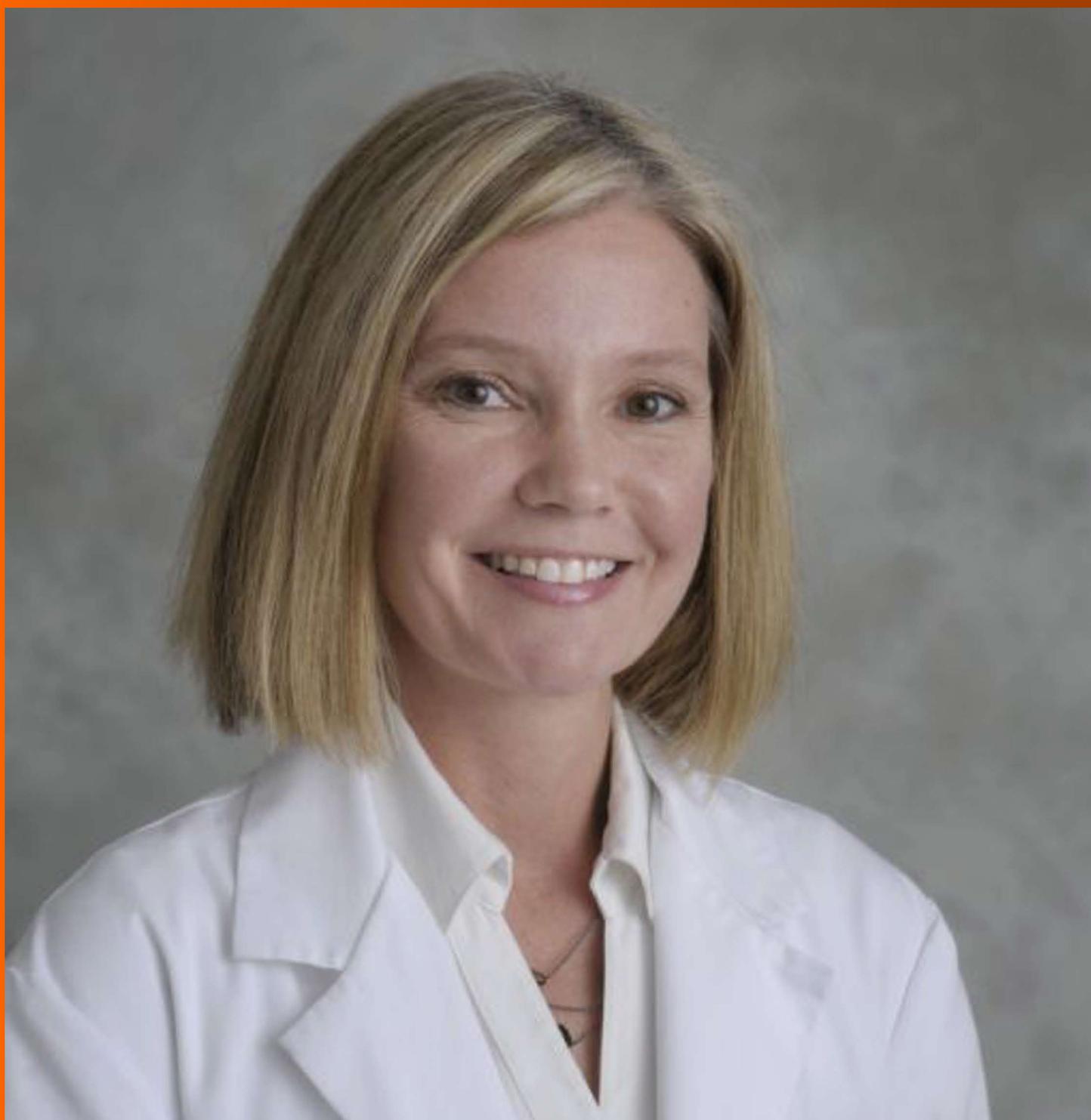


World Journal of *Diabetes*

World J Diabetes 2015 August 25; 6(10): 1122-1178



Editorial Board

2011-2015

The *World Journal of Diabetes* Editorial Board now consists of 712 members, representing a team of worldwide experts in diabetes mellitus. They are from 56 countries, including Argentina (2), Australia (27), Austria (11), Belgium (5), Brazil (13), Canada (25), Chile (3), China (40), Cuba (1), Czech Republic (3), Denmark (16), Egypt (3), Finland (5), France (12), Germany (27), Greece (17), Hungary (4), India (28), Iran (8), Iraq (2), Ireland (3), Israel (10), Italy (56), Japan (30), Jordan (1), Kuwait (3), Lebanon (1), Malaysia (1), Malta (1), Mexico (4), Netherlands (9), New Zealand (3), Nigeria (2), Norway (2), Oman (3), Pakistan (2), Poland (7), Portugal (1), Qatar (1), Romania (2), Saudi Arabia (1), Singapore (4), Slovakia (1), South Africa (1), South Korea (15), Spain (24), Sweden (5), Switzerland (4), Thailand (4), Tunisia (1), Turkey (13), United Arab Emirates (3), United Kingdom (27), United States (213), Venezuela (1), and Yemen (1).

EDITOR-IN-CHIEF

Lu Qi, *Boston*
Jingbo Zhao, *Aalborg*

STRATEGY ASSOCIATE EDITOR-IN-CHIEF

Undurti Narasimha Das, *Shaker Heights*
Min Du, *Laramie*
Gregory I Liou, *Augusta*
Zhong-Cheng Luo, *Quebec*
Demosthenes B Panagiotakos, *Athens*

GUEST EDITORIAL BOARD MEMBERS

Juei-Tang Cheng, *Tainan*
Chih-Hsung Chu, *Kaohsiung*
Low-Tone (Larry) Ho, *Taipei*
Cheng-Cheng Hsiao, *Keelung*
Yung-Hsi Kao, *Taoyuan*
Chi Feng Liu, *Taipei*
Shing-Hwa Liu, *Taipei*
Wayne H-H Sheu, *Taichung*
Eing-Mei Tsai, *Kaohsiung*
Chin-Hsiao Tseng, *Taipei*
Yen Tzung-Hai, *Taipei*
Ching-Shuang Wu, *Kaohsiung*
Wei-Chung Vivian Yang, *Taipei*
Wen-Chin Yang, *Taipei*

MEMBERS OF THE EDITORIAL BOARD



Argentina

Justo P Castaño, *Cordoba*
Eduardo Spinedi, *La Plata*



Australia

Sof Andrikopoulos, *Heidelberg Heights*
Hugh Russell Barrett, *Perth*
Bernhard T Baune, *Townsville*
Grant Brinkworth, *Adelaide*
Louise Janet Maple Brown, *Casuarina*
Melinda Therese Coughlan, *Melbourne*
Josephine Maree Forbes, *Melbourne*
Paul A Fournier, *Perth*
Angela Gialamas, *Adelaide*
Mark Douglas Gorrell, *Newtown*
Graeme Hankey, *Perth*
Anandwardhan A Hardikar, *Melbourne*
Michael Horowitz, *Adelaide*
Karin Jandeleit-Dahm, *Melbourne*
Martha Lappas, *Victoria*
Peter J Little, *Melbourne*
Xin Liu, *Brisbane*
Dianna Josephine Magliano, *Caulfield*
Robyn McDermott, *Adelaide*
Beverly Sara Muhlhausler, *Adelaide*
Christopher Nolan, *Canberra*
Luciano Pirola, *Melbourne*
Maryam Rashidi, *Victoria*
Karly Calliopi Sourris, *Victoria*
Greg Tesch, *Clayton*
Jack Ronald Wall, *Penrith*
Owen Llewellyn Woodman, *Bundooora*



Austria

Christian Heinz Anderwald, *Vienna*
Helmuth Martin Borkenstein, *Graz*
Walter Hermann Hörl, *Vienna*
Alexandra Kautzky-Willer, *Vienna*

Friedrich Mittermayer, *Vienna*
Markus Paulmichl, *Salzburg*
Stefan Pilz, *Graz*
Guntram Schernthaner, *Vienna*
Harald Sourij, *Graz*
Thomas Michael Stulnig, *Vienna*
Ludwig Wagner, *Vienna*



Belgium

Giovanni Dapri, *Brussels*
Christophe De Block, *Antwerp*
Ekaterine Tskitishvili, *Liege*
F Andre Van Assche, *Leuven*
Luc F Van Gaal, *Antwerp*



Brazil

Monica Levy Andersen, *Vila Clementino*
Claudia RL Cardoso, *Rio de Janeiro*
Ricardo Vitor Cohen, *São Paulo*
Marcelo Correia, *Rio de Janeiro*
Cassyano Januario Correr, *Curitiba*
Matheus Roriz Cruz, *Porto Alegre*
Cintia Chaves Curioni, *Rio de Janeiro*
Freddy Goldberg Eliaschewitz, *Rua Goiás*
Rodrigo Jorge, *Ribeirão Preto*
Luciana Ansaneli Naves, *Asa Norte*
Júlio César Voltarelli, *Ribeirão Preto*
Bernardo L Wajchenberg, *Pinheiros*
Jacqueline Nelisis Zanon, *Maringá*



Canada

Jean-Luc Ardilouze, *Sherbrooke*

Subrata Chakrabarti, *London*
 David Cherney, *Ontario*
 Mervyn Deitel, *Toronto*
 Jean-Pierre Després, *Quebec*
 David Joseph Hill, *London*
 Tian-Ru Jin, *Toronto*
 Arulmozhi D Kandasamy, *Edmonton*
 Jennifer L Kuk, *Toronto*
 Ismail Laher, *Vancouver*
 Roger S McIntyre, *Toronto*
 David Meyre, *Ontario*
 Joseph Fomusi Ndisang, *Saskatoon*
 Raj Padwal, *Alberta*
 Ciriaco A Piccirillo, *Montreal*
 Remi Rabasa-Lhoret, *Montreal*
 AM James Shapiro, *Edmonton*
 Guang Sun, *St. John's*
 Valerie Taylor, *Hamilton*
 Cory Toth, *Calgary*
 André Tremblay, *Montréal*
 Vincent C Woo, *Winnipeg*
 James Roscoe Wright, *Calgary*
 Xi-Long Zheng, *Calgary*



Chile

Sebastian San Martin, *Valparaiso*
 Armando Rojas-Rubio, *Talca*
 Luis Sobrevia, *Santiago*



China

Pang-Zeng Chang, *Qingdao*
 Jie Chen, *Nanjing*
 Bernard Man Yung Cheung, *Hong Kong*
 William Chi-shing Cho, *Hong Kong*
 Tian-Pei Hong, *Beijing*
 Qin Huang, *Shanghai*
 Po Sing Leung, *Hong Kong*
 Chao Liu, *Nanjing*
 Jian-Kang Liu, *Xi'an*
 Lie-Gang Liu, *Wuhan*
 Ronald Ching Wan Ma, *Hong Kong*
 Jin-Sheng Qi, *Shijiazhuang*
 Wing Yee So, *Hong Kong*
 Cheuk Chun Szeto, *Hong Kong*
 Kathryn Tan, *Hong Kong*
 Cheng-Ming Wang, *Yangzhou*
 Cong-Yi Wang, *Wuhan*
 Yu Wang, *Hong Kong*
 Guang-Da Xiang, *Wuhan*
 Bao-Feng Yang, *Harbin*
 Shu-Yu Yang, *Fujian*
 Xi-Lin Yang, *Hong Kong*
 Zai-Qing Yang, *Wuhan*
 Shan-Dong Ye, *Hefei*
 Shi-Sheng Zhou, *Dalian*
 Zhi-Guang Zhou, *Changsha*



Cuba

Luis Sarmiento-Pérez, *Havana*



Czech Republic

Martin Haluzik, *Prague*

Michal Krcma, *Plzen*
 Terezie Pelikanova, *Prague*



Denmark

Charlotte Brøns, *Gentofte*
 Jens Sandahl Christiansen, *Aarhus*
 Flemming Dela, *Copenhagen*
 Kristine Færch, *Gentofte*
 Erik L Grove, *Aarhus*
 Louise Groth Grunnet, *Gentofte*
 R Scott Heller, *Gentofte*
 Kurt Højlund, *Odense C*
 Filip K Knop, *Hellerup*
 Helle Markholst, *Måløv*
 Jens D Mikkelsen, *Copenhagen*
 Ole Hartvig Mortensen, *Copenhagen*
 Oluf Pedersen, *Copenhagen*
 Esben Thyssen Vestergaard, *Aarhus*
 Milan Zdravkovic, *Søborg*



Egypt

Mamdouh Moawad Ali Hssan, *Cairo*
 Moshira Abdel Hakim Rateb, *Cairo*
 Mona Farag Schaalán, *Cairo*



Finland

Siamak Bidel, *Helsinki*
 Gang Hu, *Helsinki*
 Thomas Kietzmann, *Oulu*
 Qing Qiao, *Helsinki*
 Karoliina Wehkalampi, *Helsinki*



France

Jean Claude Ansquer, *Dijon*
 Bertrand Cariou, *Nantes*
 Sylvie Dejager, *Rueil-Malmaison*
 Naim Akhtar Khan, *Dijon*
 Jean-Philippe Lavigne, *Nîmes*
 Michel Marre, *Paris*
 Marie-Claude Morice, *Massy*
 Riccardo Perfetti, *Paris*
 Gérard Said, *Paris*
 Sophie Visvikis Siest, *Nancy*
 Dominique Simon, *Paris*
 Didier Vieau, *Villeneuve d'Ascq*



Germany

Ioanna Gouni Berthold, *Cologne*
 Christa Buechler, *Regensburg*
 Roland Büttner, *Heidelberg*
 Michael Froehner, *Dresden*
 Hammes Hans-Peter, *Mannheim*
 Nadj Herbach, *Munich*
 Andrea Icks, *Düsseldorf*
 Thomas Jax, *Neuss*
 Ulrich Arthur Julius, *Dresden*
 Michael Kluge, *Munich*
 Florian Lang, *Tuebingen*
 Matthias Laudes, *Köln*
 Ralf Lobmann, *Stuttgart*

Rafael T Mikolajczyk, *Bremen*
 Andreas Stefan Mueller, *Halle (Saale)*
 Karsten Müssig, *Tübingen*
 Nahid Parvizi, *Neustadt am Rübenberge*
 Thomas Peter Reinehr, *Datteln*
 Michael Ristow, *Jena*
 Sven Schinner, *Duesseldorf*
 Peter Egbert Hermann Schwarz, *Dresden*
 Konstantinos Stellos, *Tubingen*
 Ovidiu Alin Stirban, *Bad Oeynhausen*
 Diego J Walther, *Berlin*
 Silvia Anette Wein, *Kiel*
 Christian Wrede, *Berlin*
 Dan Ziegler, *Düsseldorf*



Greece

George P Chrousos, *Athens*
 Moses S Elisaf, *Ioannina*
 Panagiotis Georgoulis, *Larissa*
 Nikolaos Kadoglou, *Thessaloniki*
 Gerasimos E Krassas, *Krini*
 Spilios Manolakopoulos, *Attiki*
 Nikolaos Papanas, *Alexandroupolis*
 Dimitrios Papazoglou, *Alexandroupolis*
 Sokratis Pastromas, *Athens*
 Melpomeni Peppas, *Athens*
 Christina Piperi, *Goudi*
 Nicholas K Tentolouris, *Athens*
 Konstantinos A Toulis, *Salonika*
 Apostolos Tsapas, *Thessaloniki*
 Konstantinos Tziomalos, *Thessaloniki*
 Elias Zintzaras, *Thessaly*



Hungary

Mária Bagyánszki, *Szeged*
 György Jermendy, *Budapest*
 Karoly Racz, *Budapest*
 Gyula Soltesz, *Pécs*



India

Deepak Narayan Amrapurkar, *Mumbai*
 C V Anuradha, *Tamil Nadu*
 Sarika Arora, *New Delhi*
 Pitchai Balakumar, *Sivakasi*
 Muthuswamy Balasubramanyam, *Chennai*
 Subhabrata Chakrabarti, *Hyderabad*
 Abhay Sankar Chakraborti, *Kolkata*
 Tapan K Chaudhuri, *New Delhi*
 Kanwaljit Chopra, *Chandigarh*
 Malabika Datta, *Delhi*
 Debidas Ghosh, *West Bengal*
 Ravinder Goswami, *New Delhi*
 Pappachan M Joseph, *Kerala*
 Jothydev Kesavadev, *Kerala*
 KVS Hari Kumar, *Lucknow*
 Anoop Misra, *New Delhi*
 Analava Mitra, *Kharagpur*
 Viswanathan Mohan, *Chennai*
 S P Murthy, *Bangalore*
 Pallavi Panchu, *Guntur*
 Usharani Pingali, *Hyderabad*
 Ambady Ramachandran, *Egmore Chennai*
 Vadde Ramakrishna, *Kadapa*

Geetha Vani Rayasam, *Haryana*
Rajat Sandhir, *Chandigarh*
Manju Sharma, *New Delhi*
Suman Bala Sharma, *Delhi*
Tarun Sharma, *Chennai*



Iran

Mohammad Abdollahi, *Tehran*
Mohammad Kazemi Arababadi, *Rafsanjan*
Leila Azadbakht, *Isfahan*
Hamid Baradaran, *Tehran*
Behrooz Broumand, *Tehran*
Ahmad Esmailzadeh, *Isfahan*
Majid Ghayour-Mobarhan, *Mashhad*
Mohsen Janghorbani, *Isfahan*



Iraq

Saad Abdul-Rahman Hussain, *Baghdad*
Abbas Ali Mansour, *Basrah*



Ireland

Amar Agha, *Dublin*
Mark Philip Hehir, *Dublin*
Gerald H Tomkin, *Dublin*



Israel

Michael Aviram, *Haifa*
Gal Dubnov-Raz, *Tel Hashomer*
Shimon Efrat, *Tel Aviv*
Raymond Elias Farah, *Safed*
Oren Froy, *Rehovot*
Saher Hamed, *Haifa*
Arid Nakhoul, *Haifa*
Orit Pinhas-Hamiel, *Tel Hashomer*
Haim Werner, *Tel Aviv*
Marina Shargorodsky Zimlichman, *Holon*



Italy

Luigi Angrisani, *Napoli*
Moschetta Antonio, *Bari*
Antonio Aversa, *Rome*
Roberto Baldelli, *Rome*
Giuseppe Barbaro, *Rome*
Alessandro Bartolomucci, *Parma*
Giuseppina Basta, *Pisa*
Simona Bertoli, *Milano*
Federico Bilotta, *Rome*
Fabio Broglio, *Torino*
Francesco G Chiarelli, *Chieti*
Sergio Coccheri, *Bologna*
Massimo Collino, *Torino*
Marco Aristide Comaschi, *Genoa*
Renzo Cordera, *Genova*
Francesco Dotta, *Siena*
Gagliardini Elena, *Bergamo*
Stefano Fiorucci, *Perugia*
Maurizio Galderisi, *Naples*
Amalia Gastaldelli, *Pisa*

Ezio Ghigo, *Turin*
Carla Giordano, *Palermo*
Paolo Gisondi, *Verona*
Riccarda Granata, *Turin*
Giorgio Iervasi, *Pisa*
Claudia Kusmic, *Pisa*
Carmelo La Rosa, *Catania*
Francesco Landi, *Rome*
Monica Rosa Loizzo, *Arcavacata Rende*
Paolo Magni, *Milano*
Mariano Malaguarnera, *Catania*
Melania Manco, *Rome*
Piero Marchetti, *Pisa*
Massimo Massi-Benedetti, *Perugia*
Antonio Nicolucci, *Imbaro*
Lucia Pacifico, *Rome*
Stefano Palomba, *Catanzaro*
Giampaolo Papi, *Carpi*
Renato Pasquali, *Bologna*
Piermarco Piatti, *Milano*
Dario Pitocco, *Rome*
Antonio E Pontiroli, *Milano*
Giulio Marchesini Reggiani, *Bologna*
Giuseppe Remuzzi, *Bergamo*
Manfredi Rizzo, *Palermo*
Raffaella Rosso, *Genoa*
Giuseppe Schillaci, *Perugia*
Leonardo A Sechi, *Sassari*
Imad Sheiban, *Torino*
Cesare R Sirtori, *Milano*
Giovanni Tarantino, *Naples*
Giovanni Targher, *Verona*
Donadon Valter, *Pordenone*
Alberto Verrotti, *Chieti*
Andrea Viggiano, *Napoli*
Gianvincenzo Zuccotti, *Milan*



Japan

Masato Asahina, *Chiba*
Takuya Awata, *Saitama*
Yuichiro Eguchi, *Saga*
Goji Hasegawa, *Kyoto*
Satoshi Inoue, *Tokyo*
Eiji Ishimura, *Osaka*
Masayuki Iwano, *Nara*
Takashi Kadowaki, *Tokyo*
Eisuke Kagawa, *Hiroshima*
Masahito Katahira, *Aichi*
Eiji Kawasaki, *Nagasaki*
Noriyuki Koibuchi, *Gunma*
Kazuhiko Kotani, *Tochigi*
Daisuke Koya, *Ishikawa*
Norikazu Maeda, *Osaka*
Takayuki Masaki, *Oita*
Yuji Matsuzawa, *Osaka*
Kazuaki Nishio, *Tokyo*
Kenji Okumura, *Nagoya*
Motoaki Saito, *Yonago*
Toshiyasu Sasaoka, *Toyama*
Michio Shimabukuro, *Okinawa*
Kohzo Takebayashi, *Saitama*
Hiroyuki Tamemoto, *Tochigi*
Takashi Togo, *Yokohama*
Jun Udagawa, *Izumo*
Yoshinari Uehara, *Fukuoka*
Takuya Watanabe, *Tokyo*
Toshihiko Yada, *Tochigi*

Tohru Yorifuji, *Osaka*



Jordan

Yousef S Khader, *Irbid*



Kuwait

Kamal AA Sulaiman Al-Shoumer, *Kuwait*
Ibrahim Fadil Benter, *Safat*
Abu Salim Mustafa, *Kuwait*



Lebanon

Ramzi F Sabra, *Beirut*



Malaysia

Mafauzy Mohamed, *Kota Bharu*



Malta

Charles Savona-Ventura, *Msida*



Mexico

Manuel González-Ortiz, *Guadalajara*
Fernando Guerrero-Romero, *Durango*
Jesus Alberto Olivares-Reyes, *Mexico City*
Rocío Salceda, *Mexico City*



Netherlands

Sander Kersten, *Wageningen*
Nanne Kleefstra, *Zwolle*
Edwin Mariman, *Maastricht*
Don Poldermans, *Rotterdam*
François Pouwer, *Tilburg*
Han Roelofsen, *Groningen*
Hendrik-Jan Schuurman, *Utrecht*
Suat Simsek, *Alkmaar*
Marcel Twickler, *Bergen op Zoom*



New Zealand

Paul Hofman, *Auckland*
Peter E Lobie, *Auckland*
Elaine Rush, *Auckland*



Nigeria

Adejuwon A Adeneye, *Lagos*
Anthonia Okeoghene Ogbera, *Lagos*



Norway

Akhtar Hussain, *Oslo*
Odd Erik Johansen, *Hovik*

**Oman**

Mohammed Al Shafae, *Muscat*
 Jumana S Saleh, *Muscat*
 Radha Shenoy, *Muscat*

**Pakistan**

Shahid Hameed, *Islamabad*
 Jamil A Malik, *Islamabad*

**Poland**

Marcin Baranowski, *Bialystok*
 Jerzy Beltowski, *Lublin*
 Alicia Hubalewska Dydejczyk, *Krakow*
 Maciej Owecki, *Poznań*
 Ewa Pankowska, *Warsaw*
 Agnieszka Piwowar, *Wroclaw*
 Dorota Anna Zieba, *Krakow*

**Portugal**

M Graça Pereira, *Braga*

**Qatar**

Hong Ding, *Doha*

**Romania**

Elena Ganea, *Bucharest*
 Adriana Georgescu, *Bucharest*

**Saudi Arabia**

J Fernando Arevalo, *Caracas*

**Singapore**

S Thameem Dheen, *Singapore*
 Yung Seng Lee, *Singapore*
 Daniel Ng, *Singapore*
 Rob Martinus van Dam, *Singapore*

**Slovakia**

Katarína Šebeková, *Bratislava*

**South Africa**

Md Shahidul Islam, *Durban*

**South Korea**

Huneg-Sik Choi, *Gwangju*
 Kyung Mook Choi, *Seoul*
 Won Mi Hwang, *Seoul*
 Eui-Bae Jeung, *Chungbuk*

Ju-Hee Kang, *Incheon*
 Sin Gon Kim, *Seongbuk-Gu*
 Sung-Jin Kim, *Seoul*
 Young-Gyu Ko, *Seoul*
 Kang-Beom Kwon, *Chonbuk*
 Myung Gull Lee, *Bucheon*
 Soo Lim, *Seongnam*
 Byung-Hyun Park, *Jeonbuk*
 Seungjoon Park, *Seoul*
 Kun-Ho Yoon, *Seoul*
 Jeesuk Yu, *Cheonan*

**Spain**

Vivencio Barrios, *Madrid*
 M Lusía Bonet, *Palma de Mallorca*
 Manuel Vazquez Carrera, *Barcelona*
 Maria Luz Martinez Chantar, *Derio*
 Manuel Aguilar Diosdado, *Cádiz*
 Javier Espino, *Badajoz*
 Ricardo V García-Mayor, *Vigo*
 José Manuel Gómez-Sáez, *Barcelona*
 Oreste Gualillo, *Santiago de Compostela*
 J Alfredo Martínez Hernández, *Pamplona*
 Emilio Herrera, *Madrid*
 Amelia Marti, *Pamplona*
 Merce Miranda, *Tarragona*
 JF Navarro-González, *Santa Cruz de Tenerife*
 Alberto Ortiz, *Madrid*
 Maria Javier Ramirez, *Pamplona*
 Eugenia Resmini, *Barcelona*
 Pedro Romero-Aroca, *Reus*
 Jordi Salas-Salvadó, *Reus*
 Gines M Salido, *Caceres*
 Victor Sanchez-Margalet, *Seville*
 Helmut Schröder, *Barcelona*
 Carmen Segundo, *Cádiz*
 Rafael Simó, *Barcelona*

**Sweden**

Joanna Hlebowicz, *Malmö*
 Kaj S Stenlöf, *Göteborg*
 Ann-Britt Wiréhn, *Linköping*
 Weili Xu, *Stockholm*
 Shao-Nian Yang, *Stockholm*

**Switzerland**

Kaspar Berneis, *Zurich*
 Pascal Bovet, *Lausanne*
 Luc Tappy, *Lausanne*
 Christian Toso, *Geneva*

**Thailand**

Narattaphol Charoenphandhu, *Bangkok*
 Arthorn Riewpaiboon, *Bangkok*
 Rawee Teanpaisan, *Hat-Yai*
 Viroj Wiwanitkit, *Bangkok*

**Tunisia**

Khaled Hamden, *Sfax*

**Turkey**

Ugur Cavlak, *Denizli*
 Teoman Dogru, *Ankara*
 Ersin Fadillioğlu, *Ankara*
 Abdurrahman Fatih Fidan, *Afyonkarahisar*
 Muammer Karadeniz, *Bornova-Izmir*
 Cevdet Kaya, *Istanbul*
 Fahrettin Kelestimur, *Kayseri*
 Altan Onat, *Istanbul*
 Semir Ozdemir, *Antalya*
 Mustafa Şahin, *Ankara*
 Ilker Tasci, *Ankara*
 Belma Turan, *Ankara*
 Serap Yalın, *Mersin*

**United Arab Emirates**

Ernest Akingunola Adegate, *Al Ain*
 Mukesh M Agarwal, *Al Ain*
 Samir M Awadallah, *Sharjah*

**United Kingdom**

Nisreen Alwan, *Leeds*
 Ambika P Ashraf, *Birmingham*
 Chen Bing, *Liverpool*
 Fay Crawford, *Edinburgh*
 Tim M Curtis, *Belfast*
 Umesh Dashora, *Hastings*
 Gareth Davison, *Belfast*
 Peter Raymond Flatt, *Coleraine*
 Kathleen M Gillespie, *Bristol*
 Peter John Grant, *Leeds*
 Lorna W Harries, *Exeter*
 Nigel Hoggard, *Aberdeen*
 Nigel Irwin, *Coleraine*
 Edward Jude, *Lancashire*
 Andreas F Kolb, *Aberdeen*
 Stefan Marciniak, *Cambridge*
 Moffat Joha Nyirenda, *Edinburgh*
 Jeetesh Patel, *Birmingham*
 Snorri Bjorn Rafnsson, *Edinburgh*
 Thozhukat Sathyapalan, *Yorkshire*
 Latika Sibal, *Newcastle*
 Rajagopalan Sriraman, *Lincoln*
 Ramasamyiyer Swaminathan, *London*
 Abd A Tahrani, *Birmingham*
 G Neil Thomas, *Birmingham*
 Cecil Thompson, *London*
 Paul Henry Whiting, *Leicester*

**United States**

Varun Agrawal, *Springfield*
 Mohamed Al-Shabrawey, *Augusta*
 Pascale Alard, *Louisville*
 Omar Ali, *Milwaukee*
 Judith Aponte, *New York*
 Balamurugan N Appakalai, *Minneapolis*
 Hwya A Arafat, *Philadelphia*
 Carl V Asche, *Salt Lake*
 Sanford A Asher, *Pittsburgh*
 Anthony Atala, *Winston-Salem*
 Sami Toufic Azar, *Beirut*

George Louis Bakris, *Chicago*
 Alistair J Barber, *Hershey*
 Daniel C Battle, *Chicago*
 David SH Bell, *Birmingham*
 Rita Bortell, *Worcester*
 Sebastien G Bouret, *Los Angeles*
 David Lloyd Brown, *Stony Brook*
 Lu Cai, *Louisville*
 Jack D Caldwell, *Erie*
 Anna C Calkin, *Los Angeles*
 Roberto A Calle, *Groton*
 R Keith Campbell, *Pullman*
 Carlos Campos, *New Braunfels*
 Heping Cao, *New Orleans*
 Krista Casazza, *Birmingham*
 Aaron Brandon Caughey, *Portland*
 Eileen R Chasens, *Pittsburgh*
 Munmun Chattopadhyay, *Ann Arbor*
 Xiao-Li Chen, *St Paul*
 Sheri Renee Colberg, *Norfolk*
 Craig Ian Coleman, *Hartford*
 Robert Russell Conley, *Indianapolis*
 Colleen M Croniger, *Cleveland*
 Doyle M Cummings, *Greenville*
 William C Cushman, *Memphis*
 Patricia Ann D'Amore, *Boston*
 Patricia Darbishire, *West Lafayette*
 Guillaume Darrasse-Jéze, *New York*
 Ravi M Dasu, *Sacramento*
 Michael Harvey Davidson, *Chicago*
 Prakash Deedwania, *San Francisco*
 Hong-Wen Deng, *Kansas City*
 Teresa P DiLorenzo, *Bronx*
 Scot E Dowd, *Lubbock*
 Samuel C Durso, *Baltimore*
 Krystal L Edwards, *Dallas*
 Alexander M Efanov, *Indianapolis*
 Azza B El-Remessy, *Augusta*
 Amy Zhihong Fan, *Atlanta*
 Melissa Spezia Faulkner, *Tucson*
 George S Ferzli, *Staten Island*
 Paolo Fiorina, *Boston*
 James Edward Foley, *East Hanover*
 Samuel N Forjuoh, *Temple*
 Alessia Fornoni, *Miami*
 Martha M Funnell, *Ann Arbor*
 Trudy Gaillard, *Columbus*
 Pietro Galassetti, *Irvine*
 Claudia Gragnoli, *Hershey*
 Jennifer B Green, *Durham*
 Gary J Grover, *Piscataway*
 Alok Kumar Gupta, *Baton Rouge*
 Werner K Gurr, *New Haven*
 Samy L Habib, *San Antonio*
 Abdel Rahim Hamad, *Baltimore*
 Daniel M Herron, *New York*
 Tiffany Hilton, *Rochester*
 Raimund Hirschberg, *Torrance*
 Michael Francis Holick, *Boston*
 Zhaoyong Hu, *Houston*
 Rachel Mary Hudacko, *New Brunswick*
 Yasuo Ido, *Boston*
 Brian K Irons, *Lubbock*
 Pamela Itkin-Ansari, *La Jolla*
 Hieronim Jakubowski, *Newark*
 Hong-Lin Jiang, *Blacksburg*
 Ping Jiao, *Providence*
 Shengkan Jin, *Piscataway*
 Arpita Kalla, *St Louis*
 Richard Evers Katholi, *Springfield*

Melina Rae Kibbe, *Chicago*
 Bhumsoo Kim, *Ann Arbor*
 Tomoshige Kino, *Bethesda*
 Julienne K Kirk, *Winston-Salem*
 Renu A Kowluru, *Detroit*
 Lewis H Kuller, *Pittsburgh*
 Rajesh Kumar, *Temple*
 Blandine Laferrère, *New York*
 Sang Yeoup Lee, *Mayo*
 Cong-Jun Li, *Beltsville*
 Ching-Shwun Lin, *San Francisco*
 Julie Lin, *Boston*
 Shuo Lin, *Los Angeles*
 Peter Lindgren, *San Diego*
 James F List, *Princeton*
 Dong-Min Liu, *Blacksburg*
 Zhen-Qi Liu, *Charlottesville*
 George William Lyerly, *Conway*
 Jian-Xing Ma, *Oklahoma City*
 Rong Ma, *Fort Worth*
 Xin-Laing Ma, *Philadelphia*
 David Maggs, *San Diego*
 Kenneth Maiese, *Detroit*
 Kevin C Maki, *Glen Ellyn*
 Sridhar Mani, *Bronx*
 Suresh Mathews, *Auburn*
 Lauraar McCabe, *East Lansing*
 Sarah E Messiah, *Miami*
 Thomas O Metz, *Richland*
 Shannon A Miller, *Orlando*
 Murielle Mimeault, *Omaha*
 Raghavendra G Mirmira, *Indianapolis*
 Prasun J Mishra, *Bethesda*
 Reema Mody, *Grayslake*
 Arshag D Mooradian, *Jacksonville*
 Mohammad Reza Movahed, *Tucson*
 James Mu, *Rahway*
 Muraleedharan G Nair, *East Lansing*
 Manuel F Navedo, *Seattle*
 Charles B Nemeroff, *Atlanta*
 Joshua J Neumiller, *Spokane*
 Steven Nissen, *Cleveland*
 Hirofumi Noguchi, *Fort Worth*
 Craig Nunemake, *Charlottesville*
 Patrick J O'Connor, *Minneapolis*
 Erin St Onge, *Apopka*
 Wei-Hong Pan, *Baton Rouge*
 Naushira Pandya, *Fort Lauderdale*
 Michael R Peluso, *Corvallis*
 Inga Peter, *New York*
 Axel Pflueger, *Rochester*
 Gretchen A Piatt, *Pittsburgh*
 John D Piette, *Ann Arbor*
 Leonid Poretsky, *New York*
 Walter J Pories, *Greenville*
 Parviz M Pour, *Omaha*
 Wei Qiao Qiu, *Boston*
 Teresa Quattrin, *Buffalo*
 Cristina Rabadán-Diehl, *Bethesda*
 Rajendra S Raghov, *Memphis*
 Swapnil Rajpathak, *Bronx*
 Armin Rashidi, *Norfolk*
 Mohammed S Razzaque, *Boston*
 Beverly A S Reyes, *Philadelphia*
 David Rodbard, *Potomac*
 Helena W Rodbard, *Rockville*
 June Hart Romeo, *Cleveland*
 Raul J Rosenthal, *Fort Lauderdale*
 Juan M Saavedra, *Bethesda*
 Stephen W Schaffer, *Mobile*

Frank AJL Scheer, *Boston*
 Richard E Scranton, *Tiverton*
 Vallabh (Raj) O Shah, *Albuquerque*
 Aziz Shaibani, *Houston*
 Jin-Xiong She, *Augusta*
 Guo-Ping Shi, *Boston*
 Carol Ann Shively, *Winston-Salem*
 Anders AF Sima, *Detroit*
 Pramit N Singh, *Loma Linda*
 Rajan Singh, *Los Angeles*
 Jay S Skyler, *Miami*
 Dawn Smiley, *Atlanta*
 Matthew D Solomon, *Stanford*
 Mark A Sperling, *Pittsburgh*
 Rakesh K Srivastava, *Tyler*
 Bangyan Stiles, *Los Angeles*
 Yu-Xiang Sun, *Houston*
 Salim Surani, *Corpus Christi*
 Arthur L M Swislocki, *Martinez*
 Ya-Xiong Tao, *Auburn*
 John A Tayek, *Torrance*
 John Gaylord Teeter, *New Haven*
 Carlos Marcelo Telleria, *Vermillion*
 Christopher Gordon Thanos, *Providence*
 Ronald G Tilton, *Galveston*
 Serena Tonstad, *Loma Linda*
 Michael Lawrence Traub, *Staten Island*
 Guillermo E Umpierrez, *Atlanta*
 Margrit Urbanek, *Chicago*
 Vladimir N Uversky, *Indianapolis*
 Gabriel I Uwaifo, *Baton Rouge*
 Volker Vallon, *San Diego*
 Shambhu D Varma, *Baltimore*
 Maria Virella, *Charleston*
 Hong-Jun Wang, *Boston*
 Mark E Williams, *Boston*
 Nathan D Wong, *Irvine*
 Guangyu Wu, *New Orleans*
 Zhong-Jian Xie, *San Francisco*
 Ming-Zhao Xing, *Baltimore*
 Hariom Yadav, *Bethesda*
 Lijun Yang, *Gainesville*
 Ruoqing Yang, *Rahway*
 Subhashini Yaturu, *Albany*
 Joseph Yeboah, *Charlottesville*
 Dengping Yin, *Nashville*
 Yisang Yoon, *Rochester*
 Yi-Hao Yu, *New York*
 Kevin CJ Yuen, *Portland*
 Ian Stuart Zagon, *Hershey*
 Robert Yuk-Lung Zee, *Boston*
 Cui-Lin Zhang, *Rockville*
 James Xuejie Zhang, *Richmond*
 Sarah Zhang, *Oklahoma*
 Guixiang Zhao, *Atlanta*
 Yang Zhao, *Indianapolis*
 Ming-Hui Zou, *Oklahoma City*



Venezuela

Fuad Lechin, *Caracas*



Yemen

Khaled Abdul-Aziz Ahmed, *Ibb*



EDITORIAL

- 1122 Effect of proton pump inhibitors on glyceic control in patients with diabetes
Takebayashi K, Inukai T

REVIEW

- 1132 Dysglycemia after renal transplantation: Definition, pathogenesis, outcomes and implications for management
Langsford D, Dwyer K

MINIREVIEWS

- 1152 Magnesium and type 2 diabetes
Barbagallo M, Dominguez LJ
- 1158 Vitamin paradox in obesity: Deficiency or excess?
Zhou SS, Li D, Chen NN, Zhou Y

ORIGINAL ARTICLE

Basic Study

- 1168 Simvastatin, atorvastatin, and pravastatin equally improve the hemodynamic status of diabetic rats
Crespo MJ, Quidgley J

ABOUT COVER

Editorial Board Member of *World Journal of Diabetes*, Shannon A Miller, PharmD, Pharmacotherapy Faculty, Florida Hospital East, Family Practice Residency, Orlando, FL 32822, United States

AIM AND SCOPE

World Journal of Diabetes (*World J Diabetes, WJD*, online ISSN 1948-9358, DOI: 10.4239), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJD covers topics concerning α , β , δ and PP cells of the pancreatic islet, the effect of insulin and insulinresistance, pancreatic islet transplantation, adipose cells and obesity.

We encourage authors to submit their manuscripts to *WJD*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great clinical significance.

INDEXING/ ABSTRACTING

World Journal of Diabetes is now indexed in Thomson Reuters Web of Science Emerging Sources Citation Index, PubMed Central, PubMed, Digital Object Identifier, and Directory of Open Access Journals.

FLYLEAF

I-V Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Li Xiang*
Responsible Electronic Editor: *Xiao-Kang Jiao*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Fang-Fang Ji*
Proofing Editorial Office Director: *Xiu-Xia Song*

NAME OF JOURNAL
World Journal of Diabetes

ISSN
 ISSN 1948-9358 (online)

LAUNCH DATE
 April 15, 2010

FREQUENCY
 Biweekly

EDITORS-IN-CHIEF
Lu Qi, MD, PhD, Assistant Professor, Department of Nutrition, Harvard School of Public Health, 665 Huntington Ave., Boston, MA 02115, United States

Jingbo Zhao, PhD, Associate Professor, Aalborg Hospital Science and Innovation Centre, Aalborg Hospital, Aarhus University Hospital, Aalborg 9000, Denmark

EDITORIAL OFFICE
 Jin-Lei Wang, Director

Xiu-Xia Song, Vice Director
World Journal of Diabetes
 Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China
 Telephone: +86-10-85381891
 Fax: +86-10-85381893
 E-mail: editorialoffice@wjgnet.com
 Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>
<http://www.wjgnet.com>

PUBLISHER
 Baishideng Publishing Group Inc
 8226 Regency Drive, Pleasanton, CA 94588, USA
 Telephone: +1-925-223-8242
 Fax: +1-925-223-8243
 E-mail: bpgoffice@wjgnet.com
 Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>
<http://www.wjgnet.com>

PUBLICATION DATE
 August 25, 2015

COPYRIGHT
 © 2015 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non-commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
 All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS
 Full instructions are available online at http://www.wjgnet.com/1948-9358/g_info_20100107165233.htm

ONLINE SUBMISSION
<http://www.wjgnet.com/esps/>

Effect of proton pump inhibitors on glycemic control in patients with diabetes

Kohzo Takebayashi, Toshihiko Inukai

Kohzo Takebayashi, Toshihiko Inukai, Department of Internal Medicine, Dokkyo Medical University Koshigaya Hospital, Koshigaya, Saitama 343-8555, Japan

Author contributions: Takebayashi K wrote this manuscript; Inukai T reviewed the manuscript.

Conflict-of-interest statement: There are no conflicts of interest.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Kohzo Takebayashi, MD, Department of Internal Medicine, Dokkyo Medical University Koshigaya Hospital, 2-1-50 Minamikoshigaya, Koshigaya, Saitama 343-8555, Japan. takeb@gmail.plala.or.jp
Telephone: +81-48-9651111
Fax: +81-48-9651127

Received: April 29, 2015
Peer-review started: May 3, 2015
First decision: June 24, 2015
Revised: July 6, 2015
Accepted: July 24, 2015
Article in press: July 27, 2015
Published online: August 25, 2015

Abstract

Gastrin is a linear peptide hormone which is secreted mostly in the stomach pyloric antrum G cells. Although the main role of this hormone is the promotion of the secretion of gastric acid from the stomach parietal cells, gastrin can also behave as a growth factor and

stimulate gastric cell proliferation. It is also reported that gastrin promotes β cell neogenesis in the pancreatic ductal complex, modest pancreatic β cell replication, and improvement of glucose tolerance in animal models, in which the remodeling of pancreatic tissues is promoted. These findings suggest the possibility that gastrin has the potential to promote an increase of β cell mass in pancreas, and therefore that gastrin may improve glucose tolerance. Proton pump inhibitors (PPIs) are widely used clinically for the therapy of gastro-esophageal reflux disease, gastritis due to excess stomach acid, and gastric ulcers. PPIs indirectly elevate serum gastrin levels *via* a negative feedback effect. Recent evidence has revealed the beneficial effect of PPIs on glycemic control especially in patients with type 2 diabetes mellitus (T2DM), probably *via* the elevation of the levels of serum gastrin, although the detailed mechanism remains unclear. In addition, the beneficial effects of a combination therapy of gastrin or a PPI with a glucagon-like peptide-1 receptor agonist on glycemic control in animal models have been demonstrated. Although PPIs may be possible candidates for a new approach in the therapy of diabetes, a prospective, long-term, randomized, double-blind, placebo-controlled study is needed to establish the effect of PPIs on glycemic control in a large number of patients with T2DM.

Key words: Gastrin; Proton pump inhibitors; Glycemic control; Type 2 diabetes

© The Author(s) 2015. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Recently, it is reported that gastrin may improve glucose tolerance mainly by the promotion of pancreatic β cell neogenesis. Proton pump inhibitors (PPIs) are widely used clinically for the treatment such as gastric ulcers, and it is known that PPIs indirectly elevate serum gastrin levels. Recent evidence has showed the beneficial effect of PPIs on glycemic control especially in patients with type 2 diabetes, probably

via the elevation of serum gastrin levels. Therefore, PPIs may have the potential to be candidates for a new approach in the treatment of diabetes.

Takebayashi K, Inukai T. Effect of proton pump inhibitors on glycemic control in patients with diabetes. *World J Diabetes* 2015; 6(10): 1122-1131 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v6/i10/1122.htm> DOI: <http://dx.doi.org/10.4239/wjd.v6.i10.1122>

INTRODUCTION

Gastrin is a linear peptide hormone which is secreted mostly in the stomach pyloric antrum G cells, in which high biologically active gastrin (gastrin-17 and gastrin-34) is formed^[1,2]. The secretion of gastrin is stimulated by various factors, such as considerable distension of the stomach^[3], vagal stimulation^[3,4], the presence of food (especially protein, peptides, and amino acids) in the stomach^[4-6], and high pH levels in the stomach cavity^[5,7]. Gastrin is released into the bloodstream. The main role of this hormone is the stimulation of secretion of gastric acid from the stomach parietal cells. The gastrin receptor, cholecystokinin B (CCK-B) receptor, binds to gastrin and to cholecystokinin with a similar high affinity^[8]. Gastrin can directly promote the secretion of gastric acid by binding to CCK-B receptor on parietal cells^[9,10]. However, the expression of this receptor is also found on enterochromaffin-like cells, and the binding of CCK-B receptor to gastrin on these cells promotes the secretion of the histamine resulting in subsequent promotion of the release of gastric acids by parietal cells, which may be the central mechanism of gastrin-stimulated acid secretion^[6,9-12]. Importantly, gastrin is also able to behave as a growth factor and stimulate gastric cell proliferation^[6,13]. It is reported that gastrin promotes β cell neogenesis in pancreatic ductal complex^[14], modest pancreatic β cell replication^[15], and improvement of glucose tolerance^[15] in animal models in which the remodeling of pancreatic tissues is promoted. These findings suggest the possibility that gastrin has a potential promoting effect for the increase in the pancreatic β cell mass. Therefore, gastrin improves glucose tolerance, and these effects appear to occur especially during adult pancreatic tissue remodeling but not in the normal tissue state.

Proton pump inhibitors (PPIs) are widely used clinically for the therapy of gastro-esophageal reflux disease, gastritis due to excess stomach acid, and gastric ulcers^[16]. PPIs can be orally administered as an inactive form, which enters the bloodstream from the intestine, reaches the gastric parietal cells, and is activated by crossing the cell membrane into the intracellular compartment. After converting to the active form in the unique parietal cell environment, PPIs irreversibly block the proton pump and can strongly reduce the secretion

of gastric acid promoted by either gastrin, acetylcholine, or histamine. It is well known that PPIs indirectly elevate serum gastrin levels *via* a negative feedback effect^[17-22]. Interestingly, in type 2 diabetes mellitus (T2DM) animal models, it has been reported that PPIs improved glycemic control, probably *via* possible effects on augmenting both serum levels of gastrin and β cell mass^[23]. Although some clinical studies showed negative results on glycemic control by PPIs in patients with T2DM^[24,25], most studies have demonstrated a significant improvement of glycemic control by PPI administration to these patients^[26-32]. Therefore, these agents appear to have the possibility of being a new approach for the therapy of diabetes.

BASIC STUDIES ON THE EFFECT OF GASTRIN ON THE INCREASE IN β CELL MASS

Gastrin and the CCK-B receptor are transiently expressed in fetal tissues of pancreas under period of islet neogenesis^[33-35], but no expression is observed in both adult pancreatic β cells^[36,37] and the exocrine pancreas^[34,38-40]. It has been reported that in a rat model in which the splenic portion of the pancreas is ligated (an animal model for remodeling of pancreas tissue), transdifferentiation of acinar to ductal cells is promoted, and a ductal complex consisting of a mixture transdifferentiated acinar and ductal cells is formed^[41-44]. A similar ductal complex appeared to emerge in 95% of the pancreatectomized rats (an animal model for diabetes in which pancreatic remodeling is promoted)^[15]. Although the CCK-B receptor is not expressed in adult β cells even if the pancreatic tissue is undergoing remodeling, the ductal complex shows characteristics of fetal pancreatic ductal cells in addition to those in adult, including the CCK-B receptor expression^[34]. So, it appears that gastrin is able to enhance the process of β cell neogenesis, that was already induced during the remodeling state, *via* the CCK-B receptor followed by budding from the ductal complex^[14,41]. In general, gastrin does not affect β cell replication probably because of a lack of the CCK-B receptor on β cells^[14], but there is a report suggesting that, in 95% of the pancreatectomized rats, gastrin treatment not only increased β cells neogenesis from ductal cells but also caused both a modest increase in replication and a decrease in apoptosis in β cells with the resultant improvement of glucose tolerance. The detailed mechanism for these activities remains unclear^[15]. The replication of β cells is also reported in gastrinoma patients^[45] although only β cell islets located near the gastrinomas exhibited β cell turnover despite the fact that serum levels of gastrin were elevated to the degrees to induce clinically apparent gastrointestinal symptoms. Thus, it is possible that other hormones were also involved. On other hand, the synergistic effect of other hormones, such as transforming growth factor- α ^[46], epidermal

growth factor^[47], and glucagon-like peptide-1 (GLP-1)^[48], with gastrin has also been demonstrated. For example, GLP-1 induces both β cell replication with mitogens and neogenesis of β cell from ductal cells^[49]. In combination with GLP-1, gastrin appears to enhance β cell neogenesis even when it is added in animal models, such as either *db/db* mice (a model of T2DM)^[50] or non-obese diabetic (NOD) mice [a model of type 1 diabetes mellitus (T1DM)]^[48], although, in these models, pancreatic remodeling is not necessarily occurring. In addition, an effect on regulating the autoimmune response against pancreatic β cells by combination therapy was also reported in the NOD mice model^[48]. Taken together, these effects of gastrin suggest that this hormone may possess a potential protective effect for the progression of diabetes, especially in combination with other hormones, such as GLP-1.

THE EFFECT OF PPIs ON GLYCEMIC CONTROL IN PATIENTS WITH TYPE 2 DIABETES: RESULTS OF CLINICAL STUDIES

Despite the possible effects of gastrin on both increasing β cell mass and improving glycemic control, gastrin treatment has not been used with the patients with T2DM mainly because of the difficulty with oral administration and the suggested side effects on the stomach. On the other hand, there are many publications describing the effects of PPIs on glycemic control in patients with T2DM.

Mefford *et al.*^[26] reported that a significant difference was obtained in HbA1c in patients with T2DM taking PPIs (7.0% of HbA1c, $n = 65$) vs those not taking PPIs (7.6% of HbA1c, $n = 282$, $P = 0.002$). Similarly, Boj-Carceller *et al.*^[27] reported that HbA1c was significantly different in T2DM patients who received PPIs (6.7% \pm 1.0%, $n = 54$) compared with those who did not receive PPIs (7.3% \pm 1.4%, $n = 43$, $P = 0.018$). When these patients were assigned to two groups by the treatment of diabetes, those taking insulin and concurrent PPIs had better glycemic control, compared with those taking insulin but not PPI (-0.8% reduction, $P = 0.022$). In a very recent study, Barchetta *et al.*^[28], showed that the significantly different HbA1c and FPG levels were found in the T2DM patients with PPIs for longer than 2 years ($n = 245$) compared with those who did not take PPIs ($n = 303$) (7.1% \pm 1.07% with PPIs vs 7.4% \pm 1.4% without PPIs for HbA1c, $P = 0.011$; 127 \pm 36.9 mg/dL with PPIs vs 147.6 \pm 49.6 mg/dL without PPIs for FPG, $P < 0.001$, respectively). The increase of the differences was observed in patients treated with insulin and in those treated with combination of PPIs and GLP-1 based therapy^[28]. The results of these cross-sectional studies suggest the significant association between treatment with PPIs and the improved glycemic control in patients with T2DM.

On the other hand, in a study using a retrospective analysis, patients were assigned to 2 groups: 21 patients who had taken esomeprazole (a PPI) for 11.3 \pm 3 mo and 21 control subjects^[29]. Although there was a tendency for a decline in HbA1c in the patients treated with this PPI, it was not statistically significant (8.6% to 7.9%, $P = 0.054$), while in a subgroup with HbA1c $> 9\%$, the reduction was statistically significant (9.7% to 8.5%, $n = 11$, $P = 0.004$). No change in HbA1c was found in the entire control group and in a subgroup with HbA1c $> 9.0\%$ in control group (9.2% to 9.0%, $P = 0.455$; 10.3% to 10.0%, $P = 0.287$, respectively). Furthermore, Crouch *et al.*^[30] investigated 71 individuals with T2DM who were not taking insulin. The mean HbA1c was 7.11% during periods taking either prescription or over-the-counter PPIs, vs 7.7% during periods not taking PPIs (a significant difference, $P = 0.001$). Although there was no significant difference in mean HbA1c in a metformin monotherapy (6.81 treated with PPIs vs 7.10% treated without PPIs, $P = 0.25$), mean HbA1c was significantly lower in a concomitant therapy including metformin and/or sulfonylurea and/or glitazone (7.26 treated with PPIs vs 7.80 treated without PPIs, $n = 27$, $P = 0.002$). However, in another recent retrospective study of T2DM patients with relatively low levels of HbA1c, treatment with PPIs for ≥ 2 mo (mean duration: 180 d, $n = 43$) did not significantly change HbA1c levels (6.86% \pm 1.10% to 6.77% \pm 1.07%). Metformin monotherapy did not change HbA1c compared with a combination therapy including metformin and a therapy in antidiabetic agents not including metformin^[24]. Furthermore, 3 recent prospective randomized, double-blind, placebo-controlled studies using PPIs in small number of T2DM patients showed conflicting results with its effect on glycemic control. Singh *et al.*^[31] investigated the effect of a 12-wk pantoprazole (a PPI) therapy regimen on glycemic control in patients with T2DM^[31]. Thirty one eligible patients were randomly assigned to take either pantoprazole ($n = 16$) or placebo ($n = 15$). Pantoprazole (40 mg twice daily) significantly increased both plasma levels of gastrin (54.4 \pm 14.9 to 75.6 \pm 15.1 pg/mL, $P < 0.001$) and those of insulin (10.5 \pm 4.0 to 13.9 \pm 4.5 μ U/mL, $P < 0.001$) and improved the function of β cell as calculated by the homeostasis model assessment- β (HOMA- β). HbA1c significantly decreased with pantoprazole therapy (7.60% \pm 1.17% to 6.80% \pm 1.16%, $P < 0.001$). The decrease of HbA1c was positively associated with a significant elevation in both gastrin and insulin levels. González-Ortiz *et al.*^[32] investigated the effect of pantoprazole (40 mg once daily for 45 d) on secretion of insulin in 14 drug naive patients with T2DM. Significant increases in both the late insulin phase (215 \pm 127 to 308 \pm 151 pmol/L, $P = 0.028$) and total insulin secretion (174 \pm 94 to 265 \pm 135 pmol/L, $P = 0.028$), and significant decreases in HbA1c levels (7.5% to 6.6%, $P = 0.018$) were found with pantoprazole administration ($n =$

7), while there was no significant changes in these parameters in patients treated with placebo ($n = 7$). On the other hand, Hove *et al.*^[25] investigated the effect of esomeprazole on glycemic control in 41 T2DM patients using either dietary control or treatment with oral anti-diabetic agents. These patients were randomly assigned to take either add-on esomeprazole (40 mg daily, $n = 20$) or placebo ($n = 21$) during 12 wk^[25]. In the esomeprazole group, the area under the curve (AUC) for insulin did not change, while the AUC for the placebo group significantly decreased. Esomeprazole treatment caused a nine-fold elevation in the AUC for gastrin. Contrary to the expectation, HbA1c increased from $7.0\% \pm 0.6\%$ to $7.3\% \pm 0.8\%$ ($P < 0.05$) in the esomeprazole group and from $7.0\% \pm 0.6\%$ to $7.4\% \pm 0.8\%$ ($P < 0.05$) in the placebo group with no significant difference in change between both treatments (unadjusted, $P = 0.297$). These clinical findings from all of these studies are summarized in Table 1. Based on the published data to date, the degrees of the reduction of HbA1c by PPIs therapy in the studies with positive results appears to be approximately 0.6%-0.9%. This is somewhat milder or similar compared with those by recent available anti-diabetic drugs such as dipeptidyl peptidase-4 (DPP-4) inhibitors^[51] or sodium-glucose co-transporter 2 inhibitors^[52]. This suggests that the effect of PPI for glycemic control is probably moderate and that therefore PPI may have the potential for clinical benefit on glycemic control in patients with T2DM.

THE USE OF PPIs FOR THE TREATMENT OF TYPE 2 DIABETES: INTERPRETATION OF THE RESULTS AND POSSIBLE MECHANISMS OF GLYCEMIC CONTROL

As shown in the previous section, it appears that PPIs generally have a beneficial effect on glycemic control for T2DM patients with some studies showing no effect. The results of the different studies do not appear to be dependent on the type of PPI used. Based on the results of most clinical studies in which glycemic control was improved^[26-32], it appears that the actual basal levels of HbA1c may be important for the PPIs to show the apparent glucose-lowering effect because PPIs significantly decreased HbA1c level only when the basal HbA1c level was high in 1 retrospective study^[29]. In addition, the patients in most of the studies with negative results had a tendency to be under good glycemic control (approximate 7.0% of HbA1c)^[24,25], compared with those studies that showed positive results^[26-32]. In addition, treatment with PPIs and HbA1c levels were independent from possible confounders in a multivariate regression analysis in 1 study^[28], suggesting the importance of baseline HbA1c levels for the glucose lowering effect of PPIs. Next, if the possible effect of PPIs on glycemic control is based on

the mechanism of increase of β cell mass, treatment with PPIs for a longer period may be more effective in providing the full effect on glycemic control compared with that observed in most of the previous studies. However, in fact, the mechanism of the clinical effect of PPIs on glycemic control largely remains unclear. Because gastrin does not affect β cell neogenesis from the adult pancreatic ductal cells under a non-remodeling state as previously described^[14,15], it is not apparent whether the elevation of circulating gastrin levels induced by PPIs can really promote the increase of the mass of β cell in patients with T2DM, in whom pancreatic remodeling is not necessarily occurring. Nonetheless, elevated serum gastrin levels could affect the β cell mass in animal models of T2DM although the mechanism is not fully apparent. PPI mono therapy improved glycemic control with the increase in both plasma insulin and β cells mass in *Psammomys obesus*, an animal model of T2DM^[23]. In this study, a significant effect was obtained only when the PPI was used at a very high dose (lansoprazole 10-15 mg/kg); gastrin was elevated nine-fold at this dose. Since vonoprazan (a new generation PPI: potassium-competitive acid blocker) is more effective for inhibition of secretion of gastric acid and increases serum levels of gastrin (approximate six- to seven-fold with 10-40 mg of vonoprazan) compared with that of the existing PPIs^[53], it would be interesting to investigate in a future study whether this agent is also more effective on glycemic control. However, it is important to note that such elevation of serum gastrin levels by PPIs is not always needed to exhibit the clinically apparent glucose-lowering effect in T2DM patients because, in the study by Singh *et al.*^[31], in which positive results were obtained, the increase of gastrin by a PPI (pantoprazole) was only approximately 1.5-fold^[31], which was accompanied with an increase of insulin. These findings suggest the possibility that mechanisms other than the increase of β cell mass are also involved. One possible mechanism involves a gastrin-stimulated increase in insulin secretion by pancreatic β cells. It has been reported that because the secretion of the endogenous gastrin for the oral glucose tolerance test (OGTT) in healthy subjects is very small, it is unlikely that gastrin strongly promotes insulin secretion under this condition. However, an ordinary protein-rich meal (but not glucose-rich) largely increases both circulating gastrin and insulin levels^[2]. Therefore, gastrin appears to significantly stimulate secretion of insulin during and after a meal, this may partially explain the effect of PPIs on glycemic control. Another mechanism may involve the interaction of gastrin with other gastric hormones, such as ghrelin, which is reported to have an important role in energy homeostasis and appetite regulation. There is a report showing that ghrelin was down-regulated in primary gastric cells during gastrin-stimulation, and that ghrelin and gastrin levels had a significant negative correlation in humans. For example, a long-term 3-fold increase of

Table 1 Studies showing glucose-lowering effect of proton pump inhibitors in patients with type 2 diabetes

Mefford <i>et al</i> ^[26]	<p>Outcome measures: HbA1c levels in patients with type 2 diabetes taking PPIs ($n = 65$) vs those not taking PPIs ($n = 282$) was evaluated in cross-sectional design</p> <p>Key findings: There was a significant difference in HbA1c in patients taking PPIs vs those not taking PPIs (7.0% vs 7.6%, $P = 0.002$)</p> <p>Safety information: No information is described</p>
Boj-Carceller <i>et al</i> ^[27]	<p>Outcome measures: HbA1c levels in patients with type 2 diabetes taking PPIs ($n = 54$) vs those not taking PPIs ($n = 43$) was evaluated in cross-sectional design</p> <p>Key findings: HbA1c was significantly lower in type 2 diabetic patients who take PPIs compared with those not taking PPIs ($6.7\% \pm 1.0\%$ vs $7.3\% \pm 1.4\%$, $P = 0.018$)</p> <p>Safety information: No information is described</p>
Barchetta <i>et al</i> ^[28]	<p>Outcome measures: HbA1c and FPG levels in patients with type 2 diabetes taking PPIs for longer than 2 yr ($n = 245$) vs those not taking PPIs ($n = 303$) was evaluated in cross-sectional design</p> <p>Key findings: Patients with PPIs had significantly lower HbA1c ($7.1\% \pm 1.07\%$ vs $7.4\% \pm 1.4\%$, $P = 0.011$) and FPG (127 ± 36.9 mg/dL vs 147.6 ± 49.6 mg/dL, $P < 0.001$) levels than those who did not take PPIs</p> <p>Safety information: No information is described</p>
Hove <i>et al</i> ^[29]	<p>Outcome measures: HbA1c levels were retrospectively evaluated in patients with type 2 diabetes. Patients were assigned to 2 groups: 21 patients who had taken esomeprazole (a PPI) for 11.3 ± 3 mo and 21 control subjects</p> <p>Key findings: There was a tendency for a decline in HbA1c in the patients treated with this PPI (8.6% to 7.9%, $P = 0.054$). In a subgroup with HbA1c $> 9\%$ ($n = 11$), the reduction was statistically significant (9.7% to 8.5%, $P = 0.004$). No change in HbA1c was observed in the control group (9.2% to 9.9%, $P = 0.455$)</p> <p>Safety information: No information is described</p>
Han <i>et al</i> ^[24]	<p>Outcome measures: HbA1c was retrospectively evaluated in type 2 diabetic patients treated with PPIs for ≥ 2 mo (mean duration: 180 d, $n = 43$)</p> <p>Key findings: There was no significant change in HbA1c levels ($6.86\% \pm 1.10\%$ to $6.77\% \pm 1.07\%$; $P = 0.406$)</p> <p>Safety information: No information is described</p>
Crouch <i>et al</i> ^[30]	<p>Outcome measures: 71 individuals with type 2 diabetes who were not taking insulin was retrospectively investigated for the change of HbA1c</p> <p>Key findings: The mean HbA1c was 7.11% during periods with either prescription or over-the-counter PPIs, vs 7.7% during periods without PPIs (a significant difference; $P = 0.001$)</p> <p>Safety information: No information is described</p>
Singh <i>et al</i> ^[31]	<p>Outcome measures: The effect of a 12-wk pantoprazole (40 mg twice daily) therapy regimen on HbA1c, FPG, serum insulin, serum gastrin levels was prospectively measured in patients with type 2 diabetes in randomized double-blind, placebo-controlled study design. Thirty one eligible patients were randomly assigned to receive either pantoprazole ($n = 16$) or placebo ($n = 15$)</p> <p>Key findings: HbA1c and FPG significantly decreased with pantoprazole therapy ($7.60\% \pm 1.17\%$ to $6.80\% \pm 1.16\%$, $P < 0.001$ for HbA1c and 126.3 ± 10.3 to 109.2 ± 13.0 mg/dL, $P = 0.017$ for FPG), and the differences were significant between the two groups ($P = 0.004$ for HbA1c, $P = 0.019$ for FPG). Pantoprazole significantly increased both plasma gastrin ($P < 0.001$) and insulin levels ($P < 0.001$)</p> <p>Safety information: Nine patients reported adverse events as nausea, vomiting, headache and myalgia, which were similar and mild in the both groups. None of the patients had hypoglycemia</p>
González-Ortiz <i>et al</i> ^[32]	<p>Outcome measures: The effect of pantoprazole (40 mg once daily for 45 d) on insulin secretion in 14 drug naive patients with type 2 diabetes was prospectively investigated in a randomized, double-blind, placebo-controlled study design. Insulin secretion evaluated by hyperglycemic and hyperinsulinemic clamp technique, HbA1c, FPG and serum lipids were measured</p> <p>Key findings: Significant increases in total insulin secretion ($P = 0.028$), and significant decreases in HbA1c levels (7.5% to 6.6%; $P = 0.018$) but not FPG levels ($P = 0.236$) were found with pantoprazole therapy ($n = 7$), while there was no significant changes in these parameters in patients treated with placebo ($n = 7$). There were no significant changes in serum lipids in both groups</p> <p>Safety information: Two patients had mild headache (one in each group)</p>
Hove <i>et al</i> ^[25]	<p>Outcome measures: The effect of esomeprazole on glycemic control in 41 type 2 diabetic patients using either dietary control or therapy by anti-diabetic agents was prospectively examined in a randomized double-blind placebo-controlled 2×2 factorial study. These patients were randomly assigned to receive either add-on esomeprazole (40 mg daily, $n = 20$) or placebo ($n = 21$) for 12 wk. Insulin secretion, HbA1c levels and cardiovascular risk factors were evaluated</p> <p>Key findings: In the esomeprazole-treated group, the AUC (area under the curve) for insulin did not change ($P = 0.838$), while the AUC for the placebo group significantly decreased ($P = 0.002$). HbA1c increased from $7.0\% \pm 0.6\%$ to $7.3\% \pm 0.8\%$ ($P < 0.05$) in the esomeprazole-treated group and from $7.0\% \pm 0.6\%$ to $7.4\% \pm 0.8\%$ ($P < 0.05$) in the placebo group (no significant difference in change between both treatments; unadjusted, $P = 0.297$). The differences in cardiovascular risk factors were not significant between the two groups</p> <p>Safety information: Flatulence in 2 patients and diarrhea in 1 patient was reported in lansoprazole group, and flatulence in 2 patients and intermittent diarrhea in 1 patient was reported in placebo group</p>
Takebayashi <i>et al</i> ^[72]	<p>Outcome measures: The effect of alogliptin and lansoprazole ($n = 46$) combination therapy compared with alogliptin therapy without lansoprazole ($n = 43$) on glycemic control was investigated in a randomized open-label study. After 3 mo of treatment, the changes in HbA1c, FPG, serum gastrin were evaluated</p> <p>Key findings: A significant decrease in both HbA1c and FPG (respective $7.6\% \pm 0.6\%$ to $6.8\% \pm 0.7\%$, $P < 0.0001$, 52.0 ± 35.6 to 127.3 ± 27.4 mg/dL, $P < 0.0001$ in the combination therapy group, and respective $7.7\% \pm 0.5\%$ to $6.7\% \pm 0.5\%$, $P < 0.0001$, 153.6 ± 34.4 to 128.5 ± 26.6 mg/dL, $P = 0.0001$ in the alogliptin therapy group) was obtained. There were no significant differences in changes in HbA1c, FPG ($P = 0.2945$, $P = 0.1901$, respectively) and significant elevation in change in gastrin (approximate twofold, $P = 0.0004$) before and after therapy between the combination and the alogliptin mono therapy group</p> <p>Safety information: In alogliptin group, 1 patient discontinued the drug due to epi-gastric pain. In the combination group, 1 patient withdrew due to a mild cerebral infarction, and 1 patient noticed occasional hypoglycemic symptoms</p>

PPIs: Proton pump inhibitors.

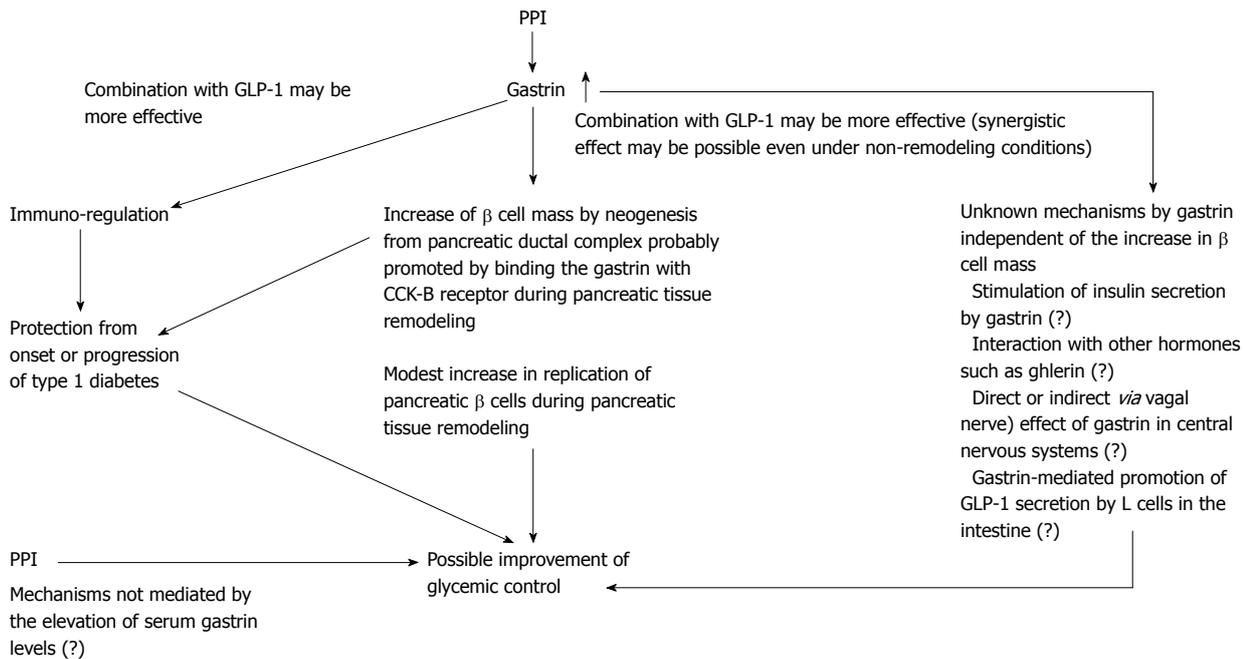


Figure 1 The possible mechanisms of proton pump inhibitors on the improvement of glycemic control. PPIs indirectly elevate serum gastrin levels. Gastrin promotes an increase in β cell mass by neogenesis of the β cells from the pancreatic ductal complex probably promoted by binding the gastrin with CCK-B receptor during pancreatic remodeling. In addition, a modest increase in the replication of pancreatic β cells during pancreatic remodeling is also reported although the mechanisms are not apparent because of the lack of a CCK-B receptor on β cells. Gastrin can enhance the effect of GLP-1 on β cell neogenesis from ductal cells. A synergistic effect may occur even under non-remodeling conditions in the pancreas. These mechanisms appear to contribute to the improvement of glycemic control in both type 1 and type 2 diabetes. Furthermore, a combination of GLP-1 and gastrin may protect from the onset or progression of type 1 diabetes by an immunoregulatory effect. Other possible gastrin-mediated mechanisms independent of the β cell mass increase may include stimulation of insulin secretion, interaction with other hormones such as ghrelin, direct or indirect (*via* vagal nerve) effects in the central nervous systems, and promotion of GLP-1 secretion by L cells in the intestine. Finally, it may be possible that PPIs affect glycemic control by unknown mechanisms independent of the elevation of serum gastrin levels. PPIs: Proton pump inhibitors; CCK-B: Cholecystokinin-B; GLP-1: Glucagon like-peptide-1.

gastrin in autoimmune gastritis significantly repressed ghrelin secretion^[54]. These findings suggest the possibility that the increase of gastrin levels is associated with less appetite and improvement of glycemic control *via* the decreased ghrelin levels although there is as yet no clinical evidence. Furthermore, it is known that the CCK-B receptor exists in the brain, especially in the hypothalamic area^[8,55]. Intracerebroventricular injection of gastrin decreases food intake, while inactivation of CCK-B receptor in mice changes the regulation of food-intake and body weight, and results in obesity^[56]. Despite the limitation of gastrin diffusion into the brain due to the blood brain barrier (BBB)^[57], there are reports suggesting that either peptide or peptide fragments might penetrate into the brain because of the lack of a BBB in the circumventricular organs^[58], and that intravenous gastrin administration activated neurons in several portions of brain^[59]. In addition, it is reported that gastrin in circulation is able to stimulate the area postrema neurons that express the CCK-B receptor and project to the nucleus of the solitary tract (NTS)^[60]. Mouse brain stem NTS-proopiomelanocortin neurons are associated with feeding-induced satiety^[61]. Therefore, we speculate that it might be possible that increased serum gastrin that is regulated by PPIs directly inhibits appetite *via* the central nervous system, although it may be possible that gastrin also

acts indirectly brain stem *via* the vagal nerve^[60]. In addition, a recent study revealed that gastrin stimulates GLP-1 secretion in L cells in the intestine^[62]. This can explain the possible effect of PPIs on glycemic control at least in part. Finally, it may also be important to consider whether PPIs potentially have a beneficial effect on glycemic control *via* unknown mechanism independent of gastrin. Taken together, the mechanisms of the possible PPI effects on glycemic control largely remain unclear, and multiple mechanisms appear to be involved. These possible mechanisms are described in Figure 1.

When treating patients, it is important to consider the potentially deleterious effects of PPIs on glycemic control, which may be more serious than the possible beneficial effect and which may modify the results. It is known that diabetes occasionally occurs with gastroesophageal reflux disease (GERD)^[63,64]. Because PPIs largely improve GERD clinical symptoms, it may be possible that the appetite of the patients with GERD is improved even if the elevation of gastrin levels by PPIs influences circulating ghrelin levels as previously described. These patients can thus potentially have worse glycemic control. In addition, it is reported that PPIs can induce dysbiosis^[65], which is connected with metabolic syndrome. Therefore, we speculate that PPIs can worsen glycemic control in this manner as well.

THE EFFECT OF COMBINATIONAL THERAPY OF PPIs (OR GASTRIN) WITH DPP-4 INHIBITORS (OR A GLP-1 RECEPTOR AGONIST) ON GLYCEMIC CONTROL IN TYPE 1 AND TYPE 2 DIABETES IN BOTH ANIMAL AND CLINICAL STUDIES

Recent evidence suggests the greater potential beneficial effect of a combination therapy of various hormones over that of a mono hormone therapy^[66]. As described in the previous section, gastrin enhances the effect of GLP-1 on β cell neogenesis, and this combination therapy more effectively improved hyperglycemia than mono therapy by each hormone in NOD mice^[48]. This result is also supported in the same animal model by combination therapy with DPP-4 inhibitors, which block degradation of GLP-1 by DPP-4 resulting in the elevation of serum active GLP-1 levels, and PPIs^[67]. Furthermore, Patel *et al*^[68], showed that combination therapy with exendin-4 (a GLP-1 receptor agonist) and omeprazole (a PPI) had better glycemic control compared with mono therapy with these drugs in *db/db* mice. Recently, Hao *et al*^[69] examined the effects of short periods of lansoprazole, sitagliptin (a DPP-4 inhibitor), and these concomitant therapy on glycemic control in mice with diet-induced obesity (DIO) and in healthy human subjects. In the DIO mice, lansoprazole therapy significantly improved glucose levels and increased both circulating insulin and C peptide levels than treatment in vehicles. Furthermore, concomitant treatment with lansoprazole and sitagliptin decreased glucose levels with higher levels in C-peptide and insulin compared to that with sitagliptin-treated mice. In a human study, the concomitant use (sitagliptin 100 mg daily and lansoprazole 30 mg daily) for 6 d resulted in significant decrease of glucose levels and increase of insulin levels in an OGTT vs the control, lansoprazole-, and sitagliptin-treated groups. Taken together, the results of these studies suggest the possibility that combination therapy with a GLP-1 receptor agonist (or DPP-4 inhibitors) and gastrin (or a PPI) may provide a more beneficial effect for glycemic control than each mono therapy. In addition, in *db/db* mice, a GLP-1-gastrin dual receptor agonist has showed a more continued regulatory effect of glucose with a significant increase in β -cell mass in pancreatic tissue than that of monotherapy in liraglutide (a GLP-1 receptor agonist)^[70]. However, the results of recent randomized, prospective studies evaluating the combination therapy with DPP-4 inhibitors and PPIs in patients with T1DM and T2DM were basically negative. Griffin *et al*^[71] reported the results of a randomized, placebo-controlled, multicenter, phase 2 trial (REPAIR-T1D) on the effect of concomitant use with sitagliptin and lansoprazole in patients with recent-onset T1DM. Patients aged 11-36 years, diagnosed with T1DM within

the past 6 mo, were recruited and were randomized (2:1) to take oral sitagliptin with lansoprazole or placebo for 12 mo. At 12 mo, the 2 h C peptide AUC was similar between the combination ($n = 40$) and placebo ($n = 18$) groups. HbA1c levels were mainly constant throughout the study period for both groups (no significant difference). HbA1c adjusted by insulin-dose was also similar (no significant difference) for both groups. Although these overall results were negative, this study is still ongoing with reassessments at both 18 and 24 mo. In T2DM, we investigated the effect of alogliptin (a DPP-4 inhibitor) and lansoprazole ($n = 46$) combination therapy compared with alogliptin therapy without a PPI ($n = 43$) on glycemic control in a randomized open-label study^[72] (Table 1). At 3 mo after the initiation of the therapy, the changes in HbA1c, FPG, HOMA- β , HOMA-insulin resistance (IR) and serum gastrin were evaluated. A significant decrease in both HbA1c ($7.6\% \pm 0.6\%$ to $6.8\% \pm 0.7\%$, $P < 0.001$ in the combination therapy group, and $7.7\% \pm 0.5\%$ to $6.7\% \pm 0.5\%$, $P < 0.001$ in the alogliptin therapy group) and FPG (152.0 ± 35.6 to 127.3 ± 27.4 mg/dL, $P < 0.001$ in the combination therapy group, and 153.6 ± 34.4 to 128.5 ± 26.6 mg/dL, $P = 0.001$ in the alogliptin therapy group), and a significant increase in HOMA- β were noted in both groups. However, significant differences were not obtained in the changes in HbA1c, FPG, and HOMA- β by therapy between the combination and the alogliptin mono therapy group ($P = 0.2945$, $P = 1901$, $P = 0.3042$, respectively). The levels of serum gastrin in the concomitant group was significantly elevated compared with those in the alogliptin mono therapy group ($P = 0.0004$). With the combination therapy, the serum gastrin levels increased approximately two-fold. Apart from the issue of the period of the administration, one of the possible reasons for these negative results may be due to the use of DPP-4 inhibitors rather than a GLP-1 receptor agonist with the PPI. The elevation of GLP-1 levels by DPP-4 inhibitors is relatively small compared with that observed with the GLP-1 receptor agonist. Therefore, despite the reports with the positive results on glycemic control using a combination of a PPI and DPP-4 inhibitors^[67,69], the effect may be small when compared to that observed with the combination of a PPI and a GLP-1 receptor agonist. The clinical data on the combination therapy of a PPI and a GLP-1 receptor agonist in patients with T1DM and T2DM are not available yet, but this therapy appears to be an attractive one, and future studies are warranted to confirm the effect of this combination therapy.

CONCLUSION

Although PPI therapy is attractive as a new approach for the therapy of diabetes (especially T2DM), the clinical effect on glycemic control of this drug is not yet fully established. The mechanisms of the clinical effect of PPIs on glycemic control are also not fully elucidated. A prospective, long term, randomized, double-blind,

placebo-controlled study on PPIs in a larger number of the T2DM patients is warranted to confirm the effect of PPIs on glyceamic control, especially in patients with relatively poor glyceamic control. The combination therapy of a PPI with a GLP-1 receptor agonist (rather than DPP-4 inhibitors) may improve glyceamic control in both T1DM and T2DM. A clinical study with a large number of patients is needed to establish the potential efficacy. At present, the clinicians' concerns are whether the patients can have better glyceamic control when PPIs are used for GERD or gastric ulcers in patients with T2DM, because the use of PPIs is not yet allowed for T2DM treatment in every country. If the treatment is for a long-term period, it is also important to consider the possible harmful effects of PPIs, including bone fracture^[73] and small intestine bacterial overgrowth^[74].

REFERENCES

- 1 **Gregory RA**, Tracy HJ. Isolation of two "big gastrins" from Zollinger-Ellison tumour tissue. *Lancet* 1972; **2**: 797-799 [PMID: 4116235]
- 2 **Rehfeld JF**. Incretin physiology beyond glucagon-like peptide 1 and glucose-dependent insulinotropic polypeptide: cholecystokinin and gastrin peptides. *Acta Physiol (Oxf)* 2011; **201**: 405-411 [PMID: 21392266 DOI: 10.1111/j.1748-1716.2010.02235.x]
- 3 **Schubert ML**, Makhlof GM. Gastrin secretion induced by distention is mediated by gastric cholinergic and vasoactive intestinal peptide neurons in rats. *Gastroenterology* 1993; **104**: 834-839 [PMID: 8095036]
- 4 **Schubert ML**, Makhlof GM. Neural, hormonal, and paracrine regulation of gastrin and acid secretion. *Yale J Biol Med* 1992; **65**: 553-560; discussion 621-623 [PMID: 1364124]
- 5 **Chueca E**, Lanas A, Piazuelo E. Role of gastrin-peptides in Barrett's and colorectal carcinogenesis. *World J Gastroenterol* 2012; **18**: 6560-6570 [PMID: 23236230 DOI: 10.3748/wjg.v18.i45.6560]
- 6 **Burkitt MD**, Varro A, Pritchard DM. Importance of gastrin in the pathogenesis and treatment of gastric tumors. *World J Gastroenterol* 2009; **15**: 1-16 [PMID: 19115463]
- 7 **Brand SJ**, Stone D. Reciprocal regulation of antral gastrin and somatostatin gene expression by omeprazole-induced achlorhydria. *J Clin Invest* 1988; **82**: 1059-1066 [PMID: 2901431]
- 8 **Dufresne M**, Seva C, Fourmy D. Cholecystokinin and gastrin receptors. *Physiol Rev* 2006; **86**: 805-847 [PMID: 16816139]
- 9 **Soll AH**, Amirian DA, Thomas LP, Reedy TJ, Elashoff JD. Gastrin receptors on isolated canine parietal cells. *J Clin Invest* 1984; **73**: 1434-1447 [PMID: 6325503]
- 10 **Bitziou E**, Patel BA. Simultaneous detection of gastric acid and histamine release to unravel the regulation of acid secretion from the guinea pig stomach. *Am J Physiol Gastrointest Liver Physiol* 2012; **303**: G396-G403 [PMID: 22595991 DOI: 10.1152/ajpgi.00548.2011]
- 11 **Waldum HL**, Hauso Ø, Fossmark R. The regulation of gastric acid secretion - clinical perspectives. *Acta Physiol (Oxf)* 2014; **210**: 239-256 [PMID: 24279703 DOI: 10.1111/apha.12208]
- 12 **Schmitz F**, Göke MN, Otte JM, Schrader H, Reimann B, Kruse ML, Siegel EG, Peters J, Herzig KH, Fölsch UR, Schmidt WE. Cellular expression of CCK-A and CCK-B/gastrin receptors in human gastric mucosa. *Regul Pept* 2001; **102**: 101-110 [PMID: 11730982]
- 13 **Hansen OH**, Pedersen T, Larsen JK, Rehfeld JF. Effect of gastrin on gastric mucosal cell proliferation in man. *Gut* 1976; **17**: 536-541 [PMID: 964686]
- 14 **Rooman I**, Lardon J, Bouwens L. Gastrin stimulates beta-cell neogenesis and increases islet mass from transdifferentiated but not from normal exocrine pancreas tissue. *Diabetes* 2002; **51**: 686-690 [PMID: 11872667]
- 15 **Téllez N**, Joanny G, Escoriza J, Vilaseca M, Montanya E. Gastrin treatment stimulates β -cell regeneration and improves glucose tolerance in 95% pancreatectomized rats. *Endocrinology* 2011; **152**: 2580-2588 [PMID: 21558313 DOI: 10.1210/en.2011-0066]
- 16 **Zhang JX**, Ji MY, Song J, Lei HB, Qiu S, Wang J, Ai MH, Wang J, Lv XG, Yang ZR, Dong WG. Proton pump inhibitor for non-erosive reflux disease: a meta-analysis. *World J Gastroenterol* 2013; **19**: 8408-8419 [PMID: 24363534 DOI: 10.3748/wjg.v19.i45.8408]
- 17 **Sheen E**, Triadafilopoulos G. Adverse effects of long-term proton pump inhibitor therapy. *Dig Dis Sci* 2011; **56**: 931-950 [PMID: 21365243 DOI: 10.1007/s10620-010-1560-3]
- 18 **Ligumsky M**, Lysy J, Siguencia G, Friedlander Y. Effect of long-term, continuous versus alternate-day omeprazole therapy on serum gastrin in patients treated for reflux esophagitis. *J Clin Gastroenterol* 2001; **33**: 32-35 [PMID: 11418787]
- 19 **Hu YM**, Mei Q, Xu XH, Hu XP, Hu NZ, Xu JM. Pharmacodynamic and kinetic effect of rabeprazole on serum gastrin level in relation to CYP2C19 polymorphism in Chinese Hans. *World J Gastroenterol* 2006; **12**: 4750-4753 [PMID: 16937451]
- 20 **Cadiot G**, Vissuzaine C, Pospai D, Ruzsniowski P, Potet F, Mignon M. [Effect of prolonged treatment with proton pump inhibitors on serum gastrin levels and the fundus mucosa. Preliminary results]. *Gastroenterol Clin Biol* 1995; **19**: 811-817 [PMID: 8566561]
- 21 **Koop H**, Klein M, Arnold R. Serum gastrin levels during long-term omeprazole treatment. *Aliment Pharmacol Ther* 1990; **4**: 131-138 [PMID: 2104080]
- 22 **Sanders SW**, Tolman KG, Greski PA, Jennings DE, Hoyos PA, Page JG. The effects of lansoprazole, a new H⁺,K⁽⁺⁾-ATPase inhibitor, on gastric pH and serum gastrin. *Aliment Pharmacol Ther* 1992; **6**: 359-372 [PMID: 1600052]
- 23 **Böðvarsdóttir TB**, Hove KD, Gotfredsen CF, Pridal L, Vaag A, Karlens AE, Petersen JS. Treatment with a proton pump inhibitor improves glycaemic control in Psammomys obesus, a model of type 2 diabetes. *Diabetologia* 2010; **53**: 2220-2223 [PMID: 20585936 DOI: 10.1007/s00125-010-1825-6]
- 24 **Han N**, Oh M, Park SM, Kim YJ, Lee EJ, Kim TK, Kim TN, Kwon MJ, Kim MK, Lee SH, Rhee BD, Park JH. The effect of proton pump inhibitors on glycated hemoglobin levels in patients with type 2 diabetes mellitus. *Can J Diabetes* 2015; **39**: 24-28 [PMID: 25305802 DOI: 10.1016/j.cjcd.2013.10.008]
- 25 **Hove KD**, Brøns C, Færch K, Lund SS, Petersen JS, Karlens AE, Rossing P, Rehfeld JF, Vaag A. Effects of 12 weeks' treatment with a proton pump inhibitor on insulin secretion, glucose metabolism and markers of cardiovascular risk in patients with type 2 diabetes: a randomised double-blind prospective placebo-controlled study. *Diabetologia* 2013; **56**: 22-30 [PMID: 23011351 DOI: 10.1007/s00125-012-2714-y]
- 26 **Mefford IN**, Wade EU. Proton pump inhibitors as a treatment method for type II diabetes. *Med Hypotheses* 2009; **73**: 29-32 [PMID: 19304401]
- 27 **Boj-Carceller D**, Bocos-Terraz P, Moreno-Vernis M, Sanz-Paris A, Trincado-Aznar P, Albero-Gamboa R. Are proton pump inhibitors a new antidiabetic drug? A cross sectional study. *World J Diabetes* 2011; **2**: 217-220 [PMID: 22174957 DOI: 10.4239/wjg.v2.i12.217]
- 28 **Barchetta I**, Guglielmi C, Bertocchini L, Calella D, Manfrini S, Secchi C, Pozzilli P, Cavallo MG. Therapy with proton pump inhibitors in patients with type 2 diabetes is independently associated with improved glycometabolic control. *Acta Diabetol* 2015; Epub ahead of print [PMID: 25716766]
- 29 **Hove KD**, Færch K, Böðvarsdóttir TB, Karlens AE, Petersen JS, Vaag A. Treatment with a proton pump inhibitor improves glycaemic control in type 2 diabetic patients - a retrospective analysis. *Diabetes Res Clin Pract* 2010; **90**: e72-e74 [PMID: 20888658 DOI: 10.1016/j.diabres.2010.09.007]
- 30 **Crouch MA**, Mefford IN, Wade EU. Proton pump inhibitor therapy associated with lower glycosylated hemoglobin levels in type 2 diabetes. *J Am Board Fam Med* 2012; **25**: 50-54 [PMID: 22218624 DOI: 10.3122/jabfm.2012.01.100161]
- 31 **Singh PK**, Hota D, Dutta P, Sachdeva N, Chakrabarti A, Srinivasan

- A, Singh I, Bhansali A. Pantoprazole improves glycemic control in type 2 diabetes: a randomized, double-blind, placebo-controlled trial. *J Clin Endocrinol Metab* 2012; **97**: E2105-E2108 [PMID: 22904177 DOI: 10.1210/jc.2012-1720]
- 32 **González-Ortiz M**, Martínez-Abundis E, Mercado-Sesma AR, Álvarez-Carrillo R. Effect of pantoprazole on insulin secretion in drug-naïve patients with type 2 diabetes. *Diabetes Res Clin Pract* 2015; **108**: e11-e13 [PMID: 25704601 DOI: 10.1016/j.diabres.2015.01.039]
- 33 **Larsson LI**, Rehfeld JF, Sundler F, Håkanson R. Pancreatic gastrin in foetal and neonatal rats. *Nature* 1976; **262**: 609-610 [PMID: 958427]
- 34 **Rooman I**, Lardon J, Flamez D, Schuit F, Bouwens L. Mitogenic effect of gastrin and expression of gastrin receptors in duct-like cells of rat pancreas. *Gastroenterology* 2001; **121**: 940-949 [PMID: 11606507]
- 35 **Brand SJ**, Fuller PJ. Differential gastrin gene expression in rat gastrointestinal tract and pancreas during neonatal development. *J Biol Chem* 1988; **263**: 5341-5347 [PMID: 3356689]
- 36 **Saïllan-Barreau C**, Dufresne M, Clerc P, Sanchez D, Corominola H, Moriscot C, Guy-Crotte O, Escricuet C, Vaysse N, Gomis R, Tarasova N, Fourmy D. Evidence for a functional role of the cholecystokinin-B/gastrin receptor in the human fetal and adult pancreas. *Diabetes* 1999; **48**: 2015-2021 [PMID: 10512367]
- 37 **Morisset J**, Julien S, Lainé J. Localization of cholecystokinin receptor subtypes in the endocrine pancreas. *J Histochem Cytochem* 2003; **51**: 1501-1513 [PMID: 14566022]
- 38 **Bouwens L**, Pipeleers DG. Extra-insular beta cells associated with ductules are frequent in adult human pancreas. *Diabetologia* 1998; **41**: 629-633 [PMID: 9662042]
- 39 **Ramiya VK**, Maraist M, Arfors KE, Schatz DA, Peck AB, Cornelius JG. Reversal of insulin-dependent diabetes using islets generated in vitro from pancreatic stem cells. *Nat Med* 2000; **6**: 278-282 [PMID: 10700229]
- 40 **Yatoh S**, Dodge R, Akashi T, Omer A, Sharma A, Weir GC, Bonner-Weir S. Differentiation of affinity-purified human pancreatic duct cells to beta-cells. *Diabetes* 2007; **56**: 1802-1809 [PMID: 17473224]
- 41 **Wang RN**, Klöppel G, Bouwens L. Duct- to islet-cell differentiation and islet growth in the pancreas of duct-ligated adult rats. *Diabetologia* 1995; **38**: 1405-1411 [PMID: 8786013]
- 42 **Bouwens L**. Transdifferentiation versus stem cell hypothesis for the regeneration of islet beta-cells in the pancreas. *Microsc Res Tech* 1998; **43**: 332-336 [PMID: 9849974]
- 43 **Iovanna JL**. Redifferentiation and apoptosis of pancreatic cells during acute pancreatitis. *Int J Pancreatol* 1996; **20**: 77-84 [PMID: 8968862]
- 44 **Bockman DE**, Merlino G. Cytological changes in the pancreas of transgenic mice overexpressing transforming growth factor alpha. *Gastroenterology* 1992; **103**: 1883-1892 [PMID: 1451981]
- 45 **Meier JJ**, Butler AE, Galasso R, Rizza RA, Butler PC. Increased islet beta cell replication adjacent to intrapancreatic gastrinomas in humans. *Diabetologia* 2006; **49**: 2689-2696 [PMID: 17016695]
- 46 **Wang TC**, Bonner-Weir S, Oates PS, Chulak M, Simon B, Merlino GT, Schmidt EV, Brand SJ. Pancreatic gastrin stimulates islet differentiation of transforming growth factor alpha-induced ductular precursor cells. *J Clin Invest* 1993; **92**: 1349-1356 [PMID: 8376589]
- 47 **Suarez-Pinzon WL**, Lakey JR, Brand SJ, Rabinovitch A. Combination therapy with epidermal growth factor and gastrin induces neogenesis of human islet {beta}-cells from pancreatic duct cells and an increase in functional {beta}-cell mass. *J Clin Endocrinol Metab* 2005; **90**: 3401-3409 [PMID: 15769977]
- 48 **Suarez-Pinzon WL**, Power RF, Yan Y, Wasserfall C, Atkinson M, Rabinovitch A. Combination therapy with glucagon-like peptide-1 and gastrin restores normoglycemia in diabetic NOD mice. *Diabetes* 2008; **57**: 3281-3288 [PMID: 18835930 DOI: 10.2337/db08-0688]
- 49 **Lavine JA**, Attie AD. Gastrointestinal hormones and the regulation of β -cell mass. *Ann N Y Acad Sci* 2010; **1212**: 41-58 [PMID: 21039588 DOI: 10.1111/j.1749-6632.2010.05802.x]
- 50 **Tamaki M**, Fujitani Y, Uchida T, Hirose T, Kawamori R, Watada H. Combination treatment of db/db mice with exendin-4 and gastrin preserves β -cell mass by stimulating β -cell growth and differentiation. *J Diabetes Investig* 2010; **1**: 172-183 [PMID: 24843429 DOI: 10.1111/j.2040-1124.2010.00044.x]
- 51 **Nauck MA**, Vilsbøll T, Gallwitz B, Garber A, Madsbad S. Incretin-based therapies: viewpoints on the way to consensus. *Diabetes Care* 2009; **32** Suppl 2: S223-S231 [PMID: 19875556 DOI: 10.2337/dc09-S315]
- 52 **Whalen K**, Miller S, Onge ES. The Role of Sodium-Glucose Co-Transporter 2 Inhibitors in the Treatment of Type 2 Diabetes. *Clin Ther* 2015; **37**: 1150-1166 [PMID: 25891804 DOI: 10.1016/j.clinthera.2015.03.004]
- 53 **Jenkins H**, Sakurai Y, Nishimura A, Okamoto H, Hibberd M, Jenkins R, Yoneyama T, Ashida K, Ogama Y, Warrington S. Randomised clinical trial: safety, tolerability, pharmacokinetics and pharmacodynamics of repeated doses of TAK-438 (vonoprazan), a novel potassium-competitive acid blocker, in healthy male subjects. *Aliment Pharmacol Ther* 2015; **41**: 636-648 [PMID: 25707624 DOI: 10.1111/apt.13121]
- 54 **Rau TT**, Sonst A, Rogler A, Burnat G, Neumann H, Oeckl K, Neuhuber W, Dimmler A, Faller G, Brzozowski T, Hartmann A, Konturek PC. Gastrin mediated down regulation of ghrelin and its pathophysiological role in atrophic gastritis. *J Physiol Pharmacol* 2013; **64**: 719-725 [PMID: 24388886]
- 55 **Mercer LD**, Le VQ, Nunan J, Jones NM, Beart PM. Direct visualization of cholecystokinin subtype2 receptors in rat central nervous system using anti-peptide antibodies. *Neurosci Lett* 2000; **293**: 167-170 [PMID: 11036187]
- 56 **Clerc P**, Coll Constans MG, Lulka H, Broussaud S, Guigné C, Leung-Theung-Long S, Perrin C, Knauf C, Carpené C, Pénicaud L, Seva C, Burcelin R, Valet P, Fourmy D, Dufresne M. Involvement of cholecystokinin 2 receptor in food intake regulation: hyperphagia and increased fat deposition in cholecystokinin 2 receptor-deficient mice. *Endocrinology* 2007; **148**: 1039-1049 [PMID: 17122076]
- 57 **Greenstein RJ**, Clain DJ, Straus E, Yalow RS. Distribution, molecular forms, and bioactivity of immunoreactive gastrin in a patient with metastatic gastrinoma. *Am J Gastroenterol* 1987; **82**: 886-889 [PMID: 3631036]
- 58 **Banks WA**, Audus KL, Davis TP. Permeability of the blood-brain barrier to peptides: an approach to the development of therapeutically useful analogs. *Peptides* 1992; **13**: 1289-1294 [PMID: 1494505]
- 59 **Yakabi K**, Iwabuchi H, Nakamura T, Endo K, Fukunaga Y, Kumaki I, Takayama K. Neuronal expression of Fos protein in the brain after intravenous injection of gastrin in rats. *Neurosci Lett* 2002; **317**: 57-60 [PMID: 11755239]
- 60 **Danzer M**, Jovic M, Samberger C, Painsipp E, Bock E, Pabst MA, Crailsheim K, Schicho R, Lippe IT, Holzer P. Stomach-brain communication by vagal afferents in response to luminal acid backdiffusion, gastrin, and gastric acid secretion. *Am J Physiol Gastrointest Liver Physiol* 2004; **286**: G403-G411 [PMID: 14592947]
- 61 **Fan W**, Ellacott KL, Halatchev IG, Takahashi K, Yu P, Cone RD. Cholecystokinin-mediated suppression of feeding involves the brainstem melanocortin system. *Nat Neurosci* 2004; **7**: 335-336 [PMID: 15034587]
- 62 **Cao Y**, Cao X, Liu XM. Expression of cholecystokinin2-receptor in rat and human L cells and the stimulation of glucagon-like peptide-1 secretion by gastrin treatment. *Acta Histochem* 2015; **117**: 205-210 [PMID: 25601282 DOI: 10.1016/j.acthis.2014.12.007]
- 63 **Feldman M**, Schiller LR. Disorders of gastrointestinal motility associated with diabetes mellitus. *Ann Intern Med* 1983; **98**: 378-384 [PMID: 6402969]
- 64 **Maser RE**, Pfeifer MA, Dorman JS, Kuller LH, Becker DJ, Orchard TJ. Diabetic autonomic neuropathy and cardiovascular risk. Pittsburgh Epidemiology of Diabetes Complications Study III. *Arch Intern Med* 1990; **150**: 1218-1222 [PMID: 2353855]
- 65 **Wallace JL**, Syer S, Denou E, de Palma G, Vong L, McKnight W, Jury J, Bolla M, Bercik P, Collins SM, Verdu E, Ongini E. Proton pump inhibitors exacerbate NSAID-induced small intestinal injury

- by inducing dysbiosis. *Gastroenterology* 2011; **141**: 1314-1322, 1314-1322, [PMID: 21745447 DOI: 10.1053/j.gastro.2011.06.075]
- 66 **Sadry SA**, Drucker DJ. Emerging combinatorial hormone therapies for the treatment of obesity and T2DM. *Nat Rev Endocrinol* 2013; **9**: 425-433 [PMID: 23478327 DOI: 10.1038/nrendo.2013.47]
- 67 **Suarez-Pinzon WL**, Cembrowski GS, Rabinovitch A. Combination therapy with a dipeptidyl peptidase-4 inhibitor and a proton pump inhibitor restores normoglycaemia in non-obese diabetic mice. *Diabetologia* 2009; **52**: 1680-1682 [PMID: 19455306 DOI: 10.1007/s00125-009-1390-z]
- 68 **Patel V**, Joharapurkar A, Gandhi T, Patel K, Dhanesha N, Kshirsagar S, Dhote V, Detroja J, Bahekar R, Jain M. Omeprazole improves the anti-obesity and antidiabetic effects of exendin-4 in db/db mice (-4 db/db)*. *J Diabetes* 2013; **5**: 163-171 [PMID: 22830490 DOI: 10.1111/j.1753-0407.2012.00227.x]
- 69 **Hao S**, Sun J, Tian X, Sun X, Zhang Z, Gao Y. Lansoprazole enhances the antidiabetic effect of sitagliptin in mice with diet-induced obesity and healthy human subjects. *J Pharm Pharmacol* 2014; **66**: 1133-1139 [PMID: 24628303 DOI: 10.1111/jphp.12237]
- 70 **Fosgerau K**, Jessen L, Lind Tolborg J, Østerlund T, Schæffler Larsen K, Rolsted K, Brorson M, Jelsing J, Skovlund Ryge Neerup T. The novel GLP-1-gastrin dual agonist, ZP3022, increases β -cell mass and prevents diabetes in db/db mice. *Diabetes Obes Metab* 2013; **15**: 62-71 [PMID: 22862961 DOI: 10.1111/j.1463-1326.2012.01676.x]
- 71 **Griffin KJ**, Thompson PA, Gottschalk M, Kylo JH, Rabinovitch A. Combination therapy with sitagliptin and lansoprazole in patients with recent-onset type 1 diabetes (REPAIR-T1D): 12-month results of a multicentre, randomised, placebo-controlled, phase 2 trial. *Lancet Diabetes Endocrinol* 2014; **2**: 710-718 [PMID: 24997559 DOI: 10.1016/S2213-8587(14)70115-9]
- 72 **Takebayashi K**, Sakurai S, Suzuki T, Hori K, Terasawa T, Naruse R, Hara K, Suetsugu M, Tsuchiya T, Aoki H, Hamasaki T, Shuutou H, Inukai T. Effect of combination therapy with alogliptin and lansoprazole on glycemic control in patients with type 2 diabetes. *Endocr J* 2014; **61**: 1031-1039 [PMID: 25185672]
- 73 **Leontiadis GI**, Moayyedi P. Proton pump inhibitors and risk of bone fractures. *Curr Treat Options Gastroenterol* 2014; **12**: 414-423 [PMID: 25209137 DOI: 10.1007/s11938-014-0030-y]
- 74 **Lo WK**, Chan WW. Proton pump inhibitor use and the risk of small intestinal bacterial overgrowth: a meta-analysis. *Clin Gastroenterol Hepatol* 2013; **11**: 483-490 [PMID: 23270866 DOI: 10.1016/j.cgh.2012.12.011]

P- Reviewer: Miller S, Neumiller JJ

S- Editor: Ji FF L- Editor: A E- Editor: Jiao XK



Dysglycemia after renal transplantation: Definition, pathogenesis, outcomes and implications for management

David Langsford, Karen Dwyer

David Langsford, Karen Dwyer, Department of Nephrology, St Vincent's Hospital Melbourne, Fitzroy 3065, Australia

Author contributions: Langsford D and Dwyer K contributed to this paper.

Conflict-of-interest statement: The authors declare that they have no competing interests.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Dr. Karen Dwyer, Department of Nephrology, St Vincent's Hospital Melbourne, 59 Victoria Parade, Fitzroy 3065, Australia. karen.dwyer@svhm.org.au
Telephone: +61-3-92883112
Fax: +61-3-92313151

Received: October 11, 2014

Peer-review started: October 11, 2014

First decision: October 28, 2014

Revised: July 6, 2015

Accepted: August 16, 2015

Article in press: August 17, 2015

Published online: August 25, 2015

Abstract

New-onset diabetes after transplantation (NODAT) is a major complication following renal transplantation. It commonly develops within 3-6 mo post-transplantation. The development of NODAT is associated with significant increase in risk of major cardiovascular events and cardiovascular death. Other dysglycemic states, such as impaired glucose tolerance are also associated

with increasing risk of cardiovascular events. The pathogenesis of these dysglycemic states is complex. Older recipient age is a consistent major risk factor and the impact of calcineurin inhibitors and glucocorticoids has been well described. Glucocorticoids likely cause insulin resistance and calcineurin inhibitors likely cause β -cell toxicity. The impact of transplantation in incretin hormones remains to be clarified. The oral glucose tolerance test remains the best diagnostic test but other tests may be validated as screening tests. Possibly, NODAT can be prevented by administering insulin early in patients identified as high risk for NODAT. Once NODAT has been diagnosed altering immunosuppression may be acceptable, but creates the difficulty of balancing immunological with metabolic risk. With regard to hypoglycemic use, metformin may be the best option. Further research is needed to better understand the pathogenesis, identify high risk patients and to improve management options given the significant increased risk of major cardiovascular events and death.

Key words: Management; Epidemiology; Pathogenesis; Renal transplantation; Diabetes

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: New-onset diabetes after transplantation (NODAT) carries a significant cardiovascular burden. Its pathogenesis is multifactorial and includes modifiable factors. New insights into glucose and insulin homeostasis may lead to improved ability to identify high risk patients and to the development of management strategies that do not require alteration in immunosuppression, whilst simultaneously reducing the risk of NODAT.

Langsford D, Dwyer K. Dysglycemia after renal transplantation: Definition, pathogenesis, outcomes and implications for manage-

ment. *World J Diabetes* 2015; 6(10): 1132-1151 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v6/i10/1132.htm> DOI: <http://dx.doi.org/10.4239/wjcd.v6.i10.1132>

INTRODUCTION

Dysglycemia post renal transplantation, encompassing new onset diabetes after transplant (NODAT), impaired fasting glucose (IGF) and impaired glucose tolerance (IGT), is a challenging clinical problem. However, despite more than two decades of research the pathogenesis of post-transplant dysglycemia is incompletely understood and a consensus on approach to screening, diagnosis and management is lacking. This review will outline the issues of defining the clinically important states, detecting and predicting their development, the progress that has been made in understanding their pathogenesis and relationship to described risk factors (particularly immunosuppression therapies) and the implications for management and further research into this significant post-transplant complication.

DEFINITION

There have been several changes in the definition of dysglycemia post transplantation over time. Initially referred to as diabetes after renal transplantation, this name failed to capture the important distinction of those who were diabetic pre-transplant from those who developed diabetes after transplant. The term post-transplant diabetes mellitus (PTDM) also failed to clearly distinguish between the two states. The most common term currently used is new-onset diabetes after transplant (NODAT); however, this too fails to capture those with new onset IGT, which is also associated with poorer outcomes (see below). Some have proposed the term "transplant associated hyperglycemia"^[1], which captures the impact of dysglycemia, as opposed to the worst category of dysglycemia alone (diabetes), however it does not make a distinction between those who came to transplant with a dysglycemic state and those who developed it after transplantation.

Prior to 2003 the most common criteria used for the diagnosis of post-transplant diabetes was use of hypoglycemic agents. However, this is reliant upon clinician awareness of the results of appropriately timed and collected glucose testing and remains an insensitive marker of NODAT. With enhanced understanding of the pathophysiology of post-transplant dysglycemia and its clinical significance a more sensitive and clinically useful definition is needed. In 2003 an international expert panel devised a consensus document^[2] that adopted the World Health Organisation/American Diabetes Association (WHO/ADA) guidelines for the testing and defining of dysglycemic states post-transplant [fasting blood glucose level (F BGL) ≥ 7.0 mol/L; 2-h BGL ≥ 11.1 mmol/L], based on the definitions used for the

general population. However, whilst there is consensus on the interpretation of blood glucose levels, there is no consensus on who to test, when to test and which test to use. Table 1 shows the wide range of tests used and timing of these in studies that have reported NODAT outcomes: F BGL, random blood glucose level (R BGL), 2-h 75 g oral glucose tolerance test (oGTT), HbA1c at 10 wk, 3 mo, 6 mo, 1 year and use of hypoglycemic agents at 30 d. Furthermore, there is little recognition in the literature of the importance of reporting and understanding the significance of dysglycemic states other than NODAT such as IGT or IFG. Few studies report incident rates and/or outcomes of such dysglycemic states. As a result, drawing conclusions based on research in this area has unavoidable caveats, which can only be addressed by large multi-centred well designed trials with post-transplant dysglycemia as the primary outcome.

EPIDEMIOLOGY

One of the confounders in any study of NODAT is the rate of pre-transplant unrecognised dysglycemia. Table 2 shows the rates of unrecognised dysglycemia in patients on the transplant waiting list. Bergrem *et al.*^[30] investigated 889 Norwegian transplant wait listed candidates who were not clinically suspected to have diabetes. The majority of patients (62%) were not on dialysis and only 12% were on glucocorticoids. All patients underwent an oGTT. Using WHO/ADA diagnostic criteria, 330 (37.1%) patients were found to have dysglycemia, in addition to which, 72 (8.1%) were found to have diabetes. Importantly, of those patients found to be diabetic on oGTT, only 22% were identified by F BGL testing alone. Further receiver operating curve (ROC) analysis demonstrated that using a cut-off of 92 mg/dL (5.1 mmol/L) for F BGL testing as the threshold for initiating an oGTT detected 90% of the diabetic patients, requiring 53% of the wait listed patients to be tested.

It is interesting to note that not all patients with dysglycemia pre-transplant develop persistent post-transplant dysglycemia (IGF, IGT or NODAT). Caillard *et al.*^[31] screened 243 patients at time of wait listing with oGTT and found 37 (15.2%) dysglycemic patients and eight (3.3%) newly diagnosed diabetic patients. The time from pre-transplant oGTT to transplantation was not documented; however, 50% of the dysglycemic patients developed NODAT, 23% remained dysglycemic and 14% become normoglycemic post transplantation. In 26% of those diagnosed with NODAT, this abnormality could only be detected by oGTT. A Japanese study in which patients with no known history of diabetes were administered an oGTT two weeks before receipt of a living donor transplant, found that 30.4% were dysglycemic with an additional 4.0% found to be diabetic^[32]. Hornum *et al.*^[33] found 33% dysglycemia rate pre-transplant ($n = 57$) and over 12-mo follow up the pre-transplant dysglycemia was not associated with the development of NODAT. Interestingly, they too

Table 1 Selection of studies that reported rates of new-onset diabetes after transplantation or other dysglycemic states

Ref.	Criteria	n	Rates
Cosio <i>et al</i> ^[5]	Use of medications, F BGL	490	13% at 1 yr 33% dysglycemic
Hjelmsaeth <i>et al</i> ^[4]	Use of medications, F BGL, oGTT	201	20% at 3 mo
Vincenti <i>et al</i> ^[5]	oGTT	682	30% at 6 mo dysglycemic
Delgado <i>et al</i> ^[6]	oGTT, F BGL	374	6.7% at 4.1 yr 25.1% dysglycemic
Ramesh Prasad <i>et al</i> ^[7]	F BGL or R BGL	151	20.5%
Luan <i>et al</i> ^[8]	oGTT	203	11.8% at 10 wk 47.8% dysglycemic
Bayer <i>et al</i> ^[9]	Use of medications, F BGL, R BGL	640	31.4% at 1 yr
Bergrem <i>et al</i> ^[10]	Use of medications, F BGL, R BGL	301	13% at 10 wk
Valderhaug <i>et al</i> ^[11]	oGTT	1410	17% at 10 wk 38% dysglycemic
Ciancio <i>et al</i> ^[12]	Use of medications	150	15%-22% at 4 yr
Israni <i>et al</i> ^[13]	Medications, F BGL	1840	13% at 5 yr
Wauters <i>et al</i> ^[14]	Use of medications, F BGL	1146	14.1% at 1 mo, 11.1% at 4 mo, 13.4% at 1 yr 27%, 34.3% and 29.8% dysglycemic
Chan <i>et al</i> ^[15]	oGTT	292	24% at 6 mo
Vacher-Coponat <i>et al</i> ^[16]	Use of medications	289	16.8%-18.8% at 3 yr
Tillman <i>et al</i> ^[17]	oGTT	200	5% at 39 mo 30.5% dysglycemic
Bonet <i>et al</i> ^[18]	F BGL, R BGL, oGTT	138	13% at 6 mo
Cole <i>et al</i> ^[19]	Use of medications, F BGL, oGTT	49	4% at 6 mo
Nagaraja <i>et al</i> ^[20]	Use of medications, F BGL	118	21% at 3 mo, 37% at 1 yr
First <i>et al</i> ^[21]	Use of medications, F BGL, HbA1c	634	17.8%-36.5% at 1 yr
Nagaraja <i>et al</i> ^[22]	oGTT	76	13% at 5 yr, 24% at 11 yr 42% and 61% dysglycemic
Tokodai <i>et al</i> ^[23]	Use of medications, F BGL, R BGL	145	11.7% at 1 yr
Viecelli <i>et al</i> ^[24]	oGTT	83	17% at 3 mo, 15% at 15 mo 31% and 21% dysglycemic
Weng <i>et al</i> ^[25]	Use of medications, F BGL, R BGL	166	29.5%
Schweer <i>et al</i> ^[26]	R BGL, HbA1c	526	16.7%
Prasad <i>et al</i> ^[27]	oGTT	439	20% at 3 mo 33% dysglycemic
Silva <i>et al</i> ^[28]	HbA1c	638	21.3%-41.1% at 4 yr
Lv <i>et al</i> ^[29]	F BGL	428	20.3% at 5.7 yr

Definitions diabetes: F BGL ≥ 7.0 mmol/L (126 mg/dL) or ≥ 11.1 mmol/L (200 mg/dL) on oGTT or R BGL ≥ 11.1 mmol/L (200 mg/dL) plus symptoms. Other dysglycemic states. IFG: ADA criteria 5.6-6.9 mmol/L (100-125 mg/dL); WHO criteria 6.1-6.9 mmol/dL (100-125 mg/dL); IGT: oGTT 7.8-11.0 mmol/L (140-199 mg/dL). F BGL: Fasting blood glucose level; R BGL: Random blood glucose level; oGTT: 2-h oral glucose tolerance test.

documented a small group of pre-transplant diabetic patients in whom the diabetic state remitted post-transplant.

The case finding described by table two highlights key differences in glucose homeostasis between end stage kidney disease (ESKD) uremic patients and the general population. Approximately 70% of general population patients can be diagnosed as diabetic *via* a F BGL^[34], as compared to 22% in the Norwegian transplant wait listed cohort. Moreover, the incidence of new diagnosis of diabetes in wait listed patients on dialysis is approximately 5%-6% per year^[33,35] (when using oGTT diagnostic criteria), compared with approximately 0.7%-1.3% per year in the general population^[36]. These figures ought to give the reader cause to be cautious with regard to the interpretation of rates of post transplantation dysglycemia and diabetes. This is particularly the case when reviewing retrospective data, in which often only a pre-transplant F BGL is available and the time from glucose testing to transplantation may extend for many months. It

may be that the denominator in the quoted rates of NODAT includes patients who were not normoglycemic at time of transplantation. This assessment is further complicated by the possibility that dysglycemia pre-transplant may not be a sufficient factor for dysglycemia post-transplant state (see below).

Further complicating the interpretation of incident rates of dysglycemia post-transplant is the spontaneous remission and normalisation of blood glucose levels observed in some patients. For example, early dysglycemia, such as in the period of hospitalisation post-transplant, is common and occurs in 75%-90% of patients within the first week^[37-39]. Luan *et al*^[8] in a prospective study of 203 non-diabetic patients showed the mean day 3 F BGL to be 124-134 mg/dL (6.9-7.4 mmol/L). Such dysglycemia should not be dismissed as due entirely to peri-operative factors, as some data suggests that day 7 F BGL may be predictive of NODAT at 1 year^[40]. A recent clinical study measured continuous capillary blood glucose levels for the first 4 d post-transplant in 43 patients. There was a considerable

Table 2 The rates of unrecognized dysglycemia in patients on the transplant waiting list

Ref.	Unrecognised on waiting list - diabetes	Unrecognised on waiting list - dysglycemia
Ramesh Prasad <i>et al</i> ^[7]	-	15%
Hornum <i>et al</i> ^[33]	-	33%
Bergrem <i>et al</i> ^[30]	8.1%	45.2%
Iida <i>et al</i> ^[32]	4%	30.4%
Caillard <i>et al</i> ^[31]	3.3%	15.2%
Bonet <i>et al</i> ^[18]	< 0.1%	8.9%

burden on hyperglycemia with 43% having blood glucose above 7.7 mmol/L for more than 12 h per day. The incidence of NODAT at 72 mo was 18.6% and the authors suggested that the day 1 capillary blood glucose may identify those at risk^[41]. Moreover, one study found that only 4% of patients normoglycemic early post-transplant later developed NODAT^[42] and a normal oGTT within the first week has been shown to have a NPV of 97.6% for later NODAT development^[43]. However, it is important to note that not all patients with early hyperglycemia develop permanent dysglycemic states, as there is a considerable degree of transience and variation in dysglycemic states^[33]. For example, a Chinese study, employing F BGL for NODAT found an incident rate of 20.32% after a mean follow up of 5.65 years in patients who survived more than one year post transplantation. Of these, 65.5% developed NODAT within 1 year and 17.2% had transient NODAT^[29]. Furthermore, such transience likely occurs within the first 3-6 mo. In an international trial comparing standard and reduced dose tacrolimus (Tac) the cumulative incidence at 6 mo of NODAT was 30.3%; however, the incidence in each group was lower at 6 mo compared to 3 mo (23.9% vs 28.4% and 13.2% vs 15.2%)^[15].

Notwithstanding the notable degree of transient dysglycemia, persistent NODAT often develops within 3 to 6 mo following renal transplantation. A mean time to diagnosis of 4.3 mo has been reported^[44]. This may help to determine the optimal time of testing. Using oGTT testing at 10 wk post transplantation, Valderhaug *et al*^[11,45] reported an incidence of NODAT of 14%-17%. Most studies find that NODAT develops early and this is confirmed by analyses of large data sets. For instance, an analysis of the organ procurement and transplantation network (OPTN) registry data has found a cumulative incidence of NODAT of only 16.2% at 3 years (registry data is limited by the nature of reporting of outcomes), the majority had developed within the first year post transplantation^[46]. Similar results have been reported in a United States cohort of 640 patients with a mean F BGL of less than 100 mg/dL (5.6 mmol/L) at time of transplantation. NODAT occurred in 31.4% of patients over 1 year, the majority of which had occurred within the first 6 mo (26.4% of total population by 6 mo). By 5 years post transplantation, 46.3% of previously believed to be non-diabetic patients had a diagnosis of NODAT^[9].

With regard to any dysglycemia (IGT/IFG or NODAT), a moderate sized ($n = 203$) prospective study of the risk of developing dysglycemia post transplantation, documented a rate of 47.8% when tested at 10 wk with an oGTT and applying WHO/ADA diagnostic criteria^[8]. Retrospective data has found rates of 39.7% who remained normoglycemic throughout the first year post-transplant^[47]. A study specifically designed to determine the rates of pre-diabetic dysglycemia found 30.5% of patients met accepted criteria using an oGTT at a median of 39 mo post-transplant^[17]. Similarly, in a large international study designed to determine the differences in diabetogenesis of cyclosporin (CsA) and Tac, at 6 mo post-transplant only 300 out of 587 patients (51.1%) remained normoglycemic^[5]; however, the criteria for definition of NODAT was need for medications at greater than 30 d. A cross sectional study of multiple Spanish centres found a rate of dysglycemia of 31.8% at almost 4 years post-transplant, the majority detected by oGTT^[6]. It is interesting to note that 58.8% of the dysglycemic patients had a simultaneous normal F BGL.

The above discussions reveal notable limitations when quoting rates of post-transplant dysglycemic states or NODAT alone. Whilst there is consensus with regard to blood glucose cut-off values, it is unclear which test should be employed and at which time post-transplant. Furthermore, the witnessed remission of some pre-transplant dysglycemia to normoglycemia post-transplant^[19,37] (although this has not been commonly documented), further complicates analyses of rates of new-onset post-transplant dysglycemia.

RISK FACTORS

Multiple risk factors have been associated with the development of NODAT (Table 3) many of which are not modifiable. The most consistently found risk factor is advancing age appreciated since the recognition of NODAT in the early period of use of CsA^[79]. Increasing age has been found to be a risk factor in small and large retrospectively analysed and prospectively collected data sets, including registry datasets in which the prevalence of NODAT may have been underestimated^[8,13,17,26,46,49,52-54]. Male gender, family history of diabetes and APCKD are documented as risk factors, but not consistently^[46,49,54-57,61,62]. With regard to genetic risk multiple polymorphisms, including mitochondrial, have been described as contributing risk to the development of NODAT^[53,54,63-67]. A closer analysis of genetic polymorphisms and their associated risk is beyond the scope of this review.

Transplant related factors: Calcineurin inhibitors

Potentially modifiable risk factors can be divided into transplant specific and generic. Of the generic, increasing body mass index (BMI) is associated with increased incidence of NODAT when categorised into intervals of 5 with < 20 as a reference, with increased

Table 3 Modifiable and non-modifiable risk factors associated with new-onset diabetes after transplantation or dysglycemic state

Variable	Ref.	Comment
ATG-divided dose	Stevens <i>et al</i> ^[48]	Increased dysglycemia compared to single dose in patients treated with Tac and sirolimus
African American	Kasiske <i>et al</i> ^[49] Shah <i>et al</i> ^[50] Johnston <i>et al</i> ^[51]	OR = 1.68 RR = 1.38 HR = 1.56
Age	Bayer <i>et al</i> ^[9] Kasiske <i>et al</i> ^[49] Cole <i>et al</i> ^[52] Ghisdal <i>et al</i> ^[53] Luan <i>et al</i> ^[8] Luan <i>et al</i> ^[46] Israni <i>et al</i> ^[13] Tillmann <i>et al</i> ^[17] McCaughan <i>et al</i> ^[54] Schweer <i>et al</i> ^[26]	HR = 1.35 Strong independent risk factor RR: 1.9-2.6 27707 registry patients OR: 1.33 If > 60 yr OR 1.03 of NODAT for each 6 mo of age Increasing age associated with dysglycemia and new onset metabolic syndrome Analysis of 25837 registry patients, increase in NODAT in each categorised group compared to reference 18-34 years old HR: 1.33 of NODAT at 60 mo Increase in dysglycemia at mean of 56 M post-transplant; RR of 1.28 for each 5 yr OR 1.4 per decade in 427 Northern Irish patients NODAT 56.1 yr <i>vs</i> 47.9 yr; <i>P</i> < 0.01
APCKD	de Mattos <i>et al</i> ^[55] Hamer <i>et al</i> ^[56] Johnston <i>et al</i> ^[51] Luan <i>et al</i> ^[46] Ruderman <i>et al</i> ^[57]	Increased 1 yr incidence in a matched cohort Multivariate analysis OR 2.4 No increase found in 21564 USRDS patients Multivariate analysis OR: 1.17 No increased risk found
Basiliximab	Aasebø <i>et al</i> ^[58] Prasad <i>et al</i> ^[27]	Basiliximab (<i>n</i> = 134) <i>vs</i> no induction historical control; increased dysglycemic state <i>P</i> = 0.017 In living recipients who elected to receive basiliximab OR 2.34 for NODAT at 3 mo
BMI	Kasiske <i>et al</i> ^[49] Cole <i>et al</i> ^[52] Luan <i>et al</i> ^[46]	Increased BMI, NODAT RR: 1.7 Multivariate analysis OR 1.76 for NODAT Analysis of 25837 registry patients. increase in NODAT in each categorised group of BMI compared to reference < 20
CMV	Israni <i>et al</i> ^[13] Hjelmsaeth <i>et al</i> ^[59]	BMI ≥ 30, HR 1.69 for NODAT at 60 mo Asymptomatic infection OR: 4.0 for NODAT at 10 wk
CNI - Higher levels	Chan <i>et al</i> ^[15] Cole <i>et al</i> ^[19] Suszynski <i>et al</i> ^[60]	NODAT 17% <i>vs</i> 31%, low dose <i>vs</i> standard dose Tac Single arm study of 49 patients with a 4% 6 mo incidence of NODAT. Early glucocorticoid reduction and low dose CsA Higher Tac levels (plus sirolimus) compared to lower Tac (plus sirolimus) or CsA/MMF higher rates of NODAT with 10 yr FU
CNI - Tac <i>vs</i> CsA	Vincenti <i>et al</i> ^[5] Cole <i>et al</i> ^[52] Luan <i>et al</i> ^[46] Vacher-Coponat <i>et al</i> ^[16] Cotovio <i>et al</i> ^[44]	RCT. Dysglycemia at 6 mo higher in Tac/MMF <i>vs</i> CsA/MMF: <i>P</i> = 0.05 27707 registry patients OR 1.51 for NODAT Analysis of 25837 registry patients. Increase in NODAT OR: 1.24 No difference in CsA/Aza <i>vs</i> Tac/MMF in RCT (<i>n</i> = 289) Retrospective multivariate analysis higher Tac not CsA levels associated with NODAT
Family history of diabetes	Bora <i>et al</i> ^[61] Santos <i>et al</i> ^[62]	Recipients from living related donors Retrospective (<i>n</i> = 303). RR: 3.6 for NODAT
Gender	Kasiske <i>et al</i> ^[49] McCaughan <i>et al</i> ^[54]	Greater risk in males in registry patients OR 2.2 for male gender in 427 Northern Irish patients
Genetic polymorphisms	Ghisdal <i>et al</i> ^[53] Ghisdal <i>et al</i> ^[63] Kurzawski <i>et al</i> ^[64] Yao <i>et al</i> ^[65] McCaughan <i>et al</i> ^[54] Nicoletto <i>et al</i> ^[66] Tavira <i>et al</i> ^[67]	rs7903146 polymorphism of TCF7L2 OR 1.6 of NODAT at 6 mol/L, but not associated with IGT Summarises known associations Polish Caucasian patients. Increasing SNPs associated with increased risk, OR = 1.37 Fok1 vitamin D polymorphism associated with NODAT OR 11.8 <i>P</i> = 0.012 7 SNPs involved with β-cell apoptosis associated with NODAT Adiponectin gene polymorphism associated with NODAT
Glucocorticoids	Boots <i>et al</i> ^[68] Ghisdal <i>et al</i> ^[53] Luan <i>et al</i> ^[46] Rizzari <i>et al</i> ^[69] Cole <i>et al</i> ^[19]	Mitochondrial haplogroup H associated with NODAT in Tac treated patients Early glucocorticoid withdrawal associated with reduced NODAT incidence in the first year OR 2.78 of NODAT at 6 mol/L if AR treated with glucocorticoids Analysis of 25837 registry patients. OR 1.42 for NODAT if discharged on maintenance. Glucocorticoid only induction associated with increase in NODAT OR: 1.31 Significant reduction in NODAT compared with historical control when glucocorticoids rapidly tapered Single arm study of 49 patients with a 4% 6 mo incidence of NODAT. Early glucocorticoid reduction and low dose CsA
HCV +	Schweer <i>et al</i> ^[26] Kasiske <i>et al</i> ^[49] Cole <i>et al</i> ^[52] Johnston <i>et al</i> ^[51] Baid-Agrawal <i>et al</i> ^[70] Luan <i>et al</i> ^[46] Lv <i>et al</i> ^[29] Prasad <i>et al</i> ^[27]	Pulse glucocorticoid for BPAR associated with increasing NODAT incidence HCV+, NODAT RR: 1.3 27707 registry patients OR for NODAT 1.82 21564 USRDS registry patients, HR: 1.7 for NODAT 14 HCV+ 24 HCV- patients. HCV+ increased insulin resistance; <i>P</i> = 0.008 Analysis of 25837 registry patients. Increase in NODAT OR: 1.43 Cohort of 428 Chinese patients. NODAT associated with HCV at mean 5.6 yr follow up, OR = 2.72 439 Indian patients, OR = 6.37

Hyper-parathyroidism post transplant	Ivarsson <i>et al</i> ^[71]	PTH > 13.8 pmol/L associated with NODAT at 1 yr, OR = 4.25
Impaired glycaemic state pre-transplant	Ramesh Prasad <i>et al</i> ^[7]	Higher within the normal range random BSL associated with NODAT
	Bora <i>et al</i> ^[61]	IGT at time of transplant associated with NODAT
	Hornum <i>et al</i> ^[33]	IGT NOT predictive of NODAT
	Cotovio <i>et al</i> ^[44]	Higher fasting BGL associated with NODAT
Magnesium post-transplant	Garg <i>et al</i> ^[72]	1 mol/L lower Mg associated with dysglycemia; no association with 1M CNI trough level
Magnesium pre-transplant	Augusto <i>et al</i> ^[73]	Lower magnesium immediately pre-transplant associated with NODAT; $P < 0.02$
Metabolic syndrome post-transplant	Israni <i>et al</i> ^[13]	MS in first 6-12 mo associated with NODAT by 60 mo, HR = 3.46
	Luan <i>et al</i> ^[8]	10 W dysglycemia associated with MS
	Nagaraja <i>et al</i> ^[22]	Development of MS predicts progressive dysglycemia
Metabolic syndrome pre-transplant	Bayer <i>et al</i> ^[9]	HR: 1.34 for NODAT at 1 yr
Sirolimus	Teutonico <i>et al</i> ^[74]	No improvement when changing from CNI to sirolimus
	Ekberg <i>et al</i> ^[75]	Low dose sirolimus may confer less risk than low dose Tac
	Johnston <i>et al</i> ^[51]	20124 registry patients. Compared to CsA + MMF/AZA: Sirolimus + CsA HR 1.61; Sirolimus + Tac HR 1.66; Sirolimus + MMF/AZA HR 1.36
	Guerra <i>et al</i> ^[76]	RCT ($n = 150$) Tac/sirolimus vs Tac/MMF vs CsA/sirolimus. No difference in NODAT
	Gyurus <i>et al</i> ^[77]	Retrospective ($n = 514$). Sirolimus HR 3.5 for NODAT over 10 yr
	Veroux <i>et al</i> ^[78]	21 NODAT converted to sirolimus, 80% remission of NODAT on basis of F BGL
	Suszynski <i>et al</i> ^[60]	Increased risk with high dose Tac/low dose sirolimus combination

F BGL: Fasting blood glucose level; oGTT: 2-h oral glucose tolerance test; NODAT: New-onset diabetes after transplantation; ATG: Antithymocyte globulin; USRDS: United States Renal Data System; BMI: Body mass index; CMV: Cytomegalovirus; CNI: Calcineurin inhibitors; Tac: Tacrolimus; MMF: Mycophenolate mofetil.

risk in the higher categories of BMI^[47]. The most significant transplant specific modifiable risk factors are immunosuppressive medications specifically the use of calcineurin inhibitors (CNI - Tac and CsA) and glucocorticoids. The diabetogenic impact of CsA has been described since the early 1980s^[79-82]. The introduction of Tac into clinical practice was associated with less acute rejection and improved graft function but at the expense of a greater incidence of NODAT^[83]. The diabetes incidence after renal transplantation trial was first large randomised study ($n = 682$; not diabetic at baseline $n = 567$) designed primarily to investigate the increase risk posed by Tac use instead of CsA. The primary endpoint was a 6-mo composite endpoint of dysglycemia (NODAT or IFG) based on oGTT administered at 90 and 180 d. They found 6-mo cumulative incidence of 33.6% in Tac treated patients and 26% in CsA treated patients ($P = 0.046$). Furthermore, more patients required hypoglycemic treatment in the Tac treated group ($P = 0.005$) and more patients in the CsA treated group who were not treated with hypoglycemic agents had an improvement in their glycaemic state by 6 mo ($P = 0.067$)^[5]. This, however, was in the era of high trough Tac targets of approximately 10-15 in the first 3 mo.

Noting that over time target drug levels have decreased, the use of therapeutic drug monitoring may assist in the management of prevention of rejection and complications of immunosuppression. There is some evidence that dysglycemic states are related the degree of CNI exposure. For example, Chan *et al*^[15] randomised 292 patients to low dose Tac (trough level 5-9 for first 3 mo then trough level 3-6 following 3 mo) or standard dose (trough level 10-15 for first 3 mo then trough level 8-12 following 3 mo). All patients received basiliximab,

similar doses of MMF and glucocorticoids over the follow up period of 6 mo. Those in the low dose Tac group had significantly less NODAT incidence over 6 mo of follow up, with a tendency towards lower incidence rate of treated diabetes^[15]. Similarly the dose response effect with respect to NODAT risk has also been described with the use of CsA with less dysglycemia post-transplant in those treated with low dose CsA (C2 600-800)^[19]. Sub-analyses of data from larger trials, such as Efficacy Limiting Toxicity Elimination-SYMPHONY, have also suggested a dose-dependent relationship. SYMPHONY found significantly higher rates of NODAT in the low-dose Tac group, compared with low-dose CsA, low-dose sirolimus or standard dose CsA without induction agent ($P = 0.02$)^[75]. Given the issues with choice of diagnostic test it is not surprising that when analysed according to F BGL there were no significant differences between the groups^[84].

As age is commonly identified as a risk factor in univariate analysis, it is important to know if older age interacts with other risk factors. In a multivariate analysis of OPTN data there is a clear increase in risk with increasing age when grouped into age groups using 18-34 years old as a reference group^[46]. Amongst the other identified risk factors use of Tac increased risk of NODAT. An analysis of the OPTN registry data compared rates of acute rejection and rates of NODAT and their impacts of graft survival. The rates of acute rejection were less in the older Tac treated patients, but the rates of NODAT were greater in the same older Tac treated group^[51]. The authors comment that targeted and individualised use of immunosuppression based on the patient's risk profile may help to ameliorate worse outcomes. Part of this may be to reconsider the use of CNI, in particular Tac, in the older recipient in whom the

development of NODAT may precipitate morbidity and mortality. However, as outlined below, other strategies may be safer and more effective.

Transplant related factors: Glucocorticoids

Oral glucocorticoids form the backbone of many immunosuppressive regimens and the diabetogenic potential of these agents is well documented. The development of diabetes is related to the cumulative exposure to glucocorticoids. The data available on glucocorticoid withdrawal, glucocorticoid free or rapid glucocorticoid tapering suggests an incidence rate of 1%-22% over a 1-5 year follow-up period^[12,19,26,46,60,69,85] which compares with rates of 15%-35% in regimens without glucocorticoid maintenance (Table 1). However, not all analyses find a benefit in glucocorticoids avoidance. For example, a meta-analysis of higher quality trials in which patients had glucocorticoid withdrawn within 14 d post-transplant and were treated with CNI/MMF did not find a reduction in NODAT^[86]. However, the largest randomised placebo-controlled trial ($n = 386$) of early glucocorticoid withdrawal within 7 d of transplantation found no difference in the rate of NODAT, although fewer of the NODAT patients required insulin therapy in the early glucocorticoid withdrawal arm^[83]. Furthermore, a matched cohort analysis of glucocorticoid free and maintenance therapy with glucocorticoid ($n = 190$ in each group) there were no differences in renal specific outcomes or any differences between F BGL or use of hypoglycemic agents. It is noteworthy that there was significantly more use of Tac and basiliximab in the glucocorticoid free group^[85]. Nonetheless, many other studies do find an advantage to glucocorticoid avoidance. Analysis of United States Renal Data System (USRDS) data found that patients discharged on a glucocorticoid containing regimen had an OR of 1.42 for NODAT compared to those discharged on a glucocorticoid free regimen^[46]. These results must be interpreted with caution, as it is not possible to capture the cumulative glucocorticoid exposure in the USRDS database. One small ($n = 62$) randomised prospective study in which glucocorticoids were ceased in one group by day 10 found a significant decrease in the incidence of NODAT when defined as used of hypoglycemic agents^[68]. A more recent pilot study ($n = 48$) of thymoglobulin induction, MMF, low dose CsA and rapid glucocorticoid reduction in low immunological risk patients found that this protocol resulted in 42 of 48 patients being normoglycemic at 6 mo^[19]. A larger single centre population ($n = 1291$) retrospectively analysed in which NODAT was defined as need for hypoglycemic agents found an incidence rate of only 2%-4% in the first year post transplantation in patients treated with glucocorticoid withdrawal after day 5 post-operative in combination with thymoglobulin induction, CNI plus sirolimus or MMF^[69]. This was a significant improvement compared to a non-matched historical control group who received a glucocorticoid containing maintenance regimen. Despite the theoretical

benefits of glucocorticoid withdrawal the studies referenced above demonstrate conflicting results^[87]. The impact of glucocorticoid exposure on the development of NODAT may be answered by a current trial in which patients of low immunological risk will be randomised to one arm including thymoglobulin induction and glucocorticoid free CNI/MMF maintenance or basiliximab induction and ongoing glucocorticoid exposure^[88].

The development of dysglycemia subsequent to the diagnosis and treatment of acute rejection may also disclose the risk of dysglycemia created by glucocorticoid exposure. A single centre review of 526 transplant recipients had a NODAT incidence of 16.7% when defined using ADA/WHO criteria for assessing random blood glucose or HbA1c. They found that there was a greater incidence of acute rejection in patients who developed NODAT and that intensified treatment with glucocorticoid and possible conversion to Tac was associated with increased risk of NODAT on multivariate analysis. However, the analysis did not treat rejection as a time varying co-variate^[26].

Transplant related factors: Sirolimus

Sirolimus, a mammalian target of rapamycin (mTOR) inhibitor, is an immunosuppressive agent used in conjunction with, or instead of, calcineurin inhibitors. Clinical data suggests that sirolimus use is not without risk for the development of NODAT^[77]. Analysis of USRDS of 2598 patients recorded as having received sirolimus, found that the combination of sirolimus with a CNI created a higher HR for cumulative 1yr incidence of NODAT compared to CNI with mycophenolate/azathioprine (MMF/AZA) or sirolimus with MMF/AZA. A sub-group multivariate analysis of USRDS data of 16861 patients known to have remained on the same immunosuppressant regimen patients treated with the combination of sirolimus and a CNI remained at increased risk of 1 year NODAT^[51]. In one study of non-NODAT renal transplant recipients who were switched from CNI to sirolimus there were no improvements noted in the glycemic state of the patients when studied robustly with oGTT. Indeed higher sirolimus levels in the absence of CNI may have increased the risk of NODAT^[74].

However, just as with the data on CNI and glucocorticoids there are inconsistent findings in the literature on sirolimus. A recent large ($n = 440$) prospectively randomised trial found that higher dose Tac, but not high or standard dose sirolimus contributed to the NODAT^[60]. A further example is a recent study of patients randomised to tacrolimus/mycophenolate, TAC/sirolimus or CsA/sirolimus. The median follow up was 8 years and the quoted cumulative incidence of NODAT was 19%-32%, with no significant differences between the groups based on the use of hypoglycemic agents^[76]. Lastly, as with CNI, it is likely that there is an important interaction between modifiable and non-modifiable risk factors. For example, a multivariate

analysis has found that older age and higher sirolimus trough levels were associated with increased hazard for NODAT^[77], once again suggesting that drug level targets in older recipients could be reviewed, for both effect and toxicity.

Transplant related factors: Other medications

Calcineurin inhibitors and glucocorticoids are the most well studied drugs in terms of impact upon glycemic control. There is no data on the contribution of MMF or AZA to the development of dysglycemia. In the transplant literature, there does not appear to be a signal that these drugs may be implicated. Recently, there has been interest in the possibility that basiliximab, a widely used induction agent particularly in the lower immunological risk patients, may be implicated in contributing to dysglycemia; although this is based on two data sets, neither of which were prospective or randomized^[27,58]. There is also little data on the contribution of thymoglobulin to the development of NODAT. A study of single dose vs divided dose anti-hymocyte globulin (ATG) induction analysed dysglycemia as a secondary outcome. In this study, fasting blood sugar levels after 1 mo to 6 mo were significantly lower ($P = 0.02$) in patients who received single dose ATG induction^[48].

PATHOGENESIS

The pathogenesis of dysglycemia post transplantation is complex and is widely assumed to be closely aligned to the pathogenesis of type 2 diabetes mellitus. However, this assumption underestimates that the impact of end stage renal failure and dialysis on glucose homeostasis. There is also little known about the histological changes in the graft over time when exposed to persistent NODAT. Small case series have found *de novo* diabetic nephropathy within 5-10 years of diagnosis of NODAT^[89,90].

Changes in both insulin resistance and insulin secretion can be shown to underlie the development of the dysglycemia post transplantation. These changes are however dynamic and sometimes transient, particularly in the early post-transplant period. Lastly, the role of changes in incretin hormones remains to be elucidated, as does the impact of the severity of chronic kidney disease (CKD) pre- and post-transplant on insulin metabolism and resistance.

Pre-transplant factors

The dynamic nature of dysglycemic states has been documented by Hornum *et al.*^[33]. They followed 57 patients from pre- to 12 mo post-transplant. Importantly, none were diabetic on an oGTT pre-transplant, however only 67% were normoglycemic. At 3 mo only 46% were normoglycemic and this increased to 56% by 12 mo. Pre-transplant, patients were compared with uremic controls. The transplanted patients were significantly younger (39 vs 47 years old) with shorter period of

time on dialysis (24 mo vs 45 mo); however, they did not differ in terms of measure of glycemic state. These measures included F BGL, oGTT and then specific validated measures of insulin resistance and secretion. It is noteworthy that both the uremic controls and transplant patients had a worse glycemic state than a small group of healthy controls - despite normal F BGL [5.1 mmol/L (all ESKD) vs 5.0 mmol/L]. The normal F BGL would suggest that hepatic gluconeogenesis was not impaired by the ESKD state; however, the ESKD patients had oGTT results of 7.4-7.5mmol/L (vs 5.4 mmol/L) and this seemed to be accounted for by increased peripheral insulin resistance. Interestingly, the increased resistance in ESKD patients was matched by increased insulin secretion compared to healthy controls (although not statistically significant). This may have been expected for two reasons. Firstly, ESKD patients will have reduced renal clearance of insulin^[91]. Secondly, as insulin resistance and insulin secretion are described as being related in a hyperbolic fashion^[92], such that changes in one parameter would be expected to drive compensator changes in the other parameter. Whilst there is evidence in these cohorts of compensatory increase in insulin secretion, it can be postulated that it was insufficient as the ESKD patients had markedly higher oGTT results and 33% were found to have IGT. At 12 mo, 14% of patients had developed NODAT and this was associated with increased insulin resistance and increased insulin secretion, which nonetheless, appeared not to be sufficient to maintain normoglycemia. The development of NODAT was not associated with pre-transplant IGT. However, those who developed dysglycemia tended to be older and have a higher pre-transplant BMI, which may co-vary (although not significant in multivariate analysis) with the noted increased pre-transplant insulin resistance and, again, higher compensatory pre-transplant insulin secretion.

Insulin resistance

Increasingly, understanding the factors responsible for insulin resistance and decreasing insulin secretion is being recognised as important for determining modifiable and treatable causes of NODAT. An increase in insulin resistance would be consistent with exposure to glucocorticoids. Glucocorticoids are believed to impair peripheral glucose uptake, impair hepatic glycogen synthesis and enhance gluconeogenesis. At higher doses they may induce β -cell apoptosis^[93]. Furthermore, it has been proposed the diabetogenic risk is not restricted to higher dose of glucocorticoid but also occurs with chronic exposure to low doses^[94]. In addition to duration and dose of glucocorticoid, older age and higher BMI also predispose to the development of diabetes in those receiving glucocorticoid treatment^[95]. Perhaps it is less well recognised that CKD and uremia may also contribute to insulin resistance. It may be that the relief from uremia, but the nonetheless persistent state of CKD post-transplant contributes to the dynamic nature of post-

transplant dysglycemia. It may also be that whilst clearly the biological stress of transplantation and exposure to diabetogenic medications is crucial in the pathogenesis, the persistence of CKD in certain older and perhaps genetically predisposed patients forms a background milieu upon which the dysglycemia can develop. There has been renewed interest in the contribution of uremia or CKD to insulin resistance and the various mechanisms are beyond the scope of this article. However, when reading literature on post-transplant dysglycemia it is important to remember that transplant patients have had periods of severe CKD/ESKD requiring dialysis and, for the most part, remain a CKD patient^[96,97]. One study of 27 diabetic and 35 non-diabetic ESKD patients using a homeostatic model assessment-insulin resistance model to assess insulin resistance found increased insulin resistance in the diabetic patients. The non-diabetic patients with increased insulin resistance had elevated C-peptide levels, indicating a compensatory response maintaining non-diabetic state^[98].

Other factors that may increase insulin resistance post-transplant include hepatitis C virus (HCV) and metabolic syndrome. Two studies have found that HCV-positive patients have increased insulin resistance compared to non-HCV transplant patients. One of these studies found a compensatory increase in insulin secretion^[99] and one did not find such compensation^[70]. On the other hand, CMV, the other recognised diabetogenic virus, seems to be associated with impaired insulin secretion; although, the exact mechanism is not well studied^[59]. Whilst metabolic syndrome has been described in the general population to be associated with insulin resistance, there is a paucity of data considering metabolic syndrome and insulin resistance in transplant recipients. A recent retrospective review of 76 patients with a mean 11.1 years post-transplant follow up found that even when adjusted for age, the presence of metabolic syndrome was associated with increased risk progression of dysglycemia^[22]. In a larger cohort of patients ($n = 640$), the presence of metabolic syndrome pre-transplant remained a significant risk factor for developing NODAT even when adjusted for age^[9]; however, there is no data available on insulin resistance in any significant cohort of transplant recipients who develop metabolic syndrome and NODAT.

Insulin secretion

It seems likely that as modifiable risk factors are altered, importantly including immunosuppressive agents, that the weights of forcing factors of NODAT will also be altered. As such, studies that repeatedly measure insulin indices throughout the post-transplant period, in particular in the higher risk first year post-transplant, are particularly valuable. Nagaraja *et al.*^[22] has recently described insulin indices pre- and 3 and 12 mo post-transplant in non-diabetic patients ($n = 118$) as defined by F BGL less than 7.0 mmol/L pre-transplant. The patients defined as NODAT had increased insulin

resistance at 3 and 12 mo, although less resistance at 12 mo when compared to 3 mo. By 12 mo, insulin secretion had fallen in patients with NODAT; however, despite the fall in insulin resistance the levels of secretion failed to be compensatory, suggesting that even in the face of falling doses of glucocorticoid and improving peripheral insulin sensitivity, impaired insulin secretion increasingly threatens normoglycemia^[20,100]. This data is supported by previous studies in which oGTT was used for diagnosis^[101,102]. Nam *et al.*^[102] first demonstrated impairment in insulin secretion as a necessary component in the pathogenesis. They followed 144 patients pre- and post-transplant and noted that higher, although normal, oGTT results pre-transplant were associated with increased risk of dysglycemia post-transplant. They also noted that those who developed post-transplant dysglycemia 9-12 mo post-transplant had significantly lower insulin secretion in the face of improved insulin resistance. A long term study found similar results when using oGTT at 10 wk and 6 years post-transplant. Patients who were dysglycemic at 10 wk and became normoglycemic had improvement in insulin resistance and a non-significant impairment of insulin secretion, thus retaining a compensatory response. On the other hand, those who remained diabetic or became diabetic over the follow-up period had a non-significant deterioration in insulin resistance and a significant fall in insulin secretion^[103].

The mechanism of impairment in insulin secretion post-transplant is thought to be related to CNI use. The mechanism of action is believed to be the impairment of pancreatic cell function due to the binding of CNI to calcineurin. Calcineurin is a systolic phosphatase that has two targets in the β -cell: the nuclear factor of activated T cells and cyclic-AMP-responsive element-binding protein transcriptional co-activator. In mice models, normal β -cell function has been shown to be dependent upon calcineurin^[104]. Calcineurin may be important for the proper response to hyperglycemia and incretin activation. Human islet cells when treated with Tac increased β -cell apoptosis, possibly mediated by the above calcineurin targets and ameliorated by the administration of incretin analogues^[105,106].

Incretins

Finally, there is no data on the impact of immunosuppression in renal transplant patients on incretin hormones. It is interesting to note that in healthy volunteers the administration of glucocorticoids in the setting of being sedentary and on a high calorie diet (not unlike the initial period of time post-transplant) have impaired responses to incretin hormones^[107]. In dialysis dependent patients, those with IGT have been shown to have a reduced incretin effect^[108], and even normoglycemic dialysis dependent patients have reduced insulin secretion with increased incretin secretion suggesting that uremia or CKD impacts upon the proper β -cell stimulation and response^[109]. However,

Table 4 Risk of mortality, cardiovascular events and graft loss associated with new-onset diabetes after transplantation or dysglycemic state

	Mortality	CV event/death	Graft loss	Ref.
Diabetes at	3 mo: 37% at 8 yr (HR = 2.1) 10 wk: 34% at 6.7 yr (HR = 2.0) 1 yr: 44% at 11 yr (HR = 2.2)	20% (death) at 8 yr (HR = 3.5)		Hjelmsaeth <i>et al</i> ^[4] Valderhaug <i>et al</i> ^[11] Nagaraja <i>et al</i> ^[20] Cosio <i>et al</i> ^[3]
Dysglycemia at	10 wk: 29% at 6.7 yr (HR = 1.78) each 1 mmol/L oGTT: 5% risk increase 4 mo: 0.5 mmol/L increase F BGL: 4% risk increase 12 mo: 0.5 mmol/L increase F BGL: 15% risk increase	Death HR: 2.72 Events increased with increased F BGL 1 mmol/L oGTT: 6% risk increase in death 12 mo: 0.5 mmol/L increase F BGL: 11% risk increase for event	3 mo: RR 3.6 at 6 yr	Valderhaug <i>et al</i> ^[11] Wauters <i>et al</i> ^[14] Wojtusciszyn <i>et al</i> ^[41]

F BGL: Fasting blood glucose level; oGTT: 2-h oral glucose tolerance test.

the dynamics of incretin hormones are yet to be described in the post-transplant setting.

OUTCOMES

There is an urgent need to develop a consensus on the best test to detect and how to manage dysglycemic states post-transplant, as there is a direct correlation with the presence of dysglycemic states and mortality predominantly from cardiovascular causes (Table 4)^[3,4,11,14,20]. An analysis of the USRDS database in which NODAT was defined according to Medicare claims analysed 27707 patients with data available greater than 1 year and not diabetic pre-transplant. Death censored graft loss was more likely in those who suffered acute rejection when compared to those who developed NODAT. Conversely, those who developed NODAT had a higher hazard ratio of death with a functioning graft compared to those with episodes of acute rejection (1.41 and 1.15 respectively) compared to patients with neither exposure^[51]. Analysis of earlier data from the same database found the development of NODAT associated with increased risk for acute myocardial infarction after a minimum 3 year follow up^[110]. Similarly, in an analysis on the International Collaborative Transplant Study database ($n = 39251$) with up to 10 years of follow up, Cox regression analysis of death with a functioning graft due to cardiovascular disease revealed an increased risk for NODAT (HR = 1.6, $P < 0.001$), which was greater than episodes of rejection within the first year (HR = 1.2, $P = 0.036$) but not as great as the risk associated with pre-transplant diabetes (HR = 2.5, $P < 0.001$)^[111].

The above datasets are large and their analyses robust, but what is needed are large prospective datasets with well-defined populations and sufficient duration of follow up. Smaller studies have found significant risk for mortality from the development of NODAT, but these findings have disappeared when adjusted for confounding factors. In one such study, major cardiac events occurred in 20% of persistent NODAT patients compared to 7% without NODAT and 21% with pre-transplant diabetes over a 8 year follow up^[4]. The outcomes of the largest prospectively followed well defined

population was described by Valderhaug *et al*^[112]. They followed 1410 patients for a mean of 6.7 years, of whom 55% were dysglycemic at 10 wk post-transplant of which 17% had NODAT. They reported a significant increase in the incidence of all cause mortality between the normoglycemic and dysglycemic groups, the rates being highest in those with NODAT. After adjusting for confounding traditional and transplant associated variables, the HR for all cause mortality was 1.54 was NODAT and 1.39 for IGT ($P < 0.05$). When analysed treating glucose as a continuous variable: on adjusted analysis, for every 1 mmol/L (18 mg/dL) increase in oGTT result there was a 5% increase risk in all cause mortality ($P < 0.05$). The main cause of death was cardiovascular disease, and those with NODAT by 10 wk were at significant increased risk on adjusted analysis (HR = 1.8 $P < 0.05$). For every 1mmol/L (18 mg/dL) increase in the oGTT result there was significant 6% increase risk in cardiovascular death ($P < 0.05$). Despite the findings of the continuous glucose analysis, other dysglycemic states were not associated with cardiovascular death. Further analysis of the same cohort found a graft failure rate of 28%, 60% of which was due to death. There was no association with death censored graft loss, but for every 1 mmol/L (18 mg/dL) increase in oGTT result there was a 3% increase risk in overall graft failure^[110]. This suggests similar conclusions as the large registry analyses described above: NODAT may not be associated with increased graft loss, but is associated with increased mortality.

In another large single centre prospectively followed group an increase in risk of all cause mortality and cardiovascular death according to the presence of NODAT at 1 year post-transplant was reported^[14]. The 12-mo rate of dysglycemia was 29.8% and NODAT 13.4%. Continuous analysis of the glucose levels revealed that for every 10 mg/dL (0.56 mmol/L) increase in F BGL there was an increase in all cause mortality censored at graft failure over a follow up period of 90.4 mo. At 12 mo, patients with IFG had a HR of 1.7 ($P = 0.009$) and those with NODAT a HR of 3.5 ($P < 0.0001$). Of note, in this study the patients on treatment for NODAT did not have a reduced mortality risk compared to the NODAT

patients not on treatment. Given the retrospective nature of the analysis it is not possible to conclude that treatment does not affect outcomes. However, such findings indicate the importance of well-defined prospectively followed transplant population analyses and potentially the need to identify early those patients at risk of dysglycemia so that directed interventions (be they aggressive glucose or metabolic risk factor control) may ameliorate the increased risk of mortality. Furthermore, such data highlights that in the transplant population clinicians do not have targets of glycemic control that can be achieved with treatment and are associated with improved outcomes. Even in the general population there is conflicting data concerning improved macrovascular outcomes achieved by treating to more intensive targets^[113,114]; however, in the transplant population, it remains unknown if meeting these same targets may improve outcomes.

SCREENING AND DIAGNOSIS

Use of oGTT remains the gold standard for diagnosis of NODAT or dysglycemia. This test, however, is not an easily completed screening test. Simple office or laboratory based tests that may be used to adequately screen for NODAT, particularly in high risk patients, include F BGL, 4 pm capillary blood glucose or HbA1c. All of these parameters have limitations. For example, a Spanish study of 374 non-diabetic pre-transplant patients found that normal F BGL in 59% of patients with an abnormal oGTT over the first 12 mo post-transplant^[6]. It is well known that changes in red cell viability, need for (due to for example, drug induced bone marrow suppression) and use of erythropoietin stimulating agents, administration of red cell transfusions and changes in hemoglobin will impact upon HbA1c levels. Notwithstanding this issue more readily encountered in ESKD, some small studies ($n = 71$) have shown concordance between oGTT and an HbA1c cut-off of 6.2% for the diagnosis of NODAT^[115]. It would be clinically more likely to find concordance between these tests after 2-3 mo post-transplant once there has been renal function recovery and the impact of uremia on erythropoiesis has resolved. However, analysis of a much larger cohort ($n = 1571$) found that using if HbA1c was used as a screening tool and oGTT as the gold standard test, then the cut-off should be 5.8%^[44]. More recently, when using a combined test of HbA1c $\geq 6.5\%$ and F BGL ≥ 7.0 a Norwegian group ($n = 1619$) have demonstrated a negative predictive value (NPV) of 97.4% for NODAT, using oGTT as gold standard test at 10 wk post transplantation^[116]. Notably, the combination of the two tests had very little additive value (NPV F BGL alone 94.2%) and the lower the HbA1c cut-off value made little difference in exclusion of NODAT (e.g., $\geq 5.5\%$ NPV 97.5 compared with $\geq 6.5\%$ NPV 93%). However, the positive predictive value of HbA1c $\geq 6.5\%$ or 6.2% or in combination with F BGL ≥ 7.0 mmol/L was poor (53.4%, 42.1%, 69.4% and 50.9%,

respectively). Thus, while HbA1c may be of use in screening for NODAT, current evidence does not support its use as a diagnostic test in transplant patients.

Determining the best test to use in transplant patients is complicated by the need to certain of the best time to administer the test. It has recently been shown that glucocorticoid administration in the morning leads to increased afternoon or evening blood glucose levels, at approximately 7-8 h after administration of glucocorticoid. Thus, reliance on F BGL may underestimate the incidence of dysglycemia. In fact, at six weeks post transplantation a 4 pm capillary blood glucose significantly outperformed oGTT, F BGL and HbA1c in detecting NODAT. Combining the tests done at 3 and 12 mo, the cumulative incidence of NODAT with oGTT was 14% and IGT 28%. Interestingly, using an HbA1c range of ≥ 5.7 and < 6.5 to detect IGT detected an incidence of 51%; but HbA1c did not perform as well as oGTT in detecting NODAT. Hence, the authors suggested using HbA1c as a screening test from 3 mo and using oGTT to determine the presence or absence of NODAT in patients detected to have dysglycemia by HbA1c. This strategy would avoid oGTT in 49% of patients and achieve a sensitivity of 94%^[117]. As yet, this data and strategy has not been replicated. Furthermore, the results of these studies suggest that the cut-offs that have been applied in the general population may not apply in CKD, ESKD or post-transplant patients. The question of the cut-off levels for any of the possible tests will only be settled by long-term large prospectively collected data sets which permit determination of the risk for poorer clinical outcomes associated with different cut-off points. Some of this data has already been described, but it is worth emphasising that only oGTT results have been shown to be associated with poorer outcomes when analysed categorically (as distinct from continuous data) and not F BGL^[11].

PREDICTING NODAT

If it is difficult to develop easy to administer diagnostic tests, it is even more challenging to develop to models that may predict the development of NODAT, based either on pre- or post- transplant data. There are very few studies able to draw conclusions about predicting NODAT using pre-transplant data. Post-transplant dysglycemia is dynamic phenomenon and there are multiple physiological changes post-transplant that may impact upon insulin and glucose handling. This is emphasised by the remarked upon cases of diabetic or dysglycemic pre-transplant patients resolving their dysglycemic state post-transplant. Hence, the pre-transplant prediction of those increasingly likely to have NODAT post-transplant is fraught with multiple difficult variables that need to be taken into account.

A range of pre-transplant variables has been described as predictors of NODAT. These include age, BMI, fasting and R BGL and metabolic syndrome. For example, one study of 139 non-diabetic patients

pre-transplant found that higher (albeit normal) pre-transplant R BGL were predictive ($P = 0.011$) of NODAT, although this data has not been replicated^[7]. A matched cohort retrospective analysis of 47 patients who developed NODAT found that a higher, albeit normal range, F BGL was associated with the development of NODAT on multivariate analysis^[44].

One reasonable sized study ($n = 640$) with a NODAT incidence at 1 year of 31.4% found an adjusted hazard for NODAT of 1.34 (1.00-1.79, $P = 0.047$) for pre-transplant metabolic syndrome. On multivariate analysis, only pre-transplant low HDL remained an independent predictor^[9]. Other groups have attempted to apply scores that are predictive of type 2 diabetes mellitus in the general population. A retrospectively analysed cohort of 191 patients in which 41 developed NODAT, two general population risk scores were found to have AUC-ROC of 0.756-0.807 for NODAT at 1 year, but the PPV for each test was poor (24.5%-31.2%). However, the authors point out that the NPV were high (92.5%-93.7%) perhaps allowing the identification of high risk patients^[118].

There is a small body of literature considering the development of predictive models that may be more unique to the transplant patient. Analyses in the general population of the patterns of oGTT results may be predictive of future type 2 diabetes^[119]; similar analyses in renal transplant patients may be useful. An analysis of a 5 time point oGTT conducted pre-transplant in 145 patients found that whilst F BGL did not predict NODAT, the AUC of the oGTT and the glucose concentrations at each time point post glucose load could be used to predict NODAT^[23]. Given the logistical difficulties in studying recipients of deceased donor organs, there is little data available that would enable us to reliably assess if pre-transplant markers for NODAT can be identified. For example, one study in which 120 transplanted patients were screened with oGTT pre-transplant found that pre-transplant IGT was significantly associated with NODAT; however, these patients were screened during the 3 mo prior to being waitlisted and there was no information provided regarding the time on the waiting list. This may introduce a potential bias in that some normoglycemic patients may have developed further dysglycemia pre-transplant^[31].

Chakkerla *et al.*^[120,121] have attempted to develop and validate a model of pre-transplant factors to predict the development of NODAT. On univariate analysis they described seven pre-transplant factors associated with increased risk of NODAT, which was defined by use of HbA1c, F BGL or requirement for treatment, including dietary changes. The seven factors were: age greater than 50 years old, use of maintenance glucocorticoids, use of gout therapies, BMI ≥ 30 , F BGL ≥ 5.6 mmol/L, fasting triglycerides ≥ 2.24 mmol/L and a family history of type diabetes. Insulin indices were not measured and pre-transplant oGTT were not done pre- or post-transplantation, potentially treating pre-transplant diabetic patients as normoglycemic. Complex statistical

methods, including bootstrapping were used. Within the limitations of this study, there were clear differences in the 1 year incidence of NODAT for those classified as low, moderate or high risk according to seven factor risk score. The results were similar in the initial and validation groups. In the higher risk group the incidence of NODAT was 44%-56% compared to the low risk group of 11%-13%. This was a first step in attempting to develop a risk score that may assist in identifying patients who could be targeted for trials of preventive therapies.

The analysis of the data from the 5 time point oGTT points towards the possibility of identifying higher risk patients by evaluating for impaired glucose and insulin regulation pre-transplant. A test that is helpful in this regard is known as the disposition index. This is a quantification of the hyperbolic balance between insulin secretion and insulin resistance. It can be measured either *via* oral or IV glucose loads and has been shown to be associated with increased risk for developing type 2 diabetes mellitus in the general population^[122,123]. There is little literature using the disposition index as a predictive pre-transplant marker. However, there are some small studies measuring insulin resistance and secretion pre-transplant and testing their relationship with NODAT. Various models that utilise data derived from oGTT or IV GTT measure insulin resistance. The homeostasis model (HOMA) is widely used, and has been validated in studies of the general population. Variations of HOMA can be used to estimate insulin resistance and secretion. There is conflicting data on whether pre-transplant insulin indices may be predictive and most studies are small^[124]. A study with the primary purpose of comparing Tac and CsA ($n = 150$) was used to retrospectively review the risk of NODAT from pre-, 3 and 12 mo indices of insulin resistance. Pre-transplant, there were no differences in insulin resistance or secretion found between those patients who developed NODAT at 3 or 12 mo^[20]. This is in contrast to an earlier study ($n = 57$) in which those patients more resistant at baseline (and older) had an increased odds of a dysglycemic state after 1 year follow up^[33]. However, as it appears increasingly more likely that falls in insulin secretion (and thus failing to compensate for insulin resistance) is crucial in the development of NODAT, it is interesting to note that measurements of insulin secretion in non-diabetic post-transplant patients can be used to predict the future development of NODAT^[98].

MANAGEMENT

The principles of management of post-transplant dysglycemia are: (1) Pre-transplant risk assessment and development of amelioration strategies; (2) Early detection and monitoring for transient or permanent dysglycemia; and (3) Appropriate therapies that may reduce the poorer outcomes in those in whom post-transplant dysglycemia develops. The issues surrounding risk assessment and detection have been discussed

above. Current advice for glucose targets during post-transplant hospitalization suggest maintaining glucose levels below diabetic range; *i.e.*, F BGL 4-7 mmol/L (72-126 mg/dL)^[125]. Following discharge, current guidelines recommend that patients be screened weekly for the first four weeks, and every 3 mo for the first year and yearly after the first year. Screening should also be commenced if there is commencement of, or substantial increase in dose of, CNI, mTOR inhibitor or glucocorticoids^[126]. There is no consensus on the best screening test to utilise; however, a combination of tests as discussed above would appear to be of greatest clinical use. This may involve weekly F BGL or 4 pm capillary blood glucose (although this is not currently part of guidelines). Detection of IFG would then prompt oGTT assessment^[125]. Perhaps use of HbA1c after the first 3 mo is warranted in stable patients. There are also few recommendations as to what the targets for blood glucose and HbA1c ought to be, as it is not known at what ranges there is substantial reduction in poorer outcomes. At present, guidelines give an ungraded suggestion to aim for an HbA1c of 7%-7.5% in United States^[126] and < 7% in Scandinavia^[125].

Adjusting immunosuppression

One approach to management is amelioration of risk. It remains difficult to identify patients at risk for dysglycemia with certainty; equally, it is challenging to know what may be done should they be identified. On the basis of data available concerning modifiable risks, physicians may wish to replace, minimise or withdraw one or more agents that form part of the maintenance immunosuppression; in particular CNI or glucocorticoids. For instance, perhaps older patients with a higher BMI and a worse (if still normal) pre-transplant oGTT may be judged to be at risk and as a result not exposed to maintenance glucocorticoids, or use of CsA in preference to Tac. This approach clearly needs to balance the immunological risk of reduced immunosuppressive exposure against the higher metabolic (and ultimately cardiovascular and infection) risk. Some authors have proposed protocols to assist in balancing the metabolic and rejection risk^[61]; however, there are no well validated methods for reliably making such assessments in a broad transplant population. In addition, the clinician is also faced with the complicated issue of applying risk assessments to individual patients with varying degrees of co-morbidities.

One potentially helpful immunosuppressive agent that has not been discussed above is belatacept. This co-stimulatory blockade agent, which remains available in for off-label use in many countries, can be used as part of a maintenance regimen in place of CNI, in combination with MMF and glucocorticoids. The BENEFIT and BENEFIT-EXT (extended criteria donors) trials have reported up to 5 year results, comparing belatacept with MMF and glucocorticoids with CsA, MMF and glucocorticoids. There is a concern that there

may be greater early acute rejection, however, over longer follow up there is no greater rejection rate. There has also been a concern about increased risk for EBV associated post transplant lymphoproliferative disease^[127-130]. With regard to NODAT, results from 1 year follow-up of BENEFIT and BENEFIT-EXT have been published. There was a significant reduction in the 1 year cumulative incidence of NODAT in the belatacept arm, with rates of NODAT in the CsA arm being comparable to that found in other studies. This was in conjunction with clinically significant reductions in blood pressure, cholesterol and triglycerides, suggesting it may have a role in management of patients at higher risk of poorer cardiovascular and metabolic outcomes^[131].

Lifestyle changes

Aside from altering immunosuppressive agents, other modifiable risk factors include reduced physical activity and poor diet. There is some data to suggest that low levels of physical activity post-transplant, particularly in patients whose appetite may now be improved, are at greater cardiovascular and all-cause mortality risk^[132]. Improved diets, increased physical activity and weight loss has also been shown to improve dysglycemia in renal transplant patients^[133]; however, this is not a well studied therapeutic approach.

Intensive and early glycaemic control

As there are many obstacles to overcome should immunosuppression be tailored to meet metabolic and immunological risk, it may be that we require strategies to "rest" β -cells in patients without changing immunosuppression in those at higher risk of metabolic complications. Hecking *et al.*^[38] in a proof of concept trial ($n = 50$) randomised patients to (non-blinded) early basal insulin or standard therapy. NODAT was defined by oGTT or need for hypoglycemic agents at study visit. All patients received maintenance Tac, glucocorticoids and MMF. Patients were given isoprene insulin if their evening blood glucose was > 140 mg/dL (7.8 mmol/L) in the treatment group; the standard of care group received short acting insulin or oral agents if their blood glucose was 180-250 mg/dL (10-13.9 mmol/L), as directed by the treating clinician. All 25 patients in the treatment group received isoprene insulin on postoperative day 3, having had high evening blood glucose the day prior. By 12 mo, no patient in the treatment group required hypoglycemic agents compared to 8 in the control group. The majority of the patients in the treatment group did not receive any hypoglycemic agent after 120 d post-operative. All patients not on hypoglycemic agents had oGTT at 3, 6 and 12 mo. By 12 mo, 5 patients in the treatment group had NODAT on oGTT compared to 4 in the control group; thus, there was a reduction in NODAT from 12 to 5. More patients in the treatment group had IGT (8 vs 5); but, overall, more patients in the treatment group were normoglycemic (12 vs 8). Furthermore, consistent with the more

recent literature on insulin secretion as a significant contributor to the pathogenesis of post-transplant dysglycemia, measures of insulin resistance between the groups did not differ at 12 mo. There was, however, a significant difference in the insulinogenic index, an oGTT derived measure of β -cell function. There was also an improvement in the disposition index (although not significant). Together these results would indicate better or more preserved insulin secretion in those whose β -cells were "rested" at time of maximal stress. Should such results be achieved in a larger study population (perhaps of higher risk patients) who are studied for a longer period of time and found to have better metabolic and cardiovascular outcomes, then it may be that early basal insulin in those with elevated evening blood glucose may become a standard of care obviating any need to tailor immunosuppression.

Standard hypoglycemic agents

Nonetheless, currently patients receive care more like the standard care administered in Hecking *et al.*^[38]. If these patients then develop NODAT, they receive hypoglycemic agents. The choice of agent is mostly guided by opinion and knowledge of risks associated with administration of these agents in CKD. This is due to the paucity of trial data on use of hypoglycemic agents within this population. There is only one small study ($n = 48$) that compares potential therapies in which a DDP IV inhibitor, vildagliptin, was compared with pioglitazone or placebo in patients with IGT at more than 6 mo post renal transplantation. Both medications reduced oGTT blood glucose levels over 3 mo, with no differences between the treatment groups^[134]. As there is concern that thiazolidinediones may be associated with poorer cardiovascular outcomes, such medications may not be considered as first line therapy. The incretin analogues, remain the only other hypoglycemic agent studied in transplant patients. Vildagliptin has been studied as part of a randomised placebo controlled trial, in which patients with oGTT defined NODAT at least 6 mo post-transplant were recruited. Thirty-three patients were recruited, all of whom were on a similar maintenance regimen of CNI/MMF and glucocorticoids. The follow up period was short, however, vildagliptin did significantly reduce oGTT and HbA1c results at 3 mo with no hypoglycemic events^[135]. Caution should be used with vildagliptin in conjunction with ACE inhibition as there is an increased risk of angioedema (OR = 4.57), albeit on the basis of a small absolute risk^[136]. Another small study ($n = 19$) has shown that sitagliptin can significantly increase insulin secretion in patients known to have NODAT^[137]. Sitagliptin, saxagliptin and vildagliptin should be dose reduced in renal impairment, linagliptin is not renal excreted. It is unclear if incretin analogues are ameliorating an impact upon the incretin effect or assisting β -cell function in other ways. There is no data in the transplant population concerning the incretin effect. In healthy people administered glucocorticoids the incretin effect has been noted to be

impaired^[103]. In favour of incretin analogues, they do not tend to produce hypoglycemia or weight gain; but they have not been shown to reduce cardiovascular events, have been associated with pancreatitis and may theoretically increase cancer risk^[138].

The incretin analogues are not widely used in the transplant population, with use of sulfonylureas and more common. Metformin may not be favoured as it can contribute to gastrointestinal side effects, potentially exacerbating the same caused by MMF use. Moreover, there is also no data on its use in transplant patients with GFR < 30 mL/min and risks of lactic acidosis. However, its lack of contribution to weight gain, its association with reduced cardiovascular events in non-transplant patients and its role as an insulin sensitiser rather than stimulating further insulin secretion from "stressed" β -cells, may make metformin more favoured than sulfonylureas^[139]. Sulfonylureas do not have the cardiovascular benefits and can contribute to weight gain. However, as long as dose adjusted to prevent hypoglycemia, their use is not associated with other serious adverse events. Nonetheless, it may be that some, if not most, transplant patients with develop dysglycemia have impaired β -cell function and that potentially a treatment strategy that induces more work from the β -cells may be counter-productive in terms of relieving dysglycemia and preventing worse cardiovascular outcomes^[140]. Problematically, the paucity of data on treatment (including treatment targeted at the underlying pathology) in this area of transplantation means it is not possible to make any firm recommendations on the choice of oral hypoglycemic agents.

CONCLUSION

In summary, dysglycemic states, not limited to NODAT, are associated with increased risk of mortality, principally as a result of cardiovascular disease. NODAT is better studied than other dysglycemic states. The natural history of dysglycemic states is not well characterized, apart from the recognition of transient dysglycemia and NODAT within the first 3-6 mo post transplantation. The majority of persistent NODAT develops within one over the first year post transplant. Whilst the diagnosis is made using the WHO/ADA criteria accepted in the general population, there is no consensus on which test should be employed, either for screening or diagnosis. At present, oGTT remains the most reliable diagnostic test in the post-transplant setting. However, predicting the development of NODAT remains challenging. Possibly, the small group of patients who remain normoglycemic within the first week post-transplant are at very low risk of developing NODAT. There are a few studies that may assist in developing tools for identifying those at high risk.

There are multiple risk factors, some of which are modifiable. The most consistently found risk factor is increasing age and there is a growing body of liter-

ature documenting the genetic risk factors. The most well described modifiable risk factor is the use of immunosuppressive agents, in particular CNI (Tac more than CsA) and glucocorticoids. These agents likely contribute to the development of NODAT *via* different mechanisms – glucocorticoids encouraging insulin resistance and CNI *via* β - cell failure. It seems that reduction in insulin secretion is more important in the pathogenesis than insulin resistance.

Any attempt to balance the metabolic and immunological risks by adjusting immunosuppression is complicated. It may be better to identify higher risk patients and utilise a preventive strategy, such as described by Hecking *et al*^[38]. As evidence emerges of the importance of β -cell failure as a major contributor to NODAT, such as strategy appears promising. In the absence of prevention, the management of NODAT in order to prevent the poorer outcomes is important. However, it is not clear which agent is most likely to successfully treat NODAT and ameliorate the poorer outcomes. A number of options exist, and it may be that metformin is the best option if insulin is not required.

Finally, further research is needed on pathogenesis, identification of higher risk patients and development of preventive and safe treatment options. Such research needs to take into account the caveats that are identified with respect to previous research: confirming normoglycemia pre-transplant, using oGTT as the primary diagnostic test (although there may be a role for capillary blood glucose early post-transplant), using WHO/ADA to define clinical states, testing regularly to detect transient and permanent states and having adequate follow up to detect the development of permanent dysglycemic states that impact upon poorer clinical outcomes. It would be ideal if future research could also map the changes in insulin secretion, resistance and the incretin effect pre- and post- transplantation in an effort to better understand the pathogenesis and further delineate targeted prevention and treatment options.

REFERENCES

- 1 **Crutchlow MF**, Bloom RD. Transplant-associated hyperglycemia: a new look at an old problem. *Clin J Am Soc Nephrol* 2007; **2**: 343-355 [PMID: 17699434 DOI: 10.2215/CJN.03671106]
- 2 **Davidson J**, Wilkinson A, Dantal J, Dotta F, Haller H, Hernández D, Kasiske BL, Kiberd B, Krentz A, Legendre C, Marchetti P, Markell M, van der Woude FJ, Wheeler DC. New-onset diabetes after transplantation: 2003 International consensus guidelines. Proceedings of an international expert panel meeting. Barcelona, Spain, 19 February 2003. *Transplantation* 2003; **75**: S3-S24 [PMID: 12775942 DOI: 10.1097/01.TP.0000069952.49242.3E]
- 3 **Cosio FG**, Kudva Y, van der Velde M, Larson TS, Textor SC, Griffin MD, Stegall MD. New onset hyperglycemia and diabetes are associated with increased cardiovascular risk after kidney transplantation. *Kidney Int* 2005; **67**: 2415-2421 [PMID: 15882287 DOI: 10.1111/j.1523-1755.2005.00349.x]
- 4 **Hjelmsaeth J**, Hartmann A, Leivestad T, Holdaas H, Sagedal S, Olstad M, Jenssen T. The impact of early-diagnosed new-onset post-transplantation diabetes mellitus on survival and major cardiac events. *Kidney Int* 2006; **69**: 588-595 [PMID: 16395250 DOI: 10.1038/sj.ki.5000116]
- 5 **Vincenzi F**, Friman S, Scheuermann E, Rostaing L, Jenssen T, Campistol JM, Uchida K, Pescovitz MD, Marchetti P, Tuncer M, Citterio F, Wiecek A, Chadban S, El-Shahawy M, Budde K, Goto N. Results of an international, randomized trial comparing glucose metabolism disorders and outcome with cyclosporine versus tacrolimus. *Am J Transplant* 2007; **7**: 1506-1514 [PMID: 17359512 DOI: 10.1111/j.1600-6143.2007.01749.x]
- 6 **Delgado P**, Diaz JM, Silva I, Osorio JM, Osuna A, Bayés B, Lauzurica R, Arellano E, Campistol JM, Dominguez R, Gómez-Alamillo C, Ibernón M, Moreso F, Benitez R, Lampreave I, Porrini E, Torres A. Unmasking glucose metabolism alterations in stable renal transplant recipients: a multicenter study. *Clin J Am Soc Nephrol* 2008; **3**: 808-813 [PMID: 18322043 DOI: 10.2215/CJN.0492110]
- 7 **Ramesh Prasad GV**, Huang M, Bandukwala F, Nash MM, Rapi L, Montada-Atin T, Meliton G, Zaltzman JS. Pre-transplantation glucose testing for predicting new-onset diabetes mellitus after renal transplantation. *Clin Nephrol* 2009; **71**: 140-146 [PMID: 19203506]
- 8 **Luan FL**, Stuckey LJ, Ojo AO. Abnormal glucose metabolism and metabolic syndrome in non-diabetic kidney transplant recipients early after transplantation. *Transplantation* 2010; **89**: 1034-1039 [PMID: 20130496 DOI: 10.1097/TP.0b013e3181d05a90]
- 9 **Bayer ND**, Cochetti PT, Anil Kumar MS, Teal V, Huan Y, Doria C, Bloom RD, Rosas SE. Association of metabolic syndrome with development of new-onset diabetes after transplantation. *Transplantation* 2010; **90**: 861-866 [PMID: 20724958 DOI: 10.1097/TP.0b013e3181f1543c]
- 10 **Bergrem HA**, Valderhaug TG, Hartmann A, Bergrem H, Hjelmsaeth J, Jenssen T. Glucose tolerance before and after renal transplantation. *Nephrol Dial Transplant* 2010; **25**: 985-992 [PMID: 19854851 DOI: 10.1093/ndt/gfp566]
- 11 **Valderhaug TG**, Hjelmsaeth J, Hartmann A, Roislien J, Bergrem HA, Leivestad T, Line PD, Jenssen T. The association of early post-transplant glucose levels with long-term mortality. *Diabetologia* 2011; **54**: 1341-1349 [PMID: 21409415 DOI: 10.1007/s00125-011-2105-9]
- 12 **Ciancio G**, Gaynor JJ, Zarak A, Sageshima J, Guerra G, Roth D, Brown R, Kupin W, Chen L, Tueros L, Hanson L, Ruiz P, Burke GW. Randomized trial of mycophenolate mofetil versus enteric-coated mycophenolate sodium in primary renal transplantation with tacrolimus and steroid avoidance: four-year analysis. *Transplantation* 2011; **91**: 1198-1205 [PMID: 21107305 DOI: 10.1097/TP.0b013e3182003d76]
- 13 **Israni AK**, Snyder JJ, Skeans MA, Kasiske BL. Clinical diagnosis of metabolic syndrome: predicting new-onset diabetes, coronary heart disease, and allograft failure late after kidney transplant. *Transpl Int* 2012; **25**: 748-757 [PMID: 22548293 DOI: 10.1111/j.1432-2277.2012.01488]
- 14 **Wauters RP**, Cosio FG, Suarez Fernandez ML, Kudva Y, Shah P, Torres VE. Cardiovascular consequences of new-onset hyperglycemia after kidney transplantation. *Transplantation* 2012; **94**: 377-382 [PMID: 22820698 DOI: 10.1097/TP.0b013e318258483]
- 15 **Chan L**, Andres A, Bunnapradist S, Gugliuzza K, Parasuraman R, Peddi VR, Cassuto E, Hart M. Renal Function and NODM in De Novo Renal Transplant Recipients Treated with Standard and Reduced Levels of Tacrolimus in Combination with EC-MPS. *J Transplant* 2012; **2012**: 941640 [PMID: 23227307 DOI: 10.1155/2012/94164]
- 16 **Vacher-Coponat H**, Moal V, Indreies M, Purgus R, Loundou A, Burtsey S, Brunet P, Moussi-Frances J, Daniel L, Dussol B, Berland Y. A randomized trial with steroids and antithymocyte globulins comparing cyclosporine/azathioprine versus tacrolimus/mycophenolate mofetil (CATM2) in renal transplantation. *Transplantation* 2012; **93**: 437-443 [PMID: 22228415 DOI: 10.1097/TP.0b013e31824215b]
- 17 **Tillmann FP**, Quack I, Schenk A, Grabensee B, Rump LC, Hetzel GR. Prevalence and risk factors of pre-diabetes after renal transplantation: a single-centre cohort study in 200 consecutive

- patients. *Nephrol Dial Transplant* 2012; **27**: 3330-3337 [PMID: 22492827 DOI: 10.1093/ndt/gfs02]
- 18 **Bonet J**, Martinez-Castelao A, Bayés B. Metabolic syndrome in hemodialysis patients as a risk factor for new-onset diabetes mellitus after renal transplant: a prospective observational study. *Diabetes Metab Syndr Obes* 2013; **6**: 339-346 [PMID: 24082792 DOI: 10.2147/DMSO.S51289]
 - 19 **Cole EH**, Prasad GV, Cardella CJ, Kim JS, Tinkam KJ, Cattran DC, Schiff JR, Landsberg DN, Zaltzman JS, Gill JS. A pilot study of reduced dose cyclosporine and corticosteroids to reduce new onset diabetes mellitus and acute rejection in kidney transplant recipients. *Transplant Res* 2013; **2**: 1 [PMID: 23369458 DOI: 10.1186/2047-1440-2-1]
 - 20 **Nagaraja P**, Ravindran V, Morris-Stiff G, Baboolal K. Role of insulin resistance indices in predicting new-onset diabetes after kidney transplantation. *Transpl Int* 2013; **26**: 273-280 [PMID: 23230898 DOI: 10.1111/tri.12026]
 - 21 **First MR**, Dhadda S, Croy R, Holman J, Fitzsimmons WE. New-onset diabetes after transplantation (NODAT): an evaluation of definitions in clinical trials. *Transplantation* 2013; **96**: 58-64 [PMID: 23619735 DOI: 10.1097/TP.0b013e318293fcf8]
 - 22 **Nagaraja P**, Sharif A, Ravindran V, Baboolal K. Long-term progression of abnormal glucose tolerance and its relationship with the metabolic syndrome after kidney transplantation. *Transplantation* 2014; **97**: 576-581 [PMID: 24398851 DOI: 10.1097/01.tp.0000438202.11971.2e]
 - 23 **Tokodai K**, Amada N, Haga I, Takayama T, Nakamura A. The 5-time point oral glucose tolerance test as a predictor of new-onset diabetes after kidney transplantation. *Diabetes Res Clin Pract* 2014; **103**: 298-303 [PMID: 24468096 DOI: 10.1016/j.diabres.2013.12.049]
 - 24 **Vicelli A**, Nguyen HT, Yong K, Chan D, Dogra G, Wong G, Lim WH. The impact of abnormal glucose regulation on arterial stiffness at 3 and 15 months after kidney transplantation. *Diabetol Metab Syndr* 2014; **6**: 52 [PMID: 24716893 DOI: 10.1186/1758-5996-6-52]
 - 25 **Weng LC**, Chiang YJ, Lin MH, Hsieh CY, Lin SC, Wei TY, Chou HF. Association between use of FK506 and prevalence of post-transplantation diabetes mellitus in kidney transplant patients. *Transplant Proc* 2014; **46**: 529-531 [PMID: 24656004 DOI: 10.1016/j.transproceed.2013.11.141]
 - 26 **Schweer T**, Gwinner W, Scheffner I, Schwarz A, Haller H, Blume C. High impact of rejection therapy on the incidence of post-transplant diabetes mellitus after kidney transplantation. *Clin Transplant* 2014; **28**: 512-519 [PMID: 24649873 DOI: 10.1111/ctr.1232]
 - 27 **Prasad N**, Gurjer D, Bhadauria D, Gupta A, Srivastava A, Kaul A, Jaiswal A, Yadav B, Yadav S, Sharma RK. Is basiliximab induction, a novel risk factor for new onset diabetes after transplantation for living donor renal allograft recipients? *Nephrology (Carlton)* 2014; **19**: 244-250 [PMID: 24447227 DOI: 10.1111/nep.12209]
 - 28 **Silva HT**, Yang HC, Meier-Kriesche HU, Croy R, Holman J, Fitzsimmons WE, First MR. Long-term follow-up of a phase III clinical trial comparing tacrolimus extended-release/MMF, tacrolimus/MMF, and cyclosporine/MMF in de novo kidney transplant recipients. *Transplantation* 2014; **97**: 636-641 [PMID: 24521771 DOI: 10.1097/01.TP.0000437669.93963.8E]
 - 29 **Lv C**, Chen M, Xu M, Xu G, Zhang Y, He S, Xue M, Gao J, Yu M, Gao X, Zhu T. Influencing factors of new-onset diabetes after a renal transplant and their effects on complications and survival rate. *PLoS One* 2014; **9**: e99406 [PMID: 24911157 DOI: 10.1371/journal.pone.0099406]
 - 30 **Bergrem HA**, Valderhaug TG, Hartmann A, Hjeltnes J, Leivestad T, Bergrem H, Jenssen T. Undiagnosed diabetes in kidney transplant candidates: a case-finding strategy. *Clin J Am Soc Nephrol* 2010; **5**: 616-622 [PMID: 20133490 DOI: 10.2215/CJN.07501009]
 - 31 **Caillard S**, Eprinchard L, Perrin P, Braun L, Heibel F, Moreau F, Kessler L, Moulin B. Incidence and risk factors of glucose metabolism disorders in kidney transplant recipients: role of systematic screening by oral glucose tolerance test. *Transplantation* 2011; **91**: 757-764 [PMID: 21336240 DOI: 10.1097/TP.0b013e31820f0877]
 - 32 **Iida S**, Ishida H, Tokumoto T, Omoto K, Shirakawa H, Shimizu T, Amano H, Setoguchi K, Nozaki T, Toki D, Tokita D, Tanabe K. New-onset diabetes after transplantation in tacrolimus-treated, living kidney transplantation: long-term impact and utility of the pre-transplant OGTT. *Int Urol Nephrol* 2010; **42**: 935-945 [PMID: 20169408 DOI: 10.1007/s11255-010-9712-0]
 - 33 **Hornum M**, Jørgensen KA, Hansen JM, Nielsen FT, Christensen KB, Mathiesen ER, Feldt-Rasmussen B. New-onset diabetes mellitus after kidney transplantation in Denmark. *Clin J Am Soc Nephrol* 2010; **5**: 709-716 [PMID: 20167685 DOI: 10.2215/CJN.05360709]
 - 34 Is fasting glucose sufficient to define diabetes? Epidemiological data from 20 European studies. The DECODE-study group. European Diabetes Epidemiology Group. *Diabetes Epidemiology: Collaborative analysis of Diagnostic Criteria in Europe. Diabetologia* 1999; **42**: 647-654 [PMID: 10382583]
 - 35 **Woodward RS**, Schnitzler MA, Baty J, Lowell JA, Lopez-Rocafort L, Haider S, Woodworth TG, Brennan DC. Incidence and cost of new onset diabetes mellitus among U.S. wait-listed and transplanted renal allograft recipients. *Am J Transplant* 2003; **3**: 590-598 [PMID: 12752315 DOI: 10.1034/j.1600-6143.2003.00082.x]
 - 36 **Tamaras SK**, Magliano DJ, Lynch B, Sethi P, Willenberg L, Polkinghorne KR, Chadban S, Dunstan D, Shaw JE. AusDiab 2012. The Australian Diabetes, Obesity and Lifestyle Study. Baker IDI Heart and Diabetes Institute 2013. Available from: URL: https://www.bakeridi.edu.au/Assets/Files/Baker%20IDI%20Ausdiab%20Rreport_interactive_FINAL.pdf
 - 37 **Chakkeri HA**, Weil EJ, Castro J, Heilman RL, Reddy KS, Mazur MJ, Hamawi K, Mulligan DC, Moss AA, Mekeel KL, Cosio FG, Cook CB. Hyperglycemia during the immediate period after kidney transplantation. *Clin J Am Soc Nephrol* 2009; **4**: 853-859 [PMID: 19339426 DOI: 10.2215/CJN.05471008]
 - 38 **Hecking M**, Haidinger M, Döller D, Werzowa J, Tura A, Zhang J, Tekoglu H, Pleiner J, Wrba T, Rasoul-Rockenschau S, Mühlbacher F, Schmaldienst S, Druml W, Hörl WH, Krebs M, Wolz M, Pacini G, Port FK, Säemann MD. Early basal insulin therapy decreases new-onset diabetes after renal transplantation. *J Am Soc Nephrol* 2012; **23**: 739-749 [PMID: 22343119 DOI: 10.1681/ASN.2011080835]
 - 39 **Federici M**, Hribal M, Perego L, Ranalli M, Caradonna Z, Perego C, Usellini L, Nano R, Bonini P, Bertuzzi F, Marlier LN, Davalli AM, Carandente O, Pontiroli AE, Melino G, Marchetti P, Lauro R, Sesti G, Folli F. High glucose causes apoptosis in cultured human pancreatic islets of Langerhans: a potential role for regulation of specific Bcl family genes toward an apoptotic cell death program. *Diabetes* 2001; **50**: 1290-1301 [PMID: 11375329 DOI: 10.2337/diabetes.50.6.1290]
 - 40 **Joss N**, Staatz CE, Thomson AH, Jardine AG. Predictors of new onset diabetes after renal transplantation. *Clin Transplant* 2007; **21**: 136-143 [PMID: 17302602 DOI: 10.1111/j.1399-0012.2006.00580.x]
 - 41 **Wojtuszczyzn A**, Mourad G, Bringer J, Renard E. Continuous glucose monitoring after kidney transplantation in non-diabetic patients: early hyperglycaemia is frequent and may herald post-transplantation diabetes mellitus and graft failure. *Diabetes Metab* 2013; **39**: 404-410 [PMID: 23999231 DOI: 10.1016/j.diabet.2012.10.007]
 - 42 **Chakkeri HA**, Knowler WC, Devarapalli Y, Weil EJ, Heilman RL, Dueck A, Mulligan DC, Reddy KS, Moss AA, Mekeel KL, Mazur MJ, Hamawi K, Castro JC, Cook CB. Relationship between inpatient hyperglycemia and insulin treatment after kidney transplantation and future new onset diabetes mellitus. *Clin J Am Soc Nephrol* 2010; **5**: 1669-1675 [PMID: 20558559 DOI: 10.2215/CJN.09481209]
 - 43 **Kuypers DR**, Claes K, Bammens B, Evenepoel P, Vanrenterghem Y. Early clinical assessment of glucose metabolism in renal allograft recipients: diagnosis and prediction of post-transplant diabetes mellitus (PTDM). *Nephrol Dial Transplant* 2008; **23**: 2033-2042 [PMID: 18174264 DOI: 10.1093/ndt/gfm875]
 - 44 **Cotovio P**, Neves M, Rodrigues L, Alves R, Bastos M, Baptista C, Macário F, Mota A. New-onset diabetes after transplantation: assessment of risk factors and clinical outcomes. *Transplant Proc* 2013; **45**: 1079-1083 [PMID: 23622631 DOI: 10.1016/j.transproceed.2013.11.141]

- d.2013.03.009]
- 45 **Valderhaug TG**, Jenssen T, Hartmann A, Midtvedt K, Holdaas H, Reisaeter AV, Hjelmesaeth J. Fasting plasma glucose and glycosylated hemoglobin in the screening for diabetes mellitus after renal transplantation. *Transplantation* 2009; **88**: 429-434 [PMID: 19667949 DOI: 10.1097/TP.0b013e3181af1f53]
 - 46 **Luan FL**, Steffick DE, Ojo AO. New-onset diabetes mellitus in kidney transplant recipients discharged on steroid-free immunosuppression. *Transplantation* 2011; **91**: 334-341 [PMID: 21242885 DOI: 10.1097/TP.0b013e318203c25f]
 - 47 **Tatar E**, Kircelli F, Demirci MS, Turan MN, Gungor O, Asci G, Ozkahya M, Ok E, Hoscoskun C, Toz H. Pre-transplant HbA1c level as an early marker for new-onset diabetes after renal transplantation. *Int Urol Nephrol* 2013; **45**: 251-258 [PMID: 23054321 DOI: 10.1007/s11255-012-0304-z]
 - 48 **Stevens RB**, Lane JT, Boerner BP, Miles CD, Rigley TH, Sandoz JP, Nielsen KJ, Skorupa JY, Skorupa AJ, Kaplan B, Wrenshall LE. Single-dose rATG induction at renal transplantation: superior renal function and glucose regulation with less hypomagnesemia. *Clin Transplant* 2012; **26**: 123-132 [PMID: 21401720 DOI: 10.1111/j.1399-0012.2011.01425.x]
 - 49 **Kasiske BL**, Snyder JJ, Gilbertson D, Matas AJ. Diabetes mellitus after kidney transplantation in the United States. *Am J Transplant* 2003; **3**: 178-185 [PMID: 12603213 DOI: 10.1034/j.1600-6143.2003.00010.x]
 - 50 **Shah T**, Kasravi A, Huang E, Hayashi R, Young B, Cho YW, Bunnapradist S. Risk factors for development of new-onset diabetes mellitus after kidney transplantation. *Transplantation* 2006; **82**: 1673-1676 [PMID: 17198258 DOI: 10.1097/01.tp.0000250756.66348.9a]
 - 51 **Johnston O**, Rose CL, Webster AC, Gill JS. Sirolimus is associated with new-onset diabetes in kidney transplant recipients. *J Am Soc Nephrol* 2008; **19**: 1411-1418 [PMID: 18385422 DOI: 10.1681/ASN.2007111202]
 - 52 **Cole EH**, Johnston O, Rose CL, Gill JS. Impact of acute rejection and new-onset diabetes on long-term transplant graft and patient survival. *Clin J Am Soc Nephrol* 2008; **3**: 814-821 [PMID: 18322046 DOI: 10.2215/CJN.04681107]
 - 53 **Ghisdal L**, Baron C, Le Meur Y, Lionet A, Halimi JM, Rerolle JP, Glowacki F, Lebranchu Y, Drouet M, Noël C, El Housni H, Cochaux P, Wissing KM, Abramowicz D, Abramowicz M. TCF7L2 polymorphism associates with new-onset diabetes after transplantation. *J Am Soc Nephrol* 2009; **20**: 2459-2467 [PMID: 19713311 DOI: 10.1681/ASN.2008121314]
 - 54 **McCaughan JA**, McKnight AJ, Maxwell AP. Genetics of new-onset diabetes after transplantation. *J Am Soc Nephrol* 2014; **25**: 1037-1049 [PMID: 24309190 DOI: 10.1681/ASN.2013040383]
 - 55 **de Mattos AM**, Olyaei AJ, Prather JC, Golconda MS, Barry JM, Norman DJ. Autosomal-dominant polycystic kidney disease as a risk factor for diabetes mellitus following renal transplantation. *Kidney Int* 2005; **67**: 714-720 [PMID: 15673321 DOI: 10.1111/j.1523-1755.2005.67132.x]
 - 56 **Hamer RA**, Chow CL, Ong AC, McKane WS. Polycystic kidney disease is a risk factor for new-onset diabetes after transplantation. *Transplantation* 2007; **83**: 36-40 [PMID: 17220788 DOI: 10.1097/01.tp.0000248759.37146.3d]
 - 57 **Ruderman I**, Masterson R, Yates C, Gorelik A, Cohny SJ, Walker RG. New onset diabetes after kidney transplantation in autosomal dominant polycystic kidney disease: a retrospective cohort study. *Nephrology (Carlton)* 2012; **17**: 89-96 [PMID: 21854501 DOI: 10.1111/j.1440-1797.2011.01507.x]
 - 58 **Aasebø W**, Midtvedt K, Valderhaug TG, Leivestad T, Hartmann A, Reisaeter AV, Jenssen T, Holdaas H. Impaired glucose homeostasis in renal transplant recipients receiving basiliximab. *Nephrol Dial Transplant* 2010; **25**: 1289-1293 [PMID: 19934089 DOI: 10.1093/ndt/gfp617]
 - 59 **Hjelmesaeth J**, Sagedal S, Hartmann A, Rollag H, Egeland T, Hagen M, Nordal KP, Jenssen T. Asymptomatic cytomegalovirus infection is associated with increased risk of new-onset diabetes mellitus and impaired insulin release after renal transplantation. *Diabetologia* 2004; **47**: 1550-1556 [PMID: 15338129 DOI: 10.1007/s00125-004-1499-z]
 - 60 **Suszynski TM**, Gillingham KJ, Rizzari MD, Dunn TB, Payne WD, Chinnakotla S, Finger EB, Sutherland DE, Najarian JS, Pruett TL, Matas AJ, Kandaswamy R. Prospective randomized trial of maintenance immunosuppression with rapid discontinuation of prednisone in adult kidney transplantation. *Am J Transplant* 2013; **13**: 961-970 [PMID: 23432755 DOI: 10.1111/ajt.12166]
 - 61 **Bora GS**, Guleria S, Reddy VS, Tandon N, Gupta N, Gupta S, Bhowmik D. Risk factors for the development of new-onset diabetes mellitus in a living related renal transplant program. *Transplant Proc* 2010; **42**: 4072-4073 [PMID: 21168630 DOI: 10.1016/j.transproceed.2010.10.008]
 - 62 **Santos L**, Rodrigo E, Piñera C, Quintella E, Ruiz JC, Fernández-Fresnedo G, Palomar R, Gómez-Alamillo C, de Francisco A, Arias M. New-onset diabetes after transplantation: drug-related risk factors. *Transplant Proc* 2012; **44**: 2585-2587 [PMID: 23146462 DOI: 10.1016/j.transproceed.2012.09.053]
 - 63 **Ghisdal L**, Van Laecke S, Abramowicz MJ, Vanholder R, Abramowicz D. New-onset diabetes after renal transplantation: risk assessment and management. *Diabetes Care* 2012; **35**: 181-188 [PMID: 22187441 DOI: 10.2337/dc11-1230]
 - 64 **Kurzawski M**, Dziewanowski K, Lapczuk J, Wajda A, Drożdżki M. Analysis of common type 2 diabetes mellitus genetic risk factors in new-onset diabetes after transplantation in kidney transplant patients medicated with tacrolimus. *Eur J Clin Pharmacol* 2012; **68**: 1587-1594 [PMID: 22569928 DOI: 10.1007/s00228-012-1292-8]
 - 65 **Yao B**, Chen X, Shen FX, Xu W, Dong TT, Chen LZ, Weng JP. The incidence of posttransplantation diabetes mellitus during follow-up in kidney transplant recipients and relationship to FokI vitamin D receptor polymorphism. *Transplant Proc* 2013; **45**: 194-196 [PMID: 23375298 DOI: 10.1016/j.transproceed.2012.08.019]
 - 66 **Nicoletto BB**, Souza GC, Fonseca NK, Centenaro A, Manfro RC, Canani LH, Gonçalves LF. Association between 276G/T adiponectin gene polymorphism and new-onset diabetes after kidney transplantation. *Transplantation* 2013; **96**: 1059-1064 [PMID: 23985723 DOI: 10.1097/TP.0b013e3182a45283]
 - 67 **Tavira B**, Coto E, Torres A, Díaz-Corte C, Díaz-Molina B, Ortega F, Arias M, Díaz JM, Selgas R, López-Larrea C, Ruiz-Ortega M, Ortiz A, González E, Campistol JM, Alvarez V. Association between a common KCNJ11 polymorphism (rs5219) and new-onset posttransplant diabetes in patients treated with Tacrolimus. *Mol Genet Metab* 2012; **105**: 525-527 [PMID: 22264780 DOI: 10.1016/j.ymgme.2011.12.020]
 - 68 **Boots JM**, Christiaans MH, Van Duijnhoven EM, Van Suylen RJ, Van Hooff JP. Early steroid withdrawal in renal transplantation with tacrolimus dual therapy: a pilot study. *Transplantation* 2002; **74**: 1703-1709 [PMID: 12499885 DOI: 10.1097/01.TP.0000040083.08795.66]
 - 69 **Rizzari MD**, Suszynski TM, Gillingham KJ, Dunn TB, Ibrahim HN, Payne WD, Chinnakotla S, Finger EB, Sutherland DE, Kandaswamy R, Najarian JS, Pruett TL, Kukla A, Spong R, Matas AJ. Ten-year outcome after rapid discontinuation of prednisone in adult primary kidney transplantation. *Clin J Am Soc Nephrol* 2012; **7**: 494-503 [PMID: 22282482 DOI: 10.2215/CJN.08630811]
 - 70 **Baid-Agrawal S**, Frei U, Reinke P, Schindler R, Kopp MA, Martus P, Berg T, Juergensen JS, Anker SD, Doehner W. Impaired insulin sensitivity as an underlying mechanism linking hepatitis C and posttransplant diabetes mellitus in kidney recipients. *Am J Transplant* 2009; **9**: 2777-2784 [PMID: 19845589 DOI: 10.1111/j.1600-6143.2009.02843.x]
 - 71 **Ivarsson KM**, Clyne N, Almquist M, Akaberi S. Hyperparathyroidism and new onset diabetes after renal transplantation. *Transplant Proc* 2014; **46**: 145-150 [PMID: 24507041 DOI: 10.1016/j.transproceed.2013.07.076]
 - 72 **Garg N**, Weinberg J, Ghai S, Bradauskaitė G, Nuhn M, Gautam A, Kumar N, Francis J, Chen JL. Lower magnesium level associated with new-onset diabetes and pre-diabetes after kidney transplantation. *J Nephrol* 2014; **27**: 339-344 [PMID: 24609888 DOI: 10.1007/s40620-014-0072-1]

- 73 **Augusto JF**, Subra JF, Duveau A, Rakotonjanahary J, Dussaussoy C, Picquet J, Croue A, Villemain F, Onno C, Sayegh J. Relation between pretransplant magnesemia and the risk of new onset diabetes after transplantation within the first year of kidney transplantation. *Transplantation* 2014; **97**: 1155-1160 [PMID: 24686469 DOI: 10.1097/01.TP.0000440950.22133.a1]
- 74 **Teutonico A**, Schena PF, Di Paolo S. Glucose metabolism in renal transplant recipients: effect of calcineurin inhibitor withdrawal and conversion to sirolimus. *J Am Soc Nephrol* 2005; **16**: 3128-3135 [PMID: 16107580 DOI: 10.1681/ASN.2005050487]
- 75 **Ekberg H**, Tedesco-Silva H, Demirbas A, Vitko S, Nashan B, Gürkan A, Margreiter R, Hugo C, Grinyó JM, Frei U, Vanrenterghem Y, Daloz P, Halloran PF. Reduced exposure to calcineurin inhibitors in renal transplantation. *N Engl J Med* 2007; **357**: 2562-2575 [PMID: 18094377 DOI: 10.1056/NEJMoa067411]
- 76 **Guerra G**, Ciancio G, Gaynor JJ, Zarak A, Brown R, Hanson L, Sageshima J, Roth D, Chen L, Kupin W, Tueros L, Ruiz P, Livingstone AS, Burke GW. Randomized trial of immunosuppressive regimens in renal transplantation. *J Am Soc Nephrol* 2011; **22**: 1758-1768 [PMID: 21807891 DOI: 10.1681/ASN.2011010006]
- 77 **Gyurus E**, Kaposztas Z, Kahan BD. Sirolimus therapy predisposes to new-onset diabetes mellitus after renal transplantation: a long-term analysis of various treatment regimens. *Transplant Proc* 2011; **43**: 1583-1592 [PMID: 21693238 DOI: 10.1016/j.transproceed.2011.05.001]
- 78 **Veroux M**, Tallarita T, Corona D, Sinagra N, Giaquinta A, Zerbo D, Guerrieri C, D'Assoro A, Cimino S, Veroux P. Conversion to sirolimus therapy in kidney transplant recipients with new onset diabetes mellitus after transplantation. *Clin Dev Immunol* 2013; **2013**: 496974 [PMID: 23762090 DOI: 10.1155/2013/496974]
- 79 **Boudreaux JP**, McHugh L, Canafax DM, Ascher N, Sutherland DE, Payne W, Simmons RL, Najarian JS, Fryd DS. The impact of cyclosporine and combination immunosuppression on the incidence of posttransplant diabetes in renal allograft recipients. *Transplantation* 1987; **44**: 376-381 [PMID: 3307061 DOI: 10.1097/00007890-198709000-00010]
- 80 **Bending JJ**, Ogg CS, Viberti GC. Diabetogenic effect of cyclosporin. *Br Med J (Clin Res Ed)* 1987; **294**: 401-402 [PMID: 3101896 DOI: 10.1136/bmj.294.6569.401]
- 81 **Scantlebury V**, Shapiro R, Fung J, Tzakis A, McCauley J, Jordan M, Jensen C, Hakala T, Simmons R, Starzl TE. New onset of diabetes in FK 506 vs cyclosporine-treated kidney transplant recipients. *Transplant Proc* 1991; **23**: 3169-3170 [PMID: 1721395]
- 82 **Mayer AD**, Dmitrewski J, Squifflet JP, Besse T, Grabensee B, Klein B, Eigler FW, Heemann U, Pichlmayr R, Behrend M, Vanrenterghem Y, Donck J, van Hooff J, Christiaans M, Morales JM, Andres A, Johnson RW, Short C, Buchholz B, Rehmert N, Land W, Schleibner S, Forsythe JL, Talbot D, Pohanka E. Multicenter randomized trial comparing tacrolimus (FK506) and cyclosporine in the prevention of renal allograft rejection: a report of the European Tacrolimus Multicenter Renal Study Group. *Transplantation* 1997; **64**: 436-443 [PMID: 9275110 DOI: 10.1097/00007890-199708150-00012]
- 83 **Webster A**, Woodroffe RC, Taylor RS, Chapman JR, Craig JC. Tacrolimus versus cyclosporin as primary immunosuppression for kidney transplant recipients. *Cochrane Database Syst Rev* 2005; **(4)**: CD003961 [PMID: 16235347]
- 84 **Claes K**, Meier-Kriesche HU, Schold JD, Vanrenterghem Y, Halloran PF, Ekberg H. Effect of different immunosuppressive regimens on the evolution of distinct metabolic parameters: evidence from the Symphony study. *Nephrol Dial Transplant* 2012; **27**: 850-857 [PMID: 21617197 DOI: 10.1093/ndt/gfr238]
- 85 **Woodle ES**, First MR, Pirsch J, Shihab F, Gaber AO, Van Veldhuisen P. A prospective, randomized, double-blind, placebo-controlled multicenter trial comparing early (7 day) corticosteroid cessation versus long-term, low-dose corticosteroid therapy. *Ann Surg* 2008; **248**: 564-577 [PMID: 18936569 DOI: 10.1097/SLA.0b013e318187d1da]
- 86 **Pascual J**, Royuela A, Galeano C, Crespo M, Zamora J. Very early steroid withdrawal or complete avoidance for kidney transplant recipients: a systematic review. *Nephrol Dial Transplant* 2012; **27**: 825-832 [PMID: 21785040 DOI: 10.1093/ndt/gfr374]
- 87 **Barracough KA**, Landsberg DN, Shapiro RJ, Gill JS, Li G, Balshaw RF, Chailimpamontree W, Keown PA. A matched cohort pharmacoepidemiological analysis of steroid free immunosuppression in renal transplantation. *Transplantation* 2009; **87**: 672-680 [PMID: 19295311 DOI: 10.1097/TP.0b013e318195aa54]
- 88 **Ekberg J**, Ekberg H, Jespersen B, Källen R, Skov K, Olausson M, Mjörnstedt L, Lindnér P. An in-progress, open-label, multi-centre study (SAILOR) evaluating whether a steroid-free immunosuppressive protocol, based on ATG induction and a low tacrolimus dose, reduces the incidence of new onset diabetes after transplantation. *Transplant Res* 2014; **3**: 12 [PMID: 24959347 DOI: 10.1186/2047-1440-3-12]
- 89 **Prasad N**, Gupta P, Jain M, Bhadauria D, Gupta A, Sharma RK, Kaul A. Outcomes of de novo allograft diabetic nephropathy in renal allograft recipients. *Exp Clin Transplant* 2013; **11**: 215-221 [PMID: 23767942 DOI: 10.6002/ect.2012.0193]
- 90 **Koselj M**, Rott T, Koselj MK, Hvala A, Arnol M, Kandus A. De novo diabetic nephropathy on renal allografts. *Transplant Proc* 2003; **35**: 2919-2921 [PMID: 14697938]
- 91 **Rabkin R**, Ryan MP, Duckworth WC. The renal metabolism of insulin. *Diabetologia* 1984; **27**: 351-357 [PMID: 6389240]
- 92 **Bergman RN**, Ader M, Huecking K, Van Citters G. Accurate assessment of beta-cell function: the hyperbolic correction. *Diabetes* 2002; **51** Suppl 1: S212-S220 [PMID: 11815482 DOI: 10.2337/diabetes.51.2007.S212]
- 93 **Delaunay F**, Khan A, Cintra A, Davani B, Ling ZC, Andersson A, Ostenson CG, Gustafsson J, Efendic S, Okret S. Pancreatic beta cells are important targets for the diabetogenic effects of glucocorticoids. *J Clin Invest* 1997; **100**: 2094-2098 [PMID: 9329975 DOI: 10.1172/JCI119743]
- 94 **van Raalte DH**, Brands M, van der Zijl NJ, Muskiet MH, Pouwels PJ, Ackermans MT, Sauerwein HP, Serlie MJ, Diamant M. Low-dose glucocorticoid treatment affects multiple aspects of intermediary metabolism in healthy humans: a randomised controlled trial. *Diabetologia* 2011; **54**: 2103-2112 [PMID: 21562755 DOI: 10.1007/s00125-011-2174-9]
- 95 **Hwang JL**, Weiss RE. Steroid-induced diabetes: a clinical and molecular approach to understanding and treatment. *Diabetes Metab Res Rev* 2014; **30**: 96-102 [PMID: 24123849 DOI: 10.1002/dmrr.2486]
- 96 **Koppe L**, Pelletier CC, Alix PM, Kalbacher E, Fouque D, Soulage CO, Guebre-Egziabher F. Insulin resistance in chronic kidney disease: new lessons from experimental models. *Nephrol Dial Transplant* 2014; **29**: 1666-1674 [PMID: 24286973 DOI: 10.1093/ndt/gft435]
- 97 **Jia T**, Huang X, Qureshi AR, Xu H, Årnlov J, Lindholm B, Cederholm T, Stenvinkel P, Risérus U, Carrero JJ. Validation of insulin sensitivity surrogate indices and prediction of clinical outcomes in individuals with and without impaired renal function. *Kidney Int* 2014; **86**: 383-391 [PMID: 24476695 DOI: 10.1038/ki.2014.1]
- 98 **Bodlaj G**, Berg J, Pichler R, Biesenbach G. Prevalence, severity and predictors of HOMA-estimated insulin resistance in diabetic and nondiabetic patients with end-stage renal disease. *J Nephrol* 2006; **19**: 607-612 [PMID: 17136689]
- 99 **Uchida J**, Iwai T, Machida Y, Kuwabara N, Kabei K, Kumada N, Nakatani T. Insulin resistance and insulin secretion in renal transplant recipients with hepatitis C. *Transplant Proc* 2013; **45**: 1540-1543 [PMID: 23726615 DOI: 10.1016/j.transproceed.2013.01.053]
- 100 **Zelle DM**, Corpeleijn E, Deinum J, Stolk RP, Gans RO, Navis G, Bakker SJ. Pancreatic β -cell dysfunction and risk of new-onset diabetes after kidney transplantation. *Diabetes Care* 2013; **36**: 1926-1932 [PMID: 23378624 DOI: 10.2337/dc12-1894]
- 101 **Hjelmessaeth J**, Hagen M, Hartmann A, Midtvedt K, Egeland T, Jenssen T. The impact of impaired insulin release and insulin resistance on glucose intolerance after renal transplantation. *Clin*

- Transplant* 2002; **16**: 389-396 [PMID: 12437616]
- 102 **Nam JH**, Mun JI, Kim SI, Kang SW, Choi KH, Park K, Ahn CW, Cha BS, Song YD, Lim SK, Kim KR, Lee HC, Huh KB. beta-Cell dysfunction rather than insulin resistance is the main contributing factor for the development of postrenal transplantation diabetes mellitus. *Transplantation* 2001; **71**: 1417-1423 [PMID: 11391229]
 - 103 **Hagen M**, Hjelmestaeth J, Jenssen T, Morkrid L, Hartmann A. A 6-year prospective study on new onset diabetes mellitus, insulin release and insulin sensitivity in renal transplant recipients. *Nephrol Dial Transplant* 2003; **18**: 2154-2159 [PMID: 13679495 DOI: 10.1093/ndt/gfg338]
 - 104 **Heit JJ**, Apelqvist AA, Gu X, Winslow MM, Neilson JR, Crabtree GR, Kim SK. Calcineurin/NFAT signalling regulates pancreatic beta-cell growth and function. *Nature* 2006; **443**: 345-349 [PMID: 16988714 DOI: 10.1038/nature05097]
 - 105 **Soleimanpour SA**, Crutchlow MF, Ferrari AM, Raum JC, Groff DN, Rankin MM, Liu C, De León DD, Naji A, Kushner JA, Stoffers DA. Calcineurin signaling regulates human islet {beta}-cell survival. *J Biol Chem* 2010; **285**: 40050-40059 [PMID: 20943662 DOI: 10.1074/jbc.M110.154955]
 - 106 **Duijnhoven EM**, Boots JM, Christiaans MH, Wolffenbuttel BH, Van Hooff JP. Influence of tacrolimus on glucose metabolism before and after renal transplantation: a prospective study. *J Am Soc Nephrol* 2001; **12**: 583-588 [PMID: 11181807]
 - 107 **Hansen KB**, Vilsbøll T, Bagger JI, Holst JJ, Knop FK. Impaired incretin-induced amplification of insulin secretion after glucose homeostatic dysregulation in healthy subjects. *J Clin Endocrinol Metab* 2012; **97**: 1363-1370 [PMID: 22319034 DOI: 10.1210/jc.2011-2594]
 - 108 **Idorn T**, Knop FK, Jørgensen M, Holst JJ, Hornum M, Feldt-Rasmussen B. Gastrointestinal factors contribute to glucometabolic disturbances in nondiabetic patients with end-stage renal disease. *Kidney Int* 2013; **83**: 915-923 [PMID: 23325073 DOI: 10.1038/ki.2012.460]
 - 109 **Idorn T**, Knop FK, Jørgensen M, Holst JJ, Hornum M, Feldt-Rasmussen B. Postprandial responses of incretin and pancreatic hormones in non-diabetic patients with end-stage renal disease. *Nephrol Dial Transplant* 2014; **29**: 119-127 [PMID: 24078334 DOI: 10.1093/ndt/gft353]
 - 110 **Lentine KL**, Brennan DC, Schnitzler MA. Incidence and predictors of myocardial infarction after kidney transplantation. *J Am Soc Nephrol* 2005; **16**: 496-506 [PMID: 15615820 DOI: 10.1681/ASN.2004070580]
 - 111 **Opelz G**, Döhler B. Cardiovascular death in kidney recipients treated with renin-angiotensin system blockers. *Transplantation* 2014; **97**: 310-315 [PMID: 24492421 DOI: 10.1097/01.TP.0000437672.78716.28]
 - 112 **Valderhaug TG**, Hjelmestaeth J, Jenssen T, Røislien J, Leivestad T, Hartmann A. Early posttransplantation hyperglycemia in kidney transplant recipients is associated with overall long-term graft losses. *Transplantation* 2012; **94**: 714-720 [PMID: 22965263 DOI: 10.1097/TP.0b013e31825f4434]
 - 113 **Zoungas S**, Chalmers J, Neal B, Billot L, Li Q, Hirakawa Y, Arima H, Monaghan H, Joshi R, Colagiuri S, Cooper ME, Glasziou P, Grobbee D, Hamet P, Harrap S, Heller S, Lisheng L, Mancia G, Marre M, Matthews DR, Mogensen CE, Perkovic V, Poulter N, Rodgers A, Williams B, MacMahon S, Patel A, Woodward M. Follow-up of blood-pressure lowering and glucose control in type 2 diabetes. *N Engl J Med* 2014; **371**: 1392-1406 [PMID: 25234206 DOI: 10.1056/NEJMoa1407963]
 - 114 **Holman RR**, Paul SK, Bethel MA, Matthews DR, Neil HA. 10-year follow-up of intensive glucose control in type 2 diabetes. *N Engl J Med* 2008; **359**: 1577-1589 [PMID: 18784090 DOI: 10.1056/NEJMoa0806470]
 - 115 **Shabir S**, Jham S, Harper L, Ball S, Borrows R, Sharif A. Validity of glycated haemoglobin to diagnose new onset diabetes after transplantation. *Transpl Int* 2013; **26**: 315-321 [PMID: 23279163 DOI: 10.1111/tri.12042]
 - 116 **Eide IA**, Halden TA, Hartmann A, Åsberg A, Dahle DO, Reisæter AV, Jenssen T. Limitations of hemoglobin A1c for the diagnosis of posttransplant diabetes mellitus. *Transplantation* 2015; **99**: 629-635 [PMID: 25162478 DOI: 10.1097/TP.0000000000000376]
 - 117 **Yates CJ**, Furlanos S, Colman PG, Cohnsey SJ. Screening for new-onset diabetes after kidney transplantation: limitations of fasting glucose and advantages of afternoon glucose and glycated hemoglobin. *Transplantation* 2013; **96**: 726-731 [PMID: 23902993 DOI: 10.1097/TP.0b013e3182a012f3]
 - 118 **Rodrigo E**, Santos L, Piñera C, Millán JC, Quintela ME, Toyos C, Allende N, Gómez-Alamillo C, Arias M. Prediction at first year of incident new-onset diabetes after kidney transplantation by risk prediction models. *Diabetes Care* 2012; **35**: 471-473 [PMID: 22279030 DOI: 10.2337/dc11-2071]
 - 119 **Hayashi T**, Boyko EJ, Sato KK, McNeely MJ, Leonetti DL, Kahn SE, Fujimoto WY. Patterns of insulin concentration during the OGTT predict the risk of type 2 diabetes in Japanese Americans. *Diabetes Care* 2013; **36**: 1229-1235 [PMID: 23275353 DOI: 10.2337/dc12-0246]
 - 120 **Chakkeri HA**, Weil EJ, Swanson CM, Dueck AC, Heilman RL, Reddy KS, Hamawi K, Khamash H, Moss AA, Mulligan DC, Katariya N, Knowler WC. Pretransplant risk score for new-onset diabetes after kidney transplantation. *Diabetes Care* 2011; **34**: 2141-2145 [PMID: 21949218 DOI: 10.2337/dc11-0752]
 - 121 **Chakkeri HA**, Chang YH, Ayub A, Gonwa TA, Weil EJ, Knowler WC. Validation of a pretransplant risk score for new-onset diabetes after kidney transplantation. *Diabetes Care* 2013; **36**: 2881-2886 [PMID: 24009296 DOI: 10.2337/dc13-0428]
 - 122 **Utzschneider KM**, Prigeon RL, Faulenbach MV, Tong J, Carr DB, Boyko EJ, Leonetti DL, McNeely MJ, Fujimoto WY, Kahn SE. Oral disposition index predicts the development of future diabetes above and beyond fasting and 2-h glucose levels. *Diabetes Care* 2009; **32**: 335-341 [PMID: 18957530 DOI: 10.2337/dc08-1478]
 - 123 **Defronzo RA**, Tripathy D, Schwenke DC, Banerji M, Bray GA, Buchanan TA, Clement SC, Henry RR, Kitabchi AE, Mudaliar S, Ratner RE, Stentz FB, Musi N, Reaven PD, Gastaldelli A. Prediction of diabetes based on baseline metabolic characteristics in individuals at high risk. *Diabetes Care* 2013; **36**: 3607-3612 [PMID: 24062330 DOI: 10.2337/dc13-0520]
 - 124 **Tokodai K**, Amada N, Haga I, Takayama T, Nakamura A, Kashiwada T. Insulin resistance as a risk factor for new-onset diabetes after kidney transplantation. *Transplant Proc* 2014; **46**: 537-539 [PMID: 24656006 DOI: 10.1016/j.transproceed.2013.10.060]
 - 125 **Hornum M**, Lindahl JP, von Zur-Mühlen B, Jenssen T, Feldt-Rasmussen B. Diagnosis, management and treatment of glucometabolic disorders emerging after kidney transplantation: a position statement from the Nordic Transplantation Societies. *Transpl Int* 2013; **26**: 1049-1060 [PMID: 23634804 DOI: 10.1111/tri.12112]
 - 126 **Kidney Disease: Improving Global Outcomes (KDIGO) Transplant Work Group**. KDIGO clinical practice guideline for the care of kidney transplant recipients. *Am J Transplant* 2009; **9** Suppl 3: S1-S155 [PMID: 19845597 DOI: 10.1111/j.1600-6143.2009.02834.x]
 - 127 **Rostaing L**, Vincenti F, Grinyó J, Rice KM, Bresnahan B, Steinberg S, Gang S, Gaité LE, Moal MC, Mondragón-Ramírez GA, Kothari J, Pupim L, Larsen CP. Long-term belatacept exposure maintains efficacy and safety at 5 years: results from the long-term extension of the BENEFIT study. *Am J Transplant* 2013; **13**: 2875-2883 [PMID: 24047110 DOI: 10.1111/ajt.12460]
 - 128 **Charpentier B**, Medina Pestana JO, Del C Rial M, Rostaing L, Grinyó J, Vanrenterghem Y, Matas A, Zhang R, Mühlbacher F, Pupim L, Florman S. Long-term exposure to belatacept in recipients of extended criteria donor kidneys. *Am J Transplant* 2013; **13**: 2884-2891 [PMID: 24103072 DOI: 10.1111/ajt.12459]
 - 129 **Vincenti F**, Charpentier B, Vanrenterghem Y, Rostaing L, Bresnahan B, Darji P, Massari P, Mondragon-Ramirez GA, Agarwal M, Di Russo G, Lin CS, Garg P, Larsen CP. A phase III study of belatacept-based immunosuppression regimens versus cyclosporine in renal transplant recipients (BENEFIT study). *Am J Transplant* 2010; **10**: 535-546 [PMID: 20415897 DOI: 10.1111/

- j.1600-6143.2009.03005.x]
- 130 **Durrbach A**, Pestana JM, Pearson T, Vincenti F, Garcia VD, Campistol J, Rial Mdel C, Florman S, Block A, Di Russo G, Xing J, Garg P, Grinyó J. A phase III study of belatacept versus cyclosporine in kidney transplants from extended criteria donors (BENEFIT-EXT study). *Am J Transplant* 2010; **10**: 547-557 [PMID: 20415898 DOI: 10.1111/j.1600-6143.2010.03016.x]
- 131 **Vanrenterghem Y**, Bresnahan B, Campistol J, Durrbach A, Grinyó J, Neumayer HH, Lang P, Larsen CP, Mancilla-Urrea E, Pestana JM, Block A, Duan T, Glicklich A, Gujrathi S, Vincenti F. Belatacept-based regimens are associated with improved cardiovascular and metabolic risk factors compared with cyclosporine in kidney transplant recipients (BENEFIT and BENEFIT-EXT studies). *Transplantation* 2011; **91**: 976-983 [PMID: 21372756 DOI: 10.1097/TP.0b013e31820c10eb]
- 132 **Zelle DM**, Corpeleijn E, Stolk RP, de Greef MH, Gans RO, van der Heide JJ, Navis G, Bakker SJ. Low physical activity and risk of cardiovascular and all-cause mortality in renal transplant recipients. *Clin J Am Soc Nephrol* 2011; **6**: 898-905 [PMID: 21372213 DOI: 10.2215/CJN.03340410]
- 133 **Sharif A**, Moore R, Baboolal K. Influence of lifestyle modification in renal transplant recipients with postprandial hyperglycemia. *Transplantation* 2008; **85**: 353-358 [PMID: 18301331 DOI: 10.1097/TP.0b013e3181605ebf]
- 134 **Worzowa J**, Hecking M, Haidinger M, Lechner F, Döller D, Pacini G, Stemer G, Pleiner J, Frantal S, Säemann MD. Vildagliptin and pioglitazone in patients with impaired glucose tolerance after kidney transplantation: a randomized, placebo-controlled clinical trial. *Transplantation* 2013; **95**: 456-462 [PMID: 23380864 DOI: 10.1097/TP.0b013e318276a20e]
- 135 **Haidinger M**, Worzowa J, Hecking M, Antlanger M, Stemer G, Pleiner J, Kopecky C, Kovarik JJ, Döller D, Pacini G, Säemann MD. Efficacy and safety of vildagliptin in new-onset diabetes after kidney transplantation--a randomized, double-blind, placebo-controlled trial. *Am J Transplant* 2014; **14**: 115-123 [PMID: 24279801 DOI: 10.1111/ajt.12518]
- 136 **Brown NJ**, Byiers S, Carr D, Maldonado M, Warner BA. Dipeptidyl peptidase-IV inhibitor use associated with increased risk of ACE inhibitor-associated angioedema. *Hypertension* 2009; **54**: 516-523 [PMID: 19581505 DOI: 10.1161/HYPERTENSIONAHA.109.134197]
- 137 **Strøm Halden TA**, Åsberg A, Vik K, Hartmann A, Jenssen T. Short-term efficacy and safety of sitagliptin treatment in long-term stable renal recipients with new-onset diabetes after transplantation. *Nephrol Dial Transplant* 2014; **29**: 926-933 [PMID: 24452849 DOI: 10.1093/ndt/gft536]
- 138 **Karagiannis T**, Boura P, Tsapas A. Safety of dipeptidyl peptidase 4 inhibitors: a perspective review. *Ther Adv Drug Saf* 2014; **5**: 138-146 [PMID: 25083269 DOI: 10.1177/2042098614523031]
- 139 **Sharif A**. Should metformin be our antiglycemic agent of choice post-transplantation? *Am J Transplant* 2011; **11**: 1376-1381 [PMID: 21564529 DOI: 10.1111/j.1600-6143.2011.03550.x]
- 140 **Hecking M**, Worzowa J, Haidinger M, Hörl WH, Pascual J, Budde K, Luan FL, Ojo A, de Vries AP, Porrini E, Pacini G, Port FK, Sharif A, Säemann MD. Novel views on new-onset diabetes after transplantation: development, prevention and treatment. *Nephrol Dial Transplant* 2013; **28**: 550-566 [PMID: 23328712 DOI: 10.1093/ndt/gfs583]

P- Reviewer: Das UN, Sasaoka T

S- Editor: Tian YL **L- Editor:** A **E- Editor:** Jiao XK



Magnesium and type 2 diabetes

Mario Barbagallo, Ligia J Dominguez

Mario Barbagallo, Ligia J Dominguez, Geriatric Unit, Department of Internal Medicine and Medical Specialties, University of Palermo, 90127 Palermo, Italy

Author contributions: Barbagallo M and Dominguez LJ were responsible for the initial plan, study design, conducting the review, interpretation, manuscript drafting, critical revision of intellectual content, and approval of the version to be published.

Conflict-of-interest statement: None of the authors has a conflict of interest to report.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Mario Barbagallo, MD, PhD, Geriatric Unit, Department of Internal Medicine and Medical Specialties, University of Palermo, Via del Vespro 129, 90127 Palermo, Italy. mario.barbagallo@unipa.it
Telephone: +39-91-6552885
Fax: +39-91-6552952

Received: April 15, 2015

Peer-review started: April 18, 2015

First decision: May 13, 2015

Revised: June 29, 2015

Accepted: August 16, 2015

Article in press: August 17, 2015

Published online: August 25, 2015

Abstract

Type 2 diabetes is frequently associated with both extracellular and intracellular magnesium (Mg) deficits. A chronic latent Mg deficit or an overt clinical hypomagnesemia is common in patients with type 2 diabetes, especially in those with poorly controlled glycemic profiles. Insulin

and glucose are important regulators of Mg metabolism. Intracellular Mg plays a key role in regulating insulin action, insulin-mediated-glucose-uptake and vascular tone. Reduced intracellular Mg concentrations result in a defective tyrosine-kinase activity, postreceptorial impairment in insulin action and worsening of insulin resistance in diabetic patients. A low Mg intake and an increased Mg urinary loss appear the most important mechanisms that may favor Mg depletion in patients with type 2 diabetes. Low dietary Mg intake has been related to the development of type 2 diabetes and metabolic syndrome. Benefits of Mg supplementation on metabolic profiles in diabetic patients have been found in most, but not all clinical studies and larger prospective studies are needed to support the potential role of dietary Mg supplementation as a possible public health strategy in diabetes risk. The aim of this review is to revise current evidence on the mechanisms of Mg deficiency in diabetes and on the possible role of Mg supplementation in the prevention and management of the disease.

Key words: Magnesium; Type 2 diabetes; Metabolic syndrome; Inflammation; Aging; Hypertension; Insulin resistance; Endothelium

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Diabetes is frequently associated with Mg deficit. The fact that most but not all diabetic subjects have low magnesium (Mg) and that no large randomised controlled trial (RCT) has been specifically focused on subjects with Mg deficit, diagnosed with a reliable technique, may help explain discrepancies of the role of supplemental Mg on glycemic control, and the impact on diabetes risk in prospective epidemiological studies. Different baseline Mg, metabolic control, and age are other potential factors that may contribute. Future prospective RCTs are needed to support the potential role of dietary Mg supplementation as a possible public health strategy to reduce diabetes risk in the population.

Barbagallo M, Dominguez LJ. Magnesium and type 2 diabetes. *World J Diabetes* 2015; 6(10): 1152-1157 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v6/i10/1152.htm> DOI: <http://dx.doi.org/10.4239/wjd.v6.i10.1152>

INTRODUCTION

Magnesium (Mg) is an electrolyte of chief physiological importance in the body, being the *most abundant* divalent intracellular cation in the cells, the second most abundant cellular ion next to potassium and the fourth cation in general in the human body^[1].

Type 2 diabetes mellitus (DM2) is often accompanied by alteration of Mg status. An increased prevalence of Mg deficits have been identified in DM2 patients, especially in those with poorly controlled glycemic profiles, with longer duration of the disease and with the presence of micro- and macrovascular chronic complications^[2-6].

Laboratory tests with a high sensitivity and specificity and easy to perform to allow an accurate clinical assessment of Mg status are missing. Patients are considered frankly hypomagnesemic with serum Mg concentrations ≤ 0.61 mmol/L or 1.5 mg/dL^[7-9]. Mg concentrations ≤ 0.75 mmol/L or 1.8 mg/dL may be considered as preclinical hypomagnesemia^[10,11].

Mg deficiency can be present without hypomagnesemia. However, hypomagnesemia, when present, is usually indicative of an important systemic Mg deficit. A depletion in intracellular and/or ionized plasma Mg can be found in individuals with normal total serum Mg^[12]. However, most of the studies in the literature have measured total serum Mg instead of the free, ionized (bioactive) or the intracellular Mg concentrations, which make it a challenge to correlate Mg deficits to diseases.

We have recently confirmed that diabetic older patients are more prone to hypomagnesemia; this condition being closely related to metabolic control as measured by glycated hemoglobin even after adjustment for relevant confounders. Ionized Mg may help to identify diabetic older adults with low concentrations of blood Mg that are not evident with the only measurement of total Mg^[12].

Intracellular free Mg levels are consistently reduced in subjects with DM2, when compared with nondiabetic subjects^[1,13,14]. Although the mechanism has not been fully elucidated, an alteration in the mechanism(s) of the Mg uptake in the cells, and/or a deficit of ATP, may help to understand the cellular Mg deficit observed in DM2^[15]. The relationship between intracellular Mg and ATP concentration is rather complex. The decrease in cellular ATP might partially explain the decrease in cellular Mg. Otherwise, a decrease in cellular ATP leads to a decreased binding of Mg to ATP in the formation of MgATP, which might increase the intracellular Mg concentration.

The aim of this review is to revise current evidence

on the mechanisms of Mg deficiency in DM2. The evidence on the role of Mg supplementation in the management of DM2 will also be discussed.

MECHANISMS OF MG DEFICIENCY IN DM2

Reduced Mg intake and/or augmented Mg urinary loss are among the most important causes of Mg deficits in DM2, while Mg absorption and retention seems to be maintained^[16-18].

A relationship between Mg levels in the plasma and the development of DM2 in the general population has been suggested^[19]. DM2 is frequently accompanied by renal calcium and Mg loss^[20,21], but the mechanism(s) of this wasting is still not completely elucidated^[22].

Both hyperglycemia and hyperinsulinemia may increase urinary Mg excretion. Urinary Mg excretion and fasting blood glucose have been found to be inversely related to serum Mg levels. Thus, hyperglycemia decreases Mg tubular reabsorption^[20]. A good metabolic control is associated with a reduction of the urinary Mg wasting^[3].

In streptozotocin-induced diabetic rats, Lee *et al.*^[22] found an increase in renal Mg transporters. The alteration was corrected by insulin administration. Insulin resistance and hyperinsulinemia may also affect Mg transport^[21].

MG AND INSULIN SENSITIVITY

Hypomagnesemia in DM2 is present only in severe (and generally long lasting) Mg deficits. A chronic latent Mg deficiency without alteration in serum total Mg is more commonly observed^[12]. These often undetected Mg insufficiencies have clinical importance, since Mg is a main co-factor in numerous enzymatic reactions (> 300 enzymatic reactions including all the enzymes of glycolysis). Mg also is deeply involved in the regulation of insulin signaling, in the phosphorylation of insulin receptor kinase, in the post-receptorial action of insulin, and in insulin-mediated cellular glucose uptake^[17,23].

The clinical consequence of a chronic Mg deficit is post-receptorial insulin resistance and consequent reduced glucose utilization in the cells, worsening the reduced insulin sensitivity present in DM2^[18].

Another possible link between Mg deficiency and reduced insulin sensitivity is the presence of oxidative stress and/or inflammation. Thus, free radicals are often increased in DM2, hypertension, metabolic syndrome and aging, conditions also associated with Mg deficits^[24,25]. In particular, we demonstrated an age-dependent deficit of cellular Mg in persons aged 65 years and over, as well as in patients with essential hypertension or DM2, independently of age^[14,25].

Nevertheless, independently of the mechanisms of Mg deficits in DM2, metabolic syndrome, essential

hypertension and aging, it is apparent that this Mg deficiency may contribute to enhance the insulin resistance status of these conditions^[17,18]. Mg deficit could precede and cause post-receptorial resistance of insulin and alter glucose tolerance.

MG DEFICIENCY AND CARDIO-METABOLIC DISEASES

Mg deficiency may be also a factor implicated in DM2 complications. We found a relation between ionic changes and echocardiographic indices alterations^[26]. We observed an significant association of reduced cellular Mg with cardiac hypertrophy in DM2 patients^[26].

Cellular Mg measured *in vivo* in skeletal muscle and in the brain with ³¹P-NMR, was directly related to aortic distensibility^[27].

Reduced Mg levels were also associated with an increased prevalence of arrhythmias in DM2 obese subjects^[6], and with a more rapid decline of renal function. Thus, hypomagnesemia is currently considered an accurate predictor of progression of diabetic nephropathy^[28-30]. Mg deficits have also been associated with cognitive decline^[31], multimorbidity^[32] and aging^[25,33].

DIETARY MG DEFICIENCY MAY PREDISPOSE TO DM2

Dietary Mg deficiency may cause insulin resistance as shown by several studies both in humans and in experimental animals^[34-40]. In sheep, Mg-deficient diet caused a significant impairment of insulin-mediated glucose uptake^[35]. In rats, Mg supplements were able to postpone the onset of diabetes^[36]. In healthy women (without DM2), the higher was the intake of Mg, the lower were fasting levels of insulin^[37]. In young, nondiabetic African Americans, low dietary Mg was associated with insulin resistance and insulin responses to an oral glucose tolerance test^[38]. A low Mg diet in rats produced an increase in triglyceride and plasma glucose levels^[39]. In rats, a maternal restriction of dietary Mg was able to cause insulin resistance in pups^[40]. Suárez *et al.*^[41] suggested that the worsening of glucose metabolism induced by Mg dietary restriction in experimental rats is due to an impairment of both, insulin secretion and insulin action.

Deficiencies of Mg status including both hypomagnesemia and/or reduced dietary Mg intake have been linked to an enhanced risk to develop DM2 or glucose intolerance^[19,42-44]. Higher Mg intakes were conversely associated with a reduced incidence of DM2^[45].

Several studies have shown a clear association of Mg intake with DM2 and with cardio-metabolic syndrome, suggesting that a higher Mg consumption is related to a reduction of the incidence of these conditions. Two meta-analyses of prospective studies concluded that Mg intake is inversely associated with the onset of DM2^[46,47]. In addition, the development of the cardio-metabolic

syndrome has been linked to dietary Mg content^[34,48]. Hypomagnesemia itself in a 10-year follow-up study was associated with glucose tolerance impairment^[49]. Conversely, higher Mg intake was associated with increased insulin sensitivity^[50] and with decreased risk of incident DM2, with a decreased risk of 0.68 in the higher compared with the lower quintiles^[51,52].

Similar findings were obtained in the CARDIA study, during a 20-year follow-up, which also confirmed the reverse relationship of dietary Mg with inflammation markers^[53].

POSSIBLE USE OF MG SUPPLEMENTS IN THE MANAGEMENT OF DM2

The detection and correction of altered Mg status in diabetic patients is clinically appropriate, although many physicians tend to ignore Mg status. The increased risk of developing impaired glucose tolerance and/or frank DM2 in persons with dietary or serum Mg deficits have suggested a potential benefit of Mg supplements in patients with DM2 or in the presence of risk factors for DM2. Mg supplements have been proposed as a complementary tool for the prevention of DM2 and its metabolic control^[54,55]. Some benefits of Mg supplements on glycemic profiles have been found in most but not all studies.

Regrettably, results from clinical trials are still limited^[56]. Thus, the clinical evidence of a clear effect of Mg supplementation on metabolic indices in persons with DM2 are controversial. Some benefit has been found in several^[8,54,57,58], but not in all clinical studies^[59]. The hypothesis of a role of supplemental Mg in the control of DM2 still needs to be ascertained by large randomized clinical trials^[60,61]. Mg supplementation may improve glycemic concentrations in fasting and postprandial states, and insulin sensitivity. We found a significant relationship between the increase in serum and cellular Mg and insulin sensitivity^[62]. We also showed that Mg supplementation is able to improve an altered endothelial function in DM2 older adults^[63]. Barragán-Rodríguez *et al.*^[64] suggested a positive effect in the treatment of depression in older persons with DM2 and hypomagnesemia. Presumably, the main problem is that all RCTs were underpowered, partially through overestimation of the treatment effect. Differences may be related to the fact that most of the existing studies have included a small number of subjects, using different Mg doses and different Mg salts.

Several studies have linked high Mg content present in fiber with the positive action of whole grains to improve insulin sensitivity^[65-68]. Oral Mg supplements have been shown to improve fasting and postprandial glucose levels and insulin sensitivity in hypomagnesemic DM2 patients^[57], to improve insulin sensitivity in non-diabetic subjects with insulin resistance^[8], and to decrease C-reactive protein levels in hypomagnesemic patients with prediabetes^[69].

In summary, oral Mg supplements appear to be useful in persons with DM2 to restore Mg deficiencies, to improve insulin resistance, oxidative stress, and systemic inflammation.

The absence of large trials in DM2 patients specifically focusing on those with Mg deficit may help to explain the inconsistency between epidemiological (mainly positive) and clinical (mostly controversial) studies. Since most, but not all, DM2 patients have Mg deficiency, it would be useful to focus on those with deficit in order to correct it. Differences in Mg balance, glycemic control, and age are other potential factors that may help to explain the differences among the studies. Most studies used total serum Mg concentration instead of the free, ionized (bioactive) Mg concentration, which make it a challenge to correlate Mg deficiency to diseases.

Future prospective large RCTs would be important to support the possible inclusion of Mg supplements in the guidelines for the management of DM2.

REFERENCES

- 1 **Barbagallo M**, Dominguez LJ. Magnesium metabolism in type 2 diabetes mellitus, metabolic syndrome and insulin resistance. *Arch Biochem Biophys* 2007; **458**: 40-47 [PMID: 16808892 DOI: 10.1016/j.abb.2006.05.007]
- 2 **Mather HM**, Levin GE. Magnesium status in diabetes. *Lancet* 1979; **1**: 924 [PMID: 86688 DOI: 10.1016/S0140-6736(79)91400-4]
- 3 **Schnack C**, Bauer I, Pregant P, Hopmeier P, Scherthaner G. Hypomagnesaemia in type 2 (non-insulin-dependent) diabetes mellitus is not corrected by improvement of long-term metabolic control. *Diabetologia* 1992; **35**: 77-79 [PMID: 1541384 DOI: 10.1007/BF00400855]
- 4 **Ramadass S**, Basu S, Srinivasan AR. SERUM magnesium levels as an indicator of status of Diabetes Mellitus type 2. *Diabetes Metab Syndr* 2015; **9**: 42-45 [PMID: 25470649 DOI: 10.1016/j.dsx.2014.04.024]
- 5 **Ma J**, Folsom AR, Melnick SL, Eckfeldt JH, Sharrett AR, Nabulsi AA, Hutchinson RG, Metcalf PA. Associations of serum and dietary magnesium with cardiovascular disease, hypertension, diabetes, insulin, and carotid arterial wall thickness: the ARIC study. Atherosclerosis Risk in Communities Study. *J Clin Epidemiol* 1995; **48**: 927-940 [PMID: 7782801 DOI: 10.1016/0895-4356(94)00200-A]
- 6 **Del Gobbo LC**, Song Y, Poirier P, Dewailly E, Elin RJ, Egeland GM. Low serum magnesium concentrations are associated with a high prevalence of premature ventricular complexes in obese adults with type 2 diabetes. *Cardiovasc Diabetol* 2012; **11**: 23 [PMID: 22405520 DOI: 10.1186/1475-2840-11-23]
- 7 **Hashizume N**, Mori M. An analysis of hypermagnesemia and hypomagnesemia. *Jpn J Med* 1990; **29**: 368-372 [PMID: 2273620 DOI: 10.2169/internalmedicine1962.29.368]
- 8 **Guerrero-Romero F**, Tamez-Perez HE, González-González G, Salinas-Martínez AM, Montes-Villarreal J, Treviño-Ortiz JH, Rodríguez-Morán M. Oral magnesium supplementation improves insulin sensitivity in non-diabetic subjects with insulin resistance. A double-blind placebo-controlled randomized trial. *Diabetes Metab* 2004; **30**: 253-258 [PMID: 15223977 DOI: 10.1016/S1262-3636(07)70116-7]
- 9 **Wong ET**, Rude RK, Singer FR, Shaw ST. A high prevalence of hypomagnesemia and hypermagnesemia in hospitalized patients. *Am J Clin Pathol* 1983; **79**: 348-352 [PMID: 6829504]
- 10 **Chernow B**, Bamberger S, Stoiko M, Vadnais M, Mills S, Hoellerich V, Warshaw AL. Hypomagnesemia in patients in postoperative intensive care. *Chest* 1989; **95**: 391-397 [PMID: 2914492 DOI: 10.1378/chest.95.2.391]
- 11 **Whang R**, Ryder KW. Frequency of hypomagnesemia and hypermagnesemia. Requested vs routine. *JAMA* 1990; **263**: 3063-3064 [PMID: 2342219 DOI: 10.1001/jama.1990.03440220087036]
- 12 **Barbagallo M**, Di Bella G, Brucato V, D'Angelo D, Damiani P, Monteverde A, Belvedere M, Dominguez LJ. Serum ionized magnesium in diabetic older persons. *Metabolism* 2014; **63**: 502-509 [PMID: 24462317 DOI: 10.1016/j.metabol.2013.12.003]
- 13 **Resnick LM**, Altura BT, Gupta RK, Laragh JH, Alderman MH, Altura BM. Intracellular and extracellular magnesium depletion in type 2 (non-insulin-dependent) diabetes mellitus. *Diabetologia* 1993; **36**: 767-770 [PMID: 8405745 DOI: 10.1007/BF00401149]
- 14 **Barbagallo M**, Gupta RK, Dominguez LJ, Resnick LM. Cellular ionic alterations with age: relation to hypertension and diabetes. *J Am Geriatr Soc* 2000; **48**: 1111-1116 [PMID: 10983912 DOI: 10.1111/j.1532-5415.2000.tb04788.x]
- 15 **Barbagallo M**, Dominguez LJ, Resnick LM. Magnesium metabolism in hypertension and type 2 diabetes mellitus. *Am J Ther* 2007; **14**: 375-385 [PMID: 17667214 DOI: 10.1097/01.mjt.0000209676.91582.46]
- 16 **Wälti MK**, Zimmermann MB, Walczyk T, Spinas GA, Hurrell RF. Measurement of magnesium absorption and retention in type 2 diabetic patients with the use of stable isotopes. *Am J Clin Nutr* 2003; **78**: 448-453 [PMID: 12936928]
- 17 **Günther T**. Magnesium in bone and the magnesium load test. *Magnes Res* 2011; **24**: 223-224 [PMID: 22192898 DOI: 10.1684/mrh.2011.0297]
- 18 **Barbagallo M**, Dominguez LJ, Galioto A, Ferlisi A, Cani C, Malfa L, Pineo A, Busardo A, Paolisso G. Role of magnesium in insulin action, diabetes and cardio-metabolic syndrome X. *Mol Aspects Med* 2003; **24**: 39-52 [PMID: 12537988 DOI: 10.1016/S0098-2997(02)00090-0]
- 19 **Kao WH**, Folsom AR, Nieto FJ, Mo JP, Watson RL, Brancati FL. Serum and dietary magnesium and the risk for type 2 diabetes mellitus: the Atherosclerosis Risk in Communities Study. *Arch Intern Med* 1999; **159**: 2151-2159 [PMID: 10527292 DOI: 10.1001/archinte.159.18.2151]
- 20 **McNair P**, Christensen MS, Christiansen C, Madsbad S, Transbøl I. Renal hypomagnesaemia in human diabetes mellitus: its relation to glucose homeostasis. *Eur J Clin Invest* 1982; **12**: 81-85 [PMID: 6802656 DOI: 10.1111/j.1365-2362.1982.tb00942.x]
- 21 **Djurhuus MS**, Skott P, Hother-Nielsen O, Klitgaard NA, Beck-Nielsen H. Insulin increases renal magnesium excretion: a possible cause of magnesium depletion in hyperinsulinaemic states. *Diabet Med* 1995; **12**: 664-669 [PMID: 7587003 DOI: 10.1111/j.1464-5491.1995.tb00566.x]
- 22 **Lee CT**, Lien YH, Lai LW, Chen JB, Lin CR, Chen HC. Increased renal calcium and magnesium transporter abundance in streptozotocin-induced diabetes mellitus. *Kidney Int* 2006; **69**: 1786-1791 [PMID: 16557223 DOI: 10.1038/sj.ki.5000344]
- 23 **Saris NE**, Mervaala E, Karppanen H, Khawaja JA, Lewenstam A. Magnesium. An update on physiological, clinical and analytical aspects. *Clin Chim Acta* 2000; **294**: 1-26 [PMID: 10727669 DOI: 10.1016/S0009-8981(99)00258-2]
- 24 **Weglicki WB**. Hypomagnesemia and inflammation: clinical and basic aspects. *Annu Rev Nutr* 2012; **32**: 55-71 [PMID: 22404119 DOI: 10.1146/annurev-nutr-071811-150656]
- 25 **Barbagallo M**, Dominguez LJ. Magnesium and aging. *Curr Pharm Des* 2010; **16**: 832-839 [PMID: 20388094 DOI: 10.2174/138161210790883679]
- 26 **Barbagallo M**, Gupta RK, Resnick LM. Cellular ions in NIDDM: relation of calcium to hyperglycemia and cardiac mass. *Diabetes Care* 1996; **19**: 1393-1398 [PMID: 8941470 DOI: 10.2337/diacare.19.12.1393]
- 27 **Resnick LM**, Militianu D, Cunnings AJ, Pipe JG, Evelhoch JL, Soulen RL. Direct magnetic resonance determination of aortic distensibility in essential hypertension: relation to age, abdominal visceral fat, and in situ intracellular free magnesium. *Hypertension* 1997; **30**: 654-659 [PMID: 9322999 DOI: 10.1161/01.HYP.30.3.654]

- 28 **Sakaguchi Y**, Shoji T, Hayashi T, Suzuki A, Shimizu M, Mitumoto K, Kawabata H, Niihata K, Okada N, Isaka Y, Rakugi H, Tsubakihara Y. Hypomagnesemia in type 2 diabetic nephropathy: a novel predictor of end-stage renal disease. *Diabetes Care* 2012; **35**: 1591-1597 [PMID: 22498805 DOI: 10.2337/dc12-0226]
- 29 **Van Laecke S**, Nagler EV, Verbeke F, Van Biesen W, Vanholder R. Hypomagnesemia and the risk of death and GFR decline in chronic kidney disease. *Am J Med* 2013; **126**: 825-831 [PMID: 23891286 DOI: 10.1016/j.amjmed.2013.02.036]
- 30 **Tin A**, Grams ME, Maruthur NM, Astor BC, Couper D, Mosley TH, Selvin E, Coresh J, Kao WH. Results from the Atherosclerosis Risk in Communities study suggest that low serum magnesium is associated with incident kidney disease. *Kidney Int* 2015; **87**: 820-827 [PMID: 25272232 DOI: 10.1038/ki.2014.331]
- 31 **Barbagallo M**, Belvedere M, Di Bella G, Dominguez LJ. Altered ionized magnesium levels in mild-to-moderate Alzheimer's disease. *Magnes Res* 2011; **24**: S115-S121 [PMID: 21951617 DOI: 10.1684/mrh.2011.0287]
- 32 **Ruel G**, Shi Z, Zhen S, Zuo H, Kröger E, Sirois C, Lévesque JF, Taylor AW. Association between nutrition and the evolution of multimorbidity: the importance of fruits and vegetables and whole grain products. *Clin Nutr* 2014; **33**: 513-520 [PMID: 23931982 DOI: 10.1016/j.clnu.2013.07.009]
- 33 **Barbagallo M**, Belvedere M, Dominguez LJ. Magnesium homeostasis and aging. *Magnes Res* 2009; **22**: 235-246 [PMID: 20228001 DOI: 10.1684/mrh.2009.0187]
- 34 **He K**, Liu K, Daviglus ML, Morris SJ, Loria CM, Van Horn L, Jacobs DR, Savage PJ. Magnesium intake and incidence of metabolic syndrome among young adults. *Circulation* 2006; **113**: 1675-1682 [PMID: 16567569 DOI: 10.1161/CIRCULATIONAHA.105.588327]
- 35 **Matsunobu S**, Terashima Y, Senshu T, Sano H, Itoh H. Insulin secretion and glucose uptake in hypomagnesemic sheep fed a low magnesium, high potassium diet. *J Nutr Biochem* 1990; **1**: 167-171 [PMID: 15539200 DOI: 10.1016/0955-2863(90)90018-G]
- 36 **Balon TW**, Gu JL, Tokuyama Y, Jasman AP, Nadler JL. Magnesium supplementation reduces development of diabetes in a rat model of spontaneous NIDDM. *Am J Physiol* 1995; **269**: E745-E752 [PMID: 7485490]
- 37 **Fung TT**, Manson JE, Solomon CG, Liu S, Willett WC, Hu FB. The association between magnesium intake and fasting insulin concentration in healthy middle-aged women. *J Am Coll Nutr* 2003; **22**: 533-538 [PMID: 14684759 DOI: 10.1080/07315724.2003.10719332]
- 38 **Humphries S**, Kushner H, Falkner B. Low dietary magnesium is associated with insulin resistance in a sample of young, nondiabetic Black Americans. *Am J Hypertens* 1999; **12**: 747-756 [PMID: 10480466 DOI: 10.1016/S0895-7061(99)00041-2]
- 39 **Chaudhary DP**, Boparai RK, Sharma R, Bansal DD. Studies on the development of an insulin resistant rat model by chronic feeding of low magnesium high sucrose diet. *Magnes Res* 2004; **17**: 293-300 [PMID: 15726905]
- 40 **Venu L**, Kishore YD, Raghunath M. Maternal and perinatal magnesium restriction predisposes rat pups to insulin resistance and glucose intolerance. *J Nutr* 2005; **135**: 1353-1358 [PMID: 15930437]
- 41 **Suárez A**, Pulido N, Casla A, Casanova B, Arrieta FJ, Rovira A. Impaired tyrosine-kinase activity of muscle insulin receptors from hypomagnesaemic rats. *Diabetologia* 1995; **38**: 1262-1270 [PMID: 8582534 DOI: 10.1007/BF00401757]
- 42 **Mather HM**, Nisbet JA, Burton GH, Poston GJ, Bland JM, Bailey PA, Pilkington TR. Hypomagnesaemia in diabetes. *Clin Chim Acta* 1979; **95**: 235-242 [PMID: 527222 DOI: 10.1016/0009-8981(79)90364-4]
- 43 **Yokota K**. [Diabetes mellitus and magnesium]. *Clin Calcium* 2005; **15**: 203-212 [PMID: 15692158]
- 44 **Longstreet DA**, Heath DL, Vink R. A potential link between magnesium intake and diabetes in Indigenous Australians. *Med J Aust* 2005; **183**: 219-220 [PMID: 16097927]
- 45 **Colditz GA**, Manson JE, Stampfer MJ, Rosner B, Willett WC, Speizer FE. Diet and risk of clinical diabetes in women. *Am J Clin Nutr* 1992; **55**: 1018-1023 [PMID: 1315120]
- 46 **Larsson SC**, Wolk A. Magnesium intake and risk of type 2 diabetes: a meta-analysis. *J Intern Med* 2007; **262**: 208-214 [PMID: 17645588 DOI: 10.1111/j.1365-2796.2007.01840.x]
- 47 **Dong JY**, Qin LQ. Dietary calcium intake and risk of type 2 diabetes: possible confounding by magnesium. *Eur J Clin Nutr* 2012; **66**: 408-410 [PMID: 22318650 DOI: 10.1038/ejcn.2012.5]
- 48 **Song Y**, Ridker PM, Manson JE, Cook NR, Buring JE, Liu S. Magnesium intake, C-reactive protein, and the prevalence of metabolic syndrome in middle-aged and older U.S. women. *Diabetes Care* 2005; **28**: 1438-1444 [PMID: 15920065 DOI: 10.2337/diacare.28.6.1438]
- 49 **Guerrero-Romero F**, Rascón-Pacheco RA, Rodríguez-Morán M, de la Peña JE, Wachter N. Hypomagnesaemia and risk for metabolic glucose disorders: a 10-year follow-up study. *Eur J Clin Invest* 2008; **38**: 389-396 [PMID: 18489400 DOI: 10.1111/j.1365-2362.2008.01957.x]
- 50 **Ma B**, Lawson AB, Liese AD, Bell RA, Mayer-Davis EJ. Dairy, magnesium, and calcium intake in relation to insulin sensitivity: approaches to modeling a dose-dependent association. *Am J Epidemiol* 2006; **164**: 449-458 [PMID: 16861328 DOI: 10.1093/aje/kwj246]
- 51 **Song Y**, Manson JE, Buring JE, Liu S. Dietary magnesium intake in relation to plasma insulin levels and risk of type 2 diabetes in women. *Diabetes Care* 2004; **27**: 59-65 [PMID: 14693967 DOI: 10.2337/diacare.27.1.59]
- 52 **Lopez-Ridaura R**, Willett WC, Rimm EB, Liu S, Stampfer MJ, Manson JE, Hu FB. Magnesium intake and risk of type 2 diabetes in men and women. *Diabetes Care* 2004; **27**: 134-140 [PMID: 14693979 DOI: 10.2337/diacare.27.1.134]
- 53 **Kim DJ**, Xun P, Liu K, Loria C, Yokota K, Jacobs DR, He K. Magnesium intake in relation to systemic inflammation, insulin resistance, and the incidence of diabetes. *Diabetes Care* 2010; **33**: 2604-2610 [PMID: 20807870 DOI: 10.2337/dc10-0994]
- 54 **Guerrero-Romero F**, Rodríguez-Morán M. Complementary therapies for diabetes: the case for chromium, magnesium, and antioxidants. *Arch Med Res* 2005; **36**: 250-257 [PMID: 15925015 DOI: 10.1016/j.arcmed.2005.01.004]
- 55 **McCarty MF**. Complementary vascular-protective actions of magnesium and taurine: a rationale for magnesium taurate. *Med Hypotheses* 1996; **46**: 89-100 [PMID: 8692051 DOI: 10.1016/S0306-9877(96)90007-9]
- 56 **Rodríguez-Morán M**, Simental Mendía LE, Zambrano Galván G, Guerrero-Romero F. The role of magnesium in type 2 diabetes: a brief based-clinical review. *Magnes Res* 2011; **24**: 156-162 [PMID: 22198525 DOI: 10.1684/mrh.2011.0299]
- 57 **Rodríguez-Morán M**, Guerrero-Romero F. Oral magnesium supplementation improves insulin sensitivity and metabolic control in type 2 diabetic subjects: a randomized double-blind controlled trial. *Diabetes Care* 2003; **26**: 1147-1152 [PMID: 12663588 DOI: 10.2337/diacare.26.4.1147]
- 58 **Yokota K**, Kato M, Lister F, Li H, Hayakawa T, Kikuta T, Kageyama S, Tajima N. Clinical efficacy of magnesium supplementation in patients with type 2 diabetes. *J Am Coll Nutr* 2004; **23**: 506S-509S [PMID: 15466952 DOI: 10.1080/07315724.2004.10719390]
- 59 **de Valk HW**, Verkaaik R, van Rijn HJ, Geerdink RA, Struyvenberg A. Oral magnesium supplementation in insulin-requiring Type 2 diabetic patients. *Diabet Med* 1998; **15**: 503-507 [PMID: 9632126 DOI: 10.1002/(SICI)1096-9136(199806)15:6<503::AID-DIA596>3.0.CO;2-M]
- 60 **McCarty MF**. Nutraceutical resources for diabetes prevention--an update. *Med Hypotheses* 2005; **64**: 151-158 [PMID: 15533633 DOI: 10.1016/j.mehy.2004.03.036]
- 61 **Schulze MB**, Hu FB. Primary prevention of diabetes: what can be done and how much can be prevented? *Annu Rev Public Health* 2005; **26**: 445-467 [PMID: 15760297 DOI: 10.1146/annurev.publhealth.26.021304.144532]
- 62 **Paolisso G**, Barbagallo M. Hypertension, diabetes mellitus, and insulin resistance: the role of intracellular magnesium. *Am J Hypertens* 1997; **10**: 346-355 [PMID: 9056694 DOI: 10.1016/

- S0895-7061(96)00342-1]
- 63 **Barbagallo M**, Dominguez LJ, Galioto A, Pineo A, Belvedere M. Oral magnesium supplementation improves vascular function in elderly diabetic patients. *Magnes Res* 2010; **23**: 131-137 [PMID: 20736142 DOI: 10.1684/mrh.2010.0214]
- 64 **Barragán-Rodríguez L**, Rodríguez-Morán M, Guerrero-Romero F. Efficacy and safety of oral magnesium supplementation in the treatment of depression in the elderly with type 2 diabetes: a randomized, equivalent trial. *Magnes Res* 2008; **21**: 218-223 [PMID: 19271419]
- 65 **McCarty MF**. Magnesium may mediate the favorable impact of whole grains on insulin sensitivity by acting as a mild calcium antagonist. *Med Hypotheses* 2005; **64**: 619-627 [PMID: 15617878 DOI: 10.1016/j.mehy.2003.10.034]
- 66 **Weickert MO**, Möhlig M, Schöfl C, Arafat AM, Otto B, Viehoff H, Koebnick C, Kohl A, Spranger J, Pfeiffer AF. Cereal fiber improves whole-body insulin sensitivity in overweight and obese women. *Diabetes Care* 2006; **29**: 775-780 [PMID: 16567814 DOI: 10.2337/diacare.29.04.06.dc05-2374]
- 67 **Liese AD**, Roach AK, Sparks KC, Marquart L, D'Agostino RB, Mayer-Davis EJ. Whole-grain intake and insulin sensitivity: the Insulin Resistance Atherosclerosis Study. *Am J Clin Nutr* 2003; **78**: 965-971 [PMID: 14594783]
- 68 **McKeown NM**. Whole grain intake and insulin sensitivity: evidence from observational studies. *Nutr Rev* 2004; **62**: 286-291 [PMID: 15384920 DOI: 10.1111/j.1753-4887.2004.tb00054.x]
- 69 **Simental-Mendia LE**, Rodríguez-Morán M, Guerrero-Romero F. Oral magnesium supplementation decreases C-reactive protein levels in subjects with prediabetes and hypomagnesemia: a clinical randomized double-blind placebo-controlled trial. *Arch Med Res* 2014; **45**: 325-330 [PMID: 24814039 DOI: 10.1016/j.arcmed.2014.04.006]

P- Reviewer: Das UN, Sriraman R, Tomkin GH
S- Editor: Tian YL **L- Editor:** A **E- Editor:** Jiao XK



Vitamin paradox in obesity: Deficiency or excess?

Shi-Sheng Zhou, Da Li, Na-Na Chen, Yiming Zhou

Shi-Sheng Zhou, Institute of Basic Medical Sciences, Medical College, Dalian University, Dalian 116622, Liaoning Province, China

Da Li, Department of Obstetrics and Gynecology, Shengjing Hospital of China Medical University, Shenyang 110004, Liaoning Province, China

Na-Na Chen, Department of Molecular Immunology, Graduate School of Medicine, Nagoya University, Nagoya 466-8550, Japan

Yiming Zhou, Renal Division, Department of Medicine, Brigham and Women's Hospital, Harvard Institutes of Medicine, Harvard Medical School, Boston, MA 02115, United States

Author contributions: Zhou SS contributed to the conception and design of the study; Zhou SS, Li D, Chen NN and Zhou Y provided substantial contributions in drafting the article or making critical revisions related to important intellectual content of the manuscript; all authors read and approved the final manuscript.

Supported by National Natural Science Foundation of China, No. 31140036.

Conflict-of-interest statement: All authors have no conflicts of interests to declare regarding this manuscript.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Shi-Sheng Zhou, MD, PhD, Professor, Institute of Basic Medical Sciences, Medical College, Dalian University, No.10 Xuefu Avenue, Dalian Economic and Technological Development Zone, Dalian 116622, Liaoning Province, China. zhouss@ymail.com
Telephone: +86-411-87402740
Fax: +86-411-87402053

Received: May 23, 2015

Peer-review started: May 23, 2015

First decision: July 6, 2015

Revised: July 19, 2015

Accepted: July 29, 2015

Article in press: August 3, 2015

Published online: August 25, 2015

Abstract

Since synthetic vitamins were used to fortify food and as supplements in the late 1930s, vitamin intake has significantly increased. This has been accompanied by an increased prevalence of obesity, a condition associated with diabetes, hypertension, cardiovascular disease, asthma and cancer. Paradoxically, obesity is often associated with low levels of fasting serum vitamins, such as folate and vitamin D. Recent studies on folic acid fortification have revealed another paradoxical phenomenon: obesity exhibits low fasting serum but high erythrocyte folate concentrations, with high levels of serum folate oxidation products. High erythrocyte folate status is known to reflect long-term excess folic acid intake, while increased folate oxidation products suggest an increased folate degradation because obesity shows an increased activity of cytochrome P450 2E1, a monooxygenase enzyme that can use folic acid as a substrate. There is also evidence that obesity increases niacin degradation, manifested by increased activity/expression of niacin-degrading enzymes and high levels of niacin metabolites. Moreover, obesity most commonly occurs in those with a low excretory reserve capacity (*e.g.*, due to low birth weight/preterm birth) and/or a low sweat gland activity (black race and physical inactivity). These lines of evidence raise the possibility that low fasting serum vitamin status in obesity may be a compensatory response to chronic excess vitamin intake, rather than vitamin deficiency, and that obesity could be one of the manifestations of chronic vitamin poisoning. In this article, we discuss vitamin paradox in obesity from the perspective of vitamin homeostasis.

Key words: Obesity; Type 2 diabetes; Developmental

origin of disease; Folic acid; Vitamin D; Niacin; Oxidative stress; Insulin resistance; Vitamin fortification

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Obesity rates have dramatically increased among the United States population, including children, since the 1980s. Considering the lag time between risk exposure and the development of child obesity, the risk must have been imposed on the whole United States population around the late 1970s. Although evidence suggests that the risk is high vitamin intake due to the update of vitamin fortification in 1974 and the implementation of the Infant Formula Act of 1980, why do obese individuals paradoxically show low levels of fasting serum vitamins? In this paper, we try to give an answer to this question based on the current understanding of vitamin homeostasis.

Zhou SS, Li D, Chen NN, Zhou Y. Vitamin paradox in obesity: Deficiency or excess? *World J Diabetes* 2015; 6(10): 1158-1167 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v6/i10/1158.htm> DOI: <http://dx.doi.org/10.4239/wjcd.v6.i10.1158>

INTRODUCTION

Obesity, a global health problem, is associated with co-morbidities such as metabolic syndrome, diabetes, hypertension, asthma, nonalcoholic fatty liver disease, renal disease, cardiovascular disease and cancer, which are thought to be of developmental origin^[1]. Since the late 1930s, when synthetic vitamins, thiamin, riboflavin and niacin (nicotinic acid and nicotinamide), were used to fortify foods or as dietary supplements, the daily intake of vitamins of the United States population has significantly increased, especially after the update of mandatory fortification in 1974^[2] and the implementation of the Infant Formula Act of 1980 (without setting an upper limit for most vitamins)^[3]. In fact, the introduction of synthetic vitamins into the diet was followed by a dramatic increase in the prevalence of obesity among all age groups in the United States^[4,5]. Similar correlations between increased obesity and vitamin fortification were observed in other vitamin-fortified countries, such as Canada and Saudi Arabia^[2]. Over the past 20-30 years, China has also been experiencing a rapid growth in the rates of obesity^[6] after having shifted from a low to a high vitamin intake, due to a combination of increased intake of animal-derived foods (rich in vitamin B₁, B₂ and niacin)^[7] and mandatory flour fortification with these vitamins, which was introduced in China in the late 1980s and was been mandatorily implemented in 1994^[2]. Paradoxically, it is frequently reported that obesity and type 2 diabetes are associated with low levels of fasting serum vitamins, including vitamin B₁, D, and folate^[8-10]. Although

the mechanism of the paradox remains unclear, it is generally thought that the low vitamin status in obesity is due to inadequate intake.

Since 1998, enriched grain products in the United States have been fortified with folic acid to prevent neural tube defects. Recent studies on folic acid fortification show that obese individuals also show lower fasting serum folate concentrations, but, paradoxically, their red blood cell (RBC) folate concentrations and MeFox (5-methyltetrahydrofolate oxidation product) are significantly higher, when compared with nonobese individuals^[11,12]. Moreover, obesity is also found to be associated with increased activity of cytochrome P450 (CYP) 2E1, a monooxygenase enzyme that can use folic acid as a substrate^[13]. Folate content in RBC is known to reflect long-term average consumption and tissue stores because RBC only accumulates folate during erythropoiesis^[14], and increased serum MeFox suggests increased degradation of folic acid. Moreover, recent evidence shows that obesity is associated with high fasting serum N¹-methylnicotinamide without significant changes in nicotinamide levels^[15] and that plasma N¹-methylnicotinamide correlates with increased tissue expression of nicotinamide N-methyltransferase (NNMT, a major enzyme responsible for the degradation of nicotinamide to N¹-methylnicotinamide) and the degree of insulin resistance^[16]. Collectively, these observations raise the possibility that the vitamin paradox in obesity may involve vitamin excess rather than deficiency. After more than seven decades of practice of vitamin fortification and painful global experience of increasing prevalence of obesity and related diseases worldwide, it is time for us to examine the relationship between vitamin fortification and vitamin paradox from the perspective of vitamin homeostasis.

VITAMIN HOMEOSTASIS AND OXIDATIVE STRESS

Vitamins are essential micronutrients needed by the body in small amounts. Vitamin homeostasis is a balance between vitamin intake and clearance. A deficiency or excess may lead to deleterious effects. Since the introduction of synthetic vitamins into food, high vitamin intake is very common during a person's lifespan from conception through to old age^[2]. In this case, the removal of excess vitamins becomes particularly important in maintaining vitamin homeostasis. This depends on the efficiency of both excretory organs and drug-metabolizing enzymes.

Excretion of vitamins

The kidneys and sweat glands are the two major excretory organs responsible for the elimination of water-soluble vitamins, and the sebaceous glands excrete lipid-soluble vitamins in the sebum^[17]. The excretion of vitamins is positively related to their intake. Aging is known to be associated with decreasing function

of excretory organs^[18,19] and thus may reduce the clearance of vitamins. It is noteworthy that sweat excretion may be particularly important in eliminating excess water-soluble vitamins, because vitamins (e.g., folate^[20], nicotinic acid and nicotinamide^[2,21]) are barely excreted in the urine before degradation due to the reabsorption by the renal tubules, but they can be easily excreted in the sweat^[22-24]. The efficiency of sweat excretion is determined by several factors, including genetic background, intrauterine and early postnatal development, environmental temperature and physical activity. Compared with whites, blacks have a high sweating threshold, manifested by lower skin conductance (*i.e.*, low insensible perspiration)^[25] and sweating rates^[26] under the same ambient temperature condition, suggesting that blacks may have lower sweat excretion of vitamins than whites.

The formation of functional sweat glands begins at week 36 of gestation and completes within 10 wk of postnatal life^[27,28]. This process is affected not only by gestational age but also by the environmental temperature during the early postnatal period. As demonstrated in the literature, preterm birth is associated not only with a lower renal reserve capacity^[29] but also with a low sweating function^[30,31]. Low temperature may cause newborn hypothermia^[32], which may occur even in summer season^[32]. Reduced sweat gland function (*i.e.*, low skin conductance) has been found to be associated with a winter birth in schizophrenia^[33]. Therefore, preterm birth and newborn hypothermia may be associated with decreased vitamin clearance.

Ambient temperature and physical activity are two important factors affecting the excretion rates of sweat and sebum. For example, a decrease in temperature from 30 °C to 22 °C reduces insensible perspiration from about 700 mL/d to 380 mL/d in adults^[34], and a one-degree decrease in local skin temperature decreases the sebum excretion rate by 10%^[35]. There is evidence showing that the levels of plasma vitamin A and E are lower in summer than in winter^[36], and a similar seasonal variation is found in blood drug concentrations^[37]. Thus, it is conceivable that physical inactivity and winter or cold weather would decrease the tolerance to high vitamin intake.

On the other hand, it should be noted that excess sweat vitamin excretion may cause or worsen water-soluble-vitamin deficiency if there is poor vitamin intake. A good example may be pellagra, a niacin-deficiency disease that affects those who live in poverty without sufficient animal-source foods (rich in nicotinamide), with the symptoms occurring during the summer^[38], a season with the highest sweat excretion rates. However, over the past decades, both natural and artificial sources (*i.e.*, vitamin fortification and supplementation) of vitamins have significantly increased^[2], while sweat excretion has significantly decreased due to physical inactivity and the widespread use of air conditioning. These dietary and lifestyle changes may increase the

risk of excess accumulation of vitamins in the body, especially in those with reduced excretory capacity and/or activity.

Degradation of vitamins

Besides being directly excreted, vitamins also undergo degradation through phase I (including oxidation, reduction, and hydrolysis) and phase II metabolisms (e.g., sulfation, methylation and glutathione conjugation), which are catalysed by phase I and phase II drug-metabolizing enzymes, respectively. After phase I and/or phase II degradation, vitamins become more water-soluble and then can be more easily excreted from the body. Excess vitamins are degraded very rapidly. For example, cumulative administration of 2000 mg nicotinic acid [166 times the estimated average daily requirement (EAR)] in 13 h 10 min is found to only increase the levels of its metabolites in the plasma, without significantly changing plasma nicotinic acid concentrations^[39]. We found that, at 5 h after oral administration of 100 mg nicotinamide (8.3 times the EAR), plasma nicotinamide had returned to near baseline levels, while its metabolite *N*¹-methylnicotinamide remained at high levels^[24]. Thus, it is clear that a transient increase in vitamin intake may not change fasting vitamin levels.

Vitamins, xenobiotics, neurotransmitters and hormones share the same drug-metabolizing enzyme system, so they may interact with one another in their metabolism by inducing and competing for the enzymes^[3,40]. For example, CYP2E1, highly expressed in obesity and type 2 diabetes^[13], has more than 50 compounds, including some vitamins and ethanol^[41]. Thus, it is conceivable that alcohol may cause low fasting vitamin levels by induced CYP2E1.

Phase II metabolism of vitamins consumes detoxification resources, such as methyl-group donors, sulphate donors and glutathione, which are also necessary for the degradation of neurotransmitters and hormones. Therefore, excess vitamins can disturb the phase II metabolism of neurotransmitters and hormones by competing for the limited detoxification resources^[3]. Here, we take niacin methylation as an example to explain how excess vitamins affect metabolism of neurotransmitters and hormones. Methylation is a methyl-group transfer reaction from a methyl donor to a substrate, which is mediated by the methionine-homocysteine cycle. Methyl donors, including betaine and choline, are non-renewable resources in the body, while other components in the methylation system, including methionine, folate, vitamin B₁₂ and relevant enzymes, can be repeatedly used in the reaction system. Choline can be used as a methyl donor only after being converted to betaine in the liver and kidneys. According to the relationship of the components in the methylation reaction system shown in Figure 1, it is quite clear that an increase in the levels of substrates will mainly increase the demand for betaine. Since niacin is degraded mainly through

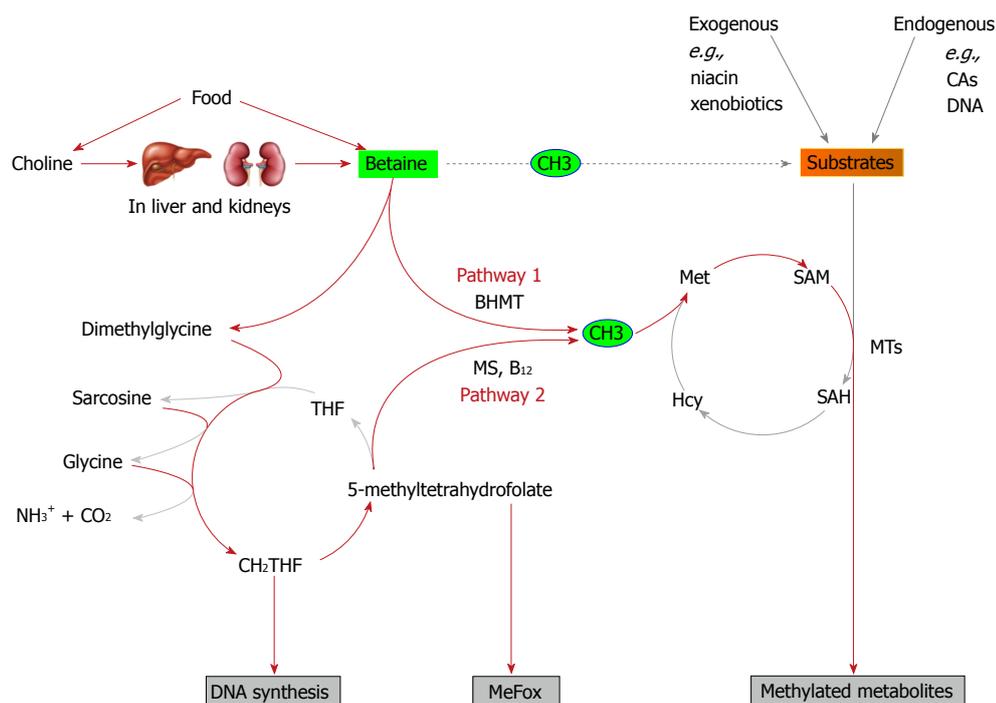


Figure 1 Relationship between methyl donors and mediators in the methylation of substrates. Methylation is a methyl-group transfer reaction from a methyl donor to a substrate, which is mediated by the methionine (Met) cycle. The deep red-arrow lines indicate the flow/transfer of methyl groups/one-carbon units from dietary sources to substrates. In this regard, methylation can be considered as a reaction between betaine and substrates (dashed line). An increase in the levels of substrates will increase the demand for betaine rather than for methylation mediators, *e.g.*, folate and vitamin B₁₂ (B₁₂), because betaine is a non-renewable resource, while the mediators can be recycled if there is an adequate supply of methyl donors. Pathway 1: Betaine-dependent homocysteine (Hcy) remethylation; Pathway 2: Folate-dependent Hcy remethylation. BHMT: Betaine-homocysteine-methyltransferase; CAs: Catecholamines; CH₂-THF: 5,10-methylene tetrahydrofolate; CH₃: Methyl groups; MeFox: An oxidation product of 5-methyltetrahydrofolate; MS: Methionine synthase; MTs: Methyltransferases; SAH: S-adenosylmethionine; SAM: S-adenosylmethionine; THF: Tetrahydrofolate.

methylation, niacin fortification/supplementation (usually using its nicotinamide form) increases the demand for methyl groups on the one hand, and on the other hand, it can reduce the utilization of choline as a methyl donor by causing hepatic and renal oxidative injury, as demonstrated in a rat model^[42]. As a result, excess nicotinamide reduces the size of betaine pool and subsequently inhibits the methylation of endogenous substrates (*e.g.*, catecholamines and DNA), leading to an increase in plasma norepinephrine levels^[43] and DNA hypomethylation, an important epigenetic alteration in human diseases^[42,44].

Relationship between vitamin excretion and degradation

There is close cooperation between the excretory system and the drug-metabolizing enzyme system in maintaining vitamin homeostasis. If the body's excretory capacity is too low to effectively eliminate excess vitamins, the activity/expression of the drug-metabolizing enzyme system will compensatorily increase due to induction by their substrates^[45]. Blacks have a lower sweat rate^[2], but have a higher drug/vitamin-metabolizing activity than whites^[46]. For example, compared with whites, blacks have a significantly higher catechol-*O*-methyltransferase (a phase II enzyme that converted norepinephrine to epinephrine)^[47] activity and norepinephrine clearance rate^[48] and, during exercise

stress, they show lower venous plasma norepinephrine and higher epinephrine^[49]. Blacks are prone to low fasting serum vitamin D and folate levels^[12,50] and need a higher vitamin D doses to achieve a desired serum 25-hydroxyvitamin D concentration^[51]. This suggests an increase in plasma vitamin clearance. Given that the levels of plasma and urinary vitamin metabolites are linked to vitamin intake and that vitamins can induce their own degrading enzymes, the findings that increased activity/expression of drug-metabolizing enzymes (*e.g.*, CYP2E1^[13,52] and NNMT^[16]) and high levels of vitamin metabolites (*e.g.*, MeFox^[12], *N*¹-methylnicotinamide^[15,16] and nicotinuric acid^[53]) can be considered as increased compensation for decreased vitamin excretion in response to high vitamin intake.

The degradation of vitamins is accompanied by the generation of reactive oxygen species (ROS). Although ROS at physiological levels functions as signalling molecules, at large levels they can induce cellular toxicity and insulin resistance. In our previous study, we found that co-administration of nicotinamide and glucose (like grain fortification with niacin) can induce insulin resistance due to excess ROS and subsequent reactive hypoglycaemia, demonstrating that vitamin-fortified grains can increase appetite^[2,5]. This may explain the sharp increase in prevalence of obesity in the United States after the levels of vitamin fortification

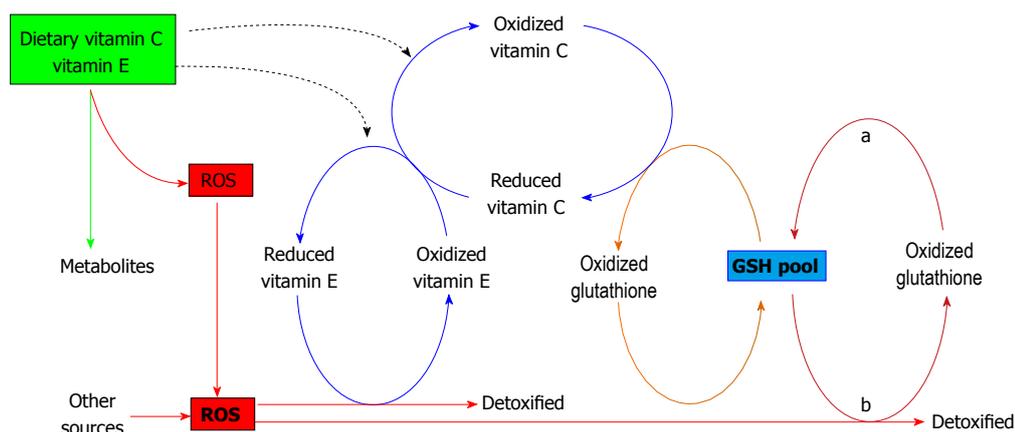


Figure 2 Glutathione-vitamin C-vitamin E interrelationship in the detoxification of reactive oxygen species. The endogenous glutathione antioxidant system maintains vitamin C and vitamin E recycling and actually determines the antioxidant effect of these vitamins. GSH: Reduced glutathione; a: Glutathione reductase; b: Glutathione peroxidase; ROS: Reactive oxygen species.

were increased in 1974^[4,5]. Because decreased sweat excretion may increase enzymatic vitamin degradation and thereby ROS generation, individuals with reduced excretory capacity are at increased risk of insulin resistance, obesity and related diseases when exposed to identical high-vitamin diets.

As shown in Figure 2, it is clear that although vitamin E and C can scavenge ROS, their antioxidant effect actually depends on the capacity of the endogenous glutathione antioxidant system, by which vitamin C and vitamin E recycling is maintained^[54]. Because the endogenous glutathione antioxidant system *per se* directly scavenges free radicals, high levels of supplementation of vitamin C and vitamin E are not only unnecessary but harmful due to increasing the burden of the glutathione antioxidant system. It is obvious that excess vitamin intake may provide an additional source of ROS. Thus, it is not surprising that some randomized clinical trials show that high-dosage vitamin E supplementation may increase, rather than decrease, cardiovascular events and all-cause mortality^[55].

FOLIC ACID FORTIFICATION-INDUCED PARADOX

Although mandatory vitamin fortification has been implemented since the early 1940s and updated in 1974, unfortunately it is hard to determine the relationship between vitamin fortification and the increased prevalence of obesity, mainly because of the lack of studies regarding the effects of vitamin fortification and excess vitamin degradation on the metabolism of the body. Fortunately, the effects of the mandatory folic acid fortification that was started in 1998 in the United States are closely monitored based on the data from National Health and Nutrition Examination Surveys (NHANES). This provides a valuable opportunity for us to understand the vitamin paradox in obesity. The major results of studies on folic acid fortification are summarized as

follows: (1) Blood folate concentrations in the United States population show first a sharp increase from pre- to postfortification (2.5 times for serum and 1.5 times for RBC folate) and then a decline over time (decreased by 17% for serum and 12% for RBC folate during 1999–2010)^[56]; (2) Unmetabolized folic acid was detected in nearly all serum samples measured, and serum unmetabolized folic acid concentrations > 1 nmol/L are associated with being older, non-Hispanic black, nonfasting (< 8 h), higher total folic acid intake (diet and supplements), and higher RBC folate concentrations^[57]; (3) Serum and RBC total folate concentrations, including MeFox (an oxidation product of folate), are high in older adults and individuals with low renal function^[12]; (4) Body mass index is associated negatively with serum unmetabolized folic acid and 5-methyltetrahydrofolate, but positively with serum MeFox and RBC folate concentrations^[12]; (5) Compared with non-Hispanic whites, non-Hispanic blacks have lower serum and RBC total folate concentrations^[12]; (6) In folic acid supplement users, it was found that non-Hispanic black users have lower serum 5-methyltetrahydrofolate concentrations than non-Hispanic-white users^[57]; and (7) Alcohol intake is negatively associated with serum unmetabolized folic acid, 5-methyltetrahydrofolate and MeFox, without significantly affecting RBC folate concentrations^[12].

Evidently, there are significant differences in response to folic acid fortification among the United States population. From the perspective of vitamin homeostasis, the differences may actually reflect differences in folic acid excretion and degradation. Because folic acid is not a natural form of folate, the detection of unmetabolized folic acid in fasting serum suggests a folic acid overload. This overload is more evident in individuals with low excretion capacity, including either low renal function or sweat excretion (in non-Hispanic blacks), or both (in older adults).

The decline in post-fortification serum and RBC folate concentration over time in the United States

population^[56], and the association between increased MeFox levels and decreased renal function^[12] suggests a compensatory increase in folic acid degradation. As mentioned above, blacks may have a higher drug-metabolizing activity to compensate for their reduced sweat excretion. This may account for the finding that non-Hispanic blacks have low serum and RBC total folate concentrations. The association between unmetabolized folic acid concentrations > 1 nmol/L and non-Hispanic blacks^[57] suggests that folic acid intake in this population may exceed their folic acid clearance capacity. Moreover, the low serum 5-methyltetrahydrofolate concentrations in non-Hispanic black users^[57] may suggest a lack of one-carbon donors (due to the increased drug-metabolizing activity in blacks), because the formation of 5-methyltetrahydrofolate consumes one-carbon donors (Figure 1).

Many obesity risk factors, such as being blacks^[11], having a low birth weight/preterm birth^[58], a winter (or cold weather) birth^[59,60], or physical inactivity^[61], are related to decreased sweat-gland function. This is also supported by the finding that an equivalent dose of folic acid (by body weight) caused a greater increase in serum folate in obese than non-obese individuals^[62]. Given that obesity is associated with folate-degrading enzyme CYP2E1^[13,52], the association of increased serum MeFox and RBC folate levels and low fasting serum folate levels in obesity may reflect a severe folic acid overload. From this point of view, the finding that the inverse association between body mass index and serum folate is no longer evident among folic acid supplement users in the United States^[63] can be considered as saturation of the compensatory capacity of the drug-metabolizing enzyme system in obesity.

Ethanol is known to induce drug-metabolizing enzymes^[64,65], including CYP2E1^[66]. This may explain the association between alcohol consumption and low fasting serum folate status. It should be pointed out that alcohol consumption-induced low fasting serum folate does not mean folate deficiency, because there is no significant decrease in RBC folate concentrations^[12].

Overall, four conclusions can be reached: (1) the current folic acid intake of Americans has exceeded their excretory capacity; (2) there is increased compensation for increased folic acid intake, especially in individuals with low excretion capacity; (3) further folic acid supplementation after fortification can saturate the drug metabolizing enzyme system; and (4) the production of MeFox suggests that excess folic acid may increase the consumption of one-carbon units (Figure 1) and provide a source of ROS.

MECHANISM BEHIND LOW VITAMIN D STATUS

There is also a paradox after vitamin D is used in fortification and as a supplement. Vitamin D, although considered a vitamin, can be produced in the skin by

sun exposure. Numerous studies have documented an association between low serum concentrations of 25-hydroxyvitamin D and many non-skeletal disorders. Many studies have examined the effect of vitamin D supplementation on the disorders^[67], including obesity^[68], diabetes^[69], hypertension^[70], dyslipidemia^[71], cardiovascular disease^[72], cancer^[73], depression^[74], and asthma^[75]. Unfortunately, most, if not all, of published meta-analyses have failed to show a significant benefit of vitamin D supplementation with or without calcium^[68-75]. It is likely that low fasting serum 25-hydroxyvitamin D status may be not the cause of these diseases.

The skin is a major determinant of 25-hydroxyvitamin D status. Besides synthesizing vitamin D, the skin also functions as a powerful excretory organ^[17]. Notably, the skin functions fluctuate with seasonal temperature fluctuation, with the highest activities in summer and lowest activities in winter. Thus, it is likely that decreased skin excretory function may be a cause of human diseases. In fact, although not directly focusing on the excretory function of the skin, many studies have suggested a direct link of between the levels of plasma compounds and skin excretory function. For example, sebum excretion decreases in winter^[76,77] and inhibition of sebum excretion increases the levels of blood triglycerides and cholesterol^[78]. Sweat-inhibiting factors (e.g., acute cold exposure^[79,80]) increases plasma norepinephrine levels. Decreased sweating function is found to be closely linked to diseases, for example, skin conductance non-response in schizophrenia and depression^[81], low skin conductance in hypertension^[82] and type 2 diabetes^[83], and the association between psoriasis and metabolic syndrome^[84]. Moreover, many well-known chronic disease risk factors, such as being of black origin, having a preterm birth or winter birth, or physical inactivity, are associated with decreased skin excretory function, as mentioned above. Taken together, it can be concluded that decreased skin excretory function may play a major role in diseases, and 25-hydroxyvitamin D status may be an indicator of skin excretory function.

Interestingly, there is a graded relationship between vitamin D status and body mass index^[85]. Sadiya *et al.*^[86] found that it is difficult to achieve target levels of 25-hydroxyvitamin D above 75 nmol/L in type 2 diabetic obese subjects with a relatively high daily dose of vitamin D₃. Recently, Didriksen *et al.*^[87] performed a 5-year intervention study with vitamin D₃ at a dose of 20000 IU (500 µg) per week vs placebo in subjects with impaired glucose tolerance and/or impaired fasting glucose, and they found that those given vitamin D₃ had significantly higher vitamin D concentration in their adipose tissue (about 6.5 times the placebo group), while their median serum 25-hydroxyvitamin D level only increased from the baseline of 61 to 99 nmol/L. This study clearly demonstrates that large amounts of vitamin D₃ are stored in adipose tissue after vitamin D₃ supplementation, and suggests that overweight and

obese subjects may store more vitamin D than normal-weight subjects because they have larger amounts of adipose tissue. Moreover, vitamin D is known to induce drug-metabolizing enzymes^[88]. Thus, it seems likely that the prevalence of low 25-hydroxyvitamin D status after the introduction of vitamin D fortification may share a similar mechanism to that of low folate status: increased degradation and storage in compensation for excess intake.

THE CLINICAL SIGNIFICANCE OF THE VITAMIN PARADOX

Understanding the vitamin paradox in obesity and related diseases is crucial in determining how to manage the low vitamin status in these diseases. From the above analysis, it is apparent that the vitamin paradox in obesity may be due to increased vitamin degradation and storage in compensation for decreased vitamin excretion. This condition will continue until drug-metabolizing enzymes are saturated by their substrates, in which high expression of vitamin-degrading enzymes and elevated vitamin-metabolite levels may serve as indicators. The vitamin paradox can be resolved by reducing vitamin intake and increasing sweat rates, rather than by giving vitamin supplementation. Indeed, a recent study shows that bariatric surgery (restricting food intake) and exercise are associated with a significant reduction in NNMT expression plasma MNA levels^[16]. This can be explained by decreased niacin intake and increased sweat excretion.

Excess vitamins have three major detrimental effects: (1) increasing ROS generation and subsequently leading to oxidative tissue damage and insulin resistance; (2) disturbing the degradation of neurotransmitters and hormones by competing for drug metabolizing enzymes and detoxification resources; and (3) causing epigenetic changes (*e.g.*, altered DNA methylation) by depleting the body's methyl-group pool^[2,89]. Thus, fortification-induced sustained excess vitamin intake may deplete the drug-metabolizing system (*e.g.*, manifested by high levels of unmetabolized vitamins) and the antioxidant system, and eventually cause a variety of metabolic disorders and oxidative tissue damage. This may play a causal role in the increased prevalence of obesity and related diseases, as hypothesized in our previous work^[2,4,5].

The association between high vitamin intake and chronic diseases can be considered as vitamin poisoning. Vitamin poisoning is dose dependent. For example, high-dosage vitamin E may increase cardiovascular events and all-cause mortality^[55]. Two recent large-scale randomized niacin trials (nicotinic acid, 1500-2000 mg/d) show that nicotinic acid has many adverse effects, including loss of glycaemic control among persons with diabetes, new-onset diabetes^[90,91] and increased risk of death, with borderline statistical significance ($P = 0.08$)^[90]. There are three factors that can increase

the risk of vitamin poisoning: (1) the function of excretory organs is too low to effectively remove excess vitamins from the body, for example, due to early-life malnutrition-induced renal insufficiency^[92]; (2) the amount of vitamin intake has exceeded the excretory capacity of individuals without any developmental defect, which may account for excess chronic diseases in blacks and those with physical inactivity; and (3) the combination of both (1) and (2), accounting for the high rates of chronic diseases in subjects born preterm after the implementation of vitamin fortification. Because the reserve capacity of excretory/detoxifying organs has been determined in early life, whether or not chronic diseases occur will depend on whether there are chemical overloads of the excretory/detoxifying organs in late life. This may be the mechanism of the origin of chronic diseases. Excess vitamin is a kind of chemical overload, accounting for the association between the prevalence of obesity and diabetes and increased B-vitamin intake^[4].

CONCLUSION

In summary, it can be concluded that the vitamin paradox in obesity may be a reflection of excess vitamin intake, rather than a vitamin deficiency. Given that there is a correlation between high vitamin intake and the increased prevalence of obesity, it can be assumed that obesity could be one of manifestations of chronic vitamin poisoning. Susceptible individuals to high vitamin intake are those with a low reserve capacity of excretory organs. Therefore, on an individual basis, prevention of obesity should focus on reducing their intake of vitamin-fortified foods, and for a country, more attention needs to be paid to the role of vitamin fortification and abuse in the increased prevalence of obesity and related diseases.

REFERENCES

- 1 **Barker DJ.** The origins of the developmental origins theory. *J Intern Med* 2007; **261**: 412-417 [PMID: 17444880 DOI: 10.1111/j.1365-2796.2007.01809.x]
- 2 **Zhou SS, Zhou Y.** Excess vitamin intake: An unrecognized risk factor for obesity. *World J Diabetes* 2014; **5**: 1-13 [PMID: 24567797 DOI: 10.4239/wjd.v5.i1.1]
- 3 **Zhou SS, Zhou YM, Li D, Ma Q.** Early infant exposure to excess multivitamin: a risk factor for autism? *Autism Res Treat* 2013; **2013**: 963697 [PMID: 23533752 DOI: 10.1155/2013/963697]
- 4 **Zhou SS, Li D, Zhou YM, Sun WP, Liu QG.** B-vitamin consumption and the prevalence of diabetes and obesity among the US adults: population based ecological study. *BMC Public Health* 2010; **10**: 746 [PMID: 21126339 DOI: 10.1186/1471-2458-10-746]
- 5 **Li D, Sun WP, Zhou YM, Liu QG, Zhou SS, Luo N, Bian FN, Zhao ZG, Guo M.** Chronic niacin overload may be involved in the increased prevalence of obesity in US children. *World J Gastroenterol* 2010; **16**: 2378-2387 [PMID: 20480523 DOI: 10.3748/wjg.v16.i19.2378]
- 6 **Gordon-Larsen P, Wang H, Popkin BM.** Overweight dynamics in Chinese children and adults. *Obes Rev* 2014; **15** Suppl 1: 37-48 [PMID: 24341757 DOI: 10.1111/obr.12121]
- 7 **Zhai F, Wang H, Du S, He Y, Wang Z, Ge K, Popkin BM.**

- Prospective study on nutrition transition in China. *Nutr Rev* 2009; **67** Suppl 1: S56-S61 [PMID: 19453679 DOI: 10.1111/j.1753-4887.2009.00160.x]
- 8 **Kant AK.** Interaction of body mass index and attempt to lose weight in a national sample of US adults: association with reported food and nutrient intake, and biomarkers. *Eur J Clin Nutr* 2003; **57**: 249-259 [PMID: 12571656 DOI: 10.1038/sj.ejcn.1601549]
 - 9 **Pereira-Santos M,** Costa PR, Assis AM, Santos CA, Santos DB. Obesity and vitamin D deficiency: a systematic review and meta-analysis. *Obes Rev* 2015; **16**: 341-349 [PMID: 25688659 DOI: 10.1111/obr.12239]
 - 10 **Kerns JC,** Arundel C, Chawla LS. Thiamin deficiency in people with obesity. *Adv Nutr* 2015; **6**: 147-153 [PMID: 25770253 DOI: 10.3945/an.114.007526]
 - 11 **Bird JK,** Ronnenberg AG, Choi SW, Du F, Mason JB, Liu Z. Obesity is associated with increased red blood cell folate despite lower dietary intakes and serum concentrations. *J Nutr* 2015; **145**: 79-86 [PMID: 25527662 DOI: 10.3945/jn.114.199117]
 - 12 **Pfeiffer CM,** Sternberg MR, Fazili Z, Lacher DA, Zhang M, Johnson CL, Hamner HC, Bailey RL, Rader JL, Yamini S, Berry RJ, Yetley EA. Folate status and concentrations of serum folate forms in the US population: National Health and Nutrition Examination Survey 2011-2. *Br J Nutr* 2015; **113**: 1965-1977 [PMID: 25917925 DOI: 10.1017/S0007114515001142]
 - 13 **Hanley MJ,** Abernethy DR, Greenblatt DJ. Effect of obesity on the pharmacokinetics of drugs in humans. *Clin Pharmacokinet* 2010; **49**: 71-87 [PMID: 20067334 DOI: 10.2165/11318100-000000000-00000]
 - 14 **Shane B.** Folate status assessment history: implications for measurement of biomarkers in NHANES. *Am J Clin Nutr* 2011; **94**: 337S-342S [PMID: 21593497 DOI: 10.3945/ajcn.111.013367]
 - 15 **Liu M,** Li L, Chu J, Zhu B, Zhang Q, Yin X, Jiang W, Dai G, Ju W, Wang Z, Yang Q, Fang Z. Serum N¹-methylnicotinamide is associated with obesity and diabetes in Chinese. *J Clin Endocrinol Metab* 2015; **100**: 3112-3117 [PMID: 26066674 DOI: 10.1210/jc.2015-1732]
 - 16 **Kannt A,** Pfenninger A, Teichert L, Tönjes A, Dietrich A, Schön MR, Klötting N, Blüher M. Association of nicotinamide-N-methyltransferase mRNA expression in human adipose tissue and the plasma concentration of its product, 1-methylnicotinamide, with insulin resistance. *Diabetologia* 2015; **58**: 799-808 [PMID: 25596852 DOI: 10.1007/s00125-014-3490-7]
 - 17 **Zhou SS,** Li D, Zhou YM, Cao JM. The skin function: a factor of anti-metabolic syndrome. *Diabetol Metab Syndr* 2012; **4**: 15 [PMID: 22537765 DOI: 10.1186/1758-5996-4-15]
 - 18 **Epstein M.** Aging and the kidney. *J Am Soc Nephrol* 1996; **7**: 1106-1122 [PMID: 8866401]
 - 19 **Fenske NA,** Lober CW. Structural and functional changes of normal aging skin. *J Am Acad Dermatol* 1986; **15**: 571-585 [PMID: 3534008]
 - 20 **Institute of Medicine (US).** Standing Committee on the Scientific Evaluation of Dietary Reference Intakes and its Panel on Folate, O. B. V., and Choline. Dietary Reference Intakes for Thiamin, Riboflavin, Niacin, Vitamin B₆, Folate, Vitamin B₁₂, Pantothenic Acid, Biotin, and Choline. National Academies Press (US), 1998
 - 21 **Mroczek JE,** Jolley RL, Young DS, Turner WJ. Metabolic response of humans to ingestion of nicotinic acid and nicotinamide. *Clin Chem* 1976; **22**: 1821-1827 [PMID: 135660]
 - 22 **Cornbleet T,** Kirch ER, Geim O, Solomon JD. Excretion of thiamine, riboflavin, niacin and pantothenic acid in human sweat. *JAMA Intern Med* 1943; **122**: 426-429 [DOI: 10.1001/jama.1943.02840240016006]
 - 23 **Johnson BC,** Hamilton TS, Mitchell HH. The excretion of "folic acid" through the skin and in the urine of normal individuals. *J Biol Chem* 1945; **159**: 425-429
 - 24 **Zhou SS,** Li D, Sun WP, Guo M, Lun YZ, Zhou YM, Xiao FC, Jing LX, Sun SX, Zhang LB, Luo N, Bian FN, Zou W, Dong LB, Zhao ZG, Li SF, Gong XJ, Yu ZG, Sun CB, Zheng CL, Jiang DJ, Li ZN. Nicotinamide overload may play a role in the development of type 2 diabetes. *World J Gastroenterol* 2009; **15**: 5674-5684 [PMID: 19960564 DOI: 10.3748/wjg.15.5674]
 - 25 **El-Sheikh M,** Keiley M, Hinnant JB. Developmental trajectories of skin conductance level in middle childhood: sex, race, and externalizing behavior problems as predictors of growth. *Biol Psychol* 2010; **83**: 116-124 [PMID: 19945501 DOI: 10.1016/j.biopsycho.2009.11.009]
 - 26 **Dill DB,** Yusef MK, Goldman A, Hillyard SD, Davis TP. Volume and composition of hand sweat of White and Black men and women in desert walks. *Am J Phys Anthropol* 1983; **61**: 67-73 [PMID: 6869514]
 - 27 **Hernes KG,** Mørkrid L, Fremming A, Ødegården S, Martinsen ØG, Storm H. Skin conductance changes during the first year of life in full-term infants. *Pediatr Res* 2002; **52**: 837-843 [PMID: 12438658 DOI: 10.1203/00006450-200212000-00005]
 - 28 **Gladman G,** Chiswick ML. Skin conductance and arousal in the newborn. *Arch Dis Child* 1990; **65**: 1063-1066 [PMID: 2241228]
 - 29 **Silverwood RJ,** Pierce M, Hardy R, Sattar N, Whincup P, Ferro C, Savage C, Kuh D, Nitsch D. Low birth weight, later renal function, and the roles of adulthood blood pressure, diabetes, and obesity in a British birth cohort. *Kidney Int* 2013; **84**: 1262-1270 [PMID: 23760284 DOI: 10.1038/ki.2013.223]
 - 30 **Collins MN,** Brawley CB, McCracken CE, Shankar PR, Schechter MS, Rogers BB. Risk factors for quantity not sufficient sweat collection in infants 3 months or younger. *Am J Clin Pathol* 2014; **142**: 72-75 [PMID: 24926088 DOI: 10.1309/AJCLPHG2BUBVBT5LY]
 - 31 **Kleyn M,** Korzeniewski S, Grigorescu V, Young W, Homnick D, Goldstein-Filbrun A, Schuen J, Nasr S. Predictors of insufficient sweat production during confirmatory testing for cystic fibrosis. *Pediatr Pulmonol* 2011; **46**: 23-30 [PMID: 20812243 DOI: 10.1002/ppul.21318]
 - 32 **Lunze K,** Bloom DE, Jamison DT, Hamer DH. The global burden of neonatal hypothermia: systematic review of a major challenge for newborn survival. *BMC Med* 2013; **11**: 24 [PMID: 23369256 DOI: 10.1186/1741-7015-11-24]
 - 33 **Katsanis J,** Ficken J, Iacono WG, Beiser M. Season of birth and electrodermal activity in functional psychoses. *Biol Psychiatry* 1992; **31**: 841-855 [PMID: 1643198 DOI: 10.1016/0006-3223(92)0316-R]
 - 34 **Lamke LO,** Nilsson GE, Reithner HL. Insensible perspiration from the skin under standardized environmental conditions. *Scand J Clin Lab Invest* 1977; **37**: 325-331 [PMID: 616059]
 - 35 **Williams M,** Cunliffe WJ, Williamson B, Forster RA, Cotterill JA, Edwards JC. The effect of local temperature changes on sebum excretion rate and forehead surface lipid composition. *Br J Dermatol* 1973; **88**: 257-262 [PMID: 4270005]
 - 36 **Cooney RV,** Franke AA, Hankin JH, Custer LJ, Wilkens LR, Harwood PJ, Le Marchand L. Seasonal variations in plasma micronutrients and antioxidants. *Cancer Epidemiol Biomarkers Prev* 1995; **4**: 207-215 [PMID: 7606195]
 - 37 **Lindh JD,** Andersson ML, Eliasson E, Björkhem-Bergman L. Seasonal variation in blood drug concentrations and a potential relationship to vitamin D. *Drug Metab Dispos* 2011; **39**: 933-937 [PMID: 21349923 DOI: 10.1124/dmd.111.038125]
 - 38 **Rajakumar K.** Pellagra in the United States: a historical perspective. *South Med J* 2000; **93**: 272-277 [PMID: 10728513]
 - 39 **Menon RM,** González MA, Adams MH, Tolbert DS, Leu JH, Cefali EA. Effect of the rate of niacin administration on the plasma and urine pharmacokinetics of niacin and its metabolites. *J Clin Pharmacol* 2007; **47**: 681-688 [PMID: 17463214 DOI: 10.1177/0091270007300264]
 - 40 **Wang Z,** Schuetz EG, Xu Y, Thummel KE. Interplay between vitamin D and the drug metabolizing enzyme CYP3A4. *J Steroid Biochem Mol Biol* 2013; **136**: 54-58 [PMID: 22985909 DOI: 10.1016/j.jsmb.2012.09.012]
 - 41 **Koop DR.** Oxidative and reductive metabolism by cytochrome P450 2E1. *FASEB J* 1992; **6**: 724-730 [PMID: 1537462]
 - 42 **Li D,** Tian YJ, Guo J, Sun WP, Lun YZ, Guo M, Luo N, Cao Y, Cao JM, Gong XJ, Zhou SS. Nicotinamide supplementation induces detrimental metabolic and epigenetic changes in developing rats.

- Br J Nutr* 2013; **110**: 2156-2164 [PMID: 23768418 DOI: 10.1017/S0007114513001815]
- 43 **Sun WP**, Li D, Lun YZ, Gong XJ, Sun SX, Guo M, Jing LX, Zhang LB, Xiao FC, Zhou SS. Excess nicotinamide inhibits methylation-mediated degradation of catecholamines in normotensives and hypertensives. *Hypertens Res* 2012; **35**: 180-185 [PMID: 21918528 DOI: 10.1038/hr.2011.151]
- 44 **Tian YJ**, Luo N, Chen NN, Lun YZ, Gu XY, Li Z, Ma Q, Zhou SS. Maternal nicotinamide supplementation causes global DNA hypomethylation, uracil hypo-incorporation and gene expression changes in fetal rats. *Br J Nutr* 2014; **111**: 1594-1601 [PMID: 24507733 DOI: 10.1017/S0007114513004054]
- 45 **Park BK**, Kitteringham NR, Pirmohamed M, Tucker GT. Relevance of induction of human drug-metabolizing enzymes: pharmacological and toxicological implications. *Br J Clin Pharmacol* 1996; **41**: 477-491 [PMID: 8799511 DOI: 10.1046/j.1365-2125.1996.03482.x]
- 46 **Johnson JA**. Ethnic differences in cardiovascular drug response: potential contribution of pharmacogenetics. *Circulation* 2008; **118**: 1383-1393 [PMID: 18809808 DOI: 10.1161/CIRCULATIONAHA.107.704023]
- 47 **McLeod HL**, Fang L, Luo X, Scott EP, Evans WE. Ethnic differences in erythrocyte catechol-O-methyltransferase activity in black and white Americans. *J Pharmacol Exp Ther* 1994; **270**: 26-29 [PMID: 8035323]
- 48 **Ziegler MG**, Mills PJ, Dimsdale J. The effects of race on norepinephrine clearance. *Life Sci* 1991; **49**: 427-433 [PMID: 1865747 DOI: 10.1016/0024-3205(91)90584-X]
- 49 **Walker AJ**, Bassett DR, Duey WJ, Howley ET, Bond V, Torok DJ, Mancuso P. Cardiovascular and plasma catecholamine responses to exercise in blacks and whites. *Hypertension* 1992; **20**: 542-548 [PMID: 1398889 DOI: 10.1161/01.HYP.20.4.542]
- 50 **Harris SS**. Vitamin D and African Americans. *J Nutr* 2006; **136**: 1126-1129 [PMID: 16549493]
- 51 **Aloia JF**, Patel M, Dimaano R, Li-Ng M, Talwar SA, Mikhail M, Pollack S, Yeh JK. Vitamin D intake to attain a desired serum 25-hydroxyvitamin D concentration. *Am J Clin Nutr* 2008; **87**: 1952-1958 [PMID: 18541590]
- 52 **Wang Z**, Hall SD, Maya JF, Li L, Asghar A, Gorski JC. Diabetes mellitus increases the in vivo activity of cytochrome P450 2E1 in humans. *Br J Clin Pharmacol* 2003; **55**: 77-85 [PMID: 12534643 DOI: 10.1046/j.1365-2125.2003.01731.x]
- 53 **Huang CF**, Cheng ML, Fan CM, Hong CY, Shiao MS. Nicotinic acid: a potential marker of metabolic syndrome through a metabolomics-based approach. *Diabetes Care* 2013; **36**: 1729-1731 [PMID: 23275373 DOI: 10.2337/dc12-1067]
- 54 **Rimbach G**, Minihane AM, Majewicz J, Fischer A, Pallauf J, Virgli F, Weinberg PD. Regulation of cell signalling by vitamin E. *Proc Nutr Soc* 2002; **61**: 415-425 [PMID: 12691170 DOI: 10.1079/PNS2002183]
- 55 **Bjelakovic G**, Nikolova D, Gluud LL, Simonetti RG, Gluud C. Antioxidant supplements for prevention of mortality in healthy participants and patients with various diseases. *Cochrane Database Syst Rev* 2012; **3**: CD007176 [PMID: 22419320 DOI: 10.1002/14651858.CD007176.pub2]
- 56 **Pfeiffer CM**, Hughes JP, Lacher DA, Bailey RL, Berry RJ, Zhang M, Yetley EA, Rader JI, Sempos CT, Johnson CL. Estimation of trends in serum and RBC folate in the U.S. population from pre- to postfortification using assay-adjusted data from the NHANES 1988-2010. *J Nutr* 2012; **142**: 886-893 [PMID: 22437563 DOI: 10.3945/jn.111.156919]
- 57 **Pfeiffer CM**, Sternberg MR, Fazili Z, Yetley EA, Lacher DA, Bailey RL, Johnson CL. Unmetabolized folic acid is detected in nearly all serum samples from US children, adolescents, and adults. *J Nutr* 2015; **145**: 520-531 [PMID: 25733468 DOI: 10.3945/jn.114.201210]
- 58 **Casey PH**, Bradley RH, Whiteside-Mansell L, Barrett K, Gossett JM, Simpson PM. Evolution of obesity in a low birth weight cohort. *J Perinatol* 2012; **32**: 91-96 [PMID: 21660083 DOI: 10.1038/jp.2011.75]
- 59 **Wattie N**, Ardern CI, Baker J. Season of birth and prevalence of overweight and obesity in Canada. *Early Hum Dev* 2008; **84**: 539-547 [PMID: 18280062 DOI: 10.1016/j.earlhumdev.2007.12.010]
- 60 **Phillips DI**, Young JB. Birth weight, climate at birth and the risk of obesity in adult life. *Int J Obes Relat Metab Disord* 2000; **24**: 281-287 [PMID: 10757620]
- 61 **Liou TH**, Pi-Sunyer FX, Laferrère B. Physical disability and obesity. *Nutr Rev* 2005; **63**: 321-331 [PMID: 16295145 DOI: 10.1111/j.1753-4887.2005.tb00110.x]
- 62 **Stern SJ**, Matok I, Kapur B, Koren G. A comparison of folic acid pharmacokinetics in obese and nonobese women of childbearing age. *Ther Drug Monit* 2011; **33**: 336-340 [PMID: 21572389 DOI: 10.1097/FTD.0b013e318219407a]
- 63 **Tinker SC**, Hamner HC, Berry RJ, Bailey LB, Pfeiffer CM. Does obesity modify the association of supplemental folic acid with folate status among nonpregnant women of childbearing age in the United States? *Birth Defects Res A Clin Mol Teratol* 2012; **94**: 749-755 [PMID: 22641603 DOI: 10.1002/bdra.23024]
- 64 **Lieber CS**, Lasker JM, Alderman J, Leo MA. The microsomal ethanol oxidizing system and its interaction with other drugs, carcinogens, and vitamins. *Ann N Y Acad Sci* 1987; **492**: 11-24 [PMID: 3474921 DOI: 10.1111/j.1749-6632.1987.tb48649.x]
- 65 **Schnellmann RG**, Wiersma DA, Randall DJ, Smith TL, Sipes IG. Hepatic mixed function oxygenase activity and glutathione S-transferase activity in mice following ethanol consumption and withdrawal. *Toxicology* 1984; **32**: 105-116 [PMID: 6540493 DOI: 10.1016/0300-483X(84)90130-6]
- 66 **Lu Y**, Cederbaum AI. CYP2E1 and oxidative liver injury by alcohol. *Free Radic Biol Med* 2008; **44**: 723-738 [PMID: 18078827 DOI: 10.1016/j.freeradbiomed.2007.11.004]
- 67 **Autier P**, Boniol M, Pizot C, Mullie P. Vitamin D status and ill health: a systematic review. *Lancet Diabetes Endocrinol* 2014; **2**: 76-89 [PMID: 24622671 DOI: 10.1016/S2213-8587(13)70165-7]
- 68 **Pathak K**, Soares MJ, Calton EK, Zhao Y, Hallett J. Vitamin D supplementation and body weight status: a systematic review and meta-analysis of randomized controlled trials. *Obes Rev* 2014; **15**: 528-537 [PMID: 24528624 DOI: 10.1111/obr.12162]
- 69 **Seida JC**, Mitri J, Colmers IN, Majumdar SR, Davidson MB, Edwards AL, Hanley DA, Pittas AG, Tjosvold L, Johnson JA. Clinical review: Effect of vitamin D supplementation on improving glucose homeostasis and preventing diabetes: a systematic review and meta-analysis. *J Clin Endocrinol Metab* 2014; **99**: 3551-3560 [PMID: 25062463 DOI: 10.1210/je.2014-2136]
- 70 **Beveridge LA**, Struthers AD, Khan F, Jorde R, Scragg R, Macdonald HM, Alvarez JA, Boxer RS, Dalbeni A, Gepner AD, Isbel NM, Larsen T, Nagpal J, Petchey WG, Stricker H, Strobel F, Tangpricha V, Toxqui L, Vaquero MP, Wamberg L, Zittermann A, Witham MD. Effect of vitamin D supplementation on blood pressure: a systematic review and meta-analysis incorporating individual patient data. *JAMA Intern Med* 2015; **175**: 745-754 [PMID: 25775274 DOI: 10.1001/jamainternmed.2015.0237]
- 71 **Challoumas D**. Vitamin D supplementation and lipid profile: what does the best available evidence show? *Atherosclerosis* 2014; **235**: 130-139 [PMID: 24835432 DOI: 10.1016/j.atherosclerosis.2014.04.024]
- 72 **Challoumas D**, Stavrou A, Pericleous A, Dimitrakakis G. Effects of combined vitamin D-calcium supplements on the cardiovascular system: should we be cautious? *Atherosclerosis* 2015; **238**: 388-398 [PMID: 25558033 DOI: 10.1016/j.atherosclerosis.2014.12.050]
- 73 **Chung M**, Lee J, Terasawa T, Lau J, Trikalinos TA. Vitamin D with or without calcium supplementation for prevention of cancer and fractures: an updated meta-analysis for the U.S. Preventive Services Task Force. *Ann Intern Med* 2011; **155**: 827-838 [PMID: 22184690 DOI: 10.7326/0003-4819-155-12-201112200-00005]
- 74 **Gowda U**, Mutowo MP, Smith BJ, Wluka AE, Renzaho AM. Vitamin D supplementation to reduce depression in adults: meta-analysis of randomized controlled trials. *Nutrition* 2015; **31**: 421-429 [PMID: 25701329 DOI: 10.1016/j.nut.2014.06.017]
- 75 **Fares MM**, Alkhaled LH, Mroueh SM, Akl EA. Vitamin D supplementation in children with asthma: a systematic review and

- meta-analysis. *BMC Res Notes* 2015; **8**: 23 [PMID: 25643669 DOI: 10.1186/s13104-014-0961-3]
- 76 **Piérard-Franchimont C**, Piérard GE, Kligman A. Seasonal modulation of sebum excretion. *Dermatologica* 1990; **181**: 21-22 [PMID: 2394299]
- 77 **Robinson D**, Bevan EA, Hinohara S, Takahashi T. Seasonal variation in serum cholesterol levels—evidence from the UK and Japan. *Atherosclerosis* 1992; **95**: 15-24 [PMID: 1642688 DOI: 10.1016/0021-9150(92)90171-C]
- 78 **Bershad S**, Rubinstein A, Paterniti JR, Le NA, Poliak SC, Heller B, Ginsberg HN, Fleischmajer R, Brown WV. Changes in plasma lipids and lipoproteins during isotretinoin therapy for acne. *N Engl J Med* 1985; **313**: 981-985
- 79 **Hiramatsu K**, Yamada T, Katakura M. Acute effects of cold on blood pressure, renin-angiotensin-aldosterone system, catecholamines and adrenal steroids in man. *Clin Exp Pharmacol Physiol* 1984; **11**: 171-179 [PMID: 6378465]
- 80 **Wagner JA**, Horvath SM, Kitagawa K, Bolduan NW. Comparisons of blood and urinary responses to cold exposures in young and older men and women. *J Gerontol* 1987; **42**: 173-179 [PMID: 3819343 DOI: 10.1093/geronj/42.2.173]
- 81 **Schnur DB**, Bernstein AS, Yeager A, Smith S, Bernstein P. The relationship of the skin conductance and finger pulse amplitude components of the orienting response to season of birth in schizophrenia and depression. *Biol Psychiatry* 1995; **37**: 34-41 [PMID: 7893856 DOI: 10.1016/0006-3223(94)00146-T]
- 82 **Kaushik RM**, Mahajan SK, Rajesh V, Kaushik R. Stress profile in essential hypertension. *Hypertens Res* 2004; **27**: 619-624 [PMID: 15750254 DOI: 10.1291/hypres.27.619]
- 83 **Freedman BI**, Bowden DW, Smith SC, Xu J, Divers J. Relationships between electrochemical skin conductance and kidney disease in Type 2 diabetes. *J Diabetes Complications* 2014; **28**: 56-60 [PMID: 24140119 DOI: 10.1016/j.jdiacomp.2013.09.006]
- 84 **Sales R**, Torres T. Psoriasis and metabolic syndrome. *Acta Dermatovenerol Croat* 2014; **22**: 169-174 [PMID: 25230056]
- 85 **Pourshahidi LK**. Vitamin D and obesity: current perspectives and future directions. *Proc Nutr Soc* 2015; **74**: 115-124 [PMID: 25359323 DOI: 10.1017/S0029665114001578]
- 86 **Sadiya A**, Ahmed SM, Carlsson M, Tesfa Y, George M, Ali SH, Siddieg HH, Abusnana S. Vitamin D3 supplementation and body composition in persons with obesity and type 2 diabetes in the UAE: A randomized controlled double-blinded clinical trial. *Clin Nutr* 2015; In press [PMID: 25892603 DOI: 10.1016/j.clnu.2015.02.017]
- 87 **Didriksen A**, Burild A, Jakobsen J, Fuskevåg OM, Jorde R. Vitamin D₃ increases in abdominal subcutaneous fat tissue after supplementation with vitamin D₃. *Eur J Endocrinol* 2015; **172**: 235-241 [PMID: 25661743 DOI: 10.1530/EJE-14-0870]
- 88 **Lindh JD**, Björkhem-Bergman L, Eliasson E. Vitamin D and drug-metabolising enzymes. *Photochem Photobiol Sci* 2012; **11**: 1797-1801 [PMID: 22903070 DOI: 10.1039/c2pp25194a]
- 89 **Zhou SS**, Zhou YM, Li D, Lun YZ. Dietary methyl-consuming compounds and metabolic syndrome. *Hypertens Res* 2011; **34**: 1239-1245 [PMID: 21814217 DOI: 10.1038/hr.2011.133]
- 90 **Landray MJ**, Haynes R, Hopewell JC, Parish S, Aung T, Tomson J, Wallendszus K, Craig M, Jiang L, Collins R, Armitage J. Effects of extended-release niacin with laropiprant in high-risk patients. *N Engl J Med* 2014; **371**: 203-212 [PMID: 25014686 DOI: 10.1056/NEJMoa1300955]
- 91 **Anderson TJ**, Boden WE, Desvigne-Nickens P, Fleg JL, Kashyap ML, McBride R, Probstfield JL. Safety profile of extended-release niacin in the AIM-HIGH trial. *N Engl J Med* 2014; **371**: 288-290 [PMID: 25014706 DOI: 10.1056/NEJMc1311039]
- 92 **McMillen IC**, Robinson JS. Developmental origins of the metabolic syndrome: prediction, plasticity, and programming. *Physiol Rev* 2005; **85**: 571-633 [PMID: 15788706 DOI: 10.1152/physrev.00053.2003]

P- Reviewer: Ji G, Masaki T, Sahu RP

S- Editor: Song XX L- Editor: A E- Editor: Jiao XK



Basic Study

Simvastatin, atorvastatin, and pravastatin equally improve the hemodynamic status of diabetic rats

María J Crespo, José Quidgley

María J Crespo, Departments of Physiology and Anesthesiology, University of Puerto Rico-School of Medicine, San Juan, PR 00936-5067, United States

María J Crespo, José Quidgley, Departments of Physiology, University of Puerto Rico-School of Medicine, San Juan, PR 00936-5067, United States

Author contributions: Crespo MJ and Quidgley J contributed equally to this work.

Supported by MBRS-RISE, No. R25-GM061838; and RCMI, No. G12-RR03051.

Institutional review board statement: Ethics of the study was approved by the Institutional Animal Care and Use Committee of the University of Puerto Rico-Medical Sciences Campus. The protocol number is 2590108.

Institutional animal care and use committee statement: Institutional animal care and use committee: All experiments were approved by the Institutional Animal Care and Use Committee of the University of Puerto Rico-Medical Sciences Campus (Protocol number: 2590108), and adhered to guidelines established by the National Institutes of Health and the American Veterinary Medical Association.

Conflict-of-interest statement: No conflict of interest to disclose.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Dr. María J Crespo, Departments of Physiology and Anesthesiology, University of Puerto Rico-School of Medicine, GPO Box 365067, San Juan, PR 00936-5067,

United States. maria.crespo3@upr.edu
Telephone: +1-787-7530120
Fax: +1-787-7530120

Received: April 25, 2015
Peer-review started: April 28, 2015
First decision: June 9, 2015
Revised: July 28, 2015
Accepted: August 16, 2015
Article in press: August 17, 2015
Published online: August 25, 2015

Abstract

AIM: To investigate if the effect of statins improving cardiovascular (CV) status of diabetics is drug-specific or class-dependent, and the underlying mechanisms involved.

METHODS: We compared the results of daily administration over a four-week period of a low dose (10 mg/kg per day) of atorvastatin (AV), simvastatin (SV), and pravastatin (PV) on cardiac performance in diabetic rats. Echocardiographic variables were tested, as well as systolic blood pressure (SBP), acetylcholine (ACh)-induced relaxation, plasma cholesterol levels, and perivascular fibrosis. Malondialdehyde (MDA) and 4-hydroxyalkenal (4-HAE), and endothelial nitric oxide synthase (eNOS) and inducible nitric oxide synthase (iNOS) protein levels were also measured in cardiac and aortic homogenates.

RESULTS: In untreated diabetic rats, cholesterol levels were higher than in control rats (CT; $n = 8$, $P < 0.05$), and the low dose of statins used did not modify these levels. In diabetic rats, SBP was higher than in CT, and was significantly reduced by all three statins ($n = 10$, $P < 0.05$). Echocardiographic parameters (EF, SV, and COI) were all lower in untreated diabetic rats than in CT ($n = 10$, $P < 0.05$). These CV parameters were equally

improved by all three statins. The maximal relaxation (E_{Max}) induced by ACh in aortic ring from diabetic rats was also improved. Moreover, this relaxation was abolished by 1 mmol/L NG-nitro-L-arginine methyl ester, suggesting the involvement of a NO-dependent mechanism.

CONCLUSION: AV, SV, and PV are equally effective in improving CV performance in diabetic rats. All three statins decreased media thickness, perivascular fibrosis, and both MDA and 4-HAE in the aortas of diabetic rats, without affecting eNOS and iNOS protein levels. The observed hemodynamic benefits are cholesterol-independent. These benefits appear to be secondary to the improved endothelial function, and to the reduced vascular tone and remodeling that result from decreased oxidative stress.

Key words: Statins; Diabetes; Oxidative stress; Cardiac function; Perivascular fibrosis

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Despite evidence that statins are useful therapeutic tools in treating diabetes, questions remain as to whether their effects are drug-specific or class-dependent, what mechanisms underlie these effects, and which statin is the most appropriate. We found that atorvastatin, simvastatin, and pravastatin are equally effective in improving cardiovascular performance in Type 1 diabetic rats, and that the observed benefits are likely to be secondary to the reduction of oxidative stress by these drugs.

Crespo MJ, Quidgley J. Simvastatin, atorvastatin, and pravastatin equally improve the hemodynamic status of diabetic rats. *World J Diabetes* 2015; 6(10): 1168-1178 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v6/i10/1168.htm> DOI: <http://dx.doi.org/10.4239/wjd.v6.i10.1168>

INTRODUCTION

Diabetes is a group of metabolic diseases primarily characterized by hyperglycemia resulting from defects in insulin production, action, or both. This condition has been associated with an increased risk of cardiovascular (CV) deterioration, which is the major cause of death in diabetic patients^[1-3]. CV complications include hypertension, ischemic heart disease, heart failure, and diabetic nephropathy. The etiology of cardiac abnormalities in diabetes has been linked to increased oxidative stress and endothelial dysfunction, although the precise mechanism for these complications remains elusive^[4-6].

The addition of statins, which inhibit 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase, to standard antiglycemic therapies decreases CV complications in diabetic patients. The American

Diabetes Association's "Standards of Medical Care in Diabetes-2015"^[7], recommends the use of statins for all diabetics under 40 years of age with additional CV risk factors, or with overt CV disease. It further recommends that diabetics over the age of 40 take statins, regardless of the absence of CV risk factors. Indeed, in Type 2 diabetics without elevated cholesterol, the risk of suffering the first CV event is reduced by atorvastatin (AV)^[8]. The statin-induced improvement of cardiac function in normo-cholesterolemic patients suggests that these drugs have pleiotropic benefits that may be independent of their ability to lower cholesterol levels^[9,10]. The mechanisms underlying these beneficial effects may include improvement of endothelial function through increased systemic NO bioavailability^[11] and endothelial nitric oxide synthase (eNOS) expression^[12], or through reduced oxidative stress^[13,14].

Despite evidence that statins are useful therapeutic tools in diabetes, questions remain as to whether their effect is drug-specific or class-dependent, which statin is most appropriate, and what mechanisms underlie this effect. In the present study, we compared the effects of three different statins (AV, SV, and PV) on the CV profile of streptozotocin (STZ)-induced diabetic rats that did not receive insulin supplementation. This animal model of Type 1 diabetes is a validated model for the study of diabetic effects on the CV system. At four weeks following diabetic induction, the rats are hypertensive and have decreased cardiac output, stroke volume, and ejection fraction, when compared to age-matched controls (CT)^[14,15]. To evaluate and compare the effects of these statins on cardiac function, we measured stroke volume, ejection fraction, and cardiac output with echocardiography. The effects of statins on endothelial function, cholesterol level, and vascular remodeling were also evaluated. The results from this study may help to identify the most effective statin for improving the CV profile in diabetics.

MATERIALS AND METHODS

Experimental animal model

Four-week-old male Sprague-Dawley rats (120-125 g average weight) were acquired from Hilltop Lab Animals, Inc. (Scottsdale, PA). A total of 160 rats were divided into two groups, diabetic and CT, with each group containing 80 animals. Diabetes was induced by injecting intraperitoneally (IP) streptozotocin (STZ, 65 mg/kg) dissolved in 0.1 mol/L citrate buffer (pH 4.5) after an overnight fast. Diabetic induction was confirmed with positive blood glucose tests twenty-four hours after STZ injection, (Accu-Chek Simplicity, Roche, Indianapolis, IN). Glucose was weekly monitored. The rats did not receive insulin and the experiments were performed at 4 wk after induction of diabetes.

Drug administration

After diabetic induction, each rat was treated daily

with the selected drug (AV or SV or PV) over a four-week period. The statins were suspended in corn oil and administered by gavage at a dose of 10 mg/kg per day. The volumes of all administered drugs were adjusted weekly according to each animal's weight in order to ensure a constant dose. Untreated diabetic and CT groups received by gavage only corn oil, as a placebo. Statin doses were selected based on previous studies on diabetic rats, and from our laboratory^[16,17]. In order to obtain cholesterol level reductions similar to those attained in humans, a 50 mg/kg per day statin administration is needed in rats^[13]. Thus, a low dose of 10 mg/kg per day allowed us to assess the effect of statins on the CV system independently of the benefits derived from cholesterol reduction.

Echocardiographic evaluation

Serial transthoracic echocardiographic evaluations were performed using an ultrasound system with a 7.5 to 9.0 MHz transducer (Sonosite Inc. WA), after anesthesia (30 mg/kg BW, IP), following a previously described protocol^[17,18]. Image analysis was performed using Sitelink Image Manager (Sonosite Inc., Bothell, WA).

Noninvasive measurement of systolic blood pressure

Noninvasive systolic blood pressure (SBP) was evaluated using a RTBP-2000 system (Kent Scientific, Litchfield, CT), and analyzed with Lab View Program (National Instruments Co. Austin, TX) as previously described^[19].

Evaluation of acetylcholine-induced relaxation

To evaluate endothelium-dependent relaxation, aortic rings (5 mm) from the descending thoracic aorta were placed in Krebs' bicarbonate solution (composition in mmol/L: 118 NaCl, 2.5 CaCl₂, 5 KCl, 1.1 MgSO₄, 25 NaHCO₃, 1.2 KH₂PO₄ and 10 glucose, pH = 7.4). The rings were suspended horizontally with a resting tension of 2.0 g, and connected to a FT03C Grass transducer, following the protocol previously described by our laboratory^[19]. The effect of statins on acetylcholine (ACh)-induced relaxation was evaluated in rings pre-contracted with norepinephrine (NE, 1.0 μmol/L). Cumulative concentration-response curves (from 0.1 nmol/L to 10 μmol/L) for ACh were generated after equilibration. An additional dose response curve was then performed after a 45-min incubation period with L-NAME (1 mmol/L). For a particular ACh concentration, the relaxation was expressed as a percentage of the maximal contraction induced by 1.0 μmol/L of NE.

Cholesterol level determination

Blood samples from both untreated and treated diabetic rats and from CT were centrifuged (5000 rpm; 5 min; 4 °C) to measure cholesterol concentration. Total cholesterol levels were quantified a cholesterol quantitation kit (Sigma-Aldrich, MAK043). A SpectraMax Microplate Reader (Molecular Devices, CA) was used to

measure sample absorbance at 570 nm. A calibration curve using cholesterol standards was used to quantify cholesterol levels.

Measurement of malondialdehyde and 4-hydroxyalkenals levels

The effect of statins on lipid peroxidation, a marker of oxidative stress, was evaluated following the previously described protocol^[20]. Malondialdehyde (MDA) and 4-hydroxyalkenals (4-HAE) levels were determined in cardiac and vascular homogenates at an absorbance of 586 nm.

Measurement of media thickness and perivascular fibrosis

Perivascular fibrosis and media thickness from the thoracic aorta from untreated and treated animals were determined to assess the effect of statin treatment. Tissues were stained with Azan-Mallory and Hematoxylin and Eosin (H and E) following the methodology previously described by our laboratory^[20]. Results (in μm) were normalized to body weight.

Western Blot for eNOS and inducible nitric oxide synthase

Western Blot studies were performed using a modified protocol described previously^[21]. Protein samples were separated by electrophoresis in a 6% SDS-PAGE gel. Proteins were transferred to a nitrocellulose membrane. Membranes were blocked with 5% Blotto for 1 h. Mouse monoclonal antibodies for eNOS (1:2000 for cardiac tissue, 1:3000 for aortic tissue; BD Biosciences, San Jose, CA), inducible nitric oxide synthase (iNOS) (1:500 for cardiac tissue, 1:750 for aortic tissue; BD Biosciences, San Jose, CA), were added to the membrane after dilution in Blotto, and incubated overnight at 4 °C. The nitrocellulose membranes were incubated with the secondary anti-mouse antibody coupled to Horseradish Peroxidase (HRP) (1:4000; Santa Cruz Biotechnology, Santa Cruz, CA). Before exposure and development, the membranes were incubated with Super Signal West Femto Maximum Sensitivity Substrate (Thermoscientific, Waltham, MA) to enhance the HRP signal derived from the secondary antibody. The Versadoc™ Imaging System and Quantity One Software (Bio-Rad Laboratories, CA) were used to develop and analyze the membranes. eNOS and iNOS levels were standardized by comparison with the β-actin housekeeping gene detected (1:4000; Sigma-Aldrich, St. Louis, MO).

Statistical analysis

All data are expressed as the mean ± SEM (GraphPad Software, Inc., San Diego, CA). Differences between experimental groups were analyzed using Student's *t* and ANOVA, followed by Student-Newman-Keuls test for posthoc analysis. Values were considered statistically significant at a *P* value less than 0.05.

Table 1 Blood glucose (mg/dL) in diabetic and control rats treated with statins

Condition	Day 0	Day 1	Day 7	Day 14	Day 28
CT none	131.25 ± 3.14	130.50 ± 1.51	126.78 ± 4.86	112.42 ± 4.55	133.88 ± 13.66
CT + AV	137.0 ± 5.86	123.33 ± 2.85	126.8 ± 2.2	128.40 ± 7.02	180.67 ± 52.21
CT + SV	143.13 ± 1.75	128.50 ± 3.10	126.80 ± 2.22	128.40 ± 7.02	152.88 ± 18.46
CT + PV	142.63 ± 5.79	127.13 ± 3.36	122.20 ± 4.79	120.80 ± 4.47	130.63 ± 4.06
Diabetic none	133.65 ± 3.51	445.41 ± 24.11 ^a	490.45 ± 34.34 ^a	530.09 ± 26.65 ^a	517.76 ± 18.11 ^a
Diabetic + AV	133.00 ± 3.30	473.82 ± 40.23 ^a	497.38 ± 47.68 ^a	485.38 ± 48.73 ^a	500.73 ± 32.65 ^a
Diabetic + SV	133.75 ± 2.70	413.19 ± 21.22 ^a	473.69 ± 27.39 ^a	483.23 ± 39.90 ^a	498.94 ± 30.62 ^a
Diabetic + PV	126.88 ± 2.08	430.44 ± 27.31 ^a	524.38 ± 19.92 ^a	564.85 ± 13.57 ^a	557.25 ± 12.92 ^a

^a*P* < 0.05 vs age-matched C. Values are means ± SEM. Rats were injected with STZ on day 0. Blood glucose for diabetic rats: *n* = average of 10 rats per group. AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; CT: Control; STZ: Streptozotocin.

Table 2 Body weight (g) of diabetic and control rats treated with statins

Condition	Day 0	Day 1	Day 7	Day 14	Day 28
CT none	179.74 ± 4.02	182.28 ± 4.15	242.78 ± 6.18	304.06 ± 13.26	391.34 ± 9.80
CT + AV	167.17 ± 4.38	172.97 ± 4.53	234.77 ± 5.27	287.67 ± 4.67	404.10 ± 12.46
CT + SV	181.89 ± 8.26	184.19 ± 8.12	248.40 ± 7.99	291.69 ± 6.93	394.14 ± 15.92
CT + PV	193.81 ± 8.25	198.85 ± 7.66	262.99 ± 9.03	301.19 ± 8.02	389.64 ± 11.48
Diabetic none	180.22 ± 5.99	172.53 ± 5.24	198.19 ± 7.41	211.94 ± 11.11 ^a	267.10 ± 27.98 ^a
Diabetic + AV	176.30 ± 3.10	159.86 ± 9.93	202.32 ± 5.13	235.45 ± 8.56 ^a	255.40 ± 17.09 ^a
Diabetic + SV	183.14 ± 4.68	178.24 ± 4.29	207.69 ± 6.81	230.13 ± 7.23 ^a	241.65 ± 12.73 ^a
Diabetic + PV	187.88 ± 5.62	182.58 ± 4.60	204.85 ± 7.18	243.69 ± 5.60 ^a	253.99 ± 11.82 ^a

^a*P* < 0.05 vs age-matched C. Values are means ± SEM. Rats were injected with STZ on day 0. Blood glucose for diabetic rats: *n* = average of 10 rats per group. STZ: Streptozotocin; AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; CT: Control.

Table 3 Total cholesterol levels in plasma from diabetic and control rats after four weeks of statin treatment (10 mg/kg per day)

Condition	Cholesterol (mg/dL)
CT none	156.01 ± 7.32
CT + AV	143.69 ± 14.21
CT + SV	169.86 ± 12.78
CT + PV	155.53 ± 7.08
Diabetic none	248.68 ± 15.78 ^a
Diabetic + AV	233.35 ± 18.44 ^a
Diabetic + SV	234.40 ± 12.11 ^a
Diabetic + PV	235.57 ± 18.20 ^a

Values shown are the means ± SEM of an average of 8 animals per group.

^a*P* < 0.05 vs age-matched treated and untreated CT. AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; CT: Control.

RESULTS

Blood glucose, body weight, and cholesterol levels are shown in Tables 1, 2 and 3. Twenty four-hours after diabetic induction, blood glucose levels were significantly higher in diabetic rats than in CT rats (445.41 ± 24.11 mg/dL vs 130.50 ± 1.51 mg/dL, respectively; *n* = 10, *P* < 0.05; Table 1). This difference was maintained throughout the course of the study and was not affected by the administration of any statin. Body weight increased in both diabetic and CT rats over the course of this study, although it was significantly lower in aged-matched diabetic rats (Table 2). This parameter also was not modified by any statin. Total cholesterol levels were significantly increased in diabetic

rats when compared to aged-matched CT (248.68 ± 15.78 mg/dL vs 156.01 ± 7.3 mg/dL; *n* = 8, *P* < 0.05; Table 3). At 10 mg/kg per day, once again, statins did not modify plasma cholesterol levels in either diabetic or CT rats (*n* = 8, *P* > 0.05).

In diabetic rats, stroke volume (Figure 1A) increased significantly after statin treatment (from 0.20 ± 0.02 mL in untreated, to 0.51 ± 0.06 mL with AV, to 0.47 ± 0.05 mL with SV, and to 0.43 ± 0.05 mL with PV; *n* = 10, *P* < 0.05). In diabetic rats ejection fraction was lower than in CT (Figure 1B; 44.93% ± 3.03% vs 70.67% ± 2.11%; *n* = 10, *P* < 0.05), but also improved after statin treatment (to 59.92% ± 2.98 % with AV, to 60.13% ± 3.55% with SV, and to 56.85% ± 4.45% with PV; *n* = 10, *P* < 0.05). Similarly, cardiac output index (mL/min per 100 g BW) improved after statins treatment in diabetic rats (from 24.74 ± 3.52 in untreated to 57.65 ± 6.59 with AV, to 60.13 ± 4.10 with SV and to 53.25 ± 6.19 with PV; *n* = 10, *P* < 0.05) (Figure 1C).

SBP (Figure 2) was higher in diabetic rats than in CT (116.52 ± 3.81 mmHg in STZ vs 82.72 ± 2.36 mmHg in CT; *n* = 10, *P* < 0.05. Administration of statins significantly reduced this variable in diabetic rats (to 100.91 ± 5.15 mmHg with AV, 93.17 ± 3.31 mmHg with SV, and 106.44 ± 4.21 mmHg with PV; *n* = 10, *P* < 0.05).

The maximal relaxation (E_{Max}) induced by ACh (Figure 3) was significantly reduced in the aortic rings from diabetic rats compared to those from aged-matched CT (53.70% ± 4.07% vs 74.61% ± 3.27%; *n* = 10, *P* < 0.05). This finding confirms that, at four weeks

Table 4 Effect of chronic statin treatment on EC₅₀ and E_{MAX} values following ach-induced relaxation in diabetic and control rats

Condition	E _{max} relaxation, %	EC ₅₀ , μmol/L
CT none	74.61 ± 3.27	0.56 ± 0.11
CT + AV	70.75 ± 3.99	0.68 ± 0.22
CT + SV	70.76 ± 4.16	1.15 ± 0.47
CT + PV	71.16 ± 4.30	0.72 ± 0.20
Diabetic none	53.70 ± 4.07 ^a	0.41 ± 0.10
Diabetic + AV	82.13 ± 7.01 ^c	0.84 ± 0.32
Diabetic + SV	84.63 ± 6.51 ^c	0.40 ± 0.21
Diabetic + PV	83.88 ± 6.83 ^c	0.66 ± 0.35

Values shown are the means ± SEM of an average of 10 animals per group. ^a*P* < 0.05 when *vs* age-matched treated and untreated CT; ^c*P* < 0.05 when *vs* age-matched untreated diabetic rats. No statistically significant differences were found between treated diabetic rats and treated CT. AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; CT: Control.

following diabetes induction, endothelial dysfunction is present in the aorta of diabetic rats. The tested statins significantly improved E_{MAX} values in diabetic rats (82.13% ± 7.01% with AV, 84.63% ± 6.51% with SV, and 83.88% ± 6.83% with PV; *n* = 10, *P* < 0.05), but did not modified this value in CT. Moreover, a 45-min incubation period with 1 mmol/L L-NAME completely abolished the ACh-induced relaxation, indicating that the effect of these statins on vascular relaxation is NO-mediated. EC₅₀ values, by contrast, were not modified by any statin in either diabetic rats or CT (Table 4).

MDA and 4-HAE (μmol/g protein), which are oxidative stress markers were higher in aortic homogenates (Figure 4A) from diabetic rats than in those from CT (6.49 ± 1.24 *vs* 3.69 ± 0.58; *n* = 8, *P* < 0.05). In diabetic rats, but not in CT, all statins significantly reduced MDA and 4-HAE levels (2.69 ± 0.42 with AV, 3.59 ± 0.47 with SV, and 4.03 ± 0.40 with PV; *n* = 8, *P* < 0.05). In cardiac homogenates (Figure 4B), by contrast, MDA and 4-HAE levels were similar in untreated diabetic (1.42 ± 0.12) and CT (1.10 ± 0.12; *n* = 8, *P* > 0.05), and statin treatment did not modify these parameters.

Similar segments of the thoracic aorta from STZ-diabetic rats and CT were investigated to assess the effects of statins on vascular remodeling. In untreated diabetic rats, perivascular fibrosis (Figure 5A) was higher than in CT (10.59 ± 0.40 μm/100 g BW *vs* 4.21 ± 0.22 μm/100 g BW; *n* = 5, *P* < 0.05). All statins reduced perivascular fibrosis in diabetic rats (8.99 ± 0.33 μm/100 g BW with AV, 8.75 ± 0.43 μm/100 g BW with SV, and 9.04 ± 0.39 μm/100 g BW with PV; *n* = 5, *P* < 0.05). Perivascular fibrosis in CT, by contrast, was not modified by any of the statins. In addition, media thickness, which was thicker in diabetic rats than in age-matched CT (49.70 ± 1.10 μm/100 g BW *vs* 46.03 ± 0.67 μm/100 g BW; *n* = 5, *P* < 0.05), was significantly reduced by all the statins in diabetic rats (44.93 ± 0.76 μm/100 g BW with AV, 47.15 ± 0.48 μm/100 g BW with SV, and 46.78 ± 0.67 μm/100 g BW with PV), but

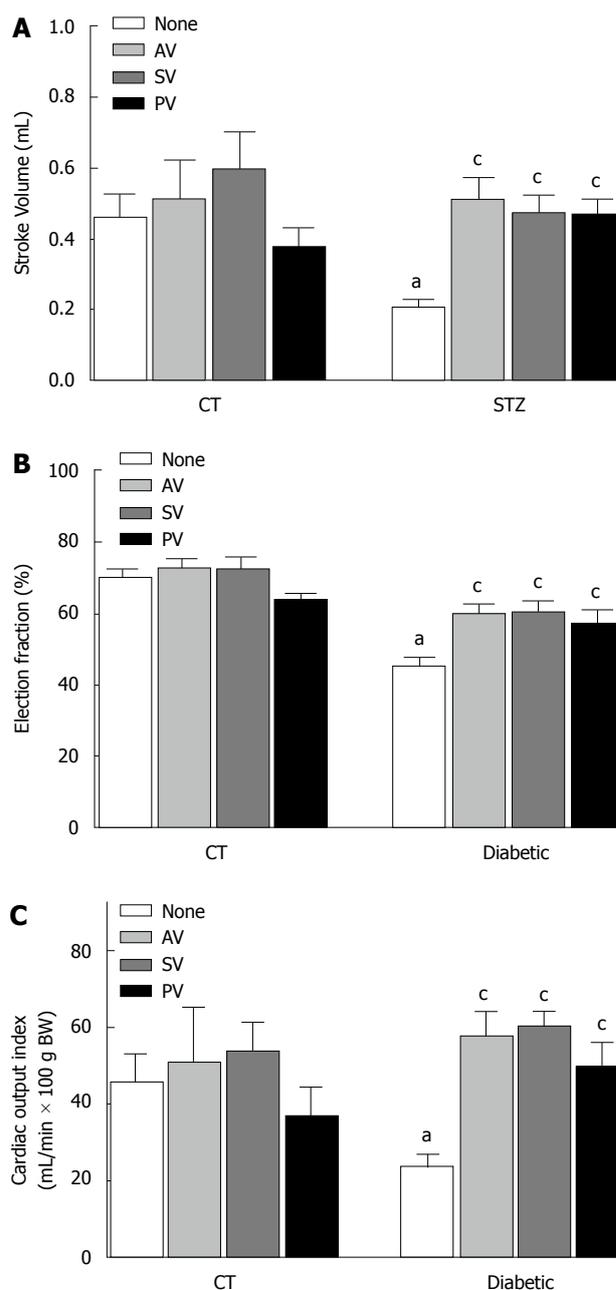


Figure 1 Effects of four weeks treatment with atorvastatin, simvastatin, and pravastatin (10 mg/kg per day) on diabetic rats and control. A: Stroke volume (mL); B: Ejection fraction (%); C: Cardiac output index (mL/min × 100 g BW). The results represent the mean ± SEM of 8 animals per group. All the statins significantly improved these CV parameters in diabetic rats. ^a*P* < 0.05 for diabetic rats *vs* CT; ^c*P* < 0.05 for untreated diabetic rats *vs* treated diabetic rats. STZ: Streptozotocin; AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; CT: Control.

not in CT.

The effect of chronic statin treatment on iNOS and eNOS protein levels (% relative to CT) was evaluated in aortic (Figure 6) and cardiac (Figure 7) tissue from diabetic rats and CT. Comparing the two groups, iNOS levels were similar in aortic tissue (115.40% ± 48.08% in diabetic *vs* 100% in CT; *n* = 5, *P* > 0.05) and in cardiac tissue (155.30% ± 54.47% in diabetic *vs* 100% in CT; *n* = 5, *P* > 0.05). Whereas eNOS levels in cardiac tissue also did not differ (92.16% ± 16.07% in diabetic

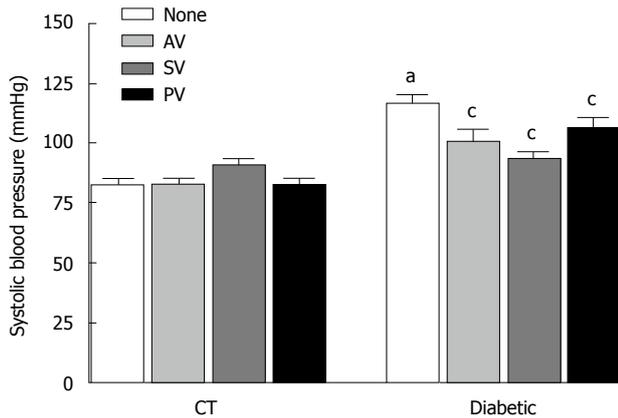


Figure 2 Effects of four weeks treatment with atorvastatin, simvastatin, and pravastatin (10 mg/kg per day) on systolic blood pressure (mmHg) in diabetic rats and control. The values shown are the means \pm SEM of 10 animals per group. All statins significantly decreased blood pressure in diabetic rats. ^a $P < 0.05$ for diabetic rats vs CT; ^c $P < 0.05$ for untreated diabetic rats vs treated diabetic rats. AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; CT: Control.

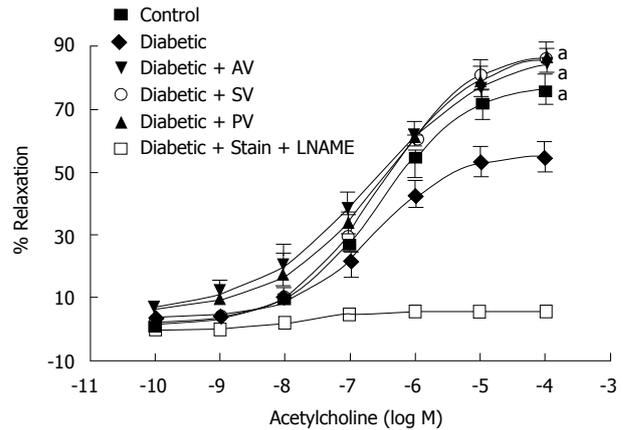


Figure 3 Cumulative concentration response curves for acetylcholine-induced relaxation of aortic rings from diabetic rats after four weeks treatment with atorvastatin, simvastatin, and pravastatin (10 mg/kg per day). Aortic rings were precontracted with 0.1 $\mu\text{mol/L}$ norepinephrine (NE) before the addition of cumulative concentrations of ACh. Note that the addition of 1 mmol/L L-NAME to the incubation bath inhibited ACh-induced relaxation. The values shown are the means \pm SEM of 10 animals per group. ^a $P < 0.05$ for E_{MAX} between untreated diabetic rats and treated diabetic rats. AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; ACh: Acetylcholine.

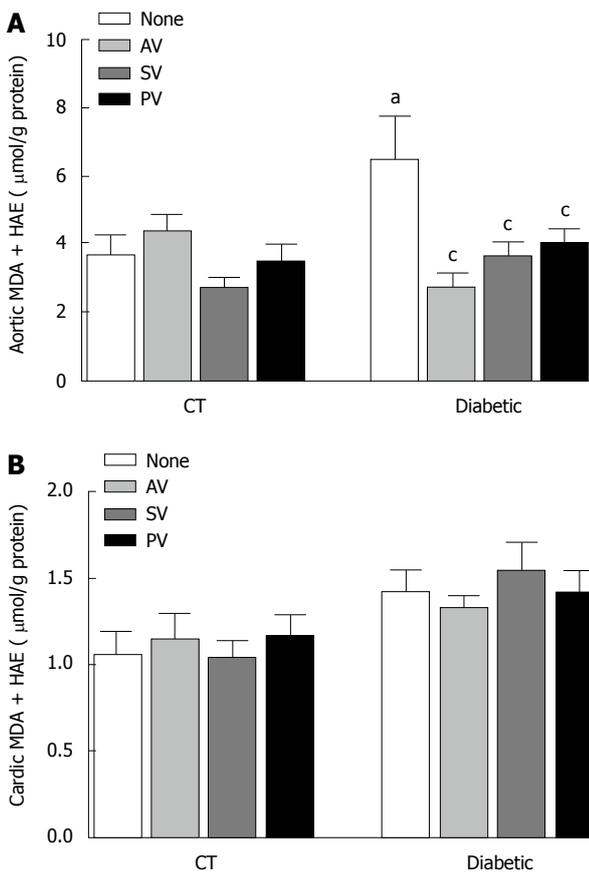


Figure 4 Effect of four weeks treatment with atorvastatin, simvastatin, and pravastatin (10 mg/kg per day) on malondialdehyde + 4-hydroxyalkenal levels in aortic homogenates (A) and in cardiac homogenates (B) from diabetic rats and control. For diabetic rats, all statins equally reduced lipid peroxidation levels in aortic homogenates, but had no effect on these levels in cardiac homogenates. For CT, no effect of statins was observed in either aortic or cardiac homogenates. The values shown are the means \pm SEM of 8 animals per group. ^a $P < 0.05$ for diabetic rats vs CT; ^c $P < 0.05$ for untreated diabetic rats vs treated diabetic rats. AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; CT: Control.

vs 100% in CT; $n = 5$, $P > 0.05$), the levels were reduced in aortic tissue ($54.37\% \pm 7.29\%$ in diabetic vs 100% in CT; $n = 5$, $P < 0.05$). Nevertheless, statin treatment had no effect on either aortic eNOS or iNOS protein levels.

For all tested variables, no significant differences were found between the effects of the three statins. AV, SV, and PV equally improved cardiac function, vascular function, and reduced perivascular fibrosis and oxidative stress.

DISCUSSION

In this study, we compared the effects of AV, SV, and PV on CV performance of Type 1 diabetic rats. For the first time, we report that these three statins similarly improve the CV function of this animal model at a low dose of 10 mg/kg per day. Each statin improves ACh-induced relaxation and CV function, and reduces aortic oxidative stress and remodeling, without lowering cholesterol levels.

In both, patients and animal models of diabetes the beneficial effects of statins improved vascular dysfunction. In diabetic rats, a 50 mg/kg per day dose of AV improves ACh-dependent relaxation^[22]. In spontaneously hypertensive rats, a lower dose of 20 mg/kg also improves vascular function^[13]. Improvements of vascular function are also observed in Type 1 diabetic patients, where both AV (40 mg/d) and PV (40 mg/d per 1 mo) normalize flow-mediated dilatation^[23,24]. Moreover, SV (40 mg/d per 8 wk) improves endothelial-dependent relaxation in hypercholesterolemic patients^[25]. In the current study, we demonstrated that all three statins tested (AV, SV,

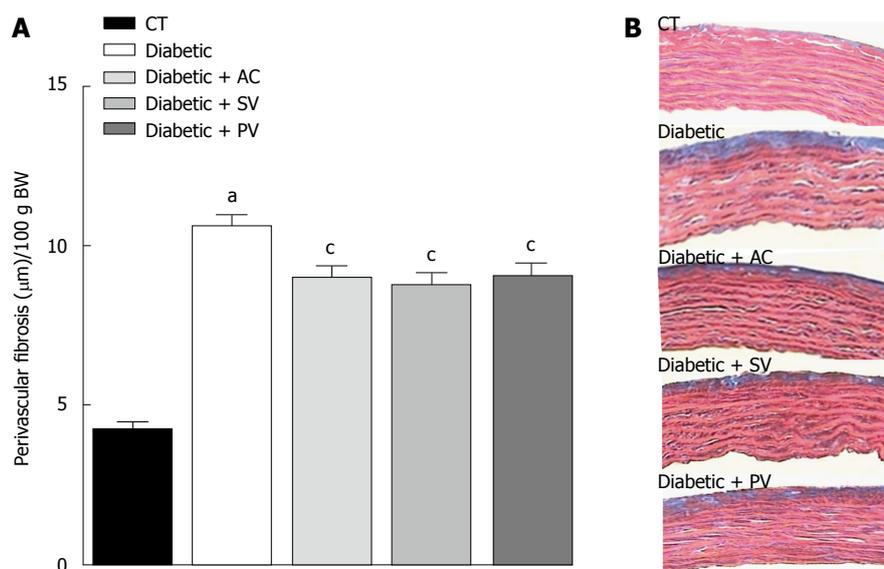


Figure 5 Representative histological sections of aortic segments from untreated and statin-treated diabetic rats, and untreated control. A: Quantified thickness of perivascular fibrosis in comparable aortic segments from treated diabetic rats and untreated diabetic rats. Perivascular fibrosis was higher in untreated diabetic rats than in CT. All statins decreased perivascular fibrosis in diabetic rats. The values shown are the means \pm SEM of 5 animals per group, with the mean value for each animal based on five measurements of its aortic segment. ^a $P < 0.05$ for untreated diabetic rats vs untreated CT; ^c $P < 0.05$ for untreated diabetic rats vs treated diabetic rats; B: Representative histological sections ($\times 40$, Azan-Mallory stain) of aortic segments from untreated diabetic rats and treated diabetic rats, demonstrating the typical reduction in perivascular fibrosis after treatment with each individual statin. AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; CT: Control.

and PV) improve endothelium-dependent relaxation equally in the aortic rings of Type 1 diabetic rats, but at a low dose of only 10 mg/kg per day.

Nevertheless, controversy still exists regarding the beneficial effects of statins on vascular function. For example, among Type 2 diabetic patients with normal cholesterol levels, endothelial function is not restored after the administration of AV (40 or 80 mg/d per 30 wk)^[26], or SV (40 mg/d per 6 wk)^[27]. Similarly PV (40 mg/d per 8 wk) was ineffective in improving endothelial-induced relaxation in patients with coronary heart disease^[28]. The lack of effect of statins in these cases may be due, at least in part, to differences among the experimental models, patient co-morbidities, statin doses, and treatment duration.

The EC₅₀ for the ACh-induced relaxation curves is not modified by any of the three statins tested, indicating that the mechanisms by which these drugs improve endothelial function do not include changes in ACh affinity for the muscarinic receptor. The improvement, however, is fully abolished by L-NAME, suggesting that AV, SV, and PV improve vascular function by increasing NO availability. Whereas all three statins reduce lipid peroxidation markers in the aorta, none modify cardiac or vascular eNOS or iNOS protein levels. Thus, the observed CV improvements at this low dose are most likely secondary to the antioxidant properties of the statins, rather than due to their direct stimulation of NO production. In addition, although the etiology of hypertension is largely unknown, oxidative stress, endothelial dysfunction, and structural alterations of the vasculature have been associated with hypertensive pathophysiology. Thus, the reduction of oxidative stress

and vascular remodeling, together with the improved endothelial dysfunction observed following statin treatment, may underlie the reduced SBP found in diabetic rats.

The results of some studies differ from ours, however. Wenzel *et al.*^[29] found that AV (20 mg/kg per day per 7 wk) decreases eNOS uncoupling in Type 1 diabetic rats. In addition, Ito and colleagues^[30] reported that in the kidney of spontaneously hypertensive rats, AV (20 mg/kg per day per 8 wk) increases eNOS and nNOS expression. Moreover, in endothelial cell cultures from human saphenous vein SV (1 μ mol/L) increases eNOS mRNA and function^[31]. It is possible that statins modify NOS activity and/or expression in a dose-dependent manner. If such is the case, the lack of effect on eNOS and iNOS activity observed in the current study may be due to dosage differences. Alternatively, or in addition, experimental models (*e.g.*, *in vivo* vs *in vitro*) and treatment duration are likely to be major factors underlying this discrepancy.

CV status is deteriorated in diabetic rats by four weeks after induction of diabetes^[19,32]. That AV, SV, and PV equally increasing ejection fraction, stroke volume, and cardiac output suggest that the cardioprotective effect of statins is class-related rather than drug-specific. In addition, this pleiotropic effect appears to be independent of the ability of these drugs to lower cholesterol levels. Improvement of systolic function may result from reductions in peripheral resistance secondary to increased endothelial function, decreased blood pressure, and the vascular remodeling regression observed with all three statins. In line with our results, SV (10 mg/kg per day per 8 wk)

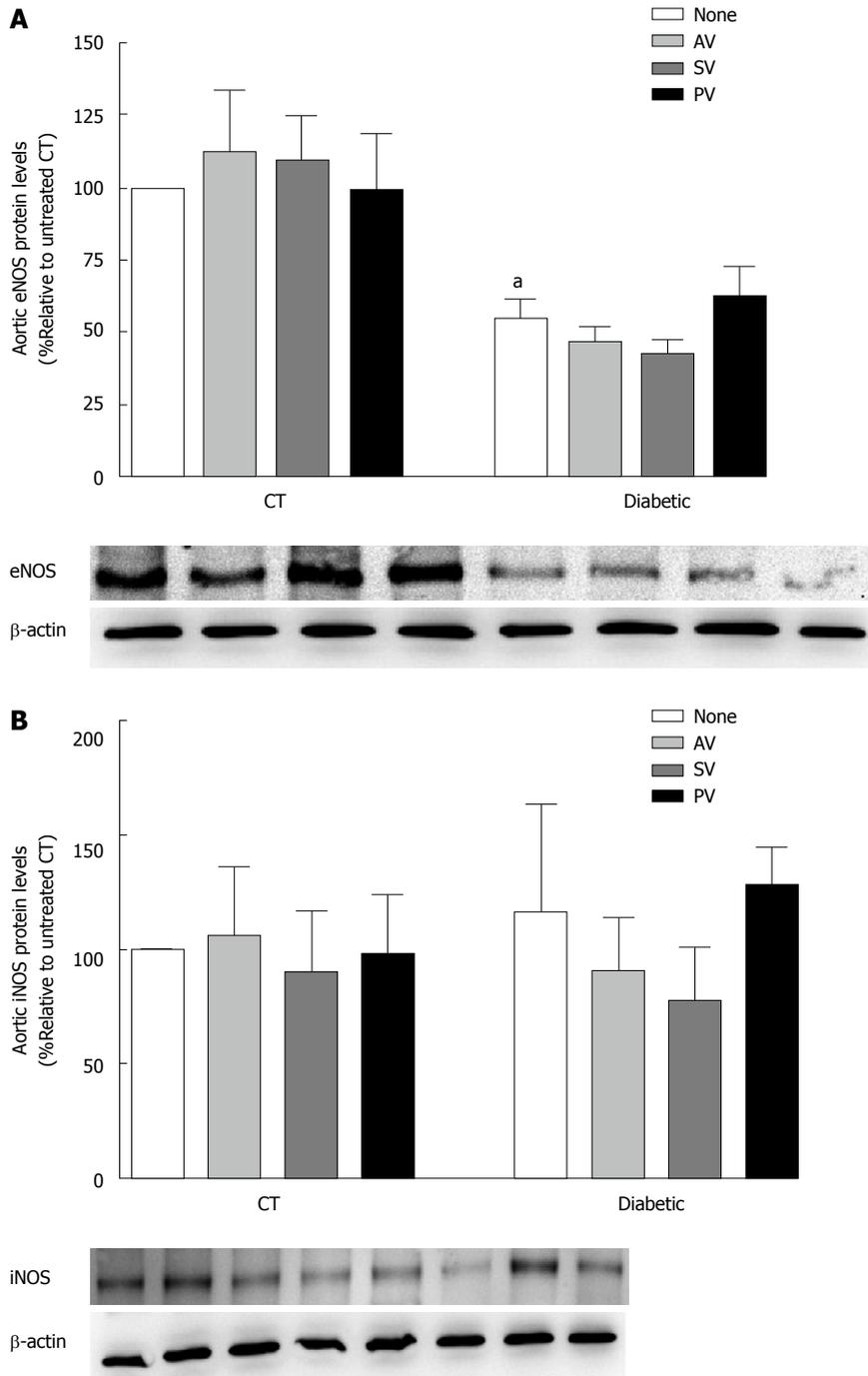


Figure 6 Effect of four weeks treatment with atorvastatin, simvastatin, and pravastatin (10 mg/kg per day) on endothelial nitric oxide synthase (A) and inducible nitric oxide synthase (B) protein levels in aortic homogenates from treated and untreated diabetic rats, and untreated control. Data represent values normalized against β -actin and expressed as percent change relative to untreated CT. The values shown are the means \pm SEM of five animals per group; ^a $P < 0.05$ for untreated diabetic rats vs untreated CT. Bottom: Representative Western blot for eNOS and iNOS of homogenized aortic tissue; AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; CT: Control; eNOS: Endothelial nitric oxide synthase; iNOS: Inducible nitric oxide synthase.

increases ejection fraction and prevents left ventricular hypertrophy and fibrosis in rabbits with non-ischemic heart failure^[33]. Improved vascular function, including augmented ACh-induced relaxation and reduced perivascular fibrosis, may increase cardiac function by reducing total peripheral resistance and reducing cardiac work. Alternatively, the beneficial effects of these statins on cardiac performance may include the preservation of myocardial contractility, which is

deteriorated in diabetes. Indeed, in hearts from diabetic hypercholesterolemic rats, SV (10 mg/kg per day per 5 d) improves cardiac contractility without reducing cholesterol levels^[34]. Statins, however, do not appear to be effective in improving particular aspects of cardiac dysfunction associated with diabetes. The appearance of diastolic dysfunction in Type 2 diabetic rats was not prevented by 100 mg/kg AV^[35]. Furthermore, although AV improves cardiac function, it does not prevent the

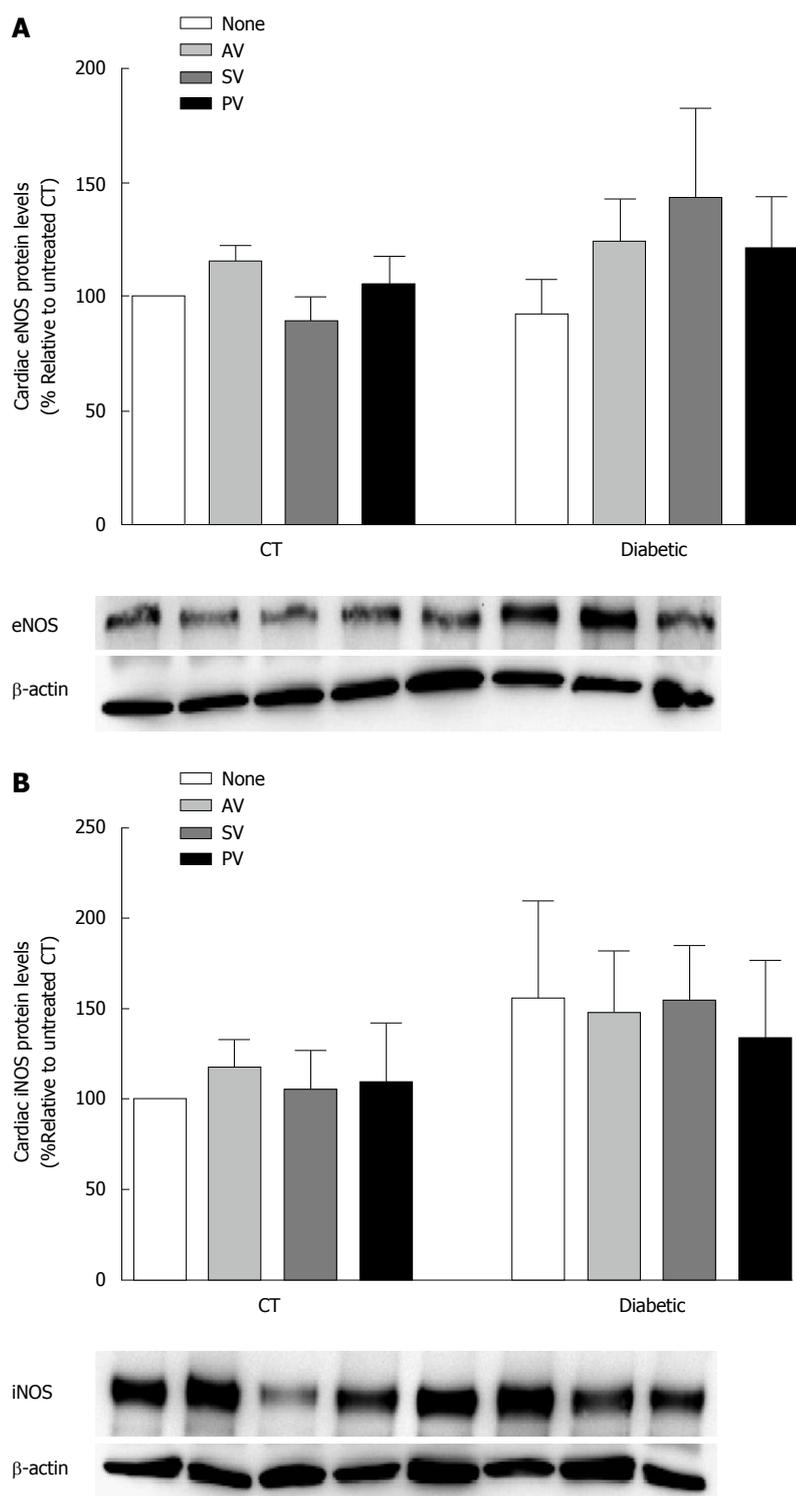


Figure 7 Effect of four weeks treatment with atorvastatin, simvastatin, and pravastatin (10 mg/kg per day) on endothelial nitric oxide synthase (A) and inducible nitric oxide synthase (B) protein levels in cardiac homogenates from treated and untreated diabetic rats, and untreated control. Data represent values normalized against β -actin and expressed as percent change relative to untreated CT. The values shown are the means \pm SEM of five animals per group. No statistically significant differences were found. Bottom: Representative Western blot for eNOS and iNOS of homogenized cardiac tissue; AV: Atorvastatin; SV: Simvastatin; PV: Pravastatin; CT: Control; eNOS: Endothelial nitric oxide synthase; iNOS: Inducible nitric oxide synthase.

onset of cardiomyopathy in Type 1 diabetic rats^[20].

Although the STZ-induced diabetic rat has proven to be an effective animal model for the study of Type 1 diabetes^[36], it has several limitations that must be taken into consideration. Reductions in effective circulating

volume due to glycosuria introduce an additional variable because cardiac and vascular RAS become activated. Autonomic dysfunction, which is present in this model, also may cause a reduction in cardiac vagal tone, without changing sympathetic tone^[37].

Moreover, due to its chemical structure, STZ down-regulates glucose and lipid metabolism genes before hyperglycemia appears, suggesting that this compound can affect gene expression in a hyperglycemia-independent manner^[38]. Despite these limitations, the STZ-diabetic rat is widely used in experimental studies because it replicates both Type 1 diabetes and poorly controlled Type 2 diabetic conditions, making it a useful model in the study of diabetes-related pathophysiology.

The current study demonstrates that AV, SV, and PV are equally effective in improving CV performance in Type 1 diabetic rats. The observed hemodynamic benefits are cholesterol-independent. These benefits appear to be secondary to improved vascular function which, in turn, results from reduced oxidative stress. Although the etiology of Type 1 and Type 2 diabetes is different, in both conditions oxidative stress is high. Thus, it is plausible to postulate that Type 2 diabetics also may benefit from statin treatment. If our findings for diabetic rats are applicable to humans, the benefits of statins to diabetics who are predisposed to develop cardiac complications may extend beyond cholesterol reduction. In addition, even at low doses, statins may be useful for improving the CV profile of diabetics.

COMMENTS

Background

Although there is evidence that statins are useful in the treatment of diabetes, whether cardiovascular (CV) improvement is class-related or drug-specific is unknown. To address the issue, this study tests how low doses of the class-related atorvastatin, simvastatin, and pravastatin improve CV performance in Type 1 diabetic rats.

Research frontiers

Knowledge of the mechanisms underlying statin improvement of CV function, whether these effects are drug-specific or class dependent, and which statin is most effective should result in significant advancements in the current treatment of diabetes.

Innovations and breakthroughs

The beneficial cardioprotective effect of statins is revealed to be class-related, rather than drug-specific. Moreover, this beneficial effect is secondary to reductions in oxidative stress and vascular remodeling, and appears to be independent of the ability of these drugs to lower cholesterol levels.

Applications

If these findings for Type 1 diabetic rats prove to be applicable to humans, the benefits of statins for diabetic patients who are prone to develop CV complications may extend beyond cholesterol reduction.

Terminology

Statins are a class of drugs used to lower cholesterol levels by inhibiting the enzyme 3-hydroxy-3-methylglutaryl-coenzyme A reductase.

Peer-review

The authors concluded the benefits appear to be secondary to the improved endothelial function, and to the reduced vascular tone and remodeling that result from decreased oxidative stress. The findings are interesting.

REFERENCES

1 Lakka HM, Laaksonen DE, Lakka TA, Niskanen LK, Kumpusalo

- E, Tuomilehto J, Salonen JT. The metabolic syndrome and total and cardiovascular disease mortality in middle-aged men. *JAMA* 2002; **288**: 2709-2716 [PMID: 12460094]
- 2 Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol* 2007; **39**: 44-84 [PMID: 16978905]
- 3 Morrish NJ, Wang SL, Stevens LK, Fuller JH, Keen H. Mortality and causes of death in the WHO Multinational Study of Vascular Disease in Diabetes. *Diabetologia* 2001; **44** Suppl 2: S14-S21 [PMID: 11587045]
- 4 Giacco F, Brownlee M. Oxidative stress and diabetic complications. *Circ Res* 2010; **107**: 1058-1070 [PMID: 21030723 DOI: 10.1161/CIRCRESAHA.110.223545]
- 5 Potenza MA, Gagliardi S, Nacci C, Carratu' MR, Montagnani M. Endothelial dysfunction in diabetes: from mechanisms to therapeutic targets. *Curr Med Chem* 2009; **16**: 94-112 [PMID: 19149564]
- 6 Hadi HA, Suwaidi JA. Endothelial dysfunction in diabetes mellitus. *Vasc Health Risk Manag* 2007; **3**: 853-876 [PMID: 18200806]
- 7 American Diabetes Association. Standards of medical care in diabetes-2015. *Diabetes Care* 2015; **38** (Suppl 1): S49-S57
- 8 Colhoun HM, Betteridge DJ, Durrington PN, Hitman GA, Neil HA, Livingstone SJ, Thomason MJ, Mackness MI, Charlton-Menys V, Fuller JH. Primary prevention of cardiovascular disease with atorvastatin in type 2 diabetes in the Collaborative Atorvastatin Diabetes Study (CARDS): multicentre randomised placebo-controlled trial. *Lancet* 2004; **364**: 685-696 [PMID: 15325833]
- 9 Yamagishi S, Imaizumi T. Diabetic vascular complications: pathophysiology, biochemical basis and potential therapeutic strategy. *Curr Pharm Des* 2005; **11**: 2279-2299 [PMID: 16022668]
- 10 Yamagishi S, Matsui T, Nakamura K. Atorvastatin and diabetic vascular complications. *Curr Pharm Des* 2006; **12**: 1549-1554 [PMID: 16611135]
- 11 Schäfer A, Fraccarollo D, Eigenthaler M, Tas P, Firmschild A, Frantz S, Ertl G, Bauersachs J. Rosuvastatin reduces platelet activation in heart failure: role of NO bioavailability. *Arterioscler Thromb Vasc Biol* 2005; **25**: 1071-1077 [PMID: 15761193]
- 12 Huang B, Li FA, Wu CH, Wang DL. The role of nitric oxide on rosuvastatin-mediated S-nitrosylation and translational proteomes in human umbilical vein endothelial cells. *Proteome Sci* 2012; **10**: 43 [PMID: 22799578 DOI: 10.1186/1477-5956-10-43]
- 13 Wassmann S, Laufs U, Bäumer AT, Müller K, Ahlborn K, Linz W, Iter G, Rösen R, Böhm M, Nickenig G. HMG-CoA reductase inhibitors improve endothelial dysfunction in normocholesterolemic hypertension via reduced production of reactive oxygen species. *Hypertension* 2001; **37**: 1450-1457 [PMID: 11408394]
- 14 van Leuven SI, Kastelein JJ. Atorvastatin. *Expert Opin Pharmacother* 2005; **6**: 1191-1203 [PMID: 15957972]
- 15 Crespo MJ, Zalacaín J, Dunbar DC, Cruz N, Arcocho L. Cardiac oxidative stress is elevated at the onset of dilated cardiomyopathy in streptozotocin-diabetic rats. *J Cardiovasc Pharmacol Ther* 2008; **13**: 64-71 [PMID: 18287592 DOI: 10.1177/1074248407307854]
- 16 Ali TK, Al-Gayyar MM, Matragoon S, Pillai BA, Abdelsaid MA, Nussbaum JJ, El-Remessy AB. Diabetes-induced peroxynitrite impairs the balance of pro-nerve growth factor and nerve growth factor, and causes neurovascular injury. *Diabetologia* 2011; **54**: 657-668 [PMID: 20957344 DOI: 10.1007/s00125-010-1935-1]
- 17 Crespo MJ, Cruz N, Quidgley J, Torres H, Hernandez C, Casiano H, Rivera K. Daily administration of atorvastatin and simvastatin for one week improves cardiac function in type 1 diabetic rats. *Pharmacology* 2014; **93**: 84-91 [PMID: 24556594 DOI: 10.1159/000358256]
- 18 Ikeda Y, Martone M, Gu Y, Hoshijima M, Thor A, Oh SS, Peterson KL, Ross J. Altered membrane proteins and permeability correlate with cardiac dysfunction in cardiomyopathic hamsters. *Am J Physiol Heart Circ Physiol* 2000; **278**: H1362-H1370 [PMID: 10749734]
- 19 Crespo MJ, Moreta S, González J. Cardiovascular deterioration in STZ-diabetic rats: possible role of vascular RAS. *Pharmacology* 2003; **68**: 1-8 [PMID: 12660473]

- 20 **Quidley J**, Cruz N, Crespo MJ. Atorvastatin improves systolic function, but does not prevent the development of dilated cardiomyopathy in streptozotocin-induced diabetic rats. *Ther Adv Cardiovasc Dis* 2014; **8**: 133-144 [PMID: 24759610]
- 21 **Cruz-Orengo L**, Figueroa JD, Torrado A, Puig A, Whittemore SR, Miranda JD. Reduction of EphA4 receptor expression after spinal cord injury does not induce axonal regeneration or return of tcMMEP response. *Neurosci Lett* 2007; **418**: 49-54 [PMID: 17418490]
- 22 **Riad A**, Du J, Stiehl S, Westermann D, Mohr Z, Sobirey M, Doehner W, Adams V, Pauschinger M, Schultheiss HP, Tschöpe C. Low-dose treatment with atorvastatin leads to anti-oxidative and anti-inflammatory effects in diabetes mellitus. *Eur J Pharmacol* 2007; **569**: 204-211 [PMID: 17669395]
- 23 **Dogra GK**, Watts GF, Chan DC, Stanton K. Statin therapy improves brachial artery vasodilator function in patients with Type 1 diabetes and microalbuminuria. *Diabet Med* 2005; **22**: 239-242 [PMID: 15717868]
- 24 **Joyce M**, Moore K, Thompson C, Fitzgerald P, Fennessy F, Kelly CJ, Bouchier-Hayes DJ. Hydroxy-methylglutaryl-coenzyme A reductase inhibition improves endothelial dysfunction in type-1 diabetes. *Eur J Vasc Endovasc Surg* 2004; **27**: 432-437 [PMID: 15015196]
- 25 **Güven GS**, Atalar E, Yavuz B, Beyazit Y, Kekilli M, Kilicarslan A, Sahiner L, Oz G, Ozer N, Aksoyek S, Haznedaroglu IC, Sozen T. Simvastatin treatment improves endothelial function and increases fibrinolysis in patients with hypercholesterolemia. *J Natl Med Assoc* 2006; **98**: 627-630 [PMID: 16623076]
- 26 **Tantikosoom W**, Thinkhamrop B, Kiatchusakul S, Jarensiripornkul N, Srinakaran J, Ojongpian S. Randomized trial of atorvastatin in improving endothelial function in diabetics without prior coronary disease and having average cholesterol level. *J Med Assoc Thai* 2005; **88**: 399-406 [PMID: 15962651]
- 27 **van de Ree MA**, Huisman MV, de Man FH, van der Vijver JC, Meinders AE, Blauw GJ. Impaired endothelium-dependent vasodilation in type 2 diabetes mellitus and the lack of effect of simvastatin. *Cardiovasc Res* 2001; **52**: 299-305 [PMID: 11684078]
- 28 **Ling MC**, Ruddy TD, deKemp RA, Ukkonen H, Duchesne L, Higginson L, Williams KA, McPherson R, Beanlands R. Early effects of statin therapy on endothelial function and microvascular reactivity in patients with coronary artery disease. *Am Heart J* 2005; **149**: 1137 [PMID: 15976803]
- 29 **Wenzel P**, Daiber A, Oelze M, Brandt M, Closs E, Xu J, Thum T, Bauersachs J, Ertl G, Zou MH, Förstermann U, Münzel T. Mechanisms underlying recoupling of eNOS by HMG-CoA reductase inhibition in a rat model of streptozotocin-induced diabetes mellitus. *Atherosclerosis* 2008; **198**: 65-76 [PMID: 18061195]
- 30 **Ito D**, Ito O, Mori N, Muroya Y, Cao PY, Takashima K, Kanazawa M, Kohzaki M. Atorvastatin upregulates nitric oxide synthases with Rho-kinase inhibition and Akt activation in the kidney of spontaneously hypertensive rats. *J Hypertens* 2010; **28**: 2278-2288 [PMID: 20724941 DOI: 10.1097/HJH.0b013e32833e0924]
- 31 **Laufs U**, La Fata V, Plutzky J, Liao JK. Upregulation of endothelial nitric oxide synthase by HMG CoA reductase inhibitors. *Circulation* 1998; **97**: 1129-1135 [PMID: 9537338]
- 32 **Crespo MJ**, Marrero M, Cruz N, Quidley J, Creagh O, Torres H, Rivera K. Diabetes alters cardiovascular responses to anaesthetic induction agents in STZ-diabetic rats. *Diab Vasc Dis Res* 2011; **8**: 299-302 [PMID: 21933844 DOI: 10.1177/1479164111421035]
- 33 **Zou C**, Qi H, Liu ZH, Han L, Zhao C, Yang X. Simvastatin activates the PPAR γ -dependent pathway to prevent left ventricular hypertrophy associated with inhibition of RhoA signaling. *Tex Heart Inst J* 2013; **40**: 140-147 [PMID: 23678211]
- 34 **Adameova A**, Harcarova A, Matejikova J, Pancza D, Kuzelova M, Carnicka S, SVEC P, Bartekova M, Styk J, Ravingerová T. Simvastatin alleviates myocardial contractile dysfunction and lethal ischemic injury in rat heart independent of cholesterol-lowering effects. *Physiol Res* 2009; **58**: 449-454 [PMID: 19627175]
- 35 **Chen Y**, Ohmori K, Mizukawa M, Yoshida J, Zeng Y, Zhang L, Shinomiya K, Kosaka H, Kohno M. Differential impact of atorvastatin vs pravastatin on progressive insulin resistance and left ventricular diastolic dysfunction in a rat model of type II diabetes. *Circ J* 2007; **71**: 144-152 [PMID: 17186993]
- 36 **De Angelis K**, Irigoyen MC, Morris M. Diabetes and cardiovascular autonomic dysfunction: application of animal models. *Auton Neurosci* 2009; **145**: 3-10 [PMID: 19054720 DOI: 10.1016/j.autneu.2008.10.013]
- 37 **Souza SB**, Flues K, Paulini J, Mostarda C, Rodrigues B, Souza LE, Irigoyen MC, De Angelis K. Role of exercise training in cardiovascular autonomic dysfunction and mortality in diabetic ovariectomized rats. *Hypertension* 2007; **50**: 786-791 [PMID: 17664387]
- 38 **Kume E**, Aruga C, Ishizuka Y, Takahashi K, Miwa S, Itoh M, Fujimura H, Toriumi W, Kitamura K, Doi K. Gene expression profiling in streptozotocin treated mouse liver using DNA microarray. *Exp Toxicol Pathol* 2005; **56**: 235-244 [PMID: 15816352]

P- Reviewer: Hssan M, Kirali K, Masaki T
S- Editor: Ji FF **L- Editor:** A **E- Editor:** Jiao XK





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

