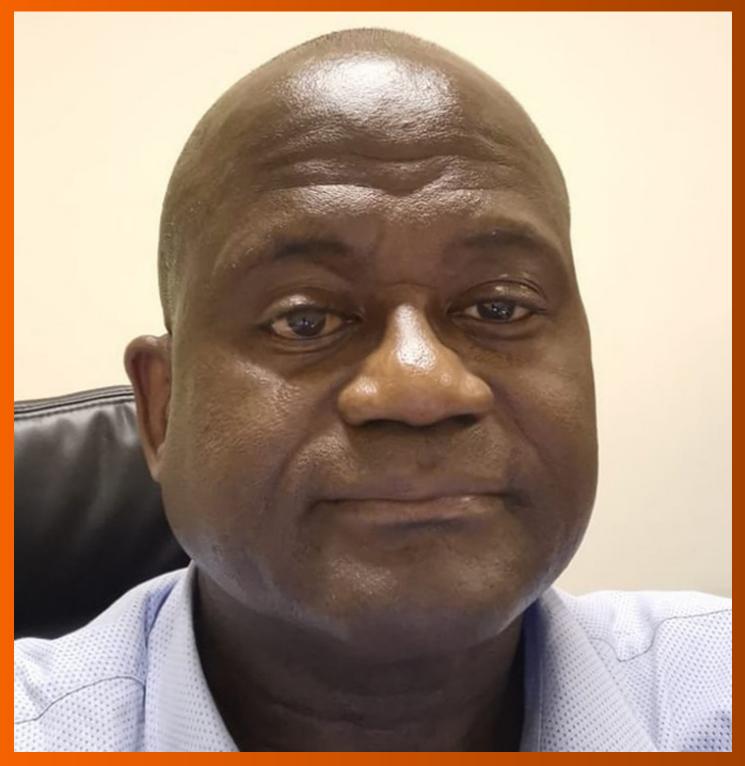
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Peer reviewer of World Journal of Virology, Zivanai Cuthbert Chapanduka, MBChB, Assistant Professor, Chairman, Chief Physician, Department of Pathology, Stellenbosch University Faculty of Medicine and Health Sciences, Cape Town 7550, South Africa. zivanai@sun.ac.za

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REVIEW

Intestinal barrier dysfunction as a key driver of severe COVID-19

Efthymios P Tsounis, Christos Triantos, Christos Konstantakis, Markos Marangos, Stelios F Assimakopoulos

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Efthymios P Tsounis, Christos Triantos, Christos Konstantakis, Division of Gastroenterology, Department of Internal Medicine, Medical School, University Hospital of Patras, Patras 26504, Greece

Markos Marangos, Stelios F Assimakopoulos, Division of Infectious Diseases, Department of Internal Medicine, Medical School, University of Patras, University Hospital of Patras, Patras 26504, Greece

Corresponding author: Stelios F Assimakopoulos, MD, PhD, Associate Professor, Division of Infectious Diseases, Department of Internal Medicine, Medical School, University of Patras, University Hospital of Patras, Rion, Patras 26504, Greece. sassim@upatras.gr

Abstract

The intestinal lumen harbors a diverse consortium of microorganisms that participate in reciprocal crosstalk with intestinal immune cells and with epithelial and endothelial cells, forming a multi-layered barrier that enables the efficient absorption of nutrients without an excessive influx of pathogens. Despite being a lung-centered disease, severe coronavirus disease 2019 (COVID-19) affects multiple systems, including the gastrointestinal tract and the pertinent gut barrier function. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) can inflict either direct cytopathic injury to intestinal epithelial and endothelial cells or indirect immune-mediated damage. Alternatively, SARS-CoV-2 undermines the structural integrity of the barrier by modifying the expression of tight junction proteins. In addition, SARS-CoV-2 induces profound alterations to the intestinal microflora at phylogenetic and metabolomic levels (dysbiosis) that are accompanied by disruption of local immune responses. The ensuing dysregulation of the gut-lung axis impairs the ability of the respiratory immune system to elicit robust and timely responses to restrict viral infection. The intestinal vasculature is vulnerable to SARS-CoV-2-induced endothelial injury, which simultaneously triggers the activation of the innate immune and coagulation systems, a condition referred to as "immunothrombosis" that drives severe thrombotic complications. Finally, increased intestinal permeability allows an aberrant dissemination of bacteria, fungi, and endotoxin into the systemic circulation and contributes, to a certain degree, to the over-exuberant immune responses and hyper-inflammation that dictate the severe form of COVID-19. In this review, we aim to elucidate SARS-CoV-2-mediated effects on gut barrier homeostasis and their implications on the progression of the disease.

Key Words: COVID-19; SARS-CoV-2; Intestinal barrier; Dysbiosis; Immunothrombosis; Gut-lung axis



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Core Tip: Severe coronavirus disease 2019 (COVID-19) is associated with a multi-layered disruption of gut barrier integrity. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) inflicts direct cytopathic or indirect immune-mediated injury to intestinal epithelial and endothelial cells and enhances paracellular permeability by downregulating tight junction proteins. SARS-CoV-2 induces profound gut microbiome alterations accompanied by dysregulation of mucosal immune responses. Gut dysbiosis attenuates, through the gut-lung axis, the ability of the respiratory immune system to elicit vigorous responses to contain SARS-CoV-2. Additionally, intestinal barrier dysfunction promotes endothelial activation and predisposes to detrimental COVID-19-related thrombotic complications. Finally, bacterial translocation and endotoxemia contribute to over-exuberant immune responses and hyper-inflammation in severe COVID-19.

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INTRODUCTION

The emergence of the novel, pathogenic, and highly transmissible severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), causing coronavirus disease 2019 (COVID-19), triggered an unprecedented public health crisis with profound socioeconomic sequelae. In most cases, COVID-19 is associated with mild-to-moderate symptoms that mainly involve the respiratory tract. However, in a subset of patients, COVID-19 may progress into a more severe disease plagued with complications such as pneumonia, acute respiratory distress syndrome (ARDS), coagulopathy, myocarditis, hepatic injury, renal dysfunction, sepsis, multiple organ failure, or even death[1]. These detrimental effects are considered to be driven by aberrant activation of the host's immune system in response to viral invasion and proliferation into the pulmonary parenchyma[2]. In particular, the virus-laden pneumonocytes secrete excessive amounts of pro-inflammatory mediators and chemoattractant molecules, such as interleukins (IL-1 β , IL-6, IL-7, IL-8, IL-12), tumor necrosis factor alpha (TNF- α), interferons (IFN- γ , IFN- λ), macrophage inflammatory protein-1 alpha (MIP-1 α), monocyte chemoattractant protein-1 (MCP-1), and granulocyte colony stimulating factor. Subsequently, the recruitment and activation of innate and adaptive immune cells into the lungs further aggravate tissue injury and inflammation[2]. In parallel, the systemic dissemination of this "cytokine storm" precipitates overactivation of the immune system beyond the sites of infection and elicits hyperinflammatory responses that impair the function of several organs including the heart, kidneys, liver, nervous system, and gastrointestinal (GI) tract^[3].

The GI system harbors an enormous interface that directly communicates with the external environment of the body and fulfills multifaceted functions. The GI mucosa serves as a semi-permeable membrane, allowing the efficient absorption of water, electrolytes, and nutrients while, in parallel, preventing the influx of xenobiotics, intraluminal microbiota, microbial components, or other inflammatory stimuli into the organism^[4]. This subtle balance is maintained thanks to intestinal barrier function, which, apart from providing a physical barrier, regulates complex immune system responses and mediates the intricate crosstalk with the gut microbiome^[5]. The integrity of the intestinal barrier can be compromised in many acute or chronic pathological conditions, leading to increased bacterial translocation and excessive penetration of pro-inflammatory signals. This dysfunction is associated with infectious complications and the establishment of a systemic pro-inflammatory status that can exacerbate or accelerate the pathophysiological processes of the underlying disease[6].

In this review, we summarize the deleterious effects of SARS-CoV-2 infection on gut barrier homeostasis. Subsequently, we discuss the mechanisms that explain how intestinal barrier dysfunction might drive severe COVID-19 or induce detrimental complications.

GI INVOLVEMENT AND THE MECHANISMS OF INTESTINAL INFECTION IN COVID-19

SARS-CoV-2 is an enveloped, positive-sense, single-stranded RNA virus with a genome of approximately 30000 nucleotides that encodes 29 proteins, including 4 structural proteins: spike (S), nucleocapsid (N), membrane (M), and envelope (E) proteins. The S protein consists of the S1 subunit, which interacts with angiotensin-converting enzyme 2 (ACE2), and the S2 subunit which promotes membrane fusion[7,8]. The principal route of transmission of SARS-CoV-2 is via respiratory droplets or aerosols



from person-to-person, and its main target is type II alveolar epithelial cells (AEC2s). The entry of SARS-CoV-2 is primarily mediated by the attachment of the S glycoprotein with ACE2 on the cellular surface, a process facilitated by proteolytic cleavage at the S1/S2 boundary by host proteins. Transmembrane protease serine 2 (TMPRSS2) is a significant host protease that mediates the cleavage of S2 protein, leading to conformational changes that precipitate viral entry through membrane fusion[7, 8]. Intriguingly, ACE2 and TMPRSS2 are widely co-expressed on the membrane of intestinal epithelial cells (IECs) at a level comparable or even higher to that of the AEC2s, rendering the GI tract a potential target for SARS-CoV-2 infection[9].

Multiple studies have demonstrated that the development of GI-related symptoms is a common extrapulmonary manifestation, affecting up to one-fifth of patients with COVID-19[10-12]. According to a meta-analysis of 108 studies comprising 17776 COVID-19 patients, the pooled prevalence estimates of GI symptoms were: 21% for loss of appetite, 13% for diarrhea, 8% for nausea or vomiting, and 4% for abdominal pain, while derangement of liver function tests was observed in 24% of the participants [10]. Whether the occurrence of GI symptoms is associated with COVID-19 severity or outcomes has been a field of debate. Most meta-analyses support that GI-symptoms and predominantly abdominal pain tend to appear at higher rates among patients with severe disease[10,13-15]; however, these symptoms are not always predictive of mortality or intensive care unit (ICU) admission[16-18]. Importantly, viral RNA can be detected in the stool of about half of COVID-19 patients and, especially, in those with GI-related symptoms[19,20]. Viral rectal shedding appears to persist long after RNA clearance in respiratory samples (mean duration: 21.8 d vs 14.7 d)[20], while RNA concentration in the stool may be as high as 10⁷ copies/g, exceeding even that in pharyngeal swabs in some cases[21]. The combination of this particularly high RNA load in the stool and the long-lasting viral presence in fecal samples strongly suggests that SARS-CoV-2 invades and proliferates in the intestinal tract. Indeed, COVID-19 elicits acute inflammatory responses in the gut, as documented by increased fecal calprotectin levels[22]. Accordingly, Livanos et al[18] provided direct evidence of direct infection of IECs by SARS-CoV-2 in vivo, using immunofluorescence staining and electron microscopy. Viral subgenomic mRNA (sgmRNA) is transcribed in infected cells, but is not encapsulated into virions, which means that sgmRNA is vulnerable to enzymatic degradation by intestinal ribonucleases. Therefore, the detection of sgmRNA in the stool of COVID-19 patients also indicates that the virus can actively and specifically replicate in the gut[21].

Although the data above corroborate the concept of SARS-CoV-2 tropism for the GI system, the routes and mechanisms of infection remain less clarified. At least theoretically, the feasibility of the fecal-oral route of transmission is supported by prolonged viral excretion in the feces. Nevertheless, although SARS-CoV-2 retains viability in stool for 1-2 d at room temperature, this is accompanied by a profound 5-log loss of its infectivity[23]. In addition, ingested SARS-CoV-2 needs to overcome the harsh gastric acidity, resist the detergent effects of bile acids in the duodenum, and avoid degradation by digestive enzymes to preserve its infectivity and spread into the intestinal epithelia[24]. The uninterrupted function of these host defense mechanisms hinders the fecal-oral transmission of enveloped viruses, such as SARS-CoV-2[25]. Besides, several lines of evidence support that intestinal infection by SARS-CoV-2 could occur via the bloodstream [19,26]. The virus replicating in pulmonary alveolar epithelial cells or in ciliary cells of the nasal cavity is capable of breaching the basement membrane and causing viremia^[27]. The GI tract comprises an extended network of capillaries harboring vascular epithelial cells, which are potential targets of circulating SARS-CoV-2. Thereafter, the proliferating virus is released from the infected cells into the gut, where it can infect neighboring enterocytes or reenter the circulation to generate new cycles of infection[27,28].

THE INTESTINAL BARRIER FUNCTION IN HEALTH

The gut barrier function is equipped with three major lines of defense that conjointly serve its complex purpose[5]. First, the mechanical barrier consists of tightly linked columnal IECs, the overlying mucus layer, and the capillary endothelial cells of the submucosa^[29]. Tight junctions (TJs) and adherens junctions (AJs) are transmembrane multiprotein complexes that confer structural stability to the mechanical barrier and engender the establishment of cell polarity. TJs connect the most apical of the lateral surfaces of the adjacent epithelial or endothelial cells (kissing points) and form channels that regulate the selective diffusion of ions and salutes through the paracellular space. AJs are more basal than TJs and contribute to the establishment and maintenance of cell-cell adhesion[30,31]. The mucus layer comprises a hydrated network of polymers, predominantly highly glycosylated mucin proteins, as well as several immune regulators, such as antimicrobial proteins (AMPs) and secretory immunoglobulin A molecules. The mucus lubricates the luminal contents and serves as a physical barrier against digestive enzymes, proliferating microorganisms, microbial components and byproducts, foodassociated toxins, or other inflammatory stimuli[32]. In addition, it nurtures a thriving biofilm of microorganisms, the gut microbiome, which typifies the second mechanism of protection, the biological barrier^[29,32]. Luminal microbiota are indispensable for the fermentation of indigestible carbohydrates, a process that provides the gut with short-chain fatty acids (SCFAs), mainly consisting of acetate,



propionate, and butyrate. Apart from being an important source of energy for host cells, SCFAs exhibit significant anti-inflammatory and anti-tumor effects and participate in various host signaling pathways, contributing to intestinal barrier integrity and metabolic homeostasis The commensal bacteria regulate choline bioavailability, promote the enterohepatic circulation of bile acids, and synthesize vitamin K and group B vitamins. In parallel, normal intestinal flora restrains the overgrowth of harmful microorganisms, a phenomenon referred to as colonization resistance, through nutrient antagonism[33]. Moreover, the microbiome-derived pathogen-associated molecular patterns (PAMPs) are recognized by pattern recognition receptors (PRRs) expressed by intestinal immune cells, indicating a relentless reciprocal dialogue between the microbiota and the intestinal immune system[6]. This leads to the third line of defense, that is, the immune barrier, which encompasses the gut-associated lymphoid tissue (GALT), effector and regulatory T cells (Tregs), immunoglobulin A (IgA)-secreting B (plasma) cells, innate lymphoid cells (ILCs), as well as macrophages and dendritic cells (DCs) of the lamina propria[5]. The cells of innate immunity carry an armamentarium of PRRs such as toll-like receptors (TLRs), nucleotide oligomerization domain-like receptors (NLRs), and retinoic acid-inducible gene-like receptors (RLRs) that recognize molecular pathogen-associated molecular patterns (PAMPs) or damaged-associated MPs (DAMPs) and orchestrate the well-tuned responses of the adaptive arm of the immune system.

The continuous crosstalk of the immune cells with the gut microbiome sustains the delicate balance between tolerance to beneficial bacteria and immunosurveillance against pathogenic species[6]. Commensal-derived signals and metabolites are recognized by myeloid cells in the lamina propria and orchestrate innate and adaptive immune responses[34]. Under homeostatic conditions, innate immune cells, such as macrophages and DCs, obtain a regulatory phenotype that promotes the secretion of antiinflammatory molecules, *i.e.*, IL-10 and TGF-β. The production of IL-22 by type 3 ILCs supports tissue homeostasis and epithelial barrier integrity[35]. Properly-regulated antigen presenting cells (APCs) remain in the GALT or migrate to the mesenteric lymph nodes, where priming of naïve CD4+ T cells occurs, and stimulate Treg cell activity that plays a central role in the suppression of intestinal inflammation. In this immunomodulatory milieu, B cell activation and effective class-switching generate large numbers of IgA-secreting plasma cells that serve multiple functions, including protection against infection and maintenance of gut microbiome homeostasis[35].

THE INTESTINAL BARRIER FUNCTION IN COVID-19

SARS-CoV-2 disrupts the intestinal mechanical barrier

SARS-CoV-2 can invade and propagate in IECs by using the vastly expressed ACE2 and TMPRSS2 receptors on those cells [18,36]. In a mouse model, ACE2 was found to be a key inducer of intestinal stem cell proliferation and differentiation under pathologic conditions, while ACE2 deficiency was associated with a significant reduction of mucin-2 expression[37]. Accordingly, ACE2 knockout mice exhibited gut barrier dysfunction with subsequent leakage of bacterial components into the circulation[38]. Therefore, it could be hypothesized that interference of SARS-CoV-2 with ACE2 signaling could destabilize the mechanical barrier by interrupting the renewal of epithelial cells or by compromising mucus composition. Mucins create a protective matrix covering the epithelium and inhibit viral invasion, presumably via steric hindrance[39]. Even though evident macroscopic alterations were usually negligible, infection of the human small bowel by SARS-CoV-2 was associated with villi blunting and an aberrant accumulation of activated intraepithelial CD8+ T cells in the epithelium[36]. Cell trafficking could be driven by direct infection of IECs or could be the result of systemic immune activation due to COVID-19. In any case, the recruitment of intraepithelial CD8+ T cells enhanced the apoptotic process of IECs, as demonstrated by an upsurge in cleaved caspase-3+ apoptotic epithelial cells. This event was accompanied by a regenerative response of the epithelium marked by an increase in Ki67+ proliferating epithelial cells that extended beyond their typical localization in the crypts and occupied the villus compartment[36]. Evidently, SARS-CoV-2 can inflict a deleterious impact on the mechanical barrier through dysregulation of the balance between cell apoptosis and proliferation.

The function and integrity of epithelia and endothelia greatly depend on TJs, reflecting a putative mechanism of intestinal barrier injury in COVID-19 patients. The fundamental transmembrane proteins that frame TJs include the family of claudins (central regulators of paracellular permeability encompassing 26 members in humans), the junctional adhesion molecules, and the three junctional MARVEL domain proteins, i.e., occludin, tricellulin, and MARVEL domain-containing protein 3. The most prominent intracellular junctional plaque components are zonula occludens (ZO) proteins (ZO1, ZO2, and ZO3), cingulin, and protein associated with LIN7 1 (PALS1)[40]. These proteins function as adaptor proteins or cytoskeletal linkers and participate in multiple extracellular and intracellular signaling pathways. Indeed, impairment of TJs occurs early in the course of COVID-19 and might represent the first hit in a multistage model of the disease[41]. SARS-CoV-2 infection decreases the expression of TJ proteins, such as occludin, claudin 5, and ZO-1 in vivo, and undermines the coherence of TJs between neighboring endothelial cells of the brain. As a result, TJs are haphazardly distributed, irregular, or gapped throughout the vascular endothelial layer, leading to derangement of the function



of the blood-brain barrier [42]. Cryo-electron microscopy and in silico modeling analyses have shown that SARS-CoV-2 uses its envelope (E) protein to interact with the TJ-associated PALS1 protein[43,44]. According to *in vitro* models of lung injury, the interplay between SARS E protein and PALS1 interrupts intracellular trafficking of E-cadherin, delays the formation of TJs and AJs, and affects epithelial polarity [45]. In a proof-of-concept study, Guo *et al*[46] developed a biomimetic human gut-on-chip model that reconstructs basic elements of the gut barrier, as it consists of IECs, endothelial, and mucin-producing cells under normal fluid flow and closely reproduces the pathophysiological processes of intestinal SARS-CoV-2 infection. Interestingly, IECs exhibit particularly high susceptibility to SARS-CoV-2 infection. Viral inoculation induces the dispersal of the physiological distribution of mucus-secreting cells and a profound reduction of both E-cadherin expression in the epithelium and VE-cadherin in the endothelium, which delineate serious impairment of AJs in the corresponding structures. As a consequence, widespread destruction of the villus-like complexes along with severe morphological remodeling of the vascular endothelium was observed [46]. IL-6 is a prominent mediator of inflammation and a reliable biomarker of disease severity in SARS-CoV-2 infection that was found to increase, in a sustained manner, endothelial permeability in a mouse model[47,48]. Administration of IL-6 induced vascular leakage and disruption of junctional localization of VE-cadherin and ZO-1 via Janus kinase-mediated signal transducer and activator of transcription 3 phosphorylation and de novo protein synthesis^[48].

Serum levels of endotoxin and ZO-1 were significantly increased in patients with COVID-19-related pneumonia on admission compared to healthy controls. Importantly, endotoxemia is positively correlated with certain markers of inflammation, such as C-reactive protein (CRP) and ferritin[49]. Endotoxins are complex lipopolysaccharides (LPS), integral parts of the membrane of gram-negative bacteria, and potent drivers of inflammation. Indeed, endotoxemia can occur as a result of gramnegative bacteremia; however, endotoxemia is most commonly caused by a compromised gut barrier [50]. ZO-1 is a peripheral membrane scaffolding protein and a basic constituent of TJs that fulfills versatile functions including establishment of cell-cell adhesion, modulation of the paracellular barrier, regulation of cell migration and angiogenesis, and induction of mucosal repair processes[51,52]. Previous studies have confirmed that serum ZO-1 represents a reliable biomarker of disrupted paracellular permeability, as it inversely correlates with intestinal ZO-1 expression in diverse pathologic conditions[53,54]. These results are consistent with another study, in which severe SARS-CoV-2 infection presented key features of gut barrier dysfunction in tandem with increased intestinal permeability. Specifically, patients with severe COVID-19 presented with endotoxemia and higher serum levels of zonulin, occludin, and regenerating family member 3 alpha, indicating severe impairment of the intestinal epithelial barrier[55].

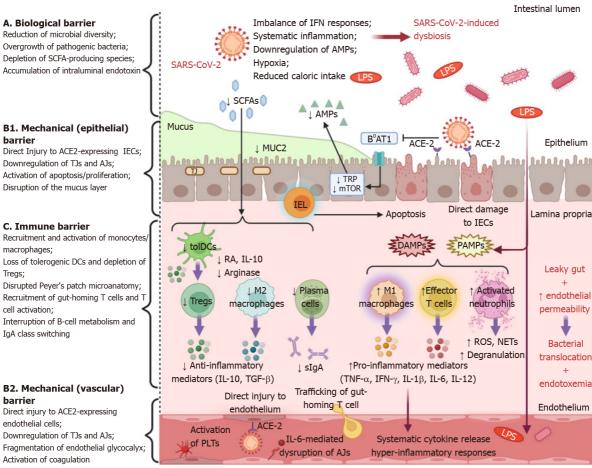
SARS-CoV-2-induced gut microbiome dysbiosis

In the course of evolution, hosts and their microbial ecosystems have mutually developed, forging an intimate relationship of interdependence. Dysbiosis refers to alterations in the composition, quantity, or distribution of the gut microbiome. This condition is characterized by a predominance of pathogenic species and has been linked to the pathophysiology of numerous intestinal and extraintestinal disorders [56]. There is a growing body of data suggesting that COVID-19 is associated with drastic alterations of the normal intestinal flora, even when removing the confounding effect of antibiotics (Figure 1)[57-59]. In addition, SARS-CoV-2-induced dysbiosis appears to persist long after the resolution of symptoms and might be a predisposing factor for long-term complications in COVID-19 patients [57,58,60].

The mechanisms underlying COVID-19-related dysbiosis have not yet been fully elucidated. The interactions between SARS-CoV-2 and the ACE2 receptor can influence the composition of gut microbiota by interfering with the secretion of AMPs. The activity of the amino acid transporter B⁰AT1, which mediates the intestinal uptake of tryptophan, depends on ACE2 signaling[61]. Tryptophan regulates AMP production through the mammalian target of rapamycin pathway[62]. Therefore, tryptophan depletion due to ACE2 blockade can interrupt AMP production, and thus, perturb the intraluminal microbial community. Commensal bacteria, in turn, contribute to mucosal homeostasis by regulating ACE2 expression in the gut[63]. The release of pro-inflammatory cytokines, particularly TNF- α , during respiratory infections has a powerful anorexigenic effect *via* hypothalamic action. The ensuing reduction in caloric intake and dietary fibers disturbs the composition of the gut microbiota and the production of their metabolites. In animal models of respiratory syncytial virus (RSV) and influenza infection, the neutralization of anorexigenic cytokines prevented weight loss and mitigated gut microbiome alterations[64]. Alternatively, SARS-CoV-2 infection can cause dysbiosis through an imbalance of systemic or intestinal IFNs[65]. In this regard, in an influenza mouse model, the microbiome was amenable to significant changes via an IFN-type I-dependent mechanism[66]. Hypoxia is a serious feature of severe COVID-19 and hypoxic stress could be an important instigator of dysbiosis by dysregulation of hypoxia-inducible factor signaling[67]. Finally, local epithelial injury, which results in leaky gut and DAMP secretion, might disrupt the immune control of microbial homeostasis and could further aggravate the dysbiotic state[68].

In their recently published meta-analysis, Farsi et al[69] offered a thorough synthesis of the gut microbiota changes in COVID-19 patients. At the phylum level, dysbiosis is typified by a decrease in the Firmicutes to Bacteroidetes ratio. More specifically, COVID-19 is associated with a decrease in important





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Figure 1 Schematic representation of intestinal barrier dysfunction in severe acute respiratory syndrome coronavirus 2 infection. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection is associated with a multifaceted dysfunction of the gut barrier as it exerts detrimental effects on all of its major levels of defense. A: Systemic inflammation, an imbalance of interferon (IFN) responses, hypoxia or low blood flow, and reduced caloric intake, due to coronavirus disease 2019-associated cachexia, contribute to intestinal microbiome alterations. Gut dysbiosis is characterized by an overgrowth of opportunistic pathogens, which are a source of harmful substances, e.g., endotoxin, peptidoglycan, β-glucan, as well as a depletion of commensal microorganisms, which synthesize beneficial metabolites such as short-chain fatty acids; B: SARS-CoV-2 induces either direct cytopathic injury to angiotensin-converting enzyme 2 (ACE2)expressing intestinal epithelial cells (IECs) and endothelial cells, or indirect immune-mediated damage, or both. Moreover, the secretion of mucin glycoproteins, the fundamental element of the mucus layer, is severely impaired. In addition, the virus undermines the expression of tight junction and adherens junction proteins, leading to increased paracellular permeability and structural destabilization of the epithelium and endothelium. Viral interference with ACE2 signaling attenuates the activity of the amino acid transporter BOAT1, which is necessary for intestinal absorption of tryptophan (TRP). As a result, TRP depletion downregulates the mammalian target of rapamycin pathway, which promotes the expression of antimicrobial peptides. The overwhelming intraepithelial CD8+ lymphocytes trigger the apoptosis of IECs, which is followed by reactive regeneration. In parallel, disease-activated and apoptotic endothelial cells are potent drivers of neutrophil/monocyte recruitment, platelet aggregation, and coagulation activation; C: Commensal-derived metabolites modulate innate immune responses by inducing tolerogenic dendritic cells and M2-polarized macrophages. SARS-CoV-2-induced dysbiosis eliminates these immunoregulatory effects, inhibits T regulatory (Treg) cell activity, and suppresses the secretion of anti-inflammatory cytokines such as interleukin 10 (IL-10) and transforming growth factor beta. The release of damage-associated molecular patterns from injured cells and the influx of pathogen-associated molecular patterns, as a result of leaky gut, orchestrate the recruitment and activation of innate immune cells that secrete pro-inflammatory mediators (IFN-y, tumor necrosis factor alpha, IL-1β, IL-6, IL-12). Subsequently, this pro-inflammatory microenvironment precipitates the derangement of adaptive immune responses, as demonstrated by increased trafficking of gut-homing T cells and effector T cell predominance. Furthermore, interruption of B-cell metabolism, plasma cell differentiation, and immunoglobulin A (IgA) class switching impede the effective secretion of protective dimeric IgA. This multi-layered disruption of intestinal barrier integrity allows the dissemination of intraluminal bacteria and endotoxin into the mesenteric lymphatic vessel or/and into the systemic circulation (created with biorender.com). IEL: Intraepithelial lymphocyte; LPS: Lipopolysaccharides; MUC2: Mucin 2; NETs: Neutrophil extracellular traps; PLTs: Platelets; RA: Retinoic acid; ROS: Reactive oxygen species; slgA: Secretory immunoglobulin A; SCFAs: Short-chain fatty acids.

> butyrate-producing bacteria, including Faecalibacterium and Roseburia[69-71]. The genus Roseburia contributes to mucosal integrity and colonic motility and exerts significant anti-inflammatory effects by modulating IL-10 production^[72]. Similarly, *Faecalibacterium prausnitzii* is a valuable gut symbiont with recognized anti-inflammatory effects in IBD patients via inhibition of the nuclear factor kappa B pathway^[73]. The gut microbiome of COVID-19 patients is also depleted of several other beneficial genera such as Alistipes, Eubacterium, and Bifidobacterium [69]. The genus Eubacterium consists of a phylogenetically diverse group of SCFA-producing bacteria that contribute to bile acid metabolism and exhibit compelling associations with intestinal health [74]. Bifidobacterium strains prompt Treg responses, induce tolerogenic DC phenotypes, and demonstrate vigorous antimicrobial and antiviral activity[75].



Of note, their ability to suppress gut dysbiosis and promote mucosal homeostasis has paved the way for researchers to investigate the therapeutic efficacy of Bifidobacterium-based probiotic preparations in several diseases, including irritable bowel syndrome, inflammatory bowel disease (IBD), or COVID-19 [76-78]. On the other hand, the COVID-19-derived gut microbiome is relatively enriched in opportunistic pathogens such as Streptococcus, Bacteroides, Rothia, Veillonella, Actinomyces, and Eggerthella [69,70]. Although the genus *Ruminococcus* is generally underrepresented, certain species such as *Rumino*coccus gnavus and Ruminococcus torques are significantly more abundant in COVID-19 patients. This is not surprising considering that Ruminococcus gnavus and Ruminococcus torques are harmful bacteria that degrade mucin glycans to harvest energy, secrete pro-inflammatory mediators, and are involved in IBD pathogenesis[79,80].

The degree of microbiome alterations correlates well with COVID-19 severity, and identification of early patterns of dysbiosis could lead to a microbiome-based stratification of patients according to their risk of progressing to severe COVID-19[81]. Indeed, the commensal genera Faecalibacterium and Roseburia are depleted in the gut microbiome of patients with critical disease [57,82]. The baseline abundance of Clostridium ramosum and Clostridium hathewayi is also associated with COVID-19 severity and could represent risk factors for portal vein thrombosis[69,82]. The genus Enterococcus is overrepresented in COVID-19 patients who necessitate ICU admission or developed bloodstream infections, whereas, surprisingly, the gut microbiome of other critically ill non-COVID-19 patients is devoid of this bacterium[83]. The reduction of the Firmicutes/Bacteroidetes ratio is indicative of severe disease, which is mainly attributed to the depletion of fiber-utilizing bacteria, namely Faecalibacterium prausnitzii, Phocaeicola plebeius, and Prevotella[84]. These findings are consistent with an interesting study exploring the role of gut microbiota as predictors of disease severity[81]. A lower Firmicutes/Bacteroidetes ratio, a higher prevalence of Proteobacteria, and an exhaustion of commensal butyrate-producing microorganisms are more evident in severely ill patients, while a lower bacterial diversity, defined by the Shannon diversity index, is identified as a prognostic biomarker of disease severity[81]. In agreement with this, the fecal microbiome of patients requiring mechanical ventilation has demonstrated low bacterial richness as assessed by Shannon or Chao1 indices[81,85,86]. Implementation of a multiomics approach to decipher the dysregulation of metabolic and microbial signatures during COVID-19 could provide a basis for the development of novel microbiome-targeted therapeutics[87].

SARS-CoV-2 deranging mucosal immune system responses

The mucosal immune system is the largest immunologically aware organ in the body, committed to maintaining the equilibrium between active protection against pathogens and immune-tolerance to commensal microorganisms, dietary substances, and self-antigens. The gut-derived metabolites orchestrate immune cell responses and differentiation and impart a critical role in mucosal homeostasis [56,88]. In particular, the transcriptional "education" of innate immune cells is strongly influenced by intestinal microbiota metabolites[56]. Butyrate, for example, induces M2-like polarization of macrophages, which shapes an immunomodulatory milieu by increasing the expression of arginase 1 and suppressing TNF-α, IL-6, IL-12b, and nitric oxide synthase 2[89]. As regards adaptive immunity, butyrate enhances transcription of the forkhead box P3 gene in naïve T cells through inhibition of histone deacetylases, and thereby, expands Treg cell populations[90]. The capsular polysaccharide A, which originates from the prominent human symbiont *fragilis*, promotes the proliferation of Treg cells in the lamina propria and shapes a pro-inflammatory microenvironment rich in IL-10[91]. Vitamin A or RA, produced by the gut commensal Bifidobacterium infantis, enhances the expression of aldehyde dehydrogenase 1 family member A2 encoding retinal dehydrogenase 2 in resident DCs of the mucosa. Subsequently, gut-modulated DCs secrete high levels of RA that drive naïve T cell differentiation into Treg cells[92]. Treg cell generation in the intestinal mucosa is also triggered by β -glucan polysaccharides deriving from the cell surface of *Bifidobacterium bifidum*[93]. In addition, B cell metabolism and differentiation in mucosal and systemic tissues are regulated by gut-derived SCFAs, emphasizing the significance of symbionts in effective antibody production[94]. Apparently, the depletion of SCFAproducing microorganisms or other beneficial species as well as gut metabolome modifications might have a detrimental impact on mucosal immunity in the course of COVID-19.

Defensins are prominent members of the AMP family with multifaceted immunomodulatory functions and broad antimicrobial and antiviral activity. Defensins provide protection against SARS-CoV-2 infection not only by maintaining gut microbiome homeostasis but also by inhibiting viral fusion via interference with ACE2 receptors[95,96]. Dysregulation in the expression of various defensin genes was evident following infection with COVID-19[97]. IFNs are multipotent cytokines of innate immunity with crucial role in the containment of viral infections. The proliferation of SARS-CoV-2 in human gut cells is effectively inhibited by type I (IFN- α , IFN- β) and type III IFNs (IFN- λ s); however, type III IFNs elicit a more profound and long-lasting antiviral effect[98,99]. In order to escape immune surveillance, SARS-CoV-2 has developed strategies to proliferate stealthily into cells without eliciting strong IFN responses [65]. Alternatively, SARS-CoV-2 can interfere with IFN- γ (type II IFN) signaling to boost its infectively in the gut. In human colonic organoids, IFN-γ drives cellular differentiation towards ACE2expessing epithelial cells, which are highly susceptible to SARS-CoV-2 infection [100]. Moreover, IFN- γ acts synergistically with TNF- α to instigate inflammatory cell death and tissue damage[101].

Elevated levels of plasma markers of inflammation and immune activation are hallmarks of severe COVID-19. Similarly, the expression of genes encoding pro-inflammatory mediators, including TNF- α , IL-6, chemokine (C-C motif) ligand 5 (CCL5), chemokine (C-X-C motif) ligand 1 (CXCL1), CXCL10, CXCL11, and CSF-3 were upregulated in digestive tissues in both in vitro and in vivo models of SARS-CoV-2 infection [46,102]. Compared to uninfected individuals, patients with COVID-19 have increased fecal levels of pro-inflammatory cytokines such as IL-8 and IL-18, whereas IL-23 is higher in patients with severe disease[103,104]. Strikingly, analysis of human ileal biopsies has demonstrated that several pro-inflammatory genes including IFNG, CXCL8, CXCL2, and IL1B are downregulated. This pro-inflammatory response is associated with milder symptoms and improved outcomes, revealing an immunomodulatory function of the GI in controlling SARS-CoV-2 infection[18]. Nevertheless, molecular events in severe COVID-19, due to increased bacterial translocation and systemic immune activation, might eliminate the immunoregulatory functions of the GI. Indeed, post-mortem evaluation of COVID-19 patients' intestinal tissues revealed the overexpression of TNF-α and IL-10, which exerts antithetical effects [105]. The presence of IL-10 in the gut and lung may have immunosuppressive effects by inhibiting the expression of the human leukocyte antigen DR isotype (HLA-DR) on APCs[105]. Depletion of HLA-DR expression on monocytes is a major characteristic of immune dysregulation in COVID-19 patients who develop severe respiratory failure[106].

Essentially, changes in the expression of cytokines and downstream dysregulation of their signaling networks mirror alterations in the composition of immune cell populations of the lamina propria. Imaging mass cytometric analysis showed that the intestinal tract of deceased patients with COVID-19 accommodated higher numbers of CD11b+ macrophages, CD11c+ DCs, natural killer T cells, and B cells compared to healthy controls[105]. Another study, which evaluated post-mortem tissues of COVID-19 patients, demonstrated severe disorganization of ileal Peyer's patches with loss of B cell/T cell zonation and depletion of the germinal center. In addition, impaired interactions between B and T cells, an enhanced number of follicular macrophages, the reduction of CD27+ memory B cells, and downregulation of CD74 expression on B cells were reported[107]. Livanos et al[18] in their study, which supports an anti-inflammatory function of the GI in SARS-CoV-2 infection, showed that the lamina propria of COVID-19 patients was depleted of conventional DCs and plasmacytoid DCs, whereas effector CD4+ and CD8+ T cells as well as tissue resident memory T cells were increased. Lehmann et al[36]. revealed that the activation and proliferation of antigen-experienced intraepithelial CD8+ T cells into the intestinal mucosa was associated with epithelial barrier dysfunction in COVID-19 patients, while monocytes and macrophages of the duodenal mucosa expressed markers indicative of their recent recruitment from the circulation. In agreement, levels of CCL25, a gut homing marker, were increased in the sera of COVID-19 patients, suggesting that intestinal inflammation might result from CCL5/CCR9mediated trafficking of gut-specific T cells into the mucosa[108,109]. Humoral immune responses are vital for counterattacking viral invasion through the production of neutralizing antibodies. Secretory dimeric IgA is the predominant mucosal antibody and an integral component of the immunological barrier[6]. IgA antibody overproduction dominates the early pre-specific humoral response to SARS-CoV-2 infection, while virus-specific IgA antibodies display more robust neutralizing capacity compared to their monomeric IgG counterparts[110]. Interestingly, the levels of mucosal SARS-CoV-2specific IgA are inversely correlated with age[111]. Therefore, it can be inferred that the inability of B cells to mount an effective IgA response contributes to excessive viral propagation in the course of severe COVID-19[112].

THE GUT-LUNG AXIS IN COVID-19

As mentioned above, homeostasis of the gut microbiome can be immensely affected in the course of respiratory tract infections such as COVID-19. Conversely, intestinal microbiota plays an important role in fine-tuning the systemic immune system and eliciting efficient antiviral responses to address lung infections[113-115]. Gut bacterial components and metabolites can enter portal circulation or mesenteric lymphatics, which drain to the cisterna chyli first, then to the thoracic duct, and finally to the left subclavian vein. Intriguingly, the pulmonary vascular bed is the first to interact with the mesenteric lymph, implicating the importance of gut-derived signals in shaping lung immune responses in health as well as in driving ARDS in critically ill patients[116]. Under normal conditions, commensalassociated stimuli provide an indispensable for optimal antiviral activity, tonic activation of the host's innate immunity through their impact on alveolar macrophages, resident DCs, and lung epithelial cells (Figure 2)[117]. In contrast, germ-free mice are unable to evoke strong innate and adaptive immune responses and, thus, experience feeble control of viral infections and unfavorable outcomes[117,118]. Although the intestinal and respiratory tracts are anatomically distinct compartments, their mucosal immune cells and microbial communities configure a bidirectional "gut-lung" axis cross-talk that is highly pertinent to COVID-19 pathogenesis.

The mechanisms through which intestinal flora reinforces lung resilience against viral invasion have only recently begun to be unraveled with the assistance of murine models. The gut microbiota orchestrates the steady-state IFN signature in lung stromal cells, which protects against early influenza virus

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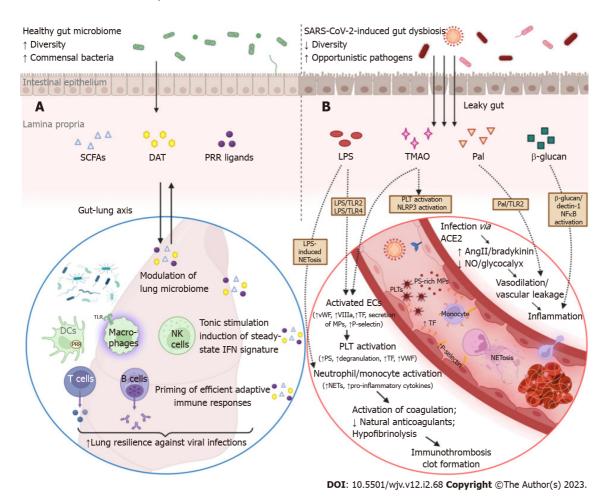


Figure 2 Overview of the sequelae of gut barrier dysfunction in severe coronavirus disease 2019: dysregulation of lung immune responses and establishment of a prothrombotic state. A: Although the intestinal and respiratory tracts are anatomically distinct compartments, their mucosal immune cells and microbial ecosystems participate in a bidirectional immunological crosstalk (gut-lung axis). An intact intestinal barrier is pivotal in maintaining lung microbiome homeostasis and fine-tuning the respiratory immune system to elicit potent antiviral responses in the case of infection. Commensal bacteria provide tonic stimulation (through the production of pattern-recognition receptor-ligands, desaminotyrosine, short-chain fatty acids, etc.) of the epithelial, stromal, and innate immune cells of the lungs and modulate the steady-state interferon-signature, which is essential for suppressing the early phase of viral proliferation. In addition, gut-derived signals and metabolites orchestrate the effective priming of adaptive immune responses by inducing the differentiation of virusspecific CD4+ and CD8+ T cells and antibody-secreting plasma cells, which are responsible for viral control and clearance in the later stages of infection. In coronavirus disease 2019 (COVID-19), gut barrier dysfunction and depletion of symbiotic microorganisms eliminate the aforementioned immunomodulatory effects and compromise the ability of the respiratory immune system to effectively contain severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection; B: The gut vascular bed has a massive endothelial surface that is susceptible to SARS-CoV-2 infection. SARS-CoV-2 can inflict direct injury to angiotensin-converting enzyme 2 (ACE2)-expressing endothelial cells; interruption of ACE2 signaling can dysregulate both the renin-angiotensin and kinin-kallikrein systems, leading to vascular leakage. The activated or apoptotic endothelial cells release phosphatidylserine-rich endothelial microparticles and secrete large amounts of tissue factor, VIIIa, von Willebrand factor, and other procoagulant cofactors. Circulating platelets accumulate at sites of vascular injury, adhere to each other, and become activated, leading to further secretion of prothrombotic substances. Overexpression of adhesion molecules, such as P-selectin, facilitates the recruitment and activation of monocytes and other leukocytes, including neutrophil extracellular trap (NET)-producing neutrophils. COVID-19-associated immunothrombosis refers to this concurrent aberrant activation of the innate immune and coagulation systems, which predisposes to serious thrombotic complications. This vicious cycle can be further exacerbated by gut barrier dysfunction. Low-grade endotoxemia, due to increased intestinal permeability, enhances the activation of endothelial cells and platelets by inducing lipopolysaccharide (LPS)/toll-like receptor 2 (TLR2) and LPS/TRL4 downstream signaling pathways. In parallel, LPS is a potent driver of NET formation. Several other bacterial lipoproteins, such as peptidoglycan-associated lipoprotein (Pal) or Pam³Cys, aggravate vascular leakage and precipitate thrombus formation through TLR2 activation. Moreover, translocation of fungal components, such as β-glucan, could directly stimulate leukocytes and promote inflammation by binding to the Dectin-1 receptor and activating the nuclear factor-kB pathway. Finally, gut dysbiosis is associated with trimethylamine N-oxide overproduction, which is a recognized risk factor for clotting events as it enhances platelet hyperresponsiveness, endothelial dysfunction, and NLR family pyrin domain containing 3 inflammasome activation (created with biorender.com). Angll: Angiotensin II; DCs: Dendritic cells; ECs: Endothelial cells; NK cells: Natural killer cells; NO: Nitric oxide; PLT: Platelet; DAT: Desaminotyrosine; SCFAs: Short-chain fatty acids; PRRs: Pattern recognition receptors.

> proliferation by driving the expression of IFN- α/β receptor[113]. In line with this, when mice were exposed to antibiotics, their macrophage-related type I and type II IFN responses were severely impaired, resulting in the uncontrolled spread of systemic lymphocytic choriomeningitis virus or mucosal (influenza virus) infection. In a murine model of influenza infection, desaminotyrosine (DAT), a degradation product of dietary flavonoids, induced an efficient amplification of the IFN type I loop, which suppressed viral gene expression and airway epithelial damage. Notably, a distinct human-

derived commensal, Clostridium orbiscindens, produced DAT and rescued antibiotically-treated mice from viral infection[119]. An outer membrane glycolipid originating from the gut symbiont Bacteroides *fragilis* protected mice against viral infection through the induction and systemic release of IFN- β by colonic DCs[120]. Acetate, another metabolite derived from the gut microbiome, is able to enter the circulation, enhance IFN- β signal transduction via stimulation of the G-protein-coupled receptor 43 in pulmonary epithelial cells, and thus limit severe RSV infection[114]. Moreover, depletion of commensal bacteria precipitated significant epigenetic modifications at the level of mononuclear phagocytes residing in non-mucosal lymphoid organs. As a result, these cells were unable to induce type I IFN secretion, which led to suboptimal NK cell priming and poor antiviral responses[121]. Admittedly, an imbalance of IFN signaling is an inherent characteristic of the pathogenesis of respiratory tract infections, including SARS-CoV-2-associated pneumonia[122]. Early severe COVID-19 is governed by overwhelming IFN- α responses as well as NK cell functional exhaustion, which is manifested by abnormal expression of interferon-stimulated genes[123]. Therefore, it is plausible to assume that metabolites and signals stemming from luminal bacteria could influence the early response to SARS-CoV-2 infection by altering the IFN signature and compromising innate immunocompetence in the lungs.

Regulation of the mucosal immune system of the respiratory tract, which supervises airway colonization, depends on reciprocal signaling with the gut. The respiratory tract is not sterile; instead, it harbors a unique microbial ecosystem and its role in homeostasis and disease is being increasingly recognized with the advent of culture-independent molecular techniques[124]. Alterations of the lung microbiome signature are associated with clinical outcomes in critically ill patients infected with SARS-CoV-2[125]. The immunomodulatory potential of airway bacteria was exemplified by a pathogen-free murine model of influenza virus. Colonization of the upper respiratory tract by Staphylococcus aureus induces the recruitment of peripheral macrophages into the alveoli and their polarization toward an M2-like phenotype via TLR2 signaling. As a result, Staphylococcus aureus creates an anti-inflammatory pulmonary milieu that attenuated immune-mediated injury and prevented lethal influenza infection [126].

The robust and timely priming of adaptive immunity is necessary to contain SAR-CoV-2 infection. Delayed activation of adaptive immune responses and depletion of virus-specific T cells are hallmarks of severe or fatal COVID-19[127]. Interestingly, multiple studies have revealed that the gut microbiome is actively involved in shaping adaptive immunity in the respiratory tract. In a murine model of West Nile virus infection, exposure to antibiotics impaired the ability to elicit potent T cell responses, decreased the number of virus-specific CD8+ T cells, and led to worse disease outcomes[118]. Ichinohe et al[115] demonstrated that intestinal microflora regulates an even broader spectrum of adaptive immunity responses in the respiratory mucosa. More specifically, neomycin-sensitive commensal bacteria were essential for the induction of effective humoral responses and the generation of virusspecific CD4+ and CD8+ T cells in the lungs. Normal intestinal flora provided signals that maintained a steady-state IL-1β, pro-IL-18, and NLR family pyrin domain containing 3 (NLRP3) expression. Antibiotic-induced immunodeficiency is attributed to impaired inflammasome activation, abnormal activation and distribution of respiratory DCs, and inadequate DC migration to draining lymph nodes of the lung[115]. In this setting, Gauguet et al[128] demonstrated that the presence of segmented filamentous bacteria in the murine microbiome was vital for priming strong Th17 immunity responses and IL-22 secretion in the respiratory system. In response to fiber supplementation, the gut microbiome generates large amounts of SCFAs, which hinders influenza-induced lung injury through a dual mechanism concerning both arms of immunity[129]. First, SCFAs enhances the cellular metabolism and the effector functions of CD8+ T cells in the respiratory tract. Second, SCFAs induce an alternative activation of macrophages, which exhibit the limited ability to express the chemokine CXCL1. As a result, SCFA-modulated macrophages reduce early neutrophil infiltration and subsequent injury in the airways[129]. In a randomized controlled trial, non-hospitalized patients with symptomatic COVID-19 were allocated 1:1 to groups receiving an oral probiotic formulation or placebo for 30 d. Remarkably, probiotic supplementation was associated with higher rates of complete remission, decreased nasopharyngeal viral levels, and shorter duration of symptoms by inducing vigorous virus-specific IgM and IgG antibody responses.

In summary, a well-preserved intestinal barrier function, harboring a diverse consortium of commensal bacteria, provides the essential signals for appropriate and effective modulation of immune system responses in the lungs. Regardless of being the cause or the result of severe SARS-CoV-2 infection, gut dysbiosis is not a silent bystander but an active orchestrator of dysregulated immune responses in the respiratory tract. Modulation of gut microbiota represents an emerging therapeutic intervention to mitigate immune-mediated lung injury and improve COVID-19 outcomes[130].

GUT BARRIER DYSFUNCTION INFLAMING COVID-19-ASSOCIATED COAGULOPATHY

COVID-19-associated coagulopathy (CAC) is a life-threatening condition that can lead to arterial thromboembolism (ATE), such as acute coronary syndrome and cerebrovascular accident, or venous



thromboembolism (VTE), which manifests as deep vein thrombosis or pulmonary embolism[131]. Mild thrombocytopenia, prolonged prothrombin time, and increased serum levels of fibrinogen, CRP, Pselectin, and d-dimers have all been linked to clotting events in COVID-19 patients; these deviations become more pronounced as the disease progresses [131,132]. A recent meta-analysis, comprised of more than 90000 patients, concluded that hospitalized COVID-19 patients have a significantly increased 90-d risk of VTE in comparison to hospitalized individuals with influenza virus (9.5% vs 5.3%), and that this peril persists even after the breakthrough of effective vaccines[133]. COVID-19-related thromboembolic events are major burdens of morbidity and mortality, and their incidence increases with disease severity, affecting up to one third of patients in ICUs[134-136]. The pathophysiology underlying CAC is multifactorial, encompassing endothelial injury, over-exuberant immune responses, and overt dysregulation of coagulation and fibrinolytic pathways, which collectively result in a procoagulant state[131]. The activation of these mechanisms can be further aggravated by defects in intestinal barrier integrity due to SARS-CoV-2 infection[27,137].

The dysfunction of the endothelium is a hallmark of COVID-19, representing a common feature in multiple clinical manifestations of the disease such as thromboembolic events, neurological complications, and renal dysfunction [138,139]. The intestinal tract accommodates an enormous vascular endothelial surface consisting of a monolayer of squamous endothelial cells. Under physiological conditions, endothelial cells regulate the vascular tone and secrete anticoagulant and antiplatelet agents that preclude clotting events[140]. Human endothelial cells express the key cofactors, namely ACE2 and TMPRSS2, which are exploited by SARS-CoV-2 to invade its target cells[141]. There is evidence of viral inclusions in endothelial cells and mononuclear cell infiltrates in the walls of small vessels, as well as markers of endothelial cell apoptosis[142]. SARS-CoV-2-induced dysfunction of gut microvasculature and fragmentation of the endothelial glycocalyx eliminate these protective effects, promote vasoconstriction due to depletion of endothelium-derived nitric oxide, and drive platelet activation and fibrin formation[131]. Indeed, mesenteric thrombosis is not uncommon in critically ill patients with COVID-19, while focal ischemic lesions, inflammation of the endothelium, vessel wall edema, microhemorrhage, and microthrombi are frequent findings in resected bowel segments[27,143-145]. SARS-CoV-2-mediated dysregulation of ACE2 signaling in intestinal vascular endothelium might create a prothrombotic microenvironment through a dual mechanism. First, ACE2 catalyzes the conversion of angiotensin (Ang II) to angiotensin 1-7 (Ang 1-7), which confers important antithrombotic and immunoregulatory effects by binding to G-protein coupled Mas receptors. Therefore, ACE2 depletion attenuates the Ang 1-7 downstream pathway and shifts the balance in favor of Ang II, which binds to its cognate receptor and exerts harmful prothrombotic and inflammatory effects [146]. Second, the kinin-kallikrein system is also directly modulated by ACE and ACE2. Thus, the ACE/ACE2 imbalance can induce a "kinin storm" and amplify vascular permeability, cell migration, platelet activation, and oxidative stress[131,147].

There are data supporting the hypothesis that SARS-CoV-2 inflicts damage on endothelial cells (endotheliitis) via both direct cytopathic and indirect immune-mediated mechanisms[148]. Activated or apoptotic intestinal endothelial cells retract their margins, release endothelial microparticles (MPs), and abolish their ability to confine phosphatidylserine (PS) into the inner layer of the cellular membrane[27, 149]. In this setting, microRNA expression in endothelial-derived extracellular vesicles has been associated with cerebrovascular events in COVID-19 patients by compromising the function of the blood-brain barrier[150,151]. Endothelial MPs and PS-positive filopods support the formation of the prothrombinase complex along with activated coagulation factors Va and Xa and thereby catalyze a pivotal step of the coagulation cascade, that is, the proteolytic activation of thrombin [27,152]. In parallel, exposure of subendothelial tissues and tissue factor (TF) to plasma procoagulants triggers the extrinsic coagulation pathway[132]. Damaged endothelial cells sustain this hypercoagulable milieu and promote platelet aggregation by secreting large amounts of factor VIII and von Willebrand factor (VWF), respectively [153-156]. Thrombotic microangiopathy is further exacerbated due to relative deficiency in metalloproteinase ADAM metallopeptidase with thrombospondin type 1 motif 13 activity in COVID-19 patients, which leads to insufficient VWF cleavage and enhanced platelet-vessel wall interactions[157]. Numerous other endothelial dysfunction markers, such as circulating endothelial cells, soluble (s)Eselectin, soluble thrombomodulin, and soluble intercellular adhesion molecule 1 are significantly increased in COVID-19 patients receiving ICU care [153,155,158]. Furthermore, critically ill patients demonstrate functional exhaustion of natural anticoagulants (protein C, protein S, and antithrombin) and develop anti-phospholipid antibodies at high rates [154]. This procoagulant state is accompanied by suboptimal fibrinolytic potential and remodeling of the clot structure, which displays a denser fibrin network as well as thinner and shorter fibrin fibers. In severe COVID-19, hypofibrinolysis is dictated by an upregulation of plasminogen activator inhibitor-1 and its stabilizing cofactor vitronectin, which reduce plasminogen generation, despite concurrent elevations of tissue plasminogen activator[159].

Immunothrombosis illustrates the intricate cross-talk between the innate immune system and the coagulation pathway, which aims to locally contain an infection by facilitating recognition and eradication of invading pathogens. An aberrant activation of immunothrombosis is associated with severe thrombotic complications in SARS-CoV-2 infection [160]. Endothelium expresses a variety of adhesion and chemoattractant molecules, promoting the recruitment of monocytes and neutrophils at sites of injury. The release of DAMPs and PAMPs stimulates innate immune cells, which subsequently express TF and pro-inflammatory mediators in large amounts and precipitates the formation of

neutrophil extracellular traps (NETs)[156]. NETs are web-like structures consisting of DNA complexed with histones, bactericidal enzymes, complement factors, and coagulants. NETosis is a potent driver of immunothrombosis in COVID-19 and contributes to the procoagulant state in IBD through various mechanisms: cleavage of natural anticoagulants, direct activation of the contact-dependent and extrinsic pathways of coagulation, and aggregation of platelets[156,161]. In turn, activated platelets overexpress P-selectin and other adhesion molecules, which enhance their interactions with monocytes and NETs [162]. Through the secretion of their intracellular prothrombotic granule substances and externalization of their PS-rich membrane, platelets trigger and sustain thromboinflammation[131].

There is evidence that this vicious cycle could be further aggravated by gut barrier dysfunction. In particular, Oliva et al [163] reported that serum LPS and zonulin were increased in hospitalized COVID-19 patients and showed that endotoxemia was an independent predictor of in-hospital thrombotic complications. Zonulin is involved in the modulation of gut permeability by orchestrating the disassembly of intercellular TJ[53]. In this regard, serum LPS positively correlated with zonulin, supporting that low-grade endotoxemia precipitating thrombosis in COVID-19 originates from the gut [163]. In addition, thrombogenesis in patients with endotoxemia was associated with the TLR4dependent activation of platelets[163]. In agreement, LPS/TLR4 and LPS/TLR2 downstream signaling pathways have been previously implicated in procoagulant conditions by promoting endothelial cell and platelet activation[164]. Several other bacterial lipoproteins, such as Pam³Cys and peptidoglycanassociated lipoprotein, promote vascular leakage and thrombus formation through TLR2 activation [165]. In a murine model, microbial translocation and release of related patterns as a result of gut barrier leakage enhanced platelet pro-aggregating capacity and thrombus growth by inducing VWF synthesis; this effect was mediated via TLR2 activation in the hepatic endothelium [166]. Restoration of endothelial function through L-arginine supplementation was followed by improved outcomes in hospitalized patients with COVID-19[167]. L-arginine, a semi-essential amino acid, stimulates endothelium relaxation by serving as a substrate for the synthesis of nitric oxide by NOS. It also plays a role in immunomodulation by controlling T cell proliferation[168]. The importance of gut microbiome homeostasis in preventing immunothrombosis and inflammatory damage was evidenced in a model of acute mesenteric ischemia/reperfusion injury. More specifically, commensal bacteria mitigated LPSinduced NETosis by providing tonic stimulation of the neutrophil-intrinsic TLR4 downstream signaling [169]. Furthermore, perturbation of intestinal microflora due to SARS-CoV-2 infection and the ensuing depletion of SCFAs might enhance thromboinflammation by eliminating the inhibitory effects of butyrate on the LPS/TLR4/NF-xB pro-inflammatory pathway [170]. Alternatively, dysregulation of microorganisms involved in the metabolism of trimethylamine and its metabolite TMA N-oxide (TMAO) could contribute to the hypercoagulative state in CAC[171]. TMA is generated by gut bacterial metabolism of choline, carnitine, and betaine and is rapidly converted into TMAO in the liver. TMAO is a recognized risk factor for clotting events by enhancing platelet hyperresponsiveness, endothelial dysfunction, and NLRP3 inflammasome activation[171,172]. Indeed, a recent study revealed significant alterations in metabolites pertaining to the choline/TMAO and carnitine/TMAO pathways in COVID-19 patients; these aberrations were associated with disease symptoms and severity^[173].

GUT BARRIER DYSFUNCTION: AN UNDERAPPRECIATED DRIVER OF SYSTEMIC INFLAMMATION

The critical form of COVID-19 is governed by multi-layered immune system dysregulation and hyperinflammatory responses. Overexpression of pro-inflammatory cytokines (IL-1, IL-6, TNF-α) and chemokines (MCP-1, MIP-1β, CCL5), as well as excessive oxidative stress and hyperactivation of the complement (C3a) and coagulation (d-dimer) systems are hallmarks of severe COVID-19[55,174]. In parallel, SARS-CoV-2 infection is accompanied by a tremendous influx of microbial components and metabolites into the systemic circulation due to intestinal barrier dysfunction. Several studies have shown that critically ill patients with COVID-19 present with endotoxemia and elevated plasma levels of zonulin and LPS-binding protein (LBP), which are surrogate markers of leaky gut[55,59,108,174-177]. Additional indices of disrupted intestinal permeability, such as fatty acid binding protein 2, an intracellular protein specifically expressed in IECs, and peptidoglycan, a core element of the cell wall in Gram-positive bacteria, were also increased in COVID-19 cases [178]. Furthermore, patients with severe COVID-19 as well as those with SARS-CoV-2 post-acute sequelae (PASC) had higher circulating levels of β -glucan, a fungal cell wall polysaccharide, indicating persistent fungal translocation [55,175,179]. Interestingly, the presence of circulating bacteriome in critically ill COVID-19 patients without evidence of secondary infections further corroborates the assumption of gut-derived bacterial translocation in severe SARS-CoV-2-infection[175]. Essentially, exuberant translocation precipitates microbial-mediated myeloid activation as demonstrated by increased serum levels of sCD14 and myeloperoxidase, which correspond to monocyte and neutrophil inflammation, respectively [55,174]. Multiple studies have revealed strong and unequivocal correlations between intestinal barrier dysfunction and biomarkers of inflammation and/or immunological activation, implicating a detrimental role of gut barrier defects in triggering or enhancing hyper-inflammatory responses in severe SARS-CoV-2 infection [55,59,108,163,



174,177]. More specifically, Giron *et al* [55] reported that LBP, β -glucan, and zonulin levels were all significantly associated with higher systemic levels of IL-6. During PASC, β -glucan could directly stimulate leukocytes and promote inflammation by binding to the Dectin-1 receptor and activating NFκB pathway[179]. In hospitalized COVID-19 patients with cardiac involvement, NLRP3 inflammasome activation and subsequent IL-18 and IL-1Ra secretion were linked with circulating LBP levels[108]. In line with this, Sun et al [59] demonstrated that plasma levels of LBP were associated with inflammation biomarkers (CRP, IL-6, IL-8) as well as with changes in relative frequencies of lymphocytes and neutrophils. Endotoxemia was significantly associated with TNF-α, CCL5, and MIP-1β in another cohort of COVID-19 inpatients, whereas sCD14 was negatively associated with TGF- β [174].

Furthermore, gut microbiome alterations, which become more pronounced as disease progresses, showed solid correlations with markers of inflammation and tissue injury. In particular, gut dysbiosis coincided with derangements in the serum levels of IL-10, TNF- α , CRP, erythrocyte sedimentation rate, aspartate aminotransferase, and lactate dehydrogenase in patients with COVID-19[60]. A negative correlation was found between CRP and the symbiotic microorganisms Faecalibacterium prausnitzii and Clostridium butyricum, which perish in critically ill patients[60]. The abundance of commensal bacteria (Lachnospiraceae, Eubacterium ventriosum, Faecalibacterium prausnitzii) was followed by an increased number of CD4+ T cells, CD8+ T cells, and NK cells[69]. Conversely, opportunistic pathogens, which are commonly overrepresented in the COVID-19-related gut microbiome, such as Bacteroides dorei and Akkermansia muciniphila, were positively correlated with pro-inflammatory cytokines, including IL-1β, IL-6, and IL-8[69]. Accordingly, the predominance of the emerging pathogen, Burkholderia contaminans, was accompanied by T cell anergy and complement activation [59]. In parallel, the prevalence of pathogenic species in SARS-CoV-2-induced gut dysbiosis predisposes to bacteremia, leading to secondary microbial bloodstream infections, and thus, to an increased risk of serious complications[178, 180]. It should be noted that dysbiosis induces multifaceted derangements in microbial-mediated metabolic functions, including regulation of amino acid, lipid, and carbohydrate metabolism, that could further contribute to worsening outcomes. In fact, severe COVID-19 disease is characterized by a drastic shift in the composition of gut-modulated biologically active molecules that engage in inflammation signaling and immune system activation[55,59]. The long-lasting alteration of the microbial signature due to SARS-CoV-2, characterized by the predominance of pathogenic species and activation of proinflammatory pathways, perpetuates intestinal inflammation and could lead to long-term complications, such as tumorigenesis and colorectal cancer[60,181]. Systemic immune activation in COVID-19, which is at least partially fueled by leaky gut, has been linked to all of the severe manifestations of the disease, including cytokine storm syndrome, ARDS, renal failure, cardiovascular events, thromboembolic disease, and neurological manifestations[182]. The co-existence of intestinal barrier dysfunction in individuals with underlying diseases, such as obesity, diabetes, colorectal cancer, or immunosuppression, could exacerbate endotoxemia and the consequent immune system overactivation, predisposing to a more severe disease course[182,183].

CONCLUSION

Although the respiratory tract is the main target organ of SARS-CoV-2 infection, severe COVID-19 is considered a complex disorder affecting multiple systems. The development of GI-related symptoms, long-lasting fecal shedding of viral RNA, and identification of the virus in human intestinal tissues have brought to the spotlight the potential effects of the GI system in COVID-19 pathophysiology. The integrity of the intestinal barrier is a *sine qua non* for the accomplishment of the diverse digestive and immunomodulatory functions of the GI tract. In this setting, SARS-CoV-2 is capable of inducing deleterious effects on the gatekeepers of paracellular transport, *i.e.*, TJs, as well as on intestinal epithelial and endothelial cells through direct ACE2-dependent or indirect immune-mediated mechanisms, or both. Apart from dismantling the mechanical structures of the mucosa, COVID-19 is accompanied by profound alterations of the intestinal microflora at taxonomic and functional levels that are associated with disease severity and the host's immune system activation. Furthermore, SARS-CoV-2 hijacks innate immune responses, principally through interference with IFN signaling, and, thus, leads to inappropriate trafficking and activation of virus-specific T and B cells. In turn, this multi-layered disruption of the gut barrier can exacerbate the underlying immunopathology of COVID-19 or precipitate serious complications.

The gut and the lungs, albeit anatomically distinct, participate in a bi-directional immunological crosstalk through their respective microbes and immune cells. A well-tuned intestinal barrier harboring a diverse community of commensal microorganisms is pivotal in modulating lung immune responses and the lung microbiome. Therefore, gut dysbiosis impairs the ability to prime vigorous immune responses in the respiratory tract to effectively contain viral infections such as SARS-CoV-2. Moreover, the gut vascular bed provides an enormous endothelial surface susceptible to SARS-CoV-2-mediated injury. Disease-activated and apoptotic endothelial cells are potent drivers of neutrophil/monocyte recruitment, platelet aggregation, and coagulation activation. In parallel, dysbiosis, endotoxemia, and systemic hyperimmune reactions shape a procoagulant state within the gut microvasculature that



possibly contributes to extraintestinal thrombotic complications or ARDS pathogenesis, which are common manifestations of severe COVID-19. A compromised gut barrier allows an excessive influx of intraluminal microbiota into otherwise sterile extraintestinal compartments. This systemic dissemination of microbial constituents and metabolites contributes, to a certain extent, to immune system activation and hyper-inflammatory responses that govern the severe form of COVID-19. Preexisting comorbidities plagued with impaired intestinal permeability, such as obesity, diabetes, cirrhosis, and autoimmune disorders, might act synergistically with SARS-CoV-2 to further aggravate endotoxemia and endotoxin-mediated immune activation, predisposing to a more complicated disease course. Modulation of the gut barrier function emerges as a promising intervention to prevent or alleviate severe COVID-19 and related complications.

FOOTNOTES

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Country/Territory of origin: Greece

ORCID number: Efthymios P Tsounis 0000-0003-2797-5070; Christos Triantos 0000-0003-3094-8209; Christos Konstantakis 0000-0001-5834-9182; Markos Marangos 0000-0001-5030-2398; Stelios F Assimakopoulos 0000-0002-6901-3681.

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REFERENCES

- Osuchowski MF, Winkler MS, Skirecki T, Cajander S, Shankar-Hari M, Lachmann G, Monneret G, Venet F, Bauer M, 1 Brunkhorst FM, Weis S, Garcia-Salido A, Kox M, Cavaillon JM, Uhle F, Weigand MA, Flohé SB, Wiersinga WJ, Almansa R, de la Fuente A, Martin-Loeches I, Meisel C, Spinetti T, Schefold JC, Cilloniz C, Torres A, Giamarellos-Bourboulis EJ, Ferrer R, Girardis M, Cossarizza A, Netea MG, van der Poll T, Bermejo-Martín JF, Rubio I. The COVID-19 puzzle: deciphering pathophysiology and phenotypes of a new disease entity. Lancet Respir Med 2021; 9: 622-642 [PMID: 33965003 DOI: 10.1016/S2213-2600(21)00218-6]
- Yang L, Liu S, Liu J, Zhang Z, Wan X, Huang B, Chen Y, Zhang Y. COVID-19: immunopathogenesis and 2 Immunotherapeutics. Signal Transduct Target Ther 2020; 5: 128 [PMID: 32712629 DOI: 10.1038/s41392-020-00243-2]
- Hu B, Huang S, Yin L. The cytokine storm and COVID-19. J Med Virol 2021; 93: 250-256 [PMID: 32592501 DOI: 3 10.1002/imv.262321
- König J, Wells J, Cani PD, García-Ródenas CL, MacDonald T, Mercenier A, Whyte J, Troost F, Brummer RJ. Human Intestinal Barrier Function in Health and Disease. Clin Transl Gastroenterol 2016; 7: e196 [PMID: 27763627 DOI: 10.1038/ctg.2016.54]
- Assimakopoulos SF, Scopa CD, Vagianos CE. Pathophysiology of increased intestinal permeability in obstructive 5 jaundice. World J Gastroenterol 2007; 13: 6458-6464 [PMID: 18161914 DOI: 10.3748/wjg.v13.i48.6458]
- Assimakopoulos SF, Triantos C, Maroulis I, Gogos C. The Role of the Gut Barrier Function in Health and Disease. 6 Gastroenterology Res 2018; 11: 261-263 [PMID: 30116424 DOI: 10.14740/gr1053w]
- 7 Jackson CB, Farzan M, Chen B, Choe H. Mechanisms of SARS-CoV-2 entry into cells. Nat Rev Mol Cell Biol 2022; 23: 3-20 [PMID: 34611326 DOI: 10.1038/s41580-021-00418-x]
- Rotondo JC, Martini F, Maritati M, Caselli E, Gallenga CE, Guarino M, De Giorgio R, Mazziotta C, Tramarin ML, 8 Badiale G, Tognon M, Contini C. Advanced Molecular and Immunological Diagnostic Methods to Detect SARS-CoV-2 Infection. Microorganisms 2022; 10 [PMID: 35744711 DOI: 10.3390/microorganisms10061193]
- Hikmet F, Méar L, Edvinsson Å, Micke P, Uhlén M, Lindskog C. The protein expression profile of ACE2 in human tissues. Mol Syst Biol 2020; 16: e9610 [PMID: 32715618 DOI: 10.15252/msb.20209610]
- 10 Dorrell RD, Dougherty MK, Barash EL, Lichtig AE, Clayton SB, Jensen ET. Gastrointestinal and hepatic manifestations of COVID-19: A systematic review and meta-analysis. JGH Open 2021; 5: 107-115 [PMID: 33363257 DOI: 10.1002/jgh3.12456]
- 11 Mao R, Qiu Y, He JS, Tan JY, Li XH, Liang J, Shen J, Zhu LR, Chen Y, Iacucci M, Ng SC, Ghosh S, Chen MH.



Manifestations and prognosis of gastrointestinal and liver involvement in patients with COVID-19: a systematic review and meta-analysis. Lancet Gastroenterol Hepatol 2020; 5: 667-678 [PMID: 32405603 DOI: 10.1016/S2468-1253(20)30126-6]

- 12 Sultan S, Altayar O, Siddique SM, Davitkov P, Feuerstein JD, Lim JK, Falck-Ytter Y, El-Serag HB; AGA Institute. AGA Institute Rapid Review of the Gastrointestinal and Liver Manifestations of COVID-19, Meta-Analysis of International Data, and Recommendations for the Consultative Management of Patients with COVID-19. Gastroenterology 2020; 159: 320-334.e27 [PMID: 32407808 DOI: 10.1053/j.gastro.2020.05.001]
- 13 Henry BM, de Oliveira MHS, Benoit J, Lippi G. Gastrointestinal symptoms associated with severity of coronavirus disease 2019 (COVID-19): a pooled analysis. Intern Emerg Med 2020; 15: 857-859 [PMID: 32303970 DOI: 10.1007/s11739-020-02329-9
- Zeng W, Qi K, Ye M, Zheng L, Liu X, Hu S, Zhang W, Tang W, Xu J, Yu D, Wei Y. Gastrointestinal symptoms are 14 associated with severity of coronavirus disease 2019: a systematic review and meta-analysis. Eur J Gastroenterol Hepatol 2022; 34: 168-176 [PMID: 33470700 DOI: 10.1097/MEG.000000000002072]
- Hayashi Y, Wagatsuma K, Nojima M, Yamakawa T, Ichimiya T, Yokoyama Y, Kazama T, Hirayama D, Nakase H. The 15 characteristics of gastrointestinal symptoms in patients with severe COVID-19: a systematic review and meta-analysis. J Gastroenterol 2021; 56: 409-420 [PMID: 33759041 DOI: 10.1007/s00535-021-01778-z]
- Shehab M, Alrashed F, Shuaibi S, Alajmi D, Barkun A. Gastroenterological and hepatic manifestations of patients with 16 COVID-19, prevalence, mortality by country, and intensive care admission rate: systematic review and meta-analysis. BMJ Open Gastroenterol 2021; 8 [PMID: 33664052 DOI: 10.1136/bmjgast-2020-000571]
- 17 Wang Y, Li Y, Zhang Y, Liu Y. Are gastrointestinal symptoms associated with higher risk of Mortality in COVID-19 patients? BMC Gastroenterol 2022; 22: 106 [PMID: 35255816 DOI: 10.1186/s12876-022-02132-0]
- 18 Livanos AE, Jha D, Cossarini F, Gonzalez-Reiche AS, Tokuyama M, Aydillo T, Parigi TL, Ladinsky MS, Ramos I, Dunleavy K, Lee B, Dixon RE, Chen ST, Martinez-Delgado G, Nagula S, Bruce EA, Ko HM, Glicksberg BS, Nadkarni G, Pujadas E, Reidy J, Naymagon S, Grinspan A, Ahmad J, Tankelevich M, Bram Y, Gordon R, Sharma K, Houldsworth J, Britton GJ, Chen-Liaw A, Spindler MP, Plitt T, Wang P, Cerutti A, Faith JJ, Colombel JF, Kenigsberg E, Argmann C, Merad M, Gnjatic S, Harpaz N, Danese S, Cordon-Cardo C, Rahman A, Schwartz RE, Kumta NA, Aghemo A, Bjorkman PJ, Petralia F, van Bakel H, Garcia-Sastre A, Mehandru S. Intestinal Host Response to SARS-CoV-2 Infection and COVID-19 Outcomes in Patients With Gastrointestinal Symptoms. Gastroenterology 2021; 160: 2435-2450.e34 [PMID: 33676971 DOI: 10.1053/j.gastro.2021.02.056]
- 19 Wang W, Xu Y, Gao R, Lu R, Han K, Wu G, Tan W. Detection of SARS-CoV-2 in Different Types of Clinical Specimens. JAMA 2020; 323: 1843-1844 [PMID: 32159775 DOI: 10.1001/jama.2020.3786]
- 20 Zhang Y, Cen M, Hu M, Du L, Hu W, Kim JJ, Dai N. Prevalence and Persistent Shedding of Fecal SARS-CoV-2 RNA in Patients With COVID-19 Infection: A Systematic Review and Meta-analysis. Clin Transl Gastroenterol 2021; 12: e00343 [PMID: 33835096 DOI: 10.14309/ctg.00000000000343]
- Wölfel R, Corman VM, Guggemos W, Seilmaier M, Zange S, Müller MA, Niemeyer D, Jones TC, Vollmar P, Rothe C, 21 Hoelscher M, Bleicker T, Brünink S, Schneider J, Ehmann R, Zwirglmaier K, Drosten C, Wendtner C. Virological assessment of hospitalized patients with COVID-2019. Nature 2020; 581: 465-469 [PMID: 32235945 DOI: 10.1038/s41586-020-2196-x
- Effenberger M, Grabherr F, Mayr L, Schwaerzler J, Nairz M, Seifert M, Hilbe R, Seiwald S, Scholl-Buergi S, Fritsche G, 22 Bellmann-Weiler R, Weiss G, Müller T, Adolph TE, Tilg H. Faecal calprotectin indicates intestinal inflammation in COVID-19. Gut 2020; 69: 1543-1544 [PMID: 32312790 DOI: 10.1136/gutjnl-2020-321388]
- 23 Chan KH, Sridhar S, Zhang RR, Chu H, Fung AY, Chan G, Chan JF, To KK, Hung IF, Cheng VC, Yuen KY. Factors affecting stability and infectivity of SARS-CoV-2. J Hosp Infect 2020; 106: 226-231 [PMID: 32652214 DOI: 10.1016/j.jhin.2020.07.009
- 24 Esseili MA. Great escape: how infectious SARS-CoV-2 avoids inactivation by gastric acidity and intestinal bile. Gut 2022 [PMID: 35672040 DOI: 10.1136]
- 25 Bushman FD, McCormick K, Sherrill-Mix S. Virus structures constrain transmission modes. Nat Microbiol 2019; 4: 1778-1780 [PMID: 31358983 DOI: 10.1038/s41564-019-0523-5]
- Deng W, Bao L, Gao H, Xiang Z, Qu Y, Song Z, Gong S, Liu J, Yu P, Qi F, Xu Y, Li F, Xiao C, Lv Q, Xue J, Wei Q, Liu 26 M, Wang G, Wang S, Yu H, Chen T, Liu X, Zhao W, Han Y, Qin C. Ocular conjunctival inoculation of SARS-CoV-2 can cause mild COVID-19 in rhesus macaques. Nat Commun 2020; 11: 4400 [PMID: 32879306 DOI: 10.1038/s41467-020-18149-6]
- Wu X, Jing H, Wang C, Wang Y, Zuo N, Jiang T, Novakovic VA, Shi J. Intestinal Damage in COVID-19: SARS-CoV-2 27 Infection and Intestinal Thrombosis. Front Microbiol 2022; 13: 860931 [PMID: 35391725 DOI: 10.3389/fmicb.2022.860931]
- 28 Hu F, Chen F, Ou Z, Fan Q, Tan X, Wang Y, Pan Y, Ke B, Li L, Guan Y, Mo X, Wang J, Luo C, Wen X, Li M, Ren P, Ke C, Li J, Lei C, Tang X, Li F. A compromised specific humoral immune response against the SARS-CoV-2 receptorbinding domain is related to viral persistence and periodic shedding in the gastrointestinal tract. Cell Mol Immunol 2020; 17: 1119-1125 [PMID: 33037400 DOI: 10.1038/s41423-020-00550-2]
- Vancamelbeke M, Vermeire S. The intestinal barrier: a fundamental role in health and disease. Expert Rev Gastroenterol Hepatol 2017; 11: 821-834 [PMID: 28650209 DOI: 10.1080/17474124.2017.1343143]
- Assimakopoulos SF, Papageorgiou I, Charonis A. Enterocytes' tight junctions: From molecules to diseases. World J 30 Gastrointest Pathophysiol 2011; 2: 123-137 [PMID: 22184542 DOI: 10.4291/wjgp.v2.i6.123]
- Meng W, Takeichi M. Adherens junction: molecular architecture and regulation. Cold Spring Harb Perspect Biol 2009; 1: 31 a002899 [PMID: 20457565 DOI: 10.1101/cshperspect.a002899]
- Schroeder BO. Fight them or feed them: how the intestinal mucus layer manages the gut microbiota. Gastroenterol Rep 32 (Oxf) 2019; 7: 3-12 [PMID: 30792861 DOI: 10.1093/gastro/goy052]
- Jandhyala SM, Talukdar R, Subramanyam C, Vuyyuru H, Sasikala M, Nageshwar Reddy D. Role of the normal gut microbiota. World J Gastroenterol 2015; 21: 8787-8803 [PMID: 26269668 DOI: 10.3748/wjg.v21.i29.8787]



- Aggeletopoulou I, Konstantakis C, Assimakopoulos SF, Triantos C. The role of the gut microbiota in the treatment of 34 inflammatory bowel diseases. Microb Pathog 2019; 137: 103774 [PMID: 31586663 DOI: 10.1016/j.micpath.2019.103774]
- 35 Okumura R, Takeda K. Maintenance of gut homeostasis by the mucosal immune system. Proc Jpn Acad Ser B Phys Biol Sci 2016; 92: 423-435 [PMID: 27840390 DOI: 10.2183/pjab.92.423]
- Lehmann M, Allers K, Heldt C, Meinhardt J, Schmidt F, Rodriguez-Sillke Y, Kunkel D, Schumann M, Böttcher C, Stahl-36 Hennig C, Elezkurtaj S, Bojarski C, Radbruch H, Corman VM, Schneider T, Loddenkemper C, Moos V, Weidinger C, Kühl AA, Siegmund B. Human small intestinal infection by SARS-CoV-2 is characterized by a mucosal infiltration with activated CD8(+) T cells. Mucosal Immunol 2021; 14: 1381-1392 [PMID: 34420043 DOI: 10.1038/s41385-021-00437-z]
- 37 Yu W, Ou X, Liu X, Zhang S, Gao X, Cheng H, Zhu B, Yan J. ACE2 contributes to the maintenance of mouse epithelial barrier function. Biochem Biophys Res Commun 2020; 533: 1276-1282 [PMID: 33097186 DOI: 10.1016/j.bbrc.2020.10.002
- Duan Y, Prasad R, Feng D, Beli E, Li Calzi S, Longhini ALF, Lamendella R, Floyd JL, Dupont M, Noothi SK, Sreejit G, Athmanathan B, Wright J, Jensen AR, Oudit GY, Markel TA, Nagareddy PR, Obukhov AG, Grant MB. Bone Marrow-Derived Cells Restore Functional Integrity of the Gut Epithelial and Vascular Barriers in a Model of Diabetes and ACE2 Deficiency. Circ Res 2019; 125: 969-988 [PMID: 31610731 DOI: 10.1161/CIRCRESAHA.119.315743]
- 39 Biering SB, Sarnik SA, Wang E, Zengel JR, Leist SR, Schäfer A, Sathyan V, Hawkins P, Okuda K, Tau C, Jangid AR, Duffy CV, Wei J, Gilmore RC, Alfajaro MM, Strine MS, Nguyenla X, Van Dis E, Catamura C, Yamashiro LH, Belk JA, Begeman A, Stark JC, Shon DJ, Fox DM, Ezzatpour S, Huang E, Olegario N, Rustagi A, Volmer AS, Livraghi-Butrico A, Wehri E, Behringer RR, Cheon DJ, Schaletzky J, Aguilar HC, Puschnik AS, Button B, Pinsky BA, Blish CA, Baric RS, O'Neal WK, Bertozzi CR, Wilen CB, Boucher RC, Carette JE, Stanley SA, Harris E, Konermann S, Hsu PD. Genomewide bidirectional CRISPR screens identify mucins as host factors modulating SARS-CoV-2 infection. Nat Genet 2022; 54: 1078-1089 [PMID: 35879412 DOI: 10.1038/s41588-022-01131-x]
- 40 Zihni C, Mills C, Matter K, Balda MS. Tight junctions: from simple barriers to multifunctional molecular gates. Nat Rev Mol Cell Biol 2016; 17: 564-580 [PMID: 27353478 DOI: 10.1038/nrm.2016.80]
- Tian W, Zhang N, Jin R, Feng Y, Wang S, Gao S, Gao R, Wu G, Tian D, Tan W, Chen Y, Gao GF, Wong CCL. Immune 41 suppression in the early stage of COVID-19 disease. Nat Commun 2020; 11: 5859 [PMID: 33203833 DOI: 10.1038/s41467-020-19706-9
- 42 Yang RC, Huang K, Zhang HP, Li L, Zhang YF, Tan C, Chen HC, Jin ML, Wang XR. SARS-CoV-2 productively infects human brain microvascular endothelial cells. J Neuroinflammation 2022; 19: 149 [PMID: 35705998 DOI: 10.1186/s12974-022-02514-x
- De Maio F, Lo Cascio E, Babini G, Sali M, Della Longa S, Tilocca B, Roncada P, Arcovito A, Sanguinetti M, Scambia G, 43 Urbani A. Improved binding of SARS-CoV-2 Envelope protein to tight junction-associated PALS1 could play a key role in COVID-19 pathogenesis. Microbes Infect 2020; 22: 592-597 [PMID: 32891874 DOI: 10.1016/j.micinf.2020.08.006]
- 44 Chai J, Cai Y, Pang C, Wang L, McSweeney S, Shanklin J, Liu Q. Structural basis for SARS-CoV-2 envelope protein recognition of human cell junction protein PALS1. Nat Commun 2021; 12: 3433 [PMID: 34103506 DOI: 10.1038/s41467-021-23533-x]
- Teoh KT, Siu YL, Chan WL, Schlüter MA, Liu CJ, Peiris JS, Bruzzone R, Margolis B, Nal B. The SARS coronavirus E protein interacts with PALS1 and alters tight junction formation and epithelial morphogenesis. Mol Biol Cell 2010; 21: 3838-3852 [PMID: 20861307 DOI: 10.1091/mbc.E10-04-0338]
- Guo Y, Luo R, Wang Y, Deng P, Song T, Zhang M, Wang P, Zhang X, Cui K, Tao T, Li Z, Chen W, Zheng Y, Qin J. 46 SARS-CoV-2 induced intestinal responses with a biomimetic human gut-on-chip. Sci Bull (Beijing) 2021; 66: 783-793 [PMID: 33282445 DOI: 10.1016/j.scib.2020.11.015]
- Santa Cruz A, Mendes-Frias A, Oliveira AI, Dias L, Matos AR, Carvalho A, Capela C, Pedrosa J, Castro AG, Silvestre 47 R. Interleukin-6 Is a Biomarker for the Development of Fatal Severe Acute Respiratory Syndrome Coronavirus 2 Pneumonia. Front Immunol 2021; 12: 613422 [PMID: 33679753 DOI: 10.3389/fimmu.2021.613422]
- Alsaffar H, Martino N, Garrett JP, Adam AP. Interleukin-6 promotes a sustained loss of endothelial barrier function via 48 Janus kinase-mediated STAT3 phosphorylation and de novo protein synthesis. Am J Physiol Cell Physiol 2018; 314: C589-C602 [PMID: 29351406 DOI: 10.1152/ajpcell.00235.2017]
- 49 Assimakopoulos SF, Mastronikolis S, DE Lastic AL, Aretha D, Papageorgiou D, Chalkidi T, Oikonomou I, Triantos C, Mouzaki A, Marangos M. Intestinal Barrier Biomarker ZO1 and Endotoxin Are Increased in Blood of Patients With COVID-19-associated Pneumonia. In Vivo 2021; 35: 2483-2488 [PMID: 34182534 DOI: 10.21873/invivo.12528]
- 50 Hurley JC, Nowak P, Öhrmalm L, Gogos C, Armaganidis A, Giamarellos-Bourboulis EJ. Endotoxemia as a diagnostic tool for patients with suspected bacteremia caused by gram-negative organisms: a meta-analysis of 4 decades of studies. J Clin Microbiol 2015; 53: 1183-1191 [PMID: 25631796 DOI: 10.1128/JCM.03531-14]
- 51 Kuo WT, Zuo L, Odenwald MA, Madha S, Singh G, Gurniak CB, Abraham C, Turner JR. The Tight Junction Protein ZO-1 Is Dispensable for Barrier Function but Critical for Effective Mucosal Repair. Gastroenterology 2021; 161: 1924-1939 [PMID: 34478742 DOI: 10.1053/j.gastro.2021.08.047]
- 52 Tornavaca O, Chia M, Dufton N, Almagro LO, Conway DE, Randi AM, Schwartz MA, Matter K, Balda MS. ZO-1 controls endothelial adherens junctions, cell-cell tension, angiogenesis, and barrier formation. J Cell Biol 2015; 208: 821-838 [PMID: 25753039 DOI: 10.1083/jcb.201404140]
- Fasano A. Intestinal permeability and its regulation by zonulin: diagnostic and therapeutic implications. Clin 53 Gastroenterol Hepatol 2012; 10: 1096-1100 [PMID: 22902773 DOI: 10.1016/j.cgh.2012.08.012]
- Sapone A, de Magistris L, Pietzak M, Clemente MG, Tripathi A, Cucca F, Lampis R, Kryszak D, Cartenì M, Generoso M, 54 Iafusco D, Prisco F, Laghi F, Riegler G, Carratu R, Counts D, Fasano A. Zonulin upregulation is associated with increased gut permeability in subjects with type 1 diabetes and their relatives. Diabetes 2006; 55: 1443-1449 [PMID: 16644703 DOI: 10.2337/db05-15931
- 55 Giron LB. Dweep H. Yin X. Wang H. Damra M. Goldman AR. Gorman N. Palmer CS. Tang HY. Shaikh MW. Forsyth CB, Balk RA, Zilberstein NF, Liu Q, Kossenkov A, Keshavarzian A, Landay A, Abdel-Mohsen M. Plasma Markers of



Disrupted Gut Permeability in Severe COVID-19 Patients. Front Immunol 2021; 12: 686240 [PMID: 34177935 DOI: 10.3389/fimmu.2021.686240]

- Levy M, Kolodziejczyk AA, Thaiss CA, Elinav E. Dysbiosis and the immune system. Nat Rev Immunol 2017; 17: 219-56 232 [PMID: 28260787 DOI: 10.1038/nri.2017.7]
- 57 Reinold J, Farahpour F, Fehring C, Dolff S, Konik M, Korth J, van Baal L, Hoffmann D, Buer J, Witzke O, Westendorf AM, Kehrmann J. A Pro-Inflammatory Gut Microbiome Characterizes SARS-CoV-2 Infected Patients and a Reduction in the Connectivity of an Anti-Inflammatory Bacterial Network Associates With Severe COVID-19. Front Cell Infect Microbiol 2021; 11: 747816 [PMID: 34869058 DOI: 10.3389/fcimb.2021.747816]
- 58 Liu Q, Mak JWY, Su Q, Yeoh YK, Lui GC, Ng SSS, Zhang F, Li AYL, Lu W, Hui DS, Chan PK, Chan FKL, Ng SC. Gut microbiota dynamics in a prospective cohort of patients with post-acute COVID-19 syndrome. Gut 2022; 71: 544-552 [PMID: 35082169 DOI: 10.1136/gutjnl-2021-325989]
- 59 Sun Z, Song ZG, Liu C, Tan S, Lin S, Zhu J, Dai FH, Gao J, She JL, Mei Z, Lou T, Zheng JJ, Liu Y, He J, Zheng Y, Ding C, Qian F, Chen YM. Gut microbiome alterations and gut barrier dysfunction are associated with host immune homeostasis in COVID-19 patients. BMC Med 2022; 20: 24 [PMID: 35045853 DOI: 10.1186/s12916-021-02212-0]
- Yeoh YK, Zuo T, Lui GC, Zhang F, Liu Q, Li AY, Chung AC, Cheung CP, Tso EY, Fung KS, Chan V, Ling L, Joynt G, 60 Hui DS, Chow KM, Ng SSS, Li TC, Ng RW, Yip TC, Wong GL, Chan FK, Wong CK, Chan PK, Ng SC. Gut microbiota composition reflects disease severity and dysfunctional immune responses in patients with COVID-19. Gut 2021; 70: 698-706 [PMID: 33431578 DOI: 10.1136/gutjnl-2020-323020]
- 61 Hashimoto T, Perlot T, Rehman A, Trichereau J, Ishiguro H, Paolino M, Sigl V, Hanada T, Hanada R, Lipinski S, Wild B, Camargo SM, Singer D, Richter A, Kuba K, Fukamizu A, Schreiber S, Clevers H, Verrey F, Rosenstiel P, Penninger JM. ACE2 Links amino acid malnutrition to microbial ecology and intestinal inflammation. Nature 2012; 487: 477-481 [PMID: 22837003 DOI: 10.1038/nature11228]
- Zhao Y, Chen F, Wu W, Sun M, Bilotta AJ, Yao S, Xiao Y, Huang X, Eaves-Pyles TD, Golovko G, Fofanov Y, D'Souza 62 W, Zhao Q, Liu Z, Cong Y. GPR43 mediates microbiota metabolite SCFA regulation of antimicrobial peptide expression in intestinal epithelial cells via activation of mTOR and STAT3. Mucosal Immunol 2018; 11: 752-762 [PMID: 29411774 DOI: 10.1038/mi.2017.118]
- 63 Edwinson A, Yang L, Chen J, Grover M. Colonic expression of Ace2, the SARS-CoV-2 entry receptor, is suppressed by commensal human microbiota. Gut Microbes 2021; 13: 1984105 [PMID: 34632957 DOI: 10.1080/19490976.2021.1984105]
- Groves HT, Higham SL, Moffatt MF, Cox MJ, Tregoning JS. Respiratory Viral Infection Alters the Gut Microbiota by 64 Inducing Inappetence. mBio 2020; 11 [PMID: 32071269 DOI: 10.1128/mBio.03236-19]
- Guimarães Sousa S, Kleiton de Sousa A, Maria Carvalho Pereira C, Sofia Miranda Loiola Araújo A, de Aguiar 65 Magalhães D, Vieira de Brito T, Barbosa ALDR. SARS-CoV-2 infection causes intestinal cell damage: Role of interferon's imbalance. Cytokine 2022; 152: 155826 [PMID: 35158258 DOI: 10.1016/j.cyto.2022.155826]
- Deriu E, Boxx GM, He X, Pan C, Benavidez SD, Cen L, Rozengurt N, Shi W, Cheng G. Influenza Virus Affects 66 Intestinal Microbiota and Secondary Salmonella Infection in the Gut through Type I Interferons. PLoS Pathog 2016; 12: e1005572 [PMID: 27149619 DOI: 10.1371/journal.ppat.1005572]
- Gopal AB, Chakraborty S, Padhan PK, Barik A, Dixit P, Chakraborty D, Poirah I, Samal S, Sarkar A, Bhattacharyya A. Silent hypoxia in COVID-19: a gut microbiota connection. Curr Opin Physiol 2021; 23: 100456 [PMID: 34250324 DOI: 10.1016/j.cophys.2021.06.010]
- Sencio V, Machado MG, Trottein F. The lung-gut axis during viral respiratory infections: the impact of gut dysbiosis on 68 secondary disease outcomes. Mucosal Immunol 2021; 14: 296-304 [PMID: 33500564 DOI: 10.1038/s41385-020-00361-8]
- Farsi Y, Tahvildari A, Arbabi M, Vazife F, Sechi LA, Shahidi Bonjar AH, Jamshidi P, Nasiri MJ, Mirsaeidi M. Diagnostic, Prognostic, and Therapeutic Roles of Gut Microbiota in COVID-19: A Comprehensive Systematic Review. Front Cell Infect Microbiol 2022; 12: 804644 [PMID: 35310853 DOI: 10.3389/fcimb.2022.804644]
- 70 Gu S, Chen Y, Wu Z, Gao H, Lv L, Guo F, Zhang X, Luo R, Huang C, Lu H, Zheng B, Zhang J, Yan R, Zhang H, Jiang H, Xu Q, Guo J, Gong Y, Tang L, Li L. Alterations of the Gut Microbiota in Patients With Coronavirus Disease 2019 or H1N1 Influenza. Clin Infect Dis 2020; 71: 2669-2678 [PMID: 32497191 DOI: 10.1093/cid/ciaa709]
- Cao J, Wang C, Zhang Y, Lei G, Xu K, Zhao N, Lu J, Meng F, Yu L, Yan J, Bai C, Zhang S, Zhang N, Gong Y, Bi Y, 71 Shi Y, Chen Z, Dai L, Wang J, Yang P. Integrated gut virome and bacteriome dynamics in COVID-19 patients. Gut Microbes 2021; 13: 1-21 [PMID: 33678150 DOI: 10.1080/19490976.2021.1887722]
- Tamanai-Shacoori Z, Smida I, Bousarghin L, Loreal O, Meuric V, Fong SB, Bonnaure-Mallet M, Jolivet-Gougeon A. 72 Roseburia spp.: a marker of health? Future Microbiol 2017; 12: 157-170 [PMID: 28139139 DOI: 10.2217/fmb-2016-0130]
- Sokol H, Pigneur B, Watterlot L, Lakhdari O, Bermúdez-Humarán LG, Gratadoux JJ, Blugeon S, Bridonneau C, Furet JP, 73 Corthier G, Grangette C, Vasquez N, Pochart P, Trugnan G, Thomas G, Blottière HM, Doré J, Marteau P, Seksik P, Langella P. Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. Proc Natl Acad Sci U S A 2008; 105: 16731-16736 [PMID: 18936492 DOI: 10.1073/pnas.0804812105]
- Mukherjee A, Lordan C, Ross RP, Cotter PD. Gut microbes from the phylogenetically diverse genus Eubacterium and 74 their various contributions to gut health. Gut Microbes 2020; 12: 1802866 [PMID: 32835590 DOI: 10.1080/19490976.2020.1802866]
- 75 Konieczna P, Akdis CA, Quigley EM, Shanahan F, O'Mahony L. Portrait of an immunoregulatory Bifidobacterium. Gut Microbes 2012; 3: 261-266 [PMID: 22572827 DOI: 10.4161/gmic.20358]
- Yao S, Zhao Z, Wang W, Liu X. Bifidobacterium Longum: Protection against Inflammatory Bowel Disease. J Immunol 76 Res 2021; 2021: 8030297 [PMID: 34337079 DOI: 10.1155/2021/8030297]
- Yuan F, Ni H, Asche CV, Kim M, Walayat S, Ren J. Efficacy of Bifidobacterium infantis 35624 in patients with irritable 77 bowel syndrome: a meta-analysis. Curr Med Res Opin 2017; 33: 1191-1197 [PMID: 28166427 DOI: 10.1080/03007995.2017.1292230]



- 78 Gautier T, David-Le Gall S, Sweidan A, Tamanai-Shacoori Z, Jolivet-Gougeon A, Loréal O, Bousarghin L. Next-Generation Probiotics and Their Metabolites in COVID-19. Microorganisms 2021; 9 [PMID: 33925715 DOI: 10.3390/microorganisms9050941]
- 79 Png CW, Lindén SK, Gilshenan KS, Zoetendal EG, McSweeney CS, Sly LI, McGuckin MA, Florin TH. Mucolytic bacteria with increased prevalence in IBD mucosa augment in vitro utilization of mucin by other bacteria. Am J Gastroenterol 2010; 105: 2420-2428 [PMID: 20648002 DOI: 10.1038/ajg.2010.281]
- Henke MT, Kenny DJ, Cassilly CD, Vlamakis H, Xavier RJ, Clardy J. Ruminococcus gnavus, a member of the human 80 gut microbiome associated with Crohn's disease, produces an inflammatory polysaccharide. Proc Natl Acad Sci USA 2019; 116: 12672-12677 [PMID: 31182571 DOI: 10.1073/pnas.1904099116]
- 81 Moreira-Rosário A, Marques C, Pinheiro H, Araújo JR, Ribeiro P, Rocha R, Mota I, Pestana D, Ribeiro R, Pereira A, de Sousa MJ, Pereira-Leal J, de Sousa J, Morais J, Teixeira D, Rocha JC, Silvestre M, Príncipe N, Gatta N, Amado J, Santos L, Maltez F, Boquinhas A, de Sousa G, Germano N, Sarmento G, Granja C, Póvoa P, Faria A, Calhau C. Gut Microbiota Diversity and C-Reactive Protein Are Predictors of Disease Severity in COVID-19 Patients. Front Microbiol 2021; 12: 705020 [PMID: 34349747 DOI: 10.3389/fmicb.2021.705020]
- 82 Zuo T, Zhang F, Lui GCY, Yeoh YK, Li AYL, Zhan H, Wan Y, Chung ACK, Cheung CP, Chen N, Lai CKC, Chen Z, Tso EYK, Fung KSC, Chan V, Ling L, Joynt G, Hui DSC, Chan FKL, Chan PKS, Ng SC. Alterations in Gut Microbiota of Patients With COVID-19 During Time of Hospitalization. Gastroenterology 2020; 159: 944-955.e8 [PMID: 32442562 DOI: 10.1053/j.gastro.2020.05.048]
- Gaibani P, D'Amico F, Bartoletti M, Lombardo D, Rampelli S, Fornaro G, Coladonato S, Siniscalchi A, Re MC, Viale P, 83 Brigidi P, Turroni S, Giannella M. The Gut Microbiota of Critically Ill Patients With COVID-19. Front Cell Infect Microbiol 2021; 11: 670424 [PMID: 34268136 DOI: 10.3389/fcimb.2021.670424]
- Khan M, Mathew BJ, Gupta P, Garg G, Khadanga S, Vyas AK, Singh AK. Gut Dysbiosis and IL-21 Response in Patients with Severe COVID-19. Microorganisms 2021; 9 [PMID: 34199203 DOI: 10.3390/microorganisms9061292]
- Mazzarelli A, Giancola ML, Farina A, Marchioni L, Rueca M, Gruber CEM, Bartolini B, Ascoli Bartoli T, Maffongelli 85 G, Capobianchi MR, Ippolito G, Di Caro A, Nicastri E, Pazienza V; INMI COVID-19 study group. 16S rRNA gene sequencing of rectal swab in patients affected by COVID-19. PLoS One 2021; 16: e0247041 [PMID: 33596245 DOI: 10.1371/journal.pone.0247041]
- Rueca M, Fontana A, Bartolini B, Piselli P, Mazzarelli A, Copetti M, Binda E, Perri F, Gruber CEM, Nicastri E, 86 Marchioni L, Ippolito G, Capobianchi MR, Di Caro A, Pazienza V. Investigation of Nasal/Oropharyngeal Microbial Community of COVID-19 Patients by 16S rDNA Sequencing. Int J Environ Res Public Health 2021; 18 [PMID: 33672177 DOI: 10.3390/ijerph18042174]
- 87 He F, Zhang T, Xue K, Fang Z, Jiang G, Huang S, Li K, Gu Z, Shi H, Zhang Z, Zhu H, Lin L, Li J, Xiao F, Shan H, Yan R, Li X, Yan Z. Fecal multi-omics analysis reveals diverse molecular alterations of gut ecosystem in COVID-19 patients. Anal Chim Acta 2021; 1180: 338881 [PMID: 34538334 DOI: 10.1016/j.aca.2021.338881]
- Wang B, Zhang L, Wang Y, Dai T, Qin Z, Zhou F. Alterations in microbiota of patients with COVID-19: potential 88 mechanisms and therapeutic interventions. Signal Transduct Target Ther 2022; 7: 143 [PMID: 35487886 DOI: 10.1038/s41392-022-00986-01
- 89 Scott NA, Andrusaite A, Andersen P, Lawson M, Alcon-Giner C, Leclaire C, Caim S, Le Gall G, Shaw T, Connolly JPR, Roe AJ, Wessel H, Bravo-Blas A, Thomson CA, Kästele V, Wang P, Peterson DA, Bancroft A, Li X, Grencis R, Mowat AM, Hall LJ, Travis MA, Milling SWF, Mann ER. Antibiotics induce sustained dysregulation of intestinal T cell immunity by perturbing macrophage homeostasis. Sci Transl Med 2018; 10 [PMID: 30355800 DOI: 10.1126/scitranslmed.aao4755
- 90 Furusawa Y, Obata Y, Fukuda S, Endo TA, Nakato G, Takahashi D, Nakanishi Y, Uetake C, Kato K, Kato T, Takahashi M, Fukuda NN, Murakami S, Miyauchi E, Hino S, Atarashi K, Onawa S, Fujimura Y, Lockett T, Clarke JM, Topping DL, Tomita M, Hori S, Ohara O, Morita T, Koseki H, Kikuchi J, Honda K, Hase K, Ohno H. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. Nature 2013; 504: 446-450 [PMID: 24226770 DOI: 10.1038/nature12721]
- 91 Mazmanian SK, Liu CH, Tzianabos AO, Kasper DL. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. Cell 2005; 122: 107-118 [PMID: 16009137 DOI: 10.1016/j.cell.2005.05.007]
- Konieczna P, Ferstl R, Ziegler M, Frei R, Nehrbass D, Lauener RP, Akdis CA, O'Mahony L. Immunomodulation by 92 Bifidobacterium infantis 35624 in the murine lamina propria requires retinoic acid-dependent and independent mechanisms. PLoS One 2013; 8: e62617 [PMID: 23704880 DOI: 10.1371/journal.pone.0062617]
- 93 Verma R, Lee C, Jeun EJ, Yi J, Kim KS, Ghosh A, Byun S, Lee CG, Kang HJ, Kim GC, Jun CD, Jan G, Suh CH, Jung JY, Sprent J, Rudra D, De Castro C, Molinaro A, Surh CD, Im SH. Cell surface polysaccharides of Bifidobacterium bifidum induce the generation of Foxp3(+) regulatory T cells. Sci Immunol 2018; 3 [PMID: 30341145 DOI: 10.1126/sciimmunol.aat6975
- Kim M, Qie Y, Park J, Kim CH. Gut Microbial Metabolites Fuel Host Antibody Responses. Cell Host Microbe 2016; 20: 94 202-214 [PMID: 27476413 DOI: 10.1016/j.chom.2016.07.001]
- Xu C, Wang A, Marin M, Honnen W, Ramasamy S, Porter E, Subbian S, Pinter A, Melikyan GB, Lu W, Chang TL. Human Defensins Inhibit SARS-CoV-2 Infection by Blocking Viral Entry. Viruses 2021; 13 [PMID: 34206990 DOI: 10.3390/v13071246
- Wang C, Wang S, Li D, Wei DQ, Zhao J, Wang J. Human Intestinal Defensin 5 Inhibits SARS-CoV-2 Invasion by 96 Cloaking ACE2. Gastroenterology 2020; 159: 1145-1147.e4 [PMID: 32437749 DOI: 10.1053/j.gastro.2020.05.015]
- Al-Bayatee NT, Ad'hiah AH. Human beta-defensins 2 and 4 are dysregulated in patients with coronavirus disease 19. 97 Microb Pathog 2021; 160: 105205 [PMID: 34547411 DOI: 10.1016/j.micpath.2021.105205]
- 98 Stanifer ML, Kee C, Cortese M, Zumaran CM, Triana S, Mukenhirn M, Kraeusslich HG, Alexandrov T, Bartenschlager R, Boulant S. Critical Role of Type III Interferon in Controlling SARS-CoV-2 Infection in Human Intestinal Epithelial Cells. Cell Rep 2020; 32: 107863 [PMID: 32610043 DOI: 10.1016/j.celrep.2020.107863]
- 99 Metz-Zumaran C, Kee C, Doldan P, Guo C, Stanifer ML, Boulant S. Increased Sensitivity of SARS-CoV-2 to Type III



Interferon in Human Intestinal Epithelial Cells. J Virol 2022; 96: e0170521 [PMID: 35262371 DOI: 10.1128/jvi.01705-21]

- 100 Heuberger J, Trimpert J, Vladimirova D, Goosmann C, Lin M, Schmuck R, Mollenkopf HJ, Brinkmann V, Tacke F, Osterrieder N, Sigal M. Epithelial response to IFN-y promotes SARS-CoV-2 infection. EMBO Mol Med 2021; 13: e13191 [PMID: 33544398 DOI: 10.15252/emmm.202013191]
- 101 Karki R, Sharma BR, Tuladhar S, Williams EP, Zalduondo L, Samir P, Zheng M, Sundaram B, Banoth B, Malireddi RKS, Schreiner P, Neale G, Vogel P, Webby R, Jonsson CB, Kanneganti TD. Synergism of TNF-α and IFN-γ Triggers Inflammatory Cell Death, Tissue Damage, and Mortality in SARS-CoV-2 Infection and Cytokine Shock Syndromes. Cell 2021; 184: 149-168.e17 [PMID: 33278357 DOI: 10.1016/j.cell.2020.11.025]
- 102 Jiao L, Li H, Xu J, Yang M, Ma C, Li J, Zhao S, Wang H, Yang Y, Yu W, Wang J, Yang J, Long H, Gao J, Ding K, Wu D, Kuang D, Zhao Y, Liu J, Lu S, Liu H, Peng X. The Gastrointestinal Tract Is an Alternative Route for SARS-CoV-2 Infection in a Nonhuman Primate Model. Gastroenterology 2021; 160: 1647-1661 [PMID: 33307034 DOI: 10.1053/j.gastro.2020.12.001]
- 103 Tao W, Zhang G, Wang X, Guo M, Zeng W, Xu Z, Cao D, Pan A, Wang Y, Zhang K, Ma X, Chen Z, Jin T, Liu L, Weng J, Zhu S. Analysis of the intestinal microbiota in COVID-19 patients and its correlation with the inflammatory factor IL-18. Med Microecol 2020; 5: 100023 [PMID: 34173452 DOI: 10.1016/j.medmic.2020.100023]
- 104 Britton GJ, Chen-Liaw A, Cossarini F, Livanos AE, Spindler MP, Plitt T, Eggers J, Mogno I, Gonzalez-Reiche AS, Siu S, Tankelevich M, Grinspan LT, Dixon RE, Jha D, van de Guchte A, Khan Z, Martinez-Delgado G, Amanat F, Hoagland DA, tenOever BR, Dubinsky MC, Merad M, van Bakel H, Krammer F, Bongers G, Mehandru S, Faith JJ. Limited intestinal inflammation despite diarrhea, fecal viral RNA and SARS-CoV-2-specific IgA in patients with acute COVID-19. Sci Rep 2021; 11: 13308 [PMID: 34172783 DOI: 10.1038/s41598-021-92740-9]
- 105 Wang C, Xu J, Wang S, Pan S, Zhang J, Han Y, Huang M, Wu D, Yang Q, Yang X, Yang Y, Shu T, Zou X, Li R, Luo Y, Yao R, Wang Y, Qiu Y, Zhang DY, Yao Q, Yan Y, Zhou X, Shang Y. Imaging Mass Cytometric Analysis of Postmortem Tissues Reveals Dysregulated Immune Cell and Cytokine Responses in Multiple Organs of COVID-19 Patients. Front Microbiol 2020; 11: 600989 [PMID: 33424804 DOI: 10.3389/fmicb.2020.600989]
- Giamarellos-Bourboulis EJ, Netea MG, Rovina N, Akinosoglou K, Antoniadou A, Antonakos N, Damoraki G, 106 Gkavogianni T, Adami ME, Katsaounou P, Ntaganou M, Kyriakopoulou M, Dimopoulos G, Koutsodimitropoulos I, Velissaris D, Koufargyris P, Karageorgos A, Katrini K, Lekakis V, Lupse M, Kotsaki A, Renieris G, Theodoulou D, Panou V, Koukaki E, Koulouris N, Gogos C, Koutsoukou A. Complex Immune Dysregulation in COVID-19 Patients with Severe Respiratory Failure. Cell Host Microbe 2020; 27: 992-1000.e3 [PMID: 32320677 DOI: 10.1016/j.chom.2020.04.009
- 107 Trevelin SC, Pickering S, Todd K, Bishop C, Pitcher M, Garrido Mesa J, Montorsi L, Spada F, Petrov N, Green A, Shankar-Hari M, Neil SJD, Spencer J. Disrupted Peyer's Patch Microanatomy in COVID-19 Including Germinal Centre Atrophy Independent of Local Virus. Front Immunol 2022; 13: 838328 [PMID: 35251032 DOI: 10.3389/fimmu.2022.838328]
- Hoel H, Heggelund L, Reikvam DH, Stiksrud B, Ueland T, Michelsen AE, Otterdal K, Muller KE, Lind A, Muller F, 108 Dudman S, Aukrust P, Dyrhol-Riise AM, Holter JC, Trøseid M. Elevated markers of gut leakage and inflammasome activation in COVID-19 patients with cardiac involvement. J Intern Med 2021; 289: 523-531 [PMID: 32976665 DOI: 10.1111/joim.13178]
- Wurbel MA, McIntire MG, Dwyer P, Fiebiger E. CCL25/CCR9 interactions regulate large intestinal inflammation in a 109 murine model of acute colitis. PLoS One 2011; 6: e16442 [PMID: 21283540 DOI: 10.1371/journal.pone.0016442]
- Sterlin D, Mathian A, Miyara M, Mohr A, Anna F, Claër L, Quentric P, Fadlallah J, Devilliers H, Ghillani P, Gunn C, 110 Hockett R, Mudumba S, Guihot A, Luyt CE, Mayaux J, Beurton A, Fourati S, Bruel T, Schwartz O, Lacorte JM, Yssel H, Parizot C, Dorgham K, Charneau P, Amoura Z, Gorochov G. IgA dominates the early neutralizing antibody response to SARS-CoV-2. Sci Transl Med 2021; 13 [PMID: 33288662 DOI: 10.1126/scitranslmed.abd2223]
- Cervia C, Nilsson J, Zurbuchen Y, Valaperti A, Schreiner J, Wolfensberger A, Raeber ME, Adamo S, Weigang S, 111 Emmenegger M, Hasler S, Bosshard PP, De Cecco E, Bächli E, Rudiger A, Stüssi-Helbling M, Huber LC, Zinkernagel AS, Schaer DJ, Aguzzi A, Kochs G, Held U, Probst-Müller E, Rampini SK, Boyman O. Systemic and mucosal antibody responses specific to SARS-CoV-2 during mild vs severe COVID-19. J Allergy Clin Immunol 2021; 147: 545-557.e9 [PMID: 33221383 DOI: 10.1016/j.jaci.2020.10.040]
- 112 Roy K, Agarwal S, Banerjee R, Paul MK, Purbey PK. COVID-19 and gut immunomodulation. World J Gastroenterol 2021; 27: 7925-7942 [PMID: 35046621 DOI: 10.3748/wjg.v27.i46.7925]
- Bradley KC, Finsterbusch K, Schnepf D, Crotta S, Llorian M, Davidson S, Fuchs SY, Staeheli P, Wack A. Microbiota-113 Driven Tonic Interferon Signals in Lung Stromal Cells Protect from Influenza Virus Infection. Cell Rep 2019; 28: 245-256.e4 [PMID: 31269444 DOI: 10.1016/j.celrep.2019.05.105]
- Antunes KH, Fachi JL, de Paula R, da Silva EF, Pral LP, Dos Santos AÁ, Dias GBM, Vargas JE, Puga R, Mayer FQ, 114 Maito F, Zárate-Bladés CR, Ajami NJ, Sant'Ana MR, Candreva T, Rodrigues HG, Schmiele M, Silva Clerici MTP, Proença-Modena JL, Vieira AT, Mackay CR, Mansur D, Caballero MT, Marzec J, Li J, Wang X, Bell D, Polack FP, Kleeberger SR, Stein RT, Vinolo MAR, de Souza APD. Microbiota-derived acetate protects against respiratory syncytial virus infection through a GPR43-type 1 interferon response. Nat Commun 2019; 10: 3273 [PMID: 31332169 DOI: 10.1038/s41467-019-11152-6
- 115 Ichinohe T, Pang IK, Kumamoto Y, Peaper DR, Ho JH, Murray TS, Iwasaki A. Microbiota regulates immune defense against respiratory tract influenza A virus infection. Proc Natl Acad Sci USA 2011; 108: 5354-5359 [PMID: 21402903] DOI: 10.1073/pnas.1019378108]
- 116 Assimakopoulos SF, Triantos C, Thomopoulos K, Fligou F, Maroulis I, Marangos M, Gogos CA. Gut-origin sepsis in the critically ill patient: pathophysiology and treatment. Infection 2018; 46: 751-760 [PMID: 30003491 DOI: 10.1007/s15010-018-1178-5
- Abt MC, Osborne LC, Monticelli LA, Doering TA, Alenghat T, Sonnenberg GF, Paley MA, Antenus M, Williams KL, 117 Erikson J, Wherry EJ, Artis D. Commensal bacteria calibrate the activation threshold of innate antiviral immunity.



Immunity 2012; 37: 158-170 [PMID: 22705104 DOI: 10.1016/j.immuni.2012.04.011]

- 118 Thackray LB, Handley SA, Gorman MJ, Poddar S, Bagadia P, Briseño CG, Theisen DJ, Tan Q, Hykes BL Jr, Lin H, Lucas TM, Desai C, Gordon JI, Murphy KM, Virgin HW, Diamond MS. Oral Antibiotic Treatment of Mice Exacerbates the Disease Severity of Multiple Flavivirus Infections. Cell Rep 2018; 22: 3440-3453.e6 [PMID: 29590614 DOI: 10.1016/j.celrep.2018.03.001]
- 119 Steed AL, Christophi GP, Kaiko GE, Sun L, Goodwin VM, Jain U, Esaulova E, Artyomov MN, Morales DJ, Holtzman MJ, Boon ACM, Lenschow DJ, Stappenbeck TS. The microbial metabolite desaminotyrosine protects from influenza through type I interferon. Science 2017; 357: 498-502 [PMID: 28774928 DOI: 10.1126/science.aam5336]
- 120 Stefan KL, Kim MV, Iwasaki A, Kasper DL. Commensal Microbiota Modulation of Natural Resistance to Virus Infection. Cell 2020; 183: 1312-1324.e10 [PMID: 33212011 DOI: 10.1016/j.cell.2020.10.047]
- 121 Ganal SC, Sanos SL, Kallfass C, Oberle K, Johner C, Kirschning C, Lienenklaus S, Weiss S, Staeheli P, Aichele P, Diefenbach A. Priming of natural killer cells by nonmucosal mononuclear phagocytes requires instructive signals from commensal microbiota. Immunity 2012; 37: 171-186 [PMID: 22749822 DOI: 10.1016/j.immuni.2012.05.020]
- 122 Akamatsu MA, de Castro JT, Takano CY, Ho PL. Off balance: Interferons in COVID-19 Lung infections. EBioMedicine 2021; 73: 103642 [PMID: 34678609 DOI: 10.1016/j.ebiom.2021.103642]
- 123 Krämer B, Knoll R, Bonaguro L, ToVinh M, Raabe J, Astaburuaga-García R, Schulte-Schrepping J, Kaiser KM, Rieke GJ, Bischoff J, Monin MB, Hoffmeister C, Schlabe S, De Domenico E, Reusch N, Händler K, Reynolds G, Blüthgen N, Hack G, Finnemann C, Nischalke HD, Strassburg CP, Stephenson E, Su Y, Gardner L, Yuan D, Chen D, Goldman J, Rosenstiel P, Schmidt SV, Latz E, Hrusovsky K, Ball AJ, Johnson JM, Koenig PA, Schmidt FI, Haniffa M, Heath JR, Kümmerer BM, Keitel V, Jensen B, Stubbemann P, Kurth F, Sander LE, Sawitzki B; Deutsche COVID-19 OMICS Initiative (DeCOI), Aschenbrenner AC, Schultze JL, Nattermann J. Early IFN-a signatures and persistent dysfunction are distinguishing features of NK cells in severe COVID-19. Immunity 2021; 54: 2650-2669.e14 [PMID: 34592166 DOI: 10.1016/j.immuni.2021.09.002
- Yagi K, Huffnagle GB, Lukacs NW, Asai N. The Lung Microbiome during Health and Disease. Int J Mol Sci 2021; 22 124 [PMID: 34639212 DOI: 10.3390/ijms221910872]
- Sulaiman I, Chung M, Angel L, Tsay JJ, Wu BG, Yeung ST, Krolikowski K, Li Y, Duerr R, Schluger R, Thannickal SA, 125 Koide A, Rafeq S, Barnett C, Postelnicu R, Wang C, Banakis S, Pérez-Pérez L, Shen G, Jour G, Meyn P, Carpenito J, Liu X, Ji K, Collazo D, Labarbiera A, Amoroso N, Brosnahan S, Mukherjee V, Kaufman D, Bakker J, Lubinsky A, Pradhan D, Sterman DH, Weiden M, Heguy A, Evans L, Uyeki TM, Clemente JC, de Wit E, Schmidt AM, Shopsin B, Desvignes L, Li H, Zhang B, Forst CV, Koide S, Stapleford KA, Khanna KM, Ghedin E, Segal LN. Microbial signatures in the lower airways of mechanically ventilated COVID-19 patients associated with poor clinical outcome. Nat Microbiol 2021; 6: 1245-1258 [PMID: 34465900 DOI: 10.1038/s41564-021-00961-5]
- 126 Wang J, Li F, Sun R, Gao X, Wei H, Li LJ, Tian Z. Bacterial colonization dampens influenza-mediated acute lung injury via induction of M2 alveolar macrophages. Nat Commun 2013; 4: 2106 [PMID: 23820884 DOI: 10.1038/ncomms3106]
- Sette A, Crotty S. Adaptive immunity to SARS-CoV-2 and COVID-19. Cell 2021; 184: 861-880 [PMID: 33497610 DOI: 127 10.1016/j.cell.2021.01.007
- Gauguet S, D'Ortona S, Ahnger-Pier K, Duan B, Surana NK, Lu R, Cywes-Bentley C, Gadjeva M, Shan Q, Priebe GP, 128 Pier GB. Intestinal Microbiota of Mice Influences Resistance to Staphylococcus aureus Pneumonia. Infect Immun 2015; 83: 4003-4014 [PMID: 26216419 DOI: 10.1128/IAI.00037-15]
- 129 Trompette A, Gollwitzer ES, Pattaroni C, Lopez-Mejia IC, Riva E, Pernot J, Ubags N, Fajas L, Nicod LP, Marsland BJ. Dietary Fiber Confers Protection against Flu by Shaping Ly6c(-) Patrolling Monocyte Hematopoiesis and CD8(+) T Cell Metabolism. Immunity 2018; 48: 992-1005.e8 [PMID: 29768180 DOI: 10.1016/j.immuni.2018.04.022]
- Chen J, Vitetta L. Modulation of Gut Microbiota for the Prevention and Treatment of COVID-19. J Clin Med 2021; 10 130 [PMID: 34209870 DOI: 10.3390/jcm10132903]
- 131 Conway EM, Mackman N, Warren RQ, Wolberg AS, Mosnier LO, Campbell RA, Gralinski LE, Rondina MT, van de Veerdonk FL, Hoffmeister KM, Griffin JH, Nugent D, Moon K, Morrissey JH. Understanding COVID-19-associated coagulopathy. Nat Rev Immunol 2022; 22: 639-649 [PMID: 35931818 DOI: 10.1038/s41577-022-00762-9]
- 132 Loo J, Spittle DA, Newnham M. COVID-19, immunothrombosis and venous thromboembolism: biological mechanisms. Thorax 2021; 76: 412-420 [PMID: 33408195 DOI: 10.1136/thoraxjnl-2020-216243]
- 133 Lo Re V 3rd, Dutcher SK, Connolly JG, Perez-Vilar S, Carbonari DM, DeFor TA, Djibo DA, Harrington LB, Hou L, Hennessy S, Hubbard RA, Kempner ME, Kuntz JL, McMahill-Walraven CN, Mosley J, Pawloski PA, Petrone AB, Pishko AM, Driscoll MR, Steiner CA, Zhou Y, Cocoros NM. Association of COVID-19 vs Influenza With Risk of Arterial and Venous Thrombotic Events Among Hospitalized Patients. JAMA 2022; 328: 637-651 [PMID: 35972486 DOI: 10.1001/jama.2022.13072
- 134 Kollias A, Kyriakoulis KG, Lagou S, Kontopantelis E, Stergiou GS, Syrigos K. Venous thromboembolism in COVID-19: A systematic review and meta-analysis. Vasc Med 2021; 26: 415-425 [PMID: 33818197 DOI: 10.1177/1358863X21995566]
- 135 Jiménez D, García-Sanchez A, Rali P, Muriel A, Bikdeli B, Ruiz-Artacho P, Le Mao R, Rodríguez C, Hunt BJ, Monreal M. Incidence of VTE and Bleeding Among Hospitalized Patients With Coronavirus Disease 2019: A Systematic Review and Meta-analysis. Chest 2021; 159: 1182-1196 [PMID: 33217420 DOI: 10.1016/j.chest.2020.11.005]
- 136 Agarwal G, Hajra A, Chakraborty S, Patel N, Biswas S, Adler MK, Lavie CJ. Predictors and mortality risk of venous thromboembolism in patients with COVID-19: systematic review and meta-analysis of observational studies. Ther Adv Cardiovasc Dis 2022; 16: 17539447221105013 [PMID: 35762736 DOI: 10.1177/17539447221105013]
- Pastorelli L, Dozio E, Pisani LF, Boscolo-Anzoletti M, Vianello E, Munizio N, Spina L, Tontini GE, Peyvandi F, Corsi 137 Romanelli MM, Vecchi M. Procoagulatory state in inflammatory bowel diseases is promoted by impaired intestinal barrier function. Gastroenterol Res Pract 2015; 2015: 189341 [PMID: 25767508 DOI: 10.1155/2015/189341]
- 138 Sardu C, Gambardella J, Morelli MB, Wang X, Marfella R, Santulli G. Hypertension, Thrombosis, Kidney Failure, and Diabetes: Is COVID-19 an Endothelial Disease? J Clin Med 2020; 9 [PMID: 32403217 DOI: 10.3390/jcm9051417]
- Gambardella J, Santulli G. What is linking COVID-19 and endothelial dysfunction? Eur Heart J Cardiovasc 139



Pharmacother 2021; 7: e2-e3 [PMID: 33377481 DOI: 10.1093/ehjcvp/pvaa145]

- 140 van Hinsbergh VW. Endothelium--role in regulation of coagulation and inflammation. Semin Immunopathol 2012; 34: 93-106 [PMID: 21845431 DOI: 10.1007/s00281-011-0285-5]
- Gambardella J, Kansakar U, Sardu C, Messina V, Jankauskas SS, Marfella R, Maggi P, Wang X, Mone P, Paolisso G, 141 Sorriento D, Santulli G. Exosomal miR-145 and miR-885 regulate thrombosis in COVID-19. J Pharmacol Exp Ther 2022 [PMID: 35772782 DOI: 10.1124/jpet.122.001209]
- Quinaglia T, Shabani M, Breder I, Silber HA, Lima JAC, Sposito AC. Coronavirus disease-19: The multi-level, multi-142 faceted vasculopathy. Atherosclerosis 2021; 322: 39-50 [PMID: 33706082 DOI: 10.1016/j.atherosclerosis.2021.02.009]
- Peshevska-Sekulovska M, Boeva I, Sekulovski M, Zashev M. Gastrointestinal Ischemia-Stumbling Stone in COVID-19 143 Patients. Gastroenterol Insigh 2022; 13: 206-217 [DOI: 10.3390/gastroent13020021]
- 144 Kaafarani HMA, El Moheb M, Hwabejire JO, Naar L, Christensen MA, Breen K, Gaitanidis A, Alser O, Mashbari H, Bankhead-Kendall B, Mokhtari A, Maurer L, Kapoen C, Langeveld K, El Hechi MW, Lee J, Mendoza AE, Saillant NN, Parks J, Fawley J, King DR, Fagenholz PJ, Velmahos GC. Gastrointestinal Complications in Critically Ill Patients With COVID-19. Ann Surg 2020; 272: e61-e62 [PMID: 32675498 DOI: 10.1097/SLA.00000000004004]
- 145 Schurink B, Roos E, Radonic T, Barbe E, Bouman CSC, de Boer HH, de Bree GJ, Bulle EB, Aronica EM, Florquin S, Fronczek J, Heunks LMA, de Jong MD, Guo L, du Long R, Lutter R, Molenaar PCG, Neefjes-Borst EA, Niessen HWM, van Noesel CJM, Roelofs JJTH, Snijder EJ, Soer EC, Verheij J, Vlaar APJ, Vos W, van der Wel NN, van der Wal AC, van der Valk P, Bugiani M. Viral presence and immunopathology in patients with lethal COVID-19: a prospective autopsy cohort study. Lancet Microbe 2020; 1: e290-e299 [PMID: 33015653 DOI: 10.1016/S2666-5247(20)30144-0]
- 146 Verdecchia P, Cavallini C, Spanevello A, Angeli F. The pivotal link between ACE2 deficiency and SARS-CoV-2 infection. Eur J Intern Med 2020; 76: 14-20 [PMID: 32336612 DOI: 10.1016/j.ejim.2020.04.037]
- 147 Carvalho PR, Sirois P, Fernandes PD. The role of kallikrein-kinin and renin-angiotensin systems in COVID-19 infection. Peptides 2021; 135: 170428 [PMID: 33065209 DOI: 10.1016/j.peptides.2020.170428]
- 148 Qin Z, Liu F, Blair R, Wang C, Yang H, Mudd J, Currey JM, Iwanaga N, He J, Mi R, Han K, Midkiff CC, Alam MA, Aktas BH, Heide RSV, Veazey R, Piedimonte G, Maness NJ, Ergün S, Mauvais-Jarvis F, Rappaport J, Kolls JK, Qin X. Endothelial cell infection and dysfunction, immune activation in severe COVID-19. Theranostics 2021; 11: 8076-8091 [PMID: 34335981 DOI: 10.7150/thno.61810]
- 149 He Z, Si Y, Jiang T, Ma R, Zhang Y, Cao M, Li T, Yao Z, Zhao L, Fang S, Yu B, Dong Z, Thatte HS, Bi Y, Kou J, Yang S, Piao D, Hao L, Zhou J, Shi J. Phosphotidylserine exposure and neutrophil extracellular traps enhance procoagulant activity in patients with inflammatory bowel disease. Thromb Haemost 2016; 115: 738-751 [PMID: 26660948 DOI: 10.1160/TH15-09-0710
- 150 Gambardella J, Coppola A, Izzo R, Fiorentino G, Trimarco B, Santulli G. Role of endothelial miR-24 in COVID-19 cerebrovascular events. Crit Care 2021; 25: 306 [PMID: 34433473 DOI: 10.1186/s13054-021-03731-1]
- Mone P, Gambardella J, Wang X, Jankauskas SS, Matarese A, Santulli G. miR-24 Targets the Transmembrane 151 Glycoprotein Neuropilin-1 in Human Brain Microvascular Endothelial Cells. Noncoding RNA 2021; 7 [PMID: 33540664 DOI: 10.3390/ncrna7010009]
- 152 Gao C, Xie R, Yu C, Ma R, Dong W, Meng H, Zhang Y, Si Y, Zhang Z, Novakovic V, Kou J, Bi Y, Li B, Gilbert GE, Zhou J, Shi J. Thrombotic Role of Blood and Endothelial Cells in Uremia through Phosphatidylserine Exposure and Microparticle Release. PLoS One 2015; 10: e0142835 [PMID: 26580207 DOI: 10.1371/journal.pone.0142835]
- Goshua G, Pine AB, Meizlish ML, Chang CH, Zhang H, Bahel P, Baluha A, Bar N, Bona RD, Burns AJ, Dela Cruz CS, 153 Dumont A, Halene S, Hwa J, Koff J, Menninger H, Neparidze N, Price C, Siner JM, Tormey C, Rinder HM, Chun HJ, Lee AI. Endotheliopathy in COVID-19-associated coagulopathy: evidence from a single-centre, cross-sectional study. Lancet Haematol 2020; 7: e575-e582 [PMID: 32619411 DOI: 10.1016/S2352-3026(20)30216-7]
- 154 Zhang Y, Cao W, Jiang W, Xiao M, Li Y, Tang N, Liu Z, Yan X, Zhao Y, Li T, Zhu T. Profile of natural anticoagulant, coagulant factor and anti-phospholipid antibody in critically ill COVID-19 patients. J Thromb Thrombolysis 2020; 50: 580-586 [PMID: 32648093 DOI: 10.1007/s11239-020-02182-9]
- 155 Vassiliou AG, Keskinidou C, Jahaj E, Gallos P, Dimopoulou I, Kotanidou A, Orfanos SE. ICU Admission Levels of Endothelial Biomarkers as Predictors of Mortality in Critically Ill COVID-19 Patients. Cells 2021; 10 [PMID: 33477776 DOI: 10.3390/cells10010186]
- Bonaventura A, Vecchié A, Dagna L, Martinod K, Dixon DL, Van Tassell BW, Dentali F, Montecucco F, Massberg S, 156 Levi M, Abbate A. Endothelial dysfunction and immunothrombosis as key pathogenic mechanisms in COVID-19. Nat Rev Immunol 2021; 21: 319-329 [PMID: 33824483 DOI: 10.1038/s41577-021-00536-9]
- 157 Martinelli N, Montagnana M, Pizzolo F, Friso S, Salvagno GL, Forni GL, Gianesin B, Morandi M, Lunardi C, Lippi G, Polati E, Olivieri O, De Franceschi L. A relative ADAMTS13 deficiency supports the presence of a secondary microangiopathy in COVID 19. Thromb Res 2020; 193: 170-172 [PMID: 32707276 DOI: 10.1016/j.thromres.2020.07.034]
- 158 Guervilly C, Burtey S, Sabatier F, Cauchois R, Lano G, Abdili E, Daviet F, Arnaud L, Brunet P, Hraiech S, Jourde-Chiche N, Koubi M, Lacroix R, Pietri L, Berda Y, Robert T, Degioanni C, Velier M, Papazian L, Kaplanski G, Dignat-George F. Circulating Endothelial Cells as a Marker of Endothelial Injury in Severe COVID -19. J Infect Dis 2020; 222: 1789-1793 [PMID: 32812049 DOI: 10.1093/infdis/jiaa528]
- Whyte CS, Simpson M, Morrow GB, Wallace CA, Mentzer AJ, Knight JC, Shapiro S, Curry N, Bagot CN, Watson H, 159 Cooper JG, Mutch NJ. The suboptimal fibrinolytic response in COVID-19 is dictated by high PAI-1. J Thromb Haemost 2022; 20: 2394-2406 [PMID: 35780481 DOI: 10.1111/jth.15806]
- Shaw RJ, Bradbury C, Abrams ST, Wang G, Toh CH. COVID-19 and immunothrombosis: emerging understanding and 160 clinical management. Br J Haematol 2021; 194: 518-529 [PMID: 34114204 DOI: 10.1111/bjh.17664]
- 161 Gillot C, Favresse J, Mullier F, Lecompte T, Dogné JM, Douxfils J. NETosis and the Immune System in COVID-19: Mechanisms and Potential Treatments. Front Pharmacol 2021; 12: 708302 [PMID: 34421600 DOI: 10.3389/fphar.2021.708302
- Hottz ED, Azevedo-Quintanilha IG, Palhinha L, Teixeira L, Barreto EA, Pão CRR, Righy C, Franco S, Souza TML, 162



Kurtz P, Bozza FA, Bozza PT. Platelet activation and platelet-monocyte aggregate formation trigger tissue factor expression in patients with severe COVID-19. Blood 2020; 136: 1330-1341 [PMID: 32678428 DOI: 10.1182/blood.2020007252]

- 163 Oliva A, Cammisotto V, Cangemi R, Ferro D, Miele MC, De Angelis M, Cancelli F, Pignatelli P, Venditti M, Pugliese F, Mastroianni CM, Violi F. Low-Grade Endotoxemia and Thrombosis in COVID-19. Clin Transl Gastroenterol 2021; 12: e00348 [PMID: 34092777 DOI: 10.14309/ctg.00000000000348]
- Hasan RA, Koh AY, Zia A. The gut microbiome and thromboembolism. Thromb Res 2020; 189: 77-87 [PMID: 32192995 164 DOI: 10.1016/j.thromres.2020.03.003]
- 165 Shin HS, Xu F, Bagchi A, Herrup E, Prakash A, Valentine C, Kulkarni H, Wilhelmsen K, Warren S, Hellman J. Bacterial lipoprotein TLR2 agonists broadly modulate endothelial function and coagulation pathways in vitro and in vivo. J Immunol 2011; 186: 1119-1130 [PMID: 21169547 DOI: 10.4049/jimmunol.1001647]
- 166 Jäckel S, Kiouptsi K, Lillich M, Hendrikx T, Khandagale A, Kollar B, Hörmann N, Reiss C, Subramaniam S, Wilms E, Ebner K, Brühl MV, Rausch P, Baines JF, Haberichter S, Lämmle B, Binder CJ, Jurk K, Ruggeri ZM, Massberg S, Walter U, Ruf W, Reinhardt C. Gut microbiota regulate hepatic von Willebrand factor synthesis and arterial thrombus formation via Toll-like receptor-2. Blood 2017; 130: 542-553 [PMID: 28572286 DOI: 10.1182/blood-2016-11-754416]
- 167 Fiorentino G, Coppola A, Izzo R, Annunziata A, Bernardo M, Lombardi A, Trimarco V, Santulli G, Trimarco B. Effects of adding L-arginine orally to standard therapy in patients with COVID-19: A randomized, double-blind, placebocontrolled, parallel-group trial. Results of the first interim analysis. EClinicalMedicine 2021; 40: 101125 [PMID: 34522871 DOI: 10.1016/j.eclinm.2021.101125]
- Adebayo A, Varzideh F, Wilson S, Gambardella J, Eacobacci M, Jankauskas SS, Donkor K, Kansakar U, Trimarco V, 168 Mone P, Lombardi A, Santulli G. I-Arginine and COVID-19: An Update. Nutrients 2021; 13 [PMID: 34836206 DOI: 10.3390/nu13113951]
- 169 Ascher S, Wilms E, Pontarollo G, Formes H, Bayer F, Müller M, Malinarich F, Grill A, Bosmann M, Saffarzadeh M, Brandão I, Groß K, Kiouptsi K, Kittner JM, Lackner KJ, Jurk K, Reinhardt C. Gut Microbiota Restricts NETosis in Acute Mesenteric Ischemia-Reperfusion Injury. Arterioscler Thromb Vasc Biol 2020; 40: 2279-2292 [PMID: 32611241 DOI: 10.1161/ATVBAHA.120.314491]
- Luo QJ, Sun MX, Guo YW, Tan SW, Wu XY, Abassa KK, Lin L, Liu HL, Jiang J, Wei XQ. Sodium butyrate protects 170 against lipopolysaccharide-induced liver injury partially via the GPR43/β-arrestin-2/NF-κB network. Gastroenterol Rep (Oxf) 2021; 9: 154-165 [PMID: 34026223 DOI: 10.1093/gastro/goaa085]
- 171 Zhu W, Gregory JC, Org E, Buffa JA, Gupta N, Wang Z, Li L, Fu X, Wu Y, Mehrabian M, Sartor RB, McIntyre TM, Silverstein RL, Tang WHW, DiDonato JA, Brown JM, Lusis AJ, Hazen SL. Gut Microbial Metabolite TMAO Enhances Platelet Hyperreactivity and Thrombosis Risk. Cell 2016; 165: 111-124 [PMID: 26972052 DOI: 10.1016/j.cell.2016.02.011]
- Boini KM, Hussain T, Li PL, Koka S. Trimethylamine-N-Oxide Instigates NLRP3 Inflammasome Activation and 172 Endothelial Dysfunction. Cell Physiol Biochem 2017; 44: 152-162 [PMID: 29130962 DOI: 10.1159/000484623]
- 173 Israr MZ, Ibrahim W, Salzano A, Sarmad S, Wilde MJ, Cordell RL, Greening NJ, Brightling CE, Siddiqui S, Suzuki T; EMBER consortium. Association of gut-related metabolites with respiratory symptoms in COVID-19: A proof-of-concept study. Nutrition 2022; 96: 111585 [PMID: 35131599 DOI: 10.1016/j.nut.2021.111585]
- 174 Teixeira PC, Dorneles GP, Santana Filho PC, da Silva IM, Schipper LL, Postiga IAL, Neves CAM, Rodrigues Junior LC, Peres A, Souto JT, Fonseca SG, Eller S, Oliveira TF, Rotta LN, Thompson CE, Romão PRT. Increased LPS levels coexist with systemic inflammation and result in monocyte activation in severe COVID-19 patients. Int Immunopharmacol 2021; 100: 108125 [PMID: 34543980 DOI: 10.1016/j.intimp.2021.108125]
- Sirivongrangson P, Kulvichit W, Payungporn S, Pisitkun T, Chindamporn A, Peerapornratana S, Pisitkun P, Chitcharoen 175 S, Sawaswong V, Worasilchai N, Kampunya S, Putcharoen O, Thawitsri T, Leelayuwatanakul N, Kongpolprom N, Phoophiboon V, Sriprasart T, Samransamruajkit R, Tungsanga S, Tiankanon K, Lumlertgul N, Leelahavanichkul A, Sriphojanart T, Tantawichien T, Thisyakorn U, Chirathaworn C, Praditpornsilpa K, Tungsanga K, Eiam-Ong S, Sitprija V, Kellum JA, Srisawat N. Endotoxemia and circulating bacteriome in severe COVID-19 patients. Intensive Care Med Exp 2020; 8: 72 [PMID: 33284413 DOI: 10.1186/s40635-020-00362-8]
- 176 Oliva A, Miele MC, Di Timoteo F, De Angelis M, Mauro V, Aronica R, Al Ismail D, Ceccarelli G, Pinacchio C, d'Ettorre G, Mascellino MT, Mastroianni CM. Persistent Systemic Microbial Translocation and Intestinal Damage During Coronavirus Disease-19. Front Immunol 2021; 12: 708149 [PMID: 34335624 DOI: 10.3389/fimmu.2021.708149]
- Kumar NP, Venkataraman A, Hanna LE, Putlibai S, Karthick M, Rajamanikam A, Sadasivam K, Sundaram B, Babu S. 177 Systemic Inflammation and Microbial Translocation Are Characteristic Features of SARS-CoV-2-Related Multisystem Inflammatory Syndrome in Children. Open Forum Infect Dis 2021; 8: ofab279 [PMID: 34322566 DOI: 10.1093/ofid/ofab279]
- 178 Prasad R, Patton MJ, Floyd JL, Fortmann S, DuPont M, Harbour A, Wright J, Lamendella R, Stevens BR, Oudit GY, Grant MB. Plasma Microbiome in COVID-19 Subjects: An Indicator of Gut Barrier Defects and Dysbiosis. Int J Mol Sci 2022; 23 [PMID: 36012406 DOI: 10.3390/ijms23169141]
- 179 Giron LB, Peluso MJ, Ding J, Kenny G, Zilberstein NF, Koshy J, Hong KY, Rasmussen H, Miller GE, Bishehsari F, Balk RA, Moy JN, Hoh R, Lu S, Goldman AR, Tang HY, Yee BC, Chenna A, Winslow JW, Petropoulos CJ, Kelly JD, Wasse H, Martin JN, Liu Q, Keshavarzian A, Landay A, Deeks SG, Henrich TJ, Abdel-Mohsen M. Markers of fungal translocation are elevated during post-acute sequelae of SARS-CoV-2 and induce NF-xB signaling. JCI Insight 2022; 7 [PMID: 35727635 DOI: 10.1172/jci.insight.160989]
- 180 Venzon M, Bernard-Raichon L, Klein J, Axelrad JE, Hussey GA, Sullivan AP, Casanovas-Massana A, Noval MG, Valero-Jimenez AM, Gago J, Wilder E; Yale IMPACT Research Team, Thorpe LE, Littman DR, Dittmann M, Stapleford KA, Shopsin B, Torres VJ, Ko AI, Iwasaki A, Cadwell K, Schluter J. Gut microbiome dysbiosis during COVID-19 is associated with increased risk for bacteremia and microbial translocation. Res Sq 2021 [PMID: 34341786 DOI: 10.21203/rs.3.rs-726620/v1]
- 181 Mozaffari SA, Salehi A, Mousavi E, Zaman BA, Nassaj AE, Ebrahimzadeh F, Nasiri H, Valedkarimi Z, Adili A,



Asemani G, Akbari M. SARS-CoV-2-associated gut microbiome alteration; A new contributor to colorectal cancer pathogenesis. Pathol Res Pract 2022; 239: 154131 [PMID: 36191449 DOI: 10.1016/j.prp.2022.154131]

- 182 Assimakopoulos SF, Eleftheriotis G, Lagadinou M, Karamouzos V, Dousdampanis P, Siakallis G, Marangos M. SARS CoV-2-Induced Viral Sepsis: The Role of Gut Barrier Dysfunction. *Microorganisms* 2022; 10 [PMID: 35630492 DOI: 10.3390/microorganisms10051050]
- 183 Alhumaid S, Al Mutair A, Busubaih JS, Al Dossary N, Alsuliman M, Baltyour SA, Alissa I, Al Hassar HI, Al Aithan NA, Albassri HA, AlOmran SA, ALGhazal RM, Busbaih A, Alsalem NA, Alagnam W, Alyousef MY, Alseffay AU, Al Aish HA, Aldiaram A, Al Eissa HA, Alhumaid MA, Bukhamseen AN, Al Mutared KM, Aljwisim AH, Twibah AM, AlSaeed MM, Alkhalaf HA, ALShakhs FM, Koritala T, Al-Tawfiq JA, Dhama K, Rabaan AA, Al-Omari A. Colorectal cancer in patients with SARS-CoV-2: a systematic review and meta-analysis. Infect Agent Cancer 2022; 17: 49 [PMID: 36096812 DOI: 10.1186/s13027-022-00459-7]



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MINIREVIEWS

The impact of COVID-19 on liver injury in various age

Amin Sadeghi Dousari, Seyed Soheil Hosseininasab, Fatemeh Sadeghi Dousari, Masoumeh Fuladvandi, Naghmeh Satarzadeh

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Amin Sadeghi Dousari, Department of Microbiology, Jiroft University of Medical Sciences, Jiroft 7861634204, Iran

Seyed Soheil Hosseininasab, Faculty of Pharmacy, Kerman University of Medical Sciences, Kerman 7616913555, Iran

Fatemeh Sadeghi Dousari, Department of Midwifery, Jiroft University of Medical Sciences, Jiroft 7861634204, Iran

Masoumeh Fuladvandi, Department of Nursing, Aligoudarz School of Nursing, Lorestan University of Medical Sciences, Khorramabad 6813833946, Iran

Naghmeh Satarzadeh, Department of Pharmaceutical Biotechnology, Kerman University of Medical Sciences, Kerman 7616913555, Iran

Corresponding author: Naghmeh Satarzadeh, PhD, Researcher, Department of Pharmaceutical Biotechnology, Kerman University of Medical Sciences, Haft-Bagh Highway, Kerman 7616913555, Iran. n.satarzadeh@kmu.ac.ir

Abstract

The coronavirus disease 2019 (COVID-19) disease was first detected in December 2019 in Wuhan, China. This disease is currently one of the most important global health problems. The novel coronavirus COVID-19 is a respiratory illness, that has caused a deadly pandemic that is spreading rapidly around the world. It is not only a respiratory system virus that causes severe lung disease, but also a systemic disease agent that can affect all systems. People with COVID-19 disease usually have respiratory signs, however, the liver disorder is not an uncommon presentation. In addition, many studies around the world have revealed that the liver is injured to various degrees in patients with severe acute respiratory syndrome coronavirus 2 disease. This review mainly focuses on the impact of COVID-19 on Liver Injury at various ages.

Key Words: Liver injury; Coronavirus disease 2019; Severe acute respiratory syndrome coronavirus 2; Minireview

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Core Tip: Studies have shown that neonates have rare evidence of liver damage, and in terms of age, they show the least amount of liver damage in the face of coronavirus disease 2019 (COVID-19) among affected people. Also, many studies reported different patterns of liver damage among children with COVID-19 much less than in adults, which is probably related to differences in their innate immune system and adaptation. The highest rate of liver damage is in adult patients and aspartate aminotransferase levels had the highest relevance with mortality compared to other indices reflecting liver injury.

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INTRODUCTION

Coronaviruses are a big family of viruses belonging to the realm Riboviria, order Nidovirales, family Coronaviridae and subfamily Coronavirinae. This virus contains an RNA genome and belongs to the Coronaviridae family [1,2]. This virus is spread in a wide spectrum of humans, other mammals, and avian species, also inducing acute respiratory infections[3]. Types of coronaviruses including HCoV-NL63, HCoV-HKU1, HCoV-229E, and HCoV-OC43 have been presented as mild virulent human viruses worldwide[4]. These viruses cause mild to severe acute respiratory illnesses in humans[3]. Coronavirus disease 2019 (COVID-19) was identified for the first time in December 2019, in Wuhan, located in the capital of Hubei Province in the People's Republic of China[1]. Coronavirus disease 2019 is an infectious illness that has caused a lethal pandemic that rapidly extends worldwide [5,6]. The signs of COVID-19 appear approximately 5.2 d after the disease and last for a minimum of 41 d and a maximum of 14 d until the end of life[4,7].

In the early stages of COVID-19, it has been found that severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is not only a respiratory system virus that generates severe lung disease but a systemic disease factor that can involve all systems[8,9]. Some extrapulmonary involvement of SARS-CoV-2 disease is in organs like the liver, heart, or kidneys^[10]. Many studies throughout the world have demonstrated that the liver is injured to differing degrees in patients affected by SARS-CoV-2 disease[8,9].

The liver is a vital member that is mostly responsible for the storage of glycogen and regulation of blood glucose levels, protein synthesis, metabolism of toxic substances, and very other physiological processes[8,9]. Liver dysfunction has been reported in 54% of hospitalized patients affected by COVID-19 disease, most of which are more severe in COVID-19[11]. Liver injuries have been documented in patients affected by COVID-19, and commonly have mild increasing liver enzymes range from 14% to 53% [12]. Patients with severe disease, especially those hospitalized in ICU, have shown a higher increase in transaminase enzymes than patients with mild to moderate severity [13]. Furthermore, few studies investigated the dynamic change of liver function during the COVID-19 pandemic. Also, no study to date has documented the incidence of a simultaneous increase in liver transaminases and total bilirubin levels in COVID-19 patients[14].

The purpose of this review is to evaluate the effect of COVID-19 on liver injury in various ages.

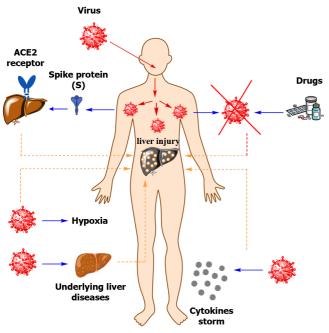
DEFINITION OF LIVER INJURY

Patients who make severe acute liver injury in the absence of preexisting chronic liver disease, usually indicate noteworthy liver dysfunction marked with coagulopathy, which is described as an international normalized ratio ≥ 1.5 and is classically defined as acute liver failure (ALF) when any degree of hepatic encephalopathy (HE) is existing[15]. The ALF types include: (1) Hyperacute: < 7 d; (2) Acute: 7-28 d, and (3) Subacute: 28 d to 6 mo, depending on latency between the beginning of signs and development of encephalopathy and coagulopathy[16,17].

HOW DOES COVID-19 CAUSE LIVER INJURY?

Liver injury is seen in patients with COVID-19, and its harshness is altered depending on the patient's age, geographical area, and disease severity[18]. Viral direct damage[19], immune damage, systemic inflammatory response, drug-induced, ischemia-reperfusion injury, mechanical ventilation, and underlying diseases may donate to liver injury[20] (Figure 1).





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Figure 1 Summary of liver injury in coronavirus disease 2019 patients. ACE2: Angiotensin-converting enzyme 2.

There is much evidence that COVID-19 causes abnormal liver function experiment outcomes with increased levels of aspartate aminotransferase (AST) and alanine aminotransferase (ALT) in people with liver damage[21,22]. Studies performed in Wuhan, China, recorded mildly elevated ALT and AST levels in 14%–53% of cases, with higher rates of both enzymes in patients with intense infection, mostly in patients requiring admission to the intensive care unit[23]. In COVID-19 patients with injured biliary tract were increased serum bilirubin, alkaline phosphatase (ALP), and gamma-glutamyl transferase (GGT) levels[24]. Also, in cases where the virus causes notable liver injury and intense clinical symptoms, varying levels of ALP and GGT along with high levels of ALT and total bilirubin have been reported in 58%-78% of patients[25].

The pathophysiology of liver damage may include the cytopathic result, in which spike (S) protein of coronaviruses 2019 attaches to the angiotensin-converting enzyme 2 (ACE2) receptor, leading to reduced liver function and hepatobiliary disease[26]. S protein viral entry into the liver cells (hepatocytes and cholangiocytes), a process that involves binding to the surface of the host cell through binding of the surface unit (S1) to a receptor[27,28]. The virus attains access to the host *via* the ACE2 receptor (a type I integral membrane protein containing zinc, which indicates enzymatic action through cleaving the vasoconstrictor peptide angiotensin II to angiotensin I, a strong vasodilator peptide, therefore decreasing blood pressure). ACE2 receptor was abundantly demonstrated in epithelial cells that line a three-dimensional network of bile ducts named cholangiocytes (60%), hepatocytes (3%) in the liver, alveolar cells of the lungs, and in various organs such as the pancreas, kidney, and heart[29,30].

FACTORS RELATED TO THE COVID-19 DISEASE THAT CAUSE LIVER DAMAGE

Drugs: There are several drugs that prescribed to manage the treatment of patients with COVID-19 and associated symptoms, including therapeutic agents such as antivirals, antibiotics, acetaminophen, immunomodulators, corticosteroids, steroids, and antipyretics, that are metabolized through the liver and their use may lead to hepatotoxicity[31,32]. It has been reported that liver damage caused by these drugs is reason of anomalies in liver experiments and histological variation like micro-vesicular steatosis and liver inflammation in COVID-19 patients. Drugs like oseltamivir, arbidol, hydroxy-chloroquine, as well as ritonavir, and lopinavir in the treatment of patients may induce variable degrees of hepatotoxicity[33].

Hypoxia: Hypoxia in patients with COVID-19 is known as a major factor that causes a decrease in oxygen saturation values and finally reduction in systemic blood pressure[34]. This will ultimately cause a reduction in liver arterial perfusion *via* liver ischemia and hypoxia reperfusion injury *via* liver cell hypoxia[35].

Cytokines storm: Another factor related to COVID-19 that causes liver damage is the occurrence of a cytokine storm. In cases of the moderate and severe phase of the disease, which includes endothelial damage, it is related *via* a strong immune response to the SARS-CoV-2 virus[36]. This step is accompanied by the stimulation of inflammasomes (cytosolic multiprotein oligomers) that are responsible for the activation of caspase-1 and the release of pro-inflammatory cytokines [Interleukin (IL)-1 β , IL-6, and IL-18][37]. In the next step, these cytokines stimulate the expression of genes relevant to the immune response, and through intracellular signaling, especially using IL-6, other pro-inflammatory cytokine biomarkers like tumor necrosis factor-alpha, IL-2, IL-8, IL-10, IL-17, granulocyte colony-stimulating factor monocyte chemoattractant protein, and interferon-inducible protein[38]. In addition, IL-6 activates numerous downstream signal pathways using creating complexes with its receptor[39], and also the reason for raised ferritin and C-reactive protein levels, reduced lymphocytes and enhanced neutrophils[40].

Underlying liver diseases: Underlying liver diseases can aggravate liver damage in the face of COVID-19. The prevalence of underlying liver diseases in patients with COVID-19 has been reported to be between 3% -11% in large observational studies[27,41,42]. From cases of these underlying diseases can be mentioned chronic liver disease and cirrhosis, non-alcoholic fatty liver disease, and liver transplantation[27].

THE ASSOCIATION DIFFERENT AGES AND LIVER INJURY CAUSED BY COVID-19

Many studies have demonstrated various patterns of disease and their outcomes between adults and children, possibly associated with the difference in their innate and adaptive immune systems. Children with or without chronic sickness are less likely to have a severe illness from COVID-19 confirmed in various studies[43]. However, children affected by COVID-19 have a milder infection than adults, possibly related to children having preserved effector and immunosuppressive components[44]. The differences in age, gender, and population are probably due to differences in immune responses and different variants of SARS-CoV-2[45]. Furthermore, children are less likely to have multiple chronic conditions than older people[44]. Children with a weakened immune system, such as liver illnesses considered at higher risk of coronavirus[43]. Some reports showed children to have higher ACE2 expression than older adults, that it conversion ang I (angiotensin I enzyme) into angiotensin 1-7 (ang 1-7) enzyme, thus ang 1-7 enzyme protecting against pulmonary capillary leak and inflammation. This issue can be the reason why children are more resistant to COVID-19 than adults. The mechanism of liver injury in cases by COVID-19 is indistinct[46]. The liver damage associated with COVID-19 is described as any liver injury happening during the progression and treatment of this disease in cases with or without underlying liver illness[47]. The most common presentation of liver damage in patients is with COVID-19 shown by increasing liver enzymes and also decreasing Serum albumin in severe cases. However, reports of death in affected by COVID-19 patients due to severe liver injury rarely happen[48,49].

THE EFFECTS OF COVID-19 ON LIVER INJURY IN NEONATES

A clinical study of 10 neonates (including twins) to 9 born to mothers with COVID-19 showed that only two infants have thrombocytopenia accompanied using abnormal liver function [50]. Clinical Analysis of 48 Neonates Born to Mothers with COVID-19 (confirmed or clinically diagnosed) or without it accomplished by Liu et al[51] polymerase chain reaction (PCR) test of all neonates was negative. Evidence of vertical transmission and liver injury was not observed. Similarly, a clinical investigation of 19 neonates born to mothers with COVID-19 was investigated at Tongji Hospital, China. The COVID-19 real-time reverse-transcription-PCR Test of all neonates was negative. In this study also, vertical transfer of SARS-CoV-2 was not found[52]. Wang et al[53] investigated a case report of neonates with positive test results for coronavirus 36 h after birth. Nevertheless, whether this Newborn is vertical transfer from the mother to the neonate is yet to be verified. In this case, was observed a significant increase in AST and abnormalities in liver function tests. Stolfi et al[54] reported a neonate of vertical transmission of COVID-19 with liver injury, confirmed using an increase in serum transaminases in Italy. The positive PCR test of COVID-19 in a neonate less than 24 h after C-section probably indicates vertical transmission, therefore proposing a transplacental transfer of SARS-CoV-2. Liver damage in this neonate was created probably using a direct virus-mediated mechanism that correlated to ACE2 receptor expression, But the details are unknown. Out of 33 neonates born to mothers affected by COVID-19 in China, three cases have positive PCR tests for COVID-19. One neonate had observed increasing transaminases[55].

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THE EFFECTS OF COVID-19 ON LIVER INJURY IN ADULTS

Guan *et al*[56] extracted information about 1099 patients with positive PCR tests for COVID-19 in 30 provinces in China (from 552 hospitals). Out of 1099 patients, 112 cases (with an average age of 47 years) had a slight increase of AST with mild illness, and 56 adults had a high increase of AST with severe illness. In 2020, in a national retrospective cohort study in France, Mallet *et al*[57] examined the danger of mortality after COVID-19 disease in adult with chronic liver disease. The study contained 259,110 of all adults with COVID-19 who were released from post-acute care and acute, public and private hospitals in France in 2020. From a total of 259,110 patients who were between 54 and 83 years old (average age 70 years) and 52% were men, including 10,006 (3.9%) and 15,746 (6.0%) patients with alcohol use disorders and chronic liver disease, respectively. The results of this study demonstrated that patients with uncompensated cirrhosis, primary liver cancer, and alcohol use disorders were at high risk for COVID-19 fatality, while patients with compensated cirrhosis, mild liver disease, organ, including liver transplant, or acquired depressive syndrome were not at risk of COVID-19 mortality. Overall, mortality was in 38,203 (15%) of the patients, including chronic liver disease 2,941 (19%) and 7,475 (28%) after mechanical ventilation.

In another study, Mantovani et al[42] evaluated the widespread outbreak of chronic liver disease among patients affected by COVID-19 with a meta-analysis of data in observational studies and investigating the association between the liver injury and COVID-19 disease. The number of 11 observational studies included 2034 adults aged between 45 and 54 years (average age of 49 years), and 57.2% were men. The results of this study revealed that the widespread outbreak of chronic liver disease was 3% and people with severe disease of COVID-19 had associated changes in liver enzymes and coagulation profiles, which were reported to be possibly due to an innate immune response to the virus. In addition, the findings of this study displayed that the gain in AST level in hospitalized severe patients was more frequent and significant than the gain in ALT, and AST levels had the highest relevance with mortality compared to other indices reflecting liver damage, and it was reported that common factors related with the increase in liver damage indicators were the enhance in the number of neutrophils, the decrease in the number of lymphocytes, and male gender. The association between liver damage and adverse events of Coronavirus disease is indistinct. In adult studies, a higher rate of liver enzymes was reported in adults with severe diseases than in milder diseases[43]. One of the limitations of this study is that it is a retrospective study, which may have inadvertently missed some studies with basic keyword searches. In addition, the mechanism of liver damage at COVID-19 patients with different ages in used studies has not been clarified. However, this study was summarized existing evidence on the effects of COVID-19 on the liver injury at various ages. Furthermore, this study might have helped in clinical diagnosis and treatment for COVID-19related liver disease.

Figure 2 shows a summary of the effects of COVID-19 on Liver Injury at various ages.

RECOMMENDATIONS AND FUTURE RESEARCHES

The mechanisms of liver damage in either adults or children with COVID-19 are not fully unclear and the impact of liver injury caused by new variants of COVID-19 in patients is unexplained. Furthermore, further investigation is required to determine liver involvement and the consequence of COVID-19 on various ages with liver disease. Also, the pathogenetic mechanisms of COVID-19 on liver injury of patients in different age groups need to be investigated.

CONCLUSION

Liver damage is seen in patients affected by COVID-19, and factors including viral direct damage, immune damage, systemic inflammatory response, drug-induced, ischemia-reperfusion injury, mechanical ventilation, and underlying diseases contribute to liver injury. The association between liver damage and adverse clinical outcomes in patients affected by COVID-19 and the mechanism of SARS-CoV-2 in creating this injury is also unclear. Studies have shown that neonates have rare evidence of liver damage, and in terms of age, they show the least amount of liver damage in the face of COVID-19 disease among affected people. Most patients with COVID-19 have maintained their normal liver function during the disease, but patients with more severe disease probably had an abnormal liver function. Also, many studies reported different patterns of liver damage among children with COVID-19 much less than in adults, which is probably related to differences in their innate immune system and adaptation. Most patients with COVID-19 have a mild increase in aspartate aminotransferase, alanine aminotransferase, or total bilirubin. The highest rate of liver damage is in adult patients and AST levels had the highest relevance with mortality compared to other indices reflecting liver injury.

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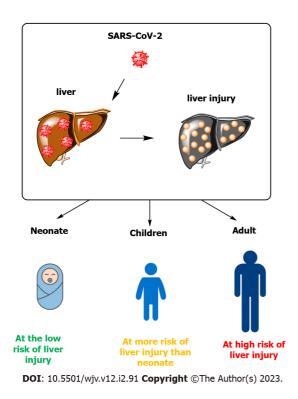


Figure 2 Summary of liver injury of coronavirus disease 2019 according to the age of patients.

FOOTNOTES

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Country/Territory of origin: Iran

ORCID number: Naghmeh Satarzadeh 0000-0002-8753-9599.

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REFERENCES

- 1 Sadeghi Dousari A, Taati Moghadam M, Satarzadeh N. COVID-19 (Coronavirus Disease 2019): A New Coronavirus Disease. Infect Drug Resist 2020; 13: 2819-2828 [PMID: 32848431 DOI: 10.2147/IDR.S259279]
- 2 Meena P, Bhargava V, Rana DS, Bhalla AK, Gupta A. COVID-19 and the kidney: A matter of concern. Curr Med Res Pract 2020; 10: 165-168 [PMID: 32839726 DOI: 10.1016/j.cmrp.2020.07.003]
- 3 Moghadam MT, Taati B, Paydar Ardakani SM, Suzuki K. Ramadan Fasting During the COVID-19 Pandemic; Observance of Health, Nutrition and Exercise Criteria for Improving the Immune System. Front Nutr 2020; 7: 570235 [PMID: 33521030 DOI: 10.3389/fnut.2020.570235]
- 4 Shakibnia P, Ahmadi RH, Fallah F, Ebrahimzadeh F, Dosari AS, Mojtahedi A. et al Iran as the Center of challenges in the Middle East for the Outbreak of COVID-19 Delta Variant. Iranian Red Crescent Medical Journal 2021; 23: 1394 [DOI: 10.32592/ircmj.2021.23.11.1394]
- Taati B, Paydar Ardakani SM, Suzuki K, Sadat Modaresi M, Taati Moghadam M, Roozbeh B. Protective Roles of Exercise 5 and Nutritional Factors for Immune System During Delta Variant-COVID-19 Outbreaks: Evidence Review and Practical Recommendations. Iranian Journal of Medical Microbiology 2022; 16: 2345-4342 [DOI: 10.30699/ijmm.16.3.178]



- 6 Moghadam MT, Babakhani S, Rajabi S, Baravati FB, Raeisi M, Dousari AS. Does stress and anxiety contribute to COVID-19? Iranian Journal of Psychiatry and Behavioral Sciences 2021; 15 [DOI: 10.5812/ijpbs.106041]
- 7 Li Q, Guan X, Wu P, Wang X, Zhou L, Tong Y, Ren R, Leung KSM, Lau EHY, Wong JY, Xing X, Xiang N, Wu Y, Li C, Chen Q, Li D, Liu T, Zhao J, Liu M, Tu W, Chen C, Jin L, Yang R, Wang Q, Zhou S, Wang R, Liu H, Luo Y, Liu Y, Shao G, Li H, Tao Z, Yang Y, Deng Z, Liu B, Ma Z, Zhang Y, Shi G, Lam TTY, Wu JT, Gao GF, Cowling BJ, Yang B, Leung GM, Feng Z. Early Transmission Dynamics in Wuhan, China, of Novel Coronavirus-Infected Pneumonia. N Engl J Med 2020; 382: 1199-1207 [PMID: 31995857 DOI: 10.1056/NEJMoa2001316]
- Kayaaslan B, Guner R. COVID-19 and the liver: A brief and core review. World J Hepatol 2021; 13: 2013-2023 [PMID: 8 35070005 DOI: 10.4254/wjh.v13.i12.2013]
- 9 Trefts E, Gannon M, Wasserman DH. The liver. Curr Biol 2017; 27: R1147-R1151 [PMID: 29112863 DOI: 10.1016/j.cub.2017.09.019]
- 10 Carvalho T. Extrapulmonary SARS-CoV-2 manifestations. Nat Med 2020; 26: 1806 [PMID: 33288941 DOI: 10.1038/s41591-020-01162-z]
- 11 Davidov-Derevynko Y, Ben Yakov G, Wieder A, Segal G, Naveh L, Orlova N, Gringauz I, Amit S, Mor O, Klempfner R, Rahav G, Ben Ari Z. The liver in severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection. Eur J Gastroenterol Hepatol 2021; 33: e313-e319 [PMID: 33653988 DOI: 10.1097/MEG.00000000002048]
- Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, Wang B, Xiang H, Cheng Z, Xiong Y, Zhao Y, Li Y, Wang X, Peng Z. 12 Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. JAMA 2020; 323: 1061-1069 [PMID: 32031570 DOI: 10.1001/jama.2020.1585]
- Shi H, Han X, Jiang N, Cao Y, Alwalid O, Gu J, Fan Y, Zheng C. Radiological findings from 81 patients with COVID-19 13 pneumonia in Wuhan, China: a descriptive study. Lancet Infect Dis 2020; 20: 425-434 [PMID: 32105637 DOI: 10.1016/S1473-3099(20)30086-4]
- 14 Wang Q, Zhao H, Liu LG, Wang YB, Zhang T, Li MH, Xu YL, Gao GJ, Xiong HF, Fan Y, Cao Y, Ding R, Wang JJ, Cheng C, Xie W. Pattern of liver injury in adult patients with COVID-19: a retrospective analysis of 105 patients. Mil Med Res 2020; 7: 28 [PMID: 32507110 DOI: 10.1186/s40779-020-00256-6]
- Koch DG, Speiser JL, Durkalski V, Fontana RJ, Davern T, McGuire B, Stravitz RT, Larson AM, Liou I, Fix O, Schilsky 15 ML, McCashland T, Hay JE, Murray N, Shaikh OS, Ganger D, Zaman A, Han SB, Chung RT, Brown RS, Munoz S, Reddy KR, Rossaro L, Satyanarayana R, Hanje AJ, Olson J, Subramanian RM, Karvellas C, Hameed B, Sherker AH, Lee WM, Reuben A. The Natural History of Severe Acute Liver Injury. Am J Gastroenterol 2017; 112: 1389-1396 [PMID: 28440304 DOI: 10.1038/aig.2017.981
- 16 Lemmer P, Pospiech JC, Canbay A. Liver failure-future challenges and remaining questions. Ann Transl Med 2021; 9: 734 [PMID: 33987432 DOI: 10.21037/atm-20-4968]
- 17 Canbay A, Tacke F, Hadem J, Trautwein C, Gerken G, Manns MP. Acute liver failure: a life-threatening disease. Dtsch Arztebl Int 2011; 108: 714-720 [PMID: 22114640 DOI: 10.3238/arztebl.2011.0714]
- Zhao JN, Fan Y, Wu SD. Liver injury in COVID-19: A minireview. World J Clin Cases 2020; 8: 4303-4310 [PMID: 18 33083389 DOI: 10.12998/wjcc.v8.i19.4303]
- Satarzadeh N, Behzadi A, Khalilabadi RM. Donors with Positive Hepatitis B and C Infections and Their Demographic 19 Characteristics, a Study in Jirof Blood Donation Center From 2011 to 2016. Int J Basic Sci Med 2022; 7: 57-60. [DOI: 10.34172/ijbsm.2022.10]
- Shousha HI, Ramadan A, Lithy R, El-Kassas M. Patterns of liver profile disturbance in patients with COVID-19. World J 20 Clin Cases 2022; 10: 2063-2071 [PMID: 35321162 DOI: 10.12998/wjcc.v10.i7.2063]
- Kukla M, Skonieczna-Żydecka K, Kotfis K, Maciejewska D, Łoniewski I, Lara LF, Pazgan-Simon M, Stachowska E, Kaczmarczyk M, Koulaouzidis A, Marlicz W. COVID-19, MERS and SARS with Concomitant Liver Injury-Systematic Review of the Existing Literature. J Clin Med 2020; 9 [PMID: 32403255 DOI: 10.3390/jcm9051420]
- 22 Krishnan A, Prichett L, Tao X, Alqahtani SA, Hamilton JP, Mezey E, Strauss AT, Kim A, Potter JJ, Chen PH, Woreta TA. Abnormal liver chemistries as a predictor of COVID-19 severity and clinical outcomes in hospitalized patients. World J Gastroenterol 2022; 28: 570-587 [PMID: 35316959 DOI: 10.3748/wjg.v28.i5.570]
- Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, Qiu Y, Wang J, Liu Y, Wei Y, Xia J, Yu T, Zhang X, Zhang L. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. Lancet 2020; 395: 507-513 [PMID: 32007143 DOI: 10.1016/S0140-6736(20)30211-7]
- 24 Cai Q, Huang D, Yu H, Zhu Z, Xia Z, Su Y, Li Z, Zhou G, Gou J, Qu J, Sun Y, Liu Y, He Q, Chen J, Liu L, Xu L. COVID-19: Abnormal liver function tests. J Hepatol 2020; 73: 566-574 [PMID: 32298767 DOI: 10.1016/j.jhep.2020.04.006]
- McGrowder DA, Miller F, Anderson Cross M, Anderson-Jackson L, Bryan S, Dilworth L. Abnormal Liver Biochemistry 25 Tests and Acute Liver Injury in COVID-19 Patients: Current Evidence and Potential Pathogenesis. Diseases 2021; 9 [PMID: 34287285 DOI: 10.3390/diseases9030050]
- 26 Lozano-Sepulveda SA, Galan-Huerta K, Martínez-Acuña N, Arellanos-Soto D, Rivas-Estilla AM. SARS-CoV-2 another kind of liver aggressor, how does it do that? Ann Hepatol 2020; 19: 592-596 [PMID: 32858226 DOI: 10.1016/j.aohep.2020.08.062]
- 27 McGrowder DA, Miller F, Anderson Cross M, Anderson-Jackson L, Bryan S, Dilworth L. Abnormal Liver Biochemistry Tests and Acute Liver Injury in COVID-19 Patients: Current Evidence and Potential Pathogenesis. Diseases 2021; 9 [PMID: 34287285 DOI: 10.3390/diseases9030050]
- Sariani OK, Dousari AS, Moghadam MT. Possible psychological consequences in public of Omicron variant (B. 1.1. 529) 28 of SARS-CoV-2 identification in Iran. Iran J Psychiatry Behav Sci 2022; 16: 485-487 [DOI: 10.30699/ijmm.16.5.485]
- Liu F, Long X, Zhang B, Zhang W, Chen X, Zhang Z. ACE2 Expression in Pancreas May Cause Pancreatic Damage After 29 SARS-CoV-2 Infection. Clin Gastroenterol Hepatol 2020; 18: 2128-2130.e2 [PMID: 32334082 DOI: 10.1016/j.cgh.2020.04.040]
- Ozkurt Z, Çınar Tanrıverdi E. COVID-19: Gastrointestinal manifestations, liver injury and recommendations. World J Clin 30 Cases 2022; 10: 1140-1163 [PMID: 35211548 DOI: 10.12998/wjcc.v10.i4.1140]



- 31 Boeckmans J, Rodrigues RM, Demuyser T, Piérard D, Vanhaecke T, Rogiers V. COVID-19 and drug-induced liver injury: a problem of plenty or a petty point? Arch Toxicol 2020; 94: 1367-1369 [PMID: 32266419 DOI: 10.1007/s00204-020-02734-1]
- 32 D'Ardes D, Boccatonda A, Cocco G, Fabiani S, Rossi I, Bucci M, Guagnano MT, Schiavone C, Cipollone F. Impaired coagulation, liver dysfunction and COVID-19: Discovering an intriguing relationship. World J Gastroenterol 2022; 28: 1102-1112 [PMID: 35431501 DOI: 10.3748/wjg.v28.i11.1102]
- 33 Marjot T, Webb GJ, Barritt AS 4th, Moon AM, Stamataki Z, Wong VW, Barnes E. COVID-19 and liver disease: mechanistic and clinical perspectives. Nat Rev Gastroenterol Hepatol 2021; 18: 348-364 [PMID: 33692570 DOI: 10.1038/s41575-021-00426-4]
- 34 Sahu T, Pande B, Pl M, Verma HK. Liver dysfunction during COVID-19 pandemic: Contributing role of associated factors in disease progression and severity. World J Hepatol 2022; 14: 1099-1110 [PMID: 35978661 DOI: 10.4254/wjh.v14.i6.1099]
- 35 Sanyaolu A, Okorie C, Marinkovic A, Patidar R, Younis K, Desai P, Hosein Z, Padda I, Mangat J, Altaf M. Comorbidity and its Impact on Patients with COVID-19. SN Compr Clin Med 2020; 2: 1069-1076 [PMID: 32838147 DOI: 10.1007/s42399-020-00363-4]
- García LF. Immune Response, Inflammation, and the Clinical Spectrum of COVID-19. Front Immunol 2020; 11: 1441 [PMID: 32612615 DOI: 10.3389/fimmu.2020.01441]
- 37 Shah A. Novel Coronavirus-Induced NLRP3 Inflammasome Activation: A Potential Drug Target in the Treatment of COVID-19. Front Immunol 2020; 11: 1021 [PMID: 32574259 DOI: 10.3389/fimmu.2020.01021]
- 38 Costela-Ruiz VJ, Illescas-Montes R, Puerta-Puerta JM, Ruiz C, Melguizo-Rodríguez L. SARS-CoV-2 infection: The role of cytokines in COVID-19 disease. Cytokine Growth Factor Rev 2020; 54: 62-75 [PMID: 32513566 DOI: 10.1016/j.cytogfr.2020.06.001]
- 39 Gonçalves Júnior J. COVID-19, liver dysfunction and pathophysiology: A conceptual discussion. World J Gastroenterol 2022; 28: 683-688 [PMID: 35317425 DOI: 10.3748/wjg.v28.i6.683]
- Samprathi M, Jayashree M. Biomarkers in COVID-19: An Up-To-Date Review. Front Pediatr 2020; 8: 607647 [PMID: 40 33859967 DOI: 10.3389/fped.2020.607647]
- 41 Fan Z, Chen L, Li J, Cheng X, Yang J, Tian C, Zhang Y, Huang S, Liu Z, Cheng J. Clinical Features of COVID-19-Related Liver Functional Abnormality. Clin Gastroenterol Hepatol 2020; 18: 1561-1566 [PMID: 32283325 DOI: 10.1016/j.cgh.2020.04.002]
- Mantovani A, Beatrice G, Dalbeni A. Coronavirus disease 2019 and prevalence of chronic liver disease: A meta-analysis. 42 Liver Int 2020; 40: 1316-1320 [PMID: 32329563 DOI: 10.1111/liv.14465]
- Di Giorgio A, Hartleif S, Warner S, Kelly D. COVID-19 in Children With Liver Disease. Front Pediatr 2021; 9: 616381 43 [PMID: 33777864 DOI: 10.3389/fped.2021.616381]
- 44 Dhochak N, Singhal T, Kabra SK, Lodha R. Pathophysiology of COVID-19: Why Children Fare Better than Adults? Indian J Pediatr 2020; 87: 537-546 [PMID: 32410003 DOI: 10.1007/s12098-020-03322-y]
- Forster P, Forster L, Renfrew C, Forster M. Phylogenetic network analysis of SARS-CoV-2 genomes. Proc Natl Acad Sci 45 USA 2020; 117: 9241-9243 [PMID: 32269081 DOI: 10.1073/pnas.2004999117]
- Wu J, Shi C, Sheng X, Xu Y, Zhang J, Zhao X, Yu J, Shi X, Li G, Cao H, Li L. Prognostic Nomogram for Patients with 46 Hepatitis E Virus-related Acute Liver Failure: A Multicenter Study in China. J Clin Transl Hepatol 2021; 9: 828-837 [PMID: 34966646 DOI: 10.14218/JCTH.2020.00117]
- 47 Sun J, Aghemo A, Forner A, Valenti L. COVID-19 and liver disease. Liver Int 2020; 40: 1278-1281 [PMID: 32251539 DOI: 10.1111/liv.14470]
- Ji D, Qin E, Xu J, Zhang D, Cheng G, Wang Y, Lau G. Implication of non-alcoholic fatty liver diseases (NAFLD) in 48 patients with COVID-19: a preliminary analysis. J Hepatol 2020; 73 [DOI: 10.1016/j.jhep.2020.03.044]
- 49 Mao R, Doyon G, Gordon IO, Li J, Lin S, Wang J, Le THN, Elias M, Kurada S, Southern B, Olman M, Chen M, Zhao S, Dejanovic D, Chandra J, Mukherjee PK, West G, Van Wagoner DR, Fiocchi C, Rieder F. Activated intestinal muscle cells promote preadipocyte migration: a novel mechanism for creeping fat formation in Crohn's disease. Gut 2022; 71: 55-67 [PMID: 33468536 DOI: 10.1136/gutjnl-2020-323719]
- Word Health Organization. Opening remarks at the media briefing on COVID-19. [Internet] [accessed March 11, 2020]. 50 Available from:https://www.who.int/director-general/speeches/detail/who-director-general-s-opening-remarks-at-themedia-briefing-on-covid-19---11-march-2020
- 51 Liu W, Cheng H, Wang J, Ding L, Zhou Z, Liu S, Chang L, Rong Z. Clinical Analysis of Neonates Born to Mothers with or without COVID-19: A Retrospective Analysis of 48 Cases from Two Neonatal Intensive Care Units in Hubei Province. Am J Perinatol 2020; 37: 1317-1323 [PMID: 32892325 DOI: 10.1055/s-0040-1716505]
- 52 Liu W, Wang J, Li W, Zhou Z, Liu S, Rong Z. Clinical characteristics of 19 neonates born to mothers with COVID-19. Front Med 2020; 14: 193-198 [PMID: 32285380 DOI: 10.1007/s11684-020-0772-y]
- Wang S, Guo L, Chen L, Liu W, Cao Y, Zhang J, Feng L. A Case Report of Neonatal 2019 Coronavirus Disease in China. 53 Clin Infect Dis 2020; 71: 853-857 [PMID: 32161941 DOI: 10.1093/cid/ciaa225]
- Stolfi I, Conti MG, Marciano A, Dito L, Natale F, Bartolucci M, Cellitti R, Regoli D, Ticchiarelli A, Pangallo I, Pagano F, Ajassa C, Brunelli R, Terrin G. Liver Involvement in SARS-CoV-2 Vertically Infected Newborn: A Case Report. Front Pediatr 2021; 9: 701722 [PMID: 34858898 DOI: 10.3389/fped.2021.701722]
- 55 Vivanti AJ, Vauloup-Fellous C, Prevot S, Zupan V, Suffee C, Do Cao J, Benachi A, De Luca D. Transplacental transmission of SARS-CoV-2 infection. Nat Commun 2020; 11: 3572 [PMID: 32665677 DOI: 10.1038/s41467-020-17436-6]
- Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, Liu L, Shan H, Lei CL, Hui DSC, Du B, Li LJ, Zeng G, Yuen KY, Chen RC, Tang CL, Wang T, Chen PY, Xiang J, Li SY, Wang JL, Liang ZJ, Peng YX, Wei L, Liu Y, Hu YH, Peng P, Wang JM, Liu JY, Chen Z, Li G, Zheng ZJ, Qiu SQ, Luo J, Ye CJ, Zhu SY, Zhong NS; China Medical Treatment Expert Group for Covid-19. Clinical Characteristics of Coronavirus Disease 2019 in China. N Engl J Med 2020; 382: 1708-1720 [PMID: 32109013 DOI: 10.1056/NEJMoa2002032]



57 Mallet V, Beeker N, Bouam S, Sogni P, Pol S; Demosthenes research group. Prognosis of French COVID-19 patients with chronic liver disease: A national retrospective cohort study for 2020. J Hepatol 2021; 75: 848-855 [PMID: 33992699 DOI: 10.1016/j.jhep.2021.04.052]



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MINIREVIEWS

Immune-mediated liver injury following COVID-19 vaccination

Georgios Schinas, Eleni Polyzou, Vasiliki Dimakopoulou, Stamatia Tsoupra, Charalambos Gogos, Karolina Akinosoglou

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Georgios Schinas, Department of Medicine, University of Patras, Patras 26504, Greece

Eleni Polyzou, Vasiliki Dimakopoulou, Stamatia Tsoupra, Charalambos Gogos, Karolina Akinosoglou, Department of Internal Medicine, University of Patras, Patras 26504, Greece

Corresponding author: Karolina Akinosoglou, MD, PhD, Associate Professor, Department of Internal Medicine, University of Patras, 5th floor, University General Hospital of Patras, Patras 26504, Greece. akin@upatras.gr

Abstract

Liver injury secondary to vaccination is a rare adverse event that has recently come under attention thanks to the continuous pharmacovigilance following the widespread implementation of coronavirus disease 2019 (COVID-19) vaccination protocols. All three most widely distributed severe acute respiratory syndrome coronavirus 2 vaccine formulations, e.g., BNT162b2, mRNA-1273, and ChAdOx1-S, can induce liver injury that may involve immune-mediated pathways and result in autoimmune hepatitis-like presentation that may require therapeutic intervention in the form of corticosteroid administration. Various mechanisms have been proposed in an attempt to highlight immune checkpoint inhibition and thus establish causality with vaccination. The autoimmune features of such a reaction also prompt an in-depth investigation of the newly employed vaccine technologies. Novel vaccine delivery platforms, e.g., mRNA-containing lipid nanoparticles and adenoviral vectors, contribute to the inflammatory background that leads to an exaggerated immune response, while patterns of molecular mimicry between the spike (S) protein and prominent liver antigens may account for the autoimmune presentation. Immune mediators triggered by vaccination or vaccine ingredients per se, including autoreactive antibodies, cytokines, and cytotoxic T-cell populations, may inflict hepatocellular damage through wellestablished pathways. We aim to review available data associated with immunemediated liver injury associated with COVID-19 vaccination and elucidate potential mechanisms underlying its pathogenesis.

Key Words: Adverse effects; COVID-19 vaccines; mRNA vaccine; Autoimmune Hepatitis; Chemical and Drug Induced Liver Injury; Autoimmunity

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Core Tip: Following the worldwide implementation of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) vaccination protocols, several reports suggest an increase in the occurrence of autoimmune phenomena involving the liver. Studies on vaccine-induced liver injury point to a specific pattern of hepatocellular injury that involves immune-mediated pathways. This minireview explores the underlying pathophysiology of immune-mediated liver injury following SARS-CoV-2 vaccination and examines the most widely distributed vaccine formulations' autoimmune and hepatotoxic potential.

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INTRODUCTION

As of August 4, 2022, approximately 5.3 billion people around the world have received at least one dose of the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) vaccine. Widespread implementation of vaccination protocols has successfully contained the spread of the pandemic and has reduced the disease burden for patients and health systems alike[1]. Newly employed vaccination platforms, e.g., mRNA- and adenovirus (AdV)- based formulations, have achieved high efficacy rates combined with a good safety profile[2]. SARS-CoV-2 vaccines have undergone the most intensive safety monitoring in the history of mankind. Both active and passive monitoring systems have been employed in order to timely detect and properly identify adverse events related to vaccination[3,4]. This worldwide vigilance has proved fruitful for epidemiological purposes and has been instrumental in ensuring public support for vaccination. Most frequently reported adverse events have been mild in nature and local in character. They primarily concern injection site-related reactions, e.g., topical pain and redness or generalized systemic symptoms, like fever and fatigue [5,6]. As far as serious, organspecific adverse events are concerned, a very low risk of myocarditis mainly in younger individuals has been linked to vaccination with an mRNA vaccine, whereas adenoviral vector vaccines have been associated with incidents of thrombosis accompanied by thrombocytopenia and possibly Guillain-Barré Syndrome (GBS) cases. A rather rare side effect that has recently come under attention is that of liver injury following vaccination with a SARS-CoV-2 vaccine.

Drug-induced liver injury (DILI), under the umbrella of which such a clinical syndrome would initially be examined, is characterized by new-onset, profound increases in liver function enzyme levels. According to the latest expert panel update, this is defined as $a \ge 5 \times$ upper limit of normal (ULN) elevation of alanine (ALT) or aspartate aminotransferase (AST) and/or $\ge 2 \times$ ULN increase in alkaline phosphatase (ALP) levels or ALT/AST $\ge 3 \times$ UNL and bilirubin $\ge 2 \times$ ULN[7]. Upon removal of the offending agent, most cases of DILI are usually self-contained; corticosteroids are sometimes added to the therapeutic regimen if autoimmune features are demonstrated. In fact, most of the reported cases' clinical and histological features closely resemble those encountered in autoimmune hepatitis (AIH)[8, 9], steering the focus of the causality investigation onto the immune-mediated background of the reaction. However, it remains unclear whether the reported association of AIH with vaccination is coincidental, represents unique SARS-CoV-2-induced antigen-specific immune activation or is associated with transient drug-induced liver injury. In this study, we aim to review underlying mechanisms driving immune-mediated liver injury following COVID-19 vaccination and discuss potential implications

METHODS

We carried out broad searches of PubMed, Scopus, and Embase between 1 January 2021 and 1 September 2022 to identify literature describing immune-mediated liver injury or autoimmune hepatitis following COVID-19 vaccination. Relevant publications were identified based on the titles and abstracts. No restriction on the type of paper or language was set, even though the main focus was put on underlying mechanisms. Two reviewers independently screened all titles/abstracts and hand-searched references of retrieved articles. Data were assessed for their quality based on overall judgement and not aggregate scores. Disagreements were discussed and resolved and duplicates were removed.

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VACCINE-INDUCED AUTOIMMUNITY

Throughout the vaccine rollout period, there have been reported cases of presumed AIH that were attributed to COVID-19 vaccination because they were observed shortly after either the first or second dose of the vaccine, with the initial case described as early as January 2021[10]. AIH is part of a diverse group of chronic inflammatory liver conditions that include primary biliary cholangitis and primary sclerosing cholangitis, and its complex pathophysiology involves underlying genetic predisposition and interactions with environmental triggers[11]. Viral infections, drug exposure, and vaccinations have been implicated in the pathogenesis of AIH[12]. AIH had fallen under the radar during the phase 3 clinical trials of all vaccines, and like every other rare adverse event, much debate has ensued over its association with the vaccination. New onset autoimmune reactions following vaccination have previously been described in the literature[13]. Both Hepatitis A and Hepatitis B vaccines have been linked to the development of AIH-like conditions[14]. Human papillomavirus, Hepatitis B, and Influenza vaccines have been held accountable for autoimmune reactions[15,16]. Molecular mimicry theory has been the platform upon which causality with vaccination has been determined[17]. It has been hypothesized that individuals with a genetic predisposition to autoimmunity undergo vaccination, and similarly to other environmental inputs, e.g., smoking and nutrition, their immune tolerance becomes compromised. Reportedly, susceptible groups include those with systemic lupus erythematosus, GBS, multiple sclerosis, and narcolepsy. Concerning SARS-CoV-2 vaccination, immune-based phenomena such as GBS, IgA nephropathy, immune thrombotic thrombocytopenia, and myocarditis have been linked to both novel vaccine platforms, *i.e.*, mRNA- and AdV-based formulations[18].

VACCINE-INDUCED LIVER INJURY

As of early 2021, case reports of documented AIH following COVID-19 vaccination have begun to emerge^[19-21]. We estimate that AIH related to COVID-19 is almost 1 in 14 million, even though we do acknowledge that many cases remain undocumented^[19]. Data mostly deriving from comprehensive case-series reporting liver injury following vaccination with SARS-CoV-2 vaccines, point to the fact that most cases are in fact, immune-related, with 57% of all patients displaying both autoantibody presence and IgG hyperglobulinemia. They mostly affected elderly females, with most of the reports originating from European countries, followed by the United States^[20]. The mean time of symptom onset is close to three weeks following the first vaccination, with some individuals presenting as early as 3 d after and others coming in as late as a month later, suggesting some heterogeneity in the underlying response mechanism. The mean duration between receiving the first or second vaccine dose and subsequent onset of liver injury was 17.3 (11.2-23.4) days and was mostly associated with mRNA vaccines, possibly to their stronger immunogenic potency[20]. The presence of underlying autoimmune diseases (e.g. Hashimoto thyroiditis, primary sclerosing cholangitis) is evident in approximately 25% of patients and could explain temporal and spatial differences in manifestations and prevalence, respectively, according to genetic predispositions^[22]. Antinuclear antibody (ANA) was by far the most prevalent autoantibody, followed by spinal muscular atrophy and anti-myocardial antibody (AMA), resembling a type 1 AIH pattern. Biopsy findings were also consistent with AIH in most individuals. Around 1/3 of those who did not undergo the diagnostic procedures fit the clinical profile of AIH. Icteric manifestations, including jaundice, choluria, and pruritus, account for around 2/3 of all presentations. Outcomes were similar in all three vaccine products, i.e., BNT162b2, mRNA-1273 and ChAdOx1 nCoV-19. Although recovery time varied greatly among study populations, the mean time for transaminase normalization was calculated at 46 d[21]. Corticosteroid treatment proved safe and effective for all those who were prescribed. No relapse was noted in the subgroup of patients whose immunosuppressant treatment was discontinued and remission was maintained in all those who spontaneously recovered.

CAUSALITY OR CASUALITY

Establishing causality is by definition, a difficult task, while the mechanism of action of such a reaction remains elusive. Several theories have been proposed in an attempt to link clinical manifestations of hepatocellular injury to patterns of immune mediation involving vaccine ingredients and products. Molecular mimicry-based reactivity and pro-inflammatory interactions involving the SARS-CoV-2 spike protein have been explored (Figure 1)[23]. The vaccine adjuvants have also come under scrutiny. The BNT162b2 and mRNA-1273 vaccines employ lipid nanoparticle (LNP) coated mRNA technology, whereas the ChAdOx1 nCoV-19 vaccine is deoxyribonucleic acid based and utilizes AdV vectors. Both AdV and mRNA vaccine platforms are newly licensed; hence, many rare *in vivo* interactions are to be explored and clarified. A deeper look into the active ingredients of the vaccines may provide us with a plausible mechanism. The mRNA itself has been carefully designed and tested as to its immunogenic properties[24,25]; however, prior to translation, it may still be recognized by cytosolic and endosomal toll-like receptors. The encoded S-spike protein elicits a strong immune reaction that involves the



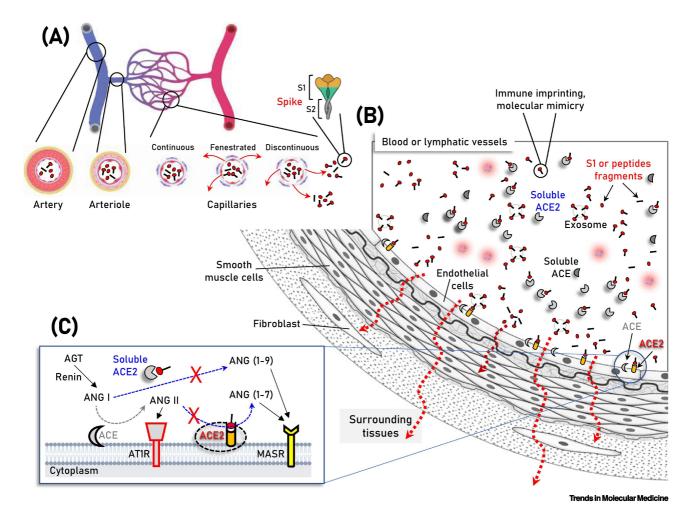


Figure 1 Schematic of the vasculature components showing vaccination-produced S protein/subunits/peptide fragments in the circulation, as well as soluble or endothelial cell membrane-attached angiotensin-converting enzyme 2. A: Parallel to immune system activation, circulating S protein/subunits/peptide fragments; B: Binding to angiotensin-converting enzyme 2 (ACE2) may occur not only to ACE2-expressing endothelial cells, but also in multiple cell types of the vasculature and surrounding tissues due to antigen diffusion (e.g., in fenestrated or discontinuous capillary beds) (A, red arrows). These series of molecular events are unlikely for any severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)-related antigen in the absence of severe coronavirus disease 2019, where SARS-CoV-2 is contained in the respiratory system; C: In the two counteracting pathways of the renin-angiotensin system (RAS), namely the 'conventional' arm, that involves ACE which generates angiotensin II (ANG II) from angiotensin I (ANG I), and the ACE2 arm which hydrolyzes ANG II to generate angiotensin (1-7) [ANG (1-7)] or ANG I to generate angiotensin (1-9) [ANG (1-9)] are depicted. ANG II binding and activation of the ANG II type 1 receptor (AT1R) promotes inflammation, fibrotic remodeling, and vasoconstriction, whereas the ANG (1-7) and ANG (1-9) peptides binding to MAS receptor (MASR) activate antifibrotic, anti-inflammatory pathways and vasodilation. Additional modules of the RAS (i.e., renin and angiotensinogen, AGT) are also shown. AT1R: Angiotensin II type 1 receptor. Citation: Trougakos IP, Terpos E, Alexopoulos H, Politou M, Paraskevis D, Scorilas A, Kastritis E, Andreakos E, Dimopoulos MA. Adverse effects of COVID-19 mRNA vaccines: the spike hypothesis. Trends Mol Med 2022; 28: 542-554. Copyright ©The Author(s) 2022. Published by Elsevier.

> activation of the innate inflammatory cascade, as well as that of the adaptive humoral response. Regarding the former, SARS-CoV-2 vaccines employ the type I interferon pathway in particular, in order to maintain an adequate and effective immune response, that in turn, may increase the probability of an autoimmune occurrence in certain individuals[26]. Concerning the latter, reactivity between anti-S protein antibodies and human tissue antigens has been confirmed by a recent report^[27]. The systemic distribution of the spike protein has also been postulated as a mechanism to explain adverse events by mRNA-based vaccines as well. Its interaction with soluble Angiotensin-converting enzyme 2 (ACE-2) and ACE-2-ligands may point to an organ-specific pattern of insult[28]. The presentation and/or production of the spike protein by the hepatocytes may induce the activation of cytotoxic T-cell subsets. Under this scope, the formation of immune complexes cannot be excluded. Their subsequent deposition on the liver may cause inflammation or exacerbation of the underlying autoimmune disease. Matyushkina et al^[27] have identified the susceptibility of human leukocyte antigen (HLA) B15:01 and HLA B39:01 allele carriers to autoimmunity following COVID-19. HLA B15 has been strongly associated with the development of infliximab-induced liver injury^[29], while HLA B39 has been recorded as one of the most prevalent alleles in AIH patients in Pakistan[30]. In the same report, although most autoreactive antibodies were associated with nuclear products, cross-reactivity with cytokeratin 18 (CK18), a prominent liver disease biomarker, was noted. Elevated anti-CK18 antibody titers have been described in AIH patients[31], and their relationship to the soluble liver antigen (SLA) has been established in the

literature. CK18's immunoreactivity has even been proposed but disproved as a potential diagnostic marker for the SLA subgroup of AIH patients[32]. The presence of SLA antibodies has been reported twice following vaccination with the mRNA-1273 vaccine, a fact that prompted investigators to conduct a genomic sequence analysis study which revealed, however, no homology between the SARS-CoV-2 spike protein and soluble liver antigen[33].

As far as AdV vectors are concerned, their clinical application as potential gene therapy delivery particles has been hindered by their hepatotoxic properties since the start of the century [34]. Recent reports have attributed this to their inherent liver tropism[34]. It should also be noted that the recombinant ChAdOx1-S AdV used in Vaxzervria formulations is likely hepatotropic since it is derived from a subset of non-human, Y25-coded adenoviruses that have been linked to viral hepatitis outbreaks in chimpanzees in the past[34]. In the same report, the authors build a case for a post-transcriptional modification taking place inside the nucleus of AdV-transduced host cells, resulting in alternate gene splicing and subsequent truncation of produced S-protein proteins that may in turn, be released in circulation. In addition to that, they demonstrated that in ChAdOx1-S-transduced hepatocytes, the truncated S-protein is the main splicing product, thereby providing us with another plausible mechanism to explain liver injury by AdV-based formulations. Of note, the spike protein produced by Vaxzervria has comparable receptor binding selection and affinity to its original counterpart[35]. Regarding common vaccine adjuvants, CpG 1018 and Aluminum, although widely used for immune response enhancement purposes and deemed safe by clinical trials[36] and regulatory authorities alike, have the potential to induce liver injury[37] and likely precipitate the development of auto-immune disease in a small percentage of the population[38]. Reportedly, none of the vaccines discussed in this review contain the aforementioned adjuvants, but future formulations may include them. Furthermore, we need to consider other vaccine formulation specificities, like the active ingredient's delivery system. The immunogenicity of the mRNA-containing LNPs has recently come under question, despite the fact that prior to their use in COVID-19 vaccines, they were being hailed as a potential genetic treatment platform for inherited liver disease[39]. The mRNA delivery particles have been linked to the development of allergic reactions^[40], suggesting a plausible, if not definite, role as immune mediators. The LNP platform mounts a strong immune response, which relies on the medium's pro-inflammatory properties for its efficacy. Such an immune response-provoking environment could potentiate a loss in self-tolerance. LNPs have been known to act as adjuvants to vaccine-induced immune reactions[41,42]. In particular, LNPs seem to trigger the NLRP3 inflammasome pathway that has been implicated in the pathogenesis of other autoimmune phenomena, like pericarditis, rheumatoid arthritis and AIH[43-46]. The intense immunogenic character of LNPs has been demonstrated both histochemically and graphically in animal models through multiple route administration of its purified form, e.g., intramuscularly and intranasally.

The hepatocellular type of injury that is predominantly associated with post-vaccination liver injury can also be attributed to the direct action of cytotoxic T-lymphocytes, as rapid and sustained activation of this cell subset has been confirmed in the context of SARS-CoV-2 vaccination[47]. In a recent report, vaccination with the BNT162b2 vaccine resulted in a CD8+ rich lymphocytic infiltrate in the liver of a patient that presented with probable AIH. The clonal expansion and peripheral activation state of this particular subset of lymphocytes correlated closely with the clinical course of hepatitis in this individual, suggesting T-cells' involvement in the development and resolution of the disease[48]. It has also been demonstrated, in animal models, that cytokine-activated, "bystander" CD8+ lymphocytes may cause hepatocellular injury even in the absence of a direct antigen[49]. Accumulation of cytotoxic infiltrates in the liver has been reported in the literature following acute infection in influenza pneumonia^[50]. All the plausible mechanisms resulting in immune-mediated liver injury discussed above are presented concisely in Table 1.

DIAGNOSIS

The immune-mediated mechanism of a clinical syndrome involving hepatocellular injury would most likely be highlighted by an elevation in ANA and/or AMA titers in a similar fashion to AIH. A report from early on in the pandemic noted the presence of elevated autoimmunity markers, including ANA and AMA, in SARS-CoV-2 antibody-rich plasma, thereby suggesting their self-reactive potential[51]. However, otherwise typical AIH auto-antibodies may be present in the acute phase of liver injury by multiple causes[52]. In order to distinguish between them, a biopsy is the preferred option, with fibrosis being the prime differentiating factor[53]. Features of widespread fibrosis would be evident in an AIHstricken liver[54], whereas evidence of acute or chronic inflammation with eosinophilic infiltration between or within the portal triads is to be expected in the case of direct liver toxicity, *i.e.*, DILI[55]. It is important to note that centrilobular necrosis is not a pathognomonic clue and should not be interpreted as such[56]. All in all, a definitive diagnosis of AIH may be challenging to make, as it relies on a constellation of clinical, serological and histological findings. Response to treatment with immunosuppressants is the only way to confirm a diagnosis [57]. AIH is a chronic condition with a high relapse rate if immunosuppression is withdrawn, whereas causes closely resembling AIH do not usually relapse[58].



Table 1 Immune-mediated phenomena concerning the liver, following severe acute respiratory syndrome coronavirus 2 vaccination			
Mechanism of injury	Liver antigens	Immune mediators	
Molecular mimicry	SLA	Autoreactive antibodies	
	CK-18		
Cytotoxicity/Humoral response	S protein (membrane expression)	Activated CD8 + clone/protective anti-S antibodies	
Humoral response	ACE-2 transmembrane receptor	Protective anti-S antibodies	
Immune complex deposition	S protein (soluble)		
	Soluble ACE-2/ACE-2 ligand + S protein		
"Bystander" toxicity		Activated CD8 + clone	
Loss of self-tolerance/Fibrosis		Type I IFN	
NLRP3 inflammasome activation		LNPs	
TLR-mediated innate immune response		mRNA	

SLA: Soluble Liver Antigen; CK-18: Cytokeratin 18; S protein: Spike protein; anti-S: Anti-Spike protein; ACE-2: Angiotensin Converting Enzyme-2; IFN: Interferon; LNPs: Lipid nanoparticles; NLRP3: NLR family Pyrin domain containing 3; TLR: Toll-like receptor.

CONCLUSION

Immune-mediated liver injury remains an elusive but rare entity following COVID-19 vaccination. It is the responsibility of investigators and scientists worldwide to maintain a vigilant eye and continue reporting rare incidents related to vaccination with a high index of suspicion. However, adverse events as such, are significantly less frequent than potentially serious complications of COVID-19 disease[59] and should by no means discourage vaccination programs worldwide.

FOOTNOTES

Author contributions: Akinosoglou K and Gogos C conceived the idea; Schinas G performed the literature search, reviewed data, and wrote the manuscript; Polyzou E, Dimakopoulou V, and Tsoupra S performed the literature search and reviewed data; Akinosoglou K and Gogos C critically corrected the manuscript; Akinosoglou K revised the manuscript

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Country/Territory of origin: Greece

ORCID number: Georgios Schinas 0000-0001-7963-1865; Charalambos Gogos 0000-0002-1165-964X; Karolina Akinosoglou 0000-0002-4289-9494.

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REFERENCES

1 Tenforde MW, Patel MM, Ginde AA, Douin DJ, Talbot HK, Casey JD, Mohr NM, Zepeski A, Gaglani M, McNeal T, Ghamande S, Shapiro NI, Gibbs KW, Files DC, Hager DN, Shehu A, Prekker ME, Erickson HL, Exline MC, Gong MN, Mohamed A, Henning DJ, Steingrub JS, Peltan ID, Brown SM, Martin ET, Monto AS, Khan A, Hough CT, Busse L, Lohuis CCT, Duggal A, Wilson JG, Gordon AJ, Qadir N, Chang SY, Mallow C, Gershengorn HB, Babcock HM, Kwon JH, Halasa N, Chappell JD, Lauring AS, Grijalva CG, Rice TW, Jones ID, Stubblefield WB, Baughman A, Womack KN, Lindsell CJ, Hart KW, Zhu Y, Olson SM, Stephenson M, Schrag SJ, Kobayashi M, Verani JR, Self WH; Influenza and



Other Viruses in the Acutely III (IVY) Network. Effectiveness of SARS-CoV-2 mRNA Vaccines for Preventing Covid-19 Hospitalizations in the United States. medRxiv 2021 [PMID: 34268515 DOI: 10.1101/2021.07.08.21259776]

- 2 Sharif N, Alzahrani KJ, Ahmed SN, Dey SK. Efficacy, Immunogenicity and Safety of COVID-19 Vaccines: A Systematic Review and Meta-Analysis. Front Immunol 2021; 12: 714170 [PMID: 34707602 DOI: 10.3389/fimmu.2021.714170]
- 3 Barda N, Dagan N, Ben-Shlomo Y, Kepten E, Waxman J, Ohana R, Hernán MA, Lipsitch M, Kohane I, Netzer D, Reis BY, Balicer RD. Safety of the BNT162b2 mRNA Covid-19 Vaccine in a Nationwide Setting. N Engl J Med 2021; 385: 1078-1090 [PMID: 34432976 DOI: 10.1056/NEJMoa2110475]
- 4 Baden LR, El Sahly HM, Essink B, Kotloff K, Frey S, Novak R, Diemert D, Spector SA, Rouphael N, Creech CB, McGettigan J, Khetan S, Segall N, Solis J, Brosz A, Fierro C, Schwartz H, Neuzil K, Corey L, Gilbert P, Janes H, Follmann D, Marovich M, Mascola J, Polakowski L, Ledgerwood J, Graham BS, Bennett H, Pajon R, Knightly C, Leav B, Deng W, Zhou H, Han S, Ivarsson M, Miller J, Zaks T; COVE Study Group. Efficacy and Safety of the mRNA-1273 SARS-CoV-2 Vaccine. N Engl J Med 2021; 384: 403-416 [PMID: 33378609 DOI: 10.1056/NEJMoa2035389]
- 5 Polack FP, Thomas SJ, Kitchin N, Absalon J, Gurtman A, Lockhart S, Perez JL, Pérez Marc G, Moreira ED, Zerbini C, Bailey R, Swanson KA, Roychoudhury S, Koury K, Li P, Kalina WV, Cooper D, Frenck RW Jr, Hammitt LL, Türeci Ö, Nell H, Schaefer A, Ünal S, Tresnan DB, Mather S, Dormitzer PR, Şahin U, Jansen KU, Gruber WC; C4591001 Clinical Trial Group. Safety and Efficacy of the BNT162b2 mRNA Covid-19 Vaccine. N Engl J Med 2020; 383: 2603-2615 [PMID: 33301246 DOI: 10.1056/NEJMoa2034577]
- Voysey M, Clemens SAC, Madhi SA, Weckx LY, Folegatti PM, Aley PK, Angus B, Baillie VL, Barnabas SL, Bhorat QE, Bibi S, Briner C, Cicconi P, Collins AM, Colin-Jones R, Cutland CL, Darton TC, Dheda K, Duncan CJA, Emary KRW, Ewer KJ, Fairlie L, Faust SN, Feng S, Ferreira DM, Finn A, Goodman AL, Green CM, Green CA, Heath PT, Hill C, Hill H, Hirsch I, Hodgson SHC, Izu A, Jackson S, Jenkin D, Joe CCD, Kerridge S, Koen A, Kwatra G, Lazarus R, Lawrie AM, Lelliott A, Libri V, Lillie PJ, Mallory R, Mendes AVA, Milan EP, Minassian AM, McGregor A, Morrison H, Mujadidi YF, Nana A, O'Reilly PJ, Padayachee SD, Pittella A, Plested E, Pollock KM, Ramasamy MN, Rhead S, Schwarzbold AV, Singh N, Smith A, Song R, Snape MD, Sprinz E, Sutherland RK, Tarrant R, Thomson EC, Török ME, Toshner M, Turner DPJ, Vekemans J, Villafana TL, Watson MEE, Williams CJ, Douglas AD, Hill AVS, Lambe T, Gilbert SC, Pollard AJ; Oxford COVID Vaccine Trial Group. Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. Lancet 2021; 397: 99-111 [PMID: 33306989 DOI: 10.1016/S0140-6736(20)32661-1]
- 7 Hayashi PH, Lucena MI, Fontana RJ, Bjornsson ES, Aithal GP, Barnhart H, Gonzalez-Jimenez A, Yang Q, Gu J, Andrade RJ, Hoofnagle JH. A revised electronic version of RUCAM for the diagnosis of DILI. Hepatology 2022; 76: 18-31 [PMID: 35014066 DOI: 10.1002/hep.32327]
- Mungmunpuntipantip R, Wiwanitkit V. Letter to the editor: "Autoimmune hepatitis after COVID-19 vaccination". 8 Hepatology 2022; 75: 756 [PMID: 34797931 DOI: 10.1002/hep.32249]
- 9 Palla P, Vergadis C, Sakellariou S, Androutsakos T. Letter to the editor: Autoimmune hepatitis after COVID-19 vaccination: A rare adverse effect? Hepatology 2022; 75: 489-490 [PMID: 34528278 DOI: 10.1002/hep.32156]
- Bril F, Al Diffalha S, Dean M, Fettig DM. Autoimmune hepatitis developing after coronavirus disease 2019 (COVID-19) 10 vaccine: Causality or casualty? J Hepatol 2021; 75: 222-224 [PMID: 33862041 DOI: 10.1016/j.jhep.2021.04.003]
- Liaskou E, Hirschfield GM, Gershwin ME. Mechanisms of tissue injury in autoimmune liver diseases. Semin 11 Immunopathol 2014; 36: 553-568 [PMID: 25082647 DOI: 10.1007/s00281-014-0439-3]
- Lammert C, Chalasani SN, Atkinson EJ, McCauley BM, Lazaridis KN. Environmental risk factors are associated with 12 autoimmune hepatitis. Liver Int 2021; 41: 2396-2403 [PMID: 33978301 DOI: 10.1111/liv.14944]
- Watad A, Bragazzi NL, McGonagle D, Adawi M, Bridgewood C, Damiani G, Alijotas-Reig J, Esteve-Valverde E, 13 Quaresma M, Amital H, Shoenfeld Y. Autoimmune/inflammatory syndrome induced by adjuvants (ASIA) demonstrates distinct autoimmune and autoinflammatory disease associations according to the adjuvant subtype: Insights from an analysis of 500 cases. Clin Immunol 2019; 203: 1-8 [PMID: 30922961 DOI: 10.1016/j.clim.2019.03.007]
- van Gemeren MA, van Wijngaarden P, Doukas M, de Man RA. Vaccine-related autoimmune hepatitis: the same disease as 14 idiopathic autoimmune hepatitis? Scand J Gastroenterol 2017; 52: 18-22 [PMID: 27565372 DOI: 10.1080/00365521.2016.1224379]
- Pellegrino P, Carnovale C, Pozzi M, Antoniazzi S, Perrone V, Salvati D, Gentili M, Brusadelli T, Clementi E, Radice S. 15 On the relationship between human papilloma virus vaccine and autoimmune diseases. Autoimmun Rev 2014; 13: 736-741 [PMID: 24468416 DOI: 10.1016/j.autrev.2014.01.054]
- 16 Wraith DC, Goldman M, Lambert PH. Vaccination and autoimmune disease: what is the evidence? Lancet 2003; 362: 1659-1666 [PMID: 14630450 DOI: 10.1016/s0140-6736(03)14802-7]
- 17 Segal Y, Shoenfeld Y. Vaccine-induced autoimmunity: the role of molecular mimicry and immune crossreaction. Cell Mol Immunol 2018; 15: 586-594 [PMID: 29503439 DOI: 10.1038/cmi.2017.151]
- Chen Y, Xu Z, Wang P, Li XM, Shuai ZW, Ye DQ, Pan HF. New-onset autoimmune phenomena post-COVID-19 18 vaccination. Immunology 2022; 165: 386-401 [PMID: 34957554 DOI: 10.1111/imm.13443]
- 19 Chow KW, Pham NV, Ibrahim BM, Hong K, Saab S. Autoimmune Hepatitis-Like Syndrome Following COVID-19 Vaccination: A Systematic Review of the Literature. Dig Dis Sci 2022; 67: 4574-4580 [PMID: 35486203 DOI: 10.1007/s10620-022-07504-w]
- Roy A, Verma N, Singh S, Pradhan P, Taneja S, Singh M. Immune-mediated liver injury following COVID-19 vaccination: 20 A systematic review. Hepatol Commun 2022; 6: 2513-2522 [PMID: 35507736 DOI: 10.1002/hep4.1979]
- Efe C, Kulkarni AV, Terziroli Beretta-Piccoli B, Magro B, Stättermayer A, Cengiz M, Clayton-Chubb D, Lammert C, 21 Bernsmeier C, Gül Ö, la Tijera FH, Anders M, Lytvyak E, Akın M, Purnak T, Liberal R, Peralta M, Ebik B, Duman S, Demir N, Balaban Y, Urzua Á, Contreras F, Venturelli MG, Bilgiç Y, Medina A, Girala M, Günşar F, Londoño MC, Androutsakos T, Kisch A, Yurci A, Güzelbulut F, Çağın YF, Avcı E, Akyıldız M, Dindar-Demiray EK, Harputluoğlu M, Kumar R, Satapathy SK, Mendizabal M, Silva M, Fagiuoli S, Roberts SK, Soylu NK, Idilman R, Yoshida EM, Montano-Loza AJ, Dalekos GN, Ridruejo E, Schiano TD, Wahlin S. Liver injury after SARS-CoV-2 vaccination: Features of immune-mediated hepatitis, role of corticosteroid therapy and outcome. Hepatology 2022; 76: 1576-1586 [PMID:



35567545 DOI: 10.1002/hep.32572]

- 22 Alhumaid S, Al Mutair A, Rabaan AA, ALShakhs FM, Choudhary OP, Yong SJ, Nainu F, Khan A, Muhammad J, Alhelal F, Al Khamees MH, Alsouaib HA, Al Majhad AS, Al-Tarfi HR, ALyasin AH, Alatiyyah YY, Alsultan AA, Alessa ME, Alissa MA, Alsayegh EH, Alshakhs HN, Al Samaeel HA, AlShayeb RA, Alnami DA, Alhassan HA, Alabdullah AA, Alhmed AH, AlDera FH, Hajissa K, Al-Tawfiq JA, Al-Omari A. New-onset and relapsed liver diseases following COVID-19 vaccination: a systematic review. BMC Gastroenterol 2022; 22: 433 [PMID: 36229799 DOI: 10.1186/s12876-022-02507-3]
- Trougakos IP, Terpos E, Alexopoulos H, Politou M, Paraskevis D, Scorilas A, Kastritis E, Andreakos E, Dimopoulos MA. 23 Adverse effects of COVID-19 mRNA vaccines: the spike hypothesis. Trends Mol Med 2022; 28: 542-554 [PMID: 35537987 DOI: 10.1016/j.molmed.2022.04.007]
- 24 Pelka K, Shibata T, Miyake K, Latz E. Nucleic acid-sensing TLRs and autoimmunity: novel insights from structural and cell biology. Immunol Rev 2016; 269: 60-75 [PMID: 26683145 DOI: 10.1111/imr.12375]
- 25 Karikó K, Muramatsu H, Welsh FA, Ludwig J, Kato H, Akira S, Weissman D. Incorporation of pseudouridine into mRNA yields superior nonimmunogenic vector with increased translational capacity and biological stability. Mol Ther 2008; 16: 1833-1840 [PMID: 18797453 DOI: 10.1038/mt.2008.200]
- 26 Teijaro JR, Farber DL. COVID-19 vaccines: modes of immune activation and future challenges. Nat Rev Immunol 2021; **21**: 195-197 [PMID: 33674759 DOI: 10.1038/s41577-021-00526-x]
- 27 Matyushkina D, Shokina V, Tikhonova P, Manuvera V, Shirokov D, Kharlampieva D, Lazarev V, Varizhuk A, Vedekhina T, Pavlenko A, Penkin L, Arapidi G, Pavlov K, Pushkar D, Kolontarev K, Rumyantsev A, Rumyantsev S, Rychkova L, Govorun V. Autoimmune Effect of Antibodies against the SARS-CoV-2 Nucleoprotein. Viruses 2022; 14 [PMID: 35746613 DOI: 10.3390/v14061141]
- Ziegler CGK, Allon SJ, Nyquist SK, Mbano IM, Miao VN, Tzouanas CN, Cao Y, Yousif AS, Bals J, Hauser BM, Feldman J, Muus C, Wadsworth MH 2nd, Kazer SW, Hughes TK, Doran B, Gatter GJ, Vukovic M, Taliaferro F, Mead BE, Guo Z, Wang JP, Gras D, Plaisant M, Ansari M, Angelidis I, Adler H, Sucre JMS, Taylor CJ, Lin B, Waghray A, Mitsialis V, Dwyer DF, Buchheit KM, Boyce JA, Barrett NA, Laidlaw TM, Carroll SL, Colonna L, Tkachev V, Peterson CW, Yu A, Zheng HB, Gideon HP, Winchell CG, Lin PL, Bingle CD, Snapper SB, Kropski JA, Theis FJ, Schiller HB, Zaragosi LE, Barbry P, Leslie A, Kiem HP, Flynn JL, Fortune SM, Berger B, Finberg RW, Kean LS, Garber M, Schmidt AG, Lingwood D, Shalek AK, Ordovas-Montanes J; HCA Lung Biological Network. HCA Lung Biological Network. SARS-CoV-2 Receptor ACE2 Is an Interferon-Stimulated Gene in Human Airway Epithelial Cells and Is Detected in Specific Cell Subsets across Tissues. Cell 2020; 181: 1016-1035.e19 [PMID: 32413319 DOI: 10.1016/j.cell.2020.04.035]
- 29 Bruno CD, Fremd B, Church RJ, Daly AK, Aithal GP, Björnsson ES, Larrey D, Watkins PB, Chow CR. HLA associations with infliximab-induced liver injury. Pharmacogenomics J 2020; 20: 681-686 [PMID: 32024945 DOI: 10.1038/s41397-020-0159-0
- Hassan N, Siddiqui AR, Abbas Z, Hassan SM, Soomro GB, Mubarak M, Anis S, Muzaffar R, Zafar MN. Clinical Profile 30 and HLA Typing of Autoimmune Hepatitis From Pakistan. Hepat Mon 2013; 13: e13598 [PMID: 24358040 DOI: 10.5812/hepatmon.13598
- 31 Murota M, Nishioka M, Fujita J, Dobashi N, Wu F, Ohtsuki Y, Hojo S, Takahara J, Kuriyama S. Anti-cytokeratin antibodies in sera of the patients with autoimmune hepatitis. Clin Exp Immunol 2001; 125: 291-299 [PMID: 11529922 DOI: 10.1046/j.1365-2249.2001.01568.x]
- Martin L, Bäurle A, Fiehn W, Volkmann M. Immunoreactivity to cytokeratin 8/18 in patients with soluble liver antigen 32 (SLA) positive autoimmune hepatitis (AIH type 3) is not sufficient for diagnostic use. Clin Lab 2000; 46: 339-344 [PMID: 10934580]
- 33 Londoño MC, Gratacós-Ginès J, Sáez-Peñataro J. Another case of autoimmune hepatitis after SARS-CoV-2 vaccination still casualty? J Hepatol 2021; 75: 1248-1249 [PMID: 34129886 DOI: 10.1016/j.jhep.2021.06.004]
- Kowarz E, Krutzke L, Külp M, Streb P, Larghero P, Reis J, Bracharz S, Engler T, Kochanek S, Marschalek R. Vaccineinduced COVID-19 mimicry syndrome. Elife 2022; 11 [PMID: 35084333 DOI: 10.7554/eLife.74974]
- 35 Watanabe Y, Mendonça L, Allen ER, Howe A, Lee M, Allen JD, Chawla H, Pulido D, Donnellan F, Davies H, Ulaszewska M, Belij-Rammerstorfer S, Morris S, Krebs AS, Dejnirattisai W, Mongkolsapaya J, Supasa P, Screaton GR, Green CM, Lambe T, Zhang P, Gilbert SC, Crispin M. Native-like SARS-CoV-2 Spike Glycoprotein Expressed by ChAdOx1 nCoV-19/AZD1222 Vaccine. ACS Cent Sci 2021; 7: 594-602 [PMID: 34056089 DOI: 10.1021/acscentsci.1c00080
- 36 Hsieh SM, Liu MC, Chen YH, Lee WS, Hwang SJ, Cheng SH, Ko WC, Hwang KP, Wang NC, Lee YL, Lin YL, Shih SR, Huang CG, Liao CC, Liang JJ, Chang CS, Chen C, Lien CE, Tai IC, Lin TY. Safety and immunogenicity of CpG 1018 and aluminium hydroxide-adjuvanted SARS-CoV-2 S-2P protein vaccine MVC-COV1901: interim results of a large-scale, double-blind, randomised, placebo-controlled phase 2 trial in Taiwan. Lancet Respir Med 2021; 9: 1396-1406 [PMID: 34655522 DOI: 10.1016/S2213-2600(21)00402-1]
- Zhang N, Li K, Liu Z, Nandakumar KS, Jiang S. A Perspective on the Roles of Adjuvants in Developing Highly Potent 37 COVID-19 Vaccines. Viruses 2022; 14 [PMID: 35215980 DOI: 10.3390/v14020387]
- 38 Cerpa-Cruz S, Paredes-Casillas P, Landeros Navarro E, Bernard-Medina AG, Martínez-Bonilla G, Gutiérrez-Ureña S. Adverse events following immunization with vaccines containing adjuvants. Immunol Res 2013; 56: 299-303 [PMID: 23576057 DOI: 10.1007/s12026-013-8400-4]
- Truong B, Allegri G, Liu XB, Burke KE, Zhu X, Cederbaum SD, Häberle J, Martini PGV, Lipshutz GS. Lipid nanoparticle-targeted mRNA therapy as a treatment for the inherited metabolic liver disorder arginase deficiency. Proc Natl Acad Sci USA 2019; 116: 21150-21159 [PMID: 31501335 DOI: 10.1073/pnas.1906182116]
- 40 Moghimi SM. Allergic Reactions and Anaphylaxis to LNP-Based COVID-19 Vaccines. Mol Ther 2021; 29: 898-900 [PMID: 33571463 DOI: 10.1016/j.ymthe.2021.01.030]
- 41 Igyártó BZ, Jacobsen S, Ndeupen S. Future considerations for the mRNA-lipid nanoparticle vaccine platform. Curr Opin Virol 2021; 48: 65-72 [PMID: 33906124 DOI: 10.1016/j.coviro.2021.03.008]
- Pardi N, Hogan MJ, Naradikian MS, Parkhouse K, Cain DW, Jones L, Moody MA, Verkerke HP, Myles A, Willis E, 42



LaBranche CC, Montefiori DC, Lobby JL, Saunders KO, Liao HX, Korber BT, Sutherland LL, Scearce RM, Hraber PT, Tombácz I, Muramatsu H, Ni H, Balikov DA, Li C, Mui BL, Tam YK, Krammer F, Karikó K, Polacino P, Eisenlohr LC, Madden TD, Hope MJ, Lewis MG, Lee KK, Hu SL, Hensley SE, Cancro MP, Haynes BF, Weissman D. Nucleosidemodified mRNA vaccines induce potent T follicular helper and germinal center B cell responses. J Exp Med 2018; 215: 1571-1588 [PMID: 29739835 DOI: 10.1084/jem.20171450]

- Ndeupen S, Qin Z, Jacobsen S, Estanbouli H, Bouteau A, Igyártó BZ. The mRNA-LNP platform's lipid nanoparticle 43 component used in preclinical vaccine studies is highly inflammatory. bioRxiv 2021 [PMID: 33688649 DOI: 10.1101/2021.03.04.430128]
- 44 Mauro AG, Bonaventura A, Vecchié A, Mezzaroma E, Carbone S, Narayan P, Potere N, Cannatà A, Paolini JF, Bussani R, Montecucco F, Sinagra G, Van Tassel BW, Abbate A, Toldo S. The Role of NLRP3 Inflammasome in Pericarditis: Potential for Therapeutic Approaches. JACC Basic Transl Sci 2021; 6: 137-150 [PMID: 33665514 DOI: 10.1016/j.jacbts.2020.11.016]
- Guo C, Fu R, Wang S, Huang Y, Li X, Zhou M, Zhao J, Yang N. NLRP3 inflammasome activation contributes to the 45 pathogenesis of rheumatoid arthritis. Clin Exp Immunol 2018; 194: 231-243 [PMID: 30277570 DOI: 10.1111/cei.13167]
- Luan J, Zhang X, Wang S, Li Y, Fan J, Chen W, Zai W, Wang Y, Chen M, Meng G, Ju D. NOD-Like Receptor Protein 3 46 Inflammasome-Dependent IL-1β Accelerated ConA-Induced Hepatitis. Front Immunol 2018; 9: 758 [PMID: 29692782 DOI: 10.3389/fimmu.2018.00758]
- 47 Oberhardt V, Luxenburger H, Kemming J, Schulien I, Ciminski K, Giese S, Csernalabics B, Lang-Meli J, Janowska I, Staniek J, Wild K, Basho K, Marinescu MS, Fuchs J, Topfstedt F, Janda A, Sogukpinar O, Hilger H, Stete K, Emmerich F, Bengsch B, Waller CF, Rieg S, Sagar, Boettler T, Zoldan K, Kochs G, Schwemmle M, Rizzi M, Thimme R, Neumann-Haefelin C, Hofmann M. Rapid and stable mobilization of CD8(+) T cells by SARS-CoV-2 mRNA vaccine. Nature 2021; 597: 268-273 [PMID: 34320609 DOI: 10.1038/s41586-021-03841-4]
- Boettler T, Csernalabics B, Salié H, Luxenburger H, Wischer L, Salimi Alizei E, Zoldan K, Krimmel L, Bronsert P, Schwabenland M, Prinz M, Mogler C, Neumann-Haefelin C, Thimme R, Hofmann M, Bengsch B. SARS-CoV-2 vaccination can elicit a CD8 T-cell dominant hepatitis. J Hepatol 2022; 77: 653-659 [PMID: 35461912 DOI: 10.1016/j.jhep.2022.03.040]
- 49 Bowen DG, Warren A, Davis T, Hoffmann MW, McCaughan GW, Fazekas de St Groth B, Bertolino P. Cytokinedependent bystander hepatitis due to intrahepatic murine CD8 T-cell activation by bone marrow-derived cells. Gastroenterology 2002; 123: 1252-1264 [PMID: 12360486 DOI: 10.1053/gast.2002.36058]
- 50 Belz GT, Altman JD, Doherty PC. Characteristics of virus-specific CD8(+) T cells in the liver during the control and resolution phases of influenza pneumonia. Proc Natl Acad Sci USA 1998; 95: 13812-13817 [PMID: 9811883 DOI: 10.1073/pnas.95.23.13812
- Vojdani A, Kharrazian D. Potential antigenic cross-reactivity between SARS-CoV-2 and human tissue with a possible link 51 to an increase in autoimmune diseases. Clin Immunol 2020; 217: 108480 [PMID: 32461193 DOI: 10.1016/j.clim.2020.108480
- Bernal W, Ma Y, Smith HM, Portmann B, Wendon J, Vergani D. The significance of autoantibodies and immunoglobulins 52 in acute liver failure: a cohort study. J Hepatol 2007; 47: 664-670 [PMID: 17602781 DOI: 10.1016/j.jhep.2007.05.011]
- Febres-Aldana CA, Alghamdi S, Krishnamurthy K, Poppiti RJ. Liver Fibrosis Helps to Distinguish Autoimmune Hepatitis from DILI with Autoimmune Features: A Review of Twenty Cases. J Clin Transl Hepatol 2019; 7: 21-26 [PMID: 30944815 DOI: 10.14218/JCTH.2018.00053]
- Suzuki A, Brunt EM, Kleiner DE, Miquel R, Smyrk TC, Andrade RJ, Lucena MI, Castiella A, Lindor K, Björnsson E. The 54 use of liver biopsy evaluation in discrimination of idiopathic autoimmune hepatitis versus drug-induced liver injury. Hepatology 2011; 54: 931-939 [PMID: 21674554 DOI: 10.1002/hep.24481]
- Weiler-Normann C, Schramm C. Drug induced liver injury and its relationship to autoimmune hepatitis. J Hepatol 2011; 55 55: 747-749 [PMID: 21396413 DOI: 10.1016/j.jhep.2011.02.024]
- 56 Hofer H, Oesterreicher C, Wrba F, Ferenci P, Penner E. Centrilobular necrosis in autoimmune hepatitis: a histological feature associated with acute clinical presentation. J Clin Pathol 2006; 59: 246-249 [PMID: 16505273 DOI: 10.1136/jcp.2005.029348]
- 57 Manns MP, Czaja AJ, Gorham JD, Krawitt EL, Mieli-Vergani G, Vergani D, Vierling JM; American Association for the Study of Liver Diseases. Diagnosis and management of autoimmune hepatitis. Hepatology 2010; 51: 2193-2213 [PMID: 20513004 DOI: 10.1002/hep.23584]
- Björnsson ES, Bergmann OM, Björnsson HK, Kvaran RB, Olafsson S. Incidence, presentation, and outcomes in patients 58 with drug-induced liver injury in the general population of Iceland. Gastroenterology 2013; 144: 1419-1425, 1425.el [PMID: 23419359 DOI: 10.1053/j.gastro.2013.02.006]
- Wong CKH, Mak LY, Au ICH, Lai FTT, Li X, Wan EYF, Chui CSL, Chan EWY, Cheng WY, Cheng FWT, Yuen MF, 59 Wong ICK. Risk of acute liver injury following the mRNA (BNT162b2) and inactivated (CoronaVac) COVID-19 vaccines. J Hepatol 2022; 77: 1339-1348 [PMID: 35817224 DOI: 10.1016/j.jhep.2022.06.032]



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MINIREVIEWS

Effect of SARS-CoV-2 infection on the liver

Adekunle Sanyaolu, Aleksandra Marinkovic, Abu Fahad Abbasi, Stephanie Prakash, Risha Patidar, Priyank Desai, Martina Williams, Abdul Jan, Kareem Hamdy, Rachael Solomon, Vyshnavy Balendra, Maaz Ansari, Omar Shazley, Nasar Khan, Rochelle Annan, Yashika Dixon, Chuku Okorie, Afolabi Antonio

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Adekunle Sanyaolu, Department of Public Health, Federal Ministry of Health, Abuja, Nigeria, Abuja 0000, FCT, Nigeria

Aleksandra Marinkovic, Stephanie Prakash, Risha Patidar, Martina Williams, Kareem Hamdy, Vyshnavy Balendra, Maaz Ansari, Department of Basic Medical Science, Saint James School of Medicine, The Quarter 2640 0000, Anguilla

Abu Fahad Abbasi, Department of Internal Medicine, Loyola University Medical Center, Maywood, Illinois, IL 60153, United States

Priyank Desai, Department of Basic Medical Science, American University of Saint Vincent School of Medicine, Saint Vincent and the Grenadines 0000, Saint Vincent and the Grenadines

Abdul Jan, Nasar Khan, Yashika Dixon, Department of Basic Medical Science, Windsor University School of Medicine, Cayon 0000, Saint Kitts and Nevis

Rachael Solomon, Department of Basic Medical Science, Caribbean Medical University School of Medicine, Willemstad 0000, Curaçao, Netherlands Antilles

Omar Shazley, Basic Medical Science, Saint James School of Medicine, Saint Vincent and the Grenadines 0000, Saint Vincent and the Grenadines

Rochelle Annan, University of Health Sciences Antigua School of Medicine, Piccadilly, St. John's Antigua

Chuku Okorie, Department of Science, Union County College, Plainfield, New Jersey, NJ 07016, United States

Afolabi Antonio, Department of Internal Medicine, Lloydminster Regional Hospital, Lloydminster S9V 1Y5, Saskatchewan, Canada

Corresponding author: Adekunle Sanyaolu, PhD, Academic Research, Director, Department of Public Health, Federal Ministry of Health, Abuja, Nigeria, New Federal Secretariat Complex, Phase III, Ahmadu Bello Way, Central Business District, Abuja 0000, FCT, Nigeria. sanyakunle@hotmail.com

Abstract

There have been numerous concerns about the disease and how it affects the human body since the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) pandemic began in December 2019. The impact of SARS-CoV-2 on the



liver is being carefully investigated due to an increase in individuals with hepatitis and other liver illnesses, such as alcoholic liver disease. Additionally, the liver is involved in the metabolism of numerous drugs used to treat comorbidities and coronavirus disease 2019 (COVID-19). Determining how SARS-CoV-2 affects the liver and what factors place individuals with COVID-19 at a higher risk of developing liver problems are the two main objectives of this study. This evaluation of the literature included research from three major scientific databases. To provide an update on the current impact of COVID-19 on the liver, data was collected and relevant information was incorporated into the review. With more knowledge about the effect of the disease on the liver, better management and therapeutics can be developed, and education can ultimately save lives and reduce the long-term impact of the pandemic on our population.

Key Words: Coronavirus; COVID-19; SARS-CoV-2; Liver; Hepatic complications

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Core Tip: We investigated the impact of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) on the liver due to an increase in individuals with hepatitis and other liver illnesses, such as alcoholic liver disease. Additionally, the liver is involved in the metabolism of numerous drugs used to treat comorbidities and coronavirus disease 2019 (COVID-19). Determining how SARS-CoV-2 affects the liver and what factors place individuals with COVID-19 at higher risk of developing hepatic issues are the two main objectives of this study.

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INTRODUCTION

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is a highly infectious pathogenic coronavirus that appeared in late 2019 causing a pandemic of acute respiratory disease, which is known today as coronavirus disease 2019 (COVID-19)[1]. At a fast rate, the virus spread worldwide, replicated, and mutated into multiple major variants, posing a threat to global public health. SARS-CoV-2 is part of the order Nidovirales, family Coronaviridae, subfamily Orthocoronavirinae, Betacoronavirus genus, and Sarbecovirus subgenus[1,2]. It is a single-stranded, positive-sense, enveloped ribonucleic acid (RNA) virus that is 79.6% identical to SARS-CoV-2 and 96.2% like a bat-derived coronavirus strain[2]. The host receptor for SARS-CoV-2 cell entry is identical to SARS-CoV-2, the angiotensin-converting enzyme 2 (ACE-2)[3]. SARS-CoV-2 binds to ACE-2 with a higher affinity to the receptor-binding domain (RBD) of its spike protein[3]. Therefore, SARS-CoV-2 is more infectious. Since the first reports, which were discovered in Wuhan, China's Hubei Province, at the end of 2019, cases have been documented on every continent^[3]. Globally, more than 500 million confirmed cases of COVID-19 from exposure to SARS-CoV-2 have been reported[3]. SARS-CoV-2 tends to replicate in the upper and lower respiratory tract and is transmitted by droplets and aerosols from asymptomatic and symptomatic infected subjects^[4]. Most infections occur between 2-14 d (about 2 wk) with an incubation period of 5-7 d[4]. These infections tend to be uncomplicated. A small percentage of patients are hospitalized due to severe inflammation and pneumonia. Complications tend to be respiratory and multiorgan failure^[4]. Risk factors for complicated diseases are older age, diabetes, hypertension, chronic cardiovascular disease, chronic pulmonary disease, and immunodeficiency^[4]. The distribution of COVID-19 cases across most countries is highest in the age group of 20-59 years old[4]. Major reductions in social interactions have been implemented in many countries with SARS-CoV-2 outbreaks, leading to rapid reductions. An estimate of the infection fatality rate that is currently reported is 0.5%-1.0% [4]. Despite a rapid worldwide spread, attack rates have been lowered in most regions, demonstrating the efficacy of control measures[4].

Based on initial COVID-19 data, both healthy individuals and those with pre-existing liver disease infected with the SARS-CoV-2 virus exhibit abnormal liver function tests (LFTs), implying that the virus may play a direct role in liver damage^[5]. The incidence of liver injury in patients with COVID-19 has been estimated to range from 14.8% to 53.0%[6]. A clinical study showed that patients with stable liver cirrhosis who contracted the SARS-CoV-2 virus can experience rapid deterioration as evidenced by an



increase in the Child-Pugh score[6]. The incidence of liver injury in cases of death from COVID-19 is 58.0%[6]. Liver injury following the contraction of the SARS-CoV-2 virus is characterized by hypoalbuminemia, hyperbilirubinemia, and an increase in alanine transaminase (ALT) and aspartate transaminase (AST)[5,6]. There may also be an increase in gamma-glutamyl transferase (GGT) and alkaline phosphatase (ALP), indicating injury to liver bile duct cells[6]. The degree of liver injury has a positive correlation with the severity of the infection. Mortality is statistically correlated with elevated AST and low albumin levels of 26.3-30.9 g/L[6]. The mechanism by which SARS-CoV-2 damages hepatocytes is still unclear; however, pathogenic mechanisms may include direct damage, immune-mediated, ischemia and hypoxia, thrombosis, and drug-induced[5,6]. This article aims to investigate the effects of SARS-CoV-2 on the liver and the risk factors for liver problems in coronavirus-infected patients.

METHODOLOGY

PubMed, Google Scholar, and Med Line Plus were used to conduct an electronic literature review. For the data compiled, the search was limited to peer-reviewed articles published between January 1, 2015, and July 1, 2022. The articles were chosen based on keywords such as coronavirus, COVID-19, SARS-CoV-2, and the effects of the virus on the liver. The articles were then examined and included depending on the topic's applicability.

REPORTED SYMPTOMS OF SARS-CoV-2 LIVER INJURY

Elevated LFTs in COVID-19

Altered LFTs have been observed in almost half of the hospitalized patients with COVID-19 infection [7]. In particular, elevated levels of liver enzymes glutamic-pyruvic transaminase (ALT), glutamic-oxaloacetic transaminase (AST), glutamyl transpeptidase (GGT), alkaline phosphatase (ALP), and bilirubin have been seen to manifest as liver injury in such patients[6,8]. Previous studies have shown that the incidence of COVID-19 liver damage with elevated ALT ranges between 9.6% and 37.6%, elevated AST between 14.8 and 36.0%, and the proportion of abnormal GGT between 13.0%-24.4%[9]. These abnormal tests are the result of increased AST and ALT, whereas AST was more common than ALT. In addition, 10.5% to 69.0% of hospitalized COVID-19 patients showed abnormal LFTs. Hypoalbuminemia has also been reported as a consequence of COVID-19-related liver injury and was observed more significantly in men with COVID-19 compared to women[10]. When comparing and analyzing those with severe and non-severe COVID-19 cases, liver function abnormalities like hypoalbuminemia, GGT, aminotransferase, and bilirubin elevations were more frequent in those with severe disease as opposed to mild/moderate forms of the infection[11]. The liver injury caused by COVID-19 was related to the degree of severity of the infection and manifested as different degrees of liver function abnormalities[12].

Pathological changes in liver biopsies

Histopathological findings from liver biopsies of COVID-19 patients showed moderate microvascular steatosis with lobular and portal vein involvement^[13]. Hepatocyte degeneration, with neutrophil infiltration of the hepatic lobes and sinusoidal enlargement of the central lobule, was observed. Congestion of hepatic sinuses with micro thrombosis and sinusoidal expansion, lymphocytic infiltration of the lobes, and hepatic necrosis in the periportal and centrilobular segments was also identified in patients [14]. Furthermore, pathological findings showed hepatocellular necrosis, cellular infiltration, an increase in the number of mitotic hepatocytes, and fatty degeneration[15]. In addition, COVID-19 Liver injury showed an elevation of eosinophilic bodies along with dilated hepatocytes[6]. Sinusothelial micro thrombosis disease was evident in approximately 20.0% of cases with focal endothelial damage[16]. Acinar atrophy was depicted in autopsy specimens in the late course of the infection^[17]. Increased liver stiffness is also correlated with increased levels of biomarkers of liver injury, such as ALT and GGT, suggesting underlying hepatocellular and cholangiocellular damage at the biochemical level[18]. Changes in liver elasticity, viscosity, and steatosis levels were also observed in liver tissues in COVID-19 patients, with increased fibrosis compared to the control group (P < 0.001)[18]. In some studies, the liver appeared pale and yellowish on sectioning, with a nutmeg appearance [19]. The infection has been studied to cause cholangiocellular injury and cholestasis and consequently bile duct proliferation, with bile plug formation. In general, analyses have revealed that as the severity of COVID-19 in a patient increase, the levels of AST, ALT, total bilirubin, GGT, and ALP increase, resulting in a greater degree of liver injury, as observed in hepatocytes[20].

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SARS-CoV-2 RELATED LIVER INJURY: PATHOGENIC MECHANISMS

Liver injury in patients infected with SARS-CoV-2 occurs via several mechanisms^[21]. Several studies indicate that liver injury in patients with SARS has manifested through the elevation of liver enzymes, mainly ALT and/or AST in the early stage of the infection^[21]. The incidence of liver injury in SARS patients ranges from 14.8% to 53.0% [21]. One hypothesis for the cause of liver injury is a direct invasion of the hepatic parenchyma by SARS-CoV-2[21]. Autopsy of patients with SARS found a large number of virus particles in the parenchyma and vascular endothelium of the liver[21]. The main receptor used by SARS to enter cells is ACE-2 which is abundantly present in cholangiocytes, endothelial cells, and the progenitor cells of the liver^[22]. This results in acute liver and hepatitis biopsies in postmortem patients showing a significant increase in macrovesicular steatosis with eosinophilic bodies and high levels of mitotic cells suggesting hepatocyte apoptosis[21]. The results suggest that SARS infection causes direct injury to the hepatic parenchyma and concomitantly compromises the regenerative capability of the liver[22]. Hepatic injury is further exacerbated by the body's immune response to severe COVID-19 infection[23]. SARS activates both the innate and acquired immune system resulting in the release of high levels of several inflammatory cytokines by immune cells[23]. The resulting cytokine storm in severe SARS infections is the cause of death in 28.0% of fatal cases of COVID[24]. Multiorgan failure is a sequela of the cytokine storm, and the liver is no exception. Critically ill patients exhibited increased levels of interferon-lambda (IFN- λ), transforming growth factor-alpha (TGF- α), thymic stromal lymphopoietin, interleukin-16 (IL-16), IL-23, IL-33, and markers linked to coagulopathy, such as thrombopoietin. Patients with severe COVID are commonly anoxic due to respiratory failure. This requires patients to be mechanically ventilated and/or on vasopressor support. Lower cardiac output has a detrimental impact on the hemodynamics of the liver[23]. The resultant reduced hepatic blood flow can lead to anoxic hypoxic hepatitis and/or cholestasis^[24]. Another complication of hepatic injury due to the high levels of inflammatory cytokines released by the body includes thrombosis and vascular congestion of the liver, which have been observed in autopsy samples of patients with severe COVID-19. Patients with severe COVID-19 were found to have elevated levels of total bilirubin and ALT, as well as elevated levels of inflammatory biomarkers such as IL-6, IL-10, C-reactive protein (CRP), and Ddimer. One of the mechanisms contributing to the damage observed in the liver of these patients is due to the SARS-CoV-2 infected cells which upregulate and produce large amounts of cytokines to help combat the virus, resulting in collateral damage to both infected and uninfected cells. This hyperstimulated systemic inflammatory response induces macro- and micro-circulatory dysfunction, leading to global hypo-perfusion resulting in hypoxia, hypo-tension, and a hypercoagulable state. Therefore, microvascular thrombosis should be considered an important cause of liver injury and dysfunction in patients with COVID-19[25]. Hepatic injury during COVID-19 infection can be exacerbated by medications leading to elevated levels of ALT and AST. Remdesivir has shown in vitro antiviral activity against SARS-CoV-2 and a shorter recovery time in clinical trials, but elevated hepatic enzymes have also been reported as a major adverse drug reaction. Although some studies attribute this abnormal increase to viral infection rather than the side effect of the drug, others proposed that, whether or not it was affected by SARS-CoV-2, remdesivir increased the risk of hepatotoxicity. Other commonly used drugs to treat COVID-19 such as lopinavir/ritonavir have also resulted in hepatic injury. ACE medications and angiotensin II receptor blockers, which take a more focused approach, have also been reported to raise liver enzymes in COVID-19 patients. In autopsies of COVID-19 deaths, moderate microvesicular steatosis and mild lobular and portal activity were observed and probably associated with drug-induced liver injury. Other medications used for patients with COVID-19 that can trigger liver injury include antibiotics such as macrolides and quinolones, antivirals such as ribavirin, and even steroids^[26].

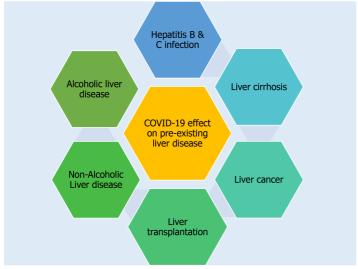
SARS-CoV-2 INFECTION AND PRE-EXISTING LIVER DISEASES

The effect of COVID-19 on pre-existing liver diseases discussed below are hepatitis B and C virus infection, liver cirrhosis, liver cancer, liver transplant, non-alcoholic fatty liver disease (NAFLD), and alcoholic liver disease (Figure 1).

COVID-19 effect on hepatitis B virus and hepatitis C virus infection

Hepatitis C virus (HCV) and hepatitis B virus (HBV) were reported as the leading causes of liver diseases, but no specific data on the prevalence of these infections were provided. Many studies involving patients with COVID-19 showed a relatively low prevalence of chronic liver disease (CLD) at baseline, equal to 3.0%. Similarly, a prevalence rate of 3.0% CLD is associated with documented underlying chronic HBV or HCV infections in specific populations^[27]. In one study of patients admitted to hospitals for COVID-19 in the northeastern United States, HCV infection was observed in P < 0.1% of patients, but information on HCV RNA levels was insufficient. In the same study, 23 cases (3.8%) presented positive HCV serology, of whom six patients (0.99%) had detectable viral HCV load at the time of hospital admission for the diagnosis of COVID-19[27]. In contrast, 2.0% of all patients





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Figure 1 Effect of coronavirus disease 2019 and preexisting liver disease.

showed hepatitis B surface antigen (HBsAg) positive chronic infection[27]. The study showed patients with recorded chronic HBV or HCV infection did not experience a more severe clinical course of COVID-19 compared to patients with HBsAg negative or undetectable HCV RNA, which measured the delay in SARS-CoV-2 clearance in HBV patients. Similarly, median viral clearance was not affected by preexisting HBV or HCV infection[27]. Lastly, chronic HBV or HCV infection (in the absence of cirrhosis) did not affect the prognosis of COVID-19 in the United States population.

COVID-19 effect on liver cirrhosis

Liver cirrhosis (Figure 2) increases the mortality of SARS-CoV-2 viral infection[28]. The pathophysiological mechanism for the SARS-CoV-2 virus begins with the spike glycoprotein (S) to allow viral entry into the target cell. The virus replicates to infect other surrounding cells through the ACE-2 receptor in cholangiocytes and hepatocytes to cause biliary and liver symptoms. Elevated liver enzymes are multifactorial and strongly associated with liver injury. A prevalent hepatic phenomenon associated with SARS-CoV-2 infection presents with elevated ALT and AST levels, with abnormal ALP and bilirubin readings^[29]. Elevated serum levels of GGT, a marker of hepatobiliary cell injury, are found in a quarter of patients hospitalized with COVID-19[28]. Higher levels of a hepatocellular enzyme associated with severe cases of COVID-19 directly affect mechanisms that include increased cytokine release in the viral presence or microthrombotic ischemic liver injury[29]. Although respiratory symptoms are the most reported among COVID-19 patients, these pulmonary manifestations are vulnerable to decompensated liver cirrhosis but are currently understudied. A cohort study with 250 patients with prior CLDs reported high mortality in patients with cirrhosis (RR: 4.6, 95.0% CI: 2.6-8.3) [28]. In an international study, a positive correlation was shown that patients with cirrhosis are predisposed to significant toxic liver injury due to SARS-CoV-2 infection, as acute-on-chronic liver failure had occurred in 20.0% of patients who experienced severe cirrhosis with COVID-19[28]. The viral ability of SARS-CoV-2 to bind to ACE-2 receptors on epithelial cells of the bile duct demonstrates its ability to affect liver regeneration capabilities and immune response^[28]. By affecting the innate immunity of the reticuloendothelial system, the immunosuppressed state causes a cytokines-mediated reaction, resulting in liver decompression[28].

COVID-19 effect on liver cancer

Pre-existing liver diseases are considered risk factors for poorer prognosis in COVID-19 as various pathophysiological processes result in liver damage due to SARS-CoV-2 infection. Biochemical presentations of liver injury include elevated levels of ALT/AST, ALP, GGT, or total bilirubin above the normal range. The decrease in lymphocyte count and the increase in neutrophil counts demonstrate the role of innate immunity in COVID-19-associated hepatic injury. Postmortem studies in liver histology in COVID-19 patients show moderate microvascular and macrovascular steatosis with mild inflammation of the lobular portal. This highlights the pathological changes observed in hepatocellular carcinoma (HCC). Elevated levels of eosinophils are observed in autopsy studies in centrilobular steatosis, in addition to the increased number of mitotic cells[30]. Patients with HCC are closely monitored, as increased inflammation due to COVID-19 may predispose patients to post-hepatectomy liver failure. Furthermore, COVID-19 can potentially exacerbate CLD and alter treatments for cancer patients with a higher risk of infection and poor outcomes[30]. The management and monitoring of patients with HCC





Figure 2 Healthy and cirrhosis liver.

are performed by imaging (magnetic resonance imaging, ultrasound) and measuring alpha-fetoprotein levels^[29]. The practice guidelines recommend the establishment of surveillance intervals to reduce the radiologic capacity of at-risk patients, with a 98.0% estimate that at-risk patients would not develop HCC during each surveillance interval. Locoregional and systematic therapies are recommended for advanced HCC treatment. However, oral therapy with tyrosine kinase inhibitors (*i.e.*, sorafenib and lenvatinib) and immunotherapy effectively serve as first-line therapies to reduce exposure^[29].

COVID-19 effect on liver transplantation

With the severity of SARS-CoV-2 infection dependent on comorbidities (i.e., cardiovascular disease and diabetes mellitus), underlying liver diseases do not necessarily influence the outcome of COVID-19 infection. Solid-organ transplants, including liver transplant (LT) recipients, are increasingly susceptible to severe infections due to chronic immunosuppression, thereby increasing the risk for severe COVID-19 infection. A meta-analysis that included 17 articles and the outcomes of 1481 COVID-19 LT patients was compared with 239704 non-LT patients infected with COVID-19. From 17 articles, a cumulative incidence of mortality of 17.4% (95.0% CI, 15.4-19.6) was found among LT recipients with COVID-19 with causes of death reported as 62.54% by COVID-19-related complications (95.0%CI, 56.24-68.55), 29.88% by pulmonary failure (95.0%CI, 24.28-36), and 1.6% liver-related (95.0%CI, 0.1-2.84). Mortality was proportionate between LT and non-LT patients [OR, 0.8 (0.6-1.08); P = 0.14][31]. Twelve studies in the same meta-analysis reported that 23.0% of LT patients who had developed a severe COVID-19 infection were positive with symptoms that included fever (49.7%), cough (43.76%), dyspnea (29.27%), and symptoms gastrointestinal (27.26%)[31]. Eight studies from the same meta-analysis reported modification change immunosuppression in 55.9% of LT recipients infected with COVID-19[31]. Comorbidities such as hypertension, diabetes, and obesity were common in infected patients where 72.0% of the patients were hospitalized, and 16.0% required care in the intensive care unit (ICU)[31]. Although hospitalization of LT recipients far exceeded non-LT patients [OR, 1.99 (1.41-2.8), P < 0.001], the ICU care requirement was comparable between groups, as the cumulative incidence of graft dysfunction was 2.3% (1.3-4.1)[31].

COVID-19 effect on non-alcoholic fatty liver disease

Many observational studies have shown that patients with comorbidities such as cardiovascular disease, arterial hypertension, diabetes mellitus, CLD, or cancer are susceptible to more severe episodes of COVID-19, as seen in NAFLD and other less common disorders[32]. Recently, several meta-analyses have shown that obesity and diabetes (both strongly associated with NAFLD) are significantly associated with the progression of more severe disease and increased mortality in patients with COVID-19[32]. They have reported a six-fold increased risk of severe COVID-19 in the presence of obesity in NAFLD. In addition, a meta-analysis showed that obesity could exacerbate COVID-19 infection. Patients with severe COVID-19 disease had higher body mass indices, and obesity was associated with the development of the disease, the need for care, and admission to an ICU[32]. The risk of severe COVID-19 in obese patients was more significant than in younger patients. This suggests that NAFLD patients are at increased risk of liver damage, although liver enzyme levels at admission or during hospitalization were generally not significantly elevated. Furthermore, NAFLD was not associated with adverse clinical outcomes in younger patients with COVID-19. Another study found that NAFLD was more common in patients with severe COVID-19 than in stable patients. However, the mean age and the number of comorbidities were also significantly higher in patients with severe COVID-19 infections[32]. Patients with NAFLD had a substantially higher risk of disease progression, more likely changes in liver enzymes, and longer viral shedding times than non-NAFLD patients. Non-NAFLD patients showed that moderate to high Fibrous-4 and NAFLD fibrosis scores were strongly and independently correlated with the severe progression of COVID-19 disease[32].

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COVID-19 effect on alcoholic liver disease

The ACE-2 receptor is exceedingly expressed in alveolar type II cells and liver and bile duct cells, making it significantly feasible for SARS-CoV-2 to infect cells in those areas. Especially, cholangiocytes have a specific ACE-2 receptor in more concentrations than hepatocytes, making them more susceptible to COVID-19 infection. However, because the liver harbors a widespread quantity of macrophages, generating an ample cytokine-mediated immune reaction, hepatocytes can also be prone to a SARS-CoV-2 infection. Patients with COVID-19 patients with liver cirrhosis have always shown elevated levels of ALT, AST, D-dimer, CRP, IL-6, and ferritin. Although the current literature is limited, research has proven that people with CLDs could have increased models for end-stage liver disease and undergo extended liver and pulmonary complications while infected with COVID-19. Specifically, the mortality rate in patients with preexisting liver disease is 1.8%. Lastly, the severity of liver harm due to COVID-19 infection tends to be substantially worse and more widespread in people with pre-existing alcoholic liver cirrhosis than in those without[33].

ANTI-SARS-CoV-2 TREATMENTS EFFECTS ON THE LIVER

Currently, various treatments for COVID-19 (SAR-CoV-2) are being investigated, some of which may be associated with hepatotoxicity[34]. Remdesivir (RDV), an antiviral medication, was initially developed and tested for the treatment of hepatitis C and later the Ebola and Marburg viruses. Amid the COVID-19 pandemic, RDV was approved for emergency use to treat COVID-19 in many countries[35]. In patients diagnosed with COVID-19, in vitro and in vivo studies indicate that RDV has an antiviral effect on SARS-CoV-2[36]. Various medication-related adverse events include but are not limited to reasonable degrees of nausea and vomiting, headache, fatigue, renal dysfunction, and rash[37]. The risk of an adverse event involving the liver exists as one of the clearest potential risks from RDV[38]. RDV therapy is administered intravenously for 3 to 10 d and is often accompanied by reversible mild to moderate elevations in serum AST levels, but has been rarely associated with clinically apparent liver injury. Effects on the liver range from asymptomatic to mild-with elevations in serum ALT and AST upon introduction of RDV therapy in patients with COVID-19. The systemic effects of COVID-19 likely overshadow the outcomes of hepatic involvement[37]. However, there is uncertainty regarding if the effect on liver enzymes is due to remdesivir, COVID-19 solely, or both [39,40]. It is recommended that patients remain under the supervision of health professionals to monitor liver health before and during remdesivir infusions[37]. To fully assess the risk of remdesivir-associated liver damage, more studies are necessary for this area[40]. Lopinavir (LPV) is an antiretroviral protease inhibitor, used together with ritonavir (booster) in the prevention and treatment of human immunodeficiency virus (HIV) infection[41]. Lopinavir/ritonavir (LPV/r) was developed to inhibit HIV protease, the primary distinction for the SARS-CoV-2 counterpart (3CLpro) lies within the varying spatial structure of the HIV aspartic protease as compared with 3CLpro cysteine protease[41]. The LPV/r combination improves LPV pharmacokinetics by decreasing liver metabolism by inhibiting the cytochrome (CYP) P450 3A4 enzyme[41].

A randomized controlled study in adult patients hospitalized with COVID-19 shows that of those adults treated with LPV/r, only one individual in the LPV/r group presented elevated ALT, more than 2.5 times above the normal limit[41,42]. When comparing patients treated with LPV/r to patients in the control group, there was no evidence of liver dysfunction noted in controls. It is important to acknowledge that the patient presenting with an elevated ALT had a pre-existing chronic liver condition, possibly contributing to the liver disturbance[42]. Another research study suggests that no observable side effects were found in the LPV/r group, except for transient elevation of ALT elevation (<125 U/L) in three patients[43]. Given that none of the patients progressed to a severe clinical status at the end of the follow-up period, it is believed that LPV/r treatment rarely causes harm in patients recovering from COVID-19[42]. LPV/r is considered an independent factor for liver injury[44]. Interferons (IFNs) are natural antiviral immune modulators that help the body's immune system defend against infection and disease, including viruses and cancer[45]. Studies have shown that during SARS and middle east respiratory syndrome, Type I IFNs are markedly suppressed and the administration of exogenous Type I IFNs has been shown to reduce the severity of the symptoms of these diseases[46]. To assess the effectiveness and safety of interferon β -1a (IFN β -1a) in patients with severe COVID-19, a randomized clinical trial was conducted [47]. Comparisons were made between patients receiving IFN and those receiving controlled standard therapy. Hepatic complication rates were measured between patients in the treatment group and those patients receiving standard care while the preexisting liver disease was considered^[44]. The frequency of hepatic failure did not differ between the IFN and control groups (11.90% vs 23.07%), suggesting that IFN- β -1a may not be a major factor in the liver damage seen by COVID-19 patients[47]. Studies show that IFNs used to treat patients with COVID-19 are unlikely to be associated with liver disease. IFNs may lead to hepatic toxicity when combined with other drugs[44]. To authenticate these results, additional studies are necessary [44]. Baricitinib is a JAK-STAT inhibitor used to treat individuals with rheumatoid arthritis who cannot tolerate more than one tumor necrosis factor (TNF) antagonist[6]. By decreasing adaptor-associated kinase 1 activity, a regulator of clathrin-



mediated endocytosis, baricitinib has been shown to affect the hyperinflammatory state that developed during SARS-CoV-2 infection and may prevent endocytosis and viral infection[6,48]. Furthermore, the oral administration of baricitinib and the excellent pharmacokinetic profile (very short half-life, low plasma protein binding, and minimal interference with CYP enzymes) make it a viable combination therapy with direct-acting antivirals such as LPV/r and RDV[48]. The growing number of reports of infections and thrombosis following the use of JAK inhibitors for the treatment of COVID-19 should be taken seriously as liver damage, cholestasis, and hepatitis unexpectedly manifested in a non-negligible fraction of individuals[6]. Furthermore, these unfavorable hepatic consequences should be evaluated. Tocilizumab is a monoclonal antibody that is used to block the inflammatory protein IL-6. Tocilizumab improves joint pain and swelling from arthritis and reduces other symptoms caused by inflammation. More recently, tocilizumab use has been indicated for the treatment of cytokine release syndrome in patients with COVID-19 infection[49]. Common side effects of tocilizumab include a runny or stuffy nose, sinus pain or sore throat, headache, or dizziness. The most common side effects of tocilizumab include headache and hypertension but, rarely, hepatotoxicity ranging from mild transaminase elevation to severe drug-induced liver injury can occur[50].

SARS-CoV-2 VACCINES EFFECT ON THE LIVER

It is not yet clear whether RNA or DNA-based vaccines have any direct effect on the liver, resulting in hepatotoxicity. Although anti-coronavirus treatments have been found to cause mitochondrial and endoplasmic reticulum dysfunction, the effects of vaccines require further testing[6]. Patients receive what are often considered benign mRNA vaccines for Crigler-Najjar syndrome and rabies, which have some form of hepatotoxicity. While there is no definitive cause, there seems to be a potential link between the two. This is not the case with DNA vaccines, which makes them strikingly different. Immune system stimulation occurs via a completely different mechanism than mRNA vaccines, with IFN-1 secretions triggering the immune response. Unlike mRNA vaccines, DNA vaccines do not require subsequent doses to maintain monoclonal antibody protection, making DNA vaccines not only potentially more efficacious than mRNA vaccines, but requiring lower amounts to achieve a less toxic overall therapeutic effect. More research is needed to fully understand the mechanisms involved and how they affect the liver. Existing studies exclude patients with chronic liver disease as they are contraindicated by mRNA vaccines, making little information available regarding the pathophysiology, and comparing that of otherwise healthy individuals 6. Patients with CLD are at an increased risk of infection, which is expected given the insufficient immune response. Vaccinations are imperative to reduce mortality in patients with CLD[51]. According to the Advisory Committee on Immunization Practices (ACIP), patients with CLD should be vaccinated against SARS-CoV-2 and influenza, pneumococcus, tetanus, diphtheria, pertussis, herpes zoster, hepatitis A, and hepatitis B[52]. Specifically, with the COVID-19 vaccine, ACIP suggests an mRNA vaccine with a booster dose five months after completing the two scheduled doses. For severely immunosuppressed patients, a booster dose is recommended after three months to strengthen the immune response[53]. A double-blind randomized trial studying the administration of a third dose of the mRNA vaccine in transplant recipients showed a strong immune response compared to the placebo group [54]. In a case report of healthy patients with no history of liver disease, patients developed jaundice and elevated liver enzymes after administration of the mRNA vaccine, either after the first or second dose. In these cases, the laboratory values and symptoms resolved without treatment after several weeks. The data suggests that this response is due to a neutrophil-predominant inflammatory response[55]. Comparatively, much data shows contraindications to multiple COVID-19 vaccines, including booster doses, specific to CLD patients. Data are still needed to assess the efficacy and long-term effects of multiple COVID-19 vaccines in patients with CLD.

The severity of the liver disease may be assessed using the Child-Pugh scale. This scale anticipates mortality in CLD and is categorized into three stages: good hepatic function, moderately impaired hepatic function, and advanced hepatic dysfunction[56]. Multiple factors, including the stage of CLD, can be a determinant of the efficacy of a vaccine, and those in later stages are more susceptible to infections and adverse events. This is possibly due to the inefficiency of the body in producing an adequate immune response[51]. However, the World Health Organization (WHO) currently recommends that COVID-19 vaccines for those with deficient immune systems be given additional boosters to help increase a sufficient immune response[57]. It is recommended that vaccinations be administered as early in the disease process as possible to gain the best performance of the vaccine[52]. In a study on the antibody response of the vaccine in CLD patients, subjects were given the recommended series of the mRNA vaccine. The results showed that 24.0% of the subjects had a poor antibody response[58]. Additional research is needed to further assess the success of the vaccine in varying severities of CLD.

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DISCUSSION

Lessons from previous coronavirus outbreaks and other viral epidemics indicate that the combination of systemic and partial inflammatory responses induced by these infections may result in severe respiratory syndromes and related complications (such as abnormal liver function, cardiac insufficiency, and renal failure)[6]. Manifestations of liver damage from SARS-CoV-2 include a decrease in albumin and an increase in ALT, AST, liver enzymes, and bilirubin[9]. Increases in GGT and ALP are also seen in COVID-19 patients, indicating liver damage to bile duct cells^[7] (Figure 3). Since liver biopsies taken from a small number of COVID-19 patients did not reveal viral inclusions, but rather a macro-vesicular steatosis, liver damage may also be the result of bile duct cell damage[59]. The pathogenic alterations frequently take the form of macro-vesicular steatosis and mild lobular and portal inflammation[59,60]. According to earlier research, severe instances of coronavirus infection had a strikingly higher frequency and severity of liver impairment than moderate cases. The mechanism by which coronaviruses harm hepatocytes and influence hepatic function is still unknown, even though multiple clinical studies have shown a high link between coronaviruses and liver damage[6]. Potential mechanisms of liver injury that have been reported include immune-mediated damage because of the severe dysregulated inflammatory response, direct cytotoxicity, systemic hypoxia with hypoxic hepatitis, drug-induced liver injury, reactivation of pre-existing liver disease, mitochondrial dysfunction, SARS-CoV-2-induced hepatic steatosis, microthrombotic disease, ischemic hepatitis, cardiomyopathy with hepatic congestion, and extrahepatic release of transaminases[61]. Ischemic, hypercoagulable, and hyperinflammatory states are independent predictors of death in patients with COVID-19 and not liver injury[62]. Coronavirus infection significantly increases immunological activation. Numerous cytokines and chemokines (IL-6, IL8, IFN, and TNF, among others) are generated by immune cells after coronavirus infection and released into the blood, causing inflammation in different organs or even acute respiratory distress syndrome and multiple organ failure, suggesting that coronavirus-induced systemic inflammatory response syndrome (SIRS) and cytokine storms are important causes of liver damage[6,25]. This shows that immunotherapy is necessary for individuals with coronavirus infection, and as a result, corticosteroids and interferons are frequently utilized due to their ability to reduce inflammation[6]. Hypoxia may result in a long-term increase in reactive oxygen species, which may encourage the release of a variety of inflammatory mediators that harm the liver[6, 25]. As a result, it will be important to keep an eye on patients' hypercoagulable conditions, such as thrombocytopenia and elevated levels of D-dimer and ALP, to prevent thrombosis and additional ischemia and hypoxia[6,23].

Immune compromise is typically caused by hepatitis B and C, liver cirrhosis, liver malignancy, and immunosuppressive medications after liver transplantation[6]. The severity and mortality rate in HBV infection patients are higher than in those with negative HBV due to delayed clearance of SARS-CoV-2 [6]. The Child-Pugh scores of those who have already developed liver cirrhosis are likely to rise due to liver injury caused by COVID-19[56]. Furthermore, COVID-19 complications occur earlier and to a greater extent in patients with systemic immunocompromised status[6]. COVID-19 also has a significant impact on the treatment of liver diseases[11]. The discontinuation of high-dose corticosteroid therapy in hepatitis B and C patients receiving anti-HBO treatment may result in HBV reactivation during SARS-CoV-2 infection[6]. In addition, lopinavir and ritonavir have been shown to increase the risk of developing liver injury in HBV or HCV infection patients[6,11]. Coronavirus infection is currently treated with redelivering, lopinavir/ritonavir, interferon-a, baricitinib, and tocilizumab. The difficulty in developing optimized drugs for coronavirus infection is mainly due to severe side effects. Remdesivir, lopinavir, and ritonavir have all been linked to an increased risk of liver injury, with the severity of the injury being closely related to the dose of these drugs. IFNs have the potential to trigger a non-specific immune response, resulting in hepatocyte damage and autoimmune hepatitis, as well as an increased risk of developing severe complications such as systemic inflammatory reaction syndrome and acute respiratory distress syndrome[6]. Baricitinib, as a JAK inhibitor, can increase the risk of thrombosis and cause liver damage[6,48]. Tocilizumab can also reactivate HBV in SARS-CoV-2 co-infection, causing both viral hepatitis and COVID-19 recovery to be delayed; whereas other studies have shown hepatotoxicity as a potential side effect^[50]. Overall, coronavirus vaccines will be critical in preventing outbreaks, but several factors must be considered to avoid an activated innate inflammatory response, an increase in the incidence of autoimmune diseases, and vaccine-induced liver injury[6].

Furthermore, a study observed 900 patients (32.2% in the 18-39 age group, 39.7% in the 40-69 age group, and 28.1% in the 70+ age group) with SARS-CoV-2[63]. It was seen that those with comorbidities, median D-dimer, and CRP levels all increased with age. AST/ALT and ALP/GGT levels also increased significantly during COVID-19[63]. Patients with elevated hepatocellular transaminases (AST/ALT) and cholestasis parameters (ALP/GGT/bilirubin) were found in 40.3% (n = 262/650) and 45.0% (n = 287/638), respectively[61]. Importantly, patients between the ages of 40 and 69 were more likely to experience COVID-19-associated liver injury (16.0%, P < 0.001), abnormal liver chemistry, and liver-related death (6.5%, P < 0.001)[61]. After the initial SARS-CoV-2 polymerase chain reaction result was positive, elevated AST and bilirubin levels independently predicted mortality in the entire population and patients aged 40 to 69 years[63].

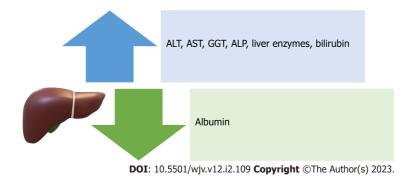


Figure 3 Manifestations of liver damage from severe acute respiratory syndrome coronavirus 2. ALT: Alanine transaminase; AST: Aspartate transaminase; GGT: Gamma-glutamyl transferase; ALP: Alkaline phosphatase.

CONCLUSION

The incidence of liver injury in patients with COVID-19 has been estimated to be as high as 53.0%. Those affected by COVID-19-associated liver injury generally fall between the ages of 40 and 69. The mechanism by which SARS-CoV-2 damages hepatocytes is still unclear. However, the SARS-CoV-2 virus may play a direct role in liver damage because both healthy individuals and those with preexisting liver disease exhibit abnormal LFTs. The liver injury caused by COVID-19 is related to the degree of severity of the infection and manifests itself with different degrees of liver abnormalities. The degree of liver injury manifested by AST, ALT, total bilirubin, GGT, and ALP has been shown to have a positive correlation with the severity of the disease. Interestingly, some studies have even shown that mortality correlates with elevated AST and low albumin levels. Furthermore, SIRS and cytokine storms augment liver injury and dysfunction. Commonly used medications may play a role in liver hepatotoxicity, however further studies are necessary. Pre-existing liver diseases are considered risk factors for worse prognosis in COVID-19, specifically, liver cirrhosis was shown to increase the mortality in these patients. Although vaccines have significantly changed the course of this pandemic, CLD is a contraindication of multiple COVID-19 vaccines. However, the increased severity of liver disease in determining the immune response to the COVID-19 vaccine is still unclear, and more studies are required in this area. As more information about the virus becomes available, it will be critical to comprehend the pandemic's effects on the liver, as well as the possible long-term consequences, especially in the immunocompromised population.

FOOTNOTES

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Country/Territory of origin: Nigeria

ORCID number: Adekunle Sanyaolu 0000-0002-6265-665X; Aleksandra Marinkovic 0000-0002-3672-0777; Abu Fahad Abbasi 0000-0003-2539-2250; Stephanie Prakash 0000-0003-2664-9775; Risha Patidar 0000-0002-5929-9348; Priyank Desai 0000-0002-4061-5144; Martina Williams 0000-0001-5136-4179; Abdul Jan 0000-0002-2768-6298; Kareem Hamdy 0000-0002-3482-3940; Rachael Solomon 0000-0001-8488-3928; Vyshnavy Balendra 0000-0002-9502-0885; Maaz Ansari 0000-0001-7362-9342; Omar Shazley 0000-0003-3456-3111; Nasar Khan 0000-0002-1647-9752; Rochelle Annan 0000-0002-4202-2633; Yashika Dixon 0000-0003-4007-9481; Chuku Okorie 0000-0001-5483-0032; Afolabi Antonio 0000-0001-7696-6782.

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REFERENCES

- 1 Ravi V, Saxena S, Panda PS. Basic virology of SARS-CoV 2. Indian J Med Microbiol 2022; 40: 182-186 [PMID: 35300895 DOI: 10.1016/j.ijmmb.2022.02.005]
- Li Y, Li JZ. SARS-CoV-2 Virology. Infect Dis Clin North Am 2022; 36: 251-265 [PMID: 35636899 DOI: 2 10.1016/j.idc.2022.01.004]
- McIntosh K, Hirsch MS, Bloom A. COVID-19: Epidemiology, virology, and prevention. 16 May 2022. UpToDate. [cited 3 12 June 2022]. In: UpToDate [DOI: 10.1097/01.cot.0000911056.28044.c7]
- Salzberger B, Buder F, Lampl B, Ehrenstein B, Hitzenbichler F, Holzmann T, Schmidt B, Hanses F. Epidemiology of 4 SARS-CoV-2. Infection 2021; 49: 233-239 [PMID: 33034020 DOI: 10.1007/s15010-020-01531-3]
- Martinez MA, Franco S. Impact of COVID-19 in liver disease progression. Hepatol Commun 2021; 5: 1138-1150 [DOI: 5 10.1002/hep4.1745]
- 6 Wang X, Lei J, Li Z, Yan L. Potential effects of coronaviruses on the liver: An update. Front Med 2021; 8: 651658 [PMID: 34646834 DOI: 10.3389/fmed.2021.651658]
- Cai Q, Huang D, Yu H, Zhu Z, Xia Z, Su Y, Li Z, Zhou G, Gou J, Qu J, Sun Y, Liu Y, He Q, Chen J, Liu L, Xu L. 7 COVID-19: Abnormal liver function tests. J Hepatol 2020; 73: 566-574 [PMID: 32298767 DOI: 10.1016/j.jhep.2020.04.006].]
- 8 Clark R, Waters B, Stanfill AG. Elevated liver function tests in COVID-19: Causes, clinical evidence, and potential treatments. Nurse Pract 2021; 46: 21-26 [PMID: 33332825 DOI: 10.1097/01.NPR.0000722316.63824.f9]
- Zhang Y, Zheng L, Liu L, Zhao M, Xiao J, Zhao Q. Liver impairment in COVID-19 patients: A retrospective analysis of 115 cases from a single centre in Wuhan city, China. Liver Int 2020; 40: 2095-2103 [PMID: 32239796 DOI: 10.1111/liv.14455
- 10 Harisha EJ, Gosavi S, Rao AA, Sahana GV, Manjunath S, Meghana TC. Liver: Function and dysfunction in COVID-19. J Family Med Prim Care 2022; 11: 758-761 [PMID: 35360815 DOI: 10.4103/jfmpc.jfmpc_7_21]
- Wong YJ, Tan M, Zheng Q, Li JW, Kumar R, Fock KM, Teo EK, Ang TL. A systematic review and meta-analysis of the 11 COVID-19 associated liver injury. Ann Hepatol 2020; 19: 627-634 [PMID: 32882393 DOI: 10.1016/j.aohep.2020.08.064]
- 12 El Ouali S, Romero-Marrero C, Regueiro M. Hepatic manifestations of COVID-19 Cleve Clin J Med 2020 [PMID: 32855179 DOI: 10.3949/ccjm.87a.ccc061]
- Díaz LA, Idalsoaga F, Cannistra M, Candia R, Cabrera D, Barrera F, Soza A, Graham R, Riquelme A, Arrese M, Leise 13 MD, Arab JP. High prevalence of hepatic steatosis and vascular thrombosis in COVID-19: A systematic review and metaanalysis of autopsy data. World J Gastroenterol 2020; 26: 7693-7706 [PMID: 33505145 DOI: 10.3748/wjg.v26.i48.7693]
- Yu D, Du Q, Yan S, Guo X-G, He Y, Zhu G, Zhao K, Ouyang S. Liver injury in COVID-19: Clinical features and 14 treatment management. Virol J 2021; 18: 121 [DOI: 10.1186/s12985-021-01593-1]
- Algahtani SA, Schattenberg JM. Liver injury in COVID-19: The current evidence. United European Gastroenterol J 2020; 15 8: 509-519 [PMID: 32450787 DOI: 10.1177/2050640620924157]
- 16 Fassan M, Mescoli C, Sbaraglia M, Guzzardo V, Russo FP, Fabris R, Trevenzoli M, Pelizzaro F, Cattelan AM, Basso C, Navalesi P, Farinati F, Vettor R, Dei Tos AP. Liver histopathology in COVID-19 patients: A mono-Institutional series of liver biopsies and autopsy specimens. Pathol Res Pract 2021; 221: 153451 [PMID: 33932720 DOI: 10.1016/j.prp.2021.153451]
- Zghal M, Bouhamed M, Mellouli M, Triki M, Kallel R, Ayedi L, Boudawara TS, Makni S. Liver injury in COVID-19: 17 pathological findings. Pan Afr Med J 2022; 41: 56 [PMID: 35317475 DOI: 10.11604/pamj.2022.41.56.31114]
- 18 Radzina M, Putrins DS, Micena A, Vanaga I, Kolesova O, Platkajis A, Viksna L. Post-COVID-19 Liver Injury: Comprehensive Imaging With Multiparametric Ultrasound. J Ultrasound Med 2022; 41: 935-949 [PMID: 34241914 DOI: 10.1002/jum.15778]
- Schmit G, Lelotte J, Vanhaebost J, Horsmans Y, Van Bockstal M, Baldin P. The Liver in COVID-19-Related Death: 19 Protagonist or Innocent Bystander? Pathobiology 2021; 88: 88-94 [PMID: 33108789 DOI: 10.1159/000512008]
- Nardo AD, Schneeweiss-Gleixner M, Bakail M, Dixon ED, Lax SF, Trauner M. Pathophysiological mechanisms of liver 20 injury in COVID-19. Liver Int 2021; 41: 20-32 [PMID: 33190346 DOI: 10.1111/liv.14730]
- Xu L, Liu J, Lu M, Yang D, Zheng X. Liver injury during highly pathogenic human coronavirus infections. Liver Int 2020; 21 40: 998-1004 [PMID: 32170806 DOI: 10.1111/liv.14435]
- Kumar P, Sharma M, Kulkarni A, Rao PN. Pathogenesis of Liver Injury in Coronavirus Disease 2019. J Clin Exp Hepatol 22 2020; 10: 641-642 [PMID: 32837092 DOI: 10.1016/j.jceh.2020.05.006]
- 23 Saviano A, Wrensch F, Ghany MG, Baumert TF. Liver Disease and Coronavirus Disease 2019: From Pathogenesis to Clinical Care. Hepatology 2021; 74: 1088-1100 [PMID: 33332624 DOI: 10.1002/hep.31684]
- 24 Lozano-Sepulveda SA, Galan-Huerta K, Martínez-Acuña N, Arellanos-Soto D, Rivas-Estilla AM. SARS-CoV-2 another kind of liver aggressor, how does it do that? Ann Hepatol 2020; 19: 592-596 [PMID: 32858226 DOI: 10.1016/j.aohep.2020.08.062
- Lizardo-Thiebaud MJ, Cervantes-Alvarez E, Limon-de la Rosa N, Tejeda-Dominguez F, Palacios-Jimenez M, Méndez-25 Guerrero O, Delaye-Martinez M, Rodriguez-Alvarez F, Romero-Morales B, Liu WH, Huang CA, Kershenobich D, Navarro-Alvarez N. Direct or Collateral Liver Damage in SARS-CoV-2-Infected Patients. Semin Liver Dis 2020; 40: 321-330 [PMID: 32886936 DOI: 10.1055/s-0040-1715108]
- 26 Lei HY, Ding YH, Nie K, Dong YM, Xu JH, Yang ML, Liu MQ, Wei L, Nasser MI, Xu LY, Zhu P, Zhao MY. Potential effects of SARS-CoV-2 on the gastrointestinal tract and liver. Biomed Pharmacother 2021; 133: 111064 [PMID: 33378966 DOI: 10.1016/j.biopha.2020.111064]
- 27 Guardigni V, Del Turco ER, Badia L, Galli S, Scolz K, Viale P, Verucchi G. Pre-existing HBV, and HCV infections do not affect COVID-19-related outcomes: An observational retrospective study. Hepatitis 2021; 21: 5 [DOI: 10.5812/hepatmon.116986]
- 28 Mohammed A, Paranji N, Chen PH, Niu B. COVID-19 in Chronic Liver Disease and Liver Transplantation: A Clinical Review. J Clin Gastroenterol 2021; 55: 187-194 [PMID: 33394628 DOI: 10.1097/MCG.00000000001481]



- Driggers KE, Sadowski BW, Shagla E, Kwok RM. Care of the Hepatology Patient in the COVID-19 Era. Curr Hepatol 29 *Rep* 2022; **21**: 9-20 [PMID: 35382426 DOI: 10.1007/s11901-021-00581-x]
- 30 Kariyawasam JC, Jayarajah U, Abeysuriya V, Riza R, Seneviratne SL. Involvement of the Liver in COVID-19: A Systematic Review. Am J Trop Med Hyg 2022; 106: 1026-1041 [PMID: 35203056 DOI: 10.4269/ajtmh.21-1240]
- 31 Kulkarni AV, Tevethia HV, Premkumar M, Arab JP, Candia R, Kumar K, Kumar P, Sharma M, Rao PN, Reddy DN. Impact of COVID-19 on liver transplant recipients-A systematic review and meta-analysis. EClinicalMedicine 2021; 38: 101025 [PMID: 34278287 DOI: 10.1016/j.eclinm.2021.101025]
- 32 Vranić L, Radovan A, Poropat G, Mikolašević I, Milić S. Non-Alcoholic Fatty Liver Disease and COVID-19-Two Pandemics Hitting at the Same Time. Medicina (Kaunas) 2021; 57 [PMID: 34684094 DOI: 10.3390/medicina57101057]
- Chowdhury T, Sultana J, Dutta J, Gousy N, Hassan KN. Rapid Turn From Cirrhosis to Encephalopathy Following 33 COVID-19 Infection: A Cautionary Tale. Cureus 2022; 14: e22089 [PMID: 35308747 DOI: 10.7759/cureus.22089]
- Ridruejo E, Soza A. The liver in times of COVID-19: What hepatologists should know. Ann Hepatol 2020; 19: 353-358 34 [PMID: 32425991 DOI: 10.1016/j.aohep.2020.05.001]
- 35 Gilead. Food and Drug Administration approves Gilead's antiviral Veklury® (remdesivir) for treatment of COVID-19. Gilead: Press Releases. 2020 Oct. [cited 20 June 2022] [DOI: 10.1186/isrctn15874265]
- 36 Zhang JJ, Dong X, Cao YY, Yuan YD, Yang YB, Yan YQ, Akdis CA, Gao YD. Clinical characteristics of 140 patients infected with SARS-CoV-2 in Wuhan, China. Allergy 2020; 75: 1730-1741 [PMID: 32077115 DOI: 10.1111/all.14238]
- 37 LiverTox: Clinical and Research Information on Drug-Induced Liver Injury. Remdesivir. National Institute of Diabetes and Digestive and Kidney Diseases. Feb, 2022. [cited 20 June 2022]. In: National Library of Medicine [DOI: 10.1002/cld.388]
- Phend C. Remdesivir safety forecast: Watch the liver, kidneys. Medpage Today. 2020 May. [cited 17 June 2022]. In: 38 Medpage Today [DOI: 10.51362/202221710]
- Moon AM, Barritt AS. Elevated liver enzymes in patients with COVID-19: Look, but not too hard. Dig Dis Sci 2021; 66: 39 1767-1769 [PMID: 32875529 DOI: 10.1007/s10620-020-06585-9]
- 40 COVID-19 Treatment Guidelines. Antiviral drugs that are approved, authorized, or under evaluation for treatment of COVID-19. Apr, 2022. [cited 27 June 2022]. Available from: https://www.empr.com/home/news/acetylcysteine-treatmentfor-acute-liver-failure-associated-with-remdesivir/
- 41 Meini S, Pagotto A, Longo B, Vendramin I, Pecori D, Tascini C. Role of Lopinavir/Ritonavir in the Treatment of Covid-19: A Review of Current Evidence, Guideline Recommendations, and Perspectives. J Clin Med 2020; 9 [PMID: 32629768 DOI: 10.3390/jcm9072050]
- 42 Li Y, Xie Z, Lin W, Cai W, Wen C, Guan Y, Mo X, Wang J, Wang Y, Peng P, Chen X, Hong W, Xiao G, Liu J, Zhang L, Hu F, Li F, Zhang F, Deng X, Li L. Efficacy and Safety of Lopinavir/Ritonavir or Arbidol in Adult Patients with Mild/ Moderate COVID-19: An Exploratory Randomized Controlled Trial. Med 2020; 1: 105-113.e4 [PMID: 32838353 DOI: 10.1016/j.medj.2020.04.001]
- 43 Cao B, Wang Y, Wen D, Liu W, Wang J, Fan G, Ruan L, Song B, Cai Y, Wei M, Li X, Xia J, Chen N, Xiang J, Yu T, Bai T, Xie X, Zhang L, Li C, Yuan Y, Chen H, Li H, Huang H, Tu S, Gong F, Liu Y, Wei Y, Dong C, Zhou F, Gu X, Xu J, Liu Z, Zhang Y, Shang L, Wang K, Li K, Zhou X, Dong X, Qu Z, Lu S, Hu X, Ruan S, Luo S, Wu J, Peng L, Cheng F, Pan L, Zou J, Jia C, Liu X, Wang S, Wu X, Ge Q, He J, Zhan H, Qiu F, Guo L, Huang C, Jaki T, Hayden FG, Horby PW, Zhang D, Wang C. A Trial of Lopinavir-Ritonavir in Adults Hospitalized with Severe Covid-19. N Engl J Med 2020; 382: 1787-1799 [PMID: 32187464 DOI: 10.1056/NEJMoa2001282]
- Jiang S, Wang R, Li L, Hong D, Ru R, Rao Y, Miao J, Chen N, Wu X, Ye Z, Hu Y, Xie M, Zuo M, Lu X, Qiu Y, Liang T. 44 Liver injury in critically ill and non-critically ill COVID-19 patients: A multicenter, retrospective, observational study. Front Med 2020; 7: 347 [PMID: 32656222 DOI: 10.3389/fmed.2020.00347]
- Scheuplein VA, Seifried J, Malczyk AH, Miller L, Höcker L, Vergara-Alert J, Dolnik O, Zielecki F, Becker B, Spreitzer I, König R, Becker S, Waibler Z, Mühlebach MD. High secretion of interferons by human plasmacytoid dendritic cells upon recognition of Middle East respiratory syndrome coronavirus. J Virol 2015; 89: 3859-3869 [PMID: 25609809 DOI: 10.1128/JVI.03607-14]
- 46 Alavi Darazam I, Shokouhi S, Pourhoseingholi MA, Irvani SSN, Mokhtari M, Shabani M, Amirdosara M, Torabinavid P, Golmohammadi M, Hashemi SP, Azimi A, Maivan MHJ, Rezaei O, Zali A, Hajiesmaeili M, Dehbsneh HS, Kusha AH, Shoushtari MT, Khalili N, Soleymaninia A, Gachkar L, Khoshkar A. Role of interferon therapy in severe COVID-19: The COVIFERON randomized controlled trial. Sci Rep 2021; 11: 8059 [PMID: 33850184 DOI: 10.1038/s41598-021-86859-y]
- Davoudi-Monfared E, Rahmani H, Khalili H, Hajiabdolbaghi M, Salehi M, Abbasian L, Kazemzadeh H, Yekaninejad MS. 47 A Randomized Clinical Trial of the Efficacy and Safety of Interferon β-1a in Treatment of Severe COVID-19. Antimicrob Agents Chemother 2020; 64 [PMID: 32661006 DOI: 10.1128/AAC.01061-20]
- Raschi E, Caraceni P, Poluzzi E, De Ponti F. Baricitinib, JAK inhibitors and liver injury: a cause for concern in COVID-48 19? Expert Opin Drug Saf 2020; 19: 1367-1369 [PMID: 32840116 DOI: 10.1080/14740338.2020.1812191]
- Serviddio G, Villani R, Stallone G, Scioscia G, Foschino-Barbaro MP, Lacedonia D. Tocilizumab and liver injury in 49 patients with COVID-19. Therap Adv Gastroenterol 2020; 13: 1756284820959183 [PMID: 33101458 DOI: 10.1177/1756284820959183]
- Muhović D, Bojović J, Bulatović A, Vukčević B, Ratković M, Lazović R, Smolović B. First case of drug-induced liver 50 injury associated with the use of tocilizumab in a patient with COVID-19. Liver Int 2020; 40: 1901-1905 [PMID: 32478465 DOI: 10.1111/liv.14516]
- Cornberg M, Buti M, Eberhardt CS, Grossi PA, Shouval D. EASL position paper on the use of COVID-19 vaccines in 51 patients with chronic liver diseases, hepatobiliary cancer and liver transplant recipients. J Hepatol 2021; 74: 944-951 [PMID: 33563499 DOI: 10.1016/j.jhep.2021.01.032]
- Alukal JJ, Naqvi HA, Thuluvath PJ. Vaccination in Chronic Liver Disease: An Update. J Clin Exp Hepatol 2022; 12: 937-947 [PMID: 34975241 DOI: 10.1016/j.jceh.2021.12.003]
- 53 Rolak S, Said A, German R, Hayney MS, Caldera F. Optimizing Immunization Strategies in Adult Patients With Chronic Liver Disease and Liver Transplant Recipients. Gastroenterol Hepatol 2022; 18: 196-206 [PMID: 35505940]



- 54 Hall VG, Ferreira VH, Ku T, Ierullo M, Majchrzak-Kita B, Chaparro C, Selzner N, Schiff J, McDonald M, Tomlinson G, Kulasingam V, Kumar D, Humar A. Randomized Trial of a Third Dose of mRNA-1273 Vaccine in Transplant Recipients. N Engl J Med 2021; 385: 1244-1246 [PMID: 34379917 DOI: 10.1056/NEJMc2111462]
- 55 Hoo CZ, Tan KC, Abdullah S, Sim BLH, Omar H, Tan SS. Severe hepatocellular liver injury after COVID-19 vaccination without autoimmune hepatitis features: A case series. ACG Case Rep J 2022; 9: pe00760 [DOI: 10.14309/crj.0000000000000760]
- 56 Tsoris A, Marlar CA. Use of the Child-Pugh score in liver disease. StatPearls. 2022 Mar. [cited 20 June 2022]. In: National Library of Medicine [DOI: 10.1016/j.crad.2022.09.021]
- 57 World Health Organization. COVID-19 advice for the public: Getting vaccinated. Apr, 2022. [cited 20 June 2022] [DOI: 10.1007/978-3-030-89125-1_6]
- 58 Thuluvath PJ, Robarts P, Chauhan M. Analysis of antibody responses after COVID-19 vaccination in liver transplant recipients and those with chronic liver diseases. J Hepatol 2021; 75: 1434-1439 [PMID: 34454993 DOI: 10.1016/j.jhep.2021.08.008]
- Tian S, Xiong Y, Liu H, Niu L, Guo J, Liao M, Xiao SY. Pathological study of the 2019 novel coronavirus disease 59 (COVID-19) through postmortem core biopsies. Mod Pathol 2020; 33: 1007-1014 [PMID: 32291399 DOI: 10.1038/s41379-020-0536-x
- Ng DL, Al Hosani F, Keating MK, Gerber SI, Jones TL, Metcalfe MG, Tong S, Tao Y, Alami NN, Haynes LM, Mutei MA, 60 Abdel-Wareth L, Uyeki TM, Swerdlow DL, Barakat M, Zaki SR. Clinicopathologic, Immunohistochemical, and Ultrastructural Findings of a Fatal Case of Middle East Respiratory Syndrome Coronavirus Infection in the United Arab Emirates, April 2014. Am J Pathol 2016; 186: 652-658 [PMID: 26857507 DOI: 10.1016/j.ajpath.2015.10.024]
- Elnaggar M, Abomhya A, Elkhattib I, Dawoud N, Doshi R. COVID-19 and liver diseases, what we know so far. World J 61 Clin Cases 2022; 10: 3969-3980 [PMID: 35665122 DOI: 10.12998/wjcc.v10.i13.3969]
- 62 Russo FP, Burra P, Zanetto A. COVID-19 and liver disease: where are we now? Nat Rev Gastroenterol Hepatol 2022; 19: 277-278 [PMID: 35301465 DOI: 10.1038/s41575-022-00607-9]
- Hartl L, Haslinger K, Angerer M, Jachs M, Simbrunner B, Bauer DJM, Semmler G, Scheiner B, Eigenbauer E, Strassl R, 63 Breuer M, Kimberger O, Laxar D, Trauner M, Mandorfer M, Reiberger T. Age-adjusted mortality and predictive value of liver chemistries in a Viennese cohort of COVID-19 patients. Liver Int 2022; 42: 1297-1307 [PMID: 35412018 DOI: 10.1111/liv.15274]



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Observational Study

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ORIGINAL ARTICLE

Demographic and risk characteristics of healthcare workers infected with SARS-CoV-2 from two tertiary care hospitals in the United Arab **Emirates**

Prashant Nasa, Payal Modi, Gladys Setubal, Aswini Puspha, Surjya Upadhyay, Syed Habib Talal

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Prashant Nasa, Syed Habib Talal, Critical Care Medicine, NMC Specialty Hospital, Dubai 7832, United Arab Emirates

Prashant Nasa, Department of Internal Medicine, College of Medicine and Health Sciences, Al Ain 15551, United Arab Emirates

Payal Modi, Department of Microbiology, NMC Royal Hospital, Dubai Investment Park, Dubai 7832, United Arab Emirates

Gladys Setubal, Prevention and Control of Infection, NMC Specialty Hospital, Dubai 7832, Dubai, United Arab Emirates

Aswini Puspha, Prevention and Control of Infection, NMC Royal Hospital, Dubai Investment Park, Dubai 7832, United Arab Emirates

Surjya Upadhyay, Department of Anaesthesiology, NMC Royal Hospital, Dubai Investment Park, Dubai 7832, United Arab Emirates

Corresponding author: Prashant Nasa, MD, Chief Doctor, Critical Care Medicine, NMC Specialty Hospital, Al Nahda 2, Amman Street, Dubai 7832, United Arab Emirates. dr.prashantnasa@hotmail.com

Abstract

BACKGROUND

Understanding the transmission dynamics of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection among healthcare workers (HCWs) and their social contacts is crucial to plan appropriate risk-reduction measures.

AIM

To analyze the socio-demographic risk factors and transmission of SARS-CoV-2 infection among HCWs in two tertiary care hospitals in Dubai, United Arab Emirates.

METHODS

The demographic and clinical characteristics were available for all HCWs in both facilities from the human resources department. A cross-sectional survey was conducted from January-April 2022 among HCWs who tested positive through



Reverse Transcriptase Polymerase Chain Reaction of the nasopharyngeal swab for SARS-CoV-2 between March 2020 and August 2021 in two tertiary-level hospitals. The survey included questions on demographics, work profile, characteristics of coronavirus disease 2019 (COVID-19), and infection among their household or co-workers. The survey also checked the knowledge and perception of participants on the infection prevention measures related to SARS-CoV-2.

RESULTS

Out of a total of 346 HCWs infected with SARS-CoV-2, 286 (82.7%) HCWs consented to participate in this study. From the sample population, 150 (52.5%) of participants were female, and a majority (230, 80.4%) were frontline HCWs, including 121 nurses (121, 42.4%). Only 48 (16.8%) participants were fully vaccinated at the time of infection. Most infected HCWs (85%) were unaware of any unprotected exposure and were symptomatic at the time of testing (225, 78.7%). Nearly half of the participants (140, 49%) had co-infection among household, and nearly one-third (29.5%) had coinfection among three or more household. Another 108 (37.8%) participants reported crossinfection among co-workers. The frontline HCWs were significantly more infected (25.1% vs 8.6%, P < 0.001) compared to non-frontline HCWs. Another significant risk factor for a high infection rate was male sex (P < 0.001). Among the infected frontline HCWs, a significantly higher proportion were male and shared accommodation with family (P < 0.001). COVID-19 vaccination significantly reduced the infection rate (83.2% vs 16.8, P < 0.001) among HCWs. Most participants (99.3%) were aware about importance of appropriate use of personal protective equipment. However, only 70% agreed with the efficacy of the COVID-19 vaccination in preventing an infection and severe disease.

CONCLUSION

The risk profiling of the HCWs infected with SARS-CoV-2 found that working at frontline and being male increase the rate of infection. COVID-19 vaccination can effectively reduce the rate of transmission of SARS-CoV-2 among HCWs.

Key Words: Coronavirus disease 2019; Risk factors; Disease transmission, infectious; Infectious disease transmission, Professional-to-patient; Health personnel; Socioeconomic factors

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Core Tip: The healthcare workers (HCWs) are vulnerable to infection with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). In the current study, the authors found that the frontline and male HCWs were at higher risk of infection. Among the infected frontline HCWs, a significantly higher proportion were male and staying in a rented accommodation with family. The coronavirus disease 2019 vaccination is effective in preventing the transmission of SARS-CoV-2 among HCWs. This information can be utilised for the healthcare workforce management and to formulate strategies to mitigate the risk of transmission of SARS-CoV-2 to the HCWs.

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INTRODUCTION

The coronavirus disease 2019 (COVID-19) pandemic has overwhelmed the healthcare resources across the globe. Since the inception of the pandemic, reports have been published on the increased vulnerabilities of healthcare workers (HCWs) compared to the general community for infection with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)[1,2]. A prospective cohort study conducted among 99795 HCWs reported that the HCWs are at a threefold higher risk of acquiring COVID-19 compared to the general community. However, the risk of exposure is not uniform and depends on multiple factors, such as the nature of work (frontline), race or ethnicity (Black, Asian, and other ethnic minorities), and access to or reuse of the personal protective equipment (PPE)[1]. Besides the risk of illness, the HCWs are at considerable risk of adverse mental health during the COVID-19 pandemic[3]. Moreover, the social and household contacts of the HCWs are also potentially vulnerable to SARS-CoV-2 infection[4].



On the other hand, the absenteeism of HCWs from work is further detrimental to the already stretched healthcare services during the pandemic^[5].

From the start of COVID-19 pandemic, the experts strongly expressed concerns regarding the nosocomial transmission of SARS-CoV-2[6,7]. HCWs were assumed to play a pivotal role in the transmission chain during a nosocomial outbreak of SARS-CoV-2. However, limited information exists on the transmission characteristics and dynamics of SARS-CoV-2 infection among the HCWs or their social contacts. In this scenario, it is crucial to explore the dynamics of SARS-CoV-2 transmission among the HCWs and their social contacts to develop and implement appropriate risk-reduction measures[6].

In the current study, the authors performed a retrospective analysis of HCWs infected with SARS-CoV-2 to analyze the socio-demographic risk factors and the characteristics of SARS-CoV-2 infection among HCWs and their social contacts.

MATERIALS AND METHODS

The demographic and clinical characteristics were available for all HCWs in both facilities from the human resources department. A cross-sectional survey was conducted between January and April 2022 among the HCWs who tested positive for reverse transcriptase polymerase chain reaction (RT-PCR) for SARS-CoV-2 between March 2020 and August 2021 in two multi-specialty tertiary-level hospitals located in Dubai. The cross-sectional survey was conducted to extract further information from the infected HCWs on their social contacts, including household. The survey included Multiple-Choice Questions and questions with 5-point Likert scale. The survey questionnaire, attached in the Supplementary material, has a total of three sections: (1) Demographic details of the participants, including age, gender, department, nature of work, and COVID-19 vaccination status; (2) Details about SARS-CoV-2 infection, the reason for RT-PCR testing, severity and duration of the symptoms, and infection among their household contacts and co-workers; and (3) Knowledge and perception among the participants on PPE and infection prevention measures related to SARS-CoV-2. The human resource department, who were not part of the data analysis, sent the survey questionnaire through e-mail. The identity of the participants was kept confidential. Frontline HCWs were those who provide care for patients with COVID-19 or worked in areas with direct patient contact during the pandemic. As per the local health regulatory requirements, the HCWs were tested with RT-PCR only in case of symptomatic infections, contact tracing, or pre-travel screening during this period. The study considered only the first SARS-CoV-2 infection for further analysis. In the United Arab Emirates (UAE), seven COVID-19 vaccines were approved for use and are made available to the public for free of cost. The data on the average number of new cases in the community was extracted from the website of National Emergency Crisis and Disaster Management Authority, UAE (https://covid19.ncema.gov.ae/en). The study was approved by the scientific and ethical committee of the hospital and Dubai Scientific Research Ethics Committee (DSREC/09/2020_32).

Statistical analysis

Descriptive statistical analysis was conducted for frequencies, percentages, medians, and ranges. Continuous data was presented as mean [standard deviation (SD)] or median with Interquartile Range (IQR). Two groups were compared in this study using the 2-sample test for equality of proportions with continuity correction (Chi-square). A comparison was made between the categorical paired data with McNemar Test. The authors used Fisher exact test to compare less than five-count cells. All the tests were 2-tailed, and P < 0.05 was considered to be significant. The statistical analyses were conducted using R version 3.4.2 from the Comprehensive R Archive Network (R Core Team, 2020).

RESULTS

Out of a total of 1568 HCWs working in both hospitals, 346 (22.1%) tested positive for SARS-CoV-2 RT-PCR during the study period. Amongst this study population, 16 (4.6%) HCWs were found to be reinfected with SARS-CoV-2. However, as mentioned earlier, only the first infection was considered for the analysis. From the 346 infected HCWs, 286 (82.7%) HCWs agreed to participate in the cross-sectional survey. Amongst the participants, 150 (52.5%) were female, whereas a majority of the participants (230, 80.4%) were frontline HCWs, including 121 nurses (121, 42.4%). Only 48 (16.8%) participants were fully vaccinated at the time of infection.

Most of the participants (225, 78.7%) were symptomatic at the time of RT-PCR testing. Among the asymptomatic HCWs, 35 (12.2%) were tested for close contact tracing. Nearly half of the participants (140, 49%) had a co-infection with their household contacts. Moreover, half (48, 51.6%) of the infection in the households occurred in a single person, while nearly one-third (29.5%) had infection among three or more households. Further, 108 (37.8%) participants reported cross-infection among their co-workers (Table 1).



Table 1 Descriptive data on demographic and risk-profile of the healthcare workers infected with severe acute respiratory syndrome

coronavirus 2		
Variables		n (%)
Age, yr		36 (IQR-10)
Female		150 (52.5)
Staff travelling in hospital accommodation		454/1467 (29)
Work profile of healthcare workers	Doctor	34 (11.9)
	Nurses	121 (42.4)
	Technician	37 (12.9)
	Pharmacy	11 (3.8)
	Paramedical staff	10 (3.5)
	Non-clinical staff	73 (25.5)
Reason for testing with RT-PCR	Symptomatic	225 (78.7)
	Contact tracing	35 (12.2)
	Travel screening	8 (2.8)
	Other	18 (6.3)
Severity of COVID-19	Asymptomatic	36 (12.6)
	Mild	220 (76.9)
	Moderate	25 (8.7)
	Severe	5 (1.7)
Place of isolation	Institutional	50 (17.4)
	Home isolation	189 (66.1)
	Hospitalization	49 (17.1)
Symptom duration	< 1 wk	99 (34.6)
	1-2 wk	128 (44.8)
	2-3 wk	49 (17.1)
	> 3 wk	10 (3.5)
Pre-existing chronic illness	Diabetes Mellitus	33 (11.5)
	Hypertension	31 (10.8)
	Chronic respiratory disease	6 (2.1)
	Chronic kidney disease	2 (0.7)
	Other	4 (1.4)
	None	210 (73.5)
HCWs with households infected within 14 d	Total	140 (49)
	Earlier (3-14 d)	49 (35)
	Same time (within 2 d)	55 (39.3)
	Later (3-14 d)	36 (25.7)
Number of infected households	1	48 (51.6)
	2	18 (18.9)
	3	16 (16.8)
	> 3	12 (12.7)
HCWs with co-workers infected within 14 d	Total	108 (37.8)
	Earlier (3-14 d)	38 (35.2)



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	Same time (within 2 d)	33 (30.5)
	Later (3-14 d)	37 (34.3)
Number of infected co-workers	1	23 (24.2)
	2	16 (16.8)
	3	3 (3.1)
	> 3	17 (17.9)

COVID-19: Coronavirus disease 2019; HCWs: Healthcare workers; RT-PCR: Reverse transcriptase polymerase chain reaction.

When compared between the infected and the uninfected HCWs, frontline HCWs (25.1% vs 8.6%, P < 0.001), who were males (54% vs 46%, P < 0.001) recorded a significantly high infection rate. The infection rate among the unvaccinated HCWs (83.2% vs 16.8%, P < 0.001) was nearly five times higher than those HCWs who were vaccinated against COVID-19. The study found that the type of accommodation (selfowned vs hospital sponsored) showed no significance effect on the infection rate (Table 2). A significantly high proportion of the infected frontline HCWs were males who stayed in rented accommodation with family (P < 0.001) (Table 3). Finally, the trend chart of a month-wise comparison of the infected HCWs and the average new cases in UAE showed three peaks.

The survey also tried to assess the knowledge and perception of the participants about safety precaution, vaccination, and the disease. Most of the participants were aware about the appropriate usage of PPE (99.3%) and did not agree to unprotected exposure to a patient with COVID-19 (85%). Around 74% of the participants agreed with the importance of social precautions like face mask, social distancing, and hand hygiene in preventing the SARS-CoV-2 infection. Only 70% agreed on the efficacy of COVID-19 vaccination in preventing infection or progression to the severe disease. The deficiency of PPE at the workplace was reported by 23.4% of the participants, whereas 29.7% participants wanted an improvement in the quality and availability of the PPE (Table 4).

DISCUSSION

This cross-sectional analysis of RT-PCR-positive HCWs from two tertiary care hospitals showed that frontline HCWs had a significantly higher infection rate. The study infers that being a male is a significant risk factors for getting infected with COVID-19. Among the infected frontline staff, a significantly higher proportion were male who shared their accommodation with family members. COVID-19 vaccination was effective in reducing the rate of infection among HCWs.

From the start of COVID-19 pandemic, various studies recorded a higher infection rate among the frontline HCWs. The risk was higher due to the reuse or inadequate availability of the PPEs and due to which the studies advocated strategies like access to high-quality PPEs and early COVID-19 vaccination to curb the spread of the virus [2,8]. Nearly one-fourth of the participants in this study reported insufficient access to the PPEs, while most were unaware of any unprotected exposure with COVID-19 patients. Limited access to adequate PPE has been linked with higher odds of infection[9-11]. Hence, ensuring access to high-quality PPEs for HCWs is an important workplace risk-reduction measure. The rate of infection was significantly higher among the male HCWs as found in other studies[2,12].

Around 13% of the study participants had asymptomatic infection. The number of asymptomatic infections could have been higher, if the hospitals had routine surveillance testing for the HCWs. However, the impact of the routine surveillance testing of asymptomatic HCWs in preventing nosocomial transmission of SARS-CoV-2 is unknown[13]. A consensus experts' panel recommended testing the HCWs to get tested for SARS-CoV-2 only when they are symptomatic or when they encountered unprotected exposure over routine testing^[14].

Around 38% of the infected participants agreed to infection among their co-workers within 14 days of their own infection, and nearly one-fifth of them agreed to have three or more infected co-workers. Moreover, sharing accommodation with family or friends was significantly higher among the infected frontline HCWs. In the absence of epidemiological investigation and genomic sequencing, these infections cannot be segregated as an outbreak. However, the absenteeism of multiple HCWs from the same department can disrupt the services of already overwhelmed frontline departments during the pandemic. Despite various published reports on a nosocomial outbreak of SARS-CoV-2, ambiguity exists regarding the role of HCWs in initiating or amplifying the nosocomial outbreaks[6,15].

Most epidemiological research on SARS-CoV-2 infection among the HCWs has focused on transmission dynamics within the hospital setting. However, the research on the impact of socialcultural and demographic factors on the transmission of SARS-CoV-2 among HCWs is lacking. Recently, a large prospective study conducted in the United Kingdom found the effect of sociodemographic characteristics on the risk of infection among the vulnerable HCWs. The study found that



Table 2 Comparison of risk-factors among infected and non-infected healthcare workers, <i>n</i> (%)			
	Uninfected HCWs, <i>n</i> = 1282	Infected HCWs, <i>n</i> = 346	<i>P</i> value
Accommodation			
Hospital sponsored	381 (29.7)	89 (25.9)	0.15
Self-owned	901 (70.3)	257 (74.1)	(χ^2 statistic value, 2.12)
Work profile			
Frontline	689 (53.7)	278 (80.4)	< 0.001
Non-frontline	593 (46.3)	67(19.6)	(χ^2 statistic value, 81.19)
COVID-19 vaccination			
Vaccinated	1198 (93.2)	58 (16.8)	< 0.001
Unvaccinated	88 (6.8)	287 (83.2)	(χ^2 statistic value, 895.49)
Sex			
Male	511 (39.9)	187 (54.0)	< 0.001
Female	771 (60.1)	159 (46.0)	(χ^2 statistic value, 22.38)

COVID-19: Coronavirus disease 2019; HCWs: Healthcare workers.

Table 3 Comparison of demographics among frontline *vs* non-frontline healthcare workers infected with severe acute syndrome coronavirus 2, *n* (%)

	Frontline healthcare workers, <i>n</i> = 238	Non-Frontline healthcare workers, <i>n</i> = 48	P value
Accommodation			
Shared with family	167 (70.2)	23 (47.9)	< 0.001
Shared with friends	60 (25.2)	24 (50)	(χ^2 statistic value, 31.07)
Non-shared	11 (4.6)	1 (2.1)	
Accommodation			
Self-rented	140 (58.8)	18 (11.4)	< 0.001
Hospital provided	49 (20.6)	25 (33.8)	(χ^2 statistic value, 42.68)
Others	49 (20.6)	5 (9.3)	
Sex			
Male	205 (86.1)	35 (72.9)	0.02
Female	33 (13.9)	13 (27.1)	(χ^2 statistic value, 5.17)

amongst the demographic and household risk factors, young age, living with a co-worker, and high religiosity are associated with high infection odds among the HCWs[9]. In another study, high odds of infection were observed among the HCWs from community contact with a suspected or a confirmed COVID-19 individual, instead of the workplace[16]. Socio-demographic risk factors may differ based on the culture and geographical differences, and the availability of resources. The cross-transmission of SARS-CoV-2 among the household is well-established concept and persists even during the low-community transmission[17].

The current study also found a significantly higher proportion of the infected frontline HCWs were staying in shared accommodation. When comparing infected HCWs per month with average new cases in the community, an agreement was observed in the peaks of two trend charts (Figure 1). This pattern reveals a synchronization in the infection rate among the HCWs and the transmission rate of SARS-CoV-2 infection in the community. Hence, the HCWs are vulnerable to contracting the infection from their households and social contacts, especially with a higher rate of SARS-CoV-2 transmission in the community. Hospital leadership can utilize this valuable insight for workforce management and to develop strategies to mitigate the risk of exposure to HCWs. Theoretically, public transport can be another risk factor for transmission. However, as reported in the literature, the current study authors did not find any increased transmission risk with public transport[18].

Table 4 Attitude and perception of the infected healthcare workers, n (%)		
Are you aware about appropriate personal protective equipment for the care of COVID-19 patients?	Yes: 99.3%; No: 0.7%	
Have you ever been exposed to a COVID-19 patient without adequate PPE?	Yes: 43 (15); No: 243 (85)	
There was always enough PPE in my workplace	Agree: 183 (64); Neutral: 36 (12.6); Disagree: 67 (23.4)	
PPE availability and quality should be improved at my workplace	Agree: 85 (29.7); Neutral 49 (17.1); Disagree: 152 (53.2)	
Proper precautions (face mask, hand hygiene, social distance) are most important tools to save you from SARS-CoV-2	Agree: 213 (74.4); Neutral: 12 (4.2); Disagree: 61 (21.3)	
Vaccines for SARS-CoV-2 can reduce infection rate and can prevent severe disease and hospitalisation	Agree: 201 (70.2); Neutral: 27 (9.6); Disagree: 58 (20.2)	

SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2; COVID-19: Coronavirus disease 2019; PPE: Personal protective equipment.

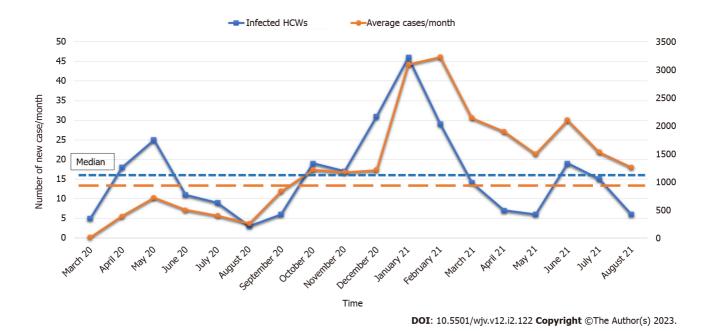


Figure 1 Trend-chart showing the month-wise distribution of infected healthcare workers and new cases in the community. Infected healthcare workers (HCWs) are displayed in absolute numbers per month (orange color) and average cases of severe acute respiratory syndrome coronavirus 2 cases in the community (blue color). Median cases per month are displayed through dashed lines. There are three peaks observed in trend-line. The peaks coincided in the trend charts, representing an increased rate of infection among HCWs with high community transmission. HCWs: Healthcare workers.

> According to a study conducted earlier, the vaccination of the HCWs effectively reduces the risk of severe disease and the transmission of SARS-CoV-2[19]. Advanced age (≥ 65 years), male sex, and other co-morbidities like diabetes mellitus, chronic respiratory disease, hypertension, chronic kidney disease, and cardiovascular disease are risk factors for severe illness and mortality^[20]. COVID-19 vaccination is highly effective in reducing the progression and the severity of disease and intensive care unit (ICU) or hospital admission, especially in the elderly population and patients with co-morbidities[21]. Vaccination is an essential intervention for the HCWs to protect them from getting infected and severe illness that may require hospital or ICU admission. However, the effectiveness of the vaccine in reducing the risk of disease reduces considerably after six months of the last dose. So, a booster dose is recommended for the vulnerable population, including HCWs^[19]. Vaccine hesitancy among the HCWs is a major issue in the successful implementation of the COVID-19 vaccination programme. Only 70.2% of the participants have agreed upon the efficacy of the COVID-19 vaccines. Other studies also found more vaccination hesitation among the previously infected people[22,23]. Hospital leadership and infection preventionist should address the issue of vaccine hesitancy strategically and through collaboration.

Strength and limitations

This is the first study to the best of the author's knowledge from the UAE or the countries in the Gulf Cooperation Council on risk profiling of RT-PCR-positive HCWs with COVID-19 using socio-



demographic factors. The study also evaluated the impact of COVID-19 vaccination on crosstransmission among the HCWs. The current study has a few limitations that are listed herewith. Firstly, the information on social contacts and households was collected through a cross-sectional survey. Hence, there exists a potential recall bias because of the time-gap between the period of infection and data collection. However, to avoid this bias, the data collected from the cross-sectional survey was validated through the human resource records maintained by the hospital.

There is missing data for about 17% of the eligible HCWs who did not participate in the crosssectional survey due to reasons like resignation and immigration to other countries. Secondly, genomic sequencing was not used to confirm the phylogenetic linkage in infection among co-workers or the household. Thirdly, the small cohort size could have missed portraying the complete statistical correlation of various socio-demographic factors. Finally, the impact of the COVID-19 vaccination booster on transmission dynamics was not assessed.

CONCLUSION

The risk profiling of the HCWs, infected with SARS-CoV-2 from two tertiary care hospitals showed that the frontline HCWs had a significantly higher infection rate. Another significant risk factor was male sex. COVID-19 vaccination can effectively reduce the rate of SARS-CoV-2 transmission among HCWs.

ARTICLE HIGHLIGHTS

Research background

There is paucity of the research on the transmission dynamics of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) among the healthcare workers (HCWs) and their co-workers and household. The current study conducted a retrospective analysis of the infected HCWs to analyze the socio-demographic risk factors and characteristics of SARS-CoV-2 transmission among HCWs and their social contacts.

Research motivation

HCWs are vulnerable to SARS-CoV-2 infection during their work, and the potential risk of transmission of SARS-CoV-2 infection from the household and co-workers of HCWs is unclear. This study provides valuable insights for workforce management and helps formulate strategies to mitigate the risk of exposure to the HCWs.

Research objectives

The current study evaluated the risk factors of SARS-CoV-2 infection among HCWs and explored the potential of transmission of SARS-CoV-2 among the household and co-workers of infected HCWs.

Research methods

The health records of all infected HCWs between March 2020 and August 2021 were analysed. The information on the coronavirus disease 2019 (COVID-19) vaccination, household and co-workers of the infected HCWs was collected through a cross-sectional survey.

Research results

The cross-sectional analysis of health records of 346 reverse transcriptase polymerase chain reaction (RT-PCR)-positive HCWs showed that the risk of infection was significantly higher among frontline HCWs. Being male was a significant risk factor for SARS-CoV-2 infection. Among infected frontline staff, a significantly higher proportion were male, and were staying with their families in rented accommodation. COVID-19 vaccination was effective in reducing the infection rate among HCWs.

Research conclusions

Working at the frontline and being male are the significant risk factors for SARS-CoV-2 infection among the HCWs. COVID-19 vaccination is effective in reducing the infection rate among HCWs.

Research perspectives

Future research should explore the role of community transmission of SARS-CoV-2 in the infection of HCWs.

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FOOTNOTES

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Country/Territory of origin: United Arab Emirates

ORCID number: Prashant Nasa 0000-0003-1948-4060; Payal Modi 0000-0001-6628-8121; Gladys Setubal 0000-0002-0512-2694; Aswini Puspha 0000-0003-0910-7085; Surjya Upadhyay 0000-0002-0766-3822; Syed Habib Talal 0000-0001-6719-4725.

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REFERENCES

- Mutambudzi M, Niedwiedz C, Macdonald EB, Levland A, Mair F, Anderson J, Celis-Morales C, Cleland J, Forbes J, Gill 1 J, Hastie C, Ho F, Jani B, Mackay DF, Nicholl B, O'Donnell C, Sattar N, Welsh P, Pell JP, Katikireddi SV, Demou E. Occupation and risk of severe COVID-19: prospective cohort study of 120 075 UK Biobank participants. Occup Environ Med 2020; 78: 307-314 [PMID: 33298533 DOI: 10.1136/oemed-2020-106731]
- 2 Nguyen LH, Drew DA, Graham MS, Joshi AD, Guo CG, Ma W, Mehta RS, Warner ET, Sikavi DR, Lo CH, Kwon S, Song M, Mucci LA, Stampfer MJ, Willett WC, Eliassen AH, Hart JE, Chavarro JE, Rich-Edwards JW, Davies R, Capdevila J, Lee KA, Lochlainn MN, Varsavsky T, Sudre CH, Cardoso MJ, Wolf J, Spector TD, Ourselin S, Steves CJ, Chan AT; COronavirus Pandemic Epidemiology Consortium. Risk of COVID-19 among front-line health-care workers and the general community: a prospective cohort study. Lancet Public Health 2020; 5: e475-e483 [PMID: 32745512 DOI: 10.1016/S2468-2667(20)30164-X
- 3 De Kock JH, Latham HA, Leslie SJ, Grindle M, Munoz SA, Ellis L, Polson R, O'Malley CM. A rapid review of the impact of COVID-19 on the mental health of healthcare workers: implications for supporting psychological well-being. BMC Public Health 2021; 21: 104 [PMID: 33422039 DOI: 10.1186/s12889-020-10070-3]
- Zheng C, Hafezi-Bakhtiari N, Cooper V, Davidson H, Habibi M, Riley P, Breathnach A. Characteristics and transmission dynamics of COVID-19 in healthcare workers at a London teaching hospital. J Hosp Infect 2020; 106: 325-329 [PMID: 32730771 DOI: 10.1016/j.jhin.2020.07.025]
- Schneider S, Piening B, Nouri-Pasovsky PA, Krüger AC, Gastmeier P, Aghdassi SJS. SARS-Coronavirus-2 cases in 5 healthcare workers may not regularly originate from patient care: lessons from a university hospital on the underestimated risk of healthcare worker to healthcare worker transmission. Antimicrob Resist Infect Control 2020; 9: 192 [PMID: 33287908 DOI: 10.1186/s13756-020-00848-w]
- Abbas M, Robalo Nunes T, Martischang R, Zingg W, Iten A, Pittet D, Harbarth S. Nosocomial transmission and outbreaks of coronavirus disease 2019: the need to protect both patients and healthcare workers. Antimicrob Resist Infect Control 2021; 10: 7 [PMID: 33407833 DOI: 10.1186/s13756-020-00875-7]
- Asad H, Johnston C, Blyth I, Holborow A, Bone A, Porter L, Tidswell P, Healy B. Health Care Workers and Patients as 7



Trojan Horses: a COVID19 ward outbreak. *Infect Prev Pract* 2020; **2**: 100073 [PMID: 34316562 DOI: 10.1016/j.infpip.2020.100073]

- 8 Fell A, Beaudoin A, D'Heilly P, Mumm E, Cole C, Tourdot L, Ruhland A, Klumb C, Rounds J, Bailey B, Liverseed G, Peterson M, Mahoehney JP, Ireland M, Bye M, Setty S, Leeds M, Taylor J, Holzbauer S; Minnesota Department of Health COVID-19 HCW Monitoring Response Team; Minnesota Department of Health COVID-19 Response Task Force. SARS-CoV-2 Exposure and Infection Among Health Care Personnel Minnesota, March 6-July 11, 2020. *MMWR Morb Mortal Wkly Rep* 2020; **69**: 1605-1610 [PMID: 33119557]
- 9 Martin CA, Pan D, Nazareth J, Aujayeb A, Bryant L, Carr S, Gray LJ, Gregary B, Gupta A, Guyatt AL, Gopal A, Hine T, John C, McManus IC, Melbourne C, Nellums LB, Reza R, Simpson S, Tobin MD, Woolf K, Zingwe S, Khunti K, Pareek M; UK-REACH Study Collaborative Group. Access to personal protective equipment in healthcare workers during the COVID-19 pandemic in the United Kingdom: results from a nationwide cohort study (UK-REACH). *BMC Health Serv Res* 2022; 22: 867 [PMID: 35790970 DOI: 10.1186/s12913-022-08202-z]
- 10 Kim H, Hegde S, LaFiura C, Raghavan M, Sun N, Cheng S, Rebholz CM, Seidelmann SB. Access to personal protective equipment in exposed healthcare workers and COVID-19 illness, severity, symptoms and duration: a population-based case-control study in six countries. *BMJ Glob Health* 2021; 6 [PMID: 33509841 DOI: 10.1136/bmjgh-2020-004611]
- 11 Dzinamarira T, Nkambule SJ, Hlongwa M, Mhango M, Iradukunda PG, Chitungo I, Dzobo M, Mapingure MP, Chingombe I, Mashora M, Madziva R, Herrera H, Makanda P, Atwine J, Mbunge E, Musuka G, Murewanhema G, Ngara B. Risk Factors for COVID-19 Infection Among Healthcare Workers. A First Report From a Living Systematic Review and meta-Analysis. *Saf Health Work* 2022; **13**: 263-268 [PMID: 35433073 DOI: 10.1016/j.shaw.2022.04.001]
- 12 Chatterjee P, Anand T, Singh KJ, Rasaily R, Singh R, Das S, Singh H, Praharaj I, Gangakhedkar RR, Bhargava B, Panda S. Healthcare workers & SARS-CoV-2 infection in India: A case-control investigation in the time of COVID-19. *Indian J Med Res* 2020; **151**: 459-467 [PMID: 32611916 DOI: 10.4103/ijmr.IJMR_22234_20]
- 13 D'Ettorre G, Pellicani V, Muratore M, Ceccarelli G. Occupational health surveillance of healthcare workers during COVID 19 pandemic: a narrative review. *Acta Biomed* 2022; 93: e2022007 [PMID: 35315429 DOI: 10.23750/abm.v93i1.10277]
- 14 Nasa P, Azoulay E, Chakrabarti A, Divatia JV, Jain R, Rodrigues C, Rosenthal VD, Alhazzani W, Arabi YM, Bakker J, Bassetti M, De Waele J, Dimopoulos G, Du B, Einav S, Evans L, Finfer S, Guérin C, Hammond NE, Jaber S, Kleinpell RM, Koh Y, Kollef M, Levy MM, Machado FR, Mancebo J, Martin-Loeches I, Mer M, Niederman MS, Pelosi P, Perner A, Peter JV, Phua J, Piquilloud L, Pletz MW, Rhodes A, Schultz MJ, Singer M, Timsit JF, Venkatesh B, Vincent JL, Welte T, Myatra SN. Infection control in the intensive care unit: expert consensus statements for SARS-CoV-2 using a Delphi method. *Lancet Infect Dis* 2022; 22: e74-e87 [PMID: 34774188 DOI: 10.1016/S1473-3099(21)00626-5]
- 15 Lindsey BB, Villabona-Arenas CJ, Campbell F, Keeley AJ, Parker MD, Shah DR, Parsons H, Zhang P, Kakkar N, Gallis M, Foulkes BH, Wolverson P, Louka SF, Christou S, State A, Johnson K, Raza M, Hsu S, Jombart T, Cori A; Sheffield COVID-19 Genomics Group; COVID-19 Genomics UK (COG-UK) consortium; CMMID COVID-19 working group, Evans CM, Partridge DG, Atkins KE, Hué S, de Silva TI. Characterising within-hospitalSARS-CoV-2 transmission events using epidemiological and viral genomic data across two pandemic waves. *Nat Commun* 2022; 13: 671 [PMID: 35115517 DOI: 10.1038/s41467-022-28291-y]
- 16 Jacob JT, Baker JM, Fridkin SK, Lopman BA, Steinberg JP, Christenson RH, King B, Leekha S, O'Hara LM, Rock P, Schrank GM, Hayden MK, Hota B, Lin MY, Stein BD, Caturegli P, Milstone AM, Rock C, Voskertchian A, Reddy SC, Harris AD. Risk Factors Associated With SARS-CoV-2 Seropositivity Among US Health Care Personnel. *JAMA Netw* Open 2021; 4: e211283 [PMID: 33688967 DOI: 10.1001/jamanetworkopen.2021.1283]
- 17 Madewell ZJ, Yang Y, Longini IM Jr, Halloran ME, Dean NE. Household Secondary Attack Rates of SARS-CoV-2 by Variant and Vaccination Status: An Updated Systematic Review and Meta-analysis. *JAMA Netw Open* 2022; 5: e229317 [PMID: 35482308 DOI: 10.1001/jamanetworkopen.2022.9317]
- 18 Mohammed I, Nauman A, Paul P, Ganesan S, Chen KH, Jalil SMS, Jaouni SH, Kawas H, Khan WA, Vattoth AL, Al-Hashimi YA, Fares A, Zeghlache R, Zakaria D. The efficacy and effectiveness of the COVID-19 vaccines in reducing infection, severity, hospitalization, and mortality: a systematic review. *Hum Vaccin Immunother* 2022; 18: 2027160 [PMID: 35113777 DOI: 10.1080/21645515.2022.2027160]
- 19 Feikin DR, Higdon MM, Abu-Raddad LJ, Andrews N, Araos R, Goldberg Y, Groome MJ, Huppert A, O'Brien KL, Smith PG, Wilder-Smith A, Zeger S, Deloria Knoll M, Patel MK. Duration of effectiveness of vaccines against SARS-CoV-2 infection and COVID-19 disease: results of a systematic review and meta-regression. *Lancet* 2022; 399: 924-944 [PMID: 35202601 DOI: 10.1016/S0140-6736(22)00152-0]
- 20 Du P, Li D, Wang A, Shen S, Ma Z, Li X. A Systematic Review and Meta-Analysis of Risk Factors Associated with Severity and Death in COVID-19 Patients. *Can J Infect Dis Med Microbiol* 2021; 2021: 6660930 [PMID: 33936349 DOI: 10.1155/2021/6660930]
- 21 Freund O, Tau L, Weiss TE, Zornitzki L, Frydman S, Jacob G, Bornstein G. Associations of vaccine status with characteristics and outcomes of hospitalized severe COVID-19 patients in the booster era. *PLoS One* 2022; 17: e0268050 [PMID: 35536849 DOI: 10.1371/journal.pone.0268050]
- 22 Martin CA, Marshall C, Patel P, Goss C, Jenkins DR, Ellwood C, Barton L, Price A, Brunskill NJ, Khunti K, Pareek M. SARS-CoV-2 vaccine uptake in a multi-ethnic UK healthcare workforce: A cross-sectional study. *PLoS Med* 2021; 18: e1003823 [PMID: 34739480 DOI: 10.1371/journal.pmed.1003823]
- 23 Caspi I, Freund O, Pines O, Elkana O, Ablin JN, Bornstein G. Effect of patient COVID-19 vaccine hesitancy on hospital care team perceptions. *World J Clin Cases* 2023; 11: 821-829 [DOI: 10.12998/wjcc.v11.i4.821]

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LETTER TO THE EDITOR

Utility of cardiac bioenzymes in predicting cardiovascular outcomes in SARS-CoV-2

Ali Osman Gulmez, Sonay Aydin

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Ali Osman Gulmez, Department of Radiology, Erzincan University, Erzincan 24100, Turkey

Sonay Aydin, Department of Radiology, Erzincan University, Erzincan 24100, Turkey

Corresponding author: Ali Osman Gulmez, MD, Academic Research, Department of Radiology, Erzincan University, Mengucek Gazi Education and Research Hospital, Başbağlar Mahallesi Hacı Ali Akın Caddesi No. 32 Erzincan/Merkez, Erzincan 24100, Turkey. aliosmangulmez.2@gmail.com

Abstract

The relationship between coronavirus disease-19 (COVID-19) and cardiovascular diseases has been an important issue. Therefore, cardiac biomarkers and cardiac imaging have an important place in the diagnostic phase. It is important to know the relationship of biomarkers in COVID-19 so that we can understand the diagnosis of the disease, the predicted course and results after diagnosis.

Key Words: Cardiac bioenzymes; Coronavirus disease - 19; Treatment; Diagnosis; Triple rule-out computed tomography angiography; Dual energy computed tomography

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Core Tip: Recommends biomarkers, especially troponin, in patients with Coronavirus disease-19-associated myocarditis and other myocardial damage; however, they have proven that in addition to traditional biomarkers, new cardiac bioenzymes such as prepesin, copeptin also increase and significantly worsen the prognosis. Knowing this, evaluation together with other imaging methods is also important in diagnosis.

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TO THE EDITOR

We read with interest and attention the review written by Muthyala *et al*[1]. One of the main effects of the coronavirus disease 2019 (COVID-19) pandemic is on the cardiovascular system. Therefore, it is important to know and use cardiac biomarkers well, and to come to an advanced point in the diagnosis stage by combining them with cardiac imaging methods. Therefore, the authors discussed the importance of these biomarkers in COVID-19 in order to determine the ways to diagnose the disease, follow-up after diagnosis and treatment. Although biomarkers are important, we also mentioned them in our evaluation since it is important to evaluate them with imaging methods. We think that when we combine cardiac biomarkers and imaging methods, a very important point will be reached in the diagnosis.

Including troponin^[2], which provides us with information about the prognosis in the diagnosis of cardiac acute coronary syndromes and myocardial damage, as well as Brain Natriuretic Peptide (BNP) [3] and pro-BNP, which gives us an advantage in the early detection of heart diseases and understanding the morbidity status of such diseases. Natriuretic peptides, especially natriuretic peptides, tend to be elevated and associated with poor prognosis in patients with heart disease, which is independently thought to be associated with COVID-19 even though the patients have no history of cardiac disease. In this review, the authors summarized the role of biomarkers in determining and diagnosing the extent of involvement of heart damage in people with COVID-19, as well as the permanent damage they may cause in the future. In the review, the researchers divided it into three main sections, considering the diagnosis, prognosis, and mortality in order to simplify the role of cardiac biomarkers in COVID-19 disease. These three sections are as follows: (1) The relationship between cardiac troponin and COVID-19-related myocardial disease; (2) The relationship between natriuretic peptides and COVID-19-related myocardial disease; and (3) The rest of the biomarkers are associated with myocardial disease.

Troponin consists of three main proteins in a complex structure^[4]; Troponin C binds calcium and regulates the work of thin filaments during contraction[5]. Troponin T provides the connection of troponin in a complex with tropomyosin[6]. Troponin I acts as an inhibitory unit, and troponin C prevents contraction in the absence of calcium[7]. The amount of troponin that rises during myocardial injury has been observed to be higher among patients with COVID-19 who died compared to those who survived. In studies concentrating on this subject, a significant relationship has been shown between troponin and mortality with additional patient and hospital-related conditions, even in patients without comorbidities[8,9,10].

BNP is first proteolytically processed from its precursor proBNP to BNP. Afterwards, it is secreted from the heart as N-terminal proBNP (NT-proBNP), undivided proBNP and mature BNP and NTproBNP in ventricular myocytes, and the amount of secretion increases in patients with heart failure [11]. In the review, it is concentrated that the main reason for the increase in natriuretic peptides in severe acute respiratory syndrome coronavirus 2 is some inflammatory processes that can lead to fulminant myocarditis. However, heart damage and hypoxia are thought to be some of the important causes of the increase in natriuretic peptides.

However, the cardiac markers mentioned in the review alone cannot rule out cardiovascular disease. Although it is supported by electrocardiogram (ECG), it may show atypical symptoms. Therefore, the differential diagnosis of acute chest pain after the new types of COVID-19 has become complicated. The viscosity increase due to COVID-19 hypoxia also causes damage to endothelial cells, resulting in increases in coagulation. In cases where biomarkers and ECG are insufficient at this stage, Triple ruleout computed tomography angiography (TRO CTA) provides us an advantage in examining the entire thoracic vascular system and detecting cardiovascular vascular diseases.

One of the important points of the new COVID-19 disease is that this disease has the potential to cause acute presentations. One of the most important of these tables is acute chest pain, which also includes respiratory tract diseases, which is the most common symptom of COVID-19. In these cases, one of the important causes of acute chest pain is diseases that affect the lung parenchyma or accompanying vascular pathologies in COVID-19 cases. In a study conducted in these cases, it is emphasized that TRO CTA is an important diagnostic method that is effective and does not require intervention to the patient in those who apply to the emergency department with sudden onset symptoms[12].

One of the important points apart from acute presentations is the long-term effects of COVID-19. In one study, persistent long-term COVID symptoms such as shortness of breath, chest pain, cough, and muscle weakness were proven to be associated with computed tomography (CT) severity values [13]. In this review, it is also emphasized that the relationship of CT with persistent symptoms yields better data than laboratory parameters. Knowing the relationship between CT severity and long-term COVID symptoms can also help to identify at-risk patients and establish follow-up programs to support these cases

COVID-19 damages the myocardium by various mechanisms. The review focused on multiple viral infections causing sympathetic activation, direct viral invasion and proinflammatory cytokines inducing heart failure. The main reason for the increase in natriuretic peptides increased as a result of myocardial damage is thought to be due to interleukin (IL)-1β and similar proinflammatory cytokines. Magnetic



Resonance Imaging (MRI), which has an important place in the diagnosis of myocarditis, also has an important place in imaging in myocarditis formed in this way; however, Cardiac MRI has some disadvantages such as not being ubiquitous, high cost, claustrophobia, incompatibility with pacemaker and its application due to prostheses. In these cases, in a study conducted, Dual Energy CT (DECT) has proven to be significantly superior to MRI[14].

The authors recommend natriuretic peptides such as troponin and BNP in patients with myocarditis and other myocardial damage mentioned in the study; however, they proved that in addition to traditional biomarkers, new cardiac bioenzymes such as prepesin, soluble ST2 and copeptin also increase and cause marked worsening of prognosis. In addition to biochemical markers, imaging methods, especially CT, have an important place in the diagnosis of myocardial damage and comorbidities in COVID-19 patients. To give an example, DECT is used in practice as an important imaging method in the diagnosis of myocarditis-like conditions with myocardial damage[14]. TRO CTA is a frequently used imaging method in the detection of hypercoagulation, which we can give as an example of comorbidity conditions[12].

In summary, we have seen that there is a significant relationship between COVID-19 and cardiovascular system findings. After understanding this relationship, we learned that we should make the best use of the data we have at the point of diagnosis. Here, we know that we need to proceed to diagnosis by combining many cardiac biomarkers such as laboratory values such as BNP and pro-BNP with imaging methods such as ECG, CT, DECT and TRO CTA. In conclusion, we should make the best use of all available methods for diagnosis and treatment in order to reduce cardiovascular-related mortality and morbidity rates and improve prognosis in these patients.

FOOTNOTES

Author contributions: Aydin S and Gulmez AO conceived the study; Aydin S and Gulmez AO were responsible for designing, materials and supervision; Aydin S and Gulmez AO did the literature search, wrote the manuscript, and reviewed the manuscript critically; All authors have read and approved the final manuscript.

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Country/Territory of origin: Turkey

ORCID number: Ali Osman Gulmez 0000-0001-7050-1765; Sonay Aydın 0000-0002-3812-6333.

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REFERENCES

- Muthyala A, Sasidharan S, John KJ, Lal A, Mishra AK. Utility of cardiac bioenzymes in predicting cardiovascular outcomes in SARS-CoV-2. World J Virol 2022; 11: 375-390 [PMID: 36188743 DOI: 10.5501/wjv.v11.i5.375]
- 2 Peacock WF 4th, De Marco T, Fonarow GC, Diercks D, Wynne J, Apple FS, Wu AH; ADHERE Investigators. Cardiac troponin and outcome in acute heart failure. N Engl J Med 2008; 358: 2117-2126 [PMID: 18480204 DOI: 10.1056/NEJMoa0706824]
- Clerico A, Del Ry S, Giannessi D. Measurement of cardiac natriuretic hormones (atrial natriuretic peptide, brain natriuretic 3 peptide, and related peptides) in clinical practice: the need for a new generation of immunoassay methods. Clin Chem 2000; 46: 1529-1534 [PMID: 11017928 DOI: 10.1007/s40262-019-00764-2]
- Sarko J, Pollack CV Jr. Cardiac troponins. J Emerg Med 2002; 23: 57-65 [PMID: 12217473 DOI: 10.1016/s0736-4679(02)00463-8]
- Parmacek MS, Leiden JM. Structure, function, and regulation of troponin C. Circulation 1991; 84: 991-1003 [PMID: 1884474 DOI: 10.1161/01.cir.84.3.991]
- 6 Raggi A, Grand RJ, Moir AJ, Perry SV. Structure-function relationships in cardiac troponin T. Biochim Biophys Acta 1989; 997: 135-143 [PMID: 2752050 DOI: 10.1016/0167-4838(89)90145-3]
- Cummins P, Perry SV. Troponin I from human skeletal and cardiac muscles. Biochem J 1978; 171: 251-259 [PMID: 417726 DOI: 10.1042/bj1710251]
- Lala A, Johnson KW, Januzzi JL, Russak AJ, Paranjpe I, Richter F, Zhao S, Somani S, Van Vleck T, Vaid A, Chaudhry F, De Freitas JK, Fayad ZA, Pinney SP, Levin M, Charney A, Bagiella E, Narula J, Glicksberg BS, Nadkarni G, Mancini DM,



Fuster V; Mount Sinai COVID Informatics Center. Prevalence and Impact of Myocardial Injury in Patients Hospitalized With COVID-19 Infection. J Am Coll Cardiol 2020; 76: 533-546 [PMID: 32517963 DOI: 10.1016/j.jacc.2020.06.007]

- 9 D'Ardes D, Boccatonda A, Cocco G, Fabiani S, Rossi I, Bucci M, Guagnano MT, Schiavone C, Cipollone F. Impaired coagulation, liver dysfunction and COVID-19: Discovering an intriguing relationship. World J Gastroenterol 2022; 28: 1102-1112 [PMID: 35431501 DOI: 10.3748/wjg.v28.i11.1102]
- 10 Al Abbasi B, Torres P, Ramos-Tuarez F, Dewaswala N, Abdallah A, Chen K, Abdul Qader M, Job R, Aboulenain S, Dziadkowiec K, Bhopalwala H, Pino JE, Chait RD. Cardiac Troponin-I and COVID-19: A Prognostic Tool for In-Hospital Mortality. Cardiol Res 2020; 11: 398-404 [PMID: 33224386 DOI: 10.14740/cr1159]
- Nakagawa Y, Nishikimi T, Kuwahara K. Atrial and brain natriuretic peptides: Hormones secreted from the heart. Peptides 11 2019; 111: 18-25 [PMID: 29859763 DOI: 10.1016/j.peptides.2018.05.012]
- Bahadir S, Aydın S, Kantarci M, Unver E, Karavas E, Şenbil DC. Triple rule-out computed tomography angiography: 12 Evaluation of acute chest pain in COVID-19 patients in the emergency department. World J Radiol 2022; 14: 311-318 [PMID: 36160833 DOI: 10.4329/wjr.v14.i8.311]
- Aydin S, Unver E, Karavas E, Yalcin S, Kantarci M. Computed tomography at every step: Long coronavirus disease. 13 Respir Investig 2021; 59: 622-627 [PMID: 34210624 DOI: 10.1016/j.resinv.2021.05.014]
- Kantarci M, Bayraktutan Ü, Akbulut A, Taydas O, Ceviz N, Güven F, Aydın S. The value of dual-energy computed 14 tomography in the evaluation of myocarditis. Diagn Interv Radiol 2022 [DOI: 10.5152/dir.2022.21749]





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