-related immune cells[117-119]. The lysophosphatidic acid acyltransferase β -AGPT4 is highly expressed in CRC patients, and the survival rate of CRC patients is reduced with high expression of AGPT4. Agpat4 knockdown can increase the expression of the proinflammatory factors IL-1 β , IL-6, and TNF- α by increasing the LPA content, inducing polarization of M1 macrophages and enhancing antitumor effects[124]. An animal experiment performed with mice treated with ethoxymethane and sodium dextran sulfate showed that[120] Lipin-1, a phospholipid acid phosphatase, could promote the infiltration of $F4/80^+$ macrophages by participating in the production of CXCL1/2 (the infiltration of other immune cells, such as T cells, was not changed), upregulating the level of Nos2/iNOS and promoting dysplasia-cancer metastasis in colorectal tumors.

Targeting tumor-related macrophages

A large number of studies have proven the role of the CSF1-CSF1R axis in TAM recruitment, and inhibition of CSF1-CSF1R signaling leads to apoptosis and death in



Lu L et al. Macrophages in colorectitis and colorectal cancer



Figure 1 Etiology of macrophages in inflammatory bowel disease and colorectal cancer. gMacs: Gut-resident macrophages; TNF-a: Tumor necrosis factor; IL-13: Interleukin-1 beta; IL-6: Interleukin-6; IL-23: Interleukin-23; IL-10: Interleukin-10; TGF-6: Transforming growth factor-beta; CCL17: C-C motif chemokine 22; CCL22: C-C motif chemokine 22.

most TAMs[121]. CSF1-CSF1R blockers can improve the efficacy of various immunotherapeutic methods, including administration of CD40 agonists or PD1 or cytotoxic T lymphocyte antigen 4 (CTLA4) antagonists and adoptive T cell therapy[122-124]. Anti-CSF1R treatment can specifically deplete C1QC⁺ TAMs but cannot deplete the entire SPP1⁺ macrophage population, which can promote tumor growth. This finding may explain why anti-CSFR1 antibodies are not effective as monotherapies in tumor patients[67]. CSF1R inhibition combined with radiotherapy or chemotherapy can improve the T cell response and enhance the therapeutic effect in a large number of animal models[125-127].

CXCR4-CXCL12 is an important signal transduction axis involved in TAM recruitment, which can promote tumor invasion and regeneration [128]. Monocytes secrete the chemokine CXCL12 and express the receptors CXCR4 and CXCR7, which lead to autocrine/paracrine loops; promote the differentiation of different types of macrophages; enhance the expression of CD4, CD14, and CD163; and decrease the ability to stimulate antigen-specific T lymphocyte responses[129]. The CXCR4 antagonist peptide R (PEP R) can reduce the growth of HCT116 cells and improve the therapeutic effect of conventional chemotherapy (CT) or chemoradiotherapy (RT-CT). This effect depends on the decreases in cell growth and mesenchymal stem cell transformation induced by CT/RT-CT[130]. PEPR can also target CXCR4⁺ stromal cells and further decrease EMT and chemoresistance[131]. Combined administration of PEP R and the CXCL12 antagonist noxa-012 can improve the function of anti-PD1 antibodies in mice with CRC[132].

Macrophages in different functional states maintain cell activity through different metabolic pathways and metabolites[133]. Mammalian target of rapamycin (mTOR) signaling via mTORC1 and mTORC2 plays a central role in tracking nutrition, oxygen, and metabolites to guide the metabolic processes of macrophages[134]. Rapamycin (an mTORC1 inhibitor) can stimulate M1 macrophages and cause them to have an antitumor effect[135]. mTORC1 inhibitors can reduce immunosuppressive inflammation and tumor occurrence. Rad001 (a rapamycin derivative) ameliorates CRC induced by AOM/DSS in mice by limiting inflammation[136]. Signaling molecules (such as PI3Ky, Akt, and PTEN) upstream of mTOR also participate in the polarization and remodeling of TAMs, making the mTOR pathway a potential anticancer target [137]. The expression of a PI3Ky inhibitor (PTEN) or silencing of AKT1 can also promote the polarization of antitumor M1 macrophages[138].

Iron participates in the interaction between tumor cells and their environment^[139]. Unlike M1 macrophages, M2 macrophages express iron transporters and downregulate ferritin and heme oxygenase, all of which promote iron release[140]. In addition, conditioned medium from M2 macrophages can promote the proliferation of tumor cells, while iron chelation can inhibit the proliferation of tumor cells[141]. Recent studies have shown that iron chelation can reverse the iron-processing function of M2 macrophages, switching from iron release to chelation, and block the tumor-



promoting effect of M2 macrophages[142].

CONCLUSION

Macrophages play a crucial role in the occurrence and development of CRC. As the disease progresses, macrophages tend to differentiate into different subsets that play different biological functions. The dual functions of TAMs and the regulatory effects of the TME on TAMs are worthy of further study. Subsets of macrophages cannot be simply classified according to the traditional M1 and M2 phenotypes. Single-cell technology will benefit the phenotypic classification of macrophages and provide further insights into their function (Figure 1).

The complexity of tumors highlights the advantages of combined therapeutic approaches. The clinical application of immune checkpoint inhibitors such as PD-1 and PD-L1 monoclonal antibodies provide additional evidence for tumor immunotherapy, and studies have shown that targeting tumor-associated macrophages can significantly improve the efficacy of existing immunotherapy. Future research needs to have a clear understanding of drug mechanisms of action and drug resistance mechanisms to design effective combined therapies. In addition, more clinical data are needed to clarify the relationships between macrophage infiltration or phenotype and the prognosis of patients and to guide whether TAM antagonists can be used in patients to overcome immunotherapy resistance. Despite these challenges, the use of macrophages to improve the prognosis of cancer patients still has great potential.

REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin 2020; 70: 7-30 [PMID: 31912902 DOI: 10.3322/caac.21590]
- 2 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- 3 Bain CC, Mowat AM. Macrophages in intestinal homeostasis and inflammation. Immunol Rev 2014; 260: 102-117 [PMID: 24942685 DOI: 10.1111/imr.12192]
- 4 Yona S, Kim KW, Wolf Y, Mildner A, Varol D, Breker M, Strauss-Ayali D, Viukov S, Guilliams M, Misharin A, Hume DA, Perlman H, Malissen B, Zelzer E, Jung S. Fate mapping reveals origins and dynamics of monocytes and tissue macrophages under homeostasis. Immunity 2013; 38: 79-91 [PMID: 23273845 DOI: 10.1016/j.immuni.2012.12.001]
- 5 De Schepper S, Verheijden S, Aguilera-Lizarraga J, Viola MF, Boesmans W, Stakenborg N, Voytyuk I, Schmidt I, Boeckx B, Dierckx de Casterlé I, Baekelandt V, Gonzalez Dominguez E, Mack M, Depoortere I, De Strooper B, Sprangers B, Himmelreich U, Soenen S, Guilliams M, Vanden Berghe P, Jones E, Lambrechts D, Boeckxstaens G. Self-Maintaining Gut Macrophages Are Essential for Intestinal Homeostasis. Cell 2018; 175: 400-415.e13 [PMID: 30173915 DOI: 10.1016/j.cell.2018.07.048]
- Guilliams M, Scott CL. Does niche competition determine the origin of tissue-resident 6 macrophages? Nat Rev Immunol 2017; 17: 451-460 [PMID: 28461703 DOI: 10.1038/nri.2017.42]
- 7 Perdiguero EG, Geissmann F. The development and maintenance of resident macrophages. Nat Immunol 2016; 17: 2-8 [PMID: 26681456 DOI: 10.1038/ni.3341]
- 8 Scott CL, Zheng F, De Baetselier P, Martens L, Saeys Y, De Prijck S, Lippens S, Abels C, Schoonooghe S, Raes G, Devoogdt N, Lambrecht BN, Beschin A, Guilliams M. Bone marrowderived monocytes give rise to self-renewing and fully differentiated Kupffer cells. Nat Commun 2016; 7: 10321 [PMID: 26813785 DOI: 10.1038/ncomms10321]
- Beattie L, Sawtell A, Mann J, Frame TCM, Teal B, de Labastida Rivera F, Brown N, Walwyn-Brown K, Moore JWJ, MacDonald S, Lim EK, Dalton JE, Engwerda CR, MacDonald KP, Kaye PM. Bone marrow-derived and resident liver macrophages display unique transcriptomic signatures but similar biological functions. J Hepatol 2016; 65: 758-768 [PMID: 27262757 DOI: 10.1016/j.jhep.2016.05.037]
- 10 David BA, Rezende RM, Antunes MM, Santos MM, Freitas Lopes MA, Diniz AB, Sousa Pereira RV, Marchesi SC, Alvarenga DM, Nakagaki BN, Araújo AM, Dos Reis DS, Rocha RM, Marques PE, Lee WY, Deniset J, Liew PX, Rubino S, Cox L, Pinho V, Cunha TM, Fernandes GR, Oliveira AG, Teixeira MM, Kubes P, Menezes GB. Combination of Mass Cytometry and Imaging Analysis Reveals Origin, Location, and Functional Repopulation of Liver Myeloid Cells in Mice. Gastroenterology 2016; 151: 1176-1191 [PMID: 27569723 DOI: 10.1053/j.gastro.2016.08.024]
- Hashimoto D, Miller J, Merad M. Dendritic cell and macrophage heterogeneity in vivo. Immunity 11 2011; 35: 323-335 [PMID: 21943488 DOI: 10.1016/j.immuni.2011.09.007]
- 12 Gautier EL, Shay T, Miller J, Greter M, Jakubzick C, Ivanov S, Helft J, Chow A, Elpek KG,



Gordonov S, Mazloom AR, Ma'ayan A, Chua WJ, Hansen TH, Turley SJ, Merad M, Randolph GJ; Immunological Genome Consortium. Gene-expression profiles and transcriptional regulatory pathways that underlie the identity and diversity of mouse tissue macrophages. Nat Immunol 2012; 13: 1118-1128 [PMID: 23023392 DOI: 10.1038/ni.2419]

- 13 Okabe Y, Medzhitov R. Tissue-specific signals control reversible program of localization and functional polarization of macrophages. Cell 2014; 157: 832-844 [PMID: 24792964 DOI: 10.1016/j.cell.2014.04.016]
- 14 Hume DA, Perry VH, Gordon S. The mononuclear phagocyte system of the mouse defined by immunohistochemical localisation of antigen F4/80: macrophages associated with epithelia. Anat Rec 1984; 210: 503-512 [PMID: 6524692 DOI: 10.1002/ar.1092100311]
- 15 McGarry MP, Stewart CC. Murine eosinophil granulocytes bind the murine macrophage-monocyte specific monoclonal antibody F4/80. J Leukoc Biol 1991; 50: 471-478 [PMID: 1721083 DOI: 10.1002/jlb.50.5.471]
- 16 Pabst O, Bernhardt G. The puzzle of intestinal lamina propria dendritic cells and macrophages. Eur J Immunol 2010; 40: 2107-2111 [PMID: 20853495 DOI: 10.1002/eji.201040557]
- Mosser DM, Edwards JP. Exploring the full spectrum of macrophage activation. Nat Rev Immunol 17 2008; 8: 958-969 [PMID: 19029990 DOI: 10.1038/nri2448]
- 18 Bain CC, Scott CL, Uronen-Hansson H, Gudjonsson S, Jansson O, Grip O, Guilliams M, Malissen B, Agace WW, Mowat AM. Resident and pro-inflammatory macrophages in the colon represent alternative context-dependent fates of the same Ly6Chi monocyte precursors. Mucosal Immunol 2013; 6: 498-510 [PMID: 22990622 DOI: 10.1038/mi.2012.89]
- 19 Gabanyi I, Muller PA, Feighery L, Oliveira TY, Costa-Pinto FA, Mucida D. Neuro-immune Interactions Drive Tissue Programming in Intestinal Macrophages. Cell 2016; 164: 378-391 [PMID: 26777404 DOI: 10.1016/j.cell.2015.12.023]
- 20 Varol C, Vallon-Eberhard A, Elinav E, Aychek T, Shapira Y, Luche H, Fehling HJ, Hardt WD, Shakhar G, Jung S. Intestinal lamina propria dendritic cell subsets have different origin and functions. Immunity 2009; 31: 502-512 [PMID: 19733097 DOI: 10.1016/j.immuni.2009.06.025]
- 21 Bogunovic M, Ginhoux F, Helft J, Shang L, Hashimoto D, Greter M, Liu K, Jakubzick C, Ingersoll MA, Leboeuf M, Stanley ER, Nussenzweig M, Lira SA, Randolph GJ, Merad M. Origin of the lamina propria dendritic cell network. Immunity 2009; 31: 513-525 [PMID: 19733489 DOI: 10.1016/j.immuni.2009.08.010
- Mortha A, Chudnovskiy A, Hashimoto D, Bogunovic M, Spencer SP, Belkaid Y, Merad M. 22 Microbiota-dependent crosstalk between macrophages and ILC3 promotes intestinal homeostasis. Science 2014; 343: 1249288 [PMID: 24625929 DOI: 10.1126/science.1249288]
- Morimura S, Oka T, Sugaya M, Sato S. CX3CR1 deficiency attenuates imiquimod-induced 23 psoriasis-like skin inflammation with decreased M1 macrophages. J Dermatol Sci 2016; 82: 175-188 [PMID: 26976687 DOI: 10.1016/j.jdermsci.2016.03.004]
- 24 Donnelly DJ, Longbrake EE, Shawler TM, Kigerl KA, Lai W, Tovar CA, Ransohoff RM, Popovich PG. Deficient CX3CR1 signaling promotes recovery after mouse spinal cord injury by limiting the recruitment and activation of Ly6Clo/iNOS+ macrophages. J Neurosci 2011; 31: 9910-9922 [PMID: 21734283 DOI: 10.1523/JNEUROSCI.2114-11.2011]
- Martin B, Hirota K, Cua DJ, Stockinger B, Veldhoen M. Interleukin-17-producing gammadelta T cells selectively expand in response to pathogen products and environmental signals. *Immunity* 2009; 31: 321-330 [PMID: 19682928 DOI: 10.1016/j.immuni.2009.06.020]
- Koscsó B, Kurapati S, Rodrigues RR, Nedjic J, Gowda K, Shin C, Soni C, Ashraf AZ, 26 Purushothaman I, Palisoc M, Xu S, Sun H, Chodisetti SB, Lin E, Mack M, Kawasawa YI, He P, Rahman ZSM, Aifantis I, Shulzhenko N, Morgun A, Bogunovic M. Gut-resident CX3CR1hi macrophages induce tertiary lymphoid structures and IgA response in situ. Sci Immunol 2020; 5 [PMID: 32276965 DOI: 10.1126/sciimmunol.aax0062]
- 27 Gu T, Li Q, Egilmez NK. IFNβ-producing CX3CR1⁺ macrophages promote T-regulatory cell expansion and tumor growth in the APC^{min/+} / Bacteroides fragilis colon cancer model. Oncoimmunology 2019; 8: e1665975 [PMID: 31741765 DOI: 10.1080/2162402X.2019.1665975]
- 28 Zhang L, Yuan S, Cheng G, Guo B. Type I IFN promotes IL-10 production from T cells to suppress Th17 cells and Th17-associated autoimmune inflammation. PLoS One 2011; 6: e28432 [PMID: 22163016 DOI: 10.1371/journal.pone.0028432]
- Tao Y, Zhang X, Chopra M, Kim MJ, Buch KR, Kong D, Jin J, Tang Y, Zhu H, Jewells V, 29 Markovic-Plese S. The role of endogenous IFN- β in the regulation of Th17 responses in patients with relapsing-remitting multiple sclerosis. J Immunol 2014; 192: 5610-5617 [PMID: 24850724 DOI: 10.4049/jimmunol.1302580]
- Muller PA, Koscsó B, Rajani GM, Stevanovic K, Berres ML, Hashimoto D, Mortha A, Leboeuf M, Li XM, Mucida D, Stanley ER, Dahan S, Margolis KG, Gershon MD, Merad M, Bogunovic M. Crosstalk between muscularis macrophages and enteric neurons regulates gastrointestinal motility. Cell 2014; 158: 300-313 [PMID: 25036630 DOI: 10.1016/j.cell.2014.04.050]
- Zigmond E, Bernshtein B, Friedlander G, Walker CR, Yona S, Kim KW, Brenner O, Krauthgamer 31 R, Varol C, Müller W, Jung S. Macrophage-restricted interleukin-10 receptor deficiency, but not IL-10 deficiency, causes severe spontaneous colitis. Immunity 2014; 40: 720-733 [PMID: 24792913 DOI: 10.1016/j.immuni.2014.03.012]
- 32 Scott CL, Bain CC, Wright PB, Sichien D, Kotarsky K, Persson EK, Luda K, Guilliams M, Lambrecht BN, Agace WW, Milling SW, Mowat AM. CCR2(+)CD103(-) intestinal dendritic cells



develop from DC-committed precursors and induce interleukin-17 production by T cells. Mucosal Immunol 2015; 8: 327-339 [PMID: 25138666 DOI: 10.1038/mi.2014.70]

- 33 Cerovic V, Houston SA, Scott CL, Aumeunier A, Yrlid U, Mowat AM, Milling SW. Intestinal CD103(-) dendritic cells migrate in lymph and prime effector T cells. Mucosal Immunol 2013; 6: 104-113 [PMID: 22718260 DOI: 10.1038/mi.2012.53]
- 34 Coombes JL, Siddiqui KR, Arancibia-Cárcamo CV, Hall J, Sun CM, Belkaid Y, Powrie F. A functionally specialized population of mucosal CD103+ DCs induces Foxp3+ regulatory T cells via a TGF-beta and retinoic acid-dependent mechanism. J Exp Med 2007; 204: 1757-1764 [PMID: 17620361 DOI: 10.1084/jem.20070590]
- 35 Shaw TN, Houston SA, Wemyss K, Bridgeman HM, Barbera TA, Zangerle-Murray T, Strangward P, Ridley AJL, Wang P, Tamoutounour S, Allen JE, Konkel JE, Grainger JR. Tissue-resident macrophages in the intestine are long lived and defined by Tim-4 and CD4 expression. J Exp Med 2018; 215: 1507-1518 [PMID: 29789388 DOI: 10.1084/jem.20180019]
- 36 Bai Y, Jia X, Huang F, Zhang R, Dong L, Liu L, Zhang M. Structural elucidation, anti-inflammatory activity and intestinal barrier protection of longan pulp polysaccharide LPIIa. Carbohydr Polym 2020; 246: 116532 [PMID: 32747231 DOI: 10.1016/j.carbpol.2020.116532]
- 37 Mowat AM. To respond or not to respond - a personal perspective of intestinal tolerance. Nat Rev Immunol 2018; 18: 405-415 [PMID: 29491358 DOI: 10.1038/s41577-018-0002-x]
- 38 Smythies LE, Sellers M, Clements RH, Mosteller-Barnum M, Meng G, Benjamin WH, Orenstein JM, Smith PD. Human intestinal macrophages display profound inflammatory anergy despite avid phagocytic and bacteriocidal activity. J Clin Invest 2005; 115: 66-75 [PMID: 15630445 DOI: 10.1172/JCI19229
- 39 Schenk M, Bouchon A, Birrer S, Colonna M, Mueller C. Macrophages expressing triggering receptor expressed on myeloid cells-1 are underrepresented in the human intestine. J Immunol 2005; 174: 517-524 [PMID: 15611278 DOI: 10.4049/jimmunol.174.1.517]
- 40 Wang F, Liu S, Wu S, Zhu Q, Ou G, Liu C, Wang Y, Liao Y, Sun Z. Blocking TREM-1 signaling prolongs survival of mice with Pseudomonas aeruginosa induced sepsis. Cell Immunol 2012; 272: 251-258 [PMID: 22055202 DOI: 10.1016/j.cellimm.2011.10.006]
- Laskin DL, Sunil VR, Gardner CR, Laskin JD. Macrophages and tissue injury: agents of defense or 41 destruction? Annu Rev Pharmacol Toxicol 2011; 51: 267-288 [PMID: 20887196 DOI: 10.1146/annurev.pharmtox.010909.105812]
- 42 Gordon S, Plüddemann A, Martinez Estrada F. Macrophage heterogeneity in tissues: phenotypic diversity and functions. Immunol Rev 2014; 262: 36-55 [PMID: 25319326 DOI: 10.1111/imr.12223]
- 43 Xiong H, Carter RA, Leiner IM, Tang YW, Chen L, Kreiswirth BN, Pamer EG. Distinct Contributions of Neutrophils and CCR2+ Monocytes to Pulmonary Clearance of Different Klebsiella pneumoniae Strains. Infect Immun 2015; 83: 3418-3427 [PMID: 26056382 DOI: 10.1128/IAI.00678-15]
- 44 Coates BM, Staricha KL, Koch CM, Cheng Y, Shumaker DK, Budinger GRS, Perlman H, Misharin AV, Ridge KM. Inflammatory Monocytes Drive Influenza A Virus-Mediated Lung Injury in Juvenile Mice. J Immunol 2018; 200: 2391-2404 [PMID: 29445006 DOI: 10.4049/jimmunol.1701543]
- 45 Desalegn G, Pabst O. Inflammation triggers immediate rather than progressive changes in monocyte differentiation in the small intestine. Nat Commun 2019; 10: 3229 [PMID: 31324779 DOI: 10.1038/s41467-019-11148-2
- 46 Ou B, Cheng X, Xu Z, Chen C, Shen X, Zhao J, Lu A. A positive feedback loop of β-catenin/CCR2 axis promotes regorafenib resistance in colorectal cancer. Cell Death Dis 2019; 10: 643 [PMID: 31501414 DOI: 10.1038/s41419-019-1906-5]
- Liu Y, Pan J, Pan X, Wu L, Bian J, Lin Z, Xue M, Su T, Lai S, Chen F, Ge Q, Chen L, Ye S, Zhu Y, 47 Chen S, Wang L. Klotho-mediated targeting of CCL2 suppresses the induction of colorectal cancer progression by stromal cell senescent microenvironments. Mol Oncol 2019; 13: 2460-2475 [PMID: 31545552 DOI: 10.1002/1878-0261.12577]
- 48 Tamoutounour S, Henri S, Lelouard H, de Bovis B, de Haar C, van der Woude CJ, Woltman AM, Reyal Y, Bonnet D, Sichien D, Bain CC, Mowat AM, Reis e Sousa C, Poulin LF, Malissen B, Guilliams M. CD64 distinguishes macrophages from dendritic cells in the gut and reveals the Th1inducing role of mesenteric lymph node macrophages during colitis. Eur J Immunol 2012; 42: 3150-3166 [PMID: 22936024 DOI: 10.1002/eji.201242847]
- Schridde A, Bain CC, Mayer JU, Montgomery J, Pollet E, Denecke B, Milling SWF, Jenkins SJ, 49 Dalod M, Henri S, Malissen B, Pabst O, Mcl Mowat A. Tissue-specific differentiation of colonic macrophages requires TGF\beta receptor-mediated signaling. Mucosal Immunol 2017; 10: 1387-1399 [PMID: 28145440 DOI: 10.1038/mi.2016.142]
- Zigmond E, Varol C, Farache J, Elmaliah E, Satpathy AT, Friedlander G, Mack M, Shpigel N, 50 Boneca IG, Murphy KM, Shakhar G, Halpern Z, Jung S. Ly6C hi monocytes in the inflamed colon give rise to proinflammatory effector cells and migratory antigen-presenting cells. Immunity 2012; 37: 1076-1090 [PMID: 23219392 DOI: 10.1016/j.immuni.2012.08.026]
- 51 Sperber K, Ogata S, Sylvester C, Aisenberg J, Chen A, Mayer L, Itzkowitz S. A novel human macrophage-derived intestinal mucin secretagogue: implications for the pathogenesis of inflammatory bowel disease. Gastroenterology 1993; 104: 1302-1309 [PMID: 8482444 DOI: 10.1016/0016-5085(93)90338-d]
- 52 Hayashi A, Sato T, Kamada N, Mikami Y, Matsuoka K, Hisamatsu T, Hibi T, Roers A, Yagita H,



Ohteki T, Yoshimura A, Kanai T. A single strain of Clostridium butyricum induces intestinal IL-10producing macrophages to suppress acute experimental colitis in mice. Cell Host Microbe 2013; 13: 711-722 [PMID: 23768495 DOI: 10.1016/j.chom.2013.05.013]

- 53 Anderson P, Souza-Moreira L, Morell M, Caro M, O'Valle F, Gonzalez-Rey E, Delgado M. Adipose-derived mesenchymal stromal cells induce immunomodulatory macrophages which protect from experimental colitis and sepsis. Gut 2013; 62: 1131-1141 [PMID: 22637701 DOI: 10.1136/gutjnl-2012-302152]
- Maloy KJ, Kullberg MC. IL-23 and Th17 cytokines in intestinal homeostasis. Mucosal Immunol 54 2008; 1: 339-349 [PMID: 19079198 DOI: 10.1038/mi.2008.28]
- 55 Makki K, Deehan EC, Walter J, Bäckhed F. The Impact of Dietary Fiber on Gut Microbiota in Host Health and Disease. Cell Host Microbe 2018; 23: 705-715 [PMID: 29902436 DOI: 10.1016/j.chom.2018.05.012
- 56 Chang PV, Hao L, Offermanns S, Medzhitov R. The microbial metabolite butyrate regulates intestinal macrophage function via histone deacetylase inhibition. Proc Natl Acad Sci US A 2014; 111: 2247-2252 [PMID: 24390544 DOI: 10.1073/pnas.1322269111]
- Singh N, Gurav A, Sivaprakasam S, Brady E, Padia R, Shi H, Thangaraju M, Prasad PD, 57 Manicassamy S, Munn DH, Lee JR, Offermanns S, Ganapathy V. Activation of Gpr109a, receptor for niacin and the commensal metabolite butyrate, suppresses colonic inflammation and carcinogenesis. Immunity 2014; 40: 128-139 [PMID: 24412617 DOI: 10.1016/j.immuni.2013.12.007
- 58 Varol C, Zigmond E, Jung S. Securing the immune tightrope: mononuclear phagocytes in the intestinal lamina propria. Nat Rev Immunol 2010; 10: 415-426 [PMID: 20498668 DOI: 10.1038/nri2778]
- Diehl GE, Longman RS, Zhang JX, Breart B, Galan C, Cuesta A, Schwab SR, Littman DR. 59 Microbiota restricts trafficking of bacteria to mesenteric lymph nodes by CX(3)CR1(hi) cells. Nature 2013; 494: 116-120 [PMID: 23334413 DOI: 10.1038/nature11809]
- 60 Ueda Y, Kayama H, Jeon SG, Kusu T, Isaka Y, Rakugi H, Yamamoto M, Takeda K. Commensal microbiota induce LPS hyporesponsiveness in colonic macrophages via the production of IL-10. Int Immunol 2010; 22: 953-962 [PMID: 21051439 DOI: 10.1093/intimm/dxq449]
- 61 Yang Y, Li L, Xu C, Wang Y, Wang Z, Chen M, Jiang Z, Pan J, Yang C, Li X, Song K, Yan J, Xie W, Wu X, Chen Z, Yuan Y, Zheng S, Huang J, Qiu F. Cross-talk between the gut microbiota and monocyte-like macrophages mediates an inflammatory response to promote colitis-associated tumourigenesis. Gut 2020 [PMID: 33122176 DOI: 10.1136/gutjnl-2020-320777]
- Wong SH, Yu J. Gut microbiota in colorectal cancer: mechanisms of action and clinical 62 applications. Nat Rev Gastroenterol Hepatol 2019; 16: 690-704 [PMID: 31554963 DOI: 10.1038/s41575-019-0209-8]
- Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. Cell 2010; 63 141: 39-51 [PMID: 20371344 DOI: 10.1016/j.cell.2010.03.014]
- Gubin MM, Esaulova E, Ward JP, Malkova ON, Runci D, Wong P, Noguchi T, Arthur CD, Meng W, Alspach E, Medrano RFV, Fronick C, Fehlings M, Newell EW, Fulton RS, Sheehan KCF, Oh ST, Schreiber RD, Artyomov MN. High-Dimensional Analysis Delineates Myeloid and Lymphoid Compartment Remodeling during Successful Immune-Checkpoint Cancer Therapy. Cell 2018; 175: 1014-1030.e19 [PMID: 30343900 DOI: 10.1016/j.cell.2018.09.030]
- 65 Farrell JA, Wang Y, Riesenfeld SJ, Shekhar K, Regev A, Schier AF. Single-cell reconstruction of developmental trajectories during zebrafish embryogenesis. Science 2018; 360 [PMID: 29700225 DOI: 10.1126/science.aar3131]
- Wolf FA, Angerer P, Theis FJ. SCANPY: large-scale single-cell gene expression data analysis. 66 Genome Biol 2018; 19: 15 [PMID: 29409532 DOI: 10.1186/s13059-017-1382-0]
- 67 Zhang L, Li Z, Skrzypczynska KM, Fang Q, Zhang W, O'Brien SA, He Y, Wang L, Zhang Q, Kim A, Gao R, Orf J, Wang T, Sawant D, Kang J, Bhatt D, Lu D, Li CM, Rapaport AS, Perez K, Ye Y, Wang S, Hu X, Ren X, Ouyang W, Shen Z, Egen JG, Zhang Z, Yu X. Single-Cell Analyses Inform Mechanisms of Myeloid-Targeted Therapies in Colon Cancer. Cell 2020; 181: 442-459.e29 [PMID: 32302573 DOI: 10.1016/j.cell.2020.03.0481
- 68 Azizi E, Carr AJ, Plitas G, Cornish AE, Konopacki C, Prabhakaran S, Nainys J, Wu K, Kiseliovas V, Setty M, Choi K, Fromme RM, Dao P, McKenney PT, Wasti RC, Kadaveru K, Mazutis L, Rudensky AY, Pe'er D. Single-Cell Map of Diverse Immune Phenotypes in the Breast Tumor Microenvironment. Cell 2018; 174: 1293-1308.e36 [PMID: 29961579 DOI: 10.1016/j.cell.2018.05.060]
- Li S, Xu F, Zhang J, Wang L, Zheng Y, Wu X, Wang J, Huang Q, Lai M. Tumor-associated macrophages remodeling EMT and predicting survival in colorectal carcinoma. Oncoimmunology 2018; 7: e1380765 [PMID: 29416940 DOI: 10.1080/2162402X.2017.1380765]
- 70 Sørensen MD, Dahlrot RH, Boldt HB, Hansen S, Kristensen BW. Tumour-associated microglia/macrophages predict poor prognosis in high-grade gliomas and correlate with an aggressive tumour subtype. Neuropathol Appl Neurobiol 2018; 44: 185-206 [PMID: 28767130 DOI: 10.1111/nan.12428]
- Gambardella V, Castillo J, Tarazona N, Gimeno-Valiente F, Martínez-Ciarpaglini C, Cabeza-71 Segura M, Roselló S, Roda D, Huerta M, Cervantes A, Fleitas T. The role of tumor-associated macrophages in gastric cancer development and their potential as a therapeutic target. Cancer Treat Rev 2020; 86: 102015 [PMID: 32248000 DOI: 10.1016/j.ctrv.2020.102015]


- 72 Salmaninejad A, Valilou SF, Soltani A, Ahmadi S, Abarghan YJ, Rosengren RJ, Sahebkar A. Tumor-associated macrophages: role in cancer development and therapeutic implications. Cell Oncol (Dordr) 2019; 42: 591-608 [PMID: 31144271 DOI: 10.1007/s13402-019-00453-z]
- 73 Li R, Zhou R, Wang H, Li W, Pan M, Yao X, Zhan W, Yang S, Xu L, Ding Y, Zhao L. Gut microbiota-stimulated cathepsin K secretion mediates TLR4-dependent M2 macrophage polarization and promotes tumor metastasis in colorectal cancer. Cell Death Differ 2019; 26: 2447-2463 [PMID: 30850734 DOI: 10.1038/s41418-019-0312-y]
- 74 Pollard JW. Tumour-educated macrophages promote tumour progression and metastasis. Nat Rev Cancer 2004; 4: 71-78 [PMID: 14708027 DOI: 10.1038/nrc1256]
- 75 Bingle L, Brown NJ, Lewis CE. The role of tumour-associated macrophages in tumour progression: implications for new anticancer therapies. J Pathol 2002; 196: 254-265 [PMID: 11857487 DOI: 10.1002/path.1027
- 76 Cui YL, Li HK, Zhou HY, Zhang T, Li Q. Correlations of tumor-associated macrophage subtypes with liver metastases of colorectal cancer. Asian Pac J Cancer Prev 2013; 14: 1003-1007 [PMID: 23621176 DOI: 10.7314/apjcp.2013.14.2.1003]
- Hamm A, Prenen H, Van Delm W, Di Matteo M, Wenes M, Delamarre E, Schmidt T, Weitz J, 77 Sarmiento R, Dezi A, Gasparini G, Rothé F, Schmitz R, D'Hoore A, Iserentant H, Hendlisz A, Mazzone M. Tumour-educated circulating monocytes are powerful candidate biomarkers for diagnosis and disease follow-up of colorectal cancer. Gut 2016; 65: 990-1000 [PMID: 25814648 DOI: 10.1136/gutjnl-2014-308988]
- 78 Biswas SK, Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. Nat Immunol 2010; 11: 889-896 [PMID: 20856220 DOI: 10.1038/ni.1937]
- 79 Ruffell B, Affara NI, Coussens LM. Differential macrophage programming in the tumor microenvironment. Trends Immunol 2012; 33: 119-126 [PMID: 22277903 DOI: 10.1016/j.it.2011.12.001
- Kim Y, Wen X, Bae JM, Kim JH, Cho NY, Kang GH. The distribution of intratumoral macrophages 80 correlates with molecular phenotypes and impacts prognosis in colorectal carcinoma. Histopathology 2018; 73: 663-671 [PMID: 29906313 DOI: 10.1111/his.13674]
- Kawachi A, Yoshida H, Kitano S, Ino Y, Kato T, Hiraoka N. Tumor-associated CD204⁺M2 81 macrophages are unfavorable prognostic indicators in uterine cervical adenocarcinoma. Cancer Sci 2018; 109: 863-870 [PMID: 29274107 DOI: 10.1111/cas.13476]
- 82 Koelzer VH, Canonica K, Dawson H, Sokol L, Karamitopoulou-Diamantis E, Lugli A, Zlobec I. Phenotyping of tumor-associated macrophages in colorectal cancer: Impact on single cell invasion (tumor budding) and clinicopathological outcome. Oncoimmunology 2016; 5: e1106677 [PMID: 27141391 DOI: 10.1080/2162402X.2015.1106677]
- 83 Malesci A, Bianchi P, Celesti G, Basso G, Marchesi F, Grizzi F, Di Caro G, Cavalleri T, Rimassa L, Palmqvist R, Lugli A, Koelzer VH, Roncalli M, Mantovani A, Ogino S, Laghi L. Tumor-associated macrophages and response to 5-fluorouracil adjuvant therapy in stage III colorectal cancer. Oncoimmunology 2017; 6: e1342918 [PMID: 29209561 DOI: 10.1080/2162402X.2017.1342918]
- 84 Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pagès C, Tosolini M, Camus M, Berger A, Wind P, Zinzindohoué F, Bruneval P, Cugnenc PH, Trajanoski Z, Fridman WH, Pagès F. Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. Science 2006; 313: 1960-1964 [PMID: 17008531 DOI: 10.1126/science.1129139]
- Porta C, Rimoldi M, Raes G, Brys L, Ghezzi P, Di Liberto D, Dieli F, Ghisletti S, Natoli G, De Baetselier P, Mantovani A, Sica A. Tolerance and M2 (alternative) macrophage polarization are related processes orchestrated by p50 nuclear factor kappaB. Proc Natl Acad Sci USA 2009; 106: 14978-14983 [PMID: 19706447 DOI: 10.1073/pnas.0809784106]
- 86 Saccani A, Schioppa T, Porta C, Biswas SK, Nebuloni M, Vago L, Bottazzi B, Colombo MP, Mantovani A, Sica A. p50 nuclear factor-kappaB overexpression in tumor-associated macrophages inhibits M1 inflammatory responses and antitumor resistance. Cancer Res 2006; 66: 11432-11440 [PMID: 17145890 DOI: 10.1158/0008-5472.CAN-06-1867]
- 87 Porta C, Ippolito A, Consonni FM, Carraro L, Celesti G, Correale C, Grizzi F, Pasqualini F, Tartari S, Rinaldi M, Bianchi P, Balzac F, Vetrano S, Turco E, Hirsch E, Laghi L, Sica A. Protumor Steering of Cancer Inflammation by p50 NF-KB Enhances Colorectal Cancer Progression. Cancer Immunol Res 2018; 6: 578-593 [PMID: 29588321 DOI: 10.1158/2326-6066.CIR-17-0036]
- Fliegauf M, Grimbacher B. Nuclear factor KB mutations in human subjects: The devil is in the 88 details. J Allergy Clin Immunol 2018; 142: 1062-1065 [PMID: 30165054 DOI: 10.1016/j.jaci.2018.06.050]
- Ruffell B, Coussens LM. Macrophages and therapeutic resistance in cancer. Cancer Cell 2015; 27: 89 462-472 [PMID: 25858805 DOI: 10.1016/j.ccell.2015.02.015]
- 90 Hao NB, Lü MH, Fan YH, Cao YL, Zhang ZR, Yang SM. Macrophages in tumor microenvironments and the progression of tumors. Clin Dev Immunol 2012; 2012: 948098 [PMID: 22778768 DOI: 10.1155/2012/948098
- Zhang S, Zhong M, Wang C, Xu Y, Gao WQ, Zhang Y. CCL5-deficiency enhances intratumoral 91 infiltration of CD8⁺T cells in colorectal cancer. Cell Death Dis 2018; 9: 766 [PMID: 29991744 DOI: 10.1038/s41419-018-0796-2]
- Liu C, Yao Z, Wang J, Zhang W, Yang Y, Zhang Y, Qu X, Zhu Y, Zou J, Peng S, Zhao Y, Zhao S, 92 He B, Mi Q, Liu X, Zhang X, Du Q. Macrophage-derived CCL5 facilitates immune escape of colorectal cancer cells via the p65/STAT3-CSN5-PD-L1 pathway. Cell Death Differ 2020; 27: 1765-



1781 [PMID: 31802034 DOI: 10.1038/s41418-019-0460-0]

- 93 Wang X, Lang M, Zhao T, Feng X, Zheng C, Huang C, Hao J, Dong J, Luo L, Li X, Lan C, Yu W, Yu M, Yang S, Ren H. Cancer-FOXP3 directly activated CCL5 to recruit FOXP3⁺ Treg cells in pancreatic ductal adenocarcinoma. Oncogene 2017; 36: 3048-3058 [PMID: 27991933 DOI: 10.1038/onc.2016.458]
- 94 Wu S, He H, Liu H, Cao Y, Li R, Zhang H, Li H, Shen Z, Qin J, Xu J. C-C motif chemokine 22 predicts postoperative prognosis and adjuvant chemotherapeutic benefits in patients with stage II/III gastric cancer. Oncoimmunology 2018; 7: e1433517 [PMID: 29872564 DOI: 10.1080/2162402X.2018.1433517]
- 95 Kumai T, Nagato T, Kobayashi H, Komabayashi Y, Ueda S, Kishibe K, Ohkuri T, Takahara M, Celis E, Harabuchi Y. CCL17 and CCL22/CCR4 signaling is a strong candidate for novel targeted therapy against nasal natural killer/T-cell lymphoma. Cancer Immunol Immunother 2015; 64: 697-705 [PMID: 25754123 DOI: 10.1007/s00262-015-1675-7]
- 96 Wei C, Yang C, Wang S, Shi D, Zhang C, Lin X, Xiong B. M2 macrophages confer resistance to 5fluorouracil in colorectal cancer through the activation of CCL22/PI3K/AKT signaling. Onco Targets Ther 2019; 12: 3051-3063 [PMID: 31114248 DOI: 10.2147/OTT.S198126]
- 97 Cao L, Hu X, Zhang J, Huang G, Zhang Y. The role of the CCL22-CCR4 axis in the metastasis of gastric cancer cells into omental milky spots. J Transl Med 2014; 12: 267 [PMID: 25245466 DOI: 10.1186/s12967-014-0267-1]
- Yeung OW, Lo CM, Ling CC, Qi X, Geng W, Li CX, Ng KT, Forbes SJ, Guan XY, Poon RT, Fan 98 ST, Man K. Alternatively activated (M2) macrophages promote tumour growth and invasiveness in hepatocellular carcinoma. J Hepatol 2015; 62: 607-616 [PMID: 25450711 DOI: 10.1016/j.jhep.2014.10.029]
- 99 Gilkes DM, Semenza GL, Wirtz D. Hypoxia and the extracellular matrix: drivers of tumour metastasis. Nat Rev Cancer 2014; 14: 430-439 [PMID: 24827502 DOI: 10.1038/nrc3726]
- Spill F, Reynolds DS, Kamm RD, Zaman MH. Impact of the physical microenvironment on tumor 100 progression and metastasis. Curr Opin Biotechnol 2016; 40: 41-48 [PMID: 26938687 DOI: 10.1016/j.copbio.2016.02.007
- 101 Henze AT, Mazzone M. The impact of hypoxia on tumor-associated macrophages. J Clin Invest 2016; 126: 3672-3679 [PMID: 27482883 DOI: 10.1172/JCI84427]
- 102 Doedens AL, Stockmann C, Rubinstein MP, Liao D, Zhang N, DeNardo DG, Coussens LM, Karin M, Goldrath AW, Johnson RS. Macrophage expression of hypoxia-inducible factor-1 alpha suppresses T-cell function and promotes tumor progression. Cancer Res 2010; 70: 7465-7475 [PMID: 20841473 DOI: 10.1158/0008-5472.CAN-10-1439]
- 103 Imtiyaz HZ, Williams EP, Hickey MM, Patel SA, Durham AC, Yuan LJ, Hammond R, Gimotty PA, Keith B, Simon MC. Hypoxia-inducible factor 2alpha regulates macrophage function in mouse models of acute and tumor inflammation. J Clin Invest 2010; 120: 2699-2714 [PMID: 20644254 DOI: 10.1172/JCI39506]
- 104 Movahedi K, Laoui D, Gysemans C, Baeten M, Stangé G, Van den Bossche J, Mack M, Pipeleers D, In't Veld P, De Baetselier P, Van Ginderachter JA. Different tumor microenvironments contain functionally distinct subsets of macrophages derived from Ly6C(high) monocytes. Cancer Res 2010; 70: 5728-5739 [PMID: 20570887 DOI: 10.1158/0008-5472.CAN-09-4672]
- 105 Casazza A, Laoui D, Wenes M, Rizzolio S, Bassani N, Mambretti M, Deschoemaeker S, Van Ginderachter JA, Tamagnone L, Mazzone M. Impeding macrophage entry into hypoxic tumor areas by Sema3A/Nrp1 signaling blockade inhibits angiogenesis and restores antitumor immunity. Cancer Cell 2013; 24: 695-709 [PMID: 24332039 DOI: 10.1016/j.ccr.2013.11.007]
- 106 Coussens LM, Zitvogel L, Palucka AK. Neutralizing tumor-promoting chronic inflammation: a magic bullet? Science 2013; 339: 286-291 [PMID: 23329041 DOI: 10.1126/science.1232227]
- 107 Eltzschig HK, Carmeliet P. Hypoxia and inflammation. N Engl J Med 2011; 364: 656-665 [PMID: 21323543 DOI: 10.1056/NEJMra0910283]
- De Palma M, Lewis CE. Macrophage regulation of tumor responses to anticancer therapies. Cancer 108 Cell 2013; 23: 277-286 [PMID: 23518347 DOI: 10.1016/j.ccr.2013.02.013]
- 109 Qi L, Chen J, Yang Y, Hu W. Hypoxia Correlates With Poor Survival and M2 Macrophage Infiltration in Colorectal Cancer. Front Oncol 2020; 10: 566430 [PMID: 33330037 DOI: 10.3389/fonc.2020.566430]
- Zhao Y, Zhao B, Wang X, Guan G, Xin Y, Sun YD, Wang JH, Guo Y, Zang YJ. Macrophage 110 transcriptome modification induced by hypoxia and lactate. Exp Ther Med 2019; 18: 4811-4819 [PMID: 31798707 DOI: 10.3892/etm.2019.8164]
- 111 Zhang Y, Yang H, Zhao J, Wan P, Hu Y, Lv K, Yang X, Ma M. Activation of MAT2A-RIP1 signaling axis reprograms monocytes in gastric cancer. J Immunother Cancer 2021: 9 [PMID: 33593829 DOI: 10.1136/jitc-2020-001364]
- 112 Kondo H, Ratcliffe CDH, Hooper S, Ellis J, MacRae JI, Hennequart M, Dunsby CW, Anderson KI, Sahai E. Single-cell resolved imaging reveals intra-tumor heterogeneity in glycolysis, transitions between metabolic states, and their regulatory mechanisms. Cell Rep 2021; 34: 108750 [PMID: 33596424 DOI: 10.1016/j.celrep.2021.108750]
- Geeraerts X, Bolli E, Fendt SM, Van Ginderachter JA. Macrophage Metabolism As Therapeutic 113 Target for Cancer, Atherosclerosis, and Obesity. Front Immunol 2017; 8: 289 [PMID: 28360914 DOI: 10.3389/fimmu.2017.00289]
- Miao H, Ou J, Peng Y, Zhang X, Chen Y, Hao L, Xie G, Wang Z, Pang X, Ruan Z, Li J, Yu L, Xue 114



B, Shi H, Shi C, Liang H. Macrophage ABHD5 promotes colorectal cancer growth by suppressing spermidine production by SRM. Nat Commun 2016; 7: 11716 [PMID: 27189574 DOI: 10.1038/ncomms11716

- 115 Ghafouri N, Tiger G, Razdan RK, Mahadevan A, Pertwee RG, Martin BR, Fowler CJ. Inhibition of monoacylglycerol lipase and fatty acid amide hydrolase by analogues of 2-arachidonoylglycerol. Br J Pharmacol 2004; 143: 774-784 [PMID: 15492019 DOI: 10.1038/sj.bjp.0705948]
- 116 Lass A, Zimmermann R, Haemmerle G, Riederer M, Schoiswohl G, Schweiger M, Kienesberger P, Strauss JG, Gorkiewicz G, Zechner R. Adipose triglyceride lipase-mediated lipolysis of cellular fat stores is activated by CGI-58 and defective in Chanarin-Dorfman Syndrome. Cell Metab 2006; 3: 309-319 [PMID: 16679289 DOI: 10.1016/j.cmet.2006.03.005]
- 117 Meana C, García-Rostán G, Peña L, Lordén G, Cubero Á, Orduña A, Győrffy B, Balsinde J, Balboa MA. The phosphatidic acid phosphatase lipin-1 facilitates inflammation-driven colon carcinogenesis. JCI Insight 2018; 3 [PMID: 30232275 DOI: 10.1172/jci.insight.97506]
- 118 Kofuji S, Kimura H, Nakanishi H, Nanjo H, Takasuga S, Liu H, Eguchi S, Nakamura R, Itoh R, Ueno N, Asanuma K, Huang M, Koizumi A, Habuchi T, Yamazaki M, Suzuki A, Sasaki J, Sasaki T. INPP4B Is a PtdIns(3,4,5)P3 Phosphatase That Can Act as a Tumor Suppressor. Cancer Discov 2015; 5: 730-739 [PMID: 25883023 DOI: 10.1158/2159-8290.CD-14-1329]
- 119 Zhang X, Zhang L, Lin B, Chai X, Li R, Liao Y, Deng X, Liu Q, Yang W, Cai Y, Zhou W, Lin Z, Huang W, Zhong M, Lei F, Wu J, Yu S, Li X, Li S, Li Y, Zeng J, Long W, Ren D, Huang Y. Phospholipid Phosphatase 4 promotes proliferation and tumorigenesis, and activates Ca²⁺-permeable Cationic Channel in lung carcinoma cells. Mol Cancer 2017; 16: 147 [PMID: 28851360 DOI: 10.1186/s12943-017-0717-51
- 120 Zhang D, Shi R, Xiang W, Kang X, Tang B, Li C, Gao L, Zhang X, Zhang L, Dai R, Miao H. The Agpat4/LPA axis in colorectal cancer cells regulates antitumor responses via p38/p65 signaling in macrophages. Signal Transduct Target Ther 2020; 5: 24 [PMID: 32296017 DOI: 10.1038/s41392-020-0117-y]
- 121 Pyonteck SM, Akkari L, Schuhmacher AJ, Bowman RL, Sevenich L, Quail DF, Olson OC, Quick ML, Huse JT, Teijeiro V, Setty M, Leslie CS, Oei Y, Pedraza A, Zhang J, Brennan CW, Sutton JC, Holland EC, Daniel D, Joyce JA. CSF-1R inhibition alters macrophage polarization and blocks glioma progression. Nat Med 2013; 19: 1264-1272 [PMID: 24056773 DOI: 10.1038/nm.3337]
- 122 Komohara Y, Jinushi M, Takeya M. Clinical significance of macrophage heterogeneity in human malignant tumors. Cancer Sci 2014; 105: 1-8 [PMID: 24168081 DOI: 10.1111/cas.12314]
- Peranzoni E, Lemoine J, Vimeux L, Feuillet V, Barrin S, Kantari-Mimoun C, Bercovici N, Guérin 123 M, Biton J, Ouakrim H, Régnier F, Lupo A, Alifano M, Damotte D, Donnadieu E. Macrophages impede CD8 T cells from reaching tumor cells and limit the efficacy of anti-PD-1 treatment. Proc Natl Acad Sci US A 2018; 115: E4041-E4050 [PMID: 29632196 DOI: 10.1073/pnas.1720948115]
- 124 Neubert NJ, Schmittnaegel M, Bordry N, Nassiri S, Wald N, Martignier C, Tillé L, Homicsko K, Damsky W, Maby-El Hajjami H, Klaman I, Danenberg E, Ioannidou K, Kandalaft L, Coukos G, Hoves S, Ries CH, Fuertes Marraco SA, Foukas PG, De Palma M, Speiser DE. T cell-induced CSF1 promotes melanoma resistance to PD1 blockade. Sci Transl Med 2018; 10 [PMID: 29643229 DOI: 10.1126/scitranslmed.aan3311
- 125 Mitchem JB, Brennan DJ, Knolhoff BL, Belt BA, Zhu Y, Sanford DE, Belavgorod L, Carpenter D, Collins L, Piwnica-Worms D, Hewitt S, Udupi GM, Gallagher WM, Wegner C, West BL, Wang-Gillam A, Goedegebuure P, Linehan DC, DeNardo DG. Targeting tumor-infiltrating macrophages decreases tumor-initiating cells, relieves immunosuppression, and improves chemotherapeutic responses. Cancer Res 2013; 73: 1128-1141 [PMID: 23221383 DOI: 10.1158/0008-5472.CAN-12-2731]
- 126 Seifert L, Werba G, Tiwari S, Giao Ly NN, Nguy S, Alothman S, Alqunaibit D, Avanzi A, Daley D, Barilla R, Tippens D, Torres-Hernandez A, Hundeyin M, Mani VR, Hajdu C, Pellicciotta I, Oh P, Du K, Miller G. Radiation Therapy Induces Macrophages to Suppress T-Cell Responses Against Pancreatic Tumors in Mice. Gastroenterology 2016; 150: 1659-1672.e5 [PMID: 26946344 DOI: 10.1053/j.gastro.2016.02.070]
- 127 Ruffell B, Chang-Strachan D, Chan V, Rosenbusch A, Ho CM, Pryer N, Daniel D, Hwang ES, Rugo HS, Coussens LM. Macrophage IL-10 blocks CD8+ T cell-dependent responses to chemotherapy by suppressing IL-12 expression in intratumoral dendritic cells. Cancer Cell 2014; 26: 623-637 [PMID: 25446896 DOI: 10.1016/j.ccell.2014.09.006]
- 128 Hughes R, Qian BZ, Rowan C, Muthana M, Keklikoglou I, Olson OC, Tazzyman S, Danson S, Addison C, Clemons M, Gonzalez-Angulo AM, Joyce JA, De Palma M, Pollard JW, Lewis CE. Perivascular M2 Macrophages Stimulate Tumor Relapse after Chemotherapy. Cancer Res 2015; 75: 3479-3491 [PMID: 26269531 DOI: 10.1158/0008-5472.CAN-14-3587]
- 129 Sánchez-Martín L, Estecha A, Samaniego R, Sánchez-Ramón S, Vega MÁ, Sánchez-Mateos P. The chemokine CXCL12 regulates monocyte-macrophage differentiation and RUNX3 expression. Blood 2011; 117: 88-97 [PMID: 20930067 DOI: 10.1182/blood-2009-12-258186]
- 130 Kang KA, Ryu YS, Piao MJ, Shilnikova K, Kang HK, Yi JM, Boulanger M, Paolillo R, Bossis G, Yoon SY, Kim SB, Hyun JW. DUOX2-mediated production of reactive oxygen species induces epithelial mesenchymal transition in 5-fluorouracil resistant human colon cancer cells. Redox Biol 2018; 17: 224-235 [PMID: 29715584 DOI: 10.1016/j.redox.2018.04.020]
- 131 D'Alterio C, Zannetti A, Trotta AM, Ieranò C, Napolitano M, Rea G, Greco A, Maiolino P, Albanese S, Scognamiglio G, Tatangelo F, Tafuto S, Portella L, Santagata S, Nasti G, Ottaiano A,



Pacelli R, Delrio P, Botti G, Scala S. New CXCR4 Antagonist Peptide R (Pep R) Improves Standard Therapy in Colorectal Cancer. Cancers (Basel) 2020; 12 [PMID: 32708431 DOI: 10.3390/cancers12071952

- 132 Zboralski D, Hoehlig K, Eulberg D, Frömming A, Vater A. Increasing Tumor-Infiltrating T Cells through Inhibition of CXCL12 with NOX-A12 Synergizes with PD-1 Blockade. Cancer Immunol Res 2017; 5: 950-956 [PMID: 28963140 DOI: 10.1158/2326-6066.CIR-16-0303]
- 133 Ghesquière B, Wong BW, Kuchnio A, Carmeliet P. Metabolism of stromal and immune cells in health and disease. Nature 2014; 511: 167-176 [PMID: 25008522 DOI: 10.1038/nature13312]
- 134 Weichhart T, Hengstschläger M, Linke M. Regulation of innate immune cell function by mTOR. Nat Rev Immunol 2015; 15: 599-614 [PMID: 26403194 DOI: 10.1038/nri3901]
- 135 Rodrik-Outmezguine VS, Okaniwa M, Yao Z, Novotny CJ, McWhirter C, Banaji A, Won H, Wong W, Berger M, de Stanchina E, Barratt DG, Cosulich S, Klinowska T, Rosen N, Shokat KM. Overcoming mTOR resistance mutations with a new-generation mTOR inhibitor. Nature 2016; 534: 272-276 [PMID: 27279227 DOI: 10.1038/nature17963]
- Thiem S, Pierce TP, Palmieri M, Putoczki TL, Buchert M, Preaudet A, Farid RO, Love C, Catimel 136 B, Lei Z, Rozen S, Gopalakrishnan V, Schaper F, Hallek M, Boussioutas A, Tan P, Jarnicki A, Ernst M. mTORC1 inhibition restricts inflammation-associated gastrointestinal tumorigenesis in mice. J Clin Invest 2013; 123: 767-781 [PMID: 23321674 DOI: 10.1172/JCI65086]
- Kaneda MM, Messer KS, Ralainirina N, Li H, Leem CJ, Gorjestani S, Woo G, Nguyen AV, 137 Figueiredo CC, Foubert P, Schmid MC, Pink M, Winkler DG, Rausch M, Palombella VJ, Kutok J, McGovern K, Frazer KA, Wu X, Karin M, Sasik R, Cohen EE, Varner JA. PI3Ky is a molecular switch that controls immune suppression. Nature 2016; 539: 437-442 [PMID: 27642729 DOI: 10.1038/nature198341
- 138 Sahin E, Haubenwallner S, Kuttke M, Kollmann I, Halfmann A, Dohnal AM, Chen L, Cheng P, Hoesel B, Einwallner E, Brunner J, Kral JB, Schrottmaier WC, Thell K, Saferding V, Blüml S, Schabbauer G. Macrophage PTEN regulates expression and secretion of arginase I modulating innate and adaptive immune responses. J Immunol 2014; 193: 1717-1727 [PMID: 25015834 DOI: 10.4049/jimmunol.1302167
- 139 Corna G, Campana L, Pignatti E, Castiglioni A, Tagliafico E, Bosurgi L, Campanella A, Brunelli S, Manfredi AA, Apostoli P, Silvestri L, Camaschella C, Rovere-Querini P. Polarization dictates iron handling by inflammatory and alternatively activated macrophages. Haematologica 2010; 95: 1814-1822 [PMID: 20511666 DOI: 10.3324/haematol.2010.023879]
- Recalcati S, Locati M, Marini A, Santambrogio P, Zaninotto F, De Pizzol M, Zammataro L, Girelli 140 D, Cairo G. Differential regulation of iron homeostasis during human macrophage polarized activation. Eur J Immunol 2010; 40: 824-835 [PMID: 20039303 DOI: 10.1002/eji.200939889]
- 141 Torti SV, Torti FM. Ironing out cancer. Cancer Res 2011; 71: 1511-1514 [PMID: 21363917 DOI: 10.1158/0008-5472.CAN-10-3614]
- Mertens C, Akam EA, Rehwald C, Brüne B, Tomat E, Jung M. Intracellular Iron Chelation 142 Modulates the Macrophage Iron Phenotype with Consequences on Tumor Progression. PLoS One 2016; 11: e0166164 [PMID: 27806101 DOI: 10.1371/journal.pone.0166164]



WJGO | https://www.wjgnet.com

WJGO

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2029-2037

DOI: 10.4251/wjgo.v13.i12.2029

ISSN 1948-5204 (online)

MINIREVIEWS

Advancement of chimeric antigen receptor-natural killer cells targeting hepatocellular carcinoma

Kai Dai, Yin Wu, Sha She, Qian Zhang

ORCID number: Kai Dai 0000-0001-7270-9965; Yin Wu 0000-0002-0987-2097; Sha She 0000-0002-1863-957X; Qian Zhang 0000-0003-1053-1588.

Author contributions: Dai K performed the majority of the writing and prepared the table; Wu Y, She S and Zhang Q carried out the literature review for data collection, and coordinated the writing of the paper; all authors have read and approve the final manuscript.

Conflict-of-interest statement: The authors declare no conflicts of interest

Supported by The National Natural Science Foundation of China, No. 81972673.

Country/Territory of origin: China

Specialty type: Oncology

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an

Kai Dai, Yin Wu, Sha She, Qian Zhang, Department of Infectious Diseases, Renmin Hospital of Wuhan University, Wuhan 430060, Hubei Province, China

Corresponding author: Kai Dai, MD, PhD, Doctor, Department of Infectious Diseases, Renmin Hospital of Wuhan University, No. 238 Jiefang Road, Wuchang District, Wuhan 430060, Hubei Province, China. daikai@whu.edu.cn

Abstract

With the advance of genome engineering technology, chimeric antigen receptors (CARs)-based immunotherapy has become an emerging therapeutic strategy for tumors. Although initially designed for T cells in tumor immunotherapy, CARs have been exploited to modify the function of natural killer (NK) cells against a variety of tumors, including hepatocellular carcinoma (HCC). CAR-NK cells have the potential to sufficiently kill tumor antigen-expressing HCC cells, independent of major histocompatibility complex matching or prior priming. In this review, we summarize the recent advances in genetic engineering of CAR-NK cells against HCC and discuss the current challenges and prospects of CAR-NK cells as a revolutionary cellular immunotherapy against HCC.

Key Words: Chimeric antigen receptors; Natural killer cells; Hepatocellular carcinoma; Immunotherapy; Genome engineering

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Chimeric antigen receptors (CARs)-based immunotherapy is an emerging therapeutic strategy for tumors. This review summarizes the recent advances in genetic engineering of CAR-natural killer (NK) cells against hepatocellular carcinoma and discuss the current challenges and prospects of CAR-NK cells as a revolutionary cellular immunotherapy against hepatocellular carcinoma.

Citation: Dai K, Wu Y, She S, Zhang Q. Advancement of chimeric antigen receptor-natural killer cells targeting hepatocellular carcinoma. World J Gastrointest Oncol 2021; 13(12): 2029-2037

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2029.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2029



open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: February 20, 2021 Peer-review started: February 20, 2021 First decision: July 29, 2021

Revised: August 4, 2021 Accepted: October 27, 2021 Article in press: October 27, 2021 Published online: December 15, 2021

P-Reviewer: Inmutto N, Trotovšek B S-Editor: Wang LL L-Editor: A P-Editor: Wang LL



INTRODUCTION

Hepatocellular carcinoma (HCC) is the predominant form of primary liver cancer, the sixth most commonly diagnosed cancer, and the third leading cause of cancer death worldwide[1,2]. The key risk factors of HCC include infection with hepatitis B virus or hepatitis C virus, alcohol consumption, non-alcoholic fatty liver disease, and aflatoxin exposure[3]. Surgery is the prevalent treatment for HCC. However, recurrence of HCC severely decreases the survival rate.

Locoregional therapies, including percutaneous ablation and intra-arterial chemotherapy, are alternative treatment approaches and rely on the grade of HCC. Unfortunately, more than 70% of patients with advanced HCC have little chance for transplant, surgery, or locoregional therapy. In the recent decade, cancer immunotherapy has become a novel emerging therapeutic strategy, improving treatment effect via mobilizing patients' immune systems to launch efficient anti-tumor reactions against cancer progression. Adoptive transfer of immune cells, as an immunotherapeutic approach, is expected to improve HCC prognosis by passively infusing autologous or allogenic leukocytes after ex vivo expansion and activation. Natural killer (NK) cells are a persistent research focus in the field of immunotherapy, due to their significant anti-tumor activity.

NK CELL BIOLOGY

NK cells, which make up 5%-15% of human blood leukocytes and 50% of hepatic innate immune cells, are the major innate lymphocytes executing anti-tumor and antiviral immunity^[4]. Based on the expression of cluster of differentiation (CD)16 and CD56, human NK cells can be divided into two subpopulations. The majority (85%-95%) of the peripheral blood NK cells are CD56^{dim}CD16^{high}, showing a developmentally mature phenotype, and mediate high cytotoxicity upon encountering target cells. The remaining minority of NK cells are the CD56^{bright}CD16^{low/-} subset, which have an immature phenotype and lower cytotoxicity upon activation[5].

Unlike T cells, the cytotoxicity of NK cells does not rely on T cell receptor (TCR)mediated specific antigen recognition and is thus independent of major histocompatibility complex (MHC) expression. Instead, NK cell activity is regulated by a wide spectrum of activating and inhibitory receptors (Table 1) that induce positive and negative signals to comprehensively control NK cell behavior upon contacting target cells[6]. In this regard, tumor cells that downregulate MHC expression are sensitive to NK cell cytotoxicity, owing to weaker killer cell immunoglobulin-like receptor (commonly known as KIR)-mediated inhibitory signals. Similar to cytotoxic T cells, activated NK cells express and release lytic molecules, such as perforin and granzyme B, to kill tumor cells. Additionally, NK cells secrete an array of cytokines, including interferon-g, tumor necrosis factor (TNF), and granulocyte-macrophage colonystimulating factor, to induce the activation of T cells and innate immune cells (dendritic cells, macrophages, and neutrophils), and result in the promotion of immunity against malignant cells and the tumorigenic microenvironment[4].

CARS FOR TUMOR TREATMENT

Thanks to genome editing technology, CARs were initially designed for T cellmediated immunotherapy against relapsed or refractory B-cell malignancies[7]. A CAR is an antibody-derived antigen-recognizing domain, linked with T cell signaling domains. When expressed by a T cell, the CAR targets a specific antigen and transduces activating signals to that T cell. A CAR consists of three domains, namely an ectodomain, a transmembrane domain, and an intracellular signaling domain[8]. The ectodomain is a single-chain variable fragment (scFv) comprising the variable regions of heavy and light chains of an antibody connected with a short linker peptide of 10-25 amino acids. The intracellular domain is conventionally derived from the TCR CD3 ζ chain and/or other key accessory proteins for relaying activating signals. Three generations of CARs have been developed to date. The first-generation CARs contain CD3 ζ alone. The second-generation CARs integrate an extra costimulatory domain from CD28 or 4-1BB. The third-generation CARs contain more than one costimulatory domain[9]. The latest ³/₄ fourth-generation ³/₄ CARs possess supplementary exogenous proteins, such as cytokines, to enhance the lifespan and effector function of NK cells [10].



Table 1 Activating and inhibitory receptors on human natural killer cells				
	Receptor	Ligand		
Activating receptors	CD16	Fc region of IgG		
	NKG2D	MICA/B, ULBP1-6		
	NKp30 (CD337)	B7-H6, BAT3, HCMV pp65		
	NKp44 (CD336)	Viral HA and HN, PCNA, MLL5, PDGF-DD		
	NKp46 (CD335)	Viral HA and HN, CFP		
	NKp65	KACL		
	NKp80	AICL1		
	CD94-NKG2C/E/H	HLA-E (for CD94-NKG2C)		
	2B4 (CD244)	CD48		
	DNAM-1 (CD226)	PVR (Necl5, CD155), nectin 2 (CD112)		
	Activating KIRs	HLA-A11, -Bw4		
	CRTAM (CD355)	Necl-2		
	Tactile (CD96)	CD155		
Inhibitory receptors	Inhibitory KIRs	HLA class I		
	CD94-NKG2A/B	HLA-E		
	NKR-P1A	LLT1		
	TIGIT	PVR (Necl5, CD155), nectin 2 (CD112)		
	LIR-1 (ILT-2/CD85j/LILRB1)	HLA (α3), HCMV UL18		
	IRp60 (CD300a)	PS, PE		
	CEACAM1 (CD66)	CEACAM1 (CD66), TIM-3 (HAVCR2)		

AICL: Activation-induced C-type lectin; BAT3: Human leukocyte antigen-B-associated transcript 3; CEACAM: Carcinoembryonic antigen-related cell adhesion molecules; CFP: Complement factor P; CRTAM: Class I-restricted T cell-associated molecule; DNAM-1: DNAX accessory molecule 1; H60: Minor histocompatibility protein 60; HA: Hemagglutinin; HAVCR2: Hepatitis A virus cellular receptor 2; HCMV: Human cytomegalovirus; HN: Hemagglutinin-neuraminidase; IgG: Immunoglobulin G; ILT-2: Immunoglobulin-like transcript 2; IRp60: Inhibitory receptor protein 60; KACL: Keratinocyte-associated C-type lectin; KIR: Killer-cell immunoglobulin-like receptor; LILRB1: Leukocyte immunoglobulin-like receptor 1; LLT1: Lectin-like transcript 1; MHC: Major histocompatibility complex; MIC: MHC class I chain-related protein; MLL5: Mixed lineage leukemia 5; MULT-1: Mouse UL-16- binding protein-like transcript 1; Necl: Nectin-like molecules; PCNA: Proliferating cell nuclear antigen; PDGF: Platelet-derived growth factor; PE: Phosphatidylethanolamine; PILR: Paired immunoglobulin-like type 2 receptor; PS: Phosphatidylserine; PVR: Poliovirus receptor; Rae-1: Retinoic acid early inducible 1; TIGIT: T cell immunoglobulin and immunoreceptor tyrosine-based activation motif domain; TIM-3: T cell immunoglobulin domain and mucin domain 3; ULBP: UL16 binding proteins.

Although T cells armed with CARs (hereinafter referred to as "CAR-T cells") have demonstrated unprecedented efficacy in boosting anti-tumor immunity, but they have intrinsic shortcomings[11,12]. First, CAR-T cells require prior antigen priming, which depends on the expression of MHC molecules on target cells. However, various types of tumor cells downregulate MHC expression and therefore escape from the attack of CAR-T cells. Second, before infusion of CAR-T cells, patients have to receive lymphodepleting treatment to facilitate the *in vivo* persistence of the CAR-T cells. However, a large number of CAR-T cells in the recipient increases the risk of graft *vs* host disease (commonly known as GVHD). Third, activated CAR-T cells produce enormous amounts of cytokines, including interleukin (IL)-1a, IL-2, IL-6, TNF- α , MCP-1, IL-8, IL-10 and IL-15, rendering the cytokine release syndrome (otherwise known as CRS) and neurotoxicity. Compared to T cells, NK cells overcome these disadvantages because they function in an MHC-independent manner, without triggering GVHD, and produce a distinct spectrum of cytokines.

Baishidena® WJGO | https://www.wjgnet.com

CAR-NK CELLS

CAR constructs for NK cells

For NK cells, the CAR structure is quite similar to that of T cells. The extracellular scFv is also derived from an antibody against a tumor antigen. So far, the tumor antigens targeted by engineered scFv involve CD19 and CD20 (B-cell acute and chronic leukemia antigen)[13], human epidermal growth factor receptor 2 (commonly known as HER2; breast cancer)[14], GD2 (neuroblastoma and melanoma)[15], CD138 (multiple myeloma)[16], CD4 (T cell lymphoma)[17], and epidermal growth factor receptor (EGFR) which is overexpressed in multiple tumors [18]. Some researchers have designed CAR-NK cells targeting certain immunosuppressive cells, such as alternatively-activated macrophages and myeloid-derived suppressor cells, to enhance antitumor immunity [19,20]. Moreover, a recent endeavor has been made to create a bispecific CAR featuring an scFv that targets two antigens simultaneously. NK cells equipped with this bispecific CAR are supposed to execute cytotoxicity more efficiently, in case one antigen is weakly expressed or absent on certain tumor cells[21].

The transmembrane domain of a CAR belongs to CD8a, CD28, CD3, or ICOS, with some exceptions derived from NKG2D[22]. Recent findings suggest that the sequences and locations of the transmembrane domain should be delicately designed to achieve the best function[23]. The intracellular domain is key to the signal transduction activity and consequent NK cell activation upon the engagement of the scFv with tumor antigens. Most CAR-NK cells exploit CD3ζ as a signaling domain to transduce activating signals because the immunoreceptor tyrosine-based activation motif (also known as ITAM) of CD3ζ initiates NK cell killing[24-26]. Additional costimulatory domains derived from CD28, CD137 (4-1BB), or CD244 (2B4) are usually required for efficient killing[22,27]. In recent research, replacing CD3ζ with the intracellular region of DAP12 has resulted in a better effect on glioblastoma[28]. The most common CAR constructs used in CAR-NK cells have been summarized elsewhere[8].

Sources of CAR-NK cells

Currently, clinical-grade NK cells are manufactured on a large scale from the following sources.

NK92 cell line: The NK-92 cell line was established from a patient with large granular lymphoma in 1994. It does not attack recipients' allogeneic cells but kills a wide range of tumor cells, such as leukemia and melanoma. The cell line is easy to grow in vitro, making it an attractive agent for adoptive cancer immunotherapy. However, its tumorigenicity potential, deficiency in CD16 and NKp44 expression, and the requirement for lethal irradiation before adoptive transfer pose concerns about clinical safety and therapeutic efficiency^[29].

Peripheral blood mononuclear cells (PBMCs): Sufficient allogeneic NK cells can be enriched from PBMCs of a patient or healthy donor, followed by ex vivo stimulation, expansion, and genetic engineering. PBMC-derived CAR-NK cells are predominantly CD56^{dim}CD16⁺ mature NK cells with high cytotoxicity, making them especially suitable for either autologous or allogeneic transfer[8].

Umbilical cord blood (UBC): As an alternative NK cell source, UBC with identified MHC types can be obtained from UBC banks. Nonetheless, the NK cell abundance in UBC is relatively low, so it takes a long ex vivo expansion and activation procedure before sufficient NK cells are generated. Another issue involves UBC-derived NK cells exhibiting an immature phenotype and being less cytotoxic to target cells[30].

Hematopoietic progenitor cells (commonly known as HPCs): CD34⁺ HPCs are sorted from bone marrow, embryonic stem cells, mobilized peripheral blood, or UBC. They are expanded and differentiated into NK cells ex vivo, under the effect of a set of cytokines. HPC-derived NK cells are mature and significantly cytotoxic to leukemia cells[31].

Induced pluripotent stem cells (iPSCs): iPSCs are derived from mature somatic cells that have been reprogrammed back into a pluripotent stem cell state. They enable the development of an unlimited source of multiple cell types needed for therapeutic purposes. Theoretically, one iPSC is competent to generate a large quantity of homogeneous CAR-NK cells. However, iPSC-derived NK cells are relatively immature and less cytotoxic, posing the challenge of inducing both phenotypically and functionally mature NK cells[8]. A recent study developed a robust and efficient manufacturing system for the differentiation and expansion of high-quality iPSC-



derived NK cells that produced inflammatory cytokines and exerted strong cytotoxicity against an array of hematologic and solid tumors[32].

CAR-NK CELLS IN HCC THERAPY

Both academia and industry have made rapid progress in developing CAR-engineered immune cells for HCC[33,34]. One of the major challenges for CAR-based immunotherapy for HCC is to target a specific, safe, and effective tumor antigen. Currently, the following tumor antigens are considered candidate targets of CAR-based immunotherapy for HCC: glypican-3 (GPC3), α -fetoprotein, epithelial cell adhesion molecule (commonly known as EpCAM), and mucin-1 (commonly known as MUC1)[35,36]. Of note, glypican-3 is a glycoprotein overexpressed on the cell surface of HCC tissues but not in healthy liver[37]. A bispecific CAR targeting GPC3 and the asialoglycoprotein receptor 1 (commonly known as ASGR1) and featuring CD3 ζ and 28BB (containing CD28 and 4-1BB signaling domains) has been tested in an HCC xenograft mouse model[38].

Although CAR-NK cells have been tested in a few clinical trials against leukemia, lymphoma, and several solid tumors, to our knowledge, no clinical trials have been performed to evaluate the efficacy of CAR-NK cells in HCC treatment. Several laboratory studies of CAR-NK cells against HCC have been conducted and showed the bright prospect of CAR-NK-based HCC immunotherapy.

The first report of CAR-NK cells against HCC was published in 2018. Yu et al[39] developed GPC3-CAR-NK-92 cells and demonstrated the potent anti-tumor properties of these cells. The CAR construct comprised a CD8a signal peptide, a humanized GPC3-specific scFv (known as "hu9F2"), a CD8a hinge region, and a CD28 transmembrane region followed by the intracellular domains of CD28 and CD3ζ. The GPC3-CAR-NK-92 cells showed potent cytotoxicity and cytokine production upon encountering HCC cells with either high or low GPC3 expression, both in vitro and in vivo. Notably, the GPC3-CAR-NK-92 cells were cytotoxic to GFP3-negative HCC cells. Huang et al[40] made a further modification to GPC3-CAR-NK-92 cells by replacing the intracellular domains of CD28 and CD3ζ with the costimulatory domains of DNAM1 and 2B4. DNAM1, also known as CD226, is a costimulatory receptor in cytotoxic T cells, NK cells, and monocytes[41]. DNAM1 cross-linking results in phosphorylation of its cytoplasmic tyrosine residues and drives NK cell cytotoxicity [42]. 2B4, as above-mentioned, is known as CD244 and harbors a cytoplasmic domain containing immunoreceptor tyrosine-based switch motifs (commonly known as ITSMs) that are responsible for interacting with multiple signaling adaptors and transmitting activating signals[43]. This modification endowed GPC3-CAR-NK-92 cells with faster expansion, lower apoptosis and higher cytotoxic abilities than their original counterparts[40].

Tseng et al[44] reported that NK cells transduced with a CAR that targets CD147, a cell surface marker that is significantly upregulated in HCC, can effectively kill various malignant HCC cell lines in vitro, as well as HCC tumors in xenograft and patient-derived xenograft mouse models. In this study, the CD147-CAR contained the scFv of the anti-CD147 antibody derived from clone 5F6 with optimization, a human IgG1-CH2CH3 spacer, a transmembrane domain of CD28, the intracellular domain of CD28-4-1BB, and intracellular signaling domains of the TCR- ζ chain. The CARencoding retrovirus was transduced into the NK-92MI cell line, which is an IL-2dependent NK cell line derived from PBMCs obtained from a patient with rapidly progressive non-Hodgkin's lymphoma. These CD147-CAR-NK-92MI cells were effectively activated by CD147+ HCC cell lines and demonstrated impressive cytotoxicity against the target HCC cell lines. When using primary PBMC-derived NK cells to prepare CD147-CAR-NK cells, the researchers observed efficient killing effects on susceptible target cells, including SKHep1, Huh7, and HepG2, etc. Furthermore, primary NK cells isolated from different zones of HCC liver tissue and engineered to express CD147-CAR can kill CD147+ HCC cell lines selectively and specifically. This study offers a valuable insight into manufacturing CD147-CAR-NK cells as either an autologous or an allogeneic off-the-shelf cell-based product.

Bouattour et al[45] developed c-MET-CAR-NK cells and tested the efficacy against HCC cell lines. c-MET is the product of the proto-oncogene MET and acts as a tumorigen in HCC, since enhanced c-MET activity initiates and contributes to the progression of HCC. The CAR construct contained an scFv of an anti-c-MET antibody fused with truncated human EGFR (referred to as huEGFRt). The intracellular domain of the CAR comprised the 4-1BB and the DAP12 cytoplasmic domain. c-MET-CAR-



encoding lentivirus was then prepared and transduced into human PBMC-derived NK cells. The resultant c-MET-CAR-NK cells remarkably killed HepG2 cells in vitro; however, no data were generated to demonstrate the effect of c-MET-CAR-NK cells in vivo. Of note, c-MET-CAR-NK cells could be potentially used to treat other solid tumors, because the upregulation of c-MET has been found in breast cancer, lung cancer, and colorectal cancer[46-48].

CHALLENGES TO BE ADDRESSED

Although CAR-NK cells have demonstrated their potency and advantages in tumor killing, technical or clinical issues remain to be addressed that will optimize CAR-NKbased immunotherapy against HCC.

First and foremost, stringent screening of HCC-specific antigens is necessary to minimize or avoid severe adverse effects due to on-target/off-tumor toxicity. Ontarget/off-tumor toxicity arises from the simultaneous expression of a target antigen on both tumor and healthy tissues[49]. This toxicity, sometimes lethal, can be reduced by careful design of CAR constructs that improve the recognition of tumor cells. One such approach is to affinity-tune CARs so that they detect tumor cells with a high density of surface antigens and do not react against normal cells that have low antigen densities[50]. Altering the scFv binding domain via mutagenesis or recombination of heavy and light chains can genetically tune the affinity of a CAR to fulfill this purpose [51,52]. The intracellular signaling domain of a CAR can also be tuned to induce a moderate activating signal to weaken CAR-mediated direct cytotoxicity.

Another issue is the promotion of CAR-NK cell expansion and persistence in vivo. Insufficient in vivo expansion of CAR-NK cells after adoptive transfer owing to unidentified factors is a particularly serious challenge. A recent clinical trial has demonstrated that incorporating IL-15 into CAR-NK cells remarkably augmented in vivo expansion[53].

The third issue involves the effect of CAR-NK cells on solid tumors like HCC, which remains unsatisfactory. The heterogeneity of solid tumors, the harsh tumor microenvironment that induces NK cell exhaustion, the limited infiltration from the bloodstream into tumor sites, and the immunosuppression caused by immunosuppressive cells and metabolic disturbances comprehensively inhibit CAR-NK cell function[54]. More efforts have to be made to deal with these challenges, one-by-one, to build more efficient CAR-NK cells against HCC and other solid tumors. With the advance of genetic engineering science, CAR-NK cells equipped with geneticallyincorporated cytokines, antibodies, survival factors, constitutively-active signaling molecules, etc. will greatly promote NK cell proliferation, persistence, migration, and penetration into HCC.

CONCLUSION

CAR-NK cell therapy is an emerging immunotherapy against tumors, including HCC. Its advantage in manufacturing off-the-shelf cellular therapy products with high clinical availability and safety makes it a promising anti-HCC approach. Recent breakthroughs in genome editing techniques have potentiated the production of novel CAR-NK cells with high anti-HCC specificity and activity and low on-target/offtumor toxicity. Innovative engineering tools including CRISPR-Cas9, zinc finger nucleases (referred to as ZFNs), transcription activator-like effector nucleases (referred to as TALENs), and meganucleases are expected to revolutionize the design and creation of CAR constructs for NK cells in the future. With the great efforts being made to enhance safety and activity through laboratory studies and clinical trials, we are confident in the eventual overcoming of the remaining challenges to CAR-NK cell therapy, bringing hope to HCC patients and their families.

REFERENCES

- Johnston MP, Khakoo SI. Immunotherapy for hepatocellular carcinoma: Current and future. World J Gastroenterol 2019; 25: 2977-2989 [PMID: 31293335 DOI: 10.3748/wjg.v25.i24.2977]
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer 2 Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185



Countries. CA Cancer J Clin 2021; 71: 209-249 [PMID: 33538338 DOI: 10.3322/caac.21660]

- Janevska D, Chaloska-Ivanova V, Janevski V. Hepatocellular Carcinoma: Risk Factors, Diagnosis 3 and Treatment. Open Access Maced J Med Sci 2015; 3: 732-736 [PMID: 27275318 DOI: 10.3889/oamjms.2015.111]
- 4 Shimasaki N, Jain A, Campana D. NK cells for cancer immunotherapy. Nat Rev Drug Discov 2020; 19: 200-218 [PMID: 31907401 DOI: 10.1038/s41573-019-0052-1]
- Stabile H, Fionda C, Gismondi A, Santoni A. Role of Distinct Natural Killer Cell Subsets in 5 Anticancer Response. Front Immunol 2017; 8: 293 [PMID: 28360915 DOI: 10.3389/fimmu.2017.00293]
- 6 Basar R, Daher M, Rezvani K. Next-generation cell therapies: the emerging role of CAR-NK cells. Blood Adv 2020; 4: 5868-5876 [PMID: 33232480 DOI: 10.1182/bloodadvances.2020002547]
- Halim L, Maher J. CAR T-cell immunotherapy of B-cell malignancy: the story so far. Ther Adv 7 Vaccines Immunother 2020; 8: 2515135520927164 [PMID: 32524070 DOI: 10.1177/2515135520927164]
- Xie G, Dong H, Liang Y, Ham JD, Rizwan R, Chen J. CAR-NK cells: A promising cellular 8 immunotherapy for cancer. EBioMedicine 2020; 59: 102975 [PMID: 32853984 DOI: 10.1016/j.ebiom.2020.102975
- 9 June CH, O'Connor RS, Kawalekar OU, Ghassemi S, Milone MC. CAR T cell immunotherapy for human cancer. Science 2018; 359: 1361-1365 [PMID: 29567707 DOI: 10.1126/science.aar6711]
- 10 Tokarew N, Ogonek J, Endres S, von Bergwelt-Baildon M, Kobold S. Teaching an old dog new tricks: next-generation CAR T cells. Br J Cancer 2019; 120: 26-37 [PMID: 30413825 DOI: 10.1038/s41416-018-0325-1
- 11 Stoiber S, Cadilha BL, Benmebarek MR, Lesch S, Endres S, Kobold S. Limitations in the Design of Chimeric Antigen Receptors for Cancer Therapy. Cells 2019; 8 [PMID: 31108883 DOI: 10.3390/cells8050472]
- 12 Brudno JN, Kochenderfer JN. Recent advances in CAR T-cell toxicity: Mechanisms, manifestations and management. Blood Rev 2019; 34: 45-55 [PMID: 30528964 DOI: 10.1016/j.blre.2018.11.002]
- Boissel L, Betancur-Boissel M, Lu W, Krause DS, Van Etten RA, Wels WS, Klingemann H. 13 Retargeting NK-92 cells by means of CD19- and CD20-specific chimeric antigen receptors compares favorably with antibody-dependent cellular cytotoxicity. Oncoimmunology 2013; 2: e26527 [PMID: 24404423 DOI: 10.4161/onci.26527]
- 14 Zhang C, Burger MC, Jennewein L, Genßler S, Schönfeld K, Zeiner P, Hattingen E, Harter PN, Mittelbronn M, Tonn T, Steinbach JP, Wels WS. ErbB2/HER2-Specific NK Cells for Targeted Therapy of Glioblastoma. J Natl Cancer Inst 2016; 108 [PMID: 26640245 DOI: 10.1093/jnci/djv375]
- Esser R, Müller T, Stefes D, Kloess S, Seidel D, Gillies SD, Aperlo-Iffland C, Huston JS, Uherek C, 15 Schönfeld K, Tonn T, Huebener N, Lode HN, Koehl U, Wels WS. NK cells engineered to express a GD2 -specific antigen receptor display built-in ADCC-like activity against tumour cells of neuroectodermal origin. J Cell Mol Med 2012; 16: 569-581 [PMID: 21595822 DOI: 10.1111/j.1582-4934.2011.01343.x]
- 16 Jiang H, Zhang W, Shang P, Zhang H, Fu W, Ye F, Zeng T, Huang H, Zhang X, Sun W, Man-Yuen Sze D, Yi Q, Hou J. Transfection of chimeric anti-CD138 gene enhances natural killer cell activation and killing of multiple myeloma cells. Mol Oncol 2014; 8: 297-310 [PMID: 24388357 DOI: 10.1016/j.molonc.2013.12.001
- Pinz KG, Yakaboski E, Jares A, Liu H, Firor AE, Chen KH, Wada M, Salman H, Tse W, Hagag N, 17 Lan F, Leung EL, Jiang X, Ma Y. Targeting T-cell malignancies using anti-CD4 CAR NK-92 cells. Oncotarget 2017; 8: 112783-112796 [PMID: 29348865 DOI: 10.18632/oncotarget.22626]
- 18 Li Y, Hermanson DL, Moriarity BS, Kaufman DS. Human iPSC-Derived Natural Killer Cells Engineered with Chimeric Antigen Receptors Enhance Anti-tumor Activity. Cell Stem Cell 2018; 23: 181-192.e5 [PMID: 30082067 DOI: 10.1016/j.stem.2018.06.002]
- Zhang P, Zhao S, Wu C, Li J, Li Z, Wen C, Hu S, An G, Meng H, Zhang X, Yang L. Effects of 19 CSF1R-targeted chimeric antigen receptor-modified NK92MI & T cells on tumor-associated macrophages. Immunotherapy 2018; 10: 935-949 [PMID: 30149762 DOI: 10.2217/imt-2018-0012]
- 20 Parihar R, Rivas C, Huynh M, Omer B, Lapteva N, Metelitsa LS, Gottschalk SM, Rooney CM. NK Cells Expressing a Chimeric Activating Receptor Eliminate MDSCs and Rescue Impaired CAR-T Cell Activity against Solid Tumors. Cancer Immunol Res 2019; 7: 363-375 [PMID: 30651290 DOI: 10.1158/2326-6066.CIR-18-0572
- 21 Klapdor R, Wang S, Morgan M, Dörk T, Hacker U, Hillemanns P, Büning H, Schambach A. Characterization of a Novel Third-Generation Anti-CD24-CAR against Ovarian Cancer. Int J Mol Sci 2019; 20 [PMID: 30717444 DOI: 10.3390/ijms20030660]
- 22 Zhao Y, Zhou X. Engineering chimeric antigen receptor-natural killer cells for cancer immunotherapy. Immunotherapy 2020; 12: 653-664 [PMID: 32436428 DOI: 10.2217/imt-2019-0139]
- 23 Guedan S, Posey AD Jr, Shaw C, Wing A, Da T, Patel PR, McGettigan SE, Casado-Medrano V, Kawalekar OU, Uribe-Herranz M, Song D, Melenhorst JJ, Lacey SF, Scholler J, Keith B, Young RM, June CH. Enhancing CAR T cell persistence through ICOS and 4-1BB costimulation. JCI Insight 2018; 3 [PMID: 29321369 DOI: 10.1172/jci.insight.96976]
- 24 Lanier LL. Up on the tightrope: natural killer cell activation and inhibition. Nat Immunol 2008; 9: 495-502 [PMID: 18425106 DOI: 10.1038/ni1581]
- Tassi I, Klesney-Tait J, Colonna M. Dissecting natural killer cell activation pathways through 25 analysis of genetic mutations in human and mouse. Immunol Rev 2006; 214: 92-105 [PMID:



17100878 DOI: 10.1111/j.1600-065X.2006.00463.x]

- Vivier E, Nunès JA, Vély F. Natural killer cell signaling pathways. Science 2004; 306: 1517-1519 26 [PMID: 15567854 DOI: 10.1126/science.1103478]
- 27 Hermanson DL, Kaufman DS. Utilizing chimeric antigen receptors to direct natural killer cell activity. Front Immunol 2015; 6: 195 [PMID: 25972867 DOI: 10.3389/fimmu.2015.00195]
- 28 Müller N, Michen S, Tietze S, Töpfer K, Schulte A, Lamszus K, Schmitz M, Schackert G, Pastan I, Temme A. Engineering NK Cells Modified With an EGFRvIII-specific Chimeric Antigen Receptor to Overexpress CXCR4 Improves Immunotherapy of CXCL12/SDF-1a-secreting Glioblastoma. J Immunother 2015; 38: 197-210 [PMID: 25962108 DOI: 10.1097/CJI.00000000000082]
- 29 Zhang J, Sun R, Wei H, Zhang J, Tian Z. Characterization of interleukin-15 gene-modified human natural killer cells: implications for adoptive cellular immunotherapy. Haematologica 2004; 89: 338-347 [PMID: 15020274]
- 30 Sarvaria A, Jawdat D, Madrigal JA, Saudemont A. Umbilical Cord Blood Natural Killer Cells, Their Characteristics, and Potential Clinical Applications. Front Immunol 2017; 8: 329 [PMID: 28386260 DOI: 10.3389/fimmu.2017.00329]
- Luevano M, Madrigal A, Saudemont A. Generation of natural killer cells from hematopoietic stem 31 cells in vitro for immunotherapy. Cell Mol Immunol 2012; 9: 310-320 [PMID: 22705914 DOI: 10.1038/cmi.2012.17]
- 32 Cichocki F, Bjordahl R, Gaidarova S, Mahmood S, Abujarour R, Wang H, Tuininga K, Felices M, Davis ZB, Bendzick L, Clarke R, Stokely L, Rogers P, Ge M, Robinson M, Rezner B, Robbins DL, Lee TT, Kaufman DS, Blazar BR, Valamehr B, Miller JS. iPSC-derived NK cells maintain high cytotoxicity and enhance in vivo tumor control in concert with T cells and anti-PD-1 therapy. Sci Transl Med 2020; 12 [PMID: 33148626 DOI: 10.1126/scitranslmed.aaz5618]
- Harrer DC, Dörrie J, Schaft N. Chimeric Antigen Receptors in Different Cell Types: New Vehicles 33 Join the Race. Hum Gene Ther 2018; 29: 547-558 [PMID: 29320890 DOI: 10.1089/hum.2017.236]
- Patel S, Burga RA, Powell AB, Chorvinsky EA, Hoq N, McCormack SE, Van Pelt SN, Hanley PJ, 34 Cruz CRY. Bevond CAR T Cells: Other Cell-Based Immunotherapeutic Strategies Against Cancer. Front Oncol 2019; 9: 196 [PMID: 31024832 DOI: 10.3389/fonc.2019.00196]
- 35 Guo J, Tang Q. Recent updates on chimeric antigen receptor T cell therapy for hepatocellular carcinoma. Cancer Gene Ther 2021 [PMID: 33500535 DOI: 10.1038/s41417-020-00259-4]
- 36 Mizukoshi E, Kaneko S. Immune cell therapy for hepatocellular carcinoma. J Hematol Oncol 2019; 12: 52 [PMID: 31142330 DOI: 10.1186/s13045-019-0742-5]
- 37 Guo M, Zhang H, Zheng J, Liu Y. Glypican-3: A New Target for Diagnosis and Treatment of Hepatocellular Carcinoma. J Cancer 2020; 11: 2008-2021 [PMID: 32127929 DOI: 10.7150/jca.39972]
- 38 Chen C, Li K, Jiang H, Song F, Gao H, Pan X, Shi B, Bi Y, Wang H, Li Z. Development of T cells carrying two complementary chimeric antigen receptors against glypican-3 and asialoglycoprotein receptor 1 for the treatment of hepatocellular carcinoma. Cancer Immunol Immunother 2017; 66: 475-489 [PMID: 28035433 DOI: 10.1007/s00262-016-1949-8]
- Yu M, Luo H, Fan M, Wu X, Shi B, Di S, Liu Y, Pan Z, Jiang H, Li Z. Development of GPC3-Specific Chimeric Antigen Receptor-Engineered Natural Killer Cells for the Treatment of Hepatocellular Carcinoma. Mol Ther 2018; 26: 366-378 [PMID: 29339014 DOI: 10.1016/i.vmthe.2017.12.012
- Huang Y, Zeng J, Liu T, Xu Q, Song X. DNAM1 and 2B4 Costimulatory Domains Enhance the Cytotoxicity of Anti-GPC3 Chimeric Antigen Receptor-Modified Natural Killer Cells Against Hepatocellular Cancer Cells in vitro. Cancer Manag Res 2020; 12: 3247-3255 [PMID: 32440221 DOI: 10.2147/CMAR.S253565]
- 41 Sanchez-Correa B, Valhondo I, Hassouneh F, Lopez-Sejas N, Pera A, Bergua JM, Arcos MJ, Bañas H, Casas-Avilés I, Durán E, Alonso C, Solana R, Tarazona R. DNAM-1 and the TIGIT/PVRIG/TACTILE Axis: Novel Immune Checkpoints for Natural Killer Cell-Based Cancer Immunotherapy. Cancers (Basel) 2019; 11 [PMID: 31234588 DOI: 10.3390/cancers11060877]
- 42 Shibuya A, Campbell D, Hannum C, Yssel H, Franz-Bacon K, McClanahan T, Kitamura T, Nicholl J, Sutherland GR, Lanier LL, Phillips JH. DNAM-1, a novel adhesion molecule involved in the cytolytic function of T lymphocytes. Immunity 1996; 4: 573-581 [PMID: 8673704 DOI: 10.1016/s1074-7613(00)70060-4]
- Agresta L, Hoebe KHN, Janssen EM. The Emerging Role of CD244 Signaling in Immune Cells of 43 the Tumor Microenvironment. Front Immunol 2018; 9: 2809 [PMID: 30546369 DOI: 10.3389/fimmu.2018.02809]
- Tseng HC, Xiong W, Badeti S, Yang Y, Ma M, Liu T, Ramos CA, Dotti G, Fritzky L, Jiang JG, Yi Q, Guarrera J, Zong WX, Liu C, Liu D. Efficacy of anti-CD147 chimeric antigen receptors targeting hepatocellular carcinoma. Nat Commun 2020; 11: 4810 [PMID: 32968061 DOI: 10.1038/s41467-020-18444-2
- Bouattour M, Raymond E, Qin S, Cheng AL, Stammberger U, Locatelli G, Faivre S. Recent 45 developments of c-Met as a therapeutic target in hepatocellular carcinoma. *Hepatology* 2018; 67: 1132-1149 [PMID: 28862760 DOI: 10.1002/hep.29496]
- 46 Yan S. Jiao X. Zou H. Li K. Prognostic significance of c-Met in breast cancer: a meta-analysis of 6010 cases. Diagn Pathol 2015; 10: 62 [PMID: 26047809 DOI: 10.1186/s13000-015-0296-y]
- 47 Pyo JS, Kang G, Cho WJ, Choi SB. Clinicopathological significance and concordance analysis of c-MET immunohistochemistry in non-small cell lung cancers: A meta-analysis. Pathol Res Pract 2016;



212: 710-716 [PMID: 27465837 DOI: 10.1016/j.prp.2016.05.006]

- Liu Y, Yu XF, Zou J, Luo ZH. Prognostic value of c-Met in colorectal cancer: a meta-analysis. World 48 J Gastroenterol 2015; 21: 3706-3710 [PMID: 25834339 DOI: 10.3748/wjg.v21.i12.3706]
- 49 Castellarin M, Sands C, Da T, Scholler J, Graham K, Buza E, Fraietta JA, Zhao Y, June CH. A rational mouse model to detect on-target, off-tumor CAR T cell toxicity. JCl Insight 2020; 5 [PMID: 32544101 DOI: 10.1172/jci.insight.136012]
- 50 Zhao Y, Wang QJ, Yang S, Kochenderfer JN, Zheng Z, Zhong X, Sadelain M, Eshhar Z, Rosenberg SA, Morgan RA. A herceptin-based chimeric antigen receptor with modified signaling domains leads to enhanced survival of transduced T lymphocytes and antitumor activity. J Immunol 2009; 183: 5563-5574 [PMID: 19843940 DOI: 10.4049/jimmunol.0900447]
- Carter P, Presta L, Gorman CM, Ridgway JB, Henner D, Wong WL, Rowland AM, Kotts C, Carver 51 ME, Shepard HM. Humanization of an anti-p185HER2 antibody for human cancer therapy. Proc Natl Acad Sci US A 1992; 89: 4285-4289 [PMID: 1350088 DOI: 10.1073/pnas.89.10.4285]
- 52 Drent E, Themeli M, Poels R, de Jong-Korlaar R, Yuan H, de Bruijn J, Martens ACM, Zweegman S, van de Donk NWCJ, Groen RWJ, Lokhorst HM, Mutis T. A Rational Strategy for Reducing On-Target Off-Tumor Effects of CD38-Chimeric Antigen Receptors by Affinity Optimization. Mol Ther 2017; 25: 1946-1958 [PMID: 28506593 DOI: 10.1016/j.ymthe.2017.04.024]
- 53 Liu E, Marin D, Banerjee P, Macapinlac HA, Thompson P, Basar R, Nassif Kerbauy L, Overman B, Thall P, Kaplan M, Nandivada V, Kaur I, Nunez Cortes A, Cao K, Daher M, Hosing C, Cohen EN, Kebriaei P, Mehta R, Neelapu S, Nieto Y, Wang M, Wierda W, Keating M, Champlin R, Shpall EJ, Rezvani K. Use of CAR-Transduced Natural Killer Cells in CD19-Positive Lymphoid Tumors. N Engl J Med 2020; 382: 545-553 [PMID: 32023374 DOI: 10.1056/NEJMoa1910607]
- 54 Yilmaz A, Cui H, Caligiuri MA, Yu J. Chimeric antigen receptor-engineered natural killer cells for cancer immunotherapy. J Hematol Oncol 2020; 13: 168 [PMID: 33287875 DOI: 10.1186/s13045-020-00998-9]



0 WJ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2038-2049

DOI: 10.4251/wjgo.v13.i12.2038

ISSN 1948-5204 (online)

MINIREVIEWS

Current status of first-line therapy, anti-angiogenic therapy and its combinations of other agents for unresectable hepatocellular carcinoma

Saleh A Alqahtani, Massimo G Colombo

ORCID number: Saleh A Alqahtani 0000-0003-2017-3526; Massimo G Colombo 0000-0001-8295-7508.

Author contributions: Both the authors contributed equally in conception, literature review, drafting, editing, revising, finalizing, and submitting the manuscript to the journal.

Conflict-of-interest statement: Authors declare no conflict of

interests for this article.

Country/Territory of origin: Italy

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): A Grade B (Very good): B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution

Saleh A Alqahtani, Division of Gastroenterology and Hepatology, Johns Hopkins University, Baltimore, MD 21287, United States

Saleh A Alqahtani, Liver Transplant Center, and Biostatistics, Epidemiology, and Scientific Computing Department, King Faisal Specialist Hospital & Research Center, Riyadh 11564, Saudi Arabia

Massimo G Colombo, Liver Center, IRCCS San Raffaele Hospital, Milan 20132, Italy

Corresponding author: Massimo G Colombo, MD, Professor, Liver Center, IRCCS San Raffaele Hospital, Via Olgettina 60, Milan 20132, Italy. mcolombo46@yahoo.it

Abstract

Globally, hepatocellular carcinoma (HCC) is a frequently diagnosed malignancy with rapidly increasing incidence and mortality rates. Unfortunately, many of these patients are diagnosed in the advanced stages when locoregional treatments are not appropriate. Before 2008, no effective drug treatments existed to prolong survival, until the breakthrough multi-tyrosine kinase inhibitor (TKI) sorafenib was developed. It remained the standard treatment option for advanced HCC for 10 years, with a battery of other candidate drugs in clinical trials failing to produce similar efficacy results. In 2018, the REFLECT trial introduced another multi-TKI, lenvatinib, which has non-inferior overall survival compared with sorafenib. Thus, offering patients and their treating physicians two effective treatment options. Recently, immunotherapy-based drugs, such as atezolizumab and bevacizumab, have shown promising results in patients with unresectable HCC. This review summarizes clinical trial and real-world data studies of sorafenib and lenvatinib in patients with unresectable HCC. We offer guidance on the optimal choice between the two treatments and discuss the potential of immunotherapy-based combination; when more data become available, this will likely make the choice between sorafenib and lenvatinib somewhat obsolete.

Key Words: Hepatocellular carcinoma; Immune checkpoint inhibitor; Lenvatinib; Multityrosine kinase inhibitor; Sorafenib

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.



NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: February 15, 2021 Peer-review started: February 15, 2021

First decision: March 15, 2021 Revised: March 24, 2021 Accepted: October 15, 2021 Article in press: October 15, 2021 Published online: December 15, 2021

P-Reviewer: Aoki H, Lee BS, Niu ZSS-Editor: Fan JR L-Editor: Filipodia P-Editor: Fan JR



Core Tip: Recently, an immunotherapy-based combination of atezolizumab and bevacizumab was shown to prolong survival compared to sorafenib in unresectable hepatocellular carcinoma patients who did not receive prior therapy. In addition, the combination of lenvatinib and pembrolizumab has yielded promising results in the same patient setting. This review article summarizes the results obtained with sorafenib and lenvatinib in patients with unresectable hepatocellular carcinoma in pivotal clinical trials and real-world studies. We offer guidance on the optimal choice between sorafenib or lenvatinib in an individual patient and discuss the immunotherapy-based combination, which will likely make the choice between sorafenib and lenvatinib somewhat obsolete.

Citation: Alqahtani SA, Colombo MG. Current status of first-line therapy, anti-angiogenic therapy and its combinations of other agents for unresectable hepatocellular carcinoma. World J Gastrointest Oncol 2021; 13(12): 2038-2049

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2038.htm **DOI:** https://dx.doi.org/10.4251/wjgo.v13.i12.2038

INTRODUCTION

Currently, liver cancer ranks as the seventh most common cancer type in the world, being the fourth most common cause of cancer-related death[1]. The vast majority of these liver cancers (approximately 80%) arise from hepatocytes and are referred to as hepatocellular carcinoma (HCC)[2]. Over the last decades, the global incidence of HCC has been increasing, with a 75% incidence increase of newly diagnosed HCC cases from 1990 to 2015[3]. Risk factors for developing HCC consist of viral hepatitis, extreme alcohol intake, and non-alcoholic fatty liver disease (NAFLD)[2]. Furthermore, obesity increases the risk of developing NAFLD, and given the ever-increasing obesity epidemic in many parts of the world, the significance of NAFLD-related HCC is predicted to have profound effects in the coming years[4].

For patients with initial-stage HCC, curative therapy strategies, such as liver resection, liver transplantation, or radiofrequency ablation, can still provide long-term survival[2,5]. However, many HCC patients receive their diagnosis at progressive stages of the disease when locoregional treatment is no longer an option. When not treated, patients with advanced-stage HCC have a very poor prognosis, with a median overall survival (OS) of 9 mo and a 6-mo OS of 56.6% [6]. Until 2008, not a single effective systemic treatment option was available for these patients, an unparalleled situation in oncology. In fact, advanced HCC proved to be notoriously difficult to treat. HCC is not only a very chemo-resistant tumor type, but the constant threat of declining liver function often compromises an effective treatment. Over the last decades, improved insights into the molecular processes that initiate and promote the tumor progression in HCC have facilitated the development of novel molecular treatment modalities that specifically target these disrupted molecular pathways.

In 2008, the multi-tyrosine kinase inhibitor (TKI) sorafenib became the first effective systemic therapy for patients with advanced HCC with a preserved liver function [7]. Following its successful introduction into the treatment paradigm for advanced HCC, several other targeted drugs were tested in this setting. Unfortunately, this resulted in a decade of disappointing phase III randomized controlled trials (RCTs). Until 2017, the oral multi-TKI regorafenib reported increased survival rates in patients who received sorafenib[8]. Shortly thereafter, the multi-TKIs cabozantinib and ramucirumab were also shown to delay the disease progression and prolong the OS of HCC patients progressing on sorafenib [for ramucirumab, this benefit was limited to patients with an increased alpha-fetoprotein (AFP) concentration][9,10]. However, until very recently, not a single clinical trial was able to demonstrate a survival benefit compared to sorafenib in the first-line treatment for patients with progressive HCC. Finally, in 2018, the multi-TKI lenvatinib emerged as a feasible first-line alternative for sorafenib in these patients. In fact, results of the phase III REFLECT trial established that the multi-TKI lenvatinib was non-inferior to sorafenib as a first-line treatment for patients with advanced HCC[11]. Thus, physicians now have a choice between two equally effective multi-TKIs in the first-line treatment of these patients. In this article, we review the clinical trial data and real-world data generated with sorafenib and



lenvatinib, with a particular focus on the differences between both agents that can be used to steer the treatment choice in an individual patient.

In recent years, the growing interest in immune checkpoint inhibition as a new pillar of the cancer treatment paradigm has also spurred the evaluation of these drugs in patients with unresectable HCC. Clinical trials using these immune checkpoint inhibitors (ICIs) in monotherapy demonstrate only a moderate clinical benefit[12,13]. In contrast, RCTs evaluating combinations of a TKI and an ICI have generated more convincing results. In fact, results of the phase III IMbrave150 trial recently highlighted that atezolizumab plus bevacizumab had a significantly prolonged OS compared to sorafenib in the first-line treatment of patients with advanced HCC[14]. Furthermore, new data are emerging for other first-line ICI-TKI combinations (e.g., pembrolizumablenvatinib). This review summarizes the results obtained from clinical trials and realworld studies of sorafenib and lenvatinib in patients with unresectable HCC. We offer guidance on the optimal choice between sorafenib or lenvatinib in an individual patient and discuss the potential of immunotherapy-based combinations, which, with more data, will likely make a choice between sorafenib and lenvatinib somewhat obsolete.

SORAFENIB: THE LONG-STANDING STANDARD

Clinical trial data

Sorafenib is an oral multi-TKI that checks and arrests several tyrosine kinases involved in tumor angiogenesis, progression, and apoptosis. It inhibits both vascular endothelial growth factor receptor and platelet-derived growth factor receptor and also targets fms-like tyrosine kinase 3, c-Kit, and several kinases involved in the mitogen activated protein kinase signaling pathway [15]. The application of sorafenib as the standard treatment choice in the first-line therapeutic management of patients with advanced HCC was based on the results of two pivotal phase III RCTs.

In the phase III SHARP trial, 602 (mainly European) patients with advanced HCC, who did not receive prior systemic treatment, were randomly assigned to receive either sorafenib (400 mg twice daily) or placebo. In order to be eligible for the trial, patients had to have an Eastern Cooperative Oncology Group performance status of \geq 2 and have a preserved liver function (Child-Pugh class A)[7]. The median age of patients in the trial was 65 years, more than 80% had Barcelona Clinic Liver Cancer (BCLC) stage C disease, and the vast majority (97%) were rated as Child-Pugh A at baseline. The study reached its primary endpoint by proving a significant OS benefit for sorafenib compared to placebo, with a median OS of 10.7 and 7.9 mo, respectively [hazard ratio (HR) 0.69; 95% confidence interval (CI): 0.55-0.87; *P* < 0.001]. Also, in terms of the time to radiological progression, sorafenib outperformed the placebo (median: 5.5 mo *vs* 2.8 mo; HR 0.58; 95% CI: 0.45-0.74; *P* < 0.001). Objective response rates (ORR) were rare in both arms of the study, with only 2% partial responses with sorafenib compared to 1% with placebo (no complete responses were reported). However, looking at the disease control rate (DCR), a significant benefit was seen for sorafenib compared to placebo (43% vs 32%; P = 0.002)[7]. In the SHARP trial, the incidence of treatment-related adverse events (TRAEs) was reported at 80% with sorafenib compared to 52% with placebo. The most common grade 3/4 TRAEs of sorafenib were diarrhea (8% with sorafenib vs 2% with placebo; P < 0.001) and handfoot skin reactions (8% vs 1%; P < 0.001). Despite the relatively low rate of high-grade TRAEs, the rate of therapy discontinuations due to adverse events (AEs) was high at 38%.

The second pivotal trial with sorafenib in patients with progressive HCC was conducted in the Asia-Pacific region and yielded fairly similar results[16]. In that trial, a total of 226 advanced HCC patients with a Child-Pugh A liver score were randomly assigned (2:1) to receive either sorafenib (400 mg twice daily) or placebo. Sorafenib also showed a significantly prolonged OS compared to placebo, with a median OS of 6.5 and 4.2 mo for sorafenib and placebo, respectively (HR 0.68; 95%CI: 0.50-0.93; P = 0.014). In addition to this, patients who received sorafenib had a significantly longer time to progression (TTP) compared to patients treated with placebo (median 2.8 mo *vs* 1.4 mo; HR 0.57; 95%CI: 0.42-0.79; *P* = 0.0005)[16].

Real-world experience

In the years following the registration of sorafenib for patients with advanced HCC, several studies were set up to evaluate the performance of sorafenib in a real-world setting. In the Italian SOFIA study, 269 advanced HCC patients were treated with



sorafenib (400 mg twice daily), resulting in a median OS of 10.5 mo (8.4 mo for patients with BCLC-C disease and 20.6 mo for BCLC-B patients)[17]. The most common grade 3/4 AEs reported in SOFIA were fatigue (25%), hand-foot skin reactions (9%), hypertension (7%), and diarrhea (6%). Similar to what was reported in SHARP, 40% of patients treated with sorafenib in the SOFIA study had to discontinue therapy due to an AE[17].

The prospective, non-interventional INSIGHT trial also evaluated the safety and efficacy of sorafenib in real-world clinical practice. In this study, including a safety set of 788 HCC patients, the rate of TRAE discontinuations was much lower than in SHARP and SOFIA, as only 15.5% of patients discontinued their therapy because of unacceptable toxicity[18]. A possible explanation for this lower rate could be the fact that this study was reported a decade after the introduction of sorafenib. Thus, it is likely that the increased experience of physicians with this agent and the increased knowledge on how to deal with its toxicity profile ultimately resulted in this lower rate of TRAE discontinuation.

Similarly, a large retrospective study from the United States reported lower rates of TRAE discontinuations with sorafenib than those reported in SHARP. In that study, published in 2017, a total of 3094 advanced HCC patients received sorafenib at the normal dose of 800 mg per day. Of them, only 22.4% had to discontinue their therapy for reasons of toxicity[19]. In an attempt to reduce further the TRAE drop-out, the possibility of introducing sorafenib at a lower dose (< 800 mg/d) was explored, resulting in a decreased pill burden that was less expensive. In addition to this, there were fewer treatment discontinuations due to safety/toxicity concerns (19.6%). With the standard dosing, the median OS reported in this cohort was 233 d (approximately 7-8 mo), which is in line with the SHARP trial[7,19]. Among patients who received a reduced dose of sorafenib, the median OS was shorter at 200 d. However, given the fact that patients who received the reduced dose were generally sicker than patients who were deemed to be eligible for the full dose, this comes as no surprise. When compensating for the differences in patient and disease characteristics between patients in the full and reduced dose cohort (propensity score-matched analysis), no difference in OS was found (HR 0.92; 95%CI: 0.83-1.01)[19].

Among the patients enrolled in these RCTs, many of them had a stable hepatic function with reference to Child-Pugh A disease (SHARP: 95% and Asia-Pacific study: 97%). However, in real-world settings, there are patients with hepatic dysfunction (Child-Pugh B or C), and for these, the RCTs do not provide a clear answer on the potential benefit of sorafenib. In this respect, real-world data can provide guidance to physicians. Not surprisingly, results of a French case-control study (n = 120) indicate that advanced HCC patients with a Child-Pugh A status have a better OS when they are treated with sorafenib than patients who have more advanced liver damage (Child-Pugh B), with a median OS of 13 and 4.5 mo, respectively (P = 0.0008)[20]. A similar observation was seen in the INSIGHT trial, where the median OS with sorafenib was reported as 17.6 mo for patients with a Child-Pugh A status, decreasing to 8.1 and 5.6 mo for patients with Child-Pugh B or C disease, respectively^[18]. Finally, the observational GIDEON registry showed that Child-Pugh A patients have a longer OS when treated with sorafenib than patients with Child-Pugh B disease (median OS: 13.6 and 5.2 mo, respectively)[21]. However, the fact that Child-Pugh B patients do worse on sorafenib than patients with a preserved liver function should not be a reason to reserve sorafenib for patients with Child-Pugh A disease alone. In fact, GIDEON also shows that the overall safety profile and dosing strategy of sorafenib are similar across the different Child-Pugh subgroups^[21]. In another prospective study by Leal *et al*^[22], in a separate prospective score, specifically focusing on the use of sorafenib in Child-Pugh B patients, a median OS of 6.5 mo was reported, which was longer than historical controls for this population. In this study, sorafenib also proved to be tolerable, with a relatively low rate of TRAE discontinuations (27.7%)[22]. As such, these results highlight that selected Child-Pugh B patients may also derive benefit from treatment with sorafenib, with a manageable toxicity profile.

Overall, a large body of real-world data convincingly validate sorafenib as a safe and effective therapy option for patients with advanced HCC and confirm the results obtained in the pivotal RCTs. Furthermore, real-world data have also indicated that given adequate patient selection, sorafenib can also be safe and effective in patients who do not meet the strict inclusion criteria of the SHARP and Asia-Pacific trial.

Zaishideng® WJGO | https://www.wjgnet.com

LENVATINIB: AT LEAST AS GOOD AS SORAFENIB

Since its introduction as a treatment option for patients with advanced HCC, sorafenib has been evaluated against several other targeted agents. However, sunitinib did not prove to be better than sorafenib, and two non-inferiority studies testing brivanib and linifanib against sorafenib turned out to be negative[23]. In addition to this, the phase III SEARCH trial, assessing the potential benefit of adding erlotinib to sorafenib in the first-line treatment of patients with advanced HCC, also failed to show a benefit[24]. In the background of these numerous negative studies, the positive outcome of the phase III REFLECT trial in 2018, showing non-inferiority of lenvatinib to sorafenib as a first-line treatment for patients with unresectable HCC, came somewhat as a surprise[11].

Clinical trial data

Similar to sorafenib, lenvatinib is a multi-TKI. It primarily inhibits the vascular endothelial growth factor receptor 1–3, fibroblast growth factor receptor 1–4, KIT, and RET (Figure 1)[25]. In a single-arm phase II trial, including 46 patients with advanced HCC, lenvatinib at a fixed dose of 12 mg/d was found to have substantial clinical activity. With this regimen, a median OS of 18.7 mo was reported, with 37% of patients obtaining a partial response. However, this came at the cost of considerable toxicity, necessitating a dose reduction and treatment discontinuation in 74% and 22% of patients, respectively[26]. Further in-depth analyses of this trial revealed a close correlation between lenvatinib treatment discontinuation and body weight. Based on this finding, the investigators opted to use a weight-adapted lenvatinib dosing in the subsequent phase III trial.

In the randomized phase III REFLECT trial, a total of 954 patients who did not receive treatment for unresectable HCC were randomly assigned (1:1) to receive either lenvatinib (12 mg/d for patients weighing \geq 60 kg; 8 mg/d for patients weighing < 60 kg) or sorafenib (400 mg twice daily). In order to be eligible for the study, patients had to have a Child-Pugh A liver status and were not allowed to have portal vein invasion at the main portal branch. In addition, patients with a platelet count below 75000 cells/µL were excluded. The study's primary outcome was to demonstrate noninferiority for lenvatinib compared to sorafenib regarding the OS, with a noninferiority margin of 1.08. The median age of the patients enrolled in REFLECT was 62 years; 69% had a body weight of \geq 60 kg, and two-thirds came from Asia-Pacific regions. Extrahepatic spread at baseline was seen in 61% of the patients, while 21% exhibited macroscopic portal vein invasion (macroscopic portal vein invasion and/or extrahepatic spread in 70%). The majority of patients (79%) were classified as having BCLC stage C disease, and 57% had more than one involved disease site. Most of the patient and disease characteristics were well-proportioned between both arms, with some important exceptions. In fact, the number of patients with a hepatitis C etiology was higher in the sorafenib arm than in patients treated with lenvatinib (19% vs 26% for lenvatinib and sorafenib, respectively), while the opposite was true for the proportion of hepatitis B-related HCC (53% vs 48%). Finally, a marked imbalance was seen in the number of patients with an AFP level of $\geq 200 \text{ ng/mL} (46\% vs 39\%)[11]$.

The median OS for patients treated with lenvatinib in the REFLECT trial was reported at 13.6 mo, which was shown to be non-inferior to the 12.3 median OS seen in patients who received sorafenib (HR 0.92; 95%CI: 0.79-1.06). Besides, lenvatinib induced a significant progression-free survival (PFS) than sorafenib. In fact, compared to sorafenib, the median PFS for patients treated with lenvatinib was more than twice as long as the median PFS obtained with sorafenib (7.4 *vs.* 3.7 mo; HR 0.66; 95%CI: 0.57-0.77; *P* < 0.0001). Importantly, lenvatinib was also shown to be associated with a significantly increased rate of ORR compared to sorafenib. With lenvatinib, an ORR of 24.1% was reported, while only 9.2% of sorafenib-treated patients obtained a partial or complete response (odds ratio; OR 3.13; 95%CI: 2.15-4.56; *P* < 0.0001) (Table 1)[11].

With respect to safety, lenvatinib was found to be associated with a slightly increased rate of grade \geq 3 treatment-emergent AEs compared to sorafenib (57% *vs* 49%). This difference was mainly fueled by a higher rate of grade \geq 3 hypertension (23% *vs* 14%) and an increased rate of grade \geq 3 decreased appetite (5% *vs* 1%) and grade \geq 3 weight loss (8% *vs* 3%) among patients treated with lenvatinib. In contrast, lenvatinib was associated with a substantially lower incidence of hand-foot skin reactions (all grade: 27% *vs* 52%; grade \geq 3: 3% *vs* 11%)[11]. The proportion of patients requiring a dose interruption (40% *vs* 32%), dose reduction (37% *vs* 38%), and treatment discontinuation (9% *vs* 7%) was similar in the lenvatinib and sorafenib arms.

WJGO | https://www.wjgnet.com

Table 1 Efficacy outcomes in the phase III REFLECT trial[11]						
	Lenvatinib (<i>n</i> = 478)	Sorafenib (<i>n</i> = 476)	HR (95%CI)	<i>P</i> value		
Median OS	13.6 mo	12.3 mo	0.92 (0.79-1.06)			
Median PFS	7.4 mo	3.7 mo	0.66 (0.57-0.77)	P < 0.0001		
Median TTP	8.9 mo	3.7 mo	0.63 (0.53-0.73)	P < 0.0001		
ORR	24.1%	9.2%	OR (95%CI): 3.13 (2.15-4.56)	P < 0.0001		
DCR	75.5%	60.5%	-			

OS: Overall survival; PFS: Progression-free survival; TTP: Time to progression; ORR: Overall response rate; DCR: Disease control rate; HR: Hazard ratio; OR: Odds ratio; CI: Confidence interval.

Real-world experience

Obviously, as lenvatinib only entered the HCC arena in 2018, the real-world evidence with this agent is more limited compared with sorafenib data. To date, the available real-world data with lenvatinib almost universally originate from Japan.

The largest real-world dataset for lenvatinib reported to date includes data from 105 unresectable HCC patients treated with lenvatinib across 48 clinics in Japan[27]. After excluding patients who started lenvatinib at a reduced dose and with a short observation time, 77 patients were eligible for a response (33 TKI-naïve, 44 TKI-exposed). Both the ORR and the DCR at 4 wk did not differ significantly between TKI-naïve and TKI-pretreated patients (38.5% *vs* 32.4% and 80.8% *vs* 70.8%, respectively). The AE profile of lenvatinib seen in this study was largely in line with what was reported in REFLECT[27].

In a second study, a total of 41 patients with unresectable HCC were treated with lenvatinib. Interestingly, of these patients, 23 (56%) would not have been eligible for the REFLECT trial, mainly because of a prior history of TKI use (n = 16), a Child-Pugh B score (n = 5), and the presence of bile duct invasion (n = 4). In this cohort, lenvatinib was associated with an ORR of 61.0% and a DCR of 90.2%. Overall, 5 patients (12.2%) experienced a complete response to lenvatinib. Interestingly, both the ORR and the DCR did not differ between patients who met the REFLECT criteria or not (P = 0.83) and 0.79, respectively). In patients with a Child-Pugh B score, the ORR was 60% (3/5), while this was 100% in the 4 patients with bile duct invasion. With respect to safety, no major differences were seen between REFLECT eligible and ineligible patients, with a similar rate of grade \geq 3 AEs. Lenvatinib in this cohort caused the following AEs most commonly: Hypertension (68.3%, grade \geq 3 12.2%), appetite loss (68.3%; 2.4%), fatigue (58.5%; 0%), and hand-foot skin reactions (56.1%, 14.6%). As such, these real-world data demonstrate that lenvatinib induces a high early response rate with good tolerability in advanced HCC patients who did and did not meet the REFLECT trial inclusion criteria^[28].

A third Japanese real-world study yielded fairly similar results, in which 57 unresectable HCC patients were treated with lenvatinib, of whom 53 were eligible for response (34 TKI-naïve, 19 TKI-exposed). In this cohort, lenvatinib therapy resulted in an ORR of 49.1% (26/53) and a DCR of 96.2% (51/53). Of note, the ORR was higher in patients receiving lenvatinib in first-line (61.8%) compared to patients receiving a second- (33.3%) or third-line (20.0%) treatment. The median TTP in the entire cohort was reported at 8.5 mo, and also for this endpoint, the outcome was better when lenvatinib was used as a first-line treatment. In addition, this real-world study revealed that patients with a better liver functional reserve had a higher response rate to lenvatinib and a longer TTP. Similar to REFLECT, the most common AEs with lenvatinib were hypertension (54.7%, grade $\geq 3: 15.1\%$), fatigue (49.1%, 7.5%), and a decreased appetite (37.7%, 0%). Hand-foot skin reactions were reported in 26.4% of the patients (all grade 1/2)[29].

In a multi-center retrospective study, including 77 patients with advanced HCC, lenvatinib was associated with an ORR of 29.9% (similar to REFLECT) and a DCR of 77.9%. Interestingly, thyroid dysfunction and appetite loss were found to be associated with a worse and shorter PFS[30]. In this respect, Hiraoka *et al*[31] also identified appetite loss as a dismal prognostic factor for advanced HCC patients treated with lenvatinib[31]. As such, these AEs should be managed with care in patients treated with lenvatinib.

Zaisbideng® WJGO | https://www.wjgnet.com

Randomized control trials Sorafenib	Randomized control trials Lenvatinib		
Sharp trial n = 602 (Europeans) advanced HCC no prior systemic treatment ECOG PS ≥ 2 & Child-Pugh A (97%) Sorafenib (400 mg twice daily) <i>vs</i> placebo OS 10.7 mo <i>vs</i> 7.9 mo TTP 5.5 mo <i>vs</i> 2.8 mo TRAEs 80% <i>vs</i> 52% (diarrhea, hand-foot skin conditions) mo Treatment discount. 38% Asia-Pacific trial n = 226 advanced HCC with Child-Pugh A Sorafenib (400 mg twice daily) <i>vs</i> placebo OS 6.5 mo <i>vs</i> 4.2 mo	Reflect trial <i>n</i> = 954 advanced HCC no prior systemic treatment. Child-Pugh A, no portal vein invasion at main branch, platelet count > 75000 cells/µL Lenvatinib (12 mg/d for ≥ 60 kg weight, or 8 mg/d ≤ 60 kg body weight) or sorafenib (400 mg twice daily) OS 13.6 mo <i>vs</i> 12.3 mo (lenvatinib <i>vs</i> sorafenib, non-inferior) PFS 8.9 mo <i>vs</i> 3.7 mo (lenvatinib <i>vs</i> sorafenib) ORR 24.1% <i>vs</i> 9.2% (lenvatinib <i>vs</i> sorafenib) TRAEs 57% <i>vs</i> 49% (lenvatinib <i>vs</i> sorafenib) Higher hypertension, decreased appetite, weight loss in lenvatinib but less hand-foot skin conditions Treatment discount. 9% <i>vs</i> 7% (lenvatinib <i>vs</i>		
Real-world data	Real-world data		
SOFIA trial n = 296 advanced HCC Sorafenib (400 mg twice daily) OS 10.5 mo (8.4 mo for BCLC-C patients & 20.6 for BCLC-B patients) Grade 3/4 AEs: Fatigue, hand-foot, skin reactions, hypertension, diarrhea. Treatment discount. 40%	Japan trial I n = 105 advanced HCC Total 77 patients included (33 TKI-naïve & 44 TKI-exposed) ORR 38.5% vs 32.4% (TKI-naïve vs TKI- exposed) DCR 80.8% vs 70.8% (TKI-naïve vs TKI- exposed) AEs profile similar to REFLECT		
INSTOLIT trial			
INSIGN that n = 788 advanced HCC Sorafenib (400 mg twice daily) OS 17.6 mo vs 8.1 mo vs 5.6 mo (Child-Pugh A vs B vs C) Grade 3/4 AEs: Fatigue, hand-foot, skin reactions, hypertension, diarrhea Treatment discount. 15.5%	Japan trial II n = 41 advanced HCC with $n = 16$ prior TKI use, n = 5 Child-Pugh B, $n = 4$ bile duct invasion ORR 61.2% DCR 90.2% AEs profile similar to REFLECT Grade 3/4 AEs: Hypertension, appetite loss, fatigue, hand-foot skin reactions		
United Chates total	Janan trial III		
once States trial n = 3094 advanced HCC Sorafenib (400 mg twice daily) then reduced <	n = 53 advanced HCC (34 TKI-naïve, 19 TKI- exposed) ORR 49.1% (higher in first-line than in second- or third-line treatment: 61.8% vs 33.3% vs 20.0% respectively) DCR 96.2% TTP 8.5 mo AEs profile similar to REFLECT and above studies		
Franch trial	Grade 3/4 AEs: Hypertension, appetite loss,		
n = 102 advanced HCC with Child-Pugh A or B Sorafenib (400 mg twice daily) OS 13 mo <i>vs</i> 4.5 mo (Child-Pugh A <i>vs</i> B)	fatigue, hand-foot skin reactions		
GIDEON trial			
n = 3213 (ITT) Sorafenih (400 mg twice daily)			

Figure 1 Evolution of clinical trials and real-world data for sorafenib and lenvatinib. AEs: Adverse events; BCLC: Barcelona Clinic Liver Cancer; DCR: Disease control rate; ECOG: Eastern Cooperative Oncology Group; HCC: Hepatocellular carcinoma; ITT: Intent-to-treat population; ORR: Objective response rate; OS: Overall survival; PFS: Progression-free survival; TKI: Tyrosine-kinase inhibitor; TRAE: Treatment-related adverse event; TTP: Time to progression.

LENVATINIB OR SORAFENIB: HOW TO CHOOSE?

As illustrated above, robust RCT data and convincing real-world results have identified both sorafenib and lenvatinib as effective and safe first-line treatment options for patients with unresectable HCC. This brings us to a logical next question: How should physicians choose between both agents? When making such a decision, several patient and disease characteristics, including liver function and concomitant medication, need to be taken into account. In addition, financial implications need to be considered, especially given the ever-increasing pressure on healthcare budgets.

Baishidena® WJGO | https://www.wjgnet.com

OS 13.6 mo vs 5.2 mo (Child-Pugh A vs B)

The phase III REFLECT trial showed non-inferiority of lenvatinib compared to sorafenib and even demonstrated a numerically longer median OS in lenvatinibtreated patients compared to patients treated with sorafenib (13.6 mo vs 12.3 mo). This difference did not meet the statistical threshold to demonstrate superiority of lenvatinib compared to sorafenib. However, in a critical appraisal of REFLECT, as acknowledged by the authors, this lack of superiority might have been influenced by elements in the study design[32]. First of all, both the baseline AFP level and the presence of macrovascular invasion were not used as a stratification factor for the randomization in REFLECT. As a result, a higher proportion of patients in the lenvatinib arm had macrovascular invasion (23% vs 19%) or an elevated AFP level (≥ 200 ng/mL: 46% vs 39%)[11]. It is likely that this higher incidence of poor prognostic factors in the lenvatinib cohort had an influence on the survival outcome of these patients. Another demographic imbalance that might have influenced the trial outcome relates to the HCC etiology. In fact, a higher proportion of patients in the sorafenib arm had HCC with a hepatitis C etiology compared to the lenvatinib arm (19% vs 26%)[11]. This difference is of clinical importance given the fact that the treatment effect of sorafenib depends on the hepatitis status of patients, with the best OS prospects for patients with hepatitis C virus-positive HCC. A third and final element from REFLECT that might have diluted the OS benefit of lenvatinib is that patients with invasion of the main portal vein and patients with a disease bulk of more than 50% of the liver were excluded from the study. As a result, the trial selected patients who were more likely to be eligible for subsequent therapy after disease progression on the study drug. This hypothesis is confirmed by the high proportion of patients in both the lenvatinib arm (33%) and sorafenib arm (39%) who received some form of post-study anticancer therapy in REFLECT[11]. These subsequent therapies have likely prolonged the post-progression survival of patients in both treatment arms, diluting the potential OS benefit obtained with one of the two agents in the firstline setting. The fact that the median OS obtained with sorafenib in REFLECT was the longest ever reported with sorafenib in a large RCT further supports the idea that poststudy therapies had an important influence on the OS analysis of this trial [7,11,16,23, 24]. As such, several elements of the REFLECT trial design might have mitigated the true OS benefit of patients treated with lenvatinib vs sorafenib. However, it is important to underscore that these statistical speculations should only be seen as hypothesis-generating. This should not be used as an argument to claim a survival superiority of lenvatinib over sorafenib in the first-line treatment of advanced HCC patients.

As such, a critical evaluation of the OS analysis of REFLECT does not help physicians to make a choice between both TKIs in their clinical practice. Perhaps, more practical advice can be derived from the detailed subgroup analysis performed in the trial. In general, the effect of lenvatinib and sorafenib on OS was consistent across all the investigated subgroups. Nevertheless, some subgroups seemed to have a slightly better OS when treated with lenvatinib instead of sorafenib. With respect to HCC etiology, particularly patients with a hepatitis B virus infection seemed to derive a more pronounced OS benefit from lenvatinib compared to sorafenib (median OS: 13.4 mo vs 10.2 mo; HR 0.83; 95% CI: 0.68-1.02). Regional differences were also seen: While patients with a Western origin had a fairly similar median OS with lenvatinib and sorafenib (13.6 mo vs 14.2 mo), patients from Asia-Pacific displayed a numerically longer median OS with lenvatinib (13.5 mo vs 11.0 mo; HR 0.86; 95% CI: 0.72-1.02). Finally, the presence of macroscopic portal vein invasion and/or extrahepatic spread (median OS: 11.5 mo vs 9.8 mo; HR 0.87; 95% CI: 0.73-1.04) and a baseline AFP level \geq 200 ng/mL (median OS: 10.4 mo vs 8.2 mo; HR 0.78; 95%CI: 0.63-0.98) seemed to be associated with a more pronounced treatment effect with lenvatinib[11].

As indicated earlier, lenvatinib was found to be superior to sorafenib in terms of response rate[11]. This finding can be used to make treatment decisions in clinical practice. It is well established that sorafenib mainly induces its survival benefit in patients with advanced HCC by stabilizing the disease. However, in some patients (*e.g.*, patients with bulky disease), a tumor response may be warranted to alleviate symptoms. When faced with such a patient, lenvatinib is probably the better choice.

Both sorafenib and lenvatinib come with a specific toxicity profile, and these differences should be taken into account when opting for one of the two agents. For example, given the high incidence of hypertension reported with lenvatinib, it seems wise to avoid this agent in patients with baseline hypertension or other cardiovascular risk factors.

Finally, the treatment cost should be considered, especially in the context of the ever-increasing pressure on healthcare budgets. In this respect, two independent costeffectiveness analyses demonstrate that lenvatinib is more cost-effective than sorafenib



in the first-line treatment of patients with unresectable HCC[33,34].

As of now, no standard therapies are available for patients who encountered lenvatinib failure. Sorafenib can be considered in such cases, given that about one-fourth of patients in the REFLECT trial received sorafenib while taking lenvatinib as the first-line medication[35]. Recently, a Japanese pilot study has suggested the potential therapeutic benefit from ramucirumab after lenvatinib failure in HCC patients; nevertheless, another study with more patients could not confirm such benefits in the post-progression treatment[36,37].

IS A CHOICE FOR TKI MONOTHERAPY STILL RELEVANT? IMMUNOTHERAPY-BASED COMBINATION THERAPIES AS A NEW STANDARD IN THE FIRST-LINE TREATMENT OF ADVANCED HCC

Since 2008, sorafenib has been the long-standing standard of care in the first-line treatment for patients with unresectable HCC. It took until 2018, with the publication of the REFLECT trial, before an alternative for sorafenib became available. Recently, results presenting the potential benefits of immunotherapy-based combinations may bring in a crucial change in therapeutic strategies for patients with advanced HCC. In the phase III IMbrave 150 trial, atezolizumab anti-PD-L1) plus bevacizumab (antivascular endothelial growth factor) was compared to sorafenib as a first-line treatment of patients with advanced HCC. The atezolizumab-bevacizumab combination showed a significant and clinically meaningful OS improvement compared to sorafenib (median OS not reached vs 13.2 mo; HR 0.58; 95% CI: 0.42-0.79; P = 0.006). At the 12-mo landmark, 67.2% of patients in the combination arm were still alive, 12% more than the 54.6% OS rate seen with sorafenib at 12 mo. Besides, the median PFS was 6.8 mo vs 4.3 mo (atezolizumab-bevacizumab *vs* sorafenib; HR 0.59; 95%CI: 0.47–0.76; *P* < 0.0001). This combination regimen had an expected drug safety profile, with a late deterioration in patients' quality of life[14]. Based on these findings, in May 2020, the United States Food and Drug Administration approved this combination for treating patients with unresectable HCC who had not previously received systemic treatment. In addition, the European Society for Medical Oncology updated its HCC guidelines and endorsed the atezolizumab-bevacizumab combination as a regimen that can be considered a first-line treatment option for advanced HCC patients^[35]. While the groundbreaking outcome of the trial led to the approval of the new treatment in 16 countries, the same has been questioned for its generalizability, owing to the short duration of the trial follow-up and the lack of both safety and efficacy data in Western patients, in whom liver cancer has a different molecular profile than in Asian patients and in patients with metabolic tumors, autoimmune disorders, and transplanted organs. Interesting results were also obtained with a second immunotherapy-based combination consisting of pembrolizumab (anti- programmed death-ligand 1) and lenvatinib. In an open-label phase Ib trial, 104 patients with advanced HCC (BCLC stage B or C, Child-Pugh A, and an Eastern Cooperative Oncology Group performance score of 0-1) were treated with lenvatinib (12 mg/d in patients weighing \geq 60 kg; 8 mg/d in < 60 kg) in combination with pembrolizumab (200 mg intravenously every 3 wk). Among the 100 eligible patients, an impressive ORR of 46% was obtained, which is markedly higher than 24% and 17% ORR obtained with lenvatinib or pembrolizumab monotherapy in the REFLECT and Keynote-224 trials, respectively [11,38,39]. The responses to the lenvatinib-pembrolizumab combination also proved to be durable, with a median response duration of 8.6 mo. The median OS reported with this combination was unprecedented at 22 mo, while patients had a median PFS of 9.3 mo[36]. Based on these findings, the United States Food and Drug Administration granted a breakthrough designation for the use of this combination in the first-line treatment of patients with advanced HCC. However, after the publication of the IMbrave 150 results, this breakthrough designation was put on hold (at that point, pembrolizumab-lenvatinib no longer showed evidence of meaningful improvement over available therapies and, as a result, no longer met the criteria for accelerated approval). Currently, the lenvatinib-pembrolizumab combination is being compared to lenvatinib alone in the randomized phase III LEAP-002 trial, which includes 750 unresectable HCC patients who have not received previous treatment for HCC (NCT03713593). The results of this study are eagerly awaited.

Zaisbidena® WJGO | https://www.wjgnet.com

CONCLUSION

Since 2008, sorafenib has been the undisputed standard of care for patients with unresectable HCC who have not received previous treatment for their advanced disease. It took until 2018 for an alternative drug to emerge. In fact, the publication of the pivotal REFLECT trial demonstrated that lenvatinib is non-inferior to sorafenib in terms of OS in the first-line treatment of patients with unresectable HCC. In addition, lenvatinib was shown to be associated with a higher ORR and significantly longer PFS than sorafenib. This leaves patients and clinicians with two equally effective first-line treatment options for patients with unresectable HCC. For physicians to choose which TKI is best to use, they need to consider the individual patient and disease characteristics and consider the specific toxicity profile of both agents. The recent publication of the IMbrave 150 trial demonstrating the superiority of atezolizumab-bevacizumab combination over sorafenib in this setting will radically change the way we treat this disease type. Additionally, the results with the pembrolizumab-lenvatinib combination are very promising but require further validation in larger, randomized trials. Overall, the results obtained with these immunotherapy-based combination regimens are very convincing and will likely make a choice between sorafenib and lenvatinib in this patient group somewhat obsolete.

REFERENCES

- 1 GLOBOCAN. GLOBOCAN cancer Fact Sheet Liver cancer. [Cited 15 February 2021]. Available from: https://gco.iarc.fr/today/data/factsheets/cancers/11-Liver-fact-sheet.pdf
- Forner A, Reig M, Bruix J. Hepatocellular carcinoma. Lancet 2018; 391: 1301-1314 [PMID: 2 29307467 DOI: 10.1016/S0140-6736(18)30010-2]
- Global Burden of Disease Liver Cancer Collaboration, Akinyemiju T, Abera S, Ahmed M, Alam 3 N, Alemayohu MA, Allen C, Al-Raddadi R, Alvis-Guzman N, Amoako Y, Artaman A, Ayele TA, Barac A, Bensenor I, Berhane A, Bhutta Z, Castillo-Rivas J, Chitheer A, Choi JY, Cowie B, Dandona L, Dandona R, Dey S, Dicker D, Phuc H, Ekwueme DU, Zaki MS, Fischer F, Fürst T, Hancock J, Hay SI, Hotez P, Jee SH, Kasaeian A, Khader Y, Khang YH, Kumar A, Kutz M, Larson H, Lopez A, Lunevicius R, Malekzadeh R, McAlinden C, Meier T, Mendoza W, Mokdad A, Moradi-Lakeh M, Nagel G, Nguyen Q, Nguyen G, Ogbo F, Patton G, Pereira DM, Pourmalek F, Qorbani M, Radfar A, Roshandel G, Salomon JA, Sanabria J, Sartorius B, Satpathy M, Sawhney M, Sepanlou S, Shackelford K, Shore H, Sun J, Mengistu DT, Topór-Mądry R, Tran B, Ukwaja KN, Vlassov V, Vollset SE, Vos T, Wakayo T, Weiderpass E, Werdecker A, Yonemoto N, Younis M, Yu C, Zaidi Z, Zhu L, Murray CJL, Naghavi M, Fitzmaurice C. The Burden of Primary Liver Cancer and Underlying Etiologies From 1990 to 2015 at the Global, Regional, and National Level: Results From the Global Burden of Disease Study 2015. JAMA Oncol 2017; 3: 1683-1691 [PMID: 28983565 DOI: 10.1001/jamaoncol.2017.3055]
- Golabi P, Rhea L, Henry L, Younossi ZM. Hepatocellular carcinoma and non-alcoholic fatty liver 4 disease. Hepatol Int 2019; 13: 688-694 [PMID: 31701393 DOI: 10.1007/s12072-019-09995-8]
- 5 European Association for the Study of the Liver. EASL Clinical Practice Guidelines: Management of hepatocellular carcinoma. J Hepatol 2018; 69: 182-236 [PMID: 29628281 DOI: 10.1016/j.jhep.2018.03.019]
- Giannini EG, Farinati F, Ciccarese F, Pecorelli A, Rapaccini GL, Di Marco M, Benvegnù L, Caturelli E, Zoli M, Borzio F, Chiaramonte M, Trevisani F; Italian Liver Cancer (ITA. LI.CA) group. Prognosis of untreated hepatocellular carcinoma. Hepatology 2015; 61: 184-190 [PMID: 25234419 DOI: 10.1002/hep.27443]
- Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, de Oliveira AC, Santoro A, Raoul 7 JL, Forner A, Schwartz M, Porta C, Zeuzem S, Bolondi L, Greten TF, Galle PR, Seitz JF, Borbath I, Häussinger D, Giannaris T, Shan M, Moscovici M, Voliotis D, Bruix J; SHARP Investigators Study Group. Sorafenib in advanced hepatocellular carcinoma. N Engl J Med 2008; 359: 378-390 [PMID: 18650514 DOI: 10.1056/NEJMoa0708857]
- Bruix J, Qin S, Merle P, Granito A, Huang YH, Bodoky G, Pracht M, Yokosuka O, Rosmorduc O, Breder V, Gerolami R, Masi G, Ross PJ, Song T, Bronowicki JP, Ollivier-Hourmand I, Kudo M, Cheng AL, Llovet JM, Finn RS, LeBerre MA, Baumhauer A, Meinhardt G, Han G; RESORCE Investigators. Regorafenib for patients with hepatocellular carcinoma who progressed on sorafenib treatment (RESORCE): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet 2017; 389: 56-66 [PMID: 27932229 DOI: 10.1016/S0140-6736(16)32453-9]
- 9 Abou-Alfa GK, Meyer T, Cheng AL, El-Khoueiry AB, Rimassa L, Ryoo BY, Cicin I, Merle P, Chen Y, Park JW, Blanc JF, Bolondi L, Klümpen HJ, Chan SL, Zagonel V, Pressiani T, Ryu MH, Venook AP, Hessel C, Borgman-Hagey AE, Schwab G, Kelley RK. Cabozantinib in Patients with Advanced and Progressing Hepatocellular Carcinoma. N Engl J Med 2018; 379: 54-63 [PMID: 29972759 DOI: 10.1056/NEJMoa1717002
- Zhu AX, Kang YK, Yen CJ, Finn RS, Galle PR, Llovet JM, Assenat E, Brandi G, Pracht M, Lim HY,



Rau KM, Motomura K, Ohno I, Merle P, Daniele B, Shin DB, Gerken G, Borg C, Hiriart JB, Okusaka T, Morimoto M, Hsu Y, Abada PB, Kudo M; REACH-2 study investigators. Ramucirumab after sorafenib in patients with advanced hepatocellular carcinoma and increased a-fetoprotein concentrations (REACH-2): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet Oncol 2019; 20: 282-296 [PMID: 30665869 DOI: 10.1016/S1470-2045(18)30937-9]

- 11 Kudo M, Finn RS, Qin S, Han KH, Ikeda K, Piscaglia F, Baron A, Park JW, Han G, Jassem J, Blanc JF, Vogel A, Komov D, Evans TRJ, Lopez C, Dutcus C, Guo M, Saito K, Kraljevic S, Tamai T, Ren M, Cheng AL. Lenvatinib versus sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomised phase 3 non-inferiority trial. Lancet 2018; 391: 1163-1173 [PMID: 29433850 DOI: 10.1016/S0140-6736(18)30207-1]
- 12 Yau T, Park JW, Finn RS, Cheng AL, Mathurin P, Edeline J, Kudo M, Han KH, Harding JJ, Merle P, Rosmorduc O, Wyrwicz L, Schott E, Choo SP, Kelley RK, Begic D, Chen G, Neely J, Anderson J, Sangro B. CheckMate 459: A randomized, multi-center phase III study of nivolumab (NIVO) vs sorafenib (SOR) as first-line (1L) treatment in patients (pts) with advanced hepatocellular carcinoma (aHCC). Annals Oncol 2019; 30: v874-v8745 [DOI: 10.1093/annonc/mdz394.029]
- Finn R, Ryoo B-Y, Merle P, Kudo M, Bouattour M, Lim H-Y, Breder VV, Edeline J, Chao Y, 13 Ogasawara S, Yau T, Garrido M, Chan SL, Knox JJ, Daniele B, Ebbinghaus S, Chen E, Siegel AB, Zhu AX, Cheng AL, and for the KEYNOTE-240 Investigators. Results of KEYNOTE-240: phase 3 study of pembrolizumab (Pembro) vs best supportive care (BSC) for second line therapy in advanced hepatocellular carcinoma (HCC). J Clin Oncol 2019; 37: 15 suppl, 4004-4004 [DOI: 10.1200/JCO.2019.37.15_suppl.4004]
- 14 Finn RS, Qin S, Ikeda M, Galle PR, Ducreux M, Kim TY, Kudo M, Breder V, Merle P, Kaseb AO, Li D, Verret W, Xu DZ, Hernandez S, Liu J, Huang C, Mulla S, Wang Y, Lim HY, Zhu AX, Cheng AL; IMbrave150 Investigators. Atezolizumab plus Bevacizumab in Unresectable Hepatocellular Carcinoma. N Engl J Med 2020; 382: 1894-1905 [PMID: 32402160 DOI: 10.1056/NEJMoa1915745]
- 15 Cervello M, Bachvarov D, Lampiasi N, Cusimano A, Azzolina A, McCubrey JA, Montalto G. Molecular mechanisms of sorafenib action in liver cancer cells. Cell Cycle 2012; 11: 2843-2855 [PMID: 22801548 DOI: 10.4161/cc.21193]
- Cheng AL, Kang YK, Chen Z, Tsao CJ, Qin S, Kim JS, Luo R, Feng J, Ye S, Yang TS, Xu J, Sun Y, 16 Liang H, Liu J, Wang J, Tak WY, Pan H, Burock K, Zou J, Voliotis D, Guan Z. Efficacy and safety of sorafenib in patients in the Asia-Pacific region with advanced hepatocellular carcinoma: a phase III randomised, double-blind, placebo-controlled trial. Lancet Oncol 2009; 10: 25-34 [PMID: 19095497 DOI: 10.1016/S1470-2045(08)70285-71
- Iavarone M, Cabibbo G, Piscaglia F, Zavaglia C, Grieco A, Villa E, Cammà C, Colombo M; SOFIA 17 (SOraFenib Italian Assessment) study group. Field-practice study of sorafenib therapy for hepatocellular carcinoma: a prospective multicenter study in Italy. Hepatology 2011; 54: 2055-2063 [PMID: 21898496 DOI: 10.1002/hep.24644]
- Ganten TM, Stauber RE, Schott E, Malfertheiner P, Buder R, Galle PR, Göhler T, Walther M, 18 Koschny R, Gerken G. Sorafenib in Patients with Hepatocellular Carcinoma-Results of the Observational INSIGHT Study. Clin Cancer Res 2017; 23: 5720-5728 [PMID: 28698202 DOI: 10.1158/1078-0432.CCR-16-0919
- 19 Reiss KA, Yu S, Mamtani R, Mehta R, D'Addeo K, Wileyto EP, Taddei TH, Kaplan DE. Starting Dose of Sorafenib for the Treatment of Hepatocellular Carcinoma: A Retrospective, Multi-Institutional Study. J Clin Oncol 2017; 35: 3575-3581 [PMID: 28872925 DOI: 10.1200/JCO.2017.73.8245
- 20 Hollebecque A, Cattan S, Romano O, Sergent G, Mourad A, Louvet A, Dharancy S, Boleslawski E, Truant S, Pruvot FR, Hebbar M, Ernst O, Mathurin P. Safety and efficacy of sorafenib in hepatocellular carcinoma: the impact of the Child-Pugh score. Aliment Pharmacol Ther 2011: 34: 1193-1201 [PMID: 21958438 DOI: 10.1111/j.1365-2036.2011.04860.x]
- 21 Marrero JA, Kudo M, Venook AP, Ye SL, Bronowicki JP, Chen XP, Dagher L, Furuse J, Geschwind JH, de Guevara LL, Papandreou C, Takayama T, Sanyal AJ, Yoon SK, Nakajima K, Lehr R, Heldner S, Lencioni R. Observational registry of sorafenib use in clinical practice across Child-Pugh subgroups: The GIDEON study. J Hepatol 2016; 65: 1140-1147 [PMID: 27469901 DOI: 10.1016/j.jhep.2016.07.020]
- 22 Leal CRG, Magalhães C, Barbosa D, Aquino D, Carvalho B, Balbi E, Pacheco L, Perez R, de Tarso Pinto P, Setubal S. Survival and tolerance to sorafenib in Child-Pugh B patients with hepatocellular carcinoma: a prospective study. Invest New Drugs 2018; 36: 911-918 [PMID: 29948358 DOI: 10.1007/s10637-018-0621-x
- Cheng AL, Kang YK, Lin DY, Park JW, Kudo M, Qin S, Chung HC, Song X, Xu J, Poggi G, Omata 23 M, Pitman Lowenthal S, Lanzalone S, Yang L, Lechuga MJ, Raymond E. Sunitinib versus sorafenib in advanced hepatocellular cancer: results of a randomized phase III trial. J Clin Oncol 2013; 31: 4067-4075 [PMID: 24081937 DOI: 10.1200/JCO.2012.45.8372]
- 24 Zhu AX, Rosmorduc O, Evans TR, Ross PJ, Santoro A, Carrilho FJ, Bruix J, Qin S, Thuluvath PJ, Llovet JM, Leberre MA, Jensen M, Meinhardt G, Kang YK. SEARCH: a phase III, randomized, double-blind, placebo-controlled trial of sorafenib plus erlotinib in patients with advanced hepatocellular carcinoma. J Clin Oncol 2015; 33: 559-566 [PMID: 25547503 DOI: 10.1200/JCO.2013.53.7746]
- Tohyama O, Matsui J, Kodama K, Hata-Sugi N, Kimura T, Okamoto K, Minoshima Y, Iwata M, 25 Funahashi Y. Antitumor activity of lenvatinib (e7080): an angiogenesis inhibitor that targets multiple



receptor tyrosine kinases in preclinical human thyroid cancer models. *J Thyroid Res* 2014; **2014**: 638747 [PMID: 25295214 DOI: 10.1155/2014/638747]

- 26 Ikeda K, Kudo M, Kawazoe S, Osaki Y, Ikeda M, Okusaka T, Tamai T, Suzuki T, Hisai T, Hayato S, Okita K, Kumada H. Phase 2 study of lenvatinib in patients with advanced hepatocellular carcinoma. J Gastroenterol 2017; 52: 512-519 [PMID: 27704266 DOI: 10.1007/s00535-016-1263-4]
- 27 Hiraoka A, Kumada T, Kariyama K, Takaguchi K, Atsukawa M, Itobayashi E, Tsuji K, Tajiri K, Hirooka M, Shimada N, Shibata H, Ishikawa T, Ochi H, Tada T, Toyoda H, Nouso K, Tsutsui A, Itokawa N, Imai M, Joko K, Hiasa Y, Michitaka K; Real-life Practice Experts for HCC (RELPEC) Study Group, HCC 48 Group (hepatocellular carcinoma experts from 48 clinics in Japan). Clinical features of lenvatinib for unresectable hepatocellular carcinoma in real-world conditions: Multicenter analysis. *Cancer Med* 2019; 8: 137-146 [PMID: 30575325 DOI: 10.1002/cam4.1909]
- Sho T, Suda G, Ogawa K, Kimura M, Shimazaki T, Maehara O, Shigesawa T, Suzuki K, Nakamura A, Ohara M, Umemura M, Kawagishi N, Natsuizaka M, Nakai M, Morikawa K, Furuya K, Baba M, Yamamoto Y, Kobayashi T, Meguro T, Saga A, Miyagishima T, Yokoo H, Kamiyama T, Taketomi A, Sakamoto N. Early response and safety of lenvatinib for patients with advanced hepatocellular carcinoma in a real-world setting. *JGH Open* 2020; **4**: 54-60 [PMID: 32055698 DOI: 10.1002/jgh3.12209]
- 29 Tomonari T, Sato Y, Tanaka H, Tanaka T, Fujino Y, Mitsui Y, Hirao A, Taniguchi T, Okamoto K, Sogabe M, Miyamoto H, Muguruma N, Kagiwada H, Kitazawa M, Fukui K, Horimoto K, Takayama T. Potential use of lenvatinib for patients with unresectable hepatocellular carcinoma including after treatment with sorafenib: Real-world evidence and *in vitro* assessment *via* protein phosphorylation array. *Oncotarget* 2020; 11: 2531-2542 [PMID: 32655838 DOI: 10.18632/oncotarget.27640]
- 30 Ohki T, Sato K, Kondo M, Goto E, Sato T, Kondo Y, Akamatsu M, Sato S, Yoshida H, Koike Y, Obi S. Impact of Adverse Events on the Progression-Free Survival of Patients with Advanced Hepatocellular Carcinoma Treated with Lenvatinib: A Multicenter Retrospective Study. *Drugs Real World Outcomes* 2020; 7: 141-149 [PMID: 32048238 DOI: 10.1007/s40801-020-00179-7]
- 31 Hiraoka A, Kumada T, Atsukawa M, Hirooka M, Tsuji K, Ishikawa T, Takaguchi K, Kariyama K, Itobayashi E, Tajiri K, Shimada N, Shibata H, Ochi H, Tada T, Toyoda H, Nouso K, Tsutsui A, Nagano T, Itokawa N, Hayama K, Imai M, Joko K, Koizumi Y, Hiasa Y, Michitaka K, Kudo M; Real-life Practice Experts for HCC (RELPEC) Study Group, HCC 48 Group (hepatocellular carcinoma experts from 48 clinics in Japan). Prognostic factor of lenvatinib for unresectable hepatocellular carcinoma in real-world conditions-Multicenter analysis. *Cancer Med* 2019; 8: 3719-3728 [PMID: 31127698 DOI: 10.1002/cam4.2241]
- 32 Kudo M. Lenvatinib in Advanced Hepatocellular Carcinoma. *Liver Cancer* 2017; 6: 253-263 [DOI: 10.1159/000479573]
- 33 Kobayashi M, Kudo M, Izumi N, Kaneko S, Azuma M, Copher R, Meier G, Pan J, Ishii M, Ikeda S. Cost-effectiveness analysis of lenvatinib treatment for patients with unresectable hepatocellular carcinoma (uHCC) compared with sorafenib in Japan. *J Gastroenterol* 2019; 54: 558-570 [PMID: 30788569 DOI: 10.1007/s00535-019-01554-0]
- 34 Kim JJ, McFarlane T, Tully S, Wong WWL. Lenvatinib Versus Sorafenib as First-Line Treatment of Unresectable Hepatocellular Carcinoma: A Cost-Utility Analysis. *Oncologist* 2019 [PMID: 31748341 DOI: 10.1634/theoncologist.2019-0501]
- 35 Han KH. Treatment of Hepatocellular Carcinoma With Lenvatinib. *Gastroenterol Hepatol (N Y)* 2018; 14: 662-664 [PMID: 30538608]
- 36 Kuzuya T, Ishigami M, Ito T, Ishizu Y, Honda T, Ishikawa T, Fujishiro M. Initial Experience of Ramucirumab Treatment After Lenvatinib Failure for Patients With Advanced Hepatocellular Carcinoma. Anticancer Res 2020; 40: 2089-2093 [PMID: 32234901 DOI: 10.21873/anticanres.14167]
- 37 Hiraoka A, Kumada T, Tada T, Ogawa C, Tani J, Fukunishi S, Atsukawa M, Hirooka M, Tsuji K, Ishikawa T, Takaguchi K, Kariyama K, Itobayashi E, Tajiri K, Shimada N, Shibata H, Ochi H, Kawata K, Toyoda H, Ohama H, Nouso K, Tsutsui A, Nagano T, Itokawa N, Hayama K, Arai T, Imai M, Koizumi Y, Nakamura S, Michitaka K, Hiasa Y, Kudo M; Real-life Practice Experts for HCC (RELPEC) Study Group and HCC 48 Group (hepatocellular-carcinoma experts from 48 clinics in Japan). Therapeutic efficacy of ramucirumab after lenvatinib for post-progression treatment of unresectable hepatocellular carcinoma. *Gastroenterol Rep (Oxf)* 2021; **9**: 133-138 [PMID: 34026220 DOI: 10.1093/gastro/goaa042]
- 38 Narayan V, Kahlmeyer A, Dahm P, Skoetz N, Risk MC, Bongiorno C, Patel N, Hwang EC, Jung JH, Gartlehner G, Kunath F. Pembrolizumab monotherapy versus chemotherapy for treatment of advanced urothelial carcinoma with disease progression during or following platinum-containing chemotherapy. A Cochrane Rapid Review. *Cochrane Database Syst Rev* 2018; 7: CD012838 [PMID: 30036453 DOI: 10.1002/14651858.CD012838.pub2]
- 39 Zhu AX, Finn RS, Edeline J, Cattan S, Ogasawara S, Palmer D, Verslype C, Zagonel V, Fartoux L, Vogel A, Sarker D, Verset G, Chan SL, Knox J, Daniele B, Webber AL, Ebbinghaus SW, Ma J, Siegel AB, Cheng AL, Kudo M; KEYNOTE-224 investigators. Pembrolizumab in patients with advanced hepatocellular carcinoma previously treated with sorafenib (KEYNOTE-224): a non-randomised, open-label phase 2 trial. *Lancet Oncol* 2018; 19: 940-952 [PMID: 29875066 DOI: 10.1016/S1470-2045(18)30351-6]

Zaishidena® WJGO | https://www.wjgnet.com

G D WJ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2050-2063

DOI: 10.4251/wjgo.v13.i12.2050

ISSN 1948-5204 (online)

MINIREVIEWS

Endoscopic or percutaneous biliary drainage in hilar cholangiocarcinoma: When and how?

Tudor Mocan, Adelina Horhat, Emil Mois, Florin Graur, Cristian Tefas, Rares Craciun, Iuliana Nenu, Mihaela Spârchez, Zeno Sparchez

ORCID number: Tudor Mocan 0000-0001-7785-6403; Adelina Horhat 0000-0002-8701-8750; Emil Mois 0000-0002-2972-3777; Florin Graur 0000-0002-5887-3408; Cristian Tefas 0000-0002-8263-7923; Rares Craciun 0000-0002-5872-8630; Iuliana Nenu 0000-0002-1690-6689; Mihaela Spârchez 0000-0001-8620-9160; Zeno Sparchez 0000-0002-3813-1677.

Author contributions: Mocan T and Sparchez Z contributed study conception and design; Mocan T and Horhat A designed the figures and tables; Mois E, Graur F, Tefas C, Craciun R, Nenu I and Spârchez M contributed draft manuscript preparation; Mocan T and Sparchez Z provided critical comments and coordinated the writing of the paper; all authors reviewed the results and approved the final version of the manuscript.

Conflict-of-interest statement: The authors have nothing to disclose.

Country/Territory of origin: Romania

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer reviewed

Peer-review report's scientific quality classification

Tudor Mocan, Adelina Horhat, Emil Mois, Florin Graur, Cristian Tefas, Rares Craciun, Iuliana Nenu, Zeno Sparchez, Third Medical Department, "Iuliu Hatieganu" University of Medicine and Pharmacy, Cluj-Napoca 400162, Romania

Tudor Mocan, Adelina Horhat, Emil Mois, Florin Graur, Cristian Tefas, Rares Craciun, Iuliana Nenu, Institute for Gastroenterology and Hepatology, Cluj-Napoca 400162, Romania

Mihaela Spârchez, Second Pediatric Department, University of Medicine and Pharmacy, "Iuliu Hatieganu", Cluj-Napoca 400162, Romania

Corresponding author: Adelina Horhat, MBBS, Research Associate, Third Medical Department, "Iuliu Hatieganu" University of Medicine and Pharmacy, Croitorilor, no 21, Cluj-Napoca 400162, Romania. adelinahorhat25@gmail.com

Abstract

Hilar cholangiocarcinoma (hCCA) is a primary liver tumor associated with a dim prognosis. The role of preoperative and palliative biliary drainage has long been debated. The most common techniques are endoscopic retrograde cholangiopancreatography (ERCP) and percutaneous transhepatic biliary drainage (PTBD); however, recently developed endoscopic ultrasound-assisted methods are gaining more atention. Selecting the best available method in any specific scenario is crucial, yet sometimes challenging. Thus, this review aimed to discuss the available techniques, indications, perks, pitfalls, and timing-related issues in the management of hCCA. In a preoperative setting, PTBD appears to have some advantages: low risk of postprocedural complications (namely cholangitis) and better priming for surgery. For palliative purposes, we propose ERCP/PTBD depending on the experience of the operators, but also on other factors: the level of bilirubin (if very high, rather PTBD), length of the stenosis and the presence of cholangitis (PTBD), ERCP failure, or altered biliary anatomy.

Key Words: Hilar cholangiocarcinoma; Endoscopic biliary drainage; Percutaneous biliary drainage; Endoscopic ultrasound biliary drainage; Surgical oncology

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.



WJGO | https://www.wjgnet.com

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: February 28, 2021 Peer-review started: February 28, 2021 First decision: April 19, 2021 Revised: April 28, 2021

Accepted: October 12, 2021 Article in press: October 12, 2021 Published online: December 15, 2021

P-Reviewer: Tonini V S-Editor: Gao CC I-Editor: A P-Editor: Gao CC



Core Tip: Hilar cholangiocarcinoma (hCCA) is a primary tumor of the liver with dim prognosis. The role of biliary drainage in curative and palliative setting has long been debated. Endoscopic retrograde cholangiopancreatography (ERCP) and percutaneous transhepatic biliary drainage (PTBD) as the most commonly used techniques. This review will highlight the available techniques, their indication, advantages or drawbacks, and also timing in the management of hCCA. In a preoperative setting, PTBD appears to win the argument as there is a lower risk of postprocedural complications and better priming for surgery. For palliative purposes, we propose ERCP/PTBD depending on the experience of the operators, biological and anatomy factors, and the presence of cholangitis.

Citation: Mocan T, Horhat A, Mois E, Graur F, Tefas C, Craciun R, Nenu I, Spârchez M, Sparchez Z. Endoscopic or percutaneous biliary drainage in hilar cholangiocarcinoma: When and how? World J Gastrointest Oncol 2021; 13(12): 2050-2063

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2050.htm

DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2050

INTRODUCTION

Cholangiocarcinoma (CCA) is, by definition, a primary tumor of the biliary duct system. Although it is relatively rare compared to the other hepatobiliary tumors, its burden is disproportionately high due to its typically dim prognosis. Per available reports, CCA amounts for approximately 20% of hepatobiliary tumors[1] and is the second most common primary hepatobiliary tumor, accounting for up to 25% of cases in some geographical areas^[2,3]. The current gold standard for treatment is surgical resection, yet only a small portion of the patients are optimal surgery candidates. Moreover, the current standard of care is less than ideal, since the five-year recurrencefree survival for radical resection barely exceeds 33% according to the most optimistic reports[4-6].

Based on its anatomic location, CCA is classified as intra- and extrahepatic, the latter accounting for up to 90% of cases. Extrahepatic CCA is further classified as either hilar CCA (hCCA), accounting for approximately two-thirds of cases and, distal CCA amassing up to 30%[6]. As the focus of our current work, hCCA is located between the emergence of the left and right hepatic ducts and the junction between the common hepatic and the cystic ducts[7].

Due to its origin and characteristics, hCCA obstructs the hepatic bile flow, leading to painless jaundice as the main clinical staple, as it occurs in 90% of cases at diagnosis.

Furthermore, accompanying systemic manifestations such as anorexia, weight loss, and fatigue are commonplace at diagnosis, affecting more than half of the patients and rendering a poor outcome due to their association with advanced or metastatic disease [7].

Depending on initial staging, patients with hCCA are typically dichotomized into the following therapeutic pathways: curative-intent surgery or palliative care.

Either path, however, must cross the same common roadblock - addressing obstructive jaundice and reestablishing adequate bile flow. At this point, the clinician might face multiple dilemmas with regards to the benefit, timing, and method of biliary drainage. The main approaches to biliary decompression are endoscopic and percutaneous. The endoscopic approach most commonly consists of bile duct stenting via endoscopic retrograde cholangiopancreatography (ERCP) and, to a smaller extent, endoscopic ultrasound (EUS)-assisted methods. The percutaneous approach typically consists of ultrasound or radiological-guided transhepatic tube drainage.

In the curative-intent setting, the jury is out on whether pre-operative drainage adds a practical benefit concerning major outcomes. However, the empirical argument appears to be straightforward, as the biliary obstruction is associated with an increased risk of liver failure and cholangitis.

In the palliative setting, the role of biliary drainage can range from allowing a patient to benefit from systemic therapy with an impact on survival, to treating and preventing cholangitis, alleviating symptoms, and, not least, reducing social stigma by resolving jaundice.



The current review aims to chart a course in the field of biliary drainage of hCCA, based on the most recently available data. In the following parts, the discussion will focus on the available techniques, their indications, advantages, potential drawbacks, and timing, to further clarify the role of endoscopic and percutaneous drainage in the therapeutic arsenal of hCCA.

ENDOSCOPIC DRAINAGE

Most patients with obstructive jaundice due to hCCA can be managed non-surgically using ERCP. Although it is less invasive compared to the percutaneous approach, selective endoscopic stenting is technically difficult and can cause severe infectious complications, such as cholangitis. Current guidelines recommend palliative drainage of malignant hilar strictures through ERCP for Bismuth types I and II, and percutaneous transhepatic biliary drainage (PTBD) or a combination of PTBD and ERCP for Bismuth types III and IV[8].

Plastic stents vs self-expanding metal stents

Plastic biliary stents come in various shapes and sizes. They are made either of polyethylene, polyurethane, or Teflon. Their diameter can range from 5F to 12F, while their length ranges between 1 and 18 cm. Furthermore, there are numerous configurations available. Pigtail stents are coiled at one (single pigtail) or both ends (double pigtail), with side holes placed along the curved ends. Flanged stents may be straight, angled, or curved. They have a single flap proximally and distally with a side hole or 4 flaps without side holes[9].

Self-expanding metal stents (SEMS) are made of different metal alloys, such as nickel and titanium. They range from 4 cm to 12 cm in length and 6 mm to 10 mm in diameter when fully expanded. Biliary metallic stents can be fully covered, partially covered, or uncovered, depending on the presence or absence of a polyurethane or silicone layer.

All plastic and metal stents are radiopaque. Most SEMS models have additional proximal and distal markers, with flared ends to prevent migration[9]. Biliary stent placement is performed under radiologic guidance. Regardless of stent type, the first step is to endoscopically locate the papilla, followed by the selective catheterization of the biliary tree. Biliary sphincterotomy is not always mandatory, as stenting without prior sphincterotomy doesn't appear to increase the incidence of post-ERCP pancreatitis[10]. Subsequently, the bile ducts are visualized using a contrast agent, which allows the characterization of the location and extent of the stenosis. The length of the stent must be carefully selected to exceed the proximal end of the stenosis.

For plastic stents, a radiopaque guidewire is placed into the intrahepatic bile ducts. The stent is then advanced over a catheter (which acts as a pusher), which is itself placed over the guidewire. Once the stent has been adequately positioned, the catheter and guidewire are withdrawn, and the stent remains in place.

Metal stents are compressed by an outer, introducer sheath. After the desired position is achieved via the guidewire, the outer sheath is withdrawn, allowing the stent to expand. The guidewire is subsequently removed.

The main goal of endoscopic stenting is to drain at least 50% of the liver, which would reduce bilirubin levels by at least 50% in patients with normal liver function [11]. Secondly, cost-effectiveness must be taken into account, as well as ensuring the stent patency for as long as possible. Given these considerations, plastic stents and SEMS have been compared in several studies and meta-analyses. SEMS are associated with longer patency [odds ratio (OR) 0.16; 95% confidence interval (CI): 0.04-0.62], lower therapeutic failure (OR 0.43; 95% CI: 0.27-0.67) occlusion (OR 0.28; 95% CI: 0.19-0.39) and re-intervention rates (mean difference, -0.49; 95%CI: -0.8 to -0.19)[12-16].

SEMS are more expensive than plastic stents. However, given the higher occlusion rates in plastic stents which impose hospitalization and performing ERCP, SEMS seem to be more cost-effective in the long run[17].

Even if the diameter of fully-expanded SEMS is larger than that of plastic stents, ensuring longer patency, they are thinner when preloaded in their delivery system (5.4-8.5 Fr)[18]. Therefore, they are easier to maneuver and pass through tight strictures than their plastic counterpart. Also, SEMS' delivery system has a sharp tip that acts as a dilatator, facilitating passage through the strictures. Uncovered SEMS also allow the drainage of the biliary tree, side branches, as opposed to plastic stents. This is especially important when performing unilateral drainage in the case of Bismuth type III-IV hilar strictures.

WJGO | https://www.wjgnet.com

Taking these into account, plastic stents are no longer considered standard of care in Bismuth III-IV type CCA, but they can still be used in Bismuth type I-II[8].

Unilateral vs bilateral stenting

Endoscopic drainage in the case of Bismuth type I-II hilar strictures is in many ways similar to distal biliary strictures, namely patients can be fully drained with a single stent, either plastic or SEMS. However, in the case of Bismuth type III-IV malignant hilar strictures, unilateral and bilateral drainage using SEMS have been compared in several randomized control trials (RCT) and meta-analyses. In one meta-analysis that included 683 patients, side by side metal stenting (n = 317) yielded better clinical success rates (CSRs) (OR: 3.56; 95%CI: 1.62-7.82, P = 0.002) and a reduced incidence of stent dysfunction (OR: 1.74; 95%CI: 1.16-2.61, P = 0.007) compared to unilateral stenting (n = 366). Complication rates seemed to be lower in the unilateral group, although they did not reach statistical significance (OR: 0.51; 95%CI: 0.30-1.00, P = 0.05) [19].

In contrast, another recent meta-analysis which included a total of 21 studies with 1292 patients demonstrated that unilateral and bilateral stenting are comparable in terms of efficacy and safety, although technical success was significantly higher in the unilateral group (P = 0.003). One of the limitations of this meta-analysis was that the authors were unable to perform subgroup analysis based on etiology or Bismuth classification[20].

Similarly, a multicenter international study of 187 patients showed that unilateral and bilateral drainage had comparable CSR irrespective of the Bismuth classification, but with a higher incidence of complications and deaths in the bilateral group (11.7% vs 0%, P = 0.007)[21].

Bilateral drainage with SEMS is technically difficult and should be reserved for patients where placing a unilateral stent does not ensure drainage of at least 50% of the liver. Injection of a contrast agent into a liver segment that cannot be subsequently drained can lead to infectious complications such as cholangitis and the formation of liver abscesses, which negatively affect patient survival rates. For this reason, preinterventional hepatobiliary imaging using computed tomography (CT) or magnetic resonance imaging (MRI) with the calculation of liver volume is paramount.

Stent-in-stent vs side-by-side stent placement

If bilateral stenting is chosen, either stent-in-stent (SIS) or stent-by-stent (SBS) drainage can be used, depending on the endoscopist's experience and preference. However, the left lobe, right anterior, or right posterior biliary tree should be selected based on preinterventional CT or MRI imaging to ensure the best drainage. Atrophied liver secondary to longstanding biliary obstruction or portal vein thrombosis should be avoided[22].

In the case of SBS stent placement, after catheterization of the common bile duct (CBD), two guidewires are advanced in the left and right hepatic ducts. The first stent is then advanced on the corresponding guidewire, with the recommendation that it should be placed in the hepatic duct where access is more difficult. The second stent is then advanced parallel to the first on the second guidewire as quickly as possible. The stents must be placed so that their distal ends are at the same level in the CBD or should cross the papilla, which will facilitate subsequent access if revision is necessary.

For SIS deployment, similarly to SBS deployment, two guidewires are inserted into both intrahepatic ducts bilaterally. The first stent is then inserted and deployed into the left or right intrahepatic duct. Subsequently, the guidewire used to deploy the first stent is retracted and passed through the central part of the deployed stent into the contralateral bile duct. The second stent is then advanced and deployed through the central portion of the wire mesh of the first stent. As is the case with SBS stenting, the branch that is more difficult for guidewire insertion should be selected as the first stent placement target.

When performing SIS placement, one thing to consider is that not all SEMS have the same structure. Wire mesh can be either small closed-cell or large open-cell, the latter being easier to dilate as it is weaker in its central part[18]. Hence, using an open-cell SEMS might allow an easier SIS placement.

Regarding their efficacy, one prospective (n = 69) and one retrospective (n = 64)study showed that SIS and SBS deployment seem to be similar in terms of clinical success, stent patency and adverse events[22,23]. In addition, a meta-analysis of four studies which included 158 patients in total revealed no significant differences with respect to the rates of successful placement (P = 0.799), successful drainage (P = 0.617), early complications (P = 0.738), late complications (P = 0.744) and stent occlusions (P = 0.744) 0.606)[24].



Complications of endoscopic stenting

A common complication of stent placement is duodenal biliary reflux, with secondary bacterial colonization of the biliary tract and sludge/stone formation. Another complication is related to stent deployment too far inside the duodenum, or its migration, with subsequent impaction of the stent flanges in the duodenal wall and perforation. If stents migrate in the bowel they can also become stuck, mostly in the ileocecal valve, leading to bowel obstruction.

Plastic stent dysfunction is managed by stent removal and replacement either with another plastic stent or a SEMS. In the case of SEMS dysfunction, if the occlusion is due to debris, this can be removed using balloon catheters. If the occlusion is caused by tissue ingrowth or overgrowth, a secondary plastic stent or SEMS can be inserted inside the existing stent.

IS THERE ANY ROOM FOR EUS-GUIDED BILIARY DRAINAGE IN THE MANAGEMENT OF HCCA?

Although ERCP and PTBD are the two established biliary decompression techniques in the management of hCCA, EUS-guided biliary drainage (EUS-BD) has gained more and more interest in the gastroenterology community^[25]. In theory, EUD-BD seems to provide multiple advantages to the management of hCCA, since it does not require the passage of the biliary stricture. However, in clinical practice, it appears to only be used when ERCP has failed, in surgically altered anatomy, or failed re-interventions for blockage of transpapillary placed stents[26]. Although scarce, the most common adverse effects associated with EUS-BD are bleeding, peritonitis, pneumoperitoneum, cholangitis, bile leak, and stent migration[27]. Moreover, its limitations also reside in the insufficient number of expert endosonographers, typically found only in tertiary referral centers.

The types of EUS-BD performed in malignant hilar obstruction are: (1) EUS-guided hepato-gastrostomy (EUS-HGS); (2) Bridging therapies; and (3) EUS-guided hepaticoduodenostomy (EUS-HDS).

EUS-HGS is one of the most commonly used EUS-BD procedures via an intrahepatic approach[27]. Technically, the EUS-HGS procedure drains only the left hepatic lobe, leaving the right biliary system undrained and thus increases the risk of potential lifethreatening cholangitis.

To address this caveat, Ogura et al[28] developed a novel technique of EUS-BD for right intrahepatic biliary obstruction by adding an uncovered metal stent to the EUS-HGS to bridge the obstruction. In brief, after catheterizing the biliary tract via the stomach and reaching the left liver lobe using a 19-gauge fine-needle aspiration and a guidewire, the needle is replaced by a standard catheter, a guidewire is passed through the hilar stricture and into the right hepatic biliary system. Functional success was reported in all patients and no severe adverse events were noted. Dismally, the bridging method is technically very challenging when passing the guidewire to the right intrahepatic biliary system and requires trained experts.

Last but not least, the right intrahepatic biliary channels could be accessed by EUS-HDS in a similar manner to EUS-HGS, the only variation being the approach via the duodenum. However, since the technique is performed on the lateral side of the duodenal bulb or proximal second duodenum, a long endoscope position might be unstable and risky. For this reason, its use is very limited in the management of hCCA [29,30].

To date, there are several studies carried out to evaluate the use of EUS-BD in clinical practice. Unfortunately, the majority used EUS-BD as a salvation technique (leading to an important selection bias) and the number of patients involved is small [28,31-33]. Nevertheless, there are some advantages of EUS-BD that could make it a more beneficial procedure in the future. The combination of ERCP and EUS-BD (CERES) appears to be more appealing than PTBD in the treatment of Bismuth III-IV CCA. In 2021, Kongkam et al[34] reported a similar technical success rate (TSR), CSR and complications rate (CR) of CERES vs PTBD as follows: TSR = 84.2% (16/19) vs 100% (17/17) (P = 0.23), CSR = 78.9% (15/19) vs 76.5% (13/17) (P = 1), and CR = 26.3 (5/19) vs 35.3 (6/17) (P = 0.56), respectively. Moreover, regarding recurrent biliary obstruction within 3 and 6 mo, authors reported improved results of the CERES procedure[34]. Several retrospective and prospective studies comparing EUS-BD vs PTBD in malignant distal obstruction favor EUS-BD as a better tool for biliary drainage [35,36]. Moreover, a multicenter survey evaluating patient preference for either EUS-BD or PTBD has shown that more than 80% of patients preferred EUS-BD, citing an



increased quality of life without the discomfort of an external drain tube (78.1%), a higher success rate with relatively lower morbidity (43.8%) and the opportunity to be performed at the same time as ERCP (28.3%)[37]. However, no study compared EUS drainage with PTBD in patients with hCCA. While certainly promising, limited experience and low availability diminish the use of EUS-BD. To this point, there is insufficient data to suggest that EUS-BD can replace PTBD as a more efficient biliary drainage tool, with current applicability in large centers with vast EUS experience.

PTBD AND PERCUTANEOUS BILIARY STENTING

PTBD can be performed in two clinical scenarios: (1) Before surgery to relieve biliary obstruction and subsequent cholestasis since an improved survival was documented [38,39]; and (2) As a palliative technique with the ultimate goal to decrease the bilirubin levels to a level that can allow chemotherapy.

Compared to endoscopic drainage, the use of the PTBD method has the advantage that a specific duct can be easily targeted to maximize the drainage of functional parenchyma[40-42]. In high bile duct obstruction, right and left hepatic ducts are typically isolated, with no distal communication. There are three types of isolation: (1) Complete: Cholangiography doesn't result in any opacification; (2) Effective: Isolated ducts are opacified but they do not drain; and (3) Impending isolation: The biliary duct is opacified and drains, but has a central narrowing that is likely to progress to complete isolation. This is important as the latter two increase the risk of subsequent cholangitis.

The first (and probably the most important) steps when assessing a patient for PTBD are: (1) To evaluate the viability of liver parenchyma by high-quality CT or MRI (e.g., drainage of a portion of the liver without an intact portal venous blood supply with ipsilateral duct obstruction will not result in the improvement of liver function; and (2) Pre-procedural antibiotic prophylaxis as cholangitis could result in serious complications that could delay or complicate further management.

Types of PTBD

Currently, there are three modalities of PTBD: (1) External biliary drainage; (2) Internal-external biliary drainage; and (3) Percutaneous self-expanding metallic stent placement.

The point of access is typically chosen depending on the location of the stenosis and the type of intervention. Typically, the right-sided access may be preferred for stent placement in high obstruction, as it offers anatomical continuity between the right hepatic duct and the CBD. When the obstruction is below the duct bifurcation, a left approach is advisable due to a lower risk of catheter displacement. When ascites or segmental isolation of the right duct are present, a left-sided approach may provide more benefit. Peripheral access is preferred because of the lower risk of bleeding and inadequate drainage.

There are two types of approaches: Fluoroscopy-guided PTBD (F-PTBD) and a combined ultrasound-guided approach (US-PTBD). The US-PTBD has more advantages: reduction of fluoroscopy time, faster access to the bile ducts, reduced number of punctures, and, consequently, significantly lowers rates of complications. Moreover, a meta-analysis showed the superiority of US-PTBD vs F-PTBD, as US-PTBD was associated with fewer severe early complications and procedure-related deaths (overall complication rates range from 5%-100% for F-PTBD (median, 21%) and from 0%-22% for US-PTBD (median, 5%)[43,44]. After localizing the best access pathway, the real-time US-guided puncture of the liver parenchyma is performed with a 21gauge Chiba needle. In the case of F-PTBD, after injecting the contrast agent, the targeted bile duct is accessed via fluoroscopy. Afterward, the inner stylet is withdrawn, and a guidewire is inserted through the needle into the collecting system.

Subsequently, progressive 6, 8, and 10 Fr coaxial sheaths are advanced over the guidewire for tract dilation, and ultimately an 8 Fr or 10 Fr biliary drainage catheter is placed. If the obstruction can be passed, the directional catheter is advanced into the small bowel. The catheter can be then exchanged over a stiffer wire for a multisidehole drainage catheter. This allows the bile to drain both externally (into a bag), and internally (into the duodenum) to preserve the normal enterohepatic circulation of bile.

Finally, a percutaneous SEMS is a third option for drainage. Stent placement can be performed as either a one-step (primary stenting technique) or a two-step (secondary stenting technique) procedure; the latter will give the clinician more time to plan and



is particularly useful in case of intraprocedural bleeding. A randomized controlled trial that compared the clinical effectiveness of percutaneous covered Viabil stents vs uncovered metallic Wallstents demonstrated improved survival in patients with hCCA who received a covered (median survival, 243 d) vs uncovered stent (median survival, 180 d) (P = 0.039). The incidence of stent dysfunction was significantly lower in the covered stent group [45].

Post transhepatic percutaneous biliary drainage complications

Virtually, PTBD complications are overlapped to those of endoscopic biliary drainage, hence we won't reiterate the matter. However, in the group of patients with resectable hCCA, a long-term complication that can be a game-changer is represented by tumor seeding. Takahashi and colleagues reported an alarming 5.2 percent catheter tract recurrence after PTBD in patients with hCCA which is much higher than previously reported. However, the duration of PTBD (over 60 d) was an important independent risk factor for tract metastasis and shortened postoperative survival. In the curative surgery setting, this points out the importance of a short delay until surgery to prevent this troublesome complication[46,47].

Combined techniques — "Rendez-vous": how and when?

Rendez-vous (RV) procedure is an appealing option for treating obstructive jaundice in the case of an unsuccessful ERCP[48-50]. Apart from ERCP failure, RV is a rescue therapy for in the case of complex biliary interventions that require combined access routes: patients with surgically altered enteric anatomy, tight hilar biliary stricture passable only by the guidewire, and in patients with a preexistent PTBD that can be easily used as an anterograde route for percutaneous RV[50,51].

Clinical scenarios: Single or dual drainage?

In the case of distal biliary obstruction, a straightforward approach is sufficient. However, in the case of Bismuth type II, III, IV hCCA controversy exists as to whether partial or total biliary drainage is more suitable in a palliative setting. There are two majors advocates for complete^[52] or incomplete^[53] drainage.

Schima et al[54] studied a group of 41 patients with hilar obstruction and compared long-term outcomes. Single stents were placed for unilateral drainage in 27 patients, while 14 patients had bilateral stents. They found no significant difference regarding mean stent patency.

Kaiho *et al*[55] performed either complete (n = 12) or partial (n = 9) drainage in a group of 21 patients with hilar obstruction. There were three, seven, and eleven patients with Bismuth types II, III, and IV obstructions, respectively. They found no difference in stent patency between complete and partial drainage.

Inal *et al*[56] evaluated the necessity of draining more than one hepatic duct in 138 patients with malignant hilar obstruction. Single-duct drainage was achieved in 74 patients (54%) by placing one stent (n = 59), two stents (n = 41) or a single transhepatic tract in a "T" configuration (n = 23). There were no differences between single and dual stenting in Bismuth type I, II, and III hCCA. However, in Bismuth type IV, the deployment of two parallel stents resulted in significantly higher patency rates.

Lee *et al*^[57] suggest that when a repeat procedure in proximal hCCA is necessary, placement of internal/external drainage catheters provides better palliation than putting in new metal stents, as life expectancy is limited in this patient group.

ENDOSCOPIC VS PERCUTANEOUS BILIARY DRAINAGE — PRO AND CONS

After decades of clinical research, it is still unclear whether endoscopic (ERCP) or percutaneous drainage is the preferred method of biliary drainage in patients with hCCA. Both can be performed before surgery (in patients suited for curative treatment) or as a palliative treatment. Choosing one technique over the other is not an easy clinical decision. A straightforward selection can only be made in patients with modified anatomy[58].

However, for the majority of patients with hCCA choosing the best technique depends on several factors.

Biliary drainage before surgery

In one retrospective study, technical success in the ERCP group (n = 87) was 78%



compared to 98% in the PTBD group (n = 42; P = 0.04). The therapeutic success rate was also higher in the PTBD group (79% vs 49%; P = 0.02)[59]. Another retrospective study showed higher technical success for PTBD vs ERCP (100 % vs 81%; P = 0.203) [60]. However, neither technical nor therapeutic success should be the sole primary outcomes when comparing the two methods. There is only one multicentric RCT comparing endoscopic and percutaneous biliary drainage. The primary outcome was the number of severe complications in the timespan between randomization and surgery. In total, 54 patients were randomly assigned to benefit from either PTBD or ERCP. The study was prematurely interrupted due to significantly higher mortality in the PTBD group [11 (41%) of 27 patients] vs the ERCP group [three (11%) of 27 patients; relative risk 3.67, 95%CI 1.15-11.69; P = 0.03][61]. Indeed this study provides the highest level of evidence we have to this point for decision making in clinical practice. However, these data should be interpreted with caution for several reasons: (1) Only one patient with Bismuth type 1 was included in the PTBD group; (2) Although not statistically significant, both technical and therapeutic success were higher in the PTBD group; (3) 55% of the patients in the ERCP group had subsequent PTBD; and (4) Only 54 patients were randomized, making the study prone to a type-I error. Nevertheless, the expertise of centers performing PTBD is highly relevant and could explain these results. In one study[62], low-volume centers showed a higher occurrence of serious complications related to PTBD, whereas high-volume centers showed a similar proportion of complications between endoscopic and percutaneous drainage. In terms of procedure-related complications (e.g., cholangitis and pancreatitis) two other studies found PTBD to be superior to ERCP[63,64]. Another critical aspect is the cost associated with each method. One recent study found ERCP to be more expensive than PTBD (P = 0.005)[63]. Some patients with hCCA are better suited for ERCP drainage, while others might be more appropriate for PTBD. Discriminating between these two categories is crucial. One study showed that patients with Bismuth 3a or 4 hCCA and a total bilirubin level above 8.8 mg/dL should be considered for initial PTBD rather than ERCP[65].

ERCP appears to perform better in Bismuth II hCCA, as it is associated with fewer postprocedural complications, namely cholangitis[66].

Until now, four meta-analyses comparing the two techniques have been published [67-70]. All of them found PTBD to be superior to some extent over ERCP. More data about the meta-analyses is provided in Table 1.

Biliary drainage in palliation

A single meta-analysis comparing ERCP and PTBD in palliation of advanced malignant hilar obstruction has been published to this point. It included a total of nine studies and 546 patients, yet not all of them had hCCA. The results showed that palliation with PTBD was associated with higher rates of successful biliary drainage and lower rates of cholangitis while palliative ERCP had lower bleeding complications [71].

A key aspect in a palliative setting is the patient's quality of life. In this light, the presence of the external drainage tube for the remainder of a patient's life (in the case of PTBD) might generate a significant alteration, especially compared to the placement of an internal stent. Surprisingly, based on a controlled study by Saluja et al[72], quality of life after PTBD was rated higher than ERCP according to the World Health Organization Quality of Life physical and psychological scores at one and three months. A potential cause might be the relatively high incidence rate of fever in the endoscopic biliary stent implementation group. Moreover, percutaneous stenting after PTBD is also possible, eliminating the burden of caring for an external drainage tube for the entire life. One study which included 85 patients with advanced Bismuth type III and IV hCCA showed that percutaneous SEMS was superior to endoscopic stenting regarding successful biliary decompression (92.7% vs 77.3%; P = 0.49)[73]. There is not enough evidence to suggest one technique over the other. Moreover, the implementation of RCTs is problematic. The results of an unsuccessful RCT were recently published. Lack of funding, provider/institutional bias in favor of one procedure, and logistical challenges were cited as possible responsible factors of failure^[74]. Therefore, until high-quality observational data or RCTs become available, one must rely on personal judgment, according to expertise and specific conditions. Based on the aforementioned discussion, we propose an algorithm on when and how to use ERCP and PTBD in patients with hCCA, depicted in Figure 1.

Zaishideng® WJGO | https://www.wjgnet.com

Table 1 Meta-analysis comparing endoscopic vs biliary drainage before surgery in patients with hilar cholangiocarcinoma				
Ref.	No studies	No. patients	Main findings	
Liu <i>et al</i> [68]	6	359-EBD; 286-PTBD	Similar technical success rate, R0 resection, incidence of total complications after resection, post-operative hospitalization time, resection time and recurrence; The incidence of total complications were higher in the EBD group ($P < 0.05$)	
Hameed <i>et</i> al[69]	15	398-EBD; 1036-PTBD	There was a trend towards higher procedure conversion (RR 7.36, $P = 0.07$) and cholangitis (RR 3.36, $P = 0.15$) in the EBD group	
Al Mahjoub et al[70]	4	275-EBD; 158-PTBD	Overall procedure related mortality was higher in EBD group ($P = 0.0009$); Similar initial technical failure; Conversion rate was higher in EBD group ($P < 0.001$); Risk of pancreatitis was higher in EBD group ($P < 0.001$); Risk of cholangitis was higher in EBD group ($P < 0.001$); Similar postoperative morbidity and mortality	
Tang <i>et al</i> [67]	9	498-EBD; 414-PTBD	PTBD was associated with a lower risk of cholangitis ($P < 0.001$); PTBD was associated with a lower risk of pancreatitis ($P = 0.003$); A higher successful rate of palliative relief of cholestasis in PTBD group ($P < 0.001$); The incidence of hemorrhage was similar ($P = 0.59$)	

EBD: Endoscopic biliary drainage; PTBD: Percutaneous biliary drainage; hCCA: Hilar cholangiocarcinoma; RR: Relative risk.



Figure 1 Management algorithm in hilar cholangiocarcinoma. ¹In high-volume centers with expertise in EUS. ²In centers specialized in ERCP and little experience in PTBD. ³Depending on the experience and preference of the patient. ERCP: Endoscopic retrograde cholangio-pancreatography; EUS: Endoscopic ultrasound; PTBD: Percutaneous transhepatic biliary drainage.

THE SURGICAL POINT OF VIEW

Biliary drainage is an established safe hCCA treatment strategy as a bridge therapy before surgery. To date, the surgical standard of treatment in hCCA is complete resection combined surgery. Although there is debate about the effect of biliary drainage on surgical outcomes in patients with hCCA, it has been demonstrated that liver failure caused by obstructive jaundice can be a significant risk factor in major liver resection. This is especially relevant in the case of hCCA, for which extended hepatectomy might be needed to provide the best chance for a cure. Therefore, it is preferable to perform the biliary decompression of the future remnant liver to preserve postoperative liver function[75].

There are two preferred methods for biliary drainage in hCCA: Endoscopic biliary drainage (ERCP) and PTBD. ERCP might be a less invasive technique, but it may come with a price: it carries an increased risk of ascending cholangitis and procedure-related complications, such as duodenal perforation and pancreatitis [59-60,64,67,76]. On the other hand, PTBD could lead to several complications such as bleeding, portal vein thrombosis, tumor seeding, patient discomfort, and has been widely reported to be associated with malpractice [64,77-80].

Several meta-analyses have compared the two methods in hCCA patients[67-70]. All of them showed some superiority of PTBD over ERCP when performed before surgery.

In our center, as surgeons, we always prefer PTBD drainage over endoscopic drainage. From our own experience several other factors must be taken into account: (1) First, the cornerstone of surgical treatment is to obtain an R0 resection, which



WJGO | https://www.wjgnet.com

translates in performing an accurate dissection of the hepatic pedicle and lymph node dissection. This is significantly easier in the absence of inflammation surrounding the main biliary duct (MBD), which might be caused by a prior ERCP[81], resulting in greater intraoperative blood loss and prolonged operative time; (2) In the case of PTBD, inflammation is absent or minimal, which leads to an easier dissection and an accurate lymph node dissection[82]; and (3) The inflammation determined by the stent could give a false appreciation of inoperability^[83]. The inflammation surrounding the MBD can be a blunder in mimicking a direct invasion of the important vascular structures, such as the portal vein or the hepatic artery, which would falsely classify the patient as inoperable. Consequently, with ERCP, we consider that the best timing for surgery is within the first seven days, to avoid MBD inflammation. A concern is that the seven-day timeline could not suffice to obtain a normal liver function, a problem that doesn't exist for PTBD.

CONCLUSION

Endoscopic or percutaneous biliary drainage? Decades of experience, a lot of research, new stents, new techniques but the same disease: hCCA remains one of the most challenging cancers. Biliary drainage, then chemotherapy, stent occlusion/external tube removal then stop chemotherapy and re-drain, and the story goes on. Treating patients according to the proposed algorithm (Figure 1), although based on lowquality data and personal experience and educated guesses might at least help the decision-making process. In the palliative setting, we would choose between ERCP and PTBD generally based on operator experience, as well as other relevant factors: stenosis length, bilirubin level (if very high, rather PTBD), cholangitis (PTBD), ERCP failure, or altered biliary anatomy. Not least, one should always consider patient preference It is not hard to understand that (from the patient perspective) there is only one answer to the question: endoscopic or percutaneous biliary drainage? EUS biliary drainage is a relatively new technique with only a few hCCA patients treated. Yet it is likely to gain more interest in the years to come, hoping to improve the current management of hCCA.

REFERENCES

- Patel N, Benipal B. Incidence of Cholangiocarcinoma in the USA from 2001 to 2015: A US Cancer Statistics Analysis of 50 States. Cureus 2019; 11: e3962 [PMID: 30956914 DOI: 10.7759/cureus.3962]
- Shaib Y, El-Serag HB. The epidemiology of cholangiocarcinoma. Semin Liver Dis 2004; 24: 115-125 2 [PMID: 15192785 DOI: 10.1055/s-2004-828889]
- 3 Tyson GL, El-Serag HB. Risk factors for cholangiocarcinoma. Hepatology 2011; 54: 173-184 [PMID: 21488076 DOI: 10.1002/hep.24351]
- Chen XP, Lau WY, Huang ZY, Zhang ZW, Chen YF, Zhang WG, Qiu FZ. Extent of liver resection 4 for hilar cholangiocarcinoma. Br J Surg 2009; 96: 1167-1175 [PMID: 19705374 DOI: 10.1002/bjs.6618]
- Jonas S, Thelen A, Benckert C, Biskup W, Neumann U, Rudolph B, Lopez-Häänninen E, Neuhaus P. 5 Extended liver resection for intrahepatic cholangiocarcinoma: A comparison of the prognostic accuracy of the fifth and sixth editions of the TNM classification. Ann Surg 2009; 249: 303-309 [PMID: 19212186 DOI: 10.1097/SLA.0b013e318195e164]
- Sapisochin G, Ivanics T, Subramanian V, Doyle M, Heimbach JK, Hong JC. Multidisciplinary treatment for hilar and intrahepatic cholangiocarcinoma: A review of the general principles. Int J Surg 2020; 82S: 77-81 [PMID: 32380231 DOI: 10.1016/j.ijsu.2020.04.067]
- Blechacz B. Cholangiocarcinoma: Current Knowledge and New Developments. Gut Liver 2017; 11: 7 13-26 [PMID: 27928095 DOI: 10.5009/gnl15568]
- Boškoski I, Schepis T, Tringali A, Familiari P, Bove V, Attili F, Landi R, Perri V, Costamagna G. 8 Personalized Endoscopy in Complex Malignant Hilar Biliary Strictures. J Pers Med 2021; 11 [PMID: 33572913 DOI: 10.3390/jpm11020078]
- Almadi MA, Barkun A, Martel M. Plastic vs. Self-Expandable Metal Stents for Palliation in Malignant Biliary Obstruction: A Series of Meta-Analyses. Am J Gastroenterol 2017; 112: 260-273 [PMID: 27845340 DOI: 10.1038/ajg.2016.512]
- 10 Frank CD, Adler DG. Post-ERCP pancreatitis and its prevention. Nat Clin Pract Gastroenterol Hepatol 2006; 3: 680-688 [PMID: 17130878 DOI: 10.1038/ncpgasthep0654]
- Kerdsirichairat T, Arain MA, Attam R, Glessing B, Bakman Y, Amateau SK, Freeman ML. 11 Endoscopic Drainage of >50% of Liver in Malignant Hilar Biliary Obstruction Using Metallic or Fenestrated Plastic Stents. Clin Transl Gastroenterol 2017; 8: e115 [PMID: 28858292 DOI:



10.1038/ctg.2017.42]

- 12 Zhou C, Li H, Huang Q, Wang J, Gao K. Biliary self-expandable metallic stent combined with Iodine-125 seeds strand in the treatment of hilar malignant biliary obstruction. J Int Med Res 2020; 48: 300060519887843 [PMID: 31884851 DOI: 10.1177/0300060519887843]
- 13 Kanno Y, Koshita S, Ogawa T, Kusunose H, Masu K, Sakai T, Yonamine K, Miyamoto K, Murabayashi T, Kozakai F, Horaguchi J, Noda Y, Ito K. Inside Plastic Stents vs Metal Stents for Treating Unresectable Malignant Perihilar Biliary Obstructions: A Retrospective Comparative Study. Clin Endosc 2020; 53: 735-742 [PMID: 32126740 DOI: 10.5946/ce.2020.003]
- 14 Xia MX, Pan YL, Cai XB, Wu J, Gao DJ, Ye X, Wang TT, Hu B. Comparison of endoscopic bilateral metal stent drainage with plastic stents in the palliation of unresectable hilar biliary malignant strictures: Large multicenter study. Dig Endosc 2021; 33: 179-189 [PMID: 32249460 DOI: 10.1111/den.13680]
- Choi JH, Lee SH, You MS, Shin BS, Choi YH, Kang J, Jang S, Paik WH, Ryu JK, Kim YT. Step-15 wise endoscopic approach to palliative bilateral biliary drainage for unresectable advanced malignant hilar obstruction. Sci Rep 2019; 9: 13207 [PMID: 31519930 DOI: 10.1038/s41598-019-48384-x]
- Hong WD, Chen XW, Wu WZ, Zhu QH, Chen XR. Metal vs plastic stents for malignant biliary 16 obstruction: an update meta-analysis. Clin Res Hepatol Gastroenterol 2013; 37: 496-500 [PMID: 23333231 DOI: 10.1016/j.clinre.2012.12.002]
- Roberts AT, Jaya J, Ha P, Thakur U, Aldridge O, Pilgrim CHC, Tan E, Wong E, Fox A, Choi J, Liew 17 D, Le STT, Croagh D. Metal stents are safe and cost-effective for preoperative biliary drainage in resectable pancreaticobiliary tumours. ANZ J Surg 2021; 91: 1841-1846 [PMID: 34309143 DOI: 10.1111/ans.17060
- 18 Lee TH, Moon JH, Park SH. Biliary stenting for hilar malignant biliary obstruction. Dig Endosc 2020; 32: 275-286 [PMID: 31578770 DOI: 10.1111/den.13549]
- 19 Chen ZK, Zhang W, Xu YS, Li Y. Unilateral Versus Side-By-Side Metal Stenting for Malignant Hilar Biliary Obstruction: A Meta-Analysis. J Laparoendosc Adv Surg Tech A 2021; 31: 203-209 [PMID: 32644848 DOI: 10.1089/lap.2020.0400]
- Aghaie Meybodi M, Shakoor D, Nanavati J, Ichkhanian Y, Vosoughi K, Brewer Gutierrez OI, 20 Kalloo AN, Singh V, Kumbhari V, Ngamruengphong S, Khashab MA. Unilateral vs bilateral endoscopic stenting in patients with unresectable malignant hilar obstruction: a systematic review and meta-analysis. Endosc Int Open 2020; 8: E281-E290 [PMID: 32118102 DOI: 10.1055/a-1067-4326]
- 21 Staub J, Siddiqui A, Murphy M, Lam R, Parikh M, Pleskow D, Papachristou G, Sharaiha R, Iqbal U, Loren D, Kowalski T, Noor A, Mumtaz T, Yasuda I, Thomas S, Hsaeeb A, Herrick J, Greene T, Adler DG. Unilateral vs bilateral hilar stents for the treatment of cholangiocarcinoma: a multicenter international study. Ann Gastroenterol 2020; 33: 202-209 [PMID: 32127742 DOI: 10.20524/aog.2020.0451]
- Zhou WZ, Liu S, Yang ZQ, Xian YT, Xu HD, Wu JZ, Shi HB. Percutaneous stent placement for 22 malignant hilar biliary obstruction: side-by-side vs stent-in-stent technique. BMC Gastroenterol 2020; 20: 174 [PMID: 32503426 DOI: 10.1186/s12876-020-01316-w]
- Ishigaki K, Hamada T, Nakai Y, Isayama H, Sato T, Hakuta R, Saito K, Saito T, Takahara N, Mizuno S, Kogure H, Ito Y, Yagioka H, Matsubara S, Akiyama D, Mohri D, Tada M, Koike K. Retrospective Comparative Study of Side-by-Side and Stent-in-Stent Metal Stent Placement for Hilar Malignant Biliary Obstruction. Dig Dis Sci 2020; 65: 3710-3718 [PMID: 32107675 DOI: 10.1007/s10620-020-06155-z
- 24 Hong W, Chen S, Zhu Q, Chen H, Pan J, Huang Q. Bilateral stenting methods for hilar biliary obstructions. Clinics (Sao Paulo) 2014; 69: 647-652 [PMID: 25318098 DOI: 10.6061/clinics/2014(09)12]
- 25 Minaga K, Kitano M. Recent advances in endoscopic ultrasound-guided biliary drainage. Dig Endosc 2018; 30: 38-47 [PMID: 28656640 DOI: 10.1111/den.12910]
- 26 Teoh AYB, Dhir V, Kida M, Yasuda I, Jin ZD, Seo DW, Almadi M, Ang TL, Hara K, Hilmi I, Itoi T, Lakhtakia S, Matsuda K, Pausawasdi N, Puri R, Tang RS, Wang HP, Yang AM, Hawes R, Varadarajulu S, Yasuda K, Ho LKY. Consensus guidelines on the optimal management in interventional EUS procedures: results from the Asian EUS group RAND/UCLA expert panel. Gut 2018; 67: 1209-1228 [PMID: 29463614 DOI: 10.1136/gutjnl-2017-314341]
- Li DF, Zhou CH, Wang LS, Yao J, Zou DW. Is ERCP-BD or EUS-BD the preferred decompression 27 modality for malignant distal biliary obstruction? Rev Esp Enferm Dig 2019; 111: 953-960 [PMID: 31729233 DOI: 10.17235/reed.2019.6125/2018]
- 28 Ogura T, Sano T, Onda S, Imoto A, Masuda D, Yamamoto K, Kitano M, Takeuchi T, Inoue T, Higuchi K. Endoscopic ultrasound-guided biliary drainage for right hepatic bile duct obstruction: novel technical tips. Endoscopy 2015; 47: 72-75 [PMID: 25264761 DOI: 10.1055/s-0034-1378111]
- Park DH. Endoscopic ultrasound-guided biliary drainage of hilar biliary obstruction. J Hepatobiliary 29 Pancreat Sci 2015; 22: 664-668 [PMID: 26178753 DOI: 10.1002/jhbp.271]
- 30 Mukai S, Itoi T, Tsuchiya T, Tanaka R, Tonozuka R. EUS-guided right hepatic bile duct drainage in complicated hilar stricture. Gastrointest Endosc 2017; 85: 256-257 [PMID: 27492714 DOI: 10.1016/j.gie.2016.07.056
- 31 Minaga K, Takenaka M, Kitano M, Chiba Y, Imai H, Yamao K, Kamata K, Miyata T, Omoto S, Sakurai T, Watanabe T, Nishida N, Kudo M. Rescue EUS-guided intrahepatic biliary drainage for malignant hilar biliary stricture after failed transpapillary re-intervention. Surg Endosc 2017; 31: 4764-4772 [PMID: 28424912 DOI: 10.1007/s00464-017-5553-6]


- Ogura T, Onda S, Takagi W, Sano T, Okuda A, Masuda D, Yamamoto K, Miyano A, Kitano M, 32 Takeuchi T, Fukunishi S, Higuchi K. Clinical utility of endoscopic ultrasound-guided biliary drainage as a rescue of re-intervention procedure for high-grade hilar stricture. J Gastroenterol Hepatol 2017; 32: 163-168 [PMID: 27161286 DOI: 10.1111/jgh.13437]
- 33 Moryoussef F, Sportes A, Leblanc S, Bachet JB, Chaussade S, Prat F. Is EUS-guided drainage a suitable alternative technique in case of proximal biliary obstruction? Therap Adv Gastroenterol 2017; 10: 537-544 [PMID: 28804514 DOI: 10.1177/1756283X17702614]
- 34 Kongkam P, Orprayoon T, Boonmee C, Sodarat P, Seabmuangsai O, Wachiramatharuch C, Auan-Klin Y, Pham KC, Tasneem AA, Kerr SJ, Romano R, Jangsirikul S, Ridtitid W, Angsuwatcharakon P, Ratanachu-Ek T, Rerknimitr R. ERCP plus endoscopic ultrasound-guided biliary drainage vs percutaneous transhepatic biliary drainage for malignant hilar biliary obstruction: a multicenter observational open-label study. Endoscopy 2021; 53: 55-62 [PMID: 32515005 DOI: 10.1055/a-1195-8197]
- Lee TH, Choi JH, Park do H, Song TJ, Kim DU, Paik WH, Hwangbo Y, Lee SS, Seo DW, Lee SK, Kim MH. Similar Efficacies of Endoscopic Ultrasound-guided Transmural and Percutaneous Drainage for Malignant Distal Biliary Obstruction. Clin Gastroenterol Hepatol 2016; 14: 1011-1019.e3 [PMID: 26748220 DOI: 10.1016/j.cgh.2015.12.032]
- Sportes A, Camus M, Greget M, Leblanc S, Coriat R, Hochberger J, Chaussade S, Grabar S, Prat F. 36 Endoscopic ultrasound-guided hepaticogastrostomy versus percutaneous transhepatic drainage for malignant biliary obstruction after failed endoscopic retrograde cholangiopancreatography: a retrospective expertise-based study from two centers. Therap Adv Gastroenterol 2017; 10: 483-493 [PMID: 28567118 DOI: 10.1177/1756283X17702096]
- Nam K, Kim DU, Lee TH, Iwashita T, Nakai Y, Bolkhir A, Castro LA, Vazquez-Sequeiros E, de la 37 Serna C, Perez-Miranda M, Lee JG, Lee SS, Seo DW, Lee SK, Kim MH, Park DH. Patient perception and preference of EUS-guided drainage over percutaneous drainage when endoscopic transpapillary biliary drainage fails: An international multicenter survey. Endosc Ultrasound 2018; 7: 48-55 [PMID: 29451169 DOI: 10.4103/eus.eus_100_17]
- Hu QL, Liu JB, Ellis RJ, Liu JY, Yang AD, D'Angelica MI, Ko CY, Merkow RP. Association of 38 preoperative biliary drainage technique with postoperative outcomes among patients with resectable hepatobiliary malignancy. HPB (Oxford) 2020; 22: 249-257 [PMID: 31350104 DOI: 10.1016/j.hpb.2019.06.011]
- Arvanitakis M, Van Laethem JL, Pouzere S, Le Moine O, Deviere J. Predictive factors for survival 39 in patients with inoperable Klatskin tumors. Hepatogastroenterology 2006; 53: 21-27 [PMID: 16506370]
- 40 Rizzo A, Ricci AD, Frega G, Palloni A, DE Lorenzo S, Abbati F, Mollica V, Tavolari S, DI Marco M, Brandi G. How to Choose Between Percutaneous Transhepatic and Endoscopic Biliary Drainage in Malignant Obstructive Jaundice: An Updated Systematic Review and Meta-analysis. In Vivo 2020; 34: 1701-1714 [PMID: 32606139 DOI: 10.21873/invivo.11964]
- Cowling MG, Adam AN. Internal stenting in malignant biliary obstruction. World J Surg 2001; 25: 355-359; discussion 359-361 [PMID: 11343193 DOI: 10.1007/s002680020384]
- 42 Yoshida H, Mamada Y, Taniai N, Mizuguchi Y, Shimizu T, Yokomuro S, Aimoto T, Nakamura Y, Uchida E, Arima Y, Watanabe M, Tajiri T. One-step palliative treatment method for obstructive jaundice caused by unresectable malignancies by percutaneous transhepatic insertion of an expandable metallic stent. World J Gastroenterol 2006; 12: 2423-2426 [PMID: 16688837 DOI: 10.3748/wjg.v12.i15.2423]
- Bednarek M, Budzyński P, Poźniczek M, Rembiasz K. Percutaneous ultrasound-guided drainage of 43 the biliary tree in palliative treatment of mechanical jaundice: 17 years of experience. Wideochir Inne Tech Maloinwazyjne 2012; 7: 193-196 [PMID: 23256025 DOI: 10.5114/wiitm.2011.28896]
- 44 Hayashi N, Sakai T, Kitagawa M, Kimoto T, Inagaki R, Ishii Y. US-guided left-sided biliary drainage: nine-year experience. Radiology 1997; 204: 119-122 [PMID: 9205232 DOI: 10.1148/radiology.204.1.92052321
- 45 Krokidis M, Fanelli F, Orgera G, Bezzi M, Passariello R, Hatzidakis A. Percutaneous treatment of malignant jaundice due to extrahepatic cholangiocarcinoma: covered Viabil stent vs uncovered Wallstents. Cardiovasc Intervent Radiol 2010; 33: 97-106 [PMID: 19495871 DOI: 10.1007/s00270-009-9604-9
- Takahashi Y, Nagino M, Nishio H, Ebata T, Igami T, Nimura Y. Percutaneous transhepatic biliary 46 drainage catheter tract recurrence in cholangiocarcinoma. Br J Surg 2010; 97: 1860-1866 [PMID: 20799295 DOI: 10.1002/bjs.7228]
- Komaya K, Ebata T, Fukami Y, Sakamoto E, Miyake H, Takara D, Wakai K, Nagino M; Nagoya 47 Surgical Oncology Group. Percutaneous biliary drainage is oncologically inferior to endoscopic drainage: a propensity score matching analysis in resectable distal cholangiocarcinoma. J *Gastroenterol* 2016; **51**: 608-619 [PMID: 26553053 DOI: 10.1007/s00535-015-1140-6]
- 48 Vettoretto N, Arezzo A, Famiglietti F, Cirocchi R, Moja L, Morino M. Laparoscopic-endoscopic rendezvous vs preoperative endoscopic sphincterotomy in people undergoing laparoscopic cholecystectomy for stones in the gallbladder and bile duct. Cochrane Database Syst Rev 2018; 4: CD010507 [PMID: 29641848 DOI: 10.1002/14651858.CD010507.pub2]
- 49 Liu YD, Wang ZQ, Wang XD, Yang YS, Linghu EQ, Wang WF, Li W, Cai FC. Stent implantation through rendezvous technique of PTBD and ERCP: the treatment of obstructive jaundice. J Dig Dis 2007; 8: 198-202 [PMID: 17970876 DOI: 10.1111/j.1751-2980.2007.00305.x]



- 50 Siripun A, Sripongpun P, Ovartlarnporn B. Endoscopic ultrasound-guided biliary intervention in patients with surgically altered anatomy. World J Gastrointest Endosc 2015; 7: 283-289 [PMID: 25789101 DOI: 10.4253/wjge.v7.i3.283]
- 51 Yang MJ, Kim JH, Hwang JC, Yoo BM, Kim SS, Lim SG, Won JH. Usefulness of combined percutaneous-endoscopic rendezvous techniques after failed therapeutic endoscopic retrograde cholangiography in the era of endoscopic ultrasound guided rendezvous. Medicine (Baltimore) 2017; 96: e8991 [PMID: 29310413 DOI: 10.1097/MD.00000000008991]
- Coons H. Metallic stents for the treatment of biliary obstruction: a report of 100 cases. Cardiovasc 52 Intervent Radiol 1992; 15: 367-374 [PMID: 1335840 DOI: 10.1007/BF02734120]
- Adam A. Metallic biliary endoprostheses. Cardiovasc Intervent Radiol 1994; 17: 127-132 [PMID: 53 8087827 DOI: 10.1007/BF00195504]
- 54 Schima W, Prokesch R, Osterreicher C, Thurnher S, Függer R, Schöfl R, Havelec L, Lammer J. Biliary Wallstent endoprosthesis in malignant hilar obstruction: long-term results with regard to the type of obstruction. Clin Radiol 1997; 52: 213-219 [PMID: 9091256 DOI: 10.1016/s0009-9260(97)80275-9]
- Kaiho T, Miyazaki M, Ito H, Nakagawa K, Ambiru S, Shimizu H, Shimizu Y, Okuno A, Nozawa S, 55 Nukui Y, Nakajima N. Treatment of unresectable hepatic hilar malignancies with self-expanding metallic stents. Hepatogastroenterology 1999; 46: 2781-2790 [PMID: 10576345]
- Inal M, Akgül E, Aksungur E, Seydaoğlu G. Percutaneous placement of biliary metallic stents in 56 patients with malignant hilar obstruction: unilobar vs bilobar drainage. J Vasc Interv Radiol 2003; 14: 1409-1416 [PMID: 14605106 DOI: 10.1097/01.rvi.0000096762.74047.a6]
- 57 Lee MJ, Dawson SL, Mueller PR, Hahn PF, Saini S, Lu DS, Goldberg MA, Gazelle GS. Failed metallic biliary stents: causes and management of delayed complications. Clin Radiol 1994; 49: 857-862 [PMID: 7530177 DOI: 10.1016/s0009-9260(05)82875-2]
- Gupta P, Maralakunte M, Rathee S, Samanta J, Sharma V, Mandavdhare H, Sinha SK, Dutta U, 58 Kochhar R. Percutaneous transhepatic biliary drainage in patients at higher risk for adverse events: experience from a tertiary care referral center. Abdom Radiol (NY) 2020; 45: 2547-2553 [PMID: 31776603 DOI: 10.1007/s00261-019-02344-1]
- Walter T, Ho CS, Horgan AM, Warkentin A, Gallinger S, Greig PD, Kortan P, Knox JJ. Endoscopic 59 or percutaneous biliary drainage for Klatskin tumors? J Vasc Interv Radiol 2013; 24: 113-121 [PMID: 23182938 DOI: 10.1016/j.jvir.2012.09.019]
- Kloek JJ, van der Gaag NA, Aziz Y, Rauws EA, van Delden OM, Lameris JS, Busch OR, Gouma 60 DJ, van Gulik TM. Endoscopic and percutaneous preoperative biliary drainage in patients with suspected hilar cholangiocarcinoma. J Gastrointest Surg 2010; 14: 119-125 [PMID: 19756881 DOI: 10.1007/s11605-009-1009-11
- Coelen RJS, Roos E, Wiggers JK, Besselink MG, Buis CI, Busch ORC, Dejong CHC, van Delden 61 OM, van Eijck CHJ, Fockens P, Gouma DJ, Koerkamp BG, de Haan MW, van Hooft JE, IJzermans JNM, Kater GM, Koornstra JJ, van Lienden KP, Moelker A, Damink SWMO, Poley JW, Porte RJ, de Ridder RJ, Verheij J, van Woerden V, Rauws EAJ, Dijkgraaf MGW, van Gulik TM. Endoscopic vs percutaneous biliary drainage in patients with resectable perihilar cholangiocarcinoma: a multicentre, randomised controlled trial. Lancet Gastroenterol Hepatol 2018; 3: 681-690 [PMID: 30122355 DOI: 10.1016/S2468-1253(18)30234-6
- Inamdar S, Slattery E, Bhalla R, Sejpal DV, Trindade AJ. Comparison of Adverse Events for 62 Endoscopic vs Percutaneous Biliary Drainage in the Treatment of Malignant Biliary Tract Obstruction in an Inpatient National Cohort. JAMA Oncol 2016; 2: 112-117 [PMID: 26513013 DOI: 10.1001/jamaoncol.2015.3670]
- 63 Ba Y, Yue P, Leung JW, Wang H, Lin Y, Bai B, Zhu X, Zhang L, Zhu K, Wang W, Meng W, Zhou W, Liu Y, Li X. Percutaneous transhepatic biliary drainage may be the preferred preoperative drainage method in hilar cholangiocarcinoma. Endosc Int Open 2020; 8: E203-E210 [PMID: 32010755 DOI: 10.1055/a-0990-9114]
- Kim KM, Park JW, Lee JK, Lee KH, Lee KT, Shim SG. A Comparison of Preoperative Biliary 64 Drainage Methods for Perihilar Cholangiocarcinoma: Endoscopic vs Percutaneous Transhepatic Biliary Drainage. Gut Liver 2015; 9: 791-799 [PMID: 26087784 DOI: 10.5009/gnl14243]
- Wiggers JK, Groot Koerkamp B, Coelen RJ, Rauws EA, Schattner MA, Nio CY, Brown KT, Gonen 65 M, van Dieren S, van Lienden KP, Allen PJ, Besselink MG, Busch OR, D'Angelica MI, DeMatteo RP, Gouma DJ, Kingham TP, Jarnagin WR, van Gulik TM. Preoperative biliary drainage in perihilar cholangiocarcinoma: identifying patients who require percutaneous drainage after failed endoscopic drainage. Endoscopy 2015; 47: 1124-1131 [PMID: 26382308 DOI: 10.1055/s-0034-1392559]
- 66 Lee SH, Park JK, Yoon WJ, Lee JK, Ryu JK, Yoon YB, Kim YT. Optimal biliary drainage for inoperable Klatskin's tumor based on Bismuth type. World J Gastroenterol 2007; 13: 3948-3955 [PMID: 17663508 DOI: 10.3748/wjg.v13.i29.3948]
- Tang Z, Yang Y, Meng W, Li X. Best option for preoperative biliary drainage in Klatskin tumor: A 67 systematic review and meta-analysis. Medicine (Baltimore) 2017; 96: e8372 [PMID: 29069029 DOI: 10.1097/MD.00000000008372
- Liu JG, Wu J, Wang J, Shu GM, Wang YJ, Lou C, Zhang J, Du Z. Endoscopic Biliary Drainage 68 Versus Percutaneous Transhepatic Biliary Drainage in Patients with Resectable Hilar Cholangiocarcinoma: A Systematic Review and Meta-Analysis. J Laparoendosc Adv Surg Tech A 2018; 28: 1053-1060 [PMID: 29641365 DOI: 10.1089/lap.2017.0744]
- 69 Hameed A, Pang T, Chiou J, Pleass H, Lam V, Hollands M, Johnston E, Richardson A, Yuen L.



Percutaneous vs. endoscopic pre-operative biliary drainage in hilar cholangiocarcinoma - a systematic review and meta-analysis. HPB (Oxford) 2016; 18: 400-410 [PMID: 27154803 DOI: 10.1016/j.hpb.2016.03.002

- 70 Al Mahjoub A, Menahem B, Fohlen A, Dupont B, Alves A, Launoy G, Lubrano J. Preoperative Biliary Drainage in Patients with Resectable Perihilar Cholangiocarcinoma: Is Percutaneous Transhepatic Biliary Drainage Safer and More Effective than Endoscopic Biliary Drainage? J Vasc Interv Radiol 2017; 28: 576-582 [PMID: 28343588 DOI: 10.1016/j.jvir.2016.12.1218]
- 71 Moole H, Dharmapuri S, Duvvuri A, Boddireddy R, Moole V, Yedama P, Bondalapati N, Uppu A, Yerasi C. Endoscopic vs Percutaneous Biliary Drainage in Palliation of Advanced Malignant Hilar Obstruction: A Meta-Analysis and Systematic Review. Can J Gastroenterol Hepatol 2016; 2016: 4726078 [PMID: 27648439 DOI: 10.1155/2016/4726078]
- Saluja SS, Gulati M, Garg PK, Pal H, Pal S, Sahni P, Chattopadhyay TK. Endoscopic or 72 percutaneous biliary drainage for gallbladder cancer: a randomized trial and quality of life assessment. Clin Gastroenterol Hepatol 2008; 6: 944-950.c3 [PMID: 18585976 DOI: 10.1016/j.cgh.2008.03.028]
- Paik WH, Park YS, Hwang JH, Lee SH, Yoon CJ, Kang SG, Lee JK, Ryu JK, Kim YT, Yoon YB. Palliative treatment with self-expandable metallic stents in patients with advanced type III or IV hilar cholangiocarcinoma: a percutaneous vs endoscopic approach. Gastrointest Endosc 2009; 69: 55-62 [PMID: 18657806 DOI: 10.1016/j.gie.2008.04.005]
- Elmunzer BJ, Smith ZL, Tarnasky P, Wang AY, Yachimski P, Banovac F, Buscaglia JM, Buxbaum 74 J, Chak A, Chong B, Coté GA, Draganov PV, Dua K, Durkalski V, Geller BS, Jamil LH, Keswani RN, Khashab MA, Law R, Lo SK, McCarthy S, Selby JB, Singh VK, Taylor JR, Willingham FF, Spitzer RL, Foster LD; INTERCPT study group and the United States Cooperative for Outcomes Research in Endoscopy (USCORE). An Unsuccessful Randomized Trial of Percutaneous vs Endoscopic Drainage of Suspected Malignant Hilar Obstruction. Clin Gastroenterol Hepatol 2021; 19: 1282-1284 [PMID: 32454259 DOI: 10.1016/j.cgh.2020.05.035]
- Miyagawa S, Makuuchi M, Kawasaki S. Outcome of extended right hepatectomy after biliary 75 drainage in hilar bile duct cancer. Arch Surg 1995; 130: 759-763 [PMID: 7611866 DOI: 10.1001/archsurg.1995.01430070081016]
- dos Santos JS, Júnior WS, Módena JL, Brunaldi JE, Ceneviva R. Effect of preoperative endoscopic 76 decompression on malignant biliary obstruction and postoperative infection. Hepatogastroenterology 2005; 52: 45-47 [PMID: 15782991]
- 77 Wiggers JK, Coelen RJ, Rauws EA, van Delden OM, van Eijck CH, de Jonge J, Porte RJ, Buis CI, Dejong CH, Molenaar IQ, Besselink MG, Busch OR, Dijkgraaf MG, van Gulik TM. Preoperative endoscopic vs percutaneous transhepatic biliary drainage in potentially resectable perihilar cholangiocarcinoma (DRAINAGE trial): design and rationale of a randomized controlled trial. BMC Gastroenterol 2015; 15: 20 [PMID: 25887103 DOI: 10.1186/s12876-015-0251-0]
- 78 Hirano S, Tanaka E, Tsuchikawa T, Matsumoto J, Kawakami H, Nakamura T, Kurashima Y, Ebihara Y, Shichinohe T. Oncological benefit of preoperative endoscopic biliary drainage in patients with hilar cholangiocarcinoma. J Hepatobiliary Pancreat Sci 2014; 21: 533-540 [PMID: 24464984 DOI: 10.1002/jhbp.76
- 79 Sakata J, Shirai Y, Wakai T, Nomura T, Sakata E, Hatakeyama K. Catheter tract implantation metastases associated with percutaneous biliary drainage for extrahepatic cholangiocarcinoma. World J Gastroenterol 2005; 11: 7024-7027 [PMID: 16437610 DOI: 10.3748/wjg.v11.i44.7024]
- Maguchi H, Takahashi K, Katanuma A, Osanai M, Nakahara K, Matuzaki S, Urata T, Iwano H. 80 Preoperative biliary drainage for hilar cholangiocarcinoma. J Hepatobiliary Pancreat Surg 2007; 14: 441-446 [PMID: 17909711 DOI: 10.1007/s00534-006-1192-3]
- Kurahara H, Maemura K, Mataki Y, Sakoda M, Iino S, Kawasaki Y, Arigami T, Uenosono Y, 81 Kijima Y, Shinchi H, Takao S, Natsugoe S. Preoperative biliary drainage-related inflammation is associated with shorter survival in biliary tract cancer patients. Int J Clin Oncol 2016; 21: 934-939 [PMID: 26894390 DOI: 10.1007/s10147-016-0961-5]
- 82 Ipek S, Alper E, Cekic C, Cerrah S, Arabul M, Aslan F, Unsal B. Evaluation of the effectiveness of endoscopic retrograde cholangiopancreatography in patients with perihilar cholangiocarcinoma and its effect on development of cholangitis. Gastroenterol Res Pract 2014; 2014: 508286 [PMID: 24982670 DOI: 10.1155/2014/508286]
- Szary NM, Al-Kawas FH. Complications of endoscopic retrograde cholangiopancreatography: how 83 to avoid and manage them. Gastroenterol Hepatol (NY) 2013; 9: 496-504 [PMID: 24719597]



0 WÛ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2064-2075

DOI: 10.4251/wjgo.v13.i12.2064

ISSN 1948-5204 (online)

MINIREVIEWS

Current status of non-surgical treatment of locally advanced pancreatic cancer

Stavros Spiliopoulos, Maria Teresa Zurlo, Annachiara Casella, Letizia Laera, Giammarco Surico, Alessia Surgo, Alba Fiorentino, Nicola de'Angelis, Roberto Calbi, Riccardo Memeo, Riccardo Inchingolo

ORCID number: Stavros Spiliopoulos 0000-0003-1860-0568; Maria Teresa Zurlo 0000-0001-6572-3654: Annachiara Casella 0000-0001-6541-7390; Letizia Laera 0000-0003-2183-8817; Giammarco Surico 0000-0002-8329-0204; Alessia Surgo 0000-0003-4502-9635; Alba Fiorentino 0000-0003-4201-5852; Nicola de'Angelis 0000-0002-1211-4916; Roberto Calbi 0000-0003-0934-8890; Riccardo Memeo 0000-0002-1668-932X; Riccardo Inchingolo 0000-0002-0253-5936.

Author contributions: Spiliopoulos S, Zurlo MT, Casella A, Laera L, Surico G, Surgo A, Fiorentino A, de'Angelis N, Calbi R, Memeo R and Inchingolo R contributed data collection and manuscript writing, data analysis, and review design and supervision; all authors equally contributed to this paper with the conception and design of the study, literature review and analysis, drafting and critical revision and editing, and final approval of the final version.

Conflict-of-interest statement: The authors state that they have no conflicts of interest.

Country/Territory of origin: Greece

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer Stavros Spiliopoulos, 2nd Radiology Department, Interventional Radiology Unit, National and Kapodistrian University of Athens, Athens 12461, Greece

Maria Teresa Zurlo, Riccardo Inchingolo, Interventional Radiology Unit, "F. Miulli" Regional General Hospital, Acquaviva delle Fonti 70021, Italy

Annachiara Casella, Riccardo Memeo, Unit of Hepato-Pancreatic-Biliary Surgery, "F. Miulli" Regional General Hospital, Acquaviva delle Fonti 70021, Italy

Letizia Laera, Giammarco Surico, Department of Oncology, General Regional Hospital "F. Miulli", Acquaviva delle Fonti 70021, Italy

Alessia Surgo, Alba Fiorentino, Department of Radiation Oncology, "F. Miulli" Regional General Hospital, Acquaviva delle Fonti 70021, Italy

Nicola de'Angelis, Unit of Minimally Invasive and Robotic Digestive Surgery, "F. Miulli" Regional General Hospital, Acquaviva delle Fonti 70021, Italy

Roberto Calbi, Department of Radiology, General Regional Hospital "F. Miulli", Acquaviva delle Fonti 70021, Italy

Corresponding author: Stavros Spiliopoulos, MD, PhD, Associate Professor, 2nd Radiology Department, Interventional Radiology Unit, National and Kapodistrian University of Athens, 1st Rimini St, Chaidari, Athens 12461, Greece. stavspiliop@med.uoa.gr

Abstract

Pancreatic cancer is the 7th leading cause of death due to cancer in industrialized countries and the 11th most common cancer globally, with 458918 new cases (2.5% of all cancers) and 432242 deaths (4.5% of all cancer deaths) in 2018. Unfortunately, 80% to 90% of the patients present with unresectable disease, and the reported 5-year survival rate range between 10% and 25%, even after successful resection with tumor-free margins. Systemic chemotherapy, radiotherapy, and minimally invasive image-guided procedures that have emerged over the past years, are used for the management of non-operable PC. This review focuses on currently available non-surgical options of locally advanced pancreatic cancer.

Key Words: Interventional radiology; Oncology; Radiotherapy; Pancreatic cancer; Ablation



reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: March 22, 2021 Peer-review started: March 22, 2021 First decision: June 16, 2021 Revised: June 28, 2021 Accepted: October 25, 2021 Article in press: October 25, 2021 Published online: December 15, 2021

P-Reviewer: Carloni R, Ogino S S-Editor: Gao CC L-Editor: A P-Editor: Gao CC



©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Currently available non-surgical treatment options for locally advanced pancreatic cancer include systemic chemotherapy, radiotherapy, and minimally invasive image-guided procedures. The latter have emerged over the past years and include radiofrequency, microwave ablation, laser ablation, cryoablation, irreversible electroporation, high intensity focused ultrasound, and trans-arterial embolization procedures. Although initial results were not deemed satisfactory, mainly due to unacceptable complication rates, cumulative experience and technological advances have led to the improvement of outcomes of image-guided procedures and their incorporation in the treatment algorithm.

Citation: Spiliopoulos S, Zurlo MT, Casella A, Laera L, Surico G, Surgo A, Fiorentino A, de'Angelis N, Calbi R, Memeo R, Inchingolo R. Current status of non-surgical treatment of locally advanced pancreatic cancer. World J Gastrointest Oncol 2021; 13(12): 2064-2075 URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2064.htm

DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2064

INTRODUCTION

Pancreatic cancer (PC) is the 7th leading cause of death due to cancer in industrialized countries and the 11th most common cancer globally with 458918 new cases (2.5% of all cancers) and 432242 deaths (4.5% of all cancer deaths) in 2018[1]. Its incidence is variable among regions (age-standardized incidence highest in Europe: 7.7 per 100000 people and lowest in Africa: 2.2 per 100000 people) and is expected to increase[1,2]. Both incidence and mortality of the disease correlate with increasing age and male gender[2].

Modifiable risk factors include alcohol (increased risk high alcohol consumption > three drinks per day, but no association with low-to-moderate alcohol intake), smoking (relative risk 1.74 for current and 1.2 for former smokers; risk persists for at least 10 years after cessation), obesity [body mass index (BMI): 25.0-29.9 kg/m² or BMI \geq 30 kg/m² in early adulthood], dietary factors (red meats, processed meats, cholesterol, foods containing nitrosamines) and exposure to metalworking, pesticides, cadmium, and arsenic[2,3]. Non-modifiable risk factors are gender (global incidence 5.5/100000 for men and 4.0/100000 for women), age (> 50 years), ethnicity (significantly higher in black people than any other racial group), diabetes mellitus (1.8-fold risk increase), family history (double risk when at least two first-degree relatives have PC and 32-fold higher in kindreds with three or more first-degree relatives) genetic factors (10% of patients have some kind of germ-line mutation), chronic infections (Helicobacter pylori), non-O blood group and chronic pancreatitis (1.8% patients longstanding pre-existing chronic pancreatitis will develop PC within 10 years from diagnosis and 4% after 20 years)[2,4].

PC symptomatology is frequently non-specific (abdominal pain, jaundice, pruritus, dark urine, and acholic stools, anorexia, early satiety, dyspepsia, nausea, and eventually weight loss), while the early-stage disease remains frequently asymptomatic and most patients present at an advanced stage with poor prognosis[5,6]. Early detection could reduce mortality and screening of selected sub-groups such as those with a family history), using blood markers could be deemed beneficial in the future. Nevertheless, screening of the general population is not currently recommended[7,8]. Diagnosis and staging include imaging modalities such as contrast-enhanced, triphasic pancreatic computed tomography (CT) protocol, magnetic resonance imaging. Endoscopic ultrasound-guided fine-needle aspiration is indicated for cytological confirmation with a sensitivity of approximately 80%), while cancer antigen 19-9 assists in the confirmation of diagnosis, prediction of prognosis, and recurrence following surgery[7].

Surgical resection remains the only curative therapy for PC. However, 80% to 90% of the patients present with unresectable tumors and the reported 5-year survival rates are low, ranging between 10% and 25%, even after successful resection with tumorfree margins^[2,9]. Over the past years, alongside with systemic chemotherapy and radiotherapy (RT), minimally invasive image-guided procedures have emerged for the



management of non-operable or recurrent locally advanced PC. These include ablative modalities such as radiofrequency ablation (RFA), microwave ablation (MWA), laser ablation, cryoablation (CA), reversible electrochemotherapy (ECT) and irreversible electroporation (IRE), and high intensity focused ultrasound, while trans-arterial embolization procedures have also been suggested and investigated [10-12]. Following the initially non-satisfactory results, which were correlated with unacceptable complication rates, outcomes of minimally-invasive image-guided procedures were significantly improved, mainly due to the cumulative experience, and technological advances of the devices used, and their incorporation in the treatment algorithm for PC has been gradually accepted in specialized centers[10].

ABLATIVE TECHNIQUE

Patients with stable or partial response RECIST (response evaluation criteria in solid tumors) disease following neoadjuvant therapy, and not eligible for surgery should be considered as potential candidates for ablative therapies[13-16]. Ablative treatment can be divided into thermal (RFA, CA, MWA) and non-thermal (RE, IRE).

Thermal ablation

RFA uses needle electrodes to apply high-frequency alternating current to solid tumors. This process generates high temperatures, resulting in thermal coagulation, necrosis, and protein denaturation within the tumor^[15]. It can be performed during laparotomy, percutaneously, or using endoscopic ultrasound. Usually, RFA is adopted for tumor debulking rather than complete ablation, as a safety margin of at least 5 mm from the ablation zone is necessary to avoid thermal damage to vital structures. This treatment is also contraindicated in small tumors with a perivascular growth pattern [17]. Early studies investigating the efficacy and safety of RFA in the treatment of LAPC reported high rates of morbidity (4%-37%) and mortality (0%-25%) due to thermal injury to bile ducts, pancreatic duct, duodenum, vital vessels, and heat-sink effect, resulting in incomplete tumor ablation. Recent literature shows an improved overall survival (OS) ranging between 19.0 and 25.6 mo when combining RFA with chemotherapy[17] by optimizing ideal parameters for temperature range (< 90 degrees), treatment time, and probe placement[15].

CA induces rapid argon-gas-based freezing and thawing of target lesions. The freeze-thaw cycles, based on the Joule-Thompson effect, cause cellular destruction by vascular-mediated cytotoxicity, endothelial damage, and cell death. CA with and without immunotherapy for LAPC treatment in term of OS was studied in a retrospective study: Median OS was higher in the cryoimmunotherapy (13 mo) and cryotherapy groups (7 mo) than in the chemotherapy group (3.5 mo; both P < 0.001) and was higher in the cryoimmunotherapy group than in the cryotherapy (P < 0.05) and immunotherapy groups (5 mo; P < 0.001)[15].

MWA promotes tissue coagulation by the oscillation of water molecules, ultimately generating tissue necrosis. This technique has advantages over RFA in the ease of setup and larger ablation zones in a shorter period but recent literature on the use of MW ablation in PC with survival data is very limited[15].

Non-thermal ablation

IRE uses ultrashort high voltage direct current pulses to create an electric field across the cell membrane. This process disrupts membrane homeostasis and irreversibly alters transmembrane potential, which activates the apoptotic pathway and leads to cell death (Figure 1). IRE has the unique ability to preserve the extracellular matrix, critical vessel structures, bile ducts, intestines and minimize heat-sink effects resulting in potentially incomplete ablation. The technology is commercially available (NanoKnife; Angiodynamics Latham, NY, United States) with 510(k) clearance by the Food and Drug Administration (FDA) for ablation of soft tissue tumors[15,18]. In addition to its cytoreductive abilities, the evidence is emerging on IRE's capability to induce systemic immunomodulation through active in vivo vaccination against PC cells. IRE induces a systemic immune response following apoptosis and necrosis of tumor cells with the release of antigens and damage-associated molecular pattern molecules (DAMPs). These DAMPs promote the maturation of dendritic cells and other antigen-presenting cells that can subsequently take up the activation of lymph nodal T-cells potentially inducing a durable antitumor T-cell response. This response could then lead to regression in distant metastases, a process known as the "abscopal effect". These effects in combination with immunotherapy may offer a new treatment





Figure 1 Pancreatic cancer percutaneous irreversible electroporation. A: Axial computed tomography (CT) showing 2 cm lesion (red arrow) in the body of the pancreas in keeping with ductal adenocarcinoma; B and C: 3D volume rendering CT with parallel needles positioning within the lesion; D: 1-mo CT follow up showing complete ablation of the tumor (white arrow).

paradigm for tumors with low immunogenic potential, like pancreatic ductal adenocarcinoma^[16]. IRE can be performed during open or laparoscopic surgical exploration or percutaneous procedure with CT guidance. For IRE, 2-6 electrodes are typically placed around the tumor, with a maximum spacing of 2.0-2.5 cm, using image guidance and an upper limit of 5 cm in tumor diameter is highly recommended [17]. Given the high-risk profile of pancreatic IRE procedures, it is important to consider absolute and relative contraindications and to carefully assessed patients for treatment: it is advised to exclude patients with irreversible bleeding disorders, epilepsy, or any other unstable condition that precludes general anesthesia like patients with a past medical history of cardiac disease (*i.e.*, cardiac arrhythmia, implantable cardioverter defibrillator, pacemaker) given the risk of inducing cardiac arrhythmias when applying the electrical pulses [18]; pervasive involvement of the duodenum is considered an absolute contra-indication; if the patient is affected by biliary obstruction, adequate biliary drainage must be guaranteed prior to treatment and if the tumor is closed to the common bile duct it is highly recommended to ensure biliary protection prior to treatment, as post-IRE swelling can impede passage through the central bile ducts; if a patient suffers from a partially occluded portal vein prior to IRE, portal vein stenting should be performed to prevent acute complete occlusion due to postprocedural swelling. Most frequent adverse events comprise GI-related symptoms including pain, diarrhea, nausea, vomiting, loss of appetite, and delayed gastric emptying[16]. Previous cohort studies report heterogeneous outcomes, with median OS rates varying between 15-32 mo when combining IRE with systemic treatment. A recent systematic review by Moris et al[16] shows an overview of all studies that have utilized IRE for LAPC, with a median OS following IRE between 7 and 27 mo. This variance may be due to selection bias, the utilized IRE approach, the diverse LAPC tumor biology, personalized (neo-) adjuvant chemo- and/or RT protocols, performance status including comorbidities, and other interpersonal patient differences. In general, major complications (e.g., portal vein thrombosis, bleeding, duodenal perforation) are reported in 0%-30% of patients, with mortality rates ranging between 0%-11%[17]. The average cumulative morbidity for surgical and percutaneous IRE was 36% vs 24%, with an average periprocedural mortality rate of 2% vs 0%, respectively^[16].

Reversible ECT is a new non-thermal ablation technique that avoids possible thermal injury to the peripancreatic vessels like portal mesenteric vein combining the use of chemotherapeutic drugs (bleomycin) with electric pulses for cell membrane electroporation. A transient cell membrane improve permeability is determined by electric pulses, permitting the exposure of the cell to chemotherapeutic drugs[19]. The procedure is divided into four steps: Laparotomy or laparoscopic or percutaneous approach and intraoperative ultrasound to confirm that the pancreatic tumor was unresectable and to exclude distant metastases, needle insertion, bleomycin infusion and electroporation. Eight minutes after the bleomycin infusion, electric pulses were applied and delivered using four single long needle electrodes having 1.2 mm in diameter, and 3 or 4 cm active part. ECT was performed mostly in young patients (mean age, 63 years), with a good performance status and normal BMI. ECT was safe, according to the absence of acute intraoperative adverse effects related to electroporation and effects related to the bleomycin[19]. Nevertheless there is few studies regarding ECT in literatures^[20,21] and additional studies should be carried out.

INTRA-ARTERIAL THERAPIES

Since the 1950s, regional intra-arterial chemotherapy (RIAC) was introduced in an attempt to increase cancer survival rates. Intra-arterial chemotherapy generates high drug concentrations in the target areas while maintaining low systemic drug levels. Clinical trials demonstrated that regional intraarterial infusion with Gemcitabine (GEM) improved the response and resectability rates for advanced PC and was well tolerated by patients [22,23]. In 2012, a systematic review and meta-analysis of six randomized controlled trials, comparing systemic chemotherapy, with RIAC, reported that the latter resulted in higher partial remission, clinical benefits and response rates with fewer complications including myelosuppression[24].

With regards to efficacy, it is known that the effect of chemotherapy is concentration-dependent, therefore intra-arterial local infusion which generates higher drug concentrations within targeted regions, could be proven more efficient. In fact, according to the results of two RCTs, RIAC improved the 1-year OS (41.2%-28.6%) compared with systemic chemotherapy [25,26]

Other Authors considered the use of RIAC as neoadjuvant regional chemotherapy with continuous infusion of GEM intending to improve resectability rates in case of locally advanced PC[27]. A study was carried out to investigate the prognostic factors in patients who received GEM-based intra-arterial infusion for advanced PC; young age, pretreatment CA19-9 value < 1000 U/mL, and tumor located at the head of the pancreas indicated better response to RIAC and improved survival[28].

A recent retrospective cohort study of 454 patients with advanced PC compared RIAC via angiographically placed celiac axis catheters vs isolated upper abdominal perfusion (upper abdominal perfusion with stop flow balloon catheters in the aorta and vena cava)[29]. The isolated perfusion group demonstrated superior survival compared to intra- arterial infusion (median survival rates: 12 and 8 mo in stage III; 8.5 and 7 mo in stage IV; respectively)[29].

Future perspectives in local chemotherapy include novel infusion techniques such as Trans-Arterial Micro-Perfusion (TAMP) using the RenovoCath catheter (RenovoRx, Inc.) which was recently approved by the FDA. The TAMP procedure involves arterial segment isolation using proximal and distal occlusion balloons generating increased intra-arterial luminal pressure above the interstitial pressure and forcing the drug across the arterial wall within the tumoral tissue. The specific catheter has the potential to deliver higher drug concentrations within the tumor, while limiting systemic exposure. The TIGeR-PaC Phase III randomized clinical trial is currently enrolling patients with unresectable LAPC to investigate TAMP vs systemic chemotherapy. The goal of the trial is to prove extended median survival and improved quality of life through targeted delivery of therapy. In Phase, I/II studies, TAMP resulted in over two years survival in more than half the patients. The TIGeR-PaC trial, currently includes approximately 30 active clinical sites, and is expected to involve 200 participants in 40 centers in the United States and Europe (clinicaltrials.gov NCT03257033).

ONCOLOGY

LAPC is in general considered incurable and the management remains unclear and controversial. Treatment includes chemotherapy, which can have a potential role as



neoadjuvant treatment or in combination with RT on a case-based approach. The aim is to control disease progression, palliation, and improvement of OS. Patients with LAPC have a poor prognosis, with a median OS of 12-14 mo following systemic therapies[14]. The first step before choosing the appropriate treatment plan is the assessment of the patient's performance status. Patients with locally advanced disease received therapies based on their performance. Patients with a poor performance status are candidates to single-agent chemotherapy or palliative radiation therapy or best supportive care; patients with a good performance status can be considered for a more intensive oncological strategy as chemotherapy or chemoradiation[30].

Formerly, standard treatment included GEM for six months. However, in 2011, the Eastern cooperative Oncology group reported improved OS with the addition of radiation therapy (for a total of 50.4 Gy) to GEM[31].

In 2016, Suker et al^[32] conducted a patient-level meta-analysis of 11 studies (315 patients) indicating that LAPC patients treated with FOLFIRINOX demonstrated a median OS of 24.2 mo, which was significantly higher compared to that achieved by GEM therapy (6-13 mo).

While systematic chemotherapy has become the standard for patients with LAPC, surgery remains the only potentially curative therapy. Therefore, the development of a new neoadjuvant treatment that improves survival rate is of the utmost importance. Neoadjuvant chemotherapy should aim in a possible resection, or radiation therapy or IRE. Clinicians and researchers should investigate cancer biology and invest to define predictive and prognostic factors, patients-related features, and biological criteria to identify patients that would benefit from aggressive surgery or chemotherapy and chemoradiation[33].

When surgery is not possible after induction chemotherapy, chemoradiotherapy (intensity modulated radiation therapy or stereotactic body radiation therapy), is a possible option to incite tumor shrinkage and enable secondary resection in a small percentage of patients, while palliation of cancer-related pain is also a significant goal in unresectable tumors. The LAP07 phase III randomized controlled trial was designed to investigate the effect of chemoradiotherapy and erlotinib in the OS of patients with LAPC controlled after 4 mo of GEM-based induction chemotherapy. Unfortunately, both chemoradiotherapy and erlotinib did not provide any benefit in patients with LAPC[34].

On the other hand, in an updated 2018 review and meta-analysis of 41 studies (1018 patients receiving consolidation chemoradiation after induction chemotherapy and 954 patients receiving chemotherapy alone) the authors noted a significant survival benefit for chemoradiation after induction chemotherapy in cases in which chemotherapy lasted for a period of at least 3 mo[35].

Currently, according to the results of the SCALOP multicenter, open-label, randomized, two-arm, phase 2 trial, a capecitabine-based regimen should be preferred over a GEM-based regimen in the context of consolidation chemoradiotherapy after a course of induction chemotherapy for locally advanced PC[36].

Only recently, novel minimally invasive approaches, such as IRE, have been evaluated for treatment of LAPC after induction chemotherapy. The PANFIRE-2 multicenter, prospective, single-arm study investigated the safety and efficacy of percutaneous CT-guided IRE alone or after induction chemotherapy with GEM alone or FOLFIRINOX. In total 50 patients were enrolled (40 with LAPC and 10 with local recurrence) and target median OS was exceeded in patients with LAPC (17 mo) and those with local recurrence (16 mo). Notably, 14 minor and 21 major complications occurred, while one probable IRE-related death was reported^[14]. Some data suggest the potential role of IRE after FOLFIRINOX chemotherapy as better results have been noted in specific subgroups of patients. IRE could be considered as a less invasive potentially curative approach for LAPC[37]. Nevertheless, randomized clinical trials are necessary to provide comparative data regarding the efficacy of these three methods.

RT

About 80%-90% of the patients with PC, present with locally advanced disease at diagnosis and consequently a very poor prognosis of less than 5% OS at 5-years[38, 39]. In this group of patients, RT associated with systemic therapy plays a crucial role in achieving satisfactory local control (LC). As noted above, the LAP 07 phase III study failed to demonstrate any significant advantage on OS, but a substantially decreased local progression rate (32% vs 46%, P = 0.03) for chemoradiotherapy was noted,



without increase in grade 3 to 4 toxicity (except for nausea)[34]. In this context, for patients demonstrating good performance status, the aim of combining RT with systemic therapy is to provide satisfactory LC and avoid or delay local disease progression. However, it is crucial that during RT a higher radiation dose should be delivered to the tumor, limiting doses to healthy tissue.

Conventionally, the standard RT dose has been set around of 50-54 Gy administered over 6 wk using the 3D conformal radiation technique (3D-CRT), a dose initially established on the tolerance of large-field radiation to organ at risks. However, these doses showed a limited LC of the disease, while higher doses often induce treatmentrelated toxicity that can significantly affect the patient's quality of life[40].

Over the past decade, modern technological advances such as intensity-modulated RT (IMRT), the introduction of respiratory management methods and improved imaging guidance during therapy, have enabled dose-escalation therapy and facilitated the possibility to reduce treatment volumes, to improve ablative doses to the tumor and consequently clinical outcomes, concurrently reducing treatment toxicity[41]. In fact, like other mobile tumors, pancreatic tumors require assessment of respiratory movements, while RT dosimetry is additionally complicated by the presence of nearby hollow digestive organs.

In a 2015 systematic review, 13 IMRT studies were analyzed and compared with 7 3D-CRT series, in order to compare toxicity and tumor outcomes between these two different RT-techniques for the management of LAPC. Even though no differences in terms of OS and PFS were obtained, IMRT was correlated with reduced risk of side effects [42]. Surely, the absence of OS advantage could be explained by the conventionally delivered dose, which is not sufficient to efficiently control the disease^[43].

In a study published by Krishnan et al[44], patients who received an intensified dose with a biological effective dose (BED) > 70 Gy demonstrated higher OS compared to those who received a standard dose BED \leq 70 Gy. In this scenario, in which the main limitation remains the risk of severe intestinal toxicity, stereotactic body RT (SBRT) has also been largely explored as a potentially effective treatment for patients with LAPC (Figure 2)[45-61]. Of note, poor prognosis related to these tumors has motivated physicians to limit the number of RT sessions and concentrate treatment within one to two weeks, thereby reducing the period of chemotherapy interruption and increasing patient's compliance.

According to recent data on SBRT treatment for LAPC, a large heterogeneity in total dose prescription and fractionations was reported. Generally, the total doses used ranged from 30 Gy to 45 Gy in 3-5 fractions resulting in effective biological dose (BED) superior to 60 Gy (assuming an $\alpha/\beta = 10$) in most of the cases[46-60]. This higher dose/fraction can theoretically produce enhanced cell destruction than conventional fractionation[62]. Due to the aggressiveness of the disease which causes a rapid evolution of the metastatic sites, OS is not greatly decreased in SBRT series.

Despite satisfactory LC and median freedom from local progression rates of approx. Eighty percent at 1 year, and very low acute toxicity obtained by SBRT single fraction treatments (25 Gy/1 fr)[45,47,48,51,52], high rates of late toxicity have limited the use of this approach. Therefore, multi-fraction treatment schemes are preferably used. In a retrospective cohort of 167 LAPC patients, 45% of whom were treated in single fraction SBRT and 54% in five-fraction SBRT, five-fraction protocol showed significantly lower gastrointestinal toxicity compared to the single fraction treatment. However, good LC rate was noted in both groups[54].

Similar conclusions were drawn following several retrospective studies that acknowledged the benefit in terms of reduced toxicity when multi-fractionated (3-5 fractions) RT regimens were applied compared to single fraction treatment, while obtaining the same efficacy in terms of LC[49,50,52,53,55-57]. All patients investigated in these studies, received systemic therapy (mainly GEM) during different time points of SBRT treatment (before, after or both before and after SBRT) and therapeutic schemes were variable based on the patients' clinical status and the grade of the disease. Notably, the indication for intensified local treatment using SBRT in LAPC patients may be abruptly compromised due to the high metastatic potential of the disease.

Only few, non-randomized, retrospective studies compared SBRT and conventional fractionated RT[63] in the local advanced PC curative or neo-adjuvant setting. SBRT was associated with significantly improved OS compared to conventional fractionated RT. Additionally, SBRT was associated with significantly increased rates of pathological complete response and margin-negative resection in neo-adjuvant setting. These are promising results and provide the basis for consideration for prospective validation.





Figure 2 Radiation treatment plan for a patient treated with stereotactic body radiotherapy for local advanced pancreatic cancer. A-C: The plans show isodose levels in the axial plane (A), coronal plane (B), and sagittal plane (C).

However, the indication for SBRT in order to intensify local treatment may be affected by the high metastatic potential of the disease.

CONCLUSION

The prognosis of LAPC remains poor although recent advancements in multimodality treatment seem to provide some improvement in clinical outcomes. Combined modality treatment using systemic chemotherapy, minimally invasive image-guided procedures and RT are currently under investigation. Neoadjuvant chemotherapy could be used to enable curative surgical resection, radiation therapy, or ablation. SBRT may be employed to obtain satisfactory LC and reduce side effects. Mounting experience with percutaneous thermal ablation and IRE provides superior outcomes with less complications. Considering the currently unsatisfactory results in terms of OS improvement, different treatment options should be investigated to optimize therapy. As the pathogenesis of LAPD is multifactorial and has been associated with genetic factors (mainly germ-line BRCA2 gene mutations, but also various syndromes such as the Lynch syndrome, hereditary breast and ovarian cancer syndrome, familial adenomatous polyposis and Li-Fraumeni syndrome) environmental factors (obesity, smoking habit, diabetes, alcohol consumption, dietary factors such as red meat consumption, and occupational exposure to nickel cadmium and arsenic, and the human microbiome), future treatment directions should focus on the investigation of these factors to provide personalized therapeutic schemes and improve survival[2,64, 65].

REFERENCES

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: 1 GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- 2 Rawla P, Sunkara T, Gaduputi V. Epidemiology of Pancreatic Cancer: Global Trends, Etiology and Risk Factors. World J Oncol 2019; 10: 10-27 [PMID: 30834048 DOI: 10.14740/wjon1166]
- 3 Shin EJ, Canto MI. Pancreatic cancer screening. Gastroenterol Clin North Am 2012; 41: 143-157 [PMID: 22341255 DOI: 10.1016/j.gtc.2011.12.001]
- American Cancer Society. Cancer Facts and Figures 2014. [cited 20 March 2021]. In: American Cancer Society [Internet]. Available from: https://www.cancer.org/research/cancer-facts-statistics/allcancer-facts-figures/cancer-facts-figures-2014.html
- Guo XZ, Cui ZM, Liu X. Current developments, problems and solutions in the non-surgical treatment 5 of pancreatic cancer. World J Gastrointest Oncol 2013; 5: 20-28 [PMID: 23556053 DOI: 10.4251/wjgo.v5.i2.20]
- 6 Krech RL, Walsh D. Symptoms of pancreatic cancer. J Pain Symptom Manage 1991; 6: 360-367 [PMID: 1880437 DOI: 10.1016/0885-3924(91)90027-2]
- 7 Cappelli G, Paladini S, D'Agata A. [Tumor markers in the diagnosis of pancreatic cancer]. Tumori 1999; 85: S19-S21 [PMID: 10235075]
- Rastogi T, Devesa S, Mangtani P, Mathew A, Cooper N, Kao R, Sinha R. Cancer incidence rates among South Asians in four geographic regions: India, Singapore, UK and US. Int J Epidemiol 2008; 37: 147-160 [PMID: 18094016 DOI: 10.1093/ije/dym219]
- Saif MW. Controversies in the adjuvant treatment of pancreatic adenocarcinoma. JOP 2007; 8: 545-



552 [PMID: 17873458]

- Ierardi AM, Lucchina N, Bacuzzi A, Marco de C, Bracchi E, Cocozza E, Dionigi G, Tsetis D, Floridi 10 C, Carrafiello G. Percutaneous ablation therapies of inoperable pancreatic cancer: a systematic review. Ann Gastroenterol 2015; 28: 431-439 [PMID: 26424487]
- 11 Tanaka T, Sho M, Nishiofuku H, Sakaguchi H, Inaba Y, Nakajima Y, Kichikawa K. Unresectable pancreatic cancer: arterial embolization to achieve a single blood supply for intraarterial infusion of 5fluorouracil and full-dose IV gemcitabine. AJR Am J Roentgenol 2012; 198: 1445-1452 [PMID: 22623561 DOI: 10.2214/AJR.11.8008]
- Das SK, Wang JL, Li B, Zhang C, Yang HF. Clinical effectiveness of combined interventional 12 therapy as a salvage modality for unresectable pancreatic carcinoma. Oncol Lett 2019; 18: 375-385 [PMID: 31289509 DOI: 10.3892/ol.2019.10323]
- 13 Lafranceschina S, Brunetti O, Delvecchio A, Conticchio M, Ammendola M, Currò G, Piardi T, de'Angelis N, Silvestris N, Memeo R. Systematic Review of Irreversible Electroporation Role in Management of Locally Advanced Pancreatic Cancer. Cancers (Basel) 2019; 11 [PMID: 31684186 DOI: 10.3390/cancers11111718]
- Ruarus AH, Vroomen LGPH, Geboers B, van Veldhuisen E, Puijk RS, Nieuwenhuizen S, Besselink 14 MG, Zonderhuis BM, Kazemier G, de Gruijl TD, van Lienden KP, de Vries JJJ, Scheffer HJ, Meijerink MR. Percutaneous Irreversible Electroporation in Locally Advanced and Recurrent Pancreatic Cancer (PANFIRE-2): A Multicenter, Prospective, Single-Arm, Phase II Study. Radiology 2020; 294: 212-220 [PMID: 31687922 DOI: 10.1148/radiol.2019191109]
- 15 Narayanan G, Ucar A, Gandhi RT, Nasiri A, Inampudi P, Wilson NM, Asbun HJ. Emerging Ablative and Transarterial Therapies for Pancreatic Cancer. Dig Dis Interv 2020; 4: 389-339 [DOI: 10.1055/s-0040-1721415]
- 16 Moris D, Machairas N, Tsilimigras DI, Prodromidou A, Ejaz A, Weiss M, Hasemaki N, Felekouras E, Pawlik TM. Systematic Review of Surgical and Percutaneous Irreversible Electroporation in the Treatment of Locally Advanced Pancreatic Cancer. Ann Surg Oncol 2019; 26: 1657-1668 [PMID: 30843163 DOI: 10.1245/s10434-019-07261-71
- van Veldhuisen E, van den Oord C, Brada LJ, Walma MS, Vogel JA, Wilmink JW, Del Chiaro M, 17 van Lienden KP, Meijerink MR, van Tienhoven G, Hackert T, Wolfgang CL, van Santvoort H, Groot Koerkamp B, Busch OR, Molenaar IQ, van Eijck CH, Besselink MG; Dutch Pancreatic Cancer Group and International Collaborative Group on Locally Advanced Pancreatic Cancer. Locally Advanced Pancreatic Cancer: Work-Up, Staging, and Local Intervention Strategies. Cancers (Basel) 2019; 11 [PMID: 31336859 DOI: 10.3390/cancers11070976]
- 18 Narayanan G, Hosein PJ, Beulaygue IC, Froud T, Scheffer HJ, Venkat SR, Echenique AM, Hevert EC, Livingstone AS, Rocha-Lima CM, Merchan JR, Levi JU, Yrizarry JM, Lencioni R. Percutaneous Image-Guided Irreversible Electroporation for the Treatment of Unresectable, Locally Advanced Pancreatic Adenocarcinoma. J Vasc Interv Radiol 2017; 28: 342-348 [PMID: 27993507 DOI: 10.1016/j.jvir.2016.10.023]
- 19 Casadei R, Ricci C, Ingaldi C, Alberici L, Di Marco M, Guido A, Minni F, Serra C. Intraoperative electrochemotherapy in locally advanced pancreatic cancer: indications, techniques and results-a single-center experience. Updates Surg 2020; 72: 1089-1096 [PMID: 32399592 DOI: 10.1007/s13304-020-00782-x]
- Jaroszeski MJ, Gilbert RA, Heller R. In vivo antitumor effects of electrochemotherapy in a hepatoma 20 model. Biochim Biophys Acta 1997; 1334: 15-18 [PMID: 9042359 DOI: 10.1016/s0304-4165(96)00147-x
- 21 Girelli R, Prejanò S, Cataldo I, Corbo V, Martini L, Scarpa A, Claudio B. Feasibility and safety of electrochemotherapy (ECT) in the pancreas: a pre-clinical investigation. Radiol Oncol 2015; 49: 147-154 [PMID: 26029026 DOI: 10.1515/raon-2015-0013]
- 22 Aigner KR, Gailhofer S. Celiac axis infusion and microembolization for advanced stage III/IV pancreatic cancer--a phase II study on 265 cases. Anticancer Res 2005; 25: 4407-4412 [PMID: 16334117]
- Li Q, Wang MQ, Duan LX, Song P, Ao GK. Regional arterial infusion chemothera-py with lipid 23 emulsion as a solvent for the treatment of advanced pancreatic can-cer: A preliminary clinical study. J Interv Radiol 2009; 18: 275-277
- Liu F, Tang Y, Sun J, Yuan Z, Li S, Sheng J, Ren H, Hao J. Regional intra-arterial vs. systemic 24 chemotherapy for advanced pancreatic cancer: a systematic review and meta-analysis of randomized controlled trials. PLoS One 2012; 7: e40847 [PMID: 22815840 DOI: 10.1371/journal.pone.0040847]
- Aigner KR, Gailhofer S, Kopp S. Regional vs systemic chemotherapy for advanced pancreatic 25 cancer: a randomized study. Hepatogastroenterology 1998; 45: 1125-1129 [PMID: 9756018]
- Hong GB, Zhou JX, Liang BL. A Clinical Study on Continuous Transarterial Infusion Chemotherapy 26 with Gemcitabine and 5-fluorouracil in Treating Patients with Advanced Pancreatic Carcinoma. Cancer Prev 2007; 33: 54-56
- Davis JL, Pandalai P, Ripley RT, Langan RC, Steinberg SM, Walker M, Toomey MA, Levy E, 27 Avital I. Regional chemotherapy in locally advanced pancreatic cancer: RECLAP trial. Trials 2011; 12: 129 [PMID: 21595953 DOI: 10.1186/1745-6215-12-129]
- Liu X, Yang X, Zhou G, Chen Y, Li C, Wang X. Gemcitabine-Based Regional Intra-Arterial Infusion 28 Chemotherapy in Patients With Advanced Pancreatic Adenocarcinoma. Medicine (Baltimore) 2016; 95: e3098 [PMID: 26986149 DOI: 10.1097/MD.00000000003098]
- Aigner KR, Gailhofer S, Selak E, Aigner K. Intra-arterial infusion chemotherapy vs isolated upper 29



abdominal perfusion for advanced pancreatic cancer: a retrospective cohort study on 454 patients. J Cancer Res Clin Oncol 2019; 145: 2855-2862 [PMID: 31506738 DOI: 10.1007/s00432-019-03019-6]

- 30 National Comprehensive Cancer Network. NCCN guideline version 1.2021 pancreatic cancer. [cited 20 March 2021]. In: National Comprehensive Cancer Network [Internet]. Available from: https://www.spg.pt/wp-content/uploads/Guidelines/NCCN/pancreatic.pdf
- Loehrer PJ Sr, Feng Y, Cardenes H, Wagner L, Brell JM, Cella D, Flynn P, Ramanathan RK, Crane 31 CH, Alberts SR, Benson AB 3rd. Gemcitabine alone vs gemcitabine plus radiotherapy in patients with locally advanced pancreatic cancer: an Eastern Cooperative Oncology Group trial. J Clin Oncol 2011; 29: 4105-4112 [PMID: 21969502 DOI: 10.1200/JCO.2011.34.8904]
- 32 Suker M, Beumer BR, Sadot E, Marthey L, Faris JE, Mellon EA, El-Rayes BF, Wang-Gillam A, Lacy J, Hosein PJ, Moorcraft SY, Conroy T, Hohla F, Allen P, Taieb J, Hong TS, Shridhar R, Chau I, van Eijck CH, Koerkamp BG. FOLFIRINOX for locally advanced pancreatic cancer: a systematic review and patient-level meta-analysis. Lancet Oncol 2016; 17: 801-810 [PMID: 27160474 DOI: 10.1016/S1470-2045(16)00172-8]
- Barcellini A, Peloso A, Pugliese L, Vitolo V, Cobianchi L. Locally Advanced Pancreatic Ductal 33 Adenocarcinoma: Challenges and Progress. Onco Targets Ther 2020; 13: 12705-12720 [PMID: 33335406 DOI: 10.2147/OTT.S220971]
- Hammel P, Huguet F, van Laethem JL, Goldstein D, Glimelius B, Artru P, Borbath I, Bouché O, 34 Shannon J, André T, Mineur L, Chibaudel B, Bonnetain F, Louvet C: LAP07 Trial Group. Effect of Chemoradiotherapy vs Chemotherapy on Survival in Patients With Locally Advanced Pancreatic Cancer Controlled After 4 Months of Gemcitabine With or Without Erlotinib: The LAP07 Randomized Clinical Trial. JAMA 2016; 315: 1844-1853 [PMID: 27139057 DOI: 10.1001/jama.2016.4324]
- 35 Chang JS, Chiu YF, Yu JC, Chen LT, Ch'ang HJ. The Role of Consolidation Chemoradiotherapy in Locally Advanced Pancreatic Cancer Receiving Chemotherapy: An Updated Systematic Review and Meta-Analysis. Cancer Res Treat 2018; 50: 562-574 [PMID: 28602054 DOI: 10.4143/crt.2017.105]
- Mukherjee S, Hurt CN, Bridgewater J, Falk S, Cummins S, Wasan H, Crosby T, Jephcott C, Roy R, 36 Radhakrishna G, McDonald A, Ray R, Joseph G, Staffurth J, Abrams RA, Griffiths G, Maughan T. Gemcitabine-based or capecitabine-based chemoradiotherapy for locally advanced pancreatic cancer (SCALOP): a multicentre, randomised, phase 2 trial. Lancet Oncol 2013; 14: 317-326 [PMID: 23474363 DOI: 10.1016/S1470-2045(13)70021-4]
- 37 He C, Sun S, Huang X, Zhang Y, Lin X, Li S. Survival Comparison of Neoadjuvant Chemotherapy Followed by Irreversible Electroporation Versus Conversional Resection for Locally Advanced Pancreatic Cancer. Front Oncol 2020; 10: 622318 [PMID: 33604301 DOI: 10.3389/fonc.2020.622318
- 38 Varadhachary GR, Tamm EP, Abbruzzese JL, Xiong HQ, Crane CH, Wang H, Lee JE, Pisters PW, Evans DB, Wolff RA. Borderline resectable pancreatic cancer: definitions, management, and role of preoperative therapy. Ann Surg Oncol 2006; 13: 1035-1046 [PMID: 16865597 DOI: 10.1245/ASO.2006.08.011]
- 39 Worni M, Guller U, White RR, Castleberry AW, Pietrobon R, Cerny T, Gloor B, Koeberle D. Modest improvement in overall survival for patients with metastatic pancreatic cancer: a trend analysis using the surveillance, epidemiology, and end results registry from 1988 to 2008. Pancreas 2013; 42: 1157-1163 [PMID: 23867367 DOI: 10.1097/MPA.0b013e318291fbc5]
- Cattaneo GM, Passoni P, Longobardi B, Slim N, Reni M, Cereda S, di Muzio N, Calandrino R. 40 Dosimetric and clinical predictors of toxicity following combined chemotherapy and moderately hypofractionated rotational radiotherapy of locally advanced pancreatic adenocarcinoma. Radiother Oncol 2013; 108: 66-71 [PMID: 23726116 DOI: 10.1016/j.radonc.2013.05.011]
- 41 Garibaldi C, Jereczek-Fossa BA, Marvaso G, Dicuonzo S, Rojas DP, Cattani F, Starzyńska A, Ciardo D, Surgo A, Leonardi MC, Ricotti R. Recent advances in radiation oncology. Ecancermedicalscience 2017; 11: 785 [PMID: 29225692 DOI: 10.3332/ecancer.2017.785]
- Bittner MI, Grosu AL, Brunner TB. Comparison of toxicity after IMRT and 3D-conformal 42 radiotherapy for patients with pancreatic cancer - a systematic review. Radiother Oncol 2015; 114: 117-121 [PMID: 25497876 DOI: 10.1016/j.radonc.2014.11.043]
- Fokas E, O'Neill E, Gordon-Weeks A, Mukherjee S, McKenna WG, Muschel RJ. Pancreatic ductal 43 adenocarcinoma: From genetics to biology to radiobiology to oncoimmunology and all the way back to the clinic. Biochim Biophys Acta 2015; 1855: 61-82 [PMID: 25489989 DOI: 10.1016/j.bbcan.2014.12.001
- Krishnan S, Chadha AS, Suh Y, Chen HC, Rao A, Das P, Minsky BD, Mahmood U, Delclos ME, 44 Sawakuchi GO, Beddar S, Katz MH, Fleming JB, Javle MM, Varadhachary GR, Wolff RA, Crane CH. Focal Radiation Therapy Dose Escalation Improves Overall Survival in Locally Advanced Pancreatic Cancer Patients Receiving Induction Chemotherapy and Consolidative Chemoradiation. Int J Radiat Oncol Biol Phys 2016; 94: 755-765 [PMID: 26972648 DOI: 10.1016/j.ijrobp.2015.12.003]
- 45 Koong AC, Le QT, Ho A, Fong B, Fisher G, Cho C, Ford J, Poen J, Gibbs IC, Mehta VK, Kee S, Trueblood W, Yang G, Bastidas JA. Phase I study of stereotactic radiosurgery in patients with locally advanced pancreatic cancer. Int J Radiat Oncol Biol Phys 2004; 58: 1017-1021 [PMID: 15001240 DOI: 10.1016/j.ijrobp.2003.11.004]
- 46 Hoyer M, Roed H, Sengelov L, Traberg A, Ohlhuis L, Pedersen J, Nellemann H, Kiil Berthelsen A, Eberholst F, Engelholm SA, von der Maase H. Phase-II study on stereotactic radiotherapy of locally advanced pancreatic carcinoma. Radiother Oncol 2005; 76: 48-53 [PMID: 15990186 DOI:



10.1016/i.radonc.2004.12.022

- 47 Schellenberg D, Goodman KA, Lee F, Chang S, Kuo T, Ford JM, Fisher GA, Quon A, Desser TS, Norton J, Greco R, Yang GP, Koong AC. Gemcitabine chemotherapy and single-fraction stereotactic body radiotherapy for locally advanced pancreatic cancer. Int J Radiat Oncol Biol Phys 2008; 72: 678-686 [PMID: 18395362 DOI: 10.1016/j.ijrobp.2008.01.051]
- Chang DT, Schellenberg D, Shen J, Kim J, Goodman KA, Fisher GA, Ford JM, Desser T, Quon A, 48 Koong AC. Stereotactic radiotherapy for unresectable adenocarcinoma of the pancreas. Cancer 2009; 115: 665-672 [PMID: 19117351 DOI: 10.1002/cncr.24059]
- 49 Polistina F, Costantin G, Casamassima F, Francescon P, Guglielmi R, Panizzoni G, Febbraro A, Ambrosino G. Unresectable locally advanced pancreatic cancer: a multimodal treatment using neoadjuvant chemoradiotherapy (gemcitabine plus stereotactic radiosurgery) and subsequent surgical exploration. Ann Surg Oncol 2010; 17: 2092-2101 [PMID: 20224860 DOI: 10.1245/s10434-010-1019-y]
- 50 Rwigema JC, Parikh SD, Heron DE, Howell M, Zeh H, Moser AJ, Bahary N, Quinn A, Burton SA. Stereotactic body radiotherapy in the treatment of advanced adenocarcinoma of the pancreas. Am J Clin Oncol 2011; 34: 63-69 [PMID: 20308870 DOI: 10.1097/COC.0b013e3181d270b4]
- Goyal K, Einstein D, Ibarra RA, Yao M, Kunos C, Ellis R, Brindle J, Singh D, Hardacre J, Zhang Y, 51 Fabians J, Funkhouser G, Machtay M, Sanabria JR. Stereotactic body radiation therapy for nonresectable tumors of the pancreas. J Surg Res 2012; 174: 319-325 [PMID: 21937061 DOI: 10.1016/j.jss.2011.07.044]
- 52 Gurka MK, Collins SP, Slack R, Tse G, Charabaty A, Ley L, Berzcel L, Lei S, Suy S, Haddad N, Jha R, Johnson CD, Jackson P, Marshall JL, Pishvaian MJ. Stereotactic body radiation therapy with concurrent full-dose gemcitabine for locally advanced pancreatic cancer: a pilot trial demonstrating safety. Radiat Oncol 2013; 8: 44 [PMID: 23452509 DOI: 10.1186/1748-717X-8-44]
- 53 Tozzi A, Comito T, Alongi F, Navarria P, Iftode C, Mancosu P, Reggiori G, Clerici E, Rimassa L, Zerbi A, Fogliata A, Cozzi L, Tomatis S, Scorsetti M. SBRT in unresectable advanced pancreatic cancer: preliminary results of a mono-institutional experience. Radiat Oncol 2013; 8: 148 [PMID: 23799996 DOI: 10.1186/1748-717X-8-148]
- 54 Herman JM, Chang DT, Goodman KA, Dholakia AS, Raman SP, Hacker-Prietz A, Iacobuzio-Donahue CA, Griffith ME, Pawlik TM, Pai JS, O'Reilly E, Fisher GA, Wild AT, Rosati LM, Zheng L, Wolfgang CL, Laheru DA, Columbo LA, Sugar EA, Koong AC. Phase 2 multi-institutional trial evaluating gemcitabine and stereotactic body radiotherapy for patients with locally advanced unresectable pancreatic adenocarcinoma. Cancer 2015; 121: 1128-1137 [PMID: 25538019 DOI: 10.1002/cncr.29161]
- Lin JC, Jen YM, Li MH, Chao HL, Tsai JT. Comparing outcomes of stereotactic body radiotherapy 55 with intensity-modulated radiotherapy for patients with locally advanced unresectable pancreatic cancer. Eur J Gastroenterol Hepatol 2015; 27: 259-264 [PMID: 25629569 DOI: 10.1097/MEG.00000000000283]
- Mellon EA, Hoffe SE, Springett GM, Frakes JM, Strom TJ, Hodul PJ, Malafa MP, Chuong MD, 56 Shridhar R. Long-term outcomes of induction chemotherapy and neoadjuvant stereotactic body radiotherapy for borderline resectable and locally advanced pancreatic adenocarcinoma. Acta Oncol 2015; 54: 979-985 [PMID: 25734581 DOI: 10.3109/0284186X.2015.1004367]
- 57 Comito T, Cozzi L, Clerici E, Franzese C, Tozzi A, Iftode C, Navarria P, D'Agostino G, Rimassa L, Carnaghi C, Personeni N, Tronconi MC, De Rose F, Franceschini D, Ascolese AM, Fogliata A, Tomatis S, Santoro A, Zerbi A, Scorsetti M. Can Stereotactic Body Radiation Therapy Be a Viable and Efficient Therapeutic Option for Unresectable Locally Advanced Pancreatic Adenocarcinoma? Technol Cancer Res Treat 2017; 16: 295-301 [PMID: 27311310 DOI: 10.1177/1533034616650778]
- 58 Gurka MK, Kim C, He AR, Charabaty A, Haddad N, Turocy J, Johnson L, Jackson P, Weiner LM, Marshall JL, Collins SP, Pishvaian MJ, Unger K. Stereotactic Body Radiation Therapy (SBRT) Combined With Chemotherapy for Unresected Pancreatic Adenocarcinoma. Am J Clin Oncol 2017; 40: 152-157 [PMID: 25171298 DOI: 10.1097/COC.00000000000118]
- Mazzola R, Fersino S, Aiello D, Gregucci F, Tebano U, Corradini S, Di Paola G, Cirillo M, Tondulli 59 L, Ruffo G, Ruggieri R, Alongi F. Linac-based stereotactic body radiation therapy for unresectable locally advanced pancreatic cancer: risk-adapted dose prescription and image-guided delivery. Strahlenther Onkol 2018; 194: 835-842 [PMID: 29696321 DOI: 10.1007/s00066-018-1306-2]
- 60 Orecchia R, Surgo A, Muto M, Ferrari A, Piperno G, Gerardi MA, Comi S, Garibaldi C, Ciardo D, Bazani A, Golino F, Pansini F, Fodor C, Romanelli P, Maestri D, Scroffi V, Mazza S, Jereczek-Fossa BA. VERO® radiotherapy for low burden cancer: 789 patients with 957 Lesions. Ecancermedicalscience 2016; 10: 677 [PMID: 27729942 DOI: 10.3332/ecancer.2016.677]
- Meduri B, Gregucci F, D'Angelo E, Alitto AR, Ciurlia E, Desideri I, Marino L, Borghetti P, Fiore M, 61 Fiorentino A; AIRO Giovani -Italian Association of Radiation Oncology-Young Members. Volume de-escalation in radiation therapy: state of the art and new perspectives. J Cancer Res Clin Oncol 2020; 146: 909-924 [PMID: 32072318 DOI: 10.1007/s00432-020-03152-7]
- Stauder MC, Miller RC. Stereotactic Body Radiation Therapy (SBRT) for Unresectable Pancreatic 62 Carcinoma. Cancers (Basel) 2010; 2: 1565-1575 [PMID: 24281173 DOI: 10.3390/cancers2031565]
- Zhong J, Patel K, Switchenko J, Cassidy RJ, Hall WA, Gillespie T, Patel PR, Kooby D, Landry J. 63 Outcomes for patients with locally advanced pancreatic adenocarcinoma treated with stereotactic body radiation therapy vs conventionally fractionated radiation. Cancer 2017; 123: 3486-3493 [PMID: 28493288 DOI: 10.1002/cncr.30706]



- 64 Pflughoeft KJ, Versalovic J. Human microbiome in health and disease. Annu Rev Pathol 2012; 7: 99-122 [PMID: 21910623 DOI: 10.1146/annurev-pathol-011811-132421]
- 65 Lepage P, Leclerc MC, Joossens M, Mondot S, Blottière HM, Raes J, Ehrlich D, Doré J. A metagenomic insight into our gut's microbiome. Gut 2013; 62: 146-158 [PMID: 22525886 DOI: 10.1136/gutjnl-2011-301805]



 \mathcal{O} W J

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2076-2087

DOI: 10.4251/wjgo.v13.i12.2076

ISSN 1948-5204 (online)

MINIREVIEWS

Prospect of lenvatinib for unresectable hepatocellular carcinoma in the new era of systemic chemotherapy

Takuya Sho, Kenichi Morikawa, Akinori Kubo, Yoshimasa Tokuchi, Takashi Kitagataya, Ren Yamada, Taku Shigesawa, Mugumi Kimura, Masato Nakai, Goki Suda, Mitsuteru Natsuizaka, Koji Ogawa, Naoya Sakamoto

ORCID number: Takuya Sho 0000-0001-7943-3842; Kenichi Morikawa 0000-0002-3891-5113: Akinori Kubo 0000-0003-3778-3531; Yoshimasa Tokuchi 0000-0001-6567-3722; Takashi Kitagataya 0000-0001-8833-8732; Ren Yamada 0000-0001-9693-4788; Taku Shigesawa 0000-0002-0124-802X; Mugumi Kimura 0000-0001-7427-4416; Masato Nakai 0000-0002-4860-1353; Goki Suda 0000-0003-0098-9106; Mitsuteru Natsuizaka 0000-0002-1819-1955; Koji Ogawa 0000-0002-9221-6815; Naoya Sakamoto 0000-0003-0061-059X.

Author contributions: Sho T and Morikawa K planned the contents of manuscript; the manuscript was drafted by Sho T and Morikawa K and was revised by all authors.

Conflict-of-interest statement:

Professor Kenichi Morikawa received research grants from Gilead Sciences, Inc., Bristol Myers Squibb, Otsuka Pharmaceutical Co., Ltd., and Takeda Pharmaceutical Co., Ltd. Professor Goki Suda received research grants from Bristol Myers Squibb and MSD K. K. Professor Naoya Sakamoto received lecture fees from Bristol Myers Squibb, Gilead Sciences, Inc., and MSD K. K., collaborative, funded research grants from Gilead Sciences, Inc. and Abbvie G. K., and research

Takuya Sho, Kenichi Morikawa, Akinori Kubo, Yoshimasa Tokuchi, Takashi Kitagataya, Ren Yamada, Taku Shigesawa, Mugumi Kimura, Masato Nakai, Goki Suda, Mitsuteru Natsuizaka, Koji Ogawa, Naoya Sakamoto, Department of Gastroenterology and Hepatology, Hokkaido University Faculty of Medicine and Graduate School of Medicine, Sapporo 060-8638, Hokkaido, Japan

Corresponding author: Kenichi Morikawa, MD, PhD, Assistant Professor, Department of Gastroenterology and Hepatology, Hokkaido University Faculty of Medicine and Graduate School of Medicine, Kita 15 Nishi 7, Kita-ku, Sapporo 060-8638, Hokkaido, Japan. kenichi.morikawa@med.hokudai.ac.jp

Abstract

The phase III clinical trial of the novel molecular targeted agent (MTA) lenvatinib for patients with advanced hepatocellular carcinoma (HCC) (REFLECT trial) found that lenvatinib was non-inferior to sorafenib in overall survival. Recently, the efficacy of multiple MTAs, including lenvatinib, in practice has been reported, and therapeutic strategies for Barcelona Clinic Liver Cancer (BCLC) intermediate stage HCC are undergoing major changes. Based on these results, lenvatinib could be recommended for patients with transcatheter arterial chemoembolization (TACE)-refractory, ALBI grade 1, within the up-to-seven criteria in the BCLC intermediate stage. Lenvatinib provides a more favorable outcome than TACE, even in cases with large or multinodular HCC beyond the up-to-seven criteria with Child-Pugh grade A. When patients meet the definitions of TACE-refractory or TACE-unsuitable, switching to systemic chemotherapy, including lenvatinib, is for favorable for preserving liver function. If initial treatment, including MTA, has a significant therapeutic effect and downstaging of HCC is obtained, additional TACE or surgical resection should be considered. Lenvatinib also has a therapeutic effect for poorly differentiated type and non-simple nodular type HCC thanks to the survival-prolonging effect of this drug. Furthermore, a significant therapeutic effect is expected in tumors with more than 50% liver involvement or main portal vein invasion, which have traditionally been considered to have a poor prognosis in patients. This suggests that at the start of lenvatinib treatment, HCC patients with ALBI grade 1 may be able to maintain liver functional reserve.

Key Words: Hepatocellular carcinoma; Lenvatinib; Molecular targeted agent; TACE-



grants from Bristol Myers Squibb, MSD K. K., Otsuka Pharmaceutical Co., Ltd., and Shionogi & Co., Ltd. Takuya Sho, Akinori Kubo, Ren Yamada, Takashi Kitagataya, Taku Shigesawa, Megumi Kimura, Masato Nakai, Goki Suda, Mitsuteru Natsuizaka, and Koji Ogawa declare that they have no conflicts of interest.

Supported by the Japan Agency for Medical Research and

Development (AMED), No. JP20fk0210053, JP20fk0310103, JP20fk0210072, JP20fk0210056, JP20fk0310101 and 20fk0210067; JSPS KAKENHI Grant Number No. JP20K08371 and JP19K18956.

Country/Territory of origin: Japan

Specialty type: Oncology

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): D Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: March 29, 2021 Peer-review started: March 29, 2021 First decision: June 16, 2021 Revised: July 8, 2021 Accepted: September 16, 2021 Article in press: September 16, 2021 Published online: December 15, 2021

refractory; TACE-unsuitable; Barcelona Clinic Liver Cancer intermediate stage

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: For about 10 years, first-line systemic chemotherapy for patients with advanced hepatocellular carcinoma (HCC) had been limited to sorafenib. The Phase III clinical trial of lenvatinib for patients with advanced HCC showed lenvatinib to be non-inferior to sorafenib with respect to overall survival (OS). The OS of patients is still far from satisfactory, and there is a great unmet medical need for more effective therapies. This review focuses on the current understanding of the therapeutic efficacy and safety of lenvatinib in the world and outlines the role of lenvatinib in the new era of chemotherapy for HCC.

Citation: Sho T, Morikawa K, Kubo A, Tokuchi Y, Kitagataya T, Yamada R, Shigesawa T, Kimura M, Nakai M, Suda G, Natsuizaka M, Ogawa K, Sakamoto N. Prospect of lenvatinib for unresectable hepatocellular carcinoma in the new era of systemic chemotherapy. World J Gastrointest Oncol 2021; 13(12): 2076-2087

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2076.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2076

INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the most common solid cancers and a major cause of cancer-related deaths globally[1]. According to the Global Cancer Observatory in 2020, HCC is ranked third in mortality, causing over 830000 deaths per year. Despite increasing global incidence as a major cause of cancer death, the development of new anticancer drugs for HCC has been inadequate. Traditionally, HCC has a poor prognosis. However, this might be partly due to the confined treatment options for patients with advanced HCC[2,3].

Since the publication of the practice guidelines of the American Association for the Study of Liver Diseases (AASLD) on the management of HCC in 2005, the Barcelona Clinic Liver Cancer (BCLC) staging system has been widely accepted and is also being used in many clinical trials of new drugs to treat HCC. These take into account factors including tumor burden, liver function, and general health conditions to determine prognosis and the best treatment. Accordingly, patients at an early stage are those with HCC \leq 5 cm or up to three nodules \leq 3 cm each (BCLC stage A). Patients exceeding these limits, without vascular invasion or extrahepatic spread, fit into the intermediate stage (BCLC stage B). Patients with evidence of a performance status ≤ 2 or an aggressive tumor pattern (vascular invasion or extrahepatic spread) correspond to the advanced stage (BCLC stage C)[4].

Systemic chemotherapy is the only therapeutic option for patients with Child-Pugh grade A at BCLC stage B with unresectable HCC and stage C. Prior to the development of the molecular targeted agent (MTA), patients in the advanced stage had a survival time of approximately 6 mo. Systemic chemotherapy for HCC has changed since the introduction of MTA sorafenib in 2007. The SHARP trial demonstrated that sorafenib prolonged median overall survival (OS) compared to placebo in patients who had not received systemic chemotherapy [10.7 mo vs 7.9 mo, hazard ratio (HR) = 0.69, 95%CI: 0.55-0.87, P < 0.001][5]. The subsequent Asia-Pacific trial confirmed these results in Asian patients[6]. Therapeutic options for extrahepatic metastases (e.g., lung, lymph node, or bone) and vascular invasion (e.g., portal vein tumor thrombus) have been demonstrated, and relatively long survival has been achieved for patients with BCLC stage C.

However, sorafenib does not shrink or induce necrosis in tumors and has relatively severe adverse events (AEs), including hand-foot-skin reactions. Therefore, the development of a novel MTA that can substitute for sorafenib is much anticipated.

For the past 10 years, sorafenib has been the only available first-line systemic chemotherapy for patients with advanced HCC. Many clinical trials of candidate systemic chemotherapeutic agents for advanced HCC have failed to demonstrate superiority or non-inferiority to sorafenib[7-9]. The phase III clinical trial of the



P-Reviewer: Antwi SO, Madian A S-Editor: Chang KL L-Editor: A P-Editor: Yu HG



lenvatinib for patients with advanced HCC (REFLECT trial)[10] showed lenvatinib to be non-inferior to sorafenib with respect to OS (13.6 mo vs 12.3 mo, HR = 0.92, 95%CI: 0.79-1.06). Furthermore, the secondary efficacy endpoints [progression-free survival (PFS) and objective response rate (ORR)] in the lenvatinib group showed a significant improvement compared with sorafenib. Based on the REFLECT trial, lenvatinib has been approved in the United States, European Union, and other countries as a first-line treatment option alongside sorafenib for advanced HCC, making it the first such drug to be used in Japan. A recent Phase III trial (IMbrave150) showed that combination immunotherapy with atezolizumab plus bevacizumab improved outcomes, including OS, PFS, ORR, and disease control rate, compared with sorafenib monotherapy[11]. Based on these results, both the 2020 AASLD and the 2021 European Society for Medical Oncology liver treatment options depends on BCLC staging and treatment guidelines which recommended atezolizumab+bevacizumab as first-line systemic therapy in stage B with transcatheter arterial chemoembolization (TACE)-unsuitable HCC and stage C. In addition, even with the latest version of the treatment algorithm in the Clinical Practice Guidelines for HCC 2020 in Japan, the first-line drug therapy for unresectable HCC is atezolizumab+bevacizumab combination therapy. Sorafenib and lenvatinib, which were previously the first-line treatments, are now second-line treatments. Regorafenib, ramucirumab, and cabozantinib can be used as third-line treatments (Figure 1).

Although various systemic chemotherapies are available for HCC, the OS of patients is still far from satisfactory, and there is a great unmet medical need for more effective therapies. Furthermore, in the future, issues are likely to arise regarding the order and combination of treatment options for HCC.

This review focuses on the current understanding of the therapeutic efficacy and safety of lenvatinib and outlines the role of lenvatinib in the new era of chemotherapy for HCC.

MOLECULAR MECHANISMS OF LENVATINIB

Lenvatinib is an MTA that suppresses vascular endothelial growth factor receptor (VEGFR), fibroblast growth factor receptor (FGFR), proto-oncogene tyrosine-protein kinase receptor RET, platelet-derived growth factor receptor α , and stem cell factor receptor^[12]. Because these targets act as drivers in cancer, lenvatinib has been reported to exhibit antitumor and immunomodulatory activities in a variety of preclinical cancer models[13].

Inhibition of the VEGF/VEGFR signaling pathway

Angiogenesis, mostly regulated by the VEGF pathway, is an essential event in tumor growth and metastasis[14]. In particular, VEGFR-2 is a high-affinity VEGF receptor in vascular endothelial cells[15,16]. Ligand binding activates certain signaling pathways, such as the phospholipase-Cy, phosphoinositide 3-kinase (PI3K)/V-akt murine thymoma viral oncogene homolog (AKT) pathways, and rat sarcoma (Ras)/mitogenactivated protein kinase (MAPK)[16]. These signaling pathways have been implicated in endothelial cells and vascular permeability during tumor enlargement[17]. Moreover, VEGFR is highly expressed in HCC cells. Lenvatinib has been shown to inhibit tumor angiogenesis in various preclinical models. In patient-derived and PLC/PRF/5 cell-transplanted tumor models, Lenvatinib administration resulted in a reduction in microvessel density of tumor^[18]. In addition, lenvatinib had been found to suppress various types of cancers[19-23], by blocking the VEGFR pathway. These data indicate that lenvatinib exhibits potent anti-angiogenic activity and may have a stronger effect than sorafenib in preclinical models.

Inhibition of the FGF/FGFR signaling pathway

Activated fibroblast growth factor (FGF) signaling can directly facilitate cell proliferation and survival, as well as promote tumor angiogenesis and progression[24]. The binding of FGF to FGFR leads to the activation of the RAS/MAPK and PI3K/AKT signaling pathways [25,26]. FGF and FGFR are typically overexpressed in HCC, and the expression of FGF19/FGFR4 contributes to HCC progression[27]. Analysis of the effects of selective FGFR inhibitors and FGFR small interfering RNAs on cancer stemlike cells (CSCs) in HCC showed that lenvatinib diminished CSCs in HCC by inhibiting FGFR1-3 signaling; however, FGFR4 signaling was not affected. FGF2 and FGF19 are involved in maintaining CD44High/CD133High CSCs in HCC, potentially via FGFR1-3[28]. Preclinical studies have shown that lenvatinib inhibits the prolif-





Figure 1 Treatment strategy for intermediate stage hepatocellular carcinoma. HCC: Epatocellular carcinoma; MWA: Microwave ablation; ORR: Objective response rate; PFS: Progression-free survival; RFA: Radiofrequency ablation; TACE: Transcatheter arterial chemoembolization.

eration of FGF19 and FGFR-overexpressing cell lines[18,28,29].

Inhibition of the RET signaling pathway

RET activates downstream signaling pathways through mutations or chromosome rearrangements, promoting tumor cell growth[30]. The autophosphorylation of specific tyrosine residues of RET allows for the recruitment adaptor proteins that connect the RET receptor to RAS/MAPK and PI3K/AKT signaling pathways, thereby promoting cell growth, proliferation, survival, and differentiation[31]. Lenvatinib can inhibit cell proliferation by blocking RET autophosphorylation[12].

Immunomodulatory activity

The immune escape of tumor cells is the main mechanism of tumorigenesis[32]. It has been suggested that VEGF-A has immunosuppressive properties[33]. VEGF-A produced by tumor cells enhances the expression of immunosuppressive receptors in CD8+ T cells and promotes immune escape[34]. Immune inhibitory receptors cause CD8+ T cell exhaustion by recognizing tumor antigens[32]. Lenvatinib reduced the infiltration of tumor-associated macrophages and increased the percentage of activated CD8+ T cells in HCC[13].

LENVATINIB THERAPY IN INTERMEDIATE STAGE HCC

Efficacy of Lenvatinib in intermediate stage HCC

In recent years, the efficacy of multiple MTAs, including lenvatinib, has been reported. As a result, therapeutic strategies for BCLC intermediate-stage HCC are undergoing major changes. In the REFLECT trial, the ORR of lenvatinib was 40.6% in the mRECIST evaluation, and in the sub-analysis, the ORR of BCLC intermediate stage HCC in the Japanese population was 61.3%[35]. Furthermore, Kudo et al[36] reported that a very high response rate (RR) of 73.3% was obtained for Child-Pugh A in BCLC intermediate stage HCC. Tomonari et al[37] reported from their real-world data that BCLC intermediate stage HCC cases had fewer AEs, could maintain the dosage amount of lenvatinib, and had a good therapeutic effect. Many reports have demonstrated the high therapeutic effect of lenvatinib in BCLC intermediate stage HCC.

Efficacy of TACE in intermediate stage HCC

TACE is the guideline-recommended standard of care for intermediate stage HCC. The AASLD Consensus Conference showed that locoregional TACE may still be the best approach if the patient's tumor volume is small and nodules can be accessed



superselectively^[38]. According to a systematic review of patients treated with lipiodol-based TACE, the survival rates at one, three, and five years were reported to be 70.3%, 40.4%, and 32.4%, respectively, with a median OS of 19.4 mo[39]. But, the efficacy of TACE was not observed equally in all cases. Intermediate-stage HCC varies widely in terms of tumor burden and liver function. The heterogeneity of tumors in the BCLC intermediate stage has led to the development of several prognostic scores, including the Kinki criteria, which are based on liver function and tumor burden that attempt to determine who may obtain the maximum benefit from TACE[40]. The upto-seven criteria, defined as the sum of the maximum tumor diameter in the liver (cm) and the number of tumors[41], has prognostic value and can predict the recurrence and maintenance of Child-Pugh grade in patients who undergo initial conventional TACE^[42]. In intermediate stage HCC, preserving liver function is as important as achieving a high objective response because the treatment goal is to prolong OS. Additional TACE has a lower RR than the initial TACE and increases the risk of liver function loss[43]. Hence, repeated TACE is not recommended, as it leads to decreased liver function and reduced therapeutic efficacy.

Recently, the characteristics of TACE-resistant tumors, which are prone to being TACE-refractory and to exacerbate to Child-Pugh grade B by TACE, have been clarified[36,44]. In cases in which the tumor is a non-simple nodular type, occupying multiple lobes, a large tumor mass, such as beyond the up-to-seven criteria, and of a histopathologically poorly differentiated type, TACE has little effect. As a result, these cases may become TACE-refractory at an early stage[45].

EFFICACY OF LENVATINIB IN TACE-REFRACTORY CASES

Therapeutic alternatives are needed for patients who are TACE-refractory. Repeated TACE could worsen liver function, thereby narrowing the time window for a switch to MTAs, which is recommended for patients with Child-Pugh grade A. In the era of sorafenib, TACE failure/refractoriness was proposed for switching to systemic chemotherapy[46]. The OPTIMIS trial showed that the survival time in TACErefractory patients was longer in patients who received sorafenib than in those who continued TACE.

A recent study showed that the median PFS times in TACE-refractory patients treated with lenvatinib, sorafenib, and TACE was 5.8, 3.2, and 2.4 mo, respectively [47]. In a Cox regression analysis, lenvatinib treatment and being within the up-toseven criteria were identified as independent factors for PFS (lenvatinib, P < 0.0001; within the up-to-seven criteria, P = 0.001). Similarly, decision-tree analysis showed that patients treated with ALBI grade 1 beyond the up-to-seven criteria had longer PFS than patients treated with ALBI grade 2 beyond the up-to-seven criteria. Therefore, lenvatinib could be recommended to patients with TACE-refractory, ALBI grade 1, and within the up-to-seven criteria in the BCLC intermediate stage. Thus, treatment with lenvatinib could give rise to good outcomes in TACE-refractory intermediatestage HCC patients.

EFFICACY OF LENVATINIB IN TACE-UNSUITABLE CASES

In patients beyond the up-to-seven criteria, TACE treatment was reported to be likely to worsen liver function, potentially resulting in losing the opportunity to be treated with MTA[42,48]. Recently, the Asia-Pacific Primary Liver Cancer Expert (APPLE) consensus statement proposed the criteria for TACE unsuitability[49].

A proof-of-concept study demonstrated that, in patients with intermediate stage HCC who exceeded the up-to-seven criteria, the lenvatinib group showed a significantly higher ORR (73.3% vs 33.3%; P < 0.001) and a significantly longer median PFS than the conventional TACE group (16.0 mo vs 3.0 mo; P < 0.001)[50]. The ALBI score was maintained in the lenvatinib group during treatment, whereas it worsened in the TACE group. Therefore, in the case of large or multinodular intermediate stage HCC with Child-Pugh grade A, lenvatinib provides a better outcome than TACE.

LENVATINIB AS AN UPFRONT SYSTEMIC CHEMOTHERAPY

When patients meet the definitions of TACE-refractory or TACE-unsuitable, switching



to systemic chemotherapy including lenvatinib may be advisable in order to preserve liver function. Upfront lenvatinib therapy may also be suitable for patients with a high tumor burden or those who are considered TACE-refractory. If the tumor responds well and downstaging is possible with lenvatinib, additional TACE (or surgical resection or ablation) could be considered. However, the efficacy and safety of this strategy have not yet been validated.

Recently, a randomized, controlled trial comparing the efficacy and safety of TACE plus sorafenib to TACE alone (TACTICS trial) found that median PFS was significantly longer in the TACE plus sorafenib group than in the TACE alone group (25.2 mo vs 13.5 mo; P = 0.006 [51]. MTA treatment is thought to improve the clinical outcome of TACE by promoting the normalization of tumor vessels and contributing to a higher density of lipiodol deposition^[49]. Lenvatinib has a higher RR than sorafenib, suggesting that Len-TACE sequential therapy may be a promising treatment for patients with moderately differentiated HCC.

EFFICACY OF LENVATINIB FOR POORLY DIFFERENTIATED HCC

In the REFLECT study, poorly differentiated HCC was diagnosed in 42 patients (4.4%), with 21 in the lenvatinib group and 21 in the sorafenib group. The ORR was 47.6% in the lenvatinib group and 14.3% in the sorafenib group. Thus, lenvatinib is expected to have an effect on poorly differentiated HCC[49].

HCC shows great diversity of tumor differentiation even in the same nodule, and it is difficult to examine the heterogeneity of all tumors by liver tumor biopsy. Therefore, it is extremely important to evaluate the degree of tumor differentiation using a noninvasive method. Poorly differentiated HCC is characterized by tumors that show a heterogeneous enhancement pattern with irregularly shaped ring structures in the arterial phase of dynamic computed tomography (CT)[52] and tumors in which lesions are detected on FDG-positron emission tomography [53]. Based on the prediction of tumor differentiation by the enhancement pattern of dynamic CT[54], the therapeutic effect of lenvatinib is also recognized for poorly differentiated type and non-simple nodular type, and it has been reported that it has a survival-prolonging effect for all of them [54]. Tumors with a heterogeneous enhancement pattern and irregularly shaped ring structures had an RR of 84%, which was significantly better than the RR of tumors with a homogeneous enhancement pattern, which was 53%. Furthermore, there was no significant difference in the PFS between the two groups. The therapeutic effect of lenvatinib, regardless of the degree of tumor differentiation, will have a significant impact on future HCC treatment[54].

Traditionally, non-simple nodular, poorly differentiated HCC has a poor prognosis with existing treatments, including hepatectomy. In particular, it was difficult to control the progression of poorly differentiated HCC using TACE. Therefore, these factors are considered to be among those that are TACE-unsuitable. It is important to identify TACE-unsuitable cases even during the course of TACE treatment and to identify Len-suitable cases while maintaining hepatic reserve.

IMPACT OF LENVATINIB ON PRESERVING LIVER FUNCTION

A sub-analysis of the REFLECT trial examined the time to progression of Child-Pugh score 6 before lenvatinib treatment to a Child-Pugh score of 7. The median time to progression to Child-Pugh score 7 was 23.7 mo in the sorafenib group, 23.9 mo in the lenvatinib 8 mg group, and 15.9 mo in the lenvatinib 12 mg group, respectively. There was no significant difference between the sorafenib and lenvatinib groups. Both sorafenib and lenvatinib affected liver functional reserve, but the difference was not significant[35].

Terashima et al^[55] reported that patients treated with lenvatinib had a Child-Pugh score that was maintained or improved after 4 and 12 wk compared with those treated with sorafenib (P = 0.048 and P = 0.036, respectively) in clinical settings. Lenvatinib was identified as one of the factors associated with maintaining Child-Pugh scores. On multivariate analysis, a worse Child-Pugh score after 4 wk was an independent predictor of poor OS. Patients treated with lenvatinib for advanced HCC maintained their liver functional reserves better than those treated with sorafenib.

Uchikawa et al[56] assessed the ALBI score as an index of liver function during sorafenib and lenvatinib treatment. The median ALBI score was -2.53 before MTA treatment and -2.45, -2.44, and -2.36 post-2, -4, and -6 mo, respectively. The ALBI



scores tended to increase during MTA treatment. When examined separately in the sorafenib and lenvatinib groups, no significant difference was observed between the two groups. However, the ALBI scores of the sorafenib group increased 2 mo after treatment initiation, and at 4 and 6 mo, significant differences were observed (P <0.01). Based on the above, although the ALBI score may gradually decrease with the course of MTA treatment, lenvatinib may have a lower effect on the deterioration of the ALBI score than sorafenib.

Hiraoka et al^[57] compared the ALBI score at the start of lenvatinib with scores after 2 and 4 wk; decreased liver function was common in the early stages after starting lenvatinib (within 4 wk, especially within 2 wk). It is important to introduce MTA to patients with as good liver function as possible, taking into account the early decrease in liver function due to lenvatinib. These results suggest that ALBI grade 1 and lenvatinib at the start of MTA treatment may be related to the maintenance of liver functional reserve.

TREATMENT OUTCOME FOR THOSE WHO DO NOT MEET THE REFLECT TRIAL ELIGIBILITY CRITERIA

In the REFLECT trial, patients were excluded if they had a treatment history of MTA, large HCC with more than 50% liver occupation, HCC with main portal vein invasion, Child–Pugh grade B, platelet count $< 75 \times 10^{\circ}/L$, or apparent bile duct invasion.

We reported the efficacy and safety of lenvatinib in patients with unresectable HCC who did not meet the REFLECT eligibility criteria in clinical settings [58,59]. The ORR and the median PFS was similar between patients who met the REFLECT inclusion criteria and those who did not. Thus, the study results support the use of lenvatinib for patients with unresectable HCC who do not meet the REFLECT inclusion criteria.

But, the efficacy and tolerability of lenvatinib treatment differed according to the eligibility criteria of the REFLECT trial.

Chuma et al[60] reported that lenvatinib treatment offers benefits in highly advanced HCC (tumors with more than 50% liver occupation or main portal vein invasion) patients with good liver function or nodular-type tumors. Maruta et al[61] also reported that lenvatinib had potential profits for patients with advanced HCC with second- or later-line therapies and a high burden of intrahepatic lesions. The various characteristics identified in these studies may be useful as indicators for lenvatinib treatment in highly advanced HCC cases, which are considered treatmentresistant cancers.

POST-PD TREATMENT AFTER LENVATINIB

In the REFLECT trial, among the 954 patients randomized to receive first-line lenvatinib (n = 478) or sorafenib (n = 476), 340 patients received subsequent anticancer medication during the survival follow-up period: 156 patients (32.6%) had received first-line lenvatinib, and 184 patients (38.7%) had received first-line sorafenib[62]. Of the patients who were treated with first-line lenvatinib, the most common subsequent carcinostatic substance was sorafenib (25.3%), and the most common subsequent nonanticancer medication treatment was TACE (56.6%). The OS of patients who were initially randomized to first-line lenvatinib (versus first-line sorafenib) and who received any subsequent anticancer medication was 20.8 mo vs 17.0 mo (HR = 0.87; 95%CI: 0.67-1.14). The OS of patients who initially received first-line lenvatinib (versus first-line sorafenib) and who did not receive any subsequent carcinostatic substance was 11.5 mo vs 9.1 mo (HR = 0.90, 95% CI: 0.75-1.09). In the -hoc analysis of all patients in the REFLECT study, the OS of those who received subsequent anticancer medication was prolonged compared with patients who did not receive any subsequent anticancer medication. In the REFLECT trial, the lenvatinib group had significantly longer PFS than the sorafenib group, but superiority in OS could not be demonstrated because of the effect of post-treatment on post-progression survival prolongation.

Lenvatinib is the preferred agent for TACE-unsuitable patients in the intermediate stage based on the high RR, survival benefit over TACE, and the possibility of conversion to resection or ablation therapy[49]. Sorafenib combined with surgical resection is a feasible option in advanced HCC patients; if sorafenib is effective, longterm survival may be achieved^[63]. Additional surgery was the most significant factor



predicting survival exceeding 3 years (P < 0.0001) and represents an independent prognostic factor (HR = 0.07, 95% CI: 0.003-0.40, P = 0.01)[64]. Long-term survival may be obtained for select patients with HCC receiving adequate additional surgical treatment, even after sorafenib induction. Therefore, lenvatinib treatment, which has a higher RR than sorafenib, is expected to increase the number of cases that can be switched to conversion therapy.

CONCLUSION

Many MTAs and immunotherapies have become available as treatment options for advanced HCC. Lenvatinib has been shown to have a good therapeutic effect, even in TACE-refractory and TACE-unsuitable cases. Intermediate-stage HCC beyond the upto-seven criteria with Child-Pugh grade A/ALBI grade 1 usually does not benefit from TACE, whereas lenvatinib provides a better outcome than TACE.

Lenvatinib also has good therapeutic performance even in cases of non-simple nodular, poorly differentiated, tumor masses with more than 50% involvement of the liver and main portal vein invasion, which are generally recognized as having a poor prognosis with existing treatments. To maximize the therapeutic effect of lenvatinib in such cases, it is necessary to preserve liver function in patients with Child-Pugh grade A and ALBI grade 1 at the start of treatment.

REFERENCES

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin 2021; 71: 209-249 [PMID: 33538338 DOI: 10.3322/caac.21660]
- Forner A, Llovet JM, Bruix J. Hepatocellular carcinoma. Lancet 2012; 379: 1245-1255 [PMID: 2 22353262 DOI: 10.1016/S0140-6736(11)61347-01
- 3 Ikeda M, Mitsunaga S, Shimizu S, Ohno I, Takahashi H, Okuyama H, Kuwahara A, Okusaka T. Current status of hepatocellular carcinoma in Japan. Chin Clin Oncol 2013; 2: 40 [PMID: 25841919 DOI: 10.3978/j.issn.2304-3865.2013.09.01]
- 4 Llovet JM, Brú C, Bruix J. Prognosis of hepatocellular carcinoma: the BCLC staging classification. Semin Liver Dis 1999; 19: 329-338 [PMID: 10518312 DOI: 10.1055/s-2007-1007122]
- Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, de Oliveira AC, Santoro A, Raoul 5 JL, Forner A, Schwartz M, Porta C, Zeuzem S, Bolondi L, Greten TF, Galle PR, Seitz JF, Borbath I, Häussinger D, Giannaris T, Shan M, Moscovici M, Voliotis D, Bruix J; SHARP Investigators Study Group. Sorafenib in advanced hepatocellular carcinoma. N Engl J Med 2008; 359: 378-390 [PMID: 18650514 DOI: 10.1056/NEJMoa0708857]
- 6 Cheng AL, Kang YK, Chen Z, Tsao CJ, Qin S, Kim JS, Luo R, Feng J, Ye S, Yang TS, Xu J, Sun Y, Liang H, Liu J, Wang J, Tak WY, Pan H, Burock K, Zou J, Voliotis D, Guan Z. Efficacy and safety of sorafenib in patients in the Asia-Pacific region with advanced hepatocellular carcinoma: a phase III randomised, double-blind, placebo-controlled trial. Lancet Oncol 2009; 10: 25-34 [PMID: 19095497 DOI: 10.1016/S1470-2045(08)70285-7]
- Cainap C, Qin S, Huang WT, Chung IJ, Pan H, Cheng Y, Kudo M, Kang YK, Chen PJ, Toh HC, 7 Gorbunova V, Eskens FA, Qian J, McKee MD, Ricker JL, Carlson DM, El-Nowiem S. Linifanib vs Sorafenib in patients with advanced hepatocellular carcinoma: results of a randomized phase III trial. J Clin Oncol 2015; 33: 172-179 [PMID: 25488963 DOI: 10.1200/JCO.2013.54.3298]
- Cheng AL, Kang YK, Lin DY, Park JW, Kudo M, Qin S, Chung HC, Song X, Xu J, Poggi G, Omata M, Pitman Lowenthal S, Lanzalone S, Yang L, Lechuga MJ, Raymond E. Sunitinib vs sorafenib in advanced hepatocellular cancer: results of a randomized phase III trial. J Clin Oncol 2013; 31: 4067-4075 [PMID: 24081937 DOI: 10.1200/JCO.2012.45.8372]
- Johnson PJ, Qin S, Park JW, Poon RT, Raoul JL, Philip PA, Hsu CH, Hu TH, Heo J, Xu J, Lu L, Chao Y, Boucher E, Han KH, Paik SW, Robles-Aviña J, Kudo M, Yan L, Sobhonslidsuk A, Komov D, Decaens T, Tak WY, Jeng LB, Liu D, Ezzeddine R, Walters I, Cheng AL. Brivanib vs sorafenib as first-line therapy in patients with unresectable, advanced hepatocellular carcinoma: results from the randomized phase III BRISK-FL study. J Clin Oncol 2013; 31: 3517-3524 [PMID: 23980084 DOI: 10.1200/JCO.2012.48.4410
- 10 Kudo M, Finn RS, Qin S, Han KH, Ikeda K, Piscaglia F, Baron A, Park JW, Han G, Jassem J, Blanc JF, Vogel A, Komov D, Evans TRJ, Lopez C, Dutcus C, Guo M, Saito K, Kraljevic S, Tamai T, Ren M, Cheng AL. Lenvatinib vs sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomised phase 3 non-inferiority trial. Lancet 2018; 391: 1163-1173 [PMID: 29433850 DOI: 10.1016/S0140-6736(18)30207-1]
- Finn RS, Qin S, Ikeda M, Galle PR, Ducreux M, Kim TY, Kudo M, Breder V, Merle P, Kaseb AO, Li D, Verret W, Xu DZ, Hernandez S, Liu J, Huang C, Mulla S, Wang Y, Lim HY, Zhu AX, Cheng



AL; IMbrave150 Investigators. Atezolizumab plus Bevacizumab in Unresectable Hepatocellular Carcinoma. N Engl J Med 2020; 382: 1894-1905 [PMID: 32402160 DOI: 10.1056/NEJMoa1915745]

- 12 Tohyama O, Matsui J, Kodama K, Hata-Sugi N, Kimura T, Okamoto K, Minoshima Y, Iwata M, Funahashi Y. Antitumor activity of lenvatinib (e7080): an angiogenesis inhibitor that targets multiple receptor tyrosine kinases in preclinical human thyroid cancer models. J Thyroid Res 2014; 2014: 638747 [PMID: 25295214 DOI: 10.1155/2014/638747]
- 13 Kato Y, Tabata K, Kimura T, Yachie-Kinoshita A, Ozawa Y, Yamada K, Ito J, Tachino S, Hori Y, Matsuki M, Matsuoka Y, Ghosh S, Kitano H, Nomoto K, Matsui J, Funahashi Y. Lenvatinib plus anti-PD-1 antibody combination treatment activates CD8+ T cells through reduction of tumor-associated macrophage and activation of the interferon pathway. PLoS One 2019; 14: e0212513 [PMID: 30811474 DOI: 10.1371/journal.pone.0212513]
- 14 Hicklin DJ, Ellis LM. Role of the vascular endothelial growth factor pathway in tumor growth and angiogenesis. J Clin Oncol 2005; 23: 1011-1027 [PMID: 15585754 DOI: 10.1200/JCO.2005.06.081]
- 15 Ferrara N. Vascular endothelial growth factor as a target for anticancer therapy. Oncologist 2004; 9 Suppl 1: 2-10 [PMID: 15178810 DOI: 10.1634/theoncologist.9-suppl 1-2]
- Koch S, Claesson-Welsh L. Signal transduction by vascular endothelial growth factor receptors. Cold 16 Spring Harb Perspect Med 2012; 2: a006502 [PMID: 22762016 DOI: 10.1101/cshperspect.a006502]
- 17 Takahashi H, Shibuya M. The vascular endothelial growth factor (VEGF)/VEGF receptor system and its role under physiological and pathological conditions. Clin Sci (Lond) 2005; 109: 227-241 [PMID: 16104843 DOI: 10.1042/CS20040370]
- 18 Matsuki M, Hoshi T, Yamamoto Y, Ikemori-Kawada M, Minoshima Y, Funahashi Y, Matsui J. Lenvatinib inhibits angiogenesis and tumor fibroblast growth factor signaling pathways in human hepatocellular carcinoma models. Cancer Med 2018; 7: 2641-2653 [PMID: 29733511 DOI: 10.1002/cam4.1517]
- 19 Matsuki M, Adachi Y, Ozawa Y, Kimura T, Hoshi T, Okamoto K, Tohyama O, Mitsuhashi K, Yamaguchi A, Matsui J, Funahashi Y. Targeting of tumor growth and angiogenesis underlies the enhanced antitumor activity of lenvatinib in combination with everolimus. Cancer Sci 2017; 108: 763-771 [PMID: 28107584 DOI: 10.1111/cas.13169]
- 20 Yamamoto Y, Matsui J, Matsushima T, Obaishi H, Miyazaki K, Nakamura K, Tohyama O, Semba T, Yamaguchi A, Hoshi SS, Mimura F, Haneda T, Fukuda Y, Kamata JI, Takahashi K, Matsukura M, Wakabayashi T, Asada M, Nomoto KI, Watanabe T, Dezso Z, Yoshimatsu K, Funahashi Y, Tsuruoka A. Lenvatinib, an angiogenesis inhibitor targeting VEGFR/FGFR, shows broad antitumor activity in human tumor xenograft models associated with microvessel density and pericyte coverage. Vasc Cell 2014; 6: 18 [PMID: 25197551 DOI: 10.1186/2045-824X-6-18]
- 21 Matsui J, Yamamoto Y, Funahashi Y, Tsuruoka A, Watanabe T, Wakabayashi T, Uenaka T, Asada M. E7080, a novel inhibitor that targets multiple kinases, has potent antitumor activities against stem cell factor producing human small cell lung cancer H146, based on angiogenesis inhibition. Int J Cancer 2008; 122: 664-671 [PMID: 17943726 DOI: 10.1002/ijc.23131]
- Matsui J, Funahashi Y, Uenaka T, Watanabe T, Tsuruoka A, Asada M. Multi-kinase inhibitor E7080 suppresses lymph node and lung metastases of human mammary breast tumor MDA-MB-231 via inhibition of vascular endothelial growth factor-receptor (VEGF-R) 2 and VEGF-R3 kinase. Clin Cancer Res 2008; 14: 5459-5465 [PMID: 18765537 DOI: 10.1158/1078-0432.CCR-07-5270]
- Ferrari SM, Bocci G, Di Desidero T, Elia G, Ruffilli I, Ragusa F, Orlandi P, Paparo SR, Patrizio A, 23 Piaggi S, La Motta C, Ulisse S, Baldini E, Materazzi G, Miccoli P, Antonelli A, Fallahi P. Lenvatinib exhibits antineoplastic activity in anaplastic thyroid cancer in vitro and in vivo. Oncol Rep 2018; 39: 2225-2234 [PMID: 29517103 DOI: 10.3892/or.2018.6306]
- Turner N, Grose R. Fibroblast growth factor signalling: from development to cancer. Nat Rev Cancer 24 2010; 10: 116-129 [PMID: 20094046 DOI: 10.1038/nrc2780]
- 25 Eswarakumar VP, Lax I, Schlessinger J. Cellular signaling by fibroblast growth factor receptors. Cytokine Growth Factor Rev 2005; 16: 139-149 [PMID: 15863030 DOI: 10.1016/j.cytogfr.2005.01.001]
- Gotoh N. Regulation of growth factor signaling by FRS2 family docking/scaffold adaptor proteins. 26 Cancer Sci 2008; 99: 1319-1325 [PMID: 18452557 DOI: 10.1111/j.1349-7006.2008.00840.x]
- 27 Raja A, Park I, Haq F, Ahn SM. FGF19-FGFR4 Signaling in Hepatocellular Carcinoma. Cells 2019; 8 [PMID: 31167419 DOI: 10.3390/cells8060536]
- 28 Shigesawa T, Maehara O, Suda G, Natsuizaka M, Kimura M, Shimazaki T, Yamamoto K, Yamada R, Kitagataya T, Nakamura A, Suzuki K, Ohara M, Kawagishi N, Umemura M, Nakai M, Sho T, Morikawa K, Ogawa K, Ohnishi S, Sugiyama M, Mizokami M, Takeda H, Sakamoto N. Lenvatinib suppresses cancer stem-like cells in HCC by inhibiting FGFR1-3 signaling, but not FGFR4 signaling. Carcinogenesis 2021; 42: 58-69 [PMID: 32449510 DOI: 10.1093/carcin/bgaa049]
- Ogasawara S, Mihara Y, Kondo R, Kusano H, Akiba J, Yano H. Antiproliferative Effect of 29 Lenvatinib on Human Liver Cancer Cell Lines In Vitro and In Vivo. Anticancer Res 2019; 39: 5973-5982 [PMID: 31704822 DOI: 10.21873/anticanres.13802]
- 30 Romei C, Ciampi R, Elisei R. A comprehensive overview of the role of the RET proto-oncogene in thyroid carcinoma. Nat Rev Endocrinol 2016; 12: 192-202 [PMID: 26868437 DOI: 10.1038/nrendo.2016.11]
- Arighi E, Borrello MG, Sariola H. RET tyrosine kinase signaling in development and cancer. Cytokine Growth Factor Rev 2005; 16: 441-467 [PMID: 15982921 DOI: 10.1016/j.cytogfr.2005.05.010]



- 32 Schreiber RD, Old LJ, Smyth MJ. Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. Science 2011; 331: 1565-1570 [PMID: 21436444 DOI: 10.1126/science.1203486]
- 33 Voron T, Marcheteau E, Pernot S, Colussi O, Tartour E, Taieb J, Terme M. Control of the immune response by pro-angiogenic factors. Front Oncol 2014; 4: 70 [PMID: 24765614 DOI: 10.3389/fonc.2014.00070]
- Voron T, Colussi O, Marcheteau E, Pernot S, Nizard M, Pointet AL, Latreche S, Bergaya S, 34 Benhamouda N, Tanchot C, Stockmann C, Combe P, Berger A, Zinzindohoue F, Yagita H, Tartour E, Taieb J, Terme M. VEGF-A modulates expression of inhibitory checkpoints on CD8+ T cells in tumors. J Exp Med 2015; 212: 139-148 [PMID: 25601652 DOI: 10.1084/jem.20140559]
- Yamashita T, Kudo M, Ikeda K, Izumi N, Tateishi R, Ikeda M, Aikata H, Kawaguchi Y, Wada Y, 35 Numata K, Inaba Y, Kuromatsu R, Kobayashi M, Okusaka T, Tamai T, Kitamura C, Saito K, Haruna K, Okita K, Kumada H. REFLECT-a phase 3 trial comparing efficacy and safety of lenvatinib to sorafenib for the treatment of unresectable hepatocellular carcinoma: an analysis of Japanese subset. J Gastroenterol 2020; 55: 113-122 [PMID: 31720835 DOI: 10.1007/s00535-019-01642-1]
- Kudo M. Extremely High Objective Response Rate of Lenvatinib: Its Clinical Relevance and -36 Changing the Treatment Paradigm in Hepatocellular Carcinoma. Liver Cancer 2018; 7: 215-224 [PMID: 30319981 DOI: 10.1159/000492533]
- Tomonari T, Sato Y, Tanaka H, Tanaka T, Fujino Y, Mitsui Y, Hirao A, Taniguchi T, Okamoto K, 37 Sogabe M, Miyamoto H, Muguruma N, Kagiwada H, Kitazawa M, Fukui K, Horimoto K, Takayama T. Potential use of lenvatinib for patients with unresectable hepatocellular carcinoma including after treatment with sorafenib: Real-world evidence and in vitro assessment via protein phosphorylation array. Oncotarget 2020; 11: 2531-2542 [PMID: 32655838 DOI: 10.18632/oncotarget.27640]
- 38 Llovet JM, Villanueva A, Marrero JA, Schwartz M, Meyer T, Galle PR, Lencioni R, Greten TF, Kudo M, Mandrekar SJ, Zhu AX, Finn RS, Roberts LR; AASLD Panel of Experts on Trial Design in HCC. Trial Design and Endpoints in Hepatocellular Carcinoma: AASLD Consensus Conference. Hepatology 2021; 73 Suppl 1: 158-191 [PMID: 32430997 DOI: 10.1002/hep.31327]
- 39 Lencioni R, de Baere T, Soulen MC, Rilling WS, Geschwind JF. Lipiodol transarterial chemoembolization for hepatocellular carcinoma: A systematic review of efficacy and safety data. Hepatology 2016; 64: 106-116 [PMID: 26765068 DOI: 10.1002/hep.28453]
- 40 Kudo M, Arizumi T, Ueshima K, Sakurai T, Kitano M, Nishida N. Subclassification of BCLC B Stage Hepatocellular Carcinoma and Treatment Strategies: Proposal of Modified Bolondi's Subclassification (Kinki Criteria). Dig Dis 2015; 33: 751-758 [PMID: 26488473 DOI: 10.1159/000439290]
- Mazzaferro V, Llovet JM, Miceli R, Bhoori S, Schiavo M, Mariani L, Camerini T, Roayaie S, 41 Schwartz ME, Grazi GL, Adam R, Neuhaus P, Salizzoni M, Bruix J, Forner A, De Carlis L, Cillo U, Burroughs AK, Troisi R, Rossi M, Gerunda GE, Lerut J, Belghiti J, Boin I, Gugenheim J, Rochling F, Van Hoek B, Majno P; Metroticket Investigator Study Group. Predicting survival after liver transplantation in patients with hepatocellular carcinoma beyond the Milan criteria: a retrospective, exploratory analysis. Lancet Oncol 2009; 10: 35-43 [PMID: 19058754 DOI: 10.1016/S1470-2045(08)70284-5
- 42 Yasui Y, Tsuchiya K, Kurosaki M, Takeguchi T, Takeguchi Y, Okada M, Wang W, Kubota Y, Goto T, Komiyama Y, Higuchi M, Takaura K, Hayashi T, Takada H, Tamaki N, Nakanishi H, Itakura J, Takahashi Y, Asahina Y, Enomoto N, Himeno Y, Izumi N. Up-to-seven criteria as a useful predictor for tumor downstaging to within Milan criteria and Child-Pugh grade deterioration after initial conventional transarterial chemoembolization. Hepatol Res 2018; 48: 442-450 [PMID: 29278654 DOI: 10.1111/hepr.13048]
- 43 Golfieri R, Renzulli M, Mosconi C, Forlani L, Giampalma E, Piscaglia F, Trevisani F, Bolondi L; Bologna Liver Oncology Group (BLOG). Hepatocellular carcinoma responding to superselective transarterial chemoembolization: an issue of nodule dimension? J Vasc Interv Radiol 2013; 24: 509-517 [PMID: 23428355 DOI: 10.1016/j.jvir.2012.12.013]
- 44 Kudo M. A New Treatment Option for Intermediate-Stage Hepatocellular Carcinoma with High Tumor Burden: Initial Lenvatinib Therapy with Subsequent Selective TACE. Liver Cancer 2019; 8: 299-311 [PMID: 31768341 DOI: 10.1159/000502905]
- Arizumi T, Ueshima K, Minami T, Kono M, Chishina H, Takita M, Kitai S, Inoue T, Yada N, Hagiwara S, Minami Y, Sakurai T, Nishida N, Kudo M. Effectiveness of Sorafenib in Patients with Transcatheter Arterial Chemoembolization (TACE) Refractory and Intermediate-Stage Hepatocellular Carcinoma. Liver Cancer 2015; 4: 253-262 [PMID: 26734579 DOI: 10.1159/000367743]
- Kudo M, Matsui O, Izumi N, Kadoya M, Okusaka T, Miyayama S, Yamakado K, Tsuchiya K, 46 Ueshima K, Hiraoka A, Ikeda M, Ogasawara S, Yamashita T, Minami T; Liver Cancer Study Group of Japan. Transarterial chemoembolization failure/refractoriness: JSH-LCSGJ criteria 2014 update. Oncology 2014; 87 Suppl 1: 22-31 [PMID: 25427730 DOI: 10.1159/000368142]
- 47 Shimose S, Kawaguchi T, Tanaka M, Iwamoto H, Miyazaki K, Moriyama E, Suzuki H, Niizeki T, Shirono T, Nakano M, Suga H, Yamaguchi T, Yokokura Y, Noguchi K, Koga H, Torimura T. Lenvatinib prolongs the progression-free survival time of patients with intermediate-stage hepatocellular carcinoma refractory to transarterial chemoembolization: A multicenter cohort study using data mining analysis. Oncol Lett 2020; 20: 2257-2265 [PMID: 32782543 DOI: 10.3892/o1.2020.11758
- 48 Eso Y, Takai A, Takahashi K, Ueda Y, Taura K, Marusawa H, Seno H. Combination of Mac-2



Binding Protein Glycosylation Isomer and Up-To-Seven Criteria as a Useful Predictor for Child-Pugh Grade Deterioration after Transarterial Chemoembolization for Hepatocellular Carcinoma. Cancers (Basel) 2019; 11 [PMID: 30909405 DOI: 10.3390/cancers11030405]

- 49 Kudo M, Han KH, Ye SL, Zhou J, Huang YH, Lin SM, Wang CK, Ikeda M, Chan SL, Choo SP, Miyayama S, Cheng AL. A Changing Paradigm for the Treatment of Intermediate-Stage Hepatocellular Carcinoma: Asia-Pacific Primary Liver Cancer Expert Consensus Statements. Liver Cancer 2020; 9: 245-260 [PMID: 32647629 DOI: 10.1159/000507370]
- 50 Kudo M, Ueshima K, Chan S, Minami T, Chishina H, Aoki T, Takita M, Hagiwara S, Minami Y, Ida H, Takenaka M, Sakurai T, Watanabe T, Morita M, Ogawa C, Wada Y, Ikeda M, Ishii H, Izumi N, Nishida N. Lenvatinib as an Initial Treatment in Patients with Intermediate-Stage Hepatocellular Carcinoma Beyond Up-To-Seven Criteria and Child-Pugh A Liver Function: A Proof-Of-Concept Study. Cancers (Basel) 2019; 11 [PMID: 31370183 DOI: 10.3390/cancers11081084]
- Kudo M, Ueshima K, Ikeda M, Torimura T, Tanabe N, Aikata H, Izumi N, Yamasaki T, Nojiri S, 51 Hino K, Tsumura H, Kuzuya T, Isoda N, Yasui K, Aino H, Ido A, Kawabe N, Nakao K, Wada Y, Yokosuka O, Yoshimura K, Okusaka T, Furuse J, Kokudo N, Okita K, Johnson PJ, Arai Y; TACTICS study group. Randomised, multicentre prospective trial of transarterial chemoembolisation (TACE) plus sorafenib as compared with TACE alone in patients with hepatocellular carcinoma: TACTICS trial. Gut 2020; 69: 1492-1501 [PMID: 31801872 DOI: 10.1136/gutjnl-2019-318934]
- Kawamura Y, Ikeda K, Hirakawa M, Yatsuji H, Sezaki H, Hosaka T, Akuta N, Kobayashi M, Saitoh 52 S. Suzuki F. Suzuki Y. Arase Y. Kumada H. New classification of dynamic computed tomography images predictive of malignant characteristics of hepatocellular carcinoma. Hepatol Res 2010; 40: 1006-1014 [PMID: 20887336 DOI: 10.1111/j.1872-034X.2010.00703.x]
- Seo S, Hatano E, Higashi T, Hara T, Tada M, Tamaki N, Iwaisako K, Ikai I, Uemoto S. Fluorine-18 53 fluorodeoxyglucose positron emission tomography predicts tumor differentiation, P-glycoprotein expression, and outcome after resection in hepatocellular carcinoma. Clin Cancer Res 2007; 13: 427-433 [PMID: 17255262 DOI: 10.1158/1078-0432.CCR-06-1357]
- 54 Kawamura Y, Kobayashi M, Shindoh J, Kobayashi Y, Kasuya K, Sano T, Fujiyama S, Hosaka T, Saitoh S, Sezaki H, Akuta N, Suzuki F, Suzuki Y, Ikeda K, Arase Y, Hashimoto M, Kumada H. Pretreatment Heterogeneous Enhancement Pattern of Hepatocellular Carcinoma May Be a Useful New Predictor of Early Response to Lenvatinib and Overall Prognosis. Liver Cancer 2020; 9: 275-292 [PMID: 32647631 DOI: 10.1159/000505190]
- Terashima T, Yamashita T, Takata N, Toyama T, Shimakami T, Takatori H, Arai K, Kawaguchi K, 55 Kitamura K, Sakai Y, Mizukoshi E, Honda M, Kaneko S. Comparative analysis of liver functional reserve during lenvatinib and sorafenib for advanced hepatocellular carcinoma. Hepatol Res 2020; 50: 871-884 [PMID: 32307874 DOI: 10.1111/hepr.13505]
- Uchikawa S, Kawaoka T, Ando Y, Yamaoka K, Kosaka Y, Suehiro Y, Fujii Y, Morio K, Nakahara 56 T, Murakami E, Tsuge M, Hiramatsu A, Imamura M, Takahashi S, Chayama K, Aikata H. Trends in Hepatic Functional Reserve of Patients with Hepatocellular Carcinoma Treated with Tyrosine Kinase Inhibitors. Oncology 2020; 98: 727-733 [PMID: 32712613 DOI: 10.1159/000507815]
- Hiraoka A, Kumada T, Atsukawa M, Hirooka M, Tsuji K, Ishikawa T, Takaguchi K, Kariyama K, Itobayashi E, Tajiri K, Shimada N, Shibata H, Ochi H, Tada T, Toyoda H, Nouso K, Tsutsui A, Nagano T, Itokawa N, Hayama K, Imai M, Joko K, Koizumi Y, Hiasa Y, Michitaka K; On behalf of the Real-Life Practice Experts for HCC (RELPEC) Study Group and HCC 48 Group (hepatocellular carcinoma experts from 48 clinics in Japan). Early Relative Change in Hepatic Function with Lenvatinib for Unresectable Hepatocellular Carcinoma. Oncology 2019; 97: 334-340 [PMID: 31466068 DOI: 10.1159/000502095
- Sho T, Suda G, Ogawa K, Kimura M, Shimazaki T, Maehara O, Shigesawa T, Suzuki K, Nakamura 58 A, Ohara M, Umemura M, Kawagishi N, Natsuizaka M, Nakai M, Morikawa K, Furuya K, Baba M, Yamamoto Y, Kobayashi T, Meguro T, Saga A, Miyagishima T, Yokoo H, Kamiyama T, Taketomi A, Sakamoto N. Early response and safety of lenvatinib for patients with advanced hepatocellular carcinoma in a real-world setting. JGH Open 2020; 4: 54-60 [PMID: 32055698 DOI: 10.1002/jgh3.12209]
- 59 Sho T, Suda G, Ogawa K, Shigesawa T, Suzuki K, Nakamura A, Ohara M, Umemura M, Kawagishi N, Natsuizaka M, Nakai M, Morikawa K, Furuya K, Baba M, Ito J, Yamamoto Y, Kobayashi T, Meguro T, Saga A, Miyagishima T, Terasita K, Takagi T, Kamiyama T, Taketomi A, Sakamoto N. Lenvatinib in patients with unresectable hepatocellular carcinoma who do not meet the REFLECT trial eligibility criteria. Hepatol Res 2020; 50: 966-977 [PMID: 32562334 DOI: 10.1111/hepr.13511]
- Chuma M, Uojima H, Hiraoka A, Kobayashi S, Toyoda H, Tada T, Hidaka H, Iwabuchi S, Numata 60 K, Itobayashi E, Itokawa N, Kariyama K, Ohama H, Hattori N, Hirose S, Shibata H, Tani J, Imai M, Tajiri K, Moriya S, Wada N, Iwasaki S, Fukushima T, Ueno M, Yasuda S, Atsukawa M, Nouso K, Fukunishi S, Watanabe T, Ishikawa T, Nakamura S, Morimoto M, Kagawa T, Sakamoto M, Kumada T, Maeda S. Analysis of efficacy of lenvatinib treatment in highly advanced hepatocellular carcinoma with tumor thrombus in the main trunk of the portal vein or tumor with more than 50% liver occupation: A multicenter analysis. Hepatol Res 2021; 51: 201-215 [PMID: 33270323 DOI: 10.1111/hepr.13592]
- 61 Maruta S, Ogasawara S, Ooka Y, Obu M, Inoue M, Itokawa N, Haga Y, Seki A, Okabe S, Azemoto R, Itobayashi E, Atsukawa M, Sugiura N, Mizumoto H, Koroki K, Kanayama K, Kanzaki H, Kobayashi K, Kiyono S, Nakamura M, Kanogawa N, Saito T, Kondo T, Suzuki E, Nakamoto S, Tawada A, Chiba T, Arai M, Kanda T, Maruyama H, Kato N. Potential of Lenvatinib for an



Expanded Indication from the REFLECT Trial in Patients with Advanced Hepatocellular Carcinoma. Liver Cancer 2020; 9: 382-396 [PMID: 32999866 DOI: 10.1159/000507022]

- 62 Alsina A, Kudo M, Vogel A, Cheng AL, Tak WY, Ryoo BY, Evans TRJ, López López C, Daniele B, Misir S, Ren M, Izumi N, Qin S, Finn RS. Effects of Subsequent Systemic Anticancer Medication Following First-Line Lenvatinib: A Post Hoc Responder Analysis from the Phase 3 REFLECT Study in Unresectable Hepatocellular Carcinoma. Liver Cancer 2020; 9: 93-104 [PMID: 32071913 DOI: 10.1159/000504624]
- Yoshimoto T, Imura S, Morine Y, Ikemoto T, Arakawa Y, Iwahashi S, Saito YU, Takasu C, Ishikawa 63 D, Teraoku H, Bando Y, Shimada M. The Outcome of Sorafenib Therapy on Unresectable Hepatocellular Carcinoma: Experience of Conversion and Salvage Hepatectomy. Anticancer Res 2018; 38: 501-507 [PMID: 29277815 DOI: 10.21873/anticanres.12250]
- Takeyama H, Beppu T, Higashi T, Kaida T, Arima K, Taki K, Imai K, Nitta H, Hayashi H, 64 Nakagawa S, Okabe H, Hashimoto D, Chikamoto A, Ishiko T, Tanaka M, Sasaki Y, Baba H. Impact of surgical treatment after sorafenib therapy for advanced hepatocellular carcinoma. Surg Today 2018; **48**: 431-438 [PMID: 29110089 DOI: 10.1007/s00595-017-1603-x]



0 WU

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2088-2100

DOI: 10.4251/wjgo.v13.i12.2088

ISSN 1948-5204 (online)

ORIGINAL ARTICLE

Basic Study Dysbiosis of the duodenal microbiota as a diagnostic marker for pancreaticobiliary cancer

Mitsuru Sugimoto, Kazumichi Abe, Tadayuki Takagi, Rei Suzuki, Naoki Konno, Hiroyuki Asama, Yuki Sato, Hiroki Irie, Ko Watanabe, Jun Nakamura, Hitomi Kikuchi, Mika Takasumi, Minami Hashimoto, Tsunetaka Kato, Ryoichiro Kobashi, Takuto Hikichi, Hiromasa Ohira

ORCID number: Mitsuru Sugimoto 0000-0002-4223-613X; Kazumichi Abe 0000-0001-5359-9465; Tadayuki Takagi 0000-0003-0696-5973; Rei Suzuki 0000-0002-4049-0484; Naoki Konno 0000-0001-9830-4317; Hiroyuki Asama 0000-0002-0102-0404; Yuki Sato 0000-0001-8000-0972; Hiroki Irie 0000-0002-4805-6244; Ko Watanabe 0000-0003-3895-7636; Jun Nakamura 0000-0001-6006-1778; Hitomi Kikuchi 0000-0003-0583-1623: Mika Takasumi 0000-0002-6025-8084; Minami Hashimoto 0000-0002-5750-7182; Tsunetaka Kato 0000-0002-2529-2463; Ryoichiro Kobashi 0000-0003-0991-6042; Takuto Hikichi 0000-0002-9815-1557; Hiromasa Ohira 0000-0003-4331-0634.

Author contributions: Sugimoto M wrote the paper and designed and performed the research and laboratory experiments; Abe K provided advice on the laboratory experiments and research; Takagi T, Suzuki R, Konno N, Asama H, Sato Y, Irie H, Watanabe K, Nakamura J, Kikuchi H, Takasumi M, Hashimoto M, Kato T, and Kobashi R provided clinical advice; Hikichi T supervised the report; Ohira H supervised the report and writing of the paper.

Institutional review board

Mitsuru Sugimoto, Kazumichi Abe, Tadayuki Takagi, Rei Suzuki, Naoki Konno, Hiroyuki Asama, Yuki Sato, Hiroki Irie, Ko Watanabe, Jun Nakamura, Hitomi Kikuchi, Mika Takasumi, Minami Hashimoto, Tsunetaka Kato, Ryoichiro Kobashi, Hiromasa Ohira, Department of Gastroenterology, Fukushima Medical University School of Medicine, Fukushima 960-1295, Japan

Ko Watanabe, Jun Nakamura, Hitomi Kikuchi, Minami Hashimoto, Tsunetaka Kato, Ryoichiro Kobashi, Takuto Hikichi, Department of Endoscopy, Fukushima Medical University Hospital, Fukushima 960-1295, Japan

Corresponding author: Mitsuru Sugimoto, MD, PhD, Assistant Professor, Doctor, Department of Gastroenterology, Fukushima Medical University School of Medicine, 1 Hikarigaoka, Fukushima 960-1295, Japan. kitachuuou335@yahoo.co.jp

Abstract

BACKGROUND

Pancreaticobiliary cancer (PB Ca) is a lethal disease, and a useful diagnostic marker is urgently needed. A correlation between the human microbiota and malignant gastrointestinal diseases was recently reported.

AIM

To investigate the efficacy of the duodenal microbiota for diagnosing PB Ca.

METHODS

We recruited 22 patients with benign pancreaticobiliary diseases (benign group) and 12 patients with PB Ca (malignant group). The duodenal microbiota of each patient was analyzed by the 16S rDNA terminal restriction fragment length polymorphism method. Patient characteristics, tumor markers, and relative abundances of the duodenal microbiota were compared between the benign and malignant groups.

RESULTS

Cancer antigen 19-9 (CA19-9), Bifidobacterium, Clostridium cluster XVIII, and Prevotella levels differed significantly between the benign and malignant groups. Clostridium cluster XVIII had the greatest area under the receiver operating characteristic curve (AUC) among the four factors with respect to diagnosing PB



Conflict-of-interest statement: The authors declare no competing interests.

Data sharing statement: The

datasets analyzed during the current study are available from the corresponding author upon reasonable request.

Country/Territory of origin: Japan

Specialty type: Gastroenterology and hepatology

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: March 22, 2021 Peer-review started: March 22, 2021 First decision: July 3, 2021 Revised: July 10, 2021 Accepted: September 16, 2021 Article in press: September 16, 2021 Published online: December 15, 2021

P-Reviewer: Yao D S-Editor: Ma YJ

Ca (cutoff value: 3.038%; sensitivity: 58.3%; specificity: 95.2%; AUC: 0.81). The combination of Clostridium cluster XVIII (cutoff value: 3.038%) and CA19-9 Levels (cutoff value: 18.8 U/mL) showed 91.7% sensitivity and 71.4% specificity for diagnosing PB Ca.

CONCLUSION

The duodenal microbiota may be useful for PB Ca screening.

Key Words: Pancreaticobiliary cancer; Diagnostic marker; Duodenal microbiota; Clostridium cluster XVIII; Cancer antigen 19-9

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Recently, a correlation between the human microbiota and malignant gastrointestinal diseases was reported. In this report, the efficacy of the duodenal microbiota for diagnosing pancreaticobiliary cancer (PB Ca) was investigated. The combination of Clostridium cluster XVIII (cutoff value: 3.038%) and cancer antigen 19-9 Levels (cutoff value: 18.8 U/mL) showed 91.7% sensitivity and 71.4% specificity for diagnosing PB Ca. In conclusion, the duodenal microbiota may be useful for PB Ca screening.

Citation: Sugimoto M, Abe K, Takagi T, Suzuki R, Konno N, Asama H, Sato Y, Irie H, Watanabe K, Nakamura J, Kikuchi H, Takasumi M, Hashimoto M, Kato T, Kobashi R, Hikichi T, Ohira H. Dysbiosis of the duodenal microbiota as a diagnostic marker for pancreaticobiliary cancer. World J Gastrointest Oncol 2021; 13(12): 2088-2100

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2088.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2088

INTRODUCTION

Pancreaticobiliary cancer (PB Ca) is a lethal disease^[1]. Surgery is the only radical treatment for pancreatic cancer, but unfortunately, many pancreatic cancer patients have advanced-stage lesions or other organ metastases, and they thus are not candidates for surgery^[2]. For those who can undergo surgical treatment, the 5-year survival rate is reported to be 20%-30%[3,4].

In general, biliary tract cancer is difficult to diagnose. Conventional diagnostic methods include evaluating tumor markers [cancer antigen 19-9 (CA19-9) or carcinoembryonic antigen (CEA)], biliary biopsy, biliary juice cytology, and brush cytology, but the diagnostic power of these methods is not sufficient[5-19]. It has been reported that serum CA19-9 elevation is observed in 85% of patients with cholangiocarcinoma, though elevation of this marker can also be found in benign obstructive jaundice. Similarly, elevated serum CEA, which is not seen in obstructive jaundice, occurs in only 30% of patients with cholangiocarcinoma[20]. Therefore, effective diagnostic methods for the early diagnosis of PB Ca are urgently needed. Recently, a correlation between the human microbiota and malignant gastrointestinal diseases was reported[21-26]. In addition, oral and salivary microbiota communities have been reported to be effective in diagnosing pancreatic cancer or predicting the onset of pancreatic cancer [27-29], and the risk of pancreatic cancer is reportedly increased in patients with a history of periodontal disease[30]. Furthermore, serum antibodies against oral microbiota are reported to be a risk factor for the onset of pancreatic cancer[31]. However, the mechanism by which this dysbiosis leads to pancreatic cancer is unknown, especially as the pancreas is relatively distant from the mouth.

Thus, we hypothesized that the duodenal microbiota would be more efficient than the oral microbiota for diagnosing PB Ca because the duodenum is closer to the bile duct and pancreas than the oral cavity. The aim of this study was to determine the efficacy of the duodenal microbiota for diagnosing PB Ca.



L-Editor: A P-Editor: Liu JH



MATERIALS AND METHODS

Ethical approval

This study was approved by the Institutional Review Board of Fukushima Medical University.

Patients

We assessed 34 patients with pancreaticobiliary disease who visited our hospital over two years. Twenty-two patients were diagnosed with benign pancreaticobiliary diseases (benign group) [chronic pancreatitis: 6; intraductal papillary mucinous neoplasm (IPMN): 5; gallbladder adenomyomatosis: 3; autoimmune pancreatitis: 3; benign common bile duct (CBD) stricture of unknown origin: 2; serous cystic neoplasm: 2; and CBD stone: 1] (Table 1). The other 12 patients were diagnosed with PB Ca (malignant group) (pancreatic cancer: 9; bile duct cancer: 3). The patients provided written informed consent to participate in this study. For all pancreatic cancer cases, the lesion was located in the head. Eight pancreatic cancer patients were diagnosed by endoscopic ultrasound-guided fine needle aspiration. One pancreatic cancer patient was diagnosed with intraductal papillary mucinous carcinoma with evident worsening of the lesion by imaging. Benign diseases were diagnosed by no histological malignancy or unchanging lesions after a clinical course of at least six months. Furthermore, the IPMN patients in the benign group did not have high-risk stigmata or worrisome features[32]. The cases of bile duct cancer were diagnosed by biliary biopsy or surgery. According to the cytology grade, classes IV and V were diagnosed as malignancies. The stage of PB Ca was determined based on the UICC classification, ver. 8.

The patients did not receive antibiotic agents for at least a week prior to duodenal juice collection, and they did not receive steroids at all.

Sample collection and DNA extraction

An endoscope was used under sedation with midazolam. The endoscope was advanced to the duodenum, and 0.5-1.0 mL of duodenal juice was collected through a catheter and stored at -20°C. The endoscope used was Q260 and Q260H, and the catheter was a PR-109Q-1 or PR-104Q-1 (Olympus, Tokyo, Japan).

Bacterial DNA was extracted from duodenal juice samples in accordance with a previous report by Takahashi et al[33].

Terminal restriction fragment length polymorphism

Terminal restriction fragment (T-RF) length polymorphism (T-RFLP) was performed by TechnoSuruga Laboratory (Shizuoka, Japan) according to Nagashima's methods[34, 35]. The 16S rRNA gene was amplified from the extracted DNA using the primers 5' FAM-labeled 516F (5'-TGCCAGCAGCCGCGGTA-3') and 1510R (5'-GGTTACCTTGT-TACGACTT-3') and HotStarTaq DNA Polymerase (Qiagen, Hilden, Germany) with a Thermal Cycler Dice (Takara, Shiga, Japan). The amplification program used was as follows: preheating at 94°C for 15 min; 35 cycles of denaturation at 94°C for 30 s, annealing at 50°C for 30 s, and extension at 72°C for 2 min; and a terminal extension at 72°C for 10 min. DNA amplification was verified by electrophoresis of the polymerase chain reaction (PCR) products (2 µL) through a 1.0% agarose gel with Tris-acetate-EDTA buffer. The amplified DNA was purified by a MultiScreen PCR96 Filter Plate (Millipore, Billerica, MA, United States).

The purified PCR product (3 µL) was digested with 10 U of Fast Digest BseLI (BslI) (Thermo Fisher Scientific) in a total volume of 15 µL at 37°C for 10 min. The restriction digestion products (0.5 µL) were mixed with 0.1 µL of a DNA fragment-length standard size marker and 10 µL of deionized formamide. The standard size marker was MapMarker X-Rhodamine Labeled 50-1000 bp (Bio Ventures, Murfreesboro, TN, United States). The samples were denatured at 95°C for 2 min and then placed immediately on ice. The T-RF length was established using an ABI PRISM 3130xl genetic analyzer (Thermo Fisher Scientific), and the length and peak area were determined using the genotyping software GeneMapper (Thermo Fisher Scientific). The fragment sizes were estimated using the Local Southern method in GeneMapper software (Thermo Fisher Scientific). If the peak height was less than 50 fluorescence units, the T-RF was excluded from the analysis. The fragments were resolved to one base pair by manual alignment of the size standard peaks from different electropherograms, and the predicted T-RFLP patterns of the 16S rDNA of known bacterial species were obtained using publicly available sequences. T-RFs were divided by operational taxonomic units (OTUs), and bacterial classification was performed



Table 1 Final patient diagnoses				
Benign group (<i>n</i> = 22)		Malignant group (<i>n</i> = 12)		
Chronic pancreatitis	6	Pancreatic cancer, stage (I/II/III/IV)	9 (2/5/1/1/)	
IPMN	5	Biliary ductal cancer, stage (I/II/III/IV)	3 (1/2/0/0)	
GB ADM	3			
Autoimmune pancreatitis	3			
CBD stricture of unknown origin	2			
Serous cystic neoplasm	2			
CBD stone	1			

IPMN: Intraductal papillary mucinous neoplasm; GB ADM: Gallbladder adenomyomatosis; CBD: Common bile duct.

according to the ratio of each OTU per total OTU area. The OTUs were identified by correspondence to a database of human intestinal flora (https://www.tecsrg.co.jp/trflp/index.html).

Analyzed traits

Patient characteristics and tumor markers (age, sex, reduction in body weight \geq 5 kg within 6 mo prior to duodenal juice sampling, intake of proton pump inhibitors, CA19-9) were compared between the two groups. The body weight marker was selected for the following reasons. The composition ratio of the microbiota has been reported to be different between subjects with obesity and those with a normal body mass index^[36]. Because the intake of high-fat foods influences the quantity and composition of bile acid, the intestinal bacterial flora might change[37]. The relative abundances of duodenal microbiota members (Bacteroides, Bifidobacterium, Lactobacillales, Prevotella, Clostridium cluster IV, Clostridium subcluster XIVa, Clostridium cluster IX, Clostridium cluster XI, Clostridium cluster XVIII, and others) were compared between the benign and malignant groups.

Statistical analysis

Normally distributed continuous variables were compared using Student's t test and nonnormally distributed continuous variables using the Mann-Whitney U test. Nominal variables were compared with Fisher's exact test. A receiver operating characteristic (ROC) curve was employed to compare the accuracy of the biomarkers. The *P* value < 0.05 was considered statistically significant.

These statistical analyses were performed using the EZR platform (Saitama Medical Center, Jichi Medical University, Saitama, Japan), which is a graphical user interface for R (The R Foundation for Statistical Computing, Vienna, Austria). EZR is a modified version of R-commander that was designed to perform functions that are frequently used in biostatistics[38].

RESULTS

Among the patient characteristics and tumor markers, age and CA19-9 Levels were significantly different between the benign and malignant groups (mean ± SD, age: 63.3 ± 12.2 vs 73.0 ± 8.3 years, P value = 0.016; median (range), CA19-9: 5.4 (2.0-54.8) vs 22.8 (2.0-9893.2), P value = 0.03) (Table 2).

Comparison of microbiome components revealed Bifidobacterium, Clostridium cluster XVIII, and *Prevotella* to be significantly different between the benign and malignant groups (median (range), *Bifidobacterium*: 0 (0-0.5)% vs 0.3 (0-4.5)%, P value < 0.05; *Clostridium* cluster XVIII: 1.3 (0-3.9)% vs 3.6 (0.8-14.9)%, P value = 0.006; Prevotella: 2.2 (0-25.9)% *vs* 0.1 (0-10.9)%, *P* value = 0.04) (Figure 1 and Table 3).

To determine the influence of age on CA19-9 Levels and microbiome composition, these factors were compared between the subgroup of patients < 69 years and the subgroup of those \geq 69 years (the median age of all patients was 69 years). According to the results, CA19-9 Levels, Bifidobacterium, Clostridium cluster XVIII, and Prevotella were not influenced by age (Table 4).

Table 2 Comparison of patient characteristics, tumor markers, and microbiomes

	Benign group (<i>n</i> = 22)	Malignant group (<i>n</i> = 12)	P value
Age, yr (mean ± SD)	63.0 ± 12.2	73.0 ± 8.3	0.016
Sex (male/female)	8/14	4/8	1.0
Reduction in body weight \geq 5 kg within 6 mo before duodenal juice sampling, <i>n</i> (%)	1 (4.5)	2 (16.7)	0.28
Intake of proton pump inhibitors, <i>n</i> (%)	4 (18.2)	2 (16.7)	1.0
CA19-9, U/mL, median (range)	5.4 (2.0-54.8)	22.8 (2.0-9893.2)	0.03

CA19-9: Cancer antigen 19-9.

Table 3 Microbiome comparison

	Benign group (<i>n</i> = 22)	Malignant group (<i>n</i> = 12)	P value
Bacteroides, %, median (range)	4.3 (0-26.1)	5.6 (0-46.4)	0.55
Bifidobacterium, %, median (range)	0 (0-0.5)	0.3 (0-4.5)	< 0.05
Clostridium cluster IV, %, median (range)	2.9 (0-10.8)	3.4 (0-8.8)	0.80
Clostridium cluster IX, %, median (range)	4.7 (0.6-19.5)	4.9 (0-17.8)	0.68
Clostridium cluster XI, %, median (range)	0 (0-0)	0 (0-0)	
Clostridium cluster XVIII, %, median (range)	1.3 (0-3.9)	3.6 (0.8-14.9)	0.006
Clostridium subcluster XIVa, %, median (range)	5.1 (0-23.1)	6.4 (2.9-13.7)	0.38
Lactobacillales, %, mean ± SD	63.0 ± 19.7	62.6 ± 18.3	0.95
Prevotella, %, median (range)	2.2 (0-25.9)	0.1 (0-10.9)	0.04
Others, %, median (range)	4.9 (2.5-20.4)	4.1 (1.5-6.3)	0.14

Table 4 Effects of age on cancer antigen 19-9 levels and the human microbiome

	Age < 69 yr (<i>n</i> = 17)	Age ≥ 69 yr (<i>n</i> = 17)	P value
CA19-9, U/mL, median (range)	7.1 (2-129.3)	4.9 (2-9893.2)	0.77
Bifidobacterium, %, median (range)	0.3 (0-0.5)	0 (0-4.5)	0.3
Clostridium cluster XVIII, %, median (range)	2.6 (0-5.7)	1.8 (0-14.9)	0.82
Prevotella, %, median (range)	1.3 (0-18.2)	1.9 (0-25.9)	0.56

CA19-9: Cancer antigen 19-9.

We assessed the ability of the microbiota to diagnose PB Ca by calculating the area under the ROC curve (AUC) and found that *Clostridium* cluster XVIII had the highest AUC (cutoff value: 3.038%, sensitivity: 58.3%, specificity: 95.2%, AUC: 0.81) among the three microbiome components (Bifidobacterium, Clostridium cluster XVIII, Prevotella) and CA19-9 Levels (Figure 2).

The combination of *Clostridium* cluster XVIII (cutoff value: 3.038%) and CA19-9 Levels (cutoff value: 18.8 U/mL) was also examined as a marker to diagnose PB Ca; the sensitivity of this combination was 91.7% (11/12), and the specificity was 71.4% (15/21) (Table 5). CA19-9 data were missing for one patient in the benign group.

DISCUSSION

In this study, we investigated which members of the duodenal microbiota could aid in diagnosing PB Ca and found Clostridium cluster XVIII to be more useful than CA19-9 Levels and other bacteria for diagnosing PB Ca. Notably, the combination of



Table 5 Diagnosis of pancreaticobiliary cancer by the combination of <i>Clostridium</i> cluster XVIII and cancer antigen 19-9 levels			
	Cutoff value	Sensitivity	Specificity
CA19-9	18.8 U/mL	66.7% (8/12)	76.2% (16/21 ¹)
Clostridium cluster XVIII	3.038%	58.3% (7/12)	95.2% (20/21 ¹)
Combination of Clostridium cluster XVIII and CA19-9		91.7% (11/12)	71.4% (15/21 ¹)

¹CA19-9 data were missing for a patient in the benign group. CA19-9: Cancer antigen 19-9.



Figure 1 Analysis of the duodenal microbiota. A, B: *Bifidobacterium* levels were significantly higher in the malignant group than in the benign group; A, C: *Clostridium* cluster XVIII levels were significantly higher in the malignant group than in the benign group; A, D: *Prevotella* levels were significantly higher in the benign group; M: Malignant group.

Clostridium cluster XVIII and CA19-9 Levels showed high sensitivity, indicating that this combination is valuable for screening patients for PB Ca.

As mentioned above, the oral microbiota has been considered to be a biomarker in pancreatic cancer. First, Michaud *et al*[30] reported that a history of periodontal disease was a risk factor for pancreatic cancer. After that, several oral microbes were reported to be more abundant and possible predictors and risk factors for survival in pancreatic cancer (Table 6)[27-29,39,40]. In addition, antibodies against *Porphyromonas gingivalis* can serve as a risk factor for pancreatic cancer onset[27].

Although the salivary microbiome may be useful for the medical care of patients with pancreatic cancer, it is influenced by differences in oral hygiene, mastication, and swallowing among individuals[27,41,42]. In contrast, the duodenal microbiota is more relevant to the pancreas and bile duct than is the salivary microbiota. Therefore, the duodenal microbiota was hypothesized to directly reflect the dysbiosis associated with PB Ca, and in fact, the results of this study reveal that the duodenal microbiota might be beneficial for screening PB Ca. The microbiota around the pancreas and biliary duct have been reported. Microbes of the duodenal mucosa, bile juice, cancer tissue, and cyst fluid of IPMN in pancreaticobiliary tumor patients have been investigated (Table 6)[30,43-47]. However, the methods used to analyze the microbiota around the pancreas and bile duct in these reports were more invasive than duodenal juice sampling.

Zaishidena® WJGO | https://www.wjgnet.com

Table 6 Past reports on microbes and pancreaticobiliary cancer				
Disease	Ref.	Microbes	Sample	Role
Pancreatic cancer	Michaud <i>et al</i> [<mark>30</mark>]	A history of periodontal diseases		Risk factor
	Farrell <i>et al</i> [28]	A combination of Neisseria elongate and Streptococcus mitis	Oral	Distinguishing from healthy controls
	Torres <i>et al</i> [29]	Ratio of Leptotrichia to Porphyromonas	Saliva	Higher in pancreatic cancer patients
	Fan <i>et al</i> [27]	Porphyromonas gingivalis	Oral, antibody	Risk factor
	Olson et al[39]	Firmicutes	Oral	More abundant
	Lu et al[<mark>40</mark>]	Leptotrichia, Fusobacterium, Rothia, Actinomyces, Corynebacterium, Atopobium, Peptostreptococcus, Catonella, Oribacterium, Filifactor, Campylobacter, Moraxella, Tannerella	Tongue coating	More prevalent
	Mei <i>et al</i> [46]	Acinetobactor, Aquabacterium, Oceanobacillus, Rahnella, Massilia, Delftia, Deinococcus, Sphingobium	Duodenal mucosa	More abundant
	Mitsuhashi et al <mark>[43</mark>]	Fusobacterium species	Cancer tissue	Poor prognosis
	Riquelme <i>et al</i> [44]	Pseudoxanthomonas, Streptomyces, Saccharopolyspora, Bacillus clausii	Cancer tissue	Long-term survival
Pancreatic and ampullary cancer	Di Calro <i>et al</i> [45]	Escherichia coli, Klebsiella pneumoniae	Bile juice	Predictor for survival
IPMN with high- grade dysplasia	Gaiser <i>et al</i> [47]	Granulicatella adiacens, Fusobacterium nucleatum	Cyst fluid	More abundant

IPMN: Intraductal papillary mucinous neoplasm.

The mechanism by which microbes lead to PB Ca remains unknown. The etiology with respect to the salivary microbiota has been considered in past reports. P. gingivalis can interrupt signaling pathways by modulating receptor expression and cytokine secretion to evade the host's immune system[48-52]. Moreover, P. gingivalis activates the Toll-like receptor signaling pathway [53,54], which has been reported to be related to pancreatic carcinogenesis^[55,56]. In other reports, oral bacteria were found outside of the oral cavity in the gastrointestinal tract. Immune responses against these bacteria can cause inflammation and carcinogenesis in the pancreas. Lipopolysaccharide has also been reported to drive pancreatic carcinogenesis by blocking the MyD88dependent, Toll-like receptor 4 and MyD88-independent pathways[57]. In another report, the mechanism was described as follows. Bacterial ligands detected by Toll-like receptors cause a Th1/Th2/Th17 imbalance in the tumor microenvironment, promoting tumorigenesis in combination with Kras mutation. In the duodenum, microbes may reach the pancreatic duct or biliary duct through the Vater papilla. In the pancreas or bile duct, pattern recognition receptors (such as Toll-like receptors) are stimulated by the pathogenic molecular patterns of bacterial ligands and induce lower levels of immune suppression, leading to the development of PB Ca[58]. These results from past reports suggest that some type of immune system response is the link between the duodenal microbiota and PB Ca.

On the one hand, the relationship between Clostridium cluster XVIII and carcinogenesis has not been reported, even though Clostridium cluster XVIII is reported to have the potential to enhance regulatory T (Treg) cells[59]. Many Treg cells exist in tumor tissue and prevent the immune response to tumors. Therefore, Tregs contribute to tumor progression and poor prognosis[60-64]. Thus, Clostridium cluster XVIII may increase in response to cancer and activate Tregs. Alternatively, *Clostridium* cluster XVIII may activate Tregs, with oncogenesis advancing.

This report has some limitations. First, this study was small and performed at a single institution. However, based on the data from *Clostridium* cluster XVIII, the average value of the malignant group was 4.5%, and that of the benign group was 1.5%. Total thirty patients were needed to achieve an α error of 5% and a β value of 0.2. When *Clostridium* cluster XVIII was the main outcome, the minimum necessary sample size was secured. Although this is the first report to describe the relationship between the duodenal juice microbiota and PB Ca, the diseases in the malignant group were not



Figure 2 Comparison of the ability of microbiome components and cancer antigen 19-9 Levels to diagnose pancreaticobiliary cancer. The area under the receiver operating characteristic curve of *Clostridium* cluster XVIII was the highest among the three microbes and cancer antigen 19-9 Levels. CA19-9: Cancer antigen 19-9; AUC: The area under the curve.

uniform. If subgroup analyses of pancreatic diseases were performed, the abundance of some duodenal microbes would be significantly different between the benign and malignant groups (Table 7). We hope that a future study with a larger number of patients will confirm our results for both pancreatic cancer and biliary cancer. Second, healthy control subjects were not enrolled in this study. However, as esophagogastroduodenoscopy under sedation is rarely performed in healthy patients, this limitation was unavoidable in the study design. Third, T-RFLP was applied. Investigations into the duodenal microbiota have been limited because duodenal juice cannot be collected in large volumes (less than 0.5 mL is typically collected). However, the measurement of the duodenal microbiota was demonstrated to be possible. Follow-up studies using next-generation sequencing are warranted[65]. Fourth, examining the duodenal microbiota requires a somewhat invasive technique. In the future, the development of serum antibody testing for the duodenal microbiota should be pursued.

CONCLUSION

In conclusion, the duodenal microbiota may contribute to PB Ca screening.

Zaishidena® WJGO | https://www.wjgnet.com

Table 7 Microbiome comparison in patients with pancreatic disease				
	Benign pancreatic diseases (<i>n</i> = 16)	Pancreatic cancer (<i>n</i> = 9)	P value	
Bacteroides, %, median (range)	2.1 (0-26.1)	5.8 (0-46.4)	0.17	
Bifidobacterium, %, median (range)	0 (0-0.5)	0.48 (0-4.5)	0.03	
<i>Clostridium</i> cluster IV, %, mean ± SD	4.0 ± 3.3	3.6 ± 2.9	0.76	
<i>Clostridium</i> cluster IX, %, mean ± SD	5.3 ± 3.6	6.4 ± 5.2	0.57	
Clostridium cluster XI, %, median (range)	0 (0-0)	0 (0-0)		
Clostridium cluster XVIII, %, median (range)	1.4 (0-3.9)	3.0 (0.8-14.9)	0.04	
Clostridium subcluster XIVa, %, median (range)	3.8 (0-21.4)	6.0 (2.9-13.7)	0.32	
Lactobacillales, %, mean ± SD	68.4 ± 19.3	59.7 ± 20.4	0.3	
Prevotella, %, median (range)	4.2 (2.5-20.4)	4.0 (1.5-6.3)	0.3	
Others, %, median (range)	2.0 (0-18.3)	0.3 (0-11.0)	0.3	

ARTICLE HIGHLIGHTS

Research background

Pancreaticobiliary cancer (PB Ca) is a lethal disease; however, there are currently no appropriate diagnostic and prognostic markers. Recently, the human microbiota was reported to be a causative factor, diagnostic marker, and prognostic marker for gastrointestinal malignant diseases.

Research motivation

The oral and fecal microbiota have been reported to be useful diagnostic markers for gastrointestinal cancer. The duodenum is located closer to the pancreas and bile duct than the oral cavity and colon. Therefore, we hypothesized that assessment of the duodenal microbiota might improve the diagnostic accuracy for PB Ca.

Research objectives

To investigate the diagnostic accuracy of duodenal microbiota evaluation for PB Ca.

Research methods

Thirty-four PB Ca and benign pancreaticobiliary disease patients were recruited for this study, and their duodenal juice was aseptically collected by endoscopy. The duodenal microbiota was analyzed, and the relative abundances of species in the duodenal microbiota were compared between PB Ca patients and benign pancreaticobiliary disease patients. The PB Ca diagnosability was compared between a conventional tumor marker and species in the duodenal microbiota with significantly different abundances in PB Ca patients vs benign pancreaticobiliary disease patients.

Research results

The abundances of cancer antigen 19-9 (CA19-9), Bifidobacterium, Clostridium cluster XVIII, and Prevotella were significantly different between PB Ca patients and benign pancreaticobiliary disease patients. The diagnostic capacity of Clostridium cluster XVIII was the highest among the four markers (CA19-9, Bifidobacterium, Clostridium cluster XVIII, and Prevotella). The combined assessment of Clostridium cluster XVIII and CA19-9 Levels was useful for PB Ca diagnosis.

Research conclusions

It was possible to investigate the microbiota of duodenal juice. Duodenal microbiota evaluation may contribute to the diagnosis of PB Ca.

Research perspectives

In the future, novel diagnostic and prognostic markers and treatments could be developed by investigating the relationship between the duodenal microbiota and PB Ca.
ACKNOWLEDGEMENTS

We thank all the staff at the Department of Gastroenterology of Fukushima Medical University, the Department of Endoscopy of Fukushima Medical University Hospital, and the Gastroenterology Ward of Fukushima Medical University Hospital. We also thank TechnoSuruga Laboratory for T-RFLP.

REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. CA Cancer J Clin 2018; 68: 7-30 [PMID: 1 29313949 DOI: 10.3322/caac.21442]
- 2 Geer RJ, Brennan MF. Prognostic indicators for survival after resection of pancreatic adenocarcinoma. Am J Surg 1993; 165: 68-72; discussion 72 [PMID: 8380315 DOI: 10.1016/s0002-9610(05)80406-4]
- Picozzi VJ, Oh SY, Edwards A, Mandelson MT, Dorer R, Rocha FG, Alseidi A, Biehl T, Traverso LW, Helton WS, Kozarek RA. Five-Year Actual Overall Survival in Resected Pancreatic Cancer: A Contemporary Single-Institution Experience from a Multidisciplinary Perspective. Ann Surg Oncol 2017; 24: 1722-1730 [PMID: 28054192 DOI: 10.1245/s10434-016-5716-z]
- White RJ, Hasan S, Monga D, Finley G, Islam M, Schiffman S, Williams HK, Kulkarni A, Thakkar S, Kirichenko AV, Wegner RE. Time to Adjuvant Systemic Therapy Following Pancreatic Cancer Resection and Effect on Outcome. Pancreas 2019; 48: 1086-1091 [PMID: 31404024 DOI: 10.1097/MPA.000000000001373]
- 5 Uchida N, Kamada H, Ono M, Aritomo Y, Masaki T, Nakatsu T, Kuriyama S. How many cytological examinations should be performed for the diagnosis of malignant biliary stricture via an endoscopic nasobiliary drainage tube? J Gastroenterol Hepatol 2008; 23: 1501-1504 [PMID: 18028351 DOI: 10.1111/j.1440-1746.2007.05214.x]
- Rösch T, Hofrichter K, Frimberger E, Meining A, Born P, Weigert N, Allescher HD, Classen M, 6 Barbur M, Schenck U, Werner M. ERCP or EUS for tissue diagnosis of biliary strictures? Gastrointest Endosc 2004; 60: 390-396 [PMID: 15332029 DOI: 10.1016/s0016-5107(04)01732-8]
- 7 Foutch PG, Kerr DM, Harlan JR, Kummet TD. A prospective, controlled analysis of endoscopic cytotechniques for diagnosis of malignant biliary strictures. Am J Gastroenterol 1991; 86: 577-580 [PMID: 2028947 DOI: 10.3109/00365529109043642]
- Kubota Y, Takaoka M, Tani K, Ogura M, Kin H, Fujimura K, Mizuno T, Inoue K. Endoscopic 8 transpapillary biopsy for diagnosis of patients with pancreaticobiliary ductal strictures. Am J Gastroenterol 1993; 88: 1700-1704 [PMID: 8213710 DOI: 10.1109/TCSI.2006.876416]
- Lee JG, Leung JW, Baillie J, Layfield LJ, Cotton PB. Benign, dysplastic, or malignant--making sense of endoscopic bile duct brush cytology: results in 149 consecutive patients. Am J Gastroenterol 1995; 90: 722-726 [PMID: 7733076]
- Ponchon T, Gagnon P, Berger F, Labadie M, Liaras A, Chavaillon A, Bory R. Value of endobiliary 10 brush cytology and biopsies for the diagnosis of malignant bile duct stenosis: results of a prospective study. Gastrointest Endosc 1995; 42: 565-572 [PMID: 8674929 DOI: 10.1016/s0016-5107(95)70012-9
- 11 Pugliese V, Conio M, Nicolò G, Saccomanno S, Gatteschi B. Endoscopic retrograde forceps biopsy and brush cytology of biliary strictures: a prospective study. Gastrointest Endosc 1995; 42: 520-526 [PMID: 8674921 DOI: 10.1016/s0016-5107(95)70004-8]
- Howell DA, Parsons WG, Jones MA, Bosco JJ, Hanson BL. Complete tissue sampling of biliary 12 strictures at ERCP using a new device. Gastrointest Endosc 1996; 43: 498-502 [PMID: 8726766 DOI: 10.1016/s0016-5107(96)70294-8]
- 13 Sugiyama M, Atomi Y, Wada N, Kuroda A, Muto T. Endoscopic transpapillary bile duct biopsy without sphincterotomy for diagnosing biliary strictures: a prospective comparative study with bile and brush cytology. Am J Gastroenterol 1996; 91: 465-467 [PMID: 8633492]
- Mansfield JC, Griffin SM, Wadehra V, Matthewson K. A prospective evaluation of cytology from 14 biliary strictures. Gut 1997; 40: 671-677 [PMID: 9203949 DOI: 10.1136/gut.40.5.671]
- Schoefl R, Haefner M, Wrba F, Pfeffel F, Stain C, Poetzi R, Gangl A. Forceps biopsy and brush 15 cytology during endoscopic retrograde cholangiopancreatography for the diagnosis of biliary stenoses. Scand J Gastroenterol 1997; 32: 363-368 [PMID: 9140159 DOI: 10.3109/00365529709007685]
- 16 Glasbrenner B, Ardan M, Boeck W, Preclik G, Möller P, Adler G. Prospective evaluation of brush cytology of biliary strictures during endoscopic retrograde cholangiopancreatography. Endoscopy 1999; **31**: 712-717 [PMID: 10604612 DOI: 10.1055/s-1999-73]
- Jailwala J, Fogel EL, Sherman S, Gottlieb K, Flueckiger J, Bucksot LG, Lehman GA. Triple-tissue 17 sampling at ERCP in malignant biliary obstruction. Gastrointest Endosc 2000; 51: 383-390 [PMID: 10744806 DOI: 10.1016/s0016-5107(00)70435-4]
- Macken E, Drijkoningen M, Van Aken E, Van Steenbergen W. Brush cytology of ductal strictures 18 during ERCP. Acta Gastroenterol Belg 2000; 63: 254-259 [PMID: 11189981 DOI: 10.1007/s002610000019
- 19 de Bellis M, Sherman S, Fogel EL, Cramer H, Chappo J, McHenry L Jr, Watkins JL, Lehman GA. Tissue sampling at ERCP in suspected malignant biliary strictures (Part 2). Gastrointest Endosc 2002;



56: 720-730 [PMID: 12397282 DOI: 10.1067/mge.2002.129219]

- Khan SA, Davidson BR, Goldin R, Pereira SP, Rosenberg WM, Taylor-Robinson SD, 20 Thillainayagam AV, Thomas HC, Thursz MR, Wasan H; British Society of Gastroenterology. Guidelines for the diagnosis and treatment of cholangiocarcinoma: consensus document. Gut 2002; 51 Suppl 6: VI1-VI9 [PMID: 12376491 DOI: 10.1136/gut.51.suppl_6.vi1]
- Kostic AD, Gevers D, Pedamallu CS, Michaud M, Duke F, Earl AM, Ojesina AI, Jung J, Bass AJ, 21 Tabernero J, Baselga J, Liu C, Shivdasani RA, Ogino S, Birren BW, Huttenhower C, Garrett WS, Meyerson M. Genomic analysis identifies association of Fusobacterium with colorectal carcinoma. Genome Res 2012; 22: 292-298 [PMID: 22009990 DOI: 10.1101/gr.126573.111]
- 22 Yamamura K, Baba Y, Nakagawa S, Mima K, Miyake K, Nakamura K, Sawayama H, Kinoshita K, Ishimoto T, Iwatsuki M, Sakamoto Y, Yamashita Y, Yoshida N, Watanabe M, Baba H. Human Microbiome Fusobacterium Nucleatum in Esophageal Cancer Tissue Is Associated with Prognosis. Clin Cancer Res 2016; 22: 5574-5581 [PMID: 27769987 DOI: 10.1158/1078-0432.CCR-16-1786]
- Ahn J, Sinha R, Pei Z, Dominianni C, Wu J, Shi J, Goedert JJ, Hayes RB, Yang L. Human gut 23 microbiome and risk for colorectal cancer. J Natl Cancer Inst 2013; 105: 1907-1911 [PMID: 24316595 DOI: 10.1093/jnci/djt300]
- 24 Mai V, Morris JG Jr. Need for prospective cohort studies to establish human gut microbiome contributions to disease risk. J Natl Cancer Inst 2013; 105: 1850-1851 [PMID: 24316594 DOI: 10.1093/jnci/djt349]
- Krishnan S, Eslick GD. Streptococcus bovis infection and colorectal neoplasia: a meta-analysis. 25 Colorectal Dis 2014; 16: 672-680 [PMID: 24824513 DOI: 10.1111/codi.12662]
- 26 Komiya Y, Shimomura Y, Higurashi T, Sugi Y, Arimoto J, Umezawa S, Uchiyama S, Matsumoto M, Nakajima A. Patients with colorectal cancer have identical strains of Fusobacterium nucleatum in their colorectal cancer and oral cavity. Gut 2019; 68: 1335-1337 [PMID: 29934439 DOI: 10.1136/gutjnl-2018-316661]
- Fan X, Alekseyenko AV, Wu J, Peters BA, Jacobs EJ, Gapstur SM, Purdue MP, Abnet CC, 27 Stolzenberg-Solomon R, Miller G, Ravel J, Hayes RB, Ahn J. Human oral microbiome and prospective risk for pancreatic cancer: a population-based nested case-control study. Gut 2018; 67: 120-127 [PMID: 27742762 DOI: 10.1136/gutjnl-2016-312580]
- Farrell JJ, Zhang L, Zhou H, Chia D, Elashoff D, Akin D, Paster BJ, Joshipura K, Wong DT. 28 Variations of oral microbiota are associated with pancreatic diseases including pancreatic cancer. Gut 2012; 61: 582-588 [PMID: 21994333 DOI: 10.1136/gutjnl-2011-300784]
- 29 Torres PJ, Fletcher EM, Gibbons SM, Bouvet M, Doran KS, Kelley ST. Characterization of the salivary microbiome in patients with pancreatic cancer. PeerJ 2015; 3: e1373 [PMID: 26587342 DOI: 10.7717/peeri.1373]
- 30 Michaud DS, Joshipura K, Giovannucci E, Fuchs CS. A prospective study of periodontal disease and pancreatic cancer in US male health professionals. J Natl Cancer Inst 2007; 99: 171-175 [PMID: 17228001 DOI: 10.1093/jnci/djk021]
- Michaud DS, Izard J, Wilhelm-Benartzi CS, You DH, Grote VA, Tjønneland A, Dahm CC, Overvad 31 K, Jenab M, Fedirko V, Boutron-Ruault MC, Clavel-Chapelon F, Racine A, Kaaks R, Boeing H, Foerster J, Trichopoulou A, Lagiou P, Trichopoulos D, Sacerdote C, Sieri S, Palli D, Tumino R, Panico S, Siersema PD, Peeters PH, Lund E, Barricarte A, Huerta JM, Molina-Montes E, Dorronsoro M, Quirós JR, Duell EJ, Ye W, Sund M, Lindkvist B, Johansen D, Khaw KT, Wareham N, Travis RC, Vineis P, Bueno-de-Mesquita HB, Riboli E. Plasma antibodies to oral bacteria and risk of pancreatic cancer in a large European prospective cohort study. Gut 2013; 62: 1764-1770 [PMID: 22990306 DOI: 10.1136/gutjnl-2012-303006]
- 32 Tanaka M, Fernández-Del Castillo C, Kamisawa T, Jang JY, Levy P, Ohtsuka T, Salvia R, Shimizu Y, Tada M, Wolfgang CL. Revisions of international consensus Fukuoka guidelines for the management of IPMN of the pancreas. Pancreatology 2017; 17: 738-753 [PMID: 28735806 DOI: 10.1016/j.pan.2017.07.007]
- Takahashi S, Tomita J, Nishioka K, Hisada T, Nishijima M. Development of a prokaryotic universal 33 primer for simultaneous analysis of Bacteria and Archaea using next-generation sequencing. PLoS One 2014; 9: e105592 [PMID: 25144201 DOI: 10.1371/journal.pone.0105592]
- Nagashima K, Hisada T, Sato M, Mochizuki J. Application of new primer-enzyme combinations to 34 terminal restriction fragment length polymorphism profiling of bacterial populations in human feces. Appl Environ Microbiol 2003; 69: 1251-1262 [PMID: 12571054 DOI: 10.1128/AEM.69.2.1251-1262.2003
- 35 Nagashima K, Mochizuki J, Hisada T, Suzuki S, Shimomura K. Phylogenetic analysis of 16S rRNA gene sequences from human fecal microbiota and improved utility of T-RFLP profiling. Biosci Microflora 2006; 25: 99-107 [DOI: 10.12938/bifidus.25.99]
- Ley RE, Turnbaugh PJ, Klein S, Gordon JI. Microbial ecology: human gut microbes associated with 36 obesity. Nature 2006; 444: 1022-1023 [PMID: 17183309 DOI: 10.1038/4441022a]
- Yatsunenko T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, Magris M, 37 Hidalgo G, Baldassano RN, Anokhin AP, Heath AC, Warner B, Reeder J, Kuczynski J, Caporaso JG, Lozupone CA, Lauber C, Clemente JC, Knights D, Knight R, Gordon JI. Human gut microbiome viewed across age and geography. Nature 2012; 486: 222-227 [PMID: 22699611 DOI: 10.1038/nature11053]
- 38 Kanda Y. Investigation of the freely available easy-to-use software 'EZR' for medical statistics. Bone Marrow Transplant 2013; 48: 452-458 [PMID: 23208313 DOI: 10.1038/bmt.2012.244]



- Olson SH, Satagopan J, Xu Y, Ling L, Leong S, Orlow I, Saldia A, Li P, Nunes P, Madonia V, Allen 39 PJ, O'Reilly E, Pamer E, Kurtz RC. The oral microbiota in patients with pancreatic cancer, patients with IPMNs, and controls: a pilot study. Cancer Causes Control 2017; 28: 959-969 [PMID: 28762074 DOI: 10.1007/s10552-017-0933-8]
- Lu H, Ren Z, Li A, Li J, Xu S, Zhang H, Jiang J, Yang J, Luo Q, Zhou K, Zheng S, Li L. Tongue 40 coating microbiome data distinguish patients with pancreatic head cancer from healthy controls. J Oral Microbiol 2019; 11: 1563409 [PMID: 30728915 DOI: 10.1080/20002297.2018.1563409]
- Lockhart PB, Brennan MT, Sasser HC, Fox PC, Paster BJ, Bahrani-Mougeot FK. Bacteremia 41 associated with toothbrushing and dental extraction. Circulation 2008; 117: 3118-3125 [PMID: 18541739 DOI: 10.1161/CIRCULATIONAHA.107.758524]
- 42 Crasta K, Daly CG, Mitchell D, Curtis B, Stewart D, Heitz-Mayfield LJ. Bacteraemia due to dental flossing. J Clin Periodontol 2009; 36: 323-332 [PMID: 19426179 DOI: 10.1111/i.1600-051X.2008.01372.x
- Mitsuhashi K, Nosho K, Sukawa Y, Matsunaga Y, Ito M, Kurihara H, Kanno S, Igarashi H, Naito T, 43 Adachi Y, Tachibana M, Tanuma T, Maguchi H, Shinohara T, Hasegawa T, Imamura M, Kimura Y, Hirata K, Maruyama R, Suzuki H, Imai K, Yamamoto H, Shinomura Y. Association of Fusobacterium species in pancreatic cancer tissues with molecular features and prognosis. Oncotarget 2015; 6: 7209-7220 [PMID: 25797243 DOI: 10.18632/oncotarget.3109]
- 44 Riquelme E, Zhang Y, Zhang L, Montiel M, Zoltan M, Dong W, Quesada P, Sahin I, Chandra V, San Lucas A, Scheet P, Xu H, Hanash SM, Feng L, Burks JK, Do KA, Peterson CB, Nejman D, Tzeng CD, Kim MP, Sears CL, Ajami N, Petrosino J, Wood LD, Maitra A, Straussman R, Katz M, White JR, Jenq R, Wargo J, McAllister F. Tumor Microbiome Diversity and Composition Influence Pancreatic Cancer Outcomes. Cell 2019; 178: 795-806.e12 [PMID: 31398337 DOI: 10.1016/j.cell.2019.07.008
- 45 Di Carlo P, Serra N, D'Arpa F, Agrusa A, Gulotta G, Fasciana T, Rodolico V, Giammanco A, Sergi C. The microbiota of the bilio-pancreatic system: a cohort, STROBE-compliant study. Infect Drug Resist 2019; 12: 1513-1527 [PMID: 31354308 DOI: 10.2147/IDR.S200378]
- Mei QX, Huang CL, Luo SZ, Zhang XM, Zeng Y, Lu YY. Characterization of the duodenal bacterial 46 microbiota in patients with pancreatic head cancer vs. healthy controls. Pancreatology 2018; 18: 438-445 [PMID: 29653723 DOI: 10.1016/j.pan.2018.03.005]
- 47 Gaiser RA, Halimi A, Alkharaan H, Lu L, Davanian H, Healy K, Hugerth LW, Ateeb Z, Valente R, Fernández Moro C, Del Chiaro M, Sällberg Chen M. Enrichment of oral microbiota in early cystic precursors to invasive pancreatic cancer. Gut 2019; 68: 2186-2194 [PMID: 30872392 DOI: 10.1136/gutjnl-2018-317458]
- Duncan L, Yoshioka M, Chandad F, Grenier D. Loss of lipopolysaccharide receptor CD14 from the 48 surface of human macrophage-like cells mediated by Porphyromonas gingivalis outer membrane vesicles. Microb Pathog 2004; 36: 319-325 [PMID: 15120158 DOI: 10.1016/j.micpath.2004.02.004]
- Stathopoulou PG, Benakanakere MR, Galicia JC, Kinane DF. The host cytokine response to 49 Porphyromonas gingivalis is modified by gingipains. Oral Microbiol Immunol 2009; 24: 11-17 [PMID: 19121064 DOI: 10.1111/j.1399-302X.2008.00467.x]
- Singh A, Wyant T, Anaya-Bergman C, Aduse-Opoku J, Brunner J, Laine ML, Curtis MA, Lewis JP. 50 The capsule of Porphyromonas gingivalis leads to a reduction in the host inflammatory response, evasion of phagocytosis, and increase in virulence. Infect Immun 2011; 79: 4533-4542 [PMID: 21911459 DOI: 10.1128/IAI.05016-11]
- Taxman DJ, Swanson KV, Broglie PM, Wen H, Holley-Guthrie E, Huang MT, Callaway JB, Eitas 51 TK, Duncan JA, Ting JP. Porphyromonas gingivalis mediates inflammasome repression in polymicrobial cultures through a novel mechanism involving reduced endocytosis. J Biol Chem 2012; 287: 32791-32799 [PMID: 22843689 DOI: 10.1074/jbc.M112.401737]
- 52 Palm E, Khalaf H, Bengtsson T. Porphyromonas gingivalis downregulates the immune response of fibroblasts. BMC Microbiol 2013; 13: 155 [PMID: 23841502 DOI: 10.1186/1471-2180-13-155]
- Hayashi C, Papadopoulos G, Gudino CV, Weinberg EO, Barth KR, Madrigal AG, Chen Y, Ning H, 53 LaValley M, Gibson FC 3rd, Hamilton JA, Genco CA. Protective role for TLR4 signaling in atherosclerosis progression as revealed by infection with a common oral pathogen. J Immunol 2012; 189: 3681-3688 [PMID: 22956579 DOI: 10.4049/jimmunol.1201541]
- Zambirinis CP, Levie E, Nguy S, Avanzi A, Barilla R, Xu Y, Seifert L, Daley D, Greco SH, Deutsch 54 M, Jonnadula S, Torres-Hernandez A, Tippens D, Pushalkar S, Eisenthal A, Saxena D, Ahn J, Hajdu C, Engle DD, Tuveson D, Miller G. TLR9 Ligation in pancreatic stellate cells promotes tumorigenesis. J Exp Med 2015; 212: 2077-2094 [PMID: 26481685 DOI: 10.1084/jem.20142162]
- 55 Zhang JJ, Wu HS, Wang L, Tian Y, Zhang JH, Wu HL. Expression and significance of TLR4 and HIF-1alpha in pancreatic ductal adenocarcinoma. World J Gastroenterol 2010; 16: 2881-2888 [PMID: 20556833 DOI: 10.3748/wjg.v16.i23.2881]
- Ochi A, Nguyen AH, Bedrosian AS, Mushlin HM, Zarbakhsh S, Barilla R, Zambirinis CP, Fallon 56 NC, Rehman A, Pylayeva-Gupta Y, Badar S, Hajdu CH, Frey AB, Bar-Sagi D, Miller G. MyD88 inhibition amplifies dendritic cell capacity to promote pancreatic carcinogenesis via Th2 cells. J Exp Med 2012; 209: 1671-1687 [PMID: 22908323 DOI: 10.1084/jem.20111706]
- Bracci PM. Oral Health and the Oral Microbiome in Pancreatic Cancer: An Overview of 57 Epidemiological Studies. Cancer J 2017; 23: 310-314 [PMID: 29189325 DOI: 10.1097/PPO.00000000000287
- Sethi V, Vitiello GA, Saxena D, Miller G, Dudeja V. The Role of the Microbiome in Immunologic



Development and its Implication For Pancreatic Cancer Immunotherapy. Gastroenterology 2019; 156: 2097-2115.e2 [PMID: 30768986 DOI: 10.1053/j.gastro.2018.12.045]

- 59 Atarashi K, Tanoue T, Oshima K, Suda W, Nagano Y, Nishikawa H, Fukuda S, Saito T, Narushima S, Hase K, Kim S, Fritz JV, Wilmes P, Ueha S, Matsushima K, Ohno H, Olle B, Sakaguchi S, Taniguchi T, Morita H, Hattori M, Honda K. Treg induction by a rationally selected mixture of Clostridia strains from the human microbiota. Nature 2013; 500: 232-236 [PMID: 23842501 DOI: 10.1038/nature12331]
- Bazewicz CG, Dinavahi SS, Schell TD, Robertson GP. Aldehyde dehydrogenase in regulatory T-cell 60 development, immunity and cancer. Immunology 2019; 156: 47-55 [PMID: 30387499 DOI: 10.1111/imm.13016
- 61 Fridman WH, Pagès F, Sautès-Fridman C, Galon J. The immune contexture in human tumours: impact on clinical outcome. Nat Rev Cancer 2012; 12: 298-306 [PMID: 22419253 DOI: 10.1038/nrc3245
- 62 Protti MP, De Monte L, Di Lullo G. Tumor antigen-specific CD4+ T cells in cancer immunity: from antigen identification to tumor prognosis and development of therapeutic strategies. Tissue Antigens 2014; 83: 237-246 [PMID: 24641502 DOI: 10.1111/tan.12329]
- Saito T, Nishikawa H, Wada H, Nagano Y, Sugiyama D, Atarashi K, Maeda Y, Hamaguchi M, 63 Ohkura N, Sato E, Nagase H, Nishimura J, Yamamoto H, Takiguchi S, Tanoue T, Suda W, Morita H, Hattori M, Honda K, Mori M, Doki Y, Sakaguchi S. Two FOXP3(+)CD4(+) T cell subpopulations distinctly control the prognosis of colorectal cancers. Nat Med 2016; 22: 679-684 [PMID: 27111280 DOI: 10.1038/nm.4086]
- Tanaka A, Sakaguchi S. Regulatory T cells in cancer immunotherapy. Cell Res 2017; 27: 109-118 64 [PMID: 27995907 DOI: 10.1038/cr.2016.151]
- 65 Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, Nielsen T, Pons N, Levenez F, Yamada T, Mende DR, Li J, Xu J, Li S, Li D, Cao J, Wang B, Liang H, Zheng H, Xie Y, Tap J, Lepage P, Bertalan M, Batto JM, Hansen T, Le Paslier D, Linneberg A, Nielsen HB, Pelletier E, Renault P, Sicheritz-Ponten T, Turner K, Zhu H, Yu C, Jian M, Zhou Y, Li Y, Zhang X, Qin N, Yang H, Wang J, Brunak S, Doré J, Guarner F, Kristiansen K, Pedersen O, Parkhill J, Weissenbach J; MetaHIT Consortium, Bork P, Ehrlich SD, Wang J. A human gut microbial gene catalogue established by metagenomic sequencing. Nature 2010; 464: 59-65 [PMID: 20203603 DOI: 10.1038/nature08821]



0 WJ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2101-2113

DOI: 10.4251/wjgo.v13.i12.2101

ISSN 1948-5204 (online)

ORIGINAL ARTICLE

Basic Study MutL homolog 1 methylation and microsatellite instability in sporadic colorectal tumors among Filipinos

Loraine Kay D Cabral, Cynthia A Mapua, Filipinas F Natividad, Caecilia H C Sukowati, Edgardo R Cortez, Ma Luisa D Enriquez

ORCID number: Loraine Kay D Cabral 0000-0001-7931-7489; Cynthia A Mapua 0000-0002-5578-7811; Filipinas F Natividad 0000-0003-2738-887X; Caecilia H C Sukowati 0000-0001-9699-7578; Edgardo R Cortez 0000-0003-0057-7793; Ma Luisa D Enriquez 0000-0002-4542-3627.

Author contributions: Enriquez MLD conceptualized the project; Cabral LKD designed and

optimized the experiments; Mapua CA screened data of the colorectal cancer databank and performed statistical analysis; Cabral LKD and Sukowati CHC analyzed data and wrote the manuscript; Cortez ER and Natividad FF coordinated specimen collection and funds allocation as study group head and division head respectively; all authors read and approved the manuscript.

Institutional review board

statement: The clinical study protocol and the revised clinical study protocol (Ref. No. 06-015) had been approved by the St. Luke's Medical Center Institutional Ethics Review Board.

Conflict-of-interest statement: All

authors declared that they have no conflicts of interest.

Loraine Kay D Cabral, Cynthia A Mapua, Filipinas F Natividad, Ma Luisa D Enriquez, Research and Biotechnology Group, St. Luke's Medical Center, Quezon City 1112, Philippines

Loraine Kay D Cabral, Caecilia H C Sukowati, Centro Studi Fegato, Fondazione Italiana Fegato ONLUS, Trieste 34149, Italy

Edgardo R Cortez, Department of Surgery, St. Luke's Medical Center, Quezon City 1112, Philippines

Ma Luisa D Enriquez, Center for Natural Science and Environmental Research, De La Salle University, Manila 1004, Philippines

Corresponding author: Loraine Kay D Cabral, BSc, MSc, Research Associate, Research and Biotechnology Group, St. Luke's Medical Center, 279 E Rodriguez Sr. Avenue, Quezon City 1112, Philippines. kay.cabral@fegato.it

Abstract

BACKGROUND

Colorectal cancer (CRC) ranks third in terms of incidence and second in mortality worldwide. In CRC, the silencing of mismatch repair genes, including the mutL homolog 1 (*hMLH1*) has been linked to microsatellite instability (MSI), the lengthening or shortening of microsatellite repeats. Very limited data have been presented so far on the link of hMLH1 methylation and MSI in Southeast Asia populations with sporadic CRC, and on its clinical significance.

AIM

To investigate the significance of the MSI status and *hMLH1* methylation in CRC Filipino patients.

METHODS

Fifty-four sporadic CRC patients with complete clinical data were included in this study. Genomic DNA from CRC tumor biopsies and their normal tissue counterparts were profiled for MSI by high resolution melting (HRM) analysis using the Bethesda Panel of Markers (BAT25, BAT26, D2S123, D5S346, and D17S250). hMLH1 methylation screening was performed using bisulfite conversion and methylation specific polymerase chain reaction. Statistical analysis was conducted to calculate their associations to clinicopathological characteristics



Data sharing statement: The authors may share the data for justified reason.

Supported by Department of Science and Technology and the Philippine Council for Health Research and Development (DOST-PCHRD) (to Cabral LKD); St. Luke's Medical Center, Manila, Philippines; and Regione Autonomo FVG in Progetti Internazionali 2021 to the FIF, No. DGR 189 dd 12/2/21.

Country/Territory of origin: Philippines

Specialty type: Oncology

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: May 11, 2021 Peer-review started: May 11, 2021 First decision: June 12, 2021 Revised: June 24, 2021 Accepted: September 14, 2021 Article in press: September 14, 2021 Published online: December 15, 2021

P-Reviewer: Lei XH S-Editor: Gao CC L-Editor: A

and survival relevance (Kaplan-Meier curves and the log-rank test).

RESULTS

hMLH1 methylation was observed in 9% and 35% of CRC and normal samples, respectively. Higher incidence of consistently methylated *hMLH1* found in both normal and CRC was noticed for relation to location of tumor (P < 0.05). As for MSI status, D2S123 the most common unstable microsatellite and MSI-high (MSI-H) was the most common MSI profile, counted for 46% and 50% of normal and CRC tissues, respectively. The presence of MSI-low (MSI-L) and microsatellite stable (MSS) was 43% and 11% for normal, and 31% and 19% for CRC samples. The mean month of patients' survival was shorter in patients whose normal and tumor tissues had methylated compared to those with unmethylated *hMLH1* and with MSI-H compared to those with MSI-L/MSS (P < 0.05). This was supported by significant difference in Kaplan-Meier with log-rank analysis. This data indicated that *hMLH1* methylation and high MSI status have prognostic value.

CONCLUSION

This study showed the clinical significance of *hMLH1* methylation and MSI status in sporadic CRC Filipino patients, especially in the normal part of the tumor.

Key Words: Sporadic colorectal cancer; DNA methylation; Microsatellite instability; Population genetic; Colorectal cancer

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Colorectal cancer (CRC) ranks third in terms of incidence and second in mortality worldwide. In CRC, the silencing of mismatch repair genes, including the mutL homolog 1 (hMLH1) has been linked to microsatellite instability (MSI). This study investigated the status of *hMLH1* methylation and MSI in normal and tumor tissues of Filipinos sporadic CRC patients and their clinical significances.

Citation: Cabral LKD, Mapua CA, Natividad FF, Sukowati CHC, Cortez ER, Enriquez MLD. MutL homolog 1 methylation and microsatellite instability in sporadic colorectal tumors among Filipinos. World J Gastrointest Oncol 2021; 13(12): 2101-2113

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2101.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2101

INTRODUCTION

Colorectal cancer (CRC) occurs when malignant tumors form in the lining of the large intestine, which includes the ascending, transverse, and descending colon, and the rectum. CRC ranks third in terms of incidence and second in mortality worldwide, for both sexes. Over 1.9 million new CRC cases and 935000 deaths were estimated to occur in 2020, accounting for about 1/10 cancer cases and deaths[1]. In the Philippines, CRC is currently the third leading site of malignancy. The incidence of CRC had almost doubled from 2010 to 2015 with survival rates under 50% [2,3]. In terms of mortality rate, Filipinos (and Chinese) ethnicity had significantly decreased risk of death compared with Caucasians[4].

CRC, like many other cancers, is a malignancy caused by DNA changes causing abnormal behavior of cells. DNA deletions, duplications, substitutions, mutations, and rearrangements can either activate or inactivate a gene or several genes and consequently affect cellular function^[5]. Depending on the origin of the mutation, CRC can be classified as sporadic (70%), inherited (5%), and familial (25%)[6]. Sporadic CRC is characterized by the carcinogenesis pathway, derived from point mutations which can occur in different genes[7].

In an inherited CRC, the hereditary non-polyposis CRC (HNPCC), germline mutation is involved in one or two of the mutator or mismatch repair (MMR) genes, including the mutL homolog 1 (hMLH1). Mutations in hMLH1 have been linked to microsatellite instability (MSI), the lengthening or shortening of microsatellite repeats



P-Editor: Yu HG



(sequences with 1-6 repeating nucleotides). When instability is left unrepaired, it may accumulate as mutations and can affect other genes that have microsatellite repeats in their coding regions[8,9]. MSI is found in 85%-90% of HNPCC patients[10].

Recent studies demonstrated that in a subset of sporadic CRC, DNA methylation, the transfer of a covalent methyl group to the C5 position of the cytosine to form 5-methylcytosine by DNA methyltransferases, may cause the loss of function of a MMR gene. Likewise, it leads to the accumulation of MSI[11,12]. Tumors with methylated *hMLH1* and high levels of MSI present with a distinct characteristic from other CRC tumors. CRC with methylated *hMLH1* had a delayed onset and was associated with female gender[13]. This linked *hMLH1* and MSI in the development of sporadic CRC, suggesting that the two molecular profiles are closely related[14].

So far, very limited data have been presented on the link of *hMLH1* methylation and MSI in Southeast Asia populations with sporadic CRC. This study presents the detection and characterization of MSI status and *hMLH1* methylation in CRC and its paired non-tumoral adjacent tissues in Filipino patients.

MATERIALS AND METHODS

Patients and samples

Fifty-four sporadic CRC patients with complete clinical data were included in this study. Diagnosis of CRC was based on the presence of malignancy in the initial biopsy. Patients should be Filipino by descent with no family history of cancer. From each patient, paired tumor and its corresponding normal tissue were obtained from surgical resection at the St. Luke's Medical Center, Quezon City, Philippines. Normal tissues were collected approximately 6 inches away from the margin of the tumor. Upon pathological confirmation, fresh frozen sections were stored in -80°C. The project of the Colorectal Cancer Study Group was approved by the Institutional Ethics Review Board of St. Luke's Medical Center (Project Code No. 06-015). All patients enrolled in the study signed an Informed Consent Form allowing the use of their tissues and clinical data in the Colorectal Cancer (CRC) Databank of St. Luke's Medical Center.

CRC cell lines SW480 (ATCC[®] CCL-228) and SW48 (ATCC[®] CCL-23) were purchased from American Type Culture Collection (ATCC) as controls for MSI and *hMLH1* methylation. SW480 is a CRC cell line that has stable microsatellite and unmethylated *hMLH1*, while SW48 is high MSI and methylated *hMLH1*. Lymphocytes from patients who underwent colonoscopy and who were found to be free of cancer and polyp were used as additional controls for stable microsatellite.

Isolation of genomic DNA

Genomic DNA (gDNA) extraction was performed using the QIAamp[®] DNA Mini Kit (Qiagen), according to the manufacturer's instructions. Briefly, tissues were finely minced and put in lysis solution and proteinase K until completely lysed. After DNA precipitation with ethanol, DNA extract was washed twice and eluted with appropriate buffer. DNA quality and quantity were assessed using Nanodrop[®] v1000 spectrophotometer (Thermo Fisher Scientific). Final working concentration of 50 ng of gDNA was used for each sample in the succeeding analysis.

Bisulfite conversion and methylation specific polymerase chain reaction for hMLH1

gDNA from normal and tumor specimens was subjected to bisulfite treatment to differentiate methylated cytosines from unmethylated ones. Bisulfite chemically modifies non-methylated cytosines into uracil, which is then converted to thymidine in polymerase chain reaction (PCR) cycles.

Bisulfite treatment of the DNA sample was performed using EZ DNA Methylation Lightning Kit (Zymo Research) according to the manufacturer's suggestion. Briefly, 200-500 ng gDNA was incubated in the conversion reagent and then treated with binding buffer in a spin column. Converted DNA was then subjected to desulphonation and clean-up using washing buffer. DNA (approximately 10 µL) was eluted and collected for methylation specific PCR (MS-PCR)[15]. Reaction was carried out using Qiagen Taq Core Kit in a reaction volume of 25 µL with 10x PCR Buffer, 2.5 mmol/L of MgCl₂, 50 pmol of primer, 1.25 mmol/L of dNTPs and 1.25 units of Taq DNA polymerase added to 1.5-2 µL of converted DNA. Primer sets used for *hMLH1* MS-PCR are taken from published work of Fox *et al*[15].

Zaishideng® WJGO | https://www.wjgnet.com

MSI test by high resolution melting analysis

MSI test was carried out using PCR for five markers from the Bethesda panel that included BAT25, BAT26, D2S123, D5S346, and D17S250, as in a previous study[16]. PCR conditions of each marker were optimized and validated in our earlier study on HNPCC (Evangelista & Enriquez, unpublished work).

In brief, PCR was performed using Qiagen Taq PCR core kit with EvaGreen dye (Biotium) in a 25 μ L reaction volume containing 50 ng of gDNA. All reactions were done in triplicates. PCR and high resolution melting (HRM) analysis were carried out using Rotor-Gene[™] 6000 (Qiagen) system with data collected over the range from 55 °C to 95 °C with ramp rising at 0.1 °C/s[17]. Melting curve was analyzed using Rotor-Gene Q (RGQ) Scanning Software version 2.0.2 (Qiagen). Raw melting-curve data were normalized by manual adjustment of linear regions before (pre-; 100% fluorescence) and after (post-; 0% fluorescence) the melting transition. The melting curve of DNA from SW480 cells which exhibits microsatellite stable (MSS) phenotype [18] was used as the normal/stable control. The RGQ software assigned the profile of each sample in reference to the stable control. The confidence percentage (cut-off), was optimized by analyzing non-cancer patients' DNA, with values not lower than 60%. Therefore, the confidence value of $\geq 60\%$ was regarded as MSS, while any confidence value of < 60% was regarded as unstable.

MSI score was defined as MSI-high (MSI-H) where HRM instability was observed in \geq 2 markers; MSI-low (MSI-L) where instability was only in one marker; and MSS if no instability was observed in any of the markers. For validation, amplified PCR products of the stable and unstable control were subjected to Sanger sequencing to verify their microsatellite repeats.

Statistical analysis

Statistical analysis was constructed using software GraphPrism version 5.01 (GraphPad Software, Inc., La Jolla, CA, United States). Associations between clinicopathological characteristics (age, sex, tumor location, grade and stage), MSI status and methylation status were determined using Pearson's chi-square test or Fisher's exact test. Survival relevance was analyzed using Kaplan-Meier curves and the logrank test based on the *hMLH1* methylation and MSI status grouping. All tests were two-tailed and values were considered significant at *P* value of less than 0.05.

RESULTS

Patients' demographic and tumor features

This study analyzed 54 Filipino CRC patients with comparable male to female ratio (30M:24F). The age mean was 56.9 ± 11.8 years old, with 5 patients under 40 years old and 49 patients above 40 years old. Regarding tumor parameters, CRC was mostly found in the distal part of the colon (38; 70%), followed by the rectum (10; 19%) and proximal part of the colon (6; 11%). By histology, moderately-differentiated grade CRC was noticed in 44 (81%), poorly-differentiated in 8 (15%), and well-differentiated in 2 (4%) patients. By disease stage, stage 1 CRC was observed in 6 (11%), stage 2 in 14 (26%), stage 3 in 30 (56%), and stage in 4 (7%) of patients. None of the patients had family history of CRC.

hMLH1 methylation status in Filipino CRC patients

We analyzed the *hMLH1* DNA methylation by MS-PCR. From 54 paired CRC and its normal adjacent tissue, most of the samples were unmethylated (91% and 65%, respectively). Representative MS-PCR gel electrophoresis is shown in Figure 1. hMLH1 methylation was noticed only in 5 (9%) and 19 (35%) of CRC and normal samples, respectively. It is interesting to note that the methylated *hMLH1* in tumor tissues was accompanied by methylation in its paired normal tissues (4/5, 80%). Only 1 sample showed tumoral hMLH1 methylation without methylation in its normal tissue (Figure 2).

The distribution of *hMLH1* methylation status in patients' demography and tumor parameters is shown in Table 1 (left panel). In normal tissues, the incidence of hMLH1 methylation was slightly higher in female compared to male (42% vs 30%), and in tumors located in the proximal colon compared to distal/rectum location (67% vs 31%). The same pattern was found for hMLH1 methylation in tumor tissues. Methylation was higher in females compared to males (13% vs 7%), and in tumors located in the proximal colon compared to those in distal colon or rectum (33% vs 6%, P = 0.031).



Table 1 hMLH1 methylation status in the Filipinos colorectal cancer patients											
		Normal tissue (%)			Tumor tissue(%)			Normal/tumor (%)			
		М	U		М	U		M/M	U/U	M/U or U/M	
	Freq	19	35	P value	5	49	P value	4	34	16	P value
Gender											
Male	30	9 (30)	21 (70)	NS	2 (7)	28 (93)	NS ¹	1 (3)	20 (67)	9 (30)	NS
Female	24	10 (42)	14 (58)		3 (12)	21 (88)		3 (13)	14 (58)	7 (29)	
Age											
≤ 40	5	3 (60)	2 (40)	NS ¹	0 (0)	5 (100)	NS ¹	0 (0)	2 (40)	3 (60)	NS
> 40	49	16 (33)	33 (67)		5 (10)	44 (90)		4 (8)	32 (65)	13 (27)	
Location of tumor											
Proximal	6	4 (67)	2 (33)	NS ¹	2 (33)	4 (67)	0.031	2 (33)	2 (33)	2 (33)	0.029
Distal/rectum	48	15 (31)	33 (69)		3 (6)	45 (94)		2 (4)	32 (67)	14 (29)	
Tumor grade											
Poor	8	5 (62)	3 (38)	NS	1 (12)	7 (88)	NS	1 (12)	3 (38)	4 (50)	NS
Moderate	44	13 (30)	31 (70)		4 (9)	40 (91)		3 (7)	30 (68)	11 (25)	
Well	2	1 (50)	1 (50)		0 (0)	2 (100)		0 (0)	1 (50)	1 (50)	
Tumor stage											
I-II	20	6 (30)	14 (70)	NS	2 (10)	18 (90)	NS ¹	2 (10)	14 (70)	4 (20)	NS
III-IV	34	13 (38)	21 (62)		3 (9)	31 (91)		2 (6)	20 (59)	12 (35)	

¹Fisher's exact test.

NS: Not significant; M: Methylated; U: Unmethylated; In paired normal/tumor tissues: M/M: Methylated/methylated; U/U: Unmethylated/ unmethylated; M/U: Methylated/unmethylated; U/M: Unmethylated/methylated.

> The signature of *hMLH1* status in paired normal and tumor tissue is presented in Table 1 (right panel). Although the incidence was not statistically significant, a higher incidence of methylated hMLH1 in both normal and tumor tissues (M/M) was noticed in females, in > 40 years old patients, and in poorly differentiated cancer. A significant difference was found for the tumor location (P = 0.029), where M/M signature was noticed in 33% of proximal tumors, while U/U was in 67% of distally located or rectal tumors.

MSI status in Filipino sporadic CRC patients

MSI status was assessed using HRM of the Bethesda panel that included BAT25, BAT26, D2S123, D5S346, and D17S250. As controls, cell lines SW480 and SW48 were used to represent a MSS and MSI-H, respectively. MSS feature was also checked in lymphocytes DNA from a normal individual. A representative HRM analysis of BAT26 gene and its direct Sanger sequencing are shown in Figure 1.

From the Bethesda panel markers, the microsatellite with the highest instability was D2S123 and this was found in 32 (59%) and 33 (61%) samples of normal and tumor, respectively. On the other hand, BAT26 instability was observed only in 1 (2%) sample of each normal and tumor tissues. In normal tissues, the instability of BAT25, D17S250, and D5S346, were 2 (4%), 14 (26%), and 29 (54%), respectively. Among the CRC tissues, instability was found in BAT25, BAT26, D17S250, and D5S346 accounting for 5 (20%), 12 (22%), and 25 (46%), respectively. There was no statistical difference in the instability of these markers between normal and tumor tissues.

The distribution of MSI status in patients' demography and tumor parameter is shown in Table 2 (left panel). In normal tissue samples, 25 (46%), 23 (43%), and 6 (11%) samples were MSI-H, MSI-L, and MSS, respectively. In female, the highest percentage was for MSI-H (54%), followed by MSI-L (33%), and MSS (13%). Similar pattern was observed in tumoral tissues with the following rates: MSI-H with 27 (50%), MSI-L with 17 (31%), and MSS with 10 (19%). Instability rates of microsatellites in tumors tissues in the female group were the exactly the same as in the normal tissues.

Table 2 Microsatellite instability status in the Filipinos colorectal cancer patients															
		Normal tissue (%)			Tumor tissue (%)				Normal/tumor (%)						
		MSI-H	MSI-L	MSS	P value	MSI-H	MSI-L	MSS	P value	H/H	L/L	S/S	L/S or S/L	L/H or H/L	P value
	Freq	25	23	6		27	17	10		19	8	3	10	14	
Gender															
Male	30	12 (40)	15 (50)	3 (10)	NS	14 (47)	9 (30)	7 (23)	NS	9 (30)	4 (13)	1 (3)	8 (27)	8 (27)	NS
Female	24	13 (54)	8 (33)	3 (13)		13 (54)	8 (33)	3 (13)		10 (42)	4 (17)	2 (8)	2 (8)	6 (25)	
Age															
≤ 40	5	3 (60)	2 (40)	0 (0)	NS	2 (40)	1 (20)	2 (40)	NS	2 (40)	0 (0)	0 (0)	2 (40)	1 (20)	NS
> 40	49	22 (45)	21 (43)	6 (12)		25 (51)	16 (33)	8 (16)		17 (35)	8 (16)	3 (6)	8 (16)	13 (27)	
Location of tumor															
Proximal	6	4 (67)	2 (33)	0 (0)	NS	4 (67)	2 (33)	0 (0)	NS	4 (67)	2 (33)	0 (0)	0 (0)	0 (0)	NS
Distal/rectum	48	21 (44)	21 (44)	6 (12)		23 (48)	15 (31)	10 (21)		15 (31)	6 (13)	3 (6)	10 (21)	14 (29)	
Tumor grade															
Poor	8	5 (62)	3 (38)	0 (0)	NS	5 (63)	2 (25)	1 (12)	NS	4 (50)	1 (13)	0 (0)	1 (13)	2 (25)	NS
Moderate	44	19 (43)	19 (43)	6 (14)		21 (48)	15 (34)	8 (18)		14 (32)	7 (16)	3 (7)	8 (18)	12 (27)	
Well	2	1 (50)	1 (50)	0 (0)		1 (50)	0 (0)	1 (50)		1 (50)	0 (0)	0 (0)	1 (50)	0 (0)	NS
Tumor stage															
I-II	20	9 (45)	9 (45)	2 (10)	NS	8 (40)	10 (50)	2 (10)	NS	7 (35)	6 (30)	0 (0)	4 (20)	3 (15)	NS
III-IV	34	16 (47)	14 (41)	4 (12)		19 (56)	7 (21)	8 (23)		12 (35)	2 (6)	3 (9)	6 (18)	11(32)	

MSI-H: Microsatellite instability-high; MSI-L: Microsatellite instability-low; MSS: Microsatellite stable; NS: Not significant; In paired normal/tumor tissues: H/H: Microsatellite instability-high/microsatellite instability-high; L/L: Microsatellite instability-low; S/S: Microsatellite stable/microsatellite stable; L/S: Microsatellite instability-low/microsatellite stable; S/L: Microsatellite stable/microsatellite instability-low; L/H: Microsatellite instability-low/microsatellite instability-high; H/L: Microsatellite instability-high/microsatellite instability-low.

> The signature of MSI status in paired normal and tumor tissue is presented in Table 2 (right panel). Analysis of data using showed that the MSI status between normal and tumor tissues was not statistically significant. Nonetheless, a slightly higher incidence of MSI-H in hMLH1 was observed in both normal and tumor tissues (H/H) among females, similar to methylated *hMLH1*.

Correlation of MSI and hMLH1 methylation status

Table 3 shows the MSI and methylation status of each sample according to the paired groupings described previously. There was no statistically significant correlation between MSI and *hMLH1* methylation status of all the sample pairs. Thirteen paired samples (24%) were unmethylated with MSI-H (H/H and U/U). The clinicopathological profile of this cancer group showed that 11 samples came from the distal colon/rectum (85%), 12 (92%) were > 40 years old, 10 (77%) were moderately differentiated, and 8 (62%) were stage III/IV cancers.

Correlation of MSI and hMLH1 methylation status with prognosis

To determine the prognostic values of both *hMLH1* methylation and MSI status, we performed Kaplan-Meier analysis on patient survival for independent groups. The survival was defined as death event (in months) post-surgery. Patients were followed up until 60 mo after surgery.

Table 3 Correlation of <i>hMLH1</i> methylation and microsatellite instability status of paired samples											
		hMLH1 methylation (%)									
		M/M	M/U	U/M	U/U	Total	<i>P</i> value				
MSI status	H/H	1 (5)	5 (26)	0 (0)	13 (68)	19	NS				
	L/L	1 (13)	4 (50)	0 (0)	3 (37)	8					
	S/S	0 (0)	1 (33)	0 (0)	2 (67)	3					
	L/S or S/L	1 (10)	3 (30)	0 (0)	6 (60)	10					
	L/H or H/L	1 (7)	2 (14)	1 (7)	10 (71)	14					
	Total	4 (7)	15 (28)	1 (2)	34 (63)	54					

MSI: Microsatellite instability; M: Methylated; U: Unmethylated; In paired normal/tumor tissues: M/M: Methylated/methylated; U/U: Unmethylated/unmethylated; M/U: Methylated/unmethylated; U/M: Unmethylated/methylated; In paired normal/tumor tissues: H/H: Microsatellite instability-high/microsatellite instability-high; L/L: Microsatellite instability-low/microsatellite instability-low; S/S: Microsatellite stable/microsatellite stable; L/S: Microsatellite instability-low/microsatellite stable; S/L: Microsatellite stable/microsatellite instability-low; L/H: Microsatellite instabilitylow/microsatellite instability-high; H/L: Microsatellite instability-high/microsatellite instability-low; NS: Not significant.

> Our results show that the mean survival was relatively shorter in patients with methylated compared to those with unmethylated hMLH1, both in tumor [18 (6-29) mo vs 44 (34-54) mo] and in normal tissues [34 (21-46) mo vs 44 (33-54) mo], expressed as mean (95% confidence interval). A significant difference was also observed in tumor tissues with MSI-H compared to those with MSI-L/MSS [27 (19-36) mo vs 56 (44-67) mo]. For the MSI status in normal tissues, again a shorter survival was found for MSI-H compared to MSI-L/MSS [27 (18-36) mo vs 54 (39-68) mo].

> Kaplan-Meier analysis was performed to associate the hMLH1 methylation and MSI status with patients' survival in both cohorts. In concordance with the mean survival above, we found the association of MSI-H status in both normal and tumor tissues with shorter probability of survival (P < 0.05), compared to that of MSI-L/MSS (Figure 3). Our data indicated that *hMLH1* methylation and high MSI status might have a prognostic value.

DISCUSSION

CRC is a classic example of disease progression accompanied by molecular changes. Its carcinogenesis is composed of stage-specific molecular changes in gene expression and the accumulation of mutations. A mutation in the MMR genes may lead to accumulation of DNA damage. Consequently, the tumor suppressor genes or oncogenes are affected, leading to the development of the cancer[8].

DNA methylation has been linked to tumor suppressor gene silencing[19], including the silencing of hMLH1 MMR gene[20]. In a subset of sporadic CRC, the loss of function of hMLH1 gene led to the accumulation of instability in microsatellite regions of the DNA[11,12].

In this study, there was a notable higher incidence of methylation of *hMLH1* in normal tissue as compared to its paired tumor tissue (35% vs 9%). This is in agreement with other studies showing elevated hMLH1 methylation in the normal gastric epithelia in patients with stomach cancer as compared to those of non-cancer patients [21]. Methylation in the normal mucosa was proposed as a marker of risk for the development of CRC[22]. As demonstrated recently, the acceleration of ageing-related DNA methylation drift in normal mucosa was correlated with an earlier age of diagnosis[23].

hMLH1 methylation in non-neoplastic epithelia was shown also to exhibit MSI[24]. It is interesting to note that the methylated *hMLH1* in tumor was accompanied by methylation in its paired normal tissue. This finding warrants further investigation on the clinical utility of *hMLH1* methylation in normal tissue as a useful marker not only in CRC but in also in other gastrointestinal cancers.

However, in this study, a high incidence of non-methylated hMLH1 in MSI-H samples (both normal and tumor) suggests that hMLH1methylation did not bring about MSI. We assumed, at least in our cohort, that unstable microsatellites can be attributed to methylation in other MMR genes, such as hSMH2[25,26] and mutation in





Figure 1 Determination of *hMLH1* methylation by methylation-specific polymerase chain reaction and microsatellite instability status by high resolution melting analysis. A: Gel images showing presence of 124 bp product indicating an unmethylated *hMLH1* and the 115 bp product for the methylated *hMLH1* in clinical samples; B: Normalized difference curves of clinical samples showing microsatellite instability-high profiles in red; C: Difference curve of the high resolution melting analysis; D: Electropherograms showing deletion of 12 adenine repeats in microsatellite instable control SW48 using BAT26 compared to the 24 adenine repeats in stable control SW480 and lymphocytes from a normal individual. MS-PCR: Methylation specific polymerase chain reaction; HRM: High resolution melting; MSI: Microsatellite instability; MSI-H: Microsatellite instability-high; CRC: Colorectal cancer.

hSMH6[27]. It was also noted that MSI may also result from other defects in the component of base excision repair of the proteins (DNA glycosylase, AP endonuclease, DNA polymerase, *etc.*)[28]. MSI tumors were also grouped to Type A (\leq 6 bp change) and Type B (\geq 8 bp change). In a report on MMR gene knock-out animals, no Type B MSI was observed, suggesting that a kind of MSI group may involve malignancies apart from MMR deficiency[29].

Baishidena® WJGO | https://www.wjgnet.com

December 15, 2021 Volume 13 Issue 12



Figure 2 hMLH1 methylation and microsatellite instability analysis results of paired normal and tumor tissues. hMLH1 methylation analysis was determined using bisulfite converted genomic DNA and methylated and unmethylated primer sets for methylation specific polymerase chain reaction. Microsatellite instability profiling was done by high-resolution melting analysis using the five Bethesda panel of markers. MSI: Microsatellite instability; MSI-H: Microsatellite instability-high; MSI-L: Microsatellite instability-low; MSS: Microsatellite stable.

Significant differences for *hMLH1* methylation were found for the location of the tumor. Colon is divided into two anatomical regions: the proximal/right colon and the distal/left colon and each with very unique features[30-32]. This may be partially attributed to their difference in embryological development and physiological circumstances. The proximal colon develops from the hindgut while the distal colon originates from the midgut[33]. Proximal and distal colon are also different in the genetic patterns[34].

Since methylation is affected by environmental exposure, the difference might also be related to function. The proximal colon is the site where breakdown of complex carbohydrates occurs while the undigested dietary proteins are broken down in the distal colon. This process of digestion in the distal colon generates by-products such as ammonia, phenols, indoles and sulfurs, that may inhibit DNA methyltransferase activity[35]. Among African-American race, higher vegetable intake diet was shown being associated with greater odds for high hMLH1 methylation[36]. Because methylation is less inhibited in proximal region, this may explain the higher incidence of methylated normal and tumor tissue (M/M), in contrast to high percentage of unmethylation in distal region.

In this study, we observed that from both the average of survival data and log-rank test of Kaplan-Meier analysis, *hMLH1* methylation in normal tissues and MSI status in both normal and CRC tissues had prognostic value. Unmethylated *hMLH1* and MSI-L/MSS status were positively associated with better patients' survival. Our data may support earlier findings where expression of *hMLH1* is considered an independent prognostic and predictive factor stage II-III CRC in Chinese population[37]. Data from this study, however, in contrast with previous studies showing the association between MSI-H with improved overall and disease-free survival[38,39], showed that population genetics and the disease stages might influence the prognosis of CRC patients.

Although there was no statistical difference, the incidence of MSI and *hMLH1* methylation showed a similar trend in association with sex and grade. MSI and *hMLH1* methylation were more common in females and in poorly differentiated tumors. This non-significant correlation may be attributed to the small sample size, thus studies using larger number of samples that are well distributed in terms of tumor clinicopathological parameters will be needed. Moreover, significance of both markers in relation to patients' prognosis, especially for Southeast Asian population, needs to be validated. Investigation of the methylation status of two or more MMR genes and their correlation with MSI status is also recommended, since this study only looked into one MMR gene out of the six. A recent data from metastatic CRC showed that a novel epigenetic signature of eight hypermethylated genes was able to identify CRC with poor prognosis. These genes were characterized with CpG-island high





Figure 3 hMLH1 methylation and microsatellite instability analysis for patients' prognosis. Kaplan-Meier analysis of colorectal cancer patients' overall survival in 60-mo of patients' follow-up after surgery. MSI: Microsatellite instability.

methylator and MSI-like phenotype[40].

The establishment of molecular biomarkers in diseases is essential for an effective management and treatment protocol for cancer patients. For the precision treatment in the future, especially for immunotherapy, it was shown that patients with MSI tumors exhibited significant response to anti-PD-1 inhibitors after failure of conventional therapy[41,42]. The role of MSI and *hMLH1* methylation is shedding light in the management of sporadic CRC in clinical practice in other countries[43,44] but not yet in the Philippines.

CONCLUSION

To conclude, we showed the clinical significance of hMLH1 methylation and MSI status in sporadic CRC Filipino patients, especially in the normal part of the organ. This study is one of the few attempts to establish the molecular profile of Filipino CRC patients in the local setting, to highlight the importance of epigenetic modification. Understanding the diverse molecular and genetic key players involved in cancer development will ultimately translate to improvement in patient care.

Zaishidena® WJGO | https://www.wjgnet.com

ARTICLE HIGHLIGHTS

Research background

A distinct molecular signature marks a particular subset of sporadic colorectal cancer (CRC). It involves the mismatch repair (MMR) genes silencing due to DNA methylation, leading to microsatellite instability (MSI).

Research motivation

To improve the management of the CRC patients based on their distinct molecular subtypes.

Research objectives

To examine the association of mutL homolog 1 (*hMLH1*) methylation (MMR gene) and the MSI phenotype in relation to cancer characteristics and patient survival among Filipino sporadic CRC patients.

Research methods

Paired tissues (normal and tumor) from sporadic CRC patients was screened for hMLH1 methylation using methylation specific polymerase chain reaction. Subsequent MSI typing was done by high resolution melting analysis.

Research results

The results of this study showed that hMLH1 methylation was mostly noticed in proximal tumors. Low overall survival was observed in methylated hMLH1 and MSI tumors.

Research conclusions

The epigenetic silencing of *hMLH1* as well as MSI may present a distinct pattern of CRC in Filipino patients.

Research perspectives

This is an initial attempt to characterize sporadic CRC in Filipino population. It may shed light in understanding molecular epigenetic modification in CRC as well as its role in tumor development and management.

ACKNOWLEDGEMENTS

The members of the Colorectal Cancer Study Group of the St. Luke's Medical Center actively and consistently participated in the study.

REFERENCES

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin 2021; 71: 209-249 [PMID: 33538338 DOI: 10.3322/caac.21660]
- Ting FIL, Sacdalan DBL, Tampo MMT, Apellido RT, Monroy HJ 3rd, Sacdalan MDP, Sacdalan DL; 2 written on behalf of the University of the Philippines, Philippine General Hospital Colorectal Polyp and Cancer Study Group. Treatment Outcomes of Patients With Colorectal Cancer Enrolled in a Comprehensive Benefits Program of the National Insurance System in the Philippines: Data From the Pilot Site. JCO Glob Oncol 2020; 6: 35-46 [PMID: 32031435 DOI: 10.1200/JGO.19.00332]
- 3 Laudico AV, Mirasol-Lumague MR, Mapua CA, Uy GB, Toral JA, Medina VM, Pukkala E. Cancer incidence and survival in Metro Manila and Rizal province, Philippines. Jpn J Clin Oncol 2010; 40: 603-612 [PMID: 20385654 DOI: 10.1093/jjco/hyq034]
- 4 Le H, Ziogas A, Taylor TH, Lipkin SM, Zell JA. Survival of distinct Asian groups among colorectal cancer cases in California. Cancer 2009; 115: 259-270 [PMID: 19109815 DOI: 10.1002/cncr.24034]
- 5 Srivastava S, Verma M, Henson DE. Biomarkers for early detection of colon cancer. Clin Cancer Res 2001; 7: 1118-1126 [PMID: 11350874]
- 6 Mármol I, Sánchez-de-Diego C, Pradilla Dieste A, Cerrada E, Rodriguez Yoldi MJ. Colorectal Carcinoma: A General Overview and Future Perspectives in Colorectal Cancer. Int J Mol Sci 2017; 18 [PMID: 28106826 DOI: 10.3390/ijms18010197]
- Fearon ER, Vogelstein B. A genetic model for colorectal tumorigenesis. Cell 1990; 61: 759-767 [PMID: 2188735 DOI: 10.1016/0092-8674(90)90186-i]



- Potter JD. Colorectal cancer: molecules and populations. J Natl Cancer Inst 1999; 91: 916-932 8 [PMID: 10359544 DOI: 10.1093/inci/91.11.916]
- Imai K, Yamamoto H. Carcinogenesis and microsatellite instability: the interrelationship between genetics and epigenetics. Carcinogenesis 2008; 29: 673-680 [PMID: 17942460 DOI: 10.1093/carcin/bgm228]
- Bianchi F, Galizia E, Catalani R, Belvederesi L, Ferretti C, Corradini F, Cellerino R. CAT25 is a 10 mononucleotide marker to identify HNPCC patients. J Mol Diagn 2009; 11: 248-252 [PMID: 19324995 DOI: 10.2353/jmoldx.2009.080155]
- Berg KD, Glaser CL, Thompson RE, Hamilton SR, Griffin CA, Eshleman JR. Detection of 11 microsatellite instability by fluorescence multiplex polymerase chain reaction. J Mol Diagn 2000; 2: 20-28 [PMID: 11272898 DOI: 10.1016/S1525-1578(10)60611-3]
- 12 Samowitz WS, Curtin K, Ma KN, Schaffer D, Coleman LW, Leppert M, Slattery ML. Microsatellite instability in sporadic colon cancer is associated with an improved prognosis at the population level. Cancer Epidemiol Biomarkers Prev 2001; 10: 917-923 [PMID: 11535541]
- Malkhosyan SR, Yamamoto H, Piao Z, Perucho M. Late onset and high incidence of colon cancer of 13 the mutator phenotype with hypermethylated hMLH1 gene in women. Gastroenterology 2000; 119: 598 [PMID: 10960275 DOI: 10.1053/gast.2000.16154]
- 14 Poynter JN, Siegmund KD, Weisenberger DJ, Long TI, Thibodeau SN, Lindor N, Young J, Jenkins MA, Hopper JL, Baron JA, Buchanan D, Casey G, Levine AJ, Le Marchand L, Gallinger S, Bapat B, Potter JD, Newcomb PA, Haile RW, Laird PW; Colon Cancer Family Registry Investigators. Molecular characterization of MSI-H colorectal cancer by MLHI promoter methylation, immunohistochemistry, and mismatch repair germline mutation screening. Cancer Epidemiol Biomarkers Prev 2008; 17: 3208-3215 [PMID: 18990764 DOI: 10.1158/1055-9965.EPI-08-0512]
- 15 Fox EJ, Leahy DT, Geraghty R, Mulcahy HE, Fennelly D, Hyland JM, O'Donoghue DP, Sheahan K. Mutually exclusive promoter hypermethylation patterns of hMLH1 and O6-methylguanine DNA methyltransferase in colorectal cancer. J Mol Diagn 2006; 8: 68-75 [PMID: 16436636 DOI: 10.2353/jmoldx.2006.050084]
- Loukola A, Eklin K, Laiho P, Salovaara R, Kristo P, Järvinen H, Mecklin JP, Launonen V, Aaltonen 16 LA. Microsatellite marker analysis in screening for hereditary nonpolyposis colorectal cancer (HNPCC). Cancer Res 2001; 61: 4545-4549 [PMID: 11389088]
- 17 Janavicius R, Matiukaite D, Jakubauskas A, Griskevicius L. Microsatellite instability detection by high-resolution melting analysis. Clin Chem 2010; 56: 1750-1757 [PMID: 20852132 DOI: 10.1373/clinchem.2010.150680]
- Brennetot C, Buhard O, Jourdan F, Flejou JF, Duval A, Hamelin R. Mononucleotide repeats BAT-26 18 and BAT-25 accurately detect MSI-H tumors and predict tumor content: implications for population screening. Int J Cancer 2005; 113: 446-450 [PMID: 15455342 DOI: 10.1002/ijc.20586]
- 19 Wong JJ, Hawkins NJ, Ward RL. Colorectal cancer: a model for epigenetic tumorigenesis. Gut 2007; 56: 140-148 [PMID: 16840508 DOI: 10.1136/gut.2005.088799]
- Yamashita K, Dai T, Dai Y, Yamamoto F, Perucho M. Genetics supersedes epigenetics in colon 20 cancer phenotype. Cancer Cell 2003; 4: 121-131 [PMID: 12957287 DOI: 10.1016/s1535-6108(03)00190-9
- Waki T, Tamura G, Tsuchiya T, Sato K, Nishizuka S, Motoyama T. Promoter methylation status of 21 E-cadherin, hMLH1, and p16 genes in nonneoplastic gastric epithelia. Am J Pathol 2002; 161: 399-403 [PMID: 12163364 DOI: 10.1016/S0002-9440(10)64195-8]
- Kawakami K, Ruszkiewicz A, Bennett G, Moore J, Grieu F, Watanabe G, Iacopetta B. DNA 22 hypermethylation in the normal colonic mucosa of patients with colorectal cancer. Br J Cancer 2006; 94: 593-598 [PMID: 16421593 DOI: 10.1038/sj.bjc.6602940]
- 23 Joo JE, Clendenning M, Wong EM, Rosty C, Mahmood K, Georgeson P, Winship IM, Preston SG, Win AK, Dugué PA, Jayasekara H, English D, Macrae FA, Hopper JL, Jenkins MA, Milne RL, Giles GG, Southey MC, Buchanan DD. DNA Methylation Signatures and the Contribution of Age-Associated Methylomic Drift to Carcinogenesis in Early-Onset Colorectal Cancer. Cancers (Basel) 2021; 13 [PMID: 34070516 DOI: 10.3390/cancers13112589]
- Sakata K, Tamura G, Endoh Y, Ohmura K, Ogata S, Motoyama T. Hypermethylation of the hMLH1 24 gene promoter in solitary and multiple gastric cancers with microsatellite instability. Br J Cancer 2002; 86: 564-567 [PMID: 11870538 DOI: 10.1038/sj.bjc.6600076]
- Morán A, Ortega P, de Juan C, Fernández-Marcelo T, Frías C, Sánchez-Pernaute A, Torres AJ, Díaz-25 Rubio E, Iniesta P, Benito M. Differential colorectal carcinogenesis: Molecular basis and clinical relevance. World J Gastrointest Oncol 2010; 2: 151-158 [PMID: 21160823 DOI: 10.4251/wjgo.v2.i3.151]
- 26 Zhang H, Fu WL, Huang Q. Mapping of the methylation pattern of the hMSH2 promoter in colon cancer, using bisulfite genomic sequencing. J Carcinog 2006; 5: 22 [PMID: 16911791 DOI: 10.1186/1477-3163-5-22]
- 27 Kawaguchi M, Banno K, Yanokura M, Kobayashi Y, Kishimi A, Ogawa S, Kisu I, Nomura H, Hirasawa A, Susumu N, Aoki D. Analysis of candidate target genes for mononucleotide repeat mutation in microsatellite instability-high (MSI-H) endometrial cancer. Int J Oncol 2009; 35: 977-982 [PMID: 19787250 DOI: 10.3892/ijo_00000411]
- 28 Brim H, Mokarram P, Naghibalhossaini F, Saberi-Firoozi M, Al-Mandhari M, Al-Mawaly K, Al-Mjeni R, Al-Sayegh A, Raeburn S, Lee E, Giardiello F, Smoot DT, Vilkin A, Boland CR, Goel A, Hafezi M, Nouraie M, Ashktorab H. Impact of BRAF, MLH1 on the incidence of microsatellite



instability high colorectal cancer in populations based study. Mol Cancer 2008; 7: 68 [PMID: 18718023 DOI: 10.1186/1476-4598-7-68]

- 29 Oda S, Maehara Y, Ikeda Y, Oki E, Egashira A, Okamura Y, Takahashi I, Kakeji Y, Sumiyoshi Y, Miyashita K, Yamada Y, Zhao Y, Hattori H, Taguchi K, Ikeuchi T, Tsuzuki T, Sekiguchi M, Karran P, Yoshida MA. Two modes of microsatellite instability in human cancer: differential connection of defective DNA mismatch repair to dinucleotide repeat instability. Nucleic Acids Res 2005; 33: 1628-1636 [PMID: 15778432 DOI: 10.1093/nar/gki303]
- 30 Bufill JA. Colorectal cancer: evidence for distinct genetic categories based on proximal or distal tumor location. Ann Intern Med 1990; 113: 779-788 [PMID: 2240880 DOI: 10.7326/0003-4819-113-10-779]
- 31 Irving MH, Catchpole B. ABC of colorectal diseases. Anatomy and physiology of the colon, rectum, and anus. BMJ 1992; 304: 1106-1108 [PMID: 1586826 DOI: 10.1136/bmj.304.6834.1106]
- Minoo P, Zlobec I, Peterson M, Terracciano L, Lugli A. Characterization of rectal, proximal and 32 distal colon cancers based on clinicopathological, molecular and protein profiles. Int J Oncol 2010; 37: 707-718 [PMID: 20664940 DOI: 10.3892/ijo_00000720]
- Szmulowicz UM, Hull TL. Colonic Physiology. In: Beck DE, Roberts PL, Saclarides TJ, Senagore 33 AJ, Stamos MJ, Wexner SD. The ASCRS Textbook of Colon and Rectal Surgery. New York, NY: Springer, 2011: 23-39 [DOI: 10.1007/978-1-4419-1584-9_2]
- Li FY, Lai MD. Colorectal cancer, one entity or three. J Zhejiang Univ Sci B 2009; 10: 219-229 34 [PMID: 19283877 DOI: 10.1631/jzus.B0820273]
- Lim U, Song MA. Dietary and lifestyle factors of DNA methylation. Methods Mol Biol 2012; 863: 35 359-376 [PMID: 22359306 DOI: 10.1007/978-1-61779-612-8_23]
- Busch EL, Galanko JA, Sandler RS, Goel A, Keku TO. Lifestyle Factors, Colorectal Tumor 36 Methylation, and Survival Among African Americans and European Americans. Sci Rep 2018; 8: 9470 [PMID: 29930328 DOI: 10.1038/s41598-018-27738-x]
- Wang SM, Jiang B, Deng Y, Huang SL, Fang MZ, Wang Y. Clinical significance of MLH1/MSH2 37 for stage II/III sporadic colorectal cancer. World J Gastrointest Oncol 2019; 11: 1065-1080 [PMID: 31798786 DOI: 10.4251/wjgo.v11.i11.1065]
- Toh JWT, Phan K, Reza F, Chapuis P, Spring KJ. Rate of dissemination and prognosis in early and 38 advanced stage colorectal cancer based on microsatellite instability status: systematic review and meta-analysis. Int J Colorectal Dis 2021; 36: 1573-1596 [PMID: 33604737 DOI: 10.1007/s00384-021-03874-1
- 39 Popat S, Hubner R, Houlston RS. Systematic review of microsatellite instability and colorectal cancer prognosis. J Clin Oncol 2005; 23: 609-618 [PMID: 15659508 DOI: 10.1200/JCO.2005.01.086]
- 40 Condelli V, Calice G, Cassano A, Basso M, Rodriquenz MG, Zupa A, Maddalena F, Crispo F, Pietrafesa M, Aieta M, Sgambato A, Tortora G, Zoppoli P, Landriscina M. Novel Epigenetic Eight-Gene Signature Predictive of Poor Prognosis and MSI-Like Phenotype in Human Metastatic Colorectal Carcinomas. Cancers (Basel) 2021; 13 [PMID: 33466447 DOI: 10.3390/cancers13010158]
- Huang Z, Chen X, Liu C, Cui L. The Clinical Significance of Microsatellite Instability in Precision Treatment. Methods Mol Biol 2020; 2204: 33-38 [PMID: 32710312 DOI: 10.1007/978-1-0716-0904-0 3]
- Diao Z, Han Y, Chen Y, Zhang R, Li J. The clinical utility of microsatellite instability in colorectal 42 cancer. Crit Rev Oncol Hematol 2021; 157: 103171 [PMID: 33290824 DOI: 10.1016/j.critrevonc.2020.103171]
- 43 García-Alfonso P, García-Carbonero R, García-Foncillas J, Pérez-Segura P, Salazar R, Vera R, Ramón Y Cajal S, Hernández-Losa J, Landolfi S, Musulén E, Cuatrecasas M, Navarro S. Update of the recommendations for the determination of biomarkers in colorectal carcinoma: National Consensus of the Spanish Society of Medical Oncology and the Spanish Society of Pathology. Clin Transl Oncol 2020; 22: 1976-1991 [PMID: 32418154 DOI: 10.1007/s12094-020-02357-z]
- Sepulveda AR, Hamilton SR, Allegra CJ, Grody W, Cushman-Vokoun AM, Funkhouser WK, 44 Kopetz SE, Lieu C, Lindor NM, Minsky BD, Monzon FA, Sargent DJ, Singh VM, Willis J, Clark J, Colasacco C, Rumble RB, Temple-Smolkin R, Ventura CB, Nowak JA. Molecular Biomarkers for the Evaluation of Colorectal Cancer: Guideline From the American Society for Clinical Pathology, College of American Pathologists, Association for Molecular Pathology, and the American Society of Clinical Oncology. J Clin Oncol 2017; 35: 1453-1486 [PMID: 28165299 DOI: 10.1200/JCO.2016.71.9807



0 WŰ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2114-2128

DOI: 10.4251/wjgo.v13.i12.2114

ISSN 1948-5204 (online)

ORIGINAL ARTICLE

Basic Study Propofol induces ferroptosis and inhibits malignant phenotypes of gastric cancer cells by regulating miR-125b-5p/STAT3 axis

Yi-Ping Liu, Zhong-Zhi Qiu, Xu-Hui Li, En-You Li

ORCID number: Yi-Ping Liu 0000-0001-9992-8789; Zhong-Zhi Qiu 0000-0001-5463-7646; Xu-Hui Li 0000-0002-7887-4440; En-You Li 0000-0001-9594-2508.

Author contributions: Liu YP and Qiu ZZ designed the study; Liu YP and Li XH performed the experiments; Qiu ZZ collected and analyzed data; Li EY wrote the manuscript.

Institutional review board

statement: This study was reviewed and approved by the Ethics Committee of the First Affiliated Hospital of Harbin Medical University.

Institutional animal care and use committee statement: All animal experiments were performed under the approval of Animal Ethics Committee of The First Affiliated Hospital of Harbin Medical University.

Conflict-of-interest statement: The authors declare no conflict of interest.

Data sharing statement: No additional data are available.

ARRIVE guidelines statement: The authors have read the ARRIVE Guidelines, and the manuscript was prepared and revised

Yi-Ping Liu, Zhong-Zhi Qiu, En-You Li, Department of Anesthesiology, First Affiliated Hospital of Harbin Medical University, Harbin 150001, Heilongjiang Province, China

Xu-Hui Li, Department of Gastroenterology, Heilongjiang Forest Industry Federation (Red Cross) Hospital, Harbin 150008, Heilongjiang Province, China

Corresponding author: En-You Li, PhD, Chief Physician, Department of Anesthesiology, First Affiliated Hospital of Harbin Medical University, No. 23 Youzheng Street, Nangang District, Harbin 150001, Heilongjiang Province, China. enyouli@sina.com

Abstract

BACKGROUND

Gastric cancer is a common malignancy with poor prognosis, in which ferroptosis plays a crucial function in its development. Propofol is a widely used anesthetic and has antitumor potential in gastric cancer. However, the effect of propofol on ferroptosis during gastric cancer progression remains unreported.

AIM

To explore the function of propofol in the regulation of ferroptosis and malignant phenotypes of gastric cancer cells.

METHODS

MTT assays, colony formation assays, Transwell assays, wound healing assay, analysis of apoptosis, ferroptosis measurement, luciferase reporter gene assay, and quantitative reverse transcription polymerase chain reaction were used in this study.

RESULTS

Our data showed that propofol was able to inhibit proliferation and induce apoptosis of gastric cancer cells. Meanwhile, propofol markedly repressed the invasion and migration of gastric cancer cells. Importantly, propofol enhanced the erastin-induced inhibition of growth of gastric cancer cells. Consistently, propofol increased the levels of reactive oxygen species, iron, and Fe2+ in gastric cancer cells. Moreover, propofol suppressed signal transducer and activator of transcription (STAT)3 expression by upregulating miR-125b-5p and propofol induced ferroptosis by targeting STAT3 in gastric cancer cells. The miR-125b-5p inhibitor or STAT3 overexpression reversed propofol-attenuated malignant phenotypes of gastric cancer cells.



according to the ARRIVE Guidelines.

Country/Territory of origin: China

Specialty type: Gastroenterology and hepatology

Provenance and peer review:

Unsolicited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: July 6, 2021 Peer-review started: July 6, 2021 First decision: July 26, 2021 Revised: September 10, 2021 Accepted: October 25, 2021 Article in press: October 25, 2021 Published online: December 15, 2021

P-Reviewer: Chisthi MM, Yashiro M S-Editor: Gong ZM L-Editor: Kerr C P-Editor: Gong ZM



CONCLUSION

Propofol induced ferroptosis and inhibited malignant phenotypes of gastric cancer cells by regulating the miR-125b-5p/STAT3 axis. Propofol may serve as a potential therapeutic candidate for gastric cancer.

Key Words: Gastric cancer; Progression; Ferroptosis; Propofol; miR-125b-5p; STAT3

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: In this study, we discovered that propofol induced ferroptosis and inhibited malignant phenotypes of gastric cancer cells by regulating the miR-125b-5p/STAT3 axis. Propofol may serve as a potential therapeutic candidate for gastric cancer.

Citation: Liu YP, Qiu ZZ, Li XH, Li EY. Propofol induces ferroptosis and inhibits malignant phenotypes of gastric cancer cells by regulating miR-125b-5p/STAT3 axis. World J Gastrointest Oncol 2021; 13(12): 2114-2128

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2114.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2114

INTRODUCTION

Gastric cancer is a severe, lethal type of cancer worldwide[1]. Despite improvement in treatment, gastric cancer remains the leading cause of cancer-associated death[2]. Adjuvant therapy, radical surgery, and early diagnosis enhance survival rates and prognosis for gastric cancer, but mortality is still unsatisfactory[3,4]. Ferroptosis is a type of regulated cell death that differs from apoptosis and is repressed by erastin, especially in RAS-mutated cancer cells[5]. It has been identified that the suppression of ferroptosis contributes to cancer progression, and ferroptosis has a crucial role in the development of gastric cancer^[6-8]. However, the exploration of a treatment that can target ferroptosis is still limited.

Propofol is a broadly applied anesthetic because of rapid recovery and has some nonanesthetic functions in cancer development[9]. It has been identified that propofol inhibits cell invasion and growth and induces apoptosis in pancreatic cancer^[10]. Meanwhile, propofol decreases cell proliferation and increases apoptosis of lung cancer cells by regulating caspases-3, Bim, forkhead box (FOX)O1, and FOXO3, in which miR-486 inhibitor can reverse this effect[11]. Propofol also represses the malignant progression of promyelocytic leukemia cells[12]. Moreover, it has been reported that propofol induces an inhibitory effect on gastric cancer cell invasion and migration in patients with gastric cancer [13,14]. However, the function of propofol in the modulation of ferroptosis during gastric cancer progression remains elusive. miRNAs can modulate gene expression in different cellular processes[15]. Previous studies have revealed that miRNAs participate in the development of gastric cancer [16,17]. Meanwhile, a recent study has shown that miR-125b-5p is a tumor suppressor and involved in the modulation of gastric cancer[18]. In addition, signal transducer and activator of transcription (STAT)3 has been identified as an oncogene in gastric cancer[19-21]. However, the effect of propofol on miR-125b-5p and STAT3 during the development of gastric cancer remains obscure.

In this study, we focused on the investigation of the function of propofol in the development of gastric cancer. We found that propofol induced ferroptosis and inhibited malignant phenotypes of gastric cancer cells by regulating the miR-125b-5p/ STAT3 axis.

MATERIALS AND METHODS

Cell culture

The SGC7901 and BGC823 cells were maintained in the laboratory. The cells were cultured in Dulbecco's modified Eagle's medium (DMEM; Solarbio, China) with 0.1 mg/mL streptomycin (Solarbio), 100 U/mL penicillin (Solarbio), and 10% fetal bovine



serum (Solarbio), under the conditions of 37 °C with 5% CO₂. The pcDNA3.1-STAT3 overexpression vector, miR-125b-5p mimic, miR-125b-5p inhibitor, and corresponding control were purchased from Genscript Biotech Corporation (Nanjing, China) and GenePharma Co. Ltd. (Shanghai, China). The transfection in the cells was performed by Liposome 3000 (Invitrogen, Carlsbad, CA, United States). Propofol was purchased from Sigma (St. Louis, MO, United States).

MTT assays

SGC7901 and BGC823 cells were treated with propofol for 48 h. MTT assays analyzed the proliferation of SGC7901 and BGC823 cells. About 2 × 10⁵ cells were plated in 96well plates and incubated for 24 h. To assess cell viability, the cells were cultured with MTT solution (5 mg/mL) and incubated for 4 h, and 150 µL dimethyl sulfoxide (DMSO) was applied to treat the cells. The cell viability was measured at 570 nm absorbance by applying an ELISA browser (Bio-Tek EL 800, Winooski, VT, United States).

Colony formation assays

The SGC7901 and BGC823 cells were treated with propofol for 48 h. Colony formation assays measured proliferation. About 104 SGC7901 and BGC823 cells were placed in six-well plates and cultured in DMEM at 37 C. The cells were cleaned with phosphatebuffered saline (PBS) after 2 wk, washed in methanol for ~30 min, and stained with 1% crystal violet dye, after which, the number of colonies was calculated.

Transwell assays

To analyze cell migration, the cells were cultured for 24 h and resuspended in serumfree culture medium, then plated into the apical chamber of a Transwell chamber at 5 \times 10³ cells/well. The culture medium was made up to 150 µL and 600 µL complete culture medium was added to the basolateral chamber. After 24 h culture at 37°C and 5% CO_{γ} the cells were fixed with 4% paraformaldehyde for 10 min, stained by crystal violet dye for 20 min, followed by analysis using the intelligent biological navigator (Olympus, Tokyo, Japan). The migrated cells were recorded and calculated by using the ImageJ software.

To analyze cell invasion, Matrigel was melted overnight at 4 °C and diluted by presold serum-free culture medium (ratio 8:1). The medium (50 µL) was plated into the Transwell polycarbonate membrane with a pore diameter of 8 µm, covering all the wells with Matrigel at 37°C for 2 h. The cells were cultured for 24 h and resuspended in serum-free culture medium, plated into the Transwell apical chamber at 10^5 cells/well, and the medium was made up to 150 µL. Complete medium with 50% FBS (600 µL) was added to the basolateral chamber. After 24 h, the cells were fixed using 4% paraformaldehyde for 15 min and stained with crystal violet dye for 10 min. The invaded cells were analyzed and calculated using the ImageJ software.

Wound healing assay

SGC7901 and BGC823 cells were treated with propofol for 48 h. Approximately 3×10^5 SGC7901 and BGC823 cells were plated into the 24-well plates and incubated overnight to reach a fully confluent monolayer. A 20-µL pipette tip was applied to slowly cut a straight line across the well. The well was washed by PBS three times and the medium was changed to serum-free medium and culture was continued. The wound healing percentage was calculated.

Analysis of apoptosis

he SGC7901 and BGC823 cells were treated with propofol for 48 h. Approximately 2 × 105 SGC7901 and BGC823 cells were plated in six-well dishes. Apoptosis was determined using the Annexin V-FITC Apoptosis Detection Kit (Cell Signaling Technology, Danvers, MA, United States). About 2×10^5 washed cells were collected by binding buffer and stained at 25 C, followed by flow cytometry analysis.

Ferroptosis measurement

SGC7901 and BGC823 cells were cotreated with erastin (5 mmol/L) or ferrostatin (1 mmol/L). After 48 h, the cell viability was analyzed by MTT assay. Elevated iron level and accumulated lipid reactive oxygen species (ROS) were representative characteristics of ferroptosis. We used an iron assay kit (Beyotime, China) to examine the level of intracellular Fe2+. The cells were homogenized to collect the supernatant, incubated with iron reducer, followed by labeling with iron probe. OD 590 nm was detected in a microplate reader (PerkinElmer, Waltham, MA, United States). For detection of lipid



ROS, cells were stained with BODIPY C-11 dye (Beyotime) for 30 min, and subsequently detected by flow cytometry (BD Biosciences, Franklin Lakes, NJ, United States). The levels of malondialdehyde (MDA) and glutathione peroxidase (GSH) was measured by the MDA detection kit (Beyotime) and GSH assay kit (Cayman, Ann Arbor, MI, United States), respectively.

Luciferase reporter gene assay

The luciferase reporter gene assays were performed using the Dual-luciferase Reporter Assay System (Promega, Madison, WI, United States). SGC7901 and BGC823 cells were treated with miR-125b-5p mimic, pmirGLO-STAT3 (contained STAT3 3'UTR), and pmirGLO-STAT3 mutant transfected into the cells using Lipofectamine 3000 (Invitrogen), followed by analysis of luciferase activities, in which Renilla was applied as a normalized control.

Quantitative reverse transcription polymerase chain reaction

Total RNA was extracted by TRIZOL (Invitrogen). The first-strand cDNA was manufactured according to the manufacturer's instructions (Thermo Fisher Scientific, Waltham, MA, United States). qRT-PCR was carried out by SYBR Real-time PCR I kit (Takara, Japan). The standard control for miRNA and mRNA/circRNA was U6 and GAPDH, respectively. Quantitative determination of the RNA levels was conducted in triplicate independent experiments. The primer sequences were as follows: miR-125b-5p forward: 5'-TCCCTGAGACCCTAACTTGTGA-3'; reverse: 5'-AGTCTCAGGGTC CGAGGTATTC-3'; STAT3 forward: 5'-GGCCATCTTGAGCACTAAGC-3', reverse: 5'-CGGACTGGATCTGGGTCTTA-3'; GAPDH forward: 5'-TATGATGATATCAAGA GGGTAGT-3', reverse: 5'-TATGATGATATCAAGAGGGTAGT-3'; U6 forward: 5'-CTCGCTTCGGCAGCACA-3', U6 reverse: 5'-AACGCTTCACGAATT TGCGT-3'.

Western blot analysis

Total proteins were isolated from the cells with RIPA buffer (Cell Signaling Technology) and analyzed by BCA Protein Quantification Kit (Abbkine, United States). The protein was separated by 12% SDS-PAGE, and transferred to polyvinylidene difluoride membranes (Millipore, Billerica, MA, United States). The membranes were treated with 5% milk and incubated overnight at 4°C with the primary antibodies for STAT3 (Rabbit monoclonal, 1:1000, diluted by 5% milk; Cell Signaling Technology), GPX4 (Rabbit monoclonal, 1:1000, diluted by 5% milk; Cell Signaling Technology); and β -actin (Mouse monoclonal, 1:1000, diluted by 5% milk; Cell Signaling Technology); and β -actin (Rabbit monoclonal, 1:1000, diluted by 5% milk; Cell Signaling Technology), E-cadherin (Rabbit monoclonal, 1:1000, diluted by 5% milk; Cell Signaling Technology) and vimentin (Rabbit monoclonal, 1:1000, diluted by 5% milk; Cell Signaling Secondary antibodies (Abcam, Cambridge, MA, United States) were incubated with the membranes for 1 h at room temperature, followed by visualization using an Odyssey CLx Infrared Imaging System.

Xenograft assays

All animal experiments were performed under the approval of Animal Ethics Committee of The First Affiliated Hospital of Harbin Medical University. Specificpathogen-free male nude mice aged 5-6 wk and weighted around 20 g were purchased from Vitalriver (China). All mice were maintained in a 12-h circadian rhythm, and had free access to water and food. SGC7901 and BGC823 cells were subcutaneously injected into the right flank of mice. Propofol (50 mg/kg/d) was administrated intraperitoneally after tumor volume approached 100 mm³ for 20 d[22]. For the control group, the mice were treated with an equal volume of DMSO. Tumor volume and body weight were monitored every 5 d. The tumor size was calculated using the formula: length × width²/2.

Statistical analysis

Data were expressed as mean \pm SD, and the statistical analysis was conducted using GraphPad Prism 7. The unpaired Student's *t* test was used to compare two groups, and one-way analysis of variance was used to compare among multiple groups. *P* < 0.05 was considered statistically significant.

Zaishideng® WJGO | https://www.wjgnet.com

RESULTS

Propofol decreases proliferation and induces apoptosis of gastric cancer cells

We evaluated the effect of propofol on proliferation and apoptosis of gastric cancer cells. Propofol repressed viability of SGC7901 and BGC823 cells in a dose-dependent manner and 10 µmol/L propofol had a greater effect, which was selected in the subsequent analysis (Supplementary Figure 1). Propofol was able to inhibit viability of SGC7901 and BGC823 cells (Figure 1A and B). Similarly, propofol markedly reduced proliferation of SGC7901 and BGC823 cells (Figure 1C and D). Apoptosis of SGC7901 and BGC823 cells was enhanced by propofol (Figure 1E and F), suggesting that propofol decreases proliferation and induces apoptosis of gastric cancer cells.

Propofol reduces invasion and migration of gastric cancer cells

We further measured the effect of propofol on the migration and invasion of gastric cancer cells. Transwell assays indicated that the migration and invasion of SGC7901 and BGC823 cells were markedly decreased by propofol (Figure 2A and B). Consistently, the treatment of propofol significantly repressed wound healing in SGC7901 and BGC823 cells (Figure 2C and D), indicating that propofol is able to attenuate the migration and invasion of gastric cancer cells. Consistently, propofol enhanced Ecadherin expression and reduced vimentin expression in SGC7901 and BGC823 cells (Figure 2E).

Propofol enhances ferroptosis in gastric cancer cells

To analyze the impact of propofol on ferroptosis, we assessed the role of propofol in the erastin-induced inhibition of cell growth and the intracellular levels of ROS, iron and Fe²⁺, and expression of GPX4 and SLC7A11, which are considered to be ferroptosis markers. Propofol enhanced the erastin-induced inhibitory effect on SGC7901 and BGC823 cell growth, in which erastin served as an activator of ferroptosis (Figure 3A and B). Iron levels were induced by propofol in SGC7901 and BGC823 cells (Figure 3C). Propofol significantly promoted the levels of ROS in SGC7901 and BGC823 cells (Figure 3D). Propofol increased accumulation of Fe²⁺ in SGC7901 and BGC823 cells (Figure 3E). Consistently, the expression of GPX4 and SLC7A11 was inhibited by propofol in SGC7901 and BGC823 cells (Figure 3F). GSH levels were reduced and MDA levels were enhanced in SGC7901 and BGC823 cells by treatment with propofol (Figure 3G and H), suggesting that propofol enhances ferroptosis in gastric cancer cells.

Propofol represses STAT3 expression by upregulating miR-125b-5p in gastric cancer cells

We explored the potential mechanisms underlying propofol-mediated gastric cancer progression. Given that propofol can regulate colon cancer metastasis by regulating STAT3 signaling[23], we assessed the correlation of propofol with STAT3 in gastric cancer. Significantly, we identified that propofol was able to upregulate expression of miR-125b-5p in SGC7901 and BGC823 cells (Figure 4A). We identified the binding site between miR-125b-5p and STAT3 mRNA 3' UTR in a bioinformatic analysis using Targetscan (http://www.targetscan.org/vert_72/) (Figure 4B). Treatment with miR-125b-5p mimic reduced luciferase activities of wild-type STAT3, but not STAT3 with miR-125b-5p-binding site mutant in SGC7901 and BGC823 cells (Figure 4C and D). mRNA and protein expression of STAT3 was significantly suppressed by miR-125b-5p mimic in SGC7901 and BGC823 cells (Figure 4E). Propofol inhibited expression of STAT3, which was reversed by miR-125b-5p inhibitor (Figure 4F), indicating that propofol represses STAT3 expression by upregulating miR-125b-5p in gastric cancer cells.

Propofol enhances ferroptosis by targeting STAT3 in gastric cancer cells

We confirmed whether propofol modulated ferroptosis by targeting STAT3 in gastric cancer cells. As expected, overexpression of STAT3 was able to rescue propofolinhibited cell growth in the erastin-treated SGC7901 and BGC823 cells (Figure 5A). Similarly, STAT3 overexpression reversed propofol-induced levels of Fe²⁺, iron and ROS (Figure 5B-D), suggesting that propofol induces ferroptosis by inhibiting STAT3 in gastric cancer cells.

Propofol attenuates gastric cancer progression by miR-125b-5p /STAT3 axis

We further investigated the role of the propofol/miR-125b-5p/STAT3 axis in regulating gastric cancer malignant phenotypes. Overexpression of STAT3 or miR-





Figure 1 Propofol decreases proliferation and induces apoptosis of gastric cancer cells. A–E: SGC7901 and BGC823 cells were treated with propofol (10 μ mol/L). A and B: MTT assays analyzed cell viability; C and D: Colony formation assays measured cell proliferation; E and F: Flow cytometry analysis tested cell apoptosis. *n* = 3, mean ± SD, ^b*P* < 0.01.

Caishideng® WJGO | https://www.wjgnet.com

December 15, 2021 Volume 13 Issue 12







125b-5p inhibitor promoted propofol-inhibited viability in SGC7901 and BGC823 cells (Figure 6A and B). Consistently, apoptosis of SGC7901 and BGC823 cells was induced by propofol, and overexpression of STAT3 or miR-125b-5p inhibitor reversed this effect in SGC7901 and BGC823 cells (Figure 6C), implying that propofol has an inhibitor effect on gastric cancer malignant phenotypes via the miR-125b-5p/STAT3 axis. We confirmed that propofol-induced levels of Fe2+, iron and ROS were reversed by inhibition of miR-125b-5p in SGC7901 and BGC823 cells (Supplementary Figure 2).

Propofol attenuates growth of gastric cancer cells in vivo

We evaluated the effect of propofol on gastric cancer cell growth in a tumorigenicity analysis in nude mice. Tumor growth of SGC7901 or BGC823 cells was attenuated by propofol in nude mice (Figure 7A-C and Supplementary Figure 3A-C), as demonstrated by the reduced tumor size, weight and volume. As expected, expression of miR-125b-5p was enhanced and STAT3 expression was reduced in the tumor tissues of propofol-treated mice compared with the control group (Figure 7D and E and Supplementary Figure 3D and E). Expression of GPX4 and SLC7A11 was also downregulated by propofol in the tumor tissues of mice (Figure 7F and Supplementary Figure 3F).

DISCUSSION

Gastric cancer is a prevalent malignancy with high mortality[1], in which ferroptosis plays a critical role in its development[6-8]. Propofol is a widely used anesthetic and has inhibitory effects on cancer progression. Nevertheless, the effect of propofol on ferroptosis during the development of gastric cancer is still unreported. In this study, we showed that propofol induced ferroptosis and inhibited malignant phenotypes of gastric cancer cells by regulating the miR-125b-5p/STAT3 axis.

Propofol has presented significant anticancer functions in several models. It has been reported that propofol inhibits the development of nonsmall cell lung cancer by downregulating the miR215p/MAPK10 axis^[24]. Propofol represses malignant progression of pancreatic cancer cells by reducing NMDA receptor[25]. Propofol induces apoptosis of cervical cancer cells through inhibition of the HOTAIR/mTOR pathway[26]. Moreover, it has been found that propofol represses proliferation, migration and invasion by enhancing miR-195 in gastric cancer cells[14]. Propofol improves cisplatin sensitivity in gastric cancer by MALAT1/miR-30e/ATG5 signaling *via* inhibiting autophagy [27]. Propofol suppresses the survival and growth of gastric cancer by inducing expression of ING3[28]. We found that propofol decreased proliferation, migration and invasion and induced apoptosis of gastric cancer cells. Importantly, propofol enhanced ferroptosis in gastric cancer cells. Our data indicate an unreported function of propofol in the modulation of ferroptosis during gastric cancer progression, elucidating the novel role of the anesthetic in the ferroptosis of cancer development. Ferroptosis is one of the critical malignant phenotypes mediated by propofol in gastric cancer progression. The importance of ferroptosis and apoptosis should be compared by more complex investigations. Meanwhile, it has been reported that the regulation of ferroptosis may benefit the inhibition of gastric cancer development, and targeting ferroptosis may be a promising strategy for gastric cancer therapy.



Figure 3 Propofol enhances ferroptosis in gastric cancer cells. A and B: SGC7901 and BGC823 were cotreated with 5 mmol/L erastin or ferrostatin (1 mmol/L) and propofol (10 µmol/L). Cell growth was analyzed by MTT assays. C-F: SGC7901 and BGC823 cells were treated with propofol (10 µmol/L). C: Flow cytometry measured the levels of ROS. D and E: Iron Assay Kit analyzed the levels of iron and Fe2+. F: Western blotting analysis tested the expression of GPX4, SLC7A11 and β -actin. G and H: Levels of GSH and MDA were analyzed by the detection kit. n = 3, mean \pm SD, ${}^{3}P < 0.05$, ${}^{b}P < 0.01$, ${}^{c}P < 0.001$.

> miRNAs function as crucial regulators and are involved in gastric cancer progression. It has been reported that miR-96-5p enhances gastric cancer cell proliferation via inhibiting FOXO3[29]. miR-27b inhibits gastric cancer metastasis by

Raishideng® WJGO | https://www.wjgnet.com



Figure 4 Propofol represses signal transducer and activator of transcription (STAT)3 expression by upregulating miR-125b-5p in gastric cancer cells. A: SGC7901 and BGC823 cells were treated with propofol (10 µmol/L). Quantitative reverse transcription polymerase chain reaction (qRT-PCR) measured expression of miR-125b-5p. B: The binding site of miR-125b-5p and STAT3 3' UTR was identified by bioinformatic analysis using Targetscan (http://www.targetscan.org/vert_72/). C-E: SGC7901 and BGC823 cells were treated with the miR-125b-5p mimic or control mimic. C and D: Luciferase reporter gene assays determined the luciferase activities. E: qRT-PCR analyzed mRNA expression of STAT3. F: SGC7901 and BGC823 cells were treated with propofol, or cotreated with propofol and miR-125b-5p inhibitor. Western blotting assessed protein expression of STAT3 and β -actin. n = 3, mean ± SD, ^bP < 0.01.

downregulating NR2F2 (nuclear receptor subfamily 2 group F member 2)[30]. miR-558 contributes to the progression of gastric cancer by repressing Smad4-regulated heparanase expression[31]. Moreover, it has been reported that miR-125b-5p represses invasion, migration and proliferation of breast cancer cells by inhibiting KIAA1522 [32]. miR-125b-5p suppresses invasion, migration and proliferation of hepatocellular carcinoma cell by downregulating thioredoxin reductase 1[33]. miR-125b-5p inhibits the progression of bladder cancer by attenuating PI3K/AKT signaling and targeting hexokinase 2[34]. In the present study, we found that propofol inhibited STAT3 expression by upregulating miR-125b-5p in gastric cancer cells. Propofol induced



Figure 5 Propofol enhances ferroptosis by targeting signal transducer and activator of transcription (STAT)3 in gastric cancer cells. A:

Salishideng® WJGO | https://www.wjgnet.com

SGC7901 and BGC823 cells were treated with 5 mmol/L erastin, cotreated with 5 mmol/L erastin and propofol, or cotreated with 5 mmol/L erastin, propofol, and pcDNA.1-STAT3. MTT assays measured cell growth. B–D: SGC7901 and BGC823 cells were treated with propofol, or cotreated with propofol and pcDNA.1-STAT3. B: Iron Assay Kit analyzed the levels of iron; C: Flow cytometry analysis tested the levels of ROS; and D: Iron Assay Kit analyzed the levels of Fe²⁺. n = 3, mean ± SD, ${}^{b}P$ < 0.01.



Figure 6 Propofol attenuates gastric cancer progression by miR-125b-5p/STAT3 axis. A–C: SGC7901 and BGC823 cells were treated propofol, or cotreated with propofol and miR-125b-5p inhibitor or pcDNA.1-STAT3. A and B: MTT assays analyzed the cell viability; C: Flow cytometry measured apoptosis. n = 3, mean \pm SD, $^{b}P < 0.01$.

ferroptosis by targeting STAT3 in gastric cancer cells. The overexpression of STAT3 and miR-125b-5p inhibitor could reverse propofol-attenuated malignant phenotypes of gastric cancer cells. It uncovers a novel mechanism involving propofol, miR-125b-5p and STAT3 in the regulation of gastric cancer, enriching the understanding of the anticancer effect of propofol.

CONCLUSION

We discovered that propofol induced ferroptosis and inhibited malignant phenotypes of gastric cancer cells by regulating the miR-125b-5p/STAT3 axis. Propofol may serve as a potential therapeutic candidate for gastric cancer therapy.

Zaishidena® WJGO | https://www.wjgnet.com



Figure 7 Propofol attenuates growth of gastric cancer cells in vivo. The nude mice were injected with SGC7901 cells and intraperitoneally treated with propofol (50 mg/kg). A: Tumor tissues; B: Tumor volume; and C: Tumor weight; D: Expression of miR-125b-5p was analyzed by quantitative reverse transcription polymerase chain reaction. E: Protein expression of STAT3 was detected by western blotting. F: Protein expression of GPX4 and SLC7A11 was measured by western blotting. n = 5, mean \pm SD, ^bP < 0.01.

ARTICLE HIGHLIGHTS

Research background

Gastric cancer is a common malignancy with poor prognosis, in which ferroptosis plays a crucial role in its development. Propofol is a widely used anesthetic and has shown antitumor potential in gastric cancer. However, the effect of propofol on ferroptosis during gastric cancer progression remains unreported.

Research motivation

This study aims to identify the function of propofol in the regulation of ferroptosis and malignant phenotypes of gastric cancer cells.

Research objectives

To explore the role of propofol in the regulation of ferroptosis and malignant phenotypes of gastric cancer cells.



Baishidena® WJGO | https://www.wjgnet.com

Research methods

MTT assays, colony formation assays, Transwell assays, wound healing assay, analysis of cell apoptosis, ferroptosis measurement, luciferase reporter gene assay and quantitative reverse transcription-PCR were used in this study.

Research results

Propofol was able to inhibit proliferation and induce apoptosis of gastric cancer cells. Propofol markedly repressed the invasion and migration of gastric cancer cells. Importantly, propofol enhanced the erastin-induced inhibitory effect on the growth of gastric cancer cells. Consistently, propofol increased the levels of ROS, iron and Fe²⁺ in gastric cancer cells. Propofol suppressed STAT3 expression by upregulating miR-125b-5p and propofol induced ferroptosis by targeting STAT3 in gastric cancer cells. The miR-125b-5p inhibitor or STAT3 overexpression could reverse propofol-attenuated malignant phenotypes of gastric cancer cells.

Research conclusions

Propofol induced ferroptosis and inhibited malignant phenotypes of gastric cancer cells by regulating the miR-125b-5p/STAT3 axis.

Research perspectives

Propofol may serve as a potential therapeutic candidate for gastric cancer therapy.

REFERENCES

- Smyth EC, Nilsson M, Grabsch HI, van Grieken NC, Lordick F. Gastric cancer. Lancet 2020; 396: 635-648 [PMID: 32861308 DOI: 10.1016/S0140-6736(20)31288-5]
- 2 Machlowska J, Baj J, Sitarz M, Maciejewski R, Sitarz R. Gastric Cancer: Epidemiology, Risk Factors, Classification, Genomic Characteristics and Treatment Strategies. Int J Mol Sci 2020; 21 [PMID: 32512697 DOI: 10.3390/ijms21114012]
- Sexton RE, Al Hallak MN, Diab M, Azmi AS. Gastric cancer: a comprehensive review of current and 3 future treatment strategies. Cancer Metastasis Rev 2020; 39: 1179-1203 [PMID: 32894370 DOI: 10.1007/s10555-020-09925-3]
- 4 Bekaii-Saab T, El-Rayes B. Identifying and targeting cancer stem cells in the treatment of gastric cancer. Cancer 2017; 123: 1303-1312 [PMID: 28117883 DOI: 10.1002/cncr.30538]
- 5 Nehring H, Meierjohann S, Friedmann Angeli JP. Emerging aspects in the regulation of ferroptosis. Biochem Soc Trans 2020; 48: 2253-2259 [PMID: 33125483 DOI: 10.1042/BST20200523]
- 6 Hao S, Yu J, He W, Huang Q, Zhao Y, Liang B, Zhang S, Wen Z, Dong S, Rao J, Liao W, Shi M. Cysteine Dioxygenase 1 Mediates Erastin-Induced Ferroptosis in Human Gastric Cancer Cells. Neoplasia 2017; 19: 1022-1032 [PMID: 29144989 DOI: 10.1016/j.neo.2017.10.005]
- 7 Lee JY, Nam M, Son HY, Hyun K, Jang SY, Kim JW, Kim MW, Jung Y, Jang E, Yoon SJ, Kim J, Seo J, Min JK, Oh KJ, Han BS, Kim WK, Bae KH, Song J, Huh YM, Hwang GS, Lee EW, Lee SC. Polyunsaturated fatty acid biosynthesis pathway determines ferroptosis sensitivity in gastric cancer. Proc Natl Acad Sci U S A 2020; 117: 32433-32442 [PMID: 33288688 DOI: 10.1073/pnas.2006828117
- Zhang H, Deng T, Liu R, Ning T, Yang H, Liu D, Zhang Q, Lin D, Ge S, Bai M, Wang X, Zhang L, 8 Li H, Yang Y, Ji Z, Wang H, Ying G, Ba Y. CAF secreted miR-522 suppresses ferroptosis and promotes acquired chemo-resistance in gastric cancer. Mol Cancer 2020; 19: 43 [PMID: 32106859 DOI: 10.1186/s12943-020-01168-8]
- 9 Vasileiou I, Xanthos T, Koudouna E, Perrea D, Klonaris C, Katsargyris A, Papadimitriou L. Propofol: a review of its non-anaesthetic effects. Eur J Pharmacol 2009; 605: 1-8 [PMID: 19248246 DOI: 10.1016/j.ejphar.2009.01.007]
- Liu Z, Zhang J, Hong G, Quan J, Zhang L, Yu M. Propofol inhibits growth and invasion of pancreatic 10 cancer cells through regulation of the miR-21/Slug signaling pathway. Am J Transl Res 2016; 8: 4120-4133 [PMID: 27829997]
- 11 Yang N, Liang Y, Yang P, Yang T, Jiang L. Propofol inhibits lung cancer cell viability and induces cell apoptosis by upregulating microRNA-486 expression. Braz J Med Biol Res 2017; 50: e5794 [PMID: 28076456 DOI: 10.1590/1414-431X20165794]
- 12 Tsuchiya M, Asada A, Arita K, Utsumi T, Yoshida T, Sato EF, Utsumi K, Inoue M. Induction and mechanism of apoptotic cell death by propofol in HL-60 cells. Acta Anaesthesiol Scand 2002; 46: 1068-1074 [PMID: 12366500 DOI: 10.1034/j.1399-6576.2002.460903.x]
- 13 Ai L, Wang H. Effects of propofol and sevoflurane on tumor killing activity of peripheral blood natural killer cells in patients with gastric cancer. J Int Med Res 2020; 48: 300060520904861 [PMID: 32216484 DOI: 10.1177/0300060520904861]
- 14 Zhang W, Wang Y, Zhu Z, Zheng Y, Song B. Propofol inhibits proliferation, migration and invasion of gastric cancer cells by up-regulating microRNA-195. Int J Biol Macromol 2018; 120: 975-984



[PMID: 30171944 DOI: 10.1016/j.ijbiomac.2018.08.173]

- Sayed D, Abdellatif M. MicroRNAs in development and disease. Physiol Rev 2011; 91: 827-887 15 [PMID: 21742789 DOI: 10.1152/physrev.00006.2010]
- 16 Huangfu L, He Q, Han J, Shi J, Li X, Cheng X, Guo T, Du H, Zhang W, Gao X, Luan F, Xing X, Ji J. MicroRNA-135b/CAMK2D Axis Contribute to Malignant Progression of Gastric Cancer through EMT Process Remodeling. Int J Biol Sci 2021; 17: 1940-1952 [PMID: 34131397 DOI: 10.7150/ijbs.58062]
- 17 Miliotis C, Slack FJ. miR-105-5p regulates PD-L1 expression and tumor immunogenicity in gastric cancer. Cancer Lett 2021; 518: 115-126 [PMID: 34098061 DOI: 10.1016/j.canlet.2021.05.037]
- 18 Deng P, Sun M, Zhao WY, Hou B, Li K, Zhang T, Gu F. Circular RNA circVAPA promotes chemotherapy drug resistance in gastric cancer progression by regulating miR-125b-5p/STAT3 axis. World J Gastroenterol 2021; 27: 487-500 [PMID: 33642823 DOI: 10.3748/wjg.v27.i6.487]
- Ashrafizadeh M, Zarrabi A, Orouei S, Zarrin V, Rahmani Moghadam E, Zabolian A, Mohammadi S, 19 Hushmandi K, Gharehaghajlou Y, Makvandi P, Najafi M, Mohammadinejad R. STAT3 Pathway in Gastric Cancer: Signaling, Therapeutic Targeting and Future Prospects. Biology (Basel) 2020; 9 [PMID: 32545648 DOI: 10.3390/biology9060126]
- Cafferkey C, Chau I. Novel STAT 3 inhibitors for treating gastric cancer. Expert Opin Investig 20 Drugs 2016; 25: 1023-1031 [PMID: 27322026 DOI: 10.1080/13543784.2016.1195807]
- 21 Pan YM, Wang CG, Zhu M, Xing R, Cui JT, Li WM, Yu DD, Wang SB, Zhu W, Ye YJ, Wu Y, Wang S, Lu YY. STAT3 signaling drives EZH2 transcriptional activation and mediates poor prognosis in gastric cancer. Mol Cancer 2016; 15: 79 [PMID: 27938379 DOI: 10.1186/s12943-016-0561-z]
- 22 Gao Y, Yu X, Zhang F, Dai J. Propofol inhibits pancreatic cancer progress under hypoxia via ADAM8. J Hepatobiliary Pancreat Sci 2019; 26: 219-226 [PMID: 30945470 DOI: 10.1002/jhbp.624]
- 23 Zhang YF, Li CS, Zhou Y, Lu XH. Effects of propofol on colon cancer metastasis through STAT3/HOTAIR axis by activating WIF-1 and suppressing Wnt pathway. Cancer Med 2020; 9: 1842-1854 [PMID: 31953926 DOI: 10.1002/cam4.2840]
- 24 Wu X, Li X, Xu G. Propofol suppresses the progression of nonsmall cell lung cancer via downregulation of the miR215p/MAPK10 axis. Oncol Rep 2020; 44: 487-498 [PMID: 32468043] DOI: 10.3892/or.2020.7619]
- Chen X, Wu Q, You L, Chen S, Zhu M, Miao C. Propofol attenuates pancreatic cancer malignant 25 potential via inhibition of NMDA receptor. Eur J Pharmacol 2017; 795: 150-159 [PMID: 27986626 DOI: 10.1016/j.ejphar.2016.12.017]
- 26 Zhang D, Zhou XH, Zhang J, Zhou YX, Ying J, Wu GQ, Qian JH. Propofol promotes cell apoptosis via inhibiting HOTAIR mediated mTOR pathway in cervical cancer. Biochem Biophys Res Commun 2015; 468: 561-567 [PMID: 26523512 DOI: 10.1016/j.bbrc.2015.10.129]
- Zhang YF, Li CS, Zhou Y, Lu XH. Propofol facilitates cisplatin sensitivity via lncRNA 27 MALAT1/miR-30e/ATG5 axis through suppressing autophagy in gastric cancer. Life Sci 2020; 244: 117280 [PMID: 31926239 DOI: 10.1016/j.lfs.2020.117280]
- 28 Yang C, Gao J, Yan N, Wu B, Ren Y, Li H, Liang J. Propofol inhibits the growth and survival of gastric cancer cells in vitro through the upregulation of ING3. Oncol Rep 2017; 37: 587-593 [PMID: 27840947 DOI: 10.3892/or.2016.5218]
- 29 He X, Zou K. MiRNA-96-5p contributed to the proliferation of gastric cancer cells by targeting FOXO3. J Biochem 2020; 167: 101-108 [PMID: 31598681 DOI: 10.1093/jb/mvz080]
- Feng Q, Wu X, Li F, Ning B, Lu X, Zhang Y, Pan Y, Guan W. miR-27b inhibits gastric cancer 30 metastasis by targeting NR2F2. Protein Cell 2017; 8: 114-122 [PMID: 27844448 DOI: 10.1007/s13238-016-0340-z]
- 31 Zheng L, Jiao W, Song H, Qu H, Li D, Mei H, Chen Y, Yang F, Li H, Huang K, Tong Q. miRNA-558 promotes gastric cancer progression through attenuating Smad4-mediated repression of heparanase expression. Cell Death Dis 2016; 7: e2382 [PMID: 27685626 DOI: 10.1038/cddis.2016.293
- 32 Li Y, Wang Y, Fan H, Zhang Z, Li N. miR-125b-5p inhibits breast cancer cell proliferation, migration and invasion by targeting KIAA1522. Biochem Biophys Res Commun 2018; 504: 277-282 [PMID: 30177391 DOI: 10.1016/j.bbrc.2018.08.172]
- Hua S, Quan Y, Zhan M, Liao H, Li Y, Lu L. miR-125b-5p inhibits cell proliferation, migration, and 33 invasion in hepatocellular carcinoma via targeting TXNRD1. Cancer Cell Int 2019; 19: 203 [PMID: 31384178 DOI: 10.1186/s12935-019-0919-6]
- Liu S, Chen Q, Wang Y. MiR-125b-5p suppresses the bladder cancer progression via targeting HK2 34 and suppressing PI3K/AKT pathway. Hum Cell 2020; 33: 185-194 [PMID: 31605287 DOI: 10.1007/s13577-019-00285-x]

0 WĴ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2129-2148

DOI: 10.4251/wjgo.v13.i12.2129

ISSN 1948-5204 (online)

ORIGINAL ARTICLE

Basic Study BRAF^{VEDE} mutant colorectal cancer cells mediate local immunosuppressive microenvironment through exosomal long noncoding RNAs

Jie Zhi, Xiao-Jing Jia, Jing Yan, Hui-Cong Wang, Bo Feng, Han-Ying Xing, Yi-Tao Jia

ORCID number: Jie Zhi 0000-0003-1727-7746; Xiao-jing Jia 0000-0003-1783-1292; Jing Yan 0000-0002-0082-2747; Hui-Cong Wang 0000-0003-2023-5176; Bo Feng 0000-0002-5758-9130; Han-Ying Xing 0000-0002-0337-9541; Yi-Tao Jia 0000-0003-2610-9330.

Author contributions: Jia YT made substantial contributions to the conception and design of the study; Zhi J searched and reviewed published articles and wrote the manuscript; Jia XJ and Yan J collected patient data and prepared the tables; Zhi J, Jia XJ, Yan J and Wang HC performed the relevant experiments in the manuscript, statistically analyzed the data, and prepared the figures; Feng B and Xing HY critically reviewed the article and made revisions to the manuscript; all authors approved the final version.

Institutional review board

statement: The study was reviewed and approved by the Hebei General Hospital Institutional Review Board (approval No. 202134).

Conflict-of-interest statement: The authors report no conflicts of interest for this manuscript.

Jie Zhi, Hui-Cong Wang, Bo Feng, Yi-Tao Jia, Department of Oncology, Hebei General Hospital, Shijiazhuang 050051, Hebei Province, China

Xiao-Jing Jia, Department of Oncology, The First Hospital of Shijiazhuang, Shijiazhuang 050051, Hebei Province, China

Jing Yan, Department of Oncology, Puyang People's Hospital, Puyang 457000, Henan Province, China

Han-Ying Xing, Clinical Medical Research Center, Hebei General Hospital, Shijiazhuang 050051, Hebei Province, China

Corresponding author: Yi-Tao Jia, MD, Professor, Department of Oncology, Hebei General Hospital, No. 348 Heping West Road, Shijiazhuang 050051, Hebei Province, China. jiayitao99@163.com

Abstract

BACKGROUND

BRAFVGOOE mutated colorectal cancer (CRC) is prone to peritoneal and distant lymph node metastasis and this correlates with a poor prognosis. The BRAF^{V600E} mutation is closely related to the formation of an immunosuppressive microenvironment. However, the correlation between BRAF^{V600E} mutation and changes in local immune microenvironment of CRC is not clear.

AIM

To explore the effect and mechanism of BRAF^{V600E} mutant on the immune microenvironment of CRC.

METHODS

Thirty patients with CRC were included in this study: 20 in a control group and 10 in a treatment group. The density of microvessels and microlymphatic vessels, and M2 subtype macrophages in tumor tissues were detected by immunohistochemistry. Screening and functional analysis of exosomal long noncoding RNAs (lncRNAs) were performed by transcriptomics. The proliferation and migration of human umbilical vein endothelial cells (HUVECs) and human lymphatic endothelial cells (HLECs) were detected by CCK-8 assay and scratch test,



Data sharing statement: No additional data are available.

Country/Territory of origin: China

Specialty type: Oncology

Provenance and peer review:

Unsolicited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: July 13, 2021 Peer-review started: July 13, 2021 First decision: August 9, 2021 Revised: August 18, 2021 Accepted: October 25, 2021 Article in press: October 25, 2021 Published online: December 15, 2021

P-Reviewer: Saber A S-Editor: Gao CC L-Editor: Wang TQ P-Editor: Gao CC



respectively. The tube-forming ability of endothelial cells was detected by tube formation assay. The macrophage subtypes were obtained by flow cytometry. The expression of vascular endothelial growth factor (VEGF)-A, basic fibroblast growth factor (bFGF), transforming growth factor (TGF)- β 1, VEGF-C, claudin-5, occludin, zonula occludens (ZO)-1, fibroblast activation protein, and α -smooth muscle actin was assessed by western blot analysis. The levels of cytokines interleukin (IL)-6, TGF- β 1, and VEGF were assessed by enzyme-linked immunosorbent assay.

RESULTS

 $\mathsf{BRAF}^{\mathsf{V600E}}$ mutation was positively correlated with the increase of preoperative serum carbohydrate antigen 19-9 (P < 0.05), and with poor tumor tissue differentiation in CRC (P < 0.01). Microvascular density and microlymphatic vessel density in BRAF^{V600E} mutant CRC tissues were higher than those in BRAF wildtype CRC (P < 0.05). The number of CD163⁺M2 macrophages in BRAF^{V600E} mutant CRC tumor tissue was markedly increased (P < 0.05). Compared with exosomes from CRC cells with BRAF gene silencing, the expression of 13 lncRNAs and 192 mRNAs in the exosomes from BRAF^{V600E} mutant CRC cells was upregulated, and the expression of 22 lncRNAs and 236 mRNAs was downregulated (P < 0.05). The biological functions and signaling pathways predicted by differential lncRNA target genes and differential mRNAs were closely related to angiogenesis, tumor cell proliferation, differentiation, metabolism, and changes in the microenvironment. The proliferation, migration, and tube formation ability of HUVECs and HLECs induced by exosomes in the 1627 cell group (HT29 cells with BRAF gene silencing) was greatly reduced compared with the HT29 cell group (P < 0.05). Compared with the HT29 cell group, the expression levels of VEGF-A, bFGF, TGF- β 1, and VEGF-C in the exosomes derived from 1627 cells were reduced. The expression of ZO-1 in HUVECs, and claudin-5, occludin, and ZO-1 in HLECs of the 1627 cell group was higher. Compared with the 1627 cell group, the exosomes of the HT29 cell group promoted the expression of CD163 in macrophages (P <0.05). IL-6 secretion by macrophages in the HT29 cell group was markedly elevated (P < 0.05), whereas TGF- β 1 was decreased (P < 0.05). The levels of IL-6, TGF-β1, and VEGF secreted by fibroblasts in the 1627 cell group decreased, compared with the HT29 cell group (P < 0.05).

CONCLUSION

BRAF^{V600E} mutant CRC cells can reach the tumor microenvironment by releasing exosomal lncRNAs, and induce the formation of an immunosuppressive microenvironment.

Key Words: Colorectal cancer; BRAF^{v600E} mutant; Exosome; Long noncoding RNA; Immunosuppressive microenvironment

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: This study revealed that BRAF^{V600E} mutant colorectal cancer (CRC) cells could lead to more angiogenesis and lymphoangiogenesis in the microenvironment by releasing exosomal long noncoding RNAs, inducing the formation of an immunosuppressive microenvironment. Our findings provide a hypothesis for finding new therapeutic strategies for BRAF^{V600E} mutant CRC.

Citation: Zhi J, Jia XJ, Yan J, Wang HC, Feng B, Xing HY, Jia YT. BRAF^{v600E} mutant colorectal cancer cells mediate local immunosuppressive microenvironment through exosomal long noncoding RNAs. *World J Gastrointest Oncol* 2021; 13(12): 2129-2148 **URL:** https://www.wjgnet.com/1948-5204/full/v13/i12/2129.htm **DOI:** https://dx.doi.org/10.4251/wjgo.v13.i12.2129

Zaisbidene® WJGO | https://www.wjgnet.com

INTRODUCTION

Colorectal cancer (CRC) is one of the most common malignant tumors, and ranks third in morbidity and mortality globally^[1]. In China, societal and lifestyle changes have tended to increase the morbidity and mortality of CRC[2]. BRAF^{V600E} gene mutation accounts for approximately 10% of patients with metastatic CRC (mCRC), and is a point mutation at nucleotide 1799 of exon 15 (T mutated into A), resulting in a change of the encoded amino acid 600, valine replaced by glutamate (V600E)[3,4]. BRAF^{V600E} mutation can continuously activate the RAS-RAF-MEK-ERK signaling pathway, promote tumor cell proliferation and migration, and induce angiogenesis, thereby reducing tumor cell apoptosis[5-8]. Clinical data reveal that BRAFV600E mutation frequently occurs in elderly women, and the related pathological type is mostly mucinous adenocarcinoma with a high level of tissue differentiation. Around 20% of patients have accompanying microsatellite instability, and most of them develop right colon cancer originating from serrated adenomas[4,9]. Compared with wild-type BRAF, patients with BRAF^{V600E} mutant CRC are prone to peritoneal metastasis and distant lymph node metastasis with a poor prognosis. Generally, the median survival time is < 12 mo. The effective treatment rate reaches only approximately 20%, even with three-drug chemotherapy and targeted combination therapy [10,11]. It is therefore important to explore the mechanism of BRAF^{V600E} mutant CRC with lymph node and peritoneal metastasis and to discover effective therapeutic strategies. This may be applicable to start with its specific immune microenvironment.

The tumor microenvironment (TME) is the local environment facilitating tumor growth and proliferation[12]. In thyroid cancer, the proportion of mast cells in BRAF^{V600E} mutant TME is markedly increased compared with wild-type BRAF, which may be involved in mediating the formation of the immunosuppressive microenvironment, suggesting that BRAFV600E mutation affects the TME[13]. Multiple cells (namely, endothelial cells, fibroblasts, and immune cells) and extracellular components (namely, cytokines, growth factors, hormones, and extracellular matrix) in the TME can induce tumor angiogenesis and lymphangiogenesis and promote chronic inflammation, thereby creating a local immunosuppressive microenvironment, which plays a vital role in tumor occurrence, invasion, and metastasis and drug resistance[14]. Tumor-associated macrophages (TAMs) and cancer-associated fibroblasts (CAFs) are the principal components of the TME. Among them, M2-type macrophages have an immunosuppressive effect. They can promote tumor cell proliferation, infiltration, and metastasis, and their expression level is intimately associated with patient prognosis [15,16]. Meanwhile, CAFs have undergone substantial changes in morphology, proliferation activity, motility, and secretory function, which can facilitate tumor proliferation, invasion, metastasis, and angiogenesis, and their expression level is closely linked to tumor stage and poor prognosis[17]. There is no current research investigating the relationship between BRAFV600E gene mutation and the formation and functional changes of blood vessels, lymphatic vessels, TAMs, and CAFs in the CRC local immune microenvironment.

Exosomes are a type of cystic microvesicle secreted by cells, and the secretion process is active. They are able to carry specific biologically active molecules including lipids, miRNA, and long noncoding RNAs (lncRNAs) into the corresponding target cells and mediate substance transportation and information exchange intercellularly [18]. LncRNAs belong to a class of single-stranded RNA molecules that do not encode proteins. They play a role *via* transcription, post-transcription, and translation, and participate in tumor occurrence, invasion, and metastasis. Exosomes secreted by tumor cells can modify the TME through lncRNAs, thereby promoting the development of tumors[19,20]. Liang *et al*[21] noted that the expression of exosome-derived lncRNA RPPH1 in CRC was markedly upregulated. It can prevent ubiquitination by binding to TUBB3 and induce epithelial-mesenchymal transition, and interact with TAMs to promote the polarization of TAMs to M2 subtype, thereby accelerating tumor progression. However, there is still a lack of relevant investigations on whether BRAF^{V600E} mutant CRC cells affect the TME through the release of exosomal lncRNAs.

The present study aimed to investigate the influence of BRAF^{V600E} mutation in CRC on the surrounding immune microenvironment, and to elucidate whether BRAF^{V600E} mutant CRC cell-derived exosomes participate in the formation of an immunosuppressive microenvironment. It is hoped that the results will provide a novel therapeutic strategy for BRAF^{V600E} mutant CRC.

Zaishideng® WJGO | https://www.wjgnet.com

MATERIALS AND METHODS

General information

Data of ten BRAF^{V600E} mutant CRC patients who underwent surgical treatment at the Hebei General Hospital from September 2014 to June 2019 were collected. Twenty BRAF wild-type CRC patients were selected as controls. There were 18 male and 12 female patients. The age range was 27-79 years, with an average of 57.57 ± 2.13 years and median of 57 years. The specimens were obtained with informed consent obtained from the patients and under the approval of the Ethics Committee of the Fourth Hospital of Hebei Medical University. The inclusion criteria were: (1) Patients with a pathological diagnosis of CRC, in whom those identified as having BRAF^{V600E} mutation were included in an experimental group, and those identified as having wild-type BRAF were included in a control group; (2) Untreated patients; and (3) Undergoing first surgical treatment for primary CRC. The exclusion criteria were: (1) History of malignant tumors; (2) Current primary tumors in other regions; and (3) Patients with incomplete pathological data.

Cells and reagents

The human colon cancer cell line HT29 (BRAF^{V600E} mutant) was purchased from the Cell Bank of the Chinese Academy of Sciences. The human colon cancer cell line 1627 was obtained from Shanghai GenePharma Company, which was a HT29 cell strain with the BRAF gene being silenced. Human umbilical vein endothelial cells (HUVECs) and human lymphatic endothelial cells (HLECs) were purchased from ScienCell, San Diego, CA, United States. Human monocytic leukemia cells (THP-1 cells) and human embryonic lung fibroblasts (MRC-5 cells) were obtained from the Cell Bank of the Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences. McCoy's 5A, RPMI 1640, minimal essential medium, and fetal bovine serum (FBS) were all purchased from Gibco (NY, United States).

Induction of macrophages

THP-1 cells in the logarithmic phase of growth were obtained and placed in a 15-mL centrifuge tube, and centrifuged at 1000 r/min for 5 min. The supernatant was discarded. Following addition of fresh RPMI 1640 culture medium, the cells were resuspended and inoculated in a 12-well plate with 5×10^5 cells per well, supplemented with pharmaceutical manufacturers association (PMA) solution (dissolved in DMSO) (Cayman, MI, United States) at a final concentration of 100 ng/mL, and cultured in a constant temperature incubator for 48 h. The cells were ultimately placed under a biological microscope (Nikon, Tokyo, Japan) and the cell morphology and adhesion were observed.

Immunohistochemistry

The postoperative tissue samples of CRC patients were collected, embedded in paraffin, and cut into 5-mm sections. After deparaffinization and hydration, antigen retrieval, and incubation in 3% H₂O₂ for 10 min, the corresponding antibodies were added, including rabbit anti-human CD31 monoclonal antibody (1:2000, Arigo, Hsinchu City, Taiwan), rabbit LYVE-1 polyclonal antibody (1:200, Arigo), mouse antihuman CD68 monoclonal antibody (1:100, BD Biosciences, NJ, United States), rabbit anti-human CD163 monoclonal antibody (1:100, HUABIO, Hangzhou, China), and rabbit anti-human α-smooth muscle actin (SMA) monoclonal antibody (1:100, HUABIO). 3,3-diaminobenzidine (DAB) color development was performed for microscopic observation. A double-blind reading method was adopted by two pathologists. The sections were initially visualized under low magnification (× 100) to determine three fields where cells were most densely distributed. The number of positive cells was counted under high magnification (× 400), and the average number was calculated.

Extraction and identification of exosomes

Cell supernatant was collected and centrifuged at 480 × g for 5 min, followed by 2000 × g for 10 min to remove cell debris. The supernatant was collected and centrifuged at $10000 \times g$ for 30 min to remove macrovesicles. The supernatant was collected again and centrifuged at $100000 \times g$ for 2 h. The supernatant was discarded and the exosomes were resuspended in 200 mL of PBS solution, and stored in a freezer at -80 °C for later use. The morphology of exosomes was identified with a transmission electron microscope (Hitachi, Tokyo, Japan). At room temperature, 10 mL of exosome suspension was dripped on a 2-nm-pore-diameter copper mesh using a pipette and


allowed to stand for 2 min. The liquid was absorbed dry using absorbent paper. A quantity of 30 mL 3% tungsten phosphate was dripped onto the copper mesh for negative staining for 5 min, and the liquid was absorbed dry using absorbent paper. After drying, photographs were taken under a transmission electron microscope.

Western blot analysis

The cell samples were removed from the -80 °C freezer and supplemented with 200 mL of RIPA lysis solution for 20 min to extract the total cell proteins. The BCA protein concentration determination kit was used to quantify the proteins. The proteins were subjected to SDS-PAGE, transferred to polyvinylidene difluoride (PVDF) membranes at 250 mA, and blocked with bovine serum albumin for 2 h. The blocked PVDF membranes were placed directly into the freshly prepared primary antibody working solution at 4 °C overnight. Primary antibodies used included rabbit anti-human CD9 monoclonal antibody (1:1000, Abcam, Cambridge, United Kingdom), rabbit antihuman CD63 monoclonal antibody (1:5000, Abcam), rabbit anti-human BRAF monoclonal antibody (1:1000, Arigo), rabbit anti-zonula occludens (ZO)-1 polyclonal antibody (1:1000, GenTex, Gentex, United States), rabbit anti-claudin-5 polyclonal antibody (1:1000, GenTex), rabbit anti-occludin polyclonal antibody (1:1000, HUABIO), rabbit anti-transforming growth factor (TGF)-1 polyclonal antibody (1:1000, Bioss, Beijing, China), rabbit anti-vascular endothelial growth factor (VEGF)-C polyclonal antibody (1:800, Bioworld, MN, United States), rabbit anti-basic fibroblast growth factor (anti-bFGF) polyclonal antibody (1:600, Bioss), mouse anti-human VEGF-A monoclonal antibody at a concentration of 6 mg/mL (Abcam), rabbit anti-human fibroblast activation protein (FAP) monoclonal antibody (1:1000, Abcam), and rabbit anti-human α-SMA monoclonal antibody (1:1000, Abcam). Following membrane washing on the next day, the corresponding secondary antibody was incubated, and chemiluminescence was detected with ECL substrate. After developing and fixing, the film was scanned using a scanner, and Tanon 1600 software was used for gray scale analysis and quantification.

CCK-8 assay

Cells in the logarithmic phase of growth were selected and seeded in a 96-well plate at 5×10^3 cells/well with a volume of 100 mL per well, and cultured at 37 °C in a 5% CO₂ incubator. After 6 h of inoculation, the experimental group was treated with 10 mL exosomes, and the control group with 10 mL PBS solution, and then cultured in the incubator again. Exosomes were cocultured with HUVECs for 24, 48, and 72 h, and cocultured with HLECs for 12, 24 and 36 h; 10 mL of CCK-8 reagent (DOJINDO, Kyushu, Japan) was added to each well and incubated at 37 °C with 5% CO₂ for 1 h. Absorbance at 560 nm was measured using a microplate reader (ThermoFisher Scientific, MA, United States).

Scratch test

HUVECs or HLECs were plated in six-well plates. When the cells were evenly spread, they were scratched using a 200- μ L pipette tip. PBS solution was used to rinse once for the removal of the suspended cells. To each well, 2 mL of FBS-free culture medium was added. The experimental group was supplemented with 20 μ L of exosomes, and the control group with 20 μ L of PBS solution. Both groups were cultured at 37 °C in a 5% CO₂ incubator. The cells were observed and photographed under a microscope at 0 and 24 h, respectively, to detect the scratch healing. Cell migration rate was calculated as [(scratch area at 0 h - scratch area at 24 h)/scratch area at 0 h] × 100%.

Tube formation assay

After freezing and thawing the Matrigel matrix (BD Biosciences, United States) containing low levels of growth factors at 4 °C, the homogenate was mixed well with a precooled pipette tip and packaged into precooled Eppendorf tubes. Matrigel matrix was diluted with serum-free medium at a ratio of 1:3, and the 96-well plate was placed on an ice pack. Fifty microliters of diluted Matrigel matrix was added to each well and left to stand at 37 °C for 30 min. Cells were seeded in a 96-well plate at 10⁴ cells/well and supplemented with 100 μ L cell suspension in each well. The experimental group was treated with 10 μ L of exosomes, and the control group with 10 μ L PBS solution, and then cultured at 37 °C in a 5% CO₂ incubator. After culturing for 24 h, the cells were visualized and photographed under a low magnification (× 100) microscope and the tube formation ability of endothelial cells was observed.

Raishideng® WJGO | https://www.wjgnet.com

Flow cytometry

THP-1 cells were induced with PMA for 48 h and the culture medium was discarded following three cycles of washing with PBS. Trypsin (0.5 mL) was supplied to each well for digestion, and 1.5 mL of culture medium containing 10% FBS was utilized to terminate the digestion. The cells were then collected and centrifuged at 1000 r/min for 5 min. The supernatant was discarded, and the cells were resuspended in PBS and centrifuged again to obtain the precipitate. Mouse anti-human CD68 monoclonal antibody (5 μ L; BD Biosciences) and rabbit anti-human CD163 monoclonal antibody (5 μ L; BD Biosciences) were added and incubated at 4 °C in the dark for 30 min. Following two cycles of washing with PBS, the cells were centrifuged at 1000 r/min for 5 min, resuspended in 300 μ L PBS, and loaded on a flow cytometer (Beckman Coulter, CA, United States) for determination.

Enzyme-linked immunosorbent assay

After cells were cultured for 48 h, the medium was collected and centrifuged at 1000 r/min for 5 min. An enzyme-linked immunosorbent assay (ELISA) kit (MULTI SCIENCES, Hangzhou, China) was used to detect the expression of interleukin (IL)-6, TGF- β 1, and VEGF in the culture medium. IL-6 antibody, TGF- β 1 antibody, VEGF antibody, and horseradish peroxidase-labeled streptavidin were all diluted 1:100.

Extraction and identification of exosomal RNA

AllPrep RNA/LncRNA kit (Qiagen, Hilden, Germany) was used to extract total RNA from exosomes. The purity of RNA was determined using QubitRNA kit (Thermo Fischer Scientific). The RNA concentration was accurately quantified utilizing Qubit. The integrity of RNA was assessed using Agilent 2100.

Transcriptome library preparation, sequencing, and data analysis

Exosome samples of HT29 and 1627 cells were initially obtained and total exosomal RNA was extracted. Ovation Solo RNA kit (NugEN) was used for library construction of exosomal lncRNAs. The constructed library was sequenced through the Illumina Hiseq 4000 platform where the double-terminal 250-300 nt transcripts were generated. The raw data in Fastq format were processed using perl to obtain sequencing data. By logging in to the TopHat2 system, the sequenced data transcripts were aligned and analyzed in light of the reference genome. Cufflinks software was used to compare the analyzed data for transcript splicing and the transcriptome was obtained. Quantitative analysis was performed on the transcript. Finally, the transcript data set was retrieved using the RefSeq database as an mRNA data set. Cuffmerge software was used to merge the obtained transcripts after splicing, delete the transcripts with unclear midstrand direction, and obtain a mRNA transcript data set. LncRNAs were ultimately screened from the combined transcript set.

Prediction of IncRNA target genes and GO and KEGG enrichment analysis

The target genes of lncRNAs were predicted through cis-acting and trans-acting, including annotating the function of its target gene mRNA. The GO seq software and KOBAS software were used to enrich differential lncRNA target genes and differential mRNAs in the three processes of biological process, cell component, and molecular function. The biological functions involved in differential lncRNA target genes and differential mRNAs were analyzed. KOBAS software was used to enrich the differential lncRNA target genes and differential lncRNA target genes and differential mRNAs were analyzed. KOBAS software was used to enrich the differential lncRNA target genes and differential lncRNA target genes and differential mRNAs with KEGG pathways. The signal transduction pathways involved in differential lncRNA target genes and differential mRNA were also analyzed.

Statistical analysis

SPSS 21.0 software was used for statistical analyses. Enumerative data are expressed as percentages and analyzed using the χ^2 test or Fisher's exact test. The enumerative data are expressed as the mean ± SD. The two groups of enumerative data were tested by two independent samples *t*-tests. One-way analysis of variance was applied for multiple groups of samples. *P* < 0.05 was considered statistically significant.

RESULTS

Relationship between BRAF^{veoue} mutation and clinicopathological parameters

We collected and analyzed the clinical data of 10 BRAF^{V600E} mutant and 20 BRAF wildtype CRC patients. In CRC, BRAF^{V600E} mutation was positively correlated with the increase of preoperative serum carbohydrate antigen (CA)19-9 (P < 0.05), and it was correlated with poor tumor tissue differentiation (P < 0.01). However, no correlation was revealed with gender, age, location, mucous tissue, T stage, TNM stage, lymph node metastasis, nerve invasion, vascular tumor thrombus, preoperative carcinoembryonic antigen (CEA) level, or preoperative platelet count of the patients (Table 1).

BRAF^{VEDE} mutation promotes formation of microvessels and microlymphatic vessels in tumor tissues and increases infiltration of M2 macrophages

To explore the influence of BRAF^{V600E} mutation on the TME, we identified the formation of tumor blood vessels and lymph vessels in BRAF^{V600E} mutation and BRAF wild-type CRC tissues. Microvascular density (MVD) and microlymphatic vessel density (MLVD) in BRAF^{V600E} mutant CRC tissues were higher than those in BRAF wild-type CRC (P < 0.05) (Figure 1A and Table 2). The number of CD163⁺M2 macrophages in BRAF^{V600E} mutant CRC tumor tissue was markedly increased (P < 0.05) (Figure 1B and Table 3), whereas the number of CD68⁺M1 macrophages was not significantly different (Figure 1B and Table 3). Additionally, the density of fibroblasts exhibited no significant difference (Figure 1C and Table 4). These results suggest that BRAF^{V600E} mutation promotes the formation of microvessels and microlymphatic vessels in tumor tissues, increases the infiltration of M2 macrophages, and induces an immunosuppressive microenvironment.

BRAF^{vision} mutant CRC cells can reach the TME by releasing exosomal IncRNAs, and induce formation of an immunosuppressive microenvironment

We performed transcriptomics analysis on exosomes derived from BRAF^{V600E} mutant CRC cells and those with BRAF gene silencing. Compared with the exosomes from cells with BRAF gene silencing, the expression of 13 lncRNAs and 192 mRNAs in the exosomes of BRAF^{V600E} mutant CRC cells was upregulated, and the expression of 22 lncRNAs and 236 mRNAs was downregulated (P < 0.05) (Figure 2A). Using cluster analysis charts, the distribution of differentially expressed lncRNAs and mRNAs in exosomes of BRAF^{V600E} mutant CRC cells and those with BRAF gene silencing was further exhibited (Figure 2B).

To illustrate the biological functions of differential lncRNAs and mRNAs and the signaling pathways involved, we conducted GO enrichment analysis and KEGG signaling pathway analysis. GO enrichment analysis included three aspects of biological process, cell component, and molecular function. Differential lncRNA target genes and mRNAs presented similar biological functions, and they are mainly involved in nucleic acid metabolism, macromolecular metabolism, nitride metabolism, and RNA metabolism. Cellular components include mostly the formation of nuclei, organelles, and nucleosomes. Molecular functions involve regulation of cell adhesion, cytoskeletal remodeling, gene expression regulation, and protein binding (Figure 2C).

KEGG results indicated that the target genes of differential lncRNAs were mainly involved in the p53 pathway, ErbB pathway, steroid synthesis pathway, actin cytoskeleton regulation pathway, pyruvate metabolism pathway, cell cycle regulation pathway, and the pathway of protein processing in the endoplasmic reticulum (Figure 2D). Differential mRNAs are mainly involved in the VEGF pathway, mammalian target of rapamycin pathway, mitogen-activated protein kinase pathway, and the pathway of protein processing in the endoplasmic reticulum (Figure 2D). The biological functions and signaling pathways predicted by differential lncRNA target genes and differential mRNAs coincided with each other. Furthermore, it was closely related to angiogenesis, tumor cell proliferation, differentiation, metabolism, and changes in the microenvironment, suggesting that BRAFV600E mutant CRC cells could reach the TME by releasing exosomal lncRNAs, and induce formation of an immunosuppressive microenvironment through mRNAs.

BRAF^{V600E} mutant CRC cell-derived exosomes promote proliferation, migration, and tube formation of HUVECs and HLECs, and induce angiogenesis and lymphoangiogenesis

Exosomes derived from HT29 and 1627 cells were cocultured with HUVECs or HLECs to detect cell proliferation. OD values of HUVECs in the HT29 group and 1627 group



Table 1 Relationship between BRAF^{veone} mutation and clinicopathological parameters in colorectal cancer

		BRAF ^{V600E} mutation		
Variable	Cases (%)	Yes (%)	No (%)	 P value
Gender				
Male	18 (60)	6 (60)	12 (60)	0.326
Female	12 (40)	4 (40)	8 (40)	
Age (yr)				
≥65	10 (33.3)	4 (40)	6 (30)	0.440
< 65	20 (66.7)	6 (60)	14 (70)	
Location				
Right colon	9 (30)	2 (20)	7 (35)	0.345
Left colon	9 (30)	5 (50)	4 (20)	
Rectum	12 (40)	3 (30)	9 (45)	
Mucous tissue				
Positive	6 (20)	1 (10)	5 (25)	0.326
Negative	24 (80)	9 (90)	15 (75)	
Differentiated degree				
High/moderate differentiation	20 (66.7)	3 (30)	17 (85)	0.005 ^a
Poor differentiation	10 (33.3)	7 (70)	3 (15)	
T stage				
Τ2	4 (13.3)	0	4 (20)	0.211
Т3	3 (10)	2 (20)	1 (5)	
Τ4	23 (76.7)	8 (80)	15 (75)	
TNM stage				
I + II	13 (43.3)	2 (20)	11 (55)	0.074
III + IV	17 (56.7)	8 (80)	9 (45)	
Lymph node				
Positive	17 (56.7)	8 (80)	9 (45)	0.074
Negative	13 (43.3)	2 (20)	11 (55)	
Nerve invasion				
Positive	7 (23.3)	4 (40)	3 (15)	0.143
Negative	23 (76.7)	6 (60)	17 (85)	
Vessel carcinoma embolus				
Positive	3 (10)	1 (10)	2 (10)	0.652
Negative	27 (90)	9 (90)	18 (90)	
CA19-9 (U/mL)				
High	12 (40)	7 (70)	5 (25)	0.024 ^a
Normal	18 (60)	3 (30)	15 (75)	
CEA (ng/mL)				
High	11 (36.7)	3 (30)	8 (40)	0.452
Normal	19 (63.3)	7 (70)	12 (60)	
PLT count (× $10^9/L$)				
High	12 (40)	5 (50)	7 (35)	0.344

	Normal 18 (6	50) 5 (50)	13 (65)	
--	--------------	------------	---------	--

 $^{a}P < 0.05.$

PLT: Platelets; CEA: Carcinoembryonic antigen; CA19-9: Carbohydrate antigen 19-9.

Table 2 Relationship between BRAF ^{veene} mutation and microvascular density/microlymphatic vessel density in colorectal cancer					
	MVD		MLVD		
	mean ± SD	P value	mean ± SD	P value	
+	38.53 ± 17.11	0.030	9.67 ± 5.63	0.0001	
-	27.54 ± 9.54		3.81 ± 2.06		

MVD: Microvascular density; MLVD: Microlymphatic vessel density.

Table 3 Relationship between BRAF ^{veone} mutation and tumor-associated macrophages in colorectal cancer						
PDAEV600E mutation	CD68⁺ macrophage		CD163⁺ M2 macrophage			
BRAF	mean ± SD	P value	mean ± SD	<i>P</i> value		
+	18.43 ± 13.53	0.664	8.33 ± 5.93	0.040		
-	20.96 ± 15.34		3.67 ± 3.02			

Table 4 Relationship between BRAF^{V600E} mutation and cancer-associated fibroblasts in colorectal cancer

DDAEV600E mutation	CAFs	B volue	
DRAF Inutation	Low expression (cases)	High expression (cases)	r value
+	2	8	1
-	3	17	

CAF: Cancer-associated fibroblast.

at 24 h were 1.215 ± 0.032 and 0.986 ± 0.046, respectively; 1.563 ± 0.035 and 1.200 ± 0.163 at 48 h, respectively; and 1.661 ± 0.031 and 1.369 ± 0.020 at 72 h, respectively. At 24, 48 and 72 h, HUVEC proliferation induced by exosomes in the 1627 cell group was reduced compared with the HT29 cell group (P < 0.05), but there was no significant difference in HUVEC proliferation between the 1627 cell group and control group (Figure 2A). The OD values of HLECs in the control group, HT29 cell group, and 1627 cell group at 12 h were 0.477 ± 0.006, 0.526 ± 0.007, and 0.500 ± 0.004, respectively; 0.622 ± 0.003, 0.728 ± 0.010, and 0.680 ± 0.010 at 24 h, respectively; and 0.644 ± 0.006, 0.725 ± 0.009, and 0.682 ± 0.014 at 36 h, respectively. At 12, 24, and 36 h, the proliferation of HLECs induced by exosomes in the 1627 cell group was greatly reduced compared with the HT29 cell group (P < 0.05) and the proliferation of HLECs in the control group was greatly reduced compared with the HT29 cell group (P < 0.05) and the proliferation of HLECs in the 1627 cell group was greater than that of the control group (P < 0.05) (Figure 3A).

Light microscopy indicated that the average migration rate of HUVECs in the HT29 cell group was 82.863% \pm 3.095% and that of the 1627 cell group was 45.067% \pm 2.895% at 24 h after scratch. The average migration rate of HLECs in the HT29 cell group was 42.393% \pm 0.247%, and that of the 1627 cell group was 23.327% \pm 1.434%. Compared with the 1627 cell group, the migration of HUVECs and HLECs induced by exosomes in the HT29 cell group was substantially elevated (*P* < 0.01) (Figure 3B).

After 24 h of incubation, the average number of tube formations by HUVECs in the HT29 cell group and 1627 cell group was 30.625 ± 0.925 and 12.750 ± 1.887 , respectively, at low magnification. The average number of tube formations by HLECs in the HT29 cell group and 1627 cell group was 26.750 ± 2.016 and 15.375 ± 1.413 , respectively, at low magnification. Compared with the 1627 cell group, the tube formation ability of HUVECs and HLECs induced by exosomes in the HT29 cell group

Zaishidena® WJGO | https://www.wjgnet.com



Figure 1 Relationship between BRAF^{V600E} mutation and microvascular density, microlymphatic vessel density, tumor-associated macrophages, and cancer-associated fibroblasts. A: Expression of CD31 and lymphatic vessel endothelial hyaluronic acid receptor-1 (LYVE-1) in

 Jaisbideng®
 WJGO
 https://www.wjgnet.com

December 15, 2021 Volume 13 Issue 12

BRAF^{V600E} mutant and BRAF wild type colorectal cancer (CRC) tissues (CD31 labeled microvascular density, and LYVE-1 labeled microlymphatic vessel density; immunohistochemistry, × 400); B: Expression of CD68 and CD163 proteins in BRAF^{VE00E} mutant and BRAF wild type CRC tissues (CD68 labeled TAMs, and CD163 labeled M2 subtype macrophages: immunohistochemistry. × 400): C: Expression of α-SMA protein in BRAF^{V600E} mutant and BRAF wild type CRC tissues (α-SMA labeled CAFs; immunohistochemistry, × 400).

was markedly higher than that in the 1627 cell group (P < 0.01) (Figure 3C).

Western blot analysis indicated that compared with the HT29 cell group, the expression of VEGF-A, bFGF, TGF-β1, and VEGF-C proteins in the exosomes derived from 1627 cells was reduced (Figure 3D). However, the expression of ZO-1 in HUVECs and that of claudin-5, occludin, and ZO-1 in HLECs in the 1627 cell group were higher than those in the HT29 cell group (Figure 3E and F). Neither the 1627 nor the HT29 group failed to display occludin and claudin-5 protein. This suggests that exosomes derived from BRAFV600E mutant CRC cells promote angiogenesis and lymphoangiogenesis.

Exosomes derived from BRAF^{veone} mutant CRC cells promote polarization of macrophages to M2 subtype and enhance secretory function of macrophages and fibroblasts

The exosomes derived from HT29 and 1627 cells were cocultured with macrophages. Flow cytometry revealed that the exosomes of the HT29 cell group promoted the expression of CD163 in macrophages compared with the control group and 1627 cell group (P < 0.05) (Figure 4A). ELISA indicated that IL-6 secreted by macrophages in the HT29 cell group was markedly elevated (P < 0.05), whereas TGF- β 1 was decreased (P< 0.05) (Figure 4B). Western blot analysis demonstrated that there was no significant difference in the expression of fibroblast FAP and α -SMA in the control, HT29 cell, and 1627 cell groups (Figure 4C). Conversely, the levels of IL-6, TGF- β 1, and VEGF secreted by fibroblasts in the 1627 cell group decreased, compared with the HT29 cell group (P < 0.05) (Figure 4D). Exosomes derived from BRAF^{V600E} mutant CRC cells promoted polarization of macrophages to M2 subtype and enhanced the secretory function of fibroblasts.

DISCUSSION

This study demonstrated that BRAF^{V600E} mutant CRC generates a unique immune microenvironment. Compared with BRAF wild type CRC, there was more angiogenesis and lymphoangiogenesis in the microenvironment. Meanwhile, the polarization of macrophages to M2 subtype was more obvious, and the immunosuppression was more prominent. Exosomes derived from BRAF^{V600E} mutant CRC cells could induce this change. Further analysis indicated that exosomes derived from BRAF^{V600E} mutant CRC cells were rich in certain lncRNAs and mRNAs, which might link to these alterations. Our findings suggested that for this particular CRC, it might be worthwhile to try to investigate the TME.

It is currently argued that BRAF^{V600E} mutation is of vital significance in predicting the prognosis of CRC patients^[22]. Several studies have indicated that BRAF^{V600E} mutation is not only correlated to poor tissue differentiation, but also to gender, advanced stage, high T stage, right colon, lymph node metastasis, mucous tissue, and high levels of platelets and CEA[22,23]. Conversely, our results were inconsistent with those reported in the literature. We recognized that BRAF^{V600E} mutation was only associated with poor tissue differentiation and increased preoperative CA19-9 levels, which might have been caused by the small sample size.

However, we analyzed the TME of such patients. The results demonstrated that the MVD and MLVD in BRAF^{V600E} mutant CRC tissues were higher than those in BRAF wild type tissues, and the number of CD163⁺M2 macrophages increased substantially. In papillary thyroid carcinoma, BRAF^{V600E} mutation upregulates the expression of VEGF-A in cancer cells and promotes angiogenesis, and increases the expression of VEGF-C in cancer cells and promotes lymphangiogenesis, whereas silencing BRAF gene can reverse these effects[24]. Under normal circumstances, HCT116 colon cancer cells can activate the VEGF signaling pathway, promote the proliferation, migration, and angiogenesis of vascular endothelial cells, and facilitate tumor metastasis by releasing exosomes[25]. Exosomes derived from CRC cells can also promote TAMs to release VEGF-C, enhance proliferation of lymphatic endothelial cells, and induce







Figure 2 Screening and functional analysis of differentially expressed long noncoding RNAs and mRNAs in exosomes of BRAF^{VG00E} mutant colorectal cancer cells and those with BRAF gene silencing. A: Transcriptomics analysis of differentially expressed long noncoding RNAs (IncRNAs) and mRNAs in the exosomes of HT29 and 1627 cells. Volcano maps are shown (the x axis represents the multiple of difference, and the y axis P values); B: Hierarchical cluster analysis of differentially expressed IncRNAs and mRNAs (red represents upregulated expression, and blue represents downregulated expression); C: GO enrichment analysis of differentially expressed IncRNA target genes and mRNAs; D: KEGG pathway analysis of differentially expressed IncRNA target genes and mRNAs. LncRNA: Long noncoding RNA.

> lymphangiogenesis[26]. However, our results indicate that BRAF^{V600E} mutation promotes angiogenesis and lymphoangiogenesis in the microenvironment.

> Simultaneously, BRAF^{V600E} mutation affected cellular components in the microenvironment. We identified that there was no difference in the content of CD68⁺ macrophages in BRAF^{V600E} mutant and BRAF wild-type CRC tissues, and the number of M2 macrophages was markedly higher than that of BRAF wild-type tissues. This is in agreement with the results reported in the literature. Compared with BRAF wild type tumor, there is no difference in CD68⁺ macrophages in BRAF^{v600E} mutant thyroid cancer, whereas M2 subtype macrophages increase [27]. Furthermore, TAMs have been positively related to lymph node and peritoneal metastasis in ovarian cancer, gastric cancer, and other tumors [28-30]. CAFs play an essential role in tumor development and metastasis. Our findings indicated no difference in the content of CAFs in CRC tissues of BRAF^{V600E} mutant and BRAF wild-type CRC. We hypothesized that BRAF^{V600E} mutant CRC might promote lymph node and peritoneal metastasis through other mechanisms, but the specific mechanism needs further exploration. This harsh microenvironment may lead to a worse prognosis and even result in resistance to traditional treatments.

> Recent research has drawn a link between exosomes and intercellular communication, which are responsible for transporting specific RNA transcripts to target organs, participating in substance exchange in the distant environment, regulating immune function, and promoting tumor angiogenesis, invasion, and metastasis, thereby directly or indirectly affecting the progression and outcome of tumors[31]. We analyzed the expression profiles of exosomal lncRNAs and mRNAs in HT29 cells and 1627 cells using high-throughput sequencing, screening out differentially expressed IncRNAs and mRNAs. GO enrichment analysis and KEGG signaling pathway analysis were performed on differentially expressed lncRNA target genes and mRNAs. The findings indicated that differential lncRNA target genes were enriched in functions related to cell components, proliferation, metabolism, and migration, which were in agreement with functions of differential mRNA enrichment. The lncRNAs and mRNAs secreted by tumor cells were transported to target cells by exosomes. On the one hand, a microenvironment suitable for tumor cell metastasis (*i.e.*, premetastatic niche) could be created at a distance; on the other hand, it could also directly act on other tumor cells, altering their characteristics, and even change the metabolic programming of the tumor, thereby ultimately accelerating the invasion, metastasis,









Figure 3 Effect of BRAFV600E mutant colorectal cancer-cell-derived exosomes on the proliferation, migration, tube formation, and protein expression of human umbilical vein endothelial cells and human lymphatic endothelial cells. A: Detection of changes in proliferation of human umbilical vein endothelial cells (HUVECs) induced by exosomes derived from HT29 and 1627 cells at 24, 48, and 72 h by CCK8 assays. Changes in the proliferation of human lymphatic endothelial cells (HLECs) induced by exosomes derived from HT29 and 1627 cells were detected at 12, 24, and 36 h (the control group was treated with PBS); B: Detection of effects of HT29 and 1627 cell-derived exosomes on the migration of HUVECs and HLECs by scratch test (× 50); C: After 24 h of incubation, the effect of HT29 and 1627 cell-derived exosomes on tube formation ability of HUVECs and HLECs was observed (× 100); D: ELISA was used to detect expression of VEGF-A, TGF-β1, bFGF, and VEGF-C proteins in exosomes derived from HT29 and 1627 cells; E: ELISA was used to detect the effect of exosomes derived from HT29 and 1627 cells on expression of ZO-1 proteins in HUVECs; F: ELISA was used to detect the effect of exosomes derived from HT29 and 1627 cells on expression of ZO-1, occludin, and claudin-5 proteins in HLECs. ^aP < 0.05. HUVECs: Human umbilical vein endothelial cells; HLECs: Human lymphatic endothelial cells.

> and drug resistance of tumors. The expression of the exosome-derived lncRNA RPPH1 in CRC was markedly upregulated and it simultaneously interacted with TAMs to promote the polarization of TAMs to M2 subtype, thereby accelerating tumor progression[21]. Gao et al[32] have found that the lncRNA 91H in tumor-cell-derived exosomes can increase the expression of heterogeneous nuclear ribonucleoprotein K, and markedly enhance the migration and invasion of CRC cells. The above evidence suggests that BRAFV600E mutant CRC cells can reach the TME by releasing exosomal IncRNAs, and participate in tumor angiogenesis and tumor cell proliferation, differentiation, and metabolism through mRNAs, which are closely associated with the TME.

> To explore the effect of exosomes secreted by BRAF^{V600E} mutant cells on the surrounding immune microenvironment, we cocultured exosomes derived from two colon cancer cell lines (HT29 and 1627) with HUVECs or HLECs. The exosomes derived from BRAF^{V600E} mutant CRC cells promoted proliferation, migration, and tube formation of endothelial cells, and induced angiogenesis and lymphoangiogenesis. Silencing BRAF gene generated corresponding inhibitory effects. BRAF^{V600E} mutation can promote the expression of matrix metalloproteinase-2 and VEGF-A in malignant melanoma cells, mediate angiogenesis, and enhance the invasiveness of tumor cells. However, BRAF gene deletion leads to a lack of VEGF-A, inhibiting angiogenesis and minimizing the permeability between endothelial cells[33]. Additional research has also proposed that BRAF^{V600E} mutant thyroid cancer cells can promote angiogenesis and lymphoangiogenesis by releasing VEGF-A and VEGF-C into the TME. Zelboraf can reduce the contents of these factors in the TME and inhibit angiogenesis and lymphoangiogenesis, thereby minimizing distant metastasis.

> The expression of VEGF-A, bFGF, TGF- β , and VEGF-C proteins in exosomes derived from BRAFV600E mutant CRC cells was also increased. Tight junction proteins ZO-1, claudin-5, and occludin participate in the formation of the endothelial barrier, affect cell permeability, and play an vital role in regulating the proliferation, migration, and tube formation of endothelial cells[34-36]. This study found that exosomes derived from BRAFV600E mutant CRC cells could inhibit the expression of ZO-1, claudin-5, and occludin in HLECs and ZO-1 in HUVECs. Due to the low concentrations of claudin-5 and occludin in HUVECs, this study failed to detect the effect of silencing BRAF gene on the expression of claudin-5 and occludin proteins in HUVECs. Despite that inhibition of ZO-1, claudin-5, and occludin can restrain the growth of endothelial cells, it may not be enough to resist BRAF^{V600E} mutation for the promotion of vascularization (namely, increased expression of VEGF-A, bFGF, TGF-β, and VEGF-C proteins). Our study suggests that exosomes derived from BRAF^{V600E} mutant CRC cells promote the expression of VEGF-A, bFGF, TGF-B, and VEGF-C to facilitate





Figure 4 Effect of exosomes derived from BRAF^{veoue} mutant colorectal cancer cells on the phenotype and secretion function of tumorassociated macrophages and cancer-associated fibroblasts. A: Effect of silencing BRAF gene on expression of CD163 in macrophages induced by exosomes derived from BRAF^{VG00E} mutant colorectal cancer cells; B: ELISA was used to detect expression of interleukin (IL)-6 and transforming growth factor (TGF)β1 in the supernatant of THP-1 cells; C: Western blotting was used to detect expression of FAP and α-SMA in MRC-5 cells; D: ELISA was used to detect expression of IL-6, TGF-B1, and vascular endothelial growth factor in the supernatant of MRC-5 cells. ^aP < 0.05. VEGF: Vascular endothelial growth factor; IL: Interleukin; TGF: Transforming growth factor.

angiogenesis and lymphangiogenesis.

We cocultured exosomes derived from HT29 and 1627 cells with macrophages and found that exosomes derived from BRAF^{V600E} mutant CRC cells enhanced the polarization of macrophages to M2 subtype. M1 subtype macrophages secrete IL-6, and M2 subtype macrophages secrete TGF- β 1[37]. IL-6 is highly expressed in diverse malignant TMEs, and it can promote tumor invasion, distant metastasis, and angiogenesis, and participate in tumor resistance[38]. Exosomes derived from BRAF^{V600E} mutant colon cancer cells can promote the secretion of IL-6 by macrophages and after silencing the BRAF gene, exosomes can inhibit the secretion of IL-6 by macrophages. Some researchers have discovered that exosomes secreted by hepatoma promote the secretion of IL-6 by macrophages, whereas exosomes secreted by melatonin-treated hepatocellular carcinoma can inhibit the secretion of IL-6 by macrophages[39]. TGF- β 1 inhibits tumor proliferation and induces apoptosis in the early stage of tumor development. Conversely, when in the advanced stage, it promotes the development and metastasis of the tumor by promoting epithelial-mesenchymal transition, regulating the microenvironment and the immune system[40,41]. Some studies have found that BRAFV600E mutant CRC-derived exosomes can inhibit macrophages from secreting more TGF-\u00b31. Therefore, we speculated that exosomes derived from BRAF^{V600E} mutant CRC cells could promote the polarization of macrophages to M2 subtype, increase the secretion of IL-6, and reduce the secretion of TGF- β 1, thereby facilitating distant metastasis.

The cell component with the highest content in the microenvironment is CAFs, which highly express α-SMA and FAP, and secrete IL-6, TGF-β1, and VEGF. Meanwhile, they assist tumor cells in immune escape, and promote angiogenesis,



tumor invasion, and metastasis[42,43]. The present study indicated that after exosomes derived from BRAFV600E mutant CRC cells were cocultured with fibroblasts, the expression of α-SMA and FAP in fibroblasts did not change markedly. Instead, they promoted the secretion of IL-6 and TGF- *β*1 and VEGF by CAFs. It was suggested that BRAF^{V600E} mutation had little effect on the number of CAFs, mainly affecting their function. Some researchers have found that exosomes derived from hepatocellular carcinoma can promote the differentiation of hepatic astrocytes into CAFs. The activated CAFs secrete cytokines VEGF, TGF- β , and IL-6, and promote angiogenesis and liver metastasis[44,45].

We selected only one BRAFVGODE mutant CRC cell line, and only performed in vitro experiments. The differentially expressed lncRNAs were not verified. Nevertheless, our research demonstrated that BRAFV600E mutant CRC had a unique immune microenvironment, which might be induced by the release of exosomes rich in certain IncRNAs. Therefore, in the future, we can consider to reshape the immune microenvironment, combined with traditional treatment, to treat this specific type of CRC.

CONCLUSION

Our study showed that, compared with wild type BRAF, BRAF^{V600E} mutation led to more angiogenesis and lymphoangiogenesis in the microenvironment. Meanwhile, the polarization of macrophages to M2 subtype was more obvious, and the immunosuppression was more prominent. Further analysis indicated that exosomes derived from BRAF^{V600E} mutant CRC cells were rich in certain lncRNAs and mRNAs, which might be linked to these alterations. This provides a hypothesis for finding new therapeutic strategies for BRAF^{V600E} mutant CRC.

ARTICLE HIGHLIGHTS

Research background

BRAF^{V600E} gene mutation accounts for approximately 10% of patients with metastatic colorectal cancer (CRC). Compared with CRC patients with wild-type BRAF, patients with BRAF^{V600E} mutant CRC are prone to peritoneal metastasis and distant lymph node metastasis with a poor prognosis. Previous findings suggest that BRAF^{V600E} mutation affects the tumor microenvironment (TME).

Research motivation

BRAF^{V600E} mutation is involved in the formation of the immunosuppressive microenvironment in thyroid cancer. However, the influence and the related mechanism of BRAF^{V600E} mutation in CRC on the surrounding immune microenvironment are not clear.

Research objectives

The study aimed to determine the influence of $BRAF^{V600E}$ mutation in CRC on the surrounding immune microenvironment, elucidating whether BRAF^{V600E} mutant CRC cell-derived exosomes participate in the formation of an immunosuppressive microenvironment

Research methods

CRC patients were divided into either a control group or a treatment group. The formation of microvessels and microlymphatic vessels and M2 subtype macrophages in tumor tissues were detected by immunohistochemistry. Screening and functional analysis of exosomal long noncoding RNAs (lncRNAs) were performed by transcriptomics. The proliferation and migration of human umbilical vein endothelial cells (HUVECs) and human lymphatic endothelial cells (HLECs) were detected by CCK-8 assays and scratch test, respectively. The tube-forming ability of endothelial cells was assessed by tube formation assay. The macrophage subtypes were obtained by flow cytometry. The expression of vascular endothelial growth factor (VEGF)-A, basic fibroblast growth factor (bFGF), transforming growth factor (TGF)-β1, VEGF-C, claudin-5, occludin, zonula occludens (ZO)-1, fibroblast activation protein (FAP), and α-smooth muscle actin was assessed by Western blot analysis. The levels of cytokines interleukin (IL)-6, TGF-β1, and VEGF were assessed by ELISA.



Research results

BRAF^{V600E} mutation was positively correlated with a poor prognosis in CRC (P < 0.01). Microvascular density and microlymphatic vessel density in BRAF^{V600E} mutant CRC tissues were higher than those in BRAF wild-type CRC (P < 0.05). The number of CD163⁺ M2 macrophages in BRAF^{V600E} mutant CRC tumor tissue was markedly increased (P < 0.05). Compared with exosomes from CRC cells with BRAF gene silencing, the expression of 13 lncRNAs and 192 mRNAs in the BRAFV600E mutant CRC cell exosomes was upregulated, and the expression of 22 lncRNAs and 236 mRNAs was downregulated (P < 0.05). The biological functions and signaling pathways predicted by differential lncRNA target genes and differential mRNA were closely related to angiogenesis, tumor cell proliferation, differentiation, metabolism, and changes in the microenvironment. The proliferation, migration, and tube formation ability of HUVECs and HLECs induced by exosomes in the 1627 cell group (HT29 cells with BRAF gene silencing) was greatly reduced compared with the HT29 cell group (P < 0.05). Compared with the HT29 cell group, the expression levels of VEGF-A, bFGF, TGF- β 1, and VEGF-C in the exosomes derived from 1627 cells were reduced. The expression of ZO-1 in HUVECs, and claudin-5, occludin, and ZO-1 in HLECs of the 1627 cell group was higher. Compared with the 1627 cell group, the exosomes of the HT29 group promoted the expression of CD163 in macrophages (P < 0.05). IL-6 secretion by macrophages in the HT29 cell group was markedly elevated (P < 0.05), whereas TGF- β 1 was decreased (P < 0.05). The levels of IL-6, TGF- β 1, and VEGF secreted by fibroblasts in the 1627 cell group decreased, compared with the HT29 group (*P* < 0.05).

Research conclusions

BRAF^{V600E} mutant CRC cells can reach the TME by releasing exosomal lncRNAs, inducing the formation of an immunosuppressive microenvironment.

Research perspectives

The study will provide a novel therapeutic strategy for BRAF^{V600E} mutant CRC.

REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. CA Cancer J Clin 2019; 69: 7-34 [PMID: 1 30620402 DOI: 10.3322/caac.21551]
- Zhou J, Zheng R, Zhang S, Zeng H, Wang S, Chen R, Sun K, Li M, Gu J, Zhuang G, Wei W. Colorectal cancer burden and trends: Comparison between China and major burden countries in the world. Chin J Cancer Res 2021; 33: 1-10 [PMID: 33707923 DOI: 10.21147/j.issn.1000-9604.2021.01.01]
- 3 Fiskus W, Mitsiades N. B-Raf Inhibition in the Clinic: Present and Future. Annu Rev Med 2016; 67: 29-43 [PMID: 26768236 DOI: 10.1146/annurev-med-090514-030732]
- 4 Ursem C, Atreya CE, Van Loon K. Emerging treatment options for BRAF-mutant colorectal cancer. Gastrointest Cancer 2018; 8: 13-23 [PMID: 29628780 DOI: 10.2147/GICTT.S125940]
- 5 Taieb J, Lapeyre-Prost A, Laurent Puig P, Zaanan A. Exploring the best treatment options for BRAFmutant metastatic colon cancer. Br J Cancer 2019; 121: 434-442 [PMID: 31353365 DOI: 10.1038/s41416-019-0526-2
- Davies H, Bignell GR, Cox C, Stephens P, Edkins S, Clegg S, Teague J, Woffendin H, Garnett MJ, 6 Bottomley W, Davis N, Dicks E, Ewing R, Floyd Y, Gray K, Hall S, Hawes R, Hughes J, Kosmidou V, Menzies A, Mould C, Parker A, Stevens C, Watt S, Hooper S, Wilson R, Jayatilake H, Gusterson BA, Cooper C, Shipley J, Hargrave D, Pritchard-Jones K, Maitland N, Chenevix-Trench G, Riggins GJ, Bigner DD, Palmieri G, Cossu A, Flanagan A, Nicholson A, Ho JW, Leung SY, Yuen ST, Weber BL, Seigler HF, Darrow TL, Paterson H, Marais R, Marshall CJ, Wooster R, Stratton MR, Futreal PA. Mutations of the BRAF gene in human cancer. Nature 2002; 417: 949-954 [PMID: 12068308 DOI: 10.1038/nature00766]
- Fedorenko IV, Paraiso KH, Smalley KS. Acquired and intrinsic BRAF inhibitor resistance in BRAF 7 V600E mutant melanoma. Biochem Pharmacol 2011; 82: 201-209 [PMID: 21635872 DOI: 10.1016/j.bcp.2011.05.015
- 8 Bhatt KV, Spofford LS, Aram G, McMullen M, Pumiglia K, Aplin AE. Adhesion control of cyclin D1 and p27Kip1 Levels is deregulated in melanoma cells through BRAF-MEK-ERK signaling. Oncogene 2005; 24: 3459-3471 [PMID: 15735667 DOI: 10.1038/sj.onc.1208544]
- Jin Z, Sinicrope FA. Advances in the therapy of BRAF^{V600E} metastatic colorectal cancer. Expert Rev 9 Anticancer Ther 2019; 19: 823-829 [PMID: 31455117 DOI: 10.1080/14737140.2019.1661778]
- Biller LH, Schrag D. Diagnosis and Treatment of Metastatic Colorectal Cancer: A Review. JAMA 10 2021; 325: 669-685 [PMID: 33591350 DOI: 10.1001/jama.2021.0106]
- Wong V, Lee M, Wong R, Tie J, Shapiro J, Desai J, Nott L, Steel S, Burge M, Ma B, Khattak A, 11



Hong W, Gibbs P. BRAFV600E Mutations Arising from a Left-Side Primary in Metastatic Colorectal Cancer: Are They a Distinct Subset? Target Oncol 2021; 16: 227-236 [PMID: 33599905 DOI: 10.1007/s11523-021-00793-7

- 12 Galon J, Bruni D. Tumor Immunology and Tumor Evolution: Intertwined Histories. Immunity 2020; 52: 55-81 [PMID: 31940273 DOI: 10.1016/j.immuni.2019.12.018]
- Means C, Clayburgh DR, Maloney L, Sauer D, Taylor MH, Shindo ML, Coussens LM, Tsujikawa T. 13 Tumor immune microenvironment characteristics of papillary thyroid carcinoma are associated with histopathological aggressiveness and BRAF mutation status. Head Neck 2019; 41: 2636-2646 [PMID: 30896061 DOI: 10.1002/hed.25740]
- 14 Denton AE, Roberts EW, Fearon DT. Stromal Cells in the Tumor Microenvironment. Adv Exp Med Biol 2018; 1060: 99-114 [PMID: 30155624 DOI: 10.1007/978-3-319-78127-3_6]
- 15 Erreni M, Mantovani A, Allavena P. Tumor-associated Macrophages (TAM) and Inflammation in Colorectal Cancer. Cancer Microenviron 2011; 4: 141-154 [PMID: 21909876 DOI: 10.1007/s12307-010-0052-5
- Herrera M, Herrera A, Domínguez G, Silva J, García V, García JM, Gómez I, Soldevilla B, Muñoz 16 C, Provencio M, Campos-Martin Y, García de Herreros A, Casal I, Bonilla F, Peña C. Cancerassociated fibroblast and M2 macrophage markers together predict outcome in colorectal cancer patients. Cancer Sci 2013; 104: 437-444 [PMID: 23298232 DOI: 10.1111/cas.12096]
- Ping Q, Yan R, Cheng X, Wang W, Zhong Y, Hou Z, Shi Y, Wang C, Li R. Cancer-associated 17 fibroblasts: overview, progress, challenges, and directions. Cancer Gene Ther 2021; 28: 984-999 [PMID: 33712707 DOI: 10.1038/s41417-021-00318-4]
- 18 Bracci L, Lozupone F, Parolini I. The role of exosomes in colorectal cancer disease progression and response to therapy. Cytokine Growth Factor Rev 2020; 51: 84-91 [PMID: 31955973 DOI: 10.1016/j.cytogfr.2019.12.004]
- 19 Galamb O, Barták BK, Kalmár A, Nagy ZB, Szigeti KA, Tulassay Z, Igaz P, Molnár B. Diagnostic and prognostic potential of tissue and circulating long non-coding RNAs in colorectal tumors. World J Gastroenterol 2019; 25: 5026-5048 [PMID: 31558855 DOI: 10.3748/wjg.v25.i34.5026]
- 20 Sun Z, Liu J, Chen C, Zhou Q, Yang S, Wang G, Song J, Li Z, Zhang Z, Xu J, Sun X, Chang Y, Yuan W. The Biological Effect and Clinical Application of Long Noncoding RNAs in Colorectal Cancer. Cell Physiol Biochem 2018; 46: 431-441 [PMID: 29614491 DOI: 10.1159/000488610]
- 21 Liang ZX, Liu HS, Wang FW, Xiong L, Zhou C, Hu T, He XW, Wu XJ, Xie D, Wu XR, Lan P. LncRNA RPPH1 promotes colorectal cancer metastasis by interacting with TUBB3 and by promoting exosomes-mediated macrophage M2 polarization. Cell Death Dis 2019; 10: 829 [PMID: 31685807 DOI: 10.1038/s41419-019-2077-01
- Wang J, Shen J, Huang C, Cao M, Shen L. Clinicopathological Significance of BRAFV600E 22 Mutation in Colorectal Cancer: An Updated Meta-Analysis. J Cancer 2019; 10: 2332-2341 [PMID: 31258736 DOI: 10.7150/jca.30789]
- 23 Caputo F, Santini C, Bardasi C, Cerma K, Casadei-Gardini A, Spallanzani A, Andrikou K, Cascinu S, Gelsomino F. BRAF-Mutated Colorectal Cancer: Clinical and Molecular Insights. Int J Mol Sci 2019; 20 [PMID: 31661924 DOI: 10.3390/ijms20215369]
- Husain A, Hu N, Sadow PM, Nucera C. Expression of angiogenic switch, cachexia and inflammation 24 factors at the crossroad in undifferentiated thyroid carcinoma with BRAF(V600E). Cancer Lett 2016; 380: 577-585 [PMID: 26189429 DOI: 10.1016/j.canlet.2015.07.012]
- Hu HY, Yu CH, Zhang HH, Zhang SZ, Yu WY, Yang Y, Chen Q. Exosomal miR-1229 derived from 25 colorectal cancer cells promotes angiogenesis by targeting HIPK2. Int J Biol Macromol 2019; 132: 470-477 [PMID: 30936013 DOI: 10.1016/j.ijbiomac.2019.03.221]
- Sun B, Zhou Y, Fang Y, Li Z, Gu X, Xiang J. Colorectal cancer exosomes induce lymphatic network 26 remodeling in lymph nodes. Int J Cancer 2019; 145: 1648-1659 [PMID: 30734278 DOI: 10.1002/ijc.32196]
- Angell TE, Lechner MG, Jang JK, Correa AJ, LoPresti JS, Epstein AL. BRAF V600E in papillary 27 thyroid carcinoma is associated with increased programmed death ligand 1 expression and suppressive immune cell infiltration. Thyroid 2014; 24: 1385-1393 [PMID: 24955518 DOI: 10.1089/thy.2014.0134]
- Wang R, Zhang T, Ma Z, Wang Y, Cheng Z, Xu H, Li W, Wang X. The interaction of coagulation 28 factor XII and monocyte/macrophages mediating peritoneal metastasis of epithelial ovarian cancer. *Gynecol Oncol* 2010; **117**: 460-466 [PMID: 20233624 DOI: 10.1016/j.ygyno.2010.02.015]
- Yamagata Y, Tomioka H, Sakamoto K, Sato K, Harada H, Ikeda T, Kayamori K. CD163-Positive Macrophages Within the Tumor Stroma Are Associated With Lymphangiogenesis and Lymph Node Metastasis in Oral Squamous Cell Carcinoma. J Oral Maxillofac Surg 2017; 75: 2144-2153 [PMID: 28399391 DOI: 10.1016/j.joms.2017.03.009]
- 30 Go Y, Tanaka H, Tokumoto M, Sakurai K, Toyokawa T, Kubo N, Muguruma K, Maeda K, Ohira M, Hirakawa K. Tumor-Associated Macrophages Extend Along Lymphatic Flow in the Pre-metastatic Lymph Nodes of Human Gastric Cancer. Ann Surg Oncol 2016; 23 Suppl 2: S230-S235 [PMID: 25743331 DOI: 10.1245/s10434-015-4458-7]
- 31 Kok VC, Yu CC. Cancer-Derived Exosomes: Their Role in Cancer Biology and Biomarker Development. Int J Nanomedicine 2020; 15: 8019-8036 [PMID: 33116515 DOI: 10.2147/IJN.S272378]
- Gao T, Liu X, He B, Nie Z, Zhu C, Zhang P, Wang S. Exosomal IncRNA 91H is associated with poor 32 development in colorectal cancer by modifying HNRNPK expression. Cancer Cell Int 2018; 18: 11



[PMID: 29410604 DOI: 10.1186/s12935-018-0506-2]

- Declercq M, Treps L. BRAF, A gatekeeper controlling endothelial permeability. FEBS J 2019; 286: 33 2273-2276 [PMID: 31081213 DOI: 10.1111/febs.14861]
- 34 Tornavaca O, Chia M, Dufton N, Almagro LO, Conway DE, Randi AM, Schwartz MA, Matter K, Balda MS. ZO-1 controls endothelial adherens junctions, cell-cell tension, angiogenesis, and barrier formation. J Cell Biol 2015; 208: 821-838 [PMID: 25753039 DOI: 10.1083/jcb.201404140]
- 35 Chidiac R, Zhang Y, Tessier S, Faubert D, Delisle C, Gratton JP. Comparative Phosphoproteomics Analysis of VEGF and Angiopoietin-1 Signaling Reveals ZO-1 as a Critical Regulator of Endothelial Cell Proliferation. Mol Cell Proteomics 2016; 15: 1511-1525 [PMID: 26846344 DOI: 10.1074/mcp.M115.053298]
- 36 Zhang H, Zhang S, Zhang J, Liu D, Wei J, Fang W, Zhao W, Chen Y, Shang D. ZO-1 expression is suppressed by GM-CSF via miR-96/ERG in brain microvascular endothelial cells. J Cereb Blood Flow Metab 2018; 38: 809-822 [PMID: 28430012 DOI: 10.1177/0271678X17702668]
- Xu YW, Xing RX, Zhang WH, Li L, Wu Y, Hu J, Wang C, Luo QL, Shen JL, Chen X. Toxoplasma 37 ROP16_{1/III} ameliorated inflammatory bowel diseases via inducing M2 phenotype of macrophages. World J Gastroenterol 2019; 25: 6634-6652 [PMID: 31832003 DOI: 10.3748/wjg.v25.i45.6634]
- Choy EH, De Benedetti F, Takeuchi T, Hashizume M, John MR, Kishimoto T. Translating IL-6 38 biology into effective treatments. Nat Rev Rheumatol 2020; 16: 335-345 [PMID: 32327746 DOI: 10.1038/s41584-020-0419-z
- Cheng L, Liu J, Liu Q, Liu Y, Fan L, Wang F, Yu H, Li Y, Bu L, Li X, Wei W, Wang H, Sun G. 39 Exosomes from Melatonin Treated Hepatocellularcarcinoma Cells Alter the Immunosupression Status through STAT3 Pathway in Macrophages. Int J Biol Sci 2017; 13: 723-734 [PMID: 28655998 DOI: 10.7150/ijbs.19642]
- Liu S, Ren J, Ten Dijke P. Targeting TGF β signal transduction for cancer therapy. Signal Transduct 40 *Target Ther* 2021; **6**: 8 [PMID: 33414388 DOI: 10.1038/s41392-020-00436-9]
- Derynck R, Turley SJ, Akhurst RJ. TGF^β biology in cancer progression and immunotherapy. Nat Rev 41 Clin Oncol 2021; 18: 9-34 [PMID: 32710082 DOI: 10.1038/s41571-020-0403-1]
- Piersma B, Hayward MK, Weaver VM. Fibrosis and cancer: A strained relationship. Biochim 42 Biophys Acta Rev Cancer 2020; 1873: 188356 [PMID: 32147542 DOI: 10.1016/j.bbcan.2020.188356]
- Lim H, Koh M, Jin H, Bae M, Lee SY, Kim KM, Jung J, Kim HJ, Park SY, Kim HS, Moon WK, 43 Hwang S, Cho NH, Moon A. Cancer-associated fibroblasts induce an aggressive phenotypic shift in non-malignant breast epithelial cells via interleukin-8 and S100A8. J Cell Physiol 2021; 236: 7014-7032 [PMID: 33748944 DOI: 10.1002/jcp.30364]
- Zhou Y, Ren H, Dai B, Li J, Shang L, Huang J, Shi X. Hepatocellular carcinoma-derived exosomal 44 miRNA-21 contributes to tumor progression by converting hepatocyte stellate cells to cancerassociated fibroblasts. J Exp Clin Cancer Res 2018; 37: 324 [PMID: 30591064 DOI: 10.1186/s13046-018-0965-21
- 45 Fang T, Lv H, Lv G, Li T, Wang C, Han Q, Yu L, Su B, Guo L, Huang S, Cao D, Tang L, Tang S, Wu M, Yang W, Wang H. Tumor-derived exosomal miR-1247-3p induces cancer-associated fibroblast activation to foster lung metastasis of liver cancer. Nat Commun 2018; 9: 191 [PMID: 29335551 DOI: 10.1038/s41467-017-02583-0]



0 WŰ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2149-2160

DOI: 10.4251/wjgo.v13.i12.2149

ISSN 1948-5204 (online)

ORIGINAL ARTICLE

Retrospective Cohort Study

Hepatocellular carcinoma surveillance and quantile regression for determinants of underutilisation in at-risk Australian patients

Elizabeth SL Low, Ross Apostolov, Darren Wong, Sandra Lin, Numan Kutaiba, Josephine A Grace, Marie Sinclair

ORCID number: Elizabeth SL Low 0000-0002-0604-3920: Ross Apostolov 0000-0002-4827-8795; Darren Wong 0000-0003-1490-0547; Sandra Lin 0000-0001-8959-1828; Numan Kutaiba 0000-0003-4627-9847; Josephine A Grace 0000-0002-8435-4740; Marie Sinclair 0000-0003-0657-3048.

Author contributions: Low ES,

Apostolov R, Lin S and Kutaiba N directly designed and performed the study, with contribution from Sinclair M, Wong D and Grace J; Low ESL, Lin S and Wong D collated and analysed the data; Low ESL wrote the paper, with revisions and editing by all other listed authors.

Institutional review board

statement: This study was approved by the Austin Health Research Ethics Committee (ID 19/114) and carried out in line with the National Statement on Ethical Conduct in Human Research (2007).

Informed consent statement: All

study participants or their legal guardian provided informed written consent about personal and medical data collection prior to study enrolment.

Conflict-of-interest statement: The

Elizabeth SL Low, Ross Apostolov, Darren Wong, Josephine A Grace, Marie Sinclair, Department of Gastroenterology and Liver Transplant Unit, Austin Health, Heidelberg 3084, Victoria, Australia

Ross Apostolov, Josephine A Grace, Marie Sinclair, Department of Medicine, University of Melbourne, Melbourne 3000, Victoria, Australia

Sandra Lin, Department of Radiology, Monash Health, Clayton 3168, Victoria, Australia

Numan Kutaiba, Department of Radiology, Austin Health, Heidelberg 3084, Victoria, Australia

Corresponding author: Elizabeth SL Low, MBBS, Doctor, Department of Gastroenterology and Liver Transplant Unit, Austin Health, 145 Studley Road, Heidelberg 3084, Victoria, Australia. elizabeth low312@hotmail.com

Abstract

BACKGROUND

While clinical guidelines recommend hepatocellular carcinoma (HCC) surveillance for at-risk individuals, reported surveillance rates in the United States and Europe remain disappointingly low.

AIM

To quantify HCC surveillance in an Australian cohort, and assess for factors associated with surveillance underutilisation.

METHODS

All patients undergoing HCC surveillance liver ultrasounds between January 1, 2018 to June 30, 2018 at a tertiary hospital in Melbourne, Australia, were followed until July 31, 2020, or when surveillance was no longer required. The primary outcome was the percentage of time up-to-date with HCC surveillance (PTUDS). Quantile regression was performed to determine the impact of factors associated with HCC surveillance underutilisation.

RESULTS

Among 775 at-risk patients followed up for a median of 27.5 months, the median PTUDS was 84.2% (IQR: 66.3%-96.3%). 85.0% of patients were followed up by specialist gastroenterologists. Amongst those receiving specialist care, quantile



authors declare no conflicts of interest or financial supports.

Data sharing statement: Patient consent was not obtained but the presented data are anonymised and risk of identification is low. No additional data are available.

Country/Territory of origin: Australia

Specialty type: Gastroenterology and hepatology

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): A Grade B (Very good): B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: April 16, 2021 Peer-review started: April 16, 2021 First decision: June 27, 2021 Revised: July 13, 2021 Accepted: September 19, 2021 Article in press: September 19, 2021 Published online: December 15, 2021

P-Reviewer: Barabino M, Parente A, Tolunay HE S-Editor: Wang LL L-Editor: A P-Editor: Guo X



regression demonstrated differential associations at various quantile levels of PTUDS for several factors. Older age at the 25th quantile (estimate 0.002 per percent, P = 0.03), and cirrhotic status at the 75th quantile (estimate 0.021, P =0.017), were significantly associated with greater percentage of time up-to-date. African ethnicity (estimate -0.089, P = 0.048) and a culturally and linguistically diverse (CALD) background (estimate -0.063, P = 0.01) were significantly associated with lower PTUDS at the 50th quantile, and again for CALD at the 75th quantile (estimate -0.026, *P* = 0.045).

CONCLUSION

While median PTUDS in this Australian cohort study was 84.2%, awareness of the impact of specific factors across PTUDS quantiles can aid targeted interventions towards improved HCC surveillance.

Key Words: Liver cirrhosis; Hepatitis, viral, human; Carcinoma, hepatocellular; Liver neoplasms; Early detection of cancer; Population surveillance

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: This study evaluated the uptake of liver cancer screening in a cohort of highrisk Australians, and found that on average, patients were up-to-date with their surveillance for 84.2% of the study time period. Certain factors, such as absence of cirrhosis, younger age, African ethnicity and a non-English speaking background were associated to varying degrees with lower time up-to-date with hepatocellular carcinoma screening.

Citation: Low ES, Apostolov R, Wong D, Lin S, Kutaiba N, Grace JA, Sinclair M. Hepatocellular carcinoma surveillance and quantile regression for determinants of underutilisation in at-risk Australian patients. World J Gastrointest Oncol 2021; 13(12): 2149-2160

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2149.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2149

INTRODUCTION

Globally, hepatocellular carcinoma (HCC) is the sixth most commonly diagnosed cancer and is the fourth leading cause of cancer-related mortality[1-3]. Over the last three decades in Australia, there has been a 306% increase in age-standardised incidence of liver cancer, and a corresponding 184% increase in attributable mortality rates[4]. These concerning escalations are mirrored in recent reports from the United States, and the age-adjusted worldwide incidence of HCC stands at 10.1 cases /100,000 person-years[5,6].

While curative treatment options offer 5-year survival rates in excess of 70%, these are only feasible at early disease stages, often when HCC is clinically silent[1,7].A meta-analysis of cohort studies demonstrated improved early-stage detection and associated survival with HCC screening in at-risk populations[8]. This is particularly accepted in the non-cirrhotic population, for whom surgical resection is often a viable and life-saving intervention for early HCC[9]. As such, international professional societies recommend that at-risk patients undergo HCC surveillance with liver ultrasound (USS) every 6 months, with or without concurrent alpha fetoprotein (AFP) measurements[10,11].

The effectiveness of HCC surveillance relies on consistent clinical uptake of screening, particularly as sensitivity rates of ultrasonography for the detection of HCC are reported to be as low as 60%, thus requiring serial scans to ensure maximum yield [6]. However, several cohort studies have supported longstanding concerns of HCC surveillance underutilisation in clinical practice, with two recent meta-analyses reporting overall pooled adherence rates of 24% and 52% [3,12]. Research into adherence to surveillance is limited by the marked heterogeneity in the definitions of adherence in the literature[3]; studies vary from requiring perfect 6-monthly



adherence, to allowing leeway periods of up to 12 mo between imaging, raising the possibility of lost interim early-stage HCC detection[3].

To better incorporate both frequency of screening and quantity of imaging performed, the concept of 'percentage of time up-to-date with surveillance' (PTUDS) has been proposed as a more robust, continuous metric to standardise the measurement of adherence to HCC surveillance[3,7]. This retrospective cohort study aimed to quantify the percentage of time up-to-date with HCC screening in an Australian cohort of 'at-risk' patients, and to identify determinants associated with higher adherence to surveillance.

MATERIALS AND METHODS

Study design

This retrospective cohort study at a large tertiary hospital in Melbourne, Australia captured all at-risk patients identified by radiology records as undergoing a liver ultrasound scan (USS) for HCC surveillance within the Austin Hospital radiology department between 1 January 2018 to 30 June 2018. All USS were requested by gastroenterologists, infectious disease physicians and nurse practitioners who manage patients deemed at-risk of HCC in outpatient specialty clinics. Most at-risk patients at our centre were managed in physician-led specialty clinics. A proportion of consenting patients, who were felt to be at low risk of progressive liver disease, were primarily managed in a nurse-led, HCC screening clinic. Eligible patients for the nurse-led clinic were patients not requiring treatment for chronic hepatitis B, and patients with wellcompensated cirrhosis from a disease aetiology that was considered adequately treated. All patients referred to the nurse-led clinic were previously seen by physicians and physician oversight was available if needed.

At-risk patients for HCC development were defined according to AASLD guidelines, and included those with cirrhosis irrespective of aetiology, and patients with chronic hepatitis B and additional HCC risk factors (i.e. family history of HCC, African descent, Asian males over 40 years old, Asian females over 50 years old)[10]. Our institution also performs surveillance for patients with chronic hepatitis C and advanced fibrosis (F3 and above), as HCC incidence in this demographic may surpass the threshold for cost-effective screening[6]. Patients who did not meet this criteria or had a documented history of HCC within the last 2 years were excluded.

Patients were followed-up from the index USS until July 31, 2020, or when surveillance was no longer required. Reasons for this included HCC diagnosis (after which patients were diverted into our liver cancer clinic), death, receipt of an orthotopic liver transplant, or when surveillance was no longer deemed appropriate, such as cases of elderly multi-comorbid patients with limited life expectancy. Followup periods were also truncated for patients who were discharged to other medical services. In these cases, follow-up end-date was documented as date of the formal discharge letter written by attending clinicians to the patient and/or their primary care physician. This acknowledged the possibility of subsequent external imaging and follow-up.

This study was approved by the Austin Health Research Ethics Committee and carried out in line with the National Statement on Ethical Conduct in Human Research (2007).

Data collection

Patient demographics, clinical history, laboratory investigations and imaging results were extracted from their electronic medical records at the time of inclusion. Diagnoses of chronic hepatitis B were verified by record review of positive hepatitis B surface antigen or HBV DNA tests at least 6 mo apart. Confirmation of chronic hepatitis C occurred if there was evidence of positive hepatitis C antibody or viral RNA. All cirrhosis diagnoses were verified by review of clinical records by a consultant gastroenterologist, where cirrhosis was confirmed using a combination of clinical, biochemical, radiological and histological findings. Scanned medical records were also reviewed to retrieve externally performed liver imaging.

Outcomes of interest

The primary outcome was adherence to HCC surveillance imaging. Accepted imaging modalities included targeted liver USS, or contrast-enhanced CT or MRI, given their routine use in clinical practice when USS images are inadequate. Adherence was defined as the percentage of time up-to-date with screening (PTUDS, %), calculated as



the percentage of an individual's total follow-up time in which they were within 6 mo of an accepted HCC surveillance test. This endpoint was chosen as it accounted for both number and timing of surveillance imaging performed during the screening period. Following surveillance imaging, patients were credited as having 6 mo of 'time up-to-date'. Any subsequent imaging that occurred within that 6-month period resulted in a re-set of the 6-month interval clock from the date that the new imaging was performed (Figure 1). AFP levels were recorded, but were not included in the primary outcome, given AFP testing is not considered a stand-alone surveillance test.

Patient demographics

Demographic variables, based on literature-reported factors, were also extracted from the database and assessed for potential associations with uptake of HCC surveillance. These variables of interest included age, gender, ethnicity, primary language spoken, aetiology of liver disease, cirrhosis status and MELD score.

Statistical analysis

Descriptive statistical analysis was reported as proportions (%) for categorical variables, and means (with standard deviations) or medians (with IQRs) for continuous variables, depending on the distribution of values. Comparative analysis was conducted using the Student t-test and Wilcoxon rank-sum tests. Univariable and multivariable quantile regression analysis was then performed to assess for factors associated with PTUDS, particularly variables with significant associations at different PTUDS quantiles. Quantile regression modelling was chosen given the non-parametric distribution of the outcome variable relative to covariates, its flexibility in assessing the relationship between determinants of surveillance and PTUDS at the upper and lower tails, and the lack of stringent model assumptions required for valid inference. The median (50th), 25th, and 75th quantiles were selected to provide a description of covariate associations across the range of the distribution of PTUDS, whilst avoiding the effect of extreme outliers. Bootstrapping was used to calculate 95% confidence intervals around the estimates. Both univariate and multivariate models were constructed separately for each of the selected PTUDS quantiles to determine if there were differential covariates associations across the distribution.

Data analysis was performed using Statistical Analysis Software (SAS) v.9.4 and R Software v.4.0.3 using the quantreg package[13,14].

RESULTS

Baseline characteristics

We identified 838 patients who underwent liver USS with radiology record requests for HCC surveillance within the 6-month recruitment period. On further review, 22 patients were excluded due to prior liver transplantation, 1 due to recent history (< 2 years) of HCC, 9 who did not meet criteria for screening (incorrectly classified as cirrhotic) and 31 with chronic hepatitis B who did not meet guideline criteria for surveillance. 775 patients were included in the final analysis. Baseline characteristics are detailed in Table 1.

The mean age of patients was 60.0 (standard deviation 12.2) years. The majority were male (59.0%), Caucasian (57.0%) and English-speaking (73.9%), with cirrhosis the most common indication for HCC surveillance (55.3%). Of those with cirrhosis, the median MELD score at inclusion was 9. The majority of patients were followed up in hepatology clinics (58.5% general liver, 16.3% pre-transplant, 10.3% nurse-led). The median follow-up time was 27.5 mo (IQR: 26.0-29.0 months). The most commonly used screening modality was ultrasound (86.9%), followed by contrast-enhanced CT liver (9.4%) and contrast-enhanced MRI (3.7%). AFP was performed in conjunction to imaging at least once in 91.9% of patients.

Twenty patients had an interim orthotopic liver transplant, 22 died and 41 were discharged from clinic surveillance, either due to service nonattendance (36%), transfer of care to other institutions (50%), or when surveillance was deemed no longer clinically appropriate (14%). HCCs developed in 22 patients (2.8%) over the course of the study. Of these 22 HCCs, 14 (64%) were detected at an early stage.

Primary outcome

The median PTUDS overall was 84.2% (IQR: 66.3%-96.3%). 13.2% of patients were upto-date for less than 50% of their surveillance time period, representing average



Table 1 Baseline Characteristics of 775 at-risk patients participating in hepatocellular carcinoma surveillance, 2018-2020					
Characteristic		n = 775 (%)	mean ± SD	Median (IQR)	
Age (years)			60.0 ± 12.2		
Sex	Male	457 (59.0)			
	Female	318 (41.0)			
Ethnicity	Caucasian	442 (57.0)			
	Asian	249 (32.1)			
	African/Middle-Eastern	68 (8.8)			
	Unreported	16 (2.1)			
Primary language spoken	English	573 (73.9)			
	Non-English	202 (26.1)			
Indication for hepatocellular carcinoma surveillance	Cirrhosis	429 (55.3)			
	Chronic HBV	343 (44.3)			
	Chronic HCV with Advanced Fibrosis	3 (0.4)			
Aetiology of Cirrhosis, $n = 429$	HBV hepatitis	58 (13.5)			
	HCV hepatitis	136 (31.7)			
	Alcoholic hepatitis	103 (24.0)			
	NASH	64 (14.9)			
	Non-viral, non-alcoholic, non-NASH cirrhosis ¹	68 (15.9)			
HBV anti-viral use, $n = 385$	Yes	206 (53.5)			
	No	179 (46.5)			
Specialty care clinic	General hepatology ²	453 (58.5)			
	Pre-transplant	126 (16.3)			
	Nurse-led surveillance	80 (10.3)			
	Non-liver ³	76 (9.8)			
	Unspecified	40 (5.2)			
MELD score, $n = 429$				9 (7-13)	

¹Includes autoimmune hepatitis, primary biliary cirrhosis, primary sclerosing cholangitis, cryptogenic cirrhosis.

²Includes general liver and outreach clinics.

³Includes clinics such as infectious diseases, renal, general medicine where patients received specialist care.

HBV: Hepatitis B virus; HCV: Hepatitis C virus; NASH: Non-alcoholic steatohepatitis; SD: Standard deviation.

screening intervals of less frequently than 12-monthly (see Figure 2). Only 40% of patients spent at least 90% of their follow-up period up-to-date with surveillance.

Univariable analyses

Older age (≥ 60 years old), cirrhotic status, Non-Asian ethnicity and English-speaking as the primary language correlated with higher continuous adherence to surveillance (see Table 2). Sex and the receipt of anti-viral therapy in chronic hepatitis B patients did not correlate with differences in surveillance adherence, nor did MELD score in cirrhotics.

Continuous adherence to HCC screening varied by aetiology of liver disease. Patients with non-alcoholic steatohepatitis (NASH) had higher median PTUDS (90.9%) compared to those with viral hepatitis (82.3%, P = 0.003) and alcohol-induced liver disease (83.8%, P = 0.07). Those with non-viral, non-alcoholic and non-NASH disease also had statistically higher continuous surveillance adherence (median PTUDS 90.8%) than their counterparts whose aetiologies were viral hepatitis (P = 0.0008), or alcohol (P = 0.07). There were no significant differences in median PTUDS between the patients whose HCCs were detected earlier (n = 14), than in those with late HCC detection (n = 14) 8) (100% *vs* 95.9%, *P* = 0.73).



Table 2 Comparison of surveillance determinants associated with greater continuous hepatocellular carcinoma surveillance among 775 at-risk patients for hepatocellular carcinoma

Factor	Hepatocellular carcinoma surveil	Hepatocellular carcinoma surveillance adherence (PTUDS)			
	< 60 yr	60 yr	<i>P</i> value		
Age	80.6%	86.8%	< 0.001		
	Non-cirrhotic	Cirrhotic			
Cirrhosis status	80.1%	87.3%	< 0.001		
	Non-Asian	Asian			
Ethnicity	85.1%	81.2%	0.04		
	English	CALD			
Primary language spoken	85.7%	80.2%	0.03		
	Male	Female			
Sex	83.3%	85.6%	0.58		
	Treatment naïve	Anti-viral therapy			
Hepatitis B treatment status	79.4%	82.1%	0.07		

PTUDS: Percentage of time up-to-date with hepatocellular carcinoma surveillance; CALD: Culturally and linguistically diverse background.



Figure 1 Scenario calculations of percentage of time up-to-date with surveillance.

Analysis demonstrated significant differences in surveillance adherence based on clinic setting, with highest PTUDS for patients attending liver transplant clinics (median PTUDS 97.4%). This was significantly greater than general hepatology clinics (median PTUDS 82.4%, P < 0.001). Of note, patients attending nurse-led screening clinics achieved continuous surveillance adherence rates that surpassed general hepatology clinics (median PTUDS 90.0%, P = 0.0001). This was not reflected in the performance of non-hepatology clinics, which reported the lowest continuous surveillance adherence rates (median PTUDS 62.3%).

Multivariable analysis

Multiple quantile regression analysis demonstrated that age and clinic type (subspecialty hepatology care vs other) had differential effects across different PTUDS quantiles. Older age (≥ 60 years) increased PTUDS across all quantiles, but this



Figure 2 Distribution of participants within percentage of time up-to-date with hepatocellular carcinoma surveillance categories.

increase was only significant at lower quantiles (25th quantile parameter estimate (PE) = 0.003, P = 0.01; 50th quantile PE = 0.002, P < 0.001). Non-liver-specific clinic care, compared with subspecialty hepatology care, was associated with lower PTUDS across all quantiles (P < 0.001).

The results from subgroup quantile regression analysis of patients attending liverspecific specialty clinics (85.0% of all patients) are demonstrated in Table 3. Primary language spoken, ethnicity, and cirrhosis status were shown to variably affect PTUDS across different quantiles. The association between older age and increased PTUDS was maintained only in the 25^{th} quantile (P = 0.03). Patients with cirrhosis had increased PTUDS compared to their non-cirrhotic counterparts at the 75th quantile (P =0.02). However, African ethnicity and culturally and linguistically diverse (CALD) backgrounds were both significantly associated with lower PTUDS at the 50th quantile (P = 0.048 and P = 0.01). CALD background was also a strong predictor of lower PTUDS at the 75th quantile (P = 0.045).

DISCUSSION

In this Australian retrospective cohort study of HCC surveillance utilisation in 775 atrisk patients, we found that average time up-to-date with 6 monthly screening was 84.2% over a median follow-up of over 2 years. However, less than half (40.8%) of patients spent at least 90% of their surveillance period up-to-date with screening, which represents a concerning proportion of patients at risk of delayed diagnosis of HCC. Younger age, non-cirrhotic status, African ethnicity and CALD background were variably associated across PTUDS quantiles with significantly decreased time upto-date with HCC surveillance on quantile regression analysis.

Our study is one of the few that presents adherence to HCC surveillance using a continuous outcome measure, contrasting with versions of the binary categorisation of 'adherent' and 'non-adherent' of other studies[7,15-19]. Depending on definitions used, these all-or-nothing classifications potentially enforce too stringent or too lax restrictions on surveillance intervals, and distort true interpretation of HCC surveillance application in real-time clinical practice. This study has described more accurate quantification of adherence and thus better identifies patients at risk.

Given heterogeneity in the published definitions of adherence, comparisons of surveillance uptake are limited. However, Goldberg et al[7] reported an analysis of patients with cirrhosis, applying the PTUDS measure to characterise adherence using United States Veterans Health Administration data. This national study found a median PTUDS for surveillance by USS only as 10% (IQR: 0%-29%)[7]. The vast discrepancy in median PTUDS compared with our finding of 84.2% is likely

Table 3 Quantile regression parameter estimates and 95% confidence intervals of factors associated with hepatocellular carcinoma surveillance adherence for the 25th, 50th and 75th guantiles of percentage of time up-to-date with surveillance

	QR Estimates (95%CI)					
	25 th	P value	50 th	P value	75 th	P value
Variables						
Age	0.002 (0.000, 0.004)	0.03				
African ethnicity			-0.089 (-0.177, -0.001)	0.048		
CALD			-0.063 (-0.110, -0.016)	0.01	-0.026 (-0.052, -0.001)	0.045
Cirrhotic status					0.021 (0.004, 0.039)	0.02

CALD: Culturally and linguistically diverse background

multifactorial. First, Goldberg's study encompassed patients followed-up in both community or specialist practices, and found that number of specialist visits was the strongest predictor for greater surveillance participation^[7]. Adherence rates for those in non-specialist care have historically been low, with international and limited local data showing optimal surveillance uptake of only 8.8%-27% [3,20,21]. Specialist input may serve as a surrogate marker for greater frequency of reminders for test follow-up, may select for a group of patients more engaged with healthcare, or indicate more unwell patients undergoing imaging for reasons other than surveillance^[22]. In particular, hepatology care as compared to other subspecialty providers may better identify the at-risk cohort eligible for HCC screening, therefore having 85% of patients in this study engaged in hepatology clinics may explain our comparatively high PTUDS. Furthermore, our study recruited patients based on attendance at an initial HCC screening scan, which likely also represents a more informed and engaged subset of all patients requiring HCC surveillance. Our findings are, however, similar to pooled adherence surveillance estimates of 73.7% for patients enrolled in specialist care from a recent meta-analysis, associating specialist care with higher likelihood of HCC surveillance^[3].

In addition to specialist gastroenterologist/hepatologist care, this study provided insight into other factors correlated with utilisation of HCC surveillance. The high continuous surveillance adherence found in nurse-led clinic-directed care lends support to the effectiveness of this model of care shown in two cohort studies[23,24]. In keeping with several prior studies, we found lower rates of surveillance in younger patients[7,19,25,26]. Studies assessing adherence by cirrhosis status echoed our finding of screening under-utilisation in patients without cirrhosis. This is potentially attributable to a higher perceived HCC risk with cirrhosis stage by patients and providers, suggesting under-recognition of screening eligibility in this group[12,22,27]. Targeted education of providers, particularly primary care physicians who follow most highrisk patients nationally, on screening guidelines that include non-cirrhotic chronic hepatitis B patients, may improve surveillance rates. Receipt of anti-viral therapy, revealed in other studies to increase the likelihood of surveillance adherence, was not significant on multivariable analysis in our cohort of chronic hepatitis B patients[16, 22]

Several studies have reported lower surveillance rates in non-Caucasians, with African American/Black patients significantly less likely to receive surveillance[7,28]. African ethnicity in our cohort was similarly associated with lower PTUDS at the 50th quantile, a concerning finding given higher rates of HCC and younger age at risk within this population[21,29]. This may correlate with our lower screening uptake in patients whose primary language was not English, although to our knowledge, no studies have yet reviewed the impact on HCC screening uptake in patients whose care is provided in a primary language that is not their native tongue. However, other cancer surveillance studies have also identified language as a key barrier in CALD patients' understanding of screening rationale and participation[30,31]. An Australian qualitative study into colorectal screening uptake within culturally and linguistically diverse groups suggested that language barriers hindered otherwise willing participation[30]. A fatalistic view on cancer diagnoses, or a fear of bad luck due to cultural beliefs, were also identified as potentially modifiable barriers to screening adherence, and suggest that culturally-tailored and language-appropriate resources may need to be employed to target diverse populations[30].

In our study, a total of 22 HCCs were detected, of which 14 were early stage and amenable to curative treatment. The annual incidence rate of HCC in our cohort was 1.3 per 100 person-years. This low incidence of HCC may be due to a high proportion in our cohort of people with treated HCV, and HBV without cirrhosis, representing patients with a relatively lower incidence rate of HCC[32]. We did not find a significant difference in overall adherence in patients with HCCs detected at an earlier compared to a later stage. While our study was not powered to answer this question, it provides interesting insight into the effect of surveillance in a cohort with adherence rates closer to the ideal, and highlights the need for quality assessment of surveillance imaging.

Strengths of this study included use of the continuous variable 'percentage of time up-to-date with surveillance' to quantify adherence to HCC screening, and quantile regression modelling to describe covariate effects across the spectrum of PTUDS[3,28]. Our data also reflects contemporary clinical practice, with confirmation of the 'at-risk' history of our surveyed population. This is also the first Australian study of a large cohort, and offers a perspective on surveillance uptake in a population not hindered by costs of screening tests. Prior observational studies have raised concerns that patient-level barriers, such as costs of surveillance investigations, may contribute to poorer surveillance receipt[22,33]. While we were unable to assess socioeconomic factors, as this data was unavailable, concerns regarding financial burden of surveillance measures are likely alleviated by their cost-free nature in Australia's healthcare system. Finally, we accounted for patient receipt of alternate, nonultrasound imaging. This reflects a more realistic practice of clinicians accepting contrast-enhanced CT or MRI liver imaging as appropriate surveillance studies, thus obviating the performance of additional ultrasonography.

Findings from our study should be interpreted within its limitations. As with retrospective study designs, our study is limited by missing data, unmeasured confounders and selection bias. Our analysis involved patients at a single healthcare centre, and may not be generalised to other practice environments, particularly primary care settings. Secondly, patient enrolment based on an initial HCC surveillance USS attendance in a 6 mo period does not incorporate eligible but nonadherent patients, nor those qualifying for screening but who have not been identified by clinicians as at risk of HCC. As such, our adherence rates may be overestimated towards a well-informed, more inherently adherent and correctly identified cohort. However, this large study provides valuable current insight into maintenance of HCC surveillance in patients already identified to be at risk.

CONCLUSION

In conclusion, average time-up-to-date with HCC surveillance in an at-risk Australian cohort was 84.2% following an index screening test performed for the purpose of surveillance. Subspecialty care was associated with higher subsequent adherence to surveillance imaging. Conversely, younger age, non-cirrhotic status, African ethnicity, and CALD background, were variably associated with significantly reduced PTUDS across different PTUDS quantiles. Further research into patient and system barriers towards HCC screening will provide further information regarding provider and patient factors to better guide development of appropriate and targeted interventions to increase adherence to HCC surveillance.

ARTICLE HIGHLIGHTS

Research background

Hepatocellular carcinoma (HCC) surveillance rates reported from the United States and Europe remain low, despite published clinical guideline recommendations. Surveillance patterns in Australia, which has the benefit a universal healthcare program, have not been clearly delineated.

Research motivation

Patients, and evaluate factors associated with greater uptake of HCC cancer screening. In incorporating both frequency of screening and quantity of imaging performed, we aimed to have a more continuous way of standardising 'adherence'. Identification of determinants associated with higher HCC screening adherence aims to guide further



areas for intervention.

Research objectives

As stated above, the objectives were characterising continuous HCC surveillance adherence. This method provides a way of standardising 'adherence', and thus allows for equal comparison between different studies evaluating the concept of HCC screening adherence.

Research methods

This was a retrospective cohort study that incorporated data electronic medical records to obtain patient demographics, clinical history, lab investigations and radiological imaging results. Data analysis was both on the univariate and multivariate level. In particular, quantile regression was performed for the non-parametric outcome variable, and provides greater description of covariate associations across the range of the outcome variable.

Research results

Follow-up of 775 at-risk patients demonstrated that median time-up-to-date with HCC surveillance was 84.2%. However, different patient factors, affected HCC surveillance adherence variably across different ranges of the outcome variable percentage of time up-to-date with HCC surveillance (PTUDS). At the 25th quantile/percentile for PTUDS, older age was associated with greater HCC surveillance. At the 50th quantile, African ethnicity had lower HCC surveillance. At the 75th quantile, cirrhotic status was associated with greater adherence to surveillance. Those of culturally and linguistically diverse backgrounds had lower continuous HCC surveillance rates at both the 50th and 75th quantiles. The ramifications of these findings and identified determinants affecting HCC surveillance participation in other settings, including the primary care setting, are less clear. However, they remain very important areas for further research. In particular, addressing the impact of ethnicity and cultural and linguistic backgrounds on screening uptake may well have beneficial consequent effects in other areas of healthcare.

Research conclusions

The study suggests specific patient and systemic factors that contribute to participation in HCC surveillance. These factors include younger age, non-cirrhotic status, African ethnicity and coming from a culturally and linguistically diverse background, which all are variably associated with lower percentage of time up-to-date with HCC surveillance.

Research perspectives

Future research should be directed at determining interventions aimed at the factors identified in this study to be associated with reduced HCC screening adherence. Those that improve participation in HCC surveillance may well benefit from widespread implementation to improve earlier diagnosis of HCCs.

REFERENCES

- Villanueva A. Hepatocellular Carcinoma. N Engl J Med 2019; 380: 1450-1462 [PMID: 30970190 DOI: 10.1056/NEJMra1713263]
- 2 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- Wolf E, Rich NE, Marrero JA, Parikh ND, Singal AG. Use of Hepatocellular Carcinoma Surveillance 3 in Patients With Cirrhosis: A Systematic Review and Meta-Analysis. Hepatology 2021; 73: 713-725 [PMID: 32383272 DOI: 10.1002/hep.31309]
- 4 Cocker F, Chien Yee K, Palmer AJ, de Graaff B. Increasing incidence and mortality related to liver cancer in Australia: time to turn the tide. Aust N Z J Public Health 2019; 43: 267-273 [PMID: 30958629 DOI: 10.1111/1753-6405.12889]
- 5 Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin 2020; 70: 7-30 [PMID: 31912902 DOI: 10.3322/caac.21590]
- 6 Forner A, Reig M, Bruix J. Hepatocellular carcinoma. Lancet 2018; 391: 1301-1314 [PMID: 29307467 DOI: 10.1016/S0140-6736(18)30010-2]
- 7 Goldberg DS, Taddei TH, Serper M, Mehta R, Dieperink E, Aytaman A, Baytarian M, Fox R, Hunt K, Pedrosa M, Pocha C, Valderrama A, Kaplan DE. Identifying barriers to hepatocellular carcinoma



surveillance in a national sample of patients with cirrhosis. Hepatology 2017; 65: 864-874 [PMID: 27531119 DOI: 10.1002/hep.28765]

- 8 Singal AG, Pillai A, Tiro J. Early detection, curative treatment, and survival rates for hepatocellular carcinoma surveillance in patients with cirrhosis: a meta-analysis. PLoS Med 2014; 11: e1001624 [PMID: 24691105 DOI: 10.1371/journal.pmed.1001624]
- 9 Kanwal F, Singal AG. Surveillance for Hepatocellular Carcinoma: Current Best Practice and Future Direction. Gastroenterology 2019; 157: 54-64 [PMID: 30986389 DOI: 10.1053/j.gastro.2019.02.049]
- 10 Marrero JA, Kulik LM, Sirlin CB, Zhu AX, Finn RS, Abecassis MM, Roberts LR, Heimbach JK. Diagnosis, Staging, and Management of Hepatocellular Carcinoma: 2018 Practice Guidance by the American Association for the Study of Liver Diseases. Hepatology 2018; 68: 723-750 [PMID: 29624699 DOI: 10.1002/hep.29913]
- 11 European Association for the Study of the Liver. EASL Clinical Practice Guidelines: Management of hepatocellular carcinoma. J Hepatol 2018; 69: 182-236 [PMID: 29628281 DOI: 10.1016/j.jhep.2012.03.006
- 12 Zhao C, Jin M, Le RH, Le MH, Chen VL, Wong GL, Wong VW, Lim YS, Chuang WL, Yu ML, Nguyen MH. Poor adherence to hepatocellular carcinoma surveillance: A systematic review and meta-analysis of a complex issue. Liver Int 2018; 38: 503-514 [PMID: 28834146 DOI: 10.1111/liv.13555
- SAS Institute Inc. SAS OnDemand for Academics. Cary, NC, USA, 2021 13
- R Development Core Team. Quantreg: Quantile Regression. R package version 5.85 edition. 14 Vienna, Austria: R Foundation for Statistical Computing, 2020
- Edenvik P, Davidsdottir L, Oksanen A, Isaksson B, Hultcrantz R, Stål P. Application of 15 hepatocellular carcinoma surveillance in a European setting. What can we learn from clinical practice? Liver Int 2015; 35: 1862-1871 [PMID: 25524812 DOI: 10.1111/liv.12764]
- Nam JY, Lee JH, Kim HY, Kim JE, Lee DH, Chang Y, Cho H, Yoo JJ, Lee M, Cho YY, Cho Y, Cho 16 E, Yu SJ, Kim YJ, Yoon JH. Oral Medications Enhance Adherence to Surveillance for Hepatocellular Carcinoma and Survival in Chronic Hepatitis B Patients. PLoS One 2017; 12: e0166188 [PMID: 28099520 DOI: 10.1371/journal.pone.0166188]
- 17 Singal AG, Tiro J, Li X, Adams-Huet B, Chubak J. Hepatocellular Carcinoma Surveillance Among Patients With Cirrhosis in a Population-based Integrated Health Care Delivery System. J Clin Gastroenterol 2017; 51: 650-655 [PMID: 27870642 DOI: 10.1097/MCG.0000000000000708]
- Tavakoli H, Robinson A, Liu B, Bhuket T, Younossi Z, Saab S, Ahmed A, Wong RJ. Cirrhosis 18 Patients with Nonalcoholic Steatohepatitis Are Significantly Less Likely to Receive Surveillance for Hepatocellular Carcinoma. Dig Dis Sci 2017; 62: 2174-2181 [PMID: 28474143 DOI: 10.1007/s10620-017-4595-x]
- Tran SA, Le A, Zhao C, Hoang J, Yasukawa LA, Weber S, Henry L, Nguyen MH. Rate of 19 hepatocellular carcinoma surveillance remains low for a large, real-life cohort of patients with hepatitis C cirrhosis. BMJ Open Gastroenterol 2018; 5: e000192 [PMID: 29607053 DOI: 10.1136/bmjgast-2017-000192]
- Allard N, Cabrie T, Wheeler E, Richmond J, MacLachlan J, Emery J, Furler J, Cowie B. The 20 challenge of liver cancer surveillance in general practice: Do recall and reminder systems hold the answer? Aust Fam Physician 2017; 46: 859-864 [PMID: 29101924]
- 21 Singal AG, Yopp A, S Skinner C, Packer M, Lee WM, Tiro JA. Utilization of hepatocellular carcinoma surveillance among American patients: a systematic review. J Gen Intern Med 2012; 27: 861-867 [PMID: 22215266 DOI: 10.1007/s11606-011-1952-x]
- 22 Wang C, Chen V, Vu V, Le A, Nguyen L, Zhao C, Wong CR, Nguyen N, Li J, Zhang J, Trinh H, Nguyen MH. Poor adherence and low persistency rates for hepatocellular carcinoma surveillance in patients with chronic hepatitis B. Medicine (Baltimore) 2016; 95: e4744 [PMID: 27583921 DOI: 10.1097/MD.00000000004744]
- 23 Aberra FB, Essenmacher M, Fisher N, Volk ML. Quality improvement measures lead to higher surveillance rates for hepatocellular carcinoma in patients with cirrhosis. Dig Dis Sci 2013; 58: 1157-1160 [PMID: 23111632 DOI: 10.1007/s10620-012-2461-4]
- Nazareth S, Leembruggen N, Tuma R, Chen SL, Rao S, Kontorinis N, Cheng W. Nurse-led 24 hepatocellular carcinoma surveillance clinic provides an effective method of monitoring patients with cirrhosis. Int J Nurs Pract 2016; 22 Suppl 2: 3-11 [PMID: 27476494 DOI: 10.1111/ijn.12472]
- 25 Davila JA, Henderson L, Kramer JR, Kanwal F, Richardson PA, Duan Z, El-Serag HB. Utilization of surveillance for hepatocellular carcinoma among hepatitis C virus-infected veterans in the United States. Ann Intern Med 2011; 154: 85-93 [PMID: 21242365 DOI: 10.7326/0003-4819-154-2-201101180-00006
- Park B, Choi KS, Suh M, Shin JY, Jun JK. Factors associated with compliance with 26 recommendations for liver cancer screening in Korea: a nationwide survey in Korea. PLoS One 2013; 8: e68315 [PMID: 23840846 DOI: 10.1371/journal.pone.0068315]
- Wong CR, Garcia RT, Trinh HN, Lam KD, Ha NB, Nguyen HA, Nguyen KK, Levitt BS, Nguyen 27 MH. Adherence to screening for hepatocellular carcinoma among patients with cirrhosis or chronic hepatitis B in a community setting. Dig Dis Sci 2009; 54: 2712-2721 [PMID: 19876735 DOI: 10.1007/s10620-009-1015-x
- 28 Singal AG, Li X, Tiro J, Kandunoori P, Adams-Huet B, Nehra MS, Yopp A. Racial, social, and clinical determinants of hepatocellular carcinoma surveillance. Am J Med 2015; 128: 90.e1-90.e7 [PMID: 25116425 DOI: 10.1016/j.amjmed.2014.07.027]



- 29 Kew MC. Epidemiology of hepatocellular carcinoma in sub-Saharan Africa. Ann Hepatol 2013; 12: 173-182 [PMID: 23396727]
- Javanparast S, Ward PR, Carter SM, Wilson CJ. Barriers to and facilitators of colorectal cancer 30 screening in different population subgroups in Adelaide, South Australia. Med J Aust 2012; 196: 521-523 [PMID: 22571311 DOI: 10.5694/mja11.10701]
- 31 Ferdous M, Goopy S, Yang H, Rumana N, Abedin T, Turin TC. Barriers to Breast Cancer Screening Among Immigrant Populations in Canada. J Immigr Minor Health 2020; 22: 410-420 [PMID: 31346839 DOI: 10.1007/s10903-019-00916-3]
- Desai A, Sandhu S, Lai JP, Sandhu DS. Hepatocellular carcinoma in non-cirrhotic liver: A 32 comprehensive review. World J Hepatol 2019; 11: 1-18 [PMID: 30705715 DOI: 10.4254/wjh.v11.i1.1]
- 33 Farvardin S, Patel J, Khambaty M, Yerokun OA, Mok H, Tiro JA, Yopp AC, Parikh ND, Marrero JA, Singal AG. Patient-reported barriers are associated with lower hepatocellular carcinoma surveillance rates in patients with cirrhosis. Hepatology 2017; 65: 875-884 [PMID: 27531684 DOI: 10.1002/hep.28770]



0 WŰ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2161-2179

DOI: 10.4251/wjgo.v13.i12.2161

ISSN 1948-5204 (online)

ORIGINAL ARTICLE

Retrospective Cohort Study

Comparison of tumor regression grading systems for locally advanced gastric adenocarcinoma after neoadjuvant chemotherapy

Zi-Ning Liu, Yin-Kui Wang, Li Zhang, Yong-Ning Jia, Shan Fei, Xiang-Ji Ying, Yan Zhang, Shuang-Xi Li, Yu Sun, Zi-Yu Li, Jia-Fu Ji

ORCID number: Zi-Ning Liu 0000-0002-4355-6899; Yin-Kui Wang 0000-0002-5607-5138; Li Zhang 0000-0001-5533-8568; Yong-Ning Jia 0000-0002-9215-7907; Shan Fei 0000-0002-4792-3385; Xiang-Ji Ying 0000-0002-1713-3788; Yan Zhang 0000-0003-2096-6526; Shuang-Xi Li 0000-0002-0566-7651; Yu Sun 0000-0003-3024-0331; Zi-Yu Li 0000-0001-5580-4979; Jia-Fu Ji 0000-0001-6878-5543.

Author contributions: Liu ZN,

Wang YK and Li ZY designed this study; Wang YK and Liu ZN enrolled patients and collected clinical data; Zhang L and Sun Y reviewed the samples; Liu ZN and Ying XJ conducted statistical analysis; Liu ZN is responsible for data visualization; Liu ZN and Wang YK drafted this article; All authors read and approved the final manuscript; Liu ZN, Wang YK and Zhang L contributed equally to this work.

Institutional review board

statement: The Ethics Committee of Peking University Cancer Hospital approved this study. All procedures followed were in accordance with the ethical standards of the responsible committee on human experimentation (institutional and national) and with the Helsinki Declaration of 1964 and later

Zi-Ning Liu, Yin-Kui Wang, Yong-Ning Jia, Shan Fei, Xiang-Ji Ying, Yan Zhang, Shuang-Xi Li, Zi-Yu Li, Jia-Fu Ji, Key Laboratory of Carcinogenesis and Translational Research (Ministry of Education/Beijing), Gastrointestinal Cancer Center, Peking University Cancer Hospital and Institute, Beijing 100142, China

Li Zhang, Yu Sun, Department of Pathology, Key Laboratory of Carcinogenesis and Translational Research (Ministry of Education/Beijing), Peking University Cancer Hospital and Institute, Beijing 100142, China

Corresponding author: Zi-Yu Li, MD, PhD, Professor, Key Laboratory of Carcinogenesis and Translational Research (Ministry of Education/Beijing), Gastrointestinal Cancer Center, Peking University Cancer Hospital and Institute, No. 52 Fucheng Road, Haidian District, Beijing 100142, China. ziyu li@hsc.pku.edu.cn

Abstract

BACKGROUND

Current tumor regression grade (TRG) evaluations are based on various systems which brings confusion for oncologists and pathologists when interpreting results. The recent six-tier system (JGCA2017-TRG) recommended by the Japanese Gastric Cancer Association (JGCA) is worth investigating, as four-tier TRG systems are favored in various parts of the world.

AIM

To compare the predictive accuracies of five published TRG systems.

METHODS

Data were retrospectively collected from patients with locally advanced gastric cancer (LAGC) who underwent neoadjuvant chemotherapy followed by D2 Lymphadenectomy between January 2005 and January 2014 at our institution. Outcomes were overall survival (OS) and disease-free survival (DFS), which were evaluated separately using the following TRG systems: JGCA2017, JGCA, Becker, AJCC/CAP, and Mandard.

RESULTS

All five published TRG systems were independent predictors for OS and DFS. Concordance indices of the JGCA2017, JGCA, Becker, AJCC/CAP-TRG, and Mandard systems were 0.651/0.648 0.652/0.649, 0.693/0.695, 0.688/0.685, and



versions. Informed consents were obtained from all patients for being included in the study. This study does not involve animal study.

Informed consent statement: All study participants or their legal guardian provided informed written consent about personal and medical data collection prior to study enrolment.

Conflict-of-interest statement: All authors declare no conflict of interest.

Data sharing statement: The datasets during and/or analyzed during the current study are available from the corresponding author on reasonable request.

STROBE statement: The authors have read the STROBE Statement - checklist of items, and the manuscript was prepared and revised according to the STROBE Statement-checklist of items.

Supported by the Beijing Municipal Health Commission, No. DFL20181103 and No. ZYLX201701.

Country/Territory of origin: China

Specialty type: Oncology

Provenance and peer review:

Unsolicited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C, C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the

0.674/0.675 for OS and DFS, respectively. The four-tier Becker system showed the highest c-index, which was significantly greater than that of the six-tier JGCA2017 and five-tier JGCA systems (P < 0.05 in OS and DFS). When residual tumor percentages were reset as: "no residual tumor", < 10%, < 100%, and "no response", the rearranged cutoff values achieved a maximum c-index with 0.728 for OS and 0.737 for DFS, which was superior to the other five systems.

CONCLUSION

The newly introduced six-tier JGCA-TRG system cannot increase prognostic stratification. The four-tier Becker system is more suitable for LAGC patients. A population-based study is warranted to define the optimal criterion for TRG in LAGC patients.

Key Words: Gastric cancer; Neoadjuvant chemotherapy; Tumor regression grade; Survival; Concordance index

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Current Tumor regression grade (TRG) evaluations are based on various systems bringing confusion to oncologists and pathologists when interpreting results in similar clinical contexts. On the other hand, the recent six-tier system tumor regression grade (JGCA2017-TRG) recommended by Japanese Gastric Cancer Association (JGCA) is investigational. This is the first report of the use of the c-index to evaluate predictive accuracies of five published TRG systems in gastric cancer. With a satisfying sample size, our results gave clinicians a better understanding of the TRG, especially the residual tumor percentage, in gastric cancer and furthermore alleviates the oncologists and pathologist's workload.

Citation: Liu ZN, Wang YK, Zhang L, Jia YN, Fei S, Ying XJ, Zhang Y, Li SX, Sun Y, Li ZY, Ji JF. Comparison of tumor regression grading systems for locally advanced gastric adenocarcinoma after neoadjuvant chemotherapy. World J Gastrointest Oncol 2021; 13(12): 2161-2179

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2161.htm DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2161

INTRODUCTION

Although surgical resection is the mainstay therapy of locally advanced gastric cancer (LAGC), neoadjuvant chemotherapy (NACT) has now been widely adopted for LAGC in Europe and most recently in China due to the solid evidence that it reduces the risk of recurrence and improves overall survival^[1-3]. Theoretical benefits of NACT are downstaging the primary tumor, increasing the R0 resection rate, and treatment of potential micrometastases.

The effects of NACT on the tumor can be histopathologically evaluated in subsequent resection specimens by applying pathological tumor regression grading (TRG) systems. There are currently more than five commonly used TRG systems for GC across the world with different principles, different layers, and different cutoff values [4,5]. These various practices in TRG evaluation place a large burden on oncologists and pathologists and make it hard to interpret results from different systems in similar clinical contexts. Pathologists may also be required to be familiar with more than one TRG system in daily practice[4].

Currently, most pathologists favor four-tier TRG systems in gastrointestinal cancer. There are the Becker system and the American Joint Committee on Cancer (AJCC)/ College of American Pathologists (CAP) system, as these have superior inter-rater agreement with no loss of discriminatory ability [4,6]. In October 2017, the 15th Japanese Classification of Gastric Carcinoma proposed a new six-tier pathological regression evaluation for GC based on its previous Japanese Gastric Cancer Association (JGCA) TRG system[7]. This added the following sub-groupings of JGCA-TRG grade 2 (residual tumor 1%-33%): grade 2a (residual tumor 10%-33%) and 2b (residual tumor <



Received: May 12, 2021 Peer-review started: May 12, 2021 First decision: July 14, 2021 Revised: July 25, 2021 Accepted: September 15, 2021 Article in press: September 15, 2021 Published online: December 15, 2021

P-Reviewer: Kao JT, Mohamed SY S-Editor: Wang LL L-Editor: Filipodia P-Editor: Guo X



10%) according to the result of JCOG1004-A[8,9]. This new classification did not draw much attention in Western countries, only in East Asia[10]. However, as both the JGCA and AJCC/CAP criteria obtained good consistency in Chinese patients, extensive validation is warranted to verify the adjustment in the new JGCA-TRG system [11]. Furthermore, an optimized histopathological evaluation system for predicting patient prognosis is urgently needed to resolve this contentious issue.

Therefore, the present study sought to validate the utility of the new JGCA-TRG system (JGCA2017-TRG). This was achieved by comparing JGCA2017-TRG with different TRG systems and exploring meaningful cutoff values of residual tumor percentage based on a current dataset comprising 413 LAGC patients who received D2 Lymphadenectomy following NACT.

MATERIALS AND METHODS

Patients

Data were obtained from a retrospective database of all patients receiving NACT followed by curative gastrectomy at the Peking University Cancer Hospital and Institute ("The Institute") from January 1, 2005 to January 1, 2014.

The inclusion criteria included: (1) Proven diagnosis of gastric adenocarcinoma by preoperative pathology; (2) No signs of distant metastasis at first visit; (3) Complete perioperative medical record and documentation of NACT in the Institute; and (4) Curative gastrectomy with D2 Lymph node resection performed at the Institute.

The exclusion criteria were as follows: (1) Insufficient record of clinicopathological information; (2) Patients who received radiotherapy or targeted therapy before surgery; (3) Specimen information was not available; (4) Patients with R1/R2 resection or suspected of having metastasis when surgery was performed; (5) Non-adenocarcinoma diagnosis based on postoperative histological findings (except for complete response cases); (6) Remnant gastric cancer; and (7) Died within 30 d post-surgery.

Regimen and radical surgery

Except for eight patients with logistic reasons, e.g., poor economic status or severe adverse events, all patients received at least two cycles of chemotherapy. In summary, 364 patients received platin-based doublet regimens, 25 patients received Taxol-based doublet regimens, and 24 patients received Taxol-platin-based triplet regimens. Supplementary Table 1 describes the detailed dosing regimens.

To assess the influence of the treatment duration, three 14 d cycles of FOLFOX or POS were regarded as two 21 d cycles of treatment. Dosage reduction or withdrawal was applied in cases of severe adverse events during chemotherapy; this was determined by the clinician according to the Common Terminology Criteria for Adverse Events (CTCAE) version 4.0, as in our previous study[12]. After two to three chemotherapy cycles, the antitumor effect was evaluated using abdominal computed tomography (CT). In most cases, two or three alignment cycles were performed. The therapy was prematurely terminated in cases of disease progression. Otherwise, gastrectomy or continued NACT was considered after obtaining informed consent and approval from patients. Subtotal or total gastrectomy plus D2 Lymphadenectomy was performed according to the JGCA guideline[13].

Histopathological examinations

The pathological preparation of the surgical specimens was commenced immediately after the operation. After recording the localization, measurement, and complete inclusion of visible tumor or suspected tumor areas, a surgeon identified the lymph node groups in the specimen. They were dissected and labeled separately from the main stomach specimen. Generally, the stomach tissue was fixed in 10% neutral buffered formalin overnight and then embedded in paraffin wax. Sections of 5 µm thickness were cut and stained with hematoxylin and eosin (H&E) for microscopic examination, all according to standard procedures. The histological patterns, degrees of differentiation, the extent of tumor invasion, number of regional lymph node metastases, and lymphovascular invasion (LVI), were recorded in each patient's pathology report. This information was then integrated according to the 8th AJCC Cancer Staging Manual and World Health Organization pathologic classifications by two oncologists (Liu ZN and Wang YK)[14,15].

All normal sections were stained with H&E and preserved in paraffin. From December 2017, two designated pathologists (Zhang L and Sun Y) were responsible for reviewing the extent of tumor regression. All patients' H&E slides were re-



examined using bright-field fluorescence microscopy for discrimination between necrotic or heat-fixed tissue and viable tissue. The extent of regressive tumors was evaluated and recorded according to: (1) The amount of viable tumor *vs* fibrotic tissue, which ranged from a total lack of tumor regression to complete response with no viable tumor identified; and (2) The percentage of the viable residual tumor, which was calculated by dividing the viable residual tumor area by the total tumor area. Tumor regression grades were then allocated according to the JGCA2017, JGCA, Becker, AJCC/CAP, and Mandard systems (Table 1)[7,8,16-18]. As for the tumor regression grade, the JGCA2017 criteria example for each grade is shown in Figure 1. The review task ended in December 2019.

Data collection

In addition to histopathological features, other included patient characteristics were age, sex, body mass index (BMI), American Society of Anesthesiologists score (ASA), ECOG performance status, tumor location, tumor diameter (on short axis), type of resection, type of NACT regimens, complications grade by Clavien-Dindo classification, NACT cycles, survival time, and survival status[19]. The follow-up methods were described in our earlier study[20]. Disease-free survival (DFS) was calculated from the date of surgery to the date of recurrence or metastasis.

Statistical analysis

Continuous variables were summarized as the median (IQR) and were compared across groups using the Kruskal-Wallis test. Categorical variables were analyzed using the Chi-squared test. The relationships between clinical and pathological factors and long-term DFS and OS were assessed using univariate log-rank tests and a multivariate Cox proportional hazard model. Tumor or treatment characteristics that achieved a *P* value < 0.10 in univariate analysis were included in the multivariate analysis. The prognostic strength and the discrimination ability of each TRG system were assessed using the concordance index (c-index \pm SE), with a concordance index of 1 indicating perfect prediction and 0.5 indicating no discrimination. The c-index was calculated and compared using the "survcomp" R package[21]. Testing for trends was based on various statistical hypotheses when necessary. For all analyses, *P* < 0.05 was considered to be statistically significant. Statistical analyses were performed using SE STATA (Stata Statistical Software, release 15.1; Stata Corp, College Station, TX, United States).

RESULTS

Patient characteristics

A total of 413 patients met the inclusion criteria and were included in this study (Figure 2). All achieved total tumor clearance (R0). The patients had a median age of 61 years (range 24-82) and were predominantly male (73.61%). Tumor localization was proximal (including esophagogastric junction Siewert III) in 166 cases, body in 51 cases, distal in 170 cases, and 26 patients had tumor involvement in the whole stomach (linitis plastica). Most patients received preoperative therapy of 5-Fu-based oxaliplatin doublet regimen (88.14%) and 105 patients did not receive adjuvant treatment after complete resection (25.42%). The demographic data of these patients are shown in Table 2, stratified by the JGCA2017-TRG system.

Tumor regression assessment

According to the JGCA system, 26 cases were grade 0 (6.30%), 205 were grade 1a (49.64%), 78 were grade 1b, 68 were grade 2 (16.46%) including 29/39 (7.02%/9.44%) in grades 2a/2b according to the JGCA2017 classification, and 36 patients were grade 3 (8.72%; Table 2, Supplementary Table 2). Similarly, the subgroup frequencies according to the Becker, AJCC/CAP, and Mandard systems are presented in Supplementary Table 3-6, respectively. Significant differences were found in the ypT, ypN, ypTNM, and LVI stages in all five systems. The correlation coefficients of ypT were 0.619, 0.587, 0.662, 0.639, and 0.616 for the JGCA017, JGCA, Becker, AJCC/CAP and Mandard systems, respectively. On the other hand, no statistical significance was found between the NACT regimen and the TRG grade or between the duration of NACT and the TRG grade in any system.

Table 1 Criteria of five tumor regression grading systems			
TRG system	Description		
JGCA/JGCA2017-TRG			
0	No response		
1a	67%-99% residual tumor/tumor bed		
1b	34%-66% residual tumor/tumor bed		
2/2a	10%-33% residual tumor/tumor bed		
/2b	< 10% residual tumor/tumor bed		
3	Complete response		
Becker-TRG			
1a	Complete response		
1b	< 10% residual tumor/tumor bed		
2	10%-50% residual tumor/tumor bed		
3	> 50% residual tumor/tumor bed		
AJCC/CAP-TRG			
0	No residual tumor		
1	Single cells or rare small groups of cancer cells		
2	More than single cells or rare small groups of cancer cells with evident tumor regression		
3	Extensive residual tumor or no response		
Mandard-TRG			
1	No residual tumor		
2	Rare residual tumor		
3	Fibrosis outgrowing residual tumor		
4	Residual tumor outgrowing fibrosis		
5	No response		

TRG: Tumor regression grade; JGCA: Japanese Gastric Cancer Association; AJCC: American Joint Committee on Cancer; CAP: College of American Pathologists.

Survival analysis and performance evaluation

The median follow-up was at 62 mo, with an IQR of 4.5 to 210 mo. At the final followup, 209 patients had recurrence, and 200 died due to cancer. Kaplan-Meier curves for OS and DFS based on each system are presented in Figures 3 and 4. In the univariate analyses, all five regression classification systems had prognostic relevance (Table 3). Although all five systems revealed statistical trends towards an increase in the risk of OS and DFS (P_{trend} < 0.001), JGCA2017 grade 2a showed a higher OS risk compared with grade 1b despite no statistical intergroup significance (HR: 1.06; 95%CI: 0.59-1.89; P = 0.855). The C-index for the six-tier JGCA2017, five-tier JGCA, four-tier Becker, four-tier AJCC/CAP, and five-tier Mandard systems was 0.651 ± 0.027 , 0.652 ± 0.027 , 0.693 ± 0.033, 0.688 ± 0.031, and 0.674 ± 0.028, respectively, for OS, and 0.648 ± 0.028, 0.649 ± 0.028 , 0.695 ± 0.034 , 0.685 ± 0.031 , and 0.675 ± 0.028 , respectively, for DFS. The four-tier Becker system had the highest c-index and was statistically significantly more accurate in predicting survival and recurrence than the six- or five-tier JGCA systems (Becker vs JGCA2017, P = 0.006 for OS, P = 0.002 for DFS; Becker vs JGCA, P = 0.007 for OS, P = 0.003 for DFS). The c-indices were comparable between the Becker and AJCC/CAP systems (P = 0.397 for OS and P = 0.273 for DFS), and between the Becker and Mandard systems (P = 0.148 for OS and P = 0.136 for DFS), while the predictive ability of the four-tier AJCC/CAP system was more accurate than the five-tier Mandard system for OS (P = 0.039) under similar evaluation principles.

Multivariate analysis for overall OS and DFS were then performed, including features that were related to poorer survival prognosis in univariate analysis (*P* <



Table 2 Clinical and demographic characteristics of the study population				
Characteristics	n (%)			
No. of patients	413			
Age, median (IQR), yr	61 (54-67)			
BMI, median (IQR), (kg/m ²)	23.04 (20.83-25.10)			
Male	304 (73.61)			
ASA score				
1	102 (24.70)			
2	255 (61.74)			
3	56 (13.56)			
ECOG				
0	229 (55.45)			
1	168 (40.68)			
2	16 (3.87)			
Location				
Upper	166 (40.19)			
Middle	51 (12.35)			
Lower	170 (41.16)			
Diffuse	26 (6.30)			
Diameter (cm)	3.0 (1.5-4.0)			
Differentiation				
Well	27 (6.54)			
Moderate	176 (42.86)			
Poor	209 (50.61)			
Mucinous or signet cell	85 (20.58)			
LVI	132 (31.96)			
Cycles of treatment	2 (2-3)			
урТ				
ypT0	37 (8.96)			
ypT1	25 (6.05)			
ypT2	55 (13.32)			
ypT3	66 (15.98)			
ypT4	230 (55.69)			
ypN				
N0	169 (40.92)			
N1	64 (15.50)			
N2	70 (16.95)			
N3	110 (26.63)			
ypStage				
pCR	32 (7.75)			
Ι	57 (13.80)			
П	116 (28.09)			
Ш	208 (50.36)			



Total gastrectomy	180 (43.58)
Regimen	
Platin-based	364 (88.14)
Taxol-based	25 (6.05)
Triplet	24 (5.81)
Adjuvant chemotherapy	308 (74.58)
Postoperative complications	
Grade 0-1	277 (67.07)
Grade 2	78 (18.89)
Grade 3-4	58 (14.04)
JGCA2017-TRG	
Grade 3 (no residual)	36 (8.72)
Grade 2b (< 10%)	39 (9.44)
Grade 2a (10%-33%)	29 (7.02)
Grade 1b (34%-66%)	78 (18.89)
Grade 1a (67%-99%)	205 (49.64)
Grade 0 (no response)	26 (6.30)
JGCA-TRG	
Grade 3 (no residual)	36 (8.72)
Grade 2 (< 33%)	68 (16.46)
Grade 1b (34%-66%)	78 (18.89)
Grade 1a (67%-99%)	205 (49.64)
Grade 0 (no response)	26 (6.30)
Becker-TRG	
1a (no residual)	36 (8.72)
1b (< 10%)	39 (9.44)
2 (10%-50%)	65 (15.74)
3 (> 50%)	273 (66.10)
AJCC-TRG	
0 (complete response)	36 (8.72)
1 (moderate response)	48 (11.62)
2 (minimal response)	89 (21.55)
3 (poor response)	240 (58.11)
Mandard-TRG	
1 (complete response)	36 (8.72)
2 (Fibrosis + scattered tumor cells)	48 (11.62)
3 (Fibrosis predominance + tumor cells)	89 (21.55)
4 (Tumor cells preponderance + fibrosis)	214 (51.82)
5 (No response)	26 (6.30)

BMI: Body mass index; ASA: American Society of Anesthesiologists; ECOG: Eastern Cooperative Oncology Group; LVI: Lymphovascular invasion; NACT: Neoadjuvant chemotherapy; TRG: Tumor regression grade.

Baisbideng® WJGO | https://www.wjgnet.com

Liu ZN et al. TRG comparison for gastric adenocarcinoma

Table 3 Univariate analyses for overall survival and progression-free survival using a Cox proportional hazards model					
Variables	OS		DFS		
	HR (95%CI)	P value	HR (95%CI)	P value	
Age					
≤ 65					
> 65	1.12 (0.84-1.51)	0.439	1.12 (0.84-1.49)	0.4351	
BMI					
≤ 23.9	1.39 (1.04-1.87)	0.027	1.31 (0.98-1.74)	0.065	
> 23.9					
Gender					
Male					
Female	1.14 (0.84-1.56)	0.393	1.06 (0.78-1.45)	0.690	
ASA score					
1-2					
3	1.00 (0.67-1.49)	0.993	0.97 (0.65-1.43)	0.874	
ECOG					
0					
1-2	1.29 (0.98-1.70)	0.073	1.30 (0.99-1.71)	0.056	
Location					
Diffuse vs Upper	3.32 (2.07-5.31)	< 0.001	2.97 (1.86-4.73)	< 0.001	
Diffuse vs Middle	2.63 (1.51-4.56)	< 0.001	2.28 (1.32-3.94)	0.003	
Diffuse vs Lower	3.94 (2.45-6.35)	< 0.001	3.52 (2.19-5.65)	< 0.001	
Diffuse	1.00		1.00		
Diameter (cm)					
≤5					
> 5	2.79 (2.00-3.88)	< 0.001	2.99 (2.17-4.13)	< 0.001	
Differentiation					
Well-Moderate					
Poor	1.43 (1.08-1.90)	0.012	1.51 (1.15-1.99)	0.003	
Histology					
Non-mucinous					
Mucinous or signet cell	1.86 (1.39-2.53)	< 0.001	1.77 (1.31-2.40)	< 0.001	
Lymphovascular invasion					
No					
Yes	2.75 (2.08-3.64)	< 0.001	2.91 (2.21-3.83)	< 0.001	
урТ					
урТ0-2					
ypT3-4	3.54 (2.35-5.36)	< 0.001	3.66 (2.44-5.49)	< 0.001	
ypN					
ypN0					
ypN+	3.50 (2.50-4.90)	< 0.001	3.59 (2.58-4.98)	< 0.001	
Resection type					
Subtotal					


Total	1.79 (1.35-2.36)	< 0.001	1.74 (1.32-2.28)	< 0.001
Cycle of NACT				
≤2				
> 2	1.18 (0.89-1.56)	0.247	1.18 (0.90-1.55)	0.233
NACT regimen				
Platin-based	1.00		1.00	
Paclitaxel-based	1.10 (0.62-1.92)	0.752	1.27 (0.75-2.15)	0.373
Triplet drug	1.05 (0.59-1.89)	0.862	1.03 (0.57-1.84)	0.930
Adjuvant chemotherapy				
Received				
Not received	1.36 (1.00-1.85)	0.050	1.18 (0.87-1.60)	0.286
Complications				
Clavien-dindo 0-2				
Clavien-dindo 3-4	1.15 (0.78-1.69)	0.491	1.11 (0.76-1.63)	0.585
JGCA2017-TRG				
Grade 3 (no residual)	1.00		1.00	
Grade 2b (< 10%)	8.97 (2.06-39.02)	0.003	8.75 (2.01-38.09)	0.004
Grade 2a (10%-33%)	13.55 (3.11-58.93)	0.001	14.03 (3.23-61.06)	< 0.001
Grade 1b (34%-66%)	12.83 (3.10-53.18)	< 0.001	14.05 (3.40-58.09)	< 0.001
Grade 1a (67%-99%)	15.15 (3.74-61.42)	< 0.001	15.55 (3.84-62.97)	< 0.001
Grade 0 (no response)	20.24 (4.67-87.68)	< 0.001	21.15 (4.88-91.67)	< 0.001
JGCA-TRG				
Grade 3 (no residual)	1.00		1.00	
Grade 2 (< 33%)	10.79 (2.59-45.05)	0.001	10.79 (2.58-45.05)	0.001
Grade 1b (34%-66%)	12.83 (3.10-53.18)	< 0.001	14.04 (3.40-58.05)	< 0.001
Grade 1a (67%-99%)	15.15 (3.74-61.42)	< 0.001	15.54 (3.84-62.93)	< 0.001
Grade 0 (no response)	20.24 (4.67-87.66)	< 0.001	21.18 (4.89-91.78)	< 0.001
Becker-TRG				
1a (no residual)	1.00		1.00	
1b (< 10%)	8.98 (2.06-39.06)	0.003	8.74 (2.01-38.05)	0.004
2 (10%-50%)	12.19 (2.92-50.87)	0.001	12.72 (3.05-53.06)	< 0.001
3 (> 50%)	15.50 (3.84-62.62)	< 0.001	16.15 (4.00-65.22)	< 0.001
AJCC-TRG				
0 (complete response)	1.00		1.00	
1 (moderate response)	10.46 (2.46-44.48)	0.001	10.31 (2.42-43.90)	0.002
2 (minimal response)	11.21 (2.71-46.34)	0.001	11.67 (2.83-48.22)	0.001
3 (poor response)	16.31 (4.03-65.97)	< 0.001	16.94 (4.19-68.49)	< 0.001
Mandard-TRG				
1 (complete response)	1.00		1.00	
2 (Fibrosis + scattered tumor cells)	10.46 (2.46-44.48)	0.001	10.33 (2.43-43.95)	0.002
3 (Fibrosis predominance + tumor cells)	11.20 (2.71-46.30)	0.001	11.66 (2.82-48.16)	0.001
4 (Tumor cells preponderance + fibrosis)	15.85 (3.91-64.19)	< 0.001	16.48 (4.07-66.71)	< 0.001
5 (No response)	20.27 (4.68-87.81)	< 0.001	21.22 (4.90-91.96)	< 0.001



BMI: Body mass index; ASA: American Society of Anesthesiologists; ECOG: Eastern Cooperative Oncology Group; NACT: Neoadjuvant chemotherapy; TRG: Tumor regression grade; HR: Hazard ratio; DFS: Disease-free survival; OS: Overall survival.



Figure 1 Tumor regression grading according to 15th Japanese Classification of Gastric Carcinoma criteria. A: Grade 3 (complete regression); B: Grade 2b (5% residual tumor); C: Grade 2a (30% residual tumor); D: Grade 1b (50% residual tumor); E: Grade 1a (70% residual tumor); F: Grade 0 (No response) (original magnification 20×).

> 0.10): BMI, ECOG, tumor location, diameter in short axis, differentiation, histology type, LVI, resection type, adjuvant chemotherapy, and ypT and ypN stages. After adjusting for potential confounders in the multivariate Cox regression model, BMI, histology type, LVI, and the ypN stage were independent predictors for OS, while LVI and the ypN stage were independent risk factors for DFS. All five TRG systems showed significant differences when setting the "complete response" group as a reference (Table 4). However, the increase in the hazard ratio was not entirely in accord with the increase in the TRG grade in the JGCA2017, AJCC/CAP, and Mandard



Raishideng® WJGO | https://www.wjgnet.com

Table 4 Multivariate Cox hazards regression model for the predictable risk of overall survival and disease-free survival in different covariate inclusion in whole patients

	Whole patients (<i>n</i> = 413)			
Covariates	OS		DFS	
	HR	P value	HR	P value
BMI ≤ 23.9	1.37 (1.01-1.87)	0.045	1.28 (0.95-1.72)	0.109
ECOG > 0	1.19 (0.87-1.61)	0.271	1.18 (0.88-1.60)	0.272
Linitis plastica	1.74 (0.97-3.13)	0.063	1.30 (0.74-2.30)	0.362
Diameter > 5 cm	1.20 (0.77-1.88)	0.426	1.43 (0.93-2.19)	0.102
Poorly differentiated	1.16 (0.85-1.57)	0.345	1.24 (0.92-1.66)	0.160
Mucinous or signet cell	1.45 (1.03-2.05)	0.036	1.32 (0.94-1.84)	0.111
Lymphovascular invasion	1.53 (1.12-2.10)	0.008	1.61 (1.18-2.19)	0.002
урТ3-4	1.45 (0.92-2.28)	0.113	1.52 (0.97-2.37)	0.065
ypN+	1.96 (1.35-2.85)	< 0.001	1.94 (1.34-2.82)	< 0.001
Total gastrectomy	1.30 (0.94-1.79)	0.118	1.23 (0.90-1.69)	0.202
Without AC	1.35 (0.98-1.87)	0.066	Not included	NA
JGCA2017-TRG (Model 1)				
Grade 3 (no residual)	1.00		1.00	
Grade 2b (< 10%)	4.69 (1.04-21.08)	0.044	4.50 (1.00-20.27)	0.050
Grade 2a (10%-33%)	5.48 (1.19-25.23)	0.029	5.50 (1.20-25.26)	0.028
Grade 1b (34%-66%)	5.32 (1.22-23.30)	0.026	5.73 (1.32-24.88)	0.020
Grade 1a (67%-99%)	6.69 (1.55-28.96)	0.011	6.22 (1.44-26.81)	0.014
Grade 0 (no response)	8.60 (1.87-39.58)	0.006	8.44 (1.84-38.76)	0.006
JGCA-TRG (Model 2)				
Grade 3 (no residual)	1.00			
Grade 2 (< 33%)	5.00 (1.15-21.78)	0.032	4.90 (1.13-21.30)	0.034
Grade 1b (34%-66%)	5.27 (1.21-23.05)	0.027	5.67 (1.30-24.61)	0.021
Grade 1a (67%-99%)	6.63 (1.53-28.66)	0.011	6.16 (1.43-26.52)	0.015
Grade 0 (no response)	8.48 (1.84-39.00)	0.006	8.31 (1.81-38.15)	0.006
Becker-TRG (Model 3)				
1a (no residual)	1.00		1.00	
1b (< 10%)	4.74 (1.05-21.30)	0.043	4.57 (1.02-20.57)	0.047
2 (10%-50%)	5.11 (1.16-22.51)	0.031	5.13 (1.17-22.49)	0.030
3 (> 50%)	6.77 (1.57-29.14)	0.010	6.64 (1.55-28.46)	0.011
AJCC-TRG (Model 4)				
0 (complete response)	1.00		1.00	
1 (moderate response)	5.39 (1.22-23.78)	0.026	5.34 (1.21-23.50)	0.027
2 (minimal response)	5.01 (1.15-21.85)	0.032	5.05 (1.16-21.93)	0.031
3 (poor response)	6.72 (1.56-28.97)	0.011	6.53 (1.52-28.05)	0.012
Mandard-TRG (Model 5)				
1 (complete response)	1.00		1.00	
2 (Fibrosis + scattered tumor cells)	5.37 (1.22-23.68)	0.026	5.31 (1.21-23.39)	0.027
3 (Fibrosis predominance + tumor cells)	4.95 (1.14-21.60)	0.033	4.98 (1.15-21.62)	0.032



Liu ZN et al. TRG comparison for gastric adenocarcinoma

4 (Tumor cells preponderance + fibrosis)	6.44 (1.49-27.87)	0.013	6.26 (1.45-26.93)	0.014
5 (No response)	8.44 (1.83-38.86)	0.006	8.41 (1.83-38.60)	0.006

BMI: Body mass index; ECOG: Eastern Cooperative Oncology Group; AC: Adjuvant chemotherapy; TRG: Tumor regression grade; HR: Hazard ratio; DFS: Disease-free survival; OS: Overall survival.



Figure 2 Selection of patients for inclusion.

systems. In fact, the intergroup differences were not statistically significant when the "complete response" group was absent in each system. Only a marginal difference was found between JGCA2017-TRG grade 0 (no response) vs 2b (< 10%) for OS (HR: 1.84; 95%CI: 0.90-3.75; *P* = 0.096) and DFS (HR: 1.87; 95%CI: 0.92-3.83; *P* = 0.085).

Rearranged cutoff values based on current residual tumor percentage

According to the previous analysis, a comparison of the five systems revealed the Becker system to enable the best prognostic differentiation between subgroups across the whole patient cohort. The AJCC/CAP system, although having the second-highest c-index, did not provide better intergroup discrimination in multivariate analysis. According to the JGCA2017 criteria, two cutoff values of residual tumor percentage -10% and 100% - were of more clinical significance than any other commonly used cutoff percentages except for total regression. Despite the intergroup differences being marginal, a higher c-index of 0.728 ± 0.035 for OS and 0.737 ± 0.035 for DFS, could be achieved based on the following rearranged residual tumor percentage cutoffs: 0 (no residual tumor; reference), < 10% (HR: 4.61; 95%CI: 1.02-20.73; *P* = 0.047 for OR; HR: 4.46; 95%CI: 0.99-20.08; P = 0.051 for DFS), 10-99% (HR: 5.98; 95%CI: 1.40-25.63; P = 0.016 for OS; HR: 5.93; 95% CI-25.29; *P* = 0.016 for DFS), no response (HR: 8.36; 95% CI: 1.82-38.44; *P* = 0.006 for OS; HR: 8.33; 95%CI: 1.82-38.23; *P* = 0.006 for DFS). There was a significant difference in the prognostic ability for DFS (P = 0.046) and a borderline significance for OS (P = 0.073) between the rearranged cutoffs and the Becker system (Table 5).

DISCUSSION

Neoadjuvant chemotherapy followed by surgery and adjuvant chemotherapy is the



Table 5 The pairwise comp	arison of C-indexes	between differei	nt tumor regressi	on grade based on C	ox regression for	overall survival
	JGCA2017	JGCA	Becker	AJCC/CAP	Mandard	Modified
Overall survival						
JGCA2017	1.000	0.308	0.006	0.018	0.053	< 0.001
JGCA		1.000	0.007	0.021	0.063	< 0.001
Becker			1.000	0.397	0.148	0.073
AJCC/CAP				1.000	0.039	0.062
Mandard					1.000	0.005
Modified						1.000
Disease-free survival						
JGCA2017	1.000	0.320	0.002	0.021	0.033	< 0.001
JGCA		1.000	0.003	0.025	0.040	< 0.001
Becker			1.000	0.273	0.136	0.046
AJCC/CAP				1.000	0.112	0.024
Mandard					1.000	0.002
Modified						1.000

JGCA: Japanese Gastric Cancer Association; AJCC: American Joint Committee on Cancer; CAP: College of American Pathologists.

current standard treatment for LAGC[1,2]. Although the benefit of this multimodality treatment was first confirmed by the MAGIC trial in 2006, the use of NACT had been adopted in GC for 30 years[22,23]. To assess the treatment response, despite the widespread use of the TRG system for gastrointestinal tract tumors, the response rates are always poor in GC compared with esophageal or colorectal cancer^[5]. This might be due to the lack of chemoradiation and sensitive regimens in preoperative settings. Due to the currently limited preoperative therapies and the limited number of responsive patients, findings on the value of TRG prognostic systems in LAGC are varied. Additional complexities arise when the study contexts are based on different TRG systems, especially on the comparison between TRG and ypTNM systems as independent predictors of patients survival[24-29]. Becker et al[16] investigated 480 patients with LAGC undergoing surgical resection and found TRG 2-3 grade (10%-100% residual tumor) to be an independent risk factor for patient OS; this reinforced the efficacy of the Becker TRG system. Ikoma et al^[25] reviewed 356 LAGC patients receiving D0-D2 Lymphadenectomy following NACT or NACRT, finding that the residual tumor < 50% group was associated with a shorter OS but not as an independent predictor^[25]. And Derieux first proved the predictive value of the Mandard system in GC, observing a poorer OS and DFS in patients with a high proportion of residual cancer cells (Mandard TRG 4) and no response (Mandard TRG 5)[29].

Therefore, when verifying the prognostic value of the histological response, considerable work should be done on determining an optimal tumor response classification for GC. Currently, two major principles are common to these systems for grading tumor regression: (1) estimating residual tumor in relation to fibrotic changes, e.g., the Mandard, AJCC/CAP, and Dworak systems[17,18,30]; and (2) proportioning the residual tumor in relation to the previous tumor site, e.g., the Becker and JGCA system[16, Although both are semiquantitative principles, the use of different systems reveals great regional disparities. The estimation of residual tumor is considered to be easier than considering therapy-induced fibrosis by the majority of pathologists[4], which potentially means a better inter-rater consistency for the residual tumor percentage method[32,33]. Most recently, an international survey was conducted and summarized preferences for using various TRG systems in gastrointestinal cancer among 173 global pathologists[4]. According to the published results, the AJCC/CAP and Mandard systems were widely adopted in North America and Europe, respectively. However, the questionnaires from East Asia - one from Japan and the other from Korea accounted for only two of the 173 valid responses, with no input from China[34]; it is doubtful whether these two contributions could fully picture the three countries that





Figure 3 Kaplan-Meier curves for overall survival of five tumor regression grade systems. A: JGCA2017-tumor regression grade (TRG); B: JGCA-TRG; C: Becker-TRG; D: AJCC/CAP-TRG; E: Mandard; F: Rearranged cutoff values. *P* value stands for log-rank test.

account for approximately one-third of the worldwide GC population. Overall, global diversity leads to obstacles in the comparison of experiments using different standards.

A comparison of different histological response systems for GC was conducted by Zhu *et al*[28]. This study included 192 patients and found that five TRG systems - including Mandard, JGCA, AJCC/CAP, Becker, and China - were not independent predictors for patient survival. Although the predictive abilities of each system were





Figure 4 Kaplan-Meier curves for progression-free survival of five tumor regression grade systems. A: JGCA2017-tumor regression grade (TRG); B: JGCA-TRG; C: Becker-TRG; D: AJCC/CAP-TRG; E: Mandard; F: Rearranged cutoff values. *P* value stands for log-rank test.

not measured, the Mandard and JGCA systems were recommended due to their superior prognosis prediction abilities. This was because a higher hazard ratio was discovered in the "no response" patients. In JCOG1004-A, 173 patients who received surgery following NACT were stratified according to different residual tumor cutoff percentages of 10%, 33%, 50%, and 67%. The 10% cutoff was found to be the best predictor of survival for various pathological types[9]. While this 10% cutoff finding was remarkable and coincided with Becker's cutoff method (described above)[35],

whether the current five- or six-tier JGCA standard provided the optimal discrimination value was not further investigated.

In the present study, c-index analysis was used to compare the discrimination value of five TRG systems including the most recent JGCA2017-TRG system. Both the fivetier JGCA and the six-tier JGCA2017 systems scored significantly lower c-indexes than the four-tier AJCC/CAP and Becker systems. Because both the JGCA2017 and the JGCA have overlapped measuring spacing compared with the four-tier Becker system, the results of the present study indicated that five- or six-tier grading systems performed no better (and even worse) than four-tier systems in evaluating GC patients. The c-index comparison suggested that the four-tier Becker system had the best predictive value for GC patients. Because of their relatively wide measuring distance, four-tier systems based on residual percentages also mean a lower workload, easier understanding of protocols, and less inter-observer disagreement propagation[4, 36].

On the other hand, based on the JCGA2017 criteria, the present study revealed that grade 2b (1%-10%) was likely to predict longer OS and DFS than grade 0 (no response). Interestingly, when the percentages of residual tumor were reset to "no residual tumor", < 10%, < 100%, and "no response", the c-index of the rearranged cutoff values scored significantly higher than the Becker system for patient survival. Similar results using these revised cutoffs were reported by Zhu *et al*[28], wherein an overt higher HR was observed for grade 3 among the other JGCA grades, and by Becker et al^[35] who demonstrated the independent predictive ability of the Becker system by using a cutoff of < 10% residual tumor. The results of the present study suggested that among moderate-to-poor (residual tumor 10-99%) responders, the response rate may not have a decisive impact on hazard stratification because NACT or chemotherapy only accounted for a small part of improving the prognosis among significant covariates in this group of GC patients. Meanwhile, a complete or subtotal response (0%-10%) often indicated a fairly good sensitivity to chemotherapy and vice versa for non-responders (no regression), who cannot receive any benefit but toxicity. Although the non-responders only accounted for 6.3% of the total patient number, it is suggested that this "break off both ends" approach provides a way for screening chemosensitivity and predicting prognosis in GC patients. However, a larger sample size is required to verify this proposal.

There were some limitations to this study. First, it was restricted by its single-center retrospective nature. Second, although histopathology was performed by two pathologists with over 10 years of experience, analysis of the inter- and intra-observer variability of the actual TRG classification was not conducted. Third, despite the involvement of many covariates, the macroscopic information may not be sufficient. According to JCOG1004-A, the TRG cutoff standard may not be recommended for Bormann type IV patients, for which a current dataset is not available[9]. Furthermore, this study did not consider intestinal and diffuse types according to the Lauren classification, which are thought to be independent prognostic factors for survival[37]. Statistically, collinearity between the TRG and ypT categories is inevitable but would have affected the multivariable analysis results: the Pearson's coefficients with ypT were 0.619, 0.587, 0.662, and 0.639 for the JGCA2017, JGCA, Becker, and AJCC/CAP systems, respectively. To reduce the impact of multicollinearity, studies with an increased sample size are warranted.

CONCLUSION

In conclusion, it was demonstrated that although all five TRG systems could be used as independent predictors for LAGC patient survival, the six-tier JGCA-TRG system did not increase prognostic stratification but may reduce the reproducibility and increase the working load on histological response evaluation. Patient survival can be effectively discriminated by the Becker system using the residual tumor percentage rather than by estimating the fibrosis/residual tumor ratio. Apart from when using the Becker classification, the group of non-responders with no regression was predicted to have a poorer prognosis. A large population-based study is still required to find the optimal criteria and validate the boundary settings of current TRG systems for LAGC patients.

ARTICLE HIGHLIGHTS

Research background

The tumor regression grade systems for gastric cancer (GC) are various, while the most suitable one is yet to be known.

Research motivation

We aimed to investigate the most accurate criteria for TRG in predicting patient's prognosis.

Research objectives

To collect 413 locally advanced GC (LAGC) patient's clinical data and their posttreatment pathological samples after neoadjuvant chemotherapy treatment.

Research methods

This is a retrospectively clinical study in which the LAGC patient's specimens were reviewed by two pathologists and the TRG grades were revalued. Then, the predictive abilities of five TRG criteria were assessed and statistically compared based on survival/risk prediction model.

Research results

The four-tier Becker system showed the highest predictive ability, among the five common TRG criteria. The TRG criteria could achieve an optimal prediction when the residual tumor percentages were reset as: "no residual tumor", < 10%, < 100%, and "no response".

Research conclusions

The four-tier Becker system is more suitable and should be recommended for LAGC patients.

Research perspectives

A population-based study is warranted to define the optimal criterion for TRG for GC.

REFERENCES

- Cunningham D, Allum WH, Stenning SP, Thompson JN, Van de Velde CJ, Nicolson M, Scarffe JH, 1 Lofts FJ, Falk SJ, Iveson TJ, Smith DB, Langley RE, Verma M, Weeden S, Chua YJ, MAGIC Trial Participants. Perioperative chemotherapy versus surgery alone for resectable gastroesophageal cancer. N Engl J Med 2006; 355: 11-20 [PMID: 16822992 DOI: 10.1056/NEJMoa055531]
- Ychou M, Boige V, Pignon JP, Conroy T, Bouché O, Lebreton G, Ducourtieux M, Bedenne L, Fabre JM, Saint-Aubert B, Genève J, Lasser P, Rougier P. Perioperative chemotherapy compared with surgery alone for resectable gastroesophageal adenocarcinoma: an FNCLCC and FFCD multicenter phase III trial. J Clin Oncol 2011; 29: 1715-1721 [PMID: 21444866 DOI: 10.1200/JCO.2010.33.0597]
- 3 Wang FH, Shen L, Li J, Zhou ZW, Liang H, Zhang XT, Tang L, Xin Y, Jin J, Zhang YJ, Yuan XL, Liu TS, Li GX, Wu Q, Xu HM, Ji JF, Li YF, Wang X, Yu S, Liu H, Guan WL, Xu RH. The Chinese Society of Clinical Oncology (CSCO): clinical guidelines for the diagnosis and treatment of gastric cancer. Cancer Commun (Lond) 2019; 39: 10 [PMID: 30885279 DOI: 10.1186/s40880-019-0349-9]
- 4 Westerhoff M, Osecky M, Langer R. Varying practices in tumor regression grading of gastrointestinal carcinomas after neoadjuvant therapy: results of an international survey. Mod Pathol 2020; 33: 676-689 [PMID: 31673084 DOI: 10.1038/s41379-019-0393-7]
- Fanelli GN, Loupakis F, Smyth E, Scarpa M, Lonardi S, Pucciarelli S, Munari G, Rugge M, Valeri N, 5 Fassan M. Pathological Tumor Regression Grade Classifications in Gastrointestinal Cancers: Role on Patients' Prognosis. Int J Surg Pathol 2019; 27: 816-835 [PMID: 31416371 DOI: 10.1177/1066896919869477]
- 6 Tong Y, Liu D, Zhang J. Connection and distinction of tumor regression grading systems of gastrointestinal cancer. Pathol Res Pract 2020; 216: 153073 [PMID: 32825946 DOI: 10.1016/j.prp.2020.153073]
- 7 Japanese Gastric Cancer Association. Japanese classification of gastric carcinoma: 3rd English edition. Gastric Cancer 2011; 14: 101-112 [PMID: 21573743 DOI: 10.1007/s10120-011-0041-5]
- Association JGC. Japanese Classification of Gastric Carcinoma, the 15th Edition (in Japanese). 8 Tokyo: Kanehara Shuppan, 2017: 43-45 [DOI: 10.1007/pl00011681]
- Nakamura K, Kuwata T, Shimoda T, Mizusawa J, Katayama H, Kushima R, Taniguchi H, Sano T, Sasako M, Fukuda H. Determination of the optimal cutoff percentage of residual tumors to define the



pathological response rate for gastric cancer treated with preoperative therapy (JCOG1004-A). Gastric Cancer 2015; 18: 597-604 [PMID: 24968818 DOI: 10.1007/s10120-014-0401-z]

- 10 Liang H. Reports of plenary session from the 89th Annual Meeting of the Japanese Gastric Cancer Association. Zhongguo Shiyong Waike Zazhi 2017; 37: 390-393 [DOI: 10.1111/j.1365-2133.2010.09814.x]
- 11 Wang FH, Zhang XT, Li YF, Tang L, Qu XJ, Ying JE, Zhang J, Sun LY, Lin RB, Qiu H, Wang C, Qiu MZ, Cai MY, Wu Q, Liu H, Guan WL, Zhou AP, Zhang YJ, Liu TS, Bi F, Yuan XL, Rao SX, Xin Y, Sheng WQ, Xu HM, Li GX, Ji JF, Zhou ZW, Liang H, Zhang YQ, Jin J, Shen L, Li J, Xu RH. The Chinese Society of Clinical Oncology (CSCO): Clinical guidelines for the diagnosis and treatment of gastric cancer, 2021. Cancer Commun (Lond) 2021; 41: 747-795. [PMID: 34197702 DOI: 10.1002/cac2.12193]
- 12 Liu Z, Wang Y, Shan F, Ying X, Zhang Y, Li S, Jia Y, Li Z, Ji J. 5-Fu-Based Doublet Regimen in Patients Receiving Perioperative or Postoperative Chemotherapy for Locally Advanced Gastric Cancer: When to Start and How Long Should the Regimen Last? Cancer Manag Res 2021; 13: 147-161 [PMID: 33469359 DOI: 10.2147/CMAR.S285361]
- Japanese Gastric Cancer Association. Japanese gastric cancer treatment guidelines 2014 (ver. 4). 13 Gastric Cancer 2017; 20: 1-19 [PMID: 27342689 DOI: 10.1007/s10120-016-0622-4]
- In H, Solsky I, Palis B, Langdon-Embry M, Ajani J, Sano T. Validation of the 8th Edition of the 14 AJCC TNM Staging System for Gastric Cancer using the National Cancer Database. Ann Surg Oncol 2017; 24: 3683-3691 [PMID: 28895113 DOI: 10.1245/s10434-017-6078-x]
- Hu B, El Hajj N, Sittler S, Lammert N, Barnes R, Meloni-Ehrig A. Gastric cancer: Classification, histology and application of molecular pathology. J Gastrointest Oncol 2012; 3: 251-261 [PMID: 22943016 DOI: 10.3978/j.issn.2078-6891.2012.021]
- 16 Becker K, Mueller JD, Schulmacher C, Ott K, Fink U, Busch R, Böttcher K, Siewert JR, Höfler H. Histomorphology and grading of regression in gastric carcinoma treated with neoadjuvant chemotherapy. Cancer 2003; 98: 1521-1530 [PMID: 14508841 DOI: 10.1002/cncr.11660]
- 17 Mandard AM, Dalibard F, Mandard JC, Marnay J, Henry-Amar M, Petiot JF, Roussel A, Jacob JH, Segol P, Samama G. Pathologic assessment of tumor regression after preoperative chemoradiotherapy of esophageal carcinoma. Clinicopathologic correlations. Cancer 1994; 73: 2680-2686 [PMID: 8194005 DOI: 10.1002/1097-0142(19940601)73:11<2680::aid-cncr2820731105>3.0.co;2-c]
- Chen HY, Feng LL, Li M, Ju HQ, Ding Y, Lan M, Song SM, Han WD, Yu L, Wei MB, Pang XL, He 18 F, Liu S, Zheng J, Ma Y, Lin CY, Lan P, Huang MJ, Zou YF, Yang ZL, Wang T, Lang JY, Orangio GR, Poylin V, Ajani JA, Wang WH, Wan XB. College of American Pathologists Tumor Regression Grading System for Long-term Outcome in Patients with Locally Advanced Rectal Cancer. Oncologist 2021 [DOI: 10.1002/onco.13707]
- Clavien PA, Barkun J, de Oliveira ML, Vauthey JN, Dindo D, Schulick RD, de Santibañes E, Pekolj 19 J, Slankamenac K, Bassi C, Graf R, Vonlanthen R, Padbury R, Cameron JL, Makuuchi M. The Clavien-Dindo classification of surgical complications: five-year experience. Ann Surg 2009; 250: 187-196 [PMID: 19638912 DOI: 10.1097/SLA.0b013e3181b13ca2]
- 20 Li Z, Wang Y, Shan F, Ying X, Wu Z, Xue K, Miao R, Zhang Y, Ji J. ypTNM staging after neoadjuvant chemotherapy in the Chinese gastric cancer population: an evaluation on the prognostic value of the AJCC eighth edition cancer staging system. Gastric Cancer 2018; 21: 977-987 [PMID: 29748876 DOI: 10.1007/s10120-018-0830-1]
- Schröder MS, Culhane AC, Quackenbush J, Haibe-Kains B. survcomp: an R/Bioconductor package 21 for performance assessment and comparison of survival models. Bioinformatics 2011; 27: 3206-3208 [PMID: 21903630 DOI: 10.1093/bioinformatics/btr511]
- 22 Reddavid R, Sofia S, Chiaro P, Colli F, Trapani R, Esposito L, Solej M, Degiuli M. Neoadjuvant chemotherapy for gastric cancer. Is it a must or a fake? World J Gastroenterol 2018; 24: 274-289 [PMID: 29375213 DOI: 10.3748/wjg.v24.i2.274]
- Songun I, Keizer HJ, Hermans J, Klementschitsch P, de Vries JE, Wils JA, van der Bijl J, van 23 Krieken JH, van de Velde CJ. Chemotherapy for operable gastric cancer: results of the Dutch randomised FAMTX trial. The Dutch Gastric Cancer Group (DGCG). Eur J Cancer 1999; 35: 558-562 [PMID: 10492627 DOI: 10.1016/s0959-8049(98)00429-8]
- Stark AP, Estrella JS, Chiang YJ, Das P, Minsky BD, Blum Murphy MA, Ajani JA, Mansfield P, 24 Badgwell BD, Ikoma N. Impact of tumor regression grade on recurrence after preoperative chemoradiation and gastrectomy for gastric cancer. J Surg Oncol 2020; 122: 422-432 [PMID: 32462681 DOI: 10.1002/jso.25984]
- Ikoma N, Estrella JS, Blum Murphy M, Das P, Minsky BD, Mansfield P, Ajani JA, Badgwell BD. 25 Tumor Regression Grade in Gastric Cancer After Preoperative Therapy. J Gastrointest Surg 2020 [DOI: 10.1007/s11605-020-04688-2]
- Blackham AU, Greenleaf E, Yamamoto M, Hollenbeak C, Gusani N, Coppola D, Pimiento JM, 26 Wong J. Tumor regression grade in gastric cancer: Predictors and impact on outcome. J Surg Oncol 2016; 114: 434-439 [PMID: 27199217 DOI: 10.1002/jso.24307]
- 27 Xu X, Zheng G, Zhang T, Zhao Y, Zheng Z. Is pathologic tumor regression grade after neo-adjuvant chemotherapy a promising prognostic indicator for patients with locally advanced gastric cancer? Cancer Chemother Pharmacol 2019; 84: 635-646 [PMID: 31230156 DOI: 10.1007/s00280-019-03893-4]
- Zhu Y, Sun Y, Hu S, Jiang Y, Yue J, Xue X, Yang L, Xue L. Comparison of five tumor regression grading systems for gastric adenocarcinoma after neoadjuvant chemotherapy: a retrospective study of



192 cases from National Cancer Center in China. BMC Gastroenterol 2017; 17: 41 [PMID: 28292272 DOI: 10.1186/s12876-017-0598-5]

- 29 Derieux S, Svrcek M, Manela S, Lagorce-Pages C, Berger A, André T, Taieb J, Paye F, Voron T. Evaluation of the prognostic impact of pathologic response to preoperative chemotherapy using Mandard's Tumor Regression Grade (TRG) in gastric adenocarcinoma. Dig Liver Dis 2020; 52: 107-114 [PMID: 31427088 DOI: 10.1016/j.dld.2019.07.010]
- Dworak O, Keilholz L, Hoffmann A. Pathological features of rectal cancer after preoperative 30 radiochemotherapy. Int J Colorectal Dis 1997; 12: 19-23 [PMID: 9112145 DOI: 10.1007/s003840050072]
- 31 Ninomiya Y, Yanagisawa A, Kato Y, Kitagawa T, Ishihara S, Nakajima T. Histological indications of a favorable prognosis with far-advanced gastric carcinomas after preoperative chemotherapy. J Cancer Res Clin Oncol 1999; 125: 699-706 [PMID: 10592104 DOI: 10.1007/s004320050337]
- 32 Karamitopoulou E, Thies S, Zlobec I, Ott K, Feith M, Slotta-Huspenina J, Lordick F, Becker K, Langer R. Assessment of tumor regression of esophageal adenocarcinomas after neoadjuvant chemotherapy: comparison of 2 commonly used scoring approaches. Am J Surg Pathol 2014; 38: 1551-1556 [PMID: 25140894 DOI: 10.1097/PAS.00000000000255]
- Puetz K, Bollschweiler E, Semrau R, Mönig SP, Hölscher AH, Drebber U. Neoadjuvant 33 chemoradiation for patients with advanced oesophageal cancer - which response grading system best impacts prognostic discrimination? Histopathology 2019; 74: 731-743 [PMID: 30636069 DOI: 10.1111/his.13811]
- **Bray F**, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- 35 Becker K, Langer R, Reim D, Novotny A, Meyer zum Buschenfelde C, Engel J, Friess H, Hofler H. Significance of histopathological tumor regression after neoadjuvant chemotherapy in gastric adenocarcinomas: a summary of 480 cases. Ann Surg 2011; 253: 934-939 [PMID: 21490451 DOI: 10.1097/SLA.0b013e318216f449
- Tong Y, Zhu Y, Zhao Y, Shan Z, Liu D, Zhang J. Evaluation and Comparison of Predictive Value of 36 Tumor Regression Grades according to Mandard and Becker in Locally Advanced Gastric Adenocarcinoma. Cancer Res Treat 2021; 53: 112-122 [DOI: 10.4143/crt.2020.516]
- 37 Reim D, Gertler R, Novotny A, Becker K, zum Büschenfelde CM, Ebert M, Dobritz M, Langer R, Hoefler H, Friess H, Schumacher C. Adenocarcinomas of the esophagogastric junction are more likely to respond to preoperative chemotherapy than distal gastric cancer. Ann Surg Oncol 2012; 19: 2108-2118 [PMID: 22130620 DOI: 10.1245/s10434-011-2147-8]



0 WŰ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2180-2189

DOI: 10.4251/wjgo.v13.i12.2180

ISSN 1948-5204 (online)

ORIGINAL ARTICLE

Retrospective Study Clinical features of intracerebral hemorrhage in patients with colorectal cancer and its underlying pathogenesis

Xu-Hui Deng, Jing Li, Shi-Jian Chen, Yi-Ju Xie, Jian Zhang, Geng-Yu Cen, Yi-Ting Song, Zhi-Jian Liang

ORCID number: Xu-Hui Deng 0000-0003-4231-7025; Jing Li 0000-0002-9867-2897; Shi-Jian Chen 0000-0001-6547-3608; Yi-Ju Xie 0000-0001-6447-3745; Jian Zhang 0000-0002-0658-8277; Geng-Yu Cen 0000-0003-3609-6633; Yi-Ting Song 0000-0003-3861-7826; Zhi-Jian Liang 0000-0003-3339-2568.

Author contributions: Deng XH

and Liang ZJ conceived and designed the study; Deng XH collected the data and drafted the manuscript; Li J, Chen SJ, Xie YJ, Zhang J, Cen GY and Song YT helped to collect the data; Li J and Chen SJ analyzed the results; Liang ZJ critically revised the manuscript and provided financial support for this work; all the authors have read the manuscript and approved the final version.

Institutional review board

statement: This study was reviewed and approved by the Ethics Committee of the First Affiliated Hospital of Guangxi Medical University.

Informed consent statement:

Patients were not required to give informed consent to the study because the analysis used anonymous clinical data that were obtained after each patient agreed to treatment by written consent. For full disclosure, the details of

Xu-Hui Deng, Shi-Jian Chen, Yi-Ju Xie, Jian Zhang, Geng-Yu Cen, Yi-Ting Song, Zhi-Jian Liang, Department of Neurology, The First Affiliated Hospital of Guangxi Medical University, Nanning 530021, Guangxi Zhuang Autonomous Region, China

Xu-Hui Deng, Department of Neurology, The Affiliated Yuebei People's Hospital of Shantou University Medical College, Shaoguan 512025, Guangdong Province, China

Jing Li, Department of General Medicine, The Affiliated Cancer Center of Guangxi Medical University, Nanning 530021, Guangxi Zhuang Autonomous Region, China

Corresponding author: Zhi-Jian Liang, MD, PhD, Professor, Department of Neurology, The First Affiliated Hospital of Guangxi Medical University, No. 22 Shuangyong Road, Nanning 530021, Guangxi Zhuang Autonomous Region, China. 1zj200415@126.com

Abstract

BACKGROUND

The diagnosis of both cancer and intracerebral hemorrhage (ICH) in the same patient is not uncommon, but the clinical features and pathogenesis of patients with colorectal cancer (CRC) and ICH are still not well known.

AIM

To investigate the clinical features and underlying pathogenesis of ICH in patients with CRC.

METHODS

A retrospective review of CRC patients complicated with ICH from three centers between January 2014 and December 2020 was performed. Clinical data such as laboratory examinations, imaging features, prognosis, and underlying pathogenesis were analyzed.

RESULTS

Of 16673 identified CRC patients, 20 (0.12%) suffered from ICH. There were 13 males and 7 females, with an average age (mean \pm SD) of 68.45 \pm 10.66 years. Fourteen patients (70%) had distant metastases and most patients (85%) showed an elevation of one or more cancer biomarkers. The hemorrhagic lesions in 13 patients (65%) were in the intracerebral lobe. Four patients were completely dependent and 4 died within 30 days after hemorrhage. Intratumoral hemorrhage (50%) and coagulopathy (50%) accounted for the majority of hemorrhages.



the study are published on the home page of the First Affiliated Hospital of Guangxi Medical University.

Conflict-of-interest statement: All authors have no conflict of interests.

Data sharing statement: No additional data are available.

Supported by The Foundation of Prevention and Control of Chronic Diseases in Central-South China (Guangxi), No. 2018YFC1311305; and the Foundation of Science and Technology Plan Projects of Qingxiu District of Nanning, No. 2020043.

Country/Territory of origin: China

Specialty type: Gastroenterology and hepatology

Provenance and peer review:

Unsolicited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: July 5, 2021 Peer-review started: July 5, 2021 First decision: July 29, 2021 Revised: August 10, 2021 Accepted: September 15, 2021 Article in press: September 15, 2021

CONCLUSION

Patients with ICH and CRC often have clinical features with lobar hemorrhage, distant metastases and poor prognosis. Intratumoral hemorrhage and coagulopathy are the main causes of ICH in patients with CRC.

Key Words: Colorectal cancer; Intracerebral hemorrhage; Clinical features; Pathogenesis

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: The association between cancer and intracerebral hemorrhage (ICH) has long been studied, however little attention has been paid to the hemorrhagic cerebrovascular events in patients with colorectal cancer (CRC). CRC has been reported to increase the risk of ICH. The present study retrospectively analyzed the clinical data of patients with CRC and ICH, and indicated that intratumoral hemorrhage and coagulopathy were the main causes of ICH in CRC patients.

Citation: Deng XH, Li J, Chen SJ, Xie YJ, Zhang J, Cen GY, Song YT, Liang ZJ. Clinical features of intracerebral hemorrhage in patients with colorectal cancer and its underlying pathogenesis. World J Gastrointest Oncol 2021; 13(12): 2180-2189

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2180.htm

DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2180

INTRODUCTION

Intracerebral hemorrhage (ICH) and cancer are both common disorders, and are significant causes of morbidity and mortality worldwide in the elderly. It is reported that ICH accounts for almost half of cerebrovascular events in cancer patients[1,2] and significantly aggravates their condition and prognosis[3,4]. Several population-based studies have demonstrated that multiple cancers are associated with an increased risk of ICH[5,6]. Despite the fact that traditional vascular risk factors for ICH are commonly observed in cancer patients, current studies have revealed that ICH in some patients shows clear-cut distinctions by its unique characteristics and pathogenesis. ICH can manifest as direct or indirect effects of cancer, namely cancer-related ICH. Cancer-related ICH might result from primary or metastatic brain malignancies, coagulopathy or systemic effects of oncological therapy[1,7,8]. However, it may lead to a variety of clinical features and pathogenesis of cancer-related ICH due to the complexity and diversity of cancers.

Colorectal cancer (CRC) is the third most frequently diagnosed cancer in the world [9,10]. It has been estimated that CRC burden rose to approximately 1.9 million new cases and 0.9 million deaths worldwide in 2020[10]. However, there are few detailed reports in the literature that focus on the hemorrhagic cerebrovascular events in CRC patients. The purpose of this study was to investigate the clinical features and underlying pathogenesis of ICH in patients with CRC.

MATERIALS AND METHODS

The present study was reviewed and approved by the Ethics Committee of the First Affiliated Hospital of Guangxi Medical University. CRC patients with ICH were recruited from the First Affiliated Hospital and the Affiliated Cancer Center of Guangxi Medical University, and the Affiliated Yuebei People's Hospital of Shantou University Medical College between January 2014 and December 2020. The diagnostic criteria for ICH were based on the 2015 Guidelines for the Diagnosis and Treatment from the American Heart Association[11]. The diagnosis of active CRC followed the definition of active cancer in the study by Lee et al[12]. Patients who met the following criteria were included: (1) Diagnosis of CRC within 6 mo before enrollment, any treatment for CRC within the previous 6 mo, or recurrent or metastatic CRC; (2) Presence of clinical symptoms, such as sudden onset of unconsciousness, headache, hemiplegic paralysis, slurred speech, or other focal neurological deficits; and (3) The



Published online: December 15, 2021

P-Reviewer: Emran TB S-Editor: Yan JP L-Editor: A P-Editor: Yuan YY



presence of ICH on intracranial computed tomography (CT) or magnetic resonance imaging (MRI) scan and susceptibility-weighted imaging, which could explain the symptoms. The exclusion criteria were as follows: (1) Combined with other systemic malignancy; (2) Presence of cerebral infarction and other central nervous system complications; (3) CRC diagnosed > 5 years ago, with no evidence of recurrence or metastasis; and (4) Patients with incomplete records (Figure 1). The selected cases were reviewed and determined by a panel which consisted of an oncologist, a neurologist and a neuroradiologist who were all blind to the study.

Collection of clinical data

General demographic data such as age and gender were obtained. Vascular risk factors for ICH including hypertension, diabetes mellitus, hypercholesterolemia, tobacco and alcohol consumption, aneurysm, arteriovenous malformation and stroke history were also documented. Moreover, data on CRC, such as pathological types of cancer cells, metastasis, prior and current treatment and information related to acute ICH, including onset form, cardinal symptoms and signs, and hemorrhagic locations were recorded. In addition, routine blood examination, blood biochemistry, coagulation indices, plasma D-dimer, cancer biomarkers, electrocardiography, Doppler echocardiography, cervical vascular Doppler, head and neck CT angiography, head CT, head MRI and magnetic resonance angiography were also carried out. The National Institute of Health Stroke Scale (NIHSS) was used to evaluate the severity of focal neurological deficits. To minimize the effects of CRC progression on physical activities, patients' functional prognosis on the 30th day after hemorrhage was measured using the modified Rankin Scale (mRS), and a mRS score > 3 was regarded as a poor prognosis[13]. According to a research study by Navi and colleagues on ICH [2], coagulopathy was identified if any of the following parameters were fulfilled: platelets $< 100 \times 10^{\circ}/L$, international normalized ratio (INR) > 1.5, activated partial thromboplastin time (APTT) > 45 s, prothrombin time (PT) > 15 s, and disseminated intravascular coagulation (DIC) (fibrinogen < 200 mg/dL and D-dimer > 290 ng/dL).

Statistical analysis

All statistical analyses were undertaken using SPSS 20.0 software (IBM, Inc., Armonk, NY, United States). Quantitative data were shown as mean ± SD and qualitative data were expressed as frequency and percentages.

RESULTS

A total of 16673 patients with CRC were identified, and 20 patients (0.12%) with ICH met the inclusion criteria. Of these 20 patients, 13 were male (65%) and 7 were female (35%). The average age (mean \pm SD) was 68.45 \pm 10.66 years (range 47-82). All patients were pathologically confirmed to have adenocarcinoma. Ten patients had vascular risk factors, including tobacco use (30%), hypertension (30%), diabetes mellitus (10%), alcohol abuse (5%), hypercholesterolemia (15%), stroke history (10%) and coronary or kidney disease (5%). No aneurysms or arteriovenous malformations were identified in these patients. None of the patients were on therapeutic anticoagulation or antiplatelet agents at the time of hemorrhage. Coagulopathy was observed in 10 patients. Thrombocytopenia was present in 6 patients (30%). Six patients (30%) had a prolonged PT value, 2 (10%) had a prolonged APPT value, and 3 (15%) displayed INR values greater than 1.5. DIC was documented in 1 patient. Most patients (17/20, 85%) had at least one elevated cancer biomarker. Six patients (30%) showed hepatic dysfunction, while 14 patients (70%) did not. Fourteen patients (70%) exhibited distant metastases, intracranial metastasis occurred in 10 patients (50%) and hepatic/osseous metastases occurred in 8 patients (40%), when ICH developed. Prior to hemorrhage, 13 patients (65%) had received oncological therapy while 7 patients (35%) had not (Table 1).

ICH occurred in 2 patients (10%) before cancer diagnosis and 18 patients (90%) after cancer diagnosis. The most frequent hemorrhagic lesion was in the cerebral lobe, occurring in 13 patients (65%). Fewer lesions were found in the basal ganglia (4/20, 20%), cerebellum (2/20, 10%), and brainstem (1/20, 5%). The etiologies of hemorrhage were ascribed to intratumoral hemorrhage (10/20, 50%), coagulopathy (10/20, 50%), both intratumoral hemorrhage and coagulopathy (5/20, 25%), hypertension (4/20, 20%), and trauma (1/20, 5%). Fifteen patients (75%) received hemorrhage targeted treatment and 5 (25%) withdrew treatment. The mean NIHSS score was 10.25 ± 7.59 (range 2-25) on the day of hemorrhage onset. On the 30th day after hemorrhage, 7 patients (35%) were completely independent, 5 (25%) were partially independent, 4



Table 1 Demographics of colorectal cancer patients with intracerebral hemorrhage		
Characteristics	mean ± SD/ <i>n</i> (%)	
Age, yr	68.45 ± 10.66	
Gender		
Male	13 (65)	
Female	7 (35)	
Vascular risk factors	10 (50)	
Tobacco	6 (30)	
Hypertension	6 (30)	
Diabetes mellitus	2 (10)	
Hypercholesterolemia	3 (15)	
Alcohol abuse	1 (5)	
Stroke history	2 (10)	
Coronary or kidney disease	1 (5)	
Coagulopathy	10 (50)	
$PLT < 100 \times 10^{9}/L$	6 (30)	
PT > 15 s	6 (30)	
APTT > 45 s	2 (10)	
INR > 1.5	3 (15)	
DIC	1 (5)	
Elevated cancer biomarkers	17 (85)	
CEA > 5 ng/mL	16 (80)	
CA125 > 35 U/mL	4 (20)	
CA153 > 31.3 U/mL	2 (10)	
CA199 > 37 U/mL	9 (45)	
Hepatic dysfunction		
Yes	6 (30)	
No	14 (70)	
Distant metastasis		
Intracranial metastasis	10 (50)	
Hepatic/osseous metastasis	8 (40)	
Metastasis of other organs	7 (35)	
None	6 (30)	
Cancer treatment before hemorrhage		
Surgery alone	2 (10)	
Chemotherapy	3 (15)	
Both of the above	8 (40)	
No treatment	7 (35)	

PLT: Platelet; PT: Prothrombin time; APTT: Activated partial thromboplastin time; INR: International normalized ratio; DIC: Disseminated intravascular coagulation; CEA: Carcinoembryonic antigen; CA125: Carbohydrate antigen 125; CA153: Carbohydrate antigen 153; CA199: Carbohydrate antigen 199.

> (20%) were completely dependent, and 4 (20%) died (Table 2). Among the 4 patients who died, the cause of death was fatal ICH induced by DIC in one patient, brain metastasis and intratumoral hemorrhage in one patient, and aspiration pneumonia in

Saisbideng® WJGO | https://www.wjgnet.com

Table 2 Data on intracerebral hemorrhage			
Characteristics	n (%)		
Location of hemorrhage			
Lobe	13 (65)		
Basal ganglia	4 (20)		
Brainstem	1 (5)		
Cerebellum	2 (10)		
Intratumoral hemorrhage	10 (50)		
Treatment of hemorrhage			
Conservative treatment	14 (70)		
Surgery	1 (5)		
Withdrawal of treatment	5 (25)		
NIHSS score on the day of hemorrhage onset			
NIHSS (0-5)	7 (35)		
NIHSS (6-10)	5 (25)		
NIHSS (11-20)	3 (15)		
NIHSS (> 20)	5 (25)		
mRS score 30 d after hemorrhage			
Completely independent (mRS = 0-1)	7 (35)		
Partially independent (mRS = 2-3)	5 (25)		
Completely dependent (mRS = 4-5)	4 (20)		
Death (mRS = 6)	4 (20)		
Etiology			
Intratumoral hemorrhage	10 (50)		
Coagulopathy	10 (50)		
Both of the above	5 (25)		
Hypertension	4 (20)		
Trauma	1 (5)		
Time interval between CRC diagnosis and hemorrhage			
ICH onset before cancer diagnosis	2 (10)		
ICH onset after cancer diagnosis			
< 6 mo	8 (40)		
6-12 mo	5 (25)		
1-5 yr	4 (20)		
5-10 yr	1 (5)		

ICH: Intracerebral hemorrhage; CRC: Colorectal cancer; NIHSS: National Institute of Health Stroke Scale; mRS: Modified Rankin Scale.

the other two patients (Figure 2).

DISCUSSION

ICH is a well-known potential complication of cancer[1,2]. A nationwide follow-up study from Sweden reported that compared with a non-cancer population, the overall risk of hemorrhage after diagnosis of cancer within 6 mo, 6 mo to 1 year and 1 year to



Baishideng® WJGO https://www.wjgnet.com

CRC patients registered December 2020 (With his	at 3 centers (<i>n</i> = 16673) between January 2014 and tological evidence and diagnosed by oncologist expert)
Excluded (<i>n</i> = 16518)) Patients without ICH
CRC patients with ICH (n = 155)
Excluded (<i>n</i> = 135)	Not an active colorectal cancer Combined with other systemic malignancy Presence of cerebral infarction and other central nervous system complications Patients with incomplete records
Patients with active CRC	and ICH $(n = 20)$

Figure 1 Patient enrollment flowchart. CRC: Colorectal cancer; ICH: Intracerebral hemorrhage.



Figure 2 Neuroimaging findings. A-F: Neuroimages of a 76-year-old patient with active colorectal cancer (CRC). Brain computed tomography axial views, showing a hemorrhagic lesion in the left parietal lobe (A-C); brain enhanced magnetic resonance views, showing a metastatic tumor in the same location (D-F); G-I: Neuroimages of a 65-year-old patient with active CRC and disseminated intravascular coagulation, showing a massive hemorrhage in the left temporal and parietal lobe.

5 years was 2.2, 1.4 and 1.3, respectively[6], indicating that cancer could increase the incidence of ICH and that cancer-related ICH theoretically exists. Patients with CRC also had a significantly increased risk of hemorrhage[6]. CRC-related ICH may also theoretically exist. In this study, although only 0.12% of CRC patients developed ICH,

Gaisbideng® WJGO | https://www.wjgnet.com

the incidence of hemorrhage was still much higher than that in the general population. Traditional pathogenesis were observed in the etiology of ICH in CRC patients, but more attention should be paid to the direct or indirect role of CRC itself.

In a retrospective review, cancer patients with ICH were more likely to present with coagulopathy, a lobar hemorrhage location, and poor short-term prognosis when compared with non-cancer patients [14]. In the present study, hemorrhagic lesions were more frequent in the cerebral lobe, occurring in 13 of 20 patients, and the shortterm prognosis was poor, as disability and death occurred in 8 of these 20 patients within 30 d after hemorrhage onset. Coagulopathy was identified in half of the patients. In some respects, the results of this study were consistent with the research stated above. In addition, most patients developed distant metastases and had significantly elevated cancer biomarkers when ICH occurred, which was another characteristic in CRC patients with ICH.

Although ICH is a late complication in most cancer patients, it may precede cancer diagnosis. It is worth noting that 2 patients were newly diagnosed with CRC during hospitalization for ICH, suggesting that ICH might be the first manifestation of CRC. Cancers with ICH as the initial symptom include choriocarcinoma, leukocythemia, and medulloblastoma[15-17]. How to quickly identify insidious cancer in patients with ICH is still a challenge. Therefore, it is necessary to conduct cancer associated examinations, including CRC associated examinations, in patients with unexplained hemorrhage.

ICH in patients with cancer often arises from unique mechanisms which are uncommon in the general population. A clinical series of 208 cancer patients with intracranial hemorrhage found that intratumoral hemorrhage and coagulopathy were the main causes, and that hypertension, the most common cause of ICH in the population, accounted for only a small proportion[2]. In the present study, ICH was caused by hypertension in a small number of patients but was mostly caused by intratumoral hemorrhage and coagulopathy, which was similar to the above results, indicating that intratumoral hemorrhage and coagulopathy might be the main pathogenesis of ICH in patients with CRC.

Pathophysiologically, factors favoring intratumoral hemorrhage include tumor necrosis, aberrant neovascularization, vascular infiltration with rupture, imbalances in the fibrinolytic cascade, and overexpression of vascular endothelial growth factor (VEGF) and metalloproteinases[1-2,18,19]. Tumor cells can directly invade blood vessels and destroy their integrity, resulting in vascular rupture. Overexpression of VEGF in tumor cells can stimulate neovascularization and increase microvessel density. However, the fine structure of newly formed blood vessels induced by VEGF tends to have high permeability and fragility leading to hemorrhagic events[19]. Abnormal high expression of metalloproteinases can degrade extracellular matrix proteins which maintain basement membrane structural/functional integrity, and cause considerable damage to capillary integrity leading to hemorrhage^[19]. Overexpression of VEGF in CRC cells has been confirmed, and anti-VEGF therapies have been proved to be effective for metastatic CRC[20,21]. The expression and activity of metalloproteinases are high in CRC cells[22], and matrix metalloproteinase inhibitors might be a new way of treating metastatic CRC.

Coagulopathy is typically related to a complex interplay of multiple mechanisms, including thrombocytopenia, coagulation factor abnormalities or both. The etiology of thrombocytopenia can be diverse and multifactorial in cancer patients. Systemic chemotherapy is the most common cause of bone marrow suppression and thrombocytopenia^[23]. In patients with solid tumors, thrombocytopenia is potentially associated with bone marrow metastasis of malignant cells and secondary myelofibrosis [24]. Metastatic malignant cells can interfere with the hematopoietic microenvironment and inhibit hematopoiesis. Myelofibrosis may be induced by a series of cytokines secreted by immune system cells as part of the inflammatory response to tumors. In addition, DIC, thrombotic thrombocytopenic purpura, immune-mediated thrombocytopenia and heparin-induced thrombocytopenia all contribute to increased platelet destruction in cancer patients^[24]. In this study, some patients had been treated with chemotherapy before hemorrhage, and hence it is reasonable to consider chemotherapy as a significant risk factor. Some patients had osseous metastases, and it was speculated that their thrombocytopenia was related to bone marrow metastasis. Abnormalities in coagulation factors generally result from hepatic dysfunction, vitamin K deficiency or DIC. Hepatic dysfunction in some patients may be related to a primary or metastatic liver tumor, and chemotherapy. The production of coagulation factors may be affected in patients with obvious hepatic dysfunction, which may lead to a deficiency in coagulation factors. Patients with advanced cancer often suffer from severe malnutrition due to insufficient food intake, and the shortage of multiple



nutrients, especially vitamin K, further aggravates their coagulopathy. DIC is the most serious form of coagulopathy. The combination of excessive consumption of coagulation factors and platelets and primary fibrinogenolysis account for the high risk of bleeding during the pathological process of DIC. It is reported that CRC can also present with DIC during treatment[25,26]. In the present study, some patients had liver metastases, obvious hepatic dysfunction and poor nutrition, which may have resulted in coagulation factor deficiency. One patient developed DIC during treatment, resulting in fatal ICH.

CA125, CA153, CA199 and CEA are widely used as common cancer biomarkers in the clinic, and are well known carcinoma mucins. Previous studies revealed that carcinoma mucins were overexpressed by malignant cells and were shown to play a multifaceted role in the initiation, progression, metastasis and subsequent colonization of multiple malignancies[27,28]. In a clinical study of lung cancer-related ICH, elevated plasma CEA and CA199 Levels were independent risk factors for ICH in patients with active lung cancer^[13]. The researchers hypothesized that elevated cancer markers could activate platelets and lead to increased platelets, hypercoagulability, and eventually thrombotic events in the early stage of cancer; however, in the later stage, elevated cancer markers could lead to decreased platelets due to consumption, coagulopathy, and finally hemorrhagic events. Significantly elevated cancer biomarkers were observed in most patients in this study, but the role of these biomarkers in ICH requires further research.

The pathogenesis of ICH induced by CRC is complex and has not been fully elucidated. To date, there is no feasible method to effectively prevent and treat ICH in patients with CRC. In view of the pathophysiological changes such as thrombocytopenia and abnormalities of coagulation factors, it is possible to take active measures, including increasing nutrition supply, and supplementing platelets and fresh plasma, to improve the coagulation state in the early stage. These measures may delay or reduce the occurrence of ICH in patients with CRC, but should be confirmed in future studies.

One of the limitations of this study is its retrospective nature; thus, information bias may exist. Although clinical data were collected from three centers, the total number of selected cases was relatively small. Prospective population-based cohort studies are warranted to better elucidate the clinical characteristics and pathogenesis of CRC patients with ICH.

CONCLUSION

Patients with ICH and CRC often have clinical features with lobar hemorrhage, distant metastases and poor prognosis. Intratumoral hemorrhage and coagulopathy are the main causes of ICH in patients with CRC. More clinical trials are needed to validate these findings in the future.

ARTICLE HIGHLIGHTS

Research background

Many studies have confirmed that cancer can increase the risk of intracerebral hemorrhage (ICH). However, most previous studies were conducted on multiple cancers, and few focused on a specific cancer. The clinical characteristics and mechanisms of ICH in colorectal cancer (CRC) patients have not been fully elucidated.

Research motivation

There are few reports on hemorrhagic cerebrovascular events in patients with CRC.

Research objectives

This retrospective study aimed to investigate the clinical features and underlying pathogenesis of ICH in patients with CRC.

Research methods

A retrospective review of 20 patients (13 males and 7 females) with CRC and ICH from three centers between January 2014 and December 2020 was conducted. The clinical data of the patients such as vascular risk factors, laboratory results, neuroimaging and



underlying pathogenesis were analyzed.

Research results

The average age (mean \pm SD) of the patients was 68.45 ± 10.66 years. Fourteen patients (70%) had distant metastases and most patients (85%) had an elevation of one or more cancer biomarkers. The hemorrhagic lesions in 13 patients (65%) were in the intracerebral lobe. Four patients were completely dependent and 4 died within 30 days after hemorrhage. Intratumoral hemorrhage (50%) and coagulopathy (50%) accounted for the majority of hemorrhages.

Research conclusions

Patients with ICH and CRC often have clinical features with lobar hemorrhage, distant metastases and poor prognosis. Intratumoral hemorrhage and coagulopathy are the main causes of ICH in patients with CRC.

Research perspectives

The detailed mechanism of ICH in CRC patients requires further elucidation. Prospective population-based studies are needed to confirm these findings in the future.

REFERENCES

- Velander AJ, DeAngelis LM, Navi BB. Intracranial hemorrhage in patients with cancer. Curr Atheroscler Rep 2012; 14: 373-381 [PMID: 22528522 DOI: 10.1007/s11883-012-0250-3]
- Navi BB, Reichman JS, Berlin D, Reiner AS, Panageas KS, Segal AZ, DeAngelis LM. Intracerebral and subarachnoid hemorrhage in patients with cancer. Neurology 2010; 74: 494-501 [PMID: 20142616 DOI: 10.1212/WNL.0b013e3181cef837]
- 3 Gon Y, Todo K, Mochizuki H, Sakaguchi M. Cancer is an independent predictor of poor outcomes in patients following intracerebral hemorrhage. Eur J Neurol 2018; 25: 128-134 [PMID: 28895254 DOI: 10.1111/ene.13456
- 4 Murthy SB, Shastri A, Merkler AE, Hanley DF, Ziai WC, Fink ME, Iadecola C, Kamel H, Navi BB. Intracerebral Hemorrhage Outcomes in Patients with Systemic Cancer. J Stroke Cerebrovasc Dis 2016; 25: 2918-2924 [PMID: 27569708 DOI: 10.1016/j.jstrokecerebrovasdis.2016.08.006]
- Navi BB, Reiner AS, Kamel H, Iadecola C, Elkind MS, Panageas KS, DeAngelis LM. Association between incident cancer and subsequent stroke. Ann Neurol 2015; 77: 291-300 [PMID: 25472885 DOI: 10.1002/ana.243251
- 6 Zöller B, Ji J, Sundquist J, Sundquist K. Risk of haemorrhagic and ischaemic stroke in patients with cancer: a nationwide follow-up study from Sweden. Eur J Cancer 2012; 48: 1875-1883 [PMID: 22296948 DOI: 10.1016/j.ejca.2012.01.005]
- 7 Rogers LR. Cerebrovascular complications in patients with cancer. Semin Neurol 2010; 30: 311-319 [PMID: 20577937 DOI: 10.1055/s-0030-1255224]
- 8 Nakahara T, Owaki Y, Kosaka T, Fukada J, Ichimura A, Jinzaki M. Fatal Intracranial Hemorrhage Due to Thrombocytopenia in a Patient With Castration-Resistant Prostate Cancer Showing Extensive Bone Uptake of Injected 223Ra Dichloride. Clin Nucl Med 2018; 43: 546-547 [PMID: 29742607 DOI: 10.1097/RLU.000000000002117]
- Johdi NA, Sukor NF. Colorectal Cancer Immunotherapy: Options and Strategies. Front Immunol 2020; 11: 1624 [PMID: 33042104 DOI: 10.3389/fimmu.2020.01624]
- 10 Xi Y, Xu P. Global colorectal cancer burden in 2020 and projections to 2040. Transl Oncol 2021; 14: 101174 [PMID: 34243011 DOI: 10.1016/j.tranon.2021.101174]
- 11 Hemphill JC 3rd, Greenberg SM, Anderson CS, Becker K, Bendok BR, Cushman M, Fung GL, Goldstein JN, Macdonald RL, Mitchell PH, Scott PA, Selim MH, Woo D; American Heart Association Stroke Council; Council on Cardiovascular and Stroke Nursing; Council on Clinical Cardiology. Guidelines for the Management of Spontaneous Intracerebral Hemorrhage: A Guideline for Healthcare Professionals From the American Heart Association/American Stroke Association. Stroke 2015; 46: 2032-2060 [PMID: 26022637 DOI: 10.1161/STR.00000000000069]
- Lee AY, Levine MN, Baker RI, Bowden C, Kakkar AK, Prins M, Rickles FR, Julian JA, Haley S, 12 Kovacs MJ, Gent M; Randomized Comparison of Low-Molecular-Weight Heparin versus Oral Anticoagulant Therapy for the Prevention of Recurrent Venous Thromboembolism in Patients with Cancer (CLOT) Investigators. Low-molecular-weight heparin versus a coumarin for the prevention of recurrent venous thromboembolism in patients with cancer. N Engl J Med 2003; 349: 146-153 [PMID: 12853587 DOI: 10.1056/NEJMoa025313]
- 13 Qin K, Chen Y, Long H, Chen J, Wang D, Chen L, Liang Z. The biomarkers and potential pathogenesis of lung cancer related cerebral hemorrhage. Medicine (Baltimore) 2019; 98: e15693 [PMID: 31096511 DOI: 10.1097/MD.000000000015693]
- 14 Zhang YY, Chan DK, Cordato D, Shen Q, Sheng AZ. Stroke risk factor, pattern and outcome in



patients with cancer. Acta Neurol Scand 2006; 114: 378-383 [PMID: 17083337 DOI: 10.1111/j.1600-0404.2006.00709.x]

- Menekse G, Gezercan Y, Demirturk P, Uysal I, Okten AI. Fatal cerebellar hemorrhage as an initial 15 presentation of medulloblastoma in a child. J Pediatr Neurosci 2015; 10: 287-289 [PMID: 26557180 DOI: 10.4103/1817-1745.165727]
- Zhou T, Zhang S, Qin Y, Zhu W. Intracerebral hemorrhage as initial presentation of metastatic 16 choriocarcinoma: A case report. Radiol Case Rep 2020; 15: 2335-2338 [PMID: 32994836 DOI: 10.1016/j.radcr.2020.09.012]
- 17 Wang H, Cao F, Li J, Sun K, Jin J, Wang M. Intracerebral Hemorrhage as the Initial Presentation of Chronic Myeloid Leukemia: A Case Report and Review of the Literature. Front Neurol 2020; 11: 571576 [PMID: 33193017 DOI: 10.3389/fneur.2020.571576]
- Katz JM, Segal AZ. Incidence and etiology of cerebrovascular disease in patients with malignancy. 18 Curr Atheroscler Rep 2005; 7: 280-288 [PMID: 15975321 DOI: 10.1007/s11883-005-0020-6]
- Jung S, Moon KS, Jung TY, Kim IY, Lee YH, Rhu HH, Sun HS, Jeong YI, Kim KK, Kang SS. 19 Possible pathophysiological role of vascular endothelial growth factor (VEGF) and matrix metalloproteinases (MMPs) in metastatic brain tumor-associated intracerebral hemorrhage. J Neurooncol 2006; 76: 257-263 [PMID: 16158215 DOI: 10.1007/s11060-005-6876-z]
- 20 Malki A, ElRuz RA, Gupta I, Allouch A, Vranic S, Al Moustafa AE. Molecular Mechanisms of Colon Cancer Progression and Metastasis: Recent Insights and Advancements. Int J Mol Sci 2020; 22 [PMID: 33374459 DOI: 10.3390/ijms22010130]
- Karpuz T, Araz M, Korkmaz L, Kılınc I, Findik S, Karaagaç M, Eryilmaz MK, Artac M. The Prognostic Value of Serum Semaphorin3A and VEGF Levels in Patients with Metastatic Colorectal Cancer. J Gastrointest Cancer 2020; 51: 491-497 [PMID: 31218581 DOI: 10.1007/s12029-019-00263-4
- 22 Morini SR, Denadai MV, Waisberg J, Lopes Filho GJ, Matos D, Saad SS. Metalloproteinases and colorectal cancer. Correlation of gene expression and clinical-pathological parameters. Acta Cir Bras 2020; 35: e202000707 [PMID: 32813775 DOI: 10.1590/s0102-865020200070000007]
- Liebman HA. Thrombocytopenia in cancer patients. Thromb Res 2014; 133 Suppl 2: S63-S69 23 [PMID: 24862148 DOI: 10.1016/S0049-3848(14)50011-4]
- 24 Eklund EA. Thrombocytopenia and cancer. Cancer Treat Res 2009; 148: 279-293 [PMID: 19377930 DOI: 10.1007/978-0-387-79962-9 16]
- 25 Desikan SP, Mclaughlin N, McClain C, Desikan R. Recurrent Colon Cancer: Presentation With Disseminated Intravascular Coagulation From Disseminated Carcinomatosis of the Bone Marrow. J Investig Med High Impact Case Rep 2021; 9: 23247096211012224 [PMID: 33966469 DOI: 10.1177/23247096211012224
- Takeyama H, Sakiyama T, Wakasa T, Kitani K, Inoue K, Kato H, Ueda S, Tsujie M, Fujiwara Y, 26 Yukawa M, Ohta Y, Inoue M. Disseminated carcinomatosis of the bone marrow with disseminated intravascular coagulation as the first symptom of recurrent rectal cancer successfully treated with chemotherapy: A case report and review of the literature. Oncol Lett 2017; 13: 4290-4294 [PMID: 28599429 DOI: 10.3892/ol.2017.5983]
- Ganguly K, Rauth S, Marimuthu S, Kumar S, Batra SK. Unraveling mucin domains in cancer and 27 metastasis: when protectors become predators. Cancer Metastasis Rev 2020; 39: 647-659 [PMID: 32488403 DOI: 10.1007/s10555-020-09896-5]
- Pothuraju R, Krishn SR, Gautam SK, Pai P, Ganguly K, Chaudhary S, Rachagani S, Kaur S, Batra 28 SK. Mechanistic and Functional Shades of Mucins and Associated Glycans in Colon Cancer. Cancers (Basel) 2020; 12 [PMID: 32168759 DOI: 10.3390/cancers12030649]



0 WJ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2190-2202

DOI: 10.4251/wjgo.v13.i12.2190

Prospective Study

ISSN 1948-5204 (online)

ORIGINAL ARTICLE

Anatomic resection improved the long-term outcome of hepatocellular carcinoma patients with microvascular invasion: A prospective cohort study

Jiang-Min Zhou, Chen-Yang Zhou, Xiao-Ping Chen, Zhi-Wei Zhang

ORCID number: Jiang-Min Zhou 0000-0001-7037-548X; Chen-Yang Zhou 0000-0002-6399-8301; Xiao-Ping Chen 0000-0001-8636-0493; Zhi-Wei Zhang 0000-0002-1240-5916.

Author contributions: Zhou JM analyzed and interpreted the patient data and wrote the manuscript; Zhou CY managed the patients, including recruiting patients, performing operations, and following-up with the patients; Zhang ZW and Chen XP designed the experiment and modified the manuscript; all authors read and approved the final manuscript.

Institutional review board

statement: The study was reviewed and approved for publication by Institutional Reviewer of Huazhong University of Science and Technology.

Informed consent statement: All study participants or their legal guardian provided informed written consent about personal and medical data collection prior to study enrolment.

Conflict-of-interest statement: All the authors have no conflict of interest related to the manuscript.

Data sharing statement: The

Jiang-Min Zhou, Chen-Yang Zhou, Zhi-Wei Zhang, Hepatic Surgery Center, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430030, Hubei Province, China

Xiao-Ping Chen, Translational Medicine Center, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430030, Hubei Province, China

Corresponding author: Zhi-Wei Zhang, PhD, Professor, Hepatic Surgery Center, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, No. 1095 Jiefang Avenue, Wuhan 430030, Hubei Province, China. zhiweizhangtjh@163.com

Abstract

BACKGROUND

The long-term effect of anatomic resection (AR) is better than that of nonanatomic resection (NAR). At present, there is no study on microvascular invasion (MVI) and liver resection types.

AIM

To explore whether AR improves long-term survival in patients with hepatocellular carcinoma (HCC) by removing the peritumoral MVI.

METHODS

A total of 217 patients diagnosed with HCC were enrolled in the study. The surgical margin was routinely measured. According to the stratification of different tumor diameters, patients were divided into the following groups: ≤ 2 cm group, 2-5 cm group, and > 5 cm group.

RESULTS

In the 2-5 cm diameter group, the overall survival (OS) of MVI positive patients was significantly better than that of MVI negative patients (P = 0.031). For the MVI positive patients, there was a statistically significant difference between AR and NAR (P = 0.027). AR leads to a wider surgical margin than NAR (2.0 ± 2.3 cm $vs 0.7 \pm 0.5$ cm, P < 0.001). In the groups with tumor diameters < 2 cm, both AR and NAR can obtain a wide surgical margin, and the surgical margins of AR are wider than that of NAR (3.5 ± 5.8 cm $vs 1.6 \pm 0.5$ cm, P = 0.048). In the groups with tumor diameters > 5 cm, both AR and NAR fail to obtain wide surgical margin



original anonymous dataset is available on request from the corresponding author at zhiweizhangtjh@163.com.

CONSORT 2010 statement: The

authors have read the CONSORT 2010 Statement, and the manuscript was prepared and revised according to the CONSORT 2010 Statement.

Supported by The National Key Research and Development Program of China, No. 2016YFC0106004.

Country/Territory of origin: China

Specialty type: Gastroenterology and hepatology

Provenance and peer review:

Unsolicited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: May 26, 2021 Peer-review started: May 26, 2021 First decision: June 24, 2021 Revised: July 5, 2021 Accepted: September 15, 2021 Article in press: September 15, 2021 Published online: December 15, 2021

P-Reviewer: Yagi H

 $(0.6 \pm 1.0 \text{ cm } vs \ 0.7 \pm 0.4 \text{ cm}, P = 0.491).$

CONCLUSION

For patients with a tumor diameter of 2-5 cm, AR can achieve the removal of peritumoral MVI by obtaining a wide incision margin, reduce postoperative recurrence, and improve prognosis.

Key Words: Microvascular invasion; Hepatocellular carcinoma; Anatomic resection; Surgical margin; Recurrence; Surgery

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: The prognosis of anatomic resection is better than that of non-anatomic resection with diameters from 2 to 5 cm. For tumor diameters smaller than 2 cm and larger than 5 cm, anatomic resection is not superior to non-anatomic resection. Anatomic resection can achieve the removal of peritumoral microvascular invasion by obtaining a wide incision margin. Both anatomic resection and non-anatomic resection can obtain wide surgical margins in the group with tumor diameters smaller than 2 cm. Both anatomic resection and non-anatomic resection failed to obtain wide surgical margins in the diameter larger than 5 cm group.

Citation: Zhou JM, Zhou CY, Chen XP, Zhang ZW. Anatomic resection improved the longterm outcome of hepatocellular carcinoma patients with microvascular invasion: A prospective cohort study. World J Gastrointest Oncol 2021; 13(12): 2190-2202 URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2190.htm

DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2190

INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the most common malignancies worldwide, and its high mortality makes it the second leading cause of cancer death[1]. Although the poor prognosis of HCC has improved significantly over the last decade due to increased knowledge of HCC behavior, improvements in staging systems, and multiple therapeutic options compared with other malignancies, HCC still has a high mortality rate^[2]. The prognosis of HCC remains very poor due to the high incidence of recurrence and metastasis, and the 5-year recurrence rate after curative treatment remains high (70%), with 15% of HCC patients developing extrahepatic metastasis[3]. One important reason is that tumor cells are able to penetrate the microvasculature, disseminate through the bloodstream to other sites, and form metastatic tumors. Studies have suggested that microvascular invasion (MVI) in HCC is one of the most significant risk factors for recurrence and metastasis in HCC following curative surgical resection[4]. MVI is defined as clusters of cancer cells observed microscopically in vessels located in the tumor capsule and surrounding liver parenchyma^[5]. Previous research reported that the incidence of MVI ranged from 15% to 57% in HCC specimens and was associated with tumor size, levels of alpha fetoprotein (AFP), and typical image features 6. Even for patients with HCC, the presence of MVI increases the risk of recurrence and dramatically shortens long-term survival [7,8]. The main reason for this is that the residual microthrombosis results in early recurrence. A safe surgical margin is a prerequisite for the complete removal of residual microtumor thrombosis. In HCC, invasion of the portal vein and intrahepatic and distant metastases are frequently observed. Resection of the portal vein invaded by the tumor is one method to decrease the risk of recurrence. Previous research has reported that both anatomic resection (AR) and non-anatomic resection (NAR), with a sufficient margin, can reduce the risk of early recurrence and improve the prognosis of HCC[9, 10]. A meta-analysis has shown that anatomical hepatectomy is more effective than non-anatomical hepatectomy[11]. We defined AR as the complete removal of at least one Couinaud segment containing the focus and the portal vein in the drainage area of the lesion. A complete tumor plus the rim of non-neoplastic liver parenchyma was considered a non-anatomic resection[12]. Studies have shown that although the width



S-Editor: Gong ZM L-Editor: Filipodia P-Editor: Yuan YY



of the resection margin does not influence the postoperative recurrence rates after liver resection, a wide margin is associated with a lower recurrence risk in patients with venous invasion or microsatellites [12,13]. Studies have shown that not all patients are suitable for AR, and controversy remains over the superiority of AR compared to NAR. In general, AR guarantees a wider surgical margin. However, a wider surgical margin means that more healthy liver tissue has to be removed. Almost all patients with HCC have liver cirrhosis, and the excessive removal of non-neoplastic liver parenchyma can lead to liver dysfunction and the morbidities of ascites, jaundice, and hypoalbuminemia. When the tumor is enormous and the remaining liver tissue is insufficient, AR may not be appropriate [14]. When the liver volume is insufficient, a surgical margin of at least 5 mm should be secured by NAR whenever possible[15]. Therefore, NAR still plays an important role in hepatectomy. Previous studies have shown that AR should be performed when the size of HCC ranges from 2 to 5 cm^[10]. However, whether AR should be recommended when the diameter is less than 2 cm or greater than 5 cm remains controversial. In addition, whether anatomical hepatectomy improves the prognosis of patients with hepatocellular carcinoma with microvascular invasion regardless of tumor size is unclear. Therefore, the aim of the study is to determine whether AR improves long-term survival in patients with hepatocellular carcinoma (HCC) by removing microvascular invasion (MVI).

MATERIALS AND METHODS

Patient selection

We consecutively enrolled 217 patients who underwent AR or NAR from November 2016 to November 2018 at the Hepatic Surgery Center, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology. The surgical margin was routinely measured. A pathological specimen of each patient was promptly sent to the pathology department and a detailed pathology report was issued. The pathological characteristics of our data, such as MVI, were derived from the report. The flow diagram of the enrolled patients is displayed in Figure 1. According to the stratification of different tumor diameters, patients were divided into the following groups: \leq 2 cm group, 2-5 cm group, and > 5 cm group.

Patient eligibility

The inclusion criteria were: (1) Definitive pathological diagnosis of HCC based on the World Health Organization criteria; (2) Curative resection, defined as complete macroscopic removal of the tumor with negative (R0) margins; (3) No prior anticancer treatment; and (4) Aged between 18 and 80 years. The exclusion criteria were: (1) Distant metastasis; (2) Portal vein tumor thrombosis (PVTT); and (3) Child-Pugh C liver disease. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the Ethical Committee of Tongji Hospital of Tongji Medical College of Huazhong University of Science and Technology, and the number of the approval was TJ-IRB20181101. Informed consent was obtained from each patient included in the study.

Terminology

Completely removing at least one Couinaud segment containing the focus and portal vein in the drainage area of the lesion was defined as an AR. A complete tumor plus a rim of non-neoplastic liver parenchyma was considered an NAR.

Surgical procedure

All surgeries were accomplished by a team who was able to professionally implement a hepatectomy. Patients were placed in supine position and under general anesthesia. The surgical principles were followed according to the corresponding the Union for International Cancer Control TNM classification. Intraoperative ultrasonography was routinely used in all patients to assess the number and size of the tumors, and their relation to nearby vascular structures. Proper hepatic vascular control techniques, including the selective inflow occlusion (SIO) maneuver and intermittent Pringle maneuvers (IPs), were used to reduce bleeding during liver resection. The SIO maneuver is described by the following procedure: dissecting the portal vein, proper hepatic artery, right and left hepatic arteries, and bile ducts followed by continuously blocking the hepatic artery in the tumor bearing lobe with a bulldog clamp. IPs encircling the hepatoduodenal ligament were performed with cycles of clamping and



Figure 1 Flow diagram for the study. AR: Anatomic resection; MVI: Microvascular invasion; NAR: Non-anatomic resection.

unclamping times of 15 min and 5 min, respectively.

Follow-up and tumor recurrence

The patients were surveilled every 1 mo with ultrasonography and AFP during the first 6 mo after surgery and every 3 mo thereafter. Patients were scheduled to have a computerized tomography (CT) scan every 6 mo and a magnetic resonance imaging (MRI) every year. Recurrence was diagnosed by computed tomography scans, magnetic resonance imaging, digital subtraction angiography, and elevated serum AFP level. We reviewed the governmental death registration and performed telephone follow-ups. Patients were excluded if they were not followed up as required, or their governmental data were incomplete. Follow-up was terminated on May 31, 2021. Patients lost to follow-up and with missing data were prematurely excluded. Ultimately, 217 eligible patients were enrolled in the study. Death was the primary endpoint and the diagnosis of intrahepatic recurrence and/or extrahepatic metastasis was the secondary endpoint.

Study design

First, all patients were randomized to receive standard anatomic or non-anatomic resection. After surgery, we measured the surgical margin and identified microvascular invasion. We divided them into three groups based on tumor size to compare the effects of anatomic and nonanatomic hepatectomy in subgroups. In addition, the relationship between tumor size and surgical margin was further analyzed.

Statistical analysis

Data are presented as the mean ± SD. The overall survival (OS) was analyzed using Kaplan-Meier survival curves and a log-rank test. Student's t-tests were used for comparison between groups where appropriate. A χ^2 test was used for comparison between groups where appropriate. P < 0.05 was considered statistically significant.





Figure 2 The long-term outcome of hepatocellular carcinoma with different tumor diameters. A: Kaplan-Meier analysis of the overall survival (OS) in microvascular invasion (MVI) positive and MVI negative patients with a tumor diameter 2-5 cm; B: In the group with a tumor diameter of 2-5 cm, Kaplan-Meier analysis of the OS in receiving MVI positive anatomic resection (AR) and non-anatomic resection (NAR) patients; C: In the group with a tumor diameter of 2-5 cm, Kaplan-Meier analysis of the OS in receiving AR and NAR patients with MVI negative. D: Kaplan-Meier analysis of the OS in MVI positive and MVI negative patients with a tumor diameter of less than 2 cm.

Statistical analyses were performed with SPSS version 19.0.

RESULTS

Patient characteristics

The mean follow-up time was 45.2 ± 6.3 mo (median: 46.0 mo; range: 30.6-53.4 mo). The cumulative survival rate for all patients was 90%, 57%, and 39% at 1, 3, and 5 years. All of the category boundaries were defined by the clinical guideline or recognized criterion when continuous variables were categorized. Table 1 demonstrates the clinical and tumor characteristics of the 217 patients with HCC. The mean patient age was 52.6 ± 12.4 years (range: 21-74 years). The patients were 90.8% (197/217) male and 9.2% (20/217) female. In total, 86.6% (178/217) were positive for hepatitis B virus (HBV) infection and 10 patients were positive for the hepatitis C virus (HCV). Overall, 42.4% (92/217) were Barcelona Clinic Liver Cancer (BCLC) 0+A, and



Zaishideng® WJGO | https://www.wjgnet.com

Table 1 Clinical characteristics of 217 patients	
Clinical characteristics	Value
Age, yr, (mean ± SD)	52.6 ± 12.4
Sex, n (%)	
Male	197 (90.8)
Female	20 (9.2)
ALT (U/mL)	30.2 ± 14.3
AST (U/mL)	38.6 ± 32.0
TBiL (µmol/L)	15.8 ± 9.6
Child-Pugh score, n (%)	
А	209 (96.3)
В	8 (3.7)
Hepatitis virus, n (%)	
HBV	178 (82.0)
HCV	10 (4.6)
No	29 (13.4)
Liver cirrhosis, n (%)	
No	43 (19.8)
Yes	174 (80.2)
ICG-R15 (%)	6.6 ± 3.8
AFP (ng/mL), <i>n</i> (%)	
≤ 4 00	144 (66.4)
> 400	73 (33.6)
Tumor diameter (cm)	4.9 ± 3.5
No. of tumor, <i>n</i> (%)	
Singe	157 (72.4)
Multiple	60 (27.6)
Microvascular invasion, n (%)	
Yes	75 (34.6)
No	142 (65.4)
BCLC stage, n (%)	
0+A	92 (42.4)
B+C	125 (57.6)
Operation method, <i>n</i> (%)	
Open	56 (25.8)
Laparoscopic	161 (74.2)
Operation time (min)	171 ± 40
Blood loss (mL)	160 ± 180
Blood transfusion, n (%)	
No	189 (87.1)
Yes	28 (12.9)
Hepatic vascular occlusion, <i>n</i> (%)	
No	64 (29.5)

Saishideng® WJGO | https://www.wjgnet.com

Υ	e	s

AFP: Alpha fetoprotein; ALT: Alanine transaminase; AST: Aspartate aminotransferase; BCLC: Barcelona Clinic Liver Cancer staging system; HBV: Hepatitis B virus; HCV: Hepatitis C virus; ICG R15 (%): Indocyanine green retention rate at 15 min; SD: Standard deviation; TBiL: Total bilirubin.

34.6% (75/217) were MVI positive. Eight patients (3.7%) had a hepatic function of Child-Pugh score B and received short-term liver protective therapy before surgery. The clinical and pathological characteristics of the AR and NAR groups are summarized in Table 2. There were no significant differences in age, sex, Child-Pugh class, *etc.* between the two groups.

Comparison of types of hepatectomy with prognosis in the 2-5 cm tumor diameter subgroup

The 217 included patients were divided into three groups: diameter $\leq 2 \text{ cm} (84/217), 2$ -5 cm (87/217) and > 5 cm (46/217). In the diameter 2-5 cm group, there was a statistically significant difference between MVI positive and MVI negative patients (median OS 32.0 mo, 95% confidence interval (CI): 13.3-50.8 mo vs not reached, P = 0.031) (Figure 2A). For the MVI positive patients, there was a statistically significant difference between AR and NAR (median OS not reached vs 29.0 mo 95%CI: 8.0-43.9 mo, P = 0.027) (Figure 2B). However, for the MVI negative patients, there were no statistically significant differences between those who underwent AR and NAR (median OS not reached *vs* 44.2 mo, 95%CI: 27.0-61.5 mo, *P* = 0.206) (Figure 2C). This suggests that AR improved OS only in MVI positive patients, but not in MVI negative patients in the diameter 2-5 cm group. In addition, we compared the surgical margin of AR and NAR in the 2-5 cm diameter group. We found that AR led to a wider surgical margin than NAR (2.0 \pm 2.3 cm vs 0.7 \pm 0.5 cm, P < 0.001) (Table 3). We speculated that AR can achieve a wide enough surgical margin, which can remove MVI in advance, reducing the risk of postoperative recurrence and improving the prognosis.

Comparison of types of hepatectomy with prognosis in the subgroup with a tumor diameter less than 2 cm

Among patients with a diameter less than 2 cm, there was no statistically significant difference between MVI positive and MVI negative patients (median OS 34 mo 95%CI: 27.6-40.4 mo *vs* 45.0 mo 95%CI: 39.4-50.6 mo, P = 0.345) (Figure 2D). In addition, there was no statistically significant difference in overall survival between patients who received AR and NAR, whether MVI positive or MVI negative (median OS 32.4 mo 95%CI: 13.0-46.3 mo *vs* not reached, P = 0.097; median OS 46.6 mo, 95%CI: 32.8-60.4 mo *vs* 41.3 mo, 95%CI: 34.6-48.1 mo, P = 0.869) (Figure 3A and B). By comparing the surgical margins of AR and NAR patients, we found that although the AR margins were wider than those of NAR patients, the margins of both were greater than 1 cm (3.5 ± 5.8 cm *vs* 1.6 \pm 0.5 cm, P = 0.048) (Table 3; Figure 4B). If a surgical margin of 1 cm is ensured, it can be clinically regarded as a R0 resection. Thus, for patients with a tumor of less than 2 cm in diameter, both AR and NAR can achieve a wide surgical margin to ensure the removal of MVI.

Comparison of types of hepatectomy with prognosis in the subgroup with a tumor diameter larger than 5 cm

In the group with a diameter > 5 cm, the prognosis of MVI positive patients was significantly worse than that of MVI negative patients (median OS 24.0 mo, 95%CI: 15.7-32.4 mo *vs* not reached, P = 0.004) (Figure 3C). However, there was no statistically significant difference in overall survival between patients who received AR and NAR, whether MVI positive or MVI negative, (median OS 27.2 mo, 95%CI: 21.9-32.5 mo *vs* 20.1 mo, 95%CI: 5.6-28.8 mo, P = 0.428; median OS not reached *vs* 38.3 mo, 95%CI: 19.5-60.5 mo, P = 0.714) (Figures 3D and 4A). In addition, there were no statistically significant differences between AR and NAR in surgical margins and the margins of both were less than 1 cm (0.6 ± 1.0 cm *vs* 0.7 ± 0.4 cm, P = 0.491) (Table 3; Figure 4B). For patients with tumors larger than 5 cm in diameter, neither AR nor NAR could obtain a wide enough surgical margin to ensure the removal of MVI.

Zaishideng® WJGO | https://www.wjgnet.com

Table 2 Clinical characteristics of anatomic	resection and non-anatomic res	section	
Clinical characteristics	AR (<i>n</i> = 103)	NAR (<i>n</i> = 114)	<i>P</i> value
Age, yr (mean ± SD)	54.1 ± 12.5	51.3 ± 12.2	0.095
Sex, n (%)			0.812
Male	93 (90.3)	104 (91.2)	
Female	10 (9.7)	10 (8.8)	
ALT (U/mL)	30.5 ± 14.5	29.8 ± 14.2	0.721
AST (U/mL)	36.3 ± 28.9	40.6 ± 34.6	0.332
TBiL (µmol/L)	17.0 ± 10.5	15.9 ± 9.0	0.098
Child-Pugh score, n (%)			0.154
А	103 (100)	106 (93.0)	
В	0 (0)	2 (7.0)	
HBsAg, n (%)			0.622
Positive	88 (85.4)	100 (87.7)	
Negative	15 (14.6)	14 (12.3)	
Liver cirrhosis, <i>n</i> (%)			0.841
No	21 (20.4)	22 (19.3)	
Yes	82 (79.6)	92 (80.7)	
AFP (ng/mL), <i>n</i> (%)			0.446
< 400	71 (68.9)	73 (64.0)	
≥ 400	32 (31.1)	41 (36.0)	
ICG-R15 (%)	5.9 ± 3.5	5.4 ± 3.1	0.578
BCLC stage, n (%)			0.867
0 + A	46 (44.7)	46 (40.4)	
B + C	57 (55.3)	68 (59.6)	
Operation method, n (%)			0.423
Open	24 (23.3)	32 (28.1)	
Laparoscopic	79 (76.7)	82 (71.9)	
Operation time (min)	171.4 ± 48.2	165.5 ± 45.3	0.335
Blood loss (mL)	210 ± 233	170 ± 175	0.233
Blood transfusion, n (%)			0.774
No	89 (86.4)	100 (87.7)	
Yes	14 (13.6)	14 (12.3)	
Hepatic vascular occlusion, n (%)			0.314
No	27 (26.2)	37 (32.5)	
Yes	76 (73.8)	77 (67.5)	
Largest tumor size, (cm)	5.1 ± 3.6	4.7 ± 3.3	0.507
No. of tumors, <i>n</i> (%)			0.884
Single	75 (72.8)	82 (71.9)	
Multiple	28 (27.2)	32 (28.1)	
Tumor encapsulation, n (%)			0.051
No	57 (55.3)	66 (57.9)	
Yes	46 (44.7)	48 (42.1)	

Zhou JM et al. AR and surgical margin

Satellite lesion, n (%)			0.822
Yes	9 (8.7)	9 (7.9)	
No	94 (91.3)	105 (92.1)	
Tumor differentiation stage, <i>n</i> (%)			0.943
Edmondson I II	51 (49.5)	57 (50.0)	
Edmondson III IV	52 (50.5)	57 (50.0)	
Microvascular invasion, n (%)			0.845
Yes	37 (35.9)	38 (33.3)	
No	66 (64.1)	76 (66.7)	

AFP: Alpha fetoprotein; ALT: Alanine transaminase; AST: Aspartate aminotransferase; AR: Anatomic resection; BCLC: Barcelona Clinic Liver Cancer staging system; HBsAg: Hepatitis B surface antigen; ICG R15 (%): Indocyanine green retention rate at 15 min; NAR: Non-anatomic resection; TBiL: Total bilirubin.

Table 3 The surgical margin of different tumor diameter groups				
Tumor diameter	Surgical margin (cm)			
rumor diameter	AR NAR	<i>P</i> value		
$D \le 2 \text{ cm}$	3.5 ± 5.8	1.6 ± 0.5	0.048	
$2 \text{ cm} \le D \le 5 \text{ cm}$	2.0 ± 2.3	0.7 ± 0.5	< 0.001	
D > 5 cm	0.6 ± 1.0	0.7 ± 0.4	0.491	

AR: Anatomic resection; D: Diameter; NAR: Non-anatomic resection

DISCUSSION

At present, surgeons consider hepatectomy and liver transplantation the optimal therapies to improve prognosis in HCC; however, tumor recurrence is still an important cause of death in patients [16]. Previous research has demonstrated that microvascular invasion is a vital risk factor for the prognosis of HCC patients after curative hepatectomy^[17]. As long as a surgical margin of 1 cm is ensured, it can be clinically regarded as an R0 resection[18]. The margin of microvascular invasion is generally no more than 1 cm. Therefore, an R0 resection enables the complete removal of the liver tissue invaded by the microvascular thrombosis. In contrast, positive margins were associated with a worse prognosis[19,20]. Previous studies have shown that a wider surgical margin has been associated with a better prognosis among patients with HCC[21-23]. In addition, a previous study has shown that AR led to a better OS than NAR. In a multivariable analysis, an AR was one of the prognostic factors[9]. AR can reduce the risk of tumor residues and recurrence due to the elimination of venous tumor thrombosis within the resected domain, when at least one complete Couinaud segment and the portal vein in the drainage area of the lesion are removed[24,25]. However, almost all patients with HCC have liver cirrhosis and excessive removal of non-neoplastic liver parenchyma can lead to liver dysfunction and the morbidities of ascites, jaundice, and hypoalbuminemia.

Our data indicated that AR improved OS only in MVI positive patients, but not in MVI negative patients in the 2-5 cm diameter group. We speculated that AR can achieve a wide enough surgical margin, which can remove MVI in advance, reduce the risk of postoperative recurrence, and improve the prognosis. For patients with a tumor diameter of less than 2 cm, both AR and NAR can obtain a wide surgical margin to ensure removal of MVI. Therefore, patients with a diameter less than 2 cm, both AR and NAR, can achieve a good prognosis. In other words, an R0 resection enabled the removal of the liver tissue invaded by the MVI regardless of whether AR or NAR was chosen by the surgeon. This suggests that AR is not necessary for tumors with a diameter of less than 2 cm, as long as sufficient surgical margin is ensured. However, in the > 5 cm group, both AR and NAR cannot guarantee sufficient surgical margin, which is one of the reasons why tumors with a diameter of more than 5 cm have a





Figure 3 The long-term outcome of hepatocellular carcinoma with different tumor diameter. A: In the group with a tumor diameter of less than 2 cm, Kaplan-Meier analysis of the overall survival (OS) in receiving anatomic resection (AR) and non-anatomic resection (NAR) patients with microvascular invasion (MVI) positive; B: In the group with a tumor diameter of less than 2 cm, Kaplan-Meier analysis of the OS in patients receiving AR and NAR patients who were MVI negative; C: Kaplan-Meier analysis of the OS in MVI positive and MVI negative patients with tumor diameters larger than 5 cm; D: In the group with a tumor diameter of larger than 5 cm, Kaplan-Meier analysis of the OS in MVI positive patients receiving AR and NAR.

worse prognosis than tumors with a diameter of less than 2 cm. We believe that the surgeon needs to consider whether the residual liver volume and liver function reserve are sufficient when faced with a very large tumor. Therefore, for tumor diameters larger than 5 cm, the width of the resection margin should be increased appropriately when a sufficient liver volume and a good liver function can be ensured.

Limitations

The limitations of this study are the relatively small samples, short follow-up time, and a single study center cohort study. A multicenter clinical trial should be designed to further validate the prognostic significance of types of hepatectomy in HCC.

CONCLUSION

For patients with a tumor diameter of 2-5 cm, AR can achieve the removal of pe-





Figure 4 Kaplan-Meier analysis of the overall survival (A) and comparison of surgical margins in patients with different tumor diameters (B). A: In the group with a tumor diameter larger than 5 cm, Kaplan-Meier analysis of the overall survival in microvascular invasion negative patients receiving anatomic resection (AR) and non-anatomic resection (NAR); B: Comparison of surgical margins in patients receiving AR and NAR with different tumor diameters.

> ritumoral MVI by obtaining a wide incision margin, reduce postoperative recurrence, and improve prognosis.

ARTICLE HIGHLIGHTS

Research background

At present, most studies suggest that anatomical resection is more effective than nonanatomical resection in the tumor diameter ranging from 2 cm to 5 cm. However, for tumors smaller than 2 cm and larger than 5 cm in diameter, the advantage of anatomic hepatectomy is not significant. Why is that? Does anatomic resection (AR) have an advantage over non-anatomic resection (NAR) in hepatocellular carcinoma (HCC) patients with microvascular invasion (MVI)?

Research motivation

Our study aimed to determine the effects of AR and NAR in different tumor diameter stratification. Further analysis shows that AR improves patient outcomes by obtaining a wider surgical margin.

Research objectives

This study compared the efficacy of AR and NAR in different tumor diameter subgroups in a prospective cohort study.

Research methods

First, all patients were randomized to receive standard anatomic or non-anatomic resection. After surgery, we measured the surgical margin and identified microvascular invasion. We divided them into three groups based on tumor size.

Research results

When the tumor is enormous and the remaining liver tissue is insufficient, AR may not be appropriate. For patients with a tumor diameter of 2-5 cm, AR can achieve the removal of peritumoral MVI by obtaining a wide incision margin, reducing postoperative recurrence and improving prognosis. For patients with a tumor of less than 2 cm in diameter, both AR and NAR can obtain a wide surgical margin to ensure the removal of MVI. AR should not be recommended for those patients. For patients with tumors larger than 5 cm in diameter, neither AR nor NAR could obtain a wide surgical margin to ensure removal of MVI.



Research conclusions

The doctor should ensure sufficient surgical margin on the premise of ensuring the safety of the operation. Therefore, for patients with a tumor diameter of 2-5 cm, AR should be strongly recommended.

Research perspectives

The study could guide doctors in their choice of surgical procedures. In general, AR guarantees a wider surgical margin. However, a wider surgical margin means that more healthy liver tissue has to be removed. Almost all patients with HCC have liver cirrhosis, and the excessive removal of non-neoplastic liver parenchyma can lead to liver dysfunction and the morbidities of ascites, jaundice, and hypoalbuminemia. When the tumor is enormous and the remaining liver tissue is insufficient, AR may not be appropriate.

ACKNOWLEDGEMENTS

The authors would like to thank Dr. Chang Shu who is a of statistics (Translational Medicine Center, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology) for her technical assistance of statistical analysis.

REFERENCES

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin 2021; 71: 209-249 [PMID: 33538338 DOI: 10.3322/caac.21660]
- Njei B, Rotman Y, Ditah I, Lim JK. Emerging trends in hepatocellular carcinoma incidence and 2 mortality. Hepatology 2015; 61: 191-199 [PMID: 25142309 DOI: 10.1002/hep.27388]
- Tabrizian P, Jibara G, Shrager B, Schwartz M, Roayaie S. Recurrence of hepatocellular cancer after 3 resection: patterns, treatments, and prognosis. Ann Surg 2015; 261: 947-955 [PMID: 25010665 DOI: 10.1097/SLA.000000000000710
- Wang L, Jin YX, Ji YZ, Mu Y, Zhang SC, Pan SY. Development and validation of a prediction 4 model for microvascular invasion in hepatocellular carcinoma. World J Gastroenterol 2020; 26: 1647-1659 [PMID: 32327913 DOI: 10.3748/wjg.v26.i14.1647]
- Shirabe K, Toshima T, Kimura K, Yamashita Y, Ikeda T, Ikegami T, Yoshizumi T, Abe K, Aishima S, Maehara Y. New scoring system for prediction of microvascular invasion in patients with hepatocellular carcinoma. Liver Int 2014; 34: 937-941 [PMID: 24393295 DOI: 10.1111/liv.12459]
- Renzulli M, Brocchi S, Cucchetti A, Mazzotti F, Mosconi C, Sportoletti C, Brandi G, Pinna AD, 6 Golfieri R. Can Current Preoperative Imaging Be Used to Detect Microvascular Invasion of Hepatocellular Carcinoma? Radiology 2016; 279: 432-442 [PMID: 26653683 DOI: 10.1148/radiol.2015150998
- Zhou J, Sun HC, Wang Z, Cong WM, Wang JH, Zeng MS, Yang JM, Bie P, Liu LX, Wen TF, Han 7 GH, Wang MQ, Liu RB, Lu LG, Ren ZG, Chen MS, Zeng ZC, Liang P, Liang CH, Chen M, Yan FH, Wang WP, Ji Y, Cheng WW, Dai CL, Jia WD, Li YM, Li YX, Liang J, Liu TS, Lv GY, Mao YL, Ren WX, Shi HC, Wang WT, Wang XY, Xing BC, Xu JM, Yang JY, Yang YF, Ye SL, Yin ZY, Zhang BH, Zhang SJ, Zhou WP, Zhu JY, Liu R, Shi YH, Xiao YS, Dai Z, Teng GJ, Cai JQ, Wang WL, Dong JH, Li Q, Shen F, Qin SK, Fan J. Guidelines for Diagnosis and Treatment of Primary Liver Cancer in China (2017 Edition). Liver Cancer 2018; 7: 235-260 [PMID: 30319983 DOI: 10.1159/000488035
- 8 Erstad DJ, Tanabe KK. Prognostic and Therapeutic Implications of Microvascular Invasion in Hepatocellular Carcinoma. Ann Surg Oncol 2019; 26: 1474-1493 [PMID: 30788629 DOI: 10.1245/s10434-019-07227-9
- Hasegawa K, Kokudo N, Imamura H, Matsuyama Y, Aoki T, Minagawa M, Sano K, Sugawara Y, Takayama T, Makuuchi M. Prognostic impact of anatomic resection for hepatocellular carcinoma. Ann Surg 2005; 242: 252-259 [PMID: 16041216 DOI: 10.1097/01.sla.0000171307.37401.db]
- 10 Eguchi S, Kanematsu T, Arii S, Okazaki M, Okita K, Omata M, Ikai I, Kudo M, Kojiro M, Makuuchi M, Monden M, Matsuyama Y, Nakanuma Y, Takayasu K; Liver Cancer Study Group of Japan. Comparison of the outcomes between an anatomical subsegmentectomy and a non-anatomical minor hepatectomy for single hepatocellular carcinomas based on a Japanese nationwide survey. Surgery 2008; 143: 469-475 [PMID: 18374043 DOI: 10.1016/j.surg.2007.12.003]
- 11 Moris D, Tsilimigras DI, Kostakis ID, Ntanasis-Stathopoulos I, Shah KN, Felekouras E, Pawlik TM. Anatomic versus non-anatomic resection for hepatocellular carcinoma: A systematic review and metaanalysis. Eur J Surg Oncol 2018; 44: 927-938 [PMID: 29751946 DOI: 10.1016/j.ejso.2018.04.018]
- 12 Couinaud C. Anatomic principles of left and right regulated hepatectomy: technics. J Chir (Paris) 1954; 70: 933-966 [PMID: 13233306]



- 13 Moris D, Ronnekleiv-Kelly S, Rahnemai-Azar AA, Felekouras E, Dillhoff M, Schmidt C, Pawlik TM. Parenchymal-Sparing Versus Anatomic Liver Resection for Colorectal Liver Metastases: a Systematic Review. J Gastrointest Surg 2017; 21: 1076-1085 [PMID: 28364212 DOI: 10.1007/s11605-017-3397-y]
- Viganò L, Procopio F, Mimmo A, Donadon M, Terrone A, Cimino M, Fabbro DD, Torzilli G. 14 Oncologic superiority of anatomic resection of hepatocellular carcinoma by ultrasound-guided compression of the portal tributaries compared with nonanatomic resection: An analysis of patients matched for tumor characteristics and liver function. Surgery 2018; 164: 1006-1013 [PMID: 30195402 DOI: 10.1016/j.surg.2018.06.030]
- 15 Field WBS, Rostas JW, Philps P, Scoggins CR, McMasters KM, Martin RCG 2nd. Wide versus narrow margins after partial hepatectomy for hepatocellular carcinoma: Balancing recurrence risk and liver function. Am J Surg 2017; 214: 273-277 [PMID: 28615138 DOI: 10.1016/j.amjsurg.2017.06.002]
- 16 Lee MW, Lim HK. Management of sub-centimeter recurrent hepatocellular carcinoma after curative treatment: Current status and future. World J Gastroenterol 2018; 24: 5215-5222 [PMID: 30581270 DOI: 10.3748/wig.v24.i46.52151
- Yamashita YI, Imai K, Yusa T, Nakao Y, Kitano Y, Nakagawa S, Okabe H, Chikamoto A, Ishiko T, 17 Yoshizumi T, Aishima S, Maehara Y, Baba H. Microvascular invasion of single small hepatocellular carcinoma ≤3 cm: Predictors and optimal treatments. Ann Gastroenterol Surg 2018; 2: 197-203 [PMID: 29863190 DOI: 10.1002/ags3.12057]
- Cherqui D, Tantawi B, Alon R, Piedbois P, Rahmouni A, Dhumeaux D, Julien M, Fagniez PL. 18 Intrahepatic cholangiocarcinoma. Results of aggressive surgical management. Arch Surg 1995; 130: 1073-1078 [PMID: 7575119 DOI: 10.1001/archsurg.1995.01430100051011]
- 19 Lang H, Sotiropoulos GC, Sgourakis G, Schmitz KJ, Paul A, Hilgard P, Zöpf T, Trarbach T, Malagó M, Baba HA, Broelsch CE. Operations for intrahepatic cholangiocarcinoma: single-institution experience of 158 patients. J Am Coll Surg 2009; 208: 218-228 [PMID: 19228533 DOI: 10.1016/j.jamcollsurg.2008.10.017]
- 20 Xu X, Zhang HL, Liu QP, Sun SW, Zhang J, Zhu FP, Yang G, Yan X, Zhang YD, Liu XS. Radiomic analysis of contrast-enhanced CT predicts microvascular invasion and outcome in hepatocellular carcinoma. J Hepatol 2019; 70: 1133-1144 [PMID: 30876945 DOI: 10.1016/j.jhep.2019.02.023]
- Kabir T, Syn NL, Tan ZZX, Tan HJ, Yen C, Koh YX, Kam JH, Teo JY, Lee SY, Cheow PC, Chow 21 PKH, Chung AYF, Ooi LL, Chan CY, Goh BKP. Predictors of post-operative complications after surgical resection of hepatocellular carcinoma and their prognostic effects on outcome and survival: A propensity-score matched and structural equation modelling study. Eur J Surg Oncol 2020; 46: 1756-1765 [PMID: 32345496 DOI: 10.1016/j.ejso.2020.03.219]
- 22 Aoki T, Kubota K, Hasegawa K, Kubo S, Izumi N, Kokudo N, Sakamoto M, Shiina S, Takayama T, Nakashima O, Matsuyama Y, Murakami T, Kudo M; Liver Cancer Study Group of Japan. Significance of the surgical hepatic resection margin in patients with a single hepatocellular carcinoma. Br J Surg 2020; 107: 113-120 [PMID: 31654406 DOI: 10.1002/bjs.11329]
- Tsilimigras DI, Sahara K, Moris D, Hyer JM, Paredes AZ, Bagante F, Merath K, Farooq AS, Ratti F, Marques HP, Soubrane O, Azoulay D, Lam V, Poultsides GA, Popescu I, Alexandrescu S, Martel G, Guglielmi A, Hugh T, Aldrighetti L, Endo I, Pawlik TM. Effect of Surgical Margin Width on Patterns of Recurrence among Patients Undergoing R0 Hepatectomy for T1 Hepatocellular Carcinoma: An International Multi-Institutional Analysis. J Gastrointest Surg 2020; 24: 1552-1560 [PMID: 31243714 DOI: 10.1007/s11605-019-04275-0]
- 24 Wakai T, Shirai Y, Sakata J, Kaneko K, Cruz PV, Akazawa K, Hatakeyama K. Anatomic resection independently improves long-term survival in patients with T1-T2 hepatocellular carcinoma. Ann Surg Oncol 2007; 14: 1356-1365 [PMID: 17252289 DOI: 10.1245/s10434-006-9318-z]
- Regimbeau JM, Kianmanesh R, Farges O, Dondero F, Sauvanet A, Belghiti J. Extent of liver 25 resection influences the outcome in patients with cirrhosis and small hepatocellular carcinoma. Surgery 2002; 131: 311-317 [PMID: 11894036 DOI: 10.1067/msy.2002.121892]



GO WJ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2203-2215

DOI: 10.4251/wjgo.v13.i12.2203

ISSN 1948-5204 (online)

SYSTEMATIC REVIEWS

Minimally invasive surgical treatment of intrahepatic cholangiocarcinoma: A systematic review

Renato Patrone, Francesco Izzo, Raffaele Palaia, Vincenza Granata, Guglielmo Nasti, Alessandro Ottaiano, Gilda Pasta, Andrea Belli

ORCID number: Renato Patrone 0000-0002-6969-4157; Francesco Izzo 0000-0003-3093-5408: Raffaele Palaia 0000-0002-4299-3448; Vincenza Granata 0000-0002-6601-3221; Guglielmo Nasti 0000-0002-3590-382X; Alessandro Ottaiano 0000-0002-2901-3855; Gilda Pasta 0000-0002-0245-9670; Andrea Belli 0000-0002-6252-573X.

Author contributions: All authors contributed significantly to the present research and reviewed the entire manuscript. Patrone R and Belli A participated substantially in the conception, design and execution of the study and in the analysis and interpretation of the data; they also participated substantially in the drafting and editing of the manuscript; Izzo F, Palaia R, Granata V, Nasti G, Ottaiano A and Pasta G participated substantially in the conception, design and execution of the study and in the review of the data.

Conflict-of-interest statement:

Authors have no conflict of interest.

PRISMA 2009 Checklist statement:

The authors have read the PRISMA 2009 Checklist, and the manuscript was prepared and revised according to the PRISMA 2009 Checklist

Renato Patrone, PhD ICTH, University of Naples Federico II, Naples 80100, Italy

Francesco Izzo, Raffaele Palaia, Andrea Belli, Department of Abdominal Oncology, Division of Hepatobiliary Surgical Oncology, Istituto Nazionale Tumori, Fondazione G. Pascale, IRCCS, Naples 80131, Italy

Vincenza Granata, Division of Radiology, Istituto Nazionale Tumori, Fondazione G. Pascale, IRCCS, Naples 80131, Italy

Guglielmo Nasti, Alessandro Ottaiano, SSD-Innovative Therapies for Abdominal Metastases, Clinical and Experimental Abdominal Oncology, Istituto Nazionale Tumori, Fondazione G. Pascale, IRCCS, Naples 80131, Italy

Gilda Pasta, Division of Anesthesia, Pain medicine and Supportive Care, Istituto Nazionale Tumori, Fondazione G. Pascale, IRCCS, Naples 80131, Italy

Corresponding author: Andrea Belli, MD, Postdoc, Surgeon, Department of Abdominal Oncology, Division of Hepatobiliary Surgical Oncology, Istituto Nazionale Tumori, Fondazione G. Pascale, IRCCS, Via Mariano Semmola, Naples 80131, Italy. a.belli@istitutotumori.na.it

Abstract

BACKGROUND

Intrahepatic cholangiocarcinoma (ICC) is the second most common primary liver cancer and is characterized by an aggressive behavior and a dismal prognosis. Radical surgical resection represents the only potentially curative treatment. Despite the increasing acceptance of laparoscopic liver resection for surgical treatment of malignant liver diseases, its use for ICC is not commonly performed. In fact, to achieve surgical free margins a major resection and/or vascular and/or biliary reconstructions is often needed, as well as an associated lymph node dissection.

AIM

To review and summarize the current evidences on the minimally invasive resection of ICC.

METHODS

A systematic review of the literature based on the criteria predetermined by the investigators was performed from the 1st of January 2009 up to the 1st of January



Country/Territory of origin: Italy

Specialty type: Surgery

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): C, C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: March 23, 2021 Peer-review started: March 23, 2021 First decision: May 3, 2021 Revised: May 30, 2021 Accepted: October 31, 2021 Article in press: October 31, 2021 Published online: December 15, 2021

P-Reviewer: Boueroy P, Saengboonmee C S-Editor: Zhang H L-Editor: A P-Editor: Zhang H



2021 in 4 databases (PubMed, Scopus, Google Scholar, and Cochrane databases). All retrospective and prospective studies reporting on the comparative outcomes of open vs minimally invasive treatment of ICC were included. An evaluation of manuscripts quality was achieved using Methodological Index for Non-Randomized Studies criteria and Newcastle-Ottawa Scale.

RESULTS

After a systematic search 9 studies fulfilled the inclusion criteria. Among the all 3012 included patients, 2450 were operated by an open approach and 562 by a minimally invasive (laparoscopic) approach. Baseline characteristics, tumor characteristics, surgical outcomes and oncological outcomes were collected and analyzed, highlighting values with a statistical significant difference between patients treated with open or laparoscopic approach. Shorter hospital stay and lower intraoperative blood losses were reported by some Authors in minimally invasive surgery, on the contrary, in the open group there was a higher number of lymphadenectomies and a higher percentage of major hepatectomies.

CONCLUSION

Minimally invasive resection of ICC has some short-term benefits and it is safe and feasible only in selected centers with a high experience in laparoscopic approach for liver surgery. Minimally invasive surgery, actually, was considered mainly in patients with a tumor with a diameter < 5 cm, without invasion of main biliary duct or main vessel and no vascular or biliary reconstructions were planned. Further studies are needed to elucidate its impact on long term oncologic outcomes.

Key Words: Cholangiocarcinoma; Minimally invasive; Laparoscopic; Liver resection; Hepatectomy; Biliary neoplasm

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Reports on the minimally invasive treatment of intrahepatic cholanciocarcinoma are scanty and no clear evidences on the feasibility, safety and oncological results are currently available. The aim of our study is to review and summarize the current evidences on the topic and to compare the short and long term outcomes to those of open surgical resection.

Citation: Patrone R, Izzo F, Palaia R, Granata V, Nasti G, Ottaiano A, Pasta G, Belli A. Minimally invasive surgical treatment of intrahepatic cholangiocarcinoma: A systematic review. World J Gastrointest Oncol 2021; 13(12): 2203-2215 URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2203.htm

DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2203

INTRODUCTION

Intrahepatic cholangiocarcinoma (ICC) is a rare gastrointestinal malignancy arising from epithelial cells of the intrahepatic bile ducts (cholangiocytes) and accounts for 10%-15% of all primary liver cancers[1]. ICC is the second most common primary liver cancer after hepatocellular carcinoma and is characterized by an aggressive behavior and a dismal prognosis[2]. Its occurrence has progressively raised worldwide during the past decades with a reported increase of more than 165% in its incidence in the last 35 years in the western world population [3,4] (from 0.49 per 100000 in 1995 to 1.49 per 100000 in 2014 in the United States)[5]. Radical surgical resection represents the only potentially curative treatment of ICC. Regrettably, less than 40% of patients are eligible for surgery mainly due to late advanced disease at the time of diagnosis[6]. Considering the lack of effective and established chemotherapeutic options, both in adjuvant and first line setting, even after radical resection 50% to 60% of patients will experience a recurrence[7], with a 5 years overall survival of ICC reported to vary from 15% to 40 % after liver resection[8], strongly depending on the presence of poor


prognostic factors such as lymph nodes involvement, multiple nodules and vascular invasion[9]. Despite the fact that minimally invasive approach to primary and metastatic liver cancer is becoming a routine approach in selected patients, showing improved perioperative outcomes and similar oncological outcomes than open surgery for the treatment of both hepatocellular carcinoma (HCC)[10,11] and colorectal liver metastases (CRLM)[12,13], reports on the minimally invasive treatment of ICC are scanty and no clear evidences on the feasibility, safety and oncological results are currently available. In a recent systematic review on laparoscopic liver resection, published in 2016, among 9527 patients only 116 underwent laparoscopic hepatectomy for ICC[14]. These data strongly reflect the reluctance, even in highly specialized centers, to embrace the minimally invasive approach for ICC. This is probably connected to the necessity of performing loco-regional lymphadenectomy, which is a technically demanding procedure to perform by a minimally invasive approach, and it is also due to the fact that ICC treatment often requires major hepatectomies or vascular and/or biliary reconstruction to achieve a R0 resection. In addition, the Southampton guidelines consensus, despite strongly supporting the adoption of the laparoscopic approach for both HCC and CRLM, did not address the role of minimally invasive approach for the surgical management of ICC[15]. Therefore, updates on the current evidences on the minimally invasive treatment of ICC are urgently needed. The aim of this study is to review and summarize the current evidences on the topic.

MATERIALS AND METHODS

The present study was accomplished in accordance with the preferred reporting Items for systematic reviews and meta-analyses (PRISMA) guidelines[16]. A systematic review of the literature, based on criteria predetermined by the investigators, was independently performed by two authors (B.A. and P.R.) from the 1st of January 2009 up to the 1st of January 2021 in 4 databases (PubMed, Scopus, Google Scholar, and Cochrane databases) in order to maximize articles capturing. Discrepancy in data collection, synthesis and analysis were solved by consensus of all authors. All retrospective and prospective studies reporting on the comparative outcomes of open vs minimally invasive treatment of ICC were included. Search terms included: "cholangiocarcinoma", "intrahepatic", "laparoscopic", "surgery", "minimally invasive", "robotic surgery" "biliary neoplasm", "liver resection" and "hepatectomy".

The following Inclusion and exclusion criteria were applied.

Inclusion criteria

(1) English language studies including patients with histologically proved ICC; (2) Use of a minimally invasive surgical approach (laparoscopic or robotic) for liver resection of ICC; (3) Comparing open surgery to minimally invasive surgery (laparoscopic or robotic) for the surgical treatment of ICC; and (4) Studies reporting on at least one intraoperative, postoperative, and long-term oncological outcomes (operative time, intraoperative complications, estimated blood loss, blood transfusion rate, length of stay, R0 resection rate, lymph nodes retrieval, postoperative morbidity and mortality rate, disease free and overall survival rates).

Exclusion criteria

(1) Non-English studies; (2) Animal studies; (3) Non-comparative studies; (4) Abstracts, expert opinions, editorials, meta-analysis, reviews, and letter to the editors; (5) Studies reporting inadequate clinical data; and (6) Studies including mixed pathologies besides ICC; The evaluation of manuscript quality was conducted using the Methodological Index for Non-Randomized Studies criteria^[17] and the Newcastle-Ottawa Scale^[18] to assess the quality of nonrandomized studies in meta-analyses because of the non-randomized nature of selected papers.

RESULTS

Study inclusion

After systematic search 4835 manuscripts were selected for initial screening. Among them 1704 papers were duplicates and therefore excluded. Based on title, abstract and keywords, the Authors selected and analyzed the full-text version of 189 papers. Main reasons for the exclusion were the absence of patients treated both with laparoscopic



and open approach (n = 114) and the inclusion of other types of tumors besides ICC (n= 36). Further causes of exclusion were population treated with palliative intent or case series or absence of specific data on the post-operative outcomes. Two studies selected after full-text analysis were then excluded because more recent studies from the same authors presented additional updated data. This led to the final selection of 9 studies which fulfilled the inclusion criteria[19-27]. The search strategy flow diagram is shown in Figure 1. There were no randomized clinical studies found. All 9 selected papers were retrospective comparative studies and 7 of them were single center series, one a bi-institutional analysis^[25] and one was based on data from a national database^[23]. Geographical distribution of the selected papers was as follows: Italy and United Kingdom (1), United States (1), Germany (1), Japan (1), Korea (2) and China (3). Characteristics of the included manuscripts and their quality assessment are summarized in Table 1.

Among all the 3012 included patients 2450 were operated by an open approach and 562 by a minimally invasive (laparoscopic) approach.

Baseline characteristics

As regards patients' baseline characteristics no statistically significant differences were detected in terms of age, sex, body mass index and American Society of Anaesthesiologists score between laparoscopic and open groups in all manuscripts. Eight studies [20-22,24-28] analyzed the presence of at least one comorbidity and no statistical difference was reported between laparoscopic and open group. Detailed data are reported in Table 2.

Tumor characteristics

Tumor size was reported in all, except one[28], of the analyzed studies and in the study by Martin et al^[23] a statistically significant difference between groups was highlighted with a smaller tumor diameter in the laparoscopic group when compared to the open group. Seven of the selected manuscripts [20-22,24-26,28] reported data on preoperative tumors, nodes and metastasis (TNM) staging and CA19.9 values with no differences between groups. CEA preoperative values were analyzed only in four studies[20,24,25,28] and no differences were found. Zhu *et al*[22], Ratti *et al*[25] and Kang *et al*[26] reported a smaller tumor size in the laparoscopic group but this difference was adjusted after propensity score matching. Kinoshita *et al*[24] found no difference in mean tumor size between the two groups but a higher percentage of patients in the open group had tumors bigger than 3 cm when compared to the laparoscopic group (71% vs 33%). Two[21,22] of the analyzed studies were focused on large (> 3 cm) or multinodular ICCs. All tumors characteristics were resumed in Table 3.

Operative outcomes

Operative time was analyzed in 8 out of 9 analyzed studies and only in the study by Zhu *et al*[22] there was a statistically significant difference in favor of the laparoscopic group. Intraoperative blood loss was reported by 7 studies and a statistically significant lower blood loss was found in the laparoscopic group in 4 of them[20,24,25, 28]

With the exception of the national database based study by Martin *et al*^[23], data on postoperative morbidity were reported in all manuscripts and a lower incidence of postoperative complications in the laparoscopic group was found in the studies by Ratti et al^[25] and by Haber et al^[27].

Laparoscopic approach significantly decreased postoperative hospital stay in four of seven study[25-28]. Days spent in intensive care unit were analyzed only by two studies[25,27] with no differences between open and laparoscopic approach.

As regards the type of liver resection, a statistically significant higher rate of major hepatectomies was reported in the open groups in the studies by Kang et al[26], Martin et al^[23] and Lee et al^[20]. Accomplishment of lymph nodes dissection was investigated by all analyzed studies and in 3 of them [23,25,26] was reported a higher rate of lymph nodes clearance in the open group but with no difference in nodal status. Six authors reported histopathological margin data with no difference between R0 and R1 in the two surgical approaches[20-23,25,26].

Detailed data are reported in Tables 2 and 3.

Oncological outcomes

Eight of the selected studies [20-22,24-28] reported comparative data on the oncological outcomes expressed as overall and disease free survival and none of them reported any differences between the open and the laparoscopic group. In the study by Martin



Table 1 Study characteristics and quality assessment										
Ref.	Country	Type of study		LS	OS	NOS				
						Selection	Selection Comparability Outcame/Exposure			
Wu et al[<mark>28</mark>], 2020	China	RetS-SC	Case control study	18	25	***	*	***	17	
Haber <i>et al</i> [27], 2020	Germany	RetS-SC	Case control study	27	31	***	*	**	16	
Kang <i>et al</i> [<mark>26</mark>], 2020	Korea	RetS-SC	Propensity score matching	30	61	***	**	***	18	
Kinoshita <i>et al</i> [24], 2020	Japan	RetS-SC	Case control study	15	21	***	**	***	18	
Ratti <i>et al</i> [<mark>25</mark>], 2020	United Kingdom- Italy	RetS-TC	Propensity score matching	104	104	***	**	***	19	
Martin <i>et al</i> [23], 2019	United States	RetS-DB	Database	312	1997	**	*	**	15	
Zhu <i>et al</i> [<mark>22</mark>], 2019	China	RetS-SC	Propensity score matching	20	63	***	**	***	19	
Wei <i>et al</i> [<mark>21</mark>], 2017	China	RetS-SC	Case control study	30	20	***	**	***	19	
Lee <i>et al</i> [<mark>20</mark>], 2016	Korea	RetS-SC	Case control study	14	23	***	**	***	20	

LS: Number of patients treated with laparoscopic surgery; OS: Number of patients treated with open surgery; NOS: Newcastle-Ottawa Scale for assessing the quality of nonrandomised studies. A study can be awarded a maximum of one star (*) for each numbered item within the selection and exposure categories. A maximum of two stars can be given for comparability. MINORS: Methodological index for non-randomized studies; RetS: Retrospective study; SC: Single center; TC: Two centres; DB: Data base.

> et al[23] the authors focused electively on the rate of administration of adjuvant treatments and found no differences related to the surgical approach.

> As regards specific variables affecting survival Wu et al[28] identified high preoperative values of CA19.9, high TNM stage and a poor tumor differentiation as independent risk factor for worst overall survival (OS) and disease-free survival (DFS) while Kang et al^[26] identified tumor size, nodularity and perineural invasion as independent factors correlated to lower DFSs.

> Kinoshita *et al*[24], instead, found tumor size (diameter \geq 3 cm), presence of vascular invasion and a high CA19.9 levels on preoperative exams to be associated with a poorer OS.

> Lee et al[20], trying to avoid bias, analyzed OS and RFS in laparoscopic liver resection and open liver resection for all patients by stratifying them by the accomplishment of lymph nodes dissection and found no difference in between groups.

> Finally, the pattern of recurrence was investigated only in 3 of the selected manuscripts[20,25,28] with no statistically significant differences between the open and the laparoscopic approach. Detailed data are reported in Table 3.

DISCUSSION

The current systematic review is focused on the comparative outcomes of open vs minimally invasive resection of ICC. In fact, even if laparoscopy proved to be an effective option for the treatment of both HCC and CRLM, offering the benefit of minimally invasiveness without compromising the oncological outcomes, reports on the operative and oncological outcomes of minimally invasive treatment of ICC are scanty and seldom reported. The uncommon adoption of the laparoscopic or robotic approach for ICC is related to various oncological and technical reasons. First, ICC has a relative low incidence when compared to others liver malignancies and due to its aggressive biological behavior is often diagnosed at an advanced stage not suitable for radical surgery which remains the only potentially curative treatment option[1]. Second, surgery for ICC is often characterized by a high degree of technical difficulty



WJGO https://www.wjgnet.com

Table 2 Preoperative and surgical data													
Ref.	SA	NP	AGE	ASA					Lymphadenectomy		AT		00111/
				I	II	III	IV	— мауогн	Yes	Number	- 01	IOBL	CONV.
Wu et al[28], 2020	OS	25	61	19		6		13 (52%)	6 (32%)	6	300 (257-392)	500 (350-750)	N/A
	LS	18	64	15		3		6 (33%)	8 (33%)	6	305 (207-390)	375 (275-500)	0
Haber <i>et al</i> [27], 2020	OS	31	63	1	21	8	1	24 (78%)	29 (94%)	8	282 (112–947)	/	N/A
	LS	27	69	0	15	12	0	19 (70%)	23 (85%)	8	314 (125-439)	/	2
Kang <i>et al</i> [26], 2020	OS	61	68	/	/	/	/	53 (88.3%)	46 (75.4%)	/	343.2 ± 106.0	979.3 ± 864.4	N/A
	LS	30	65	/	/	/	/	20 (66.7%)	9 (30%)	/	375.2 ± 204.0	1396.7 ± 2568.9	6
Kinoshita <i>et al</i> [<mark>24</mark>], 2020	OS	21	68	/	/	/	/	15	7 (33%)	3	358 (150-634)	500 (105-3710)	N/A
	LS	15	65	/	/	/	/	5	6 (40%)	2	360 (221-802)	150 (20-2500)	0
Ratti et al[25], 2020	OS	209	62	20	58	26	0	38 (36.5%)	92 (88.5%)	7 (5–14)	230 ± 60	350 ± 250	N/A
	LS	114	60	22	56	26	0	35 (33.7%)	87 (83.7%)	8 (5-11)	270 ± 65	150 ± 100	0
Martin <i>et al</i> [23], 2019	OS	1997	64	/	/	/	/	1338 (67%)	1210 (61.2%)	/	/	/	N/A
	LS	312	65	/	/	/	/	137 (44%)	312 (38.5%)	/	/	/	/
Zhu <i>et al</i> [<mark>22</mark>], 2019	OS	63	56	/	/	/	/	43 (68.3%)	27 (42.9%)	/	200 (140-320)	400 (50-2000)	N/A
	LS	20	54	/	/	/	/	11 (55%)	8 (40%)	/	225 (140-400)	200 (50-1000)	2
Wei <i>et al</i> [21], 2017	OS	20	60.5	/	/	/	/	11 (55%)	11 (55%)	/	230 (125-420)	350 (50-1200)	N/A
	LS	12	61.5	/	/	/	/	7 (58.3%)	4 (33%)	/	212.5 (60–500)	350 (30-2000)	0
Lee <i>et al</i> [20], 2016	OS	23	59	0	20	2	1	19 (82.6%)	15 (65.2%)	6 (1-16)	330.0 (140-590)	625 (250-2500)	N/A
	LS	14	66	0	12	2	0	7 (50%)	5 (35.7%)	4 (1-12)	255.0 (140-480)	325 (10-1500)	0

Results for each Author are represented divided in two lines: Open surgery and laparoscopic surgery. LS: Laparoscopic surgery; OS: Open surgery; SA: Surgical approach; NP: Number of patients; Age are expressed in year; ASA: American Society of Anaesthesiologists physical status classification; MajorH: Major hepatectomy considered as equal or more than 3 resected segments; OT: Operation time expressed in minutes; IOBL: Intra-operative blood loss expressed in mL; Conv: Number of procedure converted from laparoscopic to open approach. In bold differences with a *P* value < 0.05.

associated with the need of performing an appropriate lymphadenectomy and, especially in centrally located tumors, a vascular or biliary reconstruction as well as a major hepatic resection are often needed to achieve clear surgical margins[29]. These technical issues have probably slowed down the diffusion of ICC as a valid indication for a minimally invasive approach. In fact, major hepatectomies, hepatic hilum

Table 3 Post-operative and oncological data											
				НМ		30-d morbi	dity				
Ref.	SA	ICUS	HS	R0	R1	Grade I-II	Grade III- IV	90-d morbidity	mFU	OS	DFS
Wu et al[<mark>28</mark>], 2020	OS	/	9 (7-15)	/	/	23	2	1	/	20	4
	LS	/	6 (5-12)	/	/	17	1	0	/	47.1	0
Haber <i>et al</i> [27], 2020	OS	1 (0-6)	12 (5-33)	23	8	8	10	0	/	/	/
	LS	1 (0-81)	10 (3-94)	24	3	3	5	2	/	/	/
Kang et al[<mark>26</mark>], 2020	OS	/	18.3 ± 14.7	/	/	/	23	0	16.8	81.2	42.5
	LS	/	9.8 ± 5.1	/	/	/	8	0	39.2	76.7	65.6
Kinoshita <i>et al</i> [<mark>24</mark>], 2020	OS	/	/	20	1	/	4	/	/	36	19
	LS	/	/	14	1	/	2	/	/	32	24
Ratti <i>et al</i> [<mark>25</mark>], 2020	OS	4 (3-10)	6 (3-21)	99	5	17	8	2	50	47	34
	LS	3 (1-5)	4 (2-10)	101	3	11	4	1	39	46	36
Martin <i>et al</i> [<mark>23</mark>], 2019	OS	/	/	1451	546	/	/	/	/	/	/
	LS	/	/	247	65	/	/	/	/	/	/
Zhu et al <mark>[22]</mark> , 2019	OS	/	7 (3-33)	58	5	22	6	0	24	17	32
	LS	/	6 (3-9)	19	1	3	1	0	24	21	31
Wei <i>et al</i> [<mark>21</mark>], 2017	OS	/	11 (5-30)	19	2	10	3	0	12	32.7	27.9
	LS	/	14 (6-23)	12	0	3	2	0	17.5	56.3	43.8
Lee <i>et al</i> [<mark>20]</mark> , 2016	OS	/	20 (9-63)	/	/	1	4	0	/	75.7	/
	LS	/	15 (9–29)	/	/	0	3	0	/	84.6	/

Results for each Author are represented divided in two lines: Open surgery and laparoscopic surgery. LS: Laparoscopic surgery; OS: Open surgery; SA: Surgical approach; ICUS: Intensive care unit stay in days; HS: Hospital stay in days; HM: Histopathological margins; mFU: Median follow-up in months; OS: Overall survival expressed in months after surgery; DFS: Disease-free survival expressed in months after surgery. In bold differences with a P value < 0.05

> lymphadenectomy and biliary reconstructions are technically demanding to perform by a minimally invasive approach. In addition, to safely perform such procedures an extensive learning curve is needed[30] and since now this has been unlikely to be accomplished outside high volume centers with a steady commitment to minimal invasiveness. Notwithstanding that, recently initial data on the comparative outcomes of open vs minimally invasive resection of ICC have been published in the literature. The interest on this topic is, in fact, increasing and the surgical treatment of ICC is becoming one of the latest field of implementation of minimally invasive liver surgery. In particular, all the selected articles for this systematic review have been published in the last 5 years thus reflecting the growing interest on the topic. Nevertheless, despite the accurate search strategy applied, the current systematic review confirmed the paucity of current evidences on the minimally invasive approach for ICC. No randomized comparative studies are currently available and only 9 comparative retrospective studies were retrieved from the systematic search. Although representative of the experience of few highly specialized centers for minimally invasive liver surgery, the analyzed studies proved without doubt the feasibility and safety of the laparoscopic approach to ICC in patients with a tumor diameter < 5 cm, without main biliary duct invasion, without large vascular invasion and in which biliary and vascular reconstructions were not needed. Results from the analyzed studies also confirmed the typical benefit of minimally invasiveness already demonstrated for the laparoscopic treatment of HCC and CRLM, even when dealing with ICC. In fact, several of the analyzed studies reported a benefit of the minimally invasive approach in terms of peri-operative outcomes when compared to the open approach. In details, four studies [20,24,25,28] reported a lower intraoperative blood loss associated with the minimally invasive approach even when dealing with radical lymph nodes clearance and this is probably related to magnified view and the meticulous dissection



Patrone R et al. Minimally invasive surgical treatment of ICC



Figure 1 PRISMA flow-chart. Figure reported the diagram of our systematic review of the literature, performed in 4 databases from Jan 1, 2009 up to Jan 1, 2021. Search terms included: "cholangiocarcinoma", "intrahepatic", "laparoscopic", "surgery", "minimally invasive", "robotic surgery" "biliary neoplasm", "liver resection" and "hepatectomy". Inclusion criteria are in the big circle-box. Major reasons for exclusion were the absence of patients treated both with laparoscopic and open approach (n = 114) and the inclusion of other tumor types besides intrahepatic cholangiocarcinoma (n = 36). Further reasons for exclusion were population treated with palliative intent or case series or absence of specific data on the post-operative outcomes. This led to the final selection of 9 studies which fulfilled the inclusion criteria. ICC: Intrahepatic cholangiocarcinoma.

> achievable by laparoscopy. Nevertheless, Kang et al[26] described a higher blood loss rate in the laparoscopic group, but these difference was not statistically significant (P value = 0.393). Furthermore, four of the analyzed studies[25-28] reported a shorter hospital stay associated to the laparoscopic approach, thus confirming the benefit of minimal invasiveness in terms of a faster recovery also in this setting. Finally, despite the relative initial experience, the studies by Ratti et al^[25] and Haber et al^[27] highlighted a benefit in terms of postoperative morbidity in favor of laparoscopy. However, the reported experiences are mainly focused on mass forming type ICC without vascular and biliary involvement (away from the liver plate) and, as highlighted by the large national database-based study by Martin *et al*^[23], patients operated by laparoscopy had smaller tumor size when compared to those submitted to an open resection. In addition, a statistically significant higher rate of major hepatectomies was reported in the open groups in 3 of the analyzed studies[20,23,26]. This reflects the selection bias, which is to be expected when dealing with the appliance of laparoscopy to a new surgical indication. Indeed, the studies by Zhu *et al*[22] and Wei et al^[21] were focused on large or multinodular ICCs and both confirmed positive results similar to those reported by studies with stricter selection criteria. The benefit of performing a lymphadenectomy for ICC is a debated issue. In fact, up to 40% of



WJGO | https://www.wjgnet.com

resected patients can present with lymph nodes involvement[9] and several authors have highlighted a survival benefit in patients undergoing lymph nodes clearance associated to liver resections when compared to patients who did not[31]. On the contrary, discrepant studies reported no survival benefit and an increase in surgical morbidity associated with lymphadenectomy especially in case of patients with chronic liver disease[32,33]. Nevertheless, lymph nodes clearance for ICC is a crucial strategy for a correct staging of surgically resected patients and can both guides the administration of adjuvant chemotherapy and optimizes clinical risk stratification and prognostic outcomes. This factor is even more significant if we take into account the results of the BILCAP study which demonstrated the survival benefit of adjuvant gemcitabine for biliary tract cancers[34]. Indeed, the 8th edition of the American Joint Committee on Cancer (AJCC) guidelines recommends to perform lymphadenectomy with an optimal cut-off of six retrieved nodes for biliary tract cancers[35]. Is therefore to be expected that regional lymphadenectomy will be implemented in clinical practice and should be performed irrespectively from the open or minimally invasive surgical approach adopted. From the current systematic review, a certain under-employment of regional lymphadenectomy for ICC was highlighted. In fact, a lower rate of lymph nodes dissection in the laparoscopic group was reported in the studies by Kang et al [26] and Ratti et al[25]. These data are confirmed by the National Cancer Database analysis by Martin et al[23] which also highlighted that some form of nodal dissection was performed in only 58% of patients in the whole study cohort. Indeed, the vast majority of the published studies reports the initial experiences of selected high specialized centers and refers to a time preceding the AJCC guidelines diffusion and application. Therefore, after an initial learning curve, a major adherence to the guidelines it is likely to be accomplished. It is also to be expected that the accumulation of experience and the improvement of surgical techniques will probably promote the adoption of the minimally invasive approach for ICC.

In addition, the histopathological margin status is a crucial factor to be considered when comparing the minimally invasive approach to the standard open resection. In fact, an R0 margin represents the most significant predicting factor of oncological outcomes and results from our review show a superimposable rate of negative surgical margin in both approaches. This evidence together with the appropriateness of locoregional lymphadenectomy and the reduced intraoperative blood loss reported in the majority of the analyzed studies, allow us to consider the laparoscopic approach non inferior to the open one in terms of operative outcomes. Therefore, is not surprising that the minimally invasive approach has been recently extended to the surgical treatment of hilar type cholangiocarcinoma[36] and gallbladder cancer[37,38]. These encouraging pivotal experiences seem to demonstrate the feasibility of minimally invasive surgery in a setting often requiring the completion of a major hepatic resection in association with loco-regional lymphadenectomy and the challenge of biliary reconstructions. It is therefore likely that in the very next future the surgical research in the field of minimally invasive surgery (MIS) for biliary cancer will be concentrated on hilar type tumors and on biliary duct resection (with the aid of Indocianyne green guidance) and reconstruction via duct to duct anastomosis or hepatico-jejunostomy. In addition, the implementation of the MIS approach for the surgical treatment of ICC is likely to be promoted by the diffusion of the robotic platforms. In fact, even if it has been demonstrated by the analyzed studies that an appropriate lymphadenectomy can be performed safely and effectively by laparoscopy, it requires advanced laparoscopic skills and a long learning curve. The application of the robotic platform in this setting, thanks to the higher dexterity achievable with the robotic instruments, which, with the endowrist system, have seven degrees of freedom, could facilitate an adequate surgical manipulation and the achievement of an appropriate lymph node clearance in a confined space such as the hepatic pedicle. The magnified high-resolution 3d stereoscopic view offered by the robotic platform is also an added value in defining the anatomical structures and can facilitate biliary reconstructions when needed. As regards the oncologic outcomes, the data are scanty and not conclusive. Some form of oncological data has been reported only by eight studies[20-22,24-28] and, even though no differences have been reported in terms of disease free survival and overall survival in this systematic review, a recent meta-analysis highlighted a possible trend towards a lower 5 years overall survival for patients treated with a laparoscopic approach for ICC when compared to those operated by open approach[39]. Therefore, the interpretation of the oncologic outcomes needs to be evaluated with extreme caution. In addition, no high quality evidences are currently available and thus the need for more qualified data is urgent.

Balahidena® WJGO | https://www.wjgnet.com

CONCLUSION

In conclusion, the minimally invasive treatment of ICC is currently rarely performed but is rapidly gaining popularity. Currently available data seems to justify the implementation of the minimally invasive approach for ICC by demonstrating its safety and reproducibility and by confirming the well-known advantages of minimally invasiveness in term of perioperative outcomes also in this setting, as already proven for other liver neoplasms. Nevertheless, current evidences are based on few studies with a limited sample size and a short follow-up. In addition, selection criteria for the minimal invasive approach were highly restrictive (small tumors, generally < 3 cm, distant from the hilum and not requiring a biliary reconstruction) when compared to open series and, therefore, at high risk for selection bias. Dedicated study protocols and analysis of national and international registries are urgently needed to clarify the real role of minimally invasive surgery in the treatment of ICC and its impact on the long term oncologic outcomes.

ARTICLE HIGHLIGHTS

Research background

Intrahepatic cholangiocarcinoma represents a very aggressive tumor with poor prognosis. Nowadays surgical open approach is still the gold standard treatment but minimally invasive surgery is gaining an important role. No randomized trials are available on this topic in scientific literature.

Research motivation

Our scientific group aim to contribute to the development of the scientific research on hepatobiliary minimally invasive surgery.

Research objectives

Our research had the objective to summarize and review the scientific evidences present in the literature on minimally invasive surgical approach for intrahepatic cholangiocarcinoma.

Research methods

We performed a systematic review of the literature between 01/01/2009 and 01/01/2021. Our research keywords were: "cholangiocarcinoma", "intrahepatic", "laparoscopic", "surgery", "minimally invasive", "robotic surgery" "biliary neoplasm", "liver resection" and "hepatectomy". We selected only papers comparing open and laparoscopic approach and reporting at least one intraoperative, postoperative or oncological outcomes.

Research results

We found 9 papers that fulfilled all inclusion criteria reporting data from 3012 patients with no differences in baseline characteristic. Almost all operative outcomes were in favor of laparoscopic groups (blood losses, operative time, hospital stay, post-operative complications) except for the number of lymphonodes retrieved (higher number of lymphonodes retrieved in the open groups). No statistical differences in oncological outcomes were reported.

Research conclusions

Our research demonstrates that very few studies investigated the role of minimally invasive surgery for intrahepatic cholangiocarcinoma. Currently available data in the Literature were not consistent enough to consider the laparoscopic approach to ICC as a standard of care but a steady implementation is likely to be realized in the next future.

Research perspectives

It is likely that soon the diffusion of robotic surgery and tailored surgery, will promote the diffusion of minimally invasive approach for intrahepatic cholangiocarcinoma and will help elucidating its role and the oncological outcomes.

Zaishidenq® WJGO | https://www.wjgnet.com

ACKNOWLEDGEMENTS

The authors would like to thank Assunta Zazzaro for her appreciated work and team coordination.

REFERENCES

- Khan SA, Tavolari S, Brandi G. Cholangiocarcinoma: Epidemiology and risk factors. Liver Int 2019; 1 39 Suppl 1: 19-31 [PMID: 30851228 DOI: 10.1111/liv.14095]
- 2 Bridgewater J, Galle PR, Khan SA, Llovet JM, Park JW, Patel T, Pawlik TM, Gores GJ. Guidelines for the diagnosis and management of intrahepatic cholangiocarcinoma. J Hepatol 2014; 60: 1268-1289 [PMID: 24681130 DOI: 10.1016/j.jhep.2014.01.021]
- Patel T. Increasing incidence and mortality of primary intrahepatic cholangiocarcinoma in the United 3 States. Hepatology 2001; 33: 1353-1357 [PMID: 11391522 DOI: 10.1053/jhep.2001.25087]
- Wu L, Tsilimigras DI, Paredes AZ, Mehta R, Hyer JM, Merath K, Sahara K, Bagante F, Beal EW, Shen F, Pawlik TM. Trends in the Incidence, Treatment and Outcomes of Patients with Intrahepatic Cholangiocarcinoma in the USA: Facility Type is Associated with Margin Status, Use of Lymphadenectomy and Overall Survival. World J Surg 2019; 43: 1777-1787 [PMID: 30820734 DOI: 10.1007/s00268-019-04966-4]
- Antwi SO, Mousa OY, Patel T. Racial, Ethnic, and Age Disparities in Incidence and Survival of Intrahepatic Cholangiocarcinoma in the United States; 1995-2014. Ann Hepatol 2018; 17: 604-614 [PMID: 29893702 DOI: 10.5604/01.3001.0012.0929]
- 6 Nakeeb A, Pitt HA, Sohn TA, Coleman J, Abrams RA, Piantadosi S, Hruban RH, Lillemoe KD, Yeo CJ, Cameron JL. Cholangiocarcinoma. A spectrum of intrahepatic, perihilar, and distal tumors. Ann Surg 1996; 224: 463-473; discussion 473 [PMID: 8857851 DOI: 10.1097/0000658-199610000-00005
- Hyder O, Hatzaras I, Sotiropoulos GC, Paul A, Alexandrescu S, Marques H, Pulitano C, Barroso E, 7 Clary BM, Aldrighetti L, Ferrone CR, Zhu AX, Bauer TW, Walters DM, Groeschl R, Gamblin TC, Marsh JW, Nguyen KT, Turley R, Popescu I, Hubert C, Meyer S, Choti MA, Gigot JF, Mentha G, Pawlik TM. Recurrence after operative management of intrahepatic cholangiocarcinoma. Surgery 2013; 153: 811-818 [PMID: 23499016 DOI: 10.1016/j.surg.2012.12.005]
- Doussot A, Groot-Koerkamp B, Wiggers JK, Chou J, Gonen M, DeMatteo RP, Allen PJ, Kingham 8 TP, D'Angelica MI, Jarnagin WR. Outcomes after Resection of Intrahepatic Cholangiocarcinoma: External Validation and Comparison of Prognostic Models. J Am Coll Surg 2015; 221: 452-461 [PMID: 26206643 DOI: 10.1016/j.jamcollsurg.2015.04.009]
- de Jong MC, Nathan H, Sotiropoulos GC, Paul A, Alexandrescu S, Marques H, Pulitano C, Barroso E, Clary BM, Aldrighetti L, Ferrone CR, Zhu AX, Bauer TW, Walters DM, Gamblin TC, Nguyen KT, Turley R, Popescu I, Hubert C, Meyer S, Schulick RD, Choti MA, Gigot JF, Mentha G, Pawlik TM. Intrahepatic cholangiocarcinoma: an international multi-institutional analysis of prognostic factors and lymph node assessment. J Clin Oncol 2011; 29: 3140-3145 [PMID: 21730269 DOI: 10.1200/JCO.2011.35.6519]
- 10 Belli A, Fantini C, Cioffi L, D'Agostino A, Belli G. Mils for HCC: the state of art. Updates Surg 2015; 67: 105-109 [PMID: 26164139 DOI: 10.1007/s13304-015-0316-1]
- 11 Morise Z, Aldrighetti L, Belli G, Ratti F, Belli A, Cherqui D, Tanabe M, Wakabayashi G; ILLS-Tokyo Collaborator group. Laparoscopic repeat liver resection for hepatocellular carcinoma: a multicentre propensity score-based study. Br J Surg 2020; 107: 889-895 [PMID: 31994182 DOI: 10.1002/bjs.11436]
- 12 Fretland ÅA, Dagenborg VJ, Bjørnelv GMW, Kazaryan AM, Kristiansen R, Fagerland MW, Hausken J, Tønnessen TI, Abildgaard A, Barkhatov L, Yaqub S, Røsok BI, Bjørnbeth BA, Andersen MH, Flatmark K, Aas E, Edwin B. Laparoscopic Versus Open Resection for Colorectal Liver Metastases: The OSLO-COMET Randomized Controlled Trial. Ann Surg 2018; 267: 199-207 [PMID: 28657937 DOI: 10.1097/SLA.00000000002353]
- 13 Ratti F, Fiorentini G, Cipriani F, Catena M, Paganelli M, Aldrighetti L. Laparoscopic vs Open Surgery for Colorectal Liver Metastases. JAMA Surg 2018; 153: 1028-1035 [PMID: 30027220 DOI: 10.1001/jamasurg.2018.2107
- Ciria R, Cherqui D, Geller DA, Briceno J, Wakabayashi G. Comparative Short-term Benefits of 14 Laparoscopic Liver Resection: 9000 Cases and Climbing. Ann Surg 2016; 263: 761-777 [PMID: 26700223 DOI: 10.1097/SLA.000000000001413]
- Abu Hilal M, Aldrighetti L, Dagher I, Edwin B, Troisi RI, Alikhanov R, Aroori S, Belli G, Besselink M, Briceno J, Gayet B, D'Hondt M, Lesurtel M, Menon K, Lodge P, Rotellar F, Santoyo J, Scatton O, Soubrane O, Sutcliffe R, Van Dam R, White S, Halls MC, Cipriani F, Van der Poel M, Ciria R, Barkhatov L, Gomez-Luque Y, Ocana-Garcia S, Cook A, Buell J, Clavien PA, Dervenis C, Fusai G, Geller D, Lang H, Primrose J, Taylor M, Van Gulik T, Wakabayashi G, Asbun H, Cherqui D. The Southampton Consensus Guidelines for Laparoscopic Liver Surgery: From Indication to Implementation. Ann Surg 2018; 268: 11-18 [PMID: 29064908 DOI: 10.1097/SLA.00000000002524
- Moher D, Liberati A, Tetzlaff J, Altman DG; PRISMA Group. Preferred reporting items for 16



systematic reviews and meta-analyses: the PRISMA statement. J Clin Epidemiol 2009; 62: 1006-1012 [PMID: 19631508 DOI: 10.1016/j.jclinepi.2009.06.005]

- 17 Slim K, Nini E, Forestier D, Kwiatkowski F, Panis Y, Chipponi J. Methodological index for nonrandomized studies (minors): development and validation of a new instrument. ANZ J Surg 2003; 73: 712-716 [PMID: 12956787 DOI: 10.1046/j.1445-2197.2003.02748.x]
- Wells GA, Shea B, O'Connell D, Peterson J, Welch V, Losos M, Tugwell P. The Newcastle-Ottawa 18 Scale (NOS) for assessing the quality of nonrandomised studies in meta-analyses. Available from: http://www.ohri.ca/programs/clinical_epidemiology/oxford.asp
- Uy BJ, Han HS, Yoon YS, Cho JY. Laparoscopic liver resection for intrahepatic cholangiocarcinoma. 19 J Laparoendosc Adv Surg Tech A 2015; 25: 272-277 [PMID: 25789408 DOI: 10.1089/lap.2014.0233]
- 20 Lee W, Park JH, Kim JY, Kwag SJ, Park T, Jeong SH, Ju YT, Jung EJ, Lee YJ, Hong SC, Choi SK, Jeong CY. Comparison of perioperative and oncologic outcomes between open and laparoscopic liver resection for intrahepatic cholangiocarcinoma. Surg Endosc 2016; 30: 4835-4840 [PMID: 26902611 DOI: 10.1007/s00464-016-4817-x]
- Wei F, Lu C, Cai L, Yu H, Liang X, Cai X. Can laparoscopic liver resection provide a favorable 21 option for patients with large or multiple intrahepatic cholangiocarcinomas? Surg Endosc 2017; 31: 3646-3655 [PMID: 28032221 DOI: 10.1007/s00464-016-5399-3]
- 22 Zhu Y, Song J, Xu X, Tan Y, Yang J. Safety and feasibility of laparoscopic liver resection for patients with large or multiple intrahepatic cholangiocarcinomas: A propensity score based casematched analysis from a single institute. Medicine (Baltimore) 2019; 98: e18307 [PMID: 31804378 DOI: 10.1097/MD.00000000018307]
- 23 Martin SP, Drake J, Wach MM, Ruff S, Diggs LP, Wan JY, Brown ZJ, Ayabe RI, Glazer ES, Dickson PV, Davis JL, Deneve JL, Hernandez JM. Laparoscopic Approach to Intrahepatic Cholangiocarcinoma is Associated with an Exacerbation of Inadequate Nodal Staging. Ann Surg Oncol 2019; 26: 1851-1857 [PMID: 30895496 DOI: 10.1245/s10434-019-07303-0]
- 24 Kinoshita M, Kanazawa A, Takemura S, Tanaka S, Kodai S, Shinkawa H, Shimizu S, Murata A, Nishio K, Hamano G, Ito T, Tsukamoto T, Kubo S. Indications for laparoscopic liver resection of mass-forming intrahepatic cholangiocarcinoma. Asian J Endosc Surg 2020; 13: 46-58 [PMID: 30924307 DOI: 10.1111/ases.12703]
- Ratti F, Rawashdeh A, Cipriani F, Primrose J, Fiorentini G, Abu Hilal M, Aldrighetti L. Intrahepatic 25 cholangiocarcinoma as the new field of implementation of laparoscopic liver resection programs. A comparative propensity score-based analysis of open and laparoscopic liver resections. Surg Endosc 2021; **35**: 1851-1862 [PMID: 32342213 DOI: 10.1007/s00464-020-07588-3]
- Kang SH, Choi Y, Lee W, Ahn S, Cho JY, Yoon YS, Han HS. Laparoscopic liver resection versus 26 open liver resection for intrahepatic cholangiocarcinoma: 3-year outcomes of a cohort study with propensity score matching. Surg Oncol 2020; 33: 63-69 [PMID: 32561101 DOI: 10.1016/j.suronc.2020.01.001
- Haber PK, Wabitsch S, Kästner A, Andreou A, Krenzien F, Schöning W, Pratschke J, Schmelzle M. 27 Laparoscopic Liver Resection for Intrahepatic Cholangiocarcinoma: A Single-Center Experience. J Laparoendosc Adv Surg Tech A 2020; 30: 1354-1359 [PMID: 32503376 DOI: 10.1089/lap.2020.0215]
- Wu J, Han J, Zhang Y, Liang L, Zhao J, Han F, Dou C, Liu J, Wu W, Hu Z, Zhang C. Safety and 28 feasibility of laparoscopic versus open liver resection with associated lymphadenectomy for intrahepatic cholangiocarcinoma. Biosci Trends 2020; 14: 376-383 [PMID: 32921695 DOI: 10.5582/bst.2020.032931
- Ribero D, Pinna AD, Guglielmi A, Ponti A, Nuzzo G, Giulini SM, Aldrighetti L, Calise F, Gerunda 29 GE, Tomatis M, Amisano M, Berloco P, Torzilli G, Capussotti L; Italian Intrahepatic Cholangiocarcinoma Study Group. Surgical Approach for Long-term Survival of Patients With Intrahepatic Cholangiocarcinoma: A Multi-institutional Analysis of 434 Patients. Arch Surg 2012; 147: 1107-1113 [PMID: 22910846 DOI: 10.1001/archsurg.2012.1962]
- 30 Krenzien F, Schöning W, Brunnbauer P, Benzing C, Öllinger R, Biebl M, Bahra M, Raschzok N, Cherqui D, Geller D, Han HS, Wakabayashi G, Schmelzle M, Pratschke J; study group of the International Laparoscopic Liver Society (ILLS). The ILLS Laparoscopic Liver Surgery Fellow Skills Curriculum. Ann Surg 2020; 272: 786-792 [PMID: 32833753 DOI: 10.1097/SLA.000000000004175]
- 31 Kim SH, Han DH, Choi GH, Choi JS, Kim KS. Oncologic Impact of Lymph Node Dissection for Intrahepatic Cholangiocarcinoma: a Propensity Score-Matched Study. J Gastrointest Surg 2019; 23: 538-544 [PMID: 30112702 DOI: 10.1007/s11605-018-3899-2]
- 32 Zhou R, Lu D, Li W, Tan W, Zhu S, Chen X, Min J, Shang C, Chen Y. Is lymph node dissection necessary for resectable intrahepatic cholangiocarcinoma? HPB (Oxford) 2019; 21: 784-792 [PMID: 30878490 DOI: 10.1016/j.hpb.2018.12.011]
- Bagante F, Spolverato G, Weiss M, Alexandrescu S, Marques HP, Aldrighetti L, Maithel SK, 33 Pulitano C, Bauer TW, Shen F, Poultsides GA, Soubrane O, Martel G, Groot Koerkamp B, Guglielmi A, Itaru E, Ruzzenente A, Pawlik TM. Surgical Management of Intrahepatic Cholangiocarcinoma in Patients with Cirrhosis: Impact of Lymphadenectomy on Peri-Operative Outcomes. World J Surg 2018; 42: 2551-2560 [PMID: 29299649 DOI: 10.1007/s00268-017-4453-1]
- Primrose JN, Fox RP, Palmer DH, Malik HZ, Prasad R, Mirza D, Anthony A, Corrie P, Falk S, 34 Finch-Jones M, Wasan H, Ross P, Wall L, Wadsley J, Evans JTR, Stocken D, Praseedom R, Ma YT, Davidson B, Neoptolemos JP, Iveson T, Raftery J, Zhu S, Cunningham D, Garden OJ, Stubbs C,



Valle JW, Bridgewater J; BILCAP study group. Capecitabine compared with observation in resected biliary tract cancer (BILCAP): a randomised, controlled, multicentre, phase 3 study. Lancet Oncol 2019; 20: 663-673 [PMID: 30922733 DOI: 10.1016/S1470-2045(18)30915-X]

- Zhu AX, Pawlik TM, Kooby DA, Schefter TE, Vauthey JN. AJCC Cancer Staging Manual. 8th ed. 35 New York: Springer International, 2017
- 36 Cipriani F, Ratti F, Fiorentini G, Reineke R, Aldrighetti L. Systematic review of perioperative and oncologic outcomes of minimally-invasive surgery for hilar cholangiocarcinoma. Updates Surg 2021; 73: 359-377 [PMID: 33615423 DOI: 10.1007/s13304-021-01006-6]
- Vega EA, De Aretxabala X, Qiao W, Newhook TE, Okuno M, Castillo F, Sanhueza M, Diaz C, 37 Cavada G, Jarufe N, Munoz C, Rencoret G, Vivanco M, Joechle K, Tzeng CD, Vauthey JN, Vinuela E, Conrad C. Comparison of oncological outcomes after open and laparoscopic re-resection of incidental gallbladder cancer. Br J Surg 2020; 107: 289-300 [PMID: 31873948 DOI: 10.1002/bjs.11379]
- 38 Belli A, Patrone R, Albino V, Leongito M, Piccirillo M, Granata V, Pasta G, Palaia R, Izzo F. Robotic surgery of gallbladder cancer. Mini-invasive Surg 2020; 4: 77 [DOI: 10.20517/2574-1225.2020.70]
- 39 Regmi P, Hu HJ, Paudyal P, Liu F, Ma WJ, Yin CH, Jin YW, Li FY. Is laparoscopic liver resection safe for intrahepatic cholangiocarcinoma? Eur J Surg Oncol 2021; 47: 979-989 [PMID: 33339638 DOI: 10.1016/j.ejso.2020.11.310]



C D WŨ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2216-2218

DOI: 10.4251/wjgo.v13.i12.2216

ISSN 1948-5204 (online)

LETTER TO THE EDITOR

Gender differences in the relationship between alcohol consumption and gastric cancer risk are uncertain and not well-delineated

Henu Kumar Verma, LVKS Bhaskar

ORCID number: Henu Kumar Verma 0000-0003-1130-8783; LVKS Bhaskar 0000-0003-2977-6454.

Author contributions: Verma HK and Bhaskar LVKS both the author wrote and revised the letter.

Conflict-of-interest statement: The authors declare no conflict of interest.

Country/Territory of origin: Italy

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): D Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works

Henu Kumar Verma, Department of Immunopathology, Institute of lungs Biology and Disease, Comprehensive Pneumology Center, Helmholtz Zentrum, Neuherberg 85764, Munich, Germany

LVKS Bhaskar, Department of Zoology, Guru Ghasidas Vishwavidyalaya, Bilaspur 495001, Chhattisgarh, India

Corresponding author: Henu Kumar Verma, PhD, Research Scientist, Department of Immunopathology, Institute of lungs Biology and Disease, Comprehensive Pneumology Center, Helmholtz Zentrum, Ingolstädter Landstrasse 1, Neuherberg 85764, Munich, Germany. henu.verma@yahoo.com

Abstract

The role of alcoholic and other beverage consumption in the etiology of gastric cancer is unknown. Several studies have summarized and established a significant association between heavy alcohol consumption and gastric cancer risk, but evidence on alcohol-related cancer risk is conflicting.

Key Words: Alcohol; Gastric cancer; Meta-analysis; Gender and alcohol intake; Alcohol consumption

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Alcohol consumption among adult men and women is consistently linked to an increased risk of gastric cancer. Its role as a confounding factor in the gastric cancer burden is frequently overlooked. Although many cancers are genetically determined, most cancers are caused by interactions between the host and environmental/lifestyle factors.

Citation: Verma HK, Bhaskar L. Gender differences in the relationship between alcohol consumption and gastric cancer risk are uncertain and not well-delineated. World J Gastrointest Oncol 2021; 13(12): 2216-2218

URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2216.htm **DOI:** https://dx.doi.org/10.4251/wjgo.v13.i12.2216



on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: May 11, 2021 Peer-review started: May 11, 2021 First decision: June 16, 2021 Revised: June 17, 2021 Accepted: November 3, 2021 Article in press: November 3, 2021 Published online: December 15, 2021

P-Reviewer: Kang XJ, Tung TH S-Editor: Gao CC L-Editor: Kerr C P-Editor: Gao CC



TO THE EDITOR

Cancer is a complex disease that arises from interactions between genetic and environmental factors. There is overwhelming evidence that alcohol consumption affects cancer risk. According to the 4th edition of the European Code against Cancer, not drinking alcohol is better for cancer prevention^[1]. We read the recent publication on "Sex as an effect modifier in the association between alcohol intake and gastric cancer risk" by Bae[2]. I sincerely appreciate the author for providing relevant information about the relationship between alcohol consumption and the risk of gastric cancer in men and women. It has been of great interest to us. In this systematic review and meta-analysis, the authors included ten Asian, seven European and three American cohort studies comprising 27 cohorts, making the study more robust.

The meta-analysis showed that overall alcohol intake increased the risk of gastric cancer with a summary risk ratio of 1.13 [95% confidence interval (CI): 1.04-1.23]. However, subgroup analysis by gender demonstrated higher relative risk (RR) in male cohorts (RR = 1.18, 95%CI: 1.06-1.32, *I*² = 55.5%) than in female cohorts (RR = 1.07, 95% CI: 0.96-1.19, $I^2 = 0.0\%$). Several previous meta-analyses that have investigated the relationship between alcohol consumption and gastric cancer risk were inconclusive. Previous meta-analyses demonstrated no association between alcohol drinking and gastric cancer risk in overall and gender-stratified analyses[3]. In contrast to this, heavy alcohol consumption significantly increased the risk of gastric cancer in both men and women[4]. Subsequent meta-analyses indicated that alcohol consumption was associated with an increased risk of gastric cancer in men but not in women[5,6]. Recently, Kim *et al*[7] shown that high alcohol consumption of alcohol (≥ 20 g/d for women or \geq 40 g/d for men) significantly increased the risk of gastric cancer. Further, large-scale meta-analyses by Han et al[8] found a protective effect of alcohol consumption in Europe and a significant harmful impact in men in America. That depends on other confounding factors, including age, education level, smoking status, and body mass index.

Although the systematic review and meta-analyses have linked gastric cancer risk with alcohol consumption, gender differences in the relationship between alcohol consumption and gastric cancer risk are uncertain and not well-delineated. Indeed, hormonal, genetic and environmental differences influence how men and women consume alcohol[9]. While men are more likely to drink excessive amounts of alcohol, women are more likely to abstain for long periods. Bae[2] has not considered the frequency of heavy and light alcohol drinking habits in men and women, which is the main factor that determines modulating effect of sex on the relationship between alcohol consumption and gastric cancer risk[2]. More research with critical confounding factors such as drinking intensity is needed to provide a more precise relationship between alcohol consumption and the risk of gastric cancer in men and women.

REFERENCES

- 1 Scoccianti C, Cecchini M, Anderson AS, Berrino F, Boutron-Ruault MC, Espina C, Key TJ, Leitzmann M, Norat T, Powers H, Wiseman M, Romieu I. European Code against Cancer 4th Edition: Alcohol drinking and cancer. Cancer Epidemiol 2016; 45: 181-188 [PMID: 27816465 DOI: 10.1016/j.canep.2016.09.011]
- 2 Bae JM. Sex as an effect modifier in the association between alcohol intake and gastric cancer risk. World J Gastrointest Oncol 2021; 13: 453-461 [PMID: 34040705 DOI: 10.4251/wjgo.v13.i5.453]
- Tramacere I, Negri E, Pelucchi C, Bagnardi V, Rota M, Scotti L, Islami F, Corrao G, La Vecchia C, 3 Boffetta P. A meta-analysis on alcohol drinking and gastric cancer risk. Ann Oncol 2012; 23: 28-36 [PMID: 21536659 DOI: 10.1093/annonc/mdr135]
- 4 He Z, Zhao TT, Xu HM, Wang ZN, Xu YY, Song YX, Ni ZR, Xu H, Yin SC, Liu XY, Miao ZF. Association between alcohol consumption and the risk of gastric cancer: a meta-analysis of prospective cohort studies. Oncotarget 2017; 8: 84459-84472 [PMID: 29137439 DOI: 10.18632/oncotarget.20880]
- Wang PL, Xiao FT, Gong BC, Liu FN. Alcohol drinking and gastric cancer risk: a meta-analysis of observational studies. Oncotarget 2017; 8: 99013-99023 [PMID: 29228746 DOI: 10.18632/oncotarget.20918]
- 6 Li Y, Eshak ES, Shirai K, Liu K, Dong JY, Iso H, Tamakoshi A; JACC Study Group. Alcohol Consumption and Risk of Gastric Cancer: The Japan Collaborative Cohort Study. J Epidemiol 2021; 31: 30-36 [PMID: 31902851 DOI: 10.2188/jea.JE20190304]
- Kim MH, Kim SA, Park CH, Eun CS, Han DS, Kim YS, Song KS, Choi BY, Kim HJ. Alcohol consumption and gastric cancer risk in Korea: a case-control study. Nutr Res Pract 2019; 13: 425-433 [PMID: 31583062 DOI: 10.4162/nrp.2019.13.5.425]
- Han X, Xiao L, Yu Y, Chen Y, Shu HH. Alcohol consumption and gastric cancer risk: a meta-analysis



of prospective cohort studies. Oncotarget 2017; 8: 83237-83245 [PMID: 29137337 DOI: 10.18632/oncotarget.19177]

9 Rao VR, Bhaskar LV, Annapurna C, Reddy AG, Thangaraj K, Rao AP, Singh L. Single nucleotide polymorphisms in alcohol dehydrogenase genes among some Indian populations. Am J Hum Biol 2007; **19**: 338-344 [PMID: 17421009 DOI: 10.1002/ajhb.20589]



0 WŰ

World Journal of **Gastrointestinal** Oncology

Submit a Manuscript: https://www.f6publishing.com

World J Gastrointest Oncol 2021 December 15; 13(12): 2219-2222

DOI: 10.4251/wjgo.v13.i12.2219

ISSN 1948-5204 (online)

LETTER TO THE EDITOR

Critical biomarkers of hepatocellular carcinoma in body fluids and gut microbiota

Lekshmi R Nath, Maneesha Murali, Bhagyalakshmi Nair

ORCID number: Lekshmi R Nath 0000-0002-7726-7219; Maneesha Murali 0000-0003-2772-3930; Bhagyalakshmi Nair 0000-0002-0364-881X

Author contributions: Nath LR designed the draft; Murali M and Nair B performed a literature review; Nath LR and Nair B analyzed data; Nath LR and Murali M wrote the letter; Nath LR revised the paper.

Conflict-of-interest statement: The authors did not have any conflict of interest.

Country/Territory of origin: India

Specialty type: Oncology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): C Grade D (Fair): D, D Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative

Lekshmi R Nath, Department of Pharmacognosy, Amrita School of Pharmacy, Amrita Vishwa Vidyapeetham, AIMS Health Science Campus, Kochi 682041, Kerala, India

Maneesha Murali, Bhagyalakshmi Nair, Amrita School of Pharmacy, Amrita Vishwa Vidyapeetham, AIMS Health Science Campus, Kochi 682041, Kerala, India

Corresponding author: Lekshmi R Nath, PhD, Assistant Professor, Department of Pharmacognosy, Amrita School of Pharmacy, Amrita Vishwa Vidyapeetham, AIMS Health Science Campus, Ponekkara P.O., Kochi 682041, Kerala, India. lekshmirnath@aims.amrita.edu

Abstract

Hepatocellular carcinoma (HCC) is the most prevalent primary liver cancer and one of the major causes of cancer-related death. The development of specific noninvasive or diagnostic markers from blood, urine and feces may represent a valuable tool for detecting HCC at an early stage. Biomarkers are considered novel potential targets for therapeutic intervention. It helps in the prediction of prognosis or recurrence of HCC, and also assist in the selection of appropriate treatment modality. We summarize the most relevant existing data about various biomarkers that play a key role in the progression of HCC.

Key Words: Hepatocellular carcinoma; Biomarker; Body fluids; Blood; Gut microbiota

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Hepatocellular carcinoma (HCC) ranks fourth among the leading causes of cancer-related mortality. The development of specific noninvasive or diagnostic markers from blood, urine and feces may represent a valuable tool for detecting HCC at an early stage. Biomarkers help in the prediction of prognosis or recurrence, selection of appropriate treatment modality, and signify novel potential targets for therapeutic interventions. We summarize the most relevant existing data about various biomarkers involved in the progression of HCC.

Citation: Nath LR, Murali M, Nair B. Critical biomarkers of hepatocellular carcinoma in body fluids and gut microbiota. World J Gastrointest Oncol 2021; 13(12): 2219-2222 URL: https://www.wjgnet.com/1948-5204/full/v13/i12/2219.htm



Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: May 31, 2021 Peer-review started: May 31, 2021 First decision: June 26, 2021 Revised: July 15, 2021 Accepted: November 5, 2021 Article in press: November 5, 2021 Published online: December 15, 2021

P-Reviewer: Li X, Li Y, Sitkin S, Xu χ S-Editor: Gao CC L-Editor: Kerr C P-Editor: Gao CC



DOI: https://dx.doi.org/10.4251/wjgo.v13.i12.2219

TO THE EDITOR

We were interested to read the review reported by Guan et al[1] that clearly emphasized the substantial role of biomarkers from different body fluids such as blood, urine and feces for the early detection of primary and recurrent hepatocellular carcinoma (HCC). From the study reports, detection of biomarkers through screening of body fluids or feces is regarded as beneficial due to the quick and easy extraction procedures, stability, proper time management, cost-effectiveness and accessibility in comparison with conventional screening methods. The review highlights the clinical significance of several diagnostic biomarkers of HCC, including proteins, metabolites, circulating nucleic acids, circulating tumor cells (CTCs), extracellular vesicles (EVs), and gut microbiota from blood, urine and feces.

A large pool of evidence suggests the presence of elevated serum blood levels of bilirubin, albumin, α-fetoprotein (AFP), Lens culinaris agglutinin-reactive fraction of AFP (AFP-L3) and des-γ-carboxy prothrombin (DCP) at the time of diagnosis of HCC. These biomarkers exhibit a close relation with HCC staging and prognosis of overall survival and disease-free survival. Elevated levels of AFP in cases of liver injury above the reference range (400-500 ng/mL) can be considered crucial for the prognosis of HCC. AFP-L3 possesses better sensitivity but low specificity for the early detection of HCC. To its expression in small tumors (< 2 cm in diameter) of aggressive types, the prognosis of early-stage HCC is relevant if the AFP-L3 level is greater than 10% in comparison with AFP. DCP can be regarded as an excellent prognostic biomarker since it can differentiate nonmalignant cirrhosis and HCC with a specificity of 93% and sensitivity of 92% at a cut-off value of about 150 mAU/mL[2]. With disease progression, metabolic markers such as methionine, proline and ornithine increase, whereas the levels of pimelylcarnitine and octanoylcarnitine decrease^[3]. The applicability of phenylalanyl-tryptophan and glycocholate as a superior biomarker was demonstrated in a multicenter cohort study that indicated its diagnostic accuracy of 86.0%-92.5% in HCC[4].

The progression of HCC involves invasion, migration, proliferation and metastasis. Studies have shown that drug resistance is mainly mediated through the functional activation of miRNAs. Clinicians can predict the overall survival of patients based on the expression of miRNA. Single miRNAs like miR-130b, miR-150, miR-182, miR-215 and miR-96 are considered key candidates among all miRNAs but the use of multiple miRNAs as promising biomarkers for the prediction of early as well as recurring HCC is recent^[5]. CTCs play a significant role in the prediction of HCC recurrence, prognostic evaluation for surveillance, and promotion of suitable adjuvant therapy. CTCs are generally categorized as a small subpopulation of malignant cells secreted from primary malignant tissue and they are usually expressed at the aggressive malignancy stage; therefore, liquid biopsy of CTCs facilitates timely diagnosis of HCC [6]. Another important category of biomarker with a functional role in the prediction of HCC progression is EVs. Increased circulating levels of EVs have contributed to poor survival and disease-free survival in HCC patients. Despite their high capability of being absorbed into host cells, EVs are considered an efficient tool for targeted approaches. This is by the incorporation of therapeutic agents to improve therapeutic efficacy and reduce side effects. The incorporation of sodium/iodide symporter protein to EVs has been used as one of the systemic targeted approaches to cancer treatment with the promotion of cytotoxicity and radioiodine therapy[7].

Another potential category of biomarkers for HCC are urine-based. Among the biomarkers, higher levels of 8-oxodeoxyguanosine improve DNA repair mechanisms by overcoming oxidative DNA damage with a reduction in risk of developing HCC. Enhanced levels of 15-F2t-isoprostane are also correlated with the risks of HCC. Urinary proteins such as urinary DJ-1, chromatin assembly factor-1, heat shock protein 60 and orosomucoid, and metabolites such as ethanolamine, lactic acid, aconitinic acid, phenylalanine and ribose were found to be effective predictors for early HCC recurrence. Additionally, the overexpression of urinary trypsin inhibitor in HCC was revealed to be a risk factor for HCC recurrence^[1]. In a study reported by Hann *et al* [8], detection of urinary markers such as TP53m, mSGTP and mRASSF1A were potential tools for the early detection of HCC recurrence (Figure 1).

Inflammation significantly decreases the expression of beneficial microflora which, in turn, enhances the risk of liver malignancy by accumulating harmful compounds.





Figure 1 Pictorial representation of numerous biomarkers derived from different body fluids, namely, blood (serum), urine and feces. These biomarkers constitute a wide spectrum of proteins, nucleic acids and metabolites. Circulating tumor cells, miRNAs and gut microbiota which can be beneficial for the early detection, diagnosis and prognosis of hepatocellular carcinoma.

Translocated bacterial products such as lipopolysaccharides, peptidoglycans, muramyl-dipeptides and bacterial DNA from the infectious stage of the gut stimulate an inflammatory cascade by activation of signaling through Toll-like receptors (TLRs). Stimulation of interleukin-6, either directly or via the JAK/STAT3 pathway forces the gut microbiota to induce proliferation and progression of HCC. Gut microbiota can stimulate the generation of reactive free radical oxygen species indirectly via small molecular motifs derived from a pathogenic class of microbes by the activation of NADPH-oxidase (NOX1-NOX4). Microbial imbalance and enhancement of inflammation are directly correlated with fluctuating redox status. Modulation of farnesoid X receptor activation by gut microbiota enhances bile acid accumulation in the liver. This leads to damage of hepatocyte plasma membranes, resulting in activation of an inflammatory response and production of reactive oxygen species through stimulating the MAPK pathway. As a result, the secretion of inflammatory cytokines via the nuclear factor-B pathway is increased by induction of proliferation and immortalization of HCC cells directly or via the JAK/STAT3 pathway. Gut microbiota can exhaust the surveillance of the immune system within the tumor microenvironment of HCC through macrophage polarization via the activation of TLRs. This results in further diversification and progression of the tumor[9]. Additionally, several other biomarkers namely, glypican-3, Golgi protein complex-73, squamous cell carcinoma antigen and circulating tumor DNA are useful for early diagnosis of HCC, and might be clinically validated in the near future^[10].

The supporting evidence gives an insight into novel biomarkers for early prediction and prevention of HCC. HCC accounts for almost 90% of primary liver malignancies and has a poor prognosis due to rapid metastasis and multidrug resistance. Diagnosis of HCC at an early stage is important for overcoming the hurdles associated with the disease[11]. To conclude, it is important to identify and develop promising biomarkers for early diagnosis and prognosis as well as therapy of HCC.

ACKNOWLEDGEMENTS

We thank Dr. Abraham R, JHSPH, Baltimore for the proofreading and Ms. Diers AR, University of Florida, United States for the language editing.

MJGO | https://www.wjgnet.com

REFERENCES

- Guan MC, Ouyang W, Wang MD, Liang L, Li N, Fu TT, Shen F, Lau WY, Xu QR, Huang DS, Zhu 1 H, Yang T. Biomarkers for hepatocellular carcinoma based on body fluids and feces. World J Gastrointest Oncol 2021; 13: 351-365 [PMID: 34040698 DOI: 10.4251/wjgo.v13.i5.351]
- 2 Wang X, Zhang Y, Yang N, He H, Tao X, Kou C, Jiang J. Evaluation of the Combined Application of AFP, AFP-L3%, and DCP for Hepatocellular Carcinoma Diagnosis: A Meta-analysis. Biomed Res Int 2020; 2020: 5087643 [PMID: 33015170 DOI: 10.1155/2020/5087643]
- Kim DJ, Cho EJ, Yu KS, Jang IJ, Yoon JH, Park T, Cho JY. Comprehensive Metabolomic Search for 3 Biomarkers to Differentiate Early Stage Hepatocellular Carcinoma from Cirrhosis. Cancers (Basel) 2019; 11 [PMID: 31590436 DOI: 10.3390/cancers11101497]
- Luo P, Yin P, Hua R, Tan Y, Li Z, Qiu G, Yin Z, Xie X, Wang X, Chen W, Zhou L, Li Y, Chen H, 4 Gao L, Lu X, Wu T, Wang H, Niu J, Xu G. A Large-scale, multicenter serum metabolite biomarker identification study for the early detection of hepatocellular carcinoma. Hepatology 2018; 67: 662-675 [PMID: 28960374 DOI: 10.1002/hep.29561]
- Ning S, Liu H, Gao B, Wei W, Yang A, Li J, Zhang L. miR-155, miR-96 and miR-99a as potential 5 diagnostic and prognostic tools for the clinical management of hepatocellular carcinoma. Oncol Lett 2019; 18: 3381-3387 [PMID: 31452818 DOI: 10.3892/ol.2019.10606]
- Mann J, Reeves HL, Feldstein AE. Liquid biopsy for liver diseases. Gut 2018; 67: 2204-2212 6 [PMID: 30177542 DOI: 10.1136/gutjnl-2017-315846]
- Costanzi E, Simioni C, Varano G, Brenna C, Conti I, Neri LM. The Role of Extracellular Vesicles as 7 Shuttles of RNA and Their Clinical Significance as Biomarkers in Hepatocellular Carcinoma. Genes (Basel) 2021; 12 [PMID: 34207985 DOI: 10.3390/genes12060902]
- 8 Hann HW, Jain S, Park G, Steffen JD, Song W, Su YH. Detection of urine DNA markers for monitoring recurrent hepatocellular carcinoma. Hepatoma Res 2017; 3: 105-111 [PMID: 28795155 DOI: 10.20517/2394-5079.2017.15]
- Gupta H, Youn GS, Shin MJ, Suk KT. Role of Gut Microbiota in Hepatocarcinogenesis. Microorganisms 2019; 7 [PMID: 31060311 DOI: 10.3390/microorganisms7050121]
- 10 Pandyarajan V, Govalan R, Yang JD. Risk Factors and Biomarkers for Chronic Hepatitis B Associated Hepatocellular Carcinoma. Int J Mol Sci 2021; 22 [PMID: 33418899 DOI: 10.3390/ijms22020479]
- Tang W, Chen Z, Zhang W, Cheng Y, Zhang B, Wu F, Wang Q, Wang S, Rong D, Reiter FP, De 11 Toni EN, Wang X. The mechanisms of sorafenib resistance in hepatocellular carcinoma: theoretical basis and therapeutic aspects. Signal Transduct Target Ther 2020; 5: 87 [PMID: 32532960 DOI: 10.1038/s41392-020-0187-x]



WJGO | https://www.wjgnet.com



Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

