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## MINIREVIEWS

- 1 Role of cell-free DNA for predicting incidence and outcome of patients with ischemic stroke

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## Role of cell-free DNA for predicting incidence and outcome of patients with ischemic stroke

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### Abstract

Early diagnosis and prognosis of ischemic stroke remains a critical challenge in clinical settings. A blood biomarker can be a promising quantitative tool to represent the clinical manifestations in ischemic stroke. Cell-free DNA (cfDNA) has recently turned out to be a popular circulating biomarker due to its potential relevance for diagnostic applications in a variety of disorders. Despite bright outlook of cfDNA in clinical applications, very less is known about its origin, composition, or function. Several recent studies have identified cell-derived mitochondrial components including mitochondrial DNA (mtDNA) in the extracellular spaces including blood and cerebrospinal fluid. However, the time course of alterations in plasma mtDNA concentrations in patients after an ischemic stroke is poorly understood. DNA is thought to be freed into the plasma shortly after the commencement of an ischemic stroke and then gradually decreased. However, the importance of cell-free mtDNA (cf-mtDNA) in ischemic stroke is still unknown. This review summarizes about the utility of biomarkers which has been standardized in clinical settings and role of cfDNA including cf-mtDNA as a non-invasive potential biomarker of ischemic stroke.

**Key Words:** Ischemic stroke; Mitochondria; Circulating cell-free DNA; Plasma nucleic acid; Mortality

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**Core Tip:** Early and accurate diagnosis of ischemic stroke is critical to achieve favorable clinical outcome. Cell-free DNA can be used as a useful biomarker for early diagnosis and prognosis of ischemic stroke for saving time and increasing the likelihood of successful intervention. Discriminative quantification of cell free mitochondrial DNA instead of overall circulating DNA may provide more significant value for identifying real-time host response. The future practical adoption of this strategy may be aided by reliable and standardized quantification of cell-free mitochondrial DNAs in ischemic stroke patients to design more effective diagnosis, prognosis and therapeutic strategies.

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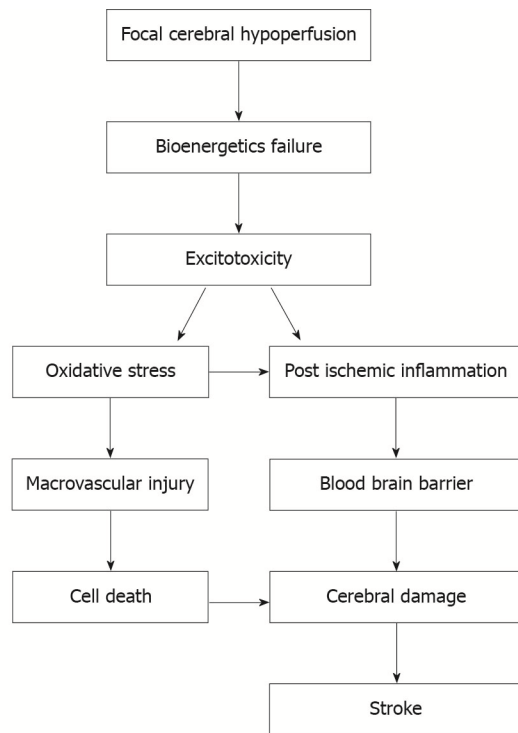
## INTRODUCTION

Stroke is associated with significant morbidity and mortality with continuously increasing incidence globally, owing to the rising prevalence of different forms of cardiovascular diseases[1]. Acute brain injuries, particularly traumatic brain injury, and brain stroke are among the main causes of death and disability worldwide[2]. Currently, stroke has become a major public health burden that is expected to rise in the next decades as a result of demographic transitions, particularly in developing countries[3]. It affects 13.7 million people and kills 5.5 million people per year. Ischemic infarctions accounts for approximately 87% of strokes, a prevalence that increased significantly between 1990 and 2016, owing to lower mortality and improved therapeutic management. Over the same period, the incidence of stroke in low- and middle-income countries increased by two-folds, while it reduced by 42% in high-income countries[4]. According to the Global Cost of Disease Study, the socio-economic burden of stroke has increased over time. The risk of stroke rises to more than 3-fold over the age of 55 years. While, younger adults of age 20 years to 54 years the incidence of stroke has been reported 12.9% to 18.6% of all cases globally between 1990 and 2016. Nonetheless, over the same time period, age-standardized attributable death rates declined by 36.2%[5,6].

Among different types of strokes, ischemic stroke is the most common type that includes cryptogenic, lacunae, and thromboembolic forms of strokes. Ischemic stroke results due to blockage in blood flow to a part of the brain and accounts for approximately 87% of all the strokes. Despite the high rate of morbidity and mortality caused by ischemic stroke, the varied etiology and intricate pathophysiology make clinical diagnosis and prognosis prediction very difficult. Efficient prognosis following an ischemic stroke remains a major obstacle in clinical settings.

Because acute ischemic stroke is often isodense on computed tomography (CT), its initial utility is mainly limited to excluding a haemorrhage[7]. Currently, magnetic resonance imaging (MRI) with diffusion has been considered a gold standard tool for acute diagnosis. However, MRI is expensive and less widely available to rural and general population[7]. The major hitches in cost-effective diagnosis, prognosis, and improved therapeutic efficacy is accompanied by the complicated pathophysiological processes including energy failure, ion homeostasis imbalance, acidosis, intracellular calcium overload, brain excitotoxicity, free radical-mediated lipid oxidation, inflammatory cell infiltration, and glial cell activation (Figure 1). As a result of the combination of aforementioned complicated pathophysiological processes alterations in the blood-brain barrier and the release of different neurological markers into circulation has been reported[8]. In addition, ischemic occlusion causes thrombotic and embolic situations in the brain[9]. The narrowing of veins due to atherosclerosis affects blood flow in thrombosis. Plaque buildup will eventually narrow the vascular chamber and cause clots, resulting in thrombotic stroke. Reduced blood supply to the brain region creates an embolism in an embolic stroke; blood flow to the brain decreases, producing acute stress and premature cell death (necrosis). Following necrosis, the plasma membrane is disrupted, organelles enlarge, and cellular contents leak into the extracellular environment, resulting in loss of neuronal function[8,10].

Owing to the lack of real-time tracking of ischemic stroke pathogenesis and treatment response, the extracellular components released into circulation or secretion may provide an important tool for evolving better approach. Recent studies have demonstrated that circulating cell-free DNA (cfDNA) molecules are significantly increased in stroke condition and act as important tool for non-invasive monitoring of disease progression and prognosis[11-13]. Although various mechanisms of release of cfDNA has been predicted, activated neutrophils have been demonstrated to produce neutrophil extracellular traps (NETs) in response to a variety of stimuli resulting in the release of detectable amount of cfDNA in thrombi. It's also likely that the link between cfDNA levels and neutrophil count is due to activated neutrophils' tendency to leak significant amounts of DNA when edifice NETs[14]. As a result, it is possible that circulating cfDNA in stroke comes from both the damaged Neurovascular Unit



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**Figure 1** Representation showing pathophysiological mechanisms leading to acute ischemic stroke in step-wise manner.

and NET formation, which could explain the link between circulating cfDNA levels and neutrophil count[15]. However, more detailed exploration is required to understand the proper mechanism for the occurrence and progression of ischemic stroke.

## CURRENTLY AVAILABLE AND EMERGING BIOMARKERS OF ISCHEMIC STROKE

Determining the cause of a stroke can be difficult. Based upon the current progress towards diagnostic investigations and classification characteristics, the cause of a stroke is usually unclear or even unknown. Natriuretic peptides, glial fibrillary acidic protein, S100b, neuron specific enolase, myelin basic protein, interleukin-6, matrix metalloproteinase (MMP-9), D-dimer, and fibrinogen have all been examined as biomarkers in stroke. Despite notable research, a troponin-like biomarker to aid in the diagnosis of stroke has been eluded by the researchers. This could be due to the fact that stroke is a heterogeneous disease with a wide range of infarct size, location, and origin[16].

In the case of ischemic stroke, a variety of biomarker panels have been tested for the diagnosis and causation. A panel of five proteins (MMP-9, BDNF, vWF, MCP-1, and S100B)[17], another panel of four proteins (MMP-9, brain natriuretic factor, D-dimer, and S100B)[18], and panel of five proteins (eotaxin, epidermal growth factor receptor, S100A12, metalloproteinase inhibitor-4, and prolactin)[19] are among them. When compared to individual indicators, the panel of numerous markers has consistently exhibited better sensitivity and specificity in diagnosing the ischemic stroke. The results support the concept of merging many markers into a panel, even though none have produced enough evidence to indicate clinical usefulness[20].

Protein assays in plasma or serum have been a frequent method of measuring biomarkers in stroke, and they have provided useful information in the establishment of suitable stroke biomarkers[7,17]. The search for biomarkers has been spurred by the need to better stroke diagnosis and to identify the more appropriate cause. Several markers have shown potential, but there is no enough evidence to warrant their use in clinical practice. The known biomarkers of stroke in patients typically reflect the magnitude of tissue damage and inflammation. A recent genome-wide meta-analysis tried to explore the crucial disease-related pathways and molecular regulatory networks after combining messenger RNA and miRNA expression analyses to identify the candidate target molecule for early diagnosis of stroke[18]. The findings of this study identified six considerably increased genes (PTGS2, IL1B, STAT3, MMP9, SOCS3, and CXCL1), as well as two significantly upregulated miRNAs (miR-320b and miR-320d), as possible clinical diagnostic indicators. These all molecules are linked with the release of extracellular molecules into the circulation and need further investigation.

Although various mechanisms such as oxidative stress, cell death, inflammation and changes in the peripheral blood have been identified, but none have found a place in clinical practice largely because of the stringent criteria that must be met by biomarkers before application. Another systematic review on scalable biomarker combinations for early stroke diagnosis also revealed the need of more comprehensive research on circulatory molecules to evaluate, identify, and systematically implement identified biomarker panels into medical practice for stroke recurrence and diagnosis[19]. Other crucial studies have also suggested that evaluating different blood-based molecules to offer diagnostic accuracy for health interventions is the most promising method. These requirements include the precision, accuracy, sensitivity and specificity of the biomarker for the outcome decided the structured data acquisition protocol and the ease of interpretation (Table 1).

## CIRCULATING CFDNA AS POTENTIAL BIOMARKER OF ISCHEMIC STROKE

Recently, a number of studies have been conducted that examine cfDNA in biological fluids during pathological processes in the human brain[28-31]. The cfDNA is highly fragmented in which released into circulation during apoptosis, cell death, necrosis, inflammation-like conditions. cfDNA contains nuclear DNA and mitochondrial DNA (mtDNA). And it has been thought that cf-DNA would be released into the bloodstream shortly after the ischemic stroke.

Although several biomarkers distinctive to stroke subtypes have been discovered; however, the association of cfDNA as a novel biomarker demands additional investigation into its alliance with stroke subtypes. Such biomarkers could also be used to non-invasively assess stroke severity, which varies between subtypes. The therapeutic window for stroke is limited, occurring only 3-6 h following the onset of symptoms, early recognition of severe patients, and prompt adoption of appropriate therapeutic interventions have enormous prognostic value. In clinical relevance, it could be critical in predicting patient death or functional outcomes, so it is necessary to elucidate further research with stroke. Even though the blood cfDNA levels rise after numerous clinical processes in the body, cfDNA has not traditionally been considered an excellent marker with specificity to a condition like a stroke.

On admission, cfDNA concentrations in the CSF, but not in plasma, are shown to be considerably higher in patients who had poorer outcomes. The diverse origins of cfDNA in both fluids, as well as differences in the dynamics of the blood-brain barrier and CSF-brain barrier integrity changes during a stroke, can explain the observed disparities in cfDNA levels between plasma and CSF at the same time point. In general, the results of different sample types, such as serum, plasma, or CSF, should not be compared[32].

Several studies have demonstrated that cfDNA concentration correlates well with the outcome of stroke intervention in acute ischemic stroke patients (Table 2). In the setting of recanalization, post-intravenous thrombolysis IVT, and mechanical thrombectomy, using cfDNA as a predictive substitute marker to envisage outcome will aid in the most efficient use of limited resources and reduce the load on the healthcare system[30].

Extracellular DNA, on the other hand, has been shown in numerous studies to perform as a hazard signal and drive immune responses. Circulating cfDNA appears to be drawn in endothelium pathophysiological changes in trauma patients, the extent of endothelium injury and an augment in cfDNA release are connected. The cfDNA levels in the blood are also discriminating in relation with greater endothelium damage after cardiac surgery, as well as epithelial and endothelial cell death in the lungs, in a dose-dependent manner[28,33]. Neutrophils use a pathogen-clearance system called NETs, and cfDNA is a key component of these traps. As a result, circulating cfDNA could be both an indicator of the severity of the damage and a contributor in the damage-causing pathways.

Regardless of years of research, the sources and processes of tissue injury that results in high cfDNA levels are only partially understood. Apoptosis and necrosis appear in contribute to circulating cfDNA, however even live cells can release DNA into the circulation under specific conditions. The biological properties of cfDNA are still to some extent understood. To improve the accuracy of stroke diagnosis, cfDNA can be used in concurrence with clinical evaluation and imaging methods[35,36]. It is noted that plasma cf-nucleic acids are increased after acute ischemic stroke and studies also showed correlation with clinical parameters like white blood cell count, diabetes mellitus, hemoglobin A1c, blood pressure. Along with this an increased sample size and follow up with duration since window period assessment may give a noninvasive prognostic implement. It can be used to supplement the diagnostic workup and aid triage patients for intervention as a stroke biomarker. With the addition of this unique marker, it is now possible to make clued-up predictions about the outcome of mechanical thrombectomy or intravenous thrombolysis in acute stroke patients[29]. Patients and their families can make informed decisions in patients with negative imaging results and before consenting to invasive or medicinal treatment using markers like cfDNA assessment with prognostic usefulness.



**Table 1 Various biomarker panels in different stroke condition**

Ref.	Panel	Source of cases	Source of marker	Panel of marker	Sensitivity/specificity (%)
Stamova <i>et al</i> [20], 2010	Gene panel	Ischemic stroke <i>vs</i> control	RNA, whole blood	BCL6, PYGL, RNASE2, S100A9, S100A12, S100P, SLC16A6, ARG1, CA4, CKAP4, ETS-2, HIST2H2AA	93/95
Montaner <i>et al</i> [21], 2008	Protein panel	Cardioembolic <i>vs</i> noncardioembolic stroke	Protein, plasma	D-dimer, BNP	87/85
Laskowitz <i>et al</i> [22], 2009	Protein panel	Ischemic + hemorrhagic stroke <i>vs</i> control	Protein, plasma	BNP, D-dimer, MMP9, S100B	86/37
Jickling and Sharp [23], 2011	Gene panel	Large vessel <i>vs</i> cardioembolic stroke	RNA, Whole Blood	FCRL1, FLJ40125, GRM5, GSTK1, HLA-DOA, IRF6, LHFP, LHFP, LOC284751, LRRC37A3, P2RX5, PIK3C2B, PTPN20A, TFDPI, TMEM19, TSKS, ZNF185, ZNF254, ADAMTSL4, AP3S2	100/96
Barr <i>et al</i> [24], 2010	Gene panel	Ischemic stroke <i>vs</i> control	RNA, whole blood	SCA4, ARG1, S100A12, ORM1, CCR7, CSPG2, IQGAP1, LY96, MMP9, ORM1	-
Montaner <i>et al</i> [25], 2011	Protein panel	Ischemic + hemorrhagic stroke <i>vs</i> control	Protein, plasma	D-dimer, MMP-9, Caspase-3, Chimerin, Secretagogen, sRAGE	17/98
Vanni <i>et al</i> [26], 2011	Protein panel	Ischemic stroke <i>vs</i> control	Protein, whole blood	BNP, D-dimer, MMP9, S100B	86/37
Sharma <i>et al</i> [27], 2014	Protein panel	Ischemic + hemorrhagic stroke <i>vs</i> control	Protein, plasma	S100A12, Metalloproteinase inhibitor-4, Prolactin, Eotaxin, EGFR	90/84

**Table 2 The tabulation of the different stroke conditions of cell-free DNA assessment**

Ref.	Disease specification	Source	Nucleic acid type	Outcome of the study
1 Tsai <i>et al</i> [29], 2011	Acute ischemic stroke	Plasma	cfDNA	Elevated plasma cf-nuclear and mitochondrial DNA in acute ischemic stroke patients than healthy controls
2 O'Connell <i>et al</i> [34], 2017	Ischemic stroke	Plasma	cfDNA	Elevated cfDNA in stroke patients relative to those diagnosed as stroke mimics ( $P = 0.001$ )
3 Naumann <i>et al</i> [28], 2017	Acute ischemic stroke	Plasma	cfDNA	IVT was associated with improved outcome in patients with cfDNA < 10000 kilogenome-equivalents/L ( $P < 0.05$ )
4 Vajpeyee <i>et al</i> [30], 2018	Acute ischemic stroke	Plasma		Higher cfDNA levels were associated with severity at the time of admission ( $P = 0.003$ ) and poor outcome as measured by modified Rankin scale 3-mo scores ( $P = 0.001$ ). Therapeutic intervention mechanical thrombectomy or IV thrombolysis associated with improved outcome in patients with cfDNA < 10000 kilogenome equivalents/L ( $P \leq 0.01$ )
5 Kananen <i>et al</i> [36], 2020	Mortality rate	Plasma	cfDNA	cfDNA associated with increased risk of mortality (hazard ratio 0.1 $\mu$ g increase in cfDNA, $P = 0.0003$ )

cfDNA: Cell-free DNA.

## CFDNA ASSOCIATION WITH MORTALITY IN ISCHEMIC STROKE

Mortality is one of the most important patient-relevant outcomes that have been explored in several earlier studies. Rainer *et al* [37] found a 100% sensitivity and > 74% specificity in using cfDNA as a predictive biomarker for both ischemic and hemorrhagic stroke. A cfDNA level of > 1400 kg-equiv/L suggested a substantial 60% increase in the probability of mortality at 6-mo in categorical analysis. Furthermore, in a 2007 update by the same group, a substantial difference in cfDNA, 48 h after a stroke was found to be a robust predictor of 6-mo mortality, with 50% lower cfDNA levels for individuals who did not have an event [38].

## ROLE OF DIFFERENT TYPES OF CIRCULATING CFDNA

There are two major types of cfDNA namely cell-free nuclear DNA (cf-ncDNA), and cell-free mtDNA



(cf-mtDNA). Recent studies have discovered that in healthy individual's plasma samples, cf-mtDNA can be present in about 50000-fold more copies than cf-ncDNA[39,40]. This indicates that intact cf-mtDNA molecules are present in circulation in higher amount, allowing for the identification and quantification of circulating cf-mtDNA from circulating cf-ncDNA, which could provide more accurate diagnostic information in a variety of physiological and pathological circumstances including ischemic stroke. Because mtDNA is a small circular genome without protective histones, it is more susceptible to breakdown in the circulation, and it is expected that ncDNA and mtDNA will have considerable configuration differences. Hence, the current focus has been on recognizing the potential of cir-mtDNA as a powerful potential biological source in the field of molecular diagnostics and prognosis which is more precise and non-invasive[41].

Recently a few studies have demonstrated the importance of studying cf-mtDNA quantification in various clinical conditions using various samples such as peripheral blood mononuclear cells, whole blood, plasma, urine, and tissue samples[42]. However, extracting pure cell-free cir-mtDNA from plasma samples without contaminating the sample with cir-ncDNA is a significant problem. In this direction, our recent study has reported a fruitful in-house protocol to extract both cf-mtDNA as well as cf-ncDNA using a single plasma sample without contamination of each type of DNA (unpublished). Our data has shown significance of this procedure to clearly identify the potential of quantifying cf-mtDNA to clearly differentiate healthy and diseased individuals at high levels of sensitivity and specificity compared to cf-ncDNA. Apart from our study, a recent report has indicated that studying mitochondrial dynamics may provide a potential therapeutic target for ischemic stroke[43]. However, such investigations are further desired in ischemic stroke to predict the significance of cf-mtDNA as precise, non-invasive diagnostic and prognostic tool.

## KEY FINDINGS AND EXPERT OPINION FOR CF-MTDNA IN ISCHEMIC STROKE

Plasma mtDNA measurement's utility in acute medicine may be enhanced by technological advancements. For example, using a column-based DNA extraction technology and real-time quantitative polymerase chain reaction (RT-qPCR) analysis, plasma mtDNA findings can currently be obtained within 3 h of sampling. Recent advances in quick capillary-based instrumentation for RT-qPCR analysis may allow this period to be cut in half, 30 min to 90 min[43]. The cfDNA is thought to be freed into the plasma shortly after the commencement of an ischemic stroke and then gradually decreased. Circulating cfDNA levels in plasma surged quickly after an acute ischemic stroke and then gradually reduced. The clinical severity of ischemic stroke is reflected in the level of plasma mtDNA in the acute stage. Circulating mtDNA in plasma has been researched in a variety of disorders during the last few decades, including sepsis, cancer, myocardial infarction, and serious trauma. Plasma cfDNA's prognostic and diagnostic value has been recognized in a number of important situations.

## WHAT THIS STUDY ADDS

Ischemic stroke is a major public health problem that is expected to worsen in the next decades as a result of population shifts, particularly in developing nations. Ischemic stroke is linked to major morbidity and mortality, and its prevalence is increasing over the world. Simple cf-mtDNA quantification test in the form of liquid biopsy can assist to estimate likely mortality or functional result when CT and/or MRI are either unavailable or show no obvious acute abnormalities.

It is apparent that cfDNA has clinical utility in predicting functional results and long-term survival. Plasma cfDNA as a prognostic marker has the advantage of being non-invasive and straight forward to use. It has also been shown to be able to distinguish between hemorrhagic and ischemic stroke and to be an independent predictor of stroke outcome in patients with negative neuroimaging. cfDNA can help doctors with patient evaluation and complement imaging technologies to improve stroke diagnostic accuracy. It can also help priorities patients for action by supplementing the diagnostic workup. When imaging is negative or not necessary, the predictive value of cfDNA can help patients and physicians make educated decisions about invasive or medicinal treatment.

## CURRENT NEEDS

The track for biomarkers evolution is prompted by the desire to early and non-invasive diagnosis and prognosis. Although numerous markers have showed promise, there is currently insufficient evidence to warrant their use in clinical practice. Patient biases and blood sample collection have not been taken into account in clinical trials using serum biomarkers. The use of cf-mtDNA as a diagnostic and therapeutic marker for important phases in the ischemic stroke cascade should improve the accuracy of acute stroke diagnosis and provide more reliable stroke prognosis predictions.

## METHODOLOGICAL CONSIDERATIONS FOR QUANTIFYING CFDNA

In some earlier studies, cfDNA was detected in serum and in some in blood plasma. It has long been recognized that cfDNA is more plentiful in serum than in plasma samples, and that the amount of cfDNA in serum varies greatly from patient to patient. The coagulation cascade, which leads to the lysis of white blood cells, may produce a significant amount of cfDNA in the blood serum. As a result, this effect has the potential to induce inaccuracies into the produced results[37]. Thus, data from different sample types, such as serum, plasma, or CSF, should not be compared.

Furthermore, sampling time discrepancies significantly limit the application and coalescence of evidence. As long as the patient is admitted, sampling frequency can range from once to daily. Given that cfDNA is a transient molecule, standardizing the collection time may be advantageous in ensuring that cfDNA is captured within the time frame of rising, peaking, or declining levels. Because the therapeutic window for stroke is small following the onset of symptoms, the timing of diagnosis is critical.

The use of multiple procedures for collecting and quantifying cfDNA is another aspect that restricts the accuracy of biomarker interpretation. For DNA extraction, the majority of the research in our review used the QIAamp circulating nucleic acid kit. The QIA kit is shown to be quite efficient and yields a lot of cfDNA. While the overall goal was to measure cell death, multiple procedures were utilized across investigations, including quantitative PCR, an enzyme-linked immunosorbent assay for cell death detection, and a nucleic acid immunofluorescent counterstain. The present cfDNA assay procedure in stroke is not standardized, and there has been little research into the consistency of the quantitative PCR methods used to quantify cfDNA.

The use of cfDNA as a biomarker for stroke diagnosis and monitoring is hampered by a lack of uniformity and suitable controls. Because of changes in sample processing methods, storage conditions, and extraction and quantification methodologies, the results from different research are not comparable. This can lead to mistakes in setting cut-off points, as well as sensitivity and specificity of assays. As a result, precise and uniform quantification of cfDNA will benefit in the clinical deployment of this strategy in the future.

## CONCLUSION

Circulating cfDNA levels are increased after insult of acute ischemic stroke and correlate with the clinical severity. Considering after thrombolysis and anti-platelets treatment evaluation of cfDNA may provide a crucial evidence to detect the disease severity in earlier phases of the stroke. The cfDNA is a non-invasive, cost-effective and easy to detect using simple procedures. A limited number of studies have shown that cfDNA has predictive significance in providing functional outcomes and hospital mortality. Comparability between experiments is hampered by inconsistencies in DNA extraction and measurement methods. This necessitates the performance of additional strong cohort studies in the future to determine the best collection periods for stroke prediction as well as the best cfDNA processing for the most accurate outcome. Further studies with follow-up and with window period are required to find exact severity and mortality prediction with ischemic stroke.

## FOOTNOTES

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- 10 Possible convulsion and electroencephalographic abnormality in a patient taking long-term oral clarithromycin: A case report

*Shiraishi W*

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## Possible convulsion and electroencephalographic abnormality in a patient taking long-term oral clarithromycin: A case report

Wataru Shiraishi

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### Abstract

#### BACKGROUND

Clarithromycin is a macrolide antibiotic commonly prescribed to patients with upper respiratory and otolaryngological infections. Neuropsychiatric adverse effects of clarithromycin include agitation, insomnia, delirium, psychosis, and seizure.

#### CASE SUMMARY

A 52-year-old man was admitted to our hospital with a convulsion. He had > 10-year history of clarithromycin intake for chronic sinusitis. One week before admission, he started to take diltiazem for angina pectoris. On admission, his convulsion subsided. His electroencephalography showed frontal intermittent rhythmic delta activity. One week after he ceased clarithromycin, his electroencephalographic abnormalities disappeared. We suggested that the patient developed convulsions due to increased blood levels of clarithromycin caused by oral administration of diltiazem, which is involved in CYP3A metabolism.

#### CONCLUSION

Clarithromycin has a relatively high safety profile and is a frequently prescribed drug. However, there are a few previous reports of clarithromycin-related convulsive disorders. Clinicians should be aware of the drug interaction and rare side effects of seizures.

**Key Words:** Clarithromycin; Convulsion; QT interval prolongation; Seizures; Case report

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**Core Tip:** Clarithromycin is a frequently used medication and has a long history of use. The rarity of reports of convulsions induced by clarithromycin suggests that the induction of convulsions by clarithromycin is a rare phenomenon or has been overlooked. Clarithromycin is a frequently prescribed drug but has many interactions. Therefore, we report this case as a cautionary statement for all neurologists. Also, the measurement of QT time may be a valuable method of assessing clarithromycin excess.

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## INTRODUCTION

Although many drugs are known to cause drug-induced seizures, there are very few reports of clarithromycin-induced seizures[1,2]. Clarithromycin is a macrolide antibiotic that is used frequently for upper respiratory tract infections and otolaryngological infections and is considered to be safe[3]. In this case report, we show a case of convulsions with electroencephalography (EEG) abnormalities in a patient who had been taking clarithromycin for over ten years. After the discontinuation of clarithromycin, the EEG abnormality disappeared, and there was no recurrence of convulsive seizures.

In this case, the QTc showed a prolonged tendency in the electrocardiography (ECG) during the convulsions, which may have reflected the excessive clarithromycin state. Thus, clarithromycin-induced epilepsy is sporadic, and we report this case based on a discussion of its pathophysiological mechanisms.

## CASE PRESENTATION

### Chief complaints

Seizure.

### History of present illness

A 52-year-old Japanese male with no past history of seizure was admitted to our hospital with generalized tonic-clonic seizure.

### History of past illness

A detailed interview revealed that the patient had been taking clarithromycin (400 mg/d) for > 10 years to treat chronic sinusitis and had recently added diltiazem (100 mg/d) for hypertension, as prescribed by his family doctor.

### Personal and family history

Nothing particular.

### Physical examination

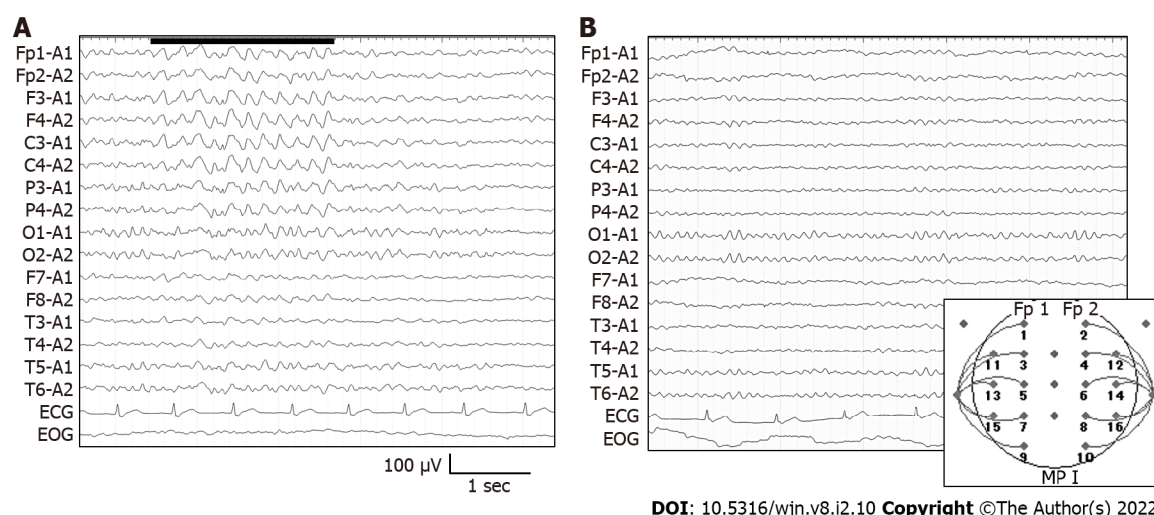
On admission, his seizure subsided. His vital signs showed no remarkable abnormalities. He had bit his tongue, but no incontinence was observed. He was alert and showed no neurological deficits.

### Laboratory examinations

Laboratory examination showed a mild increase of HbA1c (6.6%; normal, 4.6%-6.2%). Since it was a night emergency, the blood concentration of clarithromycin could not be measured. No other abnormalities, including electrolytes, serum creatine phosphokinase, and lactate, were observed. Electroencephalography (EEG) showed frequent frontal intermittent rhythmic delta activities (FIRDA) (Figure 1A). On ECG, his QTc interval was 448 milliseconds (Bazett's formula[4]; normal, < 450 milliseconds[5]).

### Imaging examinations

Head computed tomography, and magnetic resonance imaging revealed no brain abnormalities.



**Figure 1** Electroencephalography on admission and on day 7. A: Frontal intermittent rhythmic delta activities (FIRDA) were detected on admission; B: Electroencephalography at 7 d after the cessation of clarithromycin. FIRDA has disappeared and 10 Hz alpha waves were observed.

## FINAL DIAGNOSIS

We diagnosed clarithromycin-induced seizures and determined that antiepileptic drugs were unnecessary.

## TREATMENT

The cessation of clarithromycin ameliorated the EEG abnormalities. Also, he experienced no seizures to date.

## OUTCOME AND FOLLOW-UP

His EEG showed a normal pattern without FIRDA (Figure 1B), and his QTc interval was 385 milliseconds. Medications other than clarithromycin were continued. We diagnosed clarithromycin-induced seizures and determined that antiepileptic drugs were unnecessary.

## CONCLUSION

Clarithromycin has often been reported to cause psychiatric symptoms such as delirium[6] and manic episodes[7], suggesting that clarithromycin may sometimes show central nervous system side effects. However, the clarithromycin-induced seizure is quite rare, with few previous reports[1,2]. Clarithromycin is reported to increase neuronal activity, stimulate CA3 pyramidal neurons of the hippocampus by reducing GABAergic signaling, and cause neuronal hyperexcitability[8]. However, the detailed mechanism of these effects is unknown, and further study is needed. Long-term macrolide usage is common for bronchiectasis, chronic obstructive pulmonary disease, and chronic rhinosinusitis[3]. Our patient had been prescribed clarithromycin for chronic sinusitis for > 10 years. Liver CYP3A4 has been reported to metabolize clarithromycin[2]. In the present case, diltiazem was added as an antihypertensive just before the seizure. Diltiazem is also reported to inhibit CYP3A[9]. Thus, in the present case, the introduction of diltiazem may have increased the blood concentration of clarithromycin. Administering azithromycin instead of clarithromycin may be an option. Clarithromycin also causes QT interval prolongation. Our case showed a QTc time of 448 milliseconds at admission, and after discontinuation of clarithromycin, his QTc time improved to 385 milliseconds. Measurement of QT time may be a valuable method of assessing clarithromycin excess. Clarithromycin is a frequently used medication and has a long history of use. The rarity of reports of convulsions induced by clarithromycin suggests that the induction of convulsions by clarithromycin is a rare phenomenon or has been overlooked. Clarithromycin is a frequently prescribed drug but has many interactions. Therefore, we report this case as a cautionary statement for all neurologists.

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## FOOTNOTES

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