

# World Journal of *Diabetes*

*World J Diabetes* 2017 February 15; 8(2): 40-88





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2016-2019

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### AIM AND SCOPE

*World Journal of Diabetes* (*World J Diabetes*, *WJD*, online ISSN 1948-9358, DOI: 10.4239), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

*WJD* covers topics concerning  $\alpha$ ,  $\beta$ ,  $\delta$  and PP cells of the pancreatic islet, the effect of insulin and insulinresistance, pancreatic islet transplantation, adipose cells and obesity.

We encourage authors to submit their manuscripts to *WJD*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great clinical significance.

### INDEXING/ABSTRACTING

*World Journal of Diabetes* is now indexed in Emerging Sources Citation Index (Web of Science), PubMed, and PubMed Central.

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#### NAME OF JOURNAL

*World Journal of Diabetes*

#### ISSN

ISSN 1948-9358 (online)

#### LAUNCH DATE

April 15, 2010

#### FREQUENCY

Monthly

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#### PUBLICATION DATE

February 15, 2017

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## Diabetic ketoacidosis: Treatment in the intensive care unit or general medical/surgical ward?

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**Author contributions:** All authors contributed to this paper.

**Conflict-of-interest statement:** None of the authors have any conflict to disclose.

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**Manuscript source:** Invited manuscript

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**Received:** August 16, 2016

**Peer-review started:** August 16, 2016

**First decision:** September 28, 2016

**Revised:** October 11, 2016

**Accepted:** November 21, 2016

**Article in press:** November 22, 2016

**Published online:** February 15, 2017

### Abstract

Diabetic ketoacidosis (DKA) is defined as an acute metabolic disorder, which is characterized by an increased presence of circulating ketones, and the development of ketoacidosis in the presence of hyperglycemia. This syndrome occurs as a result of insulin deficiency. Patients can be dramatically ill, however, with aggressive treatment, most patients recover rapidly. Despite being a low-risk condition, the development of acidosis, is one of the admission criteria to the intensive care unit (ICU) for these patients, in order to provide close monitoring, and recognize complications that could result from the use of aggressive therapy, such as continuous infusions of insulin. In some institutions, DKA is treated in the emergency department and general medical/surgical wards to avoid ICU overcrowding.

**Key words:** Diabetic ketoacidosis; Diabetes; Hyperosmolar non-ketotic state; Clinical outcomes; Serum ketones

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**Core tip:** Diabetic ketoacidosis is a complication for some patients with insulin-dependent diabetes mellitus as well as for non-insulin dependent. It is treated commonly in the intensive care unit (ICU), even though clinical data from many studies support management in regular (medical/surgical) wards, avoiding expensive critical care unit costs and preventing bed crisis in these higher level of care units for sicker patients. Once the patient is treated, adequate follow up and education is mandatory. Noncompliance remains the primary concern for repeated admissions.

Mendez Y, Surani S, Varon J. Diabetic ketoacidosis: Treatment in the intensive care unit or general medical/surgical ward? *World J Diabetes* 2017; 8(2): 40-44 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v8/i2/40.htm> DOI: <http://dx.doi.org/10.4239/wjcd.v8.i2.40>

## INTRODUCTION

Patients with diabetes mellitus (DM) have health care costs 2.3 times higher than others without this diagnosis<sup>[1]</sup>. In a prevalence-based study, by the American Diabetes Association, in the United States in 2012, the total cost for diagnosed DM was \$245 billion United States dollars, and of it, \$176 billion was used for direct medical care costs<sup>[1]</sup>. In addition, and even more concerning, is the fact that hospitalizations for patients with DM have been increasing<sup>[2]</sup>. The National Surveillance of Diabetes Public Health Resources, reported that diabetic ketoacidosis (DKA) admissions increased from 80000/year in 1988 to 140000/year in 2009<sup>[2]</sup>.

DKA causes an acute metabolic disorder, which is primarily characterized by an increased presence of circulating ketone bodies, and the development of severe ketoacidosis in the presence of prolonged uncontrolled hyperglycemia, usually due to insulin deficiency<sup>[3]</sup>. It is more commonly seen in patients with insulin-dependent diabetes mellitus (IDDM), especially among children and young adults. Occasionally, patients with insulin resistant DM can present this complication; especially those that are noncompliant with insulin therapy or who present severe infection<sup>[3]</sup>. DKA has arbitrarily been classified by some as mild, moderate and severe, according to the initial diagnostic criteria (which includes plasma glucose, arterial pH, serum bicarbonate, urine and serum ketones, serum osmolality and anion gap; and the alteration in the mental status)<sup>[4]</sup>.

## EPIDEMIOLOGY

In 2012, 29.1 million Americans or 9.3% of the population were estimated to suffer from DM, according to the American Diabetes Association and the Center for Disease Control and Prevention<sup>[2]</sup>. Of them, approximately 1.25 million American children and adults have IDDM. This clinical condition has a cumulative incidence of 1.4 million Americans per year and it remains the 7<sup>th</sup> leading cause of death in the United States since 2010<sup>[2]</sup>. As noted above, the number of cases of DKA has steadily increased over the past 2 decades<sup>[2,3]</sup>. In one study in the United States, DKA presentations to the emergency department (ED) increased 35% from 1996 to 2006<sup>[3]</sup>. When compared to other countries like England, Austria and Germany, the United States has the highest rates of DKA in children with IDDM<sup>[5]</sup>. Mortality rates for patients with hyperglycemic syndromes (DKA and hyperosmolar non-ketotic states) have been reported as 0.02% in patients with diabetes who are 45 years or younger,

and 0.014% among older adults<sup>[6]</sup>. In some studies, the average length of stay in the hospital for patients with DKA has decreased from 5.7 to 3.4 d, being longer for patients categorized in the "severe" group<sup>[2,7]</sup>. In the authors' experience, some patients can even be discharged within 23 h of hospital admission despite an initial severe acidemia.

## IS DKA A CRITERION FOR ICU

### ADMISSION?

In many institutions, and for decades, DKA has been routinely treated in ICU environments, including recommendations by the American Diabetes Association guidelines for DKA treatment<sup>[3,4,7-9]</sup>. The primary reason for these level of care requirements, has been the presence of severe metabolic acidosis, even if patients are grouped as mild or moderate in severity<sup>[10]</sup>. Frequent blood glucose monitoring, the need for intravenous insulin infusions, and the requirement of frequent vital signs is cited as the hospital structural requirements for this ICU level of care<sup>[11]</sup>. However, several studies have shown that DKA can be safely treated in the ED or even in medical wards (Table 1)<sup>[12-17]</sup>. By taking this lower level of care approach, we can potentially avoid ICU hospitalization rate and higher costs, bed overcrowding and reserving the beds for patients who present complications such as hypotension, coma, acute myocardial ischemia, or those with several comorbidities (*i.e.*, end-stage renal disease, congestive heart failure) and anyone categorized as suffering severe DKA<sup>[12,18,19]</sup>. In some observational studies DKA patients admitted to the ICU have a shorter length of stay when compared to non-diabetic mellitus ICU patients<sup>[20,21]</sup>. A recent retrospective cohort study of 156, 842 hospitalizations among 94 acute-care hospitals, analyzed the adjusted cost of hospitalizations in lower and higher ICU utilizations groups, and concluded that the overuse of ICU only increases the cost and the utilization of invasive procedures but with no improvement in hospital mortality<sup>[22]</sup>.

In a prospective, randomized clinical trial in India, Karoli and coworkers reported that once the DKA patient is evaluated in the ED, and categorized in the severity score, direct admission to a regular ward provided no additional mortality and the only complication noted was hypoglycemia. Other groups have used other classifications to allocate resources for patients with DKA<sup>[15]</sup>. In a retrospective study, Marinac and Mesa, using laboratory criteria (serum bicarbonate, anion gap, base excess and serum osmolality), and diastolic blood pressure, patients were grouped in 5 grades (Grade 0 - IV)<sup>[19]</sup>. ICU admission was recommended only for those who had grade IV DKA<sup>[19]</sup> (Table 2).

## TREATMENT OPTIONS IN THE ED OR ICU

The treatment of acute DKA includes restoration of fluid

**Table 1 Clinical trials comparing care in the intensive care unit *vs* the emergency department or medical ward for patients with diabetic ketoacidosis**

Ref.	Country	Patients enrolled	Site of management	Therapy used	Outcome	Length of stay
Dunbar <i>et al</i> <sup>[12]</sup> Retrospective study (January 1994 - March 1995)	United States	61	15: ICU 46: Regular floor	Not mentioned	Mortality due to sepsis in only 1 patient with initial pH < 7.00	ICU: 2 d Regular floor: Not mentioned
Umpierrez <i>et al</i> <sup>[14]</sup> Prospective randomized open trial	United States	45	15: ICU 30: ED	ICU: Intravenous insulin drip ED: 15 subcutaneous insulin aspart Q1H ED: 15 subcutaneous insulin aspart Q2H	Hypoglycemic event presented in each group in only 1 patient per group. No complications, no recurrence of ketoacidosis and no mortality	ICU: 4.5 ± 3 d ED with SC Q1H: 3.4 ± 3 d ED with SC Q2H: 3.9 ± 3 d
Karoli <i>et al</i> <sup>[15]</sup> Prospective randomized open trial (January 2009 - June 2010)	India	50	25: ICU 25: ED	ICU: 25 intravenous regular insulin ED: 25 subcutaneous insulin lispro	Hypoglycemic event presented, 2 patients in the ICU group and 1 patient in the ED group. No complications, no recurrence of ketoacidosis and no mortality	ICU: 6.6 ± 1.5 d ED: 6.0 ± 1.2 d
Ersöz <i>et al</i> <sup>[16]</sup> Prospective randomized open trial	Turkey	20	20: ICU	ICU: 10 intravenous regular insulin ICU: 10 subcutaneous insulin lispro	No need to switch to IV regular insulin, no hypoglycemic events, no complications, no recurrence of ketoacidosis and no mortality	Not mentioned
Umpierrez <i>et al</i> <sup>[18]</sup> Prospective randomized open trial	United States	20	10: ICU 10: MW	ICU: 20 intravenous regular insulin IMU: 10 subcutaneous insulin lispro Regular floor: 10 subcutaneous insulin lispro	Hypoglycemic event presented in each group in only 1 patient per group, no complications, no recurrence of ketoacidosis and no mortality	IMU and Regular floor: 4 ± 2 d ICU: 4 ± 1 d
Sotiropoulos <i>et al</i> <sup>[25]</sup> Prospective study (June 2007 - May 31 2008)	Greece	21	21: ED	ED: 21 intravenous regular insulin	Myocardial infarction in only 1 patient - Mortality 4.7%	Not mentioned
Della Manna <i>et al</i> <sup>[26]</sup> Controlled clinical trial (June 2001 - June 2003)	Brazil	60	3: ICU 57: ED	ICU: 3 intravenous regular insulin ED: 27 intravenous regular insulin ED: 30 subcutaneous insulin lispro	Hypoglycemic event on 10 patients, 6 patients due to regular insulin and 4 due to lispro; no complications, no recurrence of ketoacidosis and no mortality	Not mentioned

IMU: Intermediate care unit; SC: Subcutaneous; Q1H: Every hour; Q2H: Every two hours; ICU: Intensive care unit; ED: Emergency department; MW: Medical ward; DKA: Diabetic ketoacidosis.

**Table 2 List of conditions requiring admission of patients with diabetic ketoacidosis in the intensive care unit**

Myocardial infarction  
Congestive heart failure  
Acute renal failure  
Acute respiratory failure  
Altered mental status  
Coma  
Shock  
Hypothermia  
Sepsis  
Pancreatitis  
Gastrointestinal bleeding  
Uncontrolled hypertension  
End stage renal disease  
Hyperkalemia

decreased plasma glucose<sup>[23,24]</sup>. As noted above, a few randomized, open label trials have proved good outcome and non-inferiority for patients who are managed on regular medical/surgical wards while using with rapid acting insulin, aspart or lispro<sup>[13,15,17,25-29]</sup>.

By establishing a rapid diagnosis and starting treatment in the ED, clinicians can help patients to decrease their costs and hospital stay.

The primary issue in patients with DKA remains the need for repeated hospital admissions. Non-compliance in these patients makes the outcome and prognosis worst. Indeed, medical non-compliance and adherence to the outpatient treatment is the most common precipitating factor leading to the development of moderate-to-severe DKA, requiring ICU admission secondary to complications (*i.e.*, cerebral edema, sepsis) and making the management in the ED and/or ICU very complex<sup>[21,25,30]</sup>. Life-support care, such



as mechanical ventilation, vasopressors, intravenous antibiotic therapy and mortality rates are higher in these patients, when compared to patients not requiring these interventions<sup>[30]</sup>.

## CONCLUSION

The benefit of ICU level of care for patients with DKA rather than regular medical/surgical wards is not well established for patients with mild-to-moderate DKA. Many studies suggest the utilization of the ED or the regular (medical/surgical) wards in the management of these patients. There is significant cost-benefit in managing DKA in the ED and regular wards instead of the ICU, where only patients that require life-supportive intervention should go. Once patients are discharged from the hospital adequate follow up is necessary to avoid readmissions and assure compliance.

## REFERENCES

- American Diabetes Association.** Economic costs of diabetes in the U.S. in 2012. *Diabetes Care* 2013; **36**: 1033-1046 [PMID: 23468086 DOI: 10.2337/dc12-2625]
- Diabetes Public Health Resource.** Centers of Disease Control and Prevention. [Online]. Available from: URL: <http://www.cdc.gov/diabetes/data/>
- Kitabchi AE, Umpierrez GE, Miles JM, Fisher JN.** Hyperglycemic crises in adult patients with diabetes. *Diabetes Care* 2009; **32**: 1335-1343 [PMID: 19564476 DOI: 10.2337/dc09-9032]
- Kitabchi AE, Umpierrez GE, Murphy MB, Barrett EJ, Kreisberg RA, Malone JJ, Wall BM.** Management of hyperglycemic crises in patients with diabetes. *Diabetes Care* 2001; **24**: 131-153 [PMID: 11194218 DOI: 10.2337/diacare.24.1.131]
- Maahs DM, Hermann JM, Holman N, Foster NC, Kapellen TM, Allgrove J, Schatz DA, Hofer SE, Campbell F, Steigleder-Schweiger C, Beck RW, Warner JT, Holl RW.** Rates of diabetic ketoacidosis: international comparison with 49,859 pediatric patients with type 1 diabetes from England, Wales, the U.S., Austria, and Germany. *Diabetes Care* 2015; **38**: 1876-1882 [PMID: 26283737 DOI: 10.2337/dc15-0780]
- Holman RC, Herron CA, Sinnock P.** Epidemiologic characteristics of mortality from diabetes with acidosis or coma, United States, 1970-78. *Am J Public Health* 1983; **73**: 1169-1173 [PMID: 6412575 DOI: 10.2105/AJPH.73.10.1169]
- Barski L, Nevzorov R, Rabaev E, Jotkowitz A, Harman-Boehm I, Zektser M, Zeller L, Shleyfer E, Almog Y.** Diabetic ketoacidosis: clinical characteristics, precipitating factors and outcomes of care. *Isr Med Assoc J* 2012; **14**: 299-303 [PMID: 22799061]
- Gershengorn HB, Iwashyna TJ, Cooke CR, Scales DC, Kahn JM, Wunsch H.** Variation in use of intensive care for adults with diabetic ketoacidosis\*. *Crit Care Med* 2012; **40**: 2009-2015 [PMID: 22564962 DOI: 10.1097/CCM.0b013e31824e9eae]
- Nyenwe E, Loganathan R, Blum S, Ezuteh D, Erani D, Palace M, Ogugua C.** Admissions for diabetic ketoacidosis in ethnic minority groups in a city hospital. *Metabolism* 2007; **56**: 172-178 [PMID: 17224329 DOI: 10.1016/j.metabol.2006.09.010]
- Smith G, Nielsen M.** ABC of intensive care. Criteria for admission. *BMJ* 1999; **318**: 1544-1547 [PMID: 10356016]
- Hurlock-Chorostecki C.** Managing diabetic ketoacidosis: the role of the ICU nurse in an endocrine emergency. *Dynamics* 2004; **15**: 18-22 [PMID: 15460517]
- Dunbar LM, Gonzaba WT, DeSoto D, Zaheri K, Sisley D, Thompson H.** Aggressive emergency department management of diabetic ketoacidosis can reduce ICU admissions and hospital costs without adversely impacting outcomes. *Ann Emerg Med* 1999; **34**: 83-84 [DOI: 10.1016/S0196-0644(99)80411-9]
- Cohn BG, Keim SM, Watkins JW, Camargo CA.** Does Management of Diabetic Ketoacidosis with Subcutaneous Rapid-acting Insulin Reduce the Need for Intensive Care Unit Admission? *J Emerg Med* 2015; **49**: 530-538 [PMID: 26238182 DOI: 10.1016/j.jemermed.2015.05.016]
- Umpierrez GE, Cuervo R, Karabell A, Latif K, Freire AX, Kitabchi AE.** Treatment of diabetic ketoacidosis with subcutaneous insulin aspart. *Diabetes Care* 2004; **27**: 1873-1878 [PMID: 15277410 DOI: 10.2337/diacare.27.8.1873]
- Karoli R, Fatima J, Salman T, Sandhu S, Shankar R.** Managing diabetic ketoacidosis in non-intensive care unit setting: Role of insulin analogs. *Indian J Pharmacol* 2011; **43**: 398-401 [PMID: 21844993 DOI: 10.4103/0253-7613.83109]
- Ersöz HO, Ukinc K, Köse M, Erem C, Gunduz A, Hacıhasanoglu AB, Karti SS.** Subcutaneous lispro and intravenous regular insulin treatments are equally effective and safe for the treatment of mild and moderate diabetic ketoacidosis in adult patients. *Int J Clin Pract* 2006; **60**: 429-433 [PMID: 16620355 DOI: 10.1111/j.1368-5031.2006.00786.x]
- Savoldelli RD, Farhat SC, Manna TD.** Alternative management of diabetic ketoacidosis in a Brazilian pediatric emergency department. *Diabetol Metab Syndr* 2010; **2**: 41 [PMID: 20550713 DOI: 10.1186/1758-5996-2-41]
- Umpierrez GE, Latif K, Stoevers J, Cuervo R, Park L, Freire AX, E Kitabchi A.** Efficacy of subcutaneous insulin lispro versus continuous intravenous regular insulin for the treatment of patients with diabetic ketoacidosis. *Am J Med* 2004; **117**: 291-296 [PMID: 15336577 DOI: 10.1016/j.amjmed.2004.05.010]
- Marinac JS, Mesa L.** Using a severity of illness scoring system to assess intensive care unit admissions for diabetic ketoacidosis. *Crit Care Med* 2000; **28**: 2238-2241 [PMID: 10921546 DOI: 10.1097/0003246-200007000-00009]
- Gosmanov AR, Gosmanova EO, Dillard-Cannon E.** Management of adult diabetic ketoacidosis. *Diabetes Metab Syndr Obes* 2014; **7**: 255-264 [PMID: 25061324 DOI: 10.2147/DMSO.S50516]
- Azevedo LC, Choi H, Simmonds K, Davidow J, Bagshaw SM.** Incidence and long-term outcomes of critically ill adult patients with moderate-to-severe diabetic ketoacidosis: retrospective matched cohort study. *J Crit Care* 2014; **29**: 971-977 [PMID: 25220529 DOI: 10.1016/j.jcrc.2014.07.034]
- Chang DW, Shapiro MF.** Association Between Intensive Care Unit Utilization During Hospitalization and Costs, Use of Invasive Procedures, and Mortality. *JAMA Intern Med* 2016; **176**: 1492-1499 [PMID: 27532500 DOI: 10.1001/jamainternmed.2016.4298]
- Freire AX, Umpierrez GE, Afessa B, Latif KA, Bridges L, Kitabchi AE.** Predictors of intensive care unit and hospital length of stay in diabetic ketoacidosis. *J Crit Care* 2002; **17**: 207-211 [PMID: 12501147 DOI: 10.1053/jcrc.2002.36755]
- Qari F.** Clinical characteristics of patients with diabetic ketoacidosis at the Intensive Care Unit of a University Hospital. *Pak J Med Sci* 2015; **31**: 1463-1466 [PMID: 26870116 DOI: 10.12669/pjms.316.7550]
- Sotiropoulos A, Papazafropoulou A, Skliros E, Apostolou O, Kardara M, Pappas S.** Effectiveness of management of diabetic ketoacidosis in the emergency department of a general hospital in Greece. *J Emerg Med* 2010; **39**: 341-342 [PMID: 19168308 DOI: 10.1016/j.jemermed.2008.10.001]
- Della Manna T, Steinmetz L, Campos PR, Farhat SC, Schwartsman C, Kuperman H, Setian N, Damiani D.** Subcutaneous use of a fast-acting insulin analog: an alternative treatment for pediatric patients with diabetic ketoacidosis. *Diabetes Care* 2005; **28**: 1856-1861 [PMID: 16043723 DOI: 10.2337/diacare.28.8.1856]
- Fogel N, Zimmerman D.** Management of diabetic ketoacidosis in the emergency department. *CPEM* 2009; **10**: 246-251 [DOI: 10.1016/j.cpem.2009.10.002]
- Lavoie ME.** Management of a patient with diabetic ketoacidosis in the emergency department. *Pediatr Emerg Care* 2015; **31**: 376-380; quiz 381-383 [PMID: 25931345 DOI: 10.1097/PEC.0000000000000429]



- 29 **Goyal N**, Miller JB, Sankey SS, Mossallam U. Utility of initial bolus insulin in the treatment of diabetic ketoacidosis. *J Emerg Med* 2010; **38**: 422-427 [PMID: 18514472 DOI: 10.1016/j.jemermed.2007.11.033]
- 30 **Desse TA**, Eshetie TC, Gudina EK. Predictors and treatment outcome of hyperglycemic emergencies at Jimma University Specialized Hospital, southwest Ethiopia. *BMC Res Notes* 2015; **8**: 553 [PMID: 26455633 DOI: 10.1186/s13104-015-1495-z]

**P- Reviewer:** Al-Haggar M, Rabus SA, Tzamaloukas AH  
**S- Editor:** Ji FF **L- Editor:** A **E- Editor:** Wu HL



## Integrating insulin-like growth factor 1 and sex hormones into neuroprotection: Implications for diabetes

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**Author contributions:** Huffman J designed the figure and contributed to writing the manuscript; Hoffmann C and Taylor GT contributed to writing the manuscript and editing process.

**Conflict-of-interest statement:** The authors declare no conflict of interests for this article.

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**Manuscript source:** Invited manuscript

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Telephone: +1-314-5761709

Received: July 6, 2016

Peer-review started: July 12, 2016

First decision: September 12, 2016

Revised: September 24, 2016

Accepted: November 21, 2016

Article in press: November 22, 2016

Published online: February 15, 2017

in patients with diabetes mellitus, presumably a result of the metabolic complications inherent to the disease. However, an increasing body of evidence has demonstrated the central role of insulin-like growth factor 1 (IGF1) and its relation to sex hormones in many neuroprotective processes. Both male and female patients with diabetes display abnormal IGF1 and sex-hormone levels but the comparison of these fluctuations is seldom a topic of interest. It is interesting to note that both IGF1 and sex hormones have the ability to regulate phosphoinositide 3-kinase-Akt and mitogen-activated protein kinases-extracellular signal-related kinase signaling cascades in animal and cell culture models of neuroprotection. Additionally, there is considerable evidence demonstrating the neuroprotective coupling of IGF1 and estrogen. Androgens have also been implicated in many neuroprotective processes that operate on similar signaling cascades as the estrogen-IGF1 relation. Yet, androgens have not been directly linked to the brain IGF1 system and neuroprotection. Despite the sex-specific variations in brain integrity and hormone levels observed in diabetic patients, the IGF1-sex hormone relation in neuroprotection has yet to be fully substantiated in experimental models of diabetes. Taken together, there is a clear need for the comprehensive analysis of sex differences on brain integrity of diabetic patients and the relationship between IGF1 and sex hormones that may influence brain-health outcomes. As such, this review will briefly outline the basic relation of diabetes and IGF1 and its role in neuroprotection. We will also consider the findings on sex hormones and diabetes as a basis for separately analyzing males and females to identify possible hormone-induced brain abnormalities. Finally, we will introduce the neuroprotective interplay of IGF1 and estrogen and how androgen-derived neuroprotection operates through similar signaling cascades. Future research on both neuroprotection and diabetes should include androgens into the interplay of IGF1 and sex hormones.

### Abstract

Brain integrity and cognitive aptitude are often impaired

**Key words:** Diabetes; Androgens; Estrogen; Insulin; Insulin-like growth factor 1; Neuroprotection; Brain

integrity; Cognition

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**Core tip:** Insulin-like growth factor 1 (IGF1), estrogen, and androgens are known to have neuroprotective properties. Fluctuations in these hormones is observed in patients with diabetes, varies with sex, and may contribute to abnormalities in brain integrity and cognitive impairment typical of the disease. While the neuroprotective coupling of estrogen and IGF1 has been studied extensively, little research has focused similarly on androgens. Furthermore, research investigating the IGF1-sex hormones relation to diabetes and brain-health outcomes is minimal. One avenue of approach to extend this literature may be to examine sex differences by comparison of these hormone levels, brain integrity, and cognitive aptitude.

Huffman J, Hoffmann C, Taylor GT. Integrating insulin-like growth factor 1 and sex hormones into neuroprotection: Implications for diabetes. *World J Diabetes* 2017; 8(2): 45-55 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v8/i2/45.htm> DOI: <http://dx.doi.org/10.4239/wjd.v8.i2.45>

## INTRODUCTION

Diabetes mellitus is a metabolic syndrome known for impaired insulin production. This condition is associated with an abundance of sequelae including cardiovascular disease<sup>[1,2]</sup>, brain atrophy<sup>[3,4]</sup>, and more recently, Alzheimer's disease<sup>[5-7]</sup>. Over the past thirty years, researchers have established strong evidence supporting a link between patients with diabetes and subsequent cognitive impairments and abnormalities in brain integrity.

While meta-analyses have found inconsistencies in the specifics of the literature<sup>[8-10]</sup>, general trends point to cognitive impairments and abnormalities in related structural and functional brain areas. For example, patients with type 1 diabetes (T1D) are frequently found to have decreased psychomotor speed, mental flexibility, and IQ scores<sup>[8,11-13]</sup>. T1D patients also often show reductions in the volume of regional gray matter in areas such as the prefrontal cortex, hippocampus, and thalamus<sup>[12,14,15]</sup>. On the other hand, affected skills in type 2 diabetes (T2D) are largely executive function, memory, and information processing<sup>[16,17]</sup>. Neuroimaging studies done on T2D patients indicate global brain atrophy and microstructural changes<sup>[4,7,9,18]</sup>, while findings regarding white matter hyperintensities are mixed<sup>[3]</sup>.

In both T1D and T2D these decrements are considered mild across most age groups<sup>[8,11,19]</sup>. The severity of cognitive impairments and brain abnormalities are correlated with age of onset in T1D<sup>[11]</sup> and duration

of the disease in T2D<sup>[20,21]</sup>. Age is also a risk factor as deficits in learning and memory have been reported to worsen considerably in T2D patients above 65 years of age<sup>[22]</sup>. Findings suggest the decreased brain volume in patients with T2D is correlated with increased insulin resistance<sup>[23]</sup>, and both brain atrophy and microstructural changes are associated with impaired cognitive performance<sup>[18,20]</sup>.

These data lend support to the idea that brain integrity is compromised in patients with both T1D and T2D, but also emphasize the need to integrate peripheral biomarkers associated with neuroprotection into diabetes research in humans. Various hormones altered as a result of diabetes have been recognized as neuroprotective, including insulin-like growth factor 1 (IGF1) and sex hormones. Research has revealed differences in the serum levels of IGF1 and gonadal hormones in diabetic patients<sup>[24-27]</sup>, with clear sex differences in the effects of androgens and estrogens on the brain in animal models<sup>[28]</sup>.

There is currently a movement in biomedical research to incorporate analyses of sex differences into studies<sup>[29-31]</sup>; however, studies on brain integrity of diabetic patients often fail to examine men and women separately. This is despite findings of sex-specific differences in regional brain volume between men and women<sup>[32-34]</sup>. For instance, DTI scans have also reported white matter hyperintensities are different in men and women diabetics<sup>[35]</sup>. Others have shown that, by combining the data of men and women, T2D patients had smaller gray matter volume with larger ventricular volume and white matter lesions compared to healthy controls. However, when the sexes were analyzed separately, the data for men failed to reach statistical significance<sup>[36]</sup>.

Because sex hormones can act on similar molecular pathways as IGF1, and IGF1 is functionally related to insulin and diabetes, there is a need to further investigate how these hormones interact in the brains of diabetic patients. The relationship between estrogen and IGF1 is the most extensively studied in the neuroprotection literature<sup>[37-39]</sup>, but it has yet to expand experimentally into diabetes research. Furthermore, little attention has been paid to androgen-IGF1 interactions, even in the animal literature, despite the similar mechanisms underlying estrogenic and androgenic neuroprotection.

## DIABETES AND IGF1 RELATION

IGF1 has a hypoglycemic response similar to insulin and, in some circumstances, is capable of modulating insulin receptor (IR) activities. Research has demonstrated that low IGF1 is associated with T1D and T2D<sup>[40-42]</sup>. Moreover, genetic studies suggest decreased IGF1, due to a genetic polymorphism in the promoter region of the IGF1 gene, increases the risk of glucose intolerance and T2D<sup>[43]</sup>.

On the other hand, T2D has also been correlated

with excessively high levels of IGF1. For example, people with acromegaly - a condition known for its overproduction of pituitary growth hormone - have both high levels of IGF1 and a greater risk of developing T2D<sup>[44]</sup>. These findings were corroborated by two large studies from Denmark ( $n = 3354$ ) and Germany ( $n = 7777$ ) which found U-shaped associations between IGF1 levels and the likelihood of developing insulin resistance and T2D<sup>[24,25]</sup>. Moreover, treatment with IGF1 can improve glycemic control in patients with T1D and T2D<sup>[45,46]</sup>, which may suggest an optimal range of IGF1 for normal glycemic control.

Although IGF1 is synthesized in the brain, peripheral values cannot be used to accurately infer brain levels of IGF1 in humans as local synthesis of IGF1 in the brain appears not to correlate with the quantity of IGF1 receptors (IGF1R)<sup>[47-49]</sup>. Evidence from animal models suggest that brain atrophy and loss of DNA are prevented following injection of insulin and IGF1, but not insulin alone, into cerebrospinal fluid of mice<sup>[50]</sup>. Thus, proper systemic levels of IGF1 and its transport from the periphery into the brain is likely necessary for the maintenance of various cognitive processes<sup>[51]</sup>.

Collectively, these data support the involvement of IGF1 in diabetes but also point to an "optimal range" of IGF1. Future research should examine the significance of an optimal peripheral range in the development and maintenance of diabetes and cognitive decline. Moreover, there is a need for data on the role of central vs peripheral IGF1 levels and the subsequent impact on cognitive impairment and brain atrophy.

## THE IGF1 SYSTEM

### Transportation

IGF1 is a polypeptide, structurally similar to insulin, that is released in response to growth hormones secreted by the anterior pituitary<sup>[52]</sup>. While synthesized predominantly by hepatocytes in the liver and released into general circulation, both paracrine and autocrine functions contribute through local tissue synthesis of IGF1. The concentration of IGF1 is greatest during perinatal development and decreases markedly into adulthood. IGF1R are expressed in nearly all neural cells of the CNS, being most highly expressed in the cortex, hippocampus, cerebellum, brainstem, hypothalamus, and spinal cord<sup>[53]</sup>.

The blood brain barrier and blood-cerebrospinal fluid barrier are the two primary routes involved with transporting systemic IGF1 into the brain. Both barriers utilize lipoprotein receptor-related proteins along with IGF1R as transporters to enter the brain<sup>[54,55]</sup>. However, the bioavailability of IGF1 is largely determined by the amount of hormone bound to IGF binding proteins (IGFBPs). Most circulating IGF is bound by IGFBPs, which are proteins that control the distribution and functional capabilities of IGF1 throughout the body. Six different IGFBPs modulate the activity of IGFs *via* binding affinities exceeding that of its respective receptor

and, thus, help regulate the amount of IGF1 that enters the brain<sup>[56]</sup>.

### Signaling pathways

The role of IGF1 is dependent on its binding to insulin-like peptide receptors. The three most important include the IGF1R, IR, and a hybrid receptor formed from heterodimer  $\alpha$ - $\beta$  IR and IGF1R subunits<sup>[53,57]</sup>. These receptors are important to the functional efficacy of IGF1 and have defined downstream molecular pathways. As part of the tyrosine kinase receptor family, activation of IGF1R leads to the signaling of either the mitogen-activated protein kinases-extracellular signal-related kinase (MAPK-ERK) or phosphoinositide 3-kinase (PI3K)-Akt pathways<sup>[53,57]</sup>. These pathways are involved in several important cellular processes including the regulation of gene transcription, apoptosis, oxidative stress, and cellular proliferation and differentiation.

The affinity of IGF1 varies among the three receptors with the highest affinity for IGF1R. Activation of the IGF1R is capable of directly stimulating the RAS-ERK pathway, leading to the modulation of gene transcription by way of activating ETS-like transcription factor, ELK1<sup>[57]</sup>. The capacity of insulin-like peptide receptors to initiate downstream molecular activity is modified in part by the recruitment of insulin receptor substrate (IRS) scaffolding proteins<sup>[57-59]</sup>. This scaffolding helps adjust pathway choice following receptor phosphorylation. The result is activation of PI3K-Akt and subsequent expression of downstream effectors, including glycogen synthase 3 kinase (GSK3 $\beta$ ) and mammalian target of rapamycin<sup>[53,57,60]</sup>.

### Relationship to the insulin system

IGF1 acts primarily through binding to the IGF1R, but also shares with insulin the capacity to bind the IR and hybrid receptor<sup>[53,56,57]</sup>. Insulin is produced exclusively by  $\beta$ -cells of the pancreas and, hence, is strictly transported in the systemic circulation. The amount of insulin capable of entering the brain varies considerably<sup>[54,55]</sup>. Unlike IGF1, insulin appears not to be locally synthesized in adult brain cells<sup>[53,56]</sup>. Similar to IGF1, IR located on endothelial and epithelial cell membranes allow insulin to be transported into the brain from systemic circulation. IRs are concentrated mostly in the olfactory bulb, cerebral cortex, hypothalamus, hippocampus, and cerebellum<sup>[55]</sup>. The movement of systemic insulin into the brain is not controlled by binding proteins.

Both insulin and IGF1 produced in the periphery contribute to varied physiological processes. Proper peripheral IGF1 activation is necessary for insulin secretion from the pancreas and, hence, is implicated in many facets of diabetes<sup>[61]</sup>. However, their functions differ once entering the brain. IGF1R are expressed at notably higher rates in the brain than the rate IGF1 is synthesized. This differential suggests that active transport of IGF1 into the brain is required to furnish sufficient IGF1 for proper neuronal function<sup>[47-49]</sup>. For example, peripheral IGF1 supplies the brain with

information regarding body mass, is related to neural plasticity and cognitive processes, and attenuates cognitive impairment induced by diabetes<sup>[51,62,63]</sup>. Deficiency of IGF1 can also lead to hippocampal atrophy and impaired learning<sup>[64]</sup>. Indeed, IGF1 in the brain is required for proper tissue growth in both the brain and periphery, as well as sufficient glucose regulation and insulin sensitivity<sup>[65,66]</sup>.

Insulin in the periphery is well-known for its role in glucose regulation and communication with the brain to maintain energy homeostasis. Similar to IGF1, insulin is involved in modifying BBB permeability in the brain<sup>[55]</sup> with T2D patients showing greater permeability of the BBB<sup>[67]</sup>. Insulin also acts on the PI3K and MAPK signaling cascades to enhance neuronal survival, plasticity, and subsequent cognitive processes<sup>[55,68,69]</sup>. With that said, insulin does not necessarily regulate glucose activity in neuronal cells after entering the brain. Rather, insulin modulates energy homeostasis through its actions at the level of the hypothalamus<sup>[70]</sup>.

## INTEGRATING SEX HORMONES INTO DIABETES AND IGF1

Diabetes is associated with imbalances in sex steroid hormone levels. This is not surprising as androgens and estrogens are known to play an important role in body composition<sup>[71]</sup> while maintaining glucose and lipid homeostasis<sup>[72,73]</sup>. Research into these imbalances suggests a complex relation between estradiol (E2) and insulin insensitivity. Several studies have reported that postmenopausal women with T2D have increased levels of circulating E2<sup>[27,74,75]</sup>. Elevated E2 has been correlated with the development of insulin resistance and T2D in these women<sup>[76,77]</sup>. Nevertheless, there are at least two studies that have shown inconsistencies between E2 levels and the development of diabetes in postmenopausal women<sup>[78,79]</sup>.

There is also a link between high levels of E2 and diabetes in men. Diabetic men have shown relatively high basal levels of E2<sup>[27,78]</sup>, while men with higher levels of circulating E2 have an increased risk of developing T2D<sup>[80]</sup>. Although this may simply be a product of higher body fat content as adrenal androgens are readily converted to E2 in adipose tissue<sup>[81-83]</sup>, two studies reported E2 results in men were independent of obesity<sup>[78,80]</sup>.

Findings with animal models suggest an opposite conclusion for E2 and diabetes, at least during reproductive ages. Male mice with streptozotocin-induced insulin insensitivity are more likely to develop diabetes than their female cohorts. This increased risk of diabetes in the males can be attenuated with E2 supplements<sup>[84]</sup>. Also, mice lacking the alpha subtype of estrogen receptor (ER $\alpha$ ) have been reported to develop insulin insensitivity<sup>[85]</sup>. In contrast, these data in animals mirror those from postmenopausal women in which glucose homeostasis was positively impacted with estrogen therapy in

the short term<sup>[86]</sup>.

Sex differences in androgen-diabetes relations have also been reported. Postmenopausal women with diabetes displayed elevated circulating testosterone (TS) levels<sup>[27,75]</sup>. Reports suggest that premenopausal women with higher levels of TS<sup>[76,79]</sup>, as well as female mice administered the androgen<sup>[84]</sup>, had a greater risk of developing diabetes. Another example is the link between T2D development and hyperandrogenism experienced by patients with polycystic ovarian syndrome<sup>[87]</sup>. Still, much like E2, there are also studies that dispute these reports, particularly in postmenopausal women<sup>[77,78]</sup>.

A clear sex difference is also indicated in that diabetic men tend to have either lower total, free, or bioavailable TS than healthy men<sup>[27,88,89]</sup>. Indeed, men with the highest levels of TS were at the lowest risk and men with lowest levels of TS were at highest risk for developing T2D<sup>[78,79,90]</sup>. Moreover, men undergoing androgen deprivation treatments for prostatic cancer had a greatly increased risk of developing T2D<sup>[91]</sup>. Yet again, these reports are not without contradiction<sup>[92]</sup> and some studies found this relationship to be dependent on obesity<sup>[80,93]</sup>.

Taken together, there are clear inconsistencies in the findings on sex hormones and diabetes. There is also an apparent lack of research focusing on sex hormones in premenopausal diabetic women that should be addressed<sup>[26]</sup>. It is again important to note that many studies fail to acknowledge the possible relation of sex hormones to the IGF1 system. Findings with serum E2 data are consistent with findings from meta-analyses examining IGF1<sup>[24,25]</sup>. Their proposed U-shaped association of IGF1 and T2D fits into the well-defined mechanistic relationship between E2 and IGF1, described in more detail below. The relation between sex hormones and IGF1 suggests that a delicate hormonal balance is likely an important facet of diabetes-induced brain and cognitive impairment.

## NEUROPROTECTION: SEX HORMONES AND IGF1

### Estrogen and IGF1

An intriguing feature of neuroactive hormones is their ability to protect the CNS from damage, especially in regards to estrogen. ER activation is implicated in the maintenance of various metabolic processes that are also associated with diabetes, including glucose homeostasis and obesity<sup>[94,95]</sup>. Only recently has research with animal models focused on neuroprotection from IGF1-E2 interactions. Evidence suggests that neuroprotective properties of E2 are directly related to receptor activities of insulin-like peptide receptors, mainly IGF1R. E2 and IGF1 work in tandem to reciprocally modulate and facilitate ER and IGF1R activation of the PI3K-Akt and MAPK-ERK signaling cascades<sup>[96-100]</sup>.

IGF1 shows differential sensitivities to the two



estrogen receptor subtypes with ER $\alpha$  being more sensitive than ER $\beta$ <sup>[97,101]</sup>. Selective inhibition of IGF1R, for instance, downregulates ER $\alpha$  expression in the hypothalamus, hippocampus, and cerebral cortex, with the only significant changes of ER $\beta$  occurring in the cerebellum<sup>[38]</sup>. Many glial and neuronal cells in the brain express IGF1R and both ER subtypes<sup>[102]</sup>. In particular, ER $\alpha$  is uniquely capable of increasing IGF1R activity of downstream PI3K-Akt signaling in rodent models<sup>[103,104]</sup>. ER $\alpha$  activation also increases the binding of p85 and IRS-1 regulatory subunits of PI3K and, thus, may be one mechanism assisting in Akt pro-survival signaling through the IGF1R<sup>[39,97]</sup> (Figure 1).

Administration of E2 to mice increased IGF1R and ER $\alpha$  activity in the brain, enabling activation of IGF1R and downstream PI3K-Akt pathway signaling<sup>[97]</sup>. Similarly, IGF1 and insulin modulated ER effects on gene transcription and the PI3K-Akt-GSK3 $\beta$  signaling cascade<sup>[38,98,103,105,106]</sup>. GSK3 $\beta$  is a protein kinase known particularly well for its role in glycogen synthesis. However, as reviewed by Jacobs *et al.*<sup>[60]</sup>, recent attention has turned to the dual pro- and anti-apoptosis capabilities of GSK3 $\beta$  regulated through multiple different pathways. Indeed, the neuroprotective effects of IGF1 may be consequent to Akt-derived inhibition of GSK3 $\beta$  in a hypoxic state<sup>[107]</sup> (Figure 1).

Activation of the MAPK pathway is another important signal transduction pathway involved with regulating gene transcription and cellular proliferation and differentiation, particularly in cancer<sup>[108]</sup>. However, multiple studies have demonstrated that the neuroprotective properties of estrogen are also derived from its ability to regulate MAPK signaling in the brain<sup>[38]</sup>. Both estrogen and IGF1 can facilitate MAPK signaling through the IGF1R, with IGF1 increasing ER $\alpha$  activities in the presence of E2<sup>[104]</sup>. Akt inhibitors are capable of nullifying the neuroprotective effects of IGF1 and E2 regardless of MAPK signaling<sup>[99,104]</sup>, while ERK suppression increases PI3K-Akt activity *via* ER and IGF1R heterodimers<sup>[39]</sup>. Thus, it appears the PI3K-Akt pro-survival signaling cascade is the most involved with the neuroprotective coupling of E2 and IGF1<sup>[39]</sup>.

It is important to note that IGF1 and E2 have a remarkable reciprocity. Inhibition of ER activity can downregulate IGF1R expression in the hippocampus<sup>[109]</sup>, a brain region known to atrophy in patients with diabetes and glucose intolerance<sup>[110-112]</sup>. Similarly, IGF1 has the capacity to upregulate ER $\alpha$  in the hippocampus and is impaired following administration of IGF1R antagonists<sup>[109]</sup>. Agonists or antagonists of either hormone can respectively facilitate or inhibit the neuroprotective and memory enhancing properties of the other<sup>[96,109,113-116]</sup>. This has led some to suggest that cooperation between IGF1R and ER is required for many E2-induced neuroprotective processes. The present section does not, however, do justice to the complexity of the relation between estrogen and IGF1 receptors. A fuller explanation can be found in one of several reviews<sup>[37-39,101,109,117]</sup>.

## Androgens and IGF1

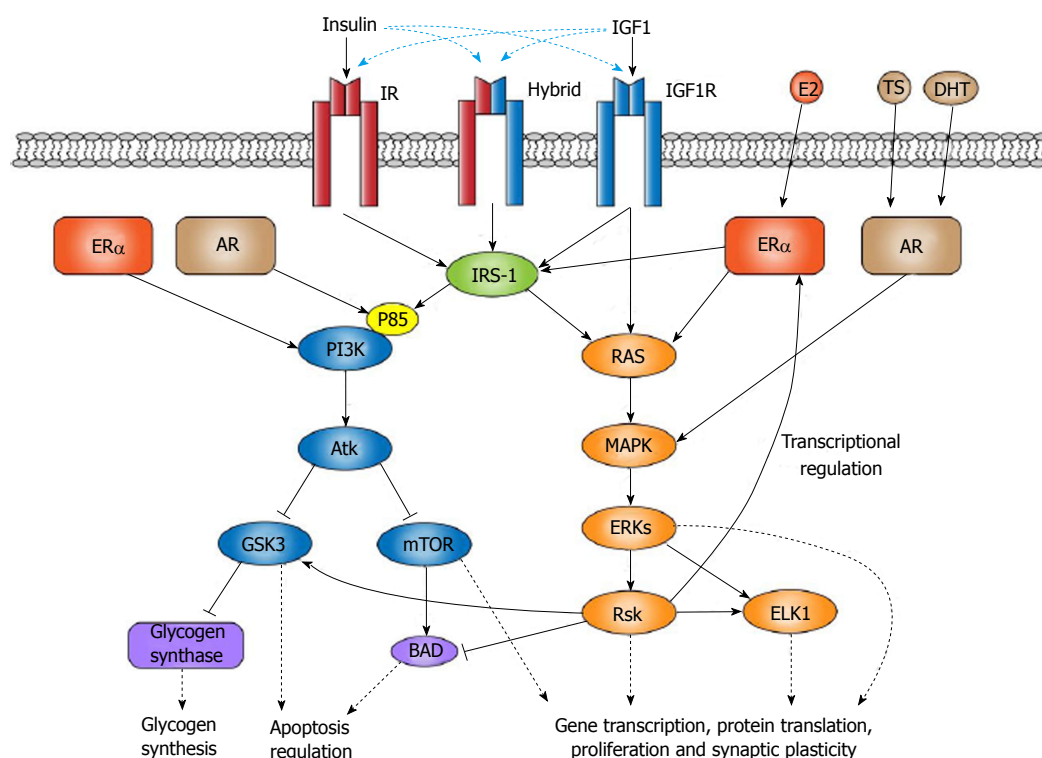
Far less research has examined a functional link between IGF1 and androgens in the brain. This is an unfortunate but common trend in neuroendocrinology. Estrogens are the most intensely studied gonadal hormone, despite estrogens and androgens sharing metabolic pathways and functional properties. Much of the current literature on IGF1-androgen relations are directed at the periphery, particularly prostate cancer and motor systems, for which there are a number of recent reviews<sup>[118,119]</sup>. Few studies have examined IGF1-androgen interactions in neuroprotection<sup>[120,121]</sup> and none, to our knowledge, have empirically examined this interaction in diabetes. Therefore, we have relied on peripheral data, often from *in vitro* experiments, to extrapolate the androgen receptor (AR) brain discussion.

There is evidence that the two main androgens, TS and dihydrotestosterone (DHT), are capable of neuroprotection through binding the AR<sup>[122-126]</sup>. Similar to ER $\alpha$ , androgen activation of the AR in mouse vas deferens epithelial cells can modulate the p85 regulatory subunits of PI3K and subsequently trigger Akt expression (Figure 1). Inhibiting the AR prevents these signaling effects<sup>[127]</sup>. Phosphorylation of MAPK and Akt can also increase AR activation in low androgen and estrogen concentrations, as well as increase the neuroprotective activities of ER $\alpha$  and AR<sup>[128]</sup>. Recent findings showed that DHT, which has a higher affinity than TS for the AR, prevents apoptosis in a C6 glial cell line through the PI3K-Akt signaling cascade<sup>[129]</sup>. These effects were also impaired by inhibition of PI3K and suggest a functional relationship between apoptosis and AR activities.

Interestingly, studies have demonstrated that binding of DHT to the transmembrane AR impairs MAPK and PI3K signaling and subsequent neuroprotection from DHT or E2<sup>[130-132]</sup>. This suggests that nuclear activation of the AR by DHT is likely one mechanism behind DHT's neuroprotective properties<sup>[130]</sup>. DHT may also interact with effectors downstream of ER and IGF1R signaling. Both TS and DHT can activate the MAPK-ERK signaling cascade<sup>[132]</sup> which has been shown to induce ribosomal S6 kinase (Rsk) expression. Rsk signaling can lead to the inhibition of the pro-apoptosis Bad protein and the activation of downstream effectors including the ER, GSK3 $\beta$  and ELK1<sup>[133]</sup> (Figure 1).

One possible explanation for the neuroprotective role of androgens is the conversion in the steroid metabolic cascade of TS into E2 by the enzyme aromatase. That is, TS may be involved in neuroprotection only to the extent that TS is a precursor for E2, which is capable of activating MAPK or PI3K signaling through the ER and IGF1R. The aromatization of TS into E2, as well as the aromatase enzyme, have been suggested to play an important role in neuroprotection<sup>[134-139]</sup>.

The ratio of endogenous TS to E2, and subsequent influences of aromatized TS, is indeed a topic of recent interest<sup>[26]</sup>. Increased local synthesis of E2 from elevated aromatase expression is seen in models



**Figure 1** Similar signaling cascades involved with neuroprotection for insulin-like peptides and sex hormones. The insulin receptor (IR), insulin-like growth factor 1 receptor (IGF1R), and insulin-IGF1 hybrid receptor enact their neuroprotection through the mitogen-activated protein kinases-extracellular signal-related kinase (MAPK-ERK) or phosphoinositide 3-kinase (PI3K)-Akt pathways signaling cascades. Although IGF1R can directly activate the RAS-ERK pathway, both the insulin-like peptide receptors and the estrogen receptor alpha (ER $\alpha$ ) firstly interact with insulin receptor substrate 1 (IRS-1) scaffolding proteins. ER $\alpha$  and the androgen receptor (AR) can also directly modulate PI3K-Akt and MAPK-ERK signaling. Both IRS-1 and p85 binding of PI3K are increased with ER $\alpha$  activation, leading to downstream Akt-derived inhibition of glycogen synthase kinase 3 (GSK3) and mammalian target of rapamycin (mTOR). GSK3, specifically, is involved with glycogen synthesis, while both effectors are involved in apoptosis. A similar effect may occur with AR's ability to modulate p85 binding to PI3K. AR-induced MAPK-ERK signaling also results in ribosomal S6 kinase (Rsk) expression that can inhibit the pro-apoptosis bcl-2-associated death promoter protein, as well as effects on the ER, GSK3, and the ETS-like transcription factor, ELK1. Solid black arrows indicate downstream interaction. Dashed black arrows represent the influence of kinases or proteins on the cellular environment. Dashed blue arrows represent the binding capabilities of IGF1 and insulin across all three receptor types.

of neuroprotection from other brain disorders, *e.g.*, stroke<sup>[140]</sup>. More pertinent to this review, streptozotocin-induced diabetes causes a considerable reduction in aromatase synthesis in female and male reproductive systems<sup>[141]</sup>. Notably, inhibition of aromatase decreases E2 and impairs insulin sensitivity and peripheral glucose disposal in healthy males<sup>[142]</sup>, although the influence this may have on brain integrity and cognitive outcomes remains debated<sup>[143]</sup>.

Another explanation places greater emphasis on the other pathway in the steroid metabolic cascade leading to DHT. Metabolites of DHT, 3 $\alpha$ -Diol and 3 $\beta$ -Diol, are also bioactive and may bind the ER or insulin-like peptide receptors to initiate MAPK or PI3K signaling cascades. Indeed, research shows that 3 $\alpha$ -Diol stimulated PI3K-Akt signaling enhances cell survival in the prostate<sup>[144]</sup>. Similarly, DHT metabolites may influence transcriptional activities of nuclear ER by modulating ER-induced MAPK or PI3K signaling cascades.

Few *in vivo* studies examining these sex steroid metabolites have focused on MAPK or PI3K signal cascades in the brain. There is, however, evidence that 3 $\alpha$ -Diol inhibits protein kinase A expression in the rat hippocampus<sup>[145]</sup>. Others have reported that strep-

tozotocin-induced diabetic mice had lower levels of TS and 3 $\alpha$ -Diol in the cerebral cortex, and lower levels of DHT and 3 $\alpha$ -Diol in the spinal cord<sup>[146]</sup>. It is still unclear, though, whether 3 $\alpha$ -Diol and 3 $\beta$ -Diol interact with or initiate the MAPK or PI3K signaling cascades following activation of the ER, AR, or, possibly, IGF1R.

None of these explanations clarify fully the ability of the AR to directly trigger these signaling cascades. We do not aim to discount the neuroprotective mechanisms of ER and AR, or the clear link between E2 and IGF1 processes in neuroprotection. Rather, we simply suggest that androgen-derived neuroprotection may be intertwined with IGF1, the activation of insulin-like peptide receptors, and/or the IGF1R and ER coupling. Given the common signaling pathways between these hormones, we suggest future research should aim to include androgens and AR activities into the ER-IGF1R neuroprotective coupling, as well as serum comparisons in brain-health outcomes of diabetic patients.

## CONCLUSION

The reciprocity of IGF1 and estrogen in neuroprotective processes is well-established in cell cultures and

animal models<sup>[38]</sup>. Interactions between androgens and IGF1 may also play an important role in the E2-IGF1 neuroprotective coupling. Both estrogens and androgens enact their neuroprotection through similar, but not identical, signal transduction pathways. Recognition of this has led us to consider the possibility that these sex hormones may work together with IGF1 and insulin-like peptide receptors to modulate MAPK and PI3K signaling and their neuroprotective properties.

Regulation of MAPK and PI3K activity may also be a driving force behind the structural changes, atrophy of brain regions, or functional changes, often observed in diabetic patients. Drawing conclusions from imaging data in humans to those found in animal models is indeed difficult. Nevertheless, there is a need for a clearer mechanistic explanation grounding the cognitive decline and brain abnormalities observed in diabetic patients.

Future studies in human research on diabetic brain integrity should integrate hormone titer measures to help substantiate sex differences in brain-health outcomes of diabetic patients. This approach may also assist in identifying region-specific brain abnormalities resulting from fluctuations in IGF1 and sex hormones between men and women. Moreover, animal models examining the E2-IGF1 coupling in neuroprotection should employ streptozotocin-induced diabetes, as well as the possible role of androgens and AR activities. These conclusions warrant further examination of the variability present in cognitive and brain-health outcomes for patients with diabetes as a result of sex hormone relations to IGF1, insulin, and the insulin-like peptide receptors.

## REFERENCES

- Ryden L, Grant PJ, Anker SD, Berne C, Cosentino F, Danchin N, Deaton C, Escaned J, Hammes HP, Huikuri H, Marre M, Marx N, Mellbin L, Ostergren J, Patrono C, Seferovic P, Uva MS, Taskinen MR, Tendera M, Tuomilehto J, Valensi P, Zamorano JL. ESC guidelines on diabetes, pre-diabetes, and cardiovascular diseases developed in collaboration with the EASD - summary. *Diab Vasc Dis Res* 2014; **11**: 133-173 [PMID: 24800783 DOI: 10.1177/1479164114525548]
- Tillin T, Hughes AD, Mayet J, Whincup P, Sattar N, Forouhi NG, McKeigue PM, Chaturvedi N. The relationship between metabolic risk factors and incident cardiovascular disease in Europeans, South Asians, and African Caribbeans: SABRE (Southall and Brent Revisited) -- a prospective population-based study. *J Am Coll Cardiol* 2013; **61**: 1777-1786 [PMID: 23500273 DOI: 10.1016/j.jacc.2012.12.046]
- van Elderen SG, de Roos A, de Craen AJ, Westendorp RG, Blauw GJ, Jukema JW, Bollen EL, Middelkoop HA, van Buchem MA, van der Grond J. Progression of brain atrophy and cognitive decline in diabetes mellitus: a 3-year follow-up. *Neurology* 2010; **75**: 997-1002 [PMID: 20837967 DOI: 10.1212/WNL.0b013e3181f25f06]
- de Bresser J, Tiehuis AM, van den Berg E, Reijmer YD, Jongen C, Kappelle LJ, Mali WP, Viergever MA, Biessels GJ. Progression of cerebral atrophy and white matter hyperintensities in patients with type 2 diabetes. *Diabetes Care* 2010; **33**: 1309-1314 [PMID: 20299484 DOI: 10.2337/dc09-1923]
- Peila R, Rodriguez BL, Launer LJ. Type 2 diabetes, APOE gene, and the risk for dementia and related pathologies: The Honolulu-Asia Aging Study. *Diabetes* 2002; **51**: 1256-1262 [PMID: 11916953 DOI: 10.2337/diabetes.51.4.1256]
- Cheng G, Huang C, Deng H, Wang H. Diabetes as a risk factor for dementia and mild cognitive impairment: a meta-analysis of longitudinal studies. *Intern Med J* 2012; **42**: 484-491 [PMID: 22372522 DOI: 10.1111/j.1445-5994.2012.02758.x]
- Moran C, Phan TG, Chen J, Blizzard L, Beare R, Venn A, Münch G, Wood AG, Forbes J, Greenaway TM, Pearson S, Srikanth V. Brain atrophy in type 2 diabetes: regional distribution and influence on cognition. *Diabetes Care* 2013; **36**: 4036-4042 [PMID: 23939539 DOI: 10.2337/dc13-0143]
- Brands AM, Biessels GJ, de Haan EH, Kappelle LJ, Kessels RP. The effects of type 1 diabetes on cognitive performance: a meta-analysis. *Diabetes Care* 2005; **28**: 726-735 [PMID: 15735218 DOI: 10.2337/diacare.28.3.726]
- van Harten B, de Leeuw FE, Weinstein HC, Scheltens P, Biessels GJ. Brain imaging in patients with diabetes: a systematic review. *Diabetes Care* 2006; **29**: 2539-2548 [PMID: 17065699 DOI: 10.2337/dc06-1637]
- Sadanand S, Balachandar R, Bharath S. Memory and executive functions in persons with type 2 diabetes: a meta-analysis. *Diabetes Metab Res Rev* 2016; **32**: 132-142 [PMID: 25963303 DOI: 10.1002/dmrr.2664]
- Ferguson SC, Blane A, Wardlaw J, Frier BM, Perros P, McCrimmon RJ, Deary IJ. Influence of an early-onset age of type 1 diabetes on cerebral structure and cognitive function. *Diabetes Care* 2005; **28**: 1431-1437 [PMID: 15920064 DOI: 10.2337/diacare.28.6.1431]
- Northam EA, Rankins D, Lin A, Wellard RM, Pell GS, Finch SJ, Werther GA, Cameron FJ. Central nervous system function in youth with type 1 diabetes 12 years after disease onset. *Diabetes Care* 2009; **32**: 445-450 [PMID: 19151204 DOI: 10.2337/dc08-1657]
- Biessels GJ, Deary IJ, Ryan CM. Cognition and diabetes: a lifespan perspective. *Lancet Neurol* 2008; **7**: 184-190 [PMID: 18207116 DOI: 10.1016/S1474-4422(08)70021-8]
- Hughes TM, Ryan CM, Aizenstein HJ, Nunley K, Gianaros PJ, Miller R, Costacou T, Strotmeyer ES, Orchard TJ, Rosano C. Frontal gray matter atrophy in middle aged adults with type 1 diabetes is independent of cardiovascular risk factors and diabetes complications. *J Diabetes Complications* 2013; **27**: 558-564 [PMID: 23994432 DOI: 10.1016/j.jdiacomp.2013.07.001]
- Foghi K, Ahmadvour S. Role of neuronal apoptosis in volumetric change of hippocampus in diabetes mellitus type 1: a predictive model. *ISRN Anat* 2013; **2013**: 958461 [PMID: 25938109 DOI: 10.5402/2013/958461]
- van den Berg E, Kloppenborg RP, Kessels RP, Kappelle LJ, Biessels GJ. Type 2 diabetes mellitus, hypertension, dyslipidemia and obesity: A systematic comparison of their impact on cognition. *Biochim Biophys Acta* 2009; **1792**: 470-481 [PMID: 18848880 DOI: 10.1016/j.bbdis.2008.09.004]
- Palta P, Schneider AL, Biessels GJ, Touradji P, Hill-Briggs F. Magnitude of cognitive dysfunction in adults with type 2 diabetes: a meta-analysis of six cognitive domains and the most frequently reported neuropsychological tests within domains. *J Int Neuropsychol Soc* 2014; **20**: 278-291 [PMID: 24555960 DOI: 10.1017/S1355617713001483]
- Reijmer YD, Brundel M, de Bresser J, Kappelle LJ, Leemans A, Biessels GJ. Microstructural white matter abnormalities and cognitive functioning in type 2 diabetes: a diffusion tensor imaging study. *Diabetes Care* 2013; **36**: 137-144 [PMID: 22961577 DOI: 10.2337/dc12-0493]
- Biessels GJ, Strachan MW, Visseren FL, Kappelle LJ, Whitmer RA. Dementia and cognitive decline in type 2 diabetes and prediabetic stages: towards targeted interventions. *Lancet Diabetes Endocrinol* 2014; **2**: 246-255 [PMID: 24622755 DOI: 10.1016/S2213-8587(13)70088-3]
- Manschot SM, Brands AM, van der Grond J, Kessels RP, Algra A, Kappelle LJ, Biessels GJ. Brain magnetic resonance imaging correlates of impaired cognition in patients with type 2 diabetes. *Diabetes* 2006; **55**: 1106-1113 [PMID: 16567535 DOI: 10.2337/diabetes.55.04.06.db05-1323]
- Saczynski JS, Siggurdsson S, Jonsson PV, Eiriksdottir G, Olafsdottir E, Kjartansson O, Harris TB, van Buchem MA,



- Gudnason V, Launer LJ. Glycemic status and brain injury in older individuals: the age gene/environment susceptibility-Reykjavik study. *Diabetes Care* 2009; **32**: 1608-1613 [PMID: 19509008 DOI: 10.2337/dc08-2300]
- 22 **Ryan CM**, Geckle M. Why is learning and memory dysfunction in Type 2 diabetes limited to older adults? *Diabetes Metab Res Rev* 2000; **16**: 308-315 [PMID: 11025555 DOI: 10.1002/1520-7560(2000)9999]
  - 23 **Tan ZS**, Beiser AS, Fox CS, Au R, Himali JJ, Debette S, Decarli C, Vasan RS, Wolf PA, Seshadri S. Association of metabolic dysregulation with volumetric brain magnetic resonance imaging and cognitive markers of subclinical brain aging in middle-aged adults: the Framingham Offspring Study. *Diabetes Care* 2011; **34**: 1766-1770 [PMID: 21680719 DOI: 10.2337/dc11-0308]
  - 24 **Friedrich N**, Thuesen B, Jørgensen T, Juul A, Spielhagen C, Wallaschofski H, Linneberg A. The association between IGF-I and insulin resistance: a general population study in Danish adults. *Diabetes Care* 2012; **35**: 768-773 [PMID: 22374641 DOI: 10.2337/dc11-1833]
  - 25 **Schneider HJ**, Friedrich N, Klotsche J, Schipf S, Nauck M, Völzke H, Sievers C, Pieper L, März W, Wittchen HU, Stalla GK, Wallaschofski H. Prediction of incident diabetes mellitus by baseline IGF1 levels. *Eur J Endocrinol* 2011; **164**: 223-229 [PMID: 21059863 DOI: 10.1530/EJE-10-0963]
  - 26 **Kim C**, Halter JB. Endogenous sex hormones, metabolic syndrome, and diabetes in men and women. *Curr Cardiol Rep* 2014; **16**: 467 [PMID: 24585109 DOI: 10.1007/s11886-014-0467-6]
  - 27 **Ding EL**, Song Y, Malik VS, Liu S. Sex differences of endogenous sex hormones and risk of type 2 diabetes: a systematic review and meta-analysis. *JAMA* 2006; **295**: 1288-1299 [PMID: 16537739 DOI: 10.1001/jama.295.11.1288]
  - 28 **Pesaresi M**, Maschi O, Giatti S, Garcia-Segura LM, Caruso D, Melcangi RC. Sex differences in neuroactive steroid levels in the nervous system of diabetic and non-diabetic rats. *Horm Behav* 2010; **57**: 46-55 [PMID: 19422828 DOI: 10.1016/j.yhbeh.2009.04.008]
  - 29 **Franconi F**, Seghieri G, Canu S, Straface E, Campesi I, Malorni W. Are the available experimental models of type 2 diabetes appropriate for a gender perspective? *Pharmacol Res* 2008; **57**: 6-18 [PMID: 18221886 DOI: 10.1016/j.phrs.2007.11.007]
  - 30 **Legato MJ**, Gelzer A, Golland R, Ebner SA, Rajan S, Villagra V, Kosowski M. Gender-specific care of the patient with diabetes: review and recommendations. *Gend Med* 2006; **3**: 131-158 [PMID: 16860272 DOI: 10.1016/S1550-8579(06)80202-0]
  - 31 **McCullum M**, Hansen LS, Lu L, Sullivan PW. Gender differences in diabetes mellitus and effects on self-care activity. *Gend Med* 2005; **2**: 246-254 [PMID: 16464736 DOI: 10.1016/S1550-8579(05)80054-3]
  - 32 **Goldstein JM**, Seidman LJ, Horton NJ, Makris N, Kennedy DN, Caviness VS, Faraone SV, Tsuang MT. Normal sexual dimorphism of the adult human brain assessed by in vivo magnetic resonance imaging. *Cereb Cortex* 2001; **11**: 490-497 [PMID: 11375910 DOI: 10.1093/cercor/11.6.490]
  - 33 **Menzler K**, Belke M, Wehrmann E, Krakow K, Lengler U, Jansen A, Hamer HM, Oertel WH, Rosenow F, Knake S. Men and women are different: diffusion tensor imaging reveals sexual dimorphism in the microstructure of the thalamus, corpus callosum and cingulum. *Neuroimage* 2011; **54**: 2557-2562 [PMID: 21087671 DOI: 10.1016/j.neuroimage.2010.11.029]
  - 34 **Nopoulos P**, Flaum M, O'Leary D, Andreasen NC. Sexual dimorphism in the human brain: evaluation of tissue volume, tissue composition and surface anatomy using magnetic resonance imaging. *Psychiatry Res* 2000; **98**: 1-13 [PMID: 10708922 DOI: 10.1016/S0925-4927(99)00044-X]
  - 35 **Sachdev PS**, Parslow R, Wen W, Anstey KJ, Eastale S. Sex differences in the causes and consequences of white matter hyperintensities. *Neurobiol Aging* 2009; **30**: 946-956 [PMID: 17950492 DOI: 10.1016/j.neurobiolaging.2007.08.023]
  - 36 **Jongen C**, van der Grond J, Kappelle LJ, Biessels GJ, Viergever MA, Pluim JP. Automated measurement of brain and white matter lesion volume in type 2 diabetes mellitus. *Diabetologia* 2007; **50**: 1509-1516 [PMID: 17492428 DOI: 10.1007/s00125-007-0688-y]
  - 37 **Arevalo MA**, Azcoitia I, Garcia-Segura LM. The neuroprotective actions of oestradiol and oestrogen receptors. *Nat Rev Neurosci* 2015; **16**: 17-29 [PMID: 25423896 DOI: 10.1038/nrn3856]
  - 38 **Garcia-Segura LM**, Arévalo MA, Azcoitia I. Interactions of estradiol and insulin-like growth factor-I signalling in the nervous system: new advances. *Prog Brain Res* 2010; **181**: 251-272 [PMID: 20478442 DOI: 10.1016/S0079-6123(08)81014-X]
  - 39 **Sohrabji F**. Estrogen-IGF-1 interactions in neuroprotection: ischemic stroke as a case study. *Front Neuroendocrinol* 2015; **36**: 1-14 [PMID: 24882635 DOI: 10.1016/j.yfrne.2014.05.003]
  - 40 **Ezzat VA**, Duncan ER, Wheatcroft SB, Kearney MT. The role of IGF-I and its binding proteins in the development of type 2 diabetes and cardiovascular disease. *Diabetes Obes Metab* 2008; **10**: 198-211 [PMID: 18269635 DOI: 10.1111/j.1463-1326.2007.00709.x]
  - 41 **Janssen JA**, Jacobs ML, Derckx FH, Weber RF, van der Lely AJ, Lamberts SW. Free and total insulin-like growth factor I (IGF-I), IGF-binding protein-1 (IGFBP-1), and IGFBP-3 and their relationships to the presence of diabetic retinopathy and glomerular hyperfiltration in insulin-dependent diabetes mellitus. *J Clin Endocrinol Metab* 1997; **82**: 2809-2815 [PMID: 9284701 DOI: 10.1210/jcem.82.9.4180]
  - 42 **Teppala S**, Shankar A, Sabanayagam C. Association between IGF-I and chronic kidney disease among US adults. *Clin Exp Nephrol* 2010; **14**: 440-444 [PMID: 20567872 DOI: 10.1007/s10157-010-0307-y]
  - 43 **Vaessen N**, Heutink P, Janssen JA, Witteman JC, Testers L, Hofman A, Lamberts SW, Oostra BA, Pols HA, van Duijn CM. A polymorphism in the gene for IGF-I: functional properties and risk for type 2 diabetes and myocardial infarction. *Diabetes* 2001; **50**: 637-642 [PMID: 11246885 DOI: 10.2337/diabetes.50.3.637]
  - 44 **Melmed S**. Medical progress: Acromegaly. *N Engl J Med* 2006; **355**: 2558-2573 [PMID: 17167139 DOI: 10.1056/NEJMra062453]
  - 45 **Carroll PV**, Christ ER, Umpleby AM, Gowrie I, Jackson N, Bowes SB, Hovorka R, Croos P, Sönksen PH, Russell-Jones DL. IGF-I treatment in adults with type 1 diabetes: effects on glucose and protein metabolism in the fasting state and during a hyperinsulinemic-euglycemic amino acid clamp. *Diabetes* 2000; **49**: 789-796 [PMID: 10905488 DOI: 10.2337/diabetes.49.5.789]
  - 46 **Zenobi PD**, Jaeggi-Groisman SE, Riesen WF, Roder ME, Froesch ER. Insulin-like growth factor-I improves glucose and lipid metabolism in type 2 diabetes mellitus. *J Clin Invest* 1992; **90**: 2234-2241 [PMID: 1469083 DOI: 10.1172/JCI116109]
  - 47 **Carro E**, Trejo JL, Núñez A, Torres-Aleman I. Brain repair and neuroprotection by serum insulin-like growth factor I. *Mol Neurobiol* 2003; **27**: 153-162 [PMID: 12777685]
  - 48 **Carro E**, Torres-Aleman I. Serum insulin-like growth factor I in brain function. *Keio J Med* 2006; **55**: 59-63 [PMID: 16830417 DOI: 10.2302/kjm.55.59]
  - 49 **Nishijima T**, Piriz J, Dufloot S, Fernandez AM, Gaitan G, Gomez-Pinedo U, Verdugo JM, Leroy F, Soya H, Nuñez A, Torres-Aleman I. Neuronal activity drives localized blood-brain-barrier transport of serum insulin-like growth factor-I into the CNS. *Neuron* 2010; **67**: 834-846 [PMID: 20826314 DOI: 10.1016/j.neuron.2010.08.007]
  - 50 **Serbedzija P**, Madl JE, Ishii DN. Insulin and IGF-I prevent brain atrophy and DNA loss in diabetes. *Brain Res* 2009; **1303**: 179-194 [PMID: 19781531 DOI: 10.1016/j.brainres.2009.09.063]
  - 51 **Lupien SB**, Bluhm EJ, Ishii DN. Systemic insulin-like growth factor-I administration prevents cognitive impairment in diabetic rats, and brain IGF regulates learning/memory in normal adult rats. *J Neurosci Res* 2003; **74**: 512-523 [PMID: 14598295 DOI: 10.1002/jnr.10791]
  - 52 **Vardatsikos G**, Sahu A, Srivastava AK. The insulin-like growth factor family: molecular mechanisms, redox regulation, and clinical implications. *Antioxid Redox Signal* 2009; **11**: 1165-1190 [PMID: 19014342 DOI: 10.1089/ars.2008.2161]
  - 53 **Fernandez AM**, Torres-Aleman I. The many faces of insulin-like peptide signalling in the brain. *Nat Rev Neurosci* 2012; **13**: 225-239 [PMID: 22430016 DOI: 10.1038/nrn3209]
  - 54 **Banks WA**. The source of cerebral insulin. *Eur J Pharmacol* 2004;

- 490: 5-12 [PMID: 15094069 DOI: 10.1016/j.ejphar.2004.02.040]
- 55 **Banks WA**, Owen JB, Erickson MA. Insulin in the brain: there and back again. *Pharmacol Ther* 2012; **136**: 82-93 [PMID: 22820012 DOI: 10.1016/j.pharmthera.2012.07.006]
- 56 **Russo VC**, Gluckman PD, Feldman EL, Werther GA. The insulin-like growth factor system and its pleiotropic functions in brain. *Endocr Rev* 2005; **26**: 916-943 [PMID: 16131630 DOI: 10.1210/er.2004-0024]
- 57 **Benarroch EE**. Insulin-like growth factors in the brain and their potential clinical implications. *Neurology* 2012; **79**: 2148-2153 [PMID: 23170013 DOI: 10.1212/WNL.0b013e3182752eef]
- 58 **Ramalingam M**, Kim SJ. Mechanisms of action of brain insulin against neurodegenerative diseases. *J Neural Transm (Vienna)* 2014; **121**: 611-626 [PMID: 24398779 DOI: 10.1007/s00702-013-1147-1]
- 59 **Werner H**, LeRoith D. Insulin and insulin-like growth factor receptors in the brain: physiological and pathological aspects. *Eur Neuropsychopharmacol* 2014; **24**: 1947-1953 [PMID: 24529663 DOI: 10.1016/j.euroneuro.2014.01.020]
- 60 **Jacobs KM**, Bhawe SR, Ferraro DJ, Jaboin JJ, Hallahan DE, Thotala D. GSK-3 $\beta$ : A Bifunctional Role in Cell Death Pathways. *Int J Cell Biol* 2012; **2012**: 930710 [PMID: 22675363 DOI: 10.1155/2012/930710]
- 61 **Withers DJ**, Burks DJ, Towery HH, Altamuro SL, Flint CL, White MF. Irs-2 coordinates Igf-1 receptor-mediated beta-cell development and peripheral insulin signalling. *Nat Genet* 1999; **23**: 32-40 [PMID: 10471495 DOI: 10.1038/12631]
- 62 **Aleman A**, Torres-Alemán I. Circulating insulin-like growth factor I and cognitive function: neuromodulation throughout the lifespan. *Prog Neurobiol* 2009; **89**: 256-265 [PMID: 19665513 DOI: 10.1016/j.pneurobio.2009.07.008]
- 63 **Torres-Aleman I**. Toward a comprehensive neurobiology of IGF-I. *Dev Neurobiol* 2010; **70**: 384-396 [PMID: 20186710 DOI: 10.1002/dneu.20778]
- 64 **Trejo JL**, Carro E, Garcia-Galloway E, Torres-Aleman I. Role of insulin-like growth factor I signaling in neurodegenerative diseases. *J Mol Med (Berl)* 2004; **82**: 156-162 [PMID: 14647921 DOI: 10.1007/s00109-003-0499-7]
- 65 **Huffman DM**, Farias Quipildor G, Mao K, Zhang X, Wan J, Apontes P, Cohen P, Barzilay N. Central insulin-like growth factor-1 (IGF-1) restores whole-body insulin action in a model of age-related insulin resistance and IGF-1 decline. *Aging Cell* 2016; **15**: 181-186 [PMID: 26534869 DOI: 10.1111/acer.12415]
- 66 **Kappeler L**, De Magalhães Filho C, Dupont J, Leneuve P, Cervera P, Périn L, Loudes C, Blaise A, Klein R, Epelbaum J, Le Bouc Y, Holzenberger M. Brain IGF-1 receptors control mammalian growth and lifespan through a neuroendocrine mechanism. *PLoS Biol* 2008; **6**: e254 [PMID: 18959478 DOI: 10.1371/journal.pbio.0060254]
- 67 **Starr JM**, Wardlaw J, Ferguson K, MacLulich A, Deary IJ, Marshall I. Increased blood-brain barrier permeability in type II diabetes demonstrated by gadolinium magnetic resonance imaging. *J Neurol Neurosurg Psychiatry* 2003; **74**: 70-76 [PMID: 12486269 DOI: 10.1136/jnnp.74.1.70]
- 68 **Duarte AI**, Moreira PI, Oliveira CR. Insulin in central nervous system: more than just a peripheral hormone. *J Aging Res* 2012; **2012**: 384017 [PMID: 22500228 DOI: 10.1155/2012/384017]
- 69 **Zeng Y**, Zhang L, Hu Z. Cerebral insulin, insulin signaling pathway, and brain angiogenesis. *Neurol Sci* 2016; **37**: 9-16 [PMID: 26442674 DOI: 10.1007/s10072-015-2386-8]
- 70 **Sánchez-Lasheras C**, Könnér AC, Brüning JC. Integrative neurobiology of energy homeostasis-neurocircuits, signals and mediators. *Front Neuroendocrinol* 2010; **31**: 4-15 [PMID: 19729032 DOI: 10.1016/j.yfine.2009.08.002]
- 71 **Power ML**, Schulkin J. Sex differences in fat storage, fat metabolism, and the health risks from obesity: possible evolutionary origins. *Br J Nutr* 2008; **99**: 931-940 [PMID: 17977473 DOI: 10.1017/S0007114507853347]
- 72 **Mauvais-Jarvis F**. Sex differences in metabolic homeostasis, diabetes, and obesity. *Biol Sex Differ* 2015; **6**: 14 [PMID: 26339468 DOI: 10.1186/s13293-015-0033-y]
- 73 **Varlamov O**, Bethea CL, Roberts CT. Sex-specific differences in lipid and glucose metabolism. *Front Endocrinol (Lausanne)* 2014; **5**: 241 [PMID: 25646091 DOI: 10.3389/fendo.2014.00241]
- 74 **Goodman-Gruen D**, Barrett-Connor E. Sex differences in the association of endogenous sex hormone levels and glucose tolerance status in older men and women. *Diabetes Care* 2000; **23**: 912-918 [PMID: 10895840 DOI: 10.2337/diacare.23.7.912]
- 75 **Phillips GB**, Tuck CH, Jing TY, Boden-Albala B, Lin IF, Dahodwala N, Sacco RL. Association of hyperandrogenemia and hyperestrogenemia with type 2 diabetes in Hispanic postmenopausal women. *Diabetes Care* 2000; **23**: 74-79 [PMID: 10857972 DOI: 10.2337/diacare.23.1.74]
- 76 **Ding EL**, Song Y, Manson JE, Rifai N, Buring JE, Liu S. Plasma sex steroid hormones and risk of developing type 2 diabetes in women: a prospective study. *Diabetologia* 2007; **50**: 2076-2084 [PMID: 17701157 DOI: 10.1007/s00125-007-0785-y]
- 77 **Kalyani RR**, Franco M, Dobs AS, Ouyang P, Vaidya D, Bertoni A, Gapstur SM, Golden SH. The association of endogenous sex hormones, adiposity, and insulin resistance with incident diabetes in postmenopausal women. *J Clin Endocrinol Metab* 2009; **94**: 4127-4135 [PMID: 19789205 DOI: 10.1210/jc.2009-0910]
- 78 **Hu J**, Zhang A, Yang S, Wang Y, Goswami R, Zhou H, Zhang Y, Wang Z, Li R, Cheng Q, Zhen Q, Li Q. Combined effects of sex hormone-binding globulin and sex hormones on risk of incident type 2 diabetes. *J Diabetes* 2016; **8**: 508-515 [PMID: 26119029 DOI: 10.1111/1753-0407.12322]
- 79 **Oh JY**, Barrett-Connor E, Wedick NM, Wingard DL. Endogenous sex hormones and the development of type 2 diabetes in older men and women: the Rancho Bernardo study. *Diabetes Care* 2002; **25**: 55-60 [PMID: 11772901 DOI: 10.2337/diacare.25.1.55]
- 80 **Vikan T**, Schirmer H, Njølstad I, Svartberg J. Low testosterone and sex hormone-binding globulin levels and high estradiol levels are independent predictors of type 2 diabetes in men. *Eur J Endocrinol* 2010; **162**: 747-754 [PMID: 20061333 DOI: 10.1530/EJE-09-0943]
- 81 **Cauley JA**, Gutai JP, Kuller LH, LeDonne D, Powell JG. The epidemiology of serum sex hormones in postmenopausal women. *Am J Epidemiol* 1989; **129**: 1120-1131 [PMID: 2729251 DOI: 10.1016/0378-5122(89)90030-3]
- 82 **Mayes JS**, Watson GH. Direct effects of sex steroid hormones on adipose tissues and obesity. *Obes Rev* 2004; **5**: 197-216 [PMID: 15458395 DOI: 10.1111/j.1467-789X.2004.00152.x]
- 83 **Turgeon JL**, Carr MC, Maki PM, Mendelsohn ME, Wise PM. Complex actions of sex steroids in adipose tissue, the cardiovascular system, and brain: Insights from basic science and clinical studies. *Endocr Rev* 2006; **27**: 575-605 [PMID: 16763155 DOI: 10.1210/er.2005-0020]
- 84 **Paik SG**, Michelis MA, Kim YT, Shin S. Induction of insulin-dependent diabetes by streptozotocin. Inhibition by estrogens and potentiation by androgens. *Diabetes* 1982; **31**: 724-729 [PMID: 6219020 DOI: 10.2337/diab.31.8.724]
- 85 **Heine PA**, Taylor JA, Iwamoto GA, Lubahn DB, Cooke PS. Increased adipose tissue in male and female estrogen receptor-alpha knockout mice. *Proc Natl Acad Sci USA* 2000; **97**: 12729-12734 [PMID: 11070086 DOI: 10.1073/pnas.97.23.12729]
- 86 **Andersson B**, Mattsson LA, Hahn L, Mårin P, Lapidus L, Holm G, Bengtsson BA, Björntorp P. Estrogen replacement therapy decreases hyperandrogenicity and improves glucose homeostasis and plasma lipids in postmenopausal women with noninsulin-dependent diabetes mellitus. *J Clin Endocrinol Metab* 1997; **82**: 638-643 [PMID: 9024268 DOI: 10.1210/jcem.82.2.3746]
- 87 **Moran LJ**, Misso ML, Wild RA, Norman RJ. Impaired glucose tolerance, type 2 diabetes and metabolic syndrome in polycystic ovary syndrome: a systematic review and meta-analysis. *Hum Reprod Update* 2010; **16**: 347-363 [PMID: 20159883 DOI: 10.1093/humupd/dmq001]
- 88 **Dhindsa S**, Prabhakar S, Sethi M, Bandyopadhyay A, Chaudhuri A, Dandona P. Frequent occurrence of hypogonadotropic hypogonadism in type 2 diabetes. *J Clin Endocrinol Metab* 2004; **89**: 5462-5468 [PMID: 15531498 DOI: 10.1210/jc.2004-0804]
- 89 **Rhoden EL**, Ribeiro EP, Teloken C, Souto CA. Diabetes mellitus is associated with subnormal serum levels of free testosterone in men.



- BJU Int* 2005; **96**: 867-870 [PMID: 16153219 DOI: 10.1111/j.1464-410X.2005.05728.x]
- 90 **Stellato RK**, Feldman HA, Hamdy O, Horton ES, McKinlay JB. Testosterone, sex hormone-binding globulin, and the development of type 2 diabetes in middle-aged men: prospective results from the Massachusetts male aging study. *Diabetes Care* 2000; **23**: 490-494 [PMID: 10857940 DOI: 10.2337/diacare.23.4.490]
  - 91 **Keating NL**, O'Malley A, Freedland SJ, Smith MR. Diabetes and cardiovascular disease during androgen deprivation therapy: observational study of veterans with prostate cancer. *J Natl Cancer Inst* 2012; **104**: 1518-1523 [PMID: 23210129 DOI: 10.1200/JCO.2006.06.2497]
  - 92 **Laaksonen DE**, Niskanen L, Punnonen K, Nyssönen K, Tuomainen TP, Valkonen VP, Salonen R, Salonen JT. Testosterone and sex hormone-binding globulin predict the metabolic syndrome and diabetes in middle-aged men. *Diabetes Care* 2004; **27**: 1036-1041 [PMID: 15111517 DOI: 10.2337/diacare.27.5.1036]
  - 93 **Corona G**, Monami M, Rastrelli G, Aversa A, Sforza A, Lenzi A, Forti G, Mannucci E, Maggi M. Type 2 diabetes mellitus and testosterone: a meta-analysis study. *Int J Androl* 2011; **34**: 528-540 [PMID: 20969599 DOI: 10.1111/j.1365-2605.2010.01117.x]
  - 94 **Foryst-Ludwig A**, Kintscher U. Metabolic impact of estrogen signalling through ERalpha and ERbeta. *J Steroid Biochem Mol Biol* 2010; **122**: 74-81 [PMID: 20599505 DOI: 10.1016/j.jsbmb.2010.06.012]
  - 95 **Fuente-Martin E**, Garcia-Caceres C, Morselli E, Clegg DJ, Chowen JA, Finan B, Brinton RD, Tschöp MH. Estrogen, astrocytes and the neuroendocrine control of metabolism. *Rev Endocr Metab Disord* 2013; **14**: 331-338 [PMID: 24009071 DOI: 10.1007/s1154-013-9263-7]
  - 96 **Azcoitia I**, Sierra A, Garcia-Segura LM. Neuroprotective effects of estradiol in the adult rat hippocampus: interaction with insulin-like growth factor-I signalling. *J Neurosci Res* 1999; **58**: 815-822 [PMID: 10583912 DOI: 10.1002/(SICI)1097-4547(19991215)58: ]
  - 97 **Mendez P**, Azcoitia I, Garcia-Segura LM. Estrogen receptor alpha forms estrogen-dependent multimolecular complexes with insulin-like growth factor receptor and phosphatidylinositol 3-kinase in the adult rat brain. *Brain Res Mol Brain Res* 2003; **112**: 170-176 [PMID: 12670715 DOI: 10.1016/S0169-328X(03)00088-3]
  - 98 **D'Astous M**, Mendez P, Morissette M, Garcia-Segura LM, Di Paolo T. Implication of the phosphatidylinositol-3 kinase/protein kinase B signaling pathway in the neuroprotective effect of estradiol in the striatum of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mice. *Mol Pharmacol* 2006; **69**: 1492-1498 [PMID: 16434614 DOI: 10.1124/mol.105.018671]
  - 99 **Quesada A**, Lee BY, Micevych PE. PI3 kinase/Akt activation mediates estrogen and IGF-1 nigral DA neuronal neuroprotection against a unilateral rat model of Parkinson's disease. *Dev Neurobiol* 2008; **68**: 632-644 [PMID: 18278798 DOI: 10.1002/dneu.20609]
  - 100 **Cardona-Gomez GP**, Mendez P, Garcia-Segura LM. Synergistic interaction of estradiol and insulin-like growth factor-I in the activation of PI3K/Akt signaling in the adult rat hypothalamus. *Brain Res Mol Brain Res* 2002; **107**: 80-88 [PMID: 12414126 DOI: 10.1016/S0169-328X(02)00449-7]
  - 101 **Marin R**, Diaz M, Alonso R, Sanz A, Arévalo MA, Garcia-Segura LM. Role of estrogen receptor alpha in membrane-initiated signaling in neural cells: interaction with IGF-1 receptor. *J Steroid Biochem Mol Biol* 2009; **114**: 2-7 [PMID: 19167493 DOI: 10.1016/j.jsbmb.2008.12.014]
  - 102 **Cardona-Gómez GP**, DonCarlos L, Garcia-Segura LM. Insulin-like growth factor I receptors and estrogen receptors colocalize in female rat brain. *Neuroscience* 2000; **99**: 751-760 [PMID: 10974438 DOI: 10.1016/S0306-4522(00)00228-1]
  - 103 **Cardona-Gomez P**, Perez M, Avila J, Garcia-Segura LM, Wadosell F. Estradiol inhibits GSK3 and regulates interaction of estrogen receptors, GSK3, and beta-catenin in the hippocampus. *Mol Cell Neurosci* 2004; **25**: 363-373 [PMID: 15033165 DOI: 10.1016/j.mcn.2003.10.008]
  - 104 **Mendez P**, Wadosell F, Garcia-Segura LM. Cross-talk between estrogen receptors and insulin-like growth factor-I receptor in the brain: cellular and molecular mechanisms. *Front Neuroendocrinol* 2006; **27**: 391-403 [PMID: 17049974 DOI: 10.1016/j.yfme.2006.09.001]
  - 105 **Mendez P**, Azcoitia I, Garcia-Segura LM. Interdependence of oestrogen and insulin-like growth factor-I in the brain: potential for analysing neuroprotective mechanisms. *J Endocrinol* 2005; **185**: 11-17 [PMID: 15817823 DOI: 10.1677/joe.1.06058]
  - 106 **Varea O**, Arevalo MA, Garrido JJ, Garcia-Segura LM, Wadosell F, Mendez P. Interaction of estrogen receptors with insulin-like growth factor-I and Wnt signaling in the nervous system. *Steroids* 2010; **75**: 565-569 [PMID: 19778547 DOI: 10.1016/j.steroids.2009.09.006]
  - 107 **Brywe KG**, Mallard C, Gustavsson M, Hedtjörn M, Leverin AL, Wang X, Blomgren K, Isgaard J, Hagberg H. IGF-I neuroprotection in the immature brain after hypoxia-ischemia, involvement of Akt and GSK3beta? *Eur J Neurosci* 2005; **21**: 1489-1502 [PMID: 15845077 DOI: 10.1111/j.1460-9568.2005.03982.x]
  - 108 **Dhillon AS**, Hagan S, Rath O, Kolch W. MAP kinase signalling pathways in cancer. *Oncogene* 2007; **26**: 3279-3290 [PMID: 17496922 DOI: 10.1038/sj.onc.1210421]
  - 109 **Cardona-Gómez GP**, Mendez P, DonCarlos LL, Azcoitia I, Garcia-Segura LM. Interactions of estrogen and insulin-like growth factor-I in the brain: molecular mechanisms and functional implications. *J Steroid Biochem Mol Biol* 2002; **83**: 211-217 [PMID: 12650718 DOI: 10.1016/S0960-0760(02)00261-3]
  - 110 **Convit A**, Wolf OT, Tarshish C, de Leon MJ. Reduced glucose tolerance is associated with poor memory performance and hippocampal atrophy among normal elderly. *Proc Natl Acad Sci USA* 2003; **100**: 2019-2022 [PMID: 12571363 DOI: 10.1073/pnas.0336073100]
  - 111 **Gold SM**, Dziobek I, Sweat V, Tirsi A, Rogers K, Bruehl H, Tsui W, Richardson S, Javier E, Convit A. Hippocampal damage and memory impairments as possible early brain complications of type 2 diabetes. *Diabetologia* 2007; **50**: 711-719 [PMID: 17334649 DOI: 10.1007/s00125-007-0602-7]
  - 112 **Kamiyama K**, Wada A, Sugihara M, Kurioka S, Hayashi K, Hayashi T, Yoshisako T, Yamamoto N, Tsuchie Y, Yamaguchi S, Sugimoto T, Kitagaki H. Potential hippocampal region atrophy in diabetes mellitus type 2: a voxel-based morphometry VSRAD study. *Jpn J Radiol* 2010; **28**: 266-272 [PMID: 20512543 DOI: 10.1007/s11604-009-0416-2]
  - 113 **Nelson BS**, Springer RC, Daniel JM. Antagonism of brain insulin-like growth factor-I receptors blocks estradiol effects on memory and levels of hippocampal synaptic proteins in ovariectomized rats. *Psychopharmacology (Berl)* 2014; **231**: 899-907 [PMID: 24146138 DOI: 10.1007/s00213-013-3310-7]
  - 114 **Perez-Martin M**, Azcoitia I, Trejo JL, Sierra A, Garcia-Segura LM. An antagonist of estrogen receptors blocks the induction of adult neurogenesis by insulin-like growth factor-I in the dentate gyrus of adult female rat. *Eur J Neurosci* 2003; **18**: 923-930 [PMID: 12925018 DOI: 10.1046/j.1460-9568.2003.02830.x]
  - 115 **Takeuchi K**, Yang Y, Takayasu Y, Gertner M, Hwang JY, Aromolaran K, Bennett MV, Zukin RS. Estradiol pretreatment ameliorates impaired synaptic plasticity at synapses of insulted CA1 neurons after transient global ischemia. *Brain Res* 2015; **1621**: 222-230 [PMID: 25463028 DOI: 10.1016/j.brainres.2014.11.016]
  - 116 **Witty CF**, Gardella LP, Perez MC, Daniel JM. Short-term estradiol administration in aging ovariectomized rats provides lasting benefits for memory and the hippocampus: a role for insulin-like growth factor-I. *Endocrinology* 2013; **154**: 842-852 [PMID: 23264616 DOI: 10.1210/en.2012-1698]
  - 117 **Cardona-Gómez GP**, Mendez P, DonCarlos LL, Azcoitia I, Garcia-Segura LM. Interactions of estrogens and insulin-like growth factor-I in the brain: implications for neuroprotection. *Brain Res Brain Res Rev* 2001; **37**: 320-334 [PMID: 11744097 DOI: 10.1016/S0165-0173(01)00137-0]
  - 118 **Aggarwal RR**, Ryan CJ, Chan JM. Insulin-like growth factor pathway: a link between androgen deprivation therapy (ADT), insulin resistance, and disease progression in patients with prostate cancer? *Urol Oncol* 2013; **31**: 522-530 [PMID: 21658978 DOI: 10.1016/j.urolonc.2011.05.001]
  - 119 **Oki K**, Law TD, Loucks AB, Clark BC. The effects of testosterone

- and insulin-like growth factor 1 on motor system form and function. *Exp Gerontol* 2015; **64**: 81-86 [PMID: 25681641 DOI: 10.1016/j.exger.2015.02.005]
- 120 **Castilla-Cortázar I**, García-Fernández M, Delgado G, Puche JE, Sierra I, Barhoum R, González-Barón S. Hepatoprotection and neuroprotection induced by low doses of IGF-II in aging rats. *J Transl Med* 2011; **9**: 103 [PMID: 21733157 DOI: 10.1186/1479-5876-9-103]
  - 121 **Lin SY**, Cui H, Yusta B, Belsham DD. IGF-I signaling prevents dehydroepiandrosterone (DHEA)-induced apoptosis in hypothalamic neurons. *Mol Cell Endocrinol* 2004; **214**: 127-135 [PMID: 15062551 DOI: 10.1016/j.mce.2003.10.064]
  - 122 **Bialek M**, Zaremba P, Borowicz KK, Czuczwar SJ. Neuroprotective role of testosterone in the nervous system. *Pol J Pharmacol* 2004; **56**: 509-518 [PMID: 15591638]
  - 123 **Creta M**, Riccio R, Chiancone F, Fusco F. Androgens exert direct neuroprotective effects on the brain: a review of pre-clinical evidences. *J Androl Sci* 2010; **17**: 49-55
  - 124 **Garcia-Segura LM**, Balthazart J. Steroids and neuroprotection: New advances. *Front Neuroendocrinol* 2009; **30**: v-ix [PMID: 19393683 DOI: 10.1016/j.yfrne.2009.04.006]
  - 125 **Hammond J**, Le Q, Goodyer C, Gelfand M, Trifiro M, LeBlanc A. Testosterone-mediated neuroprotection through the androgen receptor in human primary neurons. *J Neurochem* 2001; **77**: 1319-1326 [PMID: 11389183 DOI: 10.1046/j.1471-4159.2001.00345.x]
  - 126 **Nguyen TV**, Jayaraman A, Quaglini A, Pike CJ. Androgens selectively protect against apoptosis in hippocampal neurones. *J Neuroendocrinol* 2010; **22**: 1013-1022 [PMID: 20561156 DOI: 10.1111/j.1365-2826.2010.02044.x]
  - 127 **Baron S**, Manin M, Beaudoin C, Leotoing L, Communal Y, Veysiere G, Morel L. Androgen receptor mediates non-genomic activation of phosphatidylinositol 3-OH kinase in androgen-sensitive epithelial cells. *J Biol Chem* 2004; **279**: 14579-14586 [PMID: 14668339 DOI: 10.1074/jbc.M306143200]
  - 128 **Rochette-Egly C**. Nuclear receptors: integration of multiple signalling pathways through phosphorylation. *Cell Signal* 2003; **15**: 355-366 [PMID: 12618210 DOI: 10.1016/S0898-6568(02)00115-8]
  - 129 **Bing L**, Wu J, Zhang J, Chen Y, Hong Z, Zu H. DHT inhibits the A $\beta$ 25-35-induced apoptosis by regulation of seladin-1, survivin, XIAP, bax, and bcl-xl expression through a rapid PI3-K/Akt signaling in C6 glial cell lines. *Neurochem Res* 2015; **40**: 41-48 [PMID: 25347962 DOI: 10.1007/s11064-014-1463-3]
  - 130 **Gatson JW**, Kaur P, Singh M. Dihydrotestosterone differentially modulates the mitogen-activated protein kinase and the phosphoinositide 3-kinase/Akt pathways through the nuclear and novel membrane androgen receptor in C6 cells. *Endocrinology* 2006; **147**: 2028-2034 [PMID: 16410299 DOI: 10.1210/en.2005-1395]
  - 131 **Gatson JW**, Singh M. Activation of a membrane-associated androgen receptor promotes cell death in primary cortical astrocytes. *Endocrinology* 2007; **148**: 2458-2464 [PMID: 17303658 DOI: 10.1210/en.2006-1443]
  - 132 **Nguyen TV**, Yao M, Pike CJ. Androgens activate mitogen-activated protein kinase signaling: role in neuroprotection. *J Neurochem* 2005; **94**: 1639-1651 [PMID: 16011741 DOI: 10.1111/j.1471-4159.2005.03318.x]
  - 133 **Pike CJ**, Nguyen TV, Ramsden M, Yao M, Murphy MP, Rosario ER. Androgen cell signaling pathways involved in neuroprotective actions. *Horm Behav* 2008; **53**: 693-705 [PMID: 18222446 DOI: 10.1016/j.yhbeh.2007.11.006]
  - 134 **Azcoitia I**, Sierra A, Veiga S, Honda S, Harada N, Garcia-Segura LM. Brain aromatase is neuroprotective. *J Neurobiol* 2001; **47**: 318-329 [PMID: 11351342 DOI: 10.1002/neu.1038]
  - 135 **Azcoitia I**, Sierra A, Veiga S, Garcia-Segura LM. Aromatase expression by reactive astroglia is neuroprotective. *Ann N Y Acad Sci* 2003; **1007**: 298-305 [PMID: 14993062 DOI: 10.1196/annals.1286.028]
  - 136 **Garcia-Segura LM**, Veiga S, Sierra A, Melcangi RC, Azcoitia I. Aromatase: a neuroprotective enzyme. *Prog Neurobiol* 2003; **71**: 31-41 [PMID: 14611865 DOI: 10.1016/j.pneurobio.2003.09.005]
  - 137 **Garcia-Segura LM**. Aromatase in the brain: not just for reproduction anymore. *J Neuroendocrinol* 2008; **20**: 705-712 [PMID: 18601693 DOI: 10.1111/j.1365-2826.2008.01713.x]
  - 138 **Roselli CF**. Brain aromatase: roles in reproduction and neuroprotection. *J Steroid Biochem Mol Biol* 2007; **106**: 143-150 [PMID: 17643294 DOI: 10.1016/j.jsmb.2007.05.014]
  - 139 **Saldanha CJ**, Duncan KA, Walters BJ. Neuroprotective actions of brain aromatase. *Front Neuroendocrinol* 2009; **30**: 106-118 [PMID: 19450619 DOI: 10.1016/j.yfrne.2009.04.016]
  - 140 **Carswell HV**, Dominiczak AF, Garcia-Segura LM, Harada N, Hutchison JB, Macrae IM. Brain aromatase expression after experimental stroke: topography and time course. *J Steroid Biochem Mol Biol* 2005; **96**: 89-91 [PMID: 15896953 DOI: 10.1016/j.jsmb.2005.02.016]
  - 141 **Burul-Bozkurt N**, Pekiner C, Kelicen P. Diabetes alters aromatase enzyme levels in gonadal tissues of rats. *Naunyn Schmiedebergs Arch Pharmacol* 2010; **382**: 33-41 [PMID: 20428845 DOI: 10.1007/s00210-010-0518-5]
  - 142 **Gibb FW**, Homer NZ, Faqehi AM, Upreti R, Livingstone DE, McInnes KJ, Andrew R, Walker BR. Aromatase Inhibition Reduces Insulin Sensitivity in Healthy Men. *J Clin Endocrinol Metab* 2016; **101**: 2040-2046 [PMID: 26967690 DOI: 10.1210/jc.2015-4146]
  - 143 **Pintana H**, Chattipakorn N, Chattipakorn S. Testosterone deficiency, insulin-resistant obesity and cognitive function. *Metab Brain Dis* 2015; **30**: 853-876 [PMID: 25703239 DOI: 10.1007/s11011-015-9655-3]
  - 144 **Dozmorov MG**, Yang Q, Matwalli A, Hurst RE, Culkin DJ, Kropp BP, Lin HK. 5 $\alpha$ -androstane-3 $\alpha$ ,17 $\beta$ -diol selectively activates the canonical PI3K/AKT pathway: a bioinformatics-based evidence for androgen-activated cytoplasmic signaling. *Genomic Med* 2007; **1**: 139-146 [PMID: 18923939 DOI: 10.1007/s11568-008-9018-9]
  - 145 **Narenji SA**, Naghdi N, Azadmanesh K, Edalat R. 3 $\alpha$ -diol administration decreases hippocampal PKA (II) mRNA expression and impairs Morris water maze performance in adult male rats. *Behav Brain Res* 2015; **280**: 149-159 [PMID: 25451551 DOI: 10.1016/j.bbr.2014.11.038]
  - 146 **Caruso D**, Scurati S, Maschi O, De Angelis L, Roglio I, Giatti S, Garcia-Segura LM, Melcangi RC. Evaluation of neuroactive steroid levels by liquid chromatography-tandem mass spectrometry in central and peripheral nervous system: effect of diabetes. *Neurochem Int* 2008; **52**: 560-568 [PMID: 17686551 DOI: 10.1016/j.neuint.2007.06.004]

**P- Reviewer:** Comasco E, Egea J, Phillips J **S- Editor:** Qiu S  
**L- Editor:** A **E- Editor:** Wu HL



## Basic Study

# High fat diet dysregulates microRNA-17-5p and triggers retinal inflammation: Role of endoplasmic-reticulum-stress

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**Author contributions:** All the authors contributed to the manuscript.

**Institutional review board statement:** The study was reviewed and approved by the Charlie Norwood VA Medical Center Institutional Review Board.

**Institutional animal care and use committee statement:** All procedures involving animals were reviewed and approved by the Association for Research in Vision and Ophthalmology statement for use of animals in ophthalmic and vision research, and Charlie Norwood VA Medical Center Animal Care and Use Committee (ACORP#15-04-080)

**Conflict-of-interest statement:** There is no conflict of interest.

**Data sharing statement:** Data will be available upon request.

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**Manuscript source:** Invited manuscript

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Received: September 2, 2016

Peer-review started: September 6, 2016

First decision: September 29, 2016

Revised: October 22, 2016

Accepted: December 13, 2016

Article in press: December 14, 2016

Published online: February 15, 2017

## Abstract

### AIM

To elucidate how high diet-induced endoplasmic reticulum-stress upregulates thioredoxin interacting protein expression in Müller cells leading to retinal inflammation.

### METHODS

Male C57Bl/J mice were fed either normal diet or 60% high fat diet for 4-8 wk. During the 4 wk study, mice received phenyl-butyric acid (PBA); endoplasmic reticulum-stress inhibitor; for 2 wk. Insulin resistance was assessed by oral glucose tolerance. Effects of palmitate-bovine serum albumin (BSA) (400  $\mu$ mol/L) were examined in retinal Müller glial cell line and primary Müller cells isolated from wild type and thioredoxin interacting protein knock-out mice. Expression of thioredoxin interacting protein, endoplasmic reticulum-stress markers, miR-17-5p mRNA, as well as nucleotide-binding oligomerization domain-like receptor protein (NLRP3) and IL1 $\beta$  protein was determined.

### RESULTS

High fat diet for 8 wk induced obesity and insulin resistance evident by increases in body weight and impaired glucose tolerance. By performing quantitative real-time polymerase chain reaction, we found that high fat diet triggered the expression of retinal endoplasmic reticulum-stress markers ( $P < 0.05$ ). These effects were associated with increased thioredoxin interacting protein and decreased miR-17-5p expression, which

were restored by inhibiting endoplasmic reticulum-stress with PBA ( $P < 0.05$ ). *In vitro*, palmitate-BSA triggered endoplasmic reticulum-stress markers, which was accompanied with reduced miR-17-5p and induced thioredoxin interacting protein mRNA in retinal Müller glial cell line ( $P < 0.05$ ). Palmitate upregulated NLRP3 and IL1 $\beta$  expression in primary Müller cells isolated from wild type. However, using primary Müller cells isolated from thioredoxin interacting protein knock-out mice abolished palmitate-mediated increase in NLRP3 and IL1 $\beta$ .

### CONCLUSION

Our work suggests that targeting endoplasmic reticulum-stress or thioredoxin interacting protein are potential therapeutic strategies for early intervention of obesity-induced retinal inflammation.

**Key words:** High fat diet; Palmitate; Endoplasmic-reticulum-stress; Inflammation; Thioredoxin-interacting protein; Micro-RNA 17-5p

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**Core tip:** We previously showed that high fat diets (HFD) induced retinal inflammation and vascular dysfunction. These results were associated with an increase in thioredoxin interacting protein (TXNIP) at the mRNA and protein level. Here, we examined the mechanisms by which HFD triggers retinal TXNIP. Interestingly, we found that HFD/palmitate triggers ER-stress mediators including the inositol requiring enzyme 1, an RNase that can degrade number of mRNAs including the microRNA; miR-17-5p and sustains TXNIP expression. Inhibiting ER-stress prevented the increase in TXNIP *in vivo* and in Müller cells, the main glia in the retina. Deletion of TXNIP blunted NLRP-3 inflammasome and IL-1 $\beta$  release in Müller cells.

Coucha M, Mohamed IN, Elshaer SL, Mbata O, Bartasis ML, El-Remessy AB. High fat diet dysregulates microRNA-17-5p and triggers retinal inflammation: Role of endoplasmic-reticulum-stress. *World J Diabetes* 2017; 8(2): 56-65 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v8/i2/56.htm> DOI: <http://dx.doi.org/10.4239/wjd.v8.i2.56>

### INTRODUCTION

Obesity, recently upgraded from a mere risk factor to a disease state, is affecting one third of United States population<sup>[1]</sup>. Clinical evidence showed that obesity not only can accelerate developing type-2 diabetes and cardiovascular complications, but also induce retinal microvascular abnormalities, which eventually leads to visual impairments<sup>[2,3]</sup>. High fat diets (HFD) together with the improper physical activity are the culprit in

the obesity-induced pre-diabetes. Therefore, there is an urgent need to unravel the mechanisms involved in HFD-mediated neurovascular abnormalities. Our lab has previously shown that consumption of high caloric diet saturated fatty acids induced retinal inflammation and microvascular dysfunction *via* upregulating the expression of thioredoxin interacting protein (TXNIP); a regulator of the antioxidant thioredoxin; and activating NOD (NOD)-like receptor protein (NLRP3)-inflammasome<sup>[4]</sup>. Similar observations showed the contribution of TXNIP/NLRP3-inflammasome signaling pathway to the development of various disorders in other organs<sup>[5-7]</sup>. However, molecular mechanisms by which HFD triggers early TXNIP expression in the retina are still unclear.

MicroRNAs are small non-coding RNAs that control the translation and transcription of various genes *via* annealing to the complementary sequences in the 3' untranslated region of their target gene<sup>[8]</sup>. To date, several miR classes have been identified to be involved in development of obesity, diabetes and diabetic complications<sup>[9]</sup>. Bioinformatic analysis of the TXNIP 3' UTR identified 11 possible miRNAs that can regulate its expression including miR-130/301, miR-128, miR-148/152, miR-135, miR-106/302, miR-17-5p/20/93.mr/106/519. d, miR-128, miR-15/16/195/424/497, miR-106/302, miR-148/152. Nevertheless, levels of miR-17-5p have been reported to rapidly decline under stress condition resulting in enhancing TXNIP expression<sup>[10,11]</sup>.

Unfolded protein response (UPR) is an adaptive response, which prevents the accumulation of misfolded proteins in the lumen of the endoplasmic reticulum (ER). The UPR is transduced by three major ER-resident stress sensors, namely Protein Kinase RNA-like ER kinase (PERK), activating transcription factor 6 (ATF6), and inositol requiring enzyme 1 (IRE1). However, when protein misfolding exceeds the capacity of the UPR an ER-stress will result that triggers programmed cell death. So far, ER-stress has been shown to play a critical role in the pathogenic progression of various chronic diseases including diabetic retinopathy (reviewed in<sup>[12-14]</sup>). Among UPR pathways, IRE1 $\alpha$ , an ER bifunctional kinase/RNase has been shown to destabilize number of RNA and microRNA including miR-17-5p in pancreatic beta-cells<sup>[10,11]</sup>. Several studies reported the impact of HFD and its related metabolite such as free fatty acid in inducing ER-stress<sup>[15-17]</sup>. In the current study we were trying to decipher the underlying mechanisms that link HFD-mediated ER-stress to retinal inflammation. Here, we tested the hypothesis that HFD-mediated ER-stress upregulates TXNIP mRNA expression *via* dysregulating miR-17-5p resulting in retinal inflammation.

### MATERIALS AND METHODS

#### Animals

All animal experiments were conducted in agreement with Association for Research in Vision and Ophthalmology



**Table 1** The sequence of the polymerase chain reaction primers used in the experiments

Gene	Forward	Reverse
<i>18S</i>	CGCGGTTCTATTTTGTGGT	AGTCGGCATCGTTTATGGTC
<i>XBPI</i>	ACACGCTTGGGAATGGACAC	CCATGGGAAGATGTTCTGGG
<i>XBPI-SPLICED</i>	GAGTCCGCAGCAGGTG	GTGTCAGAGTCCATGGGA
<i>PERK</i>	AGTCCCTGCTCGAATCTTCCT	TCCCAAGGCAGAACAGATATACC
<i>IRE1α</i>	GGGTGCTGTCGTGCCTCGAG	TGGGGGCCTTCCAGCAAAGGA
<i>ATF6</i>	TGCTTGGGAGTCAGACCTAT	GCTGAGTGAAGAACACGAGTC
<i>CHOP</i>	CTGGAAGCCTGGTATGAGGAT	CAGGGTCAAGAGTAGTGAAGGT
<i>TXNIP</i>	AAGCTGTCCTCAGTCAGAGGCAAT	ATGACTTTCTTGGAGCCAGGGACA

statement for use of animals in ophthalmic and vision research, and Charlie Norwood VA Medical Center Animal Care and Use Committee (ACORP#15-04-080). 6-8 wk old male C57BL6/J mice (Stock 000664, Jackson Laboratory, ME, United States) were used in the *in vivo* studies. For the long term study, mice were fed ad libitum with normal rat chow (7% fat) or HFD [36 g %, 251 kJ (60 kcal) %fat] (F2685 Bioserv, Frenchtown, NJ, United States) for 8 wk. For the short term study, mice were fed either normal diet (ND) or 60% HFD for 2 wk. Mice were then kept on HFD for additional 2 wk while receiving an ER-stress inhibitor [Phenyl-butyric acid (PBA), 100 mg/kg] or vehicle. PBA was dissolved in DMSO/PBS and administered *via* oral gavage 5 d/wk. Mice were weighed weekly to track the increase in the body weight.

#### **Intra-peritoneal glucose tolerance test**

Mice went overnight fasting, and their fasting plasma blood glucose was measured as the baseline. Then all mice received an intraperitoneal injection of glucose (2 g/kg). Blood glucose levels were measured at different time points till 120 min after the glucose injection using a glucometer.

#### **In-vitro studies**

The rat retinal Müller glial cell line (rMC-1) was obtained originally from V. Sarthy (Department of Ophthalmology, Northwestern University, Chicago, IL, United States)<sup>[18]</sup>. Primary mouse Müller Cells from WT and TKO mice were isolated and cultured as described previously<sup>[19]</sup>. Cells were grown to confluency in complete media (DMEM, 10% vol/vol. FBS, 1% vol/vol. penicillin/streptomycin). Sodium palmitate (Cat.# P9767; Sigma-Aldrich, St. Louis, MO, United States) was dissolved in 50% ethyl alcohol, then added drop-wise to preheated 10% endotoxin- and fatty acid-free BSA (Cat.# 22070017; Bioworld, Dublin, OH) in DMEM at 50 °C to create an intermediate stock solution of palmitate coupled to BSA (Pal-BSA). Confluent cells were switched to serum-free medium for overnight then were treated for 6 h with Pal-BSA solutions (400 μmol/L final concentration). Equal volumes of 50% ethyl alcohol with BSA alone served as control. In another set of rMC-1, cells were serum starved for 4 h then treated with PBA (1 mmol/L, Cat.#P21005, Sigma-Aldrich) or IRE1α inhibitor (STF-083010, 50 μmol/L) for 2 h then palmitate was added

and kept overnight.

#### **Quantitative real-time PCR**

A one-step quantitative RT-PCR kit (Invitrogen) was used to amplify 10 ng retinal mRNA as described previously<sup>[4]</sup>. PCR primers (Table 1) were obtained from Integrated DNA Technologies (Coralville, IA, United States). Quantitative PCR was conducted using StepOnePlus qPCR system (Applied Biosystems, Life Technologies). The percent expression of various genes was normalized to 18S.

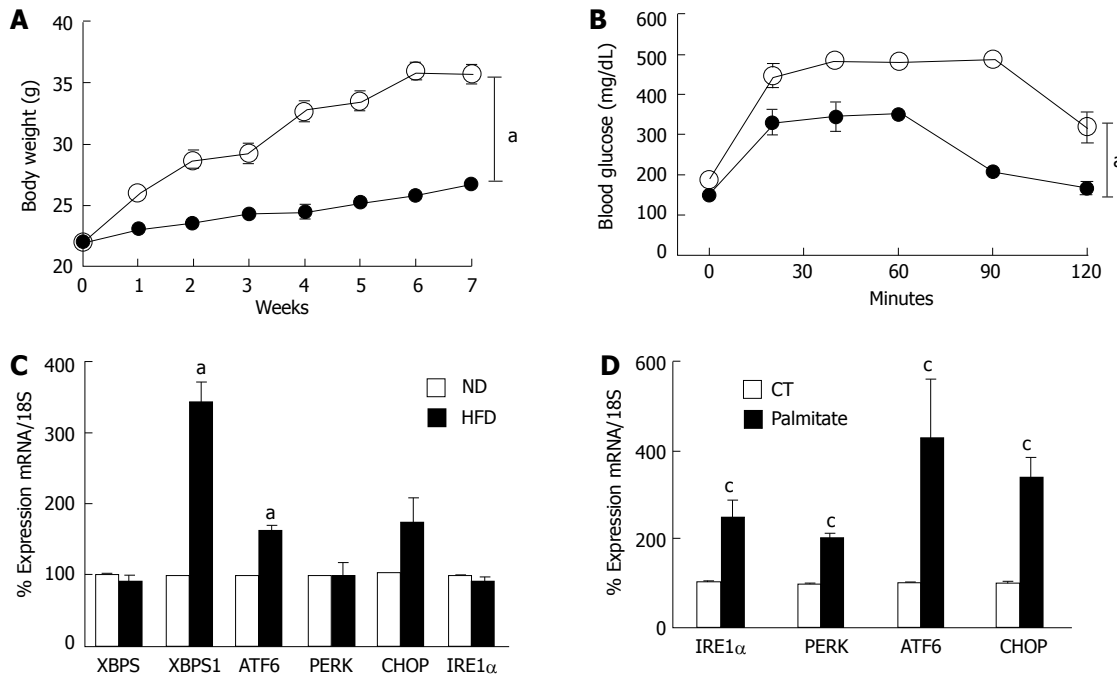
#### **Micro-RNA detection**

MirVana PARIS kit (Cat.# AM1556, Invitrogen) was used for miRNA isolation according to manufacturer's protocol. Reverse transcriptase reactions; including samples and no-template controls; were run using TaqMan® Micro-RNA Reverse Transcription Kit (Cat.# 4366596, Applied Biosystems) as described previously<sup>[20]</sup>. PCR amplification was performed using TaqMan® Universal PCR Master Mix (Cat.# 4324018, Applied Biosystems) according to manufacturer's protocol. The percent expression of miR-17-5p was normalized to U6.

#### **Western blot analysis**

Retinas were isolated and homogenized in cell disruption buffer as described previously<sup>[21]</sup>. Müller cells were harvested by scraping thoroughly with cell scraper after the addition of cell disruption buffer. Samples (25 μg protein) were separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis and transferred to a nitrocellulose membrane. Membranes were probed with the primary antibodies; anti-TXNIP (Cat.# K0205-3 MBL Abacus ALS Australia and Cat.# 403700, Invitrogen, Grand Island, NY), anti-NLRP-3 (Cat.# LS-B4321, LifeSpan Biosciences, Inc, Seattle, WA), anti-IL1β (Cat.# ab9722, Abcam, Cambridge, MA, United States) then reprobed with housekeeping gene; anti-GAPDH (Cat.# 5174, Cell Signaling, Danvers, MA, United States), anti-tubulin (Cat.# ab4074, Abcam, Cambridge, MA, United States) or anti-actin (Cat.# a5060, Sigma-Aldrich) to confirm equal loading. The primary antibody was detected using a horseradish peroxidase (HRP) and enhanced chemiluminescence. The films were scanned and the band intensity was quantified using densitometry software version 6.0.0 Software from alphaEaseFC (Santa Clara, CA) and expressed as relative





**Figure 1** High fat diet/palmitate triggered endoplasmic-reticulum-stress markers in retina and Müller cells. A: Total body weight (grams) recorded weekly was significantly higher in mice fed with HFD for 8 wk compared to ND; B: Glucose tolerance was impaired after 8 wk of HFD compared to ND; C: Realtime PCR showing increases in mRNA levels of XBP1S and ATF6, while no change in XBP1, PERK, CHOP and IRE1 $\alpha$  mRNA in retina after 8 wk of HFD compared to ND; D: Realtime PCR showing significant increases in IRE1 $\alpha$ , PERK, ATF6 and CHOP mRNA levels in rMC1 treated with palmitate compared to control (CT) (<sup>a</sup> $P < 0.05$  vs ND,  $n = 3-4$  and <sup>c</sup> $P < 0.05$  vs CT, area under the curve across all the time points was calculated,  $n = 3-4$ ). HFD: High fat diet; PERK: Protein Kinase RNA-like endoplasmic-reticulum kinase; XBP: X-box binding protein; ATF6: Activating transcription factor 6; CHOP: CCAAT-enhancer-binding protein homologous protein; IRE1: Inositol requiring enzyme 1.

optical density (OD).

### Statistical analysis

All the data are expressed as mean  $\pm$  SD or SEM. Differences between ND vs HFD and control vs palmitate were tested using two-sample *t* tests. One-way ANOVA followed by Bonferroni post-hoc multiple comparisons to assess significant differences between 3 or more groups (Graphpad-Ver.6). For body weight and blood glucose measurements, area under the curve (AUC) across all the time points was calculated. A series of 2 gene (WT vs KO)  $\times$  2 treatment (TRT) (no vs yes) ANOVAs with interaction were used to determine the effect of palmitate on NLRP3 and IL1 $\beta$ . A Bonferroni post-hoc multiple comparison test was used for significant interactions. Significance for all tests was determined at  $\alpha = 0.05$ .

## RESULTS

### HFD/palmitate triggered ER-stress markers in retina and Müller cells

Several studies showed that HFD or palmitate triggers ER-stress in different organs and cell types<sup>[17,22-24]</sup>. Therefore, we checked the levels of various ER-stress markers in the retina isolated from mice fed with HFD, and rMC1 treated with palmitate. HFD for 8 wk induced obesity and impaired glucose tolerance indicated by an increase in body weights (Figure 1A) and glucose levels (Figure 1B) across the different time points compared to ND. We also found that HFD induced an increase in

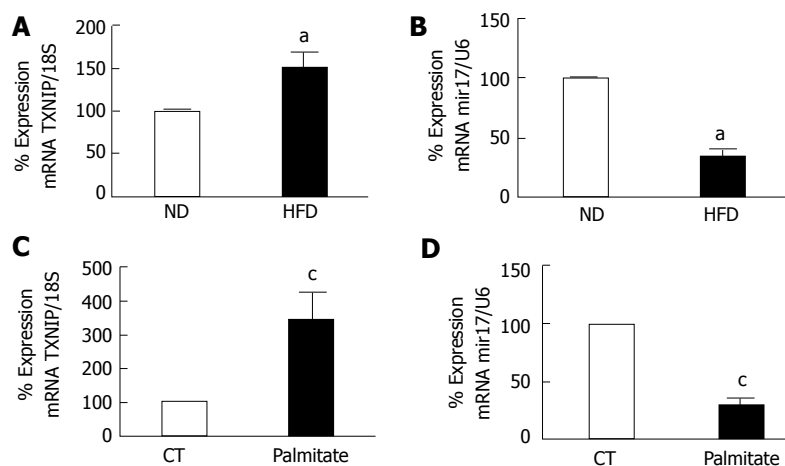
XBP1S and ATF6 mRNA levels only, while, there was no change in XBP1, PERK, CHOP and IRE1 $\alpha$  (Figure 1C). In order to study the role of Müller cells in HFD-induced inflammation, rMC-1 were treated with 400  $\mu$ mol/L palmitate coupled to bovine serum albumin (Pal-BSA) for 6hr. Palmitate; a saturated fatty acid that is increased in plasma following a HFD<sup>[25]</sup>; significantly upregulated IRE1 $\alpha$ , PERK, ATF6 and CHOP (Figure 1D).

### HFD/palmitate induced TXNIP upregulation and miR-17-5p dysregulation in retina and Müller cells

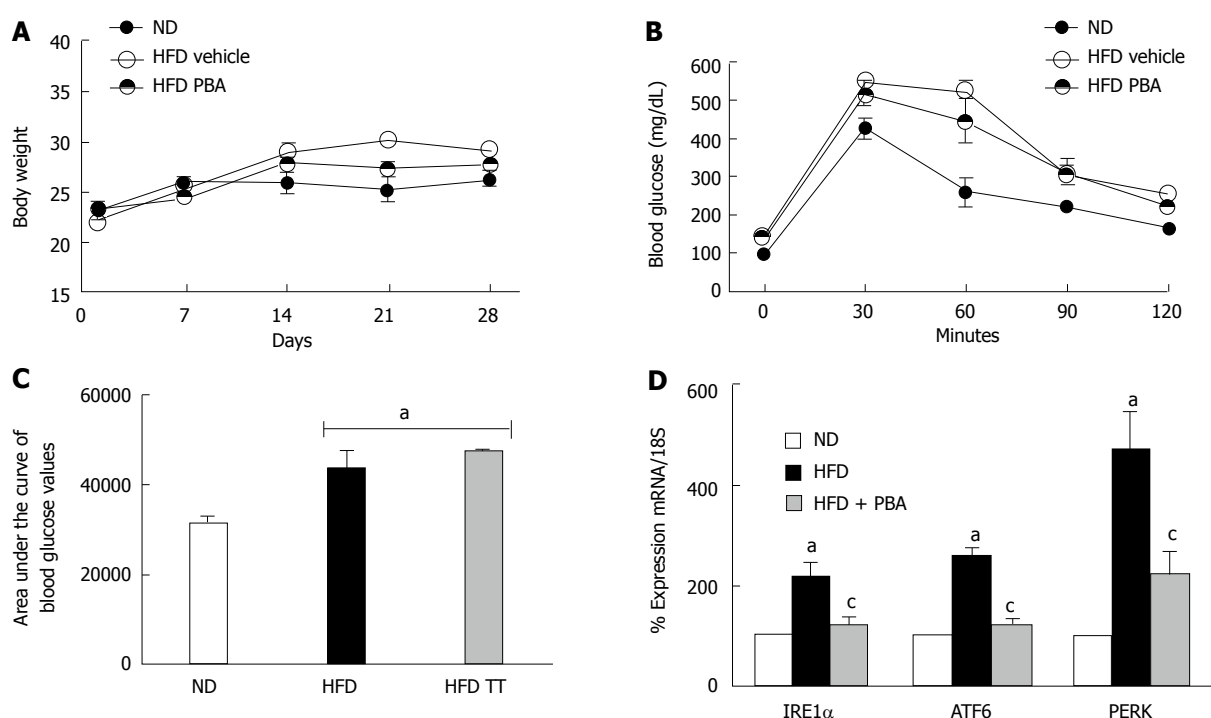
Our lab has previously reported that HFD and palmitate can induce TXNIP mRNA expression in whole retina and retina endothelial cells respectively<sup>[4]</sup>. However, the upstream events by which HFD/palmitate trigger TXNIP expression are still unclear. In agreement with the previous study, we found that 8 wk of HFD and palmitate led to an upregulation of TXNIP mRNA levels in whole retina and Müller cells (Figure 2). These results were associated with miR-17-5p dysregulation in both whole retina and Müller cells (Figure 2).

### PBA mitigated HFD-mediated ER-stress

To verify the role of ER-stress in HFD-induced TXNIP upregulation, mice were fed either ND or HFD for 2 wk. Then mice were kept on HFD for additional 2 wk while receiving PBA; an ER-stress inhibitor. Body weights were not changed by the HFD or PBA treatment (Figure 3A). However, blood glucose tolerance was significantly less in mice fed with HFD compared to ND after intra-peritoneal glucose tolerance test (Figure 3B). HFD-induced insulin



**Figure 2 Realtime polymerase chain reaction.** It shows significant (A) increase in TXNIP mRNA and (B) miR-17-5p dysregulation in retina after 8 wk of HFD compared to ND. Realtime PCR showing significant (C) increase in TXNIP mRNA levels (D) reduction in miR-17-5p in rMC1 treated with palmitate compared to control (CT) ( $^aP < 0.05$  vs ND,  $n = 3-4$  and  $^cP < 0.05$  vs CT,  $n = 3$ ). ND: Normal diet; HFD: High fat diet.



**Figure 3 PBA mitigated high fat diet-mediated endoplasmic-reticulum-stress.** A: Total body weight (g) recorded weekly for 4 wk was not changed among the different groups; B: Glucose tolerance was impaired after 4 wk of HFD compared to ND, and was not restored with PBA treatment; C: Statistical analysis of area under the curve showing an increase in blood glucose levels in HFD compared to ND, which was not reversed by the treatment; D: Realtime PCR showing significant increases in IRE1 $\alpha$ , ATF6, and PERK mRNA levels in mice kept on HFD for 4 wk compared to ND, which were nullified with PBA treatment ( $^aP < 0.05$  vs ND,  $^cP < 0.05$  vs HFD,  $n = 3-4$ ). ND: Normal diet; HFD: High fat diet; PBA: Phenyl-butyric acid; PERK: Protein Kinase RNA-like endoplasmic-reticulum kinase; IRE: Inositol requiring enzyme.

resistance suggested by marked increase in the area under the curve remained unaffected by inhibiting ER-stress with PBA (Figure 3C). HFD for 4 wk induced expression of retinal ER-stress markers mRNA including the RNase IRE1 $\alpha$ , ATF6 and PERK which were restored by PBA treatment to control level (Figure 3D).

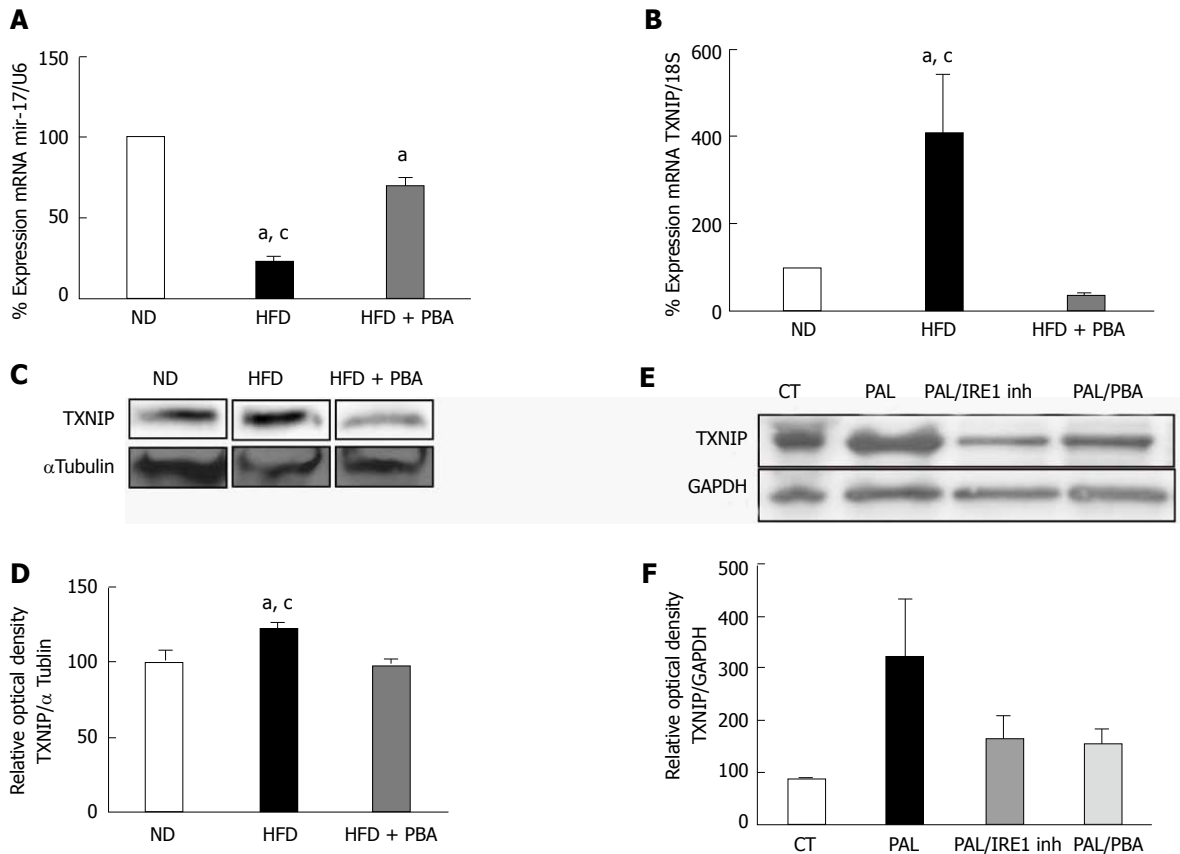
#### ER-stress inhibition prevented HFD-induced TXNIP upregulation and miR-17-5p dysregulation

To establish a causal relationship of the role of ER-stress miR-17-5p and TXNIP expression, we assessed their expression in animals that were treated with ER-stress inhibitor PBA. As shown in Figure 4A, intervention with PBA treatment in HFD partially but significantly increased

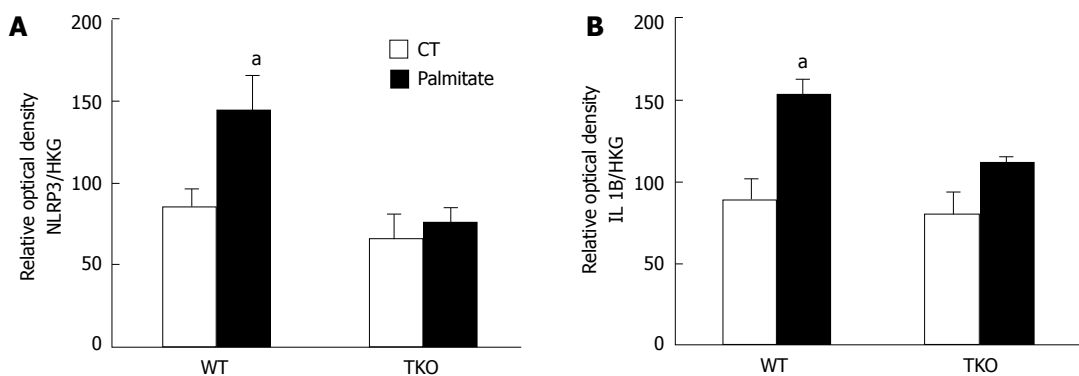
retinal miR-17-5p compared to untreated HFD. HFD triggered TXNIP mRNA and protein expression compared to ND, which were significantly inhibited in HFD-animals treated with PBA (Figure 4B-D). To establish a causal relationship of the role of ER-stress and activation of IRE1 $\alpha$  in palmitate-induced TXNIP expression, rMC1 were treated for 2 h with PBA or IRE1 $\alpha$  inhibitor prior to the addition of palmitate. As shown in Figure 4, inhibiting ER-stress or IRE1 $\alpha$  markedly reduced the increase in TXNIP protein expression in palmitate-treated cells.

#### Knocking out TXNIP abolished palmitate induced inflammation in Müller cells

We recently showed that HFD induced expression



**Figure 4 Dysregulation of realtime polymerase chain reaction.** It shows significant (A) reduction in miR-17-5p and (B) increase in TXNIP mRNA levels in mice kept on HFD for 4 wk compared to ND, which were reversed with PBA treatment ( $n = 3-4$ ); C: Representative western blot were cut from the same membrane for TXNIP and  $\alpha$ tubulin from retina (D) Statistical analysis showed an upregulation in TXNIP expression in HFD mice compared to ND, PBA treatment nullified this effect ( $n = 4-5$ ) ( $^aP < 0.05$  vs ND,  $^cP < 0.05$  vs HFD + PBA) (E) Representative western blot of TXNIP and GAPDH from rMC1 treated with palmitate (pal), after the addition of IRE inhibitor or PBA; D: Statistical analysis showed a trend increase in TXNIP expression, which is reversed by IRE inhibitor or PBA ( $P = 0.076$ ,  $n = 3$ ). HFD: High fat diet; ND: Normal diet; PBA: Phenyl-butyric acid; TXNIP: Thioredoxin-interacting protein; CT: Control; PAL: Palmitate; PAL/IRE1: Palmitate + inositol requiring enzyme 1; PAL/PBA: Palmitate + phenyl-butyric acid.



**Figure 5 Statistical analysis showed an increase in.** (A) NLRP3 and (B) IL1 $\beta$  expression following palmitate treatment in primary Müller cells isolated from WT but no effect on TKO. Actin and  $\alpha$ -tubulin were used as housekeeping genes (HKG), to which NLRP3 and IL1 $\beta$  expression was normalized ( $^aP < 0.05$  vs ND,  $n = 3$ ).

of TXNIP in Müller cells, which was associated with increased TXNIP-NLRP3 inflammasome interaction as well as the expression of cleaved caspase-1 and IL-1 $\beta$ <sup>[4]</sup>. Therefore, to dissect the role of TXNIP in palmitate-mediated inflammation in Müller cells, primary Müller cells from both WT and TKO mice were used. Primary Müller cells were serum starved overnight then treated with 400  $\mu$ mol/L palmitate coupled to bovine serum

albumin (Pal-BSA) for 6 h. We found that palmitate led to an increase in NLRP3 and IL1 $\beta$  protein expression in cells isolated from WT but has no effect on cells isolated from TKO mice (Figure 5).

## DISCUSSION

Central obesity and insulin resistance are hallmarks

of metabolic syndrome that comprises dyslipidemia, hypertriglyceridemia, hyperinsulinemia, hypertension, and reduced HDL cholesterol. Changes in lipid profile and accumulation of free fatty acids are highly significant in all forms of diabetes pointing to its possible link with inflammation and vascular complications (reviewed in<sup>[26]</sup>). Several studies showed the role of free fatty acids mainly palmitate in inducing pro-inflammatory response<sup>[27,28]</sup>. It should be noted that thorough understanding of the interaction between vascular and non-vascular cells is crucial for the management of retinal dysfunction. Müller cells are the principal glial cell found in the retina, which span the entire retinal layers and considered as resident innate immune cells (reviewed in<sup>[29]</sup>). Because of their unique morphology, Müller cells are considered a signaling hub that senses minute changes in retinal milieu, connecting retinal neuronal with retinal endothelial cells. In the current study we were interested in unraveling the mechanisms through which HFD leads to retinal inflammation. We also highlighted the critical role of Müller cells after the insult with the free fatty acid palmitate, which hasn't been reported so far. The main findings of this study are that (1) HFD or palmitate induced ER-stress dysregulates miR-17-5p in retina and Müller cells; (2) ER-stress triggers TXNIP expression in retina and Müller cells and (3) amplified TXNIP levels activate NLRP3, which contributes to inflammation.

Müller cells are considered major sources of inflammatory mediators, which become activated in response to various insults<sup>[19,30-32]</sup>. We and others have shown the increase of TXNIP expression in glial Müller cells due to chronic hyperglycemia<sup>[33-35]</sup> or HFD<sup>[4]</sup>. TXNIP is a physiological inhibitor of the thioredoxin system, which is one of the main antioxidant defense mechanisms in our body. TXNIP acts *via* binding to thioredoxin, making it unable to bind with other proteins (reviewed in<sup>[36]</sup>). In addition to the ability of TXNIP in inducing inflammatory cytokines *via* activating nuclear factor  $\kappa$ B, it can act as a direct activator of NOD-like receptor protein (NLRP3)<sup>[34,37]</sup>. NLRP3-inflammasome is a component of the innate immune system responsible for initiating obesity-induced inflammation<sup>[38]</sup>. TXNIP-NLRP3 interaction results in NLRP3 complex assembly and auto-activation of caspase-1, which eventually processes pro-IL1 $\beta$  into its mature form leading to inflammation<sup>[38,39]</sup>. Recent studies showed that HFD and palmitate trigger ER-stress in various organs and cell types<sup>[17,22-24]</sup>. However, the link between HFD/palmitate-induced ER-stress and TXNIP expression in Müller cells is yet to be determined. Here, we observed significant activation of the unfolded protein response ER-stress chaperons in retinas from 8-wk HFD mice (Figure 1). We also observed no difference in mRNA level of IRE1 $\alpha$  an ER-stress marker and a bifunctional kinase/Rnase in HFD. However, there was an increase in the splicing of XBP1; IRE1 $\alpha$  downstream target; evident by 3.5-fold increase in spliced XBP-1 in HFD compared to ND, which suggests IRE1 $\alpha$  activation. Interestingly, treatment of Müller cells with palmitate;

one of the most abundant saturated fatty acids in plasma that is significantly increased following HFD<sup>[25]</sup>; led to an increase in all ER-stress markers at the mRNA level including IRE1 $\alpha$  (Figure 1). Among UPR pathways, IRE1 $\alpha$  has been shown to degrade key cell regulators such as the neuronal cue, netrin in the retina<sup>[39,40]</sup> and miR-17-5p in pancreatic beta-cells<sup>[10,11]</sup>. MiR-17-5p is a small noncoding RNAs that binds predominantly to the 3'-UTR of TXNIP leading to down-regulation of its expression<sup>[10]</sup>. Indeed, HFD and palmitate resulted in a significant decrease in miR-17-5p in the total retina and Müller cells, respectively, an effect that coincided with TXNIP upregulation (Figure 2). These findings support the link between HFD, ER-stress and TXNIP upregulation in Müller cells.

Epidemiological studies showed a significant reduction in miR-17-5p in omental fat and blood from obese non-diabetic subjects compared to lean subjects<sup>[41,42]</sup>. In the current study, we showed that HFD or palmitate dysregulated miR-17-5p in retina and Müller cells (Figure 2). Interestingly, retinal miR-17-5p expression is not affected by hyperglycemia or diabetes compared to normal glycemic controls (data not shown). In agreement, Lerner *et al.*<sup>[10]</sup> reported similar insensitivity of miR-17-5p to high glucose treatment in pancreatic beta cells. These findings shed light on the selective sensitivity of miR-17-5p to degrade in response to HFD and palmitate. Taken together, our findings suggest that HFD-induced ER-stress uniquely triggers TXNIP expression *via* dysregulating miR-17-5p.

To dissect the role of ER-stress in regulating TXNIP expression, PBA was added to cultured rMC1 prior to palmitate treatment. PBA is an FDA approved drug for the clinical management of urea cycle disorder. PBA is a chemical chaperone that stabilizes protein conformation and in turns ER-folding (reviewed in<sup>[43,44]</sup>). Indeed, treating the cells with PBA a general ER-stress inhibitor showed a trend decrease in TXNIP expression. Similar findings were obtained by the use of a selective IRE1 $\alpha$  inhibitor (Figure 4). However, the observed reduction didn't reach significance, which could be due to the small sample size. We overcame this limitation, by treating mice kept on HFD with PBA for 2 wk. We showed that inhibiting ER-stress significantly blunted the increase in TXNIP observed in HFD group (Figure 4), without altering insulin resistance (Figure 3). Next step we tried to verify the role of TXNIP in inflammatory response in Müller cells. Building on our previous findings that silencing TXNIP reversed palmitate-induced IL1 $\beta$  release and eventually cell death in endothelial cells<sup>[4]</sup>, we isolated primary Müller cells from WT and TKO mice then exposed them to palmitate. We demonstrated that palmitate led to an increase in NLRP3 and IL1 $\beta$  expression in WT and has no effect on TKO (Figure 5), which indicates that TXNIP is responsible for inflammation in Müller cells. These results lend further support to prior findings that manifest the critical role of IL1 $\beta$  in mediating vascular injury in the pathogenesis of diabetic retinopathy. Kowluru *et al.*<sup>[45]</sup> showed that injecting IL-1 $\beta$

into the vitreous of normal rats increased cell apoptosis similar to what is observed in diabetes. Deletion of IL1 $\beta$  receptor prevented autocrine loop of inflammation<sup>[46]</sup> and protected retinas from diabetes-induced development of acellular capillaries<sup>[47]</sup>.

In summary, clinical and experimental studies have repeatedly reported the contribution of inflammation to the pathogenesis of diabetic retinopathy (reviewed in<sup>[48,49]</sup>). Similarly, suppression of inflammation has shown protective effects *via* decreasing leukostasis, blood-retinal barrier breakdown and the acellular capillaries formation<sup>[50]</sup>. Here, we provide preliminary evidence that exposure to high fat diet and palmitate trigger retinal ER-stress and glial TXNIP expression and render the retina vulnerable to inflammation. Early intervention of ER-stress or TXNIP presents potential therapeutic strategy in obesity-induced inflammation in diabetic retinopathy.

## COMMENTS

### Background

The authors have previously shown that high fat diet (HFD) induced retinal inflammation and vascular dysfunction. These results were associated with an increase in the thioredoxin interacting protein (TXNIP) at the mRNA and protein level. Here, they examined the mechanisms by which HFD triggers retinal TXNIP and regulates inflammation.

### Research frontiers

Currently, there is a great interest to understand how microRNA, the endogenous regulators of transcription can contribute to metabolic disorders. Here, they examined the impact of HFD or the free fatty acid palmitate on microRNA; miR-17-5p as it has been shown to regulate TXNIP mRNA expression. This study demonstrates the effect of HFD-induced obesity on degradation of miR-17-5p *via* activation of the ER-stress mediators including the inositol requiring enzyme 1 $\alpha$  (IRE1 $\alpha$ ). The authors also demonstrate that inhibiting ER-stress can restore miR-17-5p and TXNIP levels and hence inflammation back to comparable levels seen in normal controls.

### Innovations and breakthroughs

The results of their study delineate the contribution of Müller cells, main glia in the retina in palmitate-mediated retinal inflammation. They identify ER-stress as new therapeutic target that is involved in obesity-induced inflammation in pre-diabetic retinopathy.

### Applications

Their results suggest that inhibitors of ER-stress reversed the increase in TXNIP *in vivo* and in Müller cells, the main glia in the retina. The findings of their short-term study support the interventional use of the ER-Stress inhibitor PBA, FDA approved drug with high safety profile. This report should open the door for its future studies in diseases associated with TXNIP-NLRP3 inflammation.

### Terminology

MicroRNAs are small non-coding RNAs that contribute to the post-transcriptional regulation of various genes expressions. Inflammasome is a multiprotein oligomer responsible for the induction of inflammatory process. Unfolded protein response (UPR) is unfolded protein response, an adaptive mechanism to resolve and slow down protein processing. ER-stress is when the endoplasmic reticulum capacity to deal with UPR is overwhelmed then stress markers such as ATF6, PERK and IRE1 $\alpha$  are expressed.

### Peer-review

The paper is interesting.

## REFERENCES

- 1 **Ogden CL**, Carroll MD, Kit BK, Flegal KM. Prevalence of childhood and adult obesity in the United States, 2011-2012. *JAMA* 2014; **311**: 806-814 [PMID: 24570244 DOI: 10.1001/jama.2014.732]
- 2 **Wong TY**, Duncan BB, Golden SH, Klein R, Couper DJ, Klein BE, Hubbard LD, Sharrett AR, Schmidt MI. Associations between the metabolic syndrome and retinal microvascular signs: the Atherosclerosis Risk In Communities study. *Invest Ophthalmol Vis Sci* 2004; **45**: 2949-2954 [PMID: 15326106 DOI: 10.1167/iiov.04-0069]
- 3 **Nguyen TT**, Wong TY. Retinal vascular manifestations of metabolic disorders. *Trends Endocrinol Metab* 2006; **17**: 262-268 [PMID: 16890449 DOI: 10.1016/j.tem.2006.07.006]
- 4 **Mohamed IN**, Hafez SS, Fairaq A, Ergul A, Imig JD, El-Remessy AB. Thioredoxin-interacting protein is required for endothelial NLRP3 inflammasome activation and cell death in a rat model of high-fat diet. *Diabetologia* 2014; **57**: 413-423 [PMID: 24201577 DOI: 10.1007/s00125-013-3101-z]
- 5 **Feng H**, Gu J, Gou F, Huang W, Gao C, Chen G, Long Y, Zhou X, Yang M, Liu S, Lü S, Luo Q, Xu Y. High Glucose and Lipopolysaccharide Prime NLRP3 Inflammasome via ROS/TXNIP Pathway in Mesangial Cells. *J Diabetes Res* 2016; **2016**: 6973175 [PMID: 26881256 DOI: 10.1155/2016/6973175]
- 6 **Gao P**, He FF, Tang H, Lei CT, Chen S, Meng XF, Su H, Zhang C. NADPH oxidase-induced NALP3 inflammasome activation is driven by thioredoxin-interacting protein which contributes to podocyte injury in hyperglycemia. *J Diabetes Res* 2015; **2015**: 504761 [PMID: 25834832 DOI: 10.1155/2015/504761]
- 7 **Zhang X**, Zhang JH, Chen XY, Hu QH, Wang MX, Jin R, Zhang QY, Wang W, Wang R, Kang LL, Li JS, Li M, Pan Y, Huang JJ, Kong LD. Reactive oxygen species-induced TXNIP drives fructose-mediated hepatic inflammation and lipid accumulation through NLRP3 inflammasome activation. *Antioxid Redox Signal* 2015; **22**: 848-870 [PMID: 25602171 DOI: 10.1089/ars.2014.5868]
- 8 **Bushati N**, Cohen SM. microRNA functions. *Annu Rev Cell Dev Biol* 2007; **23**: 175-205 [PMID: 17506695 DOI: 10.1146/annurev.cellbio.23.090506.123406]
- 9 **McClelland AD**, Kantharidis P. microRNA in the development of diabetic complications. *Clin Sci (Lond)* 2014; **126**: 95-110 [PMID: 24059587 DOI: 10.1042/CS20130079]
- 10 **Lerner AG**, Upton JP, Praveen PV, Ghosh R, Nakagawa Y, Igbaria A, Shen S, Nguyen V, Backes BJ, Heiman M, Heintz N, Greengard P, Hui S, Tang Q, Trusina A, Oakes SA, Papa FR. IRE1 $\alpha$  induces thioredoxin-interacting protein to activate the NLRP3 inflammasome and promote programmed cell death under irremediable ER stress. *Cell Metab* 2012; **16**: 250-264 [PMID: 22883233 DOI: 10.1016/j.cmet.2012.07.007]
- 11 **Upton JP**, Wang L, Han D, Wang ES, Huskey NE, Lim L, Truitt M, McManus MT, Ruggero D, Goga A, Papa FR, Oakes SA. IRE1 $\alpha$  cleaves select microRNAs during ER stress to derepress translation of proapoptotic Caspase-2. *Science* 2012; **338**: 818-822 [PMID: 23042294 DOI: 10.1126/science.1226191]
- 12 **Chistiakov DA**, Sobenin IA, Orekhov AN, Bobryshev YV. Role of endoplasmic reticulum stress in atherosclerosis and diabetic macrovascular complications. *Biomed Res Int* 2014; **2014**: 610140 [PMID: 25061609 DOI: 10.1155/2014/610140]
- 13 **Dunys J**, Duplan E, Checler F. The transcription factor X-box binding protein-1 in neurodegenerative diseases. *Mol Neurodegener* 2014; **9**: 35 [PMID: 25216759 DOI: 10.1186/1750-1326-9-35]
- 14 **Ma JH**, Wang JJ, Zhang SX. The unfolded protein response and diabetic retinopathy. *J Diabetes Res* 2014; **2014**: 160140 [PMID: 25530974 DOI: 10.1155/2014/160140]
- 15 **Boden G**. Obesity, insulin resistance and free fatty acids. *Curr Opin Endocrinol Diabetes Obes* 2011; **18**: 139-143 [PMID: 21297467 DOI: 10.1097/MED.0b013e3283444b09]
- 16 **Lu Y**, Qian L, Zhang Q, Chen B, Gui L, Huang D, Chen G, Chen L. Palmitate induces apoptosis in mouse aortic endothelial cells and endothelial dysfunction in mice fed high-calorie and high-



- cholesterol diets. *Life Sci* 2013; **92**: 1165-1173 [PMID: 23680379 DOI: 10.1016/j.lfs.2013.05.002]
- 17 **Dai F**, Jiang T, Bao YY, Chen GJ, Chen L, Zhang Q, Lu YX. Fenofibrate improves high-fat diet-induced and palmitate-induced endoplasmic reticulum stress and inflammation in skeletal muscle. *Life Sci* 2016; **157**: 158-167 [PMID: 27297630 DOI: 10.1016/j.lfs.2016.06.008]
  - 18 **Sarthy VP**, Brodjan SJ, Dutt K, Kennedy BN, French RP, Crabb JW. Establishment and characterization of a retinal Müller cell line. *Invest Ophthalmol Vis Sci* 1998; **39**: 212-216 [PMID: 9430566]
  - 19 **Mysona BA**, Al-Gayyar MM, Matragoon S, Abdelsaid MA, El-Azab MF, Saragovi HU, El-Remessy AB. Modulation of p75(NTR) prevents diabetes- and proNGF-induced retinal inflammation and blood-retina barrier breakdown in mice and rats. *Diabetologia* 2013; **56**: 2329-2339 [PMID: 23918145 DOI: 10.1007/s00125-013-2998-6]
  - 20 **Chen C**, Ridzon DA, Broomer AJ, Zhou Z, Lee DH, Nguyen JT, Barbisin M, Xu NL, Mahuvakar VR, Andersen MR, Lao KQ, Livak KJ, Guegler KJ. Real-time quantification of microRNAs by stem-loop RT-PCR. *Nucleic Acids Res* 2005; **33**: e179 [PMID: 16314309 DOI: 10.1093/nar/gni178]
  - 21 **Ali TK**, Matragoon S, Pillai BA, Liou GI, El-Remessy AB. Peroxynitrite mediates retinal neurodegeneration by inhibiting nerve growth factor survival signaling in experimental and human diabetes. *Diabetes* 2008; **57**: 889-898 [PMID: 18285558 DOI: 10.2337/db07-1669]
  - 22 **Gulhane M**, Murray L, Lourie R, Tong H, Sheng YH, Wang R, Kang A, Schreiber V, Wong KY, Magor G, Denman S, Begun J, Florin TH, Perkins A, Cuív PÓ, McGuckin MA, Hasnain SZ. High Fat Diets Induce Colonic Epithelial Cell Stress and Inflammation that is Reversed by IL-22. *Sci Rep* 2016; **6**: 28990 [PMID: 27350069 DOI: 10.1038/srep28990]
  - 23 **Li SJ**, Liu CH, Chu HP, Mersmann HJ, Ding ST, Chu CH, Wang CY, Chen CY. The high-fat diet induces myocardial fibrosis in the metabolically healthy obese minipigs-The role of ER stress and oxidative stress. *Clin Nutr* 2016 Jun 16; Epub ahead of print [PMID: 27342749 DOI: 10.1016/j.clnu.2016.06.002]
  - 24 **Gwiazda KS**, Yang TL, Lin Y, Johnson JD. Effects of palmitate on ER and cytosolic Ca<sup>2+</sup> homeostasis in beta-cells. *Am J Physiol Endocrinol Metab* 2009; **296**: E690-E701 [PMID: 19141690 DOI: 10.1152/ajpendo.90525.2008]
  - 25 **Paik JS**, Cho WK, Oh EH, Lee SB, Yang SW. Palmitate induced secretion of IL-6 and MCP-1 in orbital fibroblasts derived from patients with thyroid-associated ophthalmopathy. *Mol Vis* 2012; **18**: 1467-1477 [PMID: 22736938]
  - 26 **Jaiswal M**, Schinske A, Pop-Busui R. Lipids and lipid management in diabetes. *Best Pract Res Clin Endocrinol Metab* 2014; **28**: 325-338 [PMID: 24840262 DOI: 10.1016/j.beem.2013.12.001]
  - 27 **Staiger H**, Staiger K, Stefan N, Wahl HG, Machicao F, Kellerer M, Häring HU. Palmitate-induced interleukin-6 expression in human coronary artery endothelial cells. *Diabetes* 2004; **53**: 3209-3216 [PMID: 15561952]
  - 28 **Krogmann A**, Staiger K, Haas C, Gommer N, Peter A, Heni M, Machicao F, Häring HU, Staiger H. Inflammatory response of human coronary artery endothelial cells to saturated long-chain fatty acids. *Microvasc Res* 2011; **81**: 52-59 [PMID: 21112343 DOI: 10.1016/j.mvr.2010.11.008]
  - 29 **Goldman D**. Müller glial cell reprogramming and retina regeneration. *Nat Rev Neurosci* 2014; **15**: 431-442 [PMID: 24894585 DOI: 10.1038/nrn3723]
  - 30 **Ali TK**, Al-Gayyar MM, Matragoon S, Pillai BA, Abdelsaid MA, Nussbaum JJ, El-Remessy AB. Diabetes-induced peroxynitrite impairs the balance of pro-nerve growth factor and nerve growth factor, and causes neurovascular injury. *Diabetologia* 2011; **54**: 657-668 [PMID: 20957344 DOI: 10.1007/s00125-010-1935-1]
  - 31 **Liu X**, Ye F, Xiong H, Hu DN, Limb GA, Xie T, Peng L, Zhang P, Wei Y, Zhang W, Wang J, Wu H, Lee P, Song E, Zhang DY. IL-1 $\beta$  induces IL-6 production in retinal Müller cells predominantly through the activation of p38 MAPK/NF- $\kappa$ B signaling pathway. *Exp Cell Res* 2015; **331**: 223-231 [PMID: 25239226 DOI: 10.1016/j.yexcr.2014.08.040]
  - 32 **Mizutani M**, Gerhardinger C, Lorenzi M. Müller cell changes in human diabetic retinopathy. *Diabetes* 1998; **47**: 445-449 [PMID: 9519752]
  - 33 **Devi TS**, Hosoya K, Terasaki T, Singh LP. Critical role of TXNIP in oxidative stress, DNA damage and retinal pericyte apoptosis under high glucose: implications for diabetic retinopathy. *Exp Cell Res* 2013; **319**: 1001-1012 [PMID: 23353834 DOI: 10.1016/j.yexcr.2013.01.012]
  - 34 **Perrone L**, Devi TS, Hosoya KI, Terasaki T, Singh LP. Inhibition of TXNIP expression in vivo blocks early pathologies of diabetic retinopathy. *Cell Death Dis* 2010; **1**: e65 [PMID: 21364670 DOI: 10.1038/cddis.2010.42]
  - 35 **Devi TS**, Lee I, Hüttemann M, Kumar A, Nantwi KD, Singh LP. TXNIP links innate host defense mechanisms to oxidative stress and inflammation in retinal Muller glia under chronic hyperglycemia: implications for diabetic retinopathy. *Exp Diabetes Res* 2012; **2012**: 438238 [PMID: 22474421 DOI: 10.1155/2012/438238]
  - 36 **Chong CR**, Chan WP, Nguyen TH, Liu S, Procter NE, Ngo DT, Sverdllov AL, Chirkov YY, Horowitz JD. Thioredoxin-interacting protein: pathophysiology and emerging pharmacotherapeutics in cardiovascular disease and diabetes. *Cardiovasc Drugs Ther* 2014; **28**: 347-360 [PMID: 25088927 DOI: 10.1007/s10557-014-6538-5]
  - 37 **Al-Gayyar MM**, Abdelsaid MA, Matragoon S, Pillai BA, El-Remessy AB. Thioredoxin interacting protein is a novel mediator of retinal inflammation and neurotoxicity. *Br J Pharmacol* 2011; **164**: 170-180 [PMID: 21434880 DOI: 10.1111/j.1476-5381.2011.01336.x]
  - 38 **Vandanmagsar B**, Youm YH, Ravussin A, Galgani JE, Stadler K, Mynatt RL, Ravussin E, Stephens JM, Dixit VD. The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nat Med* 2011; **17**: 179-188 [PMID: 21217695 DOI: 10.1038/nm.2279]
  - 39 **Martinon F**, Burns K, Tschopp J. The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL-beta. *Mol Cell* 2002; **10**: 417-426 [PMID: 12191486]
  - 40 **Binet F**, Mawambo G, Sitaras N, Tetreault N, Lapalme E, Favret S, Cerani A, Leboeuf D, Tremblay S, Rezende F, Juan AM, Stahl A, Joyal JS, Milot E, Kaufman RJ, Guimond M, Kennedy TE, Sapieha P. Neuronal ER stress impedes myeloid-cell-induced vascular regeneration through IRE1 $\alpha$  degradation of netrin-1. *Cell Metab* 2013; **17**: 353-371 [PMID: 23473031 DOI: 10.1016/j.cmet.2013.02.003]
  - 41 **Klötting N**, Berthold S, Kovacs P, Schön MR, Fasshauer M, Ruschke K, Stumvoll M, Blüher M. MicroRNA expression in human omental and subcutaneous adipose tissue. *PLoS One* 2009; **4**: e4699 [PMID: 19259271 DOI: 10.1371/journal.pone.0004699]
  - 42 **Heneghan HM**, Miller N, McAnena OJ, O'Brien T, Kerin MJ. Differential miRNA expression in omental adipose tissue and in the circulation of obese patients identifies novel metabolic biomarkers. *J Clin Endocrinol Metab* 2011; **96**: E846-E850 [PMID: 21367929 DOI: 10.1210/jc.2010-2701]
  - 43 **Welch WJ**, Brown CR. Influence of molecular and chemical chaperones on protein folding. *Cell Stress Chaperones* 1996; **1**: 109-115 [PMID: 9222596]
  - 44 **Kolb PS**, Ayaub EA, Zhou W, Yum V, Dickhout JG, Ask K. The therapeutic effects of 4-phenylbutyric acid in maintaining proteostasis. *Int J Biochem Cell Biol* 2015; **61**: 45-52 [PMID: 25660369 DOI: 10.1016/j.biocel.2015.01.015]
  - 45 **Kowluru RA**, Odenbach S. Role of interleukin-1beta in the development of retinopathy in rats: effect of antioxidants. *Invest Ophthalmol Vis Sci* 2004; **45**: 4161-4166 [PMID: 15505070 DOI: 10.1167/iops.04-0633]
  - 46 **Yego EC**, Vincent JA, Sarthy V, Busik JV, Mohr S. Differential regulation of high glucose-induced glyceraldehyde-3-phosphate dehydrogenase nuclear accumulation in Müller cells by IL-1beta and IL-6. *Invest Ophthalmol Vis Sci* 2009; **50**: 1920-1928 [PMID: 19060282 DOI: 10.1167/iops.08-2082]
  - 47 **Vincent JA**, Mohr S. Inhibition of caspase-1/interleukin-1beta signaling prevents degeneration of retinal capillaries in diabetes and galactosemia. *Diabetes* 2007; **56**: 224-230 [PMID: 17192486 DOI: 10.2337/db06-0427]

- 48 **Adamis AP**. Is diabetic retinopathy an inflammatory disease? *Br J Ophthalmol* 2002; **86**: 363-365 [PMID: 11914197]
- 49 **Tang J**, Kern TS. Inflammation in diabetic retinopathy. *Prog Retin Eye Res* 2011; **30**: 343-358 [PMID: 21635964 DOI: 10.1016/j.preteyeres.2011.05.002]
- 50 **Joussen AM**, Poulaki V, Le ML, Koizumi K, Esser C, Janicki H, Schraermeyer U, Kociok N, Fauser S, Kirchhof B, Kern TS, Adamis AP. A central role for inflammation in the pathogenesis of diabetic retinopathy. *FASEB J* 2004; **18**: 1450-1452 [PMID: 15231732 DOI: 10.1096/fj.03-1476fje]

**P- Reviewer:** Liou GI, Mungrue IN, Sychrova H    **S- Editor:** Qiu S  
**L- Editor:** A    **E- Editor:** Wu HL



Case Control Study

# Association of *NFKB1* gene polymorphism (rs28362491) with levels of inflammatory biomarkers and susceptibility to diabetic nephropathy in Asian Indians

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**Author contributions:** Gautam A and Gupta S contributed equally to this article; Gautam A, Gupta S, Sharma M and Singh K performed sample collection, majority of experiments, analyzed data and wrote manuscript; Mehndiratta M, Kalra OP, Agarwal S and Gambhir JK contributed to conceptual and study design, data analysis and interpretation, as well as manuscript writing.

**Institutional review board statement:** The protocol of the present study was reviewed and approved by Institutional Ethics Committee for Human Research, Delhi University, and UCMS and GTB Hospital, Delhi.

**Informed consent statement:** Informed consent was obtained from the all recruited subjects. They were briefed on the purpose of the study and its implication prior to donating peripheral blood.

**Conflict-of-interest statement:** No conflict of interest exists for this study.

**Data sharing statement:** No data sharing as this manuscript and the data were not published elsewhere.

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Manuscript source: Invited manuscript

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Telephone: +91-98-11641277

Received: September 12, 2016

Peer-review started: September 19, 2016

First decision: October 21, 2016

Revised: December 6, 2016

Accepted: December 27, 2016

Article in press: December 28, 2016

Published online: February 15, 2017

## Abstract

### AIM

To investigate the association of *NFKB1* gene -94 ATG insertion/deletion (rs28362491) polymorphism with inflammatory markers and risk of diabetic nephropathy in Asian Indians.

### METHODS

A total of 300 subjects were recruited (100 each), normoglycemic, (NG); type 2 diabetes mellitus (T2DM) without any complications (DM) and T2DM with diabetic nephropathy [DM-chronic renal disease (CRD)]. Analysis was carried out by polymerase chain reaction-restriction fragment length polymorphism and ELISA. Pearson's correlation, analysis of variance and logistic regression were

used for statistical analysis.

## RESULTS

The allelic frequencies of -94 ATTG insertion/deletion were 0.655/0.345 (NG), 0.62/0.38 (DM) and 0.775/0.225 (DM-CRD). The -94 ATTG ins allele was associated with significantly increased levels of urinary monocyte chemoattractant protein-1 (uMCP-1); uMCP-1 ( $P = 0.026$ ) and plasma tumor necrosis factor-alpha (TNF- $\alpha$ ); TNF- $\alpha$  ( $P = 0.030$ ) and almost doubled the risk of diabetic nephropathy (OR = 1.91, 95%CI: 1.080-3.386,  $P = 0.025$ ).

## CONCLUSION

-94 ATTG ins/ins polymorphism might be associated with increased risk of developing nephropathy in Asian Indian subjects with diabetes mellitus.

**Key words:** Diabetic nephropathy; Inflammation; *NFKB1* -94 ATTG ins/del polymorphism; Urinary monocyte chemoattractant protein-1; Tumor necrosis factor-alpha

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**Core tip:** Type 2 diabetes mellitus (T2DM) is considered as long standing inflammatory disease. Diabetic nephropathy (DN) is the most common micro-vascular complication of T2DM. Pro-inflammatory cytokines like Monocyte chemoattractant protein-1 (MCP-1) and tumor necrosis factor-alpha (TNF- $\alpha$ ) plays a crucial role in the pathogenesis of DN. Therefore we investigated -94 ins/del ATTG polymorphism in *NFKB1* gene and its association with the risk of DN in Asian Indians. -94 ins/del ATTG single nucleotide polymorphism was found to increase the urinary MCP-1 and plasma TNF- $\alpha$  levels. Our findings open a new area of research to explore that -94 ins/del ATTG may be considered as genetic markers for early detection of diabetic patients who are at greater risk of development of nephropathy.

Gautam A, Gupta S, Mehndiratta M, Sharma M, Singh K, Kalra OP, Agarwal S, Gambhir JK. Association of *NFKB1* gene polymorphism (rs28362491) with levels of inflammatory biomarkers and susceptibility to diabetic nephropathy in Asian Indians. *World J Diabetes* 2017; 8(2): 66-73 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v8/i2/66.htm> DOI: <http://dx.doi.org/10.4239/wjd.v8.i2.66>

## INTRODUCTION

Chronic renal disease (CRD) is an intricate pathological process, often leading to end stage renal disease. The causes of CRD are quite multi-factorial ranging from infections to heredity, but type 2 diabetes mellitus (T2DM) is the major culprit amongst them<sup>[1]</sup>. In spite of the improvement in our knowledge about the etiopathogenesis of diabetic nephropathy (DN), the

intricate mechanisms leading to the development of renal injury from chronic hyperglycemia are not yet fully understood. DN has been considered a micro-vascular complication of hyperglycemia, but various clinical and experimental studies have observed that there is a close link between hyperglycemia, inflammation and oxidative stress (OS)<sup>[2]</sup>. OS may also be involved in promoting a low grade systemic inflammation in patients with T2DM and vice versa<sup>[3]</sup>. Nuclear factor-kappa B (NF- $\kappa$ B) activation through hyperglycemia induced OS may lead to increased concentration of inflammatory cytokines<sup>[4]</sup>.

NF- $\kappa$ B was identified as a transcription factor which controls the expression of numerous genes affecting immune response, inflammation, cell-growth control, apoptosis and therefore, is an emerging candidate for studies on the pathogenesis of inflammatory diseases including DN. There are five members of the NF- $\kappa$ B family in mammals: NF- $\kappa$ B1: p105/p50, NF- $\kappa$ B2: p52/p100, RelA: p65, RelB, and c-Rel. The chief form of NF- $\kappa$ B is a hetero-dimer of the p50 and p65/RelA subunits, encoded by the *NFKB1* and *RelA* gene. Normally, inactive NF- $\kappa$ B is found in the cytoplasm bound to I $\kappa$ Bs, which are specific inhibitor proteins in cytoplasm. Cell when exposed to a variety of proinflammatory stimuli leads to the quick phosphorylation followed by ubiquitinylation, and finally proteolytic breakdown of I- $\kappa$ B. This causes transfer of NF- $\kappa$ B in nucleus and thus leading to increased transcription of gene<sup>[5]</sup>. NF- $\kappa$ B transcriptionally regulates many downstream proinflammatory genes, mainly including monocyte chemoattractant protein-1 (*MCP-1*), tumor necrosis factor-alpha (*TNF- $\alpha$* )<sup>[6]</sup>.

MCP-1 is an important proinflammatory chemokine which affects the recruitment and function of monocyte<sup>[7]</sup>. MCP-1 is synthesized in response to a various proinflammatory stimuli by kidney cell<sup>[8]</sup>. A study done by Wada *et al*<sup>[9]</sup> in 2000 has shown that expression of MCP-1 increases in inflammation induced kidney diseases including DN. Urinary MCP-1 (uMCP-1) is a potential biomarker for renal damage<sup>[10]</sup>. Hyperglycemia induced secretion of abundant MCP-1 from renal parenchymal cells, attract monocytes into the kidney stimulating myofibroblast-like properties in mesangial cells. Kidney macrophages when exposed to MCP-1 in diabetic milieu promotes activation of macrophage. Thus, leading to release of reactive oxygen species (ROS), various pro-inflammatory cytokines and profibrotic growth factors<sup>[11,12]</sup>. Thus, resulting in exaggerated inflammation that leads to renal injury through proliferation of myofibroblast, augmented production of extracellular matrix by mesangial cells and fibroblasts.

TNF- $\alpha$  is a well known proinflammatory cytokine associated with systemic inflammation<sup>[13,14]</sup>. It is produced predominantly by macrophages and monocytes<sup>[13,14]</sup>. TNF- $\alpha$  acts *via* NF- $\kappa$ B signaling and mediates the transcription of various cytokines performing roles in cell survival, proliferation, inflammatory responses, cell adhesion and inflammation<sup>[15]</sup>. A study has shown that



there is upregulation of TNF- $\alpha$  expression in glomeruli of diabetic rats<sup>[16]</sup>. TNF- $\alpha$  is well acknowledged to cause damage to renal cells by enhancing renal hypertrophy, hemodynamic imbalance, albumin permeability<sup>[17]</sup>. The harmful effects of these responses lead to the development of renal disease in patients with T2DM, hence resulting in the progression of renal failure.

In addition to poor glycemic control, OS and inflammation; genetic factors seem to be main determinants of DN in terms of both occurrence and severity<sup>[18]</sup>; however the genetic mechanism causing DN is still unexplored. In our knowledge, there is no study available regarding the polymorphisms of *NFKB1* and their correlation with levels of uMCP-1 and plasma TNF- $\alpha$ . We have reported<sup>[19]</sup> increased uMCP-1, plasma TNF- $\alpha$  levels in subjects with DN when compared to subjects with T2DM without nephropathy and observed a positive correlation between uMCP-1 and plasma TNF- $\alpha$ <sup>[20]</sup>. We have also highlighted that DN is associated with *TNFA* gene single nucleotide polymorphism (SNP)<sup>[20]</sup>. In recent times, a new functional *NFKB1* promoter SNP consisting of a insertion/deletion (-94ins/del ATTG) (rs28362491) has been identified which can elicit a regulatory effect on the *NFKB1* gene<sup>[21]</sup>. Since above mentioned polymorphism has been associated with various inflammatory diseases, autoimmune diseases and cancers<sup>[22]</sup>, therefore, it is worthwhile to further investigate the association of -94 ins/del ATTG *NFKB1* gene SNP with levels of uMCP-1, plasma TNF- $\alpha$  and nephropathy risk in subjects with T2DM.

## MATERIALS AND METHODS

### Study design

The present study comprises of total 300 subjects visiting Nephrology Outpatient Clinic and Medicine OPD at University College of Medical Sciences and Guru Teg Bahadur Hospital, Delhi. Subjects were divided into three groups of 100 each namely; Group 1: Normoglycemic (NG), Group 2: Subjects with T2DM for  $\geq 10$  years without nephropathy (DM), Group 3: Subjects with T2DM for  $\geq 5$  years with nephropathy (DM-CRD). T2DM was diagnosed according to revised ADA criteria<sup>[23]</sup>. Detailed clinical history and physical examination were recorded. Blood pressure (BP) of subjects was estimated using sphygmomanometer in the sitting position after a resting period of 10 min. The estimated glomerular filtration rate (eGFR) was measured by Modification of Diet in Renal Disease Abbreviated Equation (MDRD)<sup>[24]</sup>.

The presence of micro-albuminuria in T2DM subjects was detected by Urine Test 11 MAU dipstick (Piramal Diagnostic, sensitivity: 10-15 mg/dL), and all participants having proteinuria and micro-albuminuria were clubbed in Group 3. All participants with nephropathy were in pre-dialysis stage. Normoglycemic (Group 1) subjects were recruited from employees of UCMS and GTB Hospital with the following criteria: (1) they did not have of diabetes mellitus (fasting plasma glucose < 100 mg% or postprandial glucose < 140 mg% or

HbA1c < 5.7%) according to ADA criteria; (2) there was no presence of diabetes in their first or second degree relatives; and (3) they had normal BP, with systolic and diastolic BP not > 120 mmHg and 80 mmHg<sup>[25]</sup>.

To circumvent any possible confounding factors, patients having renal disorders (hypertensive nephropathy, chronic glomerular nephritis, chronic interstitial disease, ischemic nephropathy, obstructive nephropathy), acute and chronic infections, congestive heart failure, malignancy and liver disorder were not included into the study. All subjects in Group 3 had retinopathy; but participants with macro-vascular complications like coronary artery disease and stroke were not included into the study. Patients taking renin-angiotension aldosterone system inhibitors, aspirin and vitamin D analogues were advised to discontinue these drugs for a period of a week before inclusion in the study since they have been found to influence the synthesis of uMCP-1 and TNF- $\alpha$ . However, patients were prescribed beta-blockers to control BP in that duration of one week. The Institutional Ethics Committee for Human Research approved the protocol of this study (approval number-UCMS/IEC-HR/2010/10). Prior to the inclusion into the present study, informed written consent was taken from all participants.

### Biochemical parameters

Under aseptic conditions fasting venous blood samples were withdrawn and collected into EDTA and fluoride vials. For glycosylated hemoglobin (HbA1c) 200  $\mu$ L whole blood was preserved at 4 °C-8 °C and processed within one week of collection. Blood samples collected in EDTA vial was subjected to centrifugation at 3000 rpm for 10 min in order to separate the plasma. Early morning first mid-stream urine sample was collected and stored in aliquots at -20 °C for estimation of MCP-1, albumin and creatinine.

Routine investigations such as fasting and post-prandial plasma glucose, urea, creatinine and uric acid were carried out using commercially available kits on autoanalyser (Olympus AU-400). HbA1c was estimated by ion-exchange resin chromatography using commercially available kits (Fortress, United Kingdom). Urinary protein excretion was expressed as albumin to creatinine ratio.

### Markers of inflammation

uMCP-1 (Weldon, California; sensitivity less than 7.8 pg/mL) and plasma TNF- $\alpha$  (Diacalone, France; sensitivity less than 8 pg/mL) were estimated by commercially available ELISA kit.

### DNA extraction and polymorphism genotyping

Cellular DNA of every individual was extracted from 200  $\mu$ L EDTA-anticoagulated peripheral blood sample by means of DNA isolation kit (Zymo research, United States). The polymerase chain reaction was carried out in Thermocycler (Eppendorf Mastercycler Gradient-5331). In brief, 0.1  $\mu$ g of DNA was amplified in a reaction mixture

**Table 1** The baseline demographic and biochemical parameters in various study groups

Variables	NG (n = 100)	DM (n = 100)	DM-CRD (n = 100)
Age (yr)	46.0 ± 4.0	56.40 ± 3.5	55.7 ± 4.2
Sex ratio (male/female)	52/48	54/46	52/48
Duration of DM (yr)	-	12.7 ± 1.5	8.1 ± 2.3 <sup>d</sup>
BMI (kg/m <sup>2</sup> )	20.1 ± 1.7	21.1 ± 2.1	21.6 ± 3.4
SBP (mmHg)	118.1 ± 0.5	138.0 ± 2.1 <sup>b</sup>	137.7 ± 2.8 <sup>b</sup>
DBP (mmHg)	75.2 ± 1.0	81.6 ± 1.9 <sup>b</sup>	82.8 ± 0.0 <sup>b</sup>
Fasting glucose (mg/dL)	82.4 ± 3.1	153.5 ± 3.5 <sup>b</sup>	184.3 ± 9.2 <sup>b,d</sup>
Postprandial glucose (mg/dL)	118.2 ± 2.4	201.7 ± 10.1 <sup>b</sup>	261.1 ± 12.2 <sup>b,d</sup>
HbA1c (%)	5.11 ± 0.46	7.10 ± 0.25 <sup>b</sup>	9.16 ± 0.16 <sup>b,d</sup>
Urea (mg/dL)	31.5 ± 5.5	30.7 ± 5.8	93.2 ± 4.8 <sup>b,d</sup>
Creatinine (mg/dL)	0.83 ± 0.23	0.90 ± 0.20	3.7 ± 1.5 <sup>b,d</sup>
Uric acid (mg/dL)	4.2 ± 0.8	4.9 ± 0.6	9.1 ± 0.8 <sup>b,d</sup>
eGFR (mL/min per 1.73 m <sup>2</sup> )	99.1 ± 0.7	96.4 ± 0.6	51.2 ± 0.9 <sup>d</sup>
Urinary albumin/creatinine	-	-	0.42 ± 0.35

<sup>b</sup>Significantly different from Normoglycemic at  $P < 0.001$ ; <sup>d</sup>Significantly different from diabetic patients without nephropathy at  $P < 0.001$ . Data are expressed as mean ± SD. NG: Normoglycemic; DM: Diabetes mellitus without nephropathy; DM-CRD: Diabetic nephropathy; BMI: Body mass index; SBP and DBP: Systolic and diastolic blood pressure; eGFR: Estimated glomerular filtration rate.

of 20 µL containing 0.5 µmol/L each of the following primer pairs (Forward 5'-TGGGCACAAGTCGTTTATGA-3' and Reverse 5'-CTGGAGCCGGTAGGGAAG-3'). The reaction mixture also contained 0.5 mmol/L (dNTP mix), 2 µL (10 × PCR buffer) and 2.0 units Taq DNA polymerase, 2 mmol/L MgCl<sub>2</sub>. The PCR protocol consist an initial temperature of 94 °C (5 min) followed by 35 cycles of amplification (30 s at 94 °C, 45 s at 59 °C, and extension for 1 min at 72 °C). Final extension step was carried out for 2-min at 72 °C<sup>[22]</sup>.

For the study of the -94 insertion/deletion ATTG SNP in *NFKB1*, PCR product (281/285 bp) was subjected to fast digestion with restriction enzyme *Pf*MI. PCR products was treated with enzyme *Pf*MI in at 37 °C for 1 h and inactivated at 65 °C for 20 min. The insertion allele (ins) was cut down into two fragments of 45 bp and 240 bp by *Pf*MI restriction enzyme. But, there was no cleavage at the deletion allele (del) that has only one ATTG at its promoter<sup>[22]</sup>. The bands of digested products were visualized in 2% agarose gel electrophoresis stained with ethidium bromide.

### Statistical analysis

Demographic profiles and routine investigation was compared by  $\chi^2$  and Student's *t* test and one-way ANOVA was used. To associate all the study groups with genotype two-way ANOVA followed by *post-hoc* Tukey's test was used. For association of genotypes with uMCP-1 and plasma TNF- $\alpha$  levels, analysis of variance was used. Logistic regressions was used to evaluate the risk of development of DN at the single SNP level. Power of sample size keeping 5% significance level and 80% power was calculated by genetic power calculator. A *P* value < 0.05 was considered statistically significant

**Table 2** The genotype and allele frequencies of *NFKB1* gene for -94 insertion/deletion ATTG polymorphism in different study groups

	NG (n = 100) n (%)	DM (n = 100) n (%)	DM-CRD (n = 100) n (%)
ins/ins	41 (41)	38 (38)	61 (61)
ins/del	49 (49)	48 (48)	33 (33)
del/del	10 (10)	14 (14)	06 <sup>b</sup> (06)
ins allele	131 (65.5)	124 (62)	155 (77.5)
del allele	69 (34.5)	76 (38)	45 (22.5)

<sup>b</sup>Significantly different from diabetic patients without nephropathy at  $P < 0.001$ . NG: Normoglycemic; DM: Diabetes mellitus without nephropathy; DM-CRD: Diabetic nephropathy.

(two-tailed). All statistical tests were performed using SPSS version 20.

## RESULTS

### Characteristics of the study population

Biochemical and demographic parameters of the various study groups are shown in Table 1. There was no difference in sex distribution and BMI within all the three study groups. The subjects of Group 2 (DM) and Group 3 (DM-CRD) were older than Group 1 (NG) subjects; however the period of diabetes was more in Group 2 (DM) than Group 3 (DM-CRD) which was as per our selection criteria. Incidence of hypertension was significantly higher in Group 2 (DM) and Group 3 (DM-CRD) participants as suggested by raised SBP and DBP ( $P < 0.001$ ) when compared to NG. Poor glucose control was observed in DM-CRD as compared to DM as suggested by significantly higher ( $P < 0.001$ ) fasting, postprandial plasma glucose and HbA1c. Renal function tests suggested that blood urea, plasma creatinine, and uric acid were significantly higher ( $P < 0.001$ ) and eGFR was decreased ( $P < 0.001$ ) in Group 3 (DM-CRD) as compared to Group 2 (DM).

### Distribution of ins/del in study population

The allele frequencies and genotype of the *NFKB1* gene for -94 insertion/deletion ATTG SNP in various study groups are shown in Table 2. The distribution percentage of ins/ins, ins/del, del/del genotypes in Group 1 (NG), Group 2 (DM) and Group 3 (DM-CRD) (expressed in percentage) were 41%, 49% and 10%; 38%, 48% and 14%; and 61%, 33% and 6% respectively. The frequency of del/del genotype was significantly lower ( $P < 0.001$ ) in Group 3 (DM-CRD) as compared to Group 2 (DM). However, allele frequencies of -94 insertion/deletion ATTG were 65.5%/34.5% in Group 1 (NG), 62%/38% in Group 2 (DM) and 77.5%/22.5% in Group 3 (DM-CRD).

### Relationship between the -94 ins/del AGGT SNP with inflammatory markers and disease risk

Correlation of -94 ins/del AGGT SNP with levels of

**Table 3** Interaction analysis of -94 ins/del ATTG polymorphism with inflammatory markers

Inflammatory marker	Groups	NG (n = 100)	DM (n = 100)	DM-CRD (n = 100)	P value
uMCP-1 (pg/mg creatinine)	Total	130.00 ± 42.22	271.00 ± 120.01	5632.70 ± 1007.20 <sup>ab</sup>	
	del/del	85.1 ± 9.2	200.6 ± 66.5	4609.9 ± 900.6	P = 0.026
	ins/del	110.9 ± 15.6	278.9 ± 105.9	5879.9 ± 1016.3	
	ins/ins	166.8 ± 26.8	302.2 ± 100.1	6405.1 ± 1550.6	
Plasma TNF-α (pg/mL)	Total	15.55 ± 2.22	16.51 ± 3.75	21.38 ± 3.67 <sup>ab</sup>	
	del/del	8.27 ± 1.06	10.21 ± 1.32	17.31 ± 1.17	P = 0.030
	ins/del	11.55 ± 0.05	14.05 ± 0.18	19.31 ± 0.44	
	ins/ins	15.08 ± 1.15	16.36 ± 1.20	23.12 ± 0.70	

<sup>a</sup>Significantly different from Normoglycemic at  $P < 0.001$ ; <sup>b</sup>Significantly different from diabetic patients without nephropathy at  $P < 0.001$ . uMCP-1 levels, plasma TNF-α levels are expressed as mean + SD. NG: Normoglycemic; DM: Diabetes mellitus without nephropathy; DM-CRD: Diabetic nephropathy.

uMCP-1 and plasma TNF-α have been studied and the results are shown in Table 3. The -94 ins allele were associated with increased levels of uMCP-1 ( $P = 0.026$ ) and plasma TNF-α ( $P = 0.030$ ) in the disease study groups, *i.e.*, Group 2 (DM), Group 3 (DM-CRD).

The associations at the level of genotype is shown in Table 4. Highly significant association was observed for -94 ins/del AGGT polymorphism in subjects with Group 3 (DM-CRD) in comparison to Group 1 (NG);  $P = 0.022$ . In our present study, -94 ins SNP was found to increase risk for the development of DN by 1.91-fold in subjects with diabetes (OR = 1.91, 95%CI: 1.080-3.386,  $P = 0.025$ ).

## DISCUSSION

Polymorphism in the *NFKB1* promoter region at position -94 ins/del AGGT has been correlated with many long standing inflammatory diseases like autoimmune diseases such as rheumatoid arthritis, asthma, AIDS, cancers and various diabetic complications<sup>[26,27]</sup>. Our study is the first to report the association of above mentioned polymorphism with DN in North Indian population. In the current study, we observed that the frequency distribution of ins/del is maximum in NG and DM subjects followed by ins/ins, with least distribution of del/del in the same. However the trend was different in DM-CRD subjects with respect to ins/del genotype which was less as compared to ins/ins this group. The frequency of different genotypes observed in the present study were in accordance with studies on *NFKB1* polymorphism in healthy volunteer in different ethnic population like Turkish<sup>[22]</sup>, Caucasians<sup>[28]</sup>, English<sup>[29]</sup>, Polish<sup>[30]</sup>. But our results were not in agreement with healthy Chinese population<sup>[28]</sup>. When our findings were compared with studies on inflammatory diseases like cancer, they are in accordance with a studies conducted in Asian by Huo *et al.*<sup>[31]</sup> and Zhou *et al.*<sup>[32]</sup>. However our

**Table 4** Association between -94 ins/del ATTG polymorphism in the *NFKB1* gene and diabetic nephropathy at the genotype level

Genotype	OR	95%CI	P value
DM <i>vs</i> NG ref	1.04	0.607-4.987	0.887
DM-CRD <i>vs</i> NG ref	1.95	1.101-3.467	0.022
DM-CRD <i>vs</i> DM ref	1.91	1.080-3.386	0.025

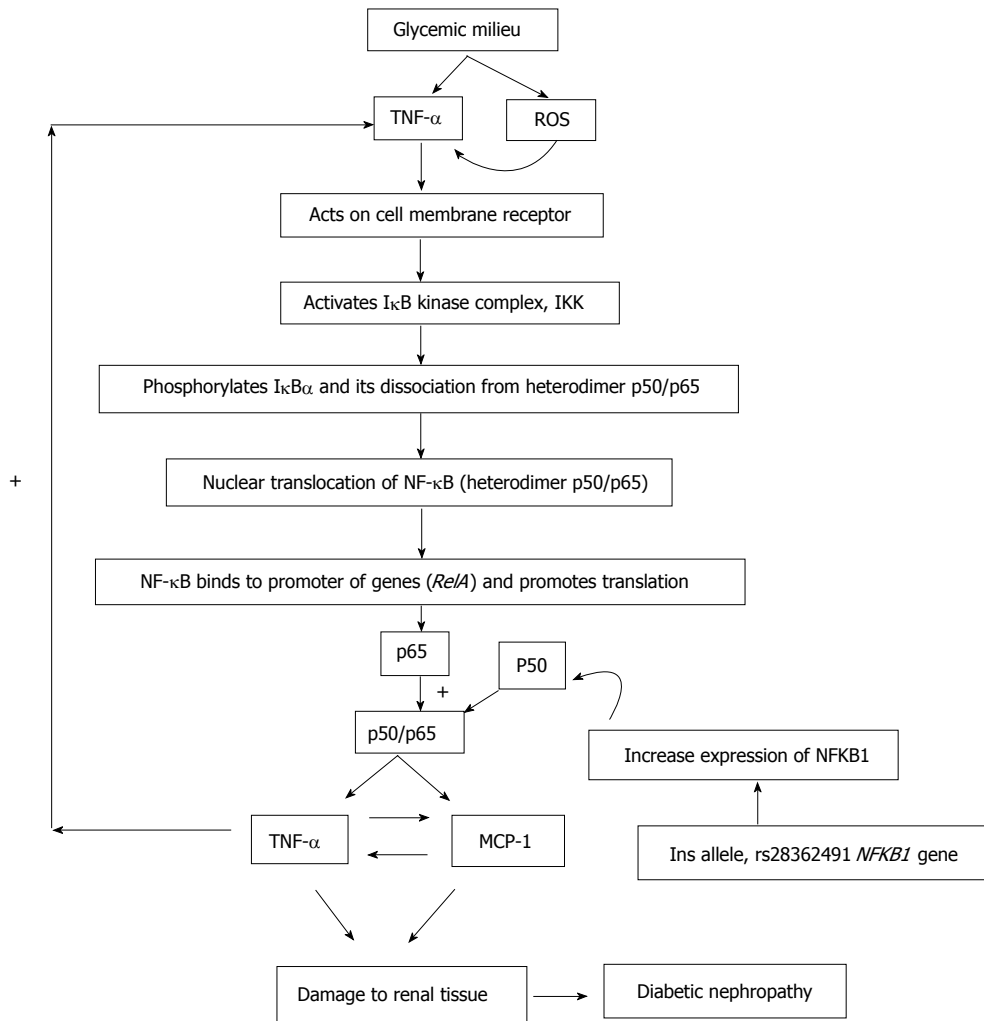
Ref: Reference group.

results were in contrast with a genomic study on cancer conducted by Yang *et al.*<sup>[28]</sup> in 2014. The dissimilarity of results could be due to diverse geographical distribution and ethnicity between our study and theirs was different, which could result in diverse genetic background.

Latest evidence has shown that the production of MCP-1 by kidney affected by diabetes along with TNF-α is a major cause of inflammation, renal injury and fibrosis in DN<sup>[10,17]</sup>. The present study is the foremost one to document the correlation of -94 ins/del AGGT SNP with levels of inflammatory markers namely uMCP-1 and plasma TNF-α in DN from North Indian patients. In our previous study, we have observed that plasma TNF-α and uMCP-1 levels were significantly raised in patients with T2DM and so more in patients with DN<sup>[19]</sup>. To explicate the role of *NFKB1* gene SNP in the development of DN, -94 ins/del AGGT SNP were analyzed in various study, *i.e.*, Group 2 (DM) and Group 3 (DM-CRD) and further correlated with measured inflammatory markers like uMCP-1 and plasma TNF-α levels. Interestingly, this study has also shown that ins allele was significantly associated with increased urinary MCP-1 and plasma TNF-α levels in NG as well as patient groups. However, there is no report in literature to compare our results.

A recent study has shown that TNF-α stimulates the MCP-1 production *via* NF-κB signalling pathway in rat astrocyte cultures<sup>[33]</sup>. TNF-α was found to increase p65 and phosphorylated p65 levels in nuclear extracts of rat astrocytes, hence augmenting MCP-1 levels<sup>[33]</sup>. This supports our finding that increased levels of TNF-α are associated with increased levels of uMCP-1.

Genetic variations are known to play a vital role in determining risk of DN. A number of studies have investigated the relationship of ins allele of -94 ins/del AGGT polymorphism with various inflammatory diseases. Till date not a single study has tried to evaluate the association between this polymorphism and DN risk. Our study is first to document that patients with T2DM having ins/ins genotype were found to have increased risk of developing nephropathy. Latest studies have reported that p50 null mice have a significantly reduced inflammatory response in various models of inflammation such as asthma<sup>[34]</sup>, arthritis<sup>[35]</sup>, and autoimmune encephalomyelitis<sup>[36]</sup>. A similar study conducted in sporadic colorectal cancer (CRC)<sup>[37]</sup> and epithelial ovarian cancer (EOC)<sup>[31]</sup> has supported



**Figure 1 NFKB1 gene and inflammatory markers: Probable mechanisms in the pathogenesis of diabetic nephropathy.** Hypoglycemia induced ROS and TNF- $\alpha$  leads to activation of IKK. IKK causes phosphorylation of I $\kappa$ B $\alpha$  bound to p50/p65. Phosphorylated I $\kappa$ B $\alpha$  dissociate from p50/p65 leading to nuclear translocation of unbound heterodimer p50/p65 (NF- $\kappa$ B). Binding of NF- $\kappa$ B to promoter gene causes translation of p65. Ins allele, rs28362491 *NFKB1* gene, if present, causes increase expression of p50. Hence there is increased production of p50/p65 heterodimer complex. This heterodimer acts on its downstream proinflammatory targets viz: MCP-1 and TNF- $\alpha$ , leading to its synthesis. MCP-1 is a positive regulator of TNF- $\alpha$  and vice versa. Both MCP-1 and TNF- $\alpha$  causes renal damage leading to development of Diabetic nephropathy. ROS: Reactive oxygen species; TNF- $\alpha$ : Tumor necrosis factor- $\alpha$ ; IKK: I $\kappa$ B kinase complex; MCP-1: Monocyte chemoattractant protein-1.

our findings which suggested that ins/ins genotype contribute to significantly increased risk of CRC and EOC. The probable mechanism of -94 ins/del AGGT polymorphism leading to increased risk of developing DN is explained in Figure 1. In almost all cell types, NF- $\kappa$ B complexes are typically localized in the cytoplasm where they bind to I $\kappa$ B inhibitory proteins. However, stimulation with hyperglycemia induced ROS and TNF- $\alpha$  leads to rapid phosphorylation of I $\kappa$ B *via* I- $\kappa$ B kinases complex which is then degraded by ubiquitin-proteasome pathway. On the other hand, simultaneously -94 ins/del AGGT polymorphism might lead to increased synthesis of p50 mRNA. Hence there will be increased production of p50/p65 heterodimer complex which is a well known proinflammatory molecule, since p50/p65 heterodimer acts on its downstream proinflammatory targets viz: MCP-1 and TNF- $\alpha$ , leading to over production of MCP-1 and TNF- $\alpha$ . Thus, there occurs a vicious cycle, *i.e.*, MCP-1 is a positive regulator of TNF- $\alpha$  and vice versa.

The above mentioned probable hypothesis might lead to increased risk of developing renal damage in T2DM. However results of a recent study from China<sup>[38]</sup> in bladder cancer is in contradiction to our findings which could be due to ethnic and geographical differences. Furthermore, the sample size of our study was fairly small than aforementioned bladder cancer study.

The results of the current study suggest that the *NFKB1* promoter -94 ins/del AGGT SNP is associated with increased possibility of developing nephropathy in patients with diabetes. This SNP may be considered as genetic markers for susceptibility to develop nephropathy in patients with T2DM. The limitation of the study is the small sample size. Therefore, further evaluation is necessary in big sample size to look for the possibility of this polymorphisms as potential genetic markers in the near future. This would help to identify patients with type 2 diabetics who may be at higher risk of developing nephropathy.



## ACKNOWLEDGMENTS

This study was financially supported by Indian Council of Medical Research and Postgraduate Research Grant, University College of Medical Sciences, New Delhi. The authors are thankful to the Department of Biostatistics and Medical Informatics, University College of Medical Sciences, Delhi for statistical analysis.

## COMMENTS

### Background

Type 2 diabetes mellitus (T2DM) is considered as a long standing inflammatory disease. Nuclear factor-kappa B (NF- $\kappa$ B) controls the expression of numerous genes affecting inflammation, immune response. Immunogenic and inflammatory cytokines like monocyte chemoattractant protein-1 (MCP-1) and tumor necrosis factor-alpha (TNF- $\alpha$ ) plays a crucial role in the pathogenesis of micro-vascular complication of T2DM, i.e., diabetic nephropathy (DN) and clinical outcome.

### Research frontiers

In spite of the present advances in our knowledge about the etiopathogenesis of DN, the intricate mechanisms leading to the development of renal injury from chronic hyperglycemia are not yet fully understood. *NFKB1* promoter polymorphism -94 ins/del ATTG has been associated with inflammatory diseases, autoimmune diseases and cancers. However, its role in the development of T2DM and DN has not been explored till date. The authors hypothesized that the -94 ins/del ATTG polymorphism would affect the levels of urinary MCP-1 and plasma TNF- $\alpha$  and therefore might be culprit in developing DN.

### Innovations and breakthroughs

The authors have recently reported that -94 ATTG ins allele was associated with significantly increased levels of urinary MCP-1, plasma TNF- $\alpha$  and was found to increase risk for the development of DN by 1.91-fold in subjects with diabetes.

### Applications

-94 ins/del AGGT polymorphisms can be considered as genetic marker for identifying those more susceptible and provide suitable interventions to delay the progression of DN. This study provides a ground for the development of newer anti-inflammatory therapeutic agents that may have potential to affect primary mechanisms contributing to the pathogenesis of DN.

### Terminology

DN: Diabetic nephropathy; NF- $\kappa$ B: Nuclear factor-kappa B; *NFKB1*: Nuclear factor-kappa B1 gene; T2DM: Type 2 diabetes mellitus; TNF- $\alpha$ : Tumor necrosis factor-alpha; uMCP-1: Urinary Monocyte chemoattractant protein-1.

### Peer-review

The manuscript is well informative.

## REFERENCES

- 1 Levey AS, Eckardt KU, Tsukamoto Y, Levin A, Coresh J, Rossert J, De Zeeuw D, Hostetter TH, Lameire N, Eknoyan G. Definition and classification of chronic kidney disease: a position statement from Kidney Disease: Improving Global Outcomes (KDIGO). *Kidney Int* 2005; **67**: 2089-2100 [PMID: 15882252]
- 2 Pan HZ, Zhang L, Guo MY, Sui H, Li H, Wu WH, Qu NQ, Liang MH, Chang D. The oxidative stress status in diabetes mellitus and diabetic nephropathy. *Acta Diabetol* 2010; **47** Suppl 1: 71-76 [PMID: 19475334 DOI: 10.1007/s00592-009-0128-1]
- 3 Arnalich F, Hernanz A, López-Maderuelo D, Peña JM, Camacho J, Madero R, Vázquez JJ, Montiel C. Enhanced acute-phase response and oxidative stress in older adults with type II diabetes. *Horm Metab Res* 2000; **32**: 407-412 [PMID: 11069205 DOI: 10.1055/s-2007-978662]
- 4 Esposito K, Nappo F, Marfella R, Giugliano G, Giugliano F, Ciotola M, Quagliaro L, Ceriello A, Giugliano D. Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans: role of oxidative stress. *Circulation* 2002; **106**: 2067-2072 [PMID: 12379575]
- 5 Karin M, Ben-Neriah Y. Phosphorylation meets ubiquitination: the control of NF-[kappa]B activity. *Annu Rev Immunol* 2000; **18**: 621-663 [PMID: 10837071 DOI: 10.1146/annurev.immunol.18.1.621]
- 6 Barnes PJ, Karin M. Nuclear factor-kappaB: a pivotal transcription factor in chronic inflammatory diseases. *N Engl J Med* 1997; **336**: 1066-1071 [PMID: 9091804 DOI: 10.1056/NEJM199704103361506]
- 7 Gu L, Tseng SC, Rollins BJ. Monocyte chemoattractant protein-1. *Chem Immunol* 1999; **72**: 7-29 [PMID: 10550927]
- 8 Rovin BH, Yoshimura T, Tan L. Cytokine-induced production of monocyte chemoattractant protein-1 by cultured human mesangial cell. *J Immunol* 1992; **148**: 2148-2153
- 9 Wada T, Furuichi K, Sakai N, Iwata Y, Yoshimoto K, Shimizu M, Takeda SI, Takasawa K, Yoshimura M, Kida H, Kobayashi KI, Mukaida N, Naito T, Matsushima K, Yokoyama H. Up-regulation of monocyte chemoattractant protein-1 in tubulointerstitial lesions of human diabetic nephropathy. *Kidney Int* 2000; **58**: 1492-1499 [PMID: 11012884 DOI: 10.1046/j.1523-1755.2000.00311.x]
- 10 Tesch GH. MCP-1/CCL2: a new diagnostic marker and therapeutic target for progressive renal injury in diabetic nephropathy. *Am J Physiol Renal Physiol* 2008; **294**: F697-F701 [PMID: 18272603 DOI: 10.1152/ajprenal.00016.2008]
- 11 Ihm CG, Park JK, Hong SP, Lee TW, Cho BS, Kim MJ, Cha DR, Ha H. A high glucose concentration stimulates the expression of monocyte chemotactic peptide 1 in human mesangial cells. *Nephron* 1998; **79**: 33-37 [PMID: 9609459]
- 12 Banba N, Nakamura T, Matsumura M, Kuroda H, Hattori Y, Kasai K. Possible relationship of monocyte chemoattractant protein-1 with diabetic nephropathy. *Kidney Int* 2000; **58**: 684-690 [PMID: 10916091 DOI: 10.1046/j.1523-1755.2000.00214.x]
- 13 Sugimoto H, Shikata K, Wada J, Horiuchi S, Makino H. Advanced glycation end products-cytokine-nitric oxide sequence pathway in the development of diabetic nephropathy: aminoguanidine ameliorates the overexpression of tumor necrosis factor-alpha and inducible nitric oxide synthase in diabetic rat glomeruli. *Diabetologia* 1999; **42**: 878-886 [PMID: 10440132]
- 14 Pamir N, McMillen TS, Kaiyala KJ, Schwartz MW, LeBoeuf RC. Receptors for tumor necrosis factor-alpha play a protective role against obesity and alter adipose tissue macrophage status. *Endocrinology* 2009; **150**: 4124-4134 [PMID: 19477937 DOI: 10.1210/en.2009-0137]
- 15 Luo SF, Fang RY, Hsieh HL, Chi PL, Lin CC, Hsiao LD, Wu CC, Wang JS, Yang CM. Involvement of MAPKs and NF-kappaB in tumor necrosis factor alpha-induced vascular cell adhesion molecule 1 expression in human rheumatoid arthritis synovial fibroblasts. *Arthritis Rheum* 2010; **62**: 105-116 [PMID: 20039412 DOI: 10.1002/art.25060]
- 16 Nakamura T, Fukui M, Ebihara I, Osada S, Nagaoka I, Tomino Y, Koide H. mRNA expression of growth factors in glomeruli from diabetic rats. *Diabetes* 1993; **42**: 450-456 [PMID: 8094359]
- 17 Rivero A, Mora C, Muros M, García J, Herrera H, Navarro-González JF. Pathogenic perspectives for the role of inflammation in diabetic nephropathy. *Clin Sci (Lond)* 2009; **116**: 479-492 [PMID: 19200057 DOI: 10.1042/CS20080394]
- 18 Makuc J, Petrović D. A review of oxidative stress related genes and new antioxidant therapy in diabetic nephropathy. *Cardiovasc Hematol Agents Med Chem* 2011; **9**: 253-261 [PMID: 21902657]
- 19 Gupta S, Gambhir JK, Kalra O, Gautam A, Shukla K, Mehndiratta M, Agarwal S, Shukla R. Association of biomarkers of inflammation and oxidative stress with the risk of chronic kidney disease in Type 2 diabetes mellitus in North Indian population. *J Diabetes Complications* 2013; **27**: 548-552 [PMID: 24012111 DOI: 10.1016/j.jdiacomp.2013.07.005]
- 20 Gupta S, Mehndiratta M, Kalra S, Kalra OP, Shukla R, Gambhir JK. Association of tumor necrosis factor (TNF) promoter polymorphisms with plasma TNF- $\alpha$  levels and susceptibility to diabetic nephropathy in North Indian population. *J Diabetes*

- Complications* 2015; **29**: 338-342 [PMID: 25704106 DOI: 10.1016/j.jdiacomp.2015.01.002]
- 21 **Karban AS**, Okazaki T, Panhuysen CI, Gallegos T, Potter JJ, Bailey-Wilson JE, Silverberg MS, Duerr RH, Cho JH, Gregersen PK, Wu Y, Achkar JP, Dassopoulos T, Mezey E, Bayless TM, Novet FJ, Brant SR. Functional annotation of a novel NFKB1 promoter polymorphism that increases risk for ulcerative colitis. *Hum Mol Genet* 2004; **13**: 35-45 [PMID: 14613970 DOI: 10.1093/hmg/ddh008]
  - 22 **Senol Tuncay S**, Okay P, Bardakci F. Identification of NF-kappaB1 and NF-kappaB1Alpha polymorphisms using PCR-RFLP assay in a Turkish population. *Biochem Genet* 2010; **48**: 104-112 [PMID: 19941056 DOI: 10.1007/s10528-009-9302-y]
  - 23 **American Diabetic Association**. Standards of medical care in diabetes-2015. *Diabetes Care* 2015; **38**: S1-S9
  - 24 **Levey AS**, Bosch JP, Lewis JB, Greene T, Rogers N, Roth D. A more accurate method to estimate glomerular filtration rate from serum creatinine: a new prediction equation. Modification of Diet in Renal Disease Study Group. *Ann Intern Med* 1999; **130**: 461-470 [PMID: 10075613]
  - 25 **James PA**, Oparil S, Carter BL, Cushman WC, Dennison-Himmelfarb C, Handler J, Lackland DT, LeFevre ML, MacKenzie TD, Ogdegebe O, Smith SC, Svetkey LP, Taler SJ, Townsend RR, Wright JT, Narva AS, Ortiz E. 2014 evidence-based guideline for the management of high blood pressure in adults: report from the panel members appointed to the Eighth Joint National Committee (JNC 8). *JAMA* 2014; **311**: 507-520 [PMID: 24352797 DOI: 10.1001/jama.2013.284427]
  - 26 **Chen F**, Castranova V, Shi X, Demers LM. New insights into the role of nuclear factor-kappaB, a ubiquitous transcription factor in the initiation of diseases. *Clin Chem* 1999; **45**: 7-17 [PMID: 9895331]
  - 27 **Romzova M**, Hohenadel D, Kolostova K, Pinterova D, Fojtikova M, Ruzickova S, Dostal C, Bosak V, Rychlik I, Cerna M. NFkappaB and its inhibitor IkappaB in relation to type 2 diabetes and its microvascular and atherosclerotic complications. *Hum Immunol* 2006; **67**: 706-713 [PMID: 17002901 DOI: 10.1016/J.humimm.2006.05.006]
  - 28 **Yang X**, Li P, Tao J, Qin C, Cao Q, Gu J, Deng X, Wang J, Liu X, Wang Z, Wu B, Gu M, Lu Q, Yin C. Association between NFKB1 -94ins/del ATTG Promoter Polymorphism and Cancer Susceptibility: An Updated Meta-Analysis. *Int J Genomics* 2014; **2014**: 612972 [PMID: 24895544 DOI: 10.1155/2014/612972]
  - 29 **Mirza MM**, Fisher SA, Onnie C, Lewis CM, Mathew CG, Sanderson J, Forbes A. No association of the NFKB1 promoter polymorphism with ulcerative colitis in a British case control cohort. *Gut* 2005; **54**: 1205-1206 [PMID: 16009698 DOI: 10.1136/gut.2005.070029]
  - 30 **Kurylowicz A**, Hiromatsu Y, Jurecka-Lubieniecka B, Kula D, Kowalska M, Ichimura M, Koga H, Kaku H, Bar-Andziak E, Nauman J, Jarzab B, Ploski R, Bednarczuk T. Association of NFKB1 -94ins/del ATTG promoter polymorphism with susceptibility to and phenotype of Graves' disease. *Genes Immun* 2007; **8**: 532-538 [PMID: 17690684 DOI: 10.1038/sj.gene.6364418]
  - 31 **Huo ZH**, Zhong HJ, Zhu YS, Xing B, Tang H. Roles of functional NFKB1 and  $\beta$ -TrCP insertion/deletion polymorphisms in mRNA expression and epithelial ovarian cancer susceptibility. *Genet Mol Res* 2013; **12**: 3435-3443 [PMID: 23546975 DOI: 10.4238/2013.March.11.6]
  - 32 **Zhou B**, Qie M, Wang Y, Yan L, Zhang Z, Liang A, Wang T, Wang X, Song Y, Zhang L. Relationship between NFKB1 -94 insertion/deletion ATTG polymorphism and susceptibility of cervical squamous cell carcinoma risk. *Ann Oncol* 2010; **21**: 506-511 [PMID: 19892748 DOI: 10.1093/annonc/mdp507]
  - 33 **Thompson WL**, Van Eldik LJ. Inflammatory cytokines stimulate the chemokines CCL2/MCP-1 and CCL7/MCP-3 through NFkB and MAPK dependent pathways in rat astrocytes [corrected]. *Brain Res* 2009; **1287**: 47-57 [PMID: 19577550 DOI: 10.1016/j.brainres.2009.06.081]
  - 34 **Yang L**, Cohn L, Zhang DH, Homer R, Ray A, Ray P. Essential role of nuclear factor kappaB in the induction of eosinophilia in allergic airway inflammation. *J Exp Med* 1998; **188**: 1739-1750 [PMID: 9802985]
  - 35 **Campbell IK**, Gerondakis S, O'Donnell K, Wicks IP. Distinct roles for the NF-kappaB1 (p50) and c-Rel transcription factors in inflammatory arthritis. *J Clin Invest* 2000; **105**: 1799-1806 [PMID: 10862795 DOI: 10.1172/JCI8298]
  - 36 **Hilliard B**, Samoilova EB, Liu TS, Rostami A, Chen Y. Experimental autoimmune encephalomyelitis in NF-kappa B-deficient mice: roles of NF-kappa B in the activation and differentiation of autoreactive T cells. *J Immunol* 1999; **163**: 2937-2943 [PMID: 10453042]
  - 37 **Mohd Suzairi MS**, Tan SC, Ahmad Aizat AA, Mohd Aminudin M, Siti Nurfatimah MS, Andee ZD, Ankathil R. The functional -94 insertion/deletion ATTG polymorphism in the promoter region of NFKB1 gene increases the risk of sporadic colorectal cancer. *Cancer Epidemiol* 2013; **37**: 634-638 [PMID: 23806437 DOI: 10.1016/j.canep.2013.05.007]
  - 38 **Li P**, Gu J, Yang X, Cai H, Tao J, Yang X, Lu Q, Wang Z, Yin C, Gu M. Functional promoter -94 ins/del ATTG polymorphism in NFKB1 gene is associated with bladder cancer risk in a Chinese population. *PLoS One* 2013; **8**: e71604 [PMID: 23977085 DOI: 10.1371/journal.pone.0071604]

**P- Reviewer:** Lehtonen SH, Sameer AS, Wada J **S- Editor:** Ji FF  
**L- Editor:** A **E- Editor:** Wu HL



## Case Control Study

# Exercise-induced albuminuria vs circadian variations in blood pressure in type 1 diabetes

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**Author contributions:** All the authors approved the final version of the manuscript.

**Institutional review board statement:** The study protocol was approved by the Ethical Committee of the Institut Supérieur des Sciences de la Santé, Université des Montagnes, Bangangté, Cameroon, and was conducted in accordance with the guidelines of the Helsinki Declaration.

**Informed consent statement:** All participants and their parents or guardians (since many were adolescents) provided informed written concern prior to study enrollment.

**Conflict-of-interest statement:** No benefits in any form have been received or will be received from a commercial party related directly or indirectly to the subject of this article.

**Data sharing statement:** Data are available from the corresponding author upon request at [sobngwieugene@yahoo.fr](mailto:sobngwieugene@yahoo.fr).

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**Manuscript source:** Invited manuscript

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Telephone: +237-675088750

**Received:** July 10, 2016

**Peer-review started:** July 14, 2016

**First decision:** September 12, 2016

**Revised:** October 19, 2016

**Accepted:** November 27, 2016

**Article in press:** November 29, 2016

**Published online:** February 15, 2017

## Abstract

### AIM

To investigated the relationship between exercise-induced ambulatory blood pressure measurement (ABPM) abnormalities in type 1 diabetes mellitus (T1DM) adolescents.

### METHODS

We conducted a case-control at the National Obesity Center of the Yaoundé Central Hospital, Cameroon. We compared 24 h ABPM and urinary albumin-to-creatinine ratio (ACR) at rest and after a standardized treadmill exercise between 20 Cameroonian T1DM patients and 20 matched controls. T1DM adolescents were aged 12-18 years, with diabetes for at least one year, without proteinuria, with normal office blood pressure (BP) and renal function according to the general reference

population. Non-diabetic controls were adolescents of general population matched for sex, age and BMI.

## RESULTS

Mean duration of diabetes was  $4.2 \pm 2.8$  years. The mean 24 h systolic blood pressure (SBP) and diastolic blood pressure (DBP) were respectively  $116 \pm 9$  mmHg in the diabetic group vs  $111 \pm 8$  mmHg in the non-diabetic ( $P = 0.06$ ), and  $69 \pm 7$  mmHg vs  $66 \pm 5$  mmHg ( $P = 0.19$ ). There was no difference in the diurnal pattern of BP in diabetes patients and non-diabetic controls (SBP:  $118 \pm 10$  mmHg vs  $114 \pm 10$  mmHg,  $P = 0.11$ ; DBP:  $71 \pm 7$  mmHg vs  $68 \pm 6$  mmHg,  $P = 0.22$ ). Nighttime BP was higher in the diabetic group with respect to SBP ( $112 \pm 11$  mmHg vs  $106 \pm 7$  mmHg,  $P = 0.06$ ) and to the mean arterial pressure (MAP) ( $89 \pm 9$  mmHg vs  $81 \pm 6$  mmHg,  $P = 0.06$ ). ACR at rest was similar in both groups ( $5.5$  mg/g vs  $5.5$  mg/g,  $P = 0.74$ ), but significantly higher in diabetes patients after exercise ( $10.5$  mg/g vs  $5.5$  mg/g,  $P = 0.03$ ). SBP was higher in patients having exercise-induced albuminuria ( $116 \pm 10$  mmHg vs  $108 \pm 10$  mmHg,  $P = 0.09$ ).

## CONCLUSION

Exercise-induced albuminuria could be useful for early diagnosis of kidney damage in adolescents with T1DM.

**Key words:** Albuminuria; Blood pressure; Ambulatory blood pressure measurement; Exercise; Type 1 diabetes

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**Core tip:** Diabetic nephropathy (DN) is a major complication of type 1 diabetes mellitus (T1DM). Therefore, strategies for early detection are of critical importance. Ambulatory blood pressure measurement is useful for detection of precocious abnormalities in the occurrence of DN and exercise-induced albuminuria has been proposed as a potential predictor of DN. Our study therefore aimed to investigate the relationship between exercise-induced albuminuria and ambulatory blood pressure measurement abnormalities in T1DM Cameroonian adolescents. We found that T1DM patients had higher nocturnal and 24 h blood pressure figures than non-diabetics suggesting that exercise-induced albuminuria could be useful early detection of diabetes kidney injuries in T1DM.

Tadida Meli IH, Tankeu AT, Dehayem MY, Chelo D, Noubiap JJN, Sobngwi E. Exercise-induced albuminuria vs circadian variations in blood pressure in type 1 diabetes. *World J Diabetes* 2017; 8(2): 74-79 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v8/i2/74.htm> DOI: <http://dx.doi.org/10.4239/wjd.v8.i2.74>

## INTRODUCTION

Diabetes nephropathy is the major life-threatening complication of type 1 diabetes mellitus (T1DM)<sup>[1,2]</sup>.

Abnormal albumin excretion has been shown to predict the development of clinically significant nephropathy in T1DM. Indeed, persistent minimal elevation of albuminuria at rest predicts the development of more severe proteinuria and clinical diabetic nephropathy, which frequently progresses to renal failure<sup>[3]</sup>. In T1DM, nephropathy develops in 30% to 40% of cases and impaired renal function or end-stage kidney disease affect up to a third of patients<sup>[4]</sup>. Thus, strategies for early detection and for preventative interventions are of critical importance since interventions at these late stages of disease may only slow but not completely arrest the inexorable progression towards renal failure<sup>[5,6]</sup>. In this direction, it has been shown that physical exercise can stimulate albuminuria in diabetes patients and can be a useful provocative test to detect early renal abnormalities<sup>[7]</sup>. However, there is still limited evidence on its value for early detection of renal disease in T1DM.

Previous studies has proven that during exercise, urinary albumin excretion rate is more increased in long term T1DM patients thus, at risk of developing diabetes nephropathy than in general population<sup>[8]</sup>. In the contrary, some evidence suggest that the level of albumin excretion during exercise is related to the quality of metabolic control; for example, exercise-induced microalbuminuria is more pronounced in newly diagnosed patients, and this abnormality is reversed by insulin treatment. Exercise-induced microalbuminuria generally is not well correlated with the duration of disease and does not predict clinical nephropathy<sup>[9]</sup>. On the other hand, the contribution of night-time blood pressure (BP) on the onset of nephropathy in diabetic patients is now established<sup>[10]</sup>. Therefore ambulatory blood pressure monitoring (ABPM) could be proposed as an useful tool for early detection of diabetic nephropathy<sup>[11,12]</sup>. This study aimed to investigate the relationship between exercise-induced albuminuria and ABPM abnormalities in early detection of diabetic nephropathy in adolescents with T1DM from Cameroon.

## MATERIALS AND METHODS

### Study subjects

This case-control study was carried out at the National Obesity Center of the Yaoundé Central Hospital, the reference diabetes center in the town. Our population was made of two groups, T1DM adolescents and non-diabetic controls. T1DM patients were aged 12-18 years, with diabetes for at least one year; without proteinuria, with normal office BP and renal function according to the general reference population. Non-diabetic controls were adolescents of general population matched for sex, age and BMI. We excluded patients and controls with an important night activity, those receiving drugs for hypertension or any other drugs able to modify albuminuria, those with contra-indication to exercise or presenting signs of urinary tract infection as well as those having fever and pregnant women.



### Procedure and investigations

The procedure was made of an inclusion visit and two exploration visits. Within 2 wk following an information visit, for all eligible participants, we performed a careful clinical exam including BP measurement and a urinary dipstick. We enrolled 40 participants, 20 in each group.

All exploration visits were conducted in the morning between 8:00 and 10:00. After arrival, participants were invited to stay in sitting position for at least five minutes. Then, clinical measurement of BP was done three times using an automated sphygmomanometer Omron HEM-705 CP (Omron Corporation, Tokyo, Japan) placed on the left arm raised itself at the heart level. The average of three measures was considered for analysis. Weight and height were respectively valued to the nearest 0.5 unit using a mechanical scale and a measuring rod and body mass index (BMI in kg/m<sup>2</sup>) calculated as  $weight\ (cm)/[height\ (m) \times height\ (m)]$ . A dipstick was done to assess proteinuria and considered positive for at least 1+.

ABPM was carried out on twenty four hours using an automatic portable, light weight monitor device the i-MAPA® CE 004 1.1 TM (High-tech Medical St Louis, Paris) which performs measurements every 15 min during daytime (07:00 to 22:00) and twice an hour during night time defined from 22:00 to 07:00. Device was activated and the two first measures performed in the laboratory to ensure functionality. Detailed information on the operation and use of the device were then given to the participant who then returned to his daily activities. At least 70% of valid measurements were considered for interpretation.

The exercise protocol was developed according to the Hierarchy of individual calibration levels for heart rate and accelerometry to measure physical activity<sup>[13]</sup>. It was made of 1 km race on a treadmill at 5.8 km/h and was divided in two phases. The first phase was made of a 3 min gathering speed up to 3.2 km/h, followed by an acceleration of 0.33 km/h every 6 min. The second phase was a walking step between 5.2-5.8 km/h on the treadmill.

Albuminuria was calculated using albumin-to-creatinine ratio in order to avoid effect of exercise on urinary concentration and expressed in mg/g. First void urine collection was used for rest albuminuria and a random sample urine was collected within the 20 min following physical exercise to measure exercise-induced albuminuria. Albuminuria or exercise-induced albuminuria was diagnosed on the basis of a urinary albumin excretion rate greater than 20 but less than 200 mg/g<sup>[14]</sup>. Adverse events such as hypoglycemia during physical exercise or exercise intolerance, were closely monitored.

### Statistical analysis

Data acquisition was done by Epi-data 3.1 software and statistical analysis was performed using Stata 12.0 software. Continuous variables are expressed as means with standard deviation (SD) where appropriate,

**Table 1 Ambulatory blood pressure measurement of the diabetes and non-diabetes patients**

Variables	Type 1 diabetic patients (n = 20)	Non-diabetic patients (n = 20)	P value
24 h BP			
SBP	116 ± 9	111 ± 8	0.06
DBP	69 ± 7	66 ± 5	0.19
PP	48 ± 8	45 ± 5	0.11
Diurnal BP			
SBP	118 ± 10	114 ± 10	0.11
MAP	92 ± 7	89 ± 7	0.15
DBP	71 ± 7	68 ± 6	0.22
Nocturnal BP			
SBP	112 ± 11	106 ± 7	0.06
MAP	85 ± 9	81 ± 6	0.06
DBP	64 ± 9	60 ± 6	0.11

SBP: Systolic blood pressure; DBP: Diastolic blood pressure; MAP: Mean arterial blood pressure; PP: Pulse pressure.

and categorical variables as count (percentage). The Spearman rank coefficient was used to test correlations. The  $\chi^2$  test and Mann-Whitney rank sum test were used to test associations between qualitative variables and difference between two respectively. A *P* value  $\leq 0.05$  was considered statistically significant. The statistical methods of this study were reviewed by Mr. Sontsa.

## RESULTS

### General characteristics

We enrolled 40 participants, 24 males, average age of  $16 \pm 2$  years. The mean BMI of diabetes patients was  $22.6 \pm 2.9$  kg/m<sup>2</sup> vs  $22.7 \pm 3.3$  kg/m<sup>2</sup> for non-diabetic. Average duration of diabetes was  $4.2 \pm 2.8$  years with mean glycated hemoglobin of  $9.9 \pm 2.8$ . Nine diabetes patients had a family history of hypertension vs six in the non-diabetic group.

### ABPM measurement of study population

Diabetes participants had lightly higher BP values compared to non-diabetic on every component (Table 1). Thus, 24 h SBP measurement in the diabetic group was  $116 \pm 9$  mmHg vs  $111 \pm 8$  mmHg for non-diabetics at borderline of significance (*P* = 0.06) while difference in DBP of two groups was non-significant ( $69 \pm 7$  mmHg vs  $66 \pm 5$  mmHg; *P* = 0.19). In keeping with that, diurnal BP figures were slightly higher in the diabetic group but with a non-significant difference (SBP:  $118 \pm 10$  mmHg vs  $114 \pm 10$  mmHg, *P* = 0.11; DBP:  $71 \pm 7$  mmHg vs  $68 \pm 6$  mmHg; *P* = 0.22). One important finding was the elevated night time BP in diabetes adolescents with a borderline significance for SBP ( $112 \pm 11$  mmHg vs  $106 \pm 7$  mmHg, *P* = 0.06) and MAP ( $85 \pm 9$  mmHg vs  $81 \pm 6$  mmHg, *P* = 0.06).

### Urinary albumin excretion of study population

In adolescents with diabetes, 06/20 (30%) developed abnormal exercise-induced albuminuria but none in the group of adolescents without diabetes. Urinary albumin

**Table 2 Comparison of blood pressure values for albuminurics and non albuminurics patients**

	UAE < 20 mg/g	UAE > 20 mg/g	P value
24 h BP			
SBP	113 ± 9	119 ± 10	0.14
DBP	67 ± 6	70 ± 8	0.51
Diurnal BP			
SBP	116 ± 10	120 ± 10	0.32
DBP	70 ± 5	72 ± 8	0.51
Nocturnal BP			
SBP	108 ± 10	116 ± 10	0.09
DBP	61 ± 8	66 ± 9	0.17

SBP: Systolic blood pressure; DBP: Diastolic blood pressure; UAE: Urinary albumin excretion.

excretion at rest was similar in both groups (5.5 mg/g vs 5.5 mg/g,  $P = 0.74$ ). After exercise, we found a significant increase in urinary albumin excretion in diabetes patients as compared to non-diabetics (10.5 mg/g vs 5.5 mg/g,  $P = 0.03$ ).

#### Relation between BP profile and albuminuria at rest and after exercise

We compared diabetes adolescents presenting exercise-induced albuminuria after exercise to those without albuminuria (Table 2). We found that diabetes patients with exercise-induced albuminuria had higher but non-significant nighttime SBP figures than those exercise-induced albuminuria (116 mmHg vs 108 mmHg,  $P = 0.09$ ) while DBP were similar. In contrast, 24 h SBP and DBP were similar in both as well as diurnal SBP and DBP.

## DISCUSSION

This study aimed to investigate the relationship between exercise-induced albuminuria and circadian BP abnormalities revealed by ABPM in non proteinuric T1DM adolescents. In order to achieve this objective, we compared young T1DM patients to non-diabetic matched controls. We found that nocturnal SBP of diabetic patients was slightly higher than that of non-diabetics as well as 24 h SBP with borderline significance. Most T1DM studies on albuminuria disease have been done in Caucasians<sup>[14-17]</sup>. This study confirms these findings in Africans. This increase in nocturnal SBP values and 24 h SBP already found by others studies suggest the existence in this group of probable subclinical kidney injuries. Indeed, it was demonstrated that diabetes patients with kidney injury or subclinical diabetic nephropathy had a tendency to higher BP than the general population<sup>[14-18]</sup>. Similarly, diabetes patients in our study have a tendency to increased nocturnal BP figures in comparison to non-diabetics leading to a reduction in the difference of day-night BP evaluated by dipping<sup>[19,20]</sup>. This anomaly is found more frequently in diabetes patients compared than in the general population and is attributed to the presence of

kidney damage, still subclinical, but already leading to an increase in renal and cardiovascular risk<sup>[18]</sup>. Thus, the studies comparing individuals with impaired nocturnal decline in BP and those with normal nocturnal BP have revealed that individuals with insufficient decrease of BP and therefore higher values of BP during the night will present in future monitoring a more rapid degradation of renal function marked by a significant decrease in creatinine clearance<sup>[21]</sup>. In the same sense, these studies did not find any difference between daytime BP as well as diastolic BP which was also to be the case in our study where daytime BP were similar in both groups of participants<sup>[18,20]</sup>. However, unlike these studies, we found 24 h BP figures slightly higher in diabetes individuals but still of borderline significance. This could be attributed to the impact of nighttime BP on the 24 h BP and would be a reflection of the nocturnal difference since for similar diurnal BP, if the nocturnal BP is elevated in one group, then it becomes logical that the 24 h BP which is the average daytime and nighttime BP appears to be also more elevated.

Secondly, our study showed that for similar or even identical values of albuminuria at rest, diabetes patients having an increase in nocturnal BP and therefore probable subclinical kidney injuries had a significantly increase in exercise-induced albuminuria in comparison to non-diabetic individuals. This suggests that exercise-induced albuminuria increases with the existence of renal alterations revealed by abnormal nocturnal BP and therefore could be used to detect patients with these abnormalities. This finding support the assumption that exercise-induced albuminuria could serve as a marker of early diabetic renal injuries and allow detection or at least help to suspect the existence of subclinical diabetic nephropathy still undetectable by albuminuria at rest. This had been suggested in 1995 by O'Brien who found during a prospective follow-up on a half-decade that patients having abnormal exercise-induced albuminuria were those who would develop a clinical albuminuria at rest and therefore faster diabetic nephropathy<sup>[22-25]</sup>. But to the best of our knowledge, nobody has so far studied the relationship between exercise-induced albuminuria and nocturnal abnormalities of BP in type 1 diabetes patients. This first finding then proves very encouraging since it opens the way to new opportunities and show new research fields to explore.

Finally, we compared the diurnal and nocturnal BP values of patients who developed exercise-induced albuminuria to those of other participants without this abnormality. We found that patients with exercise-induced albuminuria had higher non-significant figures of BP during the night than those without this abnormality. These data support the hypothesis emitted above that exercise induced-albuminuria could be used to identify T1DM patients with abnormal nocturnal BP and therefore at risk of developing diabetic nephropathy or already presenting subclinical damage due to diabetic nephropathy. However, these findings casually refer to other studies on the subject with larger population study

and ideally with a prospective follow-up in order to clearly establish the link between exercise-induced albuminuria and renal prognosis and cardiovascular evaluated by circadian BP on ABPM and especially nocturnal BP abnormalities in T1DM<sup>[26-28]</sup>.

In summary, T1DM patients having an increase in nocturnal BP exhibit an increase exercise-induced albuminuria and patients developing abnormal exercise-induced albuminuria have higher figures of nocturnal BP than others. These findings strongly suggest that exercise-induced albuminuria could be use identify diabetes patients with subclinical renal damage, therefore it would be useful in the early diagnosis of nephropathy in T1DM.

## ACKNOWLEDGMENTS

We gratefully acknowledge all the patients who have accepted to take part in this study.

## COMMENTS

### Background

Nocturnal abnormalities of blood pressure are correlated with incipient diabetes nephropathy in type 1 diabetes adolescents, but relation with exercised induced-albuminuria has not been investigated yet. Few studies have been conducted on diabetic nephropathy in Africans adolescents.

### Research frontiers

Studies on diabetic nephropathy in Africans adolescents are scarce. These data are important to determine the tie between exercise-induced albuminuria and nocturnal blood pressure abnormalities in type 1 diabetes adolescents and the possibility to use it as an earlier marker for diabetes nephropathy.

### Innovations and breakthroughs

The authors confirm data of Caucasians studies suggesting that most type 1 diabetes adolescents developed diabetes nephropathy after five years. This study was the first investigating the relationship between exercise-induced albuminuria and ambulatory blood pressure measurement measurements in type 1 diabetes adolescents in the search of early markers of diabetic nephropathy.

### Applications

This study shows that there is a relation between exercised-induced albuminuria and nocturnal abnormalities of circadian blood pressure suggesting that exercised-induced albuminuria could be useful as clinical marker for blunted night-time in type 1 diabetes adolescents.

### Peer-review

This is a nice study and well done, the topic is clear and the conclusion is novel.

## REFERENCES

- Chiarelli F, Verrotti A, Mohn A, Morgese G. The importance of microalbuminuria as an indicator of incipient diabetic nephropathy: therapeutic implications. *Ann Med* 1997; **29**: 439-445 [PMID: 9453292 DOI: 10.3109/07853899708999374]
- Bennett PH. 'Microalbuminuria' and diabetes: a critique--assessment of urinary albumin excretion and its role in screening for diabetic nephropathy. *Am J Kidney Dis* 1989; **13**: 29-34 [PMID: 2912062 DOI: 10.1016/S0272-6386(89)80111-8]
- Lehmann R, Spinass GA. [Diabetic nephropathy: significance of microalbuminuria and proteinuria in Type I and Type II diabetes mellitus]. *Praxis (Bern 1994)* 1995; **84**: 1265-1271 [PMID: 7491450]
- Perkins BA, Krolewski AS. Early nephropathy in type 1 diabetes: a new perspective on who will and who will not progress. *Curr Diab Rep* 2005; **5**: 455-463 [PMID: 16316598 DOI: 10.1007/s11892-005-0055-7]
- Steinke JM, Mauer M. Lessons learned from studies of the natural history of diabetic nephropathy in young type 1 diabetic patients. *Pediatr Endocrinol Rev* 2008; **5** Suppl 4: 958-963 [PMID: 18806710]
- Taboga C, Tonutti L, Noacco C. Effects of physical activity on microalbuminuria in type 1 diabetics without nephropathy. *G Clin Med* 1990; **71**: 569-572 [PMID: 2289652]
- Feldt-Rasmussen B, Baker L, Deckert T. Exercise as a provocative test in early renal disease in type 1 (insulin-dependent) diabetes: albuminuric, systemic and renal haemodynamic responses. *Diabetologia* 1985; **28**: 389-396 [PMID: 4043581 DOI: 10.1007/BF00280880]
- Mogensen CE. Microalbuminuria as a predictor of clinical diabetic nephropathy. *Kidney Int* 1987; **31**: 673-689 [PMID: 3550239 DOI: 10.1038/ki.1987.50]
- Garcia C, Mayaudon H, Bordier L, Le Berre JP, Dupuy O, Bauduceau B. Role of nocturnal blood pressure in the onset of diabetic nephropathy. *Arch Mal Coeur Vaiss* 2007; **100**: 668-672 [PMID: 17928773]
- Benhamou PY, Halimi S, De Gaudemaris R, Boizel R, Pitiot M, Siche JP, Bachelot I, Mallion JM. Early disturbances of ambulatory blood pressure load in normotensive type I diabetic patients with microalbuminuria. *Diabetes Care* 1992; **15**: 1614-1619 [PMID: 1468293 DOI: 10.2337/diacare.15.11.1614]
- Garcia C, Mayaudon H, Bordier L, Berre JL, Dupuy O, Bauduceau B. Responsabilité de la pression artérielle nocturne dans l'apparition d'une néphropathie chez les diabétiques. Available from: URL: <http://www.em-consulte.com/en/article/130277>
- Brage S, Ekelund U, Brage N, Hennings MA, Froberg K, Franks PW, Wareham NJ. Hierarchy of individual calibration levels for heart rate and accelerometry to measure physical activity. *J Appl Physiol* (1985) 2007; **103**: 682-692 [PMID: 17463305]
- Darcen S, Goksen D, Mir S, Serdaroglu E, Buyukinan M, Coker M, Berdeli A, Köse T, Cura A. Alterations of blood pressure in type 1 diabetic children and adolescents. *Pediatr Nephrol* 2006; **21**: 672-676 [PMID: 16568306 DOI: 10.1152/japphysiol.00092.2006]
- Ninomiya T, Perkovic V, de Galan BE, Zoungas S, Pillai A, Jardine M, Patel A, Cass A, Neal B, Poulter N, Mogensen CE, Cooper M, Marre M, Williams B, Hamet P, Mancia G, Woodward M, Macmahon S, Chalmers J. Albuminuria and kidney function independently predict cardiovascular and renal outcomes in diabetes. *J Am Soc Nephrol* 2009; **20**: 1813-1821 [PMID: 19443635 DOI: 10.1681/ASN.2008121270]
- Poulsen PL, Ebbenhøj E, Nosadini R, Fioretto P, Deferrari G, Crepaldi G, Mogensen CE. Early ACE-i intervention in microalbuminuric patients with type 1 diabetes: effects on albumin excretion, 24 h ambulatory blood pressure, and renal function. *Diabetes Metab* 2001; **27**: 123-128 [PMID: 11353877]
- Poulsen PL, Ebbenhøj E, Mogensen CE. Lisinopril reduces albuminuria during exercise in low grade microalbuminuric type 1 diabetic patients: a double blind randomized study. *J Intern Med* 2001; **249**: 433-440 [PMID: 11350567 DOI: 10.1046/j.1365-2796.2001.00821.x]
- Poulsen PL, Ebbenhøj E, Arildsen H, Knudsen ST, Hansen KW, Mølgaard H, Mogensen CE. Increased QTc dispersion is related to blunted circadian blood pressure variation in normoalbuminuric type 1 diabetic patients. *Diabetes* 2001; **50**: 837-842 [PMID: 11289050 DOI: 10.2337/diabetes.50.4.837]
- Hansen KW, Poulsen PL, Mogensen CE. Ambulatory blood pressure and abnormal albuminuria in type 1 diabetic patients. *Kidney Int Suppl* 1994; **45**: S134-S140 [PMID: 8158882]
- Cohen CN, Albanesi FM, Gonçalves MF, Gomes MB. Ambulatory blood pressure monitoring and microalbuminuria in normotensive subjects with insulin-dependent diabetes mellitus. *Arq Bras Cardiol* 2000; **75**: 195-204 [PMID: 11018805 DOI: 10.1590/S0066-782X2000000900001]
- Sochett EB, Poon I, Balfé W, Daneman D. Ambulatory blood pressure monitoring in insulin-dependent diabetes mellitus adolescents with

- and without microalbuminuria. *J Diabetes Complications* 1998; **12**: 18-23 [PMID: 9442810 DOI: 10.1016/S1056-8727(97)00050-0]
- 21 **Pańkowska E**, Golicka D. Changes in blood pressure and methods of blood pressure monitoring in patients with type-1 diabetes. *Endokrynol Diabetol Chor Przemiany Materii Wieku Rozw* 2005; **11**: 33-37 [PMID: 15850536]
  - 22 **Hansen KW**, Poulsen PL, Mogensen CE. 24-h blood pressure recordings in type I diabetic patients. *J Diabetes Complications* 1995; **9**: 237-240 [PMID: 8573736 DOI: 10.1016/1056-8727(95)80011-3]
  - 23 **Shalaby NM**, Shalaby NM. Study of ambulatory blood pressure in diabetic children: prediction of early renal insult. *Ther Clin Risk Manag* 2015; **11**: 1531-1537 [PMID: 26491340 DOI: 10.2147/TCRM.S87751]
  - 24 **Farmer CK**, Goldsmith DJ, Quin JD, Dallyn P, Cox J, Kingswood JC, Sharpstone P. Progression of diabetic nephropathy--is diurnal blood pressure rhythm as important as absolute blood pressure level? *Nephrol Dial Transplant* 1998; **13**: 635-639 [PMID: 9550639 DOI: 10.1093/ndt/13.3.635]
  - 25 **O'Brien SF**, Watts GF, Powrie JK, Shaw KM. Exercise testing as a long-term predictor of the development of microalbuminuria in normoalbuminuric IDDM patients. *Diabetes Care* 1995; **18**: 1602-1605 [PMID: 8722059 DOI: 10.2337/diacare.18.12.1602]
  - 26 **Marshall SL**, Edidin DV, Arena VC, Becker DJ, Bunker CH, Gishoma C, Gishoma F, LaPorte RE, Kaberuka V, Ogle G, Sibomana L, Orchard TJ. Glucose control in Rwandan youth with type 1 diabetes following establishment of systematic, HbA1c based, care and education. *Diabetes Res Clin Pract* 2015; **107**: 113-122 [PMID: 25458328 DOI: 10.1016/j.diabres.2014.09.045]
  - 27 **Gonzalez Suarez ML**, Thomas DB, Barisoni L, Forni A. Diabetic nephropathy: Is it time yet for routine kidney biopsy? *World J Diabetes* 2013; **4**: 245-255 [PMID: 24379914 DOI: 10.4239/wjd.v4.i6.245]
  - 28 **American Diabetes Association**. Standards of medical care in diabetes--2014. *Diabetes Care* 2014; **37** Suppl 1: S14-S80 [PMID: 24357209 DOI: 10.2337/dc14-S014]

**P- Reviewer:** Barzilay JI, Li ML, van Beers CAJ **S- Editor:** Qiu S  
**L- Editor:** A **E- Editor:** Wu HL





## Fuzzy expert system for diagnosing diabetic neuropathy

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Supported by The Iran University of Medical Sciences, No. 541.

Conflict-of-interest statement: There are no conflicts of interest arising from this work.

Data sharing statement: No further data are available.

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Received: July 27, 2016

Peer-review started: July 29, 2016

First decision: September 8, 2016

Revised: October 12, 2016

Accepted: December 1, 2016

Article in press: December 2, 2016

Published online: February 15, 2017

### Abstract

#### AIM

To design a fuzzy expert system to help detect and diagnose the severity of diabetic neuropathy.

#### METHODS

The research was completed in 2014 and consisted of two main phases. In the first phase, the diagnostic parameters were determined based on the literature review and by investigating specialists' perspectives ( $n = 8$ ). In the second phase, 244 medical records related to the patients who were visited in an endocrinology and metabolism research centre during the first six months of 2014 and were primarily diagnosed with diabetic neuropathy, were used to test the sensitivity, specificity, and accuracy of the fuzzy expert system.

#### RESULTS

The final diagnostic parameters included the duration of diabetes, the score of a symptom examination based on the Michigan questionnaire, the score of a sign examination based on the Michigan questionnaire, the glycolysis haemoglobin level, fasting blood sugar, blood creatinine, and albuminuria. The output variable was the severity of diabetic neuropathy which was shown as a number between zero and 10, had been divided into four categories: absence of the disease, (the degree of severity) mild, moderate, and severe. The interface of the system was designed by ASP.Net (Active Server Pages Network Enabled Technology) and the system function was tested in terms of sensitivity (true positive rate) (89%), specificity (true negative rate) (98%), and accuracy (a proportion of true results, both positive and negative) (93%).

#### CONCLUSION

The system designed in this study can help specialists

and general practitioners to diagnose the disease more quickly to improve the quality of care for patients.

**Key words:** Expert systems; Fuzzy logic; Artificial intelligence; Diabetes mellitus; Diabetes complications; Diabetic neuropathies

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**Core tip:** In this study, an expert system was designed for diagnosing diabetic neuropathy. This system can help specialists to diagnose the disease more quickly by using the most common diagnostic parameters. Even general practitioners can use this system in remote areas to improve the quality of care for patients with diabetes. With it, patients will no longer need to undertake complex procedures, and the care plan can be applied at the right time.

Rahmani Katigari M, Ayatollahi H, Malek M, Kamkar Haghighi M. Fuzzy expert system for diagnosing diabetic neuropathy. *World J Diabetes* 2017; 8(2): 80-88 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v8/i2/80.htm> DOI: <http://dx.doi.org/10.4239/wjd.v8.i2.80>

## INTRODUCTION

One of the biggest challenges currently experienced by healthcare organizations is the increasing burden of chronic diseases posing serious threats to public health in developing countries<sup>[1]</sup>. Diabetes is one of the world's most common and costly chronic diseases, and the number of patients suffering from diabetes has been showing an increasing trend in many countries<sup>[2]</sup>. This can be attributed to population growth, aging, urbanization, prevalence of obesity, and a sedentary lifestyle<sup>[2,3]</sup>. Long-term complications of diabetes develop gradually and might be disabling or life-threatening - for example, vascular and tissue injuries caused by the progression of diabetes can lead to serious complications, such as retinopathy, nephropathy, cardiovascular disease, cerebrovascular disease, peripheral vascular disease, metabolic disease, and diabetic foot ulcer<sup>[4,5]</sup>. However, the most common complication of diabetes is impairment of the peripheral neural system, which is known as diabetic neuropathy and a major problem with different signs and symptoms. Compared with other diabetes complications, it is one of the first reasons for hospitalizing patients with diabetes<sup>[6]</sup>. The severity of pain, decreased or lack of sensation, increased risk of foot ulceration, and amputation are the consequences of diabetic neuropathy<sup>[7]</sup>.

Diabetic peripheral neuropathy is usually seen in more than 10% of patients with type II diabetes. Early diagnosis and treatment is the first step to reduce the incidence of foot ulcers and amputations<sup>[8]</sup>. The main

cost of this disease is related to organ amputation. The risk of lower extremity amputation in patients is significantly high in case of this disease. Nevertheless, almost 85% of amputations are preventable by early detection of the disease, early intervention, good control of diabetes, and patient education<sup>[9]</sup>. Moreover, several studies show that neuropathy may negatively affect the quality of life for patients with diabetes<sup>[10,11]</sup>.

Owing to the high prevalence of neuropathy among patients with diabetes, it is necessary to conduct annual screening and further evaluation as well as to devise a plan for managing the disease. However, one of the major problems associated with the diagnosis of diabetic neuropathy is the lack of a reliable clinical scale for grading the severity of the disease<sup>[12]</sup>. A variety of methods are used to detect peripheral neuropathy. These include the nerve conduction velocity test, the vibration perception threshold, the monofilament test, the clinical neuropathy examination, the Toronto clinical scoring system, and the Michigan neuropathy screening instrument (MNSI)<sup>[13]</sup>. Other than clinical examination, laboratory tests, such as haemoglobin A1c level, fasting blood sugar, and oral glucose tolerance test, along with risk factors like age, sex, renal disease, and smoking need to be considered<sup>[14]</sup>.

It is notable that the boundary between illness and health is not clear in diabetic neuropathy, and it is difficult to express clinical diagnosis as the lack of or the existence of the disease. Since the disease develops on a continuous basis, two-valued logic cannot be used to express this continuity anymore<sup>[6]</sup>. Therefore, new methods for diagnosing the disease have been considered<sup>[15]</sup>. Among these methods, special attention has been paid to the development of information technology applications, decision support systems, and fuzzy expert systems<sup>[16,17]</sup>. The fuzzy expert system is a new version of expert systems that uses fuzzy logic for data processing. In a fuzzy expert system, the inference is conducted by a set of membership functions and fuzzy rules rather than by the rules of two-valued logic<sup>[18]</sup>. The Fuzzy expert systems are used to describe uncertain phenomena because real-world phenomena are much more complex than an exact and absolute description<sup>[19,20]</sup>. The ability to implement human science through specific linguistic concepts and fuzzy rules, non-linearity, adaptability of these systems, and the level of accuracy are the most important features of these systems<sup>[21]</sup>. Although fuzzy expert systems have been designed for different purposes in the healthcare setting, only a few studies have focused on the use of these systems with regard to the diagnosis of diabetic neuropathy<sup>[22]</sup>.

## MATERIALS AND METHODS

### Objective

To design a fuzzy expert system to categorize the severity of diabetic neuropathy based on clinical exa-

minations and results of laboratory tests.

### Setting, design, and sample size

This study was completed in 2014. The study consisted of two main phases. In the first phase, the parameters required for the diagnosis of diabetic neuropathy were determined on the basis of the literature review<sup>[23,24]</sup>. These parameters formed a questionnaire to investigate specialists' views about the importance of each of them. In the second phase, the system was tested by using real data. In the first phase, eight endocrinologists participated in the study. Owing to the limited number of specialists, no sampling method was applied in this phase. In the second phase, 244 medical records were identified from a database located in an endocrinology and metabolism research centre. These records were related to those patients who visited the centre during the first six months of 2014 and who were primarily diagnosed with diabetic neuropathy.

### Methods for data collection and distribution

The questionnaire was distributed among the specialists by one of the researchers (MRK), and their views on the importance of the diagnostic parameters were investigated. In second phase, a form was used to extract the required data from the medical records.

### Development of the questionnaire

As noted before, the questionnaire was designed based on the literature review<sup>[23,24]</sup>. It comprised two parts: The first part included the specialists' demographic information, such as age, gender, and work experience; the second part was designed based on a five-point Likert scale (5 = very important, 4 = important, 3 = relatively important, 2 = less important, 1 = unimportant) and consisted of 15 questions to identify the degree of importance of each diagnostic parameter. The face and content validity of the questionnaire was approved by experts in the field of endocrinology. Its reliability was confirmed by using the test-retest method ( $\alpha = 0.9$ ).

### Statistical analysis

A data analysis was performed by using SPSS (version 20.0) software, and parameters with a mean score of less than three were excluded to facilitate the process of writing fuzzy rules. To test the system, the sensitivity, specificity, and accuracy of the fuzzy expert system were measured and compared with the final diagnosis recorded in the database. Cohen's kappa coefficient and the receiver operating characteristic (ROC) curve were used to report data.

### Participants and recruitment

Before conducting the research, the approval of an institutional review board was obtained. In the first phase, the target population comprised endocrinologists working in an endocrinology and metabolism research

centre. They were contacted by one of the researchers (MRK) and the research facilitator (MM), and were invited to take part in the study. Their participation in the research was completely voluntary. Regarding the medical records, patient identities were excluded and only the required data was extracted so that it can be used in the process of evaluation.

## RESULTS

### Participants

As noted before, the first part of the questionnaire included the participants' demographic information. According to the results, most of the participants were men ( $n = 5$ , 62.5%) aged between 30-50 years. The highest frequency ( $n = 3$ , 37.5%) was related to the age group of 46-50 years and the specialists with more than 16 years of work experience.

### Diagnostic parameters for diagnosing diabetic neuropathy

The second part of the questionnaire was related to the diagnostic parameters required for diagnosing diabetic neuropathy. This part included the duration of diabetes, the symptom assessment based on MNSI, the sign examination based on MNSI, and the related laboratory tests. Table 1 presents the specialists' views in relation to the importance of the aforementioned diagnostic parameters.

As Table 1 shows, from the specialists' point of view, the most important diagnostic parameters were the duration of diabetes ( $4.88 \pm 0.35$ ), the glycolysis haemoglobin level ( $4.50 \pm 0.75$ ), and the score of the sign examination based on the Michigan questionnaire ( $4.38 \pm 0.51$ ). The lowest degree of importance ( $2.13 \pm 0.83$ ) was related to the amount of phosphorus in blood. After determining the diagnostic parameters of diabetic neuropathy, the semantic network of the expert system was drawn (Figure 1).

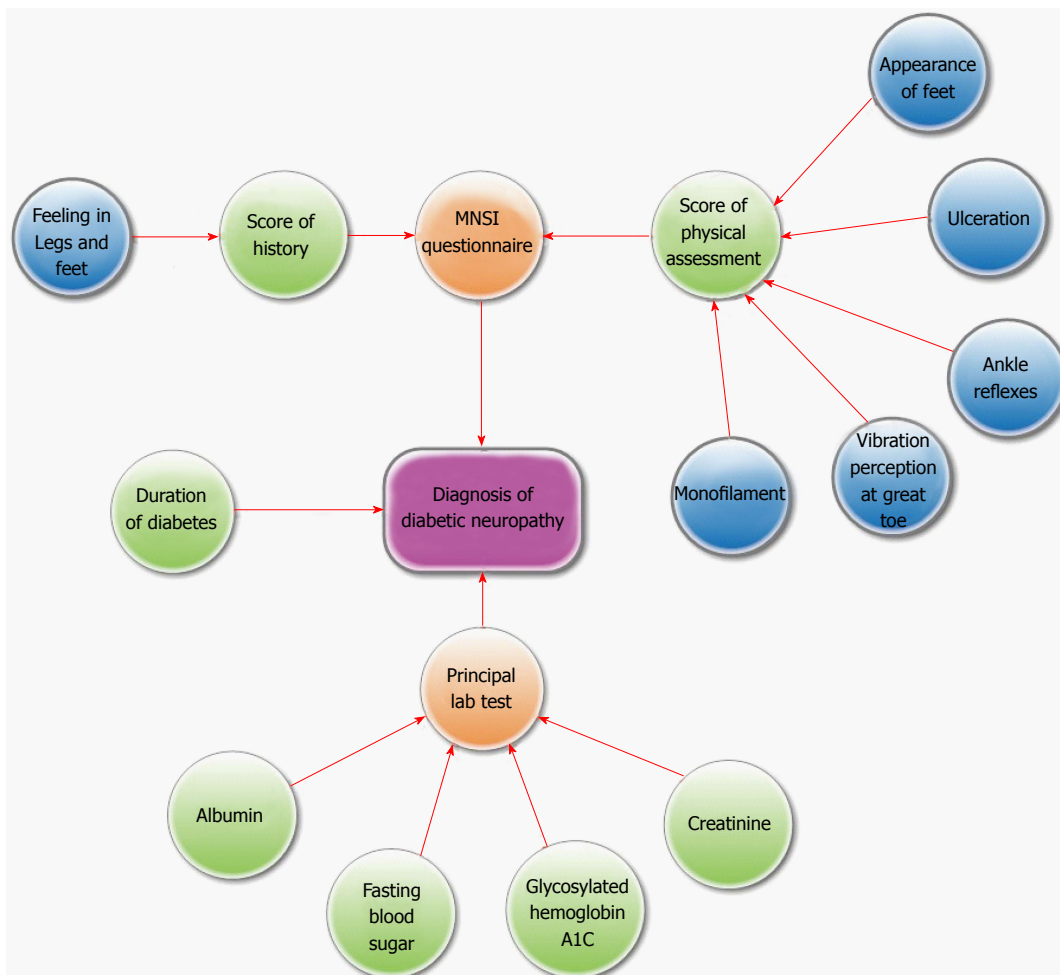
### Designing a fuzzy expert system

As can be seen in the above figure, the ultimate goal, namely diagnosing diabetic neuropathy, is shown in the centre, and the diagnostic parameters are in the leaf nodes. In order to design the fuzzy expert system, all input variables were fuzzified based on membership functions. The system had seven input variables: The duration of diabetes, the score of the symptom examination based on the Michigan questionnaire, the score of the sign examination based on the Michigan questionnaire, the glycolysis haemoglobin level, fasting blood sugar, blood creatinine, and albuminuria. The system also had one output variable, which was the severity of diabetic neuropathy. The rules of the expert system were written based on the semantic network, consulting a specialist, and giving the same weight to all rules. The inference engine of the system was designed by using the Mamdani inference method. Figure 2 provides

**Table 1** The degree of importance of the diagnostic parameters for diagnosing diabetic neuropathy from the specialists' perspectives

Degree of importance	Unimportant (1)	Less important (2)	Relatively important (3)	Important (4)	Very important (5)	Mean $\pm$ SD
Duration of diabetes	0	0	0	1 (12.5%)	7 (87.5%)	4.88 $\pm$ 0.35
Symptom assessment based on MNSI	0	0	1 (12.5%)	5 (62.5%)	2 (25%)	4.13 $\pm$ 0.64
Sign examination based on MNSI	0	0	0	5 (62.5%)	3 (37.5%)	4.38 $\pm$ 0.51
HbA1c	0	0	1 (12.5%)	2 (25%)	5 (62.5%)	4.50 $\pm$ 0.75
CBC	1 (12.5%)	3 (37.5%)	4 (50%)	0	0	2.38 $\pm$ 0.74
FBS	0	0	0	6 (75%)	2 (25%)	4.25 $\pm$ 0.46
ESR	1 (12.5%)	3 (37.5%)	3 (37.5%)	1 (12.5%)	0	2.52 $\pm$ 0.92
Oral GTT	1 (12.5%)	4 (50%)	1 (12.5%)	2 (25%)	0	2.50 $\pm$ 1.06
Albuminuria	0	1 (12.5%)	1 (12.5%)	4 (50%)	2 (25%)	3.88 $\pm$ 0.99
TSH	2 (25%)	1 (12.5%)	3 (37.5%)	2 (25%)	0	2.63 $\pm$ 1.18
B12 Vitamin	2 (25%)	1 (12.5%)	1 (12.5%)	4 (50%)	0	2.88 $\pm$ 1.35
BUN	1 (12.5%)	3 (37.5%)	3 (37.5%)	1 (12.5%)	0	2.38 $\pm$ 0.91
BCr	0	1 (12.5%)	2 (25%)	5 (62.5%)	0	3.50 $\pm$ 0.75
Calcium	2 (25%)	1 (12.5%)	4 (50%)	1 (12.5%)	0	2.50 $\pm$ 1.06
Phosphorus	2 (25%)	3 (37.5%)	3 (37.5%)	0	0	2.13 $\pm$ 0.83

BCr: Blood Creatinine; BUN: Blood urea nitrogen; TSH: Thyroid-stimulating hormone; GTT: Glucose tolerance test; ESR: Erythrocyte sedimentation rate; MNSI: Michigan Neuropathy Screening Instrument; HbA1c: Hemoglobin A1c; CBC: Complete blood count; FBS: Fasting blood sugar.


**Figure 1** The semantic network of the expert system. MNSI: Michigan Neuropathy Screening Instrument.

an overview of the fuzzy inference architecture of the system.

Finally, the graphical user interface of the expert system was designed by using Active Server Page.

Network Enabled Technology (ASP.NET). It is an open-source server-side web application framework designed for web development to produce dynamic web pages (Figure 3). The input variables, such as the duration



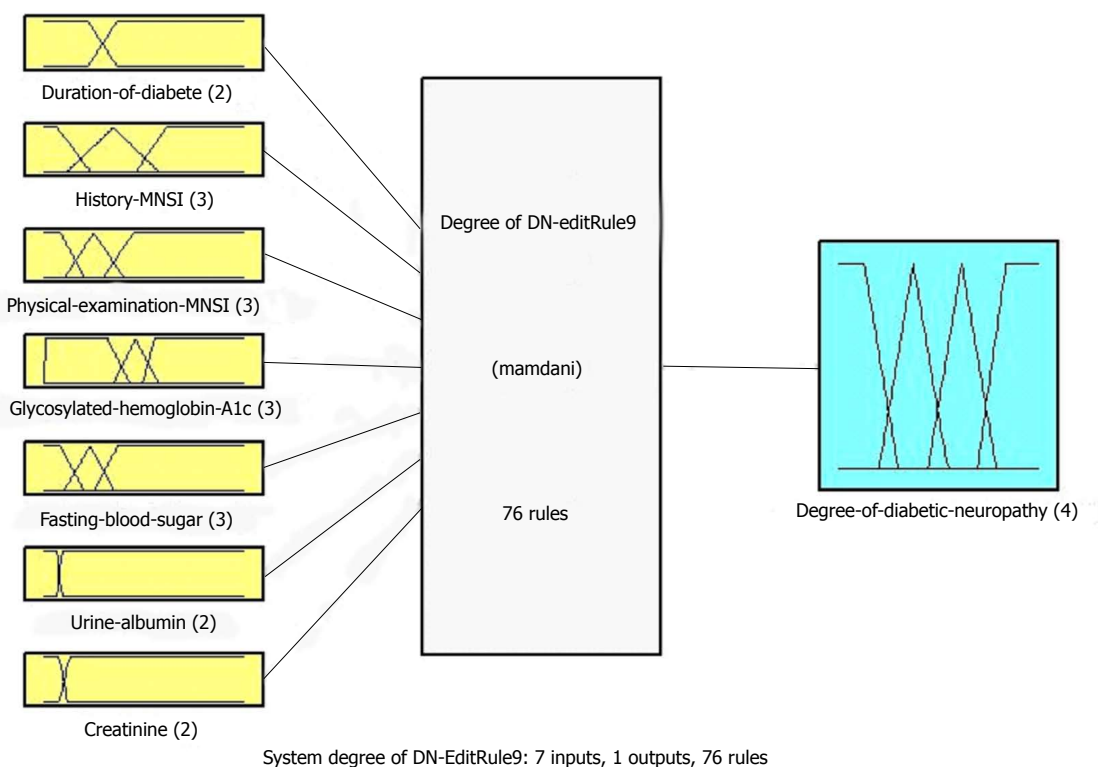


Figure 2 An overview of the fuzzy inference architecture of the system.

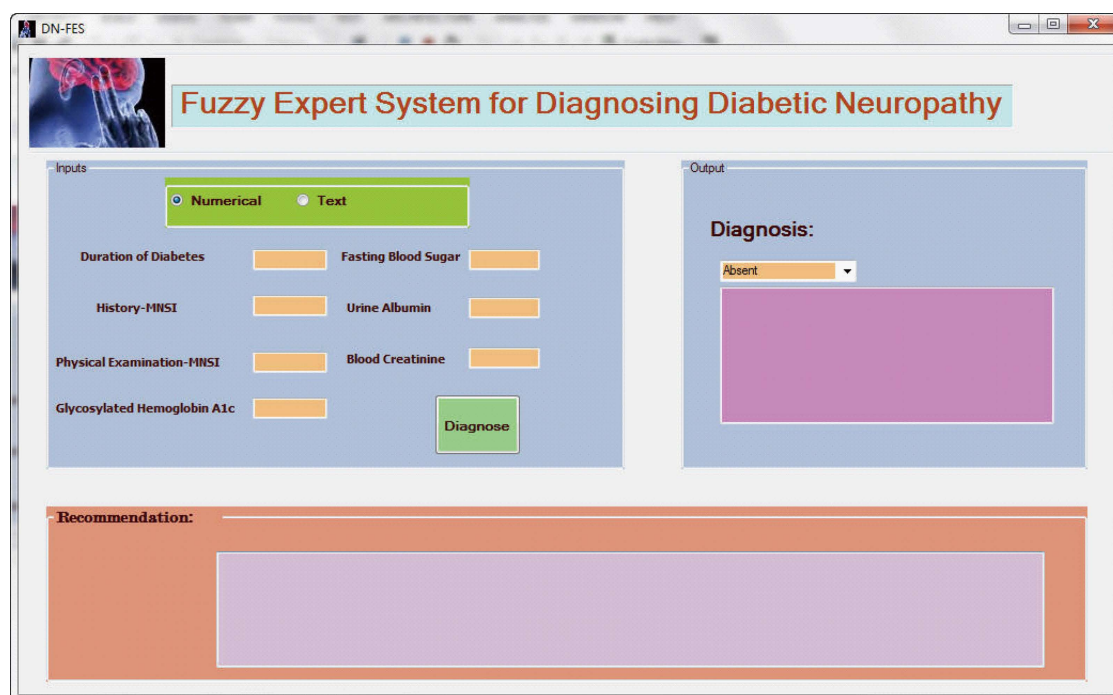
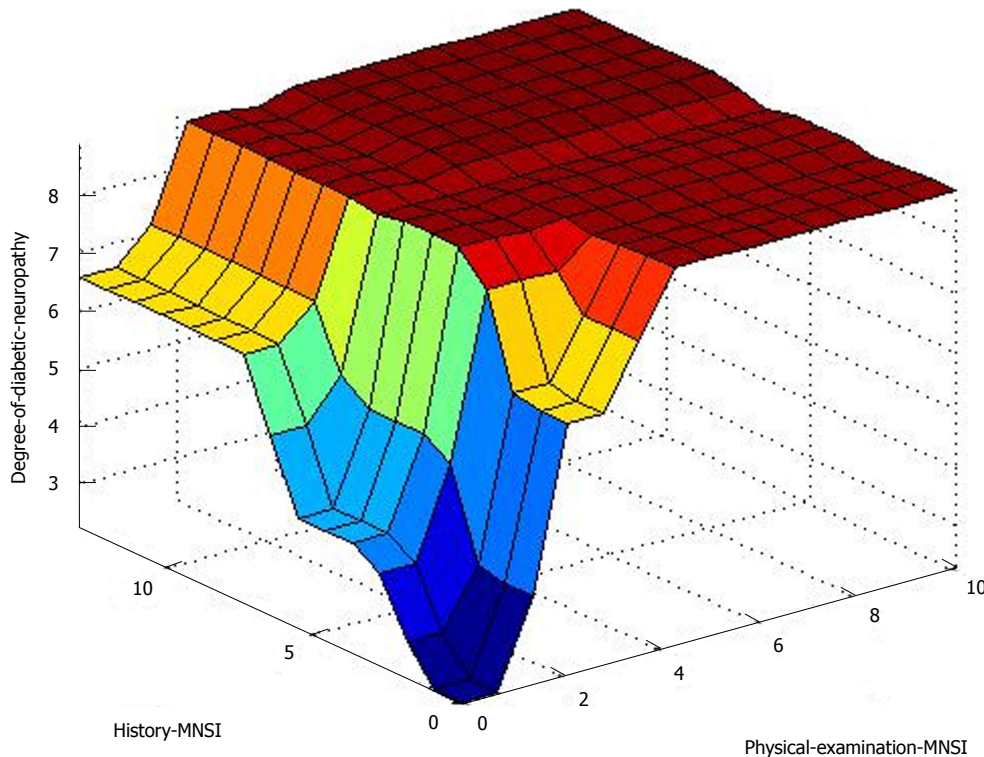


Figure 3 The graphical user interface of the fuzzy expert system.

of diabetes, the results of laboratory tests, and scores obtained from the Michigan questionnaire, could be entered into the system manually either in the textual or in the numerical format based on the user's choice. The output variable, namely the severity of the disease, which was shown as a number between zero and 10,

had been divided into four categories: absence of the disease, (the degree of severity) mild, moderate, and severe. Figure 4 shows the risk of diabetic neuropathy based on the scores obtained from the Michigan questionnaire.

According to Figure 4, by increasing the scores



**Figure 4** The risk of diabetic neuropathy based on the scores of the Michigan Neuropathy Screening Instrument questionnaire. MNSI: Michigan Neuropathy Screening Instrument.

obtained from the Michigan questionnaire, the severity of diabetic neuropathy will increase accordingly.

### System function evaluation

The system was tested by using real data. In total, the records of 244 patients with diabetic neuropathy were identified. However, 31 records were excluded due to the incompleteness of clinical data. The remaining records ( $n = 213$ ) included 118 patients who were diagnosed with diabetic neuropathy, while diagnosis was ruled out for the rest ( $n = 95$ ). The system function was tested in terms of sensitivity (true positive rate), specificity (true negative rate), and accuracy (proportion of the true results, both positive and negative), which were 89%, 98%, and 93%, respectively.

Finally, the system's output was compared with the final diagnoses made by the specialists and recorded in the patients' records. These diagnoses were made by using the nerve conduction velocity test, the vibration perception threshold, the monofilament test, and the clinical neuropathy examination. The comparison was conducted by using the Kappa coefficient and the K value was 0.6. According to Landis and Koch, a Kappa value between 0.4 and 0.75 shows a fair to good agreement<sup>[25]</sup>. Therefore, the system designed in this study showed a fair to good level of similarity between the system's function and the specialists' diagnoses. The ROC curve presents the results of testing the system (Figure 5).

As can be seen in the above figure, the ROC curve is ideal. It is close to the high point of the square that

represents an appropriate function of the system.

## DISCUSSION

As mentioned before, one of the most common long-term complications of diabetes mellitus is diabetic neuropathy. In order to control this complication, it is important to diagnose it both accurately and timely<sup>[10]</sup>. Although there are a variety of methods to detect the disease, it is difficult to diagnose it at the very early stage<sup>[13]</sup>. Therefore, the use of IT applications, such as fuzzy expert systems, is suggested.

In the present study, seven diagnostic parameters—the duration of diabetes, the symptom assessment, the sign examination based on the MNSI, the glycolysis haemoglobin level, fasting blood sugar, blood creatinine, and albuminuria—were considered as input variables, and the severity of diabetic neuropathy was considered as an output variable. These variables were selected based on the specialists' perspectives and the literature review. Similarly, the knowledge and experience of four experts in the field of diabetic neuropathy was investigated in the study conducted by Picon *et al.*<sup>[22]</sup> to determine the diagnostic parameters and to design a knowledge-based system. In their research, four inputs variables included symptom, the sign assessment based on the Michigan questionnaire, the glycolysis haemoglobin level, and the duration of diabetes. The output of the system classified the severity of diabetic neuropathy in three categories: Mild, moderate, and severe. In contrast with the study of Picon *et al.*<sup>[22]</sup> the number of input variables increased

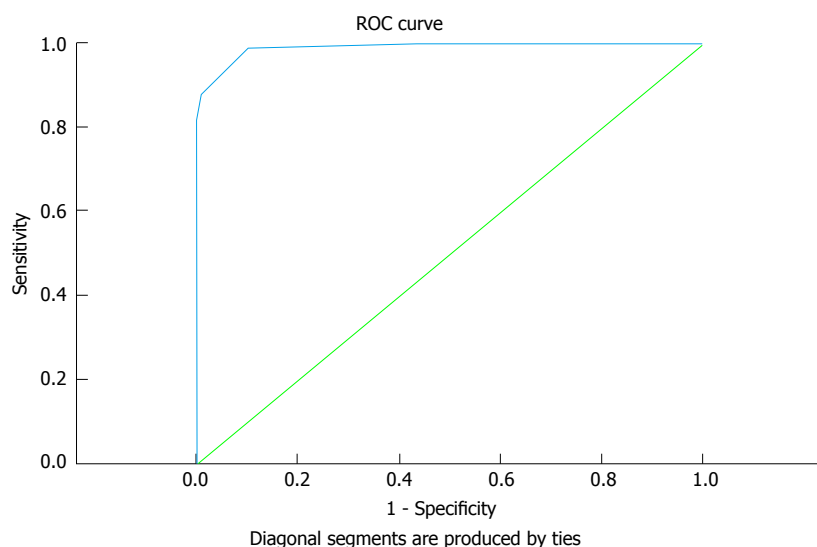


Figure 5 The receiver operating characteristic curve.

in the current research and laboratory test results were included to improve the accuracy of diagnosis. Similarly, Neshat *et al.*<sup>[26]</sup> study considered six input variables and one output variable to diagnose liver disorders. To diagnose heart ailments, Adeli *et al.*<sup>[27]</sup> used 12 input variables and considered the diagnosis of heart diseases as the output variable.

In the present study, values between zero and 10 were considered for the output variable, which was the severity of diabetic neuropathy. An increase in the value of output variable showed the level of severity for diabetic neuropathy.

In the current study, the fuzzy sets and membership functions for each of the seven input variables and the output variable were finalized after consulting a specialist. This approach can help eliminate the rules that could be covered by other rules, and finally, 76 rules were used to design the system. Similarly, DoostHoseini *et al.*<sup>[28]</sup> consulted doctors to reduce the number of rules to an appropriate number. In another study, Zolnoori *et al.*<sup>[29]</sup> developed a fuzzy expert system for diagnosing asthma. Given that the patients' records were incomplete, an indirect approach was used to develop the system's knowledge base. In this approach, the researchers reviewed books and scientific papers, and also conducted structured and unstructured interviews with doctors and patients. Having analysed the data, the most important variables useful for diagnosing asthma were identified.

In the present study, the system interface was designed by using ASP.NET rather than matrix laboratory (MATLAB). In fact, web-based applications have more flexibility and can be used by multiple users at the same time. Ease of use is another feature of these systems, which, in turn, can increase the work efficiency.

In this study, the output of the system was divided into four categories: The absence of the disease,

mild, moderate, and severe. In contrast, Picon *et al.*<sup>[22]</sup> classified the severity of neuropathy into three categories: Mild, moderate, and severe. Moreover, the specificity and sensitivity of the system were not reported in their study. In the current study, the specificity of the system was 98%, which shows a high level of system performance. Also, there was a relatively good agreement between the system's function and the diagnoses recorded by the specialists. Although other methods of diagnosis were not considered in the current study, the specificity and sensitivity of the system highly suggested that such a system could help physicians to diagnose the disease more quickly by using parameters like results of laboratory tests.

In the current study, the main aim was to develop an expert system for diagnosing diabetic neuropathy. Therefore, the clinical effectiveness of the system was not evaluated due to resource restrictions. Conducting evaluation studies after implementing the system in the actual healthcare setting would help determine the impact of the system on the health status of patients.

In conclusion, an expert system was designed for diagnosing diabetic neuropathy in this study. As diabetic neuropathy is a chronic disease that may have serious consequences, early diagnosis of the disease is important to control it. The system designed in the current study could help specialists to diagnose the disease more quickly by using the most common diagnostic parameters. General practitioners can use such a system in remote areas to improve the quality of care for patients with diabetes. With it, the disease can be diagnosed more easily and quickly. There is no need to undertake complex procedures, and the care plan can be applied at the right time. Further research is suggested to increase the number of variables to improve the accuracy, sensitivity, and specificity of the system. Moreover, the feasibility of using this method in daily clinical practice and its impact on the efficiency and

cost-effectiveness compared to those of other methods need to be investigated in future studies.

## COMMENTS

### Background

One of the major problems associated with the diagnosis of diabetic neuropathy is the lack of reliable clinical scale for grading the severity of the disease. A variety of methods, such as the nerve conduction velocity test, the vibration perception threshold, and the monofilament test, are used to detect the peripheral neuropathy. In addition to clinical examination, laboratory tests and risk factors of the disease such as age, sex, renal disease, and smoking need to be considered.

### Research frontiers

Since the disease usually develops on a continuous basis, two-valued logic cannot be used to express this continuity any more. Therefore, new methods for diagnosing the disease have been considered. Among these methods, the development of information technology applications, decision support systems, and fuzzy expert systems have received special attention.

### Innovations and breakthroughs

In order to diagnose diabetic neuropathy, clinical examinations as well as results of laboratory tests like the haemoglobin A1c level, fasting blood sugar, and the oral glucose tolerance test should be considered. In this study, information technology was used to design a fuzzy expert system to diagnose the severity of diabetic neuropathy based on clinical examinations and laboratory tests.

### Applications

The system designed in the current study can help specialists to diagnose the disease more quickly by using the most common diagnostic parameters. General practitioners, too, can use it in remote areas to improve the quality of care for patients with diabetes. With it, the disease can be diagnosed more easily and quickly. There is no need to undertake complex procedures, and the care plan can be applied at the right time.

### Terminology

The fuzzy expert system is a new version of expert systems that uses fuzzy logic for data processing. A fuzzy expert system is used to describe uncertain phenomena because the real-world phenomena are much more complex than an exact and absolute description. The most common complication of diabetes is impairment of the peripheral neural system, which is known as diabetic neuropathy.

### Peer-review

This is interesting and important paper for diagnosis of diabetic complications. The paper is well-written and focused.

## REFERENCES

- Nolte E, McKee M. Caring for people with chronic conditions: A health system perspective. England: Open University Press, 2008: 1-245
- Seino Y, Nanjo K, Tajima N, Kadowaki T, Kashiwagi A, Araki E, Ito C, Inagaki N, Iwamoto Y, Kasuga M, Hanafusa T, Haneda M, Ueki K. Report of the committee on the classification and diagnostic criteria of diabetes mellitus. *J Diabetes Investig* 2010; **1**: 212-228 [PMID: 24843435 DOI: 10.1111/j.2040-1124.2010.00074.x]
- Mehri A, Hasani-Ranjbar S, Larijani B, Abdollahi M. A systematic review of efficacy and safety of *Urtica dioica* in the treatment of diabetes. *Int J Pharmacol* 2011; **7**: 161-170 [DOI: 10.3923/ijp.2011.161.170]
- Simon GE, Katon WJ, Lin EH, Ludman E, VonKorff M, Ciechanowski P, Young BA. Diabetes complications and depression as predictors of health service costs. *Gen Hosp Psychiatry* 2005; **27**: 344-351 [PMID: 16168795 DOI: 10.1016/j.genhosppsych.2005.04.008]
- Svensson M, Eriksson JW, Dahlquist G. Early glycemic control, age at onset, and development of microvascular complications in childhood-onset type 1 diabetes: a population-based study in northern Sweden. *Diabetes Care* 2004; **27**: 955-962 [PMID: 15047655 DOI: 10.2337/diacare.27.4.955]
- Gordo A, Scuffham P, Shearer A, Oglesby A, Tobian JA. The health care costs of diabetic peripheral neuropathy in the US. *Diabetes Care* 2003; **26**: 1790-1795 [PMID: 12766111 DOI: 10.2337/diacare.26.6.1790]
- Larejani B, Zahedi F. Epidemiology of diabetes mellitus in Iran. *Iranian Journal of Diabetes and Metabolism* 2001; **1**: 1-8
- Liu F, Bao Y, Hu R, Zhang X, Li H, Zhu D, Li Y, Yan L, Li Y, Lu J, Li Q, Zhao Z, Ji Q, Jia W. Screening and prevalence of peripheral neuropathy in type 2 diabetic outpatients: a randomized multicentre survey in 12 city hospitals of China. *Diabetes Metab Res Rev* 2010; **26**: 481-489 [PMID: 20661939 DOI: 10.1002/dmrr.1107]
- Cheer K, Shearman C, Jude EB. Managing complications of the diabetic foot. *BMJ* 2009; **339**: b4905 [PMID: 19955124 DOI: 10.1136/bmj.b4905]
- Vinik AI, Park TS, Stansberry KB, Pittenger GL. Diabetic neuropathies. *Diabetologia* 2000; **43**: 957-973 [PMID: 10990072 DOI: 10.1007/s001250051477]
- Said G. Diabetic neuropathy--a review. *Nat Clin Pract Neurol* 2007; **3**: 331-340 [PMID: 17549059 DOI: 10.1038/ncpneuro0504]
- Bostani A, Homayounfar H. The Relationship between NCS Findings and Toronto Clinical Scoring System of Neuropathy in Diabetic Polyneuropathy. *Journal of Kermanshah University of Medical Sciences* 2006; **88**: 2358-2367
- Rahman M, Griffin SJ, Rathmann W, Wareham NJ. How should peripheral neuropathy be assessed in people with diabetes in primary care? A population-based comparison of four measures. *Diabet Med* 2003; **20**: 368-374 [PMID: 12752485 DOI: 10.1046/j.1464-5491.2003.00931.x]
- Ziegler D, Rathmann W, Dickhaus T, Meisinger C, Mielck A. Neuropathic pain in diabetes, prediabetes and normal glucose tolerance: the MONICA/KORA Augsburg Surveys S2 and S3. *Pain Med* 2009; **10**: 393-400 [PMID: 19207236 DOI: 10.1111/j.1526-4637.2008.00555.x]
- Toloie Ashlaqi A, Mohsen Taheri S. Designing an expert system for suggesting the blood cancer treatment. *Journal of Health Administration* 2010; **13**: 41-50
- Wei HY, Lu CS, Lin TH. Exploring the P2 and P3 ligand binding features for hepatitis C virus NS3 protease using some 3D QSAR techniques. *J Mol Graph Model* 2008; **26**: 1131-1144 [PMID: 18024210 DOI: 10.1016/j.jmgm.2007.10.005]
- Riazi H, Larijani B, Langarizadeh M, Shahmoradi L. Managing diabetes mellitus using information technology: a systematic review. *J Diabetes Metab Disord* 2015; **14**: 49 [DOI: 10.1186/s40200-015-0174-x]
- Riahi-Madvar H, Ayyoubzadeh SA, Khadangi E, Ebadzadeh MM. An expert system for predicting longitudinal dispersion coefficient in natural streams by using ANFIS. *Expert Syst Appl* 2009; **36**: 8589-8596 [DOI: 10.1016/j.eswa.2008.10.043]
- Keshwani DR, Jones DD, Meyer GE, Brand RM. Rule-based Mamdani-type fuzzy modeling of skin permeability. *Appl Soft Comput* 2008; **8**: 285-294 [DOI: 10.1016/j.asoc.2007.01.007]
- Saritas I, Ozkan IA, Allahverdi N, Argindogan M. Determination of the drug dose by fuzzy expert system in treatment of chronic intestine inflammation. *J Intell Manuf* 2009; **20**: 169-176 [DOI: 10.1007/s10845-008-0226-x]
- Sadoughi F, Sheikhtaheri A, Meidani Z, Shahmoradi L. Management information system (concepts, structure, development and evaluation). Tehran: Jafari, 2011
- Picon AP, Ortega NR, Watari R, Sartor C, Sacco IC. Classification of the severity of diabetic neuropathy: a new approach taking uncertainties into account using fuzzy logic. *Clinics (Sao Paulo)* 2012; **67**: 151-156 [PMID: 22358240 DOI: 10.6061/clinics/2012(02)10]
- Crawford F, Anandan C, Chappell FM, Murray GD, Price JF, Sheikh A, Simpson CR, Maxwell M, Stansby GP, Young MJ,



- Abbott CA, Boulton AJ, Boyko EJ, Kastenbauer T, Leese GP, Monami M, Monteiro-Soares M, Rith-Najarian SJ, Veves A, Coates N, Jeffcoate WJ, Leech N, Fahey T, Tierney J. Protocol for a systematic review and individual patient data meta-analysis of prognostic factors of foot ulceration in people with diabetes: the international research collaboration for the prediction of diabetic foot ulcerations (PODUS). *BMC Med Res Methodol* 2013; **13**: 22 [PMID: 23414550 DOI: 10.1186/1471-2288-13-22]
- 24 **Boulton AJ**, Vinik AI, Arezzo JC, Bril V, Feldman EL, Freeman R, Malik RA, Maser RE, Sosenko JM, Ziegler D. Diabetic neuropathies: a statement by the American Diabetes Association. *Diabetes Care* 2005; **28**: 956-962 [PMID: 15793206 DOI: 10.2337/diacare.28.4.956]
- 25 **Landis JR**, Koch GG. A review of statistical methods in the analysis of data arising from observer reliability studies (Part II). *Stat Neerl* 1975; **29**: 151-161 [DOI: 10.1111/j.1467-9574.1975.tb00259.x]
- 26 **Neshat M**, Yaghobi M, Naghibi M, Esmaelzadeh A. Fuzzy Expert System Design for Diagnosis of liver disorders. Knowledge Acquisition and Modeling; International Symposium on Azad University of Mashhad Iran: IEEE, 2008: 252-256 [DOI: 10.1109/kam.2008.43]
- 27 **Adeli A**, Neshat M. A fuzzy expert system for heart disease diagnosis. Proceedings of international multi conference of engineers and computer scientists. Hong Kong: Citeseer, 2010
- 28 **DoostHoseini E**, Hassanpour-ezatti M, Navidi H, Abachi T. A Fuzzy expert system for prescribing atorvastatin optimum dose. *Koomesh* 2011; **13**: Pe43-Pe49
- 29 **Zolnoori M**, Zarandi MH, Moin M. Application of intelligent systems in asthma disease: designing a fuzzy rule-based system for evaluating level of asthma exacerbation. *J Med Syst* 2012; **36**: 2071-2083 [PMID: 21399914 DOI: 10.1007/s10916-011-9671-8]

**P- Reviewer:** Ido Y, Pastromas S    **S- Editor:** Kong JX    **L- Editor:** A  
**E- Editor:** Wu HL





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