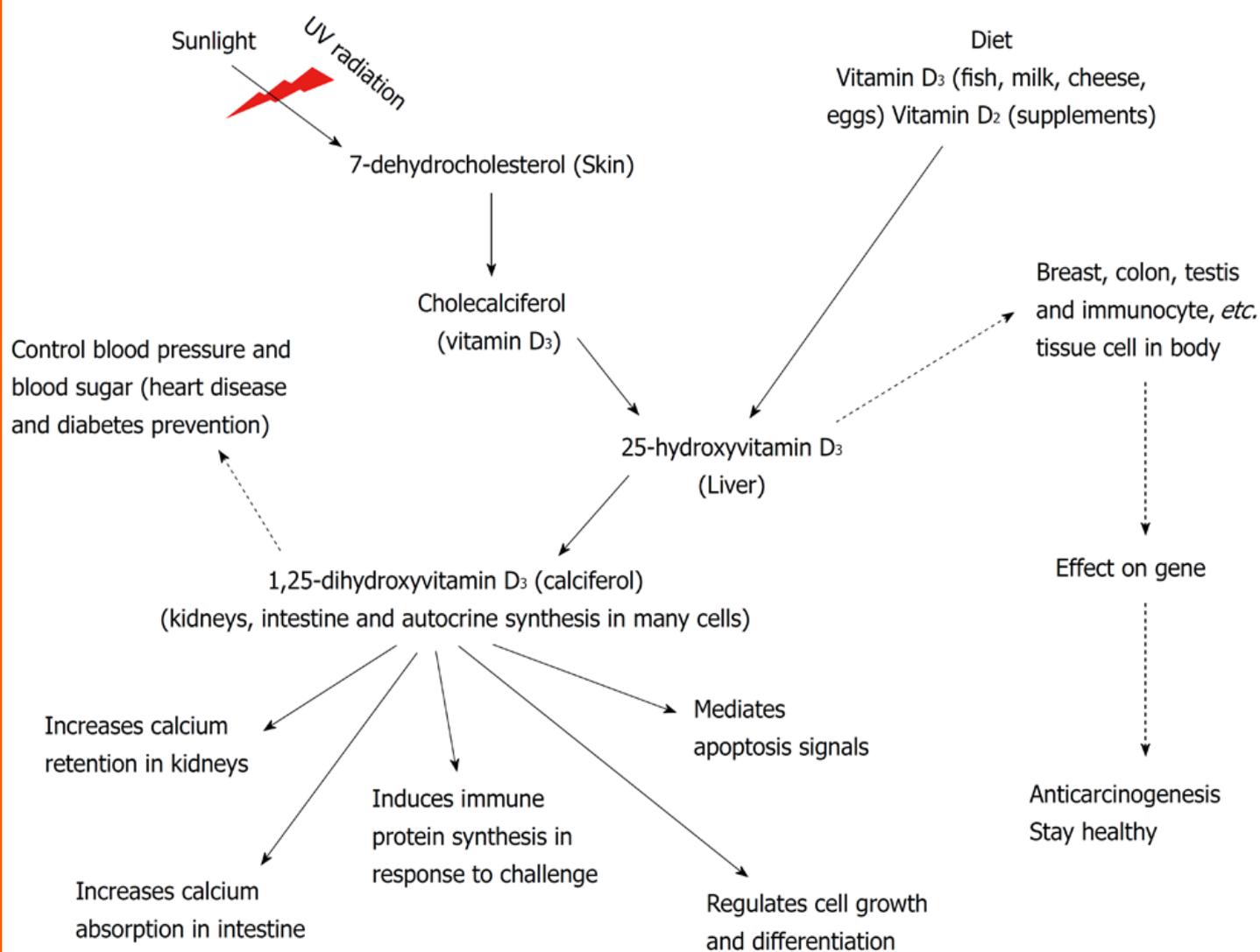


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Intracellular chloride regulates the G₁/S cell cycle progression in gastric cancer cells

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Abstract

Recent studies show that ion channels/transporters play important roles in fundamental cellular functions. Several reports indicating the important roles of Cl⁻ channels/transporters on cell proliferation suggest that the intracellular chloride concentration ([Cl⁻]_i) regulated by them would be one of critical messengers. We investigated whether the [Cl⁻]_i controls cell proliferation and cell cycle progression in human gastric cancer cells. Our studies indicated that furosemide, a blocker of Na⁺/K⁺/2Cl⁻ cotransporter (NKCC), diminished cell growth by delaying the G₁-S phase progression in gastric cancer cells with high expression and activity of NKCC. Furthermore, we found that the culture in the low Cl⁻ medium (replacement of Cl⁻ by NO₃⁻) decreased the [Cl⁻]_i and inhibited cell growth of gastric cancer cells and that this inhibition of cell growth was due to cell cycle arrest at the G₀/G₁ phase caused by diminution

of CDK2 and phosphorylated Rb. The culture of cells in the low Cl⁻ medium significantly increased expressions of p21 mRNA and protein. In addition, the low Cl⁻ medium induced phosphorylation of mitogen activated protein kinases (MAPKs). Treatment with an inhibitor of p38 or JNK significantly suppressed p21 upregulation caused by culture in a low Cl⁻ medium and rescued gastric cancer cells from the low Cl⁻-induced G₁ cell cycle arrest. These findings revealed that the [Cl⁻]_i affects the cell proliferation *via* activation of MAPKs through upregulation of p21 in gastric cancer cells. Our results suggest that the [Cl⁻]_i regulates important cellular functions in gastric cancer cells, leading to the development of novel therapeutic strategies.

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Key words: Intracellular chloride; Cell proliferation; Cell cycle; Gastric cancer; Cl⁻ channel; Cl⁻ transporter

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INTRODUCTION

Over the past two decades, many researchers have reported that ion channels and transporters play important roles in fundamental cellular functions. Particularly, their physiological roles in cell proliferation have been considered since cell volume changes, which require the participation of ion movement across the cell membrane,

are indispensable in cell cycle progression. Recently, the roles of ion transporters have been studied in cancer cells^[1-3] and various types of ion transporters have been found in cancers of digestive organs. Some types of K^+ channels, such as voltage-gated Kv1.3 channel, voltage-gated HERG channel and KCNK9 channel, have been reported to be expressed at extremely higher levels in colonic carcinoma specimens than in normal colon tissues^[4-6]. In gastric cancer, the voltage-gated HERG channel has revealed cancer-limited expression and its blocker diminishes the G_1 to S phase transition^[7]. Furthermore, increased mRNA levels of voltage-gated Cav1.2 Ca^{2+} channels and Ca^{2+} -conducting channels (TRPM8) have been reported in colorectal adenocarcinoma^[8,9].

Several reports indicate that Cl^- channels/transporters play an important role in gastrointestinal cancer cells. For instance, transcriptional downregulation of Ca^{2+} -activated $2Cl^-$ channels (CLCA1 and CLCA2) genes is detected in colorectal tumor samples^[10]. Sarosi *et al.*^[11] have reported that the Cl^-/HCO_3^- exchanger influences the proliferation of Barrett's esophageal adenocarcinoma cells through changes of intracellular pH. In addition, we previously found that mRNA and the functional expression levels of $Na^+/K^+/2Cl^-$ cotransporter (NKCC) were higher in poorly-differentiated type gastric adenocarcinoma cells than in differentiated cells^[12]. These reports indicate that the transepithelial Cl^- transport plays an important role in cell proliferation of gastrointestinal cancer cells. From this viewpoint, we investigated whether the intracellular chloride concentration ($[Cl^-]_i$) regulates cell proliferation and cell cycle progression in human gastric cancer cells.

INTRACELLULAR CHLORIDE AND SIGNAL TRANSDUCTION

Our study^[13] indicates that the intracellular chloride could act as a signal to regulate mRNA expression of ion channel in renal epithelial A6 cells, suggesting that many physiological functions are associated with the change of the $[Cl^-]_i$. Similarly, Heimlich *et al.*^[14] have shown that an alteration of the $[Cl^-]_i$ plays an important role in the activation of signaling molecules upstream of the mitochondria, specifically impairing the intrinsic apoptotic pathway. In a mouse macula densa cell line, low chloride stimulates prostaglandin E2 release and COX-2 expression through activation of mitogen activated protein kinases (MAPKs)^[15]. These reports suggest that the changes in the $[Cl^-]_i$ might act as a regulator of various types of intracellular enzymes. We previously showed that furosemide, a blocker of NKCC, diminished cell growth by delaying the G_1 -S phase progression in gastric cancer cells with high expression and activity of NKCC^[12] (Figure 1). NKCC is one of the important transporters controlling the $[Cl^-]_i$ via uptake of Cl^- into the intracellular space and, therefore, furosemide decreases the $[Cl^-]_i$ ^[16] (Figure 1). Based on these findings, we hypothesized that the $[Cl^-]_i$ would be one of critical messengers regulating cell proliferation and

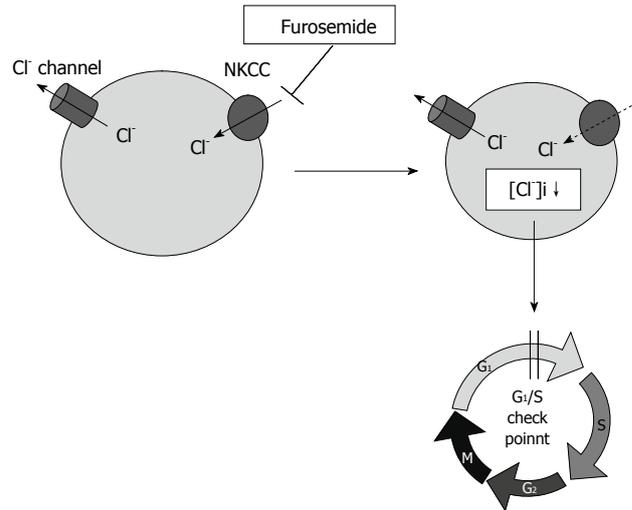


Figure 1 $Na^+/K^+/2Cl^-$ cotransporter controls the intracellular chloride concentration via uptake of Cl^- into the intracellular space. Furosemide, a blocker of $Na^+/K^+/2Cl^-$ cotransporter, delays the G_1 -S phase progression by decreasing the intracellular chloride concentration ($[Cl^-]_i$) in gastric cancer cells.

investigated whether the $[Cl^-]_i$ regulates cell cycle progression in human gastric cancer cells.

CELL CYCLE PROGRESSION AND $[Cl^-]_i$ IN GASTRIC CANCER CELLS

We directed our interest to the roles of the $[Cl^-]_i$ in cell proliferation and cell cycle progression of gastric cancer cells. We applied media containing various chloride concentrations to human gastric cancer MKN28 cells and measured the $[Cl^-]_i$ at 48 h after the application. The $[Cl^-]_i$ of gastric cancer cells incubated in the normal medium was around 30 mmol/L. When cells were incubated in the low Cl^- medium (replacement of Cl^- by NO_3^-) for 48 h, the $[Cl^-]_i$ decreased to around 0 mmol/L. Furthermore, the $[Cl^-]_i$ of cells cultured in the media containing various chloride concentrations was proportionally dependent on the chloride concentration of the cultured medium^[17,18]. These findings indicated that our experimental system using the low Cl^- medium can be used as a model of the $[Cl^-]_i$ regulation (Figure 2). The proliferation rate in MKN28 cells was significantly diminished by the culture in the low Cl^- medium compared with that in a normal one. In addition, analysis of cell proliferation of MKN28 cells cultured in the media containing various chloride concentrations indicated that the rate of cell proliferation depends on the extracellular chloride concentration^[18]. These results revealed that the $[Cl^-]_i$ plays a key role in proliferation of gastric cancer cells. Cell cycle analysis revealed that the population of MKN28 cells staying in the G_0/G_1 phase was significantly increased and that cells staying in the S or G_2/M phase were reduced by the culture in the low Cl^- medium, suggesting that the decrease of the $[Cl^-]_i$ shows an inhibitory effect on the proliferation of gastric cancer cells by mainly diminishing the transition from the G_1 phase to the S phase^[18] (Figure 3).

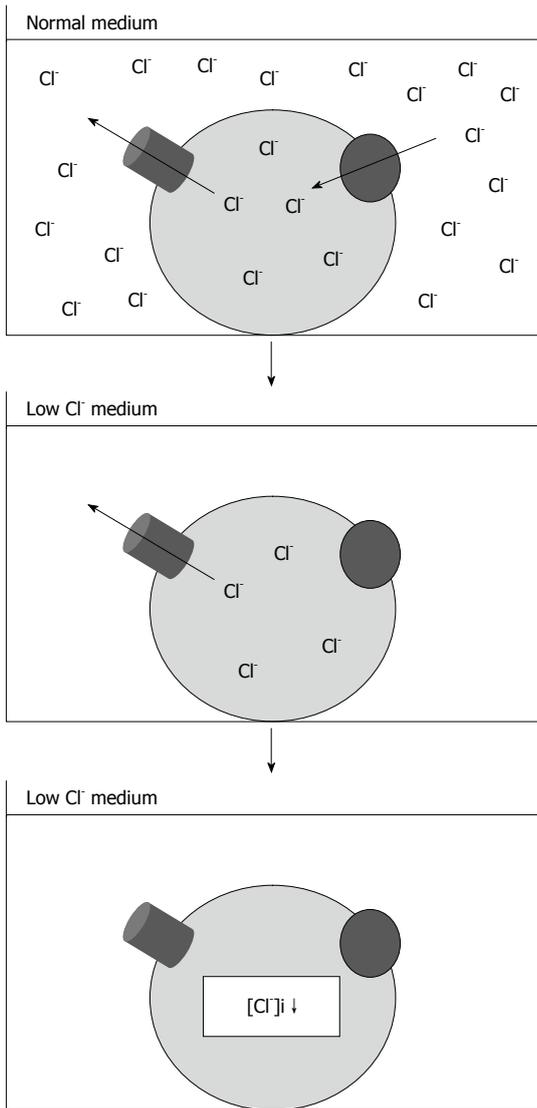


Figure 2 Experimental method for regulation of the intracellular chloride concentration of cultured cells. The intracellular chloride concentration ($[Cl^-]_i$) of gastric cancer cells is decreased by the culture in the low Cl^- medium, which were prepared by substituting, respectively, NaCl and KCl with $NaNO_3$ and KNO_3 .

$[Cl^-]_i$ CONTROLS THE G₁/S CELL CYCLE CHECK POINT BY REGULATING THE EXPRESSION OF p21 IN GASTRIC CANCER CELLS

We analyzed the expression of cell cycle-associated proteins involved in G₁-S phase transition to determine the mechanisms by which the decrease of the $[Cl^-]_i$ inhibited the proliferation of MKN28 cells. The culture in the low Cl^- medium significantly decreased phosphorylation of Rb. The expression of CDK2 protein, which is located at upstream of Rb in a signal pathway, was significantly downregulated by the culture in the low Cl^- medium. Furthermore, the low Cl^- medium elevated expression of p21^[18]. These results suggest that the $[Cl^-]_i$ of gastric cancer cells acts on the transition from the G₁ phase to

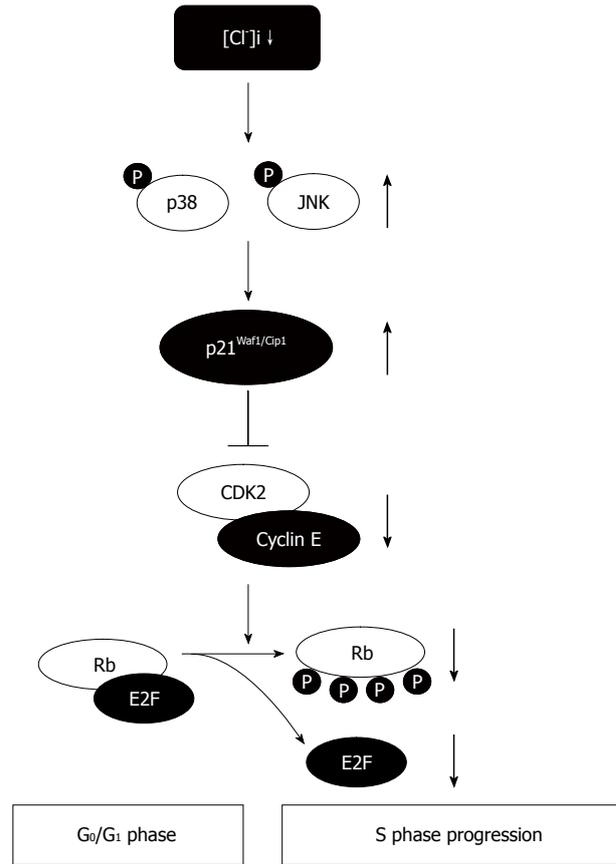


Figure 3 Roles of the intracellular chloride concentration in cell cycle progression of gastric cancer cells. The intracellular chloride concentration ($[Cl^-]_i$) affects the cell proliferation via activation of p38 and/or JNK cascades through upregulation of the p21. The decrease of the $[Cl^-]_i$ causes the G₀/G₁ phase arrest by diminishing expression of CDK2 and phosphorylated Rb due to an increase in expression of p21.

the S phase by regulating the expression of p21 and its downstream proteins in signal pathways (Figure 3). We found similar effects of the $[Cl^-]_i$ on cell cycle associated proteins in human prostatic cancer cells and mouse osteoblast cells^[16,19].

$[Cl^-]_i$ REGULATES p21 THROUGH THE ACTIVATION OF MAPKS IN GASTRIC CANCER CELLS

Generally, the induction of p21 is known to be dependent on tumor suppressor protein, p53, leading us to an idea that the upregulation of p21 induced by the decrease of the $[Cl^-]_i$ is due to activation of p53. However, the total expression and phosphorylation of p53 protein were not affected by application of the low Cl^- medium^[18], indicating that the upregulation of p21 induced by the decrease of the $[Cl^-]_i$ was not dependent on activation of p53.

Therefore, we determined that MAPKs are involved in the p21 upregulation and cell cycle arrest induced by reduction of the $[Cl^-]_i$. Culture of MKN28 cells in the low Cl^- medium significantly induced phosphorylation

of MAPKs (ERK, p38, and JNK)^[20]. Treatment with an inhibitor of p38 or JNK significantly suppressed p21 upregulation caused by culture in the low Cl⁻ medium and rescued MKN28 cells from the low Cl⁻-induced G₁ cell cycle arrest, whereas treatment with an ERK inhibitor had no significant effect on p21 expression or the growth of MKN28 cells in the low Cl⁻ medium^[20]. These results suggest that the [Cl⁻]_i affects the cell proliferation *via* activation of p38 and/or JNK cascades through upregulation of the p21 in gastric cancer cells (Figure 3).

CONCLUSION

We showed that: (1) chloride has significant effects on cell cycle progress in human gastric cancer cells; (2) the decrease of the [Cl⁻]_i causes the G₀/G₁ phase arrest by diminishing expression of CDK2 and phosphorylated Rb due to an increase in expression of p21; (3) the [Cl⁻]_i has significant effects on the activity of MAPKs cascades; and (4) the activation of p38 and JNK by a low Cl⁻ condition leads to growth inhibition *via* an increase of p21 expression. Our results suggest that the [Cl⁻]_i regulates intracellular signaling cascades participating in the control of proliferation in gastric cancer cells. A deeper understanding of these mechanisms may lead to the discovery of the [Cl⁻]_i as an important mediator in tumor development and as a novel therapeutic target for gastric cancer.

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Emerging role of vitamin D in colorectal cancer

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Abstract

Colorectal cancer is a common cancer and the fourth leading cause of death in Korea. The incidence and mortality of colorectal cancer varies according to risk factors, such as age, family history, genetic history, food habits, and physical activities. Some studies have focused on the association between vitamin D and colorectal cancer. Today, there is growing evidence that high vitamin D intake and a plasma level of 25(OH)D₃ reduce the incidence of colorectal cancer by modifying cancer angiogenesis, cell apoptosis, differentiation, and proliferation. Taken together, these results suggest that vitamin D supplementation alone, or in combination with anti-cancer agents, might reduce the incidence of colorectal cancer. In this review, we discuss the function and mechanism of vitamin D including the effect of vitamin D on colorectal cancer.

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Key words: Colorectal cancer; Vitamin D; Vitamin D receptor; 25-hydroxyvitamin D; 1,25(OH)₂D₃

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INTRODUCTION

Cancer of the colon or rectum is called colorectal cancer. It is a common cancer and the fourth leading cause of cancer-related death in the world. Colorectal cancer is also the third most common cancer in men and the second in women worldwide. It counted for an estimated, 1.2 million new cases and 0.6 million deaths in 2008^[1]. In addition, colorectal cancer is the second leading cause of cancer death in the United States. Colorectal cancer incidence rates are rapidly increasing in several historically low risk areas including Spain, and a number of countries within Eastern Asia and Eastern Europe^[2]. The risk factors for colorectal cancer are age, family history, and life-style, such as food habits and physical activities.

Vitamin D is associated with bone growth. Vitamin D insufficiency causes abnormal bone growth, while sufficiency prevents rickets and osteomalacia, as well as osteoporosis. Vitamin D enhances the immune system^[3]. In addition, vitamin D is known to prevent cancer^[4] and cardiovascular diseases^[5]. Vitamin D and its analogues have the ability to prevent cancer *in vitro* and in animal models. However, the anti-cancer effect of dietary vitamin D remains controversial. Recently, a study reported that vitamin D is not effective in preventing cancer and cardiovascular diseases^[6]. Of vitamin D forms, 1,25(OH)₂D₃ (calciferol) induces biological effects by binding to the vitamin D receptor (VDR). The activation of VDR leads

to the maintenance of calcium and phosphorus levels in the blood as well as of bone content. VDR is also involved in cell proliferation and differentiation.

For more than 20 years, epidemiological, experimental and clinical studies have shown that vitamin D has significant protective effect against the development of cancer^[4]. The mechanism of vitamin D works through several molecular pathways, such as growth-factor signaling, and transforming growth factor- β -SMAD signaling. The anti-cancer activities of vitamin D exerted by 1,25(OH)₂D₃ are produced by regulating the cell cycle, apoptosis, and adhesion, as well as by cellular differentiation and proliferation. Interestingly, vitamin D reduces the incidence of colorectal cancer, when vitamin D intake, the plasma level of 25(OH)D₃ and UV exposure is particularly high. In addition, the combination of vitamin D with other anti-cancer agents efficiently controls the development of colorectal cancer growth^[7].

Here, the function and mechanism of vitamin D is briefly introduced, and the beneficial effect of vitamin D on colorectal cancer is discussed.

CLASSIFICATION AND MOLECULAR GENETICS OF COLORECTAL CANCER

The molecular pathogenesis of colorectal cancer has been one of the most prominent study areas in recent years. Colorectal cancer exhibits two major forms: sporadic colorectal cancer and inherited colorectal cancer.

First, sporadic cancer occurs in people who have no family history or very little of the disease. Although cancer sometimes has a hereditary or familial component, it is not common. Approximately 70%-75% of colorectal cancer is sporadic cancer. Second, inherited colorectal cancer comprises familial and hereditary cancer. Familial and hereditary cancer occurs in families who have a faulty gene inherited from the father or mother. Generally, 5% of colorectal cancer is familial cancer. Familial adenomatous polyposis and hereditary nonpolyposis colorectal cancer are the two forms of inherited colorectal cancer^[8].

Ten to thirty percent of cases are attributed to familial risk and the rest to sporadic cancer. The majority of cancers are considered to be sporadic cancer. As stated above, most colorectal cancers are sporadic cancer, and only 5%-10% are inherited cancers.

FUNCTIONS OF VITAMIN D

Vitamin D exerts its various functions through molecular pathways. Vitamin D pathways are highly complex. The factors, affecting the vitamin D pathway, are P21, P27, CDKs, P53, BRCA-1.-2, β -catenin and c-myc. Depending on which factor, vitamin D is involved in cell adhesion, apoptosis, differentiation and division^[9]. Primarily, vitamin D plays an important role in muscle and bone health. Vitamin D deficiency results in impaired bone mineralization and leads to bone softening diseases in-

cluding rickets and osteomalacia^[10]. Further, vitamin D deficiency is involved in high bone turnover^[11]. Vitamin D deficiency can also play a role in the pathogenesis of auto-immune diseases such as multiple sclerosis, diabetes type 1, cancer^[12] and cardiovascular disease^[13]. Conversely, vitamin D deficiency increases parathyroid hormone levels leading to mobilization of calcium from bone, thereby compromising bone development in the adolescent^[14]. In contrast, vitamin D supplementation enhances bone density^[15]. Next, vitamin D exerts an anti-cancer activity^[16]. These activities of vitamin D functions are regulated by circulating vitamin D forms, the increasing concentration of 25(OH)D₃ and increasing activity of 1,25(OH)₂D₃. Vitamin D induces cellular proliferation, differentiation, and apoptosis of cancer and normal cells through the regulatory mechanism^[17-19]. These studies show that low intake levels of vitamin D increase the risk of colorectal cancer. Some studies show vitamin D exerts growth-restraining, anti-carcinogenic effects on colorectal cancer^[20,21]. In addition, vitamin D affects growth factors, regulation of cell division, cytokine synthesis, signaling, cell cycle control, and apoptosis pathway^[5,22]. In a study *in vitro*, a similar result was reported. When LOVO cells were treated for 8 d with various concentrations of 1,25(OH)₂D₃, cell proliferation was inhibited significantly^[23].

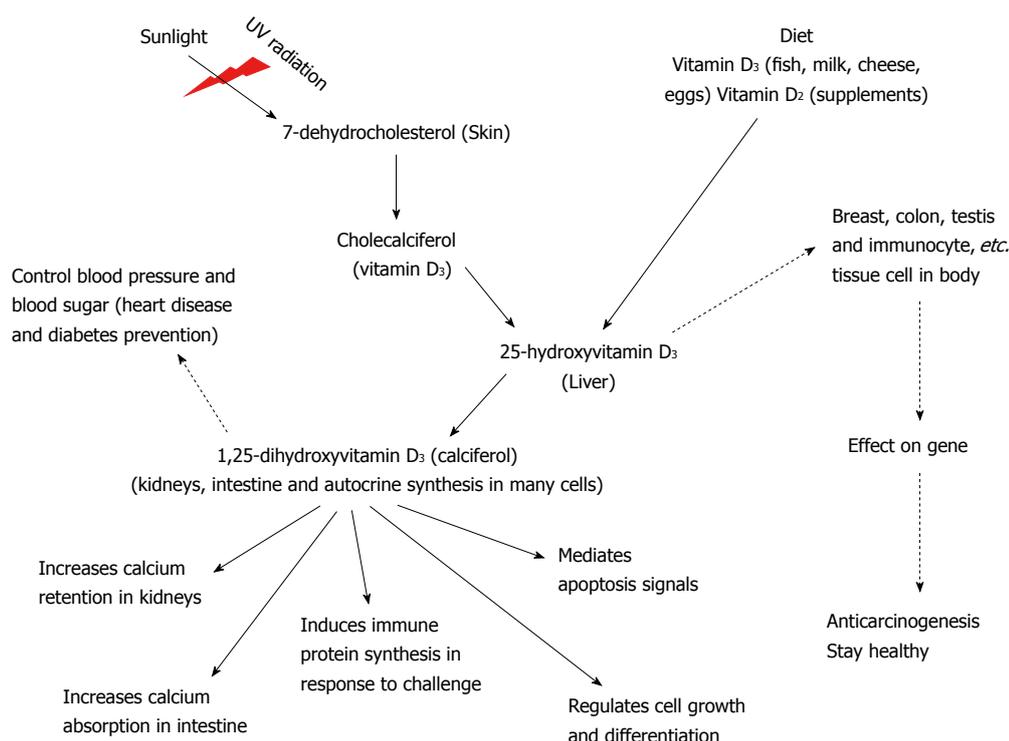
Table 1 summarizes the anti-cancer effects of vitamin D *in vivo* in mice and rats.

MECHANISM OF VITAMIN D IN COLORECTAL CANCER

Vitamin D and its metabolites reduce the incidence of various cancers by inhibiting cancer angiogenesis, stimulating normal cells^[33-37] and also by promoting the inhibition of proliferation. Vitamin D metabolites also help to maintain a standard calcium gradient in the various colonic epithelial cells. High levels of blood serum 25(OH)D₃ are associated with a noticeable decrease in proliferation of non-cancerous cells^[38,39]. The anti-proliferative effect of vitamin D is attained by inducing G1 cell-cycle arrest, which is probably mediated by up-regulation of cell cycle inhibitors. Vitamin D modulates the activation of these cell cycle related genes by various mechanisms. Vitamin D also exerts anti-carcinogenic effects by interfering with the synthesis of growth factors and cytokines and by modulating their signaling pathways. In addition to the growth inhibitory effects, vitamin D induces the differentiation of colon cancer cells. The 1,25(OH)₂D₃ and its analogs exert anti-carcinogenic activities in human colon cancer cells by inhibition of proliferation and induction of differentiation and apoptosis^[22]. The 1,25(OH)₂D₃ significantly increases the expression and activity of alkaline phosphatase, a marker of colonic differentiation. VDR activation by 1,25(OH)₂D₃ produces changes in stick junction integrity, increases differentiation and reduces oncogenic cell signaling. Induction of these genes affects cell oncogenesis, and tissue development. Thus, treatment with 1,25(OH)₂D₃ suppresses oncogenic genes in

Table 1 Anti-cancer effects of vitamin D in various rodent models

Treatment	Inducer	Species	Results	Ref.
Supplement		VDR knockout mouse	Inhibition of hyperproliferation and adenoma formation	[24]
Supplement		Apc1638 mice	Inhibition of carcinoma incidence	[25]
Deficiency		Balb/C mice	Enhancement of cancer cell growth	[26]
Supplement		Balb/C mice	Inhibition of tumor growth	[27]
Supplement		C57BL/6J mice	Inhibition of hyperproliferation	[28]
			Inhibition of tumor incidence	
Deficiency	DMH	SD rat	No effect	[29]
Deficiency		SD rat	Enhancement of carcinogenesis	[30]
Supplement	DMH	Fisher344 rat	No effect	[31]
Supplement	DMH	Fisher344 rat	Inhibition of tumor incidence	[32]

**Figure 1** Mechanisms of vitamin D in various tissues.

colon cancer cells. Finally, VDR genotypes are associated with anti-cancer activity in colorectal cancer. There are several VDR genotypes. For example, the most important VDR genotype is Bsm I, which has 3 variants: BB, Bb, and bb in America. The bb genotype is associated with lower concentrations of circulating 1,25(OH)₂D₃, leading to an increased incidence of colorectal cancer^[40,41]. Taken together, these observations demonstrate that vitamin D exerts anti-cancer activity in colon cancer.

Figure 1 describes the mechanisms of vitamin D in various tissues. In the figure, the dotted arrow shows a newly discovered function of vitamin D.

SUMMARY AND CONCLUSIONS

Previous research has shown the efficacy of taking vitamin D for reducing cancer risk^[42]. There is strong evidence that vitamin D can change and inhibit the devel-

opment of colon cancers^[22]. These protective effects are likely due to the regulatory effects of 1,25-dihydroxyvitamin D₃ (calciferol) on cellular mechanisms involved in cancer development, including apoptosis, cell adhesion, cell cycle control, regulation of cellular differentiation and proliferation. A clinical study group will set up guidelines for vitamin D intake and develop models to define levels of serum 25(OH)D₃ that prevent the growth of cancer. Elevation of vitamin D levels may protect against diverse cancers. Many studies show that vitamin D assists in prevention and therapy of cancer^[9]. The new guidelines will lead to more effective physical condition policies, resulting in substantially fewer cases of cancer of the colon in the future^[5].

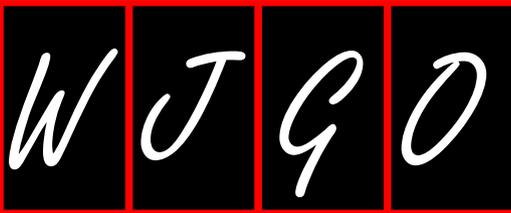
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Events Calendar 2011

January 20-22, 2011
 Gastrointestinal Cancers Symposium
 2011, San Francisco, CA,
 United States

January 27-28, 2011
 Falk Workshop, Liver and
 Immunology, Medical University,
 Regensburg, Germany

February 17-20, 2011
 APASL 2011-The 21st Conference
 of the Asian Pacific Association for
 the Study of the Liver, Bangkok,
 Thailand

February 21-21, 2011
 International Conference on
 Modern Cancer Management-Joint
 Symposium, Abuja, Nigeria,

February 26-March 1, 2011
 Canadian Digestive Diseases Week,
 Westin Bayshore, Vancouver, British
 Columbia, Canada

March 11-12, 2011
 First Integrative Care for the Future:
 The future of cancer care, Arnhem,
 The Netherlands
<http://www.integrativecareffuture.org/>

March 14-17, 2011
 British Society of Gastroenterology
 Annual Meeting 2011, Birmingham,
 England, United Kingdom

March 24-25, 2011
 Advanced Cancer Course
 "International Clinical Trials

Workshop", Punta del Este,
 Uruguay

April 6-7, 2011
 IBS-A Global Perspective,
 Milwaukee, WI, United States

April 6-8, 2011
 Third Latin American Symposium
 on Gastrointestinal Oncology-
 Chilean Foundation for Oncology
 Development Joint Symposium,
 Vina Del Mar, Chile

April 15-16, 2011
 Falk Symposium 177, Endoscopy
 Live Berlin 2011 Intestinal Disease
 Meeting, Maritim Hotel Berlin,
 Stauffenbergstr. 26, 10785 Berlin,
 Germany

April 20-23, 2011
 9th International Gastric Cancer
 Congress, COEX, World Trade
 Center, Samseong-dong, Gangnam-
 gu, Seoul 135-731, South Korea

May 8-12, 2011
 ESTRO International Oncology
 Forum, London, United Kingdom

May 19-22, 2011
 1st World Congress on Controversies
 in the Management of Viral Hepatitis
 (C-Hep), Palau de Congressos de
 Catalunya, Barcelona, Spain

May 25-27, 2011
 9th CIMT Annual Meeting,
 Targeting Cancer, Road-Maps for
 Success, Mainz, Germany

May 25-28, 2011

4th Congress of the Gastroenterology
 Association of Bosnia and
 Herzegovina with international
 participation, Sarajevo, Bosnia and
 Herzegovina

June 3-7, 2011
 2011 ASCO Annual Meeting,
 Chicago, IL, United States

June 18-24, 2011
 13th Joint ECCO-AACR-EORTC-
 ESMO Workshop on "Methods in
 Clinical Cancer Research", Flims,
 Switzerland

June 22-25, 2011
 ESMO 13th World Congress on
 Gastrointestinal Cancer, Barcelona,
 Spain

July 9-10, 2011
 Best of ASCO China, Hengzhou,
 China

July 21-23, 2011
 ASCO-JSMO Joint Symposium,
 Yokohama, Japan

August 25-28, 2011
 VII Peruvian Congress SPOM:
 Toward personalized Oncology-
 Endorsement, Lima, Peru

September 2-3, 2011
 Falk Symposium 178, Diverticular
 Disease, A Fresh Approach to a
 Neglected Disease, Martinstr. 29-37,
 50667 Cologne, Germany

September 10-14, 2011
 ICE 2011-International Congress of
 Endoscopy, Los Angeles Convention
 Center, 1201 South Figueroa Street,

Los Angeles, CA, United States

September 15-17, 2011
 2011 Gastrointestinal Oncology
 Conference, Sheraton Crystal City,
 Arlington, VA, United States

September 30-October 1, 2011
 Falk Symposium 179, Revisiting
 IBD Management: Dogmas to be
 Challenged, Place Rogier 3, 1210
 Brussels, Belgium, Germany

October 6-7, 2011
 IV InterAmerican Oncology
 Conference: Current Status and
 Future of Anti-Cancer Targeted
 Therapies, Buenos Aires, Argentina

October 14-15, 2011
 New Trends in the Medical
 Treatment of Solid Malignancy-
 Romanian Society for Medical
 Oncology Joint Symposium,
 Bucharest, Romania

October 27-29, 2011
 EORTC-NCI-ASCO Annual Meeting
 on Molecular Markers in Cancer,
 Brussels, Belgium

November 11-12, 2011
 Falk Symposium 180, IBD 2011:
 Progress and Future for Lifelong
 Management, 1-12-33 Akasaka,
 Minato-ku, Tokyo 107-0052, Japan

November 30-December 3, 2011
 8th International Cancer Conference
 "Entering the 21st Century for
 Cancer Control in Africa"-African
 Organization for Research and
 Training in Cancer Joint Symposium,
 Cairo, Egypt

GENERAL INFORMATION

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The columns in the issues of *WJGO* will include: (1) Editorial: To introduce and comment on major advances and developments in the field; (2) Frontier: To review representative achievements, comment on the state of current research, and propose directions for future research; (3) Topic Highlight: This column consists of three formats, including (A) 10 invited review articles on a hot topic, (B) a commentary on common issues of this hot topic, and (C) a commentary on the 10 individual articles; (4) Observation: To update the development of old and new questions, highlight unsolved problems, and provide strategies on how to solve the questions; (5) Guidelines for Basic Research: To provide guidelines for basic research; (6) Guidelines for Clinical Practice: To provide guidelines for clinical diagnosis and treatment; (7) Review: To review systemically progress and unresolved problems in the field, comment on the state of current research, and make suggestions for future work; (8) Original Articles: To report innovative and original findings in gastrointestinal oncology; (9) Brief Articles: To briefly report the novel and innovative findings in cardiology; (10) Case Report: To report a rare or typical case; (11) Letters to the Editor: To discuss and make reply to the contributions published in *WJGO*, or to introduce and comment on a controversial issue of general interest; (12) Book Reviews: To introduce and comment on quality monographs of gastrointestinal oncology; and (13) Guidelines: To introduce consensus and guidelines reached by international and national academic authorities worldwide on the research in gastrointestinal oncology.

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Please list 5-10 key words, selected mainly from *Index Medicus*, which reflect the content of the study.

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- 1 **Jung EM**, Clevert DA, Schreyer AG, Schmitt S, Rennert J, Kubale R, Feuerbach S, Jung F. Evaluation of quantitative contrast harmonic imaging to assess malignancy of liver tumors: A prospective controlled two-center study. *World J Gastroenterol* 2007; **13**: 6356-6364 [PMID: 18081224 DOI: 10.3748/wjg.13.6356]

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- 3 **Tian D**, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. *Proc Natl Acad Sci USA* 2006; In press

Organization as author

- 4 **Diabetes Prevention Program Research Group**. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; **40**: 679-686 [PMID: 12411462 PMCID:2516377 DOI:10.1161/01.HYP.0000035706.28494.09]

Both personal authors and an organization as author

- 5 **Vallancien G**, Emberton M, Harving N, van Moorselaar RJ; Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; **169**: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

- 6 21st century heart solution may have a sting in the tail. *BMJ* 2002; **325**: 184 [PMID: 12142303 DOI:10.1136/bmj.325.7357.184]

Volume with supplement

- 7 **Geraud G**, Spierings EL, Keywood C. Tolerability and safety of frovatriptan with short- and long-term use for treatment of migraine and in comparison with sumatriptan. *Headache* 2002; **42** Suppl 2: S93-99 [PMID: 12028325 DOI:10.1046/j.1526-4610.42.s2.7.x]

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- 8 **Banit DM**, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (**401**): 230-238 [PMID: 12151900 DOI:10.1097/00003086-200208000-00026]

No volume or issue

- 9 Outreach: Bringing HIV-positive individuals into care. *HRS A Careaction* 2002; 1-6 [PMID: 12154804]

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- 10 **Sherlock S**, Dooley J. Diseases of the liver and biliary system. 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296

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- 11 **Lam SK**. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450

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- 12 **Breedlove GK**, Schorfheide AM. Adolescent pregnancy. 2nd ed. Wiczorek RR, editor. White Plains (NY): March of Dimes Education Services, 2001: 20-34

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- 13 **Harnden P**, Joffe JK, Jones WG, editors. Germ cell tumours V. Proceedings of the 5th Germ cell tumours Conference; 2001 Sep 13-15; Leeds, UK. New York: Springer, 2002: 30-56

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- 14 **Christensen S**, Oppacher F. An analysis of Koza's computational effort statistic for genetic programming. In: Foster JA, Lutton E, Miller J, Ryan C, Tettamanzi AG, editors. Genetic programming. EuroGP 2002: Proceedings of the 5th European Conference on Genetic Programming; 2002 Apr 3-5; Kinsdale, Ireland. Berlin: Springer, 2002: 182-191

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- 15 Morse SS. Factors in the emergence of infectious diseases. *Emerg Infect Dis* serial online, 1995-01-03, cited 1996-06-05; 1(1): 24 screens. Available from: URL: <http://www.cdc.gov/ncidod/eid/index.htm>

Patent (list all authors)

- 16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1

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Write as mean \pm SD or mean \pm SE.

Statistical expression

Express *t* test as *t* (in italics), *F* test as *F* (in italics), chi square test as χ^2 (in Greek), related coefficient as *r* (in italics), degree of freedom as ν (in Greek), sample number as *n* (in italics), and probability as *P* (in italics).

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