

World Journal of *Respirology*

World J Respirol 2012 February 28; 2(1): 1-5





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2011-2015

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**BRIEF ARTICLE****1**

Effect of oxidative stress on development of silicosis

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World Journal of Respiriology (*World J Respiriol*, *WJR*, online ISSN 2218-6255, DOI: 10.5320) is a bimonthly peer-reviewed, online, open-access (OA), journal supported by an editorial board consisting of 84 experts in respirology from 30 countries.

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NAME OF JOURNAL
World Journal of Respiriology

ISSN
ISSN 2218-6255 (online)

LAUNCH DATE
December 30, 2011

FREQUENCY
Bimonthly

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PUBLISHER
Baishideng Publishing Group Co., Limited
Room 1701, 17/F, Henan Building,
No.90 Jaffe Road, Wanchai, Hong Kong, China
Fax: +852-31158812
Telephone: +852-58042046
E-mail: bpg@baishideng.com
<http://www.wjgnet.com>

PUBLICATION DATE
February 28, 2012

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ONLINE SUBMISSION
<http://www.wjgnet.com/2218-6255/esps/>

Effect of oxidative stress on development of silicosis

Rong-Ming Miao, Xue-Tao Zhang, Ping Guo, En-Qi He, Fang Zhou, Dao-Kun Zhao, Ying-Yi Zhang

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Received: December 10, 2011 Revised: February 10, 2012

Accepted: February 18, 2012

Published online: February 28, 2012

± 35.176 , 270.469 ± 39.228 and 68.209 ± 21.528 , respectively, $P = 0.004$, $P = 0.002$, $P = 0.005$). Compared with the control and dust-exposed group, T-AOC, NOS and MDA in silicosis group increased significantly (13.048 ± 4.153 , 36.201 ± 7.782 and 5.054 ± 1.204 , respectively, $P = 0.018$, $P = 0.022$, $P = 0.011$). Compared with dust-exposed group, GSH-Px in the silicosis group increased significantly (270.469 ± 39.228 , $P = 0.002$). GSH-Px in phase III silicosis was significantly higher than in phase I silicosis (290.750 ± 39.129 , $P = 0.021$). Pearson correlation analysis showed that serum GSH-Px was positively correlated with silicosis staging, length of dust exposure and type of occupation (47.109 ± 8.015 , $P = 0.001$).

CONCLUSION: The imbalance of oxidative and anti-oxidation system is associated with the development of silicosis. The surveillance of oxidative stress indicators will benefit the prognosis of silicosis patients.

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Abstract

AIM: To investigate the changes of oxidative stress indicators in the serum of silicosis patients and explore the mechanism of silicosis development.

METHODS: Two hundred workers who were exposed to silica dust for more than one year were recruited as dust-exposed group, 100 non-dust-exposed subjects served as control group, 32 patients with suspected 0^+ silicosis as observation group, and 130 silicosis patients were taken as the silicosis group. Indicators of oxidative stress, including superoxide dismutase (SOD), nitric oxide (NO), serum glutathione peroxidase (GSH-Px), total antioxidant capacity (T-AOC), nitric oxide synthase (NOS), and lipid malondialdehyde (MDA), were determined in all the groups.

RESULTS: Compared with the control group, NO and GSH-Px in dust-exposed group and silicosis group increased, and SOD decreased significantly (81.162

Key words: Silicosis; Oxidative stress; Superoxide dismutase; Glutathione peroxidase; Total antioxidant capacity; Nitric oxide synthase; Malondialdehyde

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Miao RM, Zhang XT, Guo P, He EQ, Zhou F, Zhao DK, Zhang YY. Effect of oxidative stress on development of silicosis. *World J Respirol* 2012; 2(1): 1-5 Available from: URL: <http://www.wjgnet.com/2218-6255/full/v2/i1/1.htm> DOI: <http://dx.doi.org/10.5320/wjr.v2.i1.1>

INTRODUCTION

Silicosis is the most destructive form of pneumoconiosis caused by the inhalation of SiO_2 . Exposure to silica par-

ticles smaller than 10 micrometers is considered a major cause of silicosis. According to the United States National Institute for Occupational Safety and Health, the most important factor in the development of silicosis is “the product of the concentrations of dust containing respirable silica in workplace air and the percentage of respirable silica in the total dust”^[1]. The disease is associated with inflammation of the respiratory system that eventually results in fibrosis, the hardening of the lungs, reducing the ability of the patients to breathe efficiently^[2]. The pathological manifestation includes macrophages alveolitis, silicotic nodule and dust fibration^[3]. The role of macrophages has been investigated^[4], but the pathogenesis remains unclear. The imbalance of oxidation and anti-oxidation due to long-term exposure to silica dusts may be one of the causative factors for silicosis^[5]. This study aims at clarifying the pathogenesis of silicosis by observing oxidative stress indicators in peripheral blood of silicosis patients in an attempt to search for new strategies for the prevention and treatment of silicosis.

MATERIALS AND METHODS

Subjects

This research was approved by the Ethics Committee of Wuxi Hospital for Prevention and Treatment of Occupational Disease, Wuxi, China. All subjects were male and they were divided into control group, dust-exposed group, and silicosis patient group. The control group was composed of hotel staff who had no silica dust exposure (age 48.7 ± 8.0 years, $n = 100$). The dust-exposed group consisted of workers from a local casting factory who had more than one year dust-exposure (age 50.78 ± 9.4 years, $n = 200$). The silicosis patient group was composed of in-patients and out-patients who have been admitted to our hospital since 2008 (age 52.0 ± 10.4 years, $n = 130$). There were 64 phase I, 46 phase II, and 20 phase III silicosis patients in this group. Besides, the cases of suspected 0⁺ silicosis were chosen as observation group (age 53.0 ± 9.12 years, $n = 32$). There was no significant difference in the age among the groups ($P > 0.05$). None of the subjects had working experience with radiation and toxic substance, and digestive disorders. No one had taken anti-oxidants, such as Vitamin C, Ginggo leaf or Teapol, one month before the test.

Methods

A questionnaire was designed and used to collect the general information of the subjects, including age, history of dust exposure and smoking, and past and family histories^[6]. Physical examination, electrocardiogram, high-kilovolt X-ray, pulmonary function tests including forced vital capacity (FVC), forced expiratory volume in one second (FEV1), FEV1/FVC and maximal voluntary ventilation, and abdominal ultrasonography were performed and serum fasting glucose, lipids, liver and kidney functions were examined.

Silicosis diagnosis

All the subjects were diagnosed and categorized based on the “Diagnostic Criteria of Pneumoconiosis”^[7].

Blood sample collection

Five milliliter peripheral blood was taken before breakfast, then centrifuged and preserved under -80°C .

Reagents and instruments

Superoxide dismutase (SOD), nitric oxide (NO) enzyme-linked immunosorbent assay kit (Biosource Corporation, United States). Serum glutathione peroxidase (GSH-Px), total antioxidant capacity (T-AOC), nitric oxide synthase (NOS), malondialdehyde (MDA) test kit (Nanjing Jiancheng Bioengineering Institute, Nanjing, China), automatic biochemical analyzer (Beckman Inc., United States), Spiro lab II portable spirometer (MIR Company, Italia), 550 microplate reader (BIORAD, United States), and New Century T6 ultraviolet (UV)-visible spectrophotometer (Beijing Puxi General, Beijing, China).

Test of oxidative stress indicators

SOD activity and NO levels were detected by enzyme-linked immunosorbent assay. GSH-Px activity, T-AOC and NOS were detected by chemical colorimetric assay. MDA was detected by dithiobisnitro-benzoate colorimetric assay and UV spectrophotometer was used for colorimetric assay. All the detections were performed manually according to the manufacturers' instructions.

Statistical analysis

All the data were expressed as mean \pm SD. SPSS11.5 was used for statistical analysis. Univariate analysis of variance and Pearson correlation analysis were used for the significance comparison between each group. $P < 0.05$ was considered statistically significant.

RESULTS

Comparison of general indicators between each group

There was no significant difference in the blood pressure, blood glucose, blood lipids, creatinine, urea nitrogen, and alanine aminotransferase between groups ($P > 0.05$) (Table 1).

Comparison of serum NO, SOD, MDA, T-AOC, NOS and GSH-Px between each group

NO in dust-exposed and silicosis groups was significantly higher than in the control group ($P < 0.01$), while there was no difference between dust-exposed group and silicosis patient group ($P > 0.05$). SOD in dust-exposed and silicosis group was lower than in the control group ($P < 0.01$), whereas there was no difference between silicosis and dust-exposed group ($P > 0.05$). T-AOC, NOS and MDA in silicosis group were obviously higher than in the control group and dust-exposed group ($P < 0.05$, $P < 0.01$), but there was no difference between the

Table 1 Comparison of general indicators between control group, dust-exposed group and silicosis patient group

Group	<i>n</i>	Age (yr)	Systolic pressure (mmHg)	Diastolic pressure (mmHg)	Blood glucose (mmol/L)	Triglyceride (mmol/L)	Cholesterol (mmol/L)	Creatinine (μmol/L)	Urea nitrogen (mmol/L)	Alanine transaminase (mmol/L)
Control	100	48.7 ± 8.0	130.42 ± 8.65	82.65 ± 7.68	4.93 ± 0.94	1.45 ± 0.94	5.15 ± 1.03	71.80 ± 16.50	5.53 ± 1.20	24.31 ± 17.44
Dust-exposed	200	50.8 ± 9.4	129.53 ± 9.45	84.72 ± 8.96	4.95 ± 0.79	1.52 ± 0.42	4.24 ± 0.89	69.97 ± 18.96	6.37 ± 1.16	29.16 ± 16.68
Silicosis	130	52.0 ± 10.4	131.42 ± 10.23	84.52 ± 8.14	5.06 ± 0.94	1.53 ± 0.90	4.75 ± 0.92	80.12 ± 19.10	6.47 ± 1.87	27.10 ± 16.83

Table 2 Comparison of nitric oxide, superoxide dismutase, malondialdehyde, total antioxidant capacity, nitric oxide synthase and serum glutathione peroxidase between control, dust-exposed group and silicosis patient group

Group	<i>n</i>	NO (μmol/L)	SOD (U/mL)	MDA (μmol/L)	T-AOC (U/mL)	NOS (μmol/L)	GSH-Px (U/mL)
Control	100	45.622 ± 24.081	75.239 ± 24.020	4.027 ± 0.822	11.639 ± 3.707	33.812 ± 6.398	223.360 ± 46.838
Dust-exposed	200	77.148 ± 30.188 ^b	70.515 ± 20.034 ^b	4.240 ± 1.167	11.814 ± 4.758	35.598 ± 5.664	231.164 ± 36.484 ^d
Silicosis	130	81.162 ± 35.176 ^b	68.209 ± 21.528 ^b	5.054 ± 1.204 ^{a,d}	13.048 ± 4.153 ^{a,c}	36.201 ± 7.782 ^{a,c}	270.469 ± 39.228 ^{b,d}

^a*P* < 0.05, ^b*P* < 0.01 *vs* control group; ^c*P* < 0.05, ^d*P* < 0.01 *vs* dust-exposed group. NO: Nitric oxide; SOD: Superoxide dismutase; MDA: Malondialdehyde; T-AOC: Total antioxidant capacity; NOS: Nitric oxide synthase; GSH-Px: Serum glutathione peroxidase.

Table 3 Comparison of nitric oxide, superoxide dismutase, malondialdehyde, total antioxidant capacity, nitric oxide synthase and serum glutathione peroxidase between observation group and silicosis at different stages

Group	<i>n</i>	NO (μmol/L)	SOD (U/mL)	MDA (μmol/L)	T-AOC (U/mL)	NOS (μmol/L)	GSH-Px (U/mL)
Observation	32	74.130 ± 38.049	70.414 ± 20.036	4.489 ± 1.154	12.903 ± 3.790	35.379 ± 5.504	256.906 ± 21.418
Silicosis							
Phase I	64	79.145 ± 36.149	69.215 ± 25.033	4.994 ± 1.237	12.905 ± 4.448	35.626 ± 7.234	259.594 ± 34.79
Phase II	46	80.173 ± 36.215	67.896 ± 26.018	5.038 ± 1.208	12.834 ± 3.481	36.078 ± 5.911	276.783 ± 40.969
Phase III	20	83.176 ± 34.131	67.201 ± 30.030	5.244 ± 1.127	13.993 ± 4.662	36.899 ± 4.162	290.750 ± 39.129 ^{a,c}

^a*P* < 0.05 *vs* observation group; ^c*P* < 0.05 *vs* phase I silicosis. NO: Nitric oxide; SOD: Superoxide dismutase; MDA: Malondialdehyde; T-AOC: Total antioxidant capacity; NOS: Nitric oxide synthase; GSH-Px: Serum glutathione peroxidase.

Table 4 Serum glutathione peroxidase Pearson correlation test

	Silicosis stage	Category	Length of dust exposure	Type of occupation
<i>r</i> value	0.507	0.456	0.365	0.341
<i>P</i> value	0.000	0.000	0.000	0.000

control group and dust-exposed group (*P* > 0.05). GSH-Px in dust-exposed group and silicosis group was higher (*P* < 0.05, *P* < 0.01) than in the control group, but it was even higher in silicosis group, and the difference was statistically significant (*P* < 0.01) (Table 2).

Comparison of NO, SOD, MDA, T-AOC, NOS and GSH-Px between observation group and silicosis group at different stages

Compared with observation group and phase I silicosis, GSH-Px in phase III silicosis increased, and the difference was statistically significant (*P* < 0.05). There was no significant difference in the NO, SOD, MDA, T-AOC, NOS and GSH-Px between observation group and different stages of silicosis (*P* > 0.05) (Table 3).

Pearson correlation test

Serum GSH-Px was positively correlated with the silicosis stage, category, length of dust exposure and type

of occupation (based on the exposure concentration of dust), and the differences were statistically significant (*P* < 0.01) (Table 4).

DISCUSSION

Pathogenesis of silicosis mainly involves oxidative stress, cytokines, cell apoptosis and immunity doctrine^[8-12]. Although the pathogenesis varies, the mechanism that the SiO₂ stimulates the body to secrete a variety of cytokines and other biological activity substances resulting in the fibrosis of lung tissue, is affirmative^[13-16]. The clinical diagnosis of silicosis rely mainly on the chest radiograph, but a definite diagnosis is often established when the lesion is irreversible, imposing heavy economic burdens on both the patient and society^[17,18]. If the quantitative and qualitative changes of these cytokines can be detected earlier and one or more cytokines as a relatively specific indicator can be found, it will be of great significance for the early diagnosis of silicosis and the dust-exposed population census. Many early stage biomarkers have been found in the sera of human and animals. Reactive oxygen species/reactive nitrogen species (ROS/RNS)^[5,19], cytokines^[20,21] and apoptosis-related factors^[22] are indicators related to the early biological indicators of silicosis. Clara cell protein CC16^[23,24], the lung surface ac-

tive substances D and heme monooxygenase-1 are early specific indicators for silicosis^[23,25,26]. With the progress made in the pathogenesis studies of silicosis, a variety of theories have become available. Free radical is one of the most important theories. Under physiological circumstances, the normal metabolic process continuously generates free radicals meanwhile they are eliminated by the antioxidant mechanisms in order to maintain homeostasis. When the body is subjected to different kinds of harmful stimuli, oxidative stress occurs and more bioactive molecules such as ROS and RNS are produced that exceeds the removal of oxide, resulting in the imbalance of oxidation and anti-oxidation system^[27,28]. This research shows that the imbalance of oxidation and anti-oxidation system existed in both the dust-exposed group and silicosis group, which was particularly remarkable in silicosis group. The increase of NO, NOS, T-AOC and MDA and the decrease of SOD are the prime manifestations. It suggests that silica dust can induce the occurrence of oxidative stress and increase lipid peroxidation, and then cause lung tissue damage. Since serum GSH-Px increases with stages of silicosis, it may be related to the severity of silicosis. Silicosis patients have a higher level of oxidant and anti-oxidant molecules. It may be caused by the continuous production of free radicals and lipid peroxidation with the development of silicosis^[29,30]. As the illness progresses, oxidative stress becomes more severe. Therefore, the abnormality of oxidative stress indicators not only suggests the existence of silicosis, but also the inevitable outcome of silicosis. These indicators can be used to predict the occurrence, development, severity and prognosis of silicosis in clinical practice. Due to the known effects of oxidative damage in the development of silicosis, administration of antioxidant agents is a potential strategy for the prevention and treatment of silicosis.

COMMENTS

Background

Silicosis is the most destructive form of pneumoconiosis caused by the inhalation of SiO₂. The pathological manifestation includes macrophages alveolitis, silicotic nodule and dust fibration. But the pathogenesis is still unclear.

Research frontiers

The imbalance of oxidation and anti-oxidation induced by long-term exposure to silica dusts may be one of the causes for silicosis. The authors investigated the pathogenesis of silicosis by observing oxidative stress indicators in the peripheral blood of silicosis patients in an effort to search for new therapies for the disease.

Innovations and breakthroughs

The authors concluded that the imbalance of oxidation and anti-oxidation system exists in both the dust-exposed group and silicosis group, particularly evident in silicosis group. As the illness progresses, the oxidative stress becomes more severe. So, the abnormality of oxidative stress indicators suggests not only the existence of silicosis, but also the inevitable outcome of silicosis.

Applications

The oxidative stress indicators can be used to predict the occurrence, development, severity and prognosis of silicosis in clinical practice. Due to the important effects of oxidative damage in the development of silicosis, administration of antioxidant agents will be a potential strategy for the prevention and treatment of silicosis.

Terminology

Silicosis is the most destructive form of pneumoconiosis caused by the inhalation of SiO₂. When the body is subjected to different kinds of harmful stimuli, oxidative stress occurs and more bioactive molecules such as reactive oxygen species and reactive nitrogen species will be produced that exceeds the removal of oxide, resulting in the imbalance of oxidation and anti-oxidation system.

Peer review

The paper is a short epidemiological study about oxidative stress occurrence in healthy and dust-exposed subjects and silicosis patients and addresses an important issue about the plasma level of oxidative stress-related molecules and its correlation with silicosis.

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ACKNOWLEDGMENTS

Acknowledgments to reviewers of World Journal of Respirology

We acknowledge our sincere thanks to our reviewers. Many reviewers have contributed their expertise and time to the peer review, a critical process to ensure the quality of our World Series Journals. Both the editors of the journals and authors of the manuscripts submitted to the journals are grateful to the following reviewers for reviewing the articles (either published or rejected) over the past period of time.

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Session Western Society of Allergy,
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Kauai, Hawaii

March 2-6, 2012
68th American Academy of Allergy
Asthma and Immunology Annual
Meeting
Orlando, FL, United States

March 14-17, 2012
Gulf Thoracic 2012
Dubai, United Arab Emirates

March 30-April 1, 2012
Lung Science Conference
Estoril, Portugal

April 18-21, 2012
3rd European Lung Cancer
Conference
Geneva, Switzerland

April 21-24, 2012
5th World Asthma and Chronic
Obstructive Pulmonary Disease
Forum
New York, NY, United States

April 25-28, 2012
6th International Primary Care
Respiratory Group World
Conference 2012
Edinburgh, United Kingdom

April 28-29, 2012
Houston Lung Symposium
Houston, TX, United States

May 18-23, 2012
American Thoracic Society ATS 2012
International Conference
San Francisco, CA, United States

June 15-18, 2012
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Bronchology and Interventional
Pulmonology joint with The
17th World Congress for
Bronchoesophagology
Ohio, OH, United States

June 16-20, 2012
European Academy of Allergy and
Clinical Immunology Congress 2012
Geneva, Switzerland

June 20-22, 2012
COPD8 International Conference
on Chronic Obstructive Pulmonary
Disease
Birmingham, United Kingdom

August 18-23, 2012
15th International Congress of
Immunology
Rome, Italy

August 18-21, 2012
21st World Congress of Asthma 2012
Quebec, Canada

September 1-5, 2012
22nd Annual Congress of the
European Respiratory Society
Vienna, Austria

October 11-13, 2012
26th Annual North American Cystic
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October 20-25, 2012
CHEST 2012
Atlanta, Georgia

November 8-14, 2012
Asthma and Immunology
Anaheim, CA, United States

November 13-17, 2012
The 43rd Union World Conference
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Kuala Lumpur, Malaysia

December 26-28, 2012
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Conference on Bronchology,
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Respiratory Diseases
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Name of journal

World Journal of Respirology

ISSN

ISSN 2218-6255 (online)

Editor-in-chief

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Acknowledgments

Brief acknowledgments of persons who have made genuine contributions to the manuscript and who endorse the data and conclu-

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Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; **169**: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

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- 16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1

Statistical data

Write as mean \pm SD or mean \pm SE.

Statistical expression

Express *t* test as *t* (in italics), *F* test as *F* (in italics), chi square test as χ^2 (in Greek), related coefficient as *r* (in italics), degree of freedom as *v* (in Greek), sample number as *n* (in italics), and probability as *P* (in italics).

Units

Use SI units. For example: body mass, *m* (B) = 78 kg; blood pres-

sure, p (B) = 16.2/12.3 kPa; incubation time, t (incubation) = 96 h, blood glucose concentration, c (glucose) 6.4 ± 2.1 mmol/L; blood CEA mass concentration, p (CEA) = 8.6 24.5 μ g/L; CO₂ volume fraction, 50 mL/L CO₂, not 5% CO₂; likewise for 40 g/L formaldehyde, not 10% formalin; and mass fraction, 8 ng/g, *etc.* Arabic numerals such as 23, 243, 641 should be read 23 243 641.

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Italics

Quantities: t time or temperature, c concentration, A area, l length, m mass, V volume.

Genotypes: *gyrA*, *arg 1*, *c myc*, *c fos*, *etc.*

Restriction enzymes: *EcoRI*, *HindI*, *BamHI*, *Kho I*, *Kpn I*, *etc.*

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