# World Journal of Gastrointestinal Pathophysiology

World J Gastrointest Pathophysiol 2016 November 15; 7(4): 300-319



A peer-reviewed, online, open-access journal of gastrointestinal pathophysiology

# **Editorial Board**

2016-2019

The World Journal of Gastrointestinal Pathophysiology Editorial Board consists of 503 members, representing a team of worldwide experts in gastrointestinal pathophysiology. They are from 45 countries, including Argentina (2), Australia (13), Austria (2), Belgium (7), Brazil (9), Brunei Darussalam (1), Canada (21), China (30), Croatia (1), Czech Repoublic (2), Denmark (3), Egypt (1), Estonia (1), Finland (1), France (8), Germany (24), Greece (7), Hungary (5), India (12), Indonesia (1), Iran (2), Ireland (2), Israel (8), Italy (38), Japan (45), Lebanon (3), Malaysia (1), Mexico (2), Netherlands (7), Norway (1), Poland (4), Portugal (1), Romania (1), Russia (1), Singapore (4), South Korea (12), Spain (24), Sweden (11), Switzerland (3), Thailand (2), Turkey (7), Ukraine (1), United Kingdom (10), United States (161) and Venezuela (1).

#### **EDITOR-IN-CHIEF**

Thomas Y Ma, Albuquerque

#### **ASSOCIATE EDITOR**

Naohisa Yoshida, Kyoto

# GUEST EDITORIAL BOARD MEMBERS

Jia-Ming Chang, Taipei Wai-Keung Chow, Taichung Chien-Wei Hsu, Kaohsiung Ming-Tsan Lin, Taipei Bor-Shyang Sheu, Tainan Jin-Town Wang, Taipei

# MEMBERS OF THE EDITORIAL BOARD



#### **Argentina**

Bernabé M Quesada, *Buenos Aires* Marcelo G Roma, *Rosario* 



#### **Australia**

Chris R Abbiss, Joondalup
Guy D Eslick, Sydney
Montri Gururatsakul, South Australia
Chandana B Herath, Victoria
Michael Horowitz, Adelaide
Mustafa Khasraw, Victoria
Shu-Chuen Li, New South Wales
Michael HJ Maes, Geelong
Nam Q Nguyen, South Australia
Kulmira Nurgali, Victoria
Nicholas J Spencer, South Australia

Deborah J Verran, NSW Shufeng Zhou, Victoria



#### Austria

Cord Langner, *Graz* Dietmar Ofner-Velano, *Salzburg* 



#### **Belgium**

Kathleen Blondeau, Leuven Ilse M Hoffman, Leuven Theodoor A Niewold, Heverlee Xavier Sagaert, Leuven Jean-Marie Vanderwinden, Brussels Kristin Verbeke, Belgium Mathieu Vinken, Brussels



#### Brazil

Uilian Andreis, Botucatu
Everson LDA Artifon, Sao Paulo
Joao B Calixto, Trindade
Niels OS Camara, Cidade Universitária
Júlio MF Chebli, Juiz de Fora
Clélia A Hiruma-Lima, Botucatu
Marcel CC Machado, Sao Paulo
Juarez AS Quaresma, Belem
Wagner Vilegas, Araraquara



#### Brunei Darussalam

Vui H Chong, Bandar Seri Begawan



#### Canada

Jean-Francois Beaulieu, Sherbrooke Francois Boudreau, Sherbrooke George A Bubenik, Guelph Wangxue Chen, Ottawa Fernando Fornari, Montréal Jan D Huizinga, Puslinch Ruben Hummelen, Dundas Kusum K Kharbanda, Omaha Wolfgang A Kunze, Hamilton Jianjun Li, Ottawa Roderick J MacLeod, Kingston Michele Molinari, Halifax Nathalie Rivard, Sherbrooke Nathalie Rivard, Sherbrooke Kirill Rosen, Halifax Manuela M Santos, Montreal Caroline Saucier, Québec Jean Sévigny, Québec Eldon A Shaffer, Calgary Alan BR Thomson, Edmonton Pierre H Vachon, Sherbrooke



#### China

Kai-Xing Ai, Shanghai Zhao-Xiang Bian, Hong Kong Minhu Chen, Guangzhou Chi-Hin Cho, Hong Kong Zhong-Hong Gao, Wuhan Jun-Ming Guo, Ningbo Jing-Yan Han, Beijing Jian-Dong Huang, Hong Kong Jia-Fu Ji, Beijing



Zhan-Ju Liu, Shanghai
Shi Liu, Wuhan
Zhen-Ning Wang, Shenyang
Xiao-Hong Wang, Beijing
Wei Wei, Hefei
Dong-Ping Xie, Shanghai
Wen-Xie Xu, Shanghai
Xiao Yang, Beijing
Hua Yang, Chongqing
Wei-Zhen Zhang, Beijing
Hua-Chuan Zheng, Shenyang
Min-Sheng Zhu, Nanjing
Jinxia Zhu, Beijing
Da-Ling Zhu, Harbin
Yong-Liang Zhu, Hangzhou



#### Croatia

Alen Protic, Rijeka



#### **Czech Republic**

Pavel Hladik, Semily Martin Vokurka, Prague



#### Denmark

Lars Arendt-Nielsen, *Aalborg* Frank V Schiodt, *Copenhagen* Jonas Worsoe, *Aarhus* 



#### Egypt

Mahmoud Aboelneen Khattab, Minia



#### Estonia

Enn Seppet, Tartu



#### **Finland**

Pauli Antero Puolakkainen, Turku



#### France

Bruno Bonaz, Grenoble
Pierre M Dechelotte, Rouen
Jean-Paul Lallès, Saint-Gilles
Charles-Henri Malbert, Saint-Gilles
Thierry Piche, Nice
Pascale Plaisancié, Lyon
Michelina Plateroti, Lyon
Véronique Vitton, Marseille



#### Germany

Hans G Beger, Ulm Carsten Bergmann, Ingelheim Elke Cario, Essen Arno J Dormann, Koln Nikolaus Gassler, Aachen Andreas Geier, Wuerzburg Werner Hartwig, Heidelberg Marion Hewicker-Trautwein, Hannover Jens Hoeppner, Freiburg Tobias Keck, Freiburg Jorg Kleeff, Munich Peter Malfertheiner, Magdeburg Oliver Mann, Hamburg Christoph W Michalski, Munich Andreas K Nussler, Munich Christian Pehl, Vilsbiburg Peter Schemmer, Heidelberg Manuel A Silva, Penzberg Marc Stemmler, Freiburg Frank Tacke, Aachen Sya N Ukena, Braunschweig Brigitte Vollmar, Rostock Thomas M Wex, Magdeburg Margot Zoller, Heidelberg



#### Greece

Stelios F Assimakopoulos, Patras George N Dalekos, Larissa Alkiviadis Efthymiou, thessaloniki Maria Gazouli, Athens Ioannis E Koutroubakis, Heraklion Gerassimos J Mantzaris, Athens George V Papatheodoridis, Athens



#### Hungary

Mária Bagyánszki, Szeged Mihály Boros, Szeged Laszlo Czako, Szeged Pal Miheller, Budapest Zoltan Rakonczay, Szeged



#### India

Anil K Agarwal, *Delhi*Uday Bandyopadhyay, *Kolkata*Sriparna Basu, *Varanasi*Chandra K Chakraborti, *Rourkela*Nilesh M Dagia, *Mumbai*Rajeev Garg, *Punjab*Akhtar Mahmood, *Chandigarh*Veerareddy P Reddy, *Andhra Pradesh*Chandra P Sharma, *Thiruvananthapuram*Shailesh V Shrikhande, *Mumbai*Virendra Singh, *Chandigarh*Nicholas J Skill, *Indianapolis* 



#### **Indonesia**

Laurentius A Lesmana, Jakarta



#### Iran

Gholamreza Roshandel, *Gorgan* Shahram Shahabi, *Urmia* 



#### Ireland

Billy Bourke, *Dublin* Stephen J Keely, *Dublin* 



#### Israel

Yosefa Avraham, Jerusalem Yaron Bar-Dayan, Holon Shomron Ben-Horin, Tel-Hashomer Boris Kirshtein, Beer Sheva Stephen Malnick, Rehovot Yaakov Maor, Tel-Hashomer Rifaat Safadi, Jerusalem Nachum Vaisman, Tel Aviv



## Italy

Rosaria Acquaviva, Catania Alessandro Antonelli, Pisa Giacosa Attilio, Genova Salavtore Auricchio, Naples Guido Basilisco, Milan Antonio Basoli, Rome Massimo Bellini, Pisa Luigi Bonavina, Milano Alfio Brogna, Catania Giuseppe Calamita, Bari Raffaele Capasso, Naples Ignazio Castagliuolo, Padova Francesco Cresi, Torino Rosario Cuomo, Napoli Salvatore Cuzzocrea, Gazzi Mario M D'Elios, Florence Cinzia Domeneghini, Milan Luca Elli, Milano Walter Fries, Messina Eugenio Gaudio, Rome Marco Gobbetti, Bari Fabio Grizzi, Rozzano Enzo Grossi, Milanese Enzo Ierardi, Foggia Pietro Invernizzi, Monza Angelo A Izzo, Naples Anna Kohn, Rome Giovanni Latella, L'Aquila Massimo Marignani, Rome Sergio Morini, Rome Raffaele Pezzilli, Bologna Cristiano Rumio, Milan Giovanni Sarnelli, Naples Edoardo V Savarino, Padua Pierpaolo Sileri, Roma Annamaria Staiano, Naples Giacomo C Sturniolo, Padua Claudio Tiribelli, Trieste



#### **Japan**

Hirotada Akiho, Fukuoka Akihiro Asakawa, Kagoshima Hisashi Aso, Sendai Yasu-Taka Azuma, Osaka Shotaro Enomoto, Wakayama Mikihiro Fujiya, Hokkaido Takahisa Furuta, Hamamatsu Akira Hokama, Nishihara Ryota Hokari, Saitama Yuichi Hori, Hyogo Masahiro Iizuka, Akita Shoji Ikuo, Kobe Motohiro Imano, Osaka Hajime Isomoto, Nagasaki Tatehiro Kagawa, Isehara Haruki Kitazawa, Sendai Shigeru BH Ko, Tokyo Xiao-Kang Li, Tokyo Noriaki Manabe, Okayama Atsushi Masamune, Sendai Hiroyuki Matsubayashi, Suntogun Kazuyuki Matsushita, Inohana Chuo-ku Reiko Miyazawa, Gunma Kazunari Murakami, Oita Hikaru Nagahara, Tokyo Yuji Naito, Kyoto Atsushi Nakajima, Yokohama Shoji Natsugoe, Kagoshima City Tsutomu Nishida, Suita Koji Nomoto, Tokyo Shouji Shimoyama, Tokyo Goshi Shiota, Yonago Hidekazu Suzuki, *Tokyo* Hitoshi Takagi, Gunma Toru Takahashi, Okayama Yoshihisa Takahashi, Tokyo Kan Uchiyama, Chiba Yoshiyuki Ueno, Sendai Takato Ueno, Fukuoka Hisayuki Uneyama, Kwasaki Mitsunori Yamakawa, Yamagata Takayuki Yamamoto, Mie Yutaka Yata, Maebashi-city Naohisa Yoshida, Kyoto



#### Lebanon

Costantine F Daher, *Byblos* Assaad M Soweid, *Beirut* Julnar Usta, *Beirut* 

Hitoshi Yoshiji, Kashihara



#### Malaysia

Andrew CS Boon, Ipoh



#### Mexico

José M de la Roca-Chiapas, *Leon* Maria-Raquel Huerta-Franco, *Leon* 



#### Netherland

Wouter J de Jonge, Amsterdam Aldo Grefhorst, Groningen Daniel Keszthelyi, Maastricht Cornelis FM Sier, ZA Leiden Pieter J Tanis, AZ Amsterdam Luc JW van der Laan, Rotterdam Sander van der Marel, Leiden



Anne M Bakke, Oslo



Stanistaw A Hac, Gdansk

Stanislaw J Kowalski, *Krakow* Agata Mulak, *Wroclaw* Napoleon Waszkiewicz, *Choroszcz* 



#### **Portugal**

Ricardo Marcos, Porto



#### **Romania**

Mihai Ciocirlan, Bucharest



#### Russia

Ludmila Filaretova, Petersburg



#### **Singapore**

Madhav Bhatia, Singapore Brian K Goh, Singapore Khek-Yu Ho, Singapore Cliff KS Ong, Singapore



#### **South Korea**

Jae Hee Cheon, Seoul
Myung-Haing Cho, Seoul
Ki-Baik Hahm, Seongnam
HO Jae Han, Gwangju
Chang-Duk Jun, Gwangju
Sang Geon Kim, Seoul
Hong Joo Kim, Seoul
Jin-Kyung Kim, Gyeongsan-Si
Won-Jae Lee, Seoul
Sung-Joo Park, Iksan
Seung Ha Park, Gangwon Do
Kwan-Kyu Park, Daegu



#### Spain

Raquel Abalo, Madrid Juan G Abraldes, Barcelona Dario Acuna-Castroviejo, Armilla Agustin Albillos, Madrid Maria-Angeles Aller, Madrid Fernando Azpiroz, Barcelona Ramón Bataller, Barcelona Marco Bustamante-Balén, Valencia Andres Cardenas, Barcelona Joan Claria, Barcelona Pere Clavé, Barcelona Manuel B de Acosta, Santiago Manuel Giner, Madrid Borja Hernandez-Breijo, Madrid Angel I Lanas, Zaragoza Maite T Martin, Barcelona Vicente Martinez, Bellaterra Jose M Mates, Malaga Julio Mavol, Madrid Marcal Pastor-Anglada, Barcelona María E Sáez, Seville Yolanda Sanz, Burjassot Carlos Taxonera, Madrid

Maria D Yago, Granada



#### Sweden

Marco Del Chiaro, Stockholm
Frida Fak, Gothenburg
Gunnar FA Flemstrom, Uppsala
Evangelos Kalaitzakis, Gothenburg
Kristina Lamas, Umea
Bob R Olsson, Goteborg
Sara M Regnér, Malmo
Peter T Schmidt, Stockholm
Xiao-Feng Sun, Linkoping
Henrik Thorlacius, Malmo
Curt Tysk, Orebro



#### Switzerland

Jyrki J Eloranta, Zurich Remy Meier, Liestal Catherine M Pastor, Geneva



#### Thailand

Thawatchai Akaraviputh, Bangkok Weekitt Kittisupamongkol, Bangkok



#### Turkey

Mehmet Bektas, *Ankara* Ugur Duman, *Bursa* Mukaddes Esrefoglu, *Istanbul* Ahmet Guven, *Ankara* Muammer Karadeniz, *Bornova-Izmir* Elvan Ozbek, *Sakarya* Ilhami Yuksel, *Ankara* 



#### Ukraine

Oksana S Zayavhkivska, Lviv



# United Kingdom

Geoffrey Burnstock, London
Janice E Drew, Aberdeen
Girish L Gupte, Birmingham
David C Hay, Edinburgh
Nusrat Husain, Manchester
Michael L Lucas, Glasgow
Jamie Murphy, London
Vadim Sumbayev, Chatham Maritime
Wing-Kin Syn, Birmingham
Andrea Varro, Liverpool



#### **United States**

Sami R Achem, Florida Tauseef Ali, Oklahoma City Shrikant Anant, Oklahoma Mohammed S Anwer, North Grafton Andrew Aronsohn, Chicago Toms Augustin, Sayre



Gyorgy Baffy, Boston Michael T Bailey, Columbus Kim E Barrett, San Diego Marc D Basson, Grand Forks Robert L Bell, Berkeley Heights David H Berger, Texas Urs A Boelsterli, Storrs Michael W Bradbury, Erie Qiang Cai, Atlanta Weibiao Cao, Providence Subhash C Chauhan, Sioux Falls Tao-Sheng Chen, Memphis Jian-De Chen, Galveston Chiang YL Chiang, Rootstown Mashkoor A Choudhry, Maywood Parimal Chowdhury, Little Rock Eric Cohen, Boston Robert T Cormier, Duluth Edwin A Deitch, Newark Sharon DeMorrow, Texas Dan A Dixon, Columbia James P Dolan, Portland Henry H Dong, Pittsburgh Hui Dong, La Jolla Ashkan Farhadi, Irvine Bin Feng, Pittsburgh Jenifer Fenton, East Lansing Alessandro Fichera, Chicago Mitchell P Fink, Pittsburgh Leo R Fitzpatrick, Rancho Cordova Robert A Forse, Omaha Glenn T Furuta, Aurora Juan F Gallegos-Orozco, Scottsdale Pandu R Gangula, Nasvhille Timothy B Gardner, Lebanon Shannon S Glaser, Temple Beverley Greenwood-Van Meerveld, Oklahoma City John R Grider, Richmond Yan-Fang Guan, Cincinnati Gregory M Holmes, Baton Rouge Richard Hu, Los Angeles Hartmut Jaeschke, Kansas City Robert T Jensen, Bethesda Sreenivasa S Jonnalagadda, Louis Michel Kahaleh, Charlottesville Andreas M Kaiser, Los Angeles Randeep S Kashyap, Rochester Richard Kellermayer, Houston

Chris Kevil, Shreveport

Pawel R Kiela, Tucson

Sandeep Khurana, Baltimore

Tammy L Kindel, Cincinnati

Gordana Kosutic, *Durham* David Kravetz, *San Diego* 

Ashok Kumar, Detroit Anthony Kumar, Los Angeles John H Kwon, Chicago Muriel Larauche, Los Angeles Ai-Xuan Le Holterman, Chicago I. Michael Leitman, New York Felix W Leung, Sepulveda Suthat Liangpunsakul, Indianapolis Feng-Xin Lu, Boston Pauline K Lund, Chapel Hill Guang-Xiang Luo, Lexington Jay Luther, Ann Arbor Ram I Mahato, Memphis Akhil Maheshwari, Birmingham Kenneth Maiese, Newark Adhip PN Majumdar, Detroit José E Manautou, Storrs Craig J McClain, Louisville Dermot PB McGovern, Los Angeles Douglas S Merrell, Bethesda Murielle Mimeault, Omaha Emiko Mizoguchi, Boston Huan-Biao Mo, Denton Adam J Moeser, Raleigh Ramzi M Mohammad, Detroit Satdarshan SP Monga, Pittsburgh Roger K Moreira, New York Sandeep Mukherjee, Omaha Karnam S Murthy, Richmond Michael J Nowicki, Jackson Shuji Ogino, Boston Mary F Otterson, Wisconsin Chung Owyang, Ann Arbor Helieh S Oz, Lexington Marco G Patti, Chicago Timothy M Pawlik, Baltimore Sara Peleg, Houston Li-Ya Qiao, Richmond Chao Oin, Oklahoma Parvaneh Rafiee, Milwaukee Sigrid A Rajasekaran, Wilmington Vazhaikkurichi Rajendran, Morgantown Jean P Raufman, Baltimore Ramesh M Ray, Memphis Arie Regev, Indianapolis Yehuda Ringel, Chapel Hill Richard A Rippe, Rockville Chantal A Rivera, Bossier Andrea Romani, Cleveland Praveen K Roy, Marshfield Paul A Rufo, Boston David B Sachar, New York

Muhammad Y Sheikh, Fresno Bo Shen, Cleveland Le Shen, Chicago Frank A Simmen, Little Rock Steven M Singer, Washington Shailinder J Singh, Washington Adam J Smolka, Charleston Ned Snyder, Houston Zhen-Yuan Song, Chicago Gagan K Sood, Houston Rhonda Souza, Dallas Stuart J Spechler, Dallas Subbaramiah Sridhar, Augusta Catia Sternini, Los Angeles Veedamali S Subramanian, Long Beach Jun Sun, Rochester Yvette Taché, Los Angeles Xiao-Di Tan, Chicago Paul D Terry, Atlanta Ma Thomas, Albuquerque Jennifer S Tirnauer, Farmington Andrea Todisco, Ann Arbor George C Tsokos, Boston Vic Velanovich, Detroit Raj Vuppalanchi, Indianapolis Estela Wajcberg, Cranford Arnold Wald, Madison Li-Xin Wang, Los Angeles Horst C Weber, Boston Guo-Yao Wu, Texas Christian Wunder, Bethesda Zuo-Liang Xiao, Cleveland Guang-Yin Xu, Galveston Guo-Rong Xu, East Orange Yoshio Yamaoka, Houston Guang-Yu Yang, Chicago Jay A Yelon, Valhalla Shao-Yong Yu, Hershey Yana Zavros, Cincinnati Joerg Zehetner, Los Angeles Jian X Zhang, Charlotte Zhi Zhong, Charleston Hui-Ping Zhou, Richmond Zhan-Xiang Zhou, Kannapolis Qing Zhu, Bethesda Yao-Hui Zhu, Stanford

Ron Schey, Iowa City



Fabian Michelangeli, Caracas

Bimaljit S Sandhu, Richmond

Sanjaya K Satapathy, New Hyde Park



#### **Contents**

Quarterly Volume 7 Number 4 November 15, 2016

#### **ORIGINAL ARTICLE**

#### **Basic Study**

300 Anti-Helicobacter pylori effect of CaG-NANA, a new sialic acid derivative

Rhee YH, Ku HJ, Noh HJ, Cho HH, Kim HK, Ahn JC

#### **Retrospective Study**

307 Microscopic colitis in patients with mild duodenal damage: A new clinical and pathological entity ("lymphocytic enterocolitis")?

Bonagura GA, Ribaldone DG, Fagoonee S, Sapone N, Caviglia GP, Saracco GM, Astegiano M, Pellicano R

#### **META-ANALYSIS**

314 Hepatitis C infection and renal cell carcinoma: A systematic review and meta-analysis

Wijarnpreecha K, Nissaisorakarn P, Sornprom S, Thongprayoon C, Thamcharoen N, Maneenil K, Podboy AJ, Cheungpasitporn W



#### **Contents**

#### World Journal of Gastrointestinal Pathophysiology Volume 7 Number 4 November 15, 2016

#### **ABOUT COVER**

Editorial Board Member of World Journal of Gastrointestinal Pathophysiology, Marc D Basson, MD, PhD, Professor, Department of Surgery, Pathology, and Basic Sciences, University of North Dakota School of Medicine and the Health Sciences, Grand Forks, ND 58202, United States

#### **AIM AND SCOPE**

World Journal of Gastrointestinal Pathophysiology (World J Gastrointest Pathophysiol, WJGP, online ISSN 2150-5330, DOI: 10.4291), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJGP is to report rapidly the most recent results in basic and clinical research on gastrointestinal pathophysiology, including all aspects of normal or abnormal function of the gastrointestinal tract, hepatobiliary system, and pancreas. WJGP specifically covers growth and development, digestion, secretion, absorption, metabolism and motility relative to the gastrointestinal organs, as well as immune and inflammatory processes, and neural, endocrine and circulatory control mechanisms that affect these organs. This journal will also report new methods and techniques in gastrointestinal pathophysiological research.

We encourage authors to submit their manuscripts to WJGP. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

#### INDEXING/ABSTRACTING

World Journal of Gastrointestinal Pathophysiology is now indexed in PubMed, PubMed Central.

#### **FLYLEAF**

#### I-IV **Editorial Board**

#### **EDITORS FOR** THIS ISSUE

Responsible Assistant Editor: Xiang Li Responsible Electronic Editor: Huan-Liang Wu Proofing Editor-in-Chief: Lian-Sheng Ma

Responsible Science Editor: Fang-Fang Ji Proofing Editorial Office Director: Xiu-Xia Song

#### NAME OF JOURNAL

World Journal of Gastrointestinal Pathophysiology

ISSN 2150-5330 (online)

#### LAUNCH DATE April 15, 2010

## Frequency

#### EDITOR-IN-CHIEF

Thomas Y Ma, MD, PhD, Professor, Chief, Division of Gastroenterology and Hepatology, University of New Mexico, MSC10 5550, 1 UNM, Albuquerque, NM 87131, United States

#### EDITORIAL BOARD MEMBERS

All editorial board members resources online at http:// www.wignet.com/2150-5330/editorialboard.htm

#### EDITORIAL OFFICE

Xiu-Xia Song, Director
Fang-Fang Ji, Vice Director
World Journal of Gastrointestinal Pathophysiology Baishideng Publishing Group Inc 8226 Regency Drive, Pleasanton, CA 94588, USA Telephone: +1-925-2238242 Fax: +1-925-2238243 E-mail: editorialoffice@wignet.com Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx http://www.wjgnet.com

#### **PUBLISHER**

Baishideng Publishing Group Inc 8226 Regency Drive, Pleasanton, CA 94588, USA Telephone: +1-925-223-8242 Fax: +1-925-223-8243 E-mail: bpgoffice@wjgnet.com Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx http://www.wjgnet.com

# PUBLICATION DATE November 15, 2016

#### **COPYRIGHT**

2016 Baishideng Publishing Group Inc. Articles published by this Open Access journal are distributed under the terms of the Creative Commons Attribution Noncommercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinionsof their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

# INSTRUCTIONS TO AUTHORS http://www.wignet.com/bpg/gerinfo/204

ONLINE SUBMISSION http://www.wjgnet.com/esps/



Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.4291/wjgp.v7.i4.300 World J Gastrointest Pathophysiol 2016 November 15; 7(4): 300-306 ISSN 2150-5330 (online) © 2016 Baishideng Publishing Group Inc. All rights reserved.

ORIGINAL ARTICLE

**Basic Study** 

# Anti-Helicobacter pylori effect of CaG-NANA, a new sialic acid derivative

Yun-Hee Rhee, Hyun-Jeong Ku, Hye-Ji Noh, Hyang-Hyun Cho, Hee-Kyong Kim, Jin-Chul Ahn

Yun-Hee Rhee, Hyun-Jeong Ku, Jin-Chul Ahn, Beckman Laser Institute Korea, Dankook University, Chungnam 31116, South Korea

Hye-Ji Noh, Hyang-Hyun Cho, Hee-Kyong Kim, MEDINU-TROL Co., Jeonnam 57024, South Korea

Jin-Chul Ahn, Department of Biomedical Science, College of Medicine, Dankook University, Chungnam 31116, South Korea

Jin-Chul Ahn, Medical Laser Research Center, Dankook University, Chungnam 31116, South Korea

Author contributions: Rhee YH performed the majority of experiments, analyzed the data, and wrote the manuscript; Ku HJ and Noh HJ participated in animal experiments; Cho HH, Kim HK and Ahn JC designed and coordinated the research.

Institutional animal care and use committee statement: All procedures involving animals were reviewed and approved by the Institutional Animal Care and Use Committee of the Dankook University (IACUC protocol number: DKU-14-036).

Animal care and use statement: The ICR mice (20-22 g) were housed in pathogen free environment and had free access to sterile neutral water and standard mouse feed. Oral administration was performed with conscious animals using sonde appropriate for the animal size.

Conflict-of-interest statement: The authors have no conflict of interest to declare.

Data sharing statement: Statistics and histology analysis of data are available from the corresponding author at jcahn@dankook.ac.kr.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

Manuscript source: Invited manuscript

Correspondence to: Jin-Chul Ahn, MD, Department of Biomedical Science, College of Medicine, Dankook University, 119 Dandae-ro, Cheonan, Chungnam 31116, South Korea. jcahn@dankook.ac.kr

South Korea. jcahn@dankook.ac.l Telephone: +82-1899-3700

Received: June 28, 2016

Peer-review started: June 29, 2016 First decision: August 10, 2016 Revised: August 12, 2016 Accepted: September 13, 2016 Article in press: September 18, 2016 Published online: November 15, 2016

#### **Abstract**

#### AIM

To investigate the bactericidal effects of calcium chelated N-acetylneuraminic acid-glycomacropeptide (CaG-NANA) against *Helicobacter pylori* (*H. pylori*).

#### **METHODS**

For manufacture of CaG-NANA, calcium (Ca) was combined with glycomacropeptide (GMP) by chelating, and N-acetylneuraminic acid (NANA) was produced with Ca-GMP substrate by an enzymatic method. The final concentration of each component was 5% Ca, 7% NANA, 85% GMP, and 3% water. For *in vitro* study, various concentrations of CaG-NANA were investigated under the minimal inhibitory concentration (MIC). For *in vivo* study, CaG-NANA was administered orally for 3 wk after *H. pylori* infection. The levels of inflammatory cytokines in blood were analyzed by enzyme-linked immunosorbent assay and eradication of *H. pylori* was assessed by histological observation.

#### **RESULTS**

The time-kill curves showed a persistent decrease in cell



numbers, which depended on the dose of CaG-NANA, and MIC of CaG-NANA against  $H.\ pylori$  was 0.5% in vitro. Histopathologic observation revealed no obvious inflammation or pathologic changes in the gastric mucosa in the CaG-NANA treatment group in vivo. The colonization of  $H.\ pylori$  was reduced after CaG-NANA treatment. The levels of interleukin (IL)-6, IL-1 $\beta$ , tumor necrosis factor- $\alpha$ , and IL-10 were also decreased by CaG-NANA.

#### **CONCLUSION**

CaG-NANA demonstrates effective anti-bactericidal activity against *H. pylori* both *in vitro* and *in vivo*.

**Key words:** *Helicobacter pylori*; Calcium chealated N-acetylneuraminic acid-glycomacropeptide

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Calcium chelated N-acetylneuraminic acidglycomacropeptide demonstrates effective anti-bactericidal activity against *Helicobacter pylori* both *in vitro* and *in vivo*.

Rhee YH, Ku HJ, Noh HJ, Cho HH, Kim HK, Ahn JC. Anti-Helicobacter pylori effect of CaG-NANA, a new sialic acid derivative. World J Gastrointest Pathophysiol 2016; 7(4): 300-306 Available from: URL: http://www.wjgnet.com/2150-5330/full/ v7/i4/300.htm DOI: http://dx.doi.org/10.4291/wjgp.v7.i4.300

#### INTRODUCTION

Helicobacter pylori (H. pylori) has been reported to be associated with many gastrointestinal diseases such as chronic gastritis, peptic ulcers, gastric carcinoma, and mucosa-associated lymphoid tissue lymphoma<sup>[1-3]</sup>. Until now, standard triple therapy consisting of a proton pump inhibitor and two broad-spectrum antibiotics, usually amoxicillin, clarithromycin or metronidazole, has been able to achieve eradication<sup>[3-5]</sup>. However, the Maastricht IV Consensus report recommended the careful choice of the antibiotic combination for treatment according to local H. pylori antibiotic resistance patterns due to multidrug resistance of H. pylori. For example, the concentrations of antibiotics for treatment of *H. pylori* infection were required at four times when *H. pylori* had local clarithromycin resistance<sup>[6]</sup>. Since antibiotic abuse had side effects and allowed development of resistance to antibiotics, alternative strategies have been proposed to counteract H. pylori infection. As an alternative approach, preservation of mucus from H. pylori attachment is emphasized because H. pylori infection disrupted the epithelial barrier which resulted in inflammation or cancer<sup>[7]</sup>. For example, dietary inhibitors such as lacto-oligosaccharide has been suggested as a solution for *H. pylori* infections<sup>[8]</sup>. Although oligosaccharides specific for the *H. pylori*  lectins may potentially act as inhibitors of adhesion to mucus, their production in commercial amounts as antiadhesion therapeutic agents is still a problem<sup>[9,10]</sup>. Here, we introduce a new N-acetylneuraminic acid (NANA) combined glycomacropeptide (GMP) which was made from milk serum hydrolysis protein powder as an anti-*H. pylori* agent. Wadström *et al*<sup>[11]</sup> reported that NANA derivative had an adhesion potential to *H. pylori* lectins and Hirmo *et al*<sup>[12]</sup> reported that milk glycoprotein had an anti-*H. pylori* effect. We designed a new material with NANA, GMP, and calcium (CaG-NANA), and anti-*H. pylori* activities of CaG-NANA were investigated both *in vitro* and *in vivo*.

#### **MATERIALS AND METHODS**

#### H. pylori strain and experimental animals

H. pylori strain (SS1-passed-5) was kindly provided by Helicobacter pylori Korean Type Culture Collection (HpKTCC, Jinju, South Korea) and grown in 1.5% agar added Brucella broth with 10% horse serum (No. CM0169; OXOID, Waltham, MA, United States). The pathogen-free (SPF) male ICR mice (20-22 g) were purchased from Orient Bio (Daejeon, South Korea). All the animals were housed in an SPF environment and had free access to sterile neutral water and standard mouse feed. The experimental procedures in this study were approved by the Experimental Animal Ethics Committee of Dankook University, South Korea (No. DKU14-036).

#### Supply of CaG-NANA

CaG-NANA and each component of CaG-NANA were provided by MediNutrol (Kwangju, South Korea). As shown in Figure 1A, CaG-NANA was manufactured from calcium (Ca), GMP, and NANA by chelating and enzyme methods where the component concentrations were 5%, 85%, and 7%, respectively. For comparing each produced compound, we wrote abbreviated form as standard NANA (S-NANA), GMP linked NANA (G-NANA), and calcium chelated G-NANA (CaG-NANA).

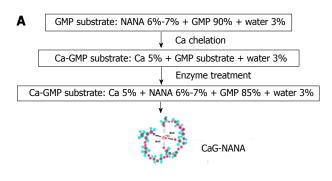
# High performance liquid chromatography for confirmation of NANA

For confirmation of NANA component, we analyzed CaG-NANA using high performance liquid chromatography (HPLC) method. HPLC analysis was performed using Agilent 1260 model equipped with a pump (G1311C), an auto sampler (G1329B), a column (G1316A), and a ultraviolet detector (G1314F), which was purchased from Agilent (Santa Clara, CA, United States). The condition of analysis was described in Table 1.

#### Culture and collection of H. pylori

H. pylori was incubated in Brucella medium contained the selective supplements (No. SR0083; OXOID, Waltham, MA, United States) under microaerobic environment (15% CO₂, 5% O₂, 80% N₂) at 37 °C for 72 h. The bacterial colonies were collected and identified





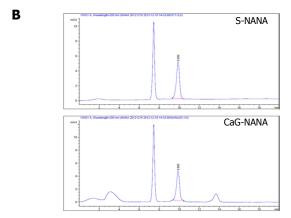


Figure 1 Confirmation of N-acetylneuraminic acid. A: The scheme of CaG-NANA manufacture; B: HPLC analysis of NANA content in CaG-NANA. The condition of HPLC analysis is described in Table 1. GMP: Glycomacropeptide; HPLC: High performance liquid chromatography; CaG-NANA: Calcium-glycomacropeptide-N-acetylneuraminic acid; S-NANA: Standard N-acetylneuraminic acid.

with a Pronto-Dry infection kit (Kokab Enterprise, Karachi, Pakistan) for bactericidal tests *in vitro*.

#### Inhibitory effect of CaG-NANA on H. pylori in vitro

The inhibitory activity of CaG-NANA against the growth of H. pylori was assessed using an agar dilution method. Briefly, 0.1%, 0.25%, or 0.5% of CaG-NANA and its components were added to the Brucella agar. The nondrug agar served as a negative control. Agar plates were inoculated with *H. pylori* at serial concentrations of  $1 \times 10^8$ ,  $1 \times 10^7$ , and  $1 \times 10^6$  colony forming units (CFU)/mL and cultured for 72 h. The minimal inhibitory concentration (MIC) was defined as the minimal concentration of CaG-NANA required for complete inhibition of H. pylori growth. The colonies were counted using image J (https://imagej.nih.gov/ij/) after 72 h incubation and the average number was calculated. Bactericidal activity was evaluated using time-kill curves with 0.5, 1.0 and 2.0 × MIC of CaG-NANA compared with the blank controls.

#### Experimental design in vivo

Mice (n=40) were randomly divided into four groups: Blank control, H.~pylori infected control, antibiotic treatment, and CaG-NANA treatment. H.~pylori (1  $\times$  10 $^9$  /mL) was administered by gastric intubation to mice three times in a week except the blank control, and H.~pylori

Table 1 High performance liquid chromatography analysis condition

Detector	UV detector
Wavelength	205 nm
Column	Aminex HPX-87H Ion Exclusion column
	$(300 \text{ mm} \times 7.8 \text{ mm}, 9 \mu\text{m})$
Mobile phase	10 mmol/L H2SO4 in water (isocratic)
Running time	20 min
Flow rate	0.5 mL/min
Injection volume	10 mL
Temperature	40 ℃

infection was detected with gastric irrigation from randomly selected mice using an *H. pylori* detection test kit which was purchased from Pronto Dry (Brignais, France; Supple 1A). The antibiotic treatment was performed with a suspension of amoxicillin (12.33 mg/kg), metronidazole (164.40 mg/kg), and clarithromycin (205.54 mg/kg) which were equivalent to clinical administration (Dankook University hospital, Korea). Treatments with CaG-NANA and each component including NANA, S-NANA, and G-NANA were performed by oral administration every day for 3 wk, and the experimental design is shown in Figure 2A.

#### Histological observation

The animals were deprived of feed but allowed free access to water for 24 h before sacrificed. At the end of sacrifice, blood was collected from mouse and gastric tissues were removed and fixed in 10% formalin. After fixation, tissues were processed and embedded in paraffin. The paraffin blocks were cut into 4  $\mu m$  sections and stained with Harris' hematoxylin and eosin for histological observation under a microscope (BX51, Olympus, Miami, FL, United States).

#### Enzyme-linked immunosorbent assay

The expression levels of interleukin (IL)-1 $\beta$ , IL-6, tumor necrosis factor (TNF)- $\alpha$ , and IL-10 in blood serum were determined using enzyme-linked immunosorbent assay (ELISA). Mouse ELISA kits were purchased from R&D Systems (Minneapolis, MN, United States). The assays were performed according to the manufacturer's instructions and repeated in triplicate.

#### Statistical analysis

Eradication rates were compared among groups by one-way analysis of variance (Kruskal-Wallis) and Dunn's multiple comparison test (GraphPad Software Inc., La Jolla, CA, United States). P < 0.05 was considered statistically significant.

#### **RESULTS**

#### Confirmation of NANA

As shown Figure 1B, the NANA component of CaG-NANA showed the same purity as standard NANA.



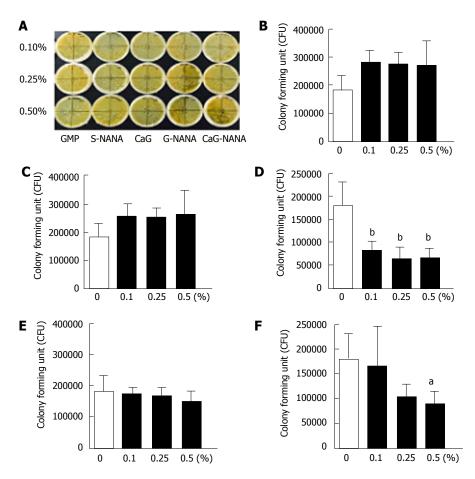


Figure 2 Inhibitory effect of calcium-glycomacropeptide-N-acetylneuraminic acid on *Helicobacter pylori in vitro*. Agar plates were inoculated with *Helicobacter pylori* (*H. pylori*) at serial concentrations of 1 × 10<sup>8</sup>, 1 × 10<sup>7</sup>, and 1 × 10<sup>6</sup> CFU/mL and cultured for 72 h. The minimal inhibitory concentration (MIC) was defined as the minimal concentration of materials required for complete inhibition of *H. pylori* growth. Bactericidal activity was evaluated using time-kill curves with 0.5, 1.0 and 2.0 × MIC of CaG-NANA compared with blank controls. All experiments were performed three times and significance was set at <sup>8</sup>*P* < 0.1 and <sup>8</sup>*P* < 0.05. A: Picture of colony forming unit assay; B: GMP; C: CaG; D: S-NANA; E: G-NANA; F: CaG-NANA. GMP: Glycomacropeptide; CaG: Calcium-glycomacropeptide; S-NANA: Ststandard N-acetylneuraminic acid; CaG-NANA: Calcium-glycomacropeptide-N-acetylneuraminic acid.

#### Inhibitory effect of CaG-NANA on H. pylori in vitro

The bactericidal effects of CaG-NANA and each component against *H. pylori* were assessed *in vitro*. We described previously that CaG-NANA is composed of NANA, GMP and calcium, thus we tested the antibacterial effects of every component used in CaG-NANA synthesis. As shown in Figure 1, S-NANA, G-NANA and CaG-NANA had an anti-bacterial effect (Figure 1D, E and F) whereas GMP and GMP with calcium (CaG) had no activity in the colony forming assay (Figure 1B and C). This result indicated that only NANA included compound had anti-bacterial activity.

#### Inhibitory effect of CaG-NANA on H. pylori in vivo

For the *in vivo* study, we divided mice into four groups and  $1 \times 10^9$ /mL CFU of *H. pylori* was administered into mice three times for one week except the negative group. *H. pylori* infection was confirmed by gastric irrigation using an *H. pylori* detection test kit (Pronto Dry). After confirmation, treatments with antibiotics, CaG-NANA and its components were orally administered every day for 3 wk. The doses of antibiotics were described in the "Material and Methods" section (Figure 3A). The doses of CaG-NANA and its components

were fixed at 0.5% (v/w). We followed the mouse weight after treatment. As shown in Figure 3B, mouse weight was decreased in the S-NANA and H. pylori positive control groups. From this result, S-NANA was considered to have toxicity at 0.5% of concentration.

Next, we observed the gastric mucus layer of mice. The majority of the *H. pylori* population reside in the mucus which binds the organisms via specific interactions such as pathogen adherence. Normal gastric mucosa has a uniform surface epithelium (Figure 4A), whereas H. pylori infected gastric mucosa has an irregular outline of the epithelium layer along with neutrophil and macrophage infiltrations (Figure 4B). In addition, bacterial attachment is mediated by outer membrane adhesions that bind to glycoconjugates present in the gastric mucus layer, lining the surface epithelium of the gastric mucosa in the H. pylori infected group. As shown in Figure 4C, the antibiotic treatment group had decreased infiltration of macrophages and adhesion of H. pylori colonization, however, the destruction of surface epithelium layers was also observed. Meanwhile, every NANA including group showed uniformed surface epithelium layers. Interestingly, many macrophages and neutrophils were observed in the S-NANA treatment

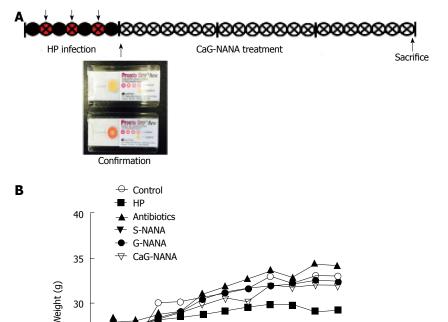


Figure 3 Inhibitory effect of calcium-glycomacropeptide-N-acetylneuraminic acid on Helicobacter pylori in vivo. Antibiotic doses are described in the "materials and methods" section. A: Schematic design of in vivo study; B: Weight changes of mice. HP: Helicobacter pyroli; S-NANA: Standard N-acetylneuraminic acid; G-NANA: Glycomacropeptide-Nacetylneuraminic acid; CaG-NANA: Calciumglycomacropeptide-N-acetylneuraminic acid.

group (Figure 4D). The damage to surface epithelium in the antibiotics treatment group and inflammation observed in the S-NANA treatment group might be related to mouse weight loses. Both the G-NANA and CaG-NANA treatment groups showed the detachment of H. pylori colonization without any damage to surface epithelium or inflammation (Figure 4E and F).

12 15 18 21 24 27 30 33

d

We also assessed the levels of IL-1 $\beta$ , IL-6, TNF- $\alpha$ , and IL-10 in blood by ELISA. PBS was used as a control because every compound was dissolved in PBS. Both of S-NANA and CaG-NANA showed a potent ability to decrease inflammatory cytokines. The level of IL-1 $\beta$  was reduced from 7.6 ng/mL to 0.8 ng/mL after S-NANA treatment and 2.36 ng/mL after CaG-NANA treatment (Figure 5A). The level of IL-6 was remarkedly decreased from 113.4 ng/mL to 33.5 ng/mL after S-NANA treatment and 1.26 ng/mL after CaG-NANA treatment (Figure 5B). The level of TNF- $\alpha$  was also reduced to 8.21 ng/mL after S-NANA treatment and 1.44 ng/mL after CaG-NANA treatment (Figure 5C). Meanwhile, the level of IL-10 was reduced only in the CaG-NANA treatment group from 7.62 ng/mL to 6.3 ng/mL (Figure 5D).

#### DISCUSSION

30

25

20

The anti-H. pylori effects of antibiotic formulas have been investigated, but the findings are limited by varying drug quality and sources, as well as the numerous and complicated components of formulas. Thus, it has been suggested that dietary inhibitors may be a solution for certain infections as an alternative approach<sup>[8]</sup>. Eradication of *H. pylori* has remained difficult for reasons that lie in the biology and environment of the organism. H. pylori populations colonize epithelial cells that line the antrum of the stomach and survive in the acidic environment<sup>[13]</sup>. Most anti-microbial agents are poorly secreted in the gastric mucosa because of the stomach environment<sup>[14]</sup>, and the residues expressed on H. pylori enable specific binding to the mucus layers. Thus, CaG-NANA was excogitated to overcome the acidic environment of the stomach and have bactericidal activity by interrupting glycol-conjugation of H. pylori on the mucosa.

CaG-NANA, a compound of N-acetylneuraminic acid with calcium combined glycomacropeptide, demonstrated an anti-bacterial effect against H. pylori. We manufactured CaG-NANA as a dietary inhibitor against H. pylori; NANA was demonstrated as an H. pylori adhesion blocker<sup>[9,15,16]</sup>, glycomacropeptide was used for modulating the acidic phase of the stomach, and calcium was inserted for improvement as functional food. NANA content in CaG-NANA was evaluated compared to S-NANA by HPLC, which confirmed its content and purity (Figure 1B). CaG-NANA at concentrations > 0.25% regulated the population of H. pylori in vitro. S-NANA also decreased the population of *H. pylori in vitro* (Figure 2D); however, S-NANA resulted in a severe weight loss in vivo (Figure 3B), which demonstrated that S-NANA treatment only had negative utility. Meanwhile, G-NANA and CaG-NANA had bactericidal and anti-adhesion effects on glyco-conjugation of the mucosa without toxicity. Bacterial attachment is mediated by outer membrane adhesions that bind to glycoconjugates present in the gastric mucus layer and lining the surface epithelium of

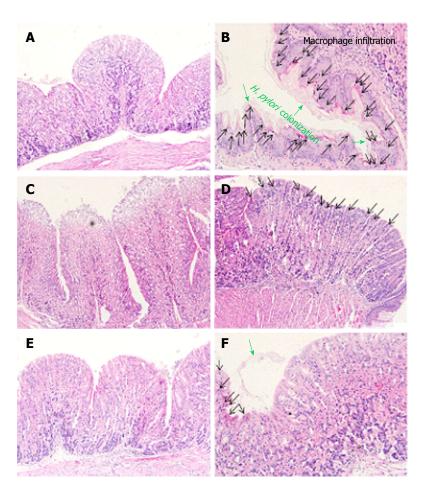


Figure 4 Histology of the gastric mucosa. Mouse stomachs were removed and underwent hematoxylin and eosin staining. *H. pylori* colonization and macrophage infiltration were observed under a light microscope. A: Normal group; B: *H. pylori* infected group; C: Antibiotics treated group; D: S-NANA treated group; E: G-NANA treated group; F: CaG-NANA treated group. S-NANA: Standard N-acetylneuraminic acid; G-NANA: Glycomacropeptide-N-acetylneuraminic acid; CaG-NANA: Calcium-glycomacropeptide-N-acetylneuraminic acid.

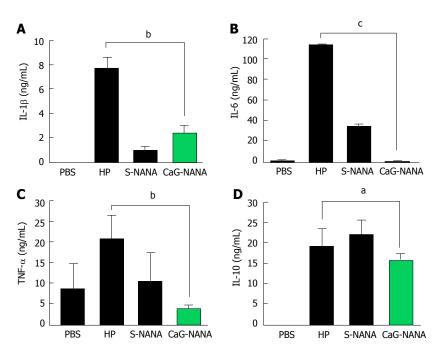


Figure 5 Expression levels of interleukine-1 $\beta$ , interleukine-6, tumor necrosis factor- $\alpha$ , and interleukine-10 in blood serum determined by enzymelinked immunosorbent assay. The assays were performed according to the manufacturer's instructions and all experiments were repeated in triplicate. Statistical analysis was performed using Dunn's multiple comparison test with significance set at  $^{8}P < 0.1$  and  $^{8}P < 0.05$ ,  $^{6}P < 0.01$ . A: IL-1 $\beta$ ; B: IL-6; C: TNF- $\alpha$ ; D: IL-10. IL: Interleukine; TNF: Tumor necrosis factor; S-NANA: Standard N-acetylneuraminic acid; G-NANA: Glycomacropeptide-N-acetylneuraminic acid; CaG-NANA: Calcium-glycomacropeptide-N-acetylneuraminic acid.

the gastric mucosa<sup>[17]</sup>.

 $H.\ pylori$  is also known to induce inflammatory response that includes the up-regulation of pro-inflammatory cytokines. CaG-NANA exerted a down-regulatory effect on pro-inflammatory cytokines such as IL-1 $\beta$ , IL-6, TNF- $\alpha$  and IL-10. Reduction of pro-inflammatory cytokines induced by  $H.\ pylori$  is responsible for the recruitment of macrophages and neutrophils in the lamina propria. CaG-NANA exerted an antagonistic effect against  $H.\ pylori$ , which is an anti-microbial effect via different mechanisms from those of antibiotics.

This study suggests that it is possible to decrease the number of *H. pylori* or its activation in the stomach through a regular ingestion of N-acetylneuraminic acid containing glycomacropeptide as dietary products. Moreover, CaG-NANA is a complex compound which can be manufactured in large scale at a low cost under good manufacturing practice (GMP). In conclusion, the anti-*H. pylori* effects of CaG-NANA were confirmed both *in vitro* and *in vivo*, which provided experimental support for future human trials.

#### **COMMENTS**

#### Background

Helicobacter pylori (H. pylori) has been reported to be associated with many gastrointestinal diseases such as chronic gastritis, peptic ulcers, gastric carcinoma, and mucosa-associated lymphoid tissue lymphoma. Until now, standard triple therapy consisting of a proton pump inhibitor and two broad-spectrum antibiotics, usually amoxicillin, clarithromycin or metronidazole, has been able to achieve eradication.

#### Research frontiers

The authors introduce a new N-acetylneuraminic acid (NANA) combined glycomacropeptide (GMP) which was made from milk serum hydrolysis protein powder as an anti-*H. pylori* agent. Wadstorm *et al* reported that NANA derivative had an adhesion potential to *H. pylori* lectins and Hirmo *et al* reported that milk glycoprotein had an anti-*H. pylori* effect. The authors designed a new material with NANA, GMP, and calcium (CaG-NANA), and anti-*H. pylori* activities of CaG-NANA were investigated both *in vitro* and *in vivo*.

#### Innovations and breakthroughs

The anti-H. pylori effects of CaG-NANA were confirmed both in vitro and in vivo, which provided experimental support for future human trials.

#### **Applications**

CaG-NANA is a complex compound which can be manufactured in a large scale at a low cost under GMP.

#### Peer-review

In the present paper, entitled "Anti-Helicobacter pylori effect of CaG-NANA, a new sialic acid derivative", Rhee H et al have investigated the effect of a compound named Cag-NANA in an animal model of H. pylori-associated gastritis and, in vitro, in bacterial cultures.

#### **REFERENCES**

1 Simon PM, Goode PL, Mobasseri A, Zopf D. Inhibition of Helico-

- bacter pylori binding to gastrointestinal epithelial cells by sialic acidcontaining oligosaccharides. *Infect Immun* 1997; **65**: 750-757 [PMID: 9009338]
- 2 Logan RP. Adherence of Helicobacter pylori. Aliment Pharmacol Ther 1996; 10 Suppl 1: 3-15 [PMID: 8730255 DOI: 10.1046/j.1365 -2036.1996.22164001.x]
- Opekun AR, El-Zaimaity HM, Osato MS, Gilger MA, Malaty HM, Terry M, Headon DR, Graham DY. Novel therapies for Helicobacter pylori infection. *Aliment Pharmacol Ther* 1999; 13: 35-42 [PMID: 9892877 DOI: 10.1046/j.1365-2036.1999.00435.x]
- 4 Kigasawa K, Ohtani H. Decomposition and stabilization of drugs. XV. Structure and stability of aminoalkylesters. (7) (author's transl). Yakugaku Zasshi 1976; 96: 6-11 [PMID: 943510]
- 5 Yang JC, Chien CT. A new approach for the prevention and treatment of Helicobacter pylori infection via upregulation of autophagy and downregulation of apoptosis. *Autophagy* 2009; 5: 413-414 [PMID: 19197143]
- 6 Malfertheiner P, Megraud F, O'Morain CA, Atherton J, Axon AT, Bazzoli F, Gensini GF, Gisbert JP, Graham DY, Rokkas T, El-Omar EM, Kuipers EJ. Management of Helicobacter pylori infection--the Maastricht IV/ Florence Consensus Report. *Gut* 2012; 61: 646-664 [PMID: 22491499 DOI: 10.1136/gutjnl-2012-302084]
- 7 Caron TJ, Scott KE, Fox JG, Hagen SJ. Tight junction disruption: Helicobacter pylori and dysregulation of the gastric mucosal barrier. World J Gastroenterol 2015; 21: 11411-11427 [PMID: 26523106 DOI: 10.3748/wjg.v21.i40.11411]
- 8 Burger O, Ofek I, Tabak M, Weiss EI, Sharon N, Neeman I. A high molecular mass constituent of cranberry juice inhibits helicobacter pylori adhesion to human gastric mucus. FEMS Immunol Med Microbiol 2000; 29: 295-301 [PMID: 11118911 DOI: 10.1111/j.1574-695X.2000.tb01537.x]
- 9 Joffe M. Epidemiology of occupational reproductive hazards: methodological aspects. *Rev Epidemiol Sante Publique* 1992; 40 Suppl 1: S17-S25 [PMID: 1626103 DOI: 10.1371/journal.ppat.0020110]
- Teneberg S. The Multiple Carbohydrate Binding Specificities of Helicobacter pylori. *Top Curr Chem* 2009; 288: 121-138 [PMID: 22328028 DOI: 10.1007/128\_2008\_14]
- Wadström T, Hirmo S, Borén T. Biochemical aspects of Helico-bacter pylori colonization of the human gastric mucosa. *Aliment Pharmacol Ther* 1996; 10 Suppl 1: 17-27 [PMID: 8730256 DOI: 10.1046/j.1365-2036.1996.22164002.x]
- Hirmo S, Kelm S, Iwersen M, Hotta K, Goso Y, Ishihara K, Suguri T, Morita M, Wadström T, Schauer R. Inhibition of Helicobacter pylori sialic acid-specific haemagglutination by human gastrointestinal mucins and milk glycoproteins. *FEMS Immunol Med Microbiol* 1998; 20: 275-281 [PMID: 9626932 DOI: 10.1111/j.1574-695X.1998. tb01137 x]
- Falk P, Roth KA, Borén T, Westblom TU, Gordon JI, Normark S. An in vitro adherence assay reveals that Helicobacter pylori exhibits cell lineage-specific tropism in the human gastric epithelium. *Proc Natl Acad Sci USA* 1993; 90: 2035-2039 [PMID: 8383333]
- Sakarya S, Gunay N. Saccharomyces boulardii expresses neuraminidase activity selective for α2,3-linked sialic acid that decreases Helicobacter pylori adhesion to host cells. APMIS 2014; 122: 941-950 [PMID: 24628732 DOI: 10.1111/apm.12237]
- Narod S. Measles vaccination in Haiti. N Engl J Med 1986; 314: 581-582 [PMID: 3945300 DOI: 10.1074/jbc.M113.513135]
- Salcedo J, Barbera R, Matencio E, Alegría A, Lagarda MJ. Gangliosides and sialic acid effects upon newborn pathogenic bacteria adhesion: an in vitro study. *Food Chem* 2013; 136: 726-734 [PMID: 23122120 DOI: 10.1016/j.foodchem.2012.08.078]
- 17 Valkonen KH, Wadström T, Moran AP. Identification of the N-acetylneuraminyllactose-specific laminin-binding protein of Helicobacter pylori. *Infect Immun* 1997; 65: 916-923 [PMID: 9038297]

P- Reviewer: Ierardi E, Pellicano R S- Editor: Qi Y L- Editor: Wang TQ E- Editor: Wu HL





Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.4291/wjgp.v7.i4.307 World J Gastrointest Pathophysiol 2016 November 15; 7(4): 307-313 ISSN 2150-5330 (online) © 2016 Baishideng Publishing Group Inc. All rights reserved.

ORIGINAL ARTICLE

#### **Retrospective Study**

# Microscopic colitis in patients with mild duodenal damage: A new clinical and pathological entity ("lymphocytic enterocolitis")?

Gabriele Antonio Bonagura, Davide Giuseppe Ribaldone, Sharmila Fagoonee, Nicoletta Sapone, Gian Paolo Caviglia, Giorgio Maria Saracco, Marco Astegiano, Rinaldo Pellicano

Gabriele Antonio Bonagura, Davide Giuseppe Ribaldone, Nicoletta Sapone, Giorgio Maria Saracco, Marco Astegiano, Rinaldo Pellicano, Unit of Gastroenterology and Hepatology, Molinette Hospital, 10126 Torino, Italy

Sharmila Fagoonee, Institute for Biostructures and Bioimages-CNR c/o Molecular Biotechnology Center, University of Torino, 10126 Turin, Italy

Gian Paolo Caviglia, Department of Medical Sciences, University of Turin, 10123 Turin, Italy

Giorgio Maria Saracco, Department of Oncology, University of Torino, 10126 Torino, Italy

Author contributions: Bonagura GA and Ribaldone DG equally contributed to this paper; Bonagura GA, Ribaldone DG, Saracco GM, Astegiano M and Pellicano R designed the research; Bonagura GA, Ribaldone DG, Fagoonee S and Sapone N performed the research; Fagoonee S and Caviglia GP analyzed the data; Bonagura GA, Ribaldone DG, Astegiano M and Pellicano R wrote the paper.

Institutional review board statement: This study was reviewed and approved by the Ethics Committee of Molinette Hospital.

Informed consent statement: Patients were not required to give informed consent to the study because the analysis used anonymous data and it was performed several years after the consultation (retrospective).

Conflict-of-interest statement: None to declare.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license,

which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

Manuscript source: Invited manuscript

Correspondence to: Davide Giuseppe Ribaldone, MD, Unit of Gastroenterology and Hepatology, Molinette Hospital, S.G.A.S., Via Cavour 31, 10126 Torino,

Italy. davrib\_1998@yahoo.com Telephone: +39-01-16335208 Fax: +39-01-16336752

Received: June 22, 2016

Peer-review started: June 22, 2016 First decision: September 1, 2016 Revised: September 3, 2016 Accepted: September 21, 2016 Article in press: September 22, 2016 Published online: November 15, 2016

#### **Abstract**

#### **AIM**

To evaluate the potential association between mild duodenal damage and microscopic colitis (MC).

#### **METHODS**

We retrospectively included 105 consecutive patients with type I Marsh-Oberhuber duodenal damage and negativity for immunoglobulin A anti-endomysium and anti-tissue transglutaminase. The following parameters were analyzed: Sex, age at execution of esophagogastroduodenoscopy, duodenal damage, and number of intraepithelial lymphocytes at biopsies, prevalence



of *Helicobacter pylori* infection, age at execution of colonoscopy, macroscopic and microscopic features of colonoscopy, family history of gastrointestinal and autoimmune diseases, smoking habits, biochemical parameters of inflammation and autoimmunity, use of proton pump inhibitors or nonsteroidal anti-inflammatory drugs, adverse reactions to drugs or foods, pathologies known to be associated with celiac disease or MC, living on a gluten-free diet or on a gluten-low diet for at least 1 mo.

#### **RESULTS**

Colonoscopy was performed in 59 patients, but only in 48 of them biopsies were taken in the entire colon. Considering the latter cohort, the diagnosis of MC was met in 25 (52.1%) patients while in 18 patients other pathologic findings were reported: 13 (27%) cases of nonspecific inflammatory bowel disease, 2 (4.2%) cases of Crohn's disease, 2 (4.2%) cases of eosinophilic gastroenteritis, and 1 (2.1%) case of autoimmune enteritis. Five (10.4%) patients had a normal colonoscopic result. Matching the groups by age, and considering only patients who underwent colonoscopy (42.7 ± 15.5 years) vs those who did not undergo colonoscopy  $(36.9 \pm 10.6 \text{ years})$ , a statistical difference was found (P = 0.039). Focusing on symptoms, diarrhea was statistically more prevalent in MC group than in patients who did not undergo colonoscopy (P = 0.03).

#### **CONCLUSION**

Mild duodenal damage is associated with MC in more than half of the cases. This association supports the hypothesis of a link between these two entities.

**Key words:** Autoimmune diseases; Celiac disease; *Helicobacter pylori*; Intraepithelial lymphocytes; Lymphocytic colitis; Lymphocytic enterocolitis; Microscopic colitis

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Scarce information is available on patients with symptoms suggestive for celiac disease but with negative serologic tests and mild duodenal damage (type I Marsh-Oberhuber classification). Our data show that mild duodenal damage is associated with microscopic colitis in more than the half of the investigated cases. This association may support the hypothesis of a new clinical and pathological entity, the "lymphocytic enterocolitis".

Bonagura GA, Ribaldone DG, Fagoonee S, Sapone N, Caviglia GP, Saracco GM, Astegiano M, Pellicano R. Microscopic colitis in patients with mild duodenal damage: A new clinical and pathological entity ("lymphocytic enterocolitis")? *World J Gastrointest Pathophysiol* 2016; 7(4): 307-313 Available from: URL: http://www.wjgnet.com/2150-5330/full/v7/i4/307.htm DOI: http://dx.doi.org/10.4291/wjgp.v7.i4.307

#### INTRODUCTION

Celiac disease (CD) is a chronic inflammatory disease characterized by a pathological reaction against gluten proteins<sup>[1]</sup>. Currently, the prevalence of CD in the general population is proximally 1%, with a ratio between diagnosed and undiagnosed cases of about 1:7<sup>[2,3]</sup>. CD presents often signs and symptoms such as chronic diarrhea, bloating, abdominal pain and malabsorption<sup>[4]</sup>. However, in a substantial number of cases, CD can manifest only extra-intestinal symptoms or signs, and it can be associated with autoimmune pathologies, as autoimmune thyroiditis, type I diabetes mellitus and rheumatoid arthritis<sup>[5,6]</sup>. The diagnosis of CD is based on the finding of positive antibody tests (anti-endomysium and anti-tissue transglutaminase), confirmed by biopsies taken during esophagogastroduodenoscopy (EGD) that reveal the characteristic duodenal damage. The Marsh-Oberhuber classification is usually used to grade the severity of duodenal lesions, with the type III representative of CD<sup>[7]</sup>. The search for human leukocyte antigen (HLA) haplotypes DQ2 and DQ8, due to its high negative predictive value, is used to exclude CD<sup>[8]</sup>. Nevertheless, there are patients with suggestive symptoms of CD, mild duodenal damage [i.e., an increase of intraepithelial lymphocytes (IEL)] defined type I, according to Marsh-Oberhuber classification, and negative antibody tests. This clinical condition, that does not conform with the diagnosis of CD, needs to be investigated for other causes<sup>[9]</sup>.

Microscopic colitis (MC) is a chronic inflammatory bowel disease, distinct in lymphocytic colitis (LC)[10] and collagenous colitis (CC)[11]. The diagnosis of MC is obtained by multiple colonic mucosal biopsies taken during colonoscopy[12]. Typically, in CC, the histological feature is a thickening of the subepithelial collagen layer beneath the basal membrane, of more than 7-10  $\mu m$ (0-3 µm in the normal colon)<sup>[13]</sup>. The histological feature of LC is the presence of more than 20 IEL/100 surface epithelial cells (< 5 IEL/100 in the normal colon)[14]. Paucicellular LC is a term used when the number of IEL is comprised between 5 IEL/100 and 20/100 surface epithelial cells. In MC, IEL are T-Lymphocyte CD3+ and CD8<sup>+</sup>, similar to those described in case of type I Marsh-Oberhuber lesions. Previously considered rare, MC is now a relatively common cause of chronic watery nonbloody diarrhea, especially in the elderly[15]. Both LC and CC are associated with autoimmune diseases and allergy[16]. Finally, it has been shown that patients with MC have an increased rate of HLA-DQ2 and HLA-DQ8 positivity<sup>[17]</sup>, even if this association is less strict than with CD.

Although some authors reported an association between MC and type I Marsh-Oberhuber duodenal damage<sup>[18-23]</sup>, the interpretation of this finding is poorly described. Nevertheless, there are few studies<sup>[24]</sup> that searched for the inverse association.

The aim of this study was to evaluate, for the first time, the association between type I Marsh-Oberhuber



duodenal damage and MC, arguing for the existence of a possible "microscopic enterocolitis" [25,26].

#### **MATERIALS AND METHODS**

We retrospectively included 105 (86 females, mean age  $40.1 \pm 13.7$ ) consecutive patients with type I Marsh-Oberhuber duodenal damage and negativity for antiendomysium (EmA) and anti-tissue transglutaminase (tTG) immunoglobulin (Ig)A antibodies. No sign of Whipple disease were reported in duodenal biopsies. Patients affected by small bowel bacterial overgrowth were excluded from the analysis. The analysis included patients observed in the period 1 January 2003-31 December 2013 in the outpatients clinic of the Unit of Gastroenterology and Hepatology, Molinette Hospital, Turin, Italy.

In 5 cases of IgA deficiency, the genetic assessment (HLA-DQ2/DQ8) was performed: In 3, the result was negative while in the remaining 2 HLA-DQ2 positivity was found.

The following parameters were analyzed: Sex, age at execution of EGD, duodenal damage with number of IEL at biopsies, age at execution of colonoscopy, macroscopic and microscopic features of colonoscopy, family history of gastrointestinal and autoimmune diseases, smoking habits, dosage of erythrocyte sedimentation rate (ESR), C-reactive protein (CRP) and anti-nuclear antibody (ANA), use of proton pump inhibitors (PPIs) or nonsteroidal anti-inflammatory drugs (NSAIDs)[27], adverse reactions to drugs or foods, pathologies associated with CD or MC, living on a glutenfree diet or on a gluten-low diet for at least 1 mo. Data on the prevalence of watery diarrhea, constipation, epigastric pain, abdominal pain, weight loss, nausea and/or vomiting, bloating, and asthenia were collected. Malabsorption was defined as the presence of at least one of these elements: Hemoglobin (Hb) < 12 g/L and low levels of serum iron or folate or vitamin B12; hypoalbuminemia; weight loss > 10% without hypocaloric diet. Helicobacter pylori (H. pylori) infection was investigated by urea breath test and gastric biopsies. The previous eradication treatment of this infection, if any, was reported.

The pharmacological anamnesis for assumption of prednisone, mesalamine, salazopyrin, budesonide, and antibiotics (rifaximin, ciprofloxacin, metronidazole) was conducted. Patients who took serotonin reuptake inhibitors or antiplatelets were excluded from the analysis.

#### Statistical analysis

For parametric data, we initially used the "normal probability plot", to value a normal data distribution; in case of positive return, the Student T test to match the two subgroups was used.

For non-parametric data, the subgroups were matched with the Yates'  $\chi^2$  test or with the Fisher's exact test if data were  $\leq 5$ .

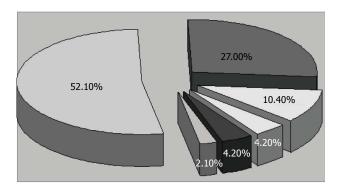
Confidence interval (CI) was set at 95%, with the statistical significance set at P value < 0.05. All statistical analyses were performed using MedCalc software (MedCalc Software, version 9.2.1.0).

#### **RESULTS**

Overall, colonoscopy was performed in 59 patients, but in only 48 cases biopsies were taken along the entire colon. In the remaining 11 cases, biopsies were not taken or taken only in the left colon. The histological findings permitted to divide the cohort into two groups: That including 25 patients with diagnosis of MC and that including 23 patients without MC (Figure 1).

Matching by age, patients who underwent colonoscopy (42.7  $\pm$  15.5 years) vs those who did not undergo colonoscopy (36.9  $\pm$  10.6 years), a statistical difference was found (P = 0.039). On the contrary, there was no significant difference between the group of patients who did not undergo colonoscopy (36.9 ± 10.6 years) vs the group who underwent colonoscopy and executed multiple biopsies (40.4  $\pm$  13.7 years) (P = 0.186). Considering the symptoms, there were no statistical differences between patients who did not undergo colonoscopy vs those who underwent colonoscopy and executed multiple biopsies (P = 0.09 and P = 0.14 for epigastric pain and diarrhea, respectively). Diarrhea was statistically more prevalent in MC group than in patients who did not undergo colonoscopy (P = 0.03). Patients who did not undergo colonoscopy vs those who underwent colonoscopy and executed multiple biopsies had not statistical differences when comparing the heterodimers HLA-DQ2 and HLA-DQ8 (P = 0.19).

Among patients who underwent colonoscopy, an inflammatory pattern was found in 89.6% of cases. Focusing on the 25 patients with MC, the females to males ratio resulted 5:1 and the mean age was 40 ± 16.3 years. The diagnosis was LC in 13 cases, paucicellular LC in 9, CC in 2, and undefined MC in the remaining patient. The average duodenal IEL were 41.6/100 epithelial cells and colonic IELs were 25.4/100 epithelial cells. Watery diarrhea was present in 17/25 (68%) patients, abdominal pain in 16/25 (64%), weight loss in 11/25 (44%), nausea or vomiting in 7/25 (28%), epigastric pain in 6/25 (24%), asthenia in 5/25 (20%), bloating in 4/25 (16%), and gastroesophageal reflux disease (GERD) in 2/25 (8%). A family history of Crohn's disease, thyroiditis, rheumatoid arthritis or spondylitis, was present in 1/25 (4%) patient for each one. Regarding smoking habits, 19/25 (76%) patients were non-smokers while the remaining 6 (24%) were smokers. ANA test resulted positive in 4/25 (16%) patients, ESR increased in 2/25 (8%), and CRP in 2/25 (8%). Four out of twenty-five (16%) patients had a positive history of PPIs use, and 1/25 (4%) of NSAIDs use. Autoimmune thyroiditis was diagnosed in 4/25 (16%) patients, asthma in 3/25 (12%), rheumatoid arthritis in 3/25 (12%). Anamnesis of adverse reactions to drugs or foods resulted in 10/25 (40%) patients.



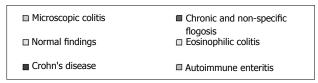


Figure 1 Microscopic findings at colonoscopy (48 cases).

Thirteen patients (52%) had HLA-DQ2 positivity, 8 (32%) HLA-DQ2/DQ8 negativity, and 4 (16%) HLA-DQ8 positivity. Regarding *H. pylori* infection, 19/25 (76%) had negativity *ab initio* while 4 out of 6 with positivity (66.6%), eradicated the infection after antibiotic treatment. None of the tested patients had positivity at coproculture or at parasitological fecal test (6 and 2 cases, respectively). Fourteen (56%) patients undertook a gluten-free diet for at least 1 mo with a clinical improvement in 3/14 (21.4%). Malabsorption was observed in 12 (48%) patients.

Among patients without MC, the female to male ratio resulted 3.8:1 and the mean age was  $40.5 \pm 13.7$ years. The diagnosis was of chronic and non-specific inflammation in 13 (56.5%) cases, there was a normal finding in 5 (21.7%), eosinophilic colitis in 2 (8.6%), Crohn's disease in 2 (8.6%), and autoimmune enteritis in the last one (4.3%). The average duodenal IEL resulted 42.1/100 epithelial cells. Based on the available data, a family history of Crohn's disease, rheumatoid arthritis or spondylitis was reported in one out of 23 (4.3%) patients for each disease. Regarding smoking habits, 15/23 (65.2%) patients were non-smokers while the remaining 8 (34.8%) were smokers. Abdominal pain was present in 10/23 (43.4%) patients, watery diarrhea in 10/23 (43.4%), epigastric pain in 5/23 (21.7%), bloating in 5/23 (21.7%), asthenia in 4/23 (17.3%), nausea or vomiting in 4/23 (17.3%), GERD in 4/23 (17.3%), constipation in 4/23 (17.3%), and weight loss in 3/23 (13.0%). ANA test resulted positive in 8/23 (34.8%) patients, ESR and CPR was increased in 6/23 (26.1%) and 5/23 (21.7%), respectively. A history of PPIs or NSAIDs use was reported in 6/23 (26.1%) and no patient, respectively. Autoimmune thyroiditis was reported in 3/23 (13%) of the patients, asthma in 2/23 (8.6%), rheumatoid arthritis in 2/23 (8.6%), systemic erythematosus lupus (SLE), autoimmune hepatitis and multiple autoimmune diseases in 1/23 (4.3%) for each one. Adverse reactions to drugs or foods resulted in

Table 1 Main clinical and laboratory parameters of enrolled patients

	Patients with MC	Patients without MC	P value
Watery diarrhea	17/25 (68%)	10/23 (43%)	0.08
Abdominal pain	16/25 (64%)	10/23 (43%)	0.15
Weight loss	11/25 (44%)	3/23 (13%)	0.01
Nausea/vomiting	7/25 (28%)	4/23 (17%)	0.29
Epigastric pain	6/25 (24%)	5/23 (21%)	0.85
Asthenia	5/25 (20%)	4/23 (17%)	0.55
Bloating	4/25 (16%)	5/23 (21%)	0.44
GERD	2/25 (8%)	4/23 (17%)	0.33
Constipation	0/25 (0%)	4/23 (17%)	0.04
History of allergy	10/25 (40%)	10/23 (43%)	0.40
PPIs use	4/25 (16%)	6/23 (26%)	0.44
NSAIDs use	1/25 (4%)	0/23 (0%)	0.34
ANA positivity	4/25 (16%)	8/23 (34%)	0.46
ESR increased	2/25 (8%)	6/23 (26%)	0.13
CRP increased	2/25 (8%)	5/23 (21%)	0.21
Helicobacter	6/25 (24%)	8/23 (34%)	0.61
pylori infection			

MC: Microscopic colitis; GERD: Gastroesophageal reflux disease; PPIs: Proton pump inhibitors; NSAIDs: Nonsteroidal anti-inflammatory drugs; ANA: Anti-nuclear antibody; ESR: Erythrocyte sedimentation rate; CRP: C-reactive protein.

10/23 (43.4%) patients. Ten (43.4%) patients had HLA-DQ2/DQ8 negativity, 9 (39.1%) had HLA-DQ2 positivity, 3 (13%) HLA-DQ8 positivity, and one (4.3%) had HLA-DQ2/DQ8 positivity. Regarding *H. pylori* infection, 15/23 (65.2%) of the tested patients were negative *ab initio*, while 8/23 (34.8%) were positive. Of the latter group, 5 out of 8 (62.5%) eradicated the infection after antibiotic treatment. None of the tested patients had positivity at coproculture or at parasitological fecal test (4 and 1 case, respectively).

Comparing patients with MC vs those without MC (Table 1), the only variables that had a statistical difference were weight loss (P=0.01), more frequent in case of MC, and constipation (P=0.04) more frequent in absence of MC. Diarrhea (P=0.08), abdominal pain (P=0.15), epigastric pain (P=0.85), GERD (P=0.33), autoimmune thyroiditis (P=0.79), smoking habits (P=0.66), asthma (P=0.72), rheumatoid arthritis (P=0.72), autoimmune hepatitis (P=0.31), multiple autoimmune diseases (P=0.35), HLA-DQ2 positivity (P=0.51), HLA-DQ8 positivity (P=0.79) did not reach statistical significance.

Among patients suffering from MC, budesonide was used in 14 patients, of whom 13 (92.9%) responded to therapy; 8 patients used mesalamine, of them 4 (50%) responded to therapy; 4 patients used salazopyrin, with response in 2 (50%); 1 patient used prednisone, with response. No difference about the response to therapy resulted from the comparison between budesonide and mesalamine (P = 0.27), budesonide and salazopyrin (P = 0.41), budesonide and prednisone (P = 0.94). Among patients without MC, 7 used budesonide and 6/7 (85.7%) responded to therapy; 4 patients used prednisone without response; 6 patients used mesalamine with response in 3 (50%).

#### DISCUSSION

In this study, we found a strong association between type I Marsh-Oberhuber duodenal damage and MC, mainly LC. More than half (52.1%) of the patients who underwent colonoscopy with multiple biopsies had MC. This percentage is significantly higher than the historical prevalence of MC in the general population (0.5%)<sup>[28]</sup>. An intriguing data was that LC and paucicellular LC, considered together, were diagnosed in much more cases than CC (22 vs 2, respectively). Usually, literature considered incidence and prevalence of CC higher than LC; however, more recent studies, according to our data, report that the incidence of LC is significantly rising<sup>[29]</sup>.

Patients who underwent colonoscopy were significantly older than those who did not undergo colonoscopy. This could be partially explained considering the age as a parameter associated to augmented risk of malignancy. Hence, clinicians recurred to endoscopy in case of unexplained symptoms and increasing age. However, there was no difference in the median age between patients who did not undergo colonoscopy *vs* those who underwent colonoscopy with multiple biopsies.

Considering biochemical results and symptoms among various groups, only chronic diarrhea was significantly higher in patients with MC than in those who did not undergo colonoscopy (P = 0.03). Thus, in patients with mild duodenal damage only this symptom could predict MC. However, due to its multifactorial pathogenesis, the presence of diarrhea cannot be the only element to decide whether this type of patients should undergo colonoscopy with multiple biopsies. On the other hand, the absence of diarrhea cannot exclude the indication for colonoscopy with multiple biopsies, because only 5 out of 48 (10.4%) patients who had a colonoscopy with biopsies had normal microscopic findings, despite suffering also from abdominal pain, weight loss, constipation, positive fecal occult blood. The search for HLA-DQ2/DQ8 haplotypes seems to be useful, although the data in our retrospective study are not broad enough to provide definitive conclusions. Since this test has a very high negative predictive value in the diagnosis of CD, in patients with mild duodenal damage, negative serological tests for CD and the above reported symptoms, the negativity of HLA-DQ2/ DQ8 haplotypes can definitively exclude this disease and propel to search for other etiologies, as MC.

The median age at which the diagnosis of MC was made in our patients (40 years) is lower than literature reports. Such finding may contrast with the idea of MC as disease of the elderly pointing out to a possible underestimation of this condition.

Another element that emerged from this study was the low prevalence of *H. pylori*-infection both in patients with MC (24%) than without MC (34%). The literature reports that *H. pylori* infection is related to duodenal lymphocytosis<sup>[9]</sup>, which disappears after bacterial eradication. At the same time, we have recently found an inverse association between MC and *H. pylori* 

infection<sup>[29]</sup>. The results of the present study agree with the fact that in case of mild duodenal damage and MC the prevalence of *H. pylori* infection is lower than the general population (in our case 24% *vs* 47%)<sup>[30]</sup>. Moreover, the rate of *H. pylori* eradication in this context, is similar to that obtained in the general population<sup>[31]</sup>.

In our study, the role of pharmacological therapy in the pathogenesis of MC is not fully clear. In fact, only 4 patients used PPIs and 1 patient used NSAIDs before the diagnosis of MC. This differs from the well-known data reporting that this type of medications are often implicated as a cause of MC $^{[32]}$ , and could be explained by a  $\beta$  error (i.e., the failure to detect an effect that is present) due to the small sample size. Considering the outcome of therapy used to treat MC, budesonide emerged as the best treatment, due to a clinical improvement, in more than 90% of patients. Mesalamine seemed to be a valid therapeutic approach for less severe cases. According to some reports, our results confirm the appropriateness of this management  $^{[32]}$ .

Although in literature an association between MC and malabsorption is not reported<sup>[33]</sup>, in our study 12 (48%) patients presented signs of it. A potential disease of the small intestine, beyond the duodenum, could explain these features. More efforts are thus needed to understand this clinical condition.

This retrospective analysis shows inadequate habits of clinicians to search for a coproculture or a parasitological test; also the search for *Giardia Lamblia* was out of routine. Such investigations should play an important role in the attempt to identify the cause of duodenal damage. In fact, literature reports that the search for *Giardia Lamblia* or other pathogens should be included in the diagnostic work up of type I Marsh-Oberhuber duodenal damage<sup>[34]</sup>.

A potential limitation of our study is its retrospective design with a theoretical loss of balance on parameters analyzed. Nevertheless, we noticed uniform diagnostic and follow-up criteria.

In conclusion, MC is frequently associated with mild duodenal damage. This association may suggest the existence of a "microscopic enterocolitis", and specifically of a "lymphocytic enterocolitis", that involves the entire gastrointestinal tract. It is advisable to perform a colonoscopy with biopsies in all patients with type I Marsh-Oberhuber duodenal damage and symptoms as chronic diarrhea, abdominal or epigastric pain, loss of weight, after exclusion of standard causes.

#### **COMMENTS**

#### Background

The diagnosis of celiac disease (CD) is based on the finding of positive antibody tests (anti-endomysium and anti-tissue transglutaminase), confirmed by biopsies that reveal the characteristic duodenal damage. The Marsh-Oberhuber classification is usually used to grade the severity of duodenal lesions, with the type III representative of CD. Nevertheless, there are patients with suggestive symptoms of CD, mild duodenal damage [i.e., an increase of intraepithelial lymphocytes (IEL)] defined type I, according to Marsh-Oberhuber classification,



WJGP | www.wjgnet.com 311 November

and negative antibody tests. This clinical condition, that does not conform with the diagnosis of CD, needs to be investigated for other causes as well as for comorbidities. Microscopic colitis (MC), previously considered rare, was demonstrated as a relatively common cause of chronic, watery, diarrhoea. While some isolated studies reported some association between MC and Marsh I duodenal damage, the interpretation of this finding is poorly described.

#### Research frontiers

To date, scarce information is available on the association between mild duodenal damage and MC.

#### Innovations and breakthroughs

This study is the first showing that type I Marsh-Oberhuber duodenal damage is strongly associated with MC, mainly lymphocytic colitis (LC). More than half (52.1%) of the patients who underwent colonoscopy with multiple biopsies had MC. This percentage is significantly higher than prevalence of MC in the general population (0.5%). This association supports the hypothesis of a link between these two entities.

#### **Applications**

These findings, of association between type I Marsh-Oberhuber duodenal damage and MC, may suggest the existence of a "microscopic enterocolitis", and specifically of a "lymphocytic enterocolitis", that involves the entire gastrointestinal tract. It is advisable to perform a colonoscopy with biopsies in all patients with type I Marsh-Oberhuber duodenal damage and symptoms as chronic diarrhea, abdominal or epigastric pain, loss of weight, after exclusion of standard causes.

#### **Terminology**

The Marsh-Oberhuber classification is usually used to grade the severity of duodenal lesions, with the type III representative of CD. There are patients with suggestive symptoms of CD, mild duodenal damage (*i.e.*, an increase of IEL) defined type I, according to Marsh-Oberhuber classification, and negative antibody tests, that do not conform with the diagnosis of CD. MC is a chronic inflammatory bowel disease, distinct in LC and collagenous colitis. The histological feature of LC is the presence of more than 20 IEL/100 surface epithelial cells (< 5 IEL/100 in the normal colon). Paucicellular LC is a term used when the number of IEL is comprised between 5 IEL/100 and 20/100 surface epithelial cells. In MC, IEL are T-Lymphocyte CD3\* and CD8\*, similar to those described in case of type I Marsh-Oberhuber lesions. Here we report for the first time the association between type I Marsh-Oberhuber duodenal damage and MC, arguing for the existence of a possible "microscopic enterocolitis".

#### Peer-review

This is an interesting manuscript.

#### **REFERENCES**

- Kagnoff MF. Celiac disease: pathogenesis of a model immunogenetic disease. J Clin Invest 2007; 117: 41-49 [PMID: 17200705 DOI: 10.1172/jci30253]
- Fasano A, Berti I, Gerarduzzi T, Not T, Colletti RB, Drago S, Elitsur Y, Green PH, Guandalini S, Hill ID, Pietzak M, Ventura A, Thorpe M, Kryszak D, Fornaroli F, Wasserman SS, Murray JA, Horvath K. Prevalence of celiac disease in at-risk and not-at-risk groups in the United States: a large multicenter study. Arch Intern Med 2003; 163: 286-292 [PMID: 12578508 DOI: 10.1001/archinte.163.3.286]
- 3 Corazza GR, Frisoni M, Treggiari EA, Valentini RA, Filipponi C, Volta U, Gasbarrini G. Subclinical celiac sprue. Increasing occurrence and clues to its diagnosis. *J Clin Gastroenterol* 1993; 16: 16-21 [PMID: 8421137 DOI: 10.1097/00004836-199301000-00 006]
- 4 Casella G, Di Bella C, Salemme M, Villanacci V, Antonelli E, Baldini V, Bassotti G. Celiac disease, non-celiac gluten sensitivity and inflammatory bowel disease. *Minerva Gastroenterol Dietol* 2015; 61: 267-271 [PMID: 26006779]

- Pellicano R, De Angelis C, Ribaldone DG, Fagoonee S, Astegiano M. 2013 update on celiac disease and eosinophilic esophagitis. Nutrients 2013; 5: 3329-3336 [PMID: 23974065 DOI: 10.3390/nu5093329]
- 6 Ribaldone DG, Astegiano M, Fagoonee S, Rizzetto M, Pellicano R. Epilepsy and celiac disease: review of literature. *Panminerva Med* 2011; 53: 213-216 [PMID: 22146418]
- Oberhuber G, Granditsch G, Vogelsang H. The histopathology of coeliac disease: time for a standardized report scheme for pathologists. *Eur J Gastroenterol Hepatol* 1999; 11: 1185-1194 [PMID: 10524652 DOI: 10.1097/00042737-199910000-00019]
- 8 Green PH, Jabri B. Coeliac disease. *Lancet* 2003; **362**: 383-391 [PMID: 12907013]
- Simondi D, Ribaldone DG, Bonagura GA, Foi S, Sapone N, Garavagno M, Villanacci V, Bernardi D, Pellicano R, Rizzetto M, Astegiano M. Helicobacter pylori in celiac disease and in duodenal intraepithelial lymphocytosis: Active protagonist or innocent bystander? Clin Res Hepatol Gastroenterol 2015; 39: 740-745 [PMID: 25956489 DOI: 10.1016/j.clinre.2015.03.005]
- 10 Lindström CG. 'Collagenous colitis' with watery diarrhoea--a new entity? Pathol Eur 1976; 11: 87-89 [PMID: 934705]
- Lazenby AJ, Yardley JH, Giardiello FM, Jessurun J, Bayless TM. Lymphocytic ("microscopic") colitis: a comparative histopathologic study with particular reference to collagenous colitis. *Hum Pathol* 1989; 20: 18-28 [PMID: 2912870 DOI: 10.1016/0046-8177(89)901 98-6]
- 12 Carpenter HA, Tremaine WJ, Batts KP, Czaja AJ. Sequential histologic evaluations in collagenous colitis. Correlations with disease behavior and sampling strategy. *Dig Dis Sci* 1992; 37: 1903-1909 [PMID: 1361906 DOI: 10.1007/bf01308086]
- Tanaka M, Mazzoleni G, Riddell RH. Distribution of collagenous colitis: utility of flexible sigmoidoscopy. *Gut* 1992; 33: 65-70 [PMID: 1740280 DOI: 10.1136/gut.33.1.65]
- 14 Fasoli R, Talbot I, Reid M, Prince C, Jewell DP. Microscopic colitis: can it be qualitatively and quantitatively characterized? *Ital J Gastroenterol* 1992; 24: 393-396 [PMID: 1392021]
- Olesen M, Eriksson S, Bohr J, Järnerot G, Tysk C. Microscopic colitis: a common diarrhoeal disease. An epidemiological study in Orebro, Sweden, 1993-1998. *Gut* 2004; 53: 346-350 [PMID: 14960513 DOI: 10.1136/gut.37.3.394]
- 16 Roth B, Manjer J, Ohlsson B. Microscopic Colitis is Associated with Several Concomitant Diseases. *Drug Target Insights* 2013; 7: 19-25 [PMID: 24003301 DOI: 10.4137/DTI.S12109]
- 17 Fernández-Bañares F, Esteve M, Farré C, Salas A, Alsina M, Casalots J, Espinós J, Forné M, Viver JM. Predisposing HLA-DQ2 and HLA-DQ8 haplotypes of coeliac disease and associated enteropathy in microscopic colitis. Eur J Gastroenterol Hepatol 2005; 17: 1333-1338 [PMID: 16292086 DOI: 10.1097/00042737-2 00512000-00011]
- Matteoni CA, Goldblum JR, Wang N, Brzezinski A, Achkar E, Soffer EE. Celiac disease is highly prevalent in lymphocytic colitis. J Clin Gastroenterol 2001; 32: 225-227 [PMID: 11246349 DOI: 10.1097/00004836-200103000-00009]
- Thijs WJ, van Baarlen J, Kleibeuker JH, Kolkman JJ. Microscopic colitis: prevalence and distribution throughout the colon in patients with chronic diarrhoea. *Neth J Med* 2005; 63: 137-140 [PMID: 15869041]
- 20 Geboes K. Lymphocytic, collagenous and other microscopic colitides: pathology and the relationship with idiopathic inflammatory bowel diseases. *Gastroenterol Clin Biol* 2008; 32: 689-694 [PMID: 18538968 DOI: 10.1016/j.gcb.2008.04.021]
- 21 Aziz I, Evans KE, Hopper AD, Smillie DM, Sanders DS. A prospective study into the aetiology of lymphocytic duodenosis. Aliment Pharmacol Ther 2010; 32: 1392-1397 [PMID: 21050242 DOI: 10.1111/j.1365-2036.2010.04477.x]
- 22 Shmidt E, Smyrk TC, Boswell CL, Enders FT, Oxentenko AS. Increasing duodenal intraepithelial lymphocytosis found at upper endoscopy: time trends and associations. *Gastrointest Endosc* 2014; 80: 105-111 [PMID: 24565068 DOI: 10.1016/j.gie.2014.01.008]
- 23 Losurdo G, Piscitelli D, Giangaspero A, Principi M, Buffelli F,



- Giorgio F, Montenegro L, Sorrentino C, Amoruso A, Ierardi E, Di Leo A. Evolution of nonspecific duodenal lymphocytosis over 2 years of follow-up. *World J Gastroenterol* 2015; **21**: 7545-7552 [PMID: 26140001 DOI: 10.3748/wjg.v21.i24.7545]
- 24 Astegiano M, Pellicano R, Verme G, Rizzetto M. High rate of microscopic colitis in patients with Marsh I-II duodenal damage. *Scand J Gastroenterol* 2009; 44: 1266-1267 [PMID: 19658019 DOI: 10.1080/00365520903144414]
- 25 Rostami K, Villanacci V. Microscopic enteritis: novel prospect in coeliac disease clinical and immuno-histogenesis. Evolution in diagnostic and treatment strategies. *Dig Liver Dis* 2009; 41: 245-252 [PMID: 18657490 DOI: 10.1016/j.dld.2008.06.008]
- 26 Rostami K, Aldulaimi D, Holmes G, Johnson MW, Robert M, Srivastava A, Fléjou JF, Sanders DS, Volta U, Derakhshan MH, Going JJ, Becheanu G, Catassi C, Danciu M, Materacki L, Ghafarzadegan K, Ishaq S, Rostami-Nejad M, Peña AS, Bassotti G, Marsh MN, Villanacci V. Microscopic enteritis: Bucharest consensus. World J Gastroenterol 2015; 21: 2593-2604 [PMID: 25759526 DOI: 10.3748/wjg.v21.i9.2593]
- 27 Beaugerie L, Pardi DS. Review article: drug-induced microscopic colitis proposal for a scoring system and review of the literature. Aliment Pharmacol Ther 2005; 22: 277-284 [PMID: 16097993 DOI: 10.1111/j.1365-2036.2005.02561.x]
- 28 Tong J, Zheng Q, Zhang C, Lo R, Shen J, Ran Z. Incidence, prevalence, and temporal trends of microscopic colitis: a systematic review and meta-analysis. Am J Gastroenterol 2015; 110: 265-276;

- quiz 277 [PMID: 25623658 DOI: 10.1038/ajg.2014.431]
- Ribaldone DG, Simondi D, Astegiano M, Pellicano R. On Inverse Association Between Helicobacter pylori Gastritis and Microscopic Colitis: The European Data. *Inflamm Bowel Dis* 2016; 22: E11-E12 [PMID: 26871398 DOI: 10.1097/MIB.00000000000000704]
- 30 Ponzetto A, Pellicano R, Morgando A, Cirillo D, Marchiaro G, Curti F, Rizzetto M. Seroprevalence of Helicobacter pylori infection among blood donors in Torino, Italy. *Minerva Gastroenterol Dietol* 2001; 47: 3-7 [PMID: 16491063]
- Ribaldone DG, Fagoonee S, Astegiano M, Saracco G, Pellicano R. Efficacy of amoxycillin and clarithromycin-based triple therapy for Helicobacter pylori eradication: a 10-year trend in Turin, Italy. *Panminerva Med* 2015; 57: 145-146 [PMID: 25971330]
- 32 Park T, Cave D, Marshall C. Microscopic colitis: A review of etiology, treatment and refractory disease. World J Gastroenterol 2015; 21: 8804-8810 [PMID: 26269669 DOI: 10.3748/wjg.v21.i29.8804]
- 33 Mellander MR, Ekbom A, Hultcrantz R, Löfberg R, Öst Å, Björk J. Microscopic colitis: a descriptive clinical cohort study of 795 patients with collagenous and lymphocytic colitis. Scand J Gastroenterol 2016; 51: 556-562 [PMID: 26679722 DOI: 10.3109/ 00365521.2015.1124283]
- Patterson ER, Shmidt E, Oxentenko AS, Enders FT, Smyrk TC. Normal villous architecture with increased intraepithelial lymphocytes: a duodenal manifestation of Crohn disease. Am J Clin Pathol 2015; 143: 445-450 [PMID: 25696804 DOI: 10.1309/AJCPBKQND4SHVX9Q]

P- Reviewer: Bonaz B S- Editor: Ji FF L- Editor: A E- Editor: Wu HL





Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.4291/wjgp.v7.i4.314 World J Gastrointest Pathophysiol 2016 November 15; 7(4): 314-319 ISSN 2150-5330 (online) © 2016 Baishideng Publishing Group Inc. All rights reserved.

META-ANALYSIS

# Hepatitis C infection and renal cell carcinoma: A systematic review and meta-analysis

Karn Wijarnpreecha, Pitchaphon Nissaisorakarn, Suthanya Sornprom, Charat Thongprayoon, Natanong Thamcharoen, Kunlatida Maneenil, Alexander J Podboy, Wisit Cheungpasitporn

Karn Wijarnpreecha, Suthanya Sornprom, Charat Thongprayoon, Natanong Thamcharoen, Department of Internal Medicine, Bassett Medical Center, Cooperstown, NY 13326, United States

Pitchaphon Nissaisorakarn, Department of Internal Medicine, Jacobi Medical Center, Albert Einstein College of Medicine, Bronx, NY 10461, United States

Kunlatida Maneenil, Division of Medical Oncology, Mayo Clinic, Rochester, MN 55905, United States

Alexander J Podboy, Department of Internal Medicine, Mayo Clinic, Rochester, MN 55905, United States

Wisit Cheungpasitporn, Division of Nephrology and Hypertension, Department of Internal Medicine, Mayo Clinic, Rochester, MN 55905, United States

Author contributions: All authors had access to the data and a role in writing the manuscript.

Conflict-of-interest statement: The authors deny any conflict of interest.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

Manuscript source: Invited manuscript

Correspondence to: Wisit Cheungpasitporn, MD, Division of Nephrology and Hypertension, Department of Internal Medicine, Mayo Clinic, 200 First Street SW, Rochester, MN 55905,

United States. wcheungpasitporn@gmail.com

Telephone: +1-507-2848450 Fax: +1-507-2667891 Received: April 6, 2016

Peer-review started: April 8, 2016 First decision: May 19, 2016 Revised: July 30, 2016 Accepted: August 17, 2016 Article in press: August 18, 2016 Published online: November 15, 2016

#### **Abstract**

#### AIM

To investigate the association between hepatitis C virus (HCV) infection and risk of renal cell carcinoma (RCC).

#### **METHODS**

A literature search was performed from inception until February 2016. Studies that reported relative risks, odd ratios, hazard ratios or standardized incidence ratio comparing the risk of RCC among HCV-infected participants  $\nu s$  those without HCV infection were included. Participants without HCV infection were used as comparators. Pooled odds ratios and 95%CI were calculated using a random-effect, generic inverse variance method.

#### RESULTS

Seven observational studies were with 196826 patients were included in the analysis to assess the risk of RCC in patients with HCV. A significantly increased risk of RCC among participants with HCV infection was found with a pooled RR of 1.86 (95%CI: 1.11-3.11). The association between RCC and HCV was marginally insignificant after a sensitivity analysis limited only to studies with adjusted analysis, with a pooled RR of 1.50 (95%CI: 0.93-2.42).

#### **CONCLUSION**

Our study demonstrated a potential association between HCV infection and RCC. Further studies of RCC



surveillance in patients with HCV are required.

**Key words:** Hepatitis C virus; Renal cancer; Kidney cancer; Systematic review; Meta-analysis

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Hepatitis C virus (HCV) is a leading cause of cirrhosis in the United States with a steadily increasing prevalence over the past two decades. Interestingly, HCV infection may also be associated with an increased risk of renal cell carcinoma (RCC) as observed in several epidemiologic studies. To further investigate this possible association, we conducted this systematic review and meta-analysis of observational studies reporting the risk of RCC among HCV-infected patients. We found a significantly increased risk of RCC among participants with HCV infection with the pooled risk ratio of 1.86 (95%CI: 1.11-3.11).

Wijarnpreecha K, Nissaisorakarn P, Sornprom S, Thongprayoon C, Thamcharoen N, Maneenil K, Podboy AJ, Cheungpasitporn W. Hepatitis C infection and renal cell carcinoma: A systematic review and meta-analysis. *World J Gastrointest Pathophysiol* 2016; 7(4): 314-319 Available from: URL: http://www.wjgnet.com/2150-5330/full/v7/i4/314.htm DOI: http://dx.doi.org/10.4291/wjgp.v7.i4.314

#### INTRODUCTION

Hepatitis C virus (HCV) remains the most common cause of chronic liver disease and cirrhosis worldwide and is one of the leading causes of chronic hepatitis and cirrhosis in the United States with a steadily increasing prevalence over the past two decades<sup>[1,2]</sup>. While commonly associated with hepatocellular carcinoma<sup>[2]</sup>, hepatitis C infection is associated with extrahepatic malignancies including cholangiocarcinoma, non-Hodgkin's lymphoma and possibly myeloma<sup>[3-5]</sup>. The oncogenic properties of hepatitis C are hypothesized to be secondary to chronic antigenic stimulation of the immune system, promotion of a chronic inflammatory state or direct oxidative stress<sup>[6]</sup>. Besides, chronic HCV infection has also been linked to a myriad of extrahepatic diseases including increasing the risk of renal disease and increasing the prevalence of chronic kidney disease up to 40% higher compared to non-infected patients<sup>[7,8]</sup>.

Renal cell carcinoma (RCC), arising from the renal cortex is responsible for 80% of all renal malignancies and accounts for approximately 14000 deaths each year in the United States<sup>[9]</sup>. The risk factors for RCC such as acquired cystic kidney disease, smoking, kidney stones, obesity, hereditary factors have been described<sup>[10-14]</sup>. The epidemiological studies have demonstrated an increasing incidence of RCC, particularly in African Americans<sup>[15]</sup>.

Secondary to the oncogenic nature of Hepatitis C,

several studies have linked chronic infection with an increased risk for development of RCC<sup>[16-22]</sup>. However, the findings from these studies were contradictory. Thus, we performed this meta-analysis to examine the risk of RCC in HCV-infected patients.

#### **MATERIALS AND METHODS**

#### Literature search

Two investigators (Karn Wijarnpreecha and Wisit Cheungpasitporn) independently reviewed published studies indexed in MEDLINE, EMBASE, and the Cochrane database from their inception to February 2016 using the search strategy that included the terms for "hepatitis" and "renal cancer" as described in Item S1 in online supplementary data. No limitation on language was applied. A manual search for additional studies using references of selected retrieved articles was also performed. Three investigators (Karn Wijarnpreecha, Charat Thongprayoon and Wisit Cheungpasitporn) independently reviewed the titles and abstracts of the studies identified in the search based on inclusion and exclusion criteria. The full text of the included studies from the first phase was reviewed independently to ascertain whether or not they matched the inclusion criteria. We also performed a manual search of conference proceedings from major gastroenterology and hepatology meetings for additional abstracts on the topic. When additional information was needed, we contacted the corresponding investigators of eligible studies.

#### Study selection

The inclusion criteria were as follows: (1) observational studies assessing the association between hepatitis C and RCC; (2) odds ratios, relative risks or hazard ratios with 95%CI were provided; and (3) individuals without HCV infection were used as comparators in cohort studies while individuals without RCC were applied as comparators in the cross-sectional and case-control studies.

Study acceptability was individually defined by the three investigators mentioned above. Disagreements in the ascertainment of study eligibility were settled by joint agreement. Also, the quality of each study was individually appraised by each investigator. We used the validated Newcastle-Ottawa quality assessment scale for cohort and case-control studies<sup>[23]</sup> and modified Newcastle-Ottawa scale<sup>[24]</sup> for the cross-sectional study.

#### Data extraction

A data collection report was utilized to derive the information from each study including name of title and the first author, year of study and publication, country, demographic data of the participants, number of participants, method used to diagnose the HCV infection and RCC, effect estimates (odds ratios, relative risks or hazard ratios) with 95%CI, and factors adjusted in the multivariate analysis. To assure the certainty, this data



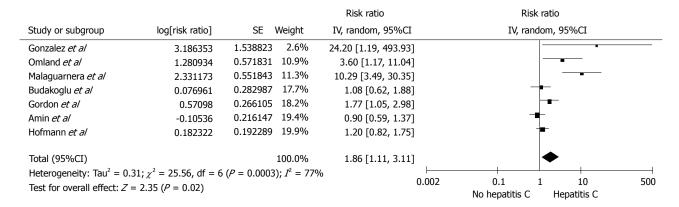


Figure 1 Forest plot of all included studies of the association between hepatitis C infection and renal cell carcinoma. Square data markers represent risk ratios (RRs); horizontal lines, the 95%Cl with marker size reflecting the statistical weight of the study using random-effects meta-analysis. A diamond data marker represents the overall RR and 95%Cl for the outcome of interest.

			Risk ratio	Risk ratio
Study or subgroup	log[risk ratio]	SE Weight	IV, random, 95%CI	IV, random, 95%CI
Gonzalez et al	3.186353	1.538823 2.4%	24.20 [1.19, 493.93]	-
Omland et al	1.280934	0.571831 12.3%	3.60 [1.17, 11.04]	
Gordon et al	0.57098	0.266105 25.9%	1.77 [1.05, 2.98]	<b></b>
Amin <i>et al</i>	-0.10536	0.216147 29.0%	0.90 [0.59, 1.37]	<del>-</del>
Hofmann <i>et al</i>	0.182322	0.192289 30.4%	1.20 [0.82, 1.75]	<del> </del>
Total (95%CI)		100.0%	1.50 [0.93, 2.42]	•
Heterogeneity: $Tau^2 = 0$ .	.16; $\chi^2 = 11.17$ , df = 4	$(P = 0.02); I^2 = 64\%$		
Test for overall effect: $Z$			0.002	No hepatitis C Hepatitis C 500

Figure 2 Forrest plot of all included studies in sensitivity analysis of the association between hepatitis C infection and renal cell carcinoma. Square data markers represent risk ratios (RRs); horizontal lines, the 95%CIs with marker size reflecting the statistical weight of the study using random-effects meta-analysis. A diamond data marker represents the overall RR and 95%CI for the outcome of interest.

extraction process was reviewed by all investigators.

#### Statistical analysis

Review Manager (RevMan) 5.3 software from the Cochrane Collaboration was utilized for meta-analysis. Generic inverse variance (DerSimonian and Laird) method was employed to combined adjusted point estimates and standard errors from each study. We used a random-effect model due to the high likelihood of between-study variance from different study designs and populations. Cochran's Q test and  $I^2$  statistic were used to determine the between-study heterogeneity. A value of  $I^2$  of 0%-25%, 25%-50%, 50%-75%, and greater than 75% embodied insignificant, low, moderate and high heterogeneity, respectively [26].

#### **RESULTS**

Of 5778 potentially relevant articles, 5582 articles were excluded by the title and abstract not fulfilling inclusion criteria due to the type of article, study design, population, or outcome of interest. Additionally, 189 articles were excluded (35 articles were not observational studies, and 154 articles did not describe the outcomes of interest, Macleod *et al*<sup>[27]</sup>'s study did not contain data on specific viral hepatitis{Macleod, 2013 #83}). Finally, seven observational studies (4 cohort and 3 case-

controlled studies) with 196826 patients were included in the meta-analysis<sup>[16-22]</sup>. Item S2 describes the study selection flow. The characteristics and quality appraisal of the included studies of HCV and RCC are shown in Table 1. Four studies were conducted in Europe, 2 in the United States, and 1 in Australia.

# The risk of renal cell carcinoma in patients with hepatitis C virus infection

Among individuals with HCV infection, there was a significantly increased risk of RCC with the pooled risk ratio (RR) of 1.86 (95%CI: 1.11-3.11,  $I^2$  =77%), as demonstrated in Figure 1. The statistical heterogeneity was high with an  $I^2$  of 77%. The association between RCC and HCV was marginally insignificant after the sensitivity analysis including only studies with confounder adjustment<sup>[16,18-20,22]</sup> with a pooled RR of 1.50 (95%CI: 0.93-2.42,  $I^2$  =64%), as shown in Figure 2.

#### Evaluation for publication bias

Two authors (Karn Wijarnpreecha and Wisit Cheungpasitporn) independently performed the assessment of the risk of bias of the included studies. Only minor disagreements between 2 reviewers were present and were resolved by discussion and consensus. A funnel plot was constructed to assess publication bias for the risk of RCC in HCV-infected patients (Figure S1). The



Ref.	Country	Study design	Year	Total	Ref. Country Study Year Total Study sample design number	Exposure	Exposure measurement	Outcome definition	Outcome	Adjusted OR	Confounder adjustment	Quality assessment (Newcastle- Ottawa scale)
Amin et al <sup>[16]</sup>	Australia	Australia Cohort 2006 study		75834 in HCV patients	People notified with HCV infection to the New South Wales Health Department's Notifiable Diseases Database between 1 January 1990 and 31 December 2002 (HCV group)	HCV infection	Detection of anti-HCV antibody or HCV RNA	Renal cancer or kidney cancer	The NSW Central Cancer Registry with ICD-10 code C64 for kidney cancer and C65 for renal cancer	Kidney cancer 0.9 (0.6-1.4)	Age, sex and calendar year	Selection: 4 Comparability: 1 Outcome: 3
Malagu arnera <i>et af</i> <sup>[21]</sup>	Italy	Case control study	2006 3 a	315 (15 case and 300 control)	and NSW population (control group)  Elderly kidney cancer patients Positive attending geriatric department (case) Anti-HCV and elderly volunteers (control)		Detection of Anti-HCV antibody using enzyme- linked immunosorbent assay. Assay positive samples were confirmed by	Kidney	N/A	10.29 (3.49-30.36)	None	Selection: 3 Comparability: 0 Outcome: 2
Omland $et  at^{ 22 }$	Denmark	Denmark Cohort 2010 4204 in study HCV patients	2010 '		Acut patients Hospita 2003 v	Acute or chronic HCV infection	immunoblotting ICD-10 code (B17.1 and B18.2) from the Danish National Hospital Registry	Kidney	The Danish Cancer 3.60 Registry with ICD-7 (0.98-9.22) code 180		Age, sex and year of diagnosis	Selection: 4 Comparability: 1 Outcome: 2
Gordon et al <sup>[19]</sup>	United	Cohort 2010 study		67063	population (control) HCV-tested patients (both positive and negative) between 1997 and 2006 from administrative data from Henry Ford hospital	HCV infection	Positive anti-HCV test using enzyme-linked immunosorbent assay, confirmed by a documented positive molecular assay for	Renal cell carcinoma	Renal cell Health system cancer carcinoma registry, confirmed by medical record and pathology report review	1.77 (1.05-2.98)	Age, race, gender, chronic kidney disease	Selection: 4 Comparability: 1 Outcome: 3
Budakoğlu <i>et al<sup>trī</sup>l</i>	Turkey	Case-control study	2011 G	6170 ] (903 1 case and 5267 control)	Patients who had histologically proven Positive renal cell carcinoma diagnosis between anti-HCV 2005 to 2010 from six tertiary cancer centers (case) and healthy people who were living in the same geographic	Positive anti-HCV	HCV KNA Positive anti-HCV test using enzyme-linked immunosorbent assay	Renal cell carcinoma	Histopathology report	1.08 (0.62-1.88)	None	Selection: 3 Comparability: 0 Outcome: 3
Hofmann et al <sup>[20]</sup> Sweden	Sweden	Cohort 2011 study		43000 in HCV patients	regions (control) All Swedish residents diagnosed with HCV infection between 1990 and 2006 (HCV group) and general population (control)	Chronic HCV infection	The national surveillance database at the Swedish Institute for Infectious Disease Control	Kidney	The national Cancer register with ICD-7 code 180.0 and 180.9 and histologic	1.2 (0.8-1.7)	Age, sex and calendar year	Selection: 4 Comparability: 1 Outcome: 3
Gonzalez et al <sup>18]</sup>	United	Case-control study	2015 2 a	2015 240 (140 case and 100 control)	Newly diagnosed renal cell carcinoma (case) and newly diagnosed colon cancer patients (control)	Chronic HCV infection	Detection of anti-HCV antibody or HCV RNA	Renal cell carcinoma	confirmation Histopathology report	Positive HCV RNA 24.20 (2.4 - > 999.9)	Sex, age, race, BMI, smoking, alcohol abuse, hypertension, diabetes mellitus, dyslipidemia, coronary artery disease,	Selection: 3 Comparability: 2 Outcome: 3

BMI: Body mass index; DNHR: Danish National Hospital Registry; ELISA: Enzyme-linked immunosorbent assay; HCV: Hepatitis C virus; N/A: Not available; NSW: New South Wales; RCC: Renal cell carcinoma; SIR: Standardized incidence ratio.



funnel plot was suggestive of a small publication bias toward studies with a positive correlation between HCV infection and RCC.

#### DISCUSSION

This meta-analysis was conducted to summarize all presently available data on the association between HCV infection and RCC. Our study demonstrated a 1.86-fold increased risk of RCC among participants who had HCV infection compared to those without HCV infection. Our analysis also illustrates that that RCC patients with HCV were significantly younger than RCC patients without HCV. The results of our study reinforce the hypothesis that HCV may accelerate the risk of developing RCC.

Although the nature in which HCV induces RCC is not entirely understood, several hypotheses exist. A recent bioinformatics study demonstrated a plausible biological relationship between HCV infection and the development of RCC via the NY-REN-54 protein. The NY-REN-54 protein which is an altered ubiquitin-related protein that plays a role in the disturbance impairs the autophagic response via to the ubiquitin-protein ligase-related self-regulatory mechanism, which in turn promotes oncogenesis<sup>[28]</sup>. Additionally, inhibition of cytotoxic T-lymphocyte-dependent apoptosis by the hepatitis virus secondarily leads to a disturbance in host immunity and normal tissue homeostasis leading to carcinoma formation<sup>[29]</sup>. Lukkonen et al<sup>[30]</sup> demonstrated an increased expression of serine protease inhibitor Kazal (SPIK), a cellular protein that inhibits serine proteaserelated apoptosis, in RCC tissue samples as an additional mechanism for HCV-induced RCC.

There are several limitations in our study. Firstly, there was a high amount of statistical heterogeneity present in the completed analysis. The potential source of this heterogeneity includes variation in confounderadjusted methods (e.g., age, sex, ethnicity, and chronic kidney disease), exposure measurement, outcome ascertainments, and follow-up duration. Secondly, it should be noted that there was a potential small publication bias with a positive association between HCV infection and RCC. The possibility of selection bias could play a role in chart-reviewed population base study. Finally, this meta-analysis of observational studies could only show an association. It cannot establish causality as an unknown number of confounders could play a role in the association between HCV and RCC.

In summary, this study demonstrates a potential association between HCV infection and RCC. In the new era of treatment of HCV infection, direct-acting antiviral agents have been demonstrated to be effective therapy and increasingly used<sup>[31]</sup>. These agents may potentially decrease the incidence of RCC in HCV patients in the long term.

#### **COMMENTS**

#### **Background**

Hepatitis C virus (HCV) is one of the leading causes of cirrhosis as the

prevalence of HCV has been steadily increasing over the past two decades in the United States. Extrahepatic manifestations of HCV are common. Interestingly, HCV infection could also increase the risk of renal cell carcinoma (RCC) as observed in several epidemiologic studies.

#### Research frontiers

The results of those epidemiologic studies were inconsistent. To further investigate this possible association of HCV and RCC, the authors conducted this systematic review and meta-analysis of observational studies reporting the risk of RCC among HCV-infected patients.

#### Innovations and breakthroughs

The authors found a significantly increased risk of RCC among participants with HCV infection with the pooled risk ratio (RR) of 1.86 (95%CI: 1.11-3.11). The sensitivity analysis including only studies with confounder adjustment also demonstrated increased risk of RCC among participants with HCV infection with the pooled RR of 1.50 (95%CI: 0.93-2.42) even though without reaching statistical significance.

#### Applications

This study demonstrated a potential association between HCV infection and RCC. This finding suggests that a history of HCV is potentially associated with RCC and may impact clinical management and cancer surveillance.

#### Peer-review

The manuscript is an interesting meta-analysis about the correlation of HCV infection and RCC development. The aim of the meta-analysis is clearly stated and methods are well-described.

#### REFERENCES

- Udompap P, Mannalithara A, Heo NY, Kim D, Kim WR. Increasing prevalence of cirrhosis among U.S. adults aware or unaware of their chronic hepatitis C virus infection. *J Hepatol* 2016; 64: 1027-1032 [PMID: 26809112 DOI: 10.1016/j.jhep.2016.01.009]
- 2 de Oliveria Andrade LJ, D'Oliveira A, Melo RC, De Souza EC, Costa Silva CA, Paraná R. Association between hepatitis C and hepatocellular carcinoma. *J Glob Infect Dis* 2009; 1: 33-37 [PMID: 20300384 DOI: 10.4103/0974-777x.52979]
- Davila JA, Morgan RO, Shaib Y, McGlynn KA, El-Serag HB. Hepatitis C infection and the increasing incidence of hepatocellular carcinoma: a population-based study. *Gastroenterology* 2004; 127: 1372-1380 [PMID: 15521006]
- 4 Giordano TP, Henderson L, Landgren O, Chiao EY, Kramer JR, El-Serag H, Engels EA. Risk of non-Hodgkin lymphoma and lymphoproliferative precursor diseases in US veterans with hepatitis C virus. *JAMA* 2007; 297: 2010-2017 [PMID: 17488966 DOI: 10.1001/jama.297.18.2010]
- 5 Hartridge-Lambert SK, Stein EM, Markowitz AJ, Portlock CS. Hepatitis C and non-Hodgkin lymphoma: the clinical perspective. Hepatology 2012; 55: 634-641 [PMID: 22120959 DOI: 10.1002/hep.25499]
- 6 Vlaar AP, Rietdijk ST, Zeerleder SS, Boerman T, Beuers U. Malignancies associated with chronic hepatitis C: case report and review of the literature. Neth J Med 2011; 69: 211-215 [PMID: 21646667]
- 7 Cacoub P, Renou C, Rosenthal E, Cohen P, Loury I, Loustaud-Ratti V, Yamamoto AM, Camproux AC, Hausfater P, Musset L, Veyssier P, Raguin G, Piette JC. Extrahepatic manifestations associated with hepatitis C virus infection. A prospective multicenter study of 321 patients. The GERMIVIC. Groupe d'Etude et de Recherche en Medecine Interne et Maladies Infectieuses sur le Virus de l'Hepatite C. Medicine (Baltimore) 2000; 79: 47-56 [PMID: 10670409]
- Fabrizi F, Verdesca S, Messa P, Martin P. Hepatitis C Virus Infection Increases the Risk of Developing Chronic Kidney Disease: A Systematic Review and Meta-Analysis. *Dig Dis Sci* 2015; 60: 3801-3813 [PMID: 26195311 DOI: 10.1007/s10620-015-3801-y]
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. CA Cancer J



- Clin 2016; 66: 7-30 [PMID: 26742998 DOI: 10.3322/caac.21332]
- Adams KF, Leitzmann MF, Albanes D, Kipnis V, Moore SC, Schatzkin A, Chow WH. Body size and renal cell cancer incidence in a large US cohort study. Am J Epidemiol 2008; 168: 268-277 [PMID: 18544571 DOI: 10.1093/aje/kwn122]
- 11 Cheungpasitporn W, Thongprayoon C, O'Corragain OA, Edmonds PJ, Ungprasert P, Kittanamongkolchai W, Erickson SB. The risk of kidney cancer in patients with kidney stones: a systematic review and meta-analysis. *QJM* 2015; 108: 205-212 [PMID: 25208892 DOI: 10.1093/qjmed/hcu195]
- 12 Cumberbatch MG, Rota M, Catto JW, La Vecchia C. The Role of Tobacco Smoke in Bladder and Kidney Carcinogenesis: A Comparison of Exposures and Meta-analysis of Incidence and Mortality Risks. Eur Urol 2016; 70: 458-466 [PMID: 26149669 DOI: 10.1016/j.eururo.2015.06.042]
- Ljungberg B, Campbell SC, Choi HY, Jacqmin D, Lee JE, Weikert S, Kiemeney LA. The epidemiology of renal cell carcinoma. *Eur Urol* 2011; 60: 615-621 [PMID: 21741761 DOI: 10.1016/j.eururo.2011.06.049]
- 14 Wiklund F, Tretli S, Choueiri TK, Signoretti S, Fall K, Adami HO. Risk of bilateral renal cell cancer. *J Clin Oncol* 2009; 27: 3737-3741 [PMID: 19597028 DOI: 10.1200/JCO.2008.20.6524]
- 15 Lipworth L, Tarone RE, McLaughlin JK. The epidemiology of renal cell carcinoma. *J Urol* 2006; **176**: 2353-2358 [PMID: 17085101 DOI: 10.1016/j.juro.2006.07.130]
- 16 Amin J, Dore GJ, O'Connell DL, Bartlett M, Tracey E, Kaldor JM, Law MG. Cancer incidence in people with hepatitis B or C infection: a large community-based linkage study. *J Hepatol* 2006; 45: 197-203 [PMID: 16684579 DOI: 10.1016/j.jhep.2006.02.014]
- Budakoğlu B, Aksoy S, Arslan Ç, Üyetürk Ü, Babacan NA, Özcan MF, Yıldız R, Öven BB, Özdemir NY, Dizdar Ö, Büyükberber S, Akıncı MB, Türker I, Öksüzoğlu B, Altundag K, Zengin N. Frequency of HCV infection in renal cell carcinoma patients. *Med Oncol* 2012; 29: 1892-1895 [PMID: 21461964 DOI: 10.1007/s12032-011-9928-6]
- 18 Gonzalez HC, Lamerato L, Rogers CG, Gordon SC. Chronic hepatitis C infection as a risk factor for renal cell carcinoma. *Dig Dis Sci* 2015; 60: 1820-1824 [PMID: 25592719 DOI: 10.1007/s10620-015-3521-3]
- 19 Gordon SC, Moonka D, Brown KA, Rogers C, Huang MA, Bhatt N, Lamerato L. Risk for renal cell carcinoma in chronic hepatitis C infection. *Cancer Epidemiol Biomarkers Prev* 2010; 19: 1066-1073 [PMID: 20332260 DOI: 10.1158/1055-9965.EPI-09-1275]
- 20 Hofmann JN, Törner A, Chow WH, Ye W, Purdue MP, Duberg AS. Risk of kidney cancer and chronic kidney disease in relation to hepatitis C virus infection: a nationwide register-based cohort

- study in Sweden. *Eur J Cancer Prev* 2011; **20**: 326-330 [PMID: 21386707 DOI: 10.1097/CEJ.0b013e32834572fa]
- 21 Malaguarnera M, Gargante MP, Risino C, Ranno S, Berretta M, Cannizzaro MA, Costanzo M, Fricia T, Rampello E, Romano M. Hepatitis C virus in elderly cancer patients. *Eur J Intern Med* 2006; 17: 325-329 [PMID: 16864006 DOI: 10.1016/j.ejim.2006.02.004]
- Omland LH, Farkas DK, Jepsen P, Obel N, Pedersen L. Hepatitis C virus infection and risk of cancer: a population-based cohort study. Clin Epidemiol 2010; 2: 179-186 [PMID: 20865115]
- Stang A. Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in metaanalyses. *Eur J Epidemiol* 2010; 25: 603-605 [PMID: 20652370 DOI: 10.1007/s10654-010-9491-z]
- 24 Herzog R, Álvarez-Pasquin MJ, Díaz C, Del Barrio JL, Estrada JM, Gil Á. Are healthcare workers' intentions to vaccinate related to their knowledge, beliefs and attitudes? A systematic review. BMC Public Health 2013; 13: 154 [PMID: 23421987 DOI: 10.1186/1471 -2458-13-154]
- 25 DerSimonian R, Laird N. Meta-analysis in clinical trials. Control Clin Trials 1986; 7: 177-188 [PMID: 3802833]
- 26 Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses. *BMJ* 2003; 327: 557-560 [PMID: 12958120 DOI: 10.1136/bmj.327.7414.557]
- Macleod LC, Hotaling JM, Wright JL, Davenport MT, Gore JL, Harper J, White E. Risk factors for renal cell carcinoma in the VITAL study. *J Urol* 2013; 190: 1657-1661 [PMID: 23665301 DOI: 10.1016/j.juro.2013.04.130]
- Wiwanitkit V. Renal cell carcinoma and hepatitis C virus infection: is there any cause-outcome relationship? *J Cancer Res Ther* 2011; 7: 226-227 [PMID: 21768723 DOI: 10.4103/0973-1482.82931]
- 29 Chisari FV. Cytotoxic T cells and viral hepatitis. *J Clin Invest* 1997; 99: 1472-1477 [PMID: 9119989 DOI: 10.1172/JCI119308]
- 30 Lukkonen A, Lintula S, von Boguslawski K, Carpén O, Ljungberg B, Landberg G, Stenman UH. Tumor-associated trypsin inhibitor in normal and malignant renal tissue and in serum of renal-cell carcinoma patients. *Int J Cancer* 1999; 83: 486-490 [PMID: 10508484]
- 31 Kalaghatgi P, Sikorski AM, Knops E, Rupp D, Sierra S, Heger E, Neumann-Fraune M, Beggel B, Walker A, Timm J, Walter H, Obermeier M, Kaiser R, Bartenschlager R, Lengauer T. Geno2pheno[HCV] A Web-based Interpretation System to Support Hepatitis C Treatment Decisions in the Era of Direct-Acting Antiviral Agents. *PLoS One* 2016; 11: e0155869 [PMID: 27196673 DOI: 10.1371/journal.pone.0155869]

P- Reviewer: Levinson A, Malnick S, Procopio G S- Editor: Kong JX L- Editor: A E- Editor: Wu HL







## Published by Baishideng Publishing Group Inc

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx http://www.wjgnet.com

