

World Journal of *Critical Care Medicine*

World J Crit Care Med 2014 May 4; 3(2): 45-67



Editorial Board

2011-2015

The *World Journal of Critical Care Medicine* Editorial Board consists of 246 members, representing a team of worldwide experts in critical care medicine. They are from 45 countries, including Argentina (2), Australia (8), Austria (2), Bangladesh (1), Belgium (3), Brazil (4), Canada (7), China (23), Croatia (1), Cuba (1), Denmark (1), Egypt (4), Finland (1), France (8), Germany (11), Greece (9), Hungary (1), India (10), Iran (2), Ireland (1), Israel (6), Italy (14), Japan (6), Jordan (1), Mexico (1), Morocco (1), Netherlands (4), New Zealand (3), Norway (1), Poland (1), Portugal (4), Russia (1), Saudi Arabia (3), Singapore (1), Slovenia (1), South Africa (1), Spain (7), Sweden (1), Switzerland (3), Thailand (1), Tunisia (1), Turkey (3), United Kingdom (8), United States (72), and Uruguay (1).

EDITOR-IN-CHIEF

Yaseen Mohamed Arabi, *Riyadh*

GUEST EDITORIAL BOARD MEMBERS

Hsing I Chen, *Hualien*
Sheng-Hsien Chen, *Tainan*
Yih-Sharnng Chen, *Taipei*
Yung-Chang Chen, *Taipei*
Der-Yang Cho, *Taichung*
Cheng-Keng Chuang, *Taoyuan*
How-Ran Guo, *Tainan*
Bang-Gee Hsu, *Hualien*
Chien-Wei Hsu, *Kaohsiung*
Wen-Jinn Liaw, *Taipei*
Yan-Ren Lin, *Changhua*
Jiunn-Jye Sheu, *Kaohsiung*

MEMBERS OF THE EDITORIAL BOARD



Argentina

Eduardo Chuluyan, *Buenos Aires*
Adrian Angel Inchauspe, *Berazategui*



Australia

Zsolt J Balogh, *Newcastle*
Zoltan Huba Endre, *Sydney*
Nam Q Nguyen, *Adelaide*
Alistair D Nichol, *Melbourne*
Srinivas Rajagopala, *Adelaide*
Georg Marcus Schmolzer, *Melbourne*
Andrew Trevitt Slack, *Southport*
Ravindranath Tiruvoipati, *Frankston*



Austria

Lars-Peter Kamolz, *Vienna*
Sylvia Knapp, *Vienna*



Bangladesh

Saidur Rahman Mashreky, *Dhaka*



Belgium

Teresinha Leal, *Brussels*
Manu Malbrain, *Antwerp*
Jean-Louis Vincent, *Brussels*



Brazil

Luciano CP Azevedo, *São Paulo*
Patricia Rieken Macedo Rocco, *Rio de Janeiro*
Marcos Antonio Rossi, *São Paulo*
Renato Seligman, *Porto Alegre*



Canada

Douglas D Fraser, *London*
Pierre A Guertin, *Quebec*
Marc Jeschke, *Toronto*
Constantine J Karvellas, *Edmonton*
Wolfgang Michael Kuebler, *Toronto*
Mingyao Liu, *Toronto*
Xi Yang, *Manitoba*



China

Xiang-Dong Chen, *Chengdu*

Xu-Lin Chen, *Hefei*
Wong Tak Chuen, *Hong Kong*
Ming-Xu Da, *Gansu*
Huang-Xian Ju, *Nanjing*
Ting-Bo Liang, *Hangzhou*
Peng-Lin Ma, *Beijing*
Chung-Wah David Siu, *Hong Kong*
Yong-Ming Yao, *Beijing*
Jia-Ping Zhang, *Chongqing*
Wei-Dong Zhou, *Beijing*



Croatia

Alan Sustic, *Rijeka*



Cuba

Jesús Pérez-Nellar, *La Habana*



Denmark

Dan Stieper Karbing, *Aalborg*



Egypt

Ibrahim Abouomira, *Cairo*
Hanan Ibrahim, *Cairo*
Amr M Moghazy, *Alexandria*
Ayman A Yousef, *Tanta*



Finland

Asko Armas Riutta, *Tampere*

**France**

Jean-Marc Cavaillon, *Paris*
 Jean-Michel Constantin, *Clermont-Ferrand*
 Marc Leone, *Marseille*
 Bruno Mégarbane, *Paris*
 Saad Nseir, *Lille*
 Nicolas Terzi, *Caen*
 Jean-François Timsit, *La Tronche Cedex*
 Benoit Vallet, *Lille*

**Germany**

Hendrik Bracht, *Ulm*
 Michael Czaplík, *Aachen*
 Gerrit Grieb, *Aachen*
 Tobias Keck, *Freiburg*
 Philipp Kobbe, *Aachen*
 Alexander Koch, *Aachen*
 Marc Maegele, *Cologne*
 Norbert Pallua, *Aachen*
 Andrzej Antoni Piatkowski, *Aachen*
 Armin Rudolf Sablotzki, *Leipzig*
 Kai D Zacharowski, *Frankfurt am Main*

**Greece**

Ioanna Dimopoulou, *Athens*
 Dimitrios Karakitsos, *Athens*
 Petros Kopterides, *Athens*
 Gregory Kouraklis, *Athens*
 Athanasios D Marinis, *Athens*
 George Nakos, *Ioannina*
 Papaioannou E Vasilios, *Alexandroupolis*
 Theodoros Xanthos, *Athens*
 Spyros G Zakynthinos, *Athens*

**Hungary**

Zoltan Rakonczay, *Szeged*

**India**

Rachna Agarwal, *Delhi*
 Ritesh Agarwal, *Chandigarh*
 Mohammad Farooq Butt, *Srinagar*
 Mohan Gurjar, *Lucknow*
 Deven Juneja, *New Delhi*
 Farhad N Kapadia, *Mumbai*
 Vikram Kate, *Pondicherry*
 Pramod Kumar, *Manipal*
 Ritesh G Menezes, *Mangalore*
 Medha Mohta, *Delhi*

**Iran**

Hemmat Maghsoudi, *Tabriz*
 Homayoun Sadeghi-Bazargani, *Tabriz*

**Ireland**

Sanjay H Chotirmall, *Dublin*

**Israel**

Alexander Becker, *Kefar Tavor*
 Yoram Kluger, *Haiifa*
 Yona Kosashvili, *Zerrifin*
 Kobi Peleg, *Tel Aviv*
 Ilan Sela, *Rehovot*
 Pierre Singer, *Tel Aviv*

**Italy**

Giacomo Bellani, *Monza*
 Giovanni Camussi, *Torino*
 Anselmo Caricato, *Rome*
 Piero Ceriana, *Pavia*
 Antonio Chiaretti, *Rome*
 Davide Chiumello, *Milano*
 Alfredo Conti, *Messina*
 Paolo Cotogni, *Torino*
 Daniele M De Luca, *Rome*
 Vincenzo De Santis, *Rome*
 Luca La Colla, *Parna*
 Giovanni Landoni, *Milano*
 Raffaele Scala, *Lucca*
 Giovanni Vento, *Rome*

**Japan**

Keishiro Aoyagi, *Kurume*
 Satoshi Hagiwara, *Yufu*
 Yuichi Hattori, *Toiyama*
 Hideo Inaba, *Kanazawa*
 Eisuke Kagawa, *Hiroshima*
 Chieko Mitaka, *Tokyo*

**Jordan**

Feras Ibrahim Hawari, *Amman*

**Mexico**

Silvio A Ñamendys-Silva, *Mexico City*

**Morocco**

Redouane Abouqal, *Rabat*

**Netherlands**

WA Burman, *Maastricht*
 Martin CJ Kneyber, *Groningen*
 Patrick Schober, *Amsterdam*
 Arie Barend Van Vugt, *Enschede*

**New Zealand**

Sultan Zayed Al-Shaqsi, *Dunedin*
 Arman Adam Kahokehr, *Whangarei*
 John William Pickering, *Christchurch*

**Norway**

Ulf R Dahle, *Oslo*

**Poland**

Maciej Owecki, *Poznań*

**Portugal**

Ernestina Rodrigues Gomes, *Porto*
 Cristina Granja, *Porto*
 José António Lopes, *Lisbon*
 Pedro M Póvoa, *Lisbon*

**Russia**

Konstantin A Popugaev, *Moscow*

**Saudi Arabia**

Imran Khalid, *Jeddah*
 Mohamed Taifour Suliman, *Tabuk*

**Singapore**

Devanand Anantham, *Singapore*

**Slovenia**

Štefek Grmec, *Maribor*

**South Africa**

DL Clarke, *Pietermaritzburg*

**Spain**

Juan Carlos Montejo González, *Madrid*
 David Jimenez, *Madrid*
 Juan Antonio Llompart-Pou, *Palma*
 Antonio Torres Mart, *Barcelona*
 Enrique Ariel Piacentini, *Barcelona*
 Alonso Mateos Rodriguez, *Madrid*
 R Rodríguez-Roisin, *Barcelona*

**Sweden**

Mihai Oltean, *Gothenburg*

**Switzerland**

Dieter Cadosch, *Zurich*
 Mihael Potocki, *Basel*
 John Friedrich Stover, *Zurich*

**Thailand**

Viroj Wiwanitkit, *Bangkok*

**Tunisia**

Mabrouk Bahloul, *Sfax*

**Turkey**

Yusuf Kenan Coban, *Malatya*
Bensu Karahalil, *Ankara*
Ali Nayci, *Mersin*

**United Kingdom**

Sammy Al-Benna, *Nottingham*
Giles N Cattermole, *London*
Frantisek Duska, *Nottingham*
James Nicholas Fullerton, *London*
Christina Jones, *Prescot*
Sameer Khan, *Middlesbrough*
George Ntoumenopoulos, *London*
Cecilia O'Kane, *Belfast*

**United States**

Edward Abraham, *Winston-Salem*
Bernard R Bendok, *Chicago*
Michael Blaivas, *Atlanta*

Charles D Boucek, *Pittsburgh*
Marcia Leigh Brackbill, *Winchester*
Ronald A Bronicki, *Houston*
Robert C Cantu, *Concord*
Marylou Cardenas-Turanzas, *Houston*
Archana Chatterjee, *Omaha*
Paul A Checchia, *St. Louis*
Rubin Issam Cohen, *New Hyde Park*
Stephen Cohn, *San Antonio*
Donald Edward Craven, *Burlington*
Ruy J Cruz Jr, *Pittsburgh*
Francis C Dane, *Roanoke*
Marc de Moya, *Boston*
Steven M Donn, *Ann Arbor*
Christopher P Farrell, *Wynnwood*
Marco Fernández, *Nashville*
Kevin Foster, *Phoenix*
Barry D Fuchs, *Philadelphia*
Richard P Gonzalez, *Mobile*
Kenneth W Gow, *Seattle*
Alan H Hall, *Laramie*
Jijo John, *Oklahoma City*
Lewis J Kaplan, *New Haven*
Jason N Katz, *Chapel Hill*
Salah Georges Keyrouz, *Little Rock*
Deborah A Kuhls, *Las Vegas*
Gregory Luke Larkin, *New Haven*
Christos Lazaridis, *Charleston*
James Anthony Lin, *Los Angeles*
Yahia M Lodi, *Syracuse*
Roger M Loria, *Richmond*
Aigang Lu, *Cincinnati*
Rudolf Lucas, *Augusta*
O John Ma, *Portland*
Robert T Mallet, *Fort Worth*
William T McGee, *Springfield*
Mark G McKenney, *Miami*

Michael Moussouttas, *Philadelphia*
Oliver Hans-Josef Muensterer, *Birmingham*
Rahul Nanchal, *Milwaukee*
Michael Steven Niederman, *Mineola*
Gary Frank Nieman, *Syracuse*
James Martin O'Brien, *Columbus*
Martin Oudega, *Pittsburgh*
Catherine Mobley Preissig, *Duluth*
Virginia Prendergast, *Phoenix*
Ramesh Raghupathi, *Philadelphia*
Miren Ava Schinco, *Jacksonville*
Carl Ivan Schulman, *Miami*
L Keith Scott, *Shreveport*
Kevin Navin Sheth, *Baltimore*
Jenni Short, *Salina*
Ronald Fong Sing, *Charlotte*
Philip Charles Spinella, *St. Louis*
Robert M Starke, *Charlottesville*
Stanislaw Peter A Stawicki, *Columbus*
David Christopher Stockwell, *Washington*
Stanislav Svetlov, *Gainesville*
Maged A Tanios, *Long Beach*
Neal James Thomas, *Hershey*
Nancy Moon Tofil, *Birmingham*
Balagangadhar R Totapally, *Miami*
Steven Nicholas Vaslef, *Durham*
Joseph Clark Watson, *Falls Church*
John Stephen Wilgis, *Orlando*
David Conrad Willms, *San Diego*
Haodong Xu, *Rochester*
Xiao-Ming Xu, *Indianapolis*
Midori Anne Yenari, *San Francisco*

**Uruguay**

William Manzanares, *Montevideo*

Contents

Quarterly Volume 3 Number 2 May 4, 2014

REVIEW

- 45 Metabolic theory of septic shock
Pravda J

BRIEF ARTICLE

- 55 Arterial vs venous blood gas differences during hemorrhagic shock
Williams KB, Christmas AB, Heniford BT, Sing RF, Messick J
- 61 Variable change in renal function by hypertonic saline
Corry JJ, Varelas P, Abdelhak T, Morris S, Hawley M, Hawkins A, Jankowski M

APPENDIX I-V Instructions to authors

ABOUT COVER Editorial Board Member of *World Journal of Critical Care Medicine*, Chieko Mitaka, MD, PhD, Associate Professor, Department of Critical Care Medicine, Tokyo Medical and Dental University Graduate School, 1-5-45, Yushima, Bunkyo-ku, Tokyo, 113-8519, Japan

AIM AND SCOPE *World Journal of Critical Care Medicine (World J Crit Care Med, WJCCM, online ISSN 2220-3141, DOI: 10.5492)* is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJCCM covers topics concerning severe infection, shock and multiple organ dysfunction syndrome, infection and anti-infection treatment, acute respiratory distress syndrome and mechanical ventilation, acute kidney failure, continuous renal replacement therapy, rational nutrition and immunomodulation in critically ill patients, sedation and analgesia, cardiopulmonary cerebral resuscitation, fluid resuscitation and tissue perfusion, coagulant dysfunction, hemodynamic monitoring and circulatory support, ICU management and treatment control, and application of bronchofiberscopy in critically ill patients.

We encourage authors to submit their manuscripts to *WJCCM*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great clinical significance.

INDEXING/ABSTRACTING *World Journal of Critical Care Medicine* is now indexed in PubMed Central, PubMed, Digital Object Identifier.

FLYLEAF I-III Editorial Board

EDITORS FOR THIS ISSUE Responsible Assistant Editor: *Xiang Li* Responsible Science Editor: *Fang-Fang Ji*
 Responsible Electronic Editor: *Huan-Liang Wu*
 Proofing Editor-in-Chief: *Lian-Sheng Ma*

NAME OF JOURNAL
World Journal of Critical Care Medicine

ISSN
 ISSN 2220-3141 (online)

LAUNCH DATE
 February 4, 2012

FREQUENCY
 Quarterly

EDITOR-IN-CHIEF
Yaseen Mohamed Arabi, MD, FCCP, FCCM, Associate Professor, Chairman, Intensive Care Department, King Saud Bin Abdulaziz University, Medical Director, Respiratory Services, King Abdulaziz Medical City, National Guard Hospital, Riyadh, PO Box 22490, Riyadh 11426, Saudi Arabia

Derek S Wheeler, MD, FAAP, FCCP, FCCM, Associate Professor, Associate Patient Safety Officer, Medical Director, Pediatric Intensive Care Unit, Division of Critical Care Medicine, James M. Anderson Center for Health Systems Excellence, The Center

for Simulation and Research, Co-Director, The Center for Acute Care Nephrology, Division of Critical Care Medicine, Cincinnati Children's Hospital Medical Center, 3333 Burnet Avenue, Cincinnati, OH 45229-3039, United States

EDITORIAL OFFICE
 Jin-Lei Wang, Director
 Xiu-Xia Song, Vice Director
World Journal of Critical Care Medicine
 Room 903, Building D, Ocean International Center, No. 62 Dongshihuan Zhonglu, Chaoyang District, Beijing 100025, China
 Telephone: +86-10-85381891
 Fax: +86-10-85381893
 E-mail: bpgoffice@wjgnet.com
 http://www.wjgnet.com

PUBLISHER
 Baishideng Publishing Group Co., Limited
 Flat C, 23/F, Lucky Plaza,
 315-321 Lockhart Road, Wan Chai,
 Hong Kong, China
 Fax: +852-6555-7188
 Telephone: +852-3177-9906

E-mail: bpgoffice@wjgnet.com
 http://www.wjgnet.com

PUBLICATION DATE
 May 4, 2014

COPYRIGHT
 © 2014 Baishideng. Articles published by this Open Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
 All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

INSTRUCTIONS TO AUTHORS
 Full instructions are available online at http://www.wjgnet.com/2220-3141/g_info_20100722180909.htm

ONLINE SUBMISSION
<http://www.wjgnet.com/esp/>

Metabolic theory of septic shock

Jay Pravda

Jay Pravda, Inflammatory Disease Research Centre, West Palm Beach, FL 33420, United States

Author contributions: Pravda J is the sole author of this manuscript and solely responsible for its content; Pravda J performed all the research, collected, analyzed and interpreted all the data; Pravda J conceived of and developed the Metabolic Theory of Septic Shock; Pravda J prepared and wrote the manuscript and performed all critical revisions; Pravda J certifies that the Metabolic Theory of Septic Shock is the product of his original research and Pravda J has overall responsibility for this manuscript.

Correspondence to: Jay Pravda, MD, MPH, Inflammatory Disease Research Centre, West Palm Beach, P.O. Box 32632, FL 33420, United States. jaypravda@yahoo.com

Telephone: +1-682-2513030 Fax: +1-888-7005813

Received: October 29, 2013 Revised: January 21, 2014

Accepted: March 3, 2014

Published online: May 4, 2014

Abstract

Septic shock is a life threatening condition that can develop subsequent to infection. Mortality can reach as high as 80% with over 150000 deaths yearly in the United States alone. Septic shock causes progressive failure of vital homeostatic mechanisms culminating in immunosuppression, coagulopathy and microvascular dysfunction which can lead to refractory hypotension, organ failure and death. The hypermetabolic response that accompanies a systemic inflammatory reaction places high demands upon stored nutritional resources. A crucial element that can become depleted early during the progression to septic shock is glutathione. Glutathione is chiefly responsible for supplying reducing equivalents to neutralize hydrogen peroxide, a toxic oxidizing agent that is produced during normal metabolism. Without glutathione, hydrogen peroxide can rise to toxic levels in tissues and blood where it can cause severe oxidative injury to organs and to the microvasculature. Continued exposure can result in microvascular dysfunction, capillary leakage and septic shock. It is the aim of this paper to present evidence that elevated systemic levels of hydrogen peroxide are present in

septic shock victims and that it significantly contributes to the development and progression of this frequently lethal condition.

© 2014 Baishideng Publishing Group Co., Limited. All rights reserved.

Key words: Septic shock; Hydrogen peroxide; Hypermetabolic; Sepsis; Systemic inflammatory response syndrome

Core tip: For decades septic shock has been attributed to an over-active immune response. However, immune modulation has failed to reduce mortality, casting doubt on a direct causal role for the immune response in the development of septic shock. A closer look suggests that septic shock is the result of a generalized build-up of hydrogen peroxide, a toxic cellular by-product generated as a consequence of the hypermetabolic state that accompanies a systemic immune response. This finding points to the systemic accumulation of hydrogen peroxide as a significant risk factor for the development of septic and non-septic shock syndromes.

Pravda J. Metabolic theory of septic shock. *World J Crit Care Med* 2014; 3(2): 45-54 Available from: URL: <http://www.wjgnet.com/2220-3141/full/v3/i2/45.htm> DOI: <http://dx.doi.org/10.5492/wjccm.v3.i2.45>

INTRODUCTION

Sepsis is a life threatening condition that is associated with a systemic inflammatory response to a microbial infection^[1]. Sepsis is the most common cause of mortality in the intensive care unit with a fatality rate that can rise to 80% for those developing multiple organ failure. The progression of an exaggerated systemic inflammatory response is thought to be responsible for the eventual development of septic shock and death^[2]. However, multiple therapeutic efforts aimed at controlling the immune

response with the intent of interrupting the process leading to organ failure have been uniformly unsuccessful^[1]. This simple fact has prompted a reappraisal of the role played by the immune system in the development of this condition that kills more than 150000 Americans yearly; more than breast, colon, prostate and brain cancer combined^[3,4].

Although immune activation is clearly evident, recent evidence suggests that the immune response may not be the direct mediator of the pathologic process that leads to septic shock. Studies conducted to define the circulating leukocyte transcriptome have revealed that there is no qualitative difference in the immunogenetic response when comparing burn or blunt trauma patients with complicated or uncomplicated outcomes. In other words, severely injured patients who die from their injuries have the same immunogenetic response as patients who recover; the only difference being the duration and intensity of systemic inflammation^[5].

The lack of a unique immunogenetic response suggests that septic shock is the phenotypic expression of a separate process that is initiated simultaneously with systemic immune activation. The multiple organ involvement, which can lead to death within a few days, suggests that this concomitant process is systemic in nature and initiated in parallel with inflammatory response. Moreover, the microvascular edema associated with multiple organ failure, which persists despite efforts at immunosuppression, suggests that a non-immune mediated angiopathic agent is being released into the systemic circulation^[1].

The high (8%) increased mortality rate for each hour of delay before instituting antibiotics after the onset of hypotension suggests that the duration of this parallel process is closely linked with a greater risk of an adverse outcome and down regulating the immune response with successful therapy simply allows this parallel process to turn off^[6].

In other words, survival is closely correlated with the early down regulation of a systemic process closely linked to systemic immune activation suggesting depletion of a crucial biochemical element that is critical for survival. Put differently, if catabasis (immune down regulation) is achieved by successful antibiotic therapy prior to depletion of this critical element the patient will survive, if not the patient is at high risk for organ failure, septic shock and death.

HYPERMETABOLIC RESPONSE

A key systemic process that is turned on and up-regulated with systemic inflammation is cellular metabolism, which becomes hypermetabolic from the onset of sepsis^[7]. The sustained high fever, highly amplified protein synthesis, tachycardia and tachypnea characteristic of a septic immune response requires supra-physiological energy supplies. It is estimated that basal energy requirements for a septic patient can reach up to 10000 calories daily^[8]. This hypermetabolic state not only requires increased nutrient

intake but also generates a large amount of toxic cellular by-products as a result of increased electron transport chain (ETC) activity required to synthesize sufficient adenosine triphosphate (ATP) to support a prolonged hypermetabolic state. This critical need for supplemental nutrients often cannot be met as it occurs at a time when caloric intake is curtailed as a result of the severe illness afflicting the patient^[8]. This suggests the progressive depletion of an element whose principal function is to metabolize a toxic cellular waste product that, upon accumulation, leads to organ dysfunction, microangiopathic edema and refractory hypotension, the characteristic pathologic findings in septic shock.

An important toxic product that is continuously generated as a result of cellular metabolism is hydrogen peroxide (H₂O₂), which is formed as a result of several metabolic activities including protein synthesis (disulfide bond formation), DNA recycling (Xanthine oxidase), ATP synthesis (ETC activity) and fatty acid oxidation (peroxisomal metabolism)^[9-12]. Most H₂O₂ is degraded to water via the enzymatic action of glutathione peroxidase (GPx), a selenium containing enzyme that has an obligate requirement for the co-factor glutathione (GSH) in order to metabolize H₂O₂. The biochemical reaction is: $2 \text{GSH} + \text{H}_2\text{O}_2 \rightarrow \text{GS-SG} + 2\text{H}_2\text{O}$ in which two molecules of GSH are converted to one molecule of glutathione disulfide (GS-SG) and two molecules of water. Glutathione is consumed during this process and must be replenished in order for the cell to prevent accumulation of H₂O₂ to toxic levels^[13,14].

Replenishment of glutathione, however, is not favored during periods of sustained hypermetabolism and caloric insufficiency, which frequently accompany critical illnesses such as sepsis leading to depletion of glutathione reserves.

Within 48 h of diagnosis critically ill children with sepsis were found to have a 60% decrease in whole blood GSH synthesis, suggesting depletion of whole body GSH stores^[15,16]. Systemic GSH depletion is supported by studies showing over 50% decrease in lung and skeletal muscle GSH in septic and critically ill patients^[17,18]. The critical importance of glutathione was demonstrated by a study which documented significantly decreased erythrocyte glutathione in septic non-survivors vs survivors ($P < 0.0001$)^[19]. This suggests high levels of circulating H₂O₂ capable of permeating erythrocyte cell membranes and oxidizing (and depleting) intracellular glutathione in septic shock non-survivors. Elegant studies have also demonstrated a significantly higher mitochondrial respiratory rate in non-survivors at three months following sepsis suggesting that failure to down regulate the hypermetabolic state (and excess H₂O₂ production) is independently associated with higher mortality even after surviving the initial infectious insult^[20].

Generalized depletion of body stores can result in cellular deficiency of GSH leading to a toxic accumulation of H₂O₂. A highly toxic oxidizing agent, H₂O₂ is the principal mediator of cellular oxidative damage. It does so by generating hydroxyl radical (OH*), the most

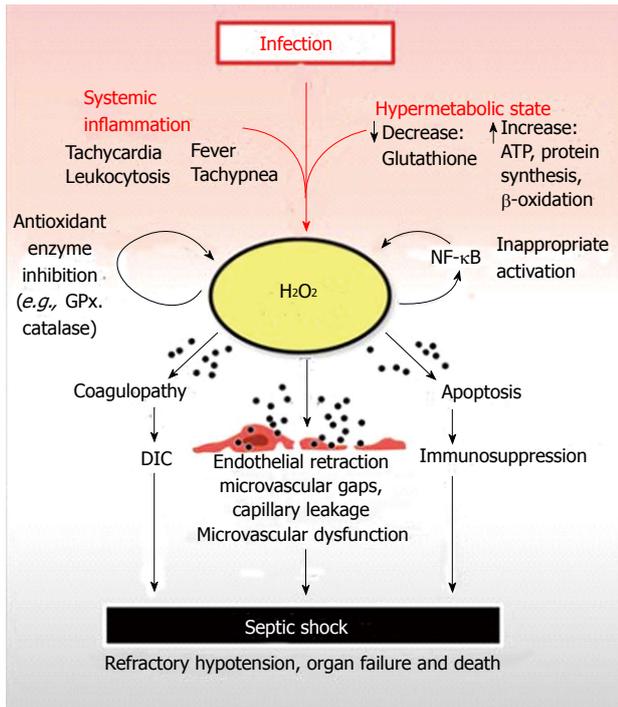


Figure 1 Septic shock begins with a systemic inflammatory reaction to an infection. A contemporaneous increase in metabolism is initiated, which can deplete reserves of critical nutrients such as glutathione. Glutathione is crucial for the neutralization of H_2O_2 , a toxic, membrane-permeable oxidizing agent generated as a by-product of cellular metabolism. Depletion of cellular glutathione results in elevation of H_2O_2 which can diffuse out of organ parenchymal cells and into capillary endothelium before reaching the bloodstream. Once in the systemic circulation, excess H_2O_2 is distributed throughout the body resulting in systemic oxidative damage to plasma components, organs and blood vessels. The net result is H_2O_2 induced coagulopathy, immunocyte apoptosis and microvascular dysfunction leading to disseminated intravascular coagulation, immunosuppression, organ failure and septic shock respectively. H_2O_2 inhibits GPx and catalase, which are critical anti-oxidant enzymes required for H_2O_2 neutralization. This prevents restoration of normal plasma and tissue redox balance while exacerbating oxidative tissue damage. H_2O_2 can also activate nuclear factor- κ B (NF- κ B) contributing to the inappropriate activation of this master pro-inflammatory transcription factor observed in septic shock. The pathologic activation of NF- κ B contributes to elevated tumor necrosis factor- α levels, another potent generator of intracellular H_2O_2 . GPx: Glutathione peroxidase.

potent reactive oxygen radical known in biological systems. Hydroxyl radical will indiscriminately disintegrate proteins, peroxidize lipids and oxidatively damage DNA leading to cell death^[21,22].

Compounding the cellular cytotoxicity of H_2O_2 is its ability to freely diffuse through biological membranes allowing it to permeate other cellular compartments and diffuse to the extracellular space from where it can pass through the capillary endothelium into the blood stream^[16,23]. Thus, the end result of a systemic GSH deficiency is the systematic discharge of excess H_2O_2 by all organs of the body into the bloodstream where it can damage distant capillary beds leading to systemic microcirculatory dysfunction, microangiopathic edema and refractory hypotension, a hallmark of septic shock.

This is supported by studies showing decreased human endothelial cell levels of GSH and eventual death after *in vitro* exposure to plasma from septic shock

patients^[24]. This implies a membrane diffusible agent capable of oxidizing intracellular GSH suggesting that a toxic level of plasma H_2O_2 was the offending oxidizing agent mediating this effect. This is consistent with the well documented oxidative damage and dose dependent cytotoxicity that occurs during human endothelial cell exposure to H_2O_2 ^[25,26].

In other studies high levels of urinary H_2O_2 were found to correlate with a fatal outcome in patients with sepsis and adult respiratory distress syndrome suggesting an important role for H_2O_2 in the pathogenesis of septic shock^[27]. Taken together the evidence suggests that H_2O_2 exerts a significant microangiopathic effect contributing to the development of microcirculatory dysfunction and the progression to refractory hypotension and fatal septic shock.

MECHANISM OF DISEASE

The above evidence supports a pathogenesis of septic shock which is initiated by the systemic depletion of glutathione as the crucial event responsible for the accumulation of H_2O_2 in tissues. Subsequent diffusion of H_2O_2 into the blood stream leads to systemic elevation of this highly toxic oxidizing agent resulting in the microvascular dysfunction and organ failure observed in septic shock (Figure 1).

At the onset, a systemic inflammatory response is accompanied by a generalized hypermetabolic state which provides the energy needed to sustain the highly up-regulated immune response switched on by the presence of a pathogen. The abrupt global increase of cellular bioenergetic reactions to several times their normal basal state presents the cell with a surge of toxic metabolic by-products that must be neutralized to avoid accumulation and cell death. Hydrogen peroxide, a toxic reactive oxygen species, is a significant metabolic by-product that is generated in increased amounts when cellular processes such as protein synthesis, DNA recycling and ATP production are upregulated during periods of hypermetabolism that accompany systemic inflammation.

The majority of cellular H_2O_2 is neutralized by GPx, a selenium containing enzyme, which utilizes the tripeptide co-factor glutathione as a donor of reducing equivalents during the enzymatic conversion of H_2O_2 to water. GSH is consumed in this reaction and must be replenished in order to prevent accumulation of H_2O_2 within the cell. However, during periods of high H_2O_2 production the availability of glutathione may be insufficient to keep up with demand leading to net H_2O_2 accumulation and glutathione depletion resulting in severe cellular dysfunction and organ failure.

Excess H_2O_2 can easily diffuse out of pericapillary parenchymal cells through capillary endothelium and into the blood stream. This augments endothelial generated H_2O_2 resulting in oxidative damage and microangiopathic dysfunction. The inability to buffer cellular H_2O_2 signals a systemic failure of reductive (anti-oxidant)

capacity as the excess oxidant load is discharged into the blood stream. Over time plasma reductive capacity is exhausted leading to severe disruption in plasma redox potential, which studies have shown is strongly associated with an unfavorable outcome^[28].

HYDROGEN PEROXIDE CAN REPRODUCE CLINICAL ABNORMALITIES OBSERVED IN SEPTIC SHOCK

Microcirculatory dysfunction

The capillary bed is not simply a conduit for the passage of cells. It is a highly dynamic and integrated system of endothelial cells that continuously interacts with its surrounding environment through a variety of displayed receptors and elaborated mediators whose functions includes vasoregulation, coagulation factors, barrier maintenance, immune cell recruitment and oxygen transport^[29]. Microcirculatory dysfunction is now considered to play a central role in the pathogenesis of sepsis and microvascular leakage has a defining role in its outcome^[1,29].

Histological analysis of microvasculature in a baboon model of lethal *Escherichia coli* (*E. coli*) sepsis revealed large gaps between endothelial cells accompanied by a significant increase in endothelial permeability^[30,31]. These changes are also observed upon exposure of human umbilical vein endothelial cells (HUVEC) to H₂O₂. Studies have demonstrated an 18x increase (from 20 to 360 gaps/mm²) in inter-endothelial cell gaps within 30 min of HUVEC exposure to H₂O₂. A time and dose dependent H₂O₂ induced endothelial contraction to about 60% of normal planar surface area was also observed^[32,33]. This provides a microanatomical basis by which excess H₂O₂ can account for the life threatening massive edema observed both in humans and experimental models of sepsis^[29,30].

Accompanying endothelial cell retraction during H₂O₂ exposure is the loss of tight junction proteins at the sites of gap formation, which strongly correlated with increased paracellular permeability^[34-36]. Extensive cytoskeletal disruption and rearrangement was also shown to occur after endothelial cell exposure to H₂O₂^[37-39]. Endothelial shape changes have been observed to occur in experimental models of sepsis and several studies have reported these pathological changes upon endothelial cell exposure to H₂O₂^[30,40-43].

The net effect of continuous H₂O₂ exposure on the systemic microvasculature is severe disruption. Barrier function is compromised, intercellular communication is blunted and signal transduction is abrogated. This leads to microvascular edema, arteriovenous shunts and vasodysregulation as a result of cumulative oxidative damage sustained from continued penetration of H₂O₂ into endothelial cells. This is supported by studies of low dose H₂O₂ perfusion into isolated rat lung, which increased pulmonary vascular bed permeability and capillary filtration coefficient^[41].

Studies of bovine brain microvascular endothelial

cells exposed to H₂O₂ revealed increased paracellular permeability of the blood brain barrier (BBB) with loss of tight junctional proteins (44)^[44]. H₂O₂ can by-pass the normally protective BBB by simply diffusing into tissues and cells^[15]. This can result in dysfunction of cerebral microvasculature and could account for the early mental changes observed in patients with sepsis as a result of impaired synaptic transmission^[8].

Immune activation

Numerous genes are activated during a systemic immune response in a critically ill or septic individual. Studies in healthy human volunteers receiving low dose endotoxin identified over 4500 activated genes, most of which were involved in the innate or adaptive immune response (5)^[5]. The simultaneous activation of this many genes is facilitated by preformed cytoplasmic signal transcription factors that serve as rapid response mediators to injury and infection. Nuclear factor kappa B (NF-κB) is a transcription factor that plays a central role in the activation and regulation of multiple genes that control immune and inflammatory reactions^[45]. NF-κB is significantly elevated in adults and children with sepsis^[46-48]. NF-κB is also a highly redox sensitive transcription factor capable of being activated by low levels of H₂O₂^[49,50] and has been proposed as a biomarker for oxidative stress^[51]. This suggests that high levels of ambient H₂O₂ may be involved in the inappropriate activation of NF-κB observed in septic shock^[45].

A central role for the innate immune system is suggested by the neutrophilic infiltration into multiple organs observed in septic shock^[45]. H₂O₂ is a highly potent neutrophilic chemo-attractant that can establish a chemotactic gradient as it diffuses out of parenchymal cells into the adjacent microvasculature. Circulating neutrophils can track this H₂O₂ gradient and enter the organ parenchyma *via* diapedesis. The net result is neutrophil infiltration into the parenchyma of multiple organs^[52-54].

Coagulopathy

Intravascular activation of the coagulation cascade with generation of fibrin and formation of diffuse microvascular thrombi is a pathologic and physiologic hallmark of sepsis^[55]. This presents clinically as disseminated intravascular coagulation (DIC) and is found in up to 50% of patients with sepsis^[56]. DIC leads to abnormal bleeding and intravascular clotting, obstructing limb and organ blood flow, and is a strong predictor of mortality^[56].

Endothelial derived tissue factor (TF) is the major physiological route by which fibrin generation is initiated in sepsis. Importantly, this process is triggered only at sites of vascular injury or endothelial disruption where plasma clotting factors can encounter the TF protein that activates this extrinsic clotting pathway^[56,57]. Studies utilizing immunohistochemistry in a lethal *E. coli* baboon sepsis model preferentially localized TF and TF mRNA at arterial branch areas, which is compatible with enhanced contact by a plasma derived oxidizing agent (*e.g.*, H₂O₂) at these sites of altered blood flow^[30].

H₂O₂ can induce vascular injury by peroxidation of cell membrane lipids and studies have shown a marked increase in endothelial cell TF and TF mRNA after 1 and 5 min exposure to Xanthine oxidase, a H₂O₂ generating enzyme^[10,58]. This indicates that TF is highly sensitive to H₂O₂ induced upregulation, which suggests with a contributory role for H₂O₂ in sepsis-associated DIC. Consistent with this mechanism is a case report describing a fatal case of sepsis with DIC and multiorgan failure in a previously healthy 37-year-old man after receiving several intravenous infusions of H₂O₂^[59].

Immunosuppression

Septic patients experience a considerable decline in lymphocyte numbers through apoptosis in the latter stages of sepsis and this is a significant contributing factor to the immunosuppression experienced by septic individuals^[60]. Studies have shown that H₂O₂ is a potent apoptosis inducing agent^[61]. B lymphocytes treated with agents that inhibit GSH synthesis experience a 95% decline in GSH concentration in 12 h. This is followed by a rise in intracellular H₂O₂ after which apoptosis occurs. By 72 h nearly 50% of B cells have died *via* apoptosis^[62]. T cells are also highly sensitive to the effects of GSH depletion. Studies have recorded a 30% decline in circulating T lymphocytes within 4 wk after glutathione levels declined to suboptimal levels in healthy volunteers^[63]. This supports a role for H₂O₂ in the development of sepsis induced immunosuppression.

Erythrocyte rigidity

Red blood cell deformability is markedly reduced in sepsis and studies have demonstrated a significant reduction in red blood cell deformability upon exposure to H₂O₂^[64]. A direct relationship was found between oxidant induced changes in erythrocyte deformability and severity of multi-organ failure in septic individuals^[65]. This suggests that plasma derived H₂O₂ is a source of oxidant-induced RBC membrane damage.

Circulating endothelial cells

Circulating endothelial cells (CEC) are a reliable, sensitive and specific indicator of vascular damage^[66]. These cells rarely exist in the peripheral blood of healthy individuals^[67]. Patients with severe sepsis and septic shock have significantly higher numbers of CECs indicating widespread vascular damage^[68,69]. Studies have shown that human endothelial cell detachment is produced by exposure to H₂O₂^[43]. The presence of CECs in patients with sepsis but without shock suggests that endothelial damage precedes the development of organ damage^[68]. This is compatible with H₂O₂ release from organ parenchymal cells into the capillary vascular bed causing microvascular dysfunction and edema with subsequent development of organ failure.

Sepsis associated encephalopathy

Sepsis associated encephalopathy (SAE) is a diffuse ce-

rebral dysfunction occurring in the setting of sepsis but without direct infection of the central nervous system^[70]. SAE is characterized by alterations in mental status and motor activity that can range from inattention, disorientation and delirium to agitation, hypoactivity and coma^[71,72]. Delirium is frequently the first manifestation of sepsis and often precedes organ failure^[73,74]. SAE is reported to occur in up to 70% of septic patients (71).

Neurons are especially sensitive to H₂O₂ induced oxidative damage. Studies have shown a concentration dependent cell death starting at 10 μmol/L when neurons are exposed to H₂O₂^[75]. The tripeptide glutathione is critically important in order to prevent oxidative damage of the brain due to H₂O₂^[76]. Glutathione is composed of amino acids glycine, cysteine and glutamate. Cysteine is the rate limiting substrate for neuronal glutathione synthesis and transsulfuration of homocysteine is a major source of cysteine in most cells. However, the brain's neuronal transsulfuration pathway is thought to be a negligible source of cysteine due to low activity of neuronal cystathionine-gamma-lyase (EC 4.4.1.1), a crucial enzyme in the transsulfuration pathway leading to the synthesis of cysteine^[77,78]. Neurons, therefore, rely mainly on the absorption of extracellular cysteine provided by astrocytes for the synthesis of glutathione^[77]. Thus, the dependence of brain neurons on extracellular cysteine in order to synthesize glutathione severely limits their ability to upregulate antioxidant defenses in response to H₂O₂ mediated oxidative stress. This makes brain neurons highly vulnerable to H₂O₂ oxidative damage and dysfunction. This is consistent with the encephalopathy that is reported to occur after accidental ingestion of H₂O₂^[79]. Encephalopathy was also a manifestation after intravenous administration of H₂O₂ during alternative medicine therapy^[59].

The main interaction site of neurons and astrocytes is the synaptic cleft^[80]. Astrocytes export glutathione directly into the synaptic cleft. Ectoenzymes present in the synapse enzymatically release cysteine from glutathione after which cysteine is transported into neurons by the membrane bound EAAT3 transporter (excitatory amino acid transporter 3)^[77-82]. H₂O₂ can react non-enzymatically with cysteine in the synaptic cleft to produce cystine^[83]. This removes cysteine from the synapse and prevents its importation into the neuron resulting in oxidative stress by decreasing the synthesis of neuronal glutathione. The presence of thiols (*i.e.*, cysteine) in the synaptic cleft suggests that this region can function as a sink for H₂O₂ resulting in disruption of synaptic transmission as a result of peroxidation of synaptic cellular membranes.

Thus, circulating H₂O₂ can permeate the brain during the initial hypermetabolic systemic inflammatory response syndrome (SIRS) phase of sepsis and disrupt brain function in the early stages of disease. Due to their limited capacity to detoxify H₂O₂, brain neurons are the first cells to be affected by H₂O₂ induced oxidative stress^[84]. This is consistent with the observation that encephalopathy is often the first sign of sepsis.

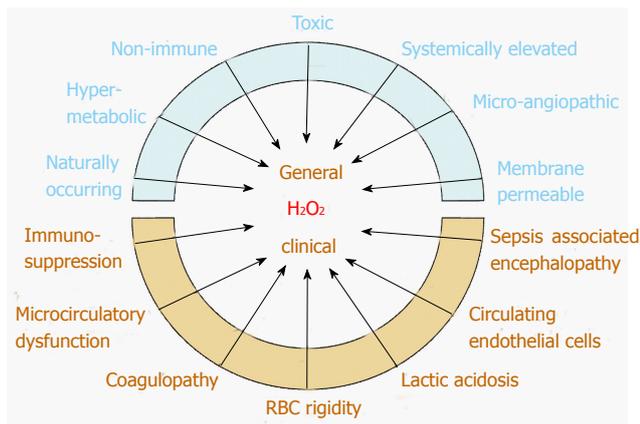


Figure 2 Pathologically elevated serum H₂O₂ levels can account for the general physiological, histological and clinical abnormalities observed in septic shock. Red blood cell glutathione accounts for a major portion of serum redox buffering capacity and is depleted in septic shock non-survivors vs. survivors. Brain neuron function is highly vulnerable to H₂O₂ oxidative stress and is manifested by electroencephalographic changes, which can appear before clinical encephalopathy is evident. Studies show that septic shock survivors upregulate serum antioxidant capacity (which decreases H₂O₂), while non-survivors are unable to do so. This suggests that elevated H₂O₂ is a necessary concomitant to the development of septic shock and recovery is preceded by decreasing H₂O₂. The individual clinical course, bookended by these extremes of H₂O₂, is influenced by parameters such as individual antioxidant capacity, susceptibility to oxidative stress, co-morbidities, age, general health and organ system involved.

LACTIC ACIDOSIS

Sepsis related lactic acidosis is generally attributed to tissue hypoxia. Although tissue hypoxia can result in lactic acidosis it is unsuitable as a general mechanism to explain the appearance of lactic acidosis in septic patients when tissue oxygenation can be normal or even increased^[85].

Under normal circumstances pyruvate, the end product of glycolysis in the cytoplasm, is transported into mitochondria where it is oxidized by the Krebs cycle. Lactate synthesis increases when the rate of pyruvate formation in the cytoplasm exceeds its rate of oxidation by the mitochondria. The excess pyruvate in the cytoplasm is then converted to lactate by lactate dehydrogenase and released into the blood stream resulting in lactic acidosis.

Inhibition of Krebs cycle enzymes will decrease pyruvate oxidation resulting in lactic acidosis. This has been observed with inherited deficiency of alpha-ketoglutarate dehydrogenase resulting in severe congenital lactic acidosis^[86]. Alpha-ketoglutarate dehydrogenase is also highly sensitive to oxidative inhibition by hydrogen peroxide^[87]. Rising systemic concentrations of H₂O₂ in sepsis can account for the observed lactic acidosis with normal tissue oxygen perfusion. This has been termed cytopathic hypoxia. In this case the lactic acidosis is an epiphenomenon of a much more serious underlying metabolic abnormality and treatment of the acidosis does not resolve the inhibition of the Krebs cycle.

DISCUSSION

A hypermetabolic state can develop very quickly after

a generalized septic or non-septic insult to the body. At the heart of the hypermetabolic state is a significantly increased bioenergetic response resulting mainly from enhanced ETC activity. The ETC is an assembly of intramitochondrial protein complexes that converts the energy of high-energy electrons into a form that is used to synthesize ATP, a high energy molecule that powers most energy requiring biosynthetic reactions and physiological functions. Thus, the high energy demands of body systems resulting from a generalized septic or non-septic insult are principally met by increased ATP production, which is manifested as a hypermetabolic state and recognized by the same parameters used to define a SIRS such as increased body temperature, heart rate, respiratory rate and increased white blood cell count.

A principle metabolic by-product of ETC activity is hydrogen peroxide; a highly toxic oxidizing agent. Hydrogen peroxide is produced when electrons spontaneously escape from the ETC and combine with available vicinal oxygen to generate superoxide that is enzymatically converted to H₂O₂ by superoxide dismutase. The increased amount of H₂O₂ generated during a hypermetabolic state can overwhelm the cell's anti-oxidant enzymatic defenses resulting in net intracellular H₂O₂ accumulation. The excess H₂O₂ can oxidatively inhibit enzyme systems including those needed to neutralize H₂O₂ resulting in a positive bio-feedback loop and a vicious cycle of ever increasing intracellular H₂O₂^[88]. Glutathione functions as a cofactor for GPx, which enzymatically neutralizes H₂O₂. GPx is inhibited by the rising concentrations of H₂O₂, which explains why exogenously supplied N-acetylcysteine has no effect on the course of septic shock since glutathione cannot be utilized by GPx to neutralize H₂O₂^[88,89].

Hydrogen peroxide is biomembrane permeable and can diffuse into the bloodstream where it is distributed to all organs of the body generating a state of severe systemic oxidative stress. Studies have documented high levels of H₂O₂ in the blood and urine of septic patients^[27,90]. This can result in the multi-organ failure and microangiopathic dysfunction characteristic of septic shock. Genetic variation in glutathione levels as well as age related decline has been reported^[91-93]. This may compromise the ability to neutralize H₂O₂ and predispose individuals to vasoplegic (*i.e.*, septic) shock and multi-organ failure during acute hypermetabolic periods, especially in older individuals. Studies have shown that glutathione is essential for cell survival^[94].

CONCLUSION

Taken together, the evidence suggests that septic shock is a primary radical induction process that has its origins early in the development of sepsis with the accumulation and generalized dispersal of cytotoxic levels of H₂O₂. This arises secondary to glutathione depletion as a result of a systemic inflammatory mediated hypermetabolic state. Studies have shown that systemic inflammation significantly reduces GSH levels, and GSH deficient animals

subjected to shock develop hypotension, kidney and liver failure, increased organ bacteria and dramatic increases in mortality rates^[95-97].

The near universal requirement of glutathione for cellular function and the pathological accumulation of H₂O₂ that ensues when glutathione is deficient can affect every organ in the body. Studies have shown that H₂O₂ can reproduce the clinico-pathological abnormalities observed in septic shock (Figure 2).

Kept in check, the high membrane diffusability of H₂O₂ allows it to fulfill its physiological role as a cellular messenger but also creates the potential for a pathophysiological response during times of metabolic stress when reductive (anti-oxidant) mechanisms can become overwhelmed as a consequence of hyper-metabolic H₂O₂ production^[98]. This is further exacerbated by nutritional deficits that may arise during the course of acute illness in addition to the effect of glutathione deficiency itself, which as master antioxidant of the cell, supplies reducing equivalents to maintain proteins in their reduced (and functional) state^[14].

REFERENCES

- 1 **Goldenberg NM**, Steinberg BE, Slutsky AS, Lee WL. Broken barriers: a new take on sepsis pathogenesis. *Sci Transl Med* 2011; **3**: 88ps25 [PMID: 21697528 DOI: 10.1126/scitranslmed.3002011]
- 2 **Marshall JC**, Vincent JL, Guyatt G, Angus DC, Abraham E, Bernard G, Bombardier C, Calandra T, Jørgensen HS, Sylvester R, Boers M. Outcome measures for clinical research in sepsis: a report of the 2nd Cambridge Colloquium of the International Sepsis Forum. *Crit Care Med* 2005; **33**: 1708-1716 [PMID: 16096445 DOI: 10.1097/01.CCM.0000174478.70338.03]
- 3 **Melamed A**, Sorvillo FJ. The burden of sepsis-associated mortality in the United States from 1999 to 2005: an analysis of multiple-cause-of-death data. *Crit Care* 2009; **13**: R28 [PMID: 19250547 DOI: 10.1186/cc7733]
- 4 **American Cancer Society, Surveillance Research**. Estimated New Cancer Cases and Deaths by Sex for All Sites, US, 2011. Available from: URL: http://seer.cancer.gov/csr/1975_2008/results_single/sect_01_table.01.pdf Accessed 20/Jan/2014
- 5 **Xiao W**, Mindrinos MN, Seok J, Cuschieri J, Cuenca AG, Gao H, Hayden DL, Hennessy L, Moore EE, Minei JP, Bankey PE, Johnson JL, Sperry J, Nathens AB, Billiar TR, West MA, Brownstein BH, Mason PH, Baker HV, Finnerty CC, Jeschke MG, López MC, Klein MB, Gamelli RL, Gibran NS, Arnoldo B, Xu W, Zhang Y, Calvano SE, McDonald-Smith GP, Schoenfeld DA, Storey JD, Cobb JP, Warren HS, Moldawer LL, Herndon DN, Lowry SF, Maier RV, Davis RW, Tompkins RG. A genomic storm in critically injured humans. *J Exp Med* 2011; **208**: 2581-2590 [PMID: 22110166 DOI: 10.1084/jem.20111354]
- 6 **Suffredini AF**, Munford RS. Novel therapies for septic shock over the past 4 decades. *JAMA* 2011; **306**: 194-199 [PMID: 21750297 DOI: 10.1001/jama.2011.909]
- 7 **Bloch KC**. Infectious Diseases. In: McPhee SJ, Hammer GD: Pathophysiology of Disease. New York: McGraw-Hill, 2010: 57-83
- 8 **Borgen L**. Total parenteral nutrition in adults. *Am J Nurs* 1978; **78**: 224-228 [PMID: 417629 DOI: 10.2307/3424283]
- 9 **Depuydt M**, Messens J, Collet JF. How proteins form disulfide bonds. *Antioxid Redox Signal* 2011; **15**: 49-66 [PMID: 20849374 DOI: 10.1089/ars.2010.3575]
- 10 **Kelley EE**, Khoo NK, Hundley NJ, Malik UZ, Freeman BA, Tarpey MM. Hydrogen peroxide is the major oxidant product of xanthine oxidase. *Free Radic Biol Med* 2010; **48**: 493-498 [PMID: 19941951 DOI: 10.1016/j.freeradbiomed.2009.11.012]
- 11 **Murphy MP**. How mitochondria produce reactive oxygen species. *Biochem J* 2009; **417**: 1-13 [PMID: 19061483 DOI: 10.1042/BJ20081386]
- 12 **Schrader M**, Fahimi HD. Mammalian peroxisomes and reactive oxygen species. *Histochem Cell Biol* 2004; **122**: 383-393 [PMID: 15241609 DOI: 10.1007/s00418-004-0673-1]
- 13 **Forman HJ**, Zhang H, Rinna A. Glutathione: overview of its protective roles, measurement, and biosynthesis. *Mol Aspects Med* 2009; **30**: 1-12 [PMID: 18796312 DOI: 10.1016/j.mam.2008.08.006]
- 14 **Wu G**, Fang YZ, Yang S, Lupton JR, Turner ND. Glutathione metabolism and its implications for health. *J Nutr* 2004; **134**: 489-492 [PMID: 14988435]
- 15 **Lyons J**, Rauh-Pfeiffer A, Ming-Yu Y, Lu XM, Zurakowski D, Curley M, Collier S, Duggan C, Nurko S, Thompson J, Ajami A, Borgonha S, Young VR, Castillo L. Cysteine metabolism and whole blood glutathione synthesis in septic pediatric patients. *Crit Care Med* 2001; **29**: 870-877 [PMID: 11373484 DOI: 10.1097/00003246-200104000-00036]
- 16 **Biolo G**, Antonione R, De Cicco M. Glutathione metabolism in sepsis. *Crit Care Med* 2007; **35**: S591-S595 [PMID: 17713414 DOI: 10.1097/01.CCM.0000278913.19123.13]
- 17 **Pacht ER**, Timerman AP, Lykens MG, Merola AJ. Deficiency of alveolar fluid glutathione in patients with sepsis and the adult respiratory distress syndrome. *Chest* 1991; **100**: 1397-1403 [PMID: 1935300 DOI: 10.1378/chest.100.5.1397]
- 18 **Hammarqvist F**, Luo JL, Cotgreave IA, Andersson K, Wernerman J. Skeletal muscle glutathione is depleted in critically ill patients. *Crit Care Med* 1997; **25**: 78-84 [PMID: 8989180 DOI: 10.1097/00003246-199701000-00016]
- 19 **Karapetsa M**, Pitsika M, Goutzourelas N, Stagos D, Tousia Becker A, Zakyntinos E. Oxidative status in ICU patients with septic shock. *Food Chem Toxicol* 2013; **61**: 106-111 [PMID: 23542126 DOI: 10.1016/j.fct.2013.03.026]
- 20 **Sjövall F**, Morota S, Hansson MJ, Friberg H, Gnaiger E, Elmér E. Temporal increase of platelet mitochondrial respiration is negatively associated with clinical outcome in patients with sepsis. *Crit Care* 2010; **14**: R214 [PMID: 21106065 DOI: 10.1186/cc9337]
- 21 **Shokolenko I**, Venediktova N, Bochkareva A, Wilson GL, Alexeyev MF. Oxidative stress induces degradation of mitochondrial DNA. *Nucleic Acids Res* 2009; **37**: 2539-2548 [PMID: 19264794 DOI: 10.1093/nar/gkp100]
- 22 **Van Houten B**, Woshner V, Santos JH. Role of mitochondrial DNA in toxic responses to oxidative stress. *DNA Repair (Amst)* 2006; **5**: 145-152 [PMID: 15878696 DOI: 10.1016/j.dnarep.2005.03.002]
- 23 **Malinouski M**, Zhou Y, Belousov VV, Hatfield DL, Gladyshev VN. Hydrogen peroxide probes directed to different cellular compartments. *PLoS One* 2011; **6**: e14564 [PMID: 21283738 DOI: 10.1371/journal.pone.0014564]
- 24 **Huet O**, Cherreau C, Nicco C, Dupic L, Conti M, Borderie D, Pene F, Vicaut E, Benhamou D, Mira JP, Duranteau J, Batteux F. Pivotal role of glutathione depletion in plasma-induced endothelial oxidative stress during sepsis. *Crit Care Med* 2008; **36**: 2328-2334 [PMID: 18664787 DOI: 10.1097/CCM.0b013e3181800387]
- 25 **Chen J**, Gu Y, Shao Z, Luo J, Tan Z. Propofol protects against hydrogen peroxide-induced oxidative stress and cell dysfunction in human umbilical vein endothelial cells. *Mol Cell Biochem* 2010; **339**: 43-54 [PMID: 20039104 DOI: 10.1007/s11010-009-0368-y]
- 26 **Li ZL**, Liu JC, Hu J, Li XQ, Wang SW, Yi DH, Zhao MG. Protective effects of hyperoside against human umbilical vein endothelial cell damage induced by hydrogen peroxide.

- J Ethnopharmacol* 2012; **139**: 388-394 [PMID: 22120016 DOI: 10.1016/j.jep.2011.11.020]
- 27 **Mathru M**, Rooney MW, Dries DJ, Hirsch LJ, Barnes L, Tobin MJ. Urine hydrogen peroxide during adult respiratory distress syndrome in patients with and without sepsis. *Chest* 1994; **105**: 232-236 [PMID: 8275738 DOI: 10.1378/chest.105.1.232]
- 28 **Cowley HC**, Bacon PJ, Goode HF, Webster NR, Jones JG, Menon DK. Plasma antioxidant potential in severe sepsis: a comparison of survivors and nonsurvivors. *Crit Care Med* 1996; **24**: 1179-1183 [PMID: 8674332 DOI: 10.1097/00003246-199607000-00019]
- 29 **Lehr HA**, Bittinger F, Kirkpatrick CJ. Microcirculatory dysfunction in sepsis: a pathogenetic basis for therapy? *J Pathol* 2000; **190**: 373-386 [PMID: 10685071]
- 30 **Lupu C**, Westmuckett AD, Peer G, Ivanciu L, Zhu H, Taylor FB, Lupu F. Tissue factor-dependent coagulation is preferentially up-regulated within arterial branching areas in a baboon model of Escherichia coli sepsis. *Am J Pathol* 2005; **167**: 1161-1172 [PMID: 16192650 DOI: 10.1016/S0002-9440(10)61204-7]
- 31 **Birukova AA**, Arce FT, Moldobaeva N, Dudek SM, Garcia JG, Lal R, Birukov KG. Endothelial permeability is controlled by spatially defined cytoskeletal mechanics: atomic force microscopy force mapping of pulmonary endothelial monolayer. *Nanomedicine* 2009; **5**: 30-41 [PMID: 18824415 DOI: 10.1016/j.nano.2008.07.002]
- 32 **Hastie LE**, Patton WF, Hechtman HB, Shepro D. H₂O₂-induced filamin redistribution in endothelial cells is modulated by the cyclic AMP-dependent protein kinase pathway. *J Cell Physiol* 1997; **172**: 373-381 [PMID: 9284957]
- 33 **López-Ongil S**, Torrecillas G, Pérez-Sala D, González-Santiago L, Rodríguez-Puyol M, Rodríguez-Puyol D. Mechanisms involved in the contraction of endothelial cells by hydrogen peroxide. *Free Radic Biol Med* 1999; **26**: 501-510 [PMID: 10218638 DOI: 10.1016/S0891-5849(98)00223-8]
- 34 **Kevil CG**, Okayama N, Alexander JS. H(2)O(2)-mediated permeability II: importance of tyrosine phosphatase and kinase activity. *Am J Physiol Cell Physiol* 2001; **281**: C1940-C1947 [PMID: 11698252]
- 35 **Kevil CG**, Oshima T, Alexander B, Coe LL, Alexander JS. H(2)O(2)-mediated permeability: role of MAPK and occludin. *Am J Physiol Cell Physiol* 2000; **279**: C21-C30 [PMID: 10898713]
- 36 **Pearse DB**, Shimoda LA, Verin AD, Bogatcheva N, Moon C, Ronnett GV, Welsh LE, Becker PM. Effect of cGMP on lung microvascular endothelial barrier dysfunction following hydrogen peroxide. *Endothelium* 2003; **10**: 309-317 [PMID: 14741846 DOI: 10.1080/174007541]
- 37 **Valen G**, Sondén A, Vaage J, Malm E, Kjellström BT. Hydrogen peroxide induces endothelial cell atypia and cytoskeleton depolymerization. *Free Radic Biol Med* 1999; **26**: 1480-1488 [PMID: 10401612 DOI: 10.1016/S0891-5849(99)00009-X]
- 38 **Bradley JR**, Thiru S, Pober JS. Hydrogen peroxide-induced endothelial retraction is accompanied by a loss of the normal spatial organization of endothelial cell adhesion molecules. *Am J Pathol* 1995; **147**: 627-641 [PMID: 7677177]
- 39 **Zhao Y**, Davis HW. Hydrogen peroxide-induced cytoskeletal rearrangement in cultured pulmonary endothelial cells. *J Cell Physiol* 1998; **174**: 370-379 [PMID: 9462699 DOI: 10.1002/(SICI)1097-4652(199803)174]
- 40 **Hirano S**, Rees RS, Yancy SL, Welsh MJ, Remick DG, Yamada T, Hata J, Gilmont RR. Endothelial barrier dysfunction caused by LPS correlates with phosphorylation of HSP27 in vivo. *Cell Biol Toxicol* 2004; **20**: 1-14 [PMID: 15119843]
- 41 **Habib MP**, Clements NC. Effects of low-dose hydrogen peroxide in the isolated perfused rat lung. *Exp Lung Res* 1995; **21**: 95-112 [PMID: 7729381 DOI: 10.3109/01902149509031747]
- 42 **Gilmont RR**, Dardano A, Young M, Engle JS, Adamson BS, Smith DJ, Rees RS. Effects of glutathione depletion on oxidant-induced endothelial cell injury. *J Surg Res* 1998; **80**: 62-68 [PMID: 9790816 DOI: 10.1006/jsre.1998.5328]
- 43 **Shingu M**, Yoshioka K, Nobunaga M, Yoshida K. Human vascular smooth muscle cells and endothelial cells lack catalase activity and are susceptible to hydrogen peroxide. *Inflammation* 1985; **9**: 309-320 [PMID: 4044027 DOI: 10.1007/BF00916279]
- 44 **Lee HS**, Namkoong K, Kim DH, Kim KJ, Cheong YH, Kim SS, Lee WB, Kim KY. Hydrogen peroxide-induced alterations of tight junction proteins in bovine brain microvascular endothelial cells. *Microvasc Res* 2004; **68**: 231-238 [PMID: 15501242 DOI: 10.1016/j.mvr.2004.07.005]
- 45 **Liu SF**, Malik AB. NF-kappa B activation as a pathological mechanism of septic shock and inflammation. *Am J Physiol Lung Cell Mol Physiol* 2006; **290**: L622-L645 [PMID: 16531564 DOI: 10.1152/ajplung.00477.2005]
- 46 **Arnalich F**, Garcia-Palomero E, López J, Jiménez M, Madero R, Renart J, Vázquez JJ, Montiel C. Predictive value of nuclear factor kappaB activity and plasma cytokine levels in patients with sepsis. *Infect Immun* 2000; **68**: 1942-1945 [PMID: 10722586 DOI: 10.1128/IAI.68.4.1942-1945.2000]
- 47 **Böhrer H**, Qiu F, Zimmermann T, Zhang Y, Jllmer T, Männel D, Böttiger BW, Stern DM, Waldherr R, Saeger HD, Ziegler R, Bierhaus A, Martin E, Nawroth PP. Role of NFkappaB in the mortality of sepsis. *J Clin Invest* 1997; **100**: 972-985 [PMID: 9276714 DOI: 10.1172/JCI119648]
- 48 **Hotta N**, Ichiyama T, Shiraishi M, Takekawa T, Matsubara T, Furukawa S. Nuclear factor-kappaB activation in peripheral blood mononuclear cells in children with sepsis. *Crit Care Med* 2007; **35**: 2395-2401 [PMID: 17944030 DOI: 10.1097/01.CCM.0000284502.38701.E6]
- 49 **Schreck R**, Albermann K, Baeuerle PA. Nuclear factor kappa B: an oxidative stress-responsive transcription factor of eukaryotic cells (a review). *Free Radic Res Commun* 1992; **17**: 221-237 [PMID: 1473734 DOI: 10.3109/10715769209079515]
- 50 **Takada Y**, Mukhopadhyay A, Kundu GC, Mahabeshwar GH, Singh S, Aggarwal BB. Hydrogen peroxide activates NF-kappa B through tyrosine phosphorylation of I kappa B alpha and serine phosphorylation of p65: evidence for the involvement of I kappa B alpha kinase and Syk protein-tyrosine kinase. *J Biol Chem* 2003; **278**: 24233-24241 [PMID: 12711606 DOI: 10.1074/jbc.M212389200]
- 51 **van den Berg R**, Haenen GR, van den Berg H, Bast A. Transcription factor NF-kappaB as a potential biomarker for oxidative stress. *Br J Nutr* 2001; **86** Suppl 1: S121-S127 [PMID: 11520430 DOI: 10.1079/BJN2001340]
- 52 **Niethammer P**, Grabher C, Look AT, Mitchison TJ. A tissue-scale gradient of hydrogen peroxide mediates rapid wound detection in zebrafish. *Nature* 2009; **459**: 996-999 [PMID: 19494811 DOI: 10.1038/nature08119]
- 53 **Klyubin IV**, Kirpichnikova KM, Gamaley IA. Hydrogen peroxide-induced chemotaxis of mouse peritoneal neutrophils. *Eur J Cell Biol* 1996; **70**: 347-351 [PMID: 8864663]
- 54 **Mathias JR**, Perrin BJ, Liu TX, Kanki J, Look AT, Huttenlocher A. Resolution of inflammation by retrograde chemotaxis of neutrophils in transgenic zebrafish. *J Leukoc Biol* 2006; **80**: 1281-1288 [PMID: 16963624 DOI: 10.1189/jlb.0506346]
- 55 **Wang L**, Bastarache JA, Ware LB. The coagulation cascade in sepsis. *Curr Pharm Des* 2008; **14**: 1860-1869 [PMID: 18691097 DOI: 10.2174/138161208784980581]
- 56 **Zeerleder S**, Hack CE, Wuillemin WA. Disseminated intravascular coagulation in sepsis. *Chest* 2005; **128**: 2864-2875 [PMID: 16236964 DOI: 10.1378/chest.128.4.2864]
- 57 **Crawley JT**, Lane DA. The haemostatic role of tissue factor pathway inhibitor. *Arterioscler Thromb Vasc Biol* 2008; **28**: 233-242 [PMID: 17951326 DOI: 10.1161/ATVBAHA.107.141606]
- 58 **Ambrosio G**, Tritto I, Golino P. Reactive oxygen metabolites and arterial thrombosis. *Cardiovasc Res* 1997; **34**: 445-452 [PMID: 9231027 DOI: 10.1016/S0008-6363(97)00101-6]

- 59 **Wetter DA**, Davis MD. Ulceration of the arm attributed to a spider bite and treated with intravenous hydrogen peroxide: a cautionary tale. *Arch Dermatol* 2006; **142**: 1658-1659 [PMID: 17179007 DOI: 10.1001/archderm.142.12.1658]
- 60 **Stearns-Kurosawa DJ**, Osuchowski MF, Valentine C, Kurosawa S, Remick DG. The pathogenesis of sepsis. *Annu Rev Pathol* 2011; **6**: 19-48 [PMID: 20887193 DOI: 10.1146/annurev-pathol-011110-130327]
- 61 **Cerella C**, Coppola S, Maresca V, De Nicola M, Radogna F, Ghibelli L. Multiple mechanisms for hydrogen peroxide-induced apoptosis. *Ann N Y Acad Sci* 2009; **1171**: 559-563 [PMID: 19723104 DOI: 10.1111/j.1749-6632.2009.04901.x]
- 62 **Armstrong JS**, Steinauer KK, Hornung B, Irish JM, Lecane P, Birrell GW, Peehl DM, Knox SJ. Role of glutathione depletion and reactive oxygen species generation in apoptotic signaling in a human B lymphoma cell line. *Cell Death Differ* 2002; **9**: 252-263 [PMID: 11859408 DOI: 10.1038/sj.cdd.4400959]
- 63 **Kinscherf R**, Fischbach T, Mihm S, Roth S, Hohenhaus-Sievert E, Weiss C, Edler L, Bärtsch P, Dröge W. Effect of glutathione depletion and oral N-acetyl-cysteine treatment on CD4+ and CD8+ cells. *FASEB J* 1994; **8**: 448-451 [PMID: 7909525]
- 64 **Snyder LM**, Fortier NL, Trainor J, Jacobs J, Leb L, Lubin B, Chiu D, Shohet S, Mohandas N. Effect of hydrogen peroxide exposure on normal human erythrocyte deformability, morphology, surface characteristics, and spectrin-hemoglobin cross-linking. *J Clin Invest* 1985; **76**: 1971-1977 [PMID: 4056060 DOI: 10.1172/JCI112196]
- 65 **Machiedo GW**, Powell RJ, Rush BF, Swislocki NI, Dikdan G. The incidence of decreased red blood cell deformability in sepsis and the association with oxygen free radical damage and multiple-system organ failure. *Arch Surg* 1989; **124**: 1386-1389 [PMID: 2589962 DOI: 10.1001/archsurg.1989.01410120032007]
- 66 **Erdbruegger U**, Dhaygude A, Haubitz M, Woywodt A. Circulating endothelial cells: markers and mediators of vascular damage. *Curr Stem Cell Res Ther* 2010; **5**: 294-302 [PMID: 20528750 DOI: 10.2174/157488810793351721]
- 67 **Wu H**, Chen H, Hu PC. Circulating endothelial cells and endothelial progenitors as surrogate biomarkers in vascular dysfunction. *Clin Lab* 2007; **53**: 285-295 [PMID: 17605403]
- 68 **Mutunga M**, Fulton B, Bullock R, Batchelor A, Gascoigne A, Gillespie JJ, Baudouin SV. Circulating endothelial cells in patients with septic shock. *Am J Respir Crit Care Med* 2001; **163**: 195-200 [PMID: 11208646 DOI: 10.1164/ajrccm.163.1.9912036]
- 69 **Schlichting DE**, Waxman AB, O'Brien LA, Wang T, Naum CC, Rubeiz GJ, Um SL, Williams M, Yan SC. Circulating endothelial and endothelial progenitor cells in patients with severe sepsis. *Microvasc Res* 2011; **81**: 216-221 [PMID: 21130783 DOI: 10.1016/j.mvr.2010.11.011]
- 70 **Goffton TE**, Young GB. Sepsis-associated encephalopathy. *Nat Rev Neurol* 2012; **8**: 557-566 [PMID: 22986430 DOI: 10.1038/nrneuro.2012.183]
- 71 **Lamar CD**, Hurley RA, Taber KH. Sepsis-associated encephalopathy: review of the neuropsychiatric manifestations and cognitive outcome. *J Neuropsychiatry Clin Neurosci* 2011; **23**: 237-241 [PMID: 21948885 DOI: 10.1176/appi.neuropsych.23.3.237]
- 72 Siami S, Polito A, Sharshar T. Sepsis-associated Encephalopathy. In Vincent JL Yearbook of Intensive Care and Emergency Medicine. Berlin-Heidelberg: Springer, 2009: 809-816
- 73 **Wilson JX**, Young GB. Progress in clinical neurosciences: sepsis-associated encephalopathy: evolving concepts. *Can J Neurol Sci* 2003; **30**: 98-105 [PMID: 12774948]
- 74 **Ringer TM**, Axer H, Romeike BF, Zinke J, Brunkhorst F, Witte OW, Günther A. Neurological Sequelae of Sepsis: I) Septic Encephalopathy. *Open Crit Care Med J* 2011; **4**: 2-7 [DOI: 10.2174/1874828701104010002]
- 75 **Desagher S**, Glowinski J, Premont J. Astrocytes protect neurons from hydrogen peroxide toxicity. *J Neurosci* 1996; **16**: 2553-2562 [PMID: 8786431]
- 76 **Dringen R**, Pawlowski PG, Hirrlinger J. Peroxide detoxification by brain cells. *J Neurosci Res* 2005; **79**: 157-165 [PMID: 15573410 DOI: 10.1002/jnr.20280]
- 77 **Aoyama K**, Watabe M, Nakaki T. Regulation of neuronal glutathione synthesis. *J Pharmacol Sci* 2008; **108**: 227-238 [PMID: 19008644 DOI: 10.1254/jphs.08R01CR]
- 78 **Deth R**, Muratore C. The redox/methylation hypothesis of autism. In: Chauhan A, Chauhan V, Brown WT : Autism: Oxidative Stress, Inflammation and immune abnormalities. Florida: Florida CRC Press, 2010: 113-130
- 79 **Cannon G**, Caravati EM, Filloux FM. Hydrogen peroxide neurotoxicity in childhood: case report with unique magnetic resonance imaging features. *J Child Neurol* 2003; **18**: 805-808 [PMID: 14696912 DOI: 10.1177/08830738030180111501]
- 80 **Cakir T**, Alsan S, Saybasili H, Akin A, Ulgen KO. Reconstruction and flux analysis of coupling between metabolic pathways of astrocytes and neurons: application to cerebral hypoxia. *Theor Biol Med Model* 2007; **4**: e48
- 81 **Wang XF**, Cynader MS. Astrocytes provide cysteine to neurons by releasing glutathione. *J Neurochem* 2000; **74**: 1434-1442 [PMID: 10737599 DOI: 10.1046/j.1471-4159.2000.0741434.x]
- 82 **Dringen R**, Hirrlinger J. Glutathione pathways in the brain. *Biol Chem* 2003; **384**: 505-516 [PMID: 12751781 DOI: 10.1515/BC.2003.059]
- 83 **Luo D**, Smith SW, Anderson BD. Kinetics and mechanism of the reaction of cysteine and hydrogen peroxide in aqueous solution. *J Pharm Sci* 2005; **94**: 304-316 [PMID: 15570599 DOI: 10.1002/jps.20253]
- 84 **Chauhan A**, Chauhan V. Oxidative stress in autism. *Pathophysiology* 2006; **13**: 171-181 [PMID: 16766163 DOI: 10.1201/9781420068870]
- 85 **Bellomo R**, Ronco C. The pathogenesis of lactic acidosis in sepsis. *Curr Opin Crit Care* 1999; **5**: 452-457 [DOI: 10.1097/00075198-199912000-00008]
- 86 **Bonnefont JP**, Chretien D, Rustin P, Robinson B, Vassault A, Aupetit J, Charpentier C, Rabier D, Saudubray JM, Munnich A. Alpha-ketoglutarate dehydrogenase deficiency presenting as congenital lactic acidosis. *J Pediatr* 1992; **121**: 255-258 [PMID: 1640293 DOI: 10.1016/S0022-3476(05)81199-0]
- 87 **Tretter L**, Adam-Vizi V. Inhibition of Krebs cycle enzymes by hydrogen peroxide: A key role of [alpha]-ketoglutarate dehydrogenase in limiting NADH production under oxidative stress. *J Neurosci* 2000; **20**: 8972-8979 [PMID: 11124972]
- 88 **Cho CS**, Lee S, Lee GT, Woo HA, Choi EJ, Rhee SG. Irreversible inactivation of glutathione peroxidase 1 and reversible inactivation of peroxiredoxin II by H₂O₂ in red blood cells. *Antioxid Redox Signal* 2010; **12**: 1235-1246 [PMID: 20070187 DOI: 10.1089/ars.2009.2701]
- 89 **Kinnula VL**, Everitt JL, Mangum JB, Chang LY, Crapo JD. Antioxidant defense mechanisms in cultured pleural mesothelial cells. *Am J Respir Cell Mol Biol* 1992; **7**: 95-103 [PMID: 1627338 DOI: 10.1165/ajrcmb/7.1.95]
- 90 **Pherwani AV**, Puri VC, V Malhotra V. Estimation of hydrogen peroxide levels in the blood and urine of normal infants and infants with sepsis. *Bombay Hosp J* 1999; **41**: 8
- 91 **van 't Erve TJ**, Wagner BA, Ryckman KK, Raife TJ, Buettner GR. The concentration of glutathione in human erythrocytes is a heritable trait. *Free Radic Biol Med* 2013; **65**: 742-749 [PMID: 23938402 DOI: 10.1016/j.freeradbiomed.2013.08.002]
- 92 **Caprari P**, Caforio MP, Cianciulli P, Maffi D, Pasquino MT, Tarzia A, Amadori S, Salvati AM. 6-Phosphogluconate dehydrogenase deficiency in an Italian family. *Ann Hematol* 2001; **80**: 41-44 [PMID: 11233775 DOI: 10.1007/s002770000233]
- 93 **Lang CA**, Naryshkin S, Schneider DL, Mills BJ, Lindeman RD. Low blood glutathione levels in healthy aging adults. *J Lab Clin Med* 1992; **120**: 720-725 [PMID: 1431500]
- 94 **Hatem E**, Berthonaud V, Dardalhon M, Lagniel G, Baudouin-Cornu P, Huang ME, Labarre J, Chédin S. Glutathione is essential to preserve nuclear function and cell survival under

Pravda J. Metabolic theory of septic shock

- oxidative stress. *Free Radic Biol Med* 2014; **67**: 103-114 [PMID: 24145121 DOI: 10.1016/j.freeradbiomed.2013.10.807]
- 95 **Keller GA**, Barke R, Harty JT, Humphrey E, Simmons RL. Decreased hepatic glutathione levels in septic shock. Predisposition of hepatocytes to oxidative stress: an experimental approach. *Arch Surg* 1985; **120**: 941-945 [PMID: 3893390 DOI: 10.1001/archsurg.1985.01390320065013]
- 96 **Ikegami K**, Lalonde C, Young YK, Picard L, Demling R. Comparison of plasma reduced glutathione and oxidized glutathione with lung and liver tissue oxidant and anti-oxidant activity during acute inflammation. *Shock* 1994; **1**: 307-312 [PMID: 7735965 DOI: 10.1097/00024382-199404000-00010]
- 97 **Robinson MK**, Rounds JD, Hong RW, Jacobs DO, Wilmore DW. Glutathione deficiency increases organ dysfunction after hemorrhagic shock. *Surgery* 1992; **112**: 140-147; discussion 148-149 [PMID: 1641757]
- 98 **Holmquist L**, Stuchbury G, Steele M, Münch G. Hydrogen peroxide is a true first messenger. *J Neural Transm Suppl* 2007; (**72**): 39-41 [PMID: 17982876 DOI: 10.1007/978-3-211-73574-9_6]

P- Reviewers: Carassiti M, Fink MP, Stover CM, Yao YM
S- Editor: Song XX **L- Editor:** A **E- Editor:** Wu HL



Arterial vs venous blood gas differences during hemorrhagic shock

Kristopher Burton Williams, Ashley Britton Christmas, Brant Todd Heniford, Ronald Fong Sing, Joseph Messick

Kristopher Burton Williams, Ashley Britton Christmas, Brant Todd Heniford, Ronald Fong Sing, Joseph Messick, Department of Surgery, Carolinas HealthCare System, Charlotte, NC 28204, United States

Author contributions: Heniford BT, Sing RF, Messick J designed research; Sing RF, Messick J performed research; Messick J contributed new reagents or analytic tools; Christmas AB, Heniford BT, Sing RF analyzed data; Williams KB, Christmas AB, Heniford BT, Sing RF wrote the paper; Messick J deceased since the completion of this study.

Supported by Carolinas HealthCare System, Department of Surgery, Charlotte, North Carolina, United States

Correspondence to: Ronald Fong Sing, DO, FACS, FCCM, Department of Surgery, Carolinas HealthCare System, 1000 Blythe Boulevard, Charlotte, NC 28203,

United States. ron.sing@carolinashealthcare.org

Telephone: +1-704-3551311 Fax: +1-704-3555619

Received: March 1, 2013 Revised: October 19, 2013

Accepted: March 3, 2014

Published online: May 4, 2014

Abstract

AIM: To characterize differences of arterial (ABG) and venous (VBG) blood gas analysis in a rabbit model of hemorrhagic shock.

METHODS: Following baseline arterial and venous blood gas analysis, fifty anesthetized, ventilated New Zealand white rabbits were hemorrhaged to and maintained at a mean arterial pressure of 40 mmHg until a state of shock was obtained, as defined by arterial pH ≤ 7.2 and base deficit ≤ -15 mmol/L. Simultaneous ABG and VBG were obtained at 3 minute intervals. Comparisons of pH, base deficit, pCO₂, and arteriovenous (a-v) differences were then made between ABG and VBG at baseline and shock states. Statistical analysis was applied where appropriate with a significance of $P < 0.05$.

RESULTS: All 50 animals were hemorrhaged to shock

status and euthanized; no unexpected loss occurred. Significant differences were noted between baseline and shock states in blood gases for the following parameters: pH was significantly decreased in both arterial (7.39 ± 0.12 to 7.14 ± 0.18) and venous blood gases (7.35 ± 0.15 to 6.98 ± 0.26 , $P < 0.05$), base deficit was significantly increased for arterial (-0.9 ± 3.9 mEq/L vs -17.8 ± 2.2 mEq/L) and venous blood gasses (-0.8 ± 3.8 mEq/L vs -15.3 ± 4.1 mEq/L, $P < 0.05$). pCO₂ trends (baseline to shock) demonstrated a decrease in arterial blood (40.0 ± 9.1 mmHg vs 28.9 ± 7.1 mmHg) but an increase in venous blood (46.0 ± 10.1 mmHg vs 62.8 ± 15.3 mmHg), although these trends were non-significant. For calculated arteriovenous differences between baseline and shock states, only the pCO₂ difference was shown to be significant during shock.

CONCLUSION: In this rabbit model, significant differences exist in blood gas measurements for arterial and venous blood after hemorrhagic shock. A widened pCO₂ a-v difference during hemorrhage, reflective of poor tissue oxygenation, may be a better indicator of impending shock.

© 2014 Baishideng Publishing Group Co., Limited. All rights reserved.

Key words: Hemorrhagic shock; pH; Base deficit; Arterial blood gases; Venous blood gases

Core tip: Recent studies regarding early goal directed therapy and damage control resuscitation have indicated a potential role for calculated arteriovenous pCO₂ differences in monitoring resuscitative efforts. In a rabbit model of hemorrhagic shock, we demonstrate significant derangements between arterial and venous blood and, while not a novel concept, explore the potential of central venous pCO₂ as an indicator of hemorrhagic shock. Our results demonstrate a widened arteriovenous pCO₂ difference is significantly associated with hemorrhagic shock and may be a more reliable

indicator of inadequate tissue perfusion and therefore impending circulatory collapse.

Williams KB, Christmas AB, Heniford BT, Sing RF, Messick J. Arterial vs venous blood gas differences during hemorrhagic shock. *World J Crit Care Med* 2014; 3(2): 55-60 Available from: URL: <http://www.wjgnet.com/2220-3141/full/v3/i2/55.htm> DOI: <http://dx.doi.org/10.5492/wjccm.v3.i2.55>

INTRODUCTION

Circulatory collapse is a definitive indicator of the shock state, but may manifest late during hemorrhage leading to delayed diagnosis, resuscitation, and treatment when clinical metrics of circulatory collapse (hypotension, tachycardia, decreased organ perfusion, altered mental status, *etc.*) are the sole measures of a patient's physiologic status. Robust compensatory responses to injury in young, healthy patients can delay treatment of hemorrhage even further as clinical parameters defining shock may not be evident until later stages in the clinical course. Any delay in diagnosis and treatment during massive hemorrhage will likely result in increased morbidity and mortality, fueling the search for adequate trauma resuscitation protocols, such as damage control resuscitation, as well as reliable early markers of impending or ongoing shock^[1].

Serologic markers including pH, base deficit, central venous oxygen saturation and lactate have been used to identify and quantitate shock^[2-6]. Arterial blood gas analysis is considered the gold standard to determine oxygenation and acid-base status in the acutely injured as well as critically ill and repeat testing offers a means of monitoring resuscitation efforts. However, serious, albeit rare, complications of arterial cannulation (pseudoaneurysm, hematoma, hemorrhage, limb ischemia, infection, neurologic injury)^[7] have led to a search for less invasive means of detecting impending shock, quantitating the degree of shock as well as measuring adequate resuscitation. As such, many studies have examined the reliability and accuracy of central venous blood gas in acid-base monitoring as an alternative to arterial blood gas analysis^[8-11]. In previous animal models of severely reduced cardiac output, venous hypercarbia has been shown to correlate with inadequacy of tissue perfusion^[12,13] and changes in venous blood were noted to occur with greater magnitude and earlier in the process of clinical deterioration than those of arterial blood^[14,15]. Similar discrepancies in arterial and venous pCO₂ have been reported in human studies of shock states, as well as the paradox of venous acidemia occurring simultaneously with arterial alkalemia, and have been suggestive of the role of serum pCO₂ differences as an indicator of tissue perfusion^[16-20]. In a recent clinical study highlighting the importance of serum pCO₂ in surgical outcomes, Silva *et al.*^[21] showed a preoperative arteriovenous pCO₂ gap greater than 5.0 mmHg in high risk patients to be predictive of increased in-hospital mortality, circulatory shock, renal failure, intensive care unit (ICU)

infection, and length of stay. These previous studies suggest the usefulness of venous blood gas analysis in identifying hemorrhagic shock earlier than other serum markers obtained from arterial blood analysis as well as the potential to accurately monitor adequate resuscitative efforts.

The purpose of this study was to examine the effectiveness of venous blood gas analysis in comparison to the gold standard of arterial blood gas analysis in a rabbit model of hemorrhagic shock.

MATERIALS AND METHODS

Following approval by the Institutional Animal Care and Use Committee of the Carolinas Medical Center, fifty New Zealand white rabbits weighing 3 to 6 kg were anesthetized with 1.0 to 1.5 mL/kg of sodium pentobarbital (25 mg/mL) through an ear vein. Anesthesia was maintained throughout the experiment with 0.5-1.0 mL/kg of intravenous sodium pentobarbital (12.5 mg/mL) as needed, determined by response to a pain stimulus. Adequately anesthetized animals then underwent a tracheotomy and endotracheal ventilation. Tidal volumes of 10 mL/kg were administered by a mechanical ventilator (Siemens 900C Servo ventilator, Berlin, Germany) and fraction of inspired oxygen (FiO₂) was maintained at 0.5.

In all animals, bilateral groin dissection was performed to adequately expose femoral vasculature. Venous access was obtained *via* right femoral vein using a 5.0 French catheter advanced into the level of the right atrium and was utilized for drug infusion as well as withdrawal of venous blood samples. Arterial access was secured *via* left femoral artery utilizing a 3.5 French catheter advanced into the distal abdominal aorta for monitoring of blood pressure, heart rate and arterial blood sampling.

Following baseline arterial and venous blood gas measurements (Radiometer analyzer, ABL-520 #2, Copenhagen, Denmark), animals were hemorrhaged to a mean arterial pressure of 40 mmHg as determined by a multichannel recorder (MT95k2, Astro-Med, Inc., West Warwick, RI). Simultaneous arterial and venous blood gases were obtained every 3 min until hemorrhagic shock was observed, as defined by an arterial pH less than 7.2 and a base deficit greater than or equal to -15 mmol/L. Once the shock state was obtained, animals were then euthanized by intravenous administration of sodium pentobarbital. To minimize procedural variation, all animals were anesthetized, instrumented, hemorrhaged, and euthanized using identical technique by the same investigator.

Statistical analysis

Data was stored and analyzed using SAS software version 9.3 (SAS Inc., Cary, North Carolina). Obtained arterial and venous blood gases were compared to baseline measurements with regard to pH, base deficit and pCO₂. Arteriovenous differences for each parameter (pH, base deficit, pCO₂) were then calculated at baseline and shock. Statistical analysis was performed using the unpaired t-test

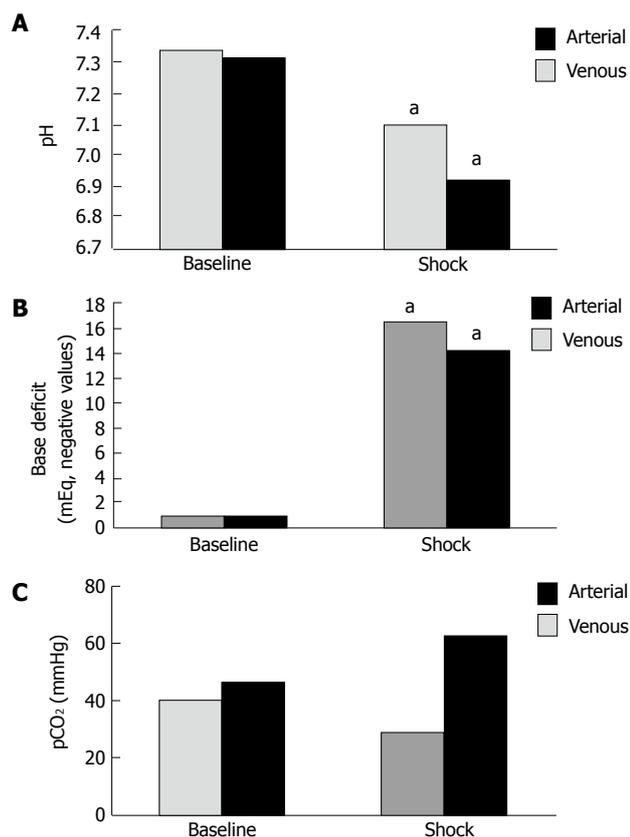


Figure 1 Arterial and venous blood gas at baseline and shock states. ^a $P < 0.05$ vs control group. A: pH; B: Base deficit; C: pCO₂.

or Wilcoxon rank sum test where appropriate. For all comparisons, statistical significance was set at a P value of less than 0.05.

RESULTSAll 50 animals underwent successful administration of anesthesia, groin dissection, instrumentation, hemorrhage to a state of shock and euthanization without any unexplained or premature losses. Data are expressed as mean \pm SD.

Arterial and venous blood gases at baseline and the hemorrhagic shock state

Mean values for pH were significantly decreased from baseline to shock ($P < 0.05$) in both arterial (7.39 ± 0.12 to 7.14 ± 0.18) and venous (7.35 ± 0.15 to 6.98 ± 0.26) blood gases (Figure 1A). Figure 1B compares mean values obtained for base deficit at the 2 physiologic states; a significant increase ($P < 0.05$) was seen in arterial (-0.9 ± 3.9 mEq/L vs -17.8 ± 2.2 mEq/L) and venous (-0.8 ± 3.8 mEq/L vs -15.3 ± 4.1 mEq/L) base deficit during shock. In comparing pCO₂ at baseline and shock, a non-significant decrease was observed in arterial pCO₂ (40.0 ± 9.1 mmHg vs 28.9 ± 7.1 mmHg, $P > 0.05$), while venous blood samples demonstrated a non-significant trend towards increased pCO₂ (46.0 ± 10.1 mmHg vs 62.8 ± 15.3 mmHg, $P > 0.05$), as shown in Figure 1C.

Blood gas arteriovenous differences at baseline and the hemorrhagic shock state

Arteriovenous differences in pH, base deficit, and pCO₂

at baseline and the hemorrhagic shock state are represented. No significant differences were seen in calculated differences at baseline for pH (0.04 ± 0.03), base deficit (0.01 ± 3.07 mEq/L), or pCO₂ (5.8 ± 7.5 mmHg), although venous pH demonstrated a larger non-significant trend toward acidosis and a larger non-significant base deficit was seen for arterial samples. In the shock state, a significant difference was noted for arteriovenous pCO₂ difference (34.0 ± 3.10 mmHg, $P < 0.05$), however, calculated differences for pH (0.16 ± 0.08) and base deficit (2.56 ± 3.10 mEq/L) were not significant.

DISCUSSION

Our results demonstrated significant parallel trends of acidosis and increased base deficit in both arterial and venous blood during hemorrhagic shock in a rabbit model (Figure 1A and B). The arteriovenous pCO₂ difference during shock was also statistically significant as venous hypercarbia was observed with simultaneous arterial hypocarbia (Figure 1C).

During hemorrhagic shock, oxygen delivery to tissues is reduced due to lack of red blood cell mass and, subsequently, hemoglobin concentration is insufficient to meet tissue oxygen demands^[22]. Contributing to the drop in oxygen carrying capacity, decreased cardiac output secondary to reduced venous return slows the delivery and elimination of venous CO₂ in the lungs and augments ongoing venous hypercarbia^[13,18,23]. Reduced oxygen delivery to tissues results in a shift from aerobic toward anaerobic cellular metabolism effecting subsequent production of organic acids, such as lactate, and ensuing acidosis and hypercarbia^[17]. When the oxygen supply can be restored quickly, metabolic function can return to normal; however, when the oxygen insufficiency is prolonged, cells become irreversibly damaged and are unable to function in normal energy metabolism^[24]. Serum and tissue acidosis develop in direct proportion to the amount and acuity of hemorrhagic shock^[2,3,6,14,25].

Studies in both animal models and humans have demonstrated a pronounced dissociation between the arterial and venous pCO₂ during periods of decreased oxygen delivery as a consequence of decreased cardiac output, such as cardiac tamponade^[15], severe hemorrhagic shock^[2,14], hemodynamic instability^[5,17,18,20] or septic shock^[16,19]. Carbon dioxide accumulates very rapidly during hemodynamic compromise, as with massive blood loss, before significant amounts of organic acids are detectable in blood since the normal liver is capable of upregulating lactate metabolism early in the hemorrhagic process^[15,21]. Mixed venous CO₂ (CvCO₂) can be represented according to the Fick equation, $CvCO_2 = VCO_2/Q + CaCO_2$, where VCO₂ represents CO₂ production in tissues, Q signifies cardiac output, and CaCO₂ denotes the arterial CO₂ content. Carbon dioxide is released into the circulation at the tissue-venous interface, represented by VCO₂/Q, and is eliminated at the alveolar-arteriole interface in the lungs, represented by CaCO₂. Under conditions of normal cardiac output and venous return,

there is adequate ventilatory elimination of CO₂ that is produced in the tissues and acid-base equilibrium is established. However, as cardiac output and venous return decrease, the increased CO₂ produced from anaerobic tissues cannot be effectively eliminated by the lungs, resulting in a disconnect between arterial and venous vascular trees whereby arterial blood gases reflect CO₂ exchange at the alveolar-arterial level while venous blood gases are indicative of acid-base status and oxygenation at the level of the tissues. Examining the Fick equation, it can be seen that as cardiac output (Q) declines with simultaneous increased tissue CO₂ production (VCO₂), mixed venous CO₂ (CvCO₂) will increase. This was demonstrated in our rabbit hemorrhage model in which the animals were adequately ventilated but hypoperfused, resulting in hypercarbia detected in venous but not arterial blood gas analysis, representing insufficient oxygen delivery to tissues. Therefore, venous blood gas values may better reflect insufficient oxygen delivery to the tissues and subsequent impending shock. Although our results did not show a statistically significant difference in the arteriovenous pH gradient, the venous samples were markedly more acidic than the arterial samples taken during the shock state (Figure 1A).

The clinical utility of arteriovenous pCO₂ differences in goal directed therapy (GDT) has recently been addressed in the literature. In a series of septic ICU patients resuscitated to a mixed venous oxygen saturation goal of 70% or greater, Vallée *et al*¹⁶¹ demonstrated those patients with arteriovenous pCO₂ differences greater than 6 mmHg had higher lactate concentrations and lower lactate clearance rates than those with arteriovenous pCO₂ differences less than 6 mmHg, subsequently reflecting the status of global tissue perfusion. They further demonstrated lower cardiac indexes in those patients with arteriovenous pCO₂ values greater than 6 mmHg following “adequate” resuscitation to a mixed venous oxygen saturation of 70% as compared to the cohort of patients with arteriovenous pCO₂ values less than 6 mmHg. Similarly, Futier *et al*²⁶¹ demonstrated larger arteriovenous pCO₂ differences to be significantly associated with post-operative complications in “adequately resuscitated” patients (to a mixed venous oxygen saturation goal greater than 71%) undergoing major abdominal surgery. These clinical results indicate further optimization of GDT may be obtained through a combination of mixed venous oxygenation and arteriovenous pCO₂ difference analysis, potentially playing vital roles in progressive damage control resuscitation models in trauma¹¹.

Some shortcomings of the current study deserve discussion. First, this study was not conducted in a spontaneously-breathing animal model, effectively eliminating the possibility of respiratory compensation which likely will occur in cases of acute injury and hemorrhage. To address this criticism, Mathias *et al*¹⁵¹ performed acid-base comparisons between arterial and venous blood gases in a spontaneously-breathing, under-anesthetized canine model of acute cardiac tamponade and found a

similar paradox of venous acidosis and hypercarbia with concomitant arterial alkalemia and hypocarbia, even in the early stages of decreased cardiac output (20%) prior to any reduction in arterial blood pressure. Although the criticisms of performing these studies in animal models with blunted or absent respiratory compensatory mechanisms are valid concerns, the results of Mathias *et al*¹⁵¹ indicate the paradoxical acid-base trends are still evident in a minimally-anesthetized, spontaneously-breathing, non-ventilated animal model. Also, performing these studies in live animal models without sufficient sedation or supportive measures, such as ventilatory support, would certainly be considered distressful to the animals. A second shortcoming of this study is the lack of a temporal metric for the onset of acid-base changes in our animal model, as well as a lack of serum lactate analysis. It would be beneficial to define the chronological relationship of arterial hypocapnea, venous hypercapnea and acidosis in comparison to the accumulation of lactate during the course of hemorrhagic shock in the rabbit model to better define the usefulness of venous blood gas in the course of hemorrhagic shock.

Our study in a rabbit model indicates hemorrhage shock results in significant acidosis and base deficit in both arterial and venous blood with a significant arteriovenous pCO₂ difference of venous hypercarbia and arterial hypocarbia, consistent with previously reported disparities between arterial and venous pCO₂ in the setting of severely hypoperfused states. These results indicate that venous blood gas analysis may be a superior indicator of cellular hypoperfusion in hemorrhagic shock, as evidenced by pronounced hypercarbia, and may be more reflective of tissue oxygenation compared to arterial blood gas analysis. Further studies are needed to determine if venous blood gas analysis is a more rapid indicator of impending circulatory collapse or is a more accurate gauge of adequate resuscitative efforts.

COMMENTS

Background

Early and adequate tissue perfusion is a key tenet of goal-directed therapy and damage control resuscitation, employed in critical care and trauma practices, respectively. Arteriovenous differences in pCO₂ have demonstrated potential in the early detection of insufficient tissue perfusion as well as the quantification of resuscitative efforts.

Research frontiers

Establishment of an early, reliable, and easily obtainable marker for impending circulatory collapse in hemorrhagic shock would contribute significantly to treatment algorithms, possibly allowing supportive measures (fluid resuscitation, blood product administration, vasopressor circulatory support, etc.) to be initiated prior to classic physiologic indicators of circulatory collapse. However, no such definitive marker has been elucidated. In this study, the authors demonstrate significant similarities and differences in arterial and venous blood gas derangements, focusing on the arteriovenous differences noted in a rabbit model of hemorrhagic shock in an effort to further define arteriovenous pCO₂ differences as a potential early indicator of inadequate tissue perfusion.

Innovations and breakthroughs

Previous studies have examined arterial and venous blood gas derangements (pH, base deficit, lactate levels, oxygen saturation) in states of hypoperfusion in animal models as well as humans. Paradoxical venous hypercarbia with arterial hypocarbia associated with decreased cardiac output has also been reported in

the literature, suggesting a role for pCO₂ monitoring in cases of hypoperfused states. In this study, authors conclusively demonstrated a widened pCO₂ difference (venous hypercarbia with concomitant arterial hypocarbia) is associated with hemorrhagic shock in a novel rabbit model.

Applications

The results of this study, viewed in light of recent work regarding venous blood gas analysis in hypoperfused states, further supports the prospect that central venous blood gas pCO₂ differences may indicate effectiveness of resuscitative efforts in the acutely injured hemorrhagic state. Certainly, further human studies in the setting of acute hemorrhage deserve attention so that a more rapid, accurate and easily obtainable mechanism of resuscitation may be elucidated.

Terminology

The term arteriovenous pCO₂ difference is used to describe the absolute value of the quantifiable variance between arterial blood gas pCO₂ and venous blood gas pCO₂. This is represented in units of mmHg, a standard unit of measurement for partial pressure.

Peer review

This is a well-written manuscript which analyzes the effects of hemorrhagic shock on arterial and venous blood gases in a rabbit animal model. An added caveat is the significant widened arteriovenous pCO₂ difference seen in the shock state. The manuscript also reviews pertinent publications on the subject, highlighting recent clinical studies which suggest a role for arteriovenous pCO₂ differences in monitoring resuscitation. Although not novel, the results certainly provide further evidence that widened pCO₂ differences are indicative of worsening shock and may therefore possibly be another tool in our armament to monitor resuscitation.

REFERENCES

- 1 **Cotton BA**, Reddy N, Hatch QM, LeFebvre E, Wade CE, Kozar RA, Gill BS, Albarado R, McNutt MK, Holcomb JB. Damage control resuscitation is associated with a reduction in resuscitation volumes and improvement in survival in 390 damage control laparotomy patients. *Ann Surg* 2011; **254**: 598-605 [PMID: 21918426 DOI: 10.1097/SLA.0b013e318230089e]
- 2 **Rixen D**, Siegel JH. Bench-to bedside review: oxygen debt and its metabolic correlates as quantifiers of the severity of hemorrhagic and post-traumatic shock. *Crit Care* 2005; **9**: 441-453 [PMID: 16277731]
- 3 **Rossaint R**, Bouillon B, Cerny V, Coats TJ, Duranteau J, Fernández-Mondéjar E, Hunt BJ, Komadina R, Nardi G, Neugebauer E, Ozier Y, Riddez L, Schultz A, Stahel PF, Vincent JL, Spahn DR. Management of bleeding following major trauma: an updated European guideline. *Crit Care* 2010; **14**: R52 [PMID: 20370902 DOI: 10.1186/cc8943]
- 4 **van Beest P**, Wietasch G, Scheeren T, Spronk P, Kuiper M. Clinical review: use of venous oxygen saturations as a goal - a yet unfinished puzzle. *Crit Care* 2011; **15**: 232 [PMID: 22047813 DOI: 10.1186/cc10351]
- 5 **Hobbs TR**, O'Malley JP, Khouangsathiene S, Dubay CJ. Comparison of lactate, base excess, bicarbonate, and pH as predictors of mortality after severe trauma in rhesus macaques (*Macaca mulatta*). *Comp Med* 2010; **60**: 233-239 [PMID: 20579439]
- 6 **Kaplan LJ**, Frangos S. Clinical review: Acid-base abnormalities in the intensive care unit -- part II. *Crit Care* 2005; **9**: 198-203 [PMID: 15774078 DOI: 10.1186/cc2912]
- 7 **Scheer B**, Perel A, Pfeiffer UJ. Clinical review: complications and risk factors of peripheral arterial catheters used for haemodynamic monitoring in anaesthesia and intensive care medicine. *Crit Care* 2002; **6**: 199-204 [PMID: 12133178]
- 8 **Malatesha G**, Singh NK, Bharija A, Rehani B, Goel A. Comparison of arterial and venous pH, bicarbonate, PCO₂ and PO₂ in initial emergency department assessment. *Emerg Med J* 2007; **24**: 569-571 [PMID: 17652681 DOI: 10.1136/emj.2007.046979]
- 9 **Rudkin SE**, Kahn CA, Oman JA, Dolich MO, Lotfipour S, Lush S, Gain M, Firme C, Anderson CL, Langdorf MI. Prospective correlation of arterial vs venous blood gas measurements in trauma patients. *Am J Emerg Med* 2012; **30**: 1371-1377 [PMID: 22169587 DOI: 10.1016/j.ajem.2011.09.027]
- 10 **Middleton P**, Kelly AM, Brown J, Robertson M. Agreement between arterial and central venous values for pH, bicarbonate, base excess, and lactate. *Emerg Med J* 2006; **23**: 622-624 [PMID: 16858095 DOI: 10.1136/emj.2006.035915]
- 11 **Malinoski DJ**, Todd SR, Slone S, Mullins RJ, Schreiber MA. Correlation of central venous and arterial blood gas measurements in mechanically ventilated trauma patients. *Arch Surg* 2005; **140**: 1122-1125 [PMID: 16342377]
- 12 **Dubin A**, Edul VS, Pozo MO, Murias G, Canullán CM, Martins EF, Ferrara G, Canales HS, Laporte M, Estensoro E, Ince C. Persistent villi hypoperfusion explains intramucosal acidosis in sheep endotoxemia. *Crit Care Med* 2008; **36**: 535-542 [PMID: 18216603 DOI: 10.1097/01.CCM.0000300083.74726.43]
- 13 **Vallet B**, Teboul JL, Cain S, Curtis S. Venous-arterial CO₂ difference during regional ischemic or hypoxic hypoxia. *J Appl Physiol* (1985) 2000; **89**: 1317-1321 [PMID: 11007564]
- 14 **Oropello JM**, Manasia A, Hannon E, Leibowitz A, Benjamin E. Continuous fiberoptic arterial and venous blood gas monitoring in hemorrhagic shock. *Chest* 1996; **109**: 1049-1055 [PMID: 8635330]
- 15 **Mathias DW**, Clifford PS, Klopfenstein HS. Mixed venous blood gases are superior to arterial blood gases in assessing acid-base status and oxygenation during acute cardiac tamponade in dogs. *J Clin Invest* 1988; **82**: 833-838 [PMID: 3417872 DOI: 10.1172/JCI113686]
- 16 **Vallée F**, Vallet B, Mathe O, Parraguet J, Mari A, Silva S, Samii K, Fourcade O, Genestal M. Central venous-to-arterial carbon dioxide difference: an additional target for goal-directed therapy in septic shock? *Intensive Care Med* 2008; **34**: 2218-2225 [PMID: 18607565 DOI: 10.1007/s00134-008-1199-0]
- 17 **Mekontso-Dessap A**, Castelain V, Anguel N, Bahloul M, Schauvliege F, Richard C, Teboul JL. Combination of venous-arterial PCO₂ difference with arteriovenous O₂ content difference to detect anaerobic metabolism in patients. *Intensive Care Med* 2002; **28**: 272-277 [PMID: 11904655 DOI: 10.1007/s00134-002-1215-8]
- 18 **Teboul JL**, Mercat A, Lenique F, Berton C, Richard C. Value of the venous-arterial PCO₂ gradient to reflect the oxygen supply to demand in humans: effects of dobutamine. *Crit Care Med* 1998; **26**: 1007-1010 [PMID: 9635647]
- 19 **Mecher CE**, Rackow EC, Astiz ME, Weil MH. Venous hypercarbia associated with severe sepsis and systemic hypoperfusion. *Crit Care Med* 1990; **18**: 585-589 [PMID: 2111753]
- 20 **Cuschieri J**, Rivers EP, Donnino MW, Katilios M, Jacobsen G, Nguyen HB, Pamukov N, Horst HM. Central venous-arterial carbon dioxide difference as an indicator of cardiac index. *Intensive Care Med* 2005; **31**: 818-822 [PMID: 15803301 DOI: 10.1007/s00134-005-2602-8]
- 21 **Silva JM**, Oliveira AM, Segura JL, Ribeiro MH, Sposito CN, Toledo DO, Rezende E, Malbouisson LM. A large Venous-Arterial PCO₂ Is Associated with Poor Outcomes in Surgical Patients. *Anesthesiol Res Pract* 2011; **2011**: 759792 [PMID: 22007204 DOI: 10.1155/2011/759792]
- 22 **Hsia CC**. Respiratory function of hemoglobin. *N Engl J Med* 1998; **338**: 239-247 [PMID: 9435331 DOI: 10.1056/NEJM199801223380407]
- 23 **Funk DJ**, Jacobsen E, Kumar A. Role of the venous return in critical illness and shock: part II-shock and mechanical ventilation. *Crit Care Med* 2013; **41**: 573-579 [PMID: 23263572 DOI: 10.1097/CCM.0b013e31827bfc25]
- 24 **Bruegger D**, Kemming GI, Jacob M, Meisner FG, Wojtczyk CJ, Packert KB, Keipert PE, Faithfull NS, Habler OP, Becker BF, Rehm M. Causes of metabolic acidosis in canine hemorrhagic shock: role of unmeasured ions. *Crit Care* 2007; **11**: R130 [PMID: 18081930 DOI: 10.1186/cc6200]
- 25 **Theusinger OM**, Thyges C, Frascarolo P, Schramm S, Seifert B, Spahn DR. Mismatch of arterial and central venous

Williams KB *et al.* Arterial vs venous blood gas differences

blood gas analysis during haemorrhage. *Eur J Anaesthesiol* 2010; **27**: 890-896 [PMID: 20601892 DOI: 10.1097/EJA.0b013e32833adea8]

26 **Futier E**, Robin E, Jabaudon M, Guerin R, Petit A, Bazin JE,

Constantin JM, Vallet B. Central venous O₂ saturation and venous-to-arterial CO₂ difference as complementary tools for goal-directed therapy during high-risk surgery. *Crit Care* 2010; **14**: R193 [PMID: 21034476 DOI: 10.1186/cc9310]

P- Reviewers: Chen XL, Mohta M, Nayci A **S- Editor:** Gou SX
L- Editor: A **E- Editor:** Wu HL



Variable change in renal function by hypertonic saline

Jesse J Corry, Panayiotis Varelas, Tamer Abdelhak, Stacey Morris, Marlisa Hawley, Allison Hawkins, Michelle Jankowski

Jesse J Corry, Panayiotis Varelas, Tamer Abdelhak, Stacey Morris, Marlisa Hawley, Allison Hawkins, Department of Neurology, Henry Ford Hospital, Marshfield, WI 54449, United States

Jesse J Corry, Panayiotis Varelas, Tamer Abdelhak, Department of Neurosurgery, Henry Ford Hospital, Marshfield, WI 54449, United States

Michelle Jankowski, Department of Biostatistics, Henry Ford Hospital, Marshfield, WI 54449, United States

Author contributions: Corry JJ contributed to study concept and design, data review, statistical analysis and review, manuscript authorship; Varelas P and Abdelhak T participated in data review, manuscript authorship and editing; Morris S, Hawley M and Hawkins A contributed to data collection, manuscript editing; Jankowski M contributed to statistical analysis, manuscript methods authorship and editing; all authors approved final revision.

Correspondence to: Jesse J Corry, MD, Department of Neurology, Henry Ford Hospital, Marshfield, 000 N. Oak Avenue, 4F3, Marshfield, WI 54449,

United States. corry.jesse@marshfieldclinic.org

Telephone: +1-715-079562 Fax: +1-715-3875727

Received: September 16, 2013 Revised: December 9, 2013

Accepted: January 13, 2014

Published online: May 4, 2014

Abstract

AIM: To investigate the effects of hypertonic saline in the neurocritical care population.

METHODS: We retrospectively reviewed our hospital's use of hypertonic saline (HS) since March of 2005, and prospectively since October 2010. Comparisons were made between admission diagnoses, creatinine change (Cr), and HS formulation (3% NaCl, 3% NaCl/sodium acetate mix, and 23.4% NaCl) to patients receiving normal saline or lactated ringers. The patients ($n = 1329$) of the retrospective portion were identified. The data presented represents the first 230 patients with data.

RESULTS: Significant differences in Acute Physiology and Chronic Health Evaluation II scores and Glasgow

Coma Scale scores occurred between different saline formulations. No significant correlation of Cl^- or Na^+ with Cr, nor with saline types, occurred. When dichotomized by diagnosis, significant correlations appear. Traumatic brain injury (TBI) patients demonstrated moderate correlation between Na^+ and Cr of 0.45. Stroke patients demonstrated weak correlations between Na^+ and Cr, and Cl^- and Cr (0.19 for both). Patients receiving HS and not diagnosed with intracerebral hemorrhage, stroke, subarachnoid hemorrhage, or TBI demonstrated a weak but significant correlation between Cl^- and Cr at 0.29.

CONCLUSION: Cr directly correlates with Na^+ or Cl^- in stroke, Na^+ in TBI, and Cl^- in other populations. Prospective comparison of HS and renal function is needed.

© 2014 Baishideng Publishing Group Co., Limited. All rights reserved.

Key words: Hypertonic saline solution; Sodium chloride; Acute kidney injury; Cerebral edema; Critical care

Core tip: This work adds to the literature that changes in Na^+ and Cl^- in the neurocritical care population correlate to adverse changes in renal function. It is critical for the neurointensivist to remain cognizant of this when choosing whether or not to use hypertonic saline, and what to monitor when doing so. Unlike previous work, this data suggests some diseases may have more or less a change in renal function from Na^+ or Cl^- . This argues for further study of how the formulations of these fluids may change outcome in the neurocritically ill.

Corry JJ, Varelas P, Abdelhak T, Morris S, Hawley M, Hawkins A, Jankowski M. Variable change in renal function by hypertonic saline. *World J Crit Care Med* 2014; 3(2): 61-67 Available from: URL: <http://www.wjgnet.com/2220-3141/full/v3/i2/61.htm> DOI: <http://dx.doi.org/10.5492/wjccm.v3.i2.61>

INTRODUCTION

A nearly ubiquitous problem in the neurocritical care population is cerebral edema (CE). Cerebral edema has been implicated in delayed neurological deterioration, and worse outcome, through the elevation of intracerebral pressure (ICP)^[1]. The role of CE in outcome is contentious, with evidence suggesting that the extent of CE may, and may not, correlate with outcome^[2-6]. Animal models of CE demonstrate increased water content of edematous tissue correlates with inflammation and neuronal death^[7]. Potentially, reduction of this edema may reduce the degree of neuronal death, potentially improving outcome and decreasing hospital length-of-stay.

The medical management of CE is not without problems. Mannitol use is common, but is complicated by deleterious effects on renal function, fluctuations in intravascular volume, and pH. Over time, mannitol's slow elimination from the cerebrospinal fluid may require progressively higher doses to control ICP and rebound CE^[8,9]. Increasingly, hypertonic saline (HS) is being used to abate cerebral edema. Used in bolus or a continuous infusion fashion, HS has been shown to be safe and effective in reducing ICP in patients with traumatic brain injury (TBI), subarachnoid hemorrhage (SAH), and stroke^[10,11]. HS shifts fluid from endothelium and surrounding tissues into the vascular compartment, normalizing the endothelial volume, increasing capillary diameter, and reducing resistance to flow^[12]. Edema can be reduced in this manner. Further, hypertonic fluids produce smooth muscle vasodilation improving regional blood flow^[12]. HS is relatively inexpensive. The early use of HS may reduce secondary cell injury caused by cerebral edema^[13]. These characteristics make HS an ideal therapeutic option in conditions such as SAH, intracerebral hemorrhage (ICH), stroke, and TBI.

However, the use of HS has not demonstrated any survival or outcome benefit despite reductions in ICP^[12,14]. Further, HS may be associated with increased risk-of blood-stream infections, and possibly increased risk-of nosocomial and urinary tract infections^[14]. A growing body of evidence suggests a possible link between HS use, renal dysfunction, and mortality^[15-17]. We hypothesize the use of HS correlates to adverse changes in renal function.

MATERIALS AND METHODS

Study setting

The Henry Ford Neurocritical Care Unit (NCCU) is a 16 bed unit with a yearly census over 1000 patients. The Henry Ford Neurocritical Database records data on stroke, TBI, ICH, SAH, SE, and spinal cord injury patient populations admitted to the NCCU. The data has been prospectively collected since October of 2010, with data added retrospectively from March of 2005 (when the first neurointensivist joined the staff) until October 2010. Between March 2005 and October 2010, 1329 patients of the retrospective cohort meet the inclusion criteria. The

data presented represents the first 230 patients with data.

Study design

With institutional review board approval, we mined the Henry Ford Neurocritical Database to identify all patients from March of 2005 to October 2010 with the aforementioned diagnosis who received HS. These patients were cross matched with the institution's pharmacy database to ascertain which saline formulation patients in the retrospective cohort received. In this retrospective sample, if patients were identified who received HS, and were not in the NCCU database, their data was retrospectively collected. Data was collected from admission until NCCU discharge, death, or post admission day (PAD) 13. Variables sought included: (1) IVF formulation: Normal saline (NS), ringer's lactate (LR), HS (3% NaCl, 3% NaCl:Na acetate, 23.4%); (2) physiologic: Mean arterial pressure; serum sodium, creatinine, chloride, HCO₃, BUN, creatinine; admission weight; (3) clinical: Admission Acute Physiology and Chronic Health Evaluation II (APACHE II) scores, admission Glasgow coma scale in all patients, Hunt and Hess grade in SAH patients, NIH stroke scale in stroke patients, presence of external ventricular drain, and duration of ICU stay. APACHE II scores were retrospectively calculated on all patients in the retrospective cohort; and (4) demographics: Age; sex; race; presence of hypertension, diabetes, pre-existing renal insufficiency and etiology of renal insufficiency, history of coronary artery disease or congestive heart failure. Patients received various formulations of HS at the discretion of the attending NCCU staff. Correlations to renal function, as measured by Cr, to the formulation of saline used, and to changes in serum sodium and chloride levels were made. Patients receiving only LR or NS served as a comparison group.

Statistical analysis

Intervariable associations were calculated between using Pearson's correlation coefficients. The *P* values for these correlation coefficients were computed using clustering methods that take into account the multiple measures from the same patient. This was done with the entire sample as well as within each saline type and within each diagnosis type.

RESULTS

Who received HS solutions?

Table 1 summarizes the baseline characteristics of this cohort. There were no significant differences in diagnosis between groups. There were significant differences in the APACHE II scores and Glasgow Coma Scale (GCS) scores between the different formulations of HS. Significant differences emerged in admission Na⁺, A-a gradient, APACHE II score, and GCS. In pairwise comparisons, patients receiving HS demonstrated higher APACHE II scores and lower GCSv scores. 3% of patients uniformly scored lower on GCS components compared to LR/NS patients, and lower in the GCSm and GCSe

Table 1 Baseline characteristics by saline type

Variable	23.40% (n = 22)	3% (n = 13)	NS/LR (n = 194)	P value
Diagnosis				0.078
ICH	15 (68)	7 (54)	67 (35)	
Other	3 (14)	3 (23)	47 (24)	
SAH	4 (18)	2 (15)	43 (22)	
Stroke	0 (0)	0 (0)	28 (14)	
TBI	0 (0)	1 (8)	9 (5)	
Age (yr)	61.9 ± 15.6	59.2 ± 19.6	56.8 ± 16.2	0.356
HCT (mmHg)	38.2 ± 6.9	38.1 ± 6.2	38.1 ± 7.0	0.999
WBC (10 ⁹ /L)	12.5 ± 6.2	11.6 ± 5.2	11.4 ± 9.1	0.865
Temperature (°C)	37.3 ± 0.9	37.2 ± 0.7	36.8 ± 2.5	0.514
HR	81.3 ± 20.3	86.5 ± 19.1	82.6 ± 16.7	0.668
RR	18.1 ± 4.6	17.5 ± 7.3	18.6 ± 4.4	0.633
MAP	98.3 ± 18.2	100.8 ± 33.2	106.5 ± 24.8	0.267
Na	141.5 ± 6.4	8.2 ± 5.3	139.0 ± 4.1	0.037
K	3.9 ± 0.6	3.7 ± 0.5	4.0 ± 2.0	0.888
Glasgow coma Scale (verbal)				0.010
1	10 (45)	6 (46)	53 (27)	
2	1 (5)	2 (15)	10 (5)	
3	2 (9)	1 (8)	15 (8)	
4	5 (23)	3 (23)	29 (15)	
5	4 (18)	1 (8)	87 (45)	
Glasgow Coma Scale (motor)	5.0 ± 1.5	3.9 ± 1.8	5.3 ± 1.3	0.002
1	1 (5)	1 (8)	5 (3)	
2	1 (5)	3 (23)	4 (2)	
3	2 (9)	2 (15)	16 (8)	
4	2 (9)	1 (8)	13 (7)	
5	4 (18)	2 (15)	27 (14)	
6	12 (55)	4 (31)	129 (66)	
Glasgow coma Scale (eyes)	2.9 ± 1.3	2.0 ± 1.0	3.3 ± 1.1	< 0.001
1	5 (23)	5 (38)	28 (14)	
2	4 (18)	4 (31)	15 (8)	
3	2 (9)	3 (23)	31 (16)	
4	11 (50)	1 (8)	120 (62)	
A aGrad ¹	6.1 ± 195.3	173.7 ± 141.2	125.3 ± 154.9	0.024
pH ²	7.4 ± 0.1	7.4 ± 0.1	7.4 ± 0.1	0.306
PCO ₂ ³	27.0 ± 1.7	27.0 ± 3.0	26.0 ± 3.3	0.756
PaO ₂	209.2 ± 114.6	189.5 ± 136.8	150.8 ± 115.1	0.053
Baseline Cr	1.3 ± 1.4	0.9 ± 0.3	1.4 ± 1.8	0.655
APACHE II	14.9 ± 7.6	17.5 ± 5.2	10.7 ± 6.5	< 0.001

¹23.4%, n = 19; 3%, n = 12; normal saline/ringer's lactate (NS/LR), n = 177; ²23.4%, n = 19; 3%, n = 10; NS/LR, n = 111; ³23.4%, n = 3; 3%, n = 3; NS/LR, n = 83. ICH: Intracerebral hemorrhage; TBI: Traumatic brain injury; SAH: Subarachnoid hemorrhage; HCT: Hematocrit; MAP: Mean arterial pressure; APACHE II: Admission acute physiology and chronic health evaluation II scores.

compared to 23.4% patients (Table 2). This would suggest patients receiving HS, particularly 3% solutions, had a greater illness burden at admission.

What are the effects of HS on renal function?

Table 3 summarizes the effects of HS on renal function in this cohort. No significant correlation occurred with Na⁺ or Cl⁻ with Cr when grouped according to saline type. The correlation between Na⁺ and Cr within each of the saline types does not differ much from the overall correlation, except for within the 3% saline group. The correlation for the 3% saline group is 0.256 but this was not statistically significantly different from zero (P = 0.26).

Table 2 Pairwise comparisons for those that were significant

Dependent	23.4% vs 3%	23.4% vs NS/LR	3% vs NS/LR
Na ⁺	0.035	0.016	0.503
A aGrad	0.370	0.009	0.306
Apache II	0.261	0.005	< 0.001
GCS v	0.579	0.034	0.019
GCS m	0.027	0.294	< 0.001
GCS E	0.028	0.123	< 0.001

NS/LR: Normal saline/ringer's lactate; APACHE II: Admission acute physiology and chronic health evaluation II scores; GCS: Glasgow coma scale.

Table 3 Correlations between Cl⁻, Na⁺ and Cr

Population	Na ⁺ and Cr			Cl ⁻ and Cr		
	n	Corr	P value	n	Corr	P value
Overall	230	0.025	0.63	229	0.074	0.16
Saline type						
23.40%	22	0.037	0.58	22	0.042	0.65
3%	13	0.256	0.26	13	0.181	0.43
NS/LR	194	0.026	0.65	194	0.097	0.09
Diagnosis type						
ICH	89	0.145	0.10	89	0.058	0.55
Other	53	0.096	0.09	53	0.287	< 0.001
SAH	50	0.125	0.08	50	0.085	0.28
Stroke	28	0.187	< 0.001	28	0.185	0.001
TBI	10	0.447	0.048	10	0.361	0.1

NS/LR: Normal saline/ringer's lactate; ICH: Intracerebral hemorrhage; TBI: Traumatic brain injury; SAH: Subarachnoid hemorrhage.

The same holds true for the correlation between Cl⁻ and Cr, it is greater in the 3% saline group but still not statistically significantly different from zero (r = 0.18, P = 0.43). When the correlations were dichotomized by the diagnosis, significant findings appear.

The strongest correlations were found in TBI, patients given HS and not diagnosed with TBI, stroke, SAH, or ICH (other), and stroke. With respect to TBI, a moderate correlation was found between rise in Cr and Na⁺. For stroke, weak correlations between rise in Cr and both increases in Na⁺ and Cl⁻ occurred. Patients in this "other" category demonstrated a significant, yet weak, correlation between increases in Cl⁻ and Cr.

DISCUSSION

Even small increases in creatinine the first two days following admission are predictive of mortality^[16,18]. Thus, therapies precipitating kidney injury are concerning. Presentation or development of diminished renal function is a predictor of poor outcome and mortality in stroke, ICH and SAH^[19-25]. In the case of ICH, this has been associated with hemorrhage volume and GCS^[21,22,26]. Similarly, development of renal dysfunction during hospitalization has been linked to increased mortality and is associated with lower GCS and higher APACHE III score in TBI^[27,28].

Frequently neurointensivists are asked to control cerebral edema *via* the use of mannitol and HS. Superi-

ority of one agent remains a matter of debate. Not yet recruiting at the time of this manuscript, investigators at Massachusetts General Hospital are investigating if induced, sustained hyponatremia to a goal of 150-160 mmol/L following traumatic brain injury will decrease the rate of cerebral edema formation and improve patient outcomes^[29]. One study, sponsored by Indiana University, currently enrolling is looking at 20% mannitol *vs* 3% saline for the treatment of intracranial hypertension^[30].

Mannitol appears to reduce ICP through reducing brain water content^[31]. However, its use may result in kidney injury and rebound edema^[8,9]. Increasingly HS saline is being used in various formulations either as a preventative or acute therapy^[12,32]. HS appears to have a number of beneficial effects. In TBI, the use of 23.4% NaCl results in ICP reductions and elevations in cerebral perfusion pressure with commensurate elevations in brain tissue oxygenation^[33,34]. These reductions in ICP are most notable in patients with the greatest elevations in ICP. Further, in cerebral hemorrhages ≥ 30 mL, the early use of HS to target a serum sodium between 145-155 mmol/L demonstrates both absolute and relative reductions of cerebral edema when compared to normonatremic patients^[32].

These effects appear to be the result of a combination of actions including reduction in brain water content *via* osmotic forces, reductions in peripheral vascular resistance, and arteriolar vasodilatation with improvement in capillary blood flow^[12,13,35]. Further, animal models treated with HS have demonstrated reduced aquaporin 4 expression on astrocytes with attenuation of brain water content^[13]. In addition, increasing evidence demonstrates HS possesses immune-modulating properties *via* reduction in cytokine production and neutrophil activation^[36,37]. Finally, animal models have demonstrated reductions in neuronal apoptosis.

Despite the ample experimental evidence, the clinical use of HS has not demonstrated any survival or outcome benefit and is not without risk^[12,14]. Hyponatremia is associated with insulin resistance, reduced hepatic gluconeogenesis and lactate clearance, delirium, rhabdomyolysis, and reduced cardiac function^[38,39]. Not surprisingly, ICU acquired elevations or reductions in serum sodium, dysnatremia, are common in neurosurgical and trauma patients, and associated with kidney injury^[40]. Further, when compared to normonatremic patients, dysnatremia is associated with increased disease severity, longer length-of-stay, and mortality^[40-42].

Our group inquired whether the formulation of saline used affected renal function in the neurocritical care population. The question of IVF formulation affecting patient outcome has been debated for some time. Evidence is increasingly suggesting formulation of saline and/or rapid change in serum sodium or chloride may adversely affect renal function^[15-17]. A study in healthy subjects has demonstrated greater natriuresis and sooner time to first post-bolus micturition in those receiving LR *vs* NS^[43]. Huang *et al*^[16] reported the use of HS in burn patients produced significant increases renal, pulmonary,

and cardiac failure compared to LR use. Patients receiving HS had less urine output. Further, the development of renal failure was heralded by a greater initial rise and slower subsequent fall in serum sodium levels during the first week of admission. Although this study was in burn patients, its findings are provocative. More recently, a study evaluating the effects of a Cl⁻ restrictive *vs* Cl⁻ liberal usage in critical ill patients demonstrated more acute kidney injury and greater use of renal-replacement therapy in the Cl⁻ liberal group^[44]. Though the populations of these studies differ from the neurocritical care population, this suggests the formulation of saline may have an effect on renal function.

With respect to our initial question, does saline type affect renal function; we found no such correlation in our sample. We found patients receiving HS have higher disease severity as assessed by lower GCS and higher APACHE II scores. Not surprisingly, we found sicker patients more frequently received HS in this sample. This correlation has been previously reported^[15]. This makes intuitive sense, with evidence suggesting early use of HS may limit the development of CE^[32]. Similar to the findings of Aiyagari *et al*^[15], Froelich *et al*^[17] reported adverse changes in renal function with serum Na⁺ > 155 mmol/L. This was not associated with the use or formulation of HS, a finding noted in our study too.

Unexpectedly, when we dichotomized by diagnosis, we found weak to moderate correlations between admitting disease and changes in Cr associated with hyperchloremia or hyponatremia. This was most noted in TBI, stroke, and non-vascular NCCU diagnosis; trends were also noted in ICH and SAH too. The explanation for this association is uncertain. Previous studies demonstrate correlations to Na⁺ increase and renal dysfunction^[15,17]. Although patients receiving a continuous HS infusion, when compared to a cohort receiving NS, do not have a higher risk of renal dysfunction, a significant correlation between severe hyponatremia and renal dysfunction does exist^[17]. This could however reflect a more severe underlying brain injury rather than effect of HS.

Both clinical and experimental literature provides insight as to how Na⁺ and Cl⁻ could adversely affect renal function. HS solutions initially cause renal vasodilatation and increased renal blood flow^[45]. It is theorized hyponatremia may produce renal injury *via* intravascular dehydration and vasoconstriction^[46]. Canine models undergoing rapid renal artery sodium elevations demonstrate reduced renal blood flow and glomerular filtration rate with inhibition of rennin secretion^[47]. Clinical studies have demonstrated hyponatremia is associated with elevations in creatinine in approximately 10% of patients^[15]. This noted increase parallels elevations in sodium and APACHE II scores, and is inversely related to admission GCS scores. However, save for sodium values > 160 mEq/L hyponatremia is not independently associated with mortality^[15,17].

While direct proof linking saline-induced hyperchloremia to nephrotoxicity is not available, a strong cir-

cumstantial case can be made^[48]. NS, with 154 mmol/L of chlorine can result in hyperchloremia and an acidosis^[43,49]. Elevation in chloride can reduce renal blood flow and decreases the excretion of sodium^[43,45]. Hyperchloremia appears to cause a renal vasoconstriction specific to renal vasculature and independent of the renal nerve^[45]. This reduction in renal blood flow could precipitate renal ischemia and reducing glomerular filtration rate^[45,50]. At the macula densa, Cl⁻ activates tubuloglomerular feedback by precipitating afferent arteriolar vasoconstriction and decreased glomerular filtration rate^[51]. Further, animal models suggest Cl⁻ increases thromboxane synthesis resulting in renal vasoconstriction and reduced renal blood flow^[52].

This single center, retrospective study has a number of limitations. First is the choice of serum creatinine as a biomarker of renal function. Though regularly used to infer kidney health and glomerular filtration, it is at best a crude measure of these. Often creatinine may be insensitive to early, deleterious changes in renal function. Next, given the time and cost of collecting retrospective data, this data represents an interim analysis to see if continued collection of these variables was warranted. As such, its small size and single center nature limit its applicability. Other centers with different demographics or practices may have different outcomes from what is represented here. The retrospective design and single center location limits what questions can be asked, data obtained, and the number of patients available. Regarding the disease specific correlations, a number of deficiencies exist. Regarding TBI, this study did not collect data on vasopressor use, blood pressure targets, or volume received, all variables noted to augment renal blood flow^[53]. Intense sympathetic stimulation alters prostaglandin-mediated vasodilatation, resulting in reduced glomerular filtration^[54]. Data on antecedent medication use was not collected. Could prior use of medications such as angiotensin converting enzyme inhibitors, in the setting of rapid changes in serum Na⁺ and Cl⁻, result in diminished renal blood flow? Finally, after dichotomizing by diagnosis, differences in baseline physiologic variables was not assessed. Perhaps these correlations occurred in patients who were inherently more ill when viewed from the perspective of admitting diagnosis. Despite these limitations, this data is provocative in suggesting the admitting disease may affect the physiologic response to a therapy.

This study adds to the literature demonstrating the use of HS is not inherently injurious to renal physiology. Further, we too note the correlation of injury severity to HS use. Finally, our data suggests when viewed from the perspective of admitting diagnosis, HS use may correlate to the development of kidney injury. However, the nature of this correlation needs further exploration. Variables to investigate include rate change of Na⁺ and Cl⁻; HS administration times over the course of disease; role of pre-morbid medications; and regional differences in population makeup. Wide variability exists in the treatment of cerebral edema among intensivists^[55]. With no clear “right answer” to the question of cerebral edema,

more investigation is needed regarding the risks/benefits of the treatments available and the patients who would be best suited for particular therapies. Prospective comparisons of HS formulation and renal function are needed to further assess if formulation affects outcome and cost. Prospective studies are warranted to better define this association and its effect on outcome.

COMMENTS

Background

The treatment and management of cerebral edema is among the duties of a neurointensivist. When and how to treat cerebral edema remain contentious. Further, a neurointensivist must remain cognizant of how their neurocentric therapies may affect the rest of the patient's body.

Research frontiers

Intravenous fluids and hypertonic saline are ubiquitous in the critical care and neurocritical care setting. Data has previously demonstrated “not all fluids are created equal.” Understanding how the formulation of intravenous fluids may affect outcome is critical to providing effective critical care. Discovery of deleterious correlations may help generate prospective, hypothesis driven, studies on patient or disease specific intravenous fluids aimed at improving outcome.

Innovations and breakthroughs

Prior work has demonstrated that the formulation of hypertonic saline (HS) may not affect renal function. However, the relative change of Na⁺, and presence in particular of hypernatremia, may correlate with development of kidney injury and worse outcomes. Much of this work was in a mixed critical care or mixed neurocritical care population. This study assessed if not only Na⁺, but if Cl⁻, HS formulation, and disease state played a role. The data here presented suggests potential rolls of Cl⁻ and disease state to adverse renal function. These findings need to be confirmed by larger, prospective trials. Potentially, such findings could form the basis for developing patient or disease specific intravenous fluids aimed at reducing cerebral edema and mitigating adverse renal effects. Further, if borne out in future studies, better understanding of what interactions occur between intravenous fluids, disease state, and comorbidities may allow for development of new therapeutic options in neurocritical care.

Applications

Data presented here, and in the context of literature to date, may suggest to the bedside clinician to be judicious with the prescription of HS to patients with cerebral edema, to closely monitor renal function, and use Cl⁻ limiting formulations of HS.

Terminology

Cerebral edema is the process whereby injured brain develops increase free water by cytotoxic or vasogenic means. Typically, these two pathologies combine in a temporal fashion. Much of the overall change of brain volume is related to this, a concern in the rigid volume provided within the skull. Potentially, cerebral edema may exacerbate inflammation. Hypertonic saline, or HS, are intravenous fluids of higher osmolarity aimed at increasing serum sodium. This has a multitude of effects including: (1) reducing brain free water and edema; (2) reducing aquaporin production and thus water entry into cells preventing/limiting the development of cerebral edema; (3) improving red blood cells malleability and ability to travel through injured tissue; and (4) potential mitigating effects on inflammation. Creatinine, Cr, is a biomarker of kidney health. Though crude, this is a readily available biomarker that can guide the clinicians management of a patient.

Peer review

This analysis provides some provocative findings that need a larger study to confirm. Further, it summarizes much of the literature on this topic to date.

REFERENCES

- 1 **Mayer SA**, Sacco RL, Shi T, Mohr JP. Neurologic deterioration in noncomatose patients with supratentorial intracerebral hemorrhage. *Neurology* 1994; **44**: 1379-1384 [PMID: 8058133]
- 2 **Kollmar R**, Staykov D, Dörfler A, Schellinger PD, Schwab S, Bardutzky J. Hypothermia reduces perihemorrhagic edema after intracerebral hemorrhage. *Stroke* 2010; **41**: 1684-1689

- [PMID: 20616317 DOI: 10.1161/STROKEAHA.110.587758]
- 3 **Ropper AH.** Lateral displacement of the brain and level of consciousness in patients with an acute hemispherical mass. *N Engl J Med* 1986; **314**: 953-958 [PMID: 3960059]
 - 4 **Ropper AH, Shafran B.** Brain edema after stroke. Clinical syndrome and intracranial pressure. *Arch Neurol* 1984; **41**: 26-29 [PMID: 6606414]
 - 5 **Zazulia AR, Diringner MN, Derdeyn CP, Powers WJ.** Progression of mass effect after intracerebral hemorrhage. *Stroke* 1999; **30**: 1167-1173 [PMID: 10356094]
 - 6 **Arima H, Wang JG, Huang Y, Heeley E, Skulina C, Parsons MW, Peng B, Li Q, Su S, Tao QL, Li YC, Jiang JD, Tai LW, Zhang JL, Xu E, Cheng Y, Morgenstern LB, Chalmers J, Anderson CS.** Significance of perihematomal edema in acute intracerebral hemorrhage: the INTERACT trial. *Neurology* 2009; **73**: 1963-1968 [PMID: 19996072 DOI: 10.1212/WNL.0b013e3181c55ed3]
 - 7 **Thiex R, Tsirka SE.** Brain edema after intracerebral hemorrhage: mechanisms, treatment options, management strategies, and operative indications. *Neurosurg Focus* 2007; **22**: E6 [PMID: 17613237]
 - 8 **Nau R, Desel H, Lassek C, Thiel A, Schinschke S, Rössing R, Kolenda H, Prange HW.** Slow elimination of mannitol from human cerebrospinal fluid. *Eur J Clin Pharmacol* 1997; **53**: 271-274 [PMID: 9476044]
 - 9 **McGraw CP, Howard G.** Effect of mannitol on increased intracranial pressure. *Neurosurgery* 1983; **13**: 269-271 [PMID: 6413884]
 - 10 **Vialet R, Albanèse J, Thomachot L, Antonini F, Bourgoign A, Alliez B, Martin C.** Isovolemic hypertonic solutes (sodium chloride or mannitol) in the treatment of refractory post-traumatic intracranial hypertension: 2 mL/kg 7.5% saline is more effective than 2 mL/kg 20% mannitol. *Crit Care Med* 2003; **31**: 1683-1687 [PMID: 12794404]
 - 11 **Suarez JL, Qureshi AI, Bhardwaj A, Williams MA, Schnitzer MS, Mirski M, Hanley DF, Ulatowski JA.** Treatment of refractory intracranial hypertension with 23.4% saline. *Crit Care Med* 1998; **26**: 1118-1122 [PMID: 9635664]
 - 12 **Strandvik GF.** Hypertonic saline in critical care: a review of the literature and guidelines for use in hypotensive states and raised intracranial pressure. *Anaesthesia* 2009; **64**: 990-1003 [PMID: 19686485 DOI: 10.1111/j.1365-044.2009.05986.x]
 - 13 **Zeng HK, Wang QS, Deng YY, Fang M, Chen CB, Fu YH, Jiang WQ, Jiang X.** Hypertonic saline ameliorates cerebral edema through downregulation of aquaporin-4 expression in the astrocytes. *Neuroscience* 2010; **166**: 878-885 [PMID: 20083168 DOI: 10.1016/j.neuroscience.2009.12.076]
 - 14 **Bulger EM, May S, Brasel KJ, Schreiber M, Kerby JD, Tisherman SA, Newgard C, Slutsky A, Coimbra R, Emerson S, Minei JP, Bardarson B, Kudenchuk P, Baker A, Christenson J, Idris A, Davis D, Fabian TC, Aufderheide TP, Callaway C, Williams C, Banek J, Vaillancourt C, van Heest R, Sopko G, Hata JS, Hoyt DB.** Out-of-hospital hypertonic resuscitation following severe traumatic brain injury: a randomized controlled trial. *JAMA* 2010; **304**: 1455-1464 [PMID: 20924011 DOI: 10.1001/jama.2010.1405]
 - 15 **Aiyagari V, Deibert E, Diringner MN.** Hyponatremia in the neurologic intensive care unit: how high is too high? *J Crit Care* 2006; **21**: 163-172 [PMID: 16769461]
 - 16 **Huang PP, Stucky FS, Dimick AR, Treat RC, Bessey PQ, Rue LW.** Hypertonic sodium resuscitation is associated with renal failure and death. *Ann Surg* 1995; **221**: 543-554; discussion 554-557 [PMID: 7748036]
 - 17 **Froelich M, Ni Q, Wess C, Ougorets I, Härtl R.** Continuous hypertonic saline therapy and the occurrence of complications in neurocritically ill patients. *Crit Care Med* 2009; **37**: 1433-1441 [PMID: 19242317 DOI: 10.1097/CCM.0b013e31819c1933]
 - 18 **Nin N, Lombardi R, Frutos-Vivar F, Esteban A, Lorente JA, Ferguson ND, Hurtado J, Apezteguia C, Brochard L, Schortgen F, Raymondos K, Tomcic V, Soto L, González M, Nightingale P, Abroug F, Pelosi P, Arabi Y, Moreno R, Anzueto A.** Early and small changes in serum creatinine concentrations are associated with mortality in mechanically ventilated patients. *Shock* 2010; **34**: 109-116 [PMID: 20634655 DOI: 10.1097/SHK.0b013e3181d671a6]
 - 19 **Yahalom G, Schwartz R, Schwammenthal Y, Merzeliak O, Toashi M, Orion D, Sela BA, Tanne D.** Chronic kidney disease and clinical outcome in patients with acute stroke. *Stroke* 2009; **40**: 1296-1303 [PMID: 19182072 DOI: 10.1161/STROKEAHA.108.520882]
 - 20 **Putala J, Haapaniemi E, Gordin D, Liebkind R, Groop PH, Kaste M, Tatlisumak T.** Factors associated with impaired kidney function and its impact on long-term outcome in young ischemic stroke. *Stroke* 2011; **42**: 2459-2464 [PMID: 21737795 DOI: 10.1161/STROKEAHA.110.612721]
 - 21 **Molshatzki N, Orion D, Tsabari R, Schwammenthal Y, Merzeliak O, Toashi M, Tanne D.** Chronic kidney disease in patients with acute intracerebral hemorrhage: association with large hematoma volume and poor outcome. *Cerebrovasc Dis* 2011; **31**: 271-277 [PMID: 21178352 DOI: 10.1159/000322155]
 - 22 **Rhoney DH, Parker D, Millis SR, Whittaker P.** Kidney dysfunction at the time of intracerebral hemorrhage is associated with increased in-hospital mortality: a retrospective observational cohort study. *Neurol Res* 2012; **34**: 518-521 [PMID: 22664363 DOI: 10.1179/1743132812Y.0000000041]
 - 23 **Lin LC, Yang JT, Weng HH, Hsiao CT, Lai SL, Fann WC.** Predictors of early clinical deterioration after acute ischemic stroke. *Am J Emerg Med* 2011; **29**: 577-581 [PMID: 20825831 DOI: 10.1016/j.ajem.2009.12.019]
 - 24 **Hao Z, Wu B, Lin S, Kong FY, Tao WD, Wang DR, Liu M.** Association between renal function and clinical outcome in patients with acute stroke. *Eur Neurol* 2010; **63**: 237-242 [PMID: 20332640 DOI: 10.1159/000285165]
 - 25 **Zacharia BE, Ducruet AF, Hickman ZL, Grobelny BT, Fernandez L, Schmidt JM, Narula R, Ko LN, Cohen ME, Mayer SA, Connolly ES.** Renal dysfunction as an independent predictor of outcome after aneurysmal subarachnoid hemorrhage: a single-center cohort study. *Stroke* 2009; **40**: 2375-2381 [PMID: 19461033 DOI: 10.1161/STROKEAHA.108.545210]
 - 26 **Misra UK, Kalita J, Srivastava M, Mandal SK.** Transient renal impairment in acute intracerebral haemorrhage. *J Neurol* 1996; **243**: 417-420 [PMID: 8741083]
 - 27 **Li N, Zhao WG, Zhang WF.** Acute kidney injury in patients with severe traumatic brain injury: implementation of the acute kidney injury network stage system. *Neurocrit Care* 2011; **14**: 377-381 [PMID: 21298359 DOI: 10.1007/s12028-011-9511-1]
 - 28 **Moore EM, Bellomo R, Nichol A, Harley N, Macisaac C, Cooper DJ.** The incidence of acute kidney injury in patients with traumatic brain injury. *Ren Fail* 2010; **32**: 1060-1065 [PMID: 20863210 DOI: 10.3109/0886022X.2010.510234] [http://www.clinicaltrials.gov/ct2/show/study/NCT01605357?term=hypertonic saline&rank=72](http://www.clinicaltrials.gov/ct2/show/study/NCT01605357?term=hypertonic%20saline&rank=72)
 - 29 [http://www.clinicaltrials.gov/ct2/show/study/NCT01215019?term=hypertonic saline&rank=84](http://www.clinicaltrials.gov/ct2/show/study/NCT01215019?term=hypertonic%20saline&rank=84)
 - 30 **Diringner MN, Scalfani MT, Zazulia AR, Videen TO, Dhar R, Powers WJ.** Effect of mannitol on cerebral blood volume in patients with head injury. *Neurosurgery* 2012; **70**: 1215-1218; discussion 1219 [PMID: 22089753 DOI: 10.1227/NEU.0b013e3182417bc2]
 - 31 **Wagner I, Hauer EM, Staykov D, Volbers B, Dörfler A, Schwab S, Bardutzky J.** Effects of continuous hypertonic saline infusion on perihemorrhagic edema evolution. *Stroke* 2011; **42**: 1540-1545 [PMID: 21512173 DOI: 10.1161/STROKEAHA.110.609479]
 - 32 **Rockswold GL, Solid CA, Paredes-Andrade E, Rockswold SB, Jancik JT, Quickel RR.** Hypertonic saline and its effect on intracranial pressure, cerebral perfusion pressure,

- and brain tissue oxygen. *Neurosurgery* 2009; **65**: 1035-1041; discussion 1041-1042 [PMID: 19934962 DOI: 10.1227/01.NEU.0000359533.16214.04]
- 34 **Al-Rawi PG**, Tseng MY, Richards HK, Nortje J, Timofeev I, Matta BF, Hutchinson PJ, Kirkpatrick PJ. Hypertonic saline in patients with poor-grade subarachnoid hemorrhage improves cerebral blood flow, brain tissue oxygen, and pH. *Stroke* 2010; **41**: 122-128 [PMID: 19910550 DOI: 10.1161/STROKEAHA.109.560698]
- 35 **Nout YS**, Mihai G, Tovar CA, Schmalbrock P, Bresnahan JC, Beattie MS. Hypertonic saline attenuates cord swelling and edema in experimental spinal cord injury: a study utilizing magnetic resonance imaging. *Crit Care Med* 2009; **37**: 2160-2166 [PMID: 19487936 DOI: 10.1097/CCM.0b013e3181a05d41]
- 36 **Rizoli SB**, Rhind SG, Shek PN, Inaba K, Filips D, Tien H, Brenneman F, Rotstein O. The immunomodulatory effects of hypertonic saline resuscitation in patients sustaining traumatic hemorrhagic shock: a randomized, controlled, double-blinded trial. *Ann Surg* 2006; **243**: 47-57 [PMID: 16371736]
- 37 **Hatanaka E**, Shimomi FM, Curi R, Campa A. Sodium chloride inhibits cytokine production by lipopolysaccharide-stimulated human neutrophils and mononuclear cells. *Shock* 2007; **27**: 32-35 [PMID: 17172977]
- 38 **Ellis SJ**. Severe hyponatraemia: complications and treatment. *QJM* 1995; **88**: 905-909 [PMID: 8593551]
- 39 **Lenz K**, Gössinger H, Laggner A, Druml W, Grimm G, Schneeweiss B. Influence of hypernatremic-hyperosmolar state on hemodynamics of patients with normal and depressed myocardial function. *Crit Care Med* 1986; **14**: 913-914 [PMID: 3757536]
- 40 **Sakr Y**, Rother S, Ferreira AM, Ewald C, Dünisch P, Riedemann N, Reinhart K. Fluctuations in serum sodium level are associated with an increased risk of death in surgical ICU patients. *Crit Care Med* 2013; **41**: 133-142 [PMID: 23128383 DOI: 10.1097/CCM.0b013e318265f576]
- 41 **Waite MD**, Fuhrman SA, Badawi O, Zuckerman IH, Franey CS. Intensive care unit-acquired hypernatremia is an independent predictor of increased mortality and length of stay. *J Crit Care* 2013; **28**: 405-412 [PMID: 23369520]
- 42 **Maggiore U**, Picetti E, Antonucci E, Parenti E, Regolisti G, Mergoni M, Vezzani A, Cabassi A, Fiaccadori E. The relation between the incidence of hypernatremia and mortality in patients with severe traumatic brain injury. *Crit Care* 2009; **13**: R110 [PMID: 19583864 DOI: 10.1186/cc7953]
- 43 **Reid F**, Lobo DN, Williams RN, Rowlands BJ, Allison SP. (Ab)normal saline and physiological Hartmann's solution: a randomized double-blind crossover study. *Clin Sci (Lond)* 2003; **104**: 17-24 [PMID: 12519083]
- 44 **Yunos NM**, Bellomo R, Hegarty C, Story D, Ho L, Bailey M. Association between a chloride-liberal vs chloride-restrictive intravenous fluid administration strategy and kidney injury in critically ill adults. *JAMA* 2012; **308**: 1566-1572 [PMID: 23073953 DOI: 10.1001/jama.2012.13356]
- 45 **Wilcox CS**. Regulation of renal blood flow by plasma chloride. *J Clin Invest* 1983; **71**: 726-735 [PMID: 6826732]
- 46 **Pérez-Pérez AJ**, Pazos B, Sobrado J, Gonzalez L, Gándara A. Acute renal failure following massive mannitol infusion. *Am J Nephrol* 2002; **22**: 573-575 [PMID: 12381962]
- 47 **Gerber JG**, Branch RA, Nies AS, Hollifield JW, Gerkens JF. Influence of hypertonic saline on canine renal blood flow and renin release. *Am J Physiol* 1979; **237**: F441-F446 [PMID: 517657]
- 48 **Prowle JR**, Bellomo R. Fluid administration and the kidney. *Curr Opin Crit Care* 2010; **16**: 332-336 [PMID: 20543683 DOI: 10.1097/MCC.0b013e32833be90b]
- 49 **Scheingraber S**, Rehm M, Sehmisch C, Finsterer U. Rapid saline infusion produces hyperchloremic acidosis in patients undergoing gynecologic surgery. *Anesthesiology* 1999; **90**: 1265-1270 [PMID: 10319771]
- 50 **Gazitúa S**, Scott JB, Chou CC, Haddy FJ. Effect of osmolarity on canine renal vascular resistance. *Am J Physiol* 1969; **217**: 1216-1223 [PMID: 5824323]
- 51 **Salomonsson M**, Gonzalez E, Kornfeld M, Persson AE. The cytosolic chloride concentration in macula densa and cortical thick ascending limb cells. *Acta Physiol Scand* 1993; **147**: 305-313 [PMID: 8386427]
- 52 **Bullivant EM**, Wilcox CS, Welch WJ. Intrarenal vasoconstriction during hyperchloremia: role of thromboxane. *Am J Physiol* 1989; **256**: F152-F157 [PMID: 2912160]
- 53 **Udy A**, Boots R, Senthuran S, Stuart J, Deans R, Lassig-Smith M, Lipman J. Augmented creatinine clearance in traumatic brain injury. *Anesth Analg* 2010; **111**: 1505-1510 [PMID: 21048095 DOI: 10.1213/ANE.0b013e3181f7107d]
- 54 **Pelayo JC**. Renal adrenergic effector mechanisms: glomerular sites for prostaglandin interaction. *Am J Physiol* 1988; **254**: F184-F190 [PMID: 3125747]
- 55 **Hays AN**, Lazaridis C, Neyens R, Nicholas J, Gay S, Chalela JA. Osmotherapy: use among neurointensivists. *Neurocrit Care* 2011; **14**: 222-228 [PMID: 21153930 DOI: 10.1007/s12028-010-9477-4]

P- Reviewers: Duan SB, Makki N, Mitaka C **S- Editor:** Zhai HH
L- Editor: A **E- Editor:** Wu HL



GENERAL INFORMATION

World Journal of Critical Care Medicine (*World J Crit Care Med*, *WJCCM*, online ISSN 2220-3141, DOI: 10.5492) is a peer-reviewed open access (OA) academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

Aim and scope

WJCCM covers topics concerning severe infection, shock and multiple organ dysfunction syndrome, infection and anti-infection treatment, acute respiratory distress syndrome and mechanical ventilation, acute kidney failure, continuous renal replacement therapy, rational nutrition and immunomodulation in critically ill patients, sedation and analgesia, cardiopulmonary cerebral resuscitation, fluid resuscitation and tissue perfusion, coagulant dysfunction, hemodynamic monitoring and circulatory support, ICU management and treatment control, and application of bronchofiberscopy in critically ill patients. The current columns of *WJCCM* include editorial, frontier, diagnostic advances, therapeutics advances, field of vision, mini-reviews, review, topic highlight, medical ethics, original articles, case report, clinical case conference (Clinicopathological conference), and autobiography.

We encourage authors to submit their manuscripts to *WJCCM*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

WJCCM is edited and published by Baishideng Publishing Group (BPG). BPG has a strong professional editorial team composed of science editors, language editors and electronic editors. BPG currently publishes 42 OA clinical medical journals, including 41 in English, has a total of 15 471 editorial board members or peer reviewers, and is a world first-class publisher.

Columns

The columns in the issues of *WJCCM* will include: (1) Editorial: The editorial board members are invited to make comments on an important topic in their field in terms of its current research status and future directions to lead the development of this discipline; (2) Frontier: The editorial board members are invited to select a highly cited cutting-edge original paper of his/her own to summarize major findings, the problems that have been resolved and remain to be resolved, and future research directions to help readers understand his/her important academic point of view and future research directions in the field; (3) Diagnostic Advances: The editorial board members are invited to write high-quality diagnostic advances in their field to improve the diagnostic skills of readers. The topic covers general clinical diagnosis, differential diagnosis, pathological diagnosis, laboratory diagnosis, imaging diagnosis, endoscopic diagnosis, biotechnological diagnosis, functional diagnosis, and physical diagnosis; (4) Therapeutics Advances: The editorial board members are invited to write high-quality therapeutic advances in their field to help improve the therapeutic skills of readers. The topic covers medication therapy, psychotherapy, physical therapy, replacement therapy, interventional therapy, minimally invasive therapy, endoscopic therapy, transplantation therapy, and surgical therapy; (5) Field of Vision: The editorial board members are invited to write commentaries on classic articles, hot topic articles, or latest articles to keep readers at the forefront of research and increase their levels of clinical research. Classic articles refer to papers that are included

in Web of Knowledge and have received a large number of citations (ranking in the top 1%) after being published for more than years, reflecting the quality and impact of papers. Hot topic articles refer to papers that are included in Web of Knowledge and have received a large number of citations after being published for no more than 2 years, reflecting cutting-edge trends in scientific research. Latest articles refer to the latest published high-quality papers that are included in PubMed, reflecting the latest research trends. These commentary articles should focus on the status quo of research, the most important research topics, the problems that have now been resolved and remain to be resolved, and future research directions. Basic information about the article to be commented (including authors, article title, journal name, year, volume, and inclusive page numbers); (6) Minireviews: The editorial board members are invited to write short reviews on recent advances and trends in research of molecular biology, genomics, and related cutting-edge technologies to provide readers with the latest knowledge and help improve their diagnostic and therapeutic skills; (7) Review: To make a systematic review to focus on the status quo of research, the most important research topics, the problems that have now been resolved and remain to be resolved, and future research directions; (8) Topic Highlight: The editorial board members are invited to write a series of articles (7-10 articles) to comment and discuss a hot topic to help improve the diagnostic and therapeutic skills of readers; (9) Medical Ethics: The editorial board members are invited to write articles about medical ethics to increase readers' knowledge of medical ethics. The topic covers international ethics guidelines, animal studies, clinical trials, organ transplantation, etc.; (10) Clinical Case Conference or Clinicopathological Conference: The editorial board members are invited to contribute high-quality clinical case conference; (11) Original Articles: To report innovative and original findings in critical care medicine; (12) Brief Articles: To briefly report the novel and innovative findings in critical care medicine; (13) Meta-Analysis: Covers the systematic review, mixed-treatment comparison, meta-regression, and overview of reviews, in order to summarize a given quantitative effect, e.g., the clinical effectiveness and safety of clinical treatments by combining data from two or more randomized controlled trials, thereby providing more precise and externally valid estimates than those which would stem from each individual dataset if analyzed separately from the others; (14) Case Report: To report a rare or typical case; (15) Letters to the Editor: To discuss and make reply to the contributions published in *WJCCM*, or to introduce and comment on a controversial issue of general interest; (16) Book Reviews: To introduce and comment on quality monographs of critical care medicine; and (17) Autobiography: The editorial board members are invited to write their autobiography to provide readers with stories of success or failure in their scientific research career. The topic covers their basic personal information and information about when they started doing research work, where and how they did research work, what they have achieved, and their lessons from success or failure.

Name of journal

World Journal of Critical Care Medicine

ISSN

ISSN 2220-3141 (online)

Launch date

February 4, 2012

Instructions to authors

Frequency

Quarterly

Editor-in-Chief

Yaseen Mohamed Arabi, MD, FCCP, FCCM, Associate Professor, Chairman, Intensive Care Department, King Saud Bin Abdulaziz University, Medical Director, Respiratory Services, King Abdulaziz Medical City, National Guard Hospital, Riyadh, PO Box 22490 Riyadh 11426, Saudi Arabia

Editorial office

Jin-Lei Wang, Director

Xiu-Xia Song, Vice Director

World Journal of Critical Care Medicine

Room 903, Building D, Ocean International Center,

No. 62 Dongsihuan Zhonglu, Chaoyang District,

Beijing 100025, China

Telephone: +86-10-85381891

Fax: +86-10-85381893

E-mail: bpgoffice@wjgnet.com

<http://www.wjgnet.com>

Publisher

Baishideng Publishing Group Co., Limited

Flat C, 23/F, Lucky Plaza, 315-321 Lockhart Road,

Wan Chai, Hong Kong, China

Telephone: +852-58042046

Fax: +852-31158812

E-mail: bpgoffice@wjgnet.com

<http://www.wjgnet.com>

Production center

Beijing Baishideng BioMed Scientific Co., Limited

Room 903, Building D, Ocean International Center,

No. 62 Dongsihuan Zhonglu, Chaoyang District,

Beijing 100025, China

Telephone: +86-10-85381892

Fax: +86-10-85381893

Representative office

USA Office

8226 Regency Drive,

Pleasanton, CA 94588-3144, United States

Instructions to authors

Full instructions are available online at http://www.wjgnet.com/2220-3141/g_info_20100722180909.htm.

Indexed and Abstracted in

Digital Object Identifier.

SPECIAL STATEMENT

All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

Biostatistical editing

Statistical review is performed after peer review. We invite an expert in Biomedical Statistics to evaluate the statistical method used in the paper, including *t*-test (group or paired comparisons), chi-squared test, Ridit, probit, logit, regression (linear, curvilinear, or stepwise), correlation, analysis of variance, analysis of covariance, *etc.* The reviewing points include: (1) Statistical methods should be described when they are used to verify the results; (2) Whether the statistical techniques are suitable or correct; (3) Only homogeneous data can be averaged. Standard deviations are preferred to standard errors. Give the number of observations and subjects (*n*). Losses in observations, such as drop-outs from the study should be reported; (4) Values such as ED50, LD50, IC50 should have their 95% confidence limits calculated and compared by weighted probit

analysis (Bliss and Finney); and (5) The word 'significantly' should be replaced by its synonyms (if it indicates extent) or the *P* value (if it indicates statistical significance).

Conflict-of-interest statement

In the interests of transparency and to help reviewers assess any potential bias, *WJCCM* requires authors of all papers to declare any competing commercial, personal, political, intellectual, or religious interests in relation to the submitted work. Referees are also asked to indicate any potential conflict they might have reviewing a particular paper. Before submitting, authors are suggested to read "Uniform Requirements for Manuscripts Submitted to Biomedical Journals: Ethical Considerations in the Conduct and Reporting of Research: Conflicts of Interest" from International Committee of Medical Journal Editors (ICMJE), which is available at: http://www.icmje.org/ethical_4conflicts.html.

Sample wording: [Name of individual] has received fees for serving as a speaker, a consultant and an advisory board member for [names of organizations], and has received research funding from [names of organization]. [Name of individual] is an employee of [name of organization]. [Name of individual] owns stocks and shares in [name of organization]. [Name of individual] owns patent [patent identification and brief description].

Statement of informed consent

Manuscripts should contain a statement to the effect that all human studies have been reviewed by the appropriate ethics committee or it should be stated clearly in the text that all persons gave their informed consent prior to their inclusion in the study. Details that might disclose the identity of the subjects under study should be omitted. Authors should also draw attention to the Code of Ethics of the World Medical Association (Declaration of Helsinki, 1964, as revised in 2004).

Statement of human and animal rights

When reporting the results from experiments, authors should follow the highest standards and the trial should conform to Good Clinical Practice (for example, US Food and Drug Administration Good Clinical Practice in FDA-Regulated Clinical Trials; UK Medicines Research Council Guidelines for Good Clinical Practice in Clinical Trials) and/or the World Medical Association Declaration of Helsinki. Generally, we suggest authors follow the lead investigator's national standard. If doubt exists whether the research was conducted in accordance with the above standards, the authors must explain the rationale for their approach and demonstrate that the institutional review body explicitly approved the doubtful aspects of the study.

Before submitting, authors should make their study approved by the relevant research ethics committee or institutional review board. If human participants were involved, manuscripts must be accompanied by a statement that the experiments were undertaken with the understanding and appropriate informed consent of each. Any personal item or information will not be published without explicit consents from the involved patients. If experimental animals were used, the materials and methods (experimental procedures) section must clearly indicate that appropriate measures were taken to minimize pain or discomfort, and details of animal care should be provided.

SUBMISSION OF MANUSCRIPTS

Manuscripts should be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Number all pages consecutively, and start each of the following sections on a new page: Title Page, Abstract, Introduction, Materials and Methods, Results, Discussion, Acknowledgements, References, Tables, Figures, and Figure Legends. Neither the editors nor the publisher are responsible for the opinions expressed by contributors. Manuscripts formally accepted for publication become the permanent property of Baishideng Publishing Group Co., Limited, and may not be reproduced by any means, in whole or in part, without the written permission of both the authors and the publisher. We reserve the right to copyedit and put onto our website accepted manuscripts. Authors should follow the relevant guidelines for the care and use of laboratory ani-

mals of their institution or national animal welfare committee. For the sake of transparency in regard to the performance and reporting of clinical trials, we endorse the policy of the ICMJE to refuse to publish papers on clinical trial results if the trial was not recorded in a publicly-accessible registry at its outset. The only register now available, to our knowledge, is <http://www.clinicaltrials.gov> sponsored by the United States National Library of Medicine and we encourage all potential contributors to register with it. However, in the case that other registers become available you will be duly notified. A letter of recommendation from each author's organization should be provided with the contributed article to ensure the privacy and secrecy of research is protected.

Authors should retain one copy of the text, tables, photographs and illustrations because rejected manuscripts will not be returned to the author(s) and the editors will not be responsible for loss or damage to photographs and illustrations sustained during mailing.

Online submissions

Manuscripts should be submitted through the Online Submission System at: <http://www.wjgnet.com/esps/>. Authors are highly recommended to consult the ONLINE INSTRUCTIONS TO AUTHORS (http://www.wjgnet.com/2220-3141/g_info_20100722180909.htm) before attempting to submit online. For assistance, authors encountering problems with the Online Submission System may send an email describing the problem to wjccm@wjgnet.com, or by telephone: +86-10-85381892. If you submit your manuscript online, do not make a postal contribution. Repeated online submission for the same manuscript is strictly prohibited.

MANUSCRIPT PREPARATION

All contributions should be written in English. All articles must be submitted using word-processing software. All submissions must be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Style should conform to our house format. Required information for each of the manuscript sections is as follows:

Title page

Title: Title should be less than 12 words.

Running title: A short running title of less than 6 words should be provided.

Authorship: Authorship credit should be in accordance with the standard proposed by ICMJE, based on (1) substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; (2) drafting the article or revising it critically for important intellectual content; and (3) final approval of the version to be published. Authors should meet conditions 1, 2, and 3.

Institution: Author names should be given first, then the complete name of institution, city, province and postcode. For example, Xu-Chen Zhang, Li-Xin Mei, Department of Pathology, Chengde Medical College, Chengde 067000, Hebei Province, China. One author may be represented from two institutions, for example, George Sgourakis, Department of General, Visceral, and Transplantation Surgery, Essen 45122, Germany; George Sgourakis, 2nd Surgical Department, Korgialenio-Benakio Red Cross Hospital, Athens 15451, Greece

Author contributions: The format of this section should be: Author contributions: Wang CL and Liang L contributed equally to this work; Wang CL, Liang L, Fu JF, Zou CC, Hong F and Wu XM designed the research; Wang CL, Zou CC, Hong F and Wu XM performed the research; Xue JZ and Lu JR contributed new reagents/analytic tools; Wang CL, Liang L and Fu JF analyzed the data; and Wang CL, Liang L and Fu JF wrote the paper.

Supportive foundations: The complete name and number of supportive foundations should be provided, e.g., Supported by National

Natural Science Foundation of China, No. 30224801

Correspondence to: Only one corresponding address should be provided. Author names should be given first, then author title, affiliation, the complete name of institution, city, postcode, province, country, and email. All the letters in the email should be in lower case. A space interval should be inserted between country name and email address. For example, Montgomery Bissell, MD, Professor of Medicine, Chief, Liver Center, Gastroenterology Division, University of California, Box 0538, San Francisco, CA 94143, United States. montgomerybissell@ucsf.edu

Telephone and fax: Telephone and fax should consist of +, country number, district number and telephone or fax number, e.g., Telephone: +86-10-85381892 Fax: +86-10-85381893

Peer reviewers: All articles received are subject to peer review. Normally, three experts are invited for each article. Decision on acceptance is made only when at least two experts recommend publication of an article. All peer-reviewers are acknowledged on Express Submission and Peer-review System website.

Abstract

There are unstructured abstracts (no less than 200 words) and structured abstracts. The specific requirements for structured abstracts are as follows:

An informative, structured abstract should accompany each manuscript. Abstracts of original contributions should be structured into the following sections: AIM (no more than 20 words; Only the purpose of the study should be included. Please write the Aim in the form of "To investigate/study/..."), METHODS (no less than 140 words for Original Articles; and no less than 80 words for Brief Articles), RESULTS (no less than 150 words for Original Articles and no less than 120 words for Brief Articles; You should present *P* values where appropriate and must provide relevant data to illustrate how they were obtained, e.g., 6.92 ± 3.86 vs 3.61 ± 1.67 , $P < 0.001$), and CONCLUSION (no more than 26 words).

Key words

Please list 5-10 key words, selected mainly from *Index Medicus*, which reflect the content of the study.

Core tip

Please write a summary of less than 100 words to outline the most innovative and important arguments and core contents in your paper to attract readers.

Text

For articles of these sections, original articles and brief articles, the main text should be structured into the following sections: INTRODUCTION, MATERIALS AND METHODS, RESULTS and DISCUSSION, and should include appropriate Figures and Tables. Data should be presented in the main text or in Figures and Tables, but not in both.

Illustrations

Figures should be numbered as 1, 2, 3, etc., and mentioned clearly in the main text. Provide a brief title for each figure on a separate page. Detailed legends should not be provided under the figures. This part should be added into the text where the figures are applicable. Keeping all elements compiled is necessary in line-art image. Scale bars should be used rather than magnification factors, with the length of the bar defined in the legend rather than on the bar itself. File names should identify the figure and panel. Avoid layering type directly over shaded or textured areas. Please use uniform legends for the same subjects. For example: Figure 1 Pathological changes in atrophic gastritis after treatment. A: ...; B: ...; C: ...; D: ...; E: ...; F: ...; G: ... etc. It is our principle to publish high resolution-figures for the E-versions.

Tables

Three-line tables should be numbered 1, 2, 3, etc., and mentioned

Instructions to authors

clearly in the main text. Provide a brief title for each table. Detailed legends should not be included under tables, but rather added into the text where applicable. The information should complement, but not duplicate the text. Use one horizontal line under the title, a second under column heads, and a third below the Table, above any footnotes. Vertical and italic lines should be omitted.

Notes in tables and illustrations

Data that are not statistically significant should not be noted. ^a*P* < 0.05, ^b*P* < 0.01 should be noted (*P* > 0.05 should not be noted). If there are other series of *P* values, ^c*P* < 0.05 and ^d*P* < 0.01 are used. A third series of *P* values can be expressed as ^e*P* < 0.05 and ^f*P* < 0.01. Other notes in tables or under illustrations should be expressed as ¹F, ²F, ³F; or sometimes as other symbols with a superscript (Arabic numerals) in the upper left corner. In a multi-curve illustration, each curve should be labeled with ●, ○, ■, □, ▲, △, etc., in a certain sequence.

Acknowledgments

Brief acknowledgments of persons who have made genuine contributions to the manuscript and who endorse the data and conclusions should be included. Authors are responsible for obtaining written permission to use any copyrighted text and/or illustrations.

REFERENCES

Coding system

The author should number the references in Arabic numerals according to the citation order in the text. Put reference numbers in square brackets in superscript at the end of citation content or after the cited author's name. For citation content which is part of the narration, the coding number and square brackets should be typeset normally. For example, "Crohn's disease (CD) is associated with increased intestinal permeability^[1,2]". If references are cited directly in the text, they should be put together within the text, for example, "From references^[19,22-24], we know that..."

When the authors write the references, please ensure that the order in text is the same as in the references section, and also ensure the spelling accuracy of the first author's name. Do not list the same citation twice.

PMID and DOI

Please provide PubMed citation numbers to the reference list, e.g., PMID and DOI, which can be found at <http://www.ncbi.nlm.nih.gov/sites/entrez?db=pubmed> and <http://www.crossref.org/SimpleTextQuery/>, respectively. The numbers will be used in E-version of this journal.

Style for journal references

Authors: the name of the first author should be typed in bold-faced letters. The family name of all authors should be typed with the initial letter capitalized, followed by their abbreviated first and middle initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR). The title of the cited article and italicized journal title (journal title should be in its abbreviated form as shown in PubMed), publication date, volume number (in black), start page, and end page [PMID: 11819634 DOI: 10.3748/wjg.13.5396].

Style for book references

Authors: the name of the first author should be typed in bold-faced letters. The surname of all authors should be typed with the initial letter capitalized, followed by their abbreviated middle and first initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR) Book title. Publication number. Publication place: Publication press, Year: start page and end page.

Format

Journals

English journal article (list all authors and include the PMID where applicable)

- 1 **Jung EM**, Clevert DA, Schreyer AG, Schmitt S, Rennert J, Kubale R, Feuerbach S, Jung F. Evaluation of quantitative con-

trast harmonic imaging to assess malignancy of liver tumors: A prospective controlled two-center study. *World J Gastroenterol* 2007; **13**: 6356-6364 [PMID: 18081224 DOI: 10.3748/wjg.13.6356]

Chinese journal article (list all authors and include the PMID where applicable)

- 2 **Lin GZ**, Wang XZ, Wang P, Lin J, Yang FD. Immunologic effect of Jianpi Yishen decoction in treatment of Pixu-diarhoea. *Shijie Huaren Xiaobua Zazhi* 1999; **7**: 285-287

In press

- 3 **Tian D**, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. *Proc Natl Acad Sci USA* 2006; In press

Organization as author

- 4 **Diabetes Prevention Program Research Group**. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; **40**: 679-686 [PMID: 12411462 PMID:2516377 DOI:10.1161/01.HYP.0000035706.28494.09]

Both personal authors and an organization as author

- 5 **Vallancien G**, Emberton M, Harving N, van Moorselaar RJ; Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; **169**: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

- 6 21st century heart solution may have a sting in the tail. *BMJ* 2002; **325**: 184 [PMID: 12142303 DOI:10.1136/bmj.325.7357.184]

Volume with supplement

- 7 **Geraud G**, Spierings EL, Keywood C. Tolerability and safety of frovatriptan with short- and long-term use for treatment of migraine and in comparison with sumatriptan. *Headache* 2002; **42** Suppl 2: S93-99 [PMID: 12028325 DOI:10.1046/j.1526-4610.42.s2.7.x]

Issue with no volume

- 8 **Banit DM**, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (**401**): 230-238 [PMID: 12151900 DOI:10.1097/00003086-200208000-00026]

No volume or issue

- 9 Outreach: Bringing HIV-positive individuals into care. *HRS A Careaction* 2002; 1-6 [PMID: 12154804]

Books

Personal author(s)

- 10 **Sherlock S**, Dooley J. Diseases of the liver and biliary system. 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296

Chapter in a book (list all authors)

- 11 **Lam SK**. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450

Author(s) and editor(s)

- 12 **Breedlove GK**, Schorfheide AM. Adolescent pregnancy. 2nd ed. Wiczorek RR, editor. White Plains (NY): March of Dimes Education Services, 2001: 20-34

Conference proceedings

- 13 **Harnden P**, Joffe JK, Jones WG, editors. Germ cell tumours V. Proceedings of the 5th Germ cell tumours Conference; 2001 Sep 13-15; Leeds, UK. New York: Springer, 2002: 30-56

Conference paper

- 14 **Christensen S**, Oppacher F. An analysis of Koza's computational effort statistic for genetic programming. In: Foster JA, Lutton E, Miller J, Ryan C, Tettamanzi AG, editors. Genetic programming. EuroGP 2002: Proceedings of the 5th European Conference on Genetic Programming; 2002 Apr 3-5; Kinsdale, Ireland. Berlin: Springer, 2002: 182-191

Electronic journal (list all authors)

- 15 Morse SS. Factors in the emergence of infectious diseases. *Emerg Infect Dis* serial online, 1995-01-03, cited 1996-06-05; 1(1): 24 screens. Available from: URL: <http://www.cdc.gov/ncidod/eid/index.htm>

Patent (list all authors)

16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1

Statistical data

Write as mean \pm SD or mean \pm SE.

Statistical expression

Express *t* test as *t* (in italics), *F* test as *F* (in italics), chi square test as χ^2 (in Greek), related coefficient as *r* (in italics), degree of freedom as ν (in Greek), sample number as *n* (in italics), and probability as *P* (in italics).

Units

Use SI units. For example: body mass, *m* (B) = 78 kg; blood pressure, *p* (B) = 16.2/12.3 kPa; incubation time, *t* (incubation) = 96 h; blood glucose concentration, *c* (glucose) 6.4 ± 2.1 mmol/L; blood CEA mass concentration, *p* (CEA) = 8.6 24.5 $\mu\text{g/L}$; CO₂ volume fraction, 50 mL/L CO₂, not 5% CO₂; likewise for 40 g/L formaldehyde, not 10% formalin; and mass fraction, 8 ng/g, *etc.* Arabic numerals such as 23, 243, 641 should be read 23 243 641.

The format for how to accurately write common units and quantum numbers can be found at: http://www.wjgnet.com/2220-3141/g_info_20100725073806.htm.

Abbreviations

Standard abbreviations should be defined in the abstract and on first mention in the text. In general, terms should not be abbreviated unless they are used repeatedly and the abbreviation is helpful to the reader. Permissible abbreviations are listed in Units, Symbols and Abbreviations: A Guide for Biological and Medical Editors and Authors (Ed. Baron DN, 1988) published by The Royal Society of Medicine, London. Certain commonly used abbreviations, such as DNA, RNA, HIV, LD50, PCR, HBV, ECG, WBC, RBC, CT, ESR, CSF, IgG, ELISA, PBS, ATP, EDTA, mAb, can be used directly without further explanation.

Italics

Quantities: *t* time or temperature, *c* concentration, *A* area, *l* length, *m* mass, *V* volume.

Genotypes: *gyrA*, *arg 1*, *c myc*, *c fos*, *etc.*

Restriction enzymes: *EcoRI*, *HindI*, *BamHI*, *Kho I*, *Kpn I*, *etc.*

Biology: *H. pylori*, *E. coli*, *etc.*

Examples for paper writing

All types of articles' writing style and requirement will be found in the link: <http://www.wjgnet.com/esps/NavigationInfo.aspx?id=15>

RESUBMISSION OF THE REVISED MANUSCRIPTS

Authors must revise their manuscript carefully according to the

revision policies of Baishideng Publishing Group Co., Limited. The revised version, along with the signed copyright transfer agreement, responses to the reviewers, and English language Grade A certificate (for non-native speakers of English), should be submitted to the online system *via* the link contained in the e-mail sent by the editor. If you have any questions about the revision, please send e-mail to esps@wjgnet.com.

Language evaluation

The language of a manuscript will be graded before it is sent for revision. (1) Grade A: priority publishing; (2) Grade B: minor language polishing; (3) Grade C: a great deal of language polishing needed; and (4) Grade D: rejected. Revised articles should reach Grade A.

Copyright assignment form

Please download a Copyright assignment form from http://www.wjgnet.com/2220-3141/g_info_20100725073726.htm.

Responses to reviewers

Please revise your article according to the comments/suggestions provided by the reviewers. The format for responses to the reviewers' comments can be found at: http://www.wjgnet.com/2220-3141/g_info_20100725073445.htm.

Proof of financial support

For papers supported by a foundation, authors should provide a copy of the approval document and serial number of the foundation.

STATEMENT ABOUT ANONYMOUS PUBLICATION OF THE PEER REVIEWERS' COMMENTS

In order to increase the quality of peer review, push authors to carefully revise their manuscripts based on the peer reviewers' comments, and promote academic interactions among peer reviewers, authors and readers, we decide to anonymously publish the reviewers' comments and author's responses at the same time the manuscript is published online.

PUBLICATION FEE

WJCCM is an international, peer-reviewed, OA online journal. Articles published by this journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium and format, provided the original work is properly cited. The use is non-commercial and is otherwise in compliance with the license. Authors of accepted articles must pay a publication fee. Publication fee: 698 USD per article. All invited articles are published free of charge.



百世登

Baishideng®

Published by **Baishideng Publishing Group Co., Limited**

Flat C, 23/F., Lucky Plaza, 315-321 Lockhart Road,

Wan Chai, Hong Kong, China

Fax: +852-31158812

Telephone: +852-58042046

E-mail: bpgoffice@wjgnet.com

<http://www.wjgnet.com>

