

World Journal of *Gastrointestinal Pathophysiology*

World J Gastrointest Pathophysiol 2023 August 24; 14(4): 71-85



ORIGINAL ARTICLE

Basic Study

- 71 Novel, non-colonizing, single-strain live biotherapeutic product ADS024 protects against *Clostridioides difficile* infection challenge *in vivo*

Murphy CK, O'Donnell MM, Hegarty JW, Schulz S, Hill C, Ross RP, Rea MC, Farquhar R, Chesnel L

Contents

World Journal of Gastrointestinal Pathophysiology
Bimonthly Volume 14 Number 4 August 24, 2023

ABOUT COVER

Editorial Board Member of *World Journal of Gastrointestinal Pathophysiology*, Paolo Aseni, MD, Academic Research, Adjunct Professor, Doctor, Dipartimento di Medicina d'Urgenza ed Emergenza, ASST Grande Ospedale Metropolitano, Niguarda Hospital, Milan 20133, Italy. paoloaseni@gmail.com

AIMS AND SCOPE

The primary aim of the *World Journal of Gastrointestinal Pathophysiology* (WJGP, *World J Gastrointest Pathophysiol*) is to provide scholars and readers from various fields of gastrointestinal pathophysiology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJGP mainly publishes articles reporting research results and findings obtained in the field of gastrointestinal pathophysiology and covering a wide range of topics including disorders of the esophagus, stomach and duodenum, small intestines, pancreas, biliary system, and liver.

INDEXING/ABSTRACTING

The WJGP is now abstracted and indexed in PubMed, PubMed Central, Reference Citation Analysis, China National Knowledge Infrastructure, China Science and Technology Journal Database, and Superstar Journals Database.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: *Ying-Yi Yuan*, Production Department Director: *Xu Guo*, Editorial Office Director: *Jia-Ping Yan*.

NAME OF JOURNAL

World Journal of Gastrointestinal Pathophysiology

ISSN

ISSN 2150-5330 (online)

LAUNCH DATE

April 15, 2010

FREQUENCY

Bimonthly

EDITORS-IN-CHIEF

Kusum K Kharbanda, Tsutomu Nishida, Somchai Amornvotin

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/2150-5330/editorialboard.htm>

PUBLICATION DATE

August 24, 2023

COPYRIGHT

© 2023 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/gerinfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/gerinfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/gerinfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>

© 2023 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

E-mail: bpgoffice@wjgnet.com <https://www.wjgnet.com>



Basic Study

Novel, non-colonizing, single-strain live biotherapeutic product ADS024 protects against *Clostridioides difficile* infection challenge *in vivo*

Christopher K Murphy, Michelle M O'Donnell, James W Hegarty, Sarah Schulz, Colin Hill, R Paul Ross, Mary C Rea, Ronald Farquhar, Laurent Chesnel

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Unsolicited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0
Grade B (Very good): B
Grade C (Good): 0
Grade D (Fair): 0
Grade E (Poor): 0

P-Reviewer: Mohapatra S, India

Received: June 1, 2023

Peer-review started: June 1, 2023

First decision: July 19, 2023

Revised: August 2, 2023

Accepted: August 7, 2023

Article in press: August 7, 2023

Published online: August 24, 2023



Christopher K Murphy, Laurent Chesnel, Research and Development, Adiso Therapeutics Inc., Concord, MA 01742, United States

Michelle M O'Donnell, Sarah Schulz, Colin Hill, R Paul Ross, APC Microbiome Ireland, University College Cork, Cork T12 K8AF, Ireland

James W Hegarty, Mary C Rea, Teagasc Food Research Centre, Moorepark Fermoy, Cork P61 C996, Ireland

Ronald Farquhar, Executive Leadership Team, Adiso Therapeutics Inc., Concord, MA 01742, United States

Corresponding author: Laurent Chesnel, PhD, Research and Development, Adiso Therapeutics, Inc., 530 Virginia Road, Suite 300, Concord, MA 01742, United States. lchesnel@adisotx.com

Abstract

BACKGROUND

The Centers for Disease Control and Prevention estimate that *Clostridioides difficile* (*C. difficile*) causes half a million infections (CDI) annually and is a major cause of total infectious disease death in the United States, causing inflammation of the colon and potentially deadly diarrhea. We recently reported the isolation of ADS024, a *Bacillus velezensis* (*B. velezensis*) strain, which demonstrated direct *in vitro* bactericidal activity against *C. difficile*, with minimal collateral impact on other members of the gut microbiota. In this study, we hypothesized that *in vitro* activities of ADS024 will translate *in vivo* to protect against CDI challenge in mouse models.

AIM

To investigate the *in vivo* efficacy of *B. velezensis* ADS024 in protecting against CDI challenge in mouse models.

METHODS

To mimic disruption of the gut microbiota, the mice were exposed to vancomycin prior to dosing with ADS024. For the mouse single-dose study, the recovery of ADS024 was assessed *via* microbiological analysis of intestinal and fecal samples

at 4 h, 8 h, and 24 h after a single oral dose of 5×10^8 colony-forming units (CFU)/mouse of freshly grown ADS024. The single-dose study in miniature swine included groups that had been pre-dosed with vancomycin and that had been exposed to a dose range of ADS024, and a group that was not pre-dosed with vancomycin and received a single dose of ADS024. The ADS024 colonies [assessed by quantitative polymerase chain reaction (qPCR) using ADS024-specific primers] were counted on agar plates. For the 28-d miniature swine study, qPCR was used to measure ADS024 levels from fecal samples after oral administration of ADS024 capsules containing 5×10^9 CFU for 28 consecutive days, followed by MiSeq compositional sequencing and bioinformatic analyses to measure the impact of ADS024 on microbiota. Two studies were performed to determine the efficacy of ADS024 in a mouse model of CDI: Study 1 to determine the effects of fresh ADS024 culture and ADS024 spore preparations on the clinical manifestations of CDI in mice, and Study 2 to compare the efficacy of single daily doses *vs* dosing 3 times per day with fresh ADS024. *C. difficile* challenge was performed 24 h after the start of ADS024 exposure. To model the human distal colon, an anaerobic fecal fermentation system was used. MiSeq compositional sequencing and bioinformatic analyses were performed to measure microbiota diversity changes following ADS024 treatment. To assess the potential of ADS024 to be a source of antibiotic resistance, its susceptibility to 18 different antibiotics was tested.

RESULTS

In a mouse model of CDI challenge, single daily doses of ADS024 were as efficacious as multiple daily doses in protecting against subsequent challenge by *C. difficile* pathogen-induced disease. ADS024 showed no evidence of colonization based on the observation that the ADS024 colonies were not recovered 24 h after single doses in mice or 72 h after single doses in miniature swine. In a 28-d repeat-dose study in miniature swine, ADS024 was not detected in fecal samples using plating and qPCR methods. Phylogenetic analysis performed in the human distal colon model showed that ADS024 had a selective impact on the healthy human colonic microbiota, similarly to the *in vivo* studies performed in miniature swine. Safety assessments indicated that ADS024 was susceptible to all the antibiotics tested, while *in silico* testing revealed a low potential for off-target activity or virulence and antibiotic-resistance mechanisms.

CONCLUSION

Our findings, demonstrating *in vivo* efficacy of ADS024 in protecting against CDI challenge in mouse models, support the use of ADS024 in preventing recurrent CDI following standard antibiotic treatment.

Key Words: Gut; Single-strain live biotherapeutic product; ADS024; *Clostridioides difficile*; *Clostridioides difficile* challenge model; Microbiota

©The Author(s) 2023. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: *Clostridioides difficile* (*C. difficile*), a Gram positive pathogen associated with life-threatening gastrointestinal disease, colonizes millions of healthy people worldwide, causing disease in individuals with disrupted gut microbiomes. Here, we demonstrate *in vivo* efficacy of recently isolated *Bacillus velezensis* ADS024, without colonization, in protecting against *C. difficile* infection (CDI) challenge in mouse models. We also show that this novel strain has minimal effects on the gut microbiome in a human distal colon model and miniature swine. This study supports further investigation of ADS024 as a single-strain, live biotherapeutic product candidate for preventing recurrent CDI following successful standard-of-care antibiotic therapy.

Citation: Murphy CK, O'Donnell MM, Hegarty JW, Schulz S, Hill C, Ross RP, Rea MC, Farquhar R, Chesnel L. Novel, non-colonizing, single-strain live biotherapeutic product ADS024 protects against *Clostridioides difficile* infection challenge *in vivo*. *World J Gastrointest Pathophysiol* 2023; 14(4): 71-85

URL: <https://www.wjgnet.com/2150-5330/full/v14/i4/71.htm>

DOI: <https://dx.doi.org/10.4291/wjgp.v14.i4.71>

INTRODUCTION

Clostridioides difficile (*C. difficile*; previously *Clostridium difficile*) is an opportunistic Gram positive pathogen that takes advantage of a disrupted gut microbiome to cause potentially life-threatening gastrointestinal (GI) disease[1,2]. *C. difficile* infects millions of people worldwide, resulting in a range of pathologies, from mild abdominal symptoms to potentially life-threatening colitis. It is the most frequently identified health care-related infection in the United States, where community-acquired infections are increasing[3]. Risk factors include prior hospitalization, residence in a nursing home care facility, advanced age, underlying disease, and alterations in the gut microbiota[3-5]. *C. difficile* infection (CDI) is mainly characterized by diarrhea, but the spectrum of disease can progress to colitis and toxic megacolon[6]. The

pathogenicity of *C. difficile* is mainly mediated by 2 exotoxins, toxin A (TcdA) and toxin B (TcdB)[2]. In addition, certain strains of *C. difficile* may also produce a binary toxin called *C. difficile* transferase[2]. The clinical symptoms described above are mediated by TcdA and TcdB, which precipitate an inflammatory response and cytotoxicity in epithelial cells of the GI tract[2]. While the relative contributions of TcdA and TcdB may be specific to the host species[7,8], TcdB is the predominant mediator of CDI[7,9] and the most clinically relevant[8].

Recurrent CDI (rCDI) following initial successful antibiotic treatment remains a clinical challenge, with each episode associated with an increased risk for recurrence (25% after the first, 45% after the second, and 65% after the third and subsequent episodes), creating a cycle of infection and antibiotic use[10,11]. The causes of rCDI are primarily attributed to the failure of antibiotic-disrupted gut microbiota to restrict the germination and subsequent overgrowth of surviving spores of *C. difficile* and an altered immune response as a result of the action of the *C. difficile* toxins[10,12]. Therefore, predicting future recurrence is crucial. Machine learning is emerging as a useful method of predicting outcomes by comparing changes in fecal microbial composition and metabolome in patients with rCDI *vs* those without[12,13].

The gut microbiome plays a critical role in preventing CDI among its many functions. Indeed, perturbation of the microbiome through frequent antibiotic administration is a critical risk factor for subsequent recurrences, presumably as a result of the time needed to recover a diverse and protective microbiota. Even with short-term antibiotic treatment, microbiota recovery takes several months[14], and perturbations can persist up to 4 years post treatment[15,16]. In addition, the standard treatment of initial and rCDI often involves broad-spectrum antibiotics, including vancomycin[1]. Even though CDI clinical cure rates are high[17,18], patients remain at risk for recurrence[18]. Thus, degrading TcdA and TcdB, killing germinating *C. difficile* spores, and reducing inflammation are vital for restoring protective intestinal microbiota and successfully preventing recurrent episodes of CDI.

In a recent study, we described the isolation of ADS024, a *Bacillus velezensis* (*B. velezensis*) strain in development as a single-strain live biotherapeutic product (SS-LBP) to prevent the recurrence of CDI after clinical cure by standard antibiotic treatment[19]. LBPs are biological medicinal products containing live microorganisms as active substances. They are being studied to prevent, treat, or cure various diseases[20–24], including allergies, dental disorders, and GI, dermatologic, and gut-brain-axis-related conditions[25]. We demonstrated that ADS024 has bactericidal activity against *C. difficile*, with minimal impact on other common members of the gut microbiota, and produces proteases that can degrade both TcdA and TcdB[19]. The current study expands on our previous findings by demonstrating the protective effects of ADS024 in a mouse model of CDI, the minimal impact of ADS024 on the gut microbiome of miniature swine and humans, and its lack of colonization/engraftment in miniature swine and mouse models. The results of these studies support further development of ADS024 as an SS-LBP for the prevention of rCDI after clinical cure following completion of standard antibiotic treatment.

MATERIALS AND METHODS

Ethics statement

Biological samples for this work were collected using Clinical Research Ethics Committee, Cork, Ireland, approved consents for the Protocol “ControlMET” numbered APC065.

Animal care and use statement

The protocol and any amendment(s) or procedures involving the care and use of animals in this study were reviewed and approved by the Institutional Animal Care and Use Committee (IACUC) at NeoSome Life Sciences. During the study, the care and use of animals were conducted with guidance from the United States National Research Council guidelines. Veterinary care was available throughout the study for animals to be examined by the veterinary staff as warranted by clinical signs or other changes. There were no changes in the animals that required veterinary intervention to ameliorate pain or distress.

Quantification of ADS024 in mice (single-dose study)

The recovery of ADS024 was assessed in intestinal and fecal samples of mice at various times after a single dose of freshly grown ADS024. Prior to dosing with ADS024, the mice were exposed to vancomycin to disrupt their microbiota in a clinically relevant manner. Further detail is provided in the [Supplementary material](#).

Quantification of ADS024 in minipigs (single-dose study)

The aim of this study was to investigate ADS024 fecal content from male miniature swine following a single dose of ADS024 capsules over the time course of 5 d post ADS024 dosing. Further detail is provided in the [Supplementary material](#).

In vivo mouse model of *C. difficile* infection

The study design was adapted from Chen *et al*[26] and modified to test if prior dosing with ADS024 could protect against subsequent challenge by *C. difficile*. Accordingly, the ADS024 treatment started 24 h prior to the CDI (spore) challenge. Upon their receipt at NeoSome Life Sciences, the mice, while not tested for *C. difficile* colonization/infection prior to inoculation with *C. difficile*, were examined by personnel to ensure acceptable health status. Veterinary care was provided by the veterinarians and staff employed by NeoSome Life Sciences. The mice were acclimated for at least 5 d prior to use. Following acclimation, the mice were infected with *C. difficile* strain VPI10463 (6.30×10^5 to 1.97×10^6 CFU, a higher dose

compared with the 10^2 to 10^5 CFU in the paper by Chen *et al* [26]), and 65 and 40 mice were treated with ADS024 or saline, respectively, across the conducted studies. The mice were then monitored for survival, weight loss, and disease-related clinical observations, including wet tail, diarrhea, hunched posture, dehydration, and lethargy. The daily score of adverse health observations was calculated based on criteria specified in [Supplementary Table 1](#). Further detail, including antibiotic pretreatment, is provided in the [Supplementary material](#).

Distal colon model

The micro-Matrix (Applikon Biotechnology, Delft, Netherlands) was used to model the human distal colon and examine the impact of ADS024 on gut microbial populations. The anaerobic fecal fermentation model of the distal colon has been previously described [27]. Further detail is provided in the [Supplementary material](#).

MiSeq compositional sequencing and bioinformatic analysis: All extracted samples from the distal colon model were prepared for MiSeq DNA compositional sequencing, and 16S data were then organized by alpha diversity, beta diversity, and phylum and genus diversity. Further detail is provided in the [Supplementary material](#).

Statistical significance determination: ADS024-treated wells were then assessed by comparing differences to wells with untreated media. Further detail is provided in the [Supplementary material](#).

Quantitative polymerase chain reaction: Absolute quantification by quantitative polymerase chain reaction (qPCR) was performed to determine total bacterial numbers and specific populations from fecal samples. Further detail is provided in the [Supplementary material](#).

ADS024 quantification in miniature swine fecal samples, 28-d study

To determine the impact of ADS024 administration on the gut composition of miniature swine, qPCR, MiSeq compositional sequencing, and bioinformatic analysis were performed in duplicate using fecal samples. Further detail is provided in the [Supplementary material](#).

ADS024 susceptibility to antibiotics

Two batches each of lyophilized drug substances (CO-33-10A2 and CO-33-12A2) and frozen glycerol stocks [research cell bank (CO-33 ART24BHI) and the master cell bank (CO-33 ART24SYD)] were tested for antimicrobial susceptibility using the reference broth microdilution method per the Clinical and Laboratory Standards Institute (CLSI) guidelines (CLSI M07 2018). Tests with metronidazole were also carried out under anaerobic conditions (CLSI M11 2018). Minimum inhibitory concentration (MIC) values obtained against ADS024 were validated by concurrently testing CLSI-recommended ATCC quality control reference strains (CLSI M100 2019). Categorical interpretations for the MIC results obtained against ADS024 used CLSI M45 (2015) [28] breakpoint criteria or European Food Safety Authority (EFSA 2012) [29] microbiological cutoff values.

Potential virulence factors and antimicrobial resistance genes of ADS024 were analyzed using IslandViewer 4. The annotated ADS024 genome was first analyzed using IslandViewer 4, and a separate web server was subsequently used to interpret the data. The output was based on mapping as detailed in [Supplementary Figure 1](#) and stored as a Microsoft Excel spreadsheet that contained the output of the integrated IslandPath-DIMOB, SIGI-HMM, and IslandPick prediction methods.

Biostatistics statement

The bioinformatics and statistical analyses relating to 16S sequencing and the micro-Matrix, as described in this manuscript, were verified by Dr. Michelle M. O'Donnell of APC Microbiome Ireland, University College Cork. The bioinformatics and statistical analyses relating to all animal-related research described in this paper were verified by Dr. Christopher Murphy of Adiso Therapeutics, Inc.

RESULTS

ADS024 is recovered in fecal samples after single-dose administration in mice and miniature swine

Microbiological analysis was conducted on fecal pellets and GI tissue from mice pre-dosed with vancomycin to mimic disruption of the gut microbiota. The mice were then administered a single oral dose of approximately 5×10^8 colony-forming units (CFU)/mouse of ADS024. ADS024 has a very distinctive morphology allowing for visual detection on agar plates [19]. ADS024 colonies were recovered in feces following plating on agar media at 4 h, 8 h, and 24 h post dose ([Figure 1A](#)). No ADS024 colonies were detected in feces after the 24-h post-dose time point. Only 1 GI tissue sample contained detectable amounts of ADS024, present in the upper intestine 24 h after ADS024 administration ([Supplementary Figure 2](#)).

The results from a single-dose study in minipigs (3 male miniature swine per group) were consistent with those from the mouse studies, showing a lack of detection in fecal and tissue samples past 72 h, supporting daily dosing for clinical use. The single-dose study in miniature swine included groups that had been pre-dosed with vancomycin to induce disruption of the gut microbiota, as described under Materials and Methods, and that had been exposed to a dose range of ADS024 (Groups 2, 3, and 4 received 6.2×10^7 , 7.1×10^8 , or 7.1×10^9 CFU, respectively), and a group that was not pre-dosed with vancomycin and received a single dose of ADS024 (Group 5 received 7.1×10^9 CFU). The control group, group

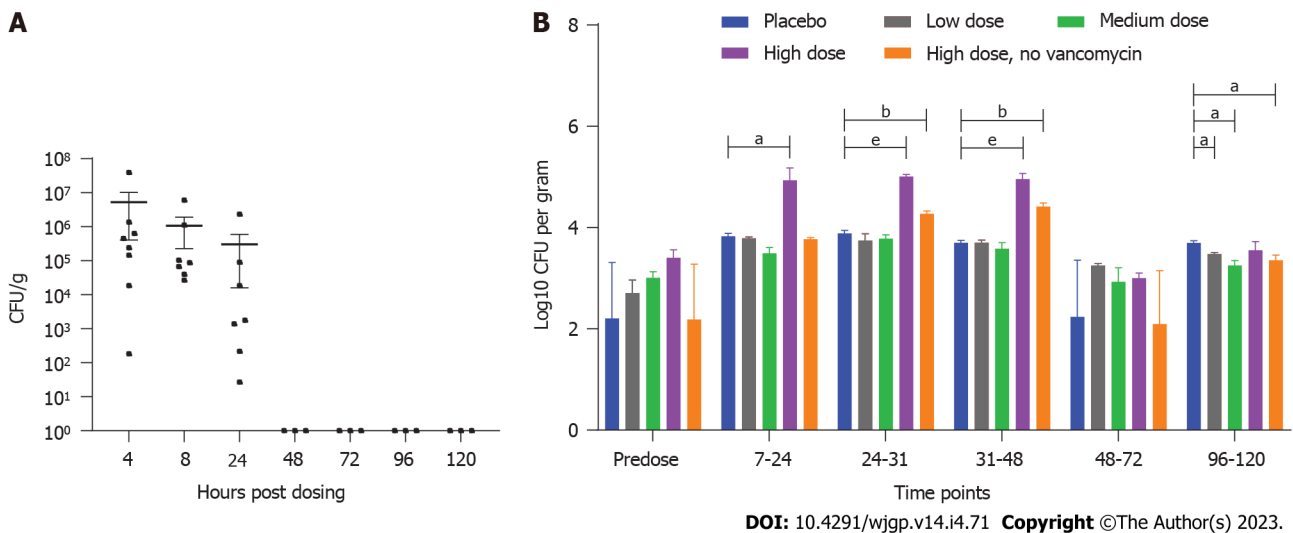


Figure 1 Quantification of ADS024 (fresh culture) in the mice and miniature swine fecal samples for the single-dose study. A: The mice were predosed with vancomycin to mimic disruption of gut microbiota and administered a single oral dose of 5×10^8 colony-forming units (CFU)/mouse of ADS024. Microbiological analysis was subsequently conducted on the mouse fecal pellets and gastrointestinal tissue. The ADS024 colonies were recovered in feces following plating on agar media at 4 h, 8 h, and 24-h post dose; B: The single-dose study in miniature swine included 3 groups predosed with vancomycin to induce disruption of gut microbiota. They were exposed to the following dose range of ADS024: Group 2 (low dose) 6.2×10^7 CFU; Group 3 (medium dose) 7.1×10^8 CFU; and Group 4 (high dose) 7.1×10^9 CFU. There was 1 group not pre-dosed with vancomycin but exposed to a single dose of ADS024: Group 5 (no vancomycin, high dose) 7.1×10^9 CFU. The control group (Group 1, placebo) did not receive vancomycin or ADS024. After single oral administration of ADS024 to the miniature swine, the ADS024 colonies, assessed by quantitative polymerase chain reaction (qPCR), were detected on agar plates in the ADS024-treated Groups 4 and 5, regardless of vancomycin therapy, at colony counts higher than placebo controls that displayed only *Bacillus*-like colonies and not ADS024-specific ones based on qPCR, at the 7-h to 48-h time points. *P* value significance: ^a $P \leq 0.05$, ^b $P \leq 0.01$, ^c $P \leq 0.001$.

1, did not receive vancomycin or ADS024. After single oral administration of ADS024 in miniature swine, the ADS024 colonies (assessed by qPCR using ADS024-specific primers) were detected on agar plates in the ADS024-treated Groups 4 and 5, regardless of vancomycin therapy, at colony counts higher than placebo controls (which grew only *Bacillus*-like, but not ADS024-specific, colonies) at the 7- to 48-h time points (Figure 1B). The qPCR data (Supplementary Figure 3) confirmed the plating colony counts (Figure 1B), as no ADS024-selective agar plates were available.

ADS024 colonies were considered to have increased if the values were higher than the highest predose value of 10^4 CFU/g and higher than the corresponding placebo value for the given time point. The 7- to 48-h post-dose period was the Discovery Window, and this term was used for subsequent qPCR analysis. The qPCR analysis of Groups 2 to 5 showed that ADS024 was present in the Discovery Window samples at 24 h to 31 h post dose for the low-dose Group 2, 24 h to 48 h for the mid-dose Group 3, and 7 to 48 h for the high-dose Groups 4 and 5 at variable concentrations higher than predose in all animals (except for the Group 5 samples) (Figure 1B).

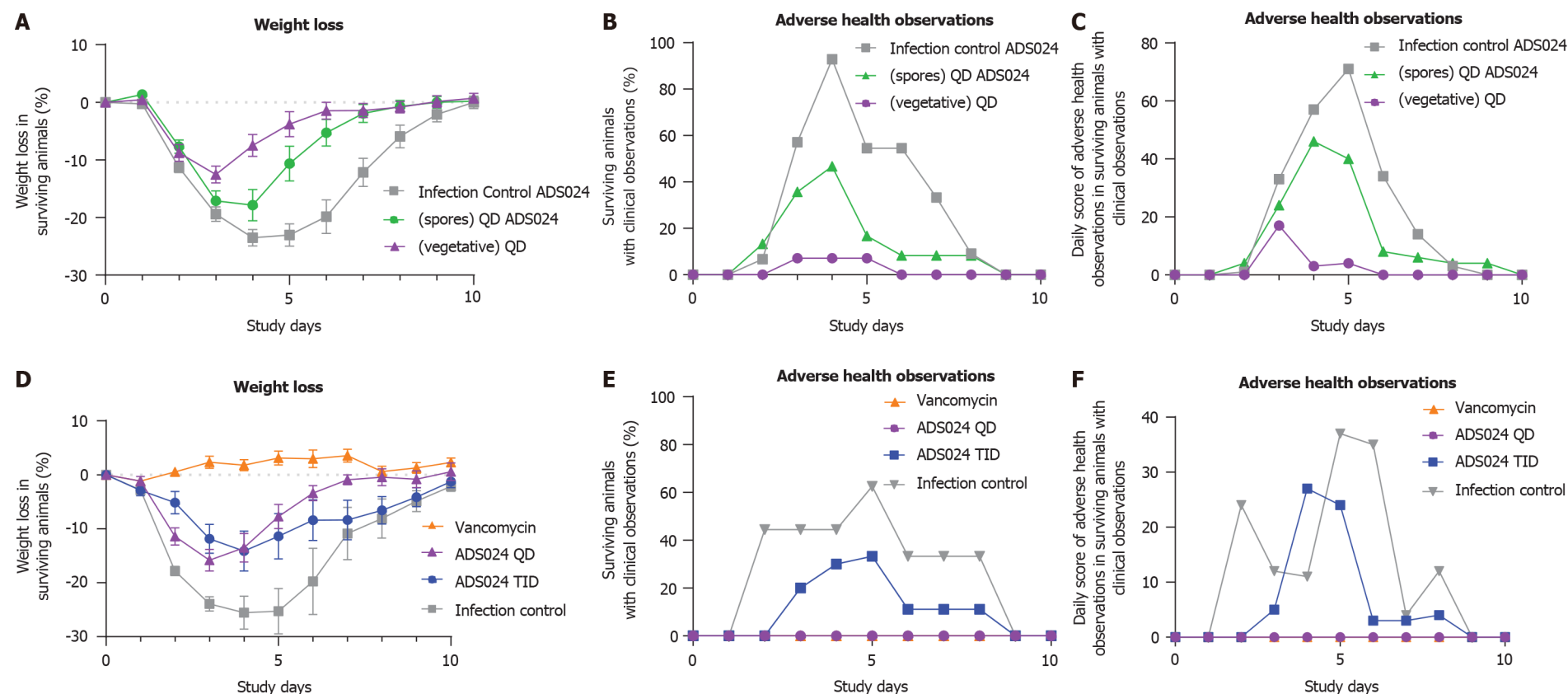
ADS024 is efficacious in a mouse model of CDI

To confirm if the toxin degradation and *C. difficile*-killing activities observed in our *in vitro* studies[19,20] translated to *in vivo* efficacy, we utilized a mouse model of CDI. The animal model was adapted from Chen *et al*[26] as described under Materials and Methods, and the *C. difficile* challenge was performed 24 h after the start of ADS024 exposure.

Two studies were performed to determine the efficacy of ADS024 in the mouse model of CDI. The first study (Study 1, Figure 2A-C) determined the effects of fresh ADS024 culture and ADS024 spore preparations on the clinical manifestations of CDI in mice, namely weight loss (Figure 2A) and adverse clinical features (Figure 2B and C). The second study (Study 2, Figure 2D-F) compared the efficacy of single daily doses (QD) *vs* dosing 3 times per day (TID) with fresh ADS024.

Study 1: ADS024 was delivered to mice ($n = 15$ per group) as resuspended spore suspensions [in phosphate-buffered saline (PBS)] or as a freshly prepared daily culture (ADS024 bacteria resuspended in PBS). The placebo (infection control) group demonstrated an average maximum weight loss of 24.4%, and the mice experienced adverse health effects for 33% of the days (Figure 2A-C). In comparison, mice that received ADS024 as a fresh culture suffered less weight loss compared with the infection (vehicle-dosed) controls (13.5% maximum average percentage of pre-infection weight loss; Figure 2A), and the mice experienced adverse health effects for only 3% of the days (Figure 2B), which was the lowest daily score of adverse health observations among the 3 groups (Figure 2C). The ADS024 spore preparation-treated mice exhibited a maximum weight loss of 20% on day 4, and for 19% of the days, the mice scored adverse health effects (Figure 2A-C). Therefore, mice treated with fresh ADS024 had lower morbidity and mortality than those treated with ADS024 spores or mice that received only placebo.

Study 2: Dose frequencies of QD and TID were compared in mice ($n = 10$ per group). The untreated (infection control) group had an average maximum weight loss of 25% on day 4 (Figure 2D), similar to observations in Study 1. In



DOI: 10.4291/wjgp.v14.i4.71 Copyright ©The Author(s) 2023.

Figure 2 *Clostridioides difficile* infection challenge mouse model studies. Study 1: The effects of fresh (vegetative) ADS024 culture vs ADS024 spore preparations on the clinical manifestations of *Clostridioides difficile* infection (CDI) over 10 d. ADS024 was delivered to the mice ($n = 15$ per group) as resuspended spore suspensions [in phosphate-buffered saline (PBS)] or as a freshly prepared daily culture (ADS024 bacteria resuspended in PBS) 24 h prior to *C. difficile*. A: The weight loss comparison between mice receiving vegetative vs spore ADS024 relative to the infection (vehicle-dosed) controls; B: The adverse health effects comparison in surviving animals among the 3 groups (control, vegetative ADS024, and spore ADS024); C: The daily score of adverse health observations among the 3 groups was determined as shown in [Supplementary Table 1](#). Study 2: The effects of single daily (QD) doses vs 3 times daily (TID) administration with fresh ADS024 in the mice ($n = 10$ per group) over the course of 10 d; D: The weight loss comparison among the 4 groups (vancomycin, untreated, QD, and TID); E: The adverse health effects comparison in the surviving animals among the 4 groups; F: The daily score of adverse health observations among the 4 groups was determined as shown in [Supplementary Table 1](#).

comparison, mice that received ADS024 QD had an average maximum weight loss of 16% on day 3 (from the pre-dose weight) before recovering to the group average pre-infection body weight at day 10 ([Figure 2D](#)). Mice receiving ADS024 TID demonstrated a maximum weight loss of 14% on day 4 ([Figure 2D](#)), similar to that in Study 1. The vancomycin control mice had no weight loss throughout the study ([Figure 2D](#)). The ADS024 QD and vancomycin groups did not display adverse clinical features, as demonstrated by overlapping values in [Figure 2E](#) and [F](#). In contrast, the ADS024 TID

group did display adverse clinical features (Figure 2E and F), perhaps due to the TID oral gavage-related stress. Therefore, there was no obvious benefit from repeat dosing because the efficacy in this model was similar; furthermore, there were fewer adverse health events in the QD-dosed group than in the TID-dosed group.

ADS024 selectively impacts the gut microbiome in an *in vitro* model of the human distal colon

We used anaerobic fecal fermentation to model the distal colon as previously described[27]. qPCR was used to analyze the effect of ADS024 exposure on 5 different components of human colonic microbiota (Enterobacteriaceae, Bacillota, Bacteroidota, Verrucomicrobiota, and *Lactobacillus*) and to determine the number of ADS024 colonies in inoculated wells (Figure 3A and B). ADS024 was detectable in the ADS024-inoculated wells (Figure 3A). Still, its concentration did not increase during the experiment and had a negligible impact on other gut microbiota components after 24 h of incubation (Figure 3B).

MiSeq 16S (Illumina, San Diego, CA, United States) compositional sequencing analyses at the phylum level (Figure 3C and Supplementary Table 2) revealed an increase in Actinomycetota (approximately 13%; $P < 0.0001$) and significant decreases in Bacillota (approximately 12%; $P < 0.01$), Fusobacteriota (approximately 0.001%; $P < 0.05$), and Verrucomicrobiota (approximately 0.4%; $P < 0.05$) compared with media-only treatment. Compositional analyses at the genus level (Figure 3D and Supplementary Table 3) revealed 12 genera showing significant differences after 24 h of treatment compared with the media-only controls. Among the 7 genera positively impacted by the exposure to ADS024, increases in *Bifidobacterium* (approximately 12%; $P < 0.00003$) and *Bacteroides* (approximately 2%; $P < 0.05$) were found compared with media-only treatment (Figure 3D). These results correlate with the MiSeq compositional sequencing and bioinformatic analyses that were performed to measure microbiota diversity changes following ADS024 treatment using Shannon and Simpson indexes for alpha diversity (Figure 4A and B) and UniFrac principal coordinates analysis (PCoA) for beta diversity (Figure 4C). After 24 h of incubation, ADS024 had less effect on alpha diversity than the media-only controls relative to baseline (Figure 4A and B). The microbial diversity between the media-exposed and ADS024-exposed samples was similar in beta diversity changes (Figure 4C).

Miniature swine gut microbiota is not impacted after 28-d dosing with ADS024

ADS024 Levels from fecal samples were determined by qPCR after QD oral administration of ADS024 capsules containing 5×10^9 CFU in male and female miniature swine for 28 consecutive days as described in Materials and Methods (Supplementary Figure 4). ADS024-specific genetic material was below the limit of detection in both the placebo and ADS024-dosed groups at all assayed time points. Subsequently, MiSeq compositional sequencing and bioinformatic analyses were performed to measure the impact of ADS024 on microbiota in miniature swine fecal samples collected pre dose (day 15), on the last day of dosing (day 29), and 14 d after the last dose (recovery) using Shannon and Simpson indexes for alpha diversity (Figure 5A and B) and UniFrac PCoA for beta diversity (Figure 5C). The alpha diversity analyses indicated no significant differences in microbiota diversity between the ADS024 and placebo treatment groups at any time (Figure 5A and B). Beta diversity using PCoA plots, based on weighted UniFrac distance matrixes, indicated minor, non-significant differences between the placebo and ADS024 groups (permutational multivariate analysis of variance statistical test in R: $P = 0.617$), and all samples tended to cluster together regardless of treatment or time point (Figure 5C). At the phylum level, no significant differences were detected in the most abundant phyla (Bacillota, Bacteroidota, and Spirochaetota) between treatment groups across time points (Figure 6A). At the genus level, no significant differences were observed across the most abundant genera (*Lactobacillus* and *Streptococcus*) at any time point (Figure 6B).

ADS024 is susceptible to a variety of antibiotics and lacks antimicrobial resistance genes

To assess the potential of ADS024 to be a source of antibiotic resistance, its susceptibility to 18 different antibiotics was tested. The frozen glycerol stock and lyophilized forms of ADS024 samples were assessed as described under materials and methods.

Table 1 summarizes the MIC results for the 18 antimicrobials tested against one lot of ADS024 (4 lots were tested with similar results). All antimicrobial agents showed low MIC values when tested against ADS024, except for metronidazole (MIC ≥ 64 mg/L, depending on atmospheric conditions). ADS024 was susceptible to all 12 drugs tested for which interpretive criteria are available[28,29].

To determine the possible presence of virulence factors or antimicrobial resistance genes, the computational genomic prediction tool IslandViewer 4 was used. Corroborating statistical analyses already published[19], no virulence factors, homologs, or operons for antibiotic resistance were identified, pointing to the low potential of ADS024 to expose the gut microbiota to virulence and antibiotic-resistance genes after oral dosing (Supplementary Figure 1).

DISCUSSION

B. velezensis ADS024, isolated from a human fecal sample, is a member of the operational group *Bacillus amyloliquefaciens*, consisting of the soilborne *B. amyloliquefaciens* and the plant-associated *Bacillus siamensis* and *B. velezensis*. Most often, *B. amyloliquefaciens* is used in literature to describe the operational group *B. amyloliquefaciens* and refers to all three closely related species[30]. In this study, we extended our *in vitro* characterization of ADS024 to show its protective effects against CDI challenge in mouse models. We demonstrated a lack of impact on the gut microbiota composition in human feces and miniature swine models. Our model aligned with a CDI challenge rather than a recurrent model. The animals used in our study were not subjected to an earlier round of CDI or pre-treated with antibiotics, which would damage

Table 1 Antimicrobial susceptibility profiles of ADS024

Antimicrobial	MIC in mg/L (interpretation) for ADS024
	CO-33 ADS024 SYD
Ampicillin	≤ 0.03 (S)
Chloramphenicol	2 (S)
Ciprofloxacin	≤ 0.03 (S)
Clindamycin	0.25 (S)
Erythromycin	0.12 (S)
Fidaxomicin	4
Gentamicin	≤ 0.06 (S)
Kanamycin	≤ 0.50 (S)
Linezolid	0.5
Metronidazole (aerobic)	> 64.00
Metronidazole (anaerobic)	64
Neomycin	≤ 0.12
Penicillin	0.06 (S)
Quinupristin–dalfopristin	2
Rifampicin	0.25 (S)
Streptomycin	2 (S)
Tetracycline	2 (S)
Trimethoprim	≤ 0.25
Vancomycin	≤ 0.25 (S)

Four samples of ADS024 were tested. Shown here are the results using ADS024 from the master cell bank (CO-33 ART24 SYD; frozen). MIC: Minimum inhibitory concentration; S: Susceptible.

their GI tract microbiota, the 2 conditions associated with CDI recurrence.

Previous studies have investigated the *B. amyloliquefaciens* operational group in a mouse model as a prophylactic treatment for *C. difficile*. In one such study by Geeraerts *et al*[21], using C57BL/6J mice and *C. difficile* strain VPI 10463, it was demonstrated that *C. difficile* toxin A and B levels were significantly lower in *B. amyloliquefaciens*-treated mice compared with untreated ones[21]. In addition, a significantly lower extent of colonic tissue damage was observed for *B. amyloliquefaciens*-treated mice compared with the untreated cohort. Consistent with these findings, ADS024 conferred protection against CDI and its clinical manifestations.

Although not directly tested in the experiments shown in this study, one possibility is that the pharmacologic effect of ADS024 was due to its bactericidal activity against *C. difficile* as well as its proteolytic degradation of TcdA and TcdB[19], resulting in fewer observed clinical symptoms. Furthermore, an anti-inflammatory role of *Bacillus*-based probiotics has been demonstrated[20,31]. To this end, the *Bacillus*-based probiotic, Bs 29784, decreased pro-inflammatory signals in a Caco-2 cell model for intestinal epithelia, prevented IkBα degradation, and reduced upregulation of iNOS protein levels, highlighting intestinal anti-inflammatory capabilities[31]. Similarly, ADS024 inhibited toxin B-mediated apoptosis and tissue injury in human colonic epithelial cells and colonic explants[20]. In addition, restoring the host gut microbiota to preexisting conditions plays an equally critical role in recovering from CDI and preventing recurrent infection after the completion of standard antibiotic treatment. We hypothesize that the selective impact of ADS024 on the human microbiota may therefore allow recovery of the normal host microbiota and prevent rCDI in clinical settings.

The aim of the mouse model (Figure 2) was to evaluate whether prior dosing with ADS024 could protect against subsequent challenge by *C. difficile*. We found that single daily (QD) doses of ADS024 were as efficacious as multiple daily (TID) doses in protecting against pathogen-induced disease (Figure 2), and, as reported in the result section of the Study 2 Mouse model of CDI, there was no significant benefit in the TID-dosed group (Figure 2D-F). While there was a slight improvement in the weight loss category in the TID-dosed group compared with the QD-dosed group (14% vs 16%), both groups showed improvement compared with the untreated group (25%) (Figure 2D). In contrast, the adverse clinical features were present only in the TID-dosed group (Figure 2E and F), possibly due to the added stress in the animals from repeated oral gavage[32], which likely negated any positive effect of ADS024. Stress would likely increase assessed disease markers if there were an infected placebo arm with the same dosing regimen.

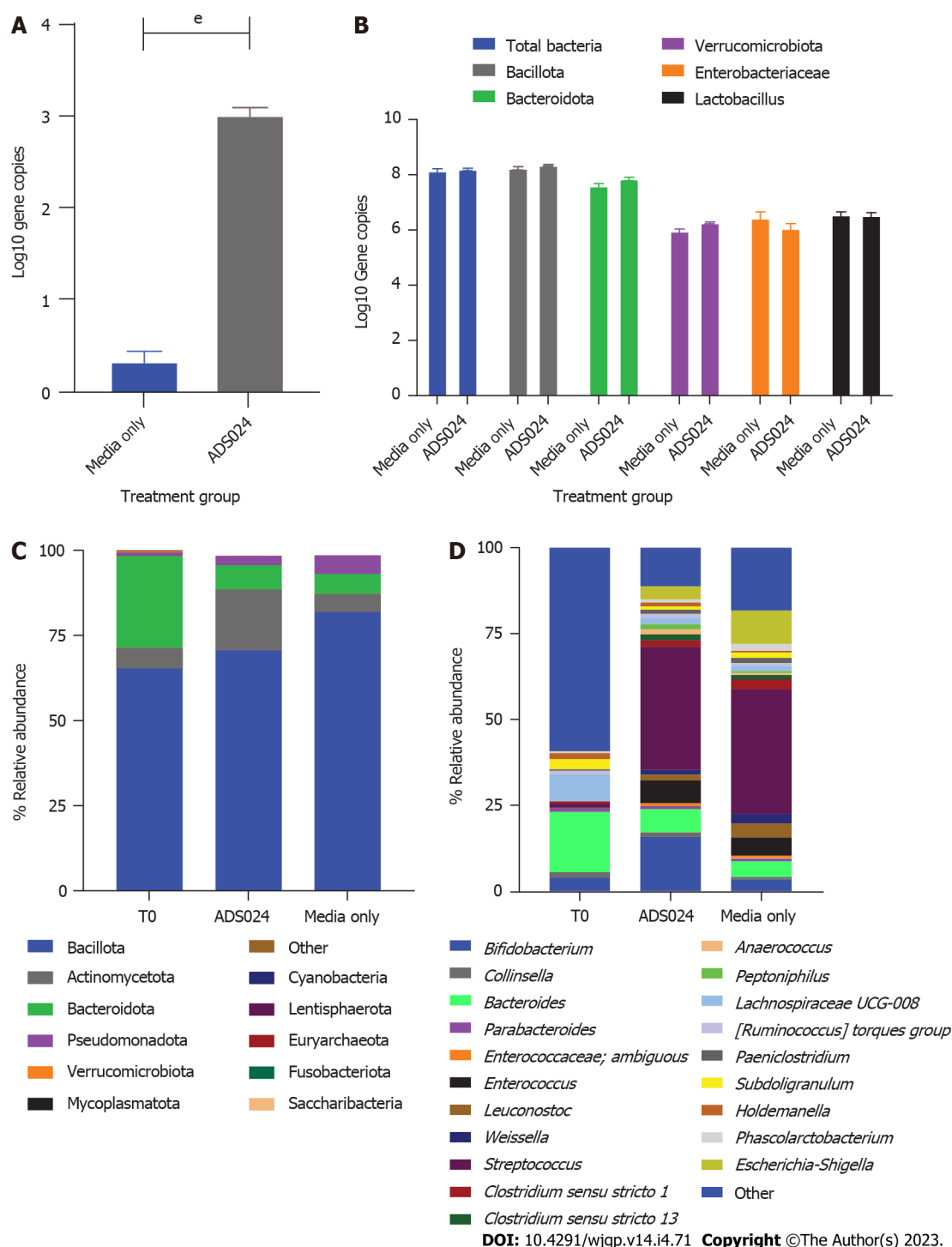


Figure 3 Distal colon model. A: Comparison of the wells inoculated with ADS024 vs media alone was done via quantitative polymerase chain reaction (qPCR) using specific primers to detect ADS024 as described in Materials and Methods; B: qPCR was performed to study the antimicrobial activity of ADS024 on 5 different phyla of the human colonic microbiota; C: MiSeq compositional sequencing analysis was performed to compare the impact of ADS024 on the gut microbiota at the phylum level with that of media alone; D: MiSeq was used to determine the ADS024 effect after 24 h of treatment at the genus level. * $P \leq 0.001$.

ADS024 showed no evidence of colonization (Figure 1) based on the observation that the ADS024 colonies were not recovered 24 h after single doses in mice or 72 h after single doses in minipigs. This suggests that the ADS024-mediated protection against *C. difficile* pathogenesis occurred independently of the colonization of ADS024 in the animals' GI tracts.

ADS024 was also assessed in a 28-d repeat-dose study in miniature swine (Figure 5 and 6), a species that offers a more clinically relevant model of the human GI tract[33]. ADS024 was not detected in fecal samples using plating and qPCR methods (Supplementary Figure 4), which again suggests that ADS024 did not colonize the gut following long-term treatment exposures. To achieve reproducible results with microbial therapeutics in humans, it may be advantageous to use transitory, non-colonizing strains[34], such as ADS024.

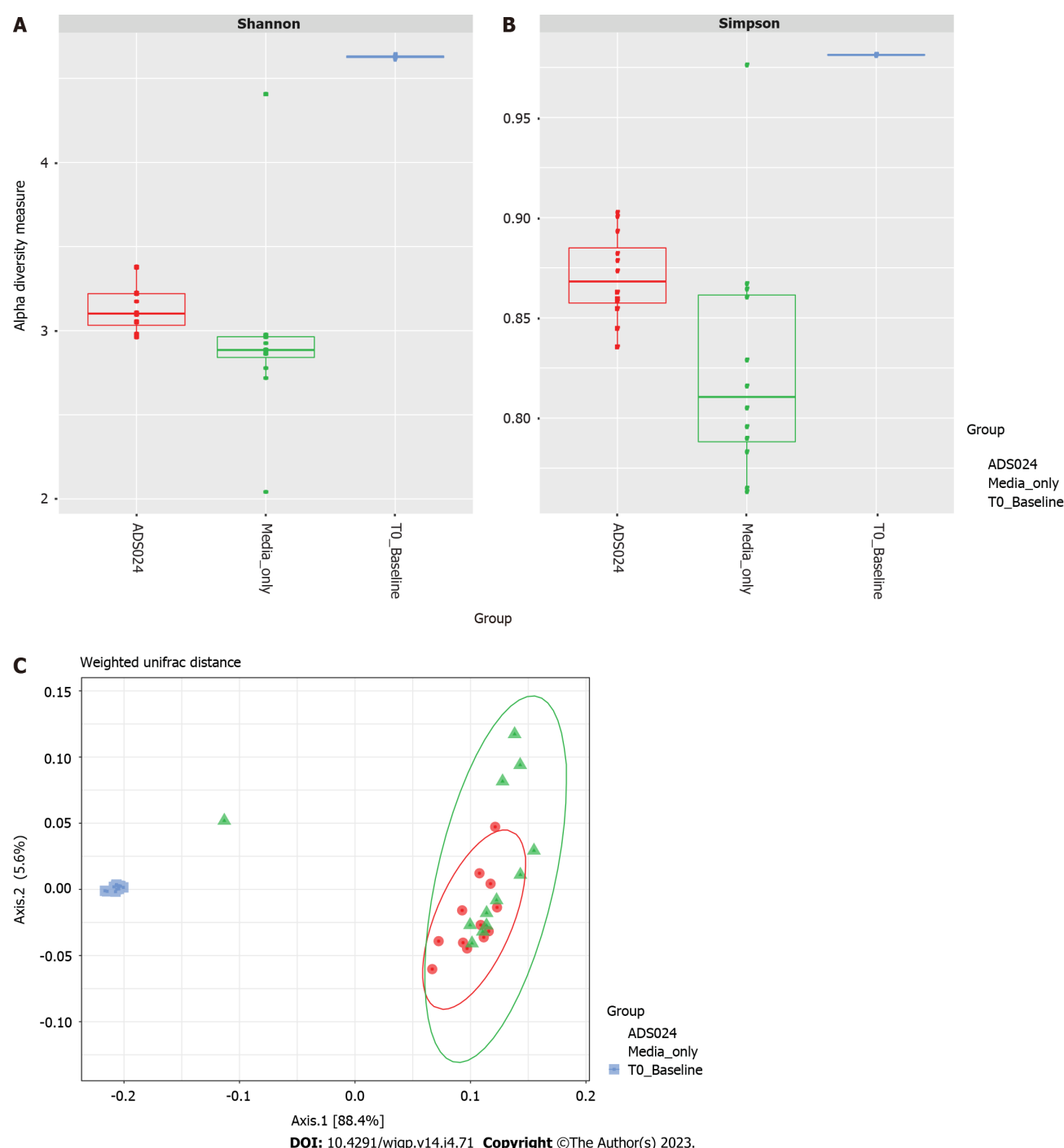


Figure 4 Distal colon model: Alpha and beta diversity analyses. MiSeq compositional sequencing and bioinformatic analysis were performed to measure the microbiota diversity changes following ADS024 treatment using the: A: Shannon index for alpha diversity; B: Simpson diversity index for alpha diversity; C: UniFrac principal coordinates analysis for beta diversity.

Restoring the resident host microbiota is an important factor in preventing rCDI. Thus, the effect of ADS024 on gut microbial composition was investigated using a human distal colon model (Figures 3 and 4). *Ex vivo* systems replicating the distal colon environment have been used to investigate the effect of antimicrobial-producing bacterial strains on the human gut microbiota[35]. To our knowledge, this is the first report examining the impact of a *B. amyloliquefaciens* operational group strain in a distal colon model using a standardized human fecal inoculum. Phylogenetic analysis performed in this model showed that ADS024 had a selective impact on the healthy human colonic microbiota (Figures 3 and 4). This finding is similar to the *in vivo* studies performed in miniature swine (Figures 5 and 6). We consider this selective impact of ADS024 on the resident commensal microbiota an important feature for its use as a therapeutic agent to prevent rCDI. Specifically, *Bifidobacterium* species expansion in ADS024-treated samples (Figure 3C and D) was noteworthy. The correlation between *Bifidobacterium* species consumption and improved human health has been well documented[35]. Various illnesses[36,37] and aging[36-38] have been associated with reduced numbers of *Bifidobacterium* species, which, in a healthy population, positively impact the immune system[39] and are thus important for health

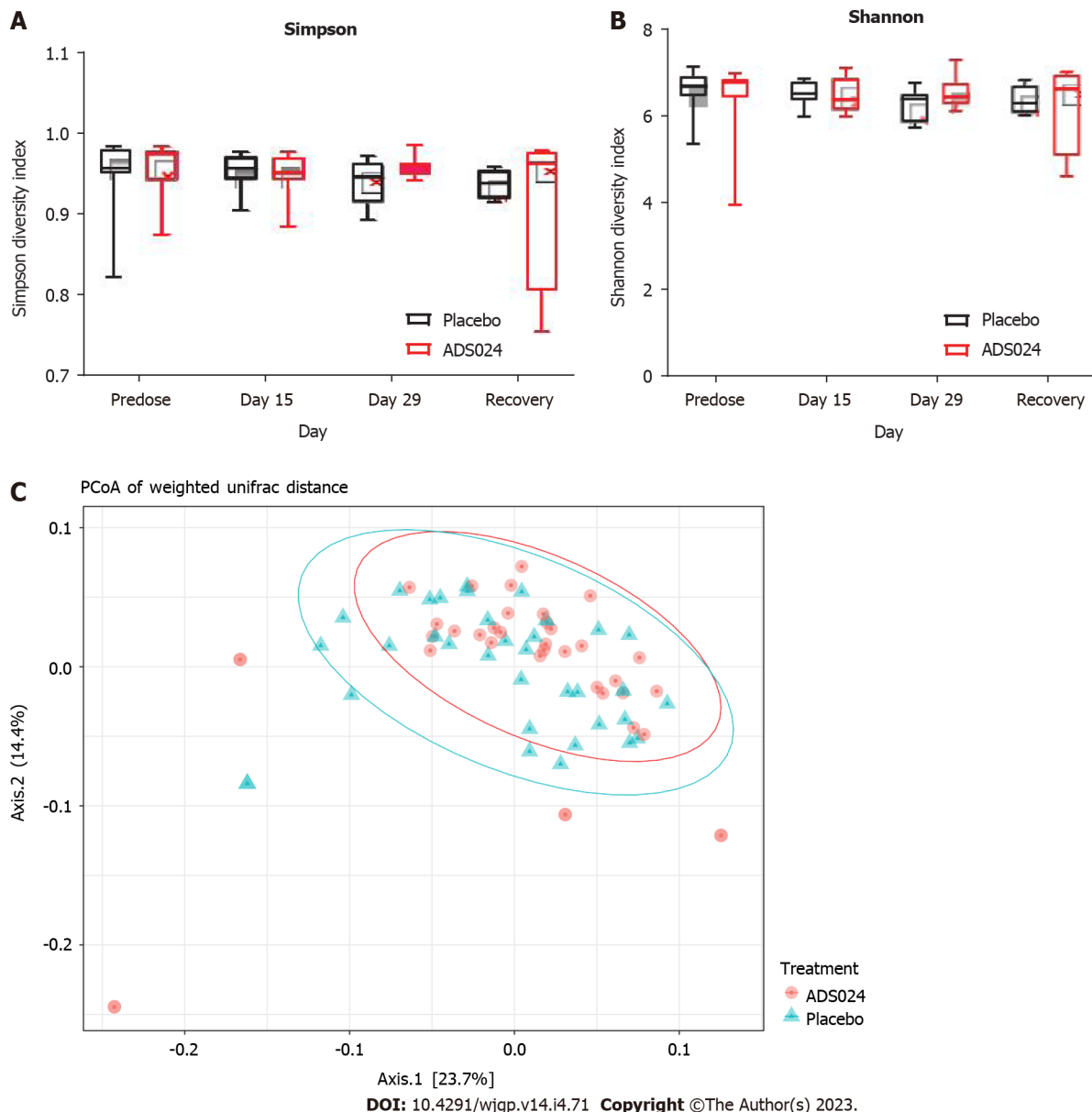


Figure 5 The impact of ADS024 on microbiota diversity in the miniature swine fecal samples during the 28-d study. MiSeq compositional sequencing and bioinformatic analysis were performed to measure the impact of ADS024 on the microbiota in miniature swine fecal samples after 28 d using the: A: Shannon index for alpha diversity; B: Simpson diversity index for alpha diversity; C: UniFrac principal coordinates analysis (PCoA) for beta diversity.

maintenance[36]. ADS024 appeared to have the added benefit of expanding this healthy bacterial population.

Recent research demonstrates the benefits of consuming *B. velezensis* isolates and their role in regulating the intestinal innate immune response and decreasing intestinal inflammation caused by pathogens in crucian carp[40]. The immunomodulatory and anti-inflammatory effects of *B. velezensis* and other related *Bacillus* species have also been studied in plants and mice[41,42], in addition to the human epithelial cells described above[31], showing their potential for treating inflammatory diseases caused by pathogens like *C. difficile*. By killing *C. difficile* and degrading its toxins[19], thereby eliminating inflammatory stimuli, ADS024 may also have offered anti-inflammatory protection, a crucial asset in tackling an inflammatory disease such as CDI.

A limitation of our studies was using the *in vitro* human distal colon model, which has limited physiological relevance. A batch fermentation system inevitably precludes the exchange of nutrients or waste removal. The accumulation of waste can potentially influence the bacterial microenvironment. Future assessments will include tissue histology and co-localization (*C. difficile*–ADS024) experiments to further characterize the mechanistic details and therapeutic effects of ADS024. Furthermore, a definitive demonstration that the direct activities of ADS024 on *C. difficile* are responsible for its biological activity in mice will require *in vivo* studies utilizing *C. difficile*–infected gnotobiotic mice.

Although safety in humans requires further study, our ADS024 findings were promising and support the continued development of ADS024 as an SS-LBP to prevent CDI recurrence following the completion of standard antibiotic treatment. In contrast to other LBPs that aim to restore diversity and prevent outgrowth by outcompeting the pathogen, we propose that ADS024 functioned by delivering bioactivities (described in O'Donnell *et al*[19,27]) directly targeting *C. difficile* and its toxins. Fecal microbiota transplantation and most LBPs may require antibiotic administration prior to

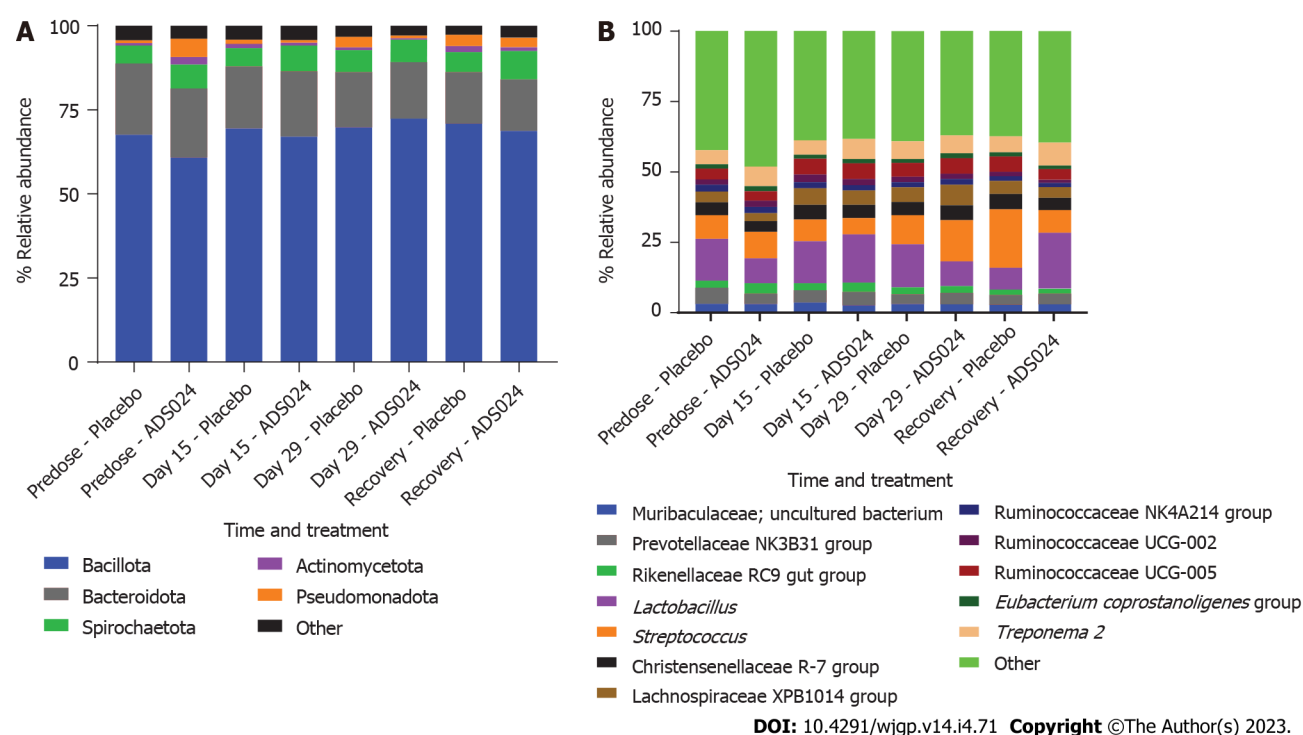


Figure 6 Effect of ADS024 on the gut microbiota of miniature swine during the 28-d study at the phylum and genus levels. A and B: MiSeq compositional sequencing and bioinformatic analysis was performed to measure the impact of ADS024 on the gut microbiota of miniature swine at the (A) phylum and (B) genus levels.

treatment to allow engraftment[43]. The lack of colonization by ADS024 contrasts with other LBPs, which depend on colonization. Given the emerging data on how microbially produced metabolites can modulate human pharmacology, the most minimally invasive, transient LBP intervention (ADS024) may be preferable. Transplanted microbiota, while effectively reducing rCDI, may cause chronic changes to the patient's natural GI microbiome as a cost of treating CDI[43, 44].

It is clear that while the burgeoning LBP field has high therapeutic potential, method optimization and the identification of selective products with fewer systemic side effects require substantially increased effort. To this end, a risk assessment should be undertaken in the early stages of LBP development[23]. The risk assessment and safety analysis of ADS024 demonstrated it to be selective, with little potential for colonizing the gut (mouse and miniature swine models), and susceptible to all the antibiotics tested for which interpretative criteria are available (Table 1). If a patient were to develop a systemic infection with ADS024 (which is not expected due to the lack of virulence factors in ADS024), a wide range of antibiotics would be available to treat the infection. *In silico* safety assessments indicated that ADS024 had a low potential for off-target activity or virulence and antibiotic-resistance mechanisms (Table 1; Supplementary Figure 1).

CONCLUSION

In vivo evaluation of ADS024 is consistent with the results of previous *in vitro* studies from our group[19,20] that showed potent *C. difficile* inhibition and maintenance of gut microbiota diversity. The totality of the evidence to date supports additional preclinical studies and early-phase clinical trials to evaluate the safety and efficacy of ADS024 in patients recovering from CDI.

ARTICLE HIGHLIGHTS

Research background

Clostridioides difficile (*C. difficile*) is a Gram positive pathogen that causes *C. difficile* infection (CDI). It infects millions of people worldwide, causing potentially life-threatening gastrointestinal disease in individuals with disrupted gut microbiomes. Following successful antibiotic treatment, recurrent CDI (rCDI) can occur in cured patients. Therefore, development of a therapeutic product for preventing rCDI following successful standard-of-care antibiotic therapy is of vital importance.

Research motivation

C. difficile, a major cause of infectious disease death in the United States, causes inflammation of the colon and potentially deadly diarrhea. ADS024, a *Bacillus velezensis* strain, has previously demonstrated direct *in vitro* bactericidal activity against *C. difficile*, without affecting other members of the gut microbiota. In this study, we investigated the efficacy of ADS024 against CDI challenge *in vivo*. Following our findings, further investigation of ADS024 as a single-strain, live biotherapeutic product (SS-LBP) for prevention of rCDI following successful standard-of-care antibiotic therapy is warranted.

Research objectives

The objectives of the research were to investigate: (1) The *in vivo* efficacy of *B. velezensis* ADS024 in protecting against CDI challenge in mouse models; (2) the capability of ADS024 to colonize the GI tract; and (3) the impact of ADS024 on the gut microbiome, finding that it was efficacious against CDI challenge without colonization, and with minimal effects on the gut microbiome, thus supporting further development of ADS024 as an SS-LBP for prevention of rCDI.

Research methods

Mouse models were used to determine: (1) The *in vivo* efficacy against CDI challenge; and (2) colonization status of ADS024 (in conjunction with miniature swine). Human distal colon model and miniature swine were utilized to determine the effects of ADS024 on the gut microbiome. To mimic disruption of the gut microbiota, the mice and miniature swine were exposed to vancomycin prior to dosing with ADS024. To model the human distal colon, an anaerobic fecal fermentation system was used.

Research results

Single oral daily doses of ADS024, similarly to multiple doses, demonstrated efficacy in protecting against subsequent challenge by *C. difficile* in a mouse model of CDI challenge. ADS024 showed no colonization based on lack of recovery ADS024 colonies in fecal samples 24 h after single doses in mice, 72 h after single doses in miniature swine, or a 28-d repeat-dose study in miniature swine. Phylogenetic analysis in the human distal colon model and *in vivo* studies performed in miniature swine demonstrated a selective impact of ADS024 on the healthy human colonic microbiota.

Research conclusions

In vivo efficacy of ADS024 in protecting against CDI challenge and minimal effects on the gut microbiome support development of ADS024 as an SS-LBP in preventing rCDI following standard antibiotic treatment.

Research perspectives

In vivo investigation of ADS024 is compatible with previous *in vitro* studies that showed efficacy against *C. difficile* and maintenance of gut microbiota diversity. Altogether, findings from these studies support initiation of clinical trials to evaluate the safety and efficacy of ADS024 in patients recovering from CDI.

ACKNOWLEDGEMENTS

The authors would like to thank Lorena Puto, PhD, of Simpson Healthcare, for her assistance in medical writing and editorial support. Her participation was provided by Adiso Therapeutics, Inc.

FOOTNOTES

Author contributions: Murphy CK, O'Donnell MM, Hill C, Ross RP, Rea MC, Farquhar R, and Chesnel L designed the experiments; O'Donnell MM, Hegarty JW, and Schulz S conducted the research; data was acquired and interpreted by O'Donnell MM, Hegarty JW, Schulz S, and Chesnel L; Chesnel L drafted the manuscript; all authors reviewed the manuscript and approved the final version.

Institutional animal care and use committee statement: All procedures involving animals were reviewed and approved by the Institutional Animal Care and Use Committee of NeoSome Life Sciences (IACUC Protocol No. NLS17-002).

Conflict-of-interest statement: Hill C, Ross RP, and Rea MC received consulting fees from Adiso Therapeutics, Inc. Murphy CK, Farquhar R, and Chesnel L are employees of Adiso Therapeutics, Inc. O'Donnell MM, Hegarty JW, and Schulz S are supported by an Adiso Therapeutics, Inc., research grant to UCC/APC.

Data sharing statement: No additional data are available.

ARRIVE guidelines statement: The authors have read the ARRIVE guidelines, and the manuscript was prepared and revised according to the ARRIVE guidelines.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to

distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

Country/Territory of origin: United States

ORCID number: Christopher K Murphy 0000-0003-3791-8854; Sarah Schulz 0000-0002-7583-1293; Colin Hill 0000-0002-8527-1445; R Paul Ross 0000-0003-4876-8839; Mary C Rea 0000-0003-3782-0232; Laurent Chesnel 0000-0002-4947-5029.

S-Editor: Chen YL

L-Editor: A

P-Editor: Yuan YY

REFERENCES

- Johnson S, Lavergne V, Skinner AM, Gonzales-Luna AJ, Garey KW, Kelly CP, Wilcox MH. Clinical Practice Guideline by the Infectious Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA): 2021 Focused Update Guidelines on Management of Clostridioides difficile Infection in Adults. *Clin Infect Dis* 2021; **73**: e1029-e1044 [PMID: 34164674 DOI: 10.1093/cid/ciab549]
- Nagy E. What do we know about the diagnostics, treatment and epidemiology of Clostridioides (Clostridium) difficile infection in Europe? *J Infect Chemother* 2018; **24**: 164-170 [PMID: 29289484 DOI: 10.1016/j.jiac.2017.12.003]
- Lessa FC, Mu Y, Bamberg WM, Beldavs ZG, Dumyati GK, Dunn JR, Farley MM, Holzbauer SM, Meek JI, Phipps EC, Wilson LE, Winston LG, Cohen JA, Limbago BM, Fridkin SK, Gerding DN, McDonald LC. Burden of Clostridium difficile infection in the United States. *N Engl J Med* 2015; **372**: 825-834 [PMID: 25714160 DOI: 10.1056/NEJMoa1408913]
- Morrison RH, Hall NS, Said M, Rice T, Groff H, Brodine SK, Slymen D, Lederman ER. Risk factors associated with complications and mortality in patients with Clostridium difficile infection. *Clin Infect Dis* 2011; **53**: 1173-1178 [PMID: 21976459 DOI: 10.1093/cid/cir668]
- Buffie CG, Jarchum I, Equinda M, Lipuma L, Gobourne A, Viale A, Ubeda C, Xavier J, Pamer EG. Profound alterations of intestinal microbiota following a single dose of clindamycin results in sustained susceptibility to Clostridium difficile-induced colitis. *Infect Immun* 2012; **80**: 62-73 [PMID: 22006564 DOI: 10.1128/IAI.05496-11]
- Guh AY, Kutty PK. Clostridioides difficile Infection. *Ann Intern Med* 2018; **169**: ITC49-ITC64 [PMID: 30285209 DOI: 10.7326/AITC201810020]
- Carter GP, Chakravorty A, Pham Nguyen TA, Mileto S, Schreiber F, Li L, Howarth P, Clare S, Cunningham B, Sambol SP, Cheknis A, Figueroa I, Johnson S, Gerding D, Rood JI, Dougan G, Lawley TD, Lytras D. Defining the Roles of TcdA and TcdB in Localized Gastrointestinal Disease, Systemic Organ Damage, and the Host Response during Clostridium difficile Infections. *mBio* 2015; **6**: e00551 [PMID: 26037121 DOI: 10.1128/mBio.00551-15]
- Wilcox MH, Gerding DN, Poxton IR, Kelly C, Nathan R, Birch T, Cornely OA, Rahav G, Bouza E, Lee C, Jenkin G, Jensen W, Kim YS, Yoshida J, Gabryelski L, Pedley A, Eves K, Tipping R, Guris D, Kartsonis N, Dorr MB; MODIFY I and MODIFY II Investigators. Bezlotoxumab for Prevention of Recurrent Clostridium difficile Infection. *N Engl J Med* 2017; **376**: 305-317 [PMID: 28121498 DOI: 10.1056/NEJMoa1602615]
- Steele J, Mukherjee J, Parry N, Tzipori S. Antibody against TcdB, but not TcdA, prevents development of gastrointestinal and systemic Clostridium difficile disease. *J Infect Dis* 2013; **207**: 323-330 [PMID: 23125448 DOI: 10.1093/infdis/jis669]
- Tsigrelis C. Recurrent Clostridioides difficile infection: Recognition, management, prevention. *Cleve Clin J Med* 2020; **87**: 347-359 [PMID: 32487555 DOI: 10.3949/ccjm.87gr.20001]
- Kelly CP. Can we identify patients at high risk of recurrent Clostridium difficile infection? *Clin Microbiol Infect* 2012; **18** Suppl 6: 21-27 [PMID: 23121551 DOI: 10.1111/1469-0691.12046]
- Dawkins JJ, Allegretti JR, Gibson TE, McClure E, Delaney M, Bry L, Gerber GK. Gut metabolites predict Clostridioides difficile recurrence. *Microbiome* 2022; **10**: 87 [PMID: 35681218 DOI: 10.1186/s40168-022-01284-1]
- Panchavati S, Zelin NS, Garikipati A, Pellegrini E, Iqbal Z, Barnes G, Hoffman J, Calvert J, Mao Q, Das R. A comparative analysis of machine learning approaches to predict *C. difficile* infection in hospitalized patients. *Am J Infect Control* 2022; **50**: 250-257 [PMID: 35067382 DOI: 10.1016/j.ajic.2021.11.012]
- Rashid MU, Zaura E, Buijs MJ, Keijser BJ, Crielaard W, Nord CE, Weintraub A. Determining the Long-term Effect of Antibiotic Administration on the Human Normal Intestinal Microbiota Using Culture and Pyrosequencing Methods. *Clin Infect Dis* 2015; **60** Suppl 2: S77-S84 [PMID: 25922405 DOI: 10.1093/cid/civ137]
- Jakobsson HE, Jernberg C, Andersson AF, Sjölund-Karlsson M, Jansson JK, Engstrand L. Short-term antibiotic treatment has differing long-term impacts on the human throat and gut microbiome. *PLoS One* 2010; **5**: e9836 [PMID: 20352091 DOI: 10.1371/journal.pone.0009836]
- Jernberg C, Löfmark S, Edlund C, Jansson JK. Long-term ecological impacts of antibiotic administration on the human intestinal microbiota. *ISME J* 2007; **1**: 56-66 [PMID: 18043614 DOI: 10.1038/ismej.2007.3]
- Zar FA, Bakkanagari SR, Moorthi KM, Davis MB. A comparison of vancomycin and metronidazole for the treatment of Clostridium difficile-associated diarrhea, stratified by disease severity. *Clin Infect Dis* 2007; **45**: 302-307 [PMID: 17599306 DOI: 10.1086/519265]
- Johnson S, Louie TJ, Gerding DN, Cornely OA, Chasan-Taber S, Fitts D, Gelone SP, Broom C, Davidson DM; Polymer Alternative for CDI Treatment (PACT) investigators. Vancomycin, metronidazole, or tolevamer for Clostridium difficile infection: results from two multinational, randomized, controlled trials. *Clin Infect Dis* 2014; **59**: 345-354 [PMID: 24799326 DOI: 10.1093/cid/ciu313]
- O'Donnell MM, Hegarty JW, Healy B, Schulz S, Walsh CJ, Hill C, Ross RP, Rea MC, Farquhar R, Chesnel L. Identification of ADS024, a newly characterized strain of Bacillus velezensis with direct Clostridioides difficile killing and toxin degradation bio-activities. *Sci Rep* 2022; **12**: 9283 [PMID: 35662257 DOI: 10.1038/s41598-022-13248-4]
- Xie Y, Chupina Estrada A, Nelson B, Feng H, Pothoulakis C, Chesnel L, Koon HW. ADS024, a Bacillus velezensis strain, protects human colonic epithelial cells against *C. difficile* toxin-mediated apoptosis. *Front Microbiol* 2022; **13**: 1072534 [PMID: 36704560 DOI: 10.3389/fmicb.2022.1072534]

- 10.3389/fmicb.2022.1072534]
- 21 **Geeraerts S**, Ducatelle R, Haesebrouck F, Van Immerseel F. *Bacillus amyloliquefaciens* as prophylactic treatment for *Clostridium difficile*-associated disease in a mouse model. *J Gastroenterol Hepatol* 2015; **30**: 1275-1280 [PMID: 25800047 DOI: 10.1111/jgh.12957]
 - 22 **US Food and Drug Administration**, Center for Biologics Evaluation and Research. Early Clinical Trials With Live Biotherapeutic Products: Chemistry, Manufacturing, and Control Information. Guidance for Industry. FDA-2010-D-0500. [cited 1 August 2023]. Available from: <https://www.fda.gov/regulatory-information/search-fda-guidance-documents/early-clinical-trials-live-biotherapeutic-products-chemistry-manufacturing-and-control-information>
 - 23 **Rouanet A**, Bolca S, Bru A, Claes I, Cvejic H, Girgis H, Harper A, Lavergne SN, Mathys S, Pane M, Pot B, Shortt C, Alkema W, Bezulowsky C, Blanquet-Diot S, Chassard C, Claus SP, Hadida B, Hemmingsen C, Jeune C, Lindman B, Midzi G, Mogna L, Movitz C, Nasir N, Oberreither M, Seegers JFML, Sterkman L, Valo A, Vieville F, Cordaillat-Simmons M. Live Biotherapeutic Products, A Road Map for Safety Assessment. *Front Med (Lausanne)* 2020; **7**: 237 [PMID: 32637416 DOI: 10.3389/fmed.2020.00237]
 - 24 **McFarland LV**. Efficacy of Single-Strain Probiotics Versus Multi-Strain Mixtures: Systematic Review of Strain and Disease Specificity. *Dig Dis Sci* 2021; **66**: 694-704 [PMID: 32274669 DOI: 10.1007/s10620-020-06244-z]
 - 25 **Cordaillat-Simmons M**, Rouanet A, Pot B. Live biotherapeutic products: the importance of a defined regulatory framework. *Exp Mol Med* 2020; **52**: 1397-1406 [PMID: 32908212 DOI: 10.1038/s12276-020-0437-6]
 - 26 **Chen X**, Katchar K, Goldsmith JD, Nanthakumar N, Cheknis A, Gerding DN, Kelly CP. A mouse model of *Clostridium difficile*-associated disease. *Gastroenterology* 2008; **135**: 1984-1992 [PMID: 18848941 DOI: 10.1053/j.gastro.2008.09.002]
 - 27 **O'Donnell MM**, Rea MC, Shanahan F, Ross RP. The Use of a Mini-Bioreactor Fermentation System as a Reproducible, High-Throughput *ex vivo* Batch Model of the Distal Colon. *Front Microbiol* 2018; **9**: 1844 [PMID: 30147684 DOI: 10.3389/fmicb.2018.01844]
 - 28 **Clinical and Laboratory Standards Institute**. M45: Methods for antimicrobial dilution and disk susceptibility testing of infrequently isolated or fastidious bacteria. 3rd ed. 2016. [cited 1 August 2023]. Available from: https://clsi.org/media/1450/m45ed3_sample.pdf
 - 29 **EFSA Panel on Additives and Products or Substances used in Animal Feed (FEEDAP)**. Guidance on the assessment of bacterial susceptibility to antimicrobials of human and veterinary importance. *EFSA Journal* 2012; **10**: 2740 [DOI: 10.2903/j.efsa.2012.2740]
 - 30 **Fan B**, Blom J, Klenk HP, Borris R. *Bacillus amyloliquefaciens*, *Bacillus velezensis*, and *Bacillus siamensis* Form an "Operational Group B. amyloliquefaciens" within the *B. subtilis* Species Complex. *Front Microbiol* 2017; **8**: 22 [PMID: 28163698 DOI: 10.3389/fmicb.2017.00022]
 - 31 **Rhayat L**, Maresca M, Nicoletti C, Perrier J, Brinch KS, Christian S, Devillard E, Eckhardt E. Effect of *Bacillus subtilis* Strains on Intestinal Barrier Function and Inflammatory Response. *Front Immunol* 2019; **10**: 564 [PMID: 30984172 DOI: 10.3389/fimmu.2019.00564]
 - 32 **Turner PV**, Brabb T, Pekow C, Vasbinder MA. Administration of substances to laboratory animals: routes of administration and factors to consider. *J Am Assoc Lab Anim Sci* 2011; **50**: 600-613 [PMID: 22330705]
 - 33 **Zhang Q**, Widmer G, Tzipori S. A pig model of the human gastrointestinal tract. *Gut Microbes* 2013; **4**: 193-200 [PMID: 23549377 DOI: 10.4161/gmic.23867]
 - 34 **Charbonneau MR**, Isabella VM, Li N, Kurtz CB. Developing a new class of engineered live bacterial therapeutics to treat human diseases. *Nat Commun* 2020; **11**: 1738 [PMID: 32269218 DOI: 10.1038/s41467-020-15508-1]
 - 35 **Dobson A**, Crispie F, Rea MC, O'Sullivan O, Casey PG, Lawlor PG, Cotter PD, Ross P, Gardiner GE, Hill C. Fate and efficacy of lacticin 3147-producing *Lactococcus lactis* in the mammalian gastrointestinal tract. *FEMS Microbiol Ecol* 2011; **76**: 602-614 [PMID: 21314706 DOI: 10.1111/j.1574-6941.2011.01069.x]
 - 36 **Arbolea S**, Watkins C, Stanton C, Ross RP. Gut Bifidobacteria Populations in Human Health and Aging. *Front Microbiol* 2016; **7**: 1204 [PMID: 27594848 DOI: 10.3389/fmicb.2016.01204]
 - 37 **O'Neill I**, Schofield Z, Hall LJ. Exploring the role of the microbiota member Bifidobacterium in modulating immune-linked diseases. *Emerg Top Life Sci* 2017; **1**: 333-349 [PMID: 33525778 DOI: 10.1042/ETLS20170058]
 - 38 **Hopkins MJ**, Macfarlane GT. Changes in predominant bacterial populations in human faeces with age and with *Clostridium difficile* infection. *J Med Microbiol* 2002; **51**: 448-454 [PMID: 11990498 DOI: 10.1099/0022-1317-51-5-448]
 - 39 **Ménard O**, Butel MJ, Gaboriau-Routhiau V, Waligora-Dupriet AJ. Gnotobiotic mouse immune response induced by Bifidobacterium sp. strains isolated from infants. *Appl Environ Microbiol* 2008; **74**: 660-666 [PMID: 18083875 DOI: 10.1128/AEM.01261-07]
 - 40 **Zhang DX**, Kang YH, Zhan S, Zhao ZL, Jin SN, Chen C, Zhang L, Shen JY, Wang CF, Wang GQ, Shan XF, Qian AD. Effect of *Bacillus velezensis* on *Aeromonas veronii*-Induced Intestinal Mucosal Barrier Function Damage and Inflammation in Crucian Carp (*Carassius auratus*). *Front Microbiol* 2019; **10**: 2663 [PMID: 31798571 DOI: 10.3389/fmicb.2019.02663]
 - 41 **Paynich ML**, Jones-Burrage SE, Knight KL. Exopolysaccharide from *Bacillus subtilis* Induces Anti-Inflammatory M2 Macrophages That Prevent T Cell-Mediated Disease. *J Immunol* 2017; **198**: 2689-2698 [PMID: 28202619 DOI: 10.4049/jimmunol.1601641]
 - 42 **Tzipilevich E**, Russ D, Dangel JL, Benfey PN. Plant immune system activation is necessary for efficient root colonization by auxin-secreting beneficial bacteria. *Cell Host Microbe* 2021; **29**: 1507-1520.e4 [PMID: 34610294 DOI: 10.1016/j.chom.2021.09.005]
 - 43 **Freitag TL**, Hartikainen A, Jouhten H, Sahl C, Meri S, Anttila VJ, Mattila E, Arkkila P, Jalanka J, Satokari R. Minor Effect of Antibiotic Pre-treatment on the Engraftment of Donor Microbiota in Fecal Transplantation in Mice. *Front Microbiol* 2019; **10**: 2685 [PMID: 31824463 DOI: 10.3389/fmicb.2019.02685]
 - 44 **Feuerstadt P**, Louie TJ, Lashner B, Wang EEL, Diao L, Bryant JA, Sims M, Kraft CS, Cohen SH, Berenson CS, Korman LY, Ford CB, Litcofsky KD, Lombardo MJ, Wortman JR, Wu H, Aunins JG, McChalicher CWJ, Winkler JA, McGovern BH, Trucksis M, Henn MR, von Moltke L. SER-109, an Oral Microbiome Therapy for Recurrent *Clostridioides difficile* Infection. *N Engl J Med* 2022; **386**: 220-229 [PMID: 35045228 DOI: 10.1056/NEJMoa2106516]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

